

Supplementary Information

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1. Experimental

1.1. General Experimental

All anhydrous solvents and reagents were obtained from commercial suppliers (Aldrich, Alfa Aesar, Fluorochem) and used without further purification. Reactions were performed in oven-dried, round-bottom flasks fitted with rubber septa under an atmosphere of argon unless otherwise stated. Analytical thin layer chromatography (TLC) was performed on pre-coated aluminium sheets of silica (60 F₂₅₄, Merck) and visualised by short-wave UV light.

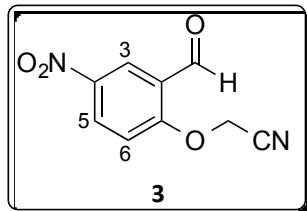
¹H NMR spectra were recorded at 500 MHz on a Bruker Avance-500 using an internal deuterium lock. Chemical shifts were measured in parts per million (ppm) relative to tetramethylsilane (δ = 0) using the following internal references for residual protons in the solvent : CDCl₃ (δ 7.26) and (CD₃)₂SO (δ 2.50). Data is presented as follows: chemical shift, multiplicity, coupling constant (*J*) in Hz, integration and assignment.

¹³C NMR spectra were recorded at 126 MHz on a Bruker Avance-500 using an internal deuterium lock. Chemical shifts were measured in parts per million (ppm) relative to tetramethylsilane (δ = 0) using the following internal references for residual protons in the solvent: CDCl₃ (δ 77.0) and (CD₃)₂SO (δ 39.5). Data is presented as follows: chemical shift and assignment.

HRMS analysis was performed on an Agilent 1200 series HPLC and diode array detector coupled to a 6520 Quadrupole-Time of flight mass spectrometer with dual multimode APCI/ESI source. Analytical separation was carried out at 30 °C in a Merck Purospher STAR column (RP-18e, 30 x 4 mm) using a flow rate of 1.5 mL/min in a 4 min gradient elution with detection at 254 nm. The mobile phase was a mixture of methanol (solvent A) and water (solvent B) both containing formic acid at 0.1%. Gradient elution was as follows: 1:9 (A/B) to 9:1 (A/B) over 2.5 min, 9:1 A/B for 1 min, and then reversion back to 1:9 (A/B) over 0.3 min, finally 1:9 (A/B) for 0.2 min. The following reference masses were used for HRMS analysis: caffeine [M+H]⁺ 195.087652; (hexakis(1H,1H,3H-tetrafluoropentoxy)phosphazene) [M+H]⁺ 922.009798 and hexakis(2,2-difluoroethoxy)phosphazene [M+H]⁺ 622.02896 or reserpine [M+H]⁺ 609.280657.

1.2. Experimental Procedures

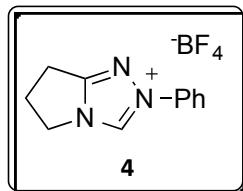
Synthesis of 2-(2-formyl-4-nitrophenoxy)acetonitrile (**3**)¹



To a solution of 2-hydroxy-5-nitrobenzaldehyde (0.50 g, 3.0 mmol) in DMF (2.5 mL) was added K₂CO₃ (0.60 g, 4.4 mmol). The resulting suspension was stirred for 15 min before the addition of bromoacetonitrile (0.40 mL, 5.8 mmol). The reaction was allowed to stir at room temperature for a period of 18 h. The reaction mixture was diluted with ethyl acetate and washed with water and brine. The organic layer was separated, dried over magnesium sulphate, and concentrated under vacuum. The crude material was purified by column chromatography (50% ethyl acetate in cyclohexane) to afford **3** (0.50 mg, 82%) as a pale yellow solid.

¹H NMR (500 MHz, CDCl₃) δ 10.46 (s, 1H, HC=O), 8.79 (d, J = 2.9 Hz, 1H, H³), 8.55 (dd, J = 9.1, 2.9 Hz, 1H, H⁵), 7.25 (d, J = 9.1 Hz, 1H, H⁶), 5.08 (s, 2H, OCH₂CN); ¹³C NMR (126 MHz, CDCl₃) δ 186.1 (C_q), 161.5 (C_q) 130.6 (CH), 125.6 (CH), 125.3 (C_q), 113.2 (C_q), 112.8 (CH), 54.0 (CH₂). N.B. Mass Spec. data could not be obtained. N.B. One C_q could not be resolved.

Synthesis of 2-phenyl-6,7-dihydro-5*H*-pyrrolo[2,1-*c*][1,2,4]triazol-2-ium tetrafluoroborate (**4**)²

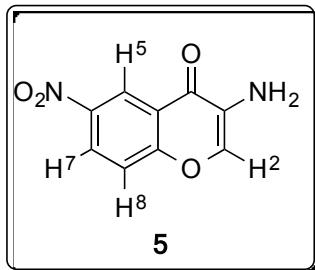


Trimethyloxonium tetrafluoroborate (0.96 g, 6.4 mmol) was added in one portion to a solution of pyrrolidin-2-one (0.50 g, 6.0 mmol) in DCM (20 mL). The reaction was allowed to stir at room temperature for 24 h. Phenylhydrazine (0.63 g, 5.8 mmol) was added and stirring was continued at room temperature for a further 48 h. The reaction mixture was then concentrated under vacuum and re-dissolved in a mixture

of methanol (2.0 mL) and triethyl orthoformate (8.0 ml). The solution was heated to reflux overnight and then allowed to cool to room temperature. The resulting precipitate was collected by filtration and washed with methanol to afford **4** (1.1 g, 70%) as a pale tan solid.

¹H NMR (500 MHz, DMSO-d₆) δ 10.70 (s, 1H, NCHNPh), 7.90 – 7.84 (m, 2H, ArCH), 7.72 – 7.66 (m, 2H, ArCH), 7.66 – 7.60 (m, 1H, ArCH), 4.66 – 4.13 (m, 2H, NCH₂), 3.23 – 3.12 (m, 2H, CH₂), 2.89 – 2.61 (m, 2H, CH₂); ¹³C NMR (126 MHz, d₆ DMSO) δ 163.5 (C_q), 138.8 (CH), 136.1 (C_q), 130.9 (CH), 130.7 (CH), 121.1 (CH), 47.4 (CH₂), 27.1 (CH₂), 21.7 (CH₂). NB. Mass spec. data could not be obtained.

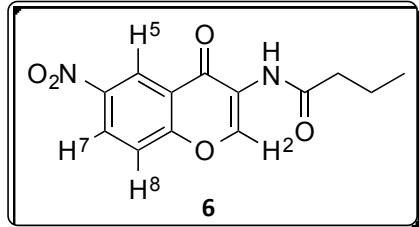
Synthesis of 3-amino-6-nitro-4*H*-chromen-4-one (**5**)¹



Compound **3** (0.47 g, 2.3 mmol) was dissolved in DCM (20 mL) before addition of catalyst **4**² (62 mg, 0.20 mmol) and DBU (30 μL, 0.20 mmol). The resulting solution was allowed to stir at room temperature for 16 h. The reaction was then concentrated under vacuum. The crude material was purified by column chromatography (40-60% ethyl acetate in cyclohexane) to afford **5** (0.27 g, 57%) as an orange solid.

¹H NMR (500 MHz, DMSO-d₆) δ 8.82 (d, *J* = 2.9 Hz, 1H, H⁵), 8.46 (dd, *J* = 9.3, 2.9 Hz, 1H, H⁷), 8.06 (s, 1H, H²), 7.84 (d, *J* = 9.3 Hz, 1H, H⁸), 4.85 (s, 2H, NH₂); ¹³C NMR (126 MHz, DMSO-d₆) δ 171.8 (C_q), 158.4 (C_q), 143.8 (C_q), 137.3 (CH), 133.7 (C_q), 127.0 (CH), 121.9 (CH), 121.3 (C_q), 121.1 (CH); HRMS (ESI) [M+H]⁺ 207.0403, C₉H₆N₂O₄ [M+H]⁺ requires 207.0406.

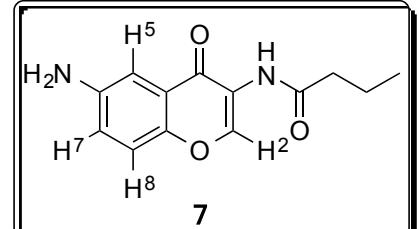
Synthesis of *N*-(6-nitro-4-oxo-4*H*-chromen-3-yl)butyramide (**6**)



Compound **5** (0.17 g, 0.83 mmol) and DIPEA (0.22 mL, 1.2 mmol) were dissolved in THF (10 mL) and cooled to 0 °C before the addition of butyryl chloride (0.10 mL, 0.90 mmol). The reaction was allowed to warm to room temperature and stirred for 1 h. The reaction mixture was then diluted with ethyl acetate and washed with sat. NaHCO₃ (aq.) and brine. The organic layer was separated, dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (50% ethyl acetate in cyclohexane) to afford **6** (0.18 g, 80%) as an orange solid.

¹H NMR (500 MHz, CDCl₃) δ 9.52 (s, 1H, H²), 9.17 (d, *J* = 2.7 Hz, 1H, H⁵), 8.54 (dd, *J* = 9.2, 2.7 Hz, 1H, H⁷), 8.01 (s, 1H, NH), 7.69 (d, *J* = 9.2 Hz, 1H, H⁸), 2.56 – 2.41 (m, 2H, CH₂CH₂CH₃), 1.88 – 1.72 (m, 2H, CH₂CH₂CH₃), 1.04 (t, *J* = 7.4 Hz, 3H, CH₂CH₂CH₃); ¹³C NMR (126 MHz, CDCl₃) δ 171.9 (C_q), 170.9 (C_q), 158.2 (C_q), 145.0 (CH), 128.0 (CH), 125.1 (C_q) 122.8 (CH), 121.7 (C_q), 120.3 (CH), 39.0 (CH₂), 18.8 (CH₂), 13.6 (CH₃); HRMS (ESI) [M+H]⁺ 277.0818 C₁₃H₁₂N₂O₃ [M+H]⁺ requires 277.0819; N.B. One C_q not resolved

Synthesis of *N*-(6-amino-4-oxo-4*H*-chromen-3-yl)butyramide (**7**)

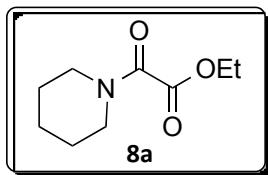


Compound **6** (0.18 g, 0.70 mmol), iron powder (0.26 g, 4.6 mmol) and ammonium chloride (0.25 g, 4.6 mmol) were heated to reflux in a mixture of ethanol and water

(3:1) (20 mL) for 16 h. The reaction mixture was diluted with ethyl acetate and filtered through celite and then washed with sat. NaHCO_3 (aq.) and brine. The organic layer was separated, dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (35-60% ethyl acetate in DCM) to afford **7** (0.12 g, 76%) as a yellow solid.

^1H NMR (500 MHz, CDCl_3) δ 9.38 (s, 1H, H^2), 8.05 (s, 1H, NH), 7.41 (d, J = 2.9 Hz, 1H, H^5), 7.37 (d, J = 9.0 Hz, 1H, H^8), 7.08 (dd, J = 9.0, 2.9 Hz, 1H, H^7), 3.89 (s, 2H, NH_2), 2.61 – 2.27 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_3$), 1.85 – 1.70 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_3$), 1.03 (t, J = 7.4 Hz, 3H, $\text{CH}_2\text{CH}_2\text{CH}_3$); ^{13}C NMR (126 MHz, CDCl_3) δ 171.6 (C_q), 171.5 (C_q), 149.7 (C_q), 144.8 (CH), 143.5 (C_q), 123.7 (C_q), 122.9 (CH), 122.6 (C_q), 119.5 (CH), 107.2 (CH), 39.1 (CH₂), 18.9 (CH₂), 13.7 (CH₃); HRMS (ESI) [M+H]⁺ 247.1077, $\text{C}_{13}\text{H}_{14}\text{N}_2\text{O}_3$ [M+H]⁺ requires 247.1077.

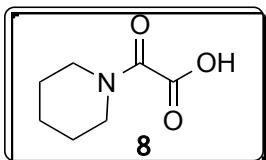
Synthesis of ethyl 2-oxo-2-(piperidin-1-yl)acetate (**8a**)



Piperidine (1.2 mL, 11.7 mmol) and DIPEA (3.1 mL, 17.6 mmol) were dissolved in DCM (15 mL) and cooled to 0 °C before the addition of ethyl 2-chloro-2-oxoacetate (1.6 mL, 14.1 mmol). The reaction was allowed to warm to room temperature and stirred for 2 h. The reaction was then diluted with DCM and washed with sat. NaHCO_3 (aq.) and brine. The organic layer was separated, dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (10-50% ethyl acetate in cyclohexane) to afford compound **8a** (1.9 g, 85 %) as a colourless oil.

^1H NMR (500 MHz, CDCl_3) δ 4.34 (q, J = 7.2 Hz, 2H, OCH_2CH_3), 3.60 – 3.54 (m, 2H, NCH_2), 3.37 – 3.32 (m, 2H, NCH_2), 1.67-1.73 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 1.59-1.66 (m, 4H, $\text{CH}_2\text{CH}_2\text{CH}_2$), 1.37 (t, J = 7.2 Hz, 3H, OCH_2CH_3); ^{13}C NMR (126 MHz, CDCl_3) δ 163.2 (C_q), 160.2 (C_q), 61.8 (CH₂), 47.2 (CH₂), 42.17 (CH₂), 26.1 (CH₂), 25.1 (CH₂), 24.3 (CH₂), 14.0 (CH₃); HRMS (ESI) [M+Na]⁺ 208.0942, $\text{C}_9\text{H}_{15}\text{NO}_3$ [M+Na]⁺ requires 208.0944.

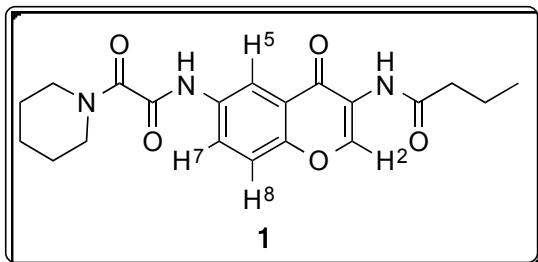
Synthesis of 2-oxo-2-(piperidin-1-yl)acetic acid (**8**)



Sodium hydroxide 0.45 M (16.0 mL, 11.0 mmol) was cooled to 0 °C before the addition of compound **8a** (1.9 g, 10.0 mmol) in THF (40 mL). The reaction was allowed to stir overnight. The reaction mixture was acidified to pH 1 with 1 M HCl (aq.) and extracted with ethyl acetate (x 3). The combined organic extracts were dried over sodium sulphate and concentrated to afford compound **8** (1.5 g, 96 %) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 4.13–4.08 (m, 2H, NCH₂), 3.69–3.64 (m, 2H, NCH₂), 1.81 – 1.42 (m, 6H, CH₂CH₂CH₂CH₂CH₂); ¹³C NMR (126 MHz, CDCl₃) δ 161.4 (C_q), 159.4 (C_q), 47.9 (CH₂), 45.2 (CH₂), 26.6 (CH₂), 25.6 (CH₂), 24.1 (CH₂); HRMS (ESI) [M+H]⁺ 158.0812, C₇H₁₁NO₃ [M+H]⁺ requires 158.0812.

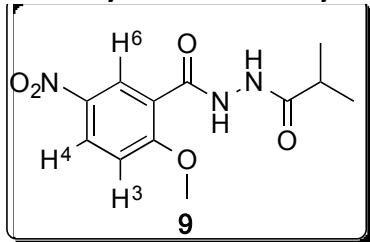
Synthesis of *N*-(4-oxo-6-(2-oxo-2-(piperidin-1-yl)acetamido)-4*H*-chromen-3-yl)butyramide (**1**)



Compound **8** (30 mg, 0.20 mmol) and DIPEA (80 μL, 0.50 mmol) were dissolved in DMF (1.5 mL) before the addition of HATU (94 mg, 0.30 mmol) and **7** (42 mg, 0.20 mmol). After a period of 16 h the reaction mixture was diluted with ethyl acetate and washed with sat. NaHCO₃ (aq.) and brine. The organic layer was separated, dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (50–80% ethyl acetate in cyclohexane) to afford **1** (48 mg, 65%) as a cream solid.

¹H NMR (500 MHz, CDCl₃) δ 9.97 (s, 1H, NH), 9.50 (s, 1H, H²), 8.51 (s, 1H, NH), 8.46 (d, J = 2.6 Hz, 1H, H⁵), 8.28 (dd, J = 9.2, 2.6 Hz, 1H, H⁷), 7.55 (d, J = 9.2 Hz, 1H, H⁸), 4.13 - 4.08 (m, 2H, NCH₂), 3.74 - 3.53 (m, 2H, NCH₂), 2.50 (t, J = 7.5 Hz, 2H, CH₂CH₂CH₃), 1.82 – 1.75 (m, 2H, CH₂CH₂CH₃), 1.74 – 1.69 (m, 6H, CH₂CH₂CH₂CH₂CH₂), 1.02 (t, J = 7.4 Hz, 3H, CH₂CH₂CH₃); ¹³C NMR (126 MHz, CDCl₃) δ 172.2 (C_q), 171.6 (C_q), 160.3 (C_q), 159.3 (C_q), 152.6 (C_q), 145.5 (CH), 134.4 (C_q), 126.6 (CH), 124.5 (C_q), 122.2 (C_q), 119.3 (CH), 115.7 (CH), 47.7 (CH₂), 45.0 (CH₂), 38.8 (CH₂), 26.8 (CH₂), 25.8 (CH₂), 24.4 (CH₂), 18.9 (CH₂), 13.7 (CH₃); HRMS (ESI) [M+H]⁺ 386.1714, C₂₀H₂₃N₃O₅ [M+H]⁺ requires 386.1710.

Synthesis of *N*-isobutyryl-2-methoxy-5-nitrobenzohydrazide (**9**)³

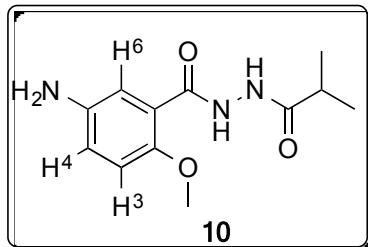


Thionyl chloride (1.2 mL), 2-methoxy-5-nitrobenzoic acid (0.20 g, 1.0 mmol) and DMF (10 μL) were heated to reflux for 2 h. The reaction mixture was concentrated to give a yellow solid. The crude acid chloride was dissolved in DCM (10 mL) and cooled to 0 °C before the addition of isobutyryl hydrazine (0.11 g, 1.1 mmol) and triethylamine (0.17 mL, 1.2 mmol) in DCM (10 mL). The reaction was allowed to warm to room temperature and stir for 2 h. The reaction mixture was then poured into sat. NaHCO₃ (aq.), the organic layer was separated and the aqueous extracted with DCM (x 3). The combined organic extracts were dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (50-100% ethyl acetate in DCM) to isolate compound **9** (0.26 g, 90% yield) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 10.82 (d, J = 7.3 Hz, 1H, NH), 9.85 (d, J = 7.4 Hz, 1H, NH), 9.07 (d, J = 2.9 Hz, 1H, H⁶), 8.39 (dd, J = 9.1, 3.0 Hz, 1H, H⁴), 7.15 (d, J = 9.2 Hz, 1H, H³), 4.19 (s, 3H, OCH₃), 2.72 (hept., J = 6.9 Hz, 1H, CH), 1.28 (d, J = 6.9 Hz, 6H, 2 x CH₃); ¹³C NMR (126 MHz, CDCl₃) δ 172.5 (C_q), 161.5 (C_q), 157.4 (C_q), 142.0 (C_q), 128.7

(CH), 128.0 (CH), 119.8 (C_q), 112.0 (CH), 57.4 (CH₃), 33.4 (CH), 19.3 (2 x CH₃); HRMS (ESI) [M+H]⁺ 282.1079, C₁₂H₁₅N₃O₅ [M+H]⁺ requires 282.1084.

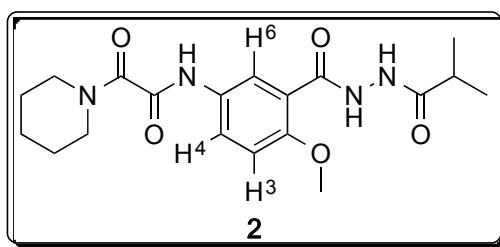
Synthesis of 5-amino-N-isobutyryl-2-methoxybenzohydrazide (**10**)³



Compound **10** (0.10 g, 0.40 mmol) was dissolved in THF (10 mL) and the reaction flask was purged by vacuum/argon cycles (x 3) before the addition of palladium 10% on carbon (10 mg). The reaction vessel was placed under an atmosphere of hydrogen (40 psi) and stirred for 2 h at room temperature. The reaction mixture was filtered through celite and concentrated under vacuum. The crude material was purified by column chromatography (0-10% EtOH in DCM) to isolate compound **10** (85 mg, 95 % yield) as a yellow solid.

¹H NMR (500 MHz, CDCl₃) δ 10.94 (d, *J* = 7.4 Hz, 1H, NH), 9.54 (d, *J* = 7.5 Hz, 1H, NH), 7.51 (dd, *J* = 2.4, 1.0 Hz, 1H, H⁴), 6.83 (m, 2H, H³+H⁶), 3.96 (s, 3H, OCH₃), 2.59 (hept., *J* = 6.9 Hz, 1H, CH), 1.23 (d, *J* = 6.9 Hz, 6H, 2xCH₃); ¹³C NMR (126 MHz, CDCl₃) δ 172.1 (C_q), 160.2 (C_q), 150.8 (C_q), 140.4 (C_q), 120.1 (CH), 119.1 (C_q), 117.9 (CH), 112.8 (CH), 56.5 (CH₃), 33.4 (CH), 19.4 (2 x CH₃); HRMS (ESI) [M+H]⁺ 252.1341, C₁₂H₁₇N₃O₃ [M+H]⁺ requires 252.1343.

Synthesis of *N*-(3-(2-isobutyrylhydrazinecarbonyl)-4-methoxyphenyl)-2-oxo-2-(piperidin-1-yl)acetamide (**2**)

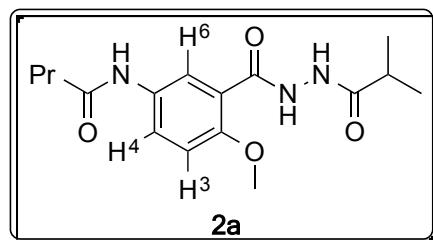


Compound **8** (0.2 g, 1.2 mmol), thionyl chloride (4.5 ml) and DMF (10 μL) were heated to reflux for 2 h. The reaction mixture was concentrated to give a yellow

solid. The crude acid chloride was dissolved in DCM (5.0 mL) and cooled to 0 °C before compound **10** (0.24 mg, 1.0 mmol) and triethylamine (0.16 mL, 1.2 mmol) in DCM (5.0 mL) were added. The reaction was allowed to warm to room temperature and stir for 1 h before it was poured into sat. NaHCO₃ (aq.). The organic layer was separated and the aqueous layer was extracted with DCM (x 3). The combined organic extracts were dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (50-100% ethyl acetate in DCM) to afford compound **2** (0.36 g, 97 % yield) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 11.34 (d, *J* = 8.4 Hz, 1H, NH), 11.07 (d, *J* = 8.3 Hz, 1H, NH), 10.71 (s, 1H, NH), 8.41 (d, *J* = 2.8 Hz, 1H, H⁶), 8.37 (dd, *J* = 8.9, 2.7 Hz, 1H, H⁴), 7.01 (d, *J* = 9.0 Hz, 1H, H³), 4.05 (s, 3H, OCH₃) 3.84 (t, *J* = 4.9 Hz, 2H, NCH₂), 3.63 (t, *J* = 5.3 Hz, 2H, NCH₂), 2.84 (hept., *J* = 6.8 Hz, 1H, CH), 1.72-1.62 (m, 6H, CH₂CH₂CH₂), 1.22 (d, *J* = 6.9 Hz, 6H, 2 x CH₃); ¹³C NMR (126 MHz, CDCl₃) δ 172.0 (C_q), 162.5 (C_q), 160.2 (C_q), 158.0 (C_q), 154.3 (C_q), 131.6 (C_q), 125.9 (CH), 124.4 (CH), 118.9 (C_q), 111.7 (CH), 56.6 (CH₃), 47.8 (CH₂), 44.2 (CH₂), 32.6 (CH), 26.6 (CH₂), 25.7 (CH₂), 24.4 (CH₂), 19.6 (2 x CH₃); HRMS (ESI) [M+H]⁺ 391.1964, C₁₉H₂₆N₄O₅ [M+H]⁺ requires 391.1976.

Synthesis of *N*-(3-(2-isobutyrylhydrazine-1-carbonyl)-methoxyphenyl)butyramide (2a)

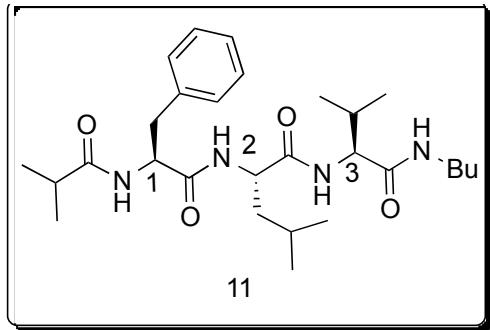


Compound **10** (0.5 g, 2.0 mmol) and pyridine (0.24 mL, 3.0 mmol) were dissolved in DCM (10 mL) and cooled to 0 °C before the addition of butyric anhydride (0.36 mL, 2.2 mmol). The reaction was allowed to warm to room temperature and stirred for 1 h. The reaction mixture was then diluted with DCM and washed with sat. NaHCO₃ (aq.) and brine. The organic layer was separated, dried over sodium sulphate and concentrated under vacuum. The crude material was purified by column chromatography (0-10% EtOH in DCM) to afford **2a** (0.53 g, 83 % yield) as a white

solid.

¹H NMR (500 MHz, CDCl₃) δ 10.41 (s, 1H, NH), 9.45 (s, 1H, NH), 8.09 (dd, *J* = 9.0, 2.8 Hz, 1H, H⁴), 7.83 (d, *J* = 2.8 Hz, 1H, H⁶), 6.89 (d, *J* = 9.0 Hz, 1H, H³), 3.97 (s, 3H, OCH₃), 2.63 (app. quint., *J* = 6.9 Hz, 1H, CH), 2.10 (s, 2H, CH₂), 1.60 (q, *J* = 8.2, 7.3 Hz, 2H, CH₂), 1.21 (d, *J* = 6.9 Hz, 6H, 2 x CH₃), 0.78 (s, 3H, CH₃); ¹³C NMR (126 MHz, CDCl₃) δ 173.5 (C_q), 172.7 (C_q), 160.5 (C_q), 153.6 (C_q), 132.5 (C_q), 126.1 (CH), 123.4 (CH), 118.5 (C_q), 111.8 (CH), 56.5 (CH₃), 38.9 (CH₂), 33.2 (CH), 19.4 (2 x CH₃), 18.9 (CH₂), 13.6 (CH₃); HRMS (ESI) [M+H]⁺ 322.1757, C₁₆H₂₃N₃O₄ [M+H]⁺ requires 391.1761.

Synthesis of (S)-N-((S)-1-(butylamino)-3-methyl-1-oxobutan-2-yl)-2-((S)-2-isobutyramido-3-phenylpropanamido)-4-methylpentanamide (11)⁴



Compound **11** was synthesized via solid phase peptide synthesis using an Activotec P14 peptide synthesizer. Fmoc-Pal-Am Resin (2.5 g, 0.53 mmol/g) was shaken with DMF (2 x 25 ml) for 5 mins. The resin was then shaken with a 20% piperidine solution in NMP (3 x 25 ml) for 12 minutes. The resin was then washed (2 x DMF, 2 x DCM, 2 x MeOH, 2 x DCM and MeOH alternately and then diethyl ether).

- To the resin was added DCM (25 mL), collidine (1.0 mL, 8.0 mmol) and 2-nitrobenzene-1-sulfonyl chloride (0.88 g, 4.0 mmol). The resulting suspension was agitated for 3 h. The resin was drained and washed as described above.
- To the resin was added DMF (25 mL), 7-methyl-1,5,7-triazabicyclo[4.4.0]dec-5-ene (MTBD) (0.80 mL, 5.3 mmol) and 1-iodobutane (1.5 mL, 13.3 mmol). The resulting suspension agitated for a period of 8 h after which the resin was drained and washed as before.
- To the resin was added DMF (25 mL), DBU (1.0 mL, 6.6 mmol) and

mercaptoethanol (0.90 mL, 13.3 mmol). The resin was agitated for 3 h and then drained. The process was repeated and then resin was washed as before.

- Fmoc-Val-OH (4.5 g, 13.3 mmol) was dissolved in a 10% solution of DIPEA in DMF (25 mL) before the addition of HATU (4.5 g, 12.0 mmol). The reaction mixture was allowed to stir for 15 mins after which time it was added to the resin and agitated for 2 h. The process was repeated and the resin was washed as before.
- The resin was then shaken with a 20% piperidine solution in NMP (3 x 25 ml) for 12 minutes. The resin was then washed (2 x DMF, 2 x DCM, 2 x MeOH, 2 x DCM and MeOH alternately and then diethyl ether).
- Fmoc-Leu-OH (2.1 g, 5.3 mmol) was dissolved in a 10% solution of DIPEA in DMF (25 mL) before the addition of HATU (1.8 g, 4.8 mmol). The reaction mixture was allowed to stir for 15 mins after which time it was added to the resin and agitated for 2 h. The process was repeated and the resin was washed as before.
- The resin was then shaken with a 20% piperidine solution in NMP (3 x 25 ml) for 12 minutes. The resin was then washed (2 x DMF, 2 x DCM, 2 x MeOH, 2 x DCM and MeOH alternately and then diethyl ether).
- Fmoc-Phe-OH (1.9 g, 5.3 mmol) was dissolved in a 10% solution of DIPEA in DMF (25 mL) before the addition of HATU (1.8 g, 4.8 mmol). The reaction mixture was allowed to stir for 15 min after which time it was added to the resin and agitated for 2 h. The process was repeated and the resin was washed as before.
- The resin was then shaken with a 20% piperidine solution in NMP (3 x 25 ml) for 12 minutes. The resin was then washed (2 x DMF, 2 x DCM, 2 x MeOH, 2 x DCM and MeOH alternately and then diethyl ether).
- To the resin was added a 10% solution of DIPEA in DMF (25 mL) and isobutyryl chloride (0.60 ml, 5.3 mmol). The suspension was agitated for 2 h before the resin was drained and washed as before.
- The resin was suspended in a 1:1 mixture of TFA and DCM (20 mL) for 1 h

without agitation. The resin was removed by filtration and the filtrate was concentrated under vacuum. The crude material was purified by HPLC to afford **11** (0.17 g, 31 %) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 8.21 (s, 1H, NH), 7.62 (s, 2H, 2 x NH), 7.24 – 7.02 (m, 6H, ArCH, NH), 5.19 (app. q, *J* = 7.0 Hz, 1H, H¹), 4.82 (app. q, *J* = 7.8 Hz, 1H, H²), 4.50 (app. t, *J* = 8.9 Hz, 1H, H³), 3.34 – 3.26 (m, 1H, NCH), 3.20 – 3.12 (m, 1H, NCH), 3.07 – 2.98 (m, 2H, PhCH₂), 2.48 (h, *J* = 6.8 Hz, 1H, HC(CH₃)₂), 2.13 – 2.05 (m, 1H, HC(CH₃)₂), 1.70 – 1.54 (m, 3H, CH₂CH(CH₃)₂, CH₂CH(CH₃)₂), 1.54 – 1.46 (m, 2H, CH₃CH₂CH₂CH₂NH), 1.34 (app. h, *J* = 7.3 Hz, 2H, CH₃CH₂CH₂CH₂NH), 1.06 (d, *J* = 7.0 Hz, 3H, CH₃), 1.04 (d, *J* = 7.1 Hz, 3H, CH₃), 0.97 (d, *J* = 6.7 Hz, 3H, CH₃), 0.94 (d, *J* = 6.8 Hz, 3H, CH₃), 0.91 (t, *J* = 7.4 Hz, 3H, CH₃CH₂CH₂CH₂NH), 0.87 (t, *J* = 6.3 Hz, 6H, 2 x CH₃); ¹³C NMR (126 MHz, CDCl₃) δ 176.8 (C_q), 172.0 (C_q), 171.1 (C_q), 136.6 (C_q), 129.5 (ArCH), 128.1 (ArCH), 126.6 (ArCH), 58.8 (CH), 53.2 (CH), 51.6 (CH), 42.5 (CH₂), 39.1 (CH₂), 38.9 (CH₂), 34.9 (CH), 31.5 (CH₂), 31.0 (CH), 29.7 (CH), 24.8 (CH₃), 22.6 (CH₃), 20.11 (CH₂), 19.7 (CH₃), 19.2 (CH₃), 19.1 (CH₃), 18.6 (CH₃), 13.7 (CH₃); HRMS (ESI) [M+H]⁺ 503.3577, C₂₈H₄₆N₄O₄ [M+H]⁺ requires 503.3592.

2. ^1H NMR Titration Data

2.1. Compound 1 Homodimer VT ^1H NMR Titration

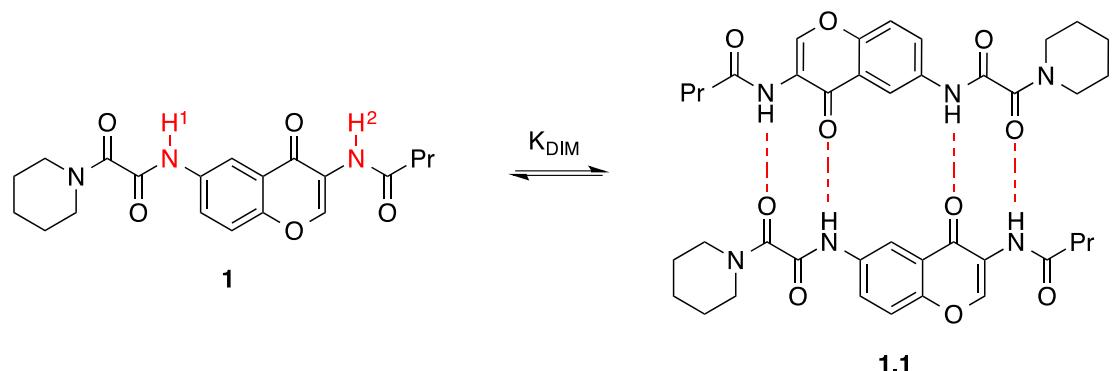


Figure 1: Homodimerisation of compound 1. Resonances (NH^1 and NH^2) used to determine K_{DIM} are highlighted in red.

2.1.1. Compound 1 Homodimer ^1H NMR Titration Experiment 1

Table 1: ^1H NMR titration of compound 1 at 278 K

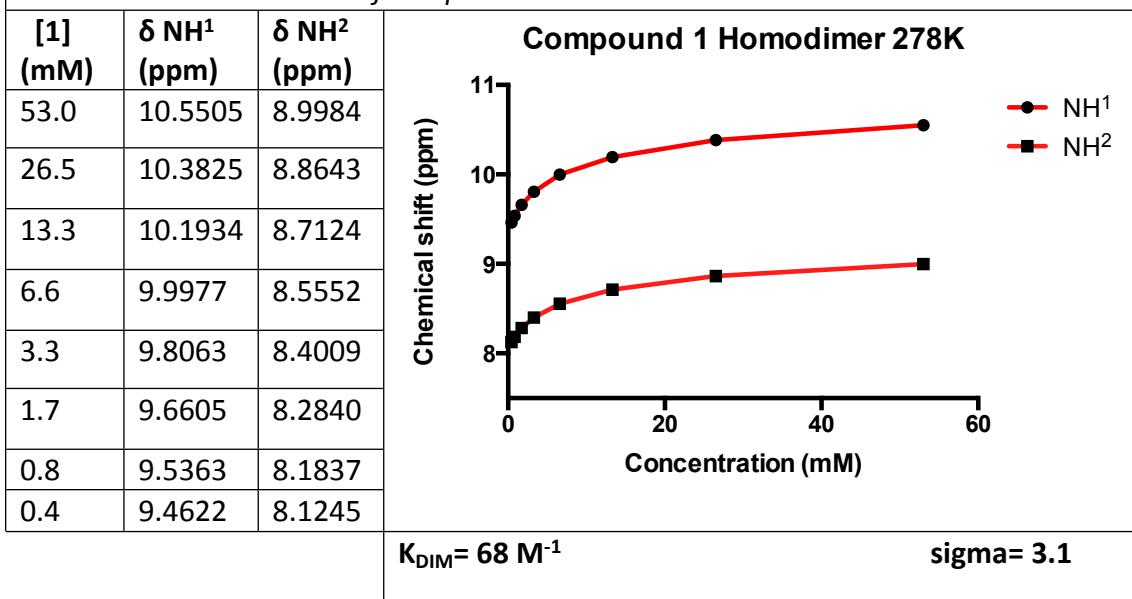


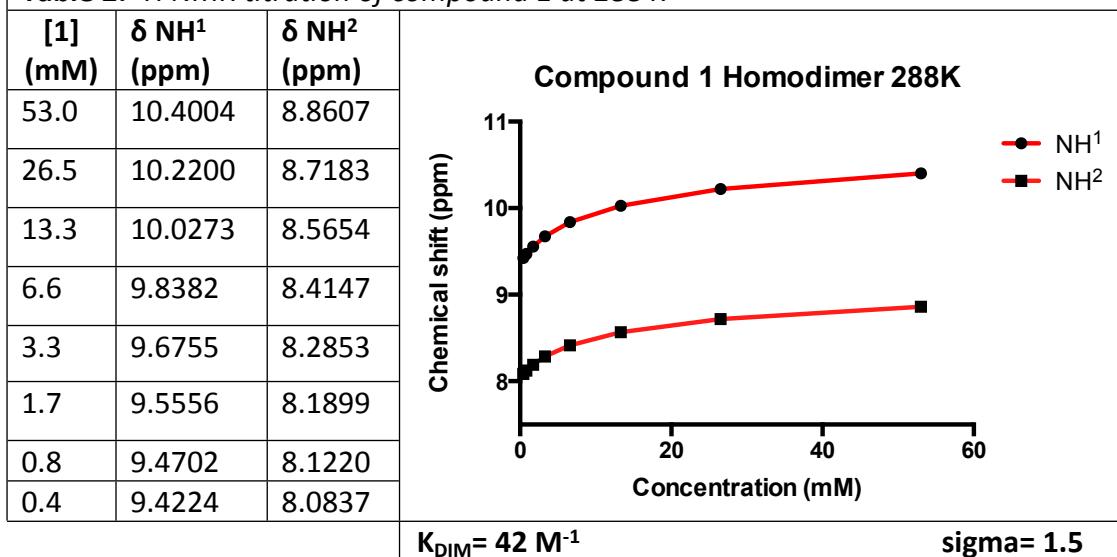
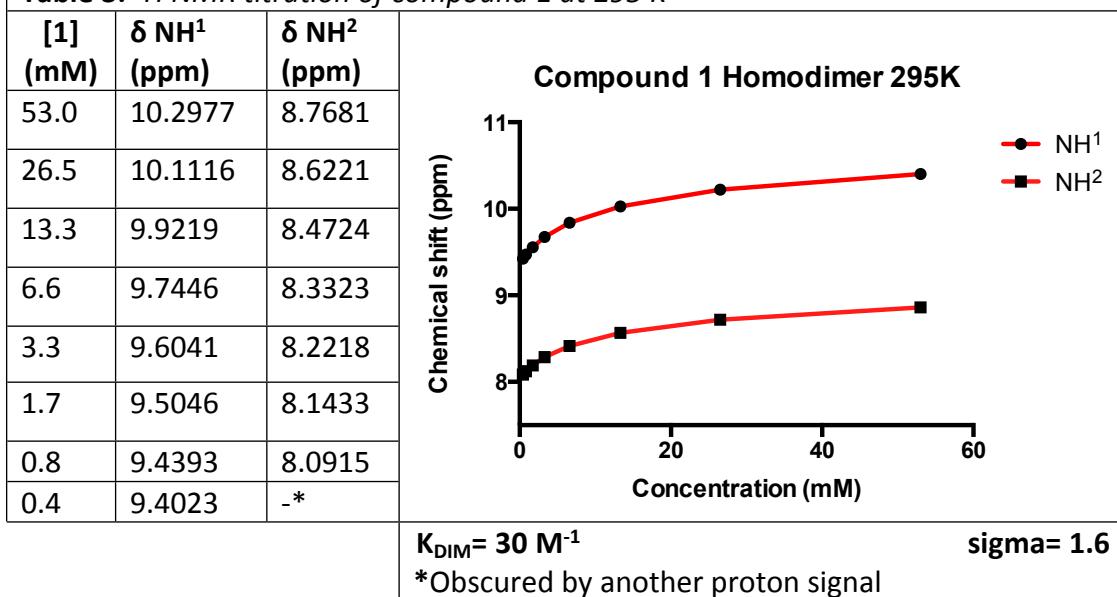
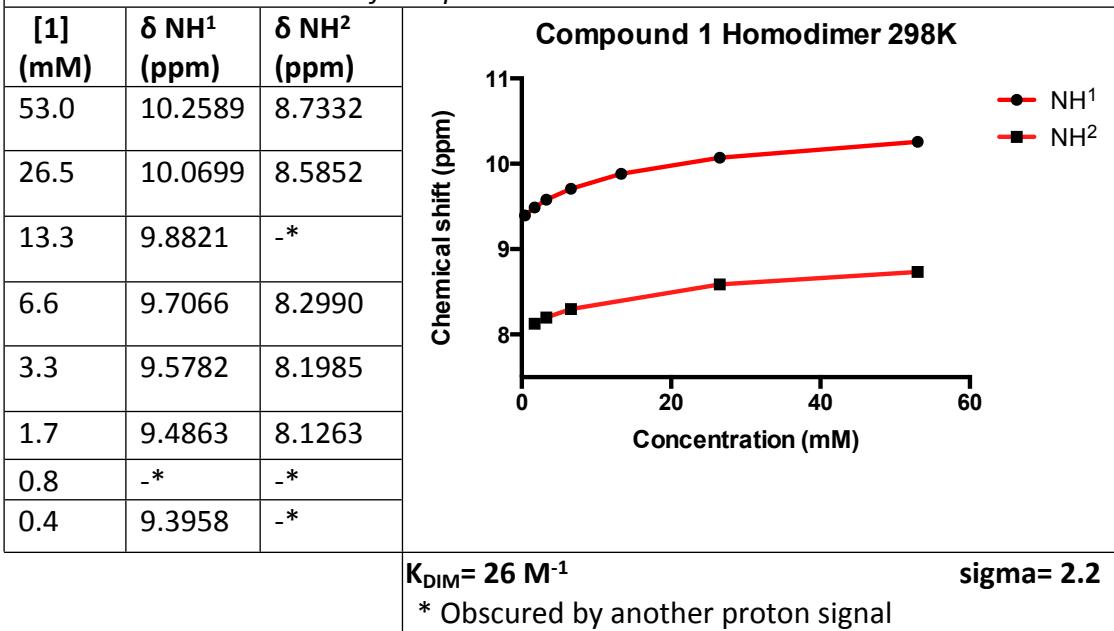
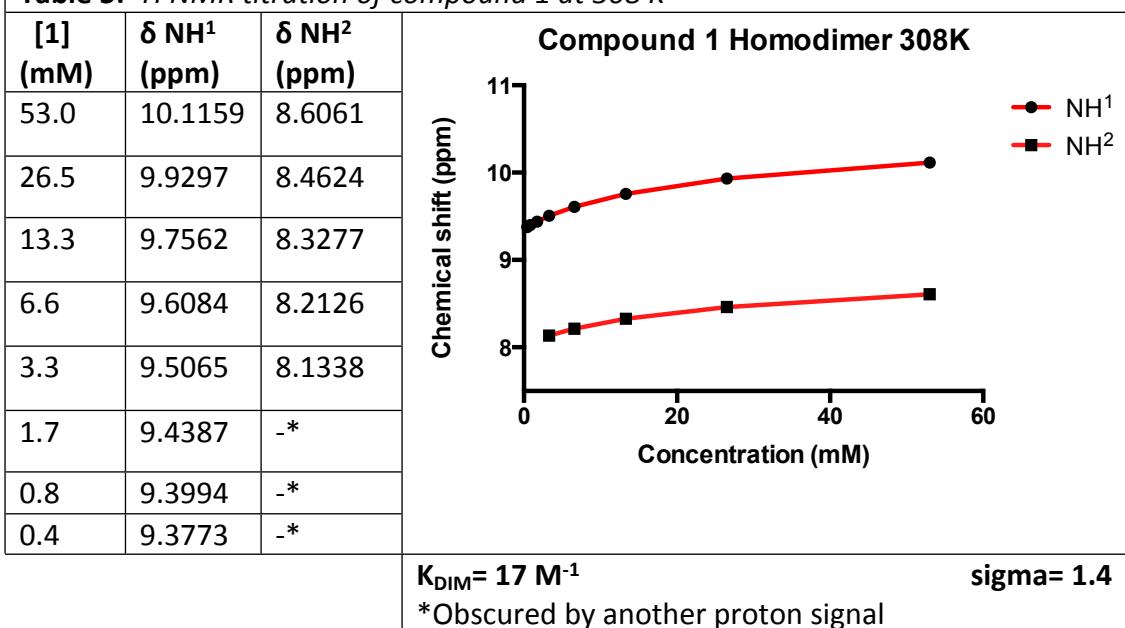
Table 2: ^1H NMR titration of compound 1 at 288 K**Table 3:** ^1H NMR titration of compound 1 at 295 K

Table 4: ^1H NMR titration of compound 1 at 298 K**Table 5:** ^1H NMR titration of compound 1 at 308 K

2.1.2. Compound 1 Homodimer ^1H NMR Titration Experiment 2

Table 6: ^1H NMR titration of compound 1 at 278 K

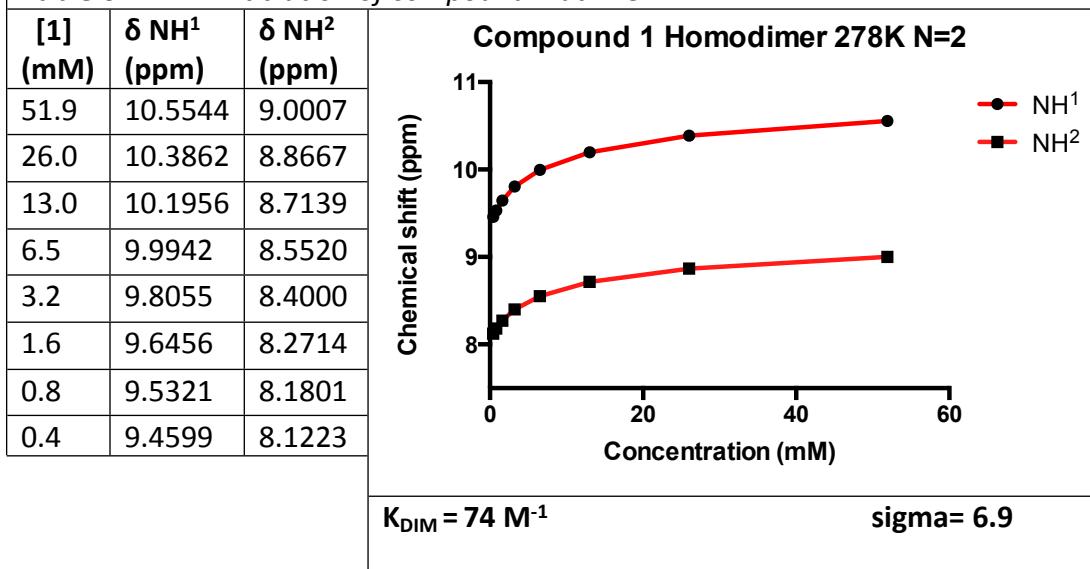


Table 7: ^1H NMR titration of compound 1 at 288 K

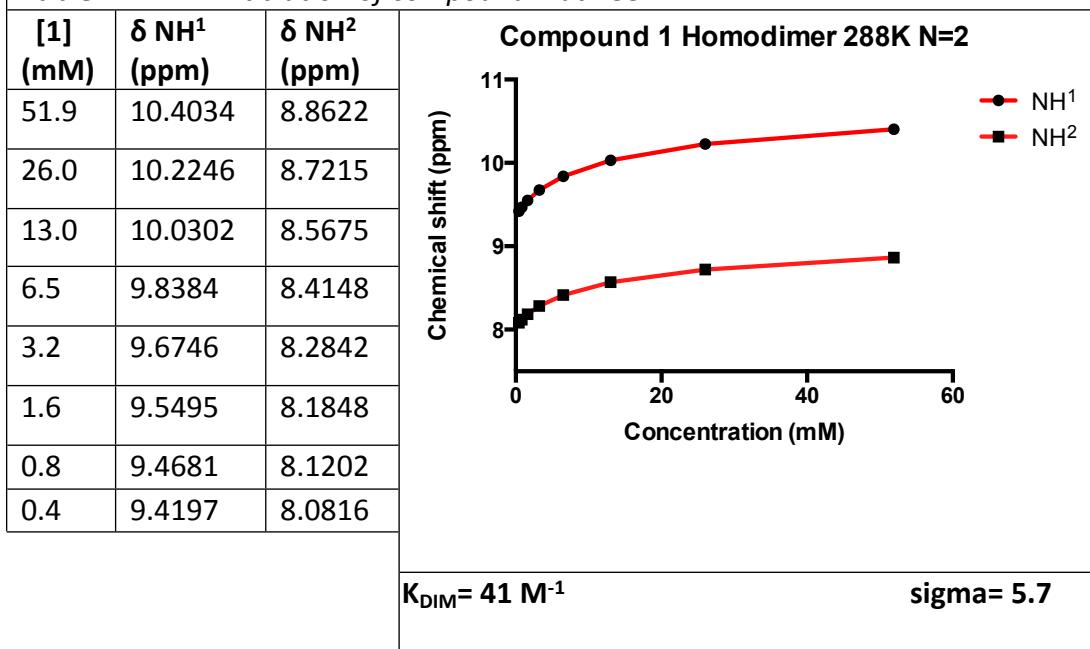


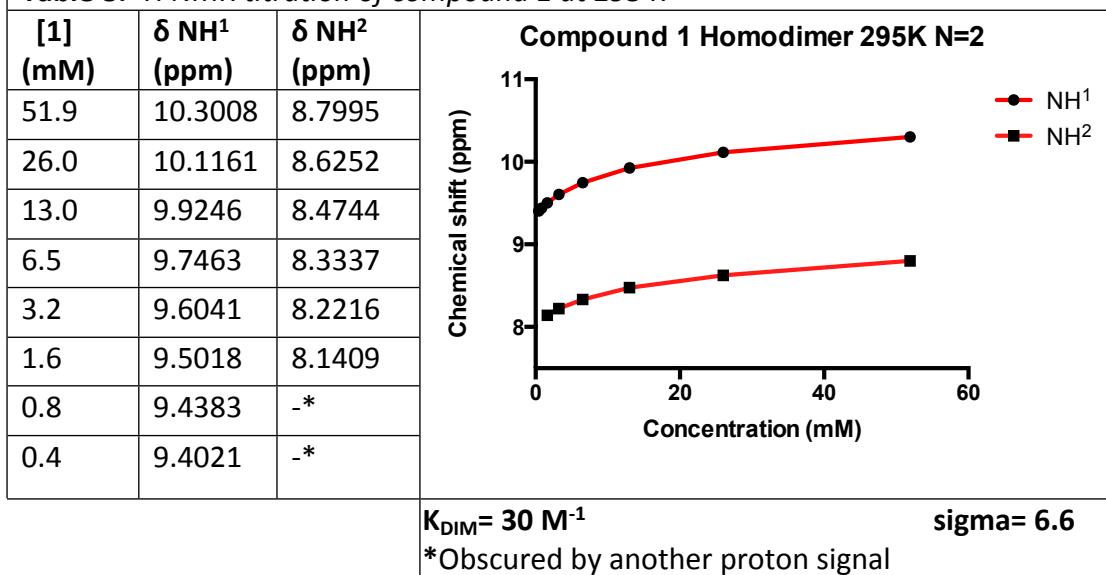
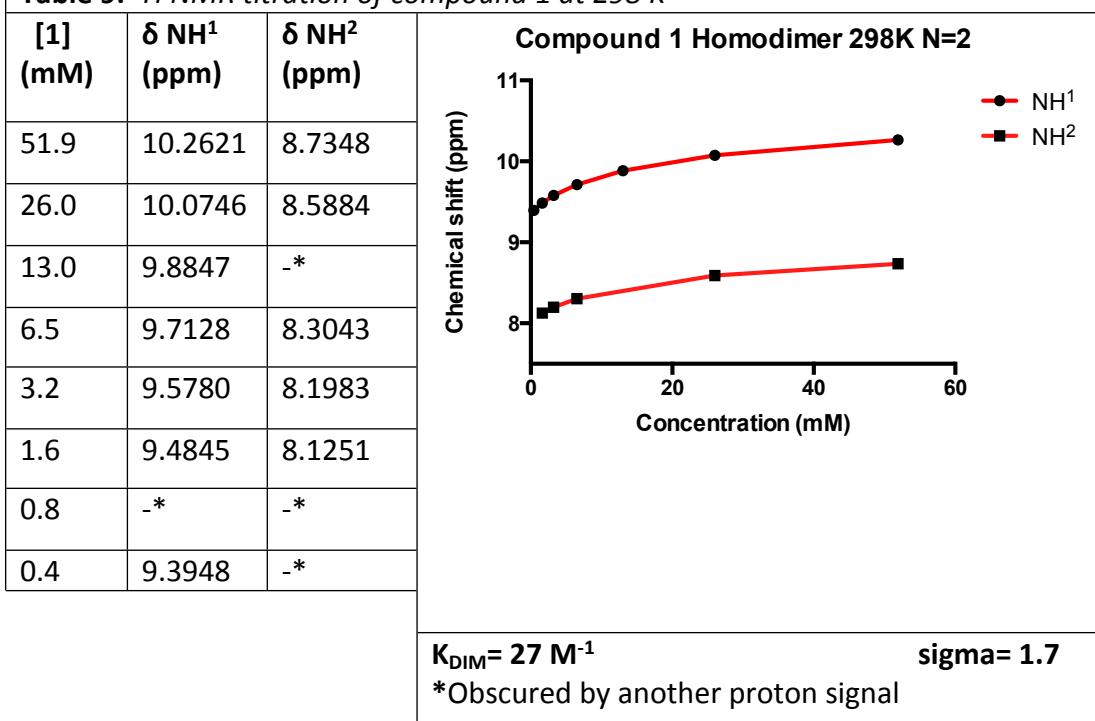
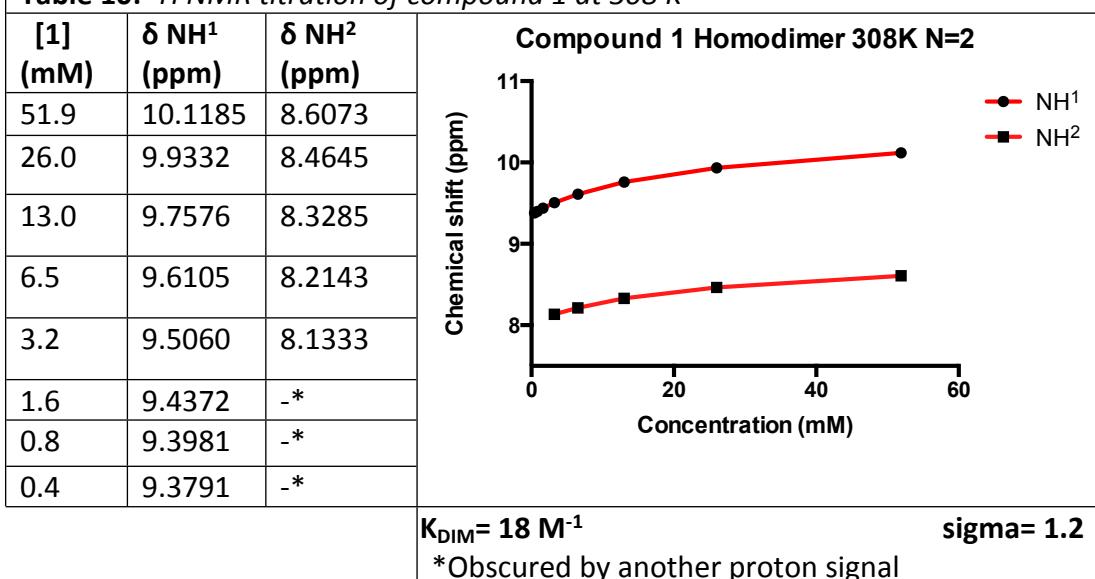
Table 8: ^1H NMR titration of compound 1 at 295 K**Table 9:** ^1H NMR titration of compound 1 at 298 K

Table 10: ^1H NMR titration of compound 1 at 308 K

2.1.3. Compound 1 Homodimer ^1H NMR Titration Experiment 3

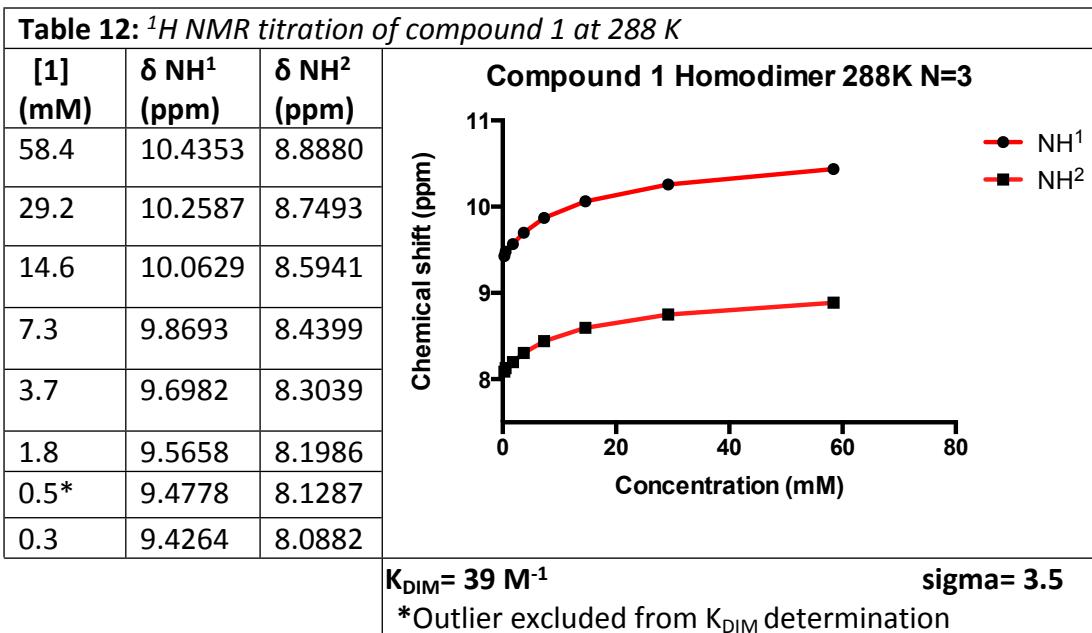
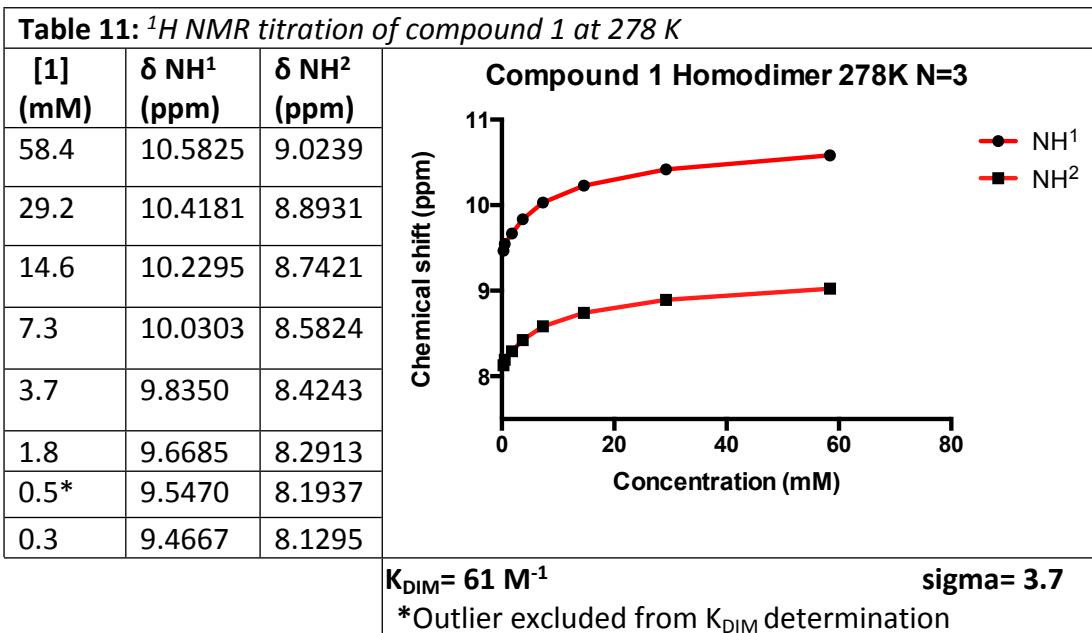


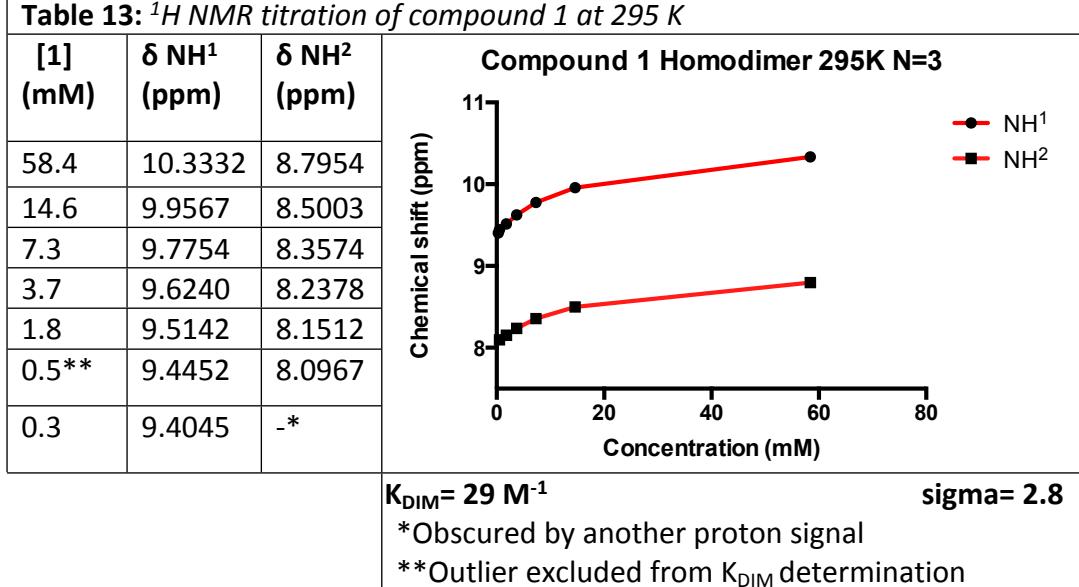
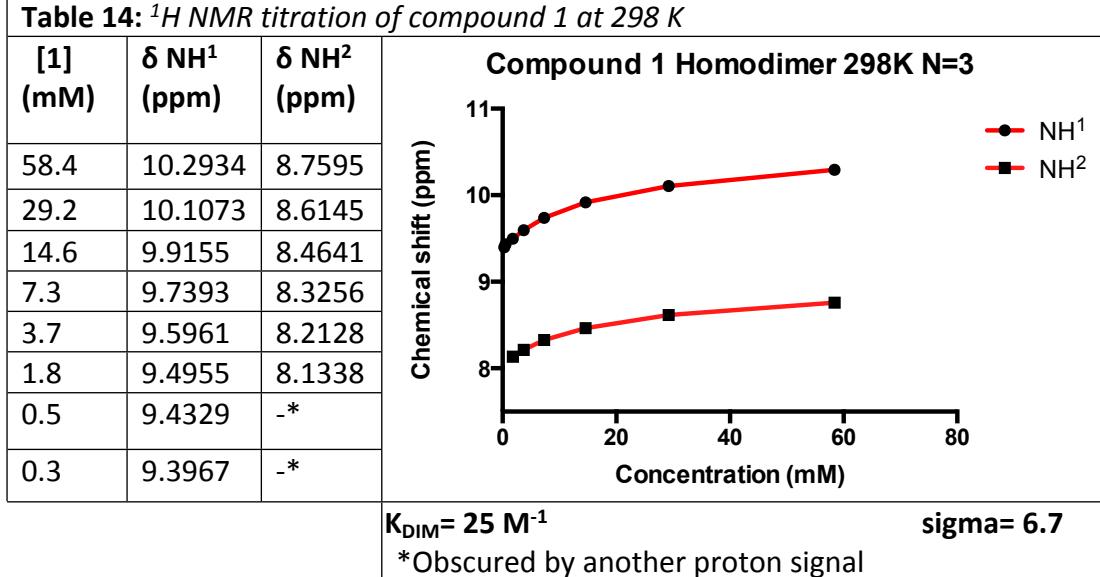
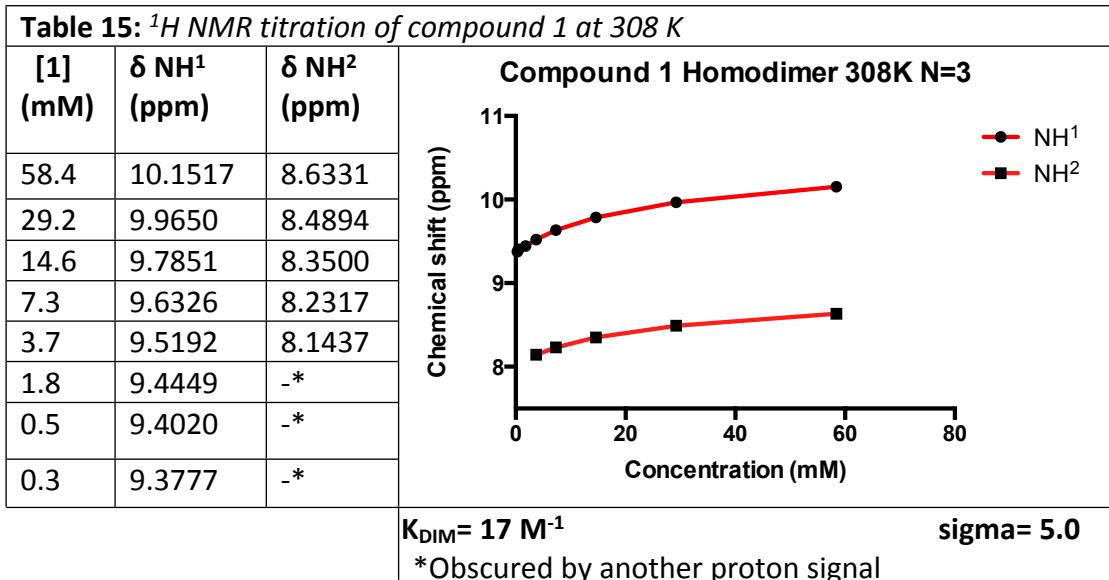
Table 13: ^1H NMR titration of compound 1 at 295 K**Table 14:** ^1H NMR titration of compound 1 at 298 K

Table 15: ^1H NMR titration of compound 1 at 308 K

2.2.4. Compound 1 Homodimer VT ^1H NMR Summary and Van't Hoff Plots

Table 16: Summary of Compound 1 Homodimer Experiment 1 and Van't Hoff Plot

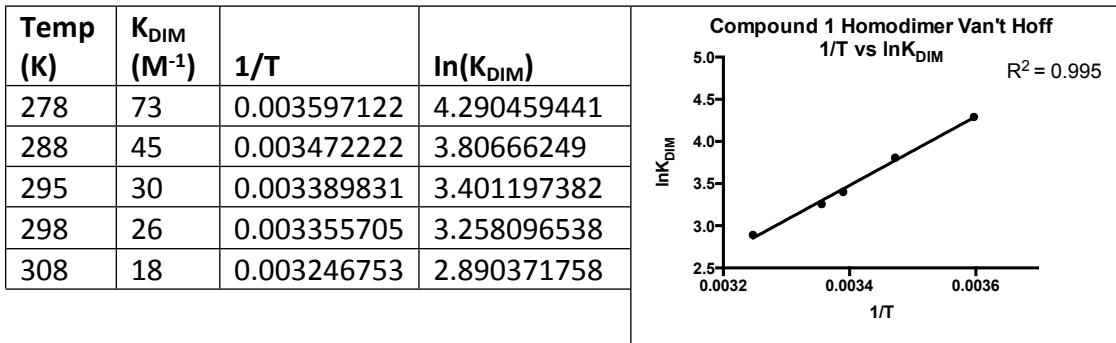


Table 17: Summary of Compound 1 Homodimer Experiment 2 and Van't Hoff Plot

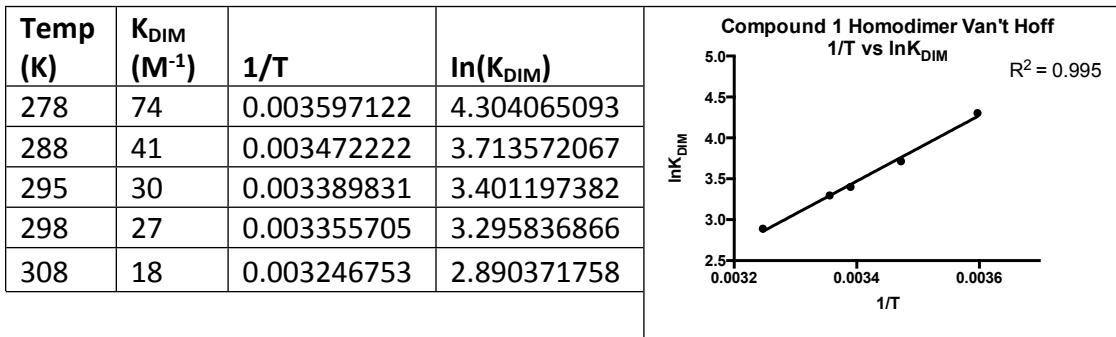


Table 18: Summary of Compound 1 Homodimer Experiment 3 and Van't Hoff Plot

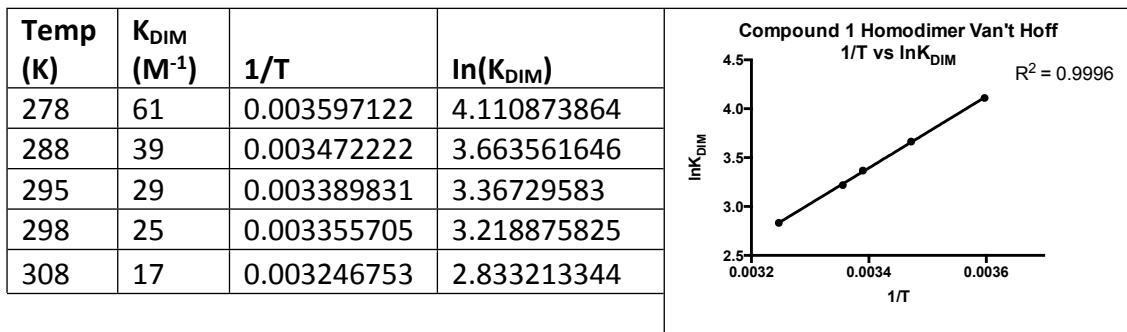


Table 19: Summary of thermodynamic parameters for compound 1 homodimer

Expt	ΔH (kcal mol $^{-1}$)	$-T\Delta S^{295 \text{ K}}$ (kcal mol $^{-1}$)	ΔG (kcal mol $^{-1}$)
1	-8.1	6.1	-2.0
2	-7.9	6.0	-1.9
3	-7.3	5.3	-2.0
Average	-7.8	5.8	-2.0

2.2. Tripeptide 11 Homodimer ^1H NMR Titration

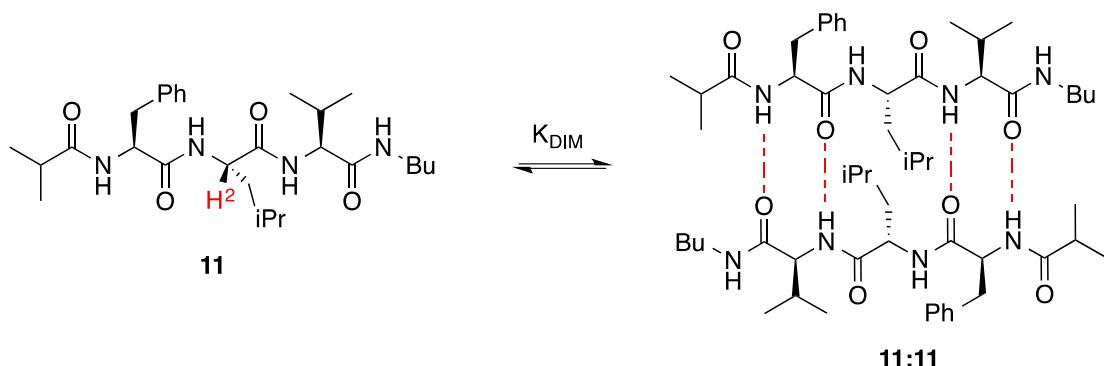


Figure 2: Homodimerisation of **11**. Resonance (H^2) used to determine K_{DIM} is highlighted in red

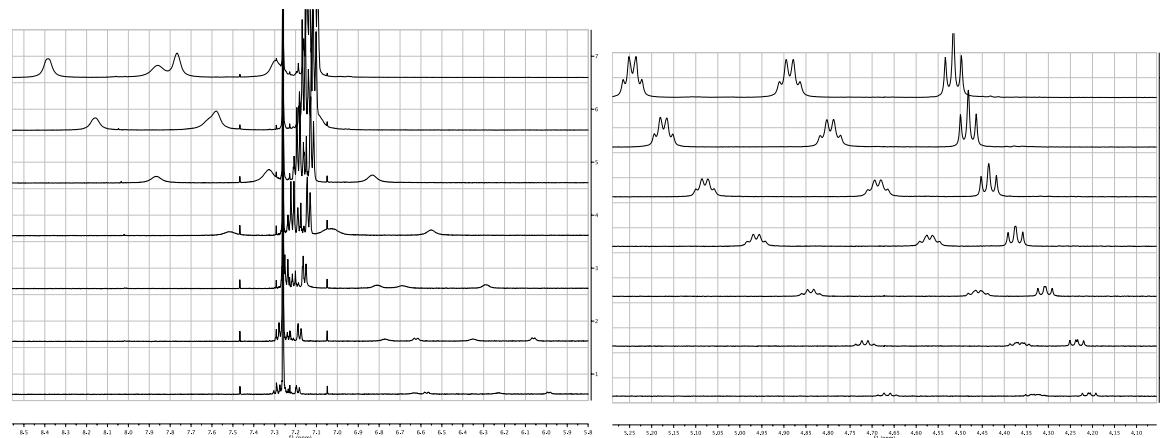


Figure 3: Comparison between NH shifts (left) and αCH shifts (right) of compound **11**. NH signals broaden into the baseline at lower concentrations making it difficult to accurately measure chemical shift. αCH signals are relatively sharp compared to NH signals and therefore were used to determine binding constants

2.2.1 Compound 11 Homodimer ^1H NMR Titration Experiment 1

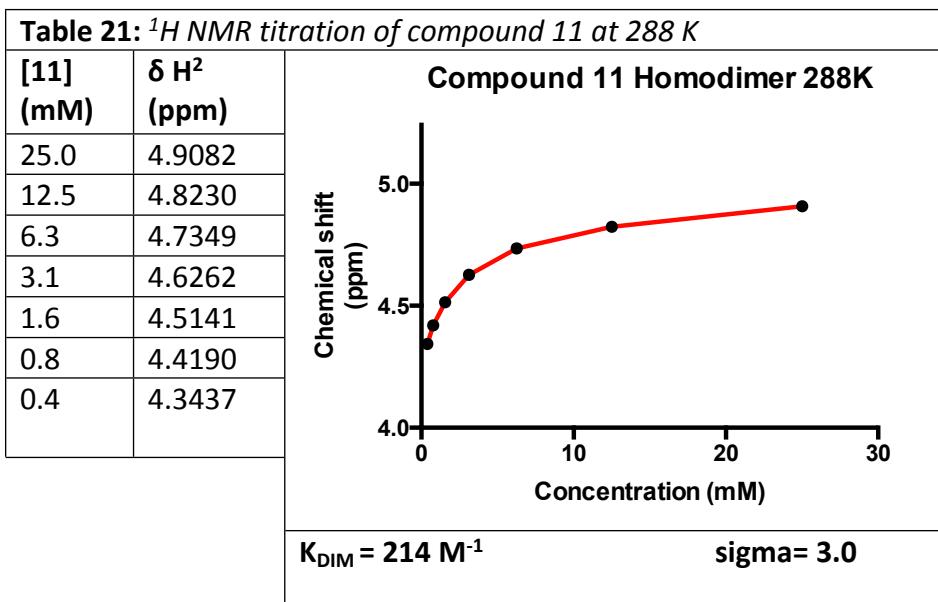
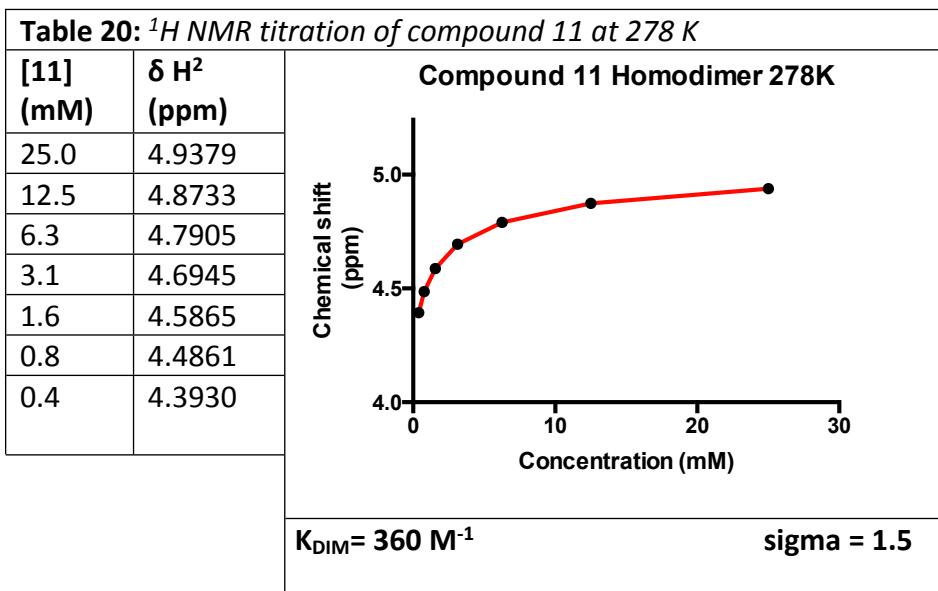


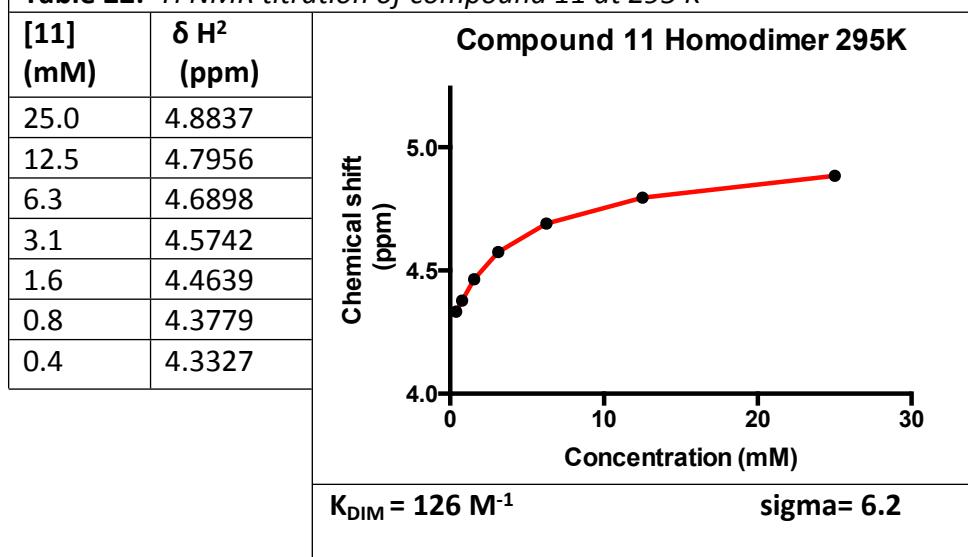
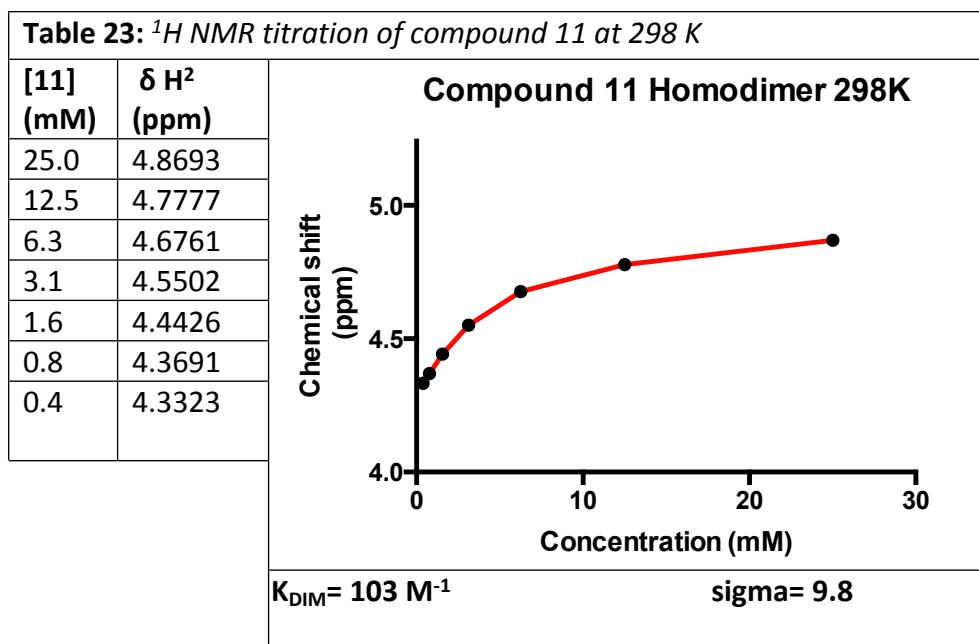
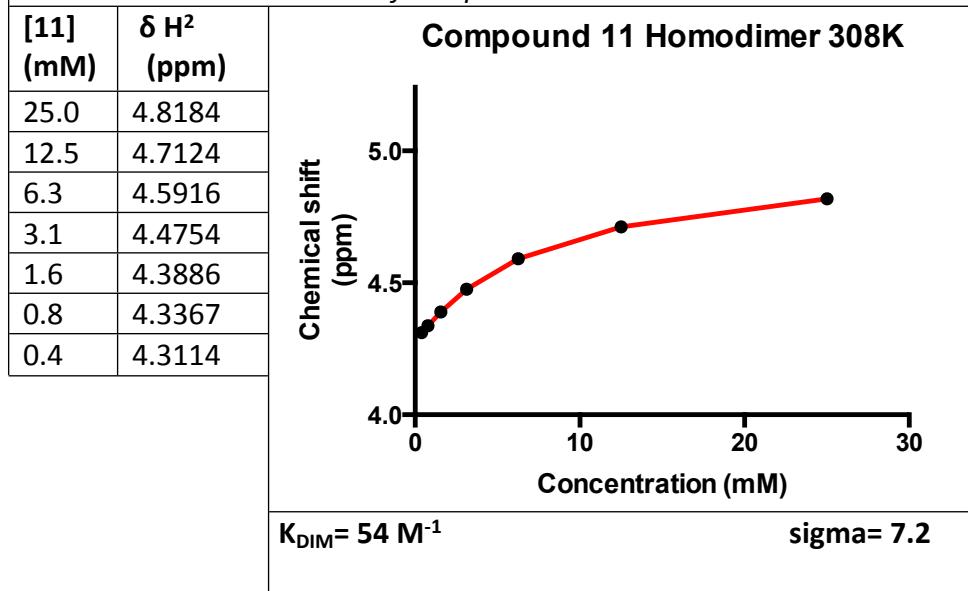
Table 22: ^1H NMR titration of compound 11 at 295 K**Table 23:** ^1H NMR titration of compound 11 at 298 K

Table 24: ^1H NMR titration of compound 11 at 308 K



2.2.2. Compound 11 Homodimer ^1H NMR Titration Experiment 2

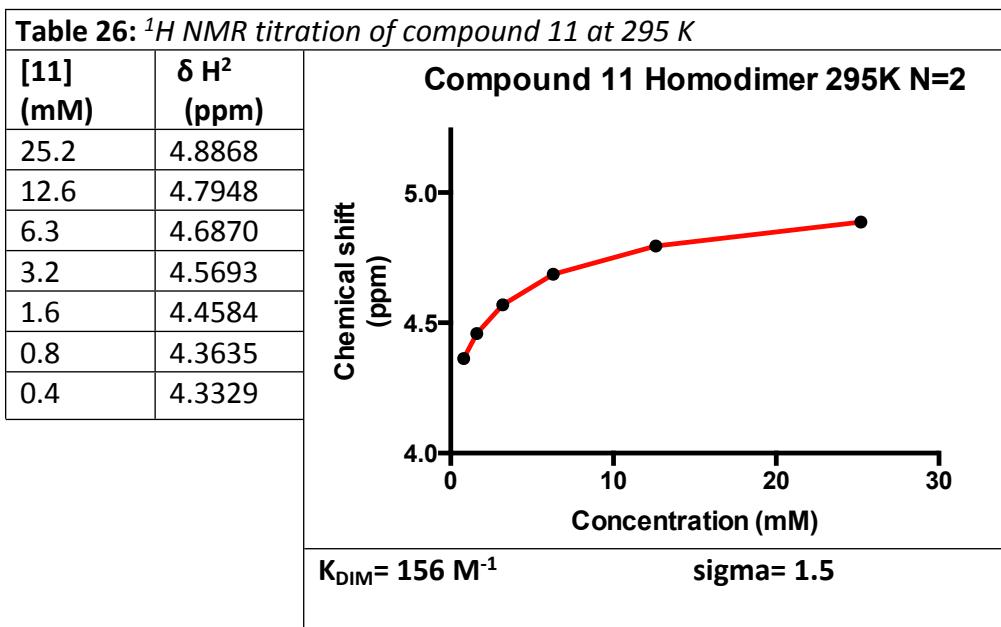
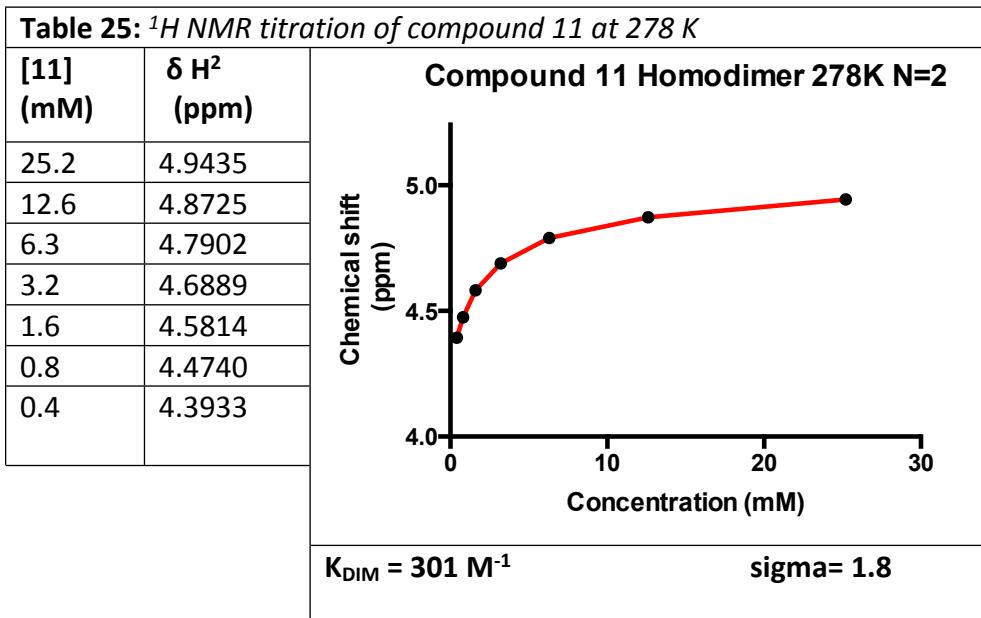
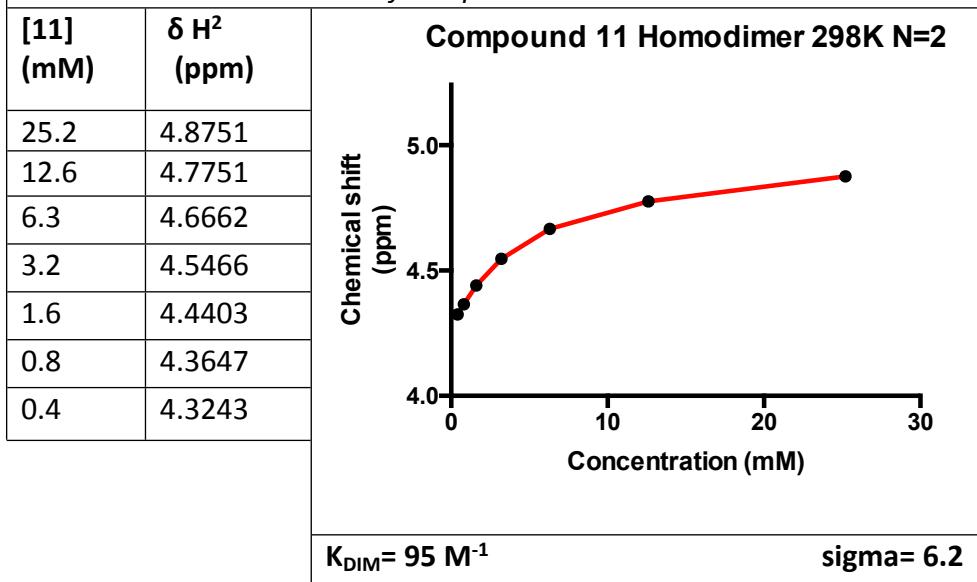
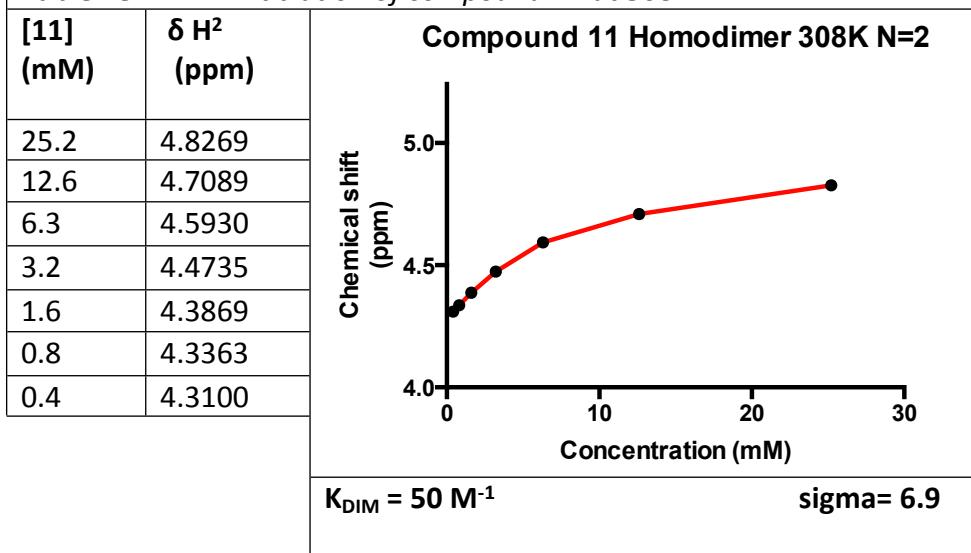


Table 27: ^1H NMR titration of compound 11 at 298 K**Table 28:** ^1H NMR titration of compound 11 at 308 K

2.2.3. Compound 11 Homodimer ^1H NMR Titration Experiment 3

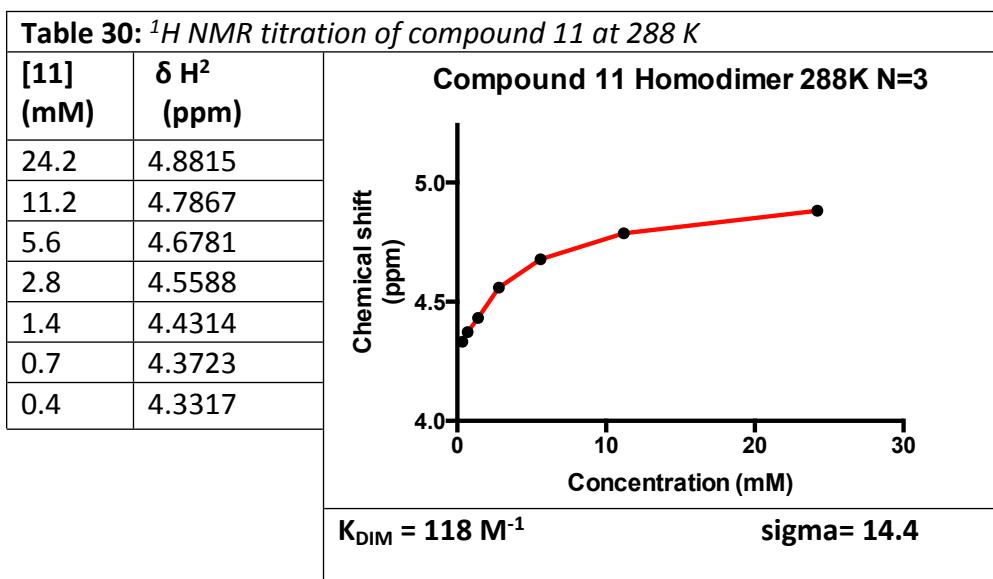
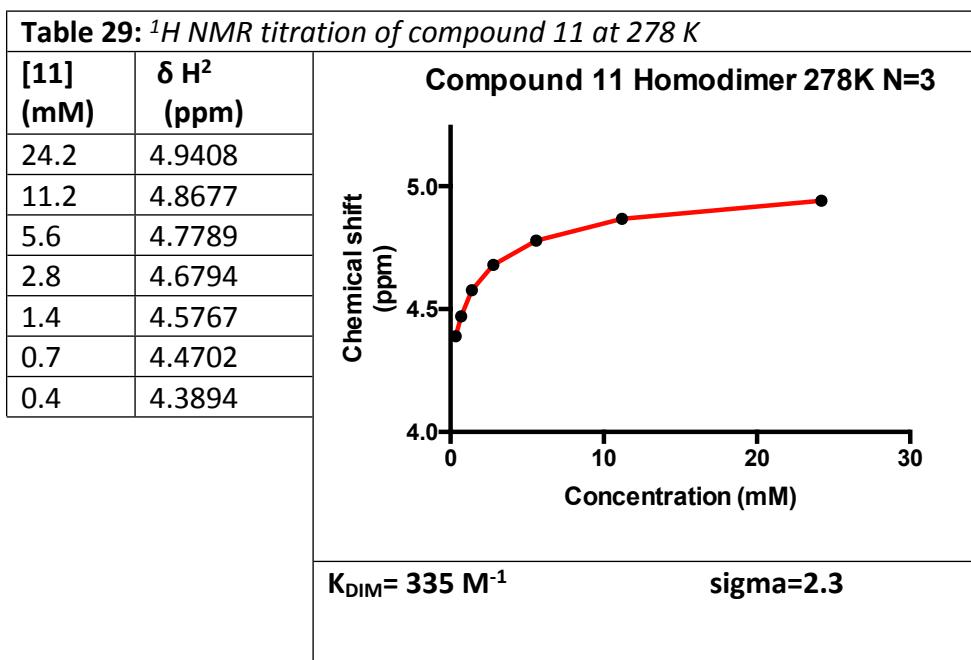
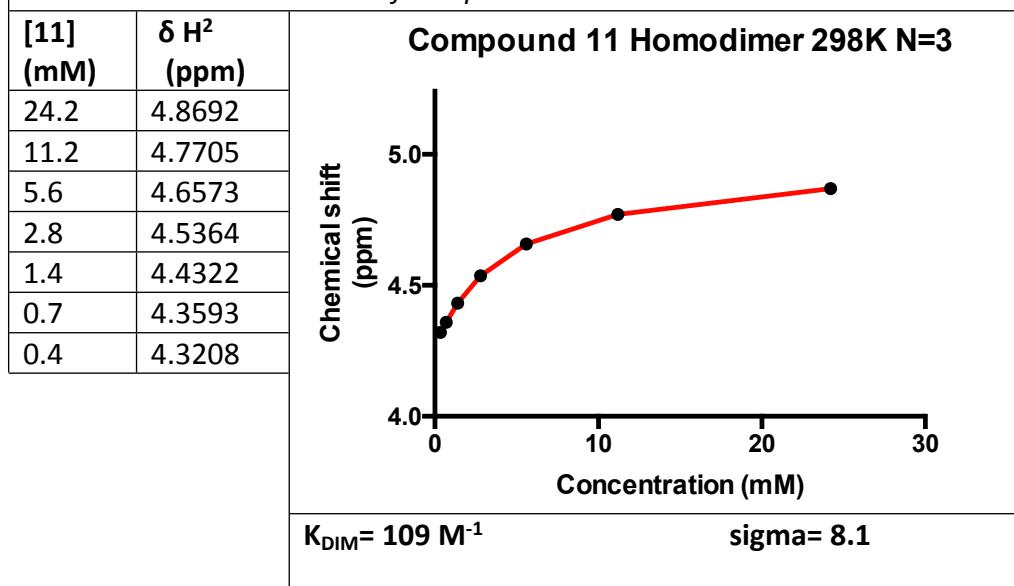
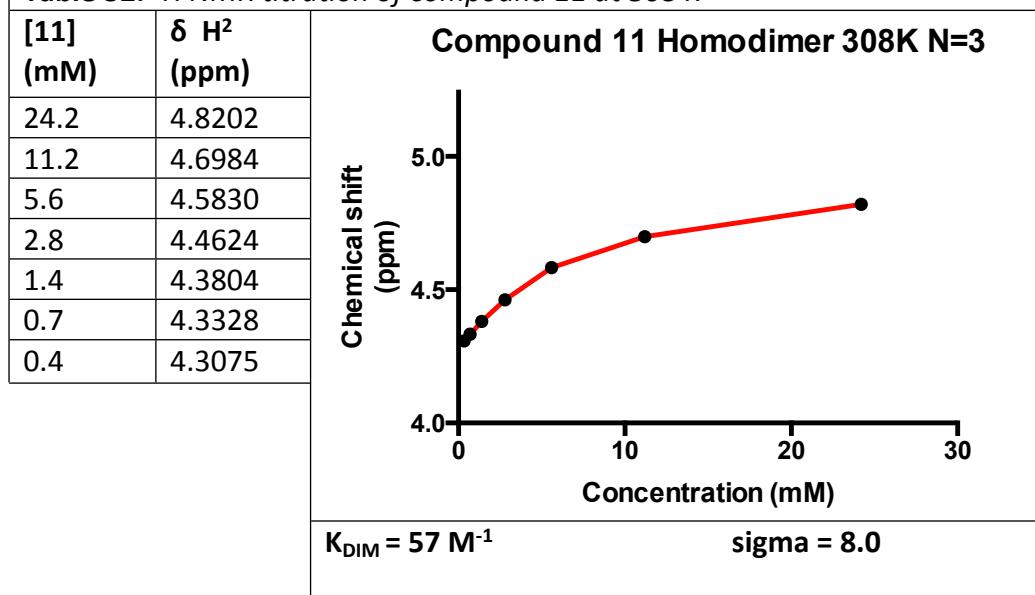


Table 31: ^1H NMR titration of compound 11 at 298 K**Table 32:** ^1H NMR titration of compound 11 at 308 K

2.2.4. Compound 11 Homodimer VT ^1H NMR Summary and Van't Hoff Plots

Table 33: Summary of compound 11 Homodimer Experiment 1 and Van't Hoff Plot

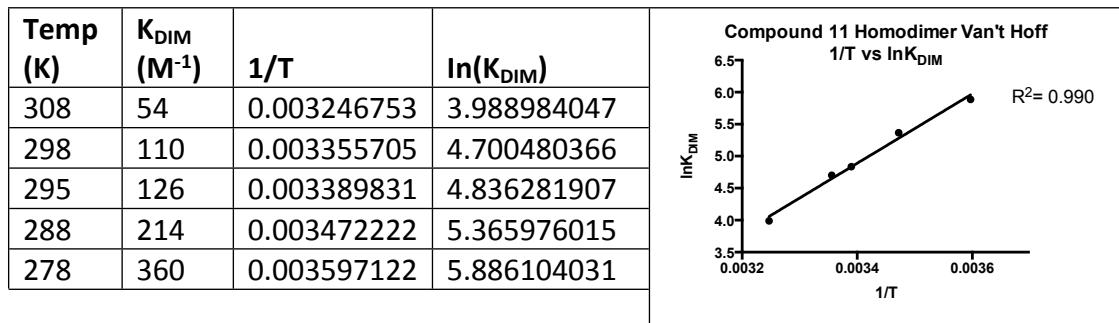


Table 34: Summary of compound 11 Homodimer Experiment 2 and Van't Hoff Plot

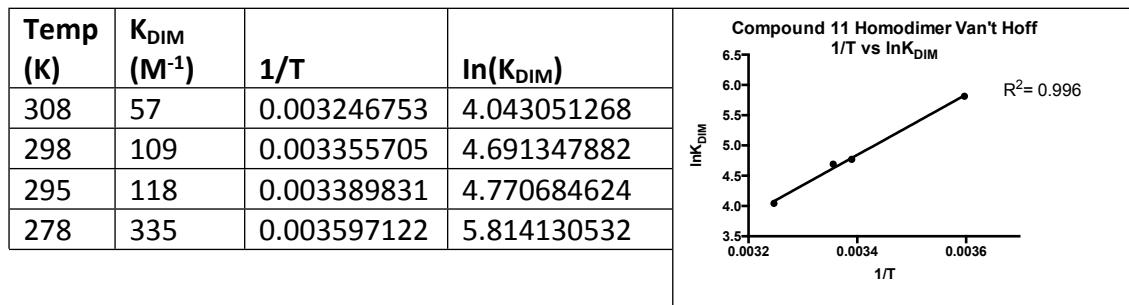


Table 35: Summary of compound 11 Homodimer Experiment 3 and Van't Hoff Plot

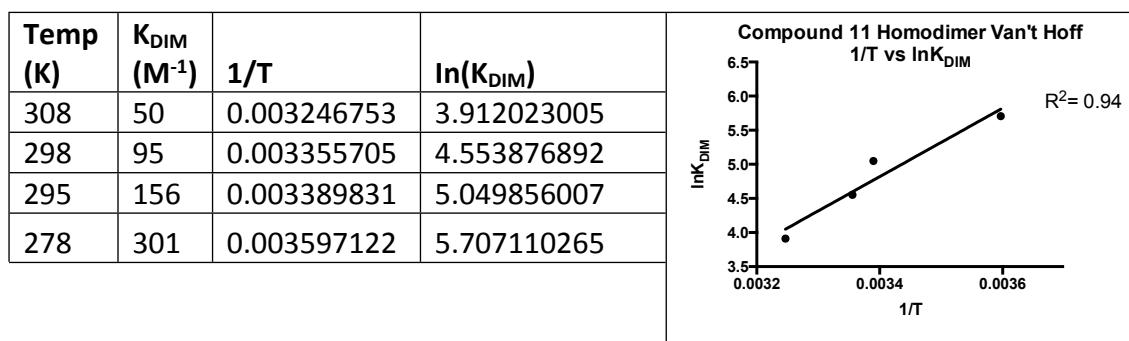


Table 36: Summary of thermodynamic parameters for compound 11 homodimer

Expt	ΔH (kcal mol $^{-1}$)	$-\Delta S^{295 \text{ K}}$ (kcal mol $^{-1}$)	ΔG (kcal mol $^{-1}$)
1	-10.8	7.9	-2.9
2	-10.0	7.2	-2.8
3	-9.9	7.1	-2.8
Average	-10.2	7.4	-2.8

2.3. Compound 2 Homodimer VT ^1H NMR Titration

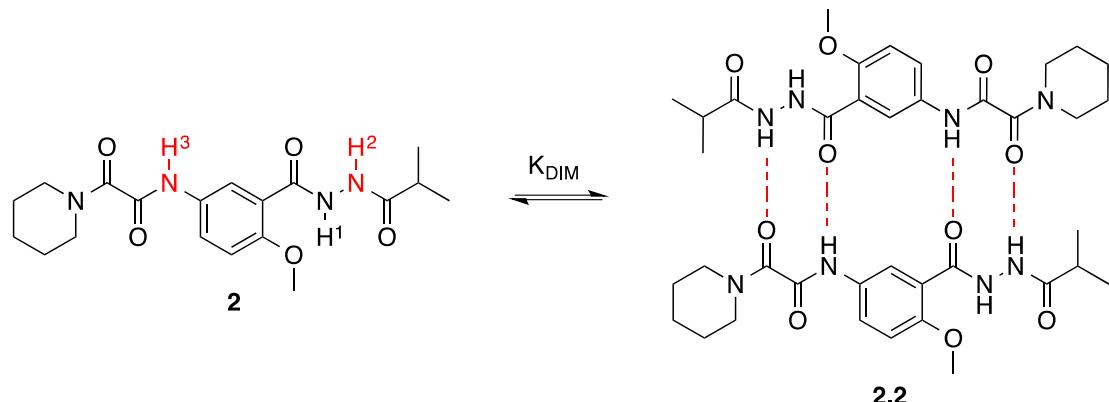


Figure 4: Homodimerisation of HAO. Resonances used to calculate K_{DIM} are highlighted in red.

N.B. K_{DIM} was calculated using NH^2 and NH^3 . The shifts of NH^1 are included for completeness.

2.3.1. Compound 2 Homodimer ^1H NMR Titration Experiment 1

Table 37: ^1H NMR titration of 2 at 278 K

[2] (mM)	$\delta \text{ NH}^1$ (ppm)	$\delta \text{ NH}^2$ (ppm)	$\delta \text{ NH}^3$ (ppm)
51.4	11.4178	11.2621	10.8163
25.7	11.4094	11.2182	10.7872
12.9	11.3964	11.1572	10.7457
6.4	11.3771	11.0726	10.6876
3.2	11.3507	10.9579	10.6093
1.6	11.3160	10.8086	10.5055
0.8	11.2709	10.6180	10.3733
0.4	11.2163	10.3790	10.2100

Compound 2 Homodimer 278K

Chemical shift (ppm)

Concentration (mM)

$K_{\text{DIM}} = 4747 \text{ M}^{-1}$

$\sigma = 0.8$

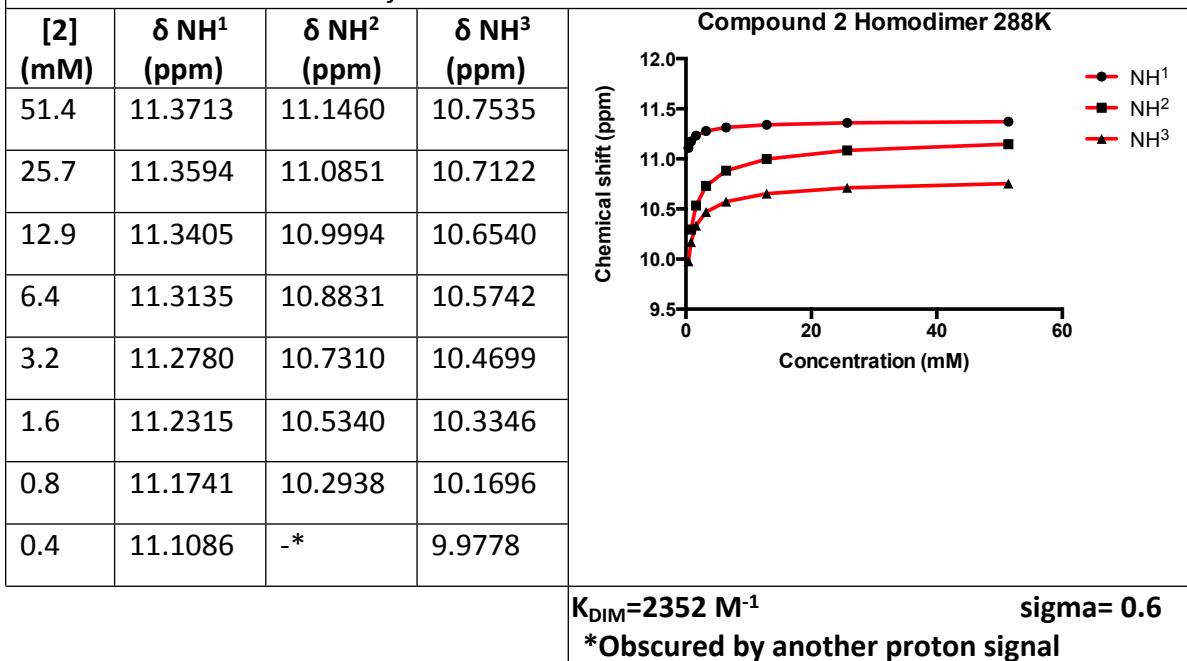
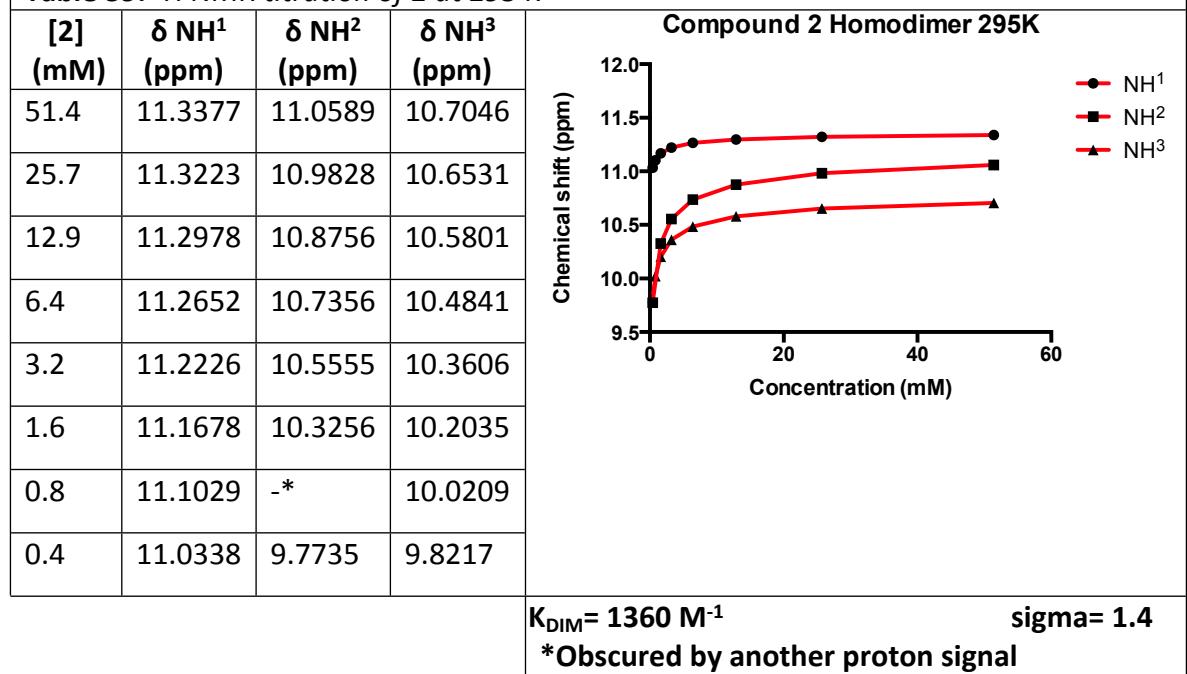
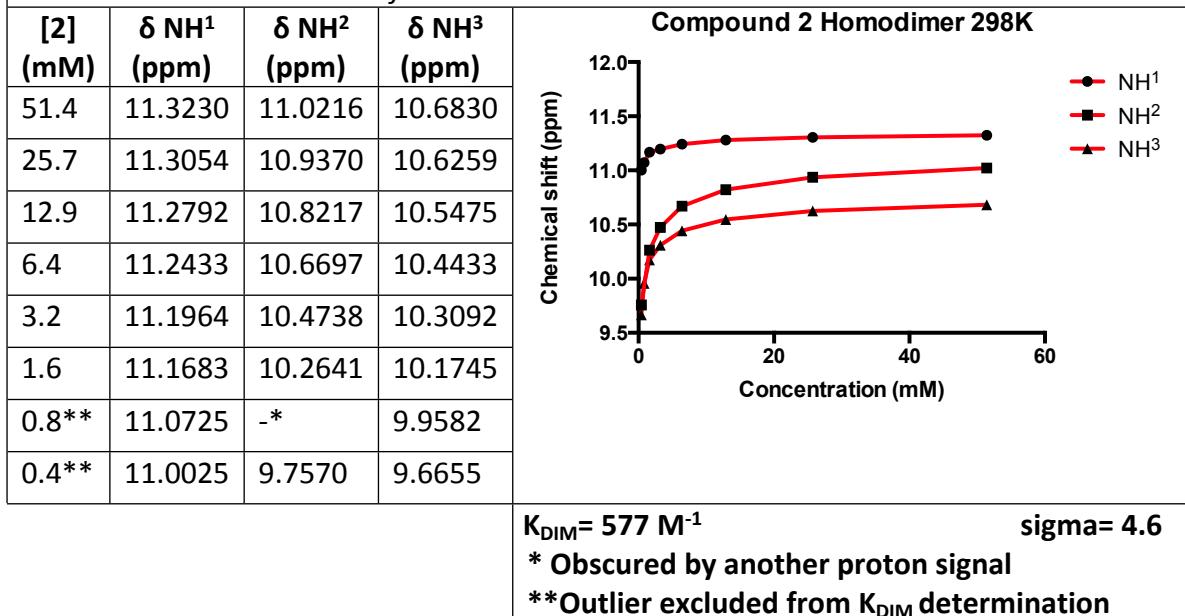
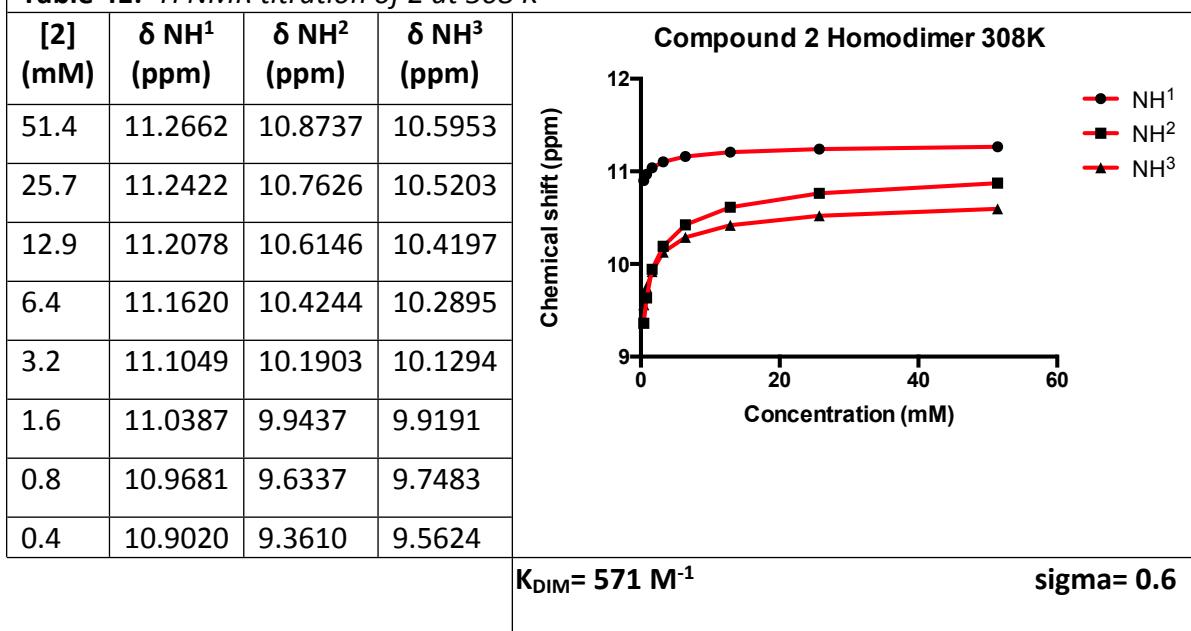
Table 38: ^1H NMR titration of 2 at 288 K**Table 39:** ^1H NMR titration of 2 at 295 K

Table 40: ^1H NMR titration of 2 at 298 K**Table 41:** ^1H NMR titration of 2 at 308 K

2.3.2. Compound 2 Homodimer ^1H NMR Titration Experiment 2

Table 42: ^1H NMR titration of 2 at 278 K

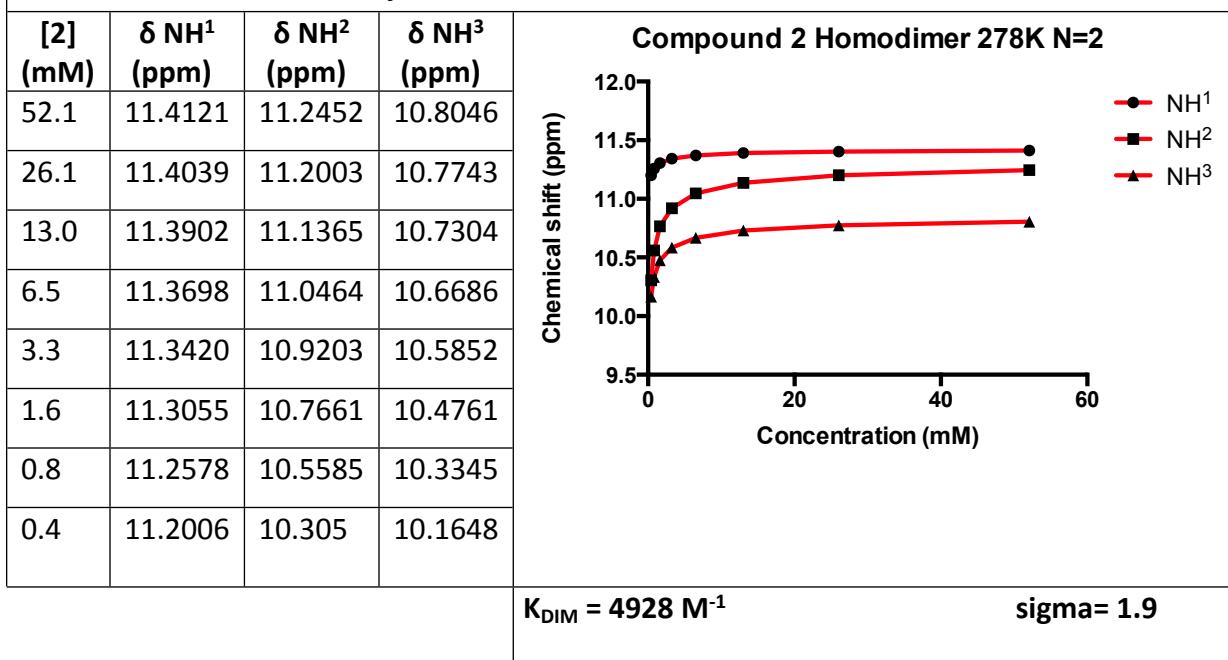


Table 43: ^1H NMR titration of 2 at 288 K

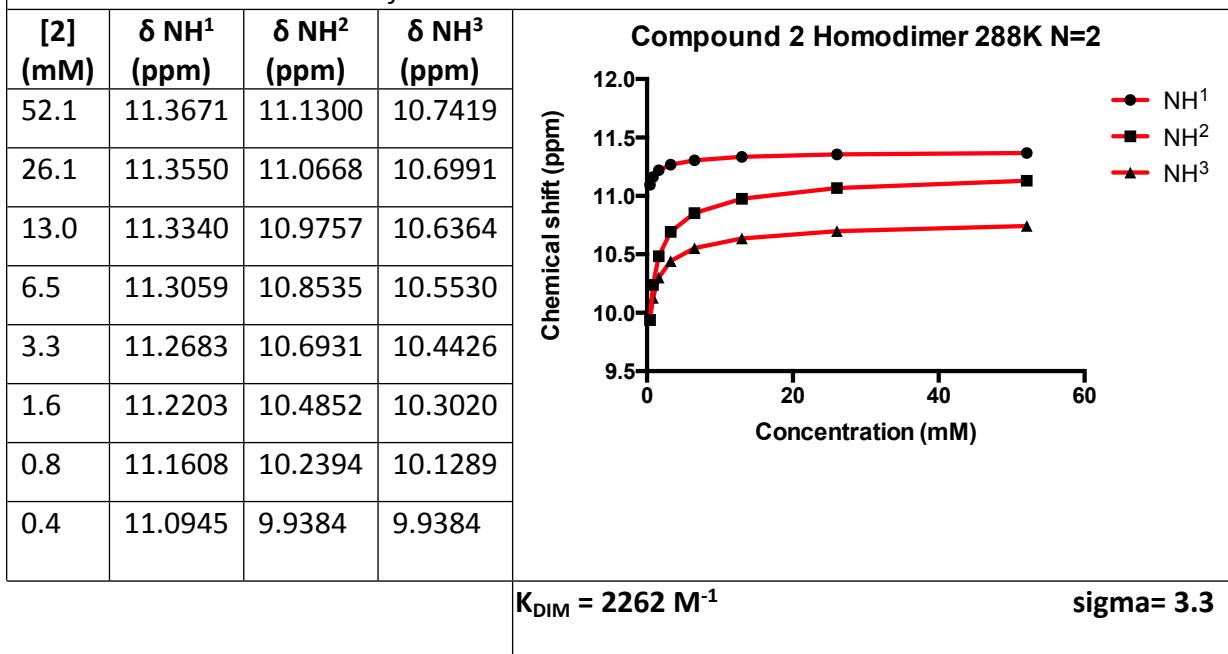


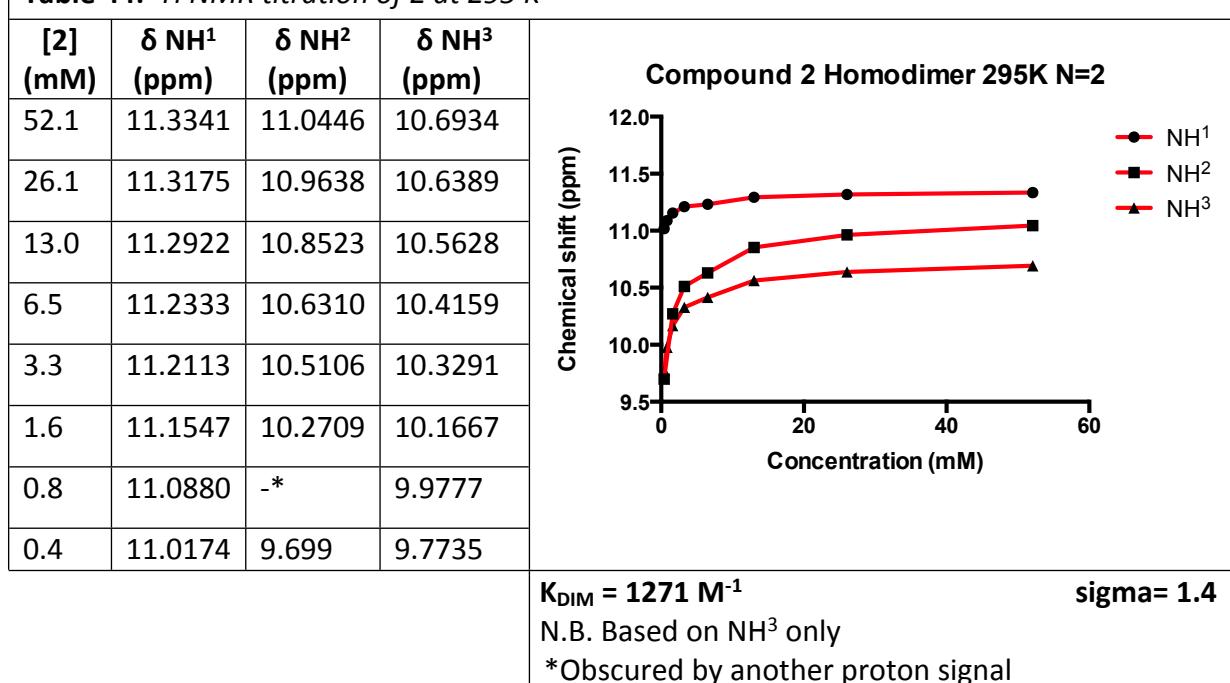
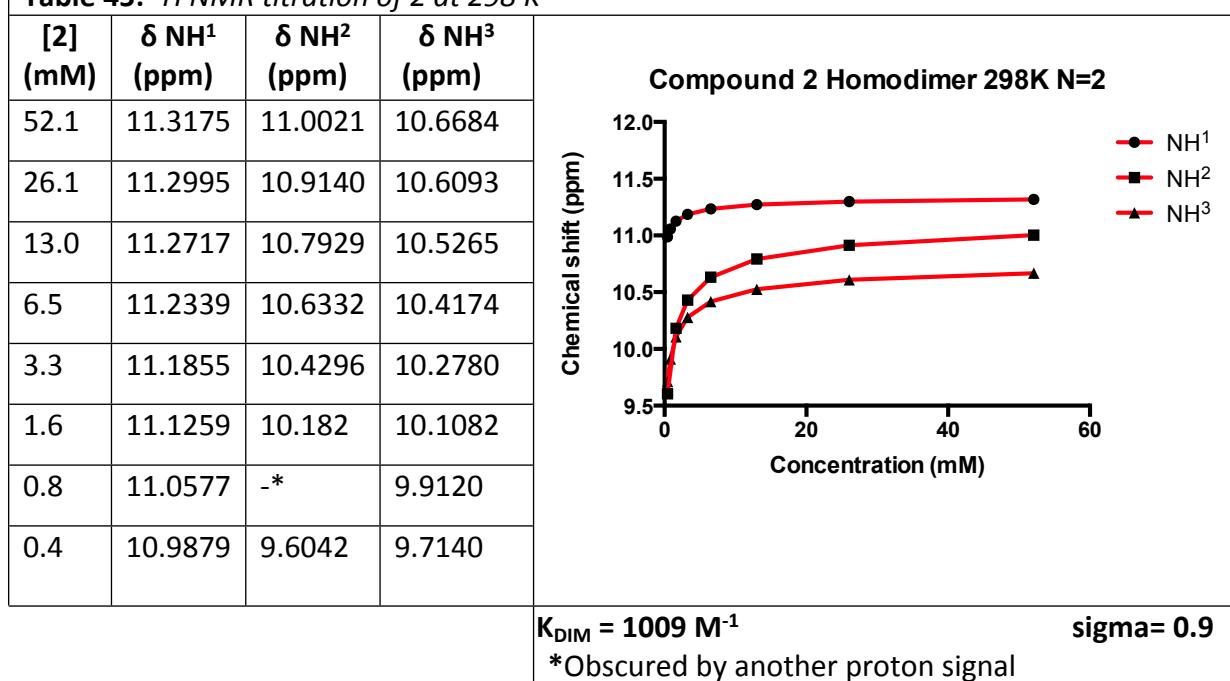
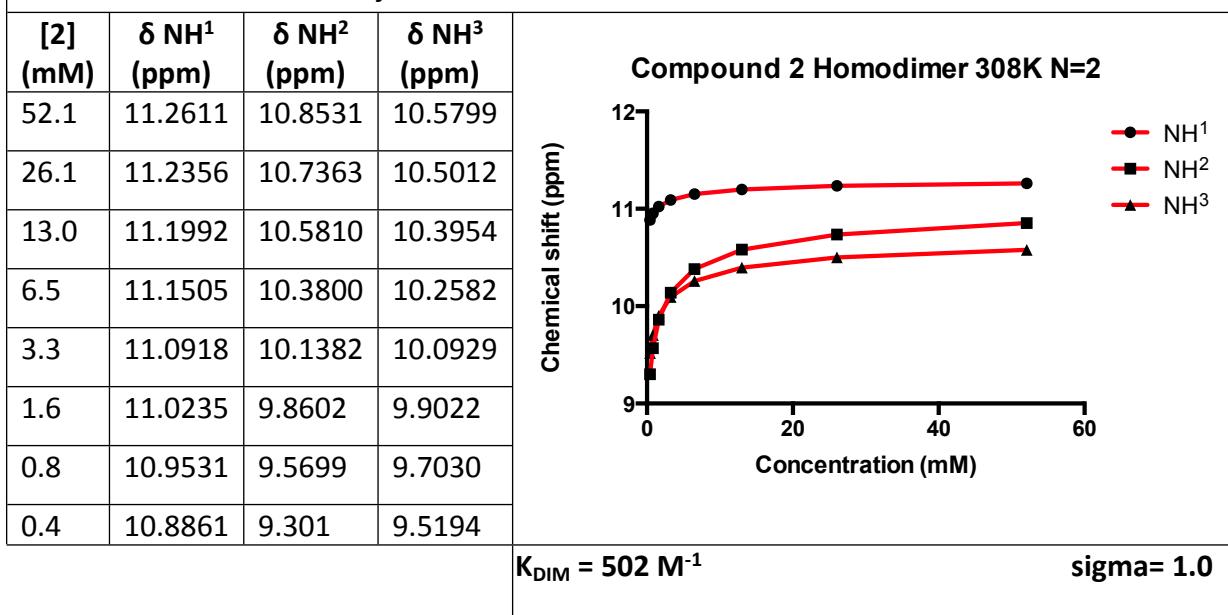
Table 44: ^1H NMR titration of 2 at 295 K**Table 45:** ^1H NMR titration of 2 at 298 K

Table 46: ^1H NMR titration of 2 at 308 K



2.3.3. Compound 2 Homodimer ^1H NMR Titration Experiment 3

Table 47: ^1H NMR titration of 2 at 278 K

[2] (mM)	$\delta \text{ NH}^1$ (ppm)	$\delta \text{ NH}^2$ (ppm)	$\delta \text{ NH}^3$ (ppm)
54.2	11.4141	11.2535	10.8103
27.1	11.4077	11.2146	10.7847
13.6	11.3951	11.1539	10.7440
6.8	11.3772	11.0729	10.6890
3.4	11.3520	10.9635	10.6132
1.7	11.3178	10.8147	10.5117
0.9	11.2747	10.6290	10.3839
0.4	11.2194	-*	10.2199

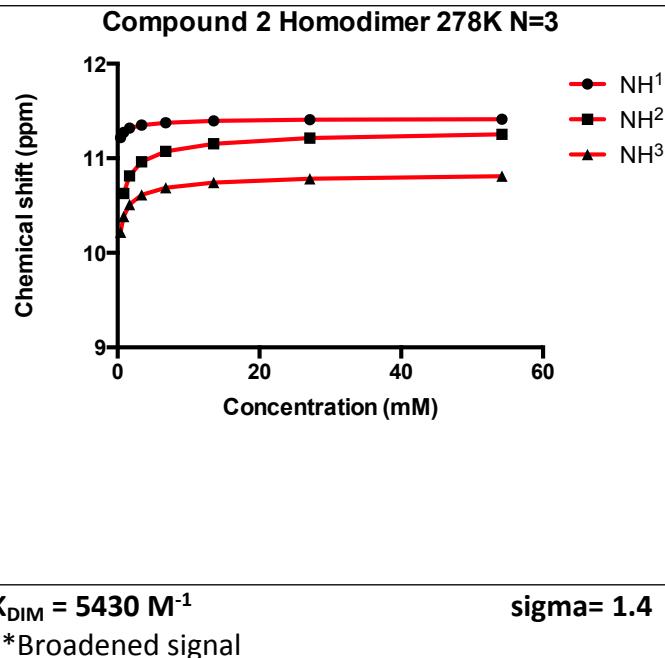


Table 48: ^1H NMR titration of 2 at 288 K

[2] (mM)	$\delta \text{ NH}^1$ (ppm)	$\delta \text{ NH}^2$ (ppm)	$\delta \text{ NH}^3$ (ppm)
54.2	11.3694	11.1407	10.7491
27.1	11.3597	11.0849	10.7124
13.6	11.3416	11.0007	10.6555
6.8	11.3161	10.8882	10.5789
3.4	11.2812	10.7384	10.4766
1.7	11.2362	10.547	10.3450
0.9	11.1813	10.3145	10.1856
0.4	11.1158	-*	9.9954

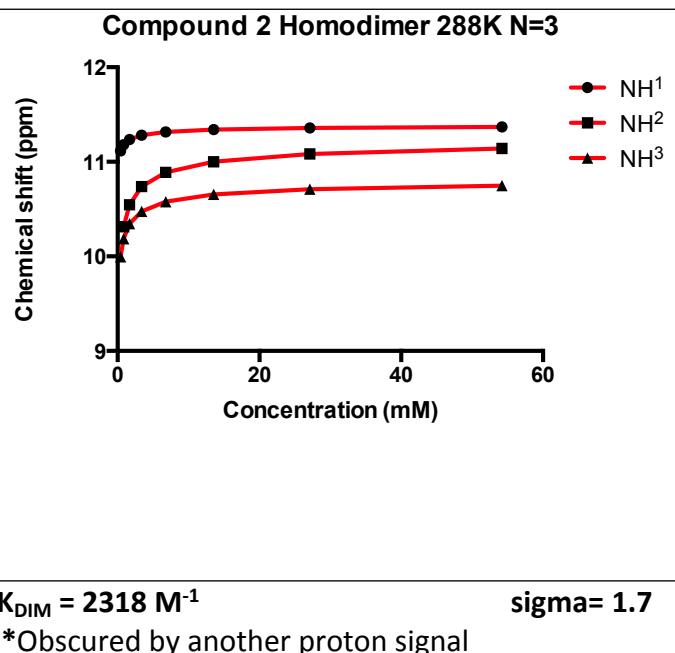


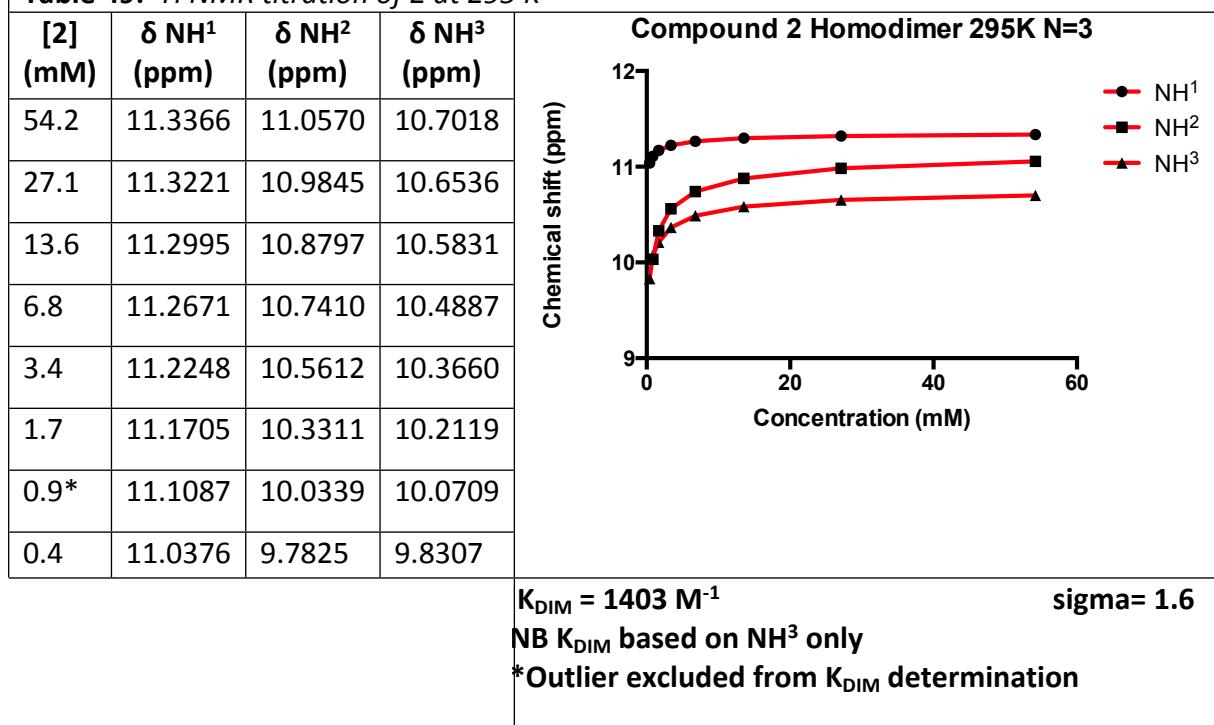
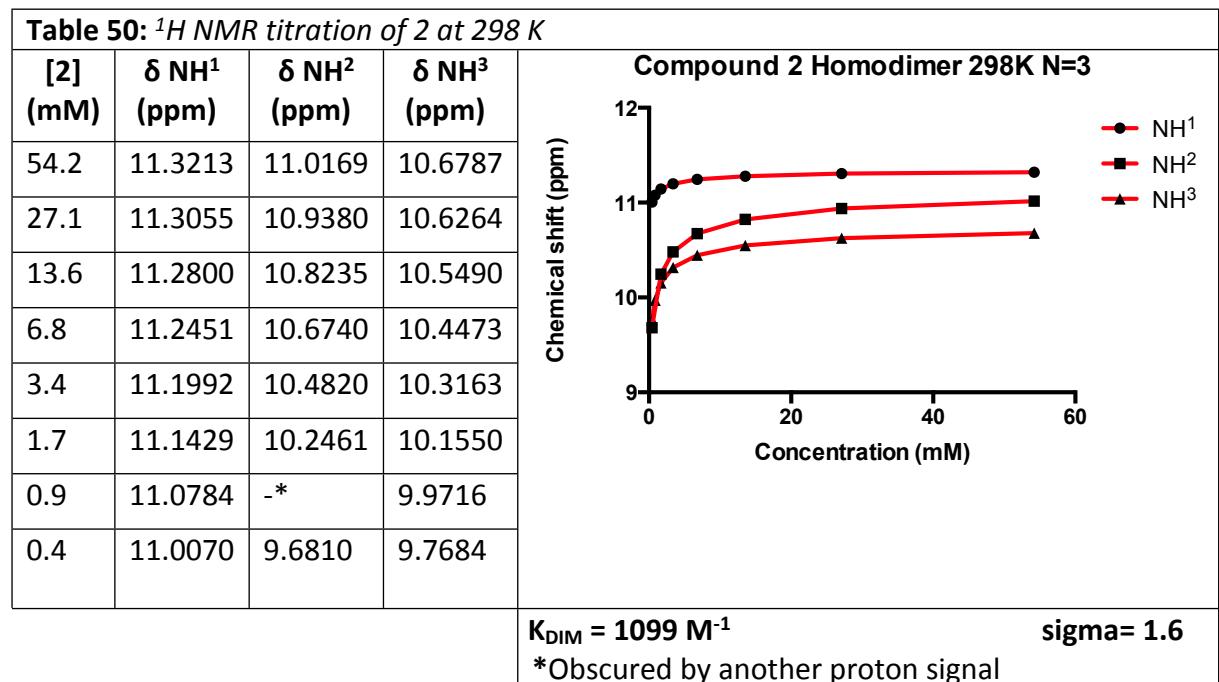
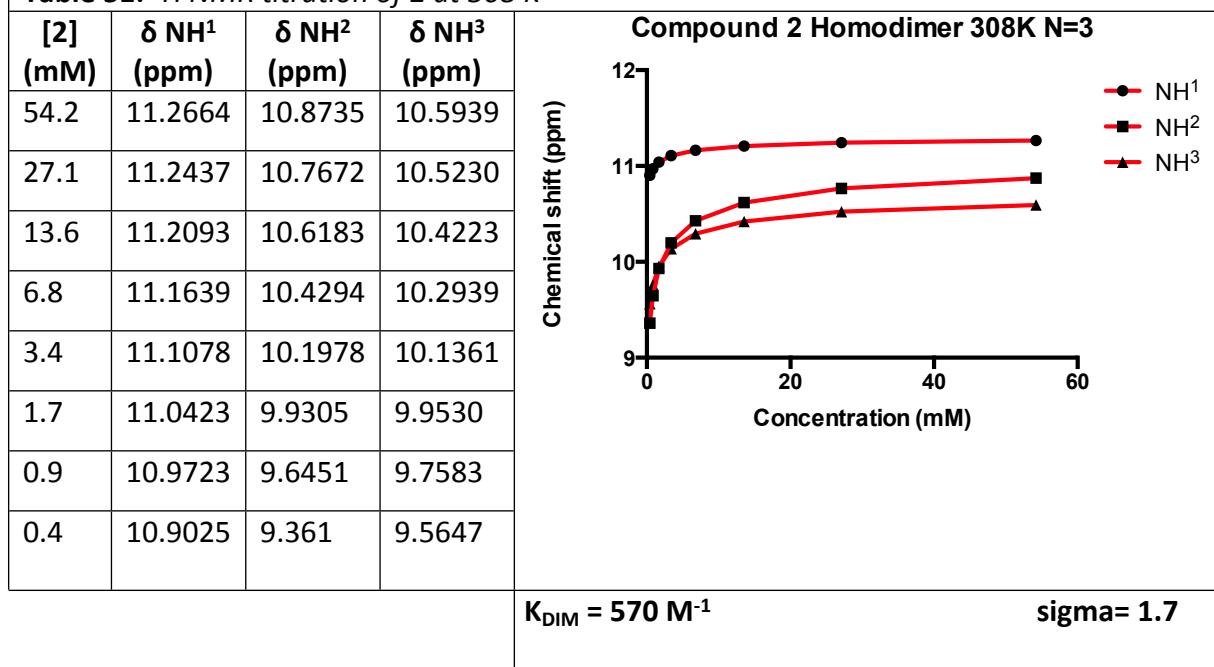
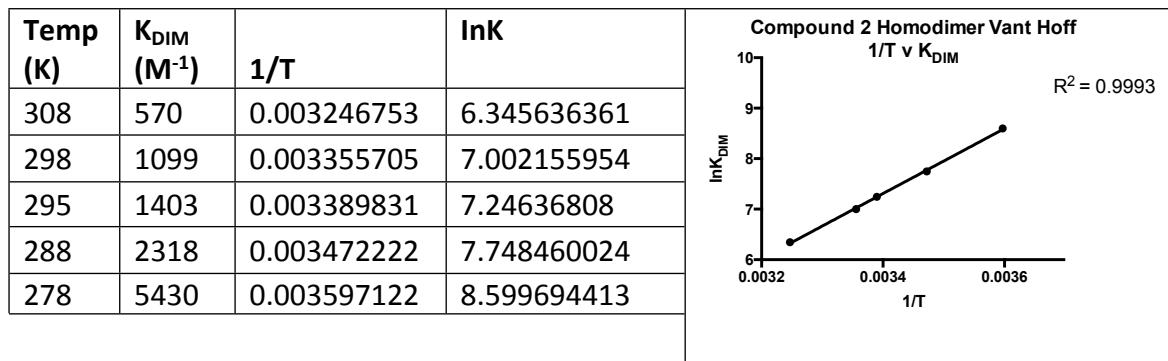
Table 49: ^1H NMR titration of 2 at 295 K**Table 50:** ^1H NMR titration of 2 at 298 K

Table 51: ^1H NMR titration of 2 at 308 K



2.3.4. Compound 2 Homodimer VT ^1H NMR Summary and Van't Hoff Plots

Table 52: Summary of compound 2 Homodimer Experiment 1 and Van't Hoff Plot

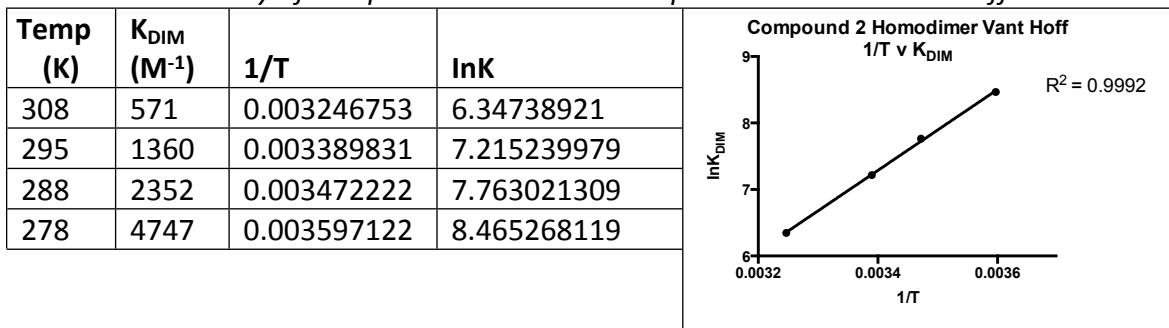


Table 53: Summary of compound 2 Homodimer Experiment 2 and Van't Hoff Plot

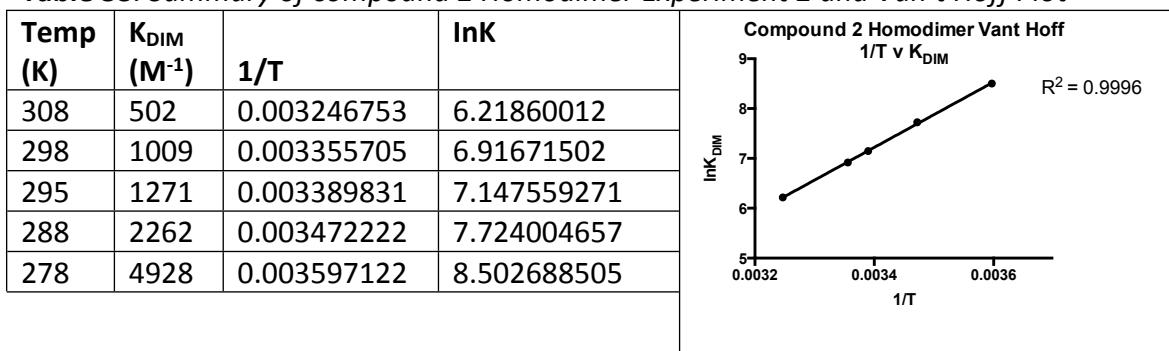


Table 54: Summary of compound 2 Homodimer Experiment 3 and Van't Hoff Plot

Table 55: Summary of thermodynamic parameters for compound 2 homodimer

Expt	ΔH (kcal mol $^{-1}$)	$-\Delta S^{295 \text{ K}}$ (kcal mol $^{-1}$)	ΔG (kcal mol $^{-1}$)
N1	- 12.1	7.8	- 4.3
N2	- 13.0	8.4	- 4.6
N3	- 12.8	8.5	- 4.3
Average	- 12.6	8.4	- 4.4

2.4. 11:1 Heterodimer VT ^1H NMR Titration Experiment

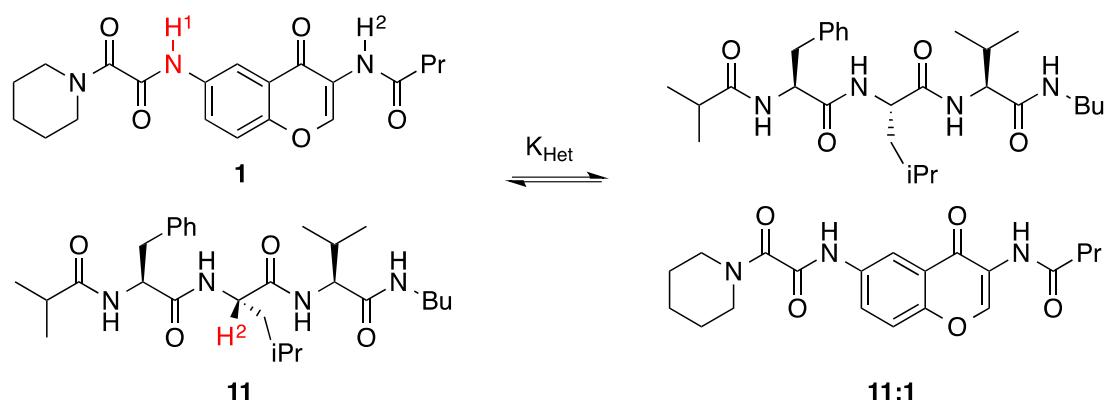


Figure 5: Heterodimerisation of 1 and 11. Resonances used to determine binding constants are highlighted in red.

N.B. K_{HET} was calculated using NH^1 and H^2 . The shift of NH^2 is included for completeness

2.4.1 11:1 Heterodimer ^1H NMR Titration Experiment 1

Peptide concentration: 4 mM

Table 56: 11:1 Heterodimer 1H NMR titration at 278 K

[1] (mM)	δ NH ¹ (ppm)	δ NH ² (ppm)	δ H ² (ppm)
49.1	10.5411	9.0581	5.0745
36.8	10.4644	9.0234	5.0581
27.6	10.4024	8.9899	5.0347
20.7	10.3342	8.9575	5.0240
15.5	10.2651	8.9323	5.0043
11.7	10.1969	8.9150	4.9798
8.7	10.1427	8.9013	4.9571
4.9*	10.0997	8.8941	4.9347
3.7	10.0422	8.8867	4.8776
2.8	10.0322	8.8973	4.8540
2.1	10.0212	8.8943	4.8345
1.6	10.0233	8.9067	4.8153
1.2	10.0189	8.9034	4.8037
0	-	-	4.7381

11:1 Heterodimer 278K

Chemical shift (ppm)

Concentration (mM)

Table 57: 11:1 Heterodimer ^1H NMR titration at 288 K

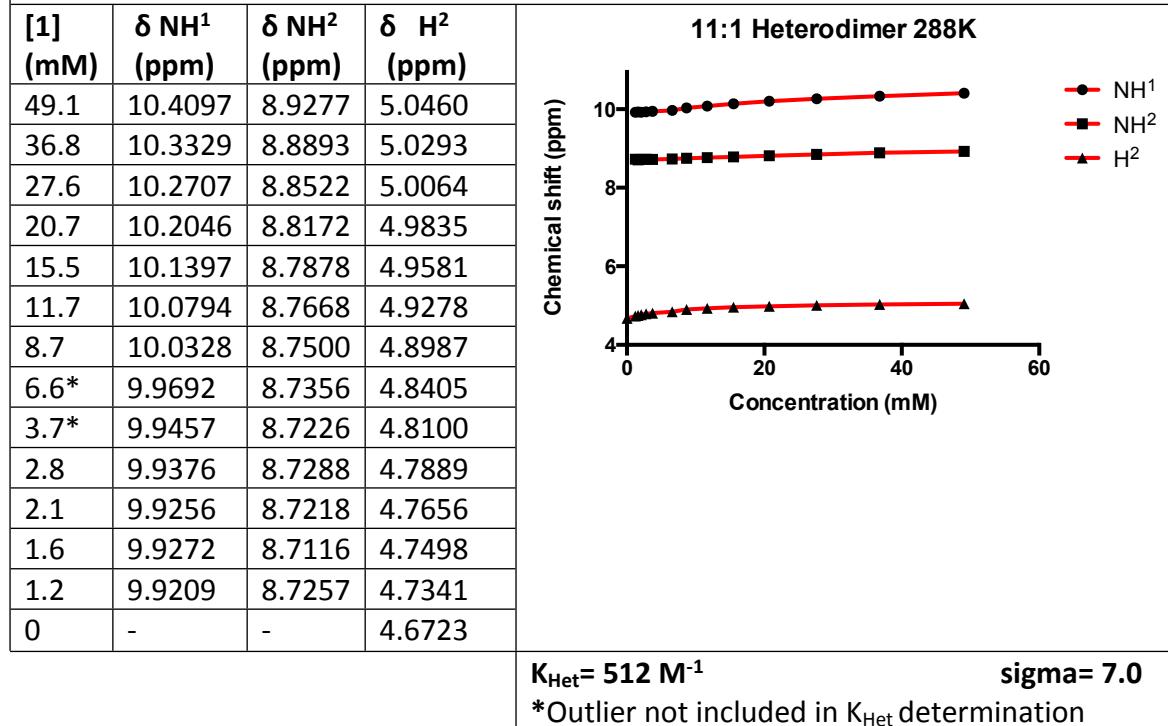


Table 58: 11:1 Heterodimer ^1H NMR titration at 295 K

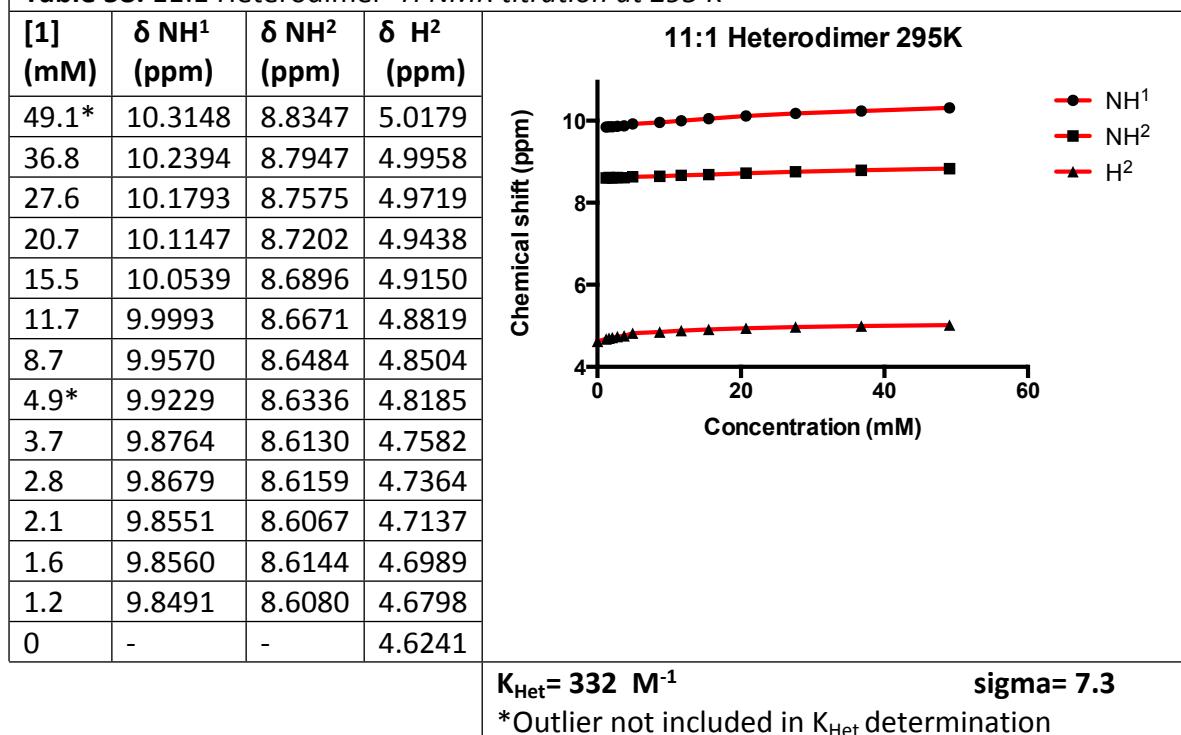
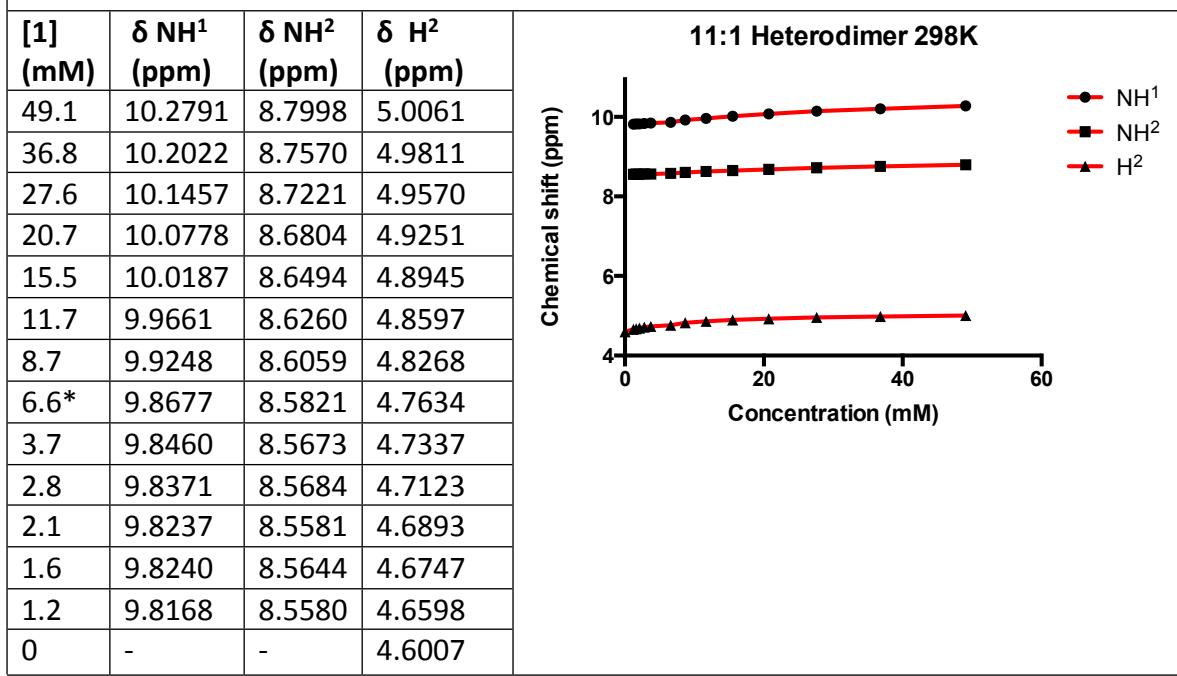
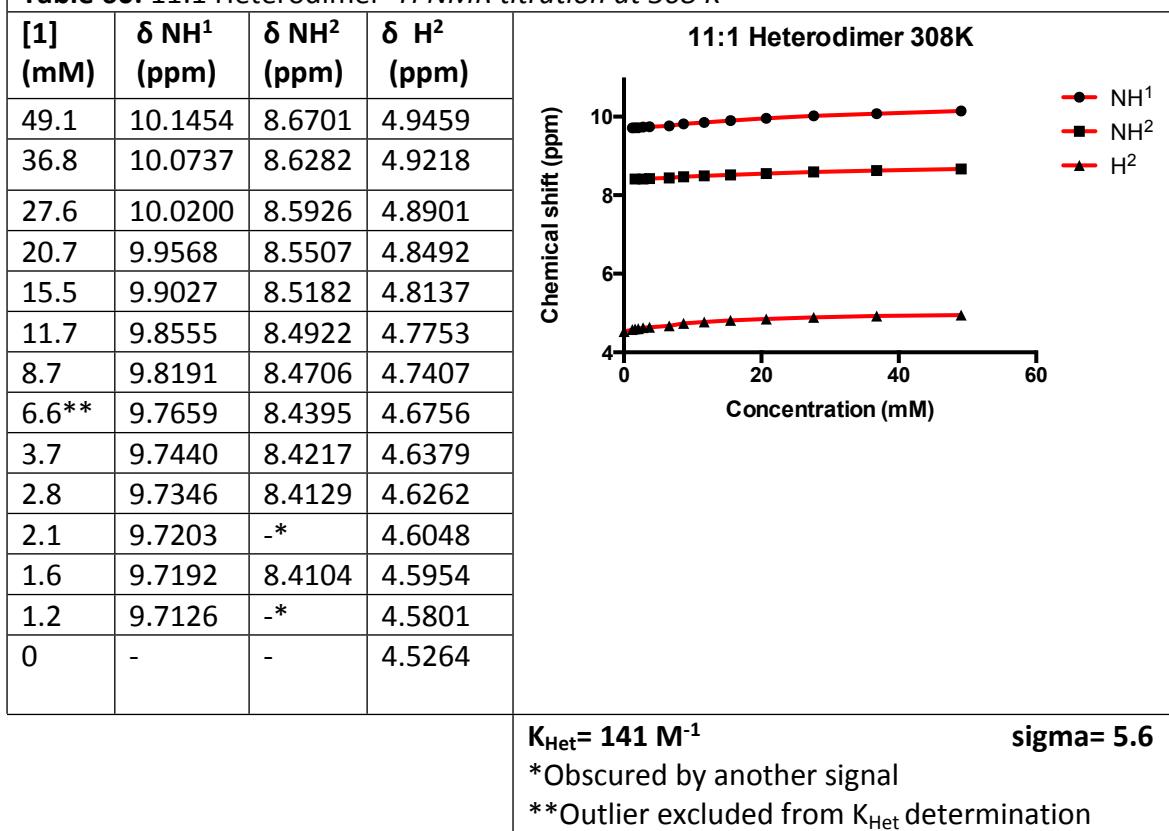


Table 59: 11:1 Heterodimer 1H NMR titration at 298 K

$$K_{\text{Het}} = 272 \text{ M}^{-1} \quad \text{sigma} = 6.8$$

*Outlier excluded from K_{Het} determination

Table 60: 11:1 Heterodimer 1H NMR titration at 308 K

$$K_{\text{Het}} = 141 \text{ M}^{-1} \quad \text{sigma} = 5.6$$

*Obscured by another signal

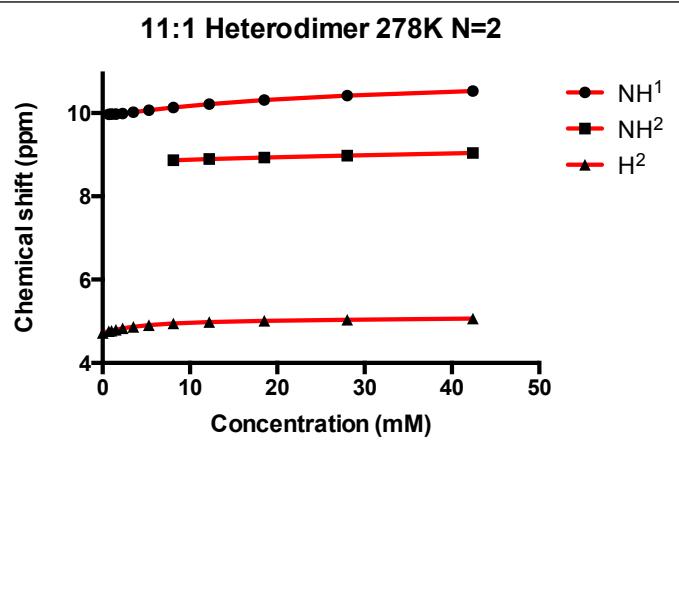
**Outlier excluded from K_{Het} determination

2.4.2. 11:1 Heterodimer ^1H NMR Titration Experiment 2

Peptide concentration: 4.3 mM

Table 61: 11:1 Heterodimer ^1H NMR titration at 278 K

[1] (mM)	$\delta \text{ NH}^1$ (ppm)	$\delta \text{ NH}^2$ (ppm)	$\delta \text{ H}^2$ (ppm)
42.4	10.5293	9.0430	5.0649
28.0	10.4168	8.9813	5.0377
18.5	10.3134	8.9344	5.0118
12.2	10.2139	8.8966	4.9801
8.1	10.1323	8.8663	4.9491
5.3	10.0673	-*	4.9057
3.5	10.0200	-*	4.8672
2.3	9.9901	-*	4.8290
1.5	9.9778	-*	4.7984
1.0	9.9747	-*	4.7739
0.7	9.9689	-*	4.7586
0		-	4.7215



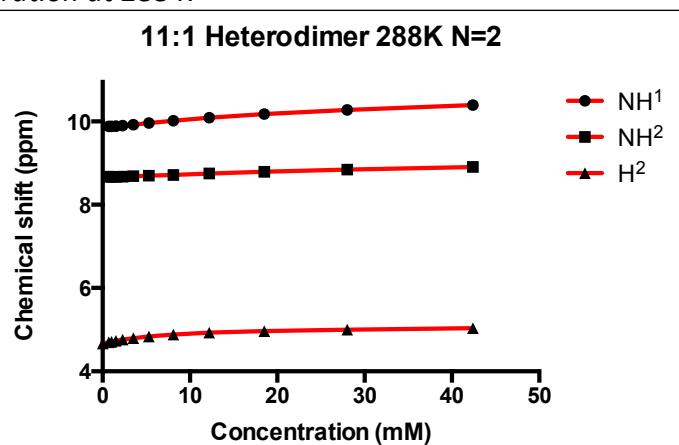
$$K_{\text{Het}} = 838 \text{ M}^{-1}$$

*Broadened signal

sigma = 9.8

Table 62: 11:1 Heterodimer ^1H NMR titration at 288 K

[1] (mM)	$\delta \text{ NH}^1$ (ppm)	$\delta \text{ NH}^2$ (ppm)	$\delta \text{ H}^2$ (ppm)
42.4	10.3939	8.9099	5.0342
28.0	10.2815	8.8443	5.0009
18.5	10.1814	8.7916	4.9671
12.2	10.0918	8.7504	4.9278
8.1	10.0177	8.7166	4.8838
5.3	9.9619	8.6962	4.8397
3.5	9.9253	8.6843	4.7977
2.3	9.9021	8.6759	4.7606
1.5	9.8875	8.6694	4.7306
1.0	9.8835	8.6722	4.7095
0.7	9.8824	8.6748	4.6950
0	-	-	4.6673



$$K_{\text{Het}} = 501 \text{ M}^{-1}$$

sigma = 5.6

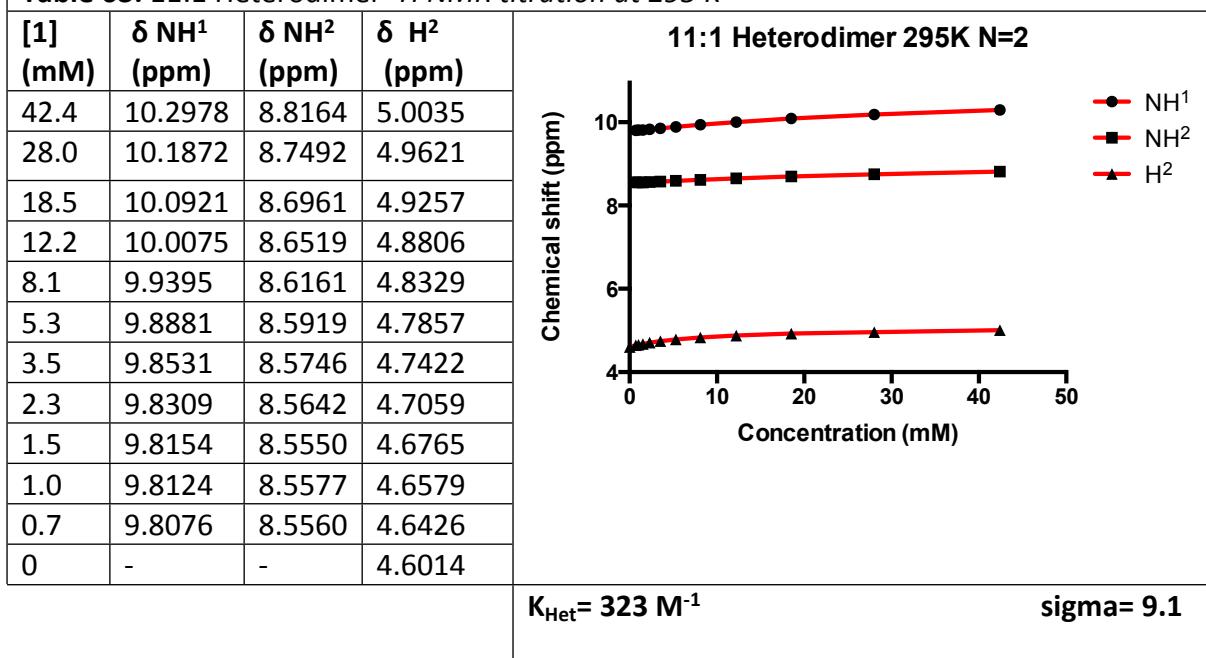
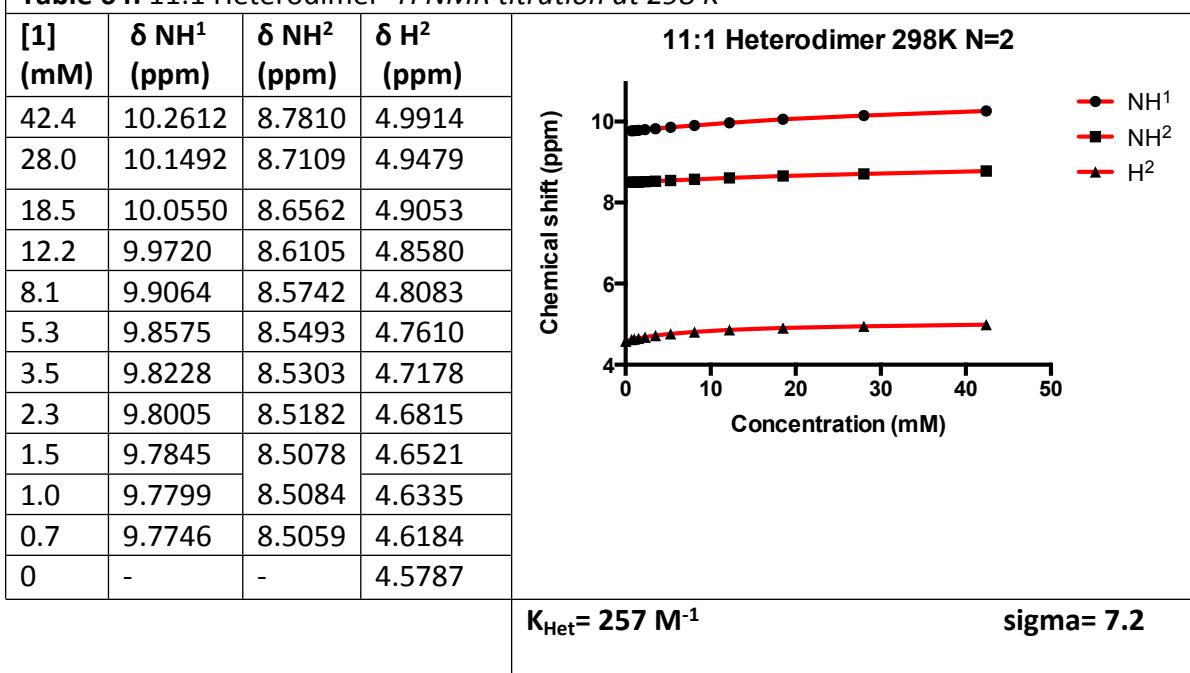
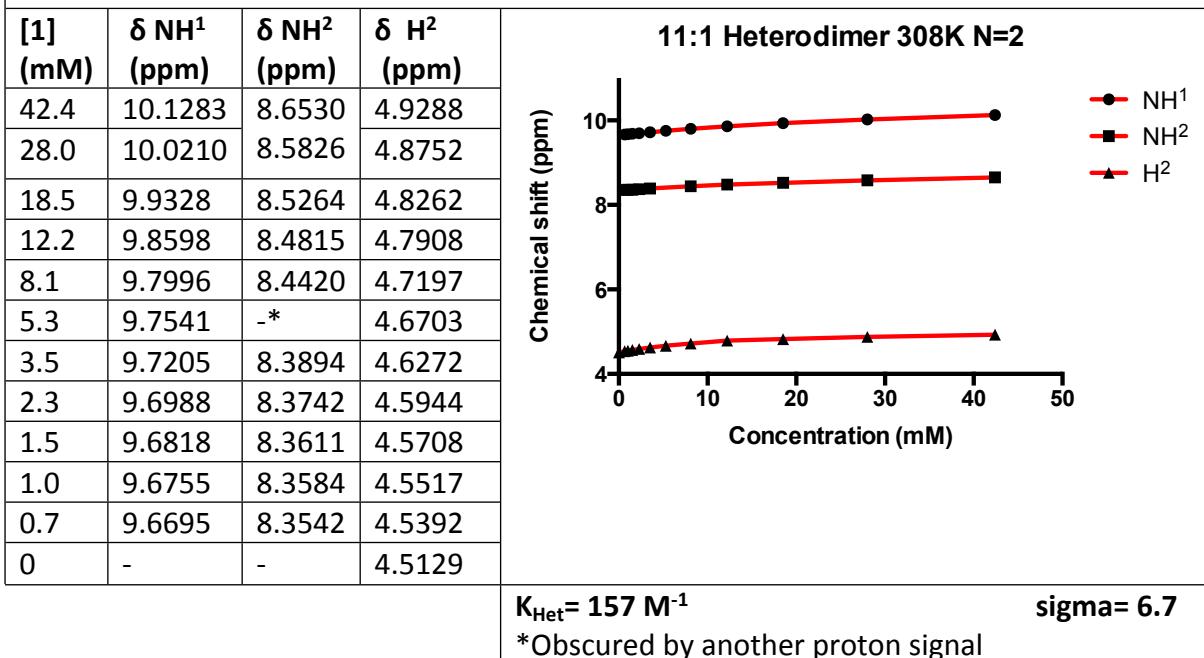
Table 63: 11:1 Heterodimer ^1H NMR titration at 295 K**Table 64:** 11:1 Heterodimer ^1H NMR titration at 298 K

Table 65: 11:1 Heterodimer ^1H NMR titration at 308 K



2.4.3. 11:1 Heterodimer ^1H NMR Titration Experiment 3

Peptide concentration: 3.3 mM

Table 66: 11:1 Heterodimer ^1H NMR titration at 278 K

[1] (mM)	$\delta \text{ NH}^1$ (ppm)	$\delta \text{ NH}^2$ (ppm)	$\delta \text{ H}^2$ (ppm)
30.3	10.4276	8.9756	5.0376
22.7	10.3525	8.9317	5.0169
17.0	10.2685	8.8932	4.9956
12.8	10.1976	8.8615	4.9703
9.6	10.1474	8.8223	4.9458
7.2	10.0888	8.7983	4.9170
5.4	10.0366	8.4192	4.8864
4.1	9.9959	-*	4.8606
3.0	9.9663	-*	4.8301
2.3	9.9487	-*	4.8044
1.7	-*	-*	4.7806
1.3	-*	-*	4.7612
0	-	-	4.6908

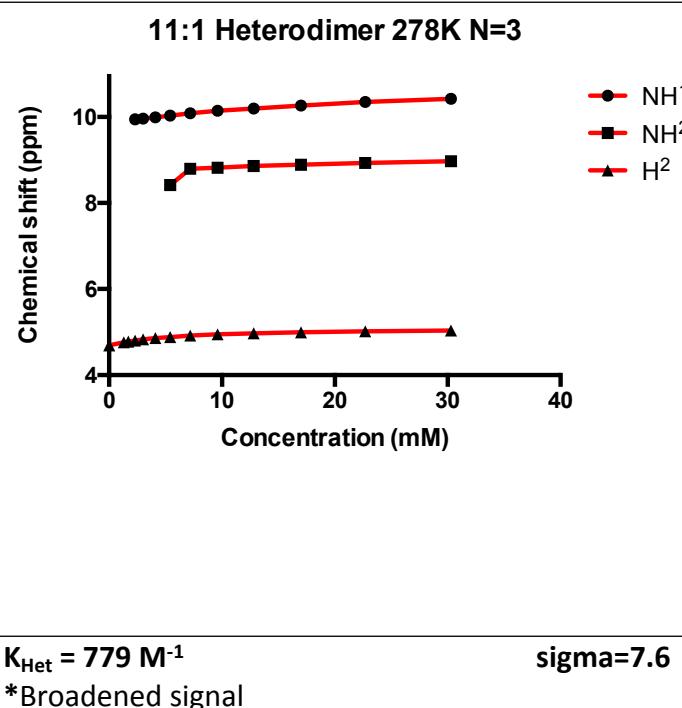


Table 67: 11:1 Heterodimer ^1H NMR titration at 288 K

[1] (mM)	$\delta \text{ NH}^1$ (ppm)	$\delta \text{ NH}^2$ (ppm)	$\delta \text{ H}^2$ (ppm)
30.3	10.2793	8.8281	4.9952
22.7	10.2100	8.7867	4.9713
17.0	10.1383	8.7475	4.9477
12.8	10.0735	8.7132	4.9128
9.6	10.0179	8.6835	4.8825
7.2	9.9683	8.6602	4.8536
5.4	9.9271	8.6426	4.8189
4.1	9.8958	8.6308	4.7877
3.0	9.8711	8.6194	4.7572
2.3	9.8559	8.6163	4.7311
1.7	9.8438	8.6113	4.7090
1.3	9.8355	8.6083	4.6887
0	-	-	4.6169

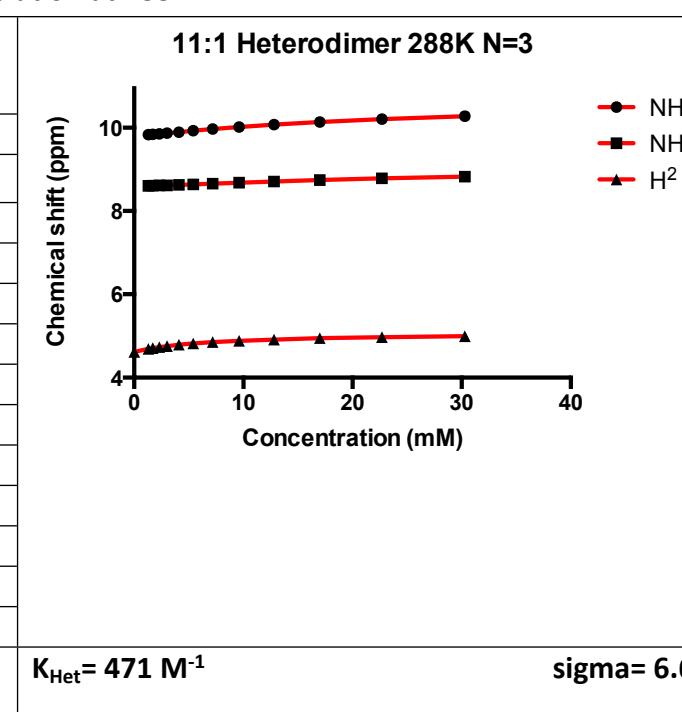


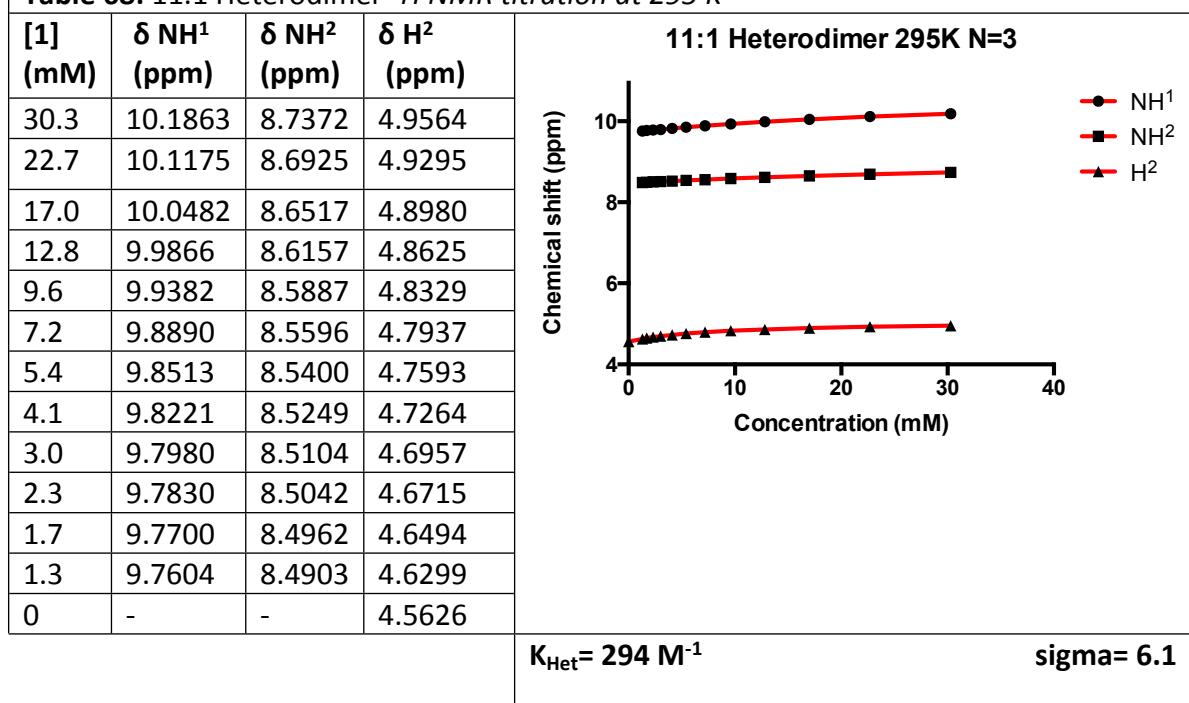
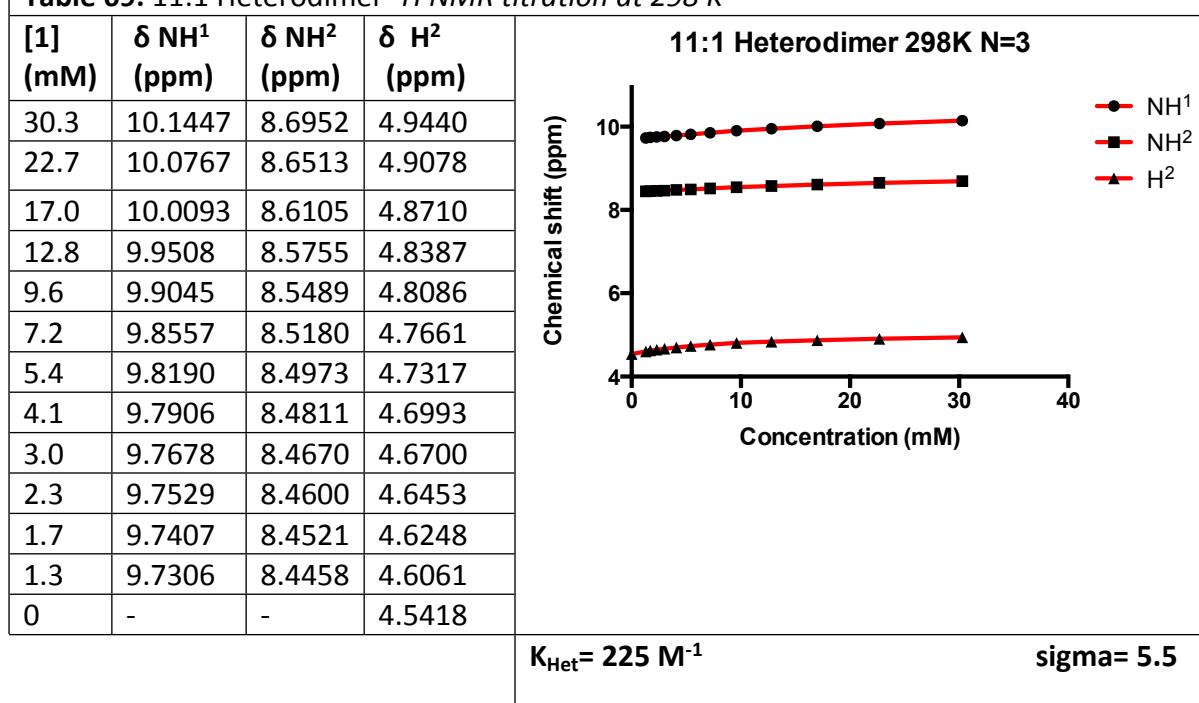
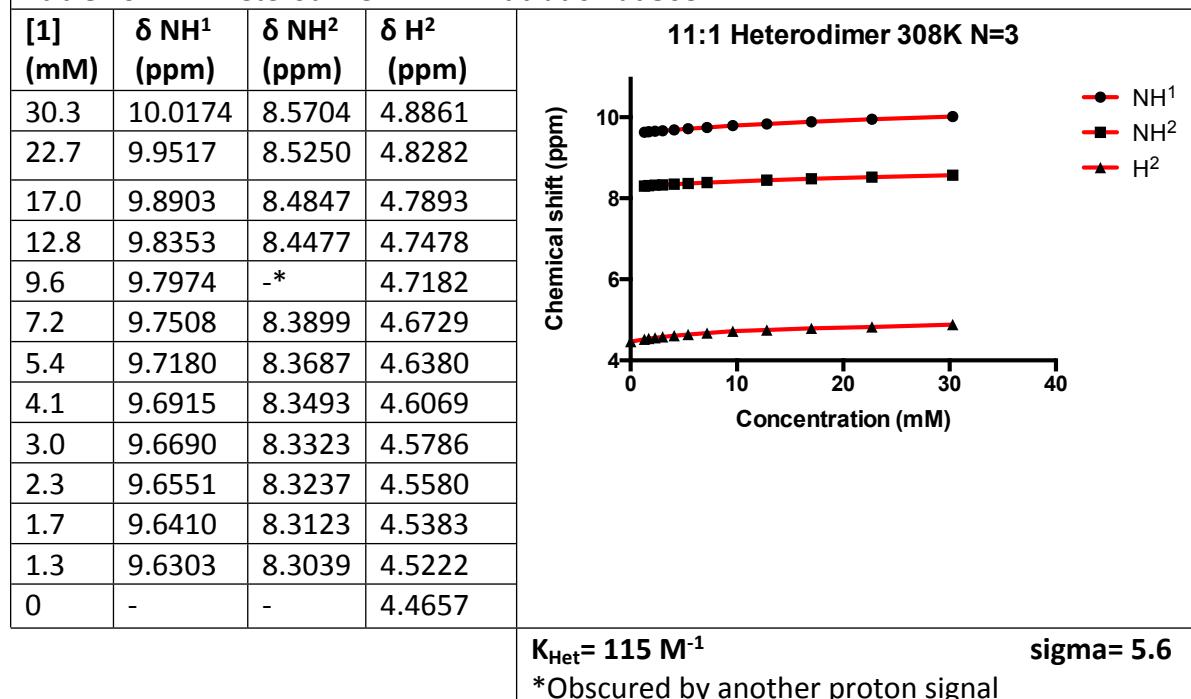
Table 68: 11:1 Heterodimer ^1H NMR titration at 295 K**Table 69:** 11:1 Heterodimer ^1H NMR titration at 298 K

Table 70: 11:1 Heterodimer 1H NMR titration at 308 K

2.4.4. 11:1 Heterodimer VT ^1H NMR Summary and Van't Hoff Plots

Table 71: Summary of 11:1 Heterodimer Experiment 1 and Van't Hoff Plot

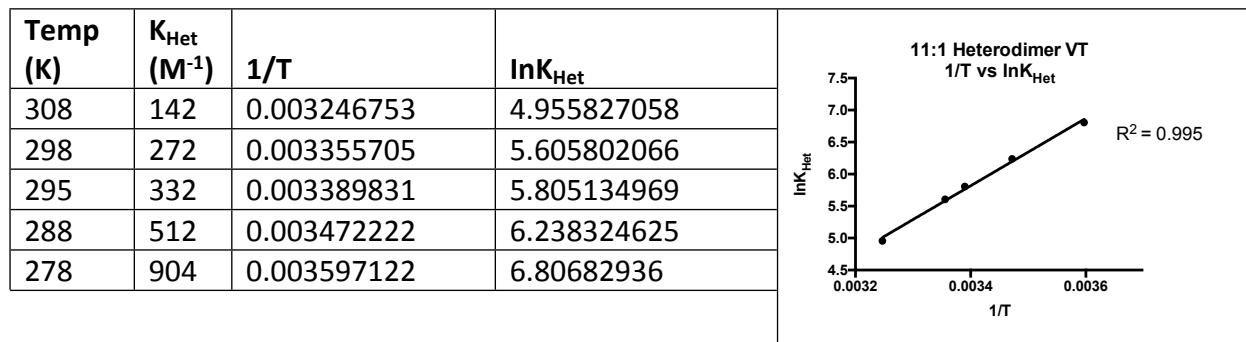


Table 72: Summary of 11:1 Heterodimer Experiment 2 and Van't Hoff Plot

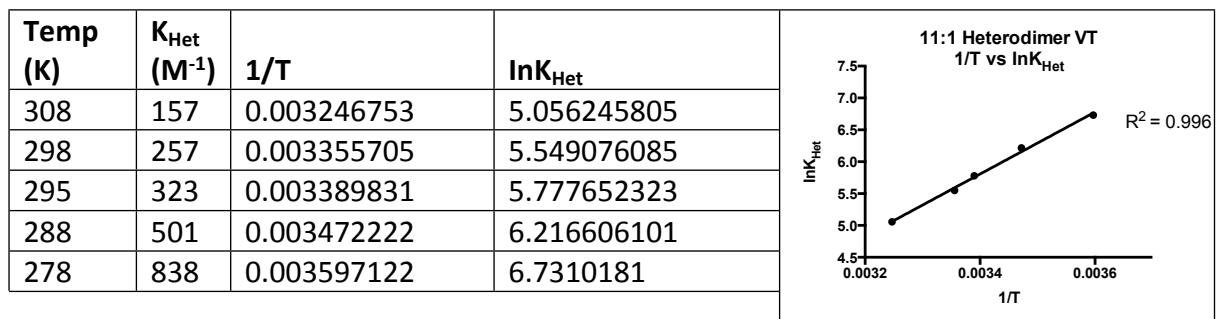


Table 73: Summary of 11:1 Heterodimer Experiment 3 and Van't Hoff Plot

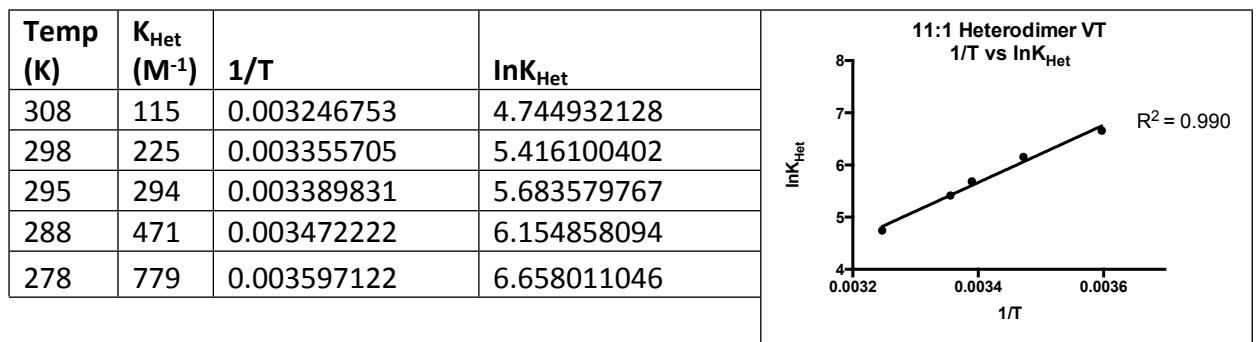


Table 74: Summary of thermodynamic parameters for 11:1 heterodimer

Expt	ΔH (kcal mol $^{-1}$)	$-T\Delta S^{295 \text{ K}}$ (kcal mol $^{-1}$)	ΔG (kcal mol $^{-1}$)
N1	- 10.9	7.6	- 3.3
N2	- 9.7	6.3	- 3.1
N3	- 10.5	7.1	- 3.4
Average	- 10.4	7.0	- 3.3

2.5. 11:2 Heterodimer VT ^1H NMR Titration Experiment

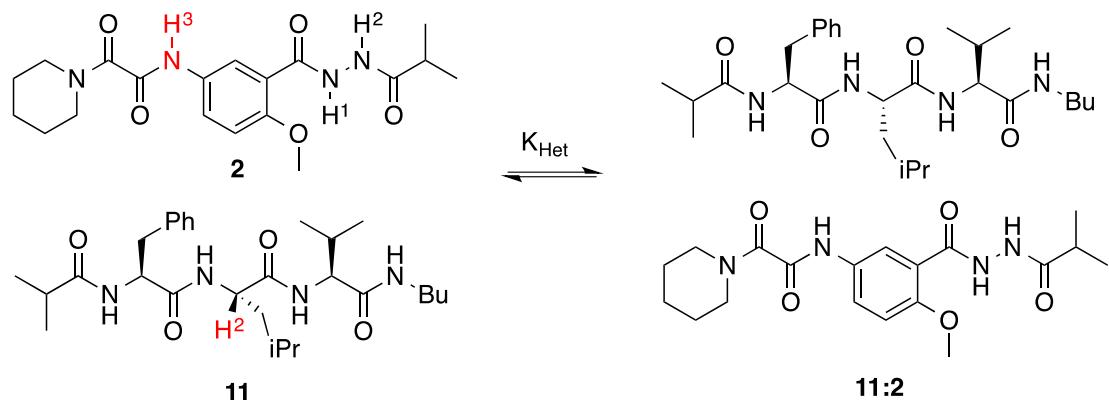


Figure 6: Heterodimerisation of 2 and 11. Resonances used to determine binding constants are highlighted in red.

N.B. K_{Het} was calculated on using NH^3 and H^2 . Shifts NH^1 and NH^2 are included for completeness

2.5.1. 11:2 Heterodimer ^1H NMR Titration Experiment 1

Peptide concentration: 3.1 mM

Table 75: 11:2 Heterodimer ^1H NMR titration at 278 K

[2] (mM)	δNH^1 (ppm)	δNH^2 (ppm)	δNH^3 (ppm)	δH^2 (ppm)
34.0	11.3932	11.2824	10.7386	5.0457
25.5	11.3870	11.2795	10.7124	5.0387
19.1	11.3783	11.2777	10.6782	5.0298
14.3	11.3705	-*	10.6443	5.0200
10.8	11.3605	-*	10.6006	5.0075
8.1	11.3493	-*	10.5508	4.9874
6.1	11.3372	-*	10.4955	4.9725
4.5	11.3259	-*	10.4395	4.9524
3.4	11.3153	-*	10.3878	4.9266
2.6	11.3076	-*	10.3444	4.8988
1.9	11.3011	-*	10.3155	4.8644
1.4	11.2955	-*	10.2884	4.8307
0	-	-	-	4.4425

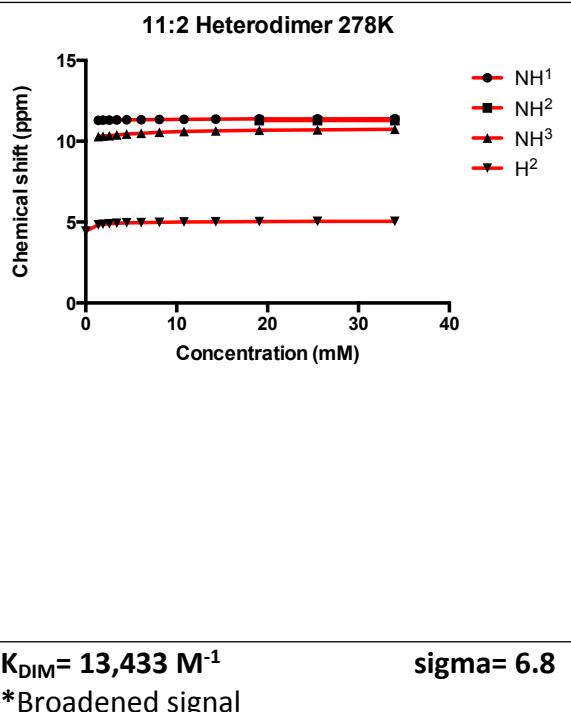


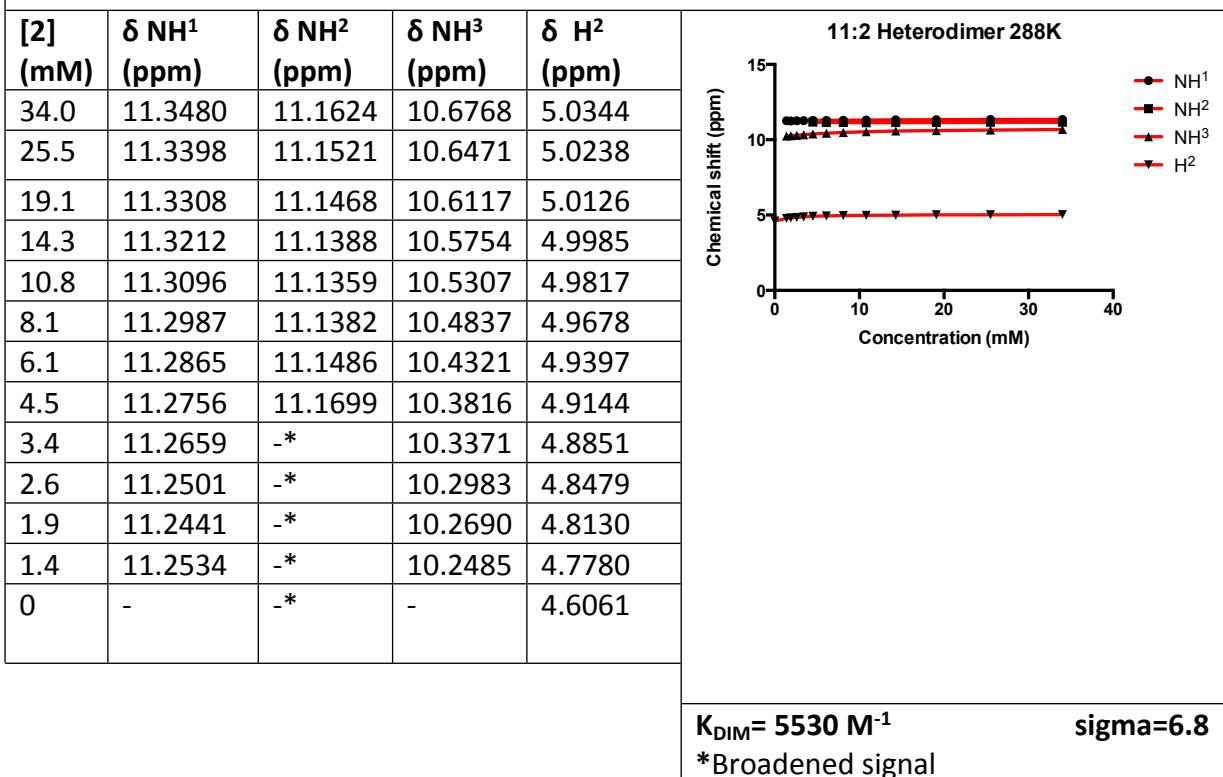
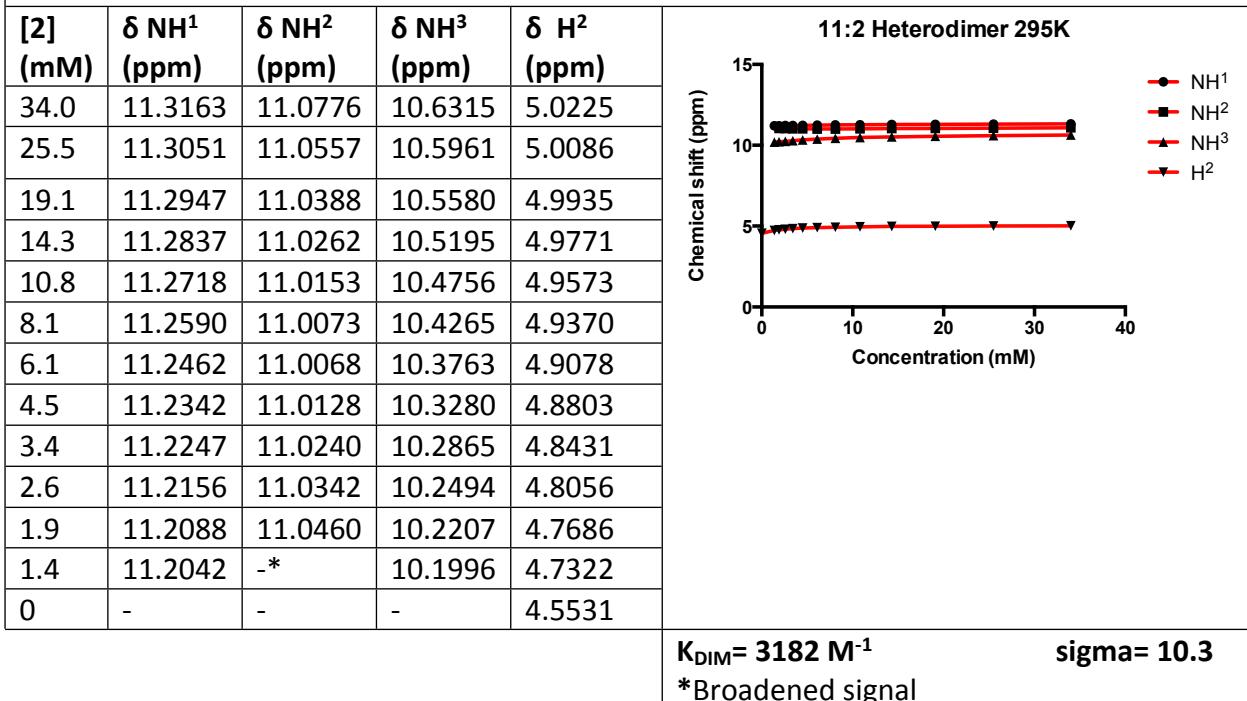
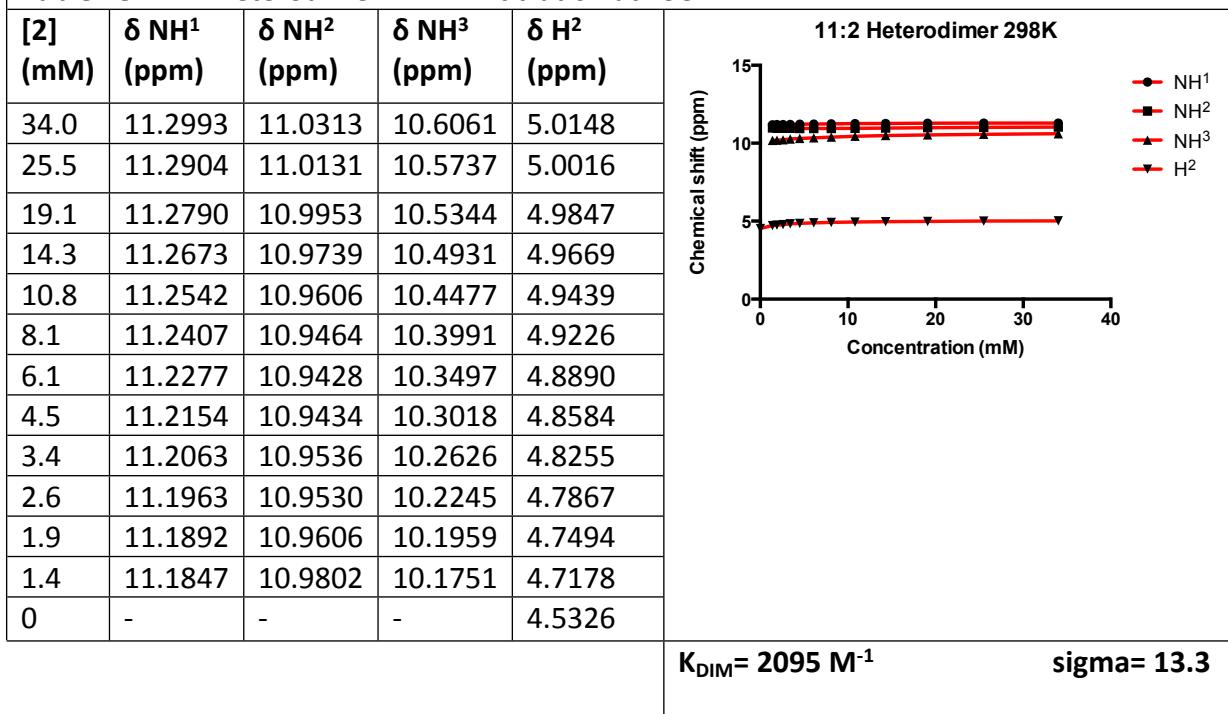
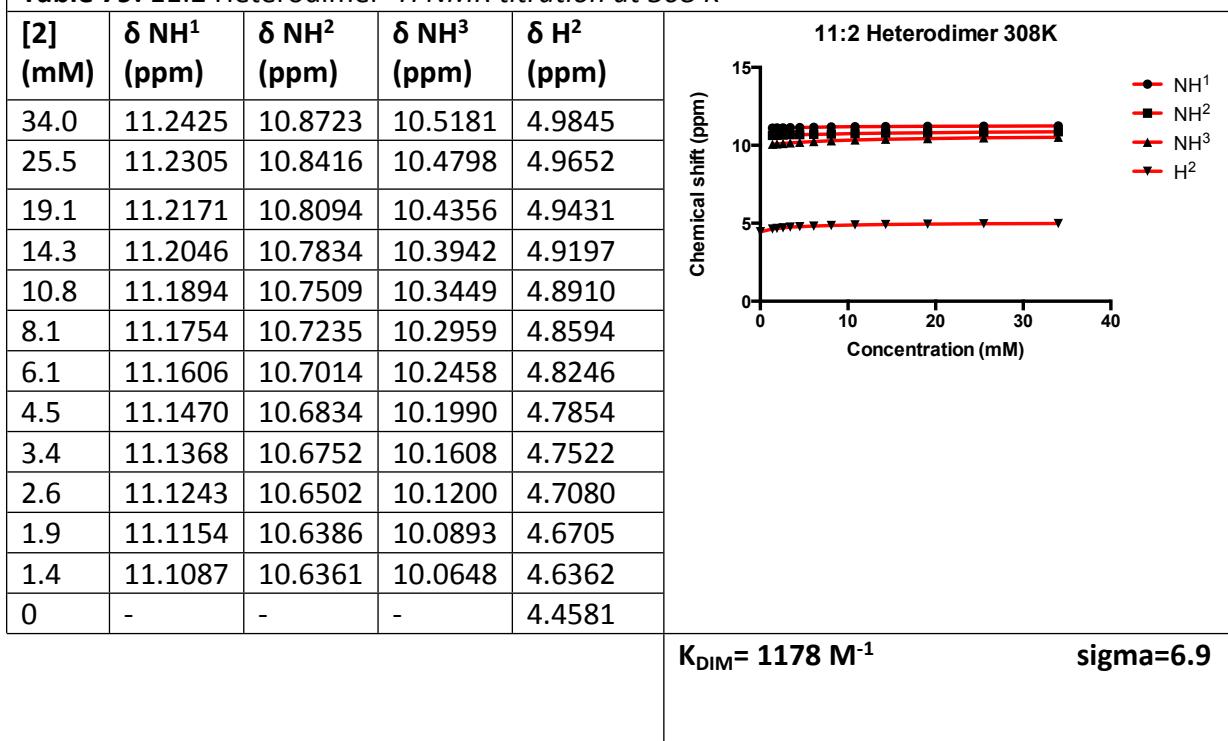
Table 76: 11:2 Heterodimer 1H NMR titration at 288 K**Table 77:** 11:2 Heterodimer 1H NMR titration at 295 K

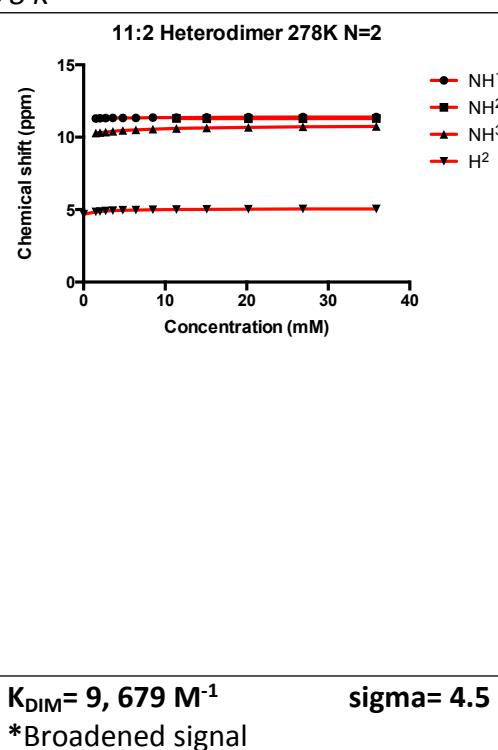
Table 78: 11:2 Heterodimer ^1H NMR titration at 298 K**Table 79:** 11:2 Heterodimer ^1H NMR titration at 308 K

2.5.2. 11:2 Heterodimer ^1H NMR Titration Experiment 2

Peptide concentration: 3.1 mM

Table 80: 11:2 Heterodimer ^1H NMR titration at 278 K

[2] (mM)	δNH^1 (ppm)	δNH^2 (ppm)	δNH^3 (ppm)	δH^2 (ppm)
35.9	11.3964	11.2951	10.7453	5.0465
26.9	11.3881	11.2854	10.7166	5.0386
20.2	11.3807	11.2817	10.6863	5.0312
15.1	11.3721	11.2837	10.6492	5.0207
11.4	11.3612	11.2936	10.6031	5.0087
8.5	11.3517	-*	10.5603	4.9938
6.4	11.3345	-*	10.5124	4.9739
4.8	11.3305	-*	10.4680	4.9580
3.6	11.3255	-*	10.4121	4.9323
2.7	11.3170	-*	10.3535	4.9040
2.0	11.3106	-*	10.3194	4.8718
1.5	11.2973	-*	10.2963	4.8413
0	-	-*	-	4.6830

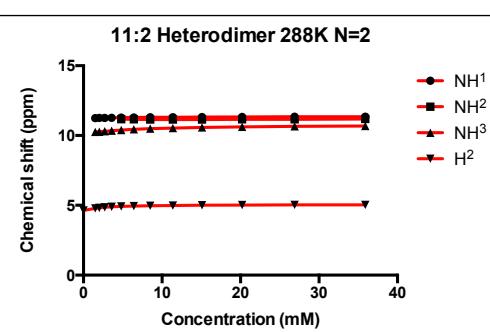


$$K_{\text{DIM}} = 9,679 \text{ M}^{-1} \quad \text{sigma} = 4.5$$

*Broadened signal

Table 81: 11:2 Heterodimer ^1H NMR titration at 288 K

[2] (mM)	δNH^1 (ppm)	δNH^2 (ppm)	δNH^3 (ppm)	δH^2 (ppm)
35.9	11.3490	11.1750	10.6814	5.0343
26.9	11.3415	11.1566	10.6521	5.0251
20.2	11.3335	11.1549	10.6204	5.0153
15.1	11.3229	11.1436	10.5812	5.0008
11.4	11.3109	11.1412	10.5355	4.9835
8.5	11.3002	11.1406	10.4923	4.9657
6.4	11.2879	11.1523	10.4392	4.9434
4.8	11.2777	11.1687	10.3920	4.9178
3.6	11.2684	-*	10.3442	4.8914
2.7**	11.2594	-*	10.3075	4.8562
2.0**	11.2517	-*	10.2724	4.8192
1.5**	11.2465	-*	10.2528	4.7862
0	-	-	-	4.6363



$$K_{\text{DIM}} = 5346 \text{ M}^{-1} \quad \text{sigma} = 3.6$$

*Obscured by another proton signal

**Outlier not included in K_{Het} determination

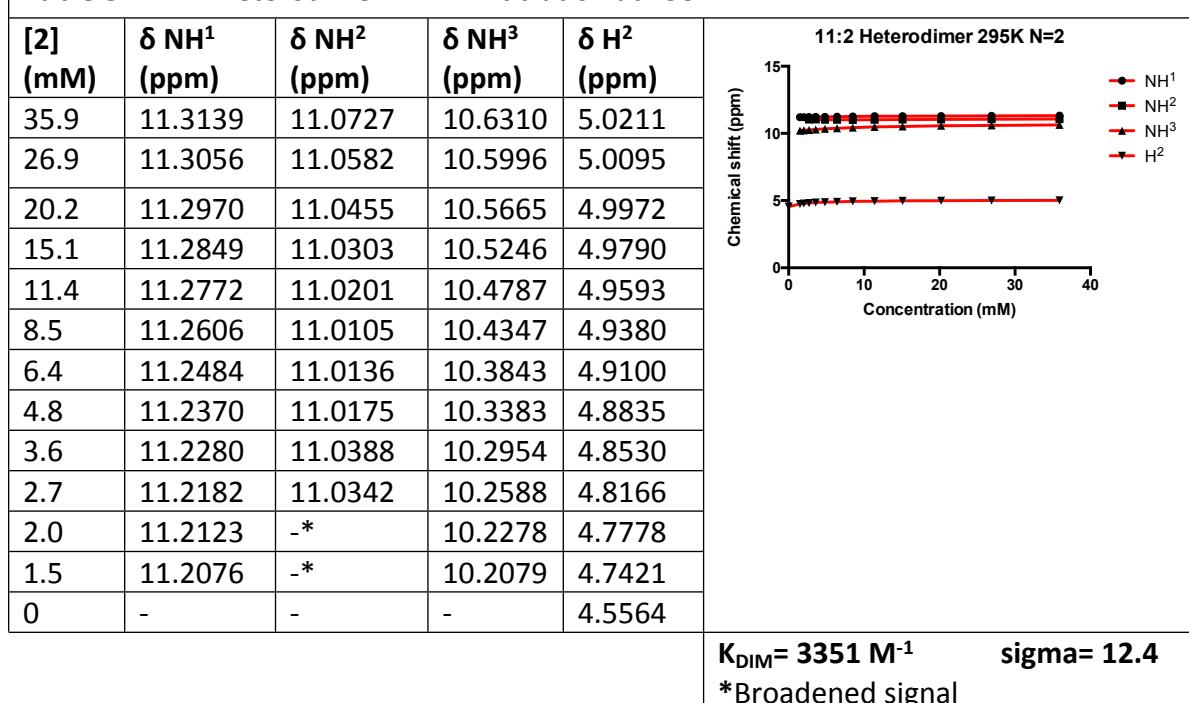
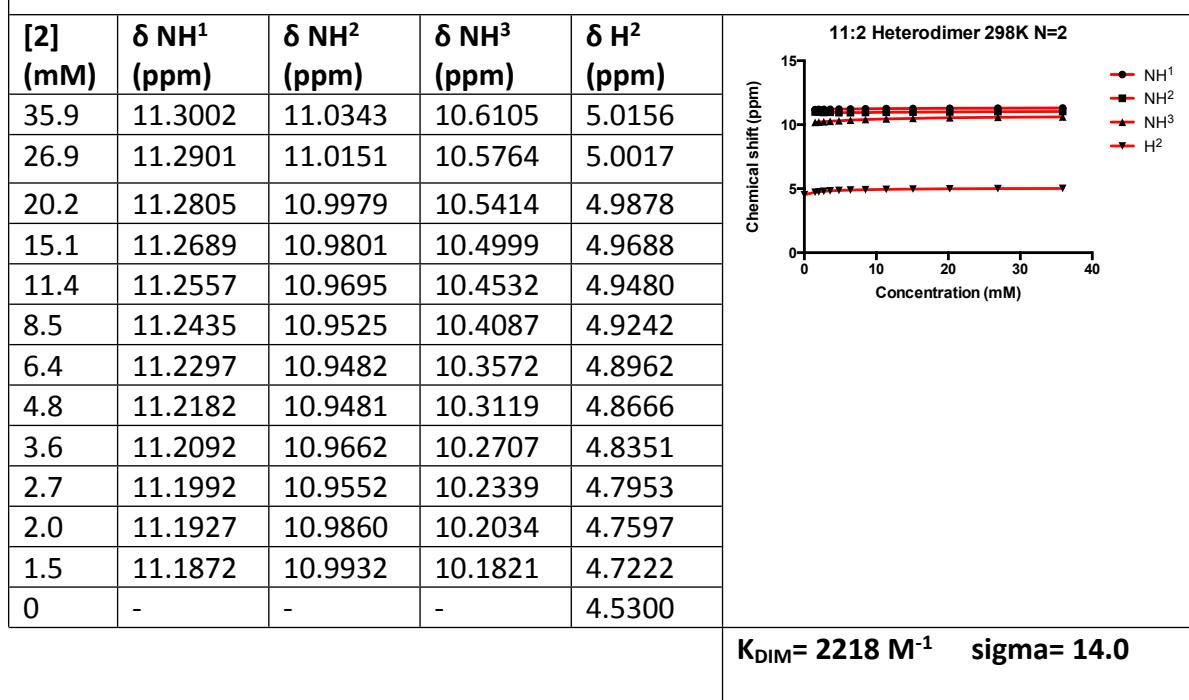
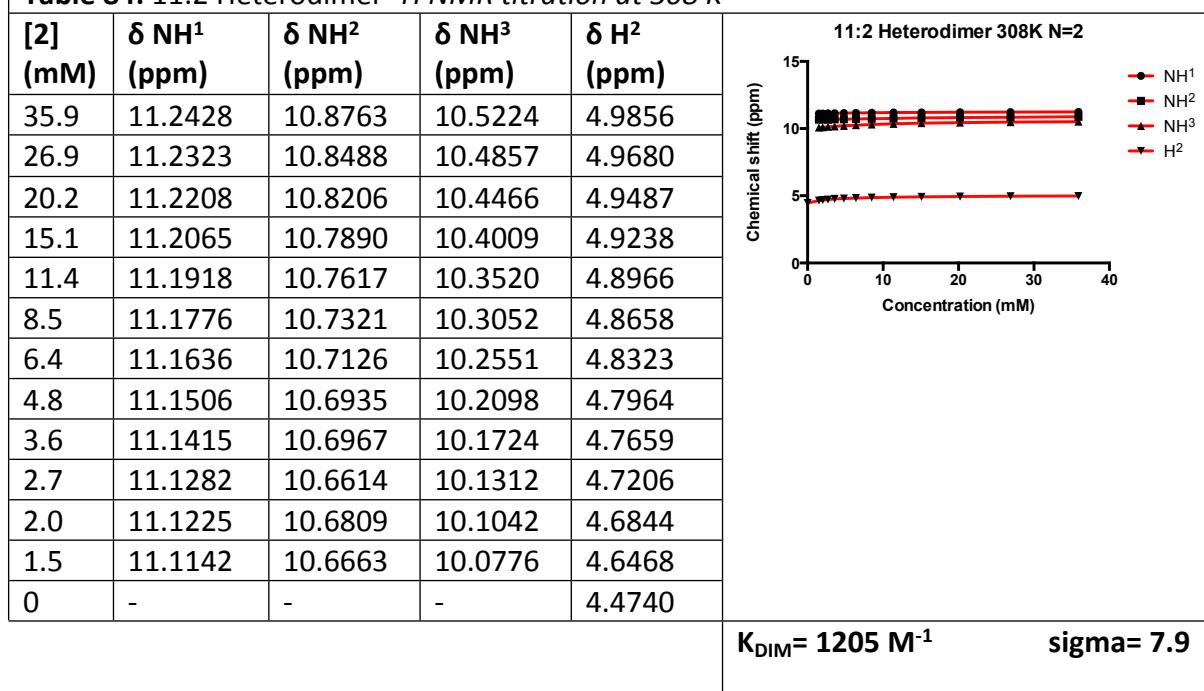
Table 82: 11:2 Heterodimer ^1H NMR titration at 295 K**Table 83:** 11:2 Heterodimer ^1H NMR titration at 298 K

Table 84: 11:2 Heterodimer 1H NMR titration at 308 K

2.5.3. 11:2 Heterodimer ^1H NMR Titration Experiment 3

Peptide concentration: 3.2 mM

Table 85: 11:2 Heterodimer ^1H NMR titration at 278 K

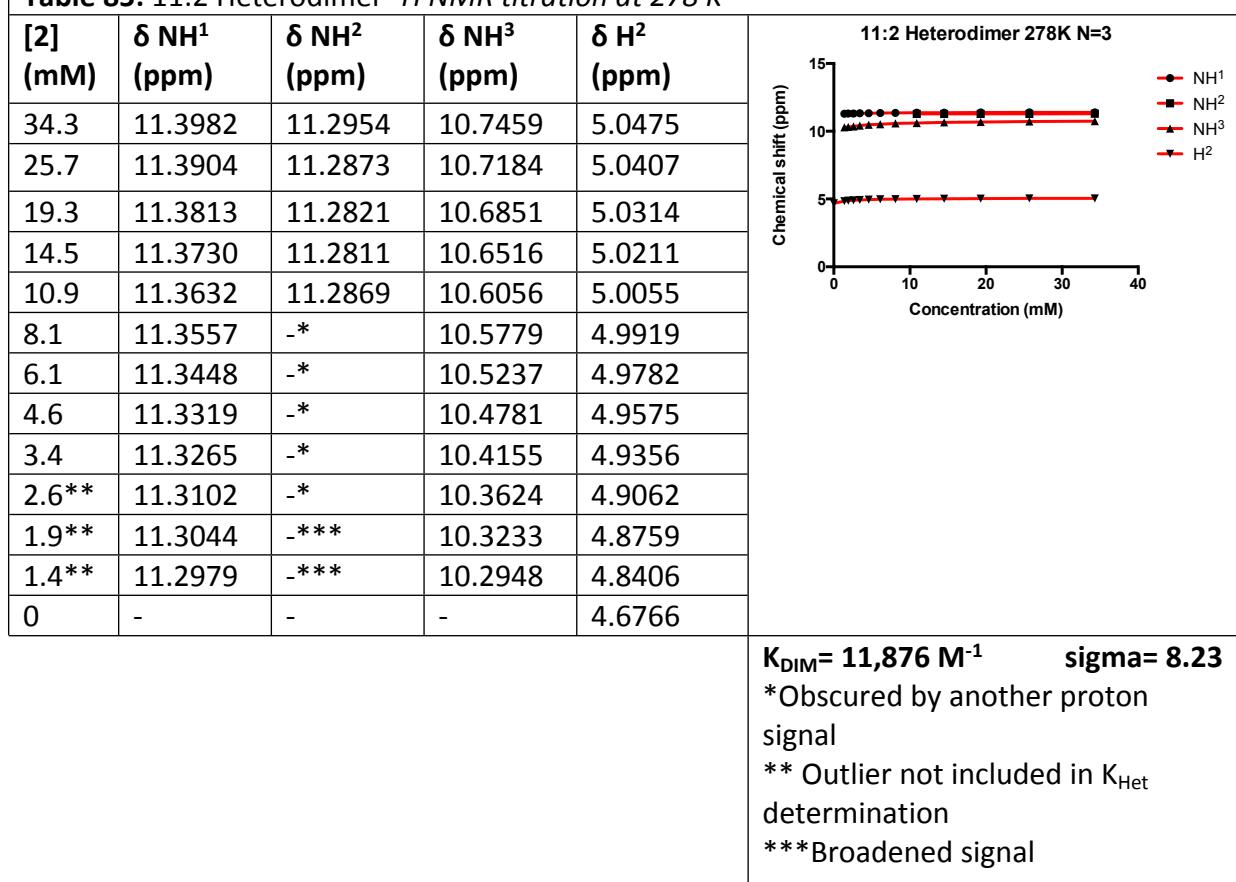


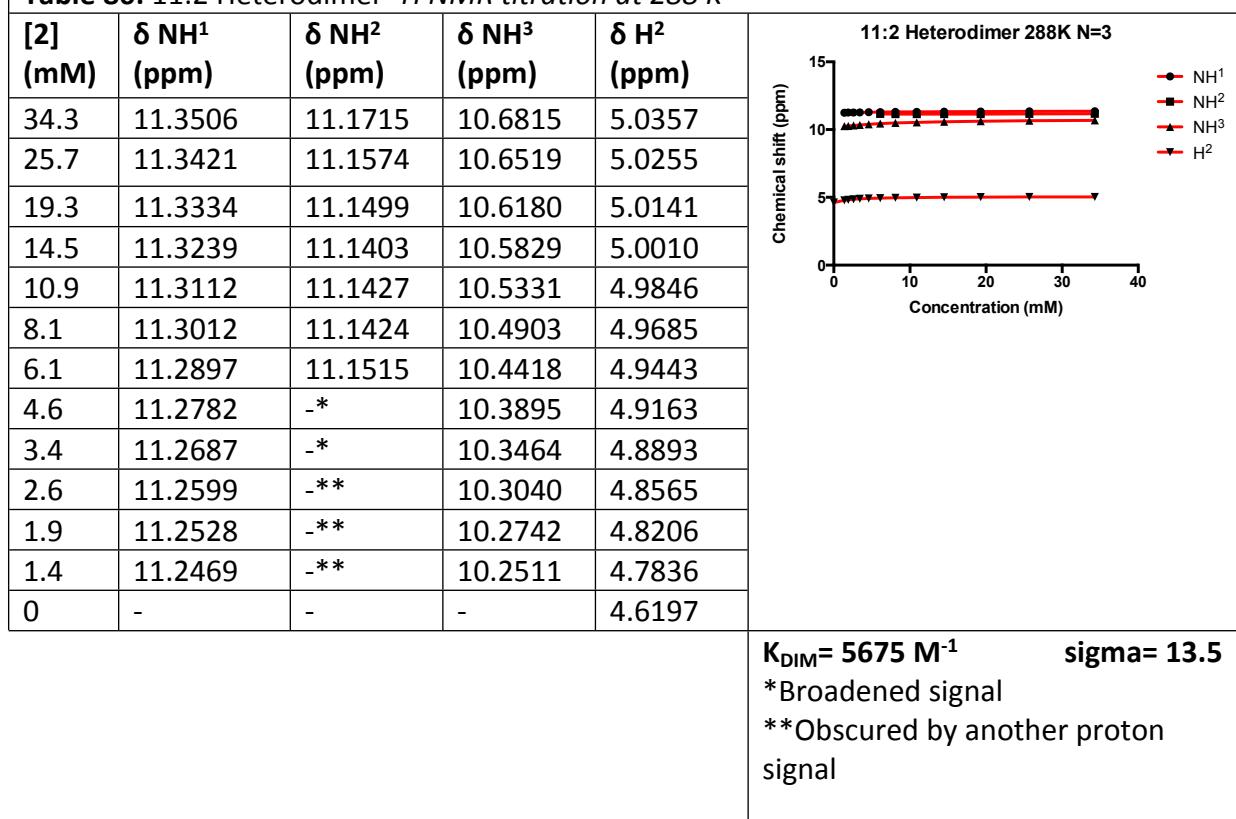
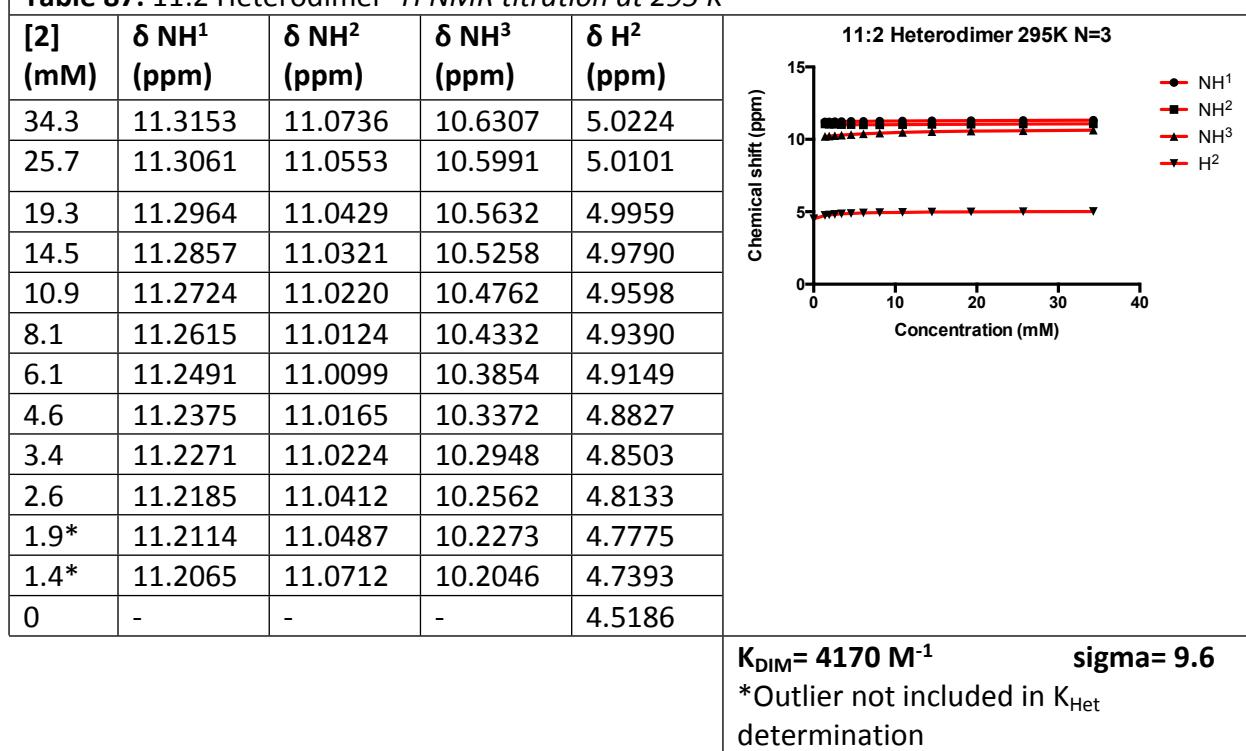
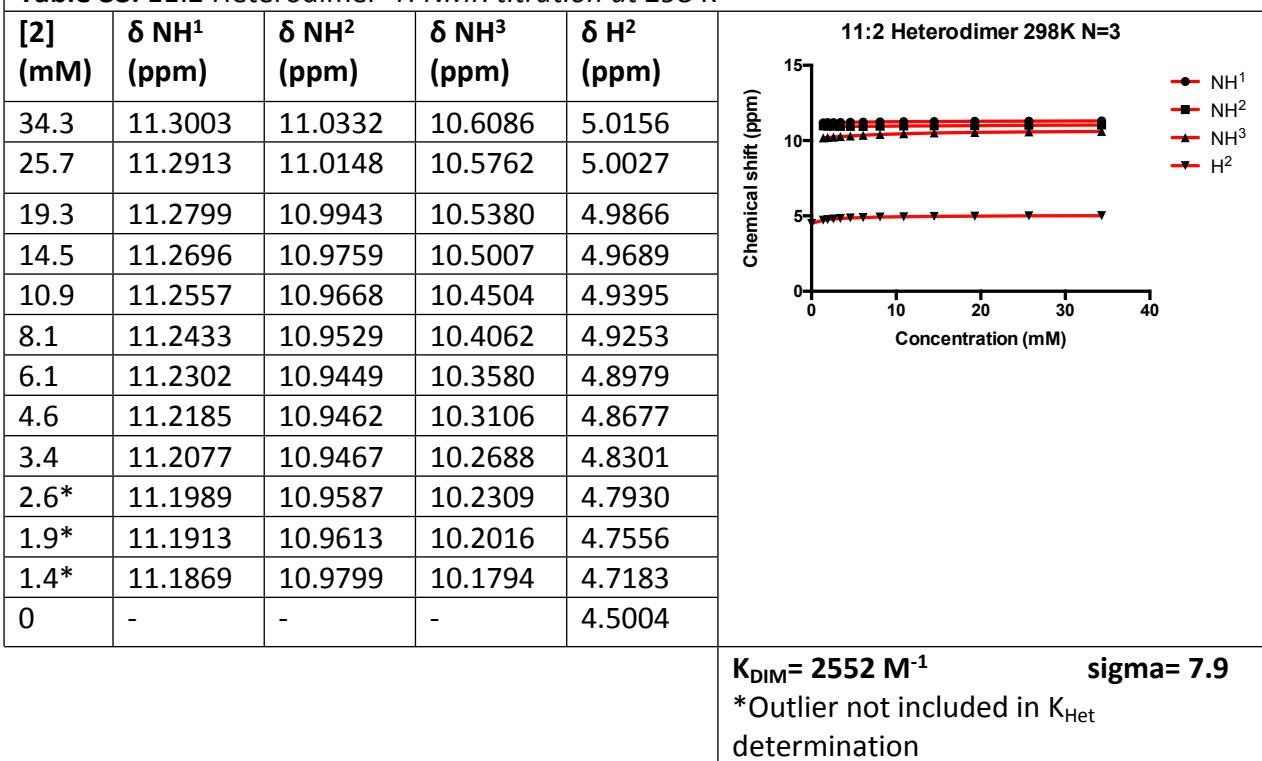
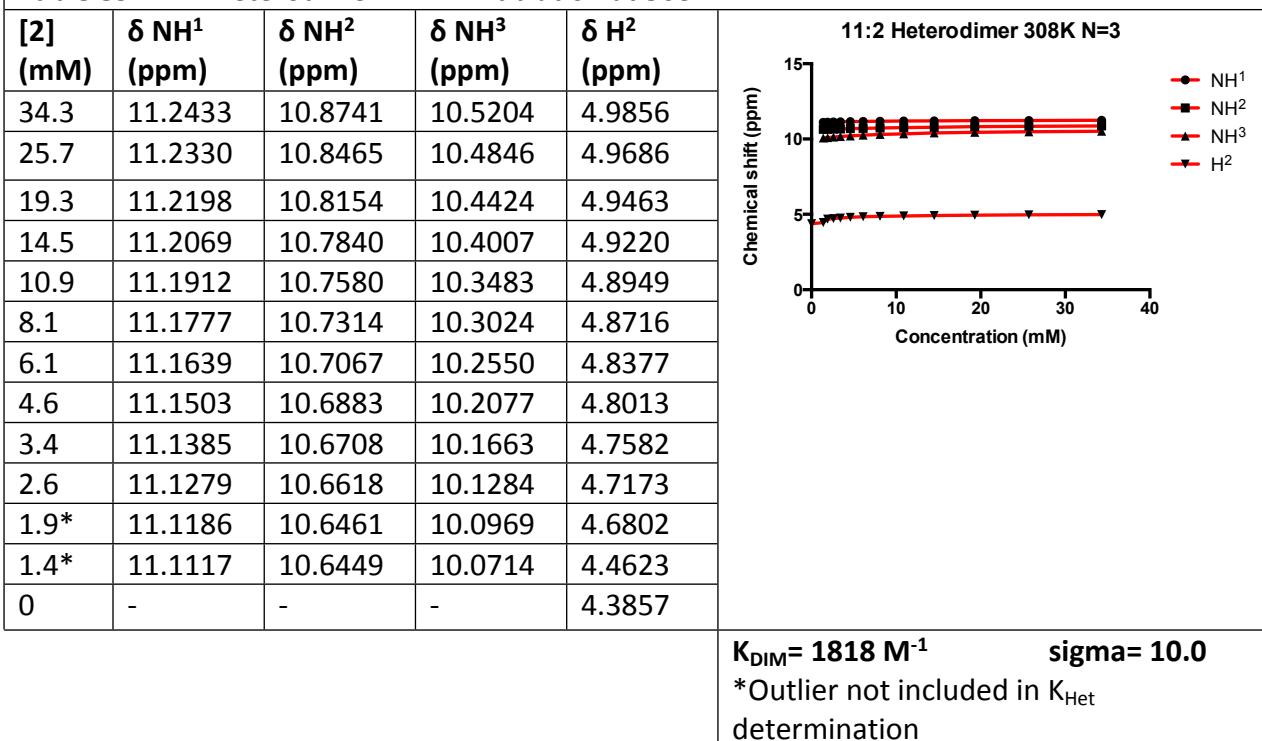
Table 86: 11:2 Heterodimer 1H NMR titration at 288 K**Table 87:** 11:2 Heterodimer 1H NMR titration at 295 K

Table 88: 11:2 Heterodimer 1H NMR titration at 298 K**Table 89:** 11:2 Heterodimer 1H NMR titration at 308 K

2.5.4. 11:2 Heterodimer VT ^1H NMR Summary and Van't Hoff Plots

Table 90: Summary of 11:2 Heterodimer Experiment 1 and Van't Hoff Plot

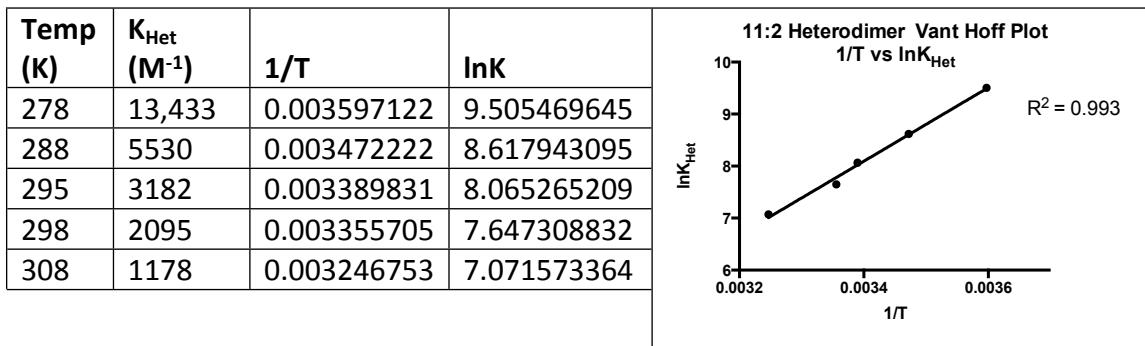


Table 91: Summary of 11:2 Heterodimer Experiment 2 and Van't Hoff Plot

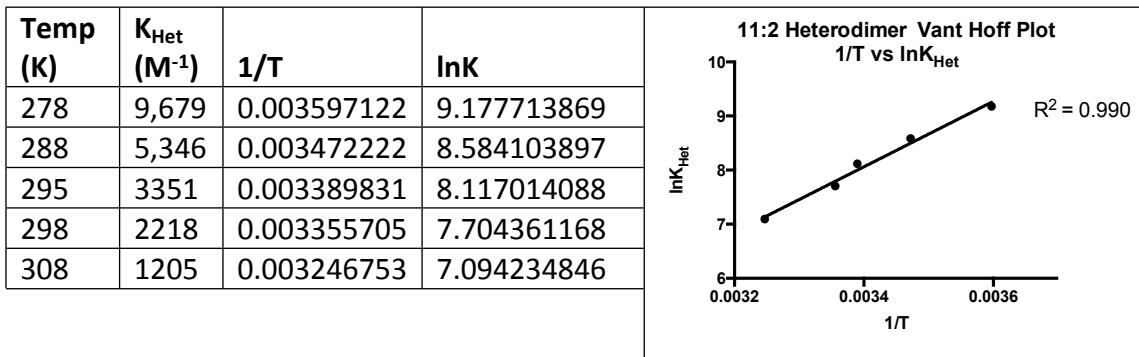


Table 92: Summary of 11:2 Heterodimer Experiment 3 and Van't Hoff Plot

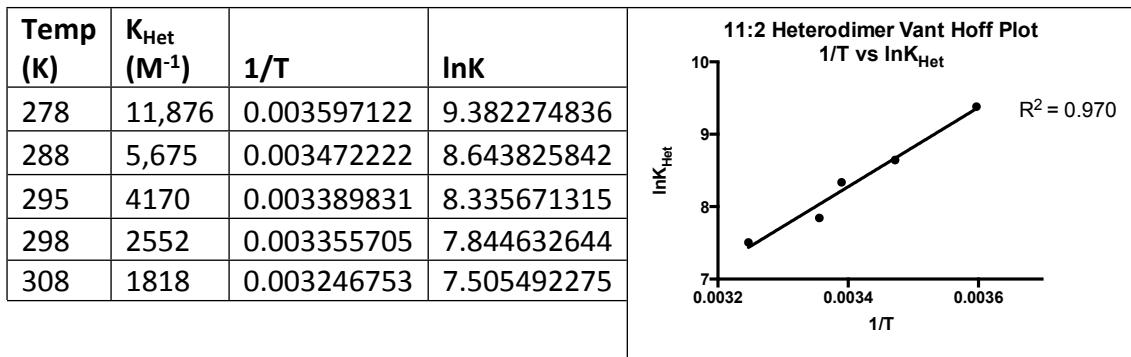


Table 93: Summary of thermodynamic parameters for 11:2 heterodimer

Expt	ΔH (kcal mol $^{-1}$)	$-T\Delta S^{295 \text{ K}}$ (kcal mol $^{-1}$)	ΔG (kcal mol $^{-1}$)
N1	-14.0	9.3	- 4.7
N2	-12.0	7.3	- 4.7
N3	-11.6	6.1	- 5.5
Average	-12.5	7.6	- 5.0

2.6. 11:2a Heterodimer VT ^1H NMR Titration Experiment

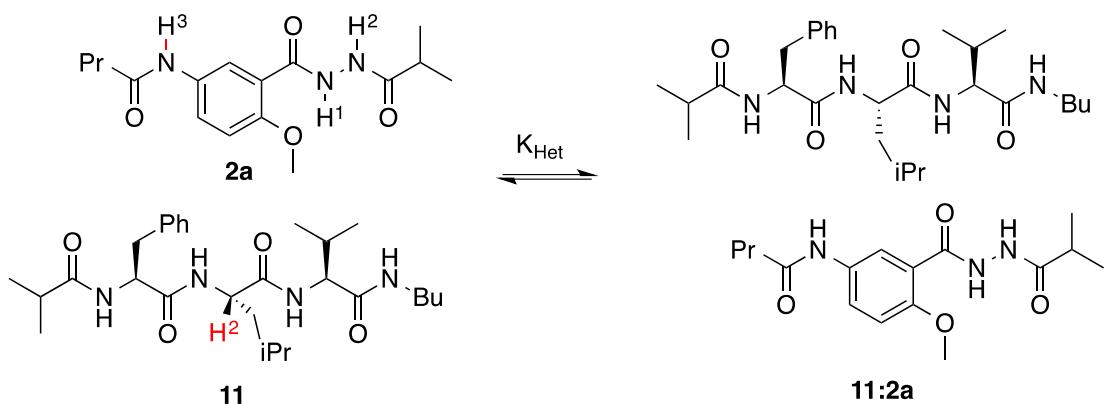


Figure 7: Heterodimerisation of **2a** and **11**. Resonances used to determine binding constants are highlighted in red.

N.B. K_{Het} was calculated on using H^2 only. Shifts of NH signals in **2a** were broad and therefore not included in K_{Het} calculation.

2.6.1. 11:2a Heterodimer ^1H NMR Titration Experiment 1

Peptide concentration: 3.1 mM

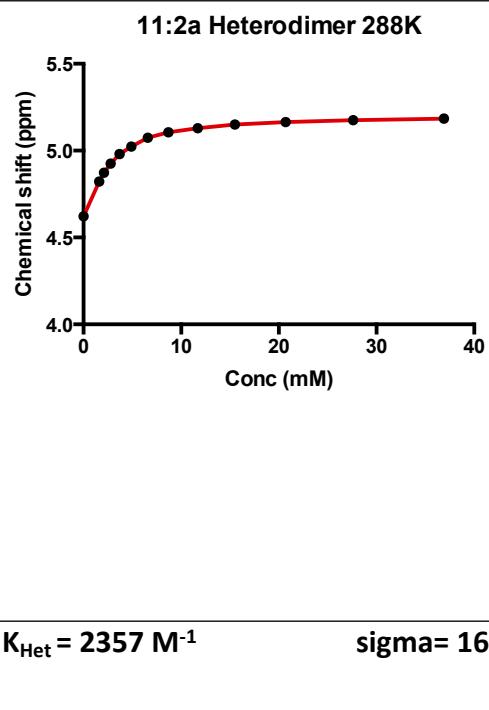
Table 94: 11:2a Heterodimer ^1H NMR titration at 278 K				
[2a] (mM)	δNH^1 (ppm)	δNH^2 (ppm)	δNH^3 (ppm)	δH^2 (ppm)
36.9	-	-	-	5.2081
27.6	-	-	-	5.2011
20.7	-	-	-	5.1957
15.5	-	-	-	5.1860
11.7	-	-	-	5.1758
8.7	-	-	-	5.1540
6.6	-	-	-	5.1336
4.9	-	-	-	5.1053
3.7	-	-	-	5.0630
2.8	-	-	-	5.0170
2.1	-	-	-	4.9541
1.6	-	-	-	4.9054
0	-	-	-	4.6751

11:2a Heterodimer 278K

$K_{\text{Het}} = 5923 \text{ M}^{-1}$
 $\sigma = 22.09$

Table 95: 11:2a Heterodimer 1H NMR titration at 288 K

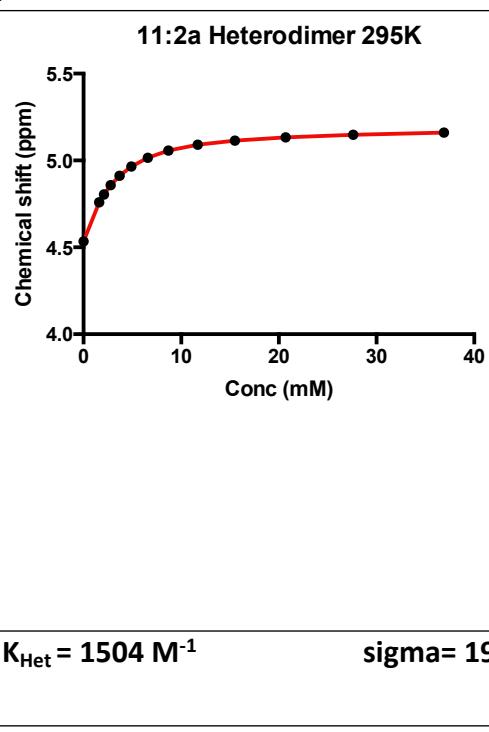
[2a] (mM)	δ NH ¹ (ppm)	δ NH ² (ppm)	δ NH ³ (ppm)	δ H ² (ppm)
36.9	-	-	-	5.1845
27.6	-	-	-	5.1750
20.7	-	-	-	5.1638
15.5	-	-	-	5.1502
11.7	-	-	-	5.1294
8.7	-	-	-	5.1060
6.6	-	-	-	5.0740
4.9	-	-	-	5.0236
3.7	-	-	-	4.9808
2.8	-	-	-	4.9262
2.1	-	-	-	4.8732
1.6	-	-	-	4.8221
0	-	-	-	4.6221



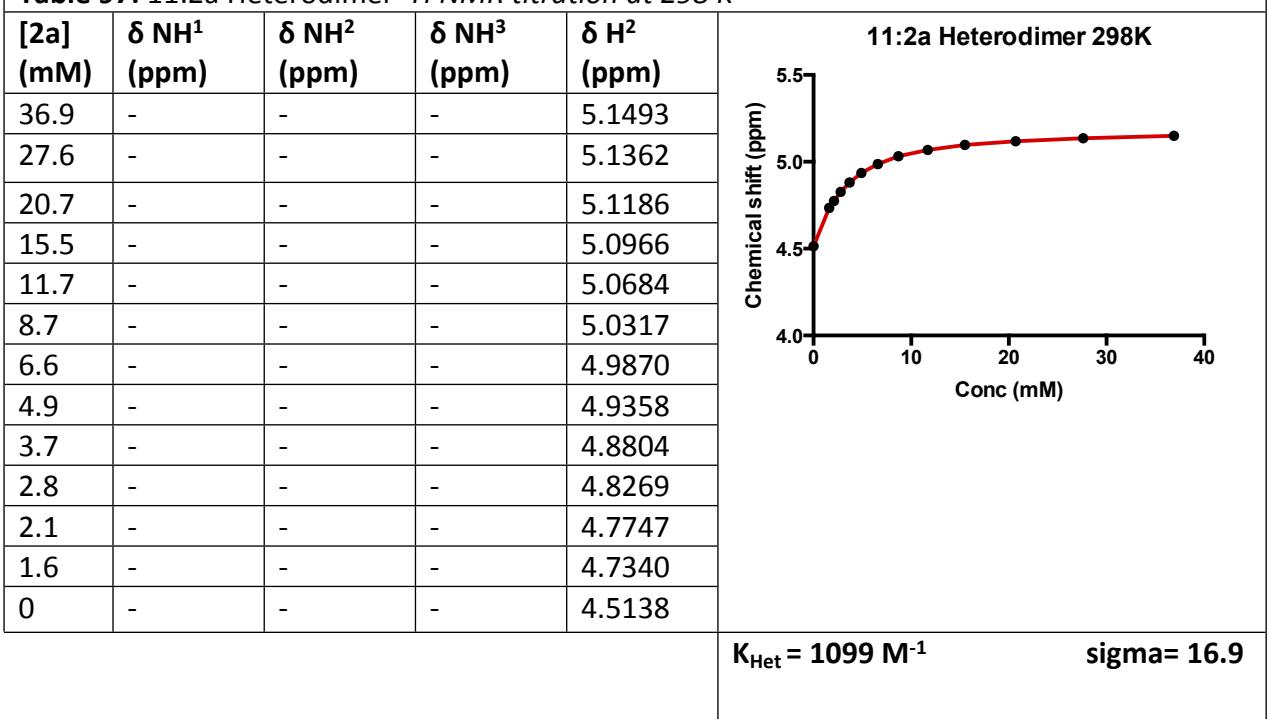
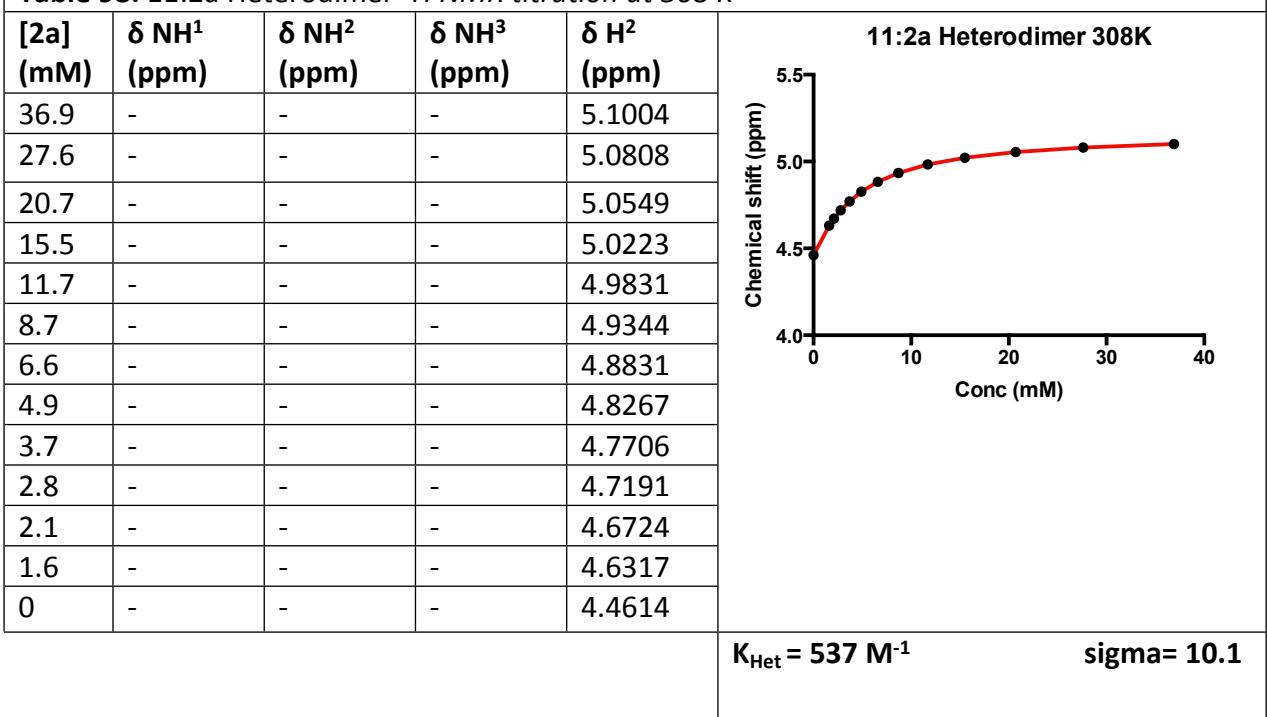
$$K_{\text{Het}} = 2357 \text{ M}^{-1} \quad \sigma = 16.5$$

Table 96: 11:2a Heterodimer 1H NMR titration at 295 K

[2a] (mM)	δ NH ¹ (ppm)	δ NH ² (ppm)	δ NH ³ (ppm)	δ H ² (ppm)
36.9	-	-	-	5.1609
27.6	-	-	-	5.1479
20.7	-	-	-	5.1335
15.5	-	-	-	5.1141
11.7	-	-	-	5.0901
8.7	-	-	-	5.0570
6.6	-	-	-	5.0160
4.9	-	-	-	4.9654
3.7	-	-	-	4.9115
2.8	-	-	-	4.8575
2.1	-	-	-	4.8050
1.6	-	-	-	4.7594
0	-	-	-	4.5339



$$K_{\text{Het}} = 1504 \text{ M}^{-1} \quad \sigma = 19.0$$

Table 97: 11:2a Heterodimer ^1H NMR titration at 298 K**Table 98:** 11:2a Heterodimer ^1H NMR titration at 308 K

2.6.2. 11:2a Heterodimer ^1H NMR Titration Experiment 2

Peptide concentration: 3.1 mM

Table 99: 11:2a Heterodimer ^1H NMR titration at 278 K N=2

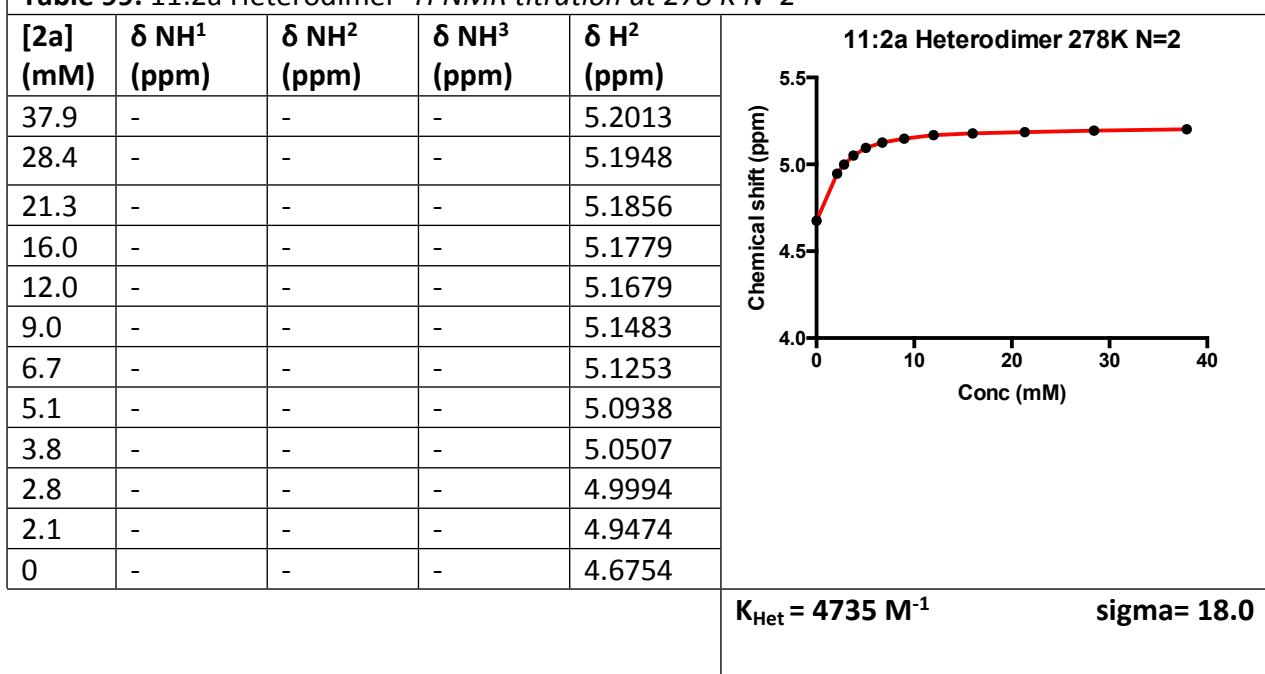


Table 100: 11:2a Heterodimer ^1H NMR titration at 288 K N=2

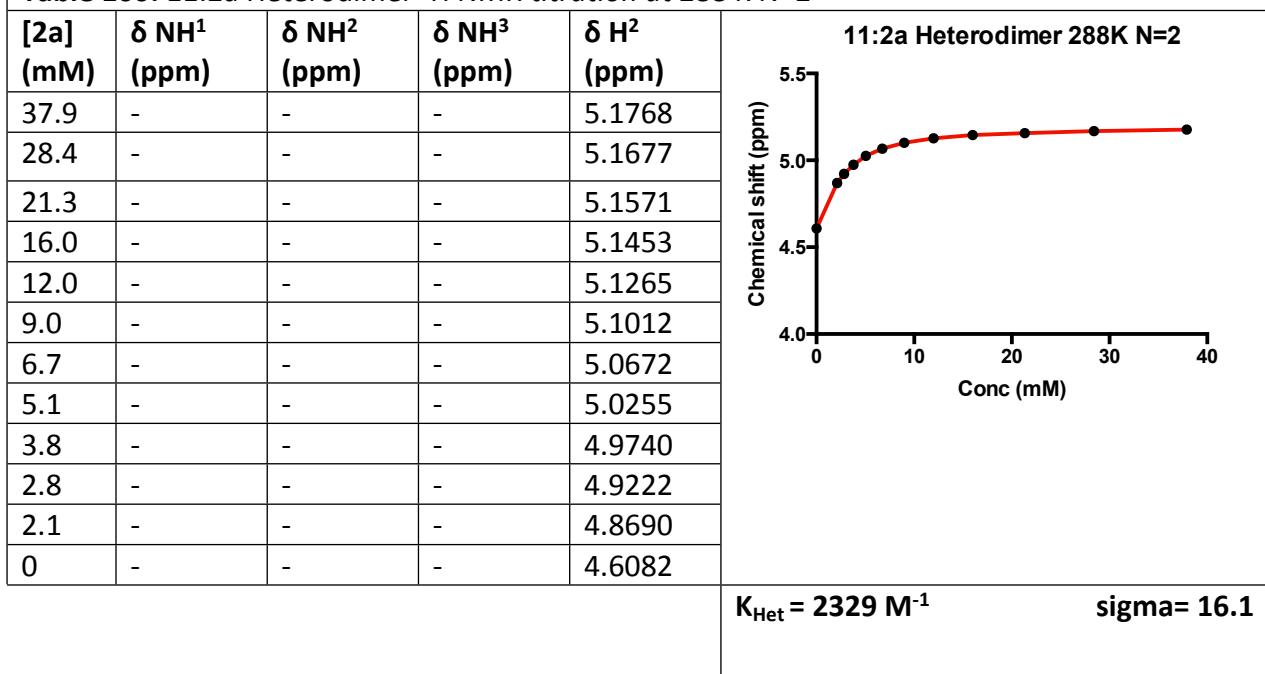


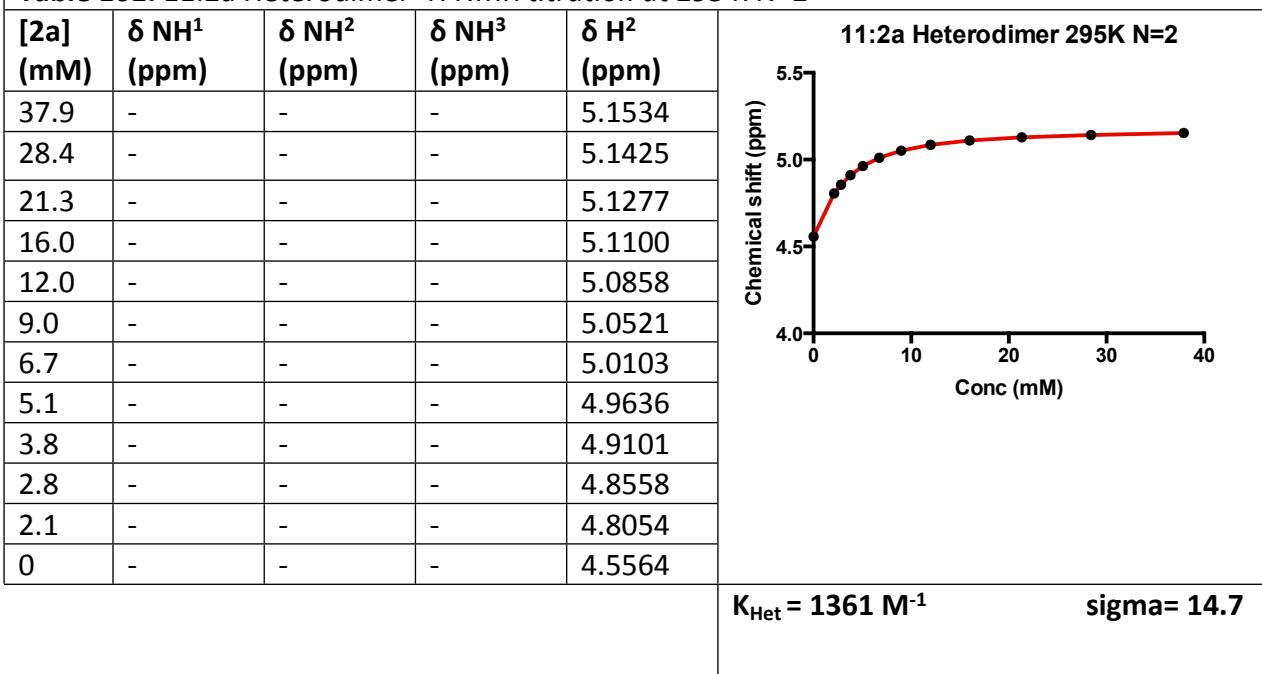
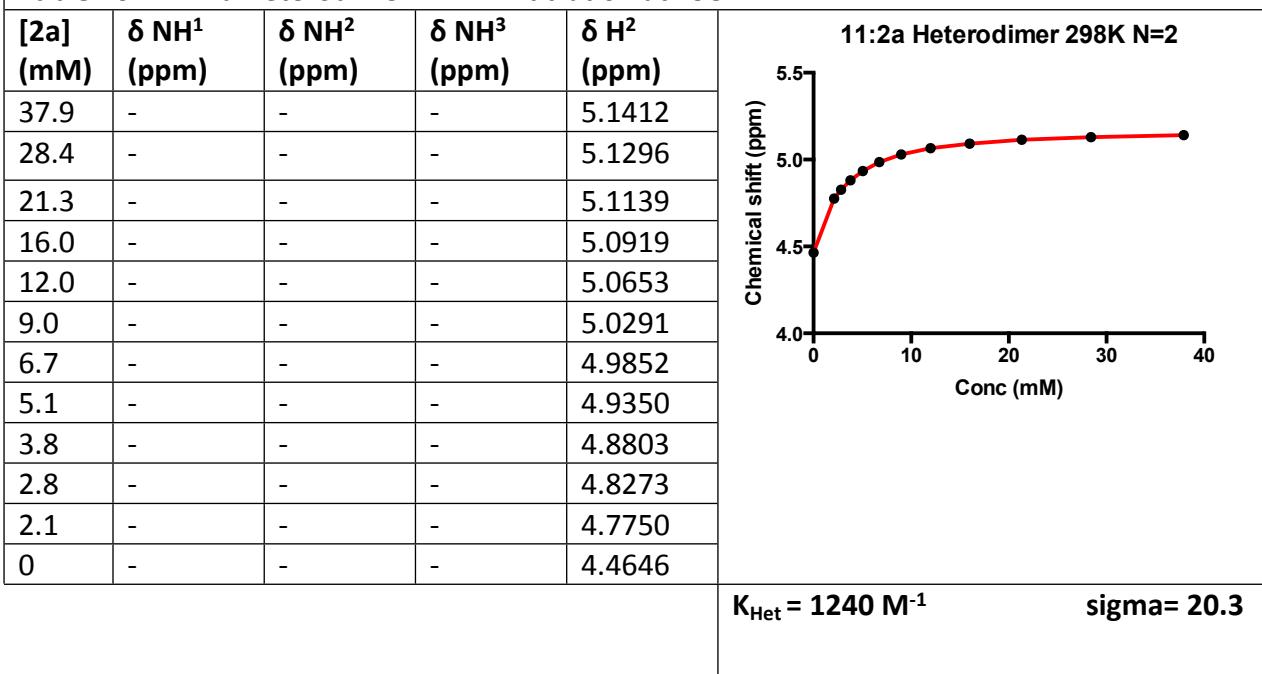
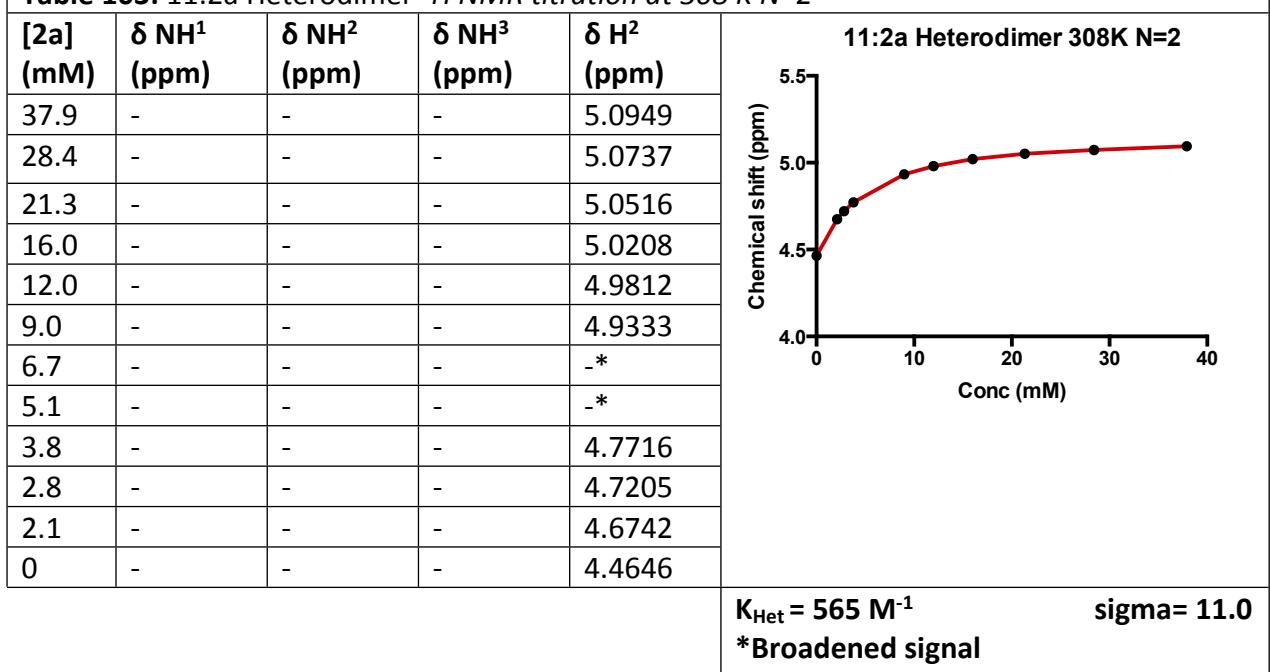
Table 101: 11:2a Heterodimer 1H NMR titration at 295 K N=2**Table 102:** 11:2a Heterodimer 1H NMR titration at 298 K N=2

Table 103: 11:2a Heterodimer 1H NMR titration at 308 K N=2



2.6.3. 11:2a Heterodimer ^1H NMR Titration Experiment 3

Peptide concentration: 3.3 mM

Table 104: 11:2a Heterodimer ^1H NMR titration at 278 K N=3

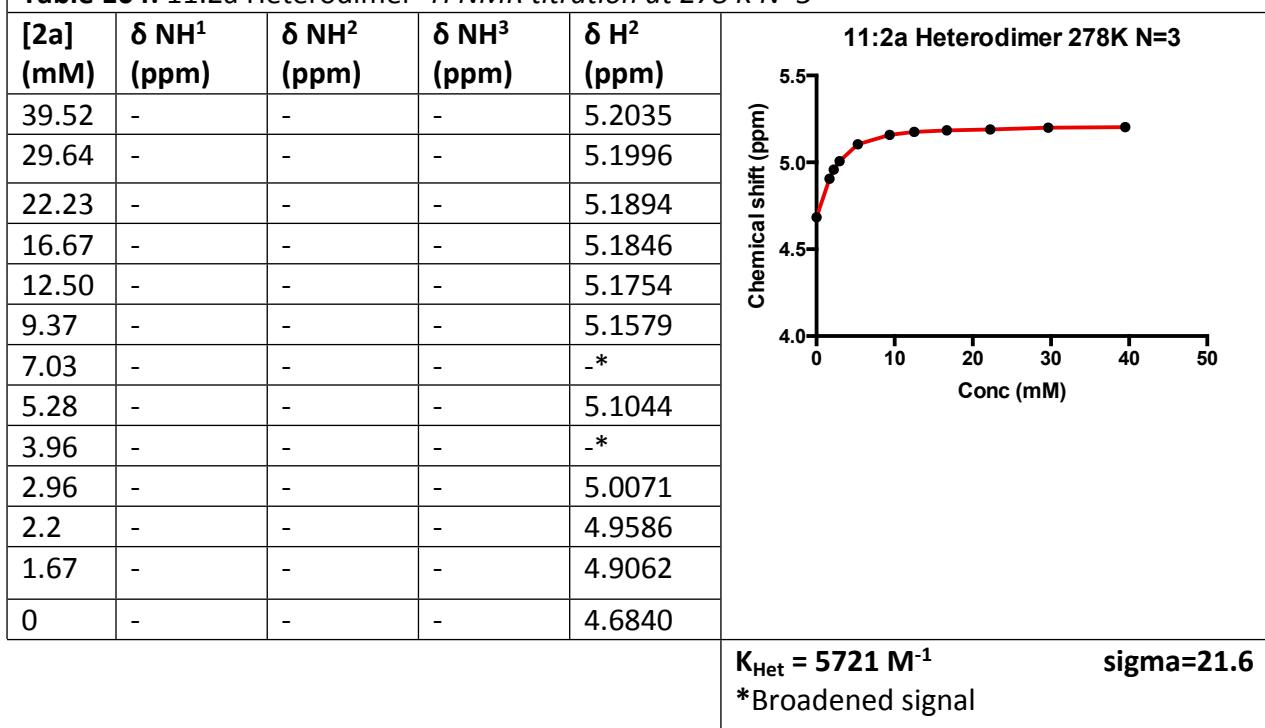


Table 105: 11:2a Heterodimer ^1H NMR titration at 288 K N=3

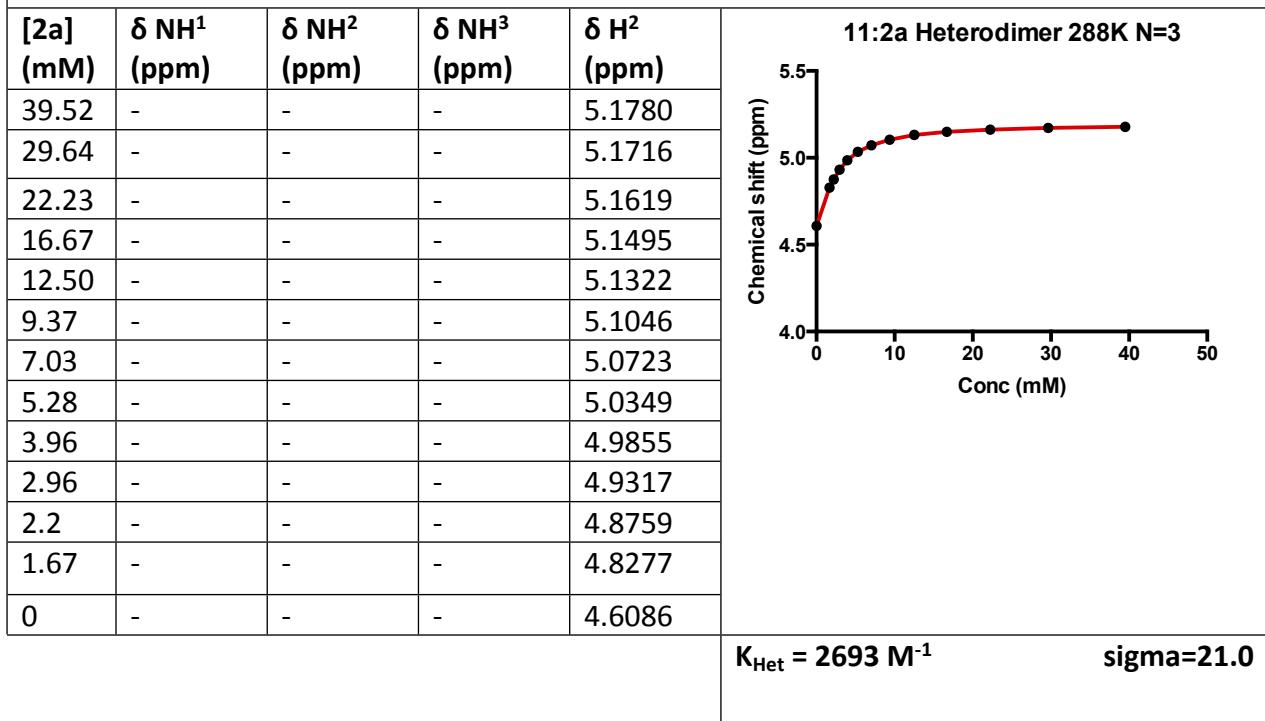


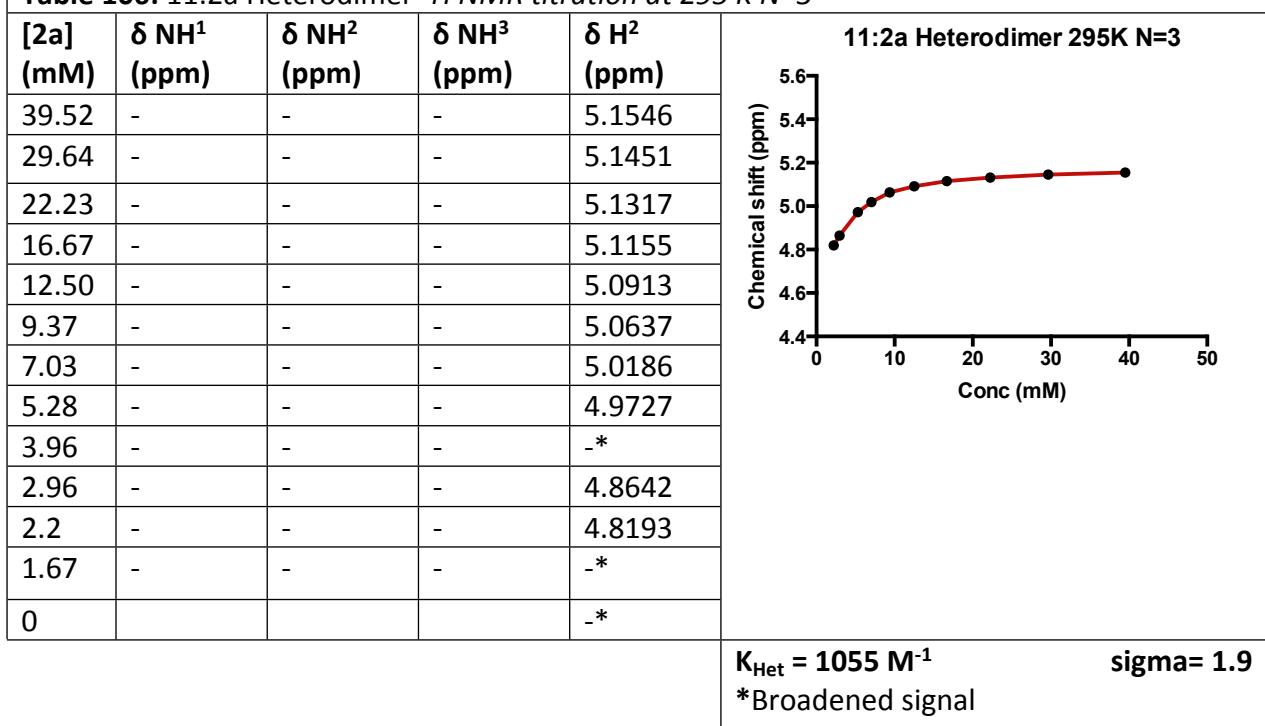
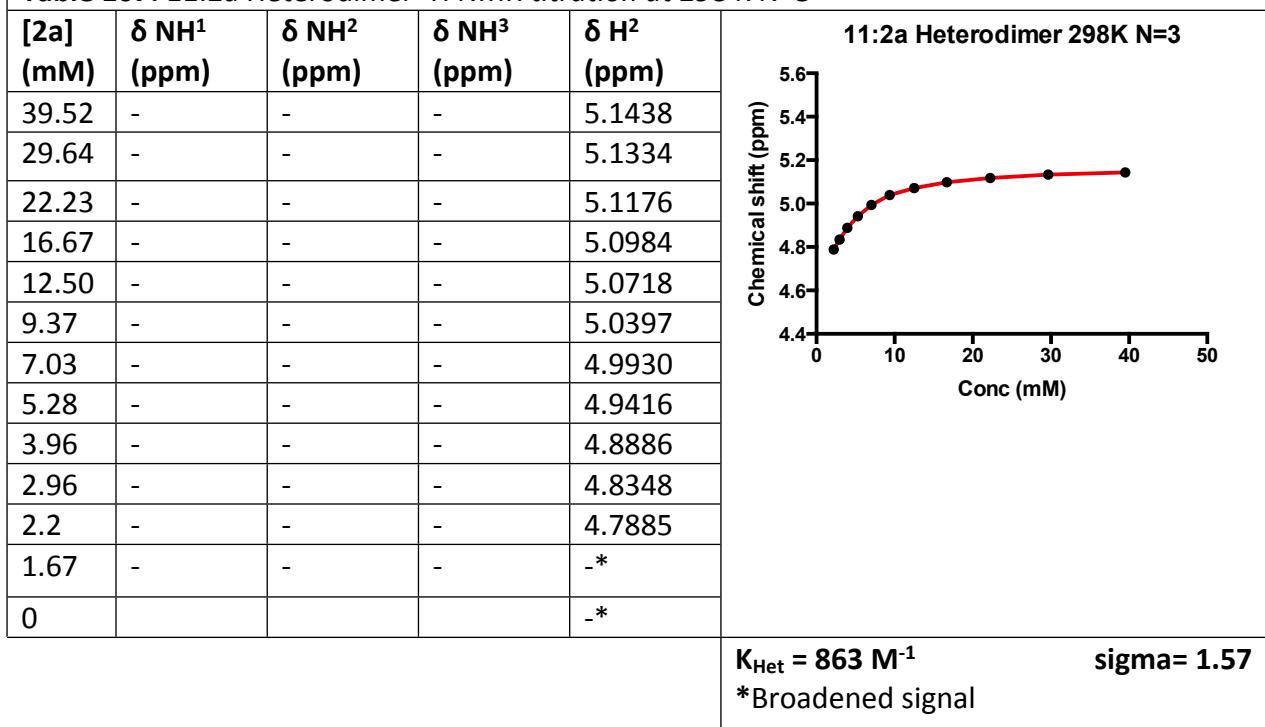
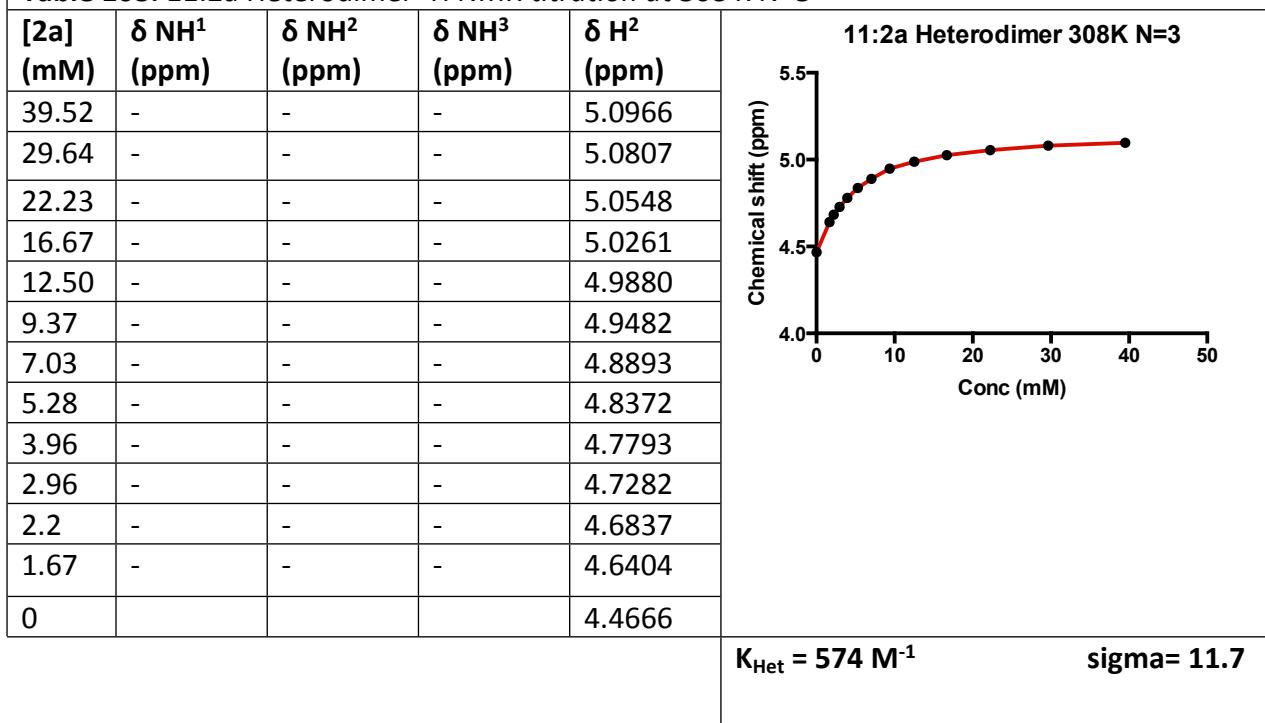
Table 106: 11:2a Heterodimer 1H NMR titration at 295 K N=3**Table 107:** 11:2a Heterodimer 1H NMR titration at 298 K N=3

Table 108: 11:2a Heterodimer ^1H NMR titration at 308 K N=3

2.6.4. 11:2a Heterodimer VT ^1H NMR Summary and Van't Hoff Plots

Table 109: Summary of 11:2a Heterodimer Experiment 1 and Van't Hoff Plot

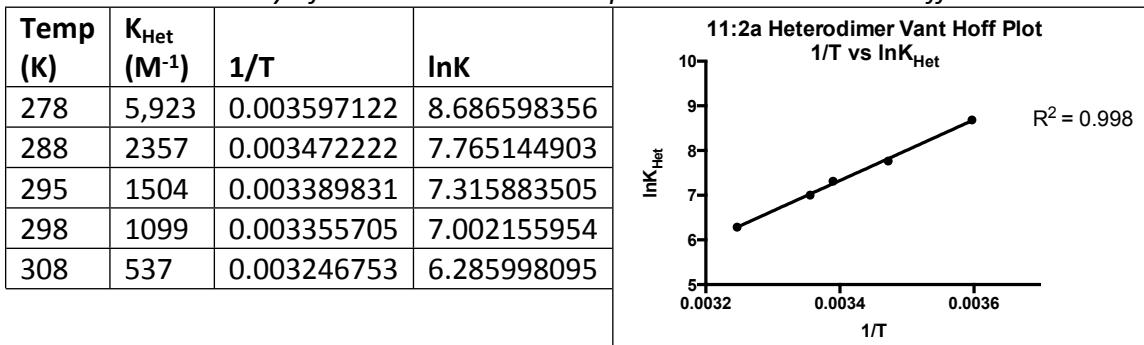


Table 110: Summary of 11:2a Heterodimer Experiment 2 and Van't Hoff Plot

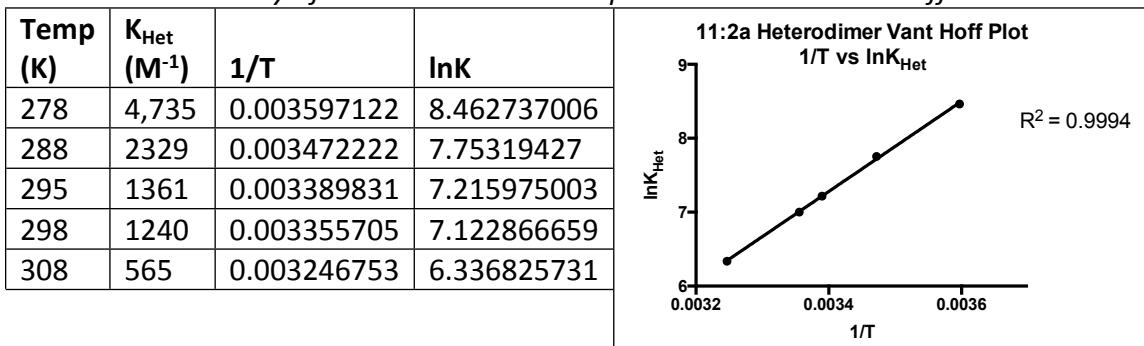


Table 111: Summary of 11:2a Heterodimer Experiment 3 and Van't Hoff Plot

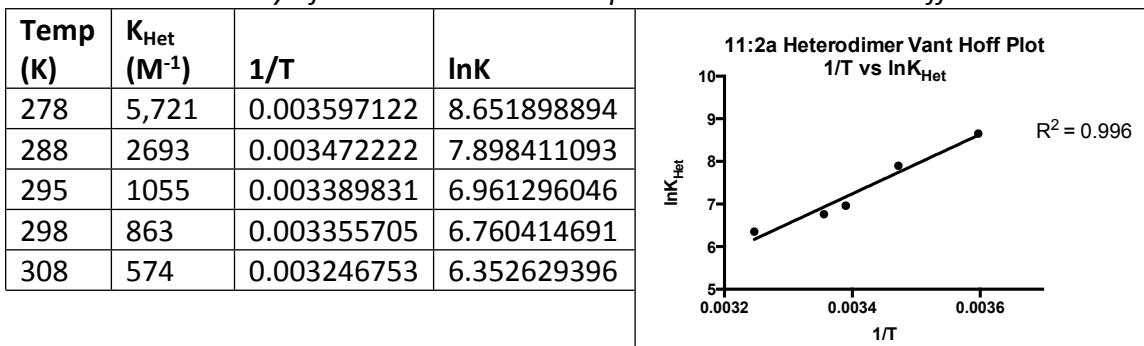
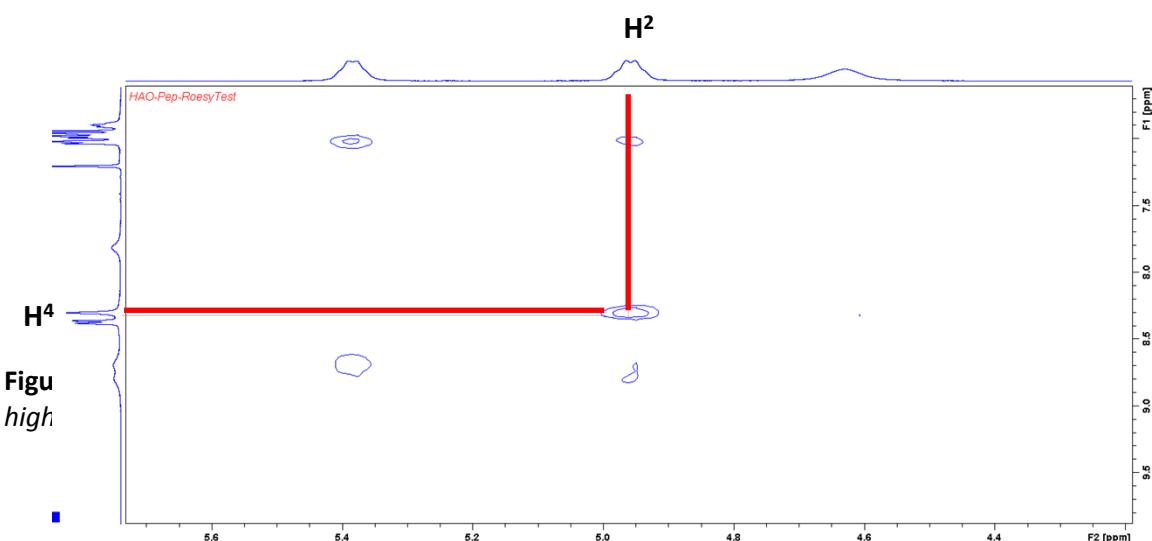
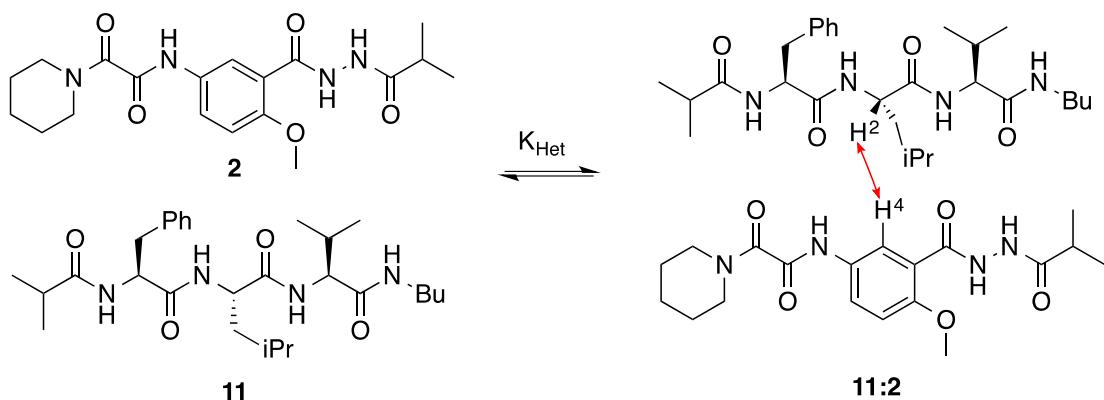


Table 112: Summary of thermodynamic parameters for 11:2 heterodimer

Expt	ΔH (kcal mol $^{-1}$)	$-\Delta S^{295 \text{ K}}$ (kcal mol $^{-1}$)	ΔG (kcal mol $^{-1}$)
N1	-13.5	9.3	- 4.2
N2	-11.9	7.7	- 4.2
N3	-13.9	9.7	- 4.2
Average	-13.1	8.9	- 4.2

2.7. ROESY Study of 2:11 Heterodimer

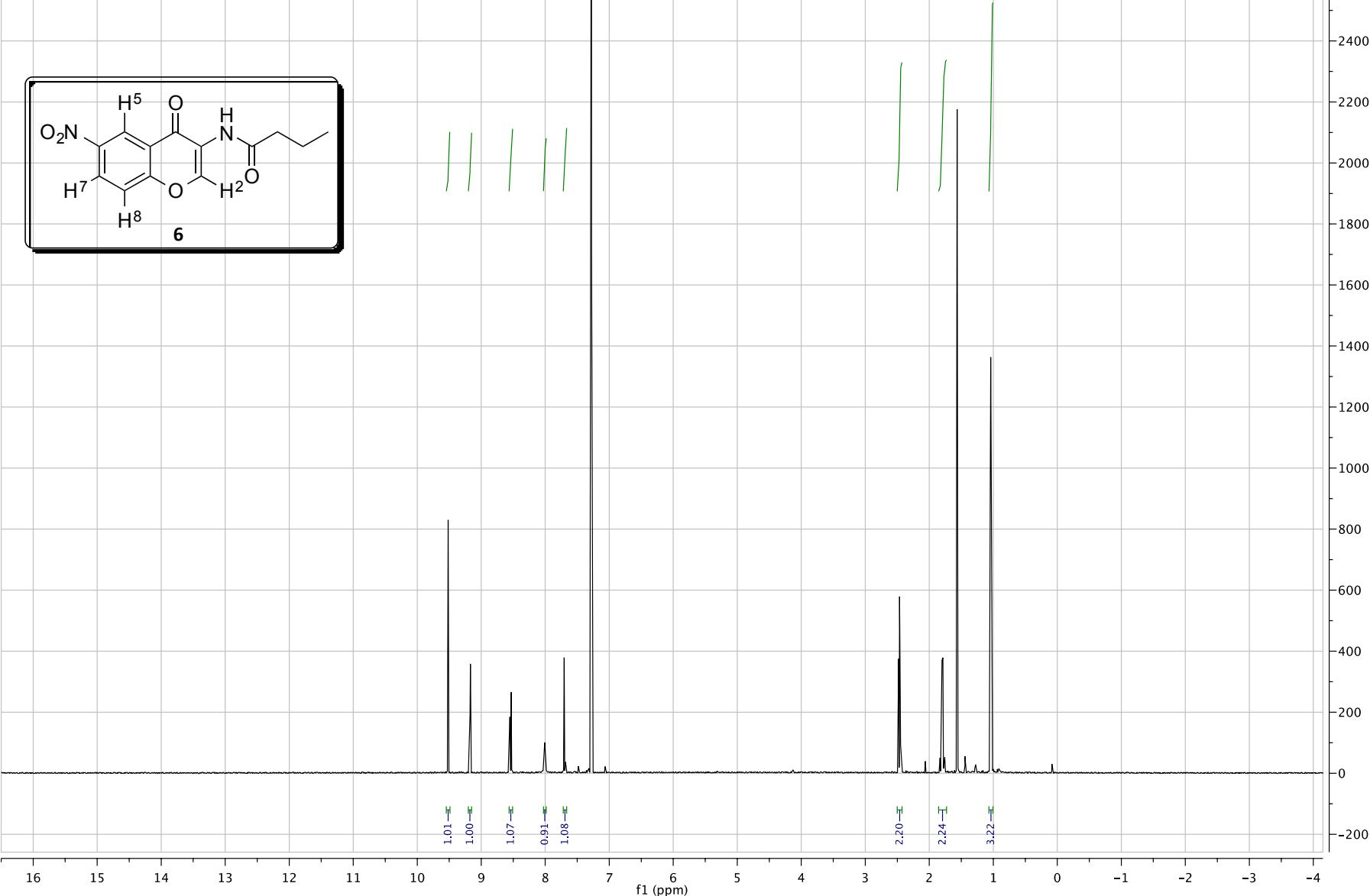


References

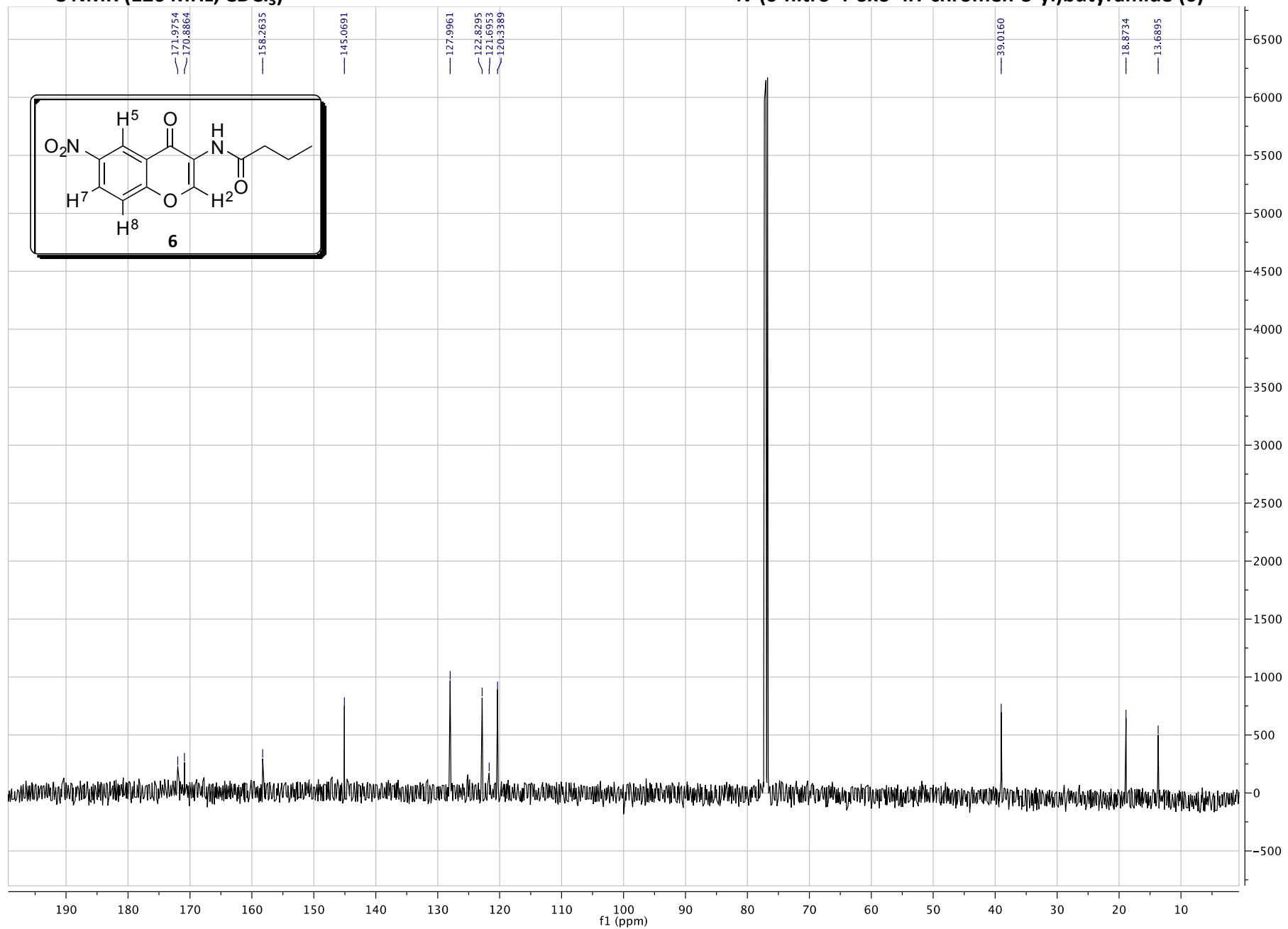
- (1) S. Vedachalam, J. Zeng, B. K. Gorityala, M. Antonio and X.Liu, *Org. Lett.*, 2010, **12**, 352.
- (2) J. E. Thomson, C. D. Campbell, C. Concellon, N. Duguet, K. Rix, A. M. Z. Slawin, and A. D. Smith, *J. Org. Chem.*, 2008, **73**, 2784.
- (3) J. S. Nowick, M.D. Chung, K. Maitra, S. Mairita, K. D. Stigers and Y. Sun, *J. Am. Chem. Soc.*, 2000, **122**, 7654.
- (4) D. L. Holmes, E. M. Smith and J. S. Nowick, *J. Am. Chem. Soc.*, 1997, **119**, 7665.

¹H and ¹³C NMR Spectra

¹H NMR (500 MHz, CDCl₃)

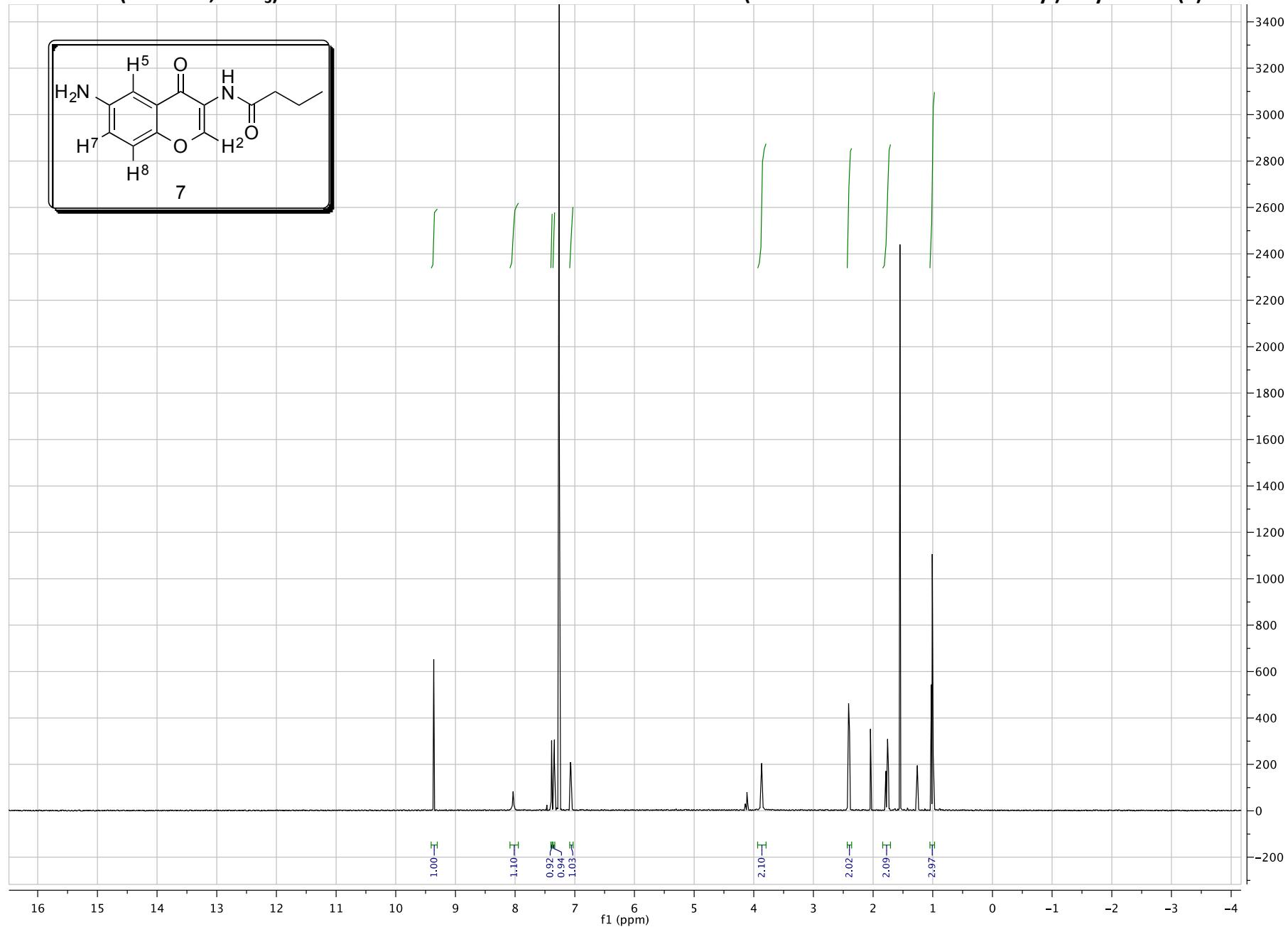


¹³C NMR (126 MHz, CDCl₃)



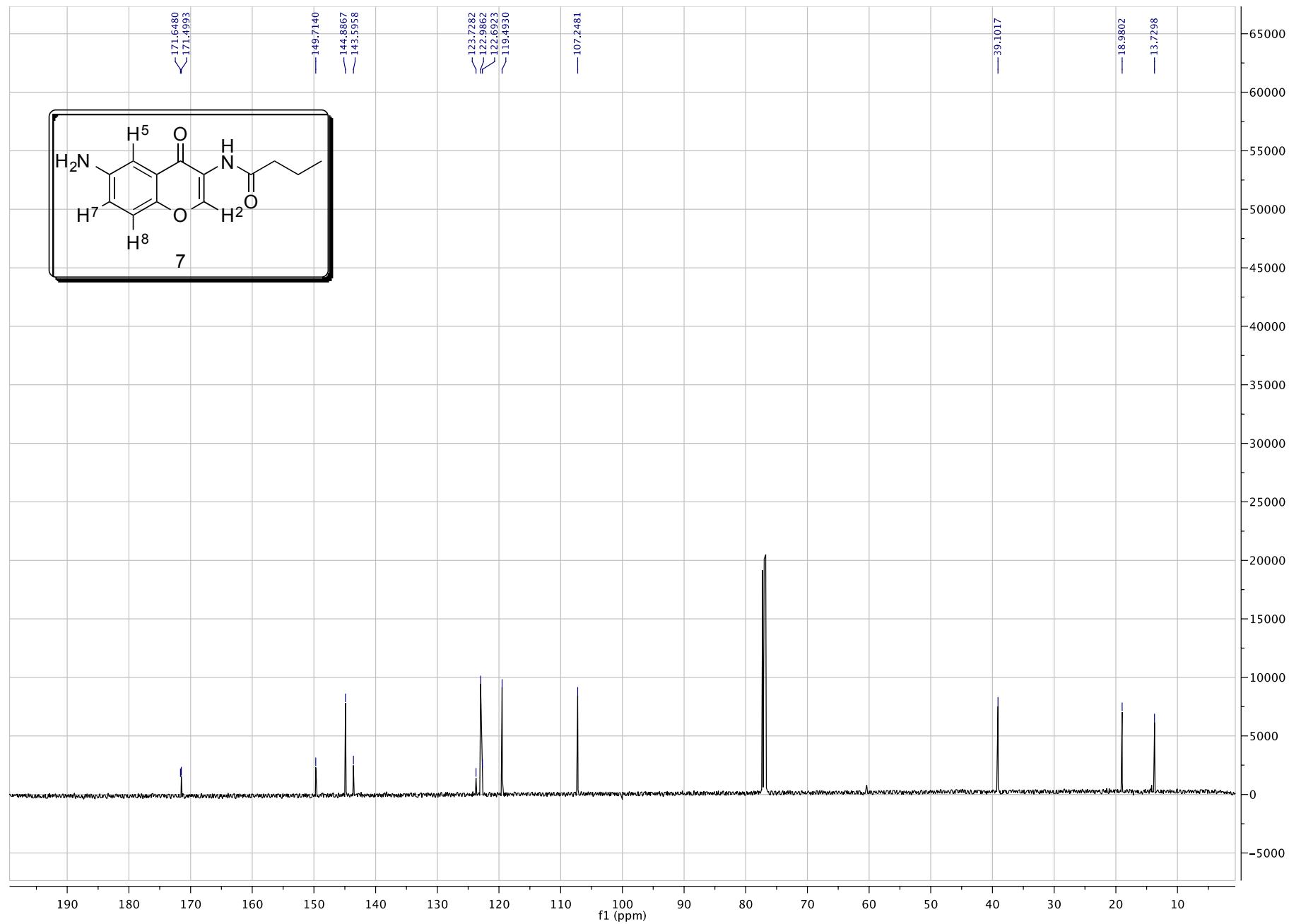
¹H NMR (500 MHz, CDCl₃)

N-(6-amino-4-oxo-4*H*-chromen-3-yl)butyramide (7)

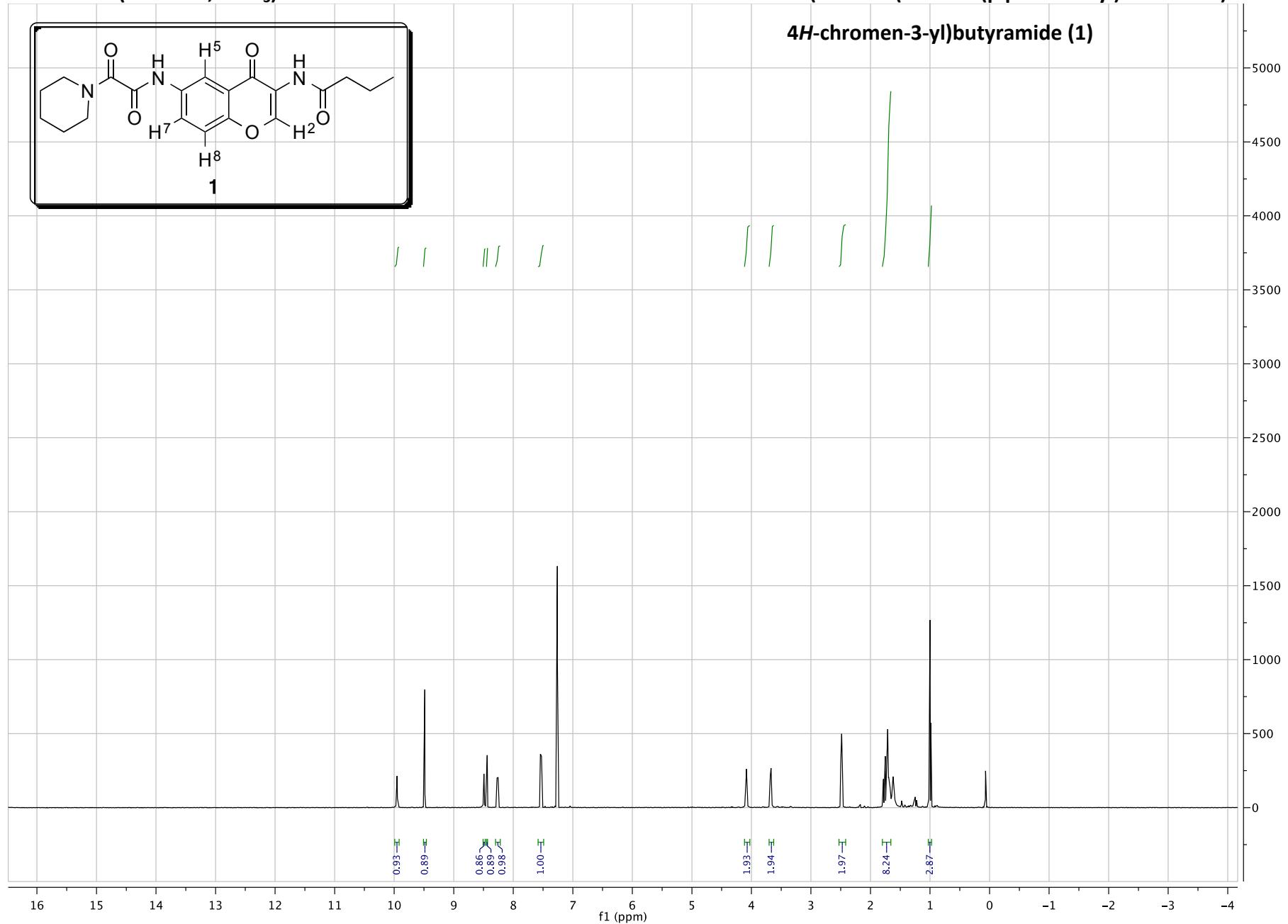


¹³C NMR (126 MHz, CDCl₃)

N-(6-amino-4-oxo-4*H*-chromen-3-yl)butyramide (7)

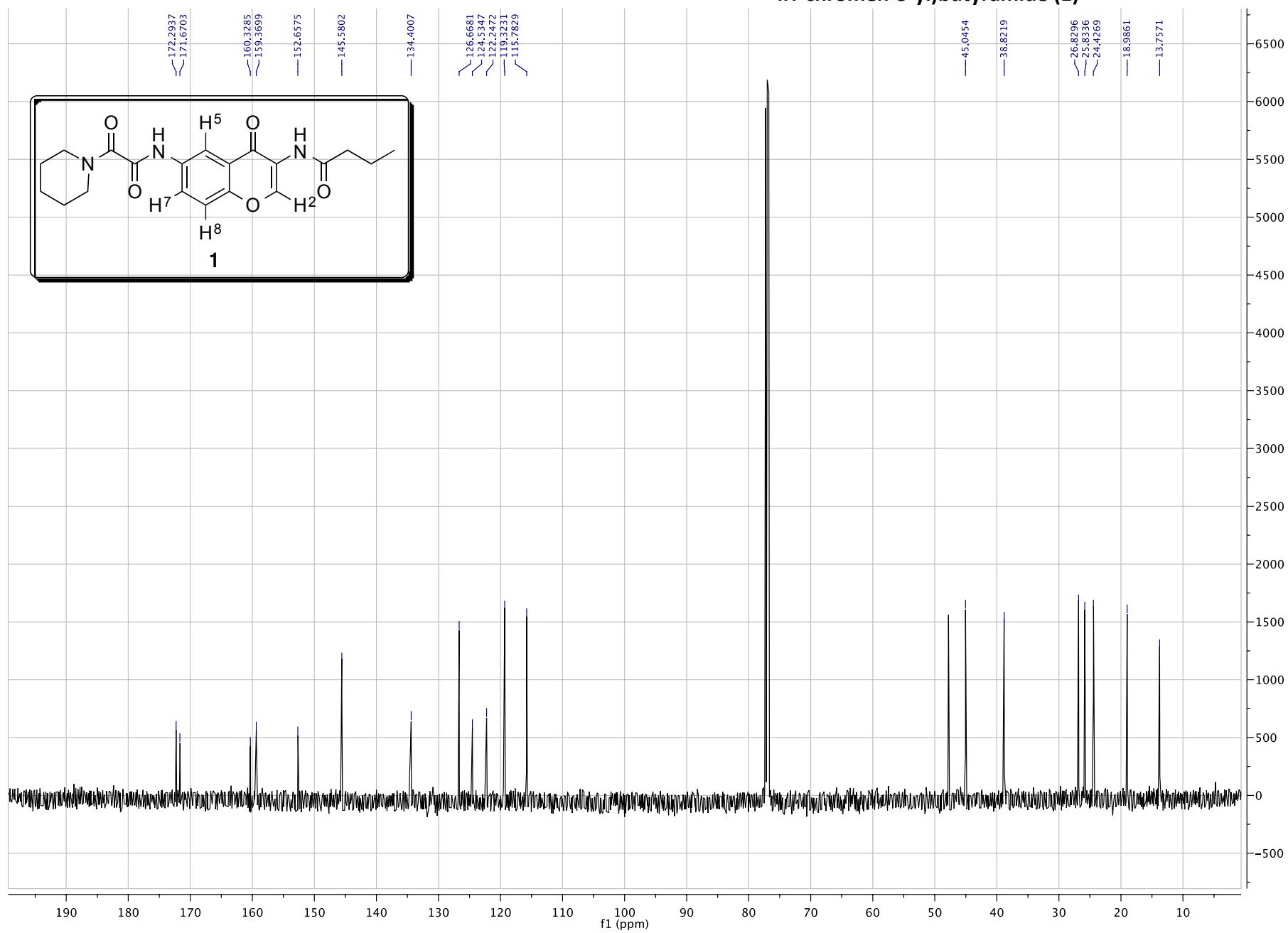


¹H NMR (500 MHz, CDCl₃)

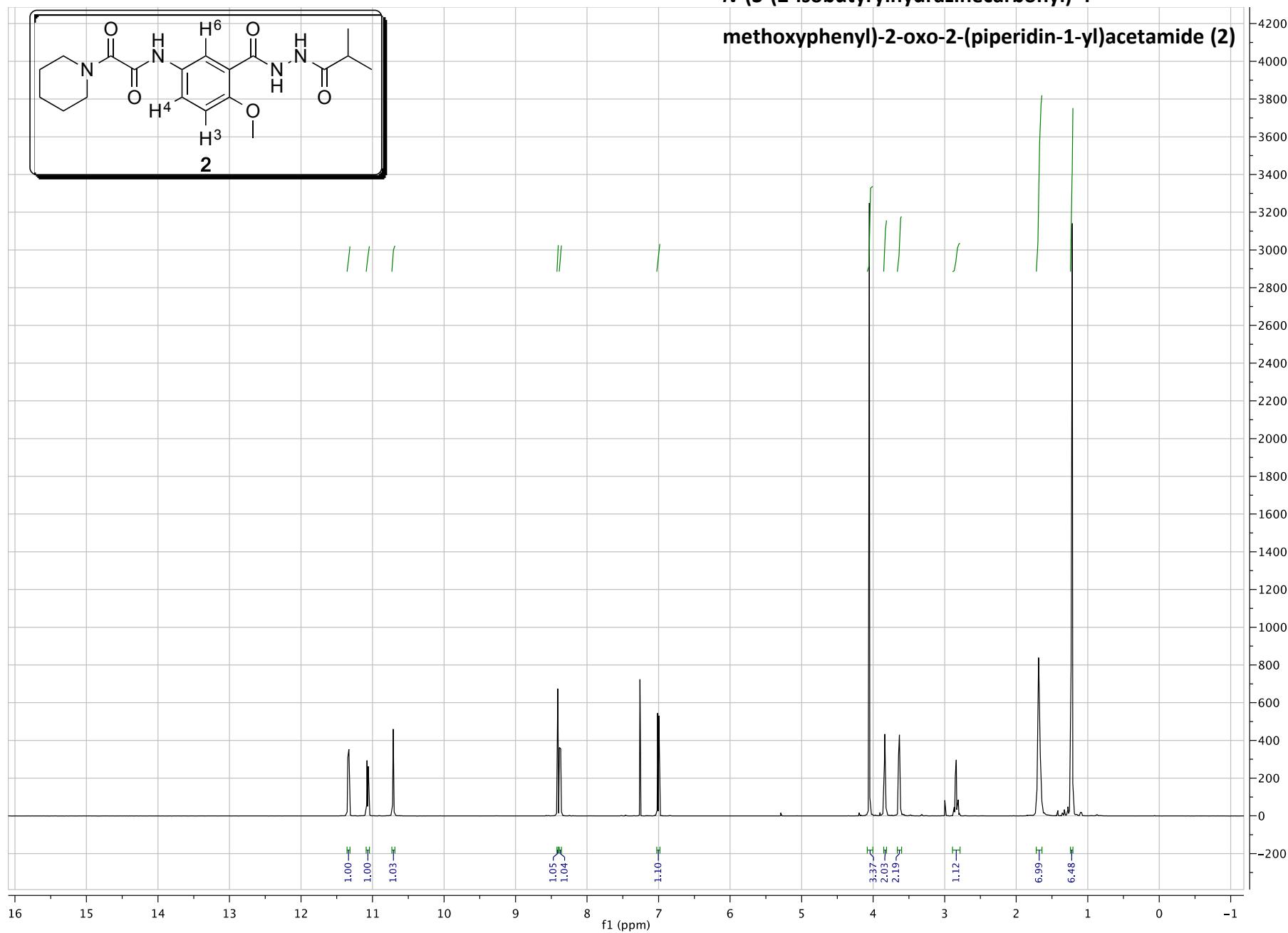


N-(4-oxo-6-(2-oxo-2-(piperidin-1-yl)acetamido)-
4*H*-chromen-3-yl)butyramide (**1**)

¹³C NMR (126 MHz, CDCl₃)

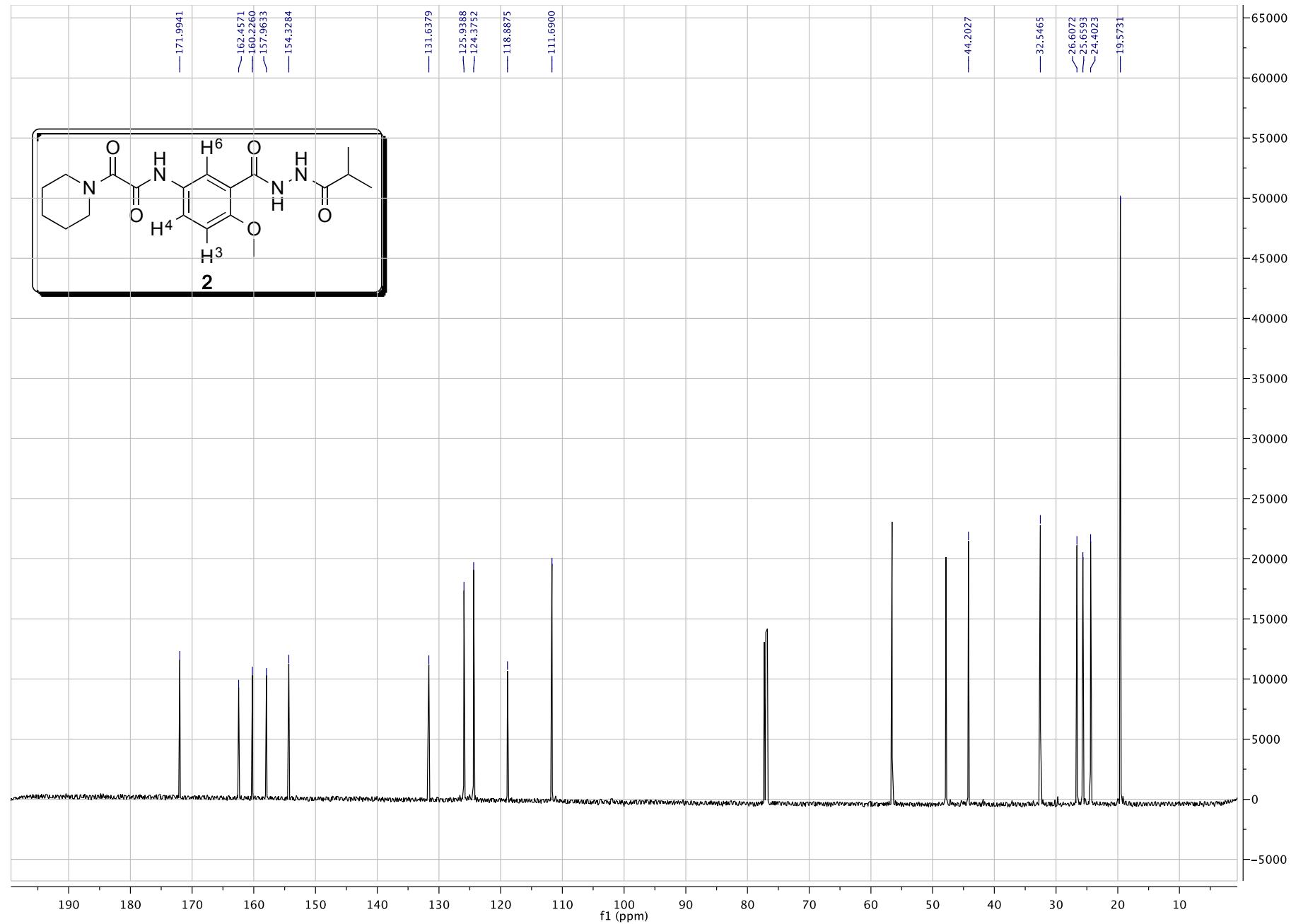


¹H NMR (500 MHz, CDCl₃)

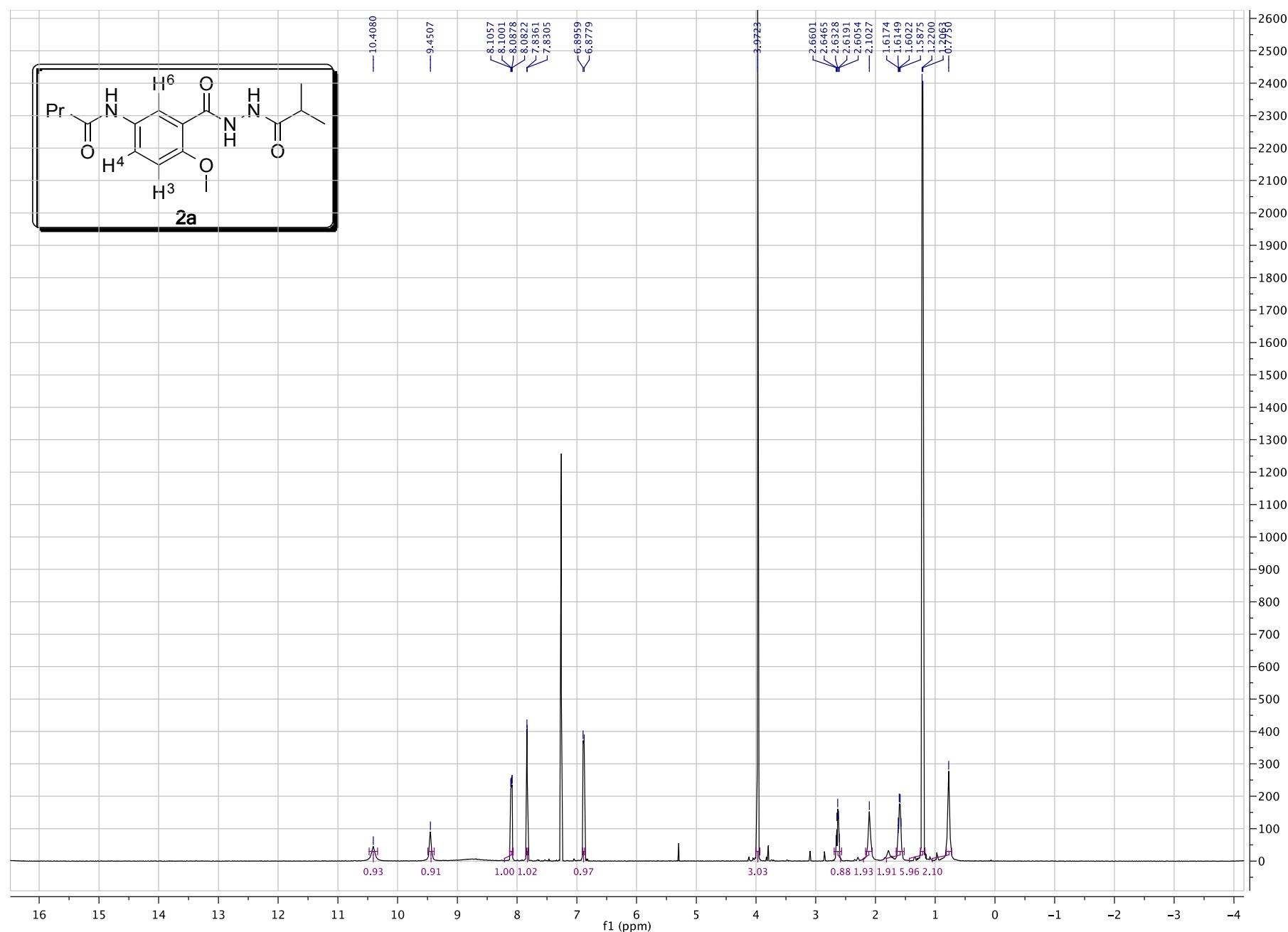


¹³C NMR (126 MHz, CDCl₃)

N-(3-(2-isobutyrylhydrazinecarbonyl)-4-methoxyphenyl)-2-oxo-2-(piperidin-1-yl)acetamide (2)

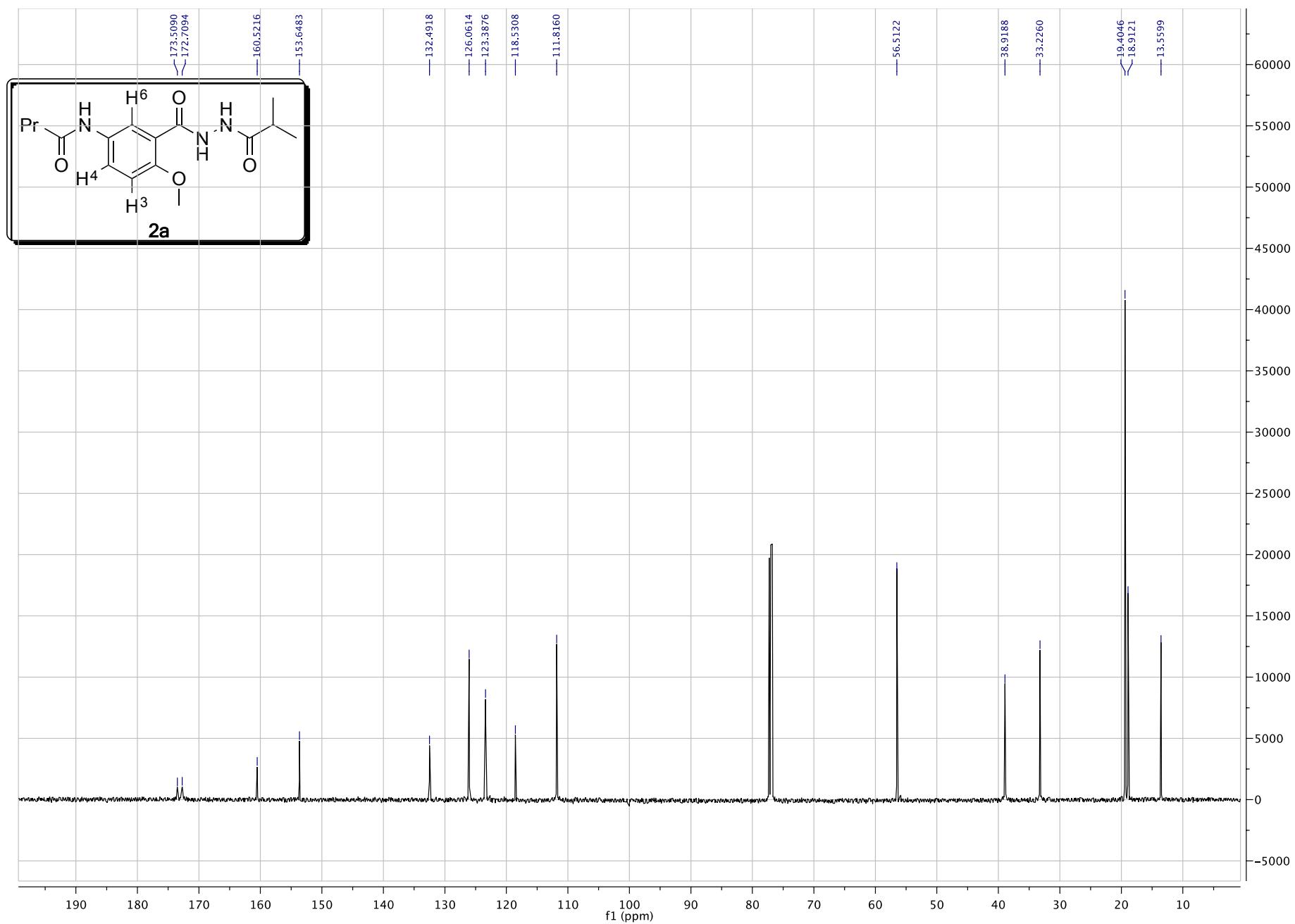


¹H NMR (500 MHz, CDCl₃)



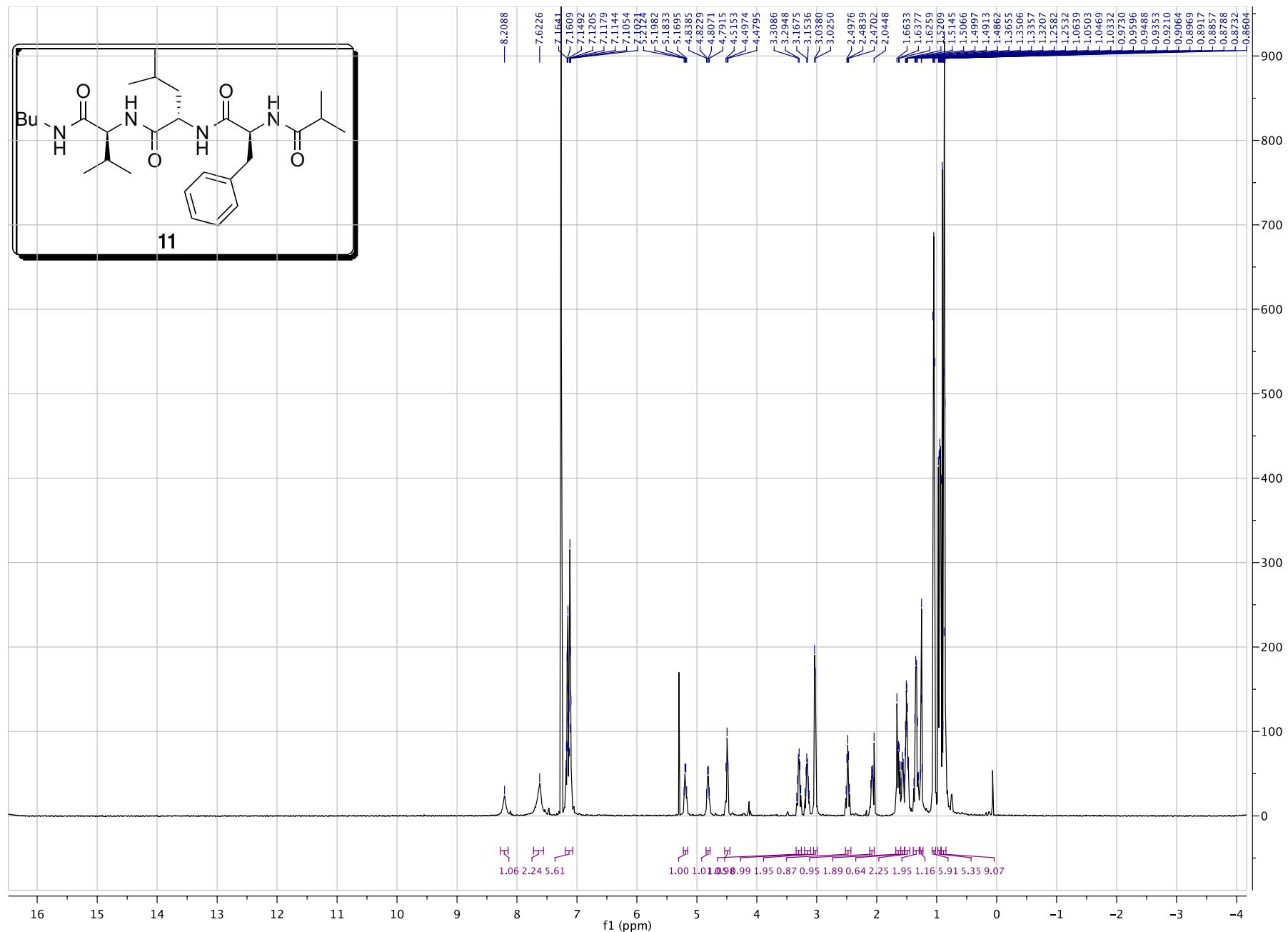
¹³C NMR (126 MHz, CDCl₃)

N-(3-(2-isobutyrylhydrazine-1-carbonyl)-methoxyphenyl)butyramide (2a)



¹H NMR (500 MHz, CDCl₃)

(S)-N-((S)-1-(butylamino)-3-methyl-1-oxobutan-2-yl)-2-((S)-2-isobutyramido-3-phenylpropanamido)-4-methylpentanamide



^{13}C NMR (126 MHz, CDCl_3)

(*S*)-*N*-((*S*)-1-(butylamino)-3-methyl-1-oxobutan-2-yl)-2-((*S*)-2-isobutyramido-3-phenylpropanamido)-4-methylpentanamide

