

A biomimetic magnetically recoverable Palladium nanocatalyst for Suzuki cross-coupling reaction

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Materials and Methods:

All reagents and starting materials were obtained commercially sources and were used without additional purification. Thin Layer Chromatography (TLC) was performed on silica (Silica Gel 60 F254) pre-coated aluminium plates and the products were visualized by UV lamp (PHILIPS TUV 8W lamp) and I₂ stain. X-ray powder diffraction (XRD) patterns were recorded on a (Shimadzu XRD-6100 using CuK radiations = 1.5405 Å) powder diffractometer instrument. Transmission Electron Microscopy studies were performed using a PHILIPS CM200 Transmission Electron Microscope (TEM) at 100 kV. ICP-AES (Inductively Coupled Plasma-Atomic Emission Spectrometer) were performed using ARCOS model instrument from M/s Spectro, Germany. The ¹H and ¹³C NMR was recorded in CDCl₃ using residual solvent peak as a reference on an Avance III and Bruker NMR spectrophotometer at 400 MHz and 100 MHz respectively.

Preparation of Fe₃O₄@Polydopamine (PDA) Core-Shell:

Fe₃O₄ nanoparticles (1 gram) were dispersed (pre-sonicated for 30 minutes) in 300 ml of de-ionized water and the pH was maintained at 8.5 by adding 10 mM Tris-HCl buffer. To this dopamine (1 gram) pre-dissolved in 200 mL of deionized water was added all at once. The solution is stirred by using a mechanical overhead stirrer fitted with a rod at room temperature for 24 hr. The polydopamine capped Fe₃O₄ nanoparticles were collected by using laboratory permanent magnet and washed 3 times with water, ethanol and acetone. The as prepared polydopamine capped Fe₃O₄ nanoparticles are dried overnight at room temperature in a desiccator.

Preparation of Pd/Fe₃O₄@PDA:

1 gram of Polydopamine capped Fe₃O₄ nanoparticles (Fe₃O₄@PDA) which were prepared by the above mentioned procedure was dispersed in 500 ml of de-ionized water by sonication for 30 min using an ultrasonication bath. To this, Na₂PdCl₄ solution [prepared by dissolving 90 mg (0.51mmol) of PdCl₂ and 60 mg (1.03 mmol) of NaCl in 15mL of water] was added. After stirring for 30 minutes, NaBH₄ solution [30 mg (0.79mmol) dissolved in 10

mL of deionized water] was slowly added dropwise and this mixture was stirred overnight at room temperature with the help of a mechanical overhead stirrer. The Pd immobilized $\text{Fe}_3\text{O}_4@\text{PDA}$ nanoparticles were collected magnetically by using laboratory permanent magnet. The particles were washed thrice with water, ethanol and acetone. The as prepared Pd/ $\text{Fe}_3\text{O}_4@\text{PDA}$ catalyst was dried overnight at room temperature in a desiccator and later completely characterized by spectroscopic and analytical techniques. The loading of Palladium was found to be 3.88wt% as revealed by ICP-AES analysis.

General experimental procedure for Suzuki reaction:

To a 25mL round bottomed flask (backfilled with nitrogen) equipped with a Teflon coated magnetic bar, aryl halide (0.5 mmol) and arylboronic acid (0.55 mmol) were added. To this, K_2CO_3 (1 mmol, 138mg) and Pd/ $\text{Fe}_3\text{O}_4@\text{PDA}$ (0.46 mol % of Pd) were added followed by 4mL of H_2O : EtOH (1:1). The reaction mixture was stirred for the indicated time at 80 °C for aryl iodides and 100°C for aryl bromides till the time taken for complete consumption of the reactants as confirmed by TLC, the solution was extracted four times with ethyl acetate (4×10 mL). The organic solvents were removed under pressure and the residue was column chromatographed using silica gel with petroleum ether/ethyl acetate as eluents to get the desired coupling product (most cases it was only filter column).

Catalyst recyclability study:

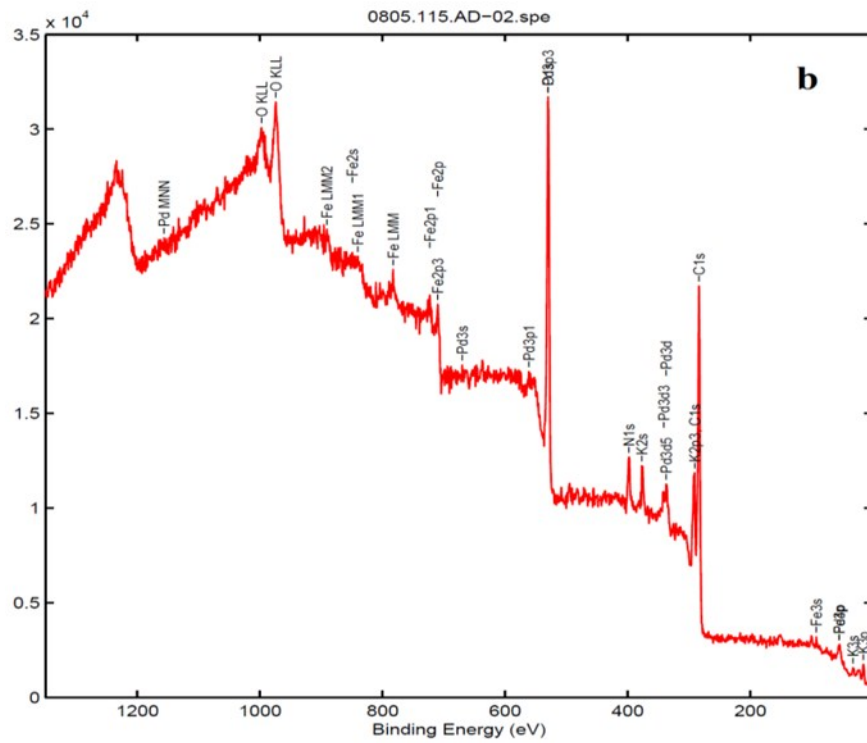
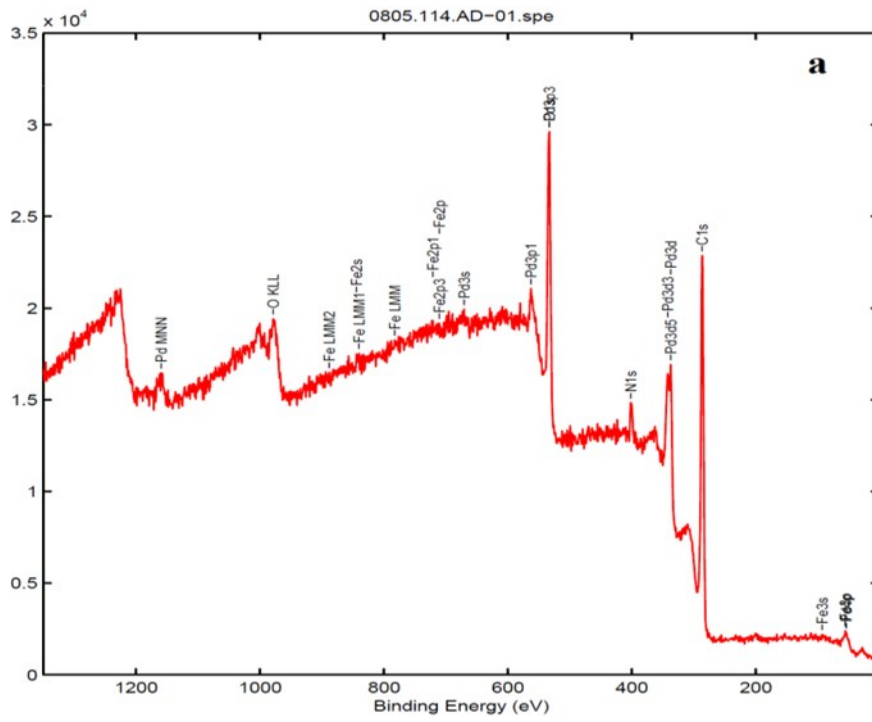
To a 25mL round bottomed flask (backfilled with nitrogen) equipped with a Teflon coated magnetic bar, aryl halide (0.5 mmol) and arylboronic acid (0.55 mmol) were added. To this, K_2CO_3 (1 mmol, 138mg) and Pd/ $\text{Fe}_3\text{O}_4@\text{PDA}$ (0.46 mol % of Pd) were added followed by 4mL of H_2O : EtOH (1:1). The reaction mixture was stirred for the indicated time at 80°C. After consumption of the reactants as confirmed by TLC, the solution was extracted four times with ethyl acetate (4×10 mL). The organic solvents were removed under pressure and the residue was column chromatographed using silica gel with petroleum ether/ethyl acetate as eluents to get the desired coupling products (most cases it was only filter column). The magnetically recovered catalyst was washed with water and acetone, dried in a desiccator overnight and subsequently used for the next cycle. As such the catalyst was reused for five more cycles.

Hot Test Experiment conditions under centrifugation conditions:

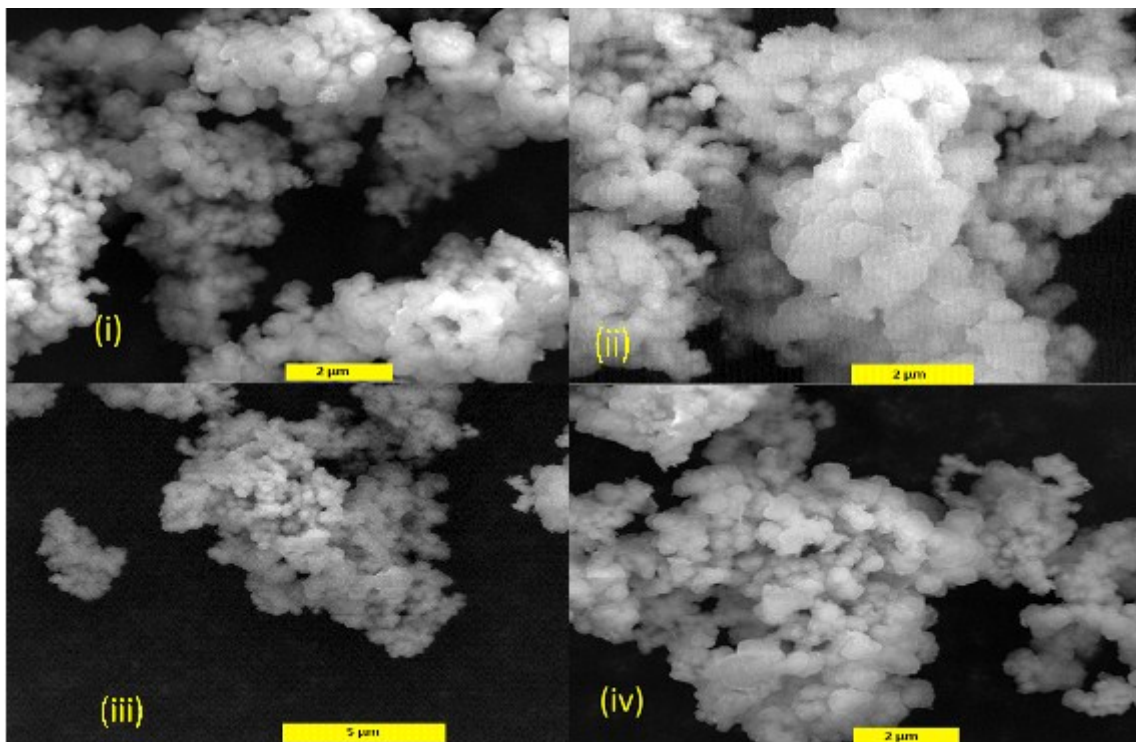
To a 25mL round bottomed flask (backfilled with nitrogen) equipped with a Teflon coated magnetic bar, aryl halide (0.5 mmol) and arylboronic acid (0.55 mmol) were added. To this, K_2CO_3 (1 mmol, 138mg) and Pd/Fe₃O₄@PDA (0.46 mol % of Pd) were added followed by 4mL of H₂O: EtOH (1:1). The reaction was stirred for 2 hours at 80 °C. The reaction mixture was centrifuged at 3000 rpm on an ultracentrifuge machine for 30 minutes and the solution was filtered with 0.44 μm, PTFE syringe filter and the afforded solution was transferred to another flask and reaction was allowed to stir at 80 °C for 24 hours.

Hot Test Experiment conditions under non-centrifugation conditions:

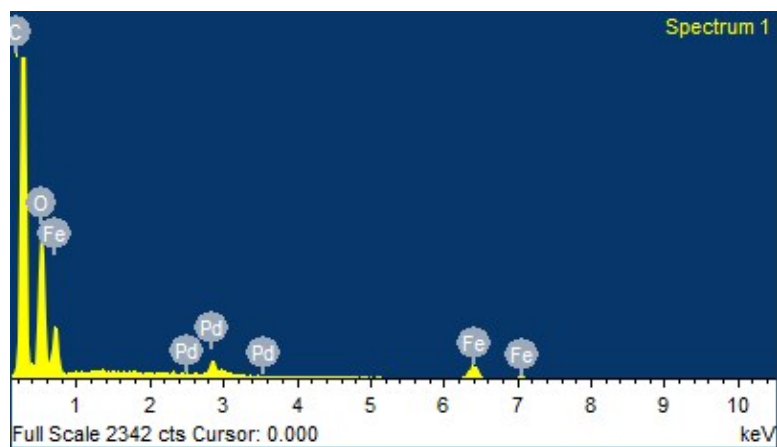
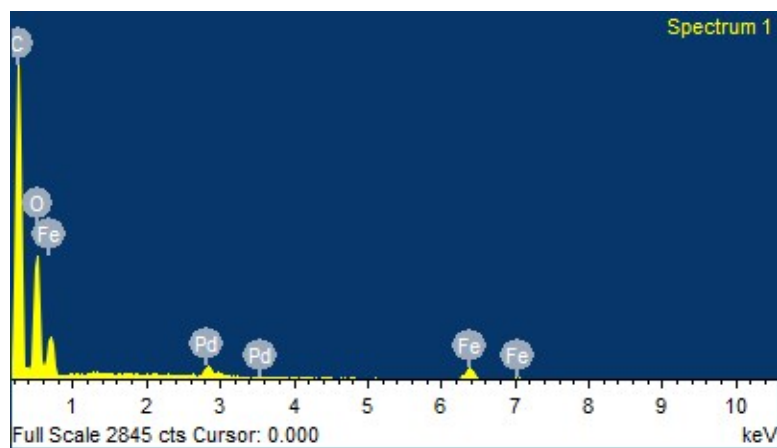
To a 25mL round bottomed flask (backfilled with nitrogen) equipped with a Teflon coated magnetic bar, aryl halide (0.5 mmol) and arylboronic acid (0.55 mmol) were added. To this, K_2CO_3 (1 mmol, 138mg) and Pd/Fe₃O₄@PDA (0.46 mol % of Pd) were added followed by 4mL of H₂O: EtOH (1:1). The reaction was stirred for 2 hours at 80 °C. The reaction mixture was filtered with 0.44 μm, PTFE syringe filter and the afforded solution was transferred to another flask and reaction was allowed to stir at 80 °C for 24 hours.



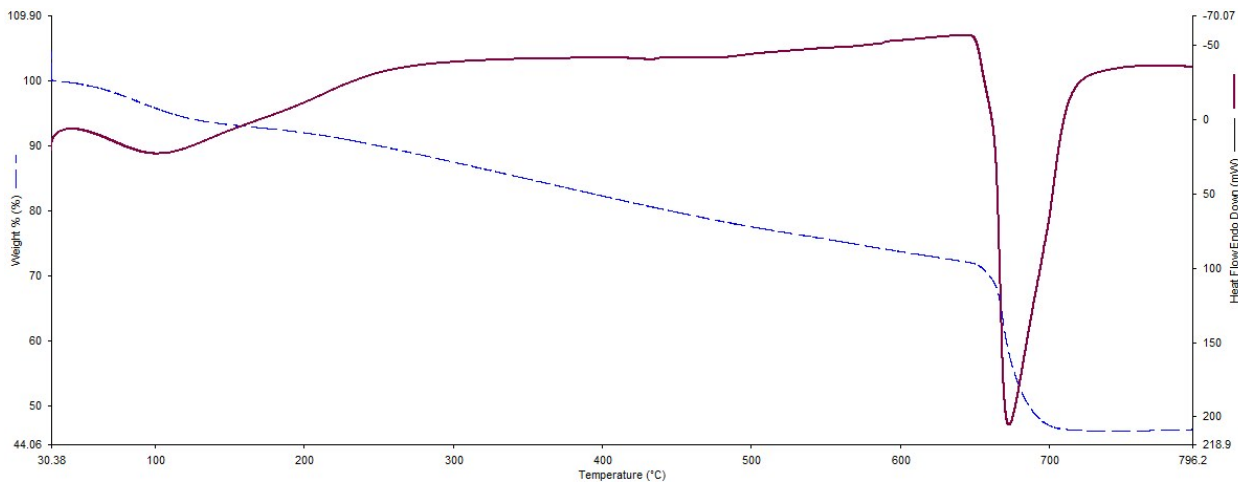
XPS of Pd/Fe₃O₄@PDA nanoparticles (a) native (b) after five cycles¹



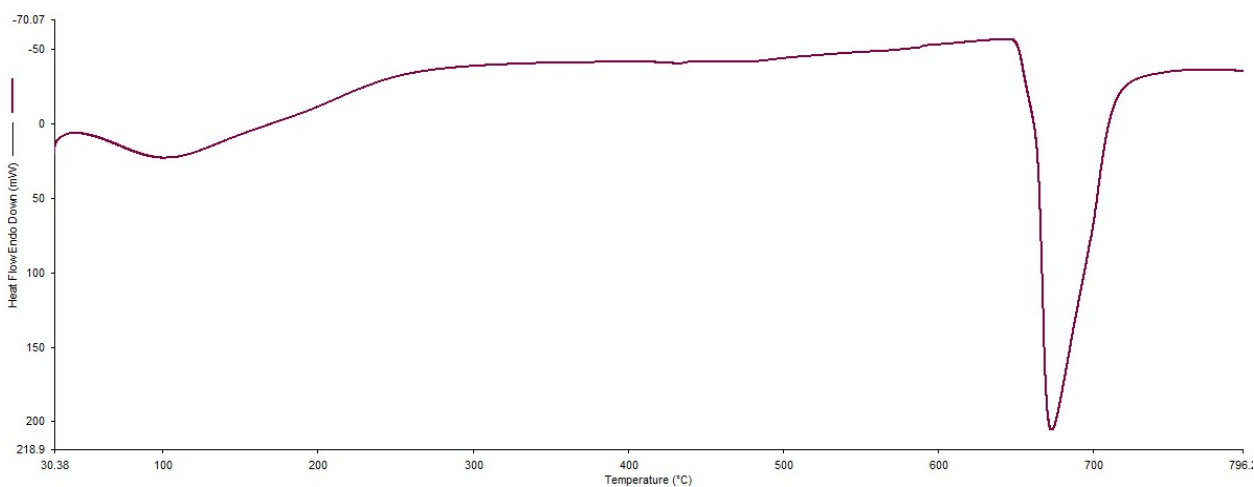
SEM images of Pd/PDA@Fe₃O₄: (i-ii) native (iii-iv) after five cycles.



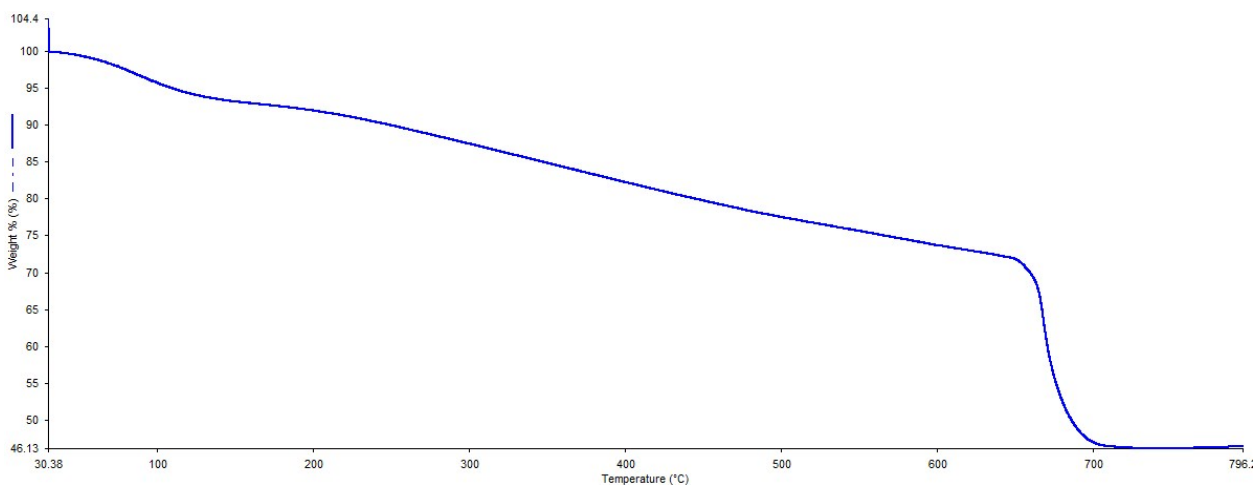
EDAX Spectra of native and recycled Pd/ Fe₃O₄@PDA



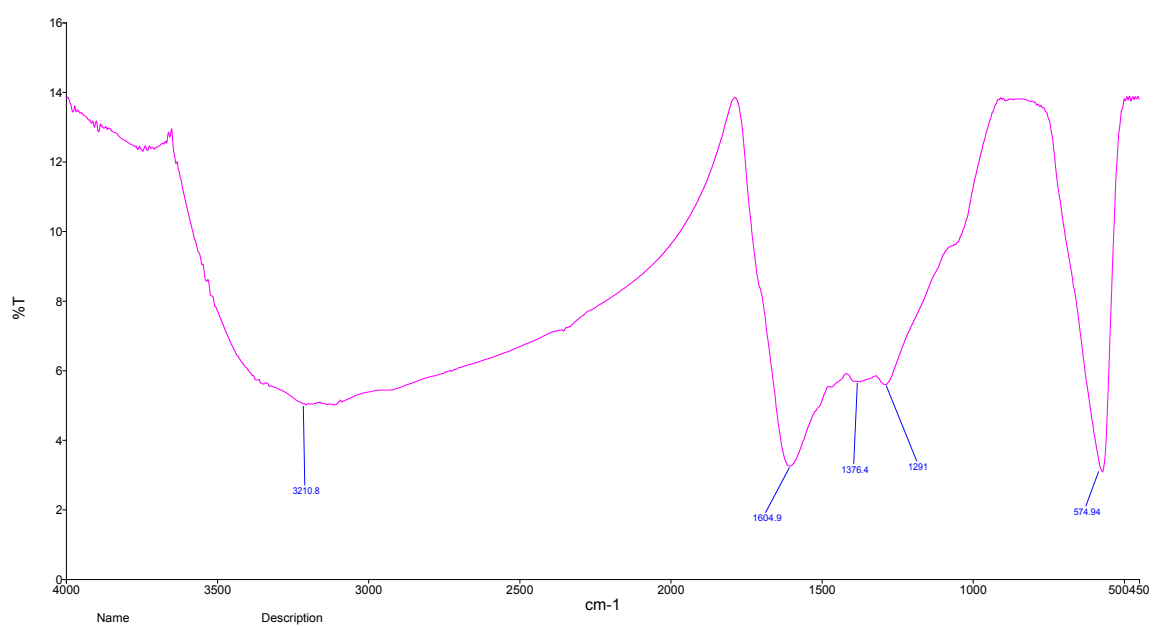
DSC TGA of Pd/Fe₃O₄@PDA native catalyst



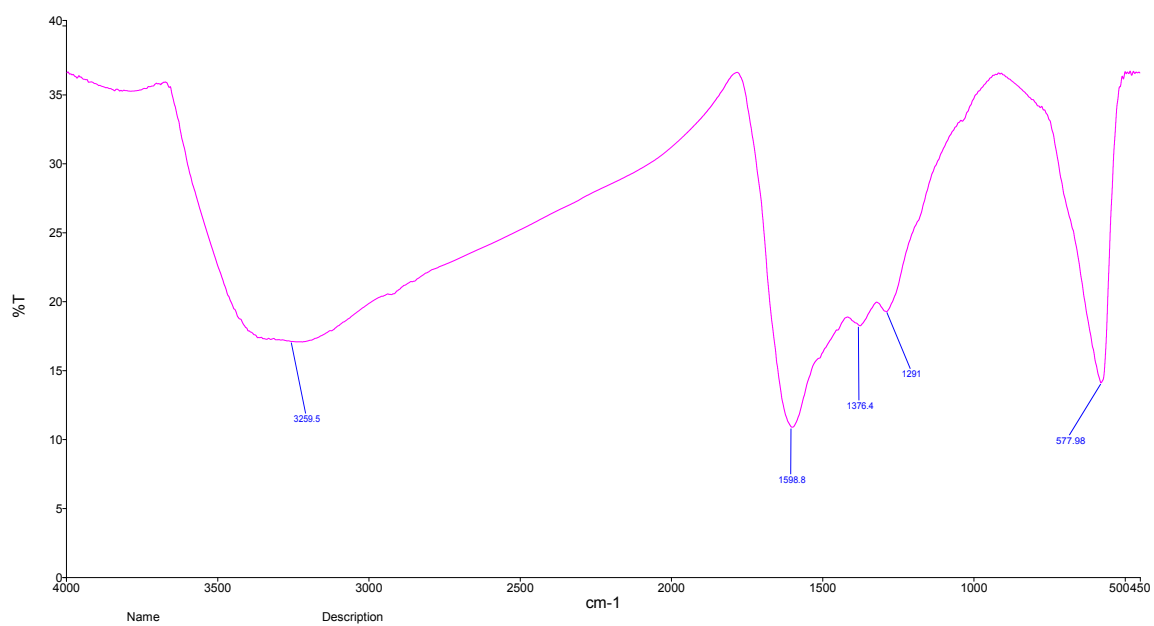
DSC of Pd/Fe₃O₄@PDA native catalyst



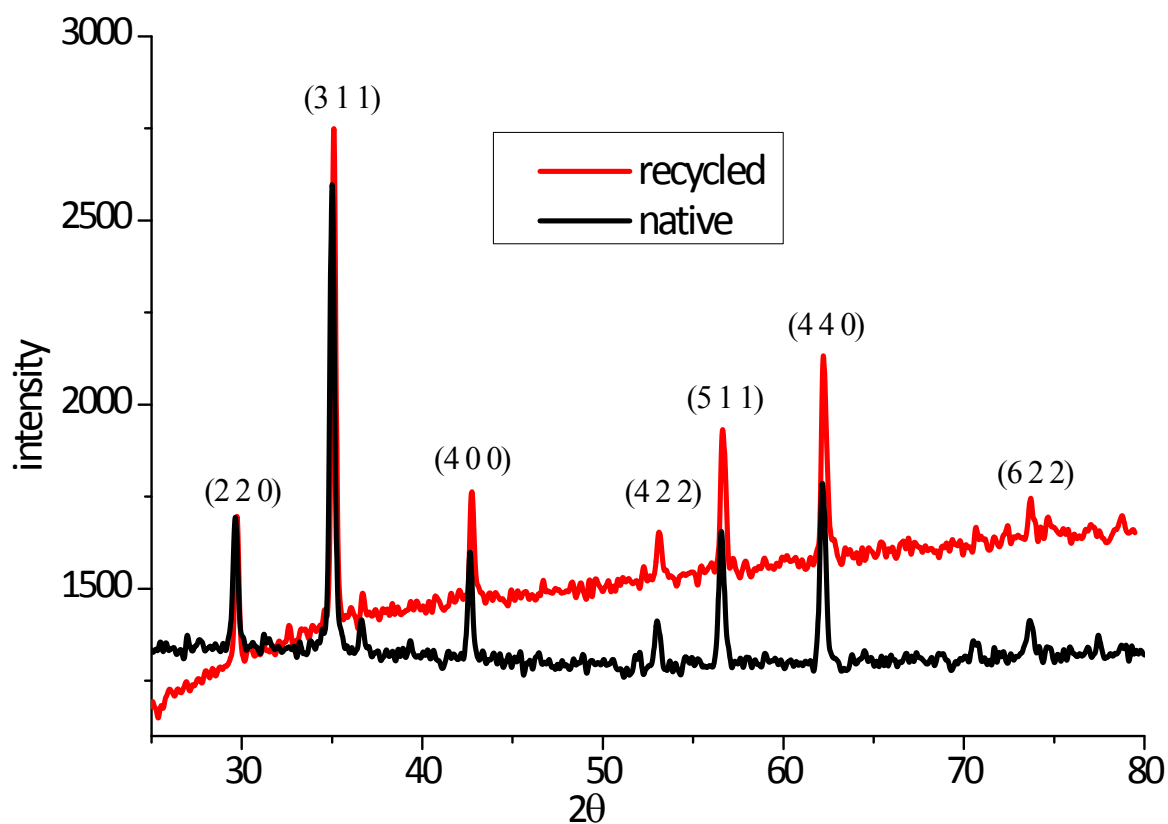
TGA of Pd/Fe₃O₄@PDA native catalyst



FT-IR of native Pd/Fe₃O₄@PDA catalyst



FT-IR of recycled Pd/Fe₃O₄@PDA catalyst

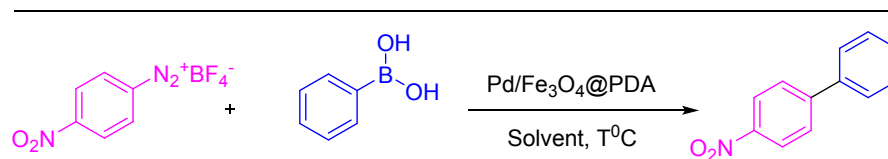


X-ray powder diffractogram of the fresh and reused Pd/Fe₃O₄@PDA nanoparticles

General experimental procedure for Suzuki reaction: (with aryldiazonium tetrafluoroborate salts)

To a 25mL round bottomed flask Teflon coated magnetic bar, aryl diazoniumtetrafluoroborate (0.5 mmol) and arylboronic acid (0.55 mmol) were added. To this 0.27 mol % of Pd were added followed by 4mL of Methanol. The reaction mixture was stirred for the indicated time at room temperature in air till the complete consumption of the reactants as confirmed by TLC. After completion of reaction 2ml of water added and solution was extracted four times with ethyl acetate (4 × 10 mL). The organic solvents were removed under pressure and the residue was column chromatographed using silica gel with petroleum ether/ethyl acetate as eluents to get the desired coupling product (most cases it was only filter column).

Table Optimization of reaction conditions of Suzuki cross-coupling of aryldiazonium salts with arylboronic acids by Pd/Fe₃O₄@PDA nanocatalyst^a



Entry	Solvent	Time (h)	Yield ^b (%)
1	CH ₃ CN	24	33
2	DMSO	24	51
3	DMF	24	46
4	CH ₂ Cl ₂	24	28
5	CHCl ₃	24	32
6	NMP	24	38
7	THF	24	24
8	1,4-dioxane	24	21
9	H ₂ O	24	10
10	H ₂ O	24	10 ^c
11	EtOH:H ₂ O(1:1)	24	26
12	EtOH	3	15
13	EtOH	24	63
14	MeOH	3	85
15	MeOH	24	85
16	MeOH	24	60 ^d
17	MeOH	24	0 ^e

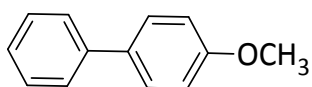
^aReaction conditions: phenylboronic acid (0.55 mmol), arenediazonium salts (0.5 mmol), Pd/Fe₃O₄@PDA (0.27 mol % of Pd).

^bisolated yield.

^creaction temperature 60 °C

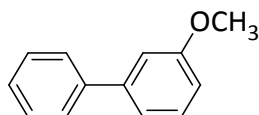
^dPd/Fe₃O₄@PDA (0.18 mol % of Pd)

^ewithout catalyst



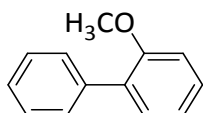
4-Methoxy-1,1'-biphenyl (1a)²

¹H NMR (400 MHz, CDCl₃) δ 7.63 – 7.56 (m, 4H), 7.50 – 7.44 (m, 2H), 7.39 – 7.33 (m, 1H), 7.06 – 7.01 (m, 2H), 3.90 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 159.19, 140.87, 133.82, 128.77, 128.20, 126.78, 126.71, 114.25, 55.37.



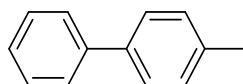
3-Methoxy-1,1'-biphenyl (1b)²

¹H NMR (400 MHz, CDCl₃) δ 7.70 – 7.63 (m, 2H), 7.54 – 7.37 (m, 4H), 7.25 (m, *J* = 17.0, 12.6, 11.9 Hz, 2H), 6.97 (dd, *J* = 8.2, 2.6 Hz, 1H), 3.92 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 160.01, 142.83, 141.17, 129.80, 128.78, 127.46, 127.25, 119.74, 112.97, 112.74, 55.33.



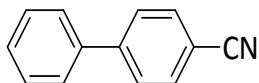
2-Methoxy-1,1'-biphenyl (1c)²

¹H NMR (400 MHz, CDCl₃) δ 7.61 – 7.56 (m, 2H), 7.46 (t, *J* = 7.6 Hz, 2H), 7.37 (m, *J* = 7.0, 4.4, 2.5 Hz, 3H), 7.12 – 7.01 (m, 2H), 3.86 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 156.48, 138.56, 130.89, 130.75, 129.55, 128.61, 127.97, 126.91, 120.84, 111.27, 55.56.



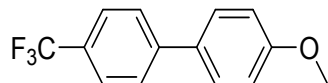
4-Methyl-1,1'-biphenyl (1d)²

¹H NMR (400 MHz, CDCl₃) δ 7.72 – 7.63 (m, 2H), 7.57 (t, *J* = 7.4 Hz, 2H), 7.78 – 7.23 (m, 2H), 7.54 – 7.47 (m, 1H), 7.44 – 7.37 (m, 2H), 2.48 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 141.24, 138.43, 137.07, 129.55, 128.78, 127.06, 21.16.



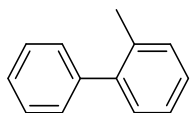
[1,1'-Biphenyl]-4-carbonitrile (1e)³

¹H NMR (400 MHz, CDCl₃) δ 7.73 (m, *J* = 8.5, 4.2 Hz, 4H), 7.62 (dd, *J* = 5.2, 3.3 Hz, 2H), 7.55 – 7.38 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 145.70, 139.20, 132.61, 129.13, 128.68, 127.75, 127.24, 118.95, 110.95.



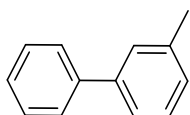
4-Methoxy-4'-(trifluoromethyl)-1,1'-biphenyl (1f) ⁴

¹H NMR (400 MHz, CDCl₃) δ 7.64 (m, 4H), 7.53 (d, J = 8.7 Hz, 2H), 6.99 (d, J = 8.7 Hz, 2H), 3.85 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 159.81, 144.24, 132.14, 128.30, 126.82, 125.68, 125.64, 125.60, 114.39, 55.34.



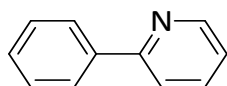
2-Methyl-1,1'-biphenyl (1g) ²

¹H NMR (400 MHz, CDCl₃) δ 7.51 – 7.25 (m, 9H), 2.33 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 142.00, 141.97, 135.37, 130.33, 129.82, 129.22, 128.78, 128.09, 127.27, 127.20, 126.78, 125.78, 20.49.



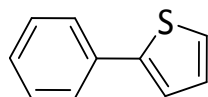
3-Methyl-1,1'-biphenyl (1h) ²

¹H NMR (400 MHz, CDCl₃) δ 7.67 – 7.60 (m, 2H), 7.42 (m, J = 10.6, 7.4, 1.5 Hz, 6H), 7.21 (d, J = 7.6 Hz, 1H), 2.47 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 141.37, 141.25, 138.32, 128.69, 128.66, 127.99, 127.18, 124.27, 21.55.



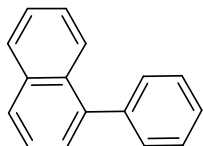
2-Phenylpyridine (1i) ⁵

¹H NMR (500 MHz, CDCl₃) δ 8.72 – 8.67 (m, 1H), 8.02 – 7.96 (m, 2H), 7.79 – 7.69 (m, 2H), 7.51 – 7.38 (m, 3H), 7.27 – 7.18 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 157.49, 149.69, 139.43, 136.76, 128.97, 128.77, 126.94, 122.12, 120.60.



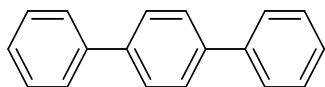
2-Phenylthiophene (1j) ⁶

¹H NMR (400 MHz, CDCl₃) δ 7.69 – 7.62 (m, 2H), 7.46 – 7.27 (m, 5H), 7.12 (dd, J = 5.0, 3.6 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 144.47, 134.44, 128.91, 128.14, 127.48, 125.88, 124.82, 123.10.



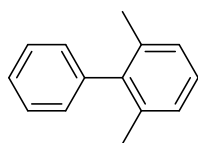
1-Phenylnaphthalene (1k) ⁶

¹H NMR (400 MHz, CDCl₃) δ 8.00 – 7.86 (m, 3H), 7.61 – 7.44 (m, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 140.79, 140.29, 133.82, 131.65, 130.10, 128.28, 127.65, 127.26, 126.95, 126.04, 125.78, 125.40.



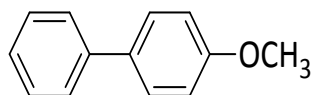
1,1':4',1''-Terphenyl (1l) ⁷

¹H NMR (500 MHz, CDCl₃) δ 7.69 – 7.62 (m, 8H), 7.49 – 7.43 (m, 4H), 7.39 – 7.34 (m, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 140.73, 140.15, 128.84, 127.53, 127.37, 127.08.



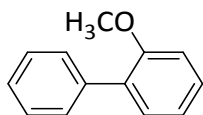
2,6-dimethyl-1,1'-biphenyl (1m) ¹¹

¹H NMR (400 MHz, CDCl₃) δ 7.43 (m, *J* = 45.4, 27.3, 7.7 Hz, 6H), 6.98 (d, *J* = 8.8 Hz, 2H), 3.84 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 159.10, 140.79, 133.74, 128.70, 128.13, 126.71, 126.63, 114.17, 55.32.



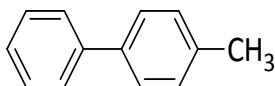
4-Methoxy-1,1'-biphenyl (1n) ²

¹H NMR (400 MHz, CDCl₃) δ 7.66 – 7.55 (m, 4H), 7.47 (m, *J* = 7.6, 3.5 Hz, 2H), 7.36 (m, *J* = 10.9, 5.6, 2.6 Hz, 1H), 7.03 (dd, *J* = 8.7, 3.2 Hz, 2H), 3.90 (s, *J* = 1.8 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 159.20, 140.87, 133.82, 128.77, 128.19, 126.78, 126.70, 114.26, 55.37.



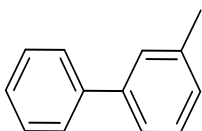
2-Methoxy-1,1'-biphenyl (1o) ²

¹H NMR (400 MHz, CDCl₃) δ 7.62 – 7.56 (m, 2H), 7.47 (dd, *J* = 10.3, 4.7 Hz, 2H), 7.38 (m, *J* = 7.6, 4.2, 1.1 Hz, 3H), 7.14 – 7.01 (m, 2H), 3.86 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 156.47, 138.54, 130.88, 130.74, 129.53, 128.59, 127.96, 126.89, 120.82, 111.25, 55.55.



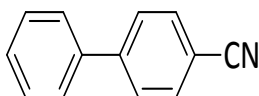
4-Methyl-1,1'-biphenyl (1p)²

¹H NMR (400 MHz, CDCl₃) δ 7.69 – 7.25 (m, 9H), 2.45 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 141.16, 138.36, 137.03, 129.50, 128.73, 126.99, 21.13.



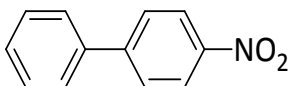
3-Methyl-1,1'-biphenyl (1q)²

¹H NMR (400 MHz, CDCl₃) δ 7.66 – 7.58 (m, 2H), 7.42 (m, *J* = 29.7, 11.1, 4.6 Hz, 6H), 7.20 (d, *J* = 7.3 Hz, 1H), 2.46 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 141.39, 141.27, 138.34, 128.71, 128.68, 128.00, 127.18, 124.30, 21.57.



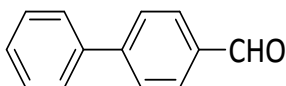
[1,1'-Biphenyl]-4-carbonitrile (1r)³

¹H NMR (400 MHz, CDCl₃) δ 7.79 – 7.68 (m, 4H), 7.66 – 7.57 (m, 2H), 7.56 – 7.42 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 145.70, 139.20, 132.61, 129.13, 128.68, 127.75, 127.25, 118.96, 110.94.



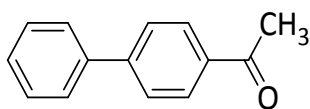
4-Nitro-1,1'-biphenyl (1s)³

¹H NMR (400 MHz, CDCl₃) δ 8.32 (d, *J* = 7.0 Hz, 2H), 7.76 (d, *J* = 7.2 Hz, 2H), 7.64 (t, *J* = 9.7 Hz, 2H), 7.57 – 7.43 (m, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 147.64, 147.11, 138.78, 129.18, 128.95, 127.81, 127.40, 124.12.



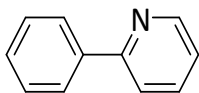
[1,1'-Biphenyl]-4-carbaldehyde (1t)⁵

¹H NMR (400 MHz, CDCl₃) δ 10.09 (s, 1H), 7.98 (d, *J* = 8.2 Hz, 2H), 7.78 (d, *J* = 8.3 Hz, 2H), 7.67 (dd, *J* = 5.3, 3.4 Hz, 2H), 7.55 – 7.41 (m, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 191.89, 147.19, 139.71, 135.21, 130.25, 129.01, 128.46, 127.68, 127.36.



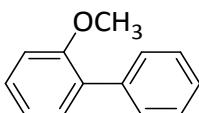
1-([1,1'-Biphenyl]-4-yl)ethanone (1u) ⁸

¹H NMR (400 MHz, CDCl₃) δ 8.11 – 8.02 (m, 2H), 7.76 – 7.62 (m, 4H), 7.54 – 7.39 (m, 3H), 2.67 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 197.76, 145.80, 139.90, 135.88, 128.97, 128.93, 128.25, 127.29, 127.24, 26.67.



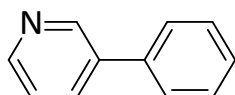
2-Phenylpyridine (1v) ⁵

¹H NMR (400 MHz, CDCl₃) δ 8.73 (d, J = 4.8 Hz, 1H), 8.03 (d, J = 7.2 Hz, 2H), 7.82 – 7.72 (m, 2H), 7.58 – 7.39 (m, 3H), 7.25 (m, J = 8.2, 7.4, 5.5 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 157.51, 149.81, 149.59, 139.45, 136.87, 136.60, 128.86, 128.66, 127.08, 126.79, 122.11, 121.75, 120.72, 120.58, 120.44.



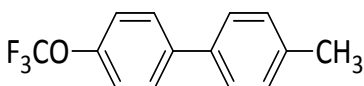
2-Methoxy-1,1'-biphenyl (1w) ²

¹H NMR (400 MHz, CDCl₃) δ 7.60 – 7.54 (m, 2H), 7.45 (t, J = 7.5 Hz, 2H), 7.40 – 7.33 (m, 3H), 7.11 – 6.99 (m, 2H), 3.85 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 156.47, 138.55, 130.89, 130.74, 129.54, 128.60, 127.96, 126.90, 120.83, 111.25, 55.56.



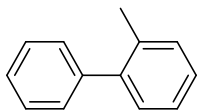
3-Phenylpyridine (1x) ⁹

¹H NMR (400 MHz, CDCl₃) δ 8.88 (s, 1H), 8.62 (d, J = 3.7 Hz, 1H), 7.94 – 7.86 (m, 1H), 7.66 – 7.35 (m, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 148.54, 148.39, 148.30, 148.21, 137.85, 136.71, 134.56, 134.28, 129.20, 128.11, 127.28, 127.05, 123.60.



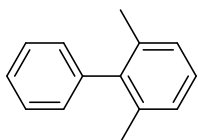
4-Methyl-4'-(trifluoromethoxy)-1,1'-biphenyl (1y) ¹⁰

¹H NMR (400 MHz, CDCl₃) δ 7.62 (d, J = 8.7 Hz, 2H), 7.50 (d, J = 8.1 Hz, 2H), 7.30 (dd, J = 8.0, 3.9 Hz, 4H), 2.44 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 148.48, 139.95, 137.55, 137.00, 129.63, 128.24, 126.96, 121.22, 21.10.



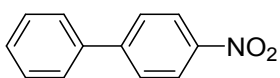
2-methyl-1,1'-biphenyl (1z)²

¹H NMR (400 MHz, CDCl₃) δ 7.48 – 7.23 (m, 9H), 2.30 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 141.95, 141.92, 135.33, 130.30, 129.79, 129.19, 128.06, 127.24, 126.75, 125.75, 20.48.



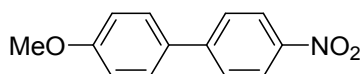
2,6-dimethyl-1,1'-biphenyl (1aa)¹¹

¹H NMR (400 MHz, CDCl₃) δ 7.51 – 7.19 (m, 6H), 6.65 (d, *J* = 8.4 Hz, 2H), 3.72 (s, 6H). ¹³C NMR (101 MHz, CDCl₃) δ 157.63, 134.09, 130.86, 128.61, 127.64, 126.74, 104.16, 55.89.



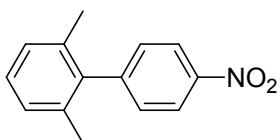
4-nitro-1,1'-biphenyl (1ab)¹²

¹H NMR (400 MHz, CDCl₃) δ 8.28 (d, *J* = 8.6 Hz, 2H), 7.78 – 7.37 (m, 7H). ¹³C NMR (101 MHz, CDCl₃) δ 147.59, 147.03, 138.72, 129.12, 128.89, 127.76, 127.35, 124.07.



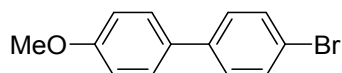
4-methoxy-4'-nitro-1,1'-biphenyl (1ac)¹²

¹H NMR (400 MHz, CDCl₃) δ 8.25 (d, *J* = 8.3 Hz, 2H), 7.61 (dd, *J* = 43.6, 8.2 Hz, 4H), 7.00 (d, *J* = 8.2 Hz, 2H), 3.85 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 160.39, 147.17, 146.49, 131.03, 128.53, 127.03, 124.11, 114.56, 55.40.



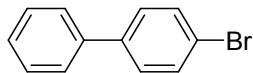
2,6-dimethyl-4'-nitro-1,1'-biphenyl (1ad)¹²

¹H NMR (400 MHz, CDCl₃) δ 8.31 (d, *J* = 9.0 Hz, 2H), 7.85 (dd, *J* = 55.9, 9.0 Hz, 3H), 6.56 (dd, *J* = 11.7, 5.5 Hz, 2H), 4.01 (s, 3H), 3.89 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 147.89, 145.67, 139.26, 136.83, 130.90, 128.80, 127.44, 123.06, 20.51.



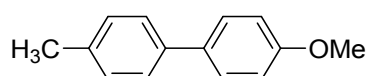
4-bromo-4'-methoxy-1,1'-biphenyl (1ae)¹³

¹H NMR (400 MHz, CDCl₃) δ 7.55 – 7.35 (m, 6H), 6.95 (d, J = 8.6 Hz, 2H), 3.83 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 159.35, 139.69, 132.44, 131.75, 128.26, 127.95, 120.75, 114.29, 55.34.



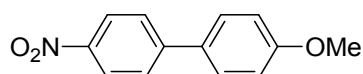
4-bromo-1,1'-biphenyl (1af)¹⁴

¹H NMR (400 MHz, CDCl₃) δ 7.63 – 7.30 (m, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 139.95, 139.63, 133.34, 128.88, 128.85, 128.36, 127.56, 126.96.



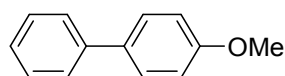
4-methoxy-4'-methyl-1,1'-biphenyl (1ag)¹⁵

¹H NMR (400 MHz, CDCl₃) δ 7.58 – 7.42 (m, 4H), 7.23 (d, J = 7.8 Hz, 2H), 7.02 – 6.91 (m, 2H), 3.84 (s, 3H), 2.39 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 158.91, 137.94, 136.33, 133.72, 129.42, 127.93, 126.56, 114.14, 55.31, 21.04.



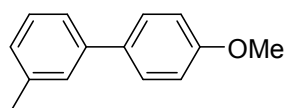
4-methoxy-4'-nitro-1,1'-biphenyl (1ah)¹²

¹H NMR (400 MHz, CDCl₃) δ 8.24 (d, J = 8.4 Hz, 2H), 7.61 (dd, J = 43.2, 8.5 Hz, 4H), 6.99 (d, J = 8.5 Hz, 2H), 3.85 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 160.39, 147.16, 146.48, 131.01, 128.53, 127.02, 124.11, 114.56, 55.40.



4-methoxy-1,1'-biphenyl (1ai)¹²

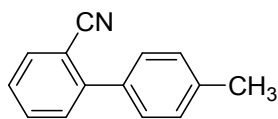
¹H NMR (400 MHz, CDCl₃) δ 7.43 (m, J = 45.4, 27.3, 7.7 Hz, 7H), 6.98 (d, J = 8.8 Hz, 2H), 3.84 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 159.10, 140.79, 133.74, 128.70, 128.13, 126.71, 126.63, 114.17, 55.32.



4-methoxy-3-methyl-1,1'-biphenyl (1aj)¹⁶

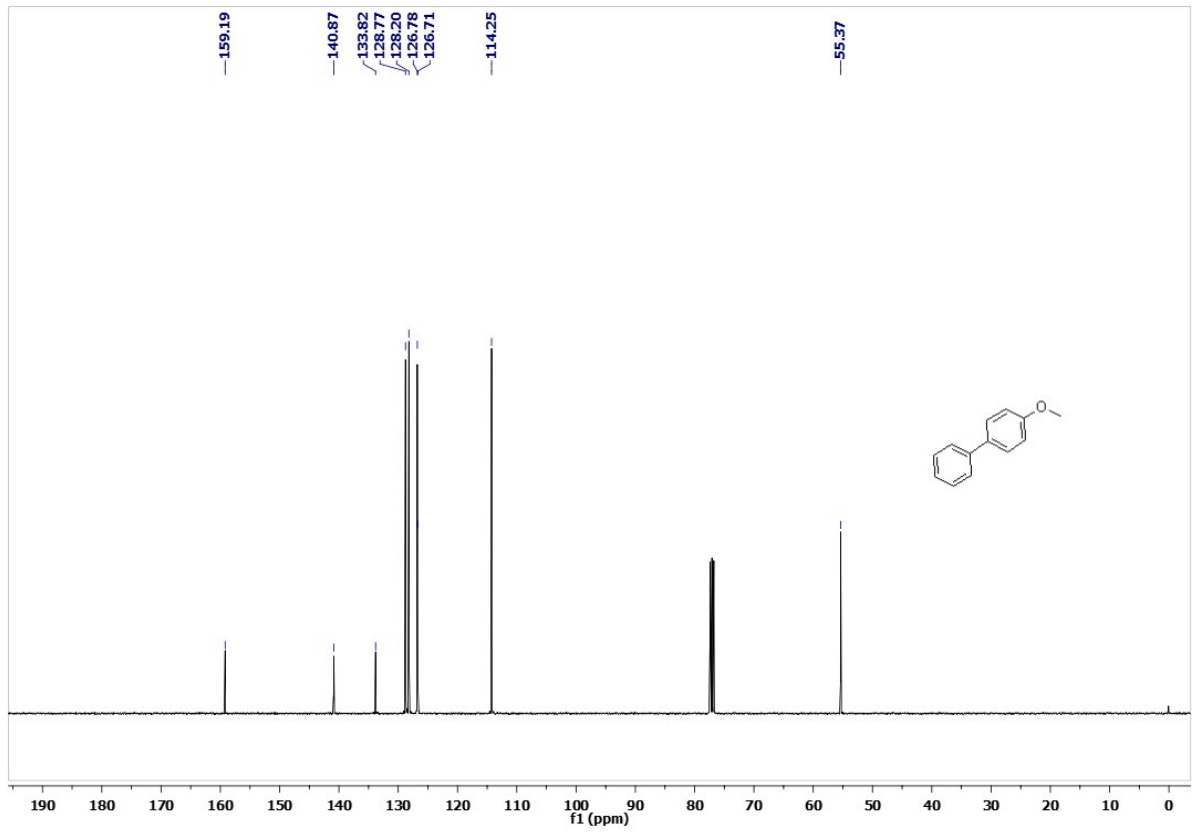
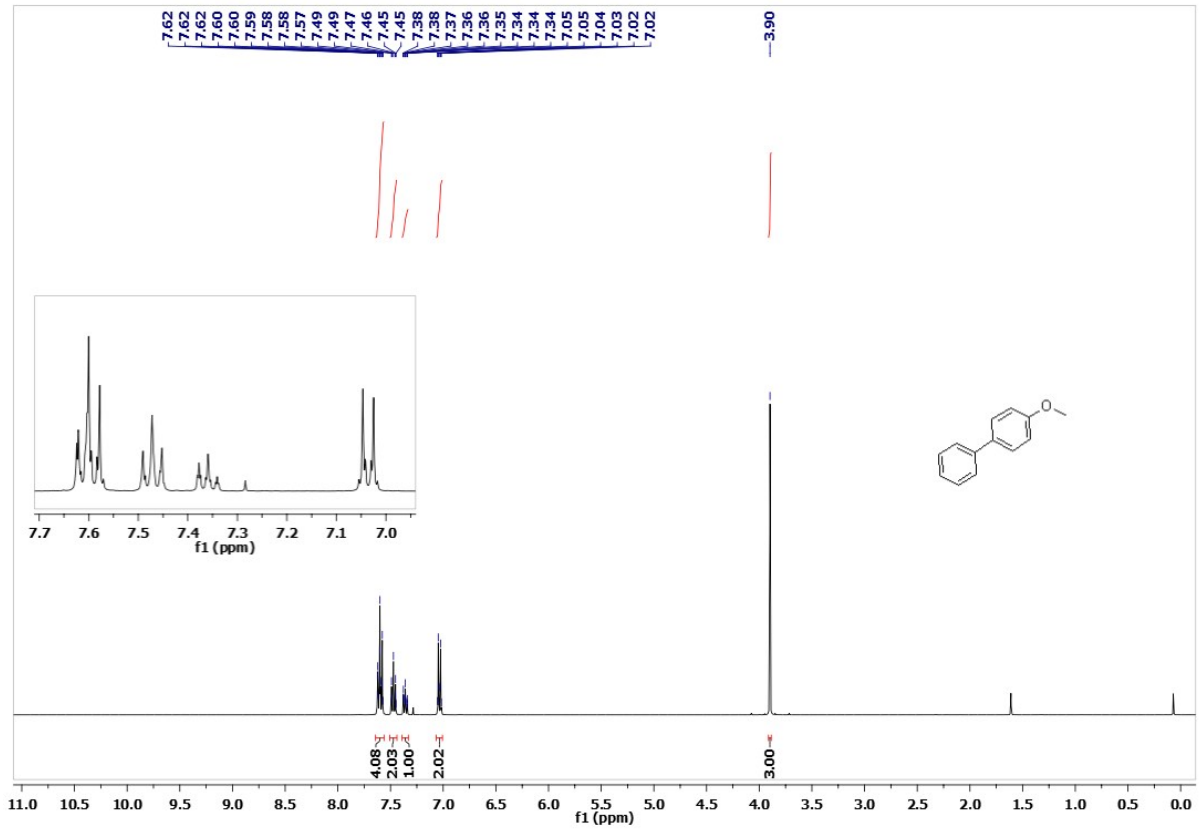
¹H NMR (400 MHz, CDCl₃) δ 7.52 (d, J = 8.5 Hz, 2H), 7.39 – 7.27 (m, 3H), 7.12 (d, J = 7.0 Hz, 1H),

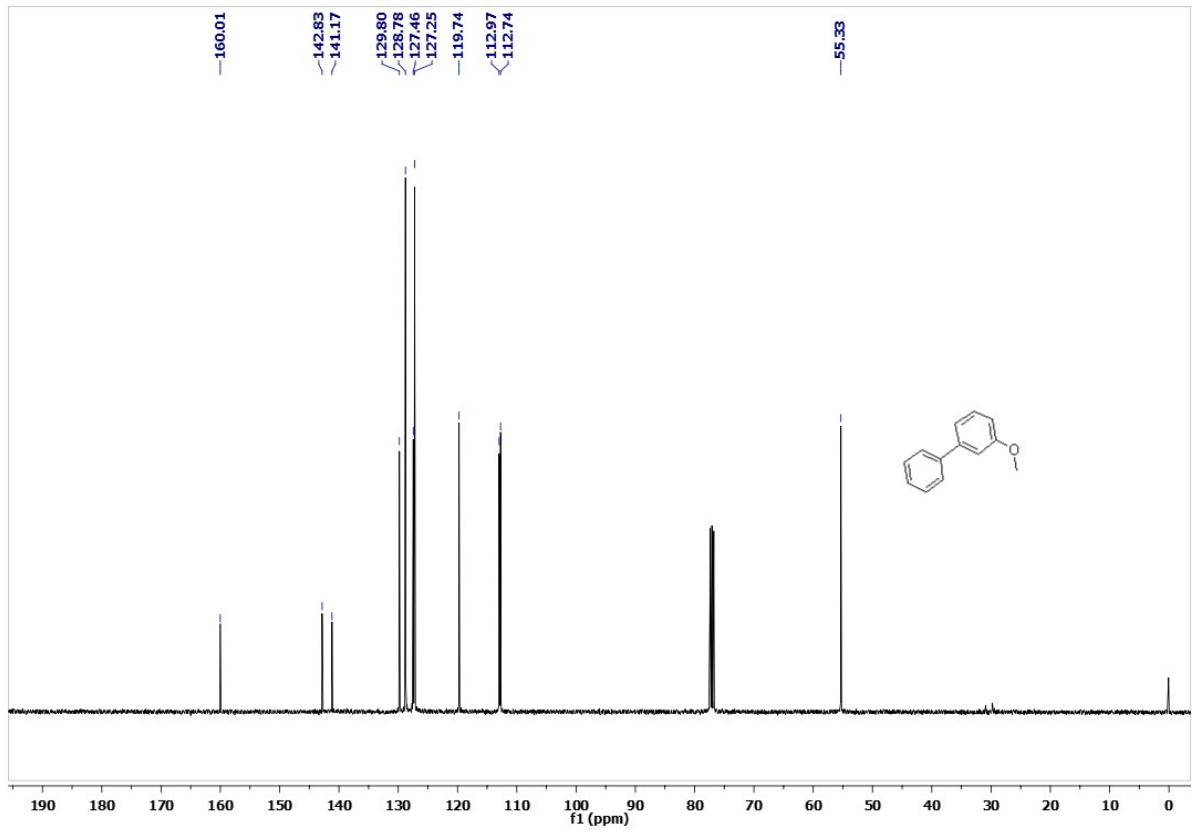
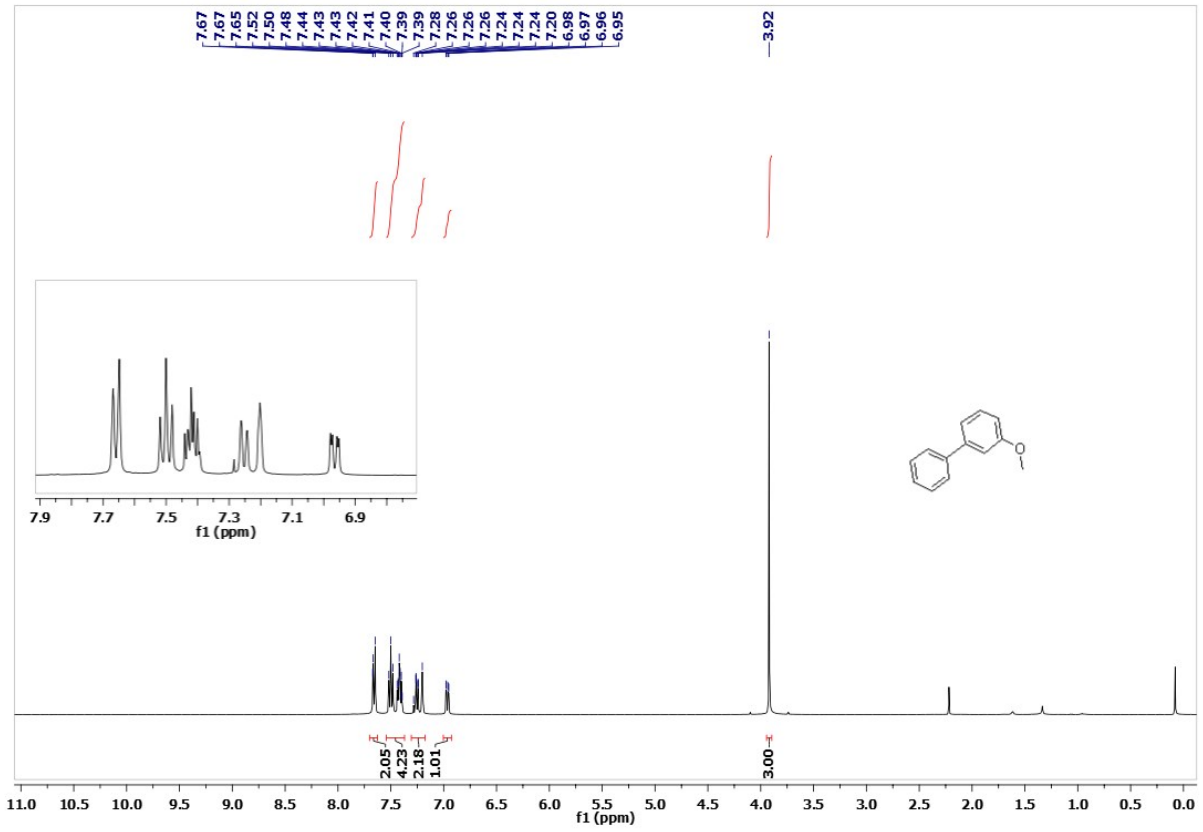
6.96 (d, $J = 8.5$ Hz, 2H), 3.84 (s, 3H), 2.41 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 159.03, 140.78, 138.24, 133.86, 128.60, 128.13, 127.53, 127.39, 123.82, 114.10, 55.31, 21.54.

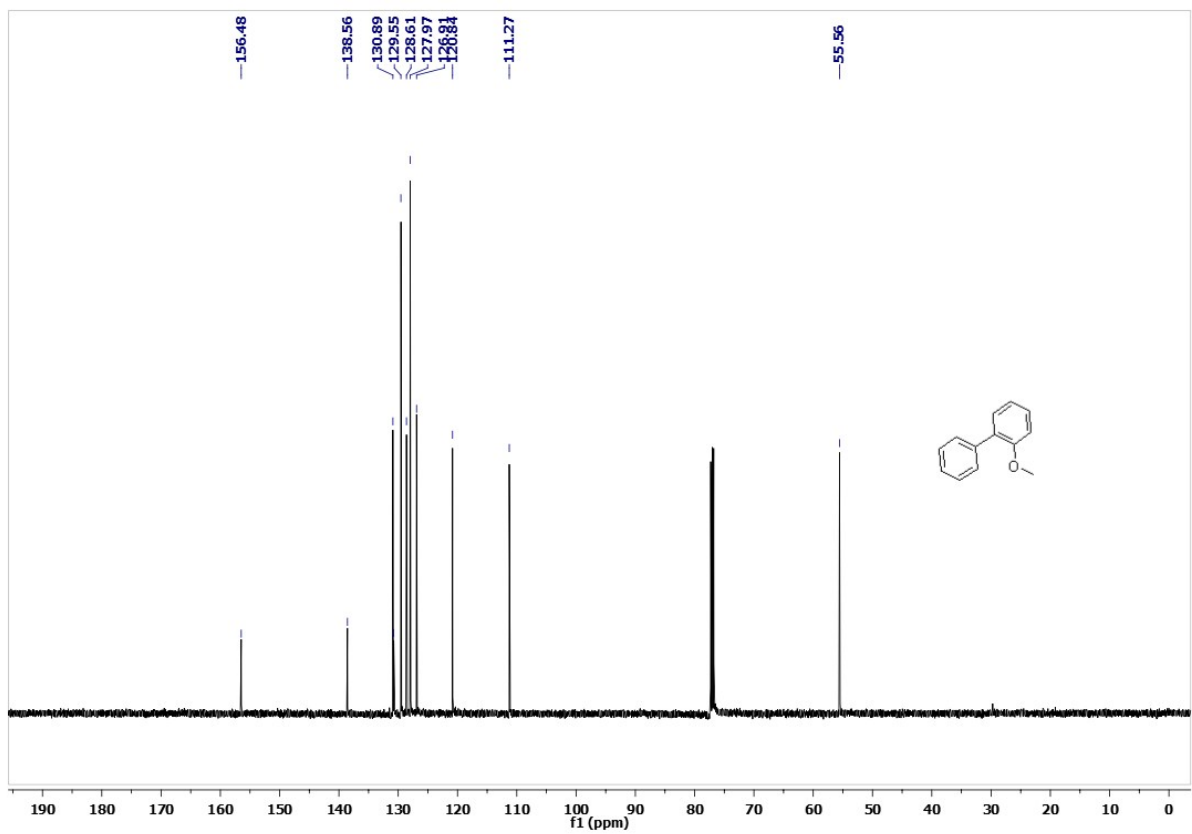
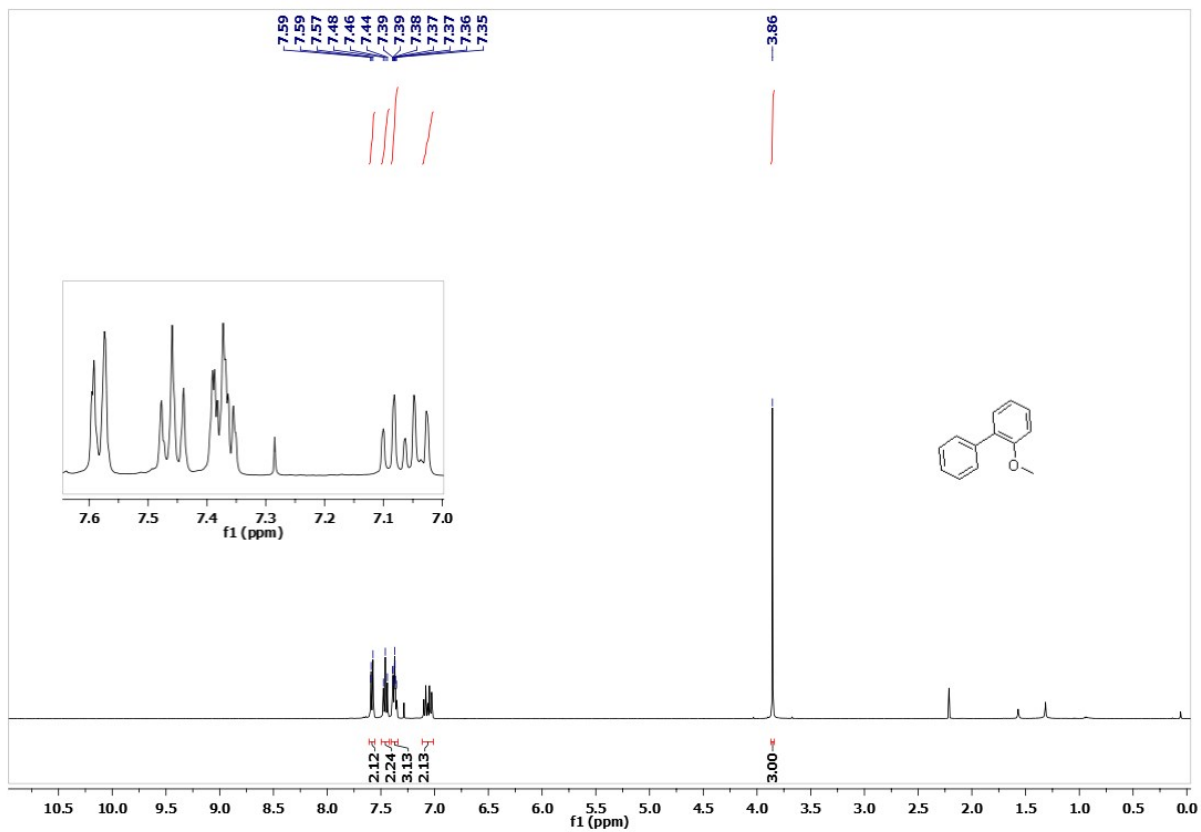


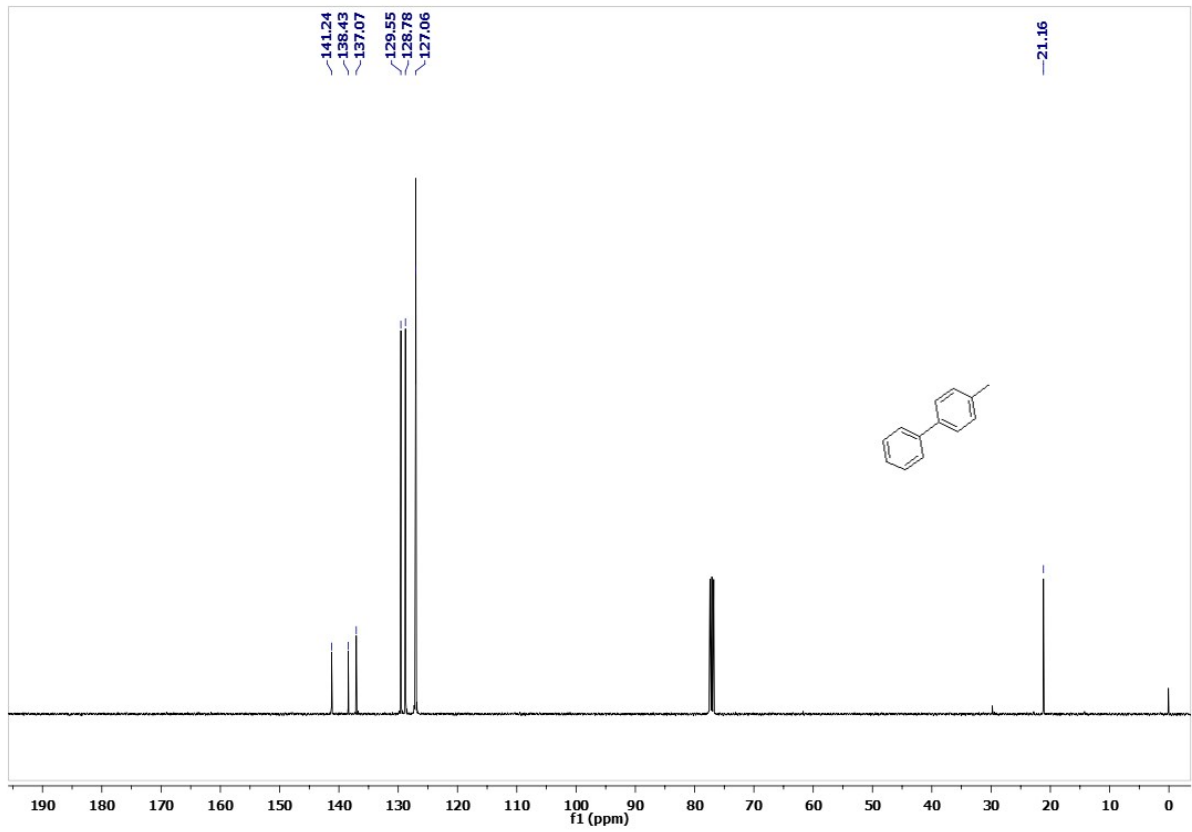
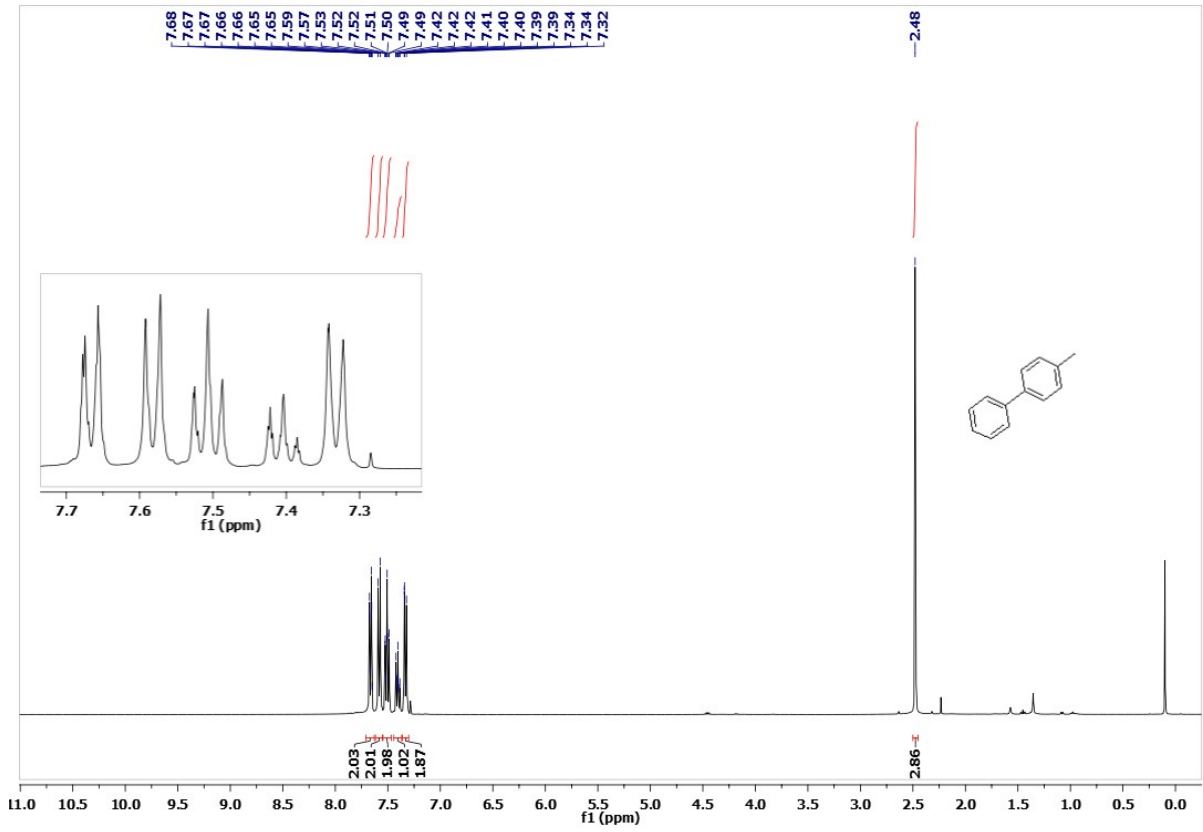
4'-Methyl-[1, 1'-biphenyl]-2-carbonitrile (1ak)¹⁰

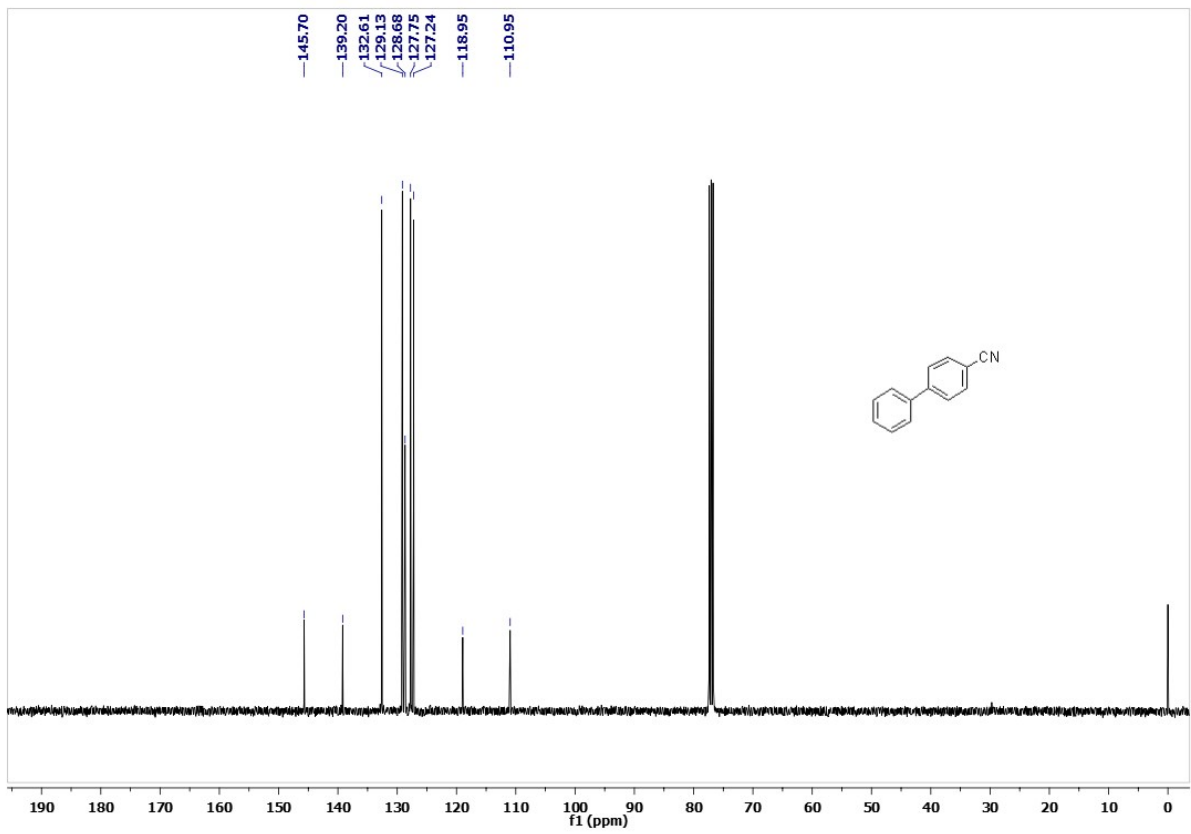
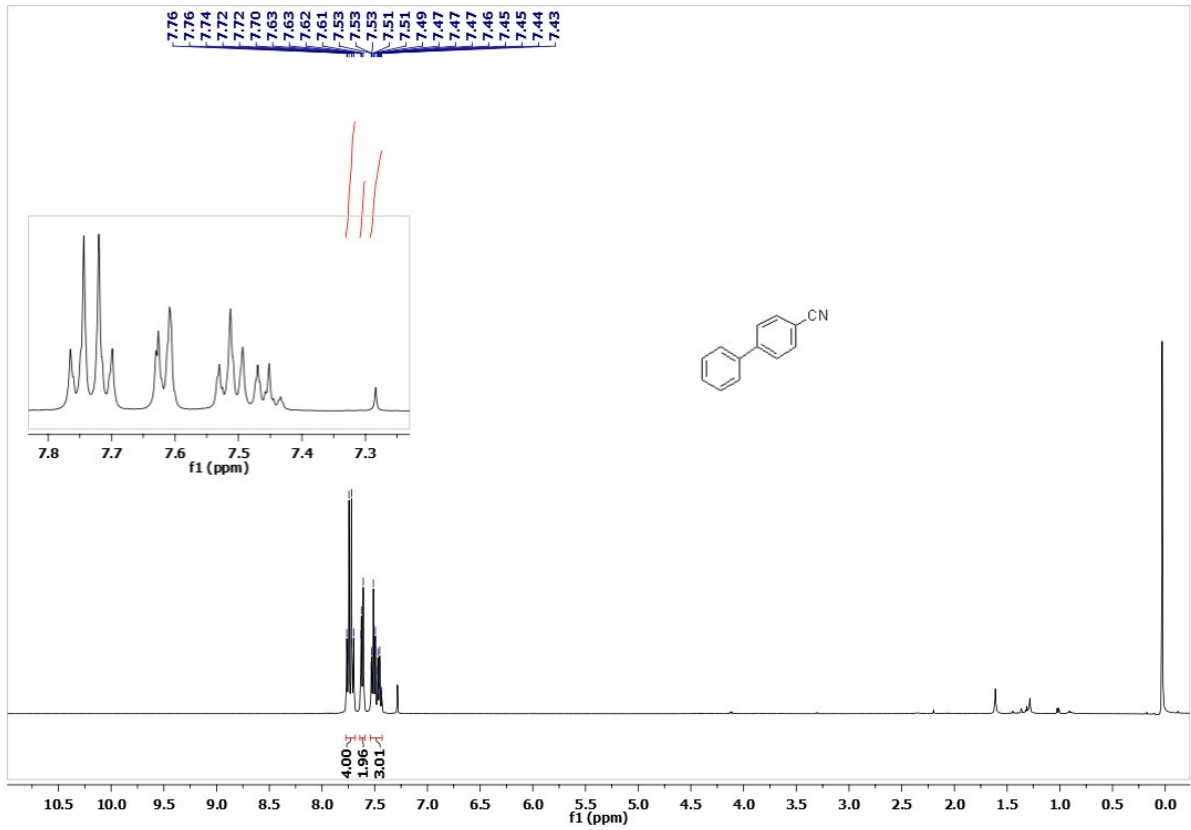
^1H NMR (400 MHz, CDCl_3) δ 7.74 (d, $J = 7.7$ Hz, 1H), 7.61 (dd, $J = 12.1, 4.5$ Hz, 1H), 7.53 – 7.37 (m, 4H), 7.33 – 7.23 (m, 3H), 2.41 (s, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 145.53, 138.68, 135.27, 133.70, 132.73, 129.96, 129.43, 128.60, 127.25, 118.84, 111.21, 21.24.

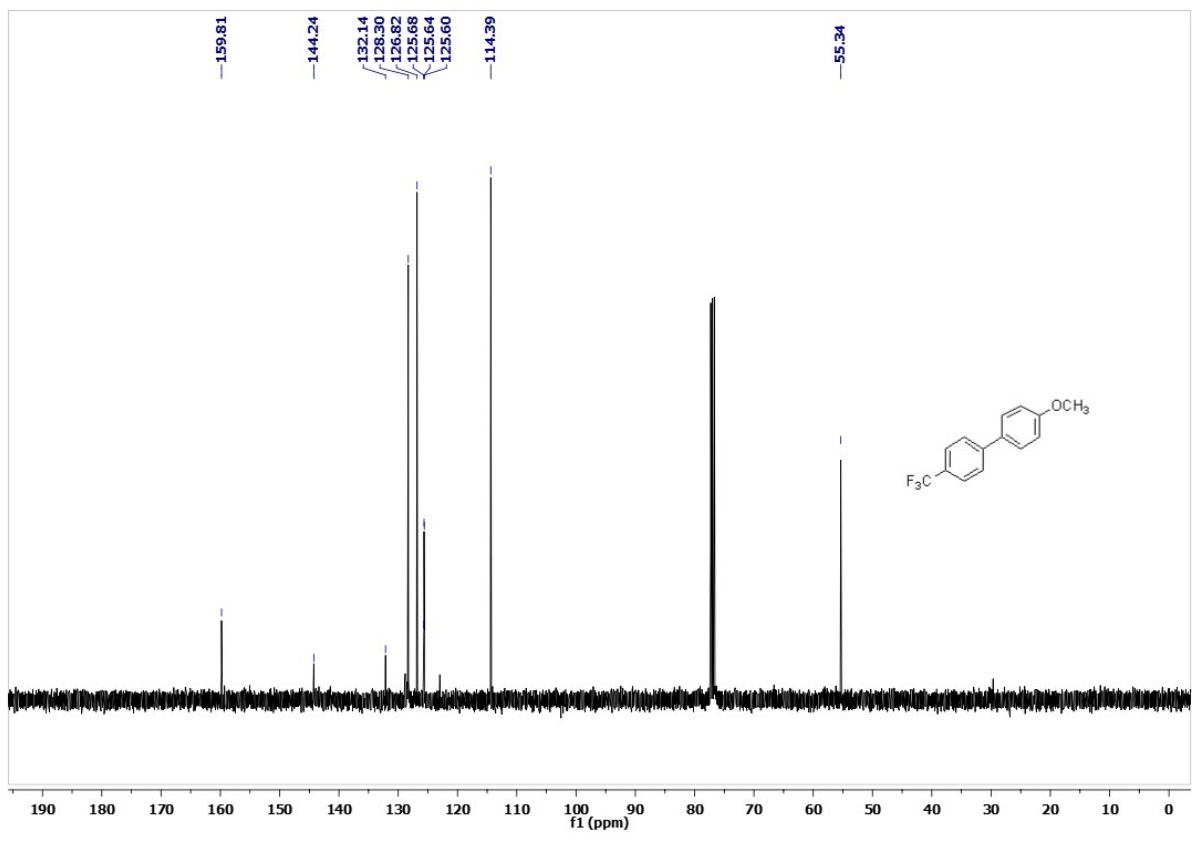
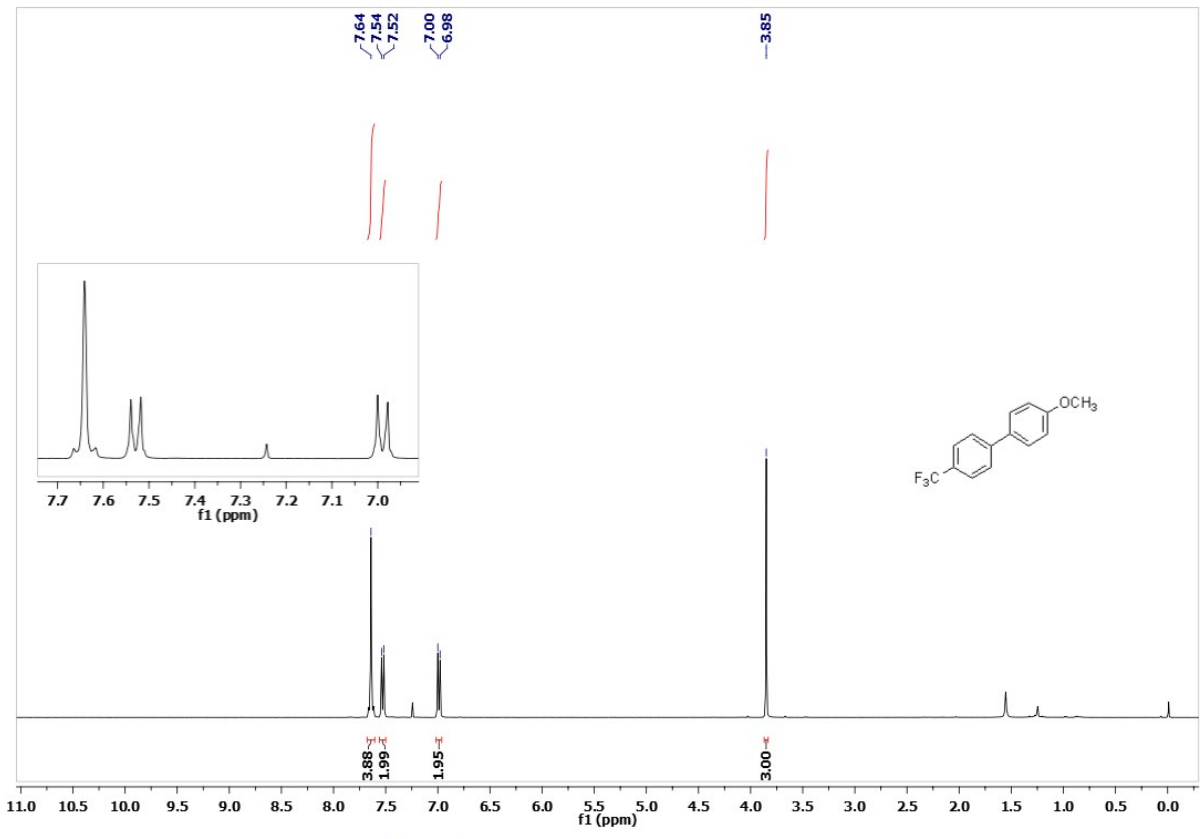


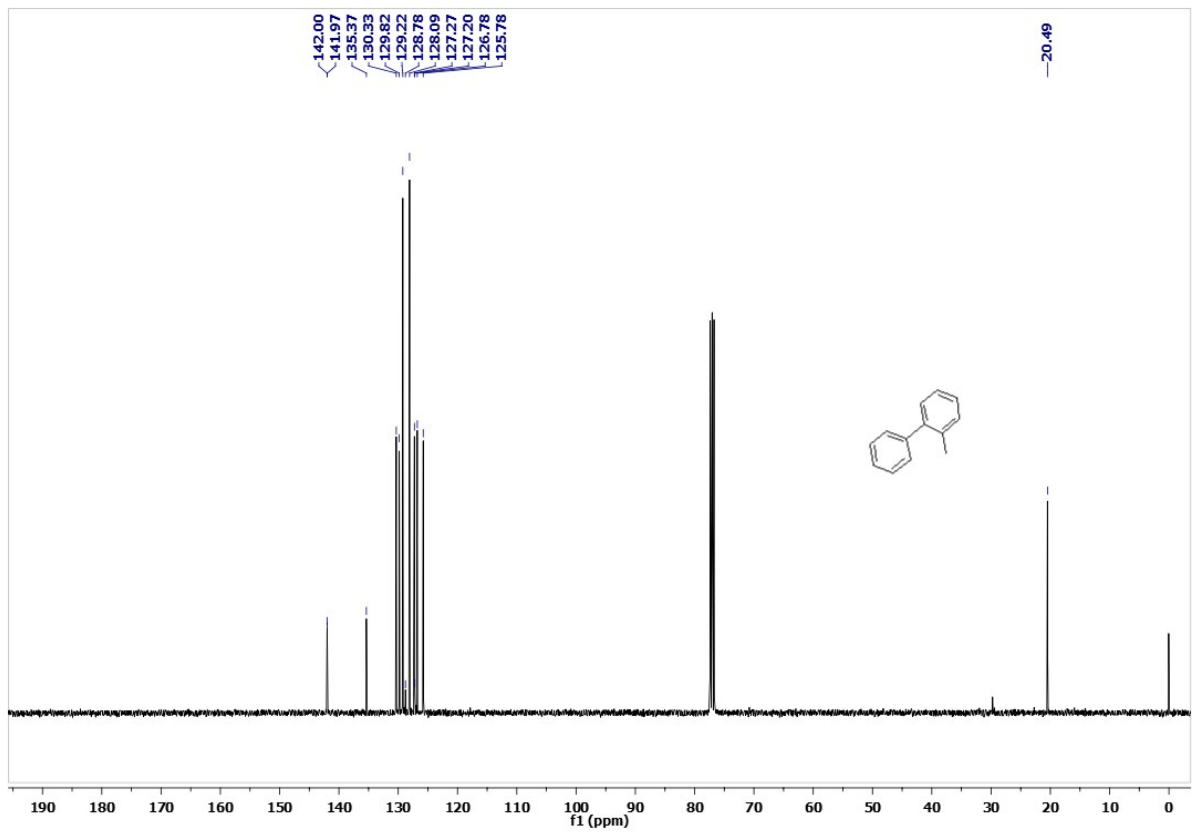
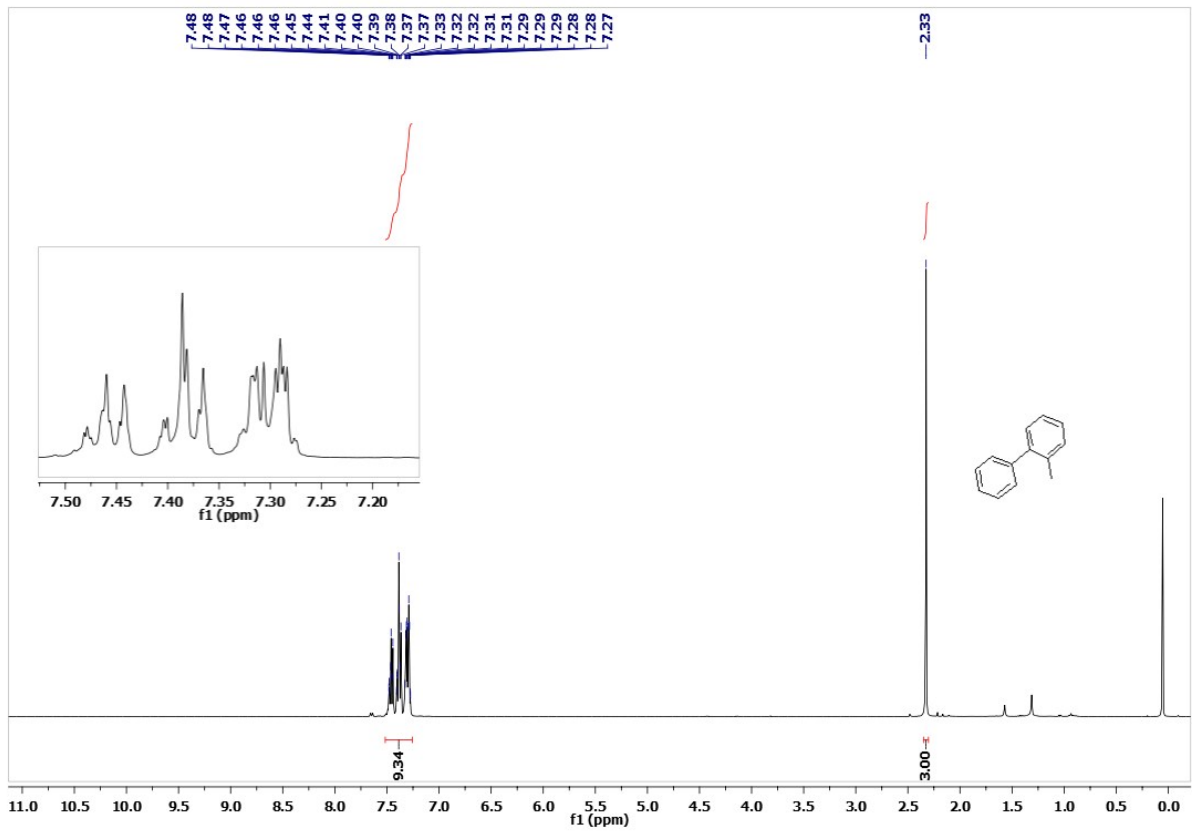


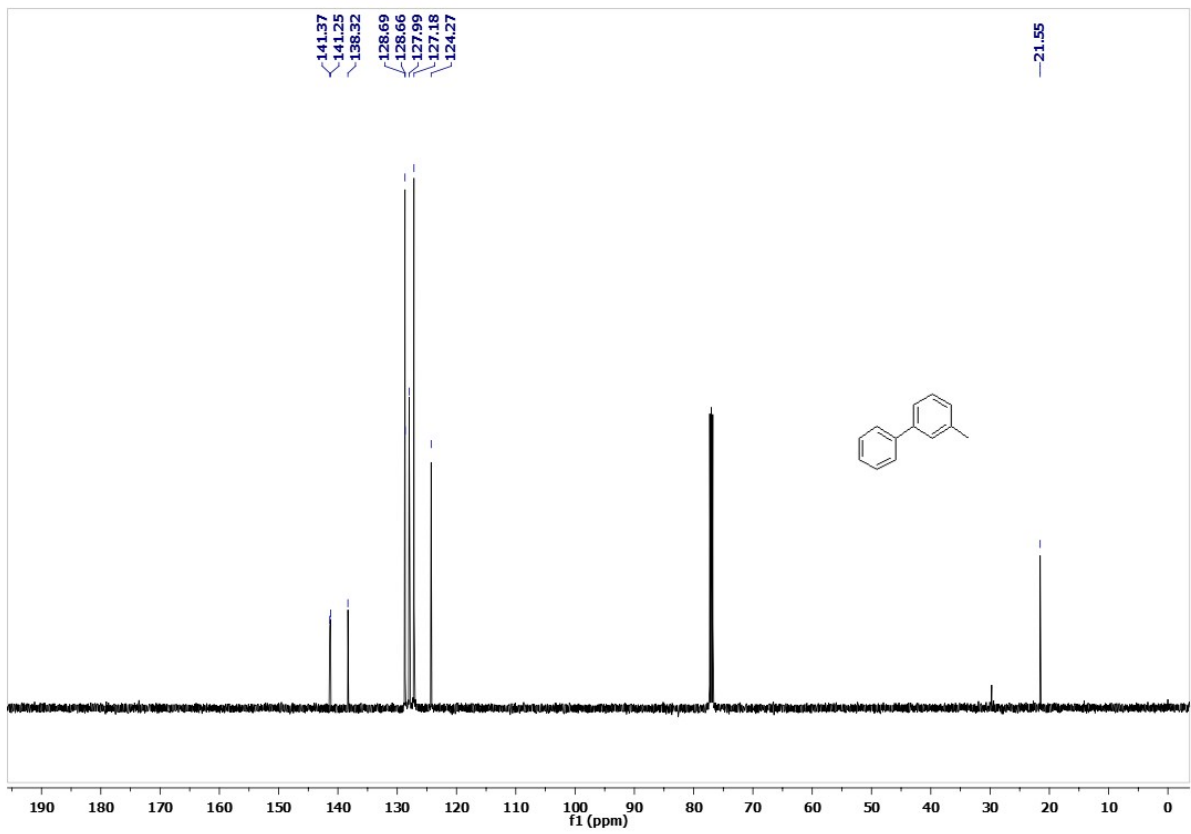
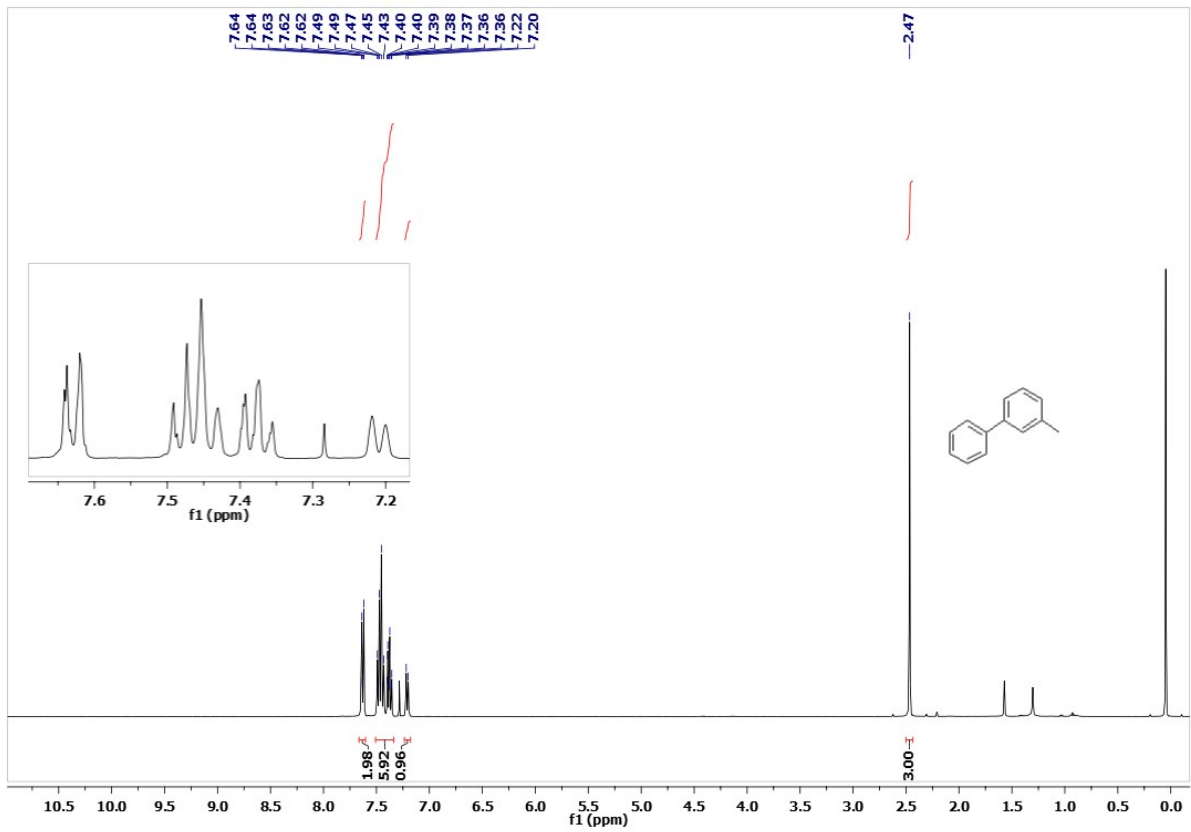


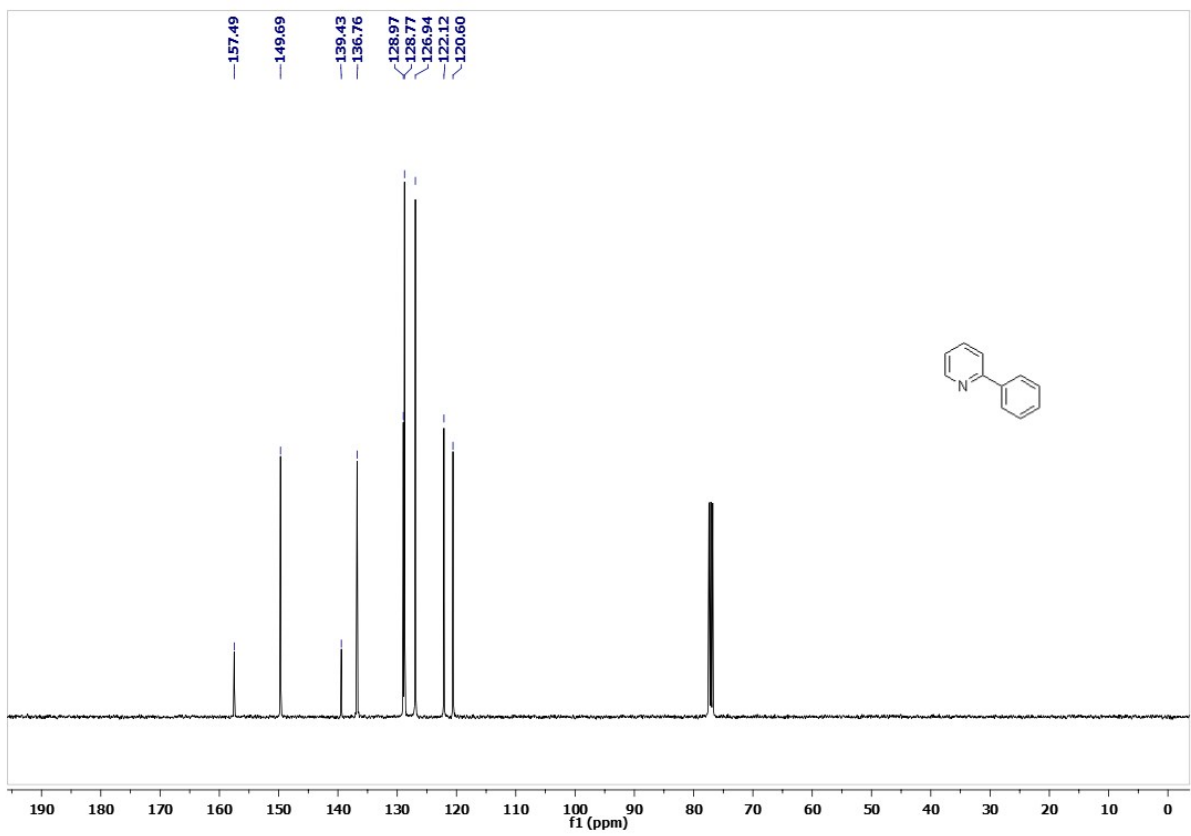
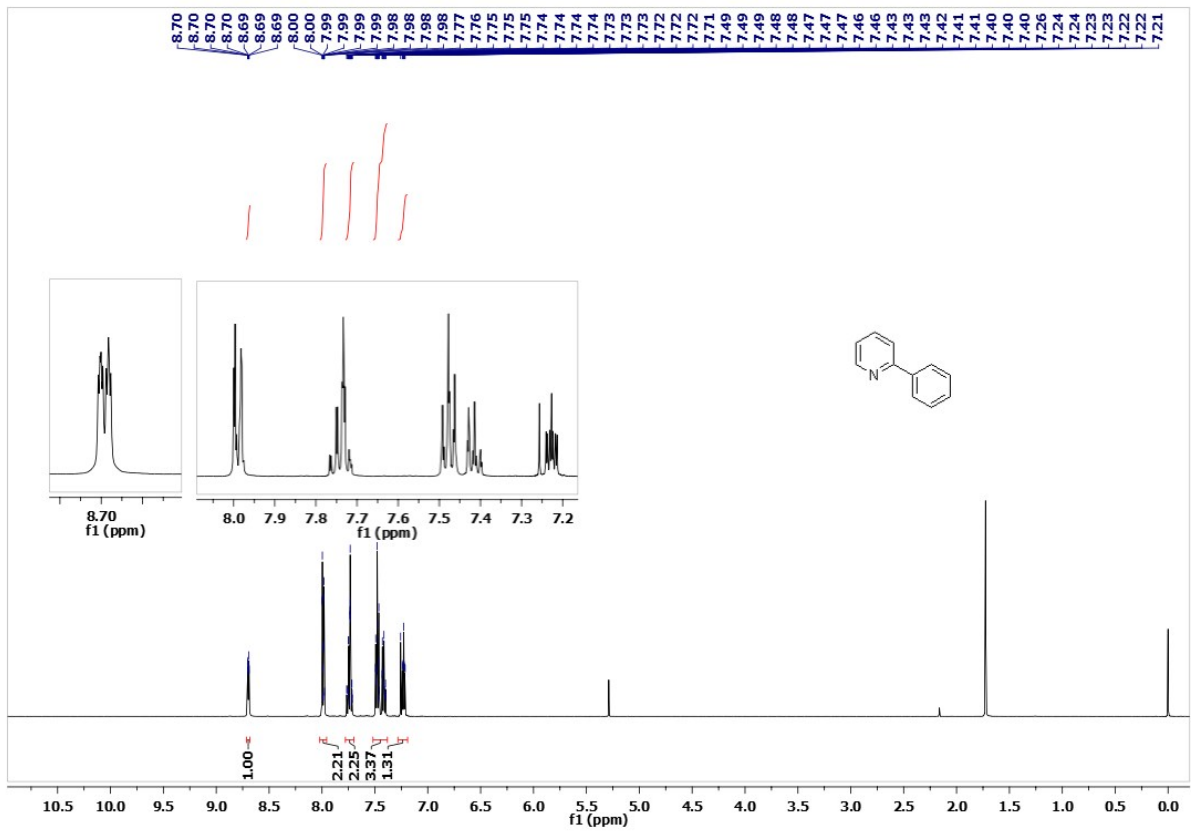


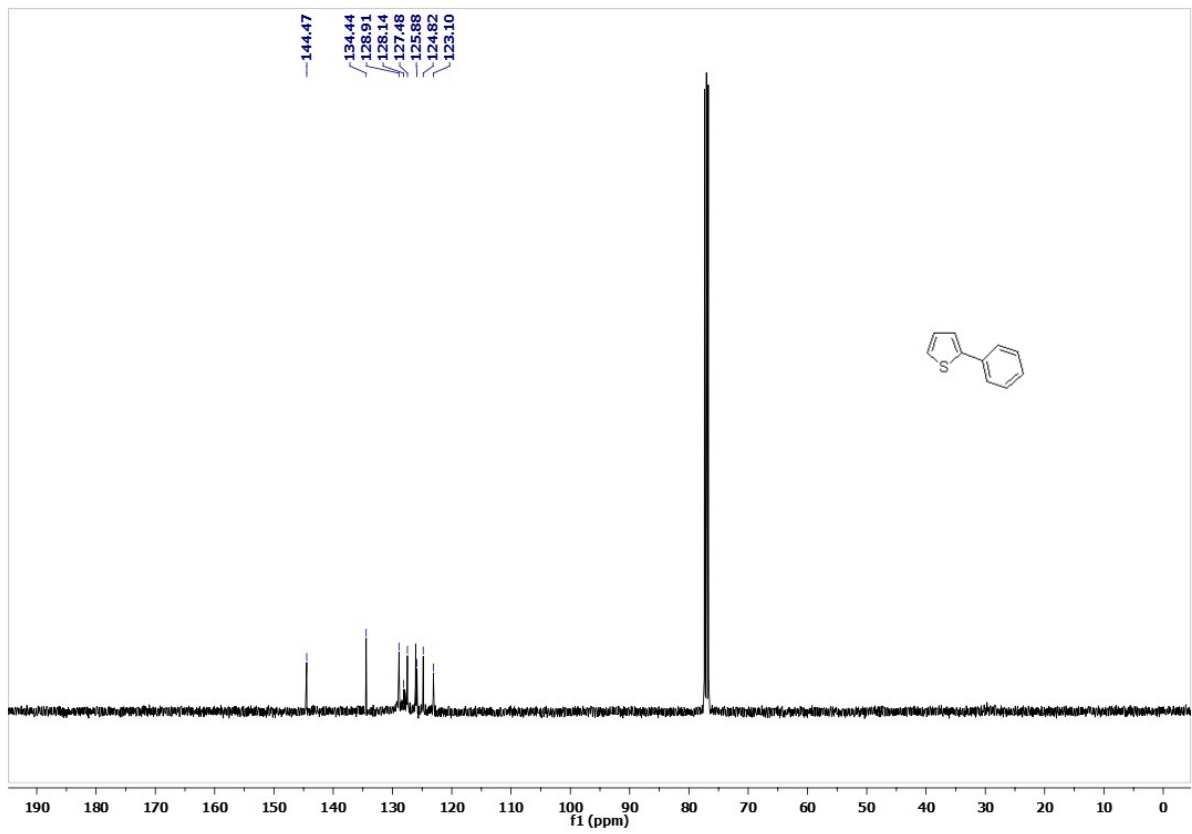
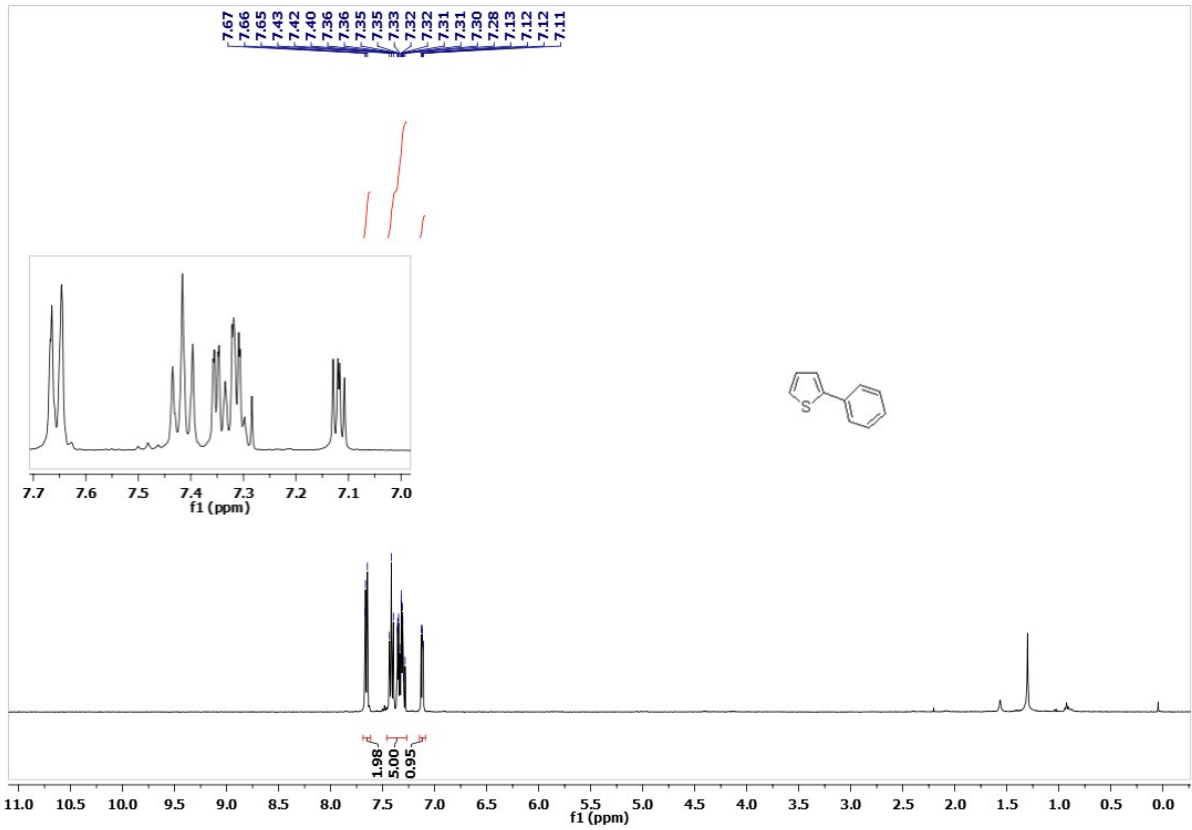


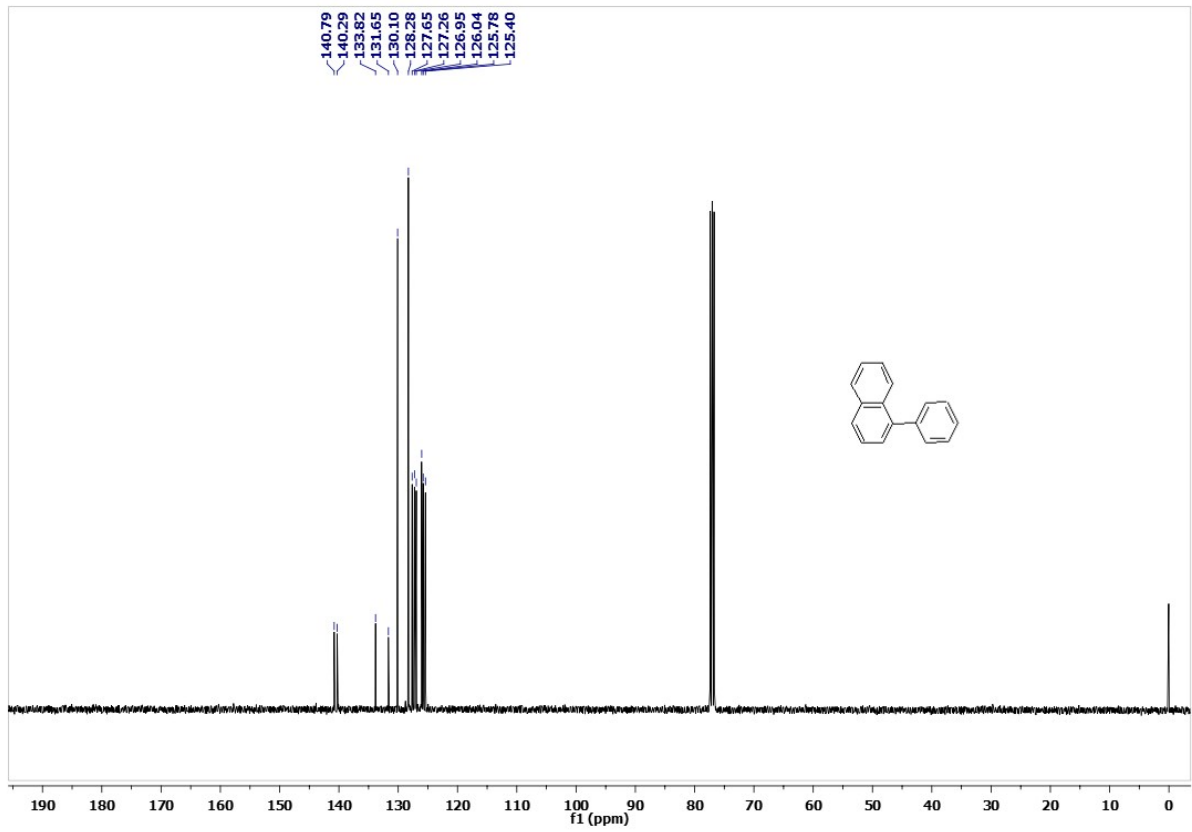
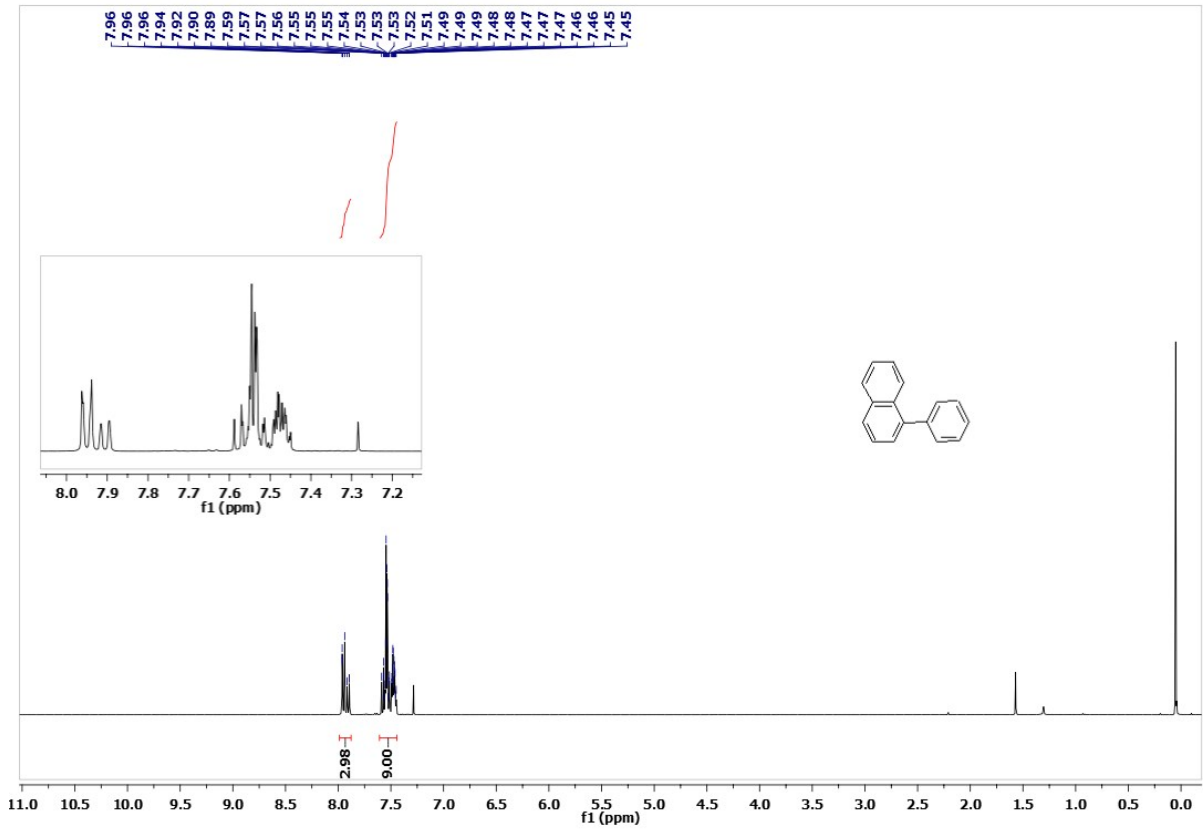


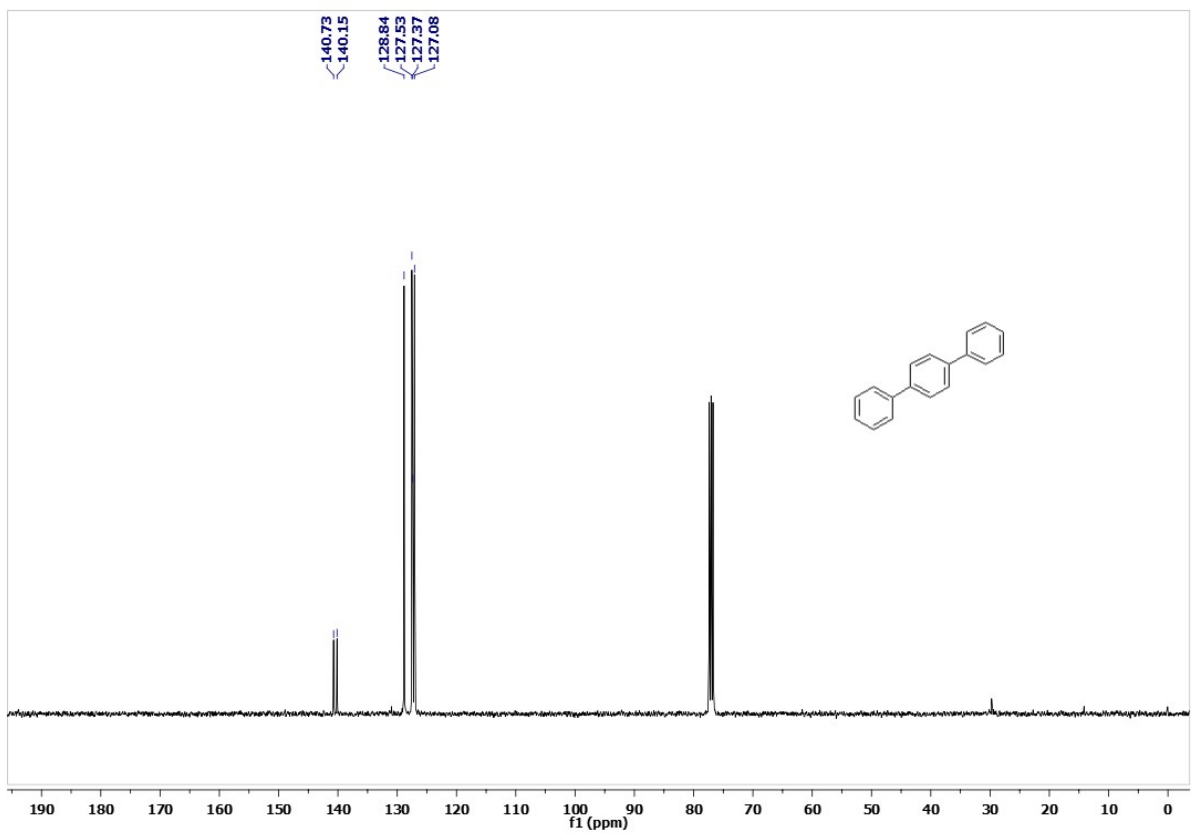
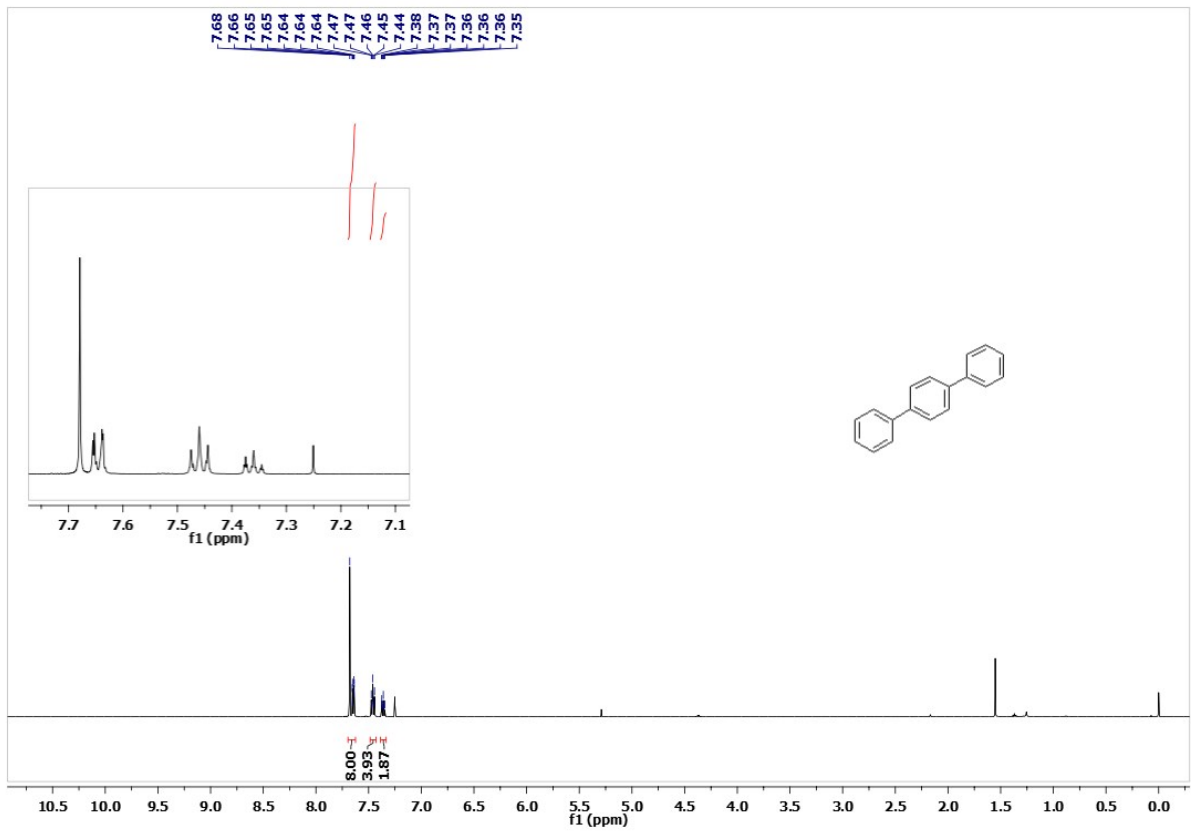


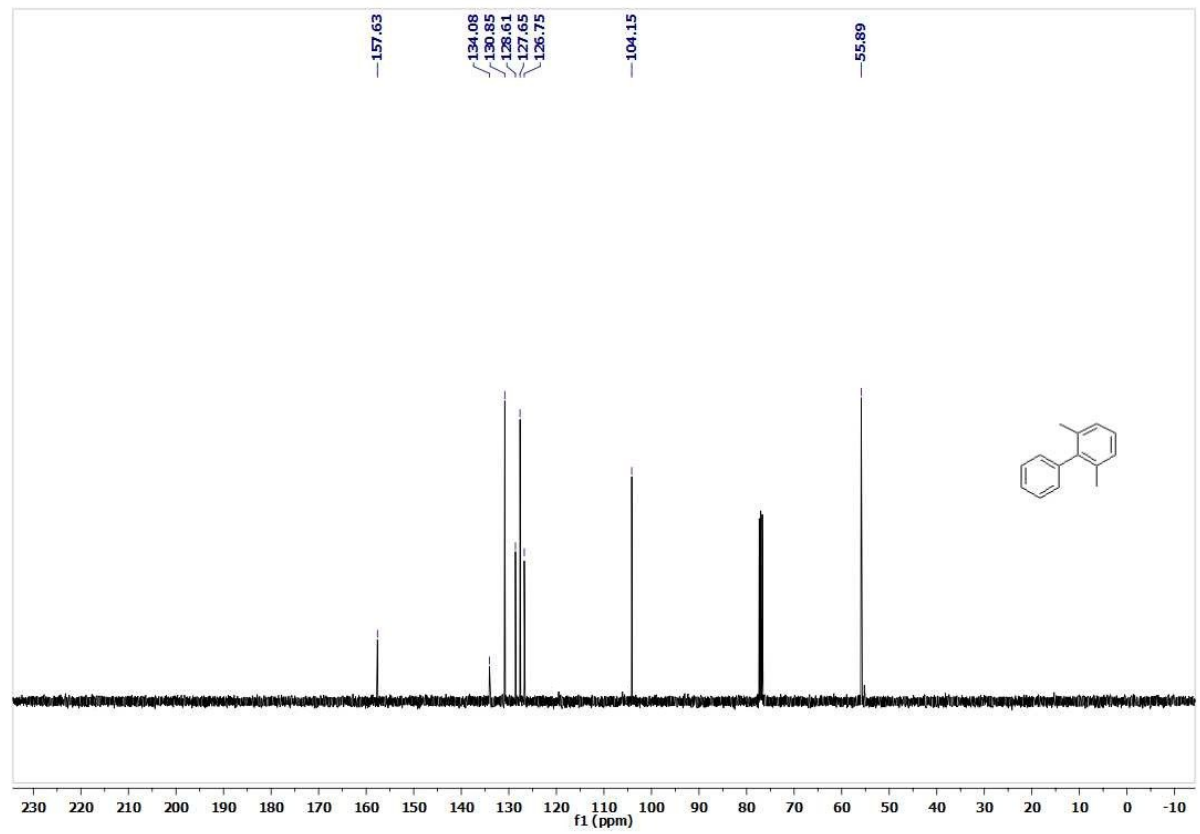
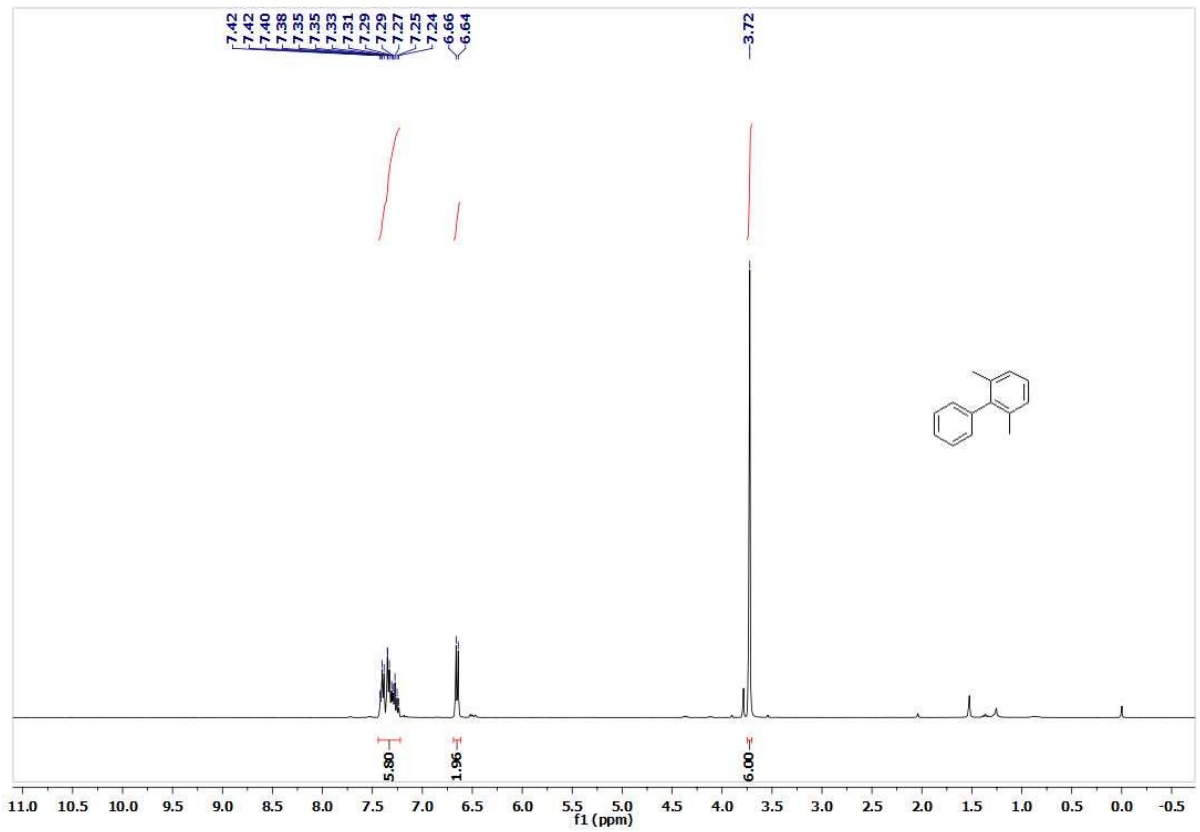


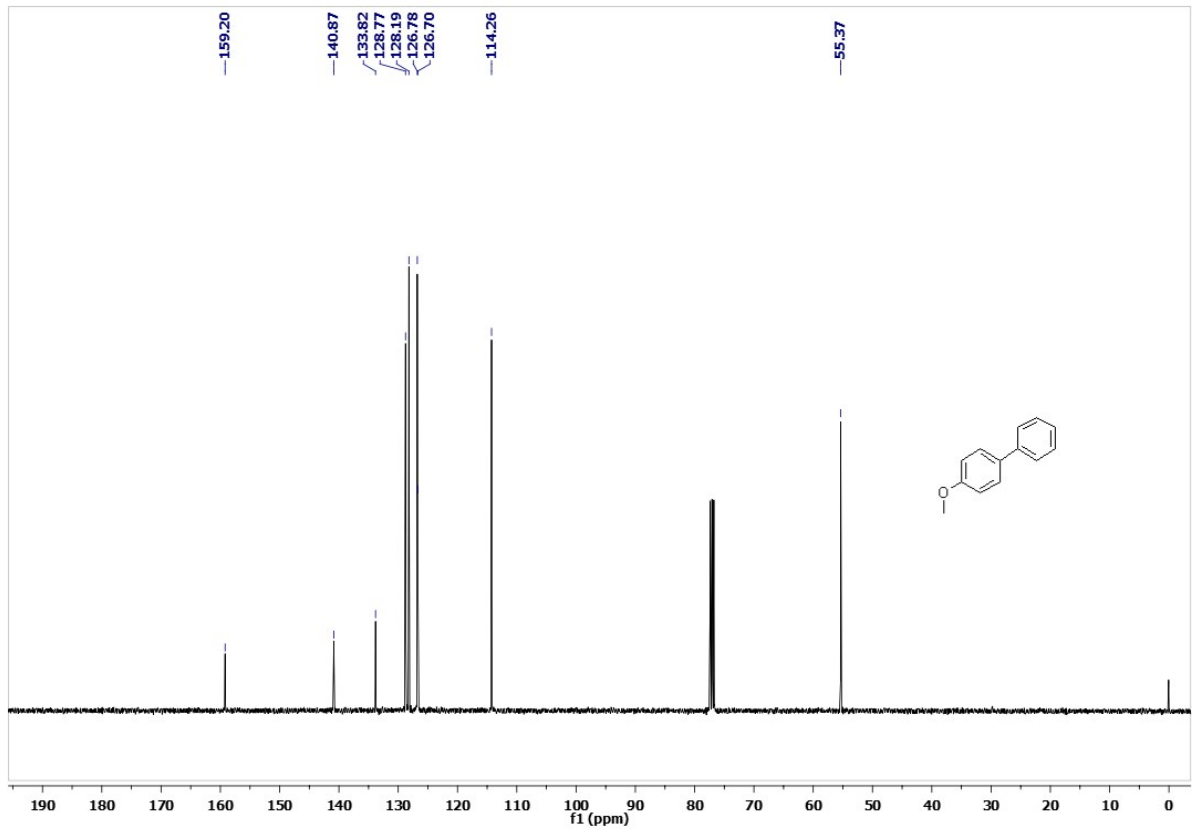
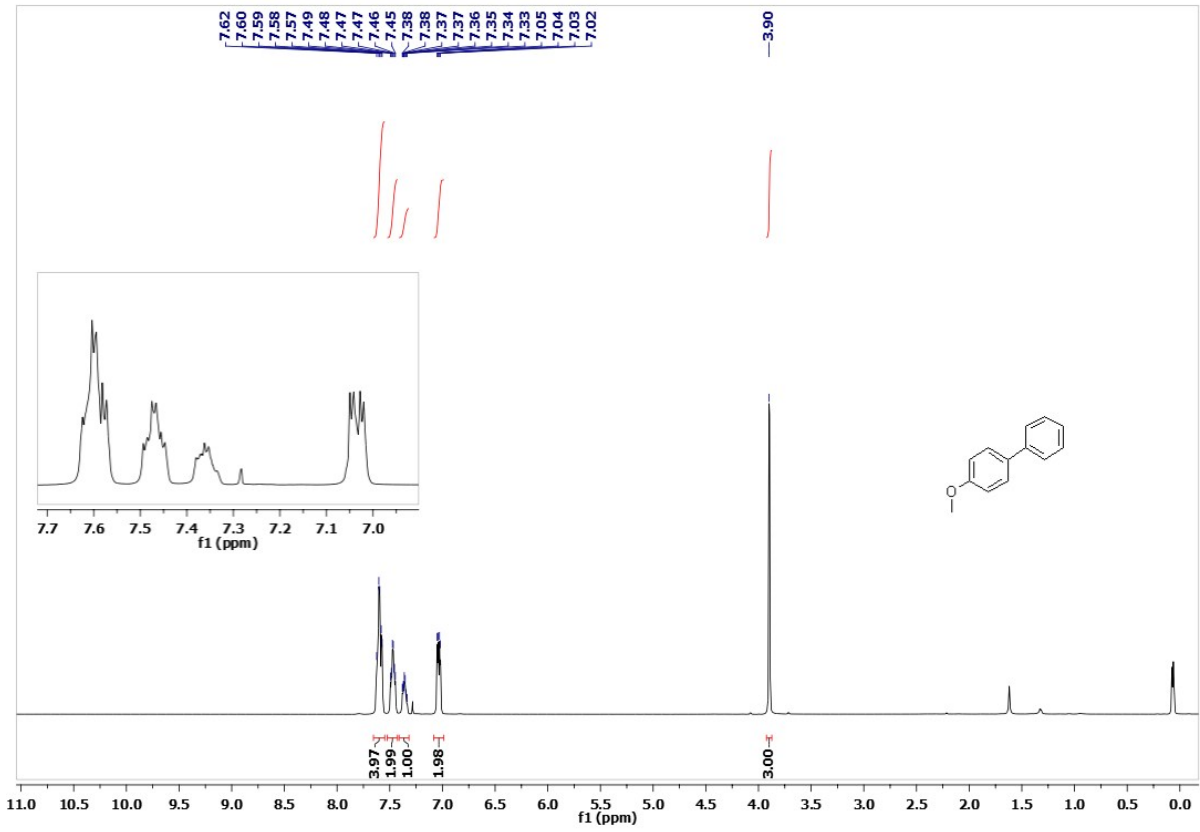


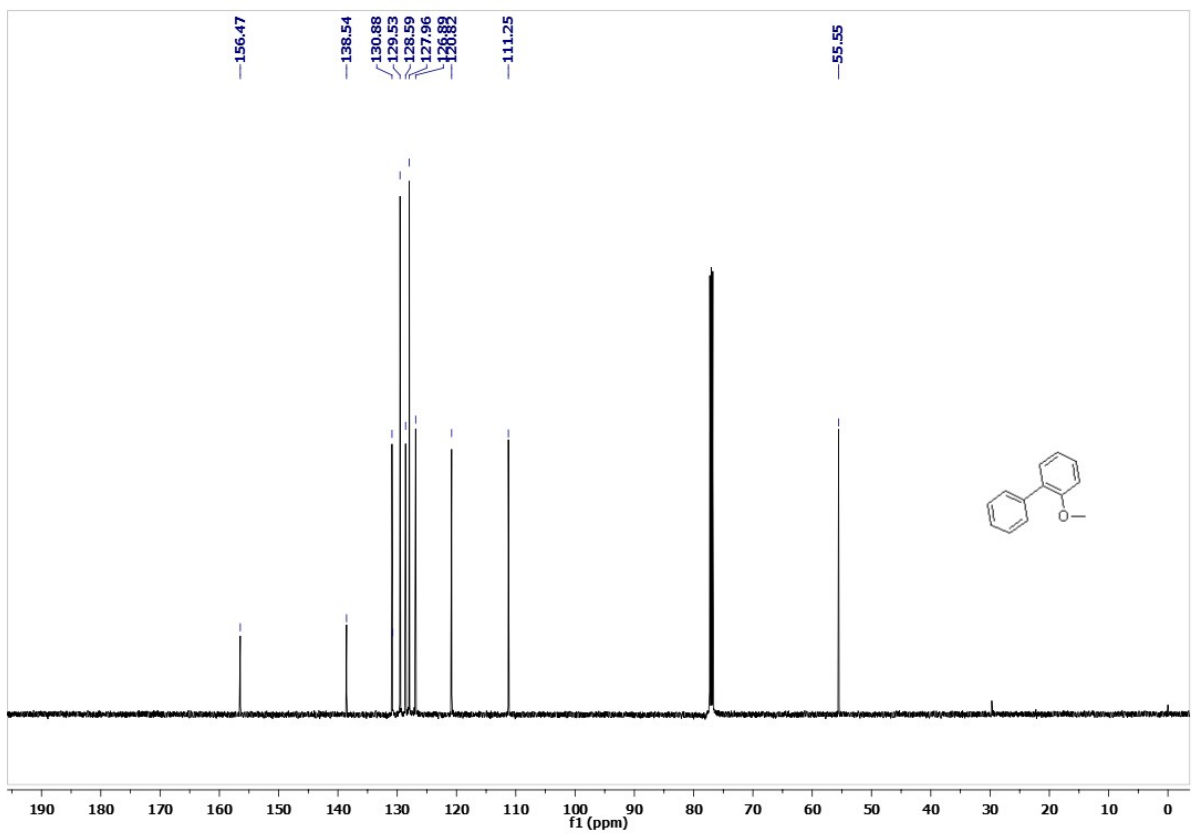
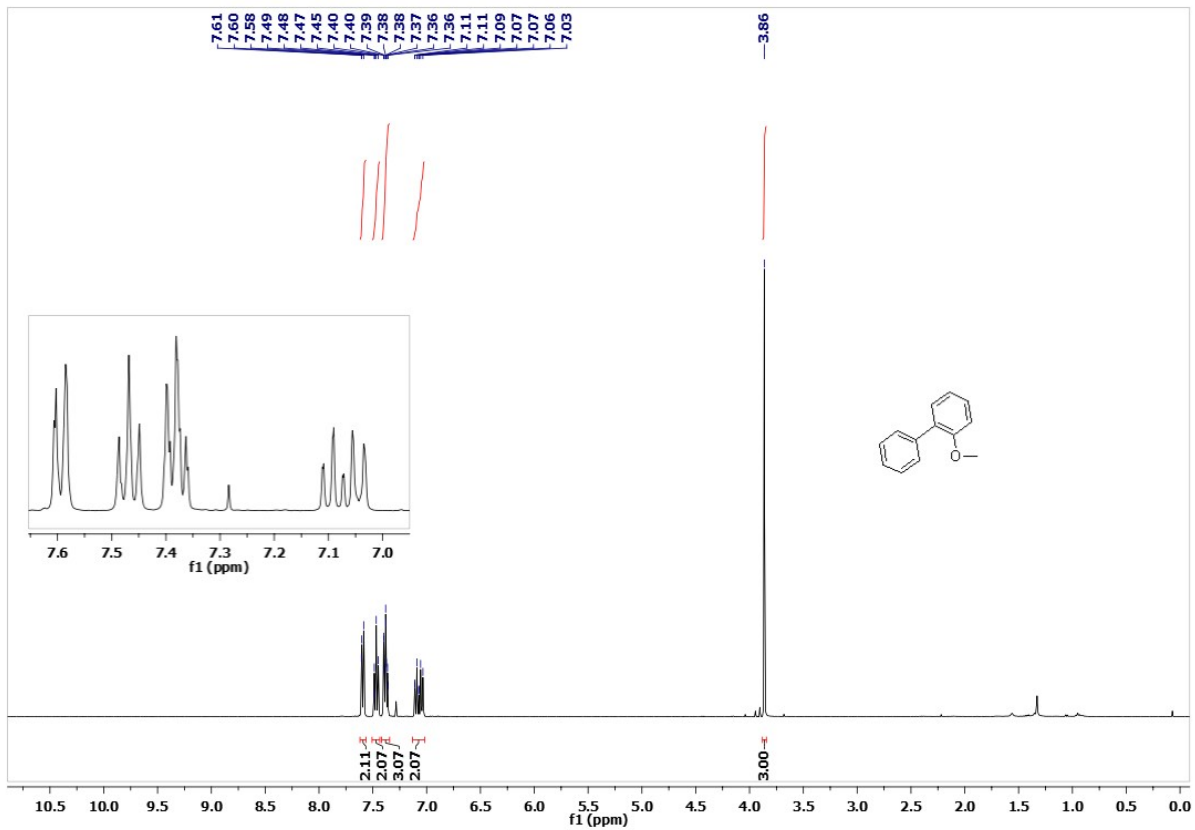


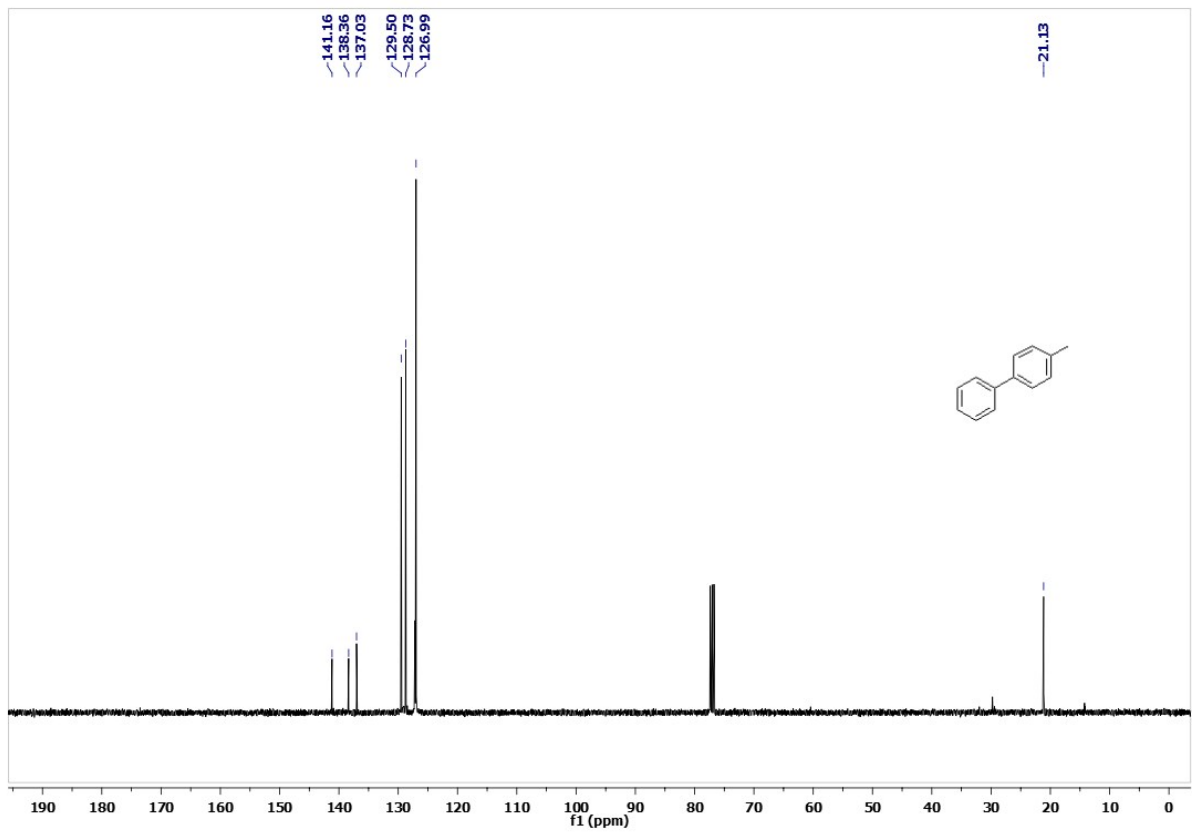
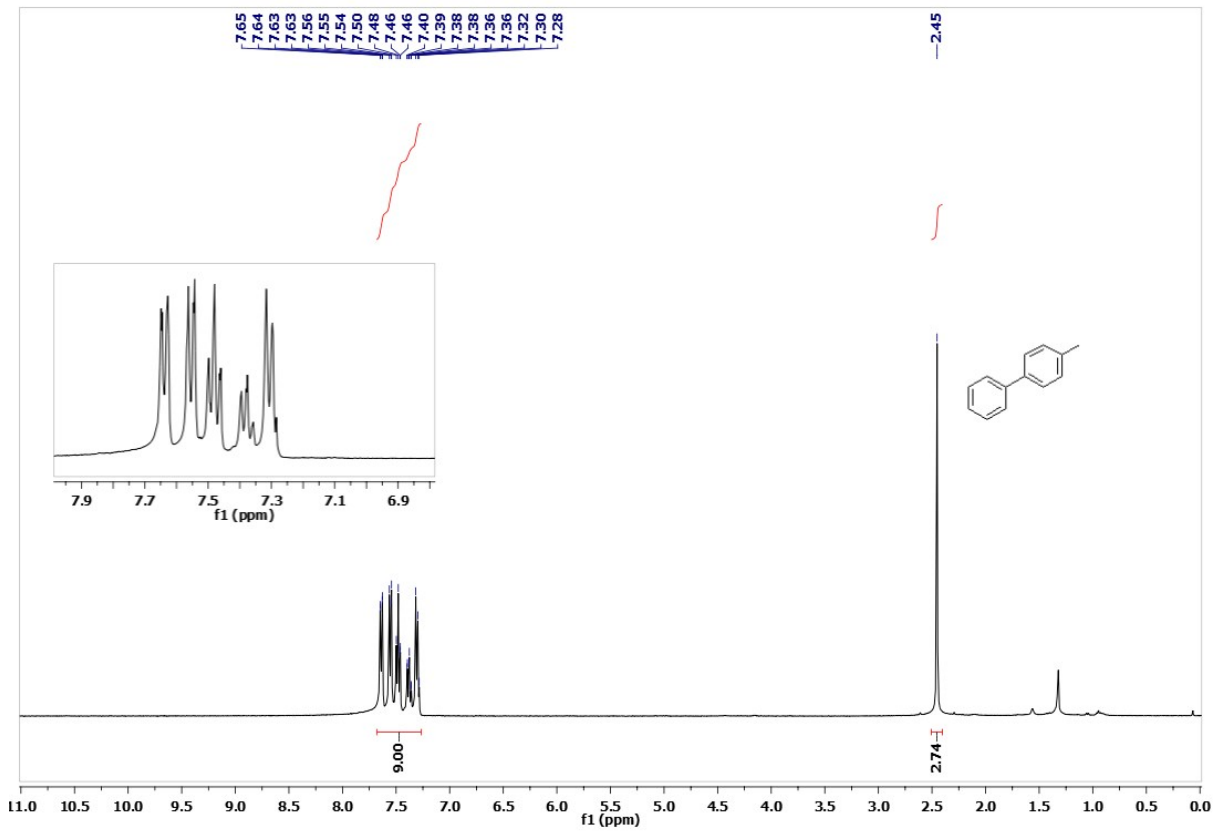


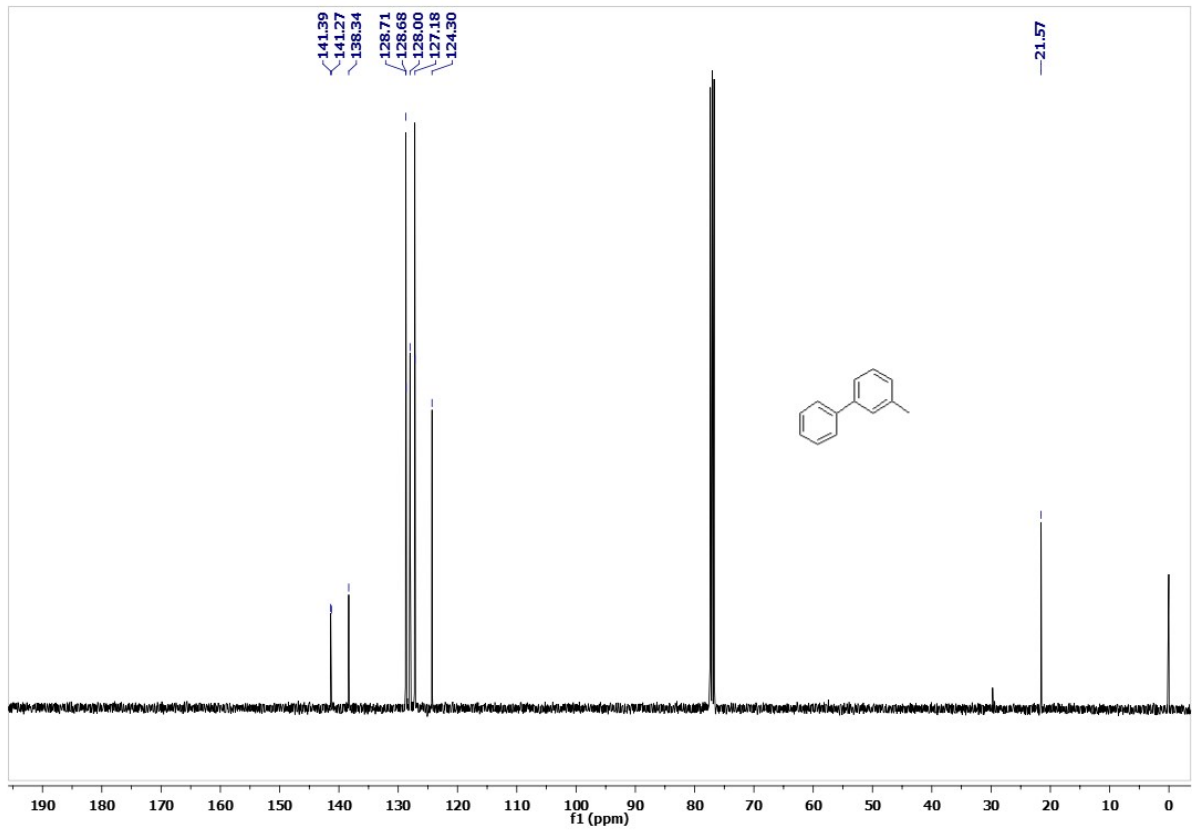
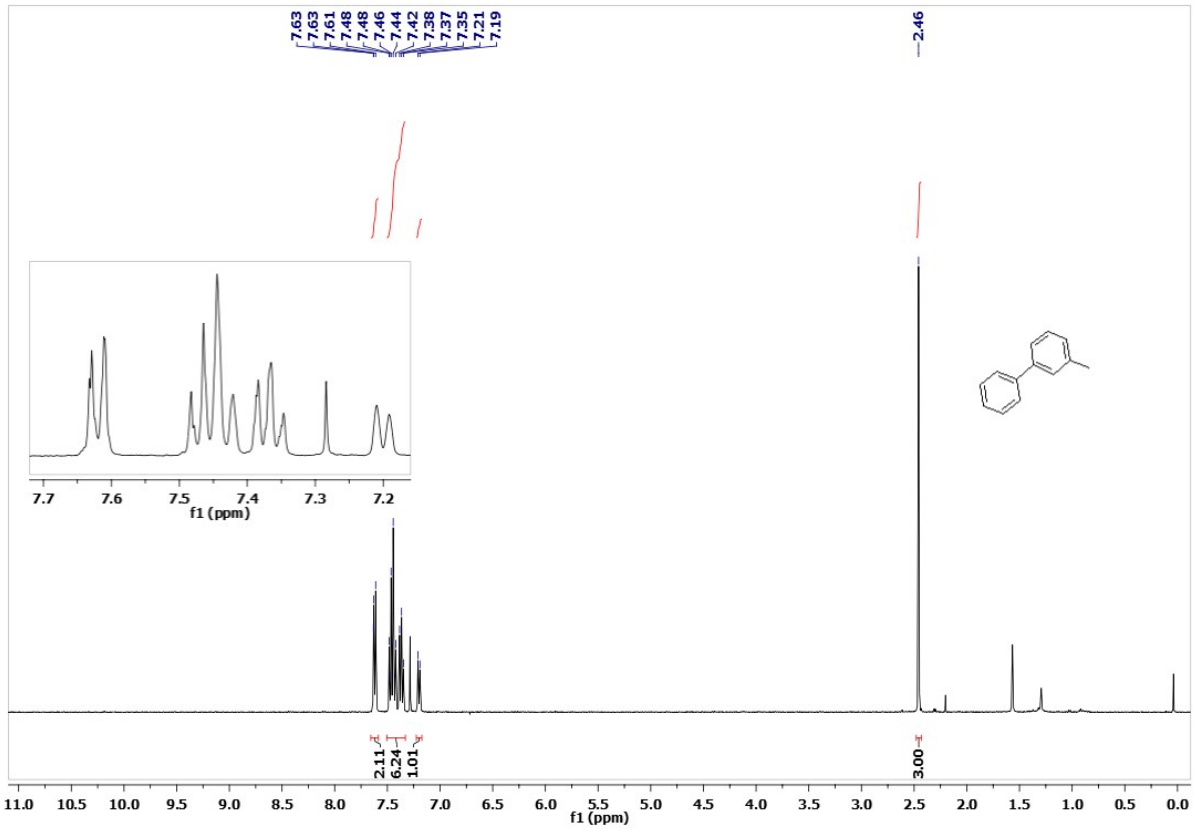


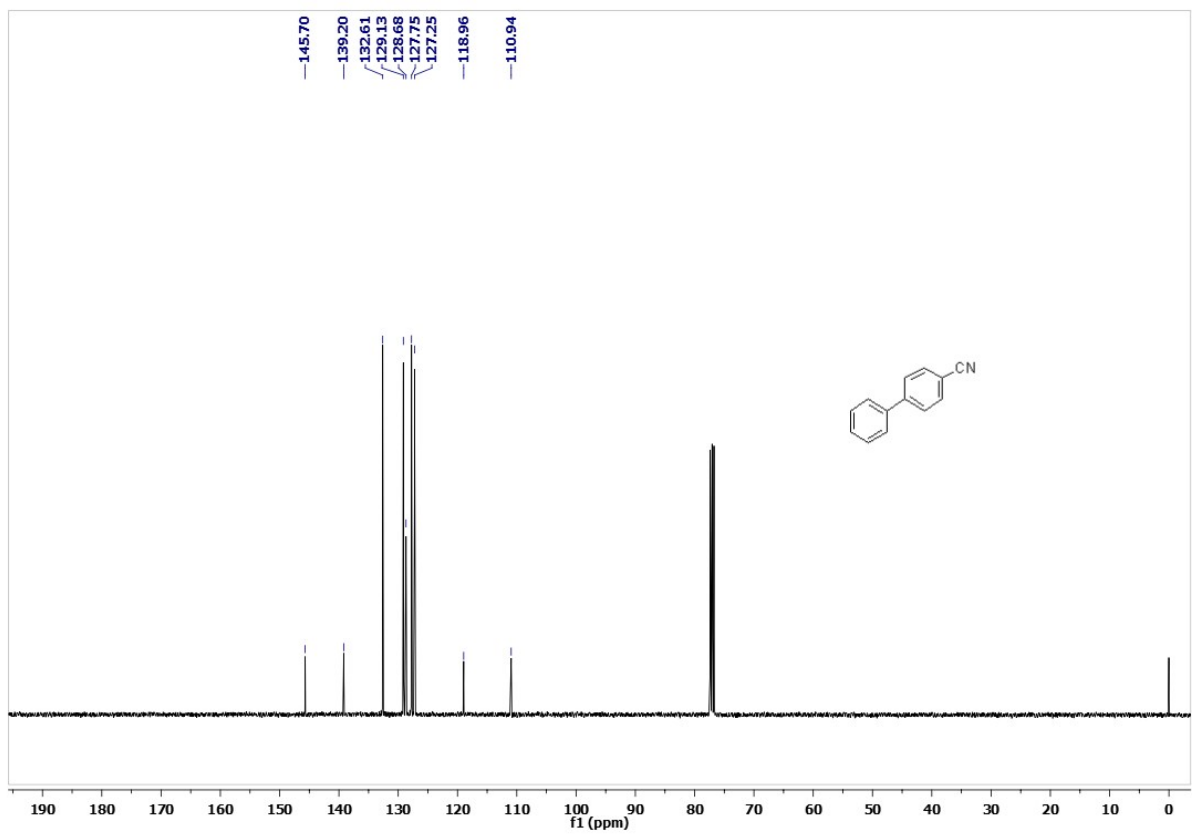
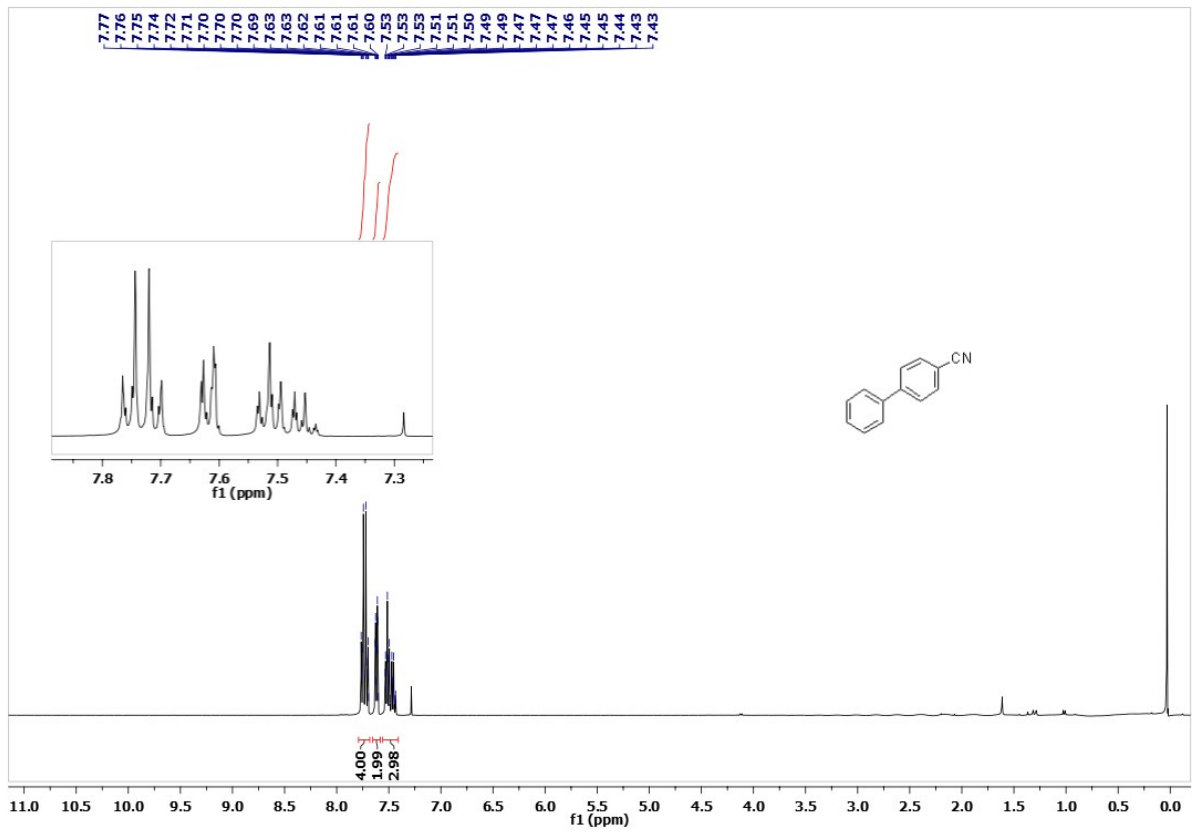


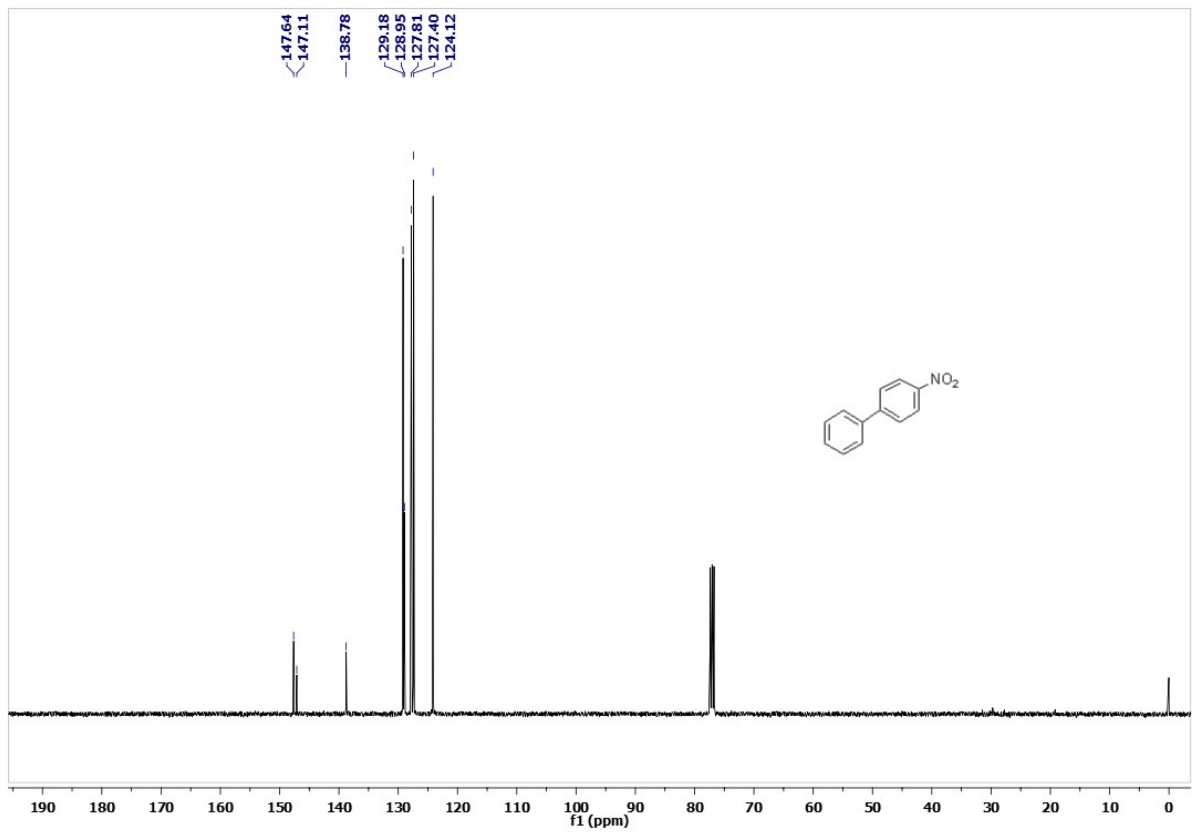
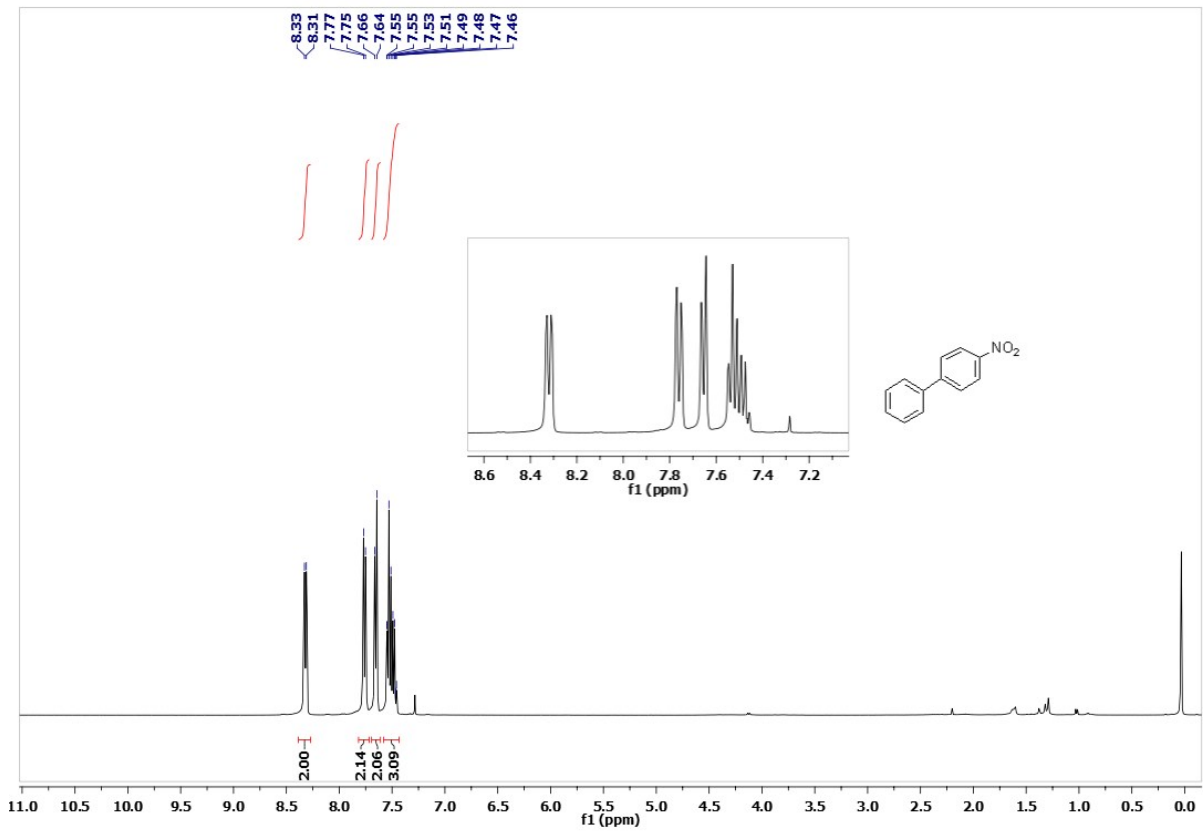


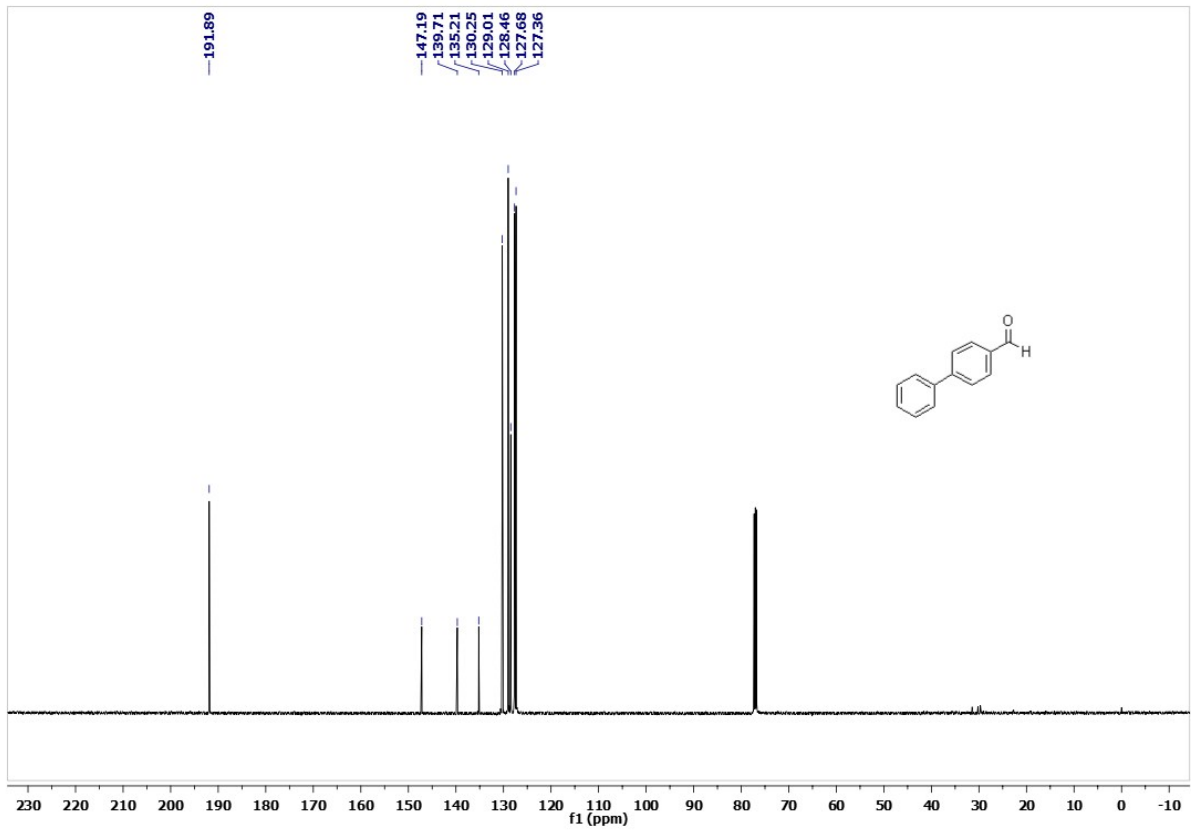
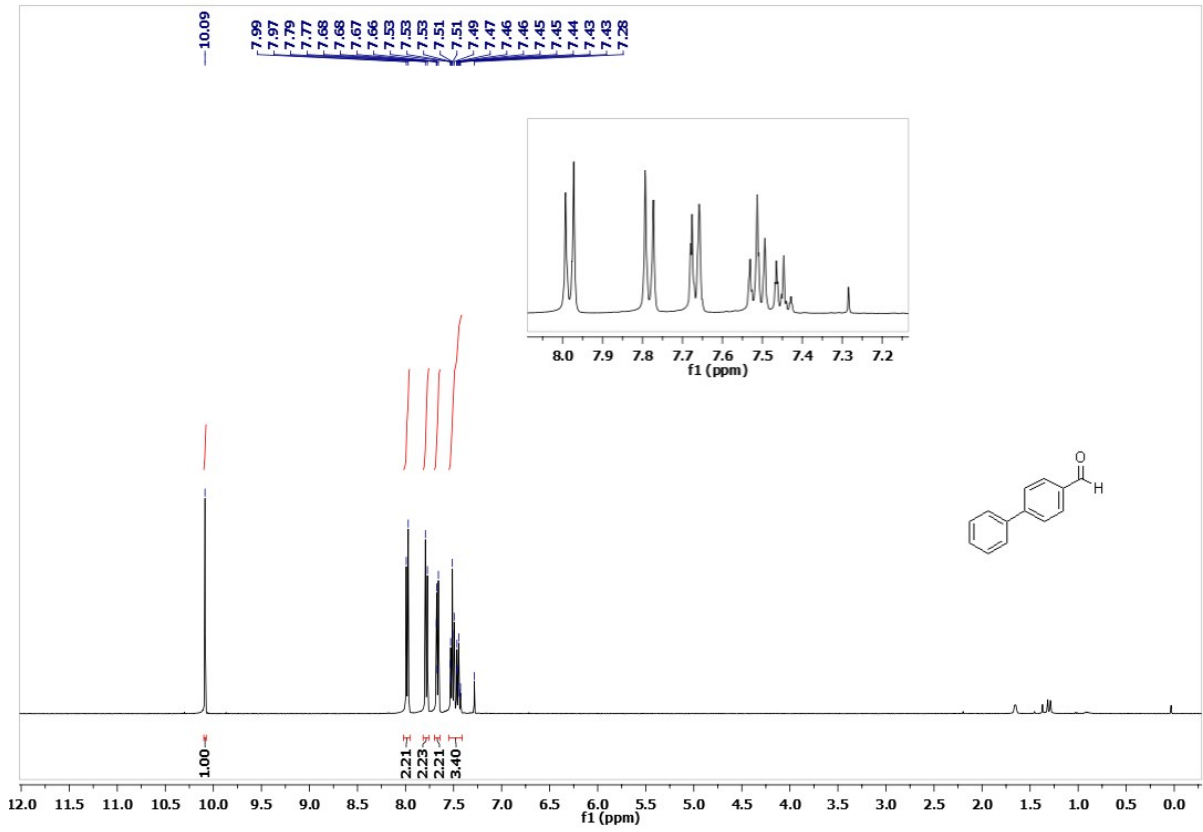


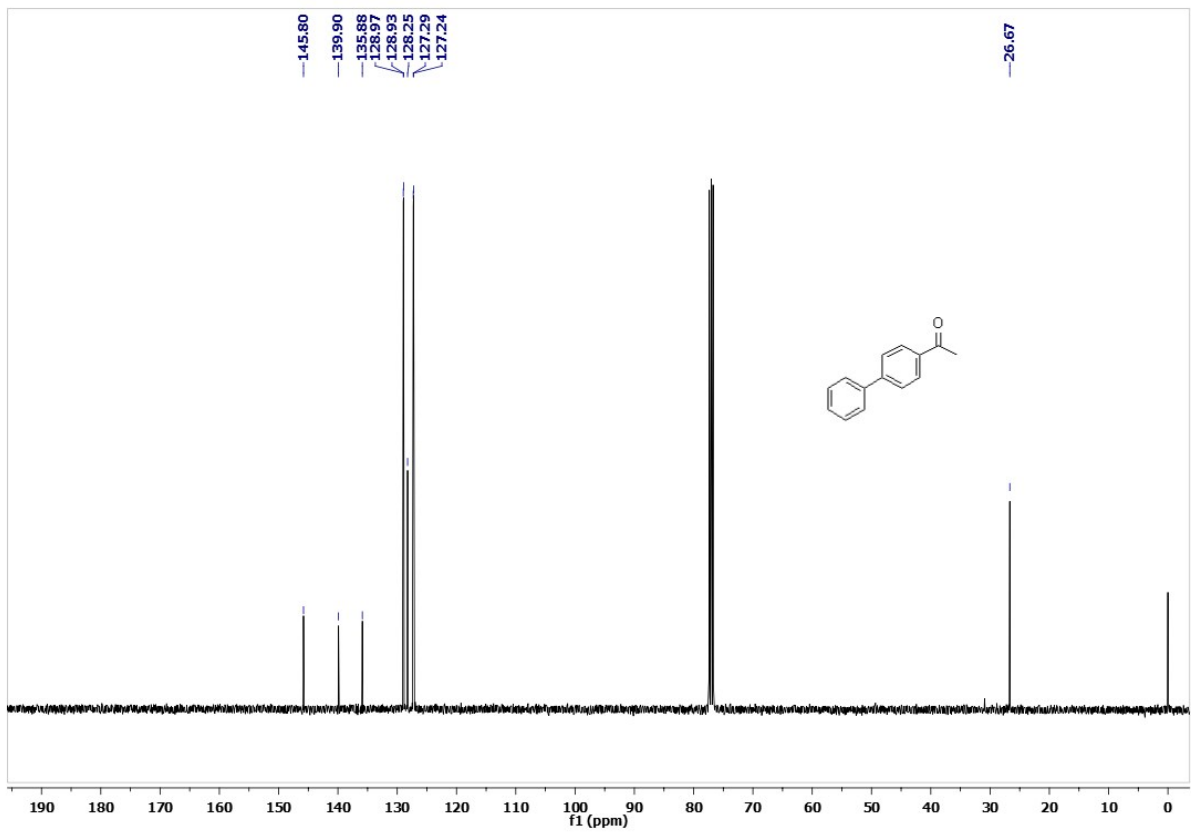
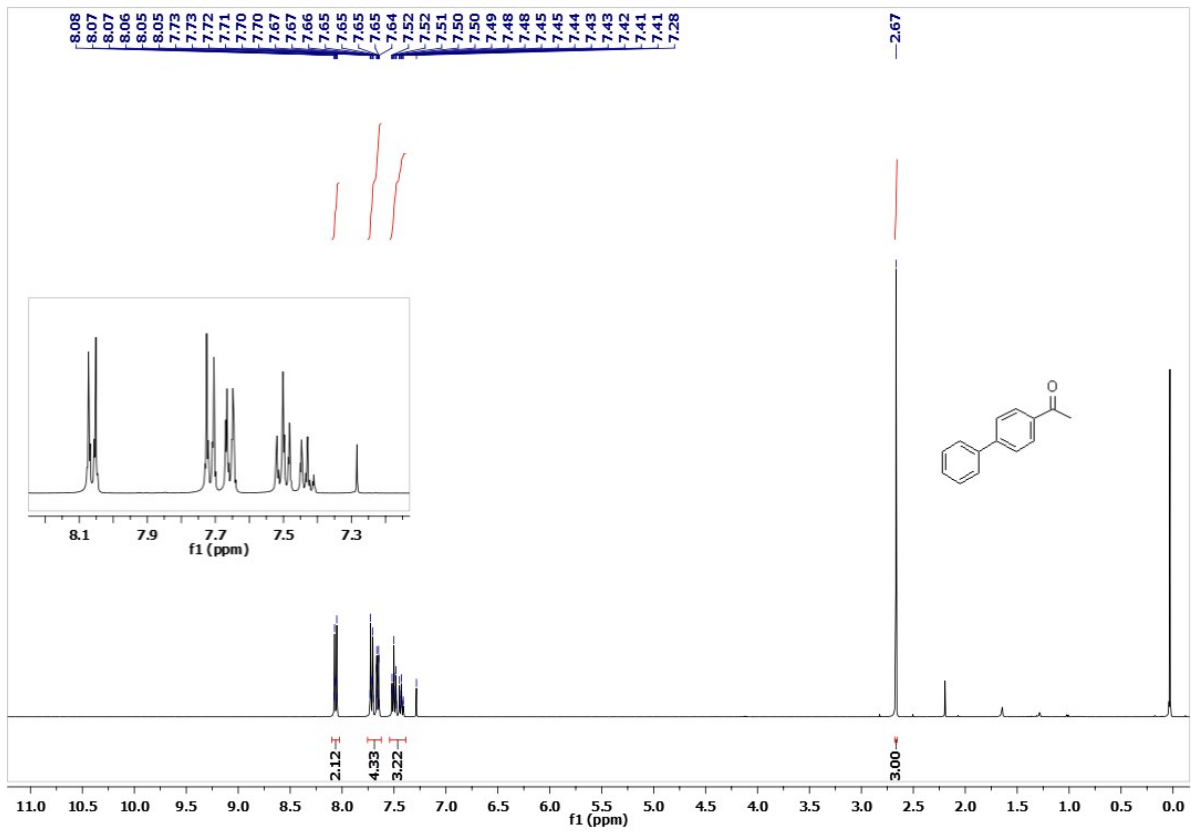


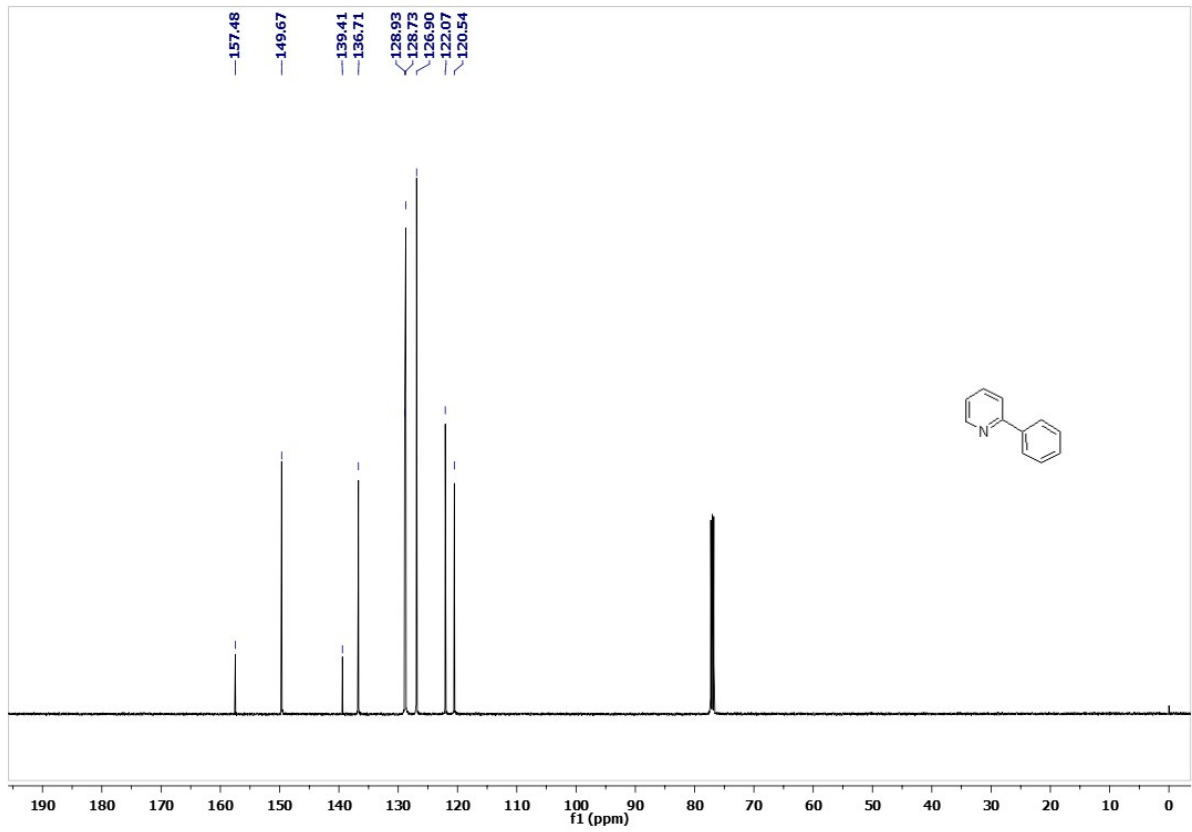
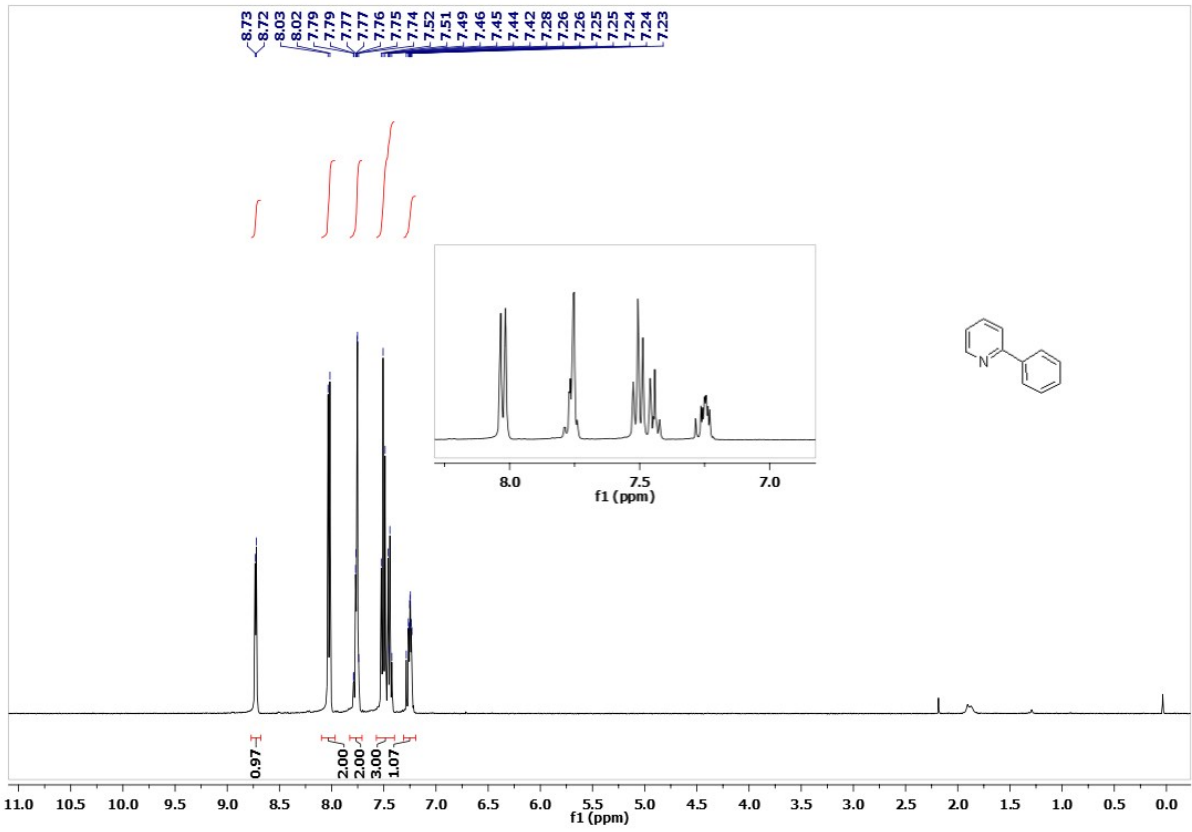


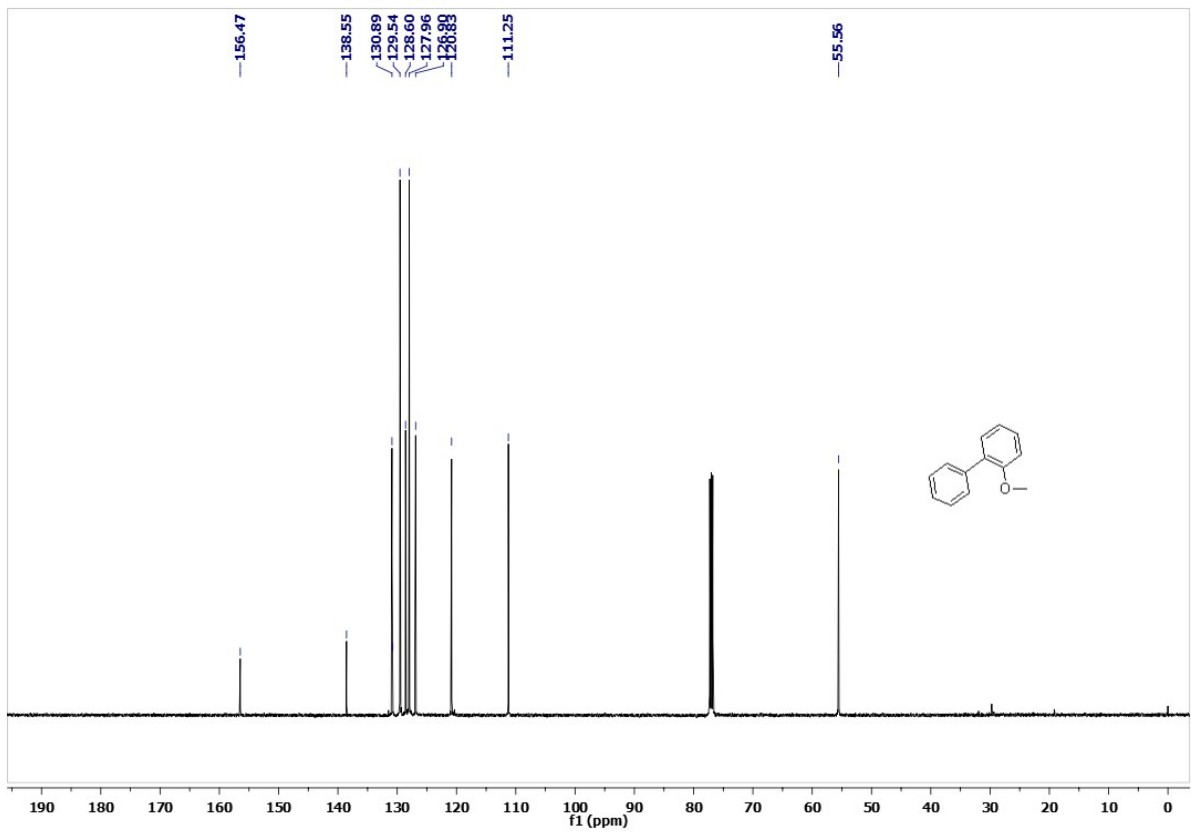
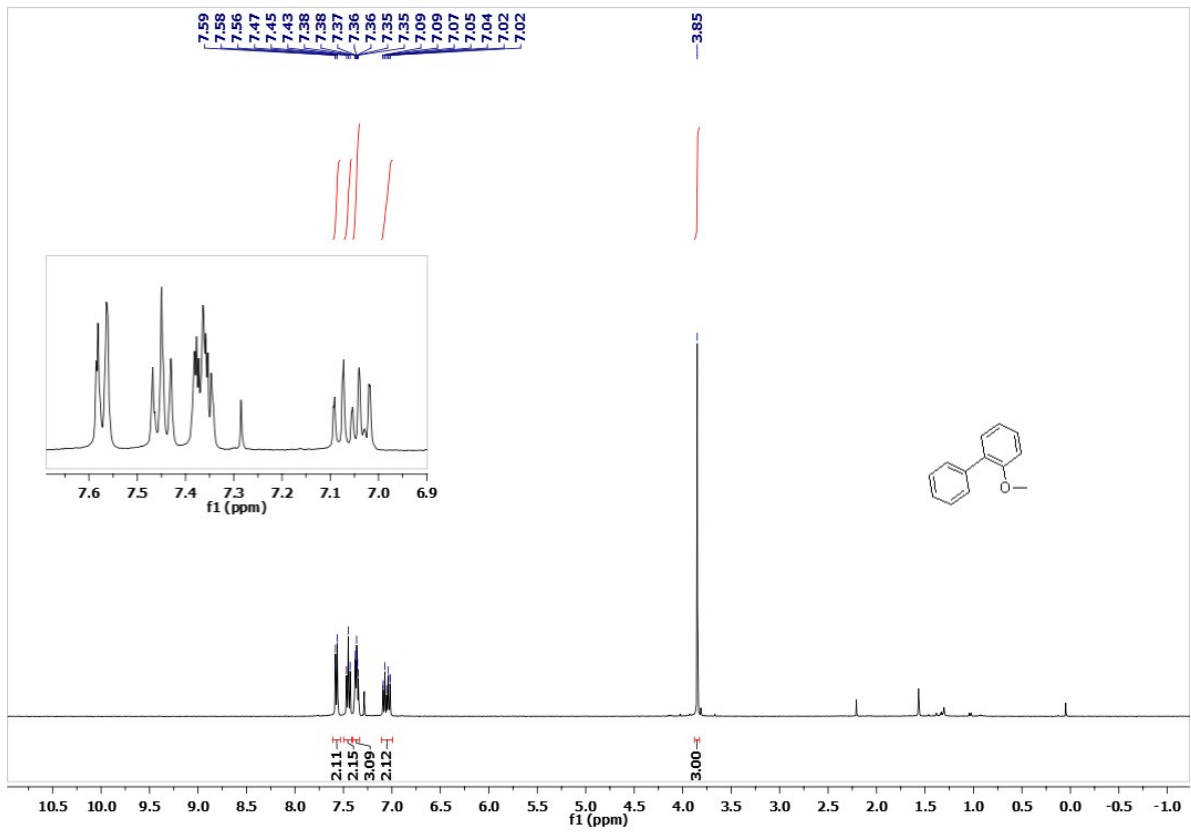


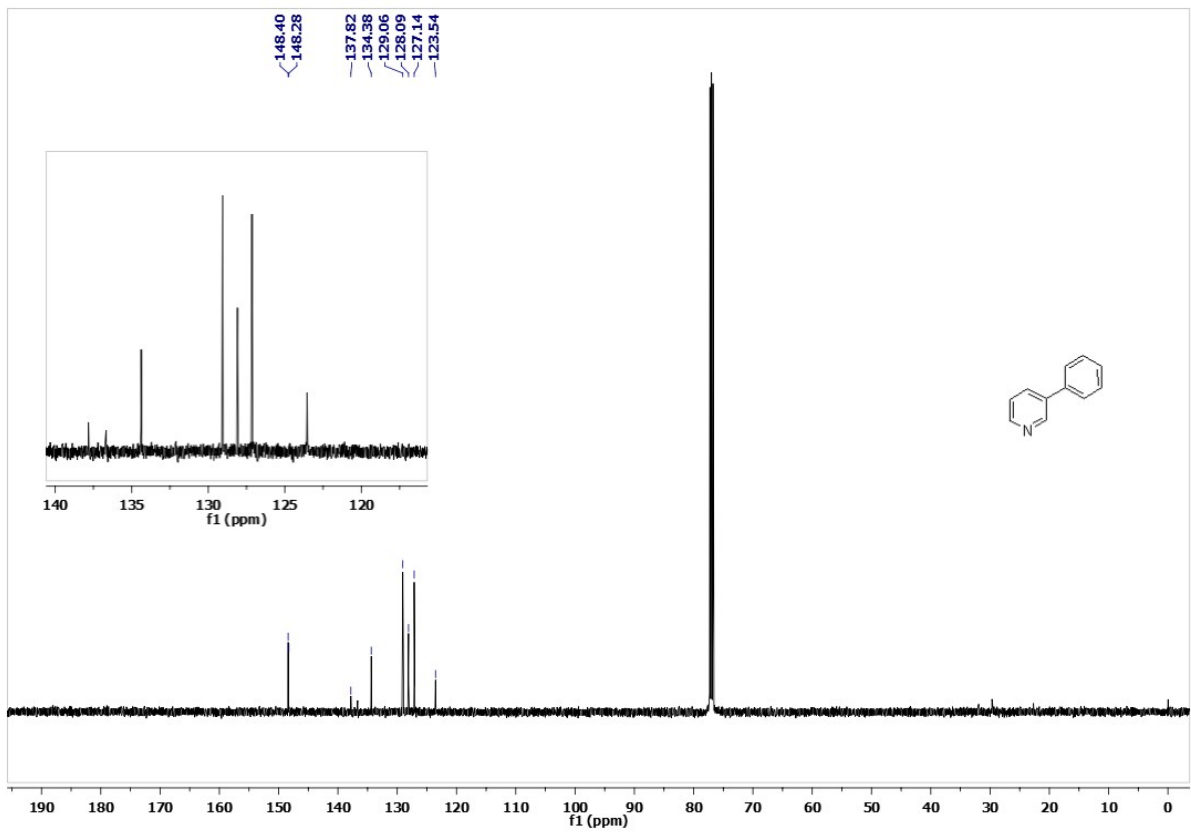
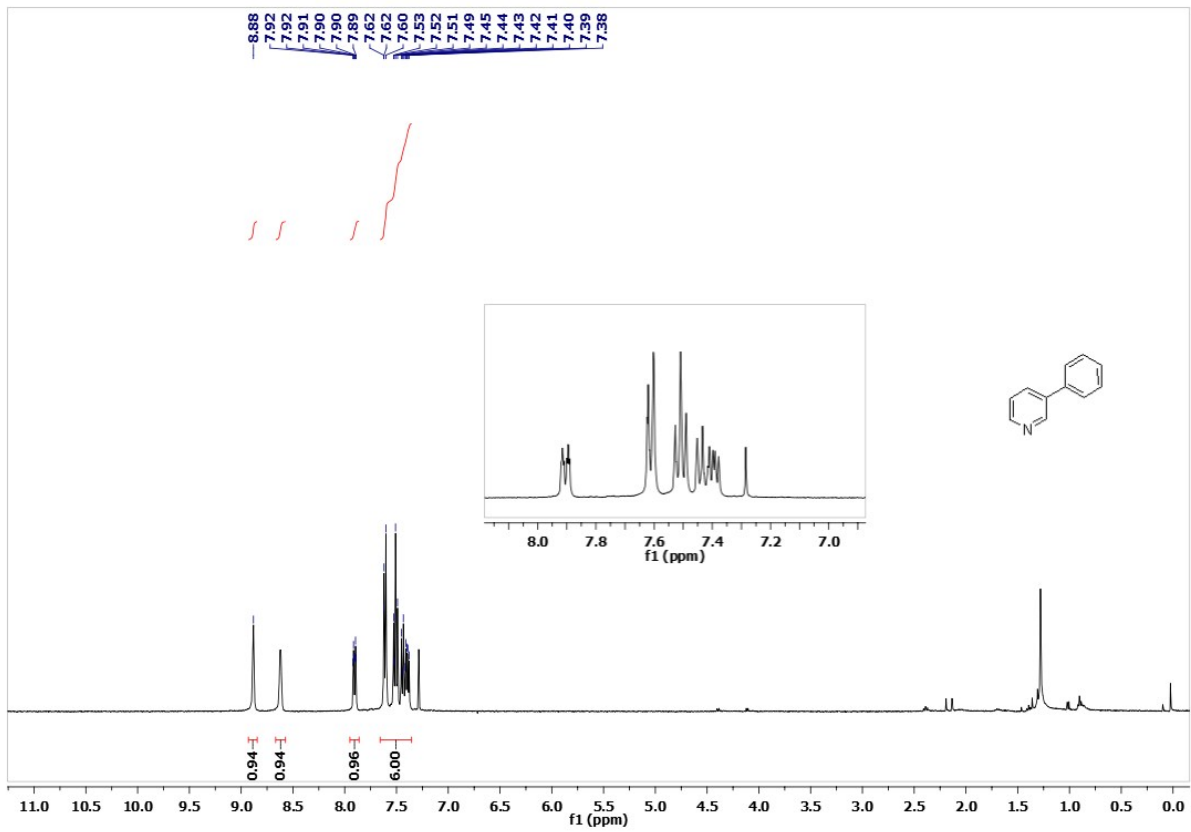


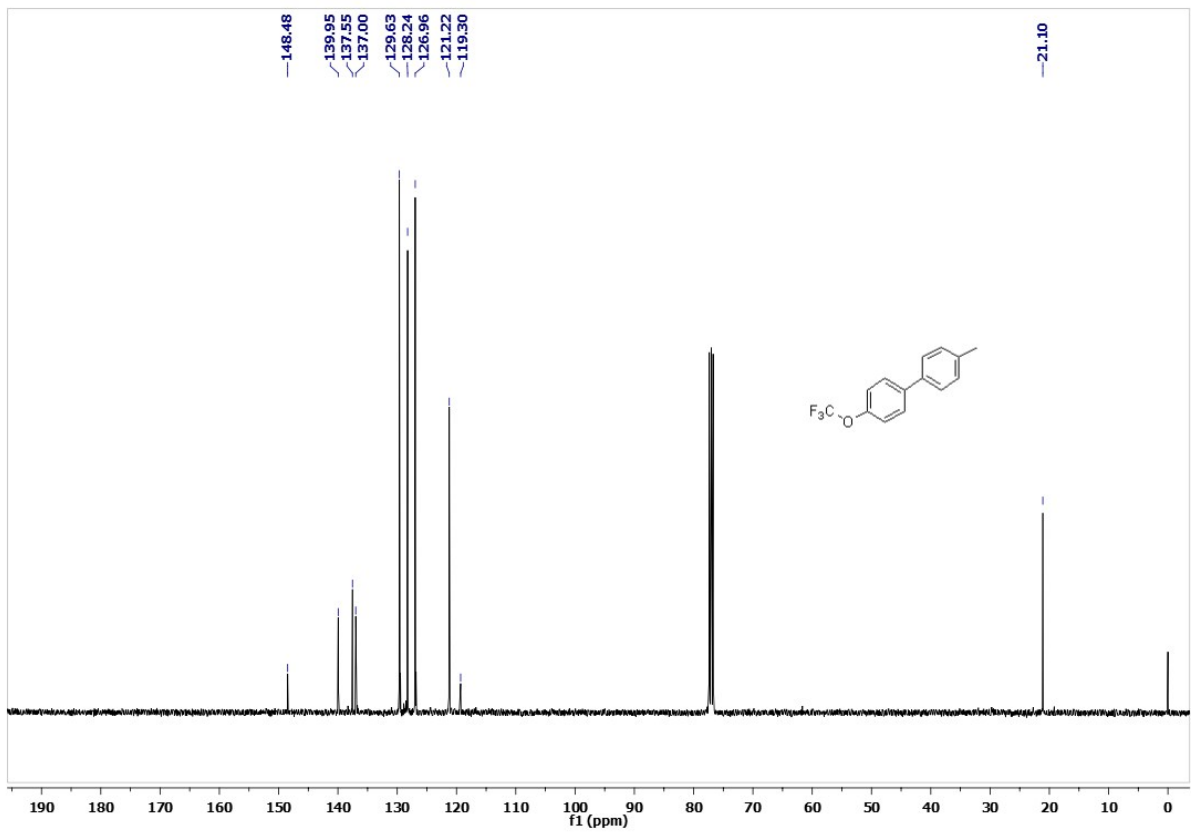
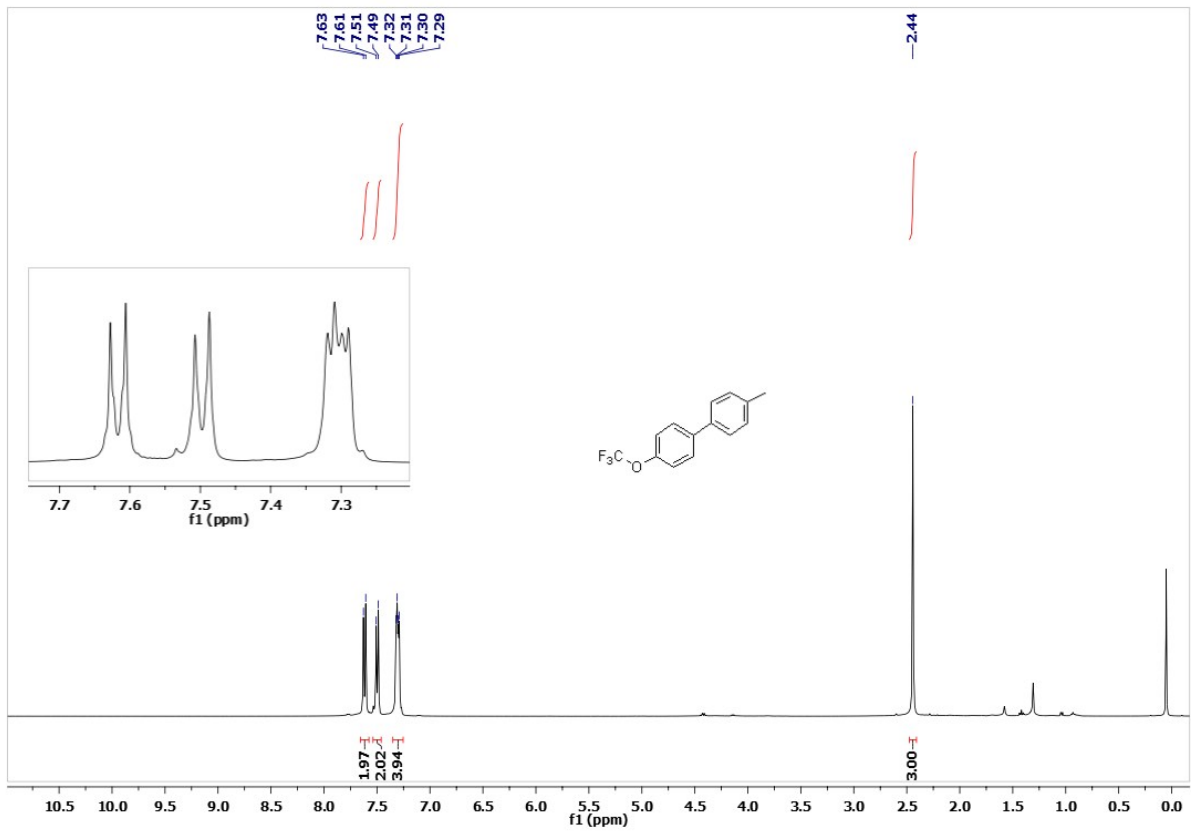


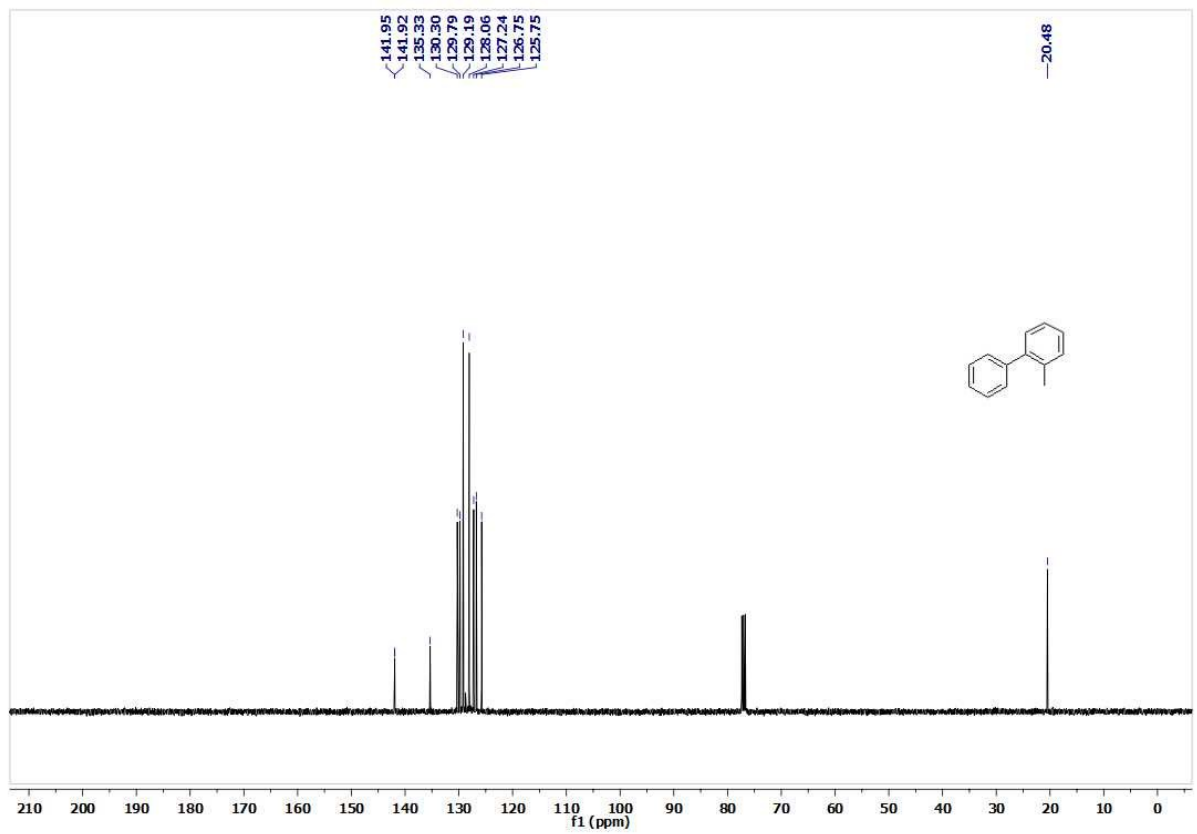
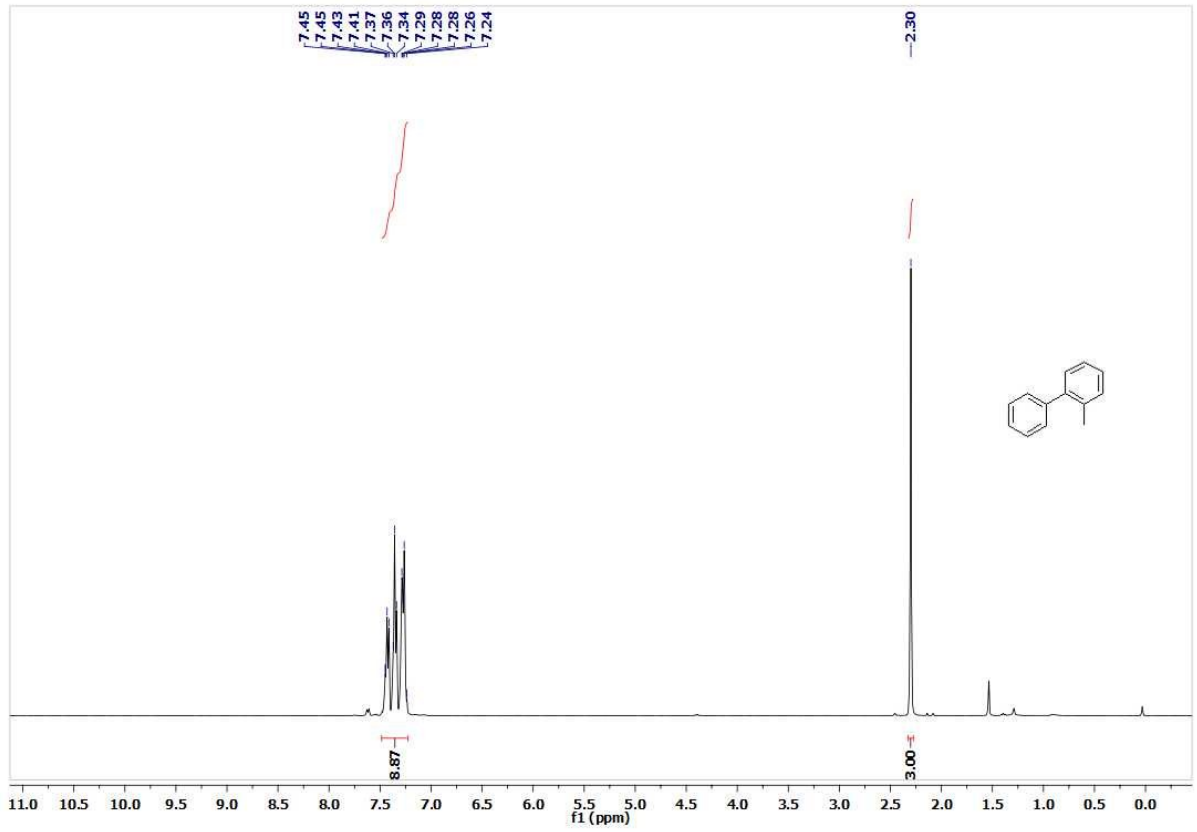


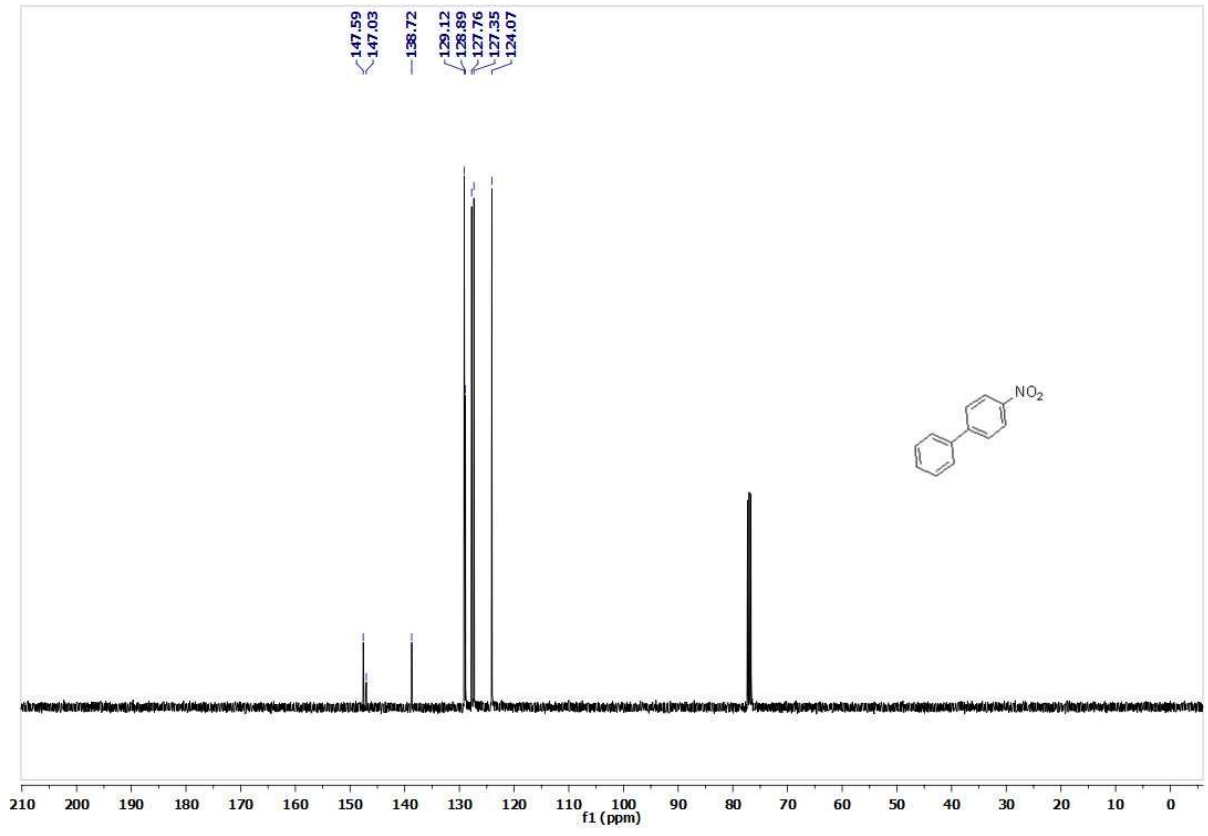
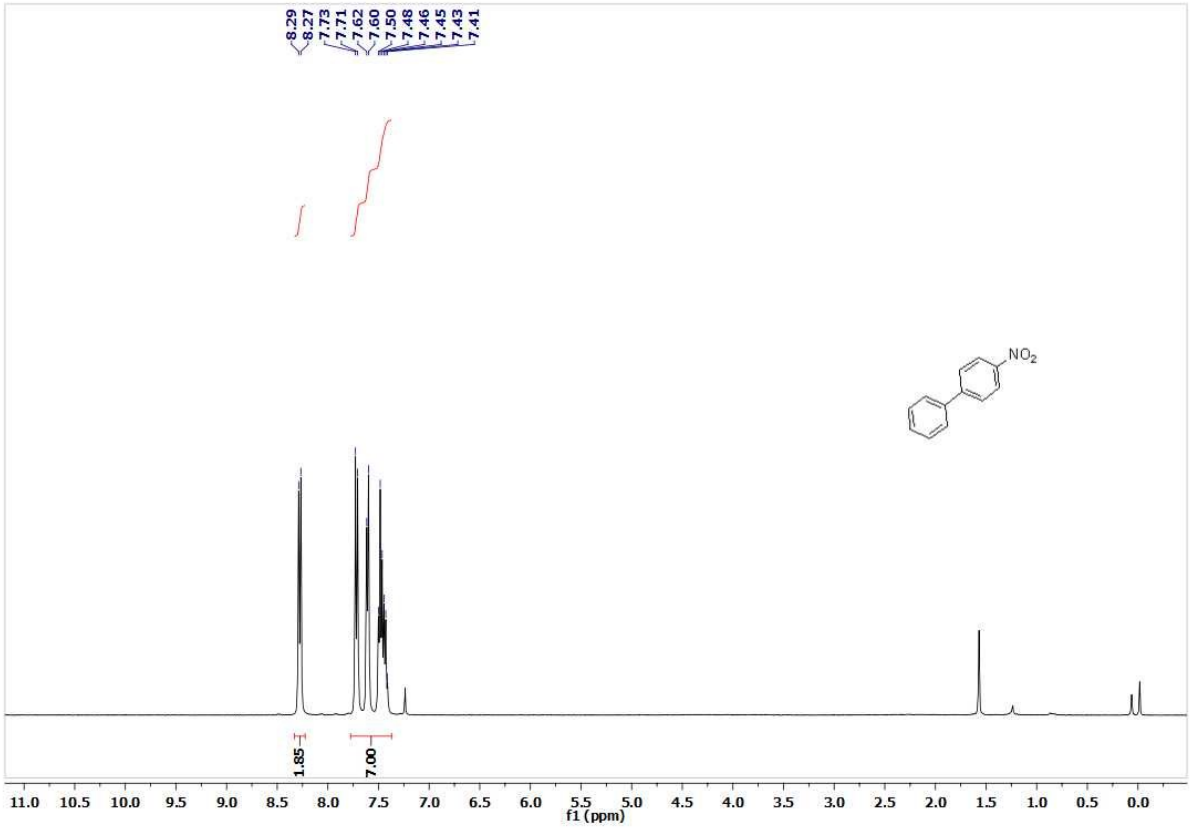


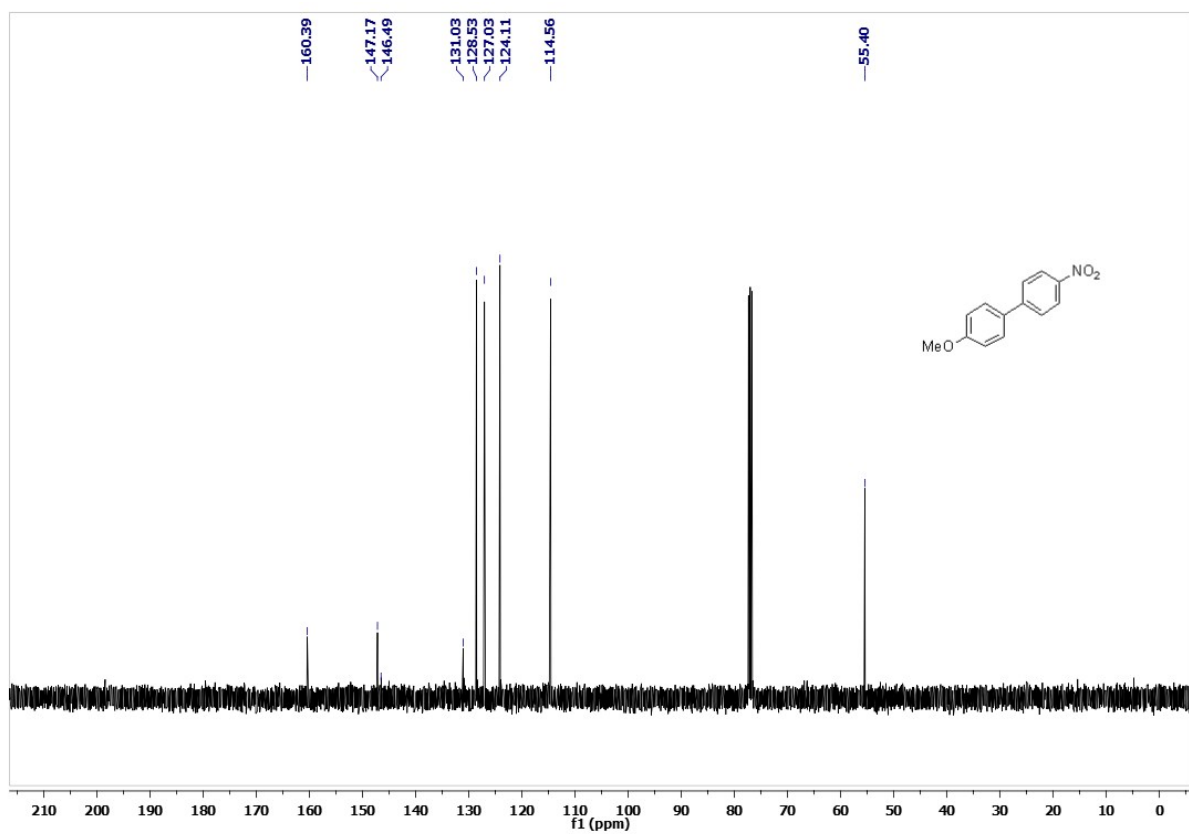
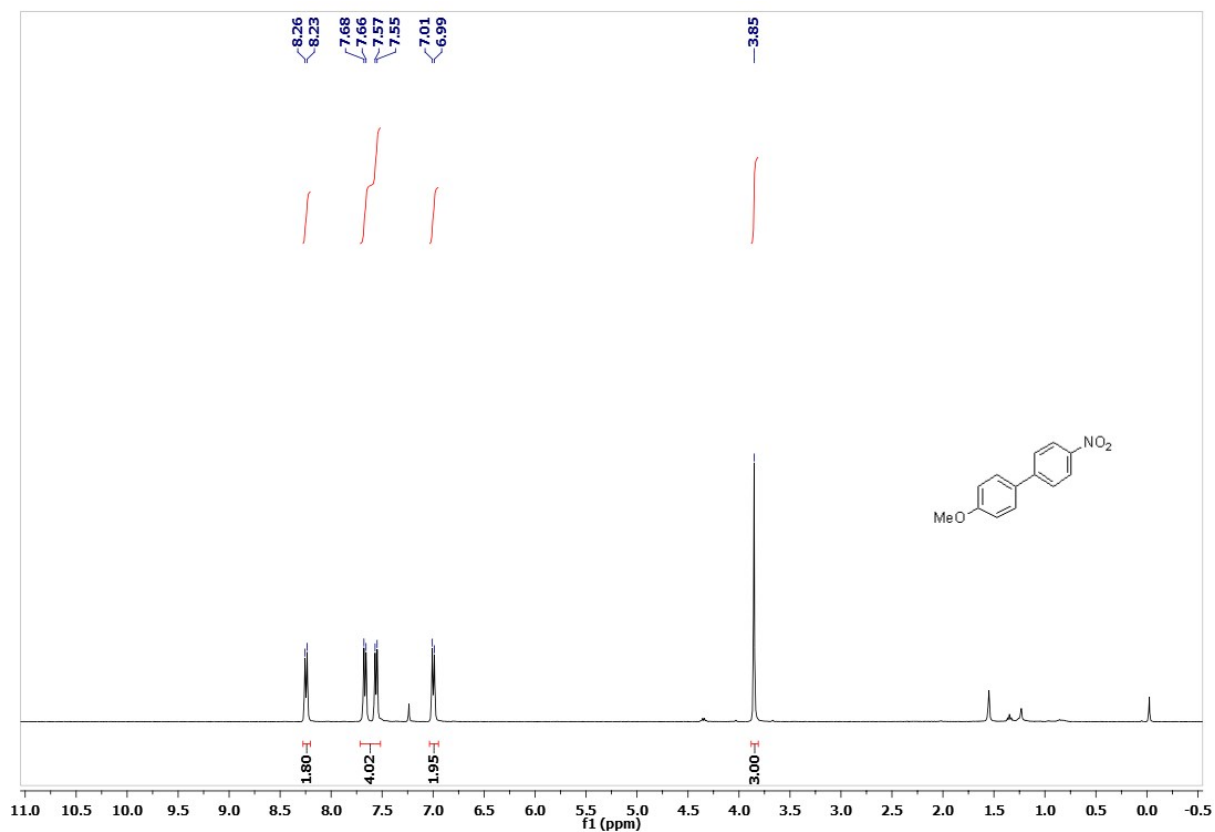


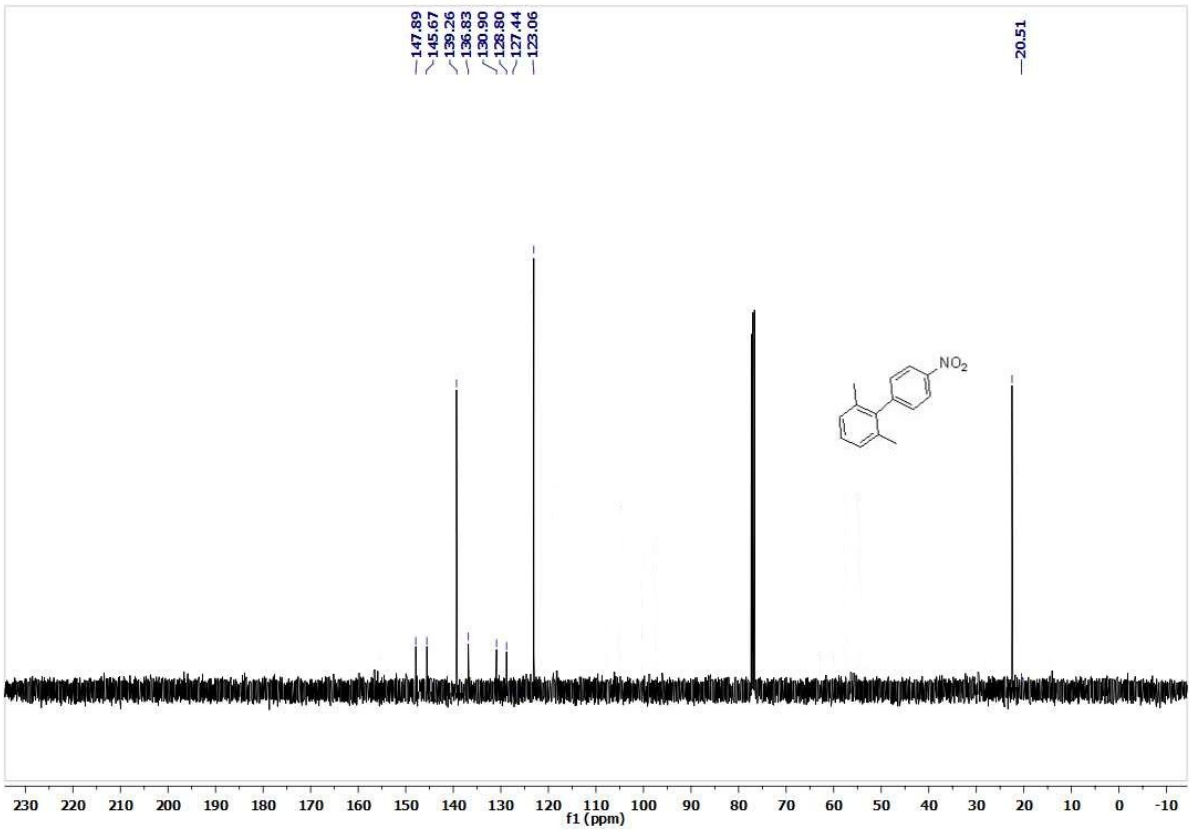
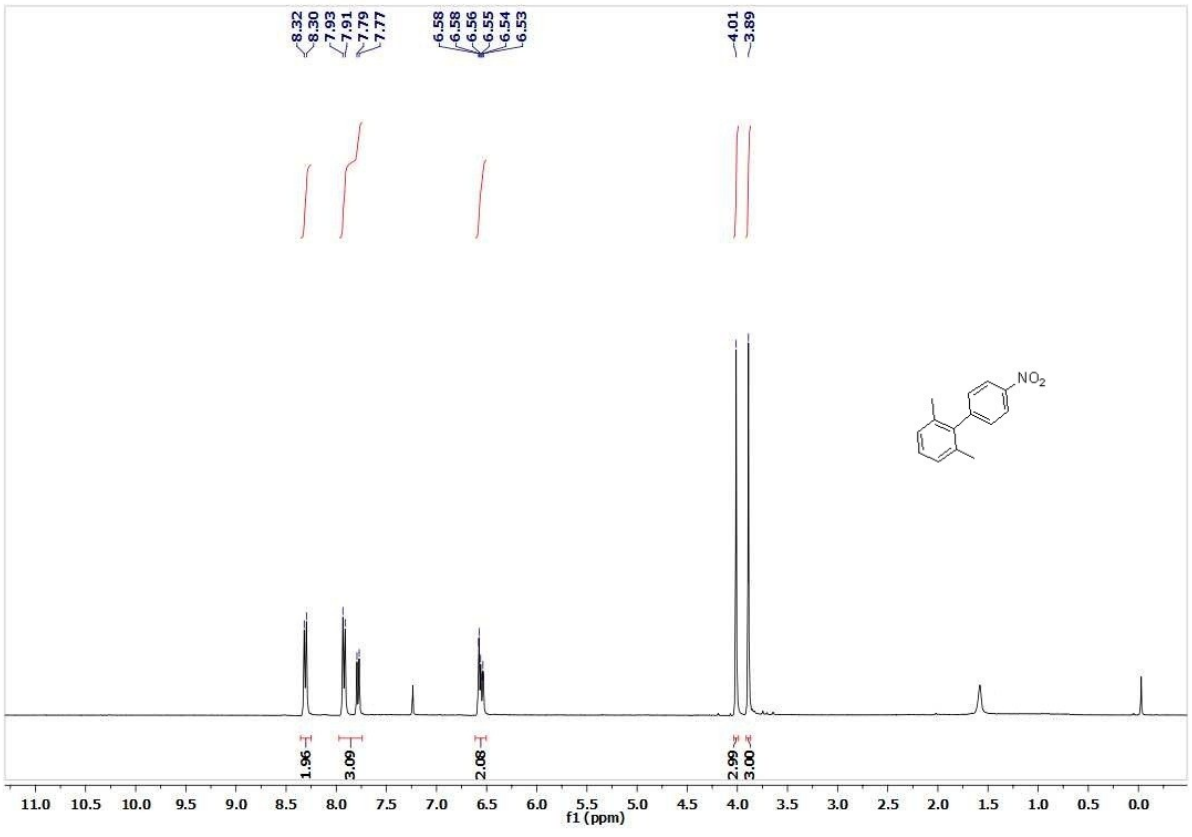


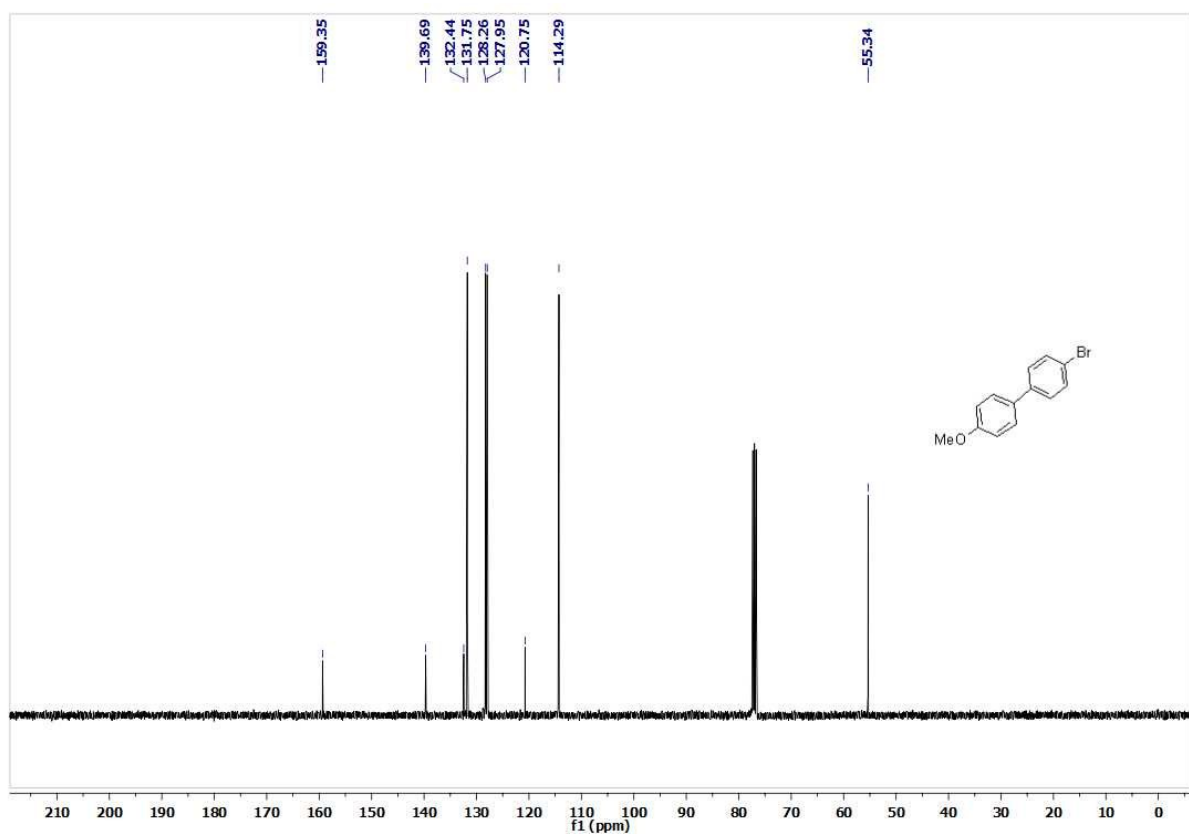
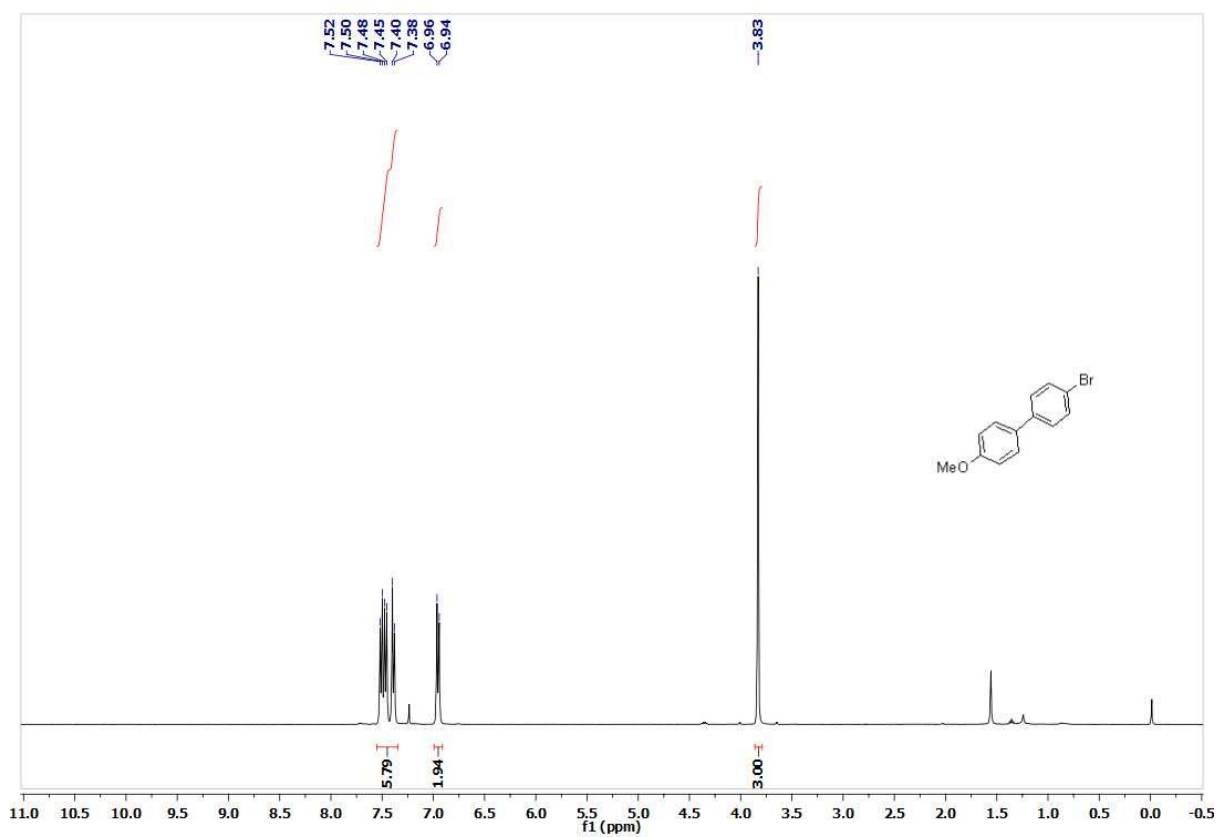


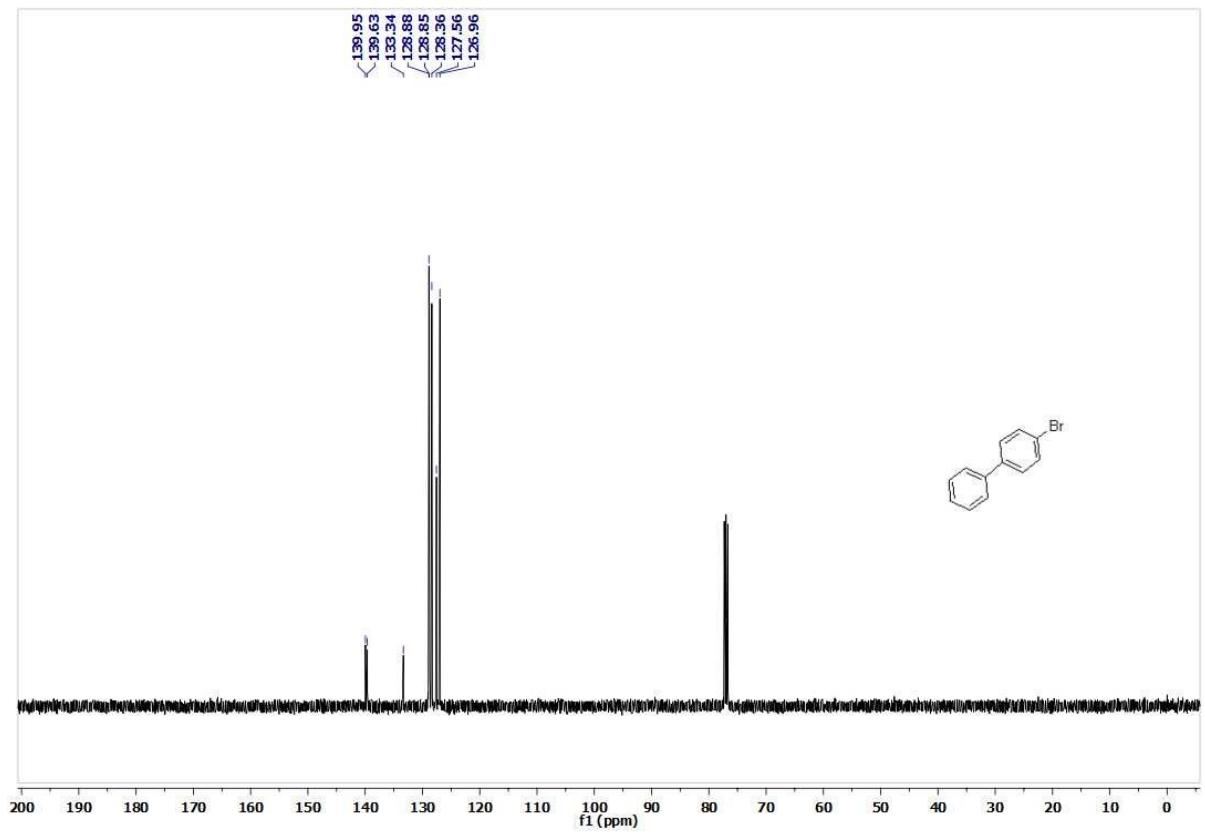
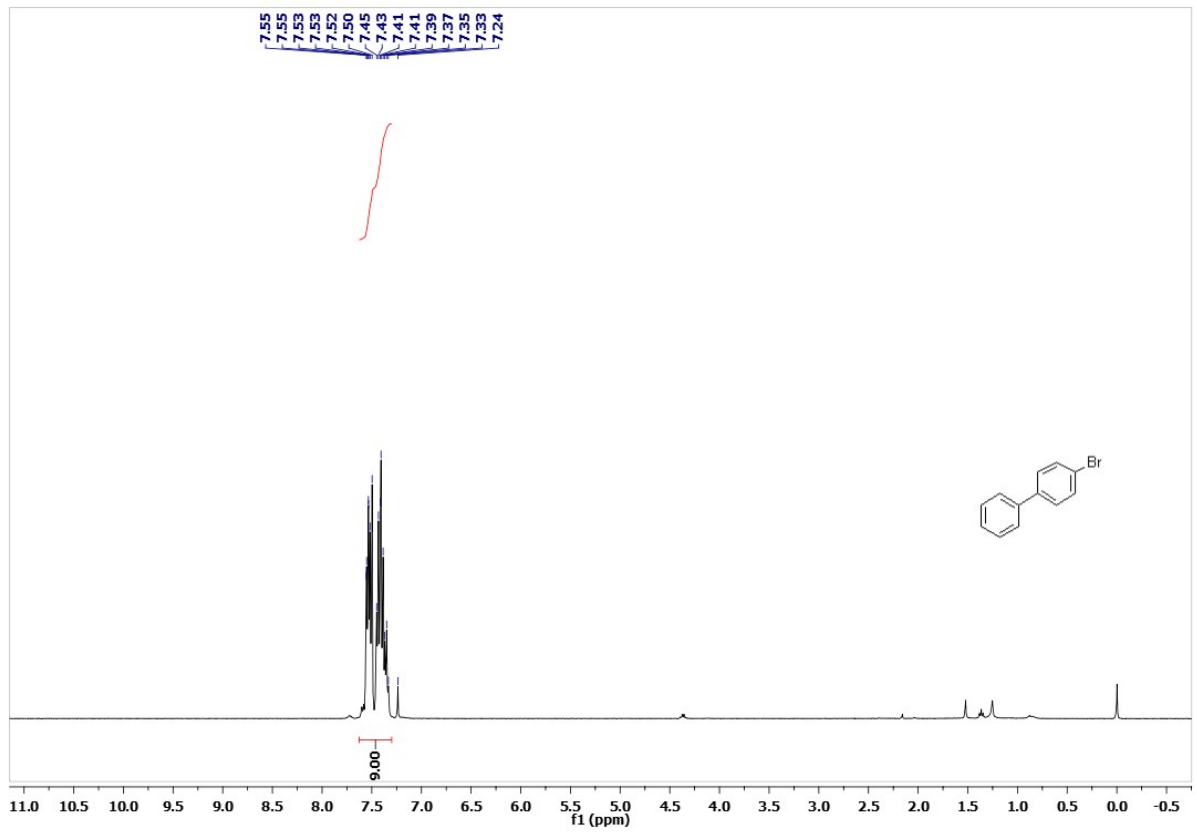


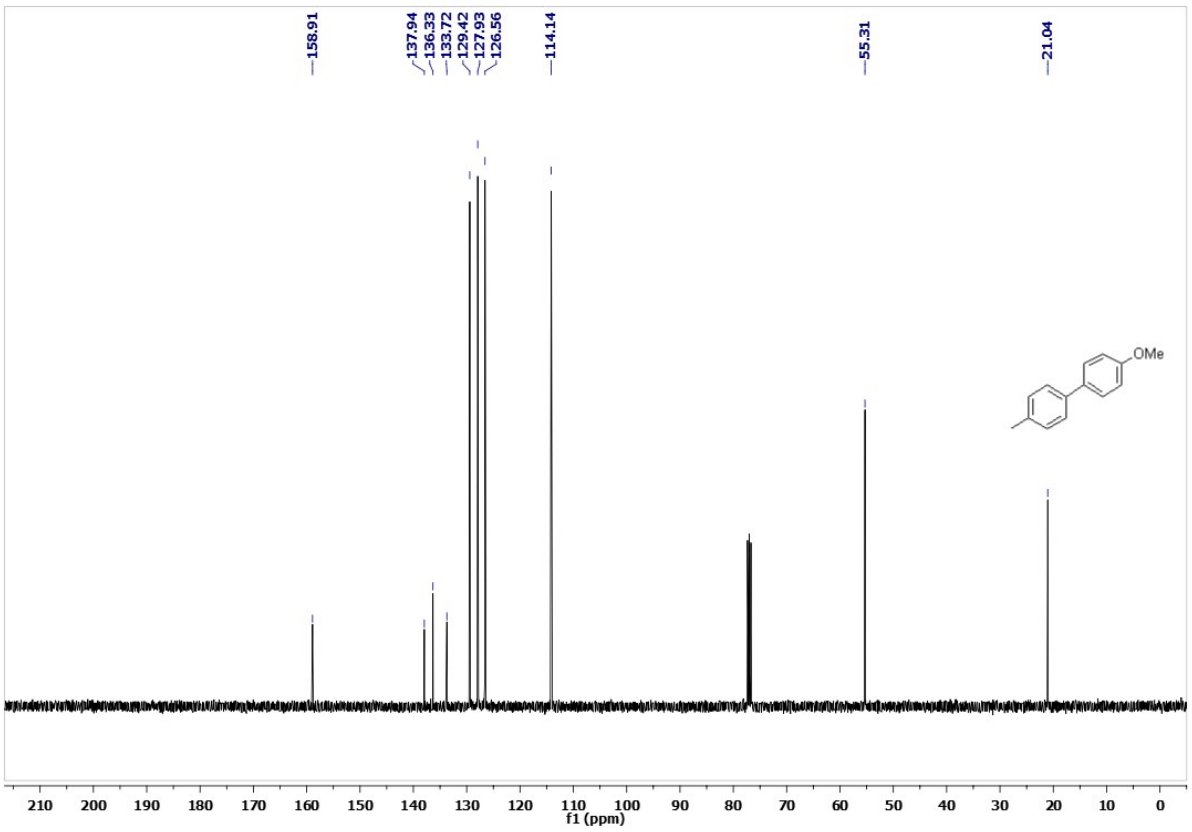
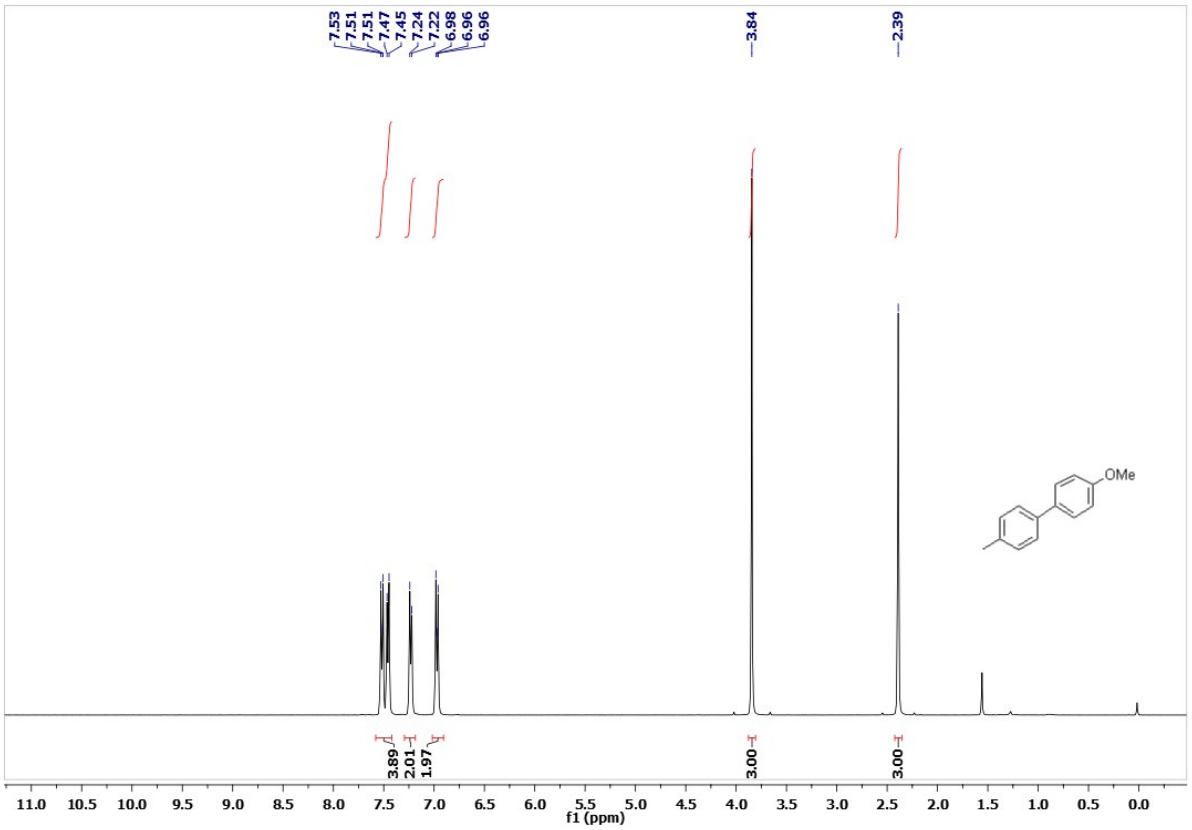


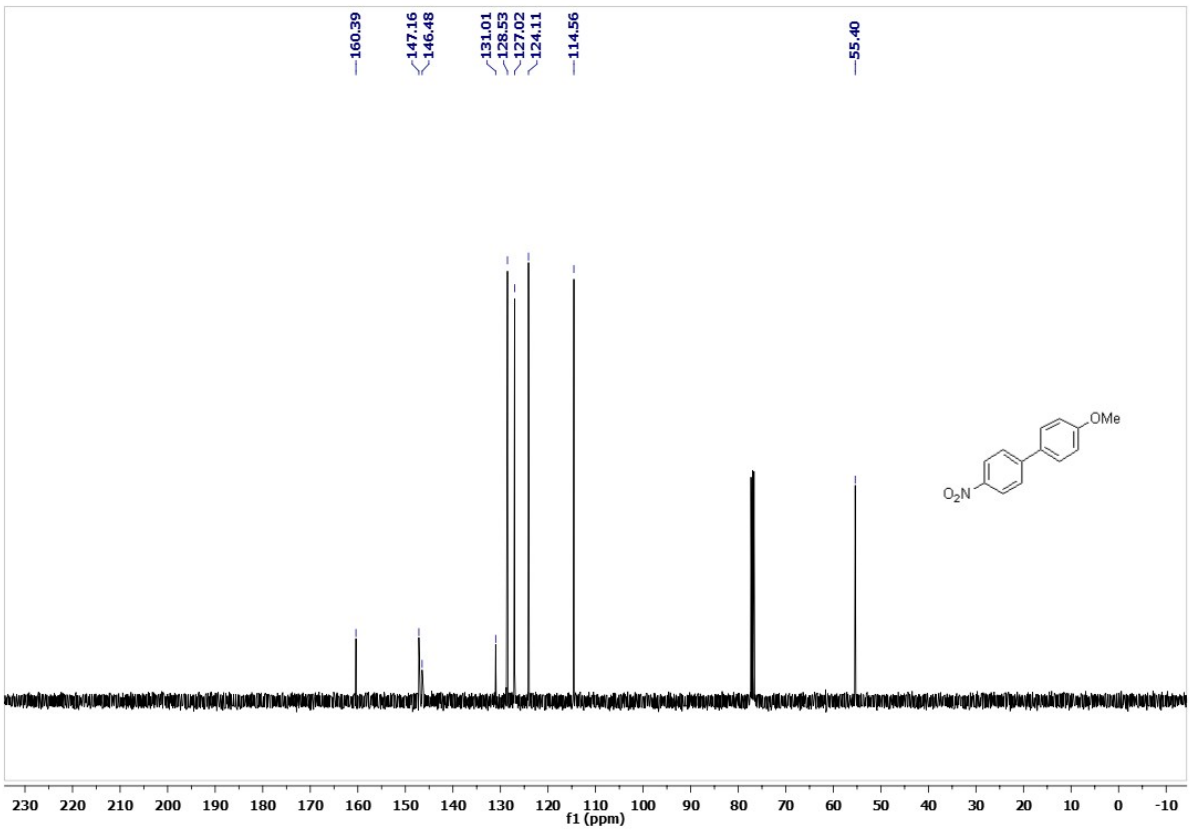
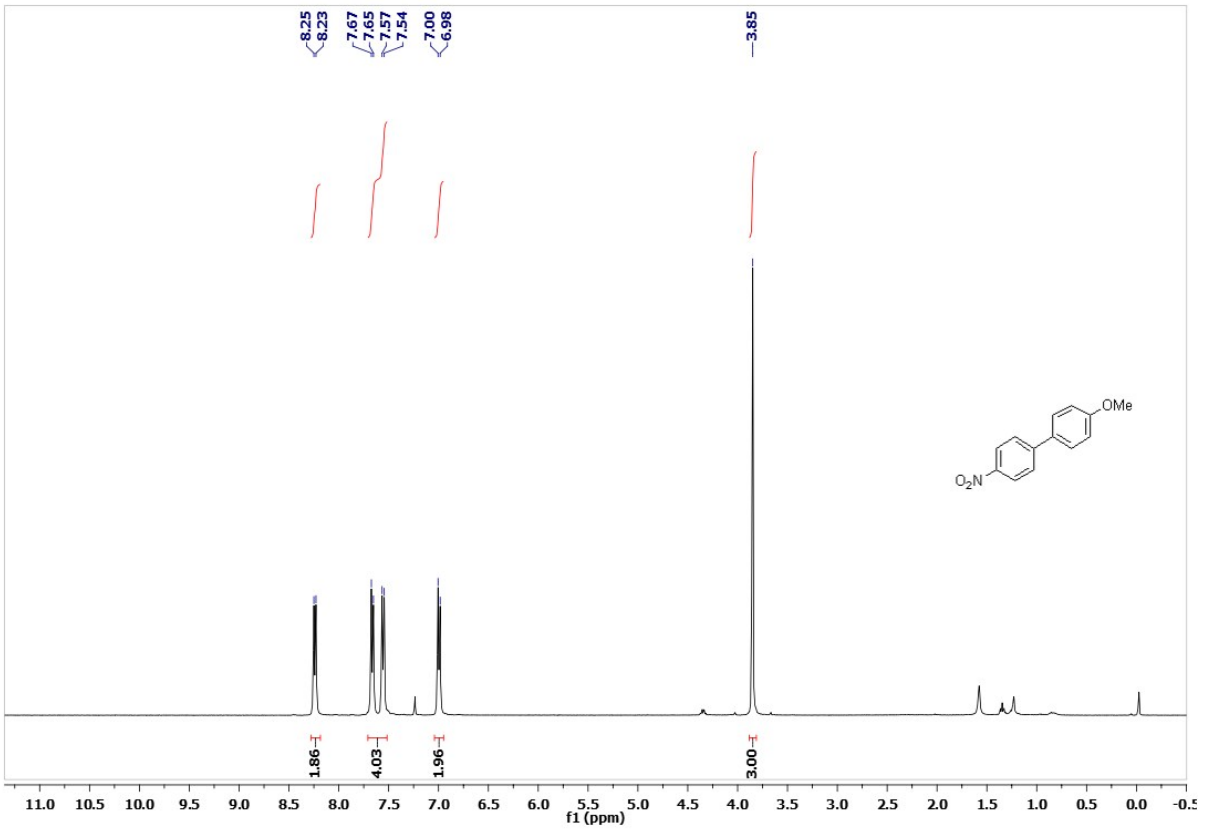


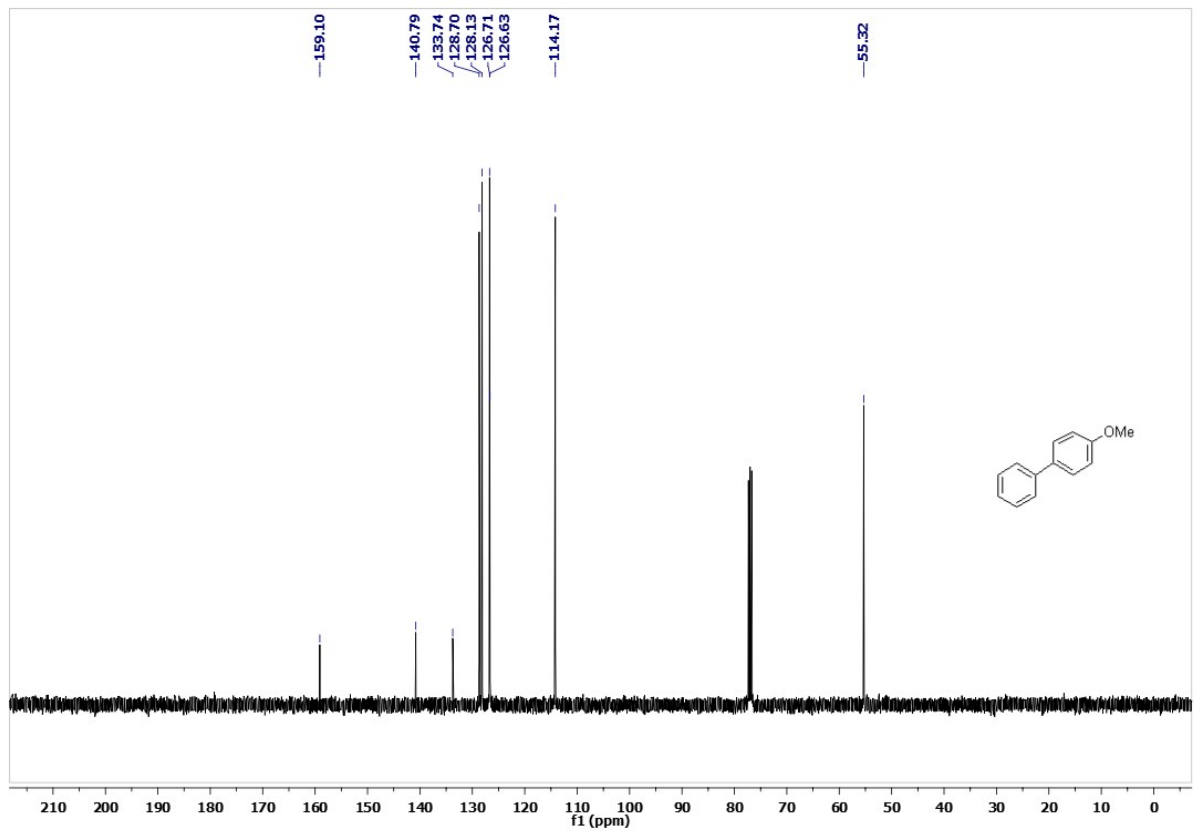
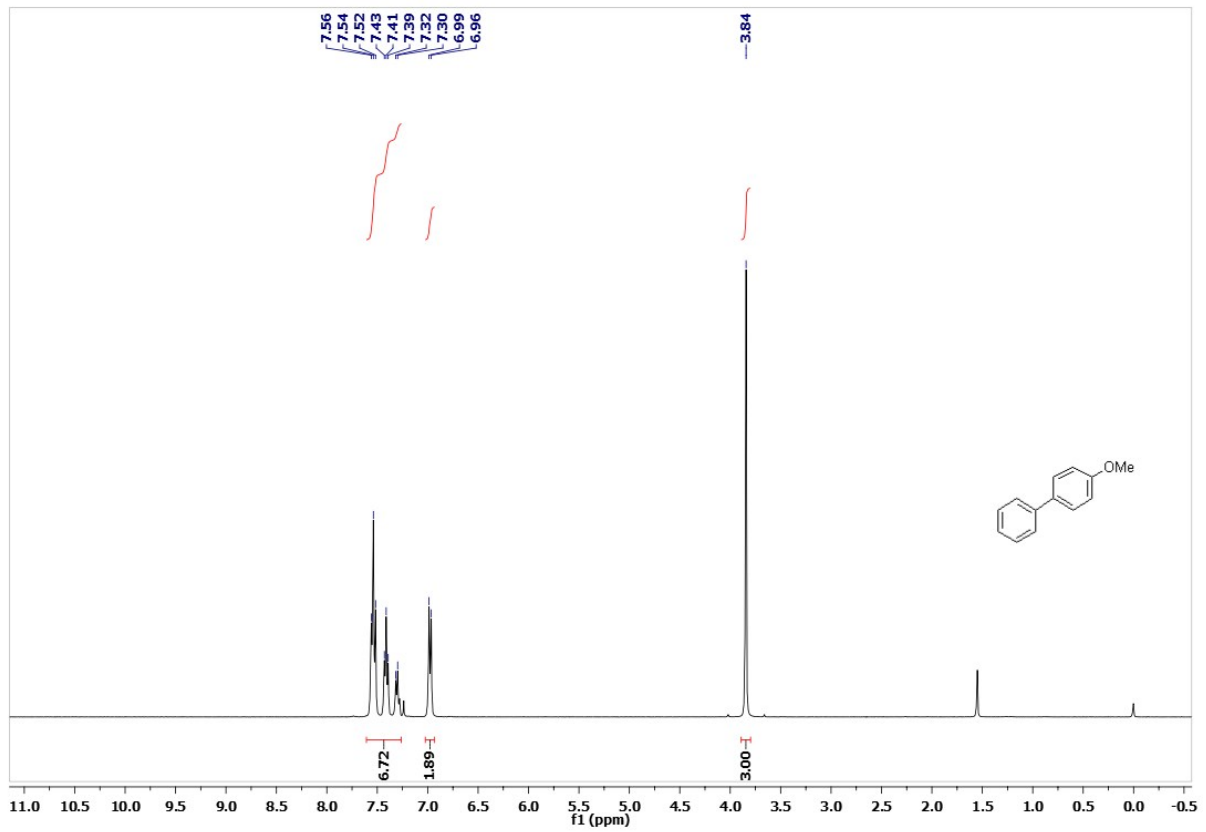


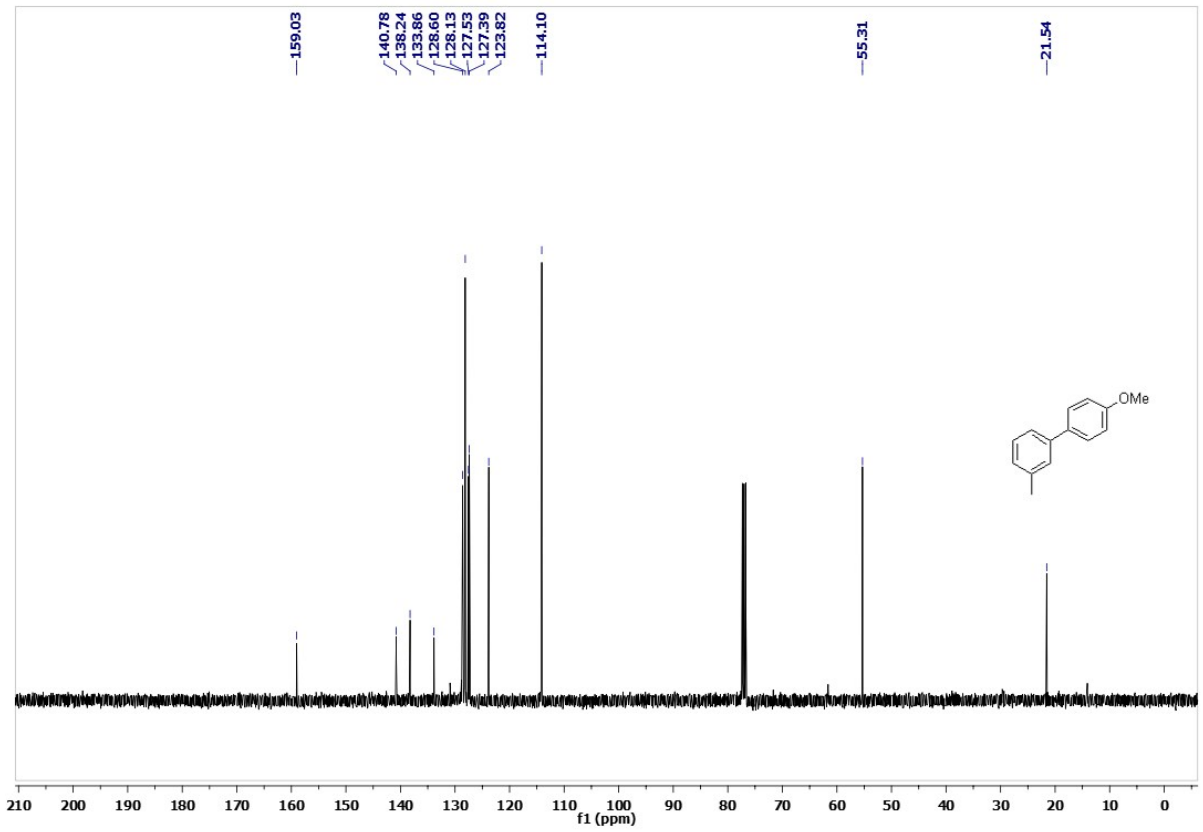
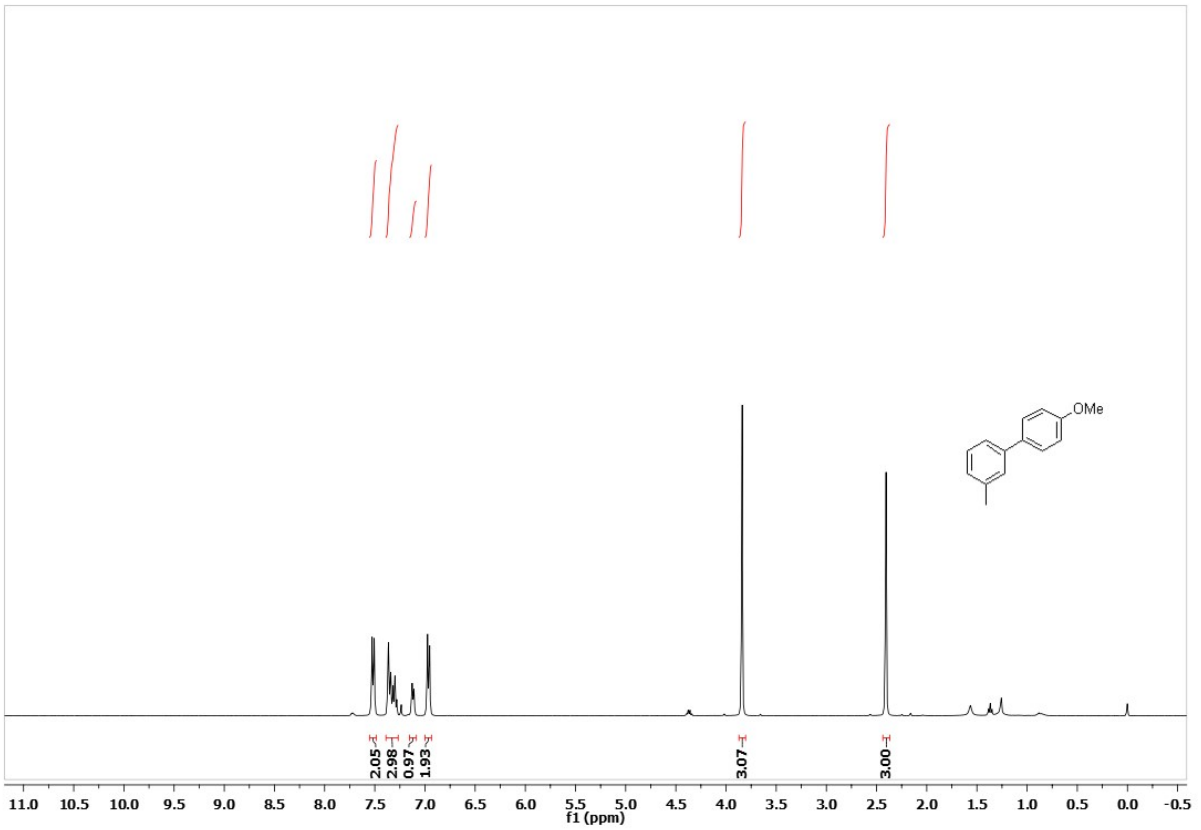


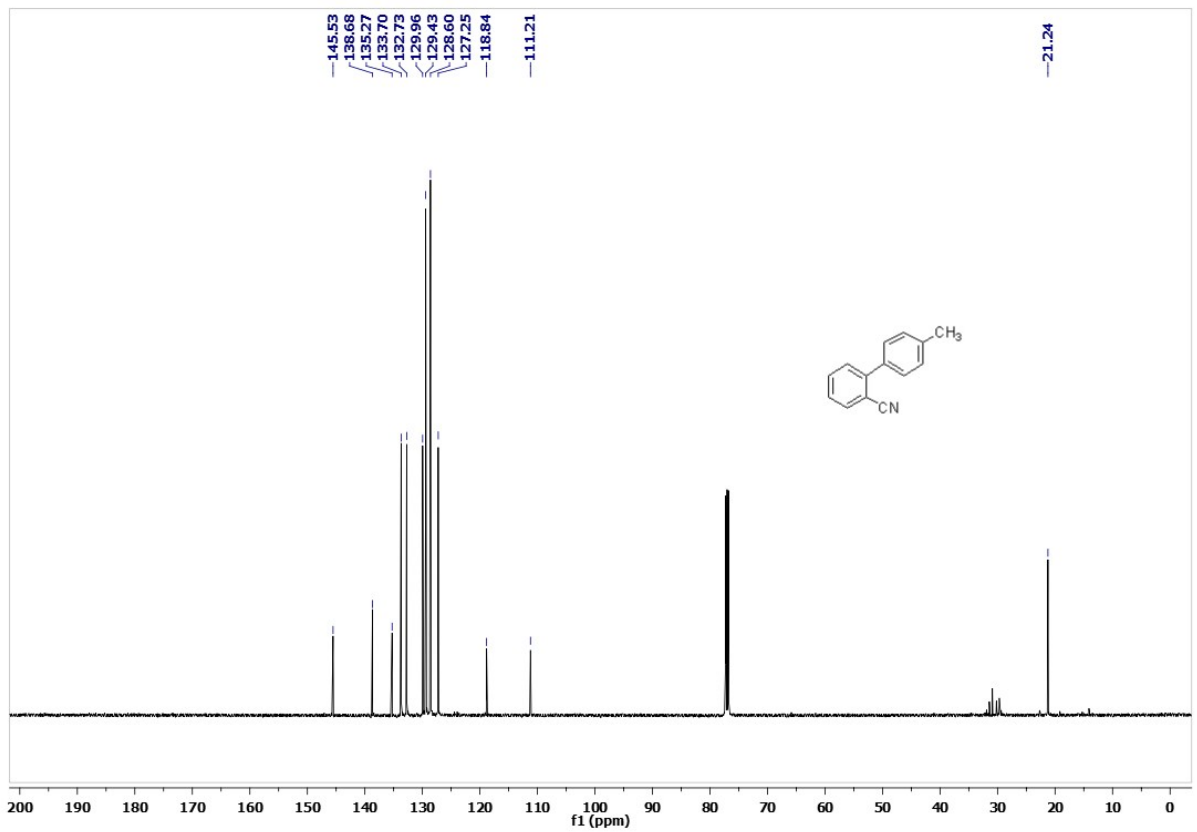
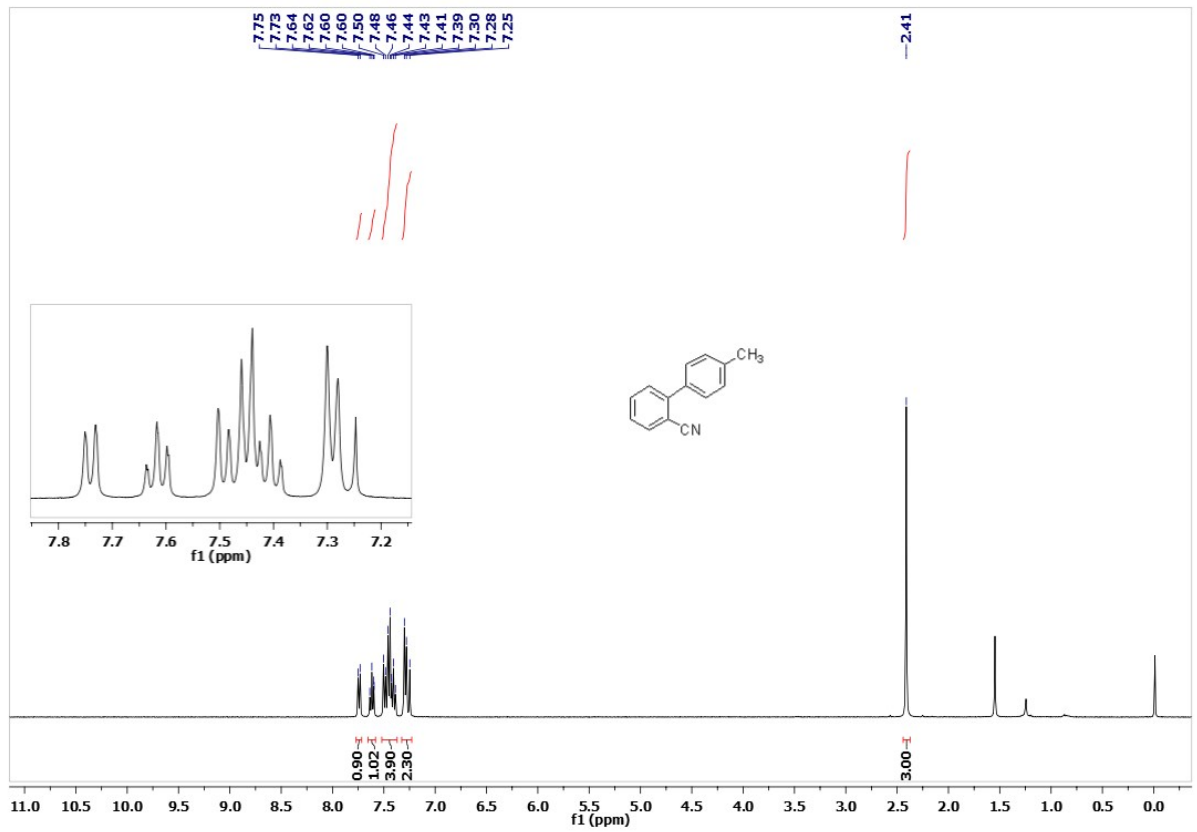












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