

## Synthesis of MacMiellan catalyst modified with ionic liquids as a recoverable catalyst for asymmetric Diels Alder reaction

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## 1. Synthetic procedure and characterization data of compound 2-4

### L-phenylalanine methyl ester hydrochloride (2) [1]

Thionyl chloride (2.2 mL, 30 mmol) was added dropwise in the ice cooled suspension of L-Phenyl alanine (4.125 g, 25 mmol) in methanol (32 mL) over a period of 15 min. The clear solution was stirred at room temperature for 28 h. Solvent was removed in vacuo and Et<sub>2</sub>O (20 mL) was added for slurried the solid residue, filtered by vacuum and washed with Et<sub>2</sub>O (10 mL) and dried to give L-phenylalanine methyl ester hydrochloride **2** as a white solid (4.0 g, 97%). mp = 159 °C;  $[\alpha]_D^{25} = + 16.7$  (*c* 1.0, MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 7.01–6.94 (m, 5H), 3.92–3.87 (m, 1H), 3.41–3.37 (m, 3H), 3.11–2.96 (m, 2H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) 168.7, 133.7, 128.9 (2C), 128.09 (2C), 126.8, 53.6, 52.0, 35.6 ppm.

### (S)-2-amino-N-(2-hydroxyethyl)-3-phenylpropanamide (3) [1]

Compound **2** (3.46 g, 15 mmol) was dissolved in ethanolamine (5.5 mL, 92.25 mmol) and allowed to stir at room temperature for 26 h. The reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (25 mL), washed with K<sub>2</sub>CO<sub>3</sub> (20% in 20 mL water). The aqueous phase was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3×15 mL), and the combined organic phase was dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and evaporated by rotavapor to give compound **3** (2.62 g, 84%) as a white solid. mp = 76 °C;  $[\alpha]_D^{25} = + 11.5$  (*c* 1.0, MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 7.66 (t, 3H, *J* = 6.22 Hz), 7.25–7.12 (m, 5H), 3.60–3.50 (m, 3H), 3.35–3.30 (m, 2H), 3.15–3.09 (m, 1H), 2.73 (brs, 2H), 2.66–2.60 (m, 1H) ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) 174.9, 137.4, 129.0 (2C), 128.4 (2C), 126.5, 60.9, 56.2, 41.7, 40.8 ppm.

### (S)-5-benzyl-3-(2-hydroxyethyl)-2,2-dimethylimidazolidin-4-one hydrochloride (4) [1]

In a 100 mL round bottom flask fitted with Dean Stark apparatus, L-phenylalanyl aminoethanol (2.49 g, 12 mmol), acetone (13 mL) and *i*-PrOH (17 mL) were taken and *p*-TsOH · H<sub>2</sub>O (0.07 g, 0.37 mmol) was added in the mixture. The resulting solution was stirred at 85 °C for 5 h. The liquid collected in the Dean-Stark trap was discarded. The solvent in the round bottom flask was removed under vacuum to give light brown oil. The oil was diluted with MeOH (1.5 mL) and an ice-cold methanolic HCl solution (prepared from 1.3 mL of acetyl chloride was added to ice-cooled 5 mL Methanol.) was added slowly. Later, Et<sub>2</sub>O (25 mL) was slowly added to give white precipitate and further stirred for 30 min, filtered and washed with Et<sub>2</sub>O (7 mL). The white solid dried to give (*5R*)-5-benzyl-2,2-dimethyl-3-(2-hydroxyethyl)-imidazolidin-4-one hydrochloride **4** mp = 150.2 °C;  $[\alpha]_D^{25} = - 92.2$  (*c* 1.2 MeOH); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400MHz) δ 7.27–7.24 (m, 2H), 7.20–7.17 (m, 3H), 3.78 (t, 1H, *J* = 5.13 Hz), 3.68–3.61 (m, 2H), 3.40–3.34 (m, 1H), 3.14–3.08 (m, 1H), 3.03 (t, 2H), 1.23 (s, 3H), 1.09 (s, 3H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) 175.4, 136.5, 129.3 (2C), 128.4 (2C), 126.7, 76.4, 61.6, 58.5, 43.6, 36.8, 27.5, 26.2 ppm.

### 5-Bromopentanoyl chloride

5-Bromo valleric acid (5 g, 27.62 mmol) was added to neat SOCl<sub>2</sub> (10.67 mL, 147.79 mmol) and stirred at room temperature for 30 min, then at 50 °C for 1 h. The excess SOCl<sub>2</sub> was evaporated under reduce pressure to give 5-Bromopentanoyl chloride (5.16 g, 94% yield). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400MHz) δ 3.40–3.34 (2H, m), 2.94–2.88 (2H, m), 1.89–1.84 (4H, m) ppm. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100MHz) 173.2, 45.9, 32.4, 30.9, 23.4 ppm.

### Determination of Leaching of catalyst during catalyst recovery

The **IL 8** (10 mg) was dissolved in 5 mL volumetric flask in acetonitrile. Different concentrations (40  $\mu$ L, 20  $\mu$ L, 10  $\mu$ L and 5  $\mu$ L) were injected to the HPLC. The area of different concentration of catalyst **8** calculated by HPLC using C18 column. After the catalytic reaction, the residue was washed with hexane/diethyl ether (1/1). Solvent was evaporated and the crude compound was make up to 5 mL acetonitrile and checked the quantity of leaching of catalyst **8** by injecting 20  $\mu$ L quantity of the crude mixture. The area of leached catalyst was determined by using reference area of **IL 8** at (2.5 mg concentration, area 261971)

Table 1. Determination of leaching of the catalyst

Entry	Cycle	Leached catalyst (mg)	Area (mv)
1	0	0.12	13478
2	1	0.1	11708
3	2	0.0	0
4	3	0.01	1480
5	4	0.02	2361
6	5	0.02	2365

## 2. $^1\text{H}$ and $^{13}\text{C}$ NMR data of catalytic products

### (1S,2S,3S,4R)-3-phenylbicyclo[2.2.1]hept-5-ene-2-carbaldehyde (**14a**) and (1R,2S,3S,4S)-3-phenylbicyclo[2.2.1]hept-5-ene-2-carbaldehyde (**15a**) (*exo/endo* : 1.2/1)

The products were converted to the corresponding alcohols with  $\text{NaBH}_4$  and enantiomeric excess was determined by HPLC using CHIRALPAK OD-H column, *n*-Hex/IPA = 95/5; flow rate 1.0 ml/min; 210 nm; *exo* isomer ( $t_{\text{major}} = 13.7$  min,  $t_{\text{minor}} = 17.8$  min), *endo* isomer ( $t_{\text{major}} = 14.8$  min,  $t_{\text{minor}} = 26.0$  min).

$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400MHz)  $\delta$  9.92 (d, 1.2H,  $J = 2.20$  Hz), 9.60 (d, 1H,  $J = 2.20$  Hz), 7.33–7.14 (m, 10H), 6.43 (dd, 1H,  $J = 5.86, 3.66$  Hz), 6.34 (dd, 1.2H,  $J = 5.86, 3.66$  Hz), 6.18 (dd, 1H,  $J = 5.13, 2.20$  Hz), 6.08 (dd, 1.2H,  $J = 5.13, 2.93$  Hz), 3.73 (t, 1.2H,  $J = 3.66$  Hz), 3.33 (s, 1H), 3.22 (d, 2.4H,  $J = 1.46$  Hz), 3.13 (s, 1H), 3.09 (d, 1H,  $J = 4.39$  Hz) 3.0–2.97 (m, 1H), 2.61–2.59 (m, 1.2H), 1.81 (d, 1.2H,  $J = 8.79$ ), 1.64–1.55 (m, 3H) ppm.  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz) 203.4, 202.7, 143.4, 142.4, 139.1, 136.4, 136.2, 133.6, 128.5 (2C), 128.0 (2C), 127.8 (2C), 127.2 (2C), 126.2, 126.1, 60.7, 59.3, 48.3, 48.2, 47.9, 47.0, 45.5, 45.39, 45.30, 45.0 ppm.

### (1S,2S,3S,4R)-3-(4-chlorophenyl)bicyclo[2.2.1]hept-5-ene-2-carbaldehyde (**14c**) and (1R,2S,3S,4S)-3-(4-chlorophenyl)bicyclo[2.2.1]hept-5-ene-2-carbaldehyde (**15c**) (*exo/endo* : 1.1/1)

The products were converted to the corresponding alcohols with  $\text{NaBH}_4$  and enantiomeric excess was determined by HPLC using CHIRALPAK OD-H column, *n*-Hex/IPA = 96/4; flow rate 0.8 ml/min; 230 nm; *exo* isomer ( $t_{\text{major}} = 17.96$  min,  $t_{\text{minor}} = 13.36$  min), *endo* isomer ( $t_{\text{major}} = 20.01$  min,  $t_{\text{minor}} = 17.96$  min).

$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400MHz)  $\delta$  9.80 (d, 1.1H,  $J = 1.83$  Hz), 9.50 (d, 1H,  $J = 1.83$  Hz), 7.19–7.13 (m, 2.1H), 7.12–7.08 (m, 4.1H), 7.0–6.96 (m, 2H), 6.32 (dd, 1H,  $J = 3.89, 3.21$  Hz), 6.26 (dd, 1.1H,  $J = 3.89, 3.21$  Hz), 6.08 (dd, 1H,  $J = 4.12, 2.75$  Hz), 5.95 (dd, 1.1H,  $J = 4.12, 2.75$  Hz), 3.62–3.60 (m, 1.1H), 3.26 (brs, 1H),

3.14 (brs, 1H), 3.09 (brs, 1.1H), 3.0 (brs, 1H), 2.99 (d, 1H,  $J = 5.50$  Hz), 2.83–2.81 (m, 1.1H), 2.45–2.44 (m, 1.1H), 1.66 (d, 1.1H,  $J = 8.70$  Hz), 1.56–1.45(m, 3H) ppm.  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz) 202.9, 202.2, 142.0, 141.0, 139.0, 136.4, 136.2, 133.7, 131.9, 131.8, 129.1 (2C), 128.6 (2C), 128.5 (2C), 128.1 (2C), 60.9, 59.4, 48.3, 48.1, 47.4, 46.9, 45.3, 45.0, 44.9, 44.6 ppm.

**(1S,2S,3S,4R)-3-(4-methoxyphenyl)bicyclo[2.2.1]hept-5-ene-2-carbaldehyde (14d) and (1R,2S,3S,4S)-3-(4-methoxyphenyl)bicyclo[2.2.1]hept-5-ene-2-carbaldehyde (15d)** (*exo/endo* : 1.1/1)

The products were converted to the corresponding alcohols with  $\text{NaBH}_4$  and enantiomeric excess was determined by HPLC using CHIRALPAK OD-H column, *n*-Hex/IPA = 95/5; flow rate 0.8 ml/min; 230 nm; *exo* isomer ( $t_{\text{major}} = 20.72$  min,  $t_{\text{minor}} = 15.62$  min).

$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400MHz)  $\delta$  9.88 (d, 1.1H,  $J = 2.29$  Hz), 9.55 (d, 1H,  $J = 2.29$  Hz), 7.16 (d, 2H,  $J = 8.70$  Hz), 7.04 (d, 2.2H,  $J = 8.70$  Hz), 6.83 (d, 2H,  $J = 8.70$  Hz), 6.77 (d, 2.2H,  $J = 8.70$  Hz), 6.39 (dd, 1H,  $J = 5.50, 3.21$  Hz), 6.31 (dd, 1.1H,  $J = 5.50, 3.21$  Hz), 6.14 (dd, 1H,  $J = 5.50, 2.75$  Hz), 6.04 (dd, 1.1H,  $J = 5.50, 2.75$  Hz), 3.77 (s, 3H), 3.75 (s, 3.1H), 3.63 (t, 1.1H,  $J = 5.04$  Hz), 3.29 (s, 1H), 3.16 (d, 2.2H,  $J = 13.74$  Hz), 3.04 (s, 1H), 3.0 (d, 1H,  $J = 4.58$  Hz), 2.91–2.90 (m, 1.2H), 2.51–2.50 (m, 1.2H), 1.77 (d, 1H,  $J = 8.70$  Hz), 1.61–1.56 (m, 3H) ppm.  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz) 203.7, 202.9, 158.0, 157.9, 139.2, 136.5, 136.2, 135.5, 134.5, 133.6, 128.7 (2C), 128.2 (2C), 113.9 (2C), 113.4 (2C), 60.8, 59.6, 55.25, 55.20, 48.6, 48.5, 47.5, 47.0, 45.4, 45.08, 45.0, 44.6 ppm.

**(1S,2S,3S,4R)-3-(4-nitrophenyl)bicyclo[2.2.1]hept-5-ene-2-carbaldehyde (14b) and (1R,2S,3S,4S)-3-(4-nitrophenyl)bicyclo[2.2.1]hept-5-ene-2-carbaldehyde (15b)** (*exo/endo* : 1.2/1)

The products were converted to the corresponding alcohols with  $\text{NaBH}_4$  and enantiomeric excess was determined by HPLC using CHIRALPAK AD-H column, *n*-Hex/IPA = 95/5; flow rate 1.0 ml/min; 230 nm; *exo* isomer ( $t_{\text{major}} = 40.40$  min,  $t_{\text{minor}} = 42.49$  min), *endo* isomer ( $t_{\text{major}} = 44.47$  min,  $t_{\text{minor}} = 57.50$  min).

$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400MHz)  $\delta$  9.83 (d, 1.2H,  $J = 1.83$  Hz), 9.56 (d, 1H,  $J = 1.83$  Hz), 8.06 (d, 2H,  $J = 8.70$  Hz), 8.0 (d, 2.2H,  $J = 8.70$  Hz), 7.34 (d, 2H,  $J = 8.70$  Hz), 7.21 (d, 2.2H,  $J = 8.70$  Hz), 6.35 (dd, 1H,  $J = 5.50, 3.21$  Hz), 6.32 (dd, 1.2H,  $J = 5.50, 3.21$  Hz), 6.12 (dd, 1H,  $J = 5.50, 2.75$  Hz), 5.96 (dd, 1.1H,  $J = 5.50, 2.75$  Hz), 3.80 (t, 1H,  $J = 3.89$  Hz), 3.35 (s, 1H), 3.22 (m, 1.2H), 3.17 (s, 1.2H), 3.13–3.10 (m, 2H), 2.89–2.87 (m, 1H), 2.55 (d, 1.2H,  $J = 4.58$  Hz), 1.70–1.67 (m, 1H), 1.63–1.60 (m, 1H), 1.53 (brs, 2.2H) ppm.  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz) 202.1, 201.5, 151.5, 150.5, 146.3, 146.2, 138.9, 136.8, 135.8, 133.8, 128.5 (2C), 128.1 (2C), 123.6 (2C), 123.2 (2C), 61.03, 59.4, 48.3, 47.8, 47.5, 47.03, 45.4, 45.3, 45.02, 44.8 ppm.

**(1S,2R,3S,4R)-3-methylbicyclo[2.2.1]hept-5-ene-2-carbaldehyde (11) and (1R,2R,3S,4S)-3-methylbicyclo[2.2.1]hept-5-ene-2-carbaldehyde (12)** (*exo/endo* : 1/1.2)

Prepared according to the general procedure with (*E*)-crotonaldehyde. The enantiomeric excess was determined by GC using  $\beta$ -DEX chiral column.  $^1\text{H}$  NMR ( $\text{C}_6\text{H}_6$ , 400MHz)  $\delta$  9.43 (brs, 1H), 9.17 (d, 1.2H,  $J = 1.91$  Hz), 5.97–5.94 (m, 1.2H), 5.88–5.85 (m, 2H), 5.77–5.76 (m, 1.2H), 2.68 (brs, 1H), 2.62 (brs, 1H), 2.38 (s, 1H), 2.16–2.12 (m, 2.2H), 1.90–1.88 (m, 1H), 1.60–1.55 (m, 1.2H), 1.33 (brs, 1H), 1.26–1.18 (m, 3H), 1.13 (d, 1.2H,  $J = 8.39$  Hz), 0.84 (d, 3.2H,  $J = 6.10$  Hz), 0.65 (d, 3H,  $J = 6.87$  Hz) ppm.  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz) 202.5, 201.7, 138.6, 136.4, 135.6, 132.7, 61.2, 59.9, 49.0, 47.6, 47.5, 45.9, 45.2, 45.1, 36.0, 35.4, 20.7, 18.7.

## References

- [1] T. O. Kristense, K. Vestli, M. G. Jakobsen, F. K. Hansen, T. Hansen, *J. Org. Chem.* 75 (2010) 162.

# Mass Spectra of Compounds

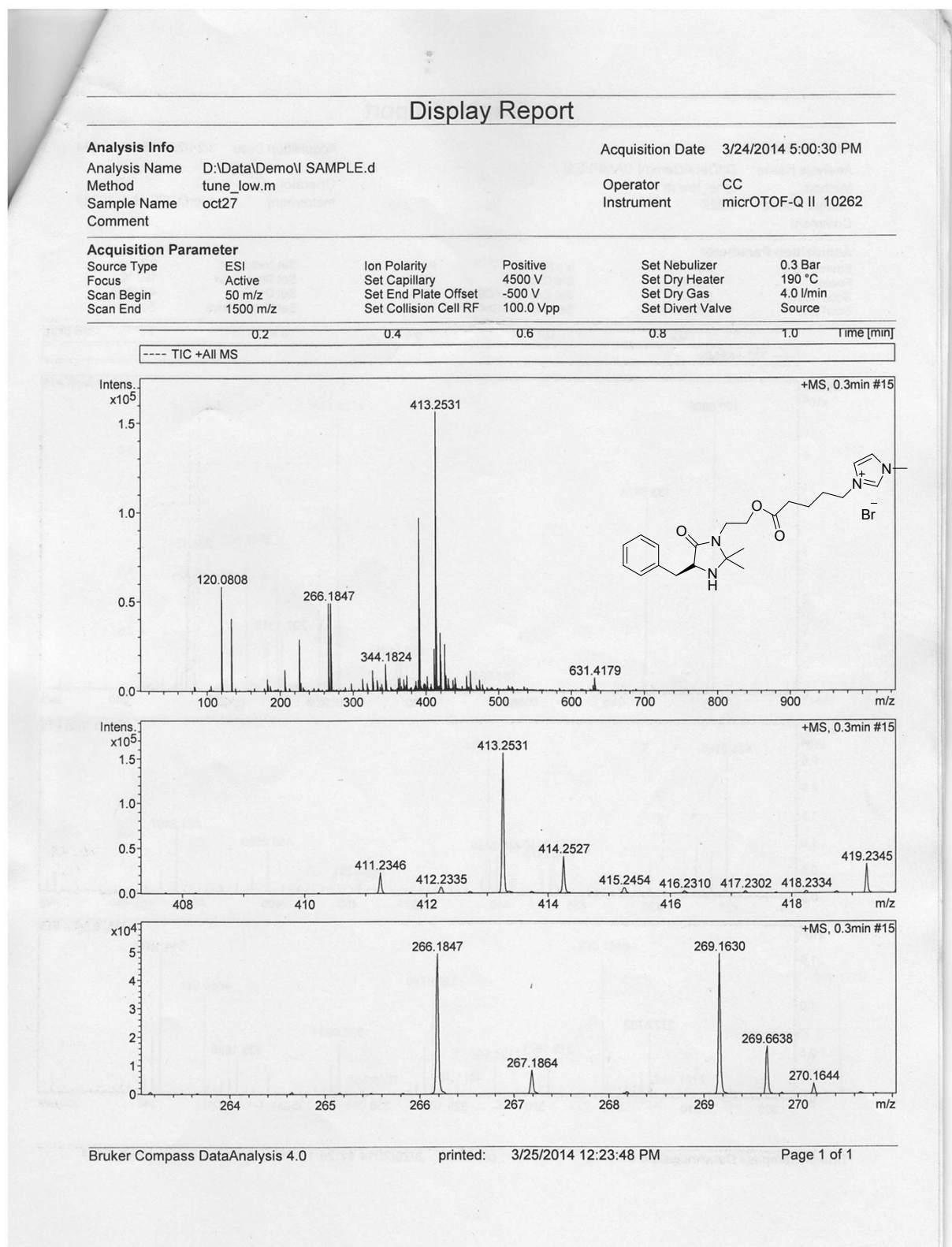


Figure 1 : HRMS data of IL 6.

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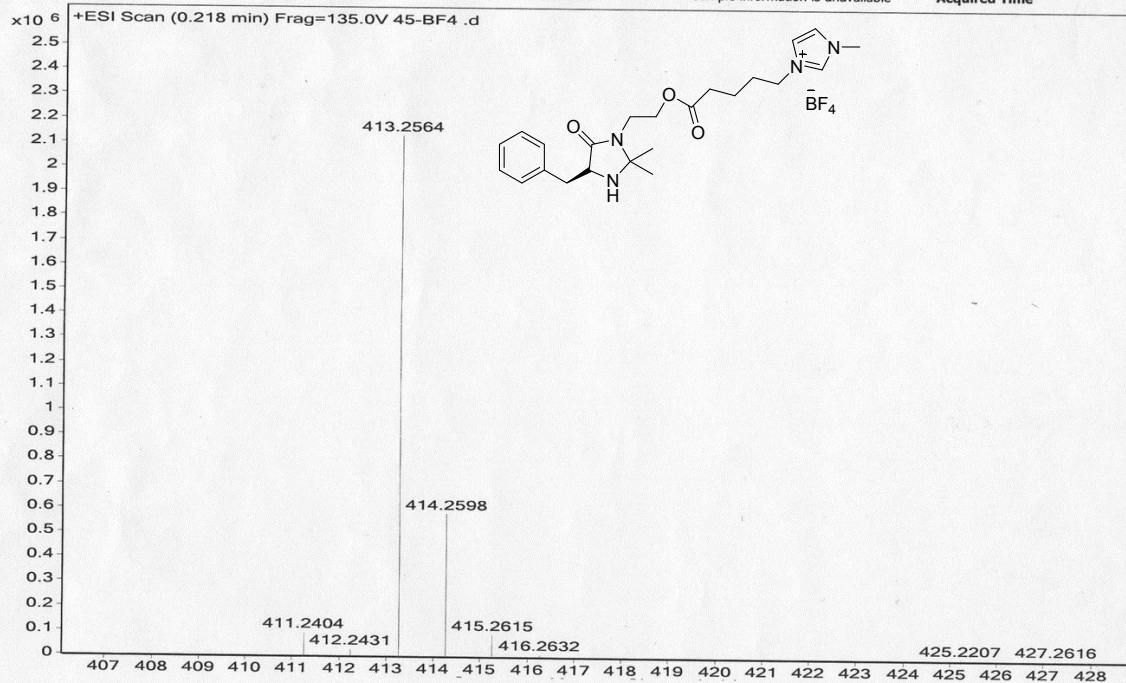


Figure 2 : HRMS data of IL 7.

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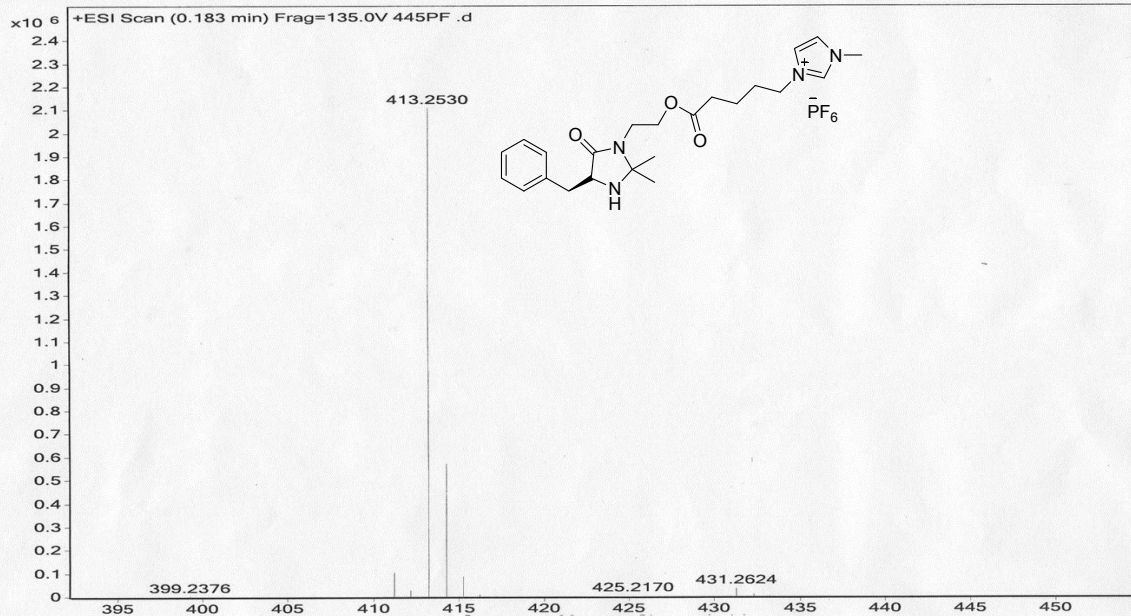


Figure 3 : HRMS data of IL 8.



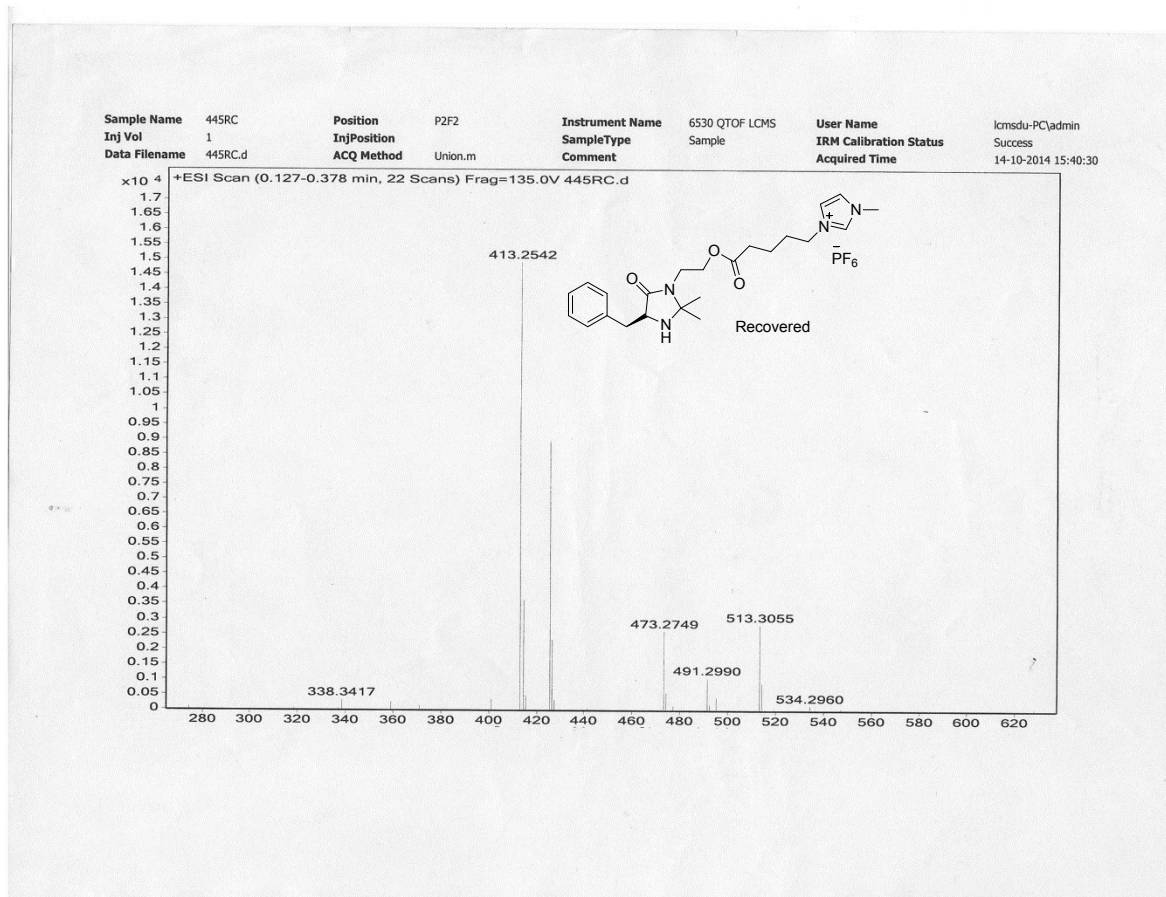


Figure 4 : HRMS data of recovered IL 8 after catalytic reaction.



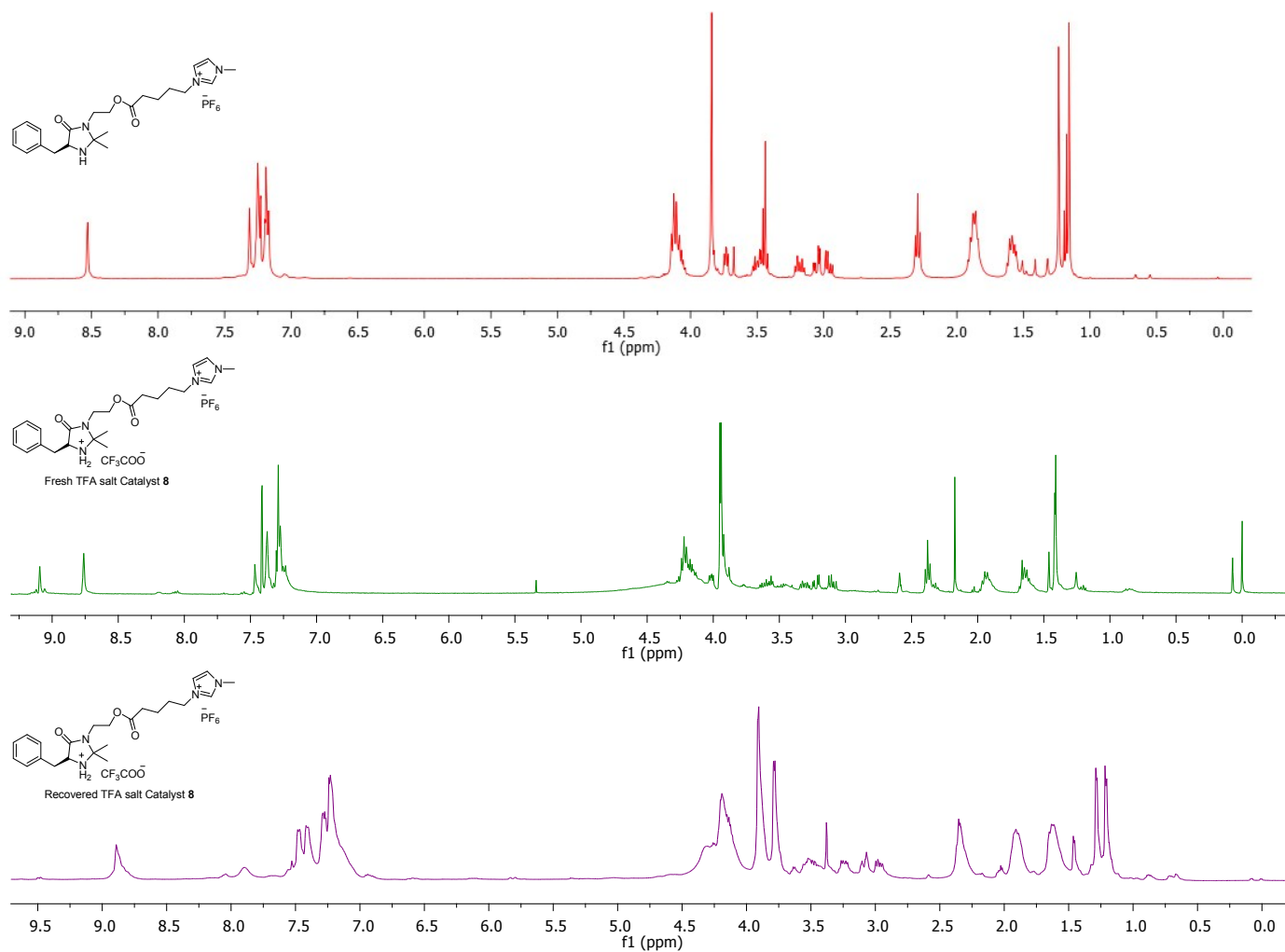


Figure 5.  $^1\text{H}$  and fresh ionic liquids, its TFA salt and recovered TFA salt

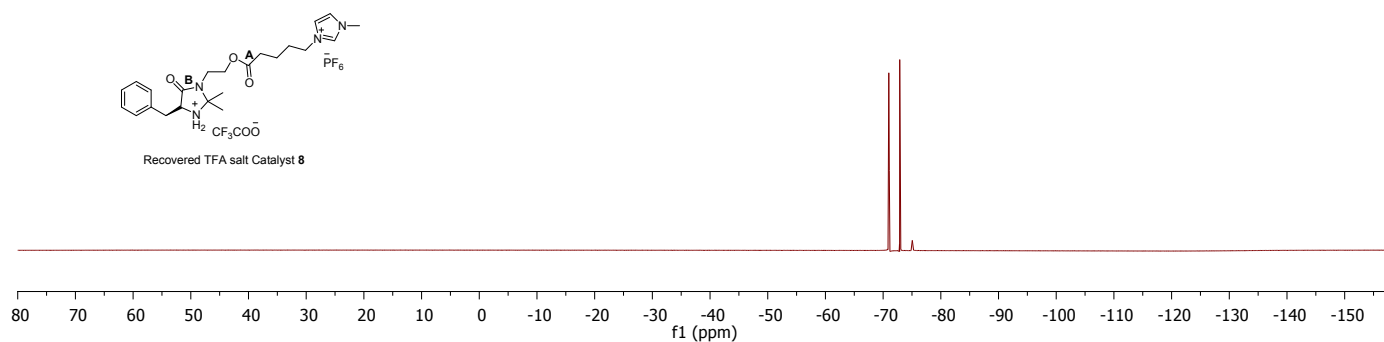
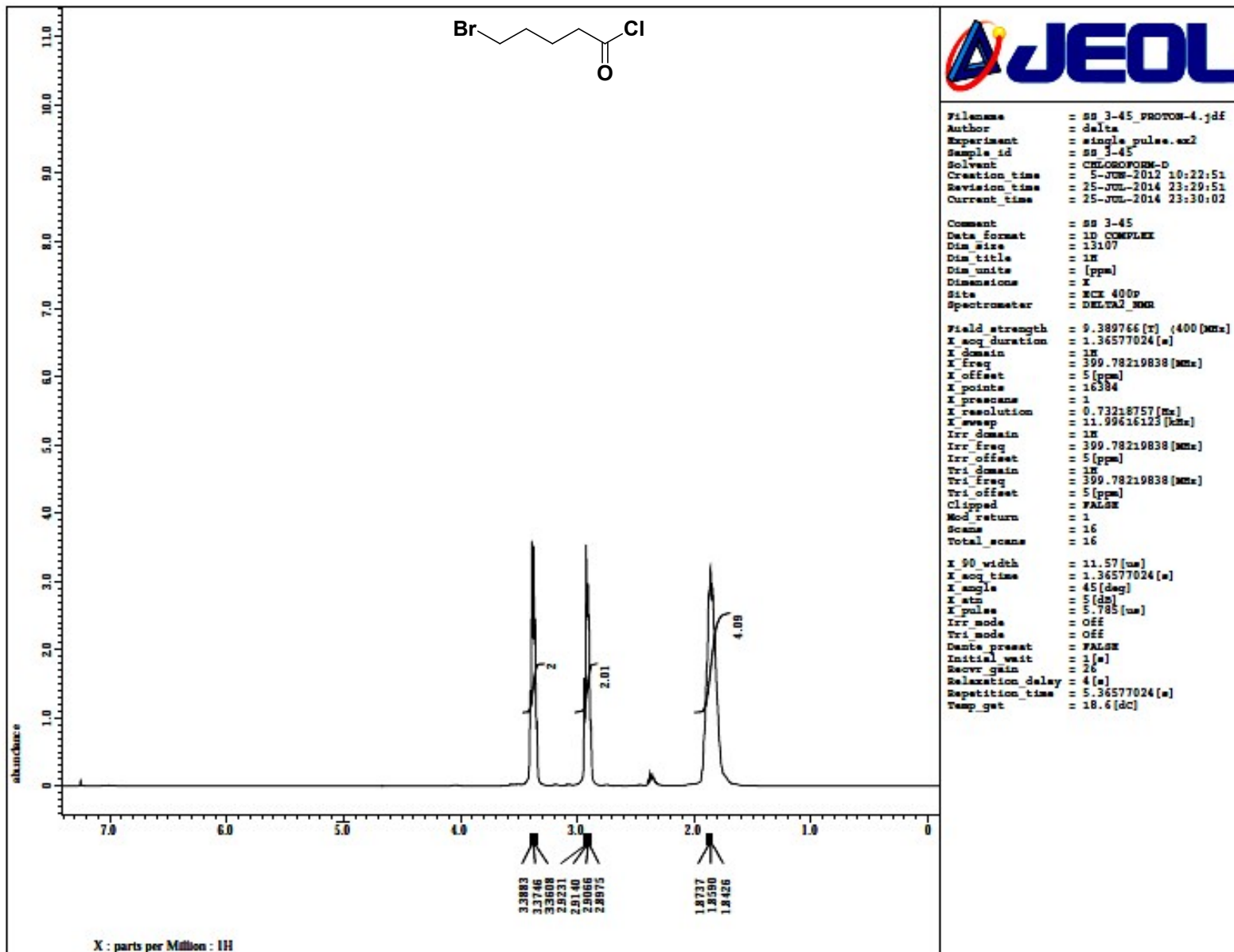
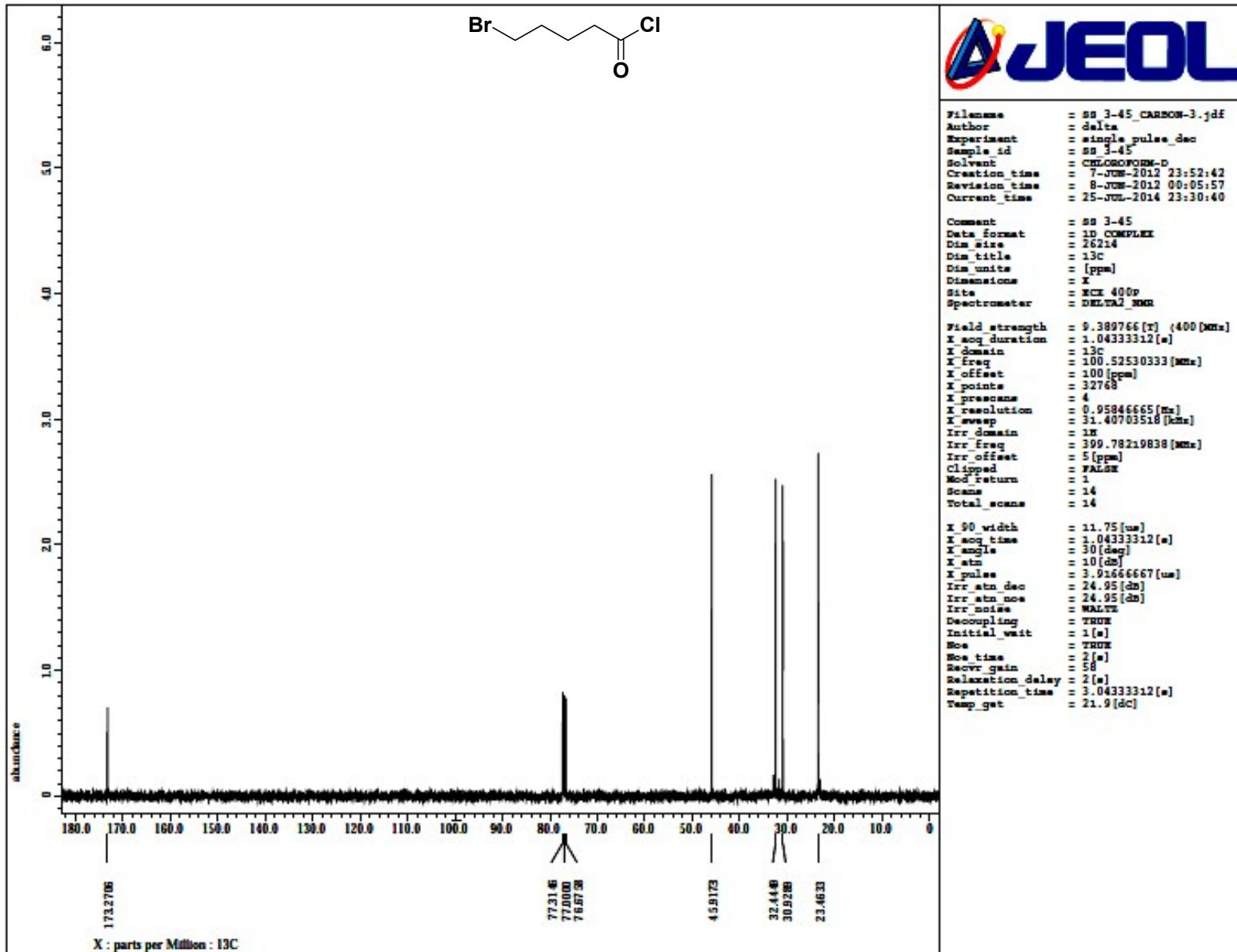
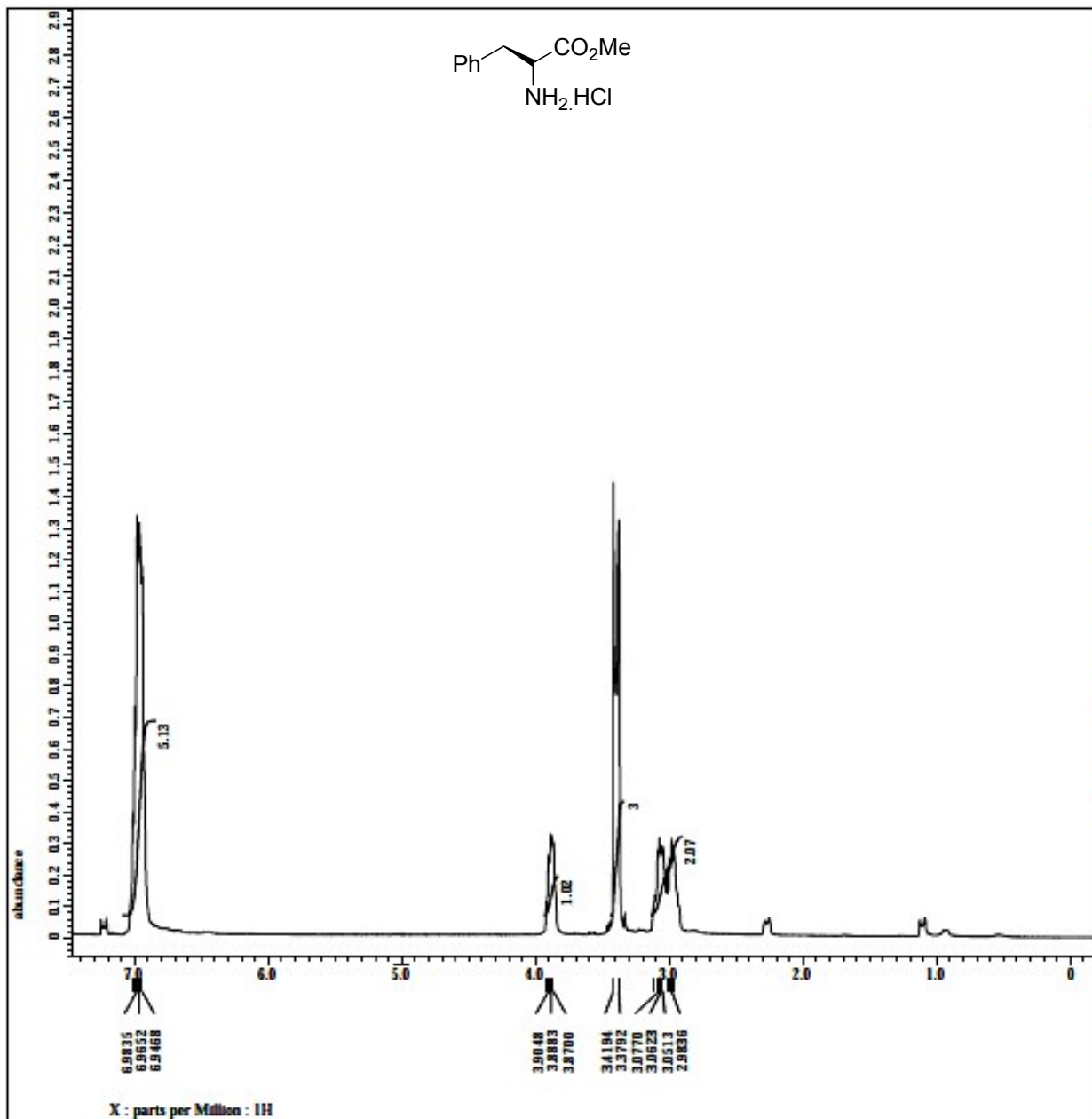
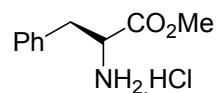


Figure 6 :  $^{19}\text{F}$  NMR of recovered IL 8.TFA salt.





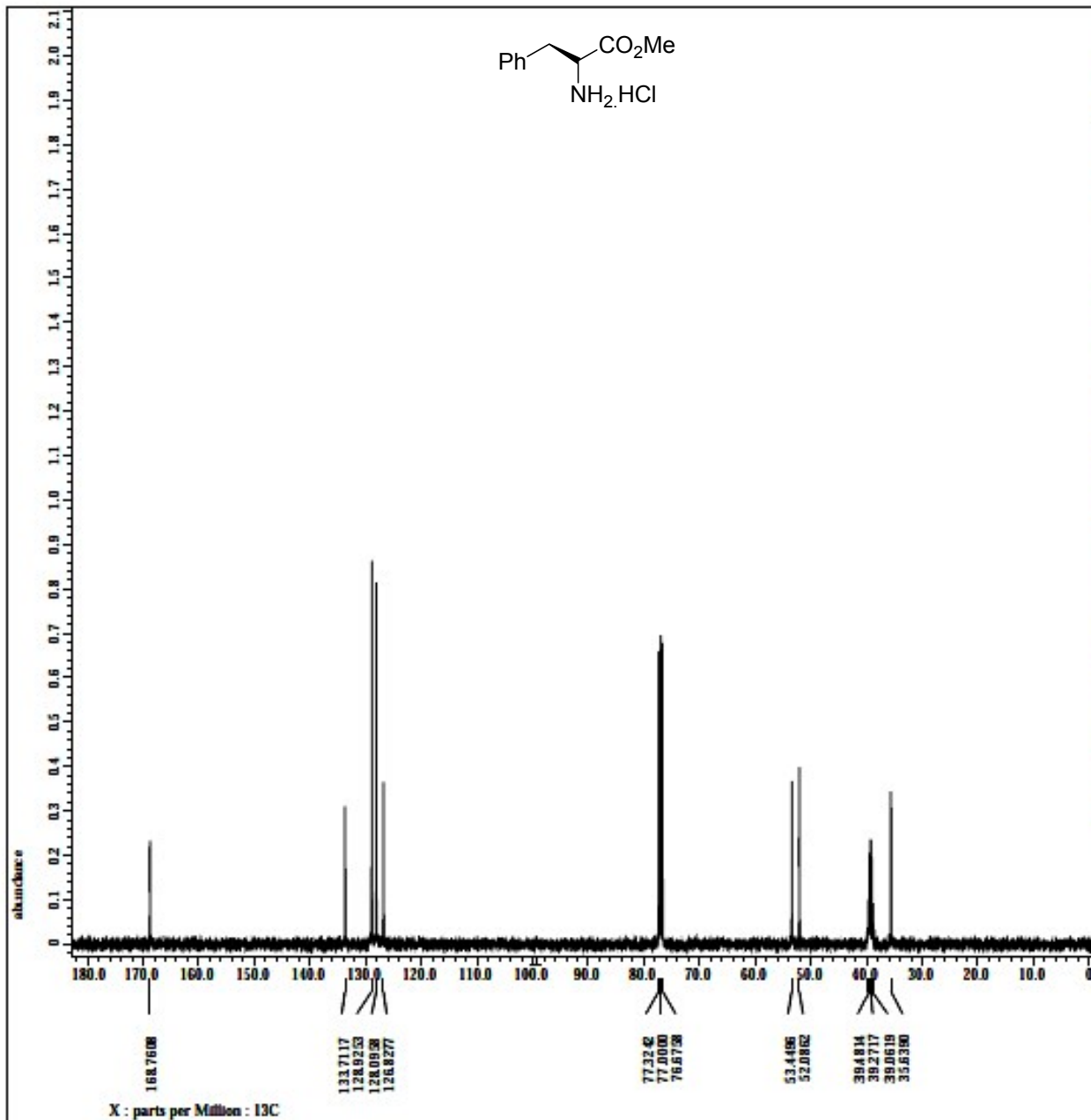
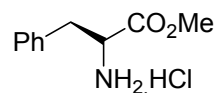


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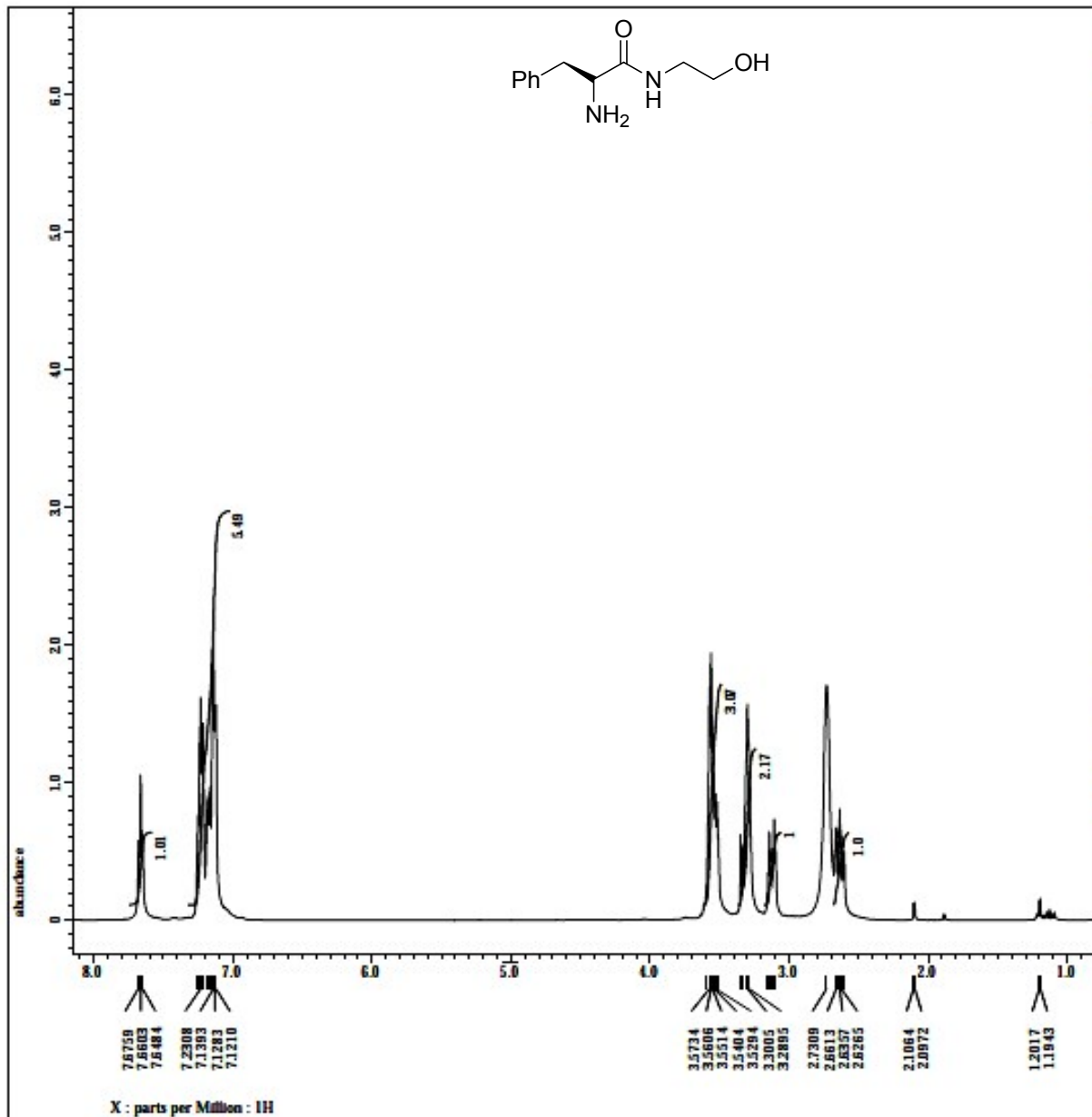
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Temp get       = 23.4 [dc]
  
```



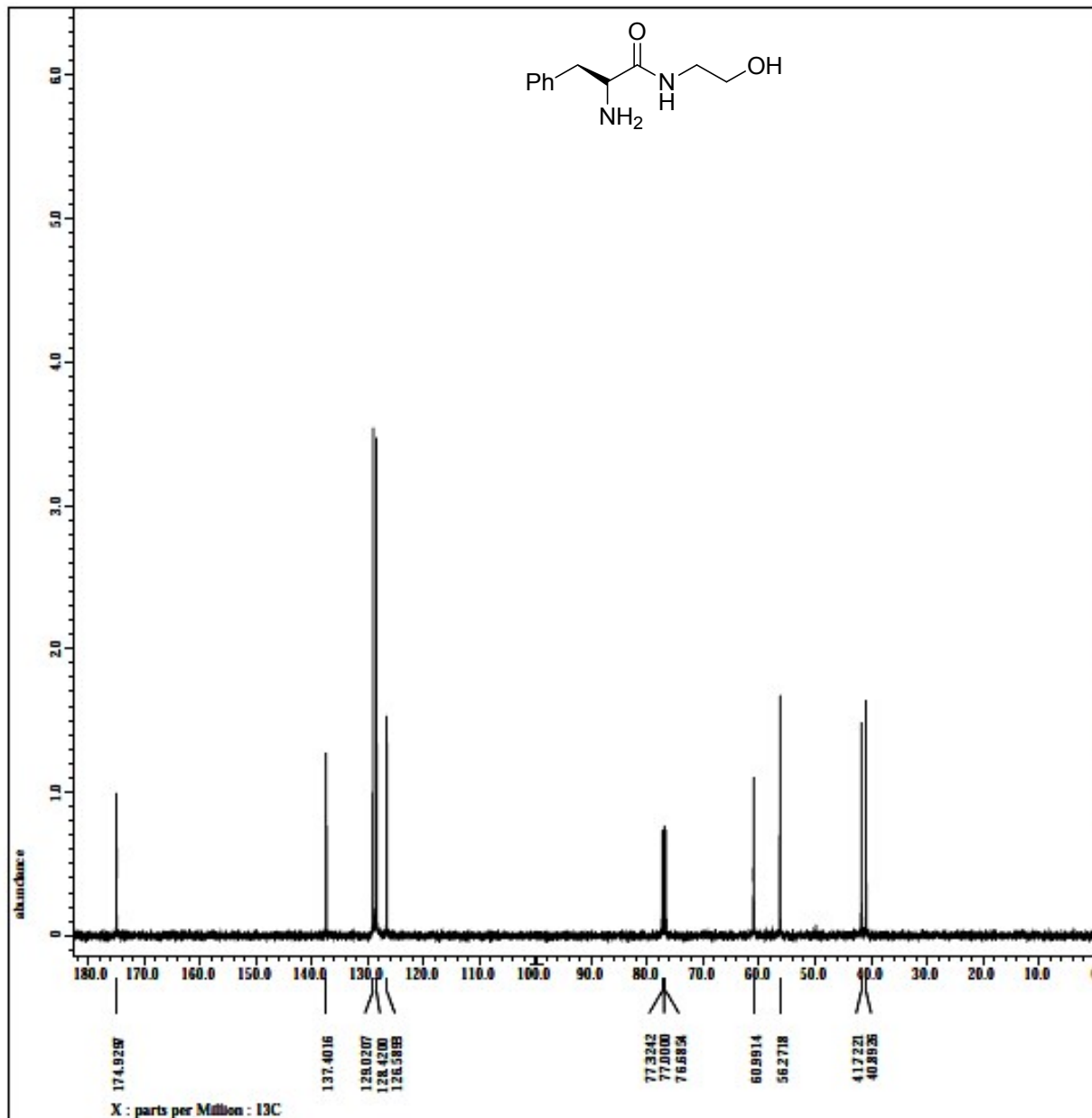
Filename = FH 4-33 N-1\_PROTON-4.  
 Author = delta  
 Experiment = single pulse.ex2  
 Sample id = FH 4-33 N-1  
 Solvent = CHLOROFORM-D  
 Creation time = 11-MAR-2013 11:40:05  
 Revision time = 27-JUL-2014 22:20:44  
 Current time = 27-JUL-2014 22:20:53

Comment = FH 4-33 N-1  
 Data format = 1D COMPLEX  
 Dir\_size = 13107  
 Dir\_title = 1H  
 Dir\_units = [ppm]  
 Dimensions = X  
 Site = NCA 400P  
 Spectrometer = DELTA3 NMR

Field strength = 9.389766 [T] (400 [MHz])  
 X acq duration = 1.36577024 [s]  
 X domain = 1H  
 X Freq = 399.78219838 [MHz]  
 X offset = 5 [ppm]  
 X points = 16584  
 X prescans = 1  
 X resolution = 0.73218757 [Hz]  
 X sweep = 11.99616123 [kHz]  
 Irr domain = 1H  
 Irr Freq = 399.78219838 [MHz]  
 Irr offset = 5 [ppm]  
 Tri domain = 1H  
 Tri Freq = 399.78219838 [MHz]  
 Tri offset = 5 [ppm]  
 Clipped = FALSE  
 Mod return = 1  
 Scans = 16  
 Total\_scans = 16

X 90 width = 11.31 [us]  
 X acq time = 1.36577024 [s]  
 X angle = 45 [deg]  
 X atm = 4.5 [dB]  
 X pulse = 5.655 [us]  
 Irr mode = OFF  
 Tri mode = OFF  
 Dents present = FALSE  
 Initial wait = 1 [s]  
 Recvr gain = 22  
 Relaxation delay = 4 [s]  
 Repetition time = 5.36577024 [s]  
 Temp\_get = 22.9 [dc]





```

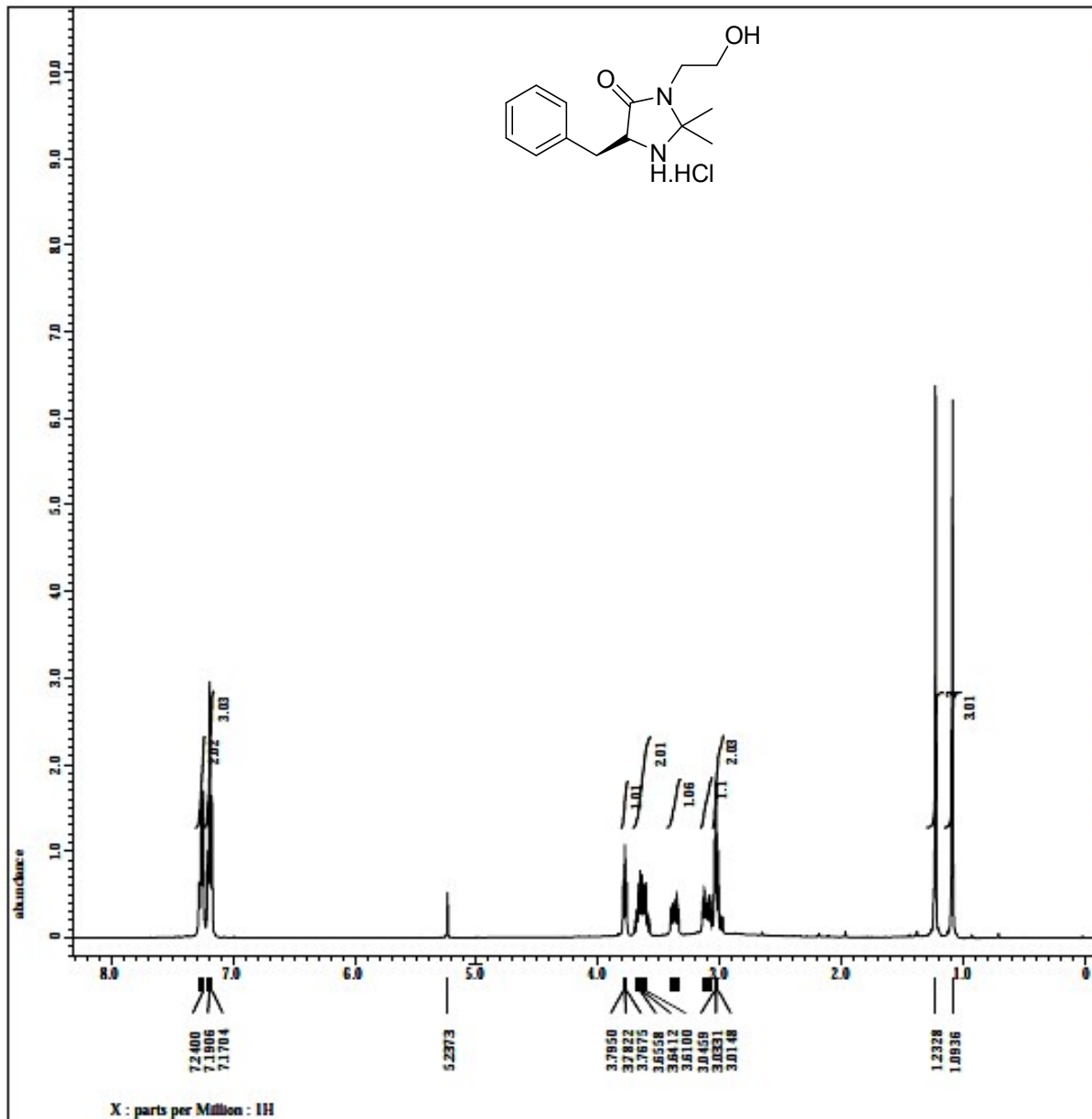
Filename      = SS 4-33 N-1 CARBON-3.
Author       = delta
Experiment   = single pulse dec
Sample id    = SS 4-33 N-1
Solvent      = CHLOROFORM-D
Creation time = 14-MAR-2013 18:50:17
Revision time = 14-MAR-2013 19:18:06
Current time  = 27-JUL-2014 22:21:26

Comment      = SS 4-33 N-1
Data format  = 1D COMPLEX
Dim size     = 26214
Dim title    = 13c
Dim units    = [ppm]
Dimensions   = X
Site         = MCI 400p
Spectrometer = DELTA2 400p

Field strength = 9.389766 [T] (400 [MHz])
X_acq duration = 1.04333312 [s]
X_domain      = 13c
X_freq        = 100.52530333 [MHz]
X_offset      = 100 [ppm]
X_points      = 32768
X_prescans    = 4
X_resolution  = 0.95846665 [Hz]
X_sweep       = 31.40703518 [kHz]
Irr_domain    = 1H
Irr_freq      = 399.78219838 [MHz]
Irr_offset    = 5 [ppm]
Clipped       = FALSE
Mod_return    = 1
Scans         = 450
Total_scans   = 450

X_90 width    = 9.13 [us]
X_acq time    = 1.04333312 [s]
X_angle       = 30 [deg]
X_atn         = 7.8 [dB]
X_pulse       = 3.04333333 [us]
Irr_atn dec   = 25.008 [dB]
Irr_atn noc   = 25.008 [dB]
Irr_noise     = WALTZ
Decoupling    = TRUE
Initial_wait  = 1 [s]
Noe           = TRUE
Noe time      = 2 [s]
Noe_gain      = 60
Relaxation_delay = 2 [s]
Repetition_time = 3.04333312 [s]
Temp_get      = 23 [dc]
  
```



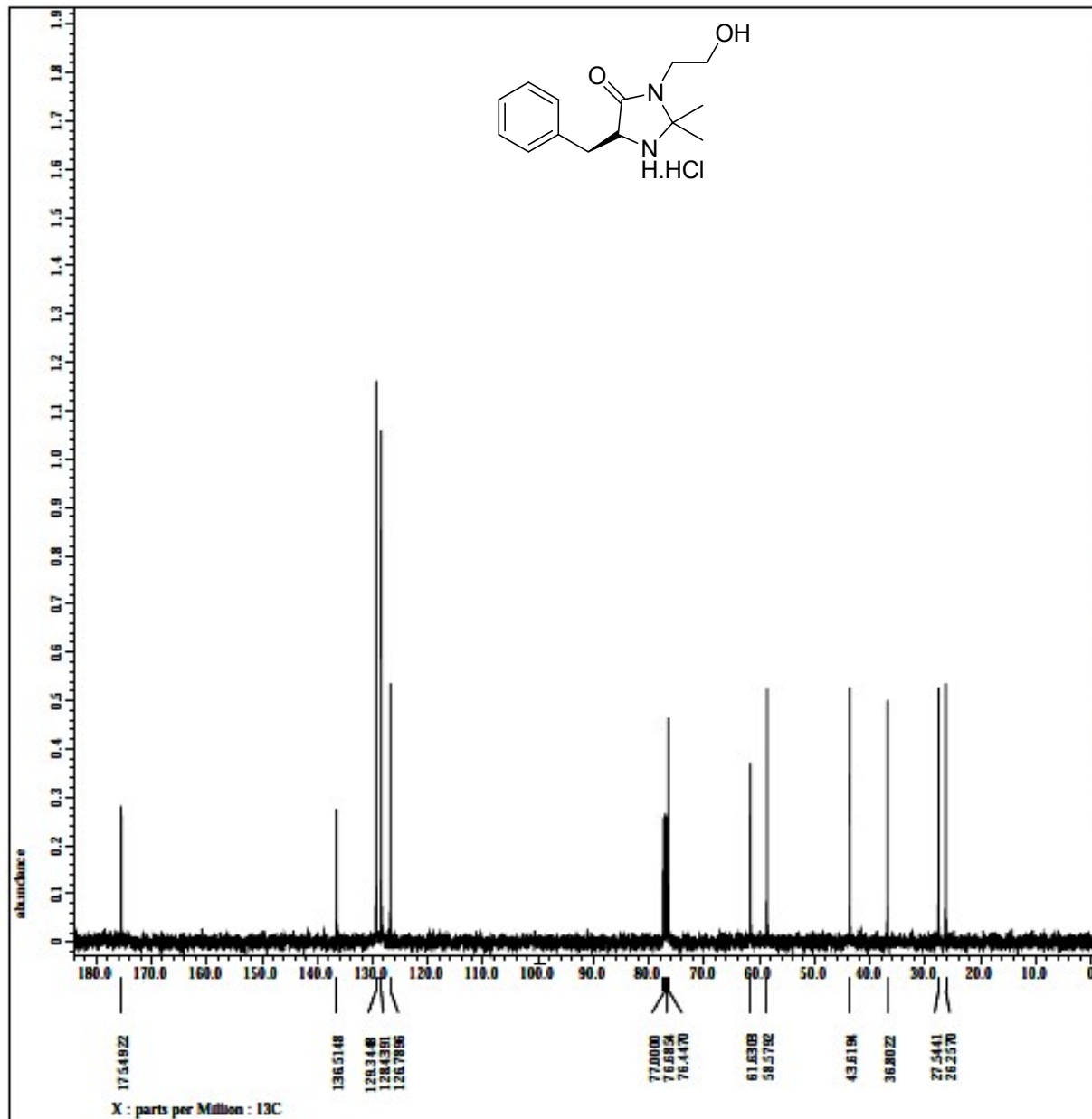


Filename = FH 4-35-N1\_PROTON-4.j  
 Author = delta  
 Experiment = single pulse.ex2  
 Sample id = FH 4-35-N1  
 Solvent = CHLOROFORM-D  
 Creation time = 4-SEP-2013 15:08:36  
 Revision time = 27-JUL-2014 22:25:48  
 Current time = 27-JUL-2014 22:26:01

Comment = FH 4-35-N1  
 Data format = 1D COMPLEX  
 File size = 13107  
 File title = 1H  
 File units = [ppm]  
 Dimensions = X  
 Site = KEX 400p  
 Spectrometer = DELTA2\_NMR

Field strength = 9.389766 [T] (400 [MHz])  
 X acq\_duration = 1.36577024 [s]  
 X domain = 1H  
 X Freq = 399.78219838 [MHz]  
 X offset = 5 [ppm]  
 X points = 16384  
 X prescans = 1  
 X resolution = 0.73218757 [Hz]  
 X sweep = 11.99616123 [kHz]  
 Irr domain = 1H  
 Irr Freq = 399.78219838 [MHz]  
 Irr offset = 5 [ppm]  
 Tri domain = 1H  
 Tri Freq = 399.78219838 [MHz]  
 Tri offset = 5 [ppm]  
 Clipped = FALSE  
 Mod return = 1  
 Scans = 16  
 Total scans = 16

X 90 width = 8.6 [us]  
 X acq\_time = 1.36577024 [s]  
 X angle = 45 [deg]  
 X atm = 3 [dm]  
 X pulse = 4.3 [us]  
 Irr mode = OFF  
 Tri mode = OFF  
 Data present = FALSE  
 Initial wait = 1 [s]  
 Recvr gain = 26  
 Relaxation delay = 4 [s]  
 Repetition time = 5.36577024 [s]  
 Temp\_get = 21.3 [dC]

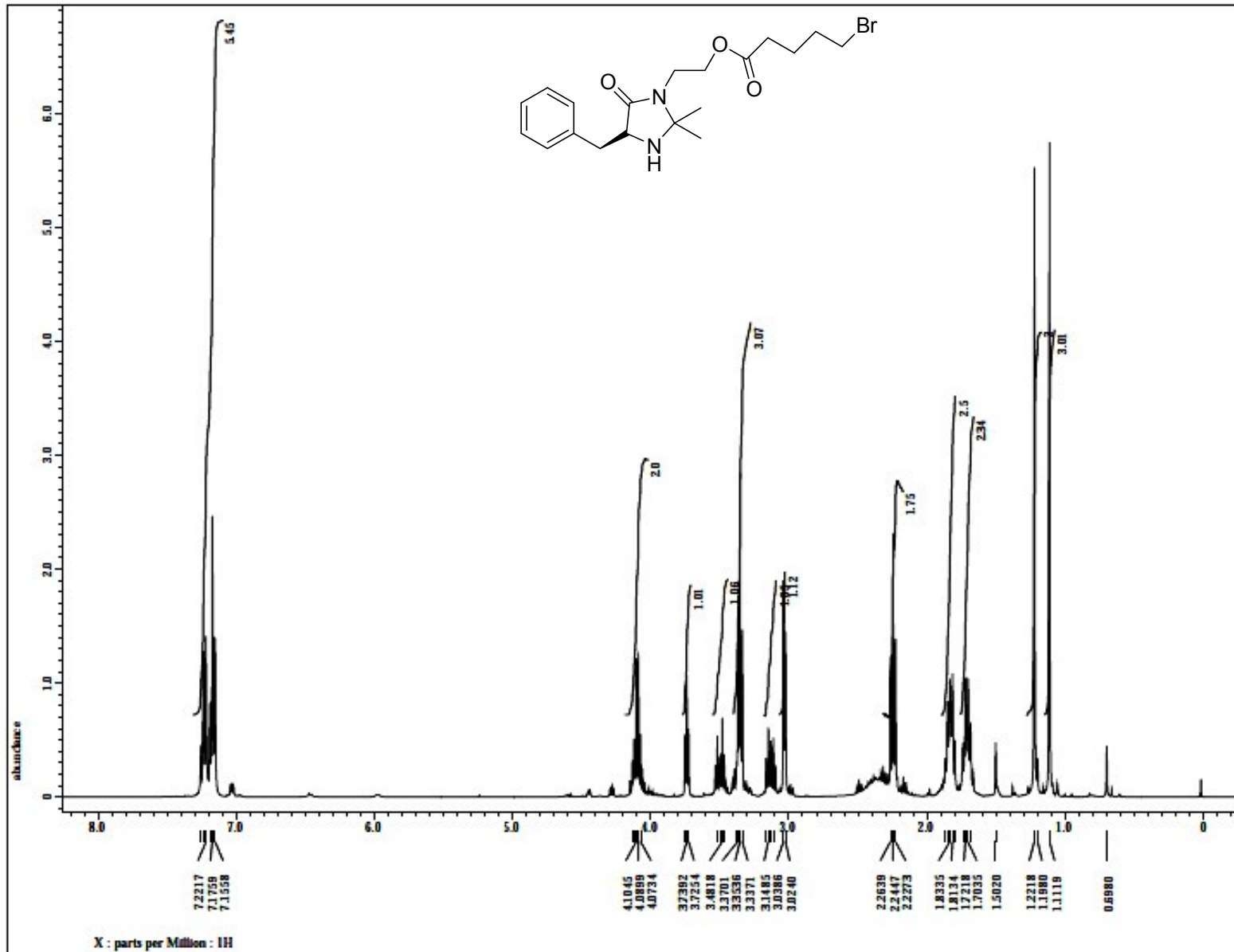


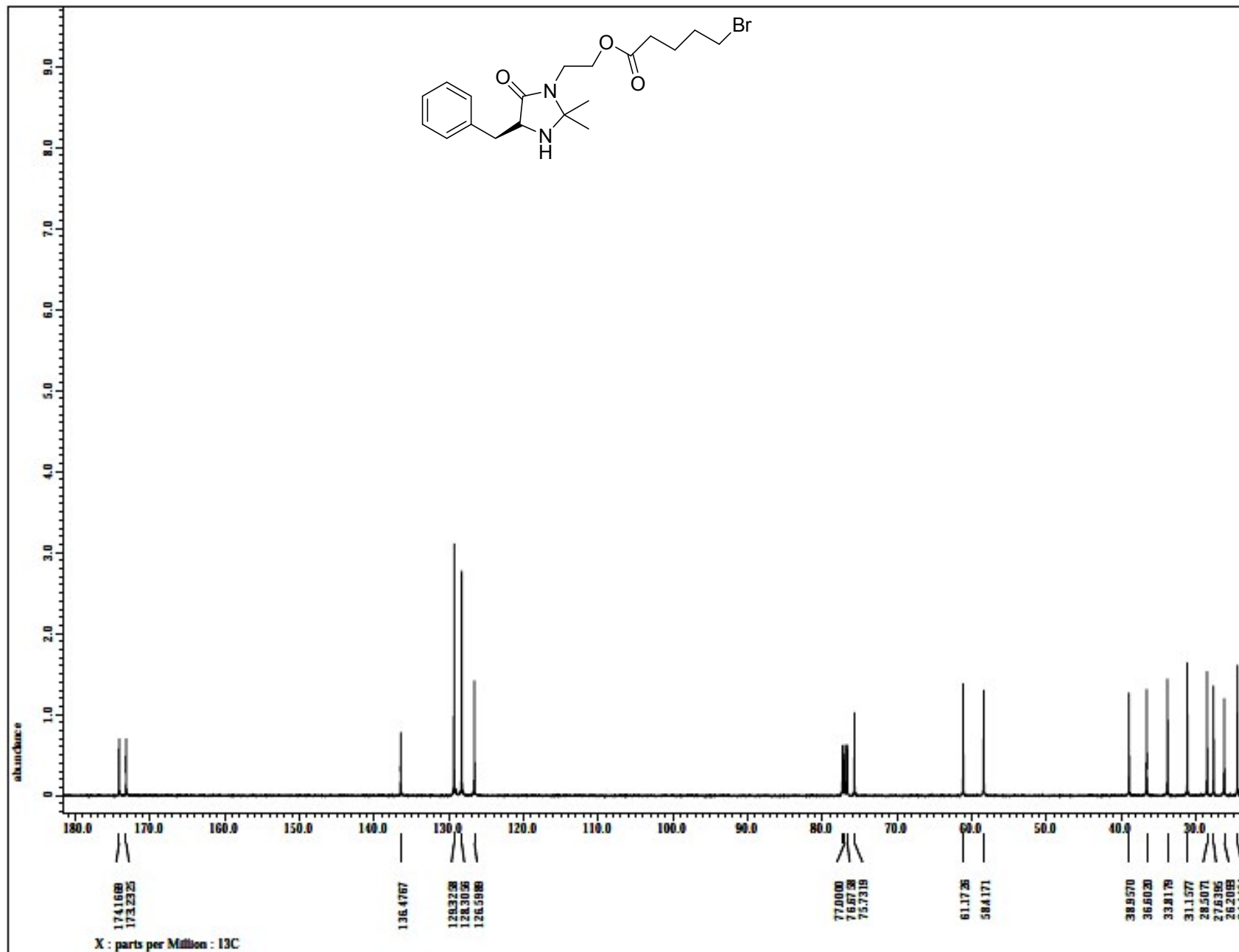
Filename = SS 4-35-N1\_CARDON-3.j  
 Author = delta  
 Experiment = single pulse\_dec  
 Sample id = SS 4-35-N1  
 Solvent = CHLOROFORM-D  
 Creation time = 6-SEP-2013 21:51:17  
 Revision time = 6-SEP-2013 22:25:55  
 Current time = 27-JUL-2014 22:26:34

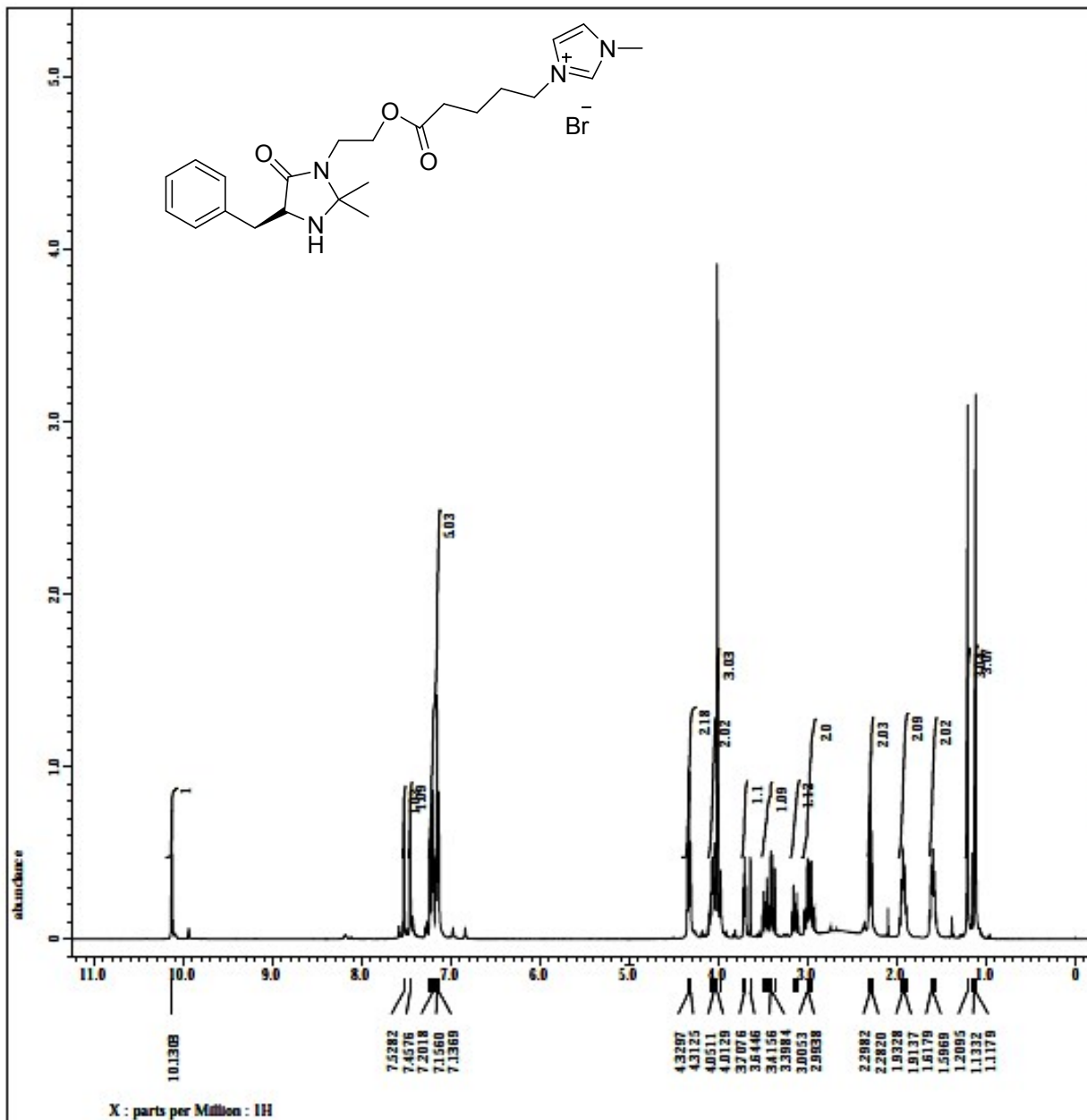
Comment = SS 4-35-N1  
 Data format = 1D COMPLEX  
 Data size = 26214  
 Data title = 13c  
 Data units = [ppm]  
 Dimensions = X  
 Site = NCI 400p  
 Spectrometer = DELTA2 400P

Field strength = 9.389766 [T] (400 [MHz])  
 X\_acq duration = 1.04333312 [s]  
 X\_domain = 13c  
 X\_freq = 100.52530333 [MHz]  
 X\_offset = 100 [ppm]  
 X\_points = 32768  
 X\_prescans = 4  
 X\_resolution = 0.95846665 [Hz]  
 X\_sweep = 31.40703518 [kHz]  
 Irr\_domain = 1H  
 Irr\_freq = 399.78219838 [MHz]  
 Irr\_offset = 5 [ppm]  
 Clipped = FALSE  
 Mod\_return = 1  
 Scans = 14  
 Total\_scans = 14

X\_90\_width = 11.98 [us]  
 X\_acq time = 1.04333312 [s]  
 X\_angle = 30 [deg]  
 X\_atn = 10 [db]  
 X\_pulse = 3.99333333 [us]  
 Irr\_atn\_dec = 25.52 [db]  
 Irr\_atn\_noc = 25.52 [db]  
 Irr\_noise = WALTZ  
 Decoupling = TRUE  
 Initial\_wait = 1 [s]  
 Nox = TRUE  
 Nox\_time = 2 [s]  
 Recvr\_gain = 52  
 Relaxation\_delay = 2 [s]  
 Repetition\_time = 3.04333312 [s]  
 Temp\_get = 20.1 [dC]





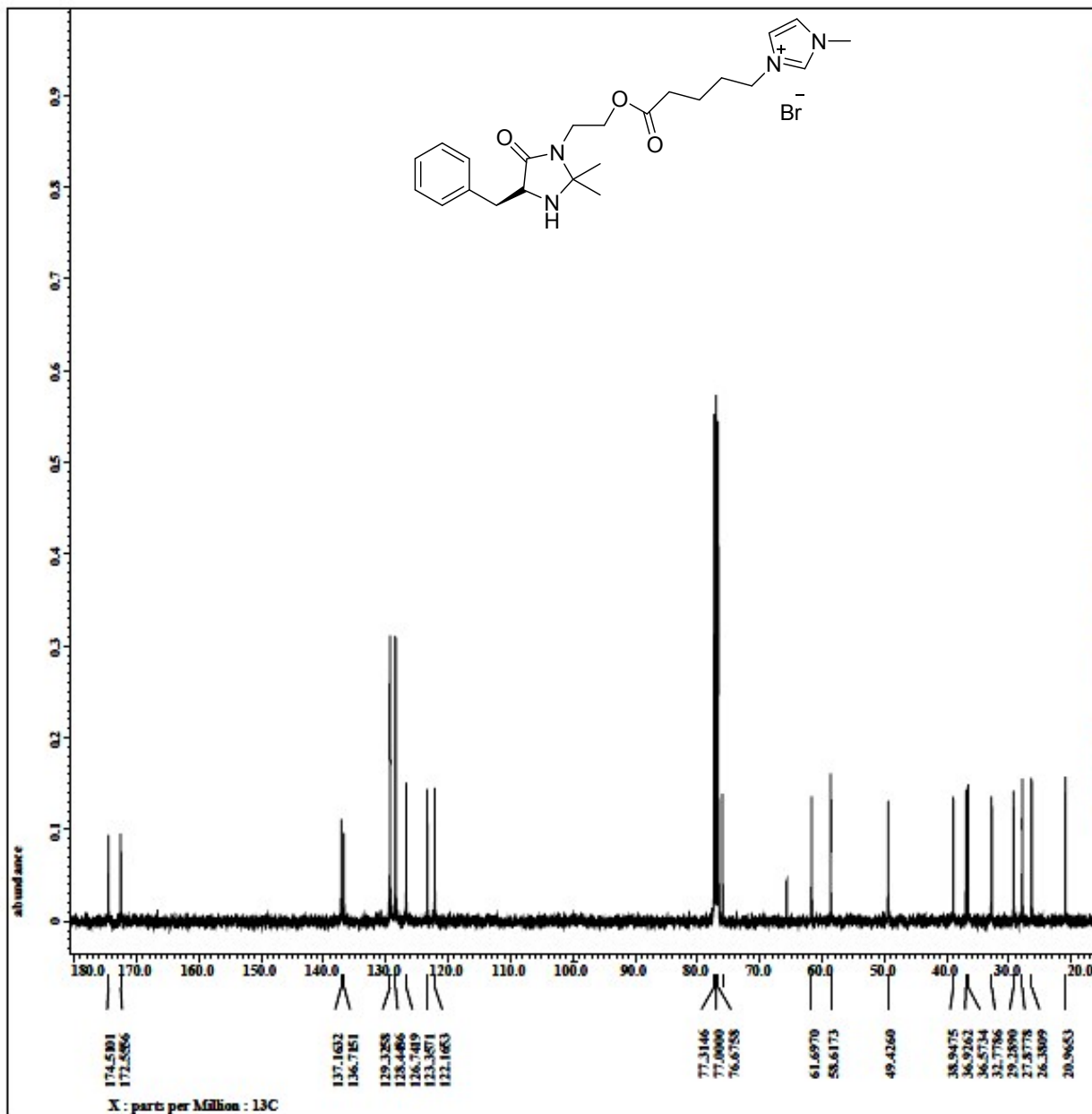


Filename = DTM 4-45-RR-8\_PROTON-  
 Author = delta  
 Experiment = single pulse.ac2  
 Sample id = DTM 4-45-RR-8  
 Solvent = CHLOROFORM-D  
 Creation time = 15-SEP-2014 11:50:55  
 Revision time = 15-JAN-2015 21:59:05  
 Current time = 15-JAN-2015 21:59:25

Comment = DTM 4-45-RR-8  
 Data format = 1D COMPLEX  
 Data size = 13107  
 Data title = 1H  
 Data units = [ppm]  
 Dimensions = X  
 Site = KXCX 400P  
 Spectrometer = DELTA2 400M

Field strength = 9.389766 [T] (400 [MHz])  
 F1 acq duration = 1.31072 [s]  
 F1 domain = 1H  
 F1 freq = 399.78219838 [MHz]  
 F1 offset = 5 [ppm]  
 F1 points = 16384  
 F1 prescans = 1  
 F1 resolution = 0.76293945 [Hz]  
 F1 sweep = 12.5 [kHz]  
 Irr domain = 1H  
 Irr freq = 399.78219838 [MHz]  
 Irr offset = 5 [ppm]  
 Tri domain = 1H  
 Tri freq = 399.78219838 [MHz]  
 Tri offset = 5 [ppm]  
 Clipped = FALSE  
 Mod return = 1  
 Scans = 16  
 Total scans = 16

F2 width = 10.1 [us]  
 F2 acq time = 1.31072 [s]  
 F2 angle = 45 [deg]  
 F2 step = 3 [dB]  
 F2 pulse = 5.05 [us]  
 Irr mode = OFF  
 Tri mode = OFF  
 Dante preset = FALSE  
 Initial wait = 1 [s]  
 Recvr gain = 30  
 Relaxation delay = 4 [s]  
 Repetition time = 5.31072 [s]  
 Temp get = 21.4 [degC]



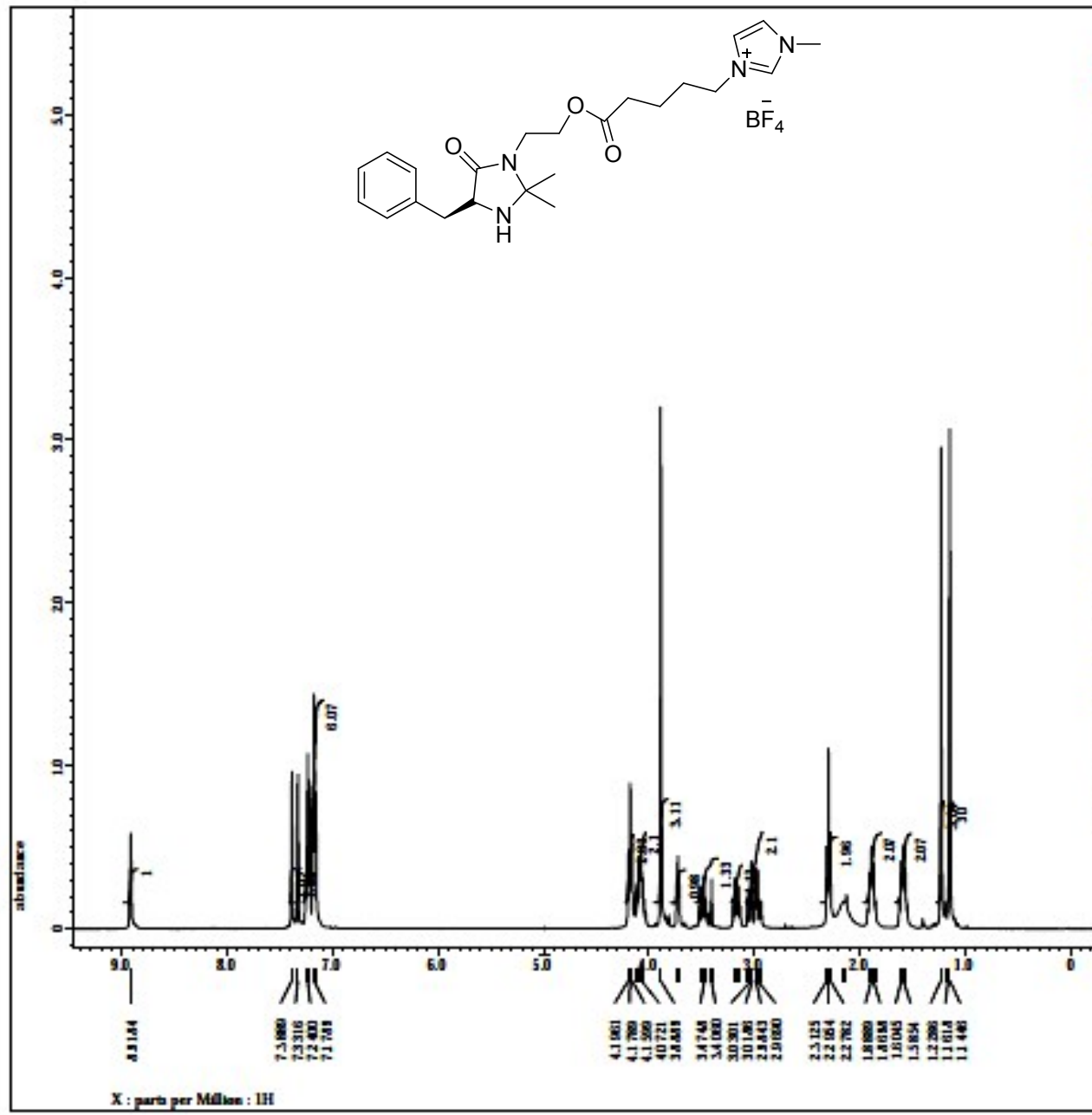
Filename = 4-45-BR-8\_CARBON-4.jd  
 Author = delta  
 Experiment = single pulse dec  
 Sample id = 4-45-BR-8  
 Solvent = CHLOROFORM-D  
 Creation time = 18-SEP-2014 01:05:27  
 Revision time = 23-SEP-2014 11:58:05  
 Current time = 23-SEP-2014 11:58:20

Comment = 4-45-BR-8  
 Data format = 1D COMPLEX  
 Dim size = 26214  
 Dim title = 13C  
 Dim units = [ppm]  
 Dimensions = X  
 Site = RCI 400P  
 Spectrometer = DELTA2 NMR

Field strength = 9.389766 [T] (400 [MHz])  
 X acq duration = 1.0433312 [s]  
 X domain = 13C  
 X freq = 100.52530333 [MHz]  
 X offset = 100 [ppm]  
 X points = 32768  
 X prescans = 4  
 X resolution = 0.95846665 [Hz]  
 X sweep = 31.40703518 [kHz]  
 Irr domain = 1H  
 Irr freq = 399.78219838 [MHz]  
 Irr offset = 5 [ppm]  
 Clipped = TRUE  
 Mod return = 1  
 Scans = 512  
 Total scans = 512

X 90 width = 11.98 [us]  
 X acq time = 1.0433312 [s]  
 X angle = 30 [deg]  
 X atn = 10 [dB]  
 X pulse = 3.99333333 [us]  
 Irr atn dec = 25.52 [dB]  
 Irr atn noe = 25.52 [dB]  
 Irr noise = WALTZ  
 Decoupling = TRUE  
 Initial wait = 1 [s]  
 Noe = TRUE  
 Noe time = 2 [s]  
 Recvr gain = 60  
 Relaxation delay = 2 [s]  
 Repetition time = 3.0433312 [s]  
 Temp get = 21.7 [dC]





```

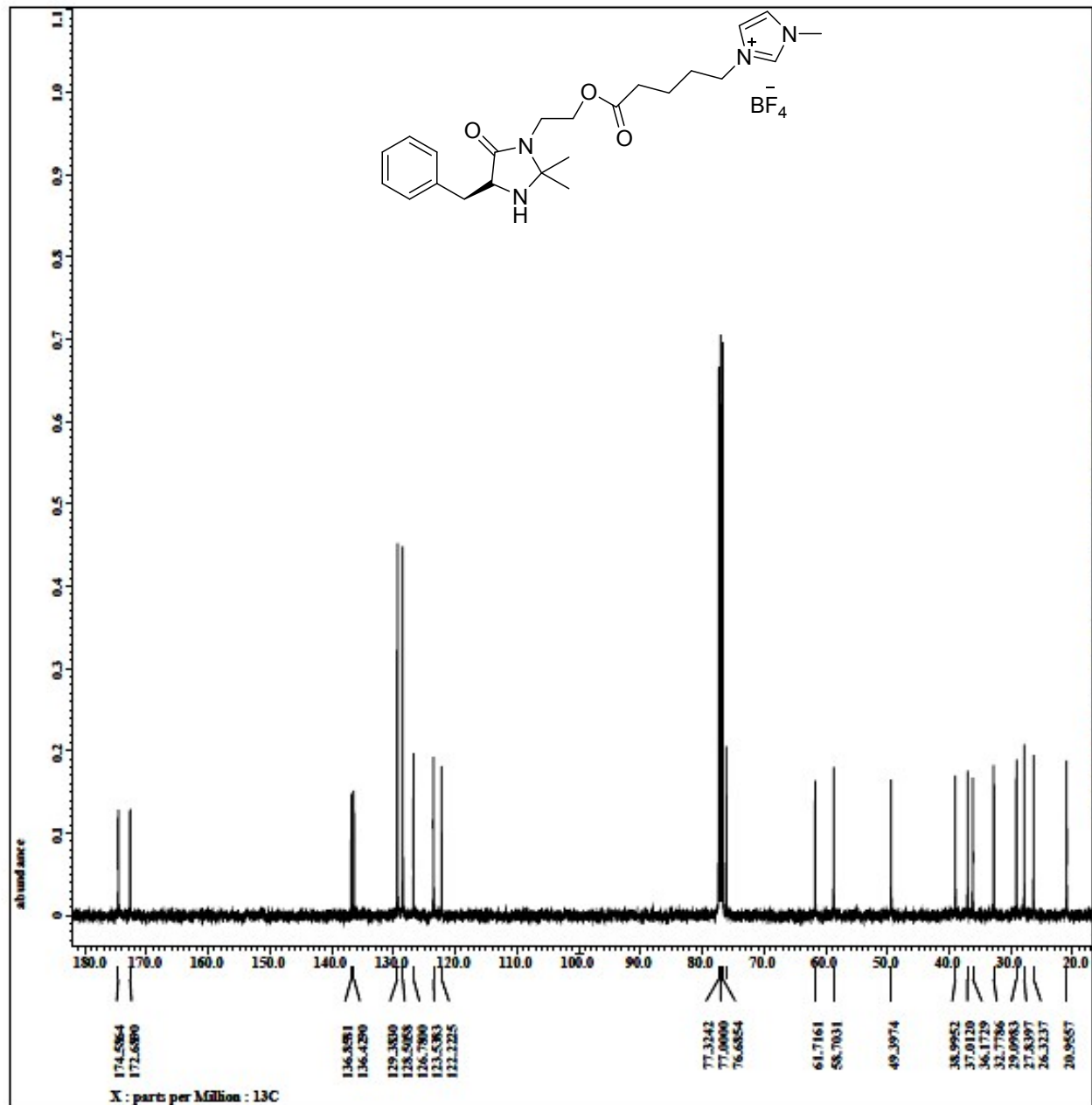
Filename      = DTM 4-45-EP-3 PROTON-
Author       = Gelta
Experiment    = single pulse.ex1
Sample id    = DTM 4-45-EP-3
Solvent      = CDCl3/CF3COOH-D
Creation time = 15-SEP-2014 11:26:56
Revision time = 15-SEP-2014 16:02:46
Current time  = 15-SEP-2014 16:02:16

Comment      = DTM 4-45-EP-3
Data format  = 1D COMPLEX
Dim size     = 13107
Dim title    = 1D
Dim units    = [ppm]
Dimensions   = 1
Site         = ECI 400P
Spectrometer = DELTA2 NMR

F1field strength = 9.389766(T) (400(MHz))
F1scq duration   = 1.31072(s)
F1domain         = 1D
F1freq           = 399.76219638(MHz)
F1offset         = 5(ppm)
F1points         = 16384
F1prescans       = 1
F1resolution     = 0.76293945(KHz)
F1sweep          = 12.5(KHz)
F1r_domain       = 1D
F1r_freq         = 399.76219638(MHz)
F1r_offset       = 5(ppm)
F1r_domain       = 1D
F1r_freq         = 399.76219638(MHz)
F1r_offset       = 5(ppm)
Clipped         = FALSE
Mod return      = 1
Scans           = 16
Total scans     = 16

F2 90 width     = 10.1(us)
F2scq time      = 1.31072(s)
F2mag           = 45(deg)
F2stb           = 1(dB)
F2pulse         = 5.05(us)
F2r_mode        = Off
F2r_mode        = Off
Dante preset    = FALSE
Initial wait    = 1(s)
Recvr gain      = 3d
Relaxation delay = 4(s)
Repetition time = 5.31072(s)
Temp_get        = 21.4(0C)
  
```





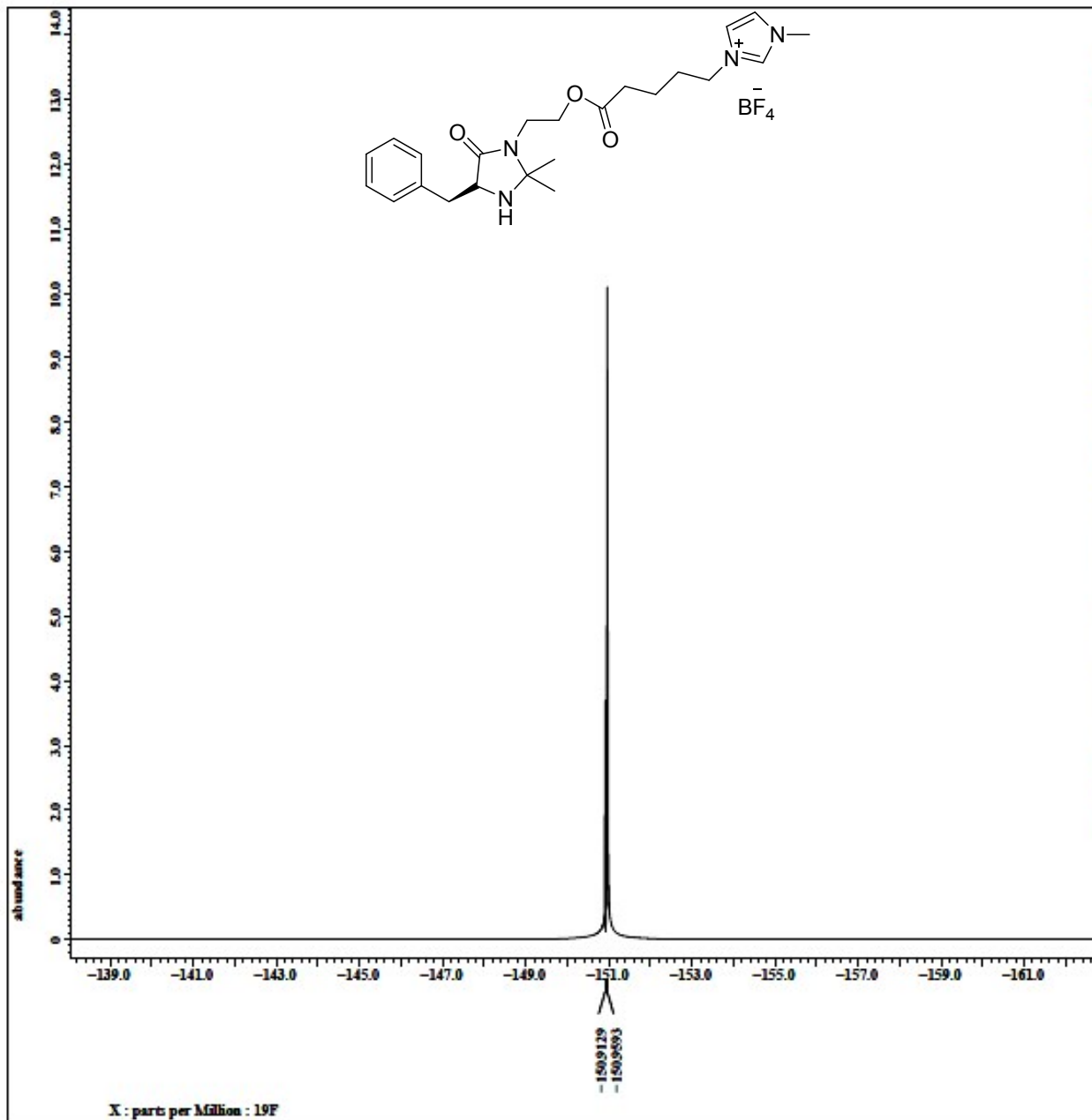
```

Filename      = 4-45-BF-8_CARBON-3.jd
Author       = delta
Experiment   = single pulse dec
Sample id    = 4-45-BF-8
Solvent      = CHLOROFORM-D
Creation time = 18-SEP-2014 00:34:59
Revision time = 18-SEP-2014 01:20:38
Current time  = 23-SEP-2014 11:57:13

Comment      = 4-45-BF-8
Data format  = 1D COMPLEX
Dim size     = 26214
Dim title    = 13C
Dim units    = [ppm]
Dimensions   = X
Site         = RFX 400P
Spectrometer = DELTA2 MGR

Field strength = 9.389766 [T] (400 [MHz])
F1 acq duration = 1.04333312 [s]
F1 domain      = 13C
F1 freq        = 100.52530333 [MHz]
F1 offset      = 100 [ppm]
F1 points      = 32768
F1 proc scans  = 4
F1 resolution  = 0.95846665 [Hz]
F1 sweep       = 31.40703518 [kHz]
Irr domain     = 1H
Irr freq       = 399.78219838 [MHz]
Irr offset     = 5 [ppm]
Clipped       = TRUE
Mod return     = 1
Scans          = 512
Total_scans   = 512

F2 90 width    = 11.98 [us]
F2 acq time    = 1.04333312 [s]
F2 angle       = 30 [deg]
F2 atn         = 10 [dB]
F2 pulse       = 3.99333333 [us]
Irr atn dec    = 25.52 [dB]
Irr atn noe    = 25.52 [dB]
Irr noise      = WALTZ
Decoupling     = TRUE
Initial wait   = 1 [s]
Noe            = TRUE
Noe time       = 2 [s]
Recvr gain     = 60
Relaxation delay = 2 [s]
Repetition time = 3.04333312 [s]
Temp gat       = 21.7 [dC]
  
```

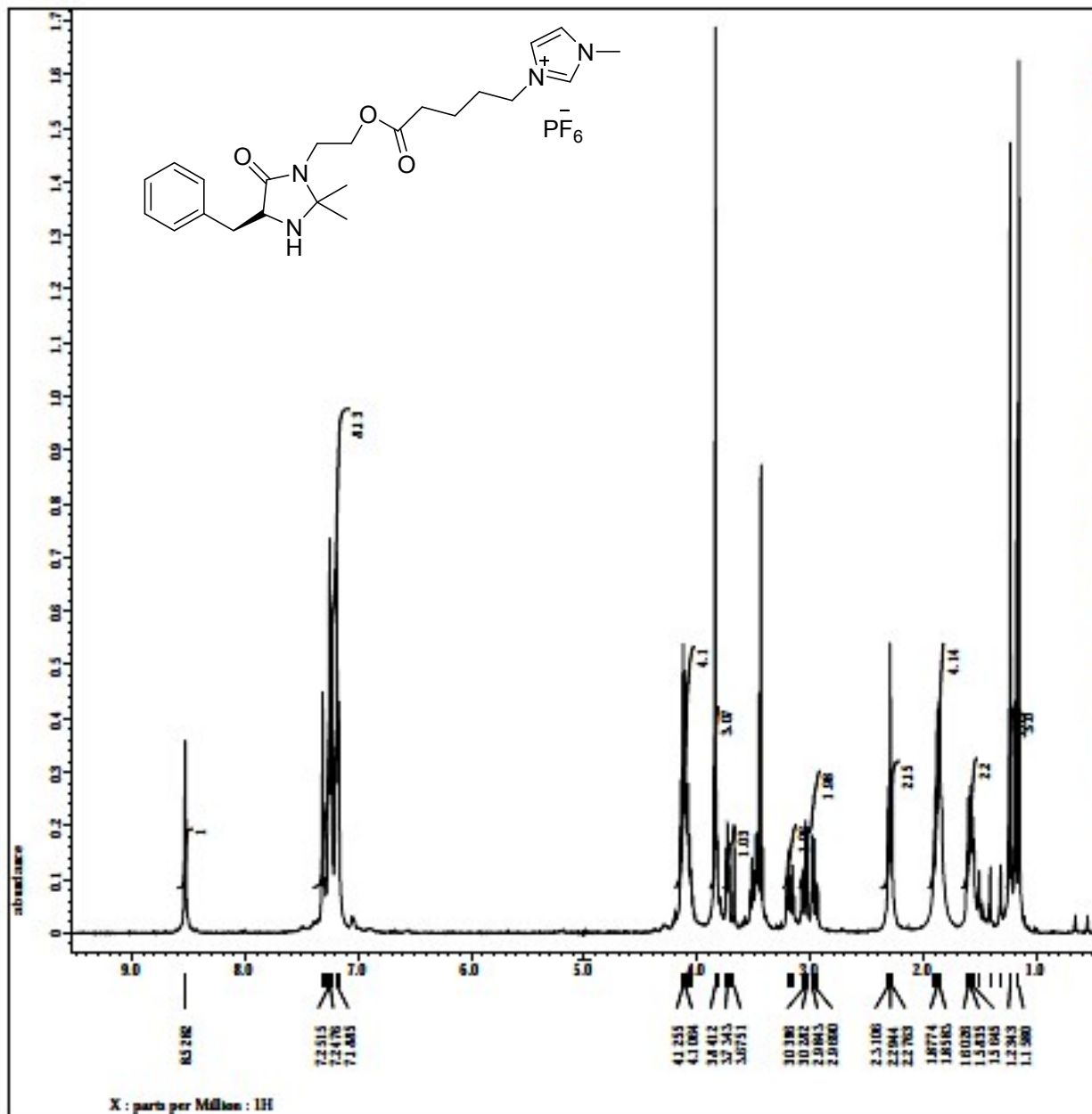


Filename = 88\_4-45-BFM\_19F-4.jdf  
 Author = delta  
 Experiment = single\_pulse.ex2  
 Sample id = 88\_4-45-BFM\_19F  
 Solvent = CHLOROFORM-D  
 Creation time = 4-DEC-2014 09:50:10  
 Revision time = 23-SEP-2014 11:53:16  
 Current time = 23-SEP-2014 11:53:42

Comment = Single pulse without  
 Data format = 1D REAL  
 Dim size = 26214  
 Dim title = 19F  
 Dim units = [ppm]  
 Dimensions = 1  
 Site = ECI 400P  
 Spectrometer = DELTA2\_NMR

Field strength = 9.389766 [T] (400 [MHz])  
 F1 acq duration = 0.229376 [s]  
 F1 domain = 19F  
 F1 freq = 376.17105393 [MHz]  
 F1 offset = -100 [ppm]  
 F1 points = 32766  
 F1 prescans = 2  
 F1 resolution = 4.35965402 [Hz]  
 F1 sweep = 142.85714286 [kHz]  
 Irr domain = 19F  
 Irr freq = 376.17105393 [MHz]  
 Irr offset = 5 [ppm]  
 Tri domain = 19F  
 Tri freq = 376.17105393 [MHz]  
 Tri offset = 5 [ppm]  
 Clipped = FALSR  
 Mod return = 1  
 Scans = 128  
 Total\_scans = 128

F2 90 width = 12.32 [us]  
 F2 acq time = 0.229376 [s]  
 F2 angle = 45 [deg]  
 F2 atn = 6 [dB]  
 F2 pulse = 6.16 [us]  
 Irr mode = Off  
 Tri mode = Off  
 Dante preset = FALSR  
 Initial wait = 1 [s]  
 Recvr gain = 36  
 Relaxation delay = 2 [s]  
 Repetition time = 2.229376 [s]  
 Temp gat = 21.9 [dC]



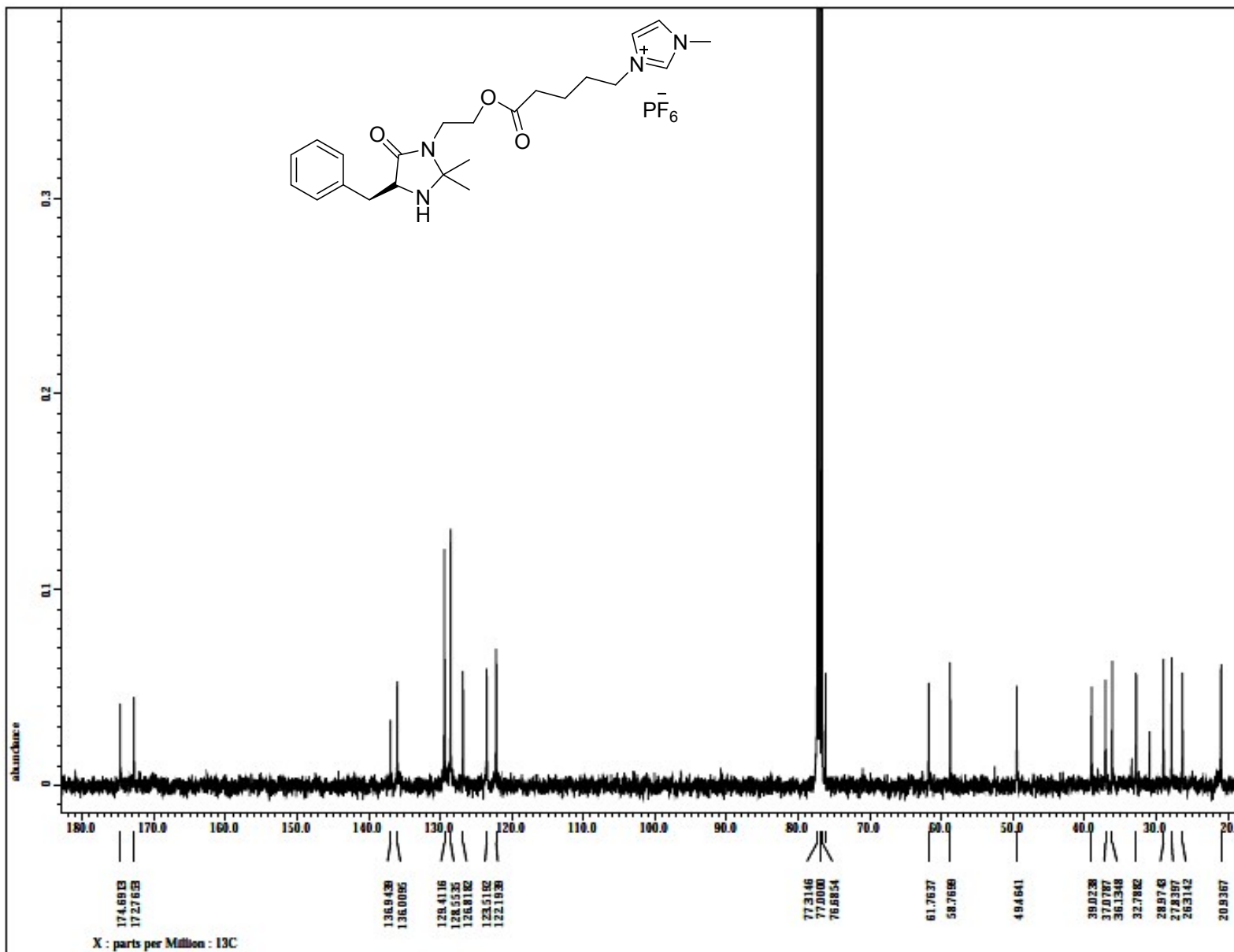
```

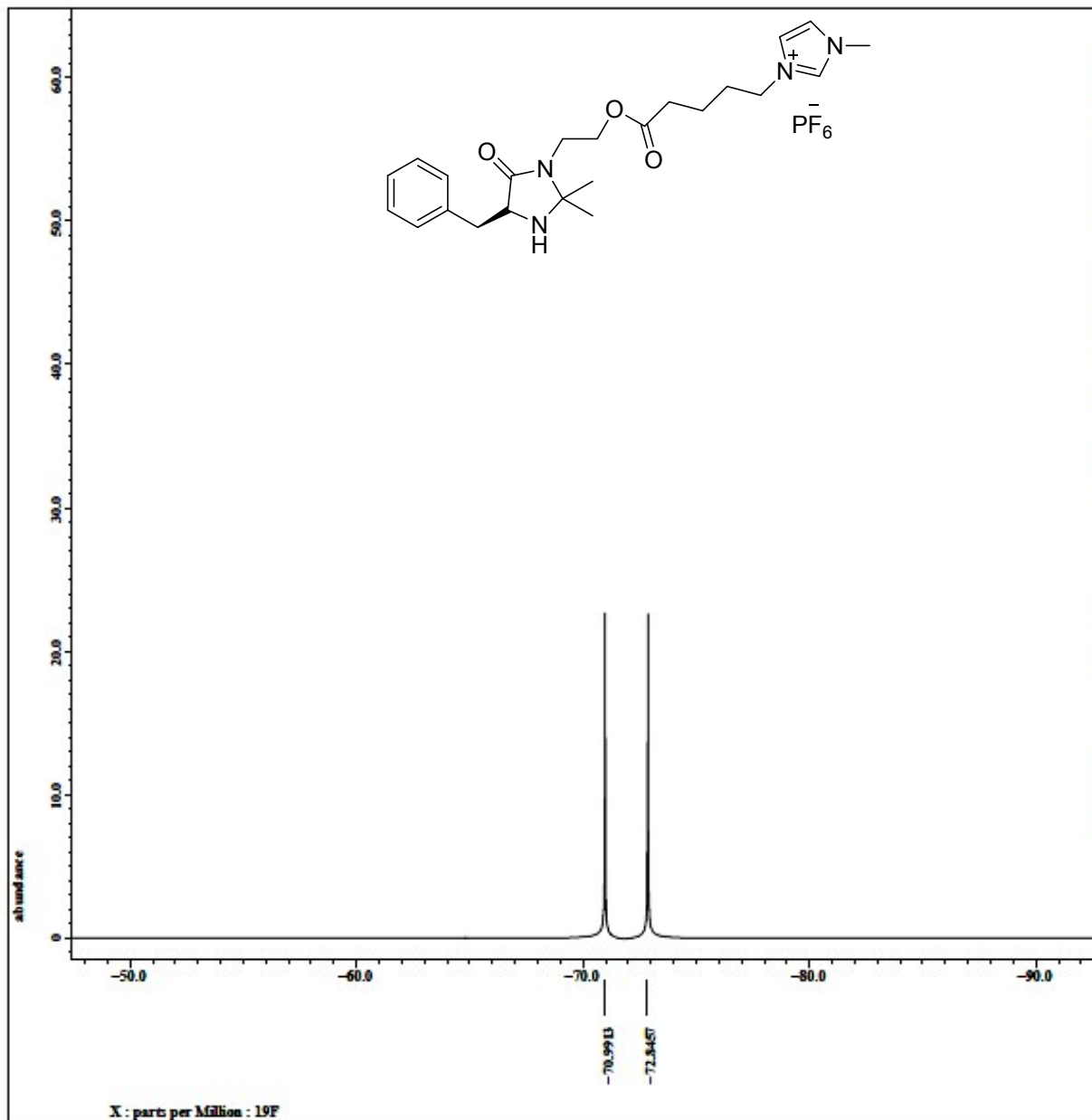
Filename      = MS 4-45-FPG_PROTON-4.
Author       = delta
Experiment    = single pulse.ex2
Sample id    = MS 4-45-FPG
Solvent      = CHLOROFORM-D
Creation time = 21-SEP-2014 13:00:13
Revision time = 15-SEP-2014 13:09:17
Current time  = 15-SEP-2014 13:09:43

Comment      = MS 4-45-FPG
Data format  = 1D COMPLEX
Dir name     = 12107
Dir title    = 1H
Dir units    = [ppm]
Dimensions   = 1
Site         = RFX 400P
Spectrometer = DELTA2 NMR

Field strength = 9.395766(T) (400(MHz))
F1 acq duration = 1.31072(s)
F1 domain      = 1H
F1 freq        = 399.78219838(MHz)
F1 offset      = 5(ppm)
F1 points      = 16384
F1 prescans    = 1
F1 resolution  = 0.76293945(Hz)
F1 sweep       = 12.5(KHz)
F1r domain     = 1H
F1r freq       = 399.78219838(MHz)
F1r offset     = 5(ppm)
F1r domain     = 1H
F1r freq       = 399.78219838(MHz)
F1r offset     = 5(ppm)
Clipped       = PALSER
Mod/return    = 1
Scale         = 8
Total scans   = 8

F1 90 width    = 10.1(us)
F1 acq time    = 1.31072(s)
F1 angle       = 45(deg)
F1 nch         = 1(ch)
F1 pulse       = 5.05(us)
F1r mode       = OFF
F1r mode       = OFF
DANTE present  = PALSER
Initial wait   = 1(s)
Recvr gain     = 10
Relaxation delay = 4(s)
Repetition time = 1.31072(s)
Temp_get      = 406.1(dC)
  
```





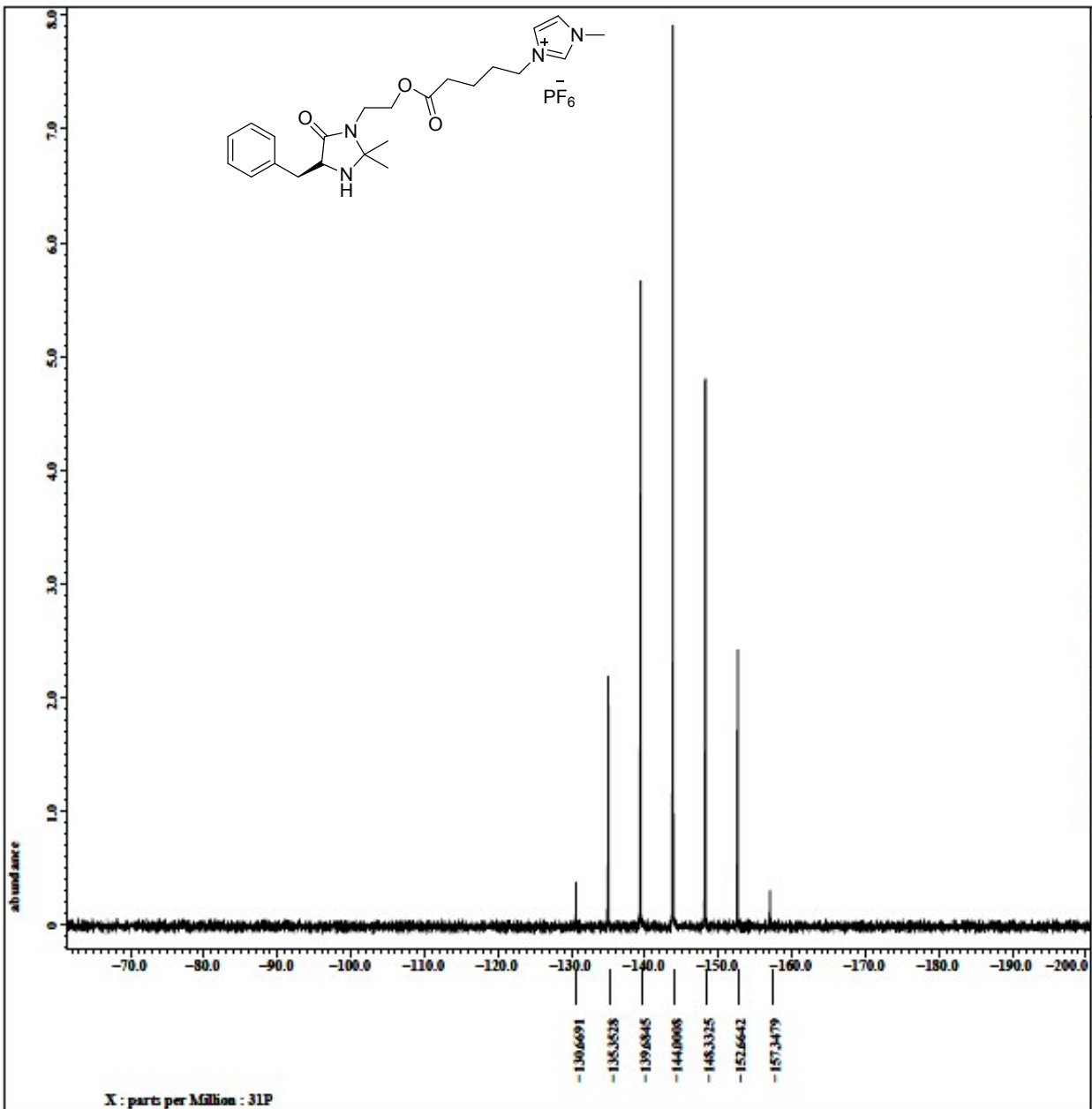
```

Filename      = 88_4-45-PPN_19F-4.jdf
Author       = delta
Experiment   = single_pulse.ex2
Sample id    = 88_4-45-PPN_19F
Solvent      = CHLOROFORM-D
Creation time = 4-ADG-2014 10:20:03
Revision time = 23-SEP-2014 11:54:50
Current time  = 23-SEP-2014 11:55:20

Comment      = Single pulse without
Data format  = 1D REAL
Dim size     = 26214
Dim title    = 19F
Dim units    = [ppm]
Dimensions   = X
Site         = ECX 400P
Spectrometer = DELTA2 NMR

Field strength = 9.389766 [T] (400 [MHz])
X_acq duration = 0.229376 [s]
X_domain      = 19F
X_freq        = 376.17105393 [MHz]
X_offset      = -100 [ppm]
X_points      = 32768
X_prescans    = 2
X_resolution  = 4.35965402 [Hz]
X_sweep       = 142.85714286 [kHz]
Irr_domain    = 19F
Irr_freq      = 376.17105393 [MHz]
Irr_offset    = 5 [ppm]
Tri_domain    = 19F
Tri_freq      = 376.17105393 [MHz]
Tri_offset    = 5 [ppm]
Clipped       = FALSR
Mod Return    = 1
Scans         = 128
Total_scans   = 128

X_90_width    = 12.32 [us]
X_acq time    = 0.229376 [s]
X_angle       = 45 [deg]
X_atn         = 6 [dB]
X_pulse       = 6.16 [us]
Irr_mode      = Off
Tri_mode      = Off
Dante preset  = FALSR
Initial wait  = 1 [s]
Recvr_gain    = 36
Relaxation delay = 2 [s]
Repetition time = 2.229376 [s]
Temp_get      = 22.6 [dC]
  
```



```

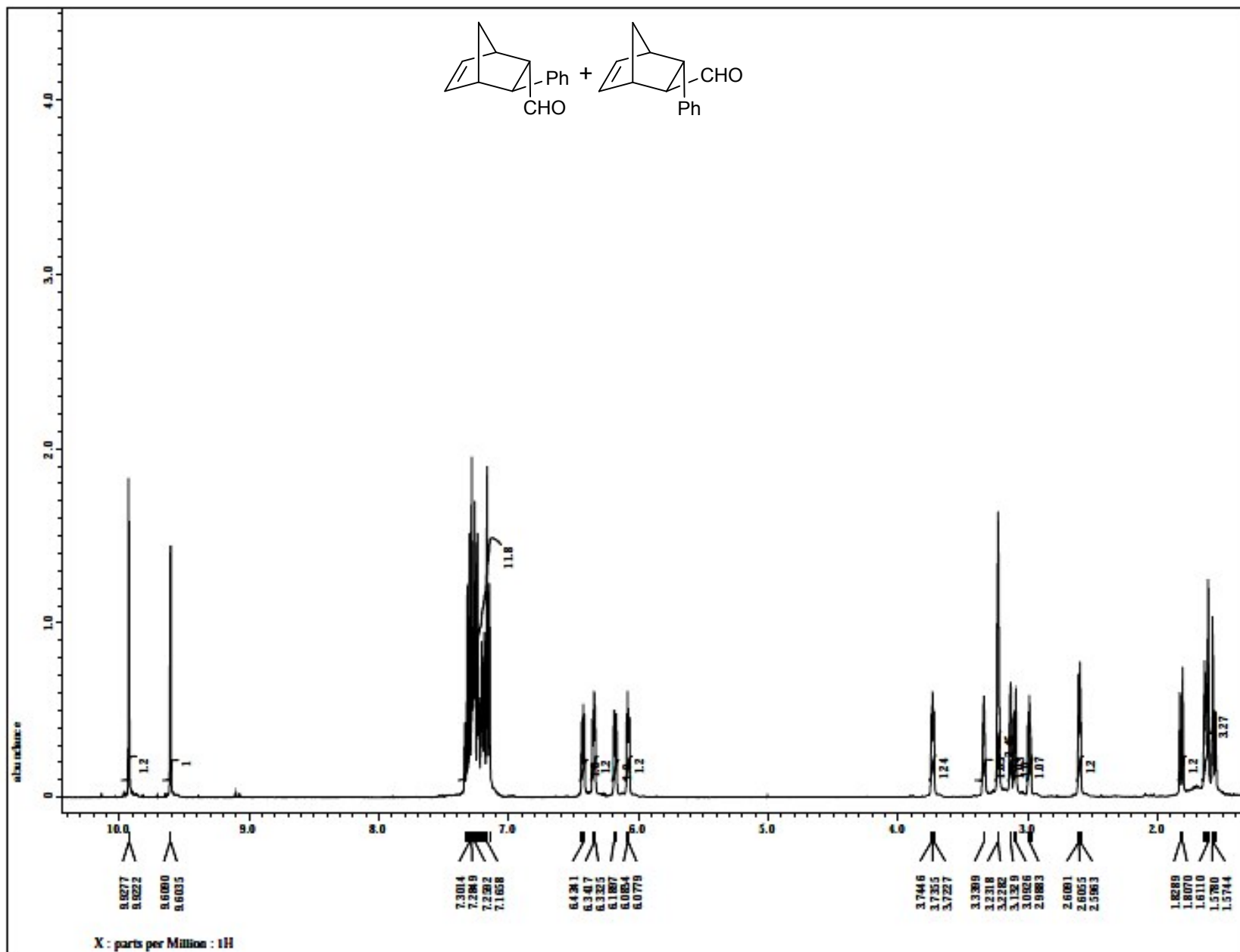
Filename      = 88 4-45-DFW 31P-3.jdf
Author       = delta
Experiment   = single pulse.ex2
Sample id    = 88 4-45-DFW 31P
Solvent      = CHLOROFORM-D
Creation time = 7-ADC-2014 11:47:52
Revision time = 23-SEP-2014 11:55:51
Current time  = 23-SEP-2014 11:56:31

Comment      = 31P Single pulse with
Data format  = 1D COMPLEX
Dim size     = 26214
Dim title    = 31P
Dim units    = [ppm]
Dimensions   = 1
Site         = RCM 400P
Spectrometer = DELTA2_NMR

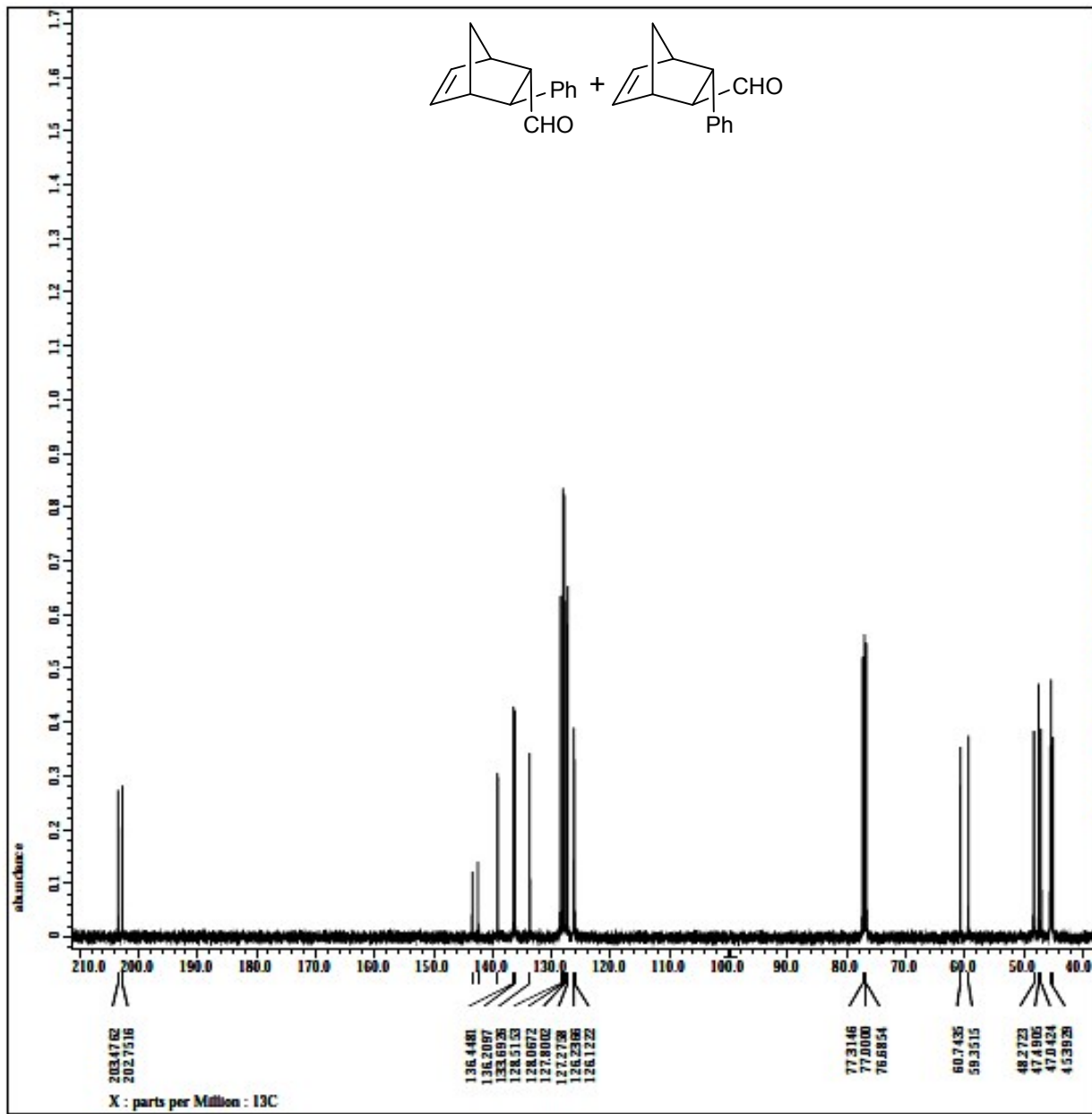
Field strength = 9.389766[T] (400[MHz])
F1 acq duration = 0.40370176[s]
F1 domain      = 31P
F1 freq        = 161.83469309[MHz]
F1 offset      = 0[ppm]
F1 points      = 32788
F1 prescans    = 2
F1 resolution  = 2.47707615[Hz]
F1 sweep       = 81.16883117[kHz]
F1r domain     = 31P
F1r freq       = 161.83469309[MHz]
F1r offset     = 5[ppm]
F1r domain     = 31P
F1r freq       = 161.83469309[MHz]
F1r offset     = 5[ppm]
Clipped       = FALSE
Mod return    = 1
Scans         = 599
Total scans   = 599

F1 90 width   = 10.15[us]
F1 acq time   = 0.40370176[s]
F1 angle      = 90[deg]
F1 gain       = 8.9[dB]
F1 pulse      = 10.15[us]
F1r mode      = Off
F1r mode      = Off
Dante preset  = FALSE
Initial wait  = 1[s]
Recvr gain    = 56
Relaxation delay = 2[s]
Repetition time = 2.40370176[s]
Temp set      = 21.6[degC]
  
```









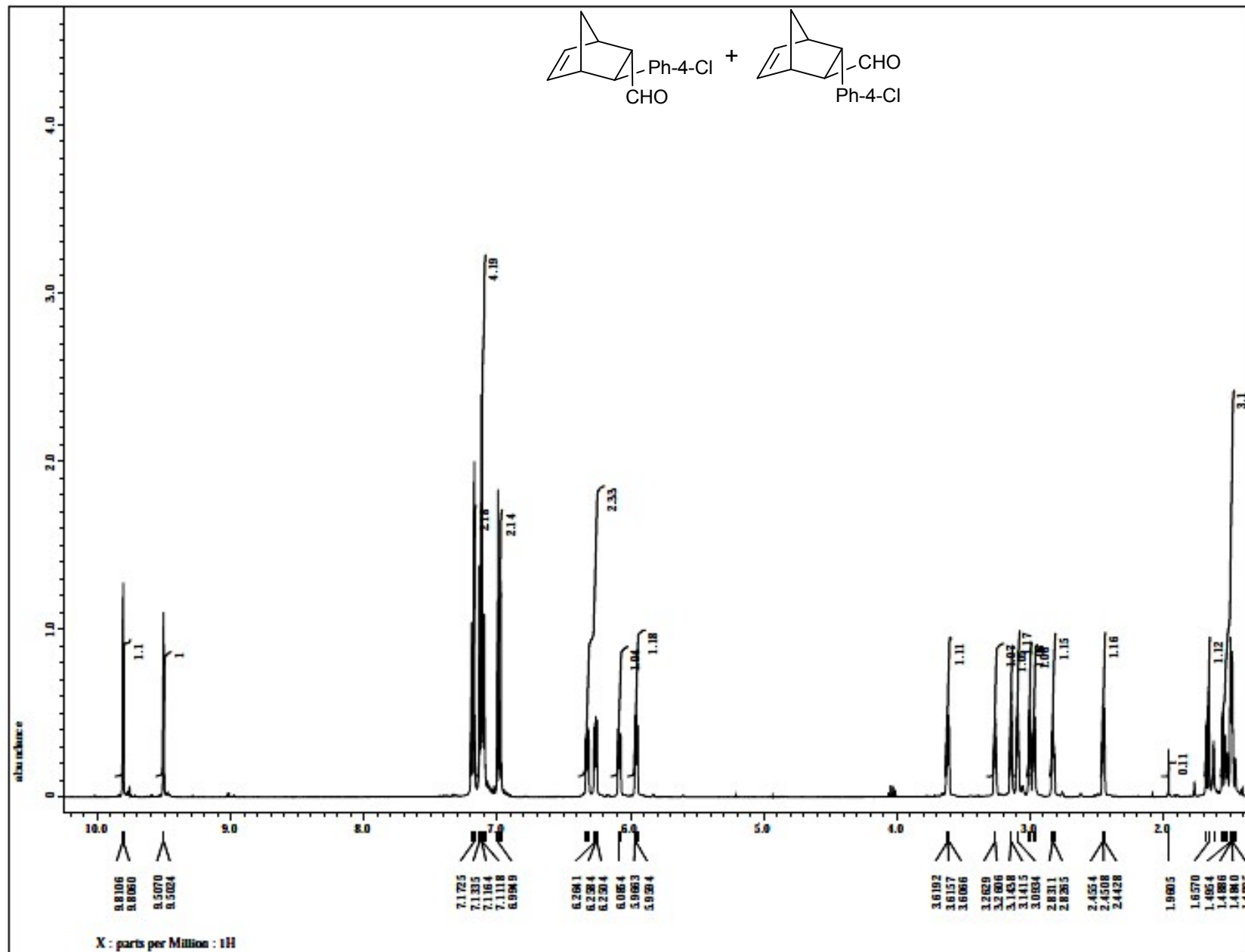
```

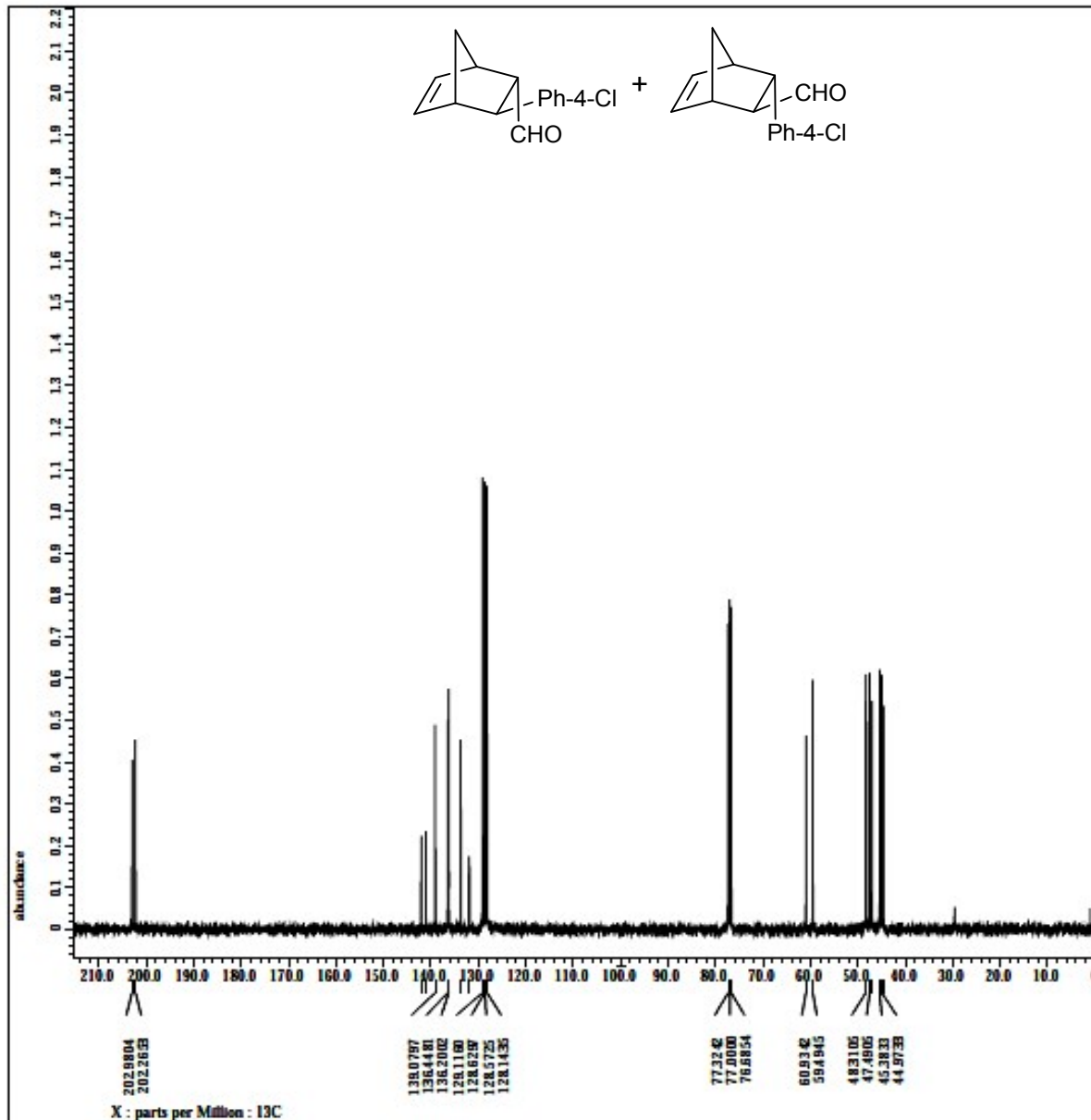
Filename      = SS 4-75_CARBON-3.jdf
Author       = delta
Experiment    = single pulse_dec
Sample id     = SS 4-75
Solvent       = CHLOROFORM-D
Creation time = 24-OCT-2012 16:12:24
Revision time = 24-OCT-2012 16:32:25
Current time  = 28-JUL-2014 20:02:35

Comment      = SS 4-75
Data format   = 1D COMPLEX
Dim size      = 26214
Dim title     = 13c
Dim units     = [ppm]
Dimensions    = X
Site          = MCI 400p
Spectrometer  = DELTA2_MMR

Field strength = 9.389766 [T] (400 [MHz])
X_acq duration = 1.04333312 [s]
X_domain      = 13c
X_freq        = 100.52530333 [MHz]
X_offset      = 100 [ppm]
X_points      = 32768
X_prescans    = 4
X_resolution  = 0.95846665 [Hz]
X_sweep       = 31.40703518 [kHz]
Irr_domain    = 1H
Irr_freq      = 399.78219838 [MHz]
Irr_offset    = 5 [ppm]
Clipped       = FALSE
Mod_return    = 1
Scans         = 750
Total_scans   = 750

X_90 width    = 11.75 [us]
X_acq time    = 1.04333312 [s]
X_angle       = 30 [deg]
X_atn         = 10 [db]
X_pulse       = 3.91666667 [us]
Irr_atn_dec   = 24.95 [db]
Irr_atn_noc   = 24.95 [db]
Irr_noise     = WALTZ
Decoupling    = TRUE
Initial_wait  = 1 [s]
Noe           = TRUE
Noe time      = 2 [s]
Noe_gain      = 58
Relaxation_delay = 2 [s]
Repetition_time = 3.04333312 [s]
Temp_get      = 19.6 [dc]
  
```





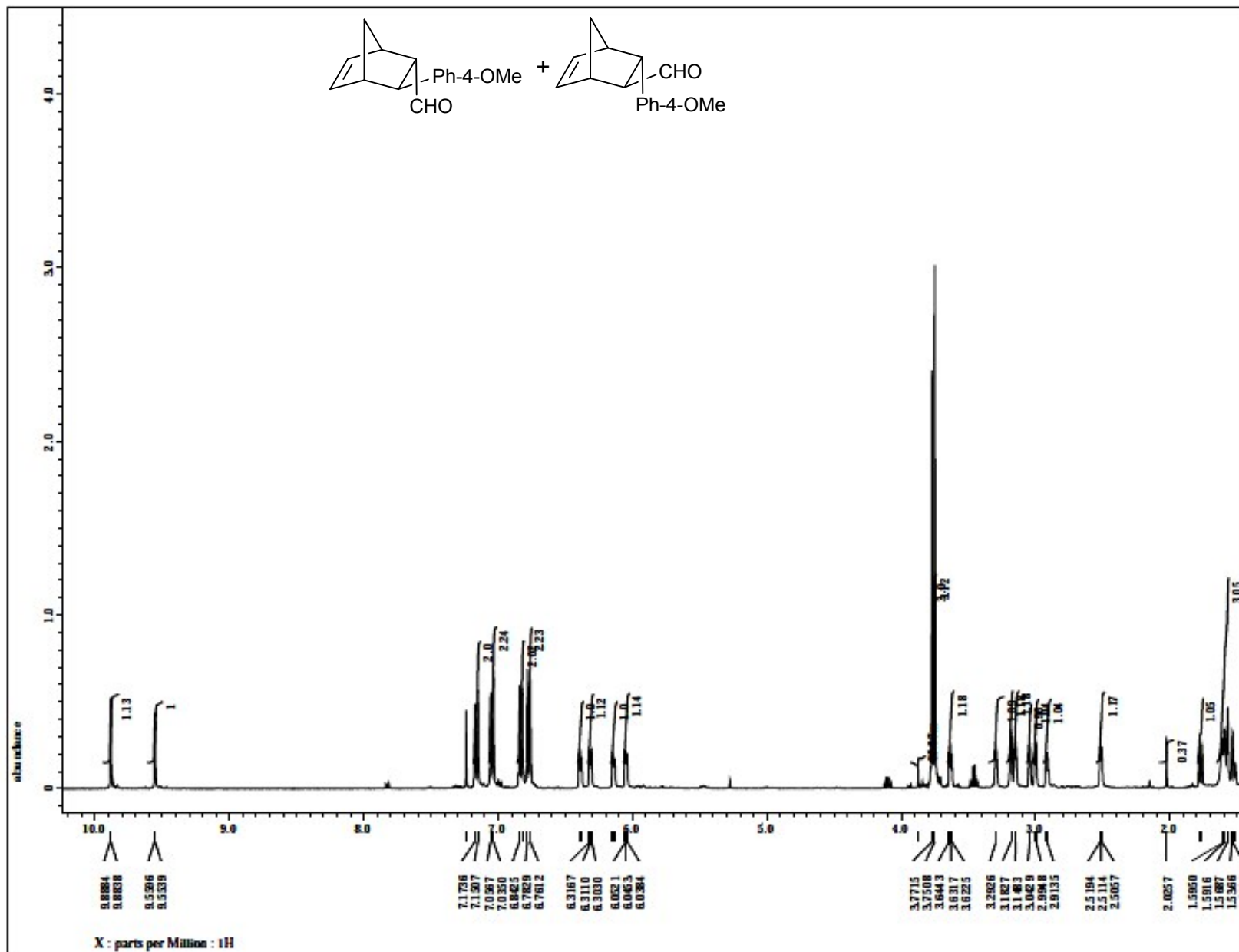
```

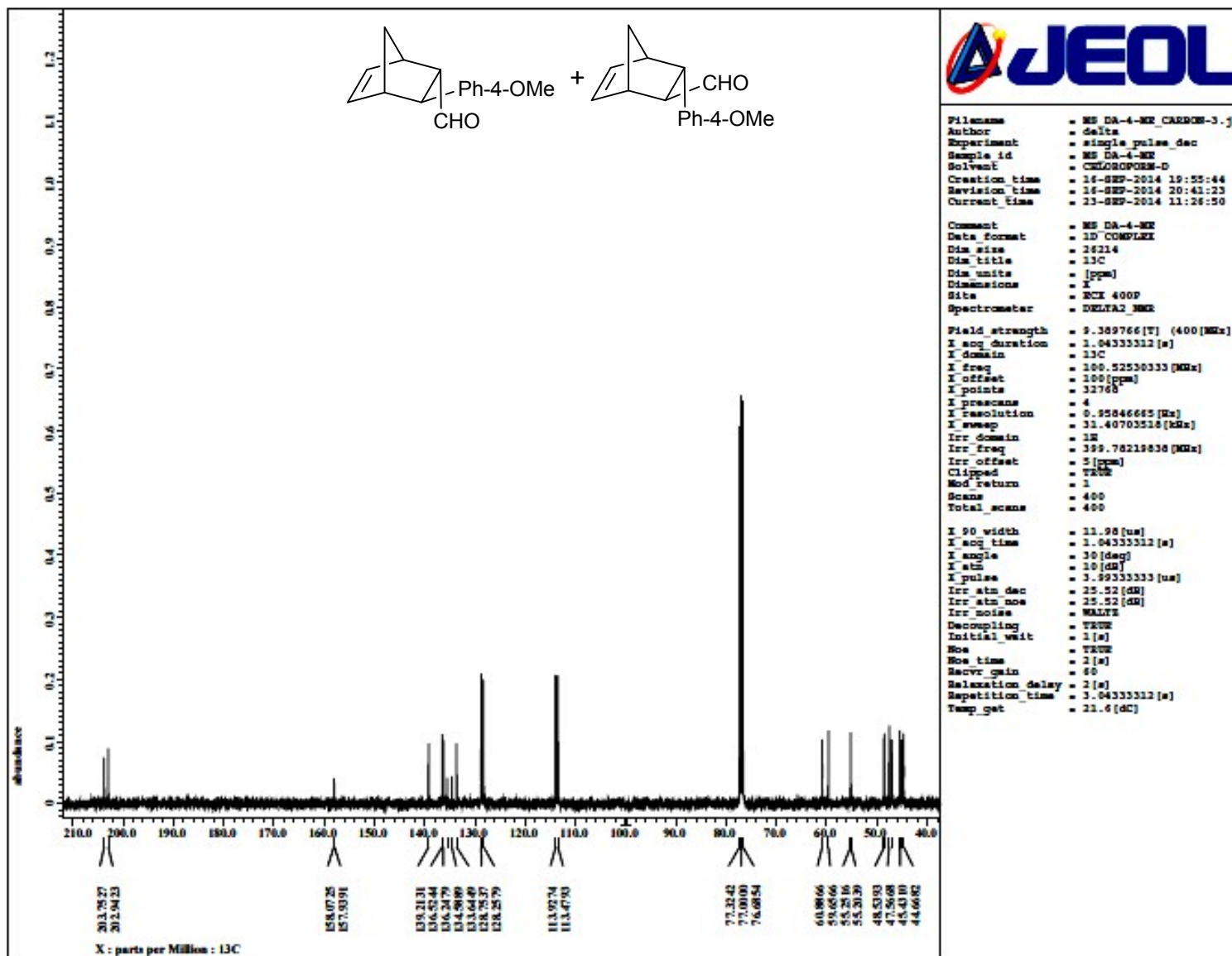
Filename      = 88 DA-4-CL CARBON-3.j
Author       = delta
Experiment   = single pulse dec
Sample id    = 88 DA-4-CL
Solvent      = CHLOROFORM-D
Creation time = 29-JUL-2014 09:21:22
Revision time = 29-JUL-2014 10:06:07
Current time  = 15-SEP-2014 12:02:49

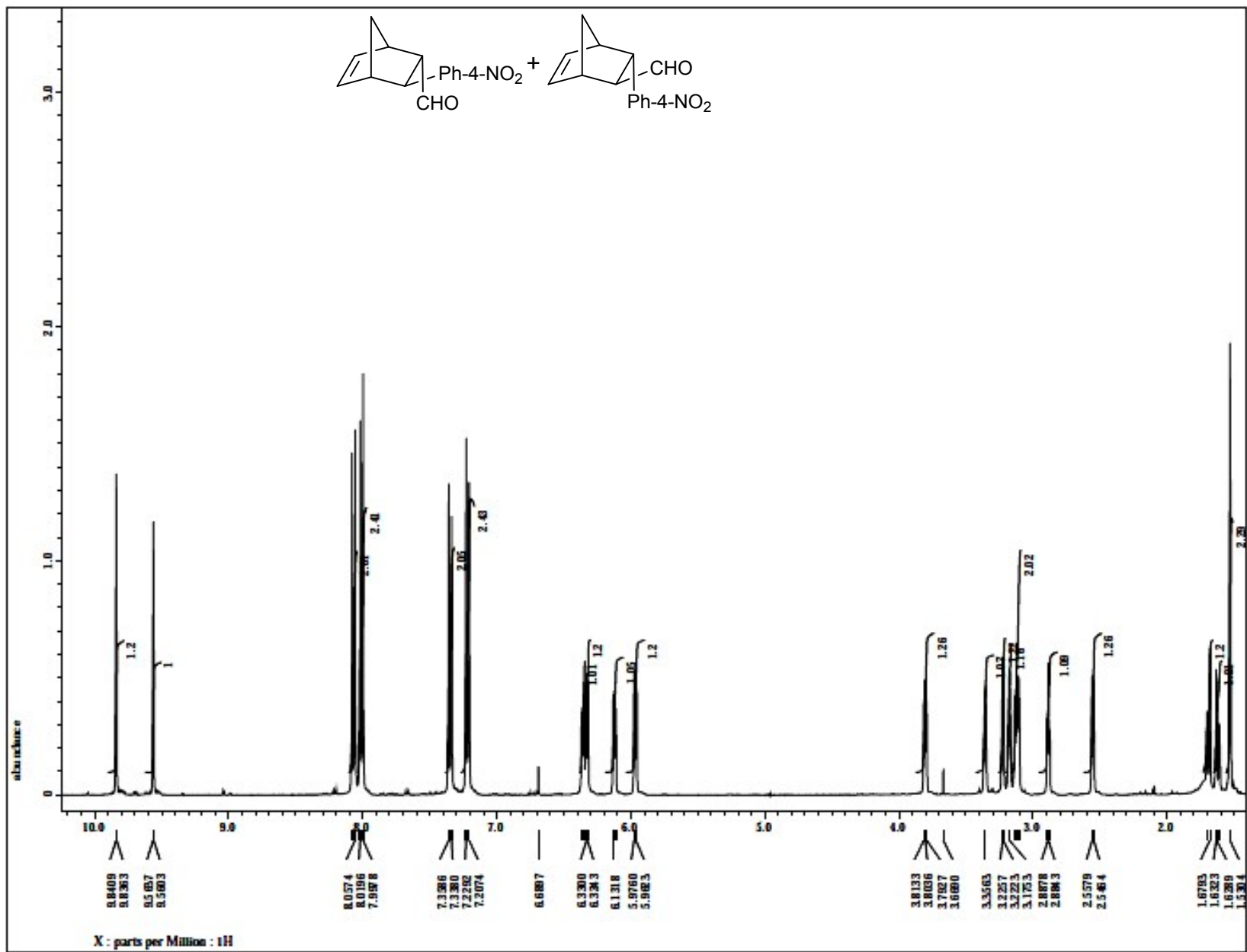
Comment      = 88 DA-4-CL
Data format  = 3D COMPLEX
Dim size     = 26214
Dim title    = 13C
Dim units    = [ppm]
Dimensions   = 1
Site         = KCC 400P
Spectrometer = DELTA2_NMR

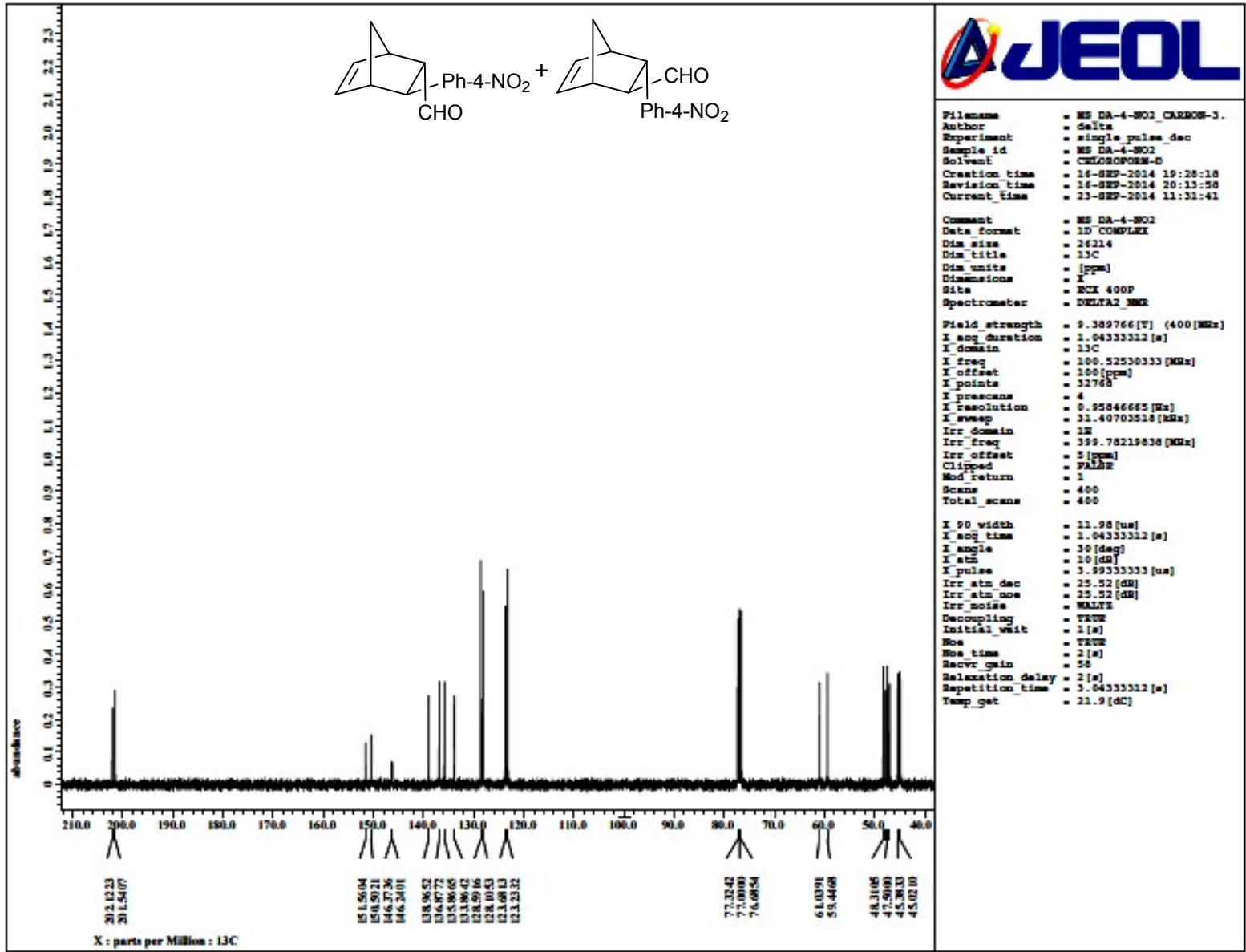
Field strength = 9.389766 [T] (400 [MHz])
F1 acq duration = 1.04333312 [s]
F1 domain      = 13C
F1 freq        = 100.52530333 [MHz]
F1 offset      = 100 [ppm]
F1 points      = 32768
F1 prescans    = 4
F1 resolution  = 0.95846665 [Hz]
F1 sweep       = 31.40703518 [kHz]
Irr domain     = 1H
Irr freq       = 399.78219838 [MHz]
Irr offset     = 5 [ppm]
Clipped       = FALSE
Mod return     = 1
Scans         = 400
Total_scans   = 400

F2 90 width   = 11.98 [us]
F2 acq time   = 1.04333312 [s]
F2 angle      = 30 [deg]
F2 stn        = 10 [dB]
F2 pulse      = 3.99333333 [us]
Irr stn dec   = 25.52 [dB]
Irr stn pow   = 25.52 [dB]
Irr noise     = WALTZ
Decoupling    = TRIP
Initial wait   = 1 [s]
Soc           = TRIP
Soc time      = 2 [s]
Socvr gain    = 60
Relaxation delay = 2 [s]
Repetition_time = 3.04333312 [s]
Temp_get      = 405.7 [dC]
  
```

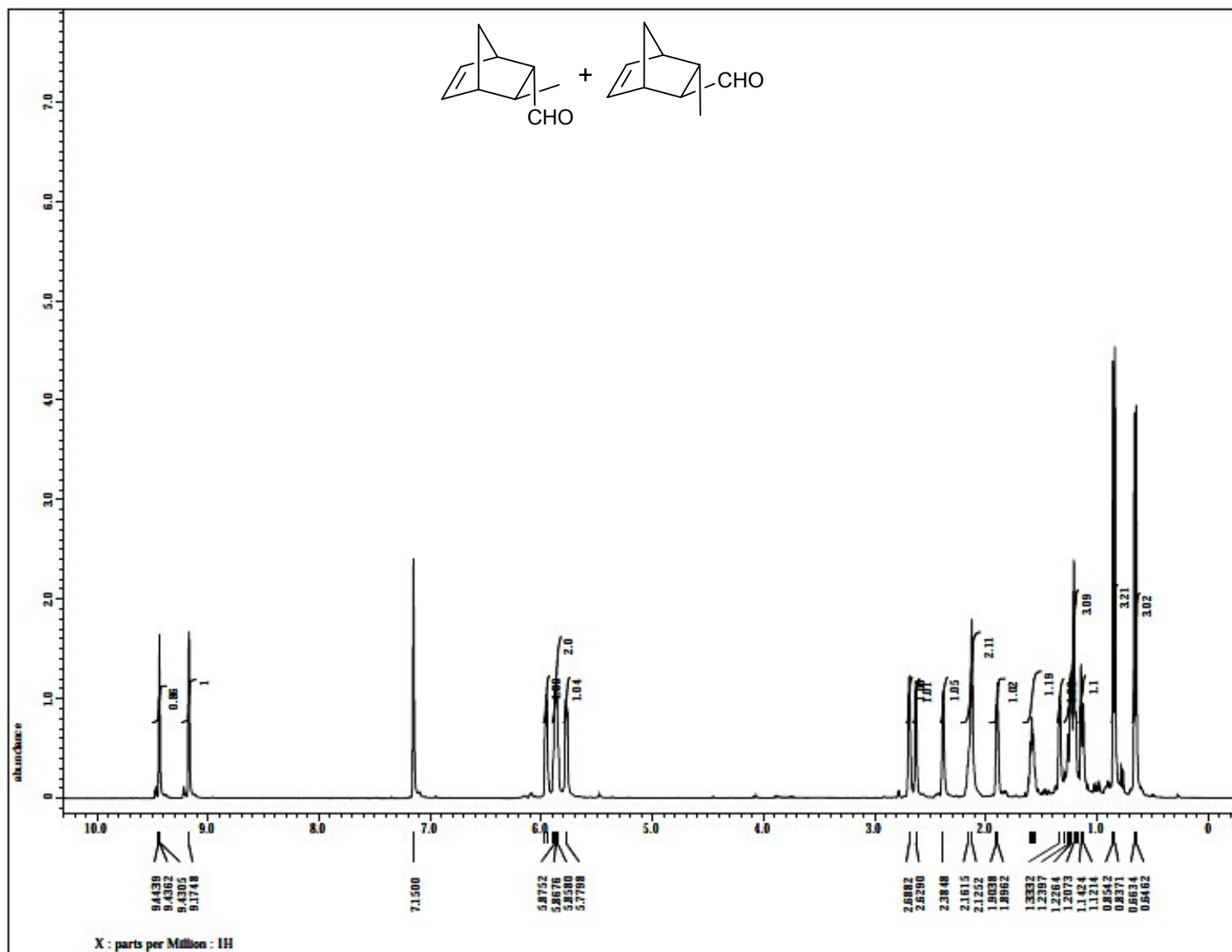


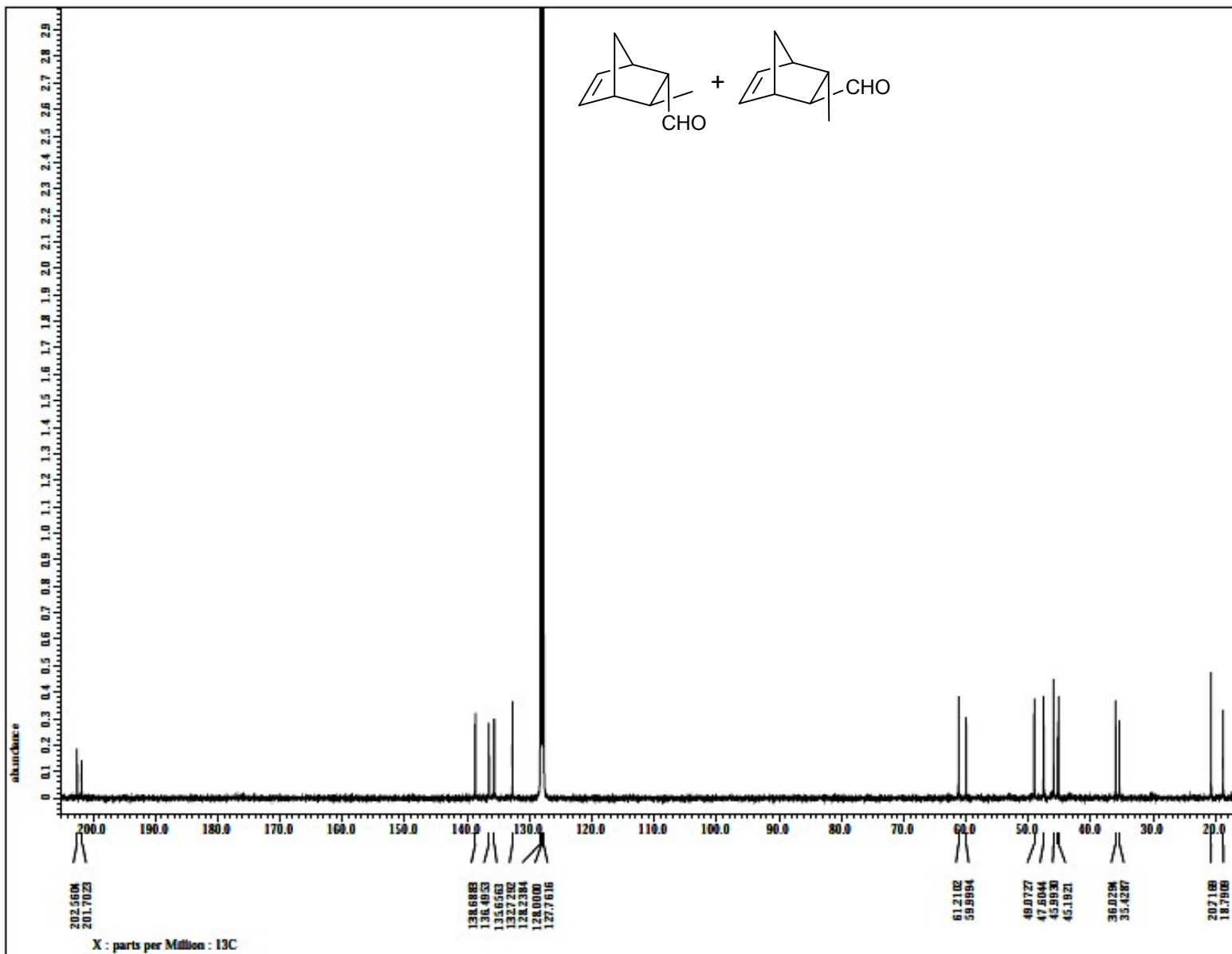






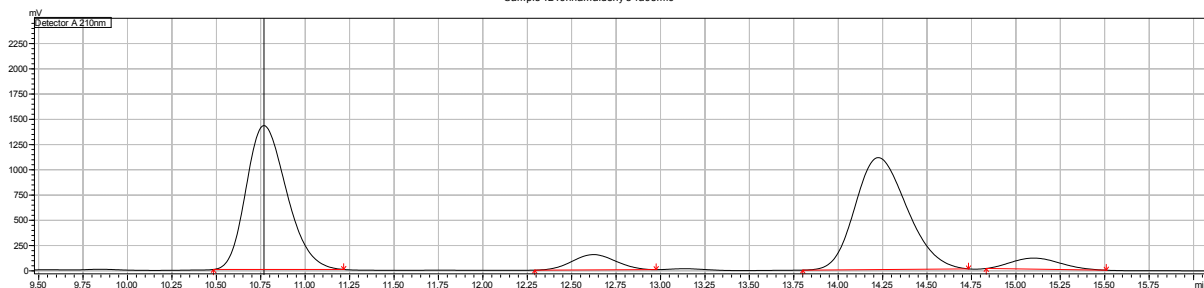






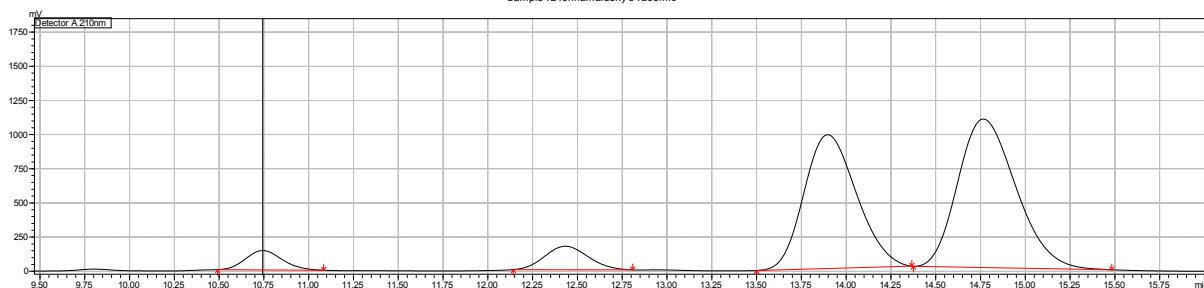
**(1R,3R,4S)-3-phenylbicyclo[2.2.1]hept-5-ene-2-carbaldehyde and (1R,2S,3R,4S)-3-phenylbicyclo[2.2.1] hept-5-ene-2-carbaldehyde** : HPLC conditions: Daicel Chiralcel OD-H column, Hexane/IPA (96 : 4), 1 ml/min., 210 nm.

Datafile Name: cnnamaldehyde racemic1.lcd  
 Sample Name: cnnamaldehyde racemic  
 Sample ID: cnnamaldehyde racemic



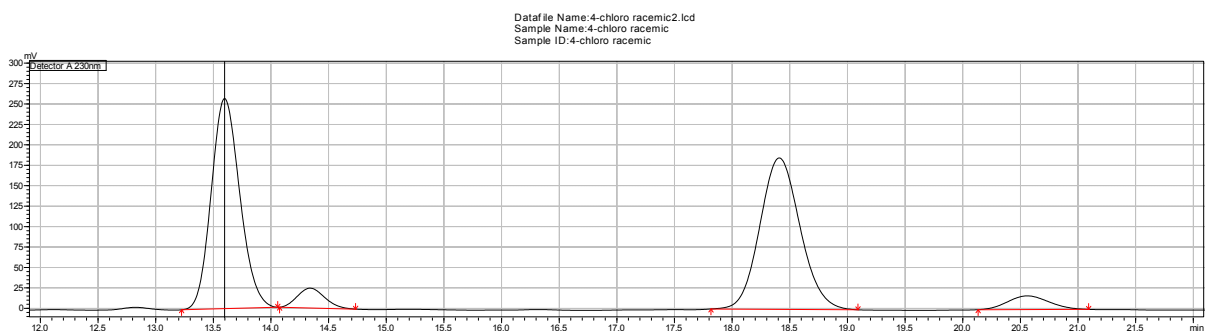
Peak	Ret. Time	Area	Height	Conc.	Area%
1	10.768	22546349	1425099	45.304	45.304 (exo)
2	12.622	2584002	151539	5.192	5.192 (endo)
3	14.226	22524433	1108584	45.260	45.260 (exo)
4	15.101	2111923	109650	4.244	4.244 (endo)
Total		49766707	2794872	100.000	100.000

Datafile Name: cnnamaldehyde Chiral2.lcd  
 Sample Name: cnnamaldehyde racemic  
 Sample ID: cnnamaldehyde racemic

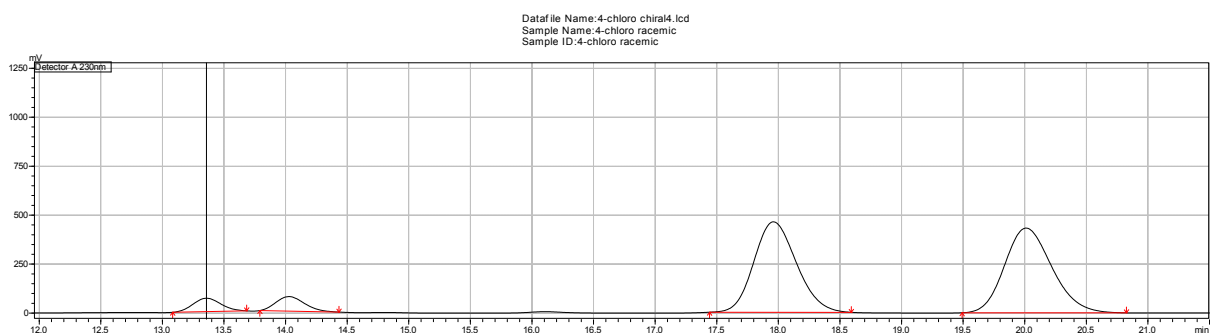


Peak	Ret. Time	Area	Height	Conc.	Area%
1	10.743	2008299	141376	4.069	4.069 (exo)
2	12.432	2895021	172031	5.866	5.866 (endo)
3	13.897	19958948	979508	40.440	40.440 (exo)
4	14.766	24492334	1087196	49.625	49.625 (endo)
Total		49354602	2380112	100.000	100.000

**(1R,3R,4S)-3-(4-chlorophenyl)bicyclo[2.2.1]hept-5-ene-2-carbaldehyde and (1R,2S,3R,4S)-3-(4-chlorophenyl)bicyclo[2.2.1]hept-5-ene-2-carbaldehyde** : HPLC conditions: Daicel Chiralcel OD-H column, Hexane/IPA (96 : 4), 0.8 ml/min., 230 nm.



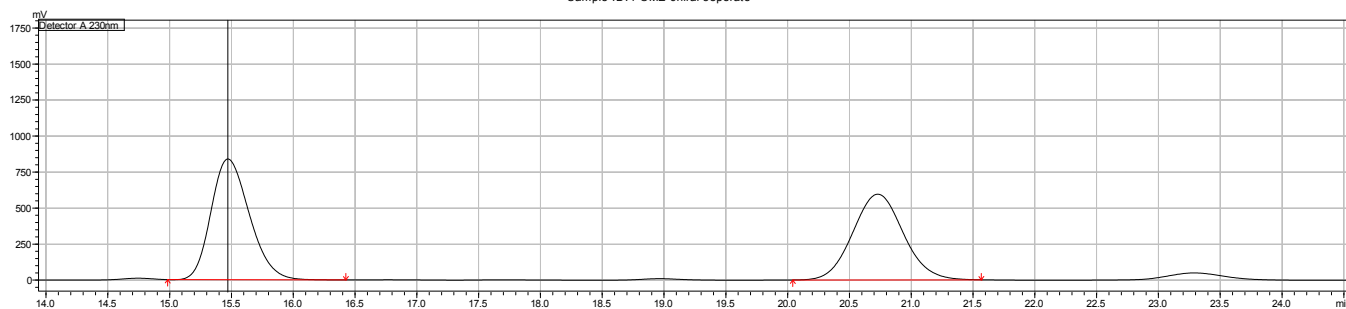
Peak	Ret. Time	Area	Height	Conc.	Area%
1	13.598	4431173	257148	45.474	45.474 (exo)
2	14.339	399294	24257	4.098	4.098 (endo)
3	18.409	4496334	185303	46.142	46.142 (exo)
4	20.560	417694	16749	4.286	4.286 (endo)
<b>Total</b>		<b>9744494</b>	<b>483458</b>	<b>100.000</b>	<b>100.000</b>



Peak	Ret. Time	Area	Height	Conc.	Area%
1	13.358	1078512	68644	4.328	4.328 (exo)
2	14.027	1193151	74026	4.788	4.788 (endo)
3	17.960	11091711	461775	44.507	44.507 (exo)
4	20.011	11558055	432663	46.378	46.378 (endo)
<b>Total</b>		<b>24921429</b>	<b>1037109</b>	<b>100.000</b>	<b>100.000</b>

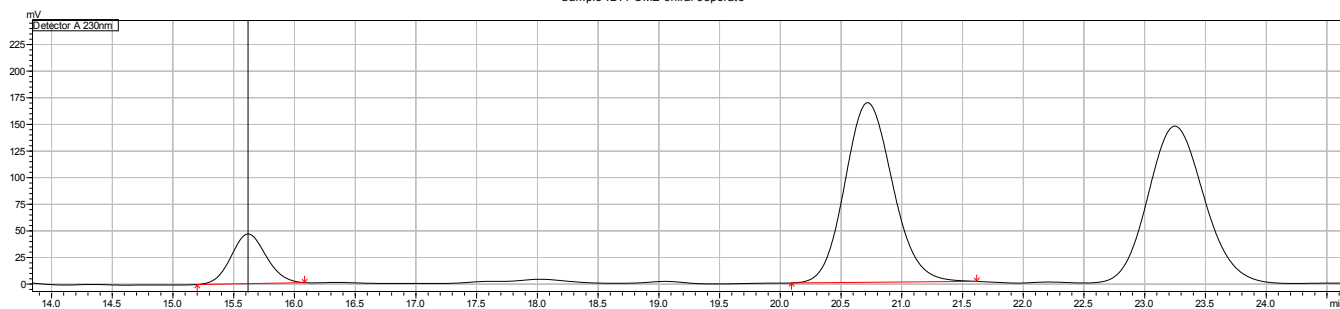
**(1S,3R,4R)-3-(4-methoxyphenyl)bicyclo[2.2.1]heptane-2-carbaldehyde and (1S,2S,3R,4R)-3-(4-methoxyphenyl)bicyclo[2.2.1]heptane-2-carbaldehyde** : HPLC conditions: Daicel Chiralcel OD-H column, Hexane/IPA (95 : 5), 0.8 ml/min., 230 nm.

Datafile Name: 4-OME chiral separate4.lcd  
 Sample Name: 4-OME chiral separate  
 Sample ID: 4-OME chiral separate



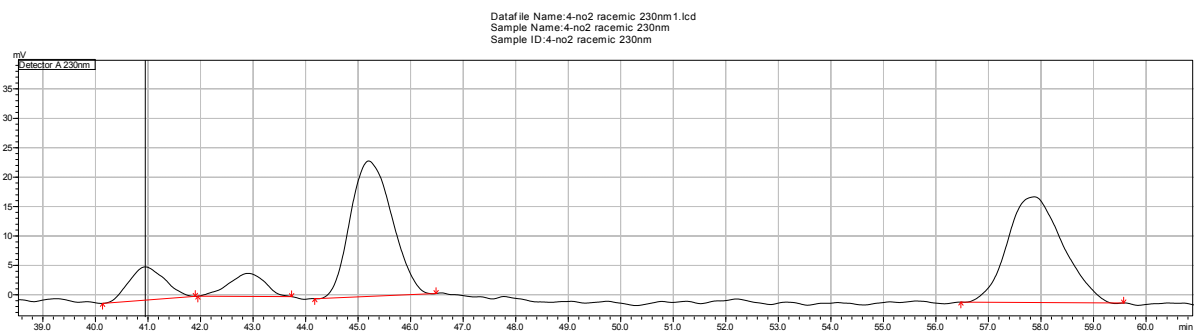
Peak	Ret. Time	Area	Height	Conc.	Area%
1	15.471	18095790	838075	51.653	51.653 (exo)
2	20.730	16937713	595246	48.347	48.347 (exo)
Total		35033502	1433321	100.000	100.000

Datafile Name: 4-OME chiral separate3.lcd  
 Sample Name: 4-OME chiral separate  
 Sample ID: 4-OME chiral separate

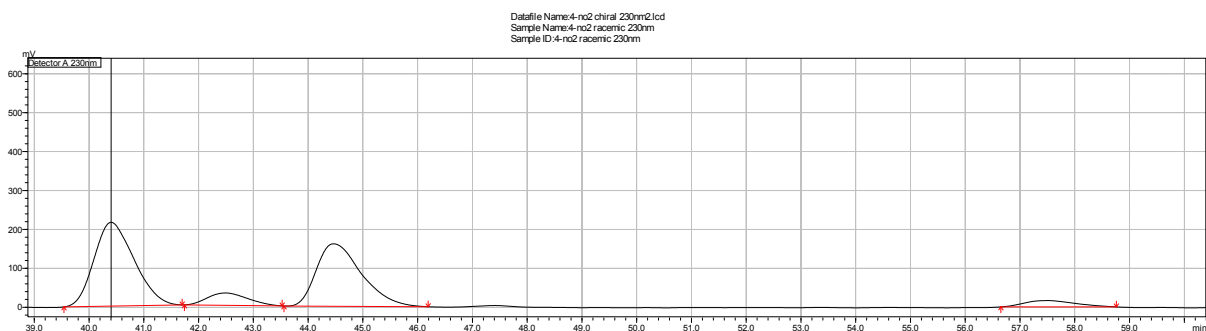


Peak	Ret. Time	Area	Height	Conc.	Area%
1	15.618	937248	46649	16.848	16.848 (exo)
2	20.719	4625840	168750	83.152	83.152 (exo)
Total		5563087	215400	100.000	100.000

**(1S,4R,6S)-5-methyl-6-(4-nitrophenyl)bicyclo[2.2.1]hept-2-ene and (1S,4R,5S,6S)-5-methyl-6-(4-nitrophenyl)bicyclo[2.2.1]hept-2-ene** : HPLC conditions: Daicel Chiralcel AD-H column, Hexane/IPA (96 : 5), 1.0 ml/min., 230 nm.



Peak	Ret. Time	Area	Height	Conc.	Area%
1	40.948	268078	5649	8.813	8.813 (exo)
2	42.900	187994	3914	6.180	6.180 (exo)
3	45.199	1269970	23027	41.748	41.748 (endo)
4	57.873	1315949	17964	43.259	43.259 (endo)
Total		3041992	50554	100.000	100.000



Peak	Ret. Time	Area	Height	Conc.	Area%
1	40.404	10932871	215431	47.978	47.978 (exo)
2	42.490	1616889	31720	7.096	7.096 (exo)
3	44.466	9186078	160450	40.312	40.312 (endo)
4	57.504	1051591	16296	4.615	4.615 (endo)
Total		22787428	423896	100.000	100.000



## GC Chromatogram:

**3-methylbicyclo[2.2.1]hept-5-ene-2-carbaldehyde** : Conversion of the product was determined using capillary RTX column (30 m × 0.25 mm): injector temperature 220°C, Pressure 88.6 kPa, column flow rate 1.10 ml/min, column temp 65 °C for 5 min. then 120 °C, temperature program 15°C/min, detector temp. = 250°C.

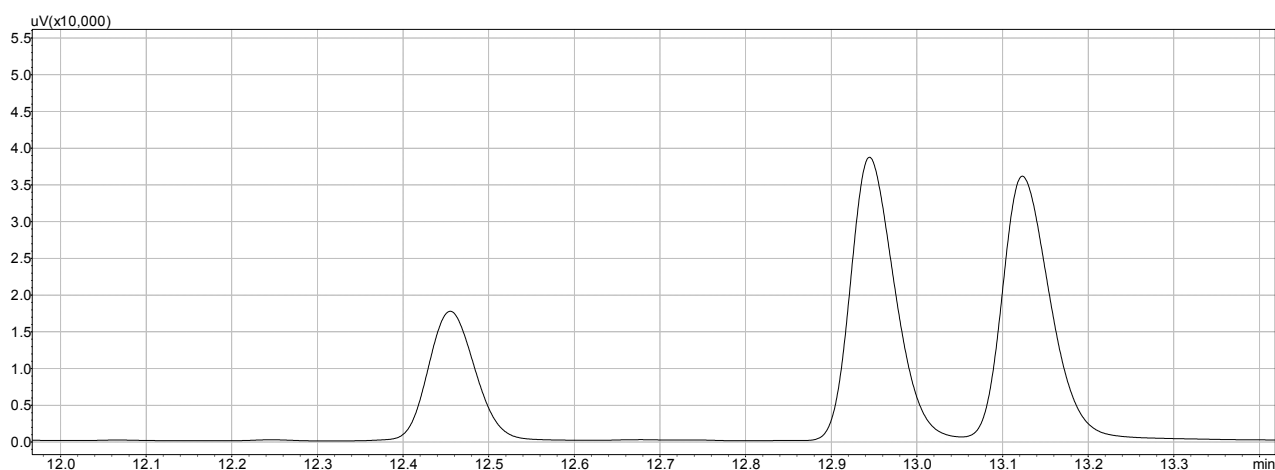


Peak	Ret.Time	Area	Height	Area%
1	2.510	124764.2	27386.6	6.7875
2	9.091	806280.5	330709.6	43.8637
3	9.361	907103.4	367640.0	49.3488

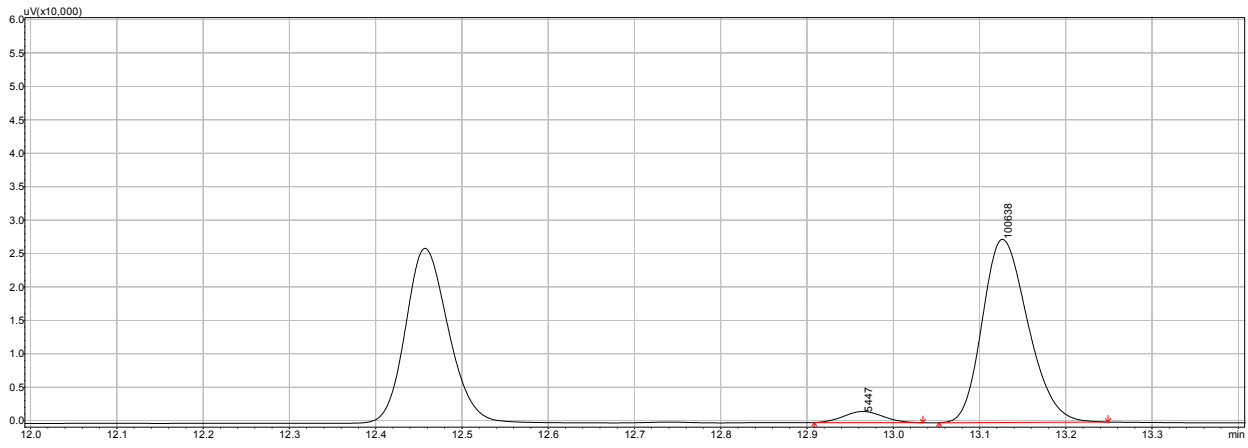
## Enantiomeric excess of the product 12

(Table 5, entry 1, fresh cycle)

GC condition : column information  $\beta$ -Dex L = 30m, 0.25mm ID, injector temperature 220°C, Pressure 88.6 kPa, column flow rate 1.10 ml/min, column temp 65 °C for 5 min. then 120 °C, temperature program 15°C/min, detector temp. = 250°C.

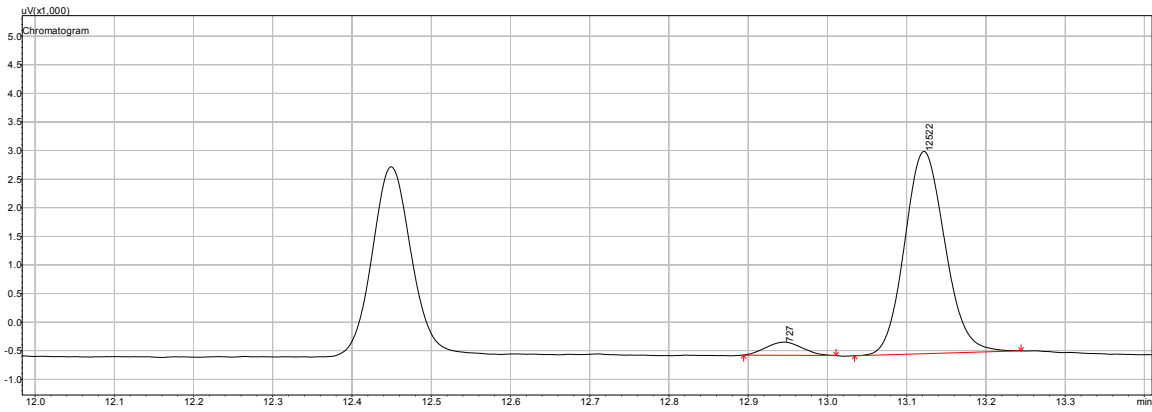


Peak	Ret.Time	Area	Height	Conc.	Area%
1	12.945	136015.8	37765.1	50.19428	50.1943 (endo)
2	13.123	134962.9	35241.2	49.80572	49.8057 (endo)



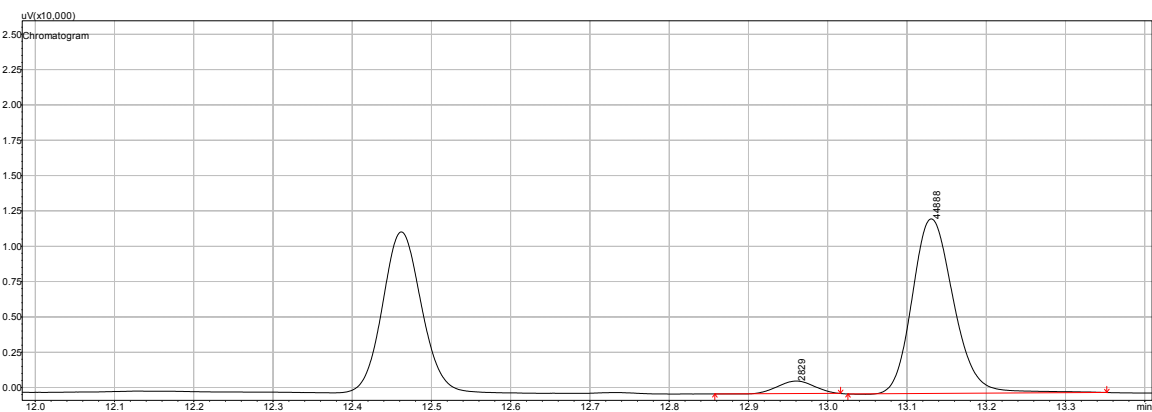
Peak	Ret.Time	Area	Height	Conc.	Area%
1	12.965	5446.8	1676.9	5.13439	5.1344
2	13.127	100637.7	27083.6	94.86561	94.8656

(Table 5, entry 2, 1<sup>st</sup> recycle)



Peak	Ret.Time	Area	Height	Conc.	Area%
1	12.945	726.8	227.3	5.48603	5.4860
2	13.122	12521.6	3482.7	94.51397	94.5140

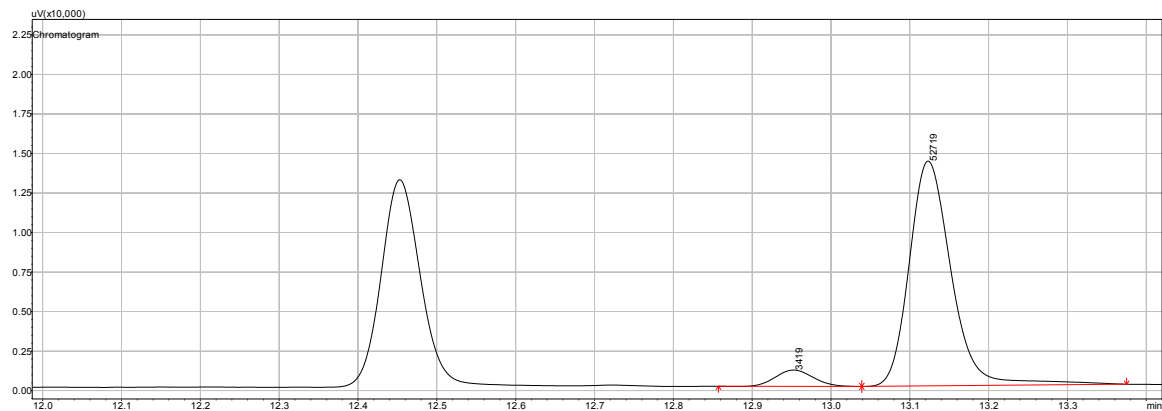
(Table 5, entry 3, 2<sup>nd</sup> recycle)



Peak	Ret.Time	Area	Height	Conc.	Area%
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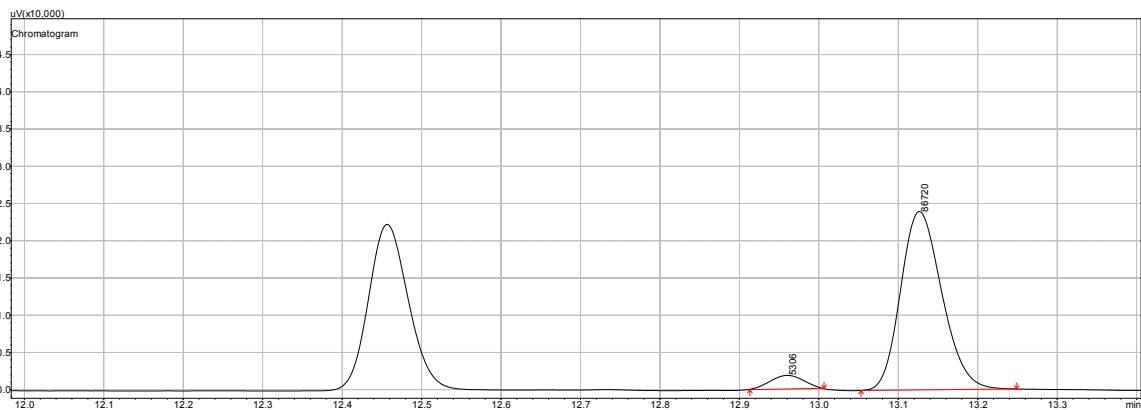
1	12.960	2829.3	886.1	5.92936	5.9294
2	13.131	44887.5	12344.4	94.07064	94.0706

(Table 5, entry 4, 3<sup>rd</sup> recycle)



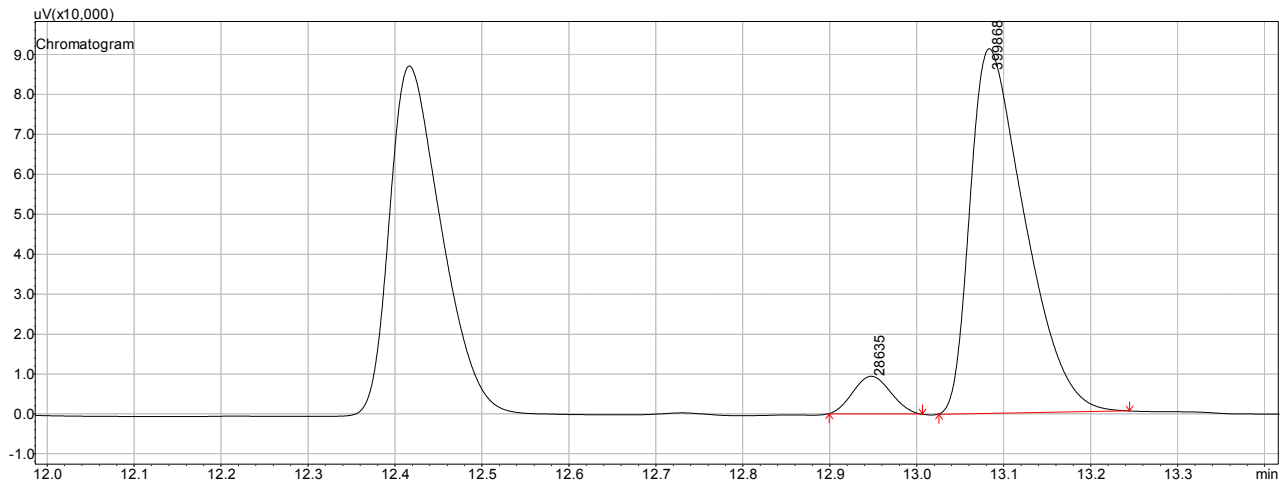
Peak	Ret.Time	Area	Height	Conc.	Area%
1	12.953	3418.5	1030.1	6.08953	6.0895
2	13.123	52719.3	14101.4	93.91047	93.9105

(Table 5, entry 5, 4<sup>th</sup> recycle)



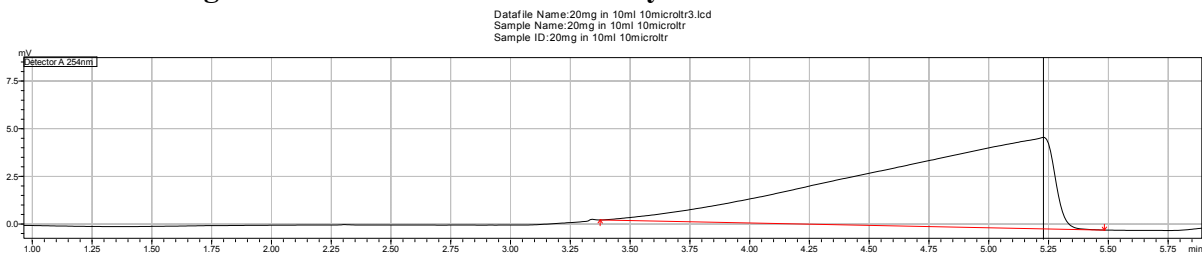
Peak	Ret.Time	Area	Height	Conc.	Area%
1	12.961	5305.9	1801.6	5.76568	5.7657
2	13.127	86719.6	23616.0	94.23432	94.2343

(Table 5, entry 6, 5<sup>th</sup> recycle)



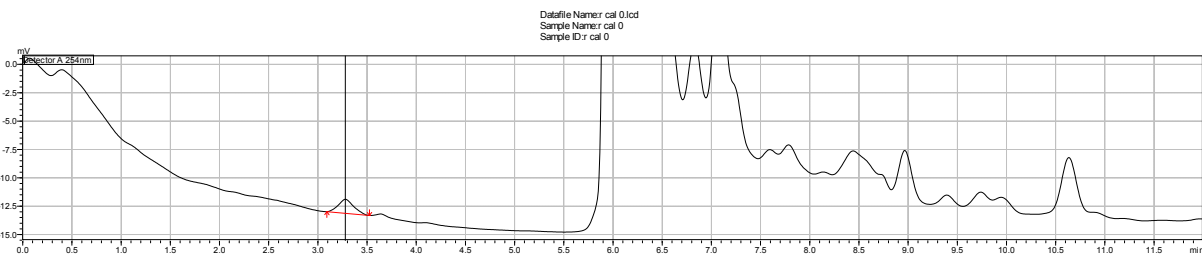
Peak	Ret.Time	Area	Height	Conc.	Area%
1	12.948	28635.1	9430.4	6.68258	6.6826
2	13.084	399868.0	91166.3	93.31742	93.3174

**HPLC chromatogram for fresh and leached catalyst:**



**Fresh catalyst**

Peak	Ret. Time	Area
1	5.228	261971
Total		261971



**Recovered catalyst and product**

Peak	Ret. Time	Area
1	3.279	13438
Total		13438