Supporting Information

An operational transformation of 3-carboxy-4quinolones into 3-nitro-4-quinolones via ipsonitration using polysaccharide supported copper nanoparticles: synthesis of 3-tetrazolyl bioisosteres of 3-carboxy-4-quinolones as antibacterial

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General Information

Reagent grade solvents were used for the extraction and flash chromatography. All the reagents and chemicals were purchased from Sigma Aldrich Chemical Co., Merck and were used directly without further purification. The progress of reactions was checked by analytical thin-layer chromatography (TLC, Merck silica gel 60F-254 plates). The plates were visualized by UV illumination. Column chromatography was performed using silica gel (60-120 mesh). The solvent compositions reported for all chromatographic separations are on a volume/volume (v/v) basis. All glassware's were dried oven before use in connection with an inert atmosphere. Solvents were evaporated under reduced pressure. Tetramethylsilane (0.0 ppm) was used as an internal standard in ¹H NMR and CDCl₃ (77.0 ppm) was used in ¹³C NMR. The abbreviations used to indicate the peak multiplicity were; s, singlet; bs, broad singlet; d, doublet; dd, double doublet; t, triplet; q, quartet; m, multiplet; Hz, Hertz. FAB MS was recorded on Jeol (Japan)/SX-102. Infrared spectrum was taken with ATR on Shimadzu IR-affinity-1s. Melting points were determined on a Buchi 565 digital melting point apparatus and were uncorrected..

Experimental procedure:

Synthesis of Polysac-Cu-NP

For the preparation of Plysac-Cu-NP cellulose and starch used as such but chitosan grind to very fine powder in mortar and pastel prior to use. The polysaccharide (1gm) were suspended in deionised water and sonicated at 40 kHz for 20 min. To polysaccharide suspension copper sulphate (250 mg) was added. The reaction mixture was stirred for 15 min under nitrogen atmosphere at room temperature and then cooled to 0°C by ice bath. The polysaccharide cupper suspension carefully reduced by sodium borohydride (100 mg) in small portion with short interwal of time. The black reaction mixture was again stirred for 1 hr till the temperature reached to rt. The black suspension again sonicated for about 15 min before filtration. The solid black mass washed with de-ionised water, vacuumed dried and stored under nitrogen atmosphere.

ED-XRF report of chit-cu-NP synthesized

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12/9/2015 4:09:00 PM
AIRF-JNU
Auto Quantify -
       Sample Id.: CEPS-01
      Date / Time: 12/9/2015 4:00:33 PM
             Type: Routine - 1/1
Initial weight (g): 1.16
 Total weight (g): 1.27
                                           Result
Compound Corr. (cps/mA) Conc.
                              Unit
                                     Status
С
                       90.472 %
        0.371
A1203
                       0.115
                               8
                                 Calibrated
        2.555
SiO2
                       0.438 %
                                  Calibrated
                       251.065 ppm Calibrated
S
        0.596
Cl
                       592.215 ppm Calibrated
        3.797
K20
       7.244
                      312.649 ppm Calibrated
CaO
       12.834
                      410.358 ppm Calibrated
TiO2 1.883
Fe2O3 34.126
                       77.566 ppm Calibrated
                       258.232 ppm Calibrated
Cu
       18181.180
                      8.682 % Calibrated
        69.927
                       281.003 ppm Calibrated
ZnO
                       690.471 ppm Calibrated
Yb
        10.298
W
        6.242
                       60.409 ppm Calibrated
Sum
                       100.000 %
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ED-XRF report of chit-cu-NP after fifth cycle

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12/9/2015 3:50:00 PM
AIRF-JNU
Auto Quantify -
       Sample Id.: CEPS-02
      Date / Time: 12/9/2015 3:33:51 PM
             Type: Routine - 1/1
Initial weight (g): 1.56
 Total weight (g): 1.78
                                            Result
Compound Corr. (cps/mA) Conc. Unit Status
CH2
                       92.057
                               00
A1203
        0.389
                       1971.364 ppm Calibrated
SiO2
        2.335
                       0.667
                               00
                                    Calibrated
        0.632
                       230.896 ppm Calibrated
S
                      501.802 ppm Calibrated
Cl
        3.710
K20
        7.121
                      266.161 ppm Calibrated
CaO
        12.400
                      343.187 ppm Calibrated
                               ppm Calibrated
TiO2
        1.362
                      48.612
Fe2O3 32.772
                      214.970 ppm Calibrated
       17819.500
                      6.865
                                    Calibrated
Cu
                               00
        91.815
                       329.780 ppm Calibrated
ZnO
Rh
        0.000
                       0.000
                               ppm Calibrated
W
        24.219
                       208.989 ppm Calibrated
                      100.000 %
Sum
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Typical experimental procedure for the synthesis of 3-nitro-4-quinolone: In a typical experiment, the quinolone acid (7) $(1 \text{ mmol})^1$, Chit-Cu-NP (37 mg, 5 mol %) and nitronium tetrafluoroborate (1.2 mmol) were taken in 50 ml round-bottom flask containing DMF (10 mL) and the reaction mixture was refluxed, till the completion of the reaction (monitored by TLC). After completion the reaction mixture was vacuum and extracted with ethyl acetate (25 ml x3) after addition of de-ionised water (50 ml), the collected EtOAc dried over sodium sulphate and evaporated under vacuum to give crude product (7). Crude was purified by silica gel (60-120 mesh) column chromatography to afford the corresponding product.

7-chloro-1-ethyl-6-fluoro-3-nitroquinolin-4(1H)-one (7a)



Yield: 92 %; pale yellow solid; mp 270-272°C (uncorrected). IR (v_{max} , ATR, cm-1): 3444, 3278, 1660, 1434, 1309, 1249, 839, 456, 180. ¹H NMR (300 MHz,) δ 9.71 (s, 1H), 8.14 (d, *J* = 8.0 Hz, 1H), 7.79 (d, *J* = 5.9 Hz, 1H), 4.50 (q, *J* = 6.9 Hz, 2H), 1.56 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃ +

DMSO*d*₆) δ 162.94 (d, *J* = 6.5 Hz), 157.95 (s), 152.75 (s), 146.18 (s), 135.61 (d, *J* = 8.5 Hz), 131.89 (s), 126.12 (*m*), 117.85 (d, *J* = 11 Hz), 113.22, 112.82, 48.66 (s), 13.64 (s). ES-MS (M+H): 271.0 m/z.

7-chloro-6-fluoro-1-isopropyl-3-nitroquinolin-4(1H)-one (7b)



Yield 85%; pale yellow solid; mp 263-256°C (uncorrected). IR (v_{max} , ATR, cm⁻¹): 3432, 3258, 1655, 1480, 1320, 1232, 842, 460; ¹H NMR (300 MHz, CDCl₃) δ 9.69 (s, 1H), 8.11 (m, 1H), 7.70 (dd, J = 3.4, 2.5 Hz, 1H), 4.74 (m, 1H), 1.56 (m, 7H); ¹³C NMR (100 MHz, CDCl₃ + DMSOd₆) δ 162.88 (d, J

= 8.7 Hz), 157.98 (s), 152.78 (s), 147.24 (s), 137.47 (d, *J* = 8.5 Hz), 132.12(s), 127.80 (d, *J* = 10 Hz), 126.19, 125.82, 118.89 (d, *J* = 13 Hz), 113.08, 112.69, 54.41 (s), 19.83 (s). ES-MS (M+H): 285.0 m/z.

<u>1-butyl-7-</u>chloro-6-fluoro-3-nitroquinolin-4(1H)-one (7c)

^{1.} Note: All the precursors were synthesized according to the published literature. (Koga, H.; Itoh A.; Murayama, S.; Suzue S. and Irikura T., *J. Med. Chem.*; 1980, **23**, 1358)

^{2.} Raghavan, K.; Lang, S. A. and Marshall M. S., J. Heterocycl. Chem, 1986, 23, 1801.



Yield: 82%; pale yellow solid; mp 275-278°C (uncorrected). IR (v_{max} , ATR, cm⁻¹): 3465, 3262, 1652, 1474, 1322, 1232, 1211, 851. ¹H NMR (300 MHz, CDCl₃) δ 9.66 (s, 1H), 8.13 (m, 1H), 7.79 (d, J = 5.7 Hz, 1H), 4.10 (t, J = 7.6 Hz, 2H), 1.81 (m, 2H), 1.59 (m, 2H), 1.10 (dd, J = 8.8, 5.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃ + DMSOd₆) δ 162.83 (d, J = 6.5 Hz), 157.90 (s), 152.70 (s),

145.21 (s), 135.4 (d, J = 6.5 Hz), 131.70 (s), 126.37, 125.9943, 124.60 (d, J = 11 Hz), 116.31 (d, J = 11 Hz), 113.26, 112.88, 48.21 (s), 29.21 (s), 18.99 (s), 13.28 (s). Anal. calcd for C₁₃H₁₂ClFN₂O₃: C, 52.27; H, 4.05; N, 9.38; Found C, 52.21; H, 4.12; N, 9.21. ES-MS (M+H): 299.1 m/z.

1-(sec-butyl)-7-chloro-6-fluoro-3-nitroquinolin-4(1H)-one (7d)



Yield 85%; pale yellow solid; mp 265-270°C (uncorrected). IR (v_{max} , ATR, cm⁻¹): 3421, 3269, 1644, 1452, 1325, 1238, 867, 380, 190. ¹H NMR (300 MHz, CDCl₃) δ 9.60 (s, 1H), 8.11 (m, 1H), 7.75 (d, *J* = 5.6 Hz, 1H), 4.78 (m, 1H), 1.48 (m, 2H), 1.17 (d, *J* = 6.1 Hz, 3H), 0.92 (m, 3H); ¹³C NMR

(100 MHz, $CDCl_3 + DMSOd_6$) δ 162.88 (d, J = 9 Hz), 157.63 (s), 152.43 (s), 145.59 (s), 137.97 (d, J = 9 Hz), 132.26 (s), 127.44 (d, J = 12 Hz), 126.25 (s), 125.88 (s), 118.92 (d, J = 11 Hz), 113.14 (s), 112.75 (s), 50.46 (s), 26.77 (s), 19.91 (s), 8.00 (s). Elemental Analysis calculated for $C_{13}H_{12}ClFN_2O_3$: C, 52.27; H, 4.05; N, 9.38; Found C, 52.11; H, 4.10; N, 9.29. ES-MS (M+H): 299.1 m/z.

7-chloro-6-fluoro-1-methyl-3-nitroquinolin-4(1H)-one (7e)



Yield 79%; yellow solid; m.p. 198-200°C, IR (vmax, ATR, cm-1): 3412, 1610, 1511, 1302, 1121, 1062, 541; ¹H NMR (300 MHz, CDCl₃): δ 9.67 (d *J*=0.5 Hz, 1H), 8.15 (m,1H), 7.87 (d *J*=5.5 Hz, 1H), 3.5 (s, 1H) ; ¹³C NMR (100 MHz, CDCl₃ + DMSO *d6*): δ 168.34 (d, *J* = 2.53 Hz),

156.97, 153.89, 151.37, 141.28 (d, *J* = 3 Hz), 127.48 (t, *J* = 20 Hz), 121.99 (d, *J* = 8.34 Hz), 115.0 (t, *J* = 7.64 Hz), 110.61 (d, *J* = 19.88 Hz), 38.23 ; ES-MS (M+H): 257.04 m/z.

7-chloro-1-cyclopropyl-6-fluoro-3-nitroquinolin-4(1H)-one (7f)



Yield 76%; yellow solid; m.p. 210-212°C. IR (vmax, ATR, cm-1): 3310, 1643, 1555, 1363, 1122, 1069, 581; ¹H NMR (300 MHz, CDCl₃): δ 9.69 (s, 1H), 8.14 (d *J*= 7.95 Hz, 1H), 7.73 (d J= 5.64 Hz, 1H), 3.75 (m, 1H), 1.04 (d *J*=4.83 Hz, 2H), 0.84 (d *J*= 2.43 Hz, 2H). ¹³C NMR (100 MHz, CDCl₃ + DMSO *d*6): δ 13C NMR (100 MHz, Common

NMR Solvents) δ 168.35 (d, J = 2.93 Hz), 158.32, 158.67, 157.99, 155.47, 140.22 (d, J = 2.84 Hz), 127.97 (d, J = 19.76 Hz), 120.20 (d, J = 8.34 Hz), 119.34, 114.05 (d, J = 7.66 Hz, H), 113.01 (d, J = 1.73 Hz), 113.0, 40.13, 10.78 ; ES-MS (M+H): 283.12 m/z.

6,7-difluoro-3-nitro-1-propylquinolin-4(1H)-one (7g)



Yield 86% pale yellow solid; mp 262-263°C (uncorrected). IR (v_{max} , ATR, cm⁻¹): 3286, 3050, 1644, 1534, 1454, 1311, 1211, 831. ¹H NMR (300 MHz, CDCl₃) δ 9.65 (m, 1H), 8.30 (t, *J* = 9.5 Hz, 1H), 7.42 (dd, *J* = 11.0, 6.5 Hz, 1H), 3.59 (t, *J* = 7.4 Hz, 2H), 1.55 (m, 2H), 1.04 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃ + DMSOd₆) δ 161.32 (d, *J* = 9 Hz), 155.71 (s),

154.93(s), 150.49 (d, J = 1.2 Hz), 150.11 (d, J = 2 Hz), 149.73 (s), 145.28 (s), 144.90 (m), 136.04 (m), 132.00 (s), 123.84 (m), 112.90 (d, J = 11 Hz), 112.52 (d, J = 11.5 Hz) 114.20 (d, J = 13 Hz) 104.01 (d, J = 13 Hz), 50.83 (s), 21.73 (s), 9.96 (s). ES-MS (M+H): 269.8 m/z.

1-(sec-butyl)-6,7-difluoro-3-nitroquinolin-4(1H)-one (7h)



Yield 88 %; yellow solid; mp 265-266°C (uncorrected). IR (v_{max} , ATR, cm⁻¹): 3276, 3105, 1631, 1510, 1432, 1331, 1232, 852. ¹H NMR (300 MHz, CDCl₃) δ 9.66 (s, 1H), 8.31 (t, *J* = 9.4 Hz, 1H), 7.43 (m, 1H), 4.79 (m, 1H), 1.49 (m, 2H), 1.17 (d, *J* = 6.0 Hz, 3H), 0.94 (dd, *J* = 9.4, 5.4 Hz, 3H); ¹³C

NMR (100 MHz, $CDCl_3 + DMSOd_6$) δ 161.23 (d, J = 6.5 Hz), 154.96 (s), 154.57 (s), 150.48 (s), 150.11 (s), 149.74 (s), 147.40 (d, J = 0.6 Hz), 145.43 (s), 145.1 (d, J = 0.5 Hz), 144.90 (d, J = 1 Hz), 136.68 (m), 132.20 (s), 124.36 (m), 122.75 (d, J = 11 Hz), 12.37 (d, J = 13 Hz), 105.05 (d, J = 14 Hz) 104.67 (d, J = 11.5 Hz), 50.29 (s), 27.38 (s), 19.73 (s), 8.00 (s). ES-MS (M+H): 283.0 m/z.

6,7-difluoro-3-nitro-1-pentylquinolin-4(1H)-one (7i) Yiled 91%; pale yellow solid; mp 269-



271°C (uncorrected). IR (v_{max} , ATR, cm⁻¹): 3406, 3209, 3109, 1648, 1551, 1469, 1325, 1212, 867. ¹H NMR (300 MHz, CDCl₃) δ 9.67 (s, 1H), 8.31 (t, *J* = 9.5 Hz, 1H), 7.58 (dd, *J* = 11.0, 6.4 Hz, 1H), 4.03 (t, *J* = 7.5 Hz,

2H), 1.30 (m, 6H), 0.87 (dd, J = 8.5, 4.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃ + DMSOd₆) δ 161.21 (d, J = 9 Hz), 155.29 (s), 154.91 (s), 150.77 (s), 150.40 (s), 150.08 (s), 149.71 (d, J = 0.75 Hz), 145.58 (s), 145.33 (s), 145.19 (s), 135.35 (m), 131.63 (s), 122.01 (m), 122.85 (d, J = 11 Hz), 122.49 (d, J = 11 Hz), 102.55 (d, J = 11 Hz), 102.16 (d, J = 13 Hz), 48.55 (s), 30.54 (s), 27.94 (s), 21.15 (s), 12.30 (s). ES-MS (M+H): 297.1 m/z.

6,7-difluoro-3-nitro-1-octylquinolin-4(1H)-one (7j)



Yield 82% pale yellow solid; mp 276-279°C (uncorrected). IR (v_{max}, ATR, cm⁻¹): 3434, 3221, 3089, 1659, 1532, 1473, 1362, 1239, 856. ¹H NMR (300 MHz, CDCl₃) δ 9.67 (s, 1H), 8.31 (t, *J* = 9.5 Hz, 1H), 7.59 (dd, *J* = 11.0, 6.4 Hz, 1H), 4.04 (dd, *J* = 11.5, 4.1 Hz, 2H),

1.42 (m, 2H), 1.22 (m, 10H), 0.84 (m, 3H); ¹³C NMR (100 MHz, $CDCl_3 + DMSOd_6$) δ 161.19 (d, J = 9 Hz), 155.26 (s), 154.88 (s), 150.76 (s), 150.39 (s), 150.07 (s), 149.68 (s), 145.95 (s), 145.56 (s), 145.18 (s), 135.36 (m), 131.62 (s), 122.02 (m), 112.85 (d, J = 13 Hz), 112.48 (d, J = 13 Hz), 102.52 (d, J = 11 Hz), 102.14 (d, J = 11 Hz), 49.77 (s), 30.40 (s), 28.68 (s), 28.59 (s), 27.71 (s), 26.67 (s), 21.91 (s), 13.37 (s); ES-MS (M+H): 339.1 m/z.

6,7-dichloro-1-ethyl-3-nitroquinolin-4(1H)-one (7k)



Yield 79%; pale yellow solid; mp 266-267°C (uncorrected). IR (v_{max} , ATR, cm⁻¹): 3421, 3211, 3090, 1651, 1512, 1485, 1352, 1231, 856, 660; ¹H NMR (300 MHz, CDCl₃) δ 9.82 (s, 1H), 8.49 (s, 1H), 7.97 (s, 1H), 4.53 (q, *J* = 6.7 Hz, 2H), 1.56 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃ + DMSOd₆)

δ 163.41, 146.26, 136.80, 136.45, 132.07, 129.76, 128.04, 126.82, 117.46, 48.05, 13. 65. ES-MS (M+H): 286.9 m/z.

6,7-dichloro-1-cyclopropyl-3-nitroquinolin-4(1H)-one (7l)



Yield 75% pale yellow solid; mp 271-271°C (uncorrected). IR (v_{max} , ATR, cm⁻¹): 3440, 3321, 3101, 1644, 1521, 1467, 1331, 1242, 866, 649. ¹H NMR (300 MHz, CDCl₃) δ 9.79 (s, 1H), 8.47 (s, 1H), 7.89 (s, 1H), 3.67 (dd, J =

2.9, 1.9 Hz, 1H), 1.02 (m, 2H), 0.82 (m, 2H); ¹³C NMR (100 MHz, CDCl₃ + DMSOd₆) δ 163.01, 147.23, 136.17, 135.26, 132.60, 129.79, 127.79, 127.75, 118.75, 33.42, 10.32. ES-MS (M+H): 298.9 m/z.

1-butyl-6,7-dichloro-3-nitroquinolin-4(1H)-one (7m)



Yield 79%; yellow solid; 268-270°C (uncorrected). IR (v_{max} , ATR, cm⁻¹): 3412, 3287, 3089, 1639, 1532, 1471, 1328, 1220, 869, 598. ¹H NMR (300 MHz, CDCl₃) δ 9.77 (s, 1H), 8.49 (s, 1H), 7.95 (s, 1H), 4.13 (t, *J* = 7.6 Hz, 2H), 1.81 (m, 2H), 1.60 (m, 2H), 1.09 (m, 3H); ¹³C NMR (100 MHz, CDCl₃ + DMSOd₆) δ 163.27, 145.26, 137.15, 136.48, 131.86, 129.70,

128.05, 125.14, 115.71, 47.60, 29.21, 18.96, 13.27. ES-MS (M+H): 315.02 m/z.

1-(sec-butyl)-6,7-dichloro-3-nitroquinolin-4(1H)-one (7n)



Yield 79%; pale yellow solid; mp 265-266°C (uncorrected). IR (ν_{max} , ATR, cm⁻¹): 3367, 3298, 3083, 1661, 1541, 1469, 1373, 1249, 841, 582, 421. ¹H NMR (300 MHz, CDCl₃) δ 9.71 (s, 1H), 8.45 (s, 1H), 7.97 (s, 1H), 4.79 (m, 1H), 1.49 (m, 2H), 1.16 (d, *J* = 6.1 Hz, 3H), 0.91 (dd, *J* = 9.6, 5.5 Hz,

3H); ¹³C NMR (100 MHz, CDCl₃ + DMSOd₆) δ 163.30, 145.65, 137.82, 136.37, 132.45, 129.43, 128.00, 127.96, 118.37, 49.83, 26.76, 19.90, 8.01; ES-MS (M+H): 315.02 m/z.

6,7-dichloro-1-heptyl-3-nitroquinolin-4(1*H*)-one (70)



Yield 89%; pale yellow solid; mp 280-282°C (uncorrected). IR (v_{max} , ATR, cm⁻¹): 3421, 3321, 3211, 3108, 1659, 1488, 1392, 1241, 860, 637, 487. ¹H NMR (300 MHz, CDCl₃) δ 9.75 (m, 1H), 8.48 (s, 1H), 7.95 (s, 1H), 4.10 (td, *J* = 7.5, 0.6 Hz, 2H), 1.45 (m, 4H), 1.21 (m, 6H), 0.85 (m, 3H); ¹³C NMR (100 MHz, CDCl₃ + DMSOd₆)

) δ 163.25, 146.18, 137.11, 136.47, 131.86, 129.69, 128.05, 125.13, 115.70, 48.26, 30.99, 27.78, 27.32, 26.65, 21.98, 13.37. ES-MS (M+H): 357.07 m/z.

1-benzyl-7-chloro-6-fluoro-3-nitroquinolin-4(1H)-one (10a)



Yield 52%; orange yellow; mp 244-246°C (uncorrected);). IR (vmax, ATR, cm-1): 3461, 3311, 3286, 3124, 1599, 1488, 1342. ¹H NMR (300 MHz, CDCl₃) δ 9.70 (s,1H), 8.15 (d *J*=7.74 Hz, 1H), 7.58-7.43 (m, 1H), 7.41-7.16 (m, 2H),

7.16-6.90 (m, 1H), 5.58 (s, 2H) ¹³C NMR (100 MHz, $CDCl_3 + DMSOd_6$) δ 168.34 (d, J = 2.93 Hz), 155.90, 155.39, 152.86, 138.78 (d, J = 3 Hz), 136.46, 127.76 (d, J=27.45 Hz), 127.32 (d, J = 16.84 Hz), 120.02 (d, J = 8.11 Hz), 116.97 (d, J = 7.6 Hz), 112.04, 110.45 (d, J = 21.08 Hz), 53.56. ES-MS (M+H): 333.62 m/z.

7-chloro-6-fluoro-3-nitro-1-(4-nitrobenzyl)quinolin-4(1H)-one (10b)



Yield 22%; canary yellow; mp 256-285°c; ¹H NMR (300 MHz, CDCl3) δ 9.7 (s, 1H), 8.36-8.03 (m, 3H), 7.71-7.46 (m, 3H), 5.67 (s, 2H); ¹³C NMR (100 MHz, CDCl3 + DMSOd6) δ 168.35 (d, J = 2.93 Hz), 155.91, 155.40, 152.87, 144.87, 144.23, 138.79 (d, J = 3 Hz), 127.66 (d, J = 6.78 Hz), 127.42, 122.92, 122.88, 120.03 (d, J = 8.11 Hz),

116.98 (d, *J* = 7.61 Hz), 112.05, 110.46 (d, *J* = 19.69 Hz, H), 53.57. ES-MS (M+H): 378.07 m/z.

General method for the preparation of 7-chloro-6-fluoro-1-methyl-3-(1H-tetrazol-1-yl)quinolin-4(1H)-one

To a pre-cooled solution of 8 (4 mmol) in conc. HCl (25 ml) was added portion wise anhydrous $SnCl_4$ (3.5 equivalent) with vigorous stirring. The resulting mixture was then allowed to rt, and stirring was continued for additional 4 h. Afterward, the reaction mixture was diluted with cold water (25ml), basified with 40 % cold aq. NaOH solution to pH ~ 12-14. The precipitated solid product was collected by suction filtration, washed with cold water, dried and recrystallized from ethanol to give 3-amine derivative, which was further purified by column chromatography using eluent 5% CH₃OH/CHCl₃. 10 mmole of 3-amine derivative was dissolved in 25 ml of glacial acetic acid and to that 11mmol of NaN₃ and 12 mmole of triethyl orthoformate were added and the resultant reaction mixture was heated to 100-110°C for 7-8 h. The flow of the reaction was monitored by TLC. After the completion of the reaction, the reaction mixture was poured into crushed ice (100gm) and the solid was filtered, washed with water and dried under vacuum to yield 12 in good yield.

7-chloro-6-fluoro-1-methyl-3-(1H-tetrazol-1-yl)quinolin-4(1H)-one



(14a)

Yield 82%; yellow solid; m.p. 252-253°C, IR (vmax, ATR, cm-1): 3417, 1608, 1534, 1300, 1122, 1012, 563; NMR (300 MHz, CDCl₃): δ 4.57 (s,

3H), 8.16 (d *J*=8Hz, 1H), 7.22 (d *J*=6 Hz), 8.19 (d, *J*=8Hz), 9.55 (s, 1H), 9.64 (s, 1H) ; ¹³C NMR (75 MHz, CDCl₃): δ 43.83, 111.69, 114.38, 114.74, 117.53, 117.65, 125.34, 125.46, 126.9, 127.29, 129.43, 138.01, 140.39, 140.47, 153.51, 158.71, 175.69, 175.77; ES-MS (M+H): 280.04 m/z.

7-chloro-1-cyclopropyl-6-fluoro-3-(1H-tetrazol-1-yl)quinolin-4(1H)one (14b)

Yield 82%; yellow solid; m.p. 268-270°C, IR (vmax, ATR, cm-1): 3409, 1605, 1532, 1304, 1115, 1043, 564; NMR (300 MHz, CDCl₃): δ 1.97–1.88 (m, 4H), 4.10-4.11 (m, 1H), 7.22 (d *J*=6 Hz), 8.20 (d, *J*=8Hz),

9.55 (s, 1H), 9.64 (s, 1H) ; ¹³C NMR (75 MHz, CDCl₃): δ 8.43, 35.44, 112.46, 113.96, 114.33, 120.06, 120.18, 126.49, 126.86, 127.54, 127.69, 136.01, 136.09, 137.81, 153.57, 158.74, 175.22, 175.3; ES-MS (M+H): 306.05 m/z.

7-chloro-1-ethyl-6-fluoro-3-(1H-tetrazol-1-yl)quinolin-4(1H)-one (14c)



Yield 82%; yellow solid; m.p. 277-278°C, IR (vmax, ATR, cm-1): 3410, 1605, 1580, 1318, 1122, 1022, 582; NMR (300 MHz, CDCl₃): δ 1.42 (t, 3H), 4.48 (d, *J*=6.8 Hz), 7.25 (d, *J*=6 Hz), 8.19 (d, *J*=8Hz), 9.55 (s, 1H), 9.64 (s, 1H); ¹³C NMR (75 MHz, CDCl₃): δ 10.7, 49.22, 111.92, 114.24,

114.61, 118.59, 118.69, 126.22, 126.35, 126.77, 127.14, 128.6, 136.63, 136.7, 137.32, 153.53, 158.74, 175.62, 175.7; ES-MS (M+H): 294.8 m/z.



7-chloro-6-fluoro-1-isopropyl-3-(1H-tetrazol-1-yl)quinolin-4(1H)-one (14d)

Yield 82%; yellow solid; m.p. 273-274°C, IR (vmax, ATR, cm-1): 3410, 1608, 1484, 1322, 1119, 1025, 566; NMR (300 MHz, CDCl₃): δ 1.55 (6H), 4.93 (s, 1H), 7.25 (d, *J*=6 Hz), 8.19 (d, *J*=8Hz), 9.55 (s, 1H), 9.64 (s, 1H);

¹³C NMR (75 MHz, CDCl₃): δ 16.87, 56.12, 112.14, 114.1, 114.48, 119.61, 119.73, 126.62, 127.01, 127.08, 127.2, 127.77, 137.96, 138.13, 138.2, 153.56, 158.76, 175.54, 175.61; ES-MS (M+H): 308.8 m/z.

General method for the preparation of 6-fluoro-1-alkyl-7-(piperazin-1-yl)-3-(1H-tetrazol-1-yl)quinolin-4(1H)-one

4 mmol of **8** was taken in a round bottom flask having 20 mm of DMSO in which 6 mmol of piperazine was added and reaction mixture was allowed to stirrer at 120-130°C for about 9-10h. After completion of reaction as evidenced by TLC, DMSO was removed in vacuum and reaction mixture partitioned in between DCM and water. DCM layer was successveily wased with 0.5 N HCl (20 ml), water (20ml X 3) and finally with brine. The DCM layer on vaccume evaporation yield crude **11** which was further purified by column chromatography using eluent 40% EtOAc/Hexane. After getting **11** same method was applied as used for the synthesis of 12a-d to yield 11a-d in good yield.



6-fluoro-1-methyl-7-(piperazin-1-yl)-3-(1H-tetrazol-1-yl)quinolin-4(1H)-one (13a)

Yield 82%; light brown solid; m.p. 326-3227°C, IR (vmax, ATR, cm-1): 3412, 1603, 1484, 1322, 1122, 1023, 756, 583; NMR (300 MHz, CDCl₃+DMSO-d6): δ 3.14 (m, 4H), 3.50 (m, 4H), 4.02 (bs, NH), 4.52

(s, 3H), 6.60 (d, J=6.73 Hz, 1H), 7.83 (d, J=12.6 Hz, 1H), 9.50 (s, 1H), 9.59 (s, 1H); ¹³C NMR (75 MHz, CDCl₃+DMSO-d6): δ 43.99, 45.1, 49.9, 49.99, 102.85, 102.98, 107.47, 112.48, 112.87, 121.33, 121.46, 129.36, 138.02, 141, 141.08, 144.59, 144.96, 150.06, 155.25, 175.06, 175.14; ES-MS (M+H): 330.2 m/z.



1-cyclopropyl-6-fluoro-7-(piperazin-1-yl)-3-(1H-tetrazol-1-

yl)quinolin-4(1H)-one (13b)

Yield 82%; light brown solid; m.p. 333-334°C, IR (vmax, ATR, cm-1): 3414, 1599, 1481, 1319, 1119, 1033, 777, 576; NMR (300 MHz,

CDCl₃+DMSO-d6): δ 0.98 (m, 2H), 1.13 (m, 2H), 3.10 (m, 4H), 3.44 (m, 4H), 4.05 (bs, NH), 7.57 (s, 1H), 7.84 (d, J=6.5 Hz), 9.51 (s, 1H), 9.60 (s, 1H); ¹³C NMR (75 MHz, CDCl₃+DMSO-d6): δ 6.78, 33.55, 45.1, 49.9, 49.98, 106.56, 106.68, 108.24, 112.06, 112.44, 122.68, 122.81, 127.64, 137.82, 138.76, 138.83, 143.18, 143.56, 150.1, 155.28, 174.6, 174.68; ES-MS (M+H): 356.3 m/z.



Yield 82%; light brown solid; m.p. 356°C, IR (vmax, ATR, cm-1): 3408, 1402, 1321, 1131, 1028, 802, 566; NMR (300 MHz, CDCl₃+DMSO-d6): δ 1.44 (t, 2H), 3.15 (m, 4H), 3.51 (m, 4H), 4.08 (bs, NH), 4.50 (dd, 2H), 7.02 (d, J=6Hz, 1H), 7.88 (d, J=12.5Hz, 1H),

9.56 (s, 1H), 9.67 (s,1H); ¹³C NMR (75 MHz, CDCl3+DMSO-d6): δ 10.93, 45.1, 48.79, 49.9, 49.97, 104.18, 104.28, 107.7, 112.34, 112.74, 121.79, 121.91, 128.52, 137.32, 138.73, 138.8, 144.97, 145.36, 150.08, 155.27, 174.99, 175.06; ES-MS (M+H): 344.3 m/z.

6-fluoro-1-isopropyl-7-(piperazin-1-yl)-3-(1H-tetrazol-1-



yl)quinolin-4(1H)-one (13d)

Yield 82%; light brown solid; m.p. 348-349°C, IR (vmax, ATR, cm-1): 3422, 1610, 1490, 1304, 1132, 1042, 768, 595; NMR (300 MHz, CDCl₃): δ 1.47 (m, 6H), 3.05 (m, 4H), 3.43 (m, 4H), 3.98 (bs, NH(,

4.85 (m, 1H), 7.52 (s, 1H), 7.76 (m, 1H), 9.46 (s, 1H), 9.55 (s, 1H); ¹³C NMR (75 MHz, CDCl3+DMSO-d6): δ 17.1, 45.11, 49.89, 49.98, 55.88, 106.11, 106.23, 107.92, 112.22, 112.6, 122.22, 122.34, 127.71, 137.98, 141.69, 141.76, 143.33, 143.71, 150.1, 155.3, 174.91, 175; ES-MS (M+H): 358.2.2 m/z.



¹H and ¹³C NMR Spectra of compound 7a



¹H and ¹³C NMR Spectra of compound **7b**



 ^1H and ^{13}C NMR Spectra of compound 7c



¹H and ¹³C NMR Spectra of compound 7d



 ^1H and ^{13}C NMR Spectra of compound 7e



¹H and ¹³C NMR Spectra of compound 7f



 ^1H and ^{13}C NMR Spectra of compound 7g



 ^1H and ^{13}C NMR Spectra of compound 7h



 ^1H and ^{13}C NMR Spectra of compound 7i



¹H and ¹³C NMR Spectra of compound 7j



 ^1H and ^{13}C NMR Spectra of compound 7k



¹H and ¹³C NMR Spectra of compound 7I



 ^1H and ^{13}C NMR Spectra of compound 7m



 ^1H and ^{13}C NMR Spectra of compound 7n



¹H and ¹³C NMR Spectra of compound 70



 ^1H and ^{13}C NMR Spectra of compound 10a



 ^1H and ^{13}C NMR Spectra of compound 10b



 ^1H and ^{13}C NMR Spectra of compound 13a



 ^1H and ^{13}C NMR Spectra of compound 13b



¹H and ¹³C NMR Spectra of compound 13c



 ^1H and ^{13}C NMR Spectra of compound 13d



 ^1H and ^{13}C NMR Spectra of compound 14a



 ^1H and ^{13}C NMR Spectra of compound 14b



 ^1H and ^{13}C NMR Spectra of compound 14c



¹H and ¹³C NMR Spectra of compound 14d

Method for biological evaluation

Solvent Used: DMSO	Standard	Antibiotic	used:
	Ciprofloxacin		

Concentrations screened: 6.25, 12.5, 25, 50 and 100 µM

Stock Sample Concentration: 1 mM

Name of the analysis method: Mirco-dilution method

Bacteria analyzed: *Staphylococcus aureus, Staphylococcus aureus (MRSA), Escherichia coli, Salmonella typhii* and *Vibrio cholerae*

Description:

Media Used (Muller Hinton broth): Beef extract 2g; Casein hydrolysate 17.5g; Starch 1.5g in 1000 ml of distilled water. Initially, the stock cultures of bacteria were revived by inoculating in broth media and grown at 37°C for 18 hrs. A 0.5ml culture was centrifuged and resuspended in 250µl volume of above sterile medium to obtain a 5x109 CFU/ml which was diluted to 5x105 CFU/ml. Ciprofloxacin was used as a standard and compounds (CSA001- CSA004) were added at the concentrations mentioned above. The bacterial solution (4x107 CFU/ml) was mixed well and this suspension was added to each well of microtitre plate so that the final volume of compounds and media reaches to 200 µl. The microtitre plate was incubated at 37°C for 24 h and bacterial growth was observed. The visual MIC was reached by comparing the turbidity with that of uninoculated broth. The complete visual similarity of test wells with that of uninoculated broth is considered as MIC.

Reference:

Methods for dilution anti-microbial susceptibility tests for bacteria that grow aerobically, January, 2012, Clinical and Laboratory Standards Institute (CLSI), M07-A9, Vol 32 (2), pp 17-18.



Figure. The Photograph of visual MIC determination of synthesized compounds.

Method for docking studies

The molecular modelling study was performed with Molegro Virtual Docker (MVD) v 4.0.0 (www.molegro.com) along with Graphical User Interface (GUI), MVD tools was used to generate grid, calculate dock score and evaluate conformers. The scoring function used by MolDock is derived from the Piecewise Linear Potential (PLP) scoring functions. The active binding site was considered as a rigid molecule, whereas the ligands were treated as being flexible, that is, all non-

ring torsions were allowed. The structures of title compounds (13 and 14) were subjected to energy minimization using molecular mechanics (MM2). The minimization was performed until the root mean square (RMS) gradient value reached a value smaller than 0.001 kcal/mol. Finally the protein target (2XCT) was prepared for molecular docking simulation in such a way that all heteroatoms (i.e., non-receptor atoms such as water, ions, etc) were removed. The active binding site region was defined as a spherical region which encompasses all protein within 15.0 Å of bound crystallographic ligand atom. The docking protocol includes the grid resolution of 0.30, for grid generation and 15 A radius from the template as the binding site. MolDock SE was used as a search algorithm and the number of runs was set to 10 Å population size of 50 and maximum interactions of 1500 were used for parameter settings. The maximum number of poses generated was 10. Since MVD works by an evolutionary algorithm, successive docking runs do not give precisely the same pose and interactions. To address this inherent randomness, three consecutive runs were done and the best three poses were used to visualize the interactions of all inhibitors.