

Supporting Information

Employment of electrostatic interactions for an amperometric detection of carbon nanoparticles in a FIA system

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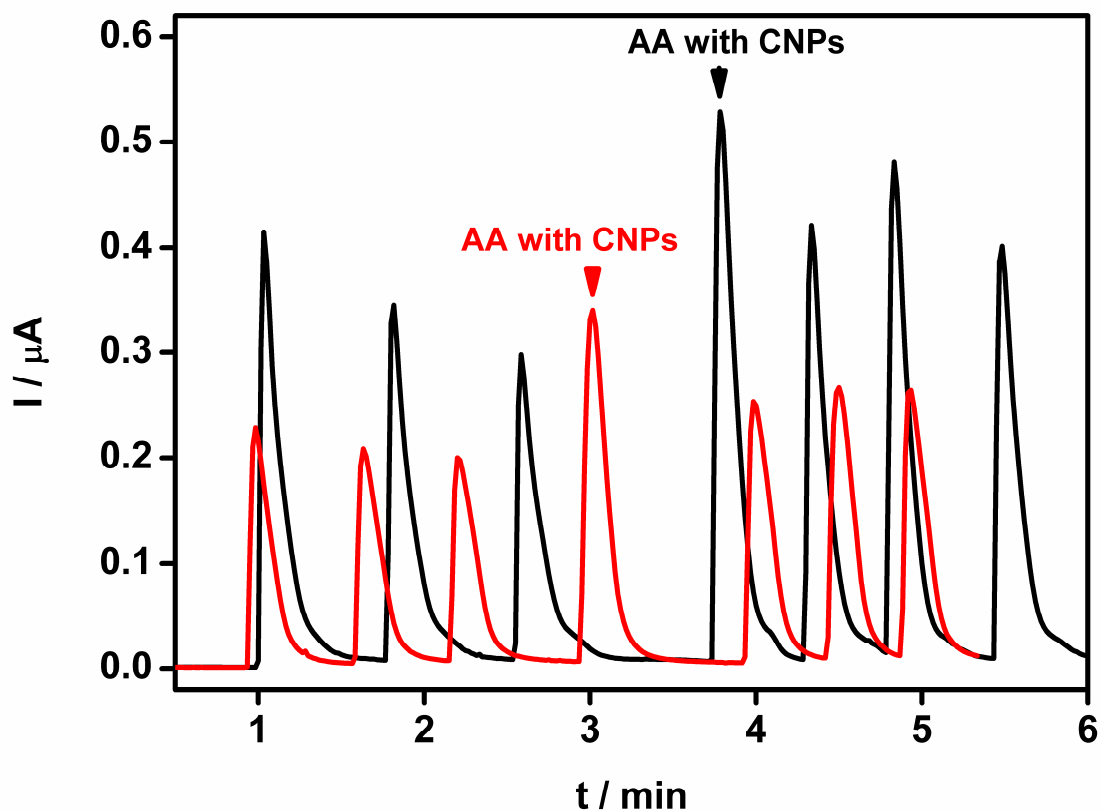


Fig.SI_1 Chronoamperometry of 1 mM AA electrooxidation performed with the use of the FIA system⁹ at 0.2 V in H₂O but not in NaH₂PO₄ with the total flow rate fixed at 3 mL min⁻¹ for samples with or without the addition of 1 mg mL⁻¹ **CNPs aqueous suspension**. The sequence of the two series (black and red) was as follows: (i) three injections of AA and (ii) one injection of AA with CNPs and (iii) three injections of AA.

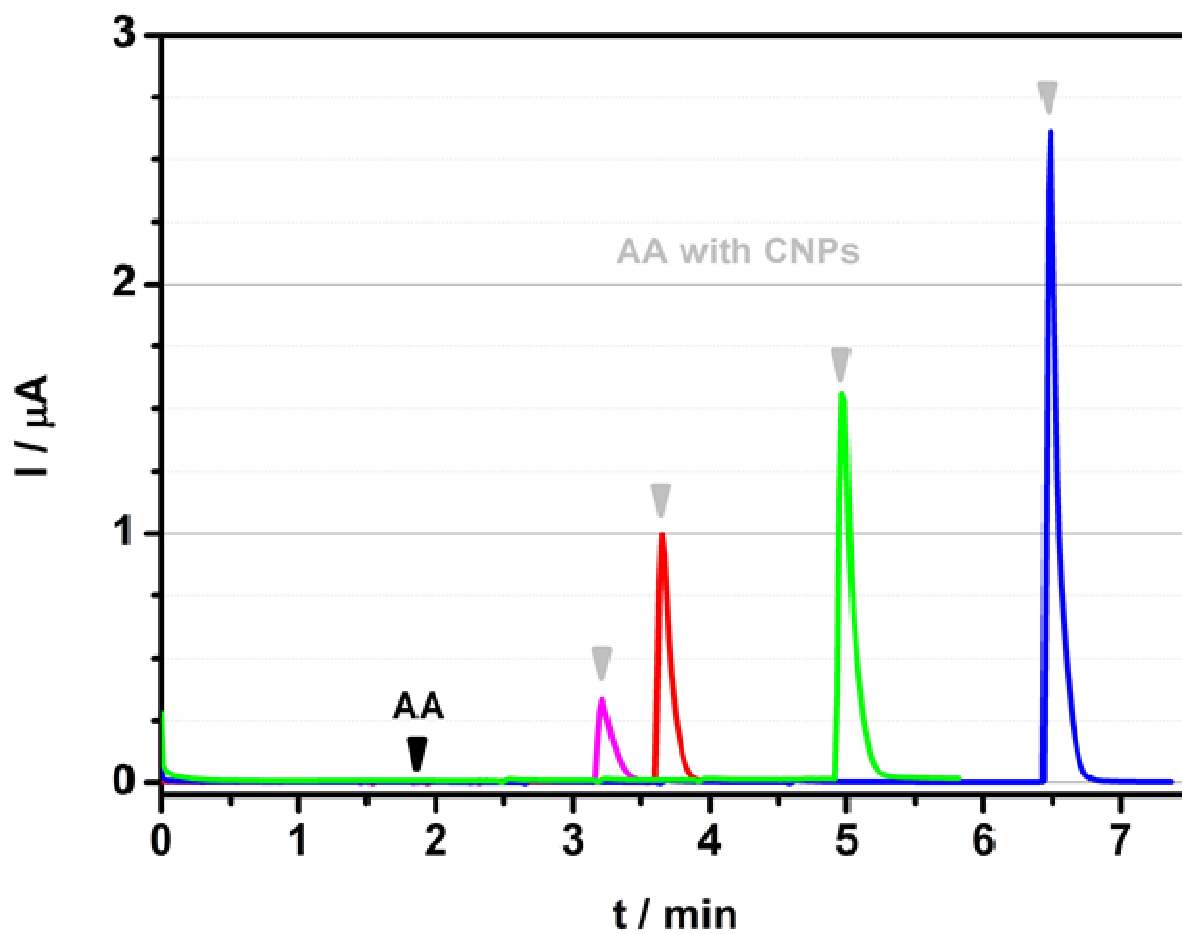


Fig.SI_2 Chronoamperometry of 1 mM AA electrooxidation performed with the use of FIA system⁹ at 0.2 V in 25 mM NaH₂PO₄ with the total flow rate fixed at 3 mL min⁻¹ for samples with or without the addition of 1 mg mL⁻¹ **CNPs aqueous suspension**. The sequence of the four series (pink, red, green and blue) was as follows: (i) three injections of 1 mM AA and (ii) one injection of AA with CNPs.

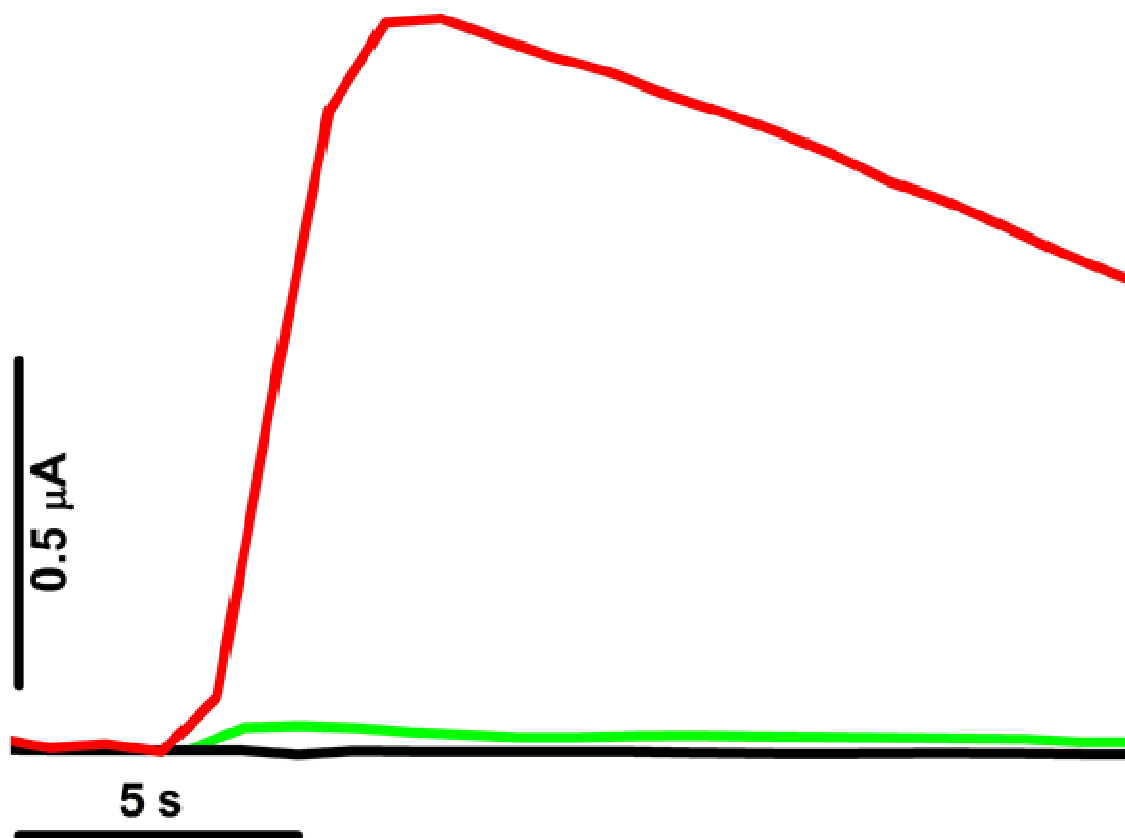


Fig.SI_3 Chronoamperometry of 20 mM deoxygenated solution of AA performed with the use of the FIA system (Fig.7B) at 0.2 V in 200 mM NaH₂PO₄ (pH 4.6) with the total flow rate fixed at 6 mL min⁻¹ for samples with addition of 0.001 (black), 0.01 (green) and 0.1 (red) mg mL⁻¹ CNPs suspension in water.

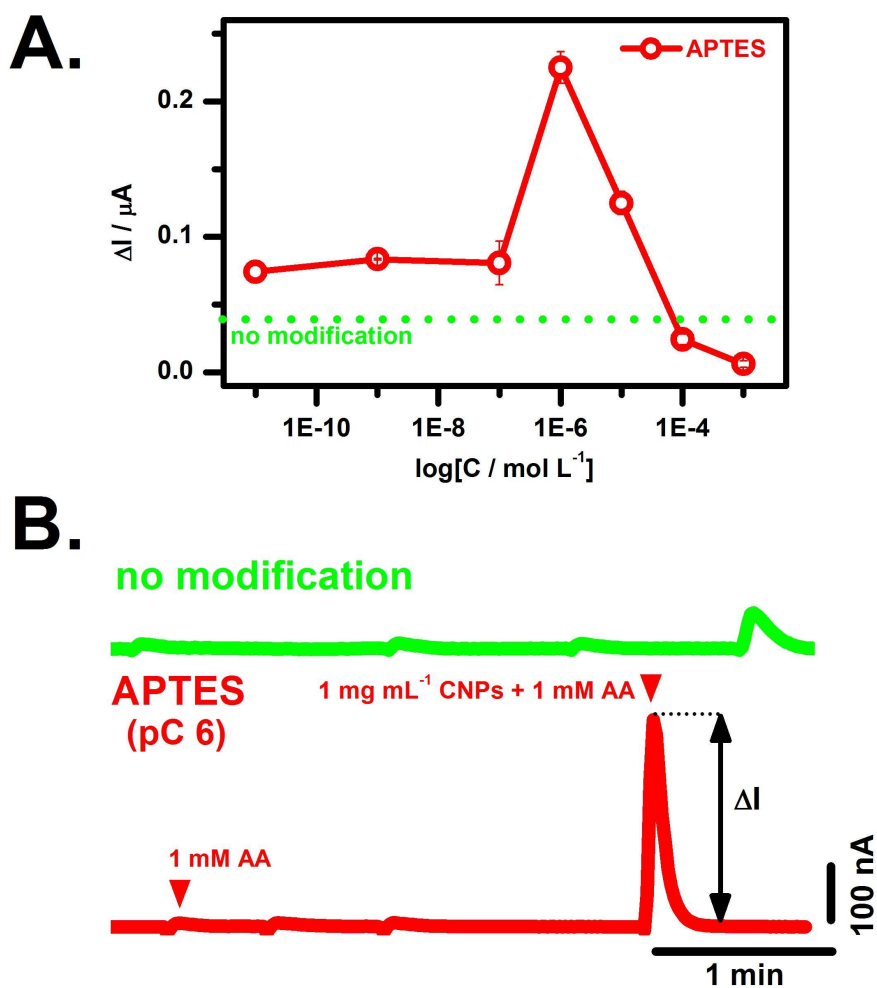


Fig.SI_4. (A) Optimization of the concentration of APTES solution (red) used for ITO modification. Baseline-corrected current (ΔI) of electrooxidation of 1 mM AA in 25 mM NaH₂PO₄ in the presence of 1 mg mL⁻¹ CNPs in 25 mM NaH₂PO₄ given as the function of APTES concentration. Chronoamperometric measurements were conducted in the FIA system⁹ at potential 0.2 V. A total flow rate was fixed at 3 mL min⁻¹. Each assay was repeated three times. **(B)** Chronoamperometric curves for bare ITO and ITO modified with the optimal concentration of APTES (10⁻⁶ mol L⁻¹).