

Supporting information for

## Hydrogelation by Bile acid-Peptide Conjugates and *in situ* Synthesis of Silver and Gold Nanoparticles in Hydrogel Matrix

Mitasree Maity, Valiparambil Sanjayan Sajisha and Uday Maitra\*

Department of Organic Chemistry, Indian Institute of Science, Bangalore- 560012,  
India

Email: maitra@orgchem.iisc.ernet.in

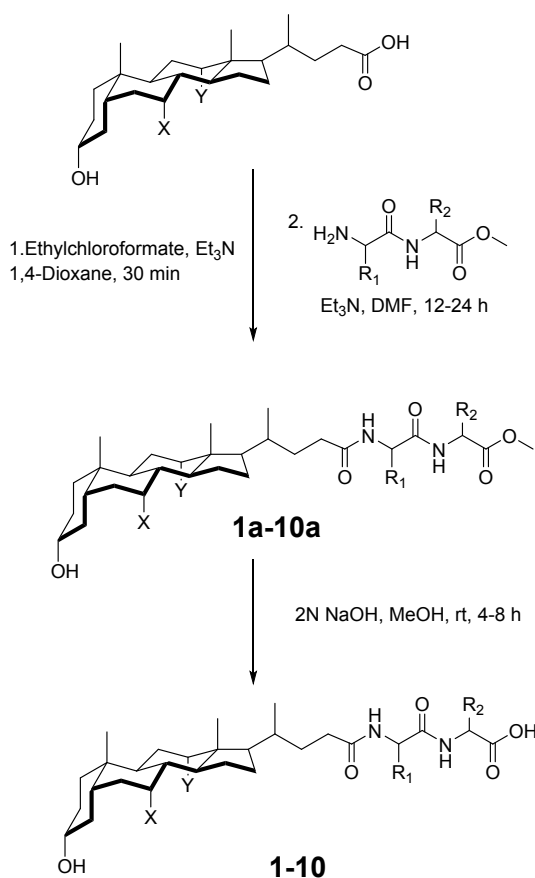
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## General considerations

All starting materials and solvents were obtained from commercial sources and were used without further purification. Distilled water was used for preparation of sodium phosphate buffer and NaOH for gelation study. Melting points of the product were determined on a Digimelt melting point apparatus.  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectra were recorded on a Bruker NMR spectrometer at 400 MHz and 100 MHz, respectively. Scanning electron micrographs were recorded on a SIRION instrument. Prior to the measurements the dried gels were coated with a thin gold layer. AFM imaging was carried out on a JPK NANO WIZARD II in non-contact mode. UV spectrums were recorded on Perkin Elmer. Transmission electron microscopy (TEM) was done on a JEM 2100F instrument. Samples were placed on a carbon-coated copper grid with subsequent drying and staining with uranyl acetate, followed by air and then vacuum drying.

## General Synthetic scheme for the bile acid peptide conjugates



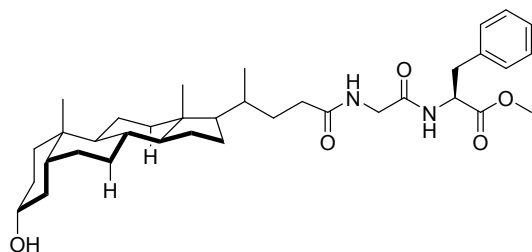
LC- Lithocholic acid (X=Y=H), DC- Deoxycholic acid (X=H, Y=OH), C- Cholic acid (X=Y=OH).

## Bile acid-peptied conjugates synthesis

All reactions were carried out in oven dried glasswares. N-terminal of di-peptied (Boc-A<sub>1</sub>-A<sub>2</sub>-OMe, 0.2 mmol, 1 eq) was deprotected by trifluoroacetic acid (TFA, 0.8 mmol, 4 eq) in DCM stirring for 2 hour. Solvent was evaporated in high vacuum to get TFA salt. TFA salt was stirred with Et<sub>3</sub>N (0.3 mmol, 1.5 eq) in DMF at room temperature for 30 min to get corresponding free amine. At the same time in another flask, bile acid (0.13 mmol, 0.67 eq) was dissolved in 1,4-Dioxane at 10 °C, then Et<sub>3</sub>N (0.2 mmol, 1 eq) was added to it and followed by ethyl chloroformate (0.2 mmol, 1 eq) and stirred it at room temperature for 30 min to prepared mixed anhydride. Free amine-dipeptied methyl ester in DMF was directly added to the freshly prepared bile acid-anhydride. Stirring was continued for 12-24h and monitor by TLC. After completion of the reaction, the mixture was diluted with ethyl acetate, was washed with water, and brine, and dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. The crude product was purified by column chromatography on silica gel (100-200 mesh), using ethyl acetate/petroleum ether as the eluent. The hydrolysis of bile acid-dipeptide methyl ester conjugates were done with aqueous NaOH in MeOH and progress of reactions were monitored by TLC. Methanol was removed under reduced pressure and the solution was acidified with 1 (N) hydrochloric acid until a precipitate was formed. The resulting suspension was filtered under vacuum. The pure product was obtained by column chromatography 100-200 mesh silica gel with MeOH/CHCl<sub>3</sub>.

## Data of compounds

### LC-G-<sup>L</sup>F-OMe (1a)



Yield- 68%, white solid, Mp- 64-70 °C.

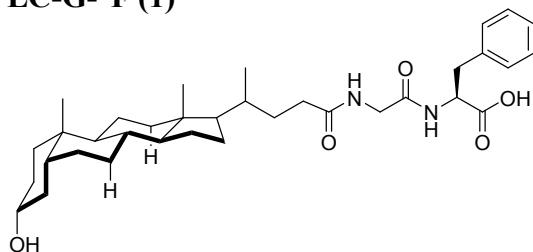
IR (KBr): 3302, 3066, 3029, 2934, 2864, 1748, 1659, 1652, 1538, 1446, 1376, 1214, 1177, 1110, 1068, 1032, 965, 945, 854, 813, 753, 701, 664, 605 cm<sup>-1</sup>.

<sup>1</sup>H NMR (400 MHz, Solvent- CDCl<sub>3</sub>) δ: 7.28-7.22 (m, 3H), 7.11-7.09 (d, 2H, *J* = 8 Hz), 6.76-6.74 (m, 1H, NH), 6.40 (s, 1H, NH), 4.87-4.82 (m, 1H), 3.96-3.83 (dq, 2H, *J* = 16.8 Hz, 4 Hz), 3.72 (s, 3H), 3.64-3.58 (m, 1H), 3.17-3.04 (m, 2H), 2.31-0.96 (m, 29H, steroidal skeleton), 0.92-0.91 (m, 6H), 0.64 (s, 3H).

<sup>13</sup>C NMR (100 MHz, Solvent- CDCl<sub>3</sub>) δ: 174.2, 171.7, 168.8, 135.7, 129.2, 128.6, 127.2, 71.9, 56.5, 56.0, 53.3, 52.5, 43.1, 42.8, 42.1, 40.5, 40.2, 37.9, 36.5, 35.9, 35.6, 35.4, 34.6, 33.3, 31.7, 30.6, 28.3, 27.2, 26.5, 24.3, 23.4, 20.9, 18.4, 12.1.

Mass Spec. (EI) *m/z*: M. Wt. calcd for C<sub>36</sub>H<sub>54</sub>N<sub>2</sub>O<sub>5</sub>Na - 617.39; observed - 617.3931 (M+Na).

### LC-G-<sup>L</sup>F (1)



Yield- 90%, white solid, Mp- 110-125 °C.

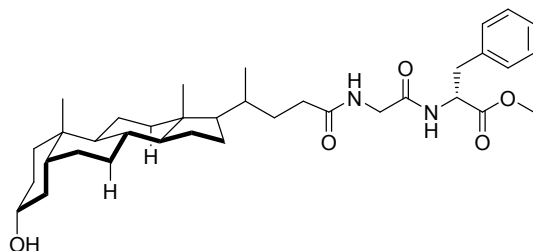
IR (KBr): 3322, 3062, 3029, 2936, 2865, 2519, 1731, 1660, 1531, 1455, 1378, 1335, 1218, 1113, 1067, 1031, 1013, 965, 945, 829, 805, 755, 700, 665, 494 cm<sup>-1</sup>.

<sup>1</sup>H MNR (400 MHz, Solvent- CD<sub>3</sub>OD) δ: 7.29-7.21 (m, 5H), 4.69-4.66 (m, 1H), 3.88-3.76 (m, 2H), 3.58-3.50 (m, 1H), 3.22-3.17 (m, 1H), 3.04-2.99 (dd, 1H, *J* = 16, 8 Hz), 2.33-0.96 (m, 29H, steroidal skeleton protons), 0.95 (s, 6H), 0.69 (s, 3H).

<sup>13</sup>C NMR (100 MHz, Solvent- CD<sub>3</sub>OD) δ: 177.1, 171.3, 138.1, 130.4, 129.25, 127.8, 79.5, 72.4, 57.9, 57.4, 54.9, 43.9, 43.5, 43.3, 41.9, 41.5, 38.4, 37.2, 37.1, 36.8, 36.5, 35.7, 33.8, 33.0, 31.2, 29.2, 28.3, 27.6, 25.3, 23.9, 21.9, 18.9, 12.5.

Mass Spec.(EI) m/z: M. Wt. calcd for C<sub>35</sub>H<sub>52</sub>N<sub>2</sub>O<sub>5</sub>Na – 603.37; observed – 603.3774 (M+Na).

### LC-G-<sup>D</sup>F-OMe (2a)



Yield- 91%, white solid, Mp- 66-74 °C.

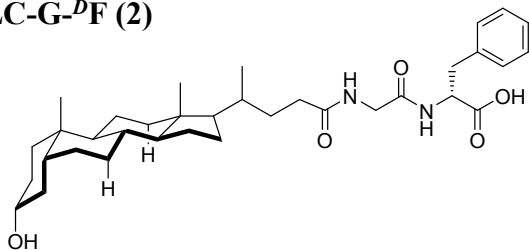
IR (KBr): 3271, 2925, 2860, 1742, 1645, 1528, 1443, 1375, 1212, 1032, 743 cm<sup>-1</sup>.

<sup>1</sup>H MNR (400 MHz, Solvent- CDCl<sub>3</sub>) δ: 7.29-7.24 (m, 3H), 7.10-7.08 (d, 2H, *J* = 8 Hz), 6.55-6.53 (d, 1H, *J* = 7.2 Hz, NH), 6.24-6.23 (d, 1H, *J* = 4 Hz, NH), 4.88-4.83 (m, 1H), 3.94-3.85 (m, 2H), 3.73 (s, 3H), 3.65-3.59 (m, 1H, C<sub>3</sub>-H), 3.17-3.05 (m, 2H), 2.31-0.986 (m, 29H, steroidal skeleton protons), 0.91 (m, 6H), 0.64 (s, 3H).

<sup>13</sup>C NMR (100 MHz, Solvent- CDCl<sub>3</sub>) δ: 174.1, 171.5, 168.6, 135.5, 129.1, 128.5, 127.1, 71.7, 56.4, 55.9, 53.1, 52.3, 42.9, 42.6, 42.0, 40.3, 40.1, 37.7, 36.3, 35.7, 35.4, 35.2, 34.5, 33.1, 31.5, 30.4, 28.1, 27.1, 26.3, 24.1, 23.3, 20.7, 18.3, 11.9.

Mass Spec.(EI) m/z: M. Wt. calcd for C<sub>36</sub>H<sub>54</sub>N<sub>2</sub>O<sub>5</sub>Na - 617.3930; observed - 617.3931 (M + Na).

### LC-G-*D*F (2)



Yield- 95%, white solid, Mp- 209-215 °C.

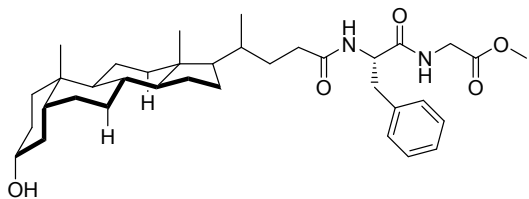
IR (KBr): 3481, 3400, 3028, 2936, 2866, 2520, 1715, 1655, 1629, 1537, 1516, 1446, 1381, 1368, 1344, 1305, 1280, 1246, 1223, 1186, 1125, 1106, 1068, 1035, 1012, 946, 861, 801, 739, 701, 617  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz, Solvent-  $\text{DMSO-d}_6$ )  $\delta$ : 8.05 (d, 1H,  $J = 7.6$  Hz), 7.95 (t, 1H,  $J =$  Hz), 7.25-7.18 (m, 5H), 4.43-4.38 (m, 2H), 3.70-3.56 (m, 2H), 3.05-3.00 (m, 1H,  $\text{C}_3\text{-H}$ ), 2.89-2.83 (m, 1H), 2.18-1.10 (m, steroid back bone protons), 0.86 (s, 6H), 0.59 (s, 3H).

$^{13}\text{C}$  NMR (100 MHz, Solvent-  $\text{DMSO-d}_6$ )  $\delta$ : 173.2, 173.1, 169.3, 137.8, 129.5, 128.5, 126.8, 70.2, 56.4, 55.9, 53.7, 42.6, 42.0, 41.9, 37.2, 36.6, 35.7, 35.3, 34.6, 32.4, 31.7, 30.7, 28.1, 27.2, 26.5, 24.2, 23.6, 20.8, 18.6, 12.2.

Mass Spec.(EI )  $m/z$ : M. Wt. calcd for  $\text{C}_{35}\text{H}_{52}\text{N}_2\text{O}_5\text{Na}$  - 603.37; observed - 603.3773 (M + Na).

### LC-*L*F-G-OMe (3a)



Yield- 95%, white solid, Mp- 83-86 °C.

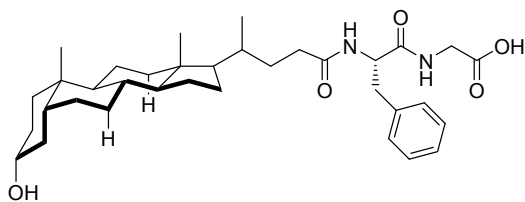
IR (KBr): 3400, 3294, 2936, 2865, 1756, 1654, 1545, 1445, 1371, 1207, 1181, 1068, 1034  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz, Solvent-  $\text{CDCl}_3$ )  $\delta$ : 7.29-7.20 (m, 5H), 6.62 (s, 1H), 6.20-6.18 (d, 1H,  $J = 8$  Hz), 4.76-4.70 (q, 1H,  $J = 8$  Hz), 4.03-3.88 (dq, 2H,  $J = 17.6$  Hz, 5.6 Hz), 3.73 (s, 3H), 3.65-3.59 (m, 1H), 3.13-3.04 (m, 2H), 2.25-0.96 (m, steroidal CH and  $\text{CH}_2$  peak) 0.92 (s, 3H), 0.89-0.87 (d, 3H,  $J = 8$  Hz), 0.65 (s, 3H).

$^{13}\text{C}$  NMR (100 MHz, Solvent-  $\text{CDCl}_3$ )  $\delta$ : 173.8, 171.3, 169.7, 136.4, 129.2, 128.5, 126.9, 71.7, 56.4, 55.8, 54.0, 52.2, 42.6, 42.0, 41.08, 40.3, 40.09, 37.9, 36.3, 35.7, 35.3, 34.5, 33.3, 31.5, 30.5, 28.1, 27.1, 26.3, 24.1, 23.3, 20.7, 18.2, 11.9.

Mass Spec.(EI )  $m/z$ : M. Wt. calcd. for  $\text{C}_{36}\text{H}_{54}\text{N}_2\text{O}_5\text{Na}$  - 617.58; observed- 617.3930 (M + Na).

### LC-<sup>L</sup>F-G (3)



Yield - 86%, white solid, Mp- 115-117 °C.

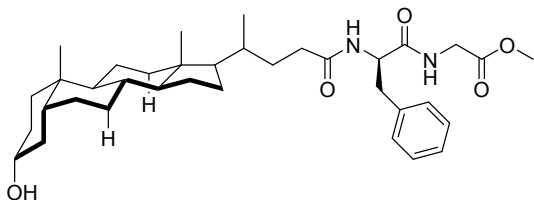
IR (KBr): 3405, 2931, 2863, 1736, 1654, 1638, 1527, 1448, 1384, 1213, 1195, 1033 cm<sup>-1</sup>.

<sup>1</sup>H NMR (400 MHz, Solvent- MeOH-d<sub>4</sub>) δ: 7.9 (s, 1H), 7.29-7.20 (m, 5H), 4.70-4.66 (dd, 1H, *J* = 8 Hz, 4.8 Hz), 3.90 (s, 2H), 3.57-3.52 (m, 1H), 3.24-3.18 (dd, 1H, *J* = 14 Hz, 5.2 Hz) 2.90-2.84 (dd, 1H, *J* = 14 Hz, 10 Hz), 2.3-0.96 (m, steroidal CH and CH<sub>2</sub> peak), 0.95 (s, 3H), 0.91-0.89 (d, 3H, *J* = 8 Hz), 0.65 (s, 3H).

<sup>13</sup>C NMR (100 MHz, Solvent- MeOH-d<sub>4</sub>) δ: 175.2, 172.7, 137.2, 128.8, 127.9, 126.2, 78.0, 70.9, 56.4, 55.9, 54.2, 42.4, 40.6, 42.1, 40.6, 40.4, 40.0, 37.3, 35.8, 35.7, 35.3, 35.0, 34.2, 32.4, 31.6, 29.7, 27.7, 26.9, 26.2, 23.8, 22.5, 20.5, 17.4, 11.1.

Mass Spec.(EI) m/z: M. Wt. calcd. for C<sub>35</sub>H<sub>52</sub>N<sub>2</sub>O<sub>5</sub>Na - 603.58; observed - 603.3776 (M+Na).

### LC-<sup>D</sup>F-G-OMe (4a)



Yield- 50%, white solid, Mp- 163.1-166.3 °C.

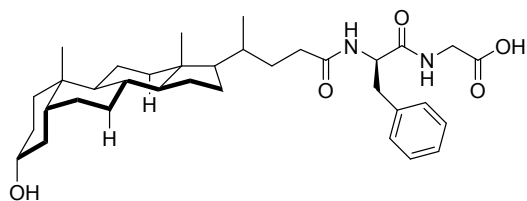
IR (KBr): 3329, 3290, 2930, 2860, 1759, 1654, 1545, 1445, 1371, 1207, 1181, 1068, 1024 cm<sup>-1</sup>.

<sup>1</sup>H NMR (400 MHz, Solvent- CDCl<sub>3</sub>) δ: 7.29-7.20 (m, 5 H), 6.69 (s, 1H, NH), 6.26-6.24 (s, 1H, *J* = 8 Hz), 4.78-4.72 (q, 1H, *J* = 8 Hz), 4.03-3.89 (dq, 2H, *J* = 18.4 Hz, 4 Hz), 3.72 (s, 3H), 3.64-3.59 (m, 1H), 3.12-3.03 (m, 2H), 2.3-0.97 (m, steroid protons), 0.96 (s, 3H), 0.87-0.85 (d, 3H), 0.62 (s, 3H).

<sup>13</sup>C NMR (100 MHz, Solvent- CDCl<sub>3</sub>) δ: 173.8, 171.5, 169.8, 136.6, 129.5, 128.8, 127.0, 71.9, 56.5, 56.0, 54.1, 52.4, 42.8, 42.2, 41.2, 40.5, 40.2, 38.1, 36.5, 35.9, 35.5, 35.4, 34.6, 33.4, 31.6, 30.6, 28.3, 27.3, 26.5, 24.2, 23.4, 20.9, 18.4, 12.1.

Mass Spec.(EI) m/z: M. Wt. calcd for C<sub>36</sub>H<sub>54</sub>N<sub>2</sub>O<sub>5</sub>Na- 617.40; observed – 617.3936 (M+Na).

#### LC-<sup>D</sup>F-G (4)



Yield- 61%, white solid, Mp- 185-187 °C.

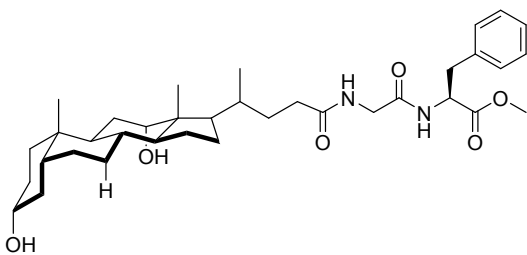
IR (KBr): 3400, 3028, 2936, 2866, 2525, 1715, 1657, 1629, 1539, 1526, 1446, 1381, 1368, 1344, 1305, 1280, 1246, 1223, 1186, 1125 cm<sup>-1</sup>.

<sup>1</sup>H NMR (400 MHz, Solvent- DMSO-d<sub>6</sub>) δ: 8.23 (s, 1H), 8.03-8.01(d, 1H, *J* = 8 Hz), 7.24-7.16 (m, 5H), 4.52 (s, 1H), 3.75-3.74 (d, 2H, *J* = 4.8 Hz), 3.03-3.01 (m, 1H), 2.74-2.68 (m, 1H), 2.1-0.92 (m, steroid protons), 0.87 (s, 3H), 0.79-0.78 (d, 3H, *J* = 8 Hz), 0.55 (s, 3H).

<sup>13</sup>C NMR (100 MHz, Solvent- DMSO-d<sub>6</sub>) δ: 172.8, 172.1, 171.5, 138.5, 129.5, 128.3, 126.5, 70.3, 56.5, 56.1, 53.9, 48.9, 42.6, 41.9, 41.3, 37.9, 36.7, 35.8, 35.5, 35.1, 34.6, 32.6, 31.7, 30.8, 28.0, 27.3, 26.5, 24.2, 23.7, 20.8, 18.6, 12.3.

Mass Spec.(EI) m/z: M. Wt. cacl. for C<sub>35</sub>H<sub>52</sub>N<sub>2</sub>O<sub>5</sub>Na, 603.38; found 603.3776 (M+Na).

#### DC-G-<sup>L</sup>F-OMe (5a)



Yield- 71%, white solid, Mp- 68-73 °C.

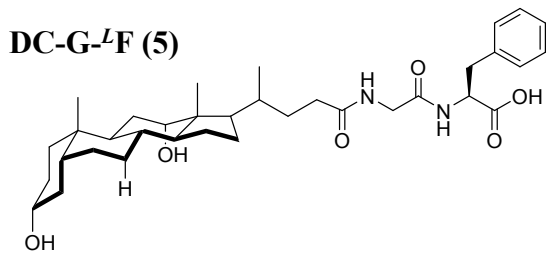
IR (KBr): 3408, 3064, 3030, 2937, 2864, 1746, 1652, 1539, 1447, 1417, 1377, 1337, 1216, 1178, 1112, 1089, 1065, 1043, 968, 943, 922, 851, 814, 755, 701 cm<sup>-1</sup>.

<sup>1</sup>H MNR (400 MHz, Solvent- CDCl<sub>3</sub>) δ: 7.29-7.24 (m, 3H), 7.11-7.09 (d, 2H, *J* = 8 Hz), 6.74-6.62 (d, 1H, *J* = 8 Hz), 6.51-6.49 (t, 1H, *J* = 4.8 Hz), 4.87-4.81 (q, 1H, *J* = 8 Hz), 3.97 (s, 1H), 3.92-3.89 (m, 1H), 3.72 (s, 3H), 3.63-3.57 (m, 1H), 3.16-3.04 (dq, 2H, *J* = 14 Hz, 5.6 Hz), 2.35-2.10 (m, 4H), 1.90-0.99 (m, steroid skeletal protons), 0.99-0.97 (d, 1H, *J* = 8 Hz), 0.91 (s, 3H, Me), 0.67 (s, 3H).

<sup>13</sup>C NMR (100 MHz, Solvent- CDCl<sub>3</sub>) δ: 174.2, 171.7, 168.8, 135.6, 129.1, 128.6, 127.2, 73.1, 71.8, 53.2, 52.4, 48.1, 46.9, 46.4, 42.9, 42.0, 37.7, 36.3, 35.9, 35.3, 35.2, 34.1, 33.5, 33.0, 31.4, 30.4, 28.6, 27.5, 27.1, 26.1, 23.7, 17.5, 12.8.

Mass Spec.(EI) m/z: M. Wt. calcd for C<sub>36</sub>H<sub>54</sub>N<sub>2</sub>O<sub>6</sub>Na - 633.3880; Observed - 633.3885 (M+Na).

#### DC-G-<sup>L</sup>F (5)



Yield- 70%, white amorphous solid.

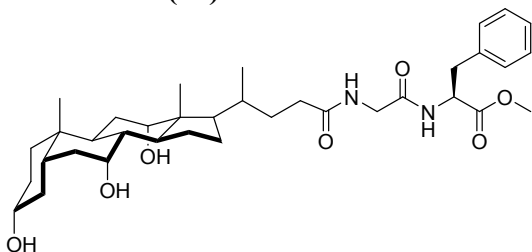
IR (KBr): 3404, 2937, 2864, 1734, 1718, 1654, 1534, 1448, 1384, 1217, 1115, 1089, 1064, 1041, 1014, 968, 944, 914, 851, 757, 700  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz, Solvent-  $\text{MeOH-d}_4$ )  $\delta$ : 7.26-7.19 (m, 5H), 4.64 (brs, 2H), 3.95 (s, 1H), 3.88-3.71 (m, 2H), 3.52-3.42 (m, 1H), 3.21-3.16 (m, 1H), 3.04-2.99 (m, 1H), 2.33-2.27 (m, 1H), 2.16-2.10 (m, 2H), 1.90-1.03 (m, steroid skeletal protons), 1.01 (d, 1H,  $J = 6$  Hz, Me), 0.93 (s, 3H, Me), 0.70 (s, 3H, Me).

$^{13}\text{C}$  MNR (100 MHz, Solvent-  $\text{MeOH-d}_4$ )  $\delta$ : 177.2, 171.3, 138.2, 130.4, 129.4, 127.7, 79.4, 74.0, 72.5, 48.0, 47.5, 43.6, 43.4, 38.5, 37.4, 37.2, 36.8, 36.4, 35.3, 34.8, 33.8, 32.9, 31.0, 29.9, 28.6, 28.4, 27.4, 24.8, 23.6, 17.7, 13.1.

Mass Spec.(EI) m/z: M. Wt. calcd for  $\text{C}_{35}\text{H}_{52}\text{N}_2\text{O}_6\text{Na} - 619.4$ ; observed- 619.3737 (M+Na).

#### C-G- $^L$ F-OMe (6a)



Yield- 61%, white solid, Mp- 84-95  $^{\circ}\text{C}$ .

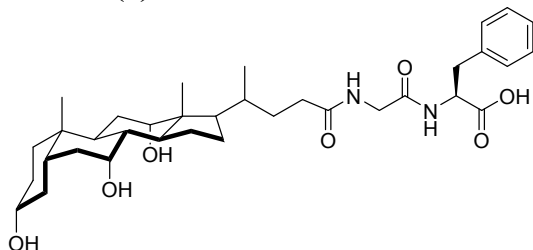
IR (KBr): 3409, 3065, 3029, 2933, 2866, 1743, 1654, 1542, 1534, 1457, 1447, 1438, 1376, 1339, 1216, 1117, 1077, 1043, 980, 950, 914, 900, 857, 812, 753, 701  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz, Solvent-  $\text{CDCl}_3$ )  $\delta$ : 7.29-7.10 (m, 5H), 4.84-4.78 (q, 1H,  $J = 8$  Hz), 3.94 (s, 1H), 3.90-3.86 (m, 2H), 3.82 (s, 1H), 3.69 (s, 3H), 3.39-3.37 (m, 1H), 3.15-2.99 (m, 2H), 2.31-2.10 (m, 4H), 1.99-0.96 (m, steroid skeletal protons), 0.98 (d, 3H,  $J = 6.4$  Hz), 0.88 (s, 3H), 0.66 (s, 3H).

$^{13}\text{C}$  NMR (100 MHz, Solvent-  $\text{CDCl}_3$ )  $\delta$ : 175.3, 171.9, 169.5, 135.9, 129.3, 128.6, 127.1, 73.2, 71.9, 68.5, 53.4, 52.5, 46.5, 46.2, 43.3, 41.6, 39.8, 39.5, 37.9, 35.5, 34.9, 34.6, 32.5, 31.5, 30.3, 28.1, 27.7, 26.2, 23.4, 22.5, 17.5, 12.4.

Mass Spec.(EI) m/z: M. Wt. calcd for  $\text{C}_{36}\text{H}_{54}\text{N}_2\text{O}_7\text{Na} - 649.38$ ; Observed - 649.3826 (M+Na).

#### C-G- $^L$ F (6)





Yield- 94%, white solid, Mp- 147-154 °C.

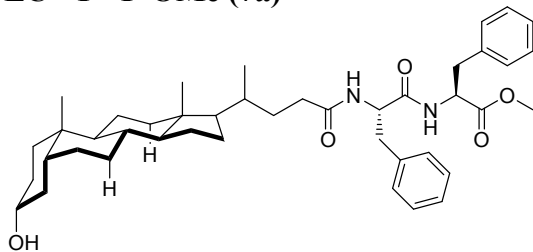
IR (KBr): 3264, 2929, 1798, 1637, 1542, 1448, 1295, 1164, 1022  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz, Solvent-  $\text{MeOH-d}_4$ )  $\delta$ : 7.29-7.19 (m, 5H), 4.69-4.65 (m, 2H), 3.95 (s, 1H), 3.88- 3.77 (m, 3H), 3.40-3.36 (m, 1H), 3.21-3.17 (dd, 1H,  $J = 16$  Hz, 5.2 Hz), 3.04-2.99 (m, 1H), 2.34-1.08 (m, steroid skeletal protons), 1.03-1.01 (d, 3H,  $J = 8$  Hz), 0.92 (s, 3H), 0.71 (s, 3H).

$^{13}\text{C}$  MNR (100 MHz, Solvent-  $\text{MeOH-d}_4$ )  $\delta$ : 175.8, 173.0, 169.9, 136.5, 128.8, 127.9, 126.3, 77.91, 72.5, 71.3, 67.6, 67.3, 53.4, 45.9, 41.8, 41.6, 41.4, 39.4, 36.8, 35.3, 34.9, 34.3, 34.2, 32.3, 31.4, 29.9, 27.1, 26.3, 22.7, 21.6, 16.2, 11.4.

Mass Spec.(EI) m/z: M. Wt. calcd for  $\text{C}_{35}\text{H}_{52}\text{N}_2\text{O}_7\text{Na}$  - 635.37; observed - 635.3672 (M+Na).

#### LC- $^1\text{F}$ - $^1\text{F}$ -OMe (7a)



Yield- 89%, white amorphous solid.

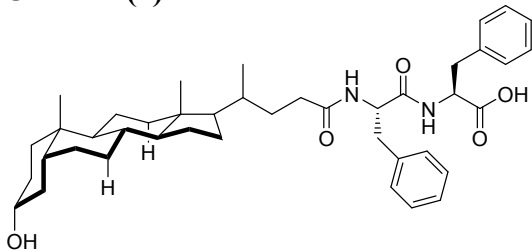
IR (KBr): 3414, 2928, 2861, 2098, 1751, 1654, 1545, 1444, 1364, 1213, 1176, 1031, 749  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz, Solvent-  $\text{CDCl}_3$ )  $\delta$ : 7.26-7.17 (m, 8H), 7.02-7.00 (d, 2H,  $J = 8$  Hz), 6.54-6.52 (d, 1H,  $J = 8$  Hz), 6.24-6.22 (d, 1H,  $J = 8$  Hz), 4.76-4.65 (m, 2H), 3.64 (s, 3H), 3.62-3.58 (m, 1H), 3.03-3.01 (m, 4H), 2.3-0.98 (m, steroid skeletal protons), 0.91-0.87 (m, 6H), 0.61 (s, 3H).

$^{13}\text{C}$  NMR (100 MHz, Solvent-  $\text{CDCl}_3$ )  $\delta$ : 173.4, 171.2, 170.5, 136.4, 135.5, 129.3, 129.1, 128.6, 128.5, 127.1, 126.9, 71.8, 56.4, 55.9, 53.9, 53.3, 52.3, 42.7, 42.0, 40.3, 40.1, 38.0, 37.8, 36.4, 35.4, 35.3, 34.5, 33.3, 31.5, 30.5, 28.2, 27.1, 26.3, 24.1, 23.3, 20.7, 18.3, 11.9.

Mass Spec.(EI) m/z: M. Wt. calcd for  $\text{C}_{43}\text{H}_{60}\text{N}_2\text{O}_5\text{Na}$  - 707.4; observed - 707.4401 (M+Na).

#### LC- $^1\text{F}$ - $^1\text{F}$ (7)



Yield- 88%, white solid, Mp- 120-130 °C.

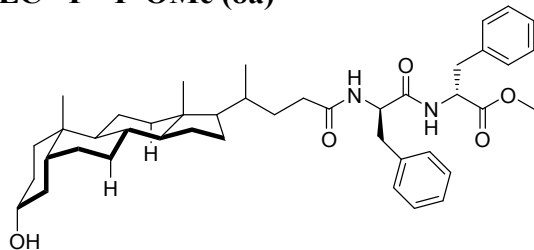
IR (KBr): 3298, 2925, 2860, 1642, 1526, 1449, 1216, 1030, 739  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz, Solvent-  $\text{MeOH-d}_4$ )  $\delta$ : 7.28-7.09 (m, 10H), 4.64-4.63 (m, 2H), 3.57-3.51 (m, 1H), 3.23-2.94 (m, 3H), 2.83-2.66 (m, 1H), 2.14-0.93 (m, steroid skeletal protons), 0.94 (s, 3H), 0.89-0.87 (d, 3H), 0.64 (s, 3H).

$^{13}\text{C}$  NMR (100 MHz, Solvent-  $\text{MeOH-d}_4$ )  $\delta$ : 175.1, 172.1, 171.8, 137.1, 136.7, 129.0, 128.8, 128.0, 127.96, 127.9, 126.4, 126.3, 126.2, 78.0, 70.9, 56.4, 55.9, 54.2, 54.1, 53.7, 42.4, 42.1, 40.4, 40.0, 37.3, 36.9, 35.75, 35.7, 35.3, 35.0, 34.2, 32.4, 31.6, 29.7, 27.7, 26.9, 26.2, 23.8, 22.5, 20.5, 17.4, 11.1.

Mass Spec.(EI) m/z: M. Wt. calcd for  $\text{C}_{42}\text{H}_{58}\text{N}_2\text{O}_5\text{Na}$  - 693.43; Observed - 693.4243 (M+Na).

#### LC- $^{2,2,2}$ F- $^{2,2,2}$ F-OMe (8a)



Yield- 74%, white amorphous solid.

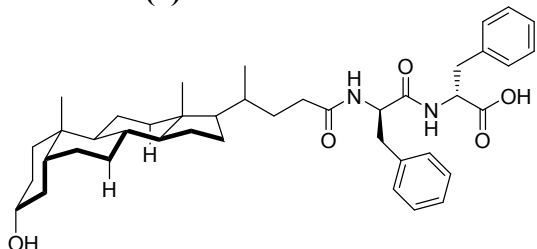
IR (KBr): 3424, 2929, 2861, 2090, 1749, 1655, 1545, 1444, 1364, 1213, 1171, 1031, 749  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.24-7.17 (m, 8 H), 7.02-7.00 (d, 2H,  $J = 8$  Hz), 6.51-6.49 (d, 1H,  $J = 8$  Hz), 6.21-6.19 (d, 1H,  $J = 8$  Hz), 4.76-4.64 (m, 2H), 3.67 (s, 3H), 3.62-3.58 (m, 1H), 3.09-2.93 (m, 4H), 2.21-0.95 (m, steroid skeletal protons), 0.91 (s, 3H), 0.87-0.85 (d, 3H), 0.62 (s, 3H).

$^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$ : 173.4, 171.2, 170.6, 136.4, 135.6, 129.3, 129.1, 128.49, 128.48, 127.0, 126.9, 71.7, 60.3, 56.4, 55.9, 53.9, 53.3, 52.2, 42.6, 42.0, 40.3, 40.1, 38.0, 37.8, 36.4, 35.7, 35.4, 35.3, 34.5, 33.3, 31.5, 30.5, 28.2, 27.1, 26.3, 24.1, 23.3, 20.9, 20.7, 18.3, 14.1, 12.0.

Mass Spec.(EI) m/z: M. Wt. calcd for  $\text{C}_{43}\text{H}_{60}\text{N}_2\text{O}_5\text{Na}$  - 707.45; observed - 707.4402 (M+Na).

#### LC- $^{2,2,2}$ F- $^{2,2,2}$ F (8)



Yield- 76%, white amorphous solid.

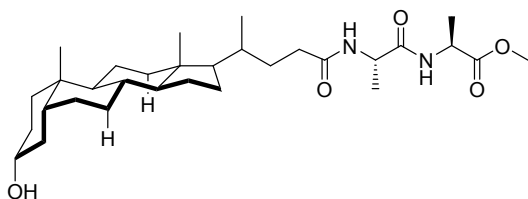
IR (KBr): 3299, 2925, 2860, 1645, 1527, 1449, 1216, 1032, 738  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz,  $\text{DMSO-d}_6$ )  $\delta$ : 8.13-8.11 (d, 1H,  $J = 8$  Hz), 7.96-7.94 (d, 1H,  $J = 8$  Hz), 7.28-7.08 (m, 10H), 4.53-4.38 (m, 2H), 3.10-3.05 (m, 1H), 2.99-2.92 (m, 2H), 2.70-2.64 (m, 1H), 2.1-0.9 (m, steroidal proton), 0.87 (s, 3H), 0.77-0.75 (d, 3H), 0.55 (s, 3H).

$^{13}\text{C}$  NMR (100 MHz,  $\text{DMSO-d}_6$ )  $\delta$ : 172.7, 172.3, 171.3, 133.0, 137.5, 129.3, 129.2, 129.1, 128.1, 128.0, 127.8, 127.7, 126.3, 126.1, 79.1, 69.8, 56.0, 55.6, 53.6, 42.2, 41.5, 37.3, 36.7, 36.3, 35.3, 34.7, 34.2, 32.1, 31.3, 30.3, 27.6, 26.8, 26.1, 23.8, 23.2, 18.1, 11.8.

Mass Spec.(EI) m/z: M. Wt. calcd for  $\text{C}_{42}\text{H}_{58}\text{N}_2\text{O}_5\text{Na}$  - 693.43; Observed - 693.4243 (M+Na).

### LC-*L*A-*L*A-OMe (9a)



Yield- 75%, white amorphous solid.

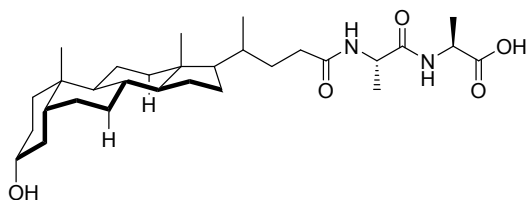
IR (KBr): 3436, 2971, 2870, 2839, 1772, 1644, 1540, 1280, 1207, 1037, 753  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz, Solvent-  $\text{CDCl}_3$ )  $\delta$ : 7.04-7.02 (d, 1H,  $J = 8$  Hz), 6.38-6.36 (d, 1H,  $J = 8$  Hz), 4.61-4.49 (m, 2H), 3.74 (s, 3H), 3.66-3.58 (m, 1H), 2.30-0.962 (m, steroidal peak and -Me group of Alanine), 0.91-0.90 (m, 6H), 0.63 (s, 3H).

$^{13}\text{C}$  NMR (100 MHz, Solvent-  $\text{CDCl}_3$ )  $\delta$ : 173.5, 173.1, 172.2, 71.8, 56.5, 56.0, 52.5, 48.5, 48.1, 42.7, 42.1, 40.4, 40.2, 36.5, 35.9, 35.5, 35.4, 34.6, 33.4, 31.7, 30.6, 29.7, 28.3, 27.2, 26.5, 24.2, 23.4, 20.9, 18.6, 18.4, 18.0, 12.1.

Mass Spec.(EI) m/z: M. Wt. calcd for  $\text{C}_{31}\text{H}_{52}\text{N}_2\text{O}_5\text{Na}$  - 555.37; Observed - 555.37 (M+Na).

### LC-*L*A-*L*A (9)



Yield- 94%, white solid, Melting point- 120-130 °C

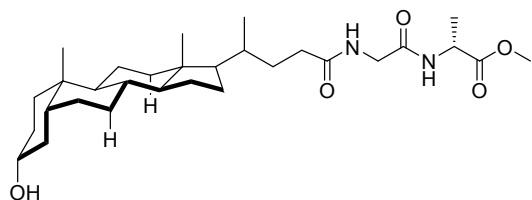
IR (KBr): 3796, 3463, 2937, 2924, 2863, 2852, 2880, 1732, 1634, 1538, 1453, 1384 cm<sup>-1</sup>.

<sup>1</sup>H MNR (400 MHz, Solvent: DMSO-d<sub>6</sub>) δ: 8.05-8.03 (d, 1H, *J* = 8 Hz), 7.93-7.91 (d, 1H, *J* = 8 Hz), 4.31-4.24 (m, 1H), 4.17-4.10 (m, 1H), 3.16 (s, 1H), 2.16-0.91 (m, steroidal peak and Me group of Alanine), 0.87-0.86 (m, 6H), 0.59 (s, 3H).

<sup>13</sup>C NMR (100 MHz, Solvent- DMSO-d<sub>6</sub>) δ: 174.4, 172.7, 172.5, 79.6, 70.3, 56.5, 55.9, 48.9, 48.0, 47.8, 42.6, 41.9, 36.7, 35.8, 35.5, 35.3, 34.6, 32.5, 31.8, 30.8, 29.4, 28.1, 27.3, 26.6, 24.2, 23.7, 20.8, 18.7, 18.5, 17.6, 12.2.

Mass Spec.(EI) m/z: M. Wt. calcd for C<sub>30</sub>H<sub>50</sub>N<sub>2</sub>O<sub>5</sub>Na - 541.36; Observed - 541.3617 (M+Na).

### LC-G-<sup>D</sup>A-OMe (10a)



Yield- 66%, white solid, Mp- 170-173 °C.

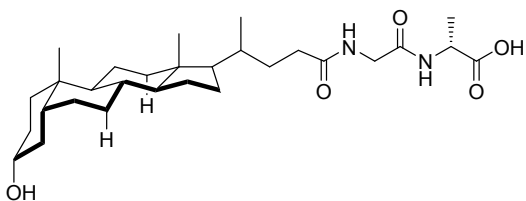
IR (KBr): 3796, 3463, 2937, 2924, 2863, 2852, 2880, 1732, 1634, 1538, 1453, 1384 cm<sup>-1</sup>.

<sup>1</sup>H MNR (400 MHz, Solvent- CDCl<sub>3</sub>) δ: 6.67-6.65 (d, 1H, *J* = 8 Hz, NH), 6.28 (s, 1H), 4.61-4.54 (m, 1H), 4.02-3.91 (m, 2H), 3.76 (s, 3H), 3.66-3.57 (m, 1H), 2.31-0.96 (m, ), 0.93-0.91 (m, 6H), 0.64 (s, 3H).

<sup>13</sup>C NMR (100 MHz, Solvent: CDCl<sub>3</sub>) δ: 174.2, 173.0, 168.6, 71.8, 56.4, 55.9, 52.5, 48.1, 43.0, 42.7, 42.0, 40.4, 40.1, 36.4, 35.8, 35.4, 35.3, 34.5, 33.2, 31.6, 30.5, 28.2, 27.1, 26.3, 24.1, 23.3, 20.7, 18.3, 18.1, 11.9.

Mass Spec.(EI) m/z: M. Wt. calcd for C<sub>30</sub>H<sub>50</sub>N<sub>2</sub>O<sub>5</sub>Na - 541.36; Observed - 541.3618 (M+Na).

### LC-G-<sup>D</sup>A (10)



Yield- 84%, white solid, Mp- 110-130 °C.

IR (KBr): 3796, 3463, 2937, 2924, 2863, 2852, 2880, 1732, 1634, 1538, 1453, 1384  $\text{cm}^{-1}$ .

$^1\text{H}$  MNR (400 MHz, Solvent-  $\text{MeOH-d}_4$ )  $\delta$ : 4.45-4.39 (m, 1H), 3.94-3.82 (m, 2H), 3.57-3.51 (m, 1H), 2.39-0.99 (m, steroidal peak and Me group of Alanine), 0.99-0.96 (m, 6H), 0.70 (s, 3H, Me).

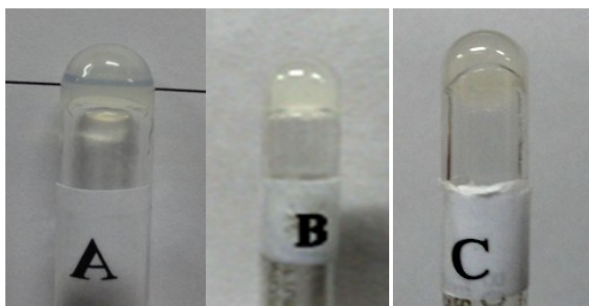
$^{13}\text{C}$  NMR (100 MHz, Solvent-  $\text{MeOH-d}_4$ )  $\delta$ : 175.7, 174.3, 169.8, 78.0, 70.9, 56.4, 55.9, 42.4, 42.07, 40.06, 35.8, 35.7, 35.4, 35.0, 34.2, 32.4, 31.5, 29.7, 27.8, 26.8, 26.2, 23.8, 22.5, 20.5, 17.4, 16.3, 11.0.

Mass Spec.(EI) m/z: M. Wt. calcd for  $\text{C}_{29}\text{H}_{48}\text{N}_2\text{O}_5\text{Na}$  - 527.72; observed - 527.3461 (M+Na).

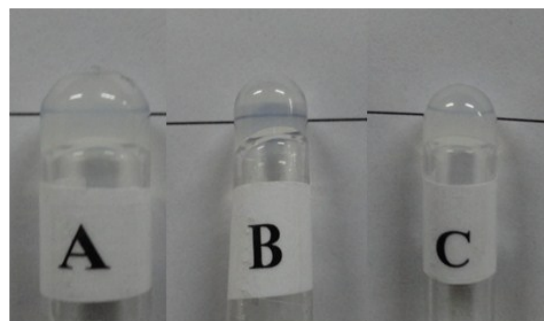
## General procedures for gelation studies

Gelation tests for compounds **1-10** were done by weighing 3 mg of a particular compound in a test tube and adding 300  $\mu\text{L}$  of the solvent in query. For checking gelation in presence of NaOH, 3 mg compounds were taken, 100  $\mu\text{L}$  0.5 N NaOH and followed by 200  $\mu\text{L}$  distilled water was added. The obtained suspension was heated until it form a clear solution. The solution was allowed to reach room temperature or sonicated for 1 minute, after that gelations were checked by inverted test tube method.

### Gel images of (1 wt%) $\text{LC-}^L\text{F-G}$ (3) and $\text{LC-}^D\text{F-G}$ (4)



**$\text{LC-}^L\text{F-G}$  (3)**

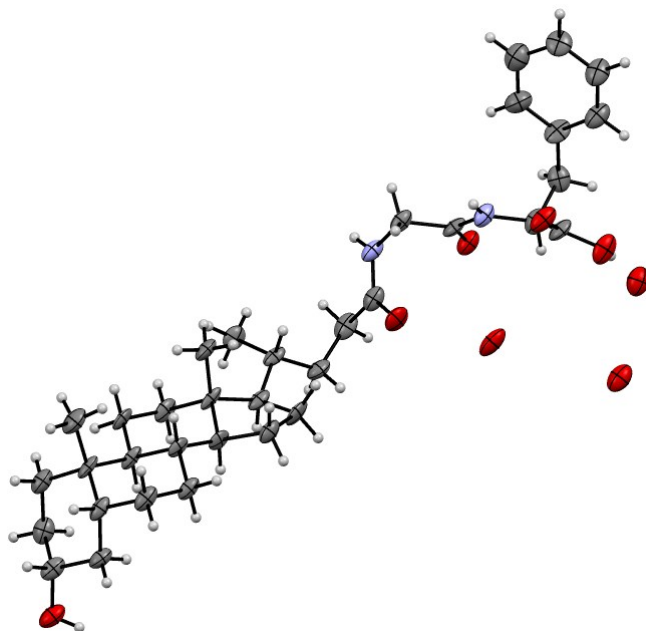


**$\text{LC-}^D\text{F-G}$  (4)**

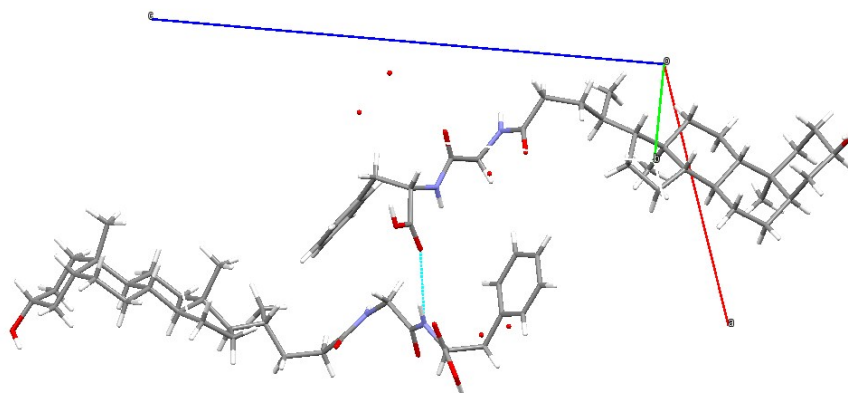
A= in NaOH, B= pH 8, C= pH 7

## Single crystal X-ray analysis

Suitable crystal of **LC-G-F (1)** for single crystal analysis was grown from MeOH/ H<sub>2</sub>O by slow evaporations.



**Figure S1:** Single Crystal structure of **LC-G-F (1)**. 3 (O)



**Figure S2:** Packing structure of **LC-G-F (1)**. 3 (O)

**Table S1: Crystal data and structure refinement for mono.**

Empirical formula	C <sub>35</sub> H <sub>52</sub> N <sub>2</sub> O <sub>8</sub> [C <sub>35</sub> H <sub>52</sub> N <sub>2</sub> O <sub>5</sub> + 3 (O)]
Formula weight	628.78
Temperature/K	100
Crystal system	monoclinic
Space group	P2 <sub>1</sub>
a/Å	12.346(5)
b/Å	7.136(3)
c/Å	20.348(7)
α/°	90
β/°	105.843(17)
γ/°	90
Volume/Å <sup>3</sup>	1724.7(11)
Z	2
ρ <sub>calc</sub> /cm <sup>3</sup>	1.211
μ/mm <sup>-1</sup>	0.085
F(000)	680.0
Crystal size/mm <sup>3</sup>	0.3 × 0.2 × 0.2
Radiation	MoKα (λ = 0.71073)
2θ range for data collection/°	3.43 to 51.998
Index ranges	-15 ≤ h ≤ 15, -8 ≤ k ≤ 5, -25 ≤ l ≤ 24
Reflections collected	23601
Independent reflections	5181 [R <sub>int</sub> = 0.1811, R <sub>sigma</sub> = 0.1554]
Data/restraints/parameters	5181/1/411
Goodness-of-fit on F <sup>2</sup>	0.993
Final R indexes [I >= 2σ (I)]	R <sub>1</sub> = 0.0895, wR <sub>2</sub> = 0.2140
Final R indexes [all data]	R <sub>1</sub> = 0.1653, wR <sub>2</sub> = 0.2456
Largest diff. peak/hole / e Å <sup>-3</sup>	0.43/-0.38
Flack parameter	1.1(10)

### Experimental

Single crystals of C<sub>35</sub>H<sub>52</sub>N<sub>2</sub>O<sub>8</sub> were LC-G-LF (**1**), 3(O). A suitable crystal was selected and mounted on a 'Bruker APEX-II CCD' diffractometer. The crystal was kept at 100 K during data collection. Using Olex2 [1], the structure was solved with the olex2.solve [2] structure solution program using Charge Flipping and refined with the ShelXL [3] refinement package using Least Squares minimisation.

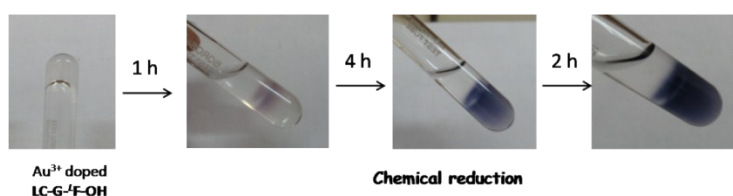
1. Dolomanov, O.V., Bourhis, L.J., Gildea, R.J., Howard, J.A.K. & Puschmann, H. (2009), *J. Appl. Cryst.* 42, 339-341.
2. Bourhis, L.J., Dolomanov, O.V., Gildea, R.J., Howard, J.A.K., Puschmann, H. (2013). in preparation.
3. Sheldrick, G.M. (2008). *Acta Cryst.* A64, 112-122.

## Crystal structure determination of [mono]

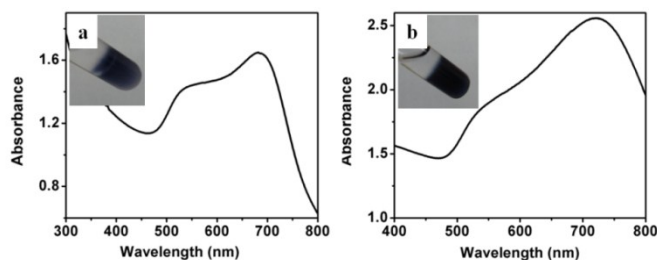
**Crystal Data** for  $C_{35}H_{52}N_2O_8$  ( $M=628.78$  g/mol): monoclinic, space group  $P2_1$  (no. 4),  $a = 12.346(5)$  Å,  $b = 7.136(3)$  Å,  $c = 20.348(7)$  Å,  $\beta = 105.843(17)^\circ$ ,  $V = 1724.7(11)$  Å<sup>3</sup>,  $Z = 2$ ,  $T = 100$  K,  $\mu(\text{MoK}\alpha) = 0.085$  mm<sup>-1</sup>,  $D_{\text{calc}} = 1.211$  g/cm<sup>3</sup>, 23601 reflections measured ( $3.43^\circ \leq 2\Theta \leq 51.998^\circ$ ), 5181 unique ( $R_{\text{int}} = 0.1811$ ,  $R_{\text{sigma}} = 0.1554$ ) which were used in all calculations. The final  $R_1$  was 0.0895 ( $I > 2\sigma(I)$ ) and  $wR_2$  was 0.2456 (all data).

## AgNPs and AuNPs synthesis in LC-G-*D/L*F gel

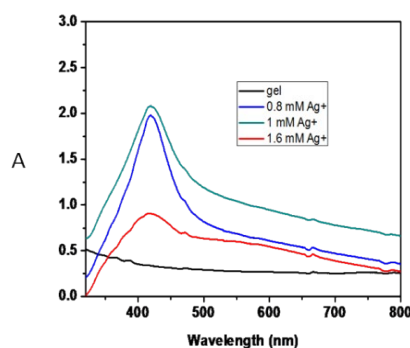
It was found that increasing the concentration of the dopant ( $> 1$  mM in 4.3 mM gel, 0.5 wt%) led to the agglomeration of small AuNPs/AgNPs to bigger clusters. Therefore, reduced doping (0.4 mM) of Au or Ag precursor in the hydrogel template was necessary for the smooth formation of silver/gold nanoparticles inside the gel fibres.



**Figure S3:** AuNPs synthesis in the hydrogel of **1** by chemical reduction of  $\text{Au}^{3+}$  (1.6 mM in 0.5 wt% gel).



**Figure S4:** Absorption spectra of AuNPs formed in the hydrogels of (a) **1** and (b) **2** by chemical reduction of  $\text{Au}^{3+}$  (1.6 mM in 0.5 wt% gel).

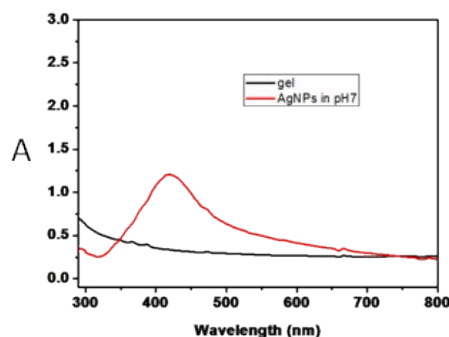


**Figure S5:** Absorption spectra of AgNPs formed in the hydrogel of **1** at different concentrations of Ag(I).



Nanoparticles formation were checked in pH 8 phosphate buffer gel for comparison as both gelator 1 and 2 formed gels in pH 8, whereas gelator 2 did not form gel in pH 7.

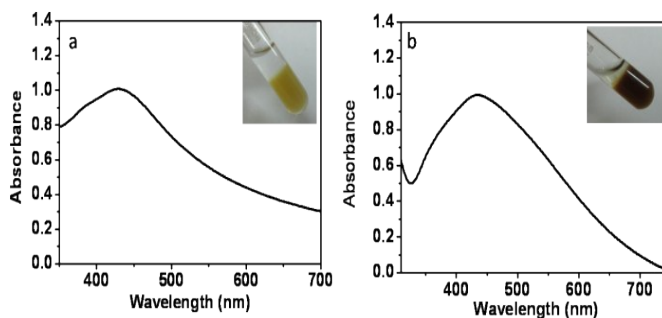
We have checked effect of pH in case of LC-G-*L*F (1). We have observed yellow AgNP formation ( $\lambda_{\max}$  420 nm) and pink color AuNP formation ( $\lambda_{\max}$  515 nm) using chemical reduction method in pH 7 which showed similar spectra as in pH 8. Also, at higher concentration of NaOH (pH 11), Ag<sup>+</sup> salt produced precipitate of AgOH or Ag<sub>2</sub>O, so we explored nanoparticle formation in pH 8.



**Figure S6:** Absorption spectra of AgNPs formed in the hydrogel of **1** by chemical reduction of Ag<sup>+</sup> (0.4 mM in 0.5 wt% gel).

### Preparation of AgNPs by chemical reduction and photoreduction in LC-*D/L*F-G gel

Silver nanoparticles were prepared in LC-*L*F-G (**3**) and LC-*D*F-G (**4**) hydrogel by the same procedure as followed for **1** and **2** (main text). For **3** a yellow colour and for **4** a brown colour appeared at the interface after 30 min. and after 12 h the entire gel turned yellow and brown, respectively (**Fig. S7**). The gels derived from compounds **3** and **4** showed absorbance peaks around 427 and 435 nm, respectively.

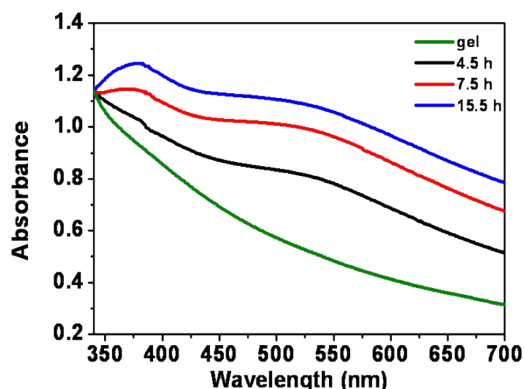


**Figure S7:** Absorption spectra of the AgNPs formed in the hydrogels of (a) **3** and (b) **4**

We have also examined photoreduction of Ag(I) in LC-*D*F-G (**3**) and LC-*L*F-G (**4**) gel by irradiating the gels by light from a 500 W tungsten lamp (Illuminance of the 500 watt tungsten bulb at a distance of 12 cm on a surface of 0.6 cm radius was 0.3 W measured using power meter). Silver doped gel **3** and **4** were turbid in nature. In case of LC-*D*F-G (**3**) gel color changes from pink to brown.

Absorptions were recorded at regular time intervals and the data show the changes in the absorption spectrum. The pink gel showed peaks around 530 nm and with time 380 nm peaks appeared (**Fig. S8**).

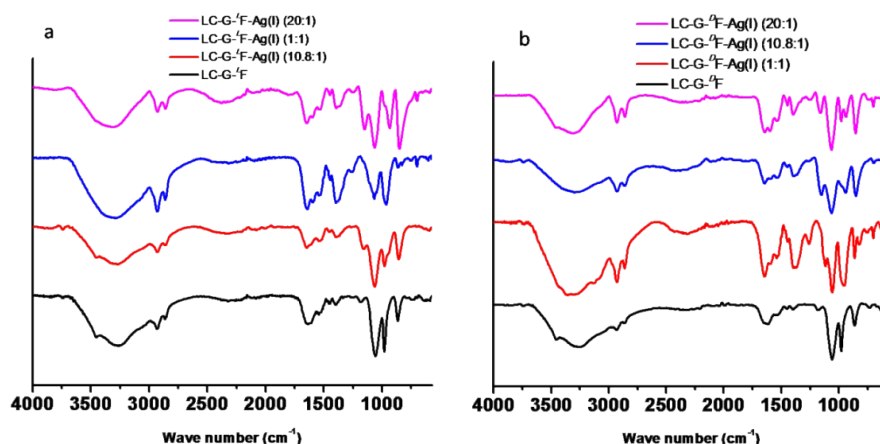
Silver doped LC-<sup>L</sup>F-G (4) gel was very turbid and after photo irradiation for 15 h, gel become little blackish color but in UV-Visible absorbance spectra there was no peak corresponding to Ag(0). Similar to LC-G-<sup>L</sup>F (2) gel, dark brown colored appeared after addition of NaBH<sub>3</sub>CN to LC-<sup>L</sup>F-G (4) gel.



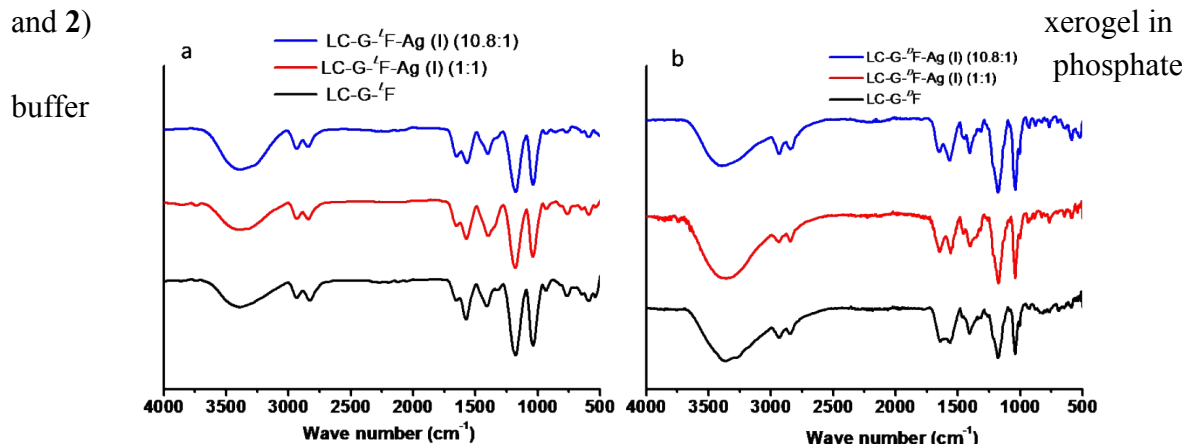
**Figure S8:** Absorption spectra as a function of irradiation time for AgNPs synthesized in LC-<sup>D</sup>F-G (4) gel

### FT-IR studies of LC-G-<sup>D/L</sup>F xerogel and Ag(I) containing LC-G-<sup>D/L</sup>F xerogel

FT-IR were recorded on Bruker putting solid xerogel directly to the sample holder. To verify the structural insight of Ag(I) doped LC-G-<sup>D/L</sup>F self assembled materials, FT-IR was used to study in xerogel states. In LC-G-<sup>D</sup>F (2) xerogel show a strong C=O stretching peak at 1632 cm<sup>-1</sup> and corresponding N-H bending and N-H stretching frequencies appearing at 1532 and 3456, 3283 cm<sup>-1</sup> (**Table S2, Fig. S9**). Interestingly, in the Ag(I) containing (gelator/Ag<sup>+</sup> ratio= 10.8) xerogel LC-G-<sup>D</sup>F (2) of C=O stretching frequency at 1632 cm<sup>-1</sup> shifted to 1652 cm<sup>-1</sup>, whereas Ag(I) containing xerogel LC-G-<sup>L</sup>F (1) no shift were observed. Infrared spectra of the xerogels indicated that Ag(I) coordinate strongly with the carbonyl group of the LC-G-<sup>D</sup>F (2) than LC-G-<sup>L</sup>F (1). Similar observation were made in different ratio of Ag(I) containing xerogel (**Table S3, Fig. S10**) also xerogel prepared in HEPES buffer (100 mM, pH 8).



**Figure S9:** FT-IR spectra of LC-G-<sup>L/D</sup>F (1 and 2) xerogel and silver(I) containing LC-G-<sup>L/D</sup>F (1 and 2)



**Figure S10:** FT-IR spectra of LC- G-<sup>L/D</sup>F (1 and 2) xerogel and silver(I) containing LC- G-<sup>L/D</sup>F (1 and 2) xerogel in HEPES buffer

**Table S2:** FT-IR of LC-G-<sup>D/L</sup>F xerogel and Ag(I) containing LC-G-<sup>D/L</sup>F xerogel in Phosphate buffer (pH 8)

Samples	C=O stretching frequency (cm <sup>-1</sup> )	N-H bending frequency (cm <sup>-1</sup> )	C(O)-N stretching frequency (cm <sup>-1</sup> )	N-H stretching frequency (cm <sup>-1</sup> )
LC-G- <sup>L</sup> F (1)	1642	1526	1396	3453, 3284
LC-G- <sup>L</sup> F- Ag(I) (17.3:17.3 mM, gelator/Ag(I) ratio= 1)	1643	1595, 1536	1388	3334
LC-G- <sup>L</sup> F- Ag(I) [17.3:1.6 mM, ratio= 10.8]	1648	1525	1391	3454, 3302
LC-G- <sup>L</sup> F- Ag(I) (17.3:0.8 mM, ratio= 20.1)	1646	1599, 1534	1389	3325
LC-G- <sup>D</sup> F (2)	1632	1532	1392	3456, 3283
LC-G- <sup>D</sup> F- Ag(I) (17.3:17.3 mM, ratio= 1)	1647	1537	1397	3344
LC-G- <sup>D</sup> F- Ag(I) (17.3:1.6 mM, ratio= 10.8)	1652	1593, 1526	1397	3332

LC-G- <sup>D/L</sup> F- Ag(I) (17.3:0.8 mM, ratio= 20.1)	1645	1599, 1537	1397	3314
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**Table S3:** FT-IR of LC-G-<sup>D/L</sup>F xerogel and Ag(I) containing LC-G-<sup>D/L</sup>F xerogel in HEPES buffer (pH 8)

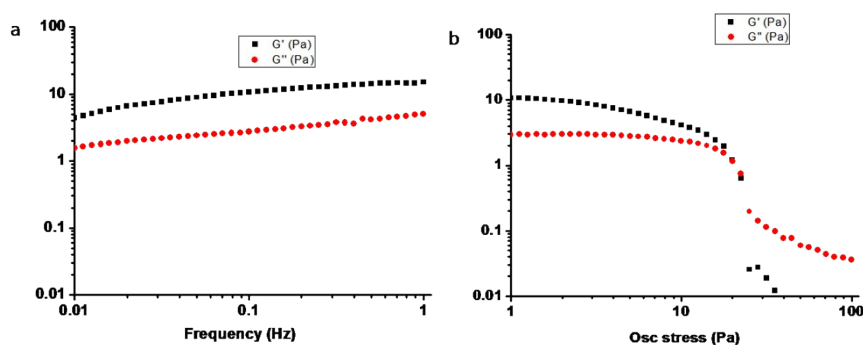
Samples	C=O stretching frequency (cm <sup>-1</sup> )	N-H bending frequency (cm <sup>-1</sup> )	C(O)-N stretching frequency (cm <sup>-1</sup> )	N-H stretching frequency (cm <sup>-1</sup> )
LC-G- <sup>L</sup> F (1)	1650	1577	1314	3376
LC-G- <sup>L</sup> F- Ag(I) (17.3:17.3 mM, gelator/Ag(I) ratio= 1)	1650	1574	1327	3362
LC-G- <sup>L</sup> F- Ag(I) [17.3:1.6 mM, gelator/Ag(I)= 10.8]	1655	1564	1314	3395
LC-G- <sup>D</sup> F (2)	1639	1565	1319	3367
LC-G- <sup>D</sup> F- Ag(I) (17.3:17.3 mM, ratio= 1)	1645	1558	1318	3368
LC-G- <sup>D</sup> F- Ag(I) (17.3:1.6 mM, ratio= 10.8)	1649	1568	1317	3395

Note- Doping Ag(I) in LC-G-<sup>D/L</sup>F gel in ratio of 1:1 in both buffer, some precipitate were formed which did not dissolved by heating, resulted turbid gel formation. This turbid gel was lyophilized and xerogel was used for FT-IR study.

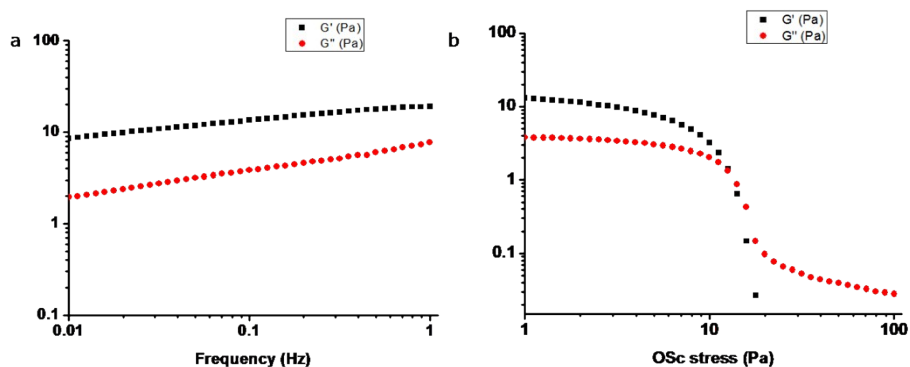
Another interesting observation was in HEPES buffer, both the derivatives got reduced by photoreduction probably HEPES facilitated to reduce Ag (I).

### Rheology studies of LC-G-<sup>D/L</sup>F gel in pH 8

Rheological measurements at pH 8 showed gel behaviour as G' (Storage modulus) was always higher than G'' (loss modulus). Rheological experiments indicated that both <sup>L</sup>F and <sup>D</sup>F derivate exhibited "soft" gel like behaviour.



**Figure S11:** Rheology of LC-G-<sup>L</sup>F (1, 2 wt%) (a) Frequency sweep (at 0.5 Pa) and (b) Stress sweep (at 0.1 Hz)



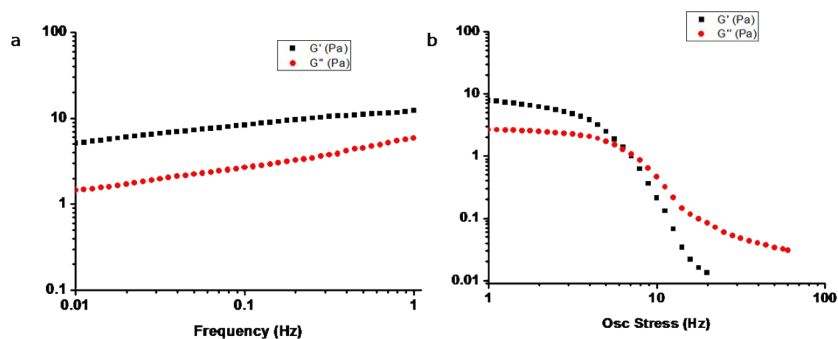
**Figure S12:** Rheology of LC-G-<sup>D</sup>F (2, 2 wt%) (a) Frequency sweep (at 0.5 Pa) and (b) Stress sweep (at 0.1 Hz)

**Table S4:** Rheology data of LC-G-<sup>L</sup>F (1) and LC-G-<sup>D</sup>F (2)

Sample	G' (Pa)	G'' (Pa)	G'/G''	$\sigma^*$ (Pa)
LC-G- <sup>L</sup> F (1)	10	2.7	3.7	22
LC-G- <sup>D</sup> F (2)	13.5	3.8	3.5	14

## Rheology studies of LC-G-<sup>D</sup>F gel in pH 8 with Ag(I) before and after photoreduction

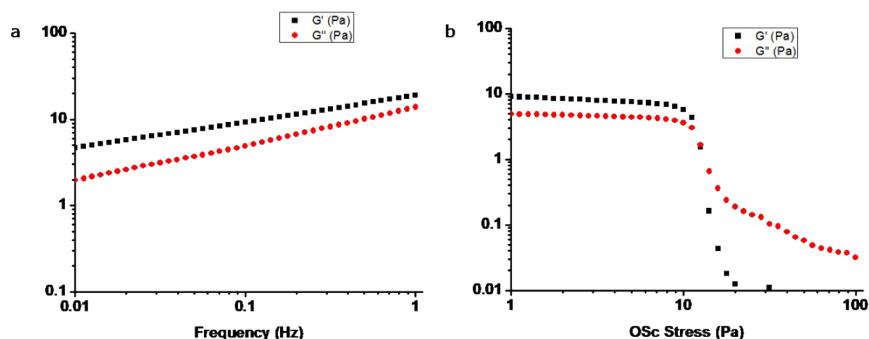
Rheological studies were done for LC-G-<sup>D</sup>F with Ag(I) doping before and after the illumination with 2 wt% of gelator at pH 8 (100 mM phosphate buffer) and 0.4 mM of Ag(I) doping. The viscoelastic behavior typical of a molecular gel was with storage modulus ( $G'$ ) values always greater than loss modulus ( $G''$ ) ones. The frequency sweep experiment and stress sweep experiments revealed that there was only a marginal increase in the dynamic storage modulus ( $G'$ ) and the loss modulus ( $G''$ ) after illumination (Table S7).



**Figure S13:** Rheology of LC-G-*D*F (2, 2 wt%) with 0.4 mM Ag (I) before illumination (a) Frequency sweep (at 0.5 Pa) and (b) Stress sweep (at 0.1 Hz)

**Table S5:** Rheology data of LC-G-*D*F with Ag(I)

Sample	G' (Pa)	G'' (Pa)	G'/G''	$\sigma^*$ (Pa)
LC-G- <i>D</i> F with Ag(I)	8.3	2.6	3.2	7



**Figure S14:** Rheology of LC-G-*D*F (2, 2 wt%) with 0.4 mM Ag(I) after illumination for 6h (a) Frequency sweep (at 0.5 Pa) and (b) Stress sweep (at 0.1 Hz)

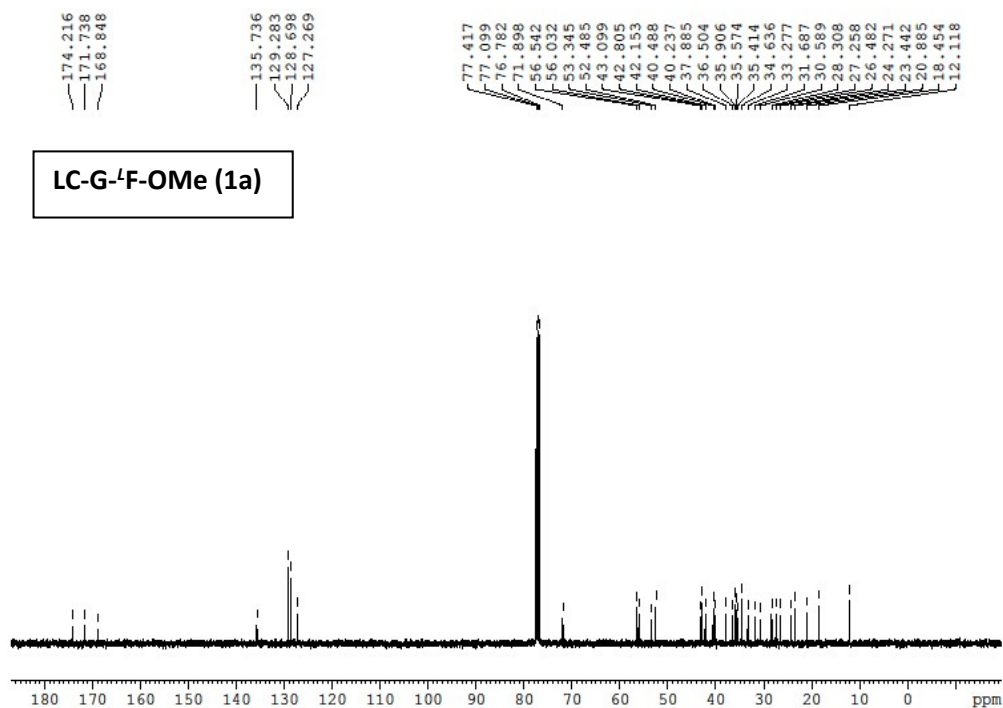
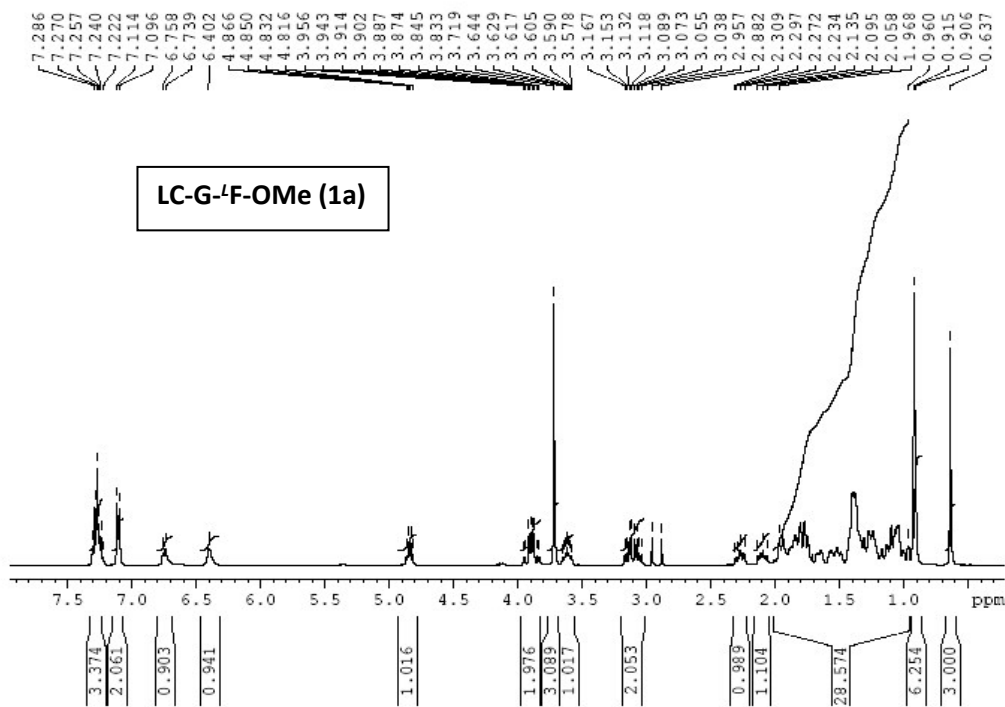
**Table S6:** Rheology data of LC-G-*D*F with AgNPs after illumination

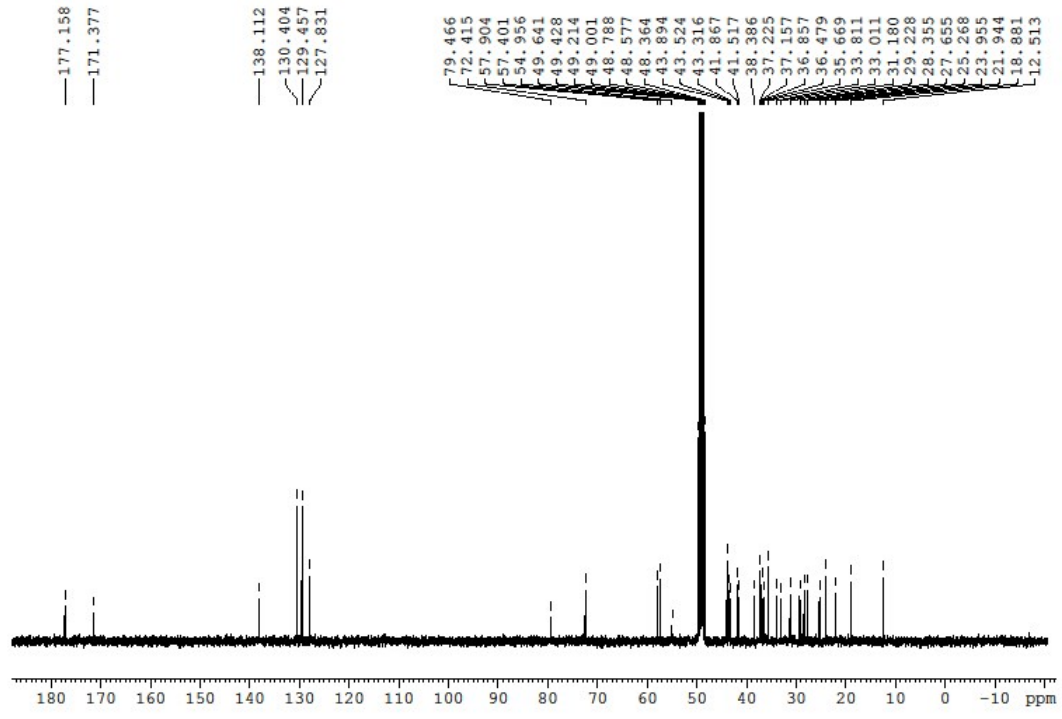
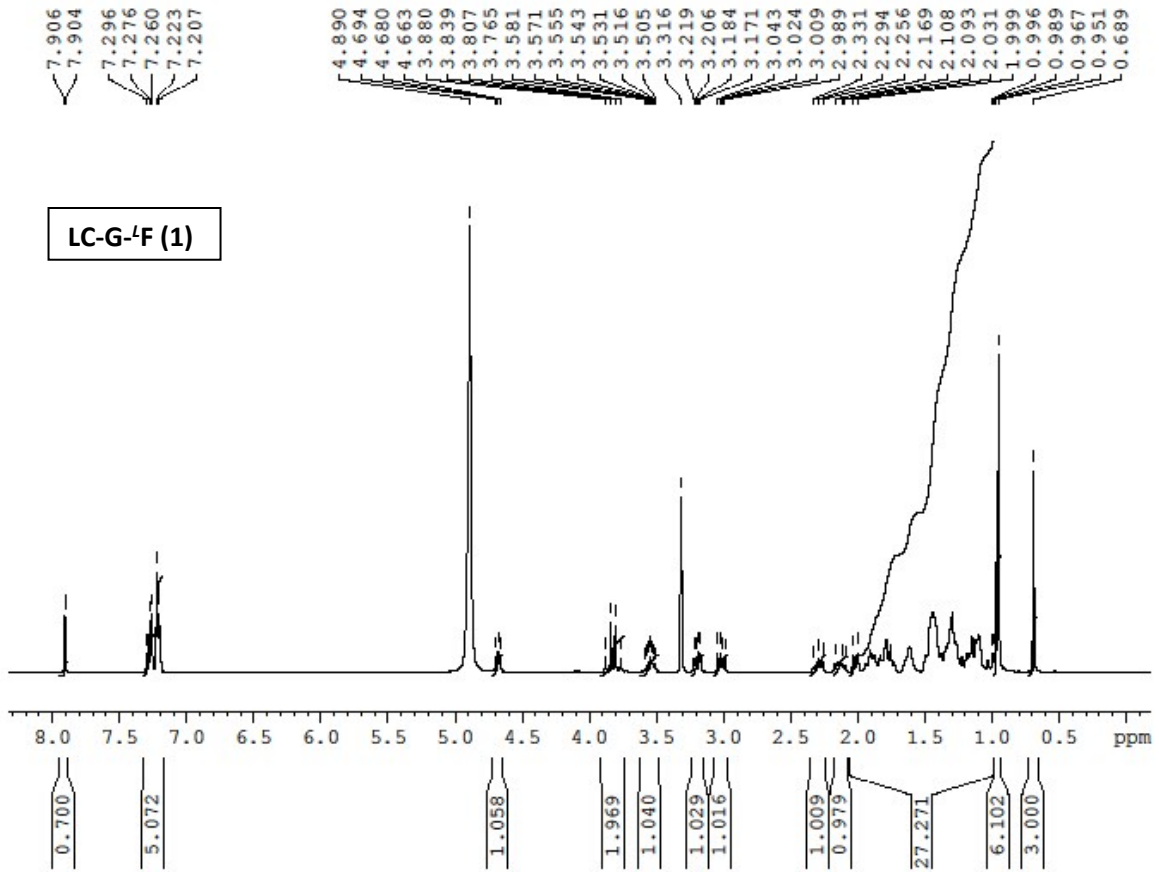
Sample	G' (Pa)	G'' (Pa)	G'/G''	$\sigma^*$ (Pa)
LC-G- <i>D</i> F with AgNPs	9.2	4.9	1.8	12.6

**Table S7:** Comparison table

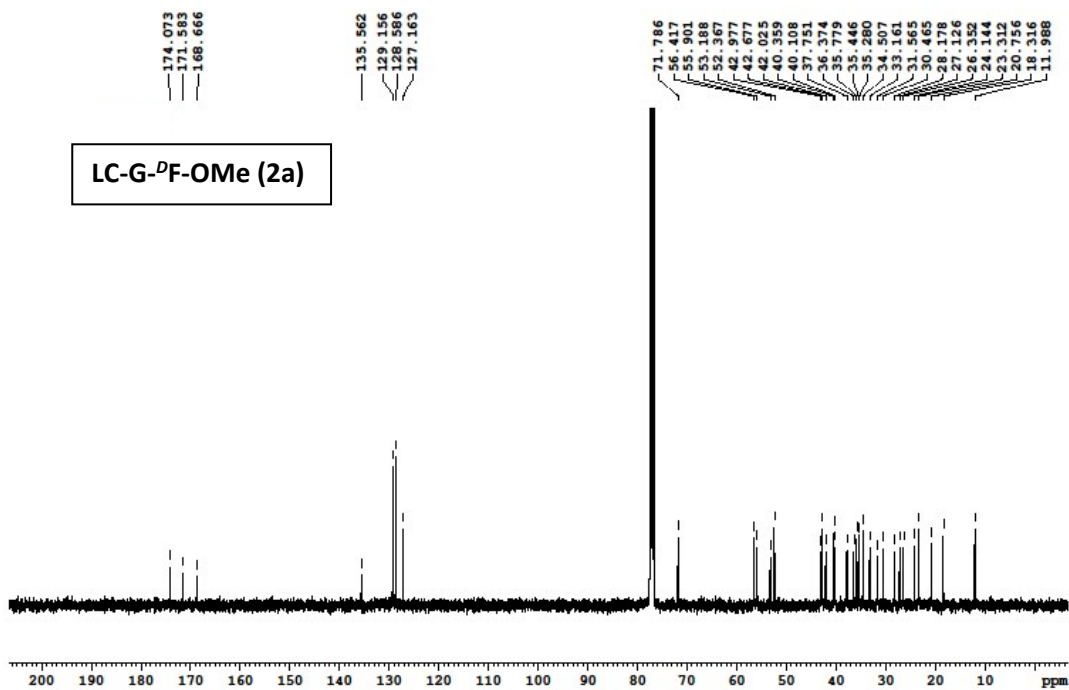
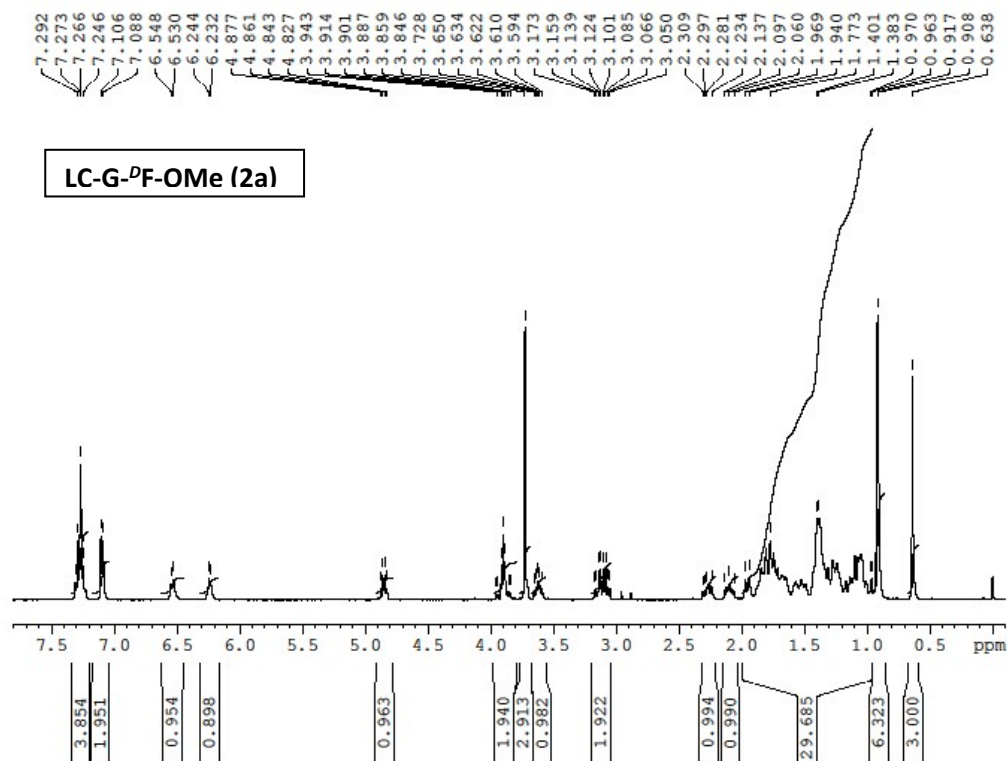
Sample	G' (Pa)	G'' (Pa)	G'/G''	$\sigma^*$ (Pa)
LC-G- <i>D</i> F (2) [Table S4]	13.5	3.8	3.5	14
LC-G- <i>D</i> F (2) with 0.4 mM Ag(I)	8.3	2.6	3.2	7
LC-G- <i>D</i> F (2) with AgNPs after illumination	9.2	4.9	1.8	12.6

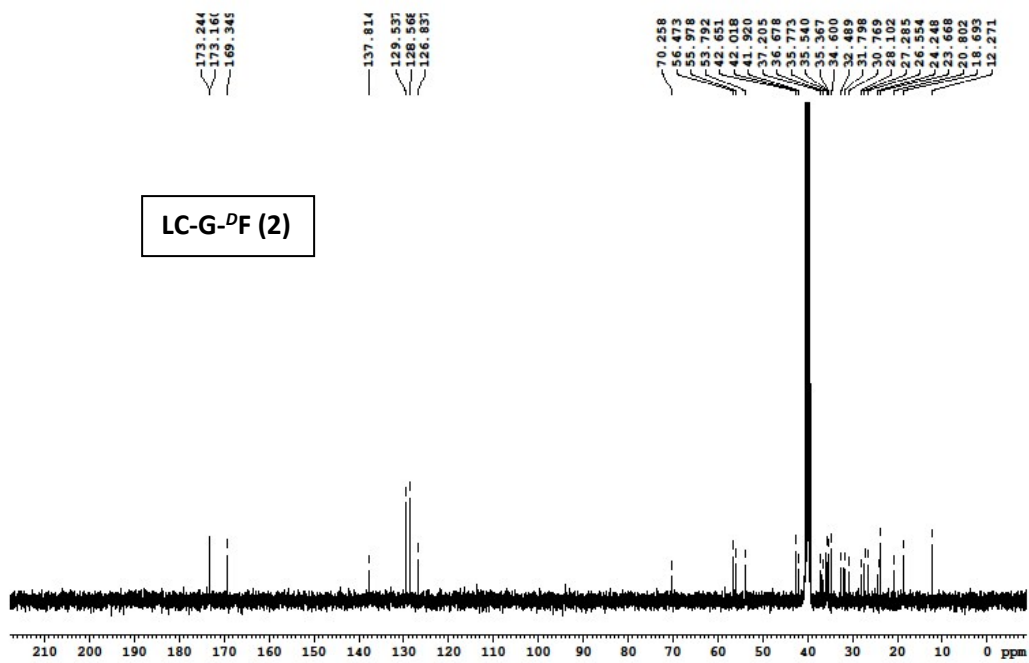
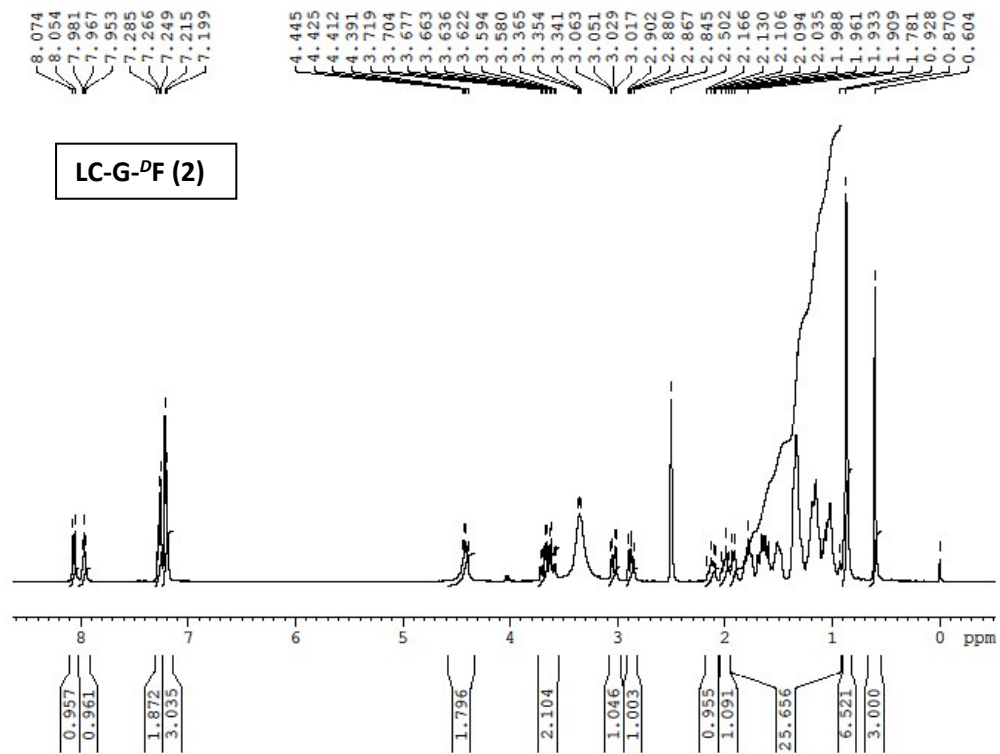
Copies of  $^1\text{H}$ ,  $^{13}\text{C}$  NMR spectra

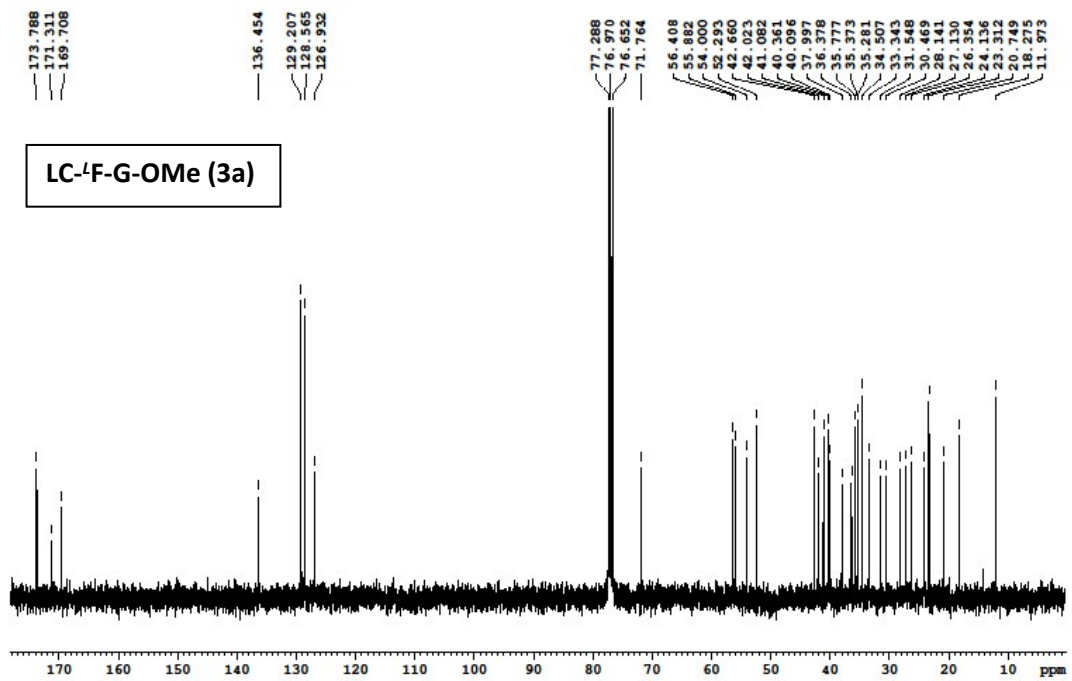
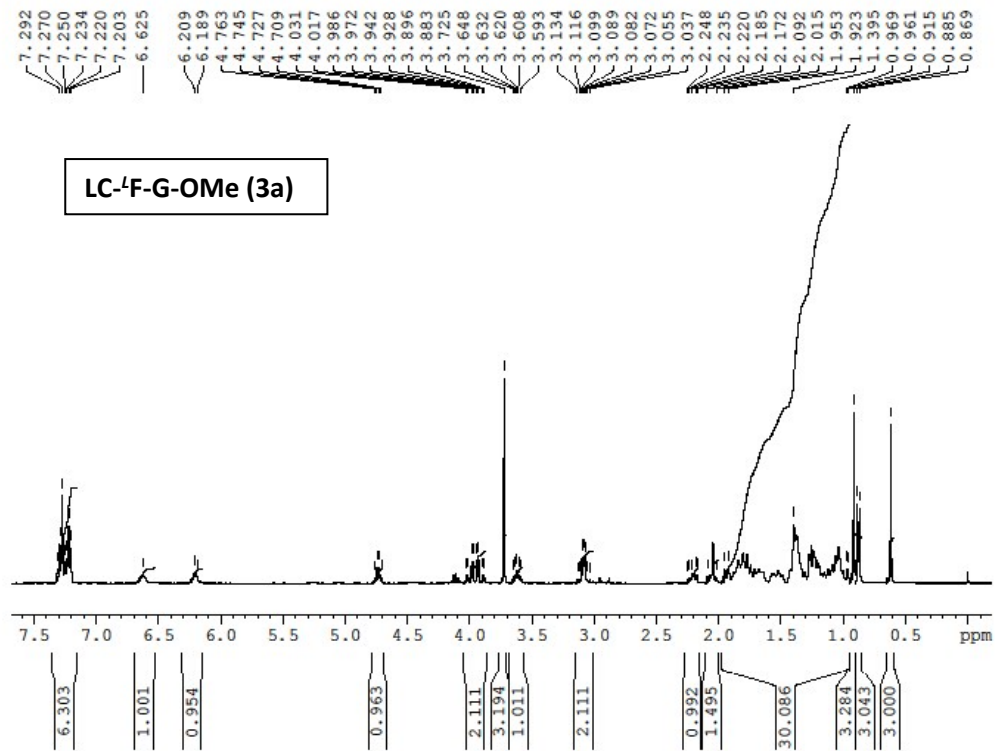


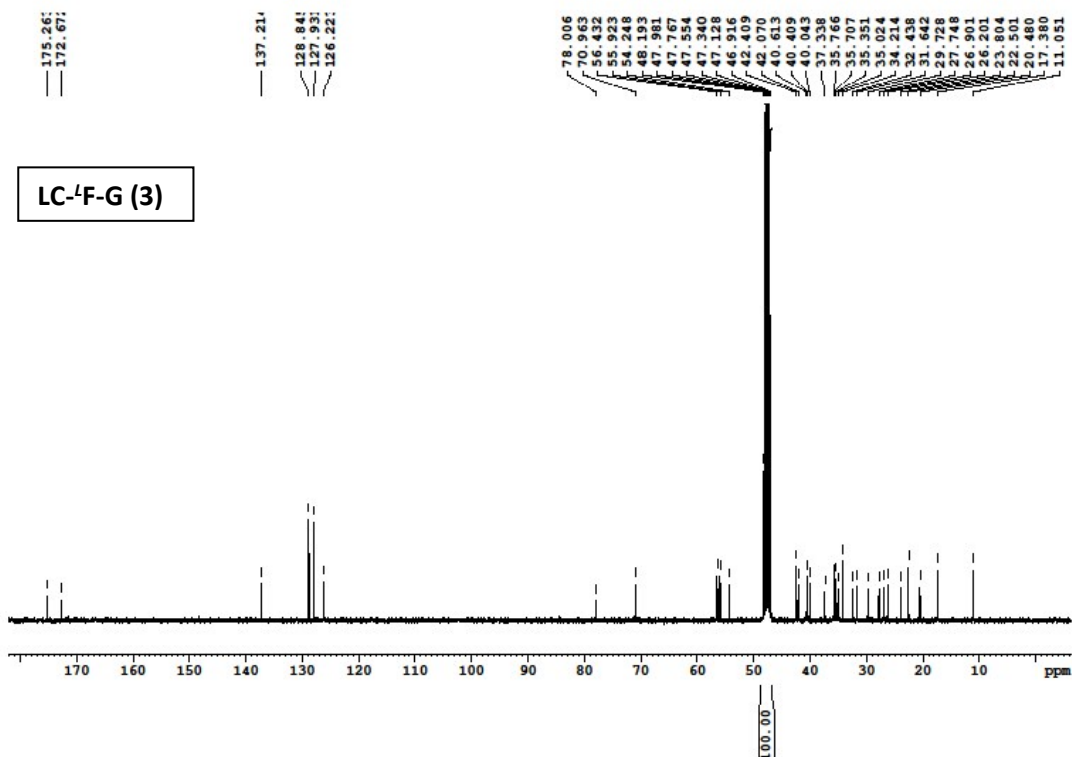
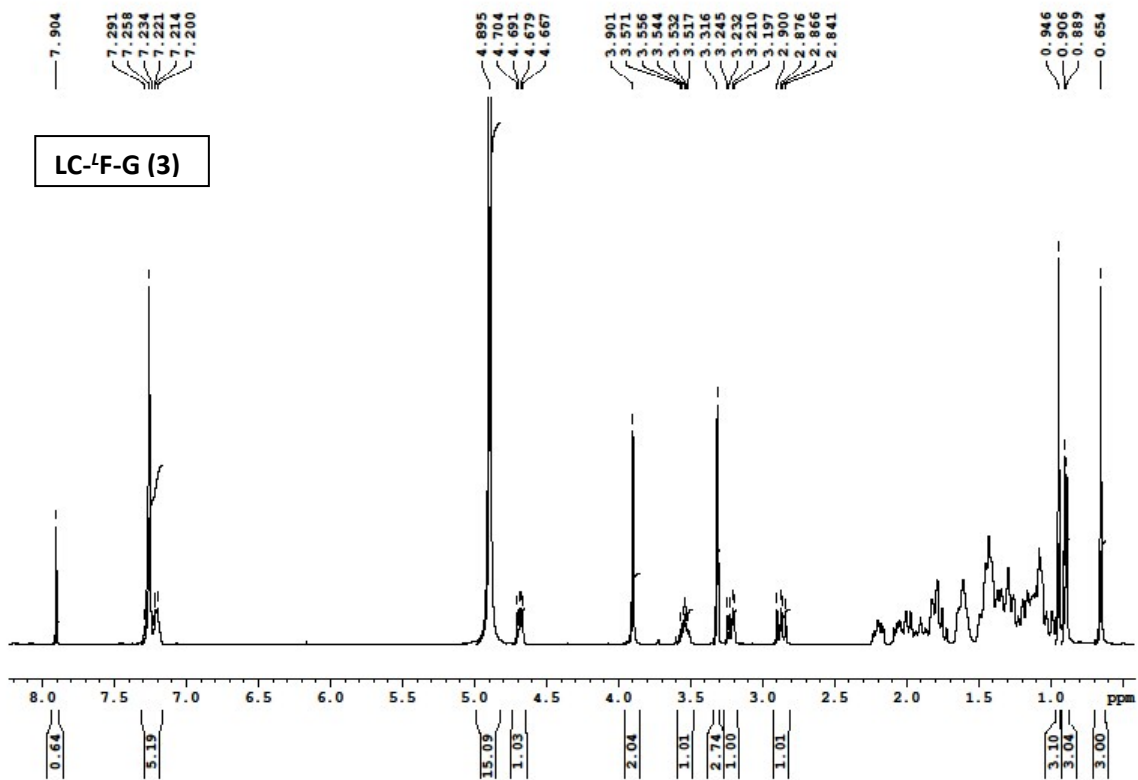


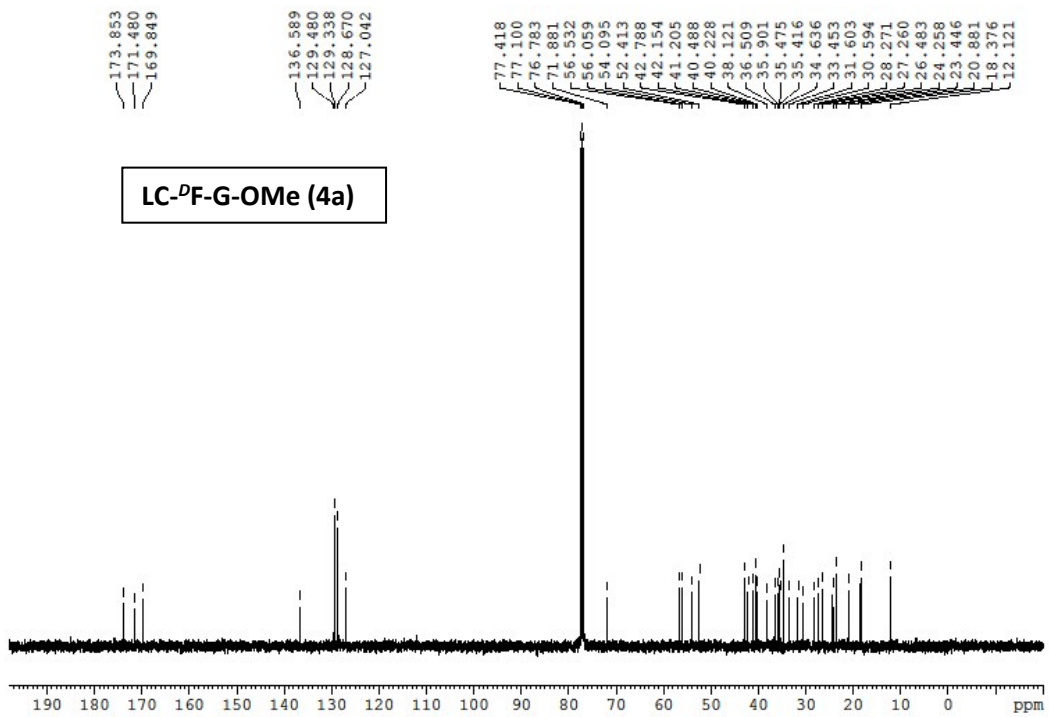
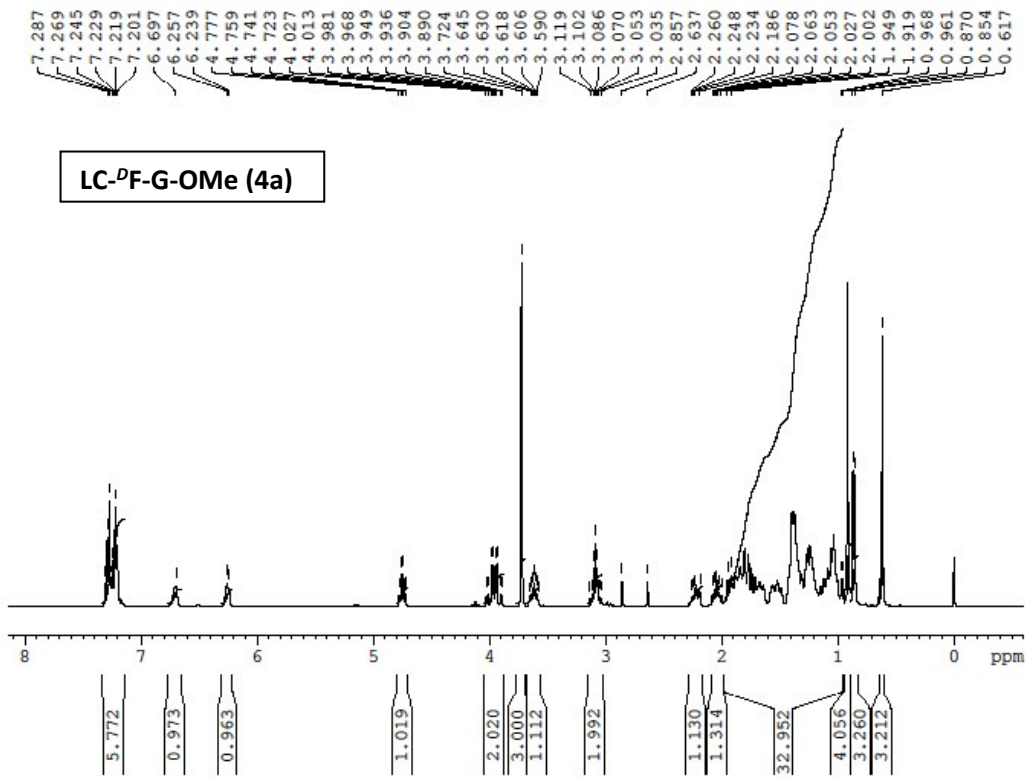


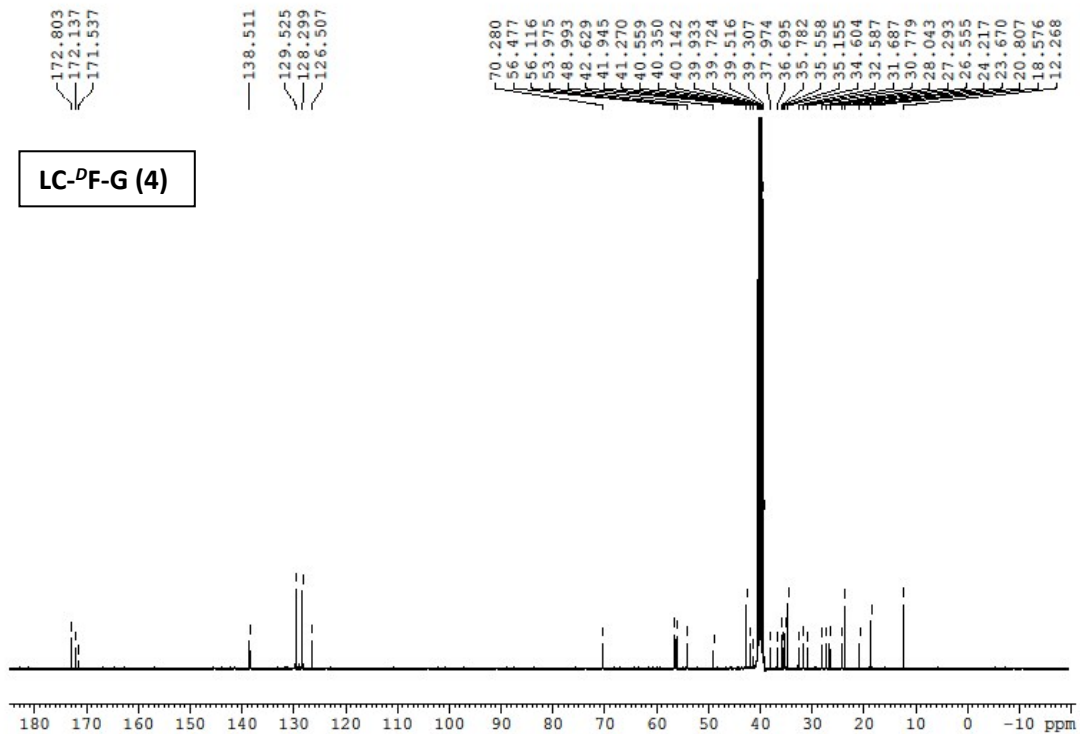
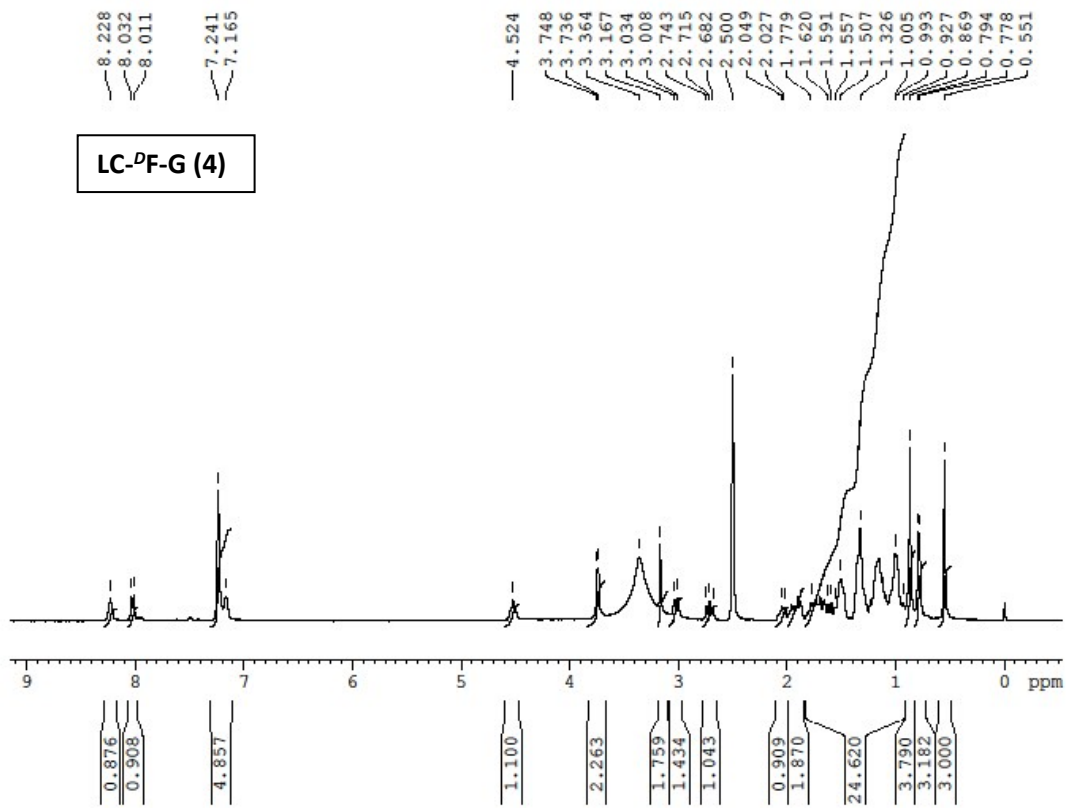


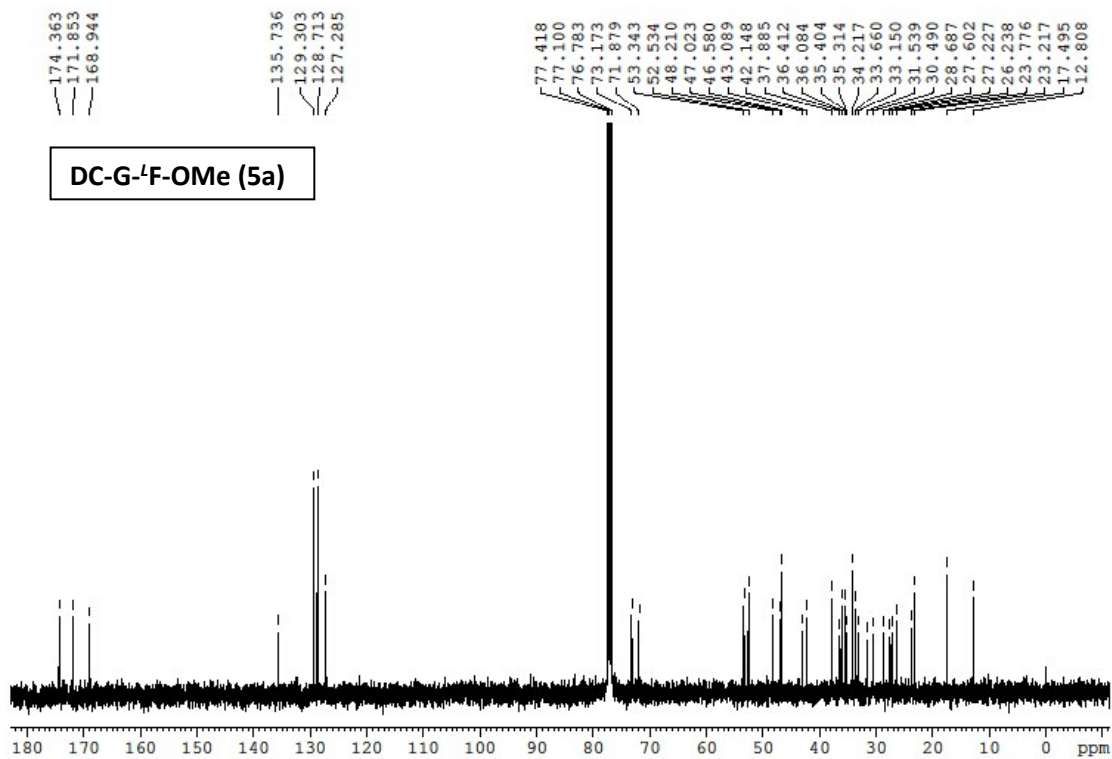
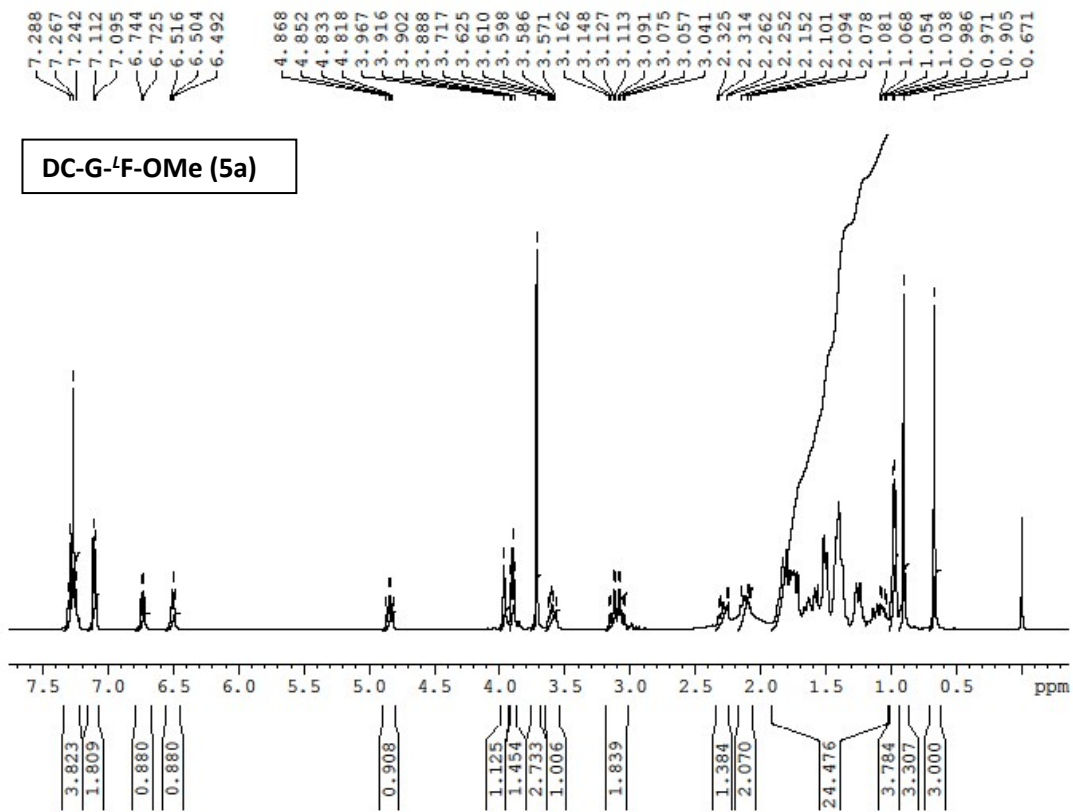


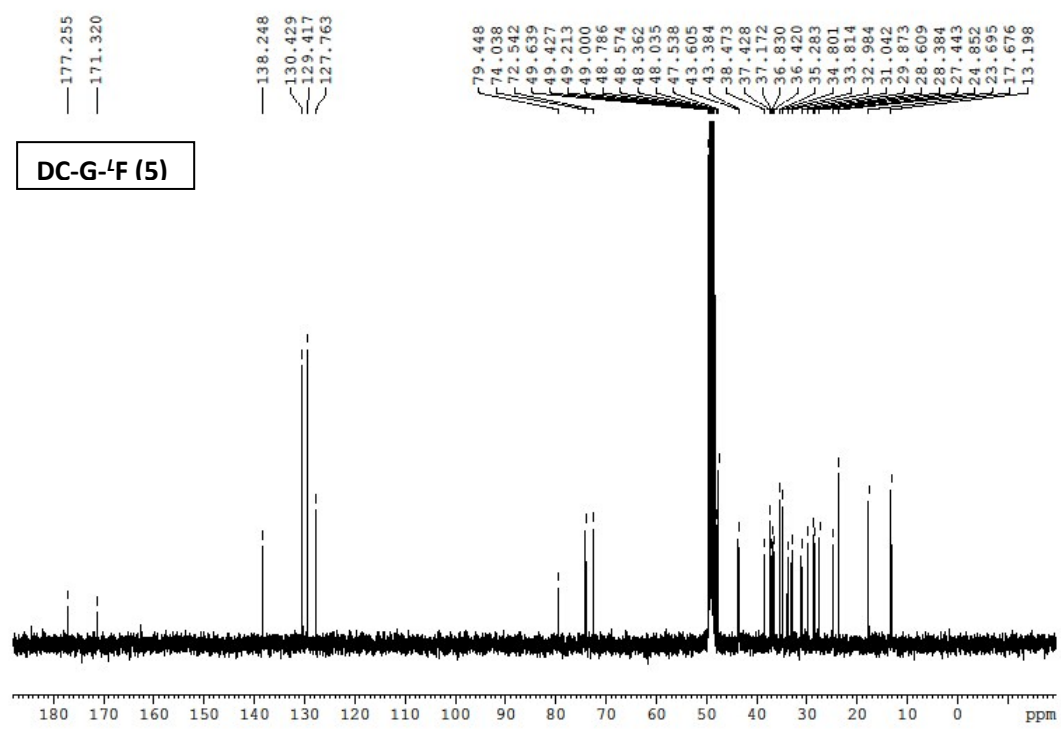
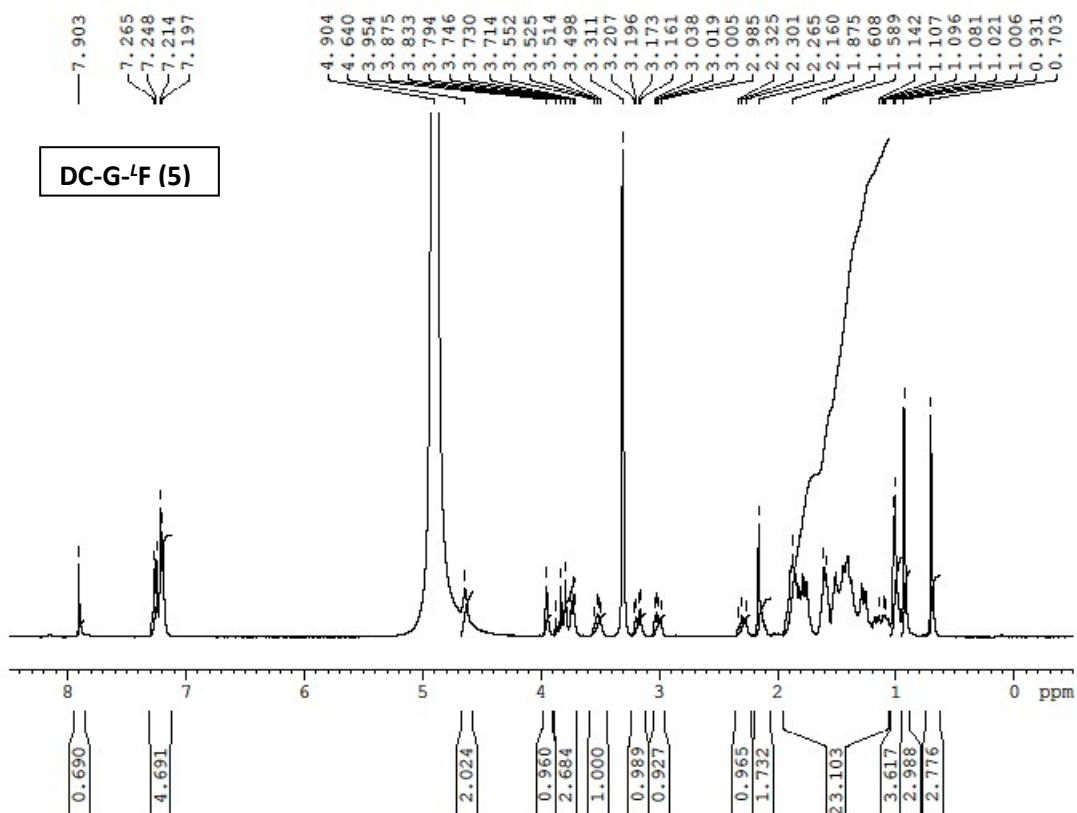




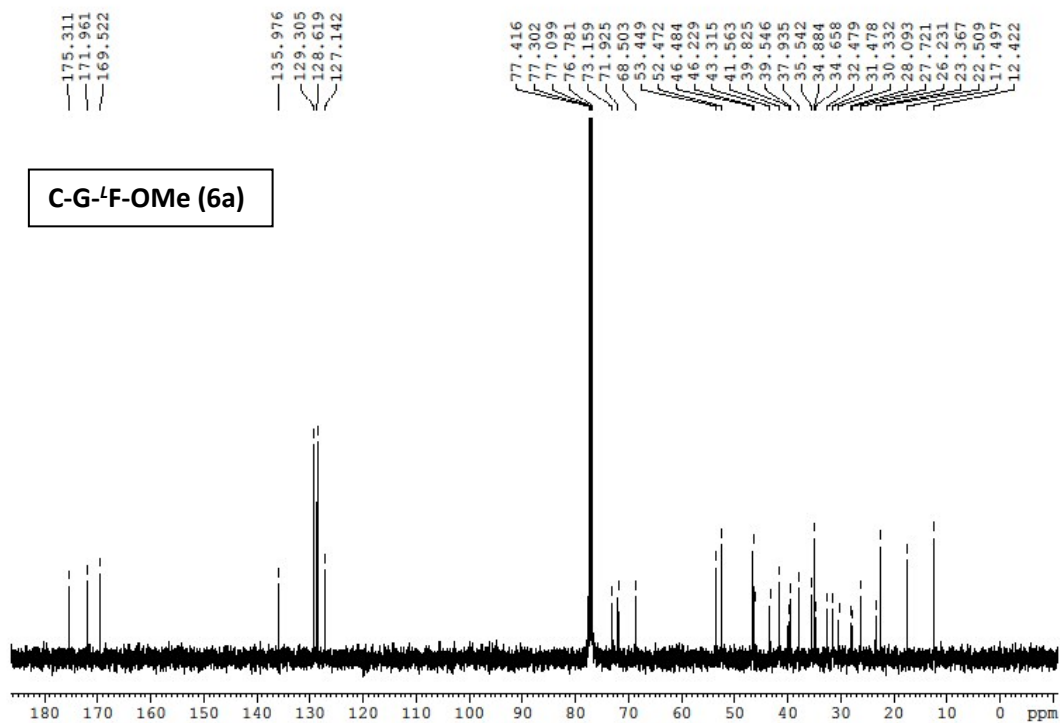
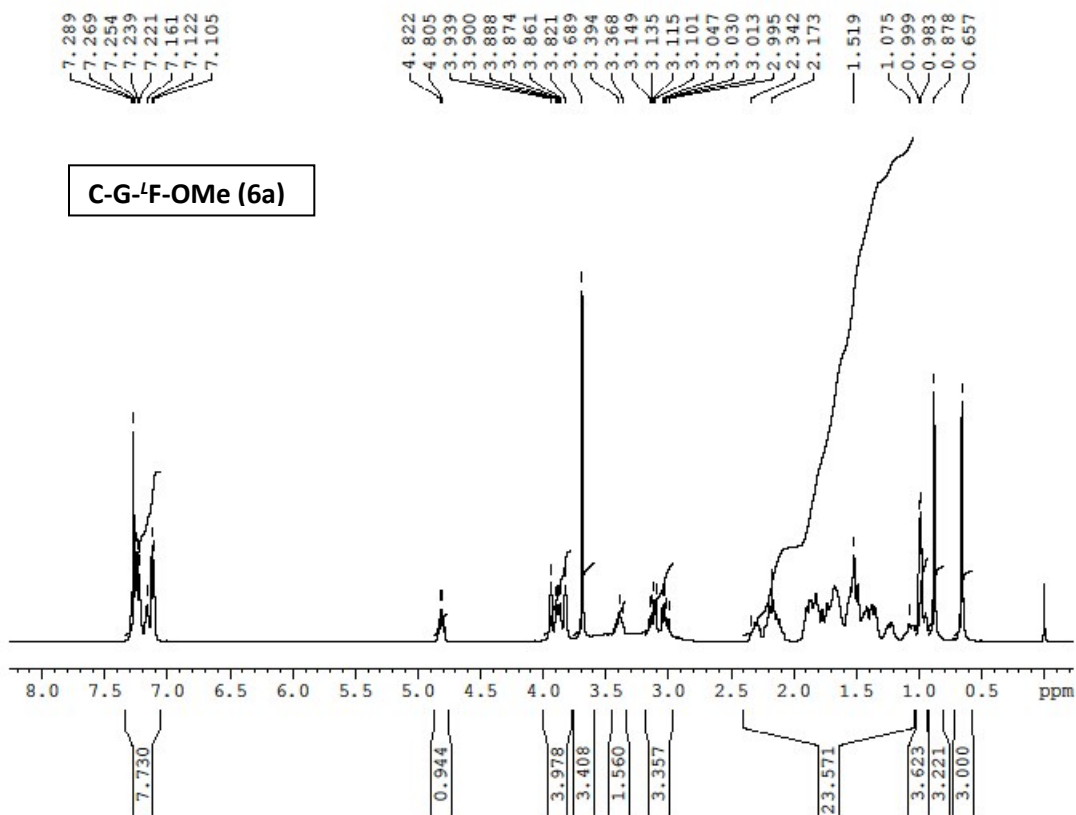


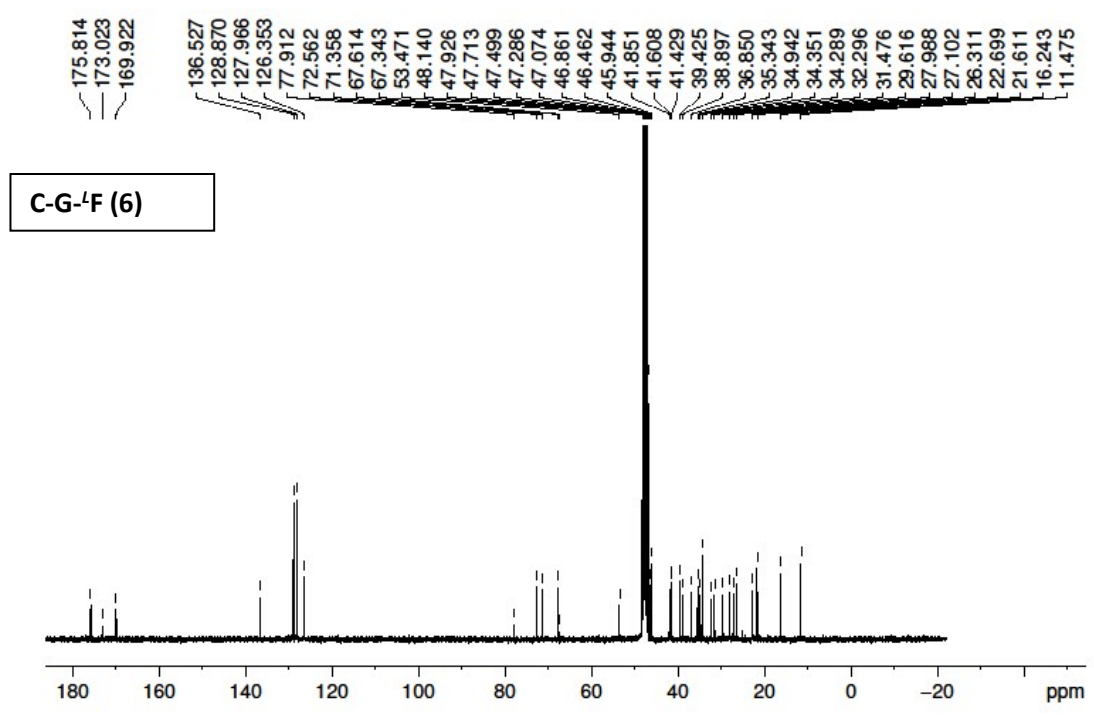
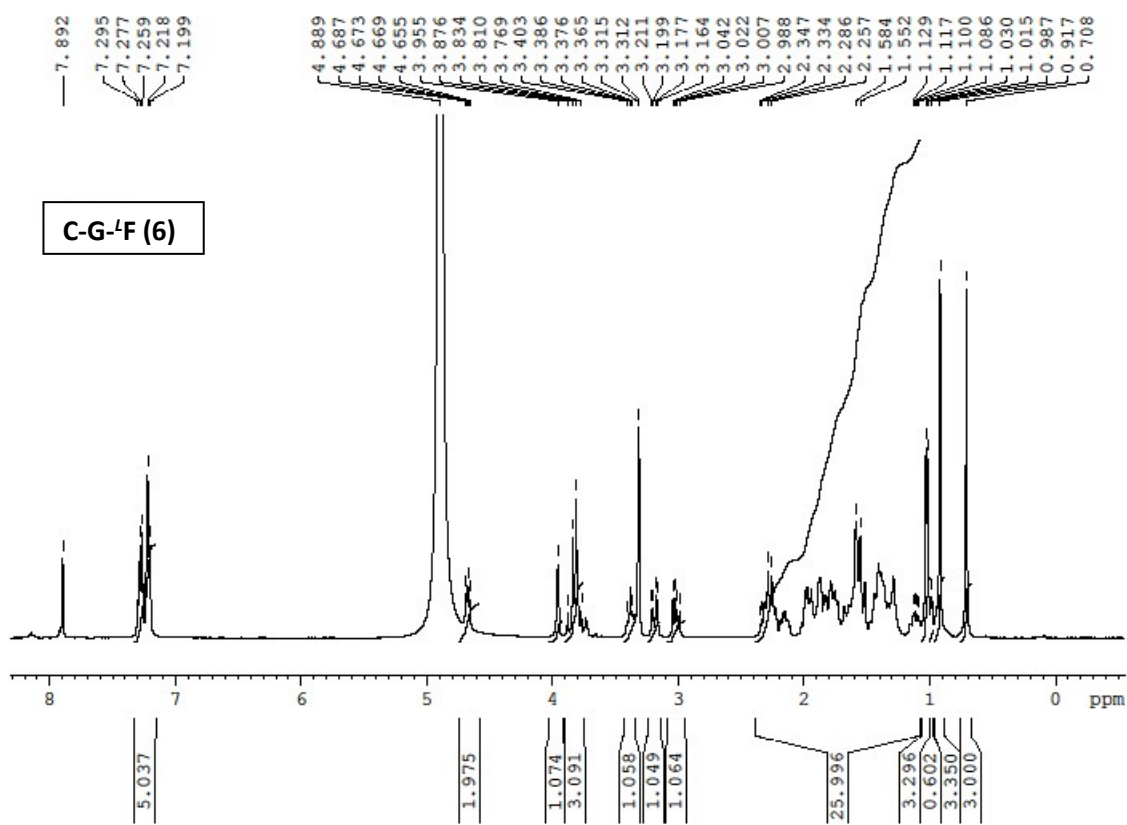


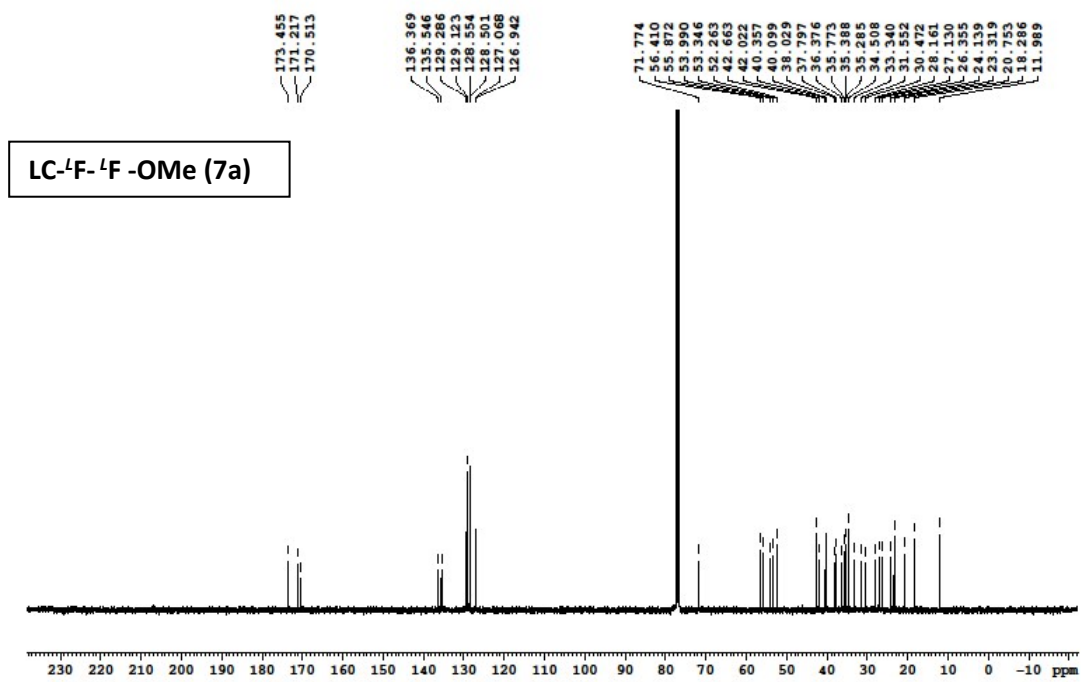
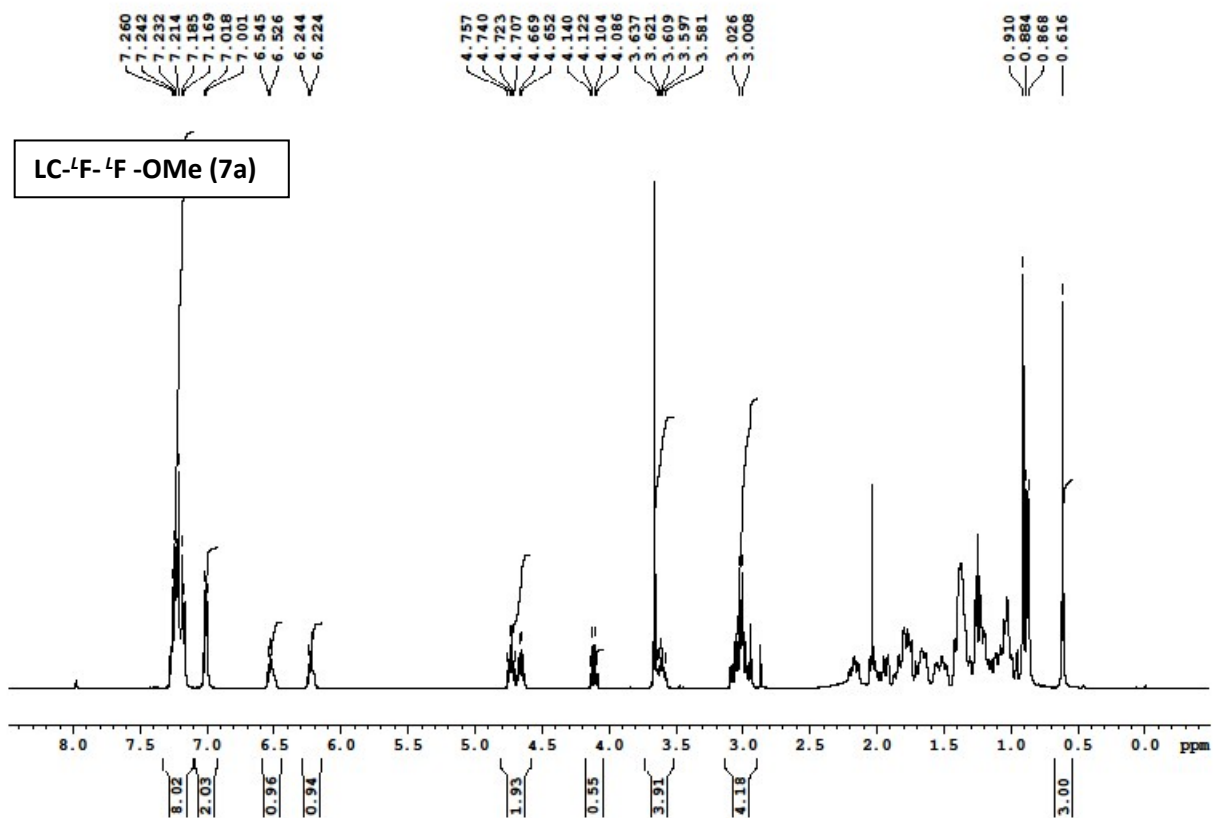


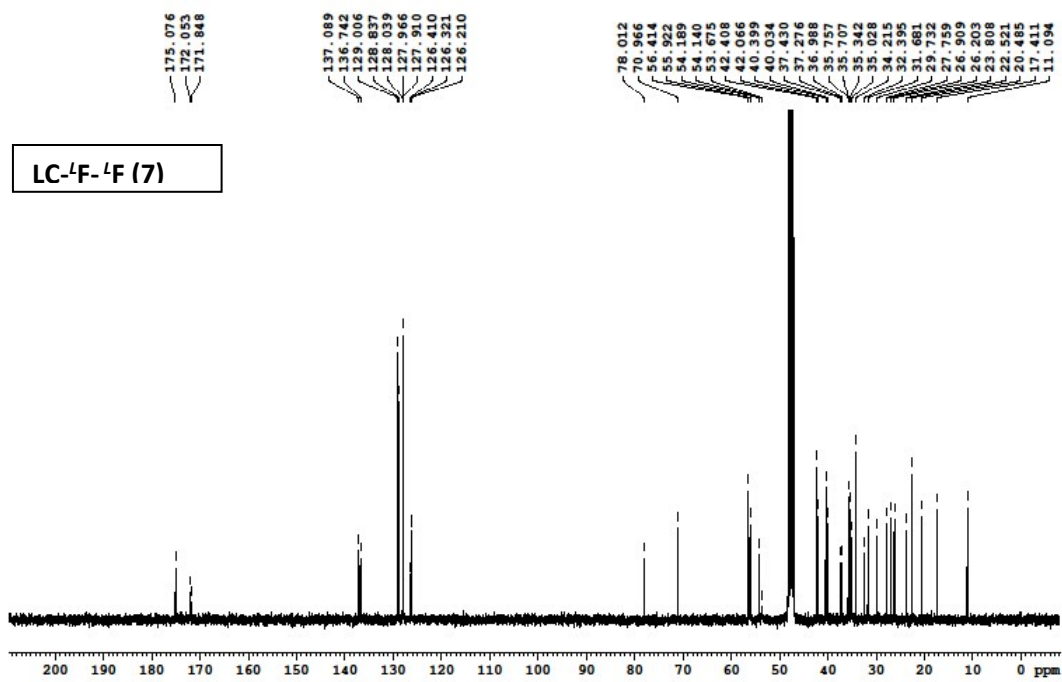
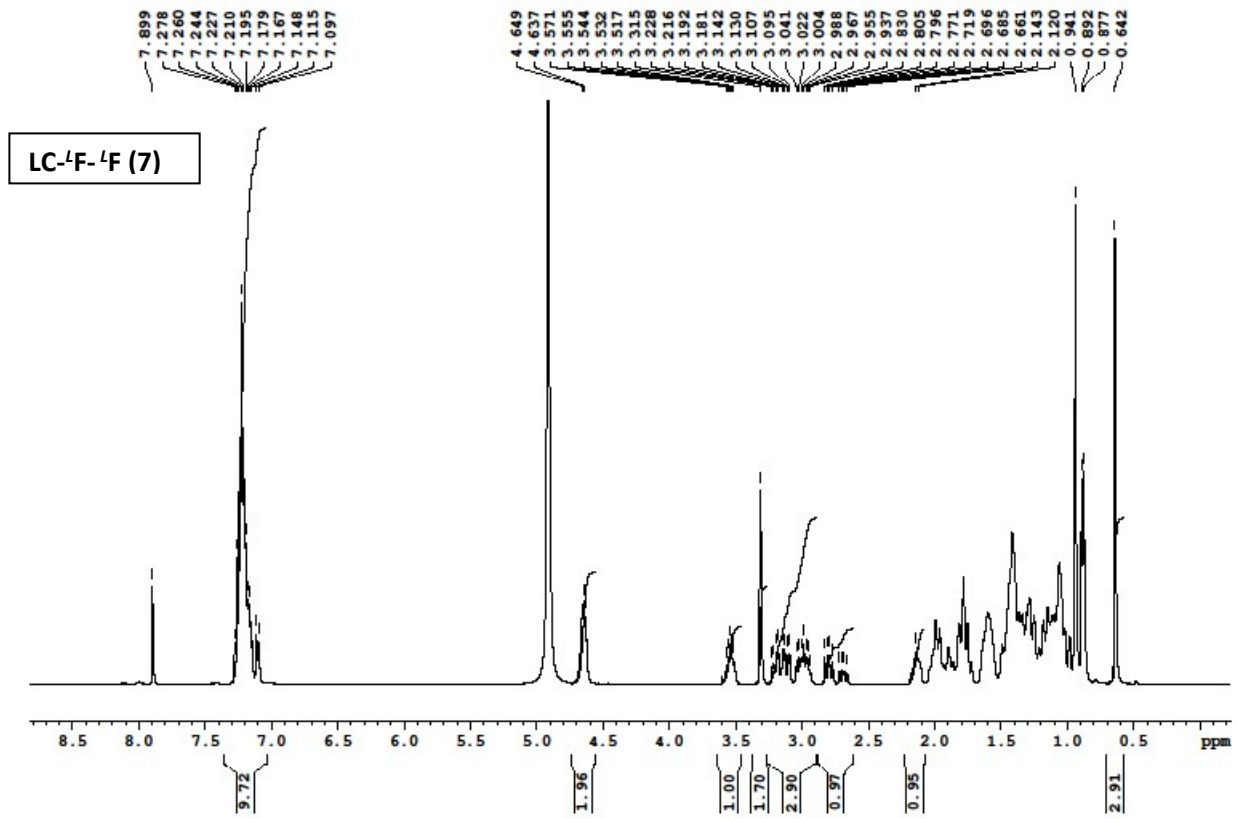


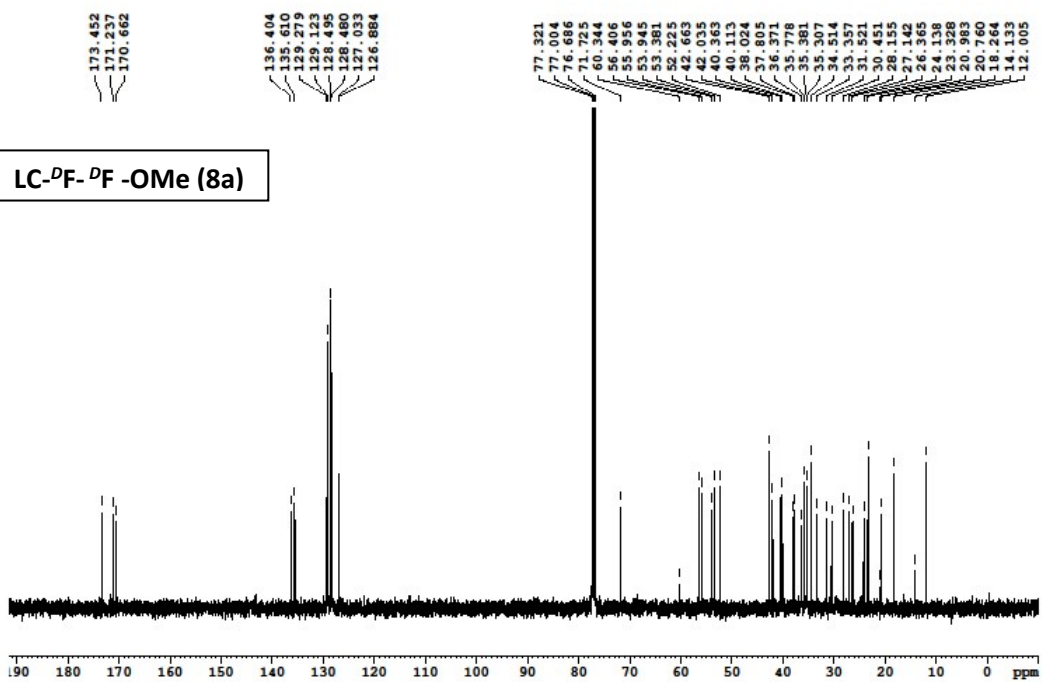
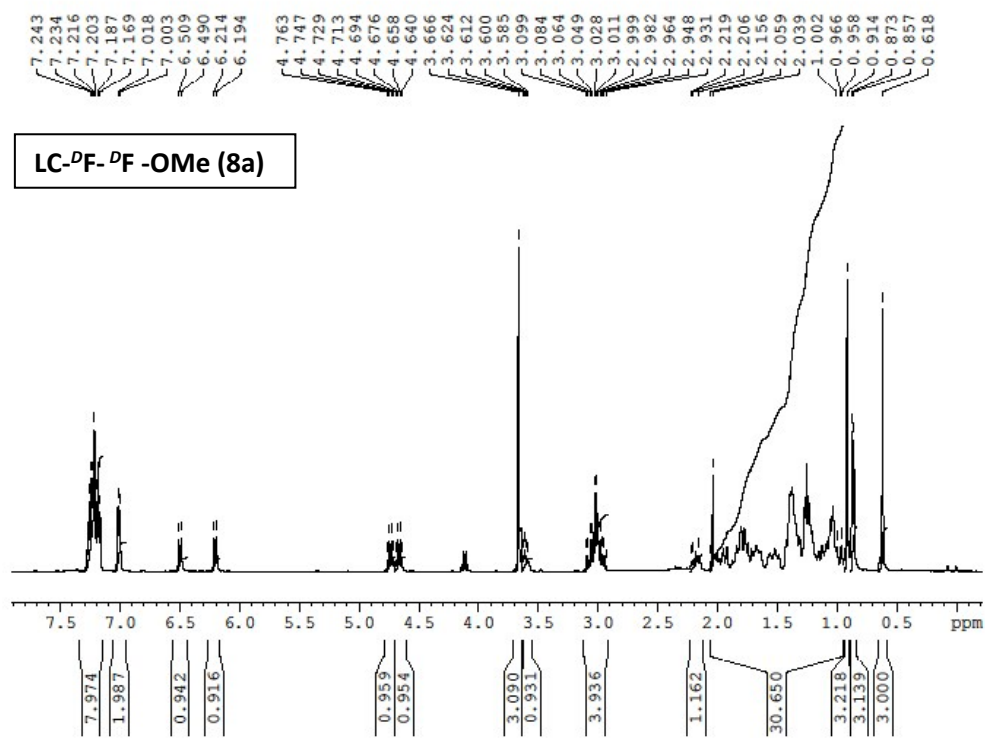




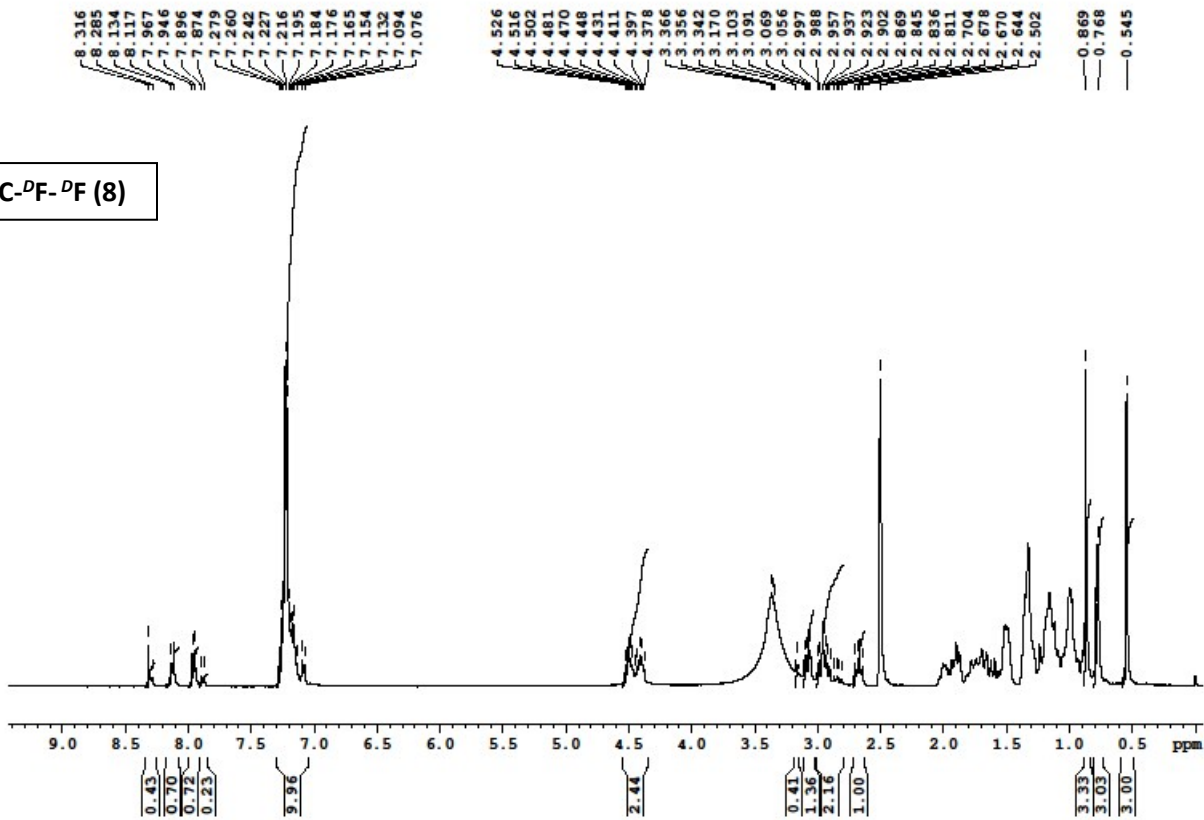








LC-DF-DF (8)



LC-DF-DF (8)

