

**A New Chiral C₁-Symmetric NHC-Catalyzed Addition to α-Aryl Substituted
α,β-Disubstituted Enals: Enantioselective Synthesis of Fully Functionalized
Dihydropyranones**

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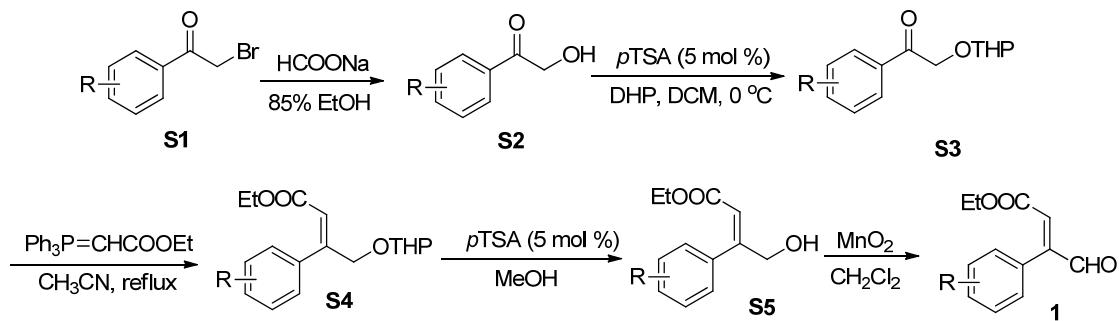
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1. General Information

Chemicals and solvents were either purchased from commercial suppliers or purified by standard techniques. Analytical thin-layer chromatography (TLC) was performed on silicycle silica gel plates with F-254 indicator and compounds were visualized by irradiation with UV light. Flash chromatography was carried out utilizing silica gel 200-300 mesh. ^1H NMR, ^{13}C NMR spectra were recorded on a Bruker AM-400 spectrometer (400 MHz ^1H , 100 MHz ^{13}C). The spectra were recorded in CDCl_3 as the solvent at room temperature, ^1H and $^{13}\text{CNMR}$ chemical shifts are reported in ppm relative to either the residual solvent peak (^{13}C) ($\delta = 77.00$ ppm) or TMS (^1H) ($\delta = 0$ ppm) as an internal standard. Data for ^1H NMR are reported as follows: chemical shift (δ ppm), multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet, dd = doublet), integration, coupling constant (Hz) and assignment. Data for ^{13}C NMR are reported as chemical shift. IR spectra were recorded using Nicolet NEXUS 670 FT-IR instrument and are reported in wavenumbers (cm^{-1}). HRMS were performed on Bruker Apex II mass instrument (ESI). Enantiomeric excess values were determined by HPLC with employing a Daicel Chirapak AD-H/OD-H/OJ-H on Agilent 1100 series and Waters 600 Deltaeluting with *i*-PrOH and *n*-hexane. Optical rotation was measured on the Perkin Elmer 341 polarimeter with $[\alpha]_D$ values reported in degrees; concentration (c) is in g/100 mL.

2. Preparation and Spectral Data of α -Aryl Substituted α,β -Disubstituted Aldehydes



2.1 General Procedure for α -Aryl Substituted α,β -Disubstituted Aldehydes

A solution of **S1** (35 mmol) of α -bromo ketone and 15.0g (220 mmol) of sodium formate in 90 mL 85% ethanol was heated to reflux for 12h. After the reaction was completed, the mixture was cool to ambient temperature, and ethanol was removed in *vacuo*, diluted with water (20 mL), aqueous layer was extracted with ethyl acetate (3×40 mL), and the combined organic phases were dried over Na_2SO_4 , filtered and concentrated. The ketols **S2** were recrystallized from 95% ethanol.

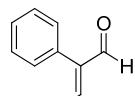
A 250 mL flame-dried round bottom flask was charged with **S2** (22 mmol) and *p*-toluenesulfonic acid 230 mg (1.1 mmol) under a dry N_2 atmosphere. After DCM (130 mL) was added to the flask, the mixture was cooled to 0°C . 3,4-2H-dihydropyran (DHP) 9.26 g (110 mmol) was then added dropwise to the mixture. After a further 2 hours of stirring at 0°C (determined by TLC), the reaction mixture was quenched with saturated NaHCO_3 . The organic layer was separated and the aqueous layer was extracted into DCM (3×20 mL). The combined organics were washed with brine, dried over Na_2SO_4 , filtered, and concentrated in *vacuo*, and purified with flash chromatography (eluent, ethyl acetate/petroleum ether 1/9) to yield crude product **S3**.

The crude product **S3** (10 mmol) was dissolved in acetonitrile (20 mL) in a 50 mL round bottom flask, ethyl 2-(triphenylphosphoranylidene)acetate 4.0 g (12 mmol) was added to the solution. The mixture was refluxed for 12 h. The mixture was evaporated to distill off one third of acetonitrile, and then continued to react for about 12 h (determined by TLC) until the reaction was complete. Solvent was distilled off, the concentrate was extracted with ether and washed with water. The combined organics were dried over Na_2SO_4 , filtered, and concentrated in *vacuo*, and purified with flash chromatography (eluent, ethyl acetate/petroleum ether 1/9) to yield crude coupling product **S4**.

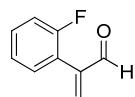
The crude product **S4** (7.5 mmol) was dissolved in MeOH (40 mL) and then cooled to 0 °C. *p*-Toluenesulfonic acid 71.3 mg (0.375 mmol) was added to the mixture. The reaction was brought to ambient temperature. After a further 3 hours of stirring at ambient temperature (determined by TLC), the reaction mixture was quenched with saturated NaHCO₃ at 0 °C. MeOH was distilled off, and the concentrate was extracted with ethyl acetate (3 × 20 mL). The combined organics were washed with brine, dried over Na₂SO₄, filtered, and concentrated in *vacuo*, yielding **S5** that was used directly without further purification in the next step.

To a stirred solution of **S5** (5 mmol) in CH₂Cl₂ (50 mL) was added MnO₂ (55 mol) at room temperature. After completion of the reaction (monitored by TLC), MnO₂ was removed, and the organic layers were concentrated in *vacuo*, the crude product was purified by silica gel column chromatography using (eluent, ethyl acetate/petroleum ether 1/8) to afford pure α,β-unsaturated aldehyde **1** as light yellow liquid.

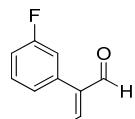
2.2 Spectral Data of Substrates



(E)-ethyl 4-oxo-3-phenylbut-2-enoate (1a) ¹H NMR (400 MHz, CDCl₃): δ = 9.73 (s, 1H), 7.40–7.36 (m, 3H), 7.25–7.20 (m, 2H), 6.69 (s, 1H), 4.11 (q, *J* = 7.2 Hz, 2H), 1.10 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 192.7, 165.0, 149.3, 136.0, 131.2, 128.9, 128.8, 127.8, 61.1, 13.6.

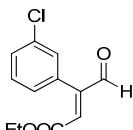


(E)-ethyl 3-(2-fluorophenyl)-4-oxobut-2-enoate (1b) ¹H NMR (400 MHz, CDCl₃): δ = 9.74 (s, 1H), 7.42–7.36 (m, 1H), 7.19–7.14 (m, 2H), 7.10 (t, 1H), 6.84 (s, 1H), 4.17–4.12 (d, *J* = 7.2 Hz, 2H), 1.14 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 191.7, 164.3, 160.7, 158.3, 144.6, 137.8, 130.9, 130.89, 130.7, 130.69, 123.64, 123.61, 119.5, 119.3, 115.4, 115.2, 61.3, 13.7.



(E)-ethyl 3-(3-chlorophenyl)-4-oxobut-2-enoate (1c) ¹H NMR (400 MHz, CDCl₃): δ = 9.75 (s, 1H), 7.40–7.34 (m, 1H), 7.13–7.08 (td, *J* = 2.4 Hz, *J* = 3.6 Hz, 1H), 7.00–6.94 (m, 2H), 6.74 (s, 1H), 4.16 (q, *J* = 7.2 Hz, 2H), 1.15 (t, *J* = 7.2 Hz, 3H); ¹³C NMR

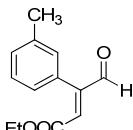
(100 MHz, CDCl₃): δ = 192.1, 164.6, 163.4, 160.9, 148.2, 137.0, 133.2, 133.1, 129.5, 129.46, 124.7, 116.2, 116.0, 115.8, 61.3, 13.7.



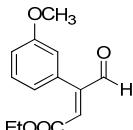
(E)-ethyl 3-(3-chlorophenyl)-4-oxobut-2-enoate (1d) ¹H NMR (400 MHz, CDCl₃): δ = 9.73 (s, 1H), 7.39–7.31 (m, 2H), 7.22 (s, 1H), 7.11 (d, *J* = 7.6 Hz, 1H), 6.73 (s, 1H), 4.16 (q, *J* = 7.2 Hz, 2H), 1.15 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 192.1, 164.5, 148.1, 137.0, 133.7, 132.8, 129.1, 129.0, 128.9, 127.1, 61.4, 13.7.



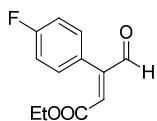
(E)-ethyl 3-(3-bromophenyl)-4-oxobut-2-enoate (1e) ¹H NMR (400 MHz, CDCl₃): δ = 9.70 (s, 1H), 7.53–7.50 (m, 1H), 7.37 (t, *J* = 2.0 Hz, 1 H), 7.26 (t, *J* = 8.0 Hz, 1H), 7.16–7.13 (m, 1H), 6.72 (s, 1H), 4.13 (q, *J* = 7.2 Hz, 2H), 1.14 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 192.0, 164.4, 147.8, 137.0, 133.0, 131.8, 131.7, 129.3, 127.5, 121.7, 61.3, 13.7.



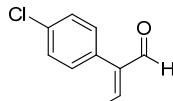
(E)-ethyl 4-oxo-3-(m-tolyl)but-2-enoate (1f) ¹H NMR (400 MHz, CDCl₃): δ = 9.74 (s, 1H), 7.27 (q, *J* = 7.6 Hz, 1H), 7.20 (d, *J* = 7.6 Hz, 1H), 7.02 (d, *J* = 7.6 Hz, 2H), 6.68 (s, 1H), 4.13 (q, *J* = 7.2 Hz, 2H), 2.36 (s, 3H), 1.13 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 192.8, 165.1, 149.5, 137.5, 135.8, 131.1, 129.8, 129.4, 127.9, 125.9, 61.1, 21.3, 13.7.



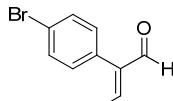
(E)-ethyl 3-(3-methoxyphenyl)-4-oxobut-2-enoate (1g) ¹H NMR (400 MHz, CDCl₃): δ = 9.74 (s, 1H), 7.30 (t, *J* = 8.0 Hz, 1H), 6.94 (dd, *J* = 2.4 Hz, *J* = 8.4 Hz, 1H), 6.81–6.77 (m, 1H), 6.70 (s, 1H), 4.13 (q, *J* = 7.2 Hz, 2H), 3.80 (s, 3H), 1.13 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 192.5, 165.0, 159.1, 149.1, 136.0, 132.5, 129.1, 121.2, 114.6, 114.5, 61.2, 55.2, 13.8.



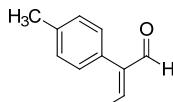
(E)-ethyl 3-(4-fluorophenyl)-4-oxobut-2-enoate (1h) ^1H NMR (400 MHz, CDCl_3): δ = 9.74 (s, 1H), 7.26–7.21 (m, 2H), 7.11–7.07 (m, 2H), 6.71 (s, 1H), 4.15 (q, J = 7.2 Hz, 2H), 1.16 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 192.6, 164.8, 164.4, 161.9, 148.6, 136.6, 131.1, 131.0, 126.9, 126.8, 115.2, 115.0, 61.3, 13.8.



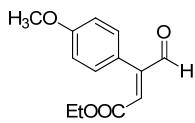
(E)-ethyl 3-(4-chlorophenyl)-4-oxobut-2-enoate (1i) ^1H NMR (400 MHz, CDCl_3): δ = 9.73 (s, 1H), 7.38 (dd, J = 2.0 Hz, J = 6.8 Hz, 2H), 7.16 (dd, J = 1.6 Hz, J = 6.8 Hz, 2H), 6.71 (s, 1H), 4.15 (q, J = 7.2 Hz, 2H), 1.17 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 192.4, 164.7, 148.5, 136.7, 135.2, 130.4, 129.4, 128.2, 61.4, 13.8.



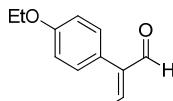
(E)-ethyl 3-(4-bromophenyl)-4-oxobut-2-enoate (1g) ^1H NMR (400 MHz, CDCl_3): δ = 9.73 (s, 1H), 7.53 (d, J = 8.4 Hz, 2H), 7.10 (d, J = 8.4 Hz, 2H), 6.72 (s, 1H), 4.15 (q, J = 7.2 Hz, 2H), 1.17 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 192.3, 164.7, 148.6, 136.7, 131.2, 130.7, 130.0, 123.5, 61.4, 13.8.



(E)-ethyl 4-oxo-3-(p-tolyl)but-2-enoate (1k) ^1H NMR (400 MHz, CDCl_3): δ = 9.73 (s, 1H), 7.20 (d, J = 7.6 Hz, 2H), 7.13 (m, 2H), 6.66 (s, 1H), 4.14 (q, J = 7.2 Hz, 2H), 2.37 (s, 3H), 1.16 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 192.9, 165.1, 149.4, 139.0, 135.5, 128.8, 128.6, 128.1, 61.1, 21.3, 13.8.

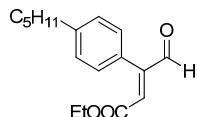


(E)-ethyl 3-(4-methoxyphenyl)-4-oxobut-2-enoate (1l) ^1H NMR (400 MHz, CDCl_3): δ = 9.70 (s, 1H), 7.21 (dd, J = 2.0 Hz, J = 6.8 Hz, 2H), 6.92 (dd, J = 2.0 Hz, J = 6.8 Hz, 2H), 6.63 (s, 1H), 4.16 (q, J = 7.2 Hz, 2H), 3.82 (s, 3H), 1.18 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 193.3, 165.2, 160.3, 148.9, 135.2, 130.6, 123.0, 113.4, 61.1, 55.2, 13.9.

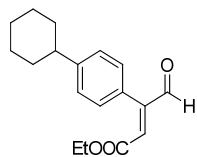


(E)-ethyl 3-(4-ethoxyphenyl)-4-oxobut-2-enoate (1m) ^1H NMR (400 MHz, CDCl_3): δ = 9.70 (s, 1H), 7.21 (d, J = 8.4 Hz, 2H), 6.92 (d, J = 8.8 Hz, 2H), 6.62 (s, 1H), 4.17

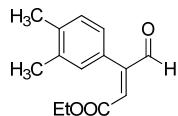
(q, $J = 7.2$ Hz, 2H), 4.06 (q, $J = 7.2$ Hz, 2H), 1.42 (t, $J = 7.2$ Hz, 3H), 1.18 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 193.3, 165.3, 159.8, 148.8, 135.1, 130.6, 122.9, 114.0, 63.4, 61.1, 14.7, 13.9$.



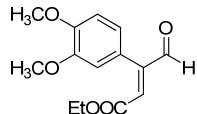
(E)-ethyl 4-oxo-3-(4-pentylphenyl)but-2-enoate (1n) ^1H NMR (400 MHz, CDCl_3): $\delta = 9.75$ (s, 1H), 7.22 (d, $J = 8.4$ Hz, 2H), 7.16 (t, $J = 1.6$ Hz, $J = 6.4$ Hz, 2H), 6.66 (s, 1H), 4.17 (q, $J = 7.2$ Hz, 2H), 2.64 (t, $J = 7.6$ Hz, $J = 8$ Hz, 2H), 1.67–1.59 (m, 2H), 1.37–1.31 (m, 4H), 1.15 (t, $J = 7.2$ Hz, 3H), 0.91 (t, $J = 6.8$ Hz, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 193.0, 165.2, 149.4, 144.1, 135.5, 128.9, 128.3, 128.0, 61.2, 35.8, 31.5, 30.9, 22.5, 14.0, 13.8$.



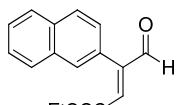
(E)-ethyl 3-(4-cyclohexylphenyl)-4-oxobut-2-enoate (1o) ^1H NMR (400 MHz, CDCl_3): $\delta = 9.76$ (s, 1H), 7.24 (d, $J = 8.4$ Hz, 2H), 7.17 (d, $J = 8$ Hz, 2H), 6.66 (s, 1H), 4.16–4.11 (q, $J = 7.2$ Hz, 2H), 2.54–2.49 (m, 1H), 1.89–1.83 (m, 4H), 1.76 (d, $J = 8.4$ Hz, 1H), 1.47–1.34 (m, 4H), 1.29–1.24 (m, 1H), 1.13 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 193.1, 165.3, 149.3, 149.2, 135.5, 129.0, 128.5, 126.5, 61.2, 44.4, 34.3, 26.8, 26.1, 13.8$.



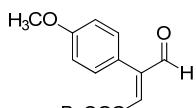
(E)-ethyl 3-(3,4-dimethylphenyl)-4-oxobut-2-enoate (1p) ^1H NMR (400 MHz, CDCl_3): $\delta = 9.73$ (s, 1H), 7.16 (d, $J = 7.6$ Hz, 2H), 7.00–6.96 (m, 2H), 6.65 (s, 1H), 4.18–4.12 (q, $J = 7.2$ Hz, 2H), 2.27 (d, 6H), 1.16 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 193.1, 165.2, 149.5, 137.8, 136.1, 135.3, 129.9, 129.3, 128.6, 126.4, 61.1, 19.7, 19.6, 13.8$.



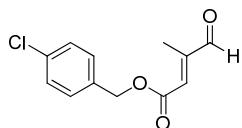
(E)-ethyl 3-(3,4-dimethoxyphenyl)-4-oxobut-2-enoate (1q) ^1H NMR (400 MHz, CDCl_3): $\delta = 9.76$ (s, 1H), 6.91–6.81 (m, 3H), 6.66 (s, 1H), 4.20–4.14 (q, $J = 7.2$ Hz, 2H), 3.91 (s, 3H), 3.87 (s, 3H), 1.19 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 193.1, 165.3, 149.8, 148.5, 148.4, 135.4, 123.5, 122.2, 112.5, 110.6, 61.1, 55.8, 13.9$.



(E)-ethyl 3-(naphthalen-2-yl)-4-oxobut-2-enoate (1r) ^1H NMR (400 MHz, CDCl_3): δ = 9.73 (s, 1H), 7.81–7.77 (m, 3H), 7.70 (s, 1H), 7.45–7.40 (m, 2H), 7.27 (dd, J = 1.2 Hz, J = 8.4 Hz, 1H), 6.67 (s, 1H), 4.04 (q, J = 7.2 Hz, 2H), 0.99 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 192.6, 164.8, 149.0, 136.1, 133.1, 132.5, 128.6, 128.5, 128.1, 127.5, 127.2, 126.5, 126.3, 126.0, 60.9, 13.5.

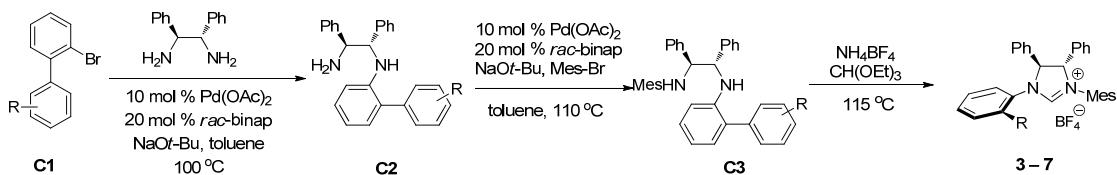


(E)-benzyl 3-(4-methoxyphenyl)-4-oxobut-2-enoate (1s) ^1H NMR (400 MHz, CDCl_3): δ = 9.71 (s, 1H), 7.31 (t, J = 3.2 Hz, 3H), 7.19–7.15 (m, 4H), 6.87–6.84 (m, 2H), 6.65 (s, 1H), 5.12 (s, 2H), 3.80 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 193.1, 165.1, 160.3, 149.1, 134.9, 134.7, 130.6, 128.4, 128.36, 123.0, 113.5, 66.9, 55.1.



(E)-4-chlorobenzyl 3-methyl-4-oxobut-2-enoate (1x) ^1H NMR (400 MHz, CDCl_3): δ = 9.55 (s, 1H), 7.38–7.82 (m, 4H), 6.54–6.53 (m, 1H), 5.21 (s, 2H), 2.88 (d, J = 1.6 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 194.3, 165.1, 151.2, 134.7, 134.5, 133.7, 129.8, 128.9, 66.0, 10.9.

3. General Procedure for the Synthesis of Chiral N-Heterocyclic Carbene Catalysts



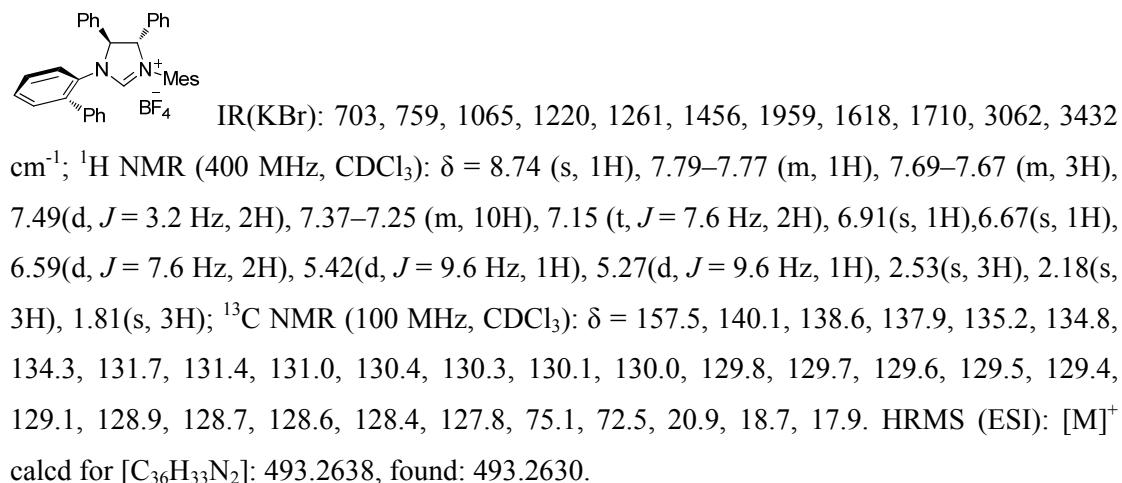
A flame-dried, round-bottom flask equipped with reflux condenser was charged with $\text{Pd}(\text{OAc})_2$ (89 mg, 0.4 mmol), *rac*-binap (498 mg, 0.8 mmol), NaOt-Bu (769 mg, 8.0 mmol), **C1** (4.0 mmol), and (*S,S*)-1,2-diphenyl ethylenediamine (1.3 g, 6.0 mmol) under a dry N_2 atmosphere. Toluene (70 mL) was added to the mixture. After 10 hours at 100 °C, the solution was allowed to cool to room temperature, passed through a plug of Celite, concentrated, and purified by silica gel chromatography (eluent, ethyl acetate/petroleum ether 1/1) to afford product diamine **C2** as light yellow oil.

Diamine **C2** (3.0 mmol) was added to a flame-dried, round-bottom flask charged with $\text{Pd}(\text{OAc})_2$ (67 mg, 0.3 mmol), *rac*-binap (373 mg, 0.6 mmol), NaOt-Bu (578 mg, 6.0 mmol), mesityl bromide (598 mg, 3.0 mmol) under a dry N_2 atmosphere. Toluene (70 mL) was added

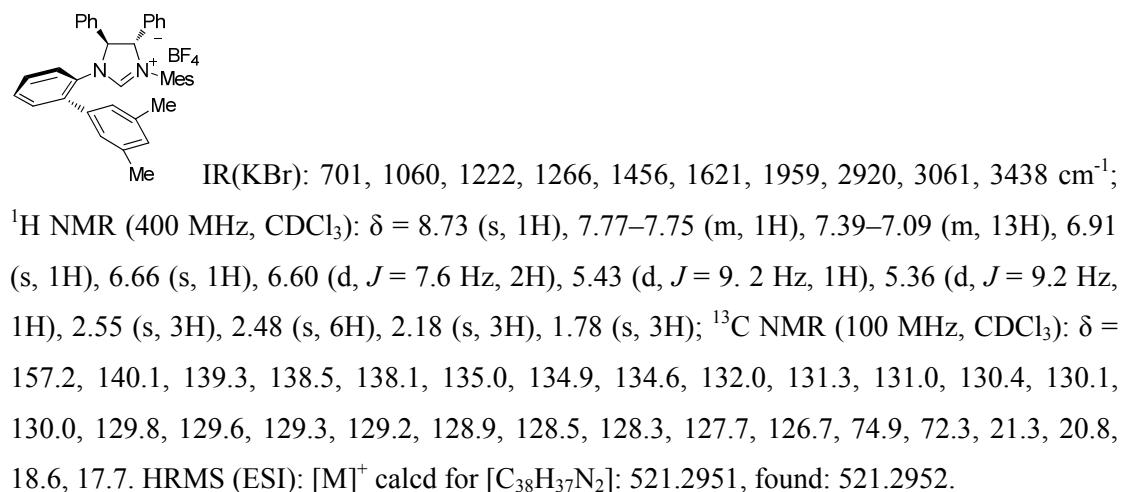
to the mixture. After 12 hours at 110 °C, the solution was allowed to cool to room temperature and then passed through a plug of Celite, concentrated, and purified by silica gel column chromatography (eluent, petroleum ether/Et₂O 20/1) to afford product **C3**.

A flame-dried, round-bottom flask equipped with a short path distillation set was charged with diamine **C3** (2.07mmol), HC(OEt)₃ (13 mL), and NH₄BF₄ (325 mg, 3.10mmol). After 3 hours at 115 °C, the solution was allowed to cool to room temperature, concentrated, and purified by silicagel column chromatography (eluent, petroleum ether/EtOAc 1/3) to afford the corresponding imidazolinium salt, which was triturated (hexane/CH₂Cl₂) to a white solid.

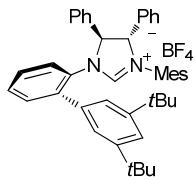
(4S,5S)-1-([1,1'-biphenyl]-2-yl)-3-mesityl-4,5-diphenyl-4,5-dihydro-1H-imidazol-3-ium (5e)



(4S,5S)-1-(3',5'-dimethyl-[1,1'-biphenyl]-2-yl)-3-mesityl-4,5-diphenyl-4,5-dihydro-1H-imidazol-3-ium (5f)

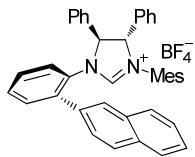


(4S,5S)-1-(3',5'-di-tert-butyl-[1,1'-biphenyl]-2-yl)-3-mesityl-4,5-diphenyl-4,5-dihydro-1H-imidazol-3-i um (5h)



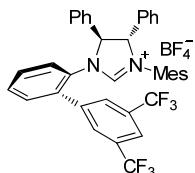
IR(KBr): 697, 1065, 1222, 1268, 1455, 1619, 1959, 2959, 3063, 3437 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 8.74 (s, 1H), 7.90 (d, *J* = 7.2 Hz, 1H), 7.75 (s, 1H), 7.41–7.13 (m, 16H), 6.91 (s, 1H), 6.63 (s, 1H), 6.47 (d, *J* = 6.4 Hz, 2H), 5.71 (d, *J* = 8.4 Hz, 1H), 5.31 (d, *J* = 8.4 Hz, 1H), 2.45 (s, 3H), 2.19 (s, 3H), 1.65 (s, 3H), 1.40 (s, 18H); ¹³C NMR (100 MHz, CDCl₃): δ = 157.4, 152.0, 140.2, 139.0, 138.1, 135.3, 134.9, 132.0, 131.6, 131.5, 130.4, 130.3, 130.0, 129.7, 129.6, 129.5, 129.1, 128.8, 128.6, 128.3, 127.4, 123.1, 123.0, 75.2, 72.1, 35.1, 31.5, 20.9, 18.5, 18.2. HRMS (ESI): [M]⁺ calcd for [C₄₄H₄₉N₂]: 605.3890, found: 605.3887.

(4S,5S)-3-mesityl-1-(2-(naphthalen-2-yl)phenyl)-4,5-diphenyl-4,5-dihydro-1H-imidazol-3-i um (5i)



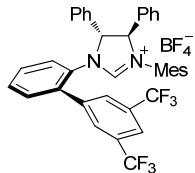
IR(KBr): 702, 784, 1061, 1220, 1266, 1455, 1620, 1959, 2922, 3057, 3437 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 8.82 (s, 1H), 8.16 (d, *J* = 8.4 Hz, 1H), 8.09 (d, *J* = 7.2 Hz, 1H), 8.02–7.96 (m, 2H), 7.78 (d, *J* = 7.2 Hz, 1H), 7.73–7.67 (m, 2H), 7.56 (d, *J* = 8.0 Hz, 1H), 7.37–7.23 (m, 9H), 7.07 (t, *J* = 7.2 Hz, 1H), 6.87 (s, 1H), 6.63 (s, 3H), 6.24 (d, *J* = 7.2 Hz, 2H), 5.33 (d, *J* = 9.2 Hz, 1H), 5.17 (d, *J* = 9.2 Hz, 1H), 2.51 (s, 3H), 2.15 (s, 3H), 1.74 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 157.4, 140.1, 138.0, 135.7, 135.3, 134.7, 134.5, 133.4, 133.0, 131.7, 131.5, 131.1, 130.3, 130.1, 130.08, 130.0, 129.7, 129.6, 129.5, 129.4, 128.6, 128.3, 128.2, 128.15, 128.08, 128.0, 127.7, 127.5, 127.3, 126.5, 74.9, 72.4, 20.8, 18.6, 17.8. HRMS (ESI): [M]⁺ calcd for [C₄₀H₃₅N₂]: 543.2795, found: 543.2791.

(4S,5S)-1-(3',5'-bis(trifluoromethyl)-[1,1'-biphenyl]-2-yl)-3-mesityl-4,5-diphenyl-4,5-dihydro-1H-imidazol-3-ium (5j)



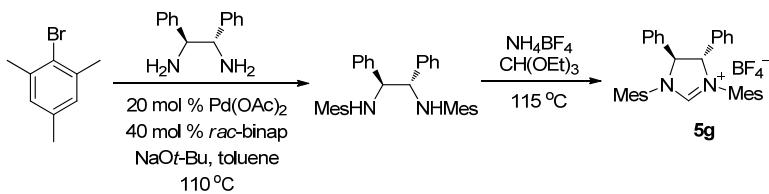
IR(KBr): 701, 1066, 1138, 1281, 1381, 1618, 1959, 2928, 3066, 3437 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 8.89 (s, 1H), 8.10 (s, 1H), 7.99 (d, *J* = 7.2 Hz, 3H), 7.37–7.33 (m, 4H), 7.32–7.25 (m, 2H), 7.21–7.13 (m, 4H), 7.09–7.06 (m, 2H), 6.93 (s, 1H), 6.73–6.71 (m, 3H), 5.51 (d, *J* = 10.4 Hz, 1H), 5.08 (d, *J* = 10 Hz, 1H), 2.52 (s, 3H), 2.22 (s, 3H), 1.87 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 158.4, 141.2, 140.1, 135.7, 134.6, 133.9, 133.4, 133.2, 132.9, 132.5, 132.2, 131.8, 131.7, 131.1, 130.8, 130.6, 130.3, 129.97, 129.95, 129.8, 129.6, 129.5, 129.2, 128.5, 128.2, 127.7, 127.0, 124.3, 122.3, 121.6, 75.6, 73.2, 20.9, 18.7, 17.7. HRMS (ESI): [M]⁺ calcd for [C₃₈H₃₁F₆N₂]: 629.2386, found: 629.2383.

(4R,5R)-1-(3',5'-bis(trifluoromethyl)-[1,1'-biphenyl]-2-yl)-3-mesityl-4,5-diphenyl-4,5-dihydro-1H-imidazol-3-ium (5k)



IR(KBr): 701, 1066, 1138, 1281, 1381, 1618, 1959, 2928, 3066, 3437 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 8.89 (s, 1H), 8.11 (s, 1H), 7.99–7.97 (m, 3H), 7.36–7.24 (m, 5H), 7.21–7.13 (m, 4H), 7.12–7.06 (m, 2H), 6.93 (s, 1H), 6.74–6.71 (m, 3H), 5.50 (d, *J* = 10.4 Hz, 1H), 5.07 (d, *J* = 10.4 Hz, 1H), 2.52 (s, 3H), 2.22 (s, 3H), 1.87 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 158.4, 141.2, 140.1, 135.7, 134.6, 133.9, 133.4, 133.2, 132.8, 132.5, 132.2, 131.8, 131.7, 131.0, 130.8, 130.6, 130.3, 129.9, 129.8, 129.6, 129.5, 129.1, 128.5, 128.2, 127.7, 127.0, 124.3, 122.3, 121.6, 118.9, 75.6, 73.2, 20.9, 18.7, 17.7. HRMS (ESI): [M]⁺ calcd for [C₃₈H₃₁F₆N₂]: 629.2386, found: 629.2383.

Representative procedure for synthesis of imidazolinium salt 5g:



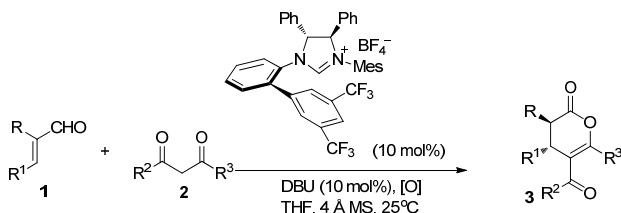
A flame-dried, round-bottom flask equipped with a reflux condenser was charged with Pd(OAc)₂ (106 mg, 0.472 mmol), *rac*-binap (587 mg, 0.944 mmol), NaOt-Bu (906 mg, 9.44 mmol), mesityl bromide (986 mg, 4.96 mmol), and (*S,S*)-1,2-diphenyl ethylenediamine (500 mg, 2.36 mmol) under a dry N₂ atmosphere. Toluene (60 mL) was added to the mixture. After 12 hours at 100 °C, the solution was allowed to cool to room temperature, passed through a plug of Celite, concentrated, and purified by silica gel chromatography (eluent, petroleum ether/Et₂O 1/20) to afford product diamine.

A flame-dried, round-bottom flask equipped with a short path distillation set was charged with diamine (2.2 mmol), HC(OEt)₃ (14 mL), and NH₄BF₄ (334 mg, 3.10 mmol). After 3 hours at 115 °C, the solution was allowed to cool to room temperature, concentrated, and purified by silicagel column chromatography (eluent, petroleum ether/EtOAc 1/3) to afford the corresponding imidazolinium salt **2**, which was triturated (hexane/CH₂Cl₂) to a white solid.

(4S,5S)-1,3-dimesityl-4,5-diphenyl-4,5-dihydro-1H-imidazol-3-ium (5g) IR (KBr): 700, 759, 854, 1060, 1224, 1267, 1456, 1959, 1620, 2823, 3040, 3438 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 8.53 (s, 1H), 7.37 (d, *J* = 7.6 Hz, 10H), 6.96 (s, 2H), 6.70 (s, 2H), 6.01 (s, 2H), 2.66 (s, 3H), 2.19 (s, 3H), 1.90 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 157.9, 140.2, 136.0, 134.3, 131.1, 130.6, 130.1, 130.0, 129.4, 128.8, 128.5, 72.8, 20.8, 18.8, 18.1. HRMS (ESI): [M]⁺ calcd for [C₃₃H₃₅N₂]: 459.2800, found: 459.2792.

4. General Procedure and Spectral Data of Products

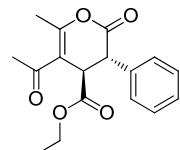
4.1 General Procedure for the synthesis of Fully Functionalized Dihydropyranones



To a flame dried vessel were successively added NHC (0.01 mmol, 7.16 mg), 4 Å Molecular sieves (40.0 mg), DBU (0.01 mmol, 1.52 mg in 0.25 mL THF), oxidant (0.1 mmol, 40.8 mg), and **2a** (0.3 mmol, 30.3 mg) under N₂ atmosphere. After 10 min, **1a** (0.1 mmol, 20.4 mg in 0.25 mL THF) was added to the mixture. The reaction was stirred at 25°C for a specified reaction time (monitored by TLC). When the reaction was completed, the solvent was evaporated under reduced pressure and the residue was purified by silica gel flash column chromatography (petroleumether/EtOAc) to give the corresponding dihydropyranones **3a**.

4.2 Analytical Data of Fully Substituted Dihydropyranones

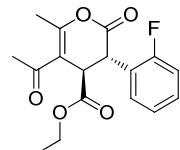
(3S,4R)-ethyl 5-acetyl-6-methyl-2-oxo-3-phenyl-3,4-dihydro-2H-pyran-4-carboxylate (3a)



Pale Yellow Liquid. $[\alpha]_D^{20} = 41$ (*c* 1.0, CH₂Cl₂, 96% ee); IR(KBr): 702, 1027, 1157, 1261, 1363, 1456, 1601, 1730, 1791, 2371, 2856, 2925, 3275 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.37–7.31 (m, 3H), 7.21–7.19 (m, 2H), 4.38 (d, *J* = 2.4 Hz, 1H), 4.22–4.16 (q, *J* = 7.2 Hz, 2H), 4.12 (d, *J* = 1.6 Hz, 1H), 2.33 (t, 6H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.6, 170.5, 165.3, 160.2, 133.9, 129.2, 128.3, 126.8, 112.6, 62.1, 45.6, 45.5, 30.2, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer *t*_R = 23.24 min, minor enantiomer *t*_R = 27.88 min. HRMS (ESI): [M+H]⁺ calcd for [C₁₇H₁₈O₅]: 303.1227, found: 303.1229.

(3S,4R)-ethyl

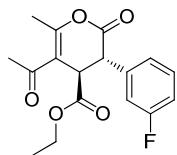
5-acetyl-3-(2-fluorophenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3b)



Pale Yellow Liquid. $[\alpha]_D^{20} = -10$ (*c* 1.0, CH₂Cl₂, 91% ee); IR(KBr): 763, 1038, 1114, 1159, 1367, 1457, 1590, 1728, 2375, 2923, 3390 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.36–7.31 (m, 1H), 7.14–7.10 (m, 2H), 7.03–7.00 (m, 1H), 4.55 (d, *J* = 4.4 Hz, 1H), 4.18–4.13 (q, *J* = 7.2 Hz, 2H), 4.04 (d, *J* = 3.6 Hz, 1H), 2.37 (s, 3H), 2.28 (s, 3H), 1.19 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.8, 170.5, 164.7, 161.5, 159.8, 159.0, 130.4, 130.3, 128.4, 128.38, 124.8, 124.7, 121.6, 116.3, 116.0, 113.1, 62.1, 45.44, 45.42, 41.21, 41.19, 30.2, 19.2, 13.8. The enantiomeric excess was determined by HPLC with an AD-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; minor enantiomer *t*_R = 7.32 min, major enantiomer *t*_R = 8.79 min. HRMS (ESI): [M+H]⁺ calcd for [C₁₇H₁₇FO₅]: 321.1133, found: 321.1137.

(3S,4R)-ethyl

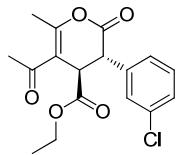
5-acetyl-3-(3-fluorophenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3c)



Pale Yellow Liquid. $[\alpha]_D^{20} = 48$ (*c* 1.0, CH₂Cl₂, 90% ee); IR(KBr): 743, 796, 1023, 1105, 1261, 1377, 1459, 1591, 1731, 2922, 2958, 3409 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.35–7.30 (m, 1H), 7.05–7.00 (m, 2H), 6.96–6.93 (dd, *J* = 2 Hz, *J* = 5.6 Hz, 1H), 4.37 (d, *J* = 2.8 Hz, 1H), 4.22–4.16 (q, *J* = 7.2 Hz, 2H), 4.12–4.11 (dd, *J* = 0.8 Hz, *J* = 2.4 Hz, 1H), 2.36 (s, 3H), 2.33 (d, *J* = 0.8 Hz, 3H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.5, 170.3, 164.8, 164.2, 161.8, 160.0, 136.2, 136.1, 130.9, 130.8, 122.64, 122.61, 115.6, 115.4, 114.4, 114.2, 112.8, 62.2, 45.39, 45.38, 45.3, 30.3, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an OD-H column. (*n*-hexane:*i*-PrOH = 90:10), 1 mL/min; minor enantiomer *t*_R = 18.80 min, major enantiomer *t*_R = 22.91 min. HRMS (ESI): [M+H]⁺ calcd for [C₁₇H₁₇FO₅]: 321.1133, found: 321.1134.

(3S,4R)-ethyl

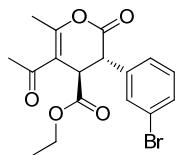
5-acetyl-3-(3-chlorophenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3d)



Yellow Liquid. $[\alpha]_D^{20} = 94$ (*c* 1.0, CH₂Cl₂, 93% ee); IR(KBr): 796, 1025, 1117, 1260, 1381, 1599, 1731, 1786, 2368, 2963, 3370 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.32–7.29 (m, 2H), 7.22 (s, 1H), 7.09–7.07 (m, 1H), 4.35 (d, *J* = 2.8 Hz, 1H), 4.21–4.16 (q, *J* = 7.2 Hz, 2H), 4.11 (d, *J* = 2 Hz, 1H), 2.37 (s, 3H), 2.34 (s, 3H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.5, 170.3, 164.8, 160.0, 135.7, 135.1, 130.4, 128.7, 127.4, 125.0, 112.8, 62.2, 45.3, 45.2, 30.3, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an OD-H column. (*n*-hexane:*i*-PrOH = 95:5), 1 mL/min; minor enantiomer *t*_R = 28.51 min, major enantiomer *t*_R = 33.50 min. HRMS (ESI): [M+Na]⁺ calcd for [C₁₇H₁₇ClO₅]: 359.0657, found: 359.0664.

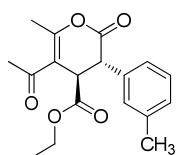
(3S,4R)-ethyl

5-acetyl-3-(3-bromophenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3e)



Yellow Liquid. $[\alpha]_D^{20} = 85$ (*c* 1.0, CH₂Cl₂, 90% ee); IR(KBr): 691, 790, 1028, 1073, 1120, 1254, 1426, 1595, 1730, 1787, 2924, 3380 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.47 (d, *J* = 8 Hz, 1H), 7.38 (s, 1H), 7.22 (t, *J* = 8 Hz, 1H), 7.14 (d, *J* = 7.6 Hz, 1H), 4.34 (d, *J* = 2.8 Hz, 1H), 4.21–4.16 (q, *J* = 7.2 Hz, 2H), 4.10 (d, *J* = 2 Hz, 1H), 2.36 (s, 3H), 2.33 (d, *J* = 0.8 Hz, 3H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.5, 170.3, 164.7, 160.0, 136.0, 131.6, 130.7, 130.3, 125.4, 123.2, 112.9, 62.3, 45.33, 45.30, 30.3, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an OD-H column. (*n*-hexane:*i*-PrOH = 90:10), 1 mL/min; minor enantiomer *t_R* = 20.54 min, major enantiomer *t_R* = 25.35 min. HRMS (ESI): [M+H]⁺ calcd for [C₁₇H₁₇BrO₅]: 381.0332, found: 381.0337.

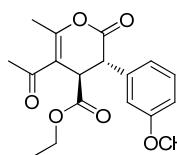
(3S,4R)-ethyl 5-acetyl-6-methyl-2-oxo-3-(m-tolyl)-3,4-dihydro-2H-pyran-4-carboxylate (3f)



Yellow Liquid. $[\alpha]_D^{20} = 56$ (*c* 1.0, CH₂Cl₂, 94% ee); IR(KBr): 737, 1024, 1155, 1262, 1362, 1462, 1608, 1732, 1788, 2926, 3196 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.22 (t, *J* = 7.6 Hz, 1H), 7.13 (d, *J* = 7.2 Hz, 1H), 7.02 (s, 1H), 6.98 (d, *J* = 7.6 Hz, 1H), 4.33 (d, *J* = 2 Hz, 1H), 4.22–4.17 (q, *J* = 7.2 Hz, 2H), 4.10 (s, 1H), 2.33 (d, 9H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.6, 170.6, 165.4, 160.2, 139.0, 133.8, 129.1, 129.0, 127.7, 123.6, 112.6, 62.1, 45.59, 45.58, 30.2, 21.4, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer *t_R* = 13.69 min, minor enantiomer *t_R* = 18.97 min. HRMS (ESI): [M+H]⁺ calcd for [C₁₈H₂₀O₅]: 317.1384, found: 317.1389.

(3S,4R)-ethyl

5-acetyl-3-(3-methoxyphenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3g)

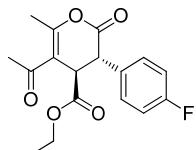


Yellow Liquid. $[\alpha]_D^{20} = 38$ (*c* 1.0, CH₂Cl₂, 94% ee); IR(KBr): 737, 1044, 1155, 1263, 1362, 1459, 1601, 1730, 2926, 3195 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.26 (t, *J* = 8 Hz, 1H), 6.86–6.83 (dd, *J* = 2.4 Hz, *J* = 4.4 Hz, 1H), 6.78–6.75 (m, 2H), 4.35 (d, *J* =

2 Hz, 1H), 4.23–4.17 (q, J = 7.2 Hz, 2H), 4.12 (d, J = 1.2 Hz, 1H), 3.78 (s, 3H), 2.35 (d, 6H), 1.25 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 196.7, 170.5, 165.3, 160.2, 160.1, 135.3, 130.2, 129.0, 118.9, 113.6, 112.9, 112.7, 62.2, 55.3, 45.5, 45.4, 30.3, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer t_R = 17.54 min, minor enantiomer t_R = 21.70 min. HRMS (ESI): [M+H] $^+$ calcd for $[\text{C}_{18}\text{H}_{20}\text{O}_6]$: 333.1333, found: 333.1331.

(3S,4R)-ethyl

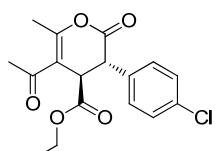
5-acetyl-3-(4-fluorophenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3h)



Yellow Liquid. $[\alpha]_D^{20} = 20$ (c 1.0, CH_2Cl_2 , 90% ee); IR(KBr): 778, 1043, 1117, 1158, 1261, 1456, 1594, 1731, 2372, 2924, 3366 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ = 7.21–7.17 (dd, J = 5.2 Hz, J = 8.4 Hz, 2H), 7.04(t, 2H), 4.34 (d, J = 2.8 Hz, 1H), 4.21–4.16(q, J = 7.2 Hz, 2H), 4.10 (d, J = 2 Hz, 1H), 2.36 (s, 3H), 2.33 (s, 3H), 1.23 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 196.6, 170.4, 165.2, 163.7, 161.3, 159.9, 129.59, 129.56, 128.8, 128.7, 116.3, 116.1, 112.9, 62.2, 45.4, 45.0, 30.3, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer t_R = 14.31 min, minor enantiomer t_R = 16.48 min. HRMS (ESI): [M+H] $^+$ calcd for $[\text{C}_{17}\text{H}_{17}\text{FO}_5]$: 321.1133, found: 321.1138.

(3S,4R)-ethyl

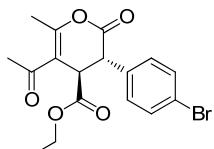
5-acetyl-3-(4-chlorophenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3i)



Yellow Liquid. $[\alpha]_D^{20} = 51$ (c 1.0, CH_2Cl_2 , 90% ee); IR(KBr): 741, 798, 1022, 1094, 1261, 1460, 1599, 1732, 2372, 2962, 3374 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): δ = 7.34–7.32 (dd, J = 2 Hz, J = 6.8 Hz, 2H), 7.04(d, J = 8.4 Hz, 2H), 4.34 (d, J = 2.8 Hz, 1H), 4.21–4.16 (q, J = 7.2 Hz, 2H), 4.11–4.10 (dd, J = 0.8 Hz, J = 2.8 Hz, 1H), 2.36 (s, 3H), 2.32 (s, 3H), 1.24 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ = 196.5, 170.3, 165.0, 159.9, 134.5, 132.3, 129.4, 128.3, 112.9, 62.2, 45.2, 45.1, 30.3, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an AD-H column. (*n*-hexane:*i*-PrOH = 90:10), 1 mL/min; major enantiomer t_R = 14.19 min, minor enantiomer t_R = 17.84 min. HRMS (ESI): [M+H] $^+$ calcd for $[\text{C}_{17}\text{H}_{17}\text{ClO}_5]$: 337.0837, found: 337.0842.

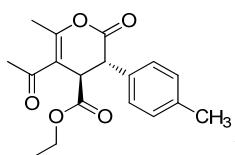
(3S,4R)-ethyl

5-acetyl-3-(4-bromophenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3j)



Yellow Liquid. $[\alpha]_D^{20} = 63$ (*c* 1.0, CH₂Cl₂, 93% ee); IR(KBr): 739, 814, 1021, 1152, 1261, 1363, 1458, 1732, 1790, 2924, 3308 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.49 (d, *J* = 8.8 Hz, 2H), 7.10 (d, *J* = 8 Hz, 2H), 4.32 (d, *J* = 2.8 Hz, 1H), 4.21–4.16 (q, *J* = 7.2 Hz, 2H), 4.11 (d, *J* = 2 Hz, 1H), 2.36 (s, 3H), 2.32 (d, *J* = 0.4 Hz, 3H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.5, 170.3, 164.9, 159.9, 132.8, 132.4, 128.6, 122.5, 112.9, 62.2, 45.2, 45.1, 30.3, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an AD-H column. (*n*-hexane:*i*-PrOH = 95:5), 1 mL/min; major enantiomer *t*_R = 26.67 min, minor enantiomer *t*_R = 33.20 min. HRMS (ESI): [M+H]⁺ calcd for [C₁₇H₁₇BrO₅]: 381.0332, found: 381.0337.

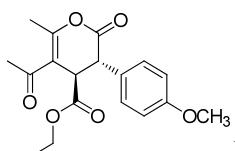
(3S,4R)-ethyl 5-acetyl-6-methyl-2-oxo-3-(p-tolyl)-3,4-dihydro-2H-pyran-4-carboxylate (3k)



Yellow Liquid. $[\alpha]_D^{20} = 111$ (*c* 1.0, CH₂Cl₂, 95% ee); IR(KBr): 737, 809, 1025, 1117, 1259, 1378, 1459, 1611, 1733, 1787, 2925, 3444 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.15 (d, *J* = 8 Hz, 2H), 7.09 (d, *J* = 8 Hz, 2H), 4.33 (d, *J* = 2.4 Hz, 1H), 4.22–4.16 (q, *J* = 7.2 Hz, 2H), 4.09 (d, *J* = 1.6 Hz, 1H), 2.33 (d, 9H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.7, 170.6, 165.5, 160.2, 138.2, 130.8, 129.9, 126.7, 112.6, 62.1, 45.6, 45.3, 30.2, 21.0, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer *t*_R = 12.74 min, minor enantiomer *t*_R = 14.11 min. HRMS (ESI): [M+H]⁺ calcd for [C₁₈H₂₀O₅]: 317.1384, found: 317.1381.

(3S,4R)-ethyl

5-acetyl-3-(4-methoxyphenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3l)

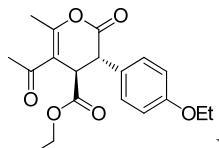


Yellow Liquid. $[\alpha]_D^{20} = 133$ (*c* 1.0, CH₂Cl₂, 95% ee); IR(KBr): 738, 1036, 1117, 1222, 1362, 1459, 1604, 1714, 2924, 3405 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.13 (d, *J* = 8.8 Hz, 2H), 6.87–6.85 (dd, *J* = 2 Hz, *J* = 6.4 Hz, 2H), 4.31 (d, *J* = 2.4 Hz, 1H),

4.21–4.16 (q, $J = 7.2$ Hz, 2H), 4.07 (d, $J = 1.6$ Hz, 1H), 3.79 (s, 3H), 2.34–2.32 (m, 6H), 1.24 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 196.8, 170.6, 165.6, 160.1, 159.5, 128.0, 125.7, 114.6, 112.7, 62.1, 55.3, 45.6, 44.9, 30.2, 19.4, 13.9$. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer $t_R = 24.64$ min, minor enantiomer $t_R = 31.60$ min. HRMS (ESI): $[\text{M}+\text{H}]^+$ calcd for $[\text{C}_{18}\text{H}_{20}\text{O}_6]$: 333.1333, found: 333.1329.

(3S,4R)-ethyl

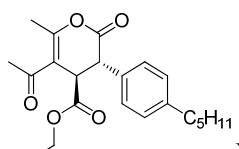
5-acetyl-3-(4-ethoxyphenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3m)



Yellow Liquid. $[\alpha]_D^{20} = 91$ ($c 1.0$, CH_2Cl_2 , 98% ee); IR(KBr): 825, 1044, 1149, 1252, 1365, 1513, 1612, 1732, 1788, 2926, 2960 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): $\delta = 7.11$ (d, $J = 8.8$ Hz, 2H), 6.86 (d, $J = 8.8$ Hz, 2H), 4.30 (d, $J = 2.4$ Hz, 1H), 4.21–4.16 (q, $J = 7.2$ Hz, 2H), 4.07 (d, $J = 1.6$ Hz, 1H), 4.03–3.98 (q, $J = 7.2$ Hz, 2H), 2.34 (d, 6H), 1.40 (t, $J = 7.2$ Hz, 3H), 1.24 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 196.8, 170.7, 165.6, 160.2, 158.8, 128.0, 125.5, 115.1, 112.7, 63.5, 62.1, 45.6, 44.9, 30.2, 19.4, 14.7, 13.9$. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer $t_R = 16.66$ min, minor enantiomer $t_R = 21.09$ min. HRMS (ESI): $[\text{M}+\text{H}]^+$ calcd for $[\text{C}_{19}\text{H}_{22}\text{O}_6]$: 347.1489, found: 347.1493.

(3S,4R)-ethyl

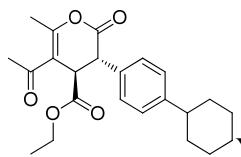
5-acetyl-6-methyl-2-oxo-3-(4-pentylphenyl)-3,4-dihydro-2H-pyran-4-carboxylate (3n)



Yellow Liquid. $[\alpha]_D^{20} = 44$ ($c 1.0$, CH_2Cl_2 , 97% ee); IR(KBr): 739, 1039, 1119, 1155, 1261, 1422, 1595, 1731, 2373, 2925, 3346 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): $\delta = 7.16$ (d, $J = 8$ Hz, 2H), 7.10 (d, $J = 8$ Hz, 2H), 4.33 (d, $J = 2$ Hz, 1H), 4.22–4.16 (q, $J = 7.2$ Hz, 2H), 4.08 (d, $J = 1.6$ Hz, 1H), 2.57 (t, $J = 8$ Hz, 2H), 2.32 (m, 6H), 1.62–1.55 (m, 2H), 1.37–1.27 (m, 4H), 1.24 (t, $J = 7.2$ Hz, 3H), 0.88 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 196.7, 170.6, 165.5, 160.2, 143.2, 131.0, 129.2, 126.7, 112.6, 62.1, 45.6, 45.3, 35.4, 31.4, 30.9, 30.2, 22.4, 19.4, 13.93, 13.91$. The enantiomeric excess was determined by HPLC with an OD-H column. (*n*-hexane:*i*-PrOH = 92:8), 1 mL/min; minor enantiomer $t_R = 9.61$ min, major enantiomer $t_R = 12.01$ min. HRMS (ESI): $[\text{M}+\text{H}]^+$ calcd for $[\text{C}_{22}\text{H}_{28}\text{O}_5]$: 373.2010, found: 373.2007.

(3S,4R)-ethyl

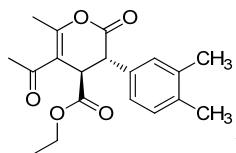
5-acetyl-3-(4-cyclohexylphenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3o)



Yellow Liquid. $[\alpha]_D^{20} = 190$ (*c* 1.0, CH₂Cl₂, 96% ee); IR(KBr): 737, 1057, 1148, 1251, 1362, 1448, 1644, 1733, 1788, 2926, 3373 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.18 (d, *J* = 8.4 Hz, 2H), 7.12 (d, *J* = 8.4 Hz, 2H), 4.33 (d, *J* = 2.4 Hz, 1H), 4.21–4.16 (q, *J* = 7.2 Hz, 2H), 4.09 (d, *J* = 1.6 Hz, 1H), 2.50 (d, *J* = 10.4 Hz, 1H), 2.34 (d, 6H), 1.84 (d, *J* = 8 Hz, 4H), 1.54 (d, *J* = 8.8 Hz, 1H), 1.42–1.32 (m, 4H), 1.21–1.21 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.7, 170.6, 165.5, 160.2, 148.2, 131.1, 127.6, 126.7, 112.5, 62.0, 45.5, 45.3, 44.0, 34.2, 30.2, 26.7, 26.0, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an OD-H column. (*n*-hexane:*i*-PrOH = 95:5), 1 mL/min; minor enantiomer *t_R* = 16.08 min, major enantiomer *t_R* = 21.30 min. HRMS (ESI): [M+H]⁺ calcd for [C₂₃H₂₈O₅]: 385.2010, found: 385.2011.

(3S,4R)-ethyl

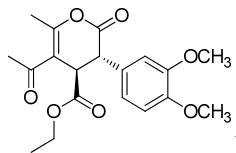
5-acetyl-3-(3,4-dimethylphenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3p)



Yellow Liquid. $[\alpha]_D^{20} = 152$ (*c* 1.0, CH₂Cl₂, 96% ee); IR(KBr): 737, 1040, 1148, 1381, 1454, 1609, 1732, 1789, 2375, 2924, 3348 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.09 (d, *J* = 8 Hz, 1H), 6.96 (s, 1H), 6.88 (d, *J* = 1.6 Hz, 1H), 4.30 (d, *J* = 2 Hz, 1H), 4.22–4.17 (q, *J* = 7.2 Hz, 2H), 4.08 (d, *J* = 1.2 Hz, 1H), 2.33 (d, 6H), 2.23 (s, 6H), 1.25 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.7, 170.7, 165.6, 160.3, 137.5, 136.8, 131.1, 130.3, 128.1, 123.8, 112.5, 62.1, 45.6, 45.2, 30.2, 19.8, 19.4, 19.3, 13.9. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer *t_R* = 12.81 min, minor enantiomer *t_R* = 15.79 min. HRMS (ESI): [M+H]⁺ calcd for [C₁₉H₂₂O₅]: 331.1540, found: 331.1538.

(3S,4R)-ethyl

5-acetyl-3-(3,4-dimethoxyphenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3q)

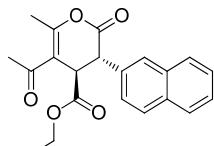


Yellow Liquid. $[\alpha]_D^{20} = 114$ (*c* 1.0, CH₂Cl₂, 96% ee); IR(KBr): 774, 1043, 1117, 1190, 1421, 1596, 1786, 2375, 2924, 3344 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 6.28 (d, *J* = 8.4 Hz, 1H), 6.74 (t, 2H), 4.32 (d, *J* = 2 Hz, 1H), 4.22–4.17 (q, *J* = 7.2 Hz, 2H), 4.13 (s,

1H), 3.86 (d, 6H), 2.37 (s, 3H), 2.31 (s, 3H), 1.25 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 196.8, 170.6, 165.6, 159.9, 149.4, 149.0, 125.9, 118.9, 113.0, 111.5, 110.3, 62.1, 55.9, 55.88, 45.1, 45.0, 30.3, 19.4, 14.0$. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer $t_R = 29.38$ min, minor enantiomer $t_R = 35.93$ min. HRMS (ESI): $[\text{M}+\text{H}]^+$ calcd for $[\text{C}_{19}\text{H}_{22}\text{O}_7]$: 363.1438, found: 363.1442.

(3S,4R)-ethyl

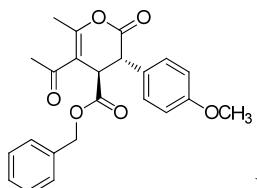
5-acetyl-6-methyl-3-(naphthalen-2-yl)-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3r)



Yellow Liquid. $[\alpha]_D^{20} = 92$ (c 1.0, CH_2Cl_2 , 96% ee); IR(KBr): 738, 1038, 1149, 1261, 1362, 1605, 1731, 1787, 2371, 2927, 3298 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): $\delta = 7.82\text{--}7.76$ (m, 3H), 7.61 (s, 1H), 7.25–7.48 (m, 2H), 7.36–7.34 (dd, $J = 1.6$ Hz, $J = 6$ Hz, 1H), 4.54 (d, $J = 2.4$ Hz, 1H), 4.27 (d, $J = 1.6$ Hz, 1H), 4.23–4.18 (q, $J = 7.2$ Hz, 2H), 2.33 (s, 6H), 1.24 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 196.6, 170.6, 165.4, 160.2, 133.2, 132.8, 131.1, 129.2, 127.9, 127.6, 126.7, 126.6, 125.9, 124.4, 112.8, 62.1, 45.8, 45.4, 30.3, 19.4, 13.9$. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer $t_R = 40.64$ min, minor enantiomer $t_R = 63.10$ min. HRMS (ESI): $[\text{M}+\text{H}]^+$ calcd for $[\text{C}_{21}\text{H}_{20}\text{O}_5]$: 353.1384, found: 353.1386.

(3S,4R)-benzyl

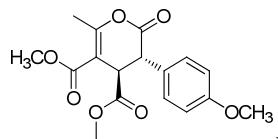
5-acetyl-3-(4-methoxyphenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3s)



Yellow Liquid. $[\alpha]_D^{20} = 115$ (c 1.0, CH_2Cl_2 , 96% ee); IR(KBr): 787, 1042, 1117, 1260, 1422, 1596, 1786, 2370, 2924, 3363 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): $\delta = 7.39\text{--}7.32$ (m, 3H), 7.31–7.25 (m, 2H), 7.11 (d, $J = 8.4$ Hz, 2H), 6.85–6.83 (m, 2H), 5.15 (s, 2H), 4.31 (d, $J = 2.8$ Hz, 1H), 4.14–4.13 (m, 1H), 3.77 (s, 3H), 2.29 (d, $J = 0.8$ Hz, 3H), 2.27 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 196.6, 170.5, 165.5, 160.1, 159.5, 134.9, 128.6, 128.5, 128.2, 128.0, 125.6, 114.6, 112.7, 67.7, 55.2, 45.5, 44.9, 30.2, 19.4$. The enantiomeric excess was determined by HPLC with an OD-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; minor enantiomer $t_R = 27.13$ min, major enantiomer $t_R = 29.99$ min. HRMS (ESI): $[\text{M}+\text{H}]^+$ calcd for $[\text{C}_{23}\text{H}_{22}\text{O}_6]$: 395.1489, found: 395.1494.

(3S,4R)-4-ethyl5-methyl

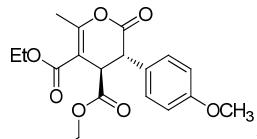
3-(4-methoxyphenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4,5-dicarboxylate (3t)



Yellow Liquid. $[\alpha]_D^{20} = 108$ (*c* 1.0, CH₂Cl₂, 95% ee); IR(KBr): 795, 1032, 1149, 1259, 1379, 1462, 1514, 1655, 1726, 1788, 2956, 3434 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.15 (d, *J* = 8.8 Hz, 2H), 6.88 (d, *J* = 8.8 Hz, 2H), 4.26 (d, *J* = 2.8 Hz, 1H), 4.19–4.14 (m, 2H), 4.10–4.09 (m, 1H), 3.79 (s, 3H), 3.77 (s, 3H), 2.38 (d, *J* = 0.8 Hz, 3H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 170.8, 165.9, 165.7, 162.0, 159.4, 128.2, 125.7, 114.5, 104.1, 61.8, 55.2, 52.0, 45.2, 45.1, 18.8, 13.9. The enantiomeric excess was determined by HPLC with an OD-H column. (*n*-hexane:*i*-PrOH = 90:10), 1 mL/min; major enantiomer *t*_R = 14.51 min, minor enantiomer *t*_R = 19.87 min. HRMS (ESI): [M+H]⁺ calcd for [C₁₈H₂₀O₇]: 349.1282, found: 349.1286.

(3S,4R)-diethyl

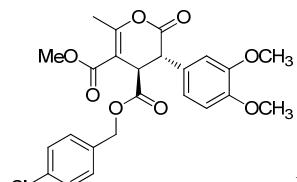
3-(4-methoxyphenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4,5-dicarboxylate (3u)



Yellow Liquid. $[\alpha]_D^{20} = 115$ (*c* 1.0, CH₂Cl₂, 96% ee); IR(KBr): 799, 1072, 1260, 1376, 1461, 1611, 1655, 1721, 1787, 2923, 3371 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.16 (d, *J* = 7.6 Hz, 2H), 6.88 (d, *J* = 8.8 Hz, 2H), 4.29–4.10 (m, 6H), 3.79 (s, 3H), 2.38 (d, *J* = 0.8 Hz, 3H), 1.30 (t, *J* = 7.2 Hz, 3H), 1.21 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 170.9, 165.9, 165.4, 161.7, 159.4, 128.2, 125.7, 114.4, 104.4, 61.8, 61.1, 55.2, 45.3, 45.0, 18.7, 14.1, 13.9. The enantiomeric excess was determined by HPLC with an OD-H column. (*n*-hexane:*i*-PrOH = 90:10), 1 mL/min; major enantiomer *t*_R = 13.34 min, minor enantiomer *t*_R = 19.39 min. HRMS (ESI): [M+H]⁺ calcd for [C₁₉H₂₂O₇]: 363.1438, found: 363.1444.

(3S,4R)-4-(4-chlorobenzyl) 5-ethyl

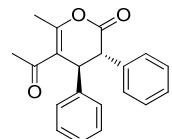
3-(3,4-dimethoxyphenyl)-6-methyl-2-oxo-3,4-dihydro-2H-pyran-4,5-dicarboxylate (3v)



Yellow Liquid. $[\alpha]_D^{20} = 69$ (*c* 1.0, CH₂Cl₂, 83% ee); IR(KBr): 801, 1076, 1261, 1460, 1597, 1688, 1735, 1786, 2369, 2925, 3371 cm⁻¹; ¹H NMR (400 MHz,

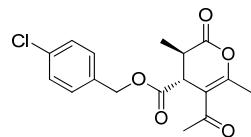
CDCl_3): $\delta = 7.33\text{--}7.29$ (m, 2H), 7.20 (d, $J = 8.4$ Hz, 2H), 6.80 (d, $J = 8.4$ Hz, 1H), 6.75–6.69 (m, 2H), 5.09 (s, 2H), 4.26 (d, $J = 3.6$ Hz, 1H), 4.12–4.20 (m, 1H), 3.85 (s, 3H), 3.84 (s, 3H), 3.72 (s, 3H), 2.36 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 170.5, 165.7, 165.5, 161.9, 149.1, 148.9, 134.2, 133.5, 129.3, 128.6, 125.5, 119.0, 111.2, 110.2, 104.1, 66.4, 55.72, 55.7, 52.0, 45.2, 44.7, 18.7$. The enantiomeric excess was determined by HPLC with an OD-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer $t_R = 34.98$ min, minor enantiomer $t_R = 54.46$ min. HRMS (ESI): $[\text{M}+\text{Na}]^+$ calcd for $[\text{C}_{24}\text{H}_{23}\text{Cl}_1\text{O}_8]$: 497.0974, found: 497.0982.

(3S,4S)-5-acetyl-6-methyl-3,4-diphenyl-3,4-dihydro-2H-pyran-2-one (3w)



Pale Yellow Liquid. $[\alpha]_D^{20} = 23$ (*c* 1.0, CH_2Cl_2 , 71% ee); IR(KBr): 740, 1042, 1140, 1268, 1356, 1621, 1740, 1785, 2375, 2933, 3331 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): $\delta = 7.38\text{--}7.28$ (m, 6H), 7.24 (d, $J = 7.2$ Hz, 2H), 7.19 (d, $J = 7.2$ Hz, 2H), 4.35 (s, 1H), 4.10 (d, $J = 2.4$ Hz, 1H), 2.42 (s, 3H), 2.09 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 197.8, 166.2, 159.3, 139.6, 135.5, 129.5, 129.2, 128.2, 128.0, 127.0, 126.9, 115.9, 52.9, 46.4, 30.0, 18.9$. The enantiomeric excess was determined by HPLC with an AD-H column. (*n*-hexane:*i*-PrOH = 90:10), 1 mL/min; minor enantiomer $t_R = 13.90$ min, major enantiomer $t_R = 19.88$ min. HRMS (ESI): $[\text{M}+\text{Na}]^+$ calcd for $[\text{C}_{20}\text{H}_{18}\text{O}_3]$: 329.1172, found: 329.1159.

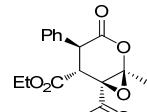
(3R,4R)-4-chlorobenzyl 5-acetyl-3,6-dimethyl-2-oxo-3,4-dihydro-2H-pyran-4-carboxylate (3x)



Pale Yellow Liquid. $[\alpha]_D^{20} = 42$ (*c* 1.0, CH_2Cl_2 , 88% ee); IR(KBr): 738, 1038, 1134, 1270, 1364, 1618, 1752, 1780, 2369, 2938, 3340 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3): $\delta = 7.36\text{--}7.32$ (m, 2H), 7.25 (d, $J = 8.8$ Hz, 2H), 5.10 (s, 2H), 3.64 (d, $J = 2.4$ Hz, 1H), 3.15–3.09 (qd, $J = 3.2$ Hz, $J = 4$ Hz, 1H), 2.36 (s, 3H), 2.33 (s, 3H), 1.29 (d, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): $\delta = 196.8, 170.7, 167.7, 160.1, 134.4, 133.4, 129.6, 128.8, 111.9, 66.6, 45.8, 35.3, 30.3, 19.4, 15.3$. The enantiomeric excess was determined by HPLC with an AS-H column. (*n*-hexane:*i*-PrOH = 85:15), 1 mL/min; minor enantiomer $t_R = 29.03$ min, major enantiomer $t_R = 34.01$ min. HRMS (ESI): $[\text{M}+\text{Na}]^+$ calcd for $[\text{C}_{17}\text{H}_{17}\text{ClO}_5]$: 359.0657, found: 359.0672.

(1R,4S,5R,6R)-ethyl

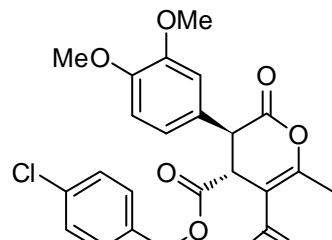
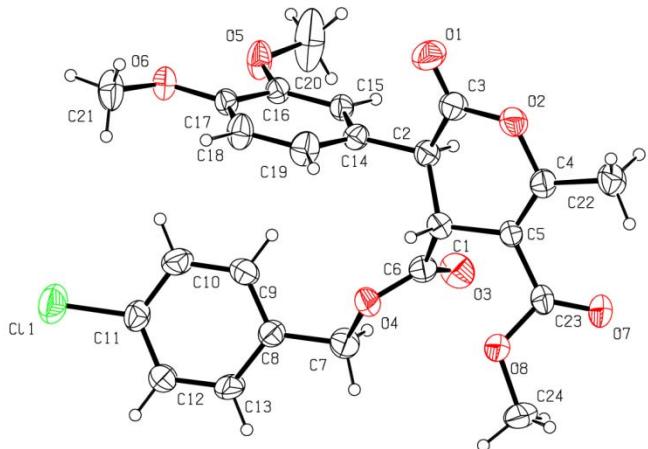
6-acetyl-1-methyl-3-oxo-4-phenyl-2,7-dioxabicyclo[4.1.0]heptane-5-carboxylate (6a)



Pale Yellow Liquid. $[\alpha]_D^{20} = 39$ (*c* 1.0, CH₂Cl₂, 95% ee); IR(KBr): 749, 1028, 1155, 1262, 1383, 1598, 1730, 2373, 2924, 3364 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ = 7.37–7.29 (m, 3H), 7.21 (d, *J* = 6.8 Hz, 2H), 4.37 (d, *J* = 2.4 Hz, 1H), 4.22–4.17 (q, *J* = 7.2 Hz, 2H), 4.11 (d, *J* = 1.2 Hz, 1H), 2.33 (d, 6H), 1.24 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ = 196.7, 170.6, 165.4, 160.2, 133.9, 129.2, 129.15, 128.4, 126.9, 112.7, 62.2, 45.7, 45.6, 30.2, 19.4, 13.9. The enantiomeric excess was determined by HPLC with an OJ-H column. (*n*-hexane:*i*-PrOH = 80:20), 1 mL/min; major enantiomer *t*_R = 15.36 min, minor enantiomer *t*_R = 17.89 min. HRMS (ESI): [M+Na]⁺ calcd for [C₁₇H₁₈O₆]: 341.0996, found: 341.0995.

5. X-ray Crystallographic Data

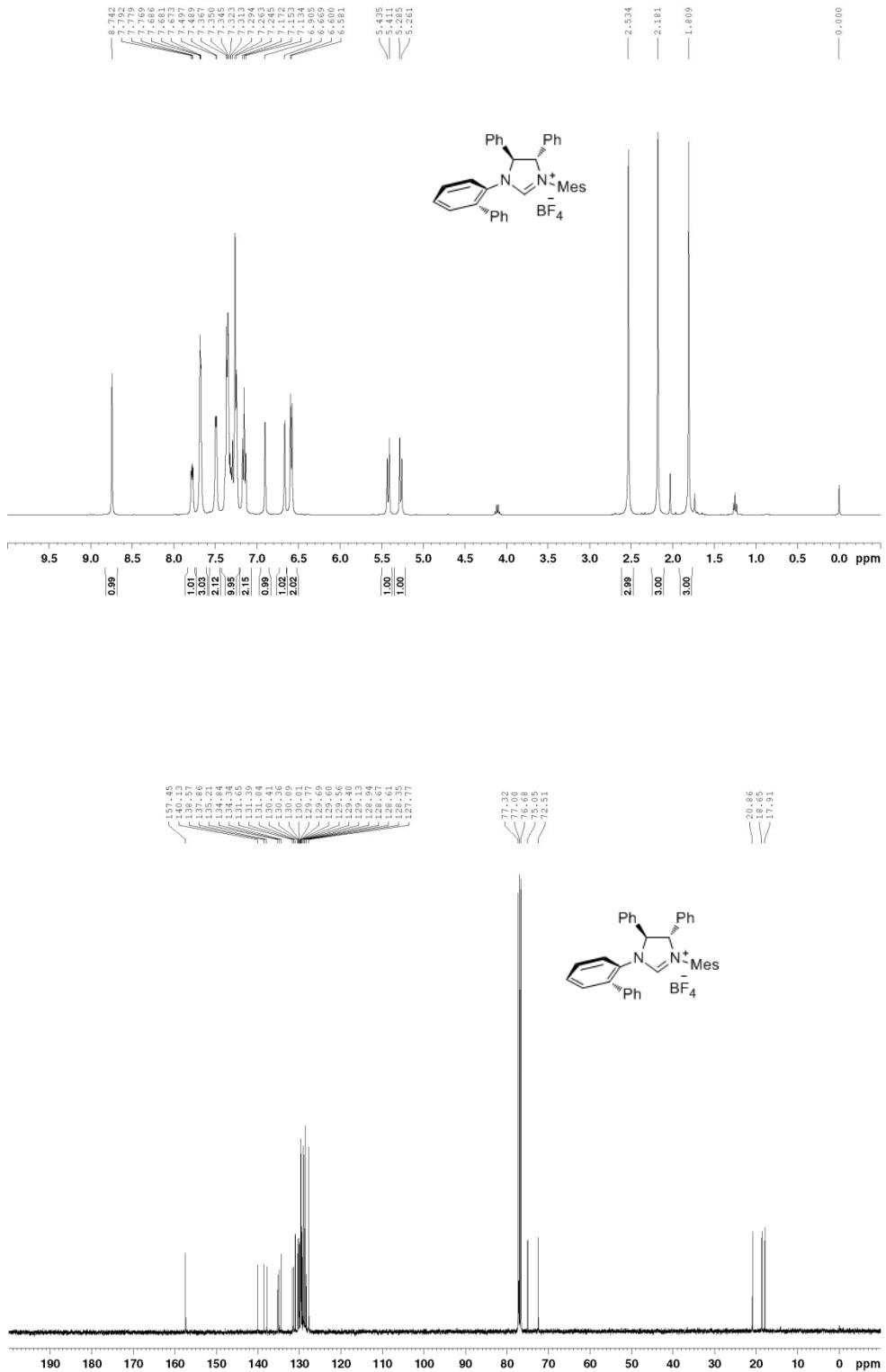
X-ray Crystallographic Data of Compound 4d

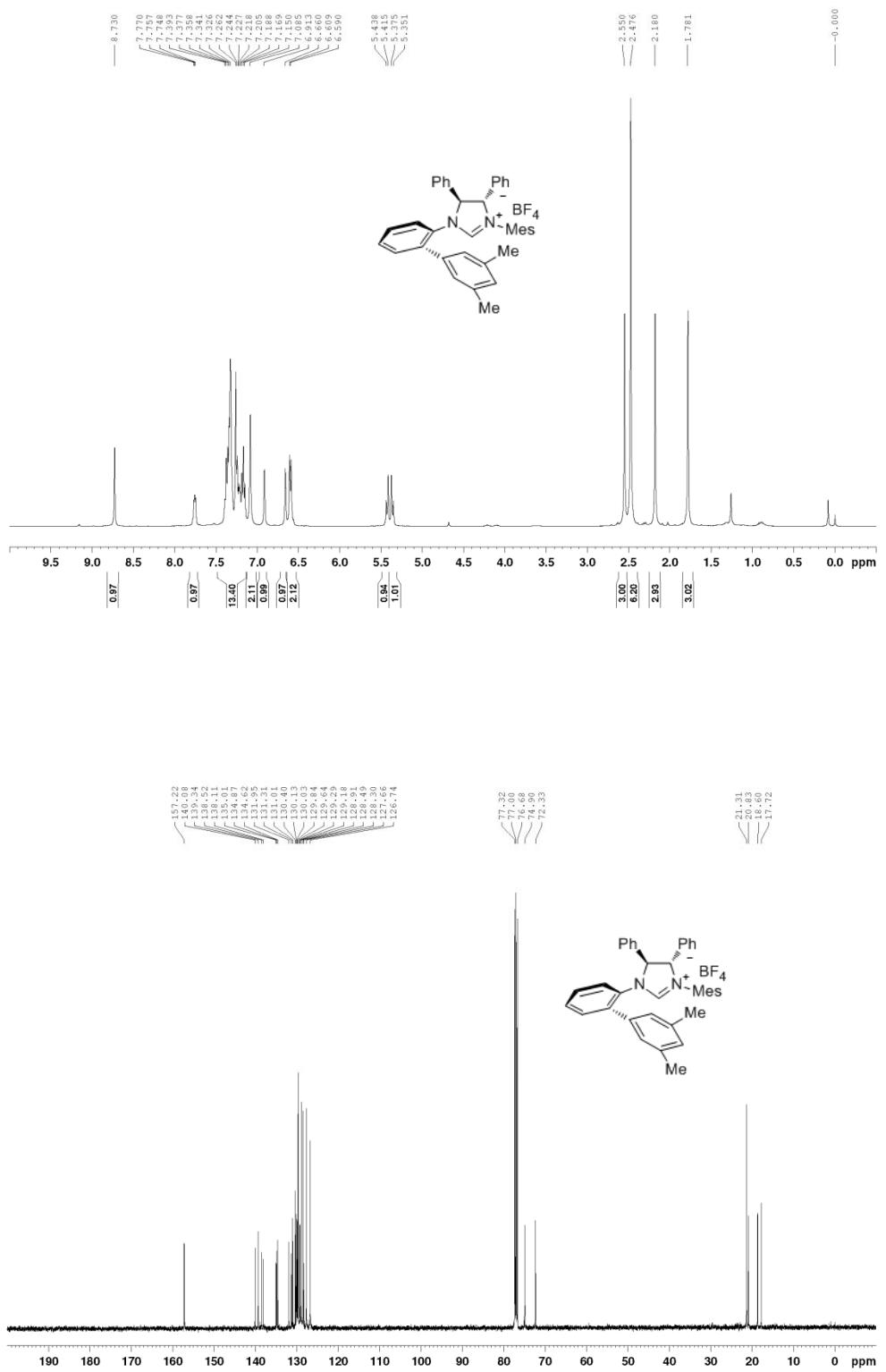


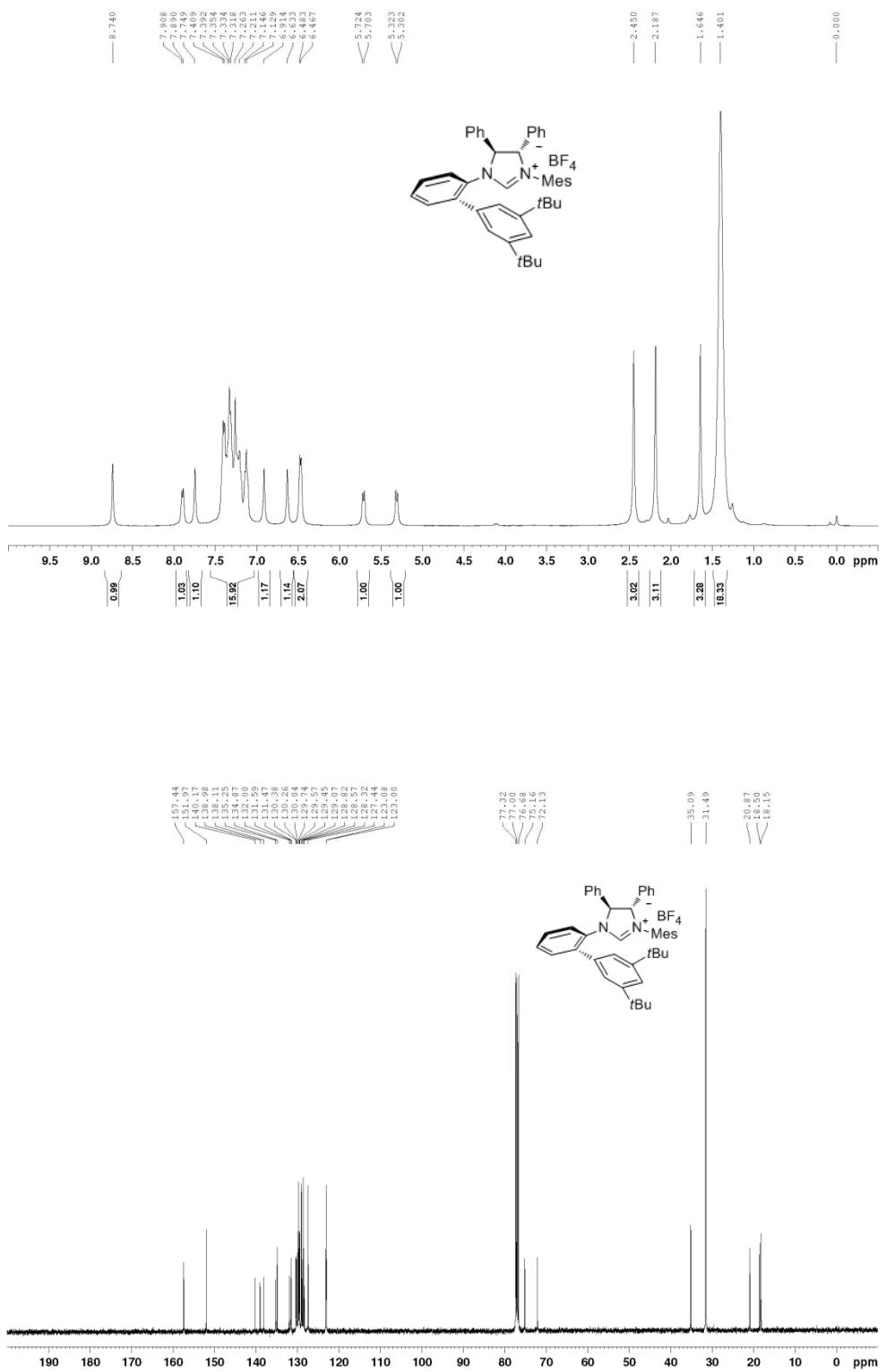
CCDC 1037041

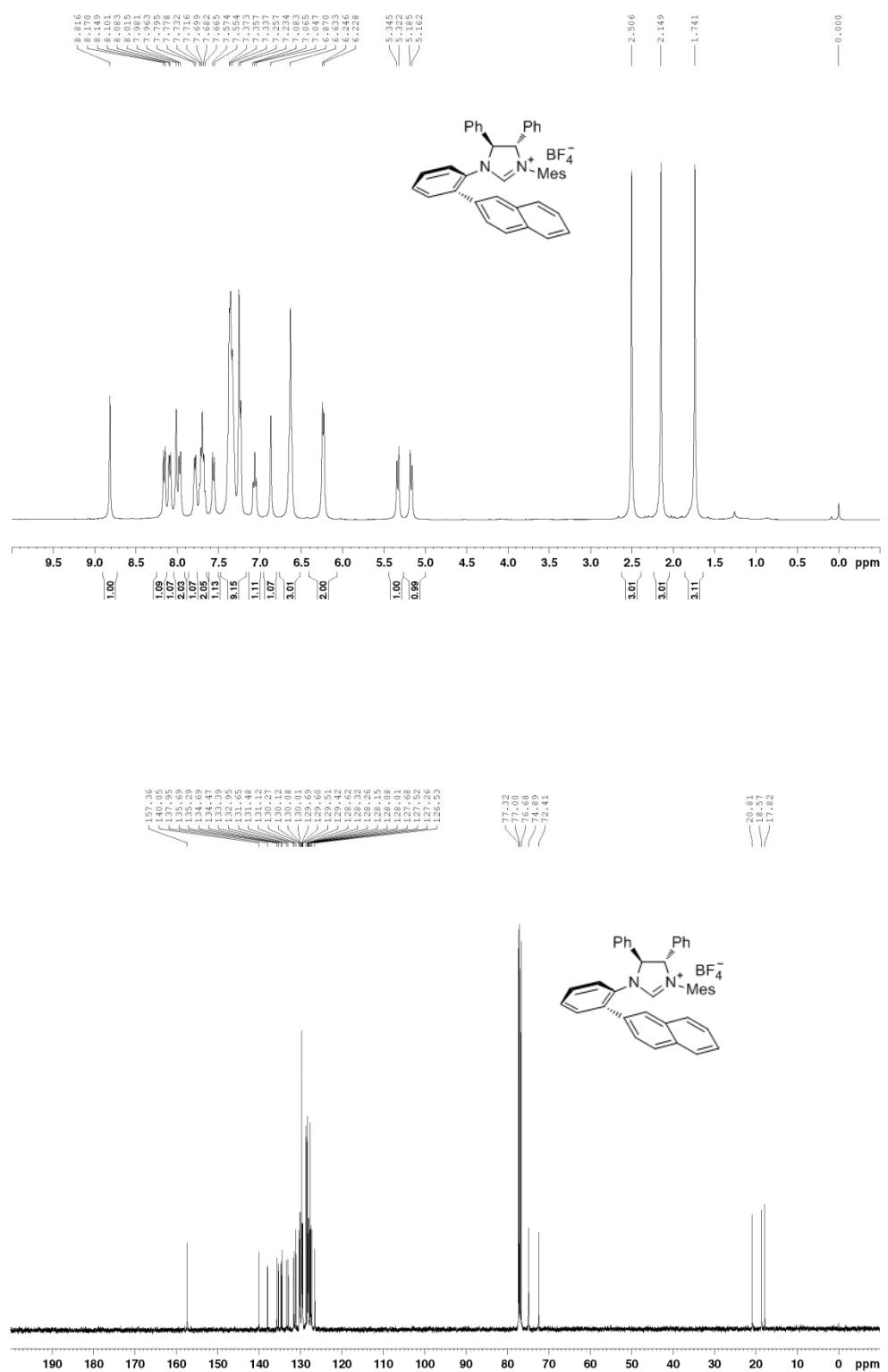
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Cell:	a=9.2814(16) b=11.7110(17)	c=12.1339(13)
	alpha=113.081(12) beta=96.350(11)	gamma=102.204(14)
Temperature:	292 K	
	Calculated	Reported
Volume	1158.1(3)	1158.1(3)
Space group	P -1	P -1
Hall group	-P 1	-P 1
Moiety formula	C ₂₄ H ₂₃ ClO ₈	C ₂₄ H ₂₃ ClO ₈
Sum formula	C ₂₄ H ₂₃ ClO ₈	C ₂₄ H ₂₃ ClO ₈
Mr	474.87	474.87
Dx,g cm ⁻³	1.362	1.362
Z	2	2
Mu (mm ⁻¹)	0.212	0.212
F000	496.0	496.0
F000'	496.58	
h,k,lmax	11,14,14	11,14,14
Nref	4569	4553
Tmin,Tmax	0.948,0.954	0.975,1.000
Tmin'	0.948	
Correction method=	MULTI-SCAN	
Data completeness=	0.996	Theta(max)= 26.020
R(reflections)=	0.0652(2049)	wR2(reflections)= 0.1818(4553)
S =	1.030	Npar= 302

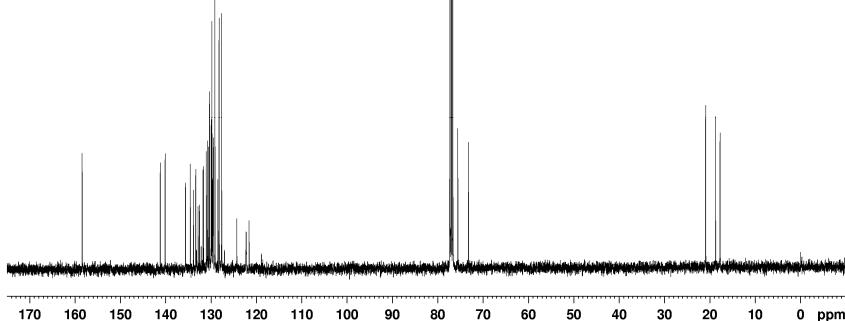
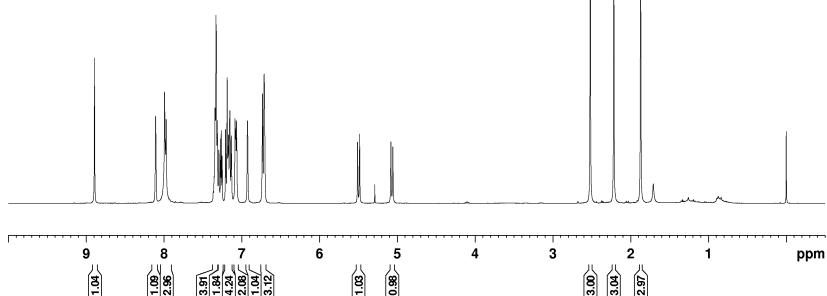
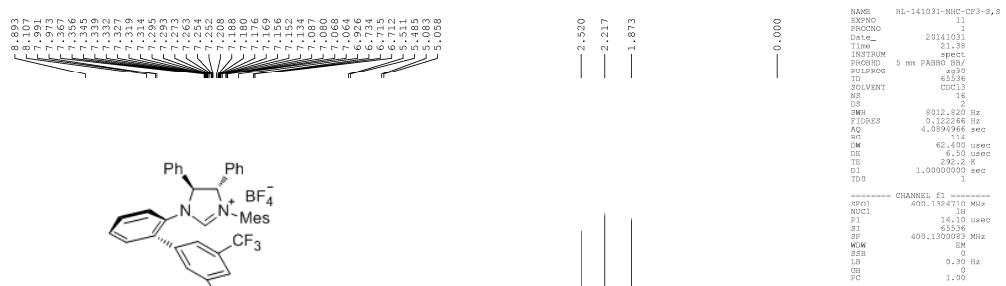
6. NMR and HPLC spectrogram

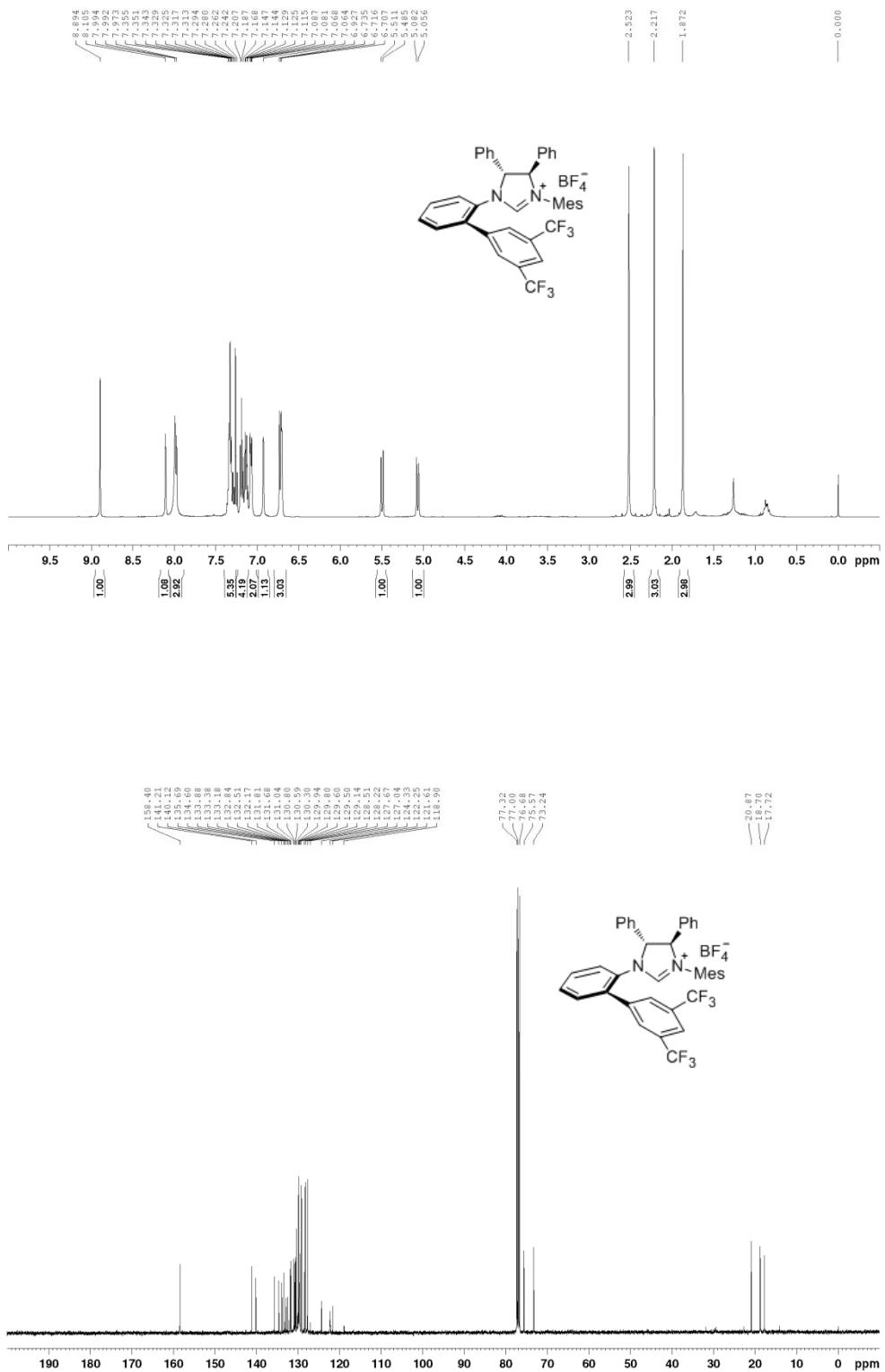


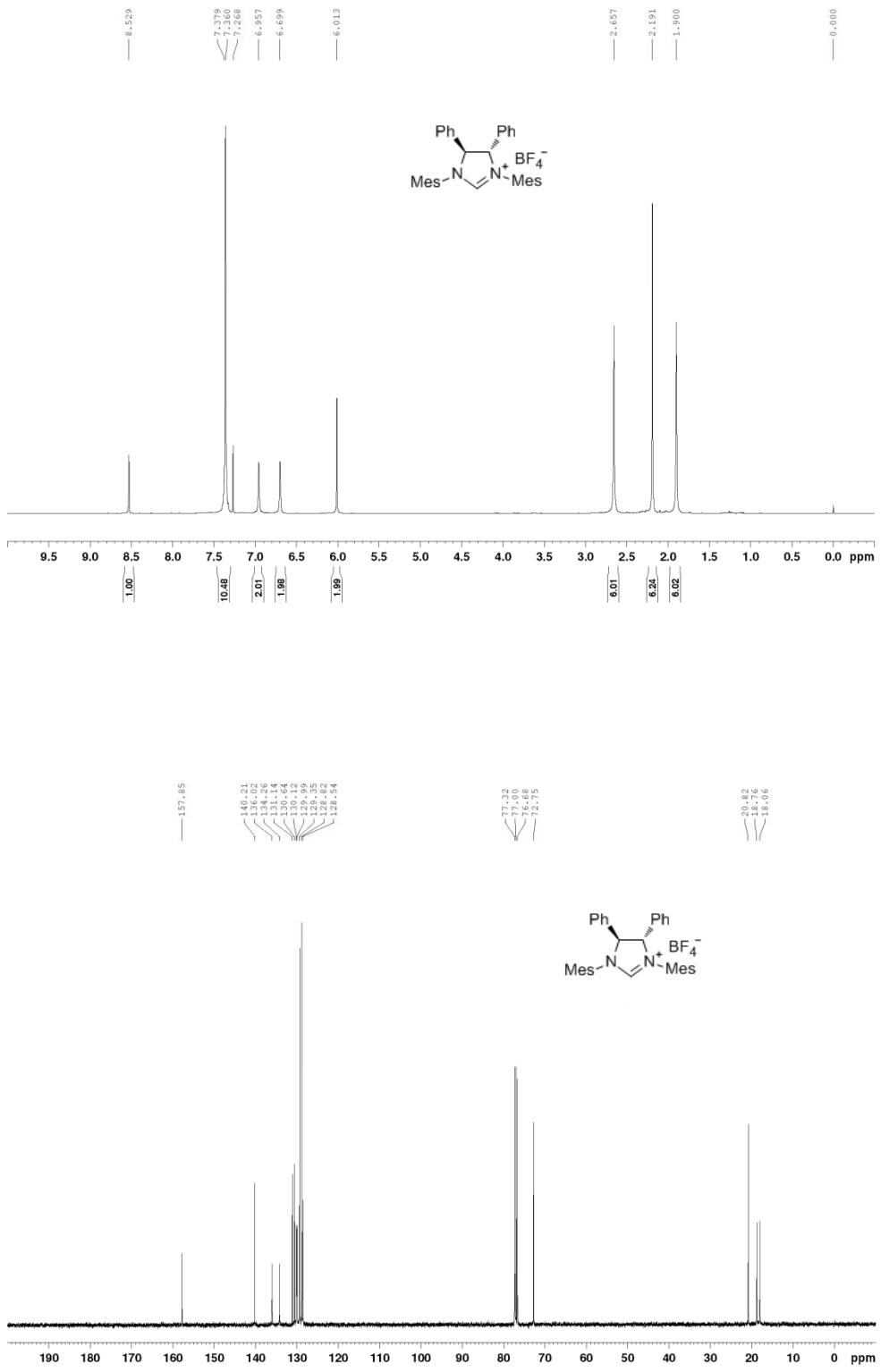


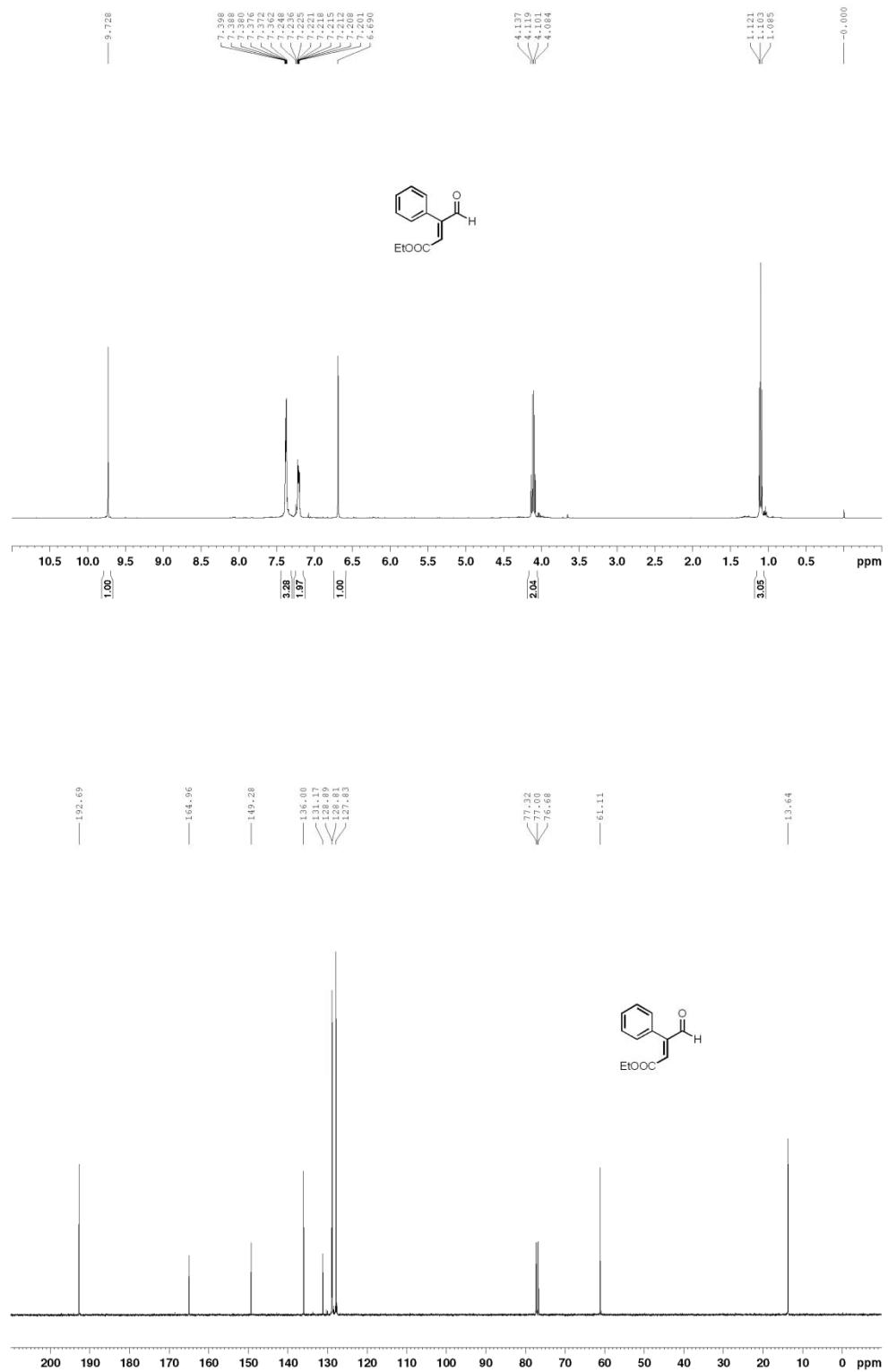


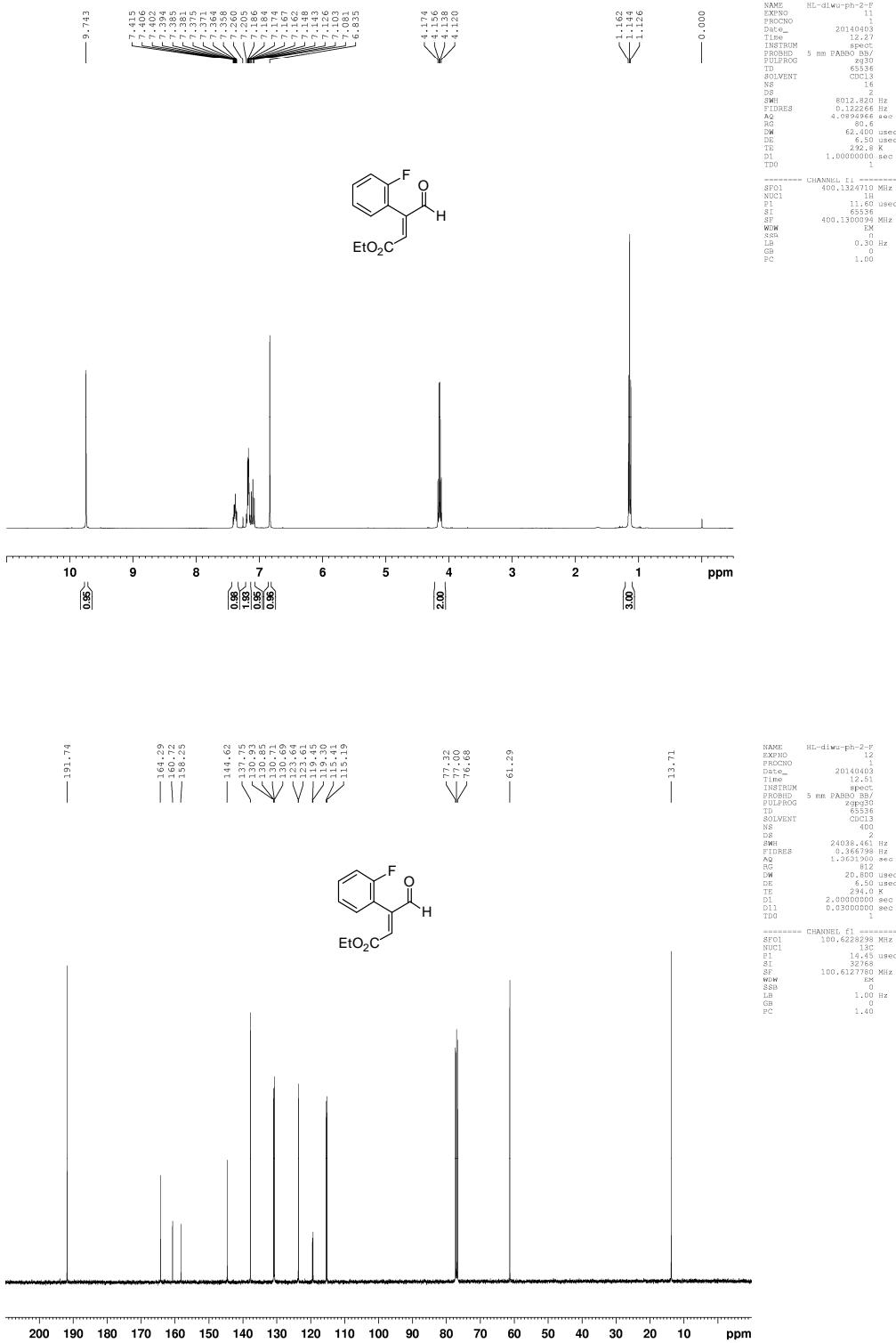


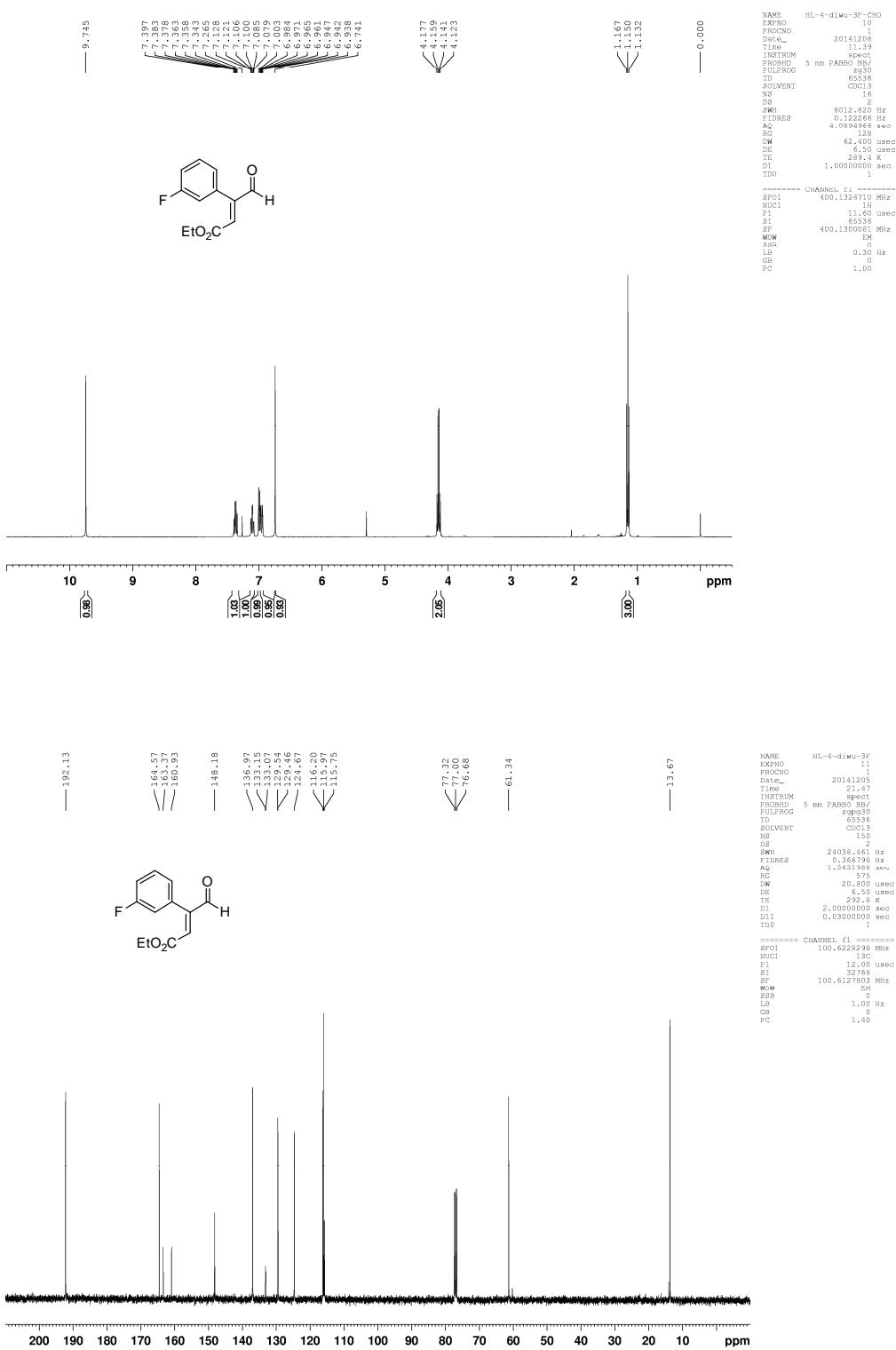


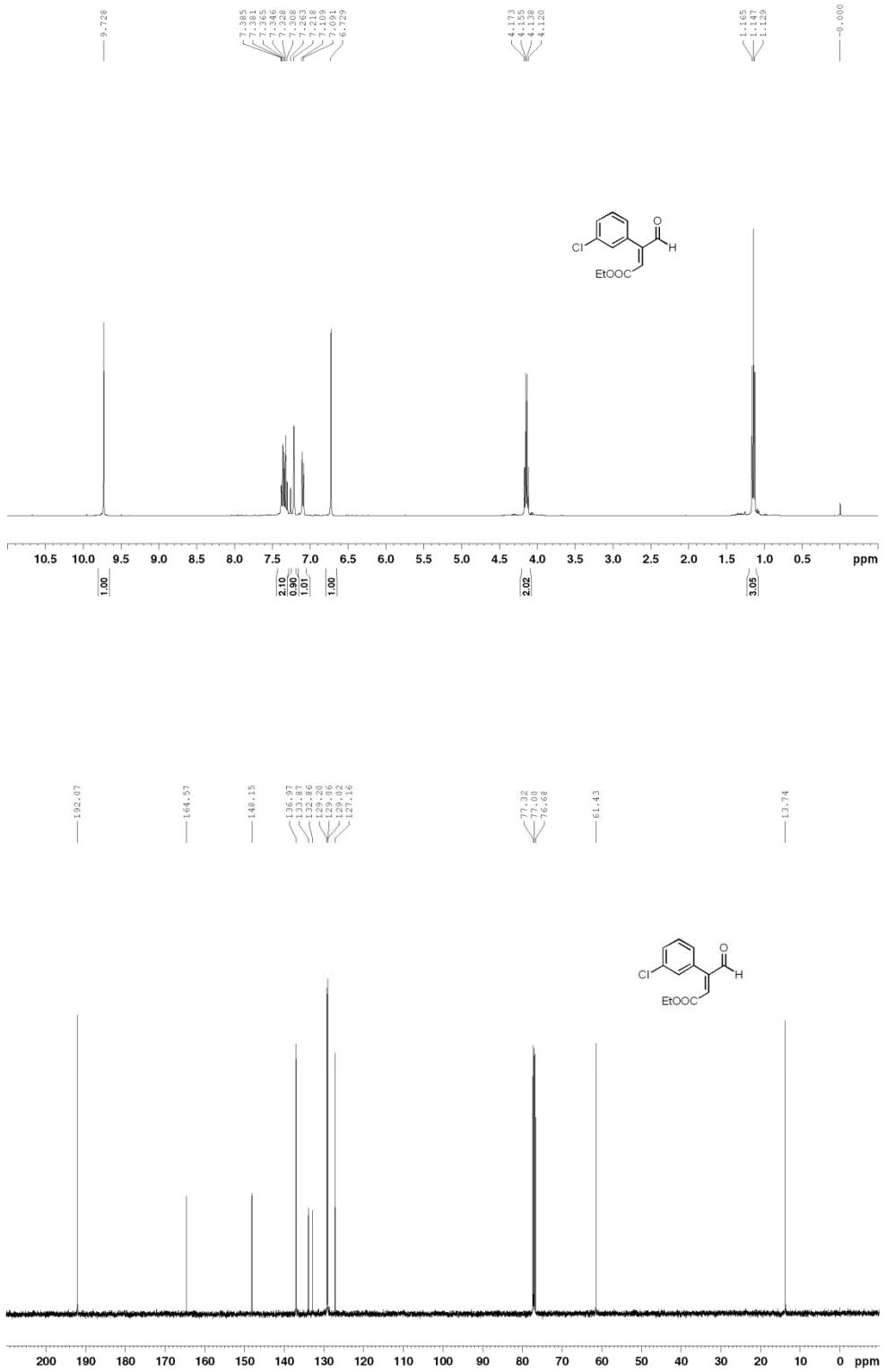


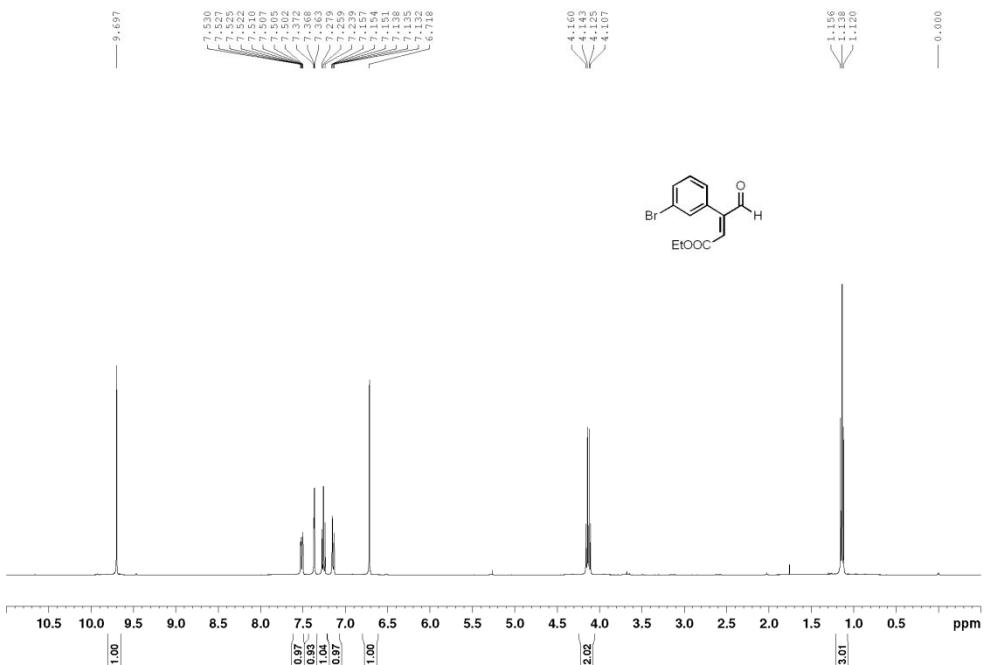


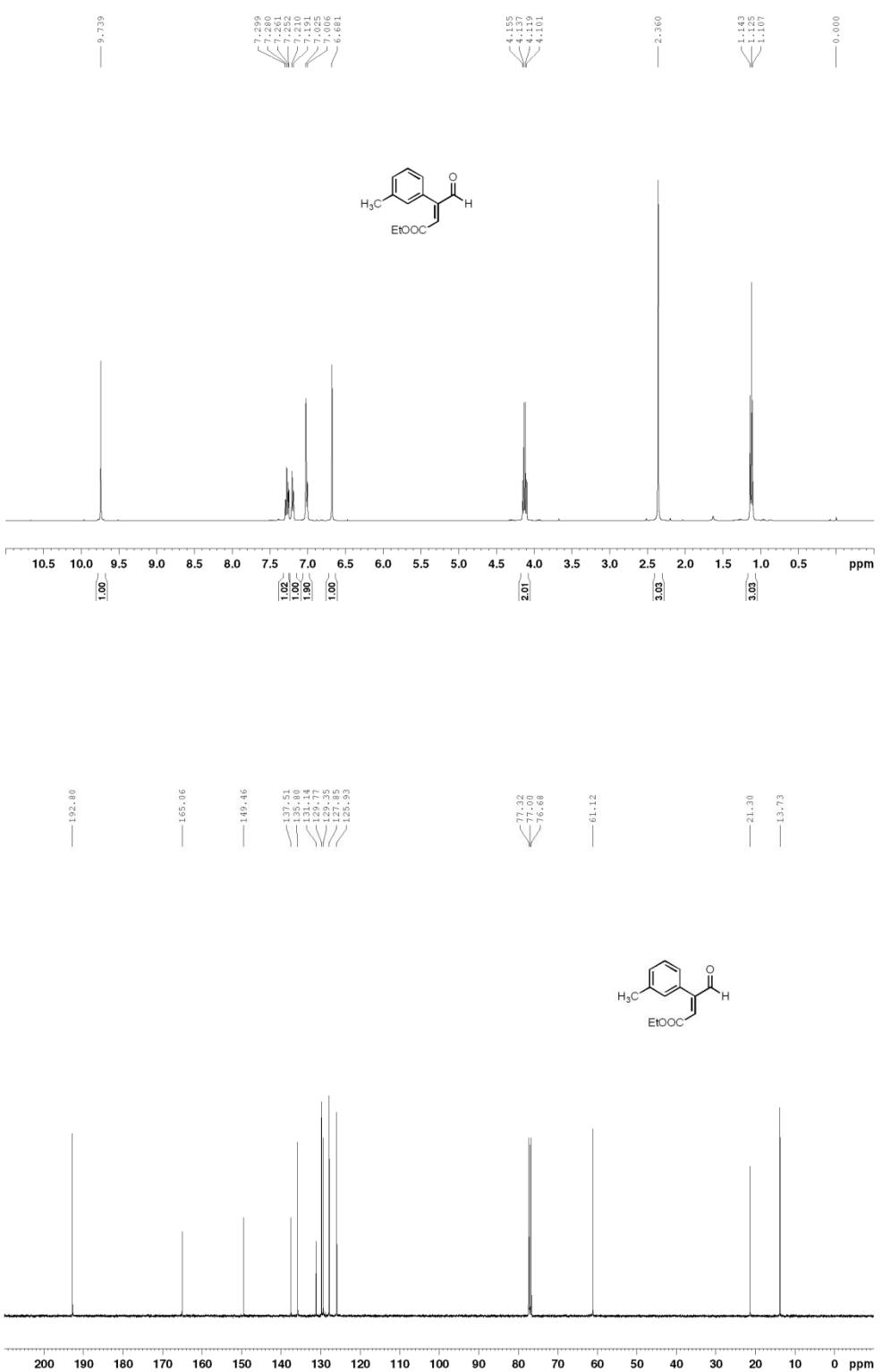


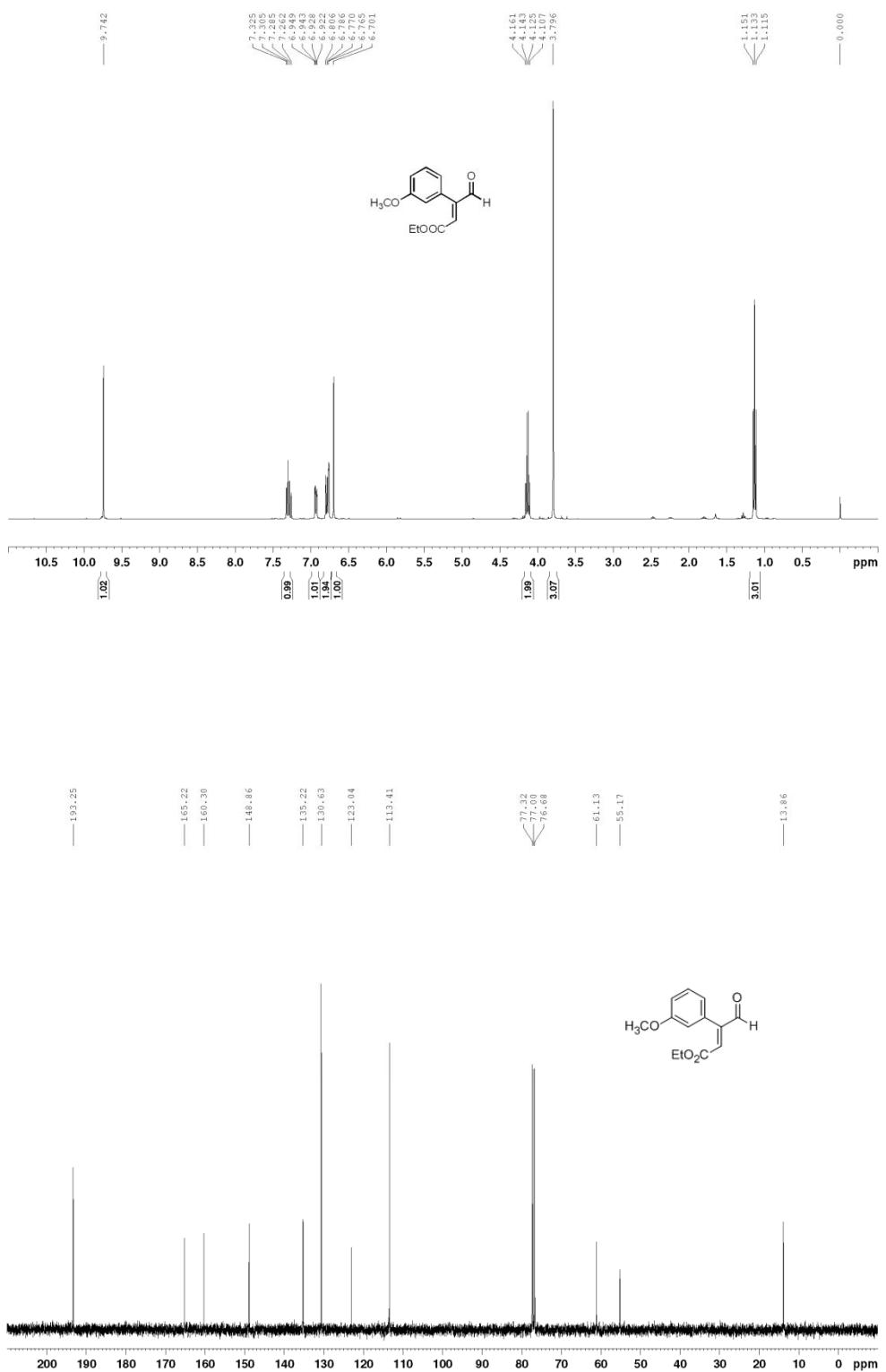


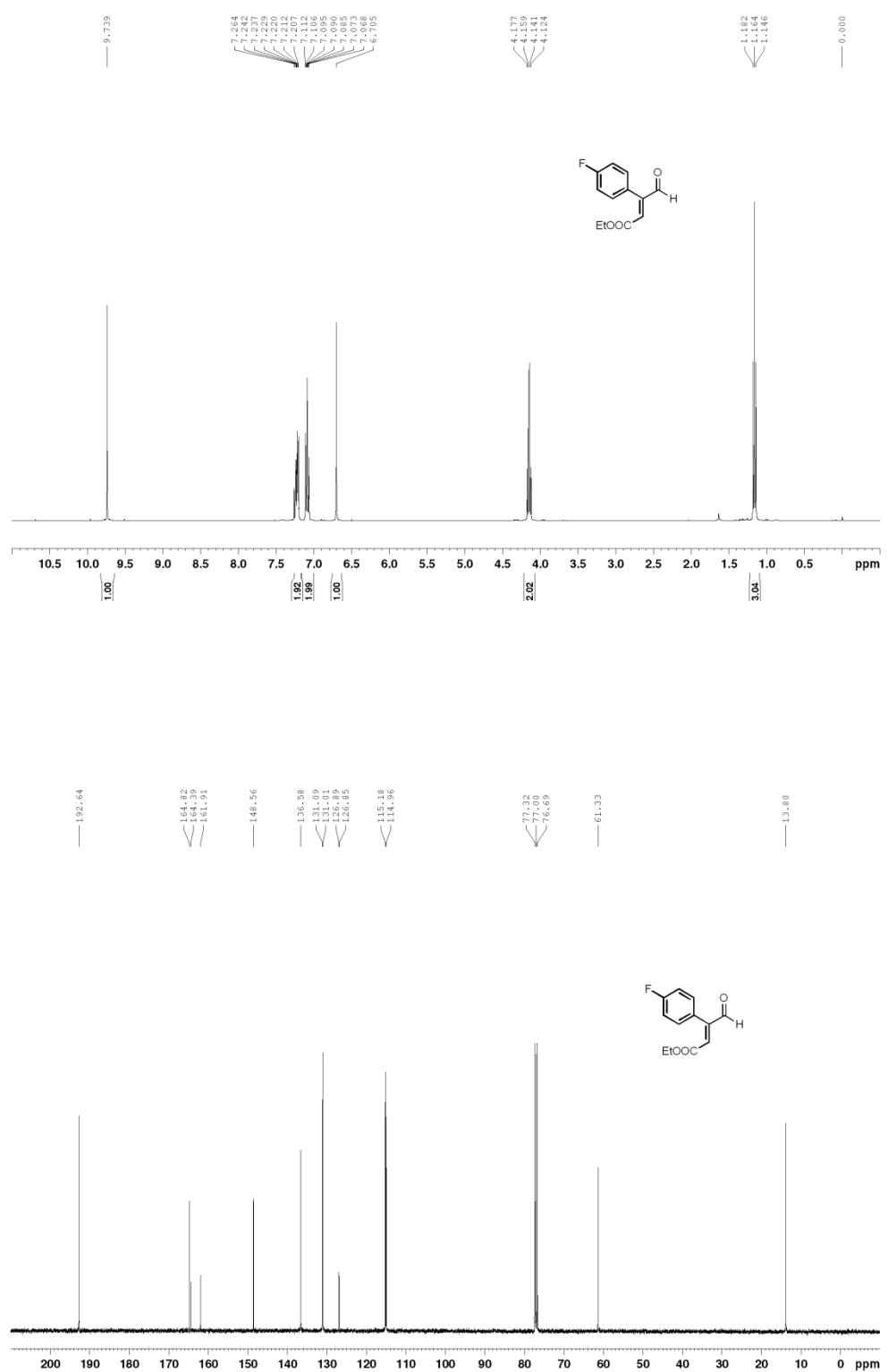


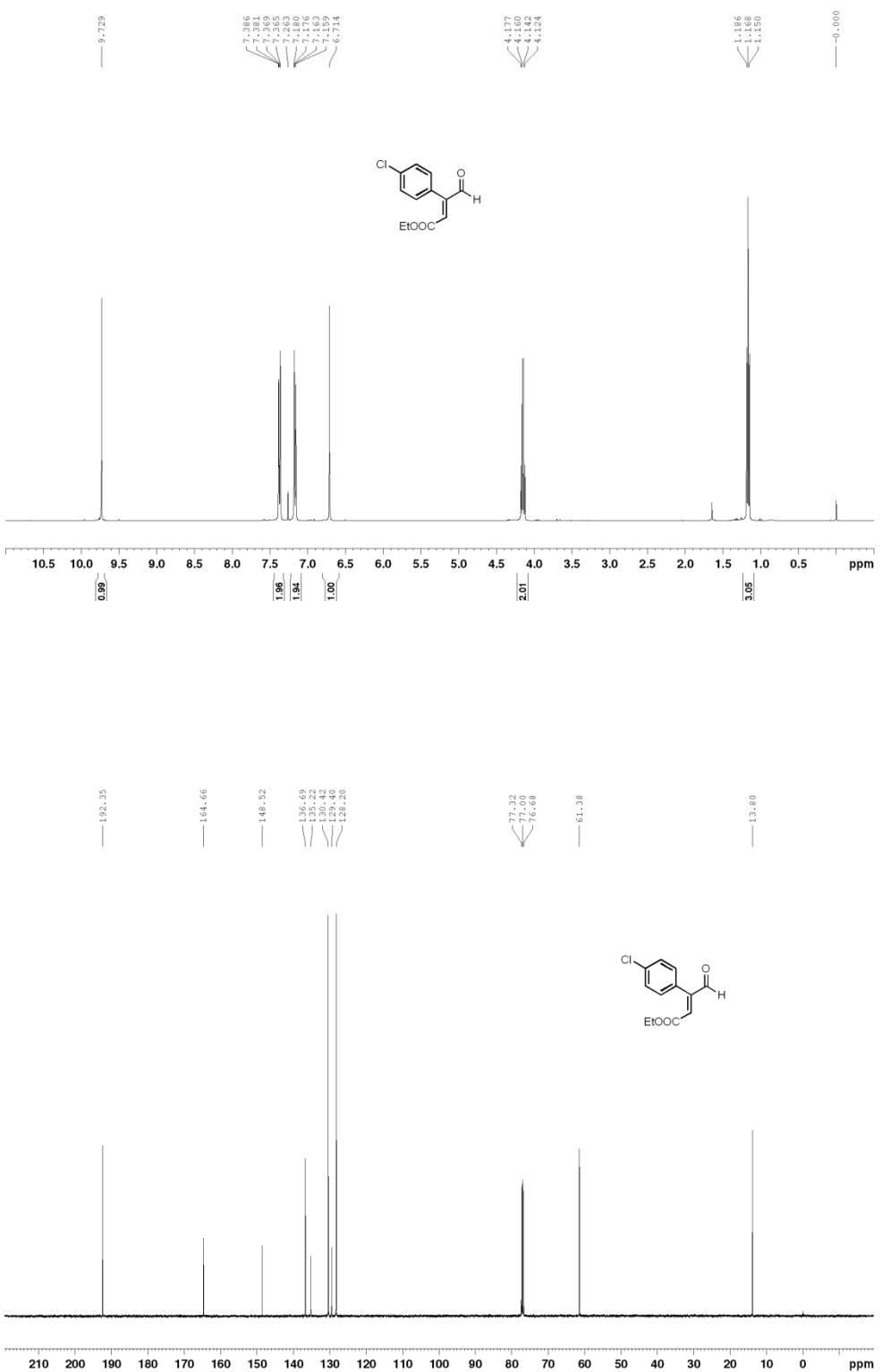


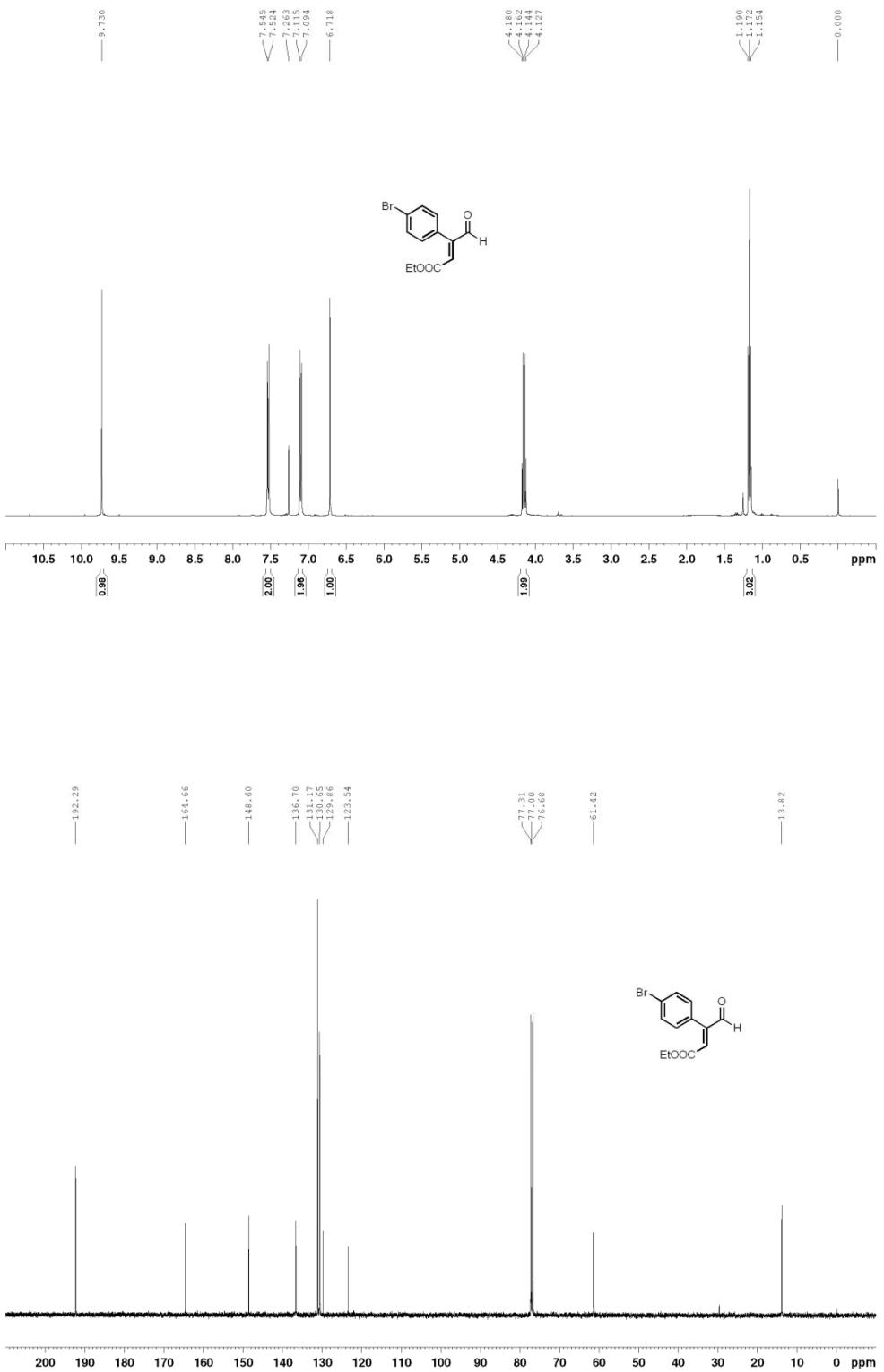


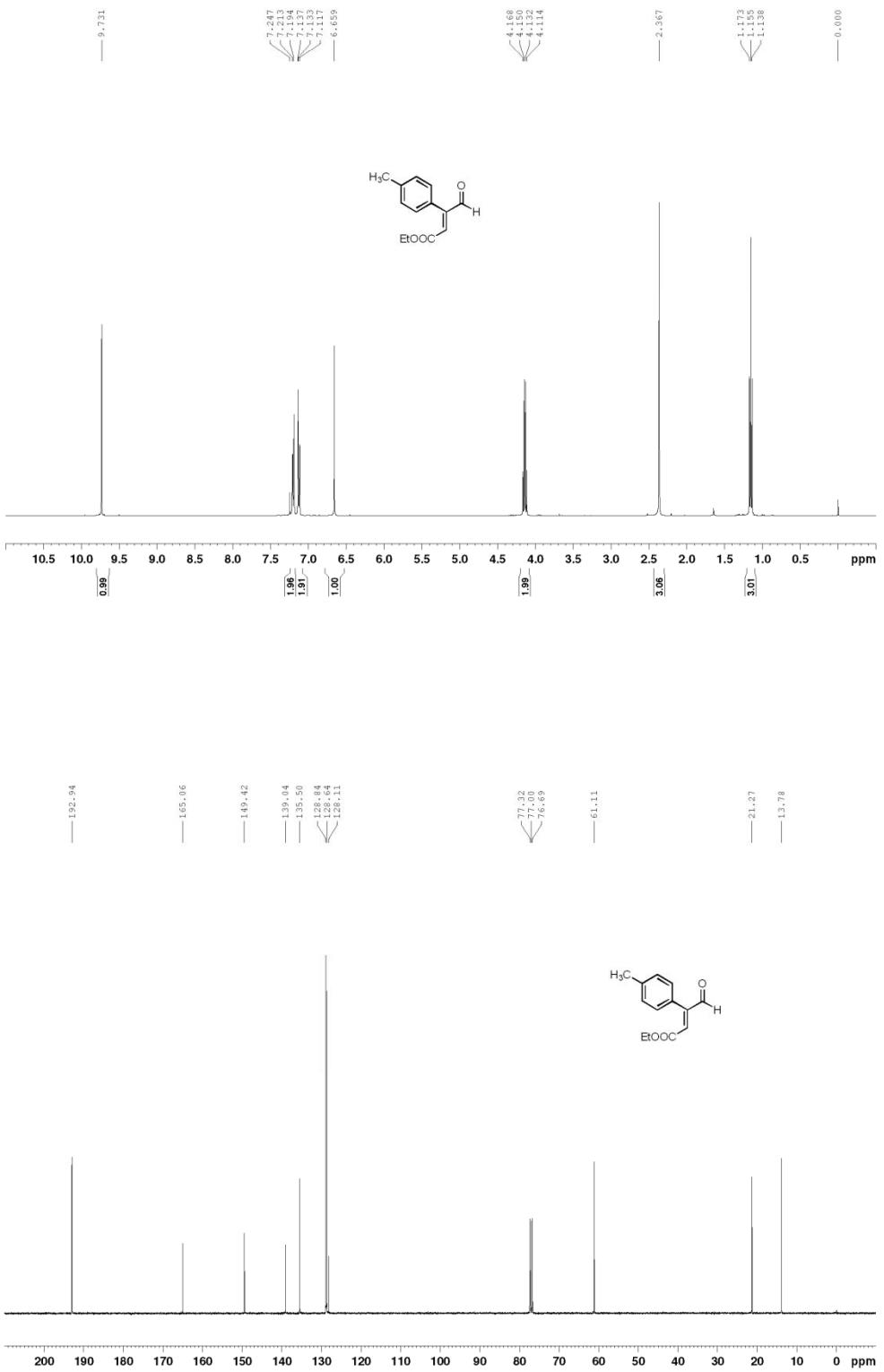


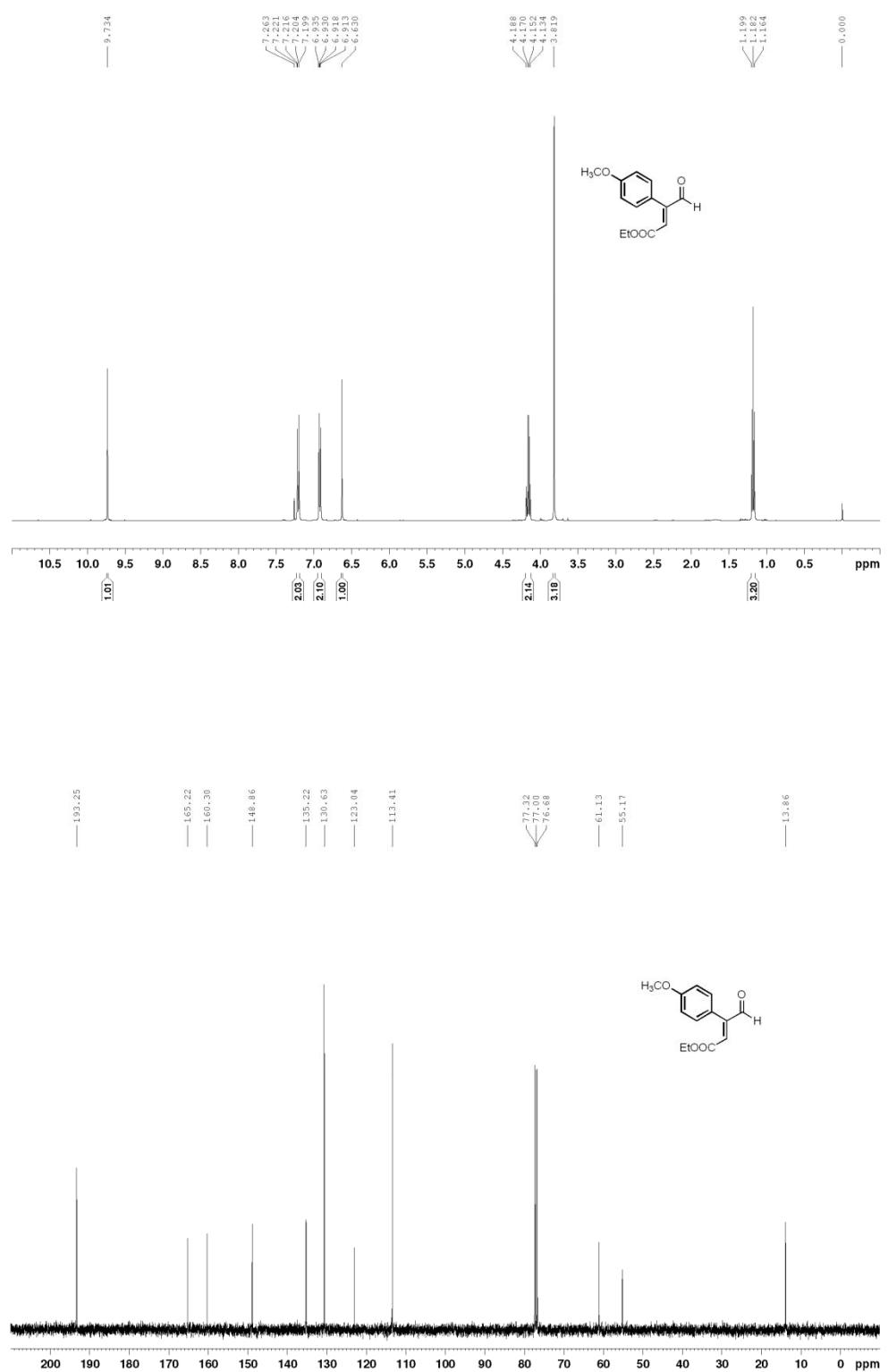


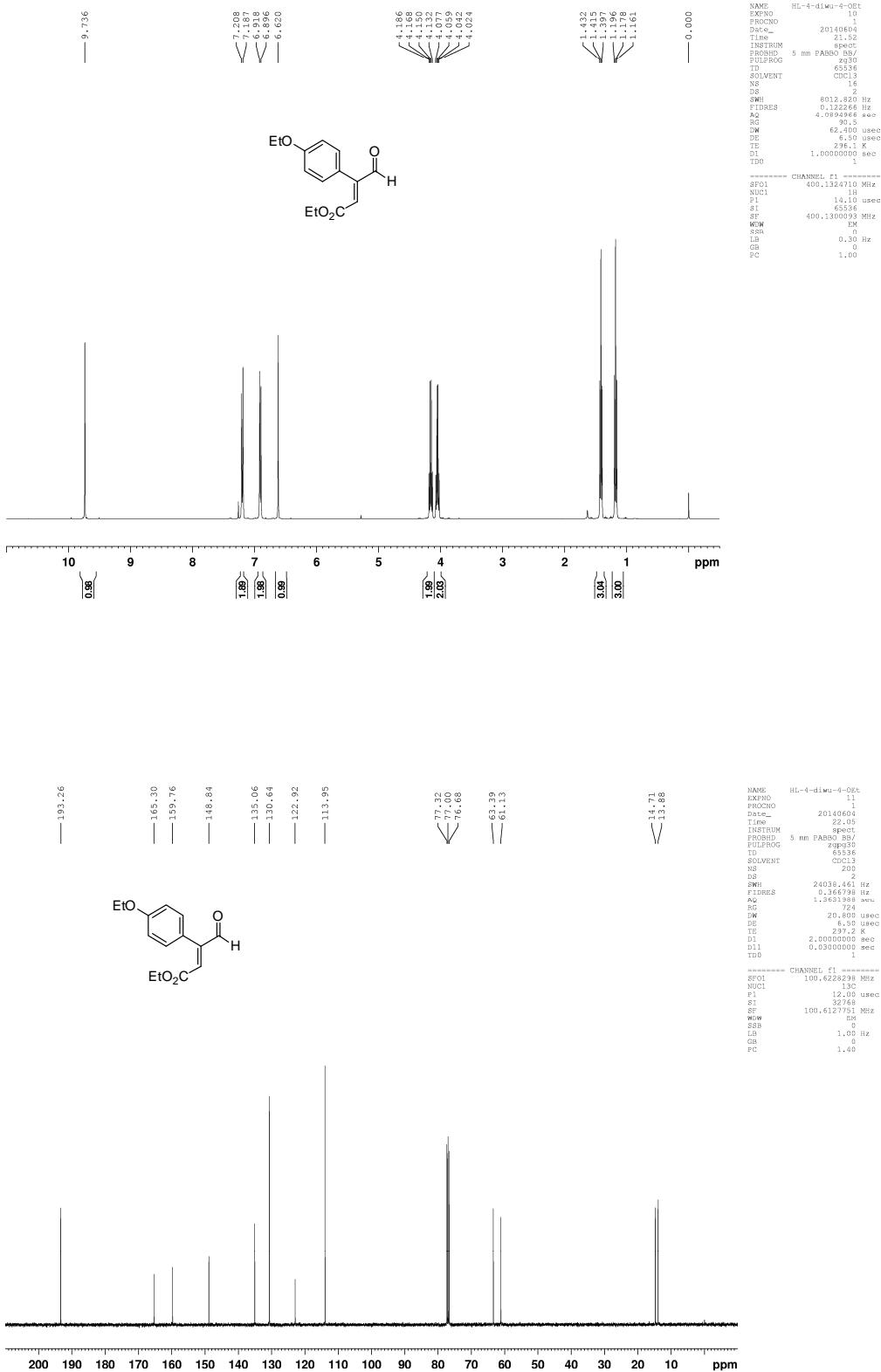


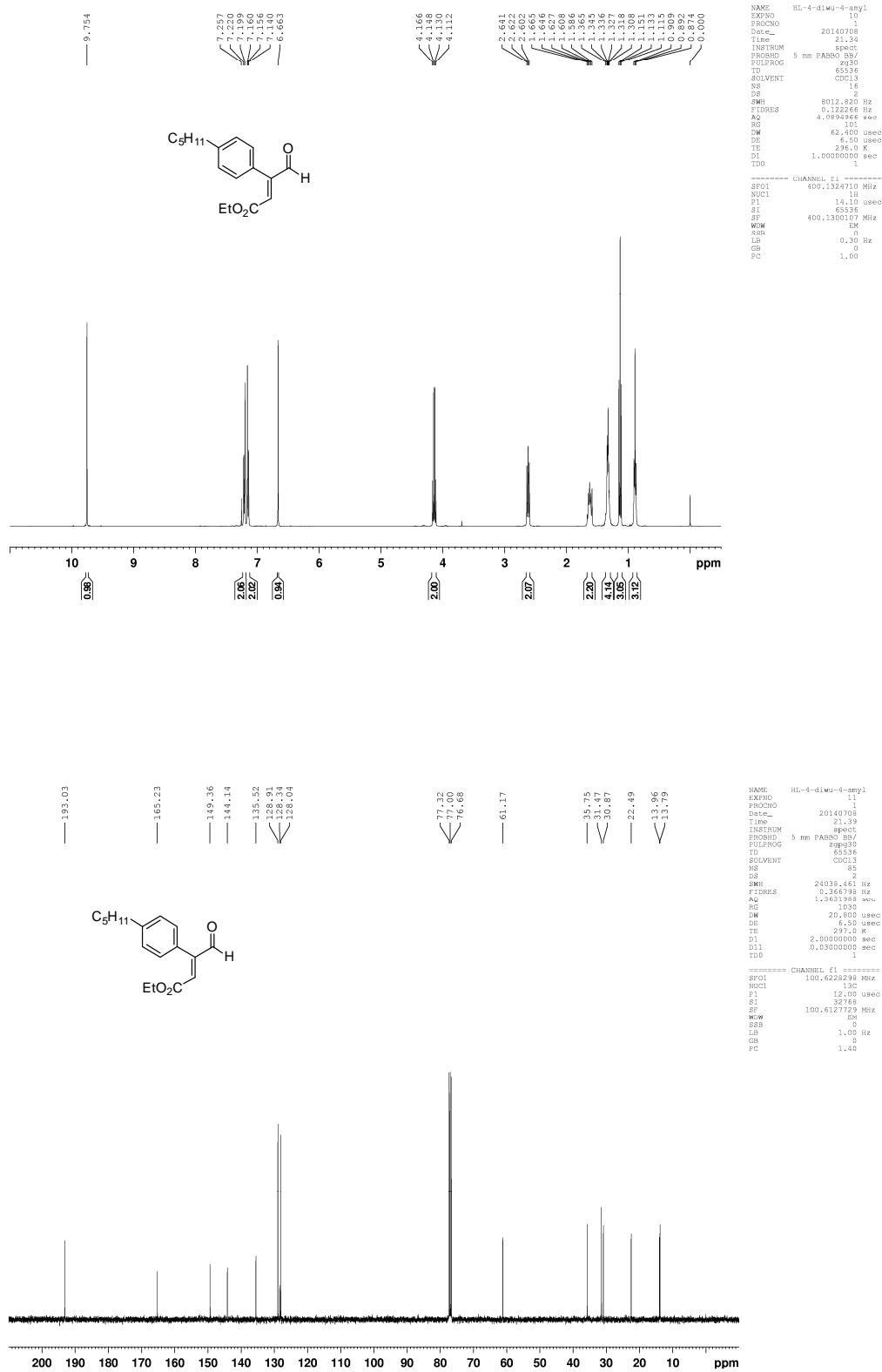


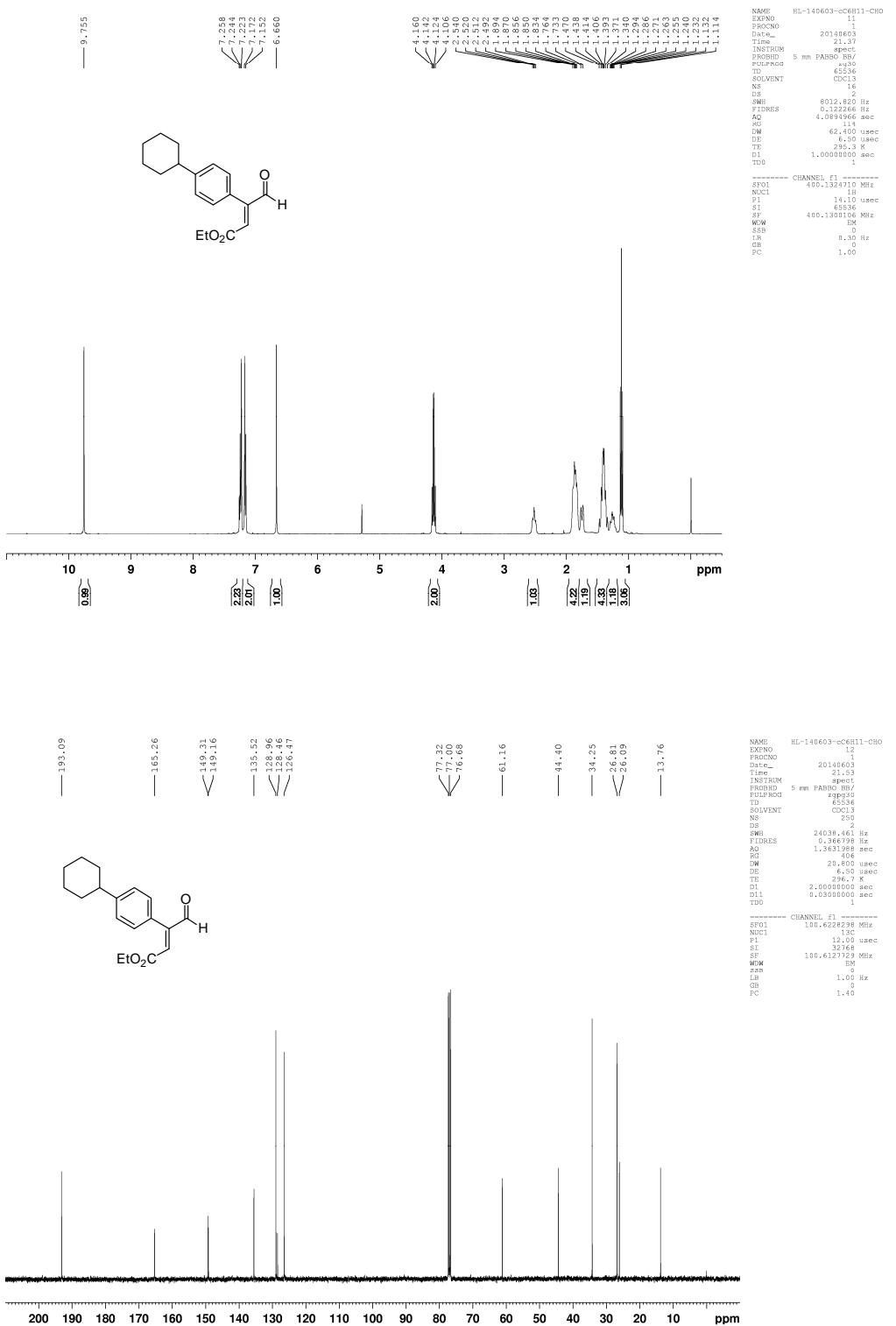


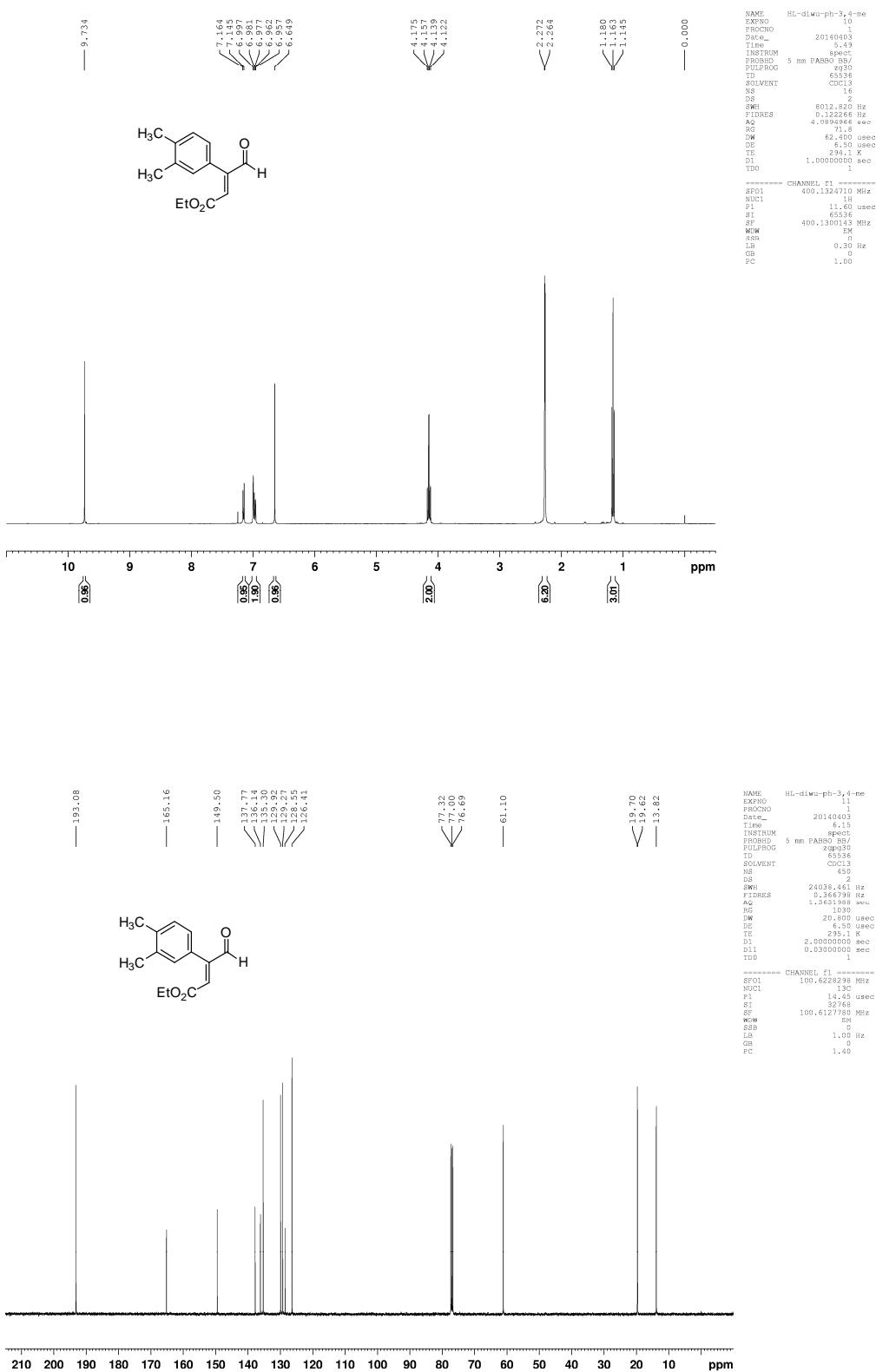


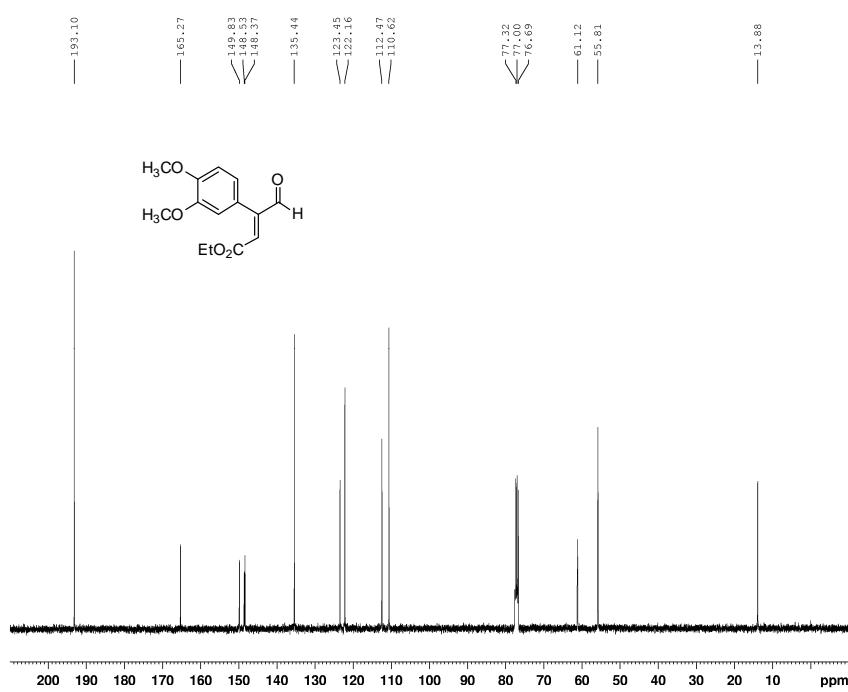
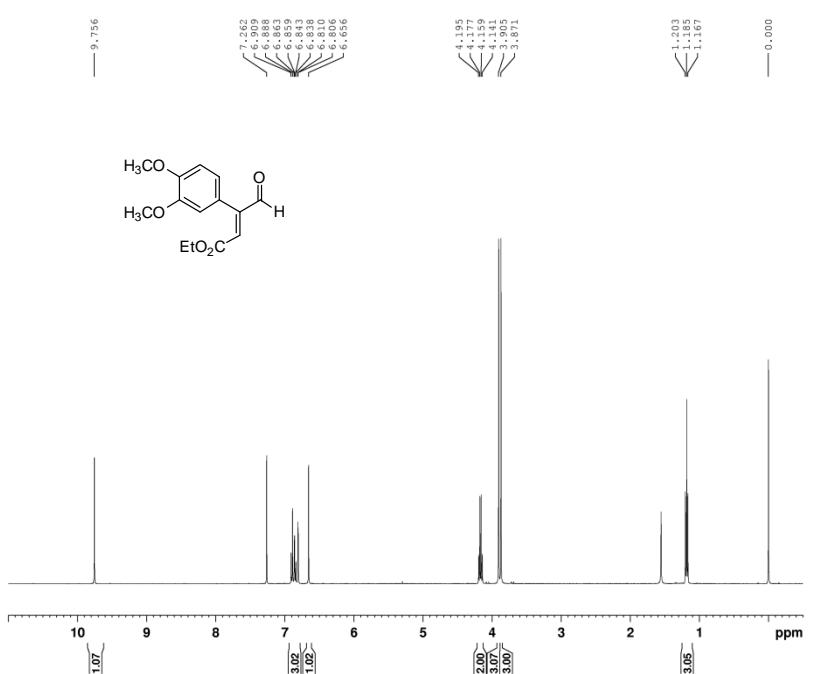


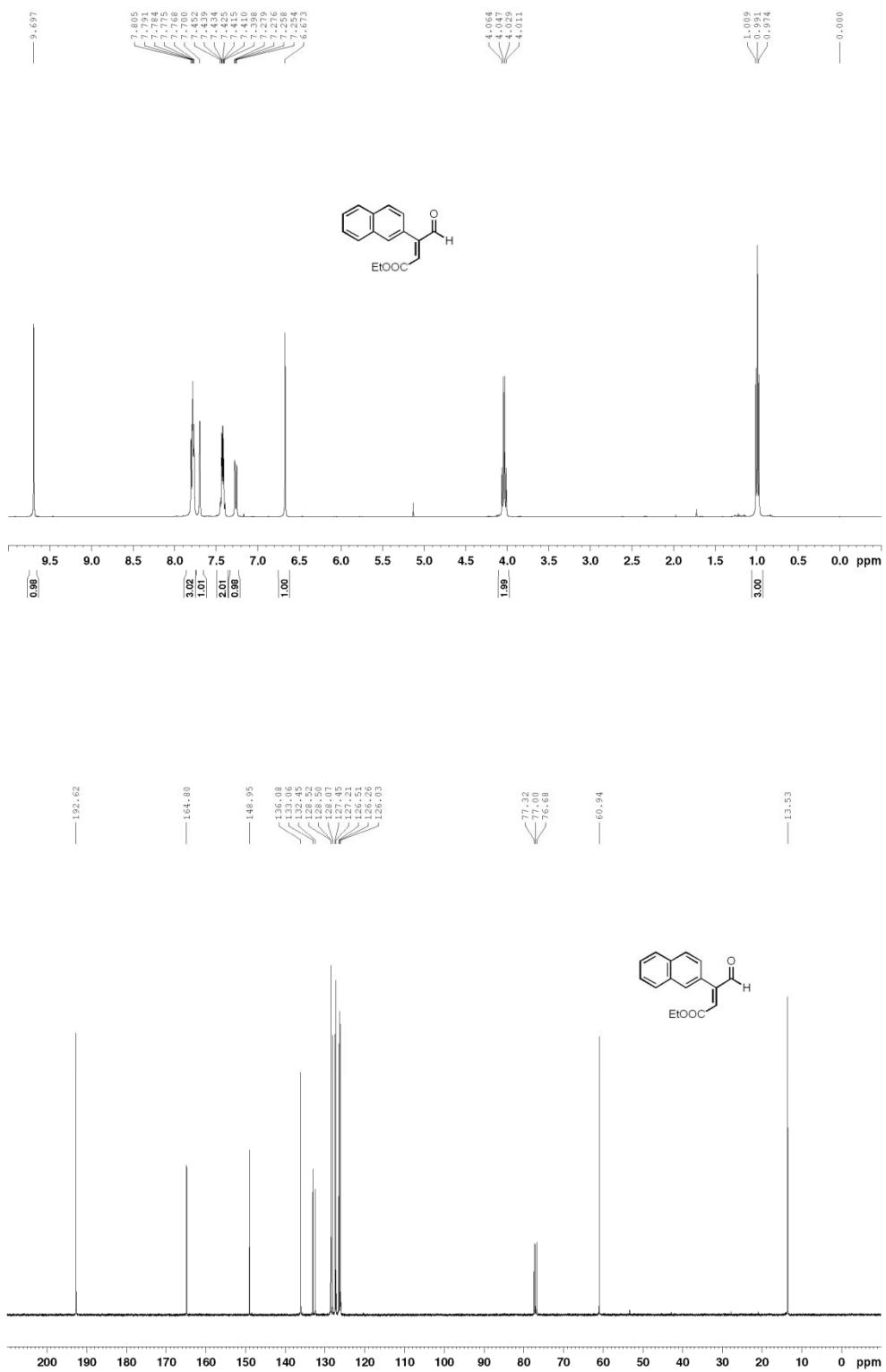


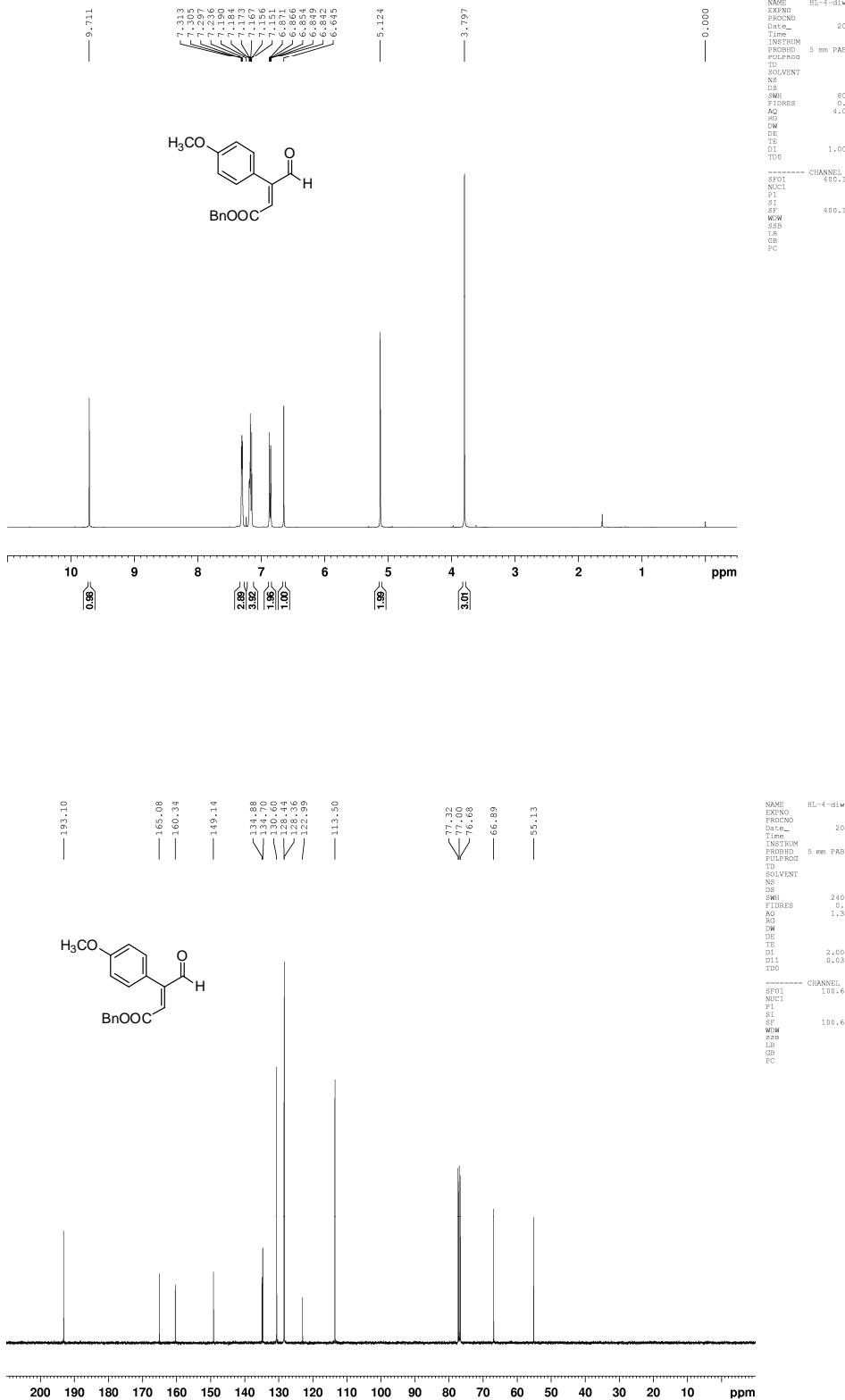


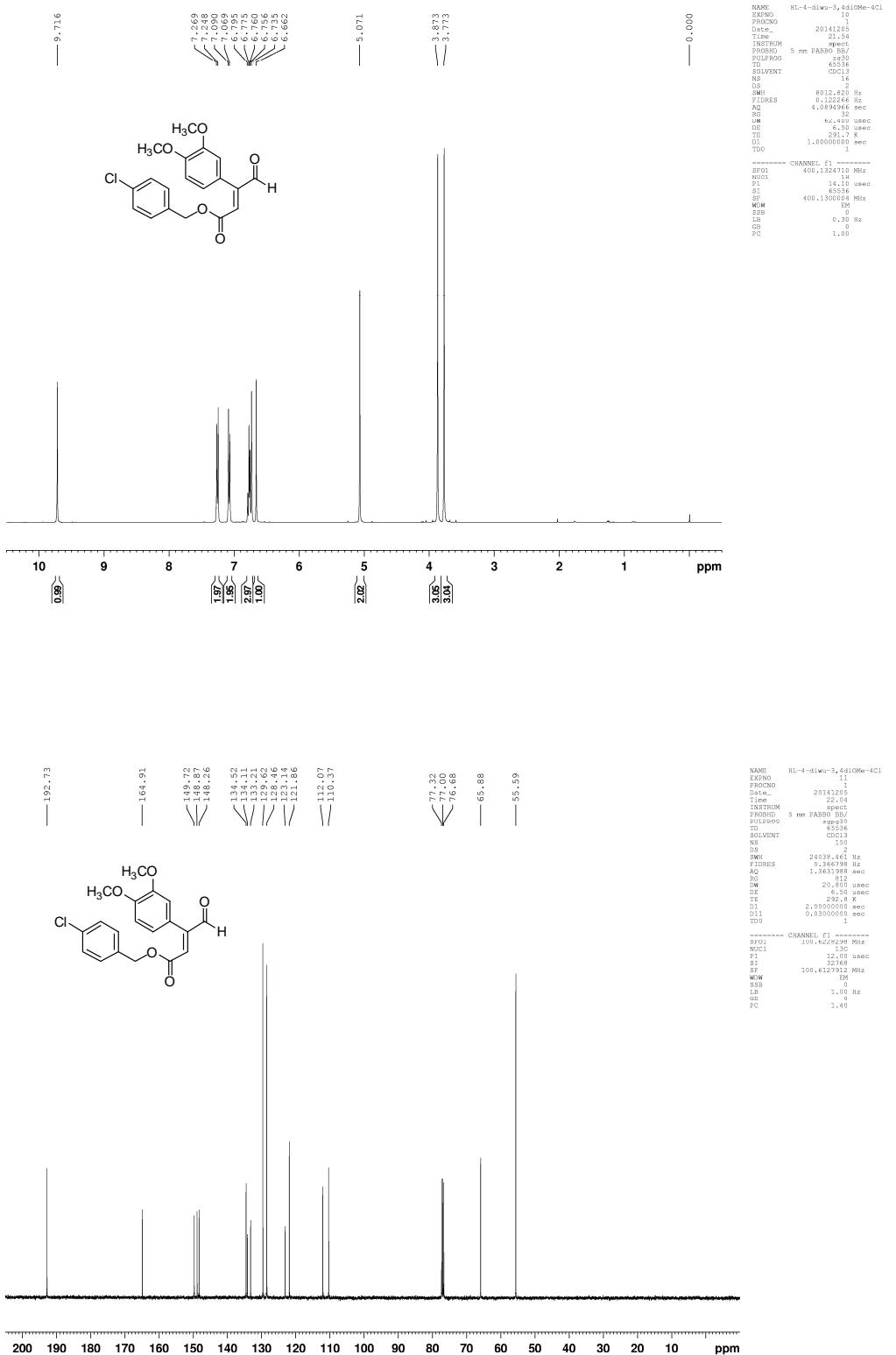


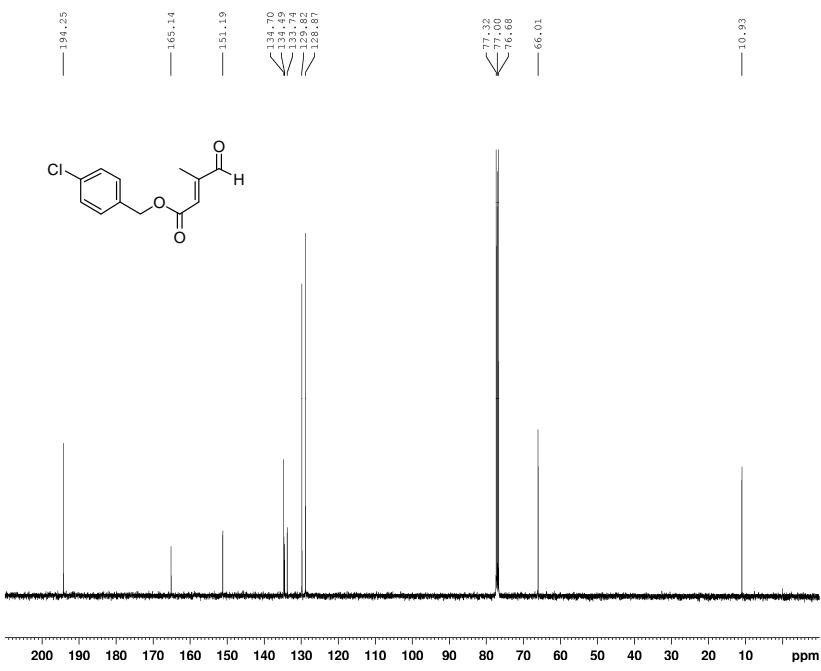
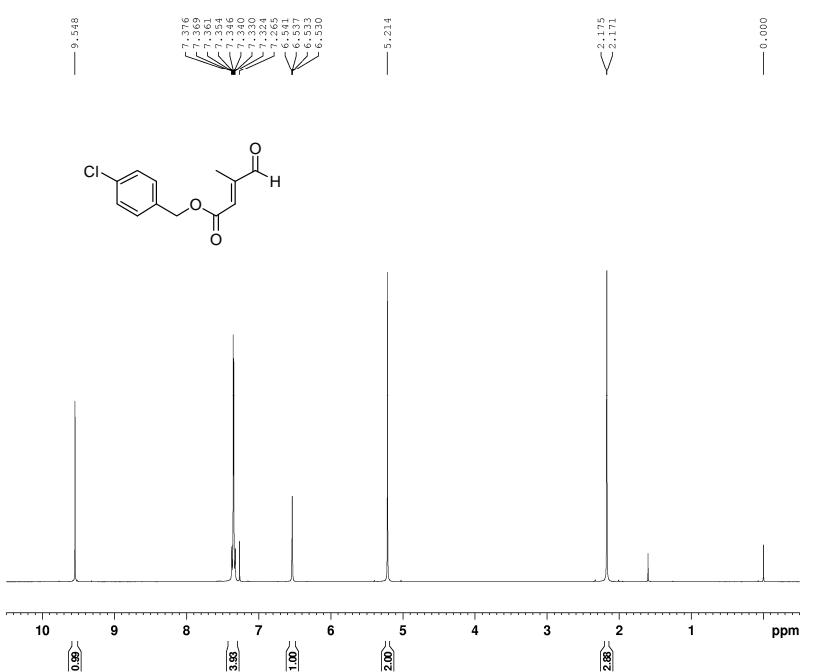


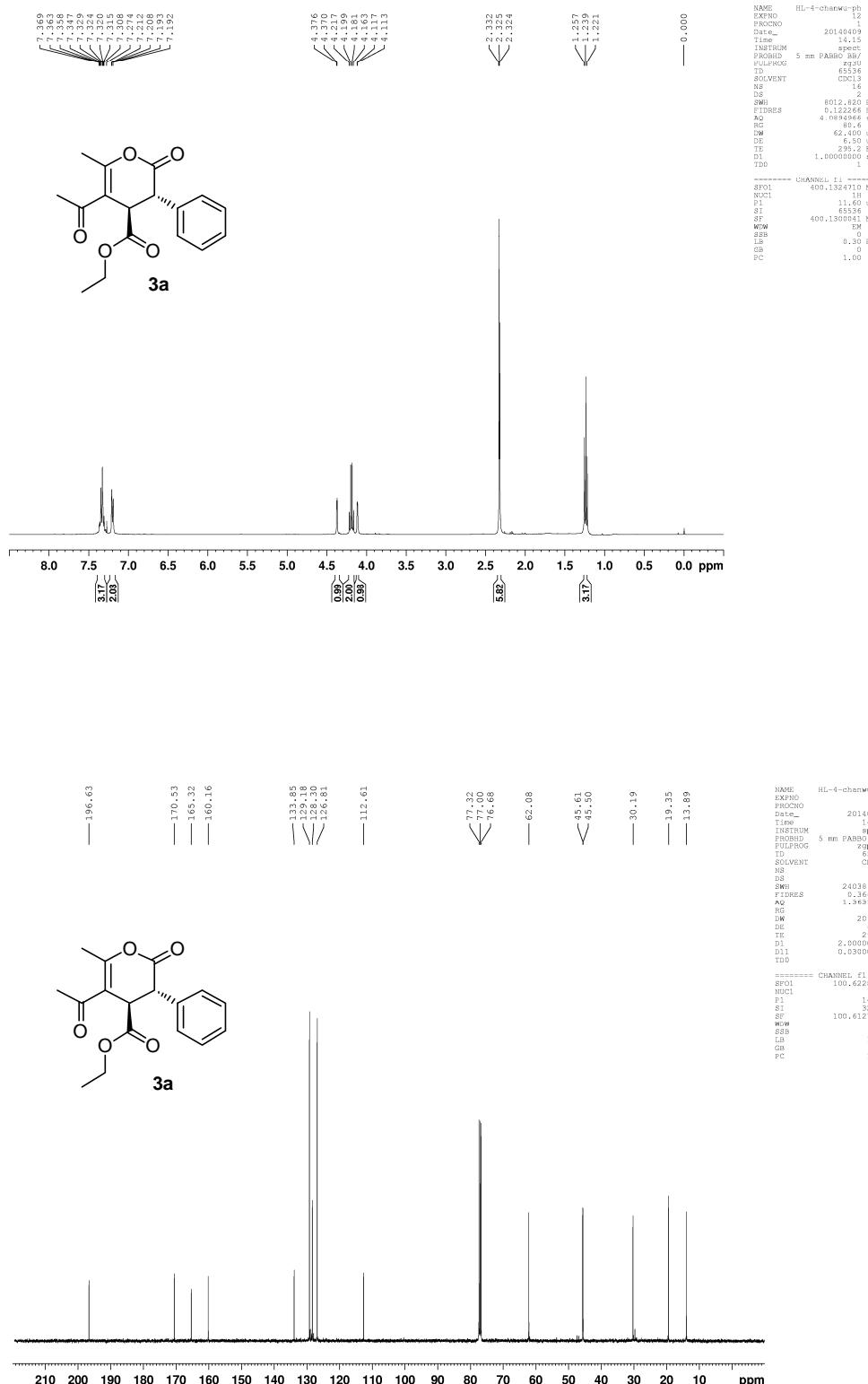


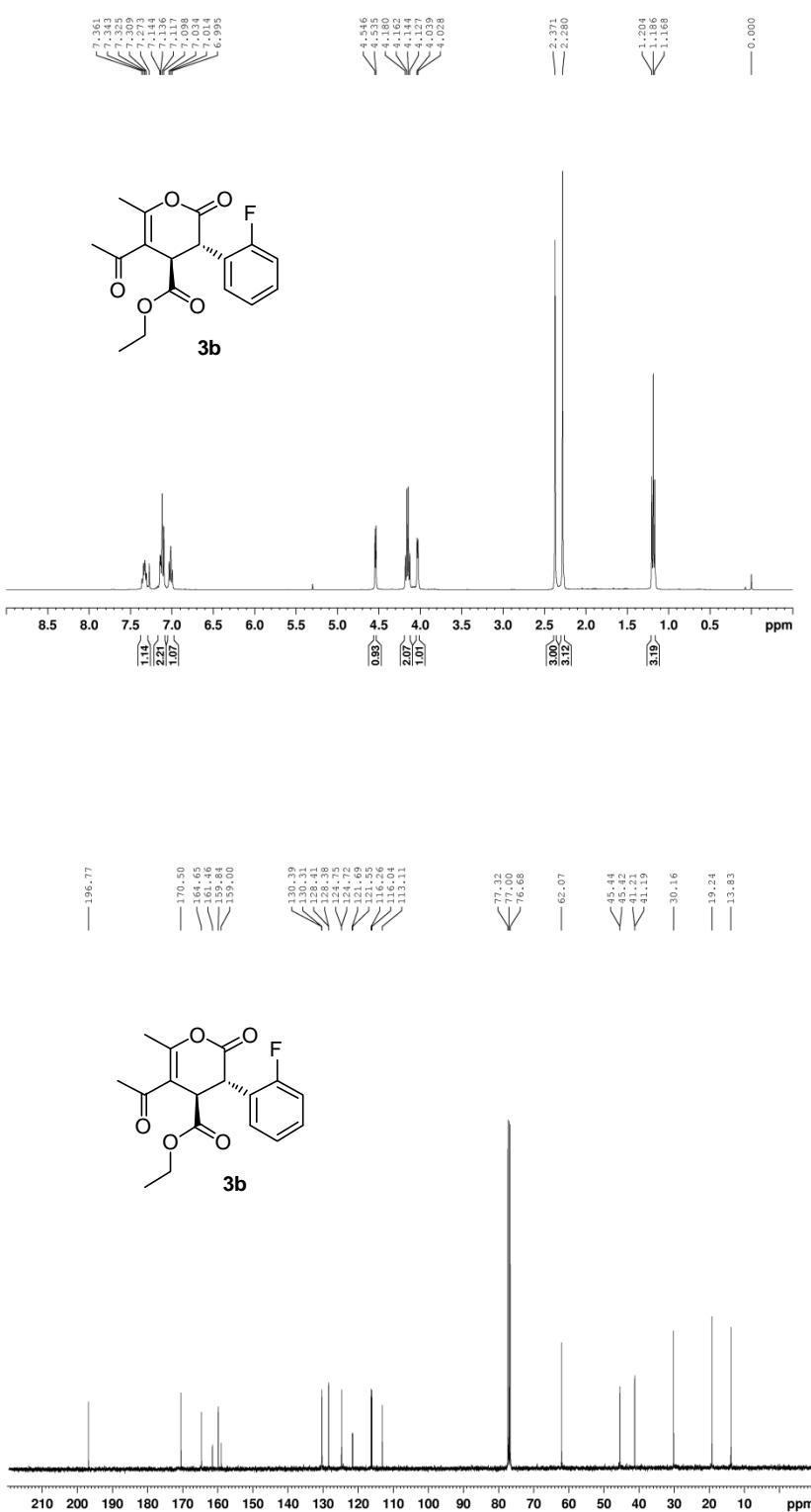


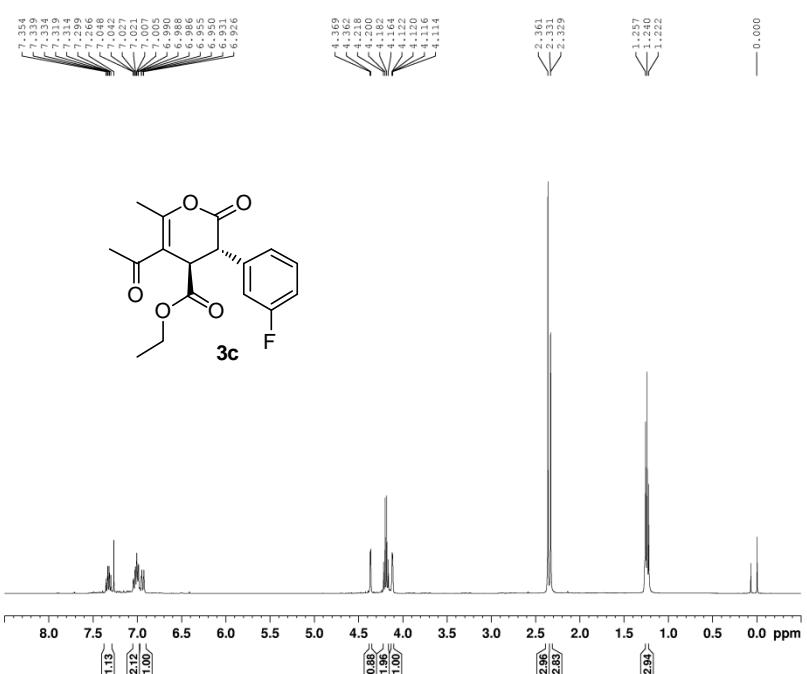


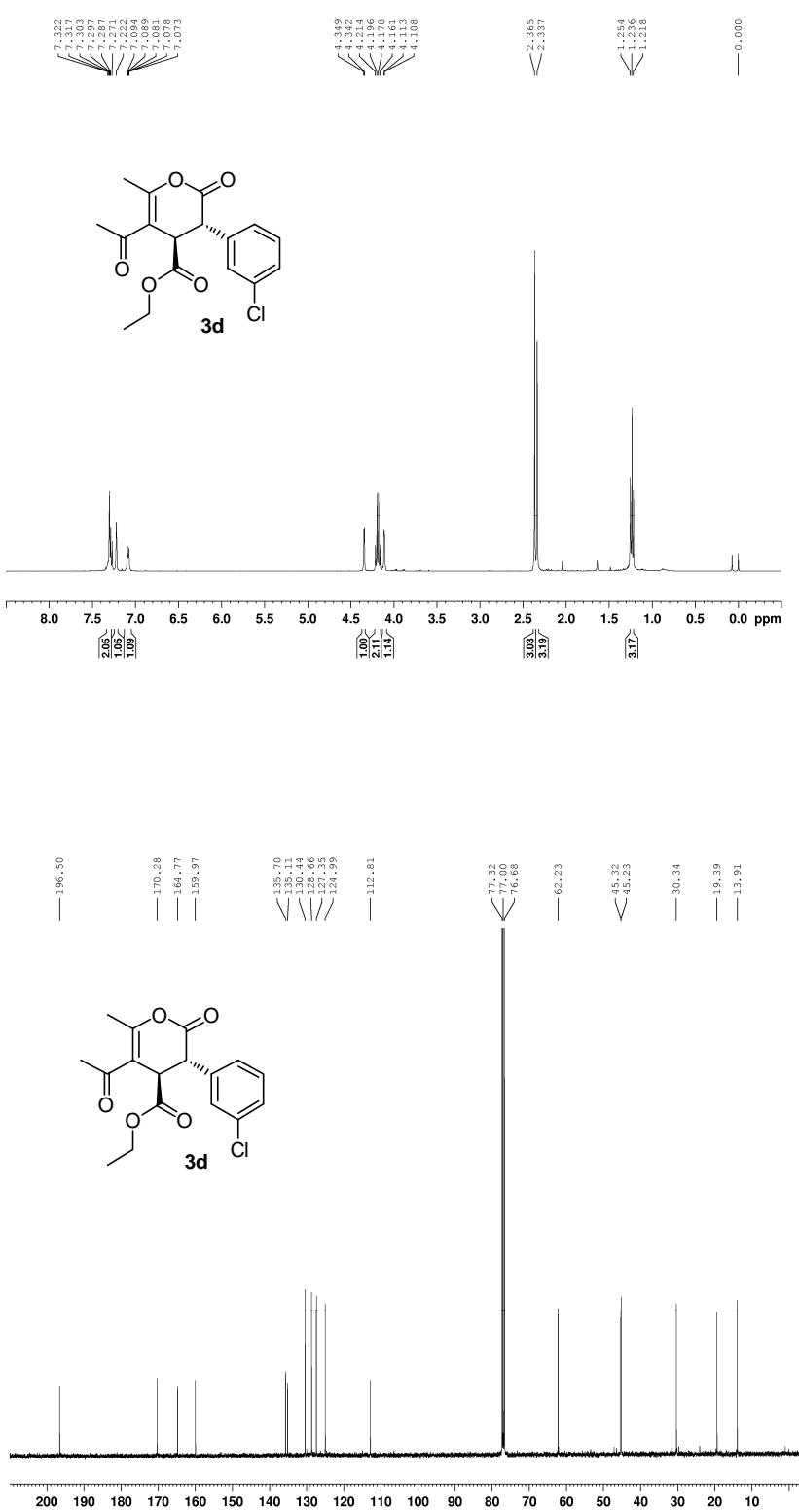


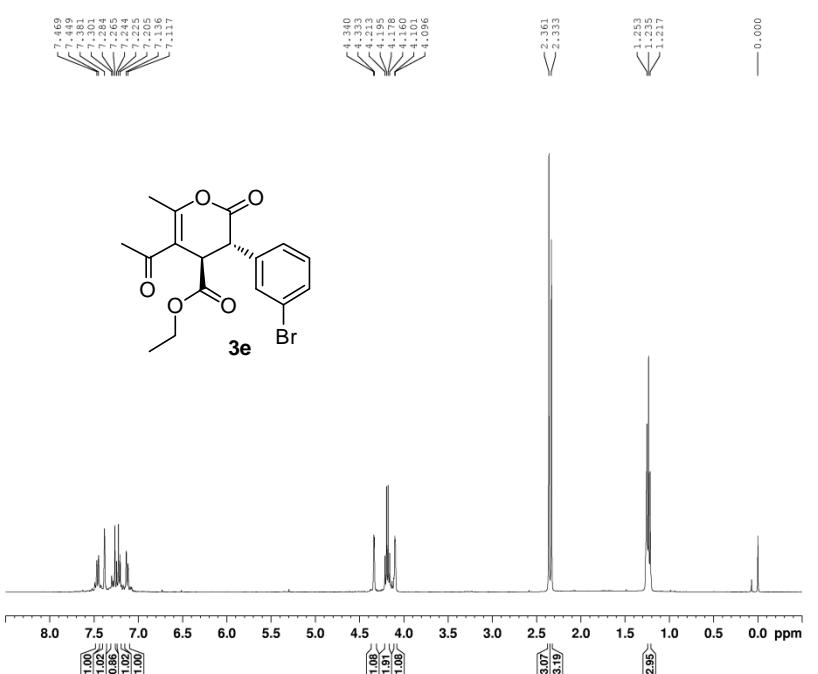












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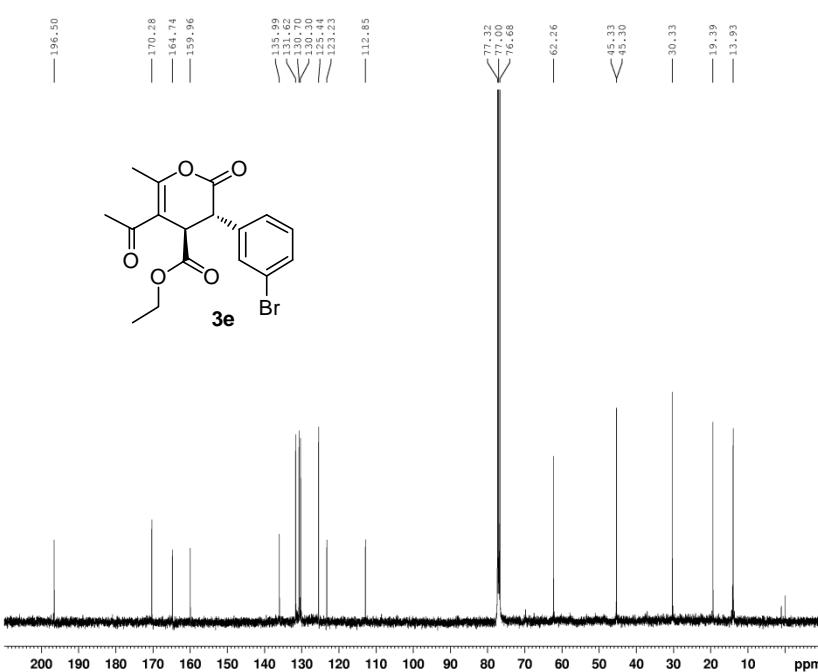
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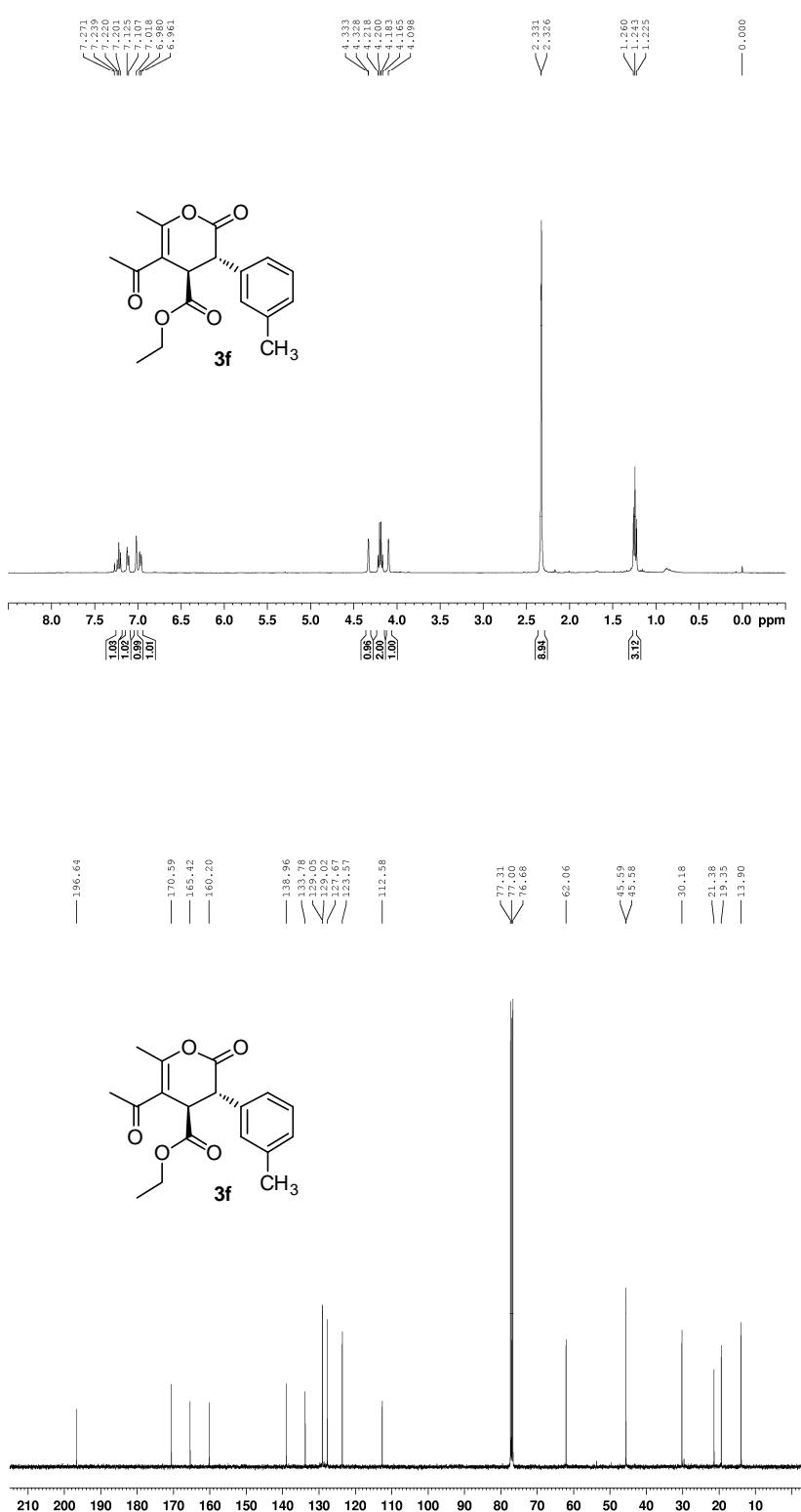
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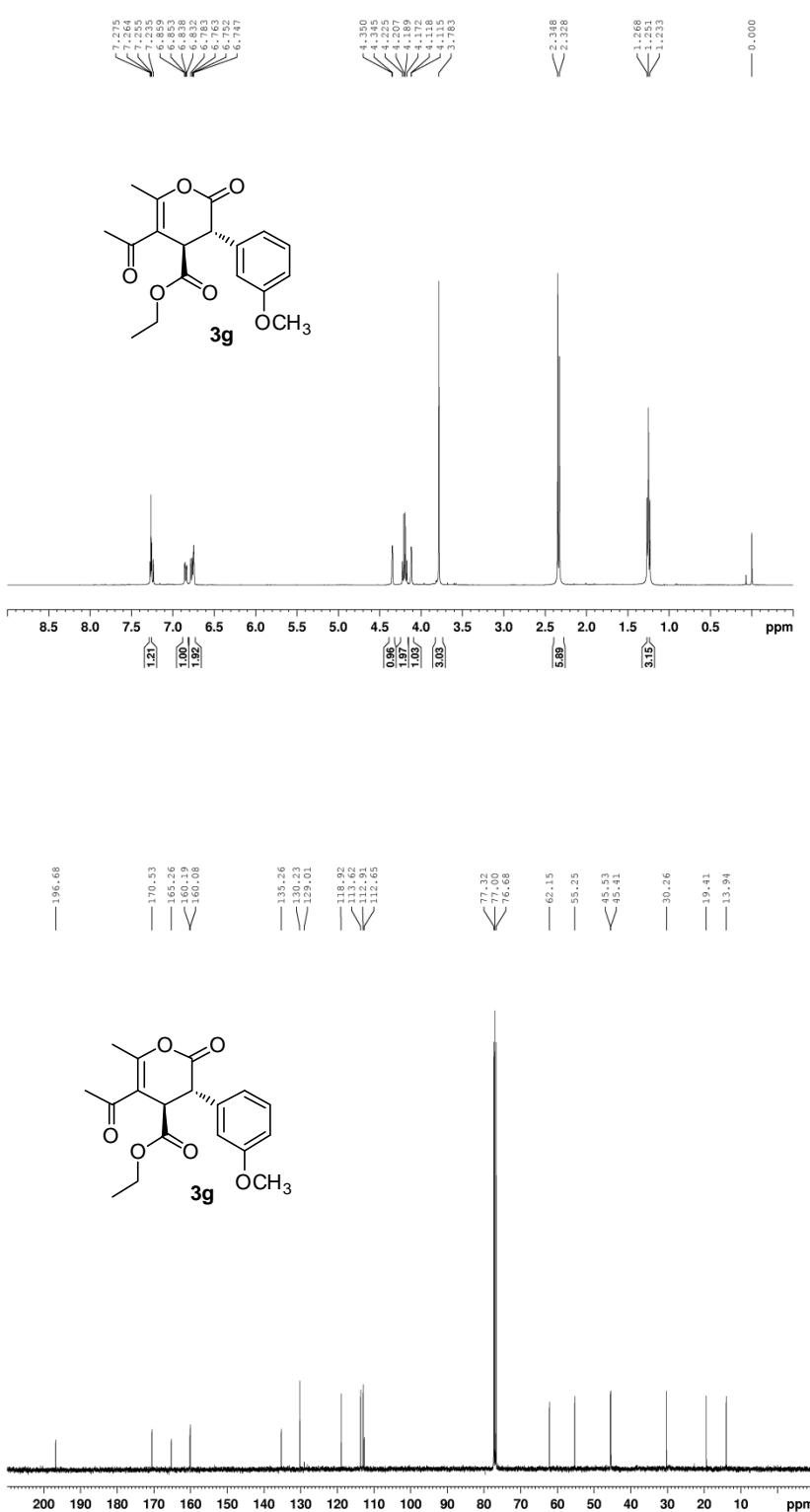
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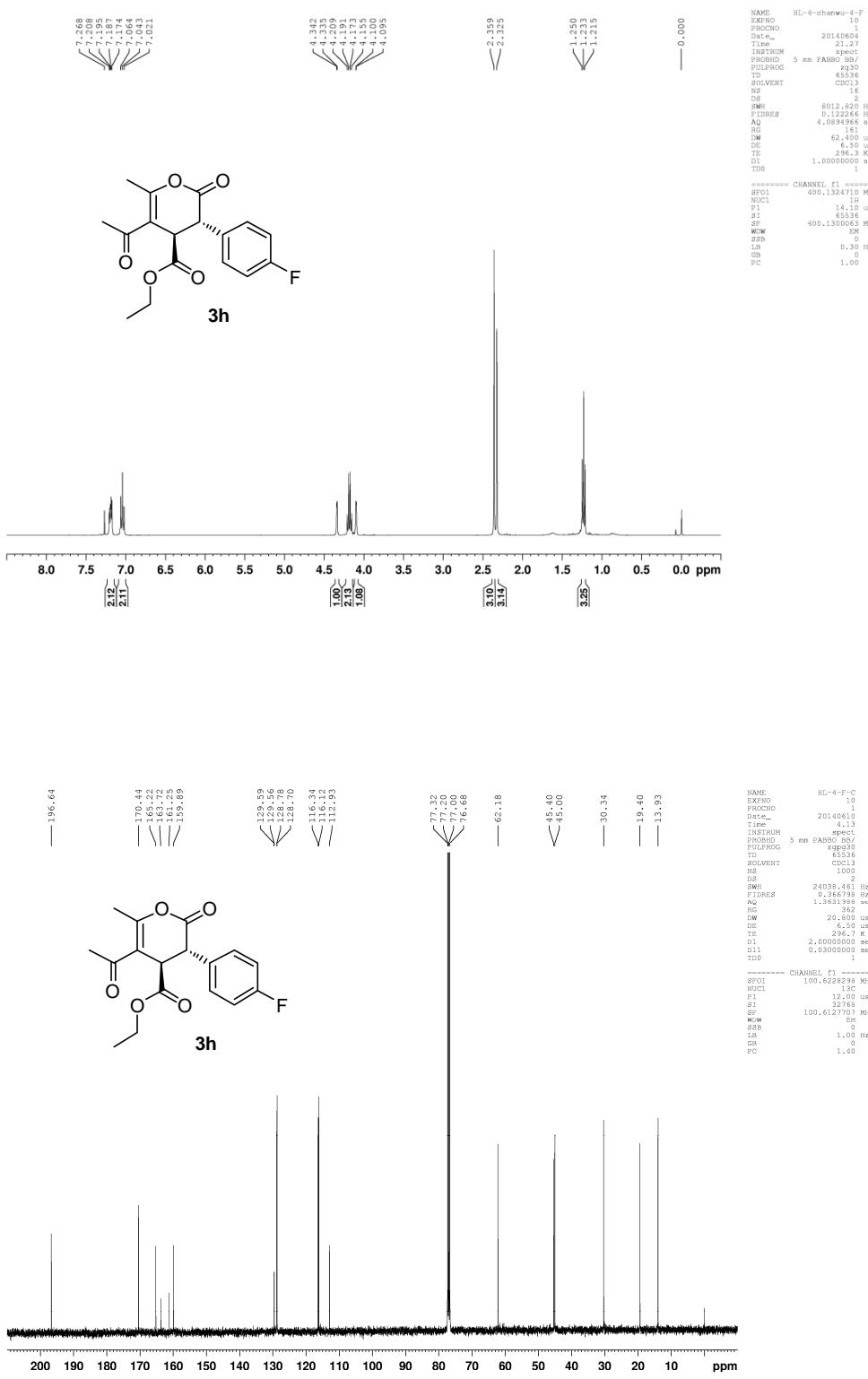
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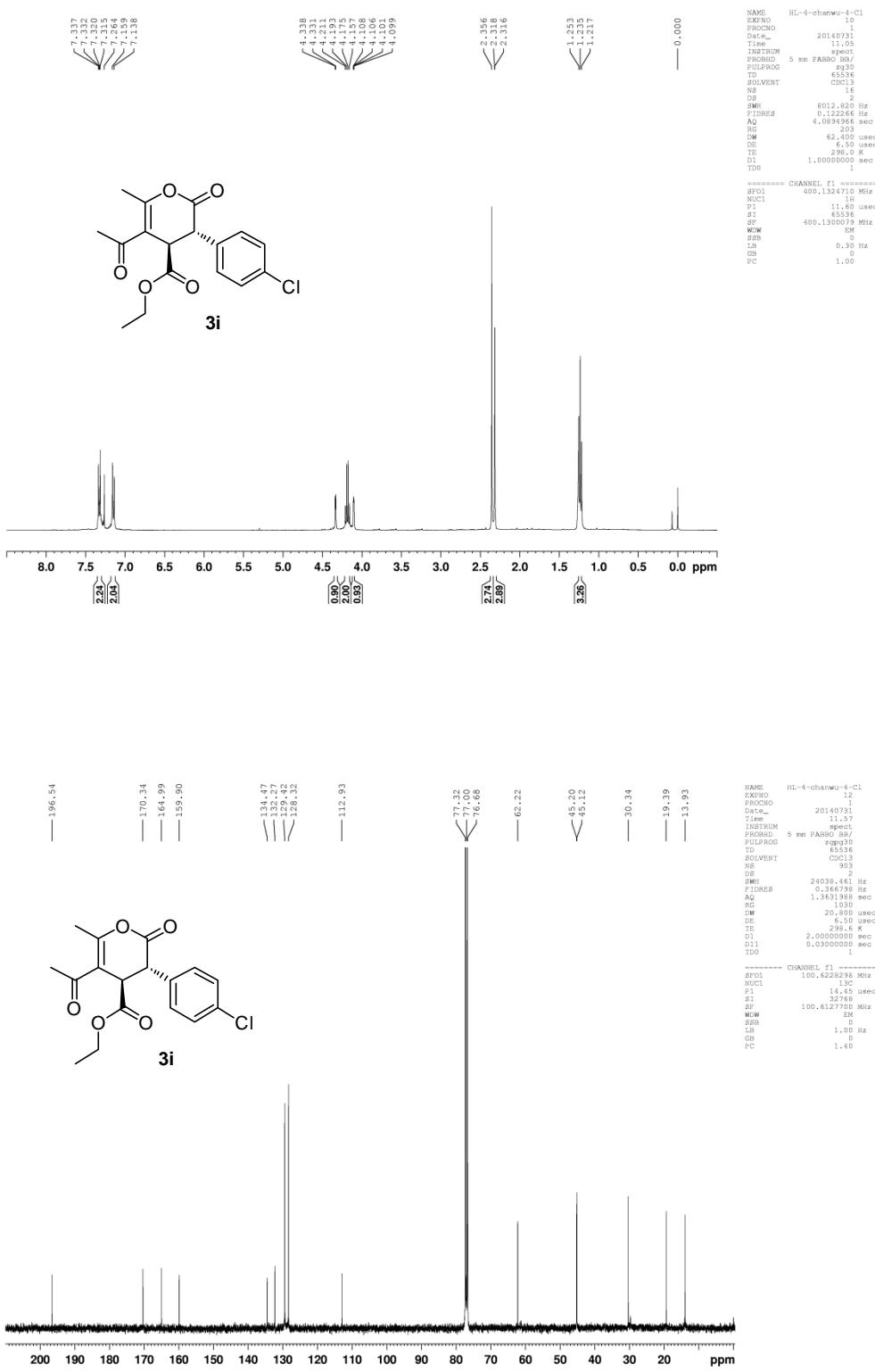
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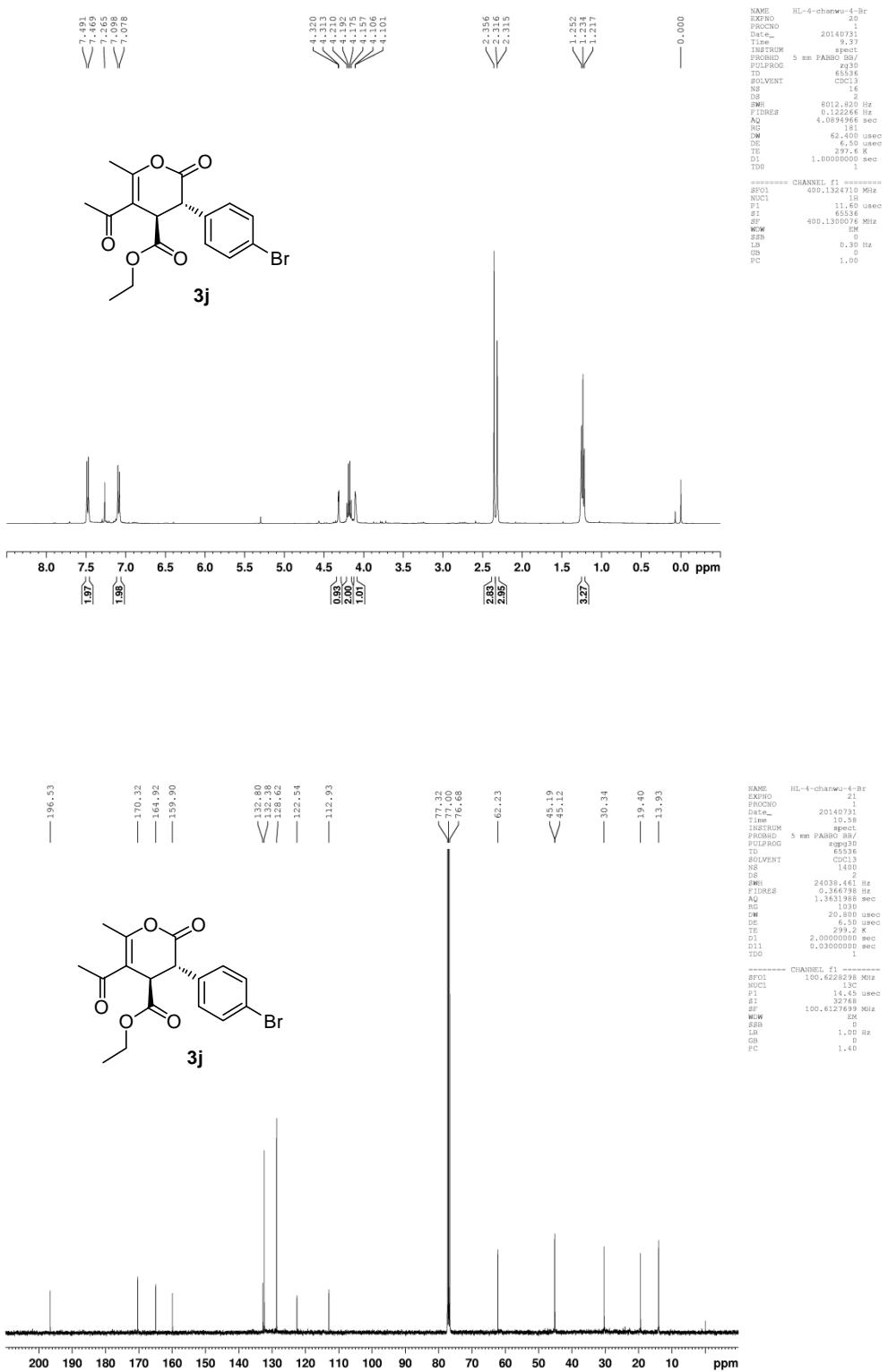
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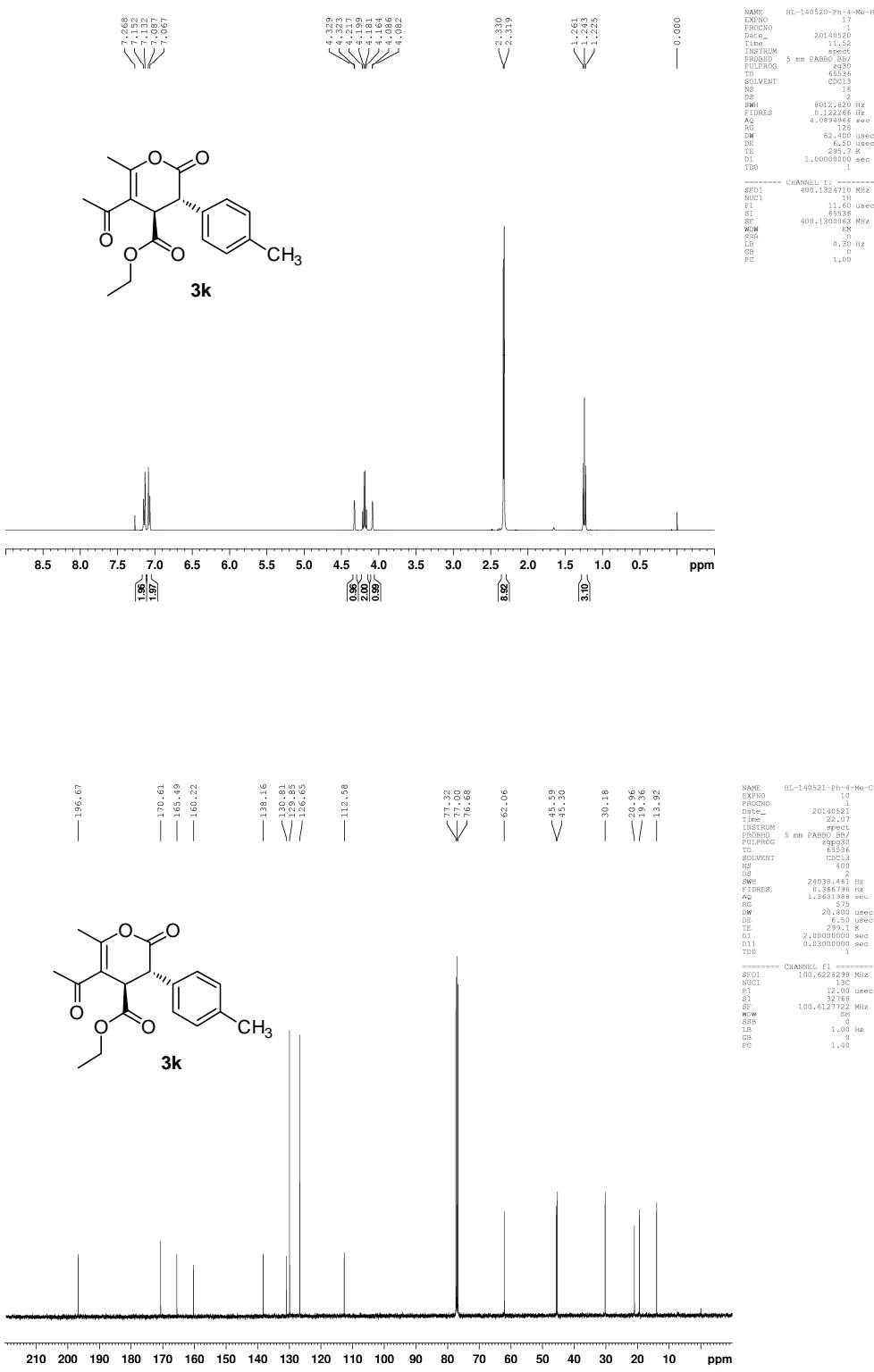


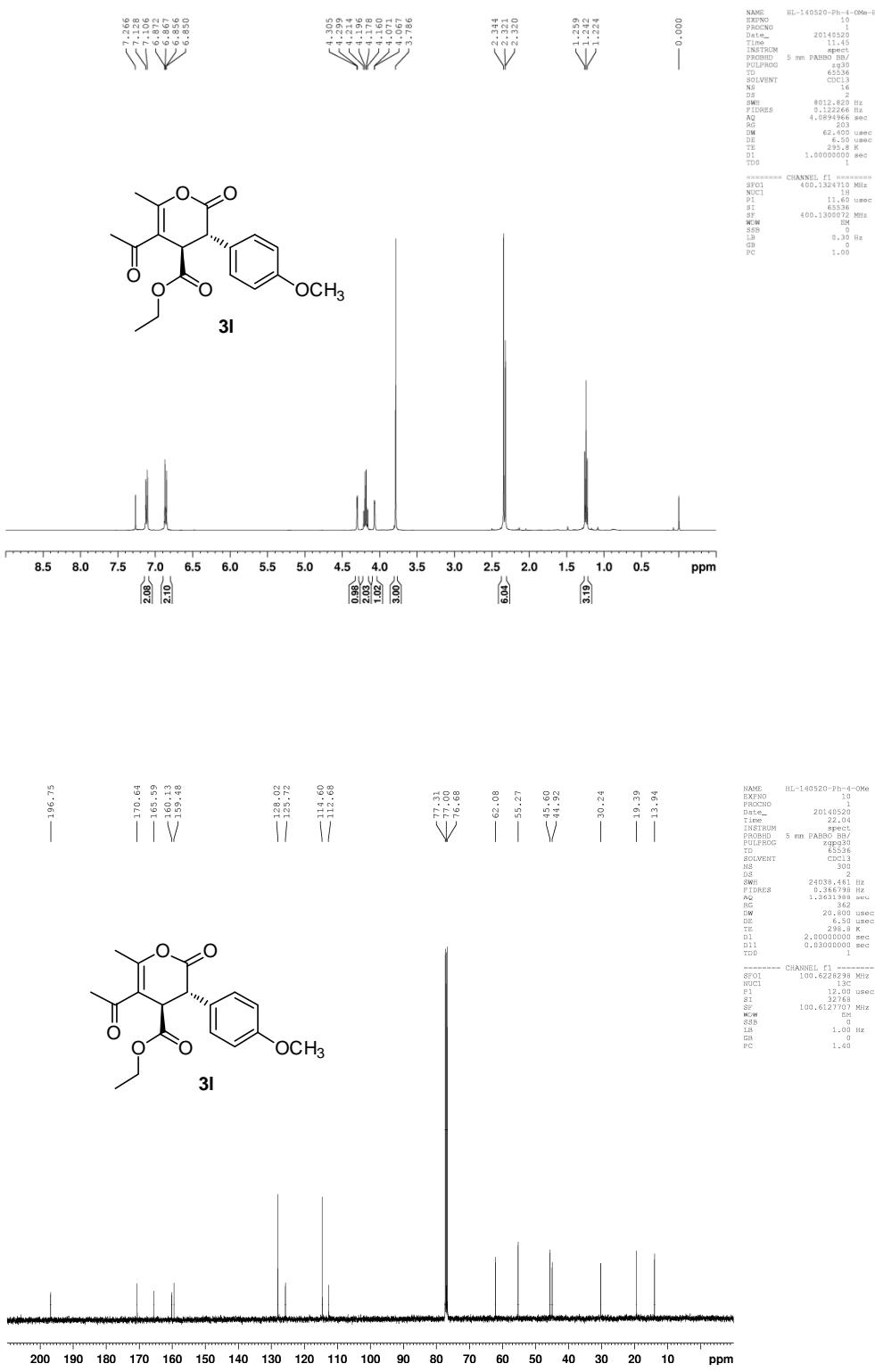


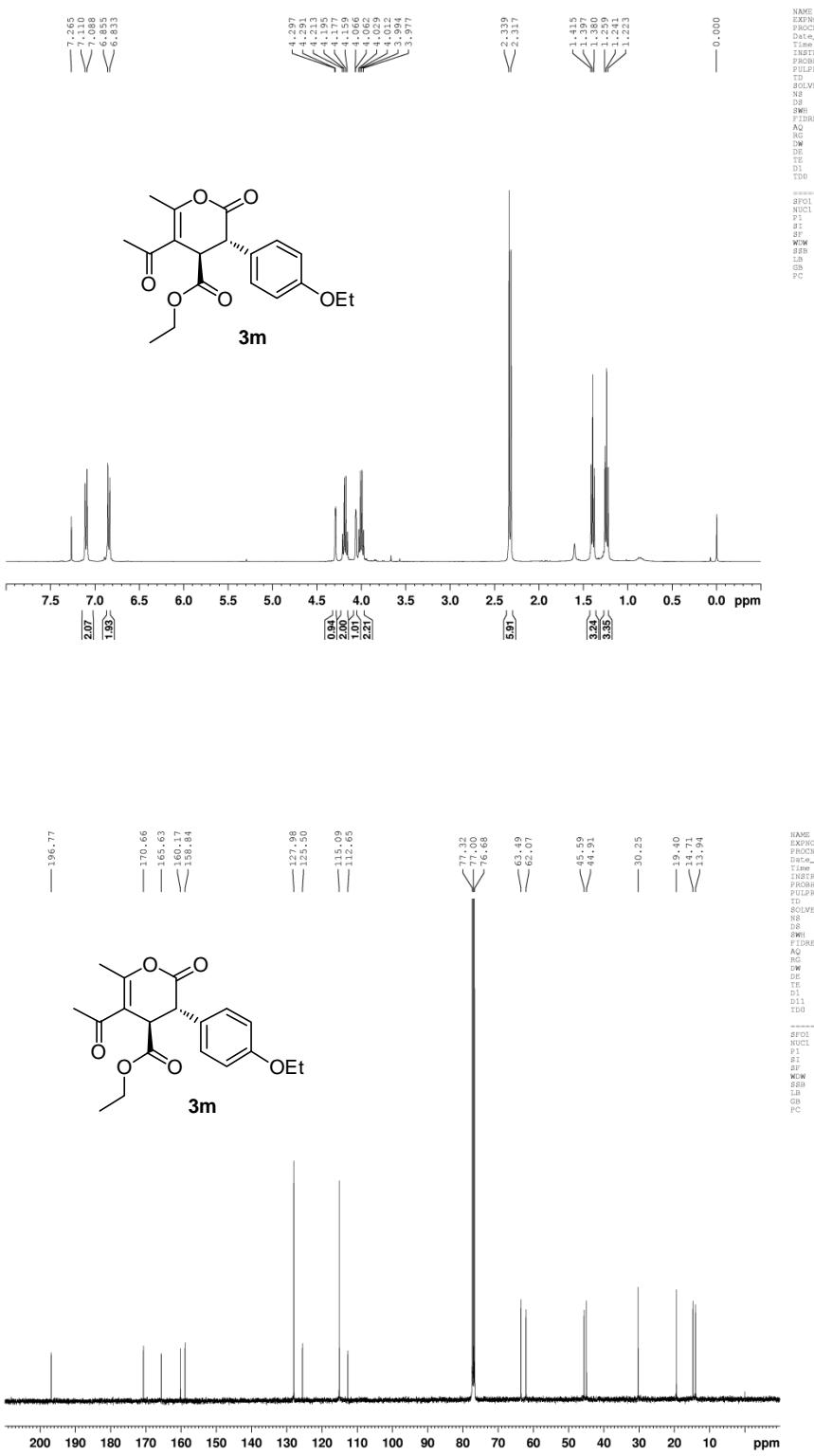


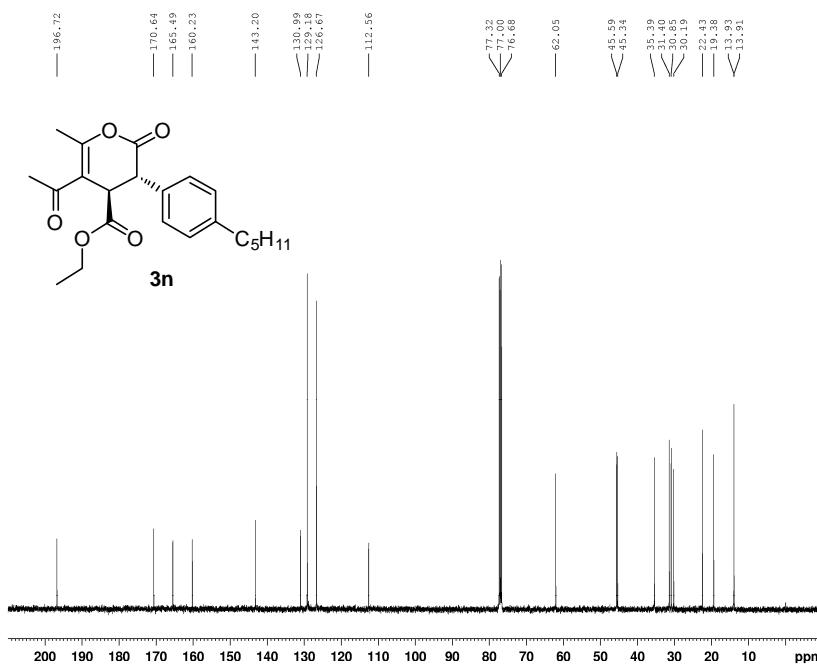
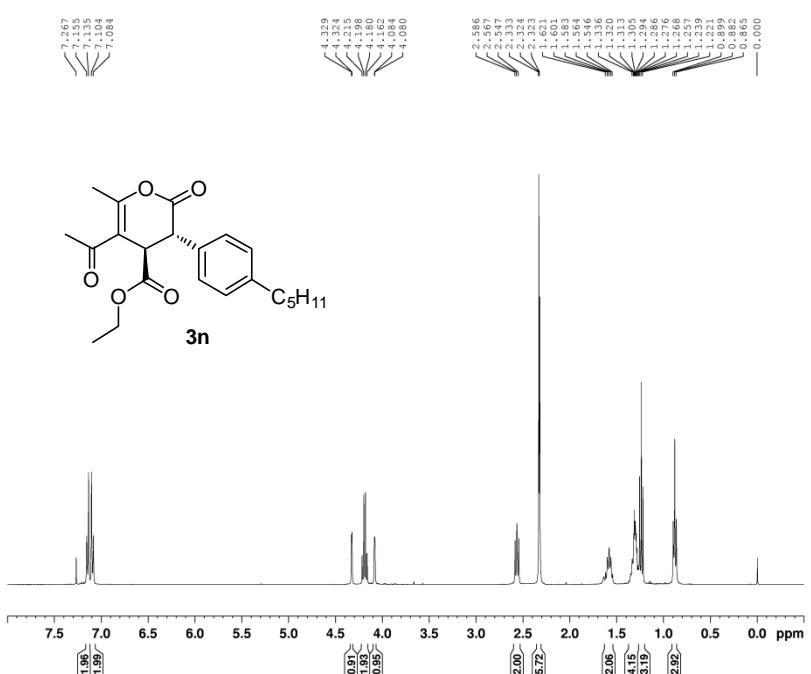


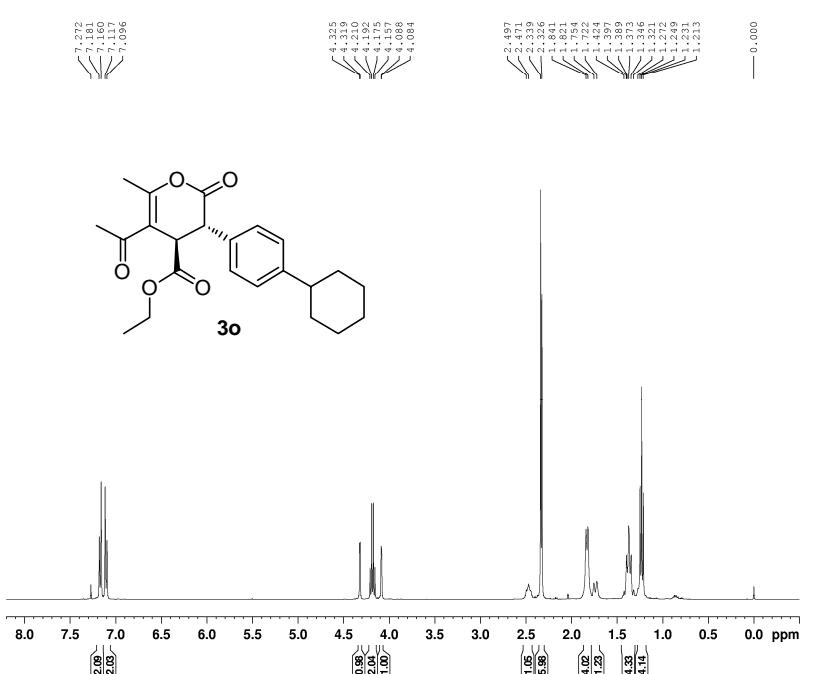


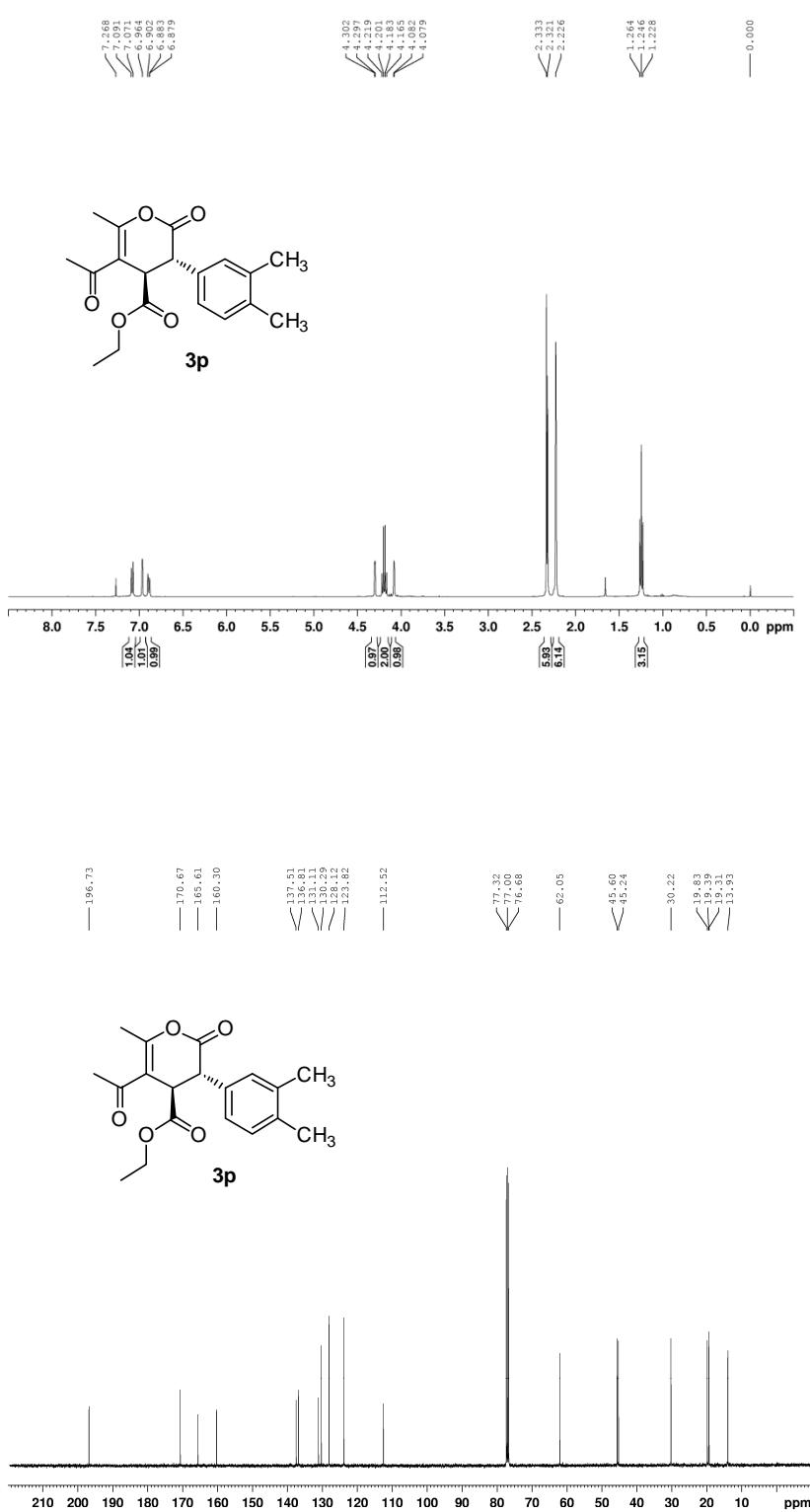


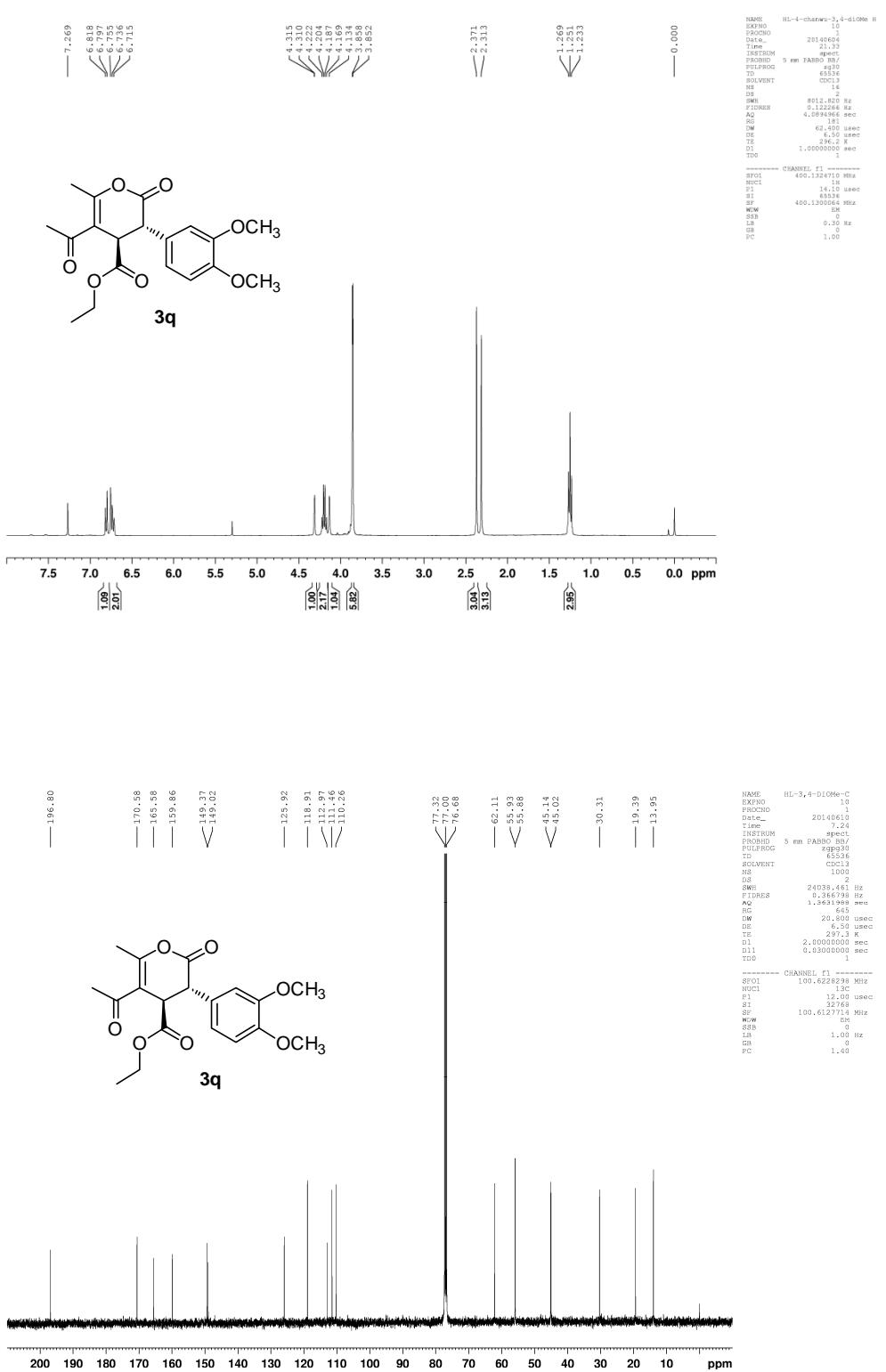




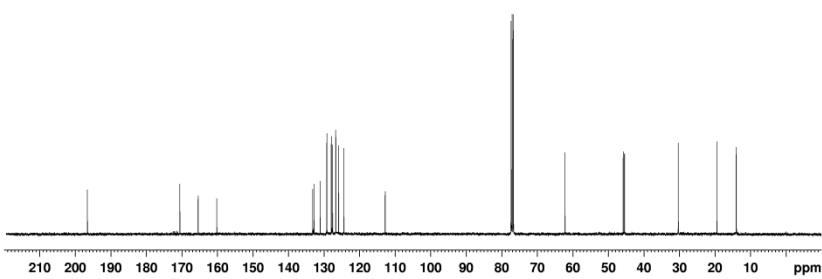
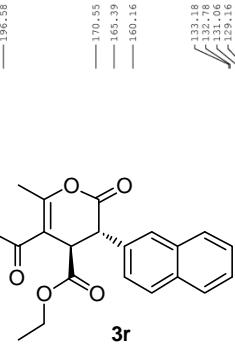
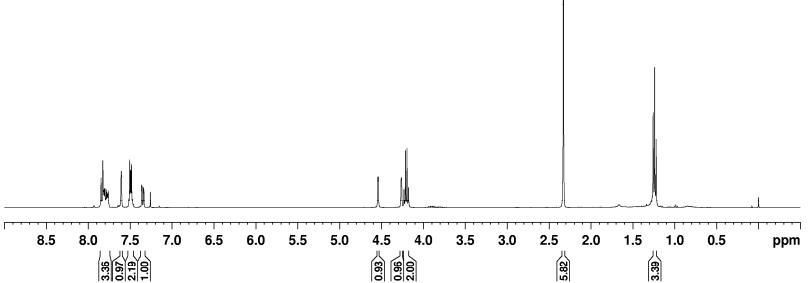
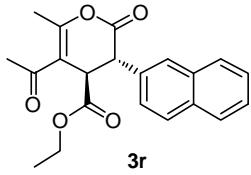






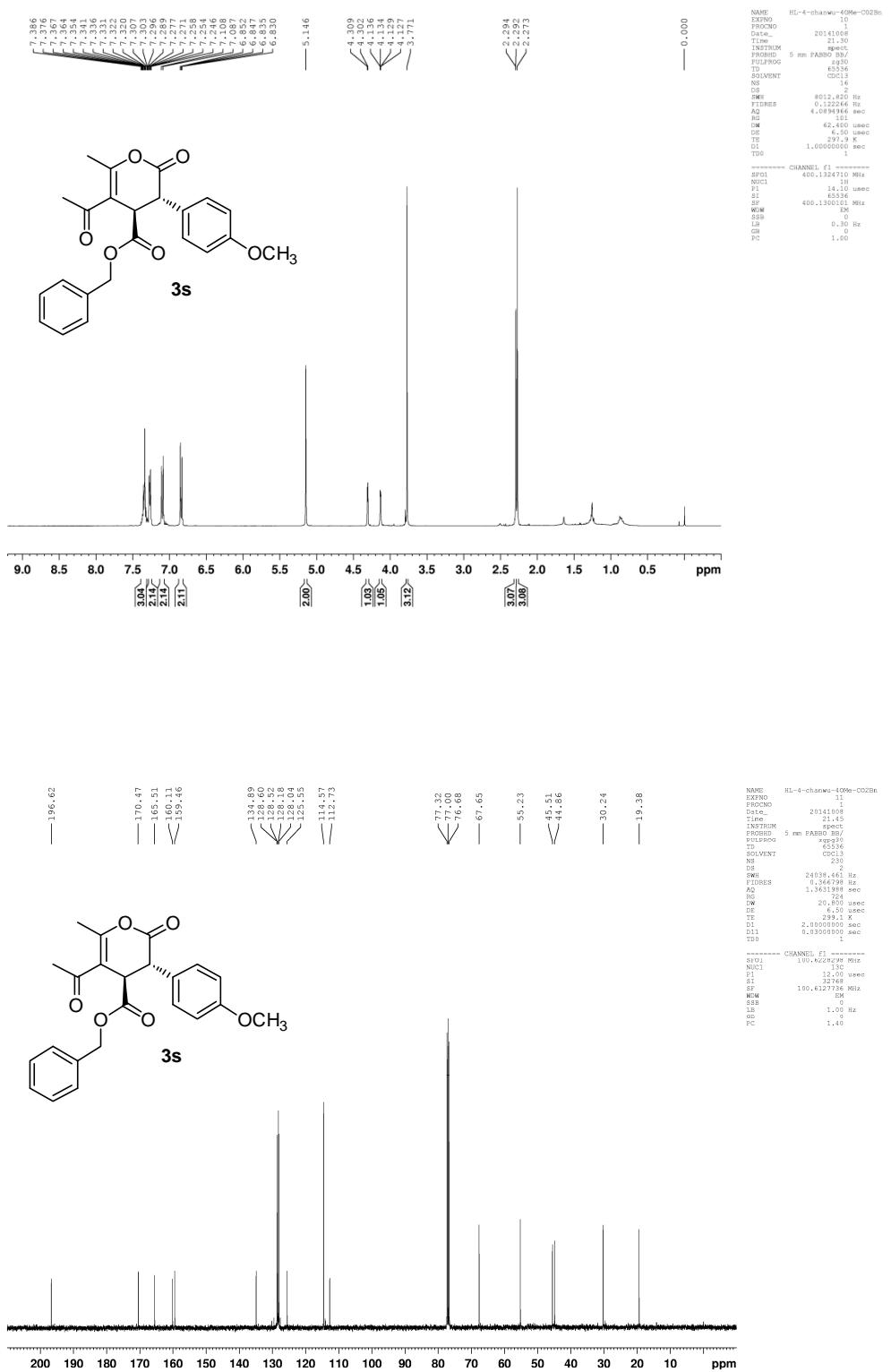


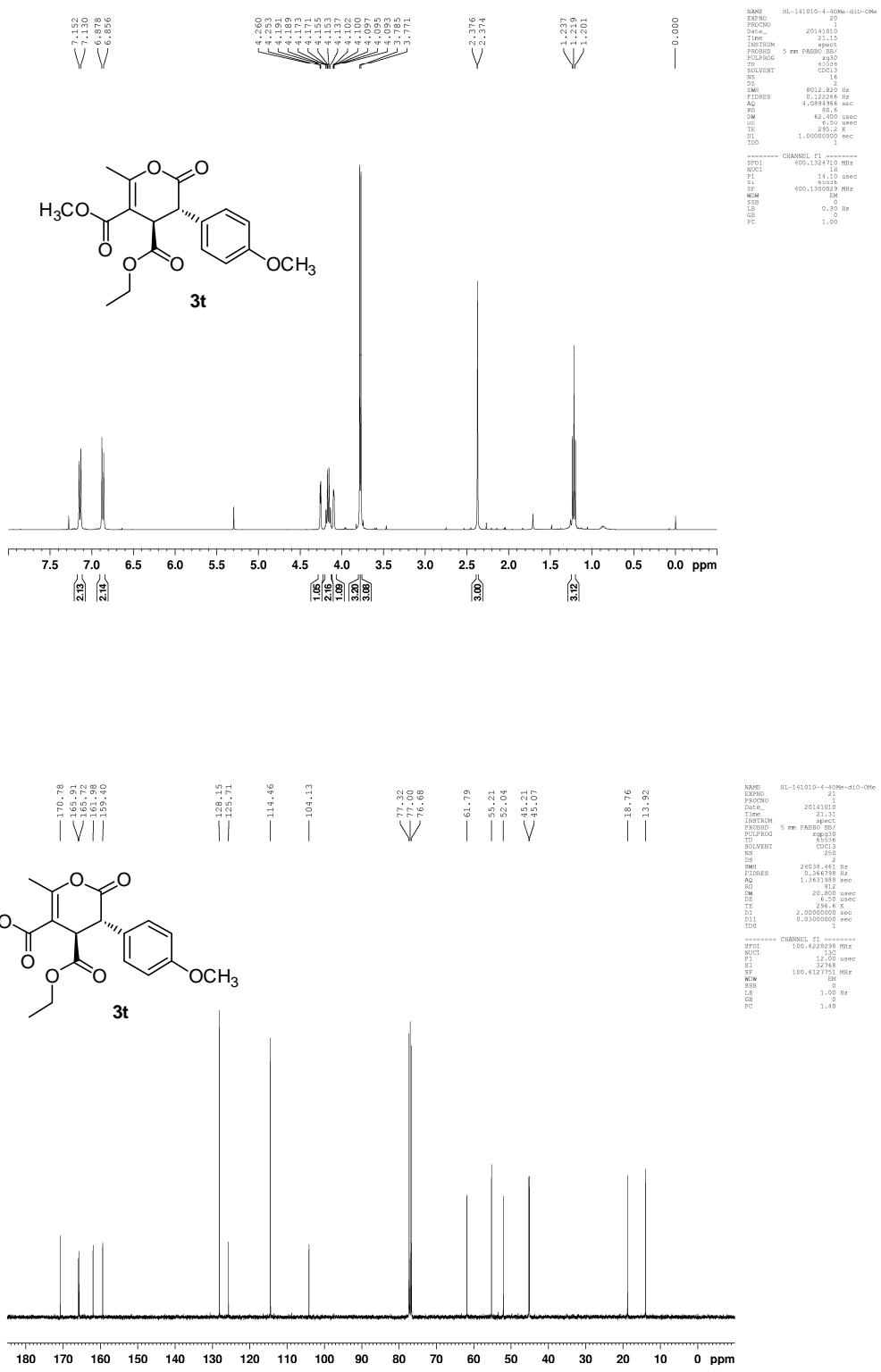
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 PULPROG: D9P
 TD: 65536
 SOLVENT: CDCl3
 NM: 6
 DS:
 SWH: 6012.650 Hz
 FIDRES: 0.122264 Hz
 AQ: 4.000000 sec
 PT: 6.000000 sec
 TM: 62,400 usec
 DE: 62.50 usec
 TE: 293.0 K
 D1: 1.0000000 sec
 TDD: 1

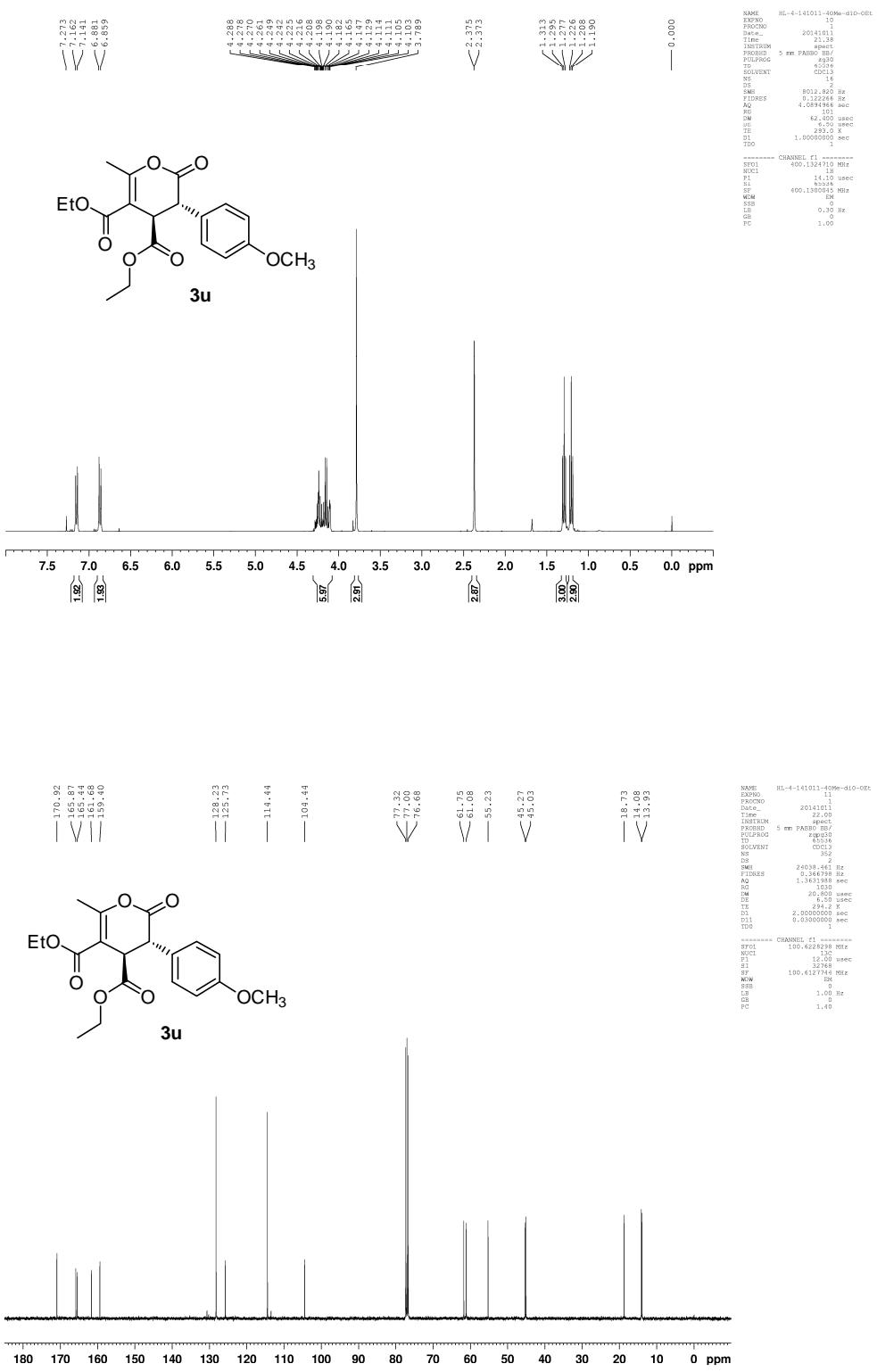


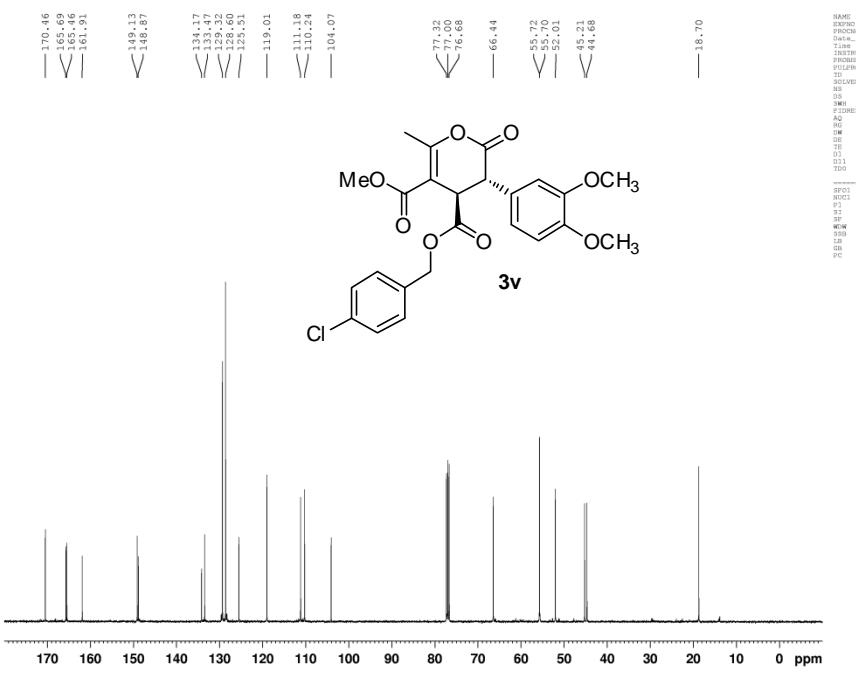
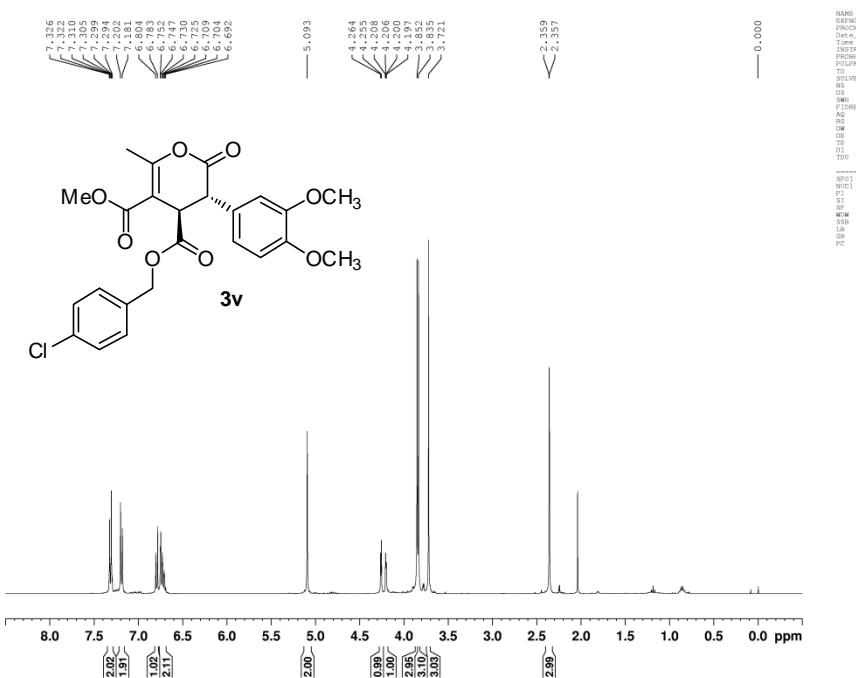
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 PCPNO: 1
 Date: 20140411
 Time: 11:13
 INSTRUM: spect
 PROBHD: 5 mm PABBO BB/
 PULPROG: D9P
 TD: 65536
 SOLVENT: CDCl3
 NM: 6
 DS:
 SWH: 24002.650 Hz
 FIDRES: 0.366798 Hz
 AQ: 1.363198 sec
 PT: 6.000000 sec
 TM: 20,800 usec
 DE: 20.000 usec
 TE: 293.0 K
 D1: 2.0000000 sec
 D11: 0.0300000 sec
 TDD: 1

NAME: HL-4-chawwu-2naph
 EXPNO: 100,6228298 MHz
 PCPNO: 1
 Date: 20140411
 Time: 14:45
 INSTRUM: spect
 PROBHD: 5 mm PABBO BB/
 PULPROG: D9P
 TD: 65536
 SOLVENT: CDCl3
 NM: 6
 DS:
 SWH: 100,6228298 Hz
 FIDRES: 32768
 AQ: 100,6127100 Hz
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 TE: 293.0 K
 D1: 1.0000000 sec
 D11: 0.0300000 sec
 TDD: 1



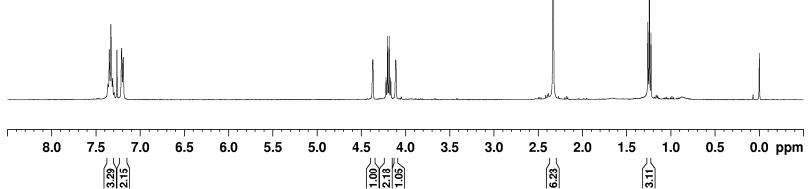
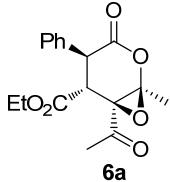






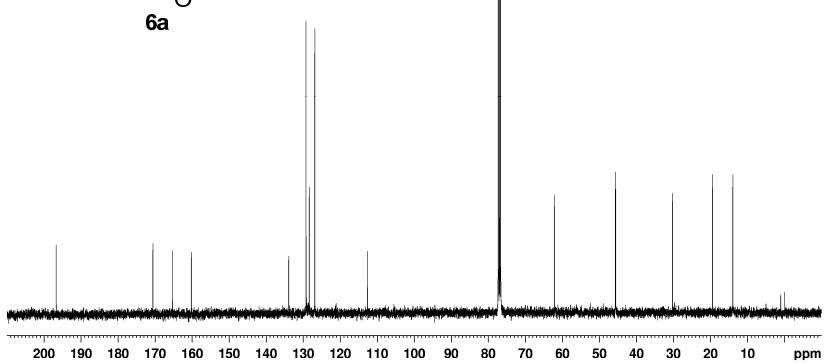
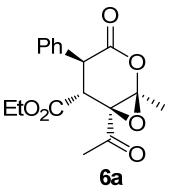
NAME HL-4-Ph-exepoxide
 EXPNO 10
 PROBNO 1
 Date_ 20140702
 Time 22:03
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 POLPROJ 0
 TDR 65336
 SOLVENT CDCl₃
 NS 2
 DS 1
 SWH 8012.500 Hz
 FIDRES 0.122266 Hz
 ACQ 4.098496 sec
 NC 65
 DM 62.400 usec
 TDM 1.530 usec
 DE 2.400 usec
 TE 293.8 K
 D1 1.0000000 sec
 TDO 1

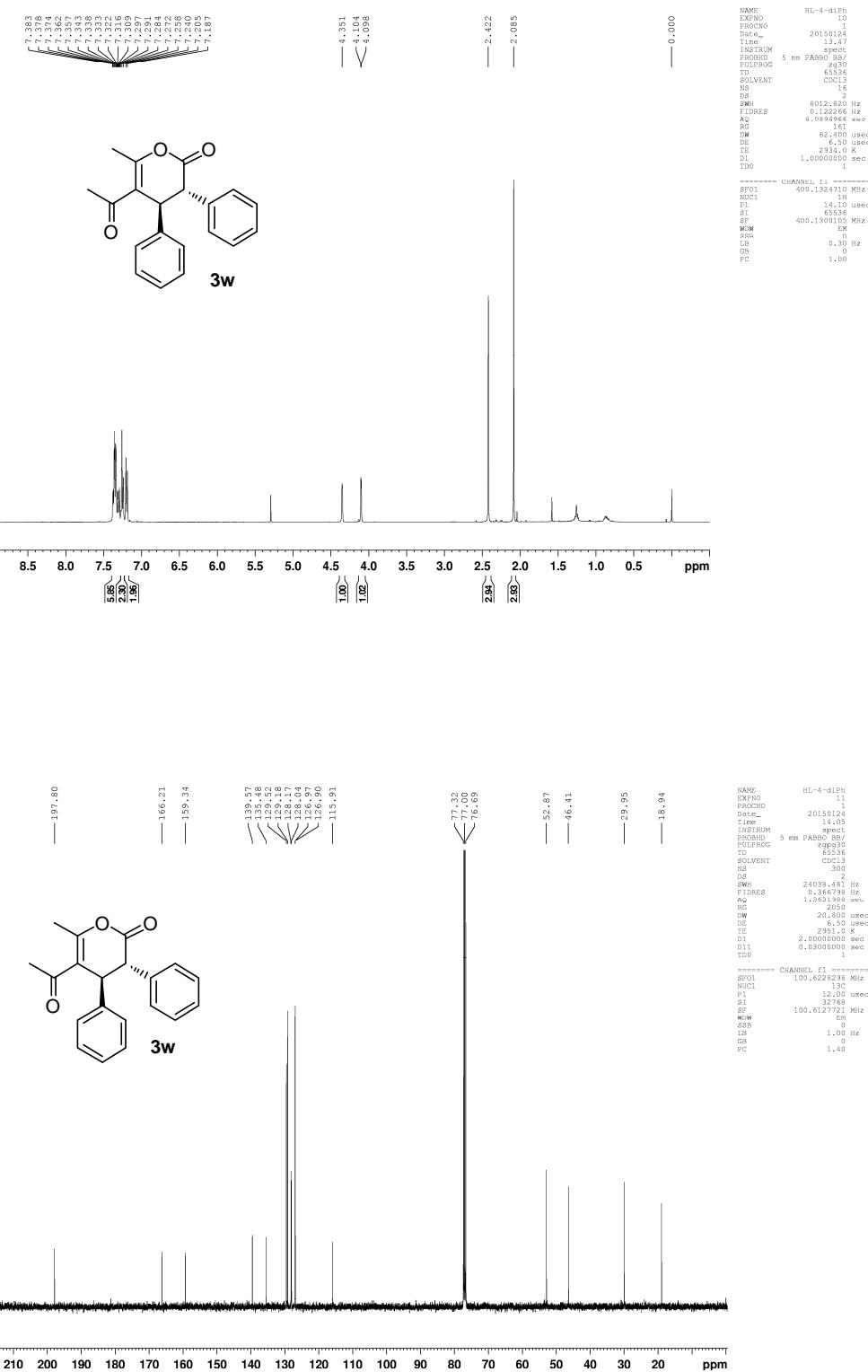
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 NUC1 1H
 P1 14.110 used
 ST 1.000 sec
 SF 400.1330085 MHz
 WDW EM
 PRG 1
 LB 0.30 Hz
 GB 0
 PC 1.00

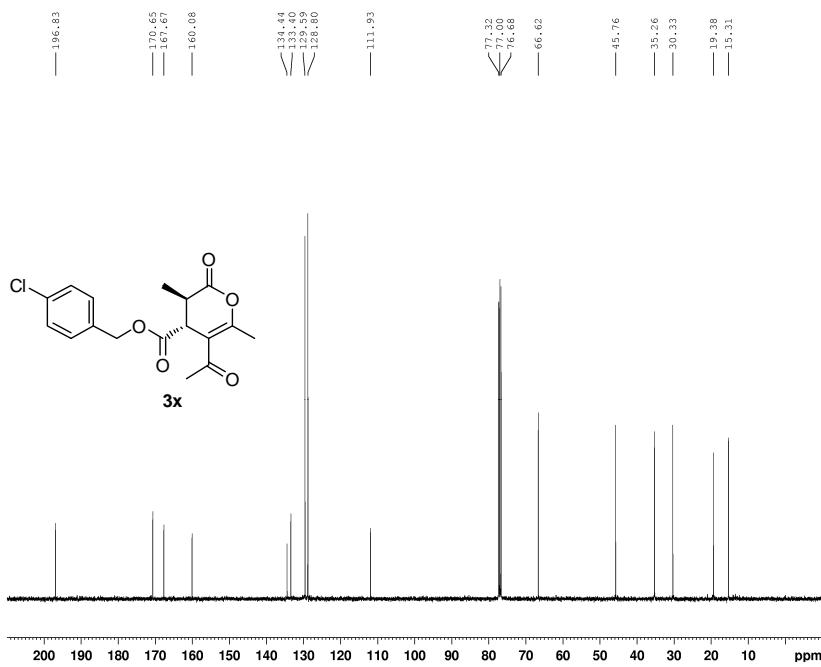
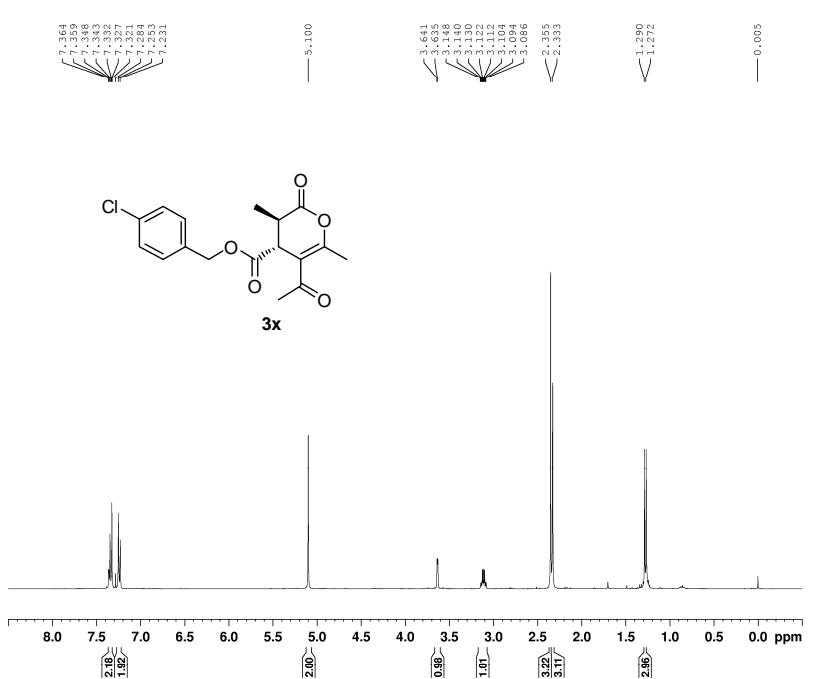


NAME HL-4-140706-Ph-exepoxide
 EXPNO 1
 PROBNO 1
 Date_ 20140706
 Time 22:18
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 POLPROJ 0.530
 TDR 65336
 SOLVENT CDCl₃
 NS 200
 DS 2
 SWH 24038.198 Hz
 FIDRES 0.366198 Hz
 ACQ 1.363100 sec
 NC 1024
 DM 62.400 usec
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 DE 2.979 K
 TE 2.0000000 sec
 D1 0.0300000 sec
 TDO 1

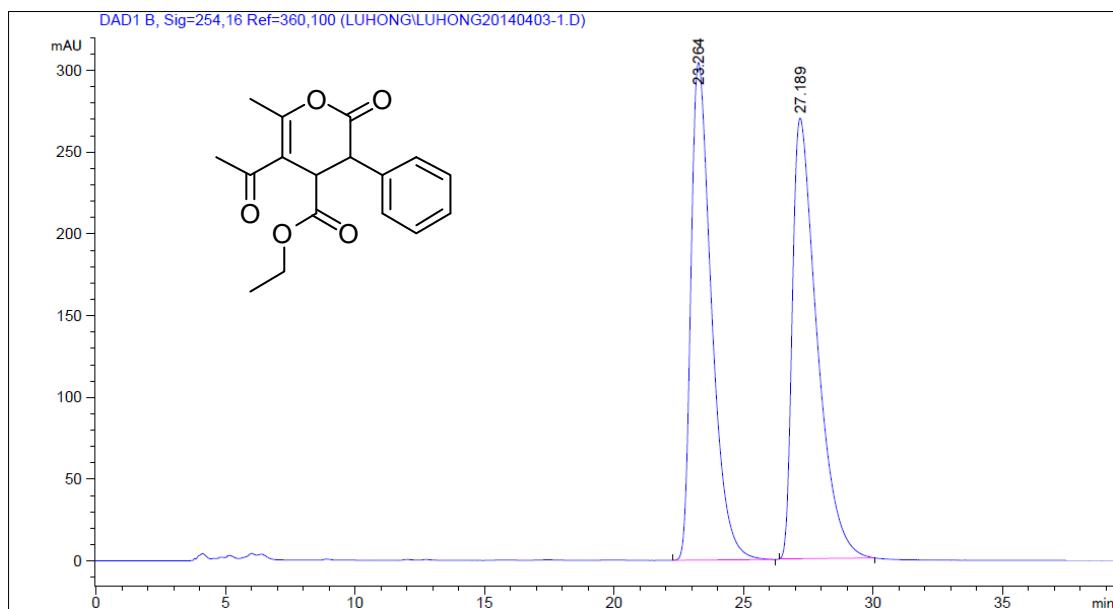
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 NUC1 13C
 P1 12.00 usec
 ST 32768
 SF 100.619778 MHz
 NMM EM
 DRS 0
 LB 1.00 Hz
 GB 0
 PC 1.40



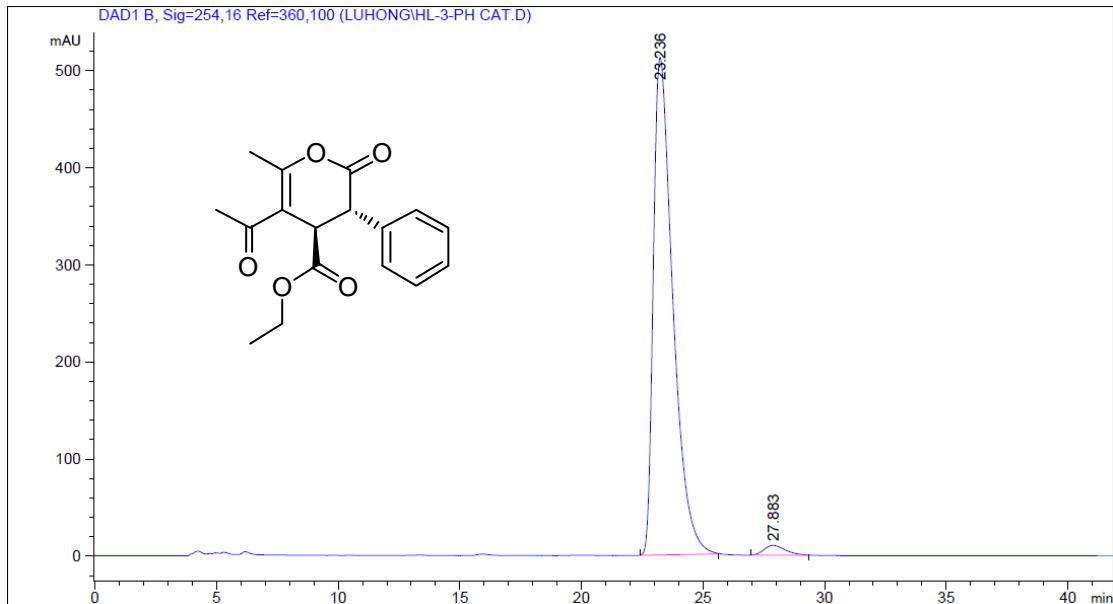




3a

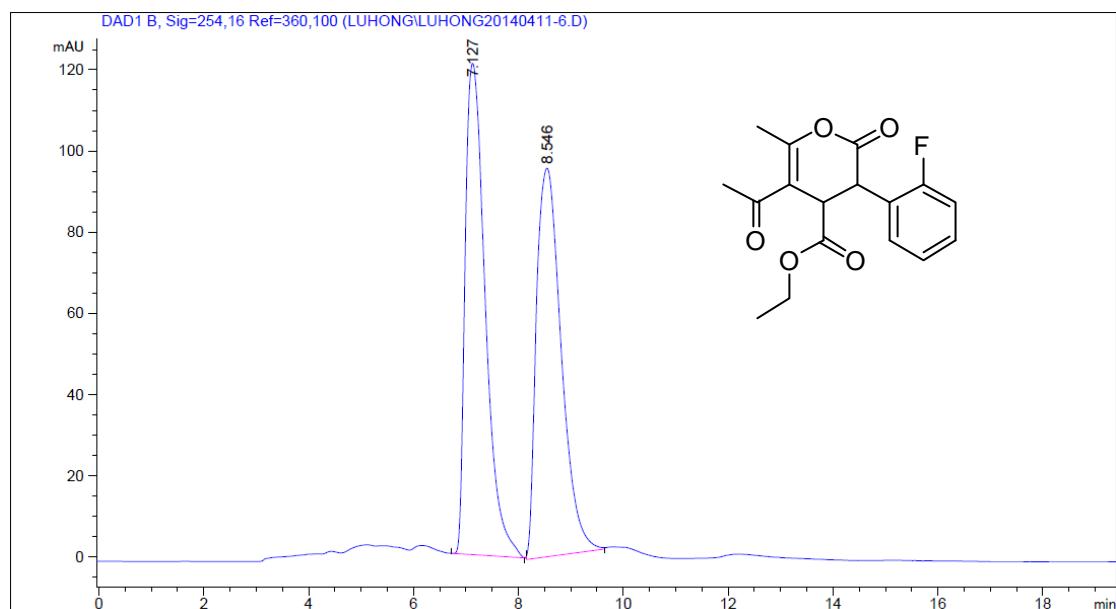


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	23.264	1.67971e4	304.28860	49.6592
2	DAD 254, 16 nm	27.189	1.77228e4	269.36551	50.3408

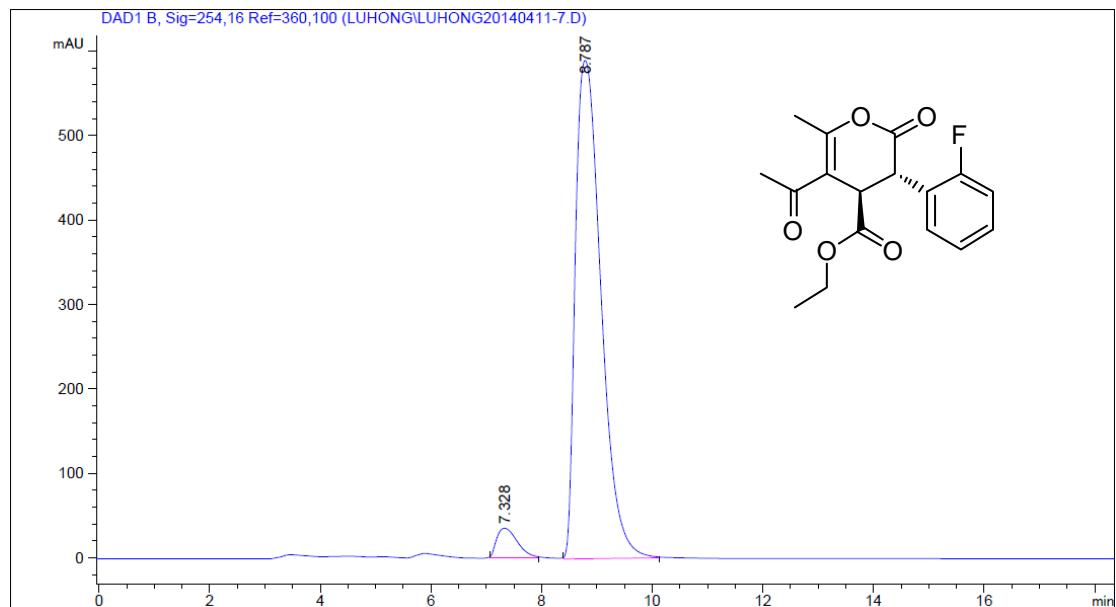


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	23.236	2.85998e4	512.54529	97.9166
2	DAD 254, 16 nm	27.883	608.53680	9.97842	2.0834

3b

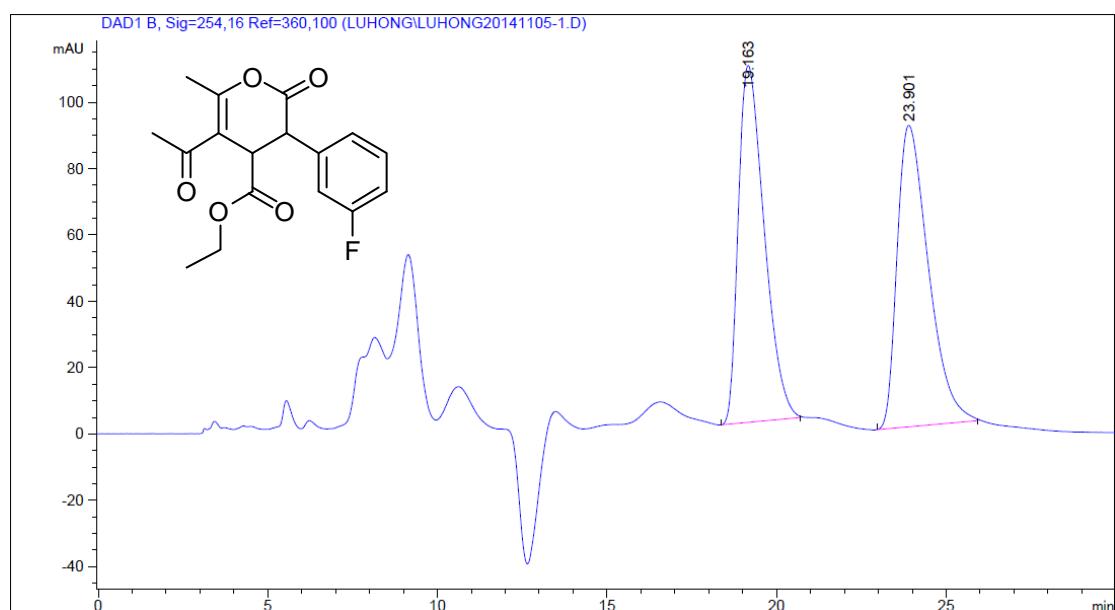


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	7.127	3217.05884	121.02797	50.5970
2	DAD 254, 16 nm	8.546	3141.13696	95.70715	49.4030

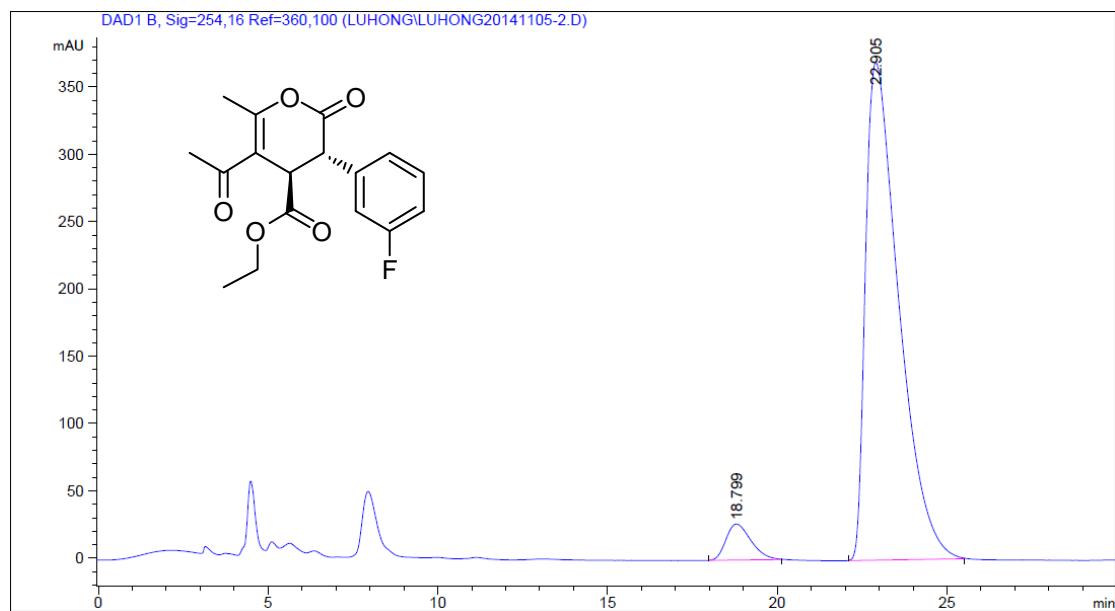


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	7.328	905.84814	34.85149	4.5650
2	DAD 254, 16 nm	8.787	1.89375e4	590.19281	95.4350

3c

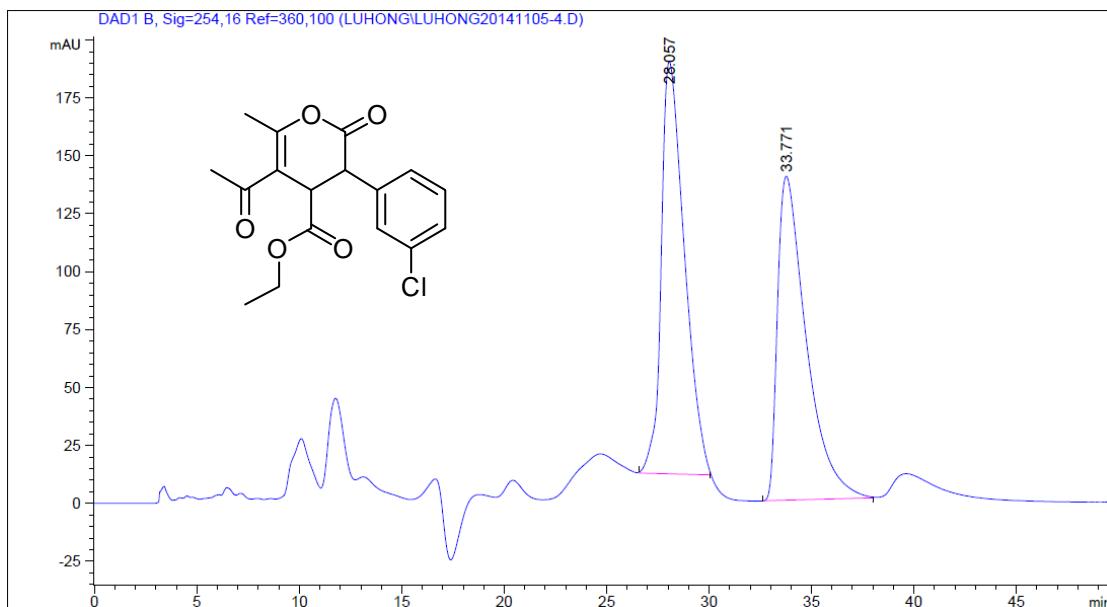


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	19.163	5708.45703	107.59546	49.5045
2	DAD 254, 16 nm	23.901	5822.72803	90.86617	50.4955

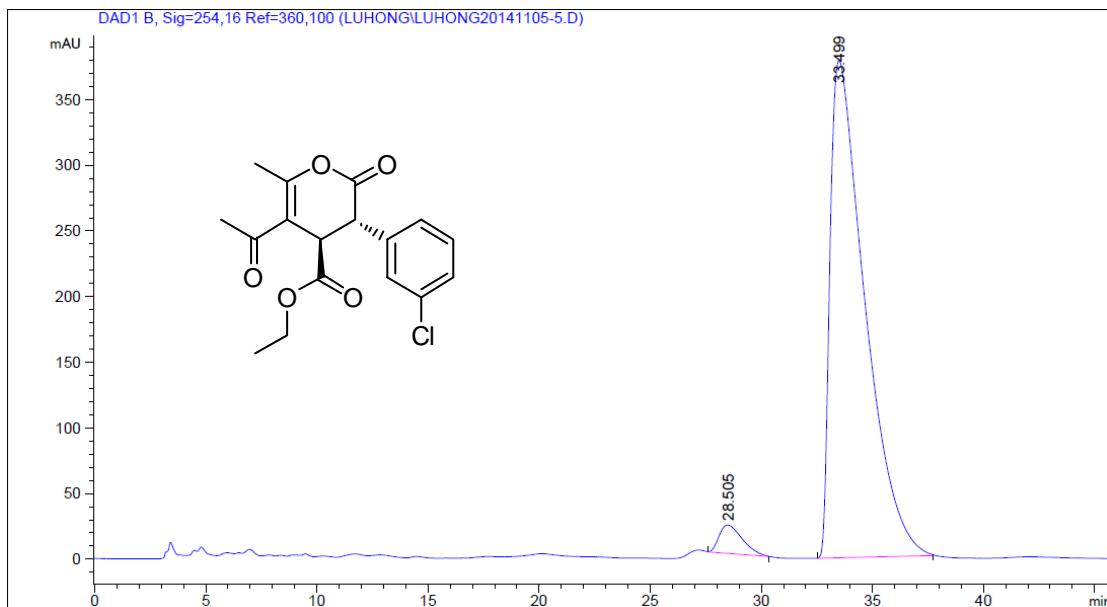


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	18.799	1369.08203	26.75384	5.1654
2	DAD 254, 16 nm	22.905	2.51358e4	369.87375	94.8346

3d

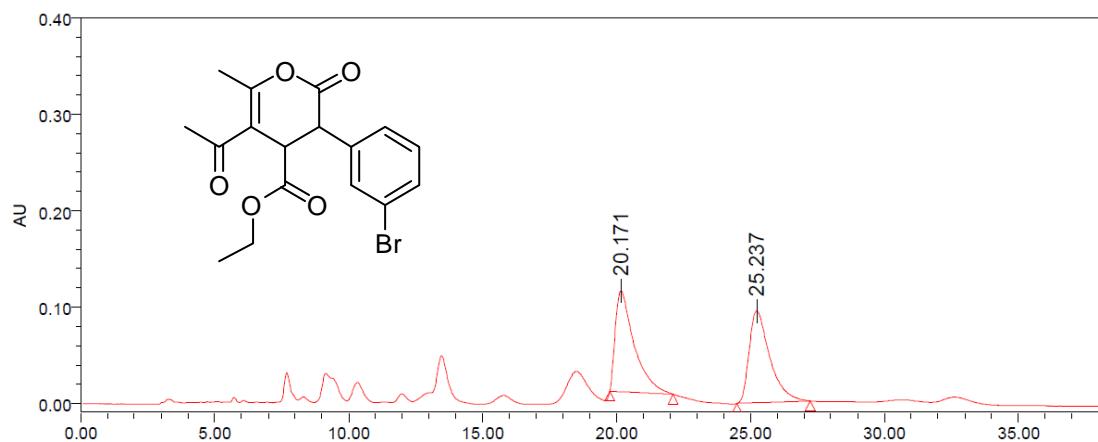


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	28.057	1.39738e4	178.06345	50.6433
2	DAD 254, 16 nm	33.771	1.36187e4	139.83339	49.3567

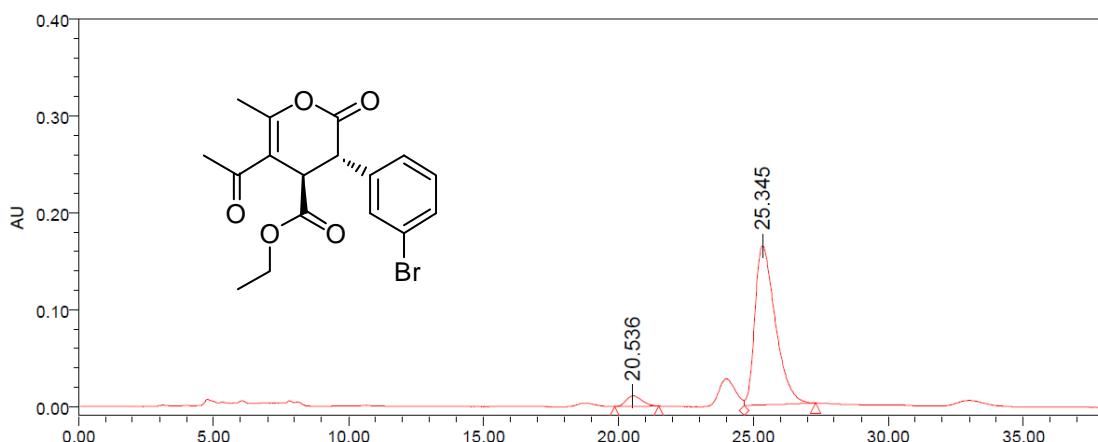


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	28.505	1565.34631	21.65634	3.6695
2	DAD 254, 16 nm	33.499	4.10933e4	378.78516	96.3305

3e

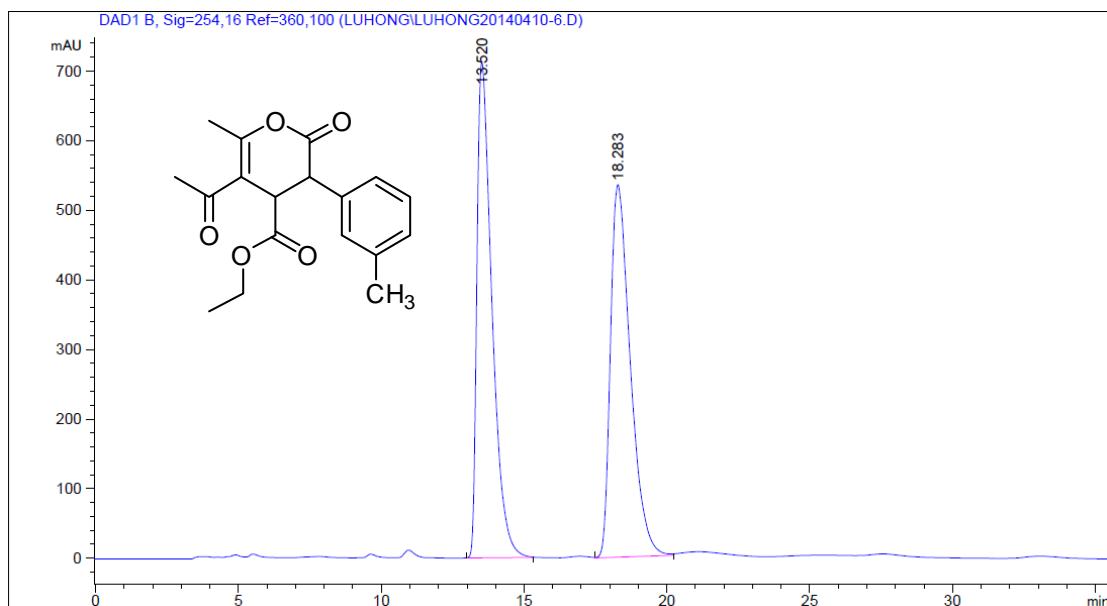


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	PDA254 nm	20.171	5129799	104702	50.13
2	PDA 254nm	25.237	5102963	95283	49.87

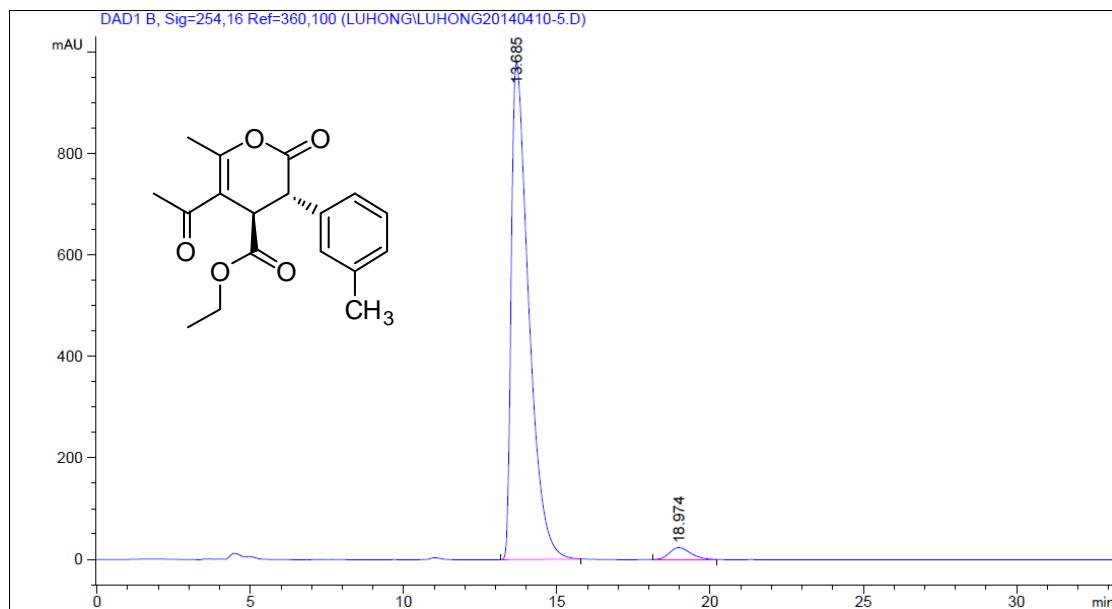


Peak	Processed Channel	Retention Time (min)	Peak Area (AU*s)	Peak Height (AU)	Peak Area (%)
1	PDA254 nm	20.536	453859	10968	5.00
2	PDA254 nm	25.345	8625663	164280	95.00

3f

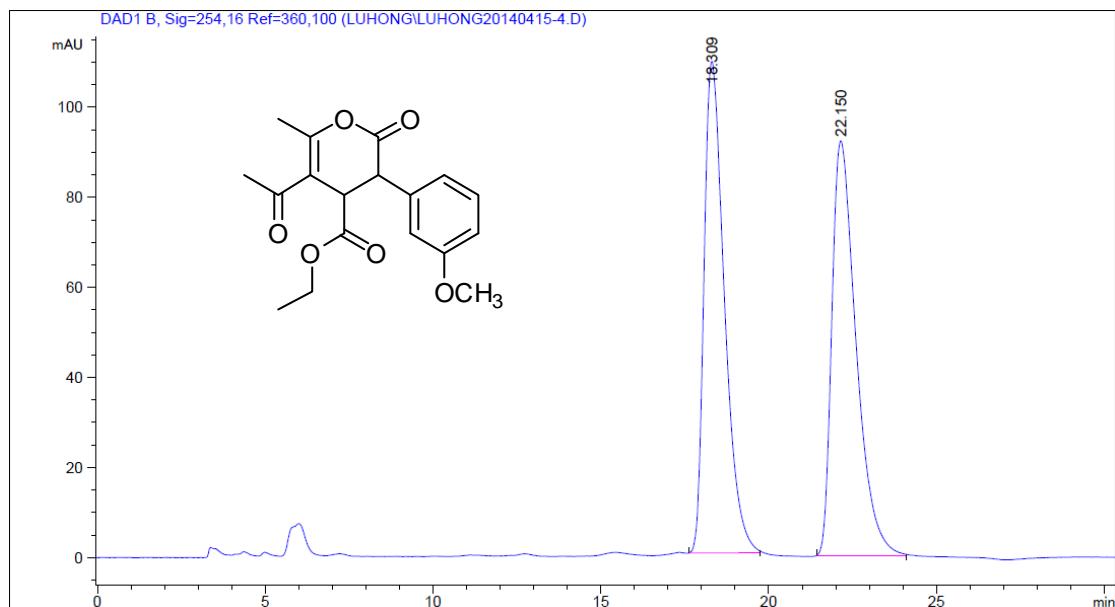


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	13.520	2.60074e4	712.30029	50.1697
2	DAD 254, 16 nm	18.283	2.58315e4	535.31207	49.8303

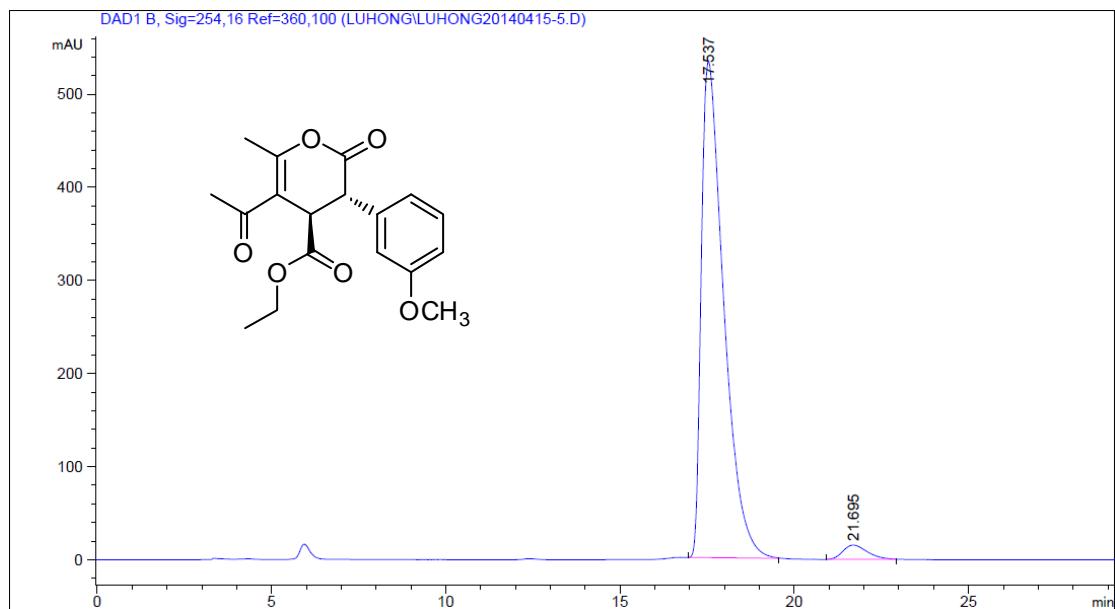


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	13.685	3.87218e4	982.26636	97.1668
2	DAD 254, 16 nm	18.974	1129.05334	23.39927	2.8332

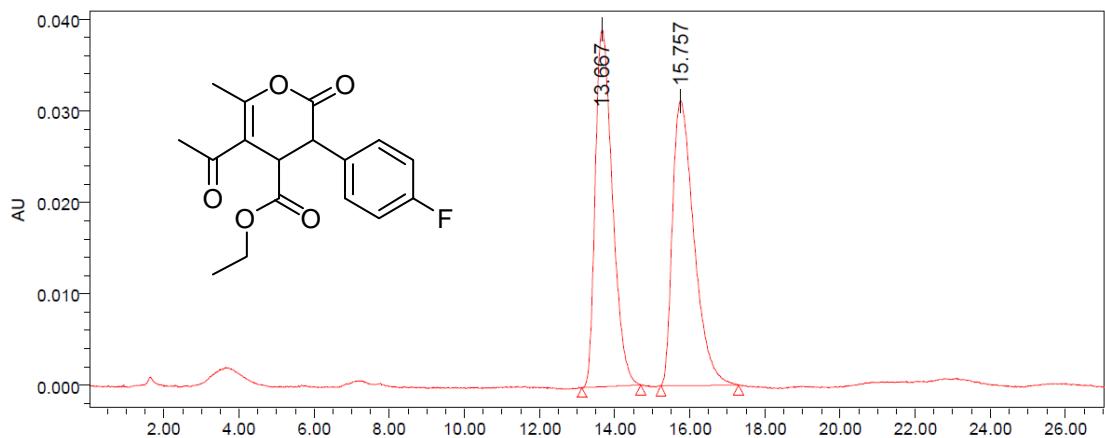
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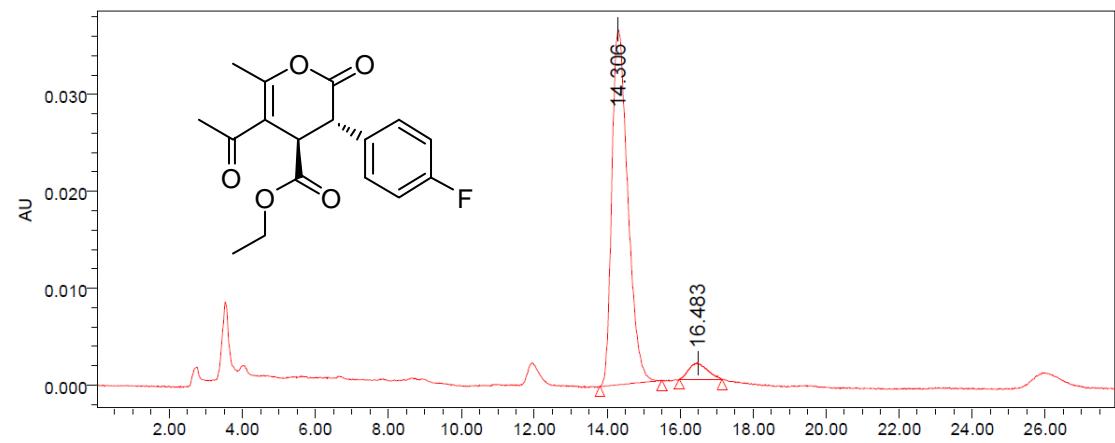
Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	18.309	4591.86377	109.14375	49.7706
2	DAD 254, 16 nm	22.150	4634.18506	92.21734	50.2294



Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	17.537	2.37991e4	533.15991	96.9647
2	DAD 254, 16 nm	21.695	744.98730	15.36586	3.0353

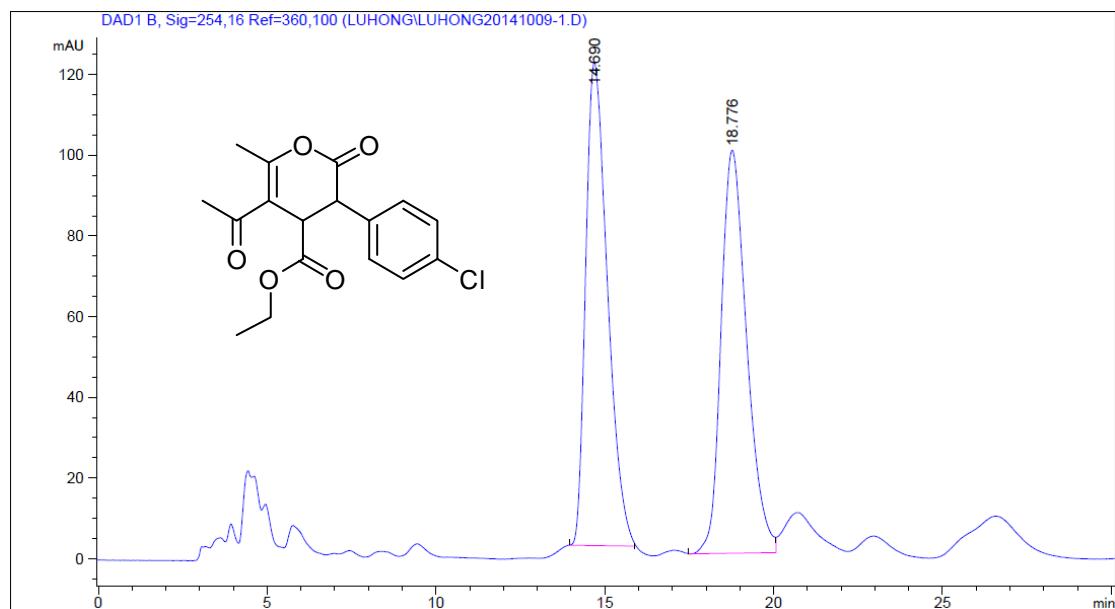
3h

Peak	Processed Channel	Retention Time (min)	Peak Area (AU*s)	Peak Height (AU)	Peak Area (%)
1	PDA246.9 nm	13.667	1240049	39046	49.90
2	PDA246.9 nm	15.757	1245039	31146	50.10

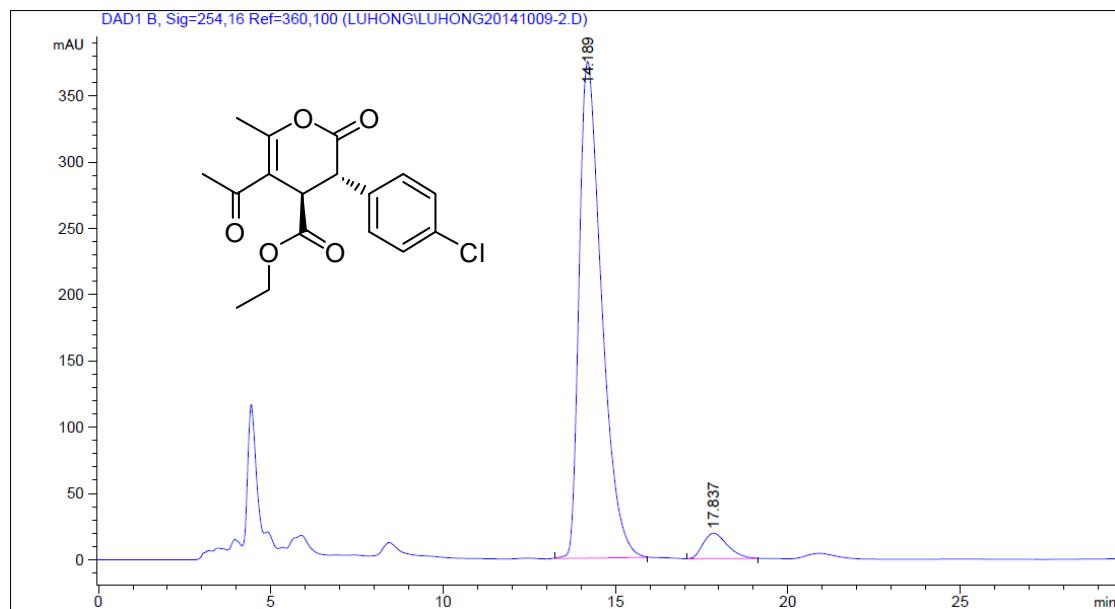


Peak	Processed Channel	Retention Time (min)	Peak Area (AU*s)	Peak Height (AU)	Peak Area (%)
1	PDA246.9 nm	14.306	1109875	36656	94.95
2	PDA246.9 nm	16.483	58971	1670	5.05

3i

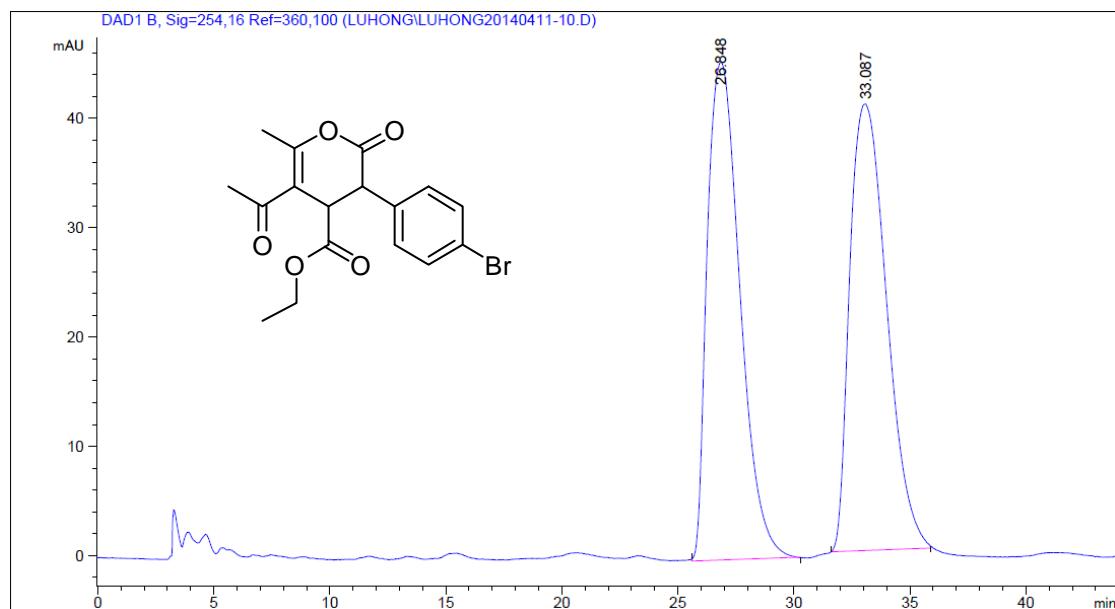


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	14.690	5360.80029	119.75430	50.4051
2	DAD 254, 16 nm	18.776	5274.63818	99.87841	49.5949

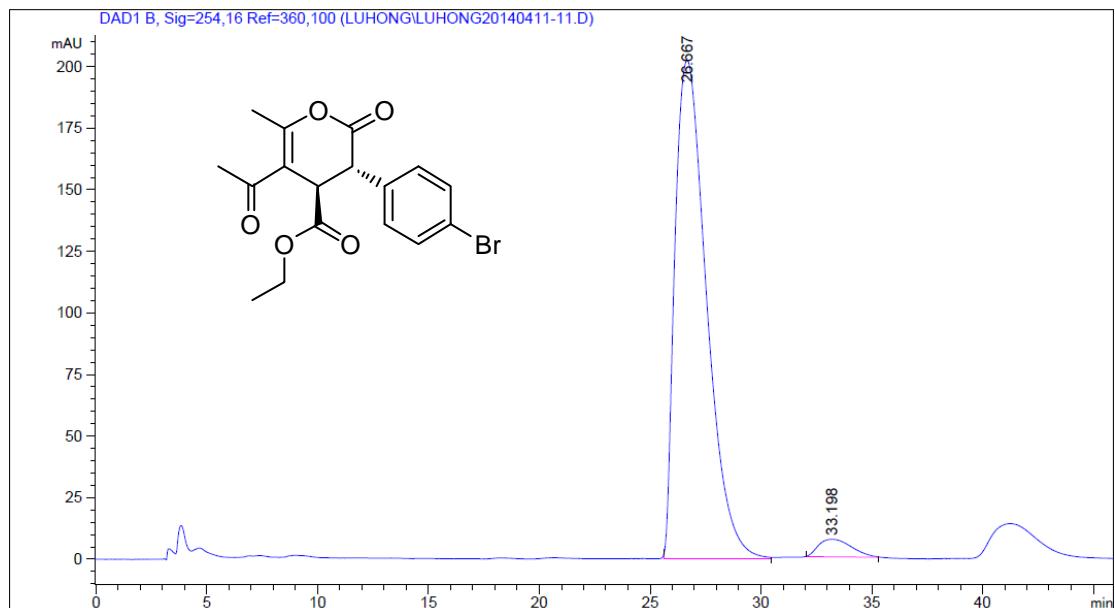


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	14.189	1.69883e4	374.87115	94.7678
2	DAD 254, 16 nm	17.837	937.93311	19.23451	5.2322

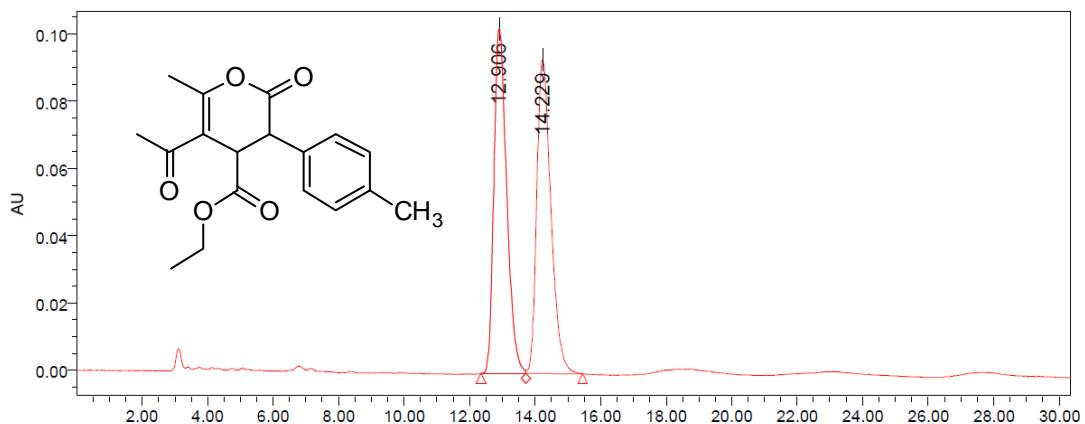
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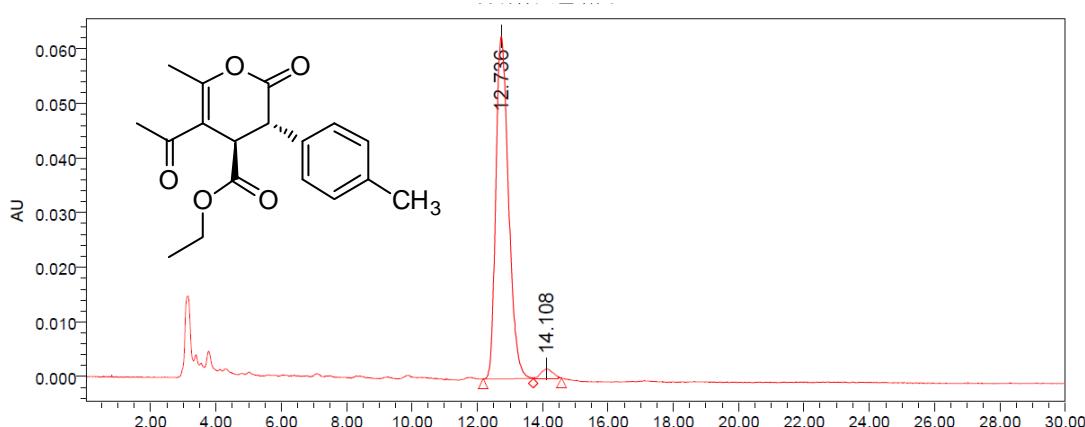
Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	26.848	4462.83496	45.53188	49.7318
2	DAD 254, 16 nm	33.087	4510.97314	40.85940	50.2682



Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	26.667	1.99684e4	202.37297	96.4331
2	DAD 254, 16 nm	33.198	738.58948	7.13121	3.5669

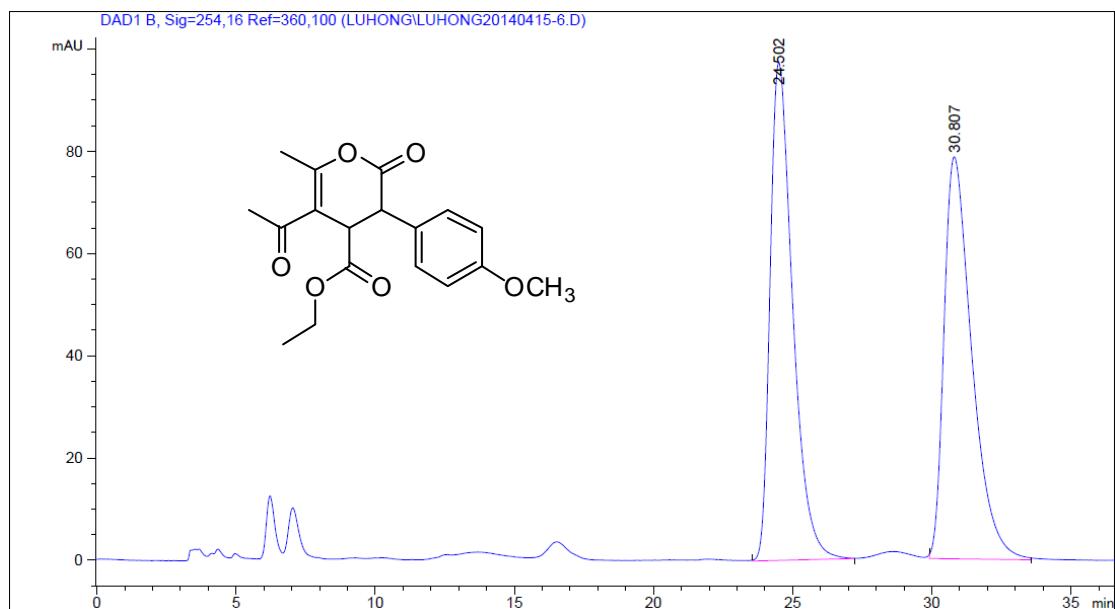
3k

Peak	Processed Channel	Retention Time (min)	Peak Area (AU*s)	Peak Height (AU)	Peak Area (%)
1	PDA246.9 nm	12.906	2790713	102444	49.88
2	PDA246.9 nm	14.229	2803883	93367	50.12

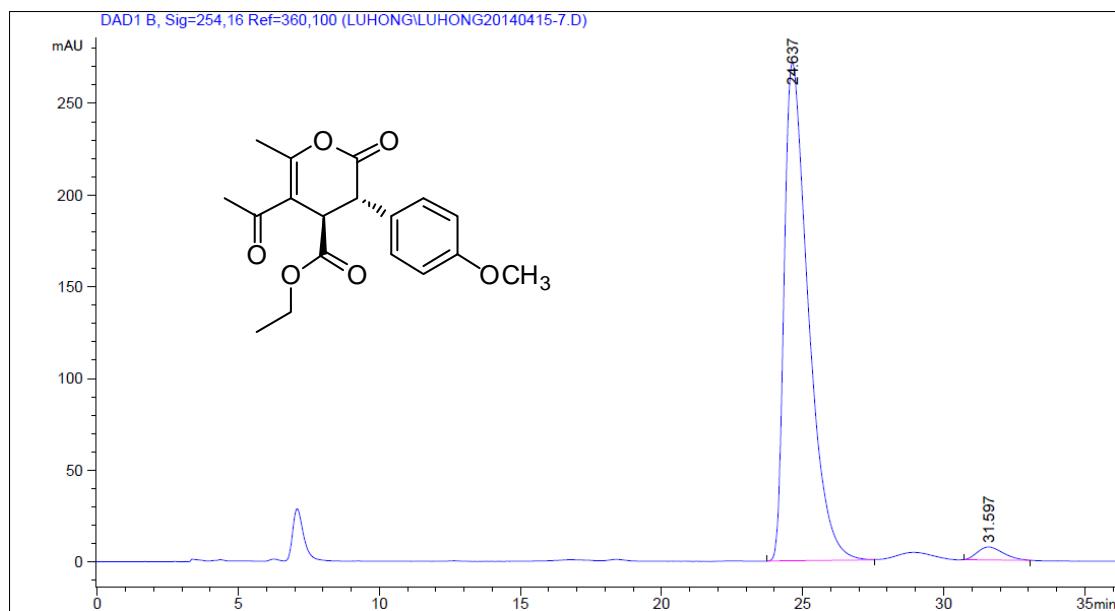


Peak	Processed Channel	Retention Time (min)	Peak Area (AU*s)	Peak Height (AU)	Peak Area (%)
1	PDA246.9 nm	12.736	1690588	62738	97.49
2	PDA246.9 nm	14.108	43514	1731	2.51

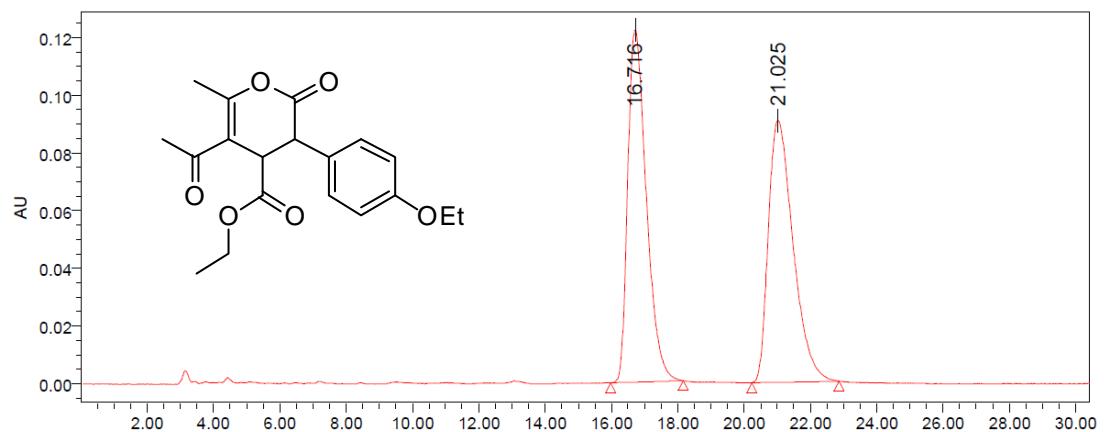
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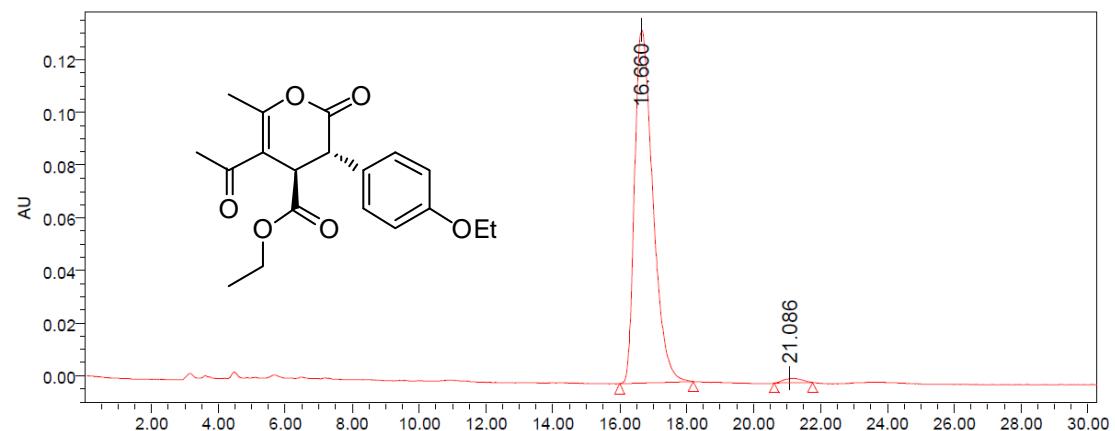
Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	24.502	5495.23730	97.43780	50.1952
2	DAD 254, 16 nm	30.807	5452.49951	78.66367	49.8048



Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	24.637	1.60160e4	271.59885	97.2659
2	DAD 254, 16 nm	31.597	450.20514	7.07779	2.7341

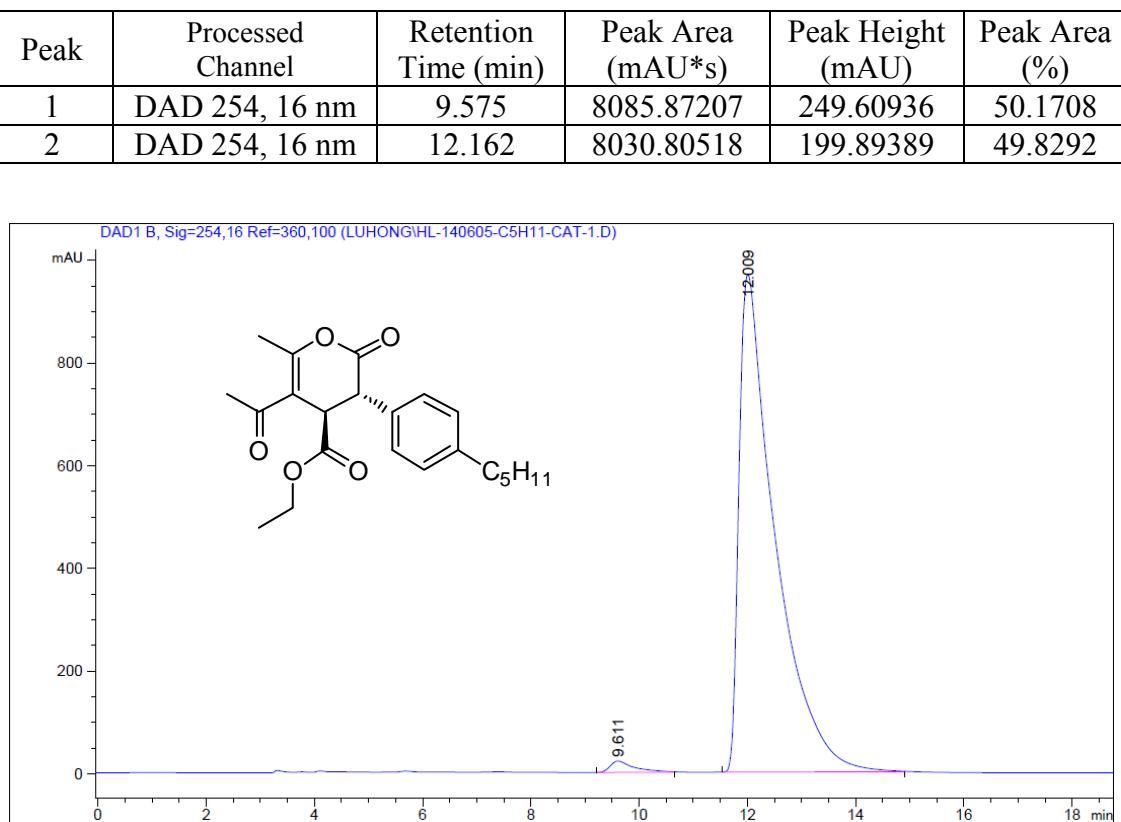
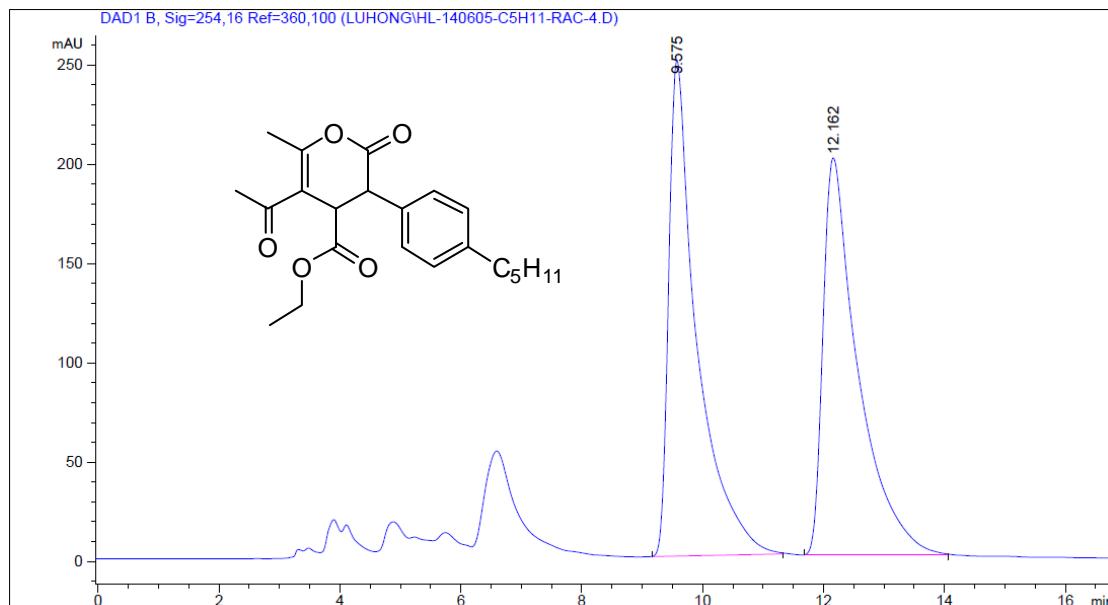
3m

Peak	Processed Channel	Retention Time (min)	Peak Area (AU*s)	Peak Height (AU)	Peak Area (%)
1	PDA246.9 nm	16.716	4743145	122087	50.19
2	PDA246.9 nm	21.025	4707496	90775	49.81

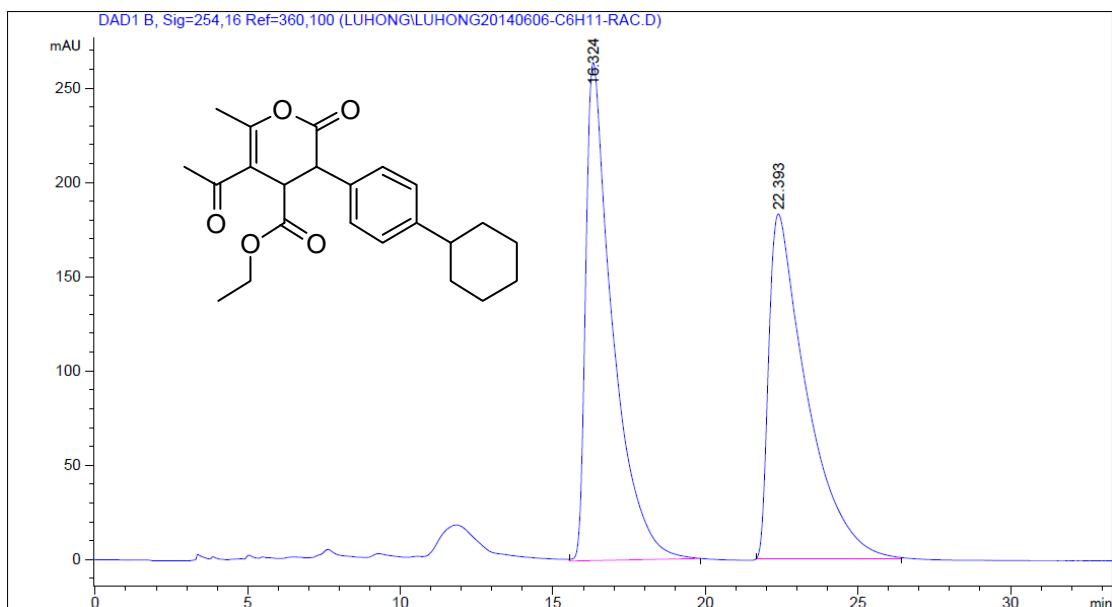


Peak	Processed Channel	Retention Time (min)	Peak Area (AU*s)	Peak Height (AU)	Peak Area (%)
1	PDA246.9 nm	16.660	5088941	134227	98.77
2	PDA246.9 nm	21.086	63474	1641	1.23

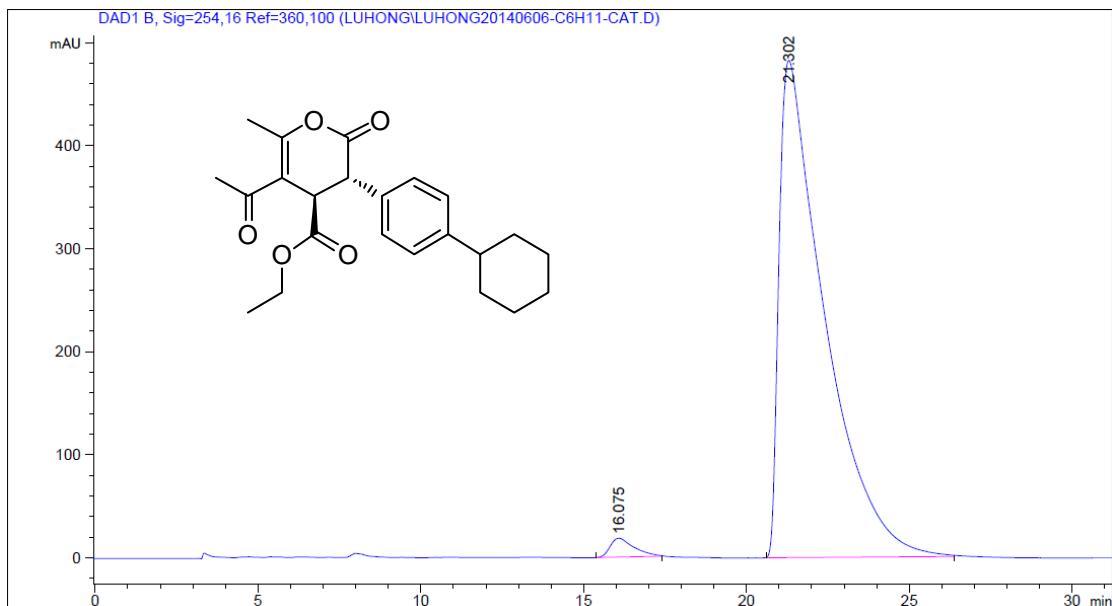
3n



3o

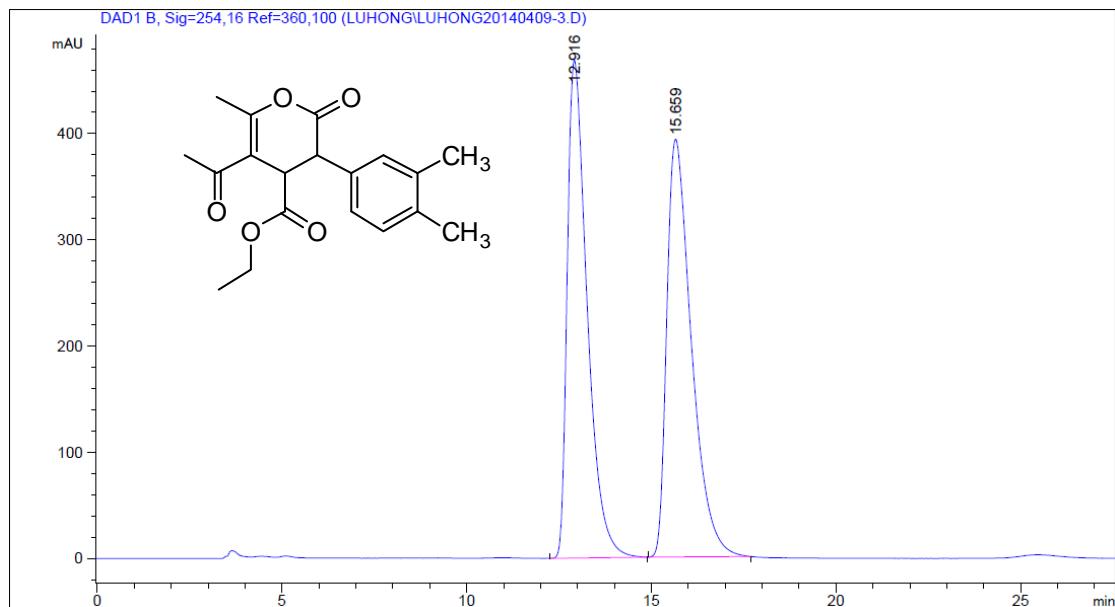


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	16.324	1.60939e4	264.19598	50.5960
2	DAD 254, 16 nm	22.393	1.57148e4	182.63591	49.4040

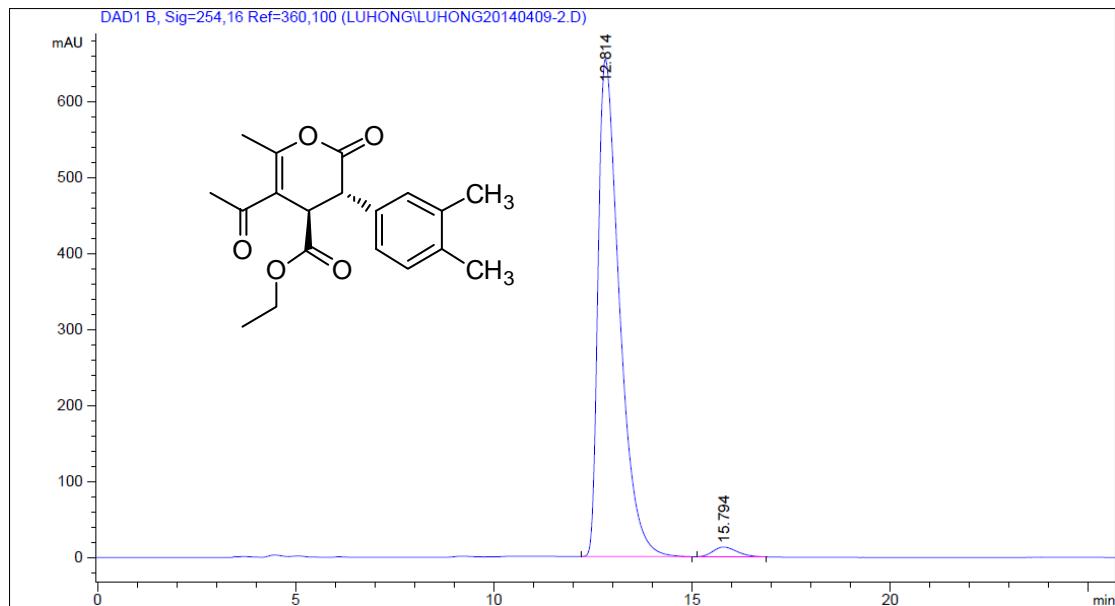


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	16.075	935.47333	18.44045	1.9669
2	DAD 254, 16 nm	21.302	4.66265e4	482.55835	98.0331

3p

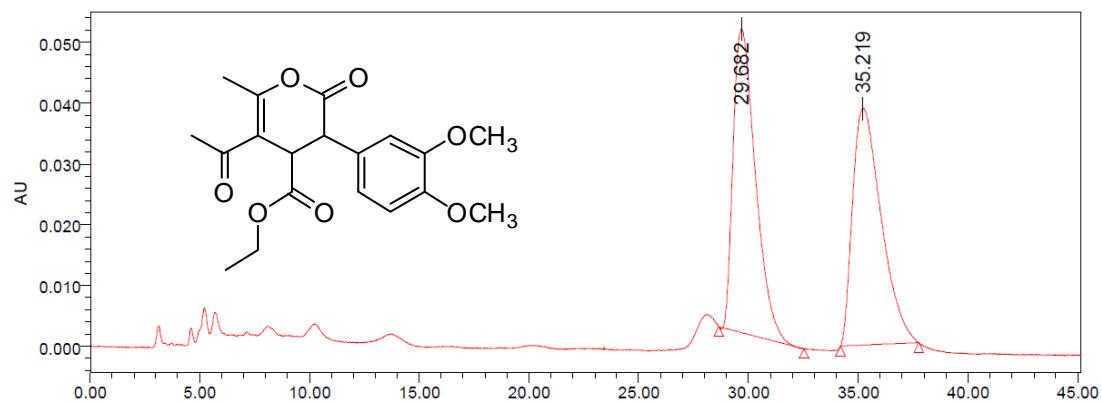


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	12.916	1.75131e4	468.87976	49.0660
2	DAD 254, 16 nm	15.659	1.81799e4	393.25751	50.9340

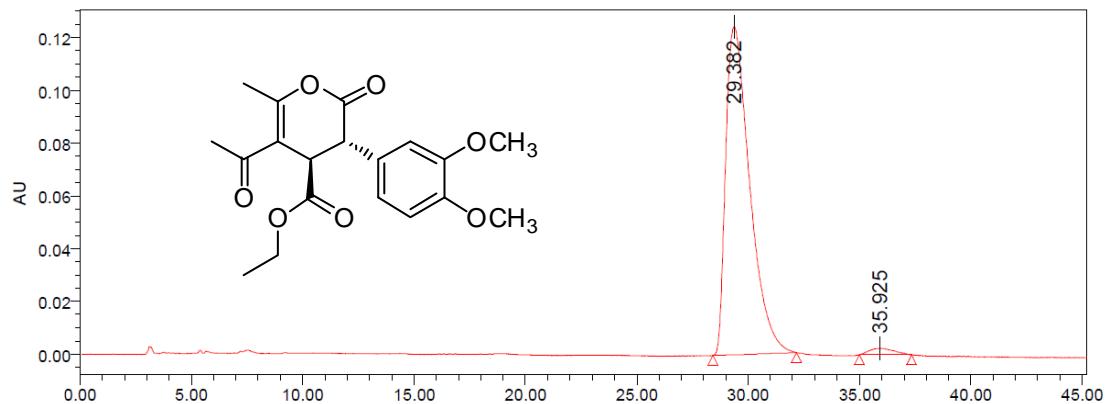


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	12.814	2.44865e4	654.27258	97.8444
2	DAD 254, 16 nm	15.794	539.47058	12.76532	2.1556

3q

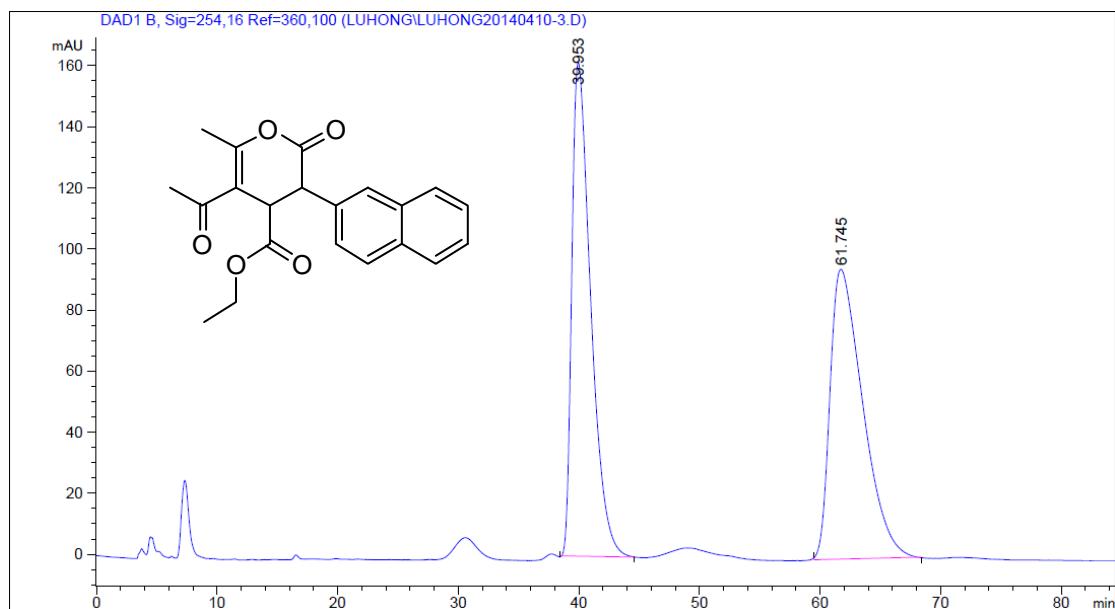


Peak	Processed Channel	Retention Time (min)	Peak Area (AU*s)	Peak Height (AU)	Peak Area (%)
1	PDA246.9 nm	29.682	3540273	49972	50.03
2	PDA246.9 nm	35.219	3535808	38939	49.97

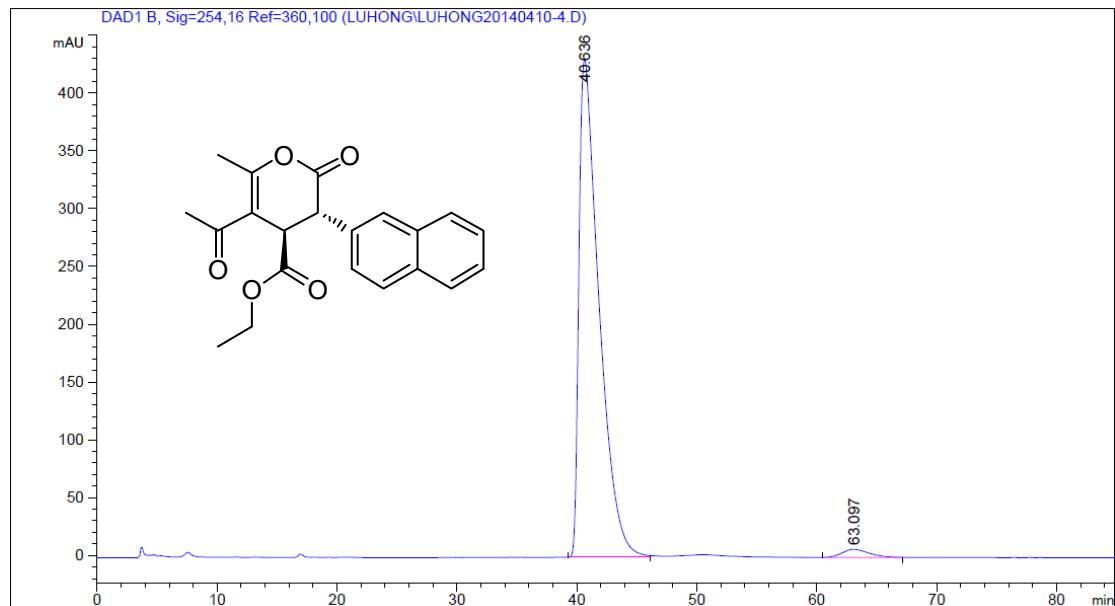


Peak	Processed Channel	Retention Time (min)	Peak Area (AU*s)	Peak Height (AU)	Peak Area (%)
1	PDA246.9 nm	29.382	9339267	124412	97.96
2	PDA246.9 nm	35.925	194163	2466	2.04

3r

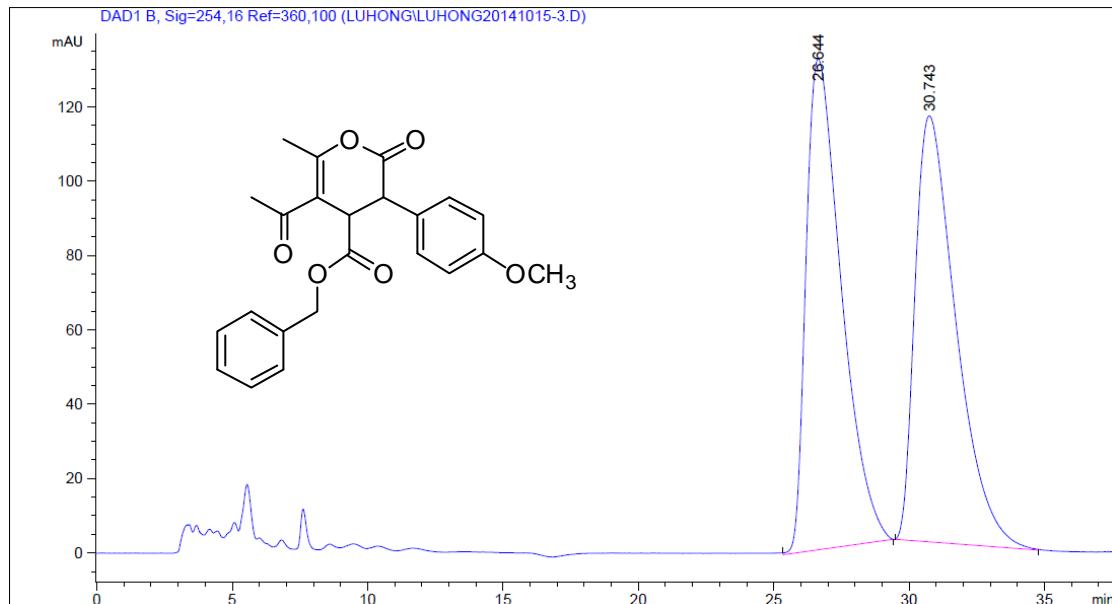


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	39.953	1.75848e4	161.70930	49.6348
2	DAD 254, 16 nm	61.745	1.78436e4	94.90437	50.3652

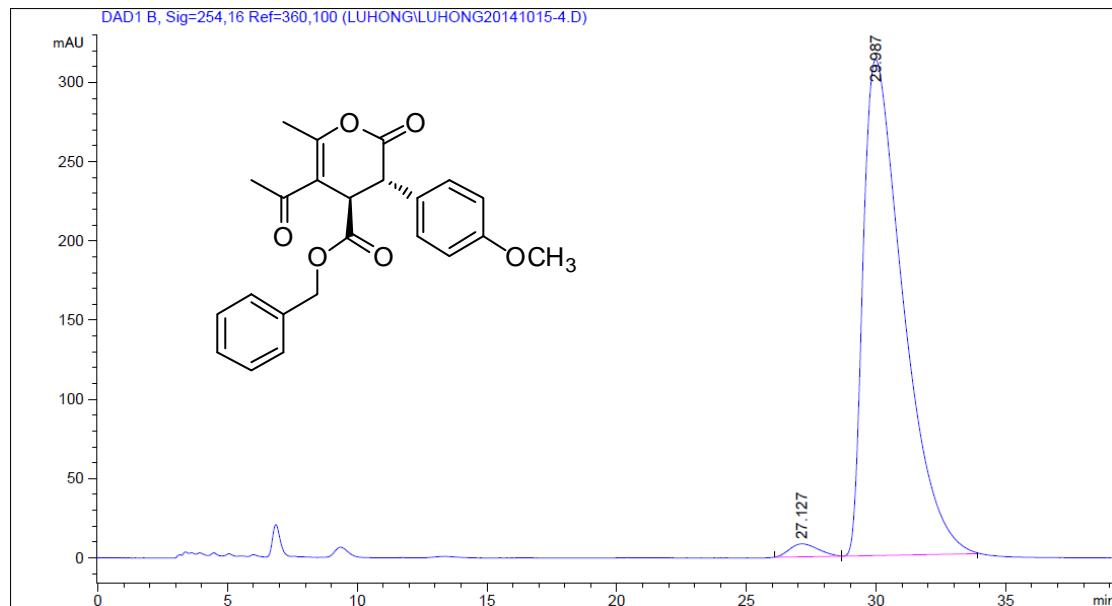


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	40.636	4.99372e4	430.04370	97.8326
2	DAD 254, 16 nm	63.097	1106.34253	7.03193	2.1674

3s

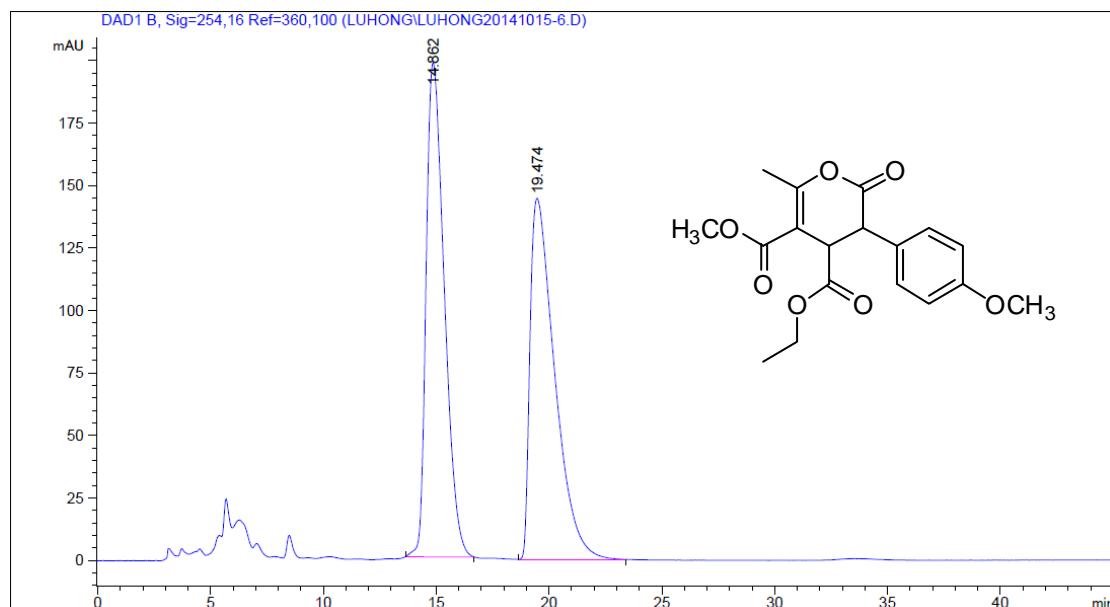


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	26.644	1.19727e4	132.14328	49.8551
2	DAD 254, 16 nm	30.743	1.20423e4	114.58701	50.1449

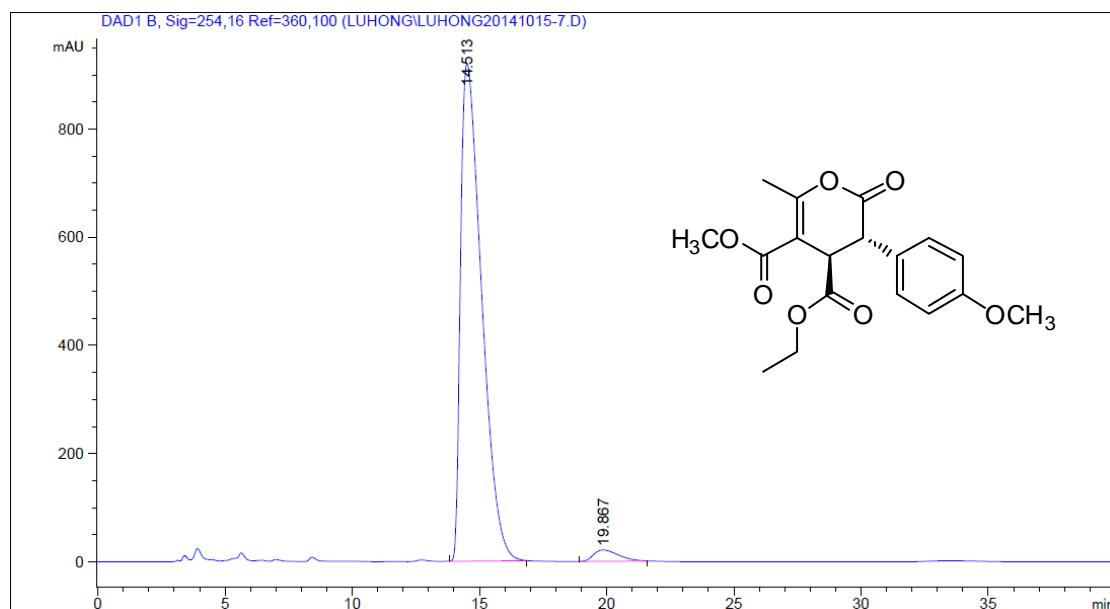


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	27.127	626.90314	8.19894	1.8409
2	DAD 254, 16 nm	29.987	3.34267e4	312.95358	98.1591

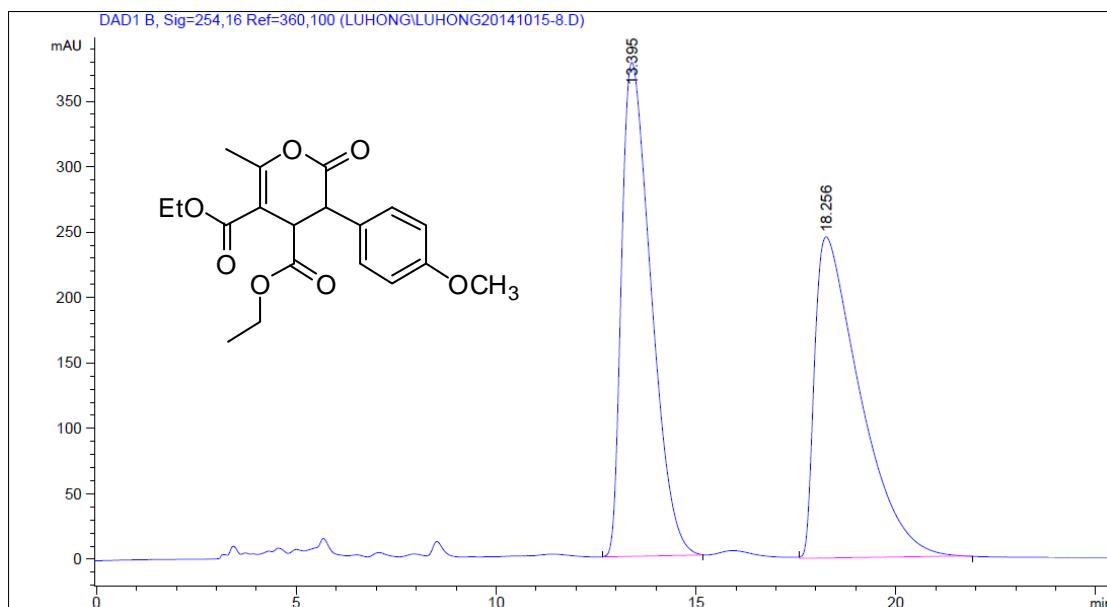
3t



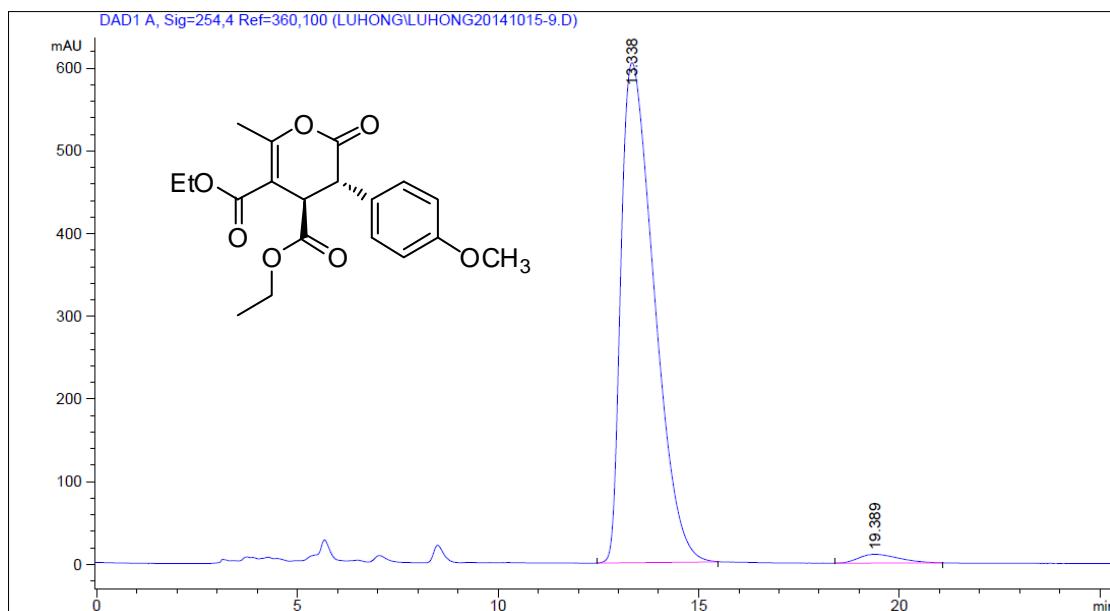
Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	14.862	1.10374e4	197.91924	50.1807
2	DAD 254, 16 nm	19.474	1.09579e4	144.61237	49.8193



Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	14.513	5.27739e4	920.35876	97.3579
2	DAD 254, 16 nm	19.867	1432.20190	20.98921	2.6421

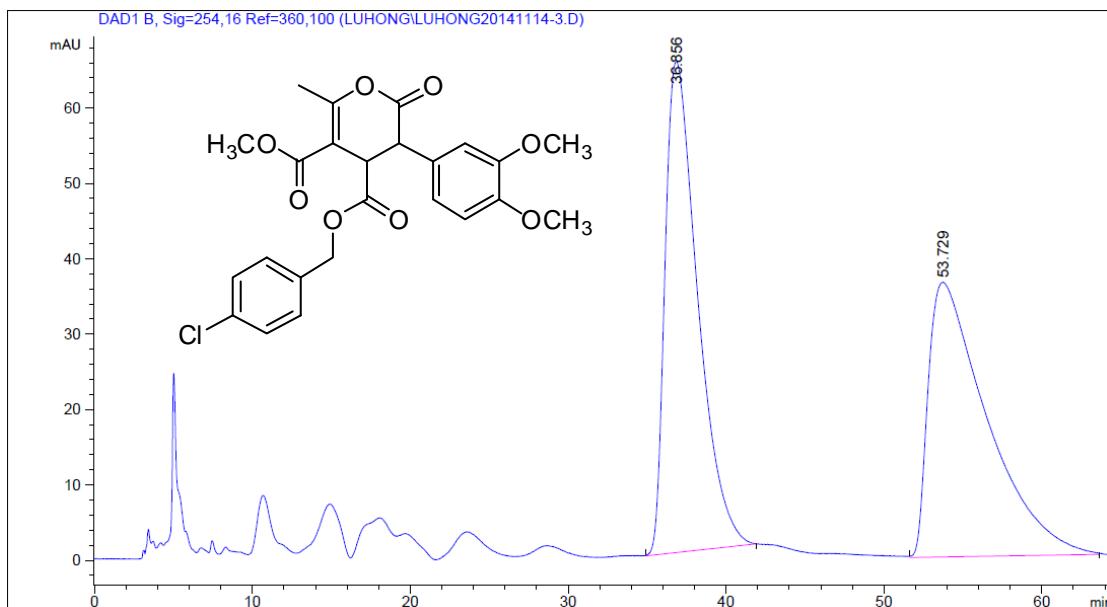
3u

Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	13.395	1.93720e4	377.46848	50.1193
2	DAD 254, 16 nm	18.256	1.92798e4	245.45526	49.8807

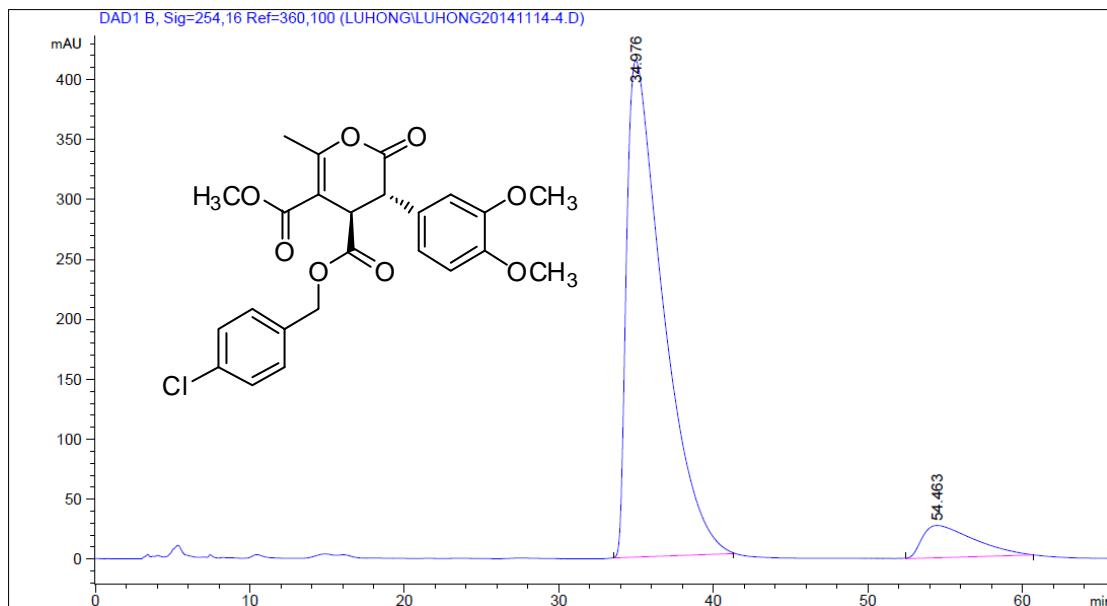


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	13.338	3.46210e4	604.72217	97.9056
2	DAD 254, 16 nm	19.389	740.59613	10.58835	2.0944

3v

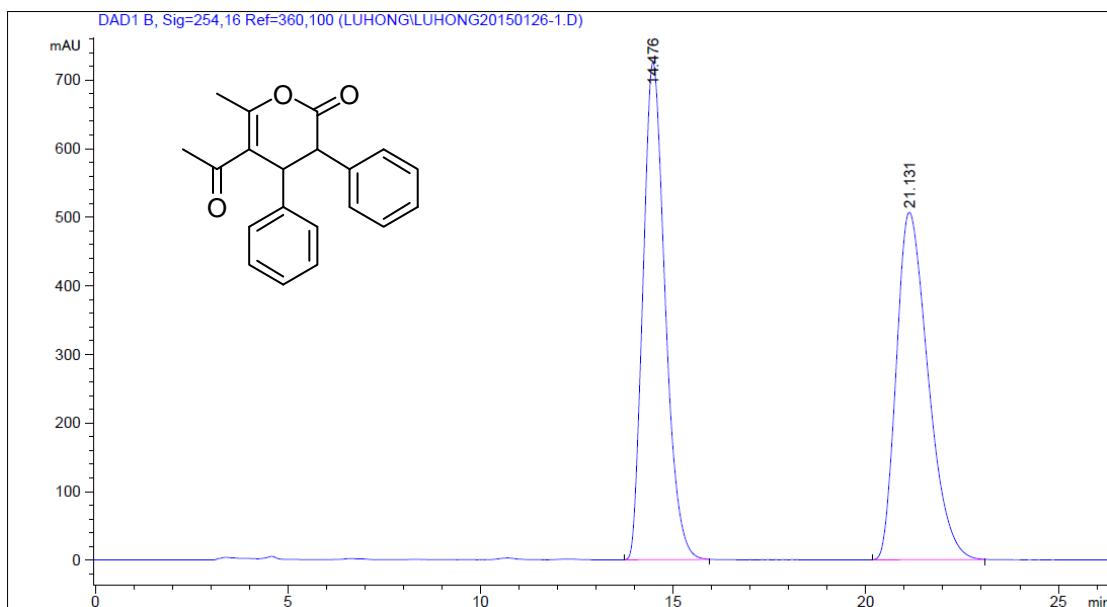


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	36.856	9330.18750	65.16678	49.7995
2	DAD 254, 16 nm	53.729	9405.30176	36.44764	50.2005

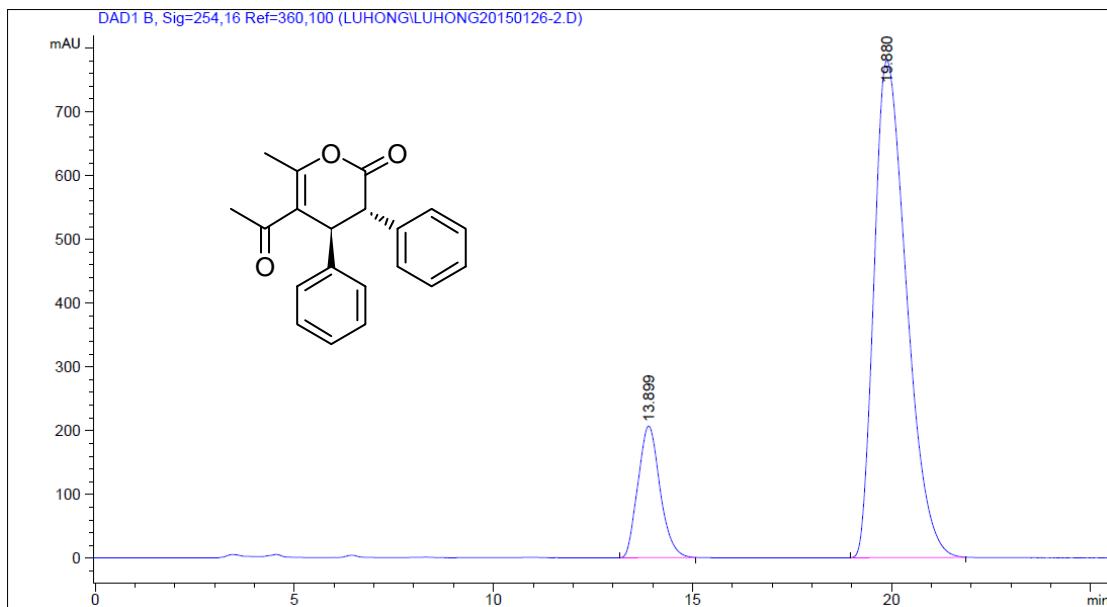


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	34.976	6.77280e4	414.29953	91.5319
2	DAD 254, 16 nm	54.463	6265.88574	27.00175	8.4681

3w

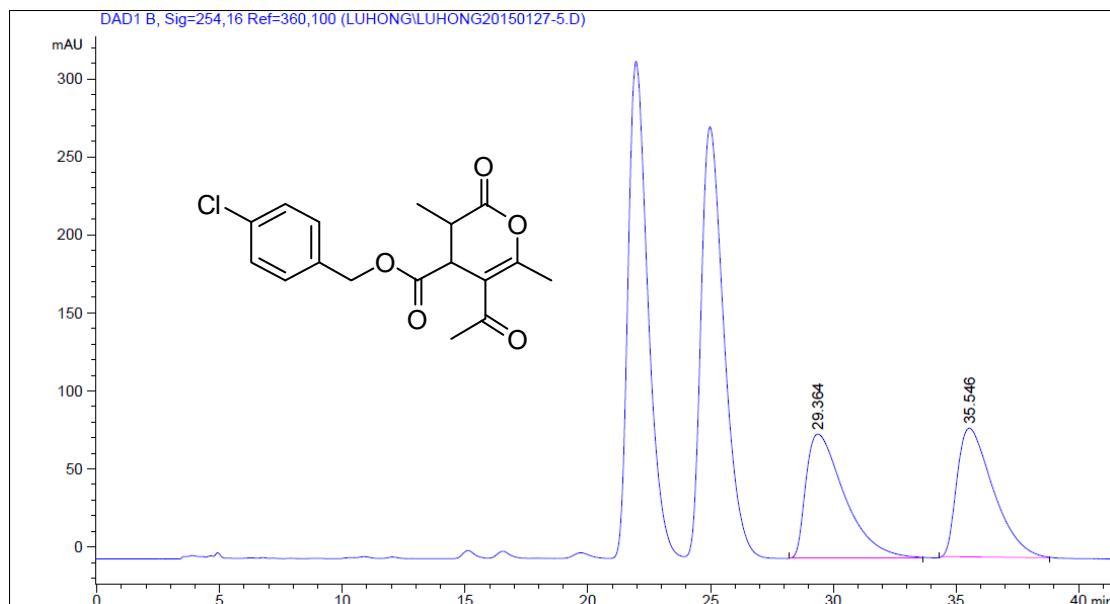


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	14.476	2.88310e4	725.20691	49.9210
2	DAD 254, 16 nm	21.131	2.89223e4	506.57614	50.0790

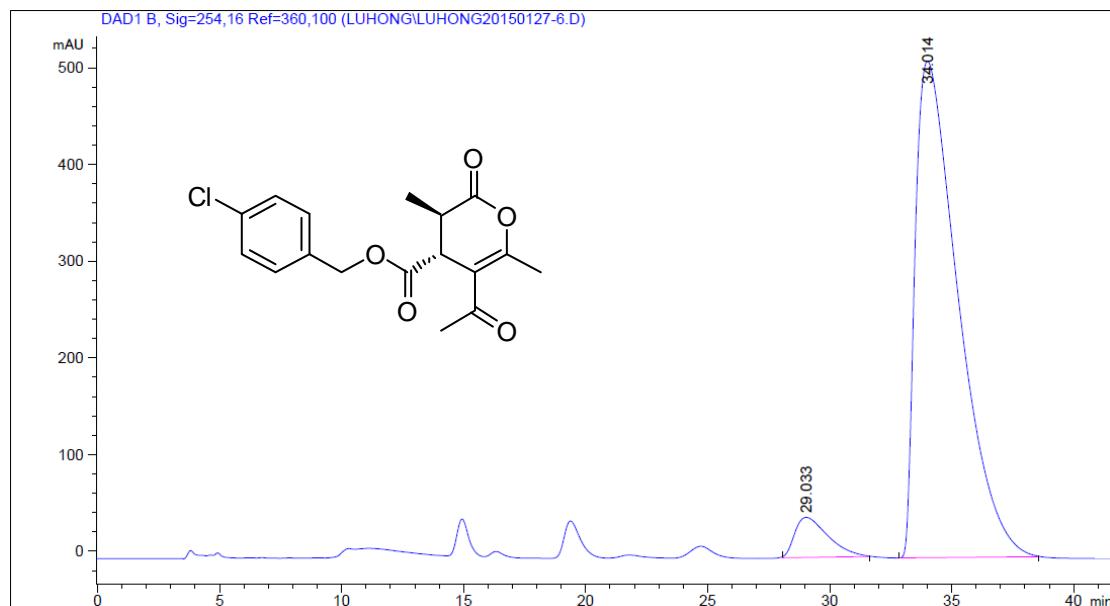


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	13.899	7955.07373	206.62357	15.2991
2	DAD 254, 16 nm	19.880	4.40420e4	779.88367	84.7009

3x

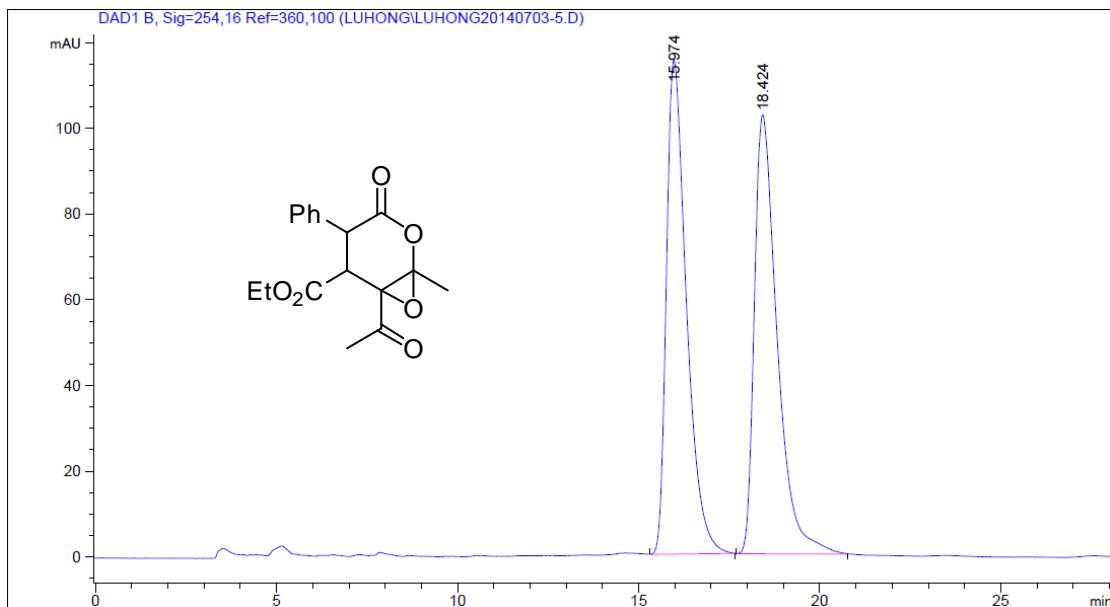


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	29.364	8643.55469	79.28326	50.5026
2	DAD 254, 16 nm	35.546	8471.50195	82.48608	49.4974

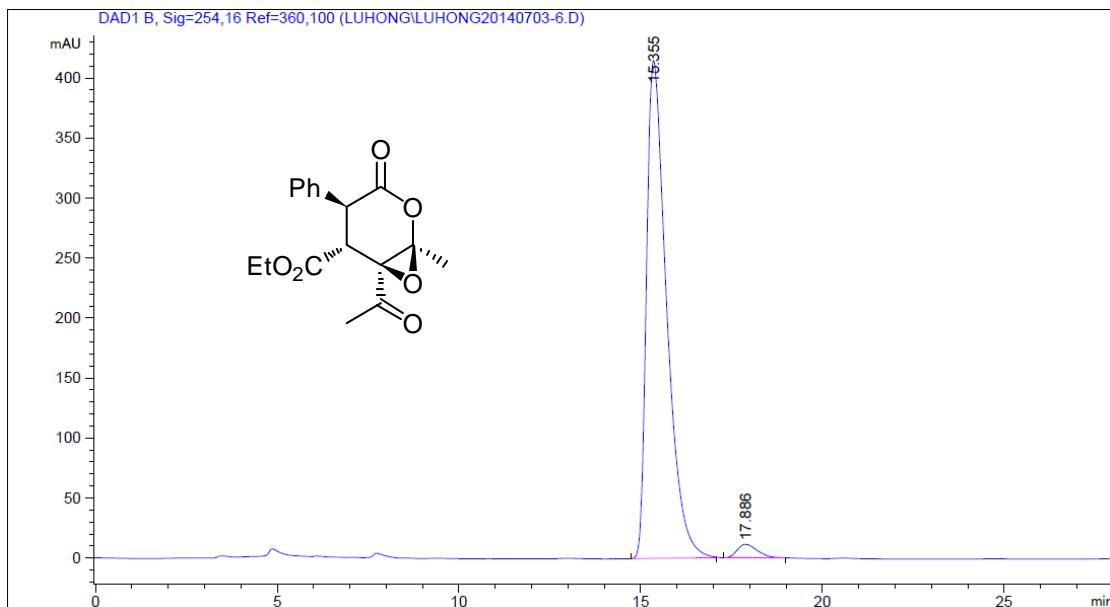


Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	29.033	3923.86377	41.47171	5.8839
2	DAD 254, 16 nm	34.014	6.27641e4	513.21808	94.1161

6a



Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	15.974	4424.85010	115.26480	49.6742
2	DAD 254, 16 nm	18.424	4482.89453	102.39242	50.3258



Peak	Processed Channel	Retention Time (min)	Peak Area (mAU*s)	Peak Height (mAU)	Peak Area (%)
1	DAD 254, 16 nm	15.355	1.59502e4	414.93729	97.3614
2	DAD 254, 16 nm	17.886	432.26294	11.25341	2.6386