

Supporting information

A Stereoselective Construction of *E*- and *Z*- Δ -Ile from E-Dehydroamino Acid Ester: the Synthesis of Phomopsin A Tripeptide Side Chain

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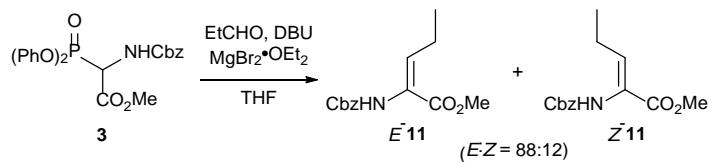
General.

All reagents and solvents were purchased from either Aldrich Chemical Company, Inc., Kanto Kagaku Co., Inc., Merck & Co., Inc., Nacalai Tesque Company, Ltd., Peptide Institute, Tokyo Kasei Kogyo Co., Ltd., or Wako Pure Chemical Industries, Ltd., and used without further purification unless otherwise indicated. Dichloromethane (CH_2Cl_2) was distilled from phosphoric pentaoxide (P_2O_5). Tetrahydrofuran (THF), chloroform (CHCl_3), and dimethylformamide (DMF) of anhydrous grade were used.

Optical rotations were taken on a JASCO P-1030 polarimeter with a sodium lamp (D line). Melting points were determined with a Yanaco MP-21 melting point apparatus and were uncorrected. FTIR spectra were measured on a JASCO FT/IR-6200 infrared spectrophotometer. ^1H NMR spectra were recorded on an either Bruker AVANCE 300 (300 MHz), JEOL JNM-LA 400 (400 MHz), or Bruker AVANCE 600 (600 MHz) spectrometer. Chemical shifts of ^1H NMR were reported in perts per million (ppm, δ) relative to CHCl_3 ($\delta = 7.26$) in CDCl_3 , CD_2HOD ($\delta = 3.31$). ^{13}C NMR spectra were recorded on an either Bruker AVANCE 300 (75 MHz), JEOL JNM-LA 400 (100 MHz), or Bruker AVANCE 600 (150 MHz) spectrometer. Chemical shifts of ^{13}C NMR were reported in ppm (δ) relative to CHCl_3 ($\delta = 77.0$) in CDCl_3 . Low resolution mass spectra (LRMS) and high resolution mass spectra (HRMS) were obtained on an JEOL JMS-AX500 for fast atom bombardment ionization (FAB), chemical ionization (CI), or electron ionization (EI). All reactions were monitored by thin layer chromatography (TLC), which was performed with precoated plates (silica gel 60 F-254, 0.25 mm thickness, manufactured by Merck). TLC visualization was accompanied using UV lamp (254 nm) or a charring solution (ethanoic phosphomolybdic acid, aqueous potassium permanganate and butanoic ninhydrin). Daisogel IR-60 1002W (40/63 μm) was used for flash column chromatography on silica gel.

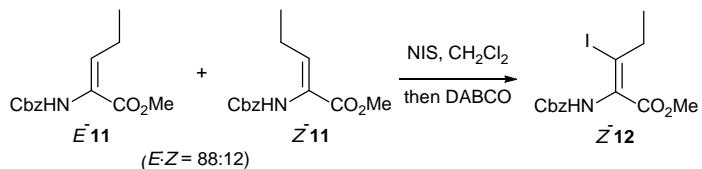
Compounds **7**, **9**, and **23** were obtained as an inseparable mixture. Amide rotamers are observed in compounds **7~9** and **18~23** under the NMR analysis conditions. These NMR signals are broaden and complicated. Therefore, NMR data assignments are not given for these compounds. Instead, actual spectra are provided.

Methyl 2-(((benzyloxy)carbonyl)amino)pent-2-enoate (11)



To a solution of **3**¹ (502 mg, 1.10 mmol) and MgBr₂·OEt₂ (258 mg, 1.00 mmol) in THF (10 mL) was added DBU (150 µL, 1.00 mmol) at 0 °C under argon. The mixture was stirred for 30 min at 0 °C, then propionaldehyde (76 µL, 1.00 mmol, 95% purity) was added to the mixture. The mixture was stirred for 12 h at room temperature, quenched with sat. NH₄Cl (10 mL), and extracted with EtOAc (10 mL × 3). The combined organic layers were washed with brine (30 mL), dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 20 : 1 to 7 : 1) to give a *E/Z* mixture of **11** (229 mg, 87%, *E* : *Z* = 88 : 12) as a colorless oil. The *E/Z* ratio was determined by comparison of the chemical shift values of the olefinic (*E*: 6.79 ppm, *Z*: 6.63 ppm) and N-H (*E*: 6.79 ppm, *Z*: 6.15 ppm) protons.² NMR data of *E/Z*-**11** were identical to those of the reported data.³

(Z)-Methyl 2-(((benzyloxy)carbonyl)amino)-3-iodopent-2-enoate (Z-12)



To a solution of **11** (229 mg, 870 μmol , $E : Z = 88 : 12$) in CH_2Cl_2 (8.7 mL) was added NIS (234 mg, 1.04 mmol) at 0 °C under argon. The mixture was stirred at room temperature for 2 h. DABCO (195 mg, 1.74 mmol) was added to the mixture at 0 °C. The mixture was stirred for 16 h at room temperature and quenched with 1*N* NaHSO_4 (10 mL). The organic layer was separated. The aqueous layer was extracted with EtOAc (10 mL \times 2). The combined organic layers were washed with sat. Na_2SO_3 (30 mL) and brine (30 mL), dried over anhydrous MgSO_4 , and filtered. The filtrate was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 15 : 1) to give **Z-12** (292 mg, 86%).

White solid;

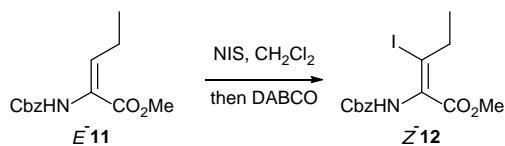
mp 75–77 °C;

FTIR (neat) 3318, 2973, 2952, 1726, 1622, 1481, 1304, 1267, 1220, 1039 cm⁻¹;

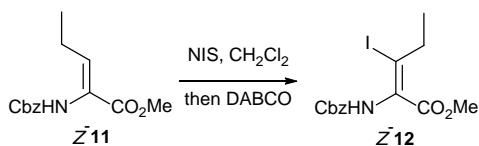
¹H NMR (400 MHz, CDCl₃) δ 7.37-7.32 (m, 5 H), 6.41 (brs, 1 H), 5.14 (s, 2 H), 3.82 (brs, 3 H), 2.76 (brq, *J* = 7.3 Hz, 2 H), 1.13 (t, *J* = 7.3 Hz, 3 H);

¹³C NMR (100 MHz, CDCl₃) δ 161.6, 153.2, 135.4, 130.6, 128.5, 128.3, 128.2, 109.3, 67.7, 52.5, 33.8, 14.8;

HRMS (FAB) calcd for C₁₄H₁₇INO₄ *m/z* 390.0202 [M+H]⁺, found 390.0201.

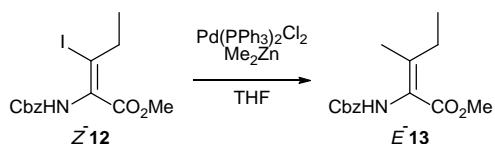


According to the same procedure, *E*-11 (156 mg, 593 μmol) purified from the 88:12 *E/Z*-mixture of **11** by repeating silicagel column chromatography was subjected to the iodination reaction to give *Z*-12 (191 mg, 83%). Spectroscopic data of the resulting *Z*-12 was identical with those of *Z*-12 described above.



According to the same procedure, *Z*-11 (157 mg, 596 μmol) purified from the 88:12 mixture of *E/Z*-11 by repeating silicagel column chromatography was subjected to the iodination reaction to give *Z*-12 (172 mg, 74%). Spectroscopic data of the resulting *Z*-12 was identical with those of *Z*-12 described above.

(*E*)-Methyl 2-(((benzyloxy)carbonyl)amino)-3-methylpent-2-enoate (*E*-13)



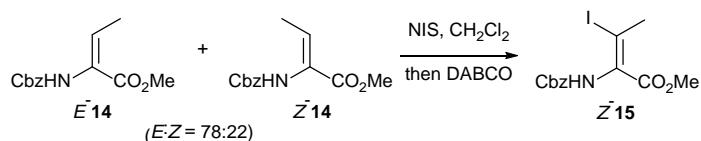
To a solution of *Z*-12 (20.0 mg, 51.4 μmol) and Pd(PPh₃)₂Cl₂ (1.8 mg, 2.6 μmol) in THF (0.5 mL) was added Me₂Zn (100 μL, 102 μmol, 1.02 M solution in hexane) at 0 °C under argon. The mixture was stirred for 13 h at room temperature, quenched with sat. NH₄Cl (5 mL), and extracted with EtOAc (5 mL × 3). The combined organic layers were washed with brine (15 mL), dried over anhydrous MgSO₄ and filtered. The filtrate was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 12 : 1 to 3 : 1) to give *E*-13 (13.2 mg, 93%) as a 7 : 2 mixture of rotamers. Stereochemistry of *E*-13 was determined by NOESY analysis shown in P13. ¹³C NMR of *E*-13 was identical with that of the reported data.⁴

White solid;

¹H NMR (600 MHz, CDCl₃) δ 7.37–7.33 (m, 5 H), 5.95 (brs, 7/9H), 5.64 (brs, 2/9H), 5.14 (s, 2 H), 3.74 (brs, 21/9H), 3.53 (brs, 6/9H), 2.52 (q, *J* = 7.2 Hz, 2 H), 1.86 (s, 3 H), 1.10 (brt, *J* = 7.2 Hz, 3 H);

¹³C NMR (75 MHz, CDCl₃) δ 164.9, 154.4, 149.7, 136.0, 128.1, 127.74, 127.68, 120.7, 66.6, 51.3, 27.2, 19.0, 12.3.

(Z)-Methyl 2-(((benzyloxy)carbonyl)amino)-3-iodobut-2-enoate (Z-15)



According to the literature,² **14** (*E* : *Z* = 78 : 22) was prepared by condensation of **3** and acetaldehyde. To a solution of **14**² (10.0 mg, 40.1 µmol, *E* : *Z* = 78 : 22) in CH₂Cl₂ (2.0 mL) was added NIS (10.8 mg, 48.0 µmol) at 0 °C under argon. The mixture was stirred at room temperature for 2 h. DABCO (8.97 mg, 80.0 µmol) was added to the mixture at 0 °C. The mixture was stirred for 7 h at room temperature. The reaction was quenched with 1*N* NaHSO₄ (5 mL) and extracted with EtOAc (5 mL × 3). The combined organic layers were washed with sat. Na₂SO₃ (15 mL) and brine (15 mL), dried over anhydrous MgSO₄, and filtered. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 15 : 1 to 3 : 1) to give **Z-15** (9.5 mg, 63%).

White solid;

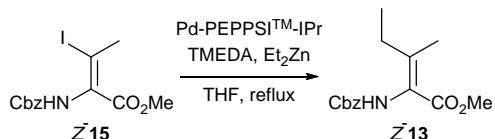
FTIR (neat) 3324, 2952, 1724, 1626, 1483, 1305, 1229, 1049 cm⁻¹;

¹H NMR (400 MHz, CDCl₃) δ 7.36 (m, 5 H), 6.35 (brs, 1 H), 5.14 (s, 2 H), 3.81 (brs, 3 H), 2.76 (s, 3 H);

¹³C NMR (100 MHz, CDCl₃) δ 161.4, 153.4, 135.5, 131.2, 128.6, 128.4, 128.3, 101.1, 67.8, 52.6, 29.1;

HRMS (EI) calcd for C₁₃H₁₄INO₄ *m/z* 374.9968 [M⁺], found 374.9977.

(Z)-Methyl 2-(((benzyloxy)carbonyl)amino)-3-methylpent-2-enoate (Z-13)



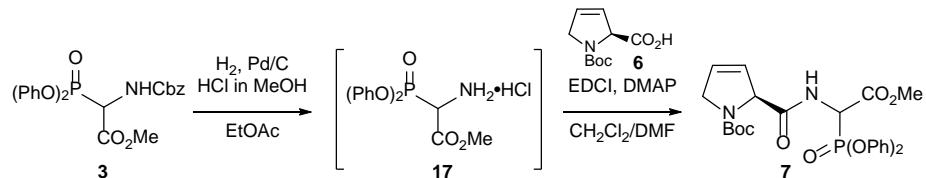
To a solution of **Z-15** (28.9 mg, 77.0 µmol), Pd-PEPPSI™-IPr (2.7 mg, 3.85 µmol, 98% purity) and TMEDA (46 µL, 308 µmol) in THF (770 µL) was added Et₂Zn (154 µL, 154 µmol, 1.0 M solution in hexane) at 0 °C under argon. The mixture was stirred for 20 min at reflux, quenched with sat. NH₄Cl (5 mL), and extracted with EtOAc (5 mL × 3). The combined organic layers were washed with brine (15 mL), dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 10 : 1 to 6 : 1) to give **Z-13** (20.8 mg, 98%) as a 3 : 1 mixture of rotamers. Stereochemistry of **Z-13** was determined by NOESY analysis shown in P14. ¹³C NMR of **Z-13** was identical with that of the reported data.⁴

White solid;

¹H NMR (600 MHz, CDCl₃) δ 7.37–7.33 (m, 5 H), 5.88 (brs, 3/4H), 5.65 (brs, 1/4H), 5.14 (s, 2 H), 3.74 (brs, 9/4H), 3.51 (brs, 3/4 H), 2.24 (q, *J* = 7.6 Hz, 2 H), 2.12 (s, 3 H), 1.04 (brt, *J* = 7.6 Hz, 3 H);

¹³C NMR (75 MHz, CDCl₃) δ 165.4, 154.9, 150.6, 136.2, 128.1, 127.7, 120.5, 66.7, 51.4, 28.1, 18.2, 11.2.

(2*S*)-*tert*-Butyl 2-((1-(diphenoxypyrophoryl)-2-methoxy-2-oxoethyl)carbamoyl)-2,5-dihydro-1*H*-pyrrole-1-carboxylate (7)



To a solution of **3**¹ (1.00 g, 2.20 mmol) in EtOAc (20 mL) was added methanolic HCl prepared from methanol (7.3 mL) and AcCl (1.2 mL, 16.5 mmol) at 0 °C. 10% Pd/C (100 mg, 10 wt%) was then added to the mixture. The mixture was stirred under hydrogen for 3.5 h at room temperature and filtrated through a thin Celite® pad. The filtrate was concentrated under reduced pressure to give crude ammonium salt **17** which was subjected to the next acylation without further purification. **6**⁵ (422 mg, 1.98 mmol), DMAP (134 mg, 1.10 mmol), and EDCI (464 mg, 2.42 mmol) were added to a solution of the crude **17** in CH₂Cl₂/DMF (2 : 1, 21 mL) at 0 °C under argon. The mixture was stirred for 13 h at room temperature, quenched with sat. NH₄Cl (25 mL), and extracted with EtOAc (25 mL × 3). The combined organic layers were washed with brine (75 mL), dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (hexane/EtOAc = 5 : 1 to 1 : 1) to give **7** (618 mg, 55% over 2 steps) as a 1 : 1 inseparable mixture of diastereomers.

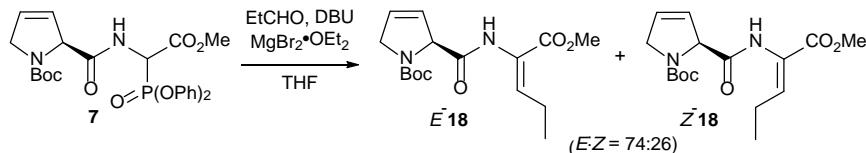
Colorless amorphous solid;

FTIR (neat) 3278, 3064, 2978, 2931, 2869, 1750, 1696, 1590, 1524, 1149, 1456, 1436, 1401, 1366, 1311, 1281, 1205, 1181, 1162, 1119, 1025, 1009 cm⁻¹;

HRMS (FAB) calcd for C₂₅H₃₀N₂O₈P m/z 517.1740 [M+H]⁺, found 517.1746.

¹H and ¹³C NMR spectra: see P15~16.

(*S,E*)-*tert*-Butyl 2-((1-methoxy-1-oxopent-2-en-2-yl)carbamoyl)-2,5-dihydro-1*H*-pyrrole-1-carboxylate (*E*-18) and (*S,Z*)-*tert*-Butyl 2-((1-methoxy-1-oxopent-2-en-2-yl)carbamoyl)-2,5-dihydro-1*H*-pyrrole-1-carboxylate (*Z*-18)



To a solution of **7** (618 mg, 1.20 mmol) and MgBr₂•OEt₂ (340 mg, 1.32 mmol) in THF (13 mL) was added DBU (181 μL, 1.32 mmol) at 0 °C under argon. The mixture was stirred for 30 min at 0 °C. Propionaldehyde (100 μL, 1.32 mmol, 95% purity) was added to the mixture. The mixture was stirred for 13 h at room temperature, quenched with sat. NH₄Cl (20 mL), and extracted with EtOAc (20 mL × 3). The combined organic layers were washed with brine (45 mL), dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated under reduced pressure. The residue was purified by flash column chromatography

on silica gel (hexane/EtOAc = 7 : 1 to 1 : 1) to give **18** (354 mg, 91%, *E* : *Z* = 74 : 26). The *E/Z* ratio was determined by comparison of the chemical shift values of olefinic proton (*E*: 7.12 ppm, *Z*: 6.68 ppm).² *E*- and *Z*-**18** was partly separated by flash column chromatography on silica gel repeatedly (hexane/EtOAc = 7 : 1 to 1 : 1);

E-**18**:

Colorless sticky oil, as a 4 : 3 mixture of rotamers;

$[\alpha]_D^{20} = -111.7$ (*c* 2.36, CHCl₃);

FTIR (neat) 3351, 2976, 2034, 2873, 1681, 1521, 1436, 1392, 1366, 1317, 1245, 1220, 1164, 1125 cm⁻¹;

¹H NMR spectrum: see P17.

¹³C NMR (75 MHz, CDCl₃) δ 168.7, 164.2, 154.7, 153.8, 134.7, 127.6, 126.1, 123.7, 80.7, 69.4, 68.4, 53.7, 52.0, 28.0, 21.5, 13.8;

HRMS (FAB) calcd for C₁₆H₂₅N₂O₅ *m/z* 325.1763 [M+H]⁺, found 325.1752.

Z-**18**:

White solid, as a 2 : 1 mixture of rotamers;

mp 44–45 °C;

$[\alpha]_D^{21} = -205.2$ (*c* 1.03, CHCl₃);

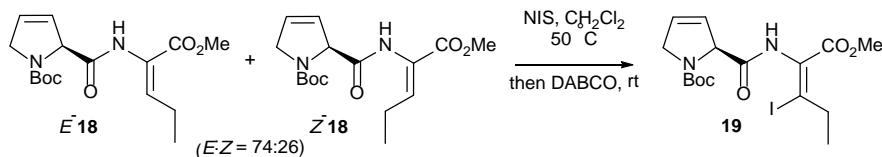
FTIR (neat) 3279, 2974, 2935, 2874, 1682, 1504, 1437, 1394, 1366, 1312, 1275, 1249, 1173, 1126, 1085 cm⁻¹;

¹H NMR spectrum: see P19.

¹³C NMR (75 MHz, CDCl₃) δ 168.7, 164.6, 154.9, 154.1, 140.4, 139.8, 127.7, 126.3, 126.0, 124.2, 123.4, 80.9, 69.0, 67.7, 53.9, 52.1, 28.0, 22.2, 21.9, 12.5;

HRMS (FAB) calcd for C₁₆H₂₅N₂O₅ *m/z* 325.1763 [M+H]⁺, found 325.1758.

(*S,Z*)-*tert*-Butyl 2-((3-iodo-1-methoxy-1-oxopent-2-en-2-yl)carbamoyl)-2,5-dihydro-1*H*-pyrrole-1-carboxylate (**19**)



To a solution of **18** (638 mg, 2.78 mmol, *E* : *Z* = 74 : 26) in CHCl₃ (11 mL) was added NIS (490 mg, 2.18 mmol) at 0 °C under argon. The mixture was stirred at 50 °C for 3 h. DABCO (611 mg, 5.45 mmol) was added to the mixture at room temperature. The mixture was stirred for 11 h and quenched with 1*N* NaHSO₄ (15 mL). The organic layer was separated and the aqueous layer was extracted with EtOAc (15 mL × 2). The combined organic layers were washed with sat. Na₂SO₃ (45 mL) and brine (45 mL), dried over

anhydrous MgSO₄, and filtered. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 5 : 1 to 3 : 1) to give **19** (364 mg, 74%) as a 2 : 1 mixture of rotamers.

Colorless amorphous solid;

$[\alpha]_D^{17} = -117.3$ (*c* 1.47, CHCl₃);

FTIR (neat) 3278, 2976, 2933, 2871, 1730, 1697, 1621, 1480, 1433, 1395, 1366, 1293, 1258, 1220, 1166, 1124, 1082 cm⁻¹;

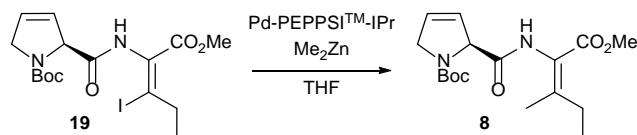
¹H NMR spectrum: see P21.

¹³C NMR (75 MHz, CDCl₃) δ 168.5, 161.4, 155.4, 154.0, 130.8, 130.0, 128.1, 127.6, 125.9, 109.7, 81.4, 68.8, 67.2, 54.0, 52.5, 33.5, 28.2, 14.8;

HRMS (FAB) calcd for C₁₆H₂₄IN₂O₅ *m/z* 451.0730 [M+H]⁺, found 451.0729.

According to the same procedure, *E*-**18** (32.0 mg, 98.7 μmol) and *Z*-**18** (29.5 mg, 90.9 μmol) were converted to **19**, respectively [35.0 mg, 79% from *E*-**18**; 21.0 mg, 51% from *Z*-**18**].

(*S,E*)-*tert*-Butyl 2-((1-methoxy-3-methyl-1-oxopent-2-en-2-yl)carbamoyl)-2,5-dihydro-1*H*-pyrrole-1-carboxylate (**8**)



To a solution of **19** (840 mg, 1.87 mmol) and Pd-PEPPSI™-IPr (38.8 mg, 56.0 μmol, 98% purity) in THF (19 mL) was added Me₂Zn (3.8 mL, 3.73 mmol, 0.99 M solution in hexane) at 0 °C under argon. The mixture was stirred for 1 h at room temperature, quenched with sat. NH₄Cl (30 mL), and extracted with EtOAc (20 mL × 3). The combined organic layers were wash with brine (60 mL), dried over anhydrous MgSO₄ and filtered. The filtrate was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 5 : 1 to 1 : 1) to give **8** (597 mg, 94%) as a 5 : 4 mixture of rotamers;

Colorless sticky oil;

$[\alpha]_D^{18} = -188.0$ (*c* 1.0, CHCl₃);

FTIR (neat) 3277, 2978, 2874, 1675, 1508, 1433, 1396, 1367, 1306, 1268, 1218, 1173, 1125, 1106 cm⁻¹;

¹H NMR spectrum: see P23.

¹³C NMR (75 MHz, CDCl₃) δ 168.8, 164.6, 155.2, 154.4, 151.3, 147.7, 127.3, 126.7, 126.1, 120.7, 119.9, 81.1, 69.1, 67.3, 54.1, 51.6, 28.2, 27.5, 19.7, 19.1, 12.6;

HRMS (FAB) calcd for C₁₇H₂₇N₂O₅ *m/z* 339.1920 [M+H]⁺, found 339.1917.

(S,E)-*tert*-Butyl 2-((1-(allyloxy)-3-methyl-1-oxopent-2-en-2-yl)carbamoyl)-2,5-dihydro-1*H*-pyrrole-1-carboxylate (20)



To a solution of **8** (169 mg, 499 μ mol) in THF/H₂O (1 : 1, 5.0 mL) was added LiOH·H₂O (105 mg, 2.50 mmol). The mixture was stirred for 14 h at 50 °C, quenched with 1*N* HCl (10 mL). The pH of the solution was adjusted to acidity (pH 1–2). The aqueous layer was extracted with EtOAc (10 mL \times 3). The combined organic layers were dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated under reduced pressure. The resulting carboxylic acid was subjected to the next esterification without further purification. To a solution of crude carboxylic acid in DMF (5.0 mL) were added Cs₂CO₃ (163 mg, 501 μ mol) and allyl bromide (51 μ L, 601 μ mol) at 0 °C. The mixture was stirred for 3.5 h at room temperature, quenched with H₂O (10 mL), and extracted with hexane/EtOAc (1 : 1, 10 mL \times 3). The combined organic layers were washed with brine (30 mL), dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 7 : 1 to 3 : 1) to give **20** (156 mg, 86% over 2 steps) as a 5 : 4 mixture of rotamers.

White solid;

mp 111–113 °C;

$[\alpha]_D^{21} = -2.7$ (*c* 1.15, CHCl₃);

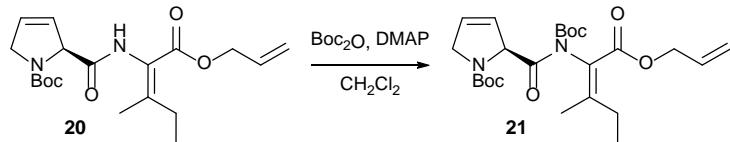
FTIR (neat) 3289, 2978, 2935, 2874, 1700, 1508, 1456, 1398, 1366, 1303, 1261, 1201, 1173, 1125, 1107 cm⁻¹;

¹H NMR spectrum: see P25.

¹³C NMR (75 MHz, CDCl₃) δ 168.8, 163.8, 155.1, 154.4, 151.5, 148.1, 132.0, 127.3, 126.6, 126.1, 120.8, 119.9, 118.3, 117.9, 81.1, 69.0, 67.3, 65.3, 54.0, 28.2, 27.5, 19.8, 19.1, 12.6;

HRMS (FAB) calcd for C₁₉H₂₉N₂O₅ *m/z* 365.2076 [M+H]⁺, found 365.2069.

(S,E)-*tert*-Butyl 2-((1-(allyloxy)-3-methyl-1-oxopent-2-en-2-yl)(tert-butoxycarbonyl)carbamoyl)-2,5-dihydro-1*H*-pyrrole-1-carboxylate (21)



To a solution of **20** (156 mg, 428 μ mol) in CH₂Cl₂ (2.1 mL) were added Boc₂O (200 μ L, 858 μ mol) and DMAP (52.4 mg, 429 μ mol) at 0 °C. The mixture was stirred for 3 h at room temperature and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 20 : 1 to 10 : 1) to give **21** (201 mg, quant.) as a colorless sticky oil;

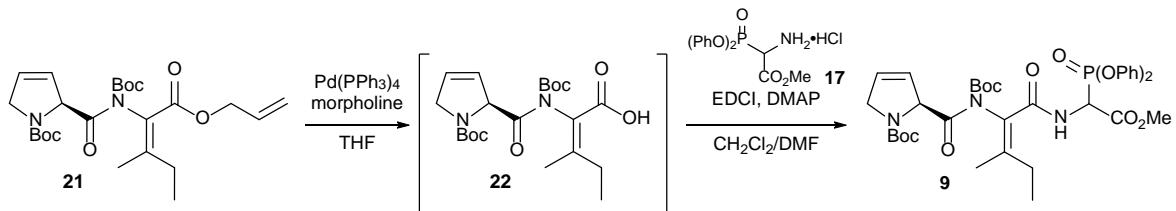
$[\alpha]_D^{21} = -1.7$ (c 1.09, CHCl_3);

FTIR (neat) 3419, 3087, 2978, 2935, 2871, 1722, 1701, 1648, 1458, 1402, 1368, 1294, 1257, 1225, 1203, 1154, 1129, 1107, 1058 cm^{-1} ;

HRMS (FAB) calcd for $\text{C}_{24}\text{H}_{37}\text{N}_2\text{O}_7$ m/z 465.2601 [$\text{M}+\text{H}]^+$, found 465.2586.

NMR spectra: see P27~28.

(2*S*)-*tert*-Butyl 2-((*tert*-butoxycarbonyl)((*E*)-1-((1-(diphenoxypyrophosphoryl)-2-methoxy-2-oxoethyl)amino)-3-methyl-1-oxopent-2-en-2-yl)carbamoyl)-2,5-dihydro-1*H*-pyrrole-1-carboxylate (9)



To a solution of **21** (162 mg, 349 μmol) in THF (3.5 mL) were added $\text{Pd}(\text{PPh}_3)_4$ (40.3 mg, 34.9 μmol) and morpholine (300 μL , 3.49 mmol) at 0 $^\circ\text{C}$. The mixture was stirred for 2 h at room temperature, quenched with 1*N* HCl (10 mL), and extracted with EtOAc (10 mL \times 3). The combined organic layers were dried over anhydrous MgSO_4 and filtered. The filtrate was concentrated under reduced pressure to give carboxylic acid **22** which was subjected to the next reaction without further purification. The crude **22** in CH_2Cl_2 (2.4 mL), EDCI (110 mg, 575 μmol), and DMAP (32.0 mg, 262 μmol) were subsequently added to a solution of α -(diphenylphosphono)glycine ammonium salt **17** in DMF (1.2 mL) prepared from **3¹** (238 mg, 523 μmol), 10% Pd/C (23.8 mg, 10 wt%), AcCl (280 μL , 3.92 mmol), and MeOH (1.2 mL) in EtOAc (3.5 mL) at 0 $^\circ\text{C}$ under argon. The mixture was stirred for 15 h at room temperature, quenched with sat. NH_4Cl (5 mL), and extracted with EtOAc (5 mL \times 3). The combined organic layers were washed with brine (15 mL), dried over anhydrous MgSO_4 , and filtered. The filtrate was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 10 : 1 to 1 : 1) to give **9** (143 mg, 56% over 2 steps) as a 1 : 1 mixture of diastereomers;

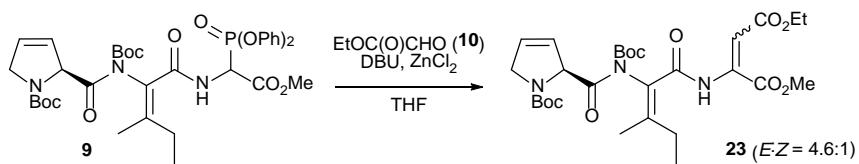
Colorless sticky oil.

FTIR (neat) 3320, 3009, 2979, 2934, 2871, 1739, 1681, 1591, 1489, 1405, 1369, 1288, 1256, 1209, 1181, 1153, 1059, 1026, 1009 cm^{-1} ;

HRMS (FAB) calcd for $\text{C}_{36}\text{H}_{47}\text{N}_3\text{O}_{11}\text{P}$ m/z 728.2948 [$\text{M}+\text{H}]^+$, found 728.2944.

NMR spectra: see P29~30.

4-Ethyl 1-methyl 2-((*E*)-2-((*S*)-*N,N*-bis(*tert*-butoxycarbonyl)-2,5-dihydro-1*H*-pyrrole-2-carboxamido)-3-methylpent-2-enamido)but-2-enedioate (23)



To a solution of **9** (140 mg, 192 µmol) and ZnCl₂ (384 µL, 384 µmol, 1.0 M solution in Et₂O) in THF (1.9 mL) was added DBU (29 µL, 192 µmol) at 0 °C under argon. The mixture was stirred for 30 min at 0 °C. Ethyl glyoxalate polymer form (**10**) (41 µL, 192 µmol, 47% solution in toluene) was added to the mixture. The mixture was stirred for 15 h at room temperature, quenched with sat. NH₄Cl (5 mL), and extracted with EtOAc (5 mL × 3). The combined organic layers were washed with brine (15 mL), dried over anhydrous MgSO₄, and filtered. The filtrate was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 9 : 1 to 5 : 1) to give **23** (88.5 mg, 80%, *E* : *Z* = 4.6 : 1) as a mixture of *E*-**23** and *Z*-**23**.

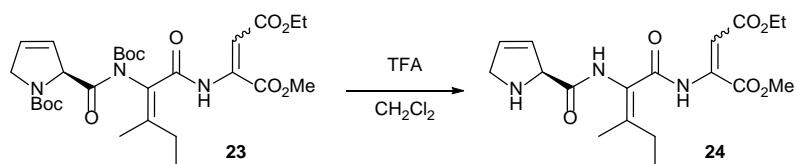
Colorless amorphous solid;

FTIR (neat) 3294, 3018, 2980, 2936, 2873, 1739, 1716, 1671, 1623, 1520, 1420, 1370, 1288, 1258, 1218, 1143, 1098, 1063, 1038 cm⁻¹;

HRMS (CI) calcd for C₂₈H₄₂N₃O₁₀ *m/z* 580.2870 [M+H]⁺, found 580.2878.

NMR spectra: see P31~32.

Synthesis of the tripeptide side chain **24**



To a solution of **23** (10.1 mg, 17.4 µmol) was added TFA (0.5 mL) in CH₂Cl₂ (0.5 mL) at 0 °C under argon. The mixture was stirred for 30 min at room temperature and concentrated under reduced pressure to give a crude 4.3:1 mixture of *E*-**24** and *Z*-**24**. The residue was purified by PLC (silica gel 60 F-254, 0.5 mm thickness, manufactured by Merck) (CHCl₃/MeOH = 10 : 1) to give *E*-**24** (2.3 mg, 35%) and a 1 : 1.6 mixture of *E*-**24** and *Z*-**24** (1.2 mg, 18%). The stereochemistry was determined by comparison of the chemical shift values of olefinic proton (*E*: 6.05 ppm, *Z*: 5.46 ppm).²

E-**24**:

¹H NMR (300 MHz, CD₃OD) δ 6.05 (s, 1 H), 6.00 (m, 1 H), 5.87 (m, 1 H), 4.59 (m, 1 H), 4.13 (q, *J* = 7.2 Hz, 2 H), 3.88 (m, 2 H), 3.81 (s, 3 H), 2.36 (q, *J* = 7.5 Hz, 2 H), 1.77 (s, 3 H), 1.25 (t, *J* = 7.2 Hz, 3 H), 1.09 (t, *J* = 7.5 Hz, 3 H).

Z-**24**:

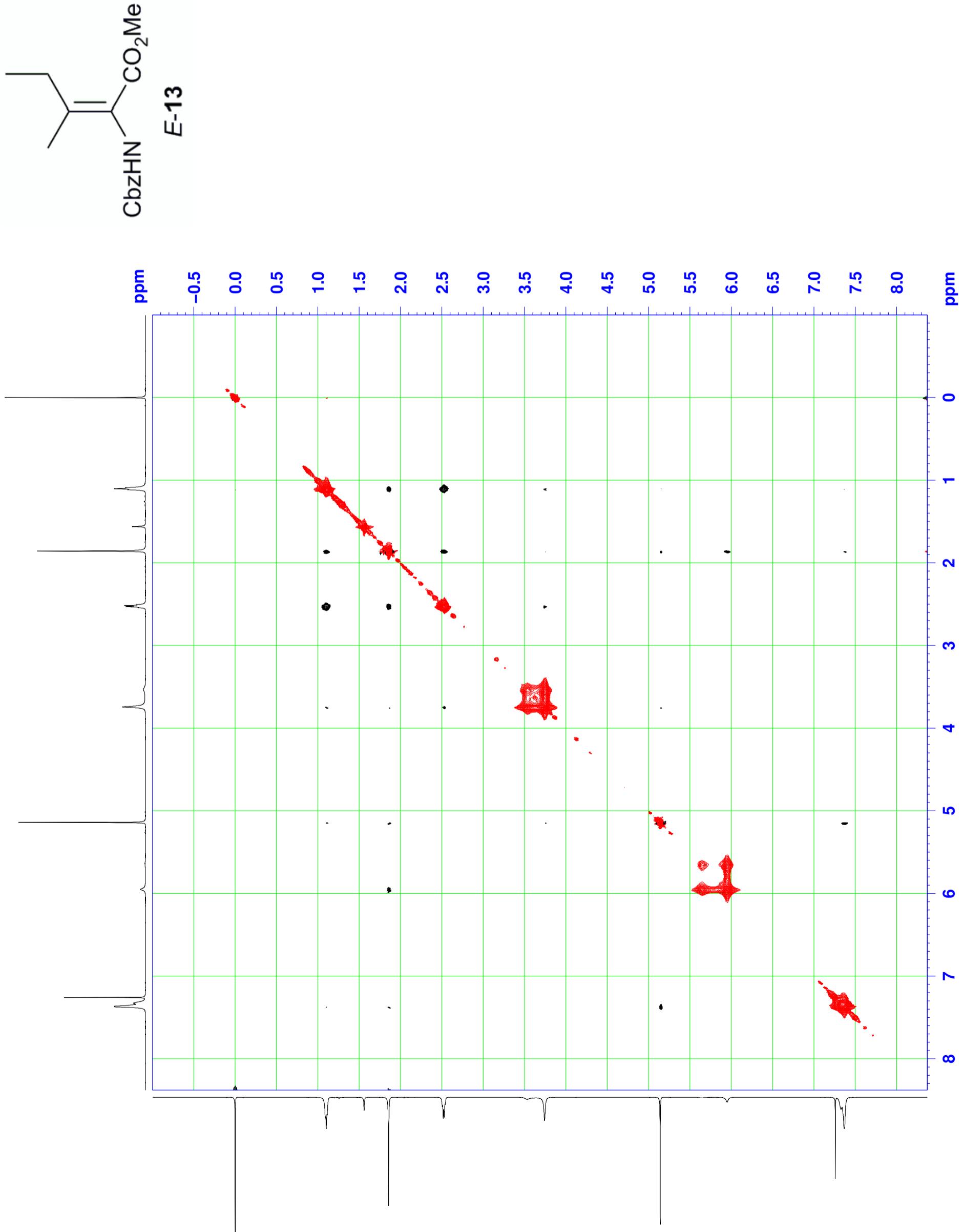
¹H NMR (300 MHz, CD₃OD) δ 6.00 (m, 1 H), 5.93 (m, 1 H), 5.46 (s, 1 H), 4.58 (m, 1 H), 4.22 (q, *J* = 7.2 Hz,

2 H), 3.90 (m, 2 H), 3.83 (s, 3 H), 2.56 (q, J = 7.5 Hz, 2 H), 1.80 (s, 3 H), 1.28 (t, J = 7.2 Hz, 3 H), 1.11 (t, J = 7.5 Hz, 3 H).

References

- 1) Hamada, M.; Shinada, T.; Ohfune, Y. *Org. Lett.* **2009**, *11*, 4664–4667.
- 2) Yasuno, Y.; Hamada, M.; Yamada, T.; Shinada, T.; Ohune, Y. *Eur. J. Org. Chem.* **2013**, 1884–1888.
- 3) Mazurkiewicz, R.; Kuźnic, A. *Tetrahedron Lett.* **2006**, *47*, 3439–3442.
- 4) Schmidt, U.; Griesser, H.; Leitenberger, V.; Lieberknecht, A. Mangold, R.; Meyer, R.; Riedl, B. *Synthesis*, **1992**, 487–490.
- 5) (a) Shangguan, N.; Joullié, M. *Tetrahedron Lett.* **2009**, *50*, 6748–6750. (b) Schumacher, K. K.; Jiang, J.; Joullié, M. M. *Tetrahedron: Asymmetry* **1998**, *9*, 47–53.

Et-Me (E)_noesy_CDCl3_temp25



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SFO1          undefined

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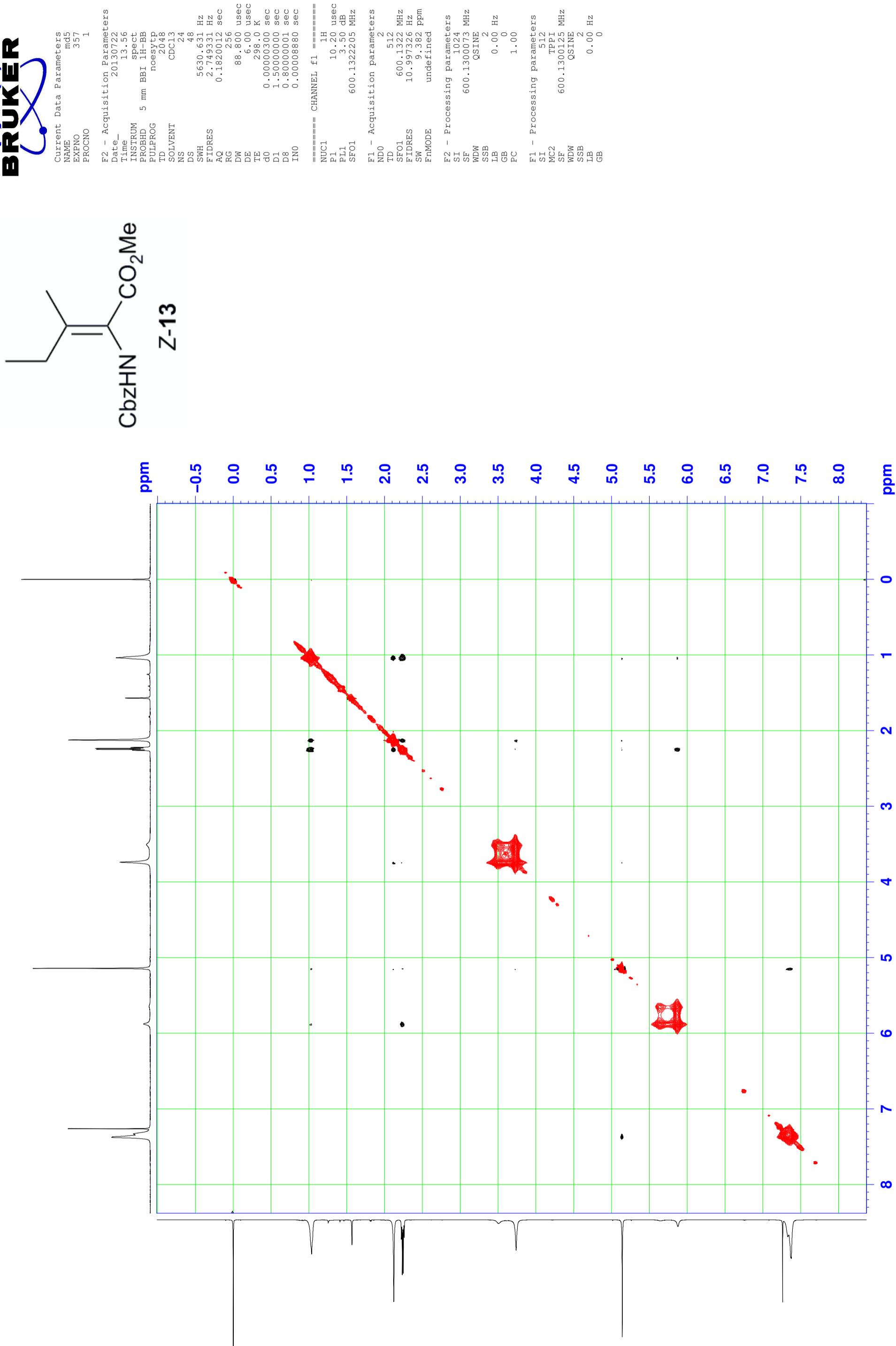
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F13 - Processing parameters
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Et-Me (Z)_noesy_CDCl₃_temp25



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SOLVENT CDCl₃
NS 24
DS 48
SWH 5630.631 Hz
FIDRES 2.749331 Hz
AQ 0.1820012 sec
RG 256
DW 88.800 usec
DE 6.00 usec
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D1 1.5000000 sec
D8 0.8000001 sec
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PL1 3.50 dB
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F1 - Acquisition Parameters
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TD 512
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FIDRES 10.997326 Hz
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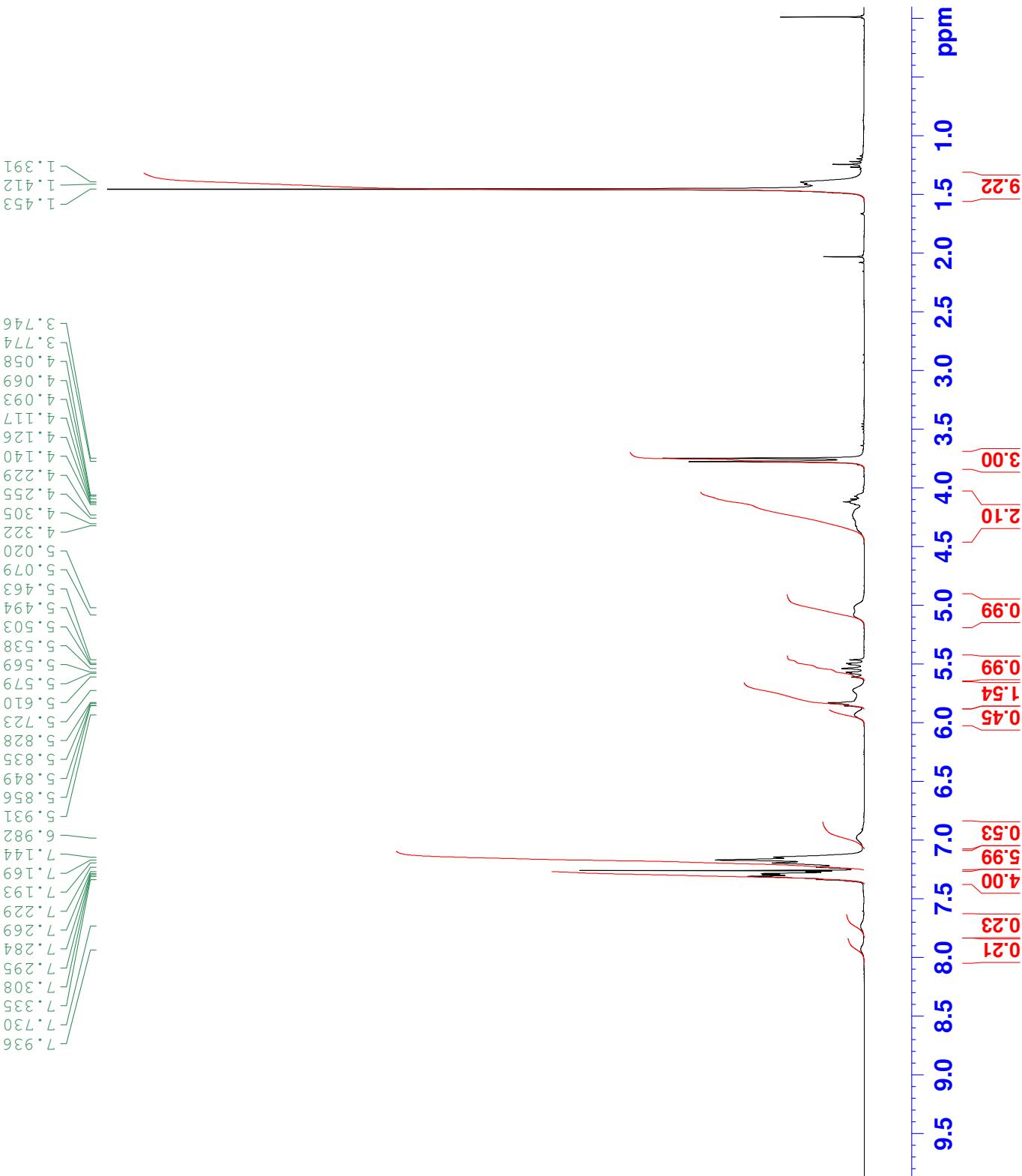
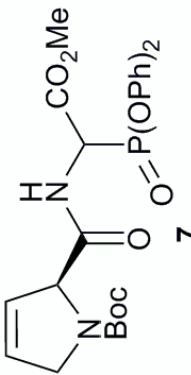
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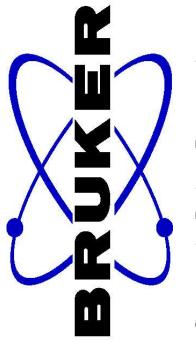


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 PROCNO 1

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 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6188.119 Hz
 FIDRES 0.094423 Hz
 AQ 5.2953587 sec
 RG 64
 DW 80.800 usec
 DE 6.50 usec
 TE 291.8 K
 D1 1.0000000 sec

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Current Data Parameters
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 PROCN0 1

F2 - Acquisition Parameters
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 Time 17.54
 INSTRUM av300
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 PULPROG CDC13
 SOLVENT 1024
 NS 2
 DS 18028.846 Hz
 SWH 0.275098 Hz
 FIDRES 1.8175818 sec
 AQ 203
 RG 27.733 usec
 DW 6.50 usec
 DE 292.8 K
 TE 2.00000000 sec
 D1 0.03000000 sec
 D11

===== CHANNEL f1 =====
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 P1 10.0 usec
 PLW1 38.05099869 W
 SFO1 75.4752953 MHz

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 NUC2 1H
 PCPD2 90.0 usec
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 PLW12 0.20833001 W
 PLW13 0.16875000 W
 SFO2 300.1312005 MHz

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 LB GB 0 1.40
 PC

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 53.10
 53.72

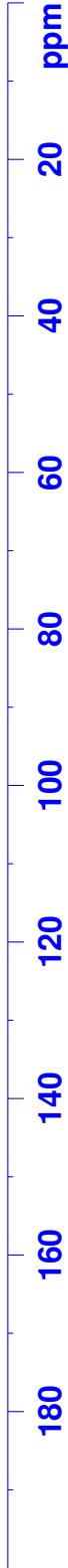
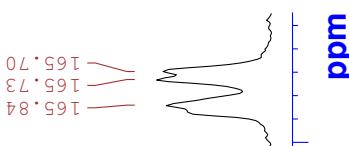
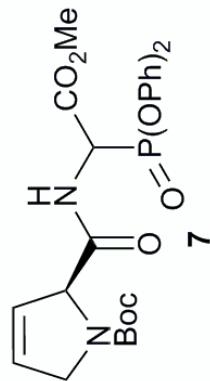
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165.70
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 170.11





Current	Data Parameters
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EXPNO	10
PROCNO	1

F2 - Acquisition Parameters

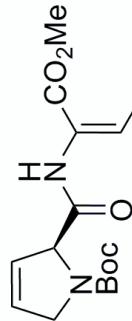
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NS	16
DS	2
SWH	61.88 .119 Hz
FIDRES	0.094423 Hz
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RG	80.6
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D1	1.0000000 sec

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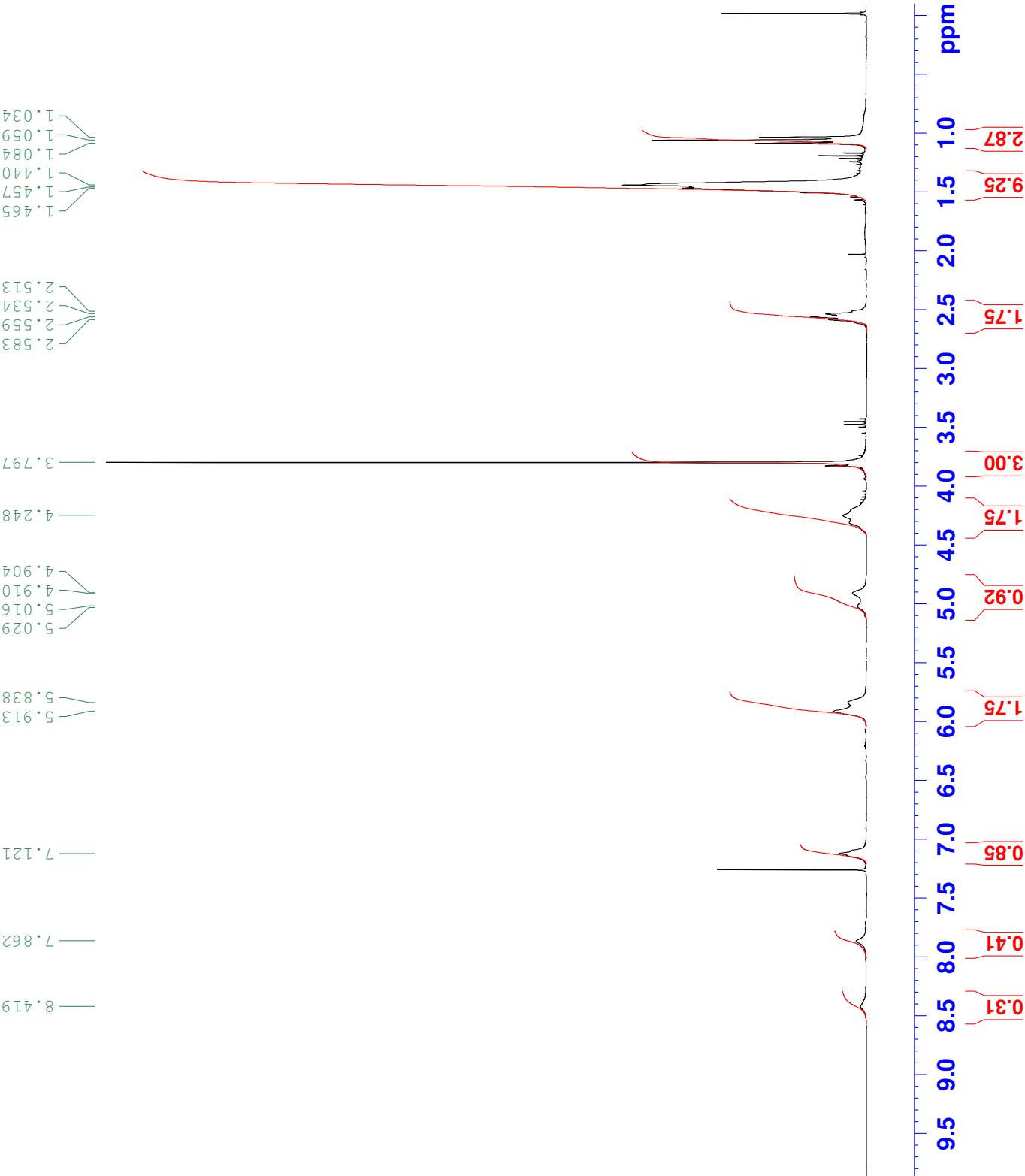
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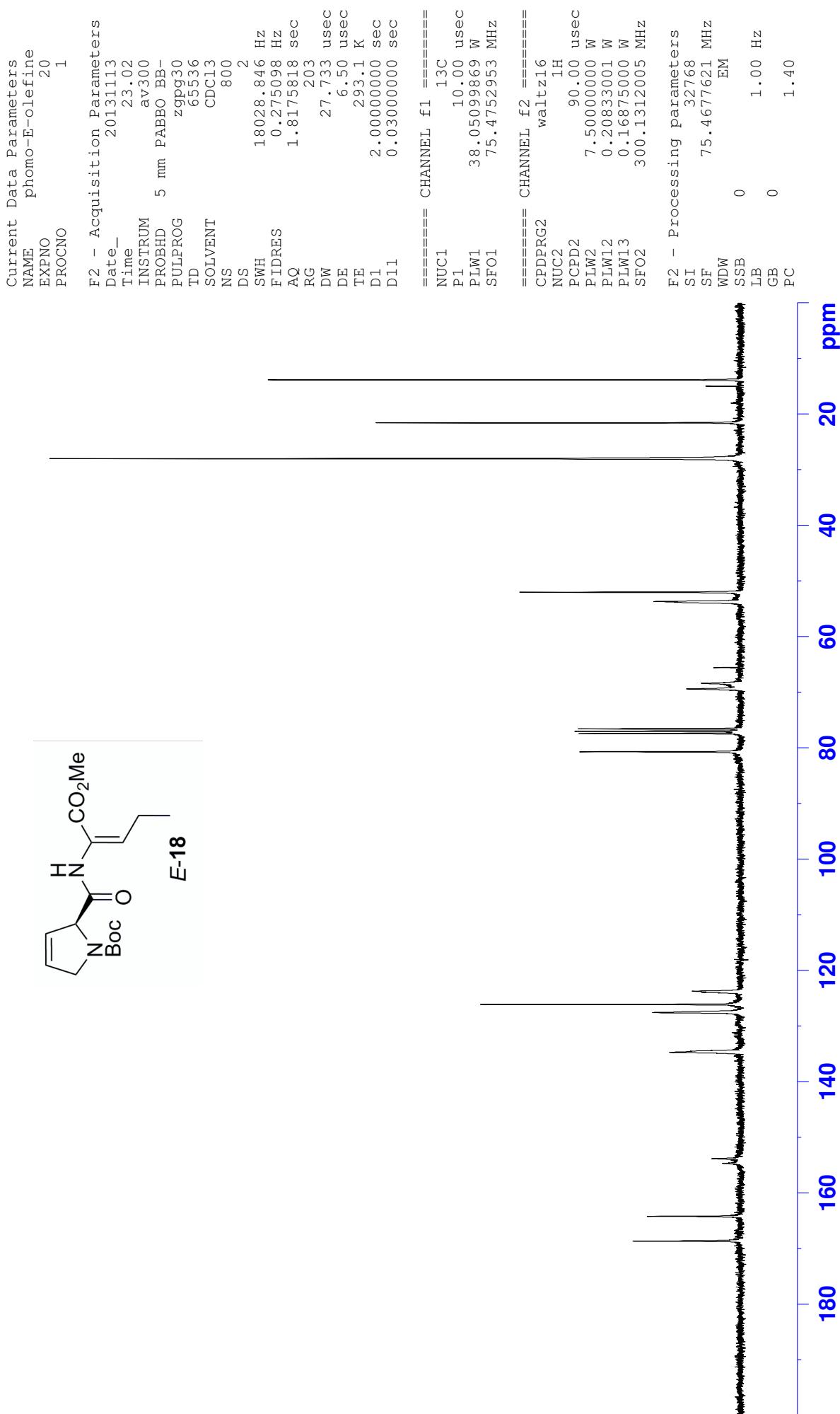
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PC

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E-18



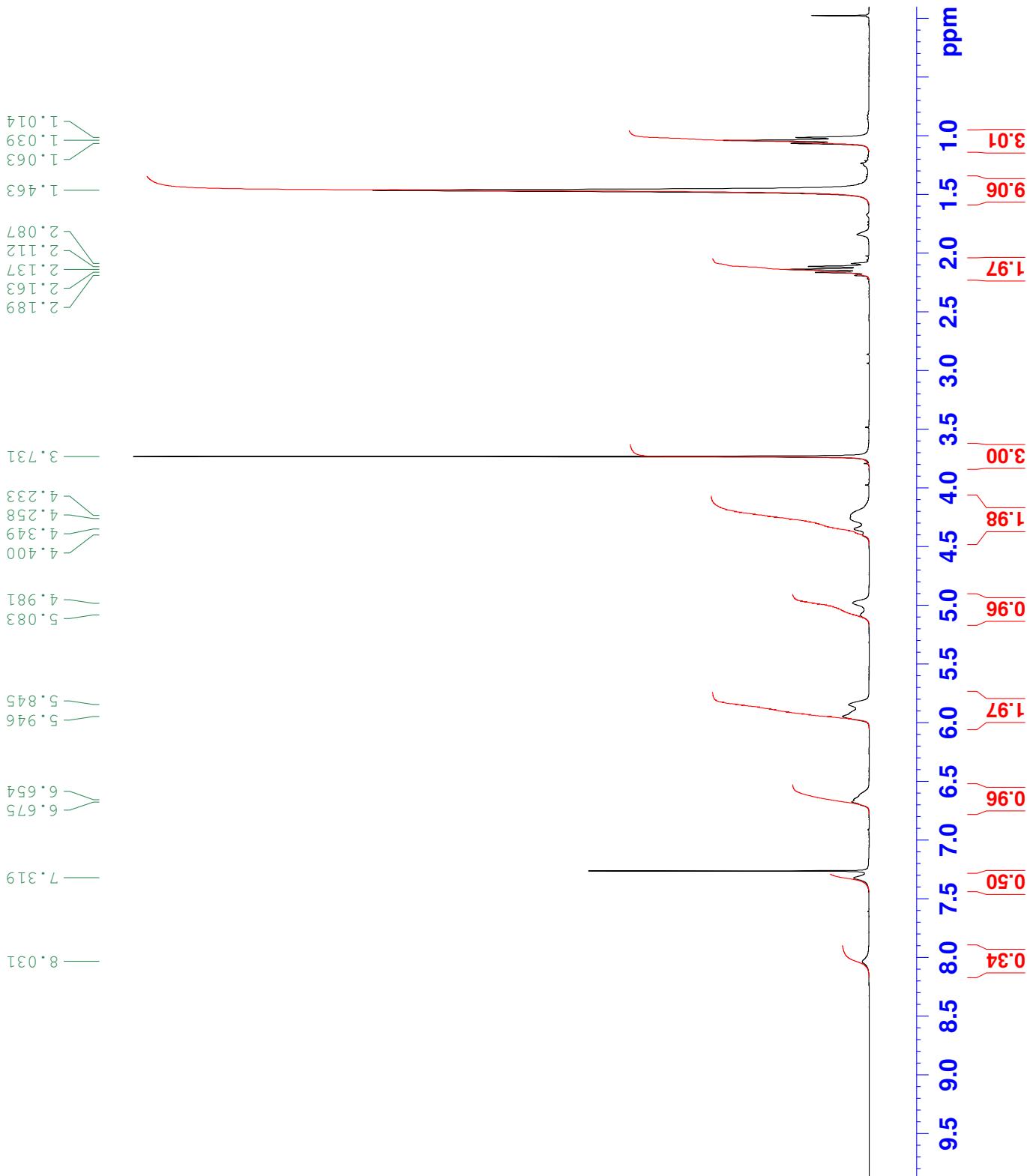


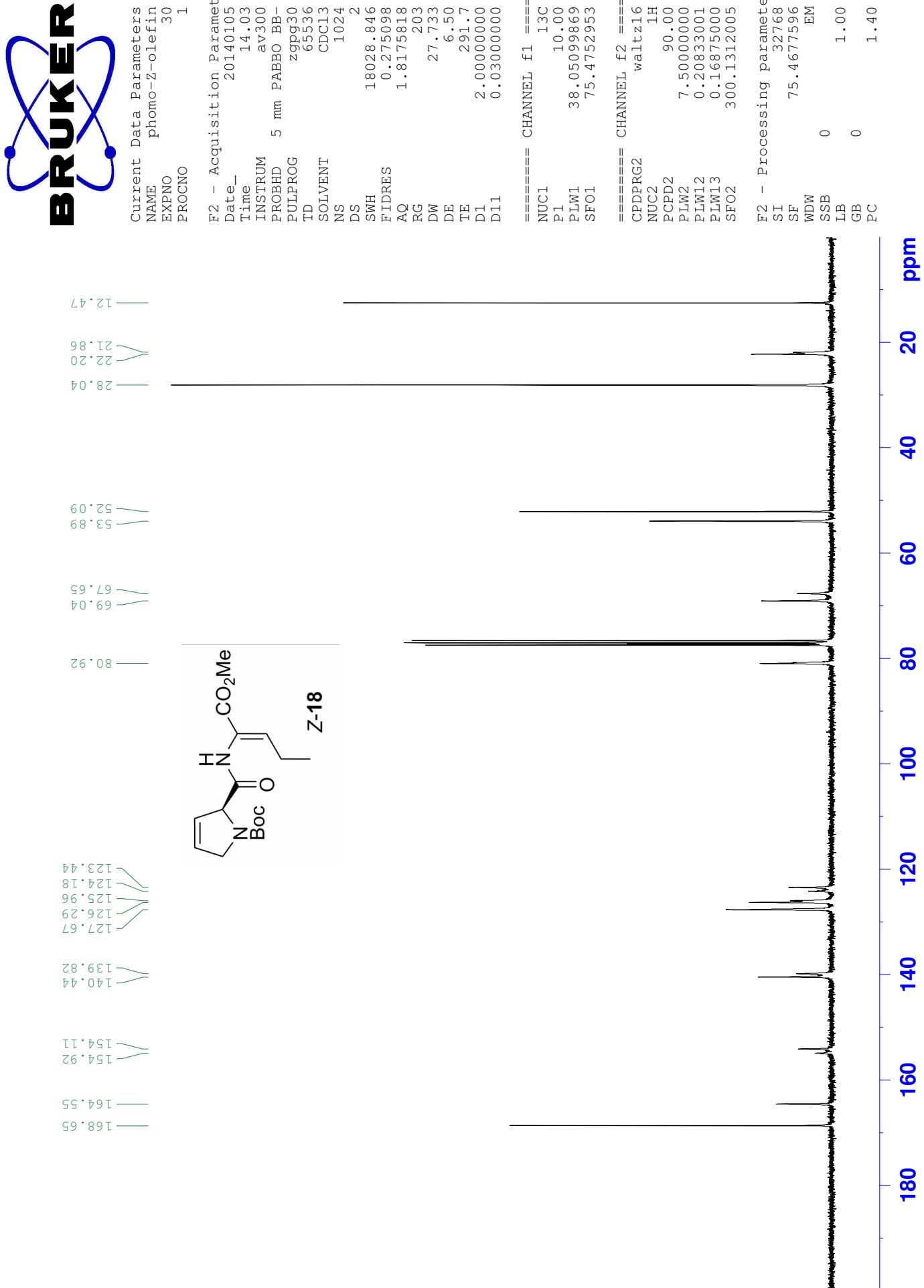


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EXPNO 20
PROCNO 1

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TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 6188.119 Hz
FIDRES 0.094423 Hz
AQ 5.2953587 sec
RG 64
DW 80.800 usec
DE 6.50 usec
TE 290.5 K
D1 1.0000000 sec

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EXPNO		10
PROCNO		1

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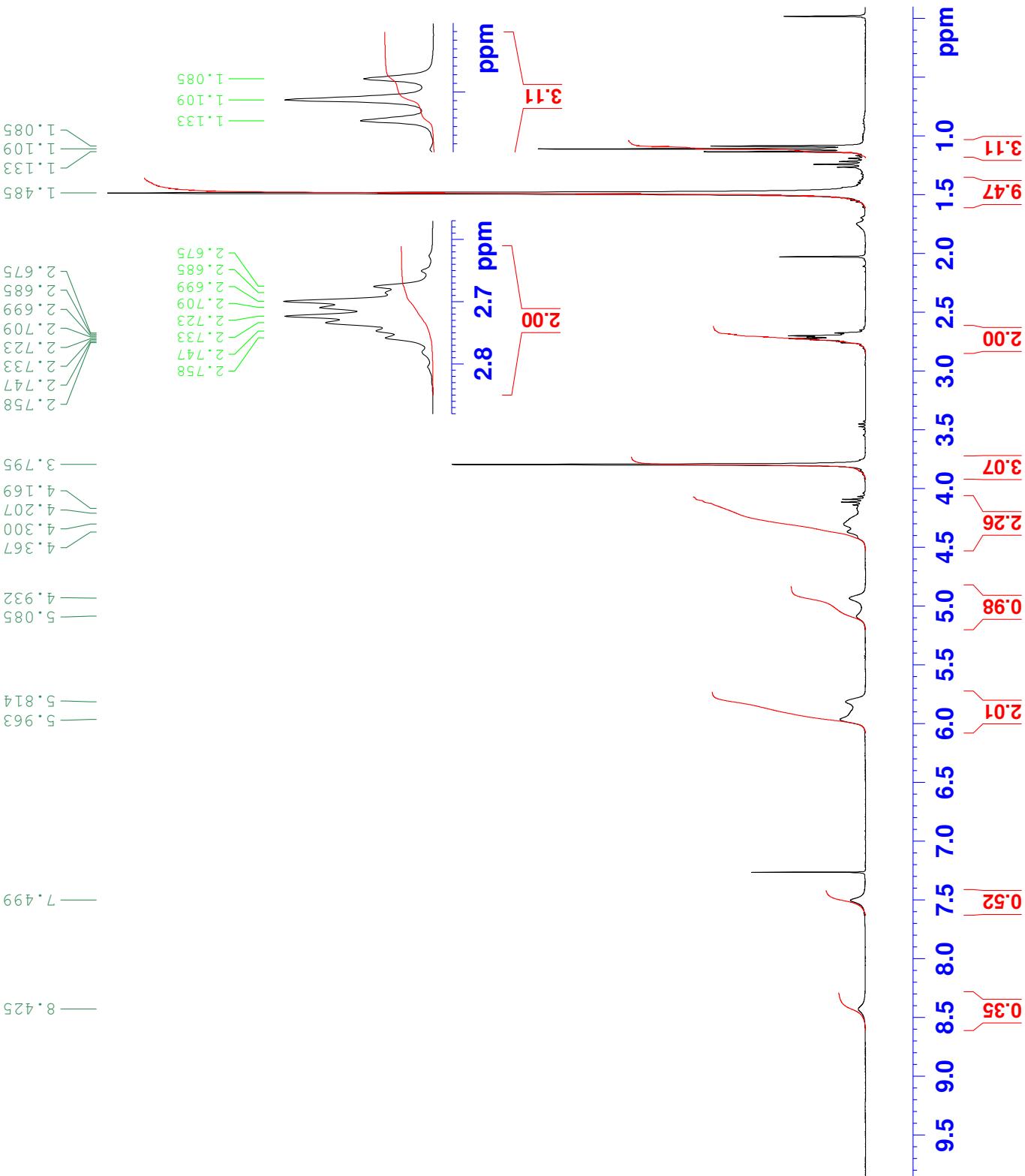
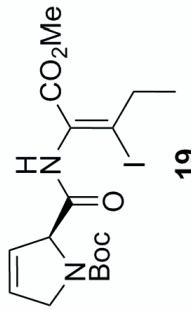
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FIDRES		5.2935587		sec
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D1				

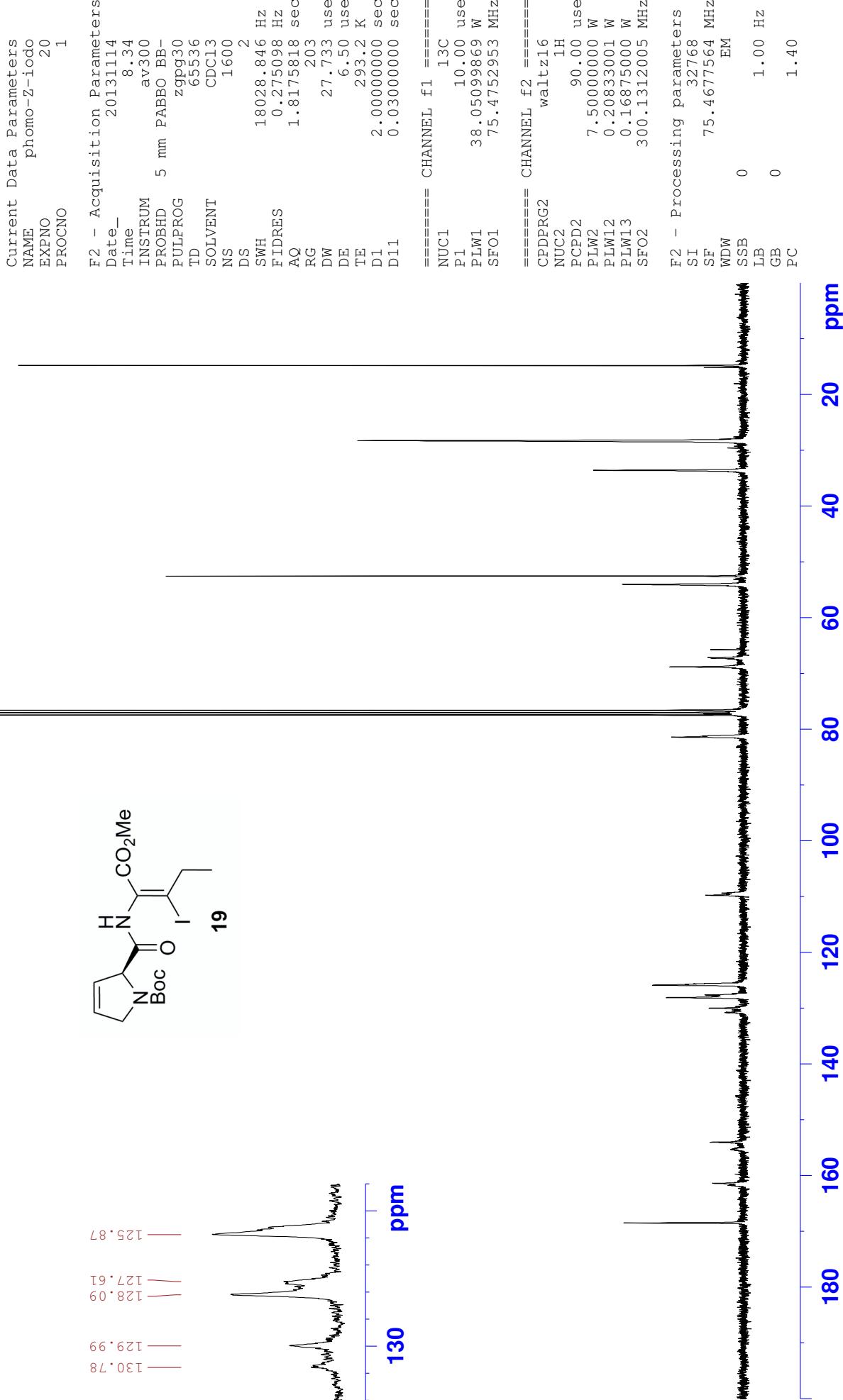
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PC

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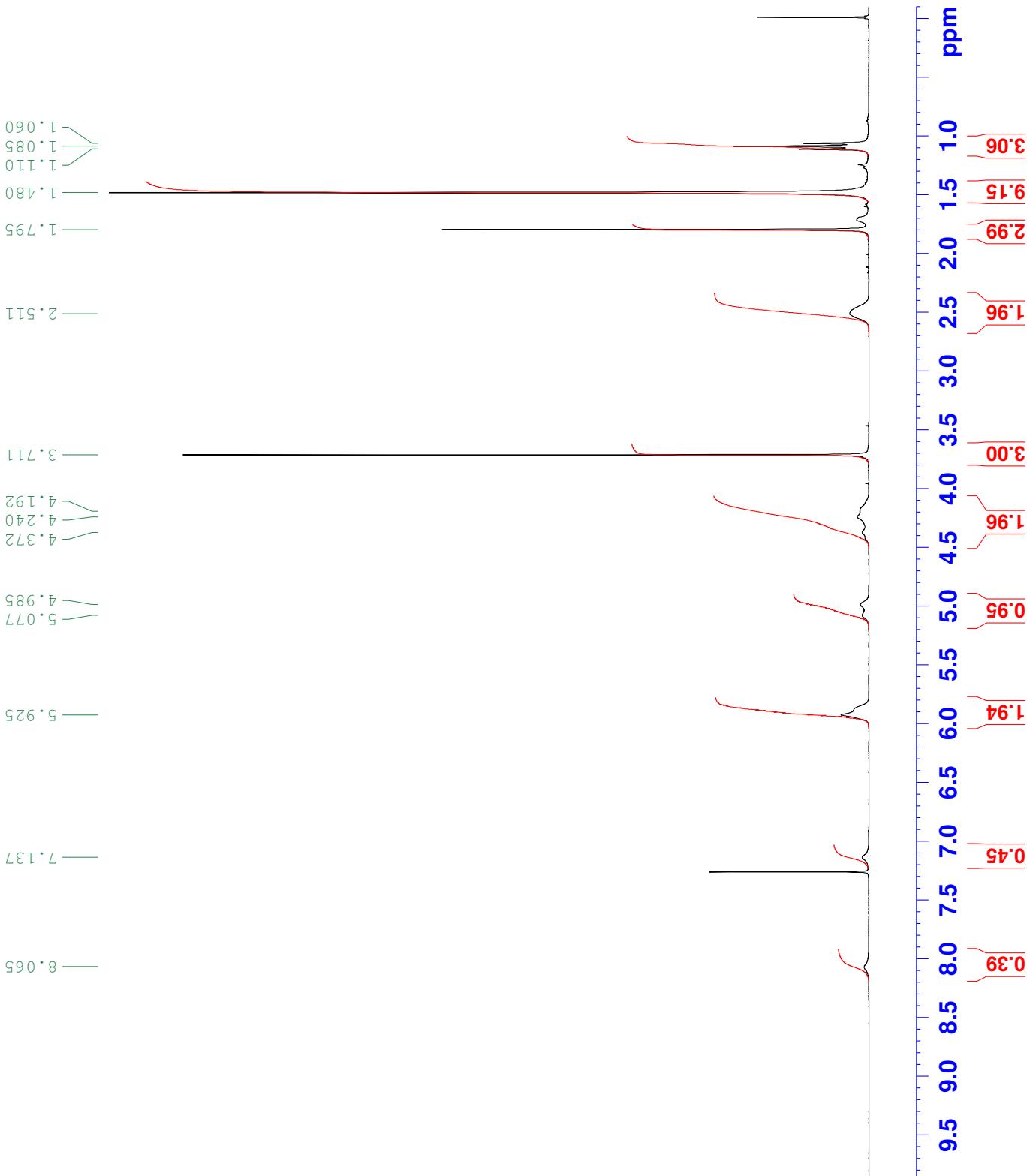
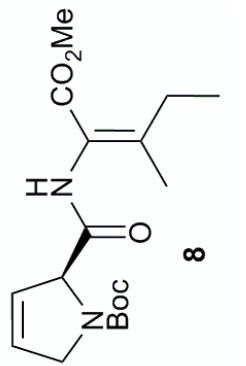


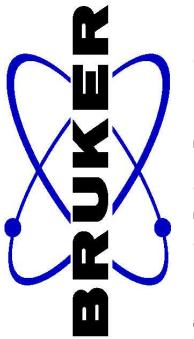


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 PROCNO 1

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 NS 16
 DS 2
 SWH 6188.119 Hz
 FIDRES 0.094423 Hz
 AQ 5.2953587 sec
 RG 128
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 DE 6.50 usec
 TE 291.8 K
 D1 1.0000000 sec

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PROCNO 1

F2 - Acquisition Parameters

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SOLVENT CDCl3
NS 7452
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FIDRES 0.275098 Hz
AQ 1.8175818 sec
RG 203
DW 6.50 usec
DE 27.733 usec
TE 293.6 K
D1 2.00000000 sec
D11 0.03000000 sec

===== CHANNEL f1 =====

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SFO1 75.4752953 MHz

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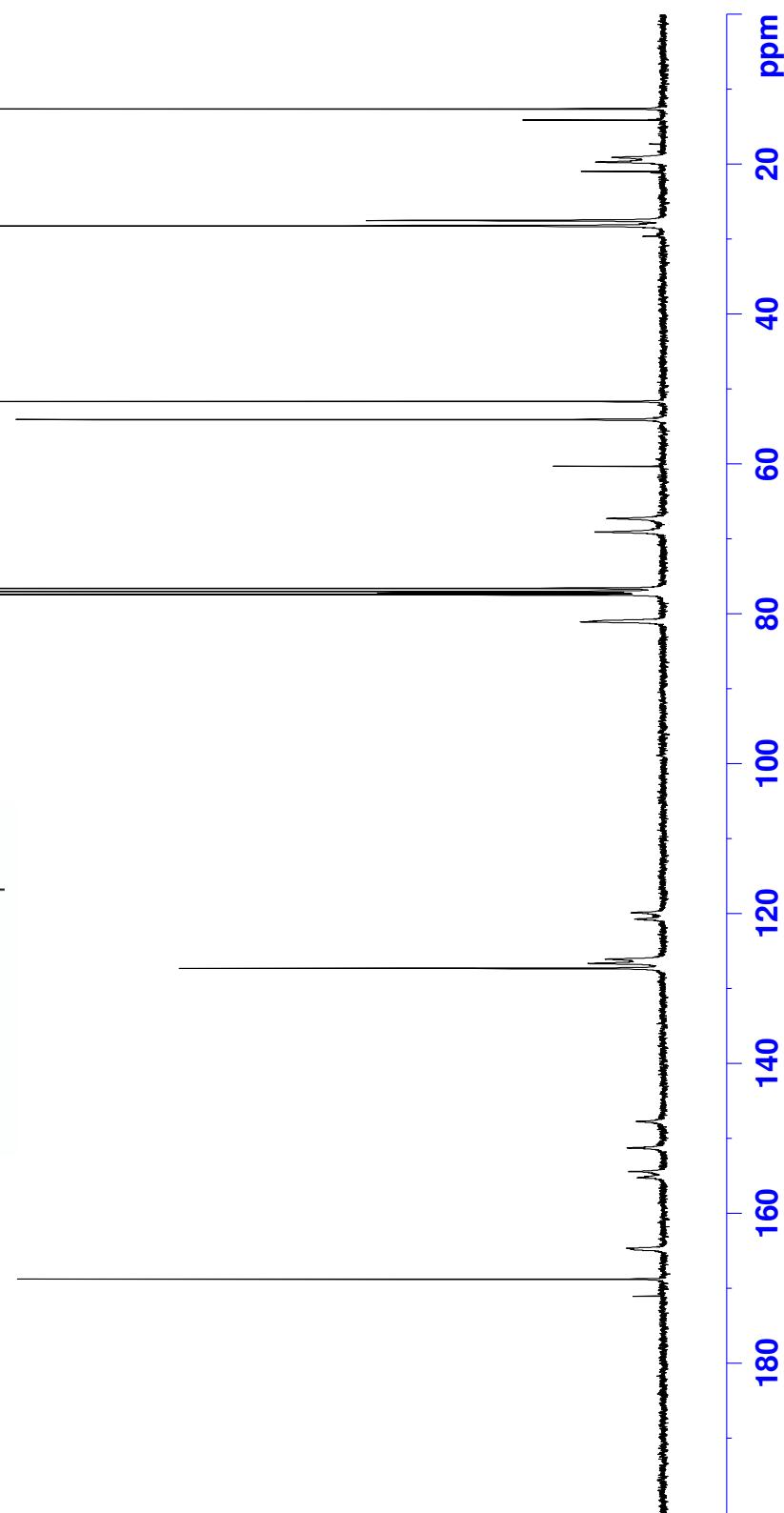
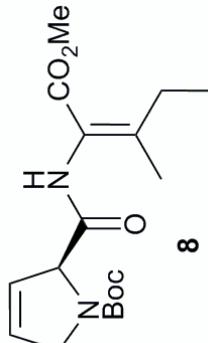
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F2 - Processing parameters

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20.93
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69.07
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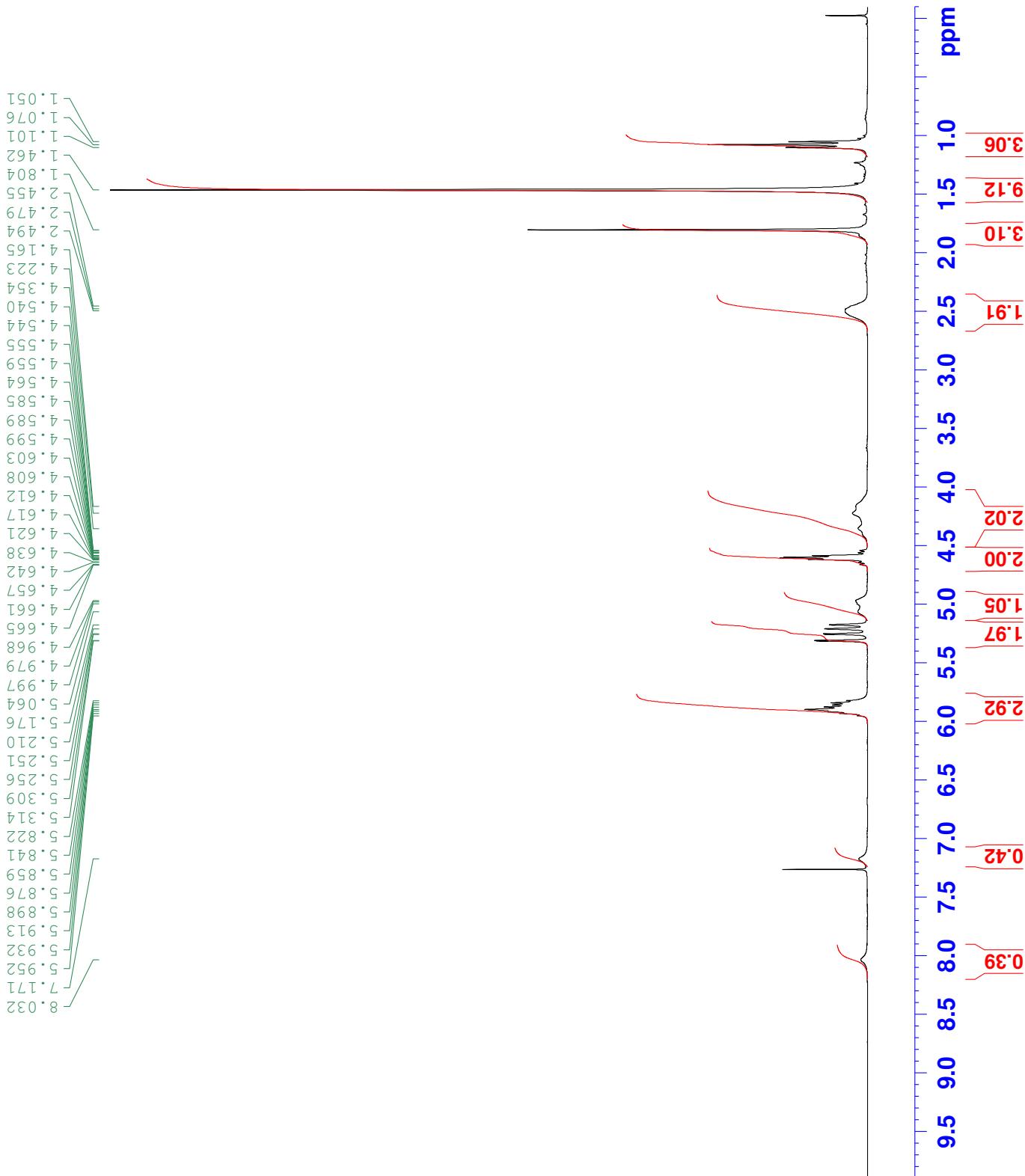




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 SOLVENT CDCl3
 NS 16
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 FIDRES 0.094423 Hz
 AQ 5.2953587 sec
 RG 50.8
 DW 80.800 usec
 DE 6.50 usec
 TE 290.7 K
 D1 1.0000000 sec

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 SF 7.5000000 W
 SFO1 300.1318534 MHz
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 SSB 0
 LB 0
 GB 0
 PC 1.00





Current Data Parameters
 NAME allylester
 EXPNO 1
 PROCNO

F2 - Acquisition Parameters
 Date_ 20140105
 Time 15.22
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 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1200
 DS 2
 SWH 18028.846 Hz
 FIDRES 0.275098 Hz
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 RG 203
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 DE 6.50 usec
 TE 292.0 K
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 D11 0.03000000 sec

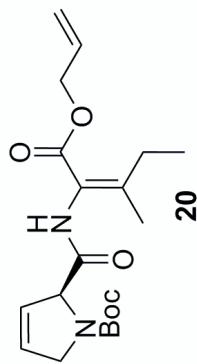
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 PLW12 0.20833001 W
 PLW13 0.16875000 W
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768
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 WDW EM
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 69.02
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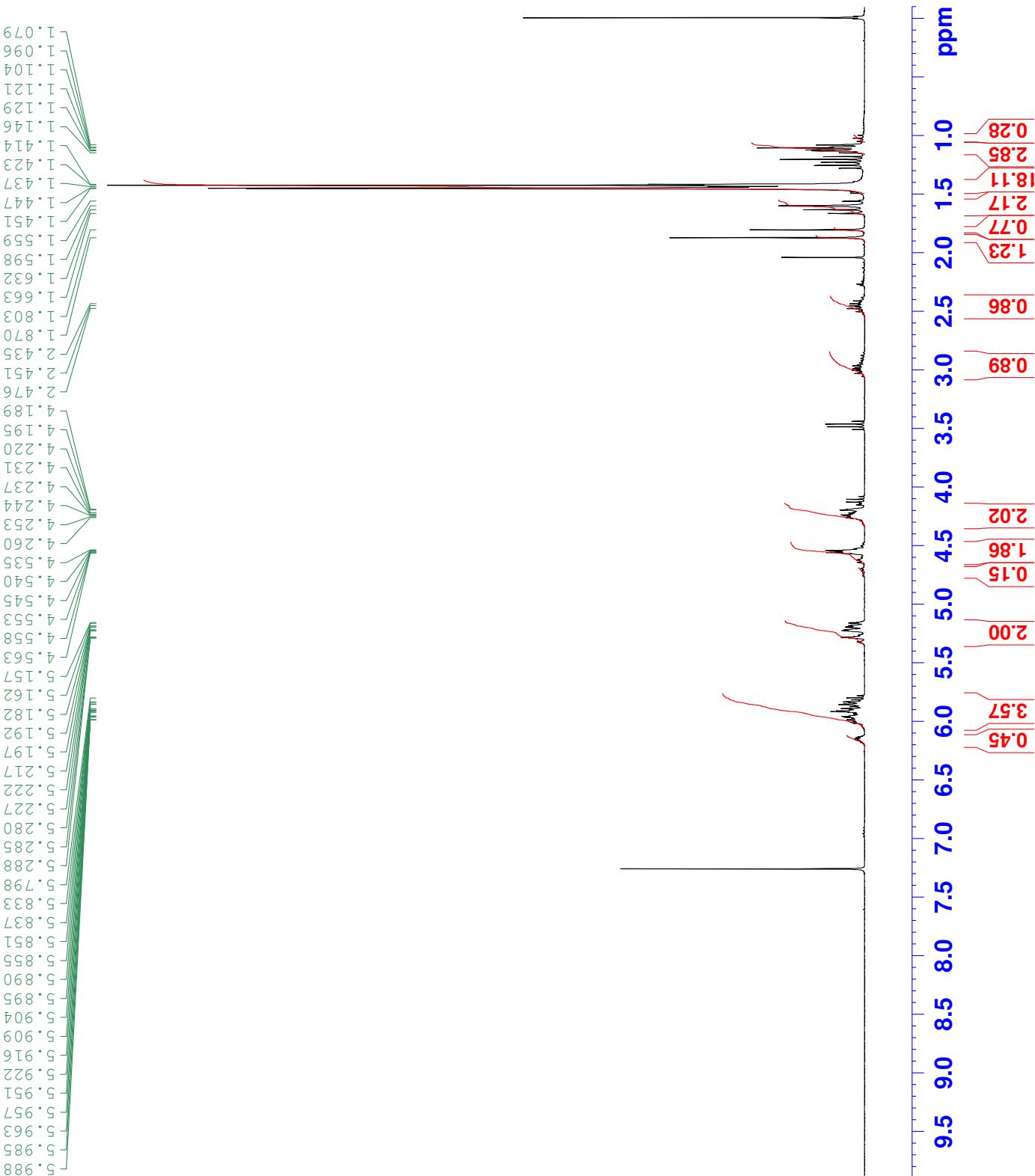
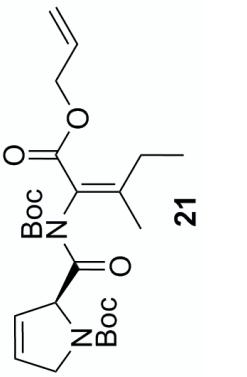




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 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6188.119 Hz
 FIDRES 0.094423 Hz
 AQ 5.2953587 sec
 RG 203
 DW 80.800 usec
 DE 6.50 usec
 TE 293.0 K
 D1 1.0000000 sec

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 P1 15.00 usec
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 SF 7.5000000 W
 SFO1 300.1318534 MHz
 WDW EM
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 PC 1.00





Current Data Parameters
 NAME pheno-Boc-Oallyl
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 PROCNO 1

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 PULPROG zgppg30
 TD 65536
 SOLVENT CDCl3
 NS 901
 DS 2
 SWH 18028.846 Hz
 FIDRES 0.275098 Hz
 AQ 1.8175818 sec
 RG 203
 DW 27.733 usec
 DE 6.50 usec
 TE 293.3 K
 D1 2.0000000 sec
 D11 0.03000000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PLW1 38.05099869 W
 SFO1 75.4752953 MHz

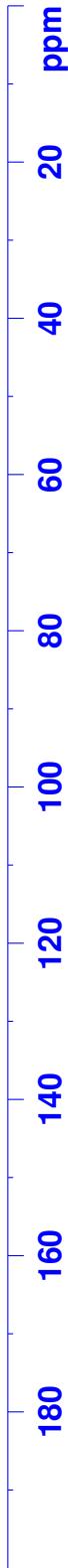
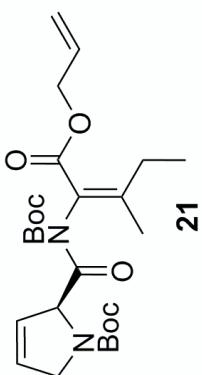
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PLW2 7.50000000 W
 PLW12 0.20833001 W
 PLW13 0.16875000 W
 SFO2 300.1312005 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4677566 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

12.06
 19.55
 19.69
 20.27
 20.36
 20.36
 27.36
 27.62
 28.08
 28.26

53.04
 53.47
 53.70
 64.80
 65.03
 68.36
 68.48
 68.80
 79.20
 79.40
 79.55
 82.99
 83.27

117.32
 117.52
 117.94
 118.03
 122.23
 122.46
 122.84
 122.96
 125.42
 125.64
 125.83
 128.46
 128.62
 129.07
 129.19
 131.77
 132.34
 132.78
 151.94
 153.34
 155.65
 155.64
 155.65
 155.85
 158.22
 158.50
 162.78
 162.88
 171.07
 171.52
 172.71
 172.79

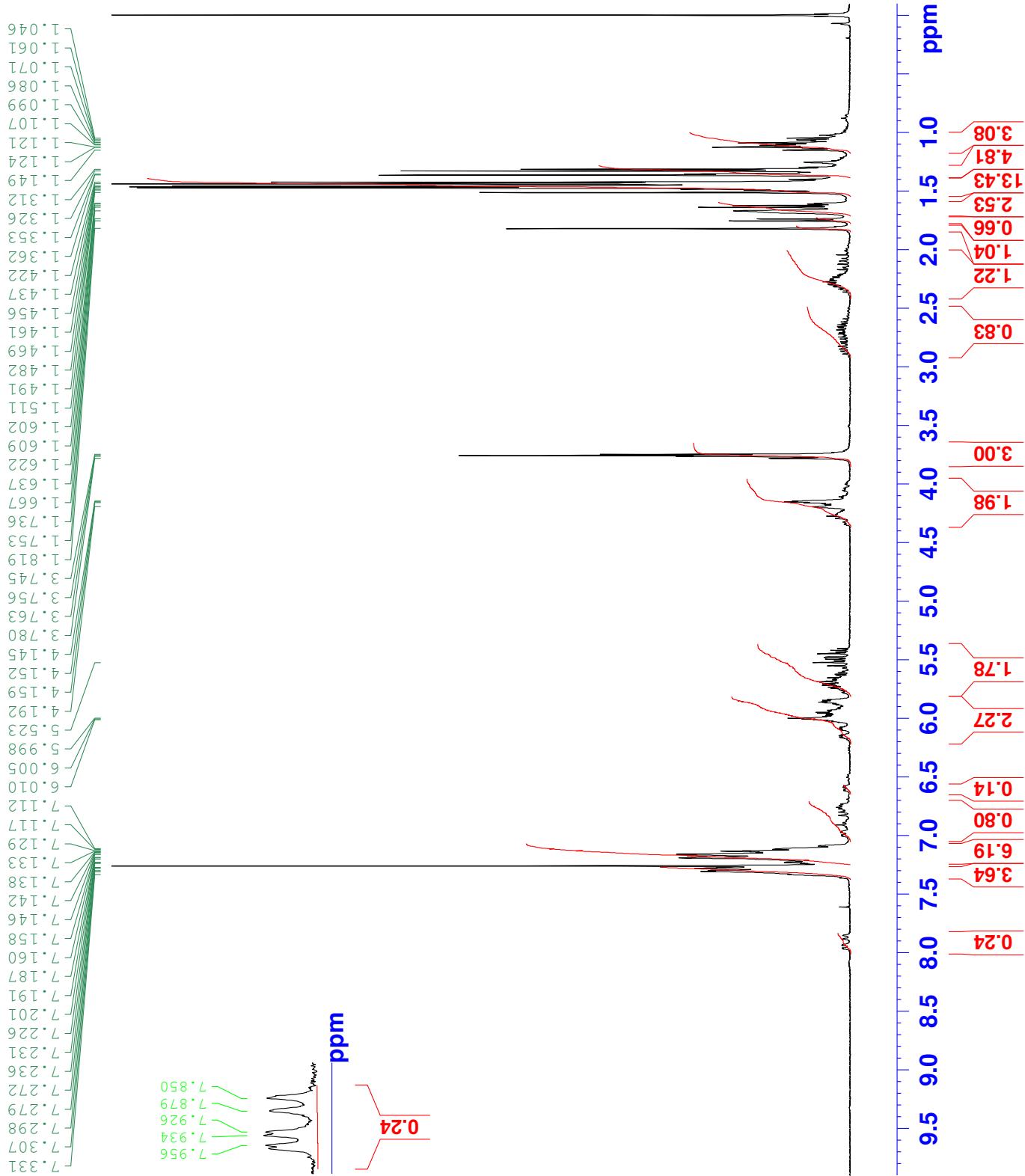




Current Data Parameters
 NAME 15-59
 EXPNO 20
 PROCN0 1

F2 - Acquisition Parameters
 Date_ 20131118
 Time 16.37
 INSTRUM av300
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6188.119 Hz
 FIDRES 0.094423 Hz
 AQ 5.2953587 sec
 RG 203
 DW 80.800 usec
 DE 6.50 usec
 TE 292.7 K
 D1 1.0000000 sec

===== CHANNEL f1 ======
 NUC1 1H
 P1 15.00 usec
 PLW1 32768
 SF 7.5000000 W
 SFO1 300.1318534 MHz
 WDW EM
 SSB 0
 LB 0
 GB 0
 PC 1.00





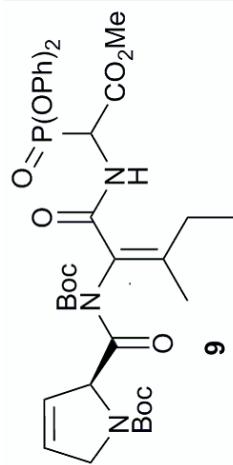
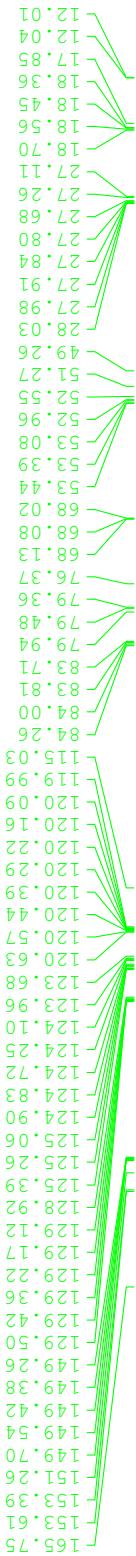
Current Data Parameters
NAME 15-59
EXPNO 40
PROCNO 1

F2 - Acquisition Parameters
Date_ 20131119
Time 1.38
INSTRUM av300
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 5120
DS 2
SWH 18028.846 Hz
FIDRES 0.275098 Hz
AQ 1.8175818 sec
RG 203
DW 6.50 usec
DE 293.5 K
TE 2.0000000 sec
D1 0.0300000 sec
D11

===== CHANNEL f1 =====
NUC1 CPDPRG2
P1 10.00 usec
PLW1 38.05099869 W
SFO1 75.4752953 MHz

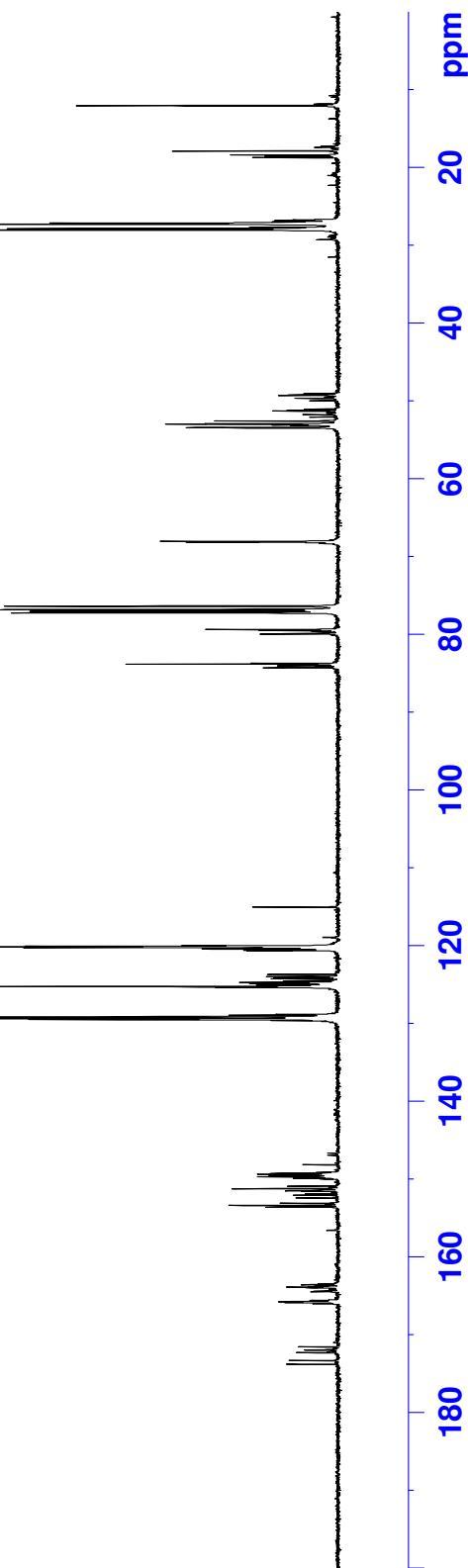
===== CHANNEL f2 =====
NUC2 waltz16
PCPD2 1H
PLW2 90.00 usec
PLW12 7.50000000 W
PLW13 0.20833001 W
SFO2 0.16875000 W
300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.4677785 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



173.80
173.31
172.29
172.00
171.95
171.56

174 ppm





Current NAME	Data EXPNO	Parameters	phomo-tripeptide	10	1
PROCNO					

F2 - Acquisition Parameters
Date_ 20131221
Time_ 15:09

INSTRUM	5	mm	PABBO	av300
PROBHD			BB-	
PULP/PROG			zg30	
TD			65536	
SOLVENT			CDC13	
NS			32	
DS			2	
SWH			61.88	.119
FIDRES			0.094423	Hz
AQ			5.2953587	sec
RG			32	
DW			80.800	usec
DE			6.50	usec
TE			292.7	K
D1			1.00000000	sec

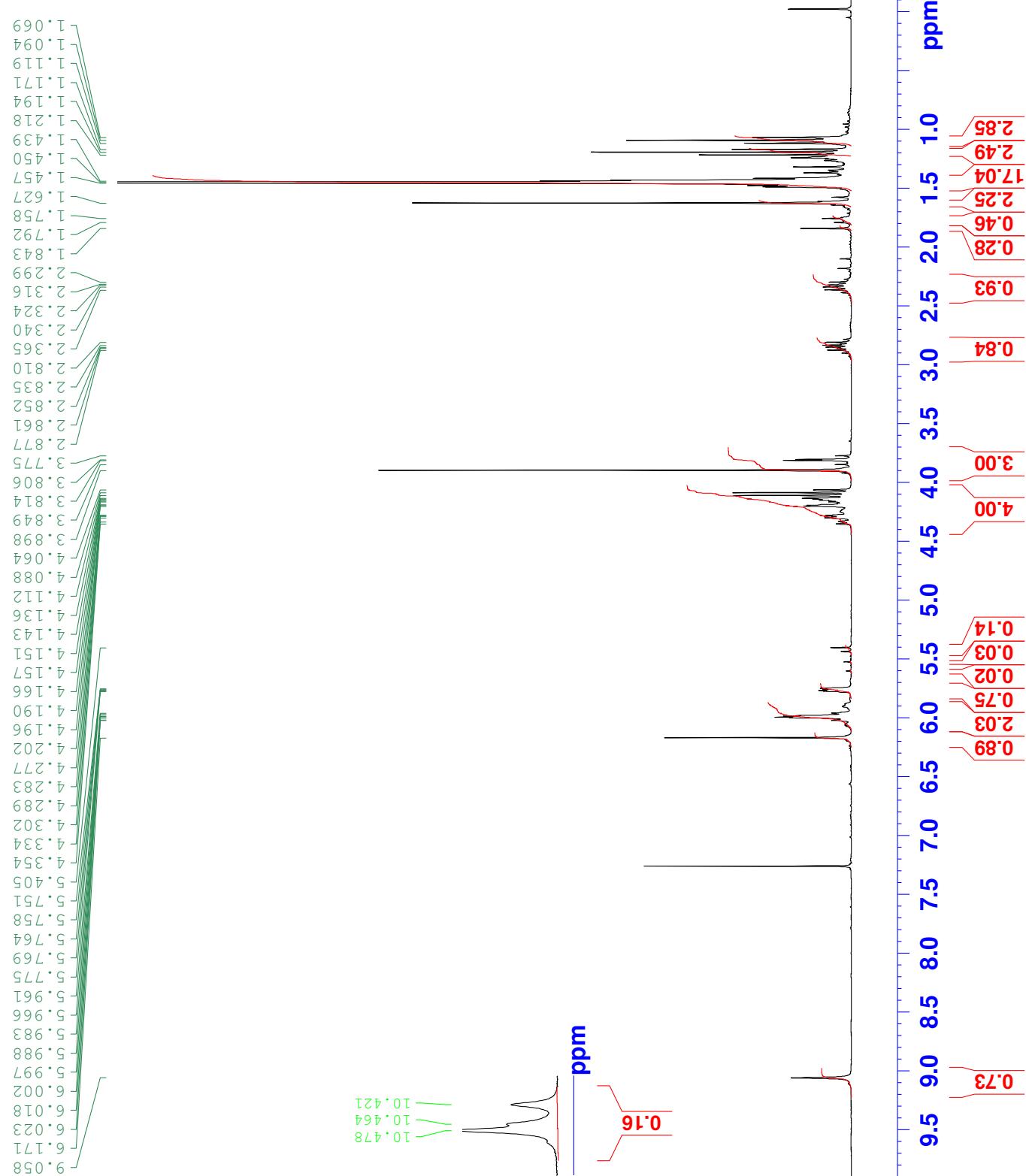
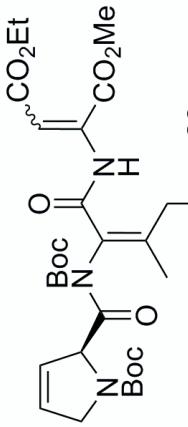
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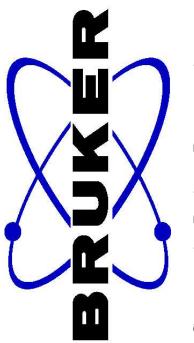
===== CHANNEL f1 =====
NUC1          1 H
P1           15.00 usec
PLW1        7.5000000 W
SFO1      300..1318534 MHz

F2 - Processing parameters
SI          32768
SF        300..1300062 MHz
EM

WDW          0
SSB          0   0..30 Hz
LB
GB
PC

```





Current Data Parameters
NAME 15-67
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters

Date_ 20131128
Time 8.12
INSTRUM av300
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 5000
DS 2
SWH 18028.846 Hz
FIDRES 0.275098 Hz
AQ 1.8175818 sec
RG 203
DW 27.733 usec
DE 6.50 usec
TE 293.3 K
D1 2.0000000 sec
D11 0.03000000 sec

===== CHANNEL f1 =====

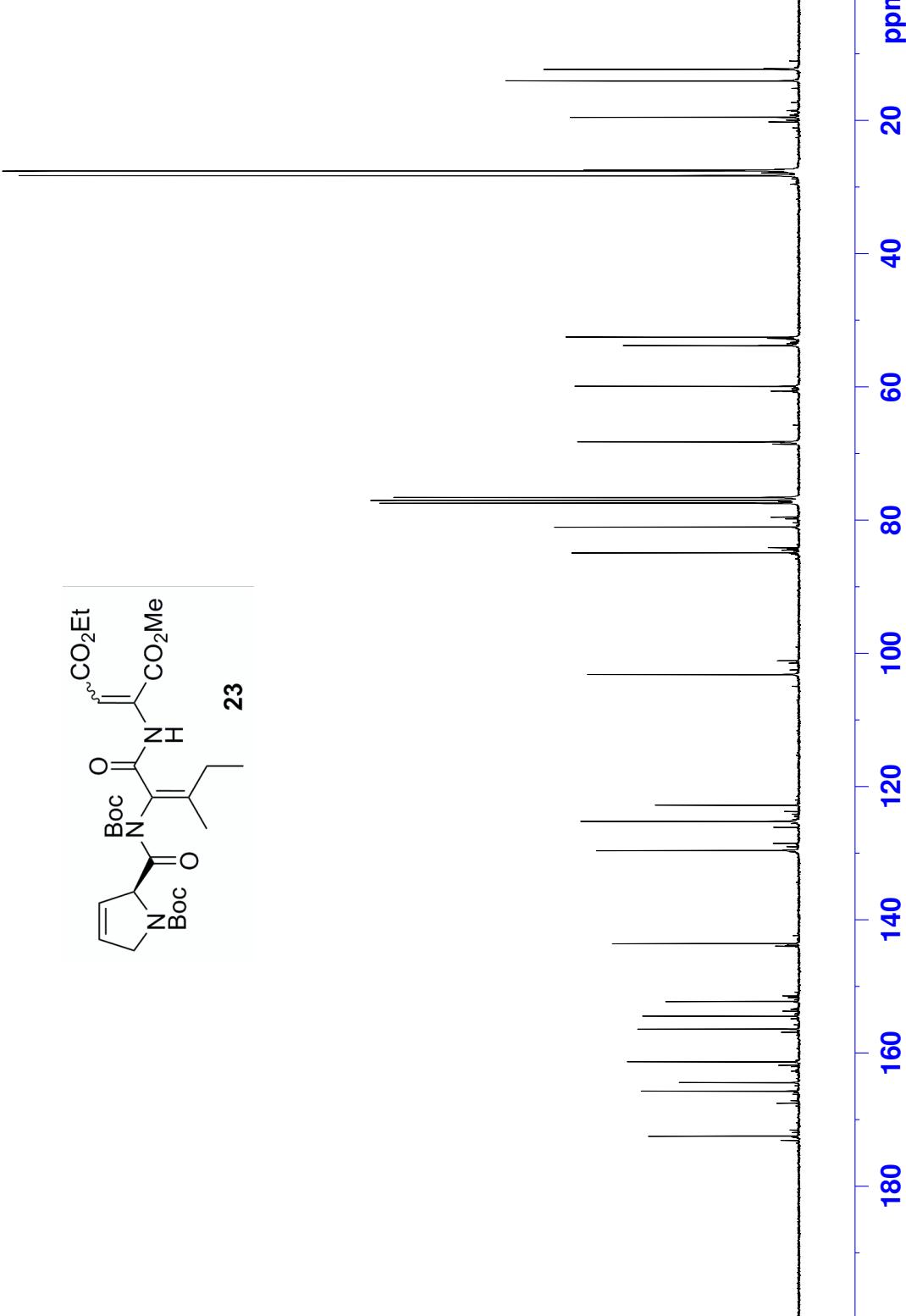
NUC1 13C
P1 10.00 usec
PLW1 38.05099869 W
SFO1 75.4752953 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 90 usec
PLW2 7.50000000 W
PLW12 0.20833001 W
PLW13 0.16875000 W
SFO2 300.1312005 MHz

F2 - Processing parameters

SI 32768
SF 75.4677559 MHz
WDW EM
SSB LB 1.00 Hz
GB 0 1.40
PC

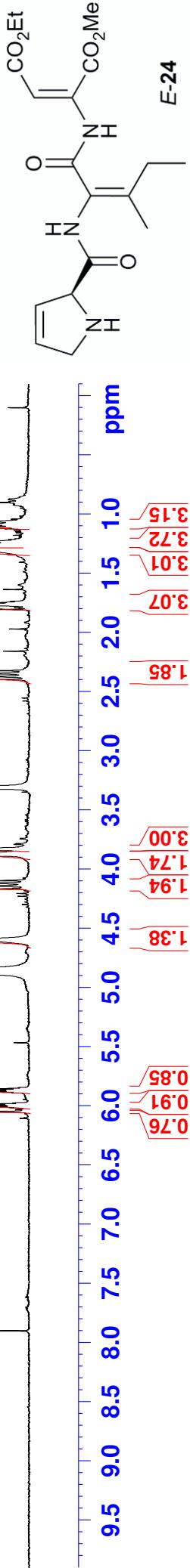




Current Data Parameters
 NAME PLC-4
 EXPNO 10
 PROCN0 1

F2 - Acquisition Parameters
 Date_ 20151011
 Time 13.56
 INSTRUM av300
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT MeOD
 NS 16
 DS 2
 SWH 6188.119 Hz
 FIDRES 0.094423 Hz
 AQ 5.2953587 sec
 RG 203
 DW 80.800 usec
 DE 6.50 usec
 TE 294.4 K
 D1 1.0000000 sec

===== CHANNEL f1 ======
 NUC1 1H
 P1 15.00 usec
 PLW1 327.68 MHz
 SF 7.5000000 W
 SFO1 300.1318534 MHz
 WDW EM
 SSB 0
 LB 0
 GB 0
 PC 1.00



1.049
 6.016
 6.002
 5.996
 5.990
 5.983
 5.977
 5.888
 5.881
 5.873
 5.862
 5.854
 5.852
 4.167
 4.143
 4.120
 4.096
 3.876
 3.831
 2.397
 2.372
 2.347
 2.322
 1.768
 1.743
 1.710
 1.687
 1.290
 1.277
 1.253
 1.229
 1.112
 1.087
 1.062