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Supporting Information

Biginelli-based Organic Nanoprobe for Simultaneous Estimation of Tyramine and 1, 2-Diaminopropane: application in real samples.

Gaganpreet Kaur,^a Tilak Raj,^b Navneet Kaur^{a,c*} and Narinder Singh^{b*}

^aCentre for Nanoscience & Nanotechnology (UIEAST), Panjab University, Chandigarh, 160014, India. E-mail: navneetkaur@pu.ac.in; Tel: +91-1722534464

^bDepartment of Chemistry, Indian Institute of Technology Ropar, Rupnagar, Punjab, India, 140001. E-mail: nsingh@iitrpr.ac.in; Fax: +91-1881223395; Tel: +91-1881242176.

^cDepartment of Chemistry, Panjab University, Chandigarh, 160014, India. E-mail: navneetkaur@pu.ac.in

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Table S1: Comparison of current sensor with existing literature.





Figure S1.¹H NMR spectrum of compound 1 and its expansion.



Figure S2.¹³C NMR spectrum of compound 1.



Figure S3. ESI Mass spectrum of compound 1.



Figure S4. Plot of variation in size of nanoparticles as a function of concentration of compound 1 in water.



Figure S5. Linear regression graph for Ag (I) titration.



Figure S6. Linear regression graphs for Tyramine titration (A) and linear regression graph for 1,2-Diaminopropane titration (B).



Figure S7. Non-linear regression graphs between Fluorescence Intensity vs. Concentrations of amines (at higher concentrations).



Figure S8. Fluorescence spectra of nano-aggregates N1 at different concentrations of (A) TBA perchlorate to evaluate the salt effect and (B) in presence of NaCl.



Figure S9. Fluorescence spectra of nano-aggregates N1 at different pH values.



Figure S10. Fluorescence intensity v/s pH graphs of A) **N1.Ag(I)** complex with 50 μ M Tyramine and B) **N1.Ag(I)** complex with 50 μ M 1,2-Diaminopropane.



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Figure S14. Fluorescence intensity variation of A) **N1** on addition of different metal ions and B) **N1.Ag(I)** on addition of different Biogenic amines in presence of HEPES buffer, pH 7.4.



Figure S15. Fluorescence intensity v/s wavelength graph of N1.Ag(I) complex with 50 μ M each of Tyramine and 50 μ M 1,2-Diaminopropane mixed together.

Determination of detection limit.

The detection limit (DL) of nano-aggregates of **1** for Ag (I) was determined from the following equation:

$$DL = \frac{KS_{b1}}{S}$$

Where K = 3; S_{b1} is the standard deviation of the blank solution; *S* is the slope of the calibration curve. The detection limits of biogenic amines were also determined in a similar fashion.

S. No.	Mode of detection	Pretreatment	Application to real sample	Detectio n limit	Reference
1	UV	Pretreatment	Urine	0.06 µM	<i>J. Sep. Sci.</i> , 2009, 32 , 4143–4147.
2	Matrix -solid -phase- dispersion using HPLC -electrospray-tandem MS	Pretreatment	Cheese	0.06 mg/kg	<i>J. Agric. Food</i> <i>Chem.</i> 2005, 53 , 3779–3783.
3	UHPLC-MS/MS	Pretreatment	Anchovy (fish)	Range: 10-750 µg/L	<i>J. Agric. Food</i> <i>Chem.</i> 2012, 60 , 5324–5329
4	RP-HPLC coupled with fluorimetry	precolumn dansylation	Wines	0.04 mg/l	<i>Food Chem.</i> , 2008, 106 , 1218–1224
5	Electrochemical sensor based on MWCNT- gold nanoparticle composites	-	Yoghurt	57 nM	Food Res. Int., 2011, 44, 276–281.
6	Cyclic voltammetry using SWCNT	Pretreatment	Fish products	0.62 µM	<i>J. Food Eng.</i> , 2015, 149 , 1–8.
7	Absorption-based Chromogenic Sensing on filter paper	-	-	0.02 mM	<i>Anal. Chem.</i> 2010, 82 , 8402- 8405
8	Chameleon dye based, microtitre plate using fluorescence spectroscopy	Pretreatment	Fish samples	3.4 µM	<i>Analyst</i> , 2011, 136 , 4492–4499
9	Micellar liquid chromatography and pulsed amperometric	-	Wine	12ng/ml	<i>J. Chromatogr.</i> <i>A</i> , 2007, 1156 , 288–295.
10	Amperometry	Pretreatment	Sauerkraut	0.57 μM	<i>Sens. Actuators</i> <i>B</i> , 2013, 178 , 40–46
11	Fluorescence Spectroscopy-using easily-engineered nanomaterials	No	Milk and Wine	3.91 nM	Current study.

 Table S1: Comparison of current sensor with existing literature.