Diols-responsive triple-component supra-amphiphile constructed from pillar[5]arene-based recognition

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1. Materials and methods

All reagents were commercially available and used as received. Compound 1^{S1} and water-soluble pillar[5]arenes^{S2} (WSP5) were synthesized according to published literature procedures. Solvents were employed as purchased or dried according to procedures described in the published literature.

NMR spectra were obtained on a Bruker Avance III-400 spectrometry or a Bruker AVANCE DMX-500 spectrometer with the internal standard TMS. Isothermal titration calorimetric (ITC) experiments were performed on a VP-ITC microcalorimeter (Microcal, USA). Low-resolution electrosprayionization mass spectra were recorded with a Bruker Esquire 3000 Plus spectrometer. High-resolution mass spectrometry experiments were performed with a BrukerDaltonics Apex III spectrometer. The melting points were collected on a SHPSIC WRS-2 automatic melting point apparatus. The critical aggregation concentration (CAC) values of **A**, **G1@A** and **WSP5⊃G1@A** were determined on a DDS-307 instrument. Transmission electron microscopy (TEM) investigation was carried out on a JEM-1200EX instrument. The fluorescence experiments were conducted on a RF-5301 spectrofluorophotometer (ShimadzuCorporation, Japan). Dynamic light scattering measurements were performed on a goniometer ALV/CGS-3 instrument.

For TEM sample preparation, a few drops of the solution were placed on the carbon coated copper grid. Then the solvent was removed by vacuum drying under low temperature to afford the TEM samples.For AFM and SEM sample preparation, a few drops of the solution were placed on the silicon wafer. Then the solvent was removed by vacuum drying under low temperature to afford the TEM samples.

2. Synthesis of compound 2



Scheme S1. Synthesis of compound 2.

Trimethylamine (33 % in ethanol, 5 mL) and compound **1** (438 mg, 1.00 mmol) were added into ethanol (50 mL), and the solution was refluxed for 12 h. The solvents and excess trimethylamine were evaporated under vacuum, and the residues were dissolved in methanol. Then the solution was acidified with aqueous HCl solution. The resulting precipitate was filtered, washed with water, and dried to afford product **2** as a white solid, mp: 233.7–234.5 °C. The ¹H NMR spectrum of **2** is shown in Fig. S1. ¹H NMR (400 MHz, CD₃OD, 298 K) δ (ppm): 7.61 (d, *J* = 8 Hz, 2H), 6.83 (d, *J* = 8 Hz, 2H), 3.93 (d, *J* = 8 Hz, 2H), 3.30–3.26 (m, 4H), 3.08 (s, 9H), 1.77–1.69 (m, 4H), 1.46–1.33 (m, 12H). The ¹³C NMR spectrum of **2** is shown in Fig. S2. ¹³C NMR (100 MHz, CD₃OD, 298 K) δ (ppm): 162.32, 136.63, 114.68, 68.70, 67.86, 53.54, 30.53, 30.43, 30.40, 30.38, 30.18, 27.35, 27.14, 25.21, 23.95. LRESIMS is shown in Fig. S3: *m/z* 336.5 [M – Br]⁺. *m/z* calcd for [M – Br]⁺ C₁₉H₃₅BNO₃⁺, 336.2705, found 336.2719, error 4 ppm.



Fig. S1. ¹H NMR spectrum (400MHz, CD₃OD, 293K) of **2**.





Fig. S2. ¹³C NMR spectrum (100MHz, CD₃OD, 293K) of **2**.

Fig. S3. Electrospray ionization mass spectrum of compound 2. Assignment of the main peak:

m/z 336.5 [M – Br]⁺.

3. Host-guest complexion between WSP5 and G2 in D_2O



Fig. S4. ¹H NMR spectrum (400 MHz, D₂O, room temperature) of solution of G2 and WSP5: (a) G2 (2.00 mM), (b) the 1:1 mixture of G2 and WSP5 (2.00 mM) and (c) WSP5 (2.00 mM).



Fig. S5. Partial NOESY NMR spectrum (500 MHz, D₂O, 298 K) of **WSP5** (10.0 mM) and **G2** (10.0 mM).



Fig. S6. Microcalorimetric titration of G2 with WSP5 in water at 298.15 K. (Top) Raw ITC data for 29 sequential injections (10 μ L per injection) of a G2 solution (2.00 mM) into a WSP5 solution (0.100 mM). (Bottom) Net reaction heat obtained from the integration of the calorimetric traces.

4 Critical aggregation comcentration (CAC) determination of A, G1@A and



Fig. S7. The concentration-dependent conductivity of **A** in aqueous solutions. The critical aggregation concentration (CAC) was determined to be 5.49×10^{-7} M.



Fig. S8. The concentration-dependent conductivity of G1@A in aqueous solutions. The critical aggregation concentration value was determined to be 7.95×10^{-6} M.



Fig. S9. The concentration-dependent conductivity of WSP5 \supset G1@A in aqueous solutions. The critical aggregation concentration value was determined to be 4.74×10^{-5} M.

5 Dynamic light scattering (DLS) result of WSP5 –G1@A



Fig. S10. DLS result of WSP5 \supset G1@A with an aqueous solution of 1.00×10^{-4} M WSP5 and 1.00×10^{-4} M G1@A.

6 Fluorscence spectra of a solution of $WSP5 \supset G1@A$ upon the titration with different

diols in aqueous solution



Fig. S11. The changes in fluorescence intensity of WSP5 \supset G1@A (1.00 × 10⁻⁴ M) upon the titration of triethanolamine (0.00, 0.10, 0.30, 0.60, 1.00, 1.50, 2.00, 2.50, 3.00, 4.00, 5.00, 6.00, 7.00 equiv.) in aqueous solution ($\lambda_{ex} = 370$ nm).



Fig. S12. The changes in fluorescence intensity of WSP5 \supset G1@A (1.00 × 10⁻⁴ M) upon the titration of diethanolamine (0.00, 0.10, 0.30, 0.60, 1.00, 1.50, 2.00, 2.50, 3.00, 4.00, 5.00, 6.00, 7.00 equiv.) in aqueous solution ($\lambda_{ex} = 370$ nm).



Fig. S13. The changes in fluorescence intensity of WSP5 \supset G1@A (1.00 × 10⁻⁴ M) upon the titration of ethylene glycol (0.00, 0.10, 0.30, 0.60, 1.00, 1.50, 2.00, 2.50, 3.00, 4.00, 5.00, 6.00, 7.00 equiv.) in aqueous solution ($\lambda_{ex} = 370$ nm).



Fig. S14. The changes in fluorescence intensity of WSP5 \supset G1@A (1.00 × 10⁻⁴ M) upon the titration of 2,2-dimethypropylene glycol (0.00, 0.10, 0.30, 0.60, 1.00, 1.50, 2.00, 2.50, 3.00, 4.00, 5.00, 6.00, 7.00 equiv.) in aqueous solution ($\lambda_{ex} = 370$ nm).



Fig. S15. The changes in fluorescence intensity of WSP5 \supset G1@A (1.00 × 10⁻⁴ M) upon the titration of propylene glycol (0.00, 0.10, 0.30, 0.60, 1.00, 1.50, 2.00, 2.50, 3.00, 4.00, 5.00, 6.00, 7.00 equiv.) in aqueous solution ($\lambda_{ex} = 370$ nm).

7. The morphology changes of G1@A upon gradual addition of WSP5



Fig. S16. The morphology changes of 1.00×10^{-4} M G1@A upon gradual addition of 1 equiv. of WSP5. Accompanied with gradual addition of WSP5, the nanosheets of G1@A first

transformed into an intermediate state nanoribbon structures and finally turned into nanoparticles.

8. The aggregate morphology changes of WSP5 \supset G1@A upon gradual addition of diols A1



Fig. S17. The self-assembly morphology of (a) 1.00×10^{-4} M WSP5 \supset G1@A; (b) 1.00×10^{-4} M WSP5 \supset G1@A with 1.0 equiv. of A1; (c) 1.00×10^{-4} M WSP5 \supset G1@A with 2.0 equiv. of A1; (d) 1.00×10^{-4} M WSP5 \supset G1@A with 3.0 equiv. of A1.

9. Scanning electron microscope (SEM) image of A



- Fig. S18. SEM images of A aggregates.
- 10. Atomic force microscope (AFM) image of G1@A



Fig. S19. AFM image of G1@A aggregates.

10 Cartoon representation of the formation of the self-assembly structure from G1@A

and WSP5



Fig. S20. Cartoon representation of the formation of the self-assembly structure from G1@A and WSP5.

11. References:

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