A novel di-triazole based peptide as a highly sensitive and selective fluorescent chemosensor for Zn²⁺ ions

General Experimental Methods

All the reactions were monitored by employing TLC technique using appropriate solvent system for development. Reactions involving air/oxygen sensitive reagents or catalysts were performed in degassed solvents. Transfer of moisture sensitive materials were carried out in a glove box, using standard syringe-septum techniques and the reactions were maintained under nitrogen atmosphere until the work up. Yields reported are isolated yields of the materials. All the commercial reagents were used as such without further purification. Infrared (IR) spectra were recorded on Nicolet Impact-400 FT IR spectrometer in KBr. Proton Nuclear Magnetic Resonance (400 MHz, ¹H NMR) spectra and Carbon Nuclear Magnetic Resonance (100.6 MHz, ¹³C NMR) spectra were recorded on Varian spectrometers. The high-resolution mass measurements were carried out using JEOL JMS-DX 303 GC-MS instrument or Micro mass Q-Tof spectrometer. Melting points were recorded on Buchi B-545. The steady state absorption spectra have been recorded on a JASCO V530 spectrophotometer and steady state fluorescence spectra are recorded on a Varian Cary with Eclipse fluorescence spectrophotometer $\lambda_{ex} = 260$ nm, with a band width of 5 nm. The spectral studies have been performed using 2µM of the ligand solution in acetonirile, HPLC grade (Spectrochem, Mumbai, India). Before performing the fluorescence studies the purity of the solvent has been assured by exciting the sample in different spectral regions, where no fluorescence has been observed. Time resolved fluorescence studies have been performed in a time correlated single photon counting (TCSPC) system, from IBH, UK, with λ_{ex} =266 nm. The full width at half maximum of the instrument response function is 840 ps. The PL decays are collected with emission polarizer at a magic angle 54.7° and is analyzed by using IBH DAS 8.2 software.

Synthesis and characterization of compounds 5, 6 and 7

Synthesis of Ac-Dprg-OH 5

To a solution of ester **4** (1.2 g, 5.8 mmol) in MeOH (20 mL) was added 2N NaOH (696 mg, 17.4 mmol) and the reaction mixture was stirred at rt for 12 h. The reaction mixture was concentrated, diluted with water (20 mL), then

acidified with 1N HCl and extracted with ethyl acetate. Evaporation of the solvent gave 39 (1.0 g, 95%) as a white solid which was used in the next step without further purification.

Rf : 0.25 (50% ethyl acetate:petroleum ether); m.p.: 138-140 °C; ¹H NMR (300 MHz, CD₃OD): δ = 1.96 (s, 3H), 2.38 (t, *J* = 2.68 Hz, 2H), 2.86-2.98 (m, 4H) ppm; ¹³C NMR (100 MHz, CD₃OD): δ = 22.5, 25.3, 61.6, 72.9, 79.4, 172.9, 173.8; I.R. (KBr pellet): 1667.9, 1730.3, 2329.3, 3304.4, 3362.2 cm⁻¹; HRMS (QTOF): Calcd. for C₁₀H₁₂NO₃ [M+H]⁺ 194.0817, found at 194.0816.

Synthesis of Ac-Dprg¹-(L)Leu²-OMe 6

To a solution of acid **5** (310 mg, 1.61 mmol) and HOBt (433 mg, 3.21 mmol) in dry THF (10 mL) was added DCC (397 mg, 1.93 mmol) at 0 °C. Then, H-Leu-OMe.HCl (350 mg, 1.93 mmol) and NMM (130 mg, 1.30 mmol, reaction mixture should have

around pH= 9) in THF (10 mL) solution was added. The reaction mixture was stirred at rt for 24 h. The solvent was evaporated and the residue was diluted with water. The aqueous layer was extracted with ethyl acetate (3 x 10 mL). The combined organic layer was washed with water, brine and dried over Na₂SO₄. Evaporation of the solvent gave the crude product, which was purified by column chromatography (30% EtOAc/petroleum ether) to give the dipeptide **6** (440 mg, 86%) as a white solid.

Rf : 0.29 (50% ethyl acetate/ petroleum ether); m.p.: 137-139 °C; ¹H NMR (400 MHz, CDCl₃): 0.93-0.96 (m, 6H), 1.64-1.68 (m, 3H), 2.07 (s, 3H), 2.10-2.15 (m, 2H), 3.01-3.20 (m, 4H), 3.74 (s, 3H), 4.59-4.65 (m, 1H), 6.55 (s, 1H), 7.15 (d, J = 7.8 Hz, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 22.1$, 22.9, 24.1, 24.9, 25.1, 41.7, 51.6, 52.5, 60.8, 72.5, 72.7, 79.0, 170.2, 170.5, 173; I.R. (KBr pellet): 1670.1, 1741.4, 2304.9 cm⁻¹; HRMS (QTOF): Calcd. for C₁₇H₂₅N₂O₄ [M+H]⁺ 321.1814, found at 321.1802; [α]_D²⁵: - 4.76(c = 0.51, CHCl₃).

Synthesis of di-trizole based peptide 7



.COOMe

AcHN

The alkyne precursor **6** (30 mg, 0.09 mmol) was dissolved in ^tBuOH/H₂O (5:5 mL) and the *p*-methoxyphenyl azide (27.9 mg, 0.19 mmol), Cu(OAc)₂ (3.7 mg, 0.02 mmol) and sodium ascorbate (7.4 mg, 0.04 mmol) were added. The



resulting mixture was stirred at rt for 24 h, until TLC indicated completion of reaction. The mixture was diluted with ethyl acetate and washed with aq NH₄OH (0.2%) and brine. The aqueous phases were extracted with ethyl acetate (2 x 10 mL). The combined organic extracts were dried (Na₂SO₄) and concentrated in vacuo. The crude product was purified by flash chromatography on silica gel with 100% ethyl acetate gave the desired peptide as white solid in 99% (57 mg) yield.

Rf : 0.16 (100% ethyl acetate); m.p. 210-213 °C; ¹H NMR (400 MHz, CDCl₃): δ = 0.88-0.92 (m, 6H), 1.57-1.64 (m, 2H), 2.07 (bs, 1H), 2.08 (s, 3H), 3.40-3.68 (m, 4H), 3.69 (s, 3H), 3.87 (s, 6H), 4.41-4.43 (m, 1H), 7.00-7.02 (m, 4H), 7.35 (bs, 1H), 7.64-7.70 (m, 4H), 8.04 (s, 1H), 8.08 (s, 1H), 8.19 (d, *J* = 6.8 Hz, 1H) ppm; ¹³C NMR (100 MHz, CDCl₃): δ = 20.8, 21.9, 22.9, 24.6, 25.1, 30.6, 31.1, 40.9, 51.8, 52.3, 55.8, 64.0, 114.8, 114.9, 122.1, 122.1, 122.5, 122.7, 130.6, 130.7, 142.9, 142.4, 159.9, 159.9, 172.3, 172.4, 173.6; I.R. (KBr pellet): 1685.1, 1736.3, 2928.7, 3054.7 cm⁻¹; HRMS (QTOF): Calcd. C₃₁H₃₉N₈O₆ [M+H]⁺ 619.2993, found: 619.2994; [α]_D²⁵: - 7.494 (c = 0.17, CHCl₃).



MHz Hz

Fig. S1 ¹H NMR (CD₃OD) of compound **5**.



Fig. S2 13 C NMR (CD₃OD) of compound **5**.



Fig. S3 1 H NMR (CDCl₃) of compound **6**.



Fig. S4 13 CNMR (CDCl₃) of compound **6**.







Fig. S6 13 CNMR (CDCl₃) of compound **7**.



Fig. S7 ¹H NMR (CD₃CN) spectra of compound **7** (10 mM solution in CD₃CN).



Fig. S8 ¹H NMR spectra of 7 (10 mM) in the presence of 0.5 equiv. of Zn^{2+} in CD₃CN



Fig. S9 ¹H NMR spectra of **7** in the presence of Zn^{2+} in CD₃CN/D₂O (To identify NH-protons, deuterium exchange).



Fig. S10 Absorption spectra of alkyne building block 6 in acetonitrile before (black) and after addition of 6.6 μ M of Zn(ClO₄)₂ (green).



 λ exc. = 282 nm

Fig. S11 Fluorescence spectra of alkyne building block 6 in acetonitrile before (black) and after addition of 6.6 μ M of Zn(ClO₄)₂ (green).



Fig. S12 Fluorescence titration in different acetonitrile-water percentage.



Fig. S13 Absorption spectra for L=7 as a function of pH (pH solutions were made in water, experimental solution was prepared by taking 20 μ L of (200 μ M solution of 7 in acetonitrile) and then volume make up to 2000 μ L by various pH solutions).



Fig. S14 Fluorescence intensity for L=7 as a function of pH (pH solutions are made in water, experimental solution was prepared by taking 20 μ L of (200 μ M solution of 7 in acetonitrile) and then volume make up to 2000 μ L by various pH solutions.



Fig. S15 Absorption spectra for L-Zn²⁺ as a function of pH (pH solutions are made in water, experimental solution was prepared by taking 20 μ L of (200 μ M solution of 7 in acetonitrile), 15 μ L of (200 μ M solution of Zn(ClO₄)₂ in acetonitrile) and then volume make up to 2000 μ L by various pH solutions.



Fig. S16 Fluorescence intensity for $L-Zn^{2+}$ as a function of pH (pH solutions are made in water, experimental solution was prepared by taking 20 µL of (200 µM solution of 7 in acetonitrile), 15 µL of (200 µM solution of $Zn(ClO_4)_2$ in acetonitrile) and then volume make up to 2000 µL by various pH solutions.



Fig. S17 ¹H NMR titration spectra of (i) **7** (10 mM); (ii) **7** in the presence of 1.23 mM of Zn^{2+} ; (iii) **7** in the presence of 2.46 mM of Zn^{2+} (iv) **7** in the presence of 4.92 mM of Zn^{2+} (0.5 equiv.) (v) **7** in the presence of 7.38 mM of Zn^{2+} (vi) **7** in the presence of 9.84 mM of Zn^{2+} (vii) **7** in the presence of 19.68 mM of Zn^{2+} (viii) **7** in the presence of 24.6 mM of Zn^{2+} in CD₃CN.



Fig. S18 Mass spectral peak observed for the 2:1 complex of **7** and Zn^{2+} .

Table S 1. Change in the fluorescece anisotropy of **7** with the increase in the concentration of Zn^{2+} .

[ZnClO ₄]/µM	r
8.69	0.025
16.66	0.017

Table S2. Fluorescence lifetime of -7 complex in presence of various concentrations of $Zn(ClO_4)_2$.

[Zn ²⁺] (μM)	τ ₁ (ns)	χ2
1.6	1.87	1.18
4.8	2.09	1.08
9.4	2.04	1.01
17.9	2.02	1.02
25.8	2.06	1.07