

## Supporting Information

for

# A Calixpyridinium-Based Supramolecular Tandem Assay for Alkaline Phosphatase and Its Application to ATP Hydrolysis Reaction

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**Table S1.** Chemical Shift Changes ( $\Delta\delta$ , ppm) of Calixpyridinium Protons in the Presence of ATP, ADP, and AMP at pD 7.2<sup>a,b</sup>

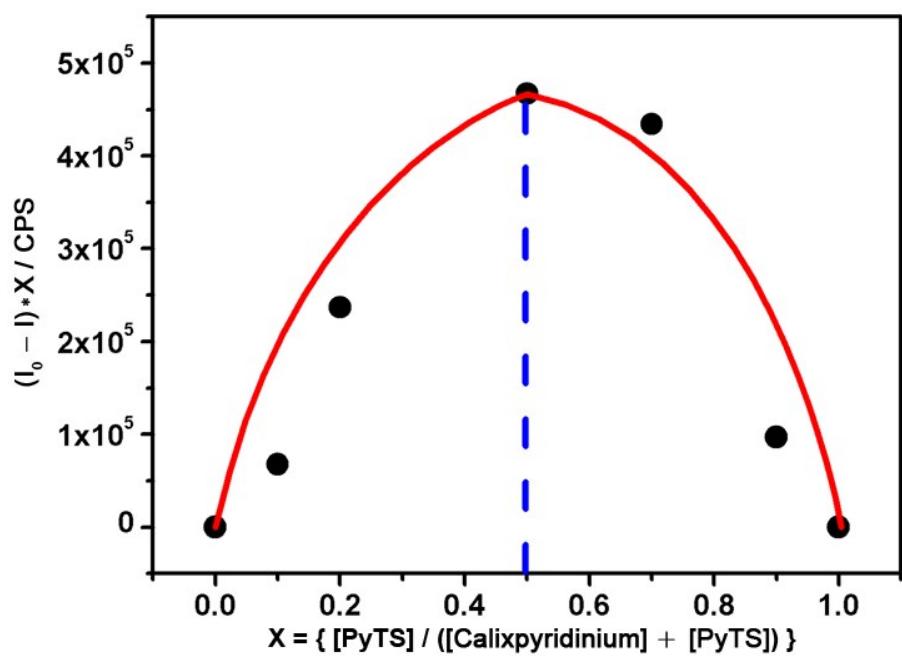
	H <sub>a</sub>	H <sub>b</sub>	H <sub>c</sub>	H <sub>d</sub>	H <sub>e</sub>
ATP	0.85	-0.14	-0.20	-0.11	0.03
ADP	0.20	-0.05	-0.07	-0.04	-0.03
AMP	0.17	-0.04	-0.05	-0.04	-0.03

<sup>a</sup>  $\Delta\delta = \delta(\text{presence of 1 equiv of guest}) - \delta(\text{free host})$ . <sup>b</sup> The host and guest were mixed in a 1:1 stoichiometry at 5 mM.

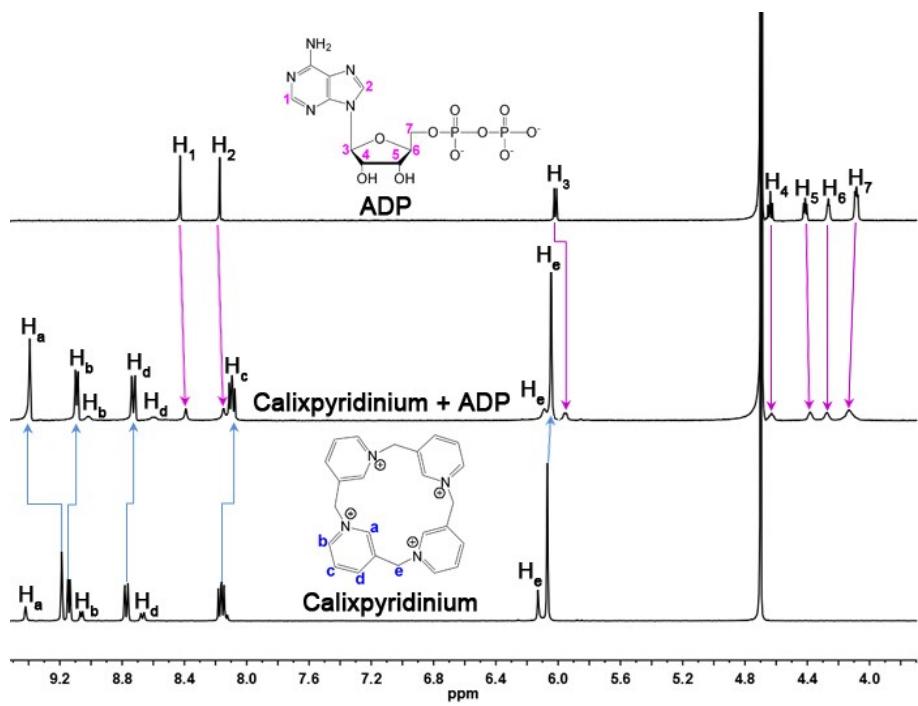
**Table S2.** Chemical Shift Changes ( $\Delta\delta$ , ppm) of ATP, ADP, and AMP Protons in the Presence of Calixpyridinium at pD 7.2<sup>a,b</sup>

guests	H <sub>1</sub>	H <sub>2</sub>	H <sub>3</sub>	H <sub>4</sub>	H <sub>5</sub>	H <sub>6</sub>	H <sub>7</sub>
ATP	-0.06	-0.06	-0.10	-0.01	-0.01	0.03	0.09
ADP	-0.04	-0.02	-0.07	-0.01	-0.03	0.01	0.05
AMP	-0.07	-0.08	-0.09	-0.02	-0.03	0.00	0.03

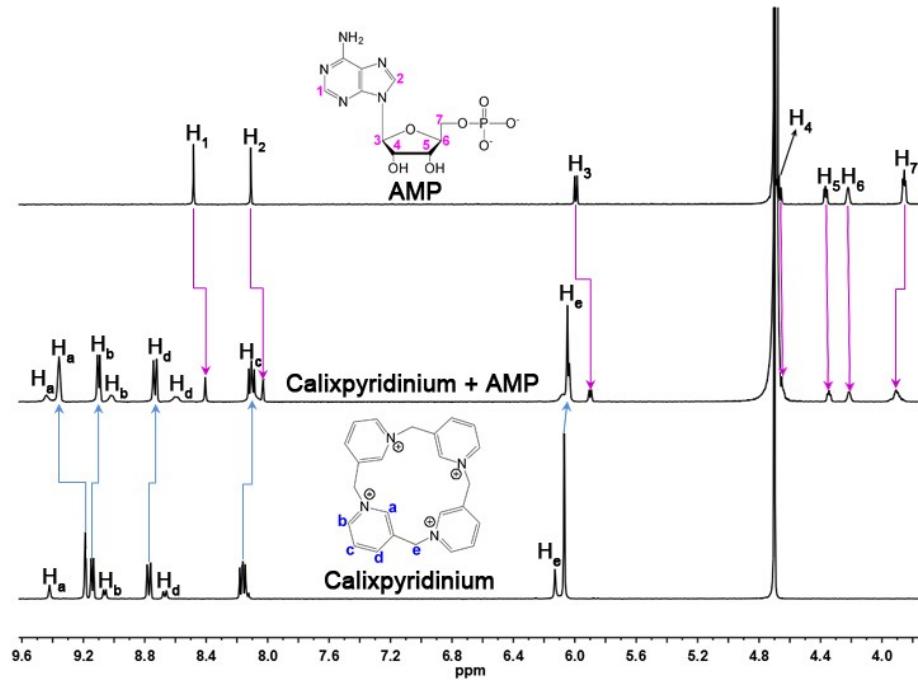
<sup>a</sup>  $\Delta\delta = \delta(\text{presence of 1 equiv of host}) - \delta(\text{free guest})$ . <sup>b</sup> The host and guest were mixed in a 1:1 stoichiometry at 5 mM.



**Figure S1.** Job's plot for calixpyridinium and PyTS in 10 mM NaOAc solution at pH 7.2 ( $\lambda_{\text{exc}} = 339$  nm,  $\lambda_{\text{obs}} = 385$  nm),  $[\text{calixpyridinium}] + [\text{PyTS}] = 4 \mu\text{M}$ .

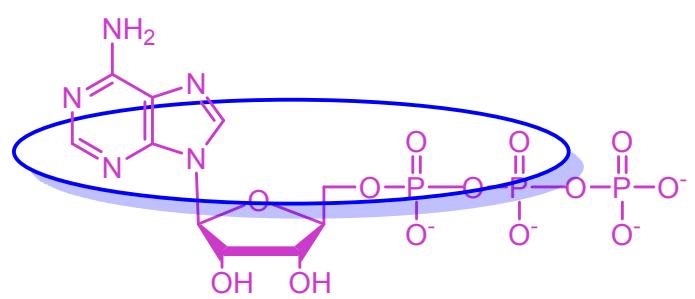


(a)

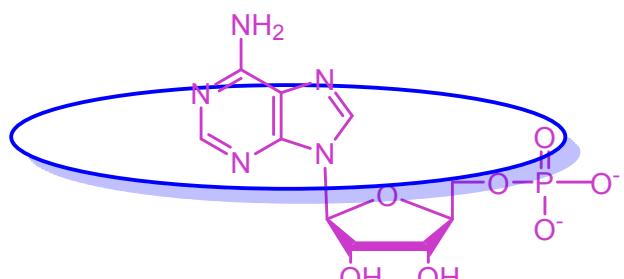


(b)

**Figure S2.** (a) <sup>1</sup>H NMR spectra of ADP, calixpyridinium, and calixpyridinium+ADP complex at pD 7.2; (b) <sup>1</sup>H NMR spectra of AMP, calixpyridinium, and calixpyridinium+AMP complex at pD 7.2. The host and guest were mixed in a 1:1 stoichiometry at 5 mM.

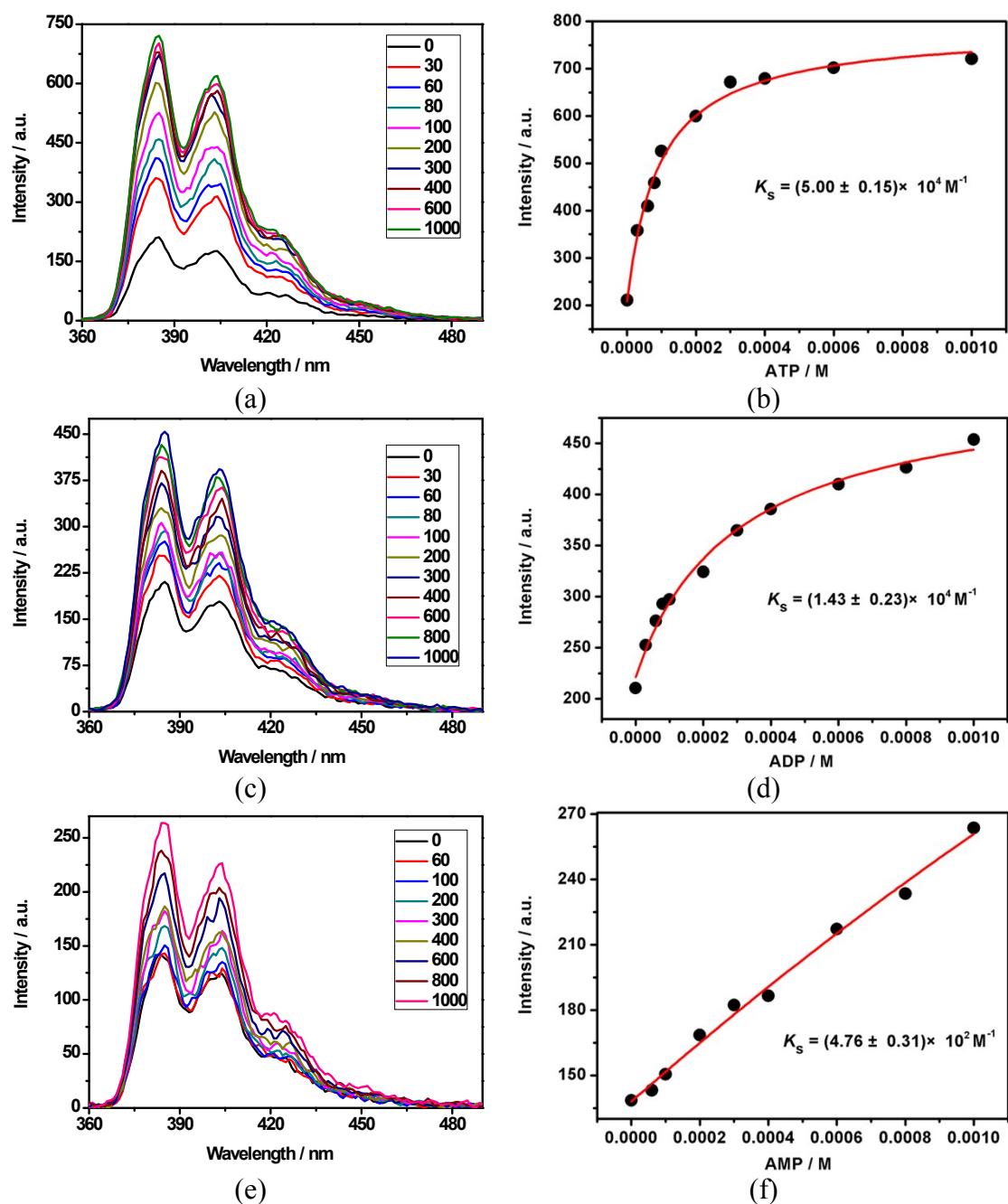


(a)

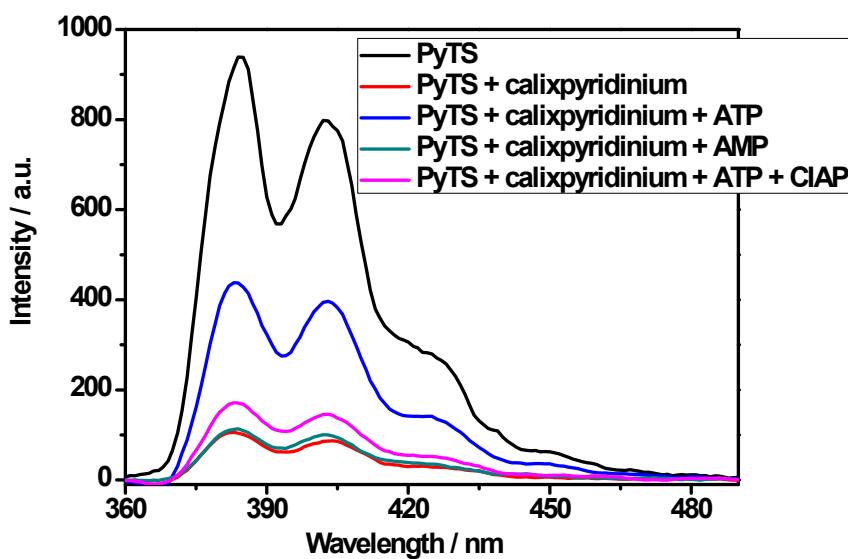


(b)

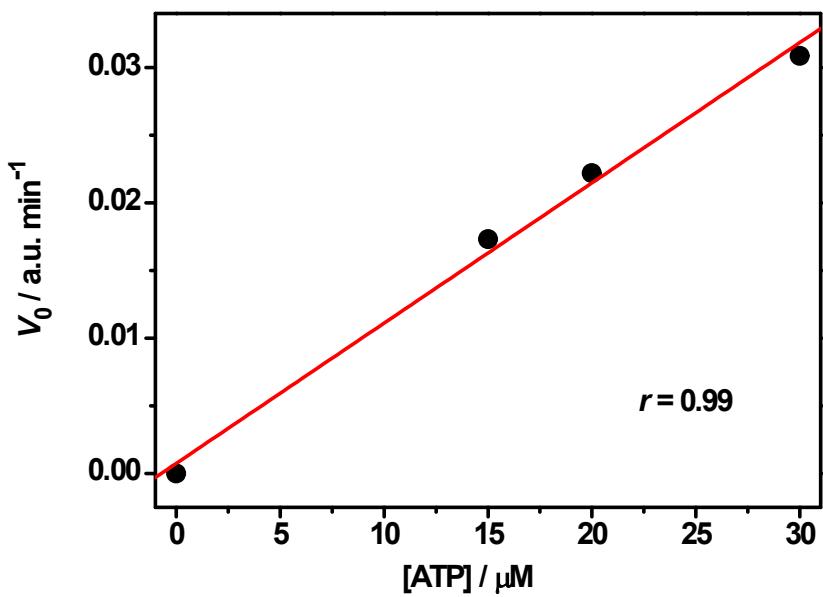
**Figure S3.** Deduced binding modes of calixpyridinium with ATP (a) and AMP (b) at pH 7.2 according to  $^1\text{H}$  NMR spectra.



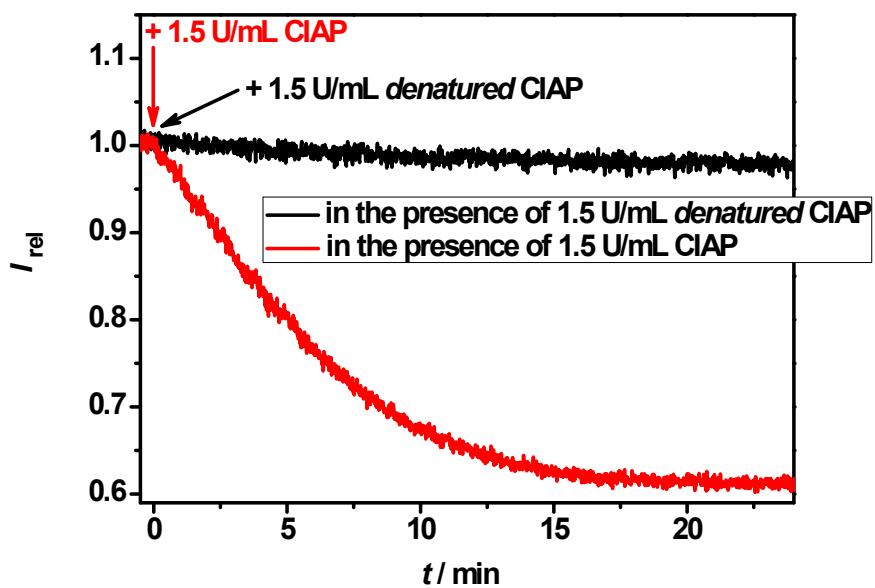
**Figure S4.** Competitive fluorescence titrations of ATP (a and b), ADP (c and d), and AMP (e and f) in the presence of PyTS (1.0  $\mu\text{M}$ ) and calixpyridinium (4.0  $\mu\text{M}$ ) in 10 mM NaOAc solution at pH 7.2 ( $\lambda_{\text{exc}} = 339 \text{ nm}$ ,  $\lambda_{\text{obs}} = 385 \text{ nm}$ ).



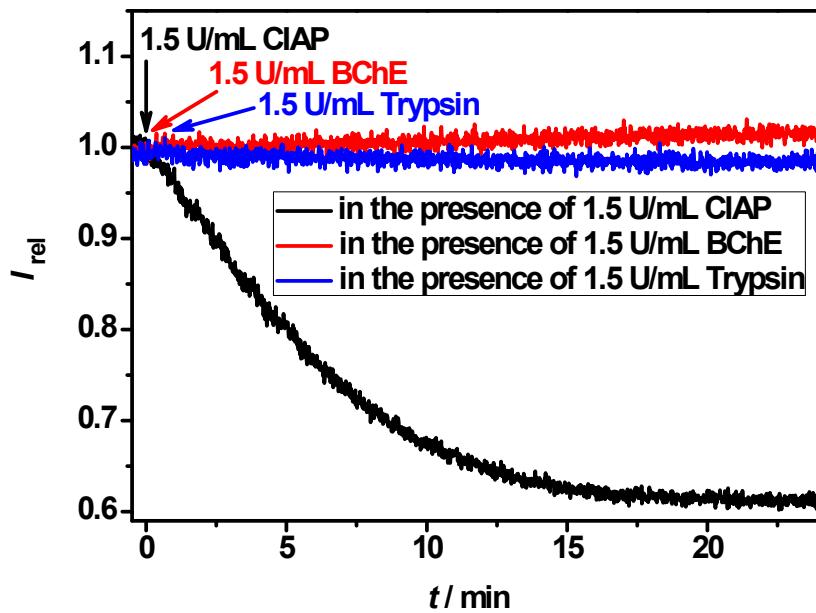
**Figure S5.** Fluorescence emission spectra of PyTS, PyTS+calixpyridinium, PyTS+calixpyridinium+ATP, PyTS+calixpyridinium+AMP, and PyTS+calixpyridinium+ATP+CIAP in 10 mM NaOAc solution at pH 7.2, excited at 339 nm.. The concentrations of PyTS, calixpyridinium, ATP, AMP, and CIAP are 1.0  $\mu$ M, 4.0  $\mu$ M, 60  $\mu$ M, 60  $\mu$ M, and 1.5 U/mL, respectively.



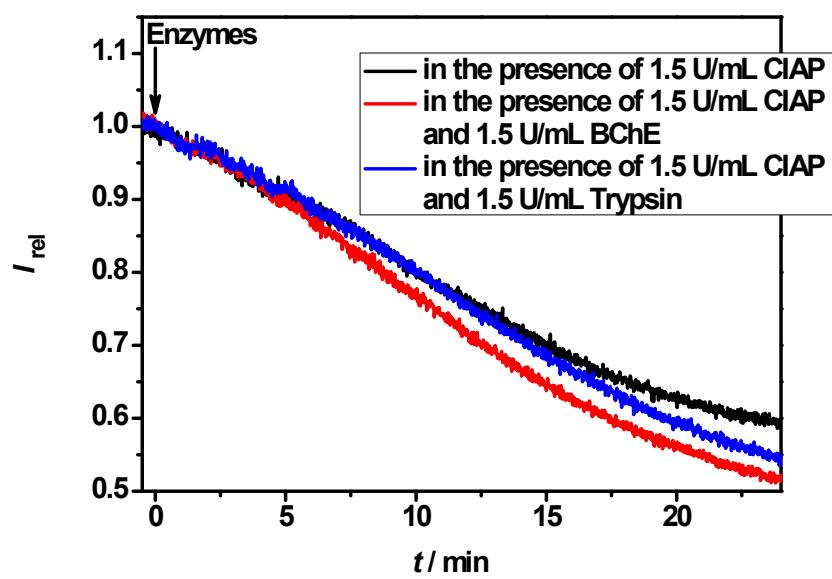
**Figure S6.** Linear relationship between the initial reaction rates and the concentrations of ATP (0–30  $\mu\text{M}$  with 1.5 U/mL CIAP, 1.0  $\mu\text{M}$  PyTS, and 4.0  $\mu\text{M}$  calixpyridinium) in 10 mM NaOAc solution at pH 7.2 ( $\lambda_{\text{exc}} = 339 \text{ nm}$ ,  $\lambda_{\text{obs}} = 385 \text{ nm}$ ).



**Figure S7.** Continuous fluorescent enzyme assays for CIAP and *denatured* CIAP (1.5 U/mL) with the calixpyridinium/PyTS reporter pair (60  $\mu$ M ATP, 1.0  $\mu$ M PyTS, and 4.0  $\mu$ M calixpyridinium in 10 mM NaOAc solution at pH 7.2;  $\lambda_{\text{exc}} = 339$  nm,  $\lambda_{\text{obs}} = 385$  nm).



**Figure S8.** Continuous fluorescent enzyme assays for CIAP, BChE, and Trypsin (1.5 U/mL) with the calixpyridinium/PyTS reporter pair (60  $\mu$ M ATP, 1.0  $\mu$ M PyTS, and 4.0  $\mu$ M calixpyridinium in 10 mM NaOAc solution at pH 7.2;  $\lambda_{\text{exc}} = 339$  nm,  $\lambda_{\text{obs}} = 385$  nm).



**Figure S9.** Continuous fluorescent enzyme assays for CIAP (1.5 U/mL) in the absence and presence of BChE (1.5 U/mL) or Trypsin (1.5 U/mL) with the calixpyridinium/PyTS reporter pair (60  $\mu\text{M}$  ATP, 1.0  $\mu\text{M}$  PyTS, and 4.0  $\mu\text{M}$  calixpyridinium in 10 mM NaOAc solution at pH 7.2;  $\lambda_{\text{exc}} = 339 \text{ nm}$ ,  $\lambda_{\text{obs}} = 385 \text{ nm}$ ).