Electronic Supplementary Material

A turn–on fluorescent sensor for relay recognition of two ions: From F––selective sensor to highly Zn2+–selective sensor by tuning electronic effects

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1. General Methods

Fresh double distilled water was used throughout the experiment. All other reagents and solvents were commercially available at analytical grade and were used without further purification. ¹H–NMR and ¹³C–NMR spectra were recorded on an Agilent DD2 at 600 MHz spectra. ¹H chemical shifts are reported in ppm downfield from tetramethylsilane (TMS, δ scale) with the solvent resonances as internal standards. UV–visible spectra were recorded on a Shimadzu UV–2550 spectrometer. Photoluminescence spectra were performed on a Shimadzu RF–5301 fluorescence spectrophotometer. Melting points were measured on an X–4 digital melting-point apparatus. The infrared spectra were performed on a Digilab FTS–3000 FT–IR spectrophotometer.

All the UV–vis experiments were carried out in DMSO on a Shimadzu UV–2550 spectrometer. Any changes in the UV–vis spectra of the synthesized compound were recorded on addition of perchlorate salts while keeping the ligand concentration constant (2.0×10⁻⁵ M) in all experiments. perchlorate salt (4.0 × 10⁻⁴ M) of anions $(Fe^{3+}, Hg^{2+}, Ag^+, Ca^{2+}, Cu^{2+}, Co^{2+}, Ni^{2+}, Cd^{2+}, Pb^{2+}, Zn^{2+}, Cr^{3+}, and Mg^{2+})$ and Tetrabutylammonium salt $(1.0 \times 10^{-3} \text{ M})$ of anions (F⁻, Cl⁻, Br⁻, I⁻, AcO⁻, H₂PO₄⁻, $HSO₄$ ⁻ and ClO₄⁻) and sodium salt (1.0 \times 10⁻³ M) of anions (CN⁻) were used for the UV–vis experiments.

All the fluorescence spectroscopy was carried out in DMSO on a Shimadzu RF– 5301 spectrometer. Any changes in the fluorescence spectra of the synthesized compound were recorded on addition of perchlorate salts while keeping the ligand concentration constant (2.0×10⁻⁵ M) in all experiments. perchlorate salt (4.0 × 10⁻⁴ M) of anions (Fe³⁺, Hg²⁺,Ag⁺, Ca²⁺,Cu²⁺, Co²⁺, Ni²⁺, Cd²⁺, Pb²⁺, Zn²⁺, Cr³⁺, and Mg²⁺) and Tetrabutylammonium salt $(1.0 \times 10^{-3} \text{ M})$ of anions (F⁻, Cl⁻, Br⁻, I⁻, AcO⁻, H₂PO₄⁻, $HSO₄$ and $ClO₄$) and sodium salt (1.0 \times 10⁻³ M) of anions (CN⁻) were used for the fluorescence experiments

For ¹H–NMR titrations, the solution of **L2** was prepared in DMSO– d_6 and the appropriate concentrated solution of guest was prepared in DMSO– d_6 . Aliquots of the two solutions were mixed directly in NMR tubes.

2. Synthesis of sensor molecule L2

Compound **L2** can be readily prepared by a simple and low–cost amide reaction of naphtha [2, 1-*b*] furan-2-carbonyl chloride and 8-aminoquinoline (Scheme S1). naphtha [2, 1-b] furan-2-carbonyl chloride (0.462 g, 2 mmol), 8-aminoquinoline $(0.360 \text{ g}, 2.5 \text{ mmol})$ and 2.5mml triethylamine (Et_3N) were combined in hot absolute tetrahydrofuran (30 mL)*.* The solution was stirred under reflux for 6 hours. After cooling to room temperature, the yellow precipitate was filtered, washed three times with hot absolute tetrahydrofuran, then recrystallized with THF to give a yellow powder product **L2** (1.56 mmol) in 78% (m. p. >300 °C), IR: (KBr, cm-1) *v*: 3338 (– NH–), 3118 (C=CH), 3046 (ArH), 1681 (C=O), 1596 (C=C), 1564 (C=N), 1531 (C=C), 1487 (C–N). ¹H–NMR (DMSO–*d*6, 600 MHz): *δ* 10.94 (1H, s, NH), 9.08 (1H, s, C=CH), 8.94 (1H, d, ArH), 8.57–8.52 (3H, m, ArH), 8.14–8.04 (3H, m, ArH), 7.82–7.62(5H, m, ArH); ¹³C–NMR (DMSO–*d*6, 150 MHz): *δ* 155.85, 152.48, 149.32, 147.84, 137.62, 137.76, 133.40, 130.11, 128.88, 128.77, 127.81, 127.31, 127.03, 125.53, 123.88, 122.55, 122.46, 116.35, 112.58, 111.10, 109.67, 109.22; Anal. calcd for $C_{22}H_{14}N_2O_2$: C, 78.11, H, 4.14, N, 8.28, O, 9.47; found C, 78.07; H, 4.11; N, 8.33; O, 9.49.

Scheme S1 Synthesis of the sensor compound **L2**.

3. Absorbance spectra of L2 in DMSO in the presence of fluoride anions

Fig. S1: Absorbance spectra of **L2** in DMSO in the presence of fluoride anions (20 equiv.). Inset: photograph showing the change in color of the solution of **L2** in DMSO after addition of fluoride

anions (20 equiv.).

4. Plot of fluorescence intensity depending on the concentration of zinc ions

Fig. S2: A plot of fluorescence intensity depending on the concentration of zinc ions in the range from 0 to 5 equivalents in **L2–F.**

5. Determination of Detection Limit

Linear Equation: $Y = 28.0669 \times X + 299.9236$ R = 0.979

$$
S = 2.8067 \times 10^{7} \qquad \delta = \sqrt{\frac{\Sigma (F - F)2}{(N - 1)}} = 1.9647 \text{ (N = 15)} \qquad \text{K}
$$

 $= 3$

$$
LOD = K \times \delta/S = 2.1 \times 10^{-7} M
$$

Fig. S3: The photograph of the fluorescent spectrum linear range.

6. The UV–vis spectroscopy and fluorescence spectroscopy for the stability and reproducibility of the sensor L2

Fig. S4: The UV–vis spectroscopy and fluorescence spectroscopy for the stability and reproducibility of the sensor **L2**.

7. IR spectra of sensor L2 and after adding fluoride anions

Fig. S5: IR spectra of sensor **L2** and after adding fluoride anions in KBr disks.

8. IR spectra of L2–F and after adding zinc ions

Fig. S6: IR spectra of **L2–F** and after adding zinc ions in KBr disks.

9. The Job's plot examined between zinc ions and L2–F

Fig. S7: The Job's plot examined between zinc ions and **L2**–**F**, indicating the 1:1 stoichiometry for **L2**–**F** and zinc ion.

10. ESI/MS of L2

Fig. S8: The ESI/MS of L2 in DMSO.

11. ESI/MS of L2-F with zinc

Fig. S9: The ESI/MS of **L2-F** with zinc in DMSO.

12. ¹H-NMR spectrum of L2

Fig. S10: ¹H-NMR spectrum of L2 in DMSO- d_6 .

13. ¹³C-NMR spectrum of L2

Fig. S11: ¹³C-NMR spectrum of L2 in DMSO- d_6 .