

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29

## Supplemental Material

# Spatiotemporal characteristics of organic contaminant concentrations and ecological risk assessment in the Songhua River, China

Ce Wang<sup>a,\*</sup>, Mike Cyterski<sup>b,\*</sup>, Yujie Feng<sup>c</sup>, Peng Gao<sup>c</sup>, Qingfang Sun<sup>c</sup>

<sup>a</sup>State Key Laboratory of Pollution Control & Resource Reuse, School of the Environment, Nanjing University, Nanjing, 210023, P.R. China

<sup>b</sup>Ecosystems Research Division, National Exposure Research Laboratory, Office of Research and Development, U.S. Environmental Protection Agency

<sup>c</sup>State Key Laboratory of Urban Water Resource and Environment, Harbin Institute of Technology, No 73 Huanghe Road, Nangang District, Harbin 150090, P.R. China

\*Corresponding Author

State Key Laboratory of Pollution Control & Resource Reuse  
School of Environment, Nanjing University  
No. 163 Xianlin Avenue, Nanjing, 210023, P.R. China

Tel: 86-25-89680535; Fax: 86-25-89680535

E-mail: wangce@nju.edu.cn

Ecosystems Research Division  
National Exposure Research Laboratory  
Office of Research and Development, U.S. Environmental Protection Agency  
960 College Station Road, Athens, GA 30605-2700

Tel: 01-706-355-8142

E-mail: cyterski.mike@epa.gov

## 30 **1. Analysis of PAHs**

### 31 **1.1 Water sample pretreatment**

32 A 1L water sample was placed into a 2L separator funnel. 30g NaCl was then  
33 added and dissolved, followed by 50mL dichloromethane. The funnel was shaken  
34 (10min) and allowed to settle (5min). The organic phase was transferred to a flask.  
35 This extraction process was repeated twice and the extracts combined. Next, 3g  
36 Na<sub>2</sub>SO<sub>4</sub> was added to the organic phase and agitated; after twenty minutes, the  
37 solution was filtered and transferred to a rotary evaporator until 3mL remained. This  
38 was further condensed to 1mL by N<sub>2</sub> stream stripping. 10mL of n-hexane was put into  
39 the extract and re-condensed to 1mL, again by N<sub>2</sub> stream stripping. The extract was  
40 put through a silica gel chromatograph, activated by 10mL acetone-n-hexane and  
41 10mL of n-hexane and cleaned using 10mL of 10% acetone-n-hexane. 1mL eluate  
42 condensed by N<sub>2</sub> stream stripping with addition of 10μL of surrogate perdeuterated  
43 naphthalene, perdeuterated acenaphthene, perdeuterated chrysene, perdeuterated  
44 phenanthrene and perdeuterated fluoranthene (10μg/mL). Finally, the solution was  
45 supplemented to 1.0mL for GC-MS determination.

### 46 **1.2 GC-MS analysis**

47 PAH concentrations were determined by a GC-6980N/MS-5973N, Agilent  
48 Technologies Inc. DB-5MS chromatogram column (J&W Scientific; 30m×0.32mm,  
49 0.25μm film thickness). Column temperature was kept at 353°K for 2min and

50 increased to 563°K for an additional 5min. The instrument was operated in selected  
51 ion mode (SIM) with a detection range of ~35-400m/z. Additional details of the  
52 analytical methods can be found in (MEP, 2002). Samples were analyzed for 8 PAHs  
53 [pyrene (PYR); fluorene (FLU); chrysene (CHR); anthracene (ANT); naphthalene  
54 (NAP); fluoranthene (FLA); acenaphthene (ACP); phenanthrene (PHE)].

### 55 **1.3 Precision and accuracy**

56 All analytical data were subjected to a strict QA/QC. Blank samples were  
57 included and all concentrations were blank corrected. All concentration data were the  
58 average of three replicates for each sample. 1L of pure water was prepared, and then  
59 added with 50ng perdeuterated PAH standard solution (solvent is acetone). After this,  
60 pretreatment and analysis procedure of water sample were the same as the content  
61 mentioned above. The recovery of each PAH in standard solution and the relative  
62 standard deviations (RSDs) were 85%-95% and 4%-8%, respectively.

## 63 **2. Analysis of Phenols**

### 64 **2.1 Water sample pretreatment**

65 A 1L water sample was placed into a 2L separator funnel, and the solution's pH  
66 was adjusted to 2-3 using 6mol/L HCl solution. 30g NaCl was added and dissolved,  
67 50mL dichloromethane was added, the vessel was then shaken for 10min and allowed  
68 to stand for 5min. The dichloromethane fraction was transferred to a flask. This  
69 process was repeated and extracts combined. 3g Na<sub>2</sub>SO<sub>4</sub> was added and the solution

70 agitated. After standing for 20min, the solution was filtrated and transferred to a  
71 rotary evaporator for concentration to 1mL. This volume was then transferred to a  
72 K-D concentrator using dichloromethane and condensed to 0.5mL using N<sub>2</sub> stream  
73 stripping. 100uL of BSTFA was added to the solution after standing for 1h at room  
74 temperature. Finally, the solution was added with 10mL of perdeuterated naphthalene  
75 (surrogate, purity 99%) and reconstituted in 1mL of dichloromethane for  
76 quantification.

## 77 **2.2 GC-MS analysis**

78 Phenolic compound concentrations were determined by a GC-6980N/MS-5973N,  
79 Agilent Technologies Inc. DB-5MS chromatogram column (J&W Scientific;  
80 30m×0.32mm, 0.2µm film thickness). Column temperature was kept at 328°K for  
81 2min, increased to 378°K, then 478°K, and finally 578°K for 5min. The pressure of  
82 He gas was kept at 40kPa for 5min, then increased to 70kPa and kept for 5min. The  
83 instrument was operated in selected ion mode (SIM) with a detection range of  
84 ~35-400m/z. Additional details of analytical methods can be found in (MEP, 2002).  
85 Samples were analyzed for 8 phenols [phenol (PHN); 2-nitrophenol (2-NP);  
86 4-nitrophenol (4-NP); 2-chlorophenol (2-CP); 2,4-dichlorophenol (2,4-DCP);  
87 2,4-dimethylphenol (2,4-DMP); 2,4,6-trichlorophenol (2,4,6-TCP); 4-chloro-*m*-cresol  
88 (4-CMC)].

## 89 **2.3 Precision and accuracy**

90 All analytical data were subjected to a strict QA/QC. Blank samples were  
91 included and all concentrations were blank corrected. All concentration data were the  
92 average of three replicates for each sample. 50 $\mu$ L of the mixed standard solution of  
93 2,4-Dichlorophenol and 2,4-Dinitrophenol was added into 1L of pure water, and then  
94 it was transferred to 2L separator funnel. After this, pretreatment and analysis  
95 procedure of water sample were the same as the content mentioned above. The  
96 recovery of phenolic compound in standard solution and the relative standard  
97 deviations (RSDs) were 98%-101% and 1.7%-6.7%, respectively.

98

99

## 100 **References**

101

102 MEP. 2002. Monitoring and analysis methods for water and wastewater (Chinese). Fourth ed. China  
103 Environmental Science Press, Beijing.

104