

**Supplementary data for**  
**Effects of Fe (II) on microbial communities and nitrogen**  
**transformation pathway of nitrogen and iron cycling in the**  
**Anammox process: kinetic, quantitative mechanism and**  
**metagenomic analysis**

**Submitted to *RSC Advance***

**Authors:**

Duntao Shu<sup>a, b §</sup>, Yanling He<sup>c\*</sup>, Hong Yue<sup>d §</sup>, Shucheng Yang<sup>e</sup>

**Affiliations of authors:**

<sup>a</sup>Center for Mitochondrial Biology and Medicine, The Key Laboratory of Biomedical Information Engineering of the Ministry of Education, School of Life Science and Technology, Xi'an Jiaotong University, Shaanxi 710049, China

<sup>b</sup>State Key Laboratory of Crop Stress Biology in Arid Areas, College of Life Sciences, Northwest A&F University, Yangling, Shaanxi 712100, China

<sup>c</sup>School of Human Settlements & Civil Engineering, Xi'an Jiaotong University, Shaanxi 710049, China

<sup>d</sup>State Key Laboratory of Crop Stress Biology in Arid Areas, College of Agronomy and Yangling Branch of China Wheat Improvement Center, Northwest A&F University, Yangling, Shaanxi 712100, China

<sup>e</sup> School of Energy and Power Engineering, Xi'an Jiaotong University, Shaanxi 710049,

---

\* Corresponding author. Email: hey1@mail.xjtu.edu.cn; Tel/Fax: 0086 029 83395128.

§ These authors contributed equally to this work.

China

## **Contents**

### **1. Tables**

**Table S1** Primers used for qPCR thermal programs in this study.

**Table S2** Raw and effective reads, plus numbers of OTUs, Good's coverage, Shannon, Chao1, ACE, and Simpson of five phases.

### **2. Figures**

**Figure S1** Rarefaction curves base on MiSeq pyrosequencing of bacterial communities in different phases. The OTUs were defined by 3% distances.

**Figure S2** Distributions of bacteria in five phases at different taxonomy level. (a) At class level; (b) at order level; (c) at family level; (d) at genus level. Taxa represented occurred at >1% frequency in at least one sample. Others refer to the taxa with their maximum abundance <1% in any sample.

### **3. References**

**Table S1** Primers used for qPCR and thermal programs in this study.

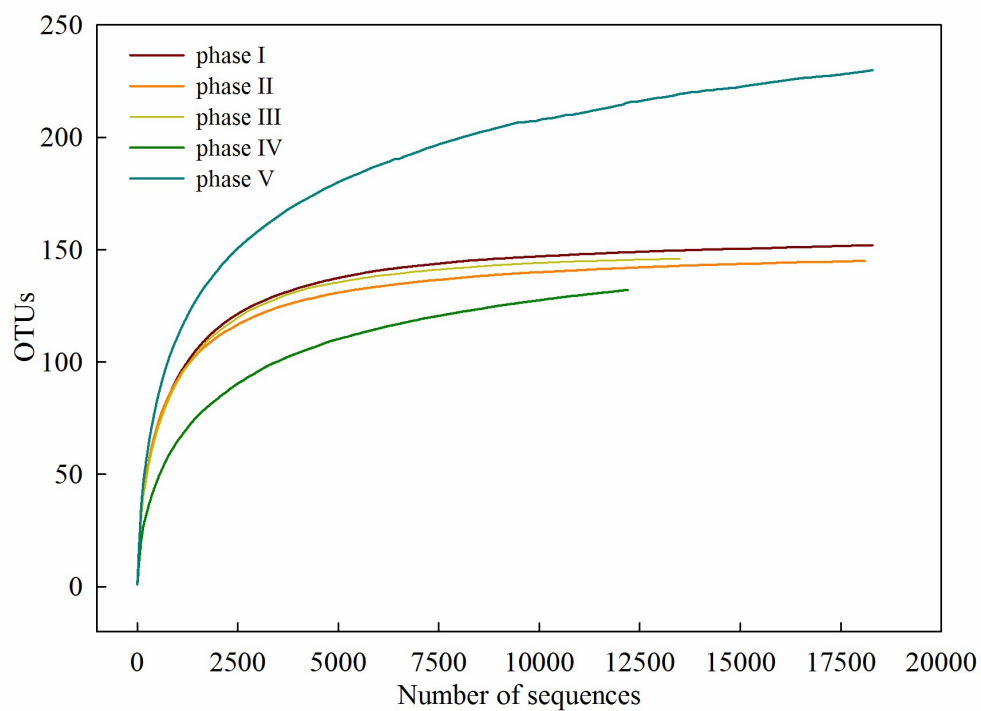
| Target prokaryote                       | Target gene | Sequence (5'-3') of primer pairs                                  | Annealing (°C) | Thermal program  | Reference    |
|---|-------------|---|----------------|--|--------------|
| Total bacteria                          | 16S rRNA    | 341F: CCTACGGGAGGCAGCAG<br>518R: ATTACCGGGCTGCTGG                 | 60             | 5 min at 95°C, 40 cycles of 30 s at 95°C, 30 s at 60°C, and 40 s at 72°C               | <sup>1</sup> |
| Anammox Bacteria                        | 16S rRNA    | Amx809f: GCCGTAAACGATGGGCACT<br>Amx1066r: AACGTCTCACGACACGAGCTG   | 60             | 10min at 95°C, followed by 35 cycles of 60 s at 95°C, 60 s at 60°C, and 45 s at 72°C   | <sup>2</sup> |
| AOA                                     | <i>amoA</i> | amoAF: STAATGGTCTGGCTTAGACG<br>amoAR: GCGGCCATCCATCTGTATGT        | 53             | 3 min at 94°C, followed by 40 cycles of 30 s at 94°C, 1 min at 53°C, and 1 min at 72°C | <sup>3</sup> |
| AOB                                     | <i>amoA</i> | amoAF: GGGGTTTCTACTGGTGGT<br>amoAR: CCCCTCKGSAAAGCCTTCTTC         | 55             | 3 min at 94°C, followed by 40 cycles of 30 s at 94°C, 30 s at 55°C, and 45 s at 72°C   | <sup>4</sup> |
| Denitrifying bacteria                   | <i>nosZ</i> | nosZ1F: WCSYTGTTTCMTCGACAGCCAG<br>nosZ1R: ATGTCGATCARCTGVKCRTTYTC | 63             | 5 min at 95°C, followed by 35 cycles of 60 s at 95°C, 60 s at 63°C, and 45 s at 72°C   | <sup>5</sup> |
| Denitrifying bacteria                   | <i>nirS</i> | nirSnF: TACCACCCGAGCCGCGCGT<br>nirSnr: GCCGCCGTCRTGVAGGAA         | 63             | 5 min at 95°C, followed by 35 cycles of 60 s at 95°C, 60 s at 63°C, and 45 s at 72°C   | <sup>6</sup> |
| Denitrifying bacteria                   | <i>nirK</i> | nirKF: ATYGGCGGVAYGGCGA<br>nirKR: GCCTCGATCAGRTRRTGG              | 57             | 5 min at 95°C, followed by 35 cycles of 60 s at 95°C, 60 s at 57°C, and 45 s at 72°C   | <sup>6</sup> |
| Dissimilarity nitrite reducing bacteria | <i>narG</i> | narG2F: CTCGAYCTGGTGGTYGA<br>narG2R: TTYTCGTACCAGGTS GC           | 55             | 5 min at 95°C, followed by 35 cycles of 60 s at 95°C, 60 s at 55°C, and 45 s at 72°C   | <sup>7</sup> |
| Dissimilarity nitrite reducing bacteria | <i>napA</i> | napA3F: CCCAATGCTCGCCACTG<br>napA3R: CATGTTKGAGCCCCACAG           | 60             | 5 min at 95°C, followed by 35 cycles of 60 s at 95°C, 60 s at 60°C, and 45 s at 72°C   | <sup>7</sup> |
| Dissimilarity nitrate reducing bacteria | <i>nrfA</i> | nrfA2F: CACGACAGCAAGACTGCCG<br>nrfA2R: CCGGCACTTTCGAGCCC          | 60             | 5 min at 95°C, followed by 35 cycles of 60 s at 95°C, 60 s at 60°C, and 45 s at 72°C   | <sup>8</sup> |
| Nitrite oxidizing                       | <i>nxrA</i> | F1norA: CAGACCGACGTGTGCGAAAG                                      | 57             | Pre-heating at 50 °C for 2 min, pre-denaturation at 95 °C                              | <sup>9</sup> |

|                       |          |   |    |  |
|-----------------------|----------|---|----|--|
| bacteria              |          | R1norA: TCYACAAGGAACGGAAGGTC  |    | for 10 min, denaturation at 95 °C for 15 s, annealing at 57 °C for 30 s, and extension at 72 °C for 30 s   |
| <i>Acidimicrobium</i> | 16S rRNA | Amf995: CTCTGCGGCTTTTCCCTCCATG<br>Uni-907R-RC: AA ACTCAA AKAATTGACGG  | 52 | Pre-heating at 50 °C for 2 min, pre-denaturation at 95 °C for 10 min, denaturation at 95 °C for 15 s, annealing at 52 °C for 45 s, and extension at 72 °C for 45 s |
| <i>Ferrovum</i>       | 16S rRNA | Ferrovum643F: ACAGACTCTAGCTTGCCAGT<br>Uni-338F-RC: ACTCCTACGGGAGGCAGC | 57 | Pre-heating at 50 °C for 2 min, pre-denaturation at 95 °C for 10 min, denaturation at 95 °C for 15 s, annealing at 57 °C for 45 s, and extension at 72 °C for 45 s |
| <i>Albidiferax</i>    | 16S rRNA | RdoR-RC: GACCTGCATTTGTGACTGYA<br>Uni-907R: CCGTCAATTCMTTTGAGTTT       | 52 | Pre-heating at 50 °C for 2 min, pre-denaturation at 95 °C for 10 min, denaturation at 95 °C for 15 s, annealing at 52 °C for 45 s, and extension at 72 °C for 45 s |
| <i>Geobacter</i>      | 16S rRNA | Geo561F: GCGTG TAGGCGGTTTBTTAA<br>Geo858R: TCAATACCCGCAACACCTAG       | 57 | Pre-heating at 50 °C for 2 min, pre-denaturation at 95 °C for 10 min, denaturation at 95 °C for 15 s, annealing at 57 °C for 45 s, and extension at 72 °C for 45 s |
| <i>Acidiphilium</i>   | 16S rRNA | ACD840: CGACACTGAAGTGCTAAGC<br>Uni-338F-RC: ACTCCTACGGGAGGCAGC        | 61 | Pre-heating at 50 °C for 2 min, pre-denaturation at 95 °C for 10 min, denaturation at 95 °C for 15 s, annealing at 61 °C for 45 s, and extension at 72 °C for 45 s |

**Table S2** Raw and effective reads, plus numbers of OTUs, Good's coverage, Shannon, Chao1, ACE, and Simpson of five phases.

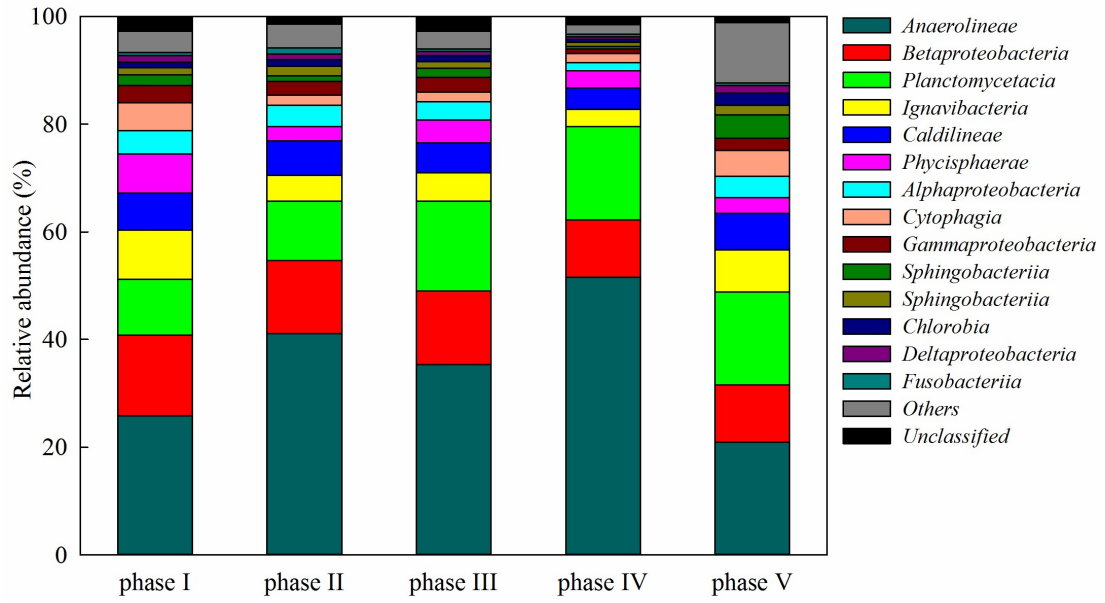
| Sample ID | Raw reads | Effective reads | OTUs | Good's coverage | Shannon | Chao 1 | ACE | Simpson |
|-----------|-----------|-----------------|------|-----------------|---------|--------|-----|---------|
| phase I   | 19527     | 18283           | 152  | 0.999562        | 3.36    | 161    | 154 | 0.0749  |
| phase II  | 19051     | 18041           | 145  | 0.999612        | 3.08    | 149    | 147 | 0.1401  |
| phase III | 15616     | 13425           | 146  | 0.999628        | 3.16    | 147    | 147 | 0.1078  |
| phase IV  | 13239     | 12125           | 132  | 0.998103        | 2.26    | 148    | 148 | 0.2605  |
| phase V   | 19925     | 17130           | 230  | 0.99784         | 3.51    | 260    | 259 | 0.0733  |

**Figure S1**

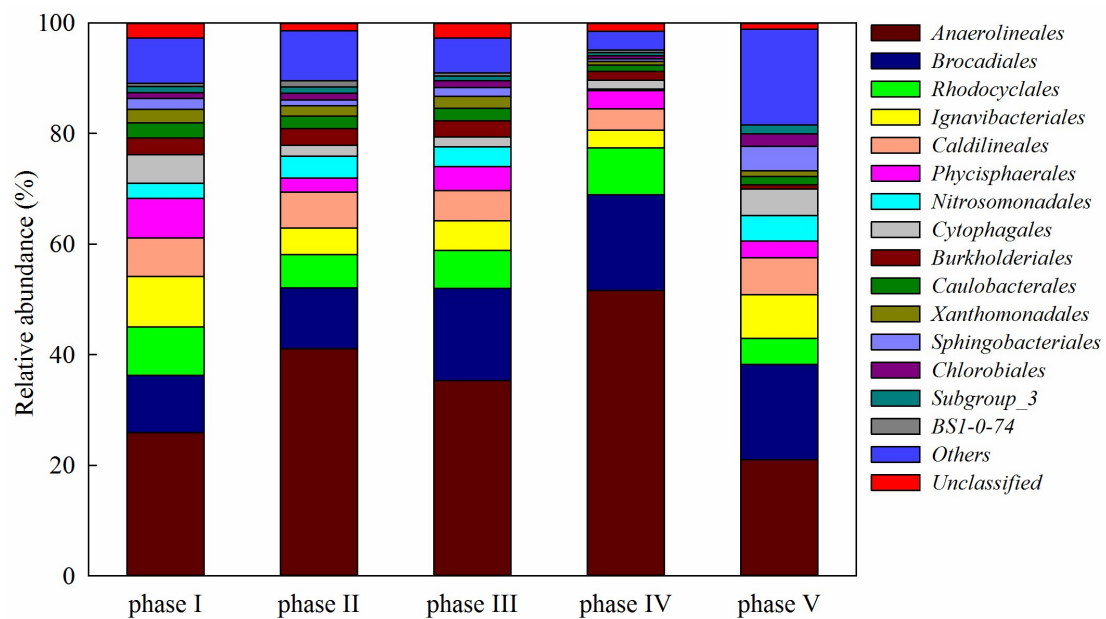


**Figure S1** Rarefaction curves base on MiSeq pyrosequencing of bacterial communities in different phases. The OTUs were defined by 3% distances.

**Figure S2a**

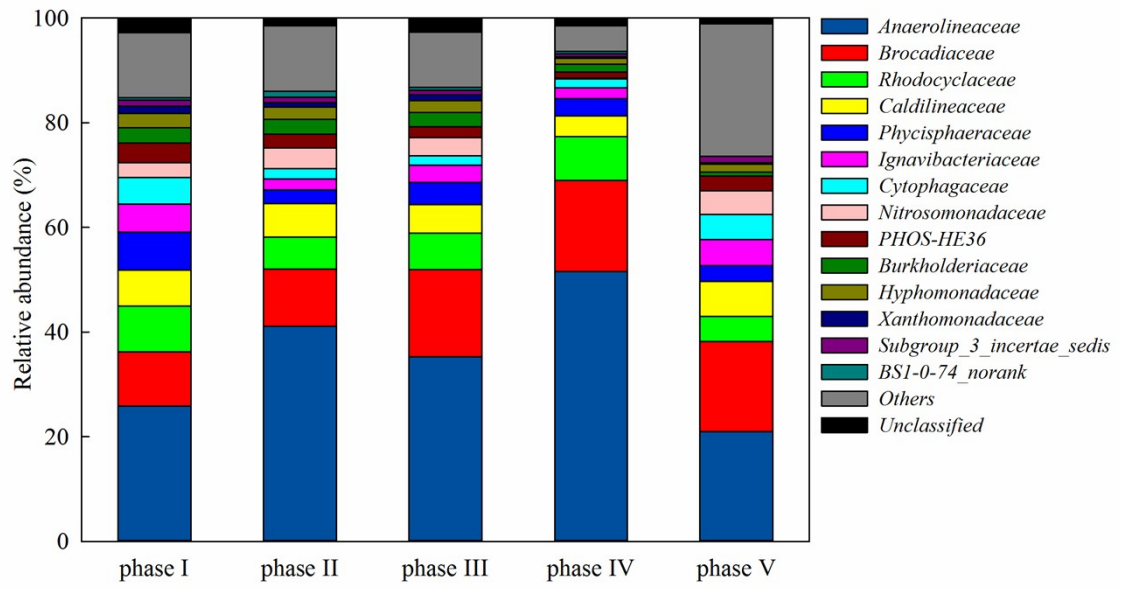


**Figure S2b**

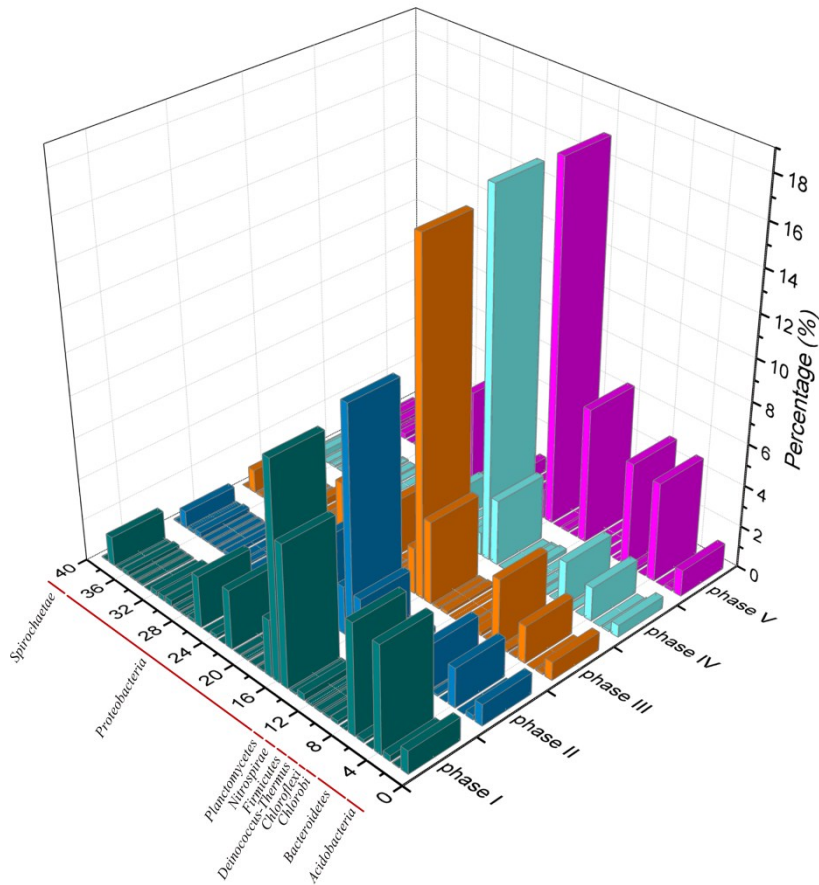




**Figure S2c**



**Figure S2d**



**Figure S2** Distributions of bacteria in five phases at different taxonomy level. (a) At class level; (b) at order level; (c) at family level; (d) at genus level. Taxa represented occurred at  $>1\%$  frequency in at least one sample. Others refer to the taxa with their maximum abundance  $<1\%$  in any sample.

## References

1. Y. Bai, Q. Sun, D. Wen and X. Tang, *FEMS microbiology ecology*, 2012, **8**, 323-330.
2. I. Tsushima, Y. Ogasawara, T. Kindaichi, H. Satoh and S. Okabe, *Water research*, 2007, **41**, 1623-1634.
3. C. A. Francis, K. J. Roberts, J. M. Beman, A. E. Santoro and B. B. Oakley, *Proceedings of the National Academy of Sciences of the United States of America*, 2005, **102**, 14683-14688.
4. J.-H. Rotthauwe, K.-P. Witzel and W. Liesack, *Applied and environmental microbiology*, 1997, **63**, 4704-4712.
5. S. Henry, D. Bru, B. Stres, S. Hallet and L. Philippot, *Applied and Environmental Microbiology*, 2006, **72**, 5181-5189.
6. E. Kandeler, K. Deiglmayr, D. Tscherko, D. Bru and L. Philippot, *Applied and Environmental Microbiology*, 2006, **72**, 5957-5962.
7. C. J. Smith, D. B. Nedwell, L. F. Dong and A. M. Osborn, *Applied and environmental microbiology*, 2007, **73**, 3612-3622.
8. P. Lam, G. Lavik, M. M. Jensen, J. van de Vossenberg, M. Schmid, D. Woebken, D. Gutiérrez, R. Amann, M. S. Jetten and M. M. Kuypers, *Proceedings of the National Academy of Sciences*, 2009, **106**, 4752-4757.
9. G. Ji, C. He and Y. Tan, *Ecological Engineering*, 2013, **55**, 35-42.
10. S. Lu, K. Chourey, M. Reiche, S. Nietzsche, M. B. Shah, T. R. Neu, R. L. Hettich and K. Küsel, *Applied and environmental microbiology*, 2013, **79**, 4272-

4281.

11. M. Fabisch, F. Beulig, D. M. Akob and K. Küsel, *Frontiers in microbiology*, 2013, 4, 1-12.