

New Journal of Chemistry

Supporting Information To:

Efficient and selective azidation of *per-O*-acetylated sugars using ultrasound activation: Application to the one-pot synthesis of 1,2,3-triazole glycosides

Hamid Marzag,^{a,b†} Soukaina Alaoui,^{a,b†} Hella Amdouni,^a Anthony R. Martin,^a Khalid Bougrin^{*b} and Rachid Benhida^{*a}

^a Institut de Chimie de Nice UMR UNS-CNRS 7272, Université Nice Sophia Antipolis, Parc Valrose, 06108 Nice Cedex 2, France.
benhida@unice.fr

^b Laboratoire de Chimie des Plantes et de Synthèse Organique et Bioorganique, URAC23, Université Mohammed V-Agdal, Faculté des Sciences B.P. 1014 Rabat, Morocco. kbougrin@yahoo.fr

Table of contents

Synthetic procedures	2
Selected spectral data ¹ H, ¹³ C	3 - 28
Selected HRMS data	29- 41

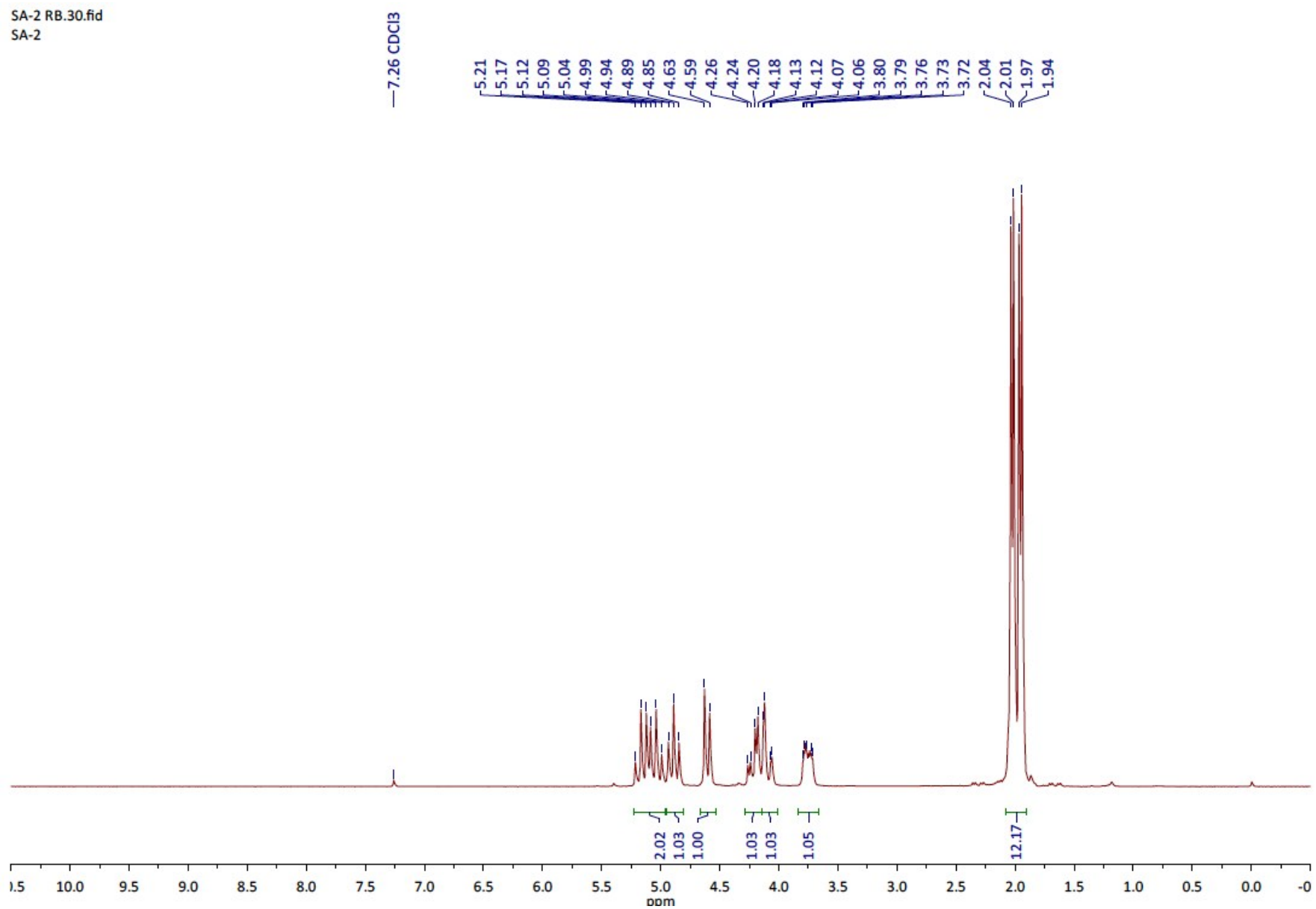
Experimental section

General. All organic solvents were purchased from commercial sources and used as received or dried using standard procedures, unless otherwise stated. All chemicals were purchased from Aldrich, Merck or Alfa Aesar and used without further purification; thin layer chromatography (TLC) was performed on precoated Merck 60 GF254 silica gel plates and revealed by spraying (p-anisaldehyde or H₂SO₄/EtOH), and detection by means of UV light at 254 and 360 nm. ¹H and ¹³C NMR spectra were recorded on a Bruker Avance 200 MHz spectrometer. Mass spectra (ESI MS) were recorded on a Bruker (Daltonics Esquire 3000+). HRMS spectra were carried out on a ThermoFisher Q Exactive plus in ESI mode positive and negative depending on the compounds to identify. We use a pump syringe at a flow of 3ul/mn and with the mass spectrometer at a resolution of 140 000 at m/z 200 for best accuracy. The purity of compounds was further verified to be >95% by HPLC analysis using analytical columns Hypersil (C18 (ELITE), 4.6 mm x 250 mm) or Nucleosil (120-5C8 (HICHROM), 4.6 mm x 250 mm) with an isocratic elution of CH₃CN/H₂O, 90/10. The ultrasound-assisted reactions were carried out in a “Branson Bransonic® 5510 DTH UltraSonic Bath Cleaner”, with a frequency of 40 kHz. The ultrasonic cleaner has a power consumption of 185W (399 × 371 × 401 mm) with liquid holding capacity of 9.5 L.

General procedure for the synthesis of azidoglycoside (2a-e). To a cold suspension of sodium azide (2 mmol) in dichloromethane (5 mL), sulfonyl chloride (1mmol) is added drop wise. After the completion of the addition the mixture is sonicated for few minutes, and then the acetylated sugar **1a-e** (2 mmol) and Lewis acid catalyst (20 mol %) are added to the mixture. The reaction mixture is sonicated during 45 min. After the completion of the reaction (TLC monitoring), the mixture was diluted with dichloromethane and washed with a saturated aqueous solution of NaHCO₃. The organic layer was washed with water (2×10 mL), dried over MgSO₄, filtered and the solvent was removed under reduced pressure. The residue was subjected to purification by silica gel column chromatography [Cyclohexane-EtOAc (9:1)] to give the pure azidoglycoside **2a-e**.

General procedure for one-pot synthesis of 1,2,3-triazolyl glycosides. To a cooled suspension of sodium azide (2 mmol) in dichloromethane (5 ml), sulfonyl chloride (1 mmol) is added drop wise. After the completion of the addition the mixture was sonicated at room temperature. The sugar derivative (2 mmol) and anhydrous FeCl₃ catalyst (20 mol %) were added and sonication continued. After reaction completion (TLC monitoring), the alkyne (4 mmol), CuI (4 mmol) and diisopropylethylamine (4 mmol) were added to the mixture and then left under sonication. After completion of the reaction (TLC monitoring), the mixture was diluted with dichloromethane and successively washed with a saturated solution of NH₄Cl and water (2×10 mL), dried over MgSO₄, filtered and concentrated under reduced pressure. The crude residue was purified by silica gel column chromatography (Cyclohexane-EtOAc 8:2 to 5:5) to afford the triazolyl glycosides **3**.

Figure S1. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **2c**.



SA-2 RB.40.fid
SA-2

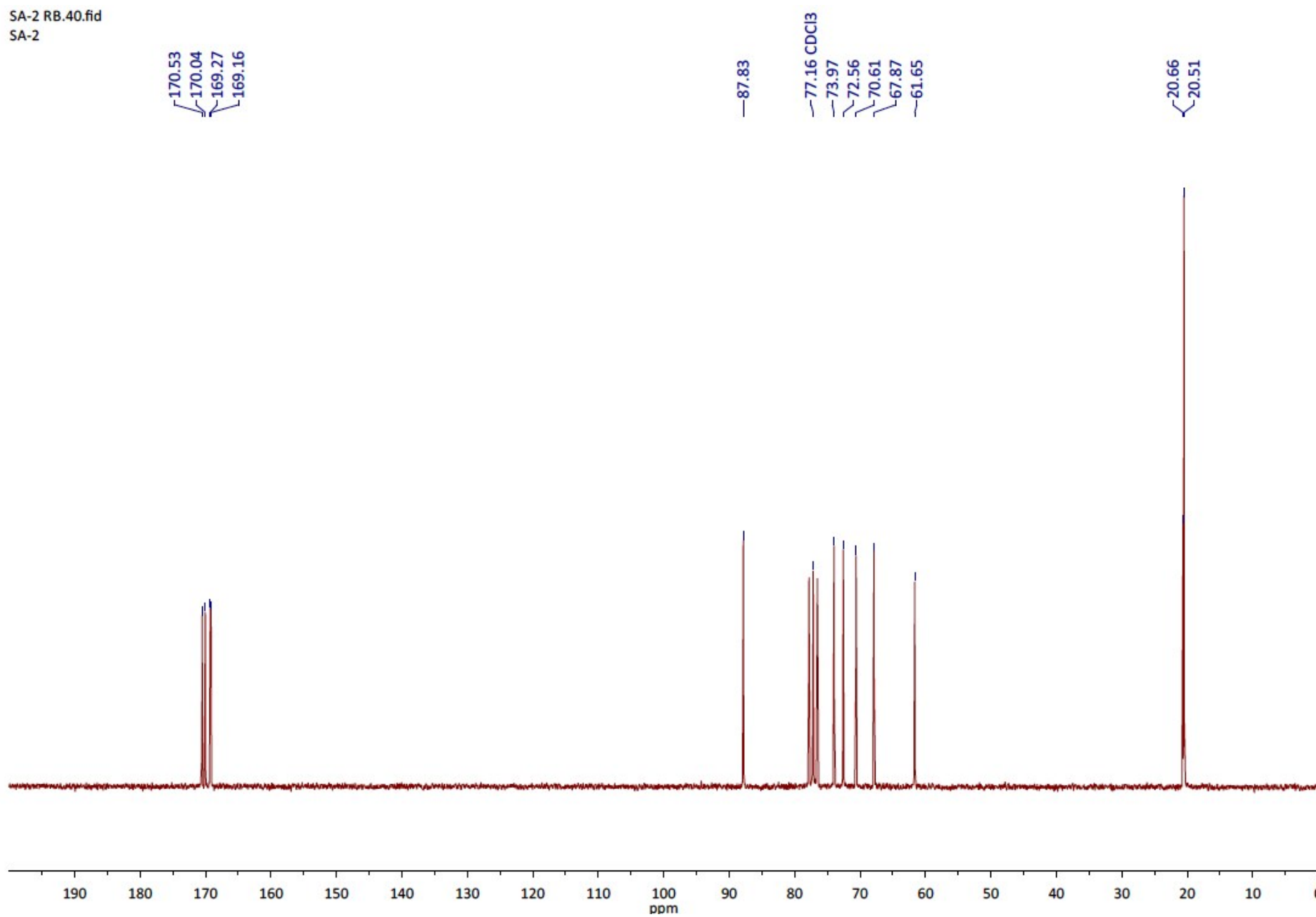
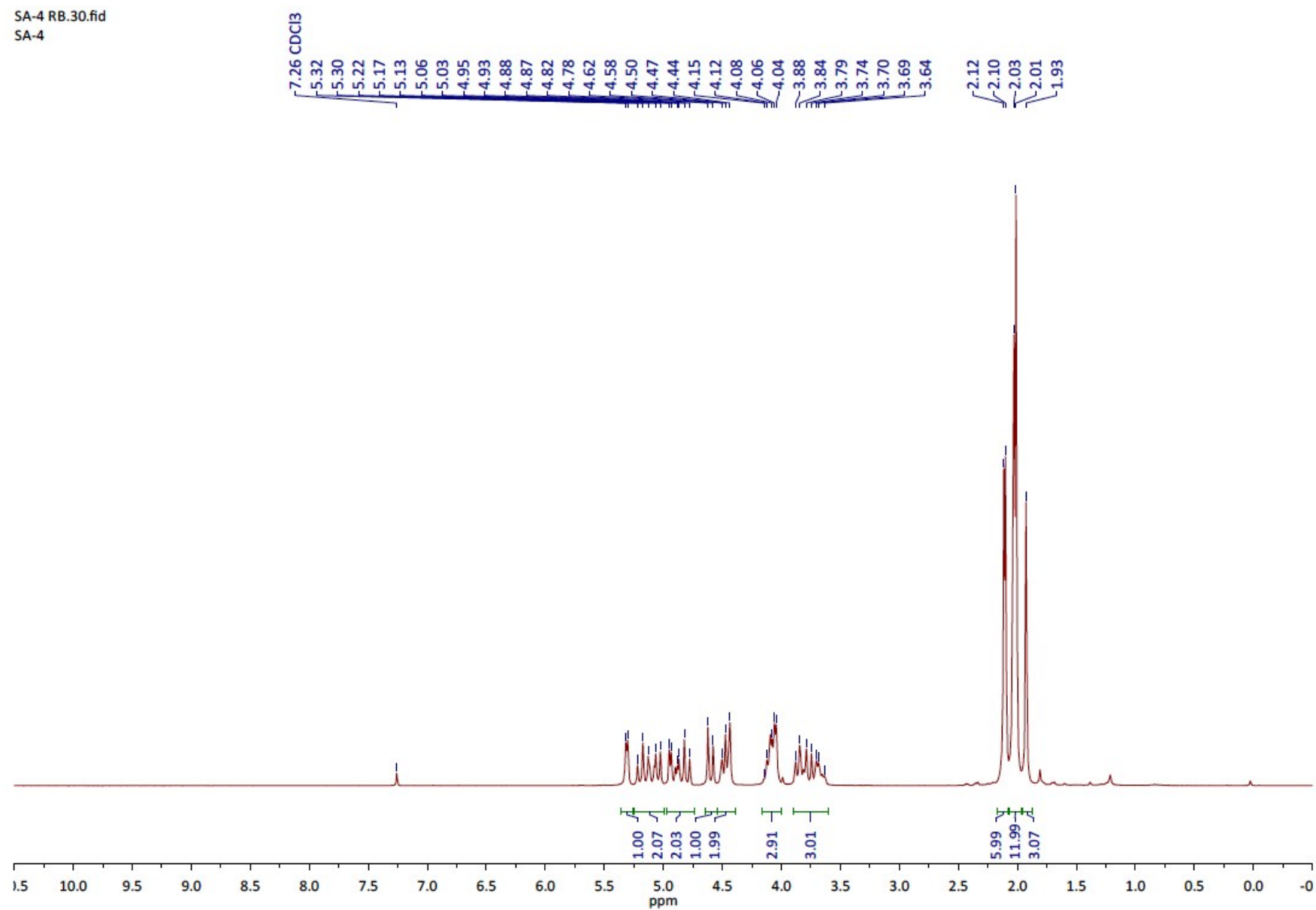


Figure S2. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **2d**.



SA-4 RB.40.fid
SA-4

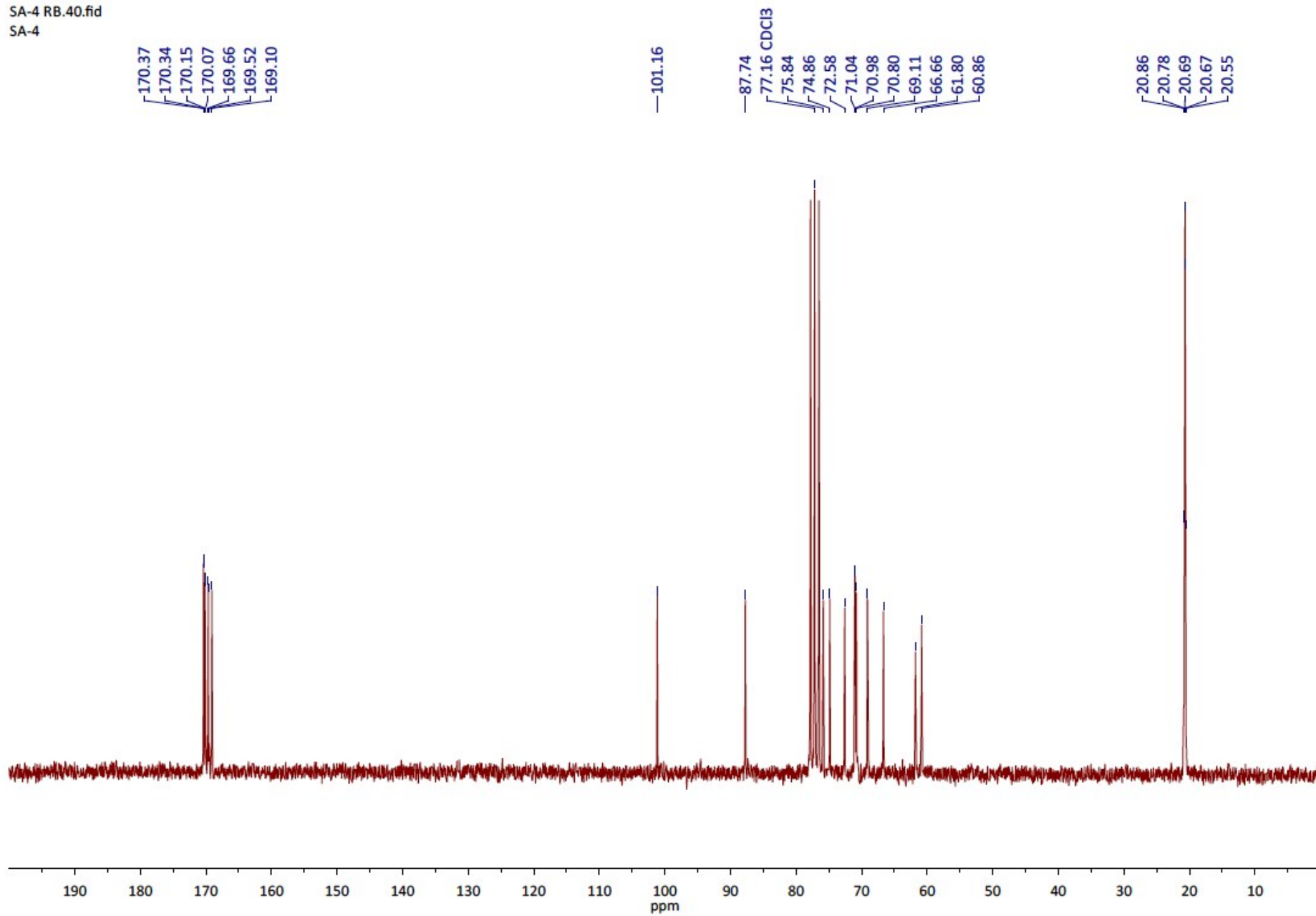
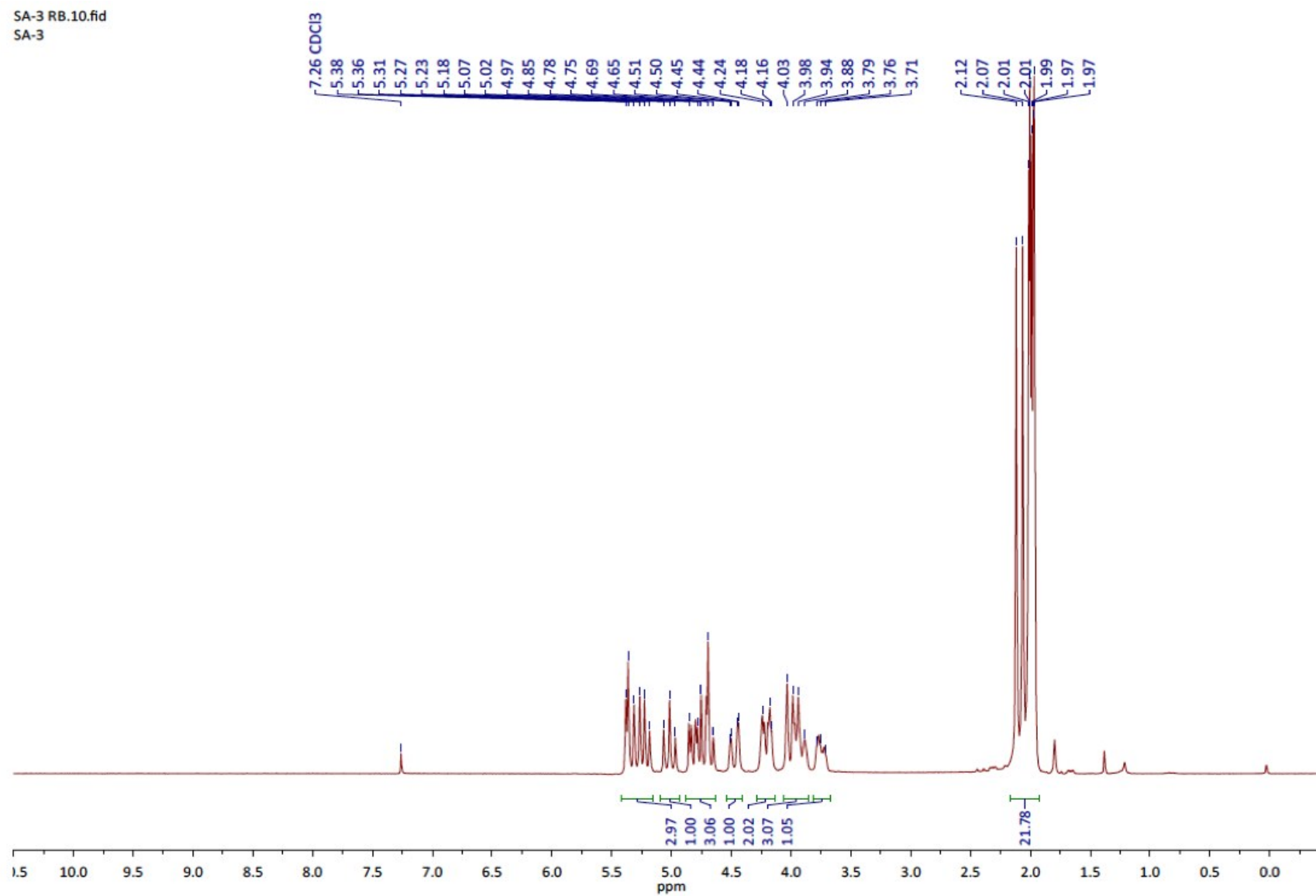


Figure S3. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **2e**.



SA-3 RB.20.fid
SA-3

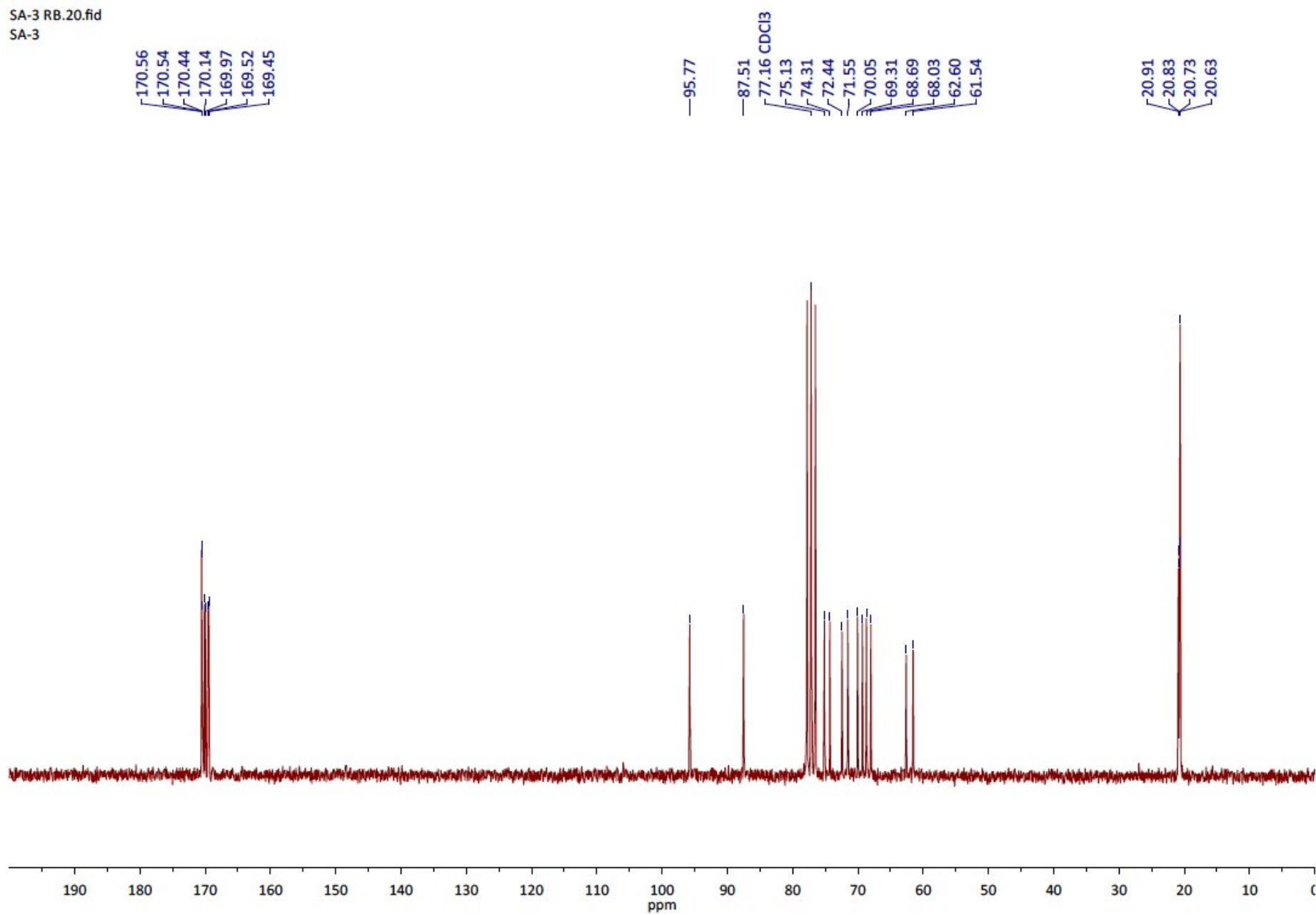
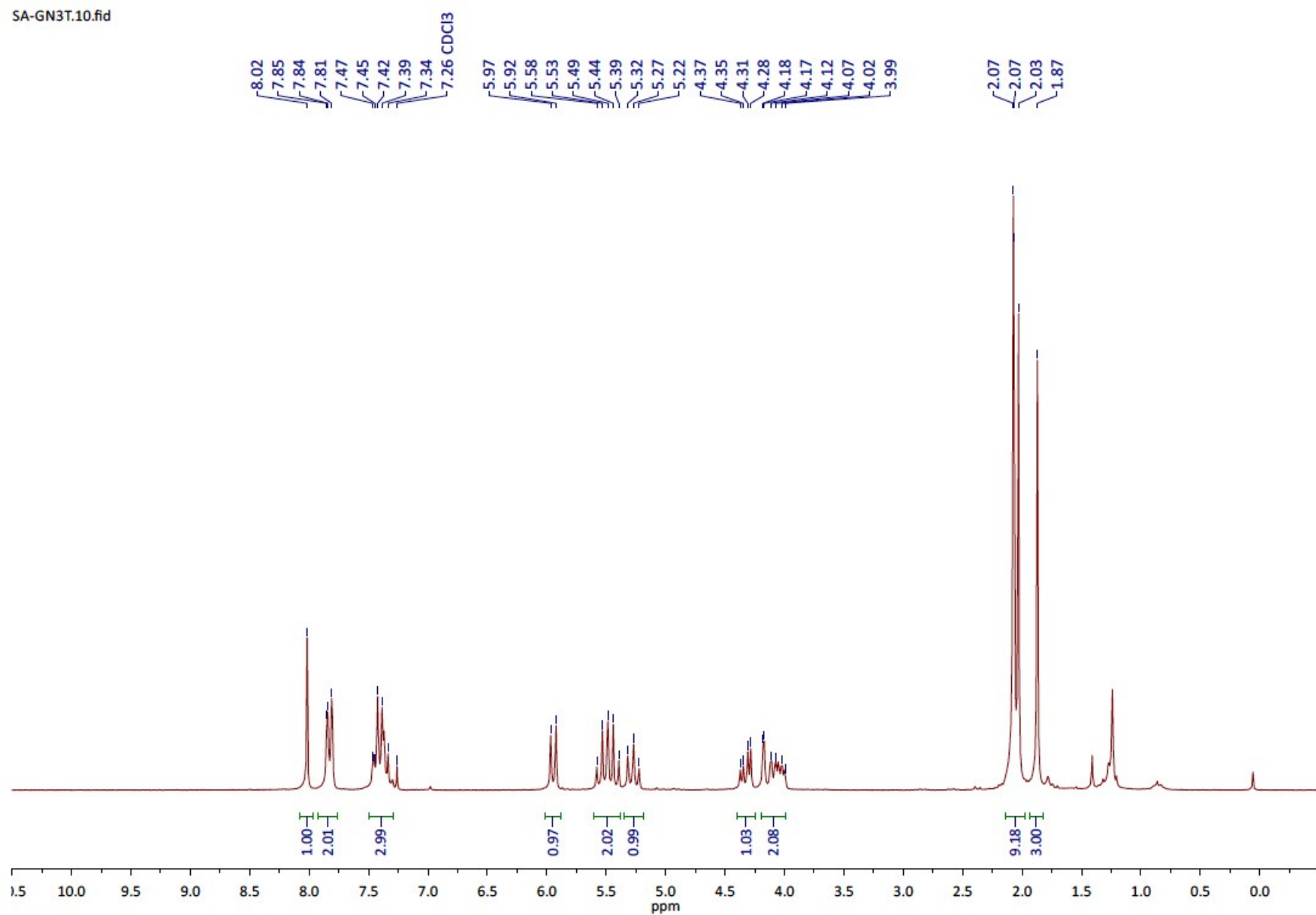


Figure S4. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **3b**.



SA GN3T.10.fid

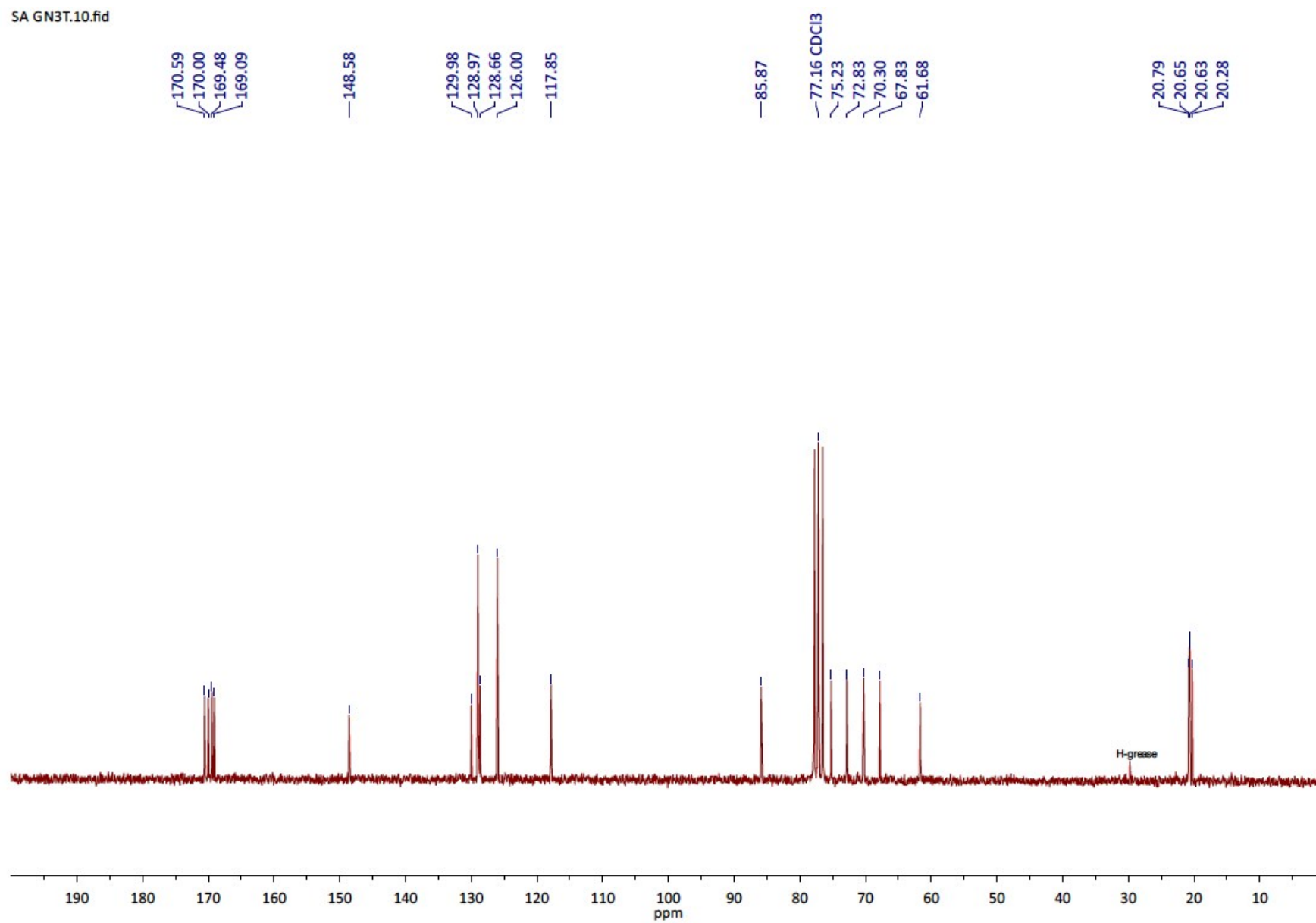
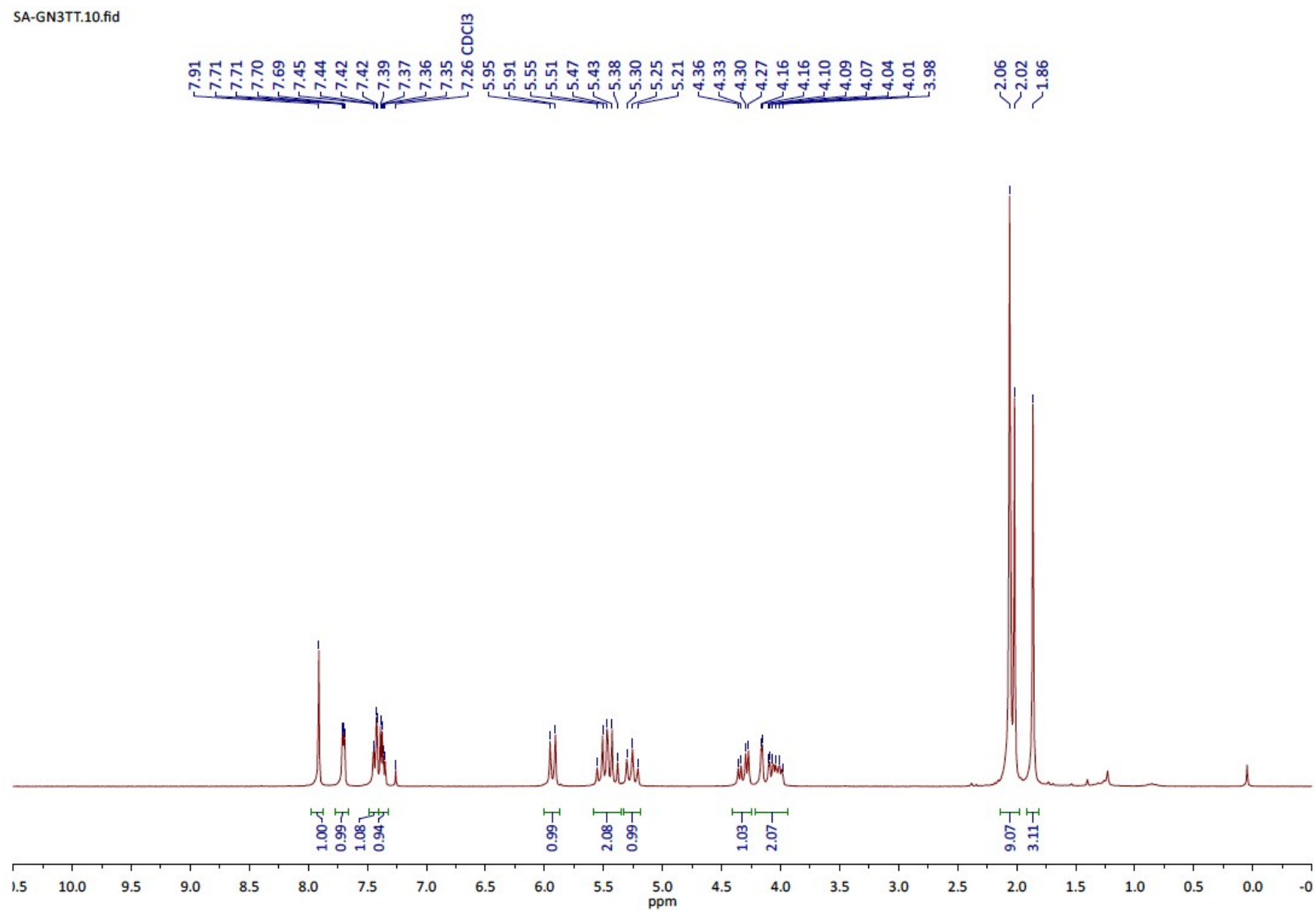


Figure S5. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **3c**.



SA GN3TT.10.fid

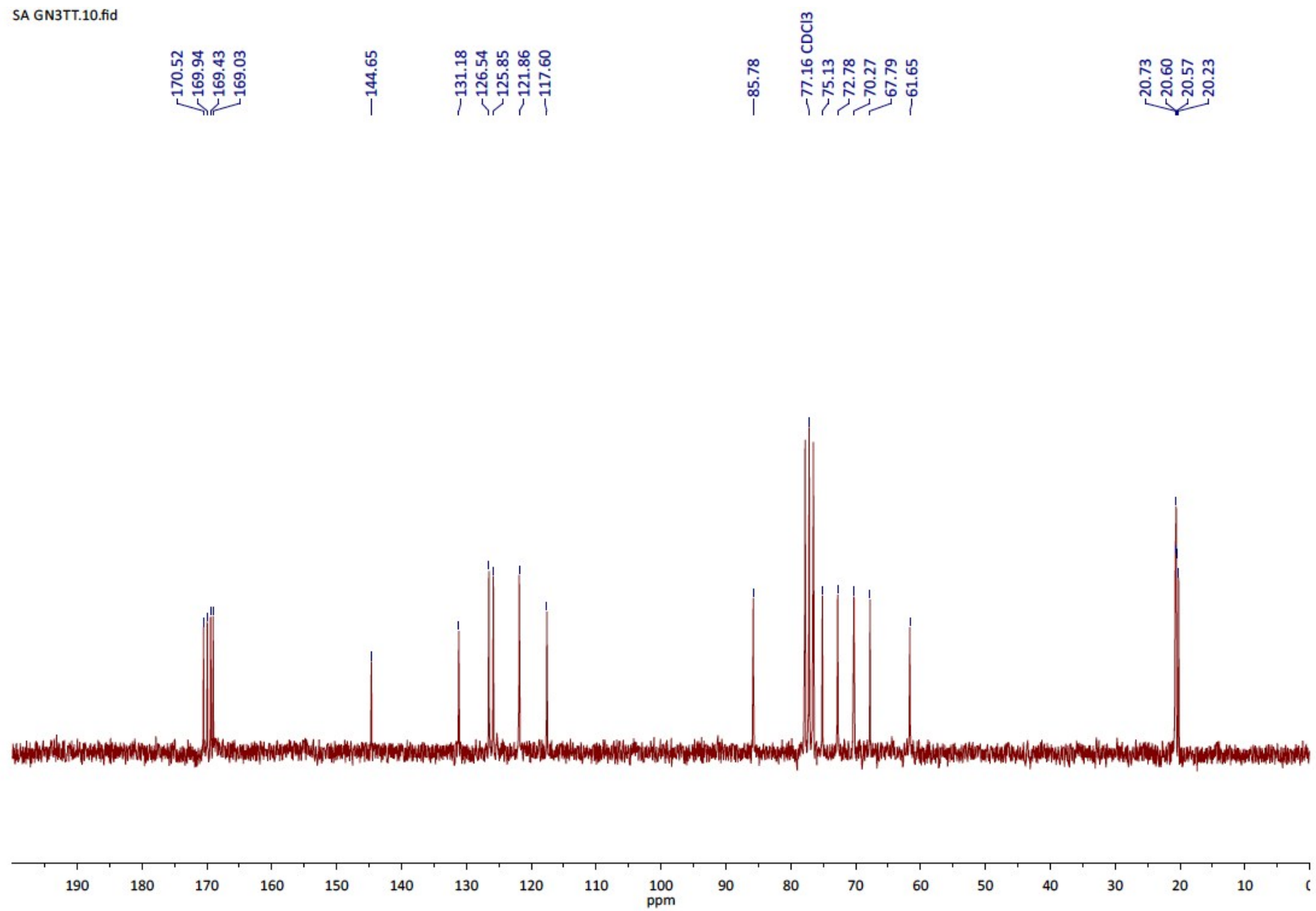
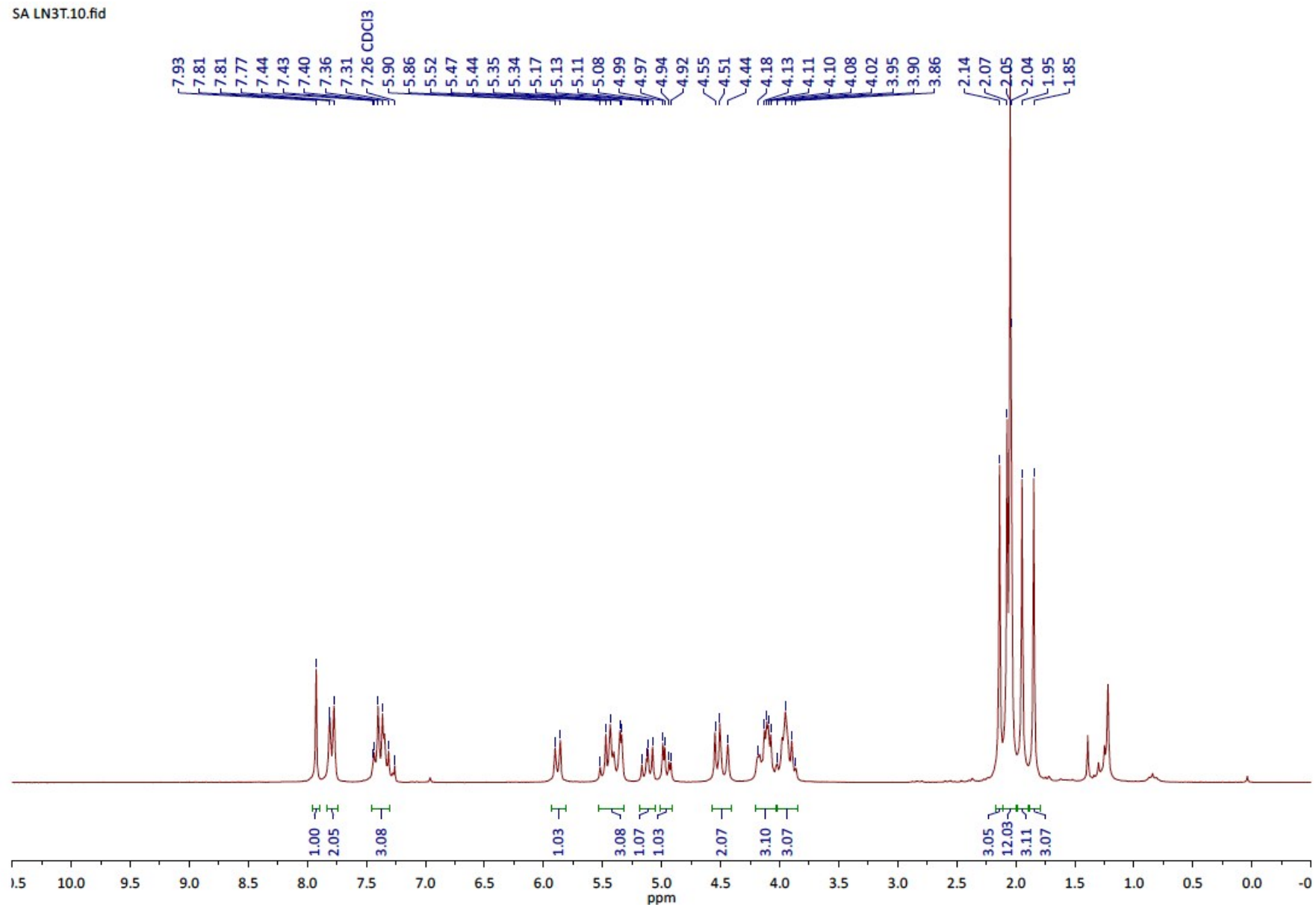


Figure S6. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **3d**.

SA LN3T.10.fid



SA LN3T.10.fid

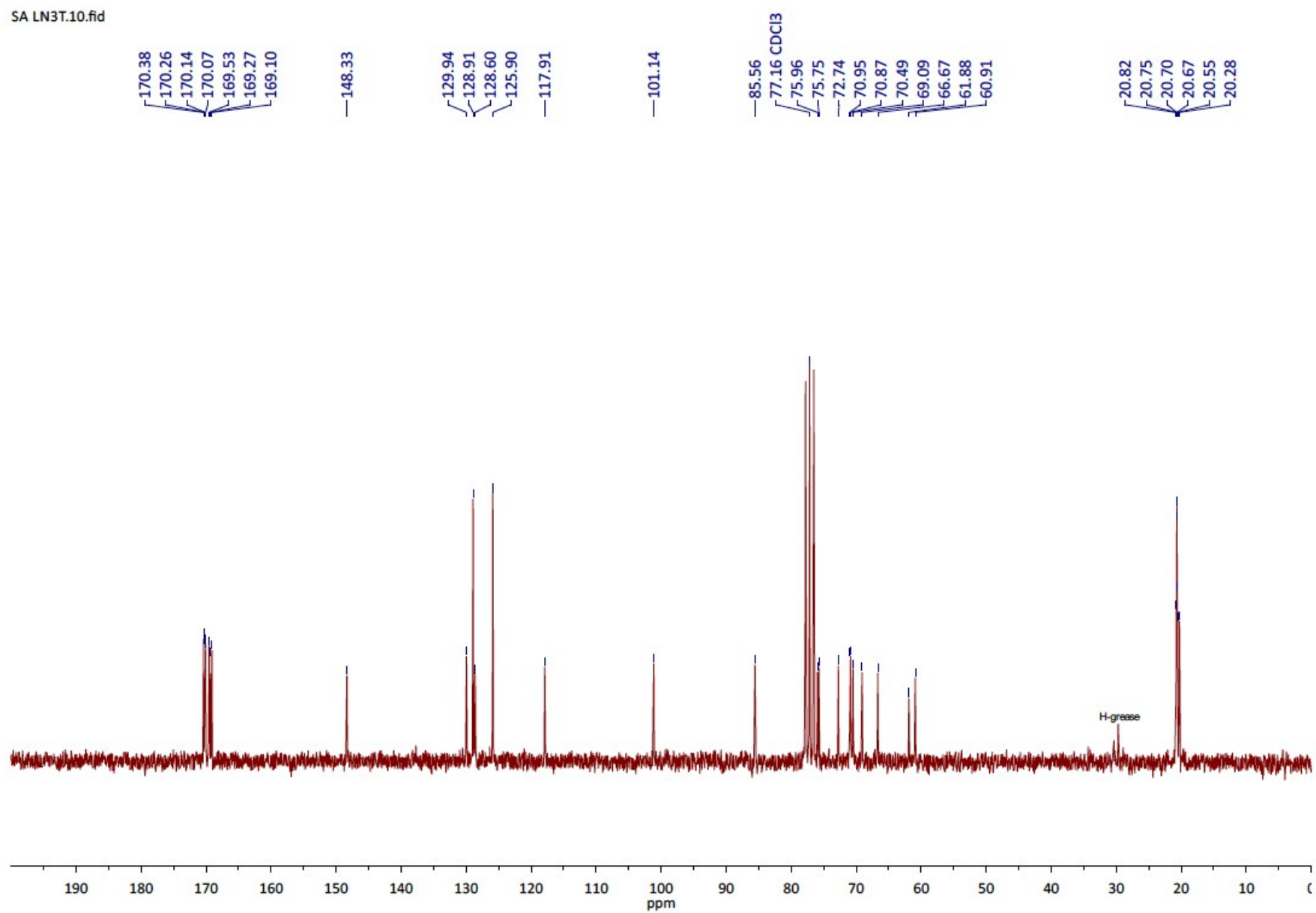
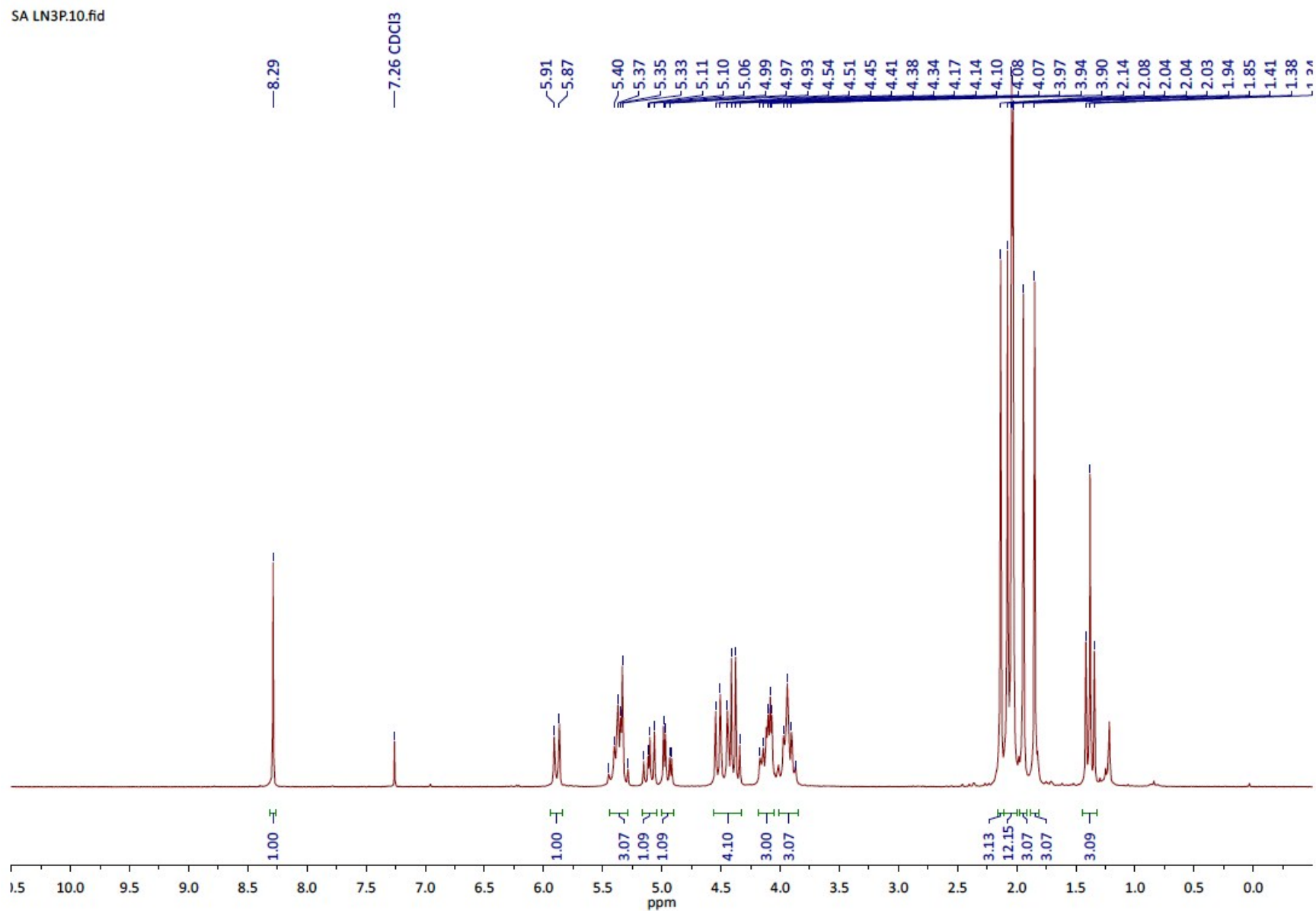


Figure S7. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **3e**.



ARM LN3P.10.fid

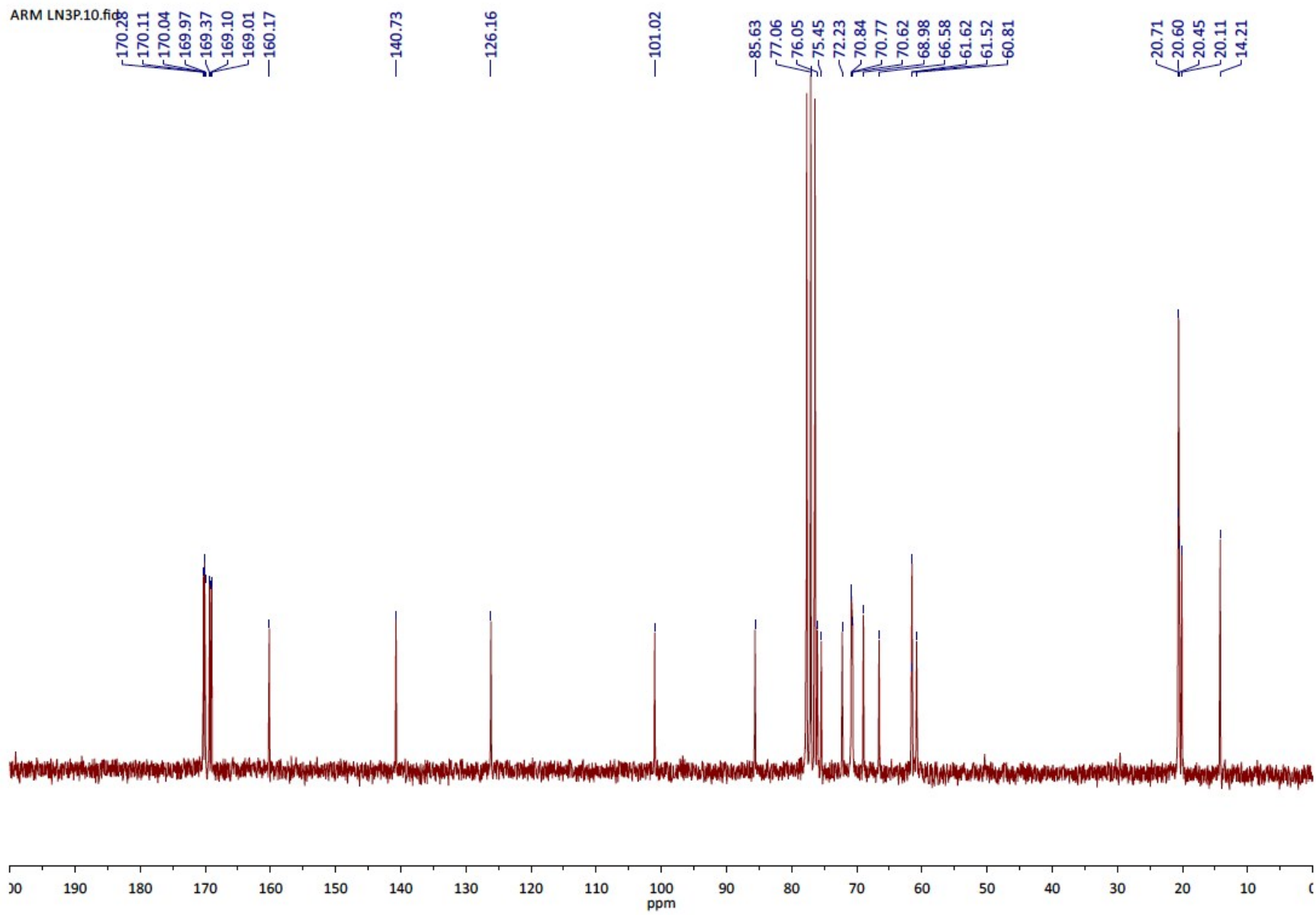
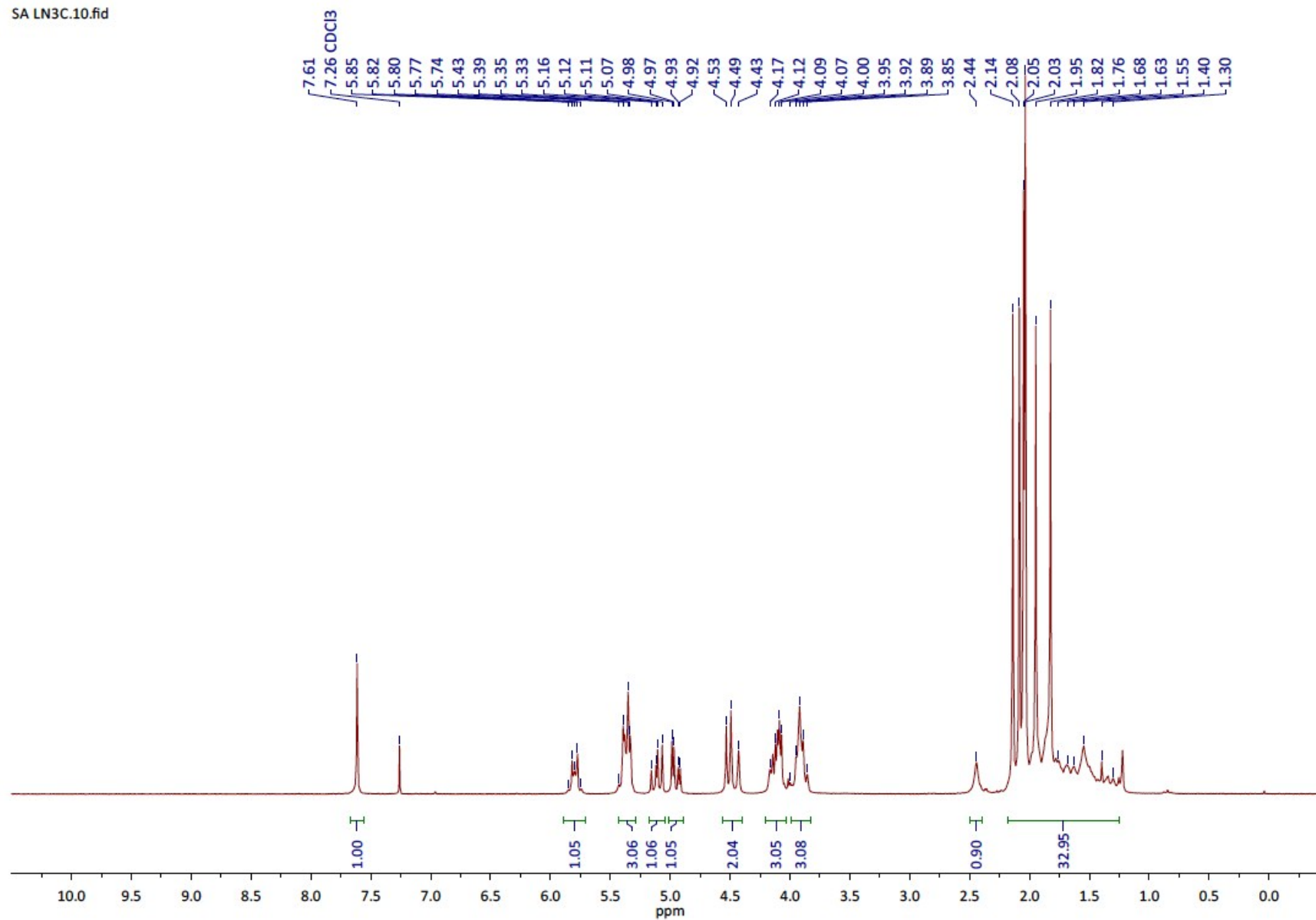


Figure S8. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **3f**.

SA LN3C.10.fid



SA LN3C.10.fid

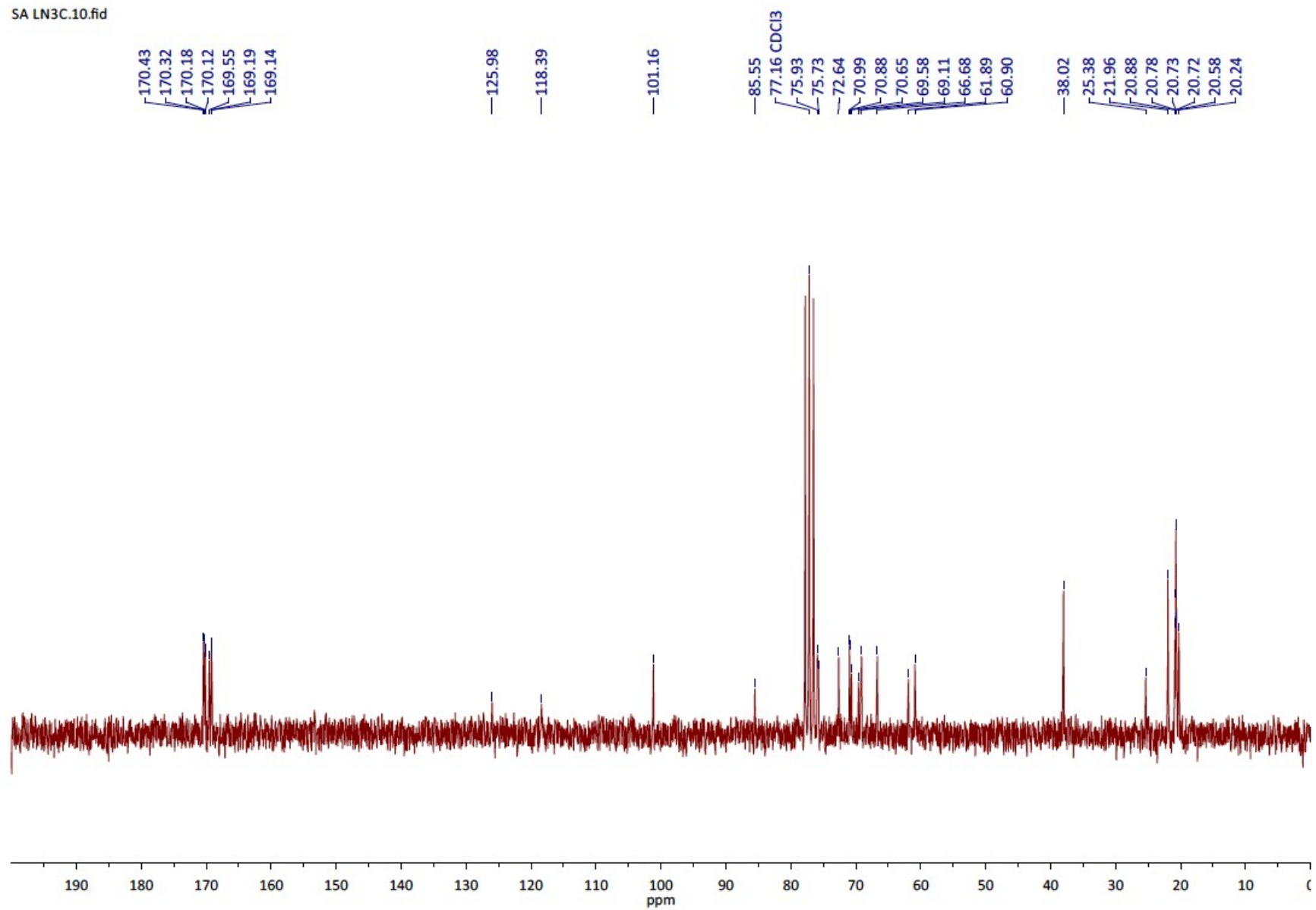
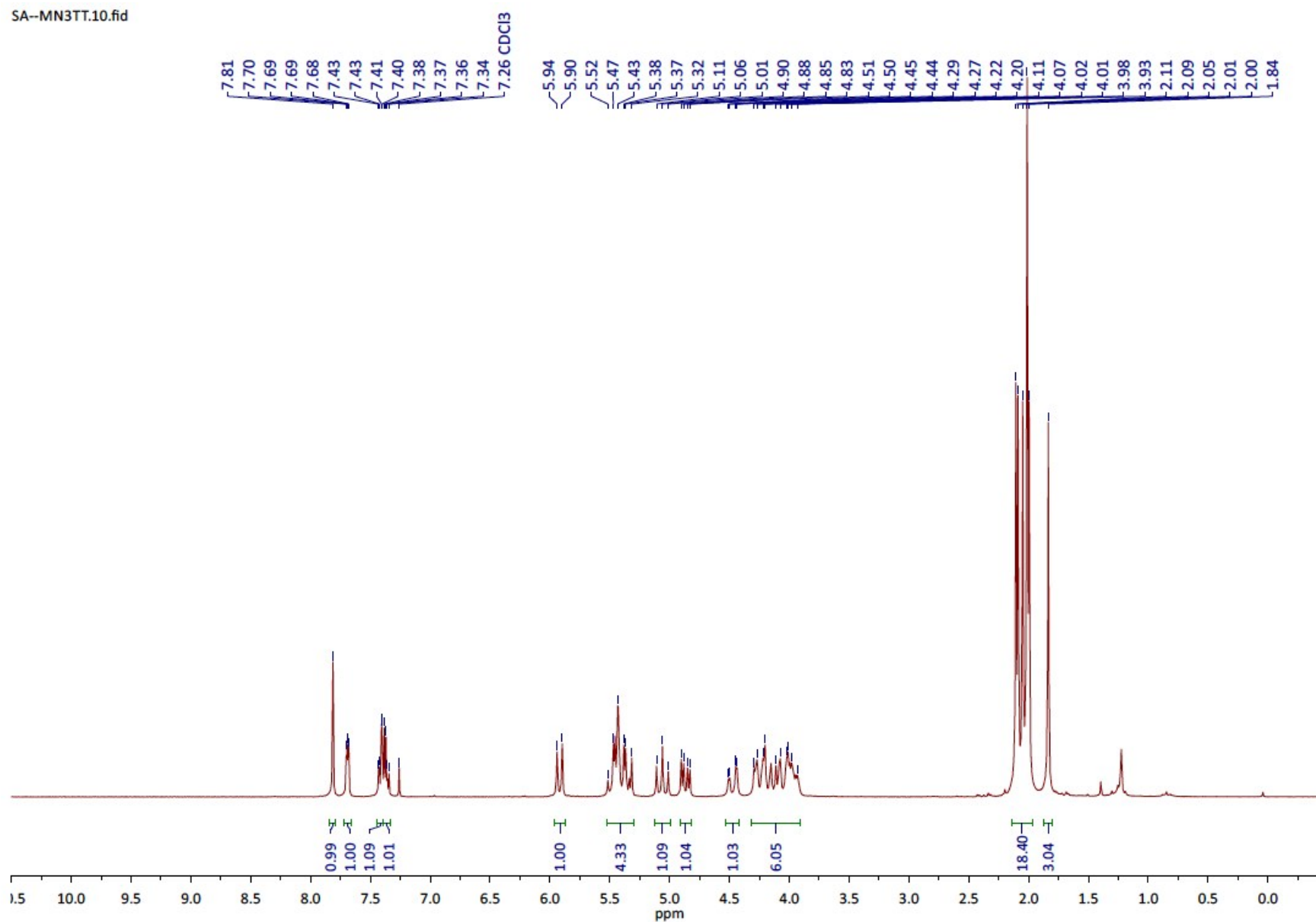


Figure S9. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **3g**.



SA MN3TT.10.fid

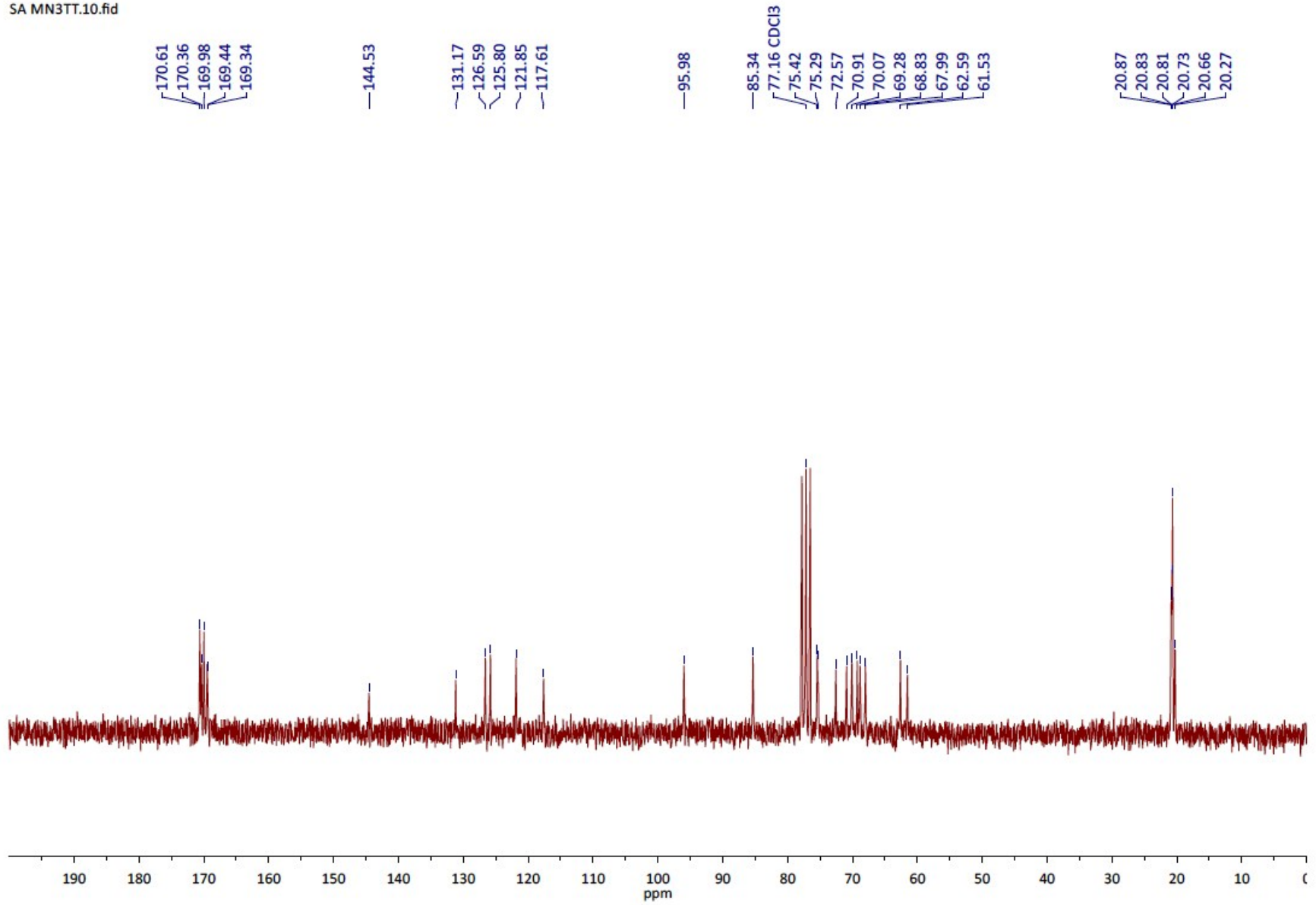
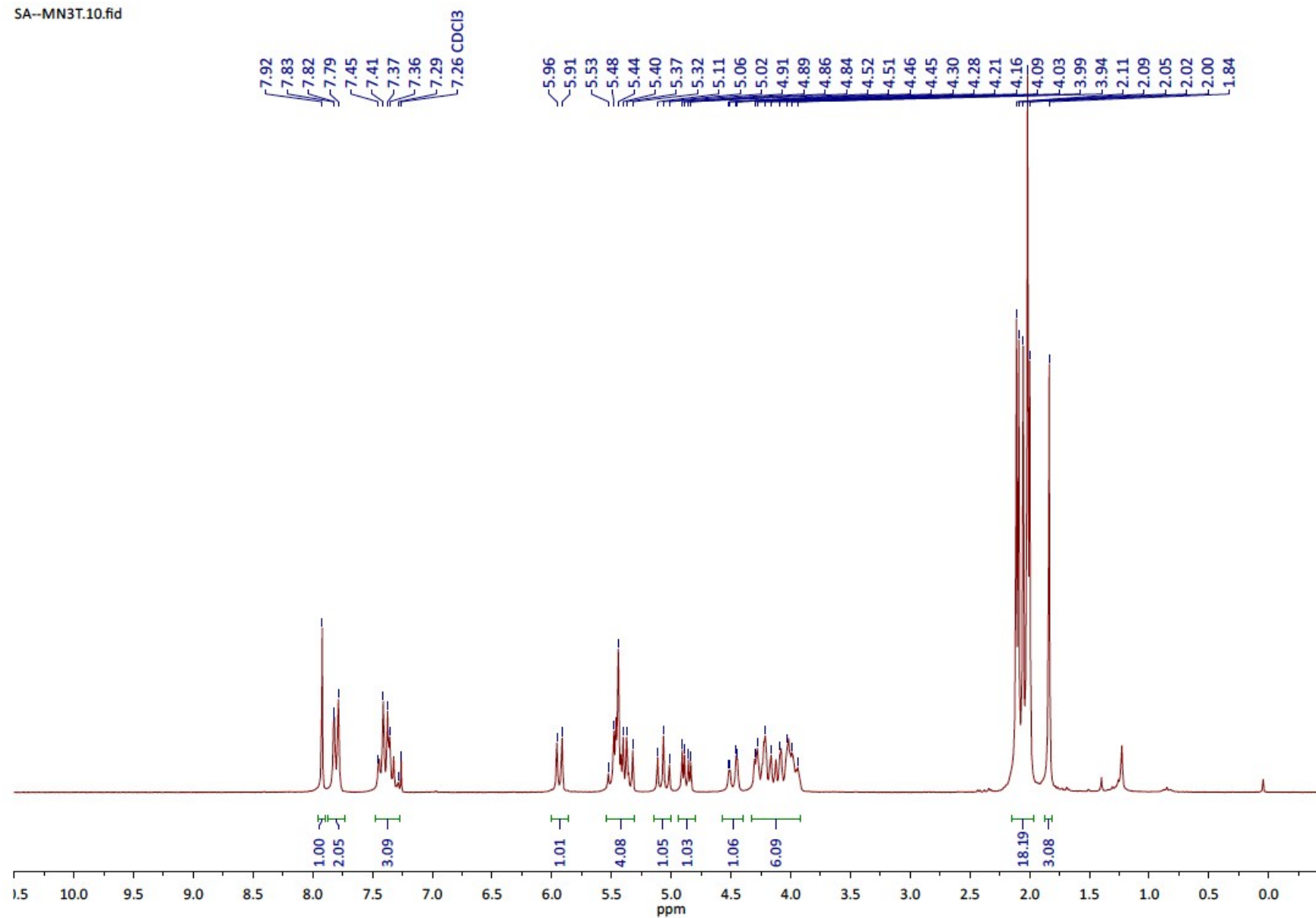


Figure S10. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **3h**.

SA--MN3T.10.fid



SA MN3T.10.fid

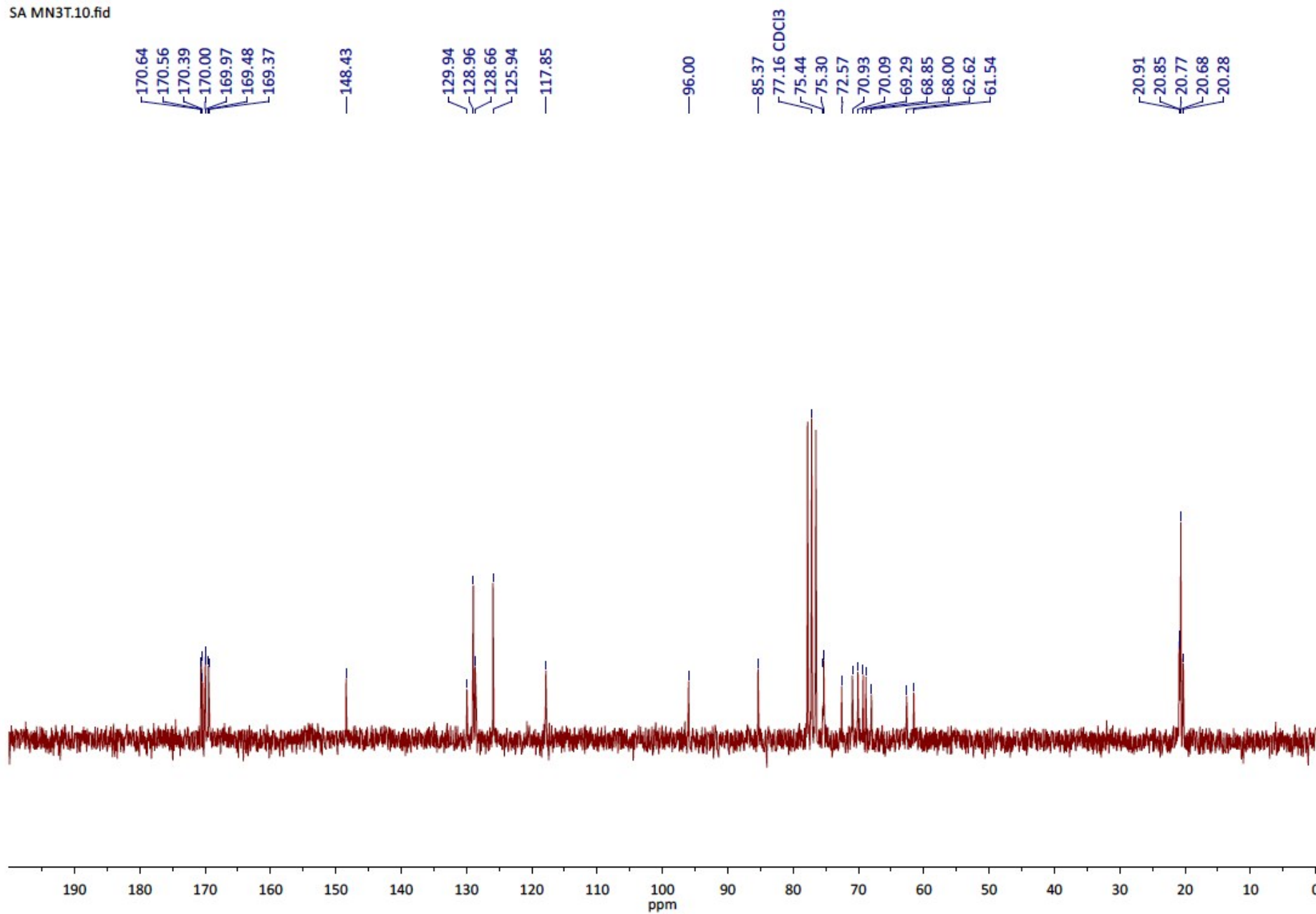
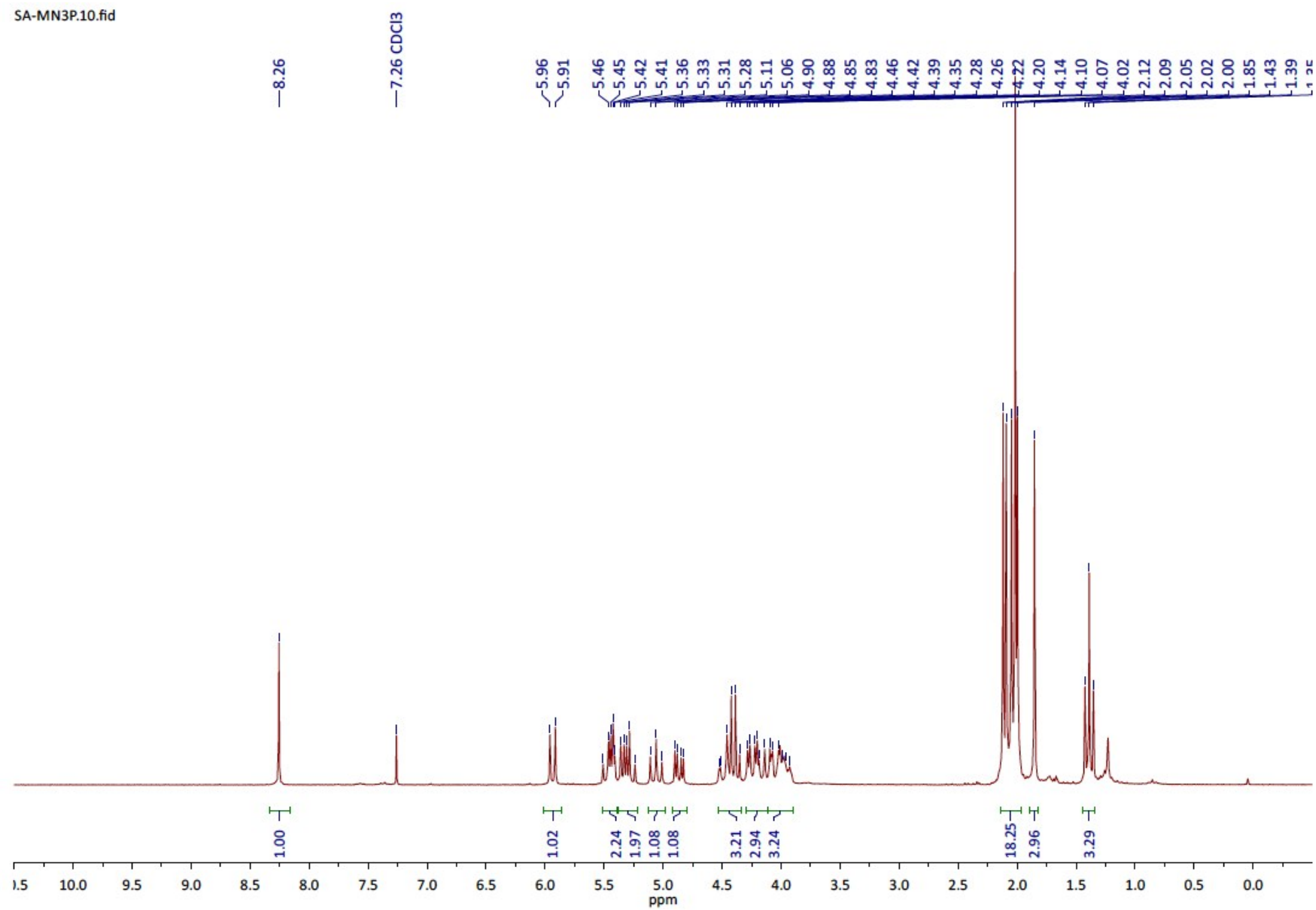


Figure S11. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **3i**.



SA14.20.fid
SA14 (DMSO)

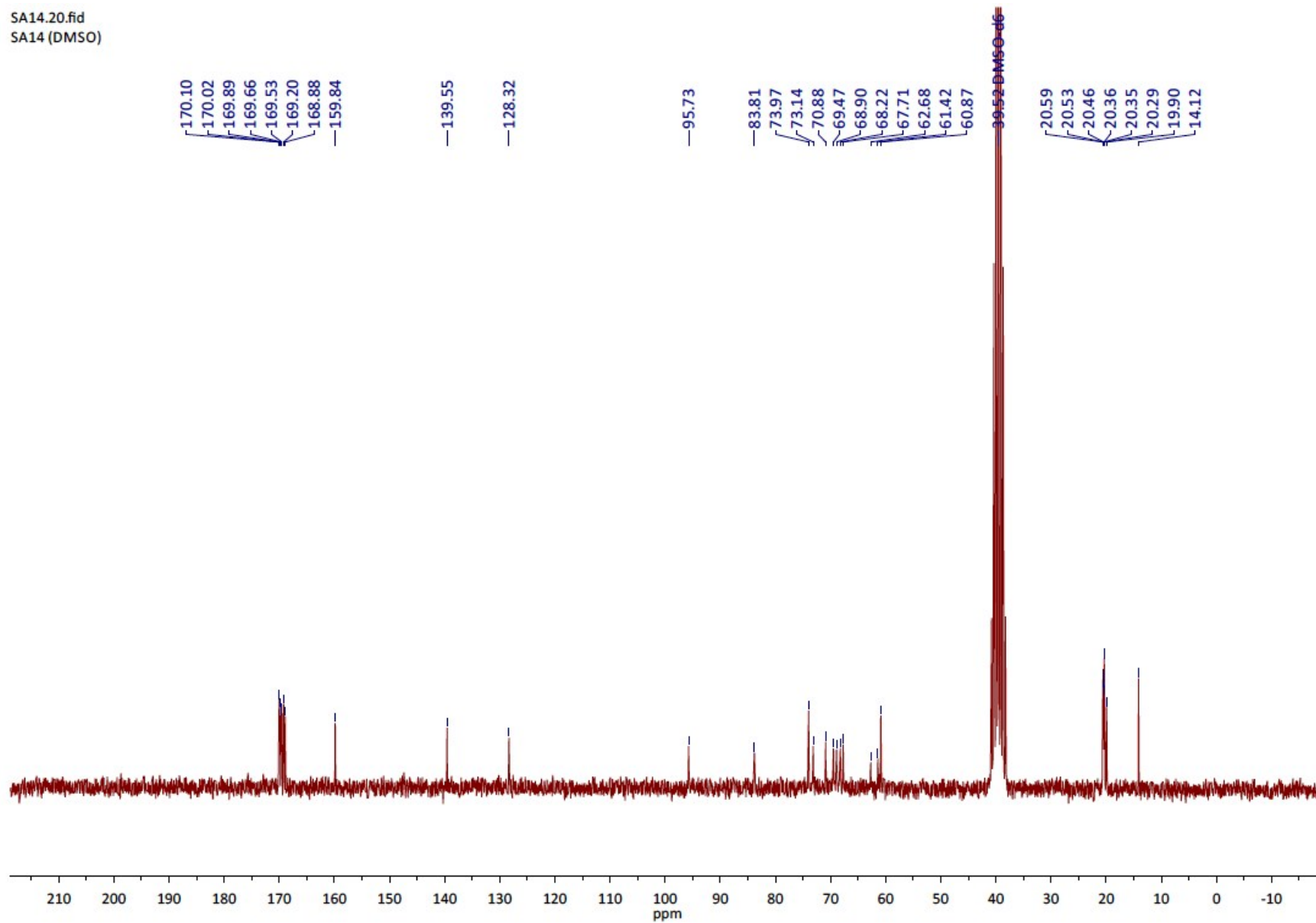
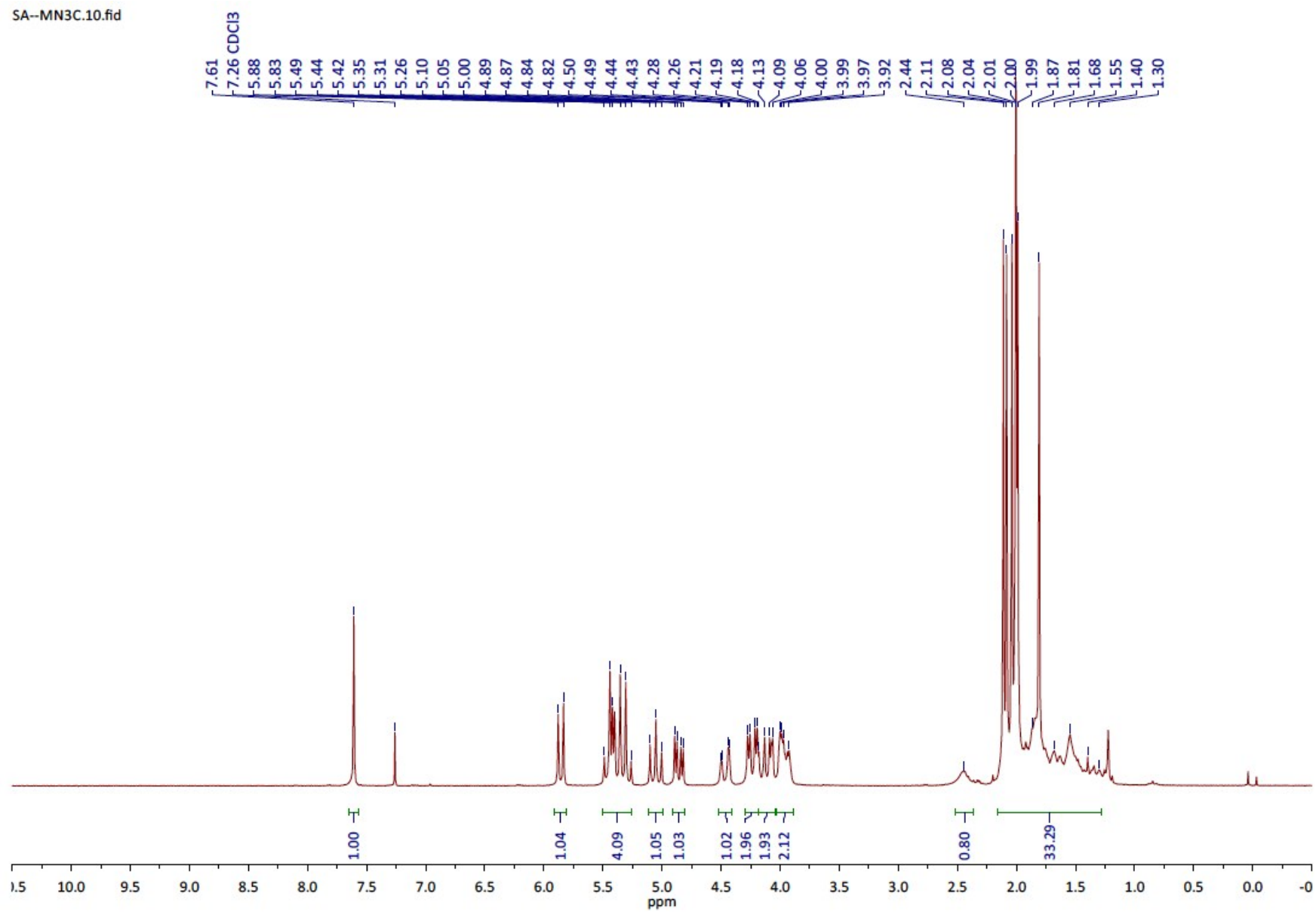


Figure S12. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **3j**.



SA15.20.fid
SA15 (DMSO)

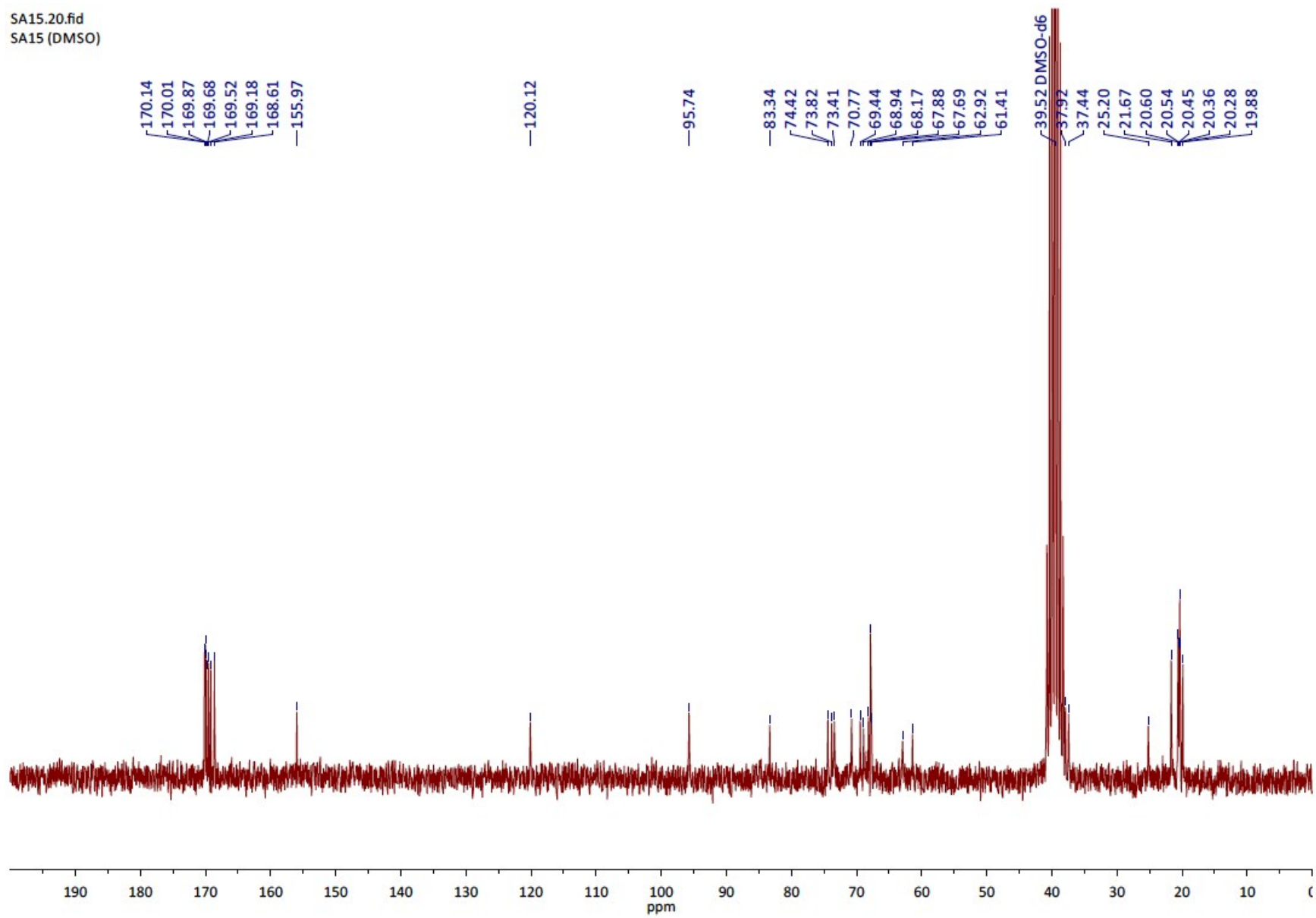
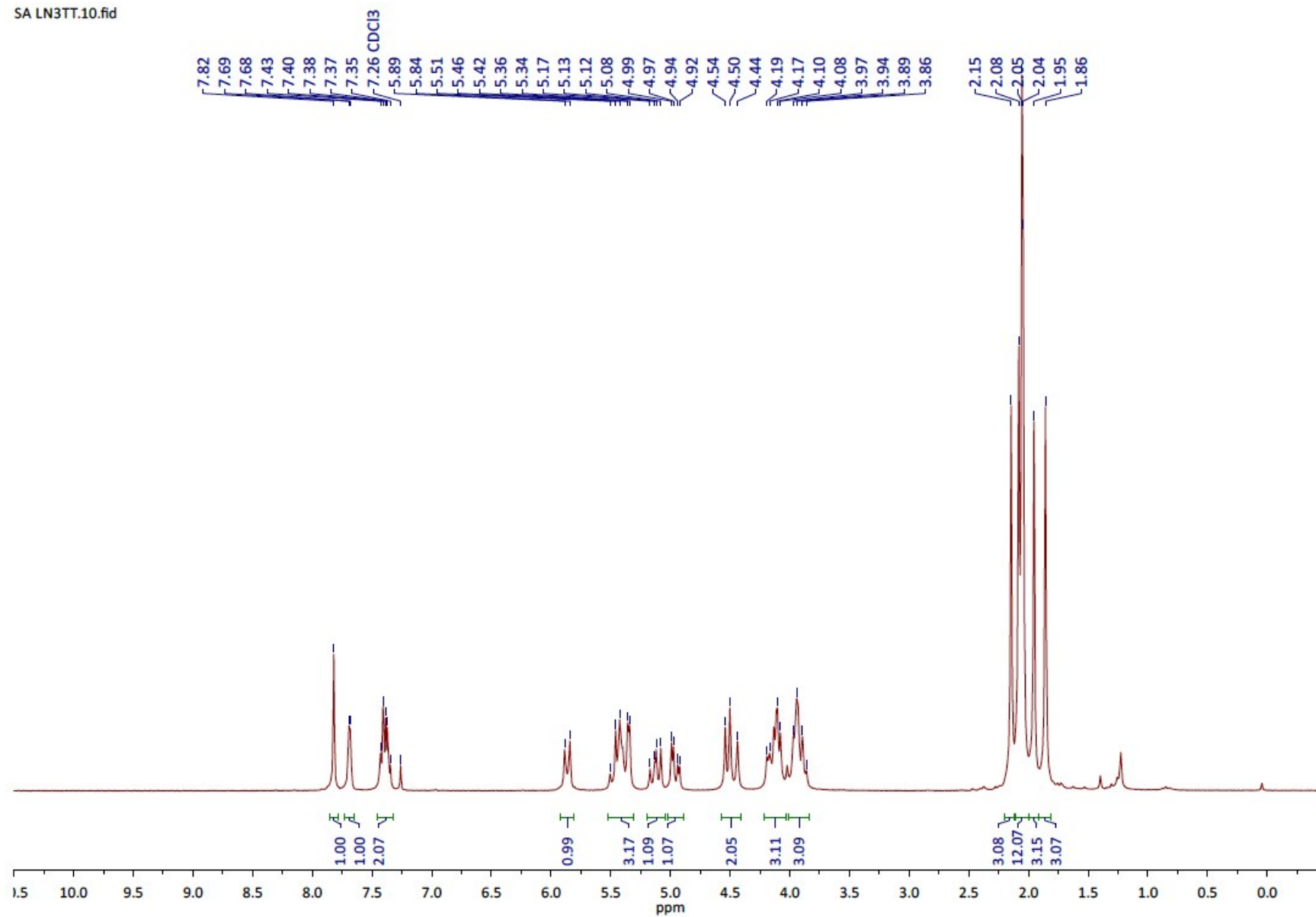
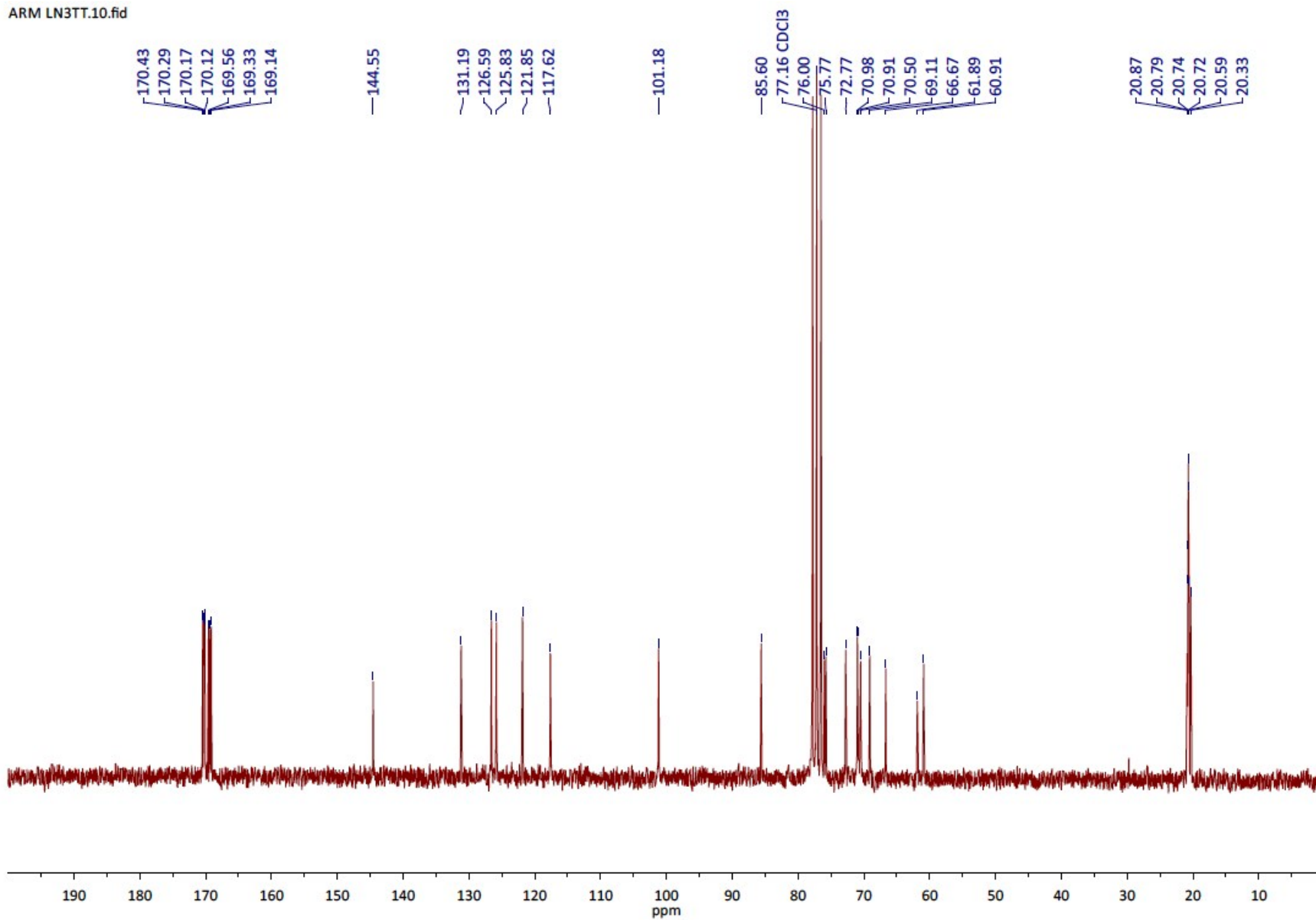


Figure S12. ^1H NMR (200 MHz) and ^{13}C NMR (50 MHz) and HRMS spectra for compound **3k**.

SA LN3TT.10.fid



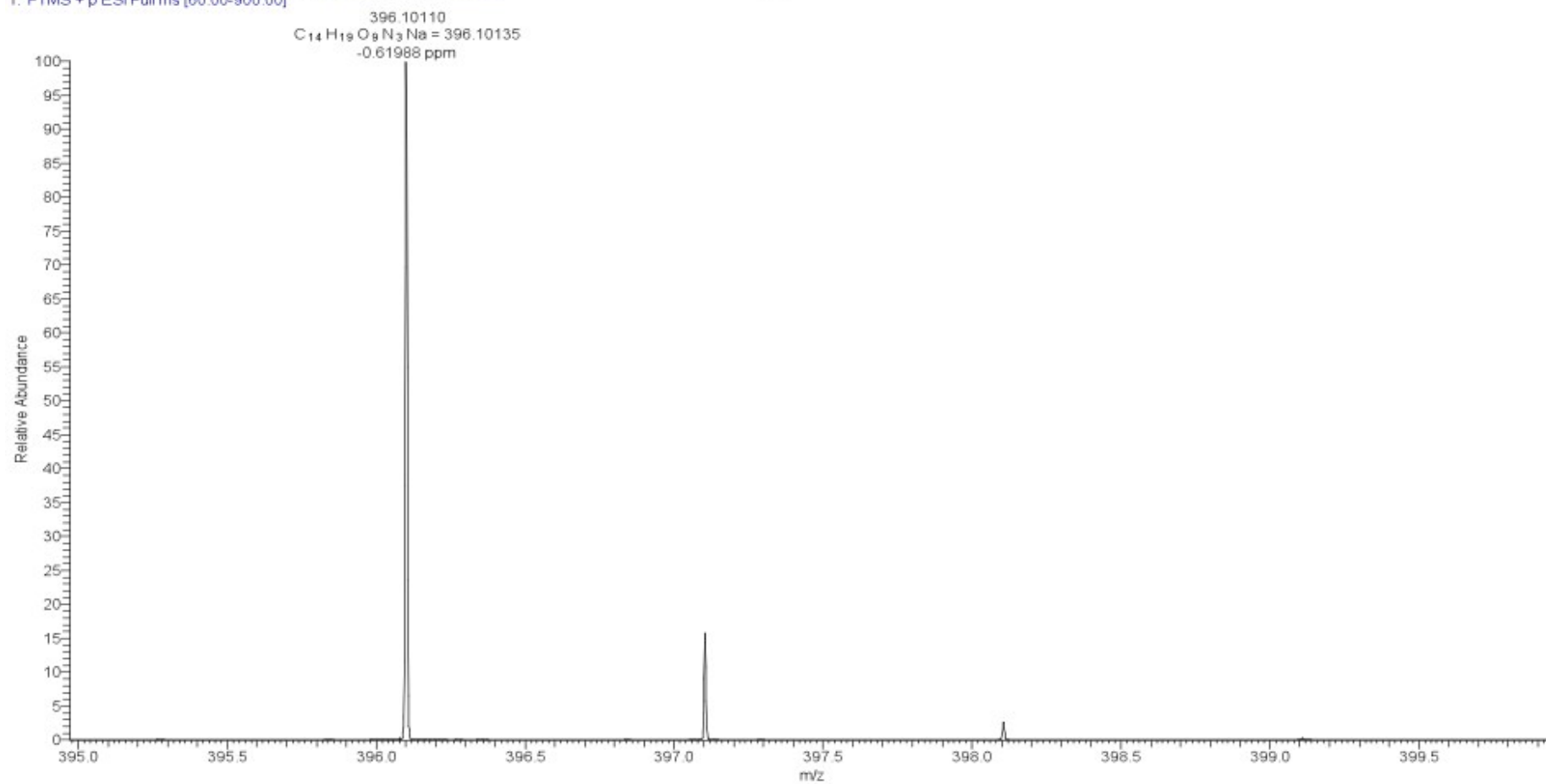
ARM LN3TT.10.fid



HRMS data

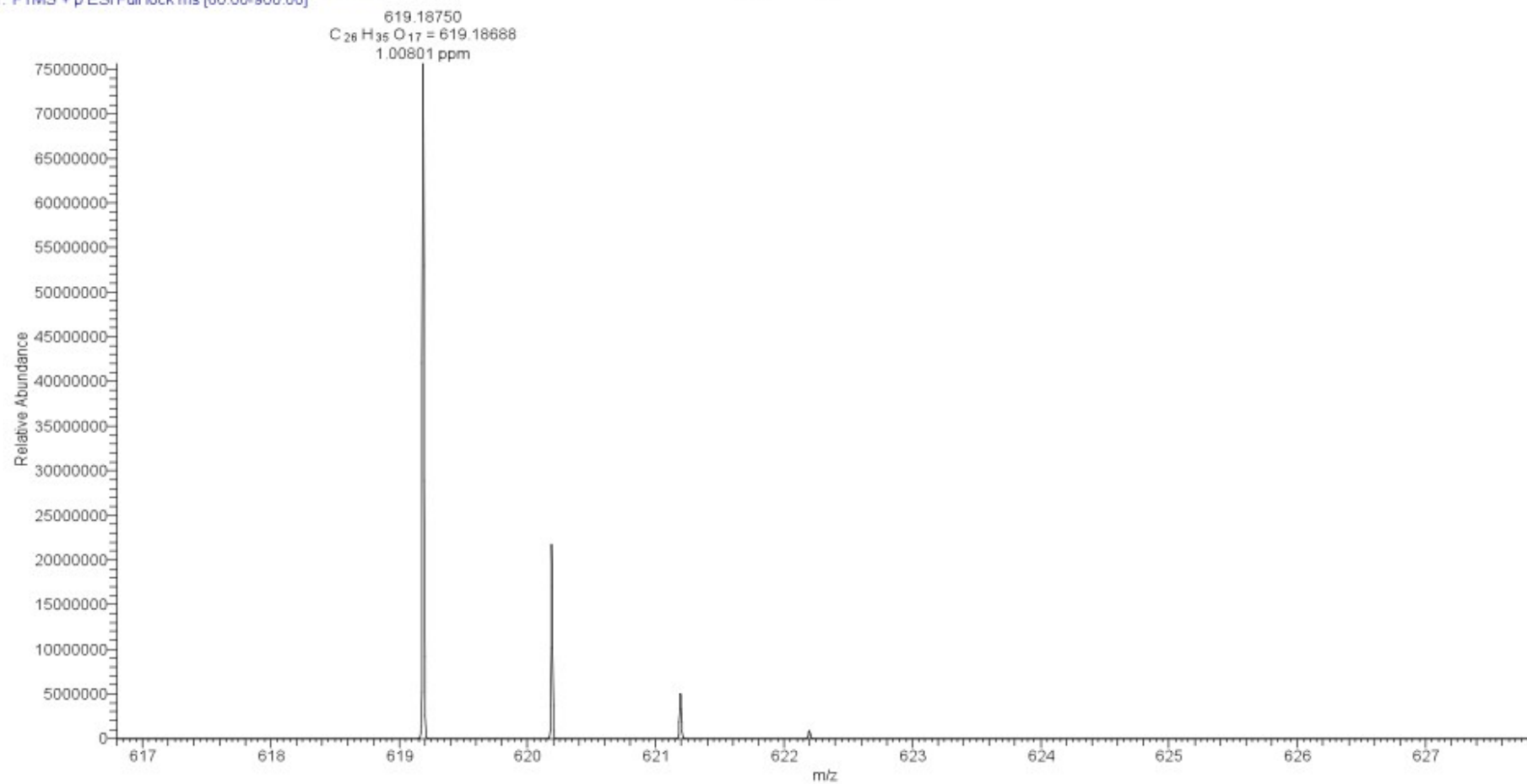
20140711ICN16_141107150257 #109 RT: 0.20 AV: 1 NL: 6.83E9
T: FTMS + p ESI Full ms [60.00-900.00]

2c



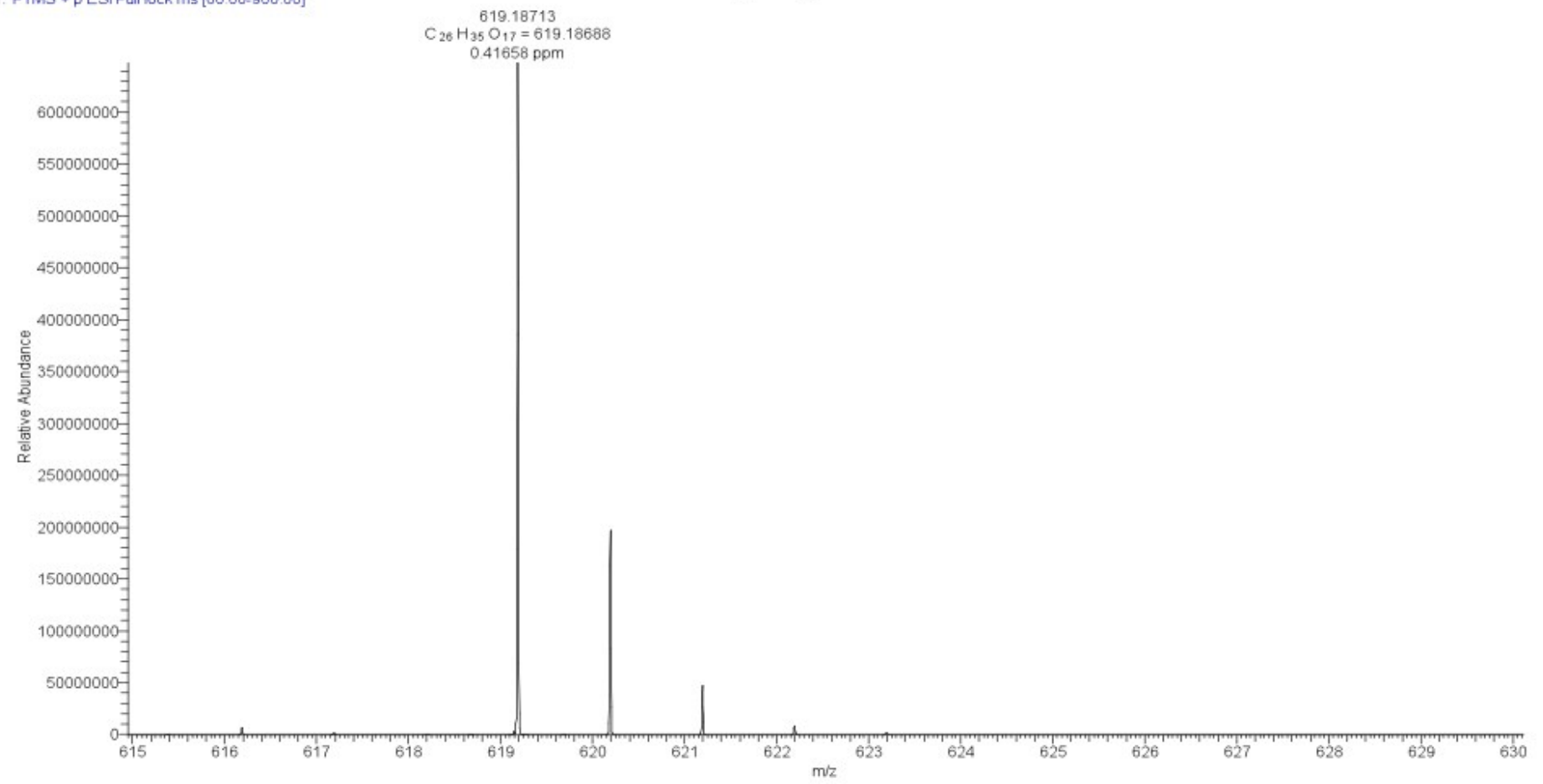
20140711ICN25 #211 RT: 0.40 AV: 1 NL: 7.56E7
T: FTMS + p ESI Full lock.ms [60.00-900.00]

2d (- N₃)



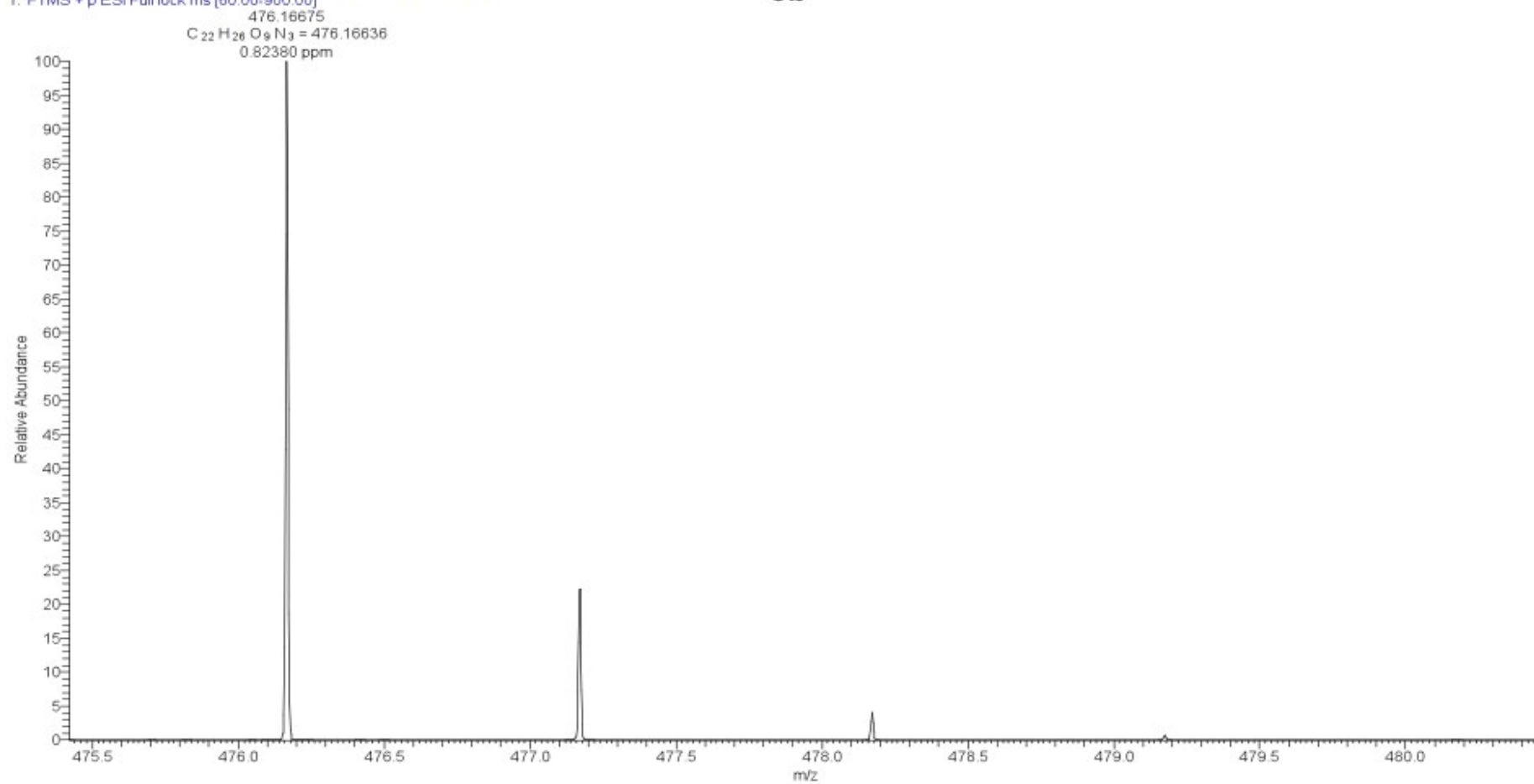
20140711ICN20 #169 RT: 0.32 AV: 1 NL: 6.47E8
T: FTMS + p ESI Full lock ms [60.00-900.00]

2e (- N₃)



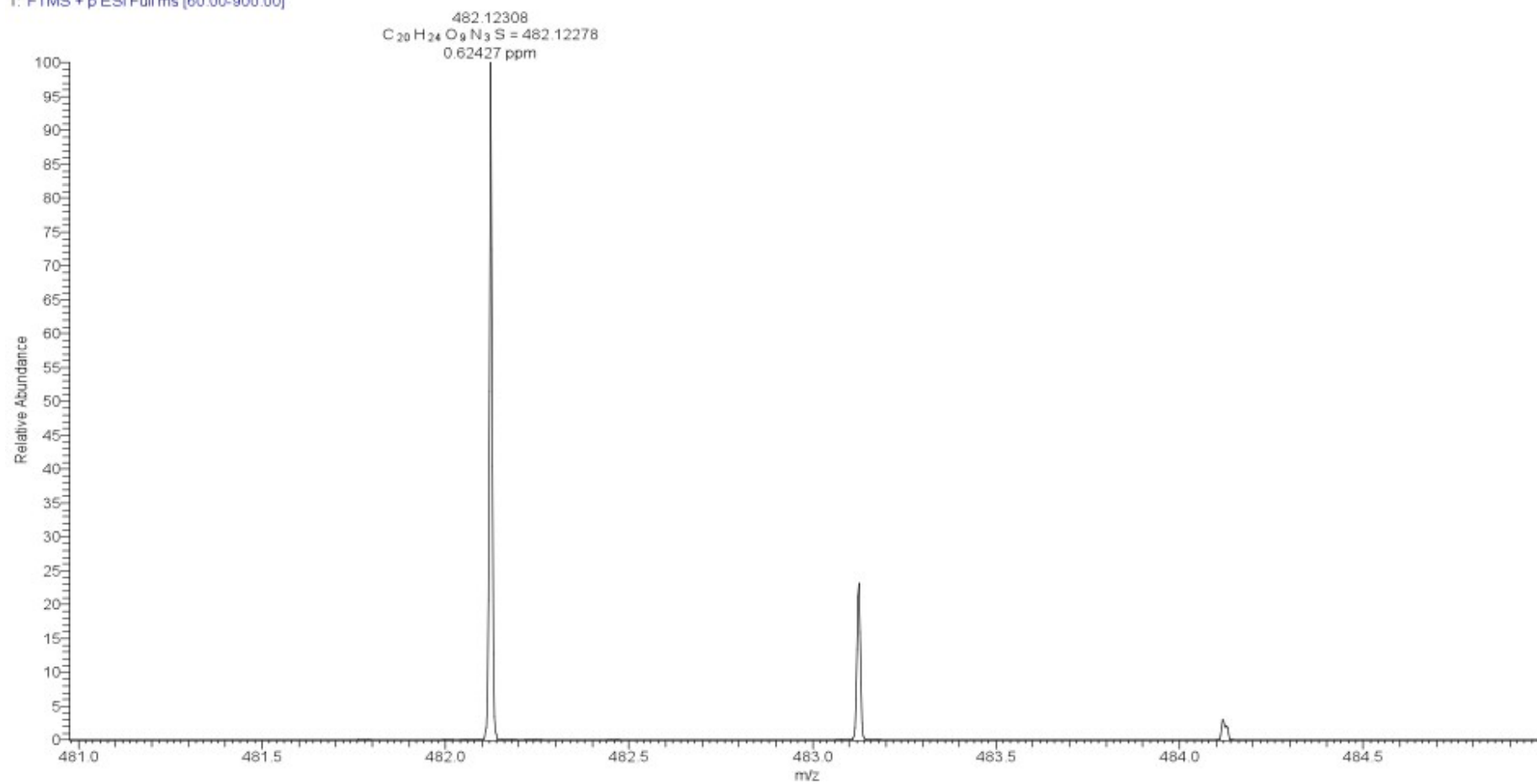
20140711CN17_141107150931 #151 RT: 0.28 AV: 1 NL: 9.54E8
T: FTMS + p ESI Full lock ms [60.00-900.00]

3b



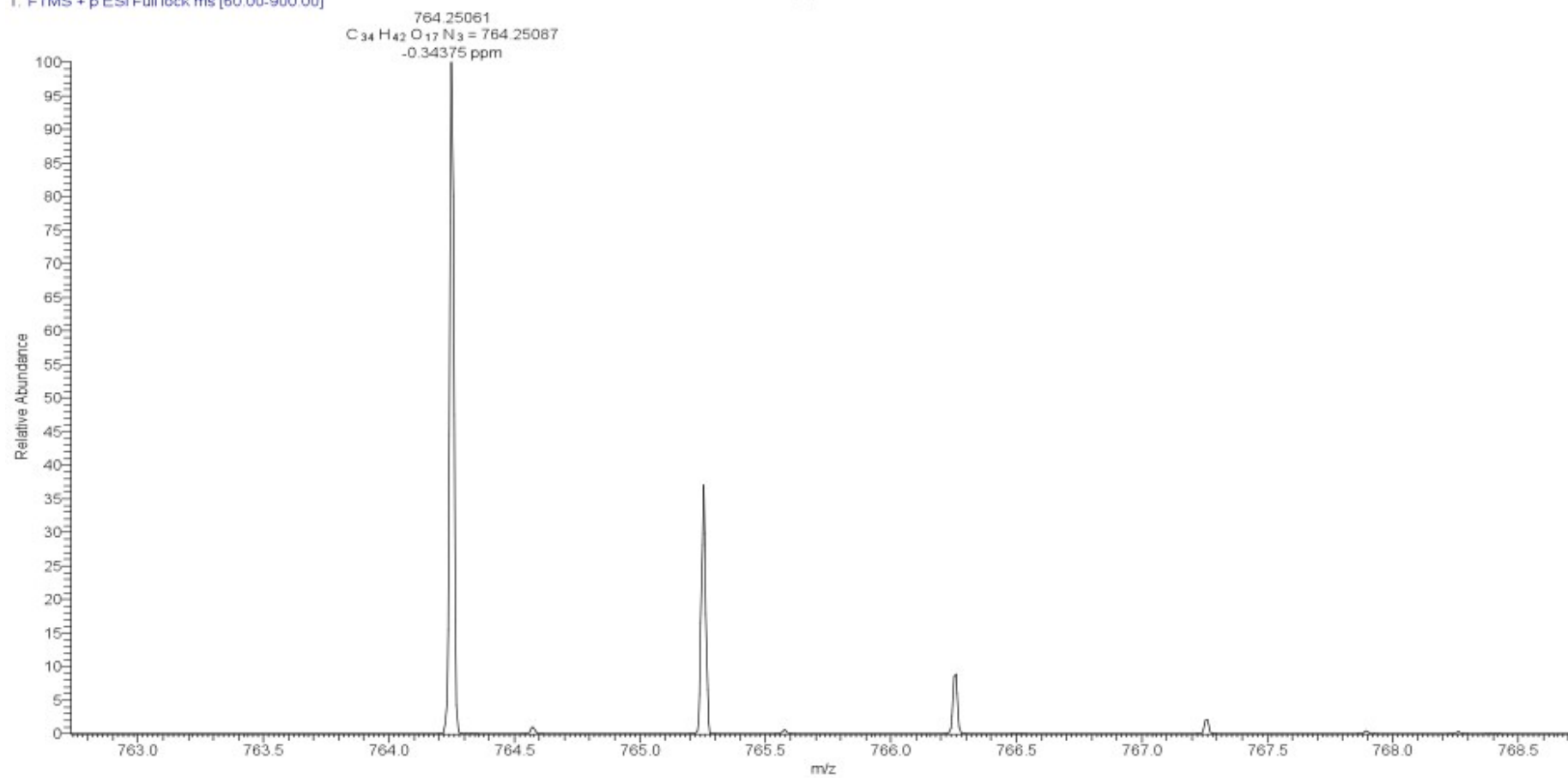
20140711ICN18_141107151605 #151 RT: 0.28 AV: 1 NL: 1.80E9
T: FTMS + p ESI Full ms (60.00-900.00)

3c



20140711CN26 #127 RT: 0.24 AV: 1 NL: 2.13E8
T: FTMS + p ESI Full lock ms [60.00-900.00]

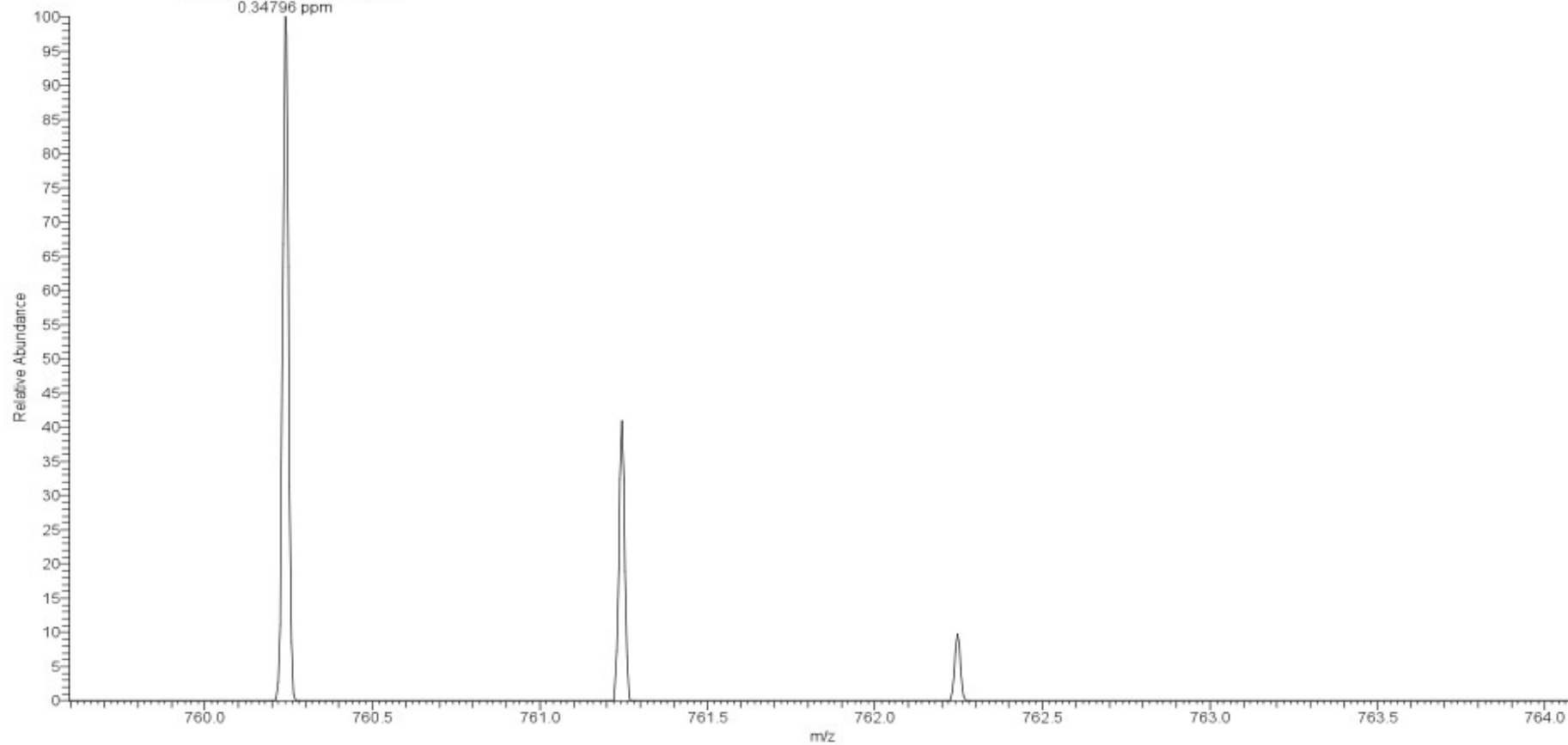
3d



20140711ICN28 #106 RT: 0.20 AV: 1 NL: 1.01E7
T: FTMS + p ESI Full lock ms [60.00-900.00]

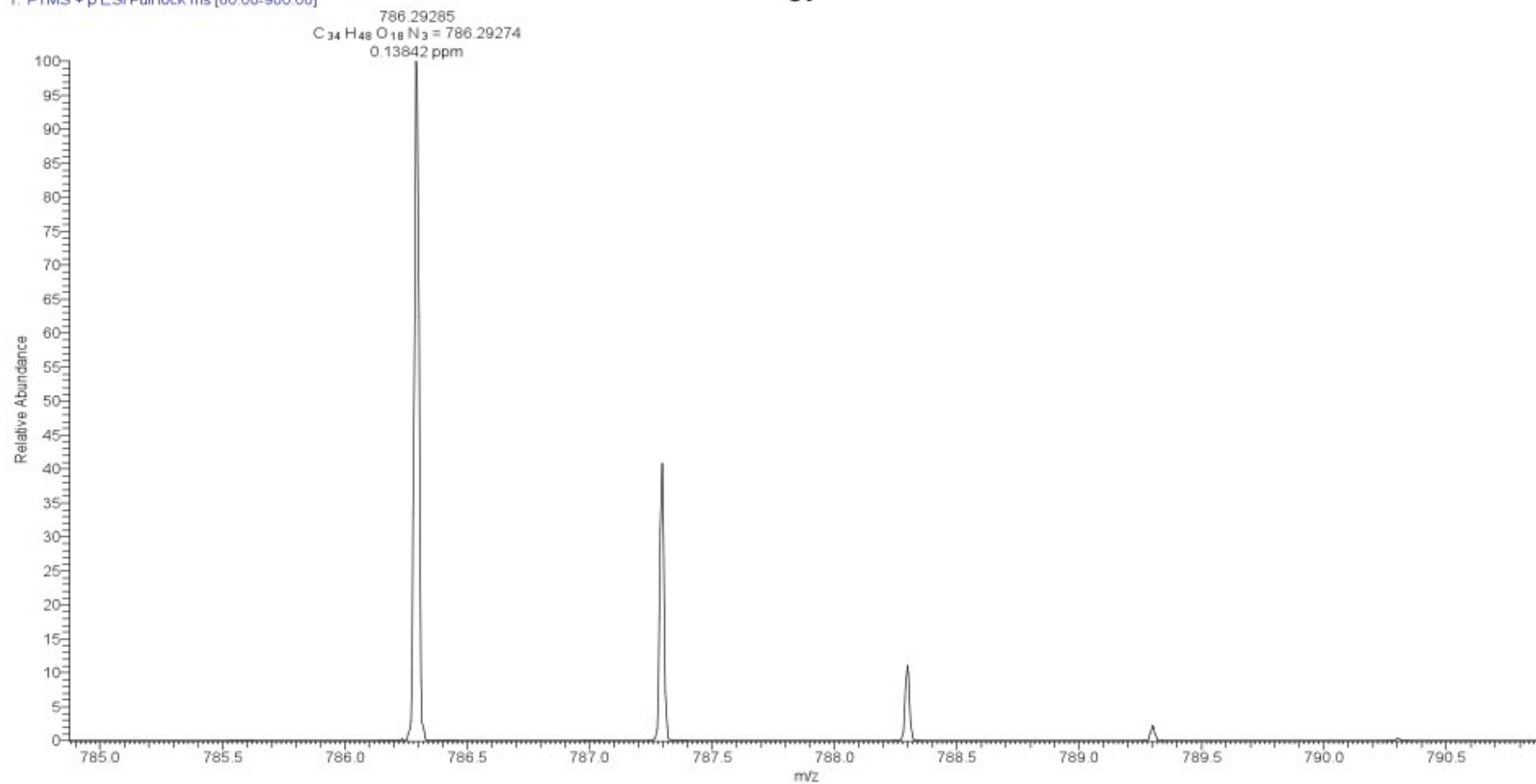
3e

760.24097
C₃₁H₄₂O₁₉N₃ = 760.24070
0.34796 ppm



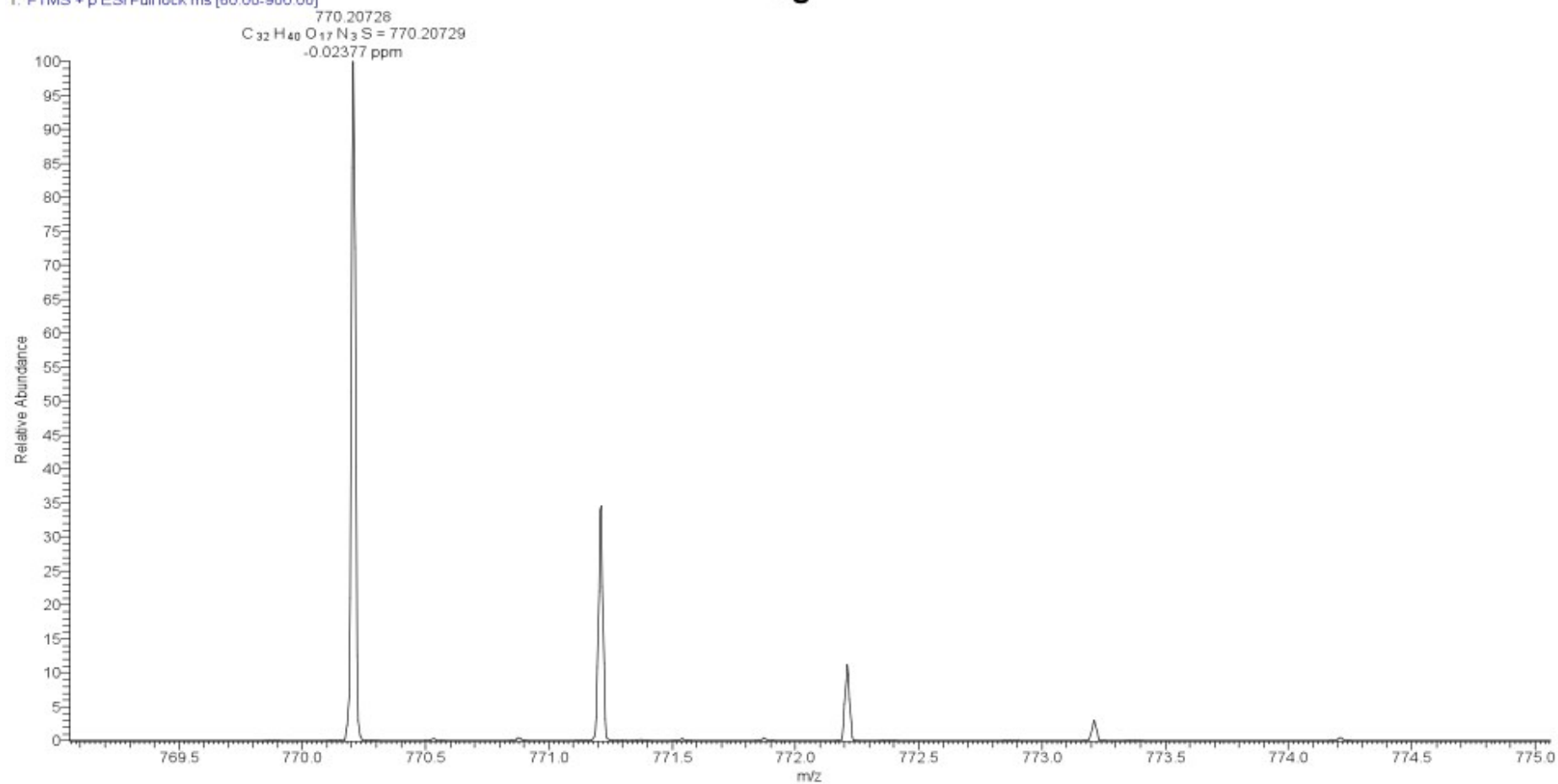
20140711ICN29 #145 RT: 0.27 AV: 1 NL: 7.46E8
T: FTMS + p ESI Full lock ms [60.00-900.00]

3f



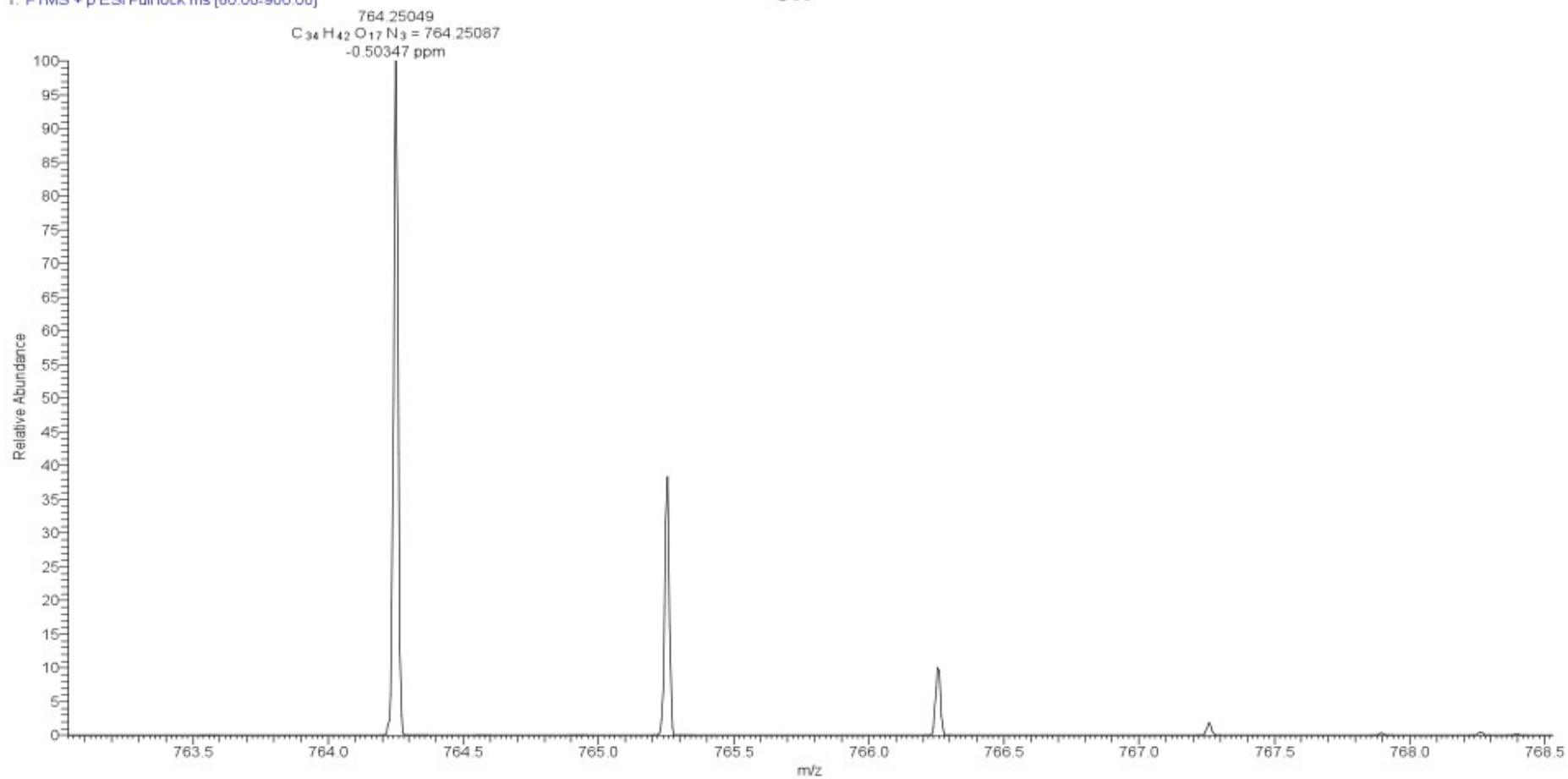
20140711CN22 #121 RT: 0.23 AV: 1 NL: 2.76E8
T: FTMS + p ESI Full lock ms [60.00-900.00]

3g



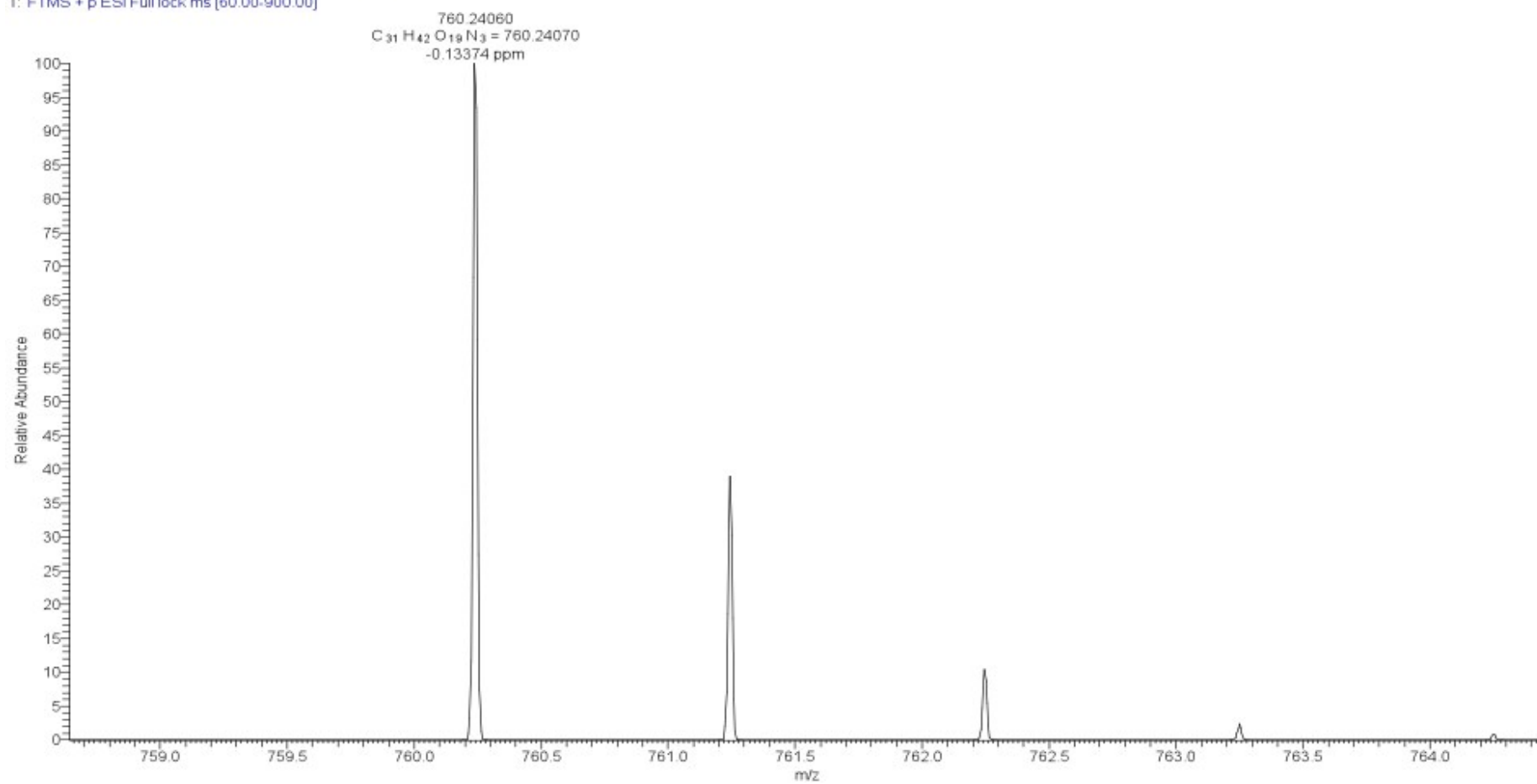
20140711CN21 #115 RT: 0.22 AV: 1 NL: 3.89E8
T: FTMS + p ESI Full lock ms [60.00-900.00]

3h



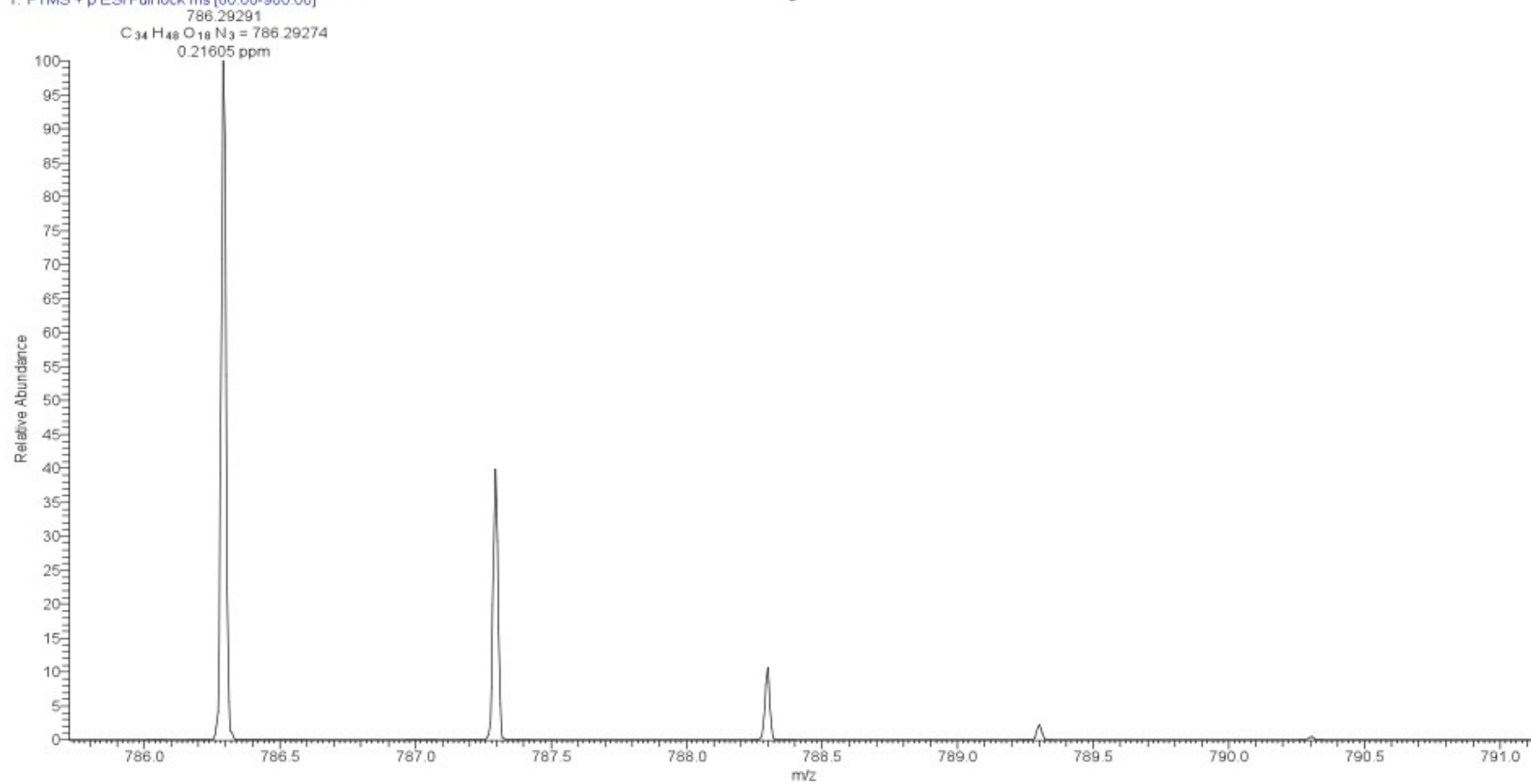
20140711CN23 #73 RT: 0.14 AV: 1 NL: 3.95E7
T: FTMS + p ESI Full lock ms [60.00-900.00]

3i



20140711ICN24 #91 RT: 0.17 AV: 1 NL: 1.05E9
T: FTMS + p ESI Full lock ms [60.00-900.00]

3j



20140711CN27 #79 RT: 0.15 AV: 1 NL: 2.74E8
T: FTMS + p ESI Full lock ms [60.00-900.00]

3k

