### **Supplementary Information**

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#### 1. Experimental

#### **1.1. General Experimental**

All anhydrous solvents and reagents were obtained from commercial suppliers (Aldrich, Alfa Aesar, Fluorochem) and used without further purification.

Reactions were performed in oven-dried, round-bottom flasks fitted with rubber septa under an atmosphere of argon unless otherwise stated. Analytical thin layer chromatography (TLC) was performed on pre-coated aluminium sheets of silica (60  $F_{254}$ , Merck) and visualised by short-wave UV light.

<sup>1</sup>H NMR spectra were recorded at 500 MHz on a Bruker Avance-500 using an internal deuterium lock. Chemical shifts were measured in parts per million (ppm) relative to tetramethylsilane ( $\delta = 0$ ) using the following internal references for residual protons in the solvent : CDCl<sub>3</sub> ( $\delta$  7.26) and (CD<sub>3</sub>)<sub>2</sub>SO ( $\delta$  2.50). Data is presented as follows: chemical shift, multiplicity, coupling constant (*J*) in Hz, integration and assignment.

<sup>13</sup>C NMR specta were recorded at 126 MHz on a Bruker Avance-500 using an internal deuterium lock. Chemical shifts were measured in parts per million (ppm) relative to tetramethylsilane ( $\delta = 0$ ) using the following internal references for residual protons in the solvent: CDCl<sub>3</sub> ( $\delta$  77.0) and (CD<sub>3</sub>)<sub>2</sub>SO ( $\delta$  39.5). Data is presented as follows: chemical shift and assignment.

HRMS analysis was performed on an Agilent 1200 series HPLC and diode array detector coupled to a 6520 Quadrupole-Time of flight mass spectrometer with dual multimode APCI/ESI source. Analytical separation was carried out at 30 °C in a Merck Purospher STAR column (RP-18e, 30 x 4 mm) using a flow rate of 1.5 mL/min in a 4 min gradient elution with detection at 254 nm. The mobile phase was a mixture of methanol (solvent A) and water (solvent B) both containing formic acid at 0.1%. Gradient elution was as follows: 1:9 (A/B) to 9:1 (A/B) over 2.5 min, 9:1 A/B for 1 min, and then reversion back to 1:9 (A/B) over 0.3 min, finally 1:9 (A/B) for 0.2 min. The following reference masses were used for HRMS analysis: caffeine [M+H]<sup>+</sup> 195.087652; (hexakis(1H,1H,3H-tetrafluoropentoxy)phosphazene [M+H]<sup>+</sup> 622.02896 or reserpine [M+H]<sup>+</sup> 609.280657.

#### **1.2. Experimental Procedures**

Synthesis of 2-(2-formyl-4-nitrophenoxy)acetonitrile (3)<sup>1</sup>



To a solution of 2-hydroxy-5-nitrobenzaldehyde (0.50 g, 3.0 mmol) in DMF (2.5 mL) was added  $K_2CO_3$  (0.60 g, 4.4 mmol). The resulting suspension was stirred for 15 min before the addition of bromoacetonitrile (0.40 mL, 5.8 mmol). The reaction was allowed to stir at room temperature for a period of 18 h. The reaction mixture was diluted with ethyl acetate and washed with water and brine. The organic layer was separated, dried over magnesium sulphate, and concentrated under vacuum. The crude material was purified by column chromatography (50% ethyl acetate in cyclohexane) to afford **3** (0.50 mg, 82%) as a pale yellow solid.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  10.46 (s, 1H, HC=O), 8.79 (d, *J* = 2.9 Hz, 1H, H<sup>3</sup>), 8.55 (dd, *J* = 9.1, 2.9 Hz, 1H, H<sup>5</sup>), 7.25 (d, *J* = 9.1 Hz, 1H, H<sup>6</sup>), 5.08 (s, 2H, OCH<sub>2</sub>CN); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  186.1 (C<sub>q</sub>), 161.5 (C<sub>q</sub>) 130.6 (CH), 125.6 (CH), 125.3 (C<sub>q</sub>), 113.2 (C<sub>q</sub>), 112.8 (CH), 54.0 (CH<sub>2</sub>). N.B. Mass Spec. data could not be obtained. N.B. One C<sub>q</sub> could not be resolved.

### Synthesis of 2-phenyl-6,7-dihydro-5*H*-pyrrolo[2,1-*c*][1,2,4]triazol-2-ium tetrafluoroborate (4)<sup>2</sup>



Trimethyloxonium tetrafluoroborate (0.96 g, 6.4 mmol) was added in one portion to a solution of pyrrolidin-2-one (0.50 g, 6.0 mmol) in DCM (20 mL). The reaction was allowed to stir at room temperature for 24 h. Phenylhydrazine (0.63 g, 5.8 mmol) was added and stirring was continued at room temperature for a further 48 h. The reaction mixture was then concentrated under vacuum and re-dissolved in a mixture of methanol (2.0 mL) and triethyl orthoformate (8.0 ml). The solution was heated to reflux overnight and then allowed to cool to room temperature. The resulting precipitate was collected by filtration and washed with methanol to afford **4** (1.1 g, 70%) as a pale tan solid.

<sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.70 (s, 1H, NCHNPh), 7.90 – 7.84 (m, 2H, ArCH), 7.72 – 7.66 (m, 2H, ArCH), 7.66 – 7.60 (m, 1H, ArCH), 4.66 – 4.13 (m, 2H, NCH<sub>2</sub>), 3.23 – 3.12 (m, 2H, CH<sub>2</sub>), 2.89 – 2.61 (m, 2H, CH<sub>2</sub>); <sup>13</sup>C NMR (126 MHz, d<sub>6</sub> DMSO)  $\delta$  163.5 (C<sub>q</sub>), 138.8 (CH), 136.1 (C<sub>q</sub>), 130.9 (CH), 130.7 (CH), 121.1 (CH), 47.4 (CH<sub>2</sub>), 27.1 (CH<sub>2</sub>), 21.7 (CH<sub>2</sub>). NB. Mass spec. data could not be obtained.

Synthesis of 3-amino-6-nitro-4H-chromen-4-one (5)<sup>1</sup>



Compound **3** (0.47 g, 2.3 mmol) was dissolved in DCM (20 mL) before addition of catalyst  $4^2$  (62 mg, 0.20 mmol) and DBU (30  $\mu$ L, 0.20 mmol). The resulting solution was allowed to stir at room temperature for 16 h. The reaction was then concentrated under vacuum. The crude material was purified by column chromatography (40-60% ethyl acetate in cyclohexane) to afford **5** (0.27 g, 57%) as an orange solid.

<sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>)  $\delta$  8.82 (d, *J* = 2.9 Hz, 1H, H<sup>5</sup>), 8.46 (dd, *J* = 9.3, 2.9 Hz, 1H, H<sup>7</sup>), 8.06 (s, 1H, H<sup>2</sup>), 7.84 (d, *J* = 9.3 Hz, 1H, H<sup>8</sup>), 4.85 (s, 2H, NH<sub>2</sub>); <sup>13</sup>C NMR (126 MHz, DMSO-d<sub>6</sub>)  $\delta$  171.8 (C<sub>q</sub>), 158.4 (C<sub>q</sub>), 143.8 (C<sub>q</sub>), 137.3 (CH), 133.7 (C<sub>q</sub>), 127.0 (CH), 121.9 (CH), 121.3 (C<sub>q</sub>), 121.1 (CH); HRMS (ESI) [M+H]<sup>+</sup> 207.0403, C<sub>9</sub>H<sub>6</sub>N<sub>2</sub>O<sub>4</sub> [M+H]<sup>+</sup> requires 207.0406.

Synthesis of N-(6-nitro-4-oxo-4H-chromen-3-yl)butyramide (6)



Compound **5** (0.17 g, 0.83 mmol) and DIPEA (0.22 mL, 1.2 mmol) were dissolved in THF (10 mL) and cooled to 0 °C before the addition of butyryl chloride (0.10 mL, 0.90 mmol). The reaction was allowed to warm to room temperature and stirred for 1 h. The reaction mixture was then diluted with ethyl acetate and washed with sat. NaHCO<sub>3</sub> (aq.) and brine. The organic layer was separated, dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (50% ethyl acetate in cyclohexane) to afford **6** (0.18 g, 80%) as an orange solid.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.52 (s, 1H, H<sup>2</sup>), 9.17 (d, *J* = 2.7 Hz, 1H, H<sup>5</sup>), 8.54 (dd, *J* = 9.2, 2.7 Hz, 1H, H<sup>7</sup>), 8.01 (s, 1H, NH), 7.69 (d, *J* = 9.2 Hz, 1H, H<sup>8</sup>), 2.56 – 2.41 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.88 – 1.72 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.04 (t, *J* = 7.4 Hz, 3H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  171.9 (C<sub>q</sub>), 170.9 (C<sub>q</sub>), 158.2 (C<sub>q</sub>), 145.0 (CH), 128.0 (CH), 125.1 (C<sub>q</sub>) 122.8 (CH), 121.7 (C<sub>q</sub>), 120.3 (CH), 39.0 (CH<sub>2</sub>), 18.8 (CH<sub>2</sub>), 13.6 (CH<sub>3</sub>); HRMS (ESI) [M+H]<sup>+</sup> 277.0818 C<sub>13</sub>H<sub>12</sub>N<sub>2</sub>O<sub>3</sub> [M+H]<sup>+</sup> requires 277.0819; N.B. One C<sub>q</sub> not resolved

Synthesis of N-(6-amino-4-oxo-4H-chromen-3-yl)butyramide (7)



Compound **6** (0.18 g, 0.70 mmol), iron powder (0.26 g, 4.6 mmol) and ammonium chloride (0.25 g, 4.6 mmol) were heated to reflux in a mixture of ethanol and water

(3:1) (20 mL) for 16 h. The reaction mixture was diluted with ethyl acetate and filtered through celite and then washed with sat. NaHCO<sub>3</sub> (aq.) and brine. The organic layer was separated, dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (35-60% ethyl acetate in DCM) to afford **7** (0.12 g, 76%) as a yellow solid.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.38 (s, 1H, H<sup>2</sup>), 8.05 (s, 1H, NH), 7.41 (d, *J* = 2.9 Hz, 1H, H<sup>5</sup>), 7.37 (d, *J* = 9.0 Hz, 1H, H<sup>8</sup>), 7.08 (dd, *J* = 9.0, 2.9 Hz, 1H, H<sup>7</sup>), 3.89 (s, 2H, NH<sub>2</sub>), 2.61 – 2.27 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.85 – 1.70 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.03 (t, *J* = 7.4 Hz, 3H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  171.6 (C<sub>q</sub>), 171.5 (C<sub>q</sub>), 149.7 (C<sub>q</sub>), 144.8 (CH), 143.5 (C<sub>q</sub>), 123.7 (C<sub>q</sub>), 122.9 (CH), 122.6 (C<sub>q</sub>), 119.5 (CH), 107.2 (CH), 39.1 (CH<sub>2</sub>), 18.9 (CH<sub>2</sub>), 13.7 (CH<sub>3</sub>); HRMS (ESI) [M+H]<sup>+</sup> 247.1077, C<sub>13</sub>H<sub>14</sub>N<sub>2</sub>O<sub>3</sub> [M+H]<sup>+</sup> requires 247.1077.

#### Synthesis of ethyl 2-oxo-2-(piperidin-1-yl)acetate (8a)



Piperidine (1.2 mL, 11.7 mmol) and DIPEA (3.1 mL, 17.6 mmol) were dissolved in DCM (15 mL) and cooled to 0 °C before the addition of ethyl 2-chloro-2-oxoacetate (1.6 mL, 14.1 mmol). The reaction was allowed to warm to room temperature and stirred for 2 h. The reaction was then diluted with DCM and washed with sat. NaHCO<sub>3</sub> (aq.) and brine. The organic layer was separated, dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (10-50% ethyl acetate in cyclohexane) to afford compound **8a** (1.9 g, 85 %) as a colourless oil.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  4.34 (q, *J* = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.60 – 3.54 (m, 2H, NCH<sub>2</sub>), 3.37 – 3.32 (m, 2H, NCH<sub>2</sub>), 1.67-1.73 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 1.59-1.66 (m, 4H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 1.37 (t, *J* = 7.2 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  163.2 (C<sub>q</sub>), 160.2 (C<sub>q</sub>), 61.8 (CH<sub>2</sub>), 47.2 (CH<sub>2</sub>), 42.17 (CH<sub>2</sub>), 26.1 (CH<sub>2</sub>), 25.1 (CH<sub>2</sub>), 24.3 (CH<sub>2</sub>), 14.0 (CH<sub>3</sub>); HRMS (ESI) [M+Na]<sup>+</sup> 208.0942, C<sub>9</sub>H<sub>15</sub>NO<sub>3</sub> [M+Na]<sup>+</sup> requires 208.0944.

#### Synthesis of 2-oxo-2-(piperidin-1-yl)acetic acid (8)



Sodium hydroxide 0.45 M (16.0 mL, 11.0 mmol) was cooled to 0 °C before the addition of compound **8a** (1.9 g, 10.0 mmol) in THF (40 mL). The reaction was allowed to stir overnight. The reaction mixture was acidified to pH 1 with 1 M HCl (aq.) and extracted with ethyl acetate (x 3). The combined organic extracts were dried over sodium sulphate and concentrated to afford compound **8** (1.5 g, 96 %) as a white solid.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  4.13-4.08 (m, 2H, NCH<sub>2</sub>), 3.69-3.64 (m, 2H, NCH<sub>2</sub>), 1.81 – 1.42 (m, 6H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  161.4 (C<sub>q</sub>), 159.4 (C<sub>q</sub>), 47.9 (CH<sub>2</sub>), 45.2 (CH<sub>2</sub>), 26.6 (CH<sub>2</sub>), 25.6 (CH<sub>2</sub>), 24.1 (CH<sub>2</sub>); HRMS (ESI) [M+H]<sup>+</sup> 158.0812, C<sub>7</sub>H<sub>11</sub>NO<sub>3</sub> [M+H]<sup>+</sup> requires 158.0812.

Synthesis of *N*-(4-oxo-6-(2-oxo-2-(piperidin-1-yl)acetamido)-4*H*-chromen-3yl)butyramide (1)



Compound **8** (30 mg, 0.20 mmol) and DIPEA (80  $\mu$ L, 0.50 mmol) were dissolved in DMF (1.5 mL) before the addition of HATU (94 mg, 0.30 mmol) and **7** (42 mg, 0.20 mmol). After a period of 16 h the reaction mixture was diluted with ethyl acetate and washed with sat. NaHCO<sub>3</sub> (aq.) and brine. The organic layer was separated, dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (50-80% ethyl acetate in cyclohexane) to afford **1** (48 mg, 65%) as a cream solid.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  9.97 (s, 1H, NH), 9.50 (s, 1H, H<sup>2</sup>), 8.51 (s, 1H, NH), 8.46 (d, *J* = 2.6 Hz, 1H, H<sup>5</sup>), 8.28 (dd, *J* = 9.2, 2.6 Hz, 1H, H<sup>7</sup>), 7.55 (d, *J* = 9.2 Hz, 1H, H<sup>8</sup>), 4.13 - 4.08 (m, 2H, NCH<sub>2</sub>), 3.74 - 3.53 (m, 2H, NCH<sub>2</sub>), 2.50 (t, *J* = 7.5 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.82 - 1.75 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.74 - 1.69 (m, 6H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 1.02 (t, *J* = 7.4 Hz, 3H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  172.2 (C<sub>q</sub>), 171.6 (C<sub>q</sub>), 160.3 (C<sub>q</sub>), 159.3 (C<sub>q</sub>), 152.6 (C<sub>q</sub>), 145.5 (CH), 134.4 (C<sub>q</sub>), 126.6 (CH), 124.5 (C<sub>q</sub>), 122.2 (C<sub>q</sub>), 119.3 (CH), 115.7 (CH), 47.7 (CH<sub>2</sub>), 45.0 (CH<sub>2</sub>), 38.8 (CH<sub>2</sub>), 26.8 (CH<sub>2</sub>), 25.8 (CH<sub>2</sub>), 24.4 (CH<sub>2</sub>), 18.9 (CH<sub>2</sub>), 13.7 (CH<sub>3</sub>); HRMS (ESI) [M+H]<sup>+</sup> 386.1714, C<sub>20</sub>H<sub>23</sub>N<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> requires 386.1710.

Synthesis of N-isobutyryl-2-methoxy-5-nitrobenzohydrazide (9)<sup>3</sup>



Thionyl chloride (1.2 mL), 2-methoxy-5-nitrobenzoic acid (0.20 g, 1.0 mmol) and DMF (10  $\mu$ L) were heated to reflux for 2 h. The reaction mixture was concentrated to give a yellow solid. The crude acid chloride was dissolved in DCM (10 mL) and cooled to 0 °C before the addition of isobutyryl hydrazine (0.11 g, 1.1 mmol) and triethylamine (0.17 mL, 1.2 mmol) in DCM (10 mL). The reaction was allowed to warm to room temperature and stir for 2 h. The reaction mixture was then poured into sat. NaHCO<sub>3</sub> (aq.), the organic layer was separated and the aqueous extracted with DCM (x 3). The combined organic extracts were dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (50-100% ethyl acetate in DCM) to isolate compound **9** (0.26 g, 90% yield) as a white solid.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  10.82 (d, *J* = 7.3 Hz, 1H, NH), 9.85 (d, *J* = 7.4 Hz, 1H, NH), 9.07 (d, *J* = 2.9 Hz, 1H, H<sup>6</sup>), 8.39 (dd, *J* = 9.1, 3.0 Hz, 1H, H<sup>4</sup>), 7.15 (d, *J* = 9.2 Hz, 1H, H<sup>3</sup>), 4.19 (s, 3H, OCH<sub>3</sub>), 2.72 (hept., *J* = 6.9 Hz, 1H, CH), 1.28 (d, *J* = 6.9 Hz, 6H, 2 x CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  172.5 (C<sub>a</sub>), 161.5 (C<sub>a</sub>), 157.4 (C<sub>a</sub>), 142.0 (C<sub>a</sub>), 128.7 (CH), 128.0 (CH), 119.8 (C<sub>q</sub>), 112.0 (CH), 57.4 (CH<sub>3</sub>), 33.4 (CH), 19.3 (2 x CH<sub>3</sub>); HRMS (ESI) [M+H]<sup>+</sup> 282.1079, C<sub>12</sub>H<sub>15</sub>N<sub>3</sub>O<sub>5</sub> [M+H]<sup>+</sup> requires 282.1084.



Synthesis of 5-amino-N-isobutyryl-2-methoxybenzohydrazide (10)<sup>3</sup>

Compound **10** (0.10 g, 0.40 mmol) was dissolved in THF (10 mL) and the reaction flask was purged by vacuum/argon cycles (x 3) before the addition of palladium 10% on carbon (10 mg). The reaction vessel was placed under an atmosphere of hydrogen (40 psi) and stirred for 2 h at room temperature. The reaction mixture was filtered through celite and concentrated under vacuum. The crude material was purified by column chromatography (0-10% EtOH in DCM) to isolate compound **10** (85 mg, 95 % yield) as a yellow solid.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  10.94 (d, *J* = 7.4 Hz, 1H, NH), 9.54 (d, *J* = 7.5 Hz, 1H, NH), 7.51 (dd, *J* = 2.4, 1.0 Hz, 1H, H<sup>4</sup>), 6.83 (m, 2H, H<sup>3</sup>+H<sup>6</sup>), 3.96 (s, 3H, OCH<sub>3</sub>), 2.59 (hept., *J* = 6.9 Hz, 1H, CH), 1.23 (d, *J* = 6.9 Hz, 6H, 2xCH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  172.1 (C<sub>q</sub>), 160.2 (C<sub>q</sub>), 150.8 (C<sub>q</sub>), 140.4 (C<sub>q</sub>), 120.1 (CH), 119.1 (C<sub>q</sub>), 117.9 (CH), 112.8 (CH), 56.5 (CH<sub>3</sub>), 33.4 (CH), 19.4 (2 x CH<sub>3</sub>); HRMS (ESI) [M+H]<sup>+</sup> 252.1341, C<sub>12</sub>H<sub>17</sub>N<sub>3</sub>O<sub>3</sub> [M+H]<sup>+</sup> requires 252.1343.

Synthesis of *N*-(3-(2-isobutyrylhydrazinecarbonyl)-4-methoxyphenyl)-2-oxo-2-(piperidin-1-yl)acetamide (2)



Compound **8** (0.2 g, 1.2 mmol), thionyl chloride (4.5 ml) and DMF (10  $\mu$ L) were heated to reflux for 2 h. The reaction mixture was concentrated to give a yellow

solid. The crude acid chloride was dissolved in DCM (5.0 mL) and cooled to 0 °C before compound **10** (0.24 mg, 1.0 mmol) and triethylamine (0.16 mL, 1.2 mmol) in DCM (5.0 mL) were added. The reaction was allowed to warm to room temperature and stir for 1 h before it was poured into sat. NaHCO<sub>3</sub> (aq.). The organic layer was separated and the aqueous layer was extracted with DCM (x 3). The combined organic extracts were dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (50-100% ethyl acetate in DCM) to afford compound **2** (0.36 g, 97 % yield) as a white solid.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  11.34 (d, *J* = 8.4 Hz, 1H, NH), 11.07 (d, *J* = 8.3 Hz, 1H, NH), 10.71 (s, 1H, NH), 8.41 (d, *J* = 2.8 Hz, 1H, H<sup>6</sup>), 8.37 (dd, *J* = 8.9, 2.7 Hz, 1H, H<sup>4</sup>), 7.01 (d, *J* = 9.0 Hz, 1H, H<sup>3</sup>), 4.05 (s, 3H, OCH<sub>3</sub>) 3.84 (t, *J* = 4.9 Hz, 2H, NCH<sub>2</sub>), 3.63 (t, *J* = 5.3 Hz, 2H, NCH<sub>2</sub>), 2.84 (hept., *J* = 6.8 Hz, 1H, CH), 1.72-1.62 (m, 6H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 1.22 (d, *J* = 6.9 Hz, 6H, 2 x CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  172.0 (C<sub>q</sub>), 162.5 (C<sub>q</sub>), 160.2 (C<sub>q</sub>), 158.0 (C<sub>q</sub>), 154.3 (C<sub>q</sub>), 131.6 (C<sub>q</sub>), 125.9 (CH), 124.4 (CH), 118.9 (C<sub>q</sub>), 111.7 (CH), 56.6 (CH<sub>3</sub>), 47.8 (CH<sub>2</sub>), 44.2 (CH<sub>2</sub>), 32.6 (CH), 26.6 (CH<sub>2</sub>), 25.7 (CH<sub>2</sub>), 24.4 (CH<sub>2</sub>), 19.6 (2 x CH<sub>3</sub>); HRMS (ESI) [M+H]<sup>+</sup> 391.1964, C<sub>19</sub>H<sub>26</sub>N<sub>4</sub>O<sub>5</sub> [M+H]<sup>+</sup> requires 391.1976.

# Synthesis of *N*-(3-(2-isobutyrylhydrazine-1-carbonyl)-methoxyphenyl)butyramide (2a)



Compound **10** (0.5 g, 2.0 mmol) and pyridine (0.24 mL, 3.0 mmol) were dissolved in DCM (10 mL) and cooled to 0 °C before the addition of butyric anhydride (0.36 mL, 2.2 mmol). The reaction was allowed to warm to room temperature and stirred for 1 h. The reaction mixture was then diluted with DCM and washed with sat. NaHCO<sub>3</sub> (aq.) and brine. The organic layer was separated, dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (0-10% EtOH in DCM) to afford **2a** (0.53 g, 83 % yield) as a white

solid.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  10.41 (s, 1H, NH), 9.45 (s, 1H, NH), 8.09 (dd, *J* = 9.0, 2.8 Hz, 1H, H<sup>4</sup>), 7.83 (d, *J* = 2.8 Hz, 1H, H<sup>6</sup>), 6.89 (d, *J* = 9.0 Hz, 1H, H<sup>3</sup>), 3.97 (s, 3H, OCH<sub>3</sub>), 2.63 (app. quint., *J* = 6.9 Hz, 1H, CH), 2.10 (s, 2H, CH<sub>2</sub>), 1.60 (q, *J* = 8.2, 7.3 Hz, 2H, CH<sub>2</sub>), 1.21 (d, *J* = 6.9 Hz, 6H, 2 x CH<sub>3</sub>), 0.78 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  173.5 (C<sub>q</sub>), 172.7 (C<sub>q</sub>), 160.5 (C<sub>q</sub>), 153.6 (C<sub>q</sub>), 132.5 (C<sub>q</sub>), 126.1 (CH), 123.4 (CH), 118.5 (C<sub>q</sub>), 111.8 (CH), 56.5 (CH<sub>3</sub>), 38.9 (CH<sub>2</sub>), 33.2 (CH), 19.4 (2 x CH<sub>3</sub>), 18.9 (CH<sub>2</sub>), 13.6 (CH<sub>3</sub>); HRMS (ESI) [M+H]<sup>+</sup> 322.1757, C<sub>16</sub>H<sub>23</sub>N<sub>3</sub>O<sub>4</sub> [M+H]<sup>+</sup> requires 391.1761.

Synthesis of (*S*)-*N*-((*S*)-1-(butylamino)-3-methyl-1-oxobutan-2-yl)-2-((*S*)-2isobutyramido-3-phenylpropanamido)-4-methylpentanamide (11)<sup>4</sup>



Compound **11** was synthesized *via* solid phase peptide synthesis using an Activotec P14 peptide synthesizer. Fmoc-Pal-Am Resin (2.5 g, 0.53 mmol/g) was shaken with DMF (2 x 25 ml) for 5 mins. The resin was then shaken with a 20% piperidine solution in NMP (3 x 25 ml) for 12 minutes. The resin was then washed (2 x DMF, 2 x DCM, 2 x MeOH, 2 x DCM and MeOH alternately and then diethyl ether).

- To the resin was added DCM (25 mL), collidine (1.0 mL, 8.0 mmol) and 2nitrobenzene-1-sulfonyl chloride (0.88 g, 4.0 mmol). The resulting suspension was agitated for 3 h. The resin was drained and washed as described above.
- To the resin was added DMF (25 mL), 7-methyl-1,5,7-triazabicyclo[4.4.0]dec-5-ene (MTBD) (0.80 mL, 5.3 mmol) and 1-iodobutane (1.5 mL, 13.3 mmol). The resulting suspension agitated for a period of 8 h after which the resin was drained and washed as before.
- To the resin was added DMF (25 mL), DBU (1.0 ml, 6.6 mmol) and

mercaptoethanol (0.90 mL, 13.3 mmol). The resin was agitated for 3 h and then drained. The process was repeated and then resin was washed as before.

- Fmoc-Val-OH (4.5 g, 13.3 mmol) was dissolved in a 10% solution of DIPEA in DMF (25 mL) before the addition of HATU (4.5 g, 12.0 mmol). The reaction mixture was allowed to stir for 15 mins after which time it was added to the resin and agitated for 2 h. The process was repeated and the resin was washed as before.
- The resin was then shaken with a 20% piperidine solution in NMP (3 x 25 ml) for 12 minutes. The resin was then washed (2 x DMF, 2 x DCM, 2 x MeOH, 2 x DCM and MeOH alternately and then diethyl ether).
- Fmoc-Leu-OH (2.1 g, 5.3 mmol) was dissolved in a 10% solution of DIPEA in DMF (25 mL) before the addition of HATU (1.8 g, 4.8 mmol). The reaction mixture was allowed to stir for 15 mins after which time it was added to the resin and agitated for 2 h. The process was repeated and the resin was washed as before.
- The resin was then shaken with a 20% piperdine solution in NMP (3 x 25 ml) for 12 minutes. The resin was then washed (2 x DMF, 2 x DCM, 2 x MeOH, 2 x DCM and MeOH alternately and then diethyl ether).
- Fmoc-Phe-OH (1.9 g, 5.3 mmol) was dissolved in a 10% solution of DIPEA in DMF (25 mL) before the addition of HATU (1.8 g, 4.8 mmol). The reaction mixture was allowed to stir for 15 min after which time it was added to the resin and agitated for 2 h. The process was repeated and the resin was washed as before.
- The resin was then shaken with a 20% piperdine solution in NMP (3 x 25 ml) for 12 minutes. The resin was then washed (2 x DMF, 2 x DCM, 2 x MeOH, 2 x DCM and MeOH alternately and then diethyl ether).
- To the resin was added a 10% solution of DIPEA in DMF (25 mL) and isobutyryl chloride (0.60 ml, 5.3 mmol). The suspension was agitated for 2 h before the resin was drained and washed as before.
- The resin was suspended in a 1:1 mixture of TFA and DCM (20 mL) for 1 h

without agitation. The resin was removed by filtration and the filtrate was concentrated under vacuum. The crude material was purified by HPLC to afford **11** (0.17 g, 31 %) as a white solid.

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.21 (s, 1H, NH), 7.62 (s, 2H, 2 x NH), 7.24 – 7.02 (m, 6H, ArCH, NH), 5.19 (app. q, *J* = 7.0 Hz, 1H, H<sup>1</sup>), 4.82 (app. q, *J* = 7.8 Hz, 1H, H<sup>2</sup>), 4.50 (app. t, *J* = 8.9 Hz, 1H, H<sup>3</sup>), 3.34 – 3.26 (m, 1H, NCH), 3.20 – 3.12 (m, 1H, NCH), 3.07 – 2.98 (m, 2H, PhCH<sub>2</sub>), 2.48 (h, *J* = 6.8 Hz, 1H, HC(CH<sub>3</sub>)<sub>2</sub>), 2.13 – 2.05 (m, 1H, HC(CH<sub>3</sub>)<sub>2</sub>), 1.70 – 1.54 (m, 3H, CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>, CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub> ), 1.54 – 1.46 (m, 2H, CH<sub>3</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>NH), 1.34 (app. h, *J* = 7.3 Hz, 2H, CH<sub>3</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>NH), 1.06 (d, J = 7.0 Hz, 3H, CH<sub>3</sub>), 1.04 (d, *J* = 7.1 Hz, 3H, CH<sub>3</sub>), 0.97 (d, *J* = 6.7 Hz, 3H, CH<sub>3</sub>), 0.94 (d, *J* = 6.8 Hz, 3H, CH<sub>3</sub>), 0.91 (t, *J* = 7.4 Hz, 3H, CH<sub>3</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>NH), 0.87 (t, *J* = 6.3 Hz, 6H, 2 x CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  176.8 (Cq), 172.0 (Cq), 171.1 (Cq), 136.6 (Cq), 129.5 (ArCH), 128.1 (ArCH), 126.6 (ArCH), 58.8 (CH), 53.2 (CH), 51.6 (CH), 42.5 (CH<sub>2</sub>), 39.1 (CH<sub>2</sub>), 38.9 (CH<sub>2</sub>), 34.9 (CH), 31.5 (CH<sub>2</sub>), 31.0 (CH), 29.7 (CH), 24.8 (CH<sub>3</sub>), 22.6 (CH<sub>3</sub>), 20.11 (CH<sub>2</sub>), 19.7 (CH<sub>3</sub>), 19.2 (CH<sub>3</sub>), 19.1 (CH<sub>3</sub>), 18.6 (CH<sub>3</sub>), 13.7 (CH<sub>3</sub>); HRMS (ESI) [M+H]<sup>+</sup> 503.3577, C<sub>28</sub>H<sub>46</sub>N<sub>4</sub>O<sub>4</sub> [M+H]<sup>+</sup> requires 503.3592.

#### 2. <sup>1</sup>H NMR Titration Data

#### 2.1. Compound 1 Homodimer VT <sup>1</sup>H NMR Titration



**Figure 1:** Homodimerisation of compound **1**. Resonances (NH<sup>1</sup> and NH<sup>2</sup>) used to determine  $K_{DIM}$  are highlighted in red.



#### 2.1.1. Compound 1 Homodimer <sup>1</sup>H NMR Titration Experiment 1























#### 2.1.3. Compound 1 Homodimer <sup>1</sup>H NMR Titration Experiment 3









## 2.2.4. Compound 1 Homodimer VT <sup>1</sup>H NMR Summary and Van't Hoff Plots

Temp	K <sub>DIM</sub>			Compound 1 Homodimer Van't Hoff
(К)	(M <sup>-1</sup> )	1/T	In(K <sub>DIM</sub> )	5.0 1/1 VS INK <sub>DIM</sub> $R^2 = 0.995$
278	73	0.003597122	4.290459441	4.5-
288	45	0.003472222	3.80666249	¥.0-
295	30	0.003389831	3.401197382	<u><u> </u></u>
298	26	0.003355705	3.258096538	3.0-
308	18	0.003246753	2.890371758	2.5
				1/Т

**Table 16**: Summary of Compound 1 Homodimer Experiment 1 and Van't Hoff Plot

**Table 17**: Summary of Compound 1 Homodimer Experiment 2 and Van't Hoff Plot



**Table 18**: Summary of Compound 1 Homodimer Experiment 3 and Van't Hoff Plot

Temp	K <sub>DIM</sub>			Compound 1 Homodimer Van't Hoff
(К)	(M <sup>-1</sup> )	1/T	In(K <sub>DIM</sub> )	4.5 1/1 vs $InK_{DIM}$ $R^2 = 0.9996$
278	61	0.003597122	4.110873864	4.0-
288	39	0.003472222	3.663561646	¥ 3.5-
295	29	0.003389831	3.36729583	
298	25	0.003355705	3.218875825	
308	17	0.003246753	2.833213344	2.5 0.0032 0.0034 0.0036
				1/1

	Table 19: Summary	ı of	f thermod	ynamic	parameters	for com	pound 1	homodimer
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Expt	ΔH (kcal mol <sup>-1</sup> )	-ΤΔS <sup>295 κ</sup> (kcal mol <sup>-1</sup> )	ΔG (kcal mol <sup>-1</sup> )
1	-8.1	6.1	-2.0
2	-7.9	6.0	-1.9
3	-7.3	5.3	-2.0
Average	-7.8	5.8	-2.0

#### 2.2. Tripeptide 11 Homodimer <sup>1</sup>H NMR Titration



**Figure 2:** Homodimerisation of **11**. Resonance ( $H^2$ ) used to determine  $K_{DIM}$  is highlighted in red



**Figure 3:** Comparison between NH shifts (left) and  $\alpha$ CH shifts (right) of compound 11. NH signals broaden into the baseline at lower concentrations making it difficult to accurately measure chemical shift.  $\alpha$ CH signals are relatively sharp compared to NH signals and therefore were used to determine binding constants















#### 2.2.2. Compound 11 Homodimer <sup>1</sup>H NMR Titration Experiment 2









#### 2.2.3. Compound 11 Homodimer <sup>1</sup>H NMR Titration Experiment 3







## 2.2.4. Compound 11 Homodimer VT <sup>1</sup>H NMR Summary and Van't Hoff Plots

Temp	K <sub>DIM</sub>			Compound 11 Homodimer Van't Hoff
(К)	(M <sup>-1</sup> )	1/T	In(K <sub>DIM</sub> )	6.5 1/1 vs InK <sub>DIM</sub>
308	54	0.003246753	3.988984047	6.0 R <sup>2</sup> = 0.990
298	110	0.003355705	4.700480366	¥ 5.0-
295	126	0.003389831	4.836281907	4.5-
288	214	0.003472222	5.365976015	4.0-
278	360	0.003597122	5.886104031	3.5 0.0032 0.0034 0.0036
				1/1

**Table 33**: Summary of compound 11 Homodimer Experiment 1 and Van't Hoff Plot

**Table 34**: Summary of compound 11 Homodimer Experiment 2 and Van't Hoff Plot



**Table 35**: Summary of compound 11 Homodimer Experiment 3 and Van't Hoff Plot

Temp	K <sub>DIM</sub>			Compound 11 Homodimer Van't Hoff
(К)	(M <sup>-1</sup> )	1/T	In(K <sub>DIM</sub> )	6.5 1/1 VS III ADIM
308	50	0.003246753	3.912023005	6.0 5 5
298	95	0.003355705	4.553876892	
295	156	0.003389831	5.049856007	E
278	301	0.003597122	5.707110265	4.0-
				3.5 <del>1</del> 0.0032 0.0034 0.0036
				1/Τ

	Table 36: Summar	y of thermod	lynamic parameters	for compoun	d 11 homodimer
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Expt	ΔH (kcal mol <sup>-1</sup> )	-TΔS <sup>295 K</sup> (kcal mol <sup>-1</sup> )	ΔG (kcal mol <sup>-1</sup> )
1	-10.8	7.9	-2.9
2	-10.0	7.2	-2.8
3	-9.9	7.1	-2.8
Average	-10.2	7.4	-2.8

#### 2.3. Compound 2 Homodimer VT <sup>1</sup>H NMR Titration



**Figure 4:** Homodimerisation of HAO. Resonances used to calculate  $K_{DIM}$  are highlighted in red.

**N.B.**  $K_{\text{DIM}}$  was calculated using NH<sup>2</sup> and NH<sup>3</sup>. The shifts of NH<sup>1</sup> are included for completeness.

2.3.1. Compound 2 Homodimer <sup>1</sup> H NMR	<b>Titration Exp</b>	periment 1
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Table 4	<b>11:</b> <sup>1</sup> H NMI	R titration	of 2 at 308	3 К	
[2]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	Compound 2 Homodimer 308	к
(mM)	(ppm)	(ppm)	(ppm)	12-	→ NH <sup>1</sup>
51.4	11.2662	10.8737	10.5953		→ NH <sup>2</sup>
25.7	11.2422	10.7626	10.5203		- NH <sup>3</sup>
12.9	11.2078	10.6146	10.4197		
6.4	11.1620	10.4244	10.2895	Che	
3.2	11.1049	10.1903	10.1294	9 <del>1 1 1</del> 0 20 40	<b>6</b> 0
1.6	11.0387	9.9437	9.9191	Concentration (mM)	
0.8	10.9681	9.6337	9.7483		
0.4	10.9020	9.3610	9.5624		
				K <sub>DIM</sub> = 571 M <sup>-1</sup>	sigma= 0.6

#### 2.3.2. Compound 2 Homodimer <sup>1</sup> H NMR Titration Experiment 2










Table 4	Table 47: <sup>1</sup> H NMR titration of 2 at 278 K					
[2]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	Compound 2 Homodimer 278K N=3		
(mM)	(ppm)	(ppm)	(ppm)	127 - NH1		
54.2	11.4141	11.2535	10.8103			
27.1	11.4077	11.2146	10.7847			
13.6	11.3951	11.1539	10.7440			
6.8	11.3772	11.0729	10.6890	Cher		
3.4	11.3520	10.9635	10.6132	9-1		
1.7	11.3178	10.8147	10.5117	Concentration (mM)		
0.9	11.2747	10.6290	10.3839			
0.4	11.2194	_*	10.2199			
			1	K <sub>DIM</sub> = 5430 M <sup>-1</sup> sigma= 1.4		
				*Broadened signal		

### 2.3.3. Compound 2 Homodimer <sup>1</sup> H NMR Titration Experiment 3









Temp (K)	К <sub>DIM</sub> (М <sup>-1</sup> )	1/T	InK	Compound 2 Homodimer Vant Hoff 1/T v K <sub>DIM</sub>
308	570	0.003246753	6.345636361	9- 9-
298	1099	0.003355705	7.002155954	° Sim
295	1403	0.003389831	7.24636808	7-
288	2318	0.003472222	7.748460024	
278	5430	0.003597122	8.599694413	0.0032 0.0034 0.0036 1/T

## 2.3.4. Compound 2 Homodimer VT <sup>1</sup>H NMR Summary and Van't Hoff Plots

 Table 52: Summary of compound 2 Homodimer Experiment 1 and Van't Hoff Plot

Temp (K)	К <sub>DIM</sub> (М⁻¹)	1/T	InK	Compound 2 Homodimer Vant Hoff <sup>9</sup> <sup>1/T</sup> v K <sub>DIM</sub>
308	571	0.003246753	6.34738921	R0.9992
295	1360	0.003389831	7.215239979	Y Y
288	2352	0.003472222	7.763021309	<sup>⊑</sup> 7
278	4747	0.003597122	8.465268119	
				6-1 0.0032 0.0034 0.0036 1/T

 Table 53: Summary of compound 2 Homodimer Experiment 2 and Van't Hoff Plot

Temp	<b>K</b> <sub>DIM</sub>		InK	Compound 2 Homodimer Vant Hoff
(К)	(M <sup>-1</sup> )	1/T		$P^{9}$
308	502	0.003246753	6.21860012	8-
298	1009	0.003355705	6.91671502	
295	1271	0.003389831	7.147559271	
288	2262	0.003472222	7.724004657	
278	4928	0.003597122	8.502688505	5 <del>1</del> 0.0032 0.0034 0.0036
				1/Т

 Table 54: Summary of compound 2 Homodimer Experiment 3 and Van't Hoff Plot

	Table 55: Summar	v o	f thermod	ynamic	parameters	for com	pound 2	? homodimer
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Expt	ΔH (kcal mol <sup>-1</sup> )	-TΔS <sup>295 K</sup> (kcal mol <sup>-1</sup> )	ΔG (kcal mol⁻¹)
N1	- 12.1	7.8	- 4.3
N2	- 13.0	8.4	- 4.6
N3	- 12.8	8.5	- 4.3
Average	- 12.6	8.4	- 4.4

## 2.4. 11:1 Heterodimer VT <sup>1</sup>H NMR Titration Experiment



**Figure 5:** *Heterodimerisation of 1 and 11. Resonances used to determine binding constants are highlighted in red.* 

**N.B.**  $K_{Het}$  was calculated using NH<sup>1</sup> and H<sup>2</sup>. The shift of NH<sup>2</sup> is included for completeness

## 2.4.1 11:1 Heterodimer <sup>1</sup> H NMR Titration Experiment 1



#### Peptide concentration: 4 mM









## **2.4.2. 11:1** Heterodimer <sup>1</sup> H NMR Titration Experiment 2 Peptide concentration: 4.3 mM

Table 6	Table 61: 11:1 Heterodimer <sup>1</sup> H NMR titration at 278 K					
[1]	$\delta  NH^1$	δ NH <sup>2</sup>	δ H <sup>2</sup>	11:1 Heterodimer 278K N=2		
(mM)	(ppm)	(ppm)	(ppm)			
42.4	10.5293	9.0430	5.0649	€ 10		
28.0	10.4168	8.9813	5.0377			
18.5	10.3134	8.9344	5.0118	5 8-		
12.2	10.2139	8.8966	4.9801	ca		
8.1	10.1323	8.8663	4.9491	- E 6-		
5.3	10.0673	-*	4.9057	້ອ		
3.5	10.0200	-*	4.8672			
2.3	9.9901	_*	4.8290	0 10 20 30 40 50 Concentration (mM)		
1.5	9.9778	_*	4.7984			
1.0	9.9747	_*	4.7739			
0.7	9.9689	_*	4.7586			
0		-	4.7215			
				K <sub>Het</sub> = 838 M <sup>-1</sup> sigma= 9.8		
				*Broadened signal		









# **2.4.3. 11:1** Heterodimer <sup>1</sup> H NMR Titration Experiment **3** Peptide concentration: 3.3 mM



 
 Table 67: 11:1 Heterodimer <sup>1</sup>H NMR titration at 288 K
  $\delta H^2$ δNH<sup>1</sup>  $\delta NH^2$ [1] 11:1 Heterodimer 288K N=3 (mM) (ppm) (ppm) (ppm) NH<sup>1</sup> 10 30.3 10.2793 4.9952 Chemical shift (ppm) 8.8281 NH<sup>2</sup> 22.7 10.2100 8.7867 4.9713  $H^2$ 8. 17.0 4.9477 10.1383 8.7475 12.8 10.0735 8.7132 4.9128 6 9.6 4.8825 10.0179 8.6835 7.2 9.9683 8.6602 4.8536 5.4 9.9271 8.6426 4.8189 20 40 10 30 4.1 9.8958 8.6308 4.7877 Concentration (mM) 3.0 9.8711 8.6194 4.7572 2.3 9.8559 8.6163 4.7311 1.7 4.7090 9.8438 8.6113 1.3 4.6887 9.8355 8.6083 0 4.6169 \_ \_ K<sub>Het</sub>= 471 M<sup>-1</sup> sigma= 6.6







## 2.4.4. 11:1 Heterodimer VT <sup>1</sup>H NMR Summary and Van't Hoff Plots

 Table 71: Summary of 11:1 Heterodimer Experiment 1 and Van't Hoff Plot

Temp (K)	K <sub>Het</sub> (M <sup>-1</sup> )	1/T	InK <sub>Het</sub>	11:1 Heterodimer VT 7.5-1/T vs InK <sub>Het</sub>
308	142	0.003246753	4.955827058	7.0-
298	272	0.003355705	5.605802066	6.5- <sup>∓</sup>
295	332	0.003389831	5.805134969	¥ 6.0-
288	512	0.003472222	6.238324625	5.0-
278	904	0.003597122	6.80682936	4.5
				1/Т

**Table 72**: Summary of 11:1 Heterodimer Experiment 2 and Van't Hoff Plot

Temp	K <sub>Het</sub>			11:1 Heterodimer VT
(K)	(M <sup>-1</sup> )	1/T	InK <sub>Het</sub>	7.5 1/T vs InK <sub>Het</sub>
308	157	0.003246753	5.056245805	$R^2 = 0.996$
298	257	0.003355705	5.549076085	± 0.5 ↓ 6.0-
295	323	0.003389831	5.777652323	<u>=</u> 5.5-
288	501	0.003472222	6.216606101	5.0-
278	838	0.003597122	6.7310181	4.5 <del>1</del> 0.0032 0.0034 0.0036
		•	•	1/π

**Table 73**: Summary of 11:1 Heterodimer Experiment 3 and Van't Hoff Plot

Temp (K)	K <sub>Het</sub> (M <sup>-1</sup> )	1/T	InK <sub>Het</sub>	11:1 Heterodimer VT 1/T vs InK <sub>Het</sub>
308	115	0.003246753	4.744932128	$R^2 = 0.990$
298	225	0.003355705	5.416100402	
295	294	0.003389831	5.683579767	
288	471	0.003472222	6.154858094	5
278	779	0.003597122	6.658011046	4 0.0032 0.0034 0.0036
				1/Т

**Table 74:** Summary of thermodynamic parameters for 11:1 heterodimer

Expt	ΔH (kcal mol⁻¹)	-TΔS <sup>295 κ</sup> (kcal mol <sup>-1</sup> )	ΔG (kcal mol <sup>-1</sup> )
N1	- 10.9	7.6	- 3.3
N2	- 9.7	6.3	- 3.1
N3	- 10.5	7.1	- 3.4
Average	- 10.4	7.0	- 3.3

## 2.5. 11:2 Heterodimer VT <sup>1</sup>H NMR Titration Experiment



**Figure 6:** *Heterodimerisation of 2 and 11. Resonances used to determine binding constants are highlighted in red.* 

 $\textbf{N.B.}~K_{Het}$  was calculated on using  $NH^3$  and  $H^2.$  Shifts  $NH^1$  and  $NH^2$  are included for completeness

## **2.5.1. 11:2** Heterodimer <sup>1</sup> H NMR Titration Experiment 1 Peptide concentration: 3.1 mM

Table 7	Table 75: 11:2 Heterodimer <sup>1</sup> H NMR titration at 278 K					
[2]	$\delta  NH^1$	$\delta  NH^2$	δ NH³	$\delta H^2$	11:2 Heterodimer 278K	
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 <b>-</b> NH <sup>1</sup>	
34.0	11.3932	11.2824	10.7386	5.0457		
25.5	11.3870	11.2795	10.7124	5.0387	₩10- ₩10- ₩10- ₩10- ₩10- ₩10- ₩10- ₩10-	
19.1	11.3783	11.2777	10.6782	5.0298		
14.3	11.3705	_*	10.6443	5.0200		
10.8	11.3605	_*	10.6006	5.0075		
8.1	11.3493	_*	10.5508	4.9874	0 10 20 30 40	
6.1	11.3372	_*	10.4955	4.9725	Concentration (mM)	
4.5	11.3259	_*	10.4395	4.9524		
3.4	11.3153	-*	10.3878	4.9266		
2.6	11.3076	_*	10.3444	4.8988		
1.9	11.3011	_*	10.3155	4.8644		
1.4	11.2955	_*	10.2884	4.8307		
0	-	-	-	4.4425		
					K <sub>DIM</sub> = 13,433 M <sup>-1</sup> sigma= 6.8	
					*Broadened signal	



*Broadened	signal
------------	--------

Table 77: 11:2 Heterodimer <sup>1</sup> H NMR titration at 295 K								
[2]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	$\delta NH^3$	δ H <sup>2</sup>	11:2 Heterodimer 295K			
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 <b>−</b> NH <sup>1</sup>			
34.0	11.3163	11.0776	10.6315	5.0225				
25.5	11.3051	11.0557	10.5961	5.0086	E 10- ↓ NH <sup>3</sup> ↓ H <sup>2</sup>			
19.1	11.2947	11.0388	10.5580	4.9935				
14.3	11.2837	11.0262	10.5195	4.9771				
10.8	11.2718	11.0153	10.4756	4.9573	ō			
8.1	11.2590	11.0073	10.4265	4.9370	0 10 20 30 40			
6.1	11.2462	11.0068	10.3763	4.9078	Concentration (mM)			
4.5	11.2342	11.0128	10.3280	4.8803				
3.4	11.2247	11.0240	10.2865	4.8431				
2.6	11.2156	11.0342	10.2494	4.8056				
1.9	11.2088	11.0460	10.2207	4.7686				
1.4	11.2042	_*	10.1996	4.7322				
0	-	-	-	4.5531				
					K <sub>DIM</sub> = 3182 M <sup>-1</sup> sigma= 10.3			
					*Broadened signal			



Table 7	<b>79:</b> 11:2 He	eterodime	r <sup>1</sup> H NMR t	308 К		
[2]	δ NH <sup>1</sup>	$\delta  NH^2$	$\delta NH^3$	δ H <sup>2</sup>	11:2 Heterodimer 308K	
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15	→ NH <sup>1</sup>
34.0	11.2425	10.8723	10.5181	4.9845		➡ NH <sup>2</sup>
25.5	11.2305	10.8416	10.4798	4.9652		- NH <sup>3</sup> - H <sup>2</sup>
19.1	11.2171	10.8094	10.4356	4.9431		
14.3	11.2046	10.7834	10.3942	4.9197		
10.8	11.1894	10.7509	10.3449	4.8910		_
8.1	11.1754	10.7235	10.2959	4.8594	0 10 20 30 Concentration (mM)	40
6.1	11.1606	10.7014	10.2458	4.8246		
4.5	11.1470	10.6834	10.1990	4.7854		
3.4	11.1368	10.6752	10.1608	4.7522		
2.6	11.1243	10.6502	10.1200	4.7080		
1.9	11.1154	10.6386	10.0893	4.6705		
1.4	11.1087	10.6361	10.0648	4.6362		
0	-	-	-	4.4581		
					K <sub>DIM</sub> = 1178 Μ <sup>-1</sup>	sigma=6.9

**2.5.2. 11:2 Heterodimer** <sup>1</sup> H NMR Titration Experiment 2 Peptide concentration: 3.1 mM

Table 80: 11:2 Heterodimer <sup>1</sup> H NMR titration at 278 K							
[2]	δ NH <sup>1</sup>	$\delta NH^2$	$\delta NH^3$	δ H <sup>2</sup>	11:2 Heterodimer 278K N=2		
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 → NH <sup>1</sup>		
35.9	11.3964	11.2951	10.7453	5.0465			
26.9	11.3881	11.2854	10.7166	5.0386			
20.2	11.3807	11.2817	10.6863	5.0312			
15.1	11.3721	11.2837	10.6492	5.0207	Š		
11.4	11.3612	11.2936	10.6031	5.0087	0 10 20 30 40		
8.5	11.3517	_*	10.5603	4.9938	Concentration (mM)		
6.4	11.3345	_*	10.5124	4.9739			
4.8	11.3305	_*	10.4680	4.9580			
3.6	11.3255	_*	10.4121	4.9323			
2.7	11.3170	_*	10.3535	4.9040			
2.0	11.3106	_*	10.3194	4.8718			
1.5	11.2973	_*	10.2963	4.8413			
0	-	_*	-	4.6830			
					K <sub>DIM</sub> = 9, 679 M <sup>-1</sup> sigma= 4.5		
					*Broadened signal		

Table 81: 11:2 Heterodimer <sup>1</sup> H NMR titration at 288 K							
[2]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2 Heterodimer 288K N=2		
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 → NH1		
35.9	11.3490	11.1750	10.6814	5.0343			
26.9	11.3415	11.1566	10.6521	5.0251			
20.2	11.3335	11.1549	10.6204	5.0153			
15.1	11.3229	11.1436	10.5812	5.0008	CP		
11.4	11.3109	11.1412	10.5355	4.9835			
8.5	11.3002	11.1406	10.4923	4.9657	Concentration (mM)		
6.4	11.2879	11.1523	10.4392	4.9434			
4.8	11.2777	11.1687	10.3920	4.9178			
3.6	11.2684	_*	10.3442	4.8914			
2.7**	11.2594	_*	10.3075	4.8562			
2.0**	11.2517	_*	10.2724	4.8192			
1.5**	11.2465	-*	10.2528	4.7862			
0	-	-	-	4.6363			
					K <sub>DIM</sub> = 5346 M <sup>-1</sup> sigma= 3.6		
					*Obscured by another proton signal		
0	-	-	-	4.6363	K <sub>DIM</sub> = 5346 M <sup>-1</sup> sigma= 3.6         *Obscured by another proton signal		

\*\*Outlier not included in K<sub>Het</sub> determination

Table 82: 11:2 Heterodimer <sup>1</sup> H NMR titration at 295 K							
[2]	δNH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2 Heterodimer 295K N=2		
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 → NH <sup>1</sup>		
35.9	11.3139	11.0727	10.6310	5.0211			
26.9	11.3056	11.0582	10.5996	5.0095			
20.2	11.2970	11.0455	10.5665	4.9972			
15.1	11.2849	11.0303	10.5246	4.9790	ö		
11.4	11.2772	11.0201	10.4787	4.9593	0 10 20 30 40		
8.5	11.2606	11.0105	10.4347	4.9380			
6.4	11.2484	11.0136	10.3843	4.9100			
4.8	11.2370	11.0175	10.3383	4.8835			
3.6	11.2280	11.0388	10.2954	4.8530			
2.7	11.2182	11.0342	10.2588	4.8166			
2.0	11.2123	_*	10.2278	4.7778			
1.5	11.2076	_*	10.2079	4.7421			
0	-	-	-	4.5564			
					K <sub>DIM</sub> = 3351 M <sup>-1</sup> sigma= 12.4		
					*Broadened signal		

Table 83: 11:2 Heterodimer <sup>1</sup> H NMR titration at 298 K							
[2]	δNH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2 Heterodimer 298K N=2		
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 → NH <sup>1</sup>		
35.9	11.3002	11.0343	10.6105	5.0156			
26.9	11.2901	11.0151	10.5764	5.0017	ੇ ਸ਼ਿੰਫ ਬੁ		
20.2	11.2805	10.9979	10.5414	4.9878			
15.1	11.2689	10.9801	10.4999	4.9688			
11.4	11.2557	10.9695	10.4532	4.9480	0 10 20 30 40 Concentration (mM)		
8.5	11.2435	10.9525	10.4087	4.9242			
6.4	11.2297	10.9482	10.3572	4.8962			
4.8	11.2182	10.9481	10.3119	4.8666			
3.6	11.2092	10.9662	10.2707	4.8351			
2.7	11.1992	10.9552	10.2339	4.7953			
2.0	11.1927	10.9860	10.2034	4.7597			
1.5	11.1872	10.9932	10.1821	4.7222			
0	-	-	-	4.5300			
					K <sub>DIM</sub> = 2218 M <sup>-1</sup> sigma= 14.0		

Table 84: 11:2 Heterodimer <sup>1</sup> H NMR titration at 308 K							
[2]	δNH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2 Heterodimer 308K N=2		
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 - NH1		
35.9	11.2428	10.8763	10.5224	4.9856			
26.9	11.2323	10.8488	10.4857	4.9680			
20.2	11.2208	10.8206	10.4466	4.9487			
15.1	11.2065	10.7890	10.4009	4.9238	Ğ		
11.4	11.1918	10.7617	10.3520	4.8966	0 10 20 30 40		
8.5	11.1776	10.7321	10.3052	4.8658	Concentration (mM)		
6.4	11.1636	10.7126	10.2551	4.8323			
4.8	11.1506	10.6935	10.2098	4.7964			
3.6	11.1415	10.6967	10.1724	4.7659			
2.7	11.1282	10.6614	10.1312	4.7206			
2.0	11.1225	10.6809	10.1042	4.6844			
1.5	11.1142	10.6663	10.0776	4.6468			
0	-	-	-	4.4740			
					K <sub>DIM</sub> = 1205 M <sup>-1</sup> sigma= 7.9		

**2.5.3. 11:2** Heterodimer <sup>1</sup> H NMR Titration Experiment 3 Peptide concentration: 3.2 mM

Table 85: 11:2 Heterodimer <sup>1</sup> H NMR titration at 278 K								
[2]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2 Heterodimer 278K N=3			
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 - NH1			
34.3	11.3982	11.2954	10.7459	5.0475				
25.7	11.3904	11.2873	10.7184	5.0407	—			
19.3	11.3813	11.2821	10.6851	5.0314				
14.5	11.3730	11.2811	10.6516	5.0211				
10.9	11.3632	11.2869	10.6056	5.0055	0 10 20 30 40 Concentration (mM)			
8.1	11.3557	_*	10.5779	4.9919				
6.1	11.3448	_*	10.5237	4.9782				
4.6	11.3319	_*	10.4781	4.9575				
3.4	11.3265	_*	10.4155	4.9356				
2.6**	11.3102	_*	10.3624	4.9062				
1.9**	11.3044	_***	10.3233	4.8759				
1.4**	11.2979	_***	10.2948	4.8406				
0	-	-	-	4.6766				
					K <sub>DIM</sub> = 11,876 M <sup>-1</sup> sigma= 8.23*Obscured by another proton			

signal

\*\* Outlier not included in K<sub>Het</sub>

determination

\*\*\*Broadened signal

Table 8	Table 86: 11:2 Heterodimer <sup>1</sup> H NMR titration at 288 K							
[2]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2 Heterodimer 288K N=3			
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 → NH <sup>1</sup>			
34.3	11.3506	11.1715	10.6815	5.0357				
25.7	11.3421	11.1574	10.6519	5.0255				
19.3	11.3334	11.1499	10.6180	5.0141				
14.5	11.3239	11.1403	10.5829	5.0010	Ö			
10.9	11.3112	11.1427	10.5331	4.9846	0 10 20 30 40			
8.1	11.3012	11.1424	10.4903	4.9685	Concentration (min)			
6.1	11.2897	11.1515	10.4418	4.9443				
4.6	11.2782	_*	10.3895	4.9163				
3.4	11.2687	_*	10.3464	4.8893				
2.6	11.2599	_**	10.3040	4.8565				
1.9	11.2528	_**	10.2742	4.8206				
1.4	11.2469	_**	10.2511	4.7836				
0	-	-	-	4.6197				
				•	K <sub>DIM</sub> = 5675 M <sup>-1</sup> sigma= 13.5			
					*Broadened signal			
					**Obscured by another proton			
					signal			

Table 87: 11:2 Heterodimer <sup>1</sup> H NMR titration at 295 K							
[2]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2 Heterodimer 295K N=3		
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 → NH1		
34.3	11.3153	11.0736	10.6307	5.0224			
25.7	11.3061	11.0553	10.5991	5.0101			
19.3	11.2964	11.0429	10.5632	4.9959			
14.5	11.2857	11.0321	10.5258	4.9790	ů S		
10.9	11.2724	11.0220	10.4762	4.9598	0 10 20 30 40		
8.1	11.2615	11.0124	10.4332	4.9390	Concentration (mM)		
6.1	11.2491	11.0099	10.3854	4.9149			
4.6	11.2375	11.0165	10.3372	4.8827			
3.4	11.2271	11.0224	10.2948	4.8503			
2.6	11.2185	11.0412	10.2562	4.8133			
1.9*	11.2114	11.0487	10.2273	4.7775			
1.4*	11.2065	11.0712	10.2046	4.7393			
0	-	-	-	4.5186			
					K <sub>DIM</sub> = 4170 M <sup>-1</sup> sigma= 9.6		
					*Outlier not included in K <sub>Het</sub>		
					determination		

Table 88: 11:2 Heterodimer <sup>1</sup> H NMR titration at 298 K							
[2]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2 Heterodimer 298K N=3		
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	15 → NH <sup>1</sup>		
34.3	11.3003	11.0332	10.6086	5.0156			
25.7	11.2913	11.0148	10.5762	5.0027			
19.3	11.2799	10.9943	10.5380	4.9866			
14.5	11.2696	10.9759	10.5007	4.9689	C.		
10.9	11.2557	10.9668	10.4504	4.9395			
8.1	11.2433	10.9529	10.4062	4.9253	Concentration (mM)		
6.1	11.2302	10.9449	10.3580	4.8979			
4.6	11.2185	10.9462	10.3106	4.8677			
3.4	11.2077	10.9467	10.2688	4.8301			
2.6*	11.1989	10.9587	10.2309	4.7930			
1.9*	11.1913	10.9613	10.2016	4.7556			
1.4*	11.1869	10.9799	10.1794	4.7183			
0	-	-	-	4.5004			
					K <sub>DIM</sub> = 2552 M <sup>-1</sup> sigma= 7.9		
					*Outlier not included in K <sub>Het</sub>		

determination



determination

## 2.5.4. 11:2 Heterodimer VT <sup>1</sup>H NMR Summary and Van't Hoff Plots

Temp	K <sub>Het</sub>			11:2 Heterodimer Vant Hoff Plot
(К)	(M <sup>-1</sup> )	1/T	lnK	10 1/T vs InK <sub>Het</sub>
278	13,433	0.003597122	9.505469645	9- R <sup>2</sup> = 0.993
288	5530	0.003472222	8.617943095	
295	3182	0.003389831	8.065265209	
298	2095	0.003355705	7.647308832	7- •
308	1178	0.003246753	7.071573364	
				1/Г

Table 90: Summary of 11:2 Heterodimer Experiment 1 and Van't Hoff Plot

Table 91: Summary of 11:2 Heterodimer Experiment 2 and Van't Hoff Plot



Table 92: Summary of 11:2 Heterodimer Experiment 3 and Van't Hoff Plot

Temp	K <sub>Het</sub>			11:2 Heterodimer Vant Hoff Plot			
(К)	(M <sup>-1</sup> )	1/T	lnK	<sup>10</sup> 1/T vs InK <sub>Het</sub>			
278	11,876	0.003597122	9.382274836	R <sup>2</sup> = 0.970			
288	5 <i>,</i> 675	0.003472222	8.643825842	y y y y y y y y y y y y y y y y y y y			
295	4170	0.003389831	8.335671315				
298	2552	0.003355705	7.844632644				
308	1818	0.003246753	7.505492275	7			
				1/Т			

Table 93: Summary of thermodynamic parameters for 11:2 heterodimer

Expt	ΔH (kcal mol <sup>-1</sup> )	-T∆S <sup>295 K</sup> (kcal mol <sup>-1</sup> )	ΔG (kcal mol <sup>-1</sup> )
N1	-14.0	9.3	- 4.7
N2	-12.0	7.3	- 4.7
N3	-11.6	6.1	- 5.5
Average	-12.5	7.6	- 5.0

## 2.6. 11:2a Heterodimer VT <sup>1</sup>H NMR Titration Experiment



**Figure 7:** *Heterodimerisation of 2a and 11. Resonances used to determine binding constants are highlighted in red.* 

**N.B.**  $K_{Het}$  was calculated on using  $H^2$  only. Shifts of NH signals in **2a** were broad and therefore not included in  $K_{Het}$  calculation.

## **2.6.1. 11:2a Heterodimer** <sup>1</sup>**H NMR Titration Experiment 1** Peptide concentration: 3.1 mM

Table 9	Table 94: 11:2a Heterodimer <sup>1</sup> H NMR titration at 278 K							
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 278K			
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	<sup>5.5</sup> 7			
36.9	-	-	-	5.2081				
27.6	-	-	-	5.2011	₫ ₩ 5.0-			
20.7	-	-	-	5.1957				
15.5	-	-	-	5.1860	55 E 4.5			
11.7	-	-	-	5.1758				
8.7	-	-	-	5.1540	4.0			
6.6	-	-	-	5.1336				
4.9	-	-	-	5.1053	Conc (mM)			
3.7	-	-	-	5.0630				
2.8	-	-	-	5.0170				
2.1	-	-	-	4.9541				
1.6	-	-	-	4.9054				
0	-	-	-	4.6751				
					K <sub>Het</sub> = 5923 M <sup>-1</sup> sigma= 22.09			

Table 9	Table 95: 11:2a Heterodimer <sup>1</sup> H NMR titration at 288 K							
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 288K			
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.57			
36.9	-	-	-	5.1845	Ê			
27.6	-	-	-	5.1750	₩ 5.0-			
20.7	-	-	-	5.1638				
15.5	-	-	-	5.1502	5 E 4.5			
11.7	-	-	-	5.1294				
8.7	-	-	-	5.1060	4.0			
6.6	-	-	-	5.0740	0 10 20 30 40			
4.9	-	-	-	5.0236	Conc (mM)			
3.7	-	-	-	4.9808				
2.8	-	-	-	4.9262				
2.1	-	-	-	4.8732				
1.6	-	-	-	4.8221				
0	-	-	-	4.6221				
					K <sub>Het</sub> = 2357 M <sup>-1</sup> sigma= 16.5			

Table 9	Table 96: 11:2a Heterodimer <sup>1</sup> H NMR titration at 295 K							
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 295K			
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	<sup>5.5</sup> 7			
36.9	-	-	-	5.1609	Ê			
27.6	-	-	-	5.1479	₩ 5.0-			
20.7	-	-	-	5.1335				
15.5	-	-	-	5.1141	<u>5</u> 4.5-			
11.7	-	-	-	5.0901				
8.7	-	-	-	5.0570	4.0			
6.6	-	-	-	5.0160	0 10 20 30 40			
4.9	-	-	-	4.9654	Conc (mM)			
3.7	-	-	-	4.9115				
2.8	-	-	-	4.8575				
2.1	-	-	-	4.8050				
1.6	-	-	-	4.7594				
0	-	-	-	4.5339				
					K <sub>Het</sub> = 1504 M <sup>-1</sup> sigma= 19.0			

Table 9	Table 97: 11:2a Heterodimer <sup>1</sup> H NMR titration at 298 K								
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 298K				
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.57				
36.9	-	-	-	5.1493	Ê				
27.6	-	-	-	5.1362	≝ 5.0- ≝ 5.0-				
20.7	-	-	-	5.1186					
15.5	-	-	-	5.0966	Ŭ 4.5				
11.7	-	-	-	5.0684	C C C C C C C C C C C C C C C C C C C				
8.7	-	-	-	5.0317	4.0				
6.6	-	-	-	4.9870	0 10 20 30 40				
4.9	-	-	-	4.9358	Conc (mM)				
3.7	-	-	-	4.8804					
2.8	-	-	-	4.8269					
2.1	-	-	-	4.7747					
1.6	-	-	-	4.7340					
0	-	-	-	4.5138					
					K <sub>Het</sub> = 1099 M <sup>-1</sup> sigma= 16.9				

Table 98: 11:2a Heterodimer <sup>1</sup> H NMR titration at 308 K								
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δH <sup>2</sup>	11:2a Heterodimer 308K			
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	<sup>5.5</sup> 7			
36.9	-	-	-	5.1004	<b>F</b>			
27.6	-	-	-	5.0808	± 5.0-			
20.7	-	-	-	5.0549				
15.5	-	-	-	5.0223				
11.7	-	-	-	4.9831				
8.7	-	-	-	4.9344	4.0			
6.6	-	-	-	4.8831	0 10 20 30 40			
4.9	-	-	-	4.8267	Conc (mM)			
3.7	-	-	-	4.7706				
2.8	-	-	-	4.7191				
2.1	-	-	-	4.6724				
1.6	-	-	-	4.6317				
0	-	-	-	4.4614				
		·			K <sub>Het</sub> = 537 M <sup>-1</sup> sigma = 10.1			

## **2.6.2.** 11:2a Heterodimer <sup>1</sup>H NMR Titration Experiment 2 Peptide concentration: 3.1 mM

Table 99: 11:2a Heterodimer <sup>1</sup> H NMR titration at 278 K N=2							
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer	278K N=2	
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.57		
37.9	-	-	-	5.2013	Ê	_ <b></b>	
28.4	-	-	-	5.1948	<sup>a</sup> ≝ 5.0-		
21.3	-	-	-	5.1856	] it s		
16.0	-	-	-	5.1779	ор д. 4.5-		
12.0	-	-	-	5.1679			
9.0	-	-	-	5.1483	4.0		
6.7	-	-	-	5.1253	0 10 20	30 40	
5.1	-	-	-	5.0938	Conc (mM)		
3.8	-	-	-	5.0507			
2.8	-	-	-	4.9994			
2.1	-	-	-	4.9474			
0	-	-	-	4.6754			
					K <sub>Het</sub> = 4735 M <sup>-1</sup>	sigma= 18.0	

Table 100: 11:2a Heterodimer <sup>1</sup> H NMR titration at 288 K N=2								
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer	288K N=2		
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.5 <b>-</b> 7			
37.9	-	-	-	5.1768	Ê			
28.4	-	-	-	5.1677				
21.3	-	-	-	5.1571				
16.0	-	-	-	5.1453				
12.0	-	-	-	5.1265				
9.0	-	-	-	5.1012	4.0			
6.7	-	-	-	5.0672	0 10 20	30 40		
5.1	-	-	-	5.0255	Conc (mM)			
3.8	-	-	-	4.9740				
2.8	-	-	-	4.9222				
2.1	-	-	-	4.8690				
0	-	-	-	4.6082				
					K <sub>Het</sub> = 2329 M <sup>-1</sup>	sigma= 16.1		

Table 1	Table 101: 11:2a Heterodimer <sup>1</sup> H NMR titration at 295 K N=2								
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 295K N=2				
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.57				
37.9	-	-	-	5.1534					
28.4	-	-	-	5.1425	± 5.0-				
21.3	-	-	-	5.1277					
16.0	-	-	-	5.1100	5 4.5-				
12.0	-	-	-	5.0858					
9.0	-	-	-	5.0521	4.0				
6.7	-	-	-	5.0103	0 10 20 30 40				
5.1	-	-	-	4.9636	Conc (mM)				
3.8	-	-	-	4.9101					
2.8	-	-	-	4.8558					
2.1	-	-	-	4.8054					
0	-	-	-	4.5564					
					K <sub>Het</sub> = 1361 M <sup>-1</sup> sigma= 14.7				

Table 1	Table 102: 11:2a Heterodimer <sup>1</sup> H NMR titration at 298 K N=2							
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 298K N=2			
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.5			
37.9	-	-	-	5.1412				
28.4	-	-	-	5.1296		•		
21.3	-	-	-	5.1139				
16.0	-	-	-	5.0919	ο Ξ.Ε. 4.5-			
12.0	-	-	-	5.0653				
9.0	-	-	-	5.0291	4.0			
6.7	-	-	-	4.9852	<u>0 10 20</u>	30 40		
5.1	-	-	-	4.9350	Conc (mM)			
3.8	-	-	-	4.8803				
2.8	-	-	-	4.8273				
2.1	-	-	-	4.7750				
0	-	-	-	4.4646	]			
					K <sub>Het</sub> = 1240 M <sup>-1</sup> s	igma= 20.3		



2.6.3. 11:2a Heterodimer <sup>1</sup> H NM	R Titration Experiment 3
Peptide concentration: 3.3 mM	

Table 104: 11:2a Heterodimer <sup>1</sup> H NMR titration at 278 K N=3						
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 278K N=3	
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.5	
39.52	-	-	-	5.2035	Ê	
29.64	-	-	-	5.1996	tu 5.0-	
22.23	-	-	-	5.1894		
16.67	-	-	-	5.1846	55 E 4.5-	
12.50	-	-	-	5.1754		
9.37	-	-	-	5.1579	4.0	
7.03	-	-	-	_*	0 10 20 30 40 50	
5.28	-	-	-	5.1044	Conc (mM)	
3.96	-	-	-	_*		
2.96	-	-	-	5.0071		
2.2	-	-	-	4.9586		
1.67	-	-	-	4.9062		
0	-	-	-	4.6840		
					K <sub>Het</sub> = 5721 M <sup>-1</sup> sigma=21.6	
					*Broadened signal	

Table 105: 11:2a Heterodimer <sup>1</sup> H NMR titration at 288 K N=3								
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 288K N=3			
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.5			
39.52	-	-	-	5.1780	Ê			
29.64	-	-	-	5.1716	<sup>d</sup> ≝ 5.0			
22.23	-	-	-	5.1619				
16.67	-	-	-	5.1495	j⊑ 4.5−			
12.50	-	-	-	5.1322	Che			
9.37	-	-	-	5.1046	4.0			
7.03	-	-	-	5.0723	0 10 20 30	40 50		
5.28	-	-	-	5.0349	Conc (mM)			
3.96	-	-	-	4.9855				
2.96	-	-	-	4.9317				
2.2	-	-	-	4.8759				
1.67	-	-	-	4.8277				
0	-	-	-	4.6086				
					K <sub>Het</sub> = 2693 M <sup>-1</sup>	sigma=21.0		

Table 106: 11:2a Heterodimer <sup>1</sup> H NMR titration at 295 K N=3									
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 295K N=3				
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.6				
39.52	-	-	-	5.1546	Ê 5.4-				
29.64	-	-	-	5.1451	± 5.2-				
22.23	-	-	-	5.1317	5.0- <b>5.0-</b>				
16.67	-	-	-	5.1155					
12.50	-	-	-	5.0913	4.6-				
9.37	-	-	-	5.0637	4.4				
7.03	-	-	-	5.0186					
5.28	-	-	-	4.9727	Conc (mM)				
3.96	-	-	-	_*					
2.96	-	-	-	4.8642					
2.2	-	-	-	4.8193					
1.67	-	-	-	_*					
0				_*					
					K <sub>Het</sub> = 1055 M <sup>-1</sup> sigma= 1.9*Broadened signal				

Table 107: 11:2a Heterodimer <sup>1</sup> H NMR titration at 298 K N=3										
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 298K N=3					
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.67					
39.52	-	-	-	5.1438	Ê 5.4-					
29.64	-	-	-	5.1334	₫ # 5.2-					
22.23	-	-	-	5.1176	5.0-					
16.67	-	-	-	5.0984						
12.50	-	-	-	5.0718	9 5 4.6-					
9.37	-	-	-	5.0397						
7.03	-	-	-	4.9930						
5.28	-	-	-	4.9416	Conc (mM)					
3.96	-	-	-	4.8886						
2.96	-	-	-	4.8348						
2.2	-	-	-	4.7885						
1.67	-	-	-	_*						
0				_*						
					K <sub>Het</sub> = 863 M <sup>-1</sup> sigma= 1.57*Broadened signal					

Table 108: 11:2a Heterodimer <sup>1</sup> H NMR titration at 308 K N=3						
[2a]	δ NH <sup>1</sup>	δ NH <sup>2</sup>	δ NH <sup>3</sup>	δ H <sup>2</sup>	11:2a Heterodimer 308K N=3	
(mM)	(ppm)	(ppm)	(ppm)	(ppm)	5.5	
39.52	-	-	-	5.0966		
29.64	-	-	-	5.0807		
22.23	-	-	-	5.0548		
16.67	-	-	-	5.0261	<u> </u>	
12.50	-	-	-	4.9880	Che	
9.37	-	-	-	4.9482	4.0	
7.03	-	-	-	4.8893		
5.28	-	-	-	4.8372	Conc (mM)	
3.96	-	-	-	4.7793		
2.96	-	-	-	4.7282		
2.2	-	-	-	4.6837		
1.67	-	-	-	4.6404		
0				4.4666		
					K <sub>Het</sub> = 574 M <sup>-1</sup> sigma= 11.7	

## 2.6.4. 11:2a Heterodimer VT <sup>1</sup>H NMR Summary and Van't Hoff Plots

Temp	K <sub>Het</sub>			11:2a Heterodimer Vant Hoff Plot				lot
(K)	(M <sup>-1</sup> )	1/T	InK	10		1/T vs InK <sub>t</sub>	let	
278	5,923	0.003597122	8.686598356	9-			مر	R <sup>2</sup> = 0.998
288	2357	0.003472222	7.765144903	-8 <sub>분</sub>				
295	1504	0.003389831	7.315883505	<u>کے</u> 1-		And and a second		
298	1099	0.003355705	7.002155954	6-	/			
308	537	0.003246753	6.285998095	5-				
				0.00	32	0.0034	0.0036	
						1/T		

 Table 109: Summary of 11:2a Heterodimer Experiment 1 and Van't Hoff Plot

 Table 110: Summary of 11:2a Heterodimer Experiment 2 and Van't Hoff Plot



Table 111: Summary of 11:2a Heterodimer Experiment 3 and Van't Hoff Plot

Temp	K <sub>Het</sub>			11:2a Heterodimer Vant Hoff Plot
(K)	(M <sup>-1</sup> )	1/T	lnK	1/T vs InK <sub>Het</sub>
278	5,721	0.003597122	8.651898894	9 <b>-</b>
288	2693	0.003472222	7.898411093	R <sup>-</sup> = 0.996
295	1055	0.003389831	6.961296046	Yu 7
298	863	0.003355705	6.760414691	
308	574	0.003246753	6.352629396	5
				0.0032 0.0034 0.0036
				1/T

Table 112: Summary of thermodynamic parameters for 11:2 heterodimer

Expt	ΔH (kcal mol <sup>-1</sup> )	-TΔS <sup>295 K</sup> (kcal mol⁻¹)	ΔG (kcal mol <sup>-1</sup> )
N1	-13.5	9.3	- 4.2
N2	-11.9	7.7	- 4.2
N3	-13.9	9.7	- 4.2
Average	-13.1	8.9	- 4.2
# 2.7. ROESY Study of 2:11 Heterodimer







## References

- (1) S. Vedachalam, J. Zeng, B. K. Gorityala, M. Antonio and X.Liu, *Org. Lett.*, 2010, **12**, 352.
- (2) J. E. Thomson, C. D. Campbell, C. Concellon, N. Duguet, K. Rix, A. M. Z. Slawin, and A. D. Smith, *J. Org. Chem.*, 2008, **73**, 2784.
- (3) J. S. Nowick, M.D. Chung, K. Maitra, S. Mairita, K. D. Stigers and Y. Sun, J. Am. Chem. Soc., 2000, **122**, 7654.
- (4) D. L. Holmes, E. M. Smith and J. S. Nowick, J. Am. Chem. Soc., 1997, **119**, 7665.



## <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)

#### N-(6-nitro-4-oxo-4H-chromen-3-yl)butyramide (6)





### <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)

#### *N*-(6-amino-4-oxo-4*H*-chromen-3-yl)butyramide (7)





N-(4-oxo-6-(2-oxo-2-(piperidin-1-yl)acetamido)-

#### <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)





#### N-(3-(2-isobutyrylhydrazinecarbonyl)-4-

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)





#### *N*-(3-(2-isobutyrylhydrazine-1-carbonyl)-methoxyphenyl)butyramide (2a)

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)



#### *N*-(3-(2-isobutyrylhydrazine-1-carbonyl)-methoxyphenyl)butyramide (2a)

#### <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)



#### <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)

(S)-N-((S)-1-(butylamino)-3-methyl-1-oxobutan-2-yl)-2-((S)-2-isobutyramido-3phenylpropanamido)-4-methylpentanamide



#### <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>)

### (S)-N-((S)-1-(butylamino)-3-methyl-1-oxobutan-2-yl)-2-((S)-2-isobutyramido-3phenylpropanamido)-4-methylpentanamide

