Enantioselective synthesis of β-substituted Chiral Allylic Amines via Rh-Catalyzed Asymmetric Hydrogenation

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1. General Information

Unless otherwise noted, all reagents and solvents were purchased from commercial suppliers and used without further purification. NMR spectra were recorded on Bruker ADVANCE III (400 MHz) spectrometers for ¹H NMR and ¹³C NMR. CDCl₃ was the solvent used for the NMR analysis, with tetramethylsilane as the internal standard. Chemical shifts were reported upfield to TMS (0.00 ppm) for ¹H NMR and relative to CDCl₃ (77.0 ppm) for ¹³C NMR. Optical rotation was determined using a Perkin Elmer 343 polarimeter. HPLC analysis was conducted on an Agilent 1260 Series instrument. Column Chromatography was performed with silica gel Merck 60 (300-400 mesh). All new products were further characterized by HRMS. A positive ion mass spectrum of sample was acquired on a Thermo LTQ-FT mass spectrometer with an electrospray ionization source.

2. General procedure for the synthesis of compound Ia-i^[1,2]



Preparation of I-b, c, g

To a round bottom flask equipped with a magnetic stir bar was charged concentrated hydrochloric acid (38 mL), H₂O (55 mL), and freshly distilled arylamine (150 mmol)was added dropwise. The reaction mixture was then cooled to 0°C and a solution of sodium nitrite (10.6 g in 7 mL H₂O) was added dropwise and keep the internal temperature did not exceed 5°C. After stirring at 0°C for 40 min, 17.04 gram fluoboric acid (40%) was added to the reaction mixture in 15 min. After stirring at 0°C for 30 min, the reaction mixture was quiescent at that temperature for 2 h. The reaction mixture was then filtered, the filter cake was washed with 30 mL of ethanol (95%) and anhydrous ether (30×2 mL). Then it was dried in vacuum oven to give aromaticdiazonium tetrafluoroborate without further purification.

To a round bottom flask equipped with a magnetic stir bar was charged anhydrous sodium acetate (12.4 g), cuprous oxide (0.8 g), isopropenyl acetate (40 mL), aromaticdiazonium tetrafluoroborate (54 mmol) was added slowly. The reaction mixture was then warmed at 40-65 °C for 5-8 h. The precipitation was separated through filter and the filter residue was washed with ethyl acetate thoroughly. Combined the filtrate and washed with water and saturated sodium bicarbonate solution. The organic layer was dried over NaSO₄, filtered. Removing the solvent by rotary evaporation, the crude material was purified by flash chromatography (*n*-hexane : ethyl acetate 50:1) to afford the target product.

Preparation of I-a, d, e, f, h, i

In a typical reaction, an aryl carboxylic acid (0.05 mol, 1 equiv.) was dissolved in Ac₂O (0.25 mol, 25mL, 5 equiv.) at room temperature, and the solution was stirred and purged with N₂ for several minutes. The reaction was initiated by the addition of 1-Methylimidazole (0.025 mol, 0.5 equiv.), and the reaction was continuously purged with a slow flow of N₂ at room temperature until the starting material was completely disappear (by TLC). After completion, water (10 mL) was added to the reaction flask to hydrolyze Ac₂O. The reaction mixture was extracted with ethyl acetate (3 \times 20mL), and the extracts combined and washed with saturated Na₂HCO₃ followed by water , then dried over MgSO₄ and filtered . Removing the solvent by rotary evaporation gave the product mixture, then purified by flash chromatography (*n*-hexane /ethyl acetate: 20:1-10:1) to give the desired products.

3. General procedure for the synthesis of compound IIa-m^[3, 4]



Preparation of IIa - j

A mixture composed of the 1-aryl-2-propanone (1. 0 equiv.), 37% formaldehyde solution (1.2 equiv.), piperidine (10 mol%), and AcOH (10 mol%) was heated in anhydrous MeOH (1.4 M) at 60°C for 10 h. Upon completion, the reaction mixture was cooled down to ambient temperature, and then diluted with water. The aqueous layer was extracted several times with diethyl ether. The combined organic layers were dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The crude residue was purified by flash chromatography on silica gel, using an appropriate *n*-hexane/ ethyl acetate (20:1-10:1) mixture as eluent to give the pure products II.

Preparation of II-k-l

A mixture of phenylacetone (30 mmol), benzaldehyde (30 mmol) and piperidine (0.8 mmol) in toluene (50 mL) was refluxed for 12 h using a Dean-Stark water separator. Then the reaction mixture was concentrated in vacuo and the residue was subjected to silica gel column chromatography (penatane /EtOAc, 20/1-10/1) to give the desired product.

Preparation of II-m

A mixture of aldehyde (30 mmol), cycloakanone (36 mmol), diethyl ether (30 mL), and 1N NaOH solution (30 mL) was stirred at room temperature for 72 h. After reaction, the mixture was diluted with 50 mL of diethyl ether and the aqueous layer was separated and extracted with ether (3×20 mL). The combined ether solution was washed to neutral with water and dried over Na₂SO₄. The solvent was evaporated and the product was purified by silica gel column chromatography with petroleum/ ethyl acetate to yield the desired product.

4. General procedure for the synthesis of compound 1^[3,5]



Preparation of 1

A methanol solution of the α , β -unsaturated ketone (1.0 equiv.), hydroxylamine hydrochloride (1.1 equiv.), and Na₂CO₃ (1.2 equiv.) was stirred at ambient temperature for 12 h. Then diluted with water. The resultant layer was extracted with

diethyl ether several times. The combined organic layer was dried over anhydrous Na_2SO_4 , filtered and concentrated under reduced pressure. The crude residue was purified by flash chromatography on silica gel (*n*-hexane/EtOAc = 5/1-2/1) to give oxime as a white solid.

Acetic anhydride (3.0 equiv.) was added, in portions, to a solution of oxime (1.0 equiv.) obtained from first step in toluene (1.3 M) under a nitrogen atmosphere. Acetic acid (3.0 equiv.) was then added, followed by Fe powder (2.0 equiv.). The mixture was then heated to 70 °C overnight. The reaction was then cooled to room temperature and filtered through celite to remove solid residues, which were then washed with ethyl acetate. The combined filtrate was washed with brine. The organic phase was separated, dried by Na₂SO₄ and evaporated. The crude material was purified by flash chromatography (*n*-hexane/ ethyl acetate 5:1-2:1) to afford the product **1** as white solid.



N-(3-phenylbuta-1,3-dien-2-yl)acetamide (1a)

White solid, 0.56g, 60% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.32-7.48 (m, 5H), 6.74 (brs, 1H), 5.93 (s, 1H), 5.44 (s, 1H), 5.34 (s, 1H) 4.96 (s, 1H), 2.02 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.9, 147.4, 140.3, 138.4, 128.4, 128.3, 128.0, 114.9, 105.3, 24.5; ESI-HRMS Calculated for C₁₂H₁₃NNaO⁺ ([M+Na]⁺): 210.0889, found: 210.08894.

N-(3-m-tolylbuta-1,3-dien-2-yl)acetamide (1b)

White solid, 0.52g, 51% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.14-7.30 (m, 5H), 6.67 (brs, 1H), 5.96 (s, 1H), 5.43 (s, 1H), 5.35 (s, 1H) 4.96 (s, 1H), 2.38 (s, 3H), 2.03 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.8, 147.5, 140.4, 138.2, 138.1, 129.1, 128.6, 128.3, 125.1, 114.8, 105.0, 24.6, 21.4; ESI-HRMS Calculated for C₁₃H₁₆NO⁺ ([M+H]⁺): 202.1226, found: 202.12240.



N-(3-p-tolylbuta-1,3-dien-2-yl)acetamide (1c)

White solid, 0.72g, 71% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.26-7.33 (m, 2H), 7.14-7.19 (m, 2H), 6.77 (brs, 1H), 5.93 (s, 1H), 5.40 (s, 1H), 5.34 (s, 1H) 4.96 (s, 1H), 2.38 (s, 3H), 2.02 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.9, 147.2, 140.5, 138.2, 135.4, 129.1, 127.8, 114.3, 104.9, 24.5, 21.2; ESI-HRMS Calculated for C₁₃H₁₅NNaO⁺ ([M+Na]⁺): 224.1046, found: 224.10461.



N-(3-(4-methoxyphenyl)buta-1,3-dien-2-yl)acetamide (1d)

White solid , 0.49g, 45% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.27-7.36 (m, 2H), 6.81-6.92 (m, 2H), 6.50 (brs, 1H), 5.94 (s, 1H), 5.34 (s, 1H), 5.28 (s, 1H), 4.95 (s, 1H), 3.82 (s, 3H), 2.01 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.76, 159.73, 146.84, 140.54, 130.59, 129.18, 113.81, 113.61, 104.58, 55.33, 24.64; ESI-HRMS Calculated for C₁₃H₁₆NO₂⁺ ([M+H]⁺): 218.1176, found: 218.1175.



N-(3-(4-fluorophenyl)buta-1,3-dien-2-yl)acetamide (1e)

White solid, 0.31g, 30% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.27-7.40 (m, 2H), 6.96-7.11 (m, 2H), 6.55 (brs, 1H), 5.90 (s, 1H), 5.40 (s, 1H), 5.30 (s, 1H), 4.91 (s, 1H), 2.03 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.71, 163.99, 161.52, 146.39, 140.13, 134.42, 129.77, 129.69, 115.48, 115.27, 114.78, 105.52, 24.58; ESI-HRMS Calculated for C₁₂H_{13F}NO⁺ ([M+H]⁺): 206.0976, found: 206.0975.



N-(3-(4-chlorophenyl)buta-1,3-dien-2-yl)acetamide (1f)

White solid, 0.40g, 35% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.28-7.35 (m, 5H), 6.55 (brs, 1H), 5.89 (s, 1H), 5.42 (s, 1H), 5.33 (s, 1H), 4.91 (s, 1H), 2.02 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.71, 146.33, 139.95, 136.87, 134.23, 129.32, 128.64, 115.27, 105.71, 24.57; ESI-HRMS Calculated for C₁₂H₁₃ClNO⁺ ([M+H]⁺): 222.0680, found: 222.0679.

CI NHAc

N-(3-(2-chlorophenyl)buta-1,3-dien-2-yl)acetamide (1g)

White solid, 0.50g, 42% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.36-7.46 (m, 1H), 7.25-7.32 (m, 3H), 6.87 (brs, 1H), 5.80 (s, 1H), 5.59 (s, 1H), 5.21 (s, 1H) 4.66 (s, 1H), 2.15 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.9, 145.2, 138.6, 138.55, 133.2, 131.4, 129.6, 129.3, 126.7, 115.4, 106.8, 24.5; ESI-HRMS Calculated for C₁₂H₁₂ClNNaO⁺ ([M+Na]⁺): 244.0500, found: 244.04996.



N-(3-(4-bromophenyl)buta-1,3-dien-2-yl)acetamide (1h)

White solid, 0.40g, 30% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.42-7.53 (m, 2H), 7.38 – 7.13 (m, 2H), 6.53 (brs, 1H), 5.90 (s, 1H), 5.44 (s, 1H), 5.35 (s, 1H), 4.92 (s, 1H), 2.03 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.69, 146.39, 139.86, 137.34, 131.61, 129.64, 122.45, 115.32, 105.75, 24.58; ESI-HRMS Calculated for C₁₂H₁₃BrNO⁺ ([M+H]⁺): 266.0175, found: 266.0175, 268.0152.



N-(3-(naphthalen-2-yl)buta-1,3-dien-2-yl)acetamide (1i)

White solid, 0.45g, 38% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.73-8.04 (m, 4H), 7.41-7.57 (m, 3H), 6.57 (brs, 1H), 6.02 (s, 1H), 5.53 (s, 1H), 5.49 (s, 1H), 5.01 (s, 1H), 2.00 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.81, 147.42, 140.35, 135.61, 133.24, 133.16, 128.24, 128.10, 127.65, 127.19, 126.44, 125.76, 115.56, 105.08, 24.67; ESI-HRMS Calculated for C₁₆H₁₆NO⁺ ([M+H]⁺): 238.1226, found: 238.1225.



N-(3-benzylbuta-1,3-dien-2-yl)acetamide (1j)

White solid, 0.48g, 48% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.27-7.36 (m, 2H), 7.16-7.25 (m, 3H), 6.52 (brs, 1H), 5.66 (s, 1H), 5.30 (s, 1H), 5.10 (s, 1H), 4.97 (s, 1H), 3.60 (s, 2H), 2.04 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.68, 144.29, 139.29, 138.95, 128.78, 128.53, 126.45, 114.20, 104.96, 40.13, 24.49; ESI-HRMS Calculated for C₁₃H₁₆NO⁺ ([M+H]⁺): 202.1226, found : 202.1225.



N-(3,4-diphenylbuta-1,3-dien-2-yl)acetamide (1k)

White solid, 0.39g, 30% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.27-7.44 (m, 3H), 7.16-7.24 (m, 2H), 7.12 – 7.05 (m, 3H), .686-6.99 (m, 2H), 6.83 (s, 1H), 6.64 (brs, 1H), 5.83 (s, 1H), 4.92 (s, 1H), 2.04 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.75, 142.12, 139.75, 137.32, 136.26, 130.16, 129.66, 128.80, 128.04, 127.85, 127.28, 106.99, 24.51; ESI-HRMS Calculated for C₁₈H₁₈NO⁺ ([M+H]⁺): 264.1383, found : 264.1381.



N-(3-phenyl-4-(p-tolyl)buta-1,3-dien-2-yl)acetamide (11)

White solid, 0.40g, 26% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.27-7.41 (m, 3H), 7.15-7.24 (m, 2H), 6.88 –6.96 (m, 2H), 6.71-6.87 (m, 3H), 6.54 (brs, 1H), 5.82 (s, 1H), 4.90 (s, 1H), 2.25 (s, 3H), 2.05 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.78, 142.14, 138.82, 137.54, 137.24, 133.32, 130.15, 129.58, 128.81, 127.93, 127.75, 106.75, 24.54, 21.20; ESI-HRMS Calculated for C₁₉H₂₀NO⁺ ([M+H]⁺): 278.1539, found : 278.1538.

(E)-N-(5-benzylidenecyclopent-1-en-1-yl)acetamide (1m)

Dark red solid, 0.38g, 36% yield; ¹H NMR (400 MHz, CDCl₃) δ 7.39-7.44 (m, 4H), 7.15-7.24 (m, 1H), 6.96 (brs, 1H), 6.73 (t, J = 2.7 Hz, 1H), 6.11 (s, 1H), 2.93 – 2.79 (m, 2H), 2.58-2.66 (m, 2H), 2.21 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.40, 143.42, 137.51, 136.33, 128.49, 128.19, 126.32, 123.56, 114.30, 29.64, 28.60, 24.41; ESI-HRMS Calculated for C₁₄H₁₆NO⁺ ([M+H]⁺): 214.1226, found : 214.1226.

5. General Procedure for Asymmetric Hydrogenation of compound 1



A stock solution was made by mixing $[Rh(cod)_2]BF_4$ with (Sc,Rp)-Duanphos in a 1:1.1 molar ratio in CH₂Cl₂ at room temperature for 30 min in a nitrogen-filled glovebox. An aliquot of the catalyst solution (0.5mL, 0.0005 mmol) was transferred by syringe into the vials charged with different substrates (0.05 mmol for each) in anhydrous MeOH (0.5 mL). The vials were subsequently transferred into an autoclave into which hydrogen gas was charged. The reaction was then stirred under H₂ (1 atm) at room temperature for 6 h. The hydrogen gas was released slowly and carefully. The solution was concentrated and passed through a short column of silica gel (eluent: EtOAc) to yield the desired products. The ee values of compounds **2** were determined by HPLC analysis on a chiral stationary phase.



(S)-N-(3-phenylbut-3-en-2-yl)acetamide (2a)

White solid, 9.3 mg, 97% yield, 95% ee; $[\alpha]_D{}^{20} = -26.0$ (c = 0.5, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak OD-H column, hexane: isopropanol = 97: 3; flow rate = 1.0 mL/min; UV detection at 220 nm; t_R = 16.8 min (major), 21.6 min (minor). ¹H NMR (400 MHz, CDCl₃) δ = 7.38-7.46 (m, 2H), 7.29-7.38 (m, 3H), 5.67 (br, 1H), 5.35 (s, 1H), 5.24 (s, 1H), 5.03-5.19 (m, 1H), 1.96 (s, 3H), 1.33 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ = 169.2, 150.5, 139.9, 128.4, 127.8, 126.6, 112.2, 47.5, 23.4, 20.3. ESI-HRMS Calculated for C₁₂H₁₅NNaO⁺ ([M+Na]⁺): 212.1046, found: 212.10463.



(S)-N-(3-m-tolylbut-3-en-2-yl)acetamide (2b)

Yellow oil, 9.6 mg, 94% yield, 95% ee; $[\alpha]_D{}^{20} = -21.0$ (c = 0.4, CH₂Cl₂); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 254 nm; t_R = 11.7 min (minor), 13.7 min (major). ¹H NMR (400 MHz, CDCl₃) δ = 7.18-7.27 (m, 3H), 7.09-7.16 (m, 1H), 5.60 (br, 1H), 5.32 (s, 1H), 5.21 (s, 1H), 5.02-5.16 (m, 1H), 2.37 (s, 3H), 1.97 (s, 3H), 1.33 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ = 169.2, 150.7, 139.9, 137.9, 128.5, 128.3, 127.4, 123.8, 111.9, 47.6, 23.4, 21.5, 20.4. ESI-HRMS Calculated for C₁₃H₁₇NNaO⁺ ([M+Na]⁺): 226.1202, found: 226.12024.



(S)-N-(3-p-tolylbut-3-en-2-yl)acetamide (2c)

White solid, 9.5 mg, 93% yield, 94% ee; $[\alpha]_D{}^{20} = -22.7$ (c = 0.4, CH₂Cl₂); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 254 nm; t_R = 13.5 min (minor), 15.7 min (major). ¹H NMR (400 MHz, CDCl₃) δ = 7.29-7.36 (m, 2H), 7.13-7.18 (m, 2H), 5.61 (br, 1H), 5.33 (s, 1H), 5.20 (s, 1H), 5.04-5.17 (m, 1H), 2.36 (s, 3H), 1.96 (s, 3H), 1.33 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ = 169.2, 150.5, 137.6, 136.9, 129.1, 126.5, 111.5, 47.4, 23.4, 21.1, 20.3. ESI-HRMS Calculated for C₁₃H₁₇NNaO⁺ ([M+Na]⁺): 226.1202, found: 226.12044.



(R)-N-(3-(4-methoxyphenyl)but-3-en-2-yl)acetamide (2d)

White solid, 10.5 mg, 95% yield, 91% ee; $[\alpha]_D{}^{20}= + 25.3$ (c = 1.0, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak OD-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 254 nm; t_R = 19.9 min (major), 29.7 min (minor); ¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.30 (m, 2H), 6.99 – 6.72 (m, 2H), 5.56 (br s, 1H), 5.27 (s, 1H), 5.14 (s, 1H), 5.01-5.11 (m, 1H), 3.80 (s, 3H), 1.93 (s, 3H), 1.31 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 169.17, 159.29, 149.66, 132.18, 127.70, 113.72, 110.83, 55.28, 47.27, 23.45, 20.28; ESI-HRMS Calculated for C₁₃H₁₈NO₂⁺ ([M+H]⁺): 220.1332, found : 220.1330.



(*R*)-N-(3-(4-fluorophenyl)but-3-en-2-yl)acetamide (2e)

White solid, 10.0 mg, 96% yield, 86% ee; $[\alpha]_D{}^{20} = +21.9$ (c = 1.0, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak OD-H column, hexane: isopropanol = 95: 5; flow rate = 0.5 mL/min; UV detection at 220 nm; t_R = 22.4 min (major), 26.4 min (minor); ¹H NMR (400 MHz, CDCl₃) δ 7.45 – 7.30 (m, 2H), 7.07 – 6.94 (m, 2H), 5.56 (brs, 1H), 5.28 (s, 1H), 5.20 (s, 1H), 5.00-5.11 (m, 1H), 1.94 (s, 3H), 1.30 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 169.18, 163.69, 161.24, 149.55, 135.96, 135.92, 128.33, 128.25, 115.35, 115.14, 112.20, 47.42, 23.40, 20.24; ESI-HRMS Calculated for C₁₂H₁₅FNO⁺ ([M+H]⁺): 208.1132, found : 208.1131.



(R)-N-(3-(4-chlorophenyl)but-3-en-2-yl)acetamide (2f)

White solid, 10.9 mg, 97% yield, 97% ee; $[\alpha]_D{}^{20}$ = +18.4 (c = 1.0, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak OD-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 220 nm; t_R = 23.9 min (major), 28.2 min (minor); ¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.31 (m, 2H), 7.31 – 7.27 (m, 2H), 5.52 (brs, 1H), 5.31 (s, 1H), 5.23 (s, 1H), 5.00-5.10 (m, 1H), 1.94 (s, 3H), 1.30 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 169.18, 149.46, 138.34, 133.64, 128.55, 127.97, 112.68, 47.25, 23.40, 20.22; ESI-HRMS Calculated for C₁₂H₁₅ClNO⁺ ([M+H]⁺): 224.0837, found : 224.0836.

(S)-N-(3-(2-chlorophenyl)but-3-en-2-yl)acetamide (2g)

Yellow oil, 11.0 mg, 98% ee; $[\alpha]_D^{20} = -20.5$ (c = 0.4, CH₂Cl₂); The enantiomeric

excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 99: 1; flow rate = 1.0 mL/min; UV detection at 210 nm; $t_R = 42.7 \text{ min (major)}$, 48.4 min (minor). ¹H NMR (400 MHz, CDCl₃) $\delta = 7.38-7.42$ (m, 1H), 7.21-7.27 (m, 3H), 5.60 (br, 1H), 5.45 (s, 1H), 5.12 (s, 1H), 4.87-5.02 (m, 1H), 1.96 (s, 3H), 1.29 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) $\delta = 159.1$, 148.3, 139.7, 132.6, 130.5, 129.6, 128.7, 126.5, 115.5, 49.3, 23.4, 19.8. ESI-HRMS Calculated for $C_{12}H_{14}CINNaO^+([M+Na]^+)$: 246.0656, found 246.06561.



(R)-N-(3-(4-bromophenyl)but-3-en-2-yl)acetamide (2h)

White solid, 13.4 mg, 99% yield, 90% ee; $[\alpha]_D{}^{20} = +11.3$ (c=1.0, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 95: 5; flow rate = 0.5 mL/min; UV detection at 220 nm; t_R = 30.5 min (major), 35.9 min (minor); H NMR (400 MHz, CDCl₃) δ 7.59 – 7.38 (m, 2H), 7.33 – 7.17 (m, 2H), 5.48 (brs, 1H), 5.33 (s, 1H), 5.24 (s, 1H), 5.01-5.11 (m, 1H), 1.95 (s, 3H), 1.31 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 169.15, 149.51, 138.80, 131.51, 128.32, 121.84, 112.75, 76.74, 47.20, 23.42, 20.22; ESI-HRMS Calculated for C₁₂H₁₅BrNO⁺ ([M+H]⁺): 268.0332, found : 268.0332, 270.0332.



(R)-N-(3-(naphthalen-2-yl)but-3-en-2-yl)acetamide (2i)

White solid, 10.4 mg, 86% yield, 84% ee; $[\alpha]_D^{20} = -0.9$ (c = 1.0, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 95: 5; flow rate = 0.5 mL/min; UV detection at 220 nm; t_R = 32.1 min (major), 35.8 min (minor); ¹H NMR (400 MHz, CDCl₃) δ 7.91 – 7.72 (m, 4H), 7.52-7.58 (m, 1H), 7.43-7.50 (m, 2H), 5.54 (br s, 1H), 5.47 (s, 1H), 5.32 (s, 1H), 5.20-5.29 (m, 1H), 1.95 (s, 3H), 1.37 (d, *J* = 6.7 Hz, 3H); ³C NMR (101 MHz, CDCl₃) δ 169.18, 150.42, 137.20, 133.28, 132.93, 128.26, 127.98, 127.55, 126.25, 126.07, 125.45, 124.98, 112.66, 47.48, 23.48, 20.43; ESI-HRMS Calculated for C₁₆H₁₈NO⁺ ([M+H]⁺): 240.1383, found : 240.1381.



(R)-N-(3-benzylbut-3-en-2-yl)acetamide (2j)

White solid, 10.0 mg, 98%yield, 96% ee; $[\alpha]_D^{20} = +53.6$ (c = 0.5, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak OJ-H column, hexane: isopropanol = 95: 5; flow rate = 0.5 mL/min; UV detection at 220 nm; t_R = 33.8 min (major), 41.0 min (minor); ¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.24 (m, 2H), 7.24 –

7.14 (m, 3H), 5.25 (brs, 1H), 5.07 (s, 1H), 4.84 (s, 1H), 4.66 – 4.45 (m, 1H), 3.39 (s, 2H), 1.85 (s, 3H), 1.24 (d, J = 6.9 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 169.09, 149.67, 139.21, 129.00, 128.47, 126.33, 112.30, 48.53, 40.76, 23.41, 19.88; ESI-HRMS Calculated for C₁₃H₁₈NO⁺ ([M+H]⁺): 204.1383, found : 204.1382.

(*R*)-N-(3,4-diphenylbut-3-en-2-yl)acetamide (2k)

White solid, 13.0 mg, 98% yield, 97% ee; $[\alpha]_D{}^{20} = +17.3$ (c=1.0, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak OD-H column, hexane: isopropanol = 90: 10; flow rate = 1.0 mL/min; UV detection at 254 nm; t_R = 19.1 min (major), 7.8 min (minor); ¹H NMR (400 MHz, CDCl₃) δ 7.27 – 7.44 (m, 3H), 7.14-7.20 (m, 2H), 6.95 –7.12 (m, 3H), 6.93 – 6.75 (m, 2H), 6.57 (s, 1H), 5.47 (d, *J* = 8.2 Hz, 1H), 5.03 – 4.73 (m, 1H), 1.96 (s, 3H), 1.31 (d, *J* = 6.9 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 169.00, 143.24, 138.54, 136.40, 129.25, 129.17, 128.79, 127.88, 127.48, 126.77, 126.74, 51.89, 23.53, 20.60 ESI-HRMS Calculated for C₁₈H₂₀NO⁺ ([M+H]⁺): 266.1539, found : 266.1537.



(R)-N-(3-phenyl-4-(p-tolyl)but-3-en-2-yl)acetamide (2l)

White solid, 13.8 mg, 99% yield, 97% ee; $[\alpha]_D^{20} = +92.6$ (c = 1.0, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak OD-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 254 nm; t_R = 13.7 min (major), 19.9 min (minor); ¹H NMR (400 MHz, CDCl₃) δ 7.29-7.41 (m, 3H), 7.23 – 7.06 (m, 2H), 6.83-6.90 (m, 2H), 6.71-6.81 (m, 2H), 6.54 (s, 1H), 5.48 (d, *J* = 8.2 Hz, 1H), 4.86-5.00 (m, 1H), 2.22 (s, 3H), 1.95 (s, 3H), 1.30 (d, *J* = 6.9 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 168.98, 142.24, 138.75, 136.52, 133.49, 129.29, 129.06, 128.79, 128.63, 127.39, 126.72, 51.89, 23.54, 21.12, 20.63; ESI-HRMS Calculated for C₁₉H₂₂NO⁺ ([M+H]⁺): 280.1696, found : 280.1696.

NHAc

(*R*)-N-(2-benzylidenecyclopentyl)acetamide (2m)

White solid, 10.1 mg, 94% yield, 84% ee; $[\alpha]_D{}^{20} = +53.6$ (c=0.5, CHCl₃); The enantiomeric excess was determined by HPLC on Chiralpak AD-H column, hexane: isopropanol = 95: 5; flow rate = 1.0 mL/min; UV detection at 220 nm; t_R = 18.4 min (major), 22.0 min (minor); ¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.16 (m, 5H), 6.37 (s,

1H), 5.54 (brs, 1H), 4.83-4.93 (m, 1H), 2.78 – 2.56 (m, 2H), 2.25 – 2.14 (m, 1H), 2.07 (s, 3H), 1.93 - 1.81 (m, 1H), 1.76 - 1.61 (m, 1H), 1.48 - 1.36 (m, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 169.87, 145.21, 137.63, 128.36, 128.26, 126.56, 122.71, 55.40, 33.62, 30.41, 23.64, 23.25; ESI-HRMS Calculated for C₁₄H₁₈NO⁺ ([M+H]⁺): 216.1383, found : 216.1382.

6. Procedure for the synthesis of compound 3d and hydrogenation of

1d and 1l with low catalyst loading



A stock solution was made by mixing [Rh(cod)₂]BF₄ with (*R*c,*S*p)-Duanphos in a 1:1.1 molar ratio in CH₂Cl₂ at room temperature for 30 min in a nitrogen-filled glovebox. An aliquot of the catalyst solution (0.5mL, 0.0005 mmol) was transferred by syringe into the vials charged with different substrates (0.05 mmol for each) in anhydrous MeOH (0.5 mL). The vials were subsequently transferred into an autoclave into which hydrogen gas was charged. The reaction was then stirred under H₂ (50 atm) at room temperature for 24 h. The hydrogen gas was released slowly and carefully. The solution was concentrated and passed through a short column of silica gel (eluent: EtOAc) to yield the desired products. The dr value of the resulting product was determined by ¹H NMR. ¹H NMR (400 MHz, CDCl₃) δ 7.20 – 7.04 (m, 2H), 6.94 – 6.76 (m,2H), 5.27 – 4.97 (m, 1H), 4.42 – 4.06 (m, 1H), 3.83 (s, 3H), 3.00 – 2.51 (m, 1H), 1.91 (s, 3H), 1.29 (d, *J* = 7.0 Hz, 3H), 1.10 (d, *J* = 6.7 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 169.34, 158.24, 134.79, 129.05, 128.81, 113.75, 55.26, 49.60, 43.67, 23.56, 18.88, 17.66.



A stock solution was made by mixing $[Rh(cod)_2]BF_4$ with (Rc,Sp)-Duanphos in a 1:1.1 molar ratio in CH_2Cl_2 at room temperature for 30 min in a nitrogen-filled

glovebox. An aliquot of the catalyst solution (0.03 mmol, 1 mol %) was transferred by syringe into the vials charged with **1d** (0.65g, 3 mmol) in anhydrous MeOH (3 mL). The vials were subsequently transferred into an autoclave into which hydrogen gas was charged. The reaction was then stirred under H₂ (1 atm) at room temperature for 5 h. The hydrogen gas was released slowly and carefully. The solution was concentrated and passed through a short column of silica gel (eluent: EtOAc) to yield the desired products **2d** as a white solid.



A stock solution was made by mixing $[Rh(cod)_2]BF_4$ with (Rc,Sp)-Duanphos in a 1:1.1 molar ratio in CH_2Cl_2 at room temperature for 30 min in a nitrogen-filled glovebox. An aliquot of the catalyst solution (0.0001 mmol, 0.1 mol %) was transferred by syringe into the vials charged with **11** (0.1 mmol) in anhydrous MeOH (1 mL). The vials were subsequently transferred into an autoclave into which hydrogen gas was charged. The reaction was then stirred under H₂ (15 atm) at room temperature for 24 h. The hydrogen gas was released slowly and carefully. The solution was concentrated and passed through a short column of silica gel (eluent: EtOAc) to yield the desired products **21** as a white solid without no loss of yield and ee.

7. References

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8. NMR spectra of 1 and 2, HPLC spectra of 2















 $1g\mathchar`{1}^{1}H$ NMR and ^{13}C NMR









1k-1H NMR and 13C NMR







11-1H NMR and 13C NMR



1m-¹H NMR and ¹³C NMR







2b - $^1\!H$ NMR and $^{13}\!C$ NMR

















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2h - $^1\!H$ NMR and $^{13}\!C$ NMR











2k - $^1\!H$ NMR and $^{13}\!C$ NMR















2a - HPLC

Data File E:\DATA\WQL\WQL-2-69-C\DAD-0DH-97-3-WQL-2-68-C 2013-05-14 16-26-27\043-0201.D Sample Name: WQL-2-54-1

Acq. Operator	: SYSTEM Seq. Line : 2	
Acq. Instrument	: 1260HPLC-DAD Location : Vial 43	
Injection Date	: 5/14/2013 4:56:25 PM INJ : 1 Inj Volume : 5.000 ul	
Acq. Method	: E:\DATA\WQL\WQL-2-69-C\DAD-ODH-97-3-WQL-2-68-C 2013-05-	14 16-26-27\DAD-
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Last changed	: 5/14/2013 4:26:27 PM by SYSTEM	
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Last changed	: 4/29/2015 3:09:38 PM by SYSTEM	
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Additional Info	: Peak(s) manually integrated	
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1260HPLC-VWD 4/29/2015 2:48:27 PM SYSTEM

2b - HPLC

Data File E:\DATA\WQL\WQL-2-78-A\VWD-0JH--95-5-WQL-2-78-A 2013-06-14 21-02-41\023-0401.D Sample Name: WQL-2-74-3



1260HPLC-VWD 4/29/2015 3:46:29 PM SYSTEM



1260HPLC-VWD 4/29/2015 3:44:40 PM SYSTEM

Data File E:\DATA\WQL\WQL-2-78-A\VWD-0JH--95-5-WQL-2-78-A 2013-06-14 21-02-41\025-0601.D Sample Name: WQL-2-78-4 Acq. Operator : SYSTEM Acq. Instrument : 1260HPLC-VWD Seg. Line : 6 Location : Vial 25 Injection Date : 6/14/2013 11:57:13 PM Inj: 1 Inj Volume : 5.000 µl : E:\DATA\WQL\WQL-2-78-A\VWD-0JH--95-5-WQL-2-78-A 2013-06-14 21-02-41\VWD-Acg. Method 0JH-95-5-1ML-254NM-40MIN(1-2).M Last changed : 6/14/2013 9:02:42 PM by SYSTEM Analysis Method : E:\DATA\WQL\WQL-2-78-A\VWD-0JH--95-5-WQL-2-78-A 2013-06-14 21-02-41\VWD-0JH-95-5-1ML-254NM-40MIN(1-2).M (Sequence Method) Last changed : 4/29/2015 3:50:38 PM by SYSTEM (modified after loading) Additional Info : Peak (3) manually integrated VWD1A, Wavelength=254 nm (E:\DATAWQ...L-2-78 A\VWD-OJH--95-5 WQL-2-78 A 2013-06-14 21-02-41\025-0601.D) mAU 15.724 80 60 ŃHAc 40 20 0 16 20 12 14 18 min Area Percent Report Sorted By Signal : Multiplier : 1.0000 1.0000 Dilution : Do not use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=254 nm Peak RetTime Type Width Area Height Area [min] [mAU*s] [mAU] ÷ # [min] ----|-----|----|-----|-----|-----| 1 13.443 BB 0.3566 2116.04980 91.23906 48.1864 2 15.724 BB 0.4393 2275.33154 77.78905 51.8136 Totals : 4391.38135 169.02811 *** End of Report ***

1260HPLC-VWD 4/29/2015 3:52:35 PM SYSTEM



1260HPLC-VWD 4/29/2015 3:50:48 PM SYSTEM





1260HPLC-DAD 2/23/2016 9:35:03 PM SYSTEM

2e - HPLC

Data File E:\DATA\GWC\GWC-16-1\DUI F 0.5 2016-01-08 17-09-44\003-0201.D Sample Name: 16-1-3

Acq. Operator : SYSTEM Acq. Instrument : 1260HPLC-DAD Seq. Line : 2 Location : Vial 3 Inj : 1 Inj Volume : 5.000 μl Injection Date : 1/8/2016 5:21:36 PM Acq. Method : E:\DATA\GWC\GWC-16-1\DUI F 0.5 2016-01-08 17-09-44\DAD-0D(1-2)-95-5-0. 5ML-220NM-254NM-50MIN.M Last changed : 1/8/2016 5:48:01 PM by SYSTEM (modified after loading) Analysis Method : E:\DATA\GWC\GWC-16-1\DUI F 0.5 2016-01-08 17-09-44\DAD-0D(1-2)-95-5-0. 5ML-220NM-254NM-50MIN.M (Sequence Method) : 2/22/2016 10:33:13 AM by SYSTEM Last changed (modified after loading) Additional Info : Peak(s) manually integrated DAD1 D, Sig=220,4 Ref=off(E:DATA\GWC\GWC-16.1DUIF05.2016-01-08.17-09-444003-0201.D) mAU] 120 100 -NHAc 80 -60 -40 -20 -0 зo 24 28 min Area Percent Report Sorted By Signal : 1.0000 Multiplier : 1.0000 Dilution Do not use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 D, Sig=220,4 Ref=off Peak RetTime Type Width Height Area Area # [min] [mAU*s] [mAŬ] % 1 22.351 BB 0.8191 6582.78223 121.29909 47.9349 2 26.172 BB 0.8550 7149.95996 125.06830 52.0651 Totals : 1.37327e4 246.36739 -----*** End of Report ***

1260HPLC-DAD 2/22/2016 10:33:36 AM SYSTEM



S52

2f - HPLC

Data File E:\DATA\GWC\GWC-16-1\DUI CL 2015-12-31 18-32-05\041-0201.D Sample Name: gwc-16-1-5

Acq. Operator :	SYSTEM			Seq. Lin	e: 2
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2g - HPLC

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1260HPLC-VWD 4/29/2015 3:17:33 PM SYSTEM

2h - HPLC

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1260HPLC-DAD 2/22/2016 10:40:35 AM SYSTEM

2i - HPLC

Data File E:\DATA\GWC\GWC-16-1\ZHIAND NAI 2016-01-25 16-12-53\003-0501.D Sample Name: NAI WAI

Acq. Operator : SYSTEM Seq. Line : 5 Acq. Instrument : 1260HPLC-VWD Location : Vial 3 Injection Date : 1/25/2016 6:59:23 PM Inj: l Inj Volume : 5.000 µl Acq. Method : E:\DATA\GWC\GWC-16-1\ZHIAND NAI 2016-01-25 16-12-53\VWD-ADH(1-6)-95-5-220NM--0.5-40MIN.M Last changed : 1/25/2016 4:12:54 PM by SYSTEM Analysis Method : E:\DATA\GWC\GWC-16-1\ZHIAND NAI 2016-01-25 16-12-53\VWD-ADH(1-6)-95-5-220NM--0.5-40MIN.M (Sequence Method) Last changed : 2/22/2016 10:54:38 AM by SYSTEM (modified after loading) Additional Info : Peak (3) manually integrated VWD1 A, Wavelength=220 nm (E:\DATA\GWC\GWC-16 1\ZHIAND NAI 2016-01-25 16 12-53\003-0501.D) mAU 175 150 -ŃHAc , BBRER 125 8 8 8.50 8 100 75 -50 · 25 -0 32 38 31 33 <u>34</u> зŚ 36 min Area Percent Report _____ Sorted By Signal Multiplier 1.0000 : 1.0000 Dilution : Do not use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=220 nm Peak RetTime Type Width Height Area Area [mAU] [min] [mAU*s] # [min] ÷ 1 32.089 BB 0.8365 5530.81592 102.99131 47.7056 2 35.739 MM 1.0085 6062.83252 100.19076 52.2944 Totals : 1.15936e4 203.18207 *** End of Report ***

1260HPLC-DAD 2/22/2016 10:54:42 AM SYSTEM



1260HPLC-DAD 2/22/2016 10:55:58 AM SYSTEM

2j - HPLC

Data File E:\DATA\GWC\GWC-16-1\ZHIAND NAI 2016-01-25 16-12-53\001-0201.D Sample Name: ZHI WAI

Acq. Operator	: SYSTEM Seq. Line : 2
Acq. Instrument	t: 1260HPLC-VWD Location: Vial 1
Injection Date	: 1/25/2016 4:24:27 FM INJ : 1 Thi Volume : 5,000 ul
Acq. Method	: E:\DATA\GWC\GWC-16-1\ZHIAND NAI 2016-01-25 16-12-53\VWD-0J(1-2)-95-5-0.
	5ML-60MIN-220NM.M
Last changed	: 1/25/2016 5:15:29 PM by SYSTEM
Analysis Method	(modified after loading) • • F•\DaTa\GMC\GMC_16_1\7HTAMD MAT 2016_01_25 16_12_53\WWD_0J(1_2)_95_5_0
Andrysis neared	5ML-60MIN-220NM.M (Sequence Method)
Last changed	: 2/23/2016 9:03:26 PM by SYSTEM
	(modified after loading)
Additional Info): Peak (3) manually integrated (melenathe 220 nm (E-DATAGOWC-)GWC-16 1/271AND NAI 2016-01-25 16 12-53/001-0201 D)
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	Area Percent Report
Sorted By	: Signal
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Do not use Mult	cipiter & Dilution Factor with ISDS
Signal 1: VWD1	A, Wavelength=220 nm
reak KetTime Ty # [min]	ppe wiath Area Height Area [min] [màll≭s] [màll] %
	[maxi, [maxi,] [maxi,] ,
1 33.768 MM	1 1.0958 4390.90186 66.78225 52.5086
2 40.634 MM	1 1.2913 3971.35083 51.25767 47.4914
Totals •	
	8362, 25269 118, 03992
	8362.25269 118.03992
	8362.25269 118.03992

1260HPLC-DAD 2/23/2016 9:03:34 PM SYSTEM



2k - HPLC

Data File E:\DATA\GWC\GWC15-1-9\15-9-15 2015-10-09 19-17-23\073-0301.D Sample Name: wai

Acg. Operator	: SYSTEM			Seg. Line	: 3		
Acq. Instrument	: 1260HPL	C-DAD		Location	: Vial 73		
Injection Date	· 10/9/20	 15 7•55•09 P	M	Tni	• 1		
injection bace	. 10,0,00			Ini Volume	· · · ·		
Acq. Method	: E:\DATA	\GWC\GWC15-1	-9\15-9-15	2015-10-09	19-17-23\DAD-OD	H-90-10-1ML-	
-	40MIN(1	-2).M					
Last changed	: 10/9/20	15 7:52:27 P	M by SYSTEM	r			
Analysis Method	• F•\DATA	GMC\GMC15-1	-9115-9-15	- 2015-10-09	19-17-23\DAD-0D	H-90-10-1ML-	
MIGIJOID IICAIOG	40MTN(1)	-2) M (Semie	nce Method)	2010 10 05	15 1, 20,000 00		
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Last changed	. 2/23/20.	10 /.42.JI F.	a by Sisier	L			
DAD4 B. Sia		ed after loa	aing) 454 0450 4500	45 40 00 40 47 7	2072 0204 0		
	-204,4 Кеноп (с		10-1-8/10-8-10-20	10-10-08-18-17-2	5075-0501.0)		
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5	7.5		12.5	15	17.5 20	22.5	n in
5	7.5	10 10 Area Percent	12.5 Report	15	17.5 20	22.5	 min
	7.5	10 Area Percent	125 Report	15		22.5	
	7.5	10 Area Percent	125 Report	15		22.5	 min
Sorted By	7.5	10 Area Percent Signal	125 Report	15	<u>17.5</u> 20	22.5	min
5 Sorted By Multiplier	<u>7.5</u>	10 Area Percent Signal 1.0000	125 Report	15		22.5	<u>, , , , , , , , , , , , , , , , , , , </u>
Sorted By Multiplier Dilution	75 75	10 Area Percent Signal 1.0000 1.0000	12.5 Report	15		22.5	<u>, , , , , , , , , , , , , , , , , , , </u>
Sorted By Multiplier Dilution Do not use Mult:	75	10 Area Percent Signal 1.0000 1.0000 ilution Fact	Report	15 		22.5	<u>, , , , , , , , , , , , , , , , , , , </u>
Sorted By Multiplier Dilution Do not use Mult:	75 75	10 Area Percent Signal 1.0000 1.0000 ilution Fact	12.5 Report	15 		22.5	 mir
5 Sorted By Multiplier Dilution Do not use Mult:	75 75 : iplier & D	10 Area Percent Signal 1.0000 1.0000 ilution Fact	Report	15 	<u>17.5</u> 20	22.5	 mir
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Sorted By Multiplier Dilution Do not use Mult: Signal 1: DAD1 1	75 	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off	Report	15 		22.5	min
Sorted By Multiplier Dilution Do not use Mult: Signal 1: DAD1 I Peak RetTime Typ	75 	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area	Report or with IST	15 TDs Area		22.5	min
5 Sorted By Multiplier Dilution Do not use Mult: Signal 1: DAD1 H Peak RetTime Typ # [min]	75 	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area [mAU*s]	12.5 Report 	TDs Area		22.5	
5 Sorted By Multiplier Dilution Do not use Mult: Signal 1: DAD1 I Peak RetTime Typ # [min]	75 ; iplier & D 3, Sig=254 pe Width [min]	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area [mAU*s]	Report Report or with IST Height [mAU]	Ds Area		22.5	
Sorted By Multiplier Dilution Do not use Mult: Signal 1: DAD1 I Peak RetTime Tyy # [min]	75 	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area [mAU*s]	Report Report or with IST Height [mAU]	15 Ds Area &		22.5	
Sorted By Multiplier Dilution Do not use Mult: Signal 1: DAD1 H Peak RetTime Typ # [min] 	75 ; iplier & D 3, Sig=254 pe Width [min] 0.3256 0.7527	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area [mAU*s] 	Report Report or with IST Height [mAU] 46.90849	15 Ds Area % 1 49.9797		22.5	min
Sorted By Multiplier Dilution Do not use Mult: Signal 1: DAD1 H Peak RetTime Typ # [min] 	75 75 iplier & D 3, Sig=254 pe Width [min] 0.3256 0.7527	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area [mAU*s] 	Report Report or with IST Height [mAU] 46.90849 20.01796	15 Ds Area % 1 49.9797 50.0203		22.5	
Sorted By Multiplier Dilution Do not use Mult: Signal 1: DAD1 H Peak RetTime Tyn # [min] 	75 75 iplier & D 3, Sig=254 pe Width [min] 1 0.3256 0.7527	Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area [mAU*s] 995.52374 996.33215	125 Report or with IST Height [mAU] 46.90849 20.01796	Ds Area % 	<u>17.5</u> 20	22.5	
Sorted By Multiplier Dilution Do not use Mult: Signal 1: DAD1 I Peak RetTime Typ # [min] 	75 ; iplier & D 3, Sig=254 pe Width [min] 0.3256 0.7527	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area [mAU*s] [Report Report or with IST Height [mAU] 46.90849 20.01796 66.92645	15 Ds Area & 1 49.9797 50.0203	<u>175</u> 20	22.5	mir
Sorted By Multiplier Dilution Do not use Mult: Sigmal 1: DADI H Peak RetTime Typ # [min] 	75 ; iplier & D 3, Sig=254 pe Width [min] 1 0.3256 0.7527	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area [mAU*s] 	Report Report or with IS7 Height [mAU] 46.90849 20.01796 66.92645	15 Ds Area & 1 49.9797 50.0203	<u>175</u> 20	22.5	nir
Sorted By Multiplier Dilution Do not use Mult: Signal 1: DADI I Peak RetTime Typ # [min] 	75 ; iplier & D 3, Sig=254 pe Width [min] 	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area [mAU*s] 	125 Report or with IST Height [mAU] 46.90849 20.01796 66.92645	15 Ds Area % 		22.5	 mir
Sorted By Multiplier Dilution Do not use Mult: Signal 1: DAD1 I Peak RetTime Typ # [min] 	75 ; iplier & D 3, Sig=254 pe Width [min] 0.3256 0.7527	10 Area Percent Signal 1.0000 1.0000 ilution Fact ,4 Ref=off Area [mAU*s] [] 995.52374 996.33215 1991.85590	125 Report or with IS7 Height [mAU] 46.90849 20.01796 66.92645	15 Ds Area * 1 49.9797 50.0203	<u>17.5</u> <u>20</u>	22.5	

1260HPLC-DAD 2/23/2016 7:42:39 PM SYSTEM

Data File E:\DATA\GWC\GWC15-1-9\15-9-15 2015-10-09 19-17-23\072-0201.D Sample Name: 15-9-15 Acq. Operator : SYSTEM Acq. Instrument : 1260HPLC-DAD Seq. Line : 2 Location : Vial 72 Injection Date : 10/9/2015 7:29:15 PM Inj: 1 Inj Volume : 5.000 µl Acq. Method : E:\DATA\GWC\GWC15-1-9\15-9-15 2015-10-09 19-17-23\DAD-0DH-90-10-1ML-40MIN(1-2).M : 10/9/2015 7:52:27 PM by SYSTEM Last changed (modified after loading) Analysis Method : E:\DATA\GWC\GWC15-1-9\15-9-15 2015-10-09 19-17-23\DAD-0DH-90-10-1ML-40MIN(1-2).M (Sequence Method) : 2/23/2016 7:43:46 PM by SYSTEM Last changed (modified after loading) Additional Info : Peak(s) manually integrated DAD1 B, Sig=254,4 Retoff (E:DATA/OWC/GWC15 1-9/15-9-15 2015-10-09 19-17-23/072-0201.D) mAU 80 Ph 60 **NHAc** 19.163 40 20 1. A. nΦ 0 20 75 <u>10</u> 12.5 15 17.5 22.5 min Area Percent Report Sorted By : Signal Multiplier : 1.0000 1.0000 : Dilution Do not use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 B, Sig=254,4 Ref=off Peak RetTime Type Width Area Height Area /eak Rectifier ---# [min] [mAU*s]
----|-----|-----|------|1 7.851 MM 0.4481 35.14143
2 19.163 BB 0.7590 2051.52808 [mAU] * -----1.30715 1.6841 41.77021 98.3159 2086.66951 43.07736 Totals : *** End of Report ***

1260HPLC-DAD 2/23/2016 7:43:50 PM SYSTEM

21 - HPLC

Data File E:\DATA\GWC\GWC15-1-9\GWC-15-9-18 2015-10-15 08-07-49\021-0301.D Sample Name: 15-9-19-2

Acq. Operator : SYSTEM Seq. Line : 3 Acq. Instrument : 1260HPLC-DAD Location : Vial 21 Inj : 1 Inj Volume : 5.000 μ1 Injection Date : 10/15/2015 8:56:04 AM : E:\DATA\GWC\GWC15-1-9\GWC-15-9-18 2015-10-15 08-07-49\DAD-0DH-95-5-1ML-Acq. Method 220-254NM-40MIN(1-2).M Last changed : 10/15/2015 9:26:39 AM by SYSTEM (modified after loading) Analysis Method : E:\DATA\GWC\GWC15-1-9\GWC-15-9-18 2015-10-15 08-07-49\DAD-0DH-95-5-1ML-220-254NM-40MIN(1-2).M (Sequence Method) : 2/23/2016 7:56:24 PM by SYSTEM Last changed (modified after loading) Additional Info : Peak(s) manually integrated DAD1 H.Sig=254.4 Re=off(E:DATAGWC\GWC15-1-9.GWC-15-9.18 2015 10-15 08-07-49021-0301.D) mAU] 13.756 40 , 5⁸38 Ph 30 -NHAc àď 20 10 0 22 ź 14 . 24 mir 12 16 18 10 _____ Area Percent Report Sorted By : Sional 1.0000 : Multiplier Dilution Do not use Multiplier & Dilution Factor with ISTDs Signal 1: DAD1 H, Sig=254,4 Ref=off Height Area Peak RetTime Type Width Area # [min] [mAU*s] [mAŬ] % 1 13.756 BB 0.6139 1391.82678 35.45882 51.2043 2 19.791 MM 0.9116 1326.35547 24.24839 48.7957 Totals : 2718.18225 59.70722 *** End of Report ***

1260HPLC-DAD 2/23/2016 7:56:53 PM SYSTEM



1260HPLC-DAD 2/23/2016 7:57:41 PM SYSTEM

2m - HPLC

Data File E:\DATA\GWC\GWC15-11\GWC-15-11-3 2015-11-17 11-05-41\062-0301.D Sample Name: SLY-3-70-1-J

Acq. Operator : SYSTEM Seq. Line : 3 Acq. Instrument : 1260HPLC-VWD Location : Vial 62 Injection Date : 11/17/2015 11:48:05 AM Inj: 1 Inj Volume : 5.000 µl Acq. Method : E:\DATA\GWC\GWC15-11\GWC-15-11-3 2015-11-17 11-05-41\VWD-ADH(1-2)-95-5-220-254NM-40MIN.M Last changed : 11/17/2015 11:42:01 AM by SYSTEM Analysis Method : E:\DATA\GWC\GWC15-11\GWC-15-11-3 2015-11-17 11-05-41\VWD-ADH(1-2)-95-5-220-254NM-40MIN.M (Sequence Method) : 2/23/2016 8:59:45 PM by SYSTEM Last changed (modified after loading) Additional Info : Peak (s) manually integrated VWD1A, Wavelength=220 nm (E:\DATA\GWC\GWC1511\GWC-1511-32015-11-1711-05-41\062-0301.D) mAU] NHAc Ph 40 18.339 21.807 30 20 10 0 16 18 20 22 24 min Area Percent Report _____ Signal Sorted By : a1،0000 1.0000 : Multiplier Dilution 1.0000 : Do not use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=220 nm Area Peak RetTime Type Width Height Area # [min] [mAU*s] [mAŬ] % 1 18.339 BB 0.4951 1067.27368 33.10320 50.2909 2 21.807 BB 0.5744 1054.92639 28.07599 49.7091 Totals : 2122.20007 61.17919 *** End of Report *** 1260HPLC-DAD 2/23/2016 8:59:50 PM SYSTEM Page 1 of 1

Data File E:\DATA\GWC\GWC15-11\GWC-15-11-3 2015-11-17 11-05-41\061-0201.D Sample Name: SLY-3-70-3-J ------Acq. Operator : SYSTEM Seq. Line : 2 Acq. Instrument : 1260HPLC-VWD Location : Vial 61 Injection Date : 11/17/2015 11:17:19 AM Inj : 1 Inj Volume : 5.000 µl : E:\DATA\GWC\GWC15-11\GWC-15-11-3 2015-11-17 11-05-41\VWD-ADH(1-2)-95-5-Acq. Method 220-254NM-40MIN.M Last changed : 11/17/2015 11:42:01 AM by SYSTEM (modified after loading) Analysis Method : E:\DATA\GWC\GWC15-11\GWC-15-11-3 2015-11-17 11-05-41\VWD-ADH(1-2)-95-5-220-254NM-40MIN.M (Sequence Method) Last changed : 2/23/2016 9:01:10 PM by SYSTEM (modified after loading) Additional Info : Peak(s) manually integrated VWD1 A, Wavelength=220 nm (E:\DATA\@WC\GWC15-11\GWC-15-11-3 2015-11-17 11-05-41\081-0201.D) mAU 쯇 NHAc 200 Ρh 150 100 50 8 3*\$* 0 22 24 16 20 min 18 Area Percent Report Sorted By : Signal Multiplier : 1.0000 1.0000 Dilution : Do not use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=220 nm Peak RetTime Type Width Height Area Area [mAU*s] # [min] [min] [mAU] ÷ 1 18.482 BB 2 22.064 MM 0.4863 7425.51709 233.97545 91.2545 0.6197 711.63751 19.13794 8.7455 8137.15460 253.11339 Totals :

1260HPLC-DAD 2/23/2016 9:01:44 PM SYSTEM