

# Construction of All-Substituted Pyrrolidine Derivative with Multiple Stereogenic Centers and Betti-Base-derived $\gamma$ -Amino Alcohols by [1,2]-Wittig Rearrangement

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## Supporting Information

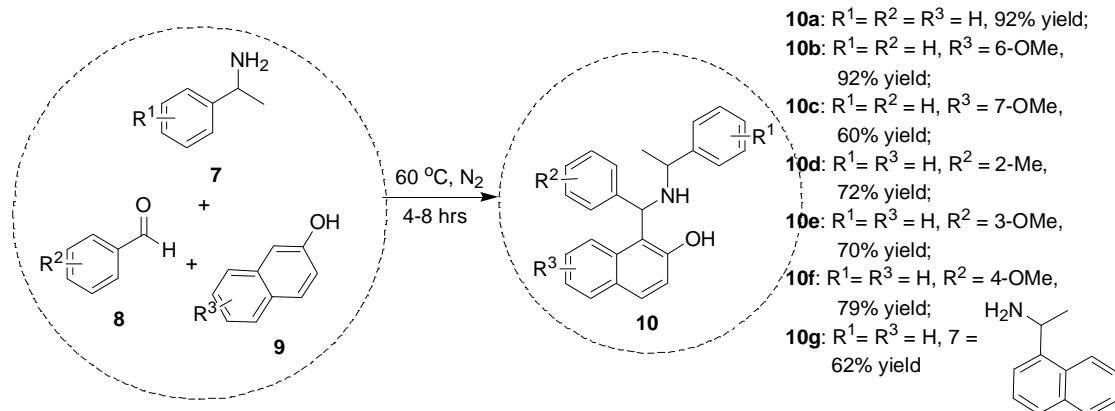
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## 1. General Information

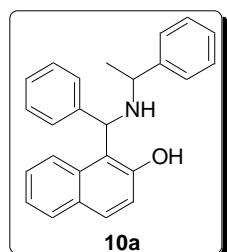
DCM was dried by  $\text{CaH}_2$ , distilled under Atmospheric pressure and stored under nitrogen. THF and  $\text{Et}_2\text{O}$  were dried by sodium benzophenone ketyl, distilled under Atmospheric pressure and stored under nitrogen. Reagents were purchased from commercial sources and were used as received unless mentioned otherwise. Reactions were monitored by thin layer chromatography using silica gel. All the reactions dealing with air or moisture sensitive compounds were carried out in a dry reaction vessel under positive pressure of argon. Air- and moisture-sensitive liquids and solutions were transferred via a syringe or a stainless steel cannula. NMR spectra were recorded on a Bruke Avance operating for  $^1\text{H}$  NMR at 400 MHz,  $^{13}\text{C}$  NMR at 100 MHz, and  $^{19}\text{F}$  NMR at 470 MHz, using TMS as internal standard. The peaks were internally referenced to TMS (0.00 ppm) or residual undeuterated solvent signal (77.16 ppm for  $^{13}\text{C}$  NMR). The following abbreviations (or combinations thereof) were used to explain multiplicities: s = singlet, d = doublet, t = triplet, m = multiplet, b = broad. Mass spectroscopy data of the products were collected on an HRMS-TOF instrument or a low-resolution MS instrument using EI ionization.

## 2. Synthesis of various Betti base **10** under the classic reaction conditions.



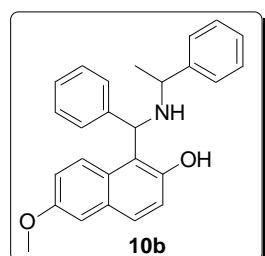
**General procedure for the synthesis of aminoalkynaphthol **10** (Betti base).** In the following typical procedure, a mixture of 2-naphthol (0.72 g, 5.0 mmol),

benzaldehyde (0.64g, 6.00 mmol), and 1-phenylethylamine (0.64 g, 5.25mmol) was stirred at 60 °C for 8 h under nitrogen atmosphere. Following the progress of the reaction by TLC and <sup>1</sup>H NMR, The reaction mixture was dispersed at room temperature with EtOH (5 mL). The white crystals separated were collected and washed with EtOH. The crystalline white residue, purified by crystallization from EtOAc/hexane, gives the pure **10**.



**1-(phenyl((1-phenylethyl)amino)methyl)naphthalen-2-ol (10a):**

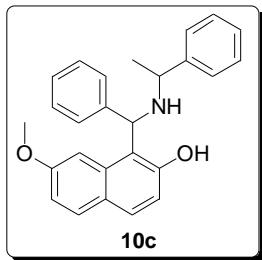
**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.76-7.72 (m, 2H), 7.38 (t, *J* = 7.1 Hz, 4H), 7.30 – 7.12 (m, 10H), 5.49 (s, 1H), 3.91 (q, *J* = 6.4 Hz, 1H), 2.31 (s, 1H), 1.50 (d, *J* = 6.8 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 157.4, 143.2, 141.6, 129.8, 129.2, 129.1, 128.9, 128.1, 128.0, 127.8, 126.8, 122.5, 121.2, 120.2, 113.2, 60.4, 56.7, 23.1. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3273, 3060, 2963, 1622, 1452, 1239, 818, 744, 671. **HRMS** (EI) calcd. for C<sub>25</sub>H<sub>24</sub>NO [M+H]<sup>+</sup>, 354.1858, found 354.1840. Isolated yield: 92 %.



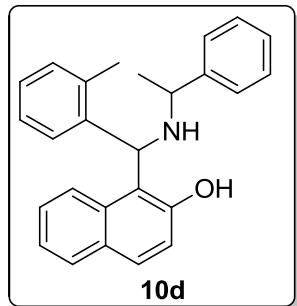
**6-methoxy-1-(phenyl((1-phenylethyl)amino)methyl)naphthalen-2-ol (10b):**

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 13.43 (s, 1H), 7.66 (d, *J* = 8.8 Hz, 1H), 7.46 – 7.34 (m, 3H), 7.34 – 7.16 (m, 10H), 7.09 (s, 1H), 6.93 (d, *J* = 9.2 Hz, 1H), 5.42 (s, 1H), 3.89 (d, *J* = 7.0 Hz, 1H), 3.86 (s, 3H), 2.28 (d, *J* = 9.8 Hz, 1H), 1.51 (d, *J* = 6.7 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 155.7, 155.2, 148.3, 141.7, 129.7, 129.2, 129.1, 128.6, 128.1, 128.0, 127.8, 126.8, 122.8, 120.6, 118.7, 113.7, 107.4, 60.5, 56.7, 55.4, 23.1. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3423, 3312, 3021, 2959, 1606, 1519, 1493, 1385,

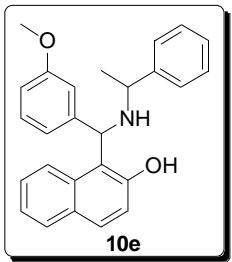
1242, 1100, 864, 700. **HRMS** (EI) calcd. for  $C_{26}H_{26}NO$   $[M+H]^+$ , 384.1964, found 384.1944. Isolated yield: 56 %.



**7-methoxy-1-(phenyl((1-phenylethyl)amino)methyl)naphthalen-2-ol (10c):**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  13.68 (s, 1H), 7.69-7.63 (m, 2H), 7.44-7.37 (m, 3H), 7.26-7.20 (m, 7H), 7.10 (d,  $J = 8.8$  Hz, 1H), 6.91 (d,  $J = 8.8$  Hz, 1H), 6.68 (s, 1H), 5.36 (s, 1H), 3.92 (q,  $J = 6.7$  Hz, 1H), 3.65 (s, 3H), 2.30 (s, 1H), 1.53 (d,  $J = 6.7$  Hz, 3H).  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  158.2, 158.0, 143.3, 141.5, 130.3, 129.5, 129.2, 129.0, 128.1, 128.0, 127.7, 127.0, 124.1, 117.6, 114.2, 112.5, 101.0, 60.5, 56.7, 55.0, 22.9. **IR** (KBr) ( $\nu_{\text{max}}$ /  $\text{cm}^{-1}$ ): 3424, 3298, 3057, 3008, 1620, 1475, 1225, 835, 699. **HRMS** (EI) calcd. for  $C_{26}H_{26}NO$   $[M+H]^+$ , 384.1964, found 384.1962. Isolated yield: 60 %.

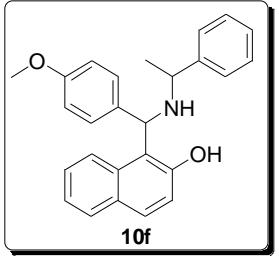


**1-(((1-phenylethyl)amino)(o-tolyl)methyl)naphthalen-2-ol (10d):**  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  13.77 (s, 1H), 7.74 (t,  $J = 8.8$  Hz, 2H), 7.41 – 6.93 (m, 14H), 5.67 (s, 1H), 3.88 (s, 1H), 1.92 (s, 3H), 1.52 (s, 3H).  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  157.9, 142.5, 138.8, 135.0, 132.7, 131.0, 129.8, 129.1, 129.0, 128.93, 128.88, 128.2, 128.1, 127.3, 127.0, 126.7, 122.6, 120.9, 120.2, 113.9, 56.9, 56.8, 21.7, 18.3. **IR** (KBr) ( $\nu_{\text{max}}$ /  $\text{cm}^{-1}$ ): 3412, 3279, 3062, 2964, 1956, 1623, 1470, 1239, 745, 720. **HRMS** (EI) calcd. for  $C_{26}H_{26}NO$   $[M+H]^+$ , 368.2014, found 368.2000. Isolated yield: 72 %.



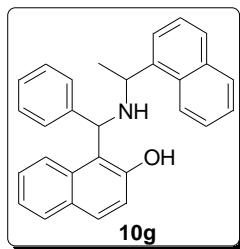
**1-((3-methoxyphenyl)((1-phenylethyl)amino)methyl)naphthalen-2-ol (10e):**

**1H NMR** (400 MHz, CDCl<sub>3</sub>) δ 13.56 (s, 1H), 7.76 (d, *J* = 8.7 Hz, 2H), 7.42-7.36 (m, 4H), 7.27-7.22 (m, 5H), 7.17 (t, *J* = 7.8 Hz, 1H), 6.82-6.74 (m, 3H), 5.46 (s, 1H), 3.92 (q, *J* = 6.6 Hz, 1H), 3.71 (s, 3H), 1.53 (d, *J* = 6.6 Hz, 3H). **13C NMR** (100 MHz, CDCl<sub>3</sub>) δ 160.1, 132.7, 130.2, 130.0, 129.1, 128.9, 128.8, 128.2, 127.0, 126.6, 122.6, 121.2, 120.1, 113.8, 113.3, 60.4, 57.0, 55.3, 22.9. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3427, 3290, 3059, 2965, 1601, 1456, 1264, 1096, 819. **HRMS** (EI) calcd. for C<sub>26</sub>H<sub>26</sub>NO [M+H]<sup>+</sup>, 384.1964, found 384.1953, **HRMS** (EI) calcd. for C<sub>26</sub>H<sub>25</sub>NONa [M+Na]<sup>+</sup>, 406.1783, found 406.1767. Isolated yield: 70 %.



**1-((4-methoxyphenyl)((1-phenylethyl)amino)methyl)naphthalen-2-ol (10f):**

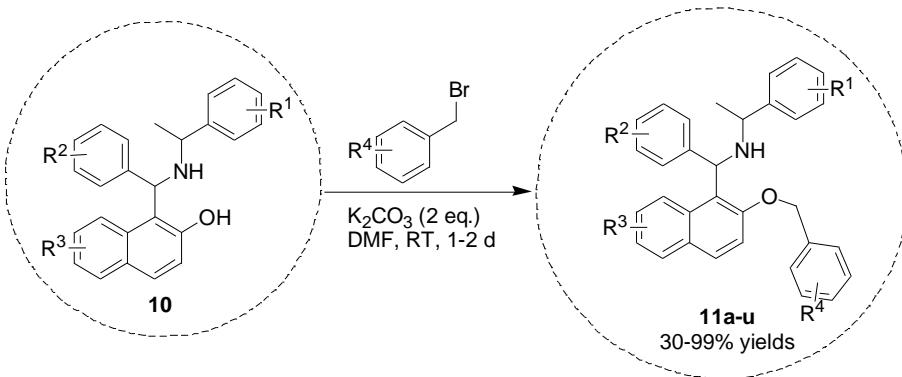
**1H NMR** (400 MHz, CDCl<sub>3</sub>) δ 13.43 (s, 1H), 7.73 (d, *J* = 8.5 Hz, 2H), 7.42 – 7.08 (m, 10H), 6.82 – 6.64 (m, 3H), 5.43 (s, 1H), 3.89 (q, *J* = 6.7 Hz, 1H), 3.67 (s, 3H), 1.50 (d, *J* = 6.7 Hz, 3H). **13C NMR** (100 MHz, CDCl<sub>3</sub>) δ 160.1, 157.2, 142.58, 130.2, 130.0, 129.1, 128.9, 128.2, 127.0, 126.6, 122.6, 121.2, 120.1, 113.8, 113.3, 112.8, 60.4, 56.9, 55.2, 22.9. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3422, 3309, 3023, 2996, 2360, 1600, 1457, 1417, 1264, 1164, 1038, 915, 816, 699. **HRMS** (EI) calcd. for C<sub>26</sub>H<sub>26</sub>NO [M+H]<sup>+</sup>, 384.1964, found 384.1942. Isolated yield: 79 %.



**1-(((1-(naphthalen-1-yl)ethyl)amino)(phenyl)methyl)naphthalene**

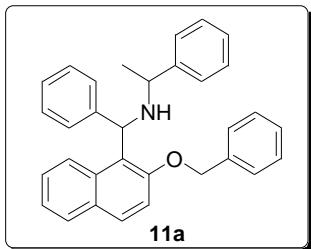
**n-2-ol (10g):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 13.94 (s, 1H), 7.82 (t, *J* = 6.8 Hz, 2H), 7.71 (d, *J* = 8.9 Hz, 1H), 7.68 – 7.42 (m, 4H), 7.35 (t, *J* = 7.3 Hz, 1H), 7.29 – 7.04 (m, 9H), 6.98 (t, *J* = 7.6 Hz, 1H), 5.47 (s, 1H), 4.84 (s, 1H), 2.55 (s, 1H), 1.56 (d, *J* = 6.3 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 157.3, 141.7, 129.9, 129.2, 128.7, 127.8, 126.4, 126.3, 125.9, 125.7, 122.4, 121.3, 120.2, 113.6, 61.3, 23.2. **IR** (KBr) (vmax/cm<sup>-1</sup>): 3422, 3320, 3049, 2961, 2361, 1621, 1454, 1379, 1239, 1100, 780, 744. **HRMS** (EI) calcd. for C<sub>29</sub>H<sub>26</sub>NO [M+H]<sup>+</sup>, 404.2014, found 404.2011. Isolated yield: 62 %.

### 3. The synthesis of Betti base-derived benzyl ethers **11**.



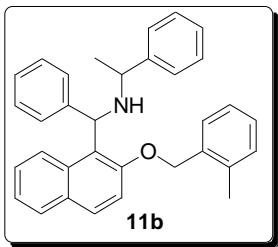
#### General procedure for the synthesis of 2-(benzyloxy)Aminoalkynaphthalene **11**.

In the following typical procedure, a mixture of aminoalkynaphthalol **10** (Betti base), benzyl bromide (1.2 eq), K<sub>2</sub>CO<sub>3</sub> (2.0 eq) in *N,N*-dimethylformamide (DMF) at room temperature for 1-2 days, The reaction was quenched with water and resulted mixture was extracted with ethyl acetate, and washed with water and saturated NaCl solution. The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated, and the residue was purified by column chromatography (hexanes/ethylacetate = 5/1) to give **11** in varied yields (see Table 1).



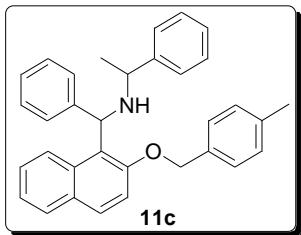
**N-((2-(benzyloxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11a):**

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.80 (s, 2H), 7.68 (s, 1H), 7.51 – 7.09 (m, 16H), 7.01 (s, 2H), 5.69 (s, 1H), 5.06 (d, *J* = 6.7 Hz, 1H), 4.87 (d, *J* = 6.7 Hz, 1H), 3.64 (s, 1H), 2.92 (s, 1H), 1.21 (s, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.7, 146.4, 145.2, 137.0, 128.7, 128.6, 128.4, 127.9, 127.4, 127.3, 126.9, 123.7, 115.1, 55.8, 55.3, 26.0. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3427, 3348, 3054, 3026, 2967, 1621, 1574, 1492, 1237, 1066, 803, 696. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>30</sub>NO [M+H]<sup>+</sup>, 444.2249, found 444.2348, **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>29</sub>NONa [M+Na]<sup>+</sup>, 466.2147, found 466.2165. Isolated yield: 85 %.

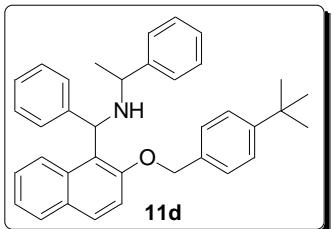


**N-((2-((2-methylbenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11b):**

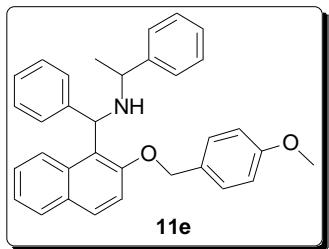
**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.80 (d, *J* = 8.2 Hz, 2H), 7.64 (s, 1H), 7.34–7.07 (m, 18H), 6.99 (d, *J* = 7.4 Hz, 1H), 5.64 (s, 1H), 5.02 (d, *J* = 11.7 Hz, 1H), 4.82 (d, *J* = 11.4 Hz, 1H), 3.62 (d, *J* = 5.3 Hz, 1H), 2.90 (s, 1H), 2.13 (s, 3H), 1.19 (d, *J* = 4.9 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.8, 136.1, 134.9, 128.7, 128.4, 128.2, 127.9, 127.3, 126.9, 126.1, 115.0, 69.5, 55.8, 55.3, 25.9, 18.7. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3421.87, 3329.81, 2966.17, 1623.30, 1599.92, 1455.77, 1257.61, 1241.11, 1080.65, 741.43. **HRMS** (EI) calcd. for C<sub>33</sub>H<sub>32</sub>NO [M+H]<sup>+</sup>, 458.2484, found 458.2473. Isolated yield: 65 %.



**N-((2-((4-methylbenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11c):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.80 – 7.70 (m, 2H), 7.64 (s, 1H), 7.40 (d, *J* = 7.0 Hz, 2H), 7.34 – 7.09 (m, 12H), 7.04 (d, *J* = 7.5 Hz, 2H), 6.86 (d, *J* = 7.4 Hz, 2H), 5.64 (s, 1H), 5.00 (d, *J* = 11.5 Hz, 1H), 4.79 (d, *J* = 11.4 Hz, 1H), 3.62 (q, *J* = 6.3 Hz, 1H), 2.97 (s, 1H), 2.30 (s, 3H), 1.19 (d, *J* = 6.2 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.7, 146.4, 145.2, 129.21, 129.16, 128.7, 128.4, 127.8, 127.5, 127.3, 127.0, 126.9, 126.5, 125.9, 123.6, 115.1, 71.0, 55.8, 55.3, 25.9, 21.3. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3422, 3334, 3056, 3016, 2921, 1595, 1511, 1492, 1234, 809, 700. **HRMS** (EI) calcd. for C<sub>33</sub>H<sub>32</sub>NO [M+H]<sup>+</sup>, 458.2484, found 458.2484. Isolated yield: 85 %.

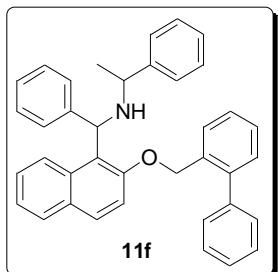


**N-((2-((4-(tert-butyl)benzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11d):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.79 (d, *J* = 8.6 Hz, 2H), 7.65 (s, 1H), 7.46 – 7.10 (m, 15H), 6.96 (d, *J* = 7.7 Hz, 2H), 5.65 (s, 1H), 5.04 (d, *J* = 11.6 Hz, 1H), 4.83 (d, *J* = 11.6 Hz, 1H), 3.62 (q, *J* = 6.2 Hz, 1H), 2.90 (s, 1H), 1.31 (s, 9H), 1.19 (d, *J* = 6.4 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.8, 150.9, 146.4, 145.2, 134.0, 128.4, 127.9, 127.3, 127.2, 127.0, 125.5, 123.6, 115.2, 71.1, 55.9, 34.7, 31.57, 26.0. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3421, 3362, 3024, 1622, 1513, 1252, 1070, 1015, 698. **HRMS** (EI) calcd. for C<sub>36</sub>H<sub>37</sub>NONa [M+Na]<sup>+</sup>, 522.2773, found 522.2784. Isolated yield: 99 %.

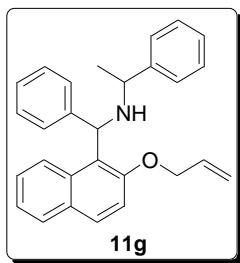


**N-((2-((4-methoxybenzyl)oxy)naphthalen-1-yl)(phenyl)m**

**ethyl)-1-phenylethanamine (11e):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.80 (d, *J* = 8.3 Hz, 2H), 7.63 (s, 1H), 7.51 – 7.07 (m, 15H), 6.90 (d, *J* = 7.7 Hz, 2H), 6.77 (d, *J* = 8.0 Hz, 2H), 5.62 (s, 1H), 5.00 (d, *J* = 11.1 Hz, 1H), 4.77 (d, *J* = 10.8 Hz, 1H), 3.78 (s, 3H), 3.61 (q, *J* = 5.8 Hz, 1H), 2.90 (s, 1H), 1.19 (d, *J* = 5.2 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 159.4, 154.7, 129.2, 129.0, 128.4, 127.9, 127.8, 127.4, 127.0, 126.6, 125.9, 123.6, 115.1, 113.9, 70.9, 55.8, 55.4, 25.9. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3423, 3337, 3057, 2999, 1618, 1513, 1246, 811, 702. **HRMS** (EI) calcd. for C<sub>33</sub>H<sub>32</sub>NO [M+H]<sup>+</sup>, 474.2433, found 474.2449. Isolated yield: 80 %.

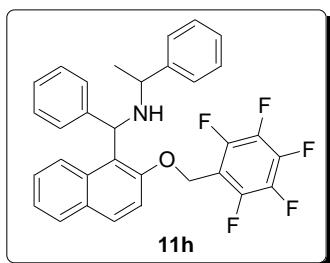


**N-((2-([1,1'-biphenyl]-2-ylmethoxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11f):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.78 (d, *J* = 7.8 Hz, 1H), 7.72 (d, *J* = 9.0 Hz, 1H), 7.62 (s, 1H), 7.44 – 7.10 (m, 21H), 7.08 (d, *J* = 8.9 Hz, 1H), 6.95 (s, 1H), 5.65 (s, 1H), 5.00 (d, *J* = 10.8 Hz, 1H), 4.73 (d, *J* = 9.0 Hz, 1H), 3.62 (s, 1H), 2.98 (s, 1H), 1.22 (s, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.5, 146.4, 145.2, 141.6, 140.5, 129.9, 129.3, 129.2, 128.7, 128.4, 127.9, 127.8, 127.4, 126.9, 123.6, 114.8, 69.0, 55.8, 55.2, 26.0. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3416, 3349, 3025, 2886, 1621, 1593, 1254, 1068, 1016, 770, 702. **HRMS** (EI) calcd. for C<sub>38</sub>H<sub>34</sub>NO [M+H]<sup>+</sup>, 520.2640, found 520.2636. Isolated yield: 90 %.

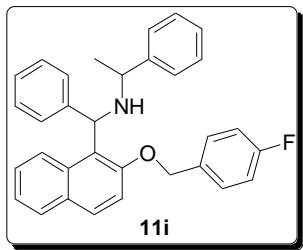


**N-((2-(allyloxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11g):**

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.79 (d, *J* = 8.1 Hz, 2H), 7.62 (s, 1H), 7.41 (d, *J* = 5.1 Hz, 2H), 7.37 – 7.04 (m, 11), 5.65 (d, *J* = 11.2 Hz, 2H), 5.13 (t, *J* = 13.9 Hz, 2H), 4.49 (s, 1H), 4.30 (s, 1H), 3.61 (s, 1H), 2.93 (s, 1H), 1.24 (s, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.7, 133.3, 129.5, 129.2, 128.7, 128.4, 127.9, 127.4, 126.9, 126.6, 127.0, 123.7, 117.5, 115.3, 70.2, 55.9, 55.2, 25.9. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3431, 3339, 3056, 2864, 2361, 1594, 1491, 1248, 805, 704. **HRMS** (EI) calcd. for C<sub>28</sub>H<sub>28</sub>NO [M+H]<sup>+</sup>, 394.2171, found 394.2169. Isolated yield: 99 %.

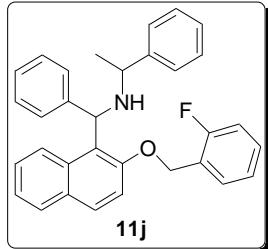


**N-((2-((perfluorophenyl)methoxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11h):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.87-7.82 (m, 2H), 7.62 (d, *J* = 6.0 Hz, 1H), 7.42 – 7.02 (m, 13H), 5.53 (s, 1H), 5.04 (d, *J* = 10.6 Hz, 1H), 4.85 (d, *J* = 10.1 Hz, 1H), 3.57 (q, *J* = 6.2 Hz, 1H), 2.64 (s, 1H), 1.21 (d, *J* = 6.0 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.1, 146.7, 146.2, 145.1, 144.2, 138.7, 136.2, 133.7, 130.0, 129.6, 128.8, 128.3, 127.8, 127.2, 126.8, 126.4, 125.9, 124.2, 123.9, 115.1, 110.1, 100.1, 55.8, 55.8, 54.9, 25.9. **<sup>19</sup>F NMR** (470 MHz, CDCl<sub>3</sub>) δ -142.07 (dd, *J* = 26.1, 5.9 Hz, 2F), -153.00 (t, *J* = 23.0 Hz, 1F), -160.33 – -162.08 (m, 2F). **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3423, 3368, 3064, 2957, 1595, 1512, 1256, 1131, 1023, 938, 618. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>24</sub>NOF<sub>5</sub> [M+Na]<sup>+</sup>, 556.1676, found: 556.1654. Isolated yield: 62 %.



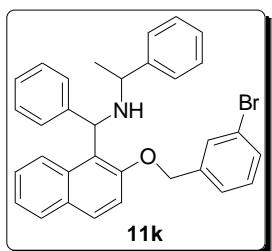
**N-((2-((4-fluorobenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11i):**

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.80 (d, *J* = 8.4 Hz, 2H), 7.64 (s, 1H), 7.47 – 7.06 (m, 13H), 6.94 – 6.82 (m, 4H), 5.64 (s, 1H), 5.00 (d, *J* = 11.3 Hz, 1H), 4.77 (d, *J* = 11.1 Hz, 1H), 3.59 (q, *J* = 5.8 Hz, 1H), 2.84 (s, 1H), 1.18 (d, *J* = 5.5 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 162.5 (d, *J*<sub>C-F</sub> = 244.0 Hz) 154.5, 146.3, 145.2, 132.6, 131.7, 129.3 (d, *J*<sub>C-F</sub> = 8.0 Hz), 128.8, 128.4, 128.0, 127.3, 126.9, 126.7, 126.0, 123.8, 115.4 (d, *J*<sub>C-F</sub> = 22 Hz), 115.0, 70.5, 55.8, 55.2, 26.0. **<sup>19</sup>F NMR** (471 MHz, CDCl<sub>3</sub>) δ -114.21 (s, 1F). **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3422, 3344, 3056, 2960, 2362, 1597, 1512, 1228, 1078, 1020, 703. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>29</sub>NOF [M+H]<sup>+</sup>, 462.2233, found 462.2235. Isolated yield: 82 %.



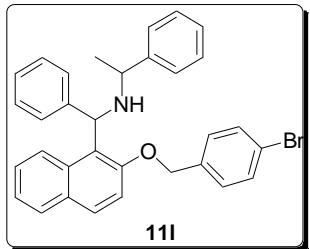
**N-((2-((2-fluorobenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11j):**

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.76 (d, *J* = 8.6 Hz, 2H), 7.67 (s, 1H), 7.42 (d, *J* = 6.5 Hz, 2H), 7.36 – 6.88 (m, 14H), 6.80 (d, *J* = 6.8 Hz, 1H), 5.69 (s, 1H), 5.16 (d, *J* = 12.1 Hz, 1H), 4.84 (d, *J* = 11.9 Hz, 1H), 3.63 (q, *J* = 5.9 Hz, 1H), 2.90 (s, 1H), 1.21 (d, *J* = 6.0 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 162.5 (d, *J*<sub>C-F</sub> = 240.0 Hz), 154.4, 145.7 (d, *J*<sub>C-F</sub> = 120.0 Hz), 129.7 (d, *J*<sub>C-F</sub> = 3 Hz), 129.6 (d, *J*<sub>C-F</sub> = 20 Hz), 128.8, 128.4, 127.9, 127.2, 126.9, 126.6, 125.9, 124.3 (d, *J*<sub>C-F</sub> = 4 Hz), 124.1, 124.0, 123.7, 115.2 (d, *J*<sub>C-F</sub> = 21 Hz), 114.8, 64.6, 55.8, 55.2, 25.9. **<sup>19</sup>F NMR** (470 MHz, CDCl<sub>3</sub>) δ -118.59 (s, 1F). **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3421, 3363, 3057, 2923, 1621, 1593, 1494, 1238, 804, 704. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>29</sub>NOF [M+H]<sup>+</sup>, 462.2233, found 462.2225. Isolated yield: 80 %.



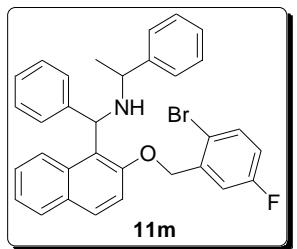
**N-((2-((3-bromobenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11k):**

**1H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.84 (d, *J* = 8.4 Hz, 2H), 7.69 (s, 1H), 7.49 – 7.06 (m, 16H), 6.89 (d, *J* = 7.4 Hz, 1H), 5.67 (s, 1H), 5.03 (d, *J* = 11.9 Hz, 1H), 4.78 (d, *J* = 11.8 Hz, 1H), 3.63 (q, *J* = 5.7 Hz, 1H), 2.85 (s, 1H), 1.23 (d, *J* = 5.7 Hz, 3H). **13C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.4, 139.2, 131.1, 130.3, 130.2, 128.4, 128.0, 127.3, 126.8, 125.9, 123.9, 115.0, 70.4, 55.8, 55.1, 26.0. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3338, 3061, 3023, 2958, 2360, 1596, 1513, 1235, 1070, 877, 801, 743, 696. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>29</sub>NOBr [M+H]<sup>+</sup>, 524.1433, found 524.1428. Isolated yield: 60 %.



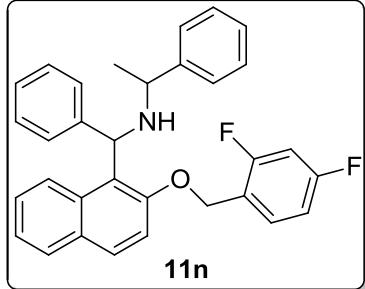
**N-((2-((4-bromobenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11l):**

**1H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.79 (d, *J* = 8.5 Hz, 2H), 7.64 (s, 1H), 7.47 – 7.09 (m, 15), 6.76 (d, *J* = 7.0 Hz, 2H), 5.64 (s, 1H), 4.98 (d, *J* = 11.5 Hz, 1H), 4.75 (d, *J* = 11.2 Hz, 1H), 3.59 (q, *J* = 5.1 Hz, 1H), 2.81 (s, 1H), 1.18 (d, *J* = 4.4 Hz, 3H). **13C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.4, 135.8, 131.7, 129.4, 129.1, 128.8, 128.4, 128.0, 127.3, 126.9, 126.7, 126.1, 123.9, 121.9, 114.8, 70.4, 55.8, 55.2, 26.0 (s). **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3421, 3370, 3063, 3023, 1594, 1490, 1215, 1069, 800, 701. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>29</sub>NOBr [M+H]<sup>+</sup>, 524.1433, found 524.1428. Isolated yield: 68 %.



**N-((2-((2-bromo-5-fluorobenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11m):**

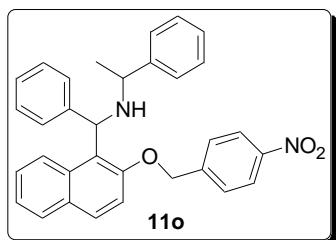
**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.81 (d, *J* = 8.6 Hz, 2H), 7.72 (s, 1H), 7.58 – 7.05 (m, 15H), 6.82 (t, *J* = 8.2 Hz, 1H), 6.55 (d, *J* = 7.9 Hz, 1H), 5.72 (s, 1H), 5.08 (d, *J* = 13.7 Hz, 1H), 4.80 (d, *J* = 13.7 Hz, 1H), 3.61 (q, *J* = 6.1 Hz, 1H), 2.77 (s, 1H), 1.23 (d, *J* = 6.1 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 162.2 (d, *J*<sub>C-F</sub> = 246.0 Hz), 154.0, 146.2, 145.0, 138.6 (d, *J*<sub>C-F</sub> = 8 Hz), 133.6 (d, *J*<sub>C-F</sub> = 8 Hz), 129.8, 129.5, 128.8, 128.4, 128.1, 127.2, 127.0, 126.8, 126.2, 124.0, 116.4, 116.1 (d, *J*<sub>C-F</sub> = 8 Hz), 115.8, 115.4, 114.8, 70.2, 55.8, 55.0, 26.1. **<sup>19</sup>F NMR** (471 MHz, CDCl<sub>3</sub>) δ -113.36 (s, 1F). **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3420, 3335, 3026, 2937, 1594, 1491, 1284, 1027, 806, 721, 701. Isolated yield: 77%.



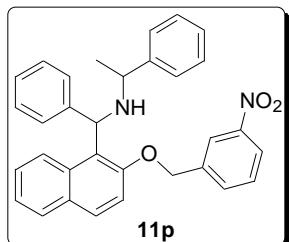
**N-((2-((2,4-difluorobenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11n):**

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.84 – 7.76 (m, 2H), 7.64 (s, 1H), 7.45 – 7.08 (m, 13H), 6.75 (t, *J* = 9.5 Hz, 1H), 6.65 (d, *J* = 6.9 Hz, 2H), 5.64 (s, 1H), 5.11 (d, *J* = 11.3 Hz, 1H), 4.77 (d, *J* = 10.7 Hz, 1H), 3.59 (q, *J* = 4.2 Hz, 1H), 2.77 (s, 1H), 1.19 (s, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 164.0 (d, *J*<sub>C-F</sub> = 12.0 Hz), 161.5 (t, *J*<sub>C-F</sub> = 20.0 Hz), 159.1 (d, *J*<sub>C-F</sub> = 12.0 Hz), 154.3, 146.3, 145.2, 133.7, 130.9, 129.7, 129.4, 128.8, 128.4, 128.0, 127.3, 126.9, 126.7, 126.0, 123.9, 120.1 (d, *J*<sub>C-F</sub> = 10.0 Hz), 114.8, 111.5 (dd, *J*<sub>C-F</sub> = 23.0 Hz, 3 Hz), 103.7 (t, *J*<sub>C-F</sub> = 25.0 Hz), 64.2, 55.8, 55.1, 26.0. **<sup>19</sup>F NMR** (470 MHz, CDCl<sub>3</sub>) δ -110.26 (s, 1F), -114.70 (s, 1F). **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3425, 3025, 2923, 2368, 1622, 1508, 1253, 1098, 962, 849, 804, 701. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>28</sub>NOF<sub>2</sub> [M+H]<sup>+</sup>, 480.2139,

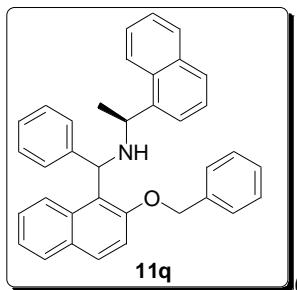
found 480.2142. Isolated yield: 73 %.



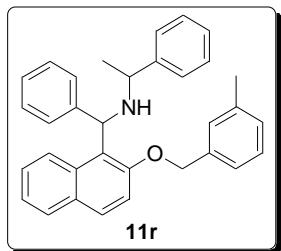
**N-((2-((4-nitrobenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11o):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.03 (d, *J* = 7.9 Hz, 2H), 7.80 (d, *J* = 8.6 Hz, 2H), 7.66 (s, 1H), 7.47 – 7.09 (m, 13H), 6.99 (d, *J* = 7.5 Hz, 2H), 5.67 (s, 1H), 5.11 (d, *J* = 12.4 Hz, 1H), 4.87 (d, *J* = 12.5 Hz, 1H), 3.57 (q, *J* = 4.9 Hz, 1H), 2.69 (s, 1H), 1.18 (d, *J* = 3.9 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.0, 147.6, 144.2, 129.5, 128.8, 128.5, 128.1, 127.7, 127.2, 126.9, 126.2, 124.1, 123.8, 114.7, 70.0, 55.8, 55.0, 26.2. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3427, 2366, 2332, 1625, 1522, 1345, 1112, 1085, 1028, 805, 701. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>29</sub>N<sub>2</sub>O<sub>3</sub> [M+H]<sup>+</sup>, 489.2178, found 489.2180. Isolated yield: 36 %.



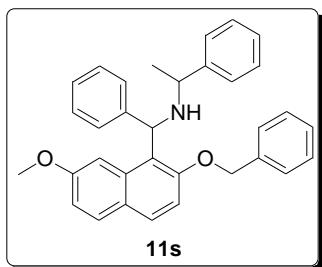
**N-((2-((3-nitrobenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11p):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.10 (d, *J* = 8.1 Hz, 1H), 7.99 (s, 1H), 7.83 (d, *J* = 8.7 Hz, 2H), 7.71 (s, 1H), 7.43 (d, *J* = 7.1 Hz, 2H), 7.40 – 7.06 (m, 13H), 5.70 (s, 1H), 5.12 (d, *J* = 12.0 Hz, 1H), 4.81 (d, *J* = 11.9 Hz, 1H), 3.63 (q, *J* = 6.2 Hz, 1H), 2.75 (s, 1H), 1.23 (d, *J* = 5.6 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.1, 148.3, 138.9, 133.4, 129.6, 128.4, 128.0, 127.2, 126.76 (s), 124.1, 122.9, 122.1, 115.0, 70.1, 55.8, 55.0, 26.0. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3446, 2918, 2850, 2363, 1734, 1653, 1559, 1508, 1348, 1240, 1093, 803. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>29</sub>N<sub>2</sub>O<sub>3</sub> [M+H]<sup>+</sup>, 489.2178, found 489.2177. Isolated yield: 30 %.



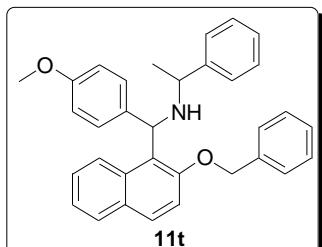
**(1S)-N-((2-(benzyloxy)naphthalen-1-yl)(phenyl)methyl)-1-(naphthalen-1-yl)ethanamine (11q):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.93 – 7.04 (m, 31H), 5.54 (d, *J* = 12.0 Hz, 1H), 5.30 (d, *J* = 12.0 Hz, 1H), 5.24 (d, *J* = 12.1 Hz, 1H), 5.03 (d, *J* = 11.6 Hz, 1H, diastereomeric isomer mixtures), 3.63 (q, *J* = 6.3 Hz, 1H), 2.83 (s, 1H), 1.21 (d, *J* = 6.5 Hz, 1H, diastereomeric isomer mixtures), 1.14 (d, *J* = 6.4 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.9, 146.4, 145.1, 128.80, 128.3, 127.8, 127.3, 126.8, 126.6, 125.9, 125.8, 125.5, 123.7, 123.4, 115.2, 69.7, 63.1, 55.9, 25.9. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3446, 3032, 2920, 2362, 1717, 1653, 1623, 1541, 1508, 1457, 1238, 1068, 801, 779, 700. **HRMS** (EI) calcd. for C<sub>36</sub>H<sub>32</sub>NO [M+H]<sup>+</sup>, 494.2484, found 494.2485. Dr = 67:37; Isolated yield: 83%.



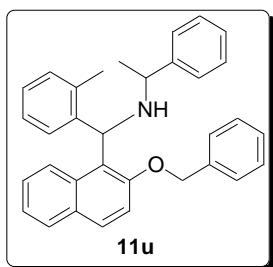
**N-((2-((3-methylbenzyl)oxy)naphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11r):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.84 (d, *J* = 8.1 Hz, 2H), 7.68 (s, 1H), 7.44 (d, *J* = 6.6 Hz, 2H), 7.41 – 7.16 (m, 13H), 7.11 (d, *J* = 7.3 Hz, 1H), 6.87 (s, 2H), 5.68 (s, 1H), 5.08 (d, *J* = 11.6 Hz, 1H), 4.88 (d, *J* = 11.5 Hz, 1H), 3.66 (q, *J* = 5.5 Hz, 1H), 3.00 (s, 1H), 2.32 (s, 3H), 1.24 (d, *J* = 5.3 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.7, 146.4, 145.2, 138.2, 136.9, 128.7, 128.5, 128.4, 128.1, 127.9, 127.4, 127.0, 124.5, 115.1, 71.2, 55.8, 55.3, 26.0, 21.5. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3422, 2961, 2866, 2362, 1623, 1559, 1490, 1457, 1253, 1068, 804, 700. Isolated yield: 86 %.



**N-((2-(benzyloxy)-7-methoxynaphthalen-1-yl)(phenyl)methyl)-1-phenylethanamine (11s):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.72 (dd, *J* = 14.2, 8.9 Hz, 2H), 7.47 (d, *J* = 6.4 Hz, 2H), 7.35 (d, *J* = 7.4 Hz, 3H), 7.32 – 7.15 (m, 10H), 7.00 (d, *J* = 8.0 Hz, 3H), 5.63 (s, 1H), 5.08 (d, *J* = 11.7 Hz, 1H), 4.87 (d, *J* = 11.4 Hz, 1H), 3.66 (q, *J* = 5.8 Hz, 1H), 3.50 (s, 3H), 2.91 (s, 1H), 1.24 (d, *J* = 5.8 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 158.3, 155.3, 146.6, 145.4, 137.0, 130.2, 128.5, 128.4, 128.0, 127.4, 127.3, 126.9, 116.9, 112.3, 70.9, 55.7, 55.3, 54.9, 26.2. **IR** (KBr) (vmax/cm<sup>-1</sup>): 3446, 3058, 3026, 2953, 2361, 1626, 1515, 1420, 1229, 1029, 970, 826, 700. **HRMS** (EI) calcd. for C<sub>33</sub>H<sub>32</sub>NO [M+H]<sup>+</sup>, 474.2355, found 474.2429. Isolated yield: 60 %.



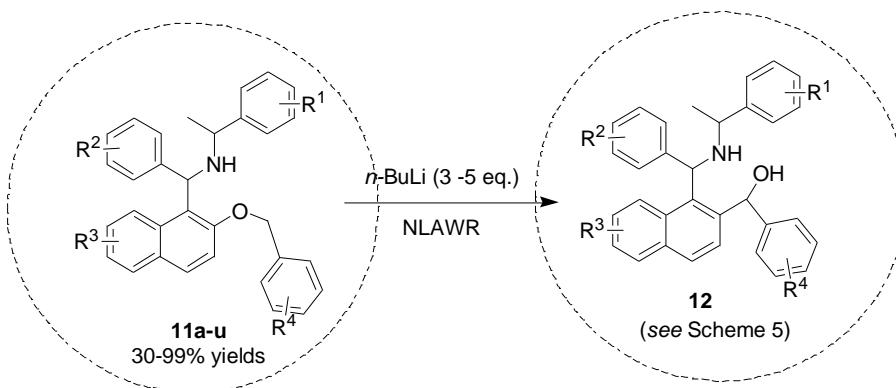
**N-((2-(benzyloxy)naphthalen-1-yl)(4-methoxyphenyl)methyl)-1-phenylethanamine (11t):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.63 (s, 1H), 7.37 (d, *J* = 9.2 Hz, 2H), 7.33-7.26 (m, 14H), 7.12 – 7.00 (m, 3H), 5.63 (d, *J* = 9.5 Hz, 1H), 5.15 (d, *J* = 11.6 Hz, 1H), 5.08 (d, *J* = 11.6 Hz, 1H), 3.69 (q, *J* = 6.4 Hz, 1H), 2.76 (s, 1H), 1.97 (s, 3H), 1.21 (d, *J* = 6.4 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 154.9, 145.7, 141.9, 130.6, 129.3, 128.7, 128.3, 128.2, 128.1, 127.6, 127.0, 126.5, 125.6, 123.5, 114.5, 71.1, 55.9, 54.8, 24.8, 19.9. **IR** (KBr) (vmax/cm<sup>-1</sup>): 3421, 2361, 1653, 1559, 1508, 1457, 1262, 1221, 1023, 806, 700. **HRMS** (EI) calcd. for C<sub>33</sub>H<sub>32</sub>NO<sub>2</sub> [M+H]<sup>+</sup>, 474.2433, found 474.2433. Isolated yield: 68 %.



**N-((2-(benzyloxy)naphthalen-1-yl)(o-tolyl)methyl)-1-phenylethananamine: (11u)**

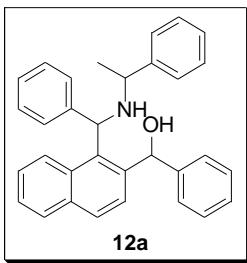
**1<sup>H</sup> NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.80 (d, *J* = 7.8 Hz, 2H), 7.66 (s, 1H), 7.36-7.06 (m, 15H), 6.86 (d, *J* = 6.8 Hz, 1H), 6.69 (d, *J* = 7.6 Hz, 1H), 5.63 (s, 1H), 5.07 (d, *J* = 11.6 Hz, 1H), 4.86 (d, *J* = 11.4 Hz, 1H), 3.70 (s, 3H), 3.62 (d, *J* = 5.9 Hz, 1H), 1.19 (d, *J* = 6.0 Hz, 3H). **13C NMR** (100 MHz, CDCl<sub>3</sub>) δ 159.6, 154.7, 147.2, 146.4, 137.0, 129.2, 128.8, 128.7, 128.5, 128.4, 127.9, 127.4, 127.3, 126.9, 126.6, 123.7, 119.5, 115.1, 113.0, 111.1, 71.2, 55.8, 55.2, 26.0. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3430, 3333, 3030, 2964, 2835, 1609, 1454, 1434, 1283, 1235, 1144, 1046, 808, 768, 703. **HRMS** (EI) calcd. for C<sub>33</sub>H<sub>32</sub>NO [M+H]<sup>+</sup>, 458.2484, found 458.2485. Isolated yield: 40 %.

#### 4. The synthesis of Betti base-derived amino alcohols 12.

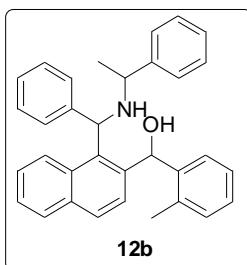


Under argon atmosphere, to a solution of **11** (2 mmol) in the dry THF (10 ml) at -78 °C, the *n*-BuLi (4 mL, 10 mmol, 2.5 M in hexane) was added slowly in 15 min. After the addition, the solution was allowed to stir at -78 °C for 1-2 h. And then the reaction solution was warmed to room temperature slowly. After quenched with saturated NH<sub>4</sub>Cl solution, the resulted mixture was extracted with ethyl acetate, and washed with water and saturated NaCl solution. The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated, and the residue was purified by column chromatography (hexanes/ethyl

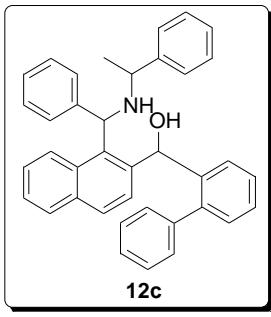
acetate = 20/1) to give **12** as a colorless powder.



**phenyl(1-(phenylethyl)amino)methyl)naphthalen-2-yl)methanol (12a):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.89 (d, *J* = 8.1 Hz, 1H), 7.71 (t, *J* = 6.8 Hz, 2H), 7.51 (d, *J* = 6.8 Hz, 1H), 7.42-7.15 (m, 17H), 6.32 (s, 1H), 6.09 (s, 1H), 3.62 (q, *J* = 5.6 Hz, 1H), 2.11 (s, 1H), 1.43 (d, *J* = 5.6 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 144.4, 143.3, 142.9, 142.1, 137.0, 133.1, 128.8, 128.7, 128.6, 128.0, 127.6, 127.5, 126.9, 126.7, 126.4, 126.3, 126.0, 125.7, 123.9, 70.8, 56.5, 56.2, 24.5. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3407, 3058, 3025, 2963, 2359, 1725, 1710, 1600, 1449, 1030, 818, 700. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>30</sub>NO [M+H]<sup>+</sup>, 444.22, found 444.2313, **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>29</sub>NO [M+Na]<sup>+</sup>, 466.2147, found 466.2118. Isolated yield: 76 %.

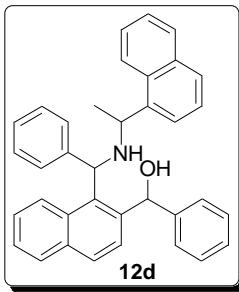


**(1-(phenyl((1-phenylethyl)amino)methyl)naphthalen-2-yl)(o-toyl)methanol (12b):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.91 (d, *J* = 7.6 Hz, 1H), 7.81 (d, *J* = 8.1 Hz, 1H), 7.64 (d, *J* = 8.5 Hz, 2H), 7.45 (d, *J* = 6.9 Hz, 3H), 7.38 – 7.13 (m, 10H), 7.00 (d, *J* = 5.6 Hz, 2H), 6.94 – 6.81 (m, 2H), 6.20 (s, 1H), 6.16 (s, 1H), 3.59 (q, *J* = 6.5 Hz, 1H), 1.50 (d, *J* = 5.7 Hz, 3H), 0.89 (s, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 141.5, 141.0, 135.1, 133.5, 133.3, 130.0, 129.3, 129.0, 128.9, 128.7, 128.5, 127.8, 127.0, 126.9, 126.6, 126.5, 126.4, 125.9, 125.9, 124.0, 68.4, 55.9, 55.5, 24.6, 18.4. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3428, 3054, 2368, 2344, 1636, 1449, 1385, 1031, 737, 700. **HRMS** (EI) calcd. for C<sub>33</sub>H<sub>32</sub>NO [M+H]<sup>+</sup>, 458.2484, found 458.2475. Isolated yield: 56 %.

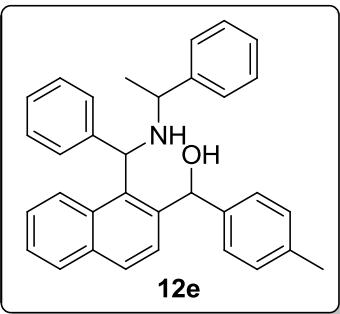


**[1,1'-biphenyl]-2-yl(1-(phenyl((1-phenylethyl)amino)methyl)**

**naphthalen-2-yl)methanol (12c):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.00 (d, *J* = 6.7 Hz, 1H), 7.88 (d, *J* = 8.1 Hz, 1H), 7.78 (d, *J* = 8.6 Hz, 2H), 7.60 – 6.59 (m, 19H), 6.06 (s, 1H), 3.51 (q, *J* = 6.4 Hz, 1H), 2.09 (s, 1H), 1.35 (d, *J* = 6.4 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 141.4, 140.8, 140.7, 133.6, 133.1, 130.7, 128.9, 128.8, 128.6, 128.5, 128.4, 127.9, 127.7, 127.6, 127.4, 126.8, 126.7, 126.2, 126.0, 125.5, 124.9, 69.7, 56.0, 31.7, 22.8. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3426, 3058, 3025, 2958, 2923, 2856, 2368, 2344, 2332, 1624, 1508, 1449, 1099, 1028, 819, 748, 701. **HRMS** (EI) calcd. for C<sub>38</sub>H<sub>34</sub>NO [M+H]<sup>+</sup>, 520.2640, found 520.2642. Isolated yield: 79 %.

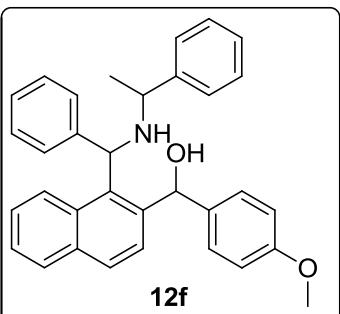


**(1-(((1-(naphthalen-1-yl)ethyl)amino)(phenyl)methyl)naphthalen-2-yl)(phenyl)methanol (12d):** **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.80 (d, *J* = 8.1 Hz, 2H), 7.69 (d, *J* = 8.4 Hz, 2H), 7.58 (d, *J* = 7.5 Hz, 1H), 7.54 – 6.99 (m, 21H), 6.76 (s, 1H, mixture of diastereomers), 6.23 (s, 2H), 4.59 (s, 1H), 0.98 (d, *J* = 6.6 Hz, 1H, mixture of diastereomers), 0.92 – 0.85 (m, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 143.4, 142.9, 141.8, 140.3, 137.2, 134.1, 133.1, 131.5, 128.8, 128.7, 128.6, 128.5, 128.1, 127.8, 127.6, 127.4, 127.0, 126.4, 126.9, 126.4, 126.1, 126.0, 125.9, 125.7, 125.6, 122.4, 122.2, 71.0, 56.5, 24.5, 14.3. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3433, 3057, 2918, 2850, 2364, 1699, 1653, 1636, 1541, 1508, 1457, 1110, 1030, 778, 728. **HRMS** (EI) calcd. for C<sub>36</sub>H<sub>31</sub>NO [M+H]<sup>+</sup>, 494.2484, found 494.2483. Isolated yield: 62 %.



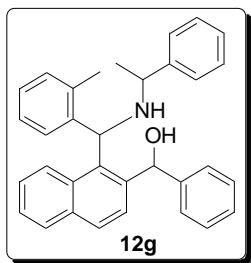
**(1-(phenyl((1-phenylethyl)amino)methyl)naphthalen-2-yl)(p-tolyl)methanol (12e):**

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.84 (d, *J* = 8.1 Hz, 1H), 7.74 (d, *J* = 8.5 Hz, 1H), 7.67 (s, 1H), 7.48 (d, *J* = 6.7 Hz, 1H), 7.42 (d, *J* = 7.2 Hz, 2H), 7.38 – 7.14 (m, 10H), 7.09 (d, *J* = 5.1 Hz, 2H), 7.02 (d, *J* = 4.1 Hz, 2H), 6.25 (s, 1H), 6.09 (s, 1H), 3.6 (b, 1H), 2.34 (d, *J* = 2.3 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 142.1, 139.9, 136.4, 133.2, 129.1, 128.93, 128.85, 128.7, 128.3, 128.0, 127.8, 127.1, 127.0, 126.5, 126.3, 126.2, 125.8, 123.8, 70.7, 56.7, 24.4, 21.2. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3421, 3025, 2965, 2362, 1653, 1508, 1457, 1262, 1030, 909, 761, 700. **HRMS** (EI) calcd. for C<sub>33</sub>H<sub>32</sub>NO [M+H]<sup>+</sup>, 458.2484, found 458.2485. Isolated yield: 42 %.



**(4-methoxyphenyl)(1-(phenyl((1-phenylethyl)amino)methyl)naphthalen-2-yl)methanol (12f):**

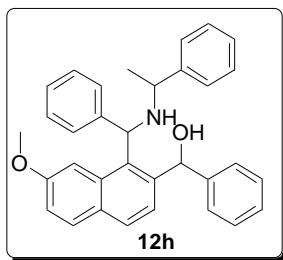
**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.83 (d, *J* = 8.1 Hz, 1H), 7.74 (d, *J* = 8.7 Hz, 1H), 7.63 (d, *J* = 8.2 Hz, 1H), 7.57 (d, *J* = 7.6 Hz, 1H), 7.50 – 7.10 (m, 11H), 7.02 (d, *J* = 8.1 Hz, 2H), 6.93 (d, *J* = 7.6 Hz, 1H), 6.81 (d, *J* = 7.8 Hz, 2H), 6.20 (s, 1H), 6.07 (s, 1H), 3.79 (s, 3H), 3.68 – 3.58 (m, 1H), 1.46 (d, *J* = 5.4 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 158.6, 142.1, 134.9, 133.2, 130.5, 129.9, 129.2, 129.0, 128.9, 128.8, 128.3, 128.0, 127.8, 127.5, 127.2, 127.1, 126.6, 126.3, 125.9, 123.8, 114.0, 113.6, 70.5, 56.8, 55.4, 24.3. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3421, 3309, 2930, 2654, 2164, 1624, 1508, 1457, 1262, 1230, 1032, 840, 700. **HRMS** (EI) calcd. for C<sub>33</sub>H<sub>32</sub>NO<sub>2</sub> [M+H]<sup>+</sup>, 474.2355, found HRMS+H: 474.2433. Isolated yield: 20 %.



**phenyl(1-((1-phenylethyl)amino)(o-tolyl)methyl)naphthalen-2-yl)methanol (12g):**

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.77 (d, *J* = 8.1 Hz, 1H), 7.67-7.61 (m, 2H), 7.40 (t, *J* = 7.4 Hz, 1H), 7.36 – 7.07 (m, 12H), 7.04 (d, *J* = 8.5 Hz, 1H), 6.91 (s, 1H), 6.84 (d, *J* = 7.7 Hz, 1H), 6.71 (d, *J* = 8.2 Hz, 1H), 6.23 (s, 1H), 5.94 (s, 1H), 3.66 (s, 3H), 3.52 (d, *J* = 6.6 Hz, 1H), 1.99 (s, 1H), 1.34 (d, *J* = 6.2 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 160.0, 143.0, 129.7, 128.9, 128.9, 128.1, 127.7, 126.9, 126.5, 126.5, 125.8, 118.7, 112.8, 111.8, 70.9, 56.6, 56.5, 55.4, 24.5. **IR** (KBr) (vmax/cm<sup>-1</sup>): 3447, 2967, 2920, 2166, 1636, 1508, 1457, 1261, 1101, 1048, 767, 698.

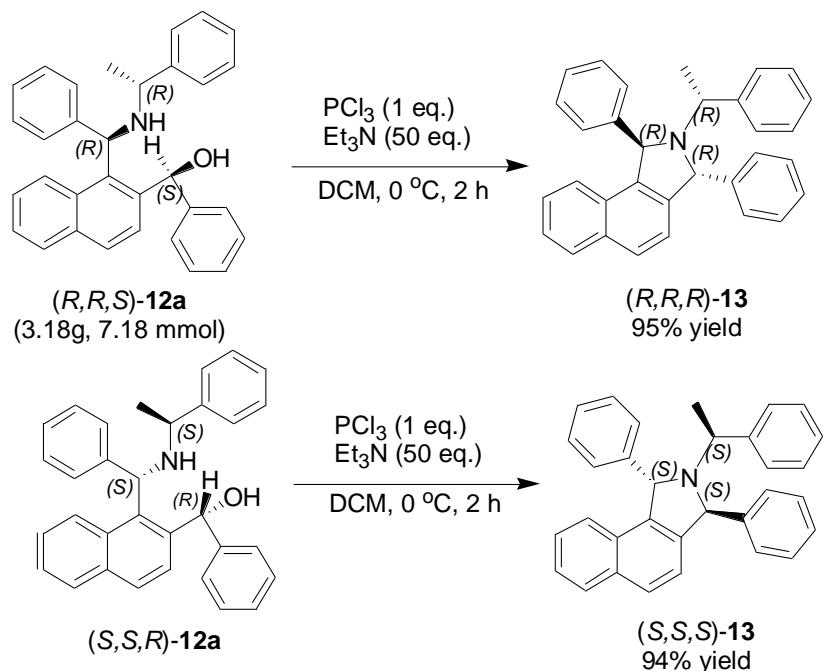
Isolated yield: 28 %.



**(7-methoxy-1-(phenyl((1-phenylethyl)amino)methyl)naphthalen-2-yl)(phenyl)methanol (12h):**

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.72 (d, *J* = 8.9 Hz, 1H), 7.65 (d, *J* = 8.5 Hz, 1H), 7.43 (d, *J* = 7.6 Hz, 2H), 7.38 – 7.19 (m, 10H), 7.15 (d, *J* = 7.4 Hz, 2H), 7.10 (d, *J* = 7.3 Hz, 2H), 6.96 – 6.86 (m, 2H), 6.22 (s, 1H), 5.98 (s, 1H), 3.60 (q, *J* = 6.5 Hz, 1H), 3.51 (s, 3H), 2.06 (s, 1H), 1.43 (d, *J* = 6.5 Hz, 3H). **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 158.2, 143.0, 130.3, 128.9, 128.8, 128.6, 128.1, 127.6, 126.9, 126.4, 126.0, 118.8, 102.1, 70.7, 56.5, 56.0, 54.9, 24.9. **IR** (KBr) (vmax/cm<sup>-1</sup>): 3421, 2164, 1624, 1559, 1507, 1457, 1262, 1230, 1032, 840. **HRMS** (EI) calcd. for C<sub>33</sub>H<sub>32</sub>NO<sub>2</sub> [M+H]<sup>+</sup>, 474.2355, found IR . HRMS+H 474.2433. Isolated yield: 20 %.

## 5. The synthesis of optically pure and all-substituted pyrrolidine derivative 13 through intramolecular cyclization.



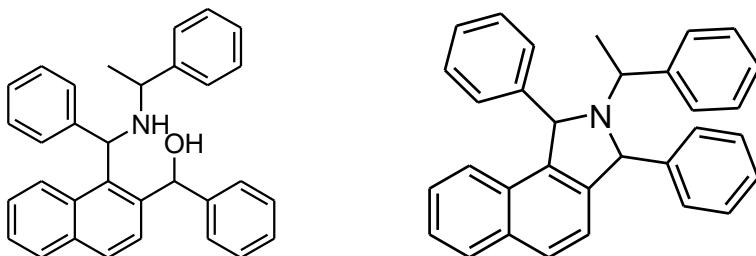
A 100 mL three-necked flask was charged with freshly distilled  $\text{PCl}_3$  (986.5 mg, 7.18 mmol) in anhydrous  $\text{CH}_2\text{Cl}_2$  (12 mL). The solution was cooled to 0 °C, and  $\text{NEt}_3$  (5 mL, 359 mmol) was added dropwise to the solution. And then a solution of **12a** (3.18g, 7.18 mol) in  $\text{CH}_2\text{Cl}_2$  (30 mL) was slowly added over 30 min to the reaction mixture, and the reaction solution was kept at 0 °C. After completing the addition, the ice bath was removed, and the resulting suspension was stirred for 90 min at room temperature. The reaction was quenched with  $\text{H}_2\text{O}$  (30 mL). The aqueous phase was extracted once with  $\text{CH}_2\text{Cl}_2$  (40 mL). The combined organic phases were dried over  $\text{Na}_2\text{SO}_4$ , the salt was removed by filtration, and the solution was evaporated to afford the crude product, which was purified by flash chromatography under pressure of argon:  $R_f = 0.4$  (3:1 (v/v) pentane/ $\text{CH}_2\text{Cl}_2$ ). The solvent was evaporated, and the product was dried under vacuum (0.04 mm) to afford **5** as a white powder.

**1,3-diphenyl-2-(1-phenylethyl)-2,3-dihydro-1H-benzo[e]isoindole:** **<sup>1</sup>H NMR** (400 MHz,  $\text{CDCl}_3$ ) δ 7.73 (d,  $J = 8.1$  Hz, 1H), 7.67 (d,  $J = 7.4$  Hz, 2H), 7.63 (d,  $J = 8.4$  Hz, 1H), 7.45 – 7.08 (m, 16H), 7.01 (d,  $J = 8.4$  Hz, 1H), 5.68 (s, 1H), 5.48 (s, 1H), 4.22 (q,  $J = 6.7$  Hz, 1H), 1.23 (d,  $J = 6.9$  Hz, 3H). **<sup>13</sup>C NMR** (100 MHz,  $\text{CDCl}_3$ ) δ 142.3, 140.6, 136.6, 133.6, 129.4, 128.8, 128.7, 128.6, 128.5, 128.3, 128.3, 128.0, 127.3, 127.0,

126.9, 126.1, 125.0, 124.2, 121.1, 70.6, 60.0, 20.4. **IR** (KBr) (vmax/ cm<sup>-1</sup>): 3420, 3059, 3028, 2970, 2369, 1680, 1600, 1492, 1453, 1145, 1026, 809, 700. **HRMS** (EI) calcd. for C<sub>32</sub>H<sub>28</sub>N [M+H]<sup>+</sup>, 426.2222, found 426.2213. Isolated yield: 95 %.

## 6. CD Spectroscopic Data of Chiral Products 12a and 13

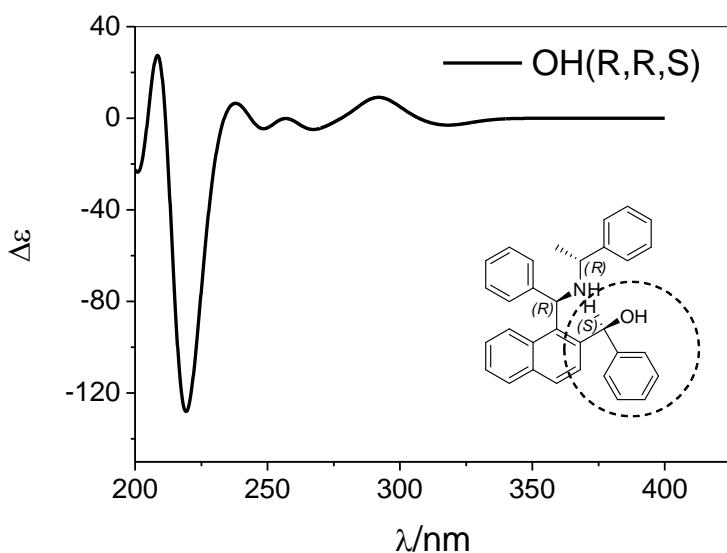
Computational details: The Gaussian 03 software package was used to carry out gas phase density functional theory (DFT) geometry optimizations<sup>[1]</sup>. The hybrid B3LYP functional was selected and 6-31G(d) basis sets were used. The same functional and basis set was used to obtain excitation energies and oscillator strengths from time dependent density functional theory (TD-DFT) calculations. The (R)- and (S)-silane are both taken as models for calculation. The calculated CD spectra were shown by the SpecDis software package (Version 1.51) software to yield higher-quality images, with comparison to the measured ones.<sup>[2]</sup>



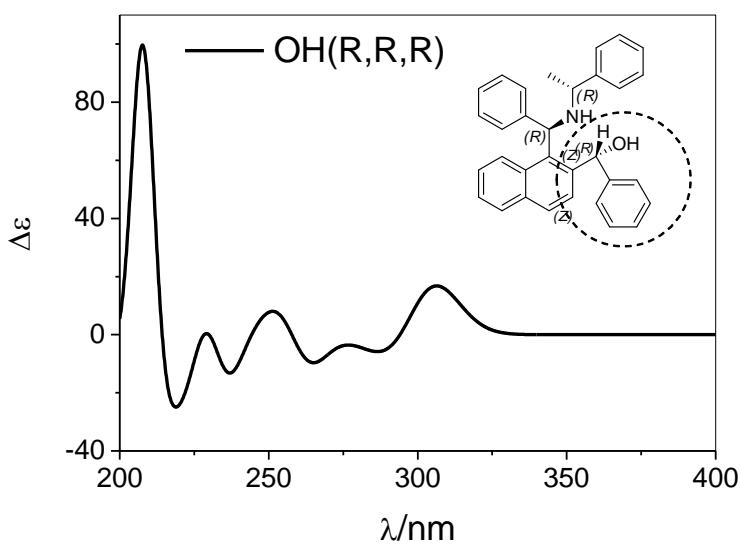
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- [1] Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Zakrzewski, V. G.; Montgomery, J. A.; Jr.; Stratmann, R. E.; Burant, J. C.; Dapprich, S.; Millam, J. M.; Daniels, A. D.; Kudin, K. N.; Strain, M. C.; Farkas, O.; Tomasi, J.; Barone, V.; Cossi, M.; Cammi, R.; Mennucci, B.; Pomelli, C.; Adamo, C.; Clifford, S.; Ochterski, J.; Petersson, G. A.; Ayala, P. Y.; Cui, Q.; Morokuma, K.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Cioslowski, J.; Ortiz, J. V.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Gomperts, R.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Gonzalez, C.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Andres, J. L.; Gonzalez, C.; Head-Gordon, M. E.; Replogle, S.; Pople, J. A. Gaussian 03, Inc., Pittsburgh PA, 2003.
- [2] a) Stephens, P.J.; Harada, N. *Chirality* 2010, 22, 229-233. b) Bringmann, G.; Bruhm, T.; Maksimenka, K.; Hemberger, Y. *Eur. J. Org. Chem.* 2009, 2717-2727. c) Bringmann, G.; Gulder, T. A. M.; Reichert, M.; Gulder, T. *Chirality* 2008, 20, 628-642.

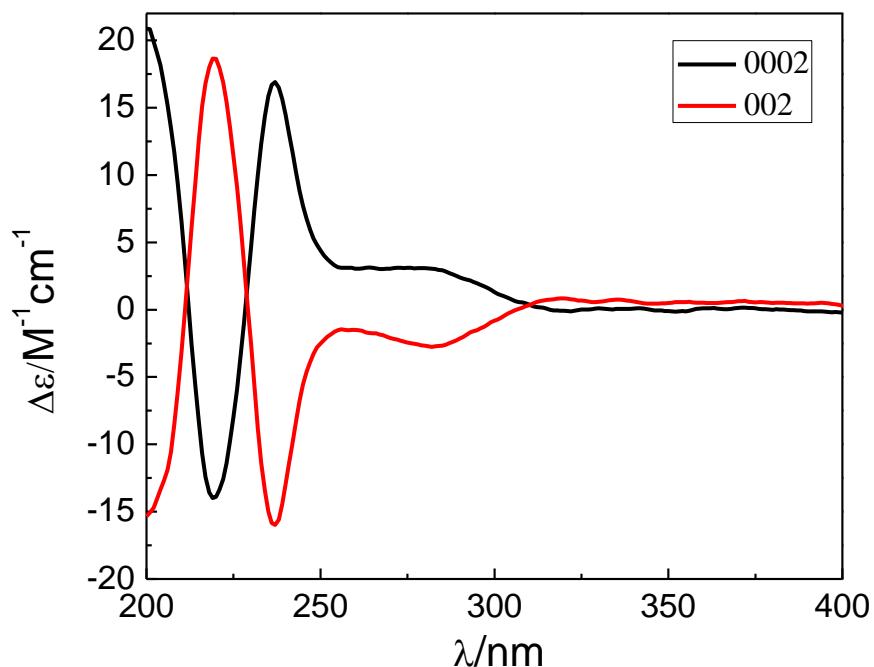
**Figure S1.** The experimental and calculated CD spectrum of chiral product **12a**: The absolute configuration of **12a** could be confirmed as corresponding configuration in comparison to that of the following calculated CD spectrum.



(a) Calculated CD spectra

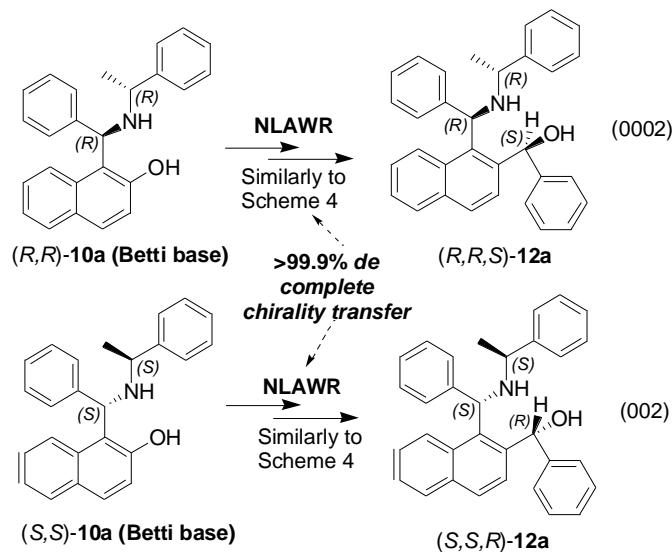


(b) Calculated CD spectra

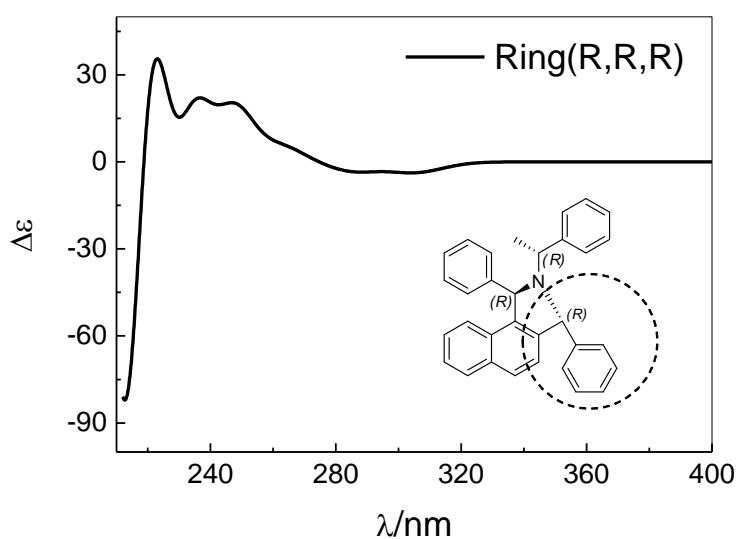


(c) Experimental CD spectra: Band width 2 nm; Response 1 sec; Sensitivity Standard; Data pitch 0.1 nm; Scanning speed 500 nm/min.  $c = 0.125\text{mmol/L}$ .

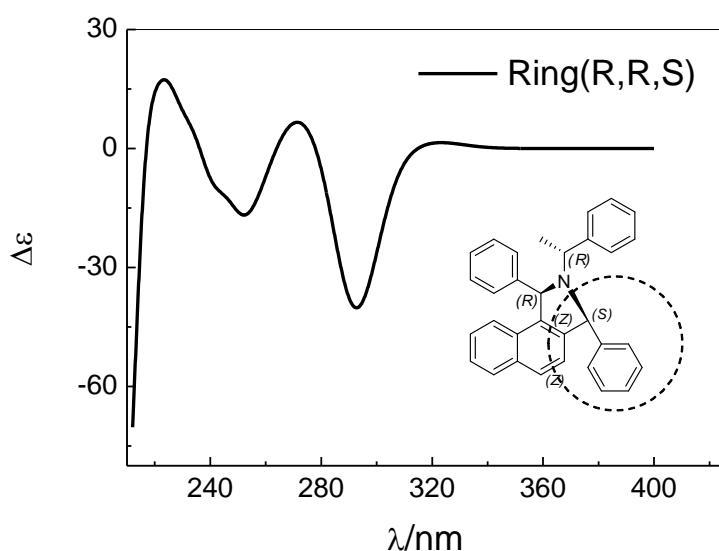
Results:



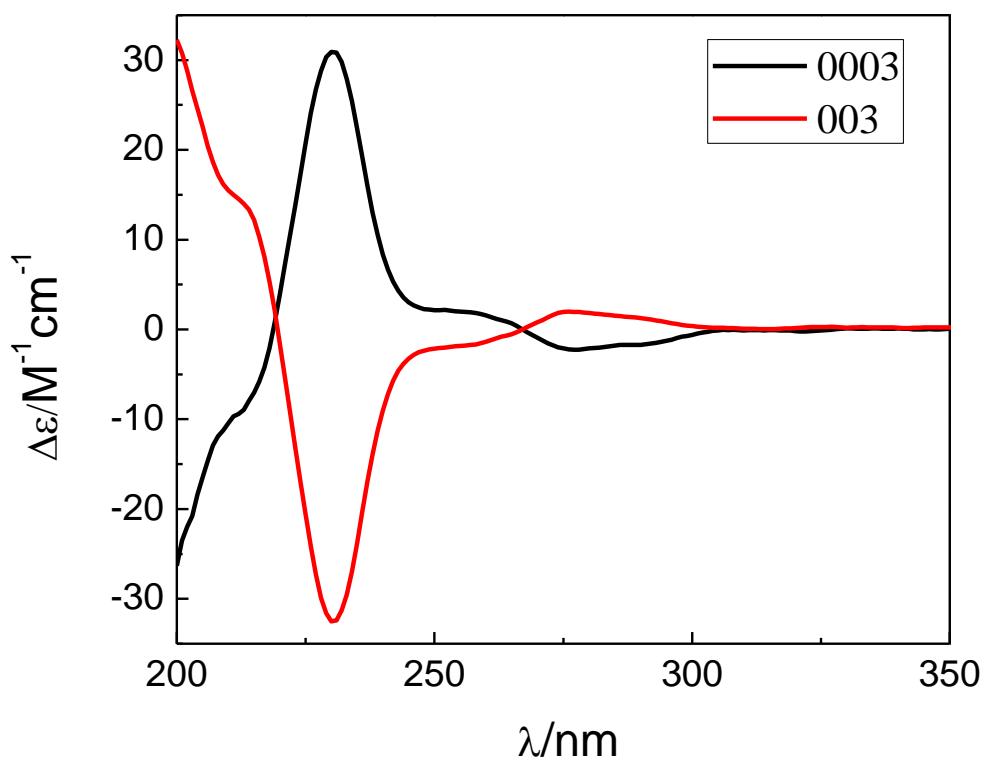
**Figure S2.** The experimental and calculated CD spectrum of chiral product **13**: The absolute configuration of **13** could be confirmed as corresponding configuration in comparison to that of the following calculated CD spectrum.



(a) Calculated spectra

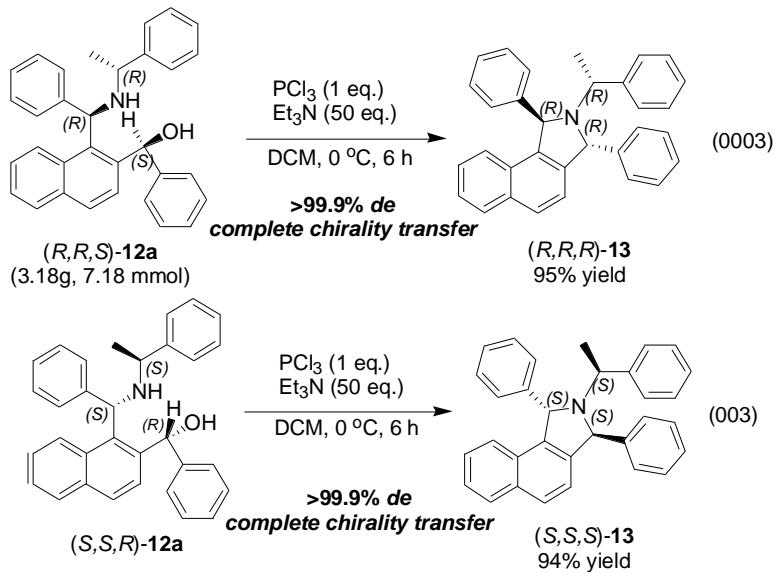


(b) Calculated spectra

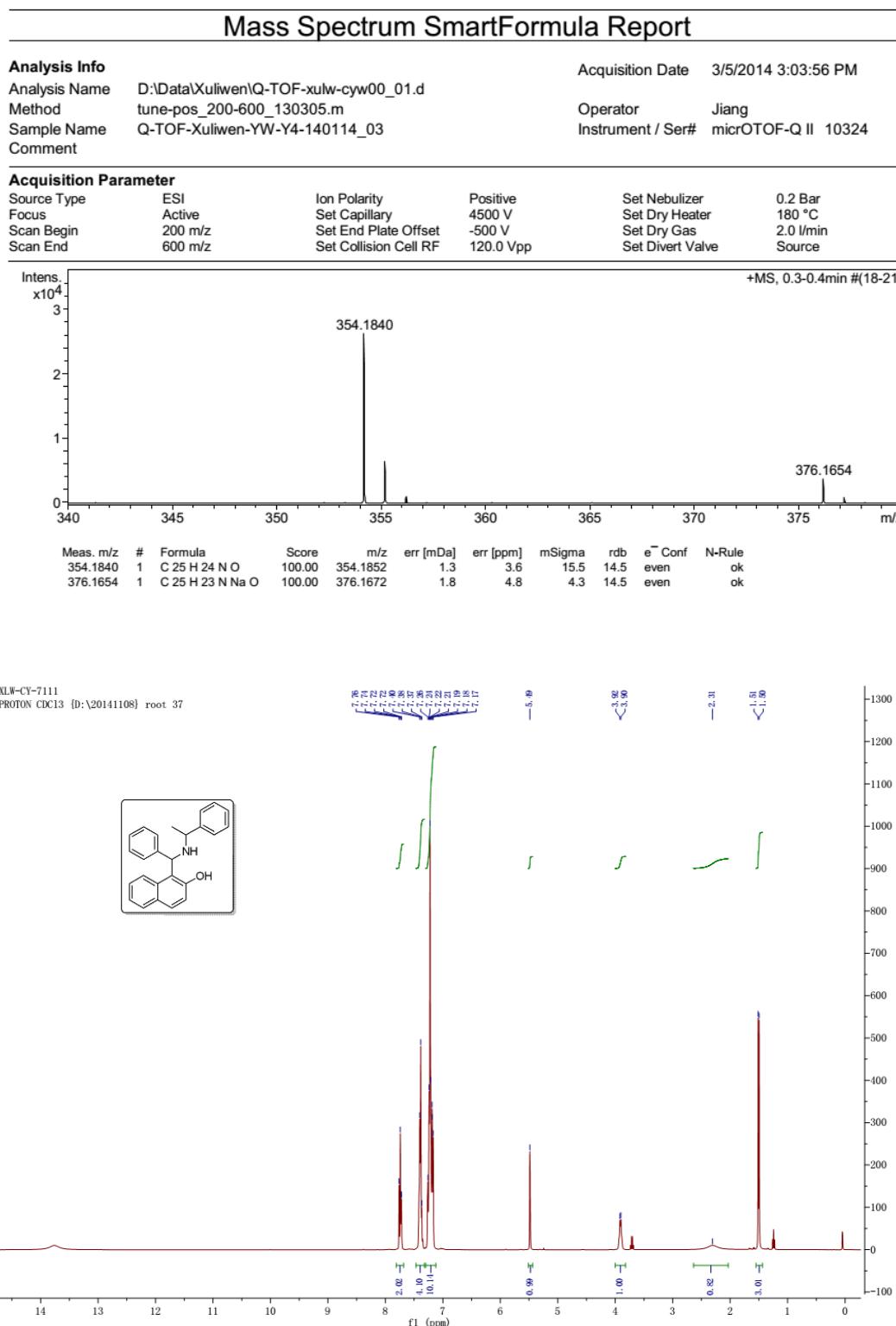


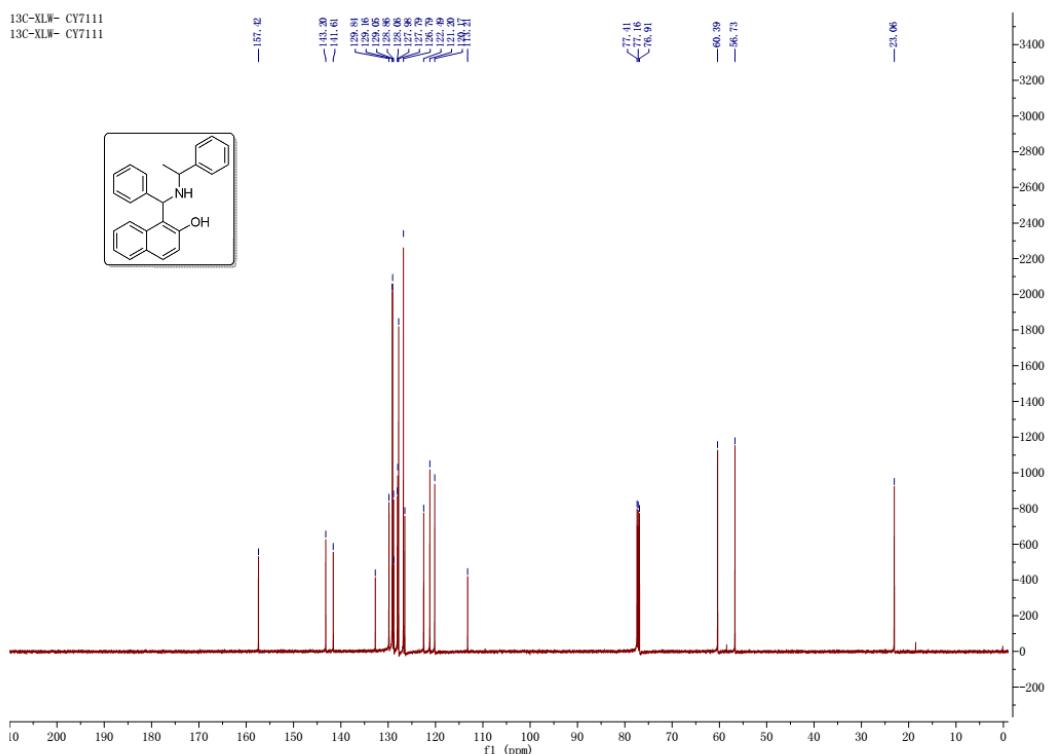
(c) Experimental CD spectra: Band width 2 nm; Response 1 sec; Sensitivity Standard; Data pitch 0.1 nm; Scanning speed 500 nm/min.  $c = 0.125\text{mmol/L}$ .

Result:



## 7. NMR and HRMS Charts of Betti bases and it derivatives





## Mass Spectrum SmartFormula Report

### Analysis Info

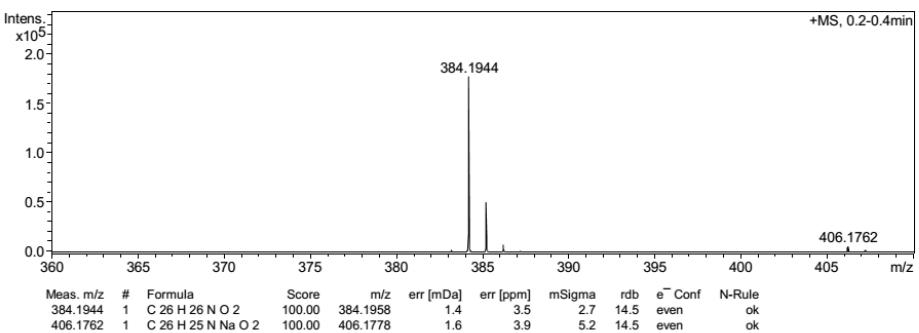
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Sample Name trz-7  
Comment

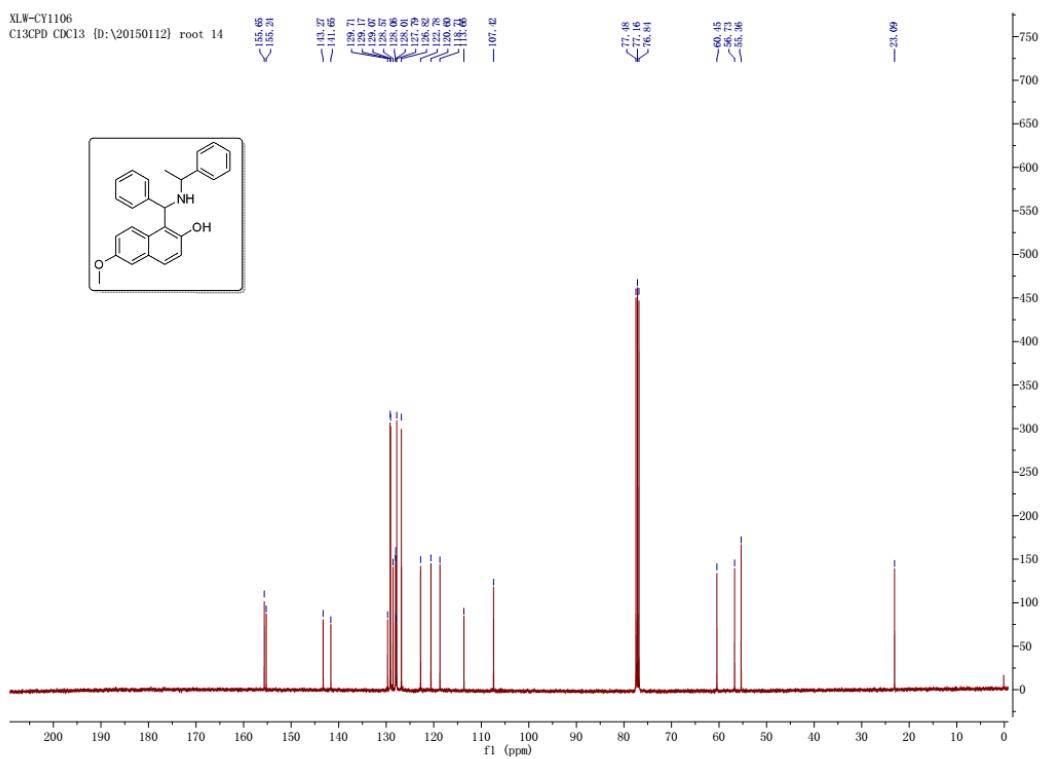
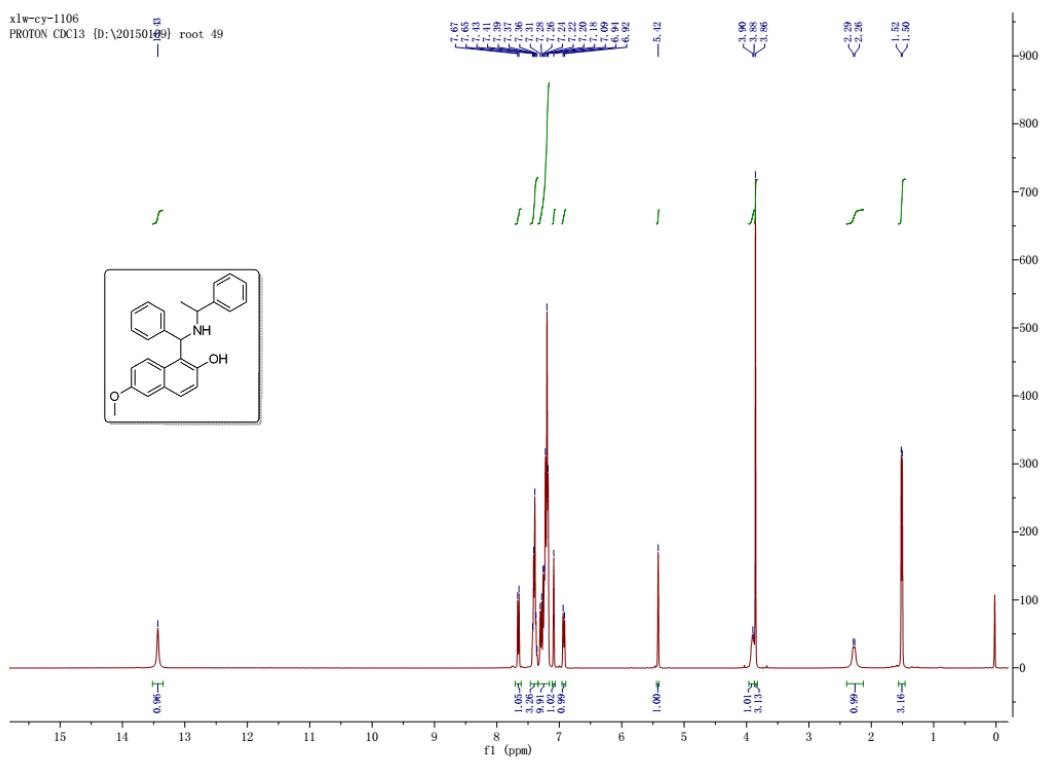
Acquisition Date 1/15/2015 2:44:16 PM

Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

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Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

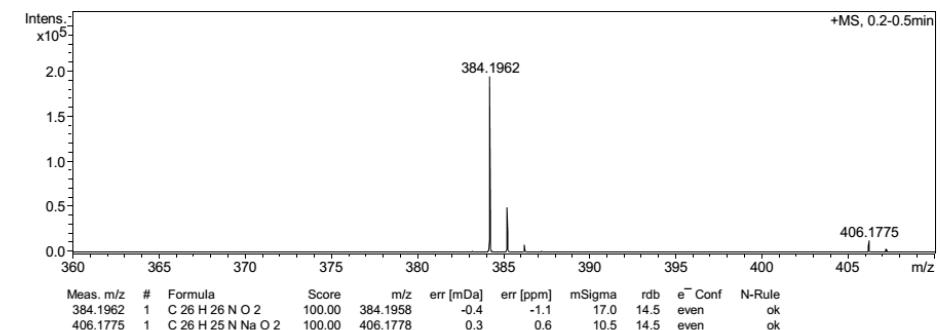
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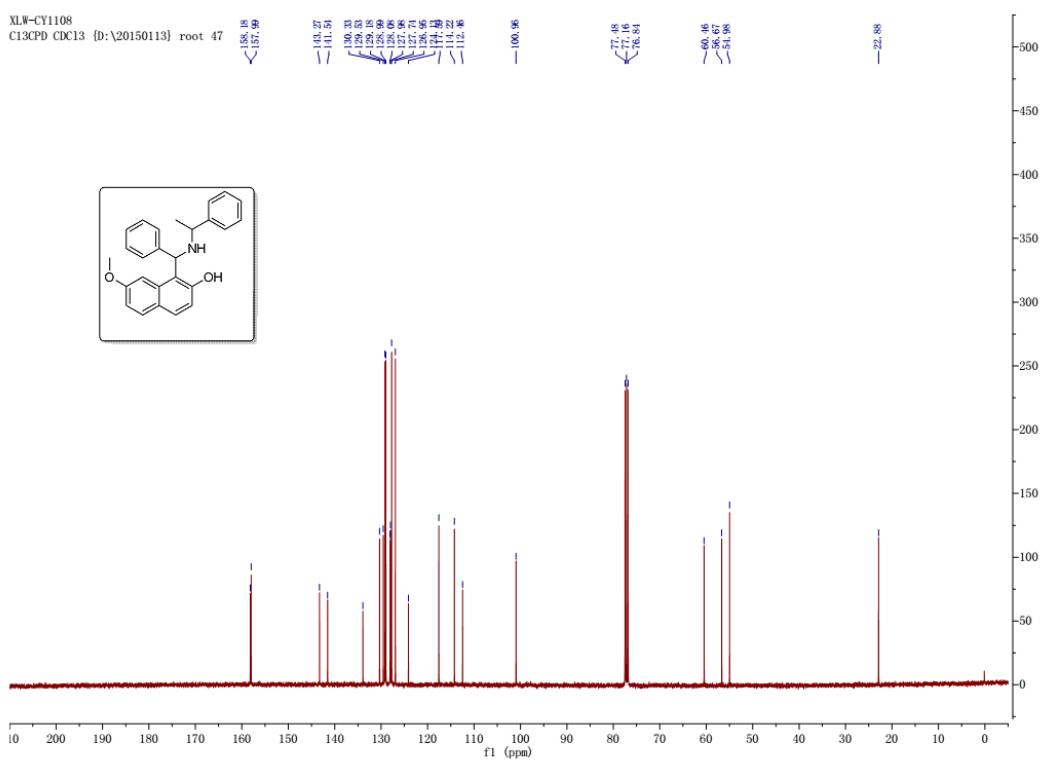
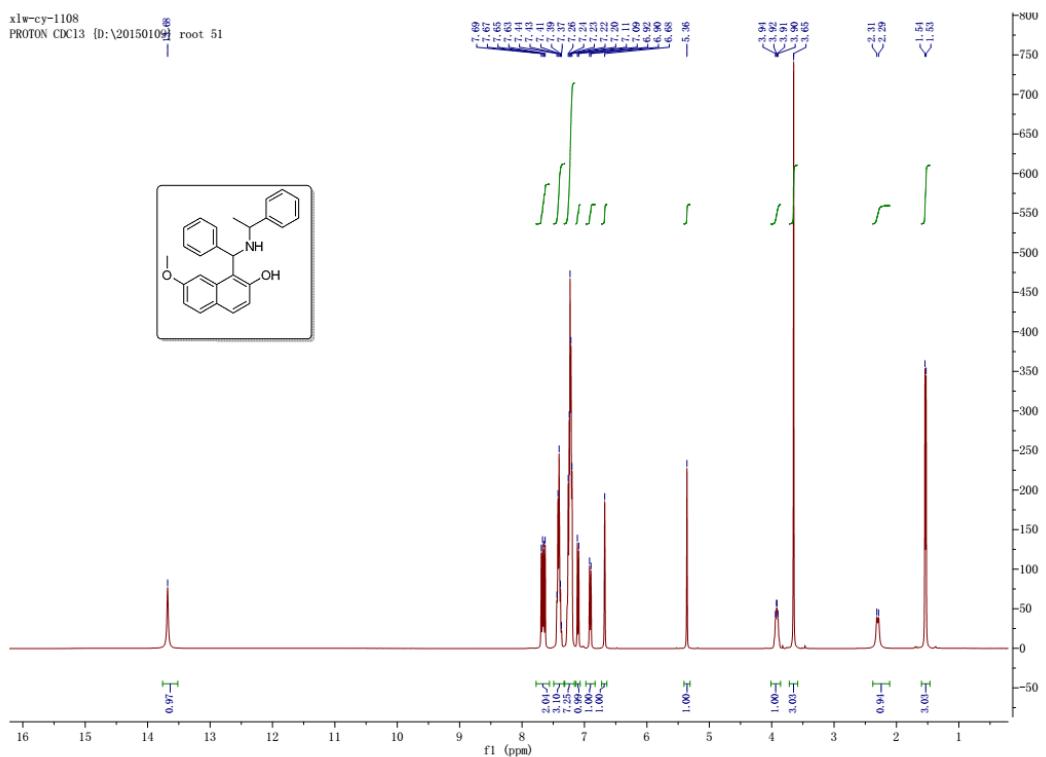
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Acquisition Date 1/15/2015 2:38:48 PM  
Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

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Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

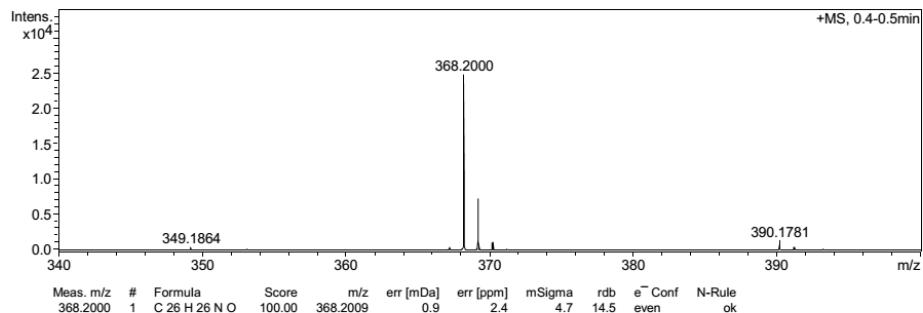
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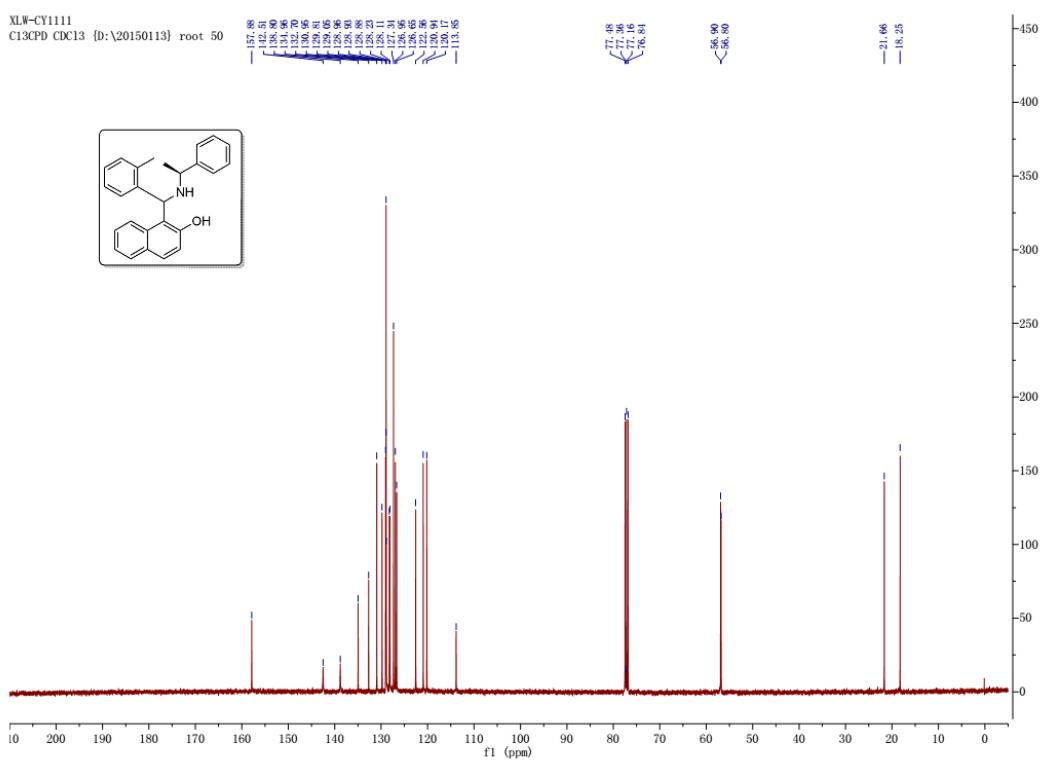
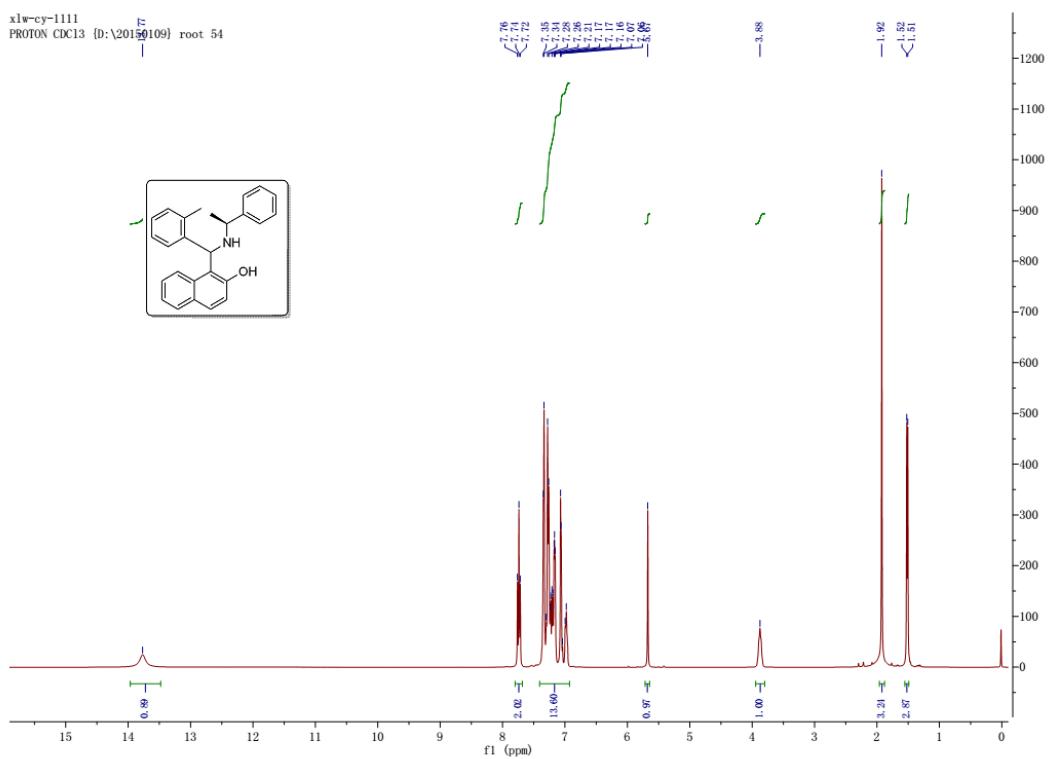
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Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

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Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

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Sample Name trz-7  
Comment

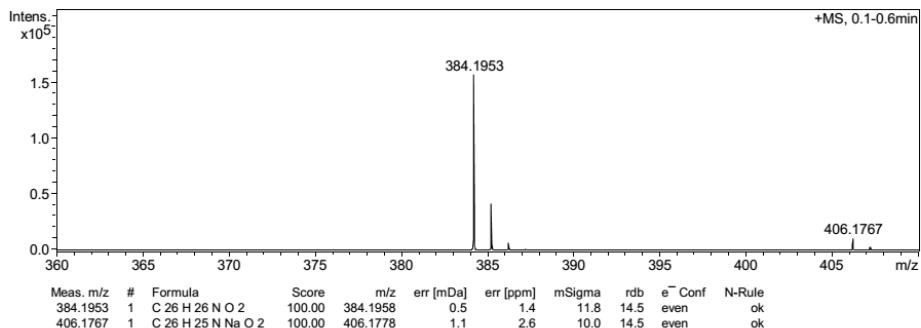
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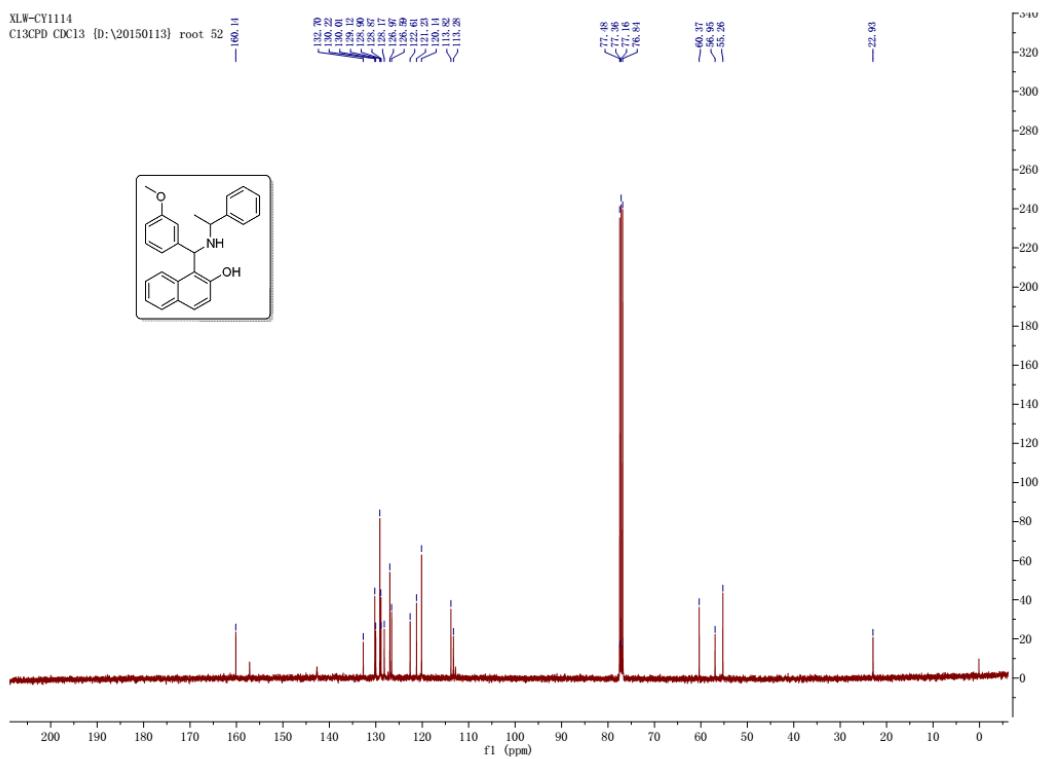
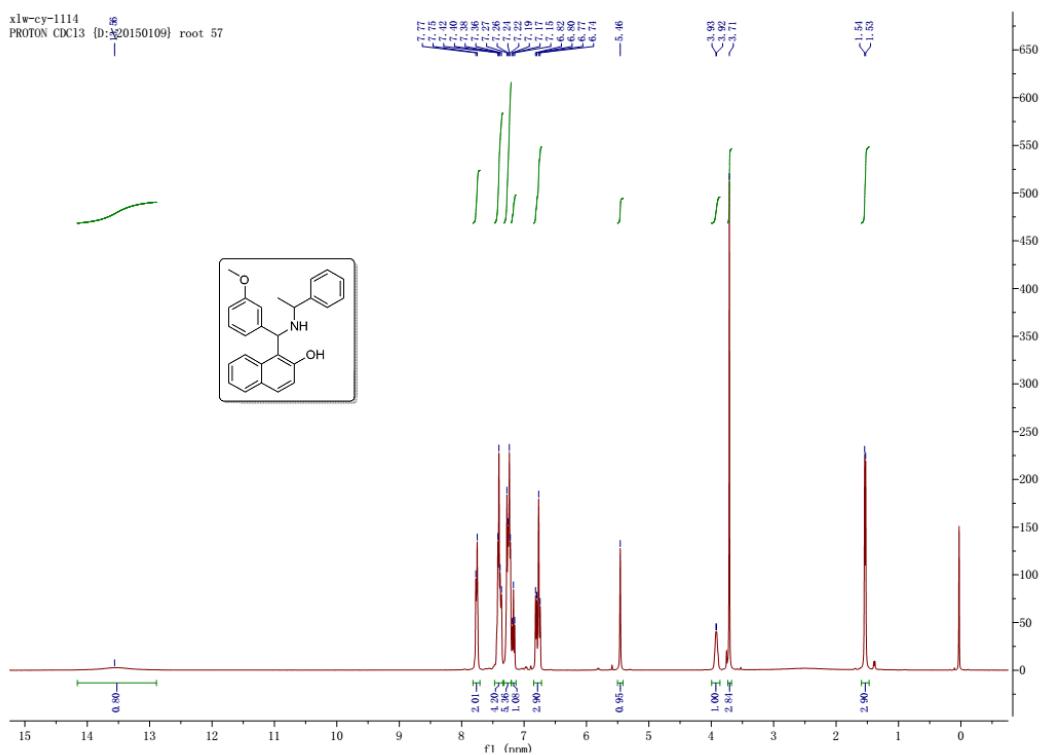
Operator Jiang

Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

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Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

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Sample Name trz-7  
Comment

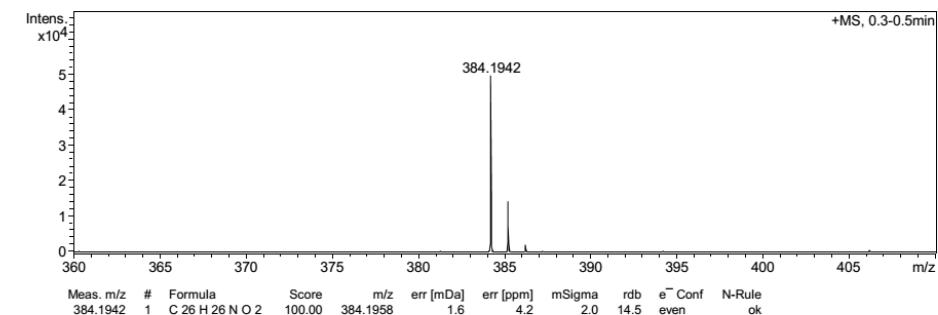
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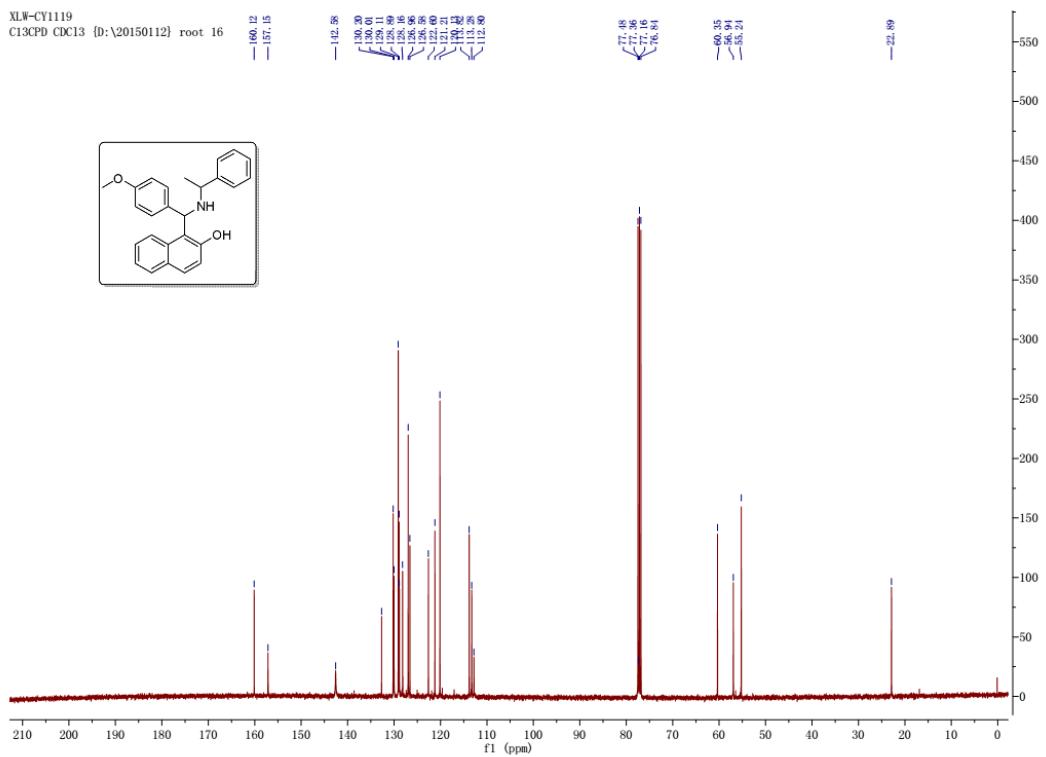
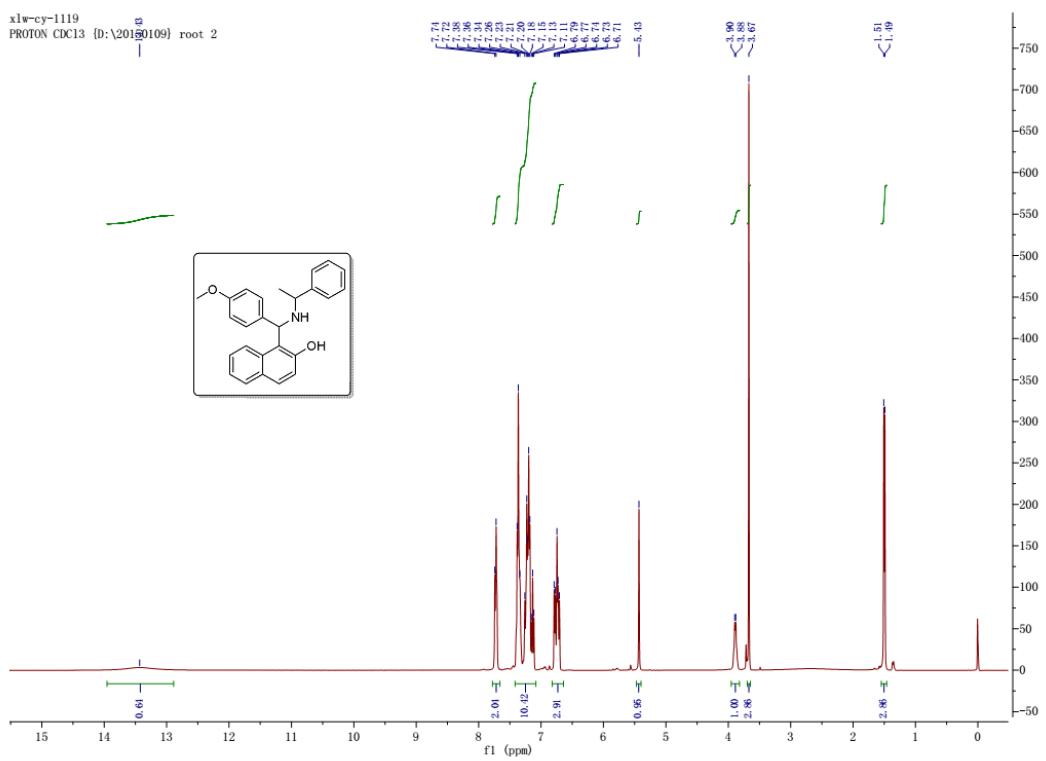
Operator Jiang

Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

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Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

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Sample Name trz-7  
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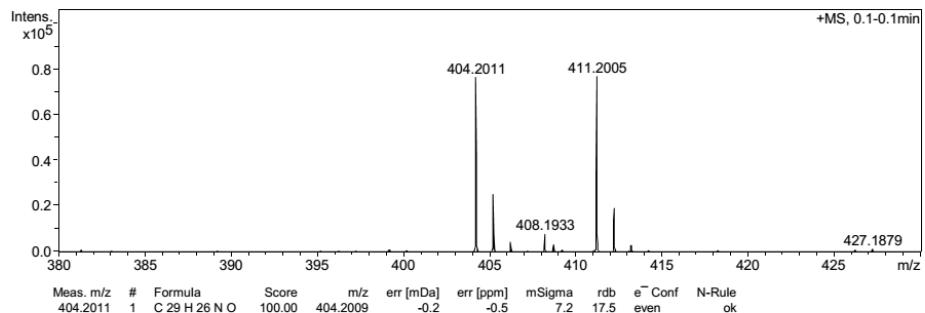
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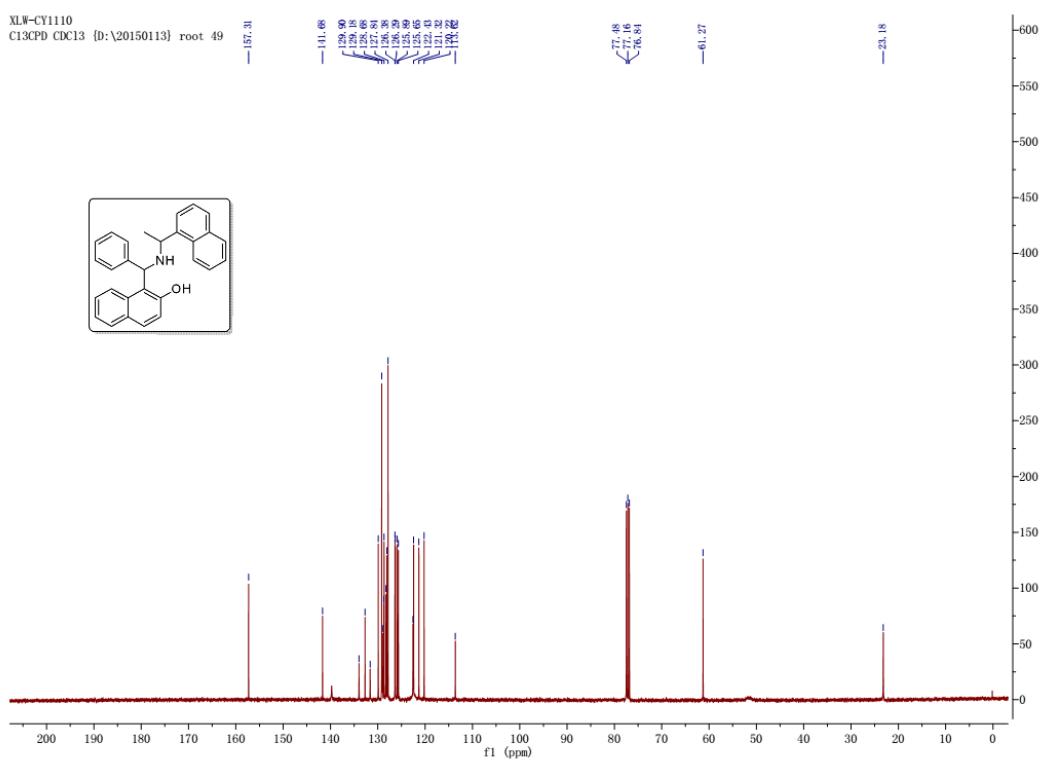
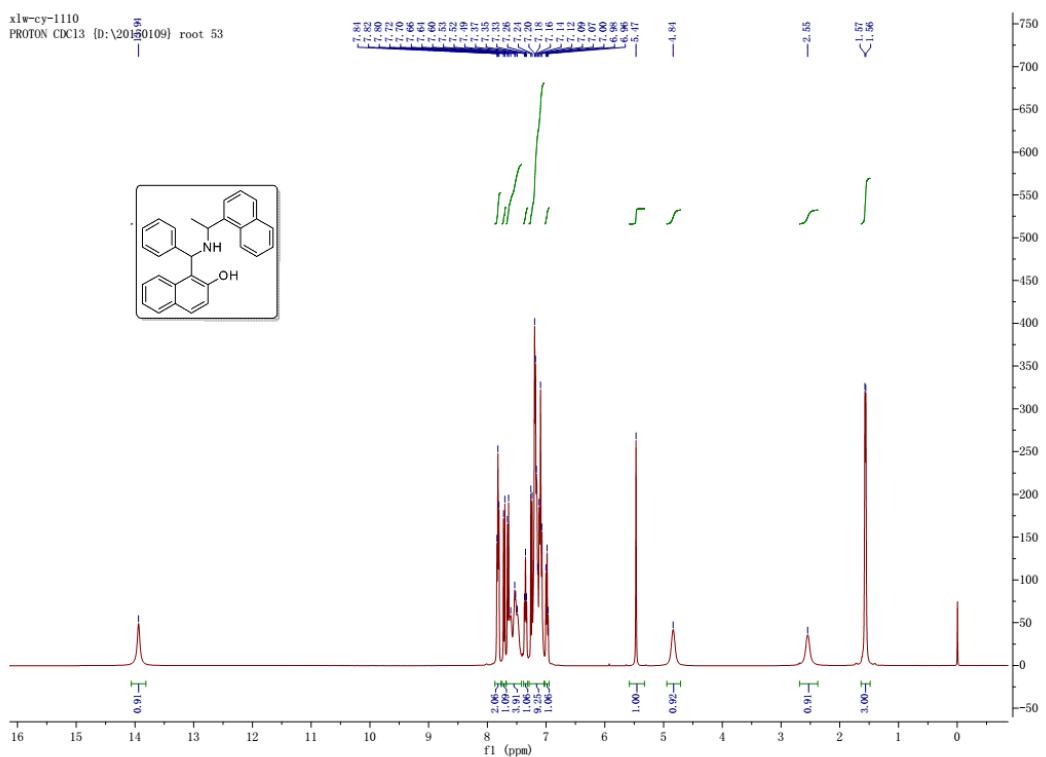
Operator Jiang

Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
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Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





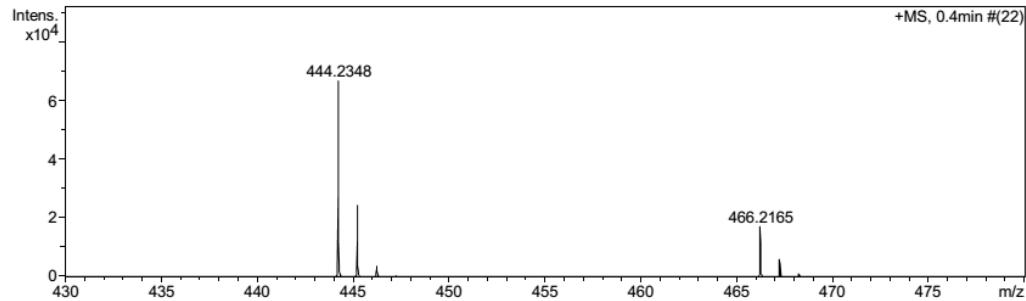
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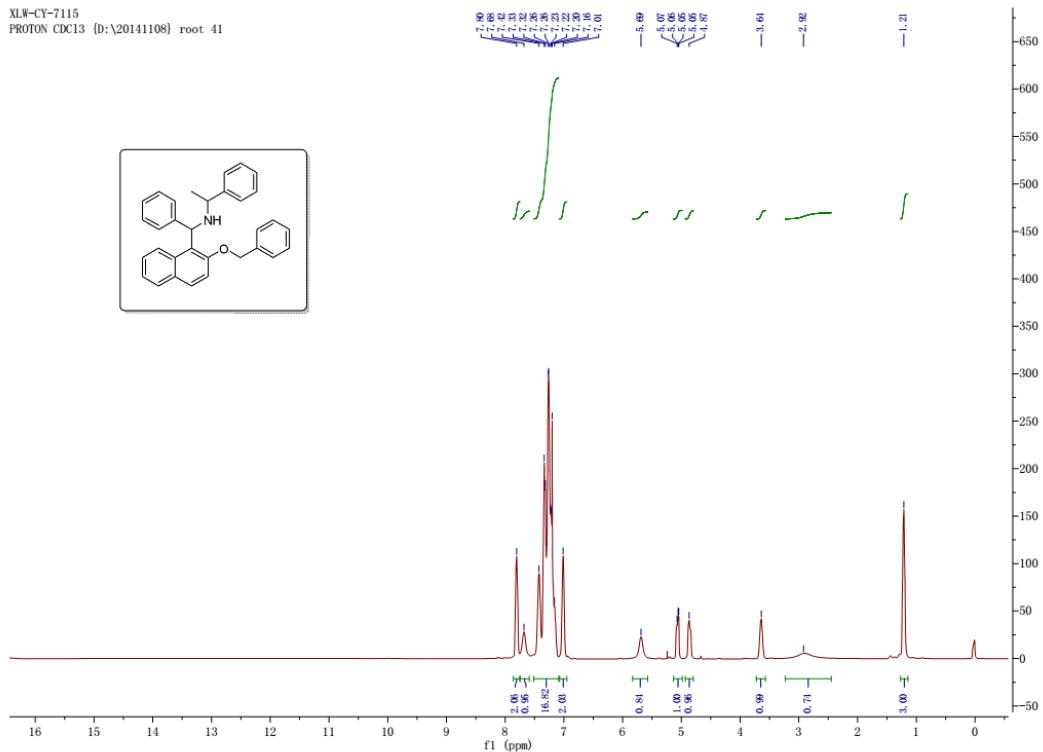
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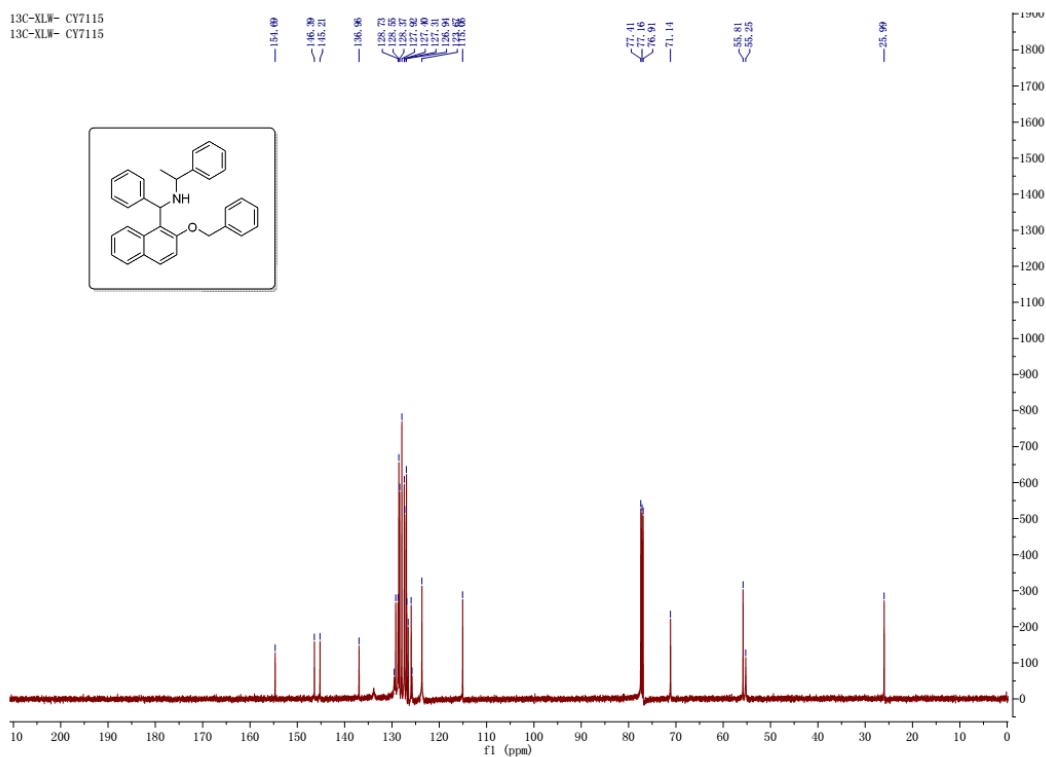
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Scan End	600 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



Meas. m/z	#	Formula	Score	m/z	err [mDa]	err [ppm]	mSigma	rdb	e <sup>-</sup> Conf	N-Rule
444.2348	1	C 32 H 30 N O	100.00	444.2322	-2.6	-5.8	8.6	18.5	even	ok
466.2165	1	C 32 H 29 N Na O	28.64	466.2141	-2.4	-5.1	1.8	18.5	even	ok

XLW-CY-7115  
PROTON CDCl<sub>3</sub> {D:\20141108} root 41





## Mass Spectrum SmartFormula Report

### Analysis Info

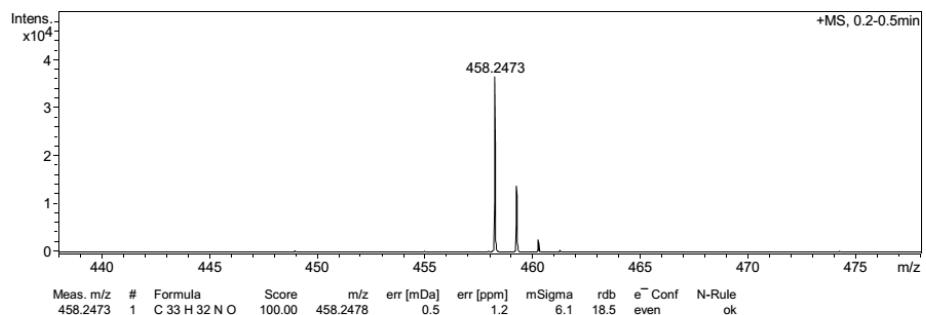
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Comment

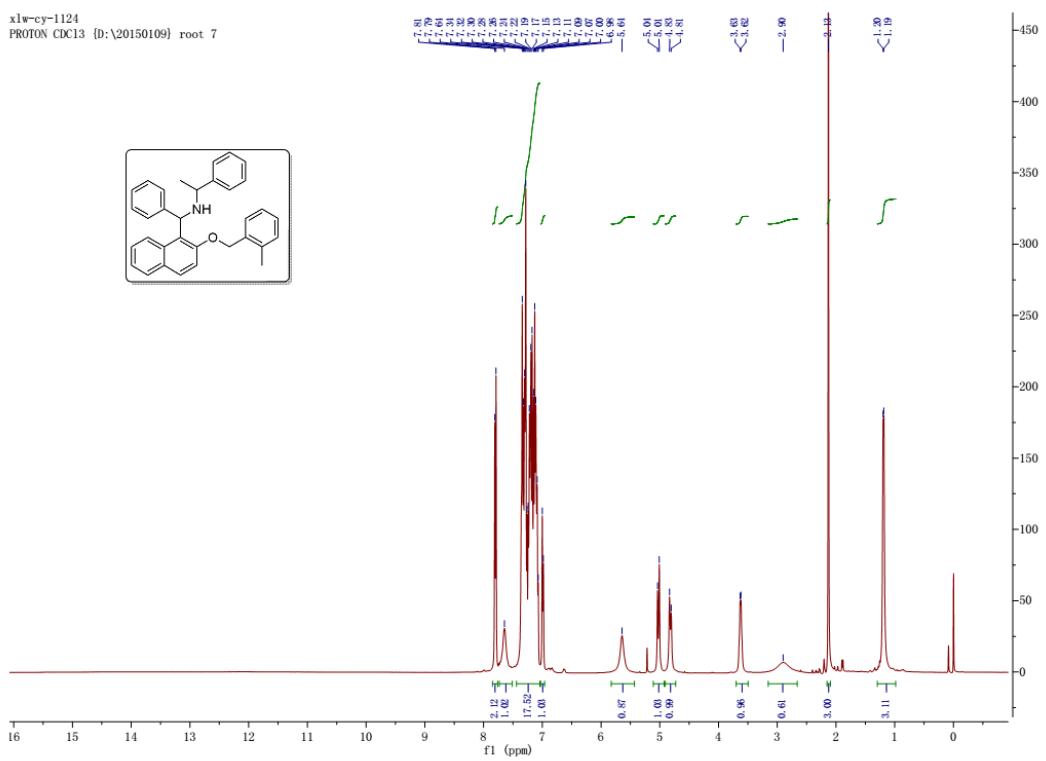
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Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

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Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

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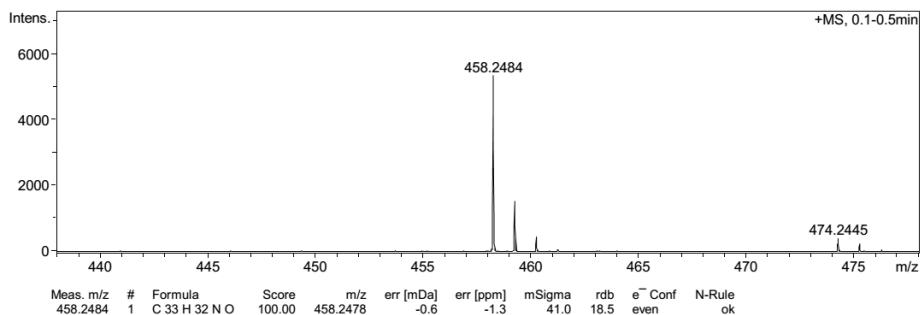
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Operator Jiang

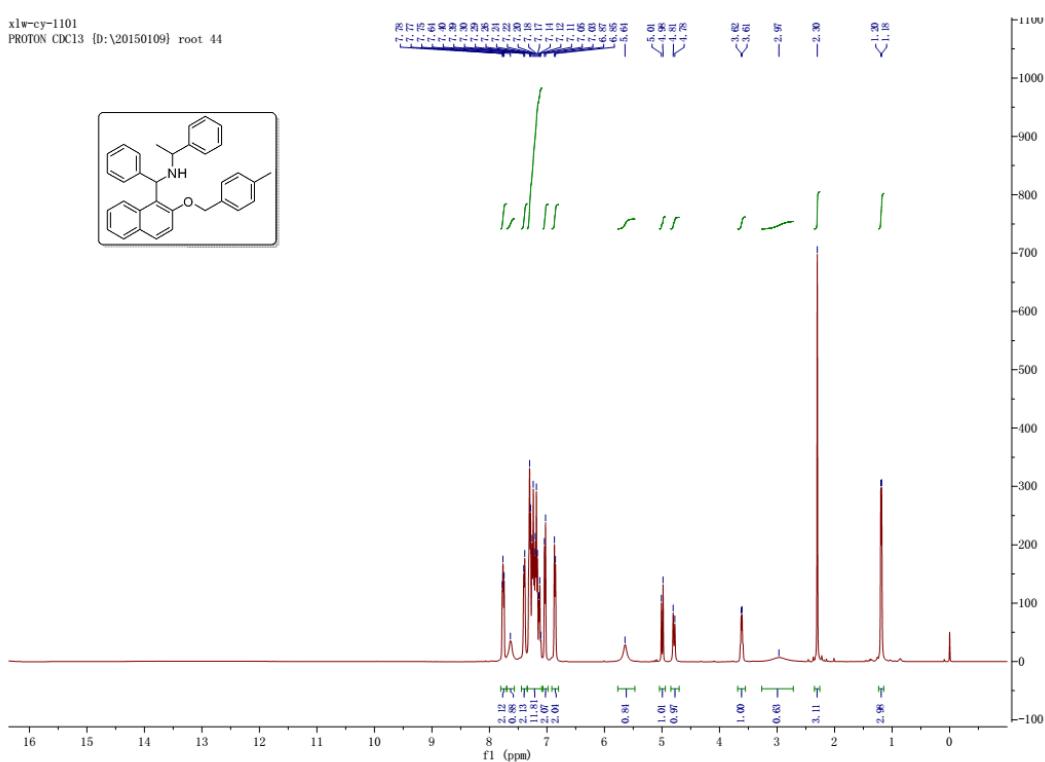
Instrument / Ser# micrOTOF-Q II 10324

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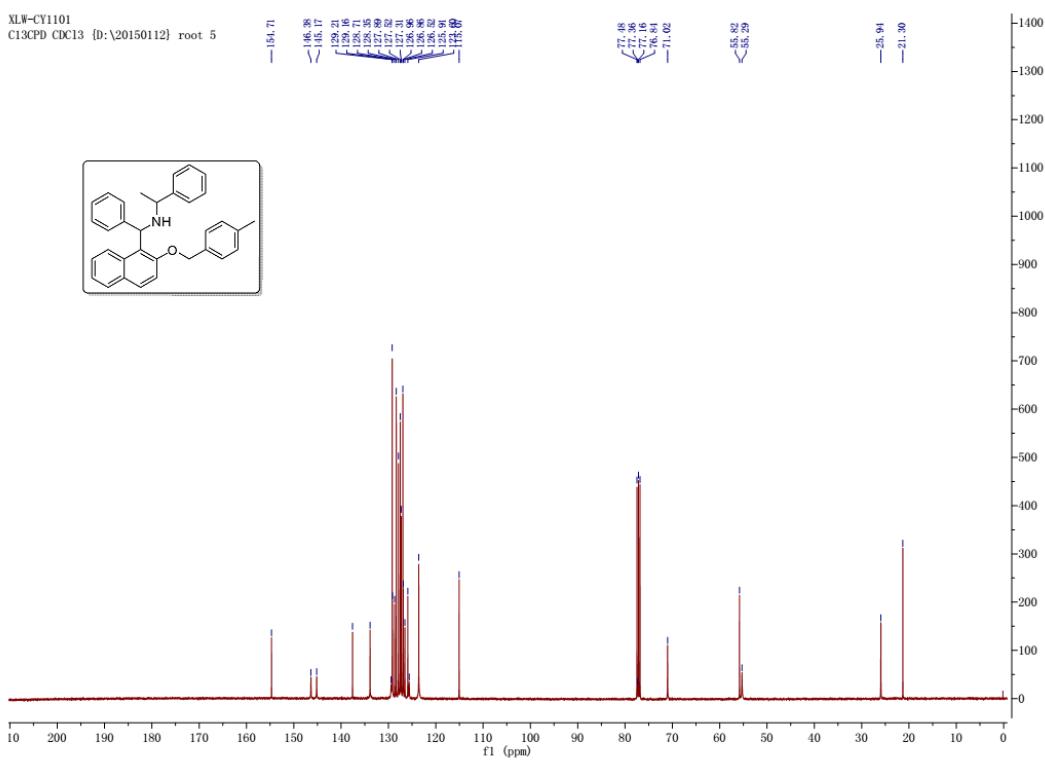
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Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



xlw-cy-1101  
PROTON CDCl<sub>3</sub> {D:\20150109} root 44



XIW-CY1101  
C13CPD CDCl<sub>3</sub> {D:\20150112} root 5



## Mass Spectrum SmartFormula Report

### Analysis Info

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Comment

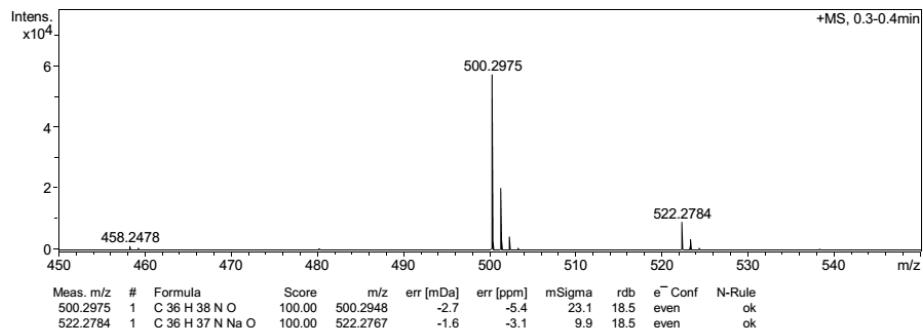
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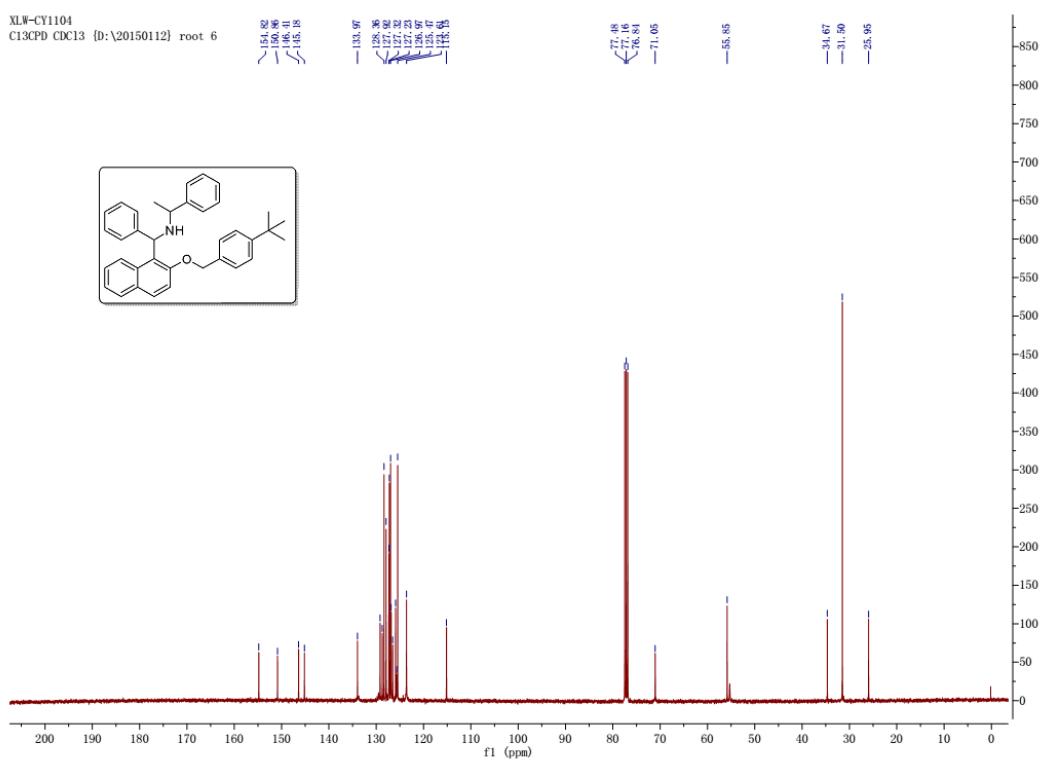
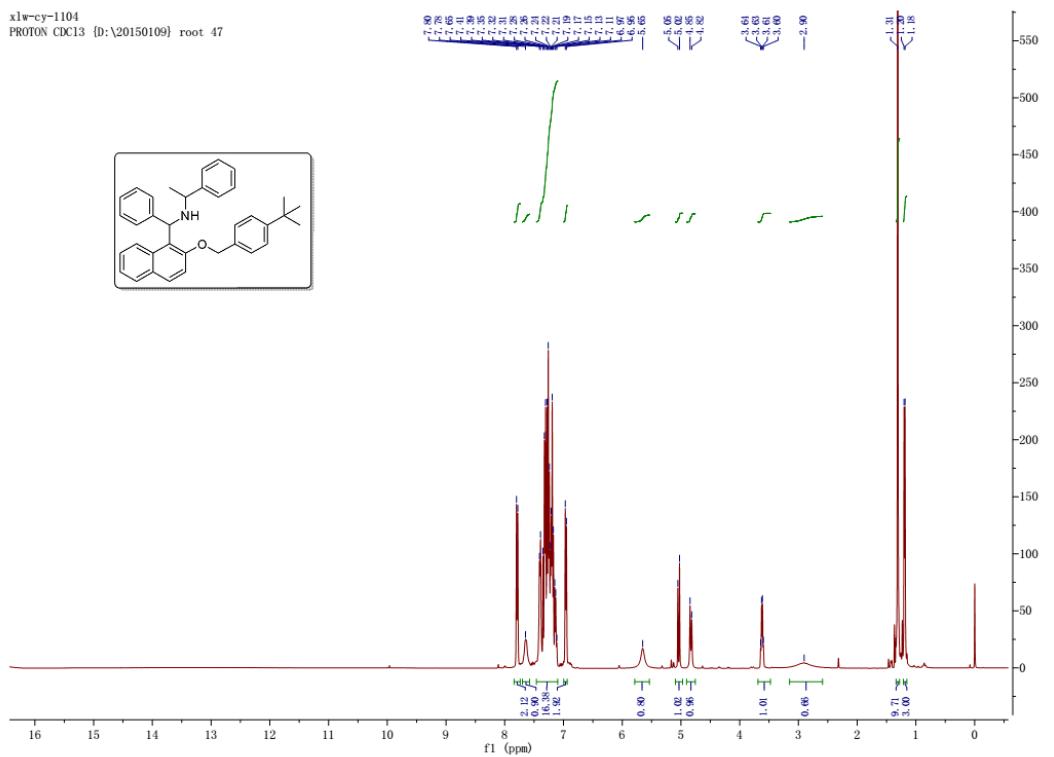
Operator Jiang

Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

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Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

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Sample Name trz-7  
Comment

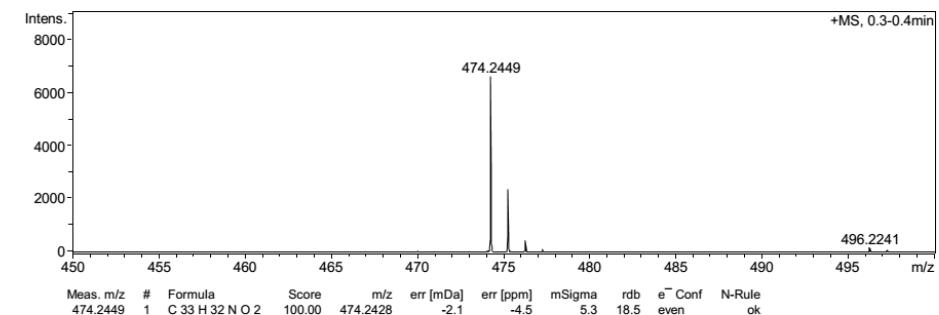
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Operator Jiang

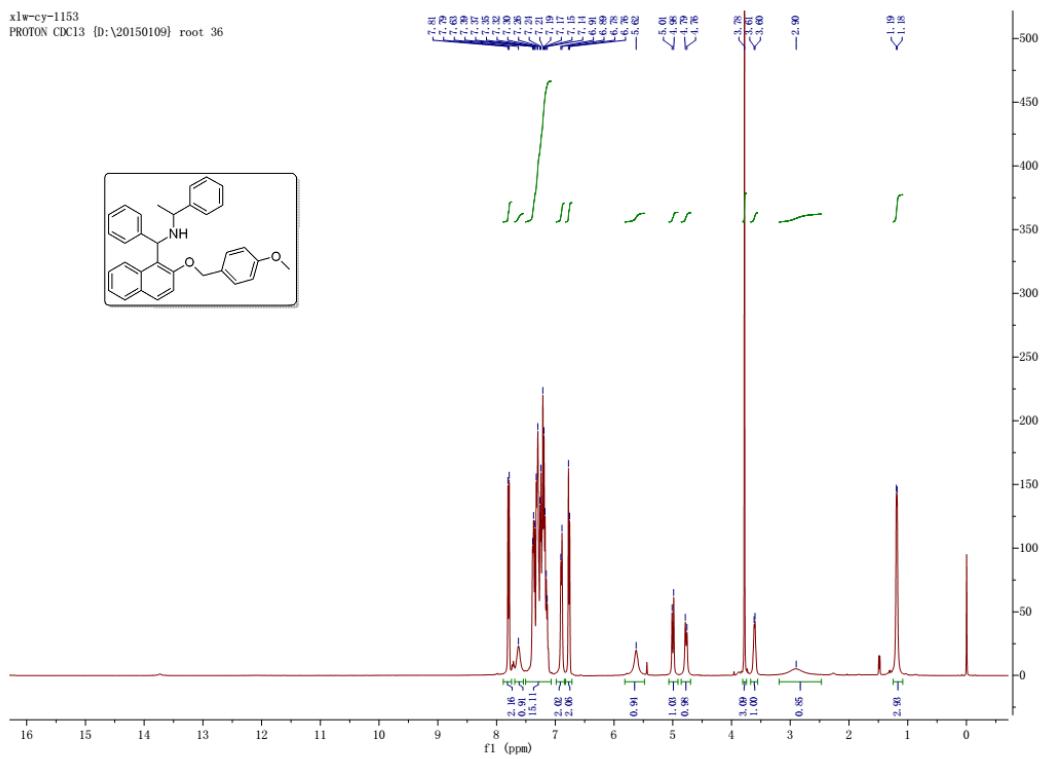
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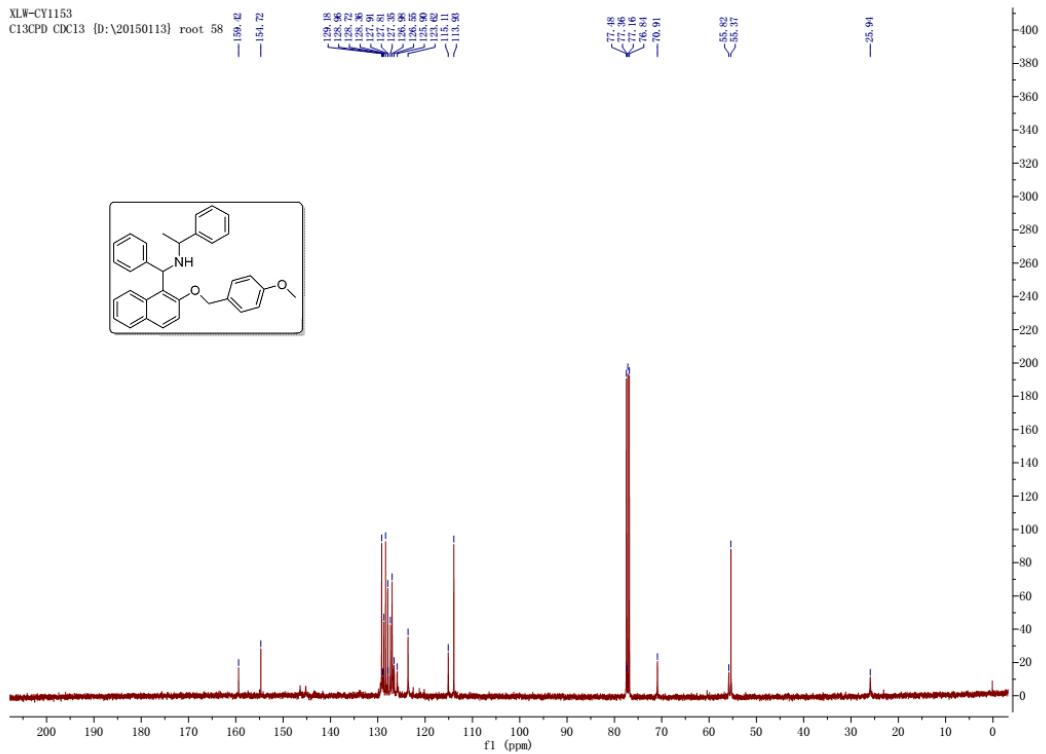
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Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



xlw-cy-1153  
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XLW-CY1153  
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## Mass Spectrum SmartFormula Report

### Analysis Info

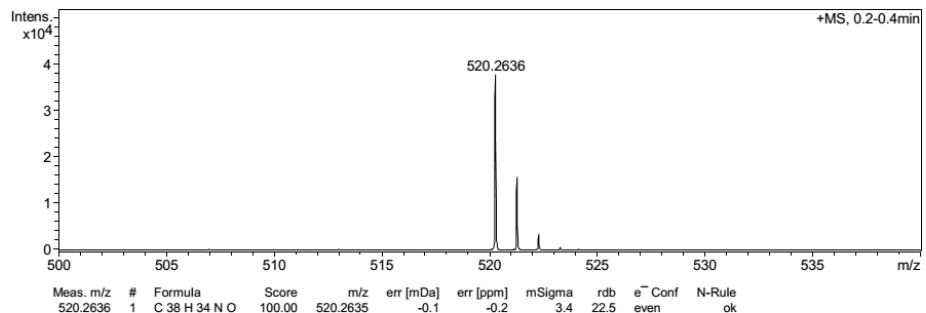
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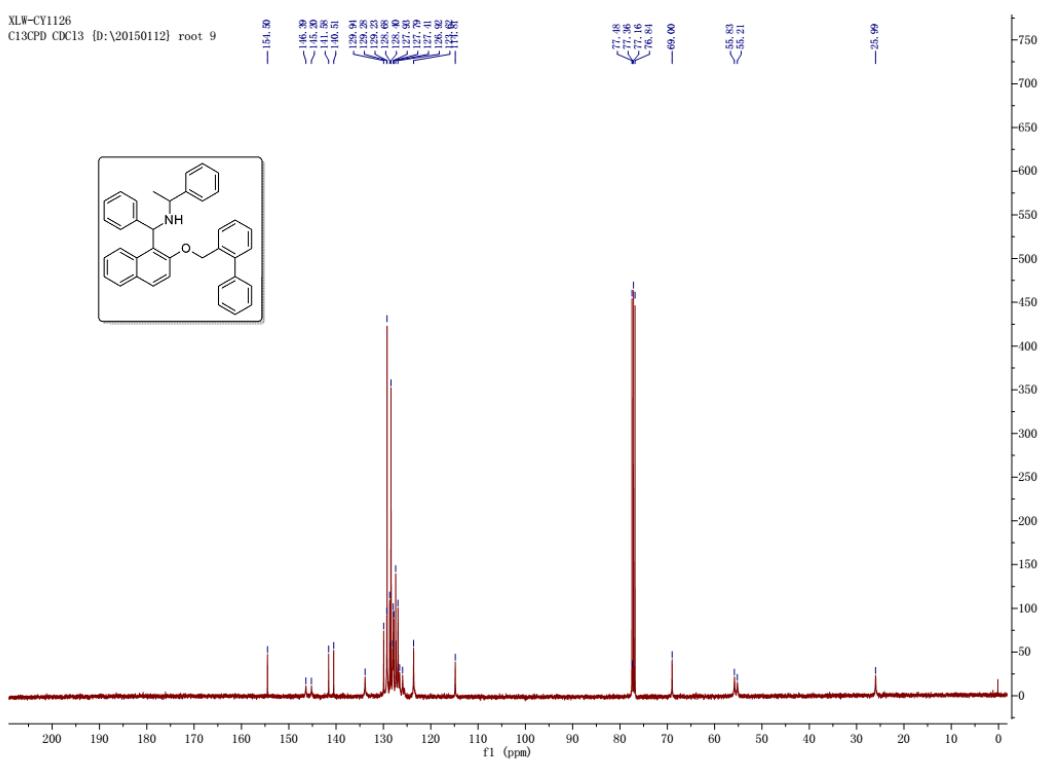
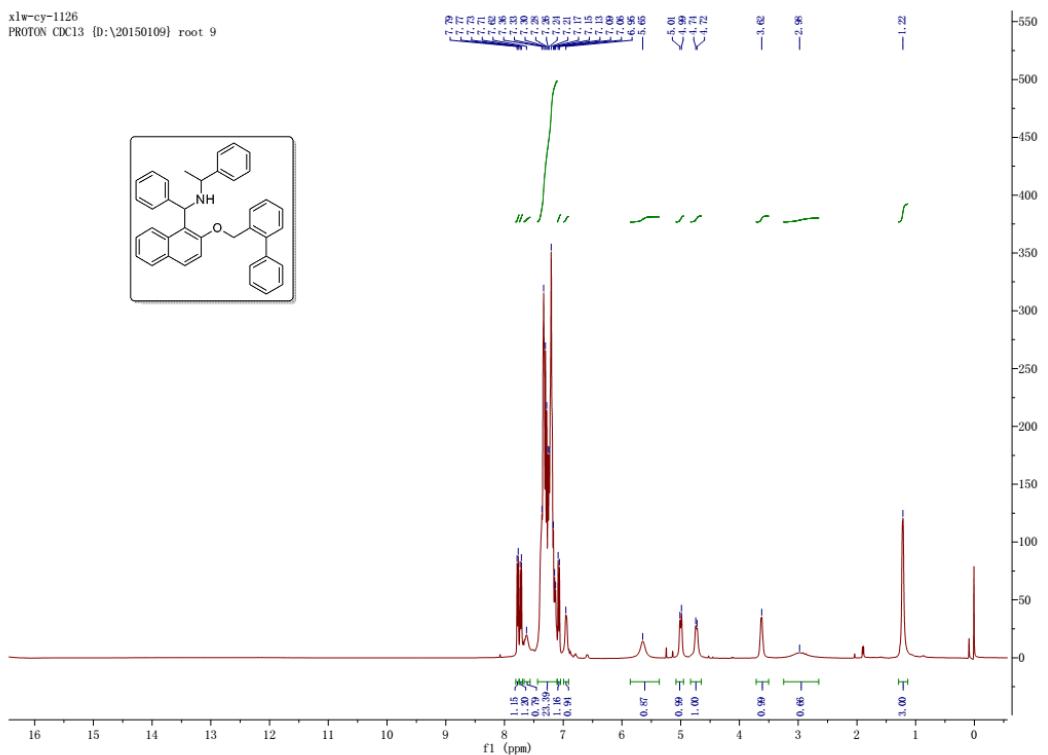
Acquisition Date 1/15/2015 2:04:27 PM

Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

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Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

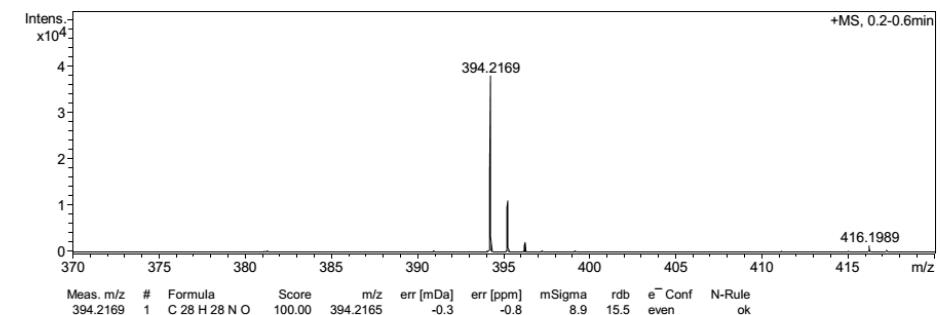
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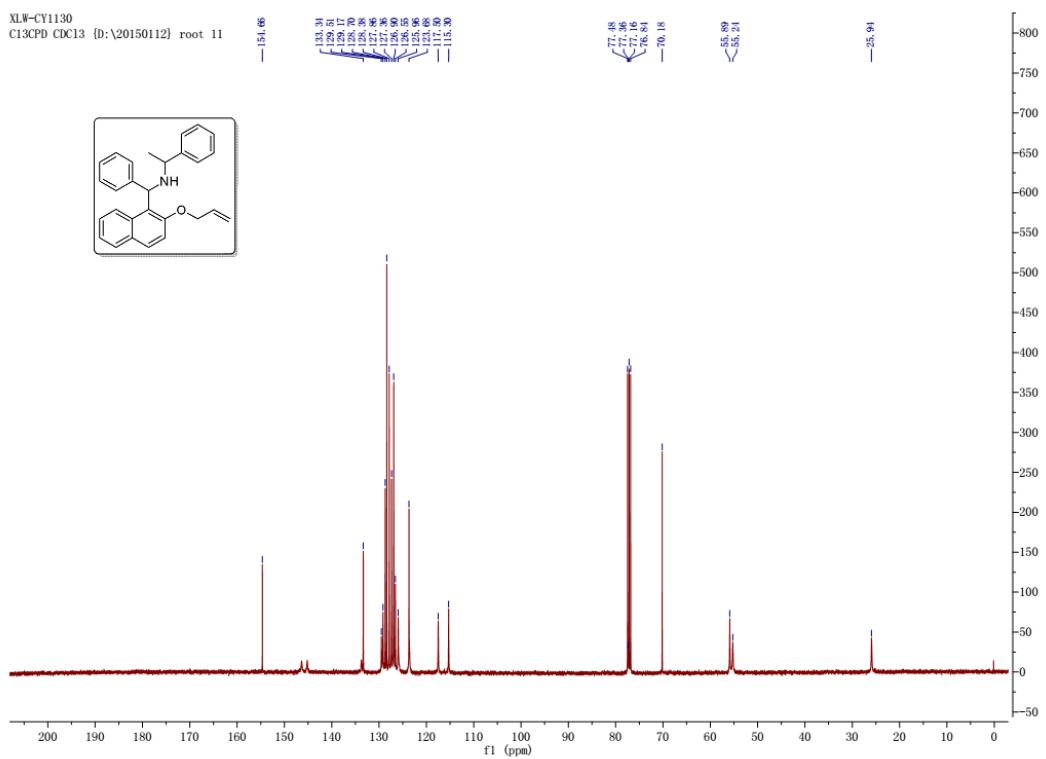
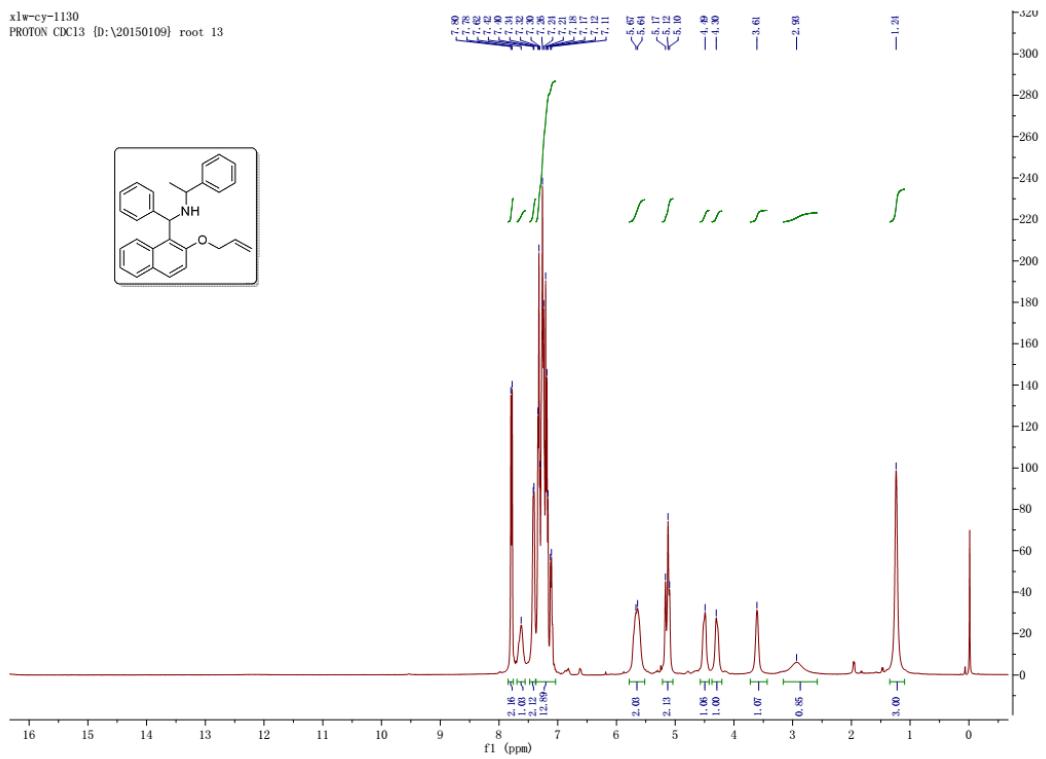
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Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

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Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
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## Mass Spectrum SmartFormula Report

### Analysis Info

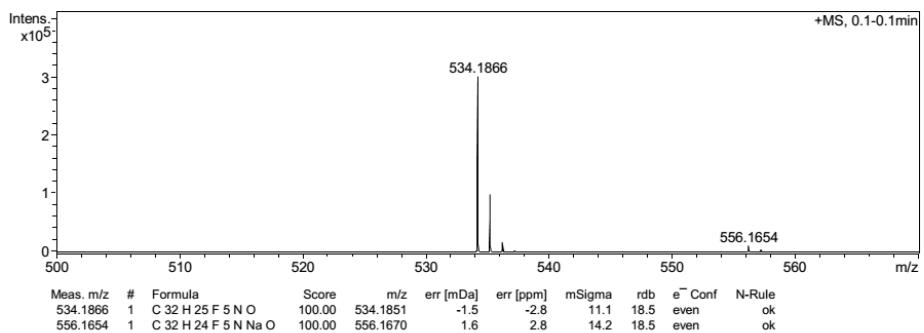
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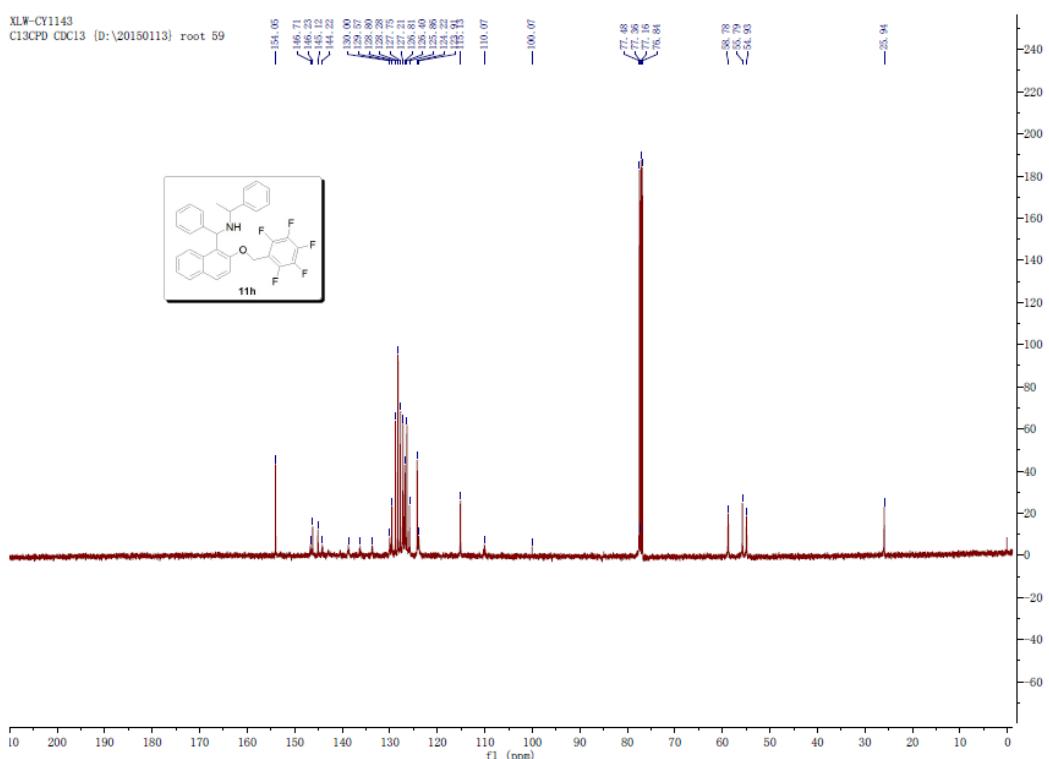
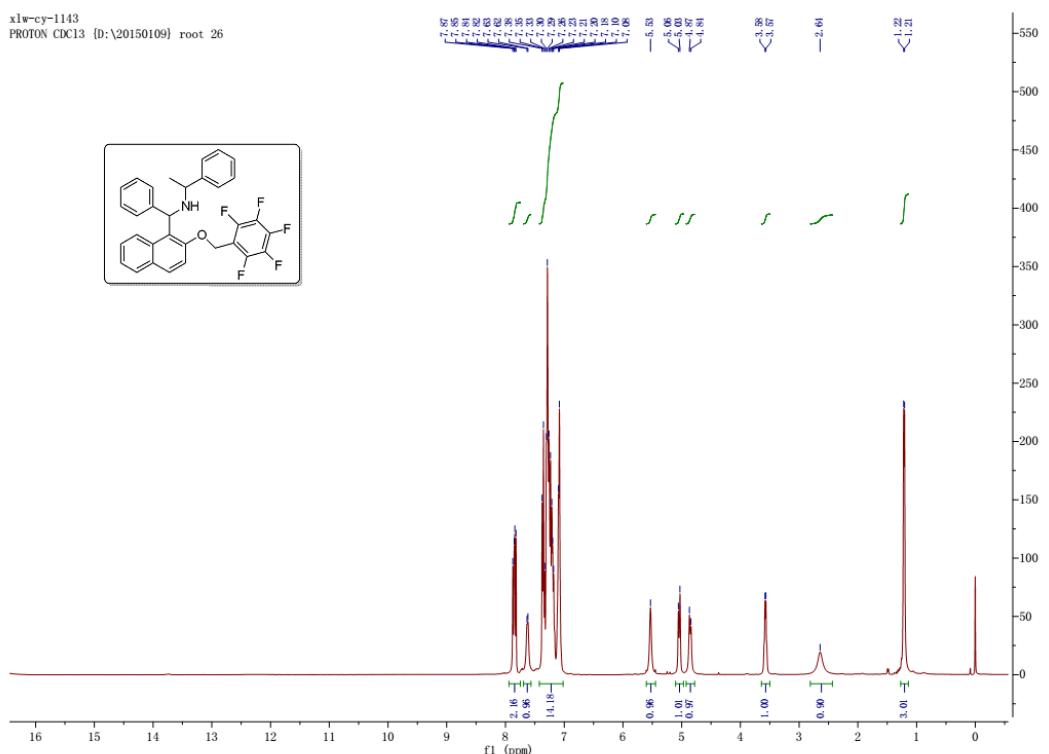
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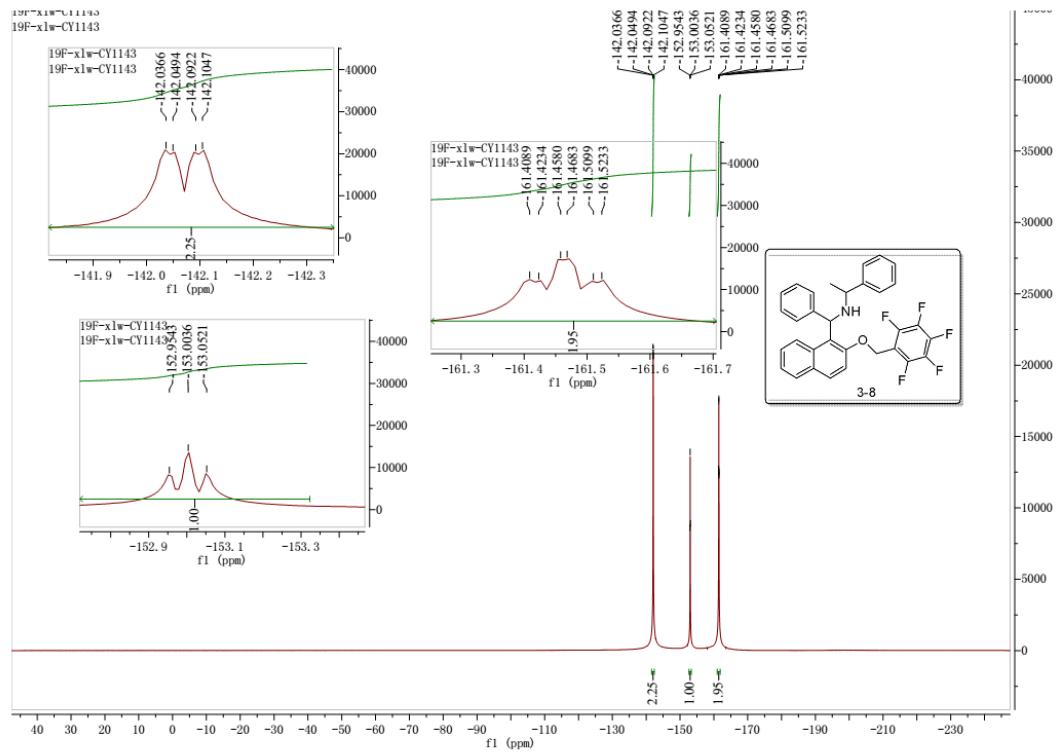
Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

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Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	600 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source







## Mass Spectrum SmartFormula Report

### Analysis Info

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Comment

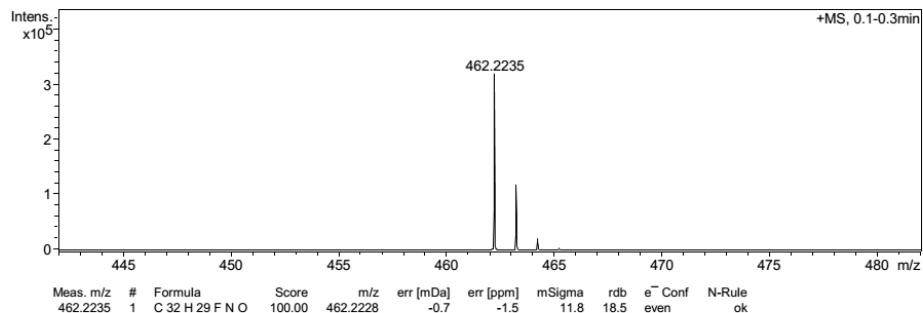
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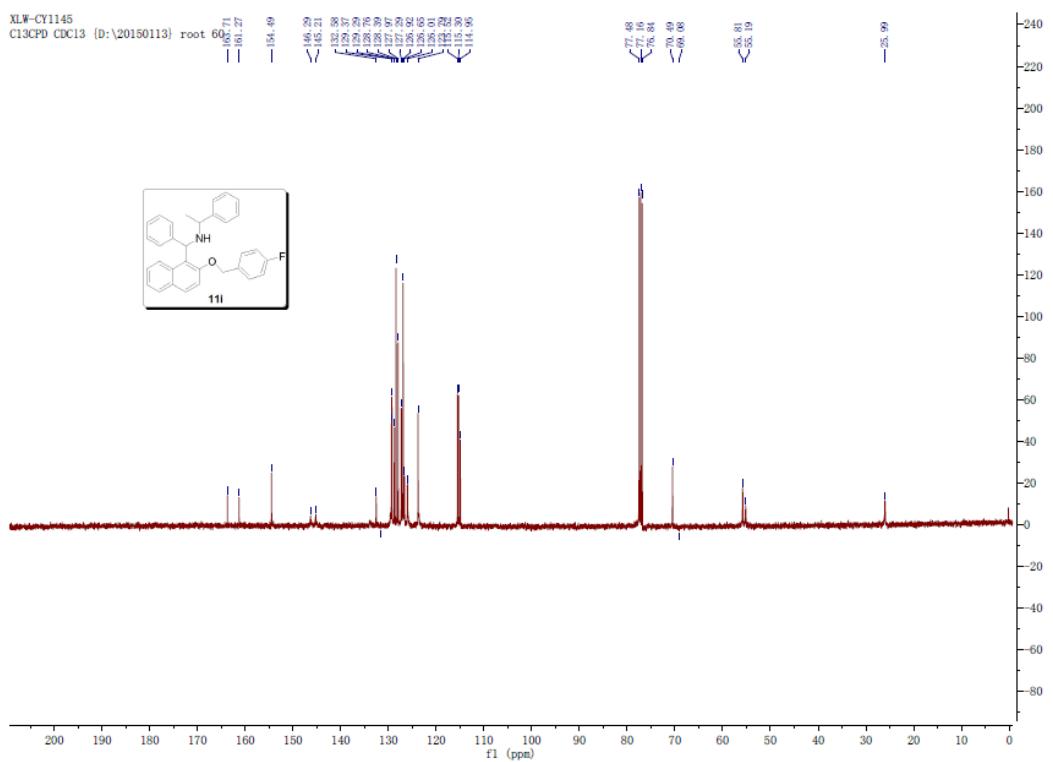
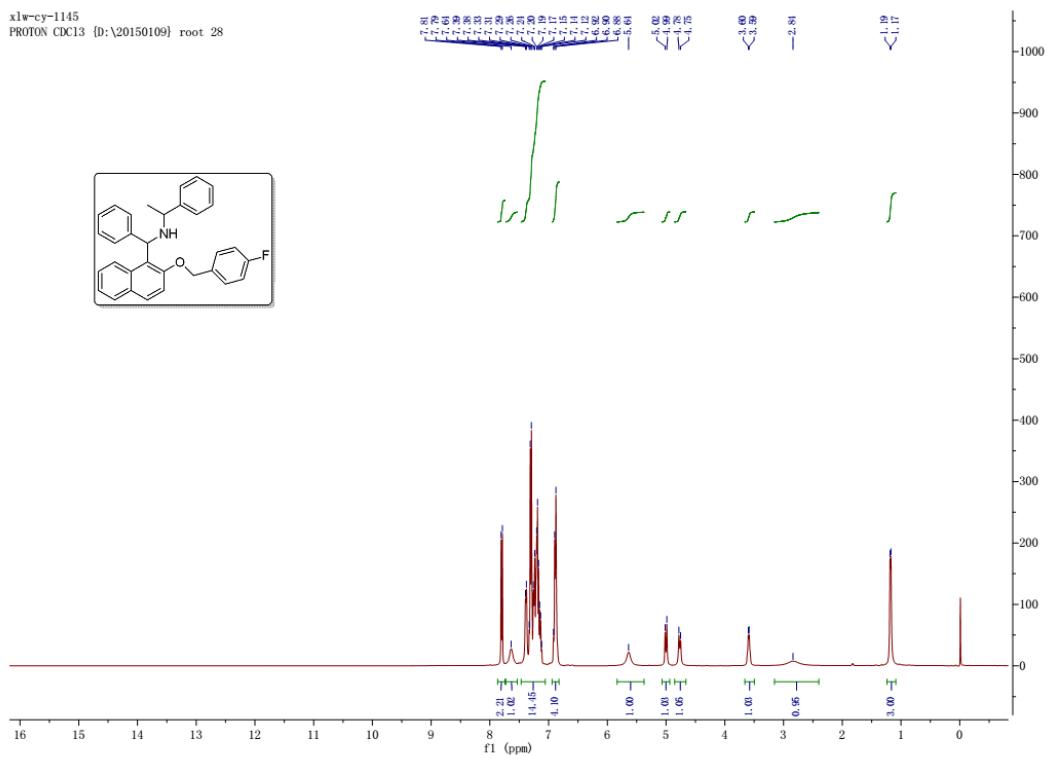
Operator Jiang

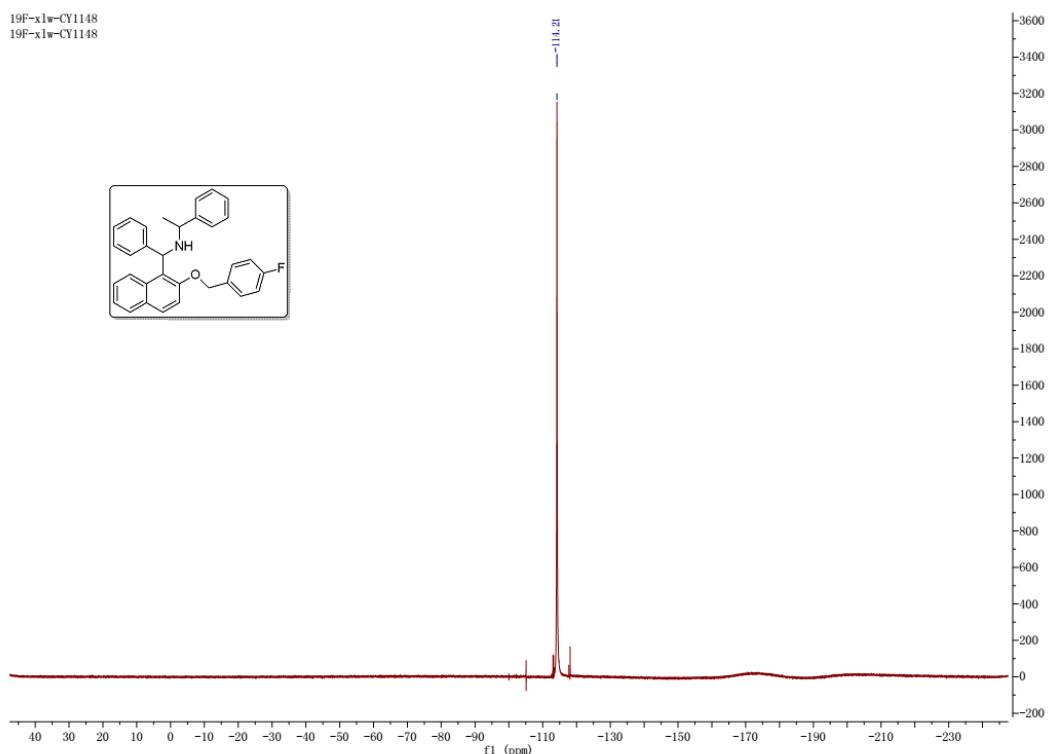
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	600 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source







## Mass Spectrum SmartFormula Report

### Analysis Info

Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150120-cy119-9\_01.d  
Method tune\_100-550\_pos150120.m  
Sample Name  
Comment

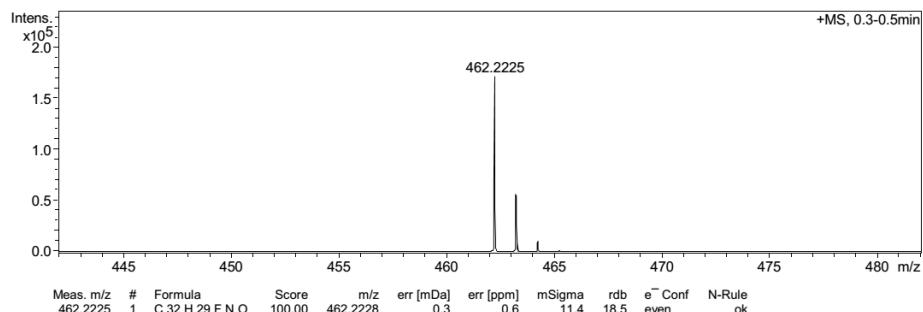
Acquisition Date 1/20/2015 3:03:21 PM

Operator Jiang

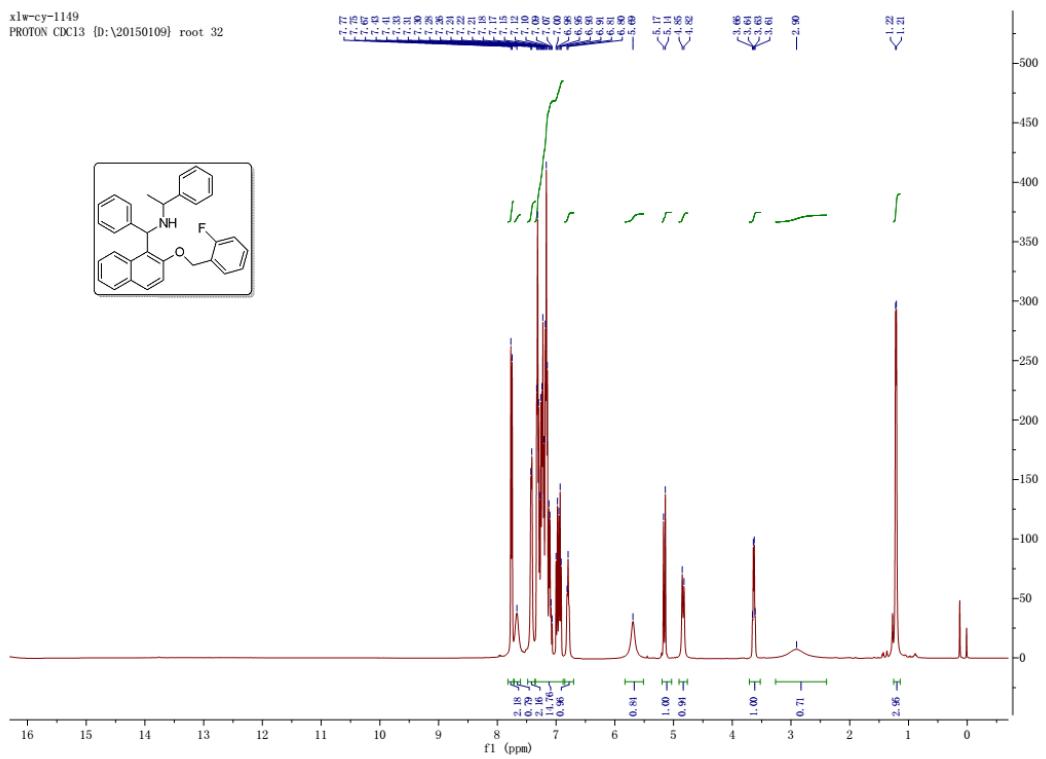
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

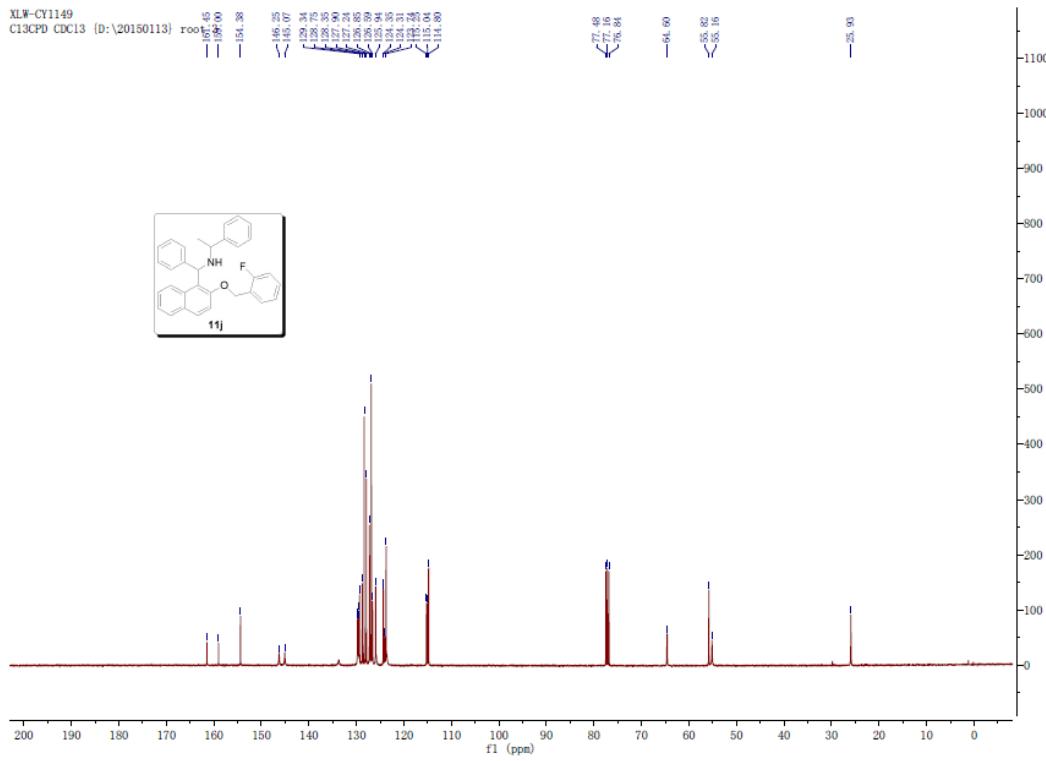
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source

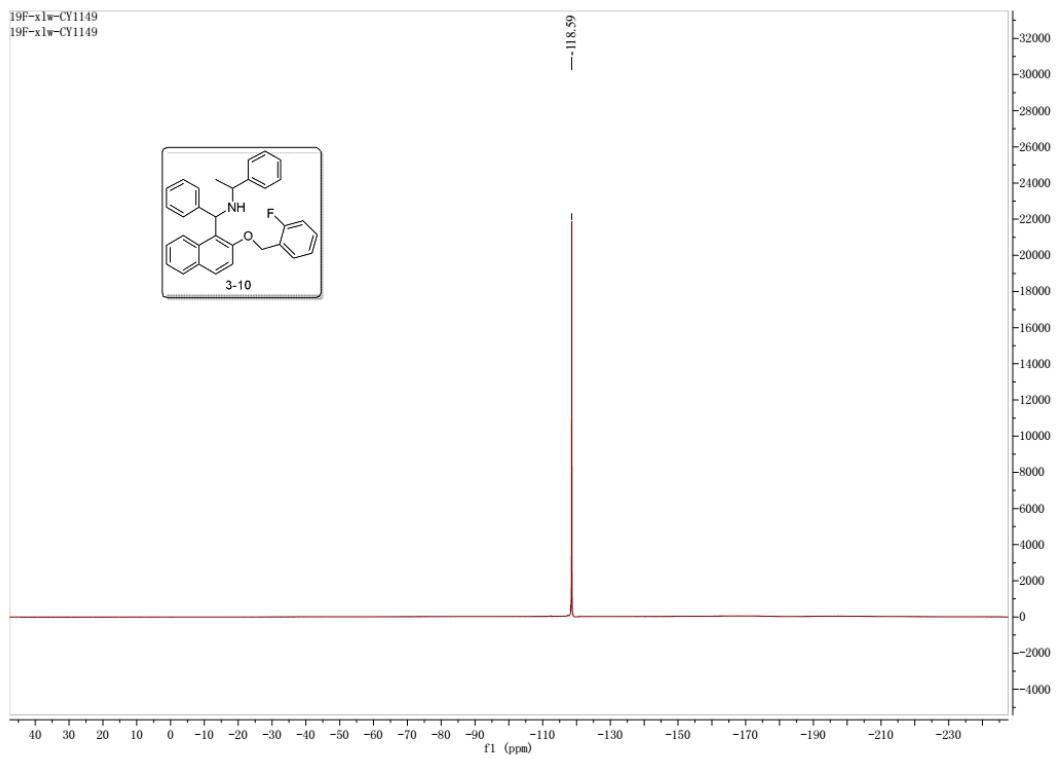


xlw-cy-1149  
PROTON CDCl<sub>3</sub> {D:\20150109} root 32



XLW-CY1149  
C13CPD CDCl<sub>3</sub> {D:\20150113} root 32





## Mass Spectrum SmartFormula Report

### Analysis Info

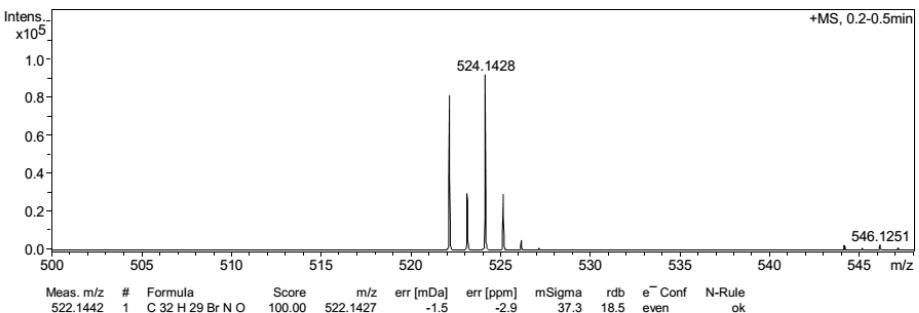
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150115-wcy-1151\_01.d  
Method tune\_100-550\_pos150115.m  
Sample Name trz-7  
Comment

Acquisition Date 1/15/2015 1:52:55 PM

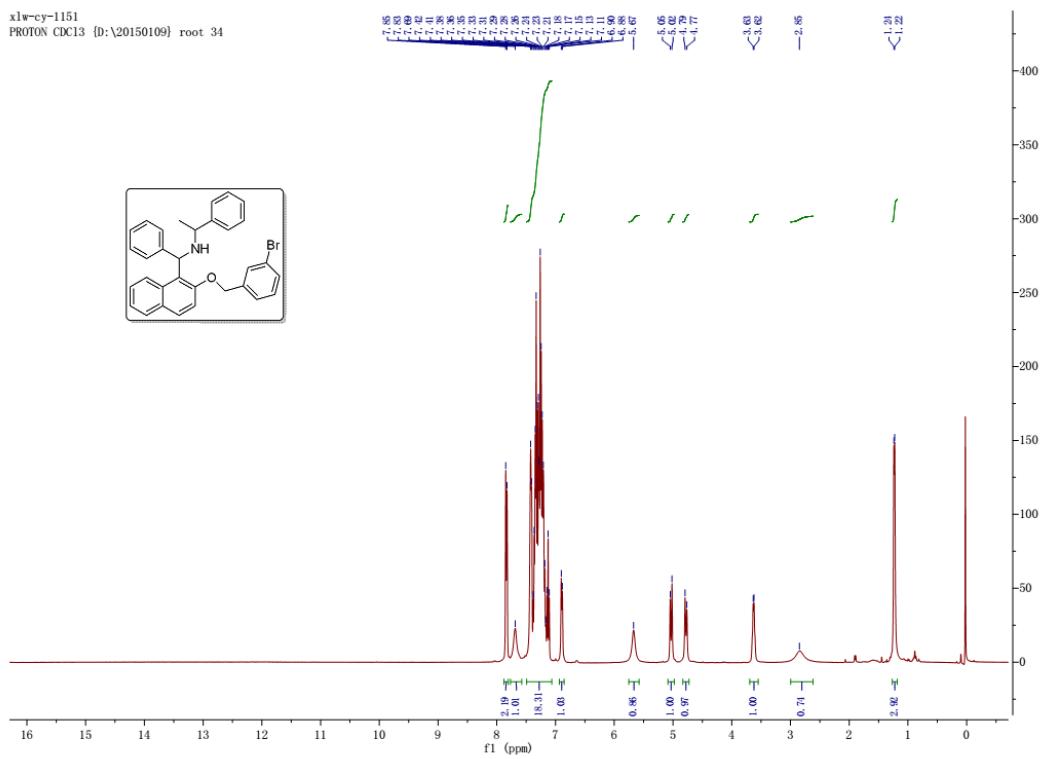
Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

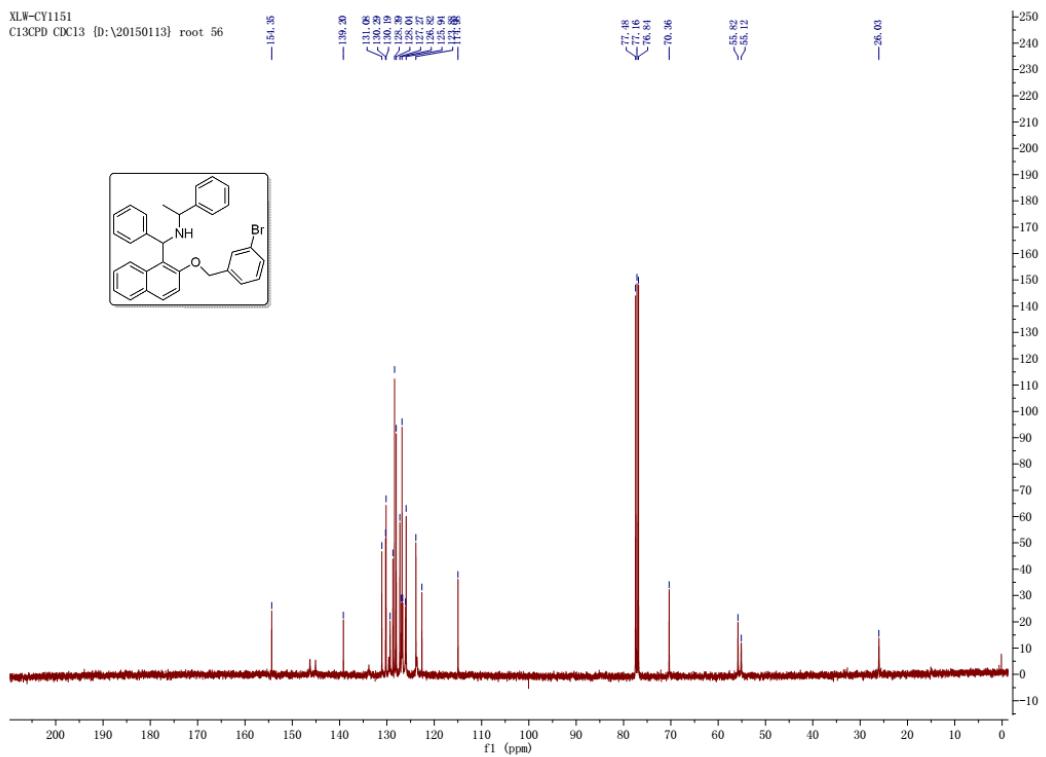
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



xlw-cy-1151  
PROTON CDCl<sub>3</sub> {D:\20150109} root 34



XLW-CY1151  
C13CPD CDCl<sub>3</sub> {D:\20150113} root 56



## Mass Spectrum SmartFormula Report

### Analysis Info

Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150115-wcy-1150\_01.d  
Method tune\_100-550\_pos150115.m  
Sample Name trz-7  
Comment

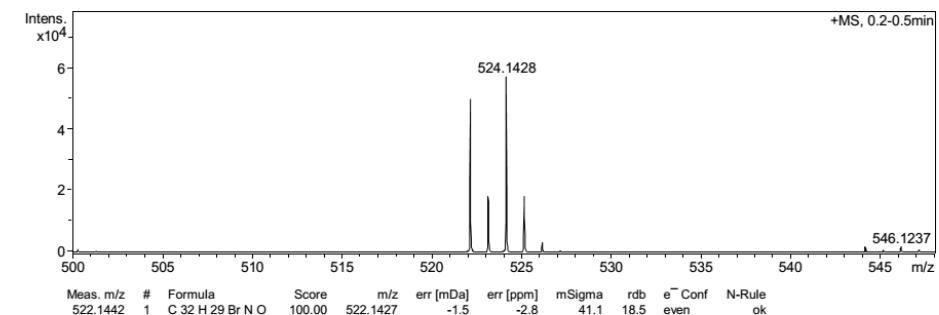
Acquisition Date 1/15/2015 1:58:44 PM

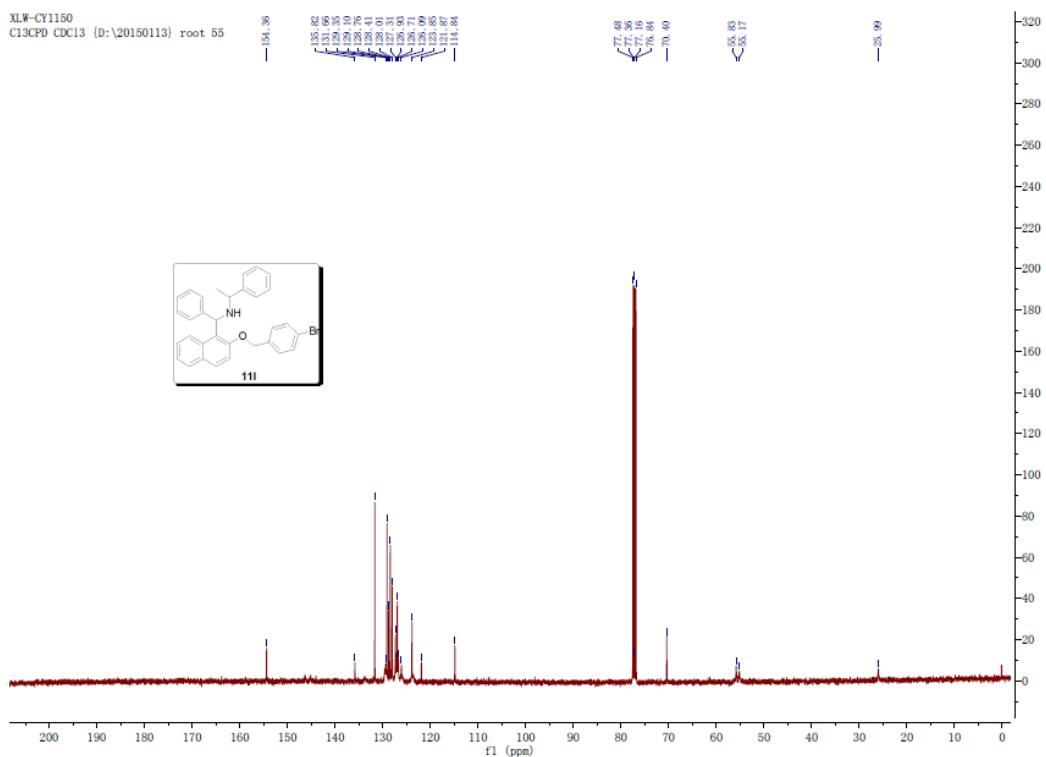
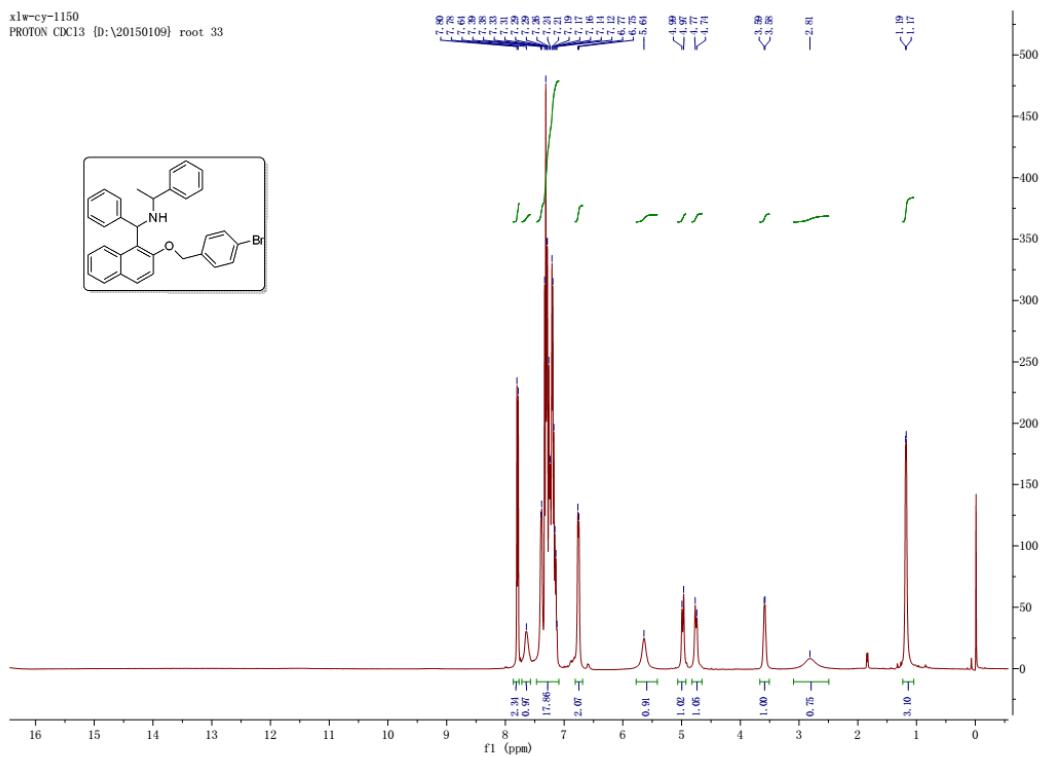
Operator Jiang

Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

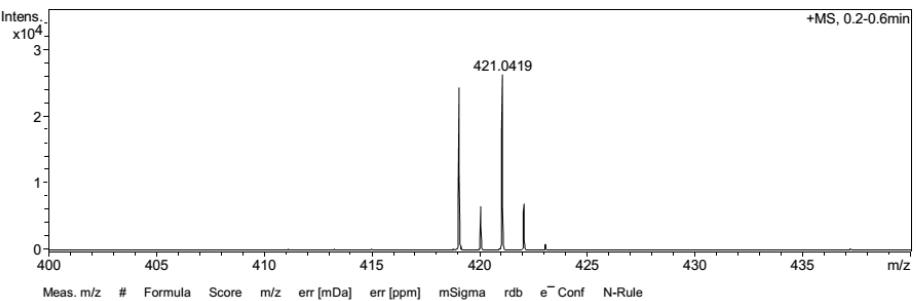
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150115-wcy-1152\_01.d  
Method tune\_100-550\_pos150115.m  
Sample Name trz-7  
Comment

Acquisition Date 1/15/2015 2:52:41 PM

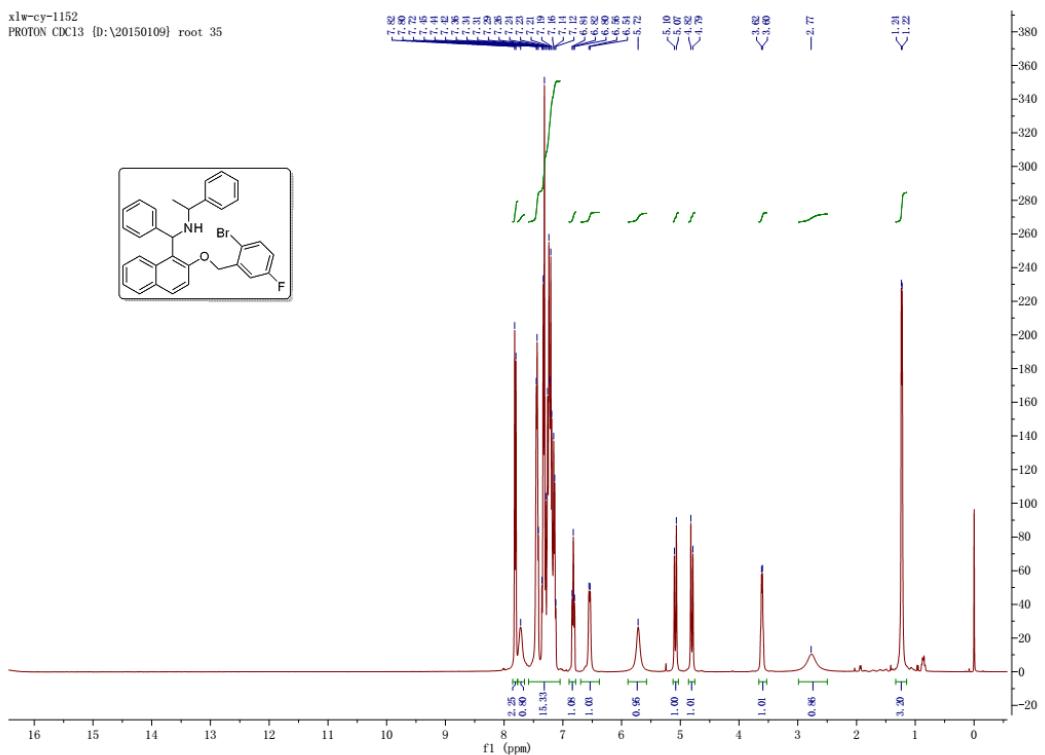
Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

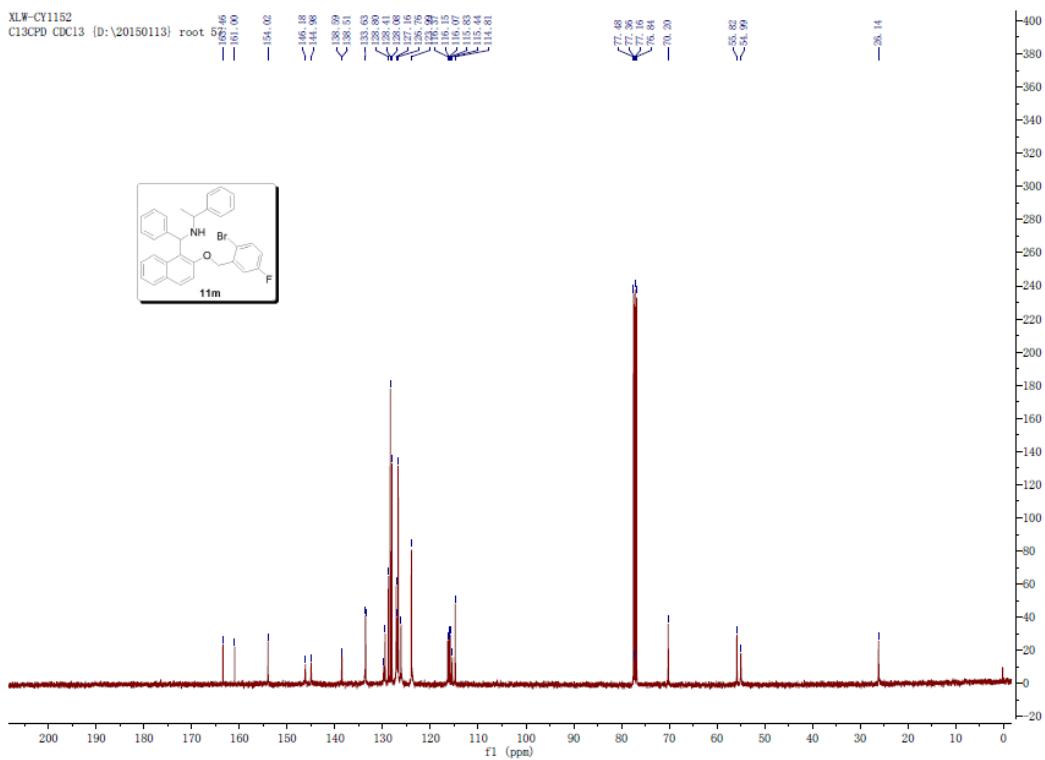
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	600 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source

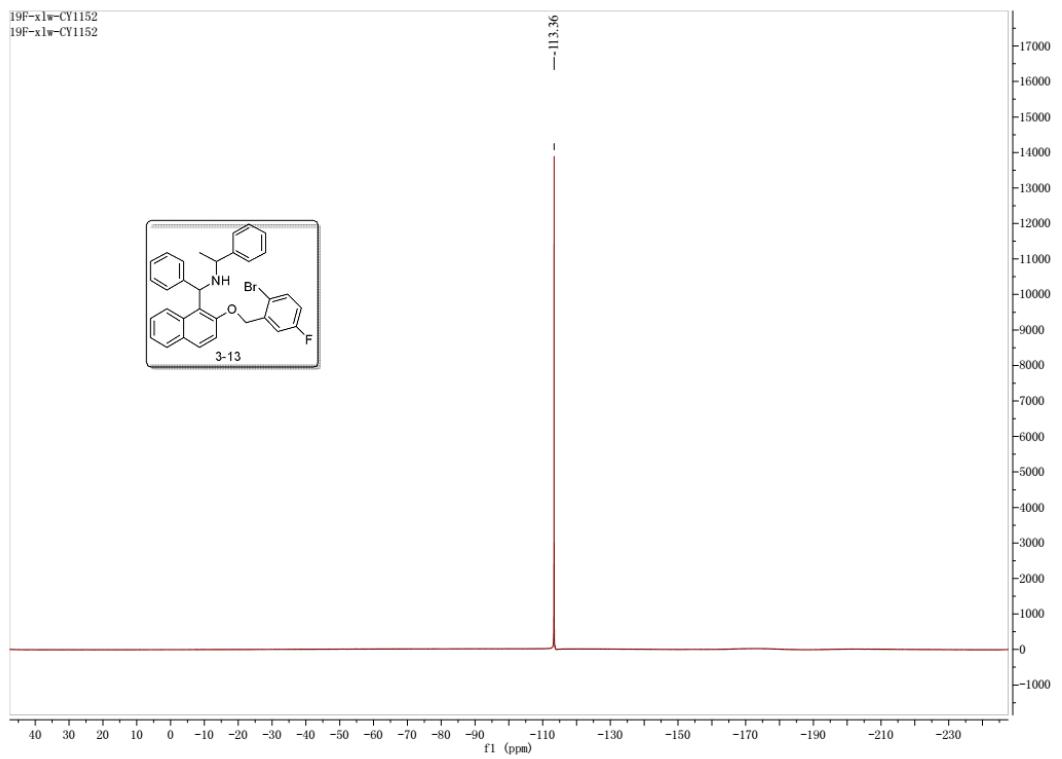


XLW-CY-1152  
PROTON CDC13 {D:\20150109} root 35



XLW-CY1152  
C13CPD CDC13 {D:\20150113} root 35





## Mass Spectrum SmartFormula Report

### Analysis Info

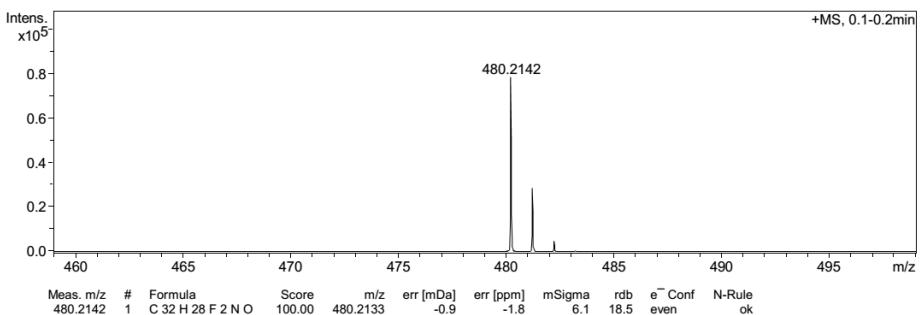
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150120-cy119-7\_01.d  
Method tune\_100-550\_pos150120.m  
Sample Name  
Comment

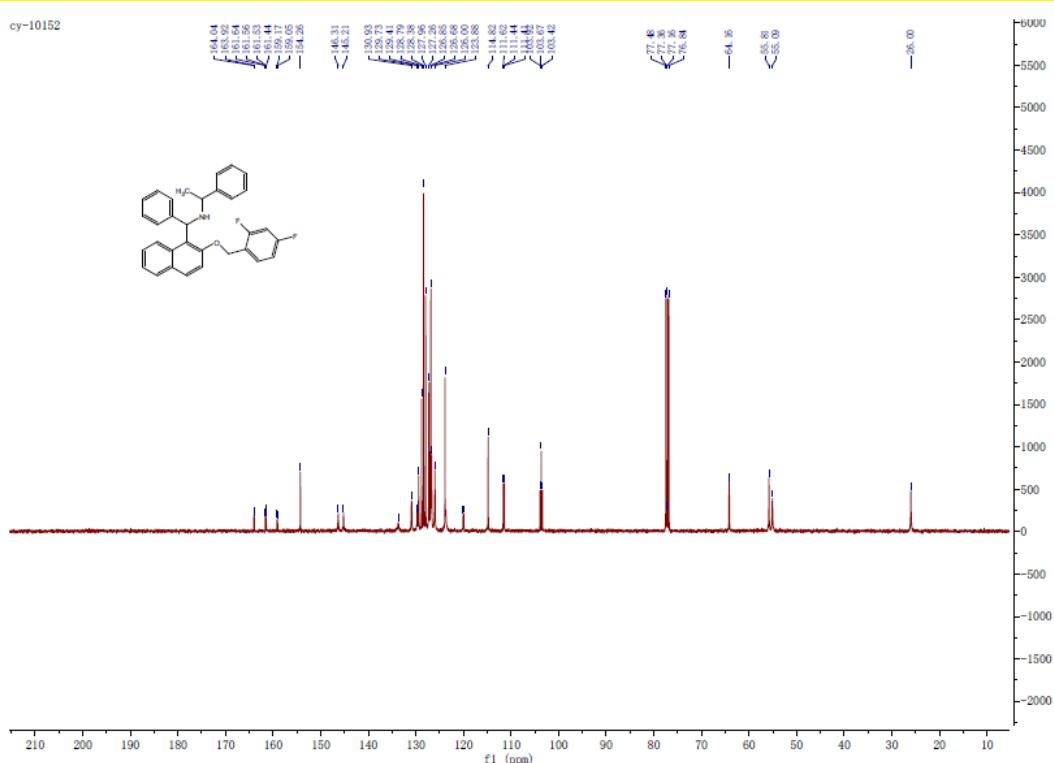
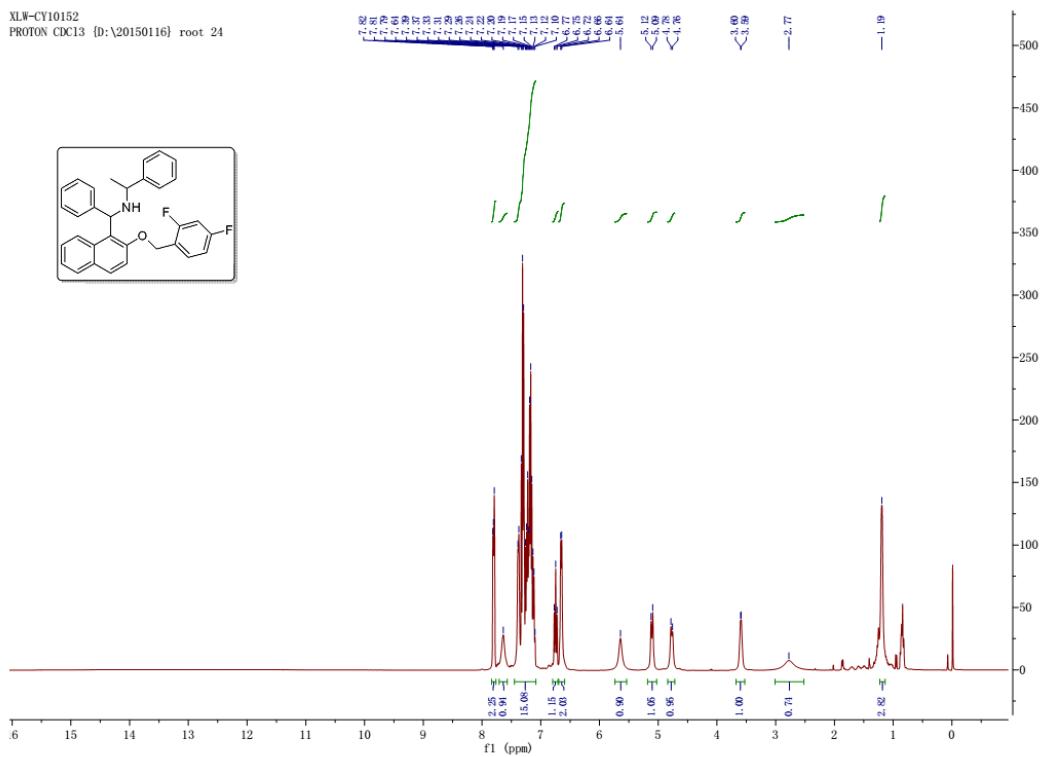
Acquisition Date 1/20/2015 2:51:37 PM

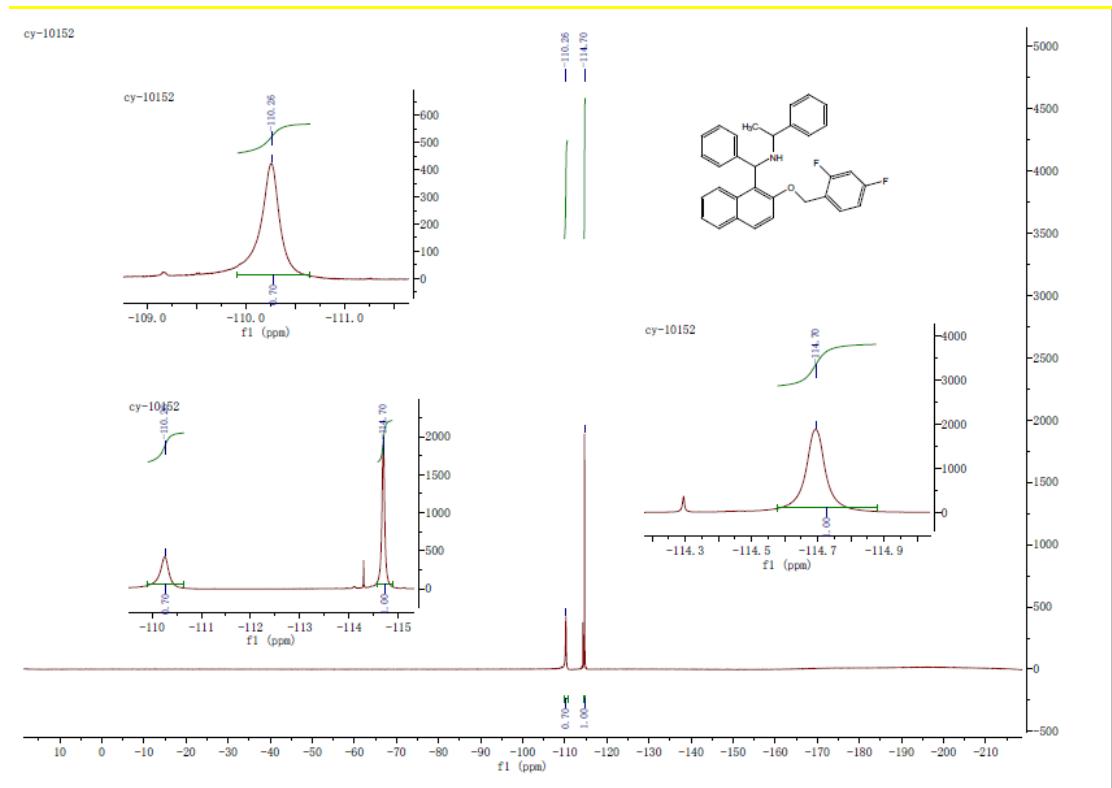
Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source







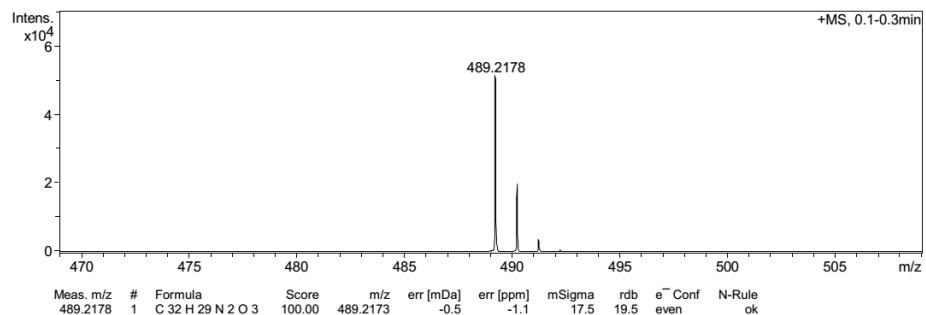
## Mass Spectrum SmartFormula Report

### Analysis Info

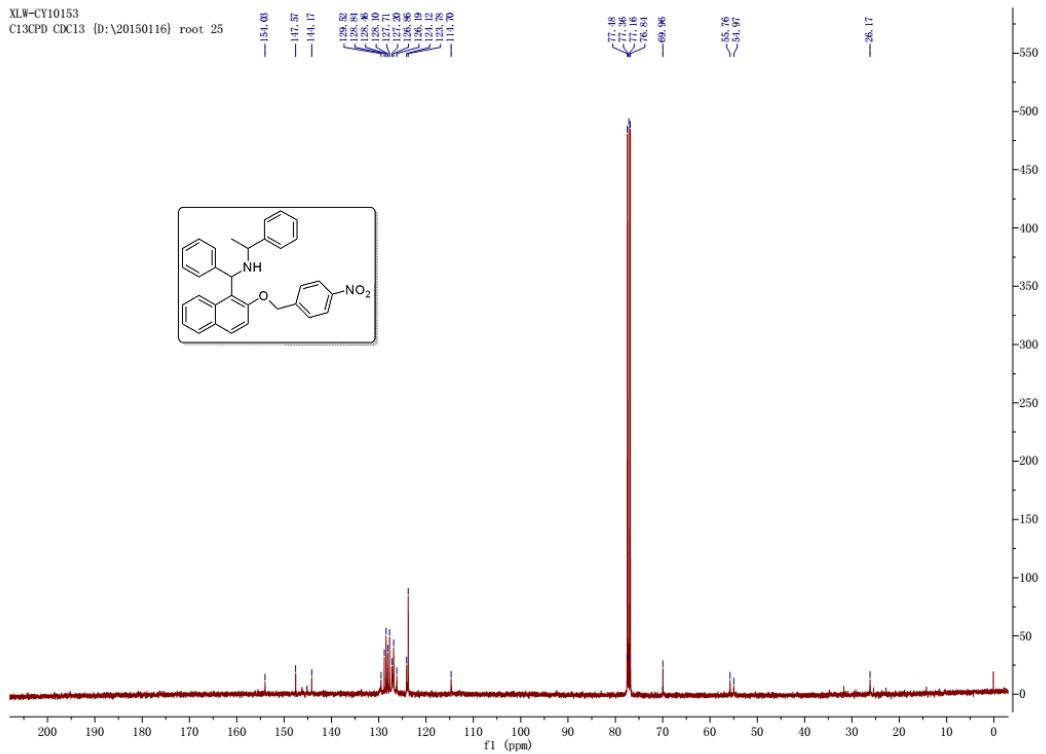
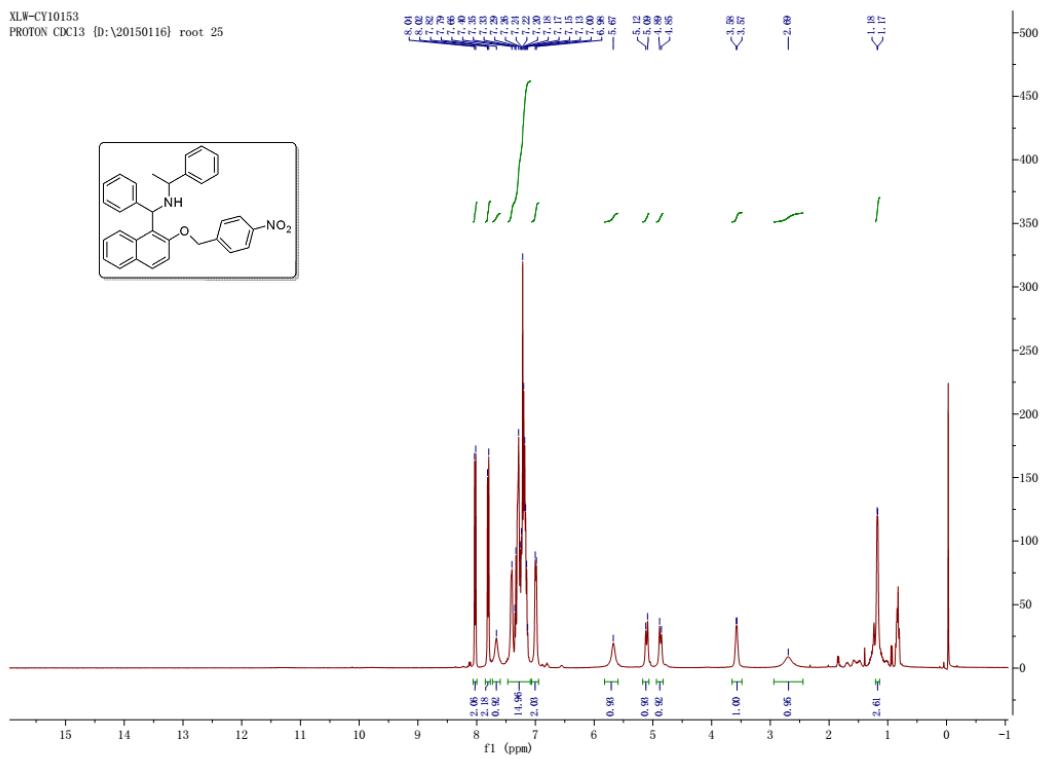
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150120-cy119-4\_01.d      Acquisition Date 1/20/2015 2:55:33 PM  
Method tune\_100-550\_pos150120.m      Operator Jiang  
Sample Name      Instrument / Ser# micrOTOF-Q II 10324  
Comment

### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



XLW-CY10153  
PROTON CDCl<sub>3</sub> {D:\20150116} root 25



## Mass Spectrum SmartFormula Report

### Analysis Info

Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150120-cy119-8\_01.d  
Method tune\_100-550\_pos150120.m  
Sample Name  
Comment

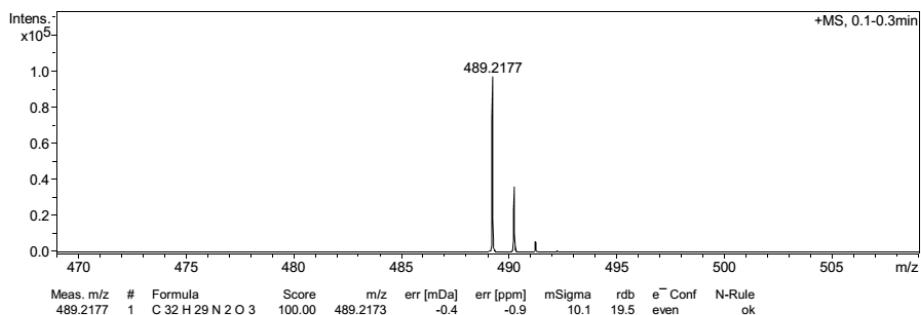
Acquisition Date 1/20/2015 3:01:31 PM

Operator Jiang

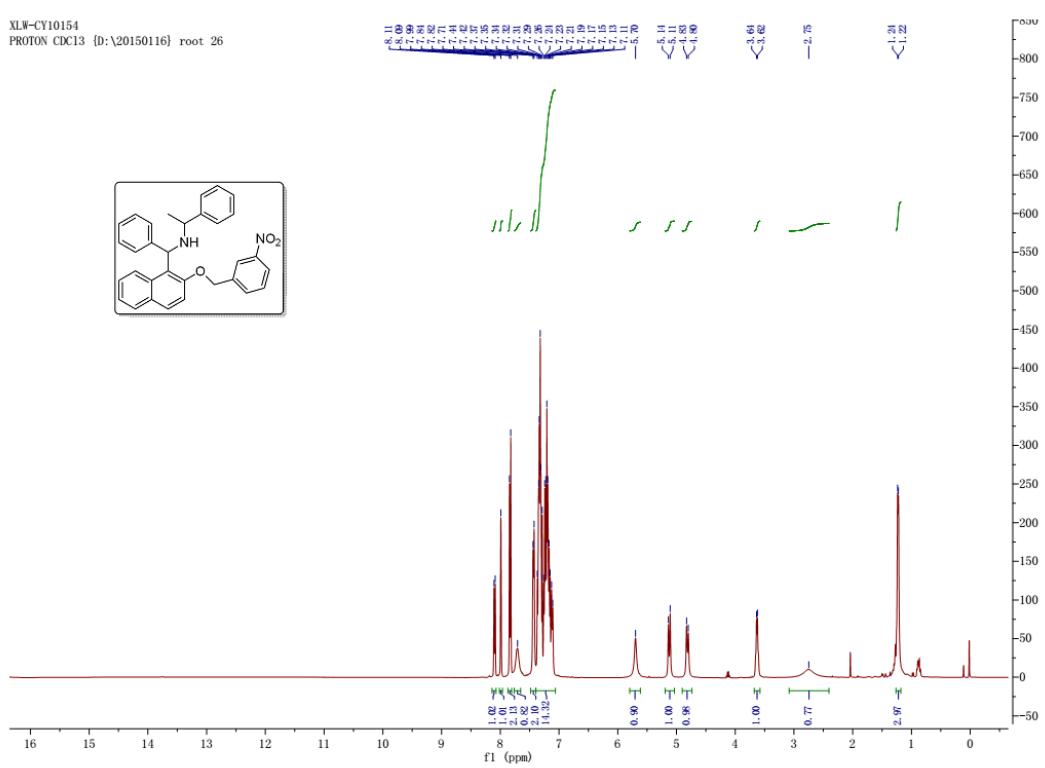
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

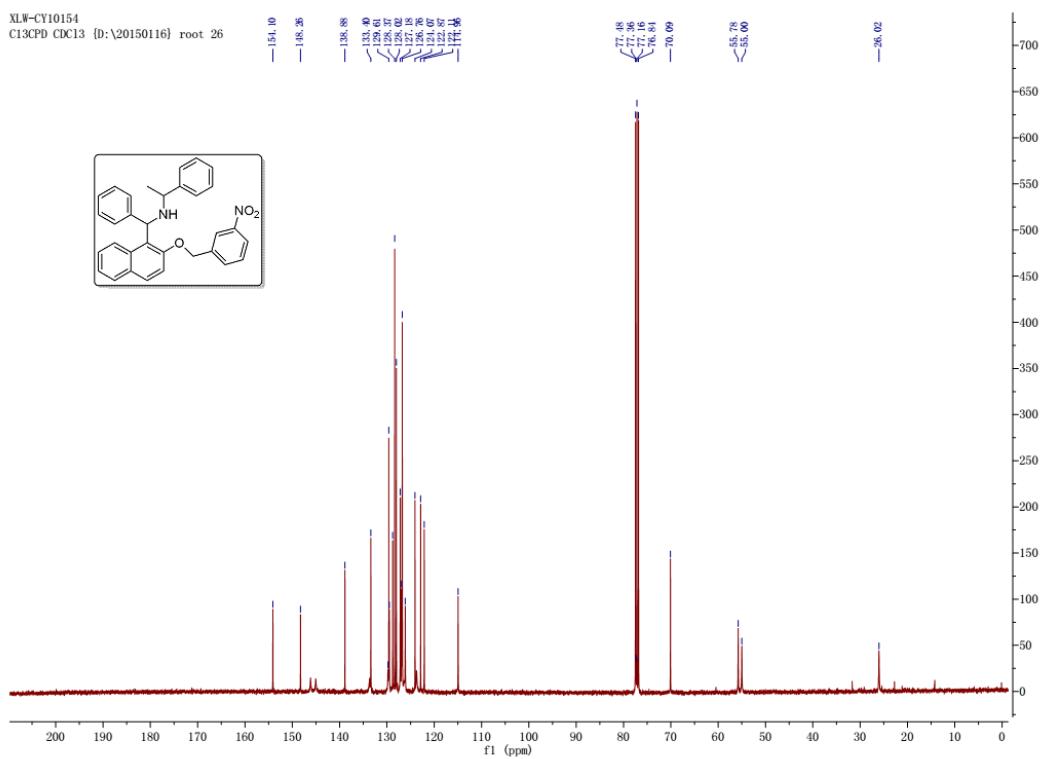
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



XLW-CY10154  
PROTON CDCl3 {D:\20150116} root 26



XLW-CY10154  
C13CPD CDCl3 {D:\20150116} root 26



## Mass Spectrum SmartFormula Report

### Analysis Info

Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150120-cy119-3\_01.d  
Method tune\_100-550\_pos150120.m  
Sample Name  
Comment

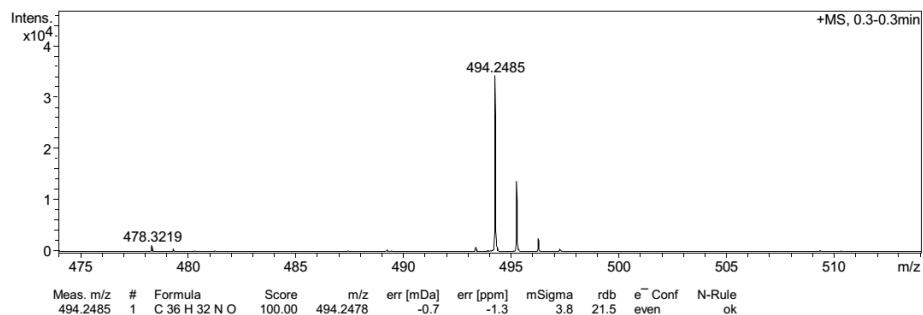
Acquisition Date 1/20/2015 2:49:07 PM

Operator Jiang

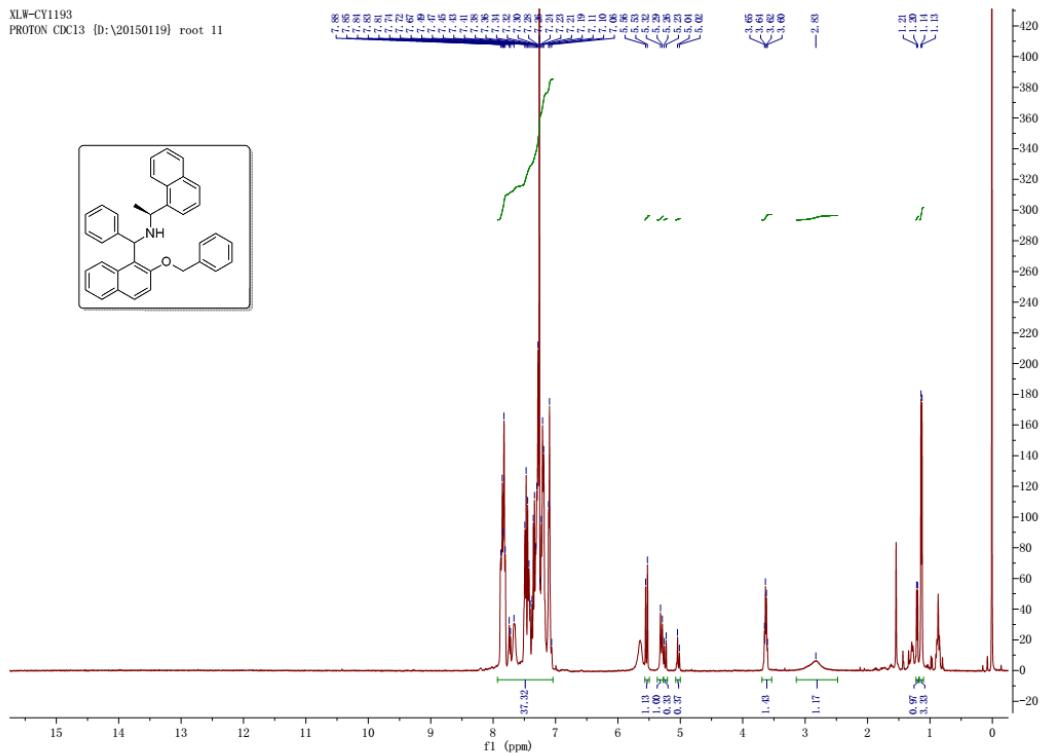
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

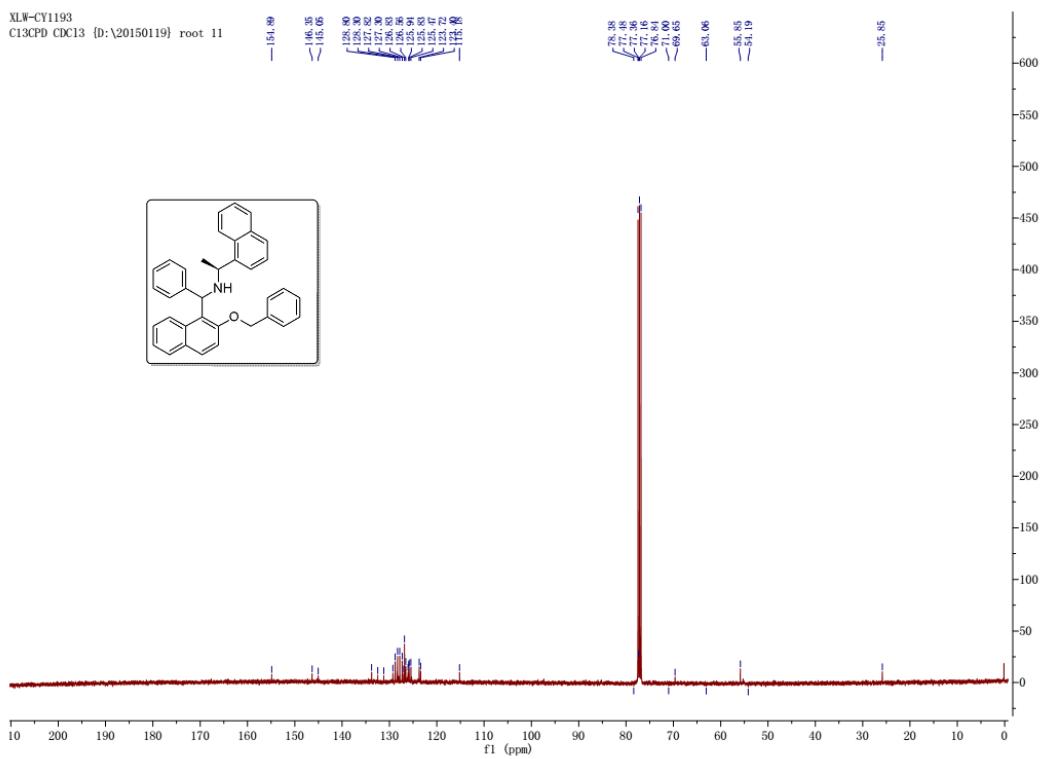
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



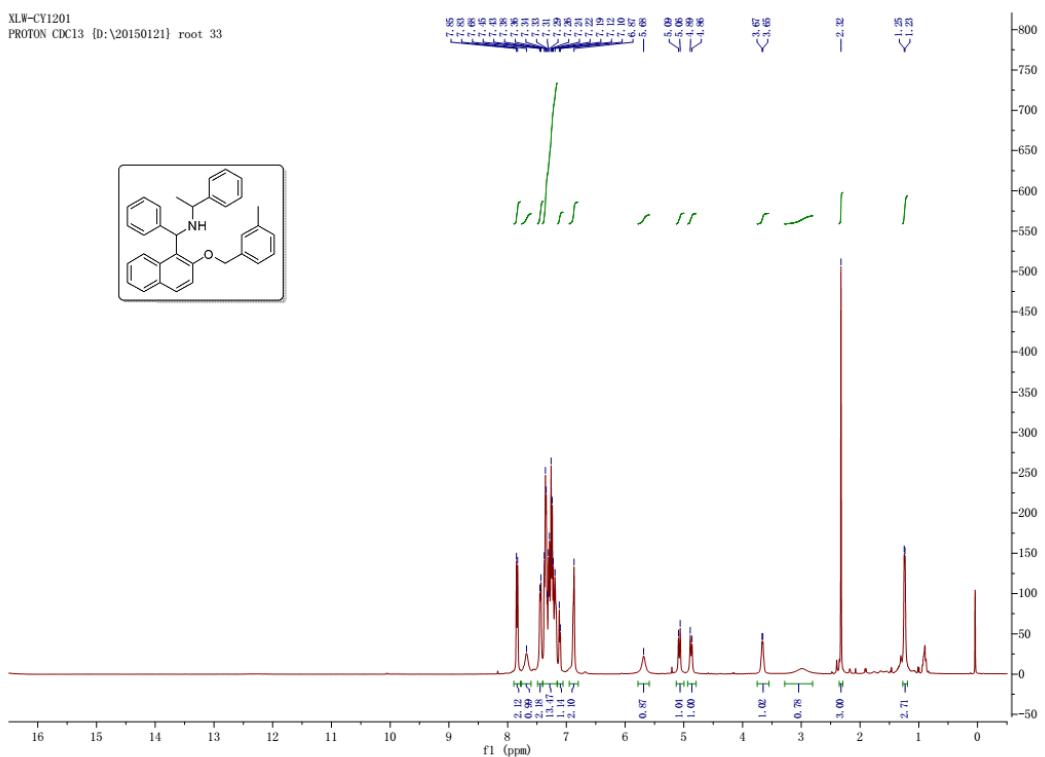
XLW-CY1193  
PROTON CDCl<sub>3</sub> {D:\20150119} root 11



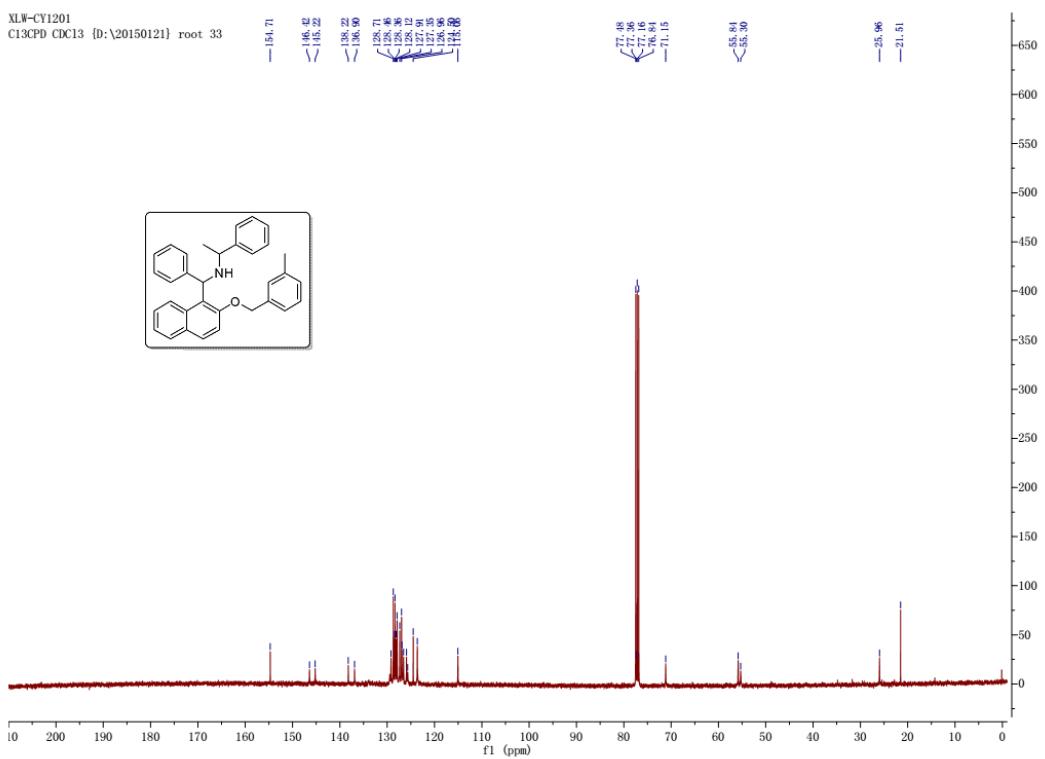
XLW-CY1193  
C13CPD CDCl<sub>3</sub> {D:\20150119} root 11



XLW-CY1201  
PROTON CDCl<sub>3</sub> {D:\20150121} root 33



XLW-CY1201  
C13CPD CDCl<sub>3</sub> {D:\20150121} root 33



## Mass Spectrum SmartFormula Report

### Analysis Info

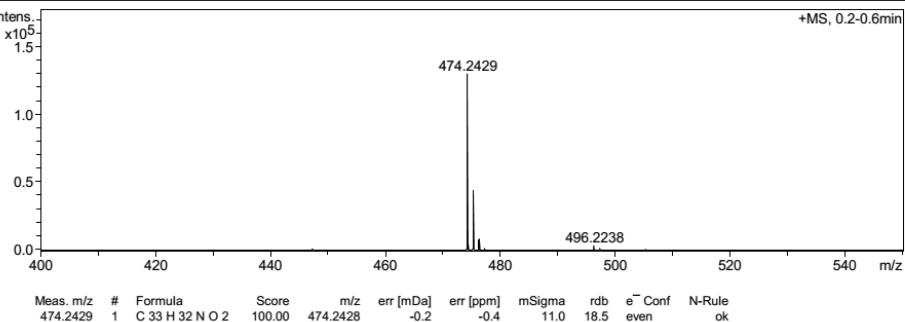
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150205-cy1252\_01.d  
Method tune\_100-550\_pos150205.m  
Sample Name wxy1501-3  
Comment

Acquisition Date 2/5/2015 9:44:08 AM

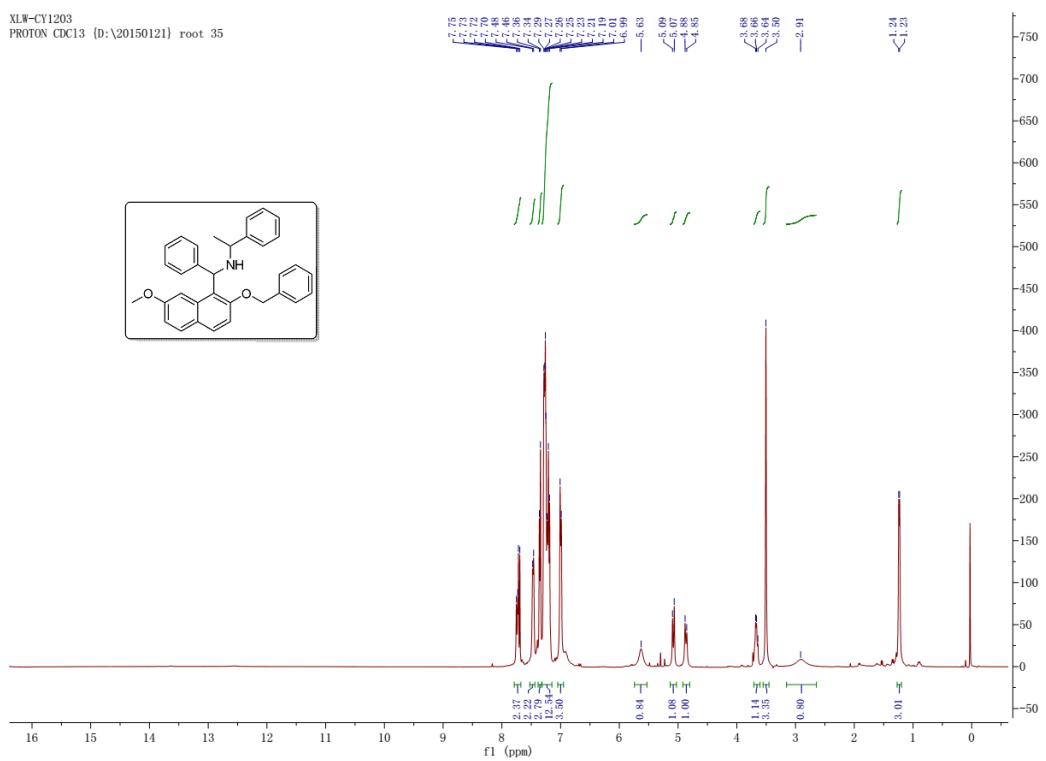
Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

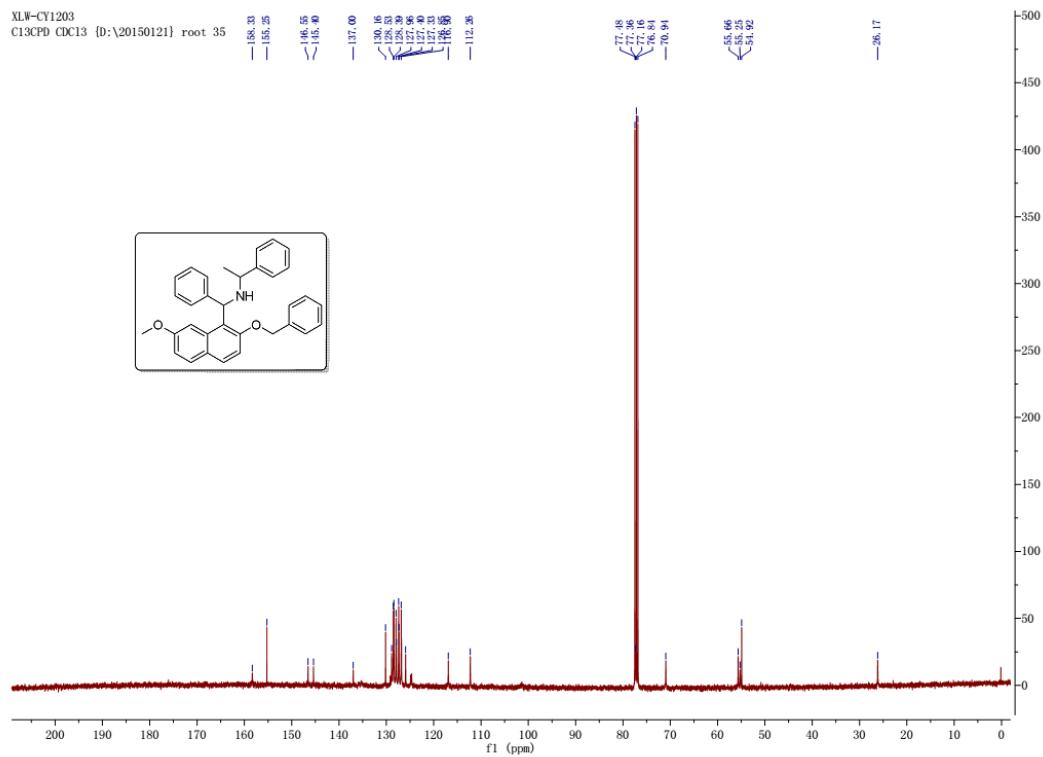
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



XLW-CY1203  
PROTON CDCl<sub>3</sub> {D:\20150121} root 35



XLW-CY1203  
C13CPD CDCl<sub>3</sub> {D:\20150121} root 35



## Mass Spectrum SmartFormula Report

### Analysis Info

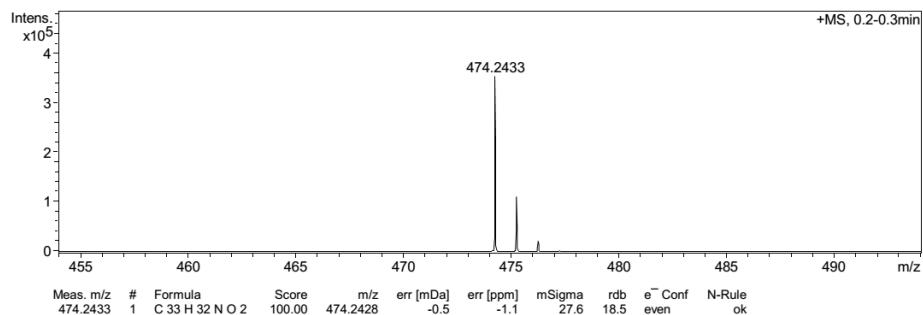
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150123-wcy-1216\_01.d  
Method tune\_100-550\_pos150120.m  
Sample Name  
Comment

Acquisition Date 1/23/2015 10:49:23 AM

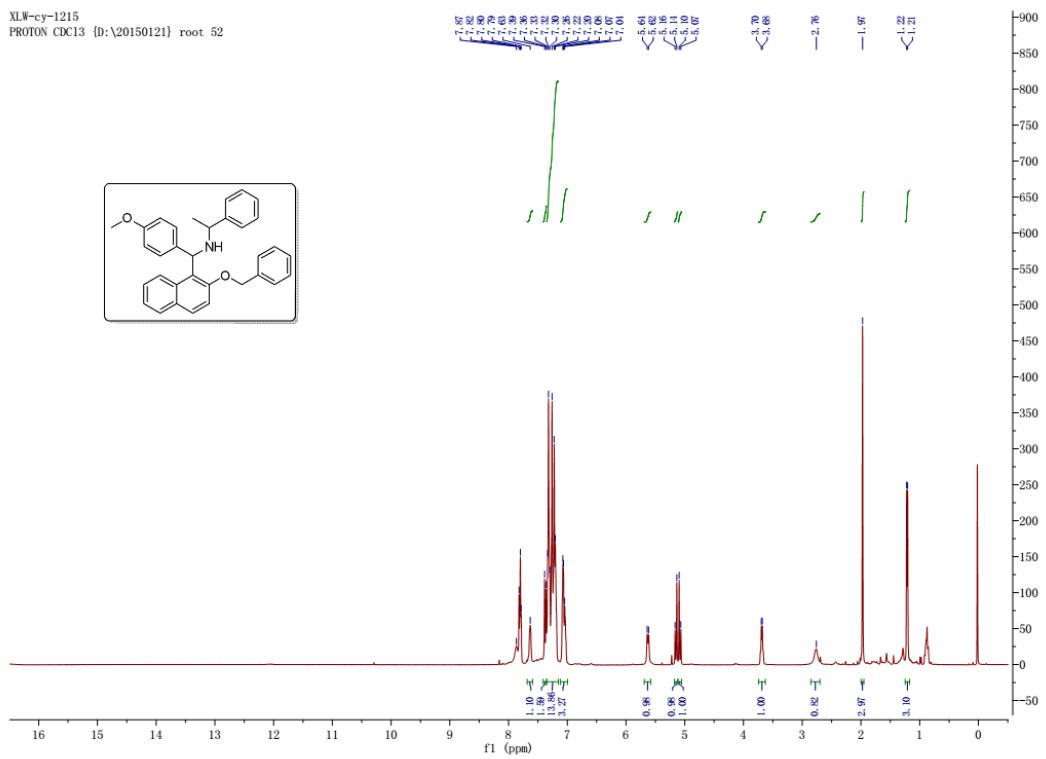
Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

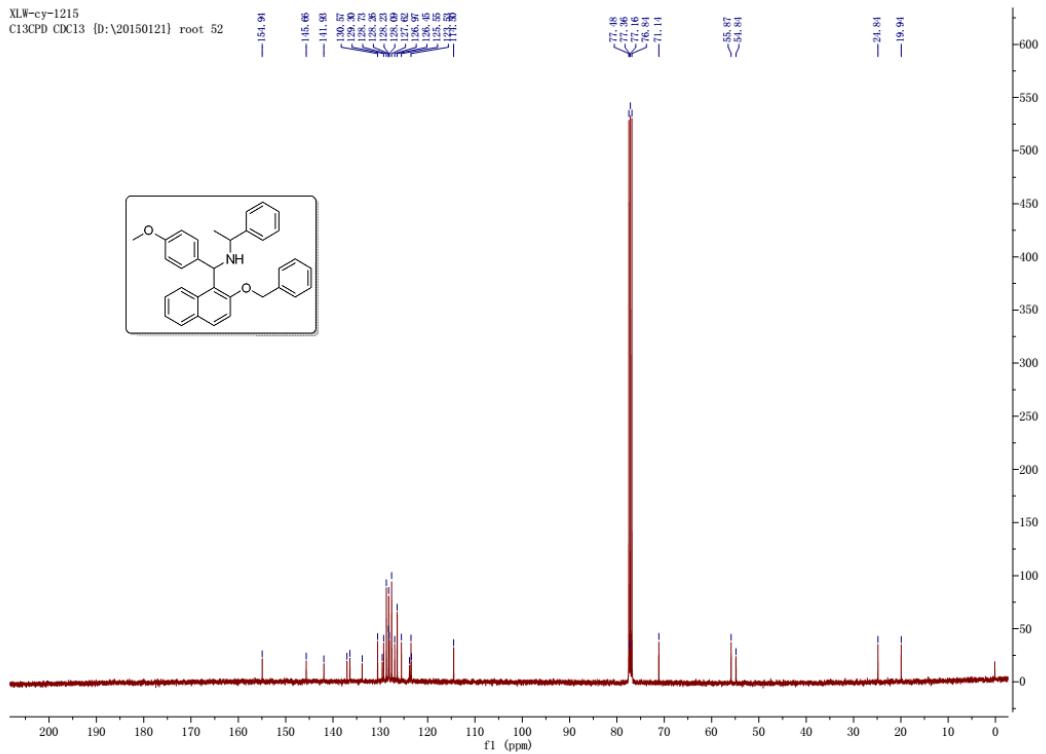
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



XLW-cy-1215  
PROTON CDCl<sub>3</sub> {D:\20150121} root 52



XLW-cy-1215  
C13CPD CDCl<sub>3</sub> {D:\20150121} root 52



## Mass Spectrum SmartFormula Report

### Analysis Info

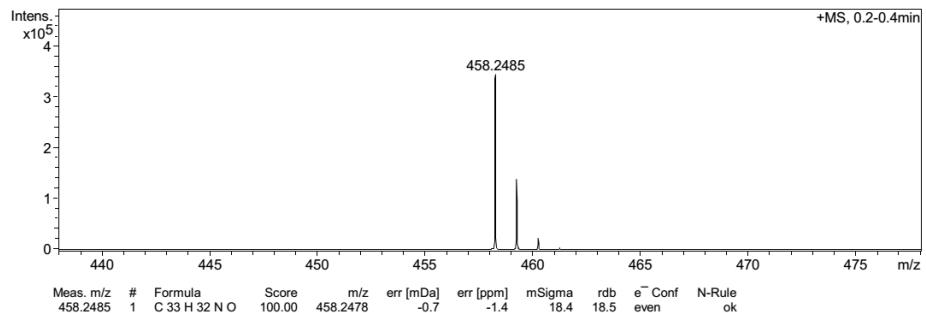
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150123-wcy-1215\_01.d  
Method tune\_100-550\_pos150120.m  
Sample Name  
Comment

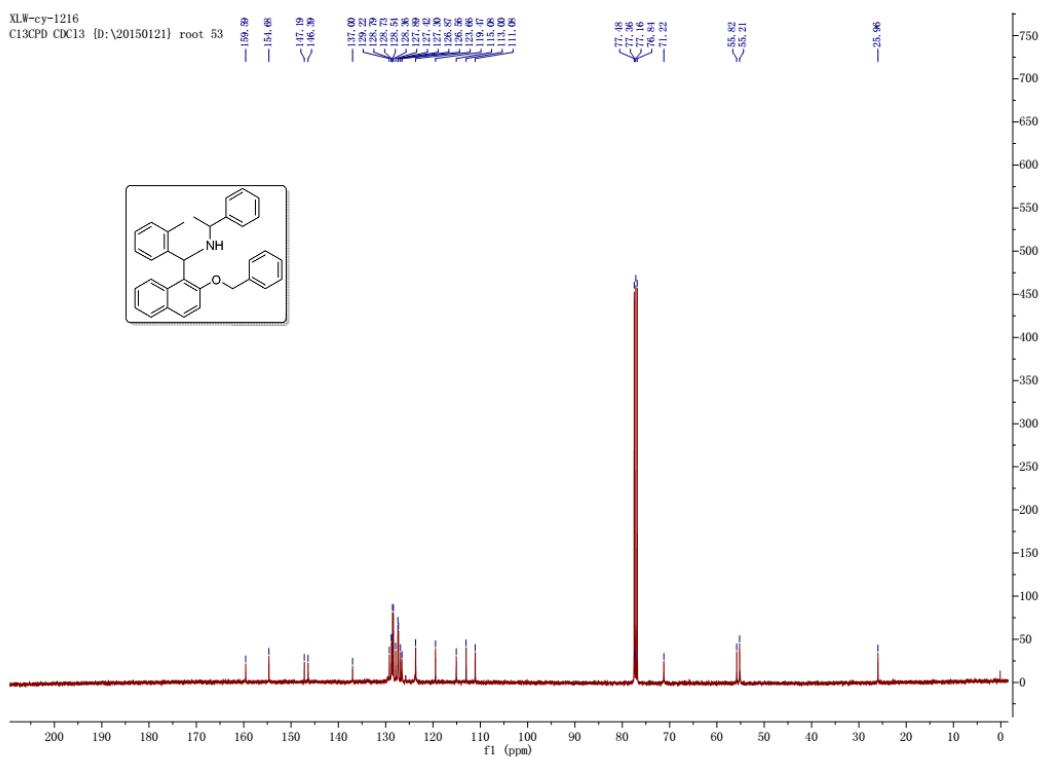
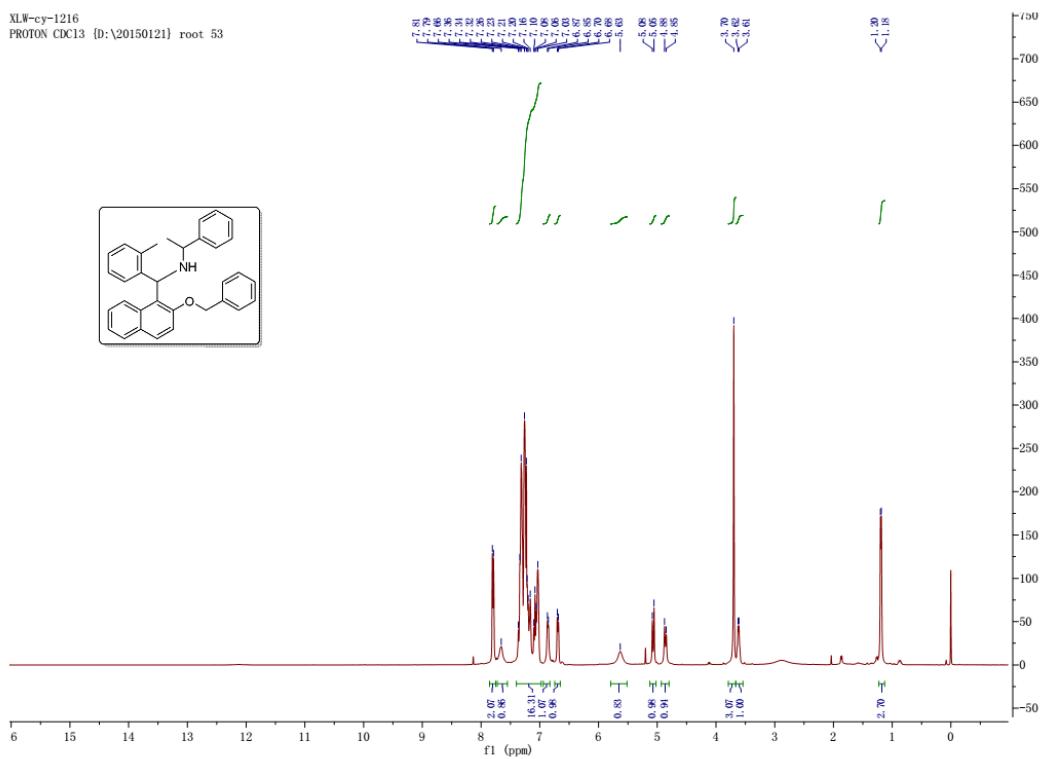
Acquisition Date 1/23/2015 10:47:31 AM

Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

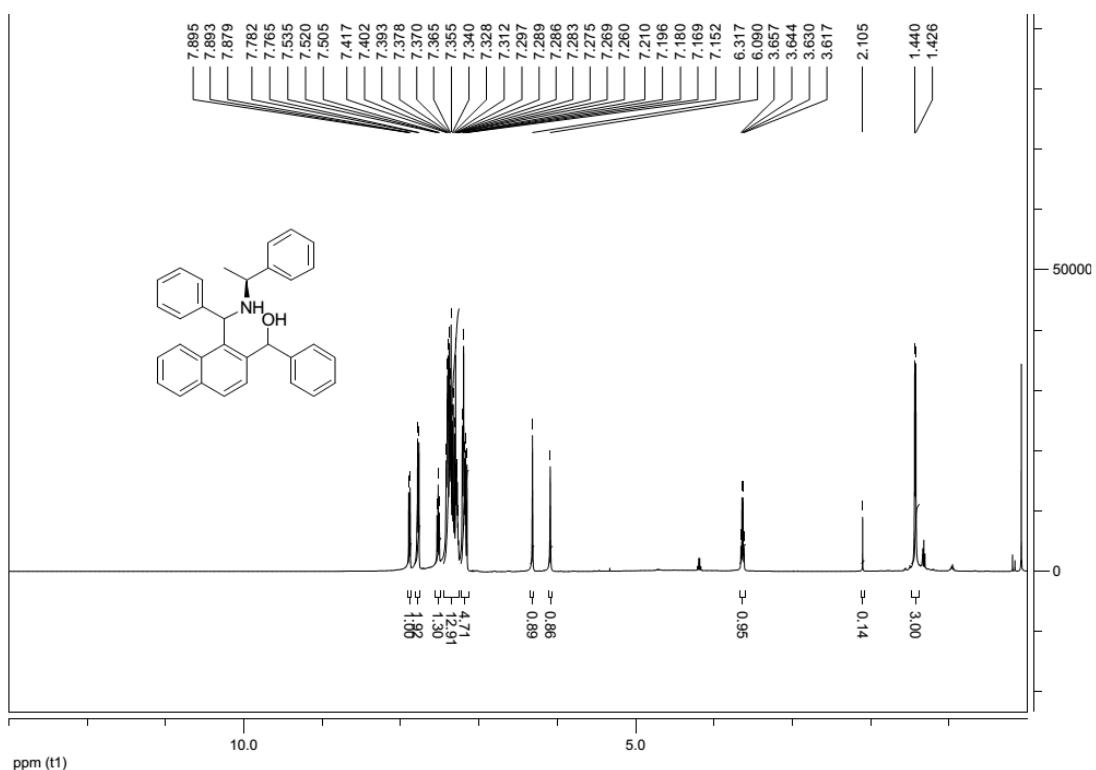
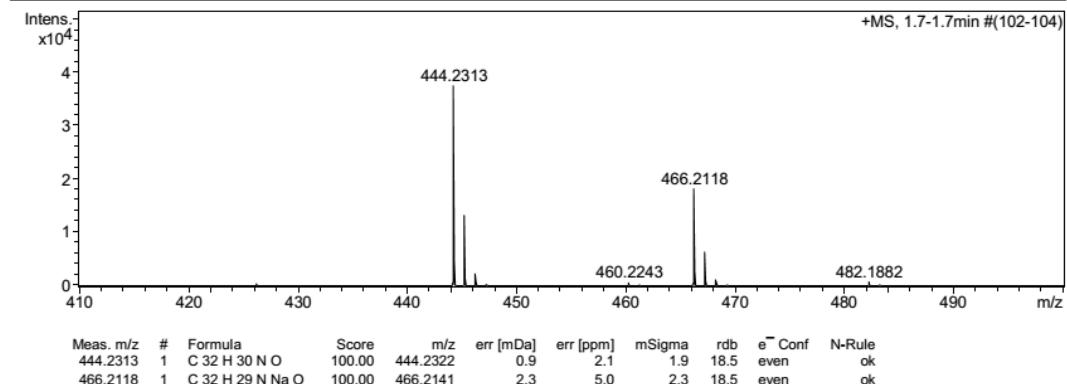
**Analysis Info**

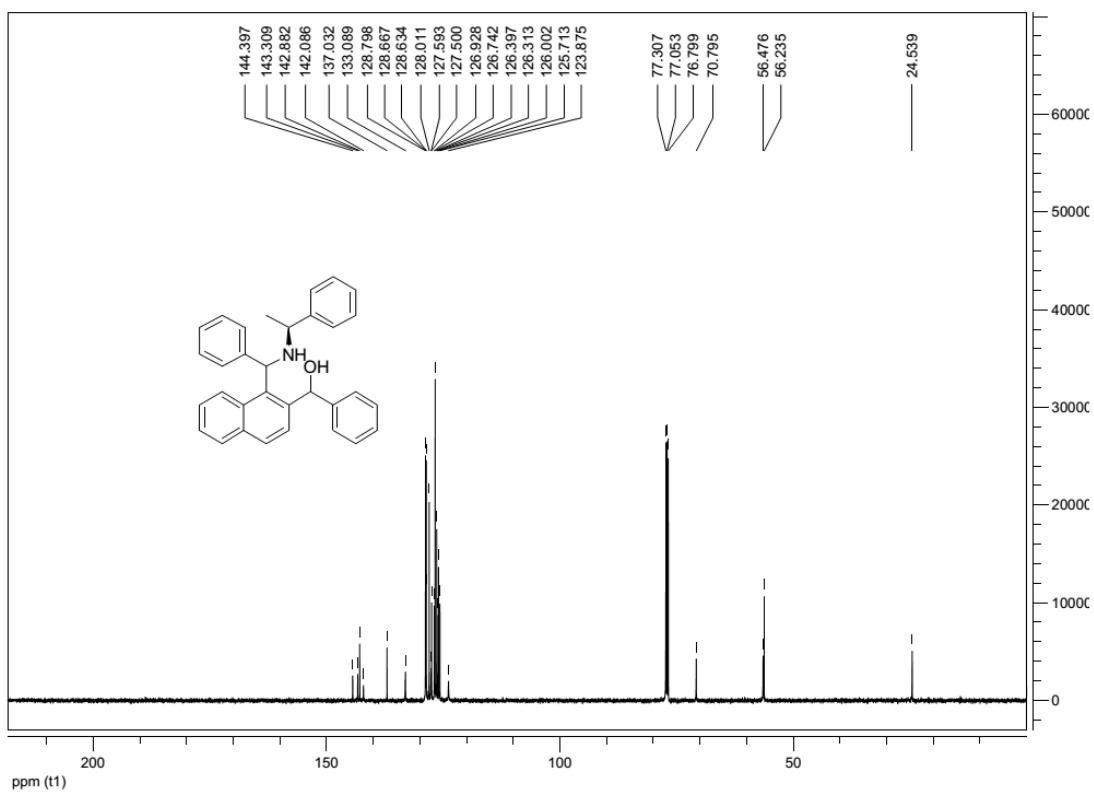
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw140106-wcy-CTWSBnCP\_01.d  
 Method tune-pos\_100-500\_140106.m  
 Sample Name CTWSBnCP  
 Comment

Acquisition Date 1/6/2014 12:57:11 PM  
 Operator Jiang  
 Instrument / Ser# micrOTOF-Q II 10324

**Acquisition Parameter**

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	100 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

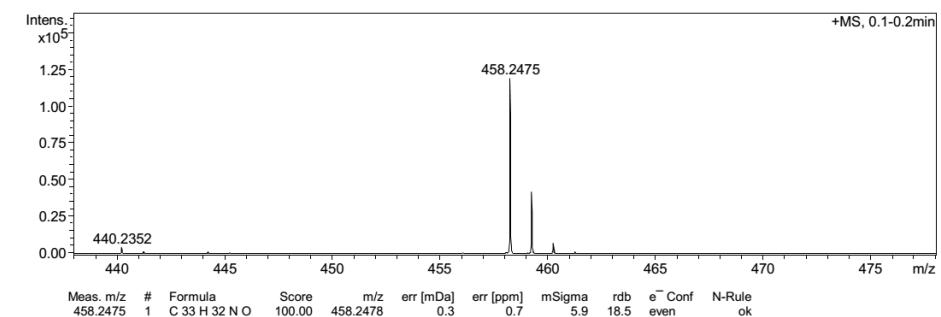
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150120-cy119-6\_01.d  
Method tune\_100-550\_pos150120.m  
Sample Name  
Comment

Acquisition Date 1/20/2015 2:53:33 PM

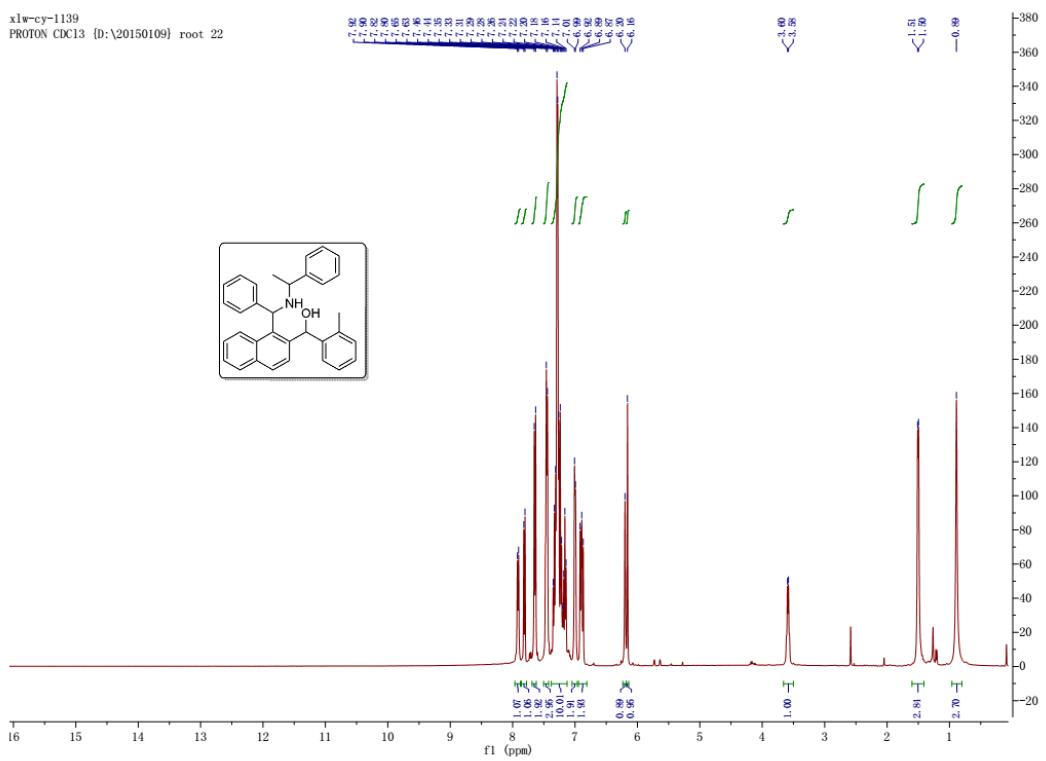
Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

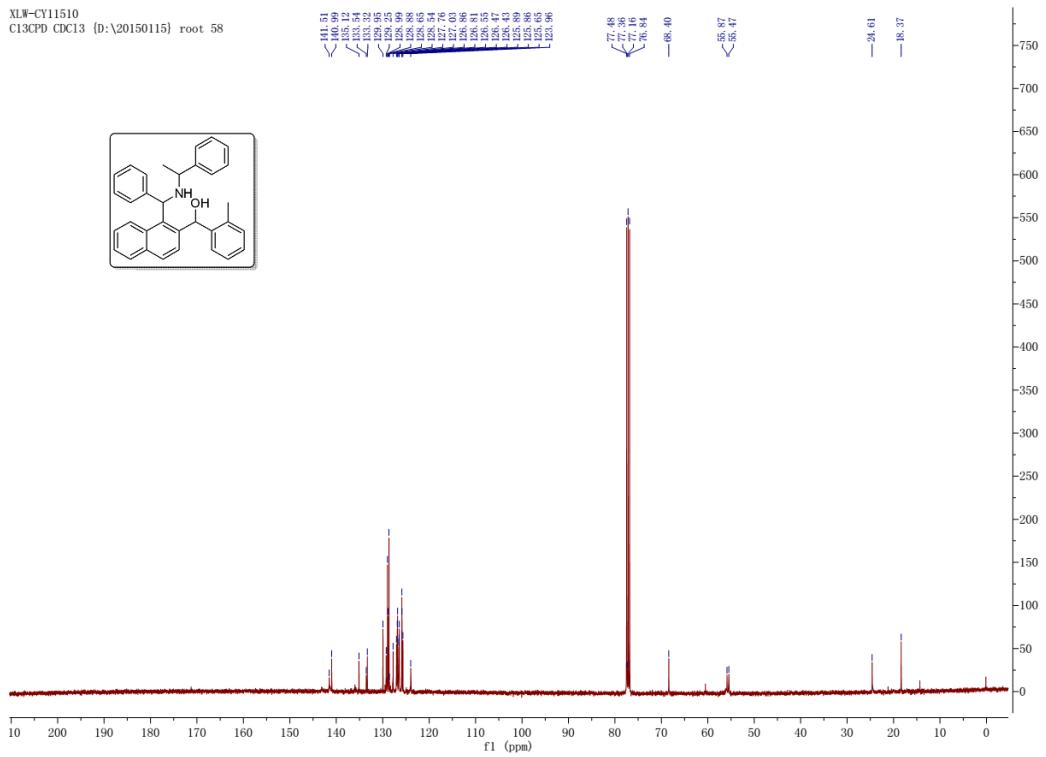
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



XLW-cy-1139  
PROTON CDCl<sub>3</sub> {D:\20150109} root 22



XLW-CY11510  
C13CPD CDCl<sub>3</sub> {D:\20150115} root 58



## Mass Spectrum SmartFormula Report

### Analysis Info

Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150120-cy119-5\_01.d  
Method tune\_100-550\_pos150120.m  
Sample Name  
Comment

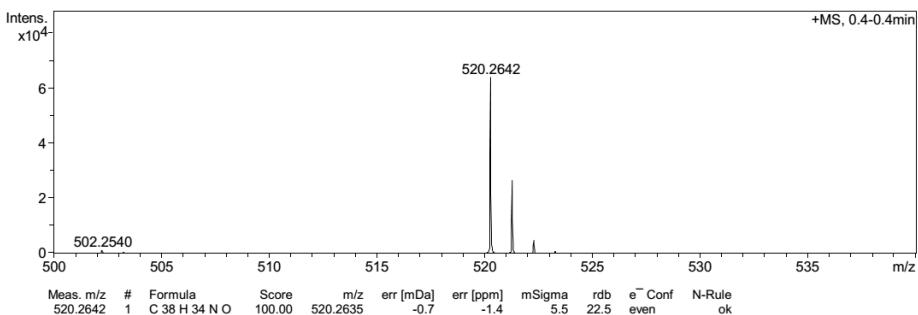
Acquisition Date 1/20/2015 2:57:36 PM

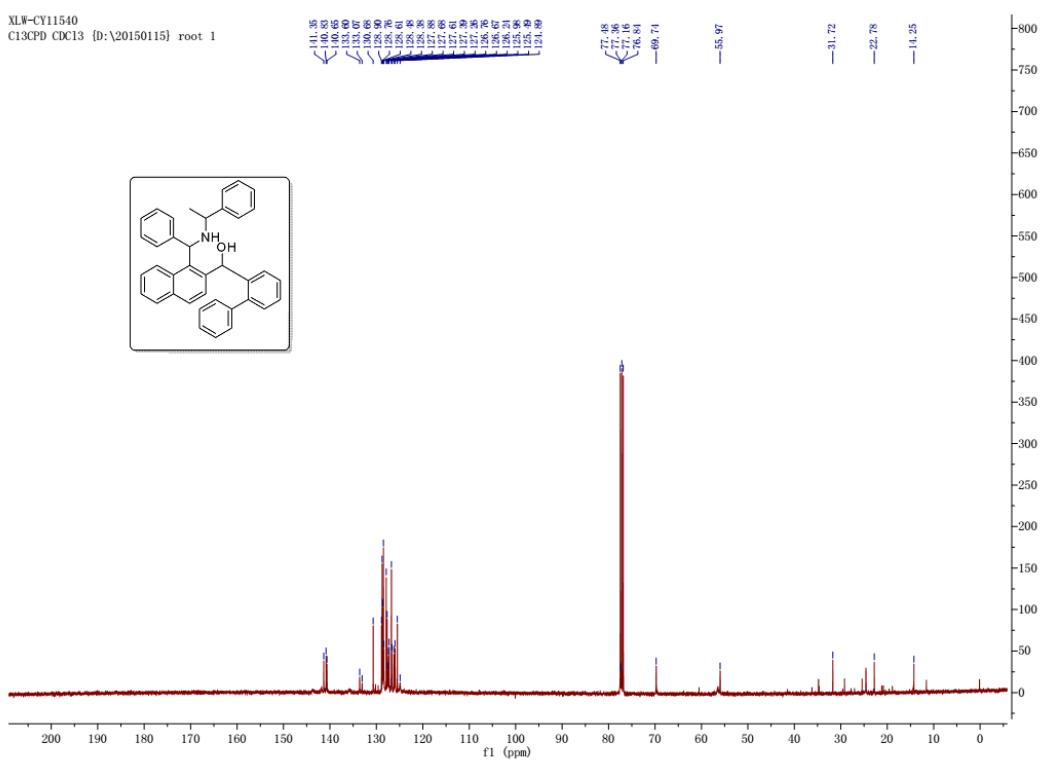
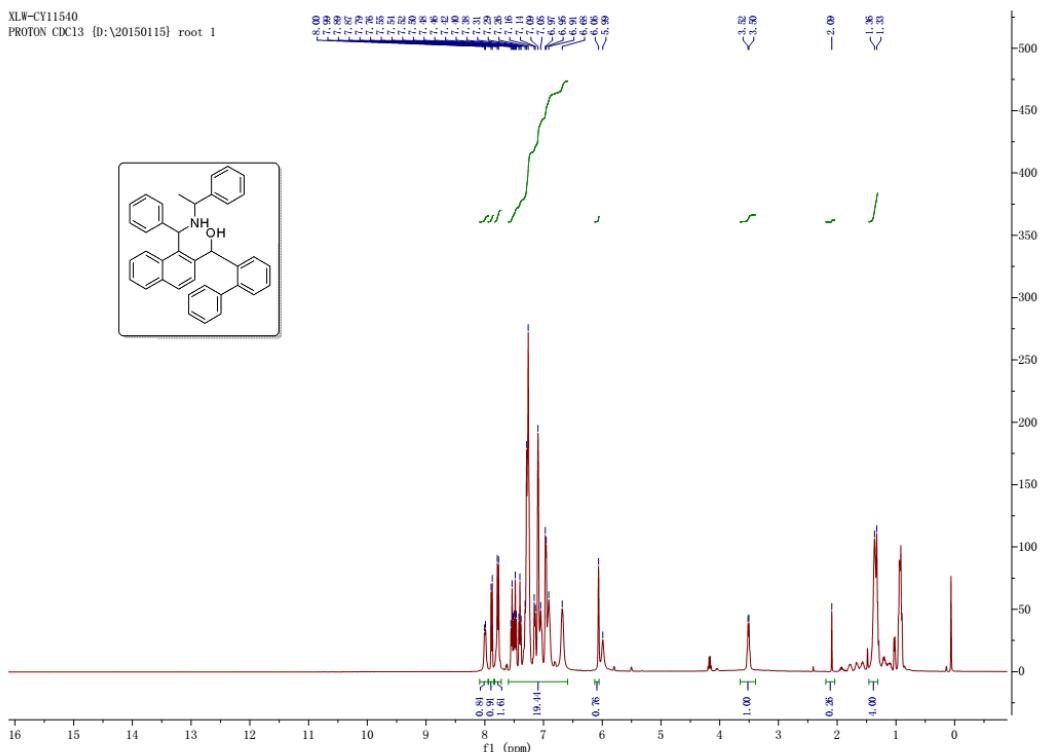
Operator Jiang

Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

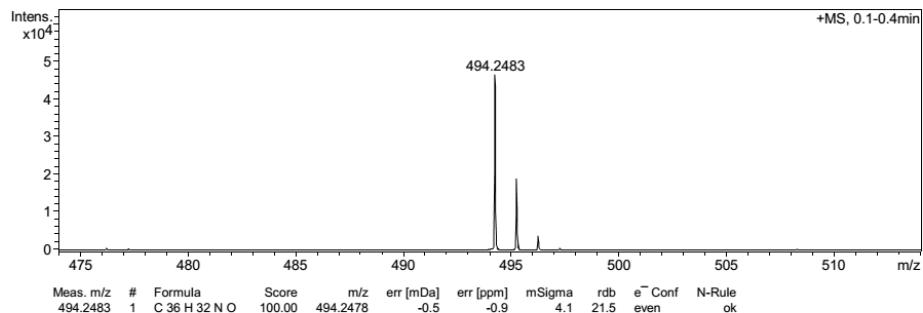
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150123-wcy-1211\_01.d  
Method tune\_100-550\_pos150120.m  
Sample Name  
Comment

Acquisition Date 1/23/2015 10:45:44 AM

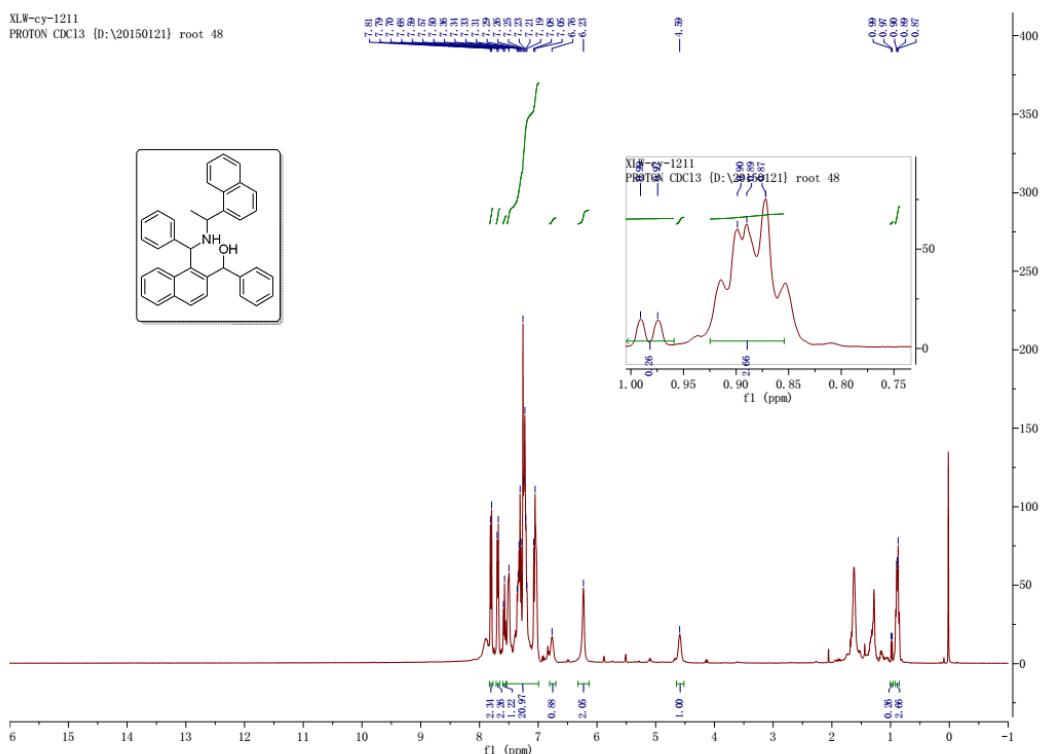
Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

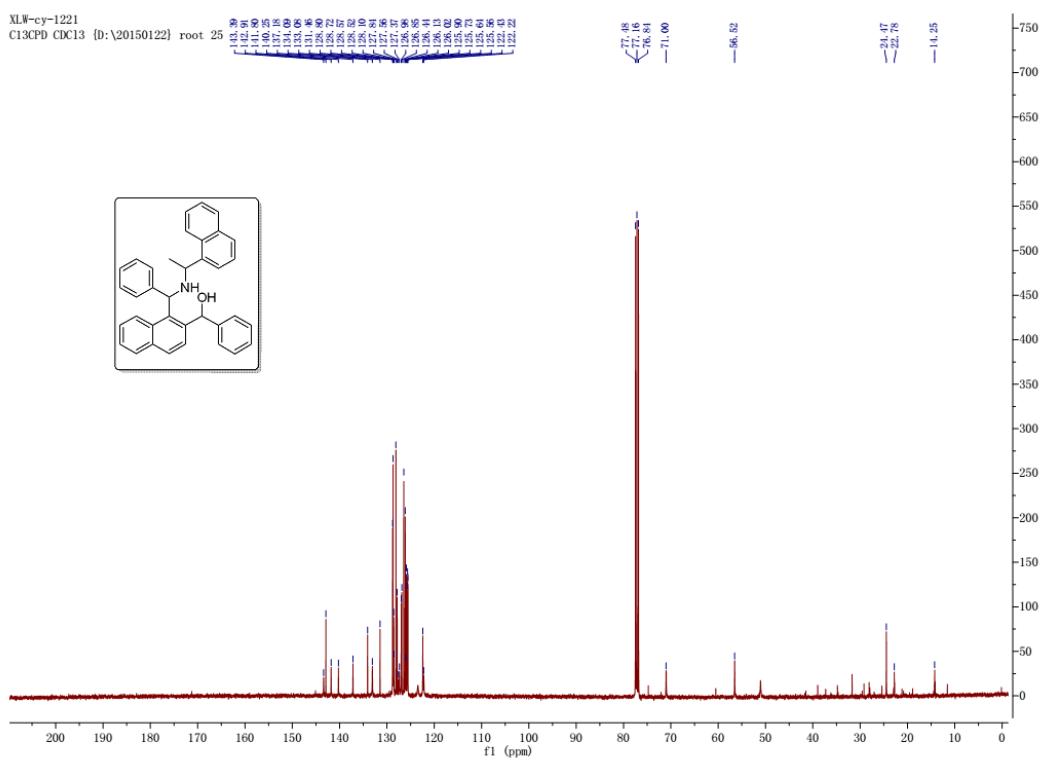
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



XLW-cy-1211  
PROTON CDCl<sub>3</sub> {D:\20150121} root 48



XLW-cy-1221  
C13CPD CDCl<sub>3</sub> {D:\20150122} root 25



## Mass Spectrum SmartFormula Report

### Analysis Info

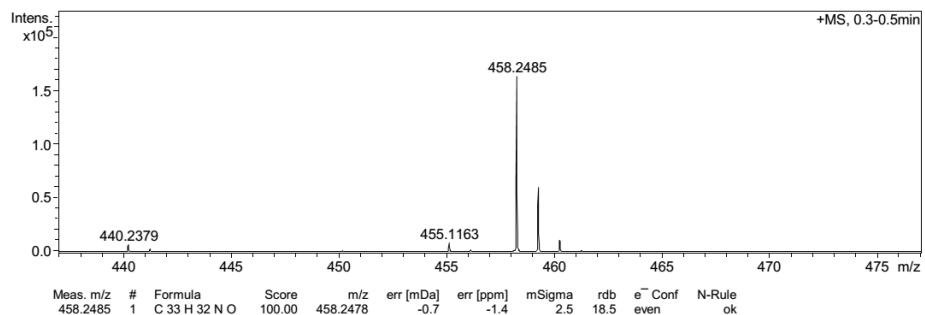
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Sample Name  
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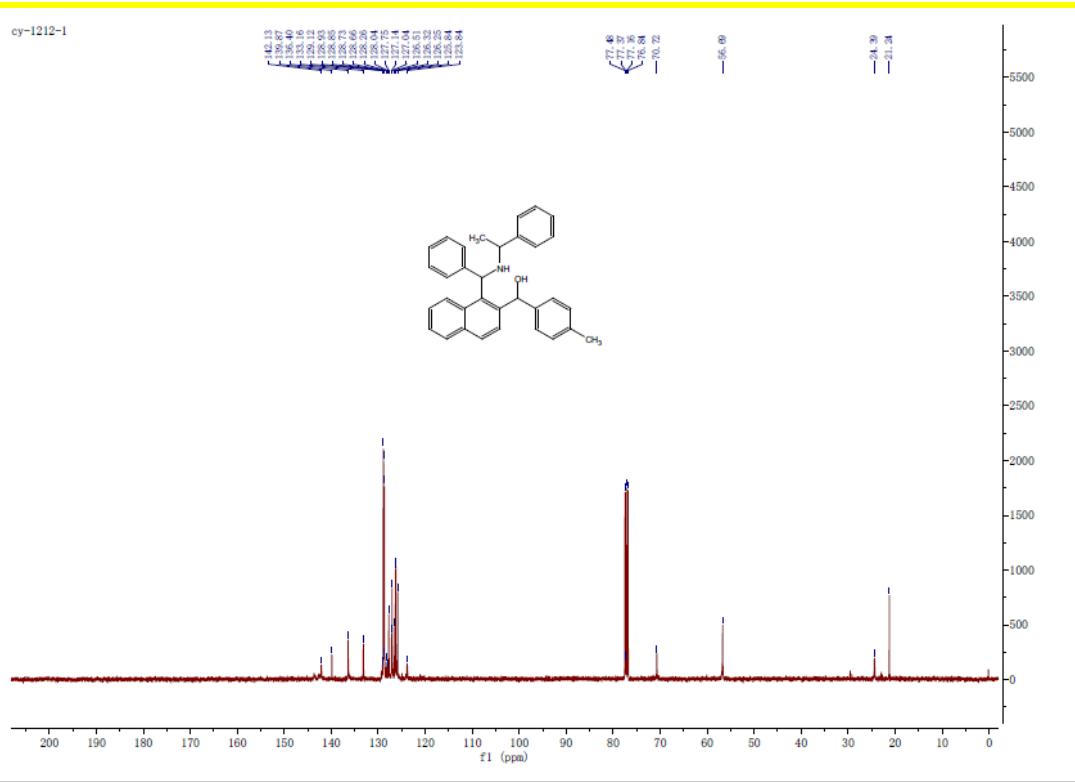
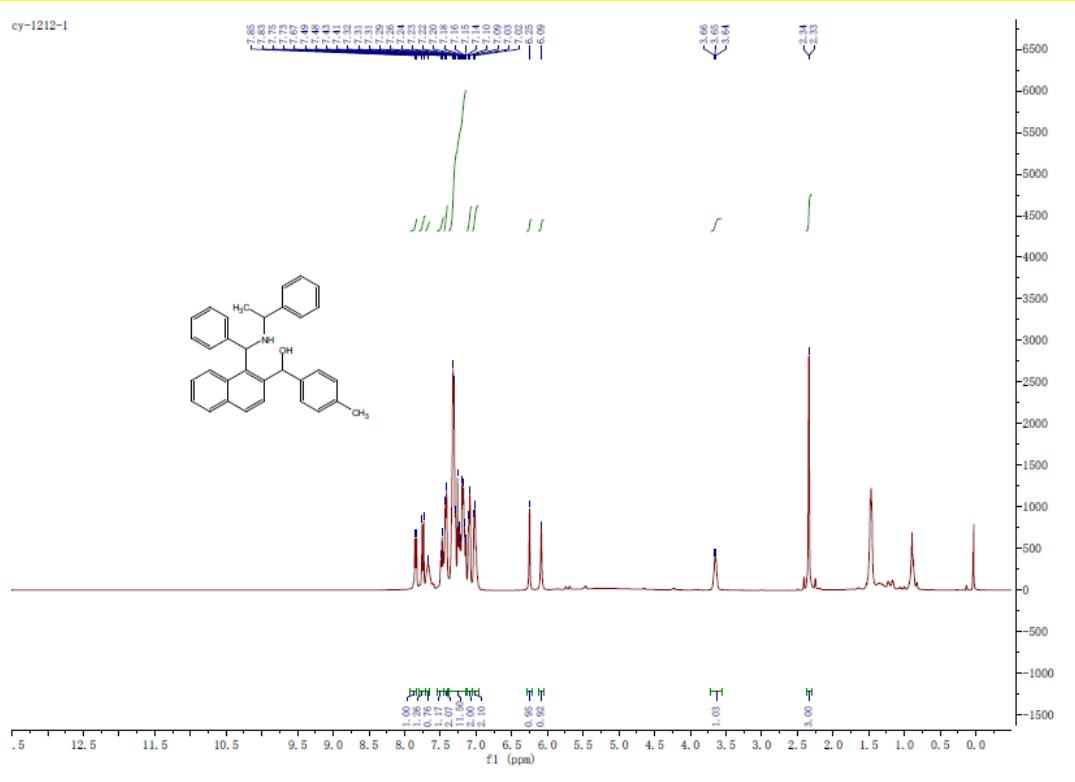
Acquisition Date 1/23/2015 10:43:38 AM

Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source





## Mass Spectrum SmartFormula Report

### Analysis Info

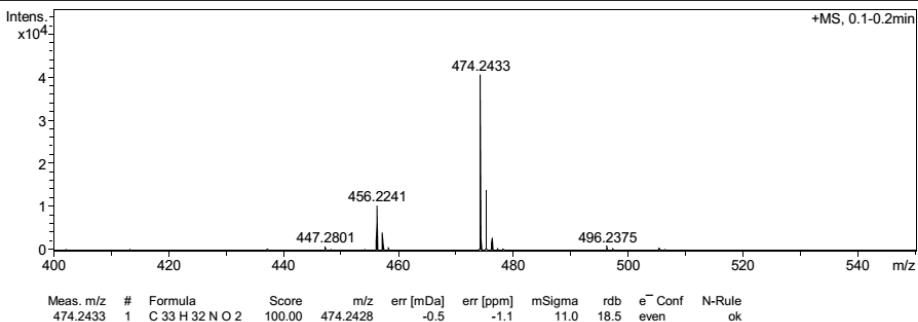
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Sample Name wxy1501-3  
Comment

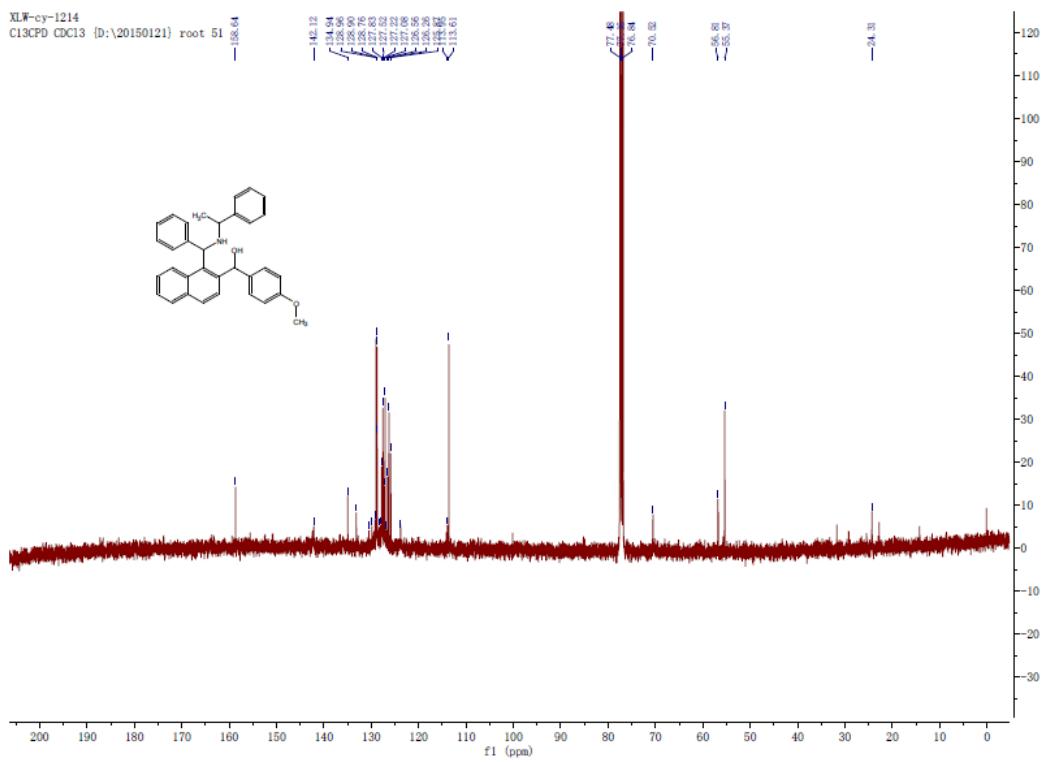
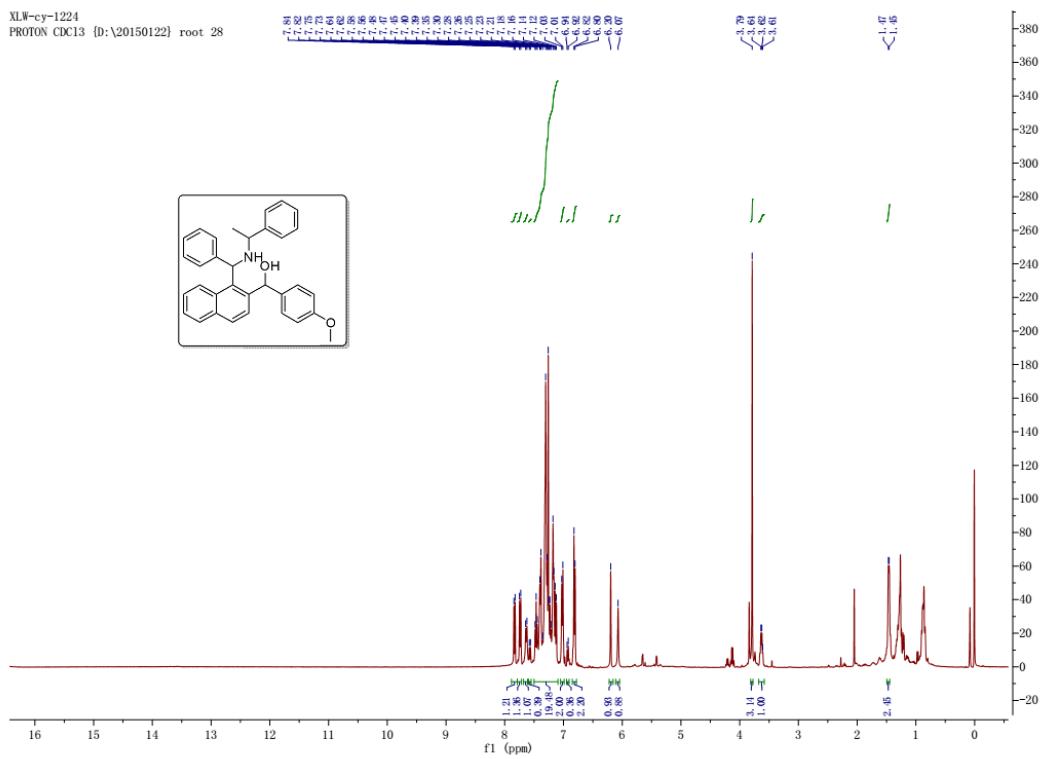
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Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

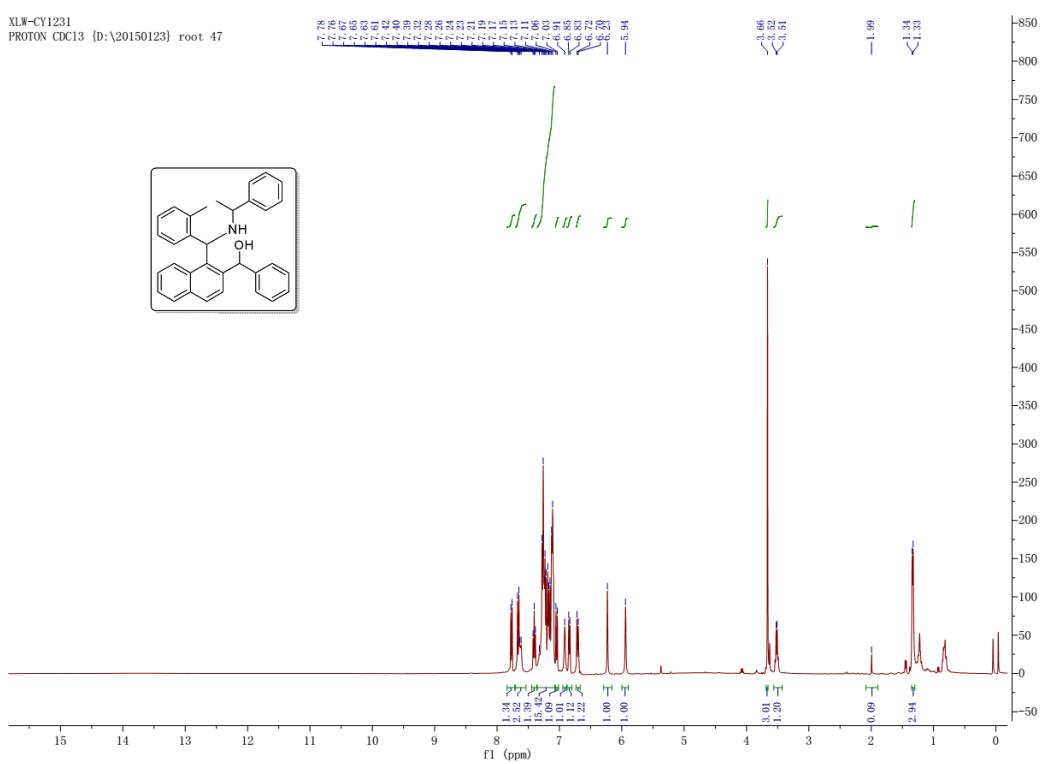
### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source

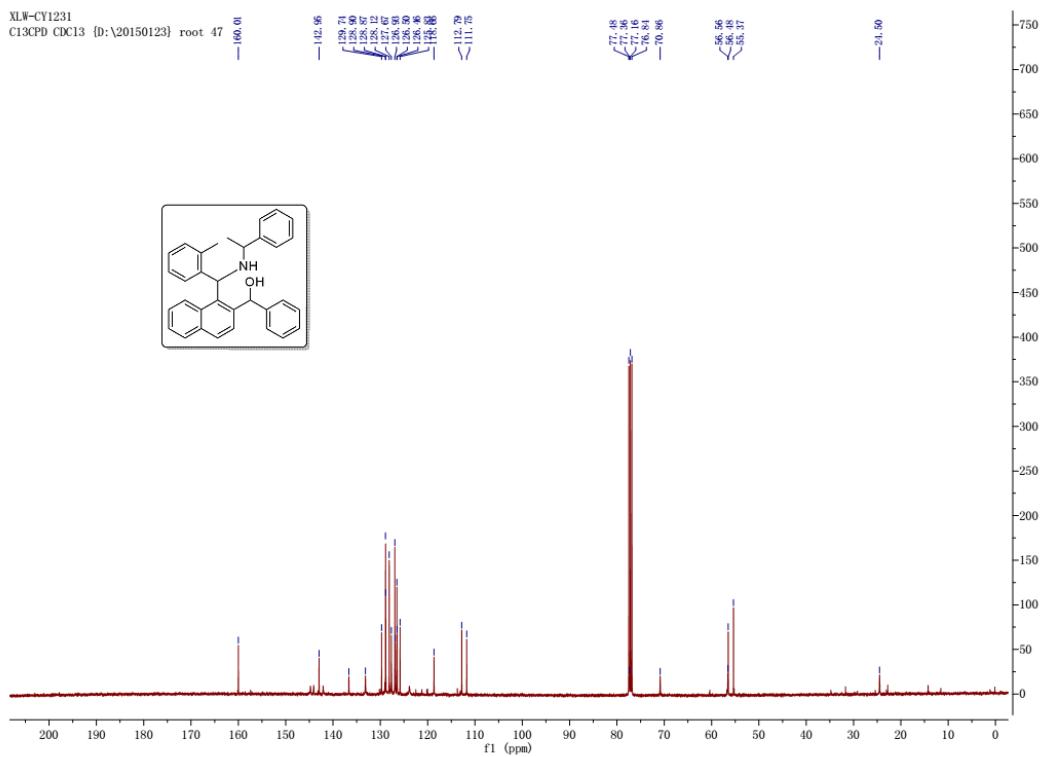




XLW-CY1231  
PROTON CDC13 {D:\20150123} root 47



XLW-CY1231  
C13CPD CDC13 {D:\20150123} root 47



## Mass Spectrum SmartFormula Report

### Analysis Info

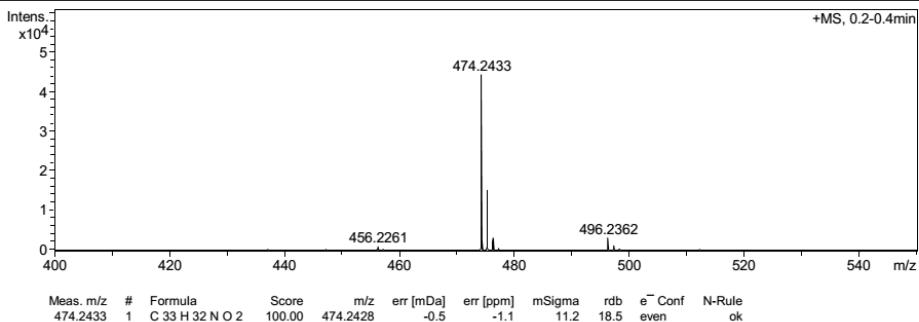
Analysis Name D:\Data\Xuliwen\Q-TOF-xulw150205-cy1255\_01.d  
Method tune\_100-550\_pos150205.m  
Sample Name wxy1501-3  
Comment

Acquisition Date 2/5/2015 9:54:58 AM

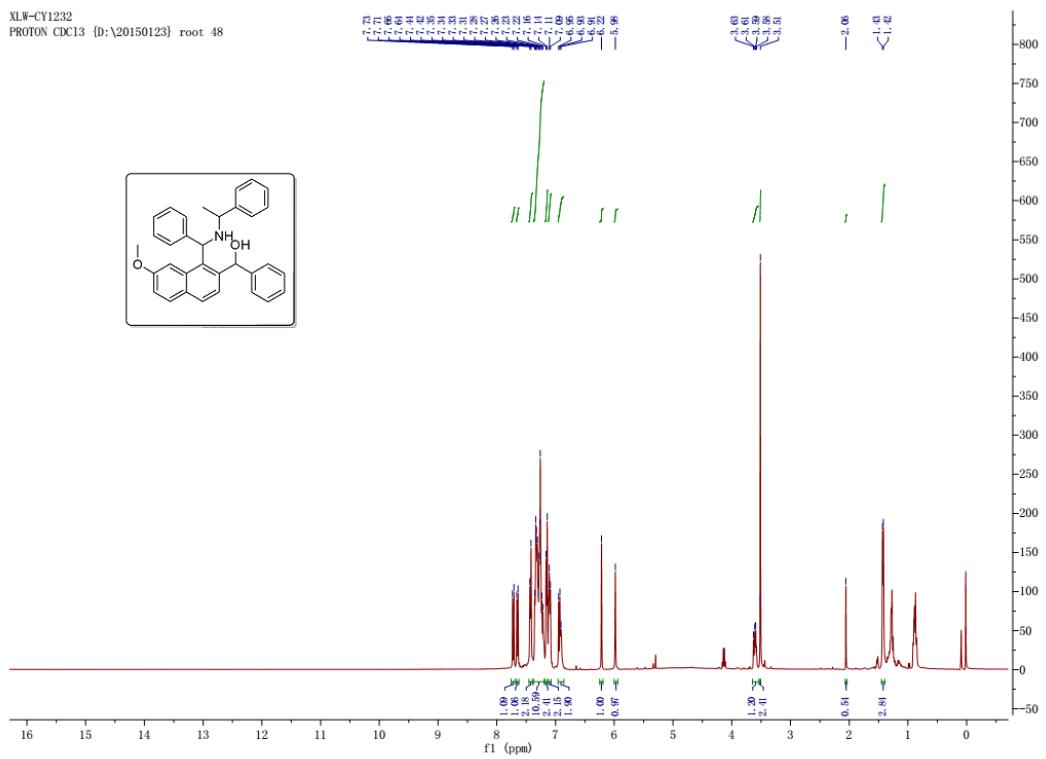
Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

### Acquisition Parameter

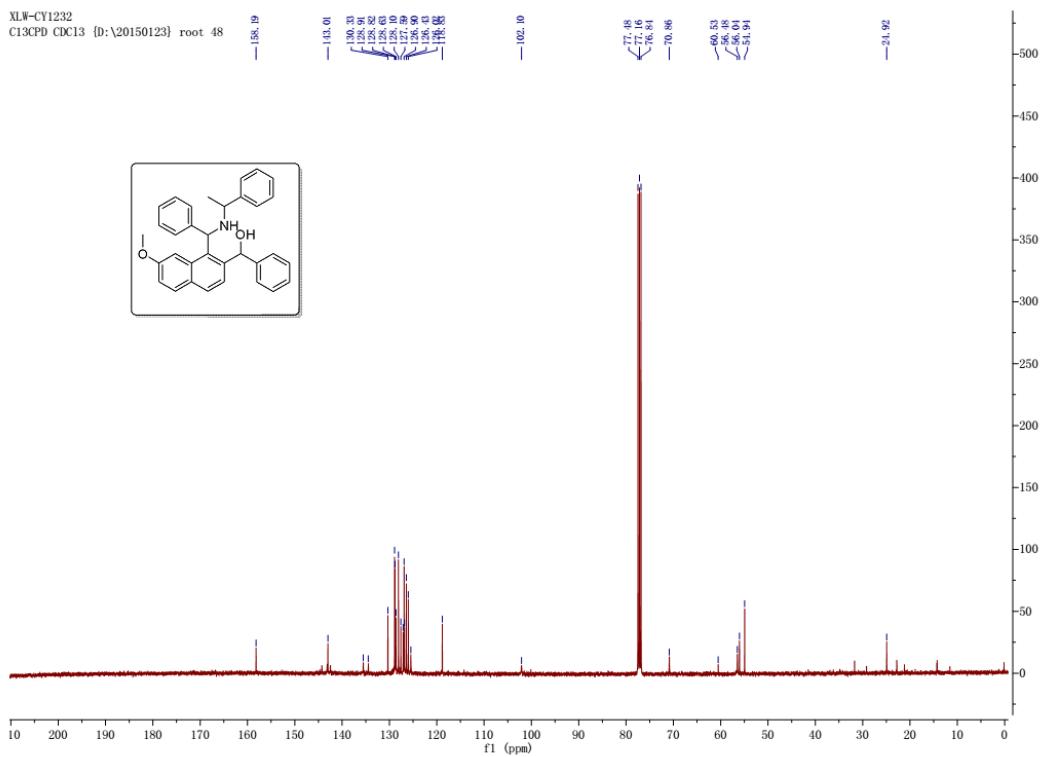
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Focus	Active	Set Capillary	4500 V	Set Dry Heater	200 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	2.2 l/min
Scan End	550 m/z	Set Collision Cell RF	120.0 Vpp	Set Divert Valve	Source



XLW-CY1232  
PROTON CDCl<sub>3</sub> {D:\20150123} root 48



XLW-CY1232  
C13CPD CDCl<sub>3</sub> {D:\20150123} root 48



## Mass Spectrum SmartFormula Report

### Analysis Info

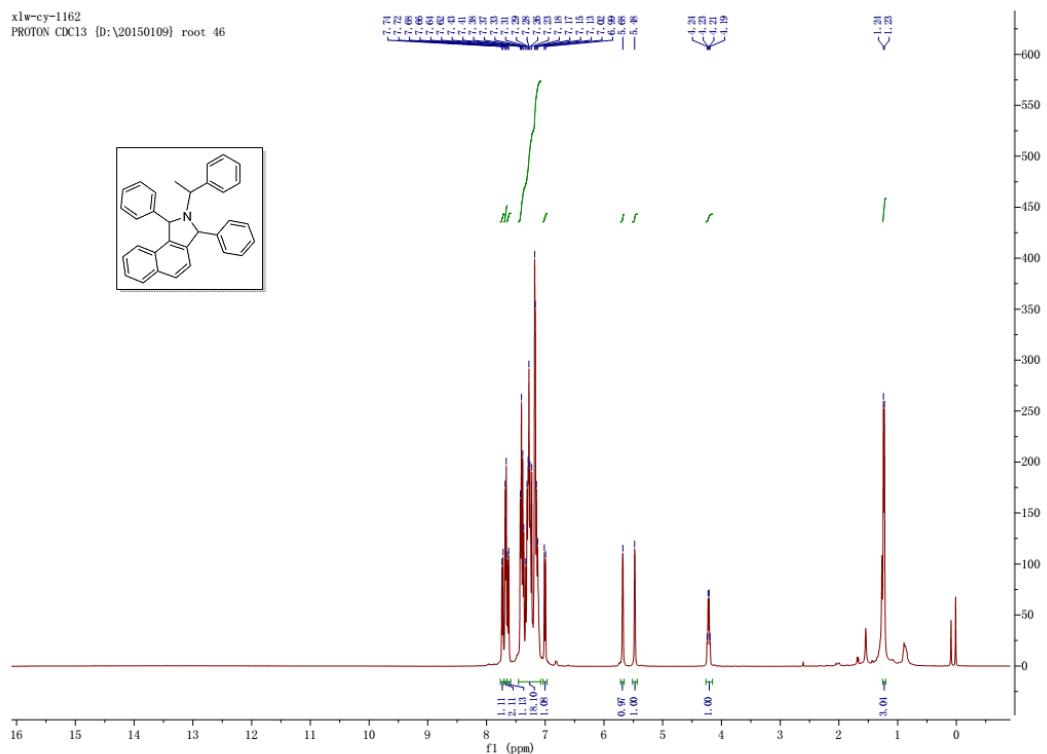
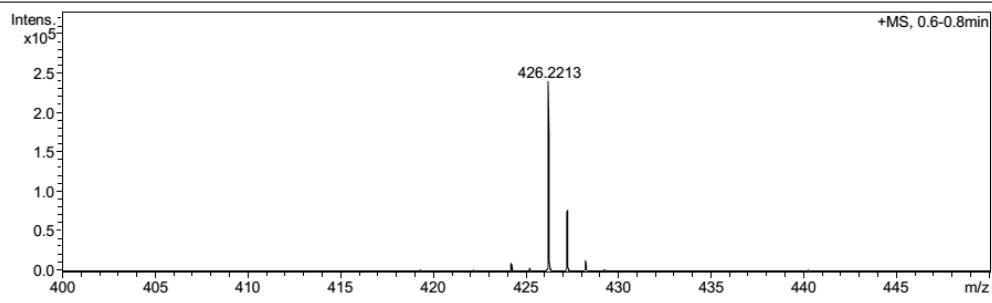
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Method tune-posAPCI-100-700\_140930.m  
Sample Name CY-2  
Comment

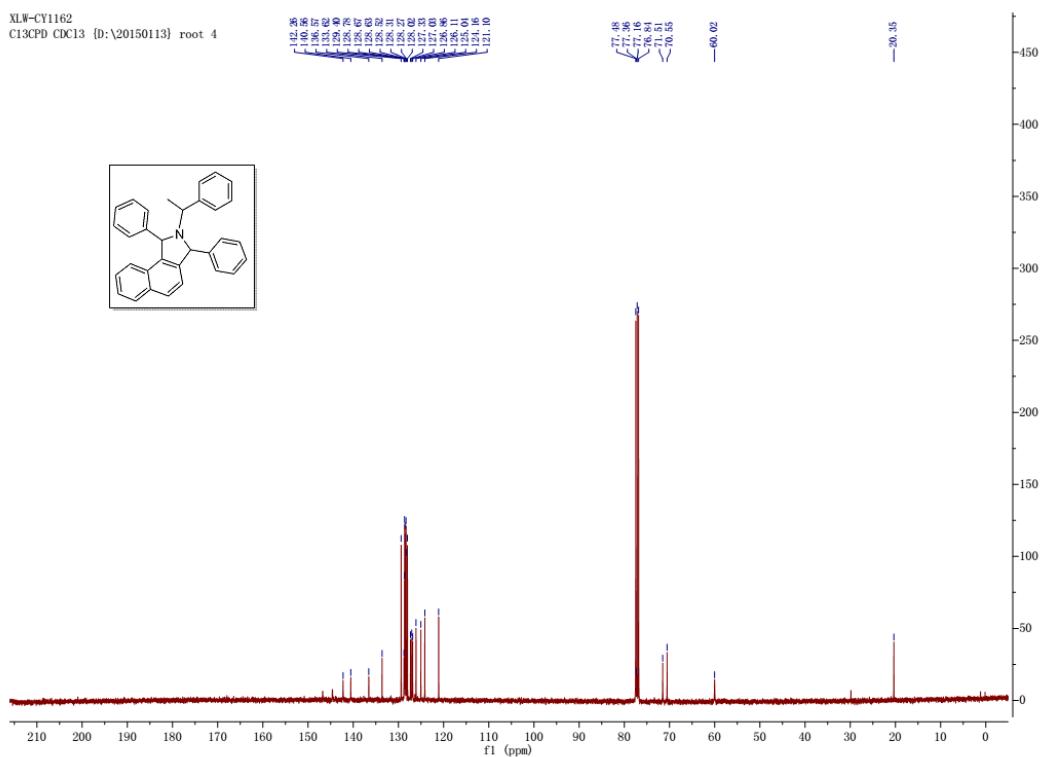
Acquisition Date 10/22/2014 6:00:05 PM

Operator Jiang  
Instrument / Ser# micrOTOF-Q II 10324

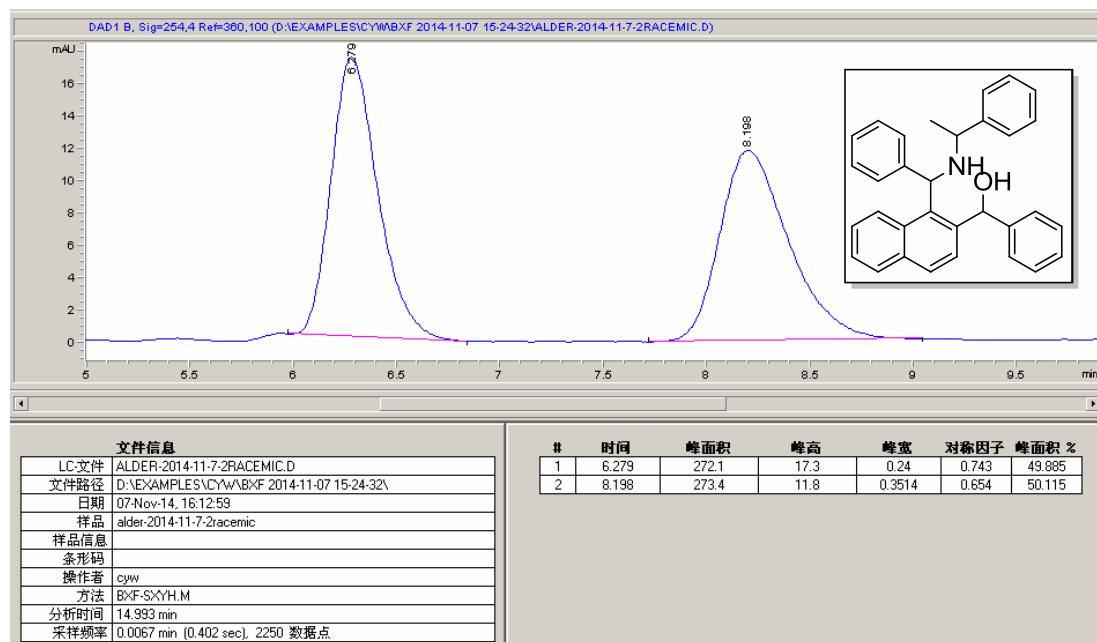
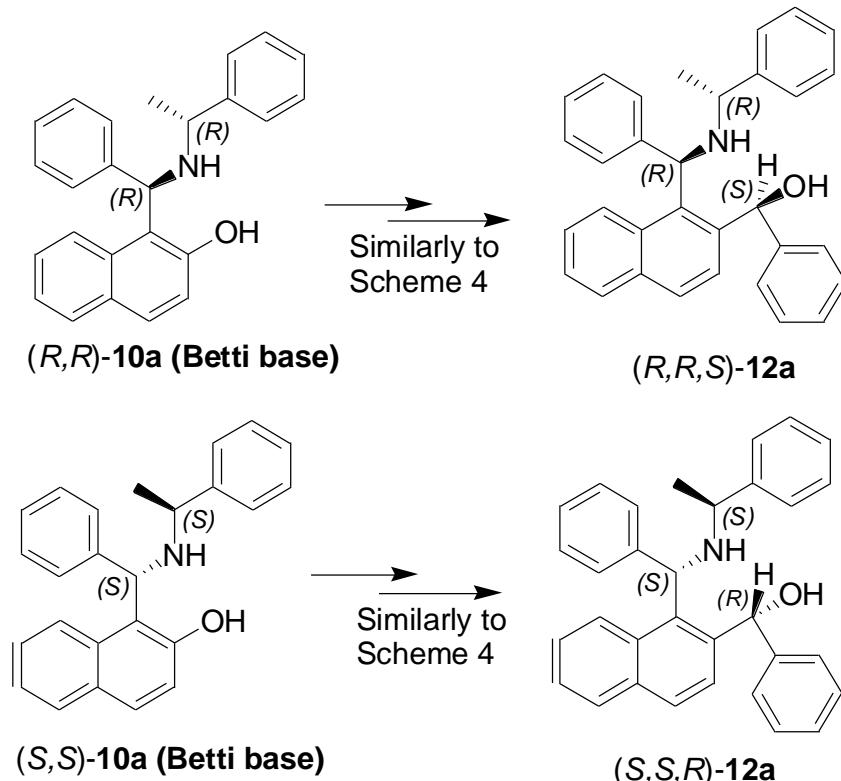
### Acquisition Parameter

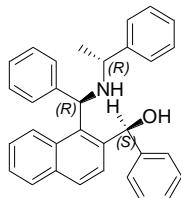
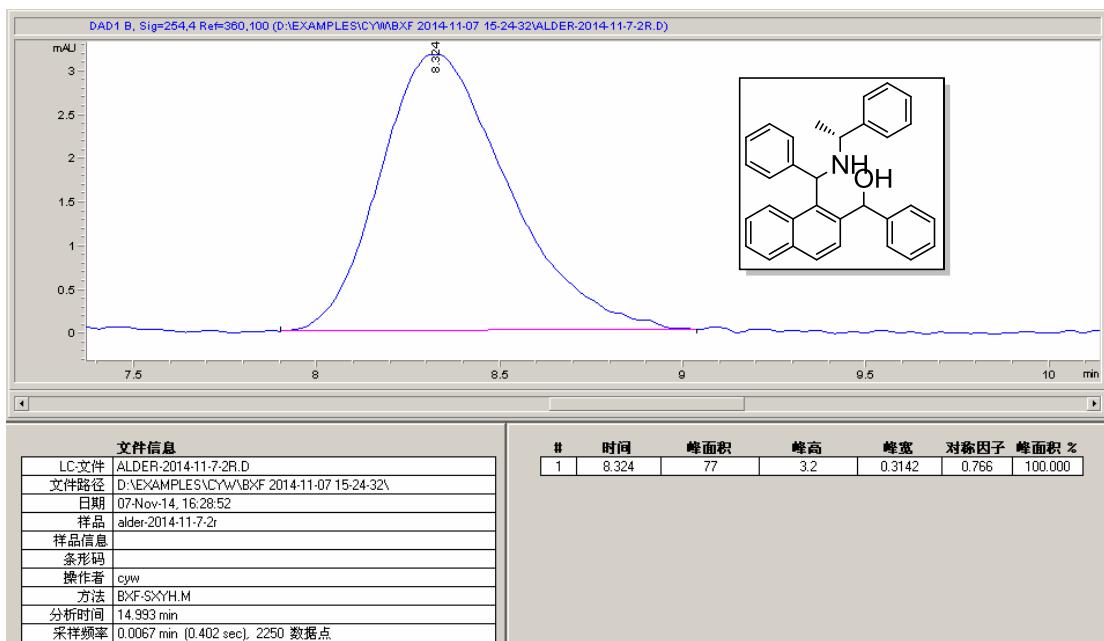
Source Type APCI  
Focus Active  
Scan Begin 100 m/z  
Scan End 1000 m/z  
Ion Polarity Set Capillary  
Set End Plate Offset -500 V  
Set Collision Cell RF 120.0 Vpp  
Positive 4500 V  
Set Nebulizer 1.0 Bar  
Set Dry Heater 200 °C  
Set Dry Gas 1.5 l/min  
Set Divert Valve Waste



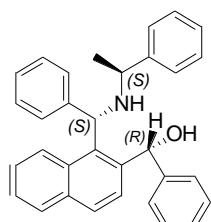
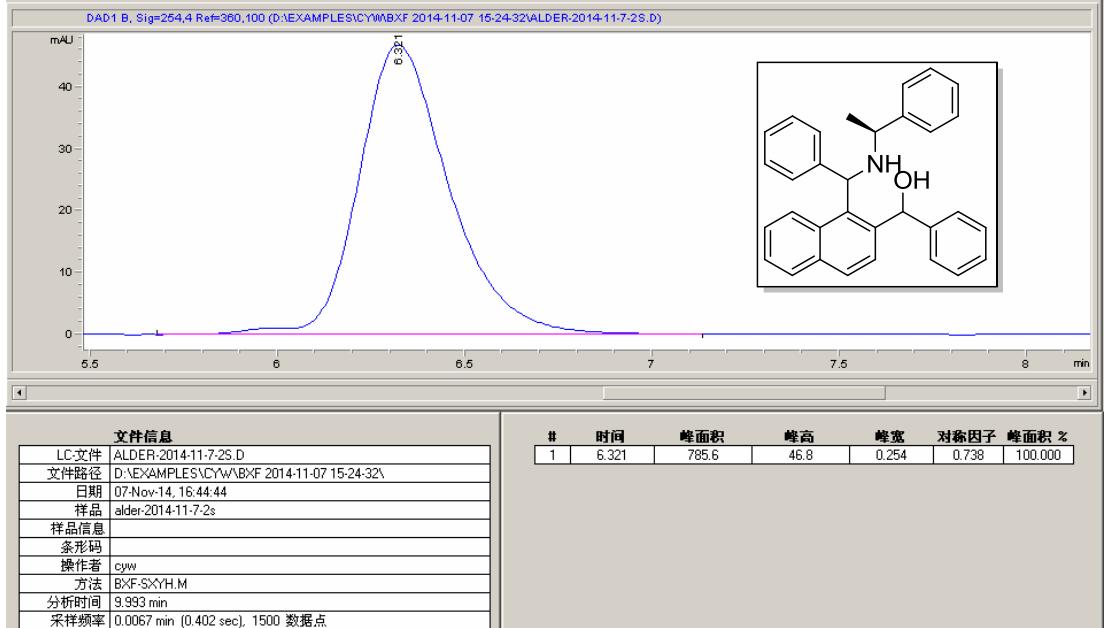


## 8. HPLC of chiral Betti base-derived amino alcohols 12a and all-substituted pyrrolidine derivative 13





(R,R,S)-12a



(S,S,R)-12a

