

Electronic Supplementary Information (ESI)

Synthesis and evaluation of near-infrared probes with barbituric acid acceptors for in vivo detection of amyloid plaques

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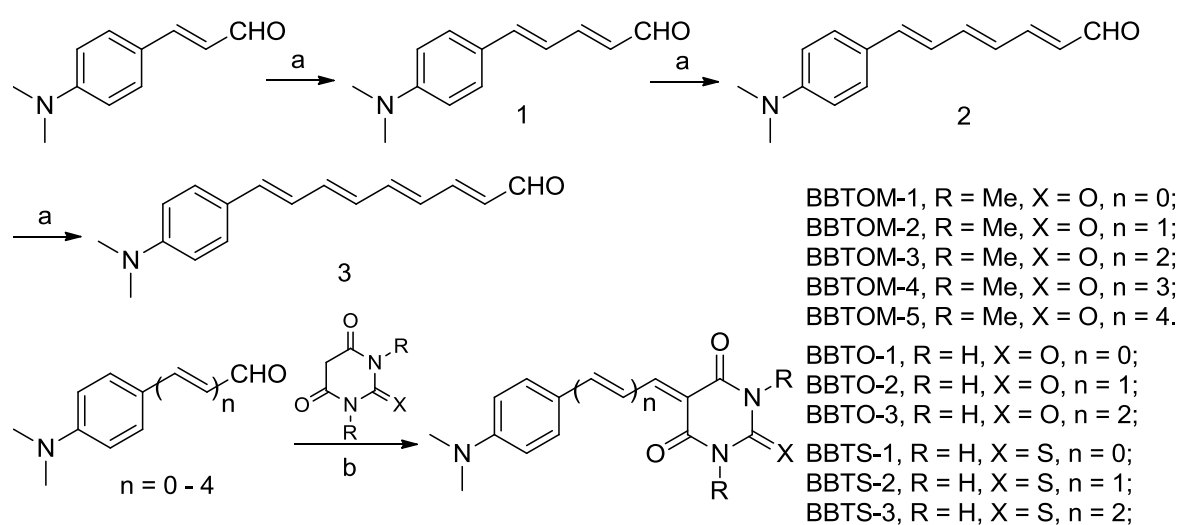
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General Procedures.

All solvents and chemicals were purchased from commercial products and used without further purification unless otherwise stated. Reactions were monitored by thin layer chromatography: Silica gel 60 F₂₅₄ plates (Merck). Flash column chromatography was manipulated using silica gel (45 - 75 μm , Yantai Industry Research Institute). Synthetic A β ₁₋₄₂ and A β ₁₋₄₀ were purchased from PEPTIDE INSTITUTE, Inc. (Japan), and the aggregation was performed using the procedures reported previously¹ for *in vitro* assays. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker Avance III (400 MHz or 100 MHz) NMR spectrometer in CDCl₃ or DMSO-*d*₆ solutions at room temperature. Chemical shift (δ) is reported in ppm downfield from tetramethylsilane and coupling constants (*J*) are reported in Hertz (Hz), and the multiplicity is defined by s (singlet), d (doublet), t (triplet), or m (multiplet). MS and HRMS spectra were obtained by the Surveyor MSQ Plus (ESI) (Waltham, MA, USA) instrument. Fluorescence spectra were measured on a SHIMADZU RF-5301PC spectrofluorophotometer (Japan). Rhodamine 6G was used as a standard ($\Phi = 0.76$ in H₂O)² to measure fluorescence quantum yields of all of the samples. UV-visible spectra were carried out on the SHIMADZU UV-3600 UV-Vis spectrophotometer (Japan). HPLC was conducted on an Agilent 1260 Infinity Quaternary LC (Agilent Technologies) system. The sample was analyzed on a Venusil MP C18 reverse column (Agela Technologies, 5 μm , 4.6 mm \times 250 mm) eluted with an isocratic system at a flow rate of 1.0 mL/min. The mobile phase A was water while B was acetonitrile. The purity of the compounds were analyzed by HPLC, and were found to be more than 95%. Fluorescence observation was performed by the Axio Observer Z1 inverted fluorescence microscope (Zeiss, Germany) equipped with DAPI, AF488, AF546 and CY5 filter sets. Male ICR

mice (5 weeks, 18-22 g) used for brain uptake studies were purchased from Vital River Laboratories (China). Double transgenic mice (C57BL6, APPsw/PSEN1, 22-month old) used as an Alzheimer's model and age matched control mice (C57BL6, 22-month old) were purchased from the Institute of Laboratory Animal Sciences, Chinese Academy of Medical Sciences. Postmortem brain tissues from autopsy confirmed cases of AD patients (91-year old, male; 68-year old, female; 64-year old, female; 70-year old, male; temporal lobe) were obtained from the Chinese Brain Bank Center (CBBC). All protocols requiring the use of animals were approved by the animal care committee of Beijing Normal University.

Chemistry.



Scheme S1. Reagents and conditions: (a) (1) ((1,3-Dioxolan-2-yl)methyl)triphenylphosphonium bromide, anhydrous THF, 18-crown-6, NaH, r.t., 12 h; (2) 1 M HCl, r.t., 30 min. (b) For BBTO 1-3 and BBTS 1-3, DMF, reflux; for BBTOM-1 and BBTOM-2, piperidine, EtOH, r.t., 15 min; for BBTOM-3, BBTOM-4 and BBTOM-5, K₂CO₃, EtOH, reflux.

(2E,4E)-5-(4-(Dimethylamino)phenyl)penta-2,4-dienal (1)

(E)-3-(4-(dimethylamino)phenyl)acrylaldehyde (371.0 mg, 2.1 mmol) was completely dissolved by anhydrous THF (20 mL) in a 100 mL round bottom flask charged a stirrer, the solution was added with ((1,3-Dioxolan-2-yl)methyl)triphenylphosphonium bromide (1.37 g, 3.2 mmol), NaH (254.0 mg, 10.6 mmol) and 18-crown-6 (50.0 mg) successively. The reaction mixture was stirred at room temperature overnight. Then 20 mL of aqueous HCl (1 M) was added to quench the reaction, the mixture was stirred for 30 min and neutralized with ammonia water then extracted with CH₂Cl₂ (3 × 50 mL). The combined organic extracts were dried over anhydrous Na₂SO₄. The solvents were evaporated under vacuum and the crude product was purified by flash column chromatography (petroleum ether/dichloromethane = 1 : 3, v/v) to yield compound **1** as yellow crystalline solids (250.3 mg, 58.6%). ¹H NMR (400 MHz, CDCl₃) δ: 9.56 (d, *J* = 8.1 Hz, 1H), 7.41 (d, *J* = 8.9 Hz, 2H), 7.32 - 7.21 (m, 1H), 6.95 (d, *J* = 15.3 Hz, 1H), 6.82 (dd, *J* = 15.3, 10.9 Hz, 1H), 6.68 (d, *J* = 8.8 Hz, 2H), 6.18 (dd, *J* = 15.0, 8.1 Hz, 1H), 3.03 (s, 6H).

(2E,4E,6E)-7-(4-(Dimethylamino)phenyl)hepta-2,4,6-trienal (2)

The same reaction as described above to prepare compound **1** was used, and the crude product was purified by flash column chromatography (petroleum ether/dichloromethane = 1 : 3, v/v). Compound **2** was obtained as dark red crystalline solids (116.8 mg, 41.4%). ¹H NMR (400 MHz, CDCl₃) δ: 9.56 (d, *J* = 8.1 Hz, 1H), 7.35 (d, *J* = 8.8 Hz, 2H), 7.18 (dd, *J* = 15.0, 11.3 Hz, 1H), 6.86 - 6.80 (m, 1H), 6.78 - 6.72 (m, 2H), 6.68 (d, *J* = 8.6 Hz, 2H), 6.47 (dd, *J* = 15.0, 8.1 Hz, 1H), 6.14 (dd, *J* = 15.0, 8.0 Hz, 1H), 3.00 (s, 6H). MS: *m/z* calcd for C₁₅H₁₈NO 228.1; found 228.1, M + H⁺.

(2E,4E,6E,8E)-9-(4-(dimethylamino)phenyl)nona-2,4,6,8-tetraenal (3)

The same reaction as described above to prepare compound **1** was used, and the crude product was

purified by flash column chromatography (petroleum ether/dichloromethane = 1 : 3, v/v). Compound **3** was obtained as purple crystalline solids (30.0 mg, 35.9%). ¹H NMR (400 MHz, CDCl₃) δ: 9.56 (d, *J* = 8.1 Hz, 1H), 7.35 (d, *J* = 8.4 Hz, 2H), 7.16 (dd, *J* = 15.0, 11.3 Hz, 1H), 6.79 - 6.61 (m, 6H), 6.48-6.35 (m, 2H), 6.14 (dd, *J* = 15.0, 8.0 Hz, 1H), 3.01 (s, 6H). MS: *m/z* calcd for C₁₇H₂₀NO 254.1; found 254.1, M + H⁺.

5-(4-(Dimethylamino)benzylidene)-1,3-dimethylpyrimidine-2,4,6(1H,3H,5H)-trione

(BBTOM-1)

4-(Dimethylamino)benzaldehyde (34.0 mg, 0.2 mmol) and 1,3-dimethylbarbituric acid (34.0 mg, 0.2 mmol) were dissolved in 10 mL ethanol. Piperidine (50 μL) was added as catalyst and the reaction mixture was stirred for 15 min at room temperature. The precipitate was collected by filtration, dried, and weighed. The final product was obtained as red crystal (43.2 mg, 72.3%). ¹H NMR (400 MHz, DMSO-*d*₆) δ: 8.41 (d, *J* = 9.2 Hz, 2H), 8.22 (s, 1H), 6.80 (d, *J* = 9.2 Hz, 2H), 3.21 (s, 6H), 3.13 (s, 6H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 163.2, 161.1, 156.3, 154.2, 151.2, 139.0, 120.0, 111.2, 109.3, 28.4, 27.8. MS: *m/z* calcd for C₁₅H₁₈N₃O₃ 288.1; found 287.9, M + H⁺.

(E)-5-(3-(4-(Dimethylamino)phenyl)allylidene)-1,3-dimethylpyrimidine-2,4,6(1H,3H,5H)-trione

(BBTOM-2)

The same reaction as described above to prepare BBTOM-1 was used, and the final product was obtained as purple crystalline solids (56.8 mg, 46.2%). ¹H NMR (400 MHz, CDCl₃) δ: 8.42 (dd, *J* = 14.8, 12.5 Hz, 1H), 8.20 (d, *J* = 12.4 Hz, 1H), 7.60 (d, *J* = 8.9 Hz, 2H), 7.40 (d, *J* = 14.8 Hz, 1H), 6.69 (d, *J* = 8.9 Hz, 2H), 3.38 (s, 6H), 3.10 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ: 163.0, 162.4,

158.6, 157.1, 153.0, 151.8, 132.1, 123.6, 120.6, 112.0, 110.0, 40.2, 28.5, 27.8. MS: m/z calcd for (M + H⁺) 314.2; found 314.0.

5-((2E,4E)-5-(4-(Dimethylamino)phenyl)penta-2,4-dien-1-ylidene)-1,3-dimethylpyrimidine-2,4,6(1H,3H,5H)-trione (BBTOM-3)

Compound 1 (30.2 mg, 0.2 mmol) and 1,3-dimethylbarbituric acid (45.3 mg, 0.3 mmol) were dissolved in 10 mL ethanol, 1 mg K₂CO₃ was added as catalyst. The reaction mixture was refluxed for 15 min, then the solvents were evaporated under vacuum and the crude product was purified by flash column chromatography (petroleum ether/dichloromethane = 1 : 2, v/v) and the final product was obtained as brown crystalline solids (74.3 mg, 53.2%). ¹H NMR (400 MHz, CDCl₃) δ: 8.11 (d, *J* = 12.5 Hz, 1H), 8.04 - 7.98 (m, 1H), 7.42 (d, *J* = 8.9 Hz, 2H), 7.31 - 7.28 (m, 1H), 7.00 - 6.92 (m, 2H), 6.69 (d, *J* = 8.8 Hz, 2H), 3.38 (s, 3H), 3.37 (s, 3H), 3.06 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ: 162.8, 162.0, 157.3, 151.8, 151.7, 145.9, 130.0, 126.9, 123.9, 123.8, 112.0, 111.1, 40.1, 28.5, 27.9. MS: m/z calcd for C₁₉H₂₂N₃O₃ 340.1661; found 340.1664, M + H⁺.

5-((2E,4E,6E)-7-(4-(Dimethylamino)phenyl)hepta-2,4,6-trien-1-ylidene)-1,3-dimethylpyrimidine-2,4,6(1H,3H,5H)-trione (BBTOM-4)

The same reaction as described above to prepare BBTOM-3 was used. The crude product was purified by flash column chromatography (petroleum ether/dichloromethane = 2 : 1, v/v) and the final produce was obtained as black crystalline solids (16.3 mg, 30.0%). ¹H NMR (400 MHz, CDCl₃) δ: 8.08 (d, *J* = 12.6 Hz, 1H), 8.00 - 7.93 (m, 1H), 7.38 (d, *J* = 8.9 Hz, 2H), 7.20 (dd, *J* = 14.0, 11.6 Hz, 1H), 6.94 - 6.87 (m, 1H), 6.81 (s, 1H), 6.79 (d, *J* = 5.4 Hz, 1H), 6.67 (d, *J* = 8.9 Hz, 2H), 6.58 (dd, *J*

= 14.0, 11.6 Hz, 1H), 3.37 (s, 3H), 3.36 (s, 3H), 3.03 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ: 162.7, 162.0, 157.0, 156.2, 151.7, 151.2, 150.2, 146.6, 141.1, 130.0, 129.0, 128.4, 127.8, 126.1, 124.5, 124.0, 119.2, 112.1, 111.7, 40.2, 28.6, 28.0. MS: m/z calcd for C₂₁H₂₄N₃O₃ 366.1818; found 366.1818, M + H⁺.

5-((2E,4E,6E,8E)-9-(4-(Dimethylamino)phenyl)nona-2,4,6,8-tetraen-1-ylidene)-1,3-dimethylpyrimidine-2,4,6(1H,3H,5H)-trione (BBTOM-5)

The same reaction as described above to prepare BBTOM-3 was used. The crude product was purified by flash column chromatography (petroleum ether/dichloromethane = 1 : 3, v/v) and the final produce was obtained as atropurpureus solid (21.0 mg, 89.4%). ¹H NMR (400 MHz, DMSO-*d*₆) δ: 7.96 (d, *J* = 12.4 Hz, 1H), 7.90 - 7.83 (m, 1H), 7.49 - 7.43 (m, 1H), 7.38 (d, *J* = 8.8 Hz, 2H), 7.00 - 6.77 (m, 4H), 6.74 - 6.64 (m, 3H), 6.57 - 6.50 (m, 1H), 3.19 (s, 3H), 3.18 (s, 3H), 2.96 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ: 162.6, 161.9, 156.7, 155.5, 151.6, 150.7, 145.5, 141.9, 138.2, 131.0, 130.2, 128.5, 128.3, 125.0, 124.4, 112.2, 112.1, 40.2, 28.6, 28.0. MS: m/z calcd for C₂₃H₂₆N₃O₃ 392.1974; found 393.1968, M + H⁺.

5-(4-(Dimethylamino)benzylidene)pyrimidine-2,4,6(1H,3H,5H)-trione (BBTO-1)

4-(Dimethylamino)benzaldehyde (33.6 mg, 0.2 mmol) and barbituric acid (28.8 mg, 0.2 mmol) were dissolved in 3 mL DMF. The reaction mixture was refluxed for 10 min, then cooled to precipitate BBTO-1 as red crystalline solids (17.2 mg, 30.1%). ¹H NMR (400 MHz, DMSO-*d*₆) δ: 8.42 (d, *J* = 9.2 Hz, 2H), 8.15 (s, 1H), 7.80 (d, *J* = 9.2 Hz, 2H), 3.12 (s, 6H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 164.6, 162.7, 155.4, 154.1, 150.3, 139.0, 120.0, 111.1, 109.5. MS: m/z

calcd for C₁₃H₁₃N₃O₃Na 282.1; found 282.1, M + Na⁺.

(E)-5-(3-(4-(Dimethylamino)phenyl)allylidene)pyrimidine-2,4,6-(1H,3H,5H)-trione (BBTO-2)

The same reaction as described above to prepare BBTO-1 was used, and the final product was obtained as atropurpureus crystalline solids (53.0 mg, 80.1%). ¹H NMR (400 MHz, DMSO-*d*₆) δ: 8.22 (dd, *J* = 15.0, 12.4 Hz, 1H), 7.98 (d, *J* = 12.3 Hz, 1H), 7.62 (d, *J* = 15.0 Hz, 1H), 7.55 (d, *J* = 8.9 Hz, 2H), 6.80 (d, *J* = 8.9 Hz, 2H), 3.06 (s, 6H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 163.6, 163.3, 156.0, 155.4, 152.8, 150.42, 131.5, 122.7, 119.0, 112.0, 110.4. MS: *m/z* calcd for C₁₅H₁₆N₃O₃ 286.1; found 285.9, M + H⁺.

5-((2E,4E)-5-(4-(Dimethylamino)phenyl)penta-2,4-dien-1-ylidene)pyrimidine-2,4,6(1H,3H,5H)-trione (BBTO-3)

The same reaction as described above to prepare BBTO-1 was used, and the final product was obtained as atropurpureus crystalline solids (40.0 mg, 64.4%). ¹H NMR (400 MHz, DMSO-*d*₆) δ: 7.93 - 7.82 (m, 2H), 7.53 (d, *J* = 8.9 Hz, 2H), 7.51 - 7.45 (m, 1H), 7.13 - 7.11 (m, 2H), 6.73 (d, *J* = 8.9 Hz, 2H), 3.01 (s, 6H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 163.6, 156.0, 152.8, 150.4, 150.4, 131.5, 130.0, 125.6, 123.7, 122.7, 119.0, 112.0, 112.0. MS: *m/z* calcd for C₁₇H₁₇N₃O₃Na 334.1; found 334.1, M + Na⁺.

5-(4-(Dimethylamino)benzylidene)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione (BBTS-1)

The same reaction as described above to prepare BBTO-1 was used, and the final product was obtained as crimson crystalline solids (13.4 mg, 21.2%). ¹H NMR (400 MHz, DMSO-*d*₆) δ: 12.12 (s,

1H), 12.02 (s, 1H), 8.46 (d, $J = 9.1$ Hz, 2H), 8.15 (s, 1H), 6.82 (d, $J = 9.1$ Hz, 2H), 3.16 (s, 6H). ^{13}C NMR (100 MHz, DMSO- d_6) δ : 177.4, 162.9, 160.4, 156.1, 154.7, 139.6, 120.4, 111.4, 109.2. MS: m/z calcd for $\text{C}_{13}\text{H}_{14}\text{N}_3\text{O}_2\text{S}$ 276.1; found 275.8, $\text{M} + \text{H}^+$.

(*E*)-5-(3-(4-(Dimethylamino)phenyl)allylidene)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione (BBTS-2)

The same reaction as described above to prepare BBTO-1 was used, and the final product was obtained as purple crystalline solids (54.0 mg, 93.6%). ^1H NMR (400 MHz, DMSO- d_6) δ : 8.25 (dd, $J = 14.6, 12.6$ Hz, 1H), 8.00 (d, $J = 12.4$ Hz, 1H), 7.72 (d, $J = 14.8$ Hz, 1H), 7.58 (d, $J = 8.8$ Hz, 2H), 6.82 (d, $J = 8.8$ Hz, 2H), 3.09 (s, 6H). ^{13}C NMR (100 MHz, DMSO- d_6) δ : 177.8, 161.8, 161.2, 157.7, 156.4, 153.3, 132.2, 122.8, 119.3, 112.2, 110.0. MS: m/z calcd for $\text{C}_{15}\text{H}_{16}\text{N}_3\text{O}_2\text{S}$ 302.1; found 301.8, $\text{M} + \text{H}^+$.

5-((2*E*,4*E*)-5-(4-(Dimethylamino)phenyl)penta-2,4-dien-1-ylidene)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione (BBTS-3)

The same reaction as described above to prepare BBTO-1 was used. The final product was obtained as dark green crystalline solids (14.5 mg, 29.5%). ^1H NMR (400 MHz, DMSO- d_6) δ : 12.15 (s, 1H), 12.09 (s, 1H), 7.96 - 7.83 (m, 2H), 7.60 - 7.56 (m, 3H), 7.22 - 7.17 (M, 2H), 6.74 (d, $J = 8.8$ Hz, 2H), 3.03(s, 6H). ^{13}C NMR (100 MHz, DMSO- d_6) δ : 177.9, 161.7, 161.0, 158.6, 155.3, 151.9, 147.4, 130.5, 126.0, 123.8, 123.4, 112.0, 111.1. MS: m/z calcd for $\text{C}_{17}\text{H}_{18}\text{N}_3\text{O}_2\text{S}$ 327.1; found 328.0, $\text{M} + \text{H}^+$.

The Purity Determination.

The purity of the compounds was determined on a HPLC system eluted with acetonitrile/water at a flow rate of 1.0 mL/min. As shown in Fig. S1, the retention time increased with the lengthening of the conjugated double bonds and the purity of these probes was higher than 95%.

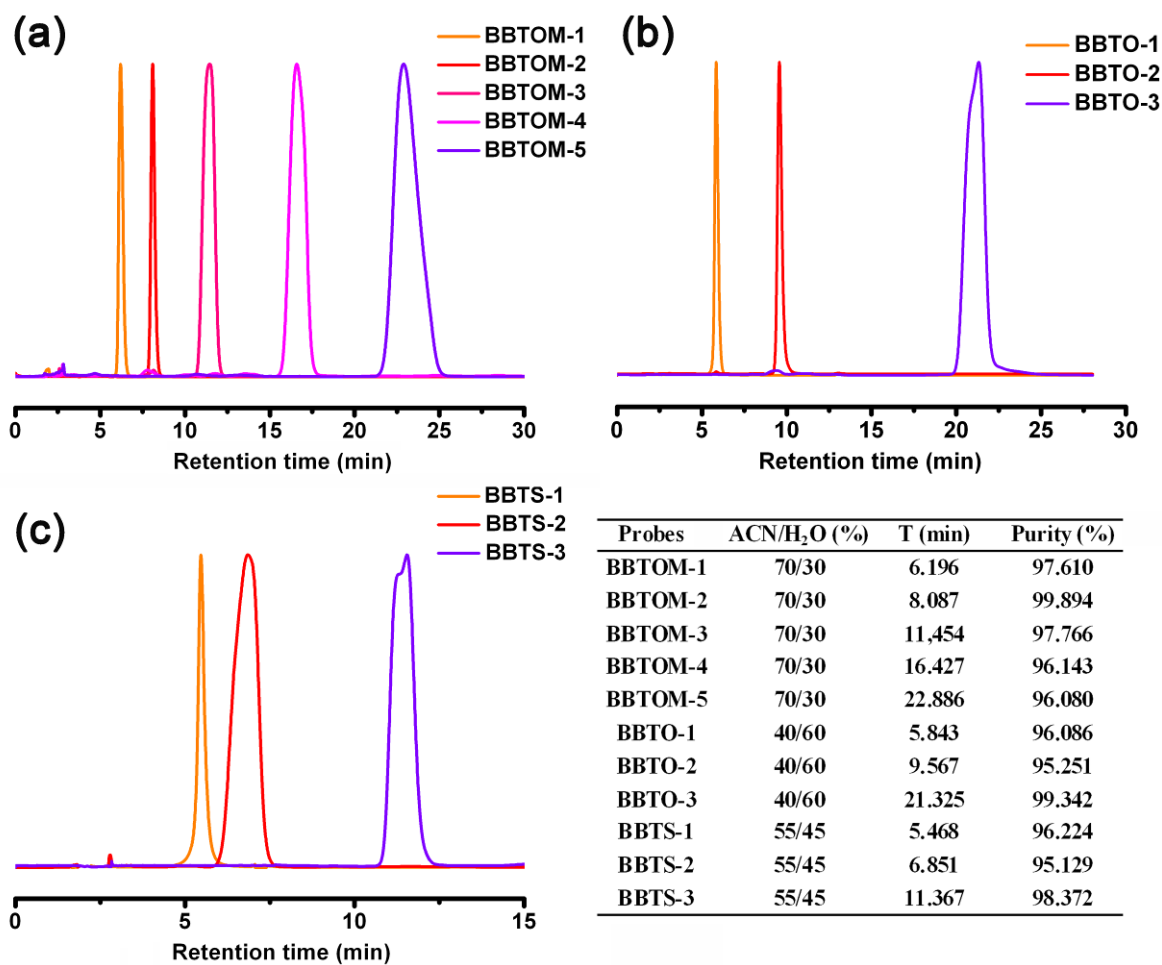
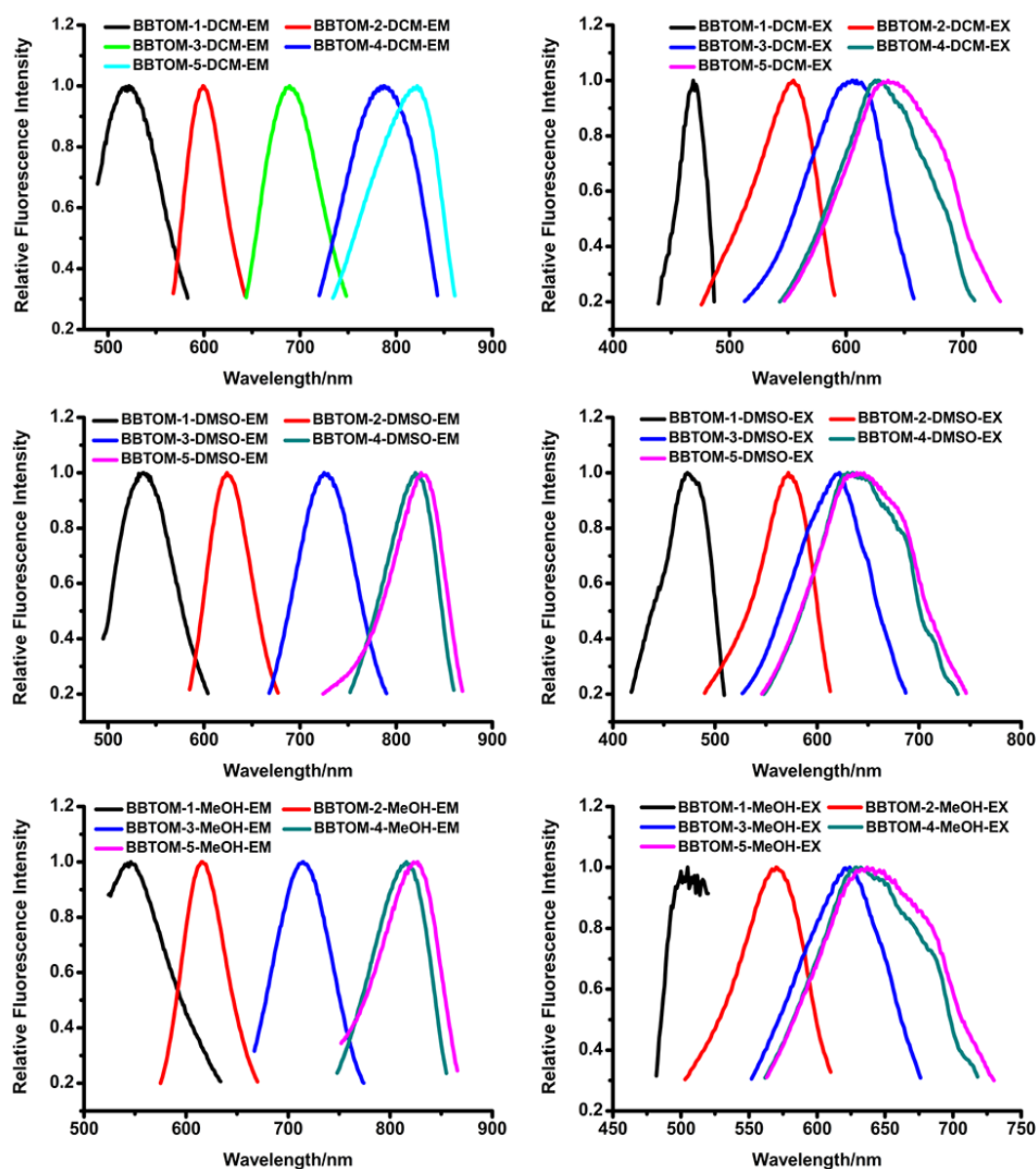


Figure. S1 HPLC profiles of BBTOMs (A), BBTOs (B), BBTSs (C) and HPLC conditions and purity (D).

Spectroscopic Determinations.

The fluorescence spectra were measured on a SHIMADZU RF-5301PC spectrofluorophotometer (Japan) in different solvents of CH₂Cl₂, DMSO, MeOH, THF and phosphate-buffered saline (PBS).

The absorption spectra are determined on the SHIMADZU UV-3600 UV-Vis spectrophotometer (Japan) in CH₂Cl₂. CH₂Cl₂ is a solvent that can simulate the hydrophobic microenvironment of the A β fibril “binding pocket”,³ then it was employed to determine the molar absorption coefficient and quantum yield. The spectroscopic data are shown in Figure S2-7 and Table S1, 2.



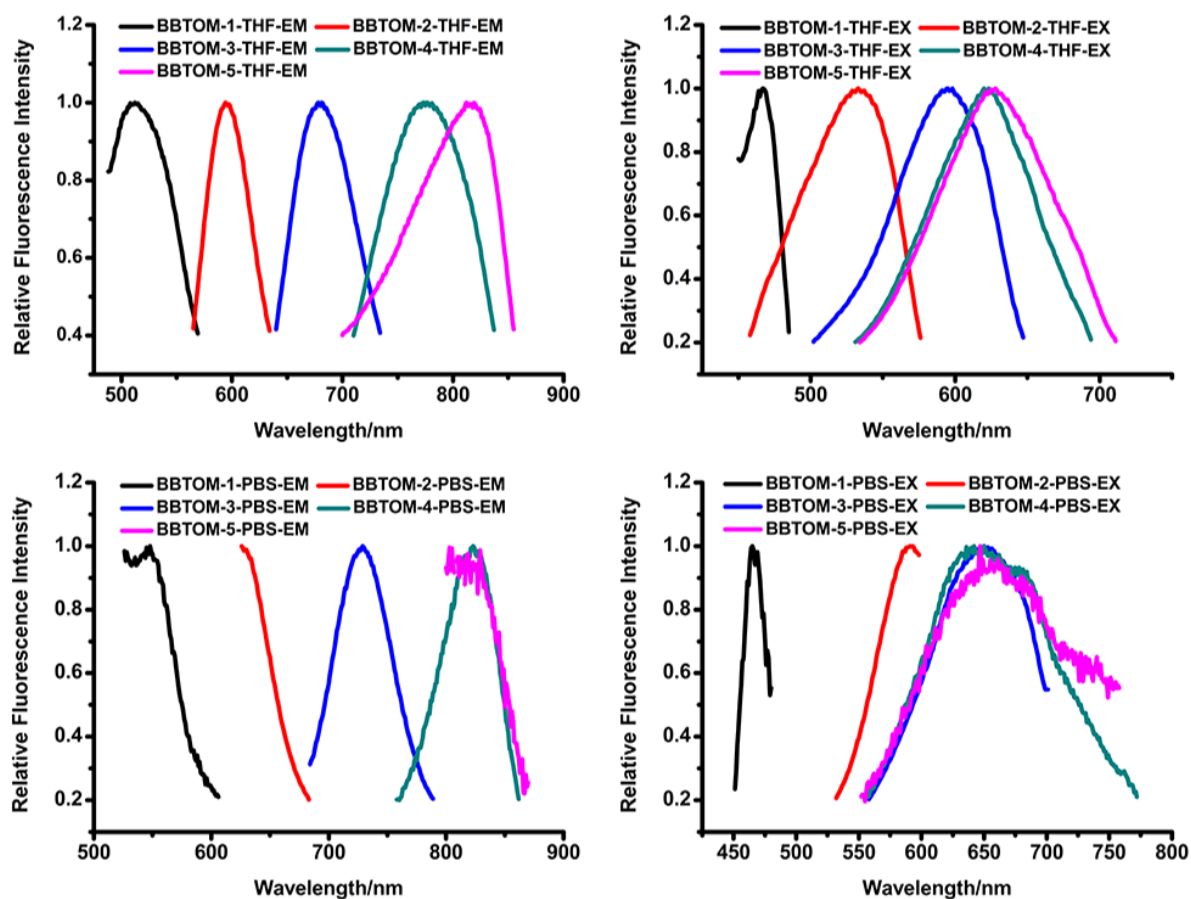
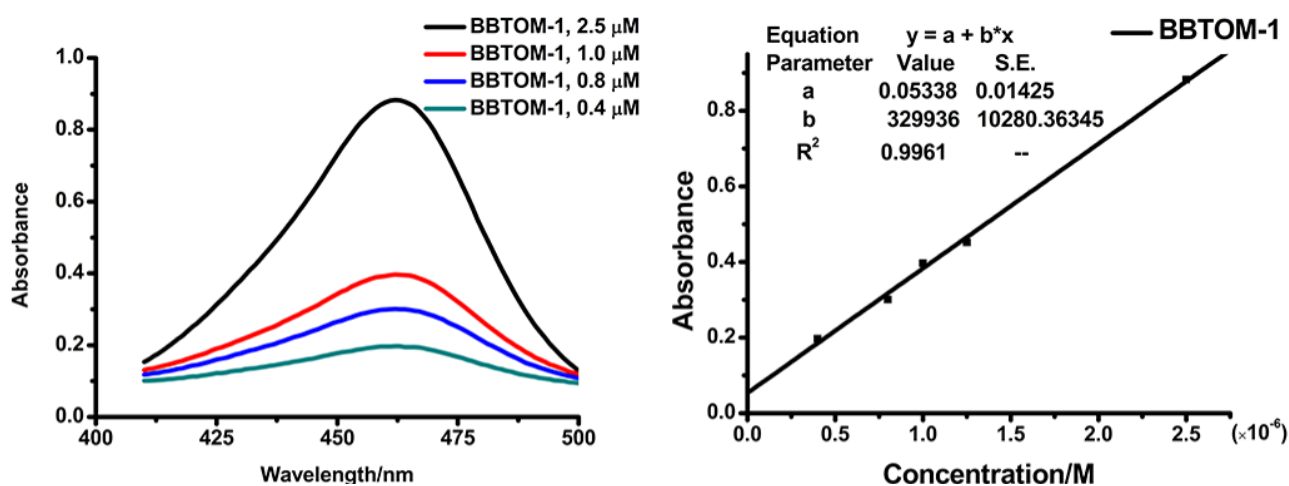


Figure. S2 The excitation (right panel) and emission (left panel) spectra of the BBTOM1-5 in different solvents: dichloromethane, DMSO, methanol, THF and PBS (10% ethanol). The emission wavelengths of the solutions were measured at a concentration of 10 μM .



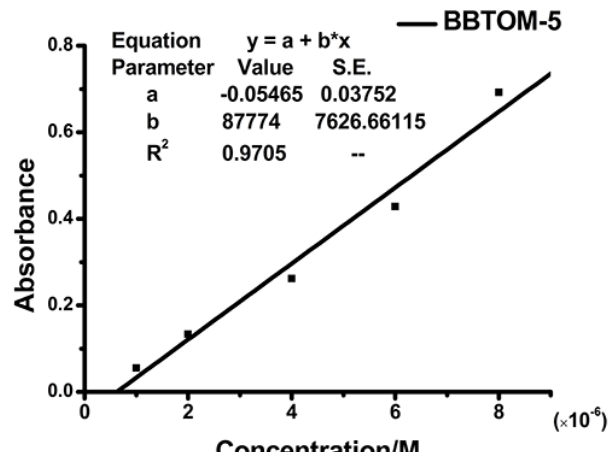
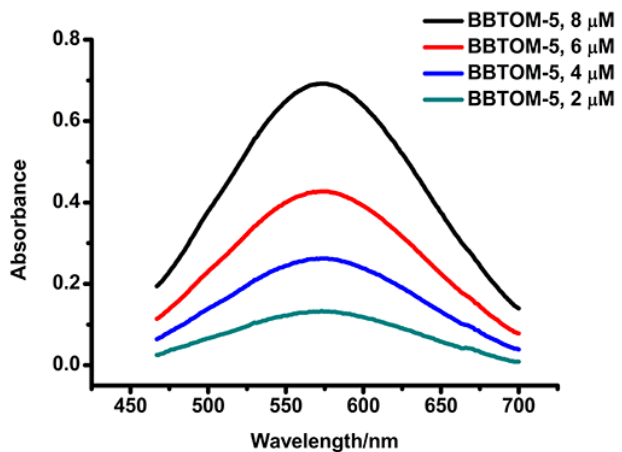
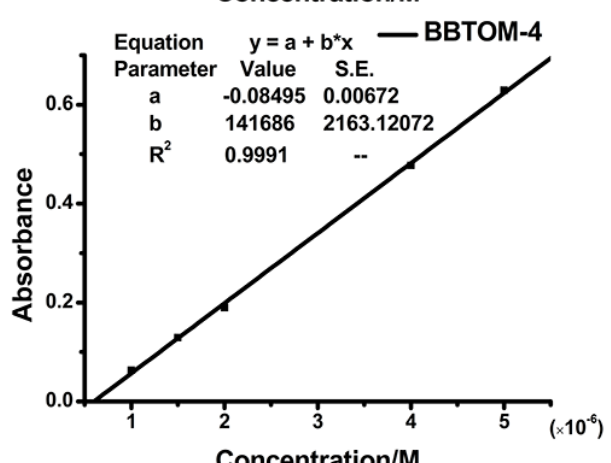
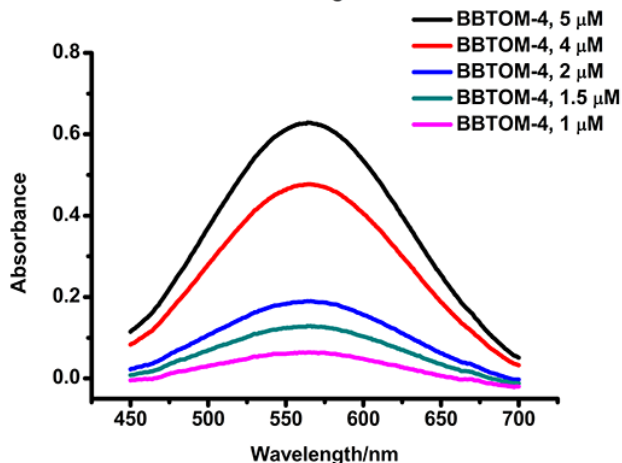
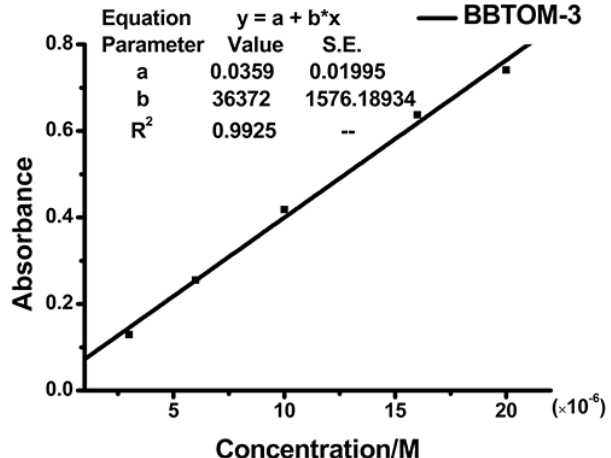
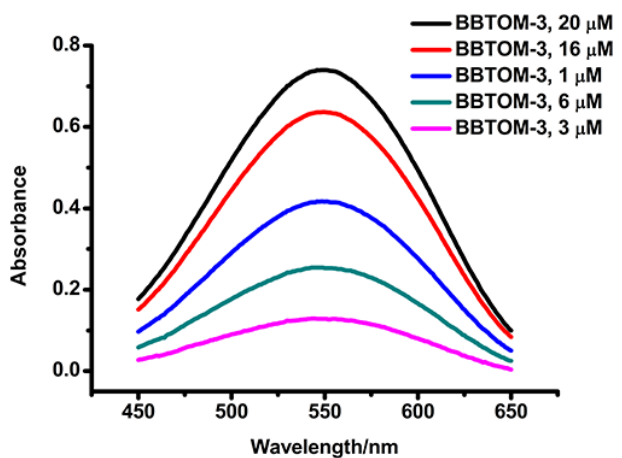
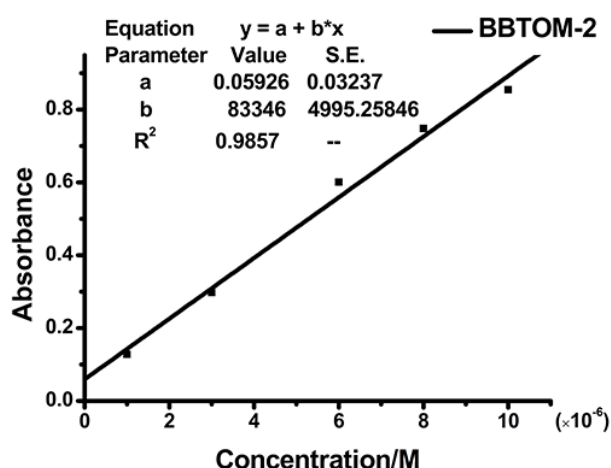
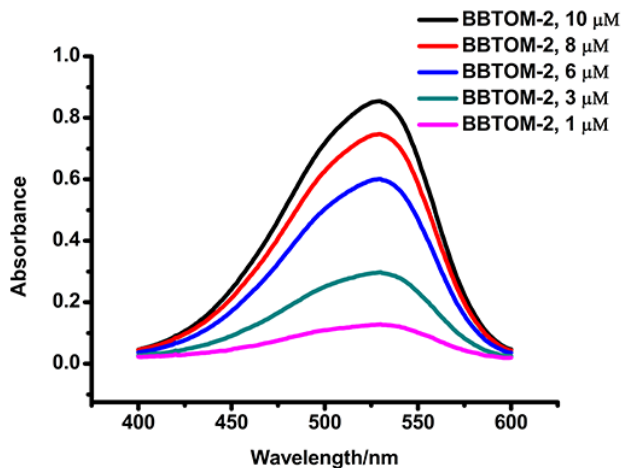
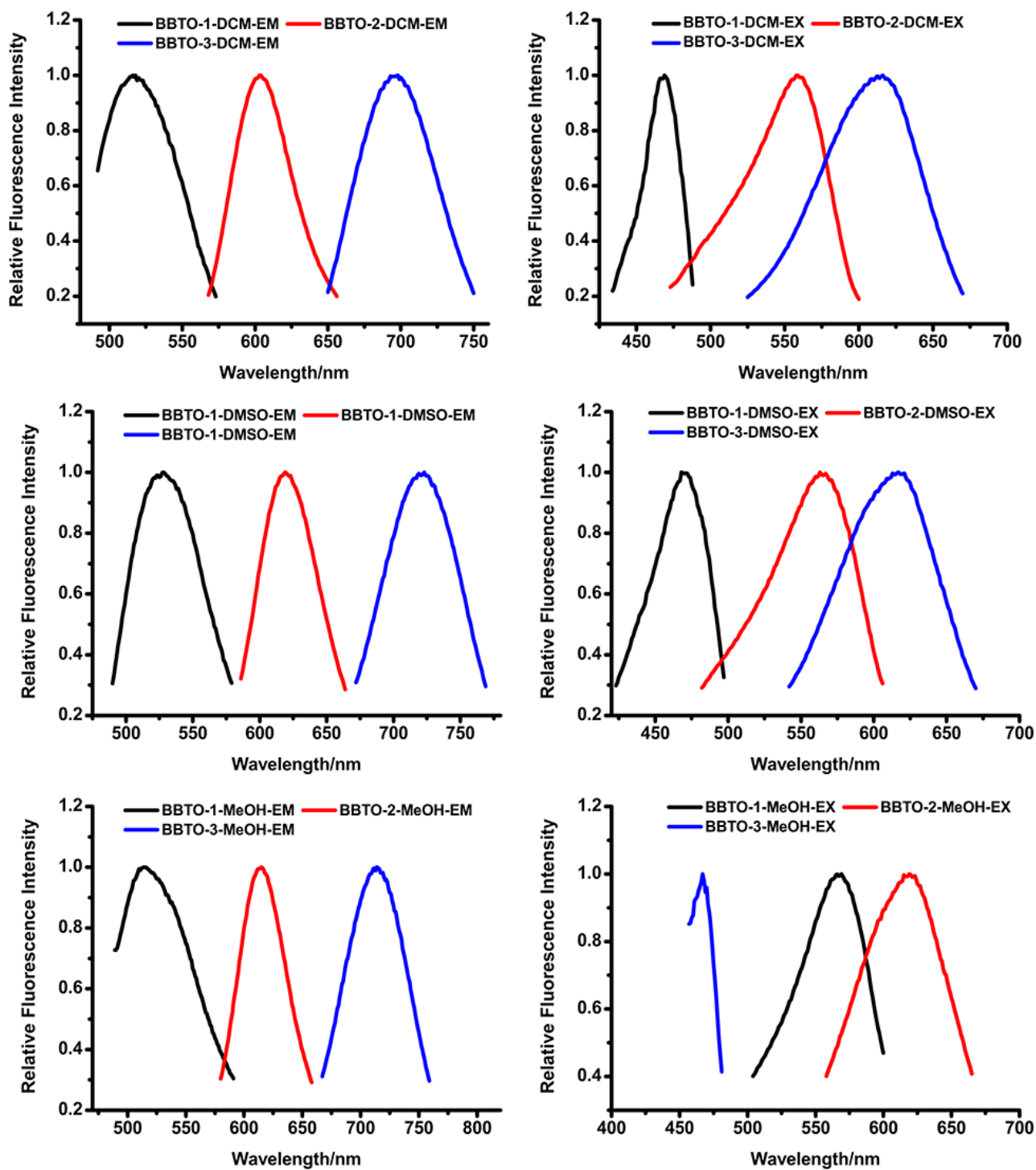


Figure. S3 Absorption spectra (left panel) and linear fitting curve between absorbance and concentration (right panel) of the BBTOM1-5 in CH₂Cl₂, molar absorption coefficients (ϵ) equals to
b.



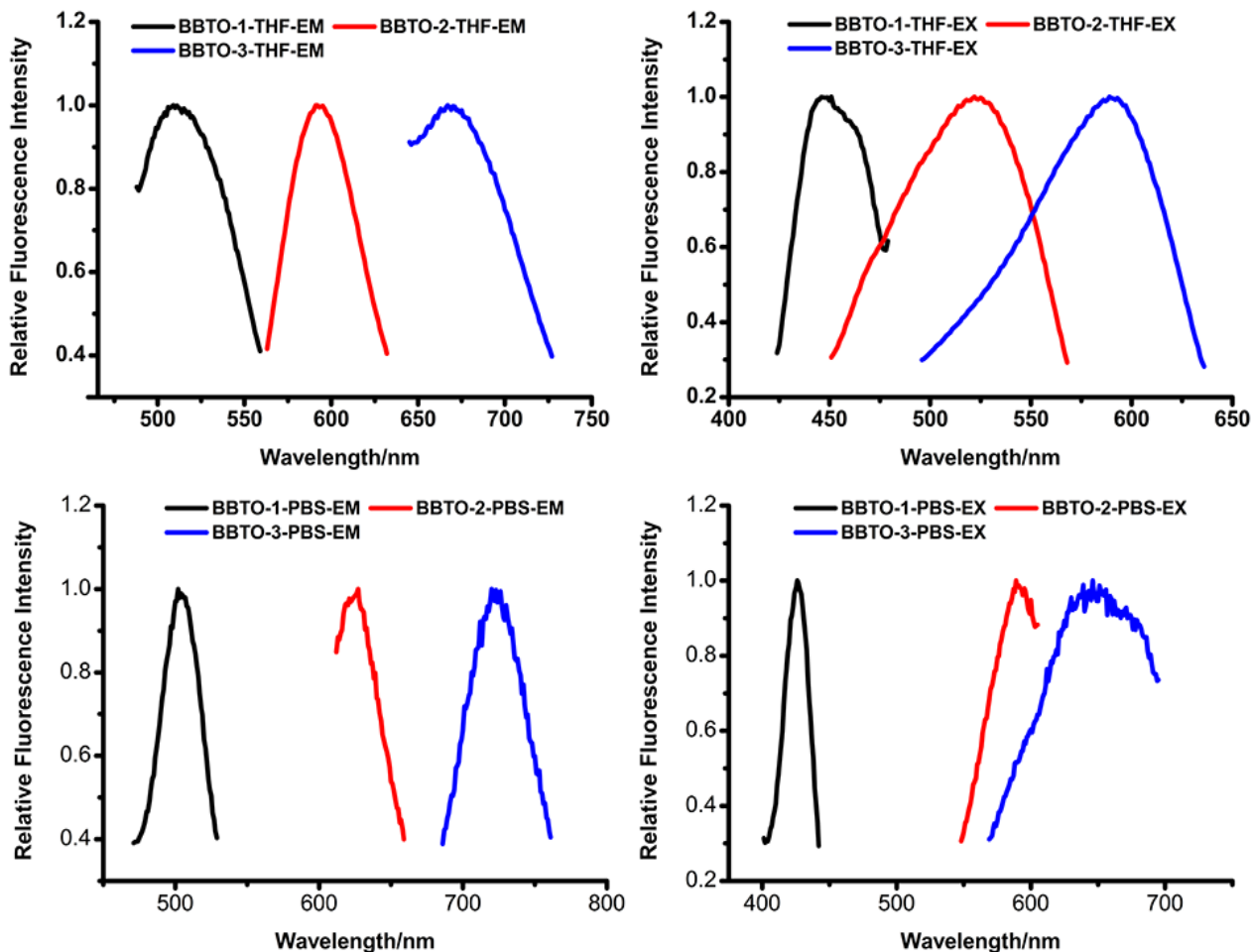
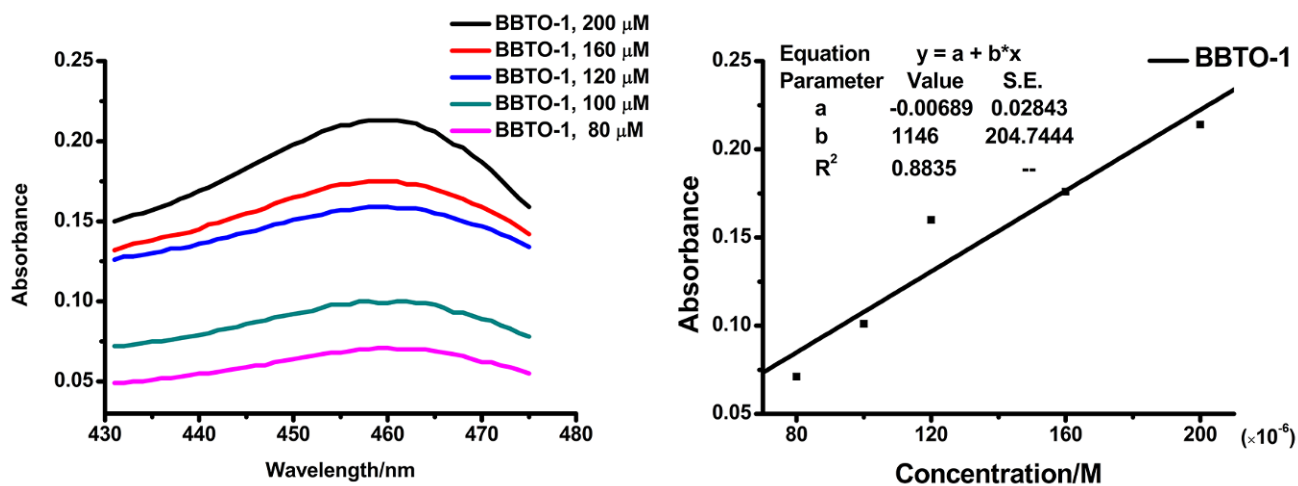


Figure. S4 The excitation (right panel) and emission (left panel) spectra of the BBTO1-3 in different solvents: dichloromethane, DMSO, methanol, THF and PBS (10% ethanol). The emission wavelengths of the solutions were measured at a concentration of 10 μM .



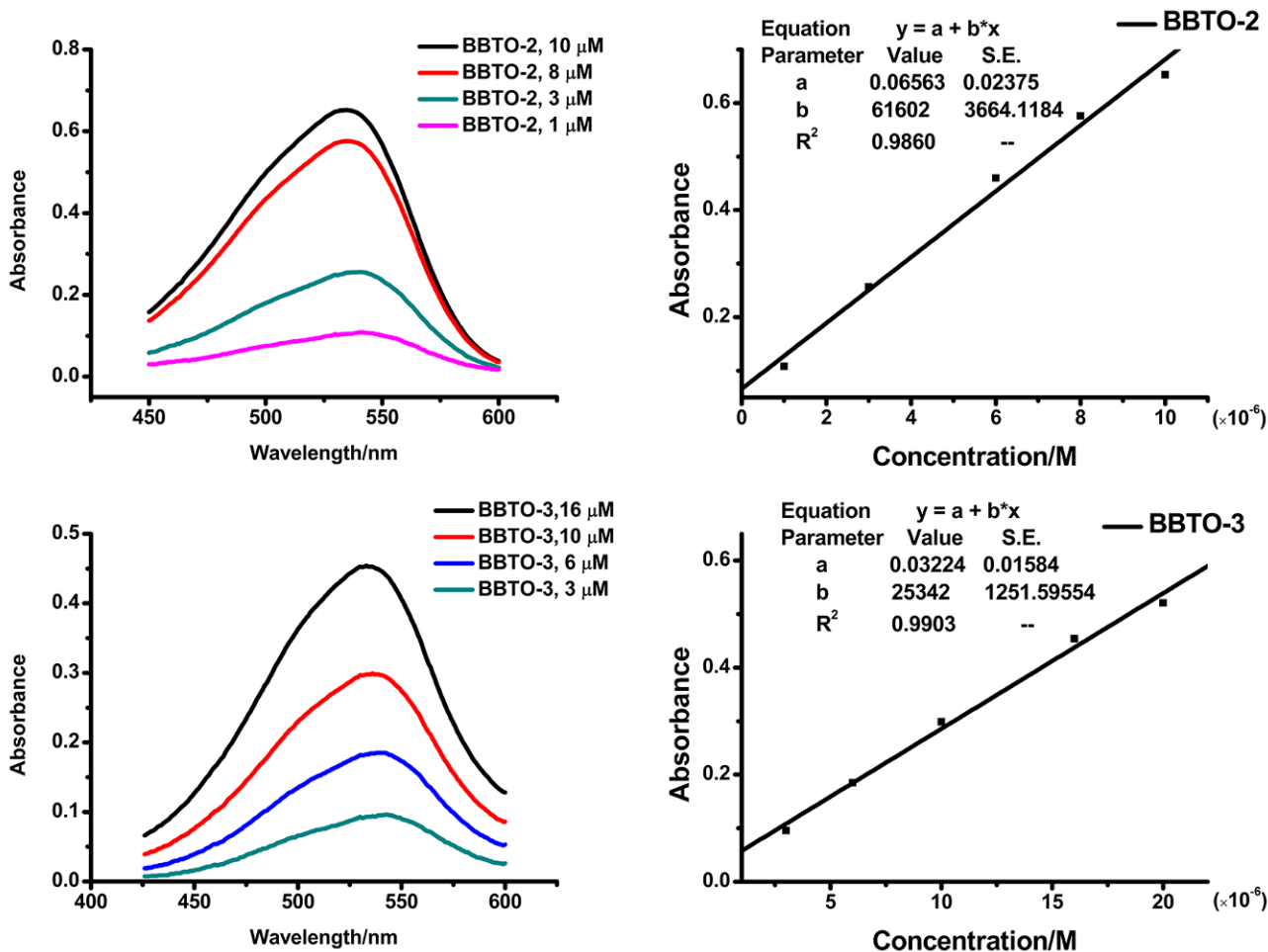
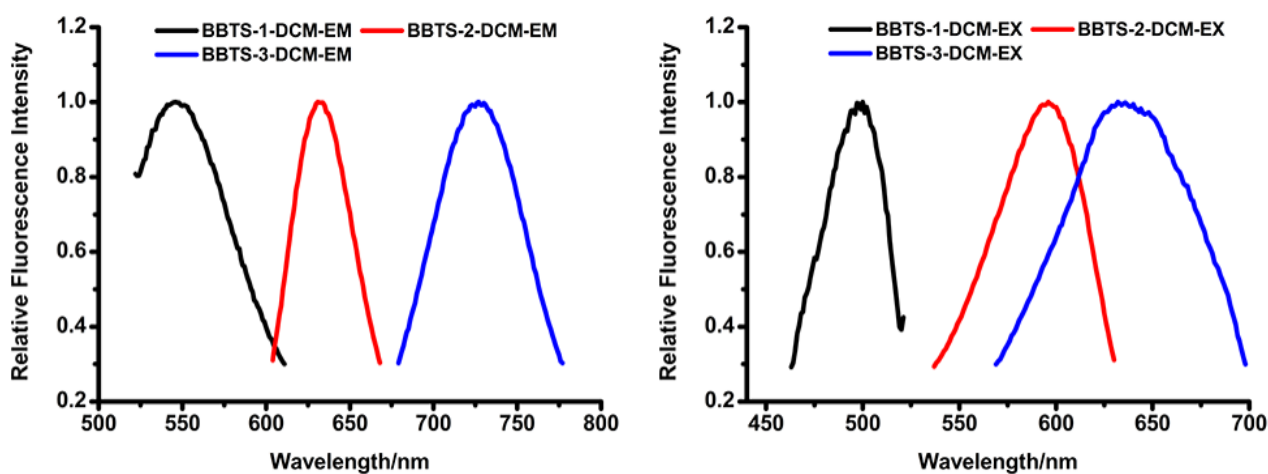


Figure. S5 Absorption spectra (left panel) and linear fitting curve between absorbance and concentration (right panel) of the BBTO1-3 in CH_2Cl_2 , molar absorption coefficients (ϵ) equals to b.



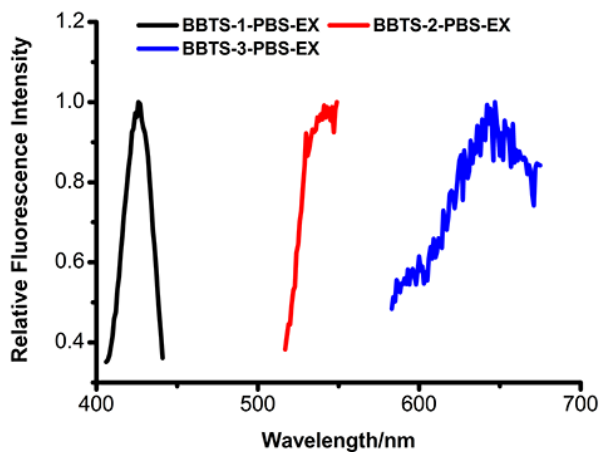
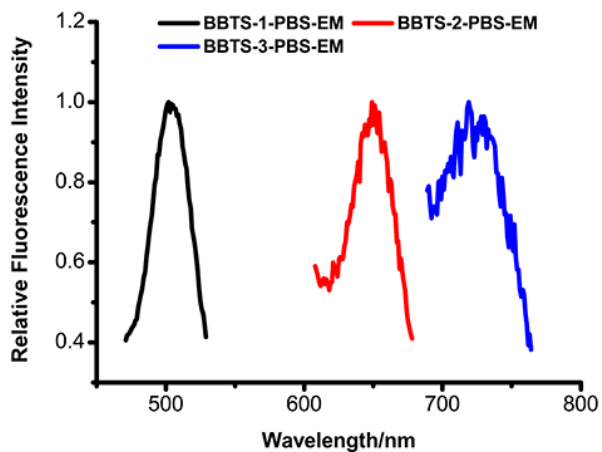
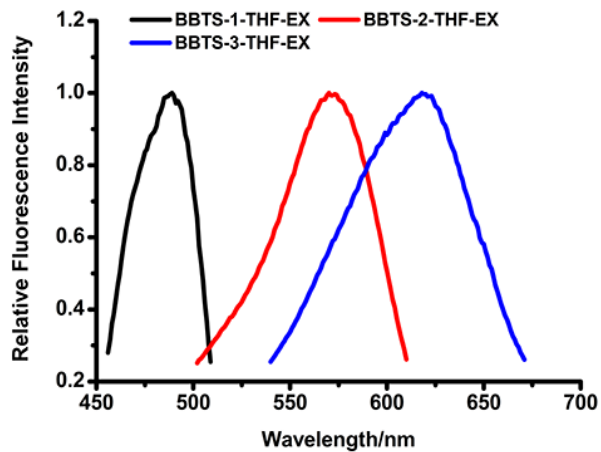
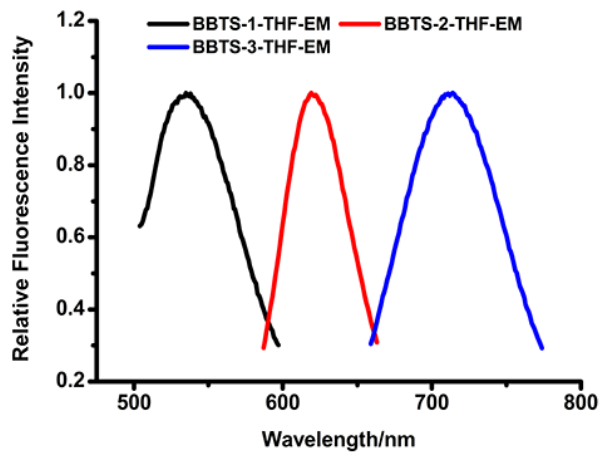
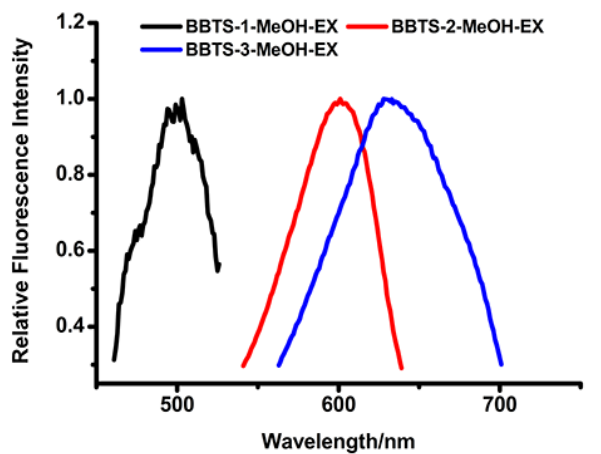
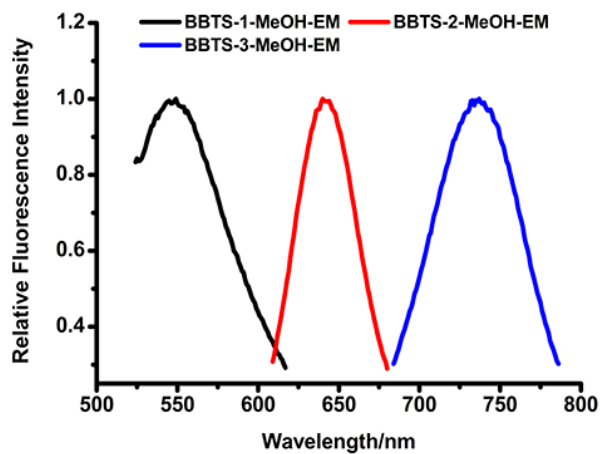
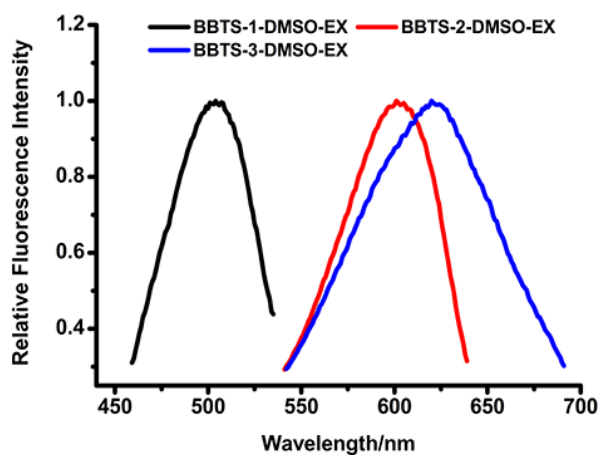
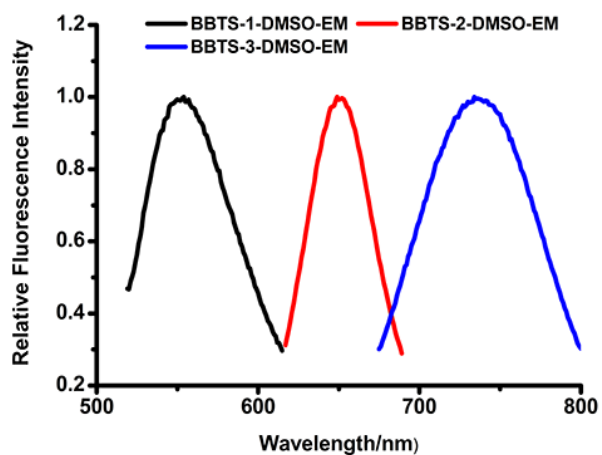


Figure. S6 The excitation (right panel) and emission (left panel) spectra of the BBTS1-3 in different solvents: dichloromethane, DMSO, methanol, THF and PBS (10% ethanol). The emission wavelengths of the solutions were measured at a concentration of 10 μM .

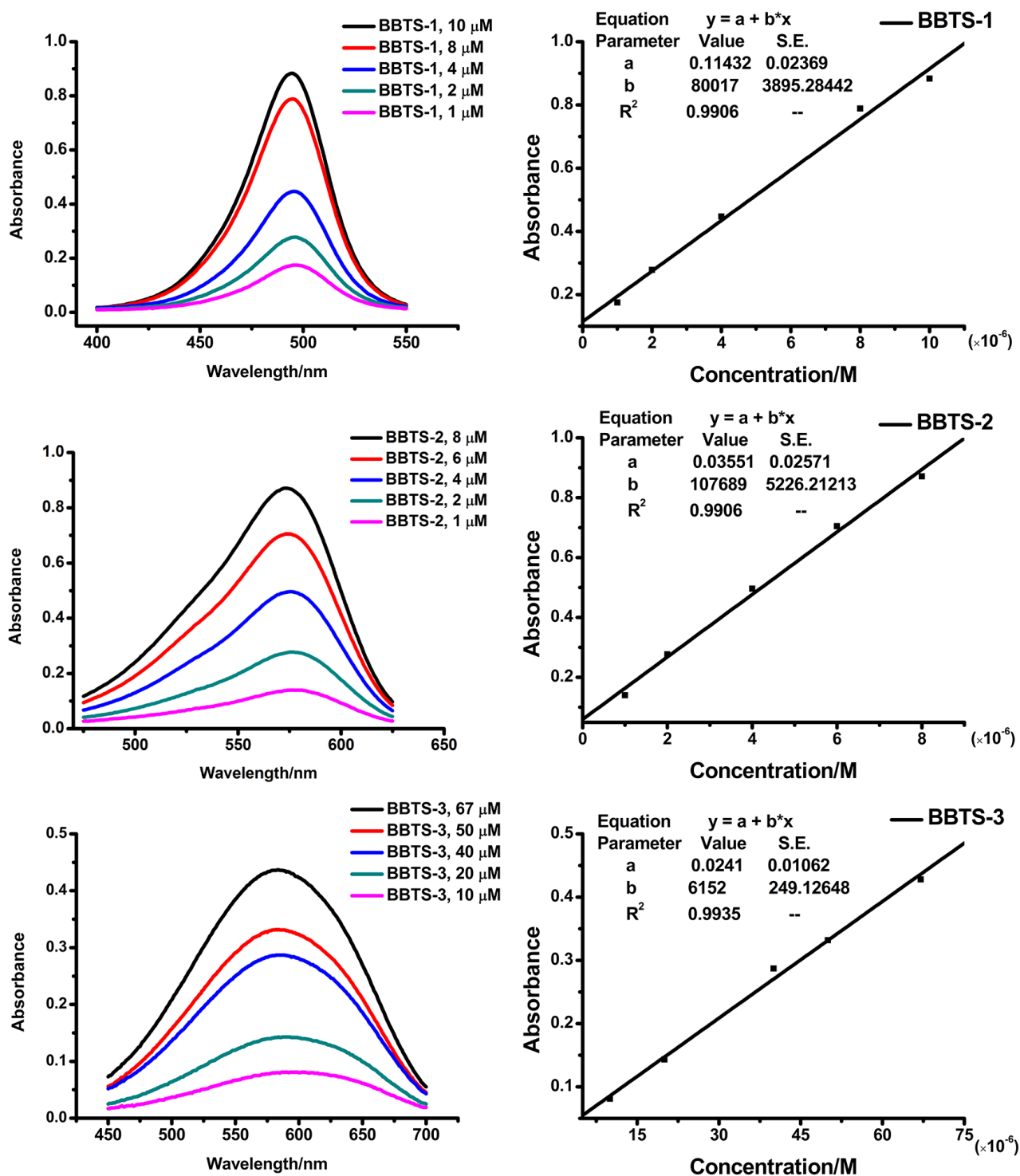


Figure. S7 Absorption spectra (left panel) and linear fitting curve between absorbance and

concentration (right panel) of the BBTS1-3 in CH₂Cl₂, molar absorption coefficients (ϵ) equals to b.

Table S1 The spectroscopic properties (absorption, excitation and emission wavelength, molar absorption coefficient, and fluorescence quantum yield) of BBTOM 1-5.

Probe	Solvent	Fluorescence spectrum			UV-vis spectrum		
		λ_{em} (nm)	λ_{ex} (nm)	Stokes shift (nm)	Φ^a %	λ_{abs} (nm)	ϵ L mol ⁻¹ cm ⁻¹
BBTOM-1	CH ₂ Cl ₂	522	468	54	0.08	461.5	329936
	DMSO	537	476	61			
	MeOH	546	505	41			
	THF	512	467	45			
	PBS	550	466	84			
BBTOM-2	CH ₂ Cl ₂	598	549	49	4.64	530	83346
	DMSO	623	567	56			
	MeOH	613	568	45			
	THF	596	534	62			
	PBS	630	593	37			
	With $A\beta$	606	574	32			
BBTOM-3	CH ₂ Cl ₂	691	593	98	7.81	548.5	36372
	DMSO	727	621	106			
	MeOH	718	619	99			
	THF	680	585	95			
	PBS	729	650	79			
	With $A\beta$	690	626	64			
BBTOM-4	CH ₂ Cl ₂	788	625	163	1.64	564	141686
	DMSO	820	635	185			
	MeOH	816	629	187			
	THF	773	620	153			
	PBS	823	642	181			
	With $A\beta$	793	632	161			
BBTOM-5	CH ₂ Cl ₂	822	636	186	0.26	574.5	87774
	DMSO	826	639	187			
	MeOH	827	634	193			
	THF	817	628	189			
	PBS	829	647	182			
	With $A\beta$	822	632	190			

^a Measured in CH₂Cl₂.

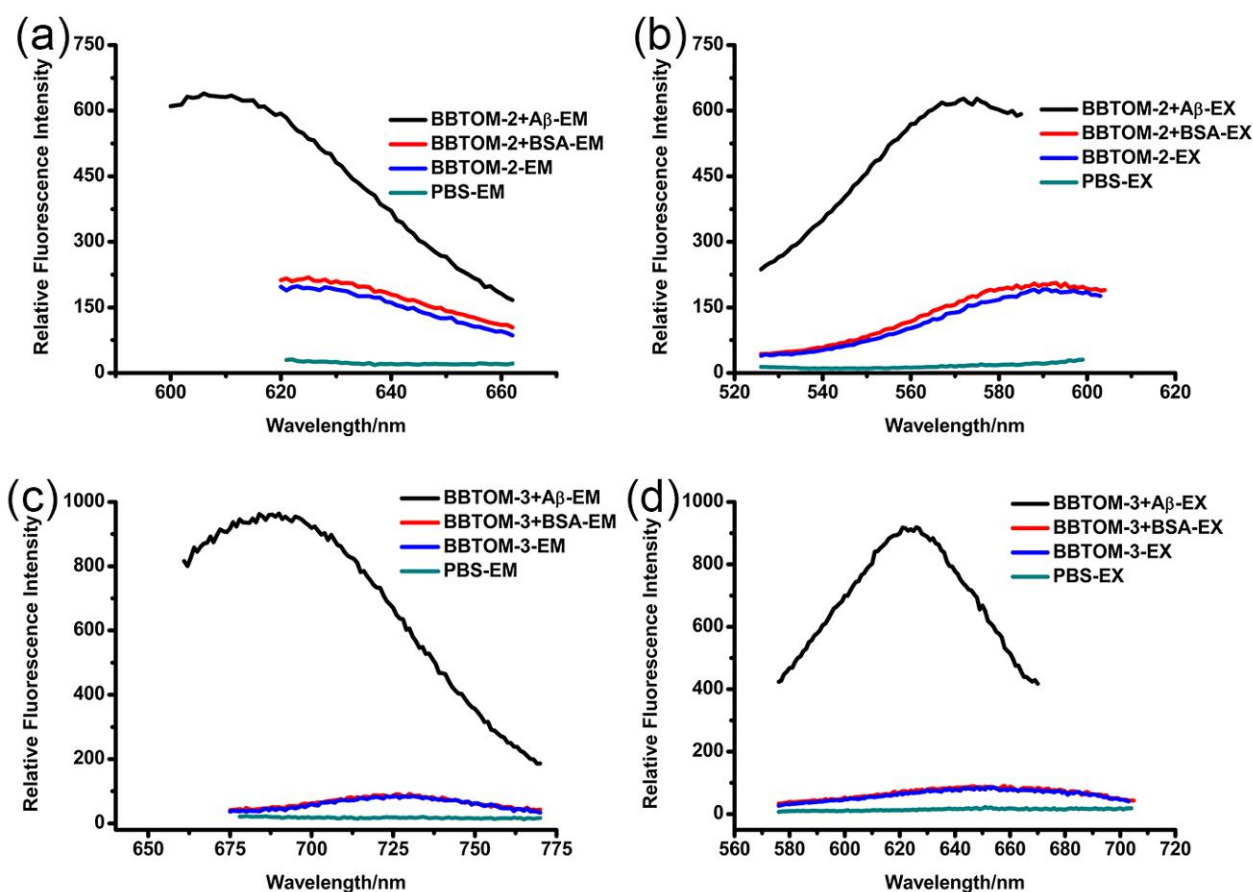
Table S2 The spectroscopic properties (absorption, excitation and emission wavelength, molar absorption coefficient, and fluorescence quantum yield) of BBTO 1-3 and BBTS 1-3.

Probe	Solvent	Fluorescence spectrum			UV-vis spectrum		
		λ_{em} (nm)	λ_{ex} (nm)	Stokes shift (nm)	Φ^a %	λ_{abs} (nm)	ϵ L mol ⁻¹ cm ⁻¹
BBTO-1	CH ₂ Cl ₂	518	469	49	0.05	459.5	1146
	DMSO	528	468	60			
	MeOH	514	467	47			
	THF	509	464	45			
	PBS	502	426	76			
BBTO-2	CH ₂ Cl ₂	602	557	45	5.71	537	61602
	DMSO	620	559	61			
	MeOH	611	566	45			
	THF	588	522	66			
	PBS	627	589	38			
BBTO-3	CH ₂ Cl ₂	696	619	77	8.16	536	25342
	DMSO	722	617	105			
	MeOH	712	620	92			
	THF	674	584	90			
	PBS	720	646	74			
	With $A\beta$	693	630	63			
BBTS-1	CH ₂ Cl ₂	546	500	46	0.79	496	80017
	DMSO	554	504	50			
	MeOH	549	494	55			
	THF	535	489	46			
	PBS	502	426	76			
BBTS-2	CH ₂ Cl ₂	632	595	37	5.93	576.5	107689
	DMSO	651	601	50			
	MeOH	641	599	42			
	THF	617	571	46			
	PBS	649	535	114			
BBTS-3	CH ₂ Cl ₂	727	632	95	6.77	584	6152
	DMSO	734	620	114			
	MeOH	737	628	109			
	THF	714	618	96			
	PBS	727	648	79			
	With $A\beta$	721	643	78			

^a Measured in CH₂Cl₂.

Fluorescence Spectral Measurements of the NIR Probes with A β Aggregates and BSA.

NIR probes (50 nM in the final test solution) in 2.9 mL PBS were added to a solution of 100 μ L of aggregated A β_{1-42} and A β_{1-40} fibrils or A β_{1-42} and A β_{1-40} monomers (30 μ g in the final test solution) or BSA (30 μ g in the final test solution), then the mixture was incubated at 37 $^{\circ}$ C with constant shaking (120 r/min) for 1 h. The mixture was then transferred to a quartz sampling cell, and the fluorescent properties (fluorescence excitation/emission wavelength and intensity) were measured on a SHIMADZU RF-5301PC spectrofluorophotometer (Japan). The fluorescence properties of the solutions of the NIRFs in PBS (50 nM) were also measured in the same manner as a blank control.



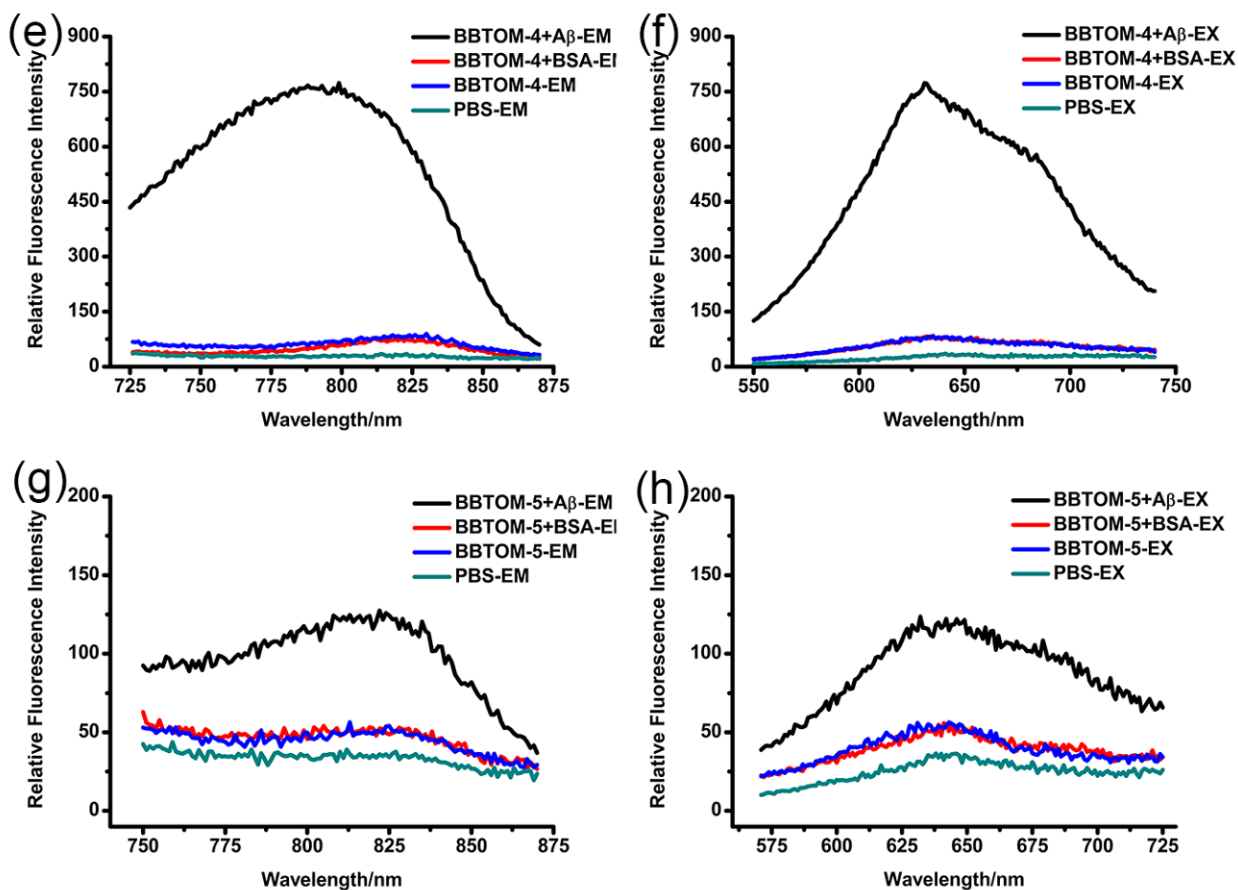
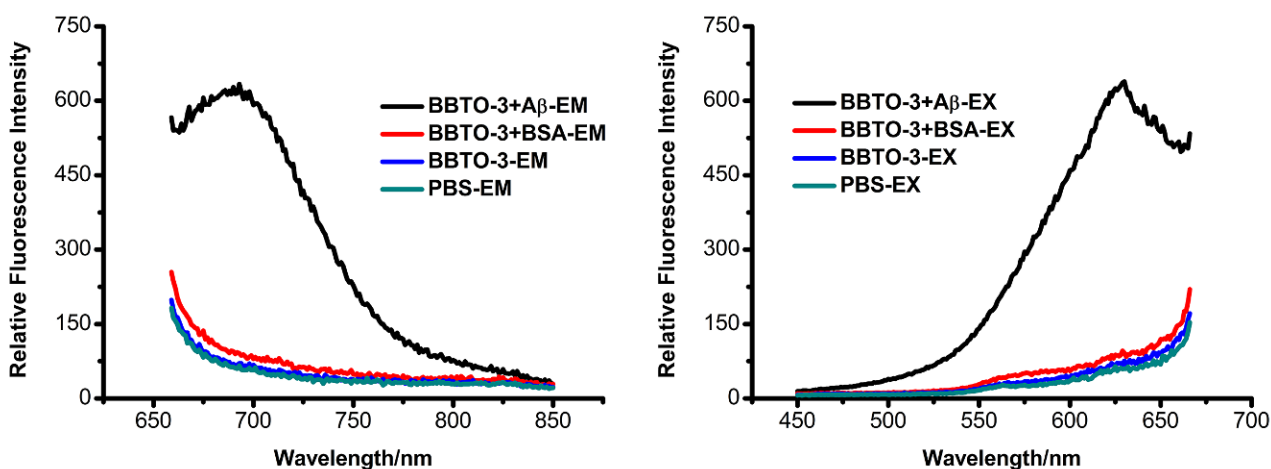


Figure. S8 The excitation (right panel) and emission (left panel) spectra of BBTOM2-5 upon interaction with $A\beta_{1-42}$ aggregates and BSA. The spectra of BBTOM2-5 solutions in PBS and PBS alone were also measured in the same condition.



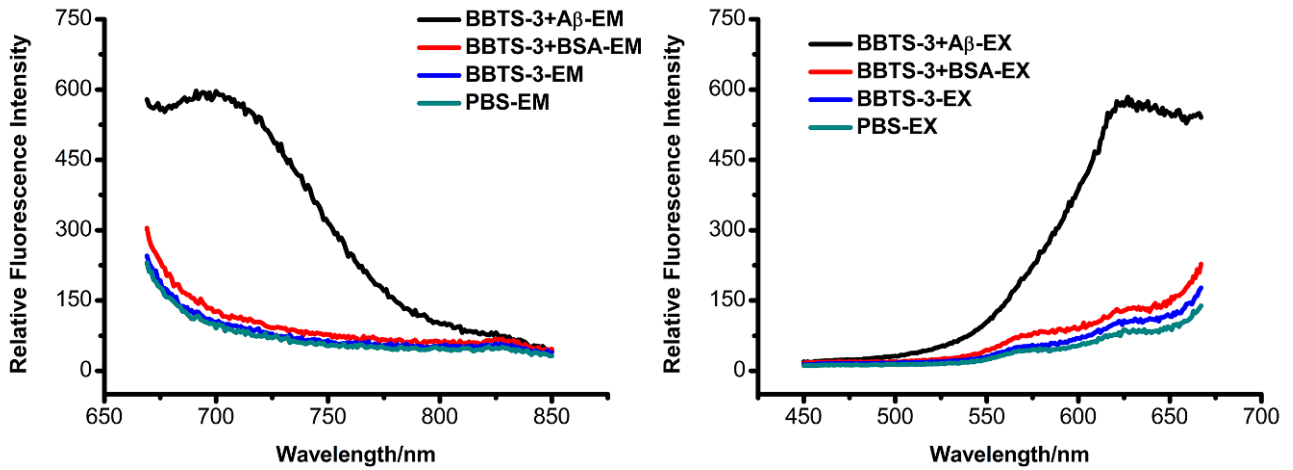


Figure. S9 The excitation (right panel) and emission (left panel) spectra of BBTO-3 and BBTS-3 upon interaction with $A\beta_{1-42}$ aggregates and BSA. The spectra of BBTO-3 and BBTS-3 solutions in PBS and PBS alone were also measured in the same condition.

***In Vitro* Fluorescent Staining.**

Paraffin-embedded brain sections from Tg mice (C57BL6, APP^{swe}/PSEN1, 12-month old, male), wild-type mice (C57BL6, 12-month old, male), AD human (91-year old, male; 64-year old, female, temporal lobe) and healthy human (72-year old, male, temporal lobe) were used for the neuropathological staining. The brain slices were deparaffinized with 10 min washes in xylene, 5 min washes in 100% ethanol, and running double distilled water for 5 min. Next, the brain sections were incubated with aqueous solutions of NIR probes (1.0 μ M, 5% DMSO, 10% ethanol) for 5 min at room temperature. The localization of plaques was confirmed by staining the adjacent sections with Th-S (0.125% in water). Fluorescent observation was carried out on an Axio Observer Z1 (Zeiss, Germany) equipped with DAPI, AF488, AF546, and Cy5 filter sets.

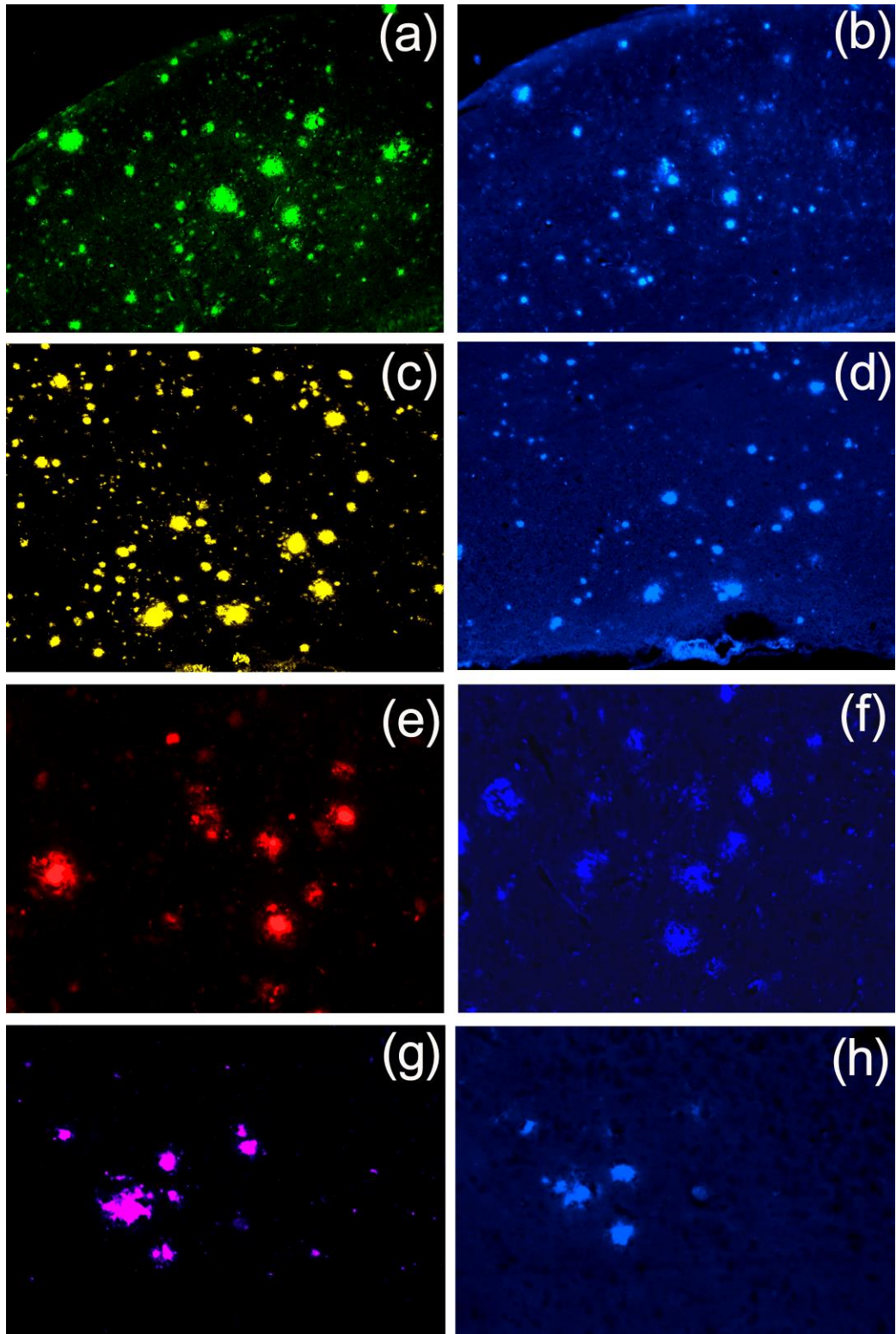


Figure. S10 *In vitro* fluorescent staining of A β plaques on brain sections of Tg mice by BBTOMs. (a, 10 \times), (c, 10 \times), (e, 10 \times) and (g, 20 \times) were stained by BBTOM-1, -2, -4, and BBTOM-5, respectively. The presence and distribution of plaques on the sections were confirmed by fluorescence staining

using Th-S on the adjacent section (b, 10×), (d, 10×), (f, 10×) and (h, 20×).

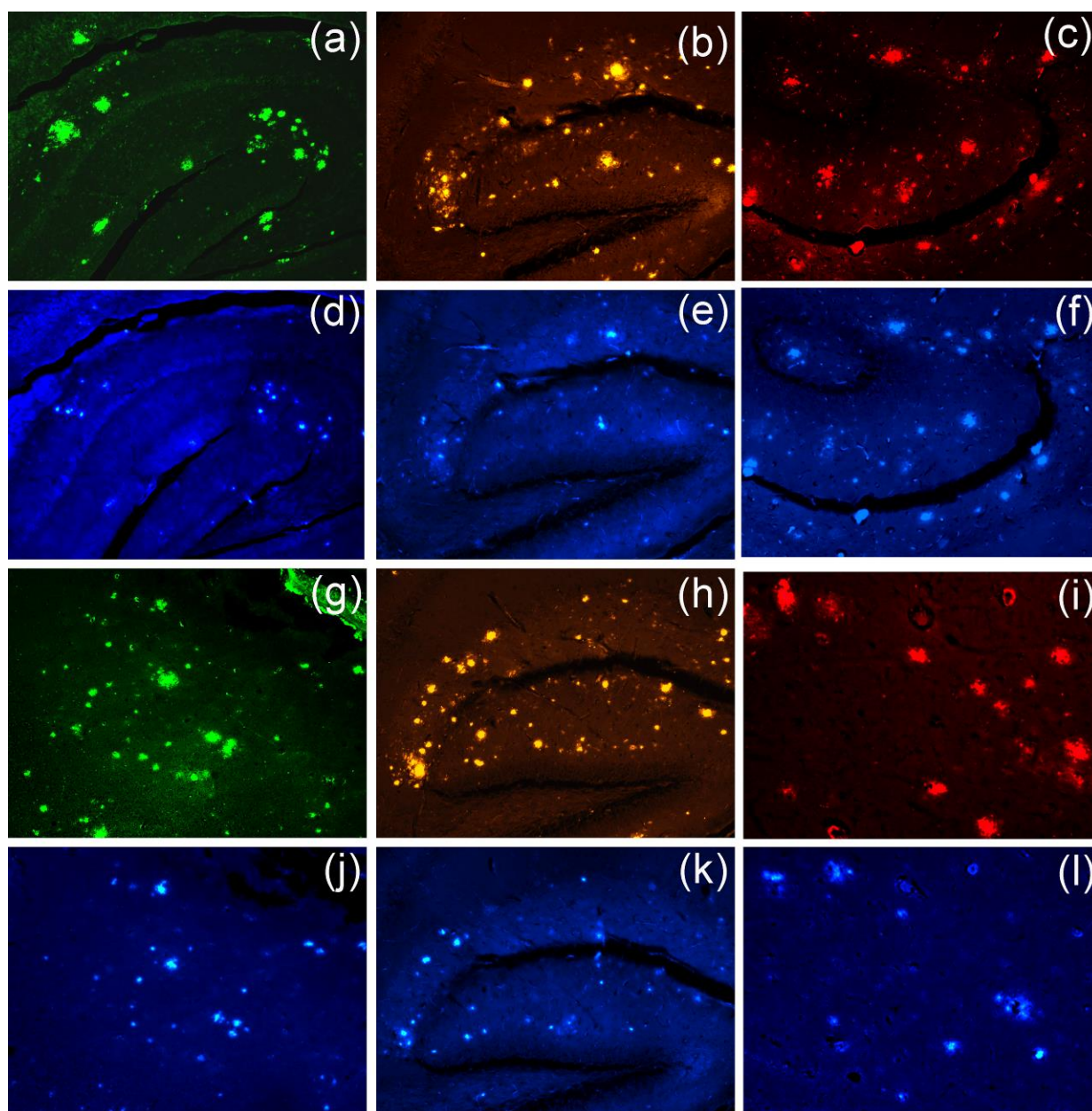


Figure. S11 *In vitro* fluorescent staining of $A\beta$ plaques on brain sections of Tg mice by BBTO1-3 and BBTS1-3. (a, 10×), (b, 10×), (c, 10×), (g, 10×), (h, 10×) and (i, 20×) were stained by BBTO-1, -2, -3, and BBTS-1, -2, -3, respectively. The presence and distribution of plaques on the sections were confirmed by fluorescence staining using Th-S on the adjacent section (d, 10×), (e, 10×), (f, 10×), (j, 10×), (k, 10×) and (l, 20×).

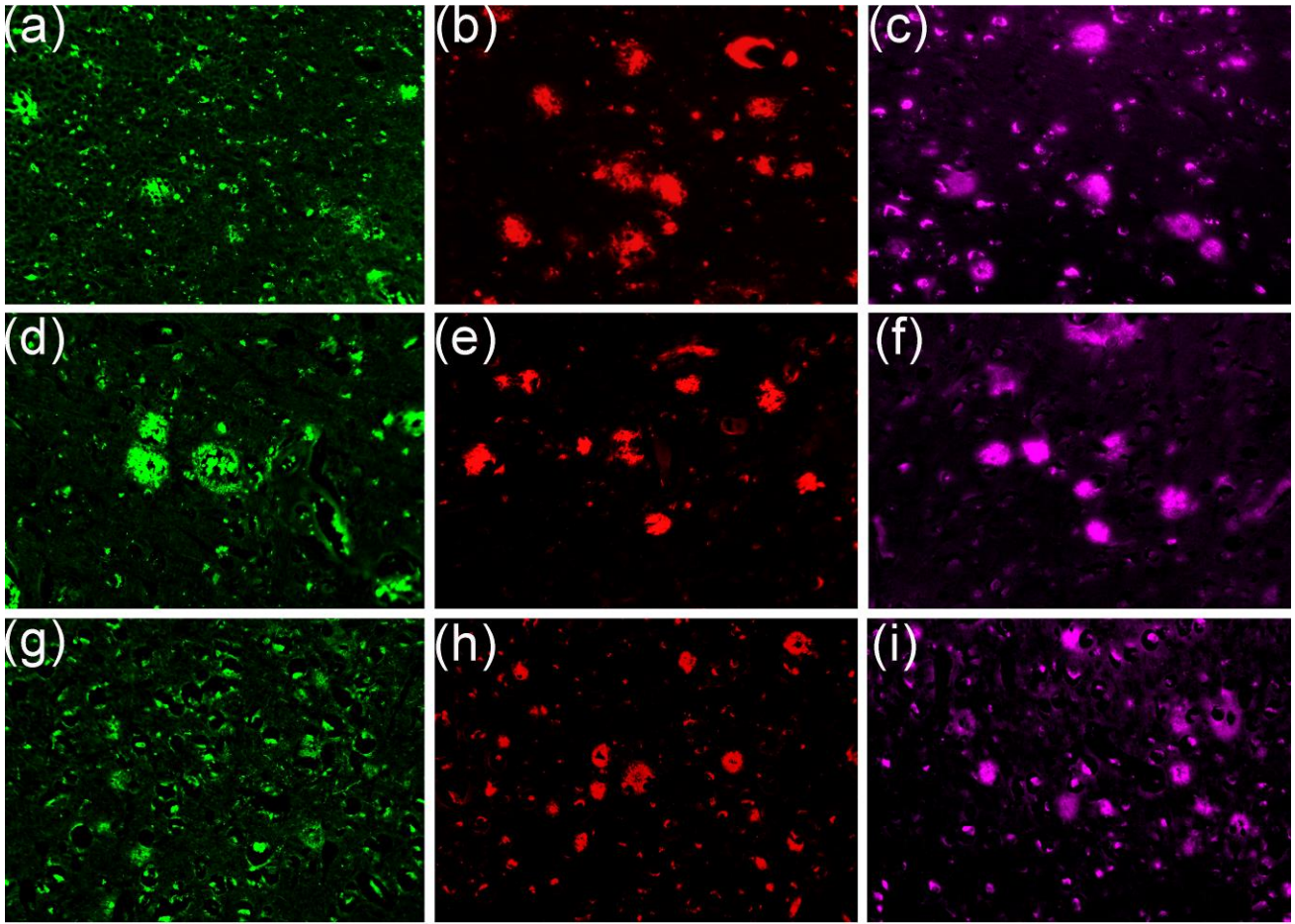


Figure. S12 *In vitro* fluorescent staining of $A\beta$ plaques on brain sections of AD patients. (a), (b) and (c) were stained by BBTOM-1, -2, -4, respectively; (d), (e) and (f) were stained by BBTO-1, -2, -3, respectively. (g), (h) and (i) were stained by BBTS-1, -2, -3, respectively. Magnification: 20 \times

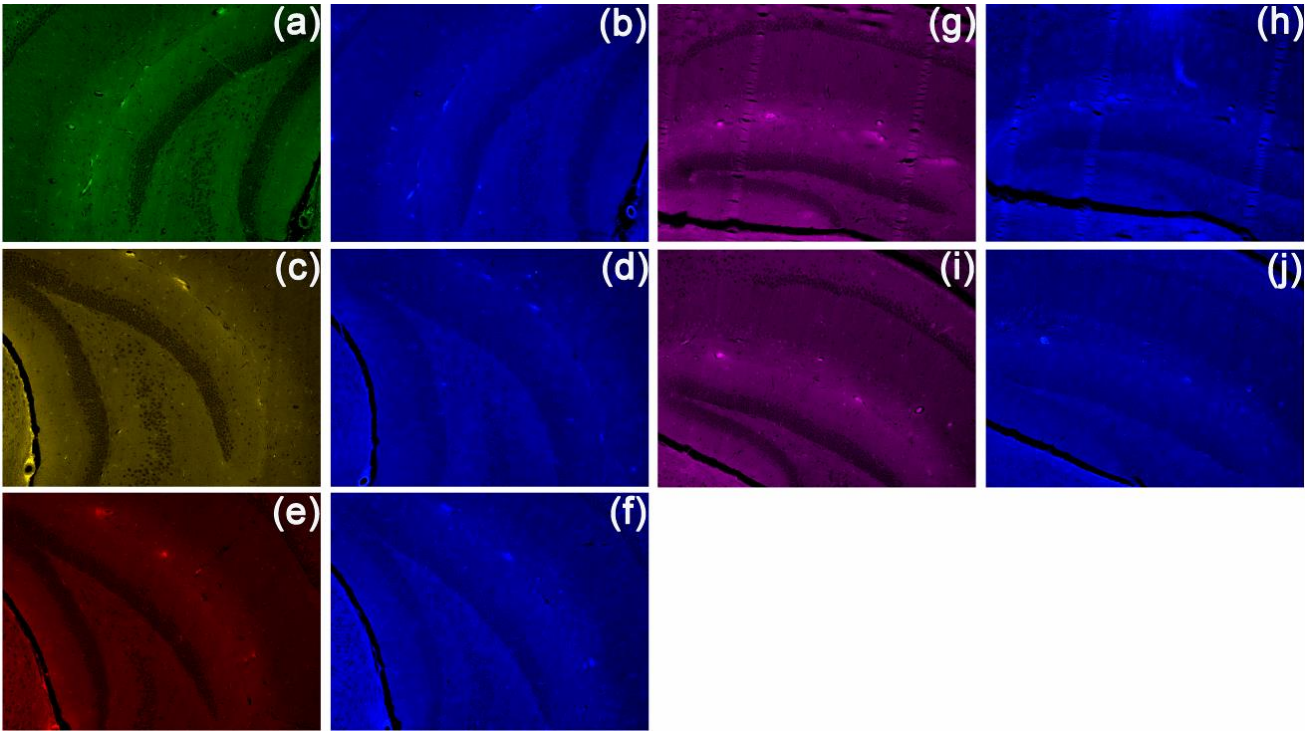


Figure. S13 *In vitro* fluorescent staining on brain sections from wild-type mouse. (a), (c), (e), (g) and (i) were stained by BBTOM-1, -2, -3, -4, and -5, respectively. The adjacent brain sections were stained with Th-S (b), (d), (f), (h) and (j). Magnification: 10 \times

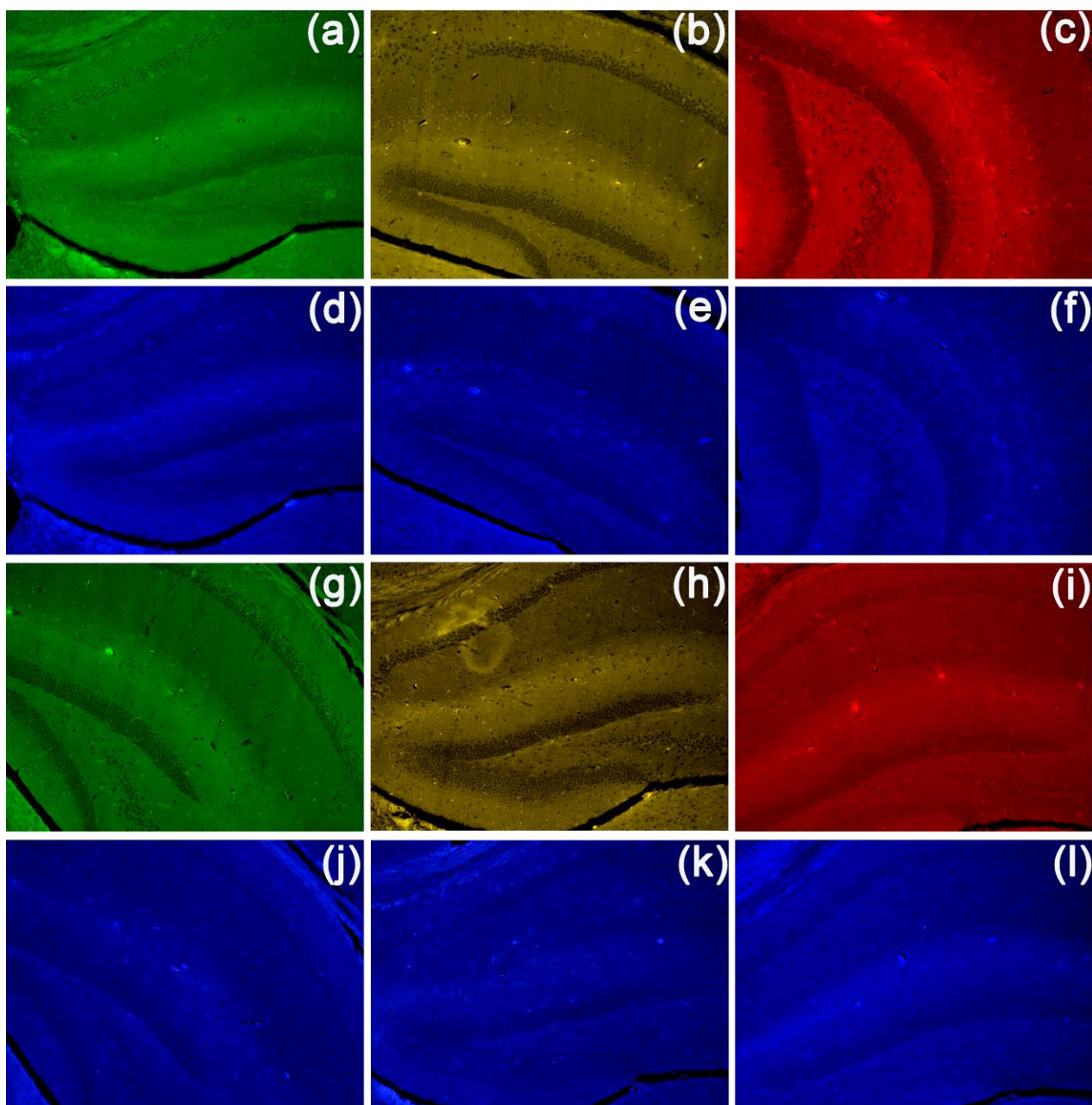


Figure. S14 *In vitro* fluorescent staining on brain sections from wild-type mouse. (a), (b) and (c) were stained by BBTO-1, -2 and -3, respectively. (g), (h) and (i) were stained by BBTS-1, -2, -3, respectively. The adjacent brain sections were stained with Th-S (d), (e), (f), (j), (k) and (l). Magnification: 10 \times .

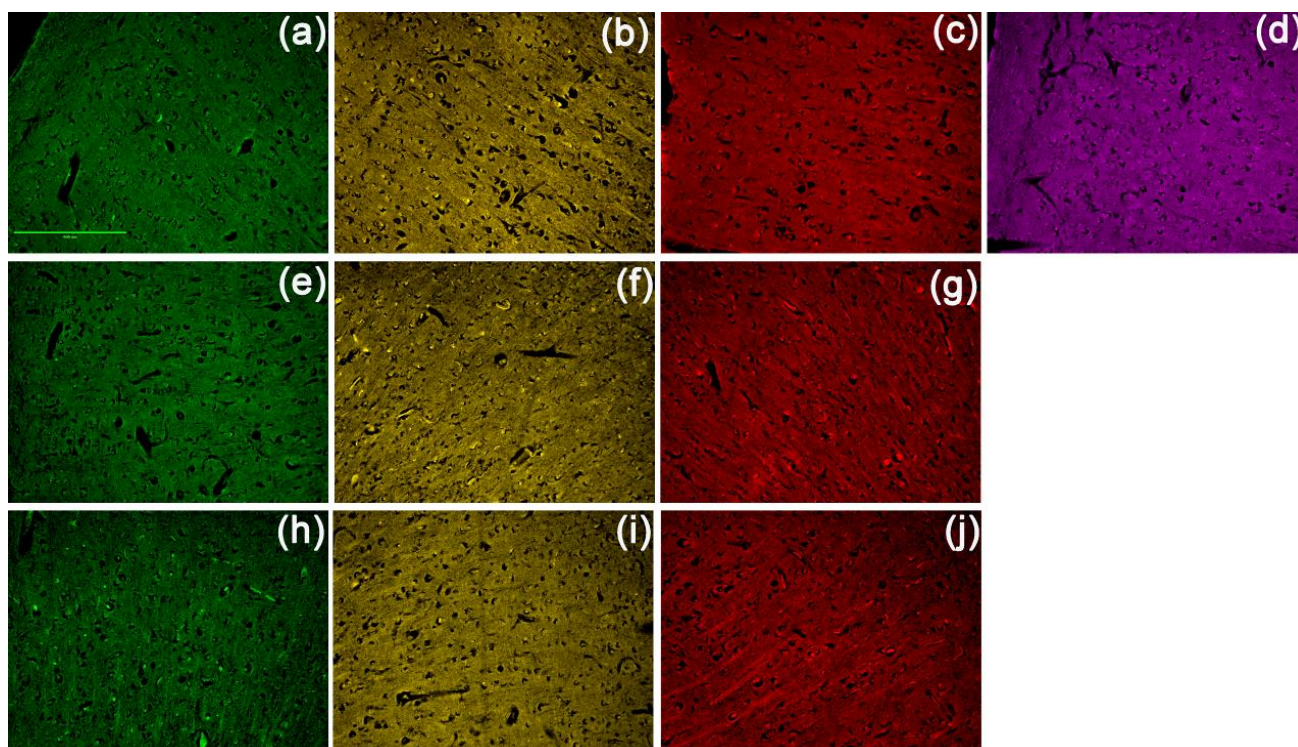


Figure. S15 *In vitro* fluorescent staining on brain sections from healthy human (72-year old, male, temporal lobe). (a), (b), (c) and (d) were stained by BBTOM-1, -2, -3 and -4, respectively. (e), (f) and (g) were stained by BBTO-1, -2, -3, respectively. (h), (i) and (j) were stained by BBTS-1, -2, -3, respectively. Magnification: 10 \times .

***In vitro* Inhibition Binding Assays Using A β ₁₋₄₂ Aggregates.**

[¹²⁵I]IMPY was employed as radioligand to determine the binding affinities of the NIR probes in *in vitro* competitive binding assays. [¹²⁵I]IMPY and A β ₁₋₄₂ aggregates were prepared according to procedures described previously.⁴ Inhibition experiments were manipulated in 12 \times 75 mm borosilicate glass tubes, 100 μ L of aggregated A β fibrils (60 nM in the final assay mixture) was added to a mixture containing 100 μ L of [¹²⁵I]IMPY at appropriate concentration, 100 μ L of probes (10⁻⁵ to 10⁻¹⁰ M in ethanol), and 700 μ L of PBS (0.2 M, pH = 7.4, containing 0.1 % BSA) in a final volume of 1 mL. The mixture was incubated at 37 $^{\circ}$ C for 2 h, then it was transferred to borosilicate

glass microfiber filters (Whatman, GF/B) using a Brandel Mp-48T cell harvester. After the bound and free [¹²⁵I]IMPY were separated, filters containing the bound [¹²⁵I]IMPY were measured by gamma counter (WALLAC/Wizard 1470, USA) with 70% counting efficiency. The half maximal inhibitory concentration (IC₅₀) was determined from the displacement curves of three independent assays using GraphPad Prism 4.0 and the inhibition constant (K_i) was calculated using the Cheng-Prusoff equation: $K_i = IC_{50}/(1 + [L]/K_d)$.⁵

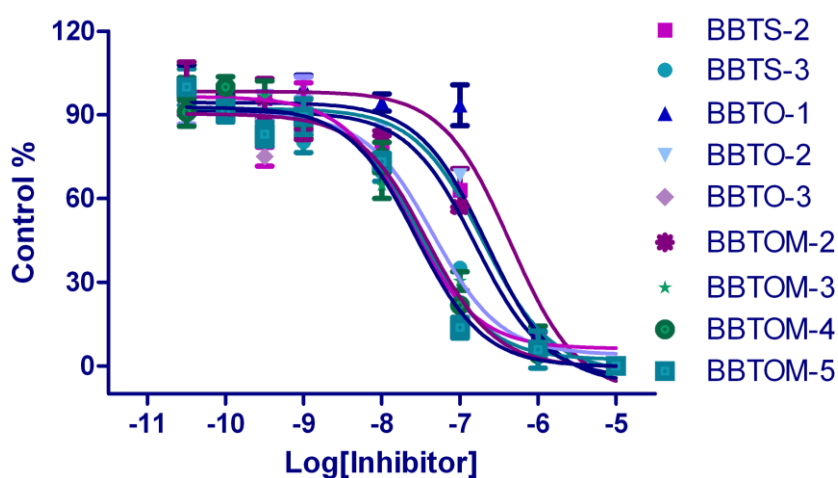


Figure. S16 Inhibition curves for the binding of [¹²⁵I]IMPY to A β ₁₋₄₂ aggregates.

Blood-Brain Barrier (BBB) Penetrating studies of BBTOM-3.

A solution of BBTOM-3 (20% DMSO and 80% propylene glycol, 50 μ L, 2 mg/Kg) was injected via tail vein of ICR mice (20-22 g, male). Next, the mice were sacrificed at proper time points of 2, 10, 30 and 60 min (n = 3 for each time point), and the brain samples were dissected out to homogenize with 1 mL acetonitrile for twice. Then, the extracts were filtered by flashing nylon membrane (0.22 μ m) and dried over anhydrous sodium sulfate and washed by 1 mL acetonitril. After that, 100 μ L of the combined acetonitrile were analyzed by Agilent 1260 Infinity Quaternary LC (Agilent

Technologies) system, HPLC conditions: Venusil MP C18 column (Agela Technologies, 5 μm , 4.6 mm \times 250 mm), $\text{CH}_3\text{CN}/\text{H}_2\text{O} = 80\%/20\%$, 1 mL/min. The blank control was obtained by analyzing the solution of the probe (10 μL diluted to 3.0 mL acetonitrile).

In Vivo Near-Infrared Imaging.

Double Tg mice (n = 3, C57BL6, APPsw/PSEN1, 14 month-old, male) and an age-matched control mice (n = 3, C57BL6, 14-month-old, male) were shaved before background imaging and were intravenous injected with BBTOM-3 (0.5 mg/kg, 15% DMSO, 15% tween-80, 70% saline, 50 μL). The fluorescence signals of the brain are recorded at different time points after intravenous injected with BBTOM-3 on an IVIS Lumina III system. A filter set (ex. at 560 nm and em. at 710 nm) was used for the measurement. The mice were kept on the imaging stage under anesthesia with 2.5% isoflurane gas in an oxygen flow (0.8 L/min) during the imaging process. Living Image Software was employed to analyze the imaging data, and an ROI was drawn around the brain section. Intensity of brain fluorescence was calculated from the radiant efficiency. The data were analyzed according to the procedures described previously.⁶

Ex Vivo Fluorescent Staining of BBTOM-3 to A β Plaques in the Brain of Transgenic Mouse.

BBTOM-3 (1 mg/kg, 20% DMSO, 80% propylene glycol, 50 μL) was intravenous injected to a double Tg mouse (C57BL6, APPsw/PSEN1, 14-month-old, male) and an age-matched control mouse (C57BL6, 14-month-old, male). Next, they were sacrificed at 20 min after injection. The brains were

excised, embedded in optimum cutting temperature compound (OCT). Frozen sections of 20 μm were cut, and fluorescent observation was performed by Axio Observer Z1 (Zeiss, Germany) equipped with an AF488 filter sets. Furthermore, the $A\beta$ plaques were further confirmed by the staining of the same section with Th-S (0.125%) using a DAPI filter set.

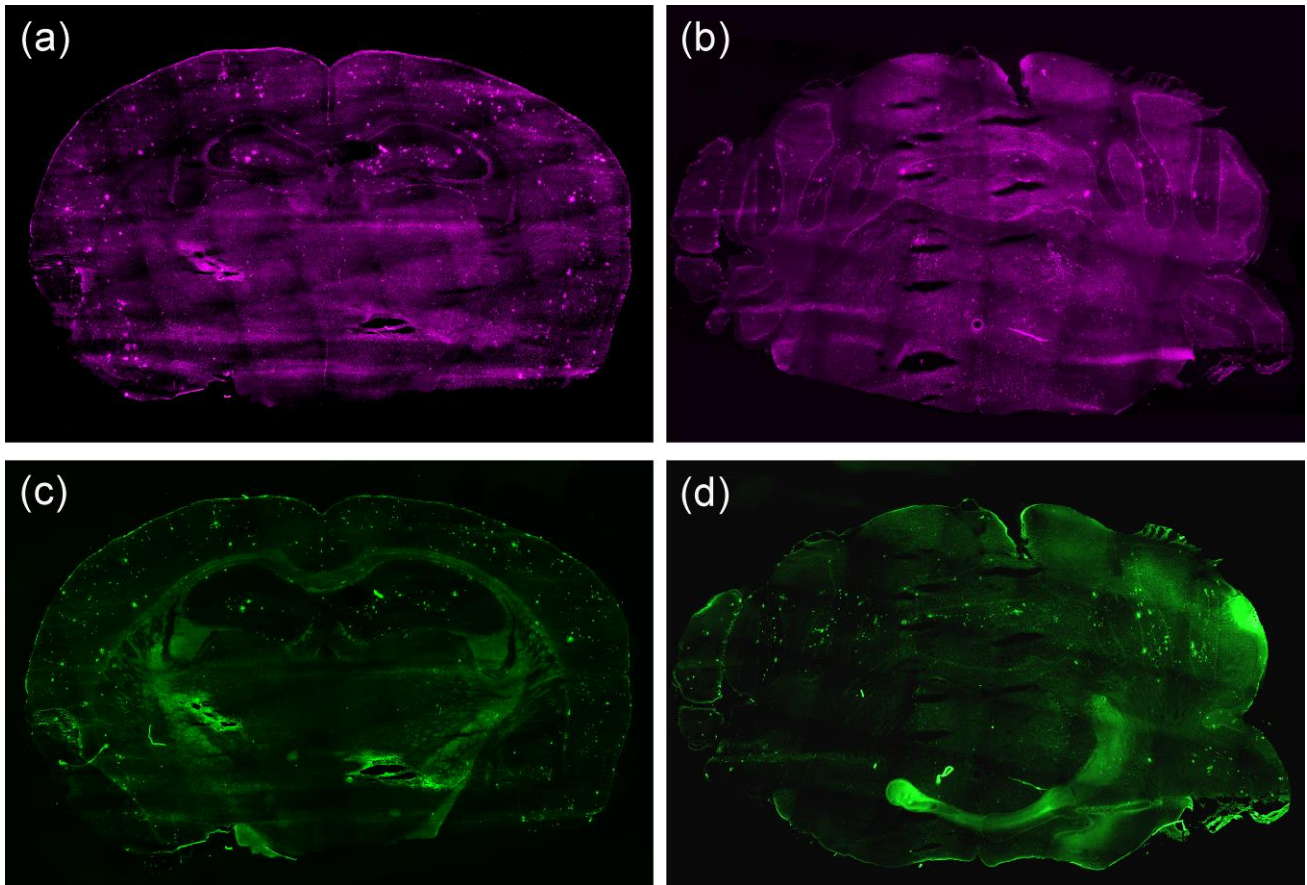


Figure. S17 *Ex vivo* fluorescence observation of coronal brain slices from a Tg mouse (a: cerebrum region; b: cerebellum region) after injection of BBTOM-3 (1 mg/kg). The $A\beta$ plaques were further confirmed by staining the same sections with Th-S (c and d).

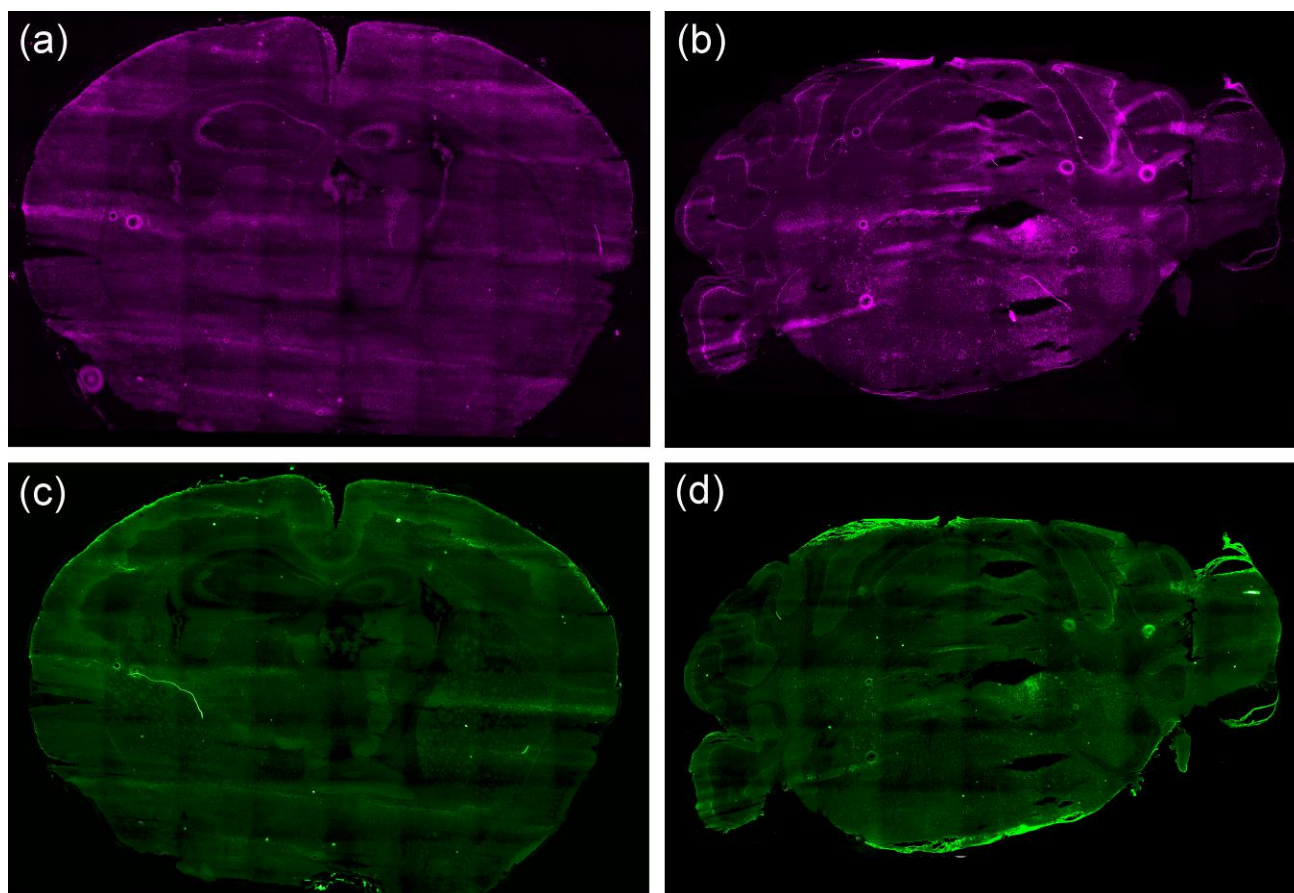


Figure. S18 *Ex vivo* fluorescence observation of coronal brain slices from a wild-type mouse (a: cerebrum region; b: cerebellum region) after injection of BBTOM-3 (1 mg/kg). The A β plaques were further confirmed by staining the same sections with Th-S (c and d).

Photo-isomerization.

BBTOM-3 was dissolved in acetonitrile and was irradiated under UV light (365 nm) for 1 hour, then the samples were analyzed by HPLC to verify the retention time and purity with or without irradiation. As shown in Fig S19, there was no retention time and purity change of BBTOM-3 after UV irradiation, which indicated that BBTOM-3 was stable towards photochemical isomerization.

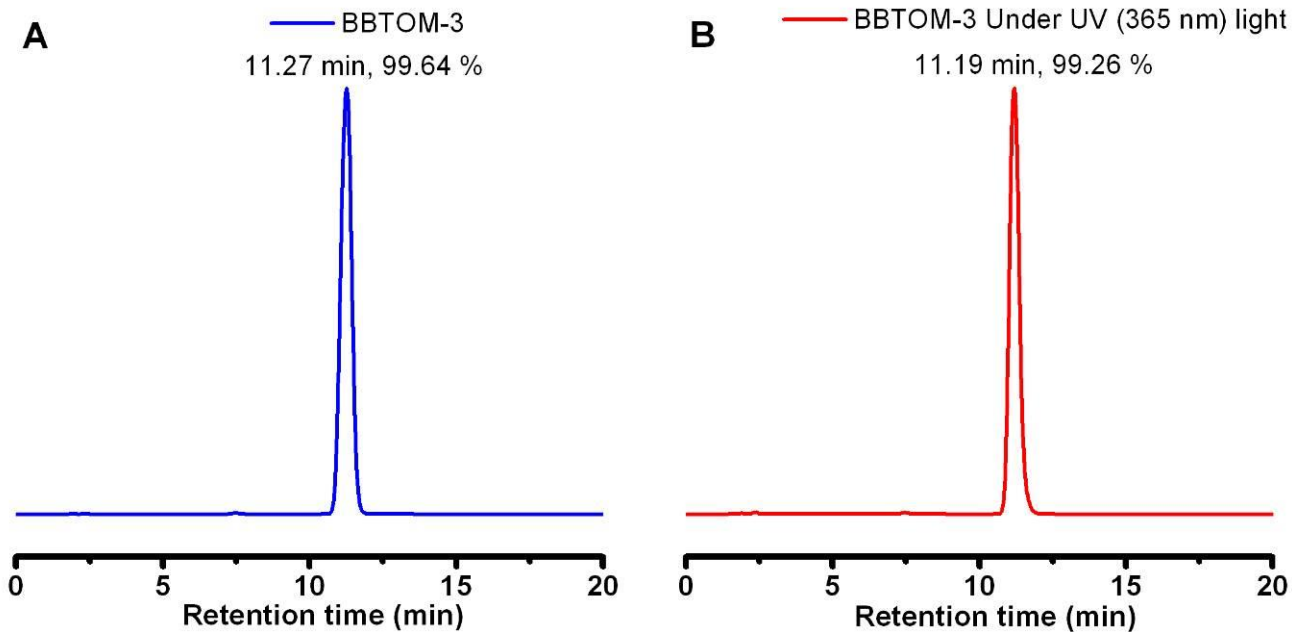
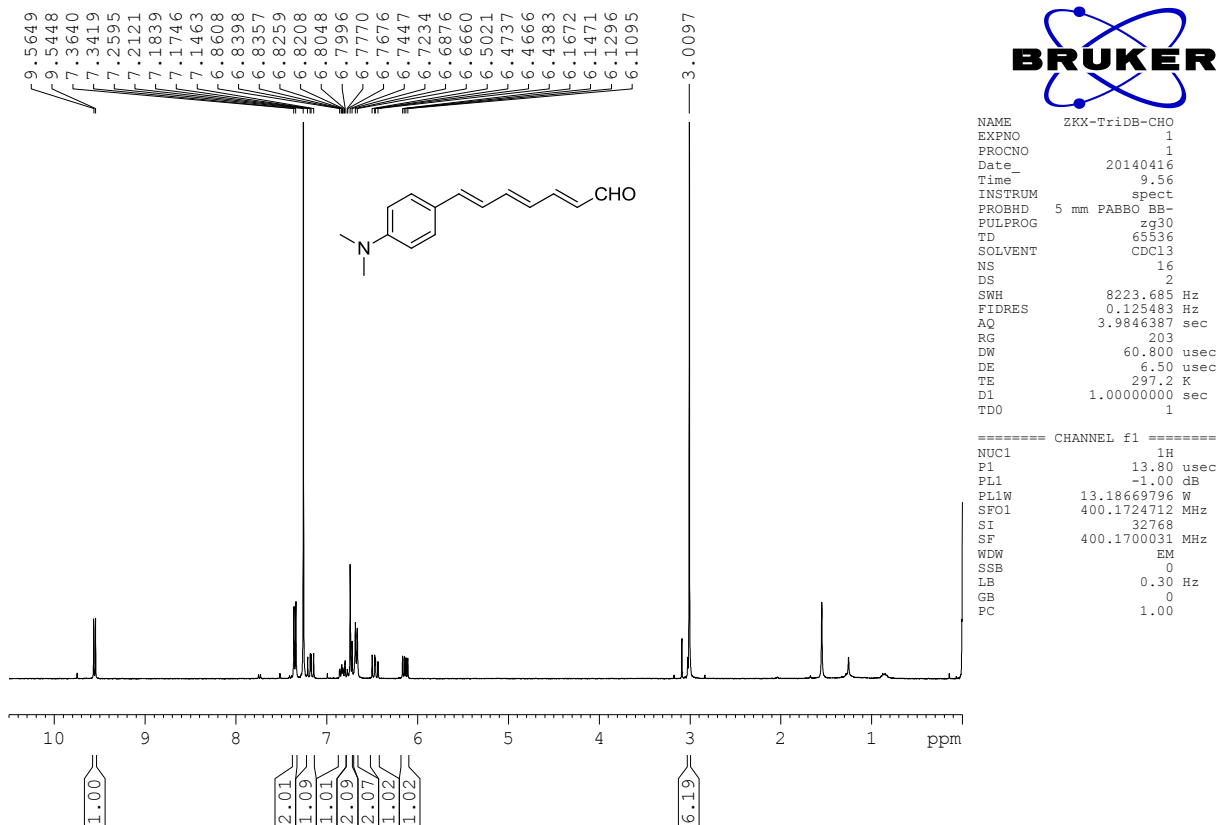
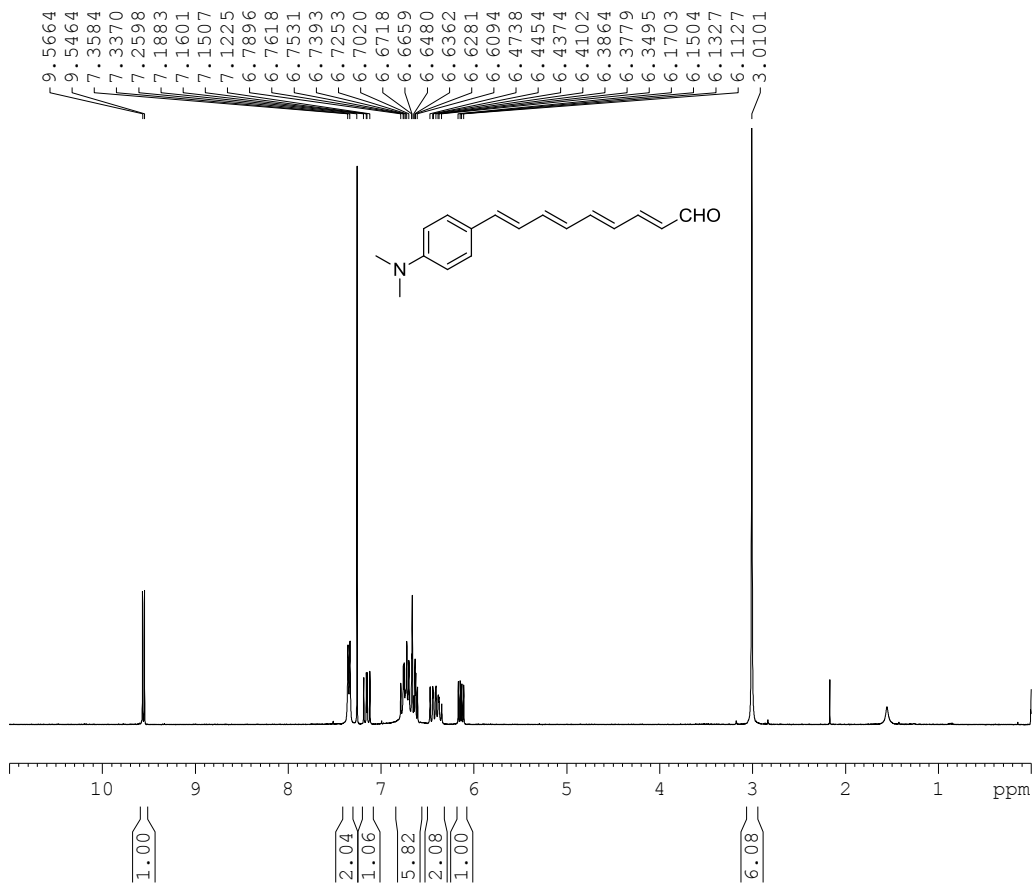


Figure. S19 HPLC profiles of BBTOM-3 (A) and BBTOM-3 under 1 hour of UV light (365 nm) irradiation (B).

NMR and MS Spectra of All Compounds.



¹H NMR (CDCl₃) spectrum of compound **2**

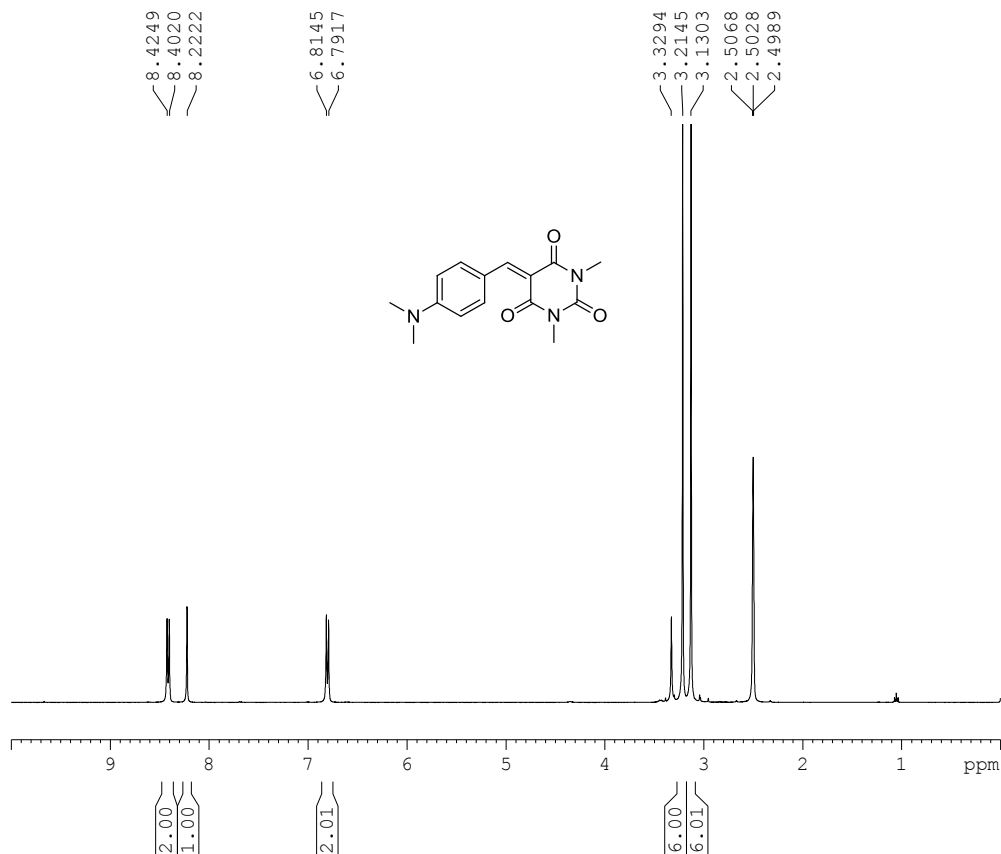


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PROCNO   1
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PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      8223.685 Hz
FIDRES   0.125483 Hz
AQ       3.9846387 sec
RG       203
DW       60.800 usec
DE       6.50 usec
TE       297.2 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     1H
P1       13.80 usec
PL1     -1.00 dB
PL1W    13.18669796 W
SFO1    400.1724712 MHz
SI       32768
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GB       0
PC       1.00
  
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¹H NMR (CDCl₃) spectrum of compound 3

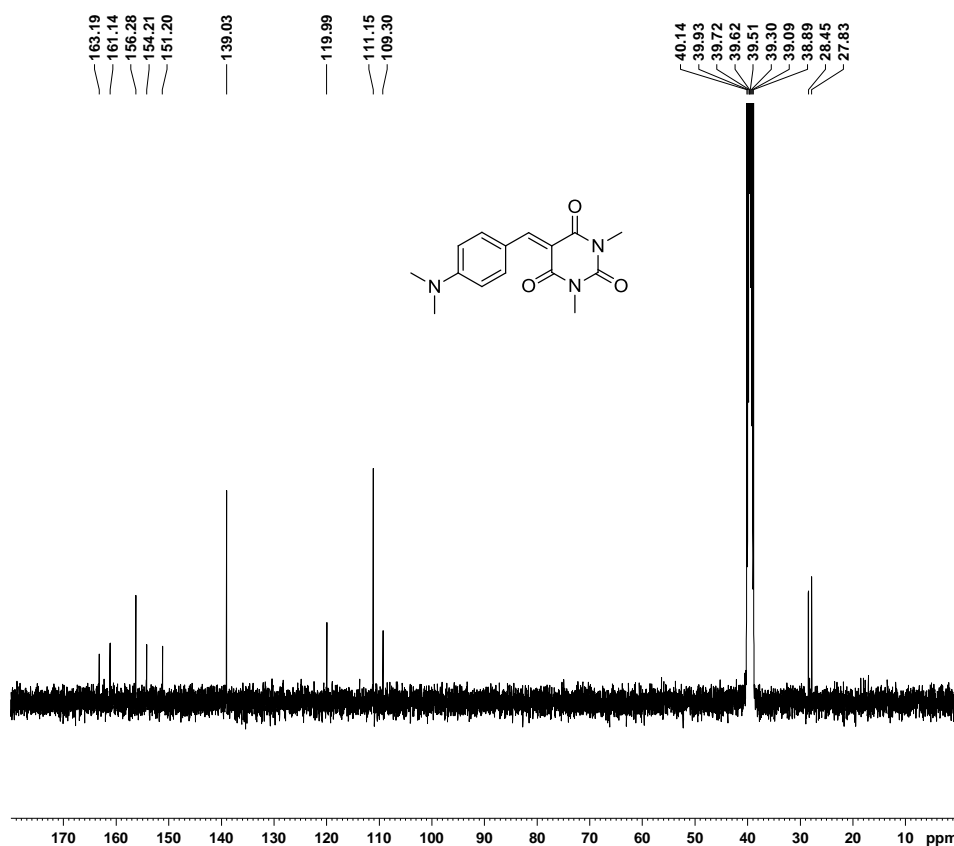


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TD       65536
SOLVENT  DMSO
NS       16
DS       2
SWH      8223.685 Hz
FIDRES   0.125483 Hz
AQ       3.9846387 sec
RG       203
DW       60.800 usec
DE       6.50 usec
TE       295.4 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     1H
P1       13.80 usec
PL1     -1.00 dB
PL1W    13.18669796 W
SFO1    400.1724712 MHz
SI       32768
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¹H NMR (DMSO-d₆) spectrum of BBTOM-1



```

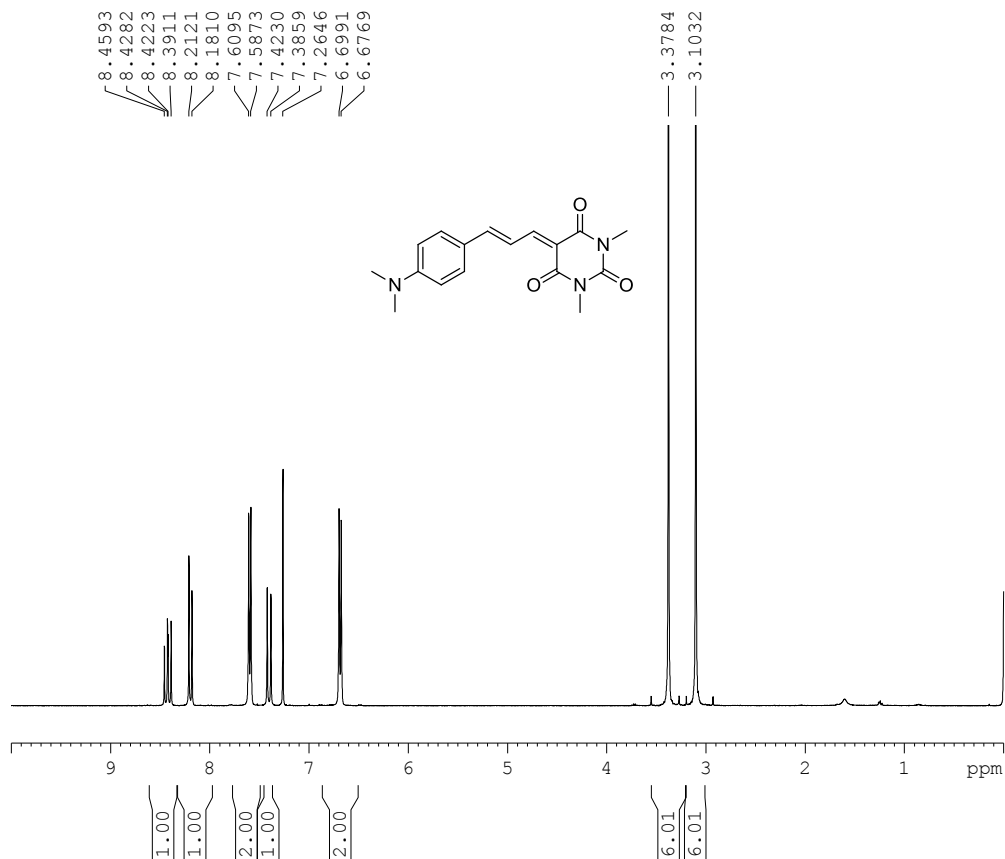
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FIDRES    0.366798 Hz
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RG        203
DW        20.800 usec
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D1        2.00000000 sec
D11       0.03000000 sec
TD0       1
    
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PL1       -2.00 dB
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===== CHANNEL f2 =====
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NUC2      1H
PCPD2     80.00 usec
PL2       -1.00 dB
PL12      14.26 dB
PL13      14.46 dB
PL2W      13.18669796 W
PL12W     0.39276794 W
PL13W     0.37509048 W
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SF        100.6228773 MHz
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¹³C NMR (DMSO-d₆) spectrum of BBTOM-1



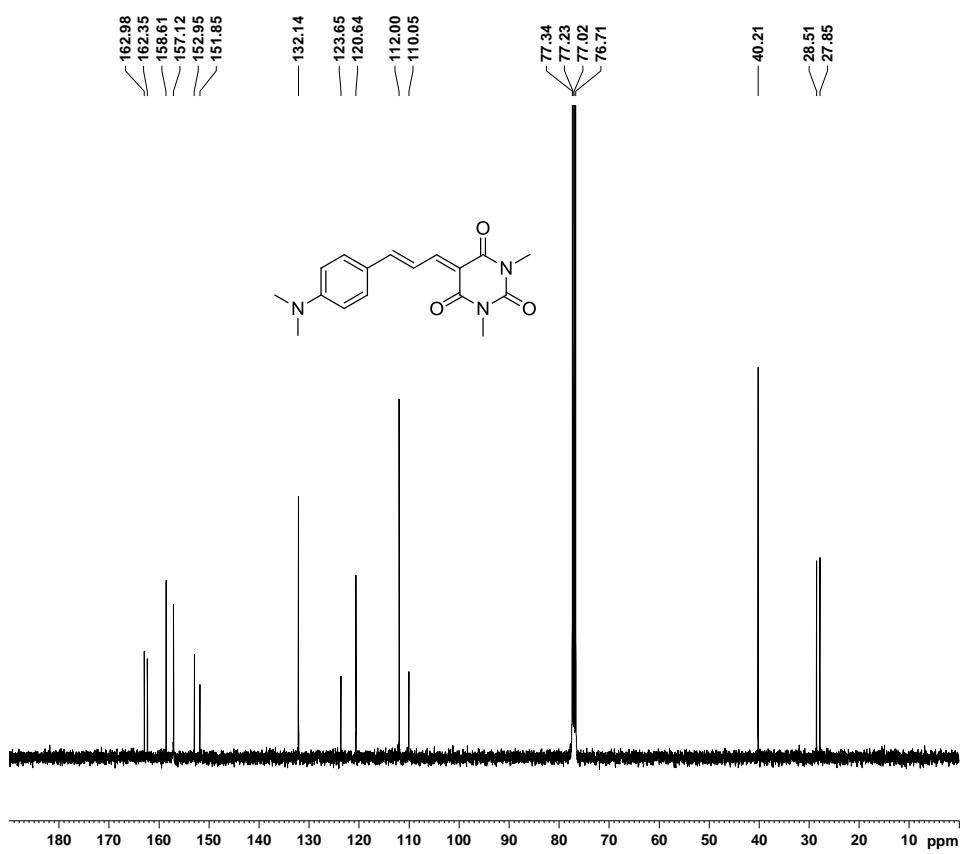
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SWH       8223.685 Hz
FIDRES    0.125483 Hz
AQ        3.9846387 sec
RG        203
DW        60.800 usec
DE        6.50 usec
TE        295.5 K
D1        1.00000000 sec
TD0       1
    
```

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===== CHANNEL f1 =====
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¹H NMR (CDCl₃) spectrum of BBTOM-2



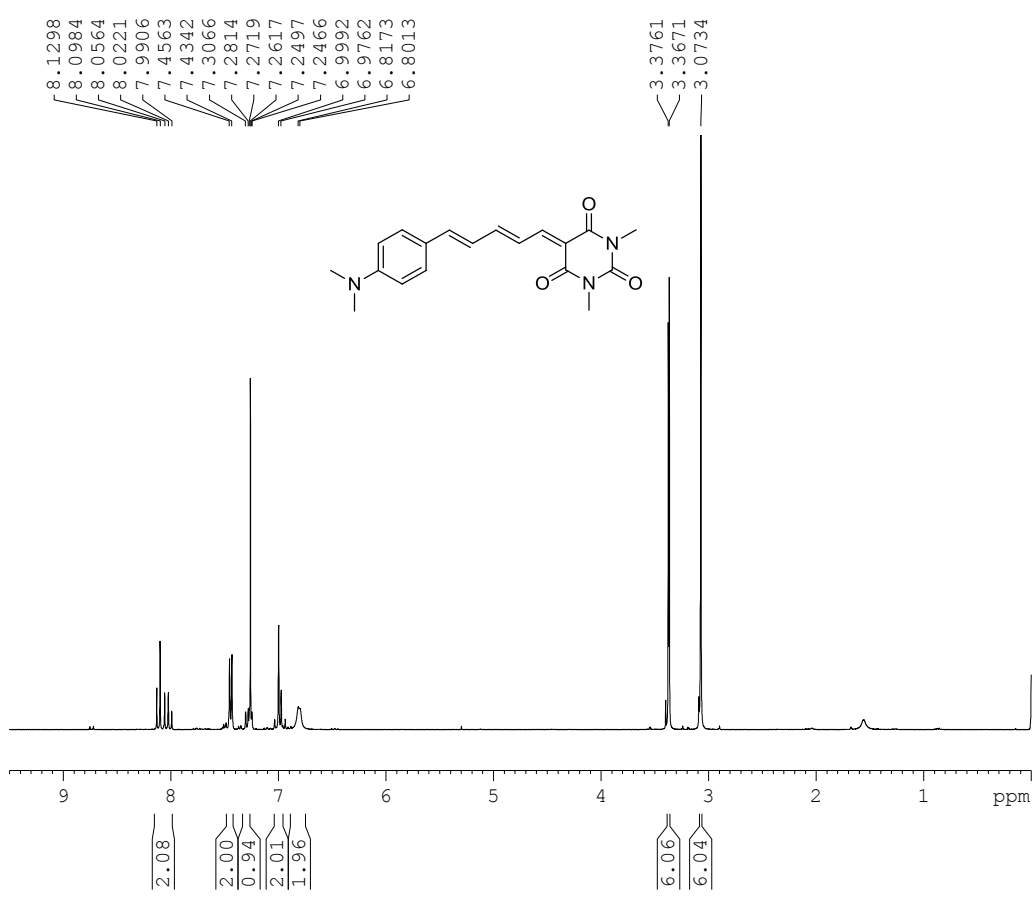
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PROBHD   5 mm PABBO BB-
FULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       2000
DS       4
SWH      24038.461 Hz
FIDRES   0.366798 Hz
AQ       1.3631988 sec
RG       203
DW       20.800 usec
DE       6.50 usec
TE       300.0 K
D1       2.00000000 sec
D11      0.03000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     13C
P1       8.50 usec
PL1     -2.00 dB
PL1W    57.32743073 W
SF01    100.6328888 MHz

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2     1H
PCPD2    80.00 usec
PL2     -1.00 dB
PL12    14.26 dB
PL13    14.46 dB
PL1W    13.18669796 W
PL2W    0.39276794 W
PL12W   0.37509048 W
SF02    400.1716007 MHz
SI       32768
SF       100.6228252 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
    
```

¹³C NMR (CDCl₃) spectrum of BBTOM-2

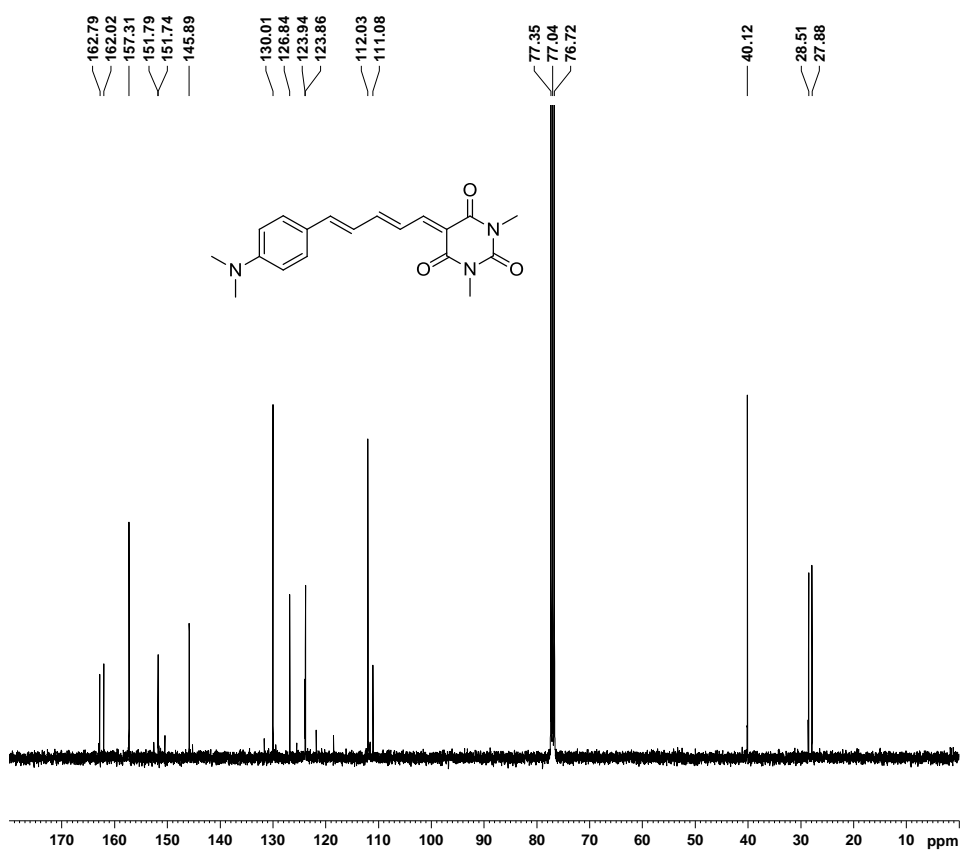


```

NAME      ZKX-BBTOM-3-20150527
EXPNO    1
PROCNO   1
Date_    20150527
Time     9.37
INSTRUM  spect
PROBHD   5 mm PABBO BB-
FULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      8223.685 Hz
FIDRES   0.125483 Hz
AQ       3.9846387 sec
RG       203
DW       60.800 usec
DE       6.50 usec
TE       297.2 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
NUC1     1H
P1       13.80 usec
PL1     -1.00 dB
PL1W    13.18669796 W
SF01    400.1724712 MHz
SI       32768
SF       400.1700026 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
```

¹H NMR (CDCl₃) spectrum of BBTOM-3



```

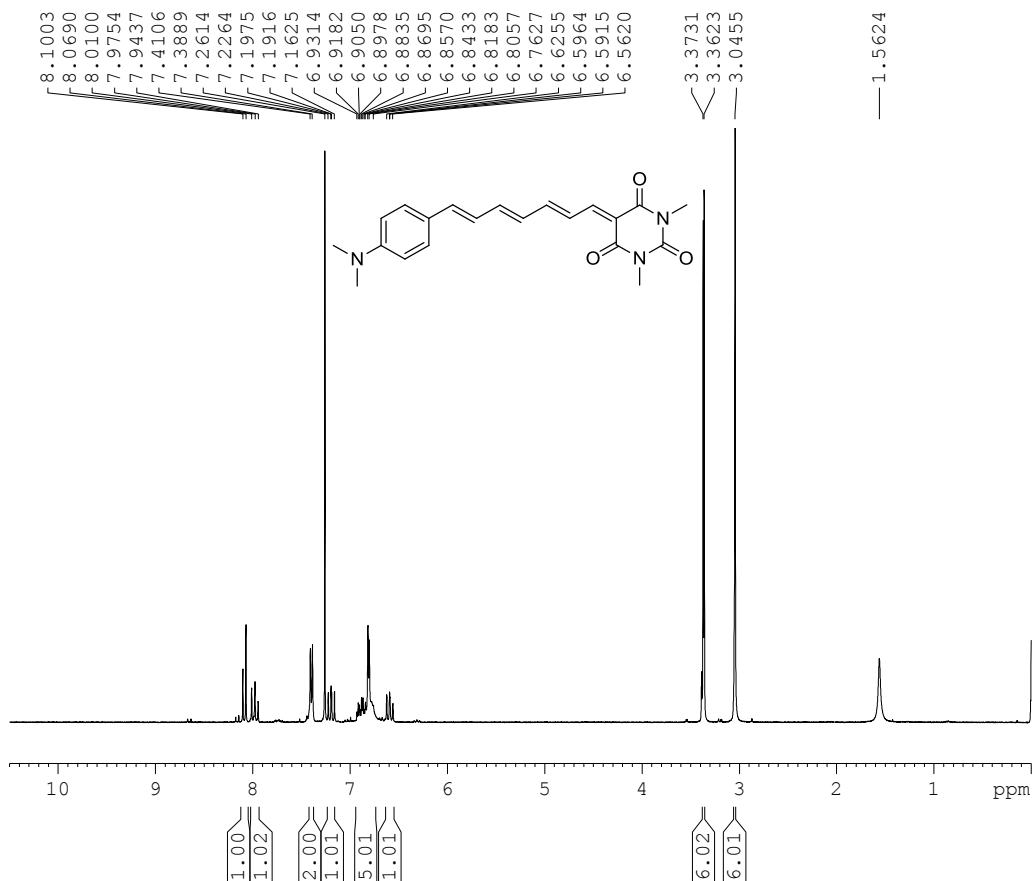
NAME      ZKX-BBATO-M-3-C
EXPNO    1
PROCNO   1
Date_    20140429
Time     20.34
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       618
DS       4
SWH      24038.461 Hz
FIDRES   0.366798 Hz
AQ       1.3631988 sec
RG       203
DW       20.800 usec
DE       6.50 usec
TE       301.0 K
D1       2.00000000 sec
D11      0.03000000 sec
TD0     1
    
```

```

===== CHANNEL f1 =====
NUC1     13C
P1       8.50 usec
PL1      -2.00 dB
PL1W     57.32743073 W
SFO1     100.6328888 MHz

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2     1H
PCPD2    80.00 usec
PL2      -1.00 dB
PL12     14.26 dB
PL13     14.46 dB
PL2W     13.18669796 W
PL12W    0.39276794 W
PL13W    0.37509048 W
SFO2     400.1716007 MHz
SI       32768
SF       100.6228244 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
    
```

¹³C NMR (CDCl₃) spectrum of BBTOM-3



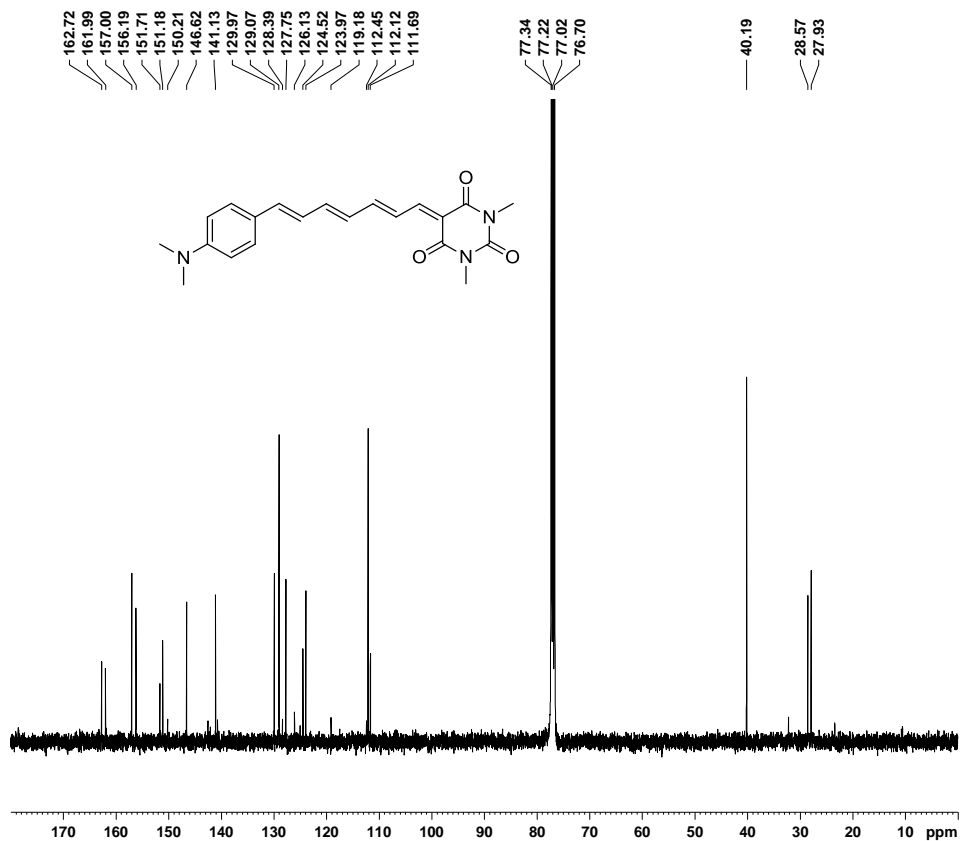
```

NAME      ZKX-BBTOM-4-20150527
EXPNO    1
PROCNO   1
Date_    20150527
Time     9.43
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      8223.685 Hz
FIDRES   0.125483 Hz
AQ       3.9846387 sec
RG       203
DW       60.800 usec
DE       6.50 usec
TE       297.2 K
D1       1.00000000 sec
TD0     1
    
```

```

===== CHANNEL f1 =====
NUC1     1H
P1       13.80 usec
PL1      -1.00 dB
PL1W     13.18669796 W
SFO1     400.1724712 MHz
SI       32768
SF       400.1700027 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
```

¹H NMR (CDCl₃) spectrum of BBTOM-4

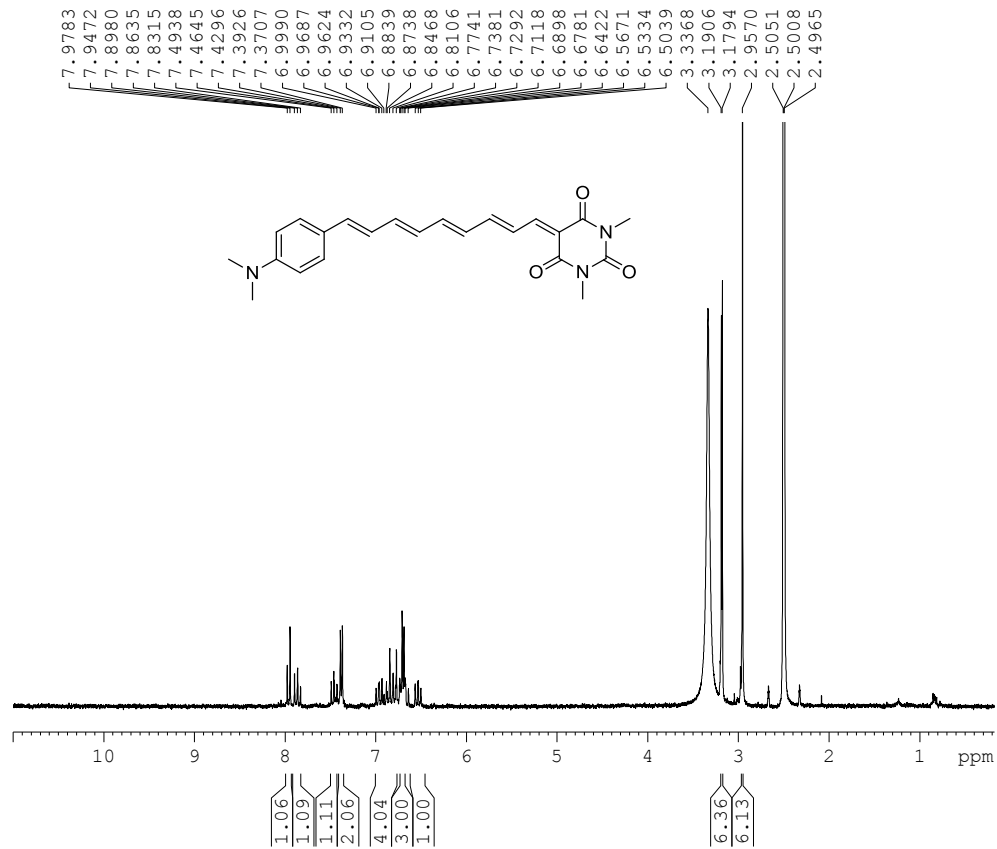


NAME ZKX-M-4-C-20140830
EXPNO 1
PROCNO 1
Date_ 20140830
Time 21.35
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT cdcl3
NS 12000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 300.6 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 8.50 usec
PL1 -2.00 dB
PL1W 57.32743073 W
SF01 100.6328888 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -1.00 dB
PL12 14.26 dB
PL13 14.46 dB
PL2W 13.18669796 W
PL12W 0.39276794 W
PL13W 0.37509048 W
SF02 400.1716007 MHz
SI 32768
SF 100.6228236 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

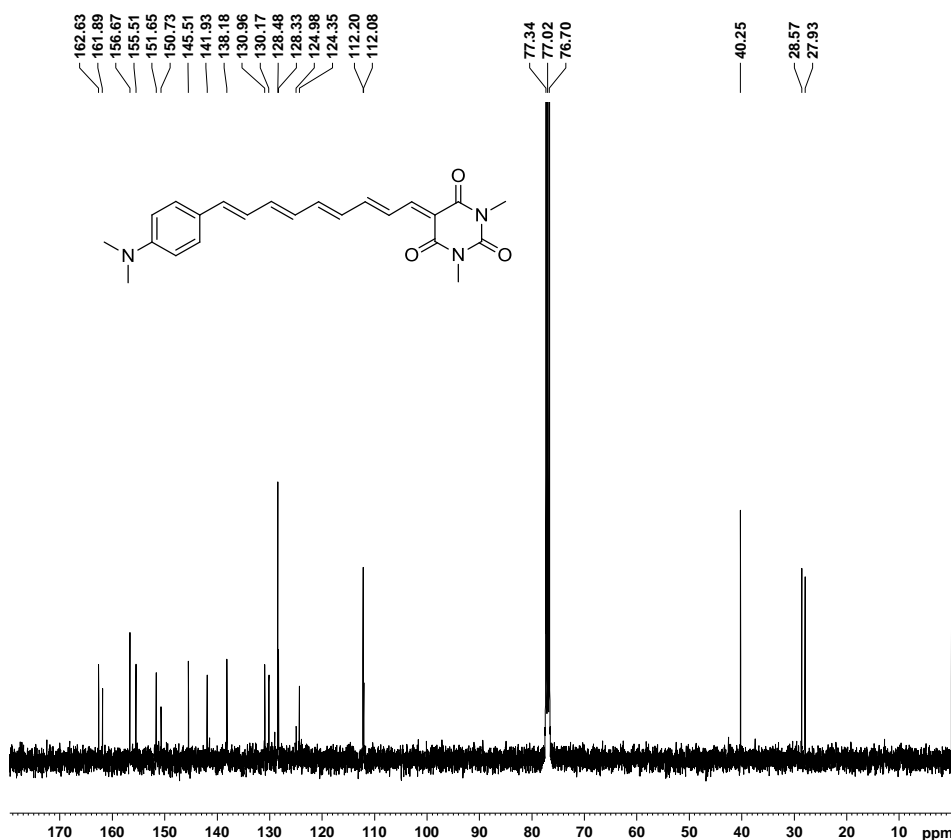
¹³C NMR (CDCl₃) spectrum of BBTOM-4



NAME ZKX-BBATO-M--S-2
EXPNO 1
PROCNO 1
Date_ 20140419
Time 9.03
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 203
DW 60.800 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 13.80 usec
PL1 -1.00 dB
PL1W 13.18669796 W
SF01 400.1724712 MHz
SI 32768
SF 400.1699940 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹H NMR (CDCl₃) spectrum of BBTOM-5



```

NAME      ZKX-BBATO-M-5-C
EXPNO    1
PROCNO   1
Date_    20140429
Time     22.24
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       1024
DS       4
SWH      24038.461 Hz
FIDRES   0.366798 Hz
AQ       1.3631988 sec
RG       203
DW       20.800 usec
DE       6.50 usec
TE       301.3 K
D1       2.00000000 sec
D11      0.03000000 sec
TD0     1
    
```

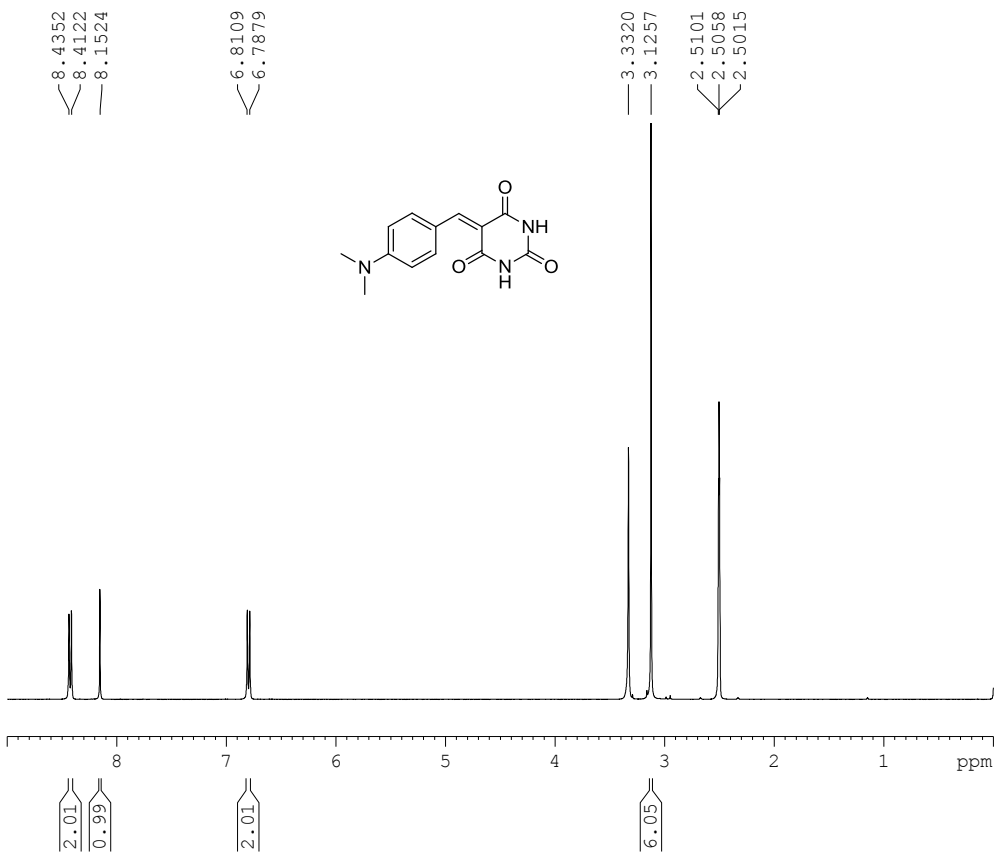
```

===== CHANNEL f1 =====
NUC1     13C
P1       8.50 usec
PL1      -2.00 dB
PL1W    57.32743073 W
SFO1    100.6328888 MHz
    
```

```

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2     1H
PCPD2    80.00 usec
PL2      -1.00 dB
PL12     14.26 dB
PL13     14.46 dB
PL2W     13.18669796 W
PL12W    0.39276794 W
PL13W    0.37509048 W
SFO2    400.1716007 MHz
SI       32768
SF       100.6228238 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
    
```

¹³C NMR (CDCl₃) spectrum of BBTOM-5



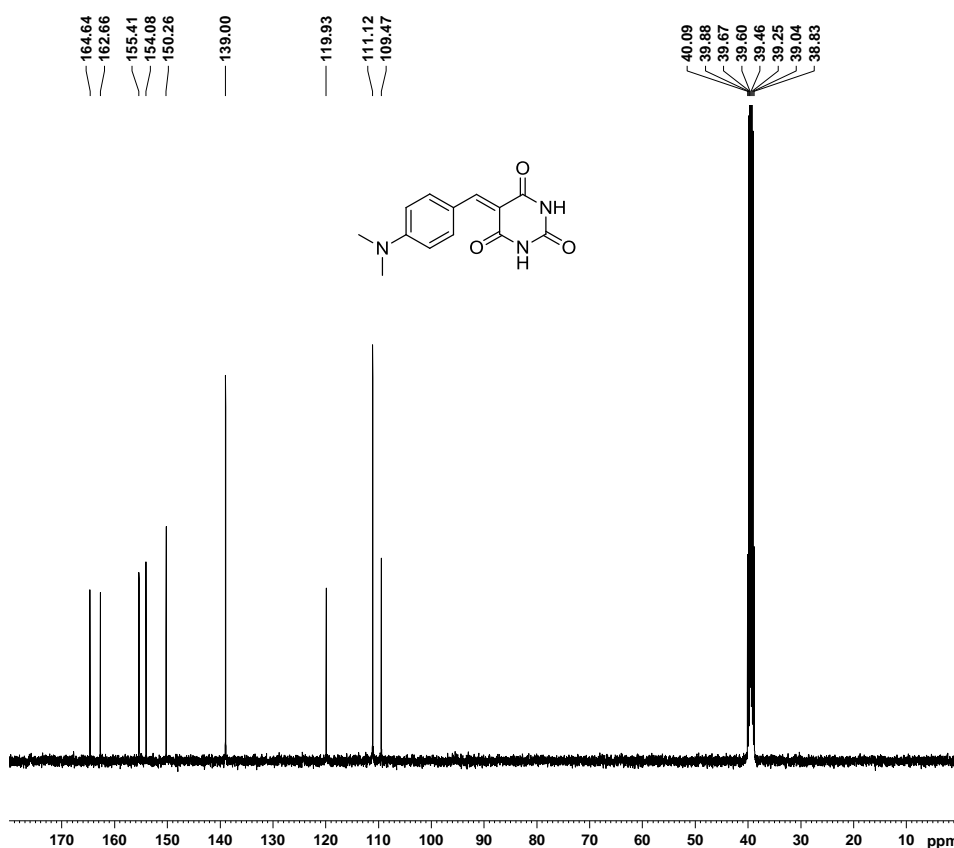
```

NAME      BBAT-1-NMe2
EXPNO    1
PROCNO   1
Date_    20110615
Time     17.03
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zg30
TD       65536
SOLVENT  DMSO
NS       16
DS       2
SWH      8223.685 Hz
FIDRES   0.125483 Hz
AQ       3.9846387 sec
RG       203
DW       60.800 usec
DE       6.50 usec
TE       299.4 K
D1       1.00000000 sec
TD0     1
    
```

```

===== CHANNEL f1 =====
NUC1     1H
P1       14.20 usec
PL1      -1.00 dB
PL1W    13.18669796 W
SFO1    400.1724712 MHz
SI       32768
SF       400.1699932 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
```

¹H NMR (DMSO-d₆) spectrum of BBTO-1



```

NAME      2KX-BBTO-1-C13
EXPNO     1
PROCNO    1
Date_     20150313
Time      23.03
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zgpg30
TD        65536
SOLVENT   DMSO
NS        4
DS        4
SWH       24038.461 Hz
FIDRES    0.366798 Hz
AQ        1.3631988 sec
RG        203
DW        20.800 usec
DE        6.50 usec
TE        296.7 K
D1        2.00000000 sec
D11       0.03000000 sec
TD0       1
    
```

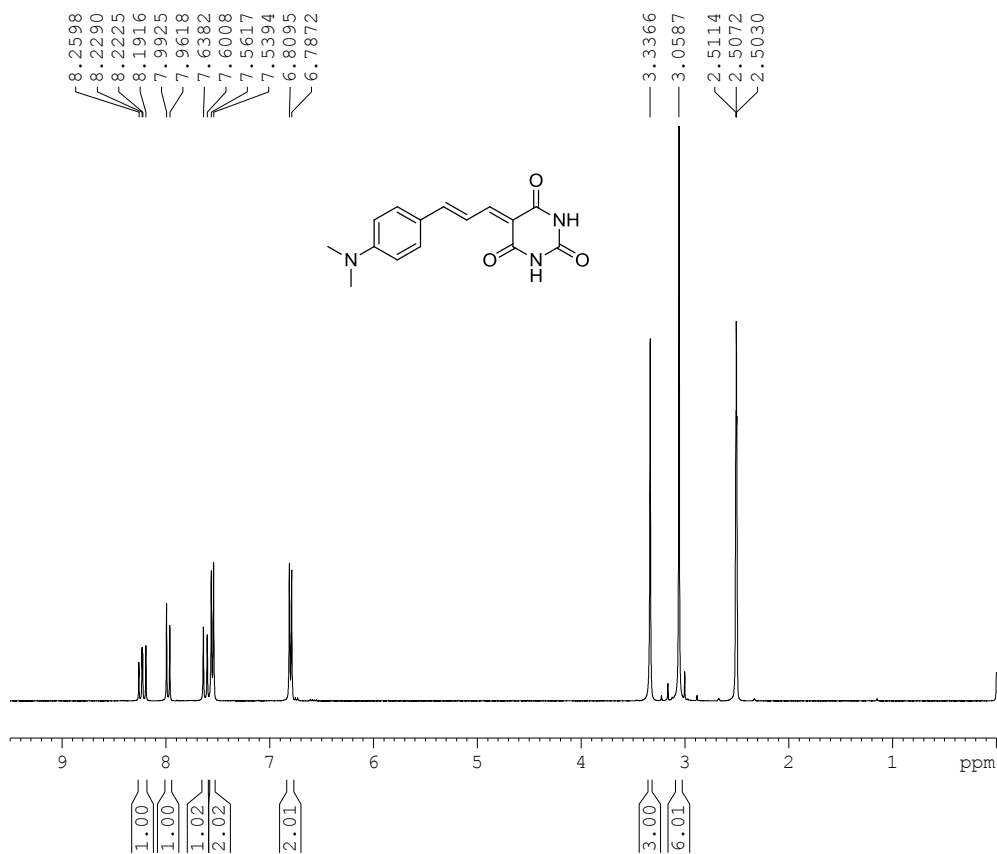
```

===== CHANNEL f1 =====
NUC1      13C
P1        8.50 usec
PL1       -2.00 dB
PL1W     57.32743073 W
SFO1     100.6328888 MHz
    
```

```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2       -1.00 dB
PL12      14.26 dB
PL13      14.46 dB
PL2W     13.18669796 W
PL12W    0.39276794 W
PL13W    0.37509048 W
SFO2     400.1716007 MHz
SI        32768
SF        100.6228773 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
    
```

¹³C NMR (DMSO-d₆) spectrum of BBTO-1



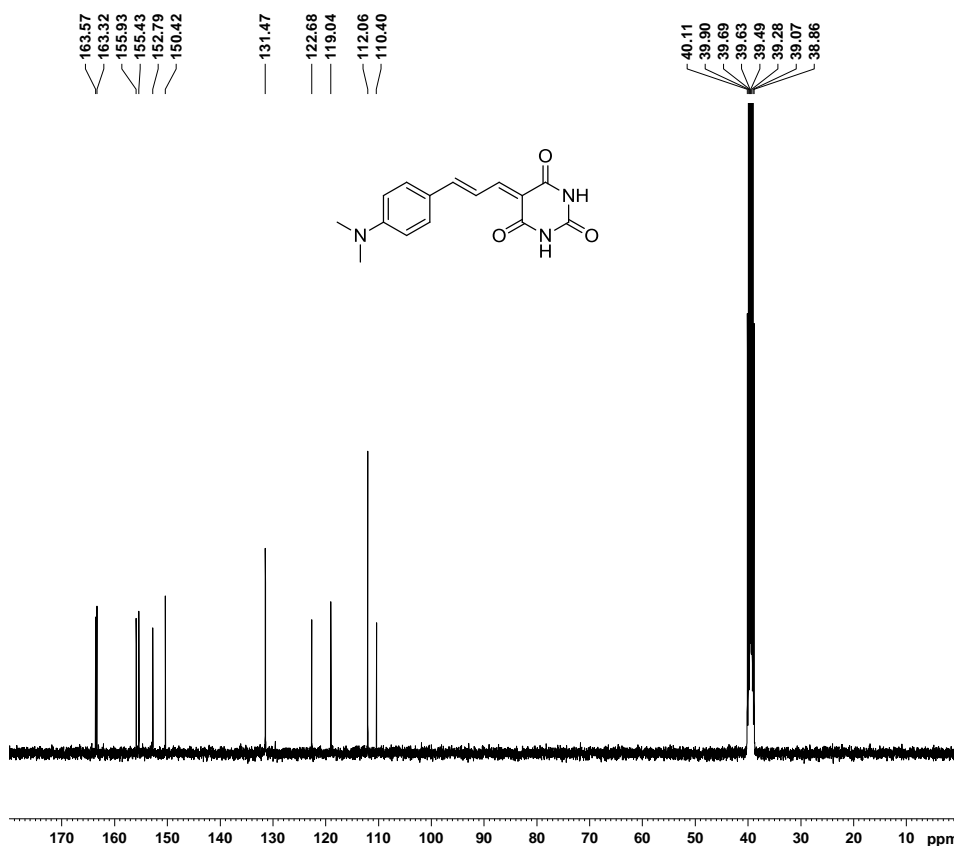
```

NAME      BBAT-2-NMe2
EXPNO     1
PROCNO    1
Date_     20110615
Time      17.07
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zg30
TD        65536
SOLVENT   DMSO
NS        16
DS        2
SWH       8223.685 Hz
FIDRES    0.125483 Hz
AQ        3.9846387 sec
RG        203
DW        60.800 usec
DE        6.50 usec
TE        299.3 K
D1        1.00000000 sec
TD0       1
    
```

```

===== CHANNEL f1 =====
NUC1      1H
P1        14.20 usec
PL1       -1.00 dB
PL1W     13.18669796 W
SFO1     400.1724712 MHz
SI        32768
SF        400.1699925 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
    
```

¹H NMR (DMSO-d₆) spectrum of BBTO-2



```

NAME      2KX-BBTO-2-C13
EXPNO     1
PROCNO    1
Date_     20150311
Time      14.36
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zgpg30
TD        65536
SOLVENT   DMSO
NS        1746
DS        4
SWH       24038.461 Hz
FIDRES    0.366798 Hz
AQ        1.3631988 sec
RG        203
DW        20.800 usec
DE        6.50 usec
TE        298.6 K
D1        2.00000000 sec
D11       0.03000000 sec
TD0       1
    
```

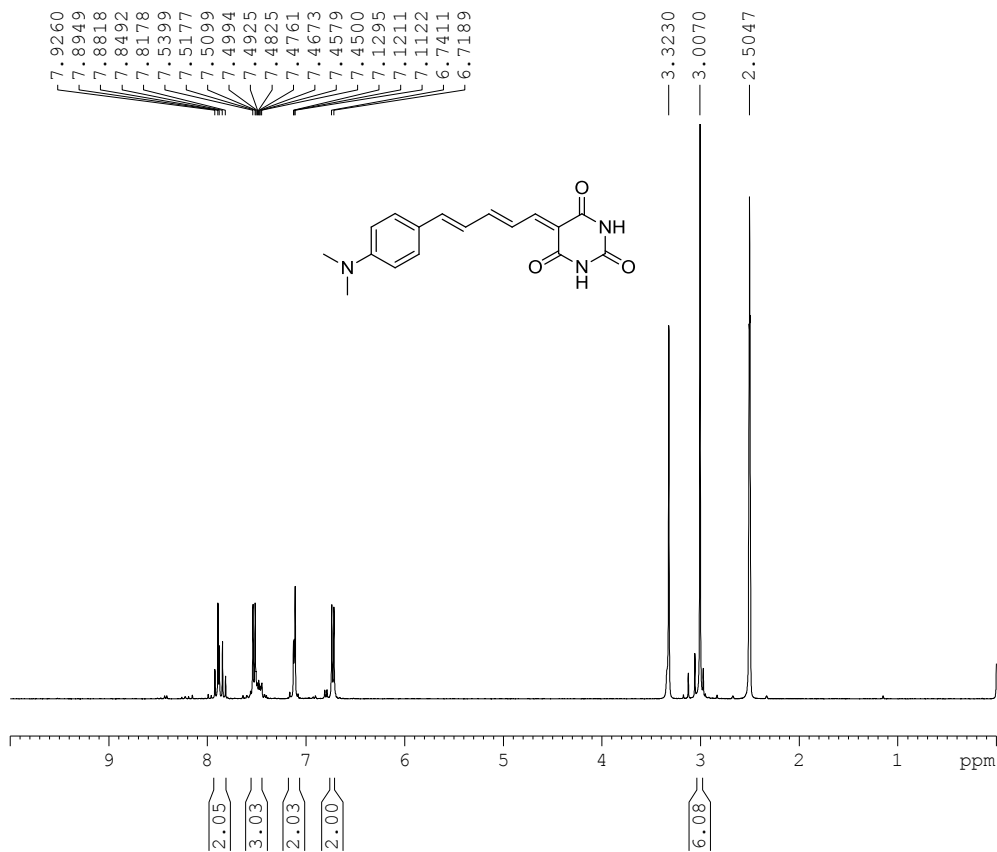
```

===== CHANNEL f1 =====
NUC1      13C
P1        8.50 usec
PL1       -2.00 dB
PL1W     57.32743073 W
SFO1     100.6328888 MHz
    
```

```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2       -1.00 dB
PL12      14.26 dB
PL13      14.46 dB
PL2W     13.18669796 W
PL12W    0.39276794 W
PL13W    0.37509048 W
SFO2     400.1716007 MHz
SI        32768
SF        100.6228773 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
    
```

¹³C NMR (DMSO-d₆) spectrum of BBTO-2



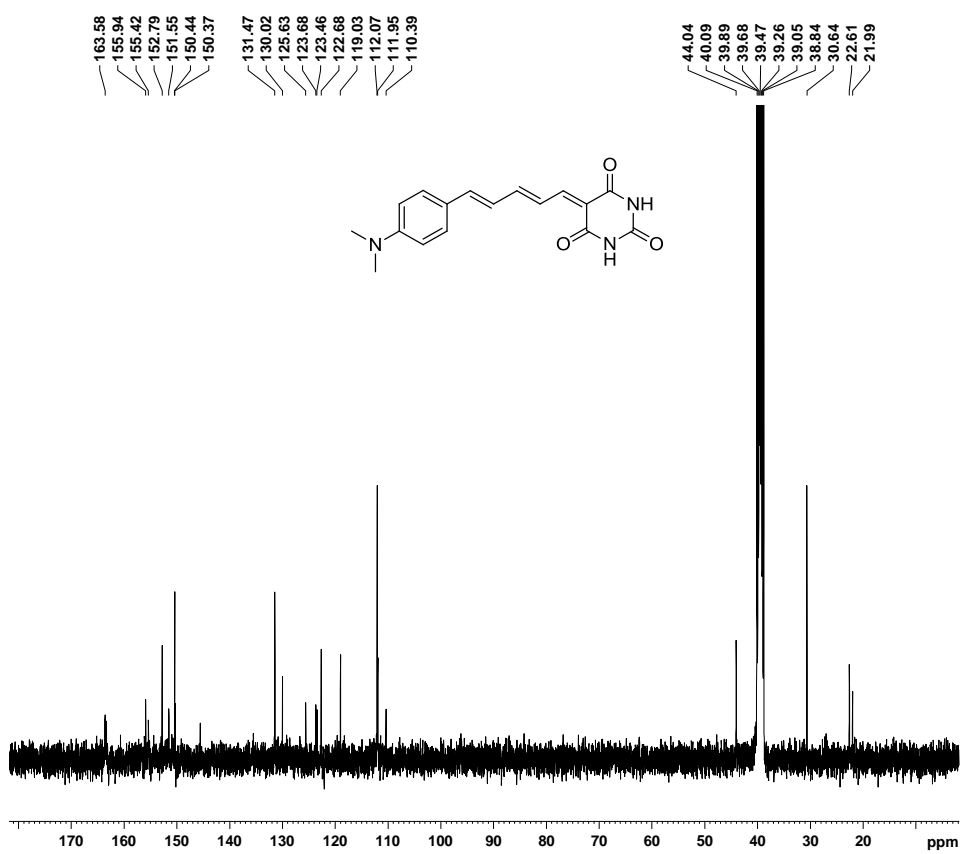
```

NAME      BBAT-3-NMe2
EXPNO     1
PROCNO    1
Date_     20110617
Time      11.19
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zg30
TD        65536
SOLVENT   DMSO
NS        16
DS        2
SWH       8223.685 Hz
FIDRES    0.125483 Hz
AQ        3.9846387 sec
RG        203
DW        60.800 usec
DE        6.50 usec
TE        300.0 K
D1        1.00000000 sec
TD0       1
    
```

```

===== CHANNEL f1 =====
NUC1      1H
P1        14.20 usec
PL1       -1.00 dB
PL1W     13.18669796 W
SFO1     400.1724712 MHz
SI        32768
SF        400.1699938 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
    
```

¹H NMR (DMSO-d₆) spectrum of BBTO-3



```

NAME      ZKX-BBTO-3-C13
EXPNO    1
PROCNO   1
Date_    20150314
Time     1.29
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zgpg30
TD       65536
SOLVENT  DMSO
NS       2500
DS       4
SWH      24038.461 Hz
FIDRES   0.366798 Hz
AQ       1.3631988 sec
RG       203
DW       20.800 usec
DE       6.50 usec
TE       296.3 K
D1       2.00000000 sec
D11      0.03000000 sec
TD0     1
    
```

```

===== CHANNEL f1 =====
NUC1     13C
P1       8.50 usec
PL1     -2.00 dB
PL1W    57.32743073 W
SFO1    100.6328888 MHz

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2     1H
PCPD2   80.00 usec
PL2     -1.00 dB
PL12    14.26 dB
PL13    14.46 dB
PL2W    13.18669796 W
PL12W   0.39276794 W
PL13W   0.37509048 W
SFO2    400.1716007 MHz
SI      32768
SF      100.6228773 MHz
WDW     EM
SSB     0
LB      1.00 Hz
GB      0
PC      1.40
    
```

¹³C NMR (DMSO-d₆) spectrum of BBTO-3



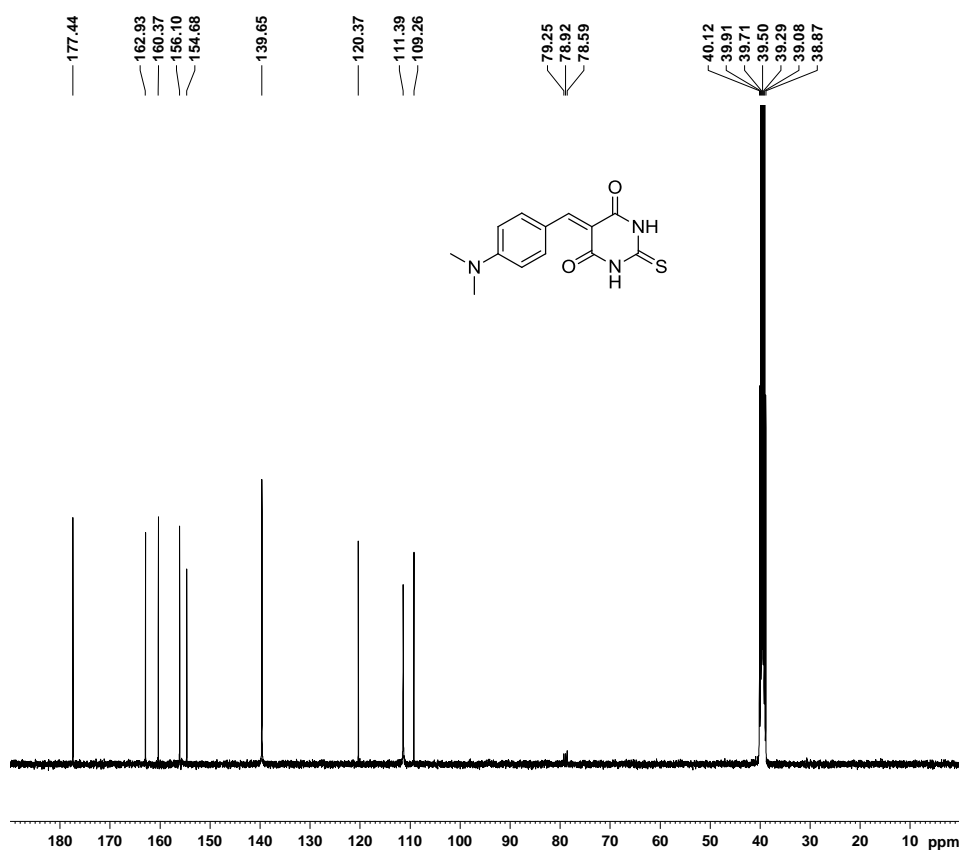
```

NAME      BBATS-1
EXPNO    1
PROCNO   1
Date_    20110818
Time     15.49
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zg30
TD       65536
SOLVENT  DMSO
NS       16
DS       2
SWH      8223.685 Hz
FIDRES   0.125483 Hz
AQ       3.9846387 sec
RG       203
DW       60.800 usec
DE       6.50 usec
TE       300.1 K
D1       1.00000000 sec
TD0     1
    
```

```

===== CHANNEL f1 =====
NUC1     1H
P1       13.80 usec
PL1     -1.00 dB
PL1W    13.18669796 W
SFO1    400.1724712 MHz
SI      32768
SF      400.1699937 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00
    
```

¹H NMR (DMSO-d₆) spectrum of BBTS-1



```

NAME      BBATS-1-C13
EXPNO     1
PROCNO    1
Date_     20110901
Time      14.06
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zgpg30
TD        65536
SOLVENT   DMSO
NS        3639
DS        4
SWH       24038.461 Hz
FIDRES    0.366798 Hz
AQ        1.3631988 sec
RG        203
DW        20.800 usec
DE        6.50 usec
TE        299.8 K
D1        2.00000000 sec
D11       0.03000000 sec
TD0       1
    
```

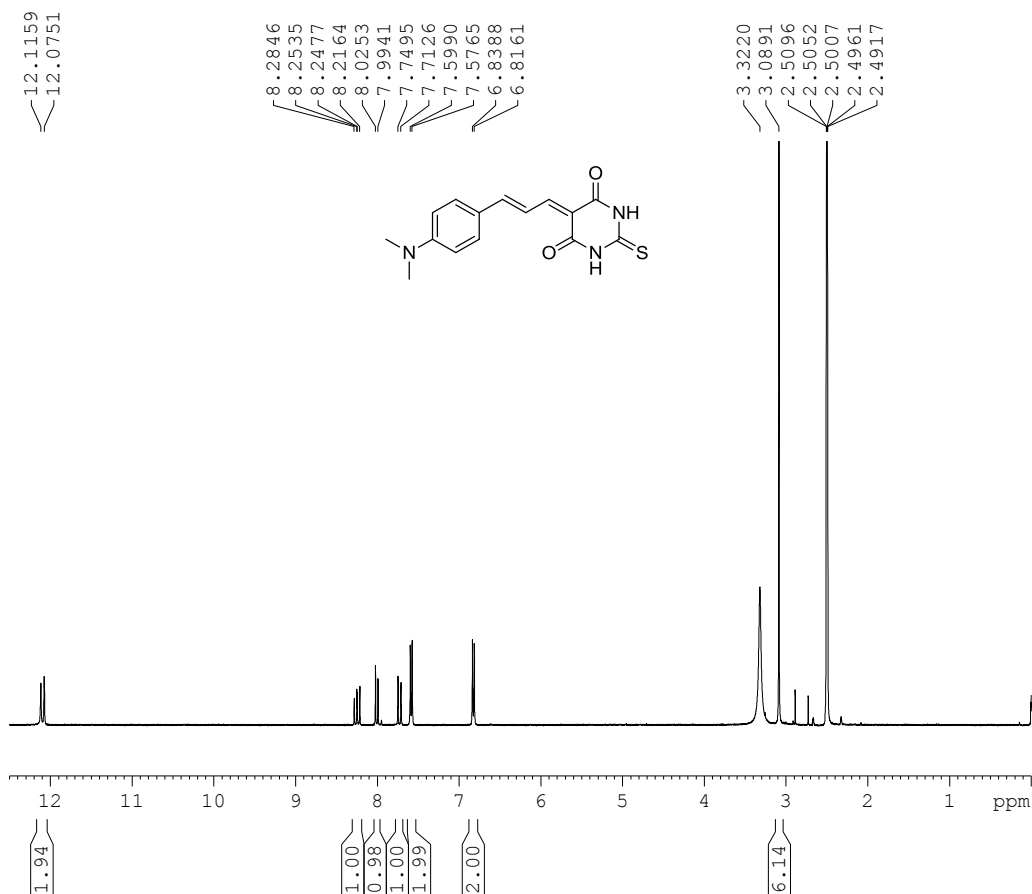
```

===== CHANNEL f1 =====
NUC1      13C
P1        8.50 usec
PL1       -2.00 dB
PL1W      57.32743073 W
SFO1      100.6328888 MHz
    
```

```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2       -1.00 dB
PL12      14.26 dB
PL13      14.46 dB
PL2W      13.18669796 W
PL12W     0.39276794 W
PL13W     0.37509048 W
SFO2      400.1716007 MHz
SI        32768
SF        100.6228773 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
    
```

¹³C NMR (DMSO-d₆) spectrum of BBTS-1



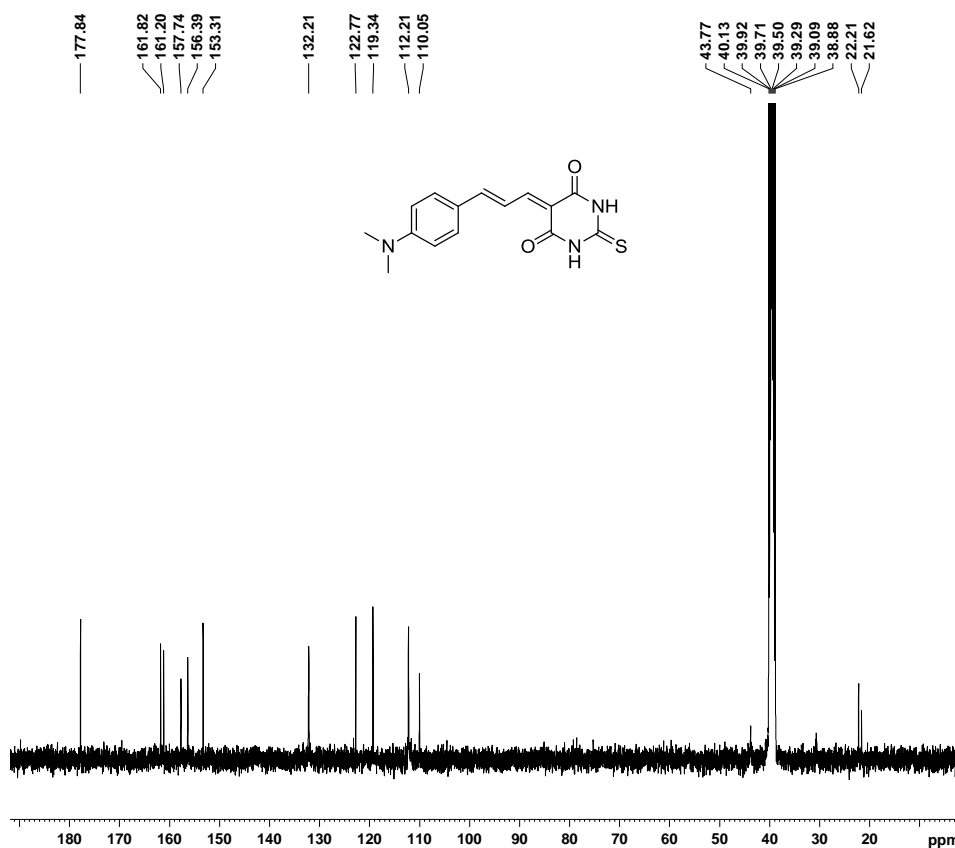
```

NAME      ZKX-BBTS-2-20150518
EXPNO     1
PROCNO    1
Date_     20150518
Time      19.04
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zg30
TD        65536
SOLVENT   DMSO
NS        16
DS        2
SWH       8223.685 Hz
FIDRES    0.125483 Hz
AQ        3.9846387 sec
RG        203
DW        60.800 usec
DE        6.50 usec
TE        298.1 K
D1        1.00000000 sec
TD0       1
    
```

```

===== CHANNEL f1 =====
NUC1      1H
P1        13.80 usec
PL1       -1.00 dB
PL1W      13.18669796 W
SFO1      400.1724712 MHz
SI        32768
SF        400.1699949 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
    
```

¹H NMR (DMSO-d₆) spectrum of BBTS-2



```

NAME      BBATS-2-C13
EXPNO    1
PROCNO   1
Date_    20110831
Time     21.50
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zgpg30
TD       65536
SOLVENT  DMSO
NS       6000
DS       4
SWH      24038.461 Hz
FIDRES   0.366798 Hz
AQ       1.3631988 sec
RG       203
DW       20.800 usec
DE       6.50 usec
TE       301.3 K
D1       2.00000000 sec
D11      0.03000000 sec
TD0     1
    
```

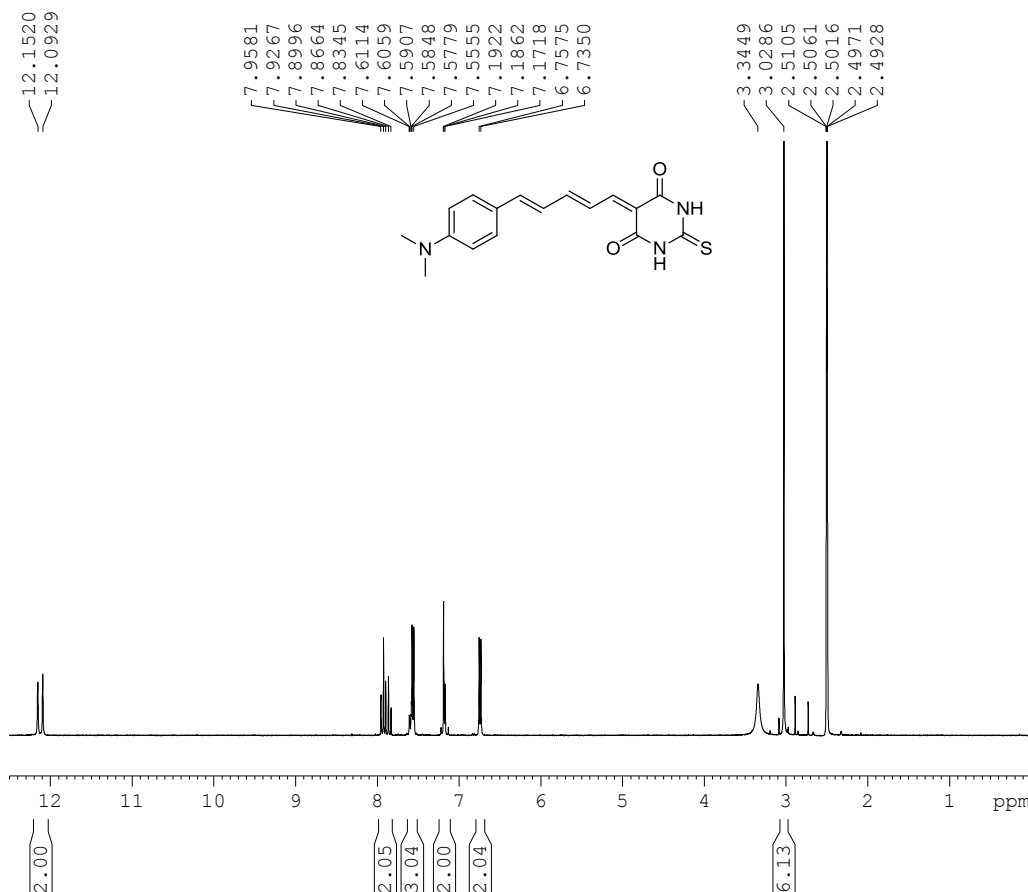
```

===== CHANNEL f1 =====
NUC1     13C
P1       8.50 usec
PL1      -2.00 dB
PL1W    57.32743073 W
SFO1    100.6328888 MHz
    
```

```

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2     1H
PCPD2    80.00 usec
PL2      -1.00 dB
PL12     14.26 dB
PL13     14.46 dB
PL2W    13.18669796 W
PL12W   0.39276794 W
PL13W   0.37509048 W
SFO2    400.1716007 MHz
SI       32768
SF       100.6228773 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
    
```

¹³C NMR (DMSO-d₆) spectrum of BBTS-2



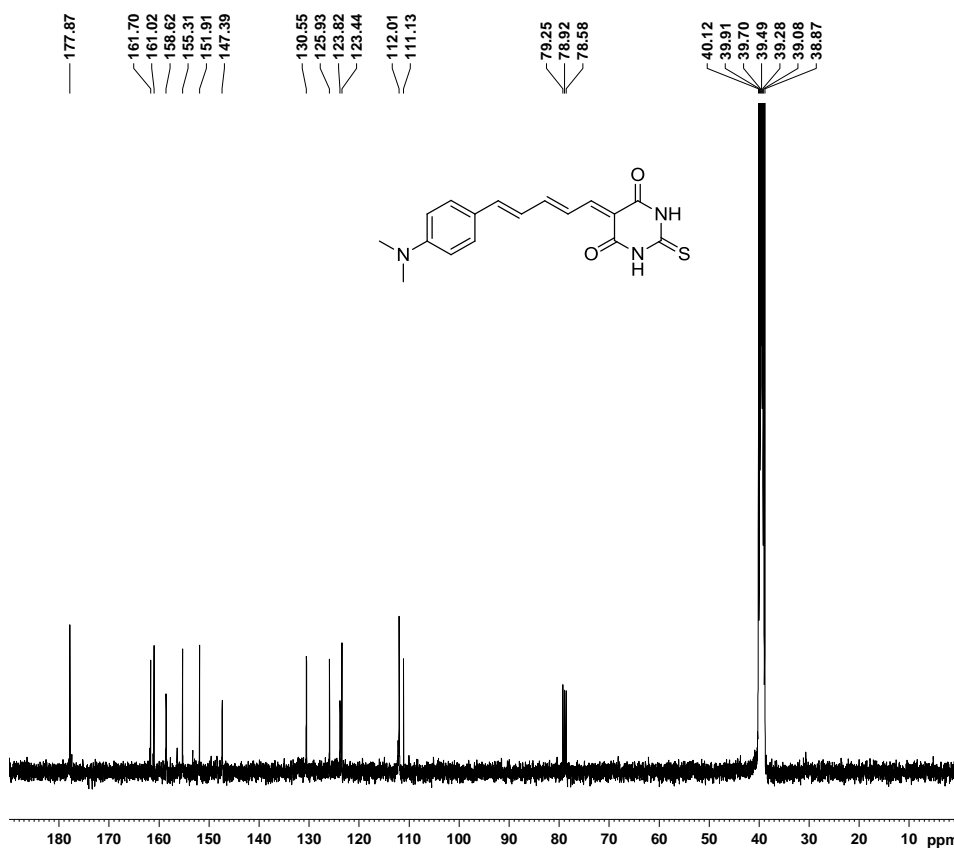
```

NAME      ZKX-BBTS-3-20150518
EXPNO    1
PROCNO   1
Date_    20150518
Time     19.08
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zg30
TD       65536
SOLVENT  DMSO
NS       16
DS       2
SWH      8223.685 Hz
FIDRES   0.125483 Hz
AQ       3.9846387 sec
RG       203
DW       60.800 usec
DE       6.50 usec
TE       297.8 K
D1       1.00000000 sec
TD0     1
    
```

```

===== CHANNEL f1 =====
NUC1     1H
P1       13.80 usec
PL1      -1.00 dB
PL1W    13.18669796 W
SFO1    400.1724712 MHz
SI       32768
SF       400.1699945 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
```

¹H NMR (DMSO-d₆) spectrum of BBTS-3



```

NAME      BBATS-3-C13
EXPNO     1
PROCNO    1
Date_     20110830
Time      6.40
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zgpg30
TD        65536
SOLVENT   DMSO
NS        8000
DS         4
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 sec
RG         203
DW         20.800 usec
DE         6.50 usec
TE         297.0 K
D1         2.00000000 sec
D11        0.03000000 sec
TD0        1
    
```

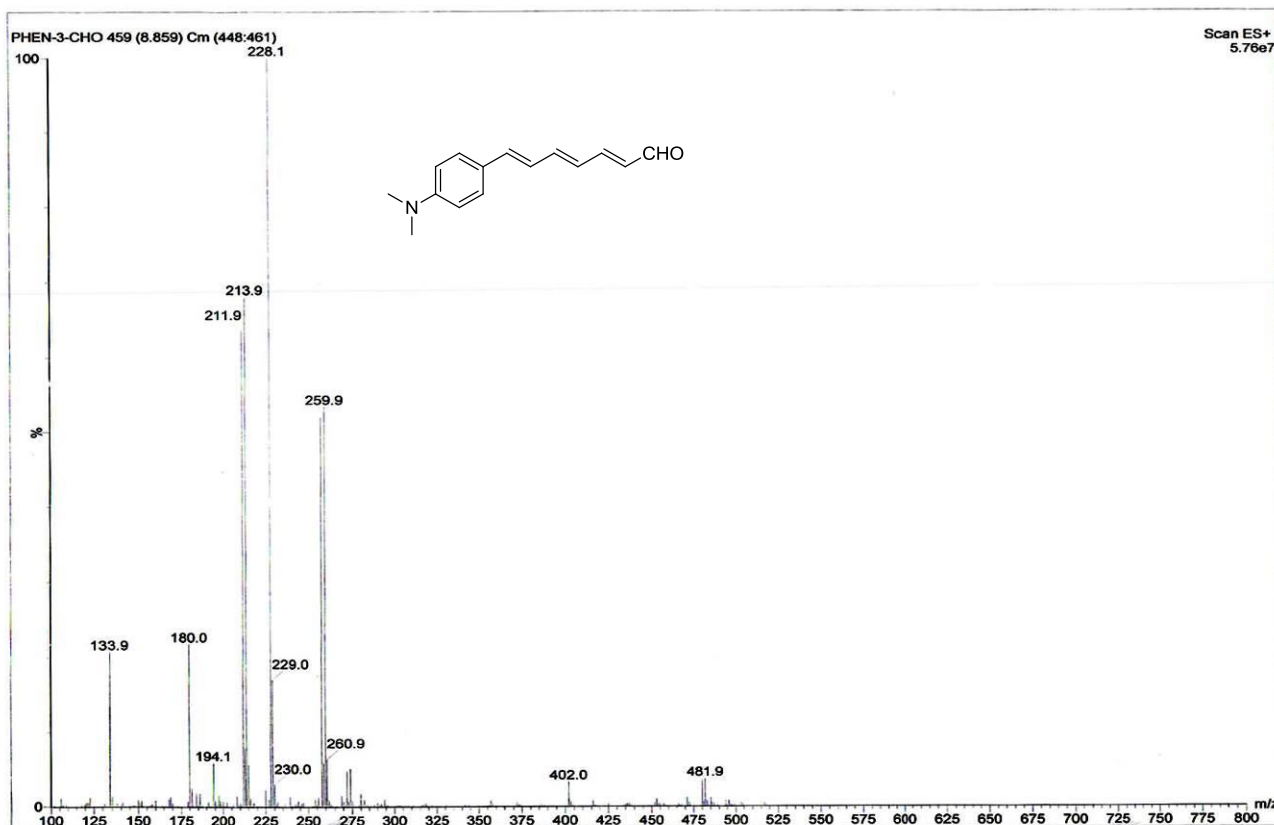
```

===== CHANNEL f1 =====
NUC1      13C
P1        8.50 usec
PL1       -2.00 dB
PL1W      57.32743073 W
SFO1      100.6328888 MHz
    
```

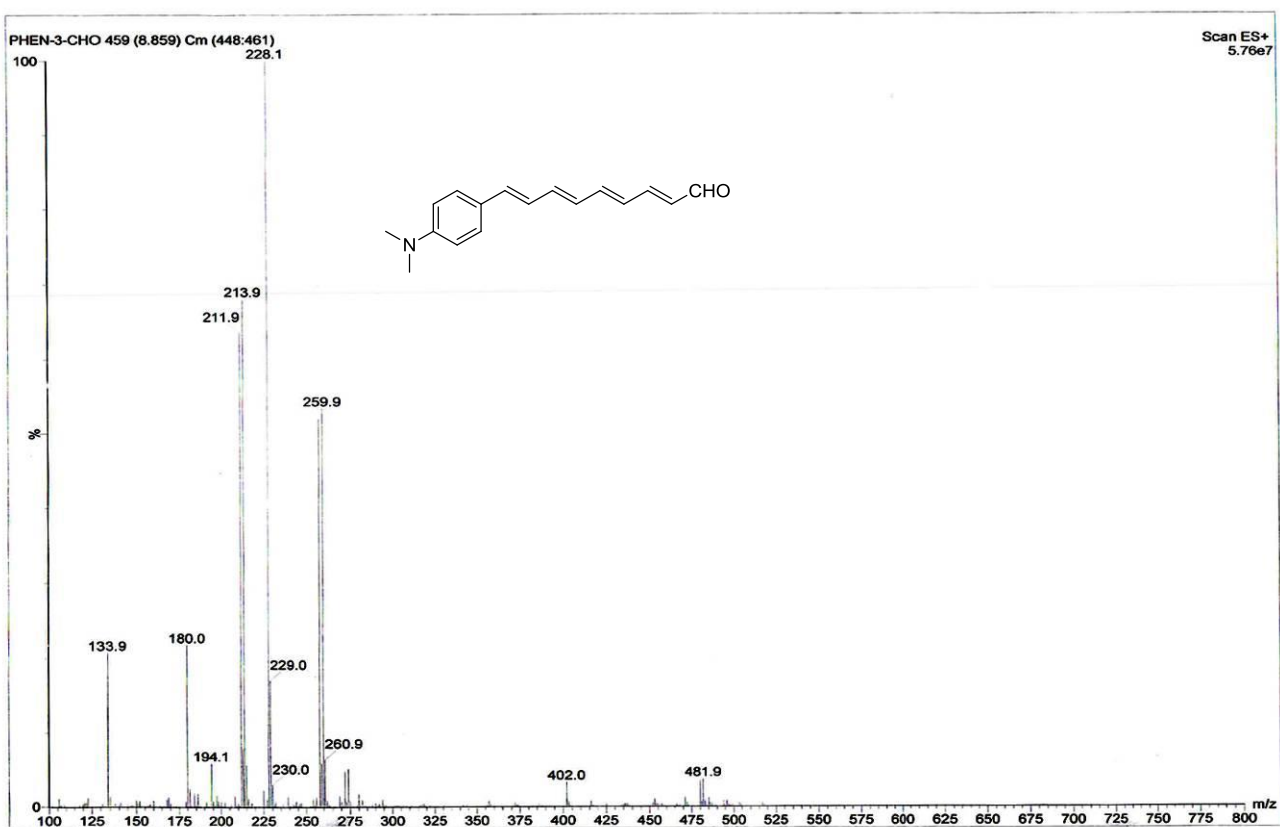
```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     80.00 usec
PL2       -1.00 dB
PL12      14.26 dB
PL13      14.46 dB
PL2W      13.18669796 W
PL12W     0.39276794 W
PL13W     0.37509048 W
SFO2      400.1716007 MHz
SI         32768
SF         100.6228773 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40
    
```

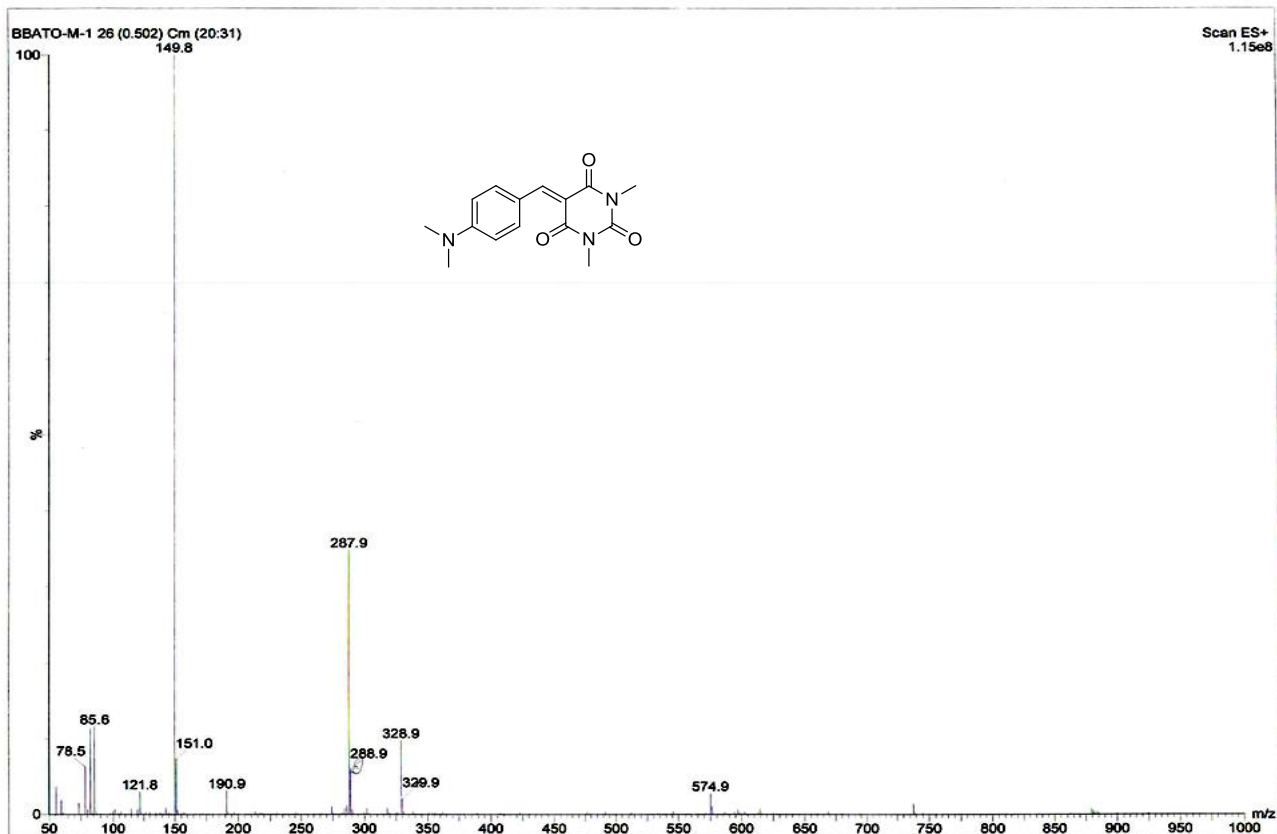
¹³C NMR (DMSO-d₆) spectrum of BBTS-3



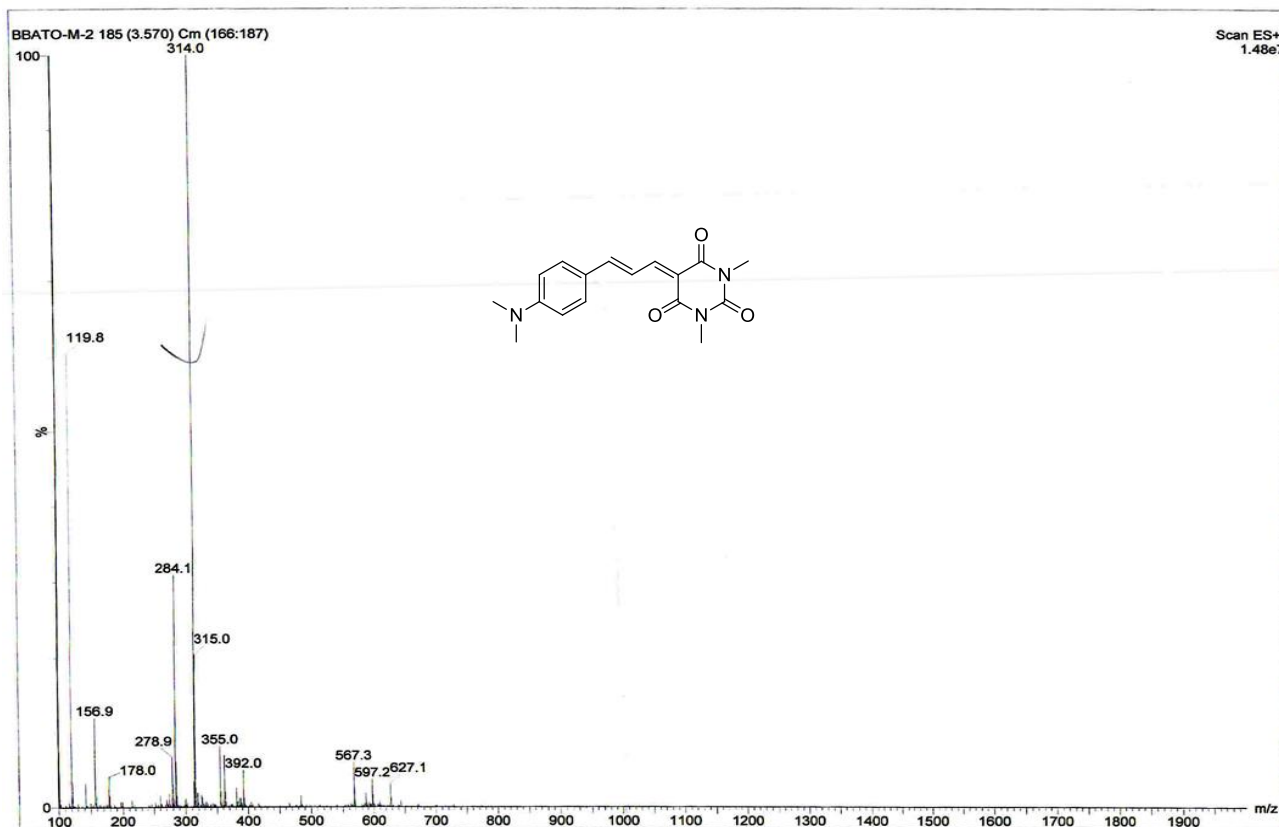
MS spectrum of compound 2



MS spectrum of compound 3



MS spectrum of BBTOM-1



MS spectrum of BBTOM-2

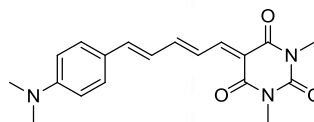
Elemental Composition Report

Single Mass Analysis

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 2



Monoisotopic Mass, Even Electron Ions

205 formula(e) evaluated with 1 results within limits (up to 100 best isotopic matches for each mass)

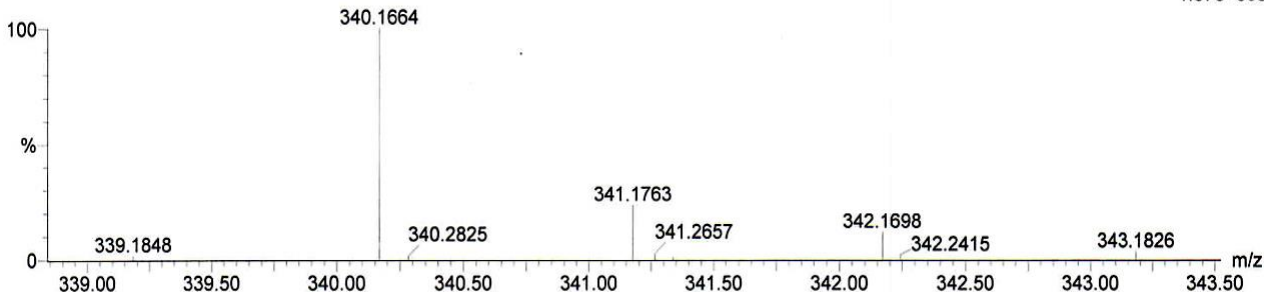
Elements Used:

C: 0-30 H: 0-30 N: 0-10 O: 0-6

BBATO-M-3 6 (0.102)

TOF MS ES+

1.57e+003



Minimum:

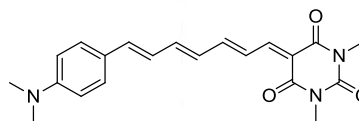
Maximum: 5.0 3.0 -1.5 100.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
340.1664	340.1661	0.3	0.9	10.5	0.6	C19 H22 N3 O3

Elemental Composition Report

Single Mass Analysis

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 100.0
 Element prediction: Off
 Number of isotope peaks used for i-FIT = 2



Monoisotopic Mass, Even Electron Ions

206 formula(e) evaluated with 1 results within limits (up to 100 best isotopic matches for each mass)

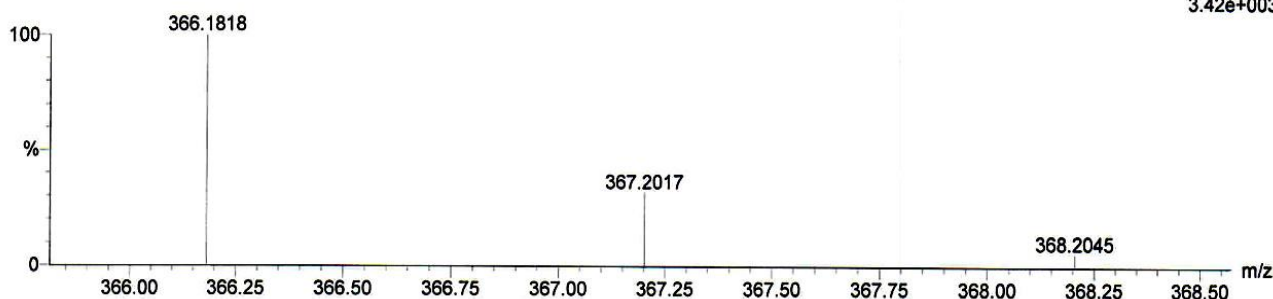
Elements Used:

C: 0-30 H: 0-30 N: 0-10 O: 0-6

BBATO-M-4 2 (0.034)

TOF MS ES+

3.42e+003



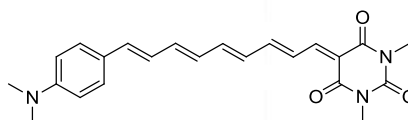
Minimum: -1.5
 Maximum: 5.0 3.0 100.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
366.1818	366.1818	0.0	0.0	11.5	25.7	C21 H24 N3 O3

Elemental Composition Report

Single Mass Analysis

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 100.0
 Element prediction: Off
 Number of isotope peaks used for i-FIT = 2



Monoisotopic Mass, Even Electron Ions

202 formula(e) evaluated with 1 results within limits (up to 100 best isotopic matches for each mass)

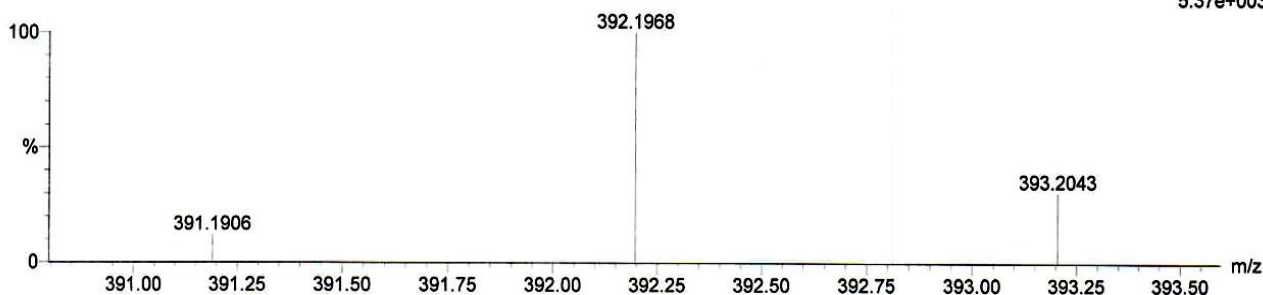
Elements Used:

C: 0-30 H: 0-30 N: 0-10 O: 0-6

BBATO-M-5 10 (0.170)

TOF MS ES+

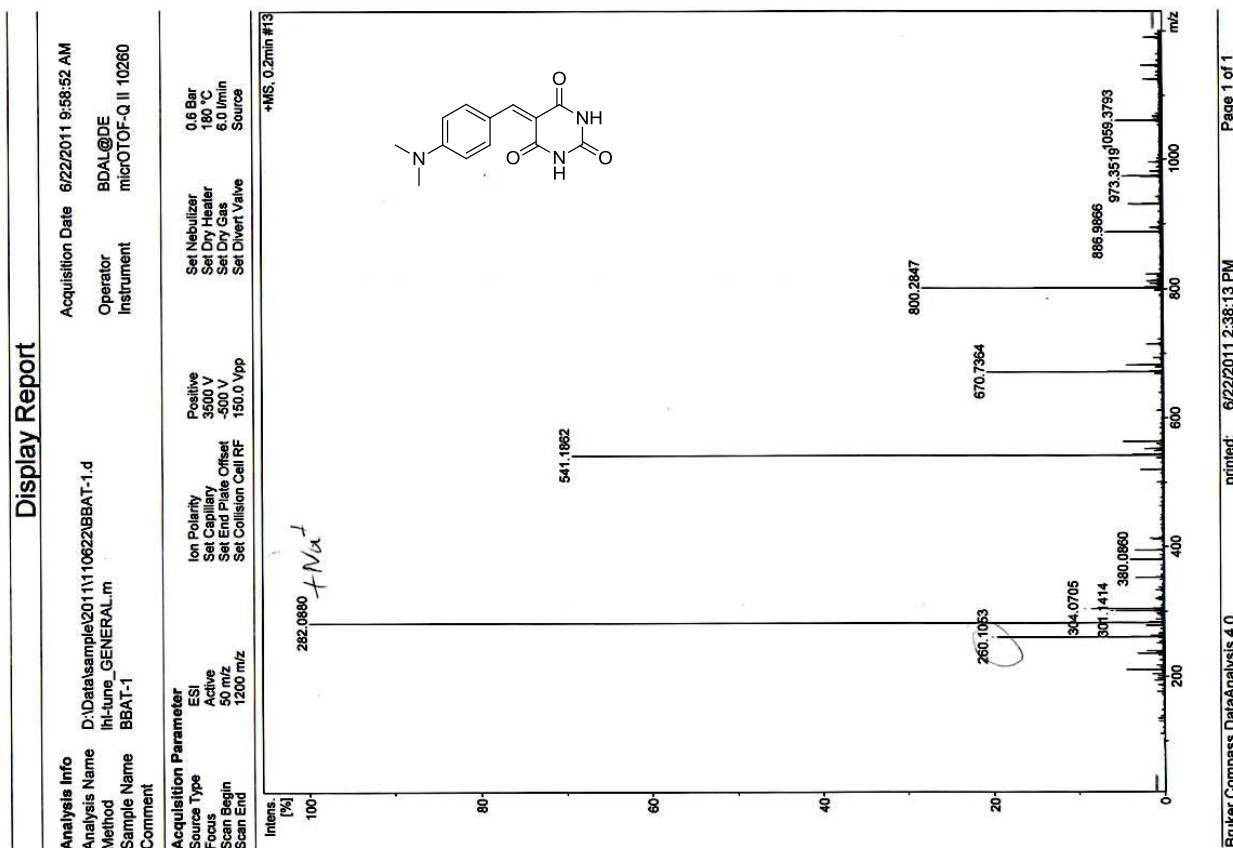
5.37e+003



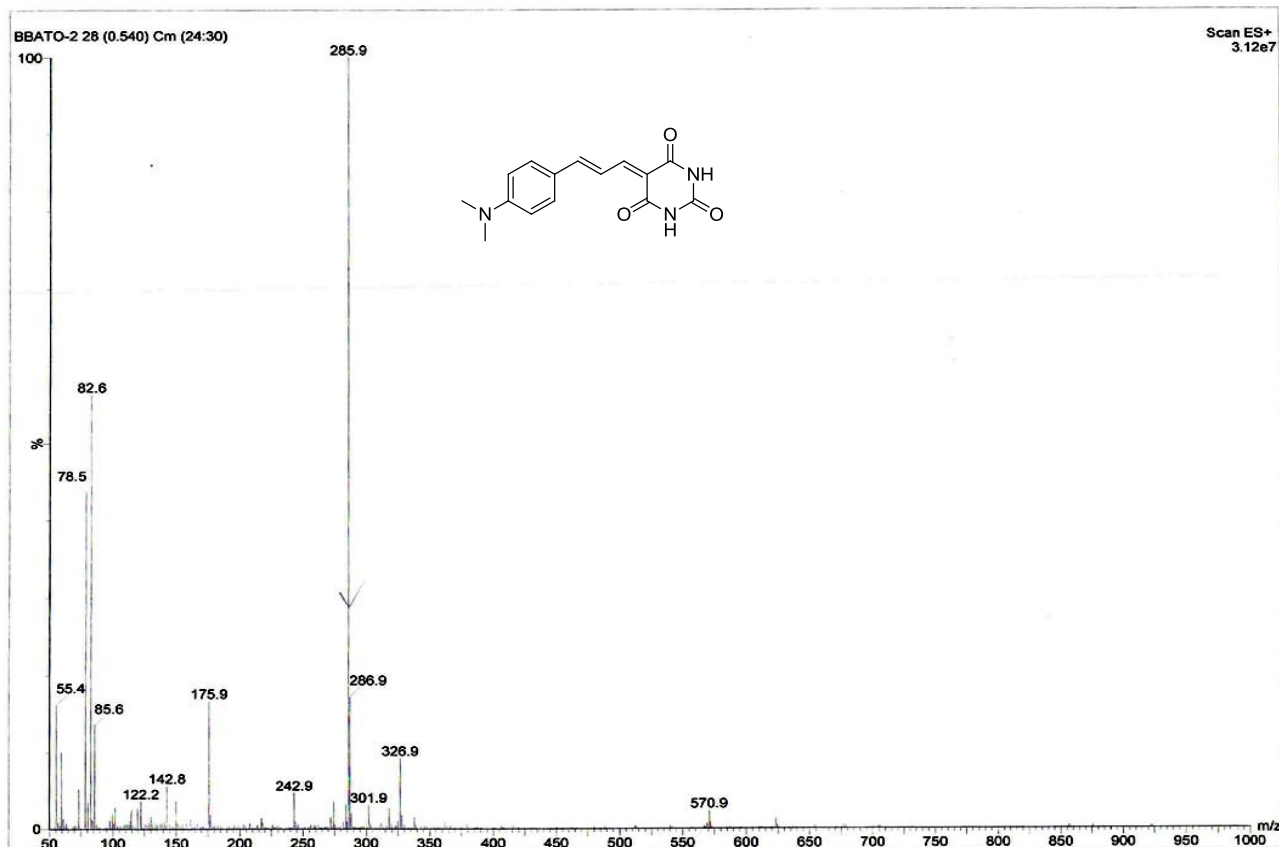
Minimum: -1.5
 Maximum: 5.0 3.0 100.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
392.1968	392.1974	-0.6	-1.5	12.5	8.4	C23 H26 N3 O3

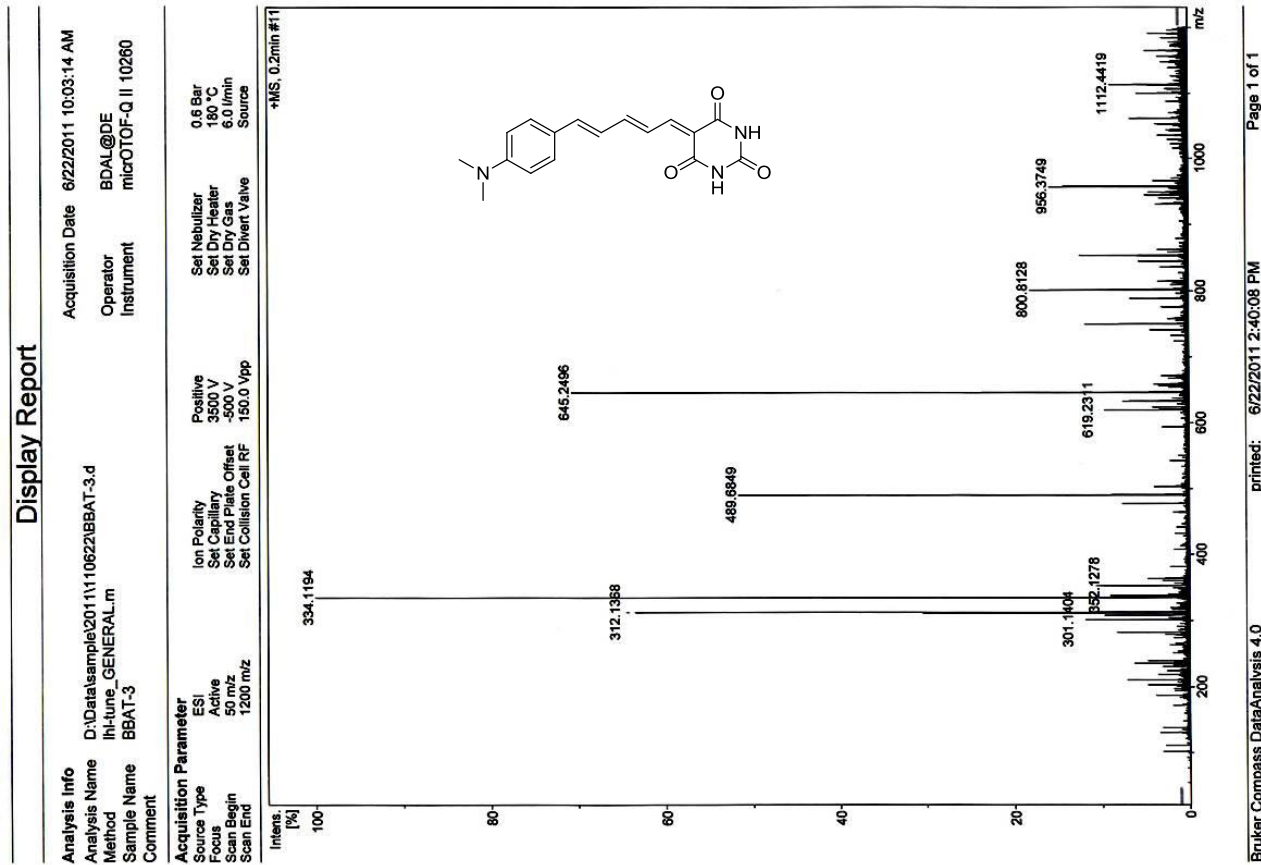
HRMS spectrum of BBTOM-5


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 Bruker Compass DataAnalysis 4.0
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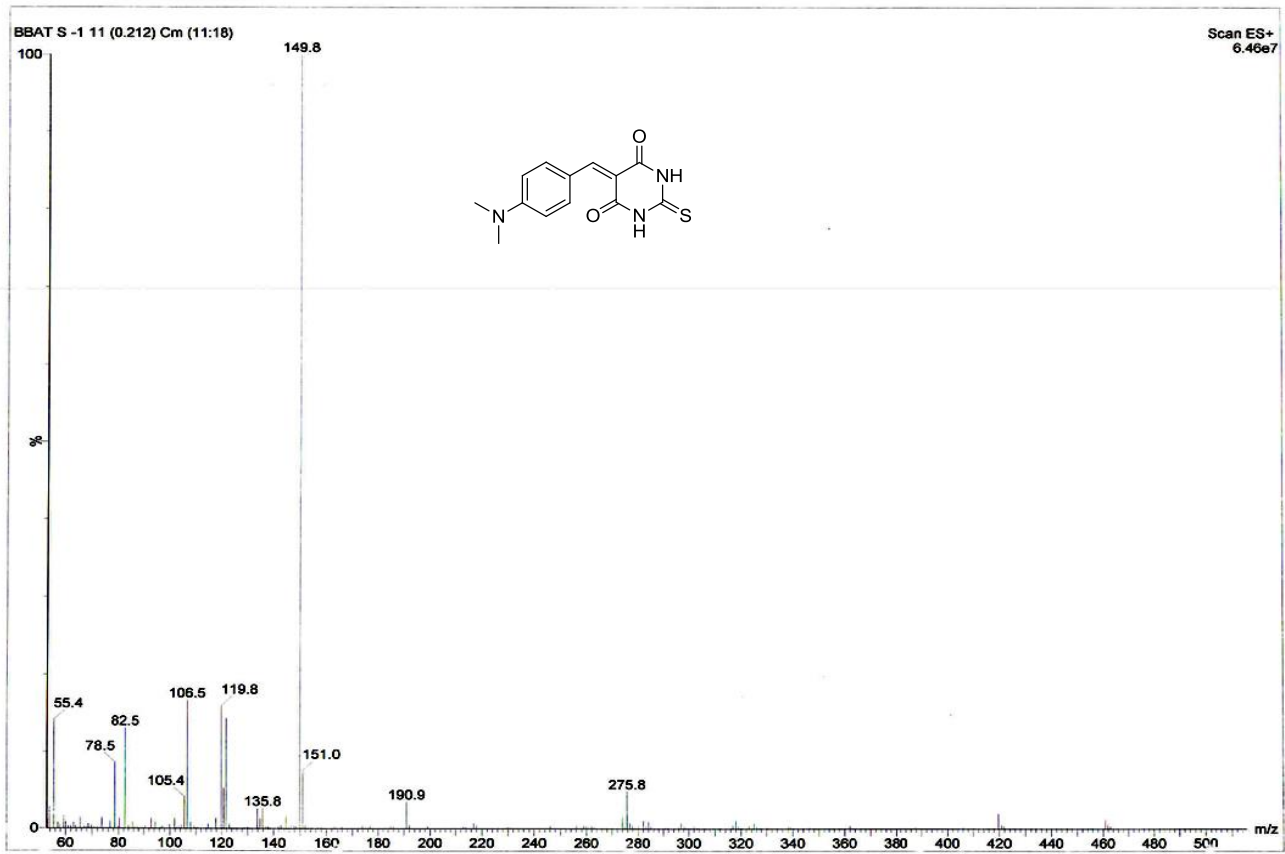
MS spectrum of BBTO-1



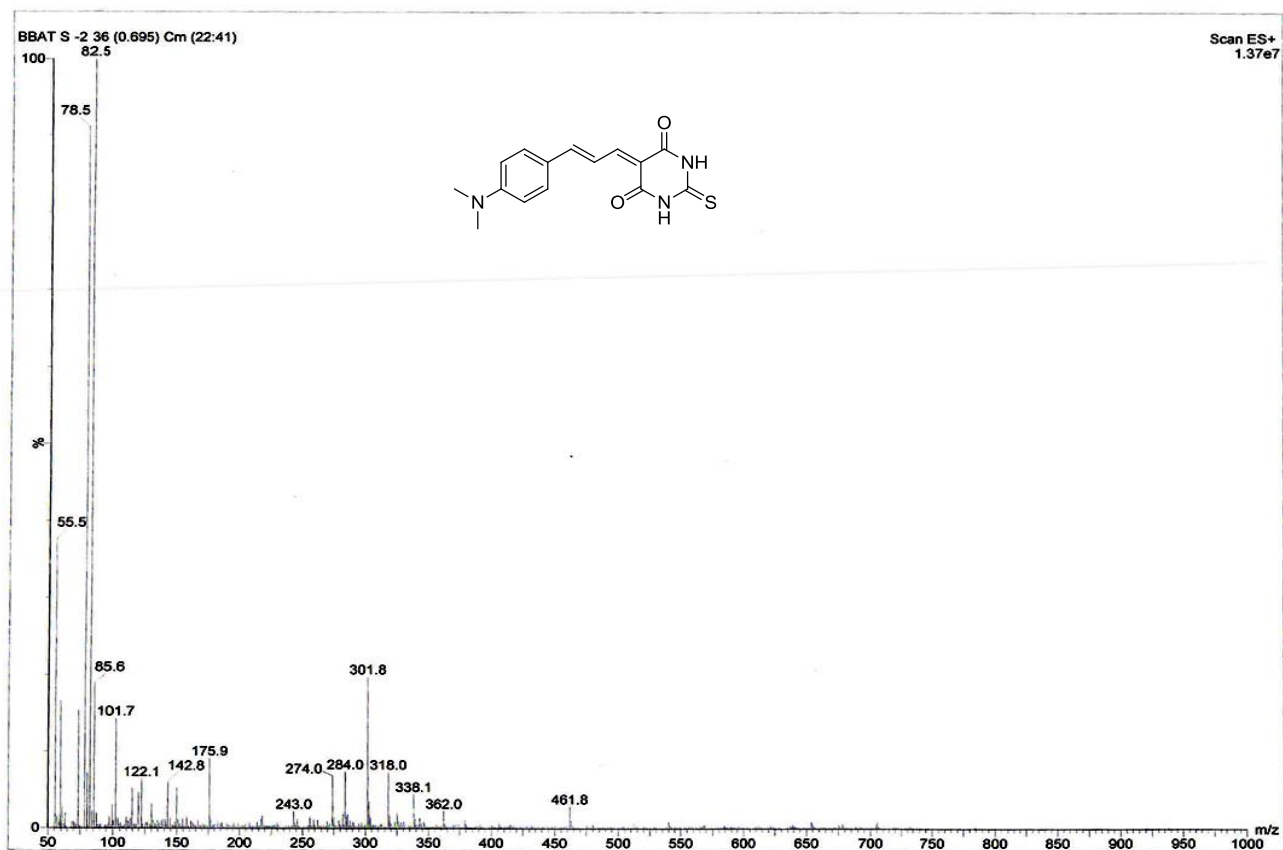
MS spectrum of BBTO-2



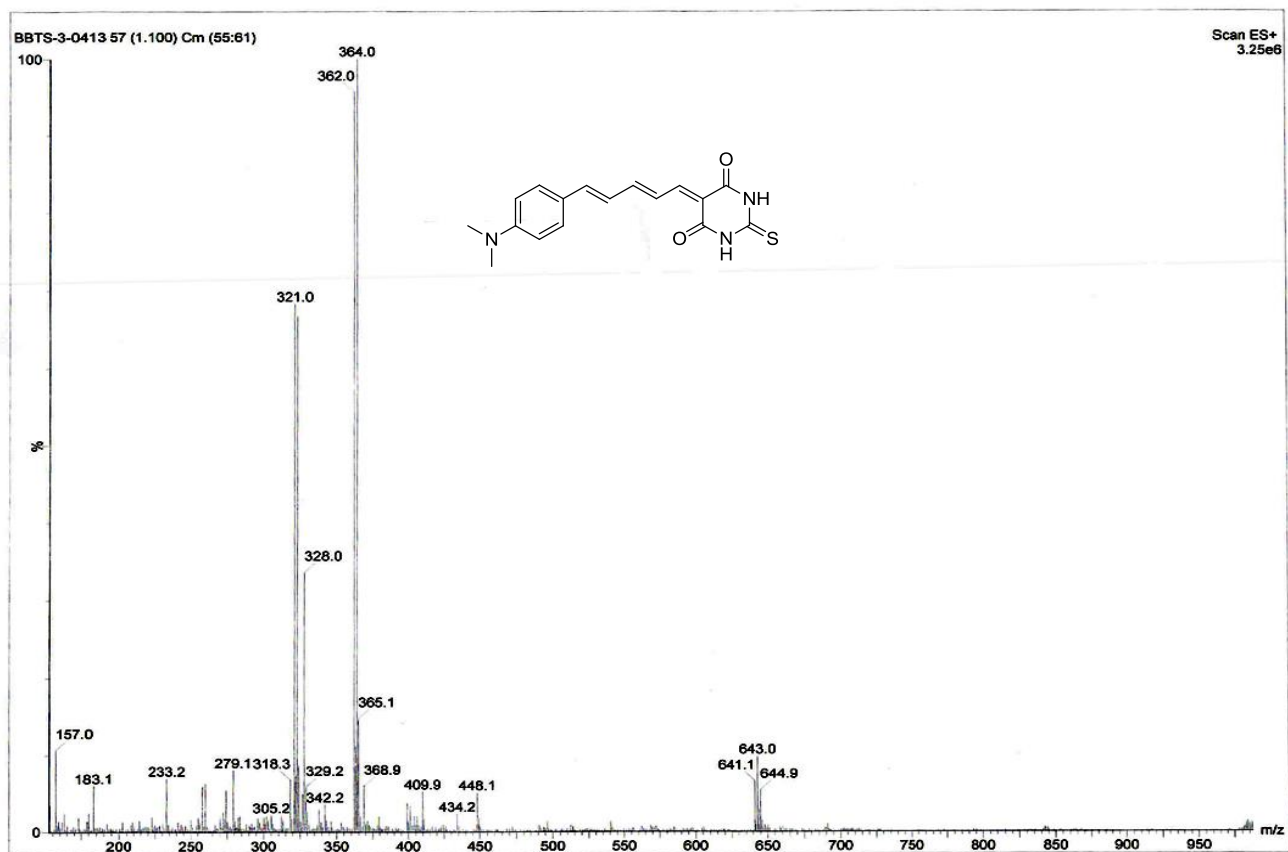
MS spectrum of BBTO-3



MS spectrum of BBTS-1



MS spectrum of BBTS-2



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