

Supporting Information

Tunable two-photon absorption near-infrared materials by different electron-donors and π -bridge center with applications in bioimaging in live cells

Ming Kong,^a Ting Wang,^b Xiaohu Tian,^a Fang Wang,^a Yanqiu Liu,^a Qiong Zhang,^a Hui
Wang,^a Hongping Zhou,^a Jieying Wu ^{*a} and Yupeng Tian ^{*a}

^a Department of Chemistry, Key Laboratory of Functional Inorganic Material
Chemistry of Anhui Province, Anhui University, Hefei 230039, P. R. China

^b School of Life Science, Anhui University, Anhui Province Key Laboratory of R&D
of Chinese Medicine, Hefei, China

E-mail: jywu1957@163.com; yptian@ahu.edu.cn

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S1. General methods

All chemicals were commercially available and used as obtained. The solvents were

purified by conventional methods before use. The ^1H NMR and ^{13}C NMR spectra recorded on at 25°C using Bruker Avance 400 spectrometer were reported as parts per million (ppm) from TMS. IR spectra were recorded on NEXUS 870 (Nicolet) spectrophotometer in the $400\text{-}4000\text{ cm}^{-1}$ region using a powder sample on a KBr plate. Mass spectra were obtained on a Micromass MALDI-TOF-MS.

X-ray crystallography and structure solution. The X-ray diffraction measurements were performed on a Bruker SMART CCD area detector using graphite monochromated $\text{MoK}\alpha$ radiation ($\lambda = 0.71069\text{ \AA}$) at $298(2)\text{ K}$. Intensity data were collected in the variable ω -scan mode. The structures were solved by direct methods and difference Fourier syntheses. The non-hydrogen atoms were refined anisotropically and hydrogen atoms were introduced geometrically. Calculations were performed with the SHELXTL-97 program package. Crystallographic data (excluding structure factors) for the structures reported in this paper have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication no. CCDC number: **X1** (965153), **XT1** (991562), **XT2** (991563), **XT3** (991564). Details of the crystal parameters, data collections and refinements for the complexes are summarized in **Table 1** and selected bond distances and angles are listed in **supporting**.

Absorption and fluorescence spectra. Absorption spectra were obtained on a UV-265 spectrophotometer, Fluorescence measurements were performed using a Hitachi F-7000 fluorescence spectrophotometer excited at the longest absorption band with a 1 cm standard quartz cell. The concentration of sample solution was $1.0 \times 10^{-5}\text{ mol/L}$. The excitation and emission slit widths were all 5 nm. For time-resolved fluorescence measurements, the fluorescence signals were collimated and focused onto the entrance slit of a monochromator with the output plane equipped with a photomultiplier tube (HORIBA HuoroMax-4P). The decays were analyzed by 'least-squares'. The quality of the exponential fits was evaluated by the goodness of fit (χ^2).

Theoretical calculation. To better identify the charge transition, time-dependent density functional theory (TD-DFT) calculations were carried out in ethanol. Geometry optimizations (**X2** and **X3**) were carried out with B3LYP functional

without any symmetry restraint, and the TD-DFT calculations were performed on the optimized structure with B3LYP functional. For **X1**, **XT1**, **XT2** and **XT3**, the crystal structures were directly adopted for TD-DFT calculations. All calculations, including optimizations and TD-DFT, were performed with the G03 software. Geometry optimization of the singlet ground state and the TD-DFT calculation of the lowest 50 singlet-singlet excitation energies were calculated with a basis set composed of 6-31 G for C H N O S atoms. An analytical frequency confirmed evidence that the calculated species represents a true minimum without imaginary frequencies on the respective potential energy surface. The lowest 40 spin-allowed singlet-singlet transitions, up to energy of about 5 eV, were taken into account in the calculation of the absorption spectra.

Fluorescence quantum yield

The fluorescence quantum yield was measured by using

$$\Phi_s = \Phi_r \left(\frac{A_r \eta_s^2 D_s}{A_s \eta_r^2 D_r} \right)$$

in which the subscripts *s* and *r* stand for designate the sample and reference sample, respectively, *A* is the absorbance at λ_{exc} , η is the average refractive index of the appropriate solution, and *D* is the integrated fluorescence area under the corrected emission spectrum. Here, quinine sulfate in 1 N H₂SO₄ was used as the reference ($\Phi=0.54$). All the measurements were at room temperature.

Microscopy. HepG2 cells were imaged on a Zeiss LSM 710 META upright confocal laser scanning microscope using magnification 40× and 100× water-dipping lenses for monolayer cultures. Image data acquisition and processing was performed using Zeiss LSM Image Browser, Zeiss LSM Image Expert and Image J.

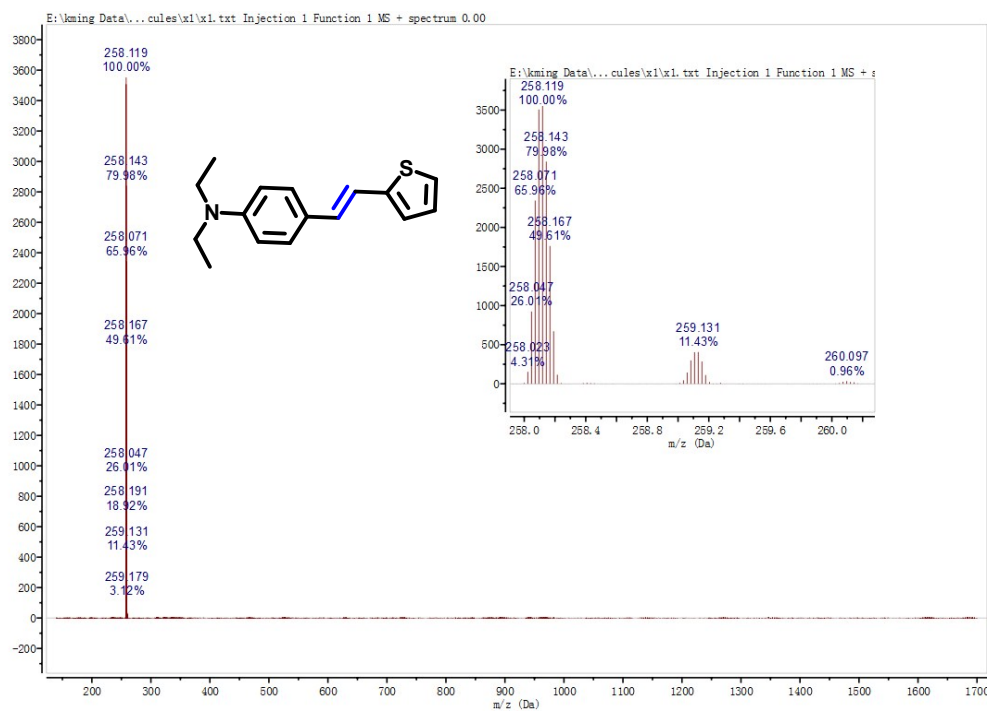


Figure S1. MALDI-TOF-MS of X1

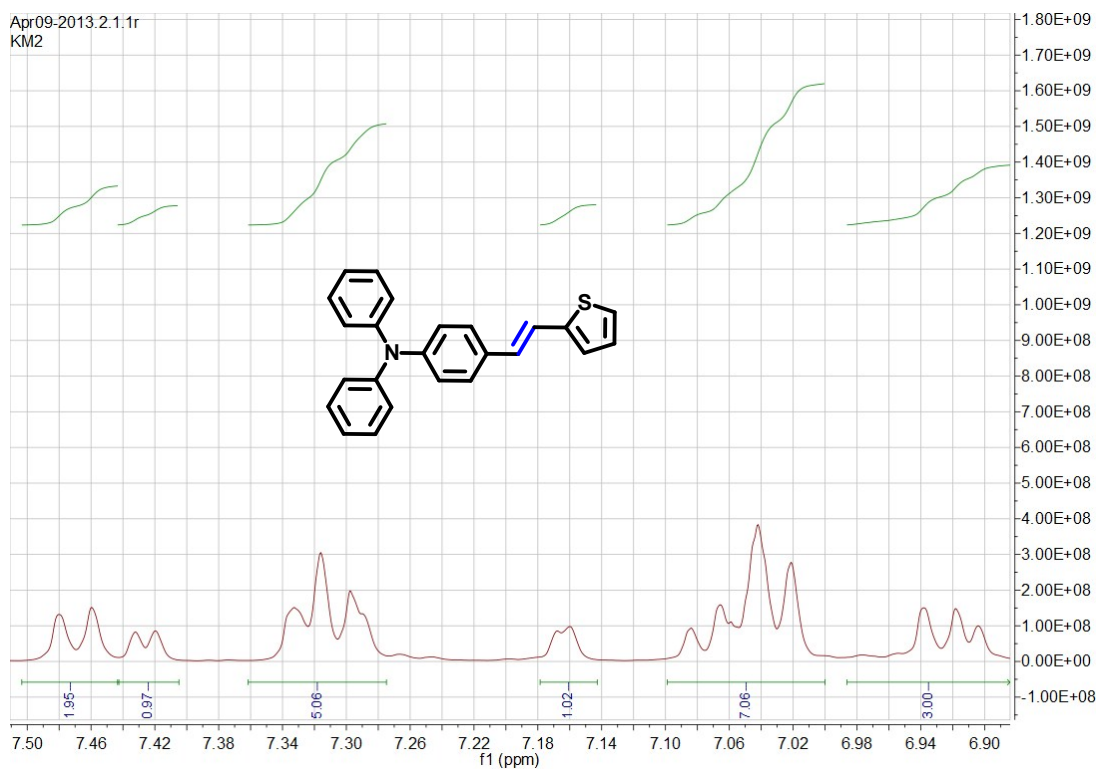


Figure S2. ¹H NMR of X2

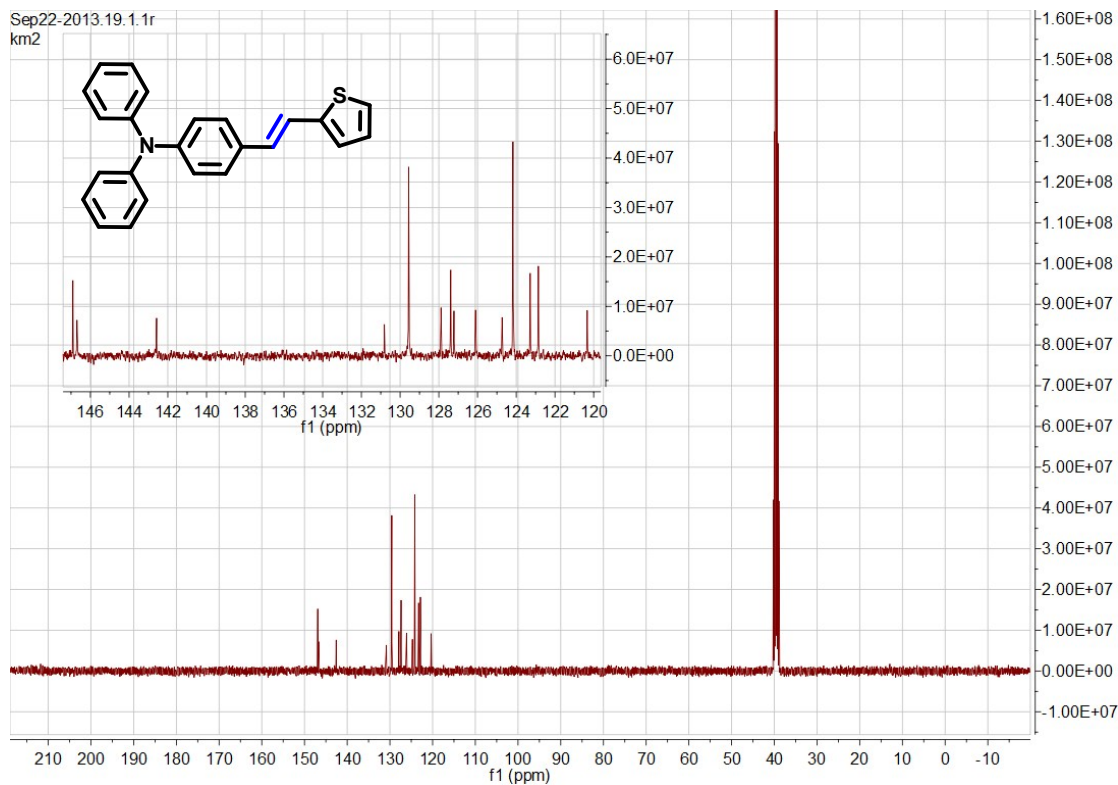


Figure S3. ^{13}C NMR of X2

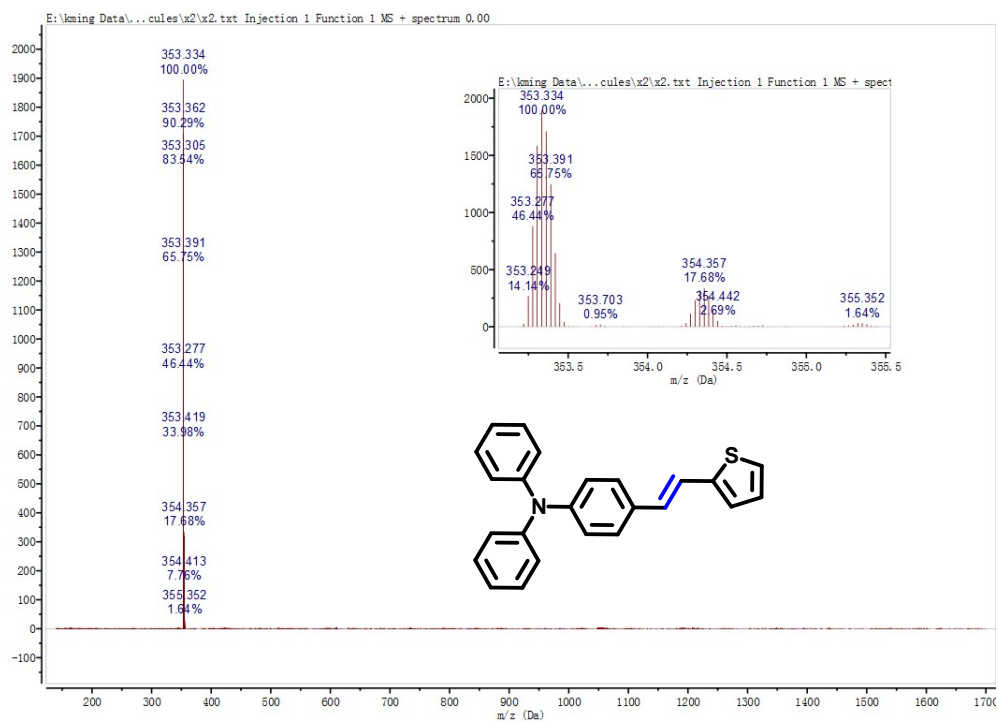


Figure S4. MALDI-TOF-MS of X2

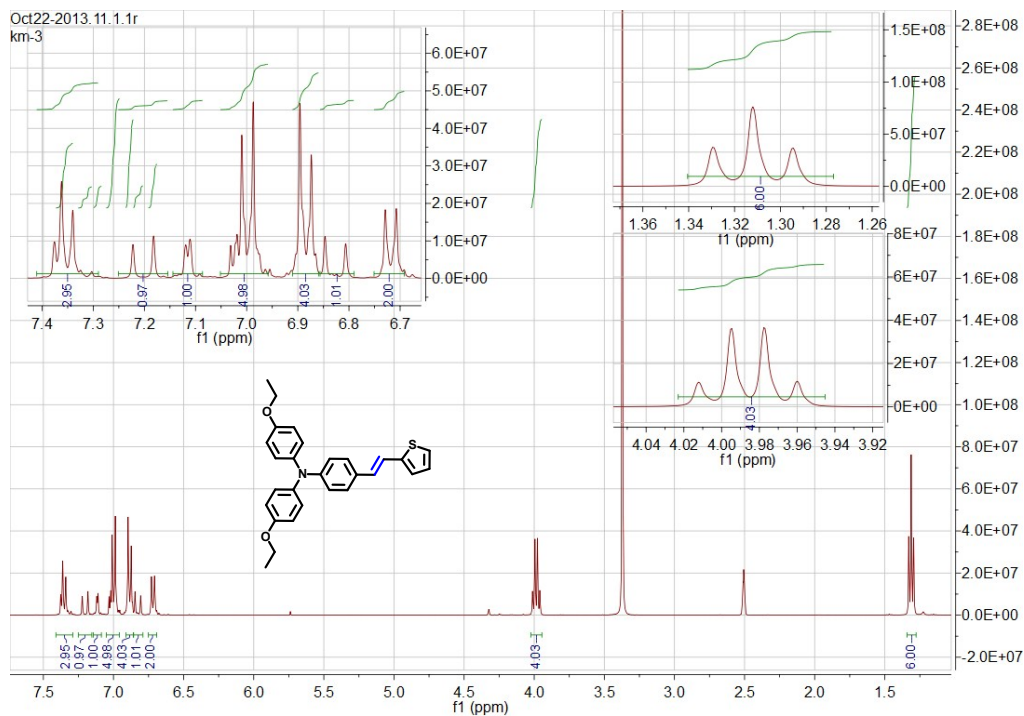


Figure S5. ¹H NMR of X3

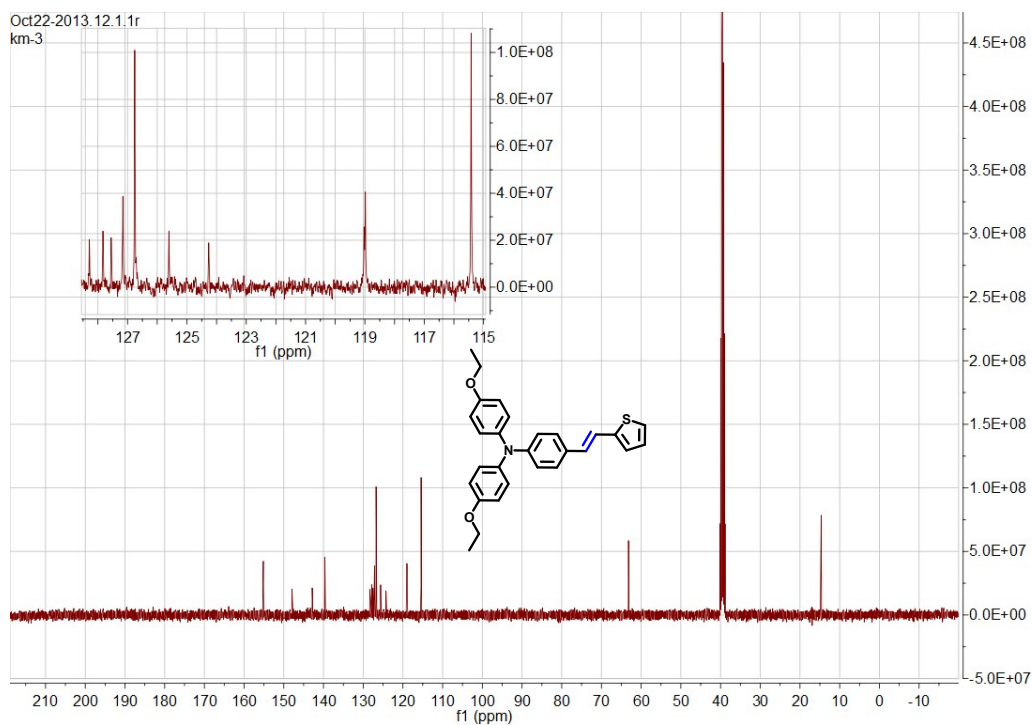
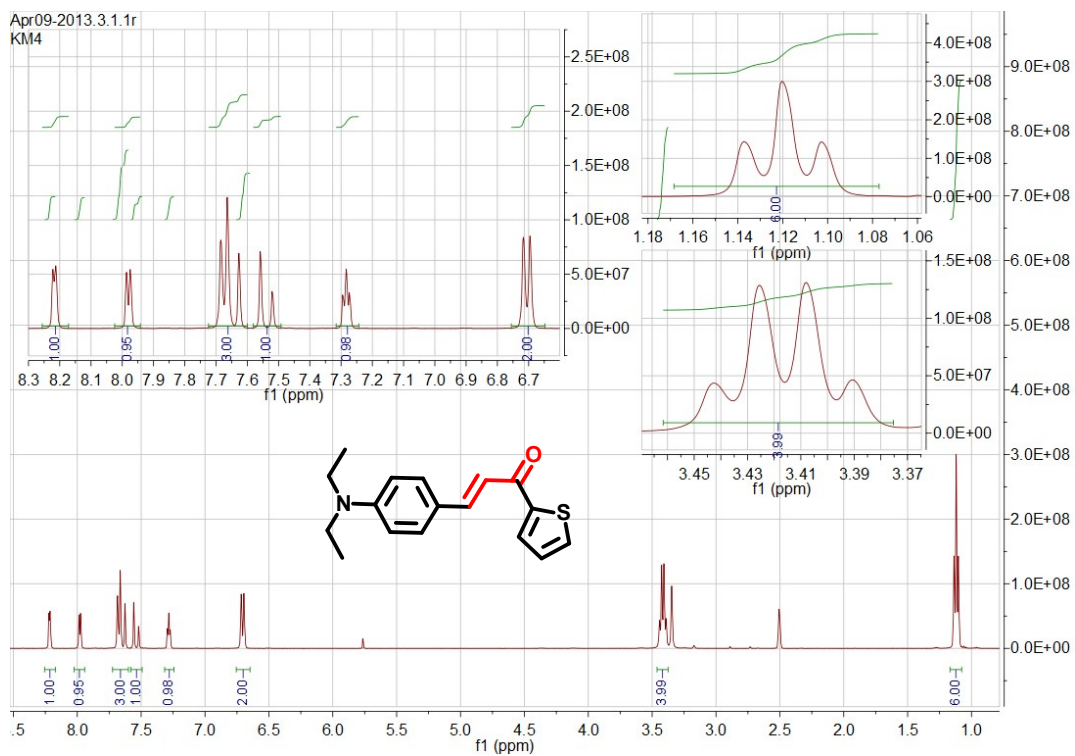
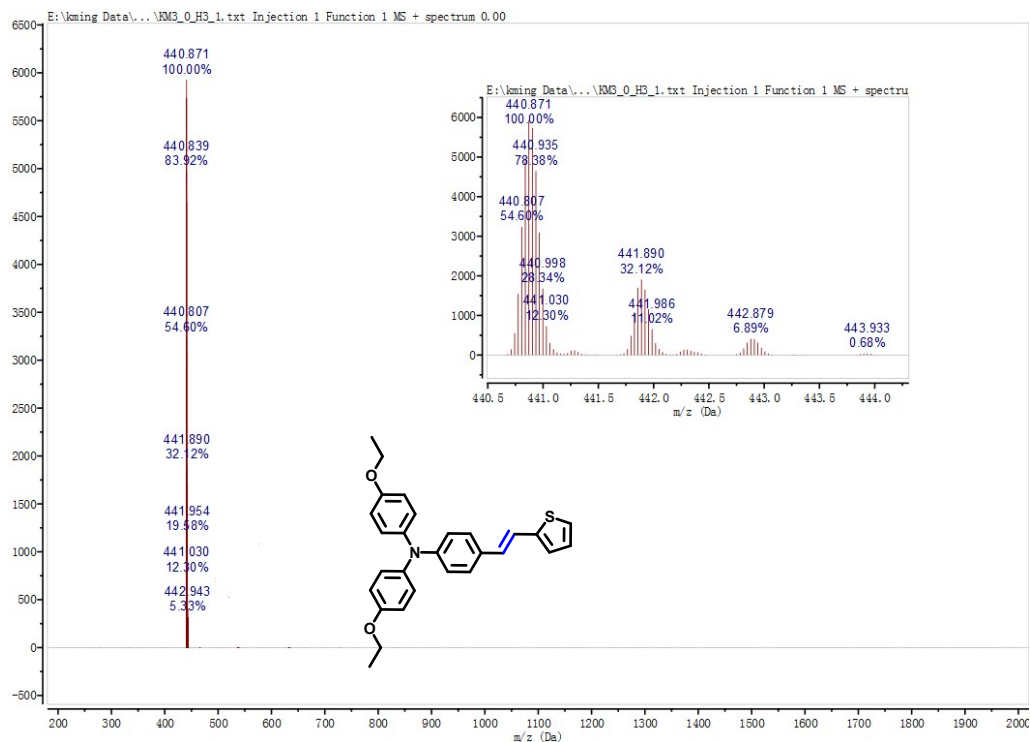


Figure S6. ¹³C NMR of X3



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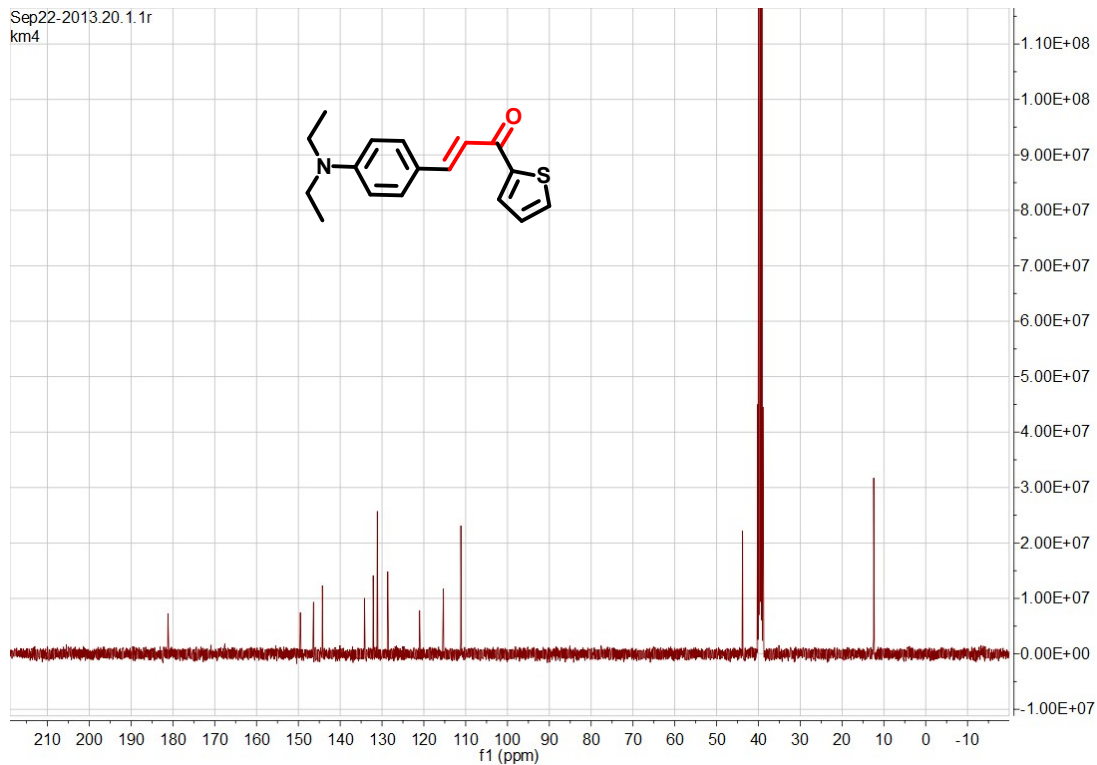


Figure S9. ^{13}C NMR of XT1

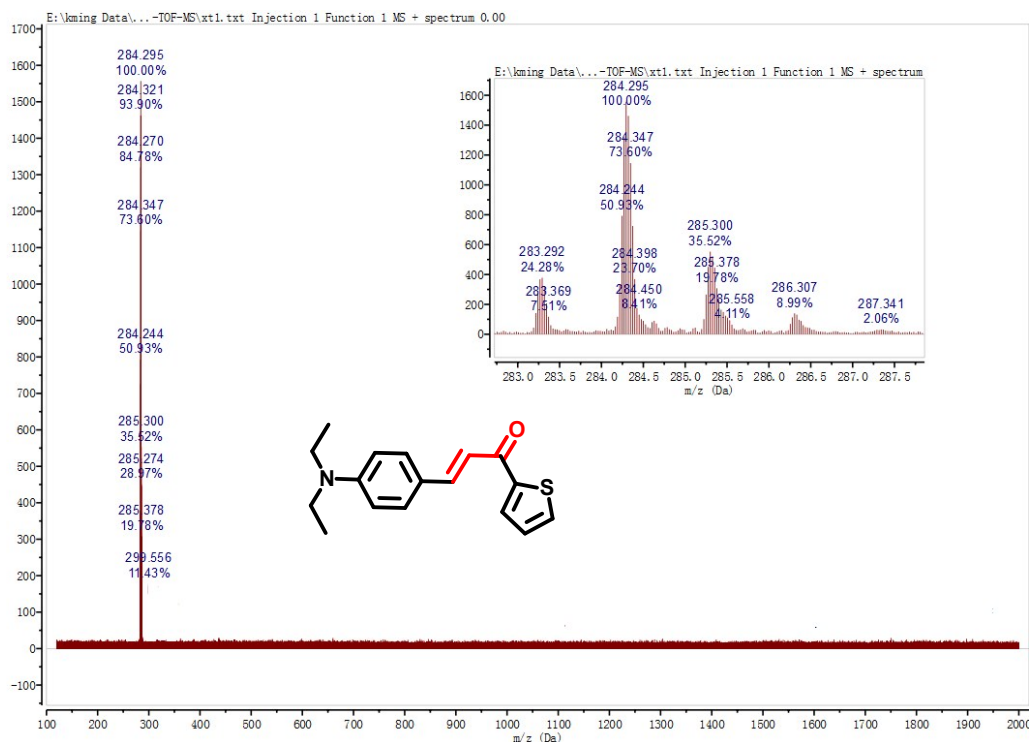
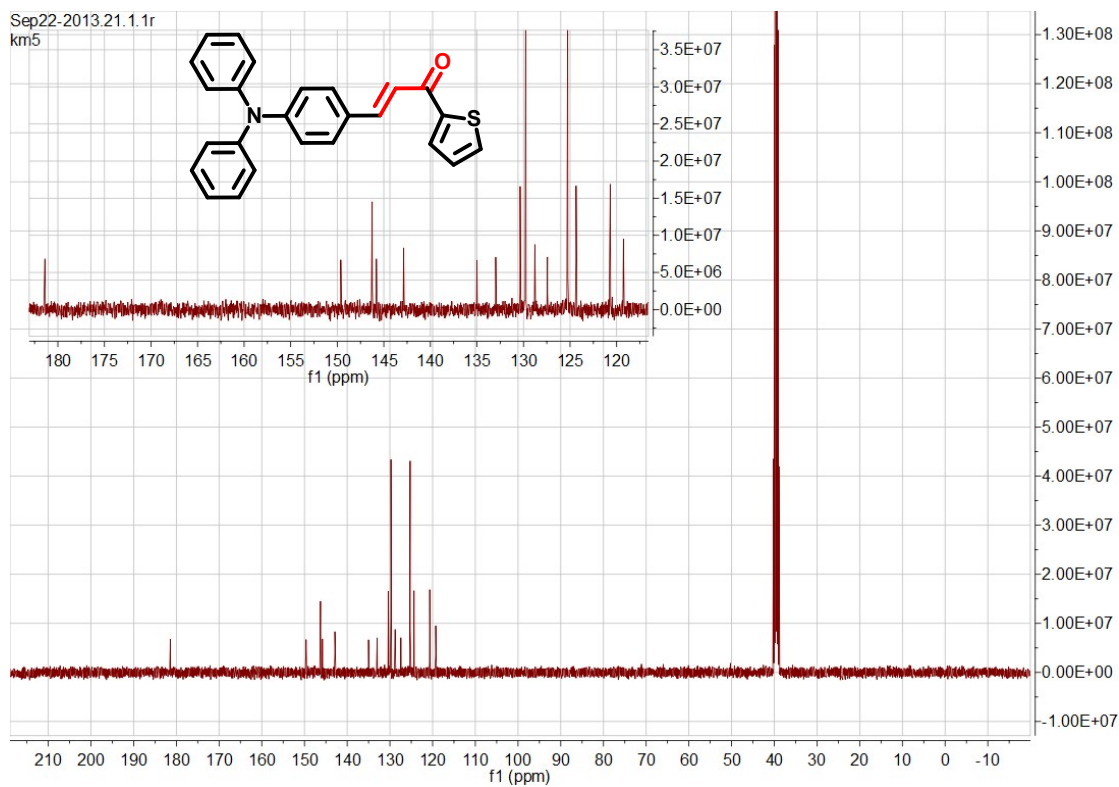
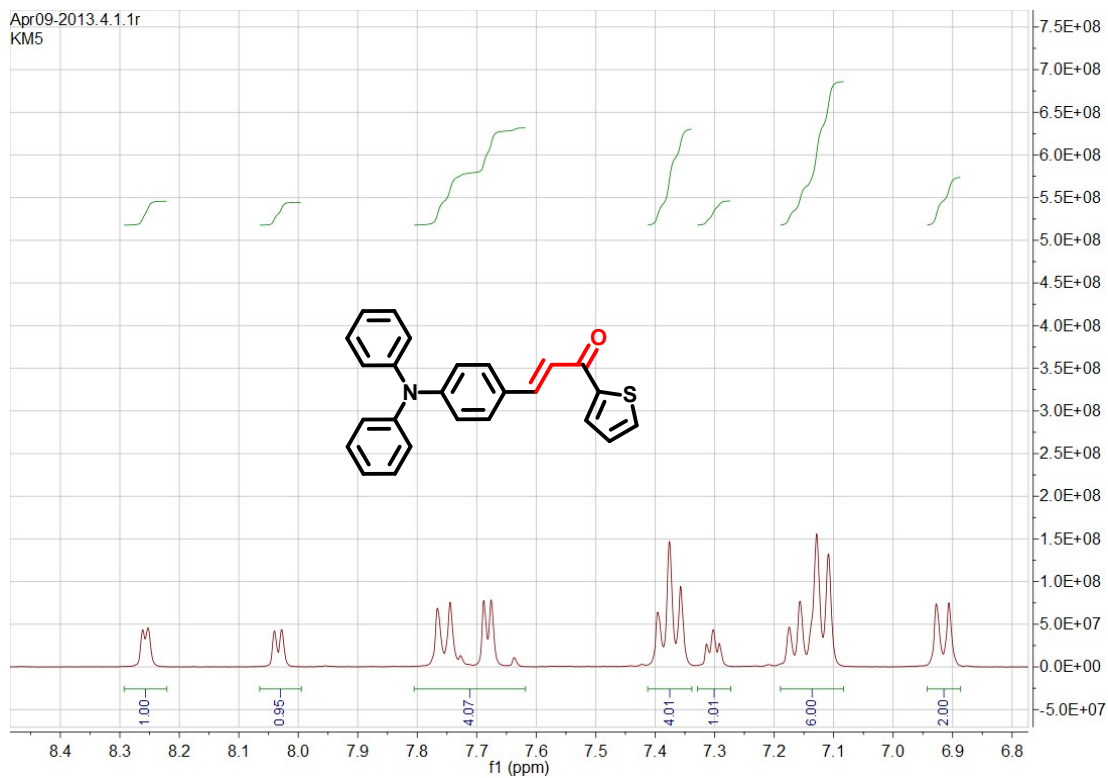


Figure S10. MALDI-TOF-MS of XT1



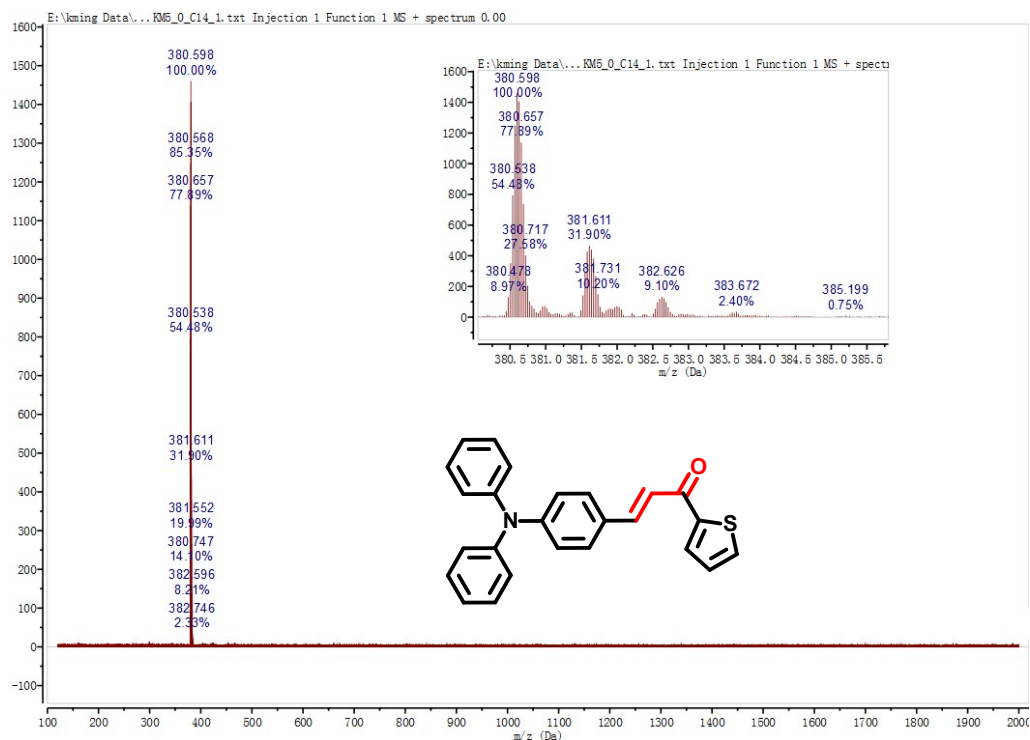


Figure S13. MALDI-TOF-MS of XT2

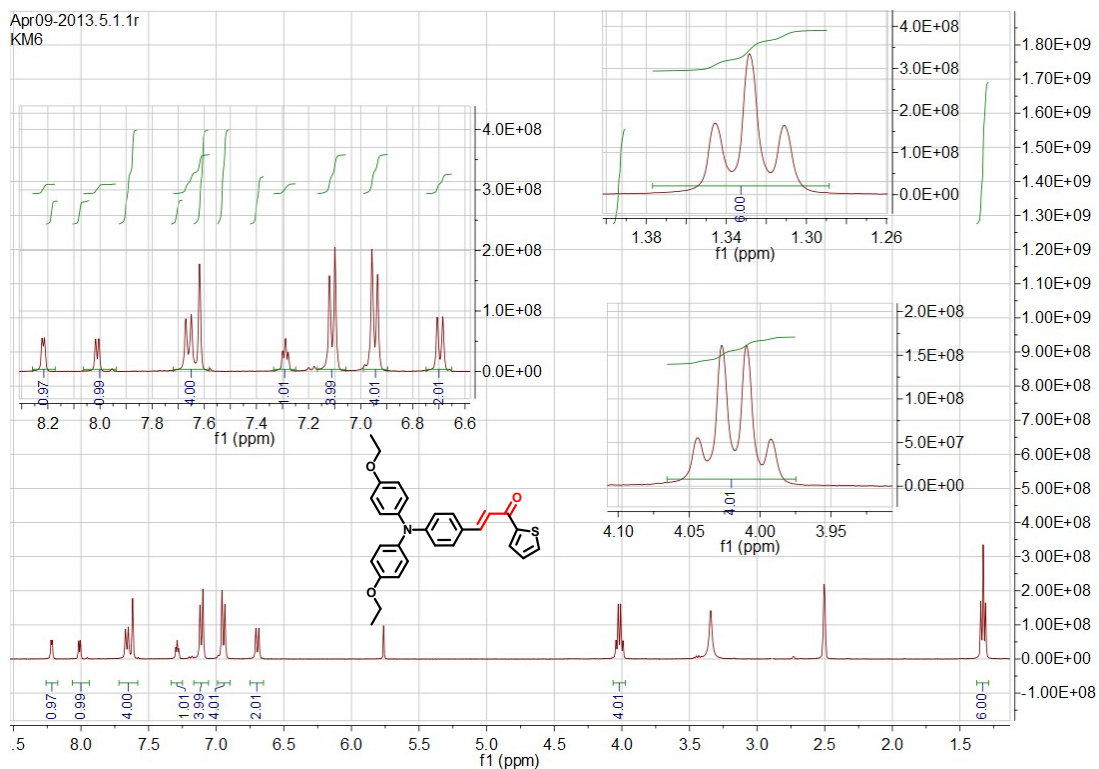
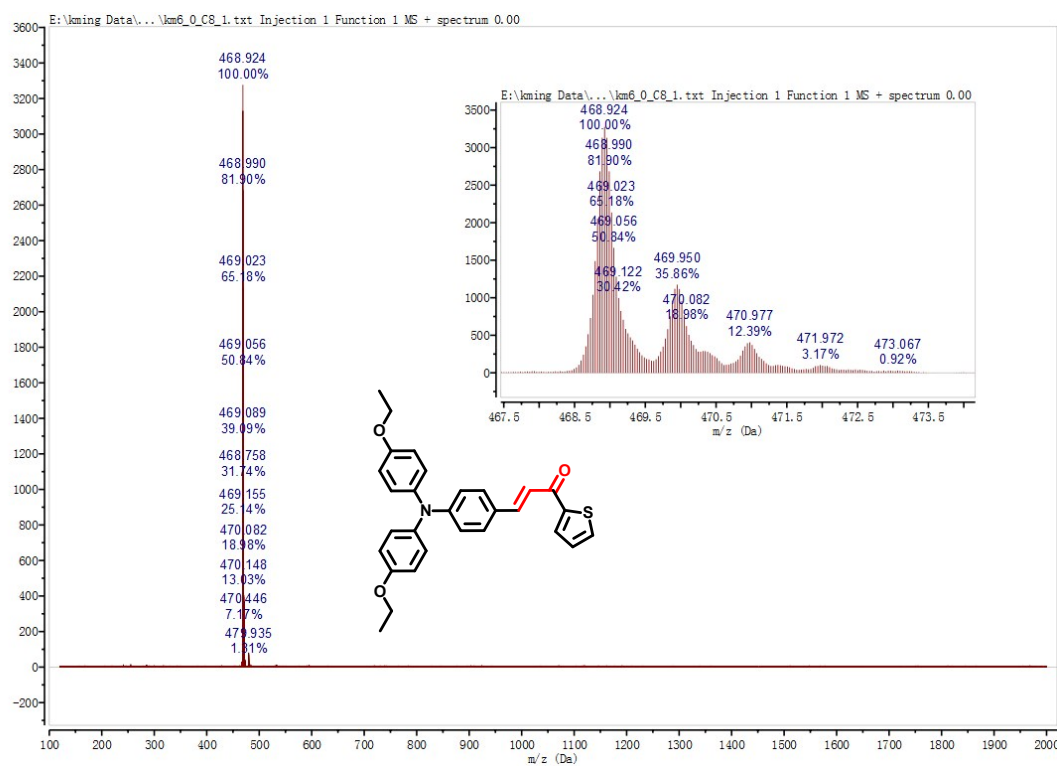
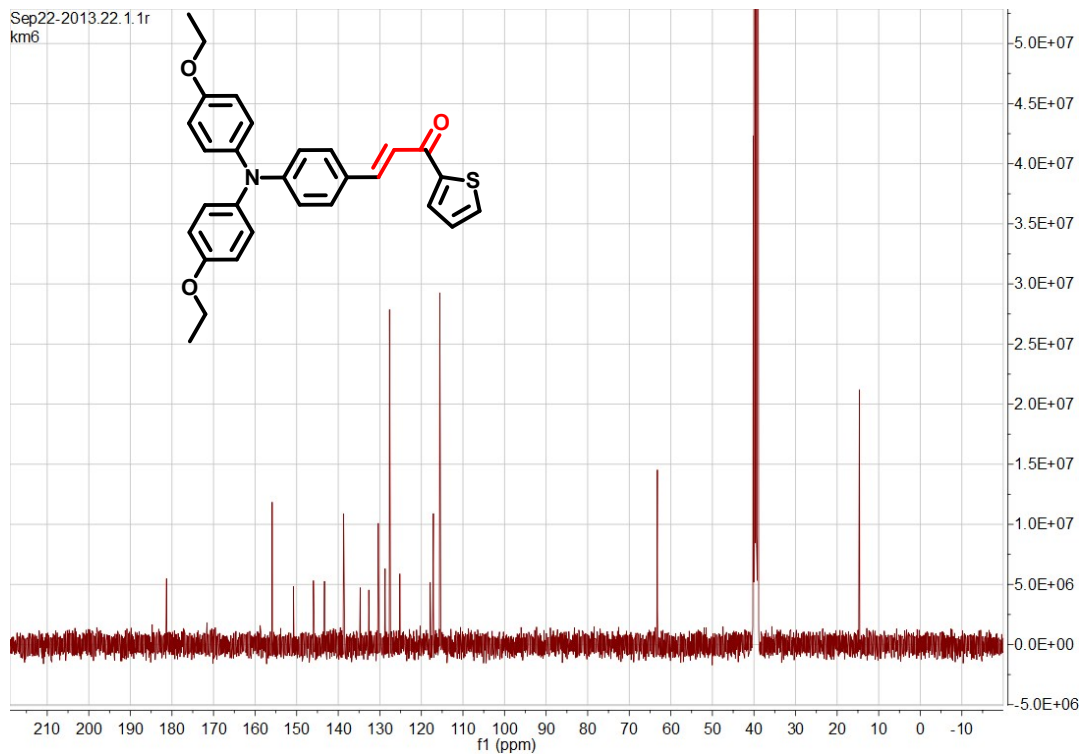


Figure S14. ¹H NMR of XT3



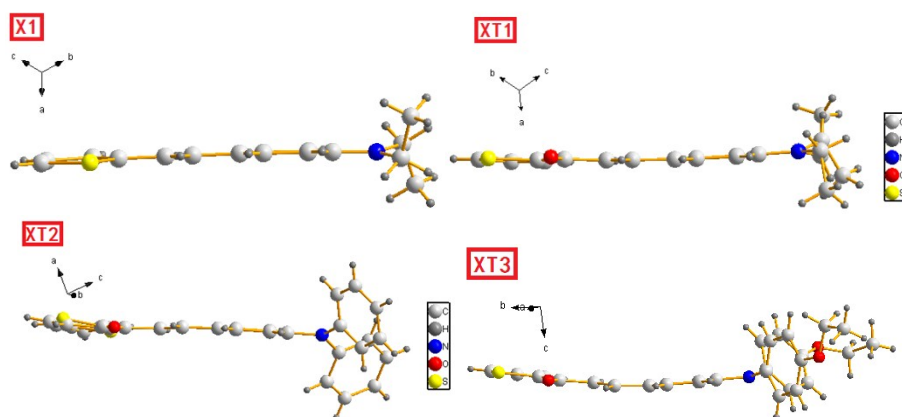


Figure S17. Alternative views of **X1**, **XT1**, **XT2** and **XT3** showing the quasi-planarity of terminal moieties.

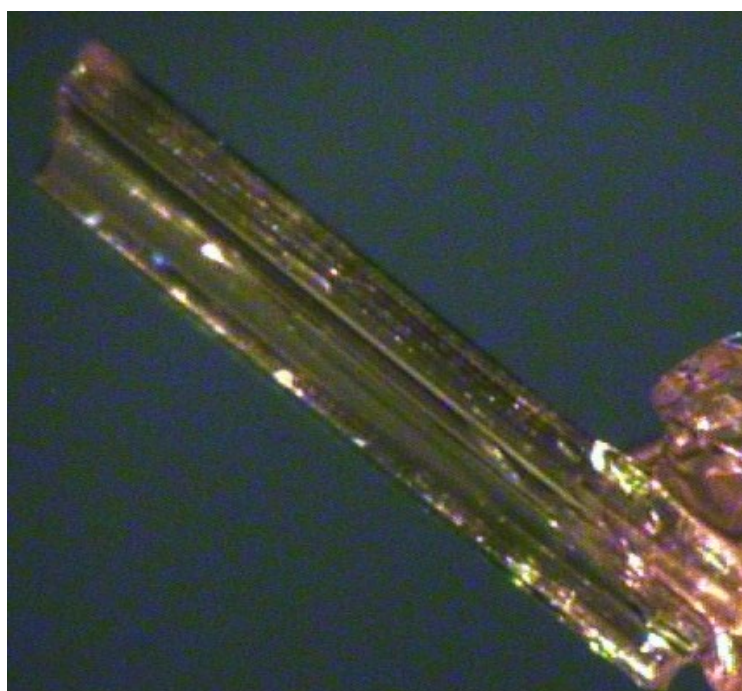


Figure S18. The crystal photograph of **XT1**

Table S1 Selected bond lengths (Å) and angles (°) for **X1**

C(1)-H(1A)	0.9600	C(5)-N(1)-C(2)	121.6(2)
C(2)-H(2A)	0.9700	C(5)-N(1)-C(4)	122.6(2)
C(2)-C(1)	1.511(4)	C(2)-N(1)-C(4)	115.8(2)
C(4)-C(3)	1.513(5)	C(6)-C(7)-C(10)	122.9(2)
C(4)-H(4A)	0.9700	C(6)-C(7)-H(7)	118.6
C(6)-H(6)	0.9300	C(7)-C(10)-C(9)	115.9(3)
C(10)-C(9)	1.398(4)	C(7)-C(10)-C(11)	118.8(2)
C(10)-C(11)	1.472(4)	C(9)-C(10)-C(11)	125.3(3)
C(11)-C(12)	1.333(4)	C(12)-C(11)-C(10)	126.6(3)
C(12)-C(13)	1.462(4)	C(12)-C(11)-H(11)	116.7
C(12)-H(12)	0.9300	C(10)-C(11)-H(11)	116.7

C(11)-H(11)	0.9300	N(1)-C(4)-C(3)	113.9(3)
C(9)-H(9)	0.9300	N(1)-C(4)-H(4A)	108.8
C(8)-H(8)	0.9300	C(11)-C(12)-C(13)	126.0(3)
C(12)-C(13)-S(1)	124.7(2)	C(11)-C(12)-H(12)	117.0
C(14)-C(13)-C(12)	123.9(3)	C(13)-C(12)-H(12)	117.0

Table S2 Selected $\pi\cdots\pi$, C-H $\cdots\pi$ Interactions and potential hydrogen bonds in **X1**

$\pi\cdots\pi$ interactions			
R(i)-R(j)	Dihedral angle(i,j)($^{\circ}$)	\perp distance between the centroid(\AA)	
R(1)-R(1) ^(I)	47	5.977	
R(1)-R(2) ^(II)	3.07	5.914	
R(1)-R(2) ^(III)	74.61	5.035	
R(2)-R(1) ^(IV)	3.07	5.913	
R(2)-R(1) ^(III)	74.61	4.718	
R(2)-R(2) ^(V)	72	5.023	
hydrogen bond			
	d(H \cdots S)	d(C \cdots S)	\angle CHS($^{\circ}$)
C(11)-H(11) \cdots S(1)	2.85	3.248	107
C-H$\cdots\pi$ Interactions			
	H \cdots R(2)(\AA)	\angle C-H \cdots R(2)($^{\circ}$)	C \cdots R(2)(\AA)
C(14)-H(14) \rightarrow R(2)(VI)	2.69	153	3.563
Symmetry code:(I)1-X,1/2+Y,5/2-Z;(II)X,-1+Y,Z;(III)1/2+X,-1/2-Y,2-Z;(IV)X,1+Y,Z;(V)1/2+X,1/2-Y,2-Z;(VI):-1/2+X,-1/2-Y,2-Z;R(1):S(1)-C(13)-C(14)-C(15)-C(16);R(2):C(5)-C(6)-C(7)-C(10)-C(9)-C(8)			

Table S3 Selected bond lengths (\AA) and angles ($^{\circ}$) for **XT1**

C(4)-C(5)	1.469(5)	C(5)-C(4)-S(1)	118.9(2)
C(5)-O(1)	1.232(4)	O(1)-C(5)-C(6)	122.3(3)
C(5)-C(6)	1.449(5)	O(1)-C(5)-C(4)	119.5(3)
C(6)-C(7)	1.330(5)	C(6)-C(5)-C(4)	118.2(3)
C(7)-C(8)	1.450(5)	C(7)-C(6)-C(5)	121.2(3)
C(6)-H(6)	0.9300	C(7)-C(6)-H(6)	119.4
C(7)-H(7)	0.9300	C(5)-C(6)-H(6)	119.4
C(8)-C(9)	1.385(4)	C(6)-C(7)-C(8)	129.2(3)
C(9)-C(10)	1.370(5)	C(6)-C(7)-H(7)	115.4
C(9)-H(9)	0.9300	C(8)-C(7)-H(7)	115.4
C(10)-C(11)	1.406(5)	C(9)-C(8)-C(7)	124.3(3)
C(11)-N(1)	1.370(5)	C(13)-C(8)-C(7)	119.2(3)
C(16)-N(1)	1.453(4)	C(10)-C(9)-C(8)	122.2(3)
C(16)-C(17)	1.490(6)	C(8)-C(9)-H(9)	118.9

Table S4 Selected $\pi\cdots\pi$, C-H $\cdots\pi$ Interactions and potential hydrogen bonds in **XT1**

$\pi\cdots\pi$ interactions			
R(i)-R(j)	Dihedral angle(i,j)(°)	\perp distance between the centroid(Å)	
R(1)-R(1) ^(I)	67	5.079	
R(1)-R(2) ^(II)	89.98	5.236	
R(2)-R(1) ^(III)	58.31	4.782	
R(2)-R(1) ^(IV)	89.98	5.236	
hydrogen bonds			
	d(H \cdots O)	d(C \cdots O)	\angle CHS(°)
C(7)-H(7) \cdots O(1)	2.45	2.799	102
C(16)-H(16B) \cdots O(1)	2.55	3.497	166
C-H$\cdots\pi$ Interactions			
	H \cdots R(2)(Å)	\angle C-H \cdots R(2)(°)	C \cdots R(2)(Å)
C(15 [*])-H(15E) \rightarrow R(2)(V)	2.89	128	3.559

Symmetry code:(I)-1/2+X,Y,1/2-Z;(II)1/2-X,1/2+Y,Z;(III)1-X,-1/2+Y,1/2-Z;(IV) 1/2-X,-1/2+Y,Z;(V)1/2+X,1/2-Y,1-Z;S(1)-C(1)-C(2)-C(3)-C(4);R(2) :C(8)-C(9)-C(10)-C(11)-C(12)-C(13)

Table S5 Selected bond lengths (Å) and angles (°) for **XT2**

C(1)-C(6)	1.378(2)	C(20)-H(20)	0.9300
C(1)-C(2)	1.381(3)	C(20)-C(21)	1.469(2)
C(1)-N(1)	1.423(2)	C(21)-O(1)	1.2273(19)
C(7)-N(1)	1.429(2)	C(21)-C(22)	1.505(18)
C(2)-C(3)	1.384(3)	C(23)-C(22)-S(1)	113.6(13)
C(13)-C(14)	1.394(2)	C(23)-C(22)-C(21)	121.3(12)
C(14)-C(15)	1.376(2)	C(21)-C(22)-S(1)	125.0(10)
C(15)-C(16)	1.393(2)	O(1)-C(21)-C(22)	119.2(6)
C(14)-H(14)	0.9300	O(1)-C(21)-C(20)	121.87(15)
C(15)-H(15)	0.9300	C(21)-C(20)-H(20)	119.7
C(16)-C(19)	1.457(2)	C(19)-C(20)-C(21)	120.55(15)
C(19)-H(19)	0.9300	C(20)-C(19)-H(19)	115.3
C(19)-C(20)	1.329(2)	C(20)-C(19)-C(16)	129.35(15)
C(17)-C(16)-C(19)	123.96(14)	C(15)-C(16)-C(19)	118.71(14)
		C(16)-C(19)-H(19)	115.3

Table S6 Selected $\pi\cdots\pi$, C-H $\cdots\pi$ Interactions and potential hydrogen bonds in **XT2**

$\pi\cdots\pi$ interactions			
R(i)-R(j)	Dihedral angle(i,j)(°)	\perp distance between the centroid(Å)	
R(1)-R(1) ^(I)	0	4.976	
R(1)-R(2) ^(II)	5.5	4.874	
R(1)-R(3) ^(III)	63.4	4.973	
R(1)-R(5) ^(III)	5.0	5.451	
R(1)-R(5) ^(III)	5.0	4.851	
R(1)-R(5) ^(I)	5.0	5.492	
R(2)-R(2) ^(I)	0	4.77	
R(2)-R(5) ^(III)	9.4	4.815	
R(4)-R(4) ^(IV)	0	5.333	
R(5)-R(4) ^(V)	61.76	5.134	

hydrogen bonds			
D-H \cdots A	d(H \cdots A)	d(D \cdots A)	\angle DHA(°)
C(12)-H(12) \cdots O(1)	2.56	3.489	173
C(19)-H(19) \cdots O(1)	2.43	2.795	103
C(20)-H(20) \cdots S(1)	2.79	3.175	106
C(25)-H(25) \cdots O(1)	2.39	3.289	162

C-H$\cdots\pi$ Interactions			
	H \cdots R(4)(Å)	\angle C-H \cdots R(4)(°)	C \cdots R(2)(Å)
C(5)-H(5) \rightarrow R(4)(VI)	2.79	149	3.613

Symmetry code:(I)2-X,-1-Y,1-Z;(II)1-X,-Y,1-Z;(III)1-X,-1-Y,1-Z;(IV)-X,-Y,2-Z;(V)1+X,Y,Z;(VI) X,1+Y,Z;R(1) S(1)-C(22)-C(23)-C(24)-C(25);R(2) : S(1') C(22') C(23') C(24') C(25');R(3): C(1)-C(2)-C(3)-C(4)-C(5)-C(6);R(4):C(7)-C(8)-C(9)-C(10)-C(11)-C(12);R(5):C(13)-C(14)-C(15)-C(16)-C(17)-C(18);

Table S7 Selected bond lengths (Å) and angles (°) for **XT3**

S(1)-C(26)	1.707(4)	C(27)-C(26)-S(1)	110.1(3)
C(26)-C(27)	1.358(5)	C(25)-C(26)-S(1)	118.9(3)
C(26)-C(25)	1.454(5)	C(27)-C(26)-C(25)	131.0(4)
C(25)-O(3)	1.230(4)	O(3)-C(25)-C(26)	120.5(4)
C(25)-C(24)	1.474(5)	C(26)-C(25)-C(24)	117.4(4)
C(23)-C(24)	1.323(5)	O(3)-C(25)-C(24)	122.1(4)
C(23)-C(22)	1.448(5)	C(25)-C(24)-H(24)	118.9
C(24)-H(24)	0.9300	C(23)-C(24)-H(24)	118.9
C(23)-H(23)	0.9300	C(24)-C(23)-H(23)	115.9
N(1)-C(17)	1.382(5)	C(22)-C(23)-H(23)	115.9
N(1)-C(16)	1.440(5)	C(21)-C(22)-C(23)	122.1(4)
N(1)-C(8)	1.433(5)	C(19)-C(22)-C(23)	121.5(4)
C(24)-C(23)-C(22)	128.2(4)	C(23)-C(24)-C(25)	122.2(4)

Table S8 Selected $\pi\cdots\pi$, C-H $\cdots\pi$ Interactions and potential hydrogen bonds in **XT3**

$\pi\cdots\pi$ interactions			
R(i)-R(j)	Dihedral angle(i,j)(°)	\perp distance between the centroid(Å)	
R(1)-R(2) ^(I)	57.4	5.358	
R(1)-R(4) ^(II)	14.1	4.208	
R(1)-R(5) ^(III)	15.7	5.656	
R(5)-R(5) ^(IV)	0	5.273	
R(6)-R(6) ^(V)	0	5.424	

hydrogen bonds			
D-H \cdots A	d(H \cdots A)	d(D \cdots A)	\angle DHA(°)
C(23)-H(23) \cdots O(3)	2.50	2.835	101
C(43)-H(43) \cdots O(1)	2.58	3.410	149
C(52)-H(52) \cdots O(6)	2.48	2.821	102

C-H$\cdots\pi$ Interactions			
	H \cdots R(Å)	\angle C-H \cdots R(°)	C \cdots R(Å)
C(1)-H(1C) \rightarrow R(1)	2.90	163	3.828
C(15)-H(15) \rightarrow R(8)	2.97	157	3.848
C(36)-H(36) \rightarrow R(4)	2.95	151	3.791

Symmetry code:(I)3/2-X,-1/2+Y,3/2-Z;(II)1-X,-Y,1-Z;(III)-1/2+X,1/2-Y,1/2+Z;(IV) 1-X,1-Y,2-Z;(V)1-X,-Y,2-Z;R(1):S(1)-C(26)-C(27)-C(28)-C(29);R(2):C(3)-C(4)-C(5)-C(8)-C(7)-C(6);R(3):C(11)-C(12)-C(13)-C(16)-C(15)-C(14);R(4):C(17)-C(18)-C(19)-C(22)-C(21)-C(20);R(5): S(2)-C(55)-C(56)-C(57)-C(58);R(6) C(32)-C(33)-C(34)-C(37)-C(36)-C(35);R(8) C(46)-C(47)-C(48)-C(51)-C(50)-C(49).

Table S9 Single-photon-related photophysical properties of six dyes in different solvents.

Compounds	Solvents	λ_{\max}^a (ϵ_{\max}^b)	λ_{\max}^c	$\Delta\nu^d$	Φ^e	τ/ns
X1	Benzene	360(1.84)	419	59	0.023	0.59
	Dichloromethane	361(1.74)	443	82	0.017	0.37
	Tetrahydrofuran	362(1.77)	433	71	0.030	~
	Ethyl acetate	357(1.72)	432	75	0.023	0.42
	Ethanol	353(1.61)	442	89	0.040	0.37
	Acetonitrile	357(1.67)	451	94	0.017	0.38
	DMF	363(1.09)	453	90	0.029	~
X2	Benzene	302(2.43) 373(2.25)	425	52	0.240	1.42
	Dichloromethane	303(2.26) 374(2.82)	453	79	0.265	2.33
	Tetrahydrofuran	302(2.40) 372(2.78)	440	68	0.260	~
	Ethyl acetate	301(1.94) 368(2.05)	438	70	0.225	1.78
	Ethanol	301(1.79) 368(1.97)	445	77	0.326	2.12
	Acetonitrile	301(2.17) 369(2.88)	462	93	0.218	2.80
	DMF	303(1.68) 373(2.57)	461	88	0.263	~
	Benzene	303(1.70) 388(2.59)	463	75	0.147	2.89
	Dichloromethane	302(1.81) 387(2.99)	511	124	0.077	4.65

X3	Tetrahydrofuran	301(1.71)	386(2.76)	491	105	0.116	~
	Ethyl acetate	300(1.95)	382(3.16)	490	108	0.095	3.83
	Ethanol	301(1.62)	382(2.72)	506	124	0.039	2.75
	Acetonitrile	300(1.69)	381(3.11)	521	140	0.018	2.31
	DMF	301(1.57)	385(3.10)	520	135	0.034	~
XT1	Benzene	282(2.22)	428(3.20)	480	52	0.016	0.65
	Dichloromethane	287(1.76)	435(3.75)	525	90	0.055	2.62
	Tetrahydrofuran	285(1.17)	425(2.57)	508	83	0.069	~
	Ethyl acetate	283(1.66)	421(3.79)	518	97	0.036	1.96
	Ethanol	280(1.59)	437(4.06)	550	113	0.002	0.15
	Acetonitrile	280(1.24)	429(3.46)	539	110	0.018	0.74
	DMF	286(1.26)	433(3.92)	534	101	0.038	~
XT2	Benzene	300(2.95)	422(2.95)	497	75	0.091	2.62
	Dichloromethane	300(3.74)	426(4.53)	557	131	0.022	3.74
	Tetrahydrofuran	299(2.76)	416(3.21)	523	107	0.073	~
	Ethyl acetate	298(2.70)	413(3.36)	530	117	0.027	3.02
	Ethanol	298(2.59)	425(3.23)	533	108	0.0006	0.14
	Acetonitrile	297(2.73)	417(3.53)	575	158	0.002	0.62
	DMF	300(2.45)	421(3.28)	564	143	0.006	~
XT3	Benzene	299(2.97)	435(3.21)	546	111	0.020	2.27
	Dichloromethane	299(2.98)	439(3.64)	—	—	—	~
	Tetrahydrofuran	295(2.85)	426(3.38)	574	148	0.003	~
	Ethyl acetate	295(2.58)	426(3.19)	576	150	0.0003	0.51
	Ethanol	296(2.41)	441(3.08)	—	—	—	~
	Acetonitrile	295(2.81)	428(3.85)	—	—	—	~
	DMF	299(2.01)	433(2.97)	—	—	—	~

^a Absorption peak position in nm. ^b Maximum molar extinction coefficient in $10^4 \text{ mol}^{-1} \text{ L cm}^{-1}$. ^c Peak position of SPEF, excited at the maximum wavelength of absorption. ^d Stokes shift in nm. ^e Quantum yields determined by using quinine sulfate as standard.

—Fluorescence quenching.

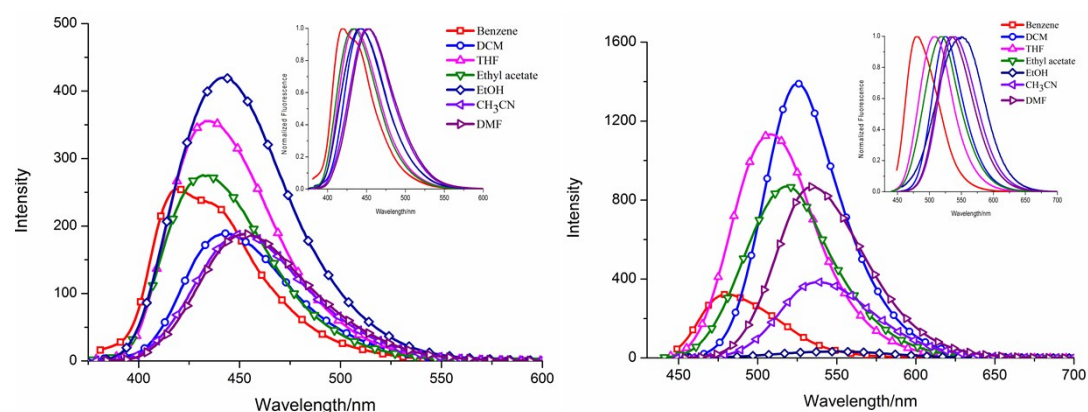


Figure S19. Fluorescence spectra of dyes **X1**(left) and **XT1**(right) in different solvents of different polarities with a concentration of $1.0 \times 10^{-5} \text{ mol/L}$.

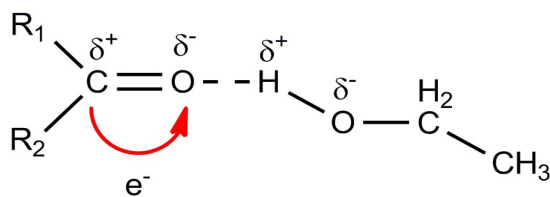


Figure S20. The formation process of hydrogen bond.

Table S10 Selected experimental and calculated optical data for the dyes

Compd.	OI ^[a]	$\Delta E(\text{eV})$ ^[b]	Cal. λ_{max} (nm) ^[c]	Obs. λ_{max} (nm) ^[d]	f ^[e]	Character
X1	HOMO→LUMO	3.4008	365	353	1.0508	ICT/ $\pi \rightarrow \pi^*$
X2	HOMO→LUMO+2	4.0663	305	301	0.1533	$\pi \rightarrow \pi^*$
	HOMO→LUMO	3.2578	381	368	1.1358	ICT/ $\pi \rightarrow \pi^*$
X3	HOMO→LUMO+3	4.2192	294	301	0.0988	$\pi \rightarrow \pi^*$
	HOMO→LUMO	3.3595	369	382	1.2417	ICT/ $\pi \rightarrow \pi^*$
XT1	HOMO→LUMO+1	4.4290	280	280	0.0644	$n \rightarrow \pi^*/\pi \rightarrow \pi^*$
	HOMO→LUMO	2.8728	432	437	0.8660	ICT/ $\pi \rightarrow \pi^*$
XT2	HOMO→LUMO+3	4.2534	291	298	0.1694	$\pi \rightarrow \pi^*$
	HOMO→LUMO	2.7400	452	425	0.8023	ICT/ $\pi \rightarrow \pi^*$
XT3	HOMO→LUMO+3	4.2671	291	296	0.0878	$\pi \rightarrow \pi^*$
	HOMO→LUMO	2.7798	446	441	0.8393	ICT/ $\pi \rightarrow \pi^*$

[a]Orbitals involved in the excitations, [b]Excitation energies (eV), [c]Calculated peak position

of the longest absorption band in ethanol, [d] Observed peak position of the longest absorption

band in ethanol, [f]Oscillator Strengths

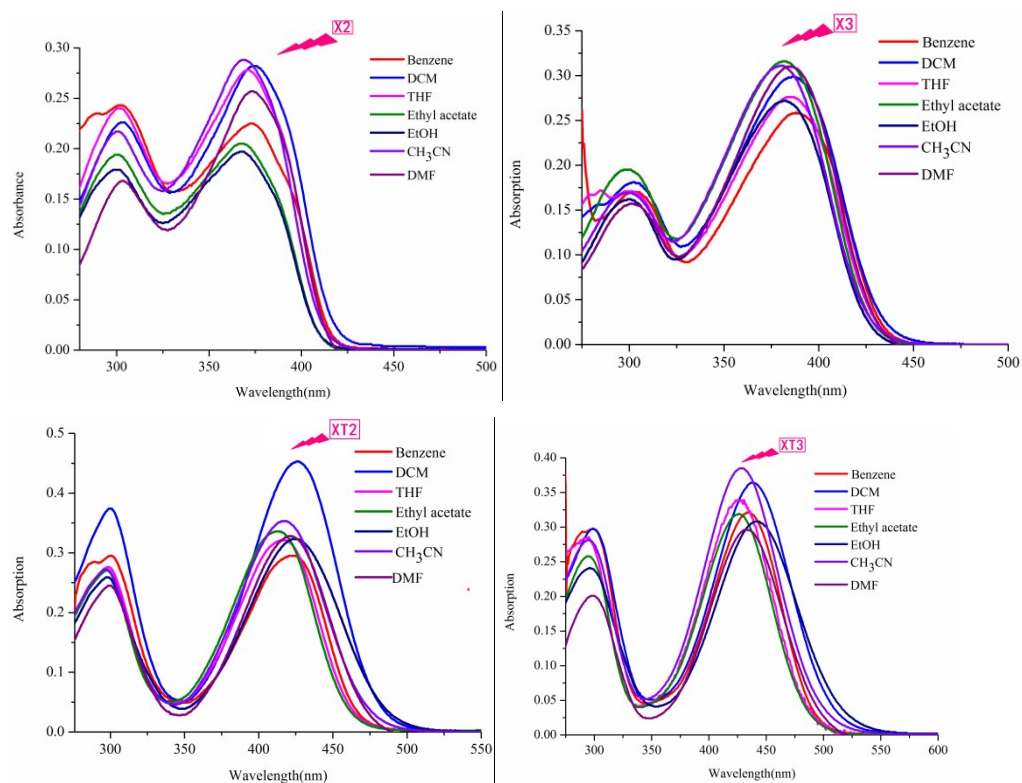


Figure S21. Absorption spectra of dyes **X2**, **X3**, **XT2** and **XT3** in different solvents of different polarities with a concentration of 1.0×10^{-5} mol/L.

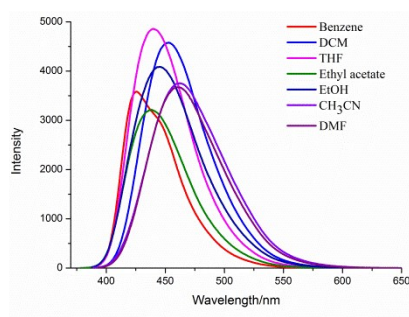


Figure S22. Fluorescence spectra of dye **X2** in different solvents of different polarities with a concentration of 1.0×10^{-5} mol/L.

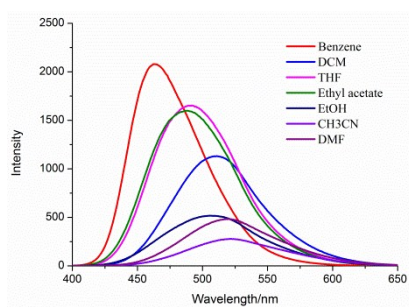


Figure S23. Fluorescence spectra of dye **X3** in different solvents of different polarities with a concentration of 1.0×10^{-5} mol/L

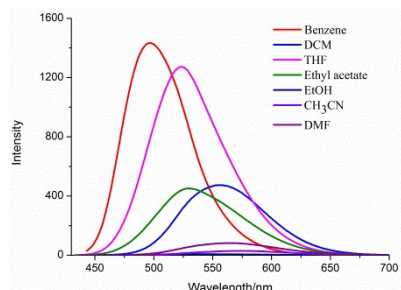


Figure S24. Fluorescence spectra of dye **XT2** in different solvents of different polarities with a concentration of 1.0×10^{-5} mol/L

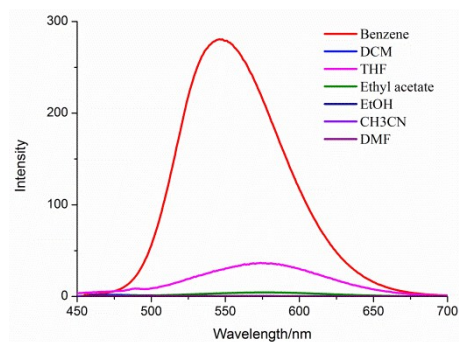
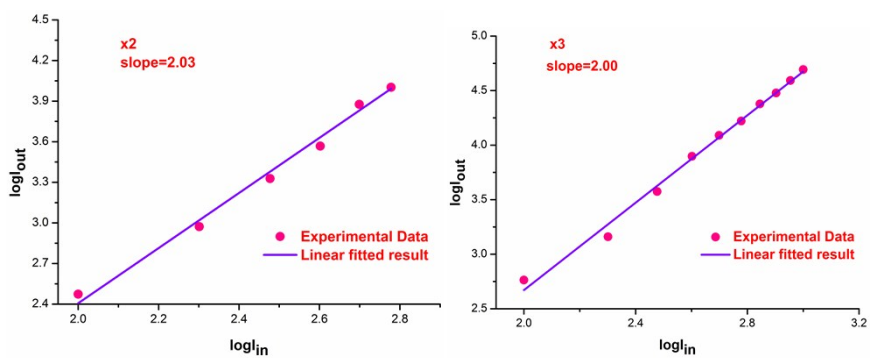


Figure S25. Fluorescence spectra of dye **XT3** in different solvents of different polarities with a concentration of 1.0×10^{-5} mol/L



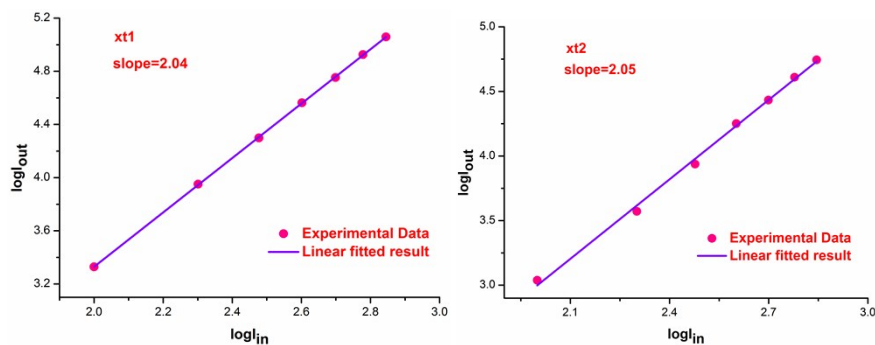


Figure S26. Two-photon absorption verification of X2, X3, XT1 and XT2 which I_{in} and I_{out} represent the input laser power and output fluorescence, respectively.

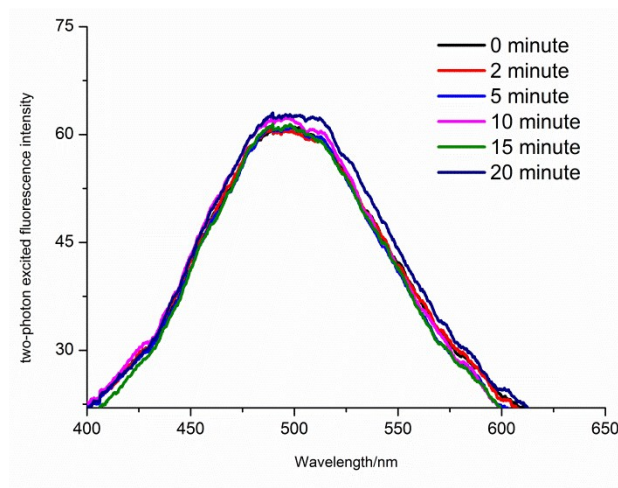


Figure S27. Time-dependent two-photon excited fluorescence of X2 excited at 790 nm in DMF.

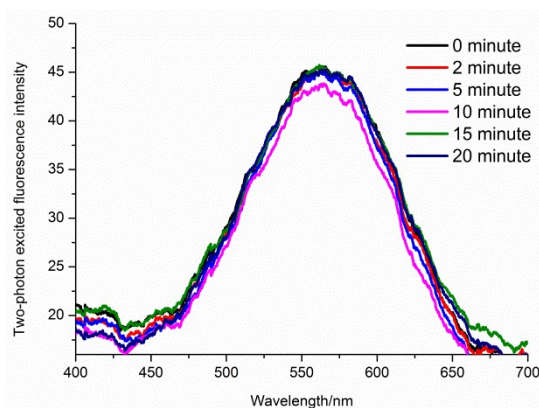


Figure S28. Time-dependent two-photon excited fluorescence of X3 excited at 790 nm in DMF.

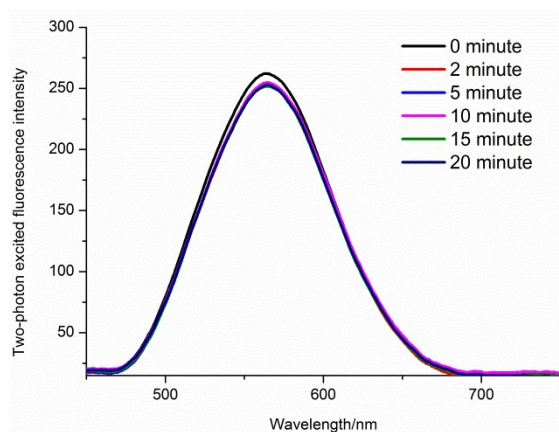


Figure S29. Time-dependent two-photon excited fluorescence of **XT1** excited at 830 nm in DMF.

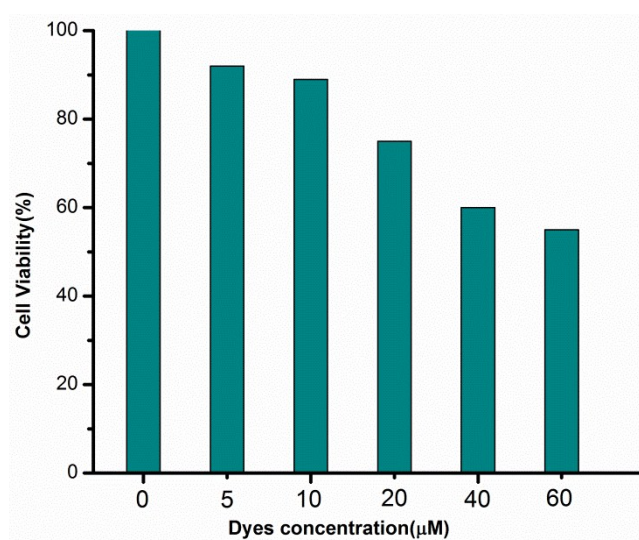


Figure S30. MTT assay of HepG2 cells incubate with dyes **X2** at different concentrations for 24h.

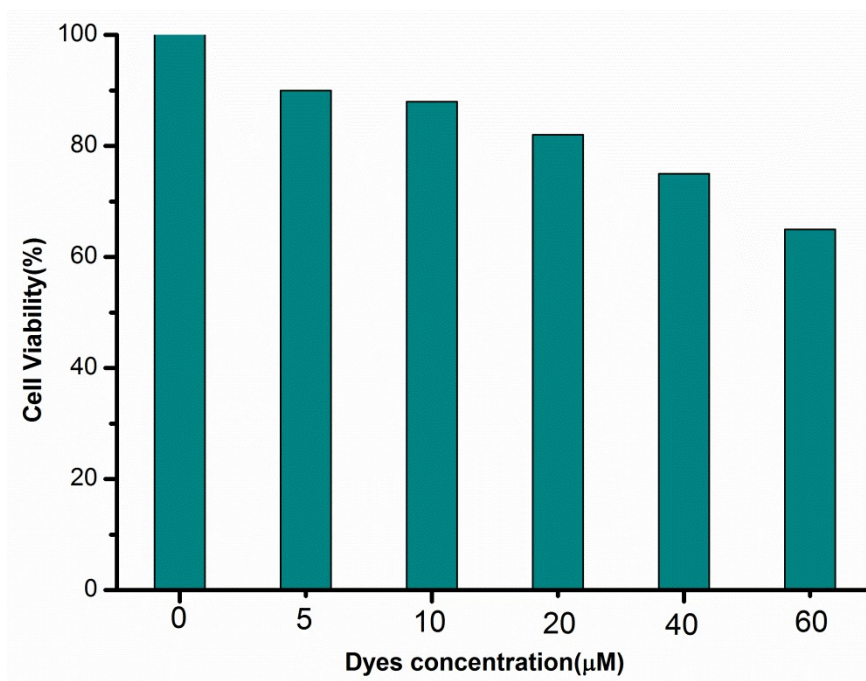


Figure S31. MTT assay of HepG2 cells incubate with dyes **XT1** at different concentrations for 24h.