## **Electronic Supplementary Information (ESI)**

Functionalized  $Ni@SiO_2$  core/shell magnetic nanoparticles as a chemosensor and adsorbent for  $Cu^{2+}$  ion in drinking water and human blood

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**Preparation of compound 4.** A solution of **3** (3g, 0.029mol) and 3-(triethoxylyl) propyl-isocyanate (14.38ml, 0.058mol) was reacted in anhydrous methylenechloride stirred for overnight at 0 °C. The solvent was removed under vacuum.(86.65%) <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>-d<sub>6</sub>)  $\delta$  = 6.02 (s, 4H; NH), 3.8 (m, 12H; CH<sub>2</sub>), 3.58 (t, <sup>3</sup>J (H,H) = 6.78 Hz, 2H; CH<sub>2</sub>), 3.28 (m, 6H; CH<sub>2</sub>), 2.8(m, 4H; CH<sub>2</sub>), 1.6 (m, 4H; CH<sub>2</sub>), 1.21 (m, 18H; CH<sub>2</sub>), 0.7 (t, <sup>3</sup>J (H,H) = 8.14 Hz, 4H; CH<sub>2</sub>), <sup>13</sup>C NMR (CDCl<sub>3</sub>-d<sub>6</sub>)  $\delta$  =160, 59, 48, 46, 40, 23, 18, 7.4 . IR (KBr, cm<sup>-1</sup>): 3350, 2976, 2893, 2361, 1885, 1641, 1564, 1271, 1084, 954, 782, 473. HRMS (FAB<sup>+</sup>) m/z 597.3589 [(M+H)<sup>+</sup> calcd for C<sub>24</sub>H<sub>55</sub>N<sub>5</sub>O<sub>8</sub>Si<sub>2</sub>:598.4301]. Anal. calcd for C<sub>24</sub>H<sub>55</sub>N<sub>5</sub>O<sub>8</sub>Si<sub>2</sub>: C, 48.21; H, 9.27; N, 11.71. found: C, 49.23; H, 11.02; N, 11.25.

**Preparation of compound 2.** A solution of **4** (2.13 g, 0.0035mol) toluene at 120 °C was added to 4-chloro-7-nitrobenzo[c][1,2,5]oxadiazole (0.85ml, 0.0042mol). The reaction mixture was stirred for 2 hour at 120 °C, and then cooled to room temperature. The solvent was removed under vacuum, and the residue was purified by silica gel column chromatography (methanol/methylenechloride = 1:20) to afford yellow solid. (22.5 %): <sup>1</sup>H NMR (300 MHz, DMSO-d<sub>6</sub>) δ = 8.5 (d, <sup>2</sup>J (H,H) = 8.78 Hz, 1H; Ar-H), 6.5 (d, <sup>2</sup>J(H,H) = 8.9Hz, 1H; Ar-H), 6.02 (s, 4H; NH), 3.8 (m, 12H; CH<sub>2</sub>), 3.65(m, 8H; CH<sub>2</sub>), 3.38 (t, <sup>3</sup>J (H,H) = 5.98 Hz, 4H; CH<sub>2</sub>), 1.6 (m, 4H; CH<sub>2</sub>), 1.25 (m, 18H; CH<sub>3</sub>), 0.6 (t, <sup>3</sup>J (H,H) = 6.72Hz, 4H; CH<sub>2</sub>), <sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ = 160, 144, 140.2, 138, 131, 122.2, 98.1, 58, 56.4, 45.6, 39.1, 24.8, 18.4, 7.4 . IR (KBr, cm<sup>-1</sup>): 3418, 2930, 2846, 2335, 1663, 1072, 786, 645, 451. HRMS (FAB<sup>+</sup>) m/z 760.3607 [(M+H)<sup>+</sup> calcd for C<sub>30</sub>H<sub>56</sub>N<sub>8</sub>O<sub>11</sub>Si<sub>2</sub>: 761.3641]. Anal. calcd for C<sub>30</sub>H<sub>56</sub>N<sub>8</sub>O<sub>11</sub>Si<sub>2</sub>: C, 47.35; H, 7.42; N, 14.72. found: C, 48.98; H, 9.58; N, 14.23.

**Preparation of nickel nanoparticles.** A solution of poly(vinyl pyrrolidone) (5.35 g,  $9.73 \times 10^{-5}$  mol, Mw = 55,000) and Ni(acetylacetonate)<sub>2</sub> (1.0 g,  $3.89 \times 10^{-3}$  mol) in 1,5-pentanediol (50 mL) was refluxed at 200 °C for 4 h, and the reflux was continued at 240

<sup>o</sup>C for another 1 h, and then the reaction mixture was cooled in an ice–water bath. The product was separated by centrifugation at 12000 rpm for 20 min and was washed with ethanol by a repetitive dispersion/precipitation cycle in ethanol (100 mL) to remove excess poly(vinyl pyrrolidone). Monodisperse nickel nanoparticles were obtained via a careful size selection in 40% yield.

**Preparation of Ni@SiO<sub>2</sub> core/shell particles:** 1.0 mL of ammonium hydroxide (28% in water, Junsei) in 1.0 mL of water was added to the nickel colloidal solution ( $6.20 \times 10^{-2}$  M with respect to the original nickel precursor concentration). After 30 s, 0.1 mL of tetraethyl orthosilicate (TEOS) and 0.1 mL of octadecyltrimethoxy-silane (C<sub>18</sub>TMS, 9%) were added, and the mixture was vigorously stirred for 1 h to ensure complete polymerization of silica. The product was separated by centrifugation at 6000 rpm for 10 min. Ni@SiO<sub>2</sub> core/shell particles with a shell thickness of 40 nm were chosen for **1** synthesis.

**Preparation of nitrobenzofurane-functionalized core/shell magnetic nanoparticles 1.** Compound **2** (100 mg, 0.054 mmol) was dissolved in anhydrous toluene (5 mL) to which Ni@SiO<sub>2</sub> core/shell particles (100 mg) was added, and it was stirred under reflux in N<sub>2</sub> for 24 h. The collected solid was washed several times with dichloromethane and acetone to rinse away any excess **2** and then dried under vacuum.

**Characterization.** <sup>1</sup>H and <sup>13</sup>C NMR spectra were measured with a Bruker ARX 300 MHz sepctrometer. MS spectra were obtained with a JEOL JMS-700 mass spectrometer. Transmission electron microscopy (TEM) images were taken with a JEOL JEM-2100 F instrument operated at 150 keV. IR spectra were obtained for KBr pellets, in the range 400–4000 cm<sup>-1</sup>, with a Shimadzu FTIR 8400S instrument, and the MS spectrum was obtained with a JEOL JMS-700 mass spectrometer. Time-of-flight Second Ion Mass Spectrometer (TOF-SIMS) was analyzed on Model PHI 7200 equipped with Cs and Ga ion guns for positive and negative ion mass detection.

**Photospectroscopy.** Fluorescence emission spectra were recorded with a Shimadzu RF-5301-PC instrument. Stock solutions (0.01 M) of the hydrated metal perchlorate salts were prepared in H<sub>2</sub>O at pH 7. Stock solutions of **1** were prepared in H<sub>2</sub>O. For all measurements, excitation was at 350 nm, with excitation and emission slit widths of 1.5 nm. The pH value was adjusted by using 0.2 M MOPS. Fluorescence quantum yields were determined by reference to methylene blue ( $\Phi = 0.04$ ).<sup>1</sup>

Separation of  $Cu^{2+}$  from drinking water. The column (1 cm x 5 cm) was prepared by using core/shell nanoparticle 1. The column as adsorbent was connected before analytical column in ion chromatography. Then,  $Cu^{2+}$  contained drinking water was injected. The eluent was flowed with 1.0 ml/min into columns. The solution was analyzed by ICP-MS (ELAN DRC II, Perkin Elmer) and Ion chromatography.

## References

1. J. Olmsted III, J. Phy. Chem., 1979, 83, 2581.



Scheme S1. Synthetic route of 2.



Fig. S1 FT-Infrared spectra of (a)  $Ni@SiO_2$  and (b) 1.



Fig. S2 TOF-SIMS spectrum of 1.



Fig. S3 Job's plot of 1:1 complex of receptor 2 attached onto 1 and  $Cu^{2+}$ .



Fig. S4 Time course of the fluorescence intensity of 1 (1.0  $\mu$ M) in water at pH 7 at the addition of Cu<sup>2+</sup> (10.0 equiv).



**Fig. S5** Fluorescence intensity of **1** (1.0  $\mu$ M) with Cu<sup>2+</sup> (10.0 equiv) in the presence of other metal ions such as Na<sup>+</sup>, Ca<sup>2+</sup>, Cu<sup>2+</sup>, Ag<sup>+</sup>, Zn<sup>2+</sup>, Mn<sup>2+</sup>, Hg<sup>2+</sup>, Cu<sup>+</sup> and Pb<sup>2+</sup> ions (10.0 equiv) in water at pH 7.



Fig. S6 Fluorescence changes of 2 (1.0  $\mu$ M) upon the addition of metal ions (10.0 equiv) in acetonitrile.



Fig. S7 Plot of pH values against fluorescence intensity of 1 (1.0  $\mu$ M) in the (a) absence and the presence of (b) Hg<sup>2+</sup> (10.0 equiv) in water.