

CIELab Chromaticity Evolution to Measure Binding Free Energy of Non-colored Biomolecules to Gold Nanoparticles.

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Supporting Information

Obtention of CIELab parameters

XYZ colorimetric parameters were obtained from experimental measurements by using the following mathematical expressions:

$$X = K \sum_{\lambda} T_{\lambda} S_{\lambda} X_{10(\lambda)} \Delta_{\lambda}$$

$$Y = K \sum_{\lambda} T_{\lambda} S_{\lambda} Y_{10(\lambda)} \Delta_{\lambda}$$

$$Z = K \sum_{\lambda} T_{\lambda} S_{\lambda} Z_{10(\lambda)} \Delta_{\lambda}$$

$$K = 100 / \sum_{\lambda} S_{\lambda} \bar{Y}_{10(\lambda)} \Delta_{\lambda}$$

where T_{λ} is the transmittance of the sample; S_{λ} is a coefficient which depends on both λ and the illuminant (in our case, a D65 illuminant was employed) and $X_{10(\lambda)}$, $Y_{10(\lambda)}$, $Z_{10(\lambda)}$ are functions of both λ and the observer. Conversion from XYZ values to $L^*a^*b^*$ was done directly by using white point values for the D65 illuminant and 10° observer:²

$$X_n = 94.825; Y_n = 100; Z_n = 107.38$$

$L^*a^*b^*$ values were calculated as follows:³

$$L^* = 116 (Y/Y_n)^{1/3} - 16$$

$$a^* = 500[f(X/X_n) - f(Y/Y_n)]$$

$$b^* = 200[f(Y/Y_n) - f(Z/Z_n)]$$

where:

$$f(X/X_n) = (X/X_n)^{1/3}$$

$$f(Y/Y_n) = (Y/Y_n)^{1/3}$$

$$f(Z/Z_n) = (Z/Z_n)^{1/3}$$

Figures

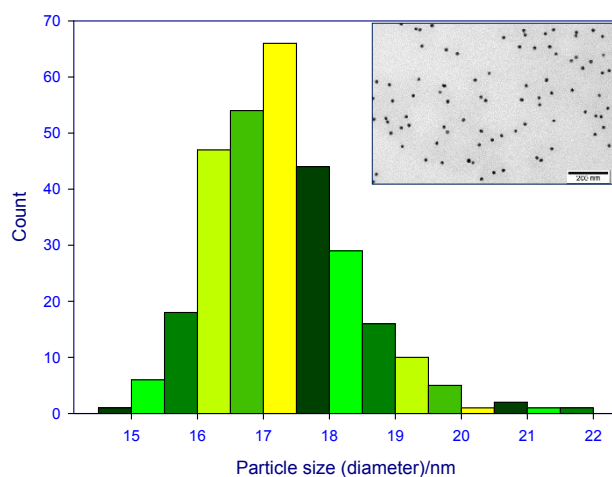


Figure S1. Size distribution of synthesized AuNPs.

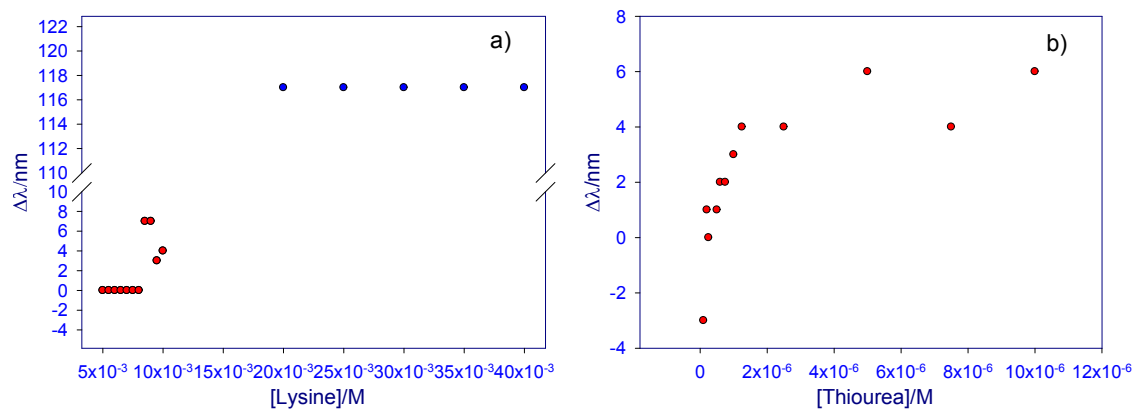


Figure S2. Wavelength shift ($\Delta\lambda$) of the maximum intensity absorbance peak for solutions containing $[\text{AuNPs}] = 3.2 \times 10^{-10} \text{ M}$ and varying concentrations of a) lysine, b) thiourea.

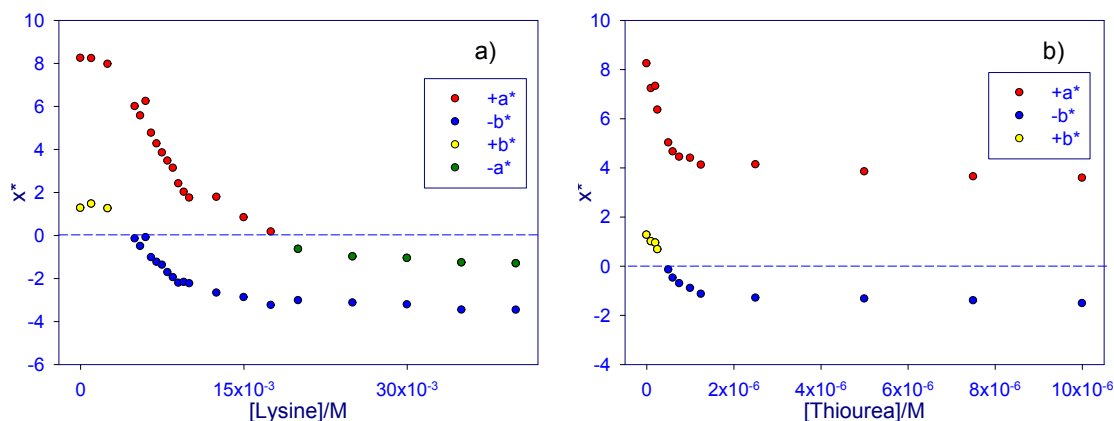


Figure S3. a^* and b^* parameters for a series of $[\text{AuNPs}] = 3.2 \times 10^{-10} \text{ M}$ solutions containing a) lysine and b) thiourea. Green-colored points indicate negative values of a^* which account for a green tone in the CIELab color system, and are indicative of fully blue (as opposed to purple) nanoparticle solutions.

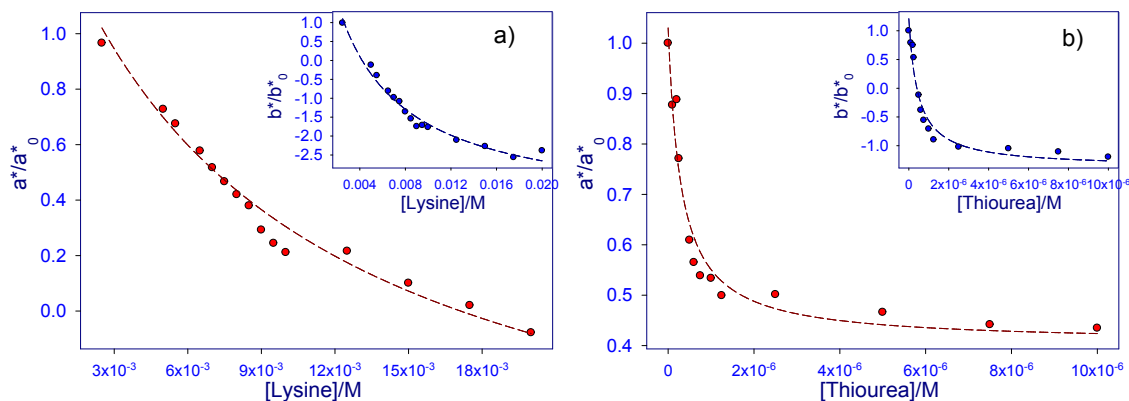


Figure S4. Two-state model fitting for normalized a^* and b^* (shown in the inset) parameters of AuNPs/biomolecule solutions. a) $[\text{AuNPs}] = 3.2 \times 10^{-10} \text{ M}$; $[\text{Lysine}] = 2.5 \times 10^{-3} - 2 \times 10^{-2} \text{ M}$, b) $[\text{AuNPs}] = 3.2 \times 10^{-10} \text{ M}$; $[\text{Thiourea}] = 0 - 1 \times 10^{-5} \text{ M}$.

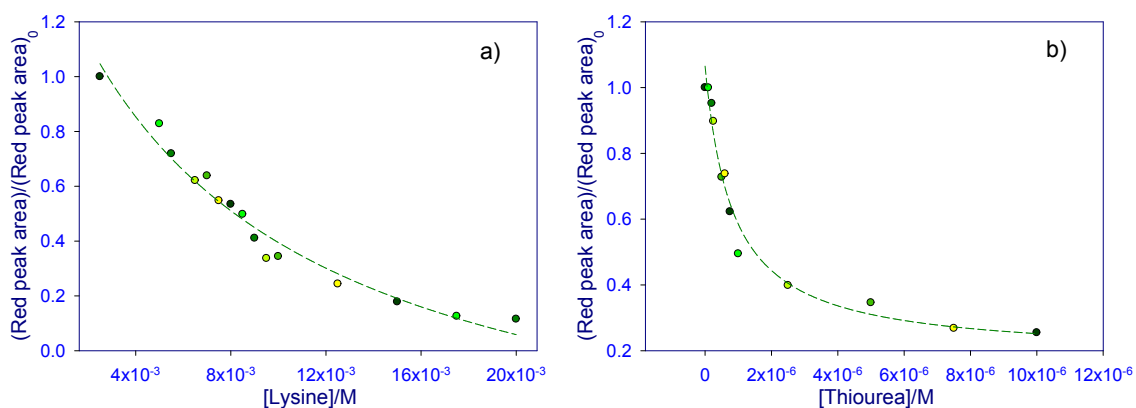


Figure S5. Two-state model fit for the red (non-aggregated) deconvolution peak area for a) a series of AuNPs/lysine solutions ranging from $[\text{Lysine}] = 2.5 \times 10^{-3} - 2 \times 10^{-2} \text{ M}$ and b) a series of AuNPs/thiourea solutions ranging from $[\text{Thiourea}] = 0 - 1 \times 10^{-5} \text{ M}$.

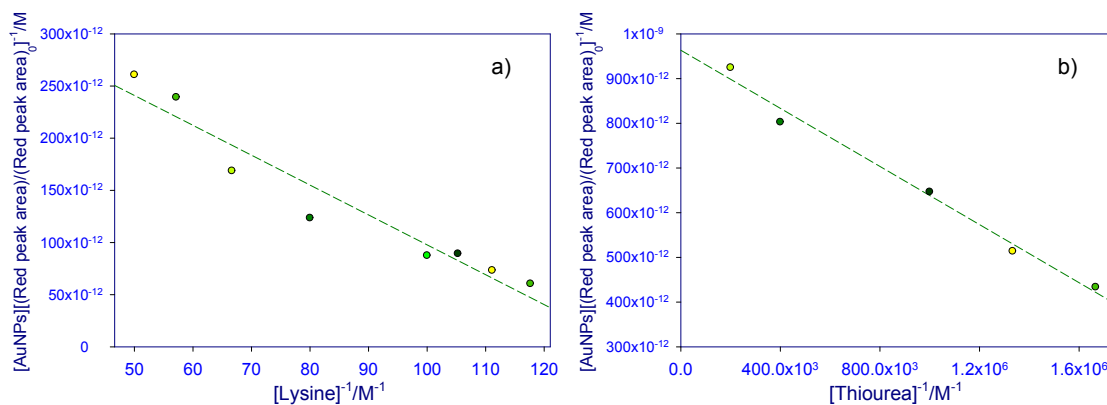


Figure S6. Benesi-Hildebrand fit for the normalized red peak area obtained from deconvolution procedures for a) AuNPs/lysine solutions ranging from $[\text{Lysine}] = 8.5 \times 10^{-3}$ to 2×10^{-2} M and b) AuNPs/thiourea solutions ranging from $[\text{Thiourea}] = 6 \times 10^{-7}$ to 5×10^{-6} M.

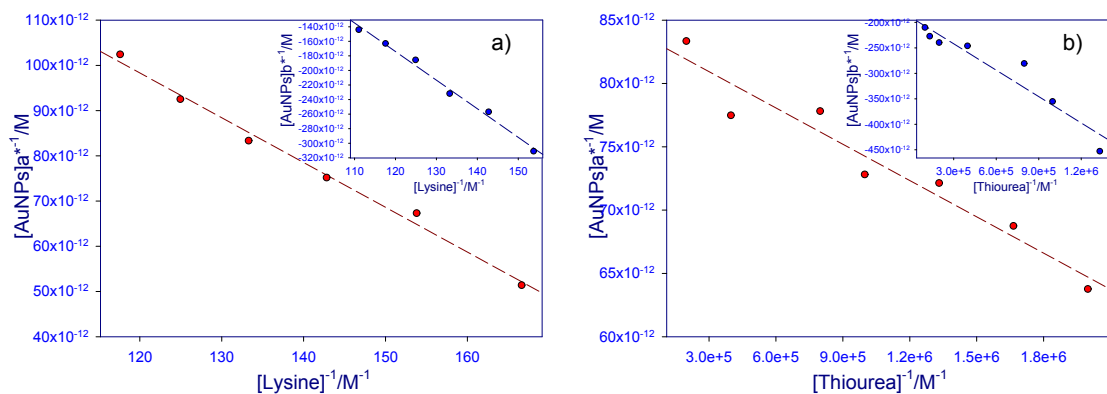


Figure S7. Benesi-Hildebrand fit for a^* and b^* (shown on inset) for a series of AuNPs solutions ranging from a) $[\text{Lysine}] = 6 \times 10^{-3}$ to 9×10^{-3} M and b) $[\text{Thiourea}] = 5 \times 10^{-7}$ to 1×10^{-5} M.