

Supplementary Information

Discovery of Lipoic acid-4-phenyl-1H-pyrazole Hybrids as Novel Bifunctional ROCK Inhibitors with Antioxidant Activity

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General information

Reagents and solvents were purchased from commercial sources and were used without further purification unless stated. The progress of the reactions was monitored by thin-layer chromatography on a glass plate coated with silica gel with fluorescent indicator (GF254) and visualized with UV light or by treatment with phosphomolybdic acid (PMA) or ninhydrin. Flash chromatography was performed on silica gel (200-300 mesh) using solvents as indicated. The ^1H and ^{13}C nuclear magnetic resonance (NMR) spectra were recorded on a Bruker Avance spectrometer 400 at 400 and 100 MHz respectively. Chemical shifts are given in ppm (δ) referenced to CDCl_3 with 7.26 for ^1H and 77.10 for ^{13}C , and to d_6 -DMSO with 2.50 for ^1H and 4.90 for ^{13}C . In the case of multiplet, the signals are reported as intervals. Signals are abbreviated as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet. Coupling constants are expressed in hertz. High-resolution mass spectrometric data (HRMS) were collected on a Shimadzu LCMS-IT-TOF mass spectrometer. The purity of all final compounds was determined by Agilent 1260 HPLC system using an Eclipse XDB-C18 column. In HPLC conditions, flow rate was set at 1 mL/min, and gradient elution was used 70%MeOH/H₂O with 0.1% TFA for 15mins.

HPLC-grade acetonitrile and methanol purchased from Merck (Darmstadt, Germany) were used for HPLC analysis. Dullbecco's modified Eagle's medium

(DMEM), and fetal bovine serum (FBS) were purchased from Gibico-BRL (Grand Island, NY, USA). 3-(3, 4- dimethylthiazol-2-yl)-2, 5-diphenyl-tetrazolium bromide (MTT), and all other chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA) unless stated.

Synthesis and characterization

General procedure A to synthesize 1-8: The synthesis of compound 6 was exemplified here.

To a solution of 2-(dimethylamino)ethanol (1.15 mL, 11.48 mmol) in THF (30 mL) at 0 °C was added NaH (60%, 636.31 mg, 15.91 mmol). After stirring for 20 min, the mixture was allowed to room temperature and stirred overnight. The solvent was removed by rotary evaporation and the residue was added addition of saturated aq. NaHCO₃ and then extracted with EtOAc (20 ml × 3) . The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated to give the crud 2-(5-bromo-2-nitrophenoxy)-N,N-dimethylethanamine (18*6) as a brown oil (2.97 g, 90.4%).

To a mixture of crude 18*6 (300.00 mg, 1.04 mmol), pyridine-4-boronic acid pinacol ester (234.05 mg, 1.14 mmol), Pd[P(Ph)₃]₄ (23.98 mg, 20.75 μmol) and K₂CO₃ (573.61 mg, 4.15 mmol) in a sealed tube, a mixed solvent (toluene/H₂O/EtOH=7/2/1, 9 ml) were then injected under Argon and the mixture was stirred at 100 °C for 8h. After cooling to room temperature, the solvent was removed

by rotary evaporation and the residue was added addition of water and extracted with EtOAc (15 ml × 3) . The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄ and filtered. The filtrate was concentrated and the residue was purified by flash chromatography on silica gel (MeOH/DCM 1/30-1/20) to give N,N-dimethyl-2-(2-nitro-5-(pyridin-4-yl)phenoxy)ethanamine (19*6) as a brown-orange solid (311.00 mg, 100%).

To a solution of 19*6 (155 mg, 539.48 μmol) in MeOH (5 ml) was added Raney nickel (20 mg) and Hydrazine (200 ul), the reaction mixture was stirred at rt for 3h. The mixture was filtered, and the filtrate was evaporated by rotary evaporation to give 2-(2-(dimethylamino)ethoxy)-4-(pyridin-4-yl)aniline (20*6) as a brown solid (140 mg, 100%).

To a solution of 20*6 (140.00 mg, 544.05 μmol), LA (123.48 mg, 598.45 μmol) and DIEA (269.75 μl, 1.63 mmol) in DMF (5 ml) was added HATU (268.92 mg, 707.26 μmol) and the solution was stirred at rt overnight. After the solvent was removed by rotary evaporation, saturated aq. NaHCO₃ was added and then extracted with EtOAc (15 ml × 3). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄ and filtered, the filtrate was concentrated and the residue was purified by flash chromatography on silica gel (MeOH/DCM 1/50-1/20) to give compound 6.

General Procedure B to synthesize 9-16: The synthesis of compound 14 was exemplified here.

To a mixture of crude 18*6 (300.00 mg, 1.04 mmol), 1-Boc-4-pyrazoleboronic acid pinacol ester (305.22 mg, 1.04 mmol), PdCl₂(dppf) (75.92 mg, 103.76 μmol) and Cs₂CO₃ (676.15 mg, 2.08 mmol) in a sealed tube, a mixed solvent (dioxane/H₂O =10/1, 6ml) were then injected under Argon and the mixture was stirred at 70 °C for 6h. After cooling to room temperature, the solvent was removed by rotary evaporation and the residue was added addition of water and extracted with EtOAc (15 ml × 3) . The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄ and filtered, the filtrate was concentrated and the residue was purified by flash chromatography on silica gel (MeOH/DCM 1/50-1/20) to give tert-butyl 4-(3-(2-(dimethylamino)ethoxy)-4-nitrophenyl)-1H-pyrazole-1-carboxylate (21*14) as a white solid (249.00 mg, 63.8%).

To a solution of 21*14 (360.00 mg, 956.41 μmol) in MeOH (8 ml) was added 10%Pd/C, the reaction mixture was stirred at rt under H₂ balloon atmosphere for 1h. The mixture was filtered, and the filtrate was evaporated by rotary evaporation to give tert-butyl -(4-amino-3-(2-(dimethylamino)ethoxy)phenyl)-1H-pyrazole-1-carboxylate (22*14) as a brown solid (320.00 mg, 96.6%) .

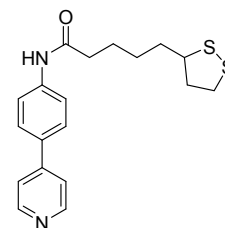
To a solution of 22*14 (228.00 mg, 658.15 μmol), LA (176.53 mg, 855.60 μmol) and DIEA (375.37 mg, 987.23 μmol) in DMF (2 ml) was added HATU (326.32 μl, 1.97 mmol) and the solution was stirred at rt overnight. After the solvent was removed by rotary evaporation, saturated aq. NaHCO₃ was added and then extracted with EtOAc (15ml × 3). The combined organic layers were washed with brine, dried over anhydrous Na₂SO₄ and filtered, the filtrate was concentrated and the residue was

purified by flash chromatography on silica gel (MeOH/DCM 1/50-1/20) to give tert-butyl4-(4-(5-(1,2-dithiolan-3-yl)pentanamido)-3-(2-(dimethylamino)ethoxy)phenyl)-1H-pyrazole-1-carboxylate (23*14) as a brown-orange solid (200.00 mg, 56.8%).

To a solution of 23*14 in DCM (2 mL) was injected TFA (0.4 mL) at rt. The mixture was stirred at rt for 2h and concentrated under a reduced pressure to remove TFA, then saturated aq. NaHCO₃ was added and the aqueous phase was extracted with DCM (15 ml × 3). The combined organic layers were washed with brine (10 mL), dried over anhydrous Na₂SO₄, filtered. The filtrate was concentrated and the residual solid was purified by flash chromatography on silica gel (MeOH/DCM 1/20) to give 14 as a faint yellow solid.

5-(1,2-dithiolan-3-yl)-N-(4-(pyridin-4-yl)phenyl)pentanamide

(1):

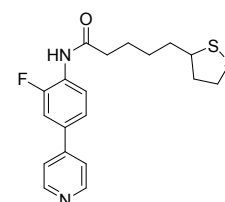


Similar to the General Procedure A, 1 was obtained as a faint

yellow solid at 52% yield for three steps. ¹H NMR (400 MHz, CDCl₃) δ 8.62 (d, *J* = 4.7 Hz, 2H), 7.96 (s, 1H), 7.68 (d, *J* = 8.1 Hz, 2H), 7.60 (d, *J* = 8.4 Hz, 2H), 7.48 (d, *J* = 5.2 Hz, 2H), 3.64 – 3.49 (m, 1H), 3.22 – 3.04 (m, 2H), 2.53 – 2.33 (m, 3H), 1.90 (dt, *J* = 19.7, 6.8 Hz, 1H), 1.84 – 1.62 (m, 4H), 1.61 – 1.43 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 171.4, 150.1, 147.7, 139.2, 133.4, 127.6, 121.3, 120.2, 56.4, 40.3, 38.5, 37.4, 34.7, 28.9, 25.2. ESI-HRMS for [C₁₉H₂₃N₂OS₂]⁺, calcd: 359.1246; found: 359.1232. Purity: 100%.

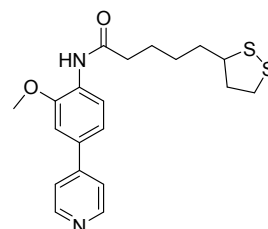
5-(1,2-dithiolan-3-yl)-N-(2-fluoro-4-(pyridin-4-

yl)phenyl)pentanamide (2):



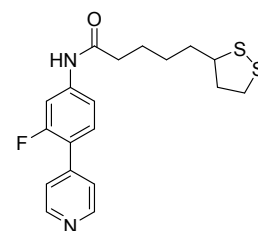
Similar to the General Procedure A, 2 was obtained as a faint yellow solid at 32.8% yield for three steps. ^1H NMR (400 MHz, CDCl_3) δ 8.65 (d, $J = 6.0$ Hz, 2H), 8.48 (t, $J = 8.2$ Hz, 1H), 7.51 – 7.35 (m, 5H), 3.66 – 3.53 (m, 1H), 3.25 – 3.06 (m, 2H), 2.57 – 2.37 (m, 3H), 1.93 (td, $J = 13.6, 6.9$ Hz, 1H), 1.87 – 1.72 (m, 4H), 1.62 – 1.47 (m, 2H). ^{13}C NMR (100 MHz, CDCl_3) δ 170.2, 152.7, 150.3, 149.3, 145.5 (d, $J = 1.8$ Hz), 133.0 (d, $J = 7.5$ Hz), 126.3, 126.2, 122.1 (d, $J = 3.0$ Hz), 121.1, 120.1, 112.3, 112.1, 55.3, 39.2, 37.5, 36.4, 33.6, 27.8, 24.1. ESI-HRMS for $[\text{C}_{19}\text{H}_{22}\text{FN}_2\text{OS}_2]^+$, calcd: 377.1158; found: 377.1151. Purity: 99%.

5-(1,2-dithiolan-3-yl)-N-(2-methoxy-4-(pyridin-4-yl)phenyl)pentanamide (3) :



Similar to the General Procedure A, 3 was obtained as a faint yellow solid at 22% yield for three steps. ^1H NMR (400 MHz, CDCl_3) δ 8.62 (dd, $J = 4.5, 1.6$ Hz, 2H), 8.50 (d, $J = 8.4$ Hz, 1H), 7.83 (s, 1H), 7.47 (dd, $J = 4.5, 1.6$ Hz, 2H), 7.29 – 7.23 (m, 1H), 7.11 (d, $J = 1.8$ Hz, 1H), 3.97 (s, 3H), 3.64 – 3.52 (m, 1H), 3.24 – 3.06 (m, 2H), 2.54 – 2.39 (m, 3H), 1.97 – 1.85 (m, 2H), 1.83 – 1.69 (m, 4H), 1.60 – 1.47 (m, 2H). ^{13}C NMR (100MHz, CDCl_3) δ 170.0, 149.2, 147.1, 147.0, 132.3, 127.7, 120.3, 119.0, 107.3, 55.4, 54.9, 39.3, 37.5, 36.7, 33.7, 27.8, 24.2. ESI-HRMS for $[\text{C}_{20}\text{H}_{25}\text{N}_2\text{O}_2\text{S}_2]^+$, calcd: 389.1352; found: 389.1350. Purity: 99%.

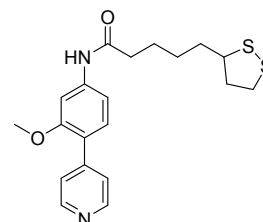
5-(1,2-dithiolan-3-yl)-N-(3-fluoro-4-(pyridin-4-yl)phenyl)pentanamide (4):



Similar to the General Procedure A, 4 was obtained as a faint yellow solid at 45.2% yield for three steps. ^1H NMR (400 MHz, CDCl_3) δ 8.66 (s, 1H),

8.63 (d, $J = 5.8$ Hz, 2H), 7.68 (d, $J = 12.9$ Hz, 1H), 7.47 (d, $J = 4.7$ Hz, 2H), 7.40 (t, $J = 8.4$ Hz, 1H), 7.33 (d, $J = 8.4$ Hz, 1H), 3.59 – 3.48 (m, 1H), 3.21 – 3.02 (m, 2H), 2.46 – 2.36 (m, 3H), 1.87 (td, $J = 13.6, 6.9$ Hz, 1H), 1.81 – 1.61 (m, 4H), 1.56 – 1.43 (m, 2H). ^{13}C NMR (100 MHz, CDCl_3) δ 171.8, 161.3, 158.8, 149.7, 143.5, 140.8, 140.6, 130.3 (d, $J = 4.3$ Hz), 123.5 (d, $J = 3.7$ Hz), 121.1, 121.0, 115.6, 108.1, 107.8, 56.4, 40.3, 38.5, 37.3, 34.6, 28.9, 25.2. ESI-HRMS for $[\text{C}_{19}\text{H}_{22}\text{FN}_2\text{OS}_2]^+$, calcd: 377.1152; found: 377.1136. Purity: 98%.

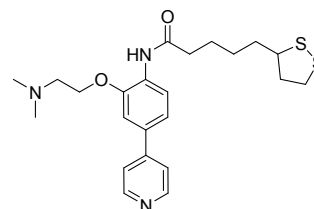
5-(1,2-dithiolan-3-yl)-N-(3-methoxy-4-(pyridin-4-yl)phenyl)pentanamide (5):



Similar to the General Procedure A, 5 was obtained as a yellow

solid at 78.8% yield for three steps. ^1H NMR (400 MHz, CDCl_3) δ 8.59 (dd, $J = 4.6, 1.6$ Hz, 2H), 7.78 (s, 1H), 7.65 (s, 1H), 7.47 (dd, $J = 4.5, 1.6$ Hz, 2H), 7.28 (d, $J = 8.3$ Hz, 1H), 6.96 (dd, $J = 8.3, 2.0$ Hz, 1H), 3.84 (s, 3H), 3.58 (td, $J = 12.7, 6.4$ Hz, 1H), 3.26 – 3.06 (m, 2H), 2.51 – 2.39 (m, 3H), 1.91 (dt, $J = 19.7, 6.9$ Hz, 1H), 1.84 – 1.69 (m, 4H), 1.58 – 1.48 (m, 2H). ^{13}C NMR (100 MHz, CDCl_3) δ 171.5, 157.2, 149.2, 146.2, 140.3, 130.5, 124.2, 122.9, 111.7, 103.4, 56.4, 55.6, 40.3, 38.5, 37.4, 34.7, 28.9, 25.2. ESI-HRMS for $[\text{C}_{20}\text{H}_{25}\text{N}_2\text{O}_2\text{S}_2]^+$, calcd: 389.1352; found: 389.1342. Purity: 99%.

N-(2-(2-(dimethylamino)ethoxy)-4-(pyridin-4-yl)phenyl)-5-(1,2-dithiolan-3-yl)pentanamide (6):

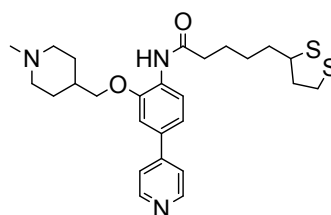


A faint yellow solid at 67.8% yield for four steps. ^1H NMR

(400 MHz, CDCl_3) δ 9.34 (s, 1H), 8.54 (dd, $J = 4.5, 1.6$ Hz, 2H), 8.40 (d, $J = 8.5$ Hz, 1H), 7.39 (dd, $J = 4.5, 1.6$ Hz, 2H), 7.24 (dd, $J = 8.5, 2.0$ Hz, 1H), 7.14 (d, $J = 2.0$ Hz,

1H), 4.12 (t, J = 5.2, 2H), 3.57 – 3.48 (m, 1H), 3.17 – 2.99 (m, 2H), 2.63 (t, J = 5.2, 2H), 2.41 – 2.34 (m, 3H), 2.29 (s, 6H), 1.85 (td, J = 13.8, 7.0 Hz, 1H), 1.76 – 1.62 (m, 4H), 1.53 – 1.40 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 170.7, 149.1, 147.0, 146.8, 132.2, 130.0, 120.2, 112.1, 67.0, 57.0, 55.4, 44.2, 39.2, 37.5, 36.2, 33.7, 28.0, 24.3. ESI-HRMS for [C₂₃H₃₂N₃O₂S₂]⁺, calcd: 446.1930; found: 446.1929. Purity: 99%.

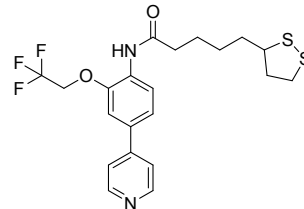
5-(1,2-dithiolan-3-yl)-N-(2-((1-methylpiperidin-4-yl)methoxy)-4-(pyridin-4-yl)phenyl)pentanamide (7):



Similar to the General Procedure A, 7 was obtained as a

brown yellow solid at 64.8% yield for four steps. ¹H NMR (400 MHz, CDCl₃) δ 8.62 (d, J = 4.6 Hz, 2H), 8.48 (d, J = 8.0 Hz, 1H), 7.81 (s, 1H), 7.47 (d, J = 4.7 Hz, 2H), 7.10 (s, 1H), 3.99 (d, J = 5.6 Hz, 2H), 3.64 – 3.53 (m, 1H), 3.22 – 3.08 (m, 2H), 3.02 – 2.91 (m, 3H), 2.50 – 2.42 (m, 3H), 2.33 (s, 3H), 2.08 – 1.97 (m, 6H), 1.89 – 1.82 (m, 4H), 1.58 – 1.51 (m, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 171.0, 150.2, 148.2, 147.5, 133.4, 128.7, 121.4, 120.2, 120.0, 109.4, 73.2, 56.4, 55.2, 46.2, 40.3, 38.5, 37.8, 35.2, 34.7, 29.0, 28.9, 25.3. ESI-HRMS for [C₂₆H₃₆N₃O₂S₂]⁺, calcd: 486.2243; found: 486.2234. Purity: 98%.

5-(1,2-dithiolan-3-yl)-N-(4-(pyridin-4-yl)-2-(2,2,2-trifluoroethoxy)phenyl)pentanamide (8):

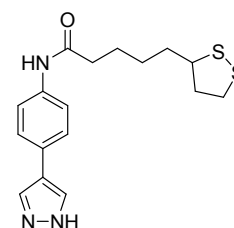


Similar to the General Procedure A, 8 was obtained as a white

solid at 40.4% yield for four steps. ¹H NMR (400 MHz, CDCl₃) δ 8.64 (d, J = 4.9 Hz, 2H), 8.52 (d, J = 8.3 Hz, 1H), 7.73 (s, 1H), 7.45 (d, J = 5.2 Hz, 2H), 7.36 (d, J = 8.1 Hz, 1H), 7.13 (s, 1H), 4.52 (dd, J = 15.7, 7.8 Hz, 2H), 3.69 – 3.52 (m, 1H), 3.29 –

3.04 (m, 2H), 2.59 – 2.37 (m, 3H), 1.92 (dt, $J = 19.7, 6.8$ Hz, 1H), 1.88 – 1.67 (m, 4H), 1.64 – 1.45 (m, 2H). ^{13}C NMR (100 MHz, CDCl_3) δ 170.0, 149.3, 146.2, 145.2, 132.7, 128.2, 123.5, 121.2, 120.7, 120.2, 120.1, 109.7, 66.5, 66.1, 65.8, 65.4, 55.3, 39.2, 37.5, 36.7, 33.7, 27.8, 24.1. ESI-HRMS for $[\text{C}_{21}\text{H}_{24}\text{F}_3\text{N}_2\text{O}_2\text{S}_2]^+$, calcd: 457.1226; found: 457.1225. Purity: 99%.

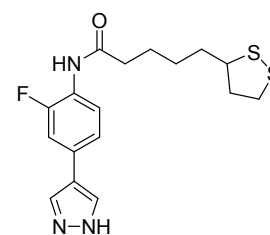
N-(4-(1H-pyrazol-4-yl)phenyl)-5-(1,2-dithiolan-3-yl)pentanamide (9):



Similar to the General Procedure B, 9 was obtained as a white

solid at 5.6% yield for four steps. ^1H NMR (400 MHz, DMSO) δ 12.81 (s, 1H), 9.86 (s, 1H), 7.97 (s, 2H), 7.57 (d, $J = 8.6$ Hz, 2H), 7.51 (d, $J = 8.7$ Hz, 2H), 3.74 – 3.56 (m, 1H), 3.26 – 3.05 (m, 2H), 2.41 (td, $J = 12.5, 6.3$ Hz, 1H), 2.30 (t, $J = 7.3$ Hz, 2H), 1.87 (dt, $J = 19.7, 6.8$ Hz, 1H), 1.77 – 1.55 (m, 4H), 1.46 – 1.37 (m, 2H). ^{13}C NMR (100 MHz, DMSO) δ 170.9, 137.2, 127.7, 125.3, 120.9, 119.4, 56.1, 38.1, 36.2, 34.1, 28.3, 24.9. ESI-HRMS for $[\text{C}_{17}\text{H}_{22}\text{N}_3\text{OS}_2]^+$, calcd: 348.1199; found: 348.1195. Purity: 97%.

5-(1,2-dithiolan-3-yl)-N-(2-fluoro-4-(1H-pyrazol-4-yl)phenyl)pentanamide(10):

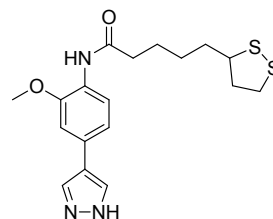


Similar to the General Procedure B, 10 was obtained as a faint

yellow solid at 7% yield for four steps. ^1H NMR (400 MHz, DMSO) δ 12.94 (s, 1H), 9.60 (s, 1H), 8.06 (s, 2H), 7.77 (t, $J = 8.4$ Hz, 1H), 7.49 (dd, $J = 12.3, 1.8$ Hz, 1H), 7.38 (dd, $J = 8.3, 1.5$ Hz, 1H), 3.62 (td, $J = 12.3, 6.2$ Hz, 1H), 3.22 – 3.05 (m, 2H), 2.45 – 2.31 (m, 3H), 1.87 (dq, $J = 13.5, 6.8$ Hz, 1H), 1.75 – 1.52 (m, 4H), 1.48 – 1.33 (m, 5H). ^{13}C NMR (100 MHz, DMSO) δ 171.4, 155.4, 152.9, 130.4 (d, $J = 7.6$ Hz),

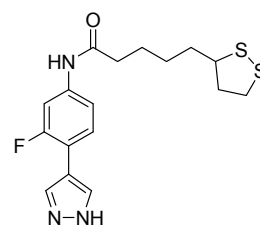
124.7, 123.7, 123.6, 120.6 (d, $J = 2.8$ Hz), 120.0 (d, $J = 1.9$ Hz), 111.9, 111.7, 56.1, 38.1, 35.5, 34.1, 28.2, 24.9. ESI-HRMS for $[C_{17}H_{21}FN_3OS_2]^+$, calcd: 366.1105; found: 366.1098. Purity: 97%.

5-(1,2-dithiolan-3-yl)-N-(2-methoxy-4-(1H-pyrazol-4-yl)phenyl)pentanamide (11):



Similar to the General Procedure B, 11 was obtained as a white solid at 14.3% yield for four steps. 1H NMR (400 MHz, DMSO) δ 12.90 (s, 1H), 9.02 (s, 1H), 8.34 – 7.76 (m, 3H), 7.23 (s, 1H), 7.13 (d, $J = 8.2$ Hz, 1H), 3.88 (s, 3H), 3.62 (td, $J = 12.5, 6.2$ Hz, 1H), 3.26 – 3.06 (m, 2H), 2.47 – 2.32 (m, 3H), 1.87 (dq, $J = 13.5, 6.8$ Hz, 1H), 1.77 – 1.50 (m, 4H), 1.48 – 1.33 (m, 2H). ^{13}C NMR (100 MHz, DMSO) δ 171.6, 150.5, 129.7, 125.8, 122.9, 121.7, 117.3, 108.6, 56.7, 56.2, 38.6, 36.3, 34.6, 28.8, 25.5. ESI-HRMS for $[C_{18}H_{24}N_3O_2S_2]^+$, calcd: 378.1304; found: 378.1306. Purity: 99%.

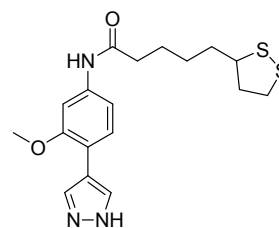
5-(1,2-dithiolan-3-yl)-N-(3-fluoro-4-(1H-pyrazol-4-yl)phenyl)pentanamide (12):



Similar to the General Procedure B, 12 was obtained as a faint yellow solid at 15.3% yield for four steps. 1H NMR (400 MHz, DMSO) δ 13.00 (s, 1H), 10.08 (s, 1H), 7.98 (s, 2H), 7.73 – 7.57 (m, 2H), 7.28 (dd, $J = 8.5, 1.8$ Hz, 1H), 3.67 – 3.56 (m, 1H), 3.24 – 3.06 (m, 2H), 2.47 – 2.37 (m, 1H), 2.33 (dd, $J = 18.4, 11.0$ Hz, 2H), 1.87 (td, $J = 13.4, 6.8$ Hz, 1H), 1.74 – 1.52 (m, 4H), 1.46 – 1.35 (m, 2H). ^{13}C NMR (100 MHz, DMSO) δ 171.8, 159.9, 157.4, 139.0, 138.9, 128.3 (d, $J = 5.9$ Hz), 115.5 (d, $J = 2.1$ Hz), 115.4, 115.3, 115.1 (d, $J = 1.8$ Hz), 107.0, 106.7, 56.6, 38.6,

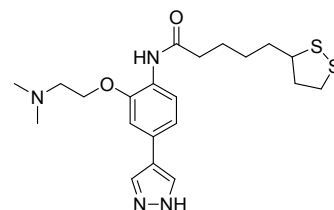
36.7, 34.6, 28.8, 25.2. ESI-HRMS for $[C_{17}H_{21}FN_3OS_2]^+$, calcd: 366.1105; found: 366.1090. Purity: 98%.

5-(1,2-dithiolan-3-yl)-N-(3-methoxy-4-(1H-pyrazol-4-yl)phenyl)pentanamide (13):



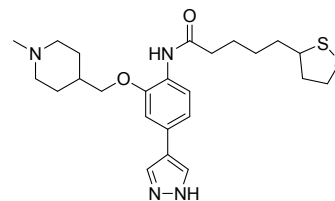
Similar to the General Procedure B, 13 was obtained as a orange solid at 4.3% yield for four steps. 1H NMR (400 MHz, DMSO) δ 12.78 (s, 1H), 9.89 (s, 1H), 7.96 (s, 2H), 7.50 (d, $J = 8.3$ Hz, 1H), 7.44 (d, $J = 1.6$ Hz, 1H), 7.15 (dd, $J = 8.4, 1.8$ Hz, 1H), 3.82 (s, 3H), 3.67 – 3.57 (m, 1H), 3.23 – 3.07 (m, 2H), 2.41 (dt, $J = 18.7, 6.3$ Hz, 1H), 2.30 (t, $J = 7.3$ Hz, 2H), 1.87 (td, $J = 13.5, 6.8$ Hz, 1H), 1.75 – 1.54 (m, 4H), 1.46 – 1.36 (m, 2H). ^{13}C NMR (100 MHz, DMSO) δ 171.0, 155.4, 138.4, 127.0, 116.8, 116.2, 111.2, 102.7, 56.1, 55.2, 38.1, 36.3, 34.1, 28.3, 24.8. ESI-HRMS for $[C_{18}H_{24}N_3O_2S_2]^+$, calcd: 378.1304; found: 378.1301. Purity: 98%.

N-(2-(2-(dimethylamino)ethoxy)-4-(1H-pyrazol-4-yl)phenyl)-5-(1,2-dithiolan-3-yl)pentanamide (14):



A faint yellow solid at 16.1% yield for four steps. 1H NMR (400 MHz, DMSO) δ 12.89 (s, 1H), 9.34 (s, 1H), 8.14 (s, 2H), 7.92 (d, $J = 8.0$ Hz, 1H), 7.31 (s, 1H), 7.18 (d, $J = 7.9$ Hz, 1H), 4.16 (t, $J = 4.8$ Hz, 2H), 3.71 – 3.53 (m, 1H), 3.22 – 3.11 (m, 2H), 2.63 (t, $J = 4.8$ Hz, 2H), 2.46 – 2.38 (m, 1H), 2.38 – 2.30 (m, 2H), 2.27 (s, 6H), 1.87 (dt, $J = 19.2, 6.5$ Hz, 1H), 1.80 – 1.50 (m, 4H), 1.50 – 1.30 (m, 2H). ^{13}C NMR (100 MHz, DMSO) δ 171.3, 149.4, 136.9, 129.6, 127.5, 125.9, 122.5, 121.5, 118.5, 112.3, 68.3, 58.0, 56.6, 55.4, 45.7, 38.6, 36.6, 34.7, 28.8, 25.5. ESI-HRMS for $[C_{21}H_{31}N_4O_2S_2]^+$, calcd: 435.1883; found: 435.1883. Purity: 99%.

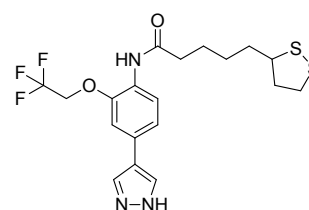
5-(1,2-dithiolan-3-yl)-N-(2-((1-methylpiperidin-4-yl)methoxy)-4-(1H-pyrazol-4-yl)phenyl)pentanamide



(15):

Similar to the General Procedure B, 15 was obtained as a white solid at 24.6% yield for four steps. ¹H NMR (400 MHz, CDCl₃) δ 8.34 (d, *J* = 8.2 Hz, 1H), 7.80 (s, 2H), 7.73 (s, 1H), 7.09 (dd, *J* = 8.3, 1.7 Hz, 1H), 6.97 (d, *J* = 1.7 Hz, 1H), 3.96 (d, *J* = 5.9 Hz, 2H), 3.64 – 3.52 (m, 1H), 3.24 – 3.08 (m, 2H), 3.06 – 2.96 (m, 2H), 2.46 – 2.40 (m, 3H), 2.36 (s, 3H), 2.15 – 2.05 (m, 3H), 1.95 – 1.83 (m, 4H), 1.79 – 1.71 (m, 3H), 1.63 – 1.52 (m, 4H). ¹³C NMR (100 MHz, CDCl₃) δ 170.8, 147.6, 131.0, 128.3, 126.3, 122.5, 120.5, 118.5, 108.7, 73.1, 56.4, 55.2, 46.1, 40.3, 38.5, 37.7, 35.2, 34.7, 29.7, 28.9, 25.4. ESI-HRMS for [C₂₄H₃₅N₄O₂S₂]⁺, calcd: 475.2196; found: 475.2179. Purity: 99%.

N-(4-(1H-pyrazol-4-yl)-2-(2,2,2-trifluoroethoxy)phenyl)-5-(1,2-dithiolan-3-yl)pentanamide (16)



Similar to the General Procedure B, 16 was obtained as a white solid at 8.7% yield for four steps. ¹H NMR (400 MHz, DMSO) δ 12.91 (s, 1H), 9.01 (s, 1H), 8.09 (s, 2H), 7.66 (d, *J* = 7.0 Hz, 1H), 7.40 (s, 1H), 7.25 (d, *J* = 7.8 Hz, 1H), 4.81 (d, *J* = 8.6 Hz, 2H), 3.68 – 3.60 (m, 1H), 3.25 – 2.98 (m, 2H), 2.46 – 2.22 (m, 3H), 1.96 – 1.80 (m, 1H), 1.80 – 1.52 (m, 4H), 1.49 – 1.36 (m, 2H). ¹³C NMR (100 MHz, DMSO) δ 171.7, 149.7, 130.7, 125.9, 125.8, 125.2, 123.1, 121.2, 119.2, 111.2, 66.6, 66.3, 65.9, 65.6, 56.6, 38.6, 36.3, 34.7, 28.7, 25.5. ESI-HRMS for

$[\text{C}_{19}\text{H}_{23}\text{F}_3\text{N}_3\text{O}_2\text{S}_2]^+$, calcd: 446.1178; found: 446.1161. Purity: 98%.

Cell Culture HT22 cells were maintained in DMEM supplemented with 10% (v/v) fetal bovine serum and incubated at 37 °C under 5% CO₂. Test compounds were dissolved in DMSO and diluted in DMEM supplemented with 10% (v/v) FBS. The final concentration of DMSO in the medium was less than 0.01% (vol/vol) which showed no influence on cell growth.

MTT assay To study the neuroprotective effects of tested compounds, HT22 cells were seeded in 96-well plates at a density of 4×10^3 in triplicate cultures and incubated overnight. Cells were pretreated with tested compounds or the vehicle DMSO for 30 min followed with / without 2 mM glutamate for 24 h. 10 μL of 5 mg/mL MTT was added to each well and cells were incubated for 2 h at 37 °C. To dissolve purple formazan crystal, 100 μL DMSO was carefully added to replace the medium. After vigorously shaking for 15 min at 37°C, the absorbance at 570 nm was measured using a microculture plate reader (Bio-Tek, USA).

Kinase activity assay Kinase inhibition by tested compounds were evaluated by a fluorescence resonance energy transfer based Z'-LYTE kinase assay kit following our described protocols¹. The IC₅₀ value of compound 15 for inhibition of ROCK 1/2 kinases to determine the selectivity of 15 between ROCK1 and ROCK2 was carried out by SelectScreen® Kinase Profiling Services (Life Technologies). Measurements were performed using a Km app Z'-LYTE assay (Assay ID: 784, 787) in duplicates at each concentration (10 points were detected, Fig. S1).

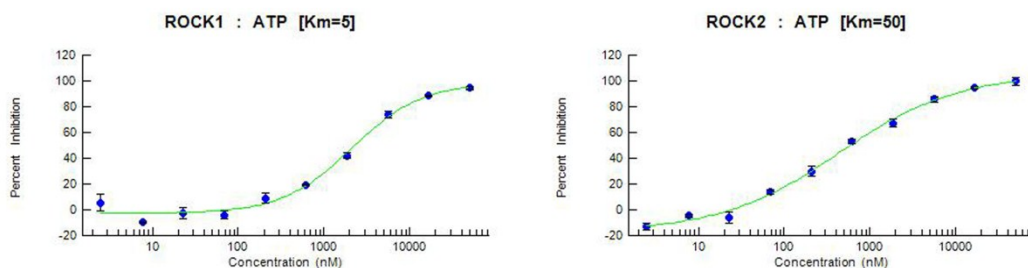


Fig. S1 Dose response data for ROCK inhibition by compound 15.

PAMPA-BBB assay Prediction of the brain penetration of compounds was evaluated using a parallel artificial membrane permeation assay (PAMPA) and porcine brain lipid (Avanti Polar Lipids) in a similar manner as our previously described¹.

Table S1 Permeability ($P_e \times 10^{-6} \text{ cm s}^{-1}$) in the PAMPA-BBB assay for the selected compounds and their predicted penetration into the CNS.

Comp.	Permeability ($P_e \times 10^{-6} \text{ cm s}^{-1}$) ¹⁾	Prediction	Comp.	Permeability ($P_e \times 10^{-6} \text{ cm s}^{-1}$) ¹⁾	Prediction
1	3.69±0.05	CNS+/-	9	2.93±0.09	CNS+/-
2	12.3±0.83	CNS+	10	7.65±0.17	CNS+
3	NT ^a	-	11	NT ^a	-
4	7.71±1.31	CNS+	12	4.78±0.62	CNS+
5	9.15±0.65	CNS+	13	12.23±0.25	CNS+
6	12.87±0.98	CNS+	14	6.72±0.70	CNS+
7	8.78±0.79	CNS+	15	5.70±0.48	CNS+
8	13.26±0.33	CNS+	16	13.49±0.49	CNS+

^aNT, not tested (insoluble in the buffer).

Table S2 Ranges of Permeability of PAMPA-BBB Assays ($P_e, 10^{-6} \text{ cm s}^{-1}$)²

Compounds of high BBB permeation (CNS+)	$P_e > 4.7$
Compounds of uncertain BBB permeation (CNS+/-)	$4.7 > P_e > 1.8$
Compounds of low BBB permeation (CNS-)	$P_e < 1.8$

Measurement of intracellular ROS HT22 cells were grown in Corning 96-well plates at a cell density 4×10^3 cells/well. After overnight attachment, cells were

pretreated with 15, LA or the vehicle control DMSO for 30 min and then incubated with / without 2 mM glutamate for 10 h. Cells were washed twice with phosphate-buffered saline (PBS) and then incubated with 10 μ M non-fluorescent dye DHE in serum-free medium for 30 min at 37 °C in the dark. Cells were subsequently washed twice with PBS, photographed and analyzed by a high content system (ArrayScanVTI, Thermo Fisher). Values were expressed as a percentage of the fluorescence relative to the vehicle control.

Glutathione (GSH) assay kit Intracellular GSH concentration was tested by a GSH assay kit (48 T, Nanjing Jiancheng, China). By reacting with dithiobisnitrobenzoic acid, reduced GSH could form a yellow compound, which is quantifiable at 405 nm and reflect the content of the reduced GSH indirectly. In brief, after treatment, whole-cell lysate was prepared and tested according to manufacturer's instructions. All GSH values were normalized to per μ g protein of each sample.

Rat aorta assay Male Sprague-Dawley (SD) rats (200 ± 20 g) were supplied by the Center of Experimental Animals, Sun Yat-sen University (Guangzhou, China, Certificate No.SCXK 20110029). The rats were housed in standard cages under controlled temperature conditions of 21 °C with free access to food and water until 12 h prior to experiments. The vasorelaxant effects were examined by a rat thoracic aorta assay. Briefly, animals were anesthetized with 0.1% sodium pentobarbital (50 mg/kg). The thoracic aorta was quickly isolated and cleaned of fat and adherent connective tissue. Aortic rings 2–3 mm in length were prepared and mounted horizontally in 5

mL organ bath containing the Krebs–Henseleit (K-H) solution at 37 °C, bubbled with 95% O₂ and 5% CO₂. The ring was allowed to equilibrate for 30min under 2 g resting tension. Then, KCl (60 mM) was used to induce a similar sustained contraction of thoracic aorta rings with a peak tension in each group. Following, fasudil, compound 15 were cumulatively added into the organ bath in 15min intervals respectively. DMSO was also added into the bath as a control. The contractile responses were expressed as a percentage of the responses following and before the application of the testing compounds. Results are expressed as mean ± S.D. Statistical analysis was performed with repeated measures of the ANOVA. Differences were accepted as statistically significant at *P* values < 0.05.

Molecular docking In order to understand the interactions between 15 and ROCK2 in the active site in detail, a Molecular Operating Environment (MOE 2010) (Chemical Computing Group Inc, Montreal, Quebec, Canada) was used for molecular docking. 15 was constructed using the builder module and were energy minimized using Force Field MMFF94x and saved as MDB file. The crystal structure of ROCK2 with fasudil was downloaded from the protein data bank (PDB ID: 2F2U), water molecules were removed, and one of the chain was deleted as the ROCK2 is a dimer. The residual crystal structure was prepared using ligX with the the default parameters of MOE [gradient: 0.1, Force Field: MMFF94X]. Define 5Å radius around the bound inhibitor (fasudil) in the crystal structure as active site. The prepared ligand structure was flexibly docked into the ROCK2 binding site, triangle matcher as the placement methodology, LondonΔG as scoring methodology, Forcefield Refinement was

selected and dock calculations was run automatically. The obtained 30 conformations were generated and stored in database, the best conformation was analyzed for the binding interaction.

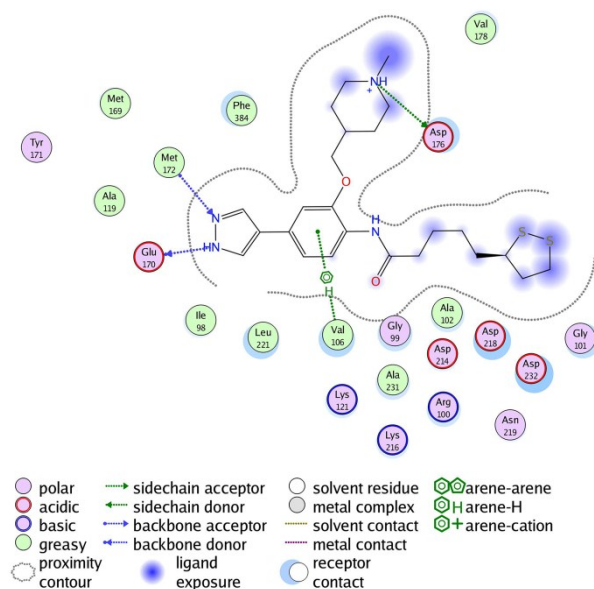


Fig. S2 2D representation of the binding mode of **15** in ROCK-II (2F2U).

Statistical analysis All quantitative data and experiments described in this study were repeated at least three times. Data were presented as mean \pm S.D. of multiple independent experiments. Statistics were analyzed with one-way ANOVA followed by a least significant difference test (Graphpad Prism 5.0 software (US)). Statistical difference was considered at $P < 0.05$.

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