

T-JUNCTION SPLITTING OF DROPLETS FROM NANOLITER TO FEMTOLITER AND MANIPULATION OF SINGLE NANOPARTICLES ON MICROFLUIDIC CHIPS

Xuan Feng, Ying Yi, Dai-Wen Pang, Zhi-Ling Zhang*

Key Laboratory of Analytical Chemistry for Biology and Medicine (Ministry of Education)
College of Chemistry and Molecular Sciences, Wuhan University, Wuhan 430072, P. R. China

ABSTRACT

In this paper, we got femtoliter volume droplet by splitting of nanoliter droplet at a T-junction channel, and the volume of the daughter droplet was not limited by the channel size. We use this droplet splitting system to entrap several fluorescent nanospheres, which show great potential to manipulate and observe single nanoparticles independently. The influences of fluid flow rate, channel geometry, addition of surfactant on droplet splitting were carefully studied. The droplet splitting in our T-junction chip has a small volume, monodisperse size, and can be produced efficiently, orderly, controllably.

KEYWORDS: Microfluidic Chips, Droplets, Femtoliter

INTRODUCTION

Carrying out chemical and biochemical reactions in compartmentalized containers on droplet-based microfluidic chip has shown great advantages, such as saving reagent, fast, independent and high-throughput[1]. Microreactor containing several molecules or particles offer opportunities for discovering and characterizing new chemical and biochemical phenomena and can greatly improved signal/background ratio in femtoliter-volume droplets[2]. As we know, sizes of water-in-oil droplets always scale with microchannel dimensions in the range 10-100 μm . Liu et al. first reported of a microfluidic platform for splitting off daughter droplets ranging in volumes of 34-523 fL (10^{-15} L) from mother water-in-oil plugs, where the sizes of the droplets were not limited by the channel width of the device[3]. Tang et.al described a novel method of generating monodisperse aqueous droplets with volumes down to 200 aL (10^{-18} L) by means of piezoelectric injection, and use optical tweezers for droplets manipulation and mixing[4].

EXPERIMENTAL

Microfluidic chip design and fabrication

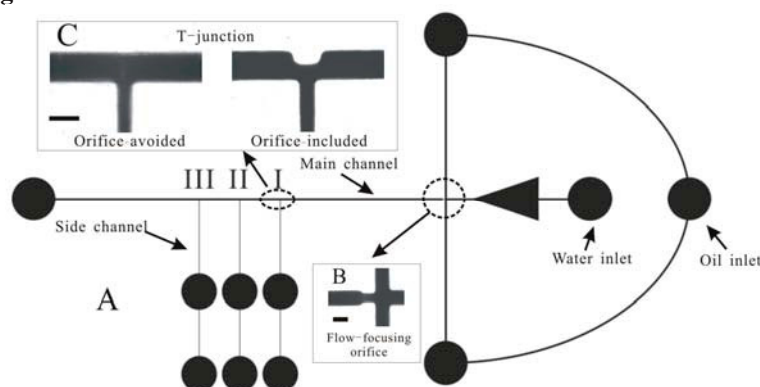


Figure 1. Schematic of microfluidic chip. (A) Schematic of the whole chip. (B) The image of flow-focusing regions with orifices, the orifice was 50 μm in length and 30 μm in width. (C) Schematic of orifice-avoided and orifice-included T-junction channel. Scale bar was 50 μm .

In our work, we designed a microfluidic chip containing a flow-focusing main channel for generation of nanoliter-volume droplet (water-in-oil, mother droplet), and three T-junction side channels for splitting of femtoliter-volume daughter droplet from mother droplet, as shown in Fig. 1A. The main channels are 50 μm in width, and 30 μm in height, the side channels are 25 μm in width. For the orifice-included T-junction chip, the orifice is 50 μm in length, and 25 μm in width, which can obviously facilitate the splitting of mother droplet, and reduce the daughter droplet volume compared with the orifice-avoided T-junction channel.

Soft-lithography was used to fabricate the PDMS chips. Briefly, the designed structure printed on a transparent film was transferred onto a silicon wafer (Institute of Microelectronics of Chinese Academy of Sciences) with 30 nm thick AZ 50 XT photoresist (AZ Electronic Materials USA Corp, USA). After casting about 5 nm thick poly-(dimethylsiloxane) (PDMS, RTV615 GE Toshiba Silicones Co. Ltd) with a 10 : 1 polymer/curing agent ratio, onto the silicon master and waiting until the bubbles disappeared, PDMS was cured at 75 $^{\circ}\text{C}$ for 3 h. Then the cured PDMS was peeled off from the master, and inlet and outlet holes were drilled by a metal pipe. After exposed to a plasma treatment for 1 min, the PDMS was bonded to a glass slide on which a 50 nm thick PDMS film was coated previously. The PDMS chips were baked at 120 $^{\circ}\text{C}$ overnight to get the hydrophobic channels.

Formation of droplets

The two flow stream, soybean oil and water were pumped constantly by syringe pumps (pump 11 Pico Plus, Harvard Apparatus, USA) into the well-fabricated PDMS hydrophobic PDMS chips, in which soybean oil was used as the continuous phase to wrap water. The flow behavior was observed on an inverted fluorescence microscope (TE2000-U, Nikon Corp, Japan), coupled with an chargecoupled device (CCD, Retiga 2000R, Qimaging Corp, Canada) for the recording process.

RESULTS AND DISCUSSION

At the first T-junction, the mother droplet was split into two daughter droplets, the smaller one was driven into the side channel, and the bigger one continued move forward in the main channel (as shown in Figure 2), and at the second T-junction, it was split again, so was the third T-junction.

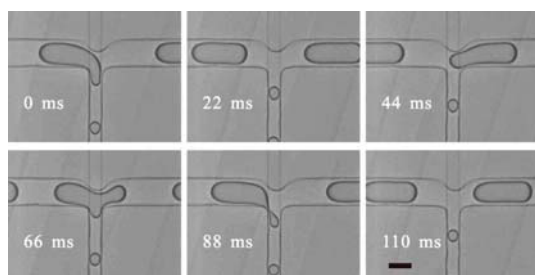


Figure 2. A single period of splitting of daughter droplet from mother droplet

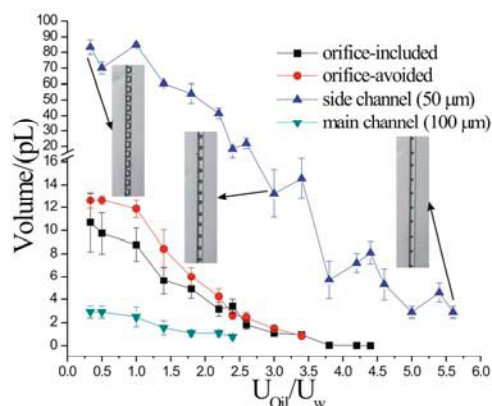


Figure 3. Influence of microchannel geometry on droplet splitting

We studied influences of four different channel geometry on droplet splitting, including orifice-avoided T-junction channel, orifice-included T-junction channel, main channel of 100 μm , side of 50 μm . We found that the introduction of orifice into the T-junction region can obviously reduced the volume of daughter droplet, the smallest volume of daughter droplet generated at the first orifice-included T-junction was 6.5 fL. As the increase of the U_{oil}/U_w (U_{oil} —flow rate of soybean oil, U_w —water), the volume of the daughter droplet decreased (as shown in Figure 3).

As the introduction of surfactant, the droplet volume can be reduced to 0.65 fL, also it can prevent small droplet fusing (as shown in Figure 4). We use this droplet splitting system to entrap several fluorescent nanospheres, which show great potential to manipulate and observe single nanoparticles independently (as shown in Figure 5).

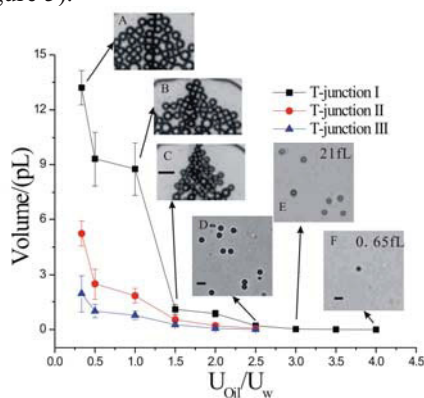


Figure 4. Influence of introduction of surfactant on droplet splitting

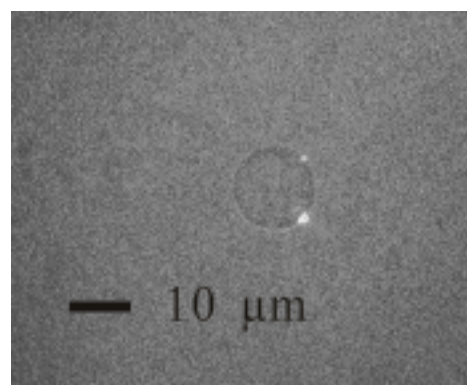


Figure 5. Manipulation of fluorescent nanospheres

CONCLUSION

In our work, we generated femtoliter droplets by droplet splitting from nanoliter ones, and studied influences of fluid flow rate, channel geometry, addition of surfactant on droplet splitting. The smallest volume of droplets can be reduced to 0.65 fL by addition of surfactant. Using this droplet splitting system, we realized entrapment of single fluorescent nanospheres.

ACKNOWLEDGEMENTS

We thank the support from the National Basic Research Program of China (973 Program, 2007CB714507), the Science Fund for Creative Research Groups (20621502), the National Natural Science Foundation of China (20875072), and the Ministry of Public Health (2009ZX10004-107 and 2008ZX10004-004).

REFERENCES

- [1] F. Microtas and S.A. Fluidic, The principles of manuscript writing, Proc. Micro Total Analysis Systems 2000, Kluwer Academic Publisher, Dordrecht, The Netherlands, pp. 1000-1001, (2000).
- [1] S. Y. Teh, R. Lin, L. H. Hung and A. P. Lee. Droplet microfluidics. Lab on a Chip, pp. 198-220, (2008).
- [2] T. Li, L. Su, W. P. Hu, H. L. Dong, Y. F. Li and L. Q. Mao. Femtoliter and Attoliter Electrochemical Cells on Chips. Analytical Chemistry, pp. 1521-1526, (2010).
- [3] Y. Liu, S. Y. Jung and C. P. Collier. Shear-Driven Redistribution of Surfactant Affects Enzyme Activity in Well-Mixed Femtoliter Droplets. Analytical Chemistry, pp. 4922-4928, (2009).
- [4] J. Y. Tang, A. M. Jofre, R. B. Kishore, J. E. Reiner, M. E. Greene, G. M. Lowman, J. S. Denker, C. C. C. Willis, K. Helmerson and L. S. Goldner. Generation and Mixing of Subfemtoliter Aqueous Droplets On Demand. Analytical Chemistry, pp. 8041-8047, (2009).

CONTACT

* Zhi-Ling Zhang, tel: +86 27 68756759; zlzhang@whu.edu.cn