

Oligonucleotide length- and probe number-dependent assembly of gold nanoparticle on triangular DNA origami

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1. Materials

1.1 Abbreviations

TAE: tris acetate-EDTA buffer

BSPP: bis (p-sulfonatophenyl) phenylphosphine dihydrate dipotassium salt

TCEP: tris (2-carboxyethyl) phosphine hydrochloride

0.5 × TBE: 89 mM tris, 89 mM boric acid, 2mM EDTA, pH=8.0

AAM: atomic force microscopy

NAS: nucleic acid stain

1.2 Chemicals

Single-stranded M13mp18 DNA was purchased from New England Biolabs (Catalog number: #N4040S). All staple strands and thiolated ssDNA were purchased from Shanghai Sangon Biotech Inc. (Shanghai, China). TAE was purchased from Beyotime Institute of Biotechnology (Shanghai, China). Magnesium acetate tetrahydrate ($C_4H_6MgO_4 \cdot 4H_2O$), sodium citrate tribasic dehydrate ($C_6H_5Na_3O_7 \cdot 2H_2O$), Tris (2-carboxyethyl) phosphine hydrochloride (TCEP), chloroauric acid ($HAuCl_4$), sodium chloride were bought from Aladdin (Shanghai, China). Bis (p-sulfonatophenyl) phenylphosphine dihydrate dipotassium salt (BSPP) was obtained from Sigma-Aldrich (USA).

EDTA, Tris, boric acid, CH₃OH, nucleic acid stain (NAS) are of analytical reagent grade. All solutions were prepared with deionized water ($\geq 18\text{M}\Omega\text{ cm}$) generated from a Millipore Q water purification system.

1.3 Sequence of the staple ssDNA and thiolated oligonucleotides

DNA origami was prepared according to Rothemund's approach with minor modifications^[1].

Staple name	(DNA sequences, from 5' to 3')
t1s18h,D1,	AATACTGCGGAATCGTAGGGGGTAATAGTAAAATGTTTAGACT
t1s28h,E1,	TCTTTGATTAGTAATAGTCTGTCCATCACGCAAATTAACCGTT
t1s8h,A1,	CAGAAGGAAACCGAGGTTTTTAAGAAAAGTAAGCAGATAGCCG
t1s10g,B2,	GACGGGAGAATTAAGTTCGGAATAAGTTTATTTCCAGCGCC
t1s12i,C2,	TCATATGTGTAATCGTAAAAGTATGTCATTTTC
t1s14i,D2,	GTGAGAAAATGTGTAGGTAAAGATACAACCTTT
t1s16i,E2,	GGCATCAAATTTGGGGCGCGAGCTAGTTAAAG
t1s18i,A2,	TTCGAGCTAAGACTTCAAATATCGGGAACGAG
t1s20g,G2,	GAATACCACATTCAACTTAAGAGGAAGCCCGATCAAAGCG
t1s22i,H2,	TCGGGAGATATACAGTAACAGTACAAATAATT
t1s24i,A3,	CCTGATTAAAGGAGCGGAATTATCTCGGCCTC
t1s26i,B3,	GCAAATCACCTCAATCAATATCTGCAGGTCGA
t1s28i,C3,	CGACCAGTACATTGGCAGATTCACCTGATTGC
t1s2i,D3,	CGGGGTTTCCTCAAGAGAAGGATTTTGAATTA
t1s30g,E3,	TTGACGAGCACGTATACTGAAATGGATTATTTAATAAAAAG
t1s4i,A3,	AGCGTCATGTCTCTGAATTTACCGACTACCTT
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t2s17f,D4,	AACCAGACGTTTAGCTATATTTTCTTCTACTA

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t2s23g,G4, TGGCAATTTTTAACGTCAGATGAAAACAATAACGGATTTCG
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t3s14e,A5, CAATATGACCCTCATATATTTTAAAGCATTAA
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t3s20g,A6, CGCCAAAAGGAATTACAGTCAGAAGCAAAGCGCAGGTCAG
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t4s17f,D7, GATTAGAGATTAGATACATTTTCGCAAATCATA
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t4s21g,A7, GCGCAGAGGCGAATTAATTATTTGCACGTAAATTCTGAAT
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t4s5f,C8, CTCAGAGCATATTCACAAACAAATTAATAAGT
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t-5s12e-t6s3c-3T,E6, TGTAGCATTCTTTTATAAACAGTT

2. Preparation of DNA origami

The single-strand M13mp18 virus DNA and the staple strands (including the modified ones) were mixed in a molar ratio of 1:10 in the $1 \times \text{TAE} \sim \text{Mg}^{2+}$ buffer (40 mM Tris, 20 mM acetic acid, 1 mM EDTA, 12.5 mM magnesium acetate). The mixture was annealed from 95°C to 20°C in a PCR instrument (1°C/100s).^[1] The products were filtered with centrifugal filters using the washing buffer ($1 \times \text{TAE} \sim \text{Mg}^{2+}$ buffer) for three times to remove excess staple strands.^[2]

3. Modification of AuNPs

AuNP-DNA conjugates were synthesized according to Ding's route.^[3] Briefly, 15 mg BSPP was dissolved in a 50 mL AuNPs solution, followed by a stirring in dark at room temperature for 12 hours. Solid sodium chloride was added in slowly until the color of the solution changes from wine red to lilac. The products were collected by centrifugation at 10000 rpm for 15 min and re-suspended in a mixture of BSPP and CH₃OH. After a second round of centrifugation, the AuNPs were suspended in 1mL BSPP (2.5mM). Thiolated oligonucleotides were activated with TCEP (10mM) for several hours and then incubated with the AuNPs at a molar ratio of 200:1 in a $0.5 \times$ TBE buffer containing 50 mM NaCl for 40 hours at room temperature. After washing with the buffer for three times, the oligonucleotides-conjugated AuNPs were harvested via centrifugation. The UV-vis spectra of the final products were measured to determine the concentration according to Wolfgang Haiss's method.^[4]

4. Assembly of AuNPs with the modified DNA origami templates

The conjugated AuNPs were mixed with the modified DNA origami templates. The mixture was cooled from 43 to 20 °C to allow the hybridization of the complementary ssDNA for the assembly of AuNPs with the modified DNA origami templates.^[3]

5. AFM characterization

A freshly cleaved mica was pre-treated with a MgCl₂ solution. The adsorbed Mg²⁺ ions could facilitate the attachment of negatively charged DNA origami and AuNPs. Then, 10 μL sample were dropped on the mica surface, drying in air for 6 hours.^[5] The AAM images were scanned with a Dimension Icon scanning probe microscope (Bruker, USA) at ambient conditions.

6. Gel electrophoresis

The resulting DNA origami-AuNPs were loaded on 1.0% agarose gels with (5 μ L NAS) and run for three hours at 100V. ^{[6] [7]} The gel images were photographed using a digital camera system (Chemi XR5).

7. Data analysis

Data were collected from at least 4 runs of independent experiments. Differences between different groups were tested for significance using Student's t test (Origin 6.0).

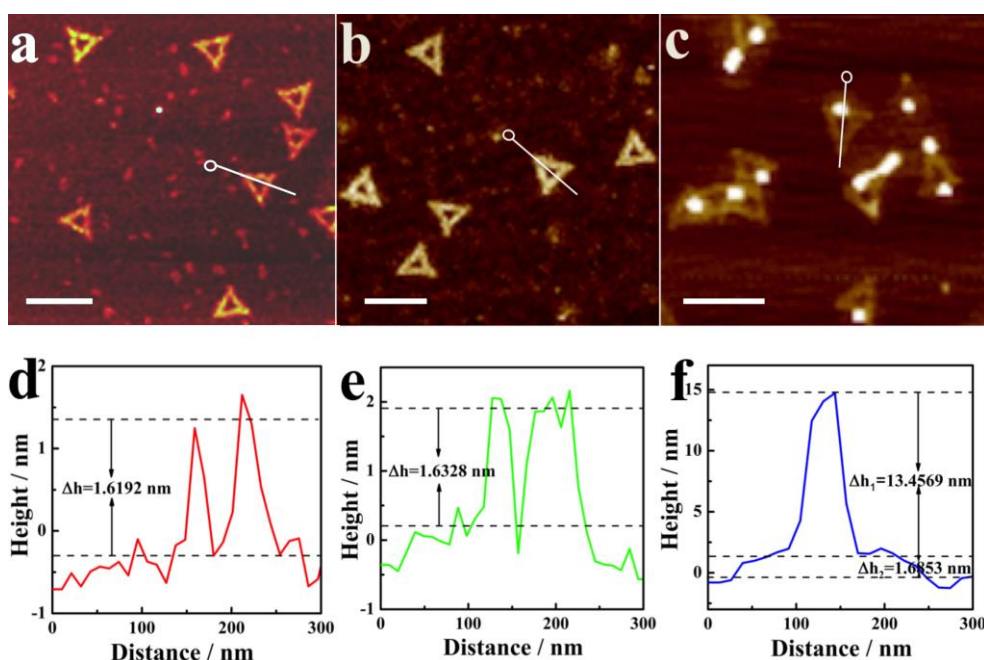


Fig. S2 AAM images of (a) DNA origami triangles, (b) DNA origami triangles modified with a single strand DNA probe and (c) AuNP-attached DNA origami triangles. Height profiles of (d) a DNA origami triangle, (e) a DNA origami triangle modified with a single strand DNA probe and (f) a AuNP-attached DNA origami triangle.

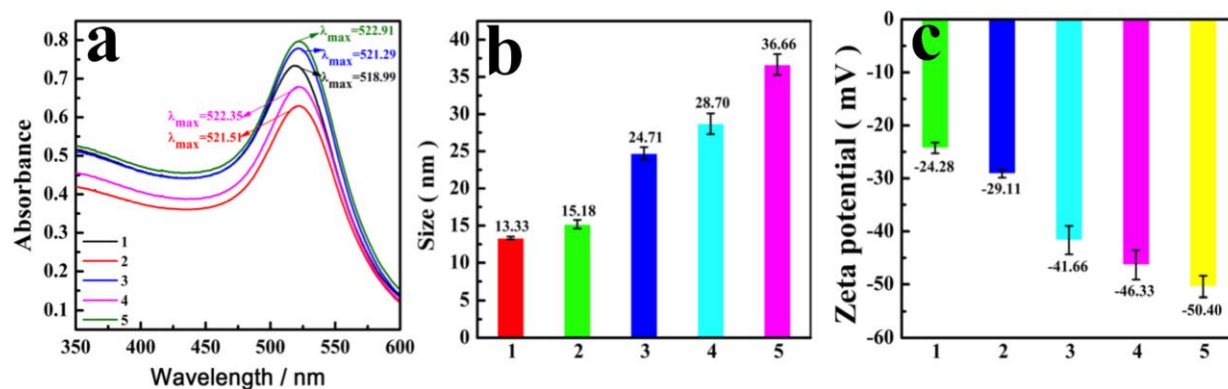


Fig. S3 UV-vis spectra (a), hydrodynamic size (b) and zeta potential (c) of AuNPs before and after modification with BSPP and ssDNA. Data in (b) and (c) are presented as the average \pm standard deviation from three independent measurements. The numbers 1, 2, 3, 4 and 5 correspond to AuNPs, BSPP-AuNPs, 20-mer DNA-BSPP-AuNPs, 40-mer DNA-BSPP-AuNPs and 60-mer DNA-BSPP-AuNPs, respectively.

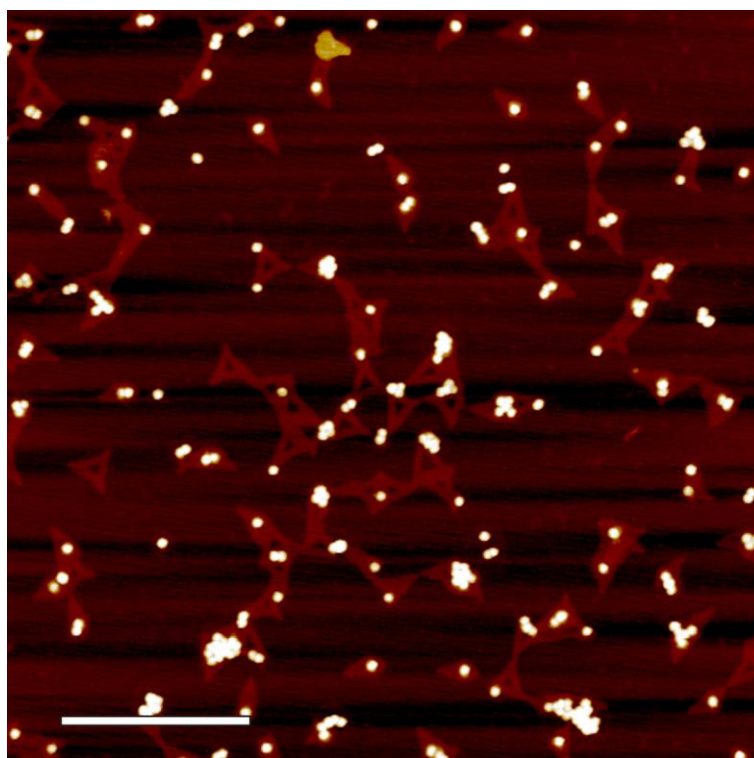


Fig. S4 AAM image of the DNA origami attached 40-mer DNA-BSPP-AuNPs.

References

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