Sensing of glycoprotein via a biomimetic sensor based on molecularly imprinted polymers and graphene-Au nanoparticles

Xindong Wang ^a, Jing Dong ^{a,b}, Huami Ming ^a, Shiyun Ai ^{a *}

a College of Chemistry and Material Science, Shandong Agricultural

University, Taian, Shandong, 271018, China

b College of Resources and Environment, Shandong Agricultural University, Taian, Shandong, 271018, China

* Corresponding authors.

Tel: +86 538 8249248

Fax: +86 538 8242251

E-mail address: ashy@sdau.edu.cn (S.Y. Ai)



Fig. S1 CVs of the electropolymerization on bare GCE (A), NIP/Gr-AuNPs/GCE (B), MIP/Gr-AuNPs/GCE (C) for 20 cycles in ABS (pH 5.0). The voltage range: 0.0 to 0.8 V; scan rate: 0.1 V/s; OPD: 5 mM; ABA: 10 mM.



Fig. S2 The DPV curves for NIP/Gr-AuNPs/GCE (a), NIP/Gr-AuNPs/GCE with 6-ferrocenylhexanethiol (b), and MIP/Gr-AuNPs/GCE with 6-ferrocenylhexanethiol (c).

In Fig.S2, it shows obviously that the peak current of curve a is almost completely invisible. We use I_1 , and I_2 to represent the peak current of curve b and curve c, respectively, and $\triangle I$ was used to denote the difference of I_1 and I_2 as follows: $\triangle I = I_2 - I_1$. So $\triangle I$ correlates with the concentration of BSA, and eliminates the interference of physically absorption of probe moleculars in the polymer film.



Fig. S3 The effect of monomers molar ratio on the response current



Fig. S4 The effect of the number of potential cycles on the response current



Fig. S5 The effect of accumulation time on the response current



Fig. S6 calibration plots of MIP and NIP.

BSA standard solutions (g/mL)	1.0×10 ⁻⁵	5.0×10 ⁻⁶	1.0×10 ⁻⁶	5.0×10 ⁻⁷
(I) Response currents (μA) ^a	3.151	3.022	2.738	2.620
(II) Response currents (μA) b	2.923	2.802	2.547	2.436
(III) Response currents shift (%) ^c	7.236	7.280	6.976	7.023

Table S1 Regeneration experiments of the sensors

^a: BSA were detected by new prepared sensors.

^b: BSA were detected by regenerated sensors. ^c: III = (I – II) / I × 100%.