A novel 3, 6-diamino-1, 8-naphthalimide derivative as a highly selective fluorescent "turn-on" probe for thiols

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Supporting Information

Experimental

Generals. ¹H and ¹³C NMR spectra were recorded in *d*₆-DMSO, CD₃OD, CDCl₃ or *d*₆-DMSO-D₂O (5/2, v/v) on a Varian Mercury 400 spectrometer. LR-ESI-MS and HR-ESI-MS spectra were measured on Waters UPLC/Quattro Premier XE and Agilent 6460 Triple Quadrupole mass spectrometers, respectively. Silica gel 60 Å (reagent pure, Qingdao Haiyang Chemical Co. Ltd) was used for column chromatography. Analytical thin-layer chromatography was performed on silica gel plates 60 GF254 (chemical pure, Qingdao Haiyang Chemical Co. Ltd). Detection on TLC was made by use of a UV lamp (254 or 365 nm). Fluorescence spectra were measured on a Shimadzu RF-5301PC spectrofluorimeter. UV-Vis spectra were undertaken with Olympus FV1000 confocal laser scanning microscopy.

N-butyl-3, 6-dinitro-1, 8-naphthalimide **3** was prepared according to the reported literature. ¹ All reagents and solvents were purchased from commercial sources and were of analytical grade. Solvents were dried according to standard procedures.

Synthesis of compounds 1 and 2

Compound **2**. To a solution of compound **3** (155 mg, 0.45 mmol) in ethanol (10 mL) was added 20 mg of 10% Pd/C in ethanol (3 mL). The resulting mixture was stirred under the atmosphere of hydrogen at 40 ° C for 3 h, and then concentrated under reduced pressure. The obtained residue was purified by chromatography on a silica gel column, eluted with CHCl₃/CH₃OH (400/1, v/v) to afford compound **2** (108 mg, 85%) as a yellowish solid having ¹H NMR (400 MHz, *d*₆-DMSO) δ 7.58 (s, 2H), 6.93 (s, 2H), 5.68 (s, 4H), 3.98 (t, *J* = 7.2 Hz, 2H), 1.57 (m, 2H), 1.35 (m, 2H), 0.92 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, *d*₆-DMSO) δ 164.3, 148.0, 135.9, 122.7, 117.4, 110.0, 30.1, 20.2, 14.1 and ESI-MS *m/z*: 284.2 ([M+H]⁺).

Compound **1**. A solution of compound **2** (28 mg, 0.10 mmol) and 2, 6-dimethylpyridine (54 mg, 0.50 mmol) in anhydrous CH₂Cl₂ (20 mL) was stirred at 0 °C for 15 min and then added to a solution of 2, 4-dinitrobenzenesulfonyl chloride (133 mg, 0.50 mmol) in anhydrous CH₂Cl₂ (10 mL). The resulting mixture was stirred at room temperature for 45 h and concentrated under reduced pressure. The obtained residue was purified by chromatography on a silica gel column, eluted with CHCl₃/CH₃OH (200/1, v/v) to afford compound **1** (14 mg, 28%) as a yellow solid having ¹H NMR (400 MHz, *d*₆-DMSO) δ 8.89 (d, *J* = 2.0 Hz, 1H), 8.58 (m, 1H), 8.30 (d, *J* = 3.2 Hz, 1H), 7.87 (d, *J* = 2.4 Hz, 1H), 7.86 (d, *J* = 2.4 Hz, 1H), 7.69 (d, *J* = 2.0 Hz, 1H), 7.14 (d, *J* = 2.0 Hz, 1H), 3.97 (t, *J* = 7.2 Hz, 2H), 1.56 (m, 2H), 1.32 (m, 2H), 0.91 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, *d*₆-DMSO) δ 163.7, 163.5, 150.6, 149.0, 148.3, 136.4, 135.4, 134.6, 132.1, 127.7, 123.6, 123.0, 121.6, 120.8, 120.6, 119.4, 118.5, 111.2, 30.0, 20.2, 14.1; ESI-MS *m/z*: 514.2 ([M+H]⁺) and HR-ESI-MS for C₂₂H₁₉N₅O₈S ([M+H]⁺) calcd: 514.1033, found: 514.1029.

Determination of quantum yields

The quantum yield of compound **2** was measured by using the protocol described in literature. ² Thus, the absorption and emission spectra of compound **2** were measured in ethanol. It showed absorption and emission maxima at 415 nm ($\varepsilon = 8300 \text{ M}^{-1} \cdot \text{cm}^{-1}$) and 560 nm, respectively. Coumarin **4** was chosen as a standard because its absorption and emission spectra largely overlap those of compound **2** in ethanol. Fluorescence quantum yield Φ_X of compound **2** was calculated according to eq (1).

$$\Phi_{\rm X} = \Phi_{\rm S} \times [{\rm Abs}_{\rm S} / {\rm Abs}_{\rm X}] \times [{\rm A}_{\rm FX} / {\rm A}_{\rm FS}] \times [{\rm N}_{\rm X} / {\rm N}_{\rm S}]^2$$
(1)

Where, Φ_S (= 0.78) is the reported quantum yield of coumarin 4 in ethanol; Abs_S and Abs_X are the absorbance at the excitation wavelengths of coumarin 4 and compound 2, respectively; A_{FS} and A_{FX} are the areas under the emission spectra of coumarin 4 and compound 2, respectively; and N_S and N_X are the refractive indexes of the solvents used for coumarin 4 and compound 2, respectively. Here ethanol was used in both cases, thus N_S = N_X = 1.3611. It has been reported that *N*-butyl-4-amino-1, 8-naphthalimide has the absorption and emission maxima at 430 nm (ϵ = 11600 M⁻¹·cm⁻¹) and 541 nm (Φ = 0.13), respectively. ³

Sample	Abs	\mathbf{A}_{F}	Φ
Coumarin 4	0.0464	31189	0.78
Compound 2	0.0160	819	0.0594

Table 1 Results for coumarin 4 and compound 2

Procedures of thiols sensing

A stock solution of compound 1 (1 mM) was prepared in DMSO and then diluted to the corresponding concentration (25 μ M) with 1 : 1 DMSO-PBS buffer (0.01 M, pH 7.4). Fluorescence spectra were recorded in an indicated time after the addition of amino acids and different kinds of common species ($\lambda_{ex} = 415$ nm, ex/em = 5 nm/5 nm).

Effect of pH value

The effect of pH on the fluorescence intensities of compound 1 (25 μ M) was determined in the absence and presence of Cys (250 μ M), respectively. The reaction of compound 1 and Cys was carried out at room temperature for 120 min, in 1 : 1 DMSO-PBS buffer (0.01 M) of varying pH from 3.0 to 9.0. The corresponding fluorescence spectrum at each pH was recorded ($\lambda_{ex} = 415$ nm.). The fluorescence intensities at 560 nm were plotted against pH, which is shown in Fig. S9.

Kinetic study

The kinetic profiles of the reaction were examined under *pseudo-first-order* conditions with a large excess of Cys, Hcy, or GSH over compound **1** in 1 : 1 DMSO-PBS (0.01 M, pH 7.4) at room temperature. The *pseudo-first-order* rate constant k was calculated according to eqn (2):

$$\ln[F_{\max}/(F_{\max} - F_{t})] = kt$$
⁽²⁾

Wherein F_t and F_{max} are the fluorescence intensity at 560 nm at time *t* and when the reaction is complete, respectively. *k* is the *pseudo-first-order* rate constant.⁴

Detection limit

The detection limit was calculated based on the fluorescence titration. The slit was adjusted to 10 nm. To determine the S/N ratio, the emission intensity of compound 1 (25 μ M) in the absence of Cys was measured by 10 times and the standard deviation of blank measurements was determined. Under the present conditions, a good linear relationship between the fluorescence intensity and the Cys concentration was obtained in the range of 0~100 μ M (R = 0.9998, Fig. S11). The detection limit was then calculated with the equation: detection limit = $3\sigma_{bi}/m$, wherein σ_{bi} is the standard deviation of blank measurements and *m* is the slope of the straight line between the fluorescence intensity and the Cys Concentration of Cys. ⁵ The detection limit was calculated to be 2.0×10^{-7} M at S/N = 3 (signal-to-noise ratio of 3 : 1) for Cys. Similarly, the detection limit was calculated to be 4.3×10^{-7} M for GSH (Fig. S12) and 1.2×10^{-6} M for Hcy (Fig. S13) at S/N = 3, respectively.

Fluorescence Imaging

HepG2 cells were cultured in Dulbecco's modified Eagle's medium (DMEM) with 10% fetal bovine serum (FBS) and 1% double resistant (50 U/mL penicillin, 50 μ g/mL streptomycin) at 37 °C under a humidified atmosphere containing 5% CO₂. Exponentially growing cells were placed in 15 mm culture dish (1×10⁶ cells/mL) and incubated at 37 °C for 12 h for attachment. HepG2 cells were treated with 5 μ M of compound **1** in culture media for 2 h at 30 °C and washed 3 times with PBS buffer (0.01 M, pH 7.4). For the control experiment, the cells were

treated with *N*-ethylmaleimide (NEM, 20 μ M) in culture media for 30 min at 30 °C. After washed with PBS buffer to remove the remaining NEM, the cells were further incubated with 5 μ M of compound 1 in culture media for 2 h at 30 °C. ⁶

References

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Fig. S1 ¹H NMR (400 MHz) of compound 2 in d_6 -DMSO.



Fig. S2 ¹³C NMR (100 MHz) of compound 2 in d_6 -DMSO.



Fig. S3 ¹H NMR (400 MHz) of compound 1 in d_6 -DMSO.



Fig. S4 ¹³C NMR (100 MHz) of compound 1 in d_6 -DMSO.



Fig. S5 HR-ESI-MS of compound 1.



Fig. S6 ¹H NMR spectra of compound **1** in the absence (a), presence of Cys for 18 h (b) and for 72 h (c), and of compound **2** (d) in d_6 -DMSO/D₂O (5/2, v/v).







Fig. S8 LR-ESI-MS of compound 1 titrated with Cys.



Fig. S9 Effect of pH on the fluorescence intensities of compound 1 (25 μ M) in the absence (•) and presence (**A**) of Cys (250 μ M). The reaction of compound **1** and Cys was carried out at room temperature for 120 min, in 1 : 1 DMSO-PBS buffer (0.01 M) of varying pH from 3 to 9. $\lambda_{ex}/\lambda_{em} = 415/560$ nm.



Fig. S10 (a) Fluorescence response of compound 1 (25 μ M) to Cys of varying concentrations in 1 : 1 DMSO-PBS buffer (0.01 M, pH 7.4) ($\lambda_{ex}/\lambda_{em} = 415/560$ nm, ex/em 5/5 nm). (b) Fluorescence intensity at 560 nm of compound 1 (25 μ M) upon the addition of Cys (0~100 μ M).



Fig. S11 (a) Fluorescence response of compound 1 (25 μ M) to GSH of varying concentrations in 1 : 1 DMSO-PBS buffer (0.01 M, pH 7.4) ($\lambda_{ex}/\lambda_{em} = 415/560$ nm, ex/em 5/5 nm). (b) Fluorescence intensity at 560 nm of compound 1 (25 μ M) upon the addition of GSH (0~150 μ M).



Fig. S12 (a) Fluorescence response of compound 1 (25 μ M) to Hcy of varying concentrations in 1 : 1 DMSO-PBS buffer (0.01 M, pH 7.4) ($\lambda_{ex}/\lambda_{em} = 415/560$ nm, ex/em 5/5 nm). (b) Fluorescence intensity at 560 nm of compound 1 (25 μ M) upon the addition of Hcy (0~150 μ M).



Fig. S13 Time-dependent fluorescence response of compound 1 (25 μ M) to GSH (1 mM) at room temperature in 1 : 1 DMSO-PBS buffer (0.01 M, pH 7.4).



Fig. S14 Time-dependent fluorescence response of compound **1** (25 μ M) to Hcy (1 mM) at room temperature in 1 : 1 DMSO-PBS buffer (0.01 M, pH 7.4).