

Supporting Information

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EXPNO    1
PROCNO    1

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NS         16
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FIDRES     0.152588 Hz
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RG         30.72
RW         50.000 usec
DE         6.50 usec
TE         294.9 K
D1         1.00000000 sec
TEC        1

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NUC1       1H
P1         13.00 usec
PLW1      12.00000000 W

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SF         500.1300124 MHz
WDW        EM
SSB        0
LB         0.30 Hz
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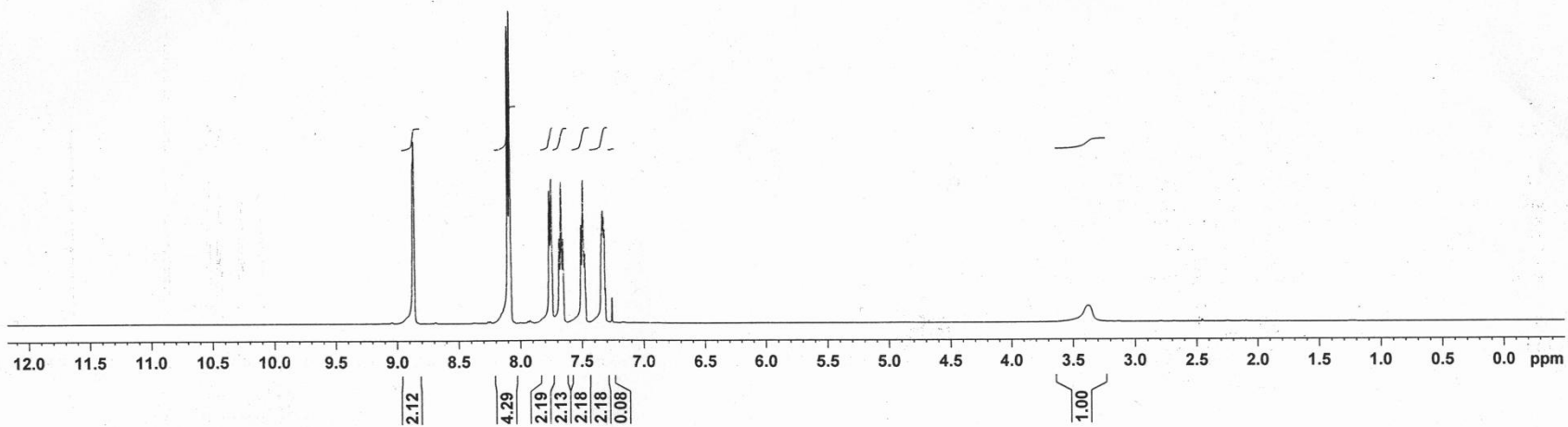


Figure S1. ¹H NMR Spectrum of 2a

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PROCNO 1

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PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 64
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010048 sec
RG 197.27
DW 16.800 usec
DE 6.50 usec
TE 299.6 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

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NUC1 13C
P1 8.90 usec
PLW1 103.00000000 W

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NUC2 1H
CPDPRG[2] waltz16
PCPD2 80.00 usec
PLW2 13.00000000 W
PLW12 0.34327999 W
PLW13 0.21969999 W

F2 - Processing parameters
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WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

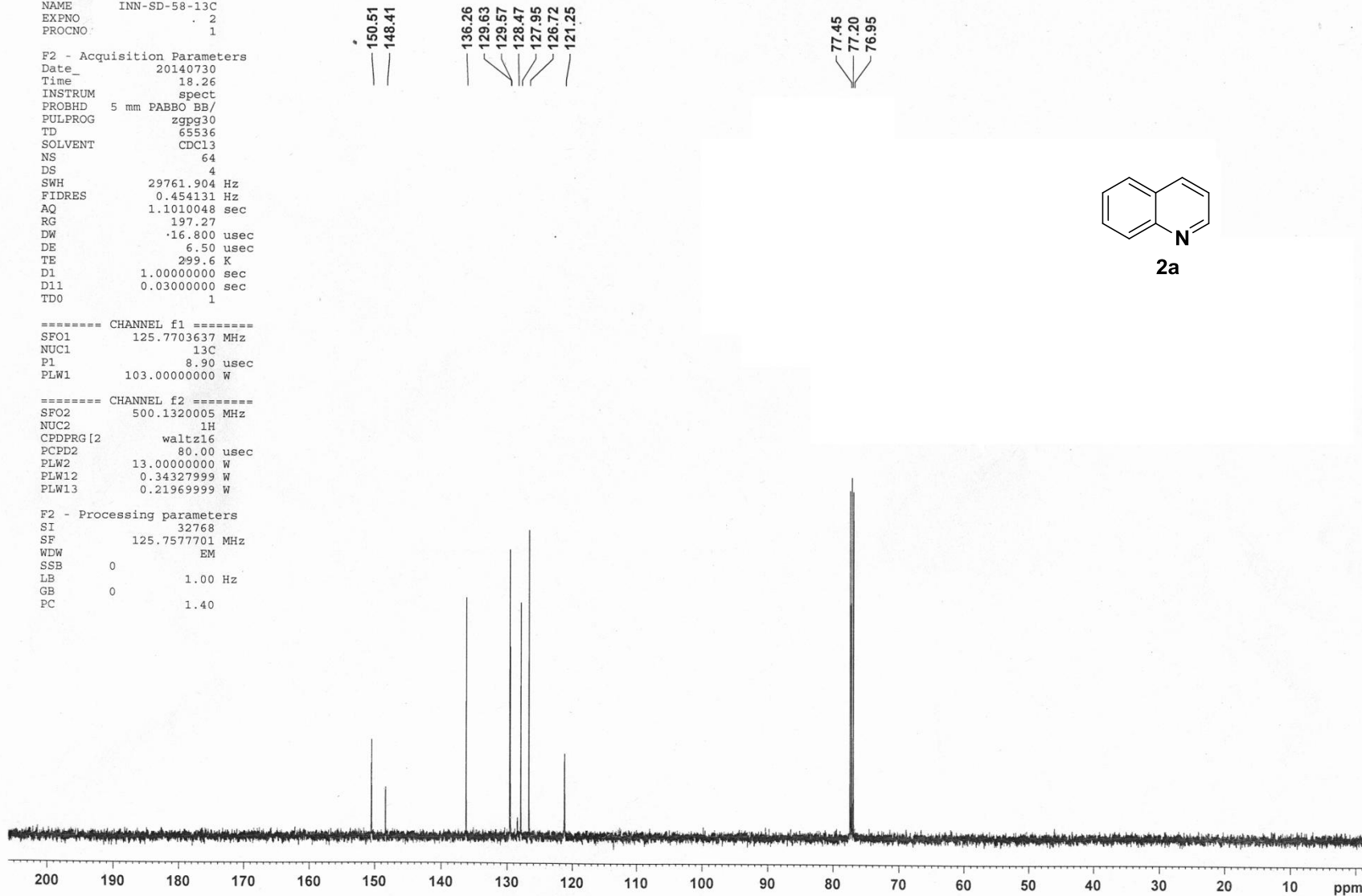


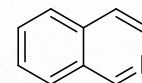
Figure S2. ^{13}C NMR Spectrum of 2a

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EXPNO 16
PROCNO 1

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Time_ 22.25
INSTRUM spect
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PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10000.000 Hz
FIDRES 0.152588 Hz
AQ 3.2767999 sec
RG 69.35
DW 50.000 usec
DE 6.50 usec
TE 296.6 K
D1 1.0000000 sec
TDO 1

==== CHANNEL f1 =====
SF01 500.1330885 MHz
NUC1 1H
F1 13.00 usec
PLW1 13.0000000 W

F2 - Processing parameters
SI 65536
SF 500.1300120 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
FC 1.00



2b

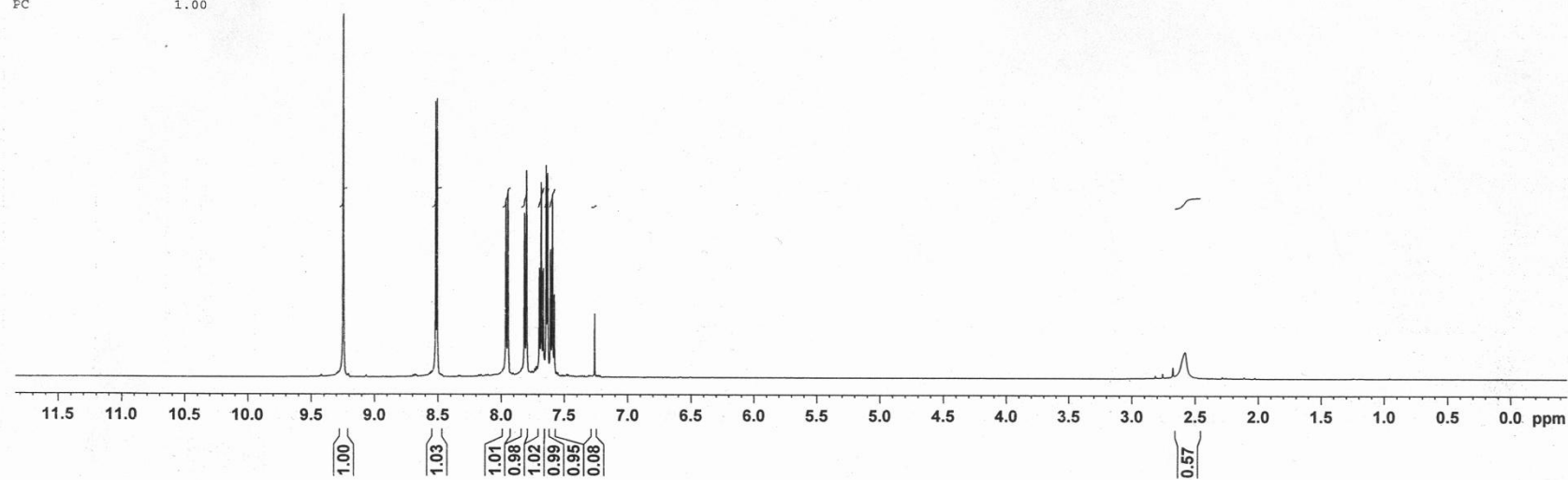


Figure S3. ¹H NMR Spectrum of 2b

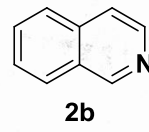
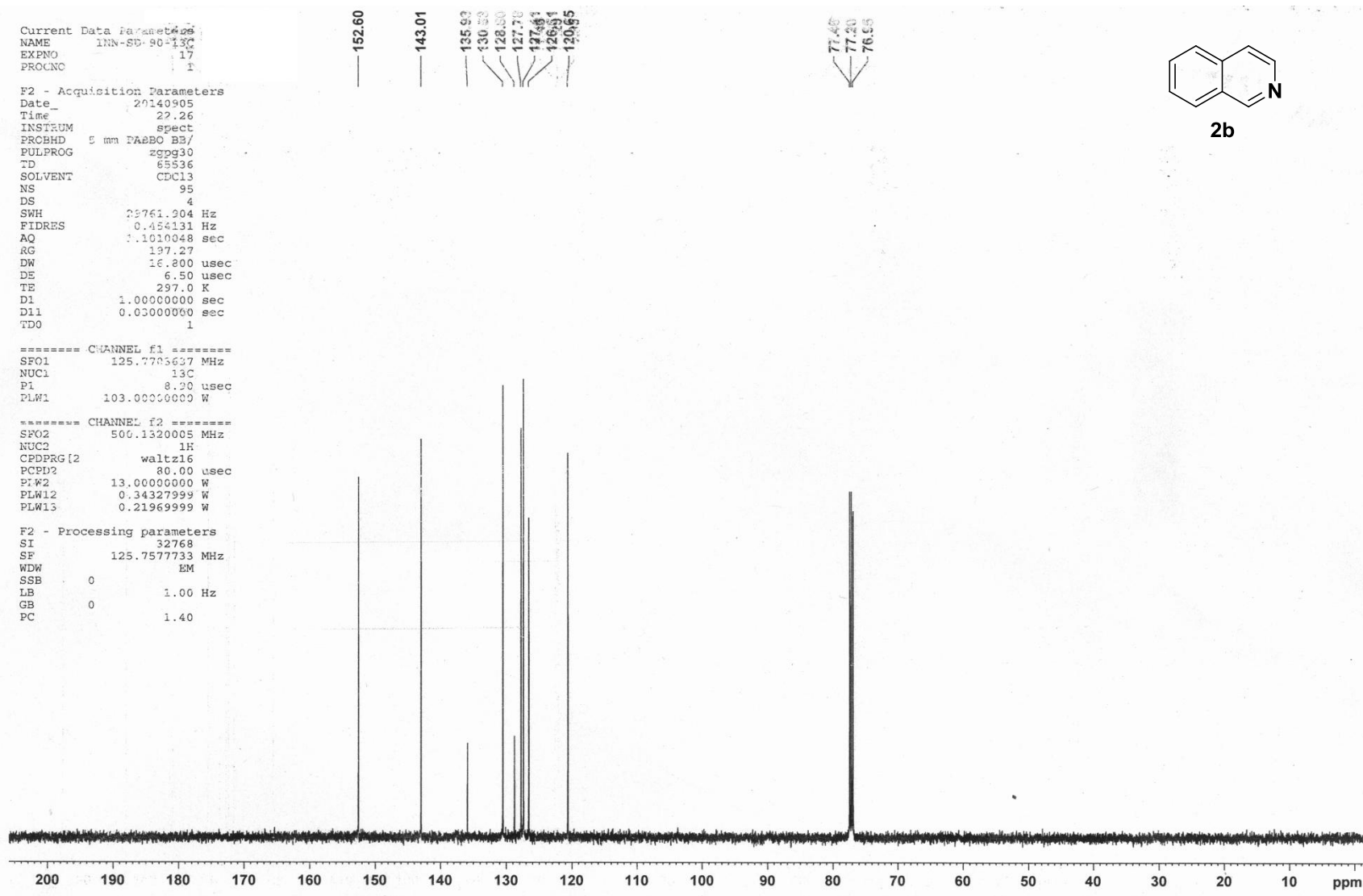
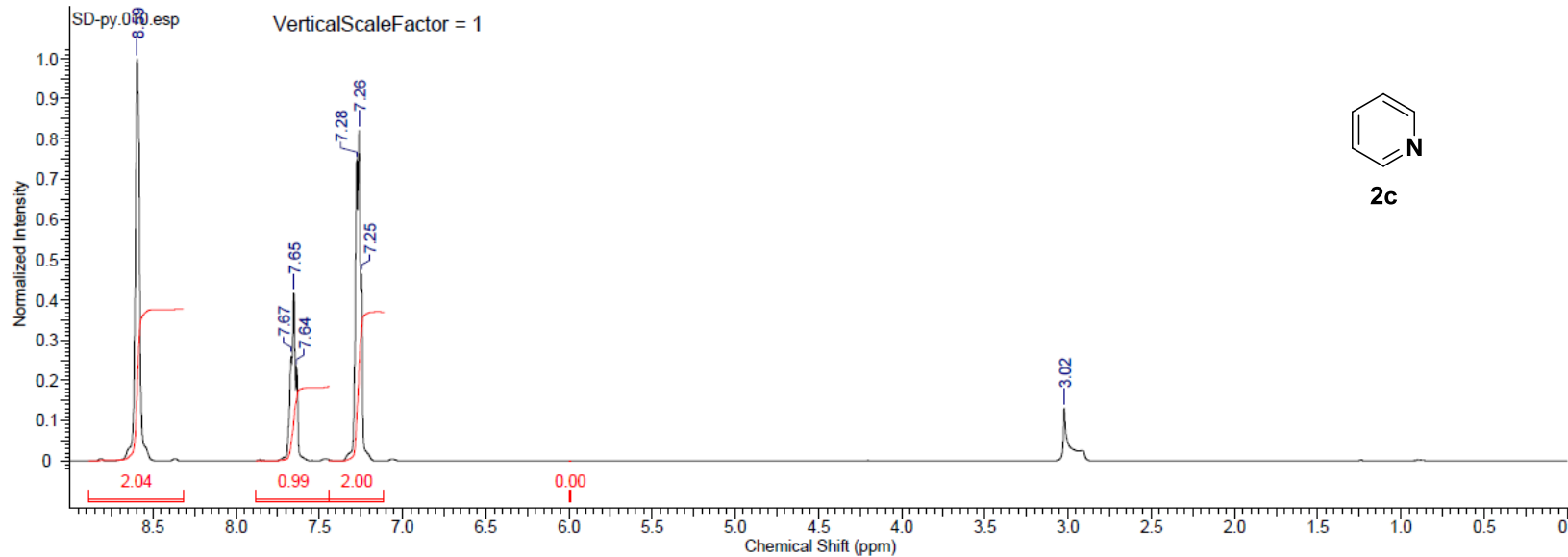
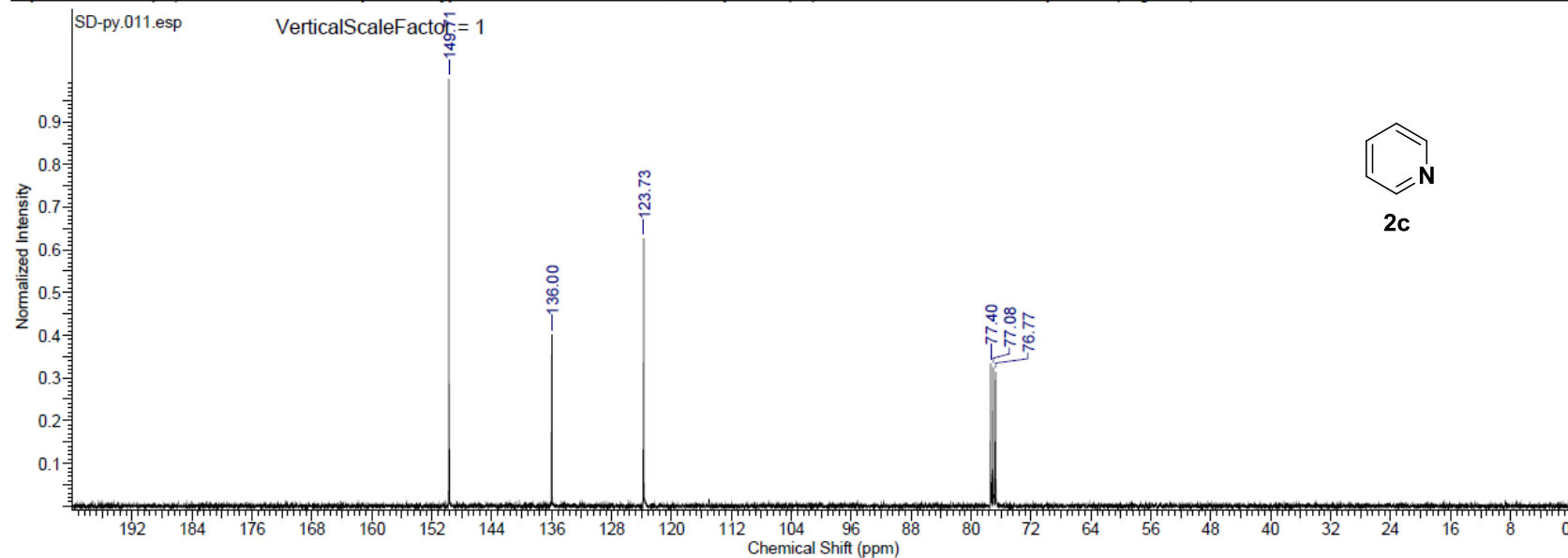


Figure S4. ¹³C NMR Spectrum of 2b

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Date Stamp	16 Oct 2014 16:26:08	File Name	\\labies\Bitec-s\partages\scbm\SMACB_RMNTritium\data\tritium\nmr\SD-py\10\fid		
Frequency (MHz)	400.13	Nucleus	1H	Number of Transients	8
Original Points Count	32768	Owner	BRUKER	Points Count	32768
Receiver Gain	114.00	SW(cyclical) (Hz)	5995.20	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2400.0000	Spectrum Type	STANDARD	Sweep Width (Hz)	5995.02
				Temperature (degree C)	25.400

Figure S5. ¹H NMR Spectrum of 2c

Acquisition Time (sec)	0.6521	Comment	5 mm PABBO BB-1H Z-GRD Z824801/0109	Date	16 Oct 2014 16:43:12		
Date Stamp	16 Oct 2014 16:43:12	File Name	\\labies\Bitec-s\partages\scbm\SMCB RMN\Tritium\data\tridium\nmr\SD-py\11\fid				
Frequency (MHz)	100.62	Nucleus	13C	Number of Transients	512	Origin	spect
Original Points Count	16384	Owner	BRUKER	Points Count	16384	Pulse Sequence	zpgq30
Receiver Gain	2896.30	SW(cyclical) (Hz)	25125.63	Solvent	CHLOROFORM-d		
Spectrum Offset (Hz)	11570.4688	Spectrum Type	STANDARD	Sweep Width (Hz)	25124.09	Temperature (degree C)	25.800

Figure S6. ^{13}C NMR Spectrum of 2c

NAME INN-SD-97-1H
EXPNO 3
PROCNO 1
Date_ 20140911
Time_ 19.46
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 54274
SOLVENT CDCl3
NS 11
DS 0
SWH 8223.685 Hz
FIDRES 0.151522 Hz
AQ 3.2999091 sec
RG 90.5
DW 60.800 usec
DE 6.50 usec
TE 297.1 K
D1 1.00000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 1H
P1 14.75 usec
PL1 -1.00 dB
PL1W 10.56200695 W
SFO1 400.1324710 MHz
SI 32768
SF 400.1300100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

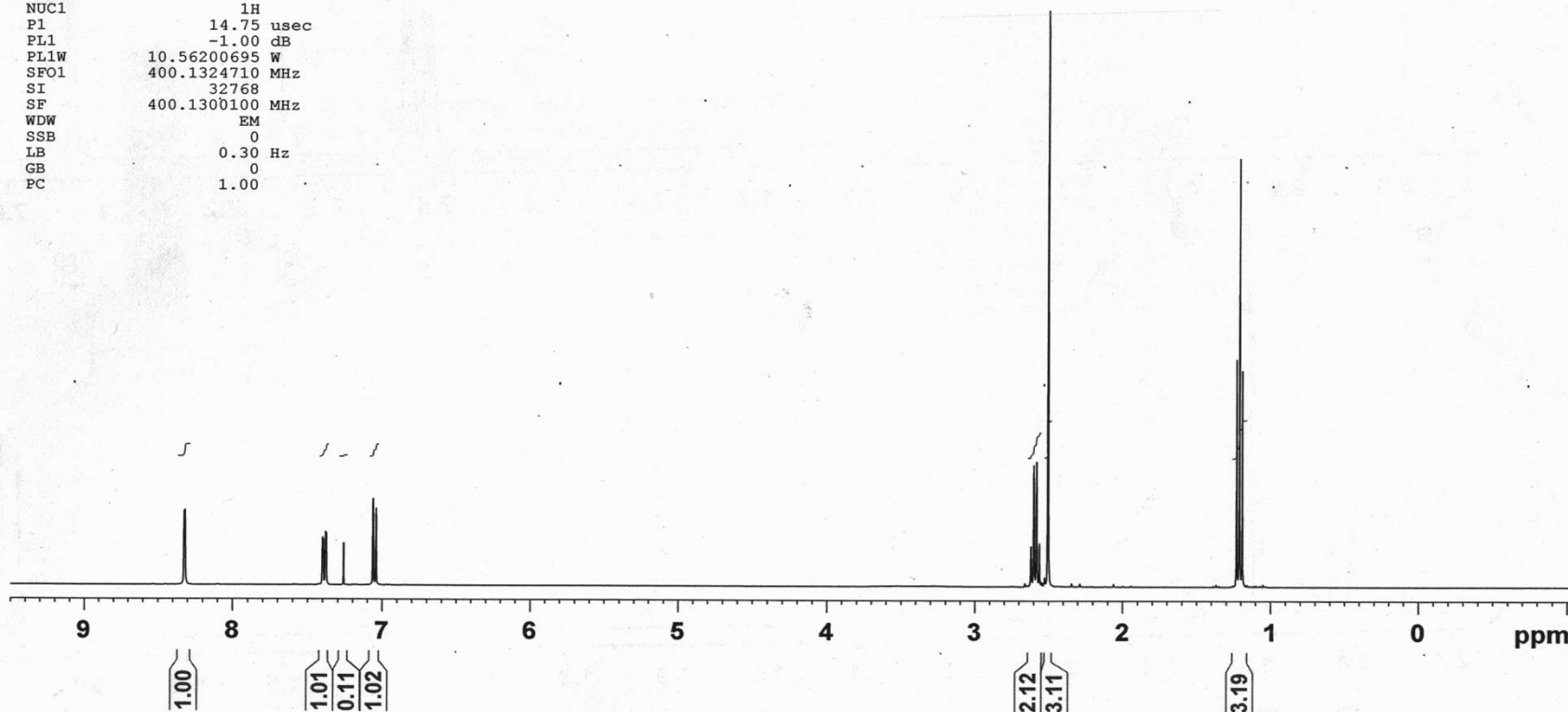
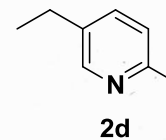


Figure S7. ¹H NMR Spectrum of 2d


```

NAME      INN-SD-97-13C
EXPNO    4
PROCNO   1
Date_    20140911
Time     19.48
INSTRUM  spect
PROBHD   5 mm PABBO BB-
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       100
DS       4
SWH      26041.666 Hz
FIDRES   0.397364 Hz
AQ       1.2583412 sec
RG       2050
DW       19.200 usec
DE       6.50 usec
TE       297.6 K
D1       1.00000000 sec
D11      0.03000000 sec
TD0      1

```

```

----- CHANNEL f1 -----
NUC1     13C
P1       8.50 usec
PL1      -2.00 dB
PL1W     56.53121948 W
SFO1     100.6238364 MHz

```

```

----- CHANNEL f2 -----
CPDPRG2  waltz16
NUC2     1H
PCPD2    80.00 usec
PL2      -1.00 dB
PL12     13.69 dB
PL13     14.50 dB
PL2W     10.56200695 W
PL12W    0.35871249 W
PL13W    0.29767781 W
SFO2     400.1316005 MHz
SI       32768
SF       100.6127536 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40

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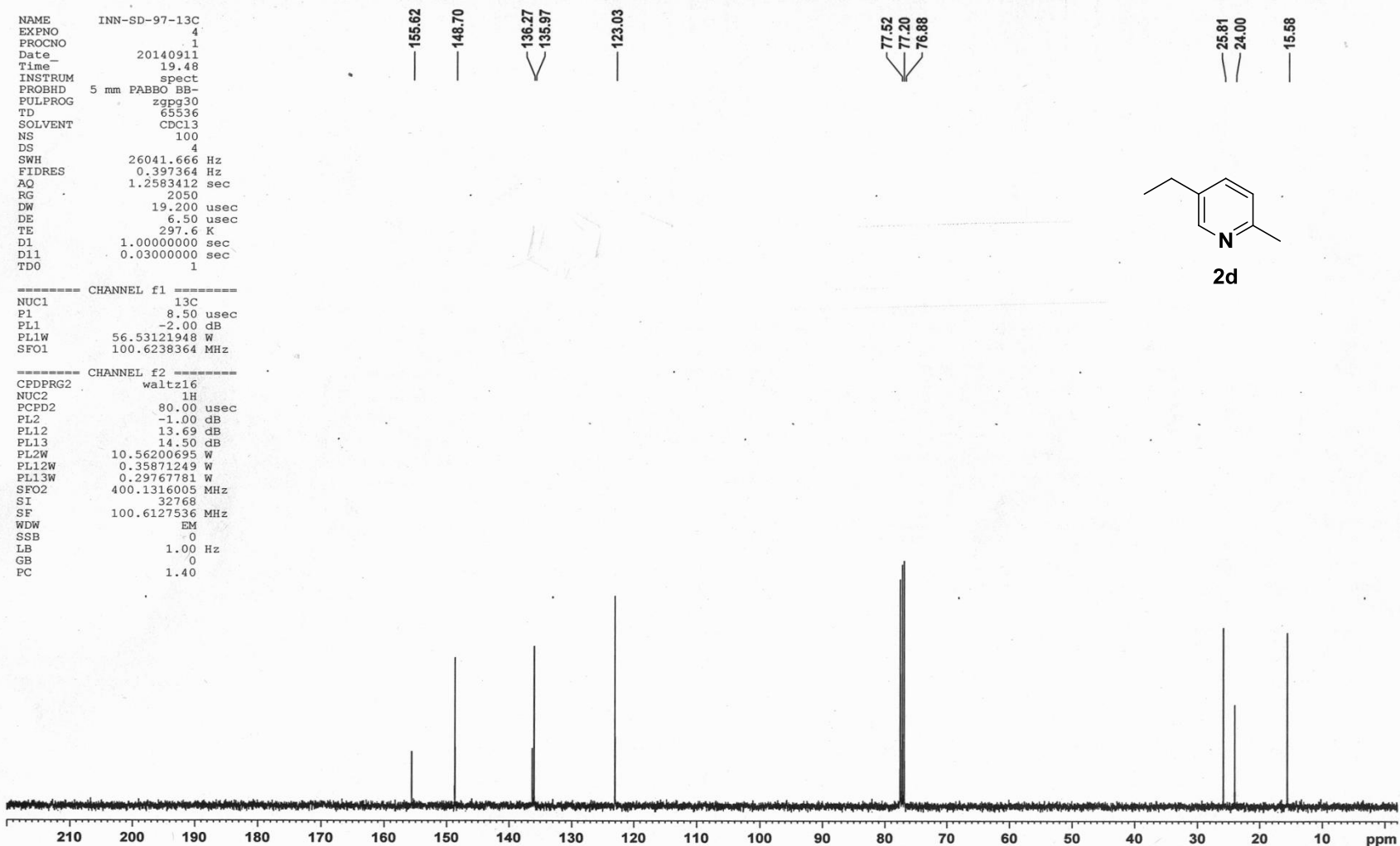


Figure S8. ¹³C NMR Spectrum of 2d

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Date Stamp	08 Oct 2014 11:36:00	File Name	\\labies\Bitec-s\partages\scbm\SMMCB_RMNI\Tritium\data\tridium\nmr\SD-361\20\fid		
Frequency (MHz)	400.13	Nucleus	¹ H	Number of Transients	16
Original Points Count	32768	Owner	BRUKER	Points Count	32768
Receiver Gain	228.10	SW(cyclical) (Hz)	5995.20	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2400.0000	Spectrum Type	STANDARD	Sweep Width (Hz)	5995.02
				Temperature (degree C)	23.500

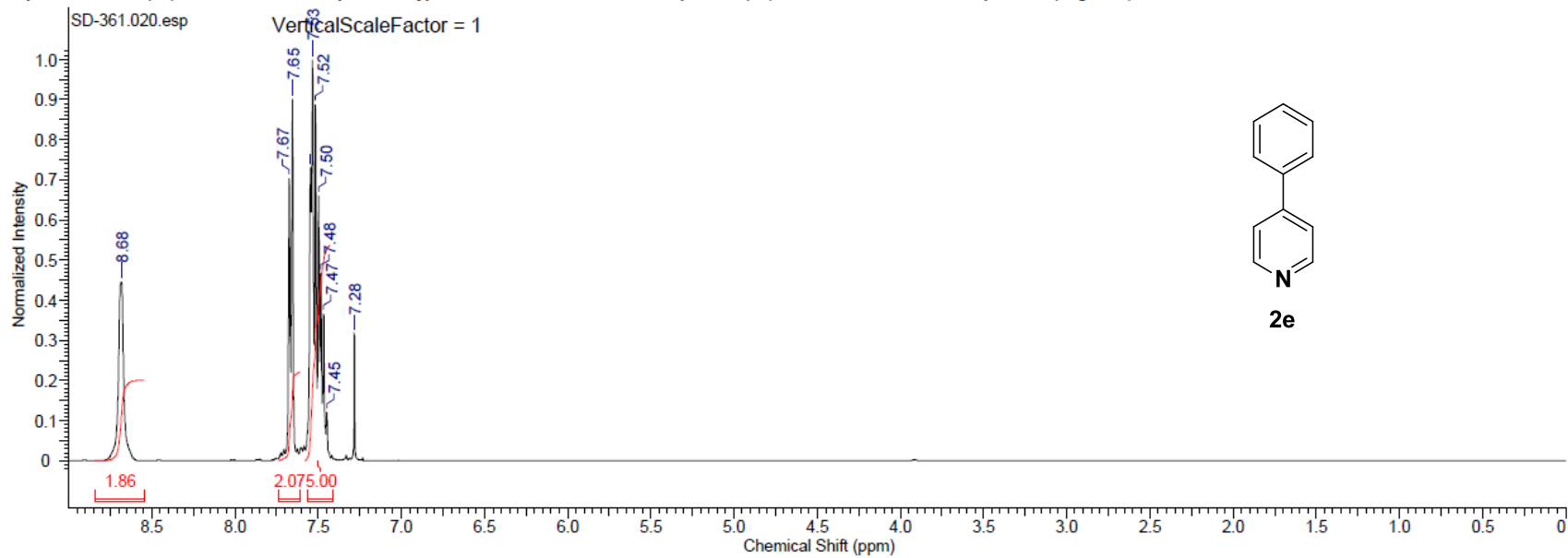


Figure S9. ¹H NMR Spectrum of 2e

Acquisition Time (sec)	0.6521	Comment	SD-361	Date	09 Oct 2014 07:30:40
Date Stamp	09 Oct 2014 07:30:40	File Name	\\labies\l\bitec-s\partages\scbm\SMCB RMN\Tritium\data\tridium\nmr\SD-361\10\fid		
Frequency (MHz)	100.62	Nucleus	¹³ C	Number of Transients	750
Original Points Count	16384	Owner	BRUKER	Points Count	16384
Receiver Gain	2896.30	SW(cyclical) (Hz)	25125.63	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11570.4688	Spectrum Type	STANDARD	Sweep Width (Hz)	25124.09
				Temperature (degree C)	24.700

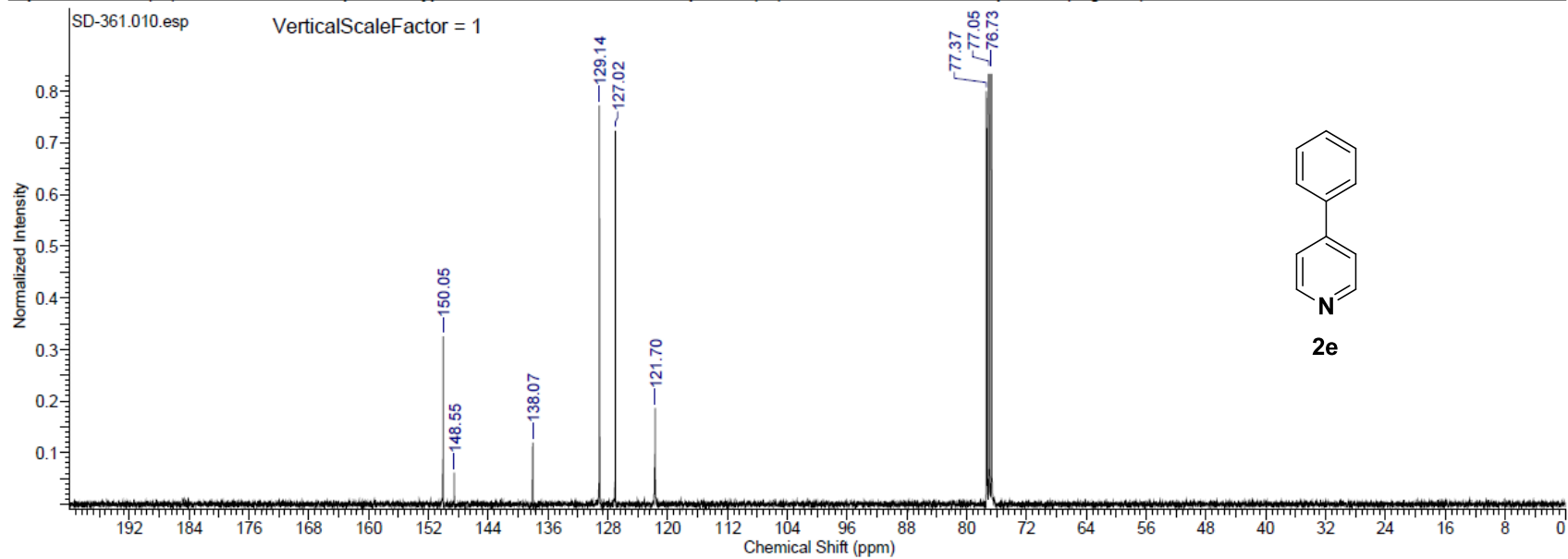
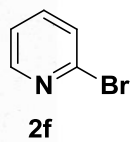


Figure S10. ¹³C NMR Spectrum of 2e



```

NAME      INN-SD-71-PURE-1H
EXPNO     1
PROCNO    1
Date_     20140902
Time_     18.11
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zg30
TD        54274
SOLVENT   CDC13
NS        11
DS        0
SWH       8223.685 Hz
FIDRES    0.151522 Hz
AQ        3.2999091 sec
RG        32
DW        60.800 usec
DE        6.50 usec
TE        297.6 K
D1        1.00000000 sec
TD0       1

```

```

===== CHANNEL f1 =====
NUC1      1H
P1        14.75 usec
PL1       -1.00 dB
PL1W      10.56200695 W
SFO1      400.1324710 MHz
SI        32768
SF        400.1300099 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

```

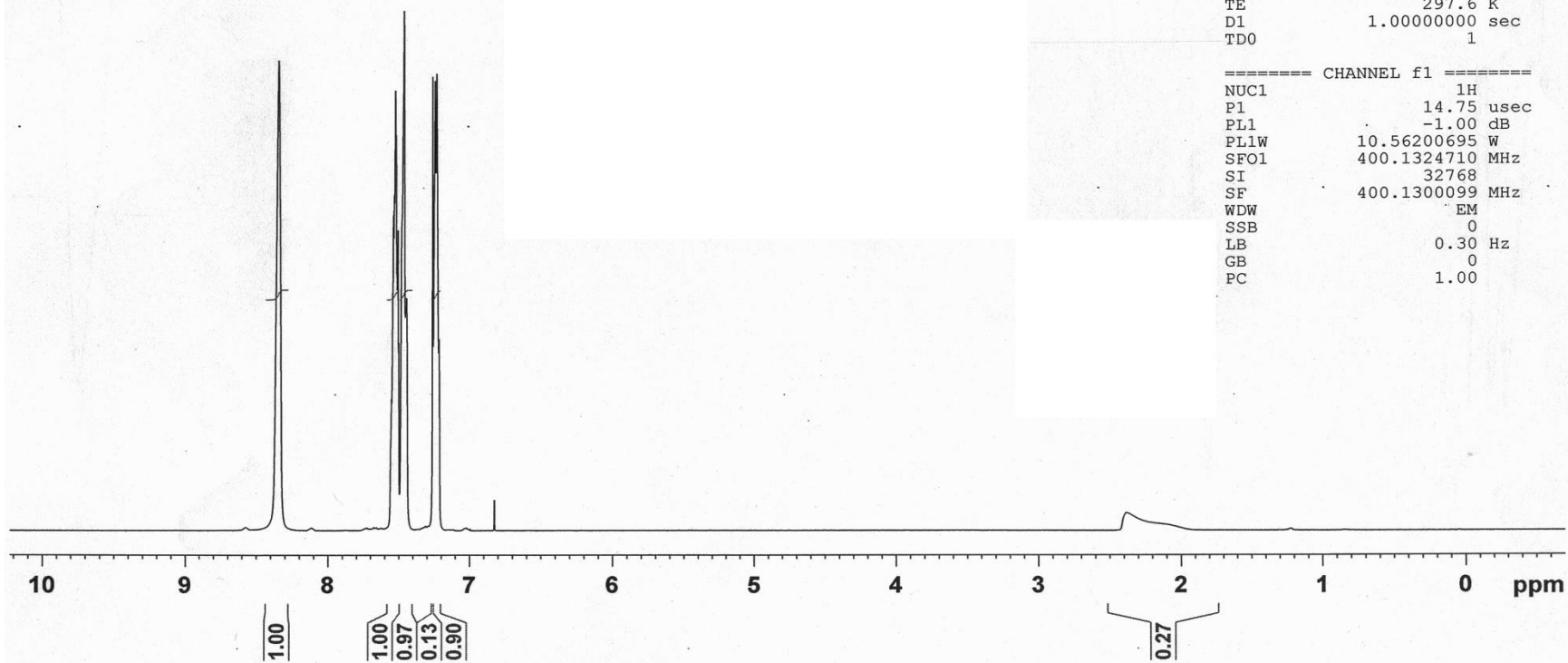


Figure S11. ¹H NMR Spectrum of 2f

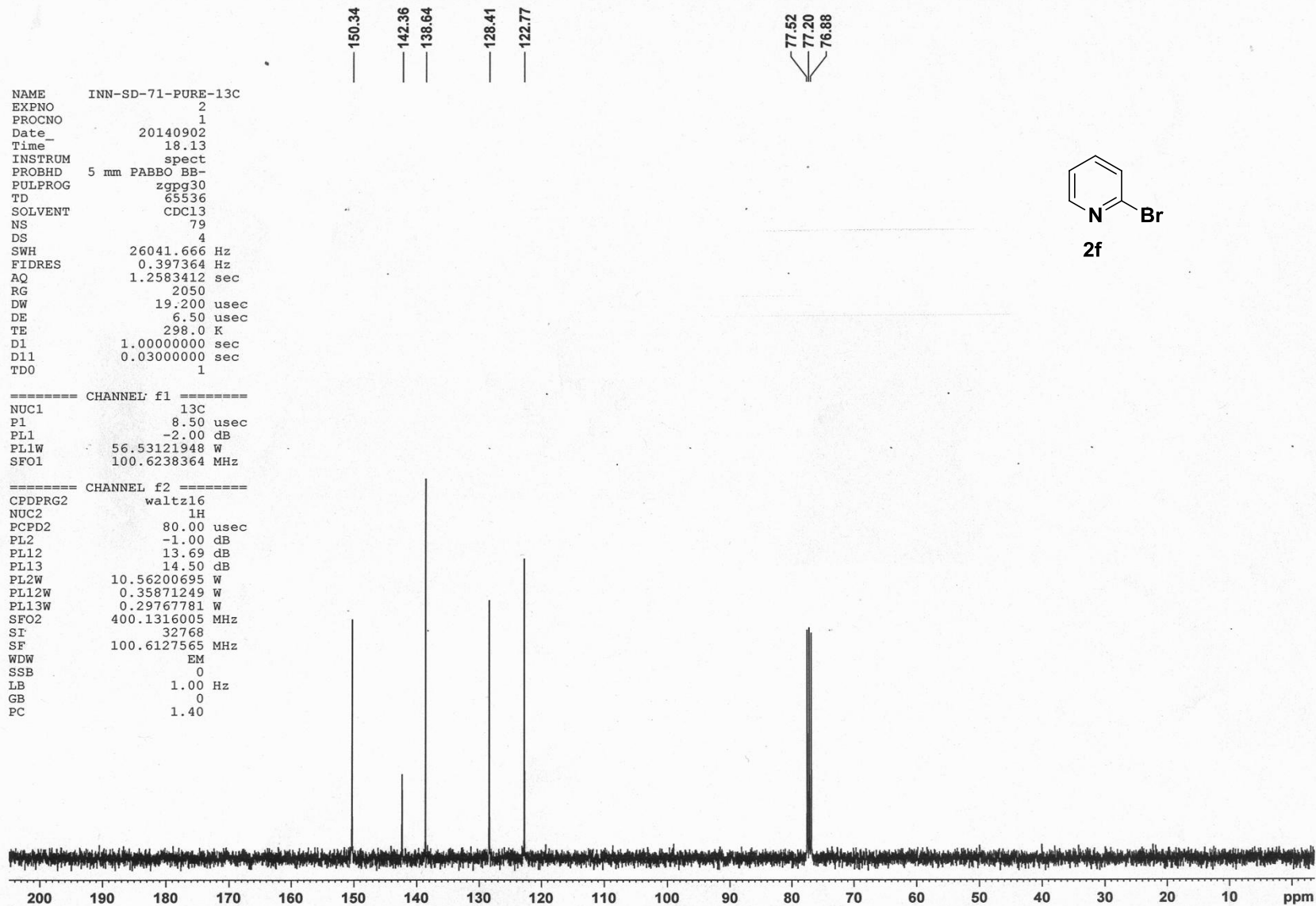


Figure S12. ¹³C NMR Spectrum of 2f

NAME INN-SD-76-PURE-1H
EXPNO 3
PROCNO 1
Date_ 20140902
Time_ 18.19
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 54274
SOLVENT DMSO
NS 16
DS 0
SWH 8223.685 Hz
FIDRES 0.151522 Hz
AQ 3.2999091 sec
RG 32
DW 60.800 usec
DE 6.50 usec
TE 297.5 K
D1 1.00000000 sec
TDO 1

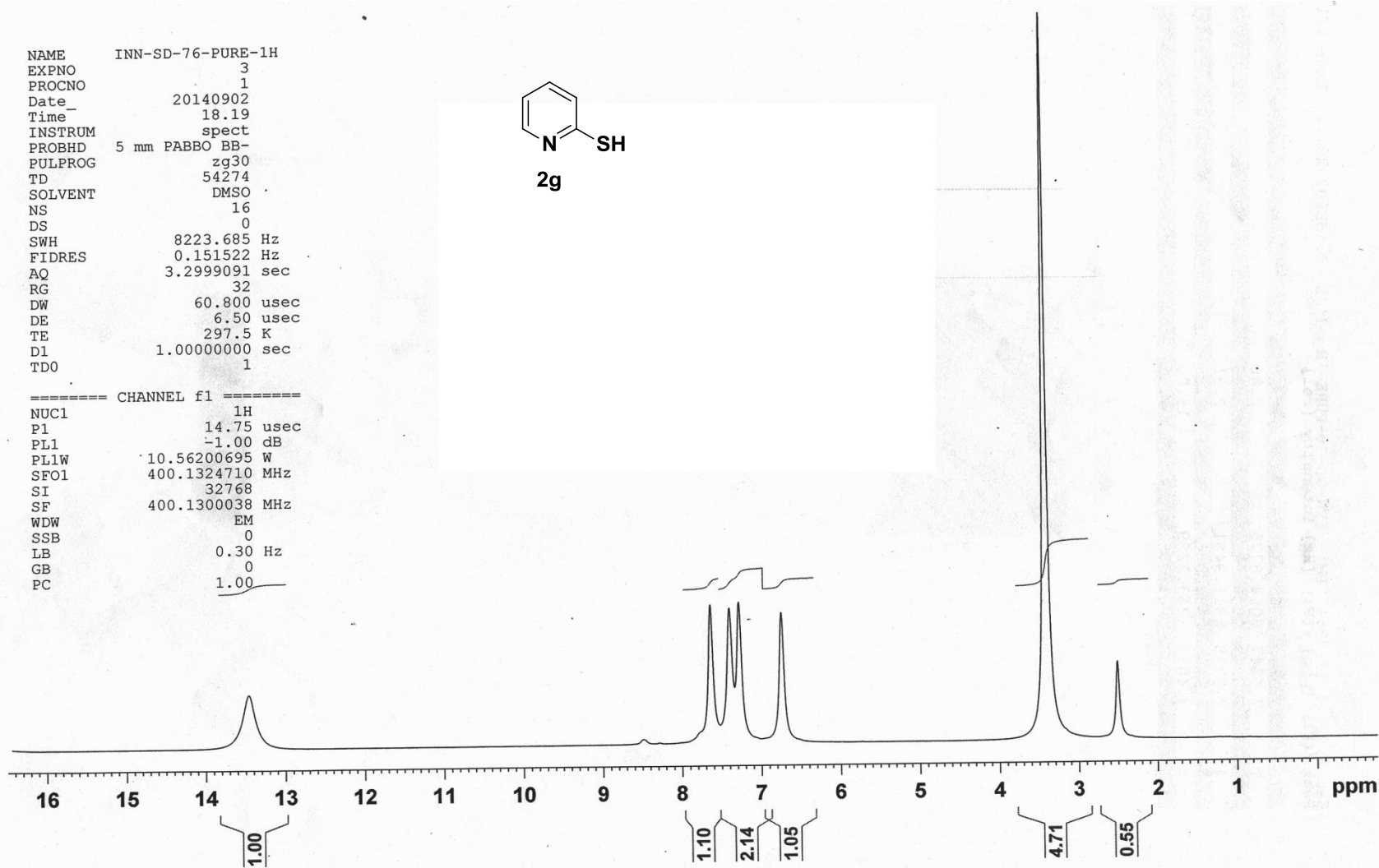
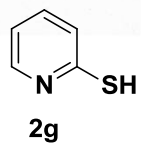


Figure S13. ¹H NMR Spectrum of 2g

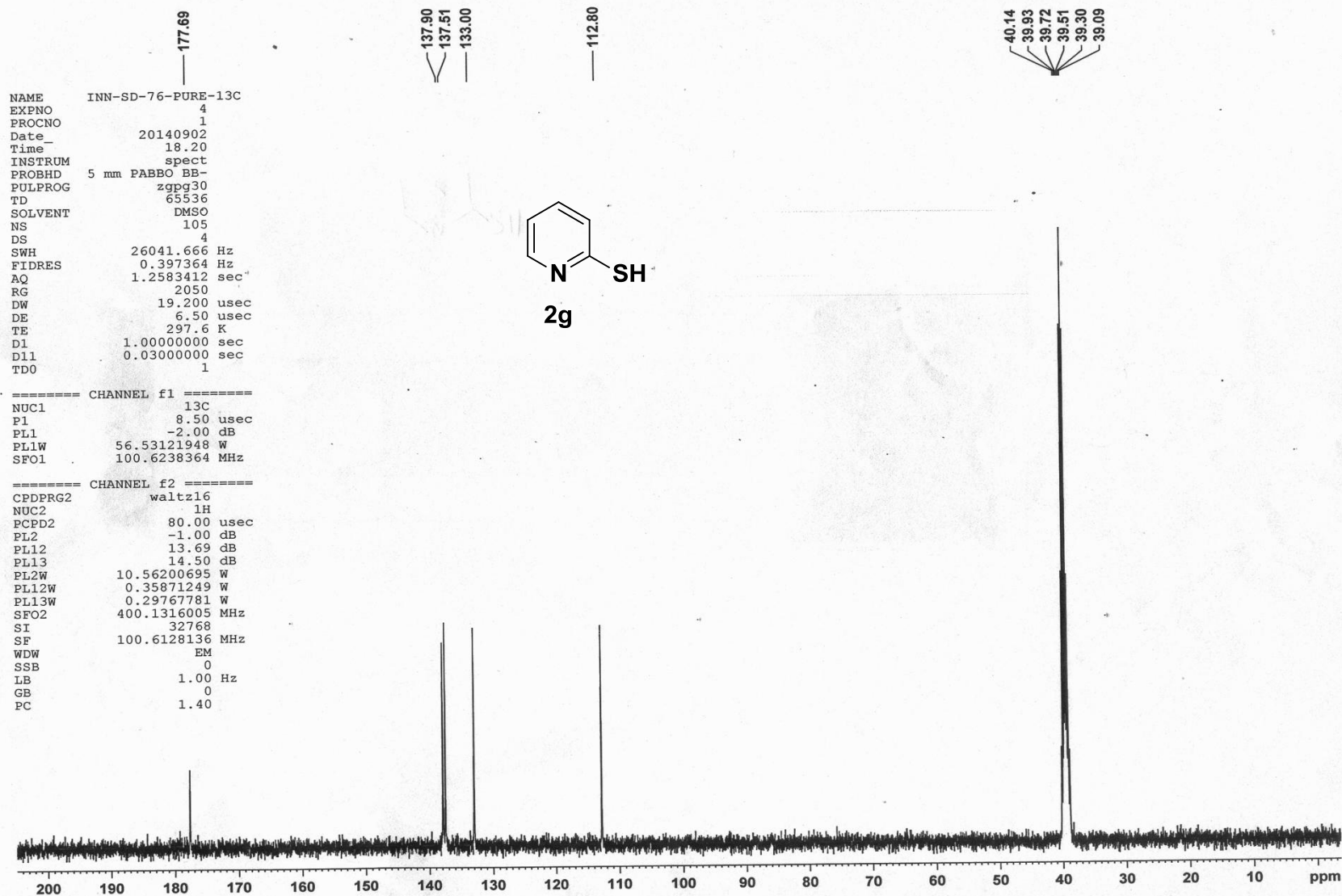


Figure S14. ¹³C NMR Spectrum of 2g

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Frequency (MHz)	400.13	Nucleus	¹ H	Number of Transients	16	Origin	spect	Original Points Count	32768
Owner	BRUKER	Points Count	32768	Pulse Sequence	zq45	Receiver Gain	228.10	SW(cyclical) (Hz)	5995.20
Solvent	DMSO-d6	Spectrum Offset (Hz)	2400.0000	Spectrum Type	STANDARD	Sweep Width (Hz)	5995.02	Temperature (degree C)	27.000

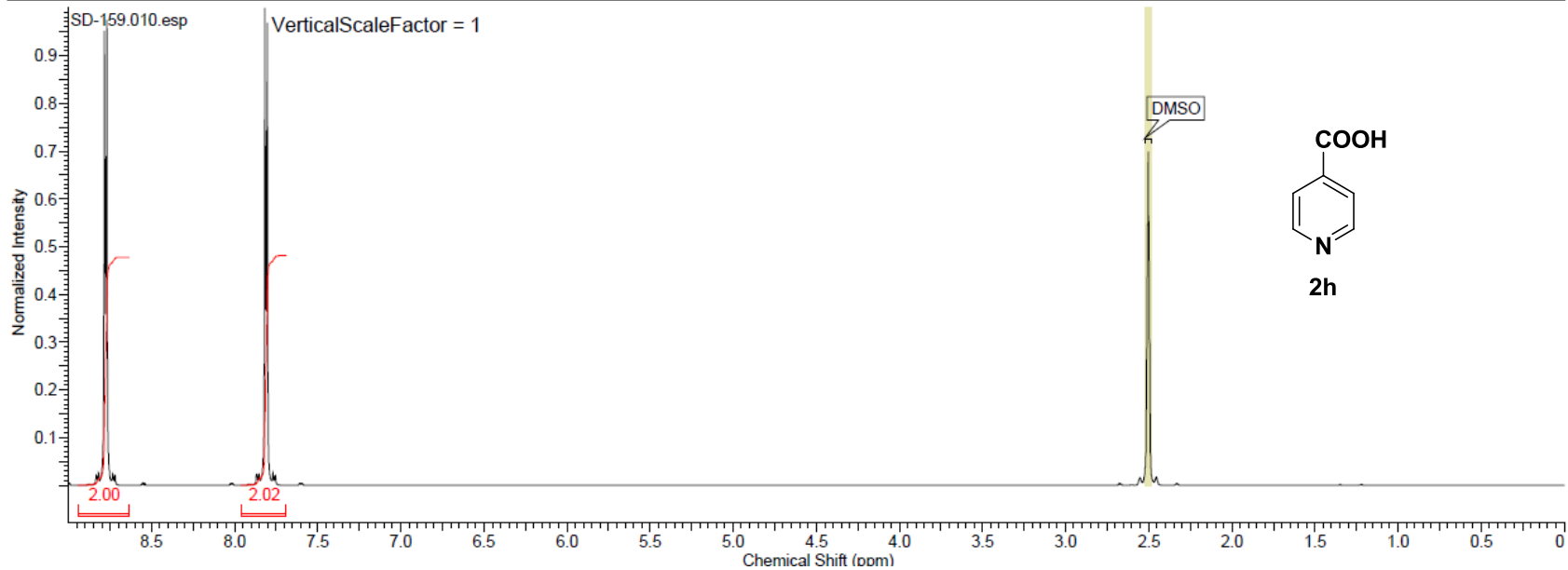


Figure S15. ¹H NMR Spectrum of 2h

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Frequency (MHz)	100.62	Nucleus	¹³ C	Number of Transients	750
Original Points Count	16384	Owner	BRUKER	Points Count	16384
Receiver Gain	2896.30	SW(cyclical) (Hz)	25125.63	Solvent	DMSO-d6
Spectrum Type	STANDARD	Sweep Width (Hz)	25124.09	Spectrum Offset (Hz)	11570.4688
		Temperature (degree C)	27.000		

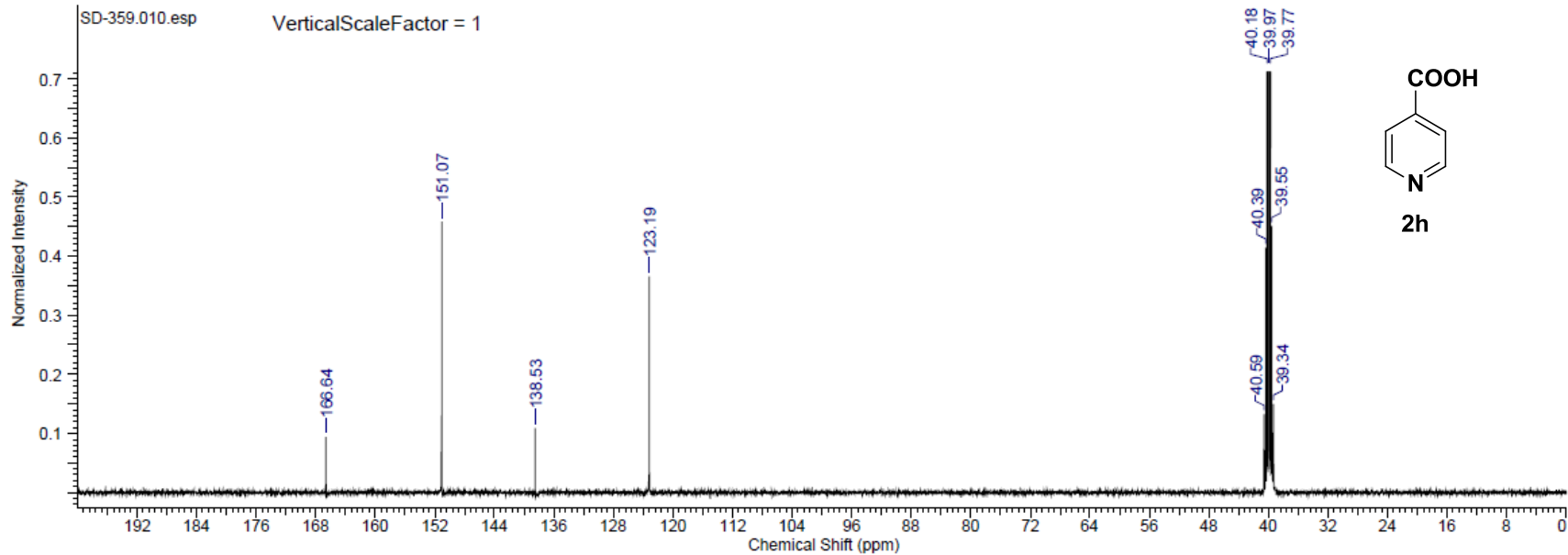


Figure S16. ¹³C NMR Spectrum of 2h

Acquisition Time (sec)	5.4657	Comment	SD-360a	Date	09 Oct 2014 11:23:12		
Date Stamp	09 Oct 2014 11:23:12			File Name	\\labies\Bitec-s\partages\scbm\SMCB_RMNI\Tritium\data\tritium\nmr\SD-360a\10\fid		
Frequency (MHz)	400.13	Nucleus	1H	Number of Transients	16	Origin	spect
Owner	BRUKER	Points Count	32768	Pulse Sequence	zg45	Receiver Gain	228.10
Solvent	DMSO-d6	Spectrum Offset (Hz)	2400.0000	Spectrum Type	STANDARD	Sweep Width (Hz)	5995.02
							Temperature (degree C) 23.900

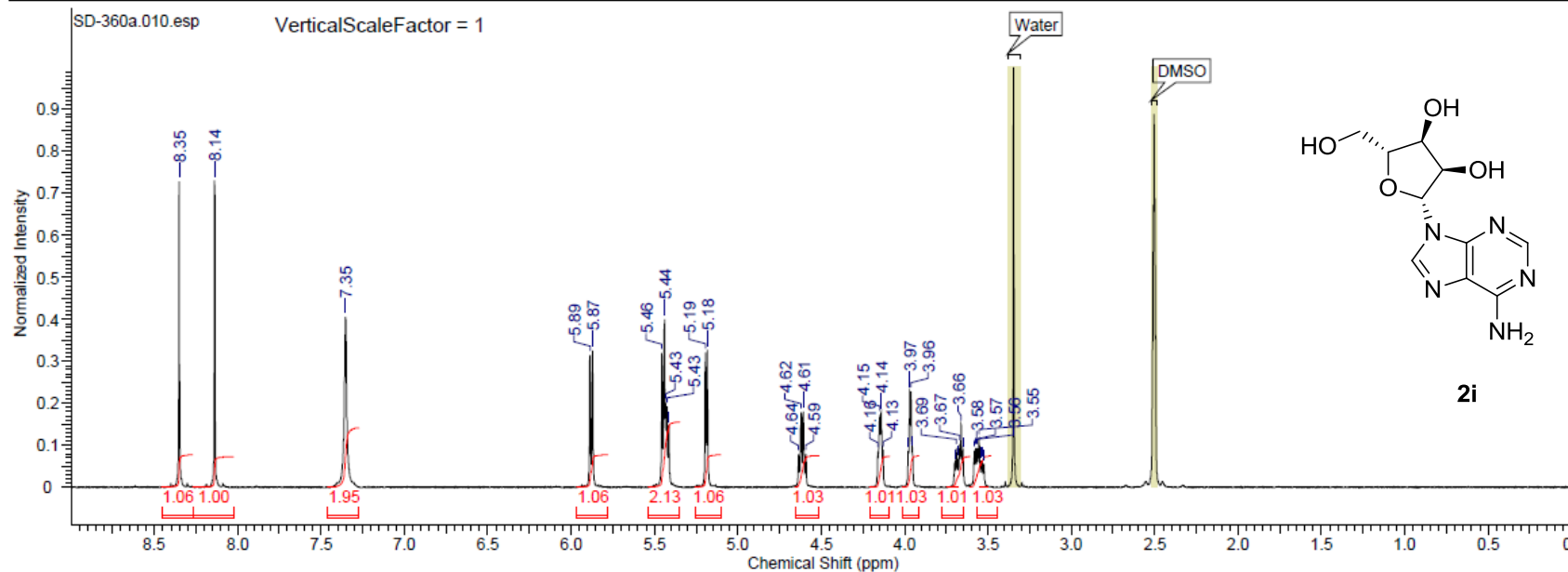


Figure S17. ^1H NMR Spectrum of 2i

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Date Stamp	09 Oct 2014 21:58:56	File Name	\\labies\Bitec-s\partages\scbm\SMMCB_RMNI\Tritium\data\tridium\nmr\SD-360a\11\fid		
Frequency (MHz)	100.62	Nucleus	13C	Number of Transients	700
Original Points Count	16384	Owner	BRUKER	Points Count	16384
Receiver Gain	2896.30	SW(cyclical) (Hz)	25125.63	Solvent	DMSO-d6
Spectrum Type	STANDARD	Sweep Width (Hz)	25124.09	Temperature (degree C)	25.200
				Pulse Sequence	zqpg30
				Spectrum Offset (Hz)	11570.4688

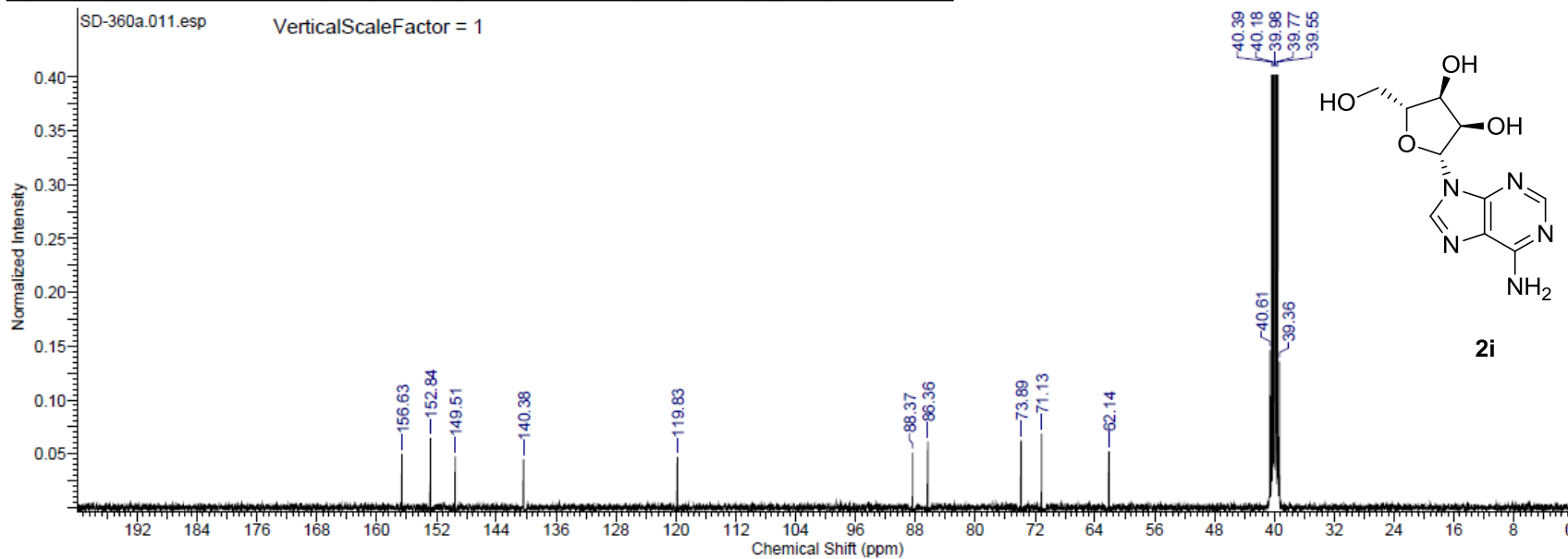
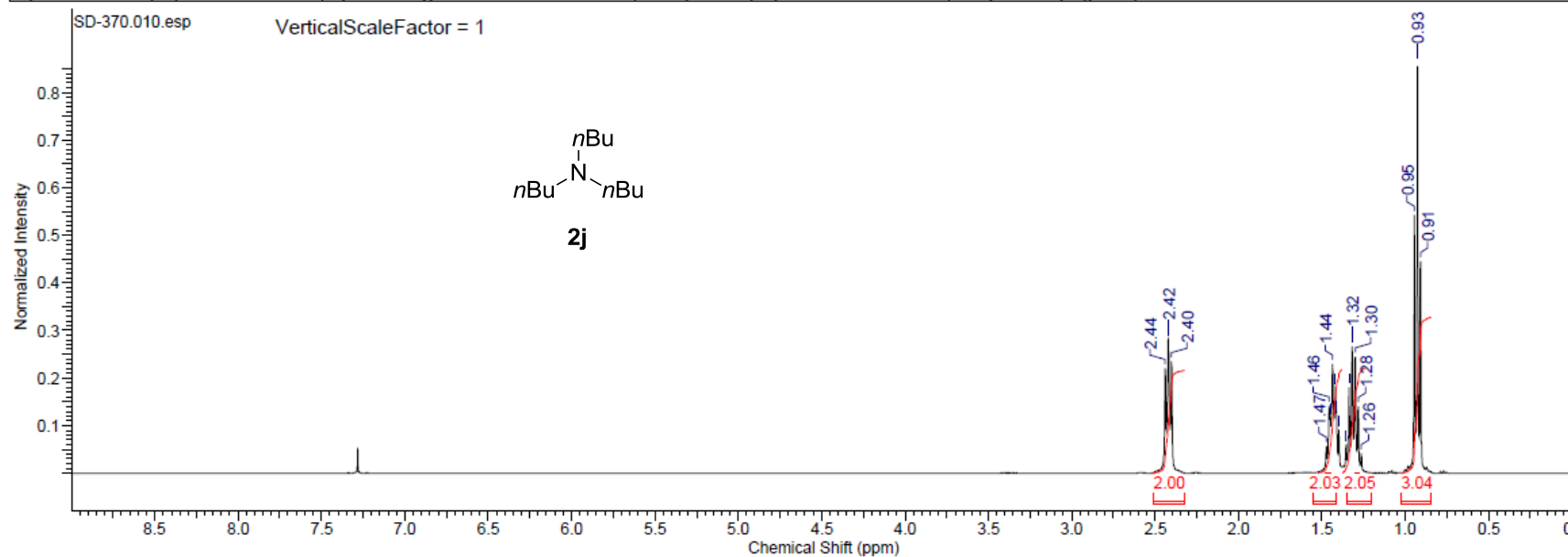
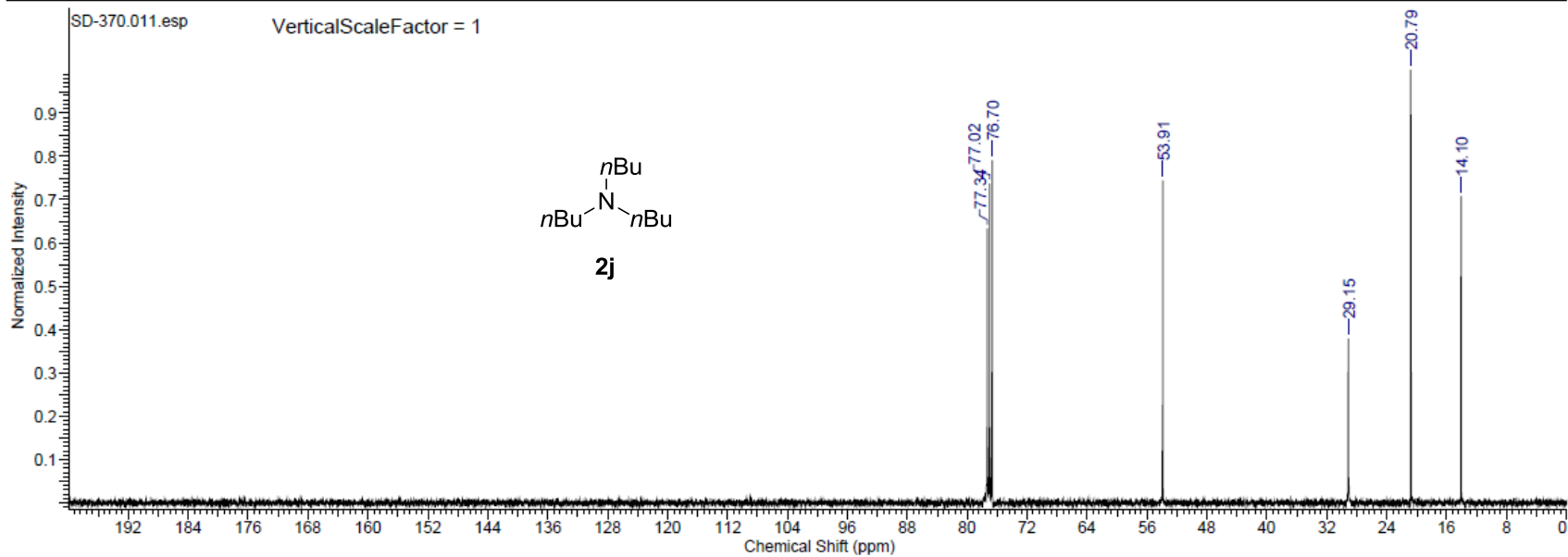


Figure S18. ¹³C NMR Spectrum of 2i

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Date Stamp	15 Oct 2014 14:41:36	File Name	\\abies\Bitec-s\partages\scbm\SMMCB_RMNI\tritium\data\tritium\nmr\SD-370\10\fid				
Frequency (MHz)	400.13	Nucleus	1H	Number of Transients	16	Origin	spect
Original Points Count	32768	Owner	BRUKER	Points Count	32768	Pulse Sequence	zg45
Receiver Gain	101.60	SW(cyclical) (Hz)	5995.20	Solvent	CHLOROFORM-d		
Spectrum Offset (Hz)	2400.0000	Spectrum Type	STANDARD	Sweep Width (Hz)	5995.02	Temperature (degree C)	25.300

Figure S19. ¹H NMR Spectrum of **2j**

Acquisition Time (sec)	0.6521	Comment	SD-370	Date	15 Oct 2014 15:02:56		
Date Stamp	15 Oct 2014 15:02:56	File Name	\\abies\Bitec-s\partages\scbm\SMACB_RMNI\tritium\data\tritium\nmr\SD-370\11\fid				
Frequency (MHz)	100.62	Nucleus	13C	Number of Transients	400	Origin	spect
Original Points Count	16384	Owner	BRUKER	Points Count	16384	Pulse Sequence	zpgq30
Receiver Gain	2896.30	SW(cyclical) (Hz)	25125.63	Solvent	CHLOROFORM-d		
Spectrum Offset (Hz)	11570.4688	Spectrum Type	STANDARD	Sweep Width (Hz)	25124.09	Temperature (degree C)	27.000

Figure S20. ¹³C NMR Spectrum of 2j

Acquisition Time (sec)	5.4657	Comment	5 mm PABBO BB-1H Z-GRD Z824801/0109	Date	23 Mar 2015 08:49:20
Date Stamp	23 Mar 2015 08:49:20	File Name	\\abies\iBiTec-S\PARTAGES\SCBMSMMCB_RMNI\Tritium\data\tridium\nmr\sd-nmM\10\fid		
Frequency (MHz)	400.13	Nucleus	1H	Number of Transients	8
Original Points Count	32768	Owner	BRUKER	Points Count	32768
Receiver Gain	14.30	SW(cyclical) (Hz)	5995.20	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	2400.0000	Spectrum Type	STANDARD	Sweep Width (Hz)	5995.02
				Temperature (degree C)	22.600

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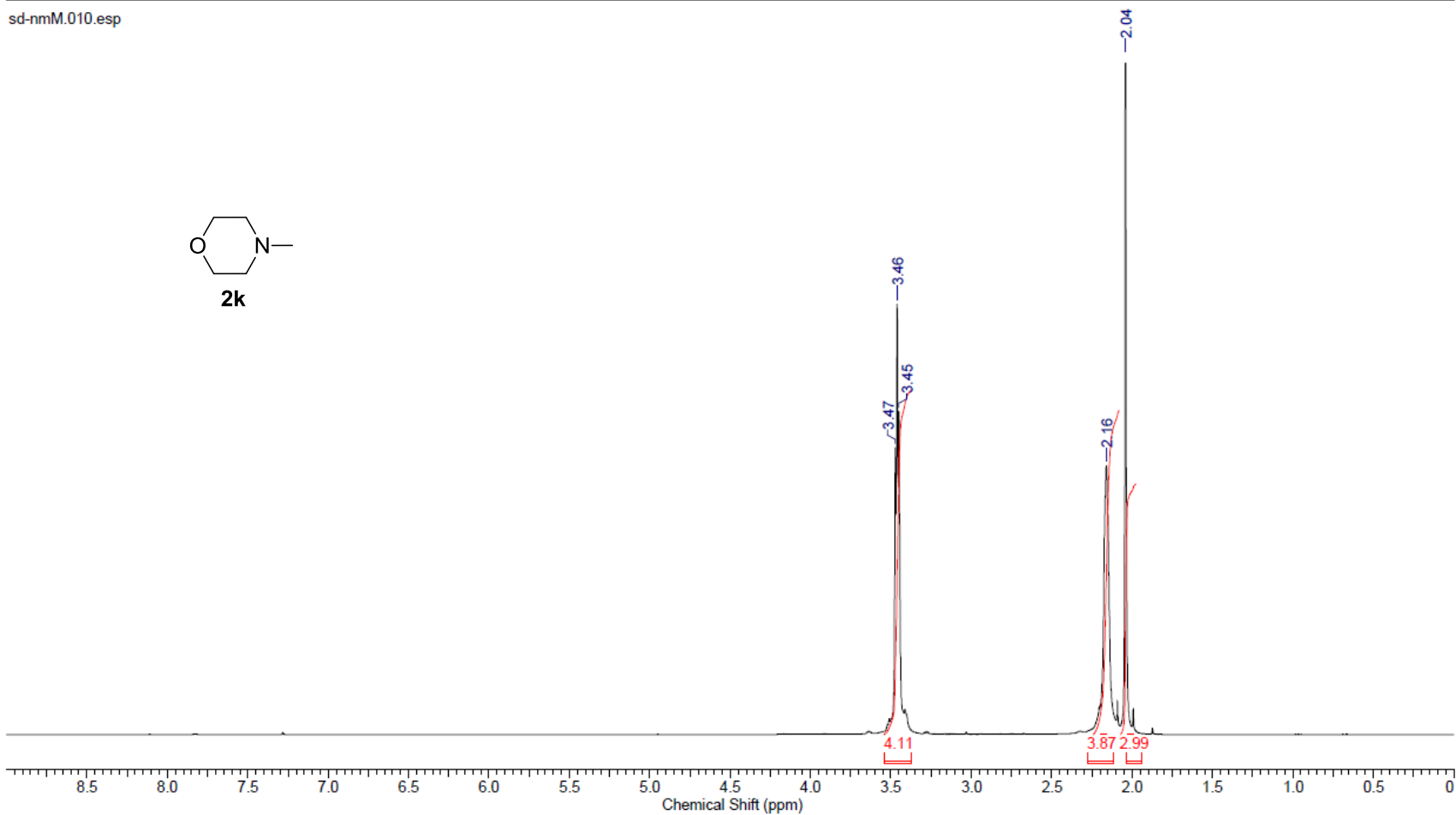
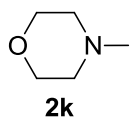


Figure S21. ¹H NMR Spectrum of 2k

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Date Stamp	23 Mar 2015 09:00:00	File Name	\\labies\iBiTec-S\PARTAGES\SCBM\SMMCB_RMNI\Tritium\data\tridium\nmr\sd-nmM11\fid		
Frequency (MHz)	100.62	Nucleus	13C	Number of Transients	126
Original Points Count	16384	Owner	BRUKER	Points Count	16384
Receiver Gain	2896.30	SW(cyclical) (Hz)	25125.63	Solvent	CHLOROFORM-d
Spectrum Type	STANDARD	Sweep Width (Hz)	25124.09	Temperature (degree C)	27.000
				Pulse Sequence	zgpg30
				Spectrum Offset (Hz)	11570.4688

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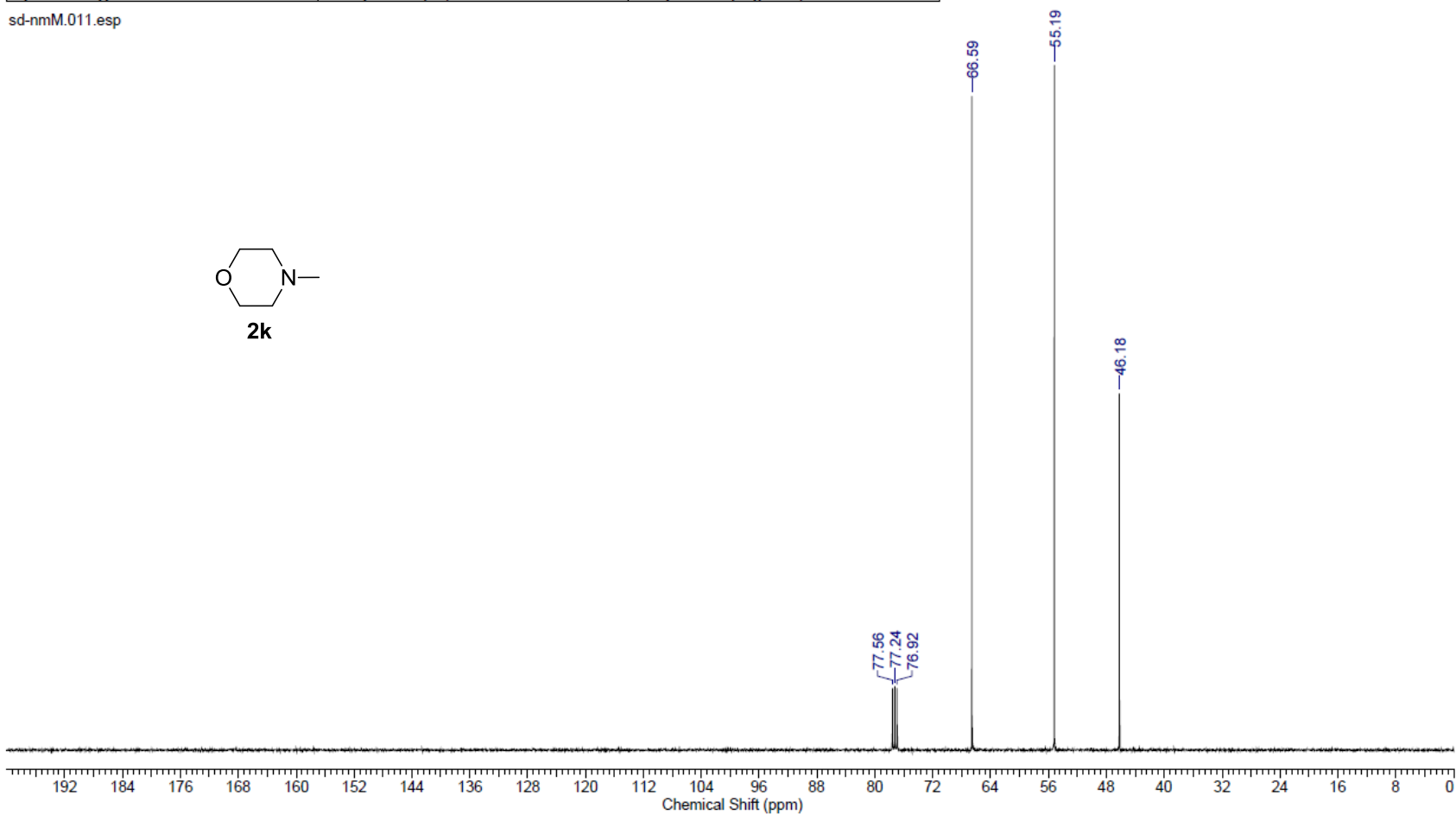
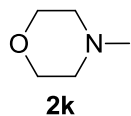


Figure S22. ¹³C NMR Spectrum of 2k

This report was created by ACD/NMR Processor Academic Edition. For more information go to www.acdlabs.com/nmrproc/

Acquisition Time (sec)	5.4657	Comment	SD-505	Date	26 Mar 2015 08:36:32		
Date Stamp	26 Mar 2015 08:36:32	Nucleus	1H	File Name	\\labes\Bitec-s\partages\scbm\SMCB RMN\TRITIUM\DATA\TRITIUM\NMR\SD-505PUR\2\FID		
Frequency (MHz)	400.13	Owner	BRUKER	Number of Transients	8	Origin	spect
Original Points Count	32768	SW(cyclical) (Hz)	5995.20	Points Count	32768	Pulse Sequence	zq45
Receiver Gain	40.30	Spectrum Type	STANDARD	Solvent	CHLOROFORM-d		
Spectrum Offset (Hz)	2400.0000			Sweep Width (Hz)	5995.02	Temperature (degree C)	27.000

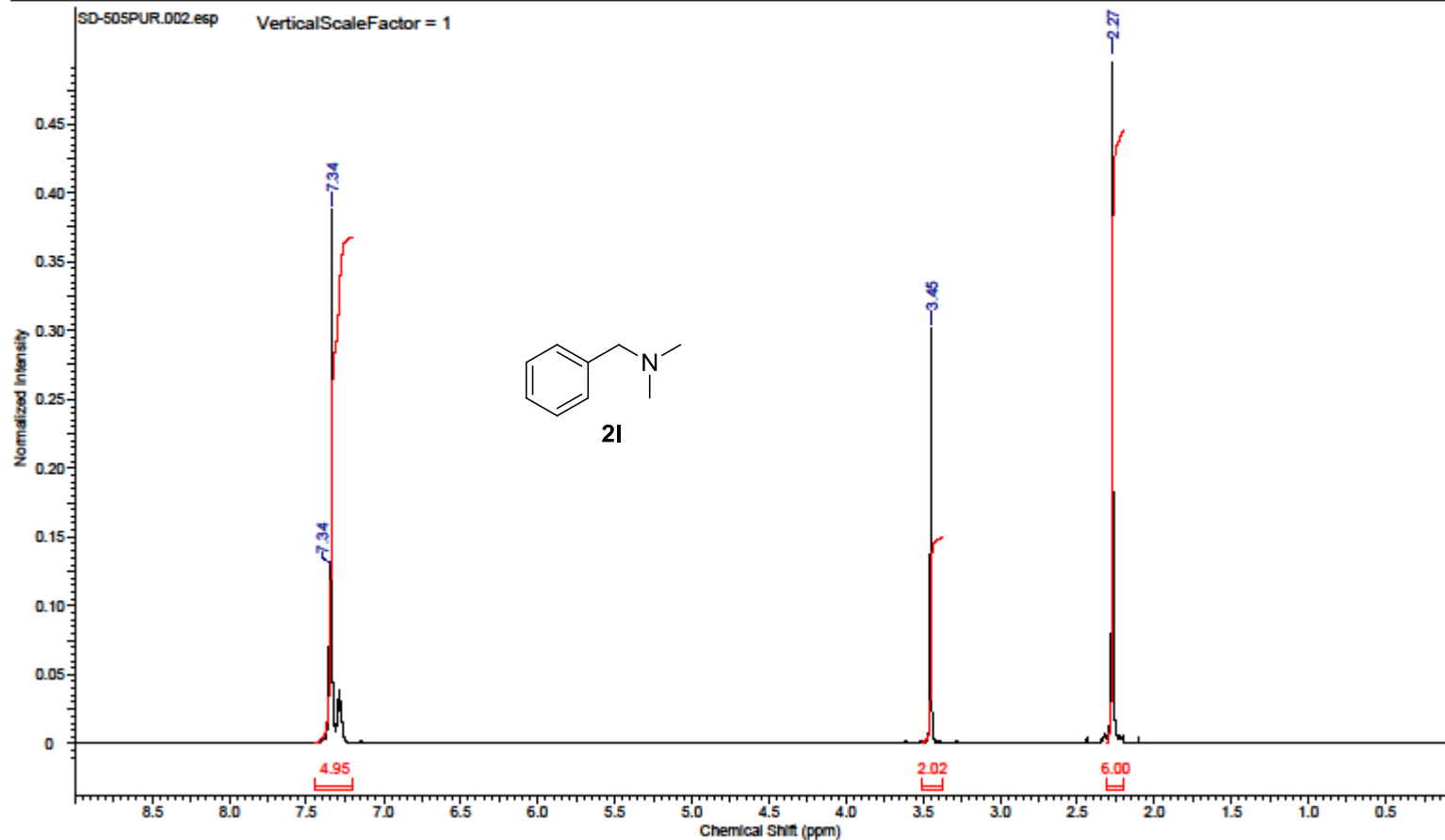


Figure S23. ^1H NMR Spectrum of 21

This report was created by ACD/NMR Processor Academic Edition. For more information go to www.acdlabs.com/nmrproc/

Acquisition Time (sec)	0.6521	Comment	SD-505	Date	26 Mar 2015 08:47:12
Date Stamp	26 Mar 2015 08:47:12	Nucleus	13C	File Name	\\labes\Bitec-s\partages\scbm\SMCB_RMNI\TRITIUM\DATA\TRITIUM\NMR\SD-505PUR\3\FID
Frequency (MHz)	100.62	Owner	BRUKER	Number of Transients	126
Original Points Count	16384	SW(cyclical) (Hz)	25125.63	Points Count	16384
Receiver Gain	2896.30	Spectrum Type	STANDARD	Solvent	CHLOROFORM-d
Spectrum Offset (Hz)	11561.2373			Pulse Sequence	zpgq30
				Sweep Width (Hz)	25124.09
				Temperature (degree C)	23.000

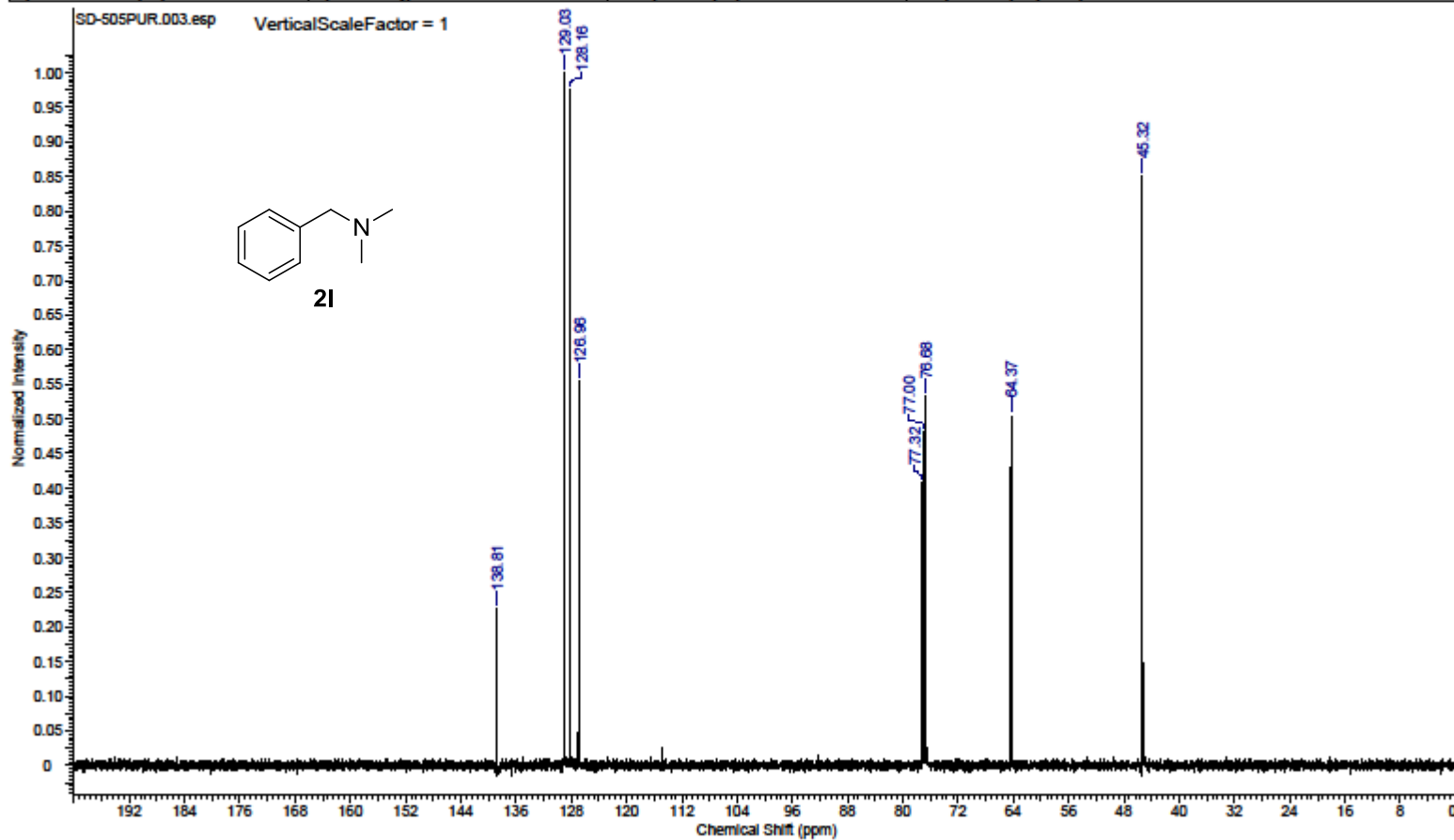


Figure S24. ^{13}C NMR Spectrum of 21

This report was created by ACD/NMR Processor Academic Edition. For more information go to www.acdlabs.com/nmrproc/

Acquisition Time (sec)	0.2785	Comment	5 mm PABBO BB-1H Z-GRD Z824801/0109	Date	13 Apr 2015 11:40:16
Date Stamp	13 Apr 2015 11:40:16				
File Name	\\labies\Bitec-s\partages\scbm\SMACB_RM\NTRITUM\DATA\TRITUM\NMR\SD512-DEUTERIUM\11\FID			Frequency (MHz)	61.42
Nucleus	2H	Number of Transients	3230	Origin	spect
Owner	root	Points Count	512	Pulse Sequence	zgpg
SW(cyclical) (Hz)	1838.24	Solvent	CHCl ₃	Receiver Gain	12.70
Spectrum Type	STANDARD	Sweep Width (Hz)	1834.65	Temperature (degree C)	23.800

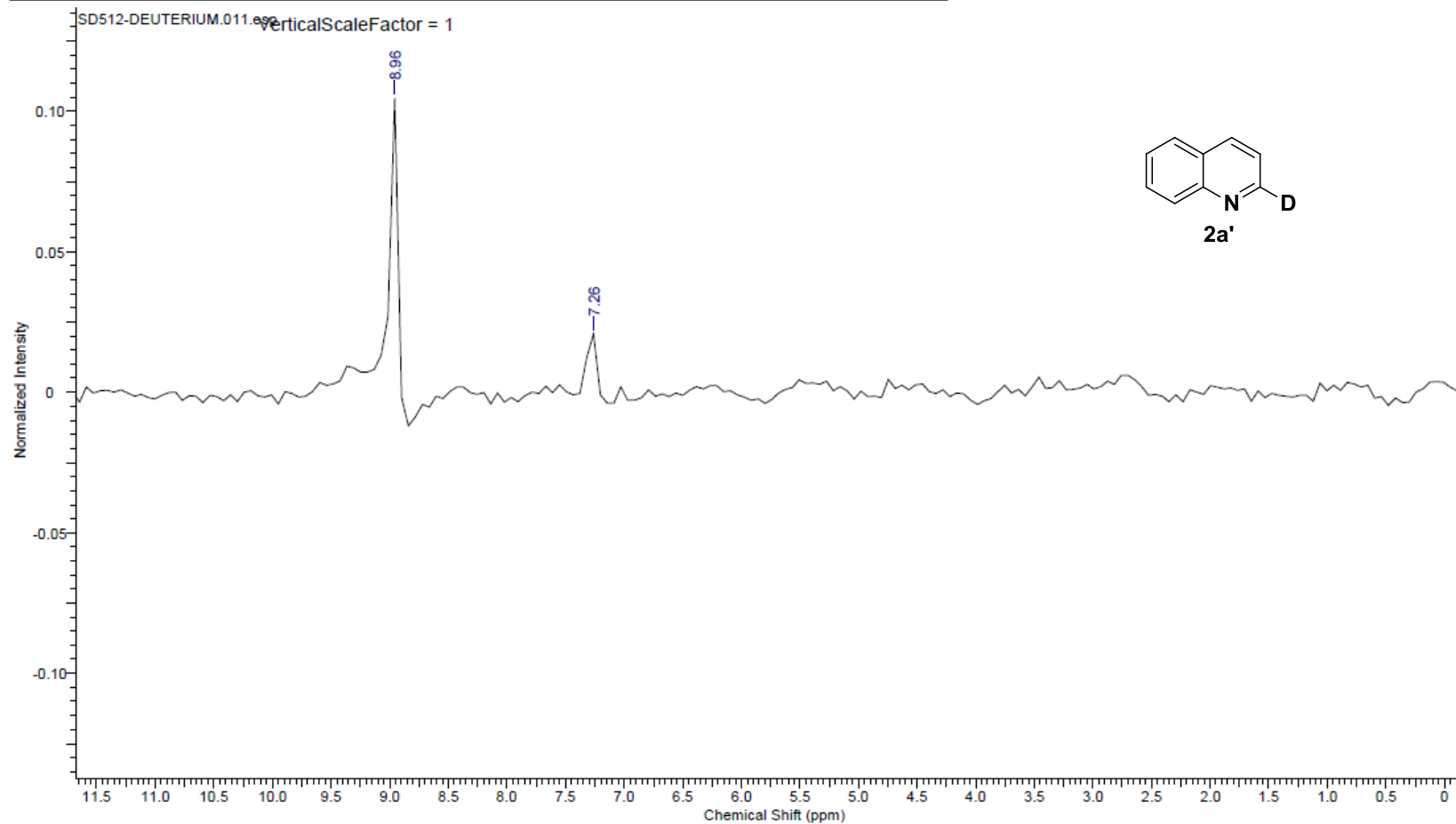
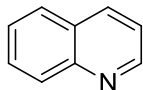


Figure S25. ²H NMR Spectrum of 2a'

A typical procedure for the deoxygenation of amine *N*-oxides is given for quinoline *N*-oxide:

The AuCNT catalyst (0.4 mol%) aqueous suspension was centrifuged and washed three times with dry THF prior to use. Under N₂, to a solution of quinoline *N*-oxide (0.1 mmol) in dry THF (1 mL) was added the AuCNT catalyst and dimethylphenylsilane (0.11 mmol). The resulting mixture was stirred at room temperature and the progress of the reaction was monitored by TLC. After completion, water (2 mL) was added and the aqueous phase was extracted with dichloromethane (3 × 5 mL). The combined organic layers were dried over anhydrous Na₂SO₄, filtered, and concentrated under vacuum. The crude material was then purified by silica gel column chromatography (cyclohexane/EtOAc, 95:5) to afford quinoline as colorless oil (84% yield).

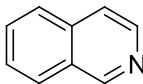
Quinoline (2a)



¹H NMR (500 MHz, CDCl₃) δ 8.95-8.90 (m, 1H), 8.15 (d, *J* = 8.3 Hz, 1H), 8.11 (d, *J* = 8.1 Hz, 1H), 7.81 (d, *J* = 7.9 Hz, 1H), 7.71 (td, *J* = 8.1, 1.3 Hz, 1H), 7.54 (td, *J* = 7.9, 1.3 Hz, 1H), 7.38 (dd, *J* = 8.3 Hz, *J* = 4.2 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 150.5, 148.4, 136.3, 129.6, 129.6, 128.5, 128.0, 126.7, 121.3.

Isoquinoline (2b)



¹H NMR (500 MHz, CDCl₃) δ 9.25 (s, 1H), 8.51 (d, *J* = 5.8 Hz, 1H), 7.95 (d, *J* = 7.8 Hz, 1H), 7.81 (d, *J* = 7.9 Hz, 1H), 7.68 (t, *J* = 7.9 Hz, 1H), 7.64 (d, *J* = 5.8 Hz, 1H), 7.59 (t, *J* = 7.8 Hz, 1H).

¹³C NMR (125 MHz, CDCl₃) δ 152.6, 143.0, 135.9, 130.5, 128.8, 127.8, 127.4, 126.6, 120.7.

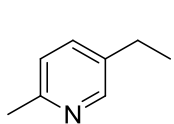
Pyridine (2c)



¹H NMR (400 MHz, CDCl₃) δ 8.56 (s, 2H), 7.65-7.62 (m, 1H), 7.26-7.23 (m, 2H).

¹³C NMR (100 MHz, CDCl₃) δ 149.6 (2C), 135.9, 123.7 (2C).

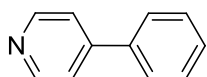
5-Ethyl-2-methyl pyridine (2d)



$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.32 (d, $J = 2.1$ Hz, 1H), 7.38 (dd, $J = 7.9$ Hz, $J = 2.1$ Hz, 1H), 7.05 (d, $J = 7.9$ Hz, 1H), 2.58 (q, $J = 7.6$ Hz, 2H), 2.50 (s, 3H), 1.21 (t, $J = 7.6$ Hz, 3H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 155.6, 148.7, 136.3, 136.0, 123.0, 25.8, 24.0, 15.6.

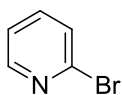
4-Phenylpyridine (2e)



$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.67 (s, 2H), 7.64 (d, $J = 8.0$ Hz, 2H), 7.53-7.45 (m, 5H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 150.0 (2C), 148.5, 138.0, 129.1 (3C), 127.0 (2C), 121.6 (2C).

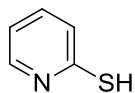
2-Bromopyridine (2f)



$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.30-8.40 (m, 1H), 7.55-7.50 (m, 1H), 7.46-7.44 (m, 1H), 7.26-7.21 (m, 1H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 150.3, 142.4, 138.6, 128.4, 122.8.

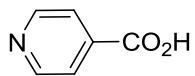
Mercaptopyridine (2g)



$^1\text{H NMR}$ (400 MHz, $\text{DMSO-}d_6$) δ 13.46 (br s, 1H), 7.60-7.65 (m, 1H), 7.38-7.42 (m, 1H), 7.25-7.30 (m, 1H), 6.72-6.76 (m, 1H).

$^{13}\text{C NMR}$ (100 MHz, $\text{DMSO-}d_6$) δ 177.7, 137.9, 137.5, 133.0, 112.8.

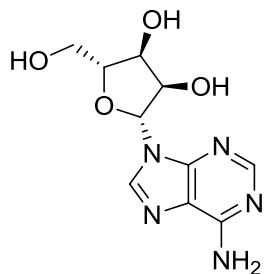
Isonicotinic acid (2h)



$^1\text{H NMR}$ (400 MHz, $\text{DMSO-}d_6$) δ 8.75 (d, $J = 6.0$ Hz, 2H), 7.79 (d, $J = 6.0$ Hz, 2H).

$^{13}\text{C NMR}$ (100 MHz, $\text{DMSO-}d_6$) δ 166.6, 151.0, 138.5, 123.1.

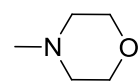
Adenosine (2i)



^1H NMR (400 MHz, DMSO- d_6) δ 8.33 (s, 1H), 8.12 (s, 1H), 7.33 (s, 2H), 5.86 (d, $J = 6$ Hz, 1H), 5.44-5.39 (m, 2H), 5.17 (d, $J = 4.4$ Hz, 1H), 4.59 (q, $J = 5.9$ Hz, 1H), 4.12 (q, 4.1 Hz, 1H), 3.94 (q, $J = 3.2$ Hz, 1H), 3.67 (m, 1H), 3.53 (m, 1H).

^{13}C NMR (100 MHz, DMSO- d_6) δ 156.6, 152.8, 149.5, 140.3, 119.8, 88.3, 86.3, 73.8, 71.1, 62.1.

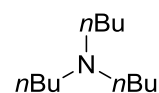
N-Methyl morpholine (2j)



^1H NMR (400 MHz, CDCl_3) δ 3.58 (s, 4H), 2.28 (s, 4H), 2.16 (s, 3H).

^{13}C NMR (100 MHz, CDCl_3) δ 66.9 (2C), 55.4 (2C), 46.4.

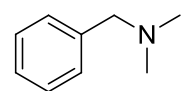
Tri-*n*-butylamine (2k)



^1H NMR (400 MHz, CDCl_3) δ 2.40 (t, $J = 7.6$ Hz, 6H), 1.45-1.38 (m, 6H), 1.28 (m, 6H), 0.91 (t, $J = 7.4$ Hz, 9H).

^{13}C NMR (100 MHz, CDCl_3) δ 53.8 (3C), 29.1 (3C), 20.7 (3C), 14.0 (3C).

N,N-dimethyl-1-phenylmethanamine (2l)



^1H NMR (400 MHz, CDCl_3) δ 7.35-7.24 (m, 5H), 3.43 (s, 2H), 2.52 (s, 6H).

^{13}C NMR (100 MHz, CDCl_3) δ 138.8, 129.1 (2C), 128.2 (2C), 127.0, 64.4, 45.3 (2C).