

Supplementary Information

# Synthesis in living cells with the assistance of supramolecular nanocarriers

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• Experimental Procedures.....	S2
• Nuclear Magnetic Resonance Spectra .....	S4
• Crystallographic Data.....	S5
• Absorption and Emission Spectra .....	S6
• Fluorescence Images .....	S7

## Experimental Procedures

**Materials and methods.** Chemicals were purchased from commercial sources and used as received with the exception of MeCN, which was distilled over CaH<sub>2</sub>, and H<sub>2</sub>O, which was purified with a Barnstead International NANOpure Diamond Analytical system. Compounds **1** and **3** were prepared according to literature procedures.<sup>S1,S2</sup> EISMS was performed with a Bruker micrOTO-Q II spectrometer. NMR spectra were recorded with a Bruker Avance 400 spectrometer. Fourier transform infrared spectra (FTIR) spectra were recorded on neat samples with a Perkin Elmer Frontier spectrometer.

**7-Nitro-*N*-(triphenylphosphoranylidene)benzo[*c*][1,2,5]oxadiazol-4-amine (**2**).** Ph<sub>3</sub>P (65 mg 0.4 mmol) was added to a solution of **1** (52 mg, 0.4 mmol) in MeCN (20 mL) and the mixture was stirred for 30 min at ambient temperature. The resulting precipitate was filtered off, washed with hexane (20 mL) and dried to afford **2** (88 mg, 80%) as a red solid. ESIMS:  $m/z = 441.1117$  [M + H]<sup>+</sup> ( $m/z$  calcd. for C<sub>24</sub>H<sub>18</sub>N<sub>4</sub>O<sub>3</sub>P = 441.1133); <sup>1</sup>H NMR [(CD<sub>3</sub>)<sub>2</sub>SO]:  $\delta = 5.97$  (1H, d, 8 Hz), 7.64–7.69 (6H, m), 7.79–7.74 (3H, m), 7.85–7.90 (6H, m), 8.25 (1H, d, 8 Hz) ppm; <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 110.1, 110.3, 123.1, 126.2, 127.2, 128.3, 129.3, 129.5, 132.6, 132.7, 133.3, 133.4, 135.7, 144.7, 144.8, 150.3, 150.5, 153.9$  ppm; <sup>31</sup>P NMR [(CD<sub>3</sub>)<sub>2</sub>SO]:  $\delta = 15.03$  ppm; FTIR:  $\bar{\nu} = 1610, 1512, 1479, 1437, 1294, 1249, 1098, 995, 916, 802$  cm<sup>-1</sup>.

**Crystallographic analysis.** Red single crystals suitable for X-ray diffraction analysis were obtained after vapor diffusion of Et<sub>2</sub>O into a CHCl<sub>3</sub> solution **2**. The data crystal was glued onto the end of a thin glass fiber. X-Ray intensity data were measured with a Bruker SMART APEX2 CCD-based diffractometer, using Mo K $\alpha$  radiation ( $\lambda = 0.71073$  Å).<sup>S3</sup> The raw data frames were integrated with the SAINT+ program by using a narrow-frame integration algorithm. Corrections for Lorentz and polarization effects were also applied with SAINT+. An empirical absorption correction based on the multiple measurement of equivalent reflections was applied using the program SADABS. The structure was solved by a combination of direct methods and difference Fourier syntheses and refined by full-matrix least-squares on F<sup>2</sup> with the SHELXTL software package.<sup>S4</sup> Crystal data, data collection parameters and results of the analyses are listed in Table S1. The compound crystallized in the monoclinic crystal system and the space group *P2*<sub>1</sub>/*n* was chosen based on the systematic absences in the intensity data. All non-hydrogen atoms were refined with anisotropic displacement parameters. Hydrogen atoms were placed in geometrically idealized positions and included as standard riding atoms during the least-squares refinements.

**Absorption and emission spectroscopies.** CH<sub>2</sub>Cl<sub>2</sub> solutions of **3** (2.5 mg mL<sup>-1</sup>, 400  $\mu$ L) and either **1** (83  $\mu$ g mL<sup>-1</sup>, 100  $\mu$ L) or Ph<sub>3</sub>P (104  $\mu$ g mL<sup>-1</sup>, 100  $\mu$ L) were mixed and the solvent was distilled off under reduced pressure. The residue was dispersed in PBS (1.0 mL) and the resulting dispersions were sonicated for 5 min, stored for 10 min at ambient temperature, passed through a syringe filter (200 nm) and used for the spectroscopic measurements without further purification. Absorption spectra were recorded with a Varian Cary 100 Bio spectrometer in quartz cells with a path length of 1.0 cm. Emission spectra were recorded with a Varian Cary Eclipse spectrometer in aerated solutions. The fluorescence quantum yield of **2** was determined against a fluorescein standard, following a literature protocol.<sup>S5</sup>

**Fluorescence imaging.** *Drosophila melanogaster* S2 cells were cultured in Shields and Sang M3 Insect Medium (39.4 gL<sup>-1</sup>) with fetal bovine serum (15%, v/v), penicillin-streptomycin solution (1% v/v at a final concentration of 100 k units L<sup>-1</sup> penicillin and 100 mg L<sup>-1</sup> streptomycin) and KHCO<sub>3</sub> (0.5 g L<sup>-1</sup>) and incubated at 22 °C. The cells were seeded in glass-bottom plates at a density of 5 × 10<sup>4</sup> cells mL<sup>-1</sup> and incubated for 30 min at 22 °C.

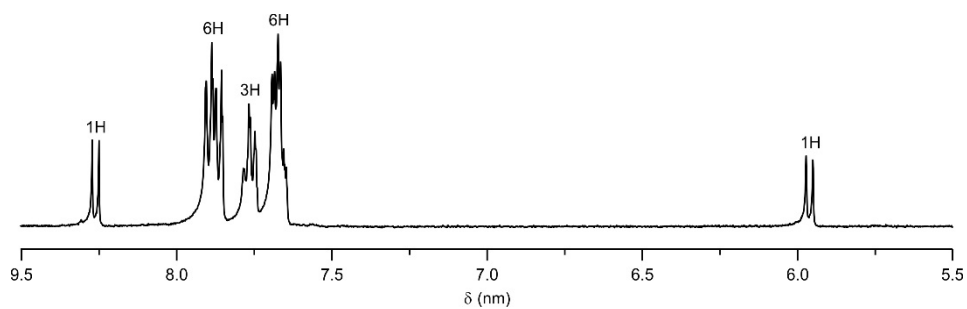
CH<sub>2</sub>Cl<sub>2</sub> solutions of **3** (16 mg mL<sup>-1</sup>, 1.0 mL) and either **1** (2 mg mL<sup>-1</sup>, 60  $\mu$ L) or Ph<sub>3</sub>P (2 mg mL<sup>-1</sup>, 80  $\mu$ L) were mixed and the solvent was distilled off under reduced pressure. The residue was dispersed in PBS (1.0 mL) and the resulting dispersions were sonicated for 5 min and incubated with the cultured cells without further purification.

The cultured cells were incubated with a PBS solution (5%, v/v) of **1** (120  $\mu$ g mL<sup>-1</sup>) and **3** (16 mg mL<sup>-1</sup>) for 30 min washed twice with PBS (100  $\mu$ L), imaged, incubated further with Ph<sub>3</sub>P (160  $\mu$ g mL<sup>-1</sup>) and **3** (16 mg mL<sup>-1</sup>) for

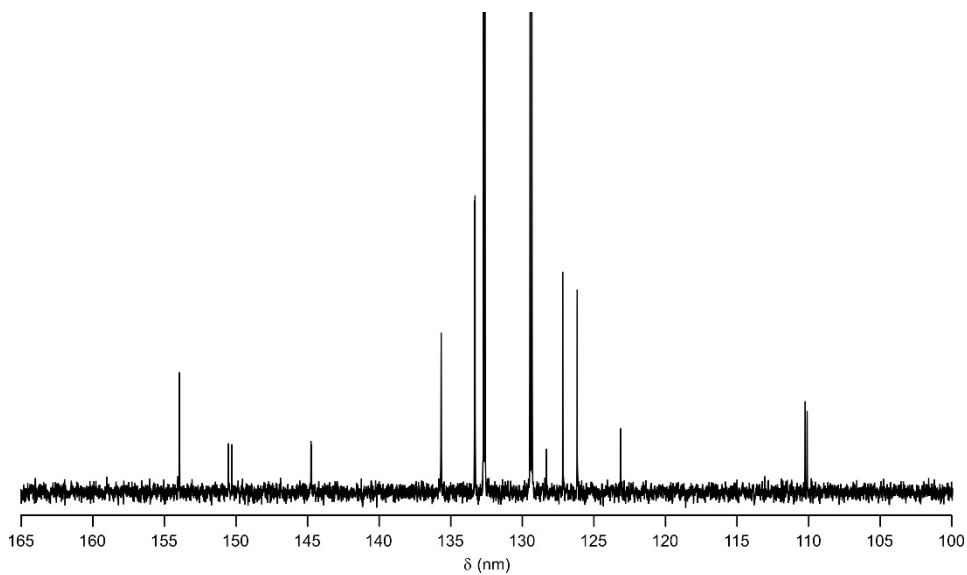
another 30 min, washed twice with PBS (100  $\mu\text{L}$ ) and imaged again. The same protocol was repeated inverting the order of the two incubation steps. Alternatively, the cultured cells were incubated with a PBS solution (5%, v/v) of **1** (120  $\mu\text{g mL}^{-1}$ ) for 30 min, washed twice with PBS (100  $\mu\text{L}$ ), incubated further with a PBS dispersion (5%, v/v) of Ph<sub>3</sub>P (160  $\mu\text{g mL}^{-1}$ ) and **3** (16  $\text{mg mL}^{-1}$ ) for another 30 min and imaged. Once again, the same protocol was repeated inverting the order of the two incubation steps. All images were recorded with a Leica SP5 confocal laser-scanning microscope.

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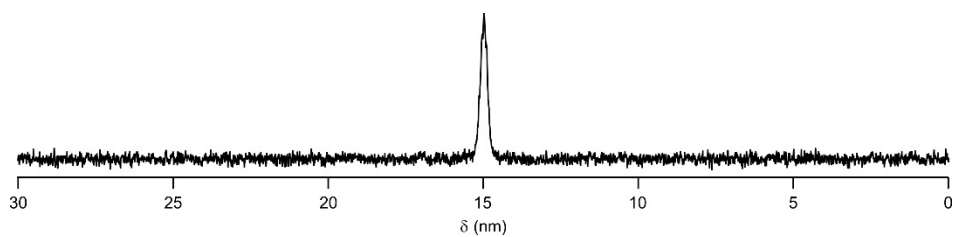
- S1 S. J. Lord, H. D. Lee, R. Samuel, R. Weber, N. Liu, N. R. Conley, M. A. Thompson, R. J. Twieg, W. E. Moerner, *J. Phys. Chem. B*, 2010, **114**, 14157.
- S2 I. Yildiz, S. Impellizzeri, E. Deniz, B. McCaughan, J. F. Callan, F. M. Raymo, *J. Am. Chem. Soc.*, 2011, **133**, 871.
- S3 Apex2 Version 2.2-0 and SAINT+ Version 7.46A; Bruker Analytical X-Ray System, Inc., Madison, Wisconsin, USA, 2007.
- S4 (a) G. M. Sheldrick, SHELXTL Version 6.1; Bruker Analytical X-Ray Systems, Inc., Madison, Wisconsin, USA, 2000. (b) G. M. Sheldrick, *Acta Cryst.*, 2008, **A64**, 112.
- S5 J. R. Lakowicz, *Principles of Fluorescence Spectroscopy*; Springer: New York, 2006.



**Fig. S1.**  $^1\text{H}$  NMR spectrum (400 MHz) of **2** in  $(\text{CD}_3)_2\text{SO}$  at 25 °C.



**Fig. S2.**  $^{13}\text{C}$  NMR spectrum (400 MHz) of **2** in  $\text{CDCl}_3$  at 25 °C.

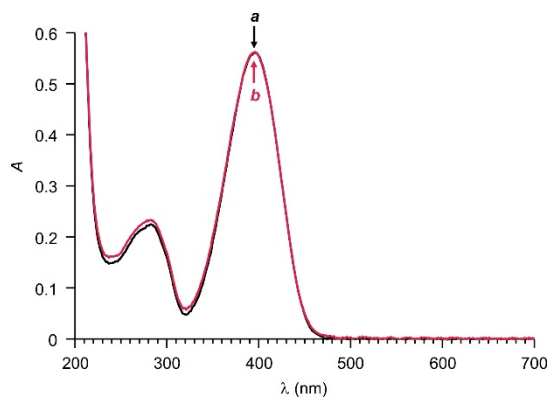


**Fig. S3.**  $^{31}\text{P}$  NMR spectrum (400 MHz) of **2** in  $(\text{CD}_3)_2\text{SO}$  at 25 °C.

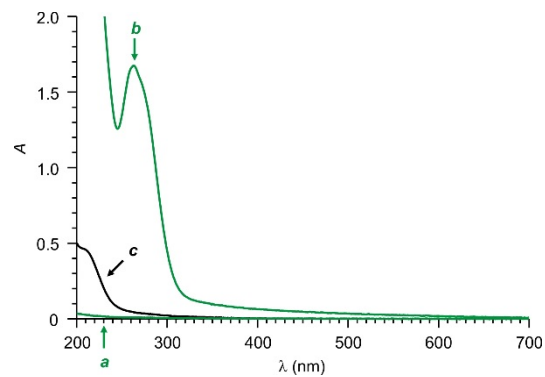
**Table S1.** Crystallographic Data for **2**.

<i>Empirical Formula</i>	C <sub>24</sub> H <sub>17</sub> N <sub>4</sub> O <sub>3</sub> P
<i>Formula Weight</i>	440.39
<i>Crystal System</i>	Monoclinic
<i>Lattice Parameters:</i>	
<i>a</i> (Å)	11.9130(5)
<i>b</i> (Å)	8.4877(3)
<i>c</i> (Å)	21.1543(9)
<i>β</i> (°)	104.391(1)
<i>V</i> (Å <sup>3</sup> )	2071.88(14)
<i>Space Group</i>	<i>P</i> 2 <sub>1</sub> / <i>n</i> (# 14)
<i>Z</i> Value	4
<i>ρ</i> <sub>calc</sub> (g cm <sup>-3</sup> )	1.412
<i>μ</i> (Mo Kα) (mm <sup>-1</sup> )	0.168
<i>T</i> (K)	296
2 $\Theta$ <sub>max</sub> (°)	60.00
<i>No. Obs.</i> ( <i>I</i> > 2σ( <i>I</i> ))	4975
<i>No. Parameters</i>	289
<i>Goodness of Fit</i>	1.035
<i>Max. Shift in Cycle</i>	0.001
<i>Residuals</i> *: R1; wR2	0.0391; 0.1078
<i>Absorption Correction</i> ,	Multi-Scan
Max/min	0.9834/0.9388
<i>Largest Peak in Final Diff. Map</i> (e <sup>-</sup> Å <sup>-3</sup> )	0.300

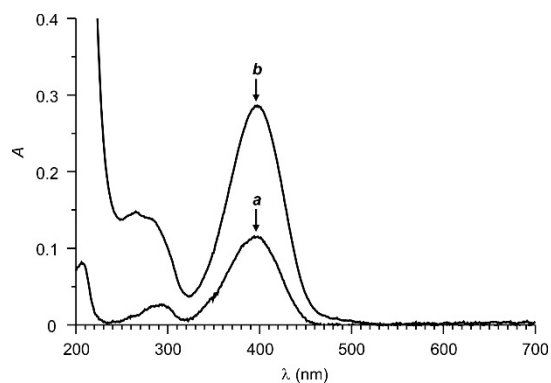
\*  $R = \sum_{hkl} (|F_{obs}| - |F_{calc}|) / \sum_{hkl} |F_{obs}|$ ;  $R_w = [\sum_{hkl} w(|F_{obs}| - |F_{calc}|)^2 / \sum_{hkl} w F_{obs}^2]^{1/2}$ ,  
 $w = 1/\sigma^2(F_{obs})$ ;  $GOF = [\sum_{hkl} w(|F_{obs}| - |F_{calc}|)^2 / (n_{data} - n_{vari})]^{1/2}$ .



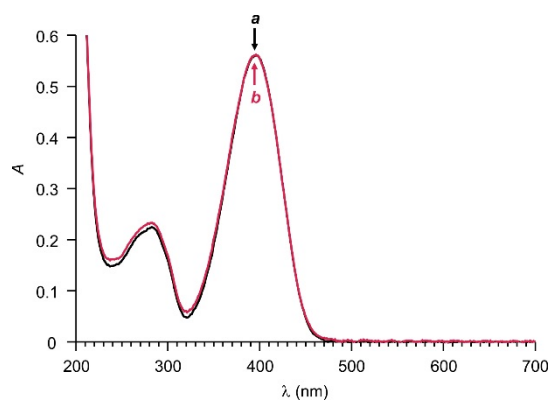
**Fig. S4.** Absorption spectra of a MeCN solution of **1** ( $40 \mu\text{M}$ ) recorded before (*a*) and after (*b*) storage in the dark for 24 hours at  $25 \text{ }^\circ\text{C}$ .



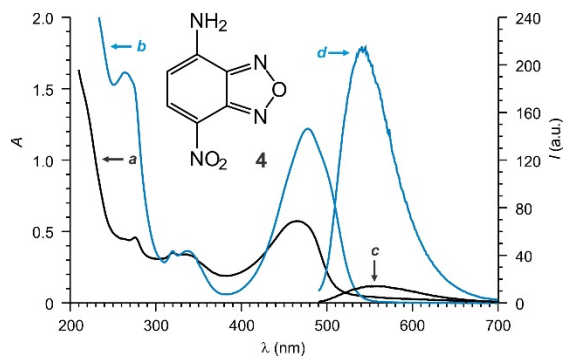
**Fig. S5.** Absorption spectra recorded at  $25 \text{ }^\circ\text{C}$  after treating  $\text{PPh}_3$  ( $10 \mu\text{g}$ ) with PBS ( $1.0 \text{ mL}$ ) in the absence (*a*) or presence (*b*) of **3** ( $1.0 \text{ mg mL}^{-1}$ ). Absorption spectrum (*c*) of **3** ( $1.0 \text{ mg mL}^{-1}$ ) in PBS at  $25 \text{ }^\circ\text{C}$ .



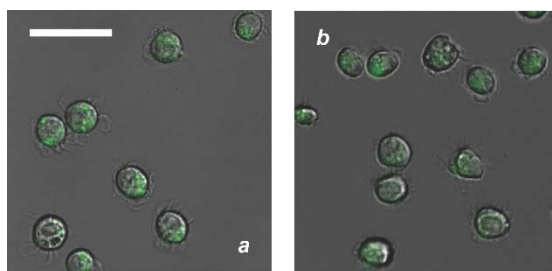
**Fig. S6.** Absorption spectra recorded at  $25 \text{ }^\circ\text{C}$  after treating **1** ( $8 \mu\text{g}$ ) with PBS ( $1.0 \text{ mL}$ ) in the absence (*a*) or presence (*b*) of **3** ( $1.0 \text{ mg mL}^{-1}$ ).



**Fig. S7.** Absorption spectra of a PBS solution of **1** ( $40 \mu\text{g mL}^{-1}$ ) and **3** ( $1.0 \text{ mg mL}^{-1}$ ) recorded before (*a*) and after (*b*) storage in the dark for 3 hours at  $25 \text{ }^\circ\text{C}$ .



**Fig. S8.** Absorption and emission ( $\lambda_{\text{Ex}} = 480 \text{ nm}$ ) spectra of PBS solutions ( $25 \text{ }^\circ\text{C}$ ) of **3** ( $1.0 \text{ mg mL}^{-1}$ ) and either **2** ( $8 \text{ } \mu\text{g mL}^{-1}$ , *a* and *c*) or **4** ( $8 \text{ } \mu\text{g mL}^{-1}$ , *b* and *d*).



**Fig. S9.** Overlaid fluorescence and transmittance images ( $\lambda_{\text{Ex}} = 488 \text{ nm}$ ,  $\lambda_{\text{Em}} = 510\text{--}700 \text{ nm}$ , scale bar =  $25 \text{ } \mu\text{m}$ ) of S2 cells recorded after incubation with either a PBS solution (5%, v/v) of **1** ( $120 \text{ } \mu\text{g mL}^{-1}$ ) for 30 min, washing, further incubation with a PBS solution (5%, v/v) of  $\text{Ph}_3\text{P}$  ( $160 \text{ } \mu\text{g mL}^{-1}$ ) and **3** ( $16 \text{ mg mL}^{-1}$ ) for 30 min and washing (*a*) or a PBS solution (5%, v/v) of **3** ( $16 \text{ mg mL}^{-1}$ ) and  $\text{Ph}_3\text{P}$  ( $160 \text{ } \mu\text{g mL}^{-1}$ ) for 30 min, washing, further incubation with a PBS solution (5%, v/v) of **1** ( $120 \text{ } \mu\text{g mL}^{-1}$ ) for 30 min and washing (*b*).