

Supporting Information

Porphyrinic MOFs for reversible fluorescent and colorimetric sensing of mercury (II) ions in aqueous-phase

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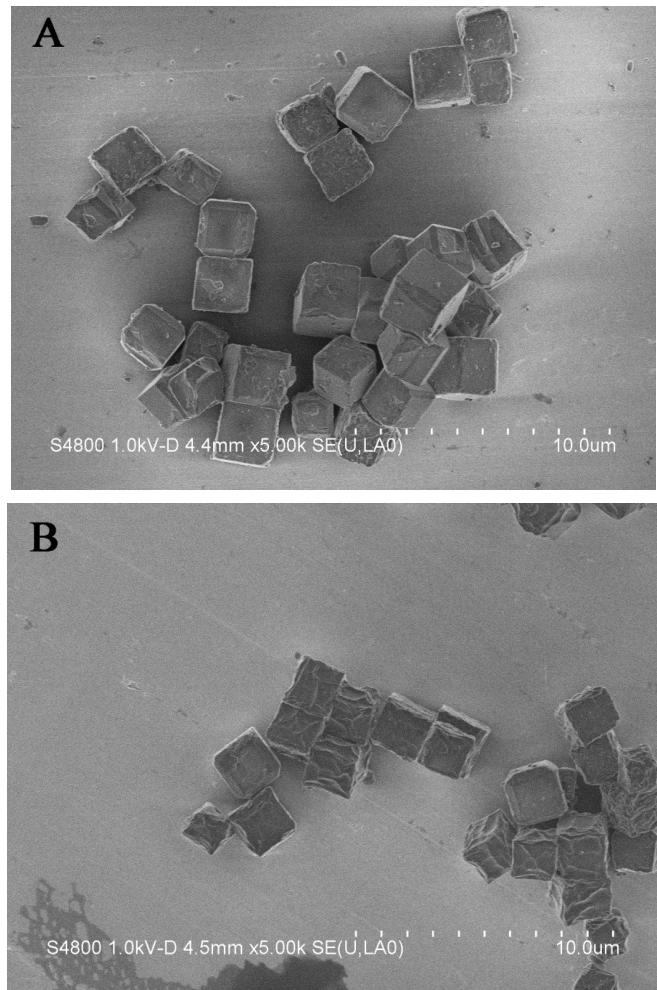


Fig. S1 Scanning electron microscopy (SEM) images of (A) the as-synthesized PCN-224 and (B) PCN-224-KI particles regenerated by KI solution.

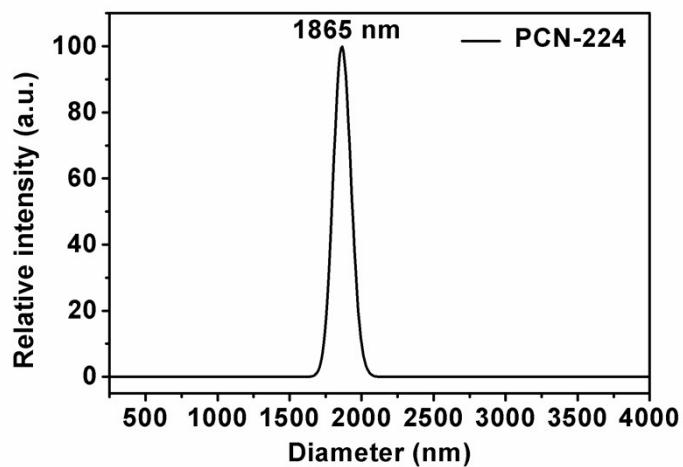


Fig. S2 Typical DLS profile of PCN-224 particles measured in aqueous solution.

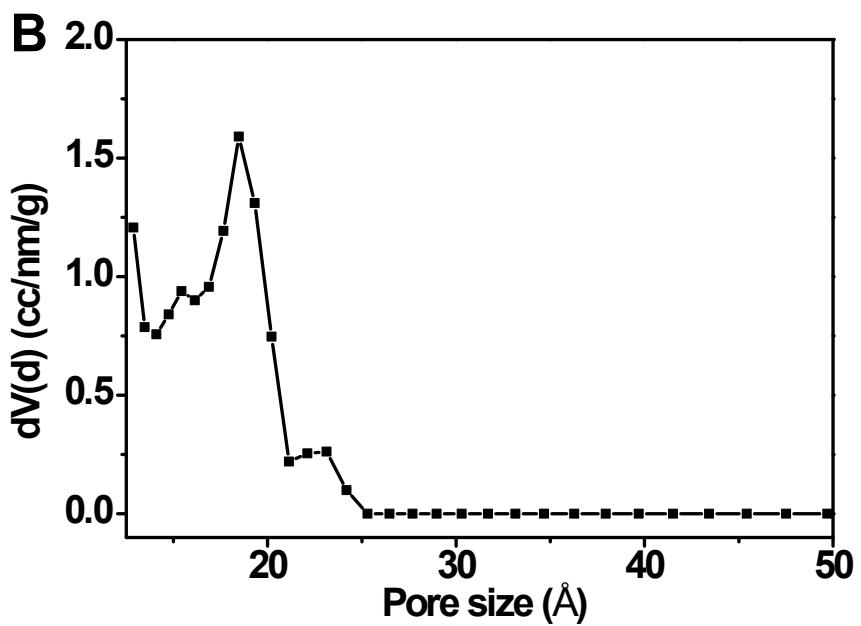
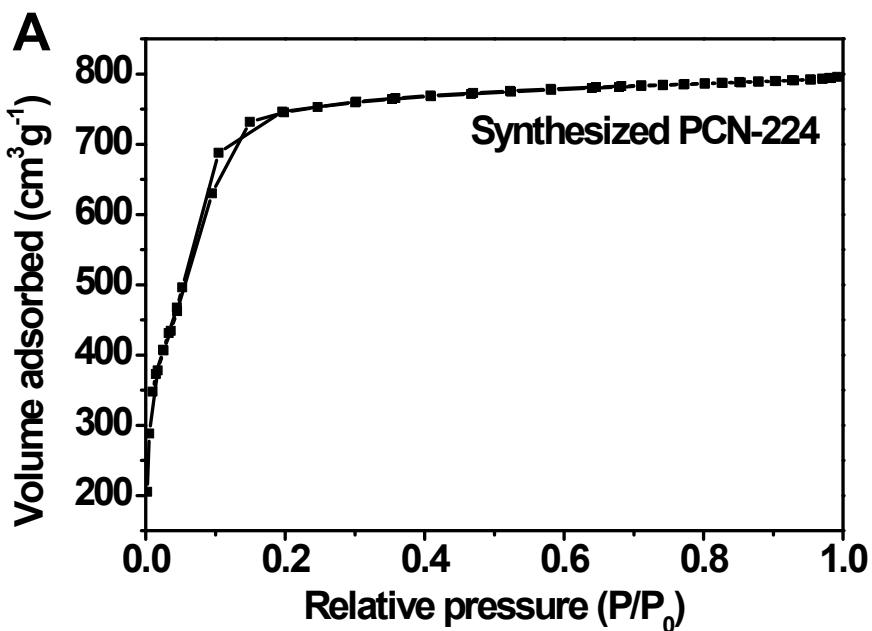


Fig. S3 (A) N₂ sorption isotherm at 77 K for the as-synthesized PCN-224 particles and (B) is the corresponding pore size distribution calculated by the DFT method.

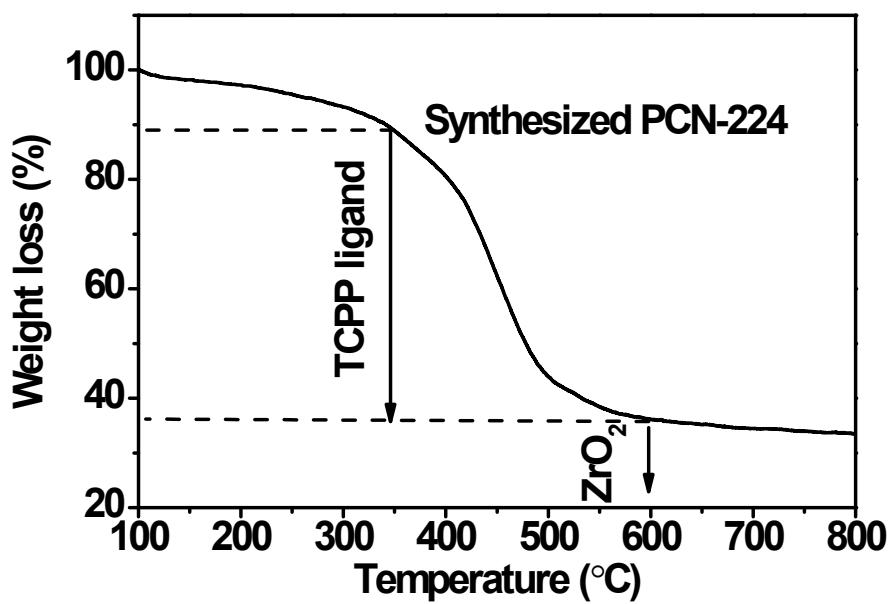


Fig. S4 TGA profile for the as-synthesized PCN-224 particles recorded under air flow.

The calculation for the limit of detection (LOD)

$$\text{Detection Limit} = 3\sigma/\text{slope}$$

$$= (3 \times 0.17)/81.1 \times 1000$$

$$= 6 \text{ nM}$$

Multiple number of PL spectra ($n = 10$) were recorded for the blank sample of PCN-224 suspension. Sample standard deviation σ for the blank probe without the addition of Hg^{2+} was calculated to be 0.17.

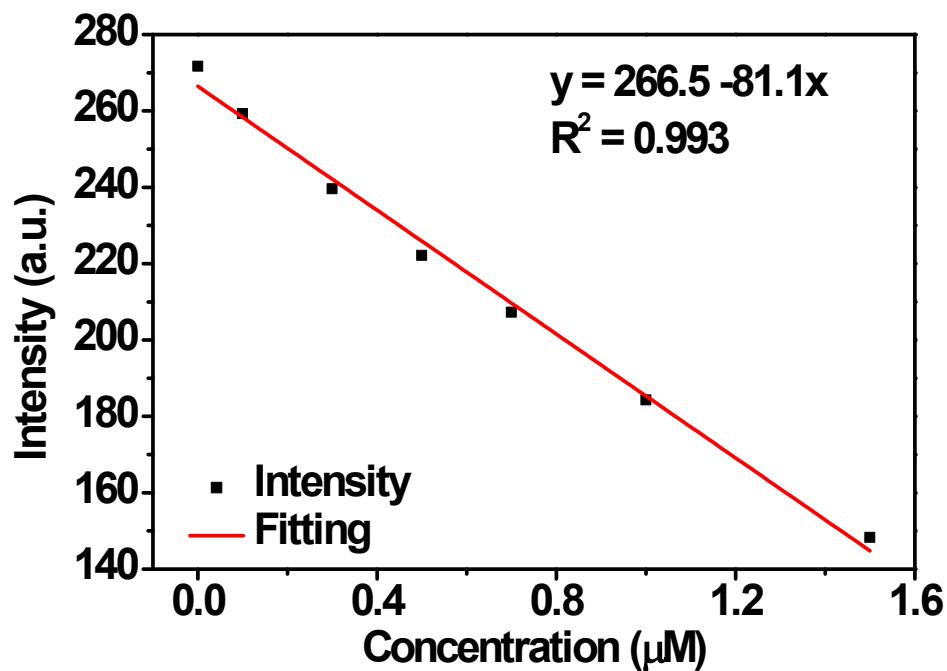


Fig. S5 Relation of fluorescence intensity against Hg^{2+} added into PCN-224 suspension and their linear fitting curve for the estimation of LOD.

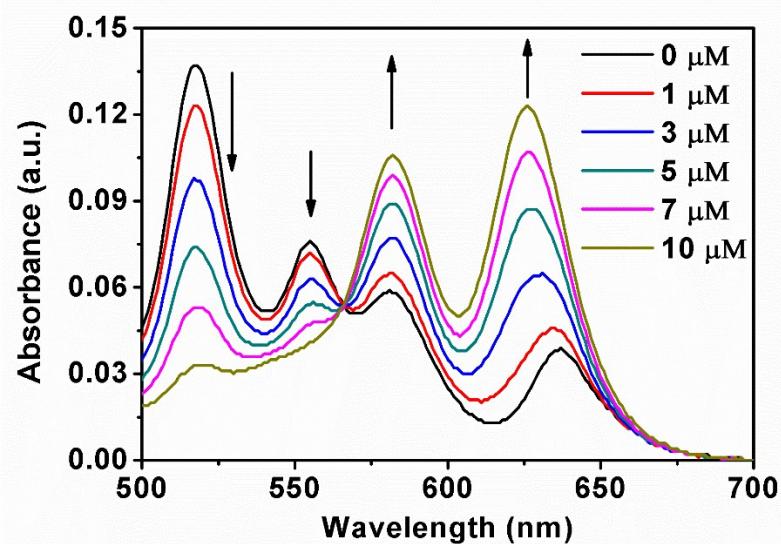
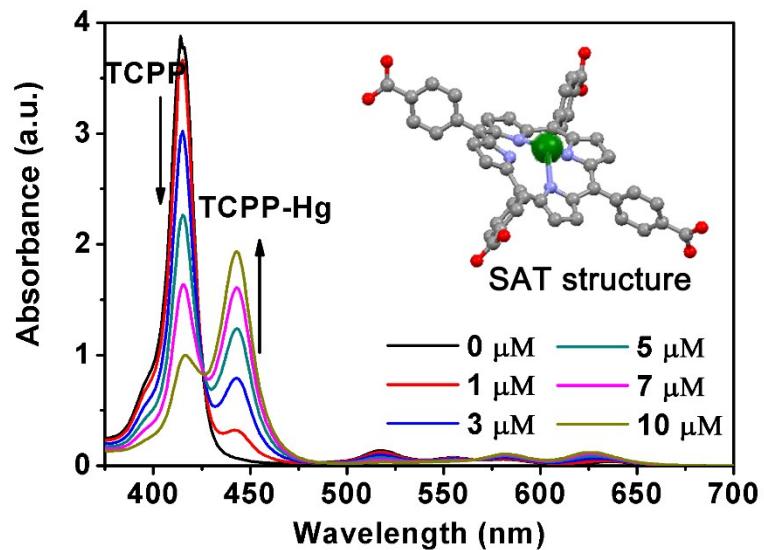


Fig. S6 The evolvement of UV-Vis spectra of (A) the Soret band and (B) the enlargement of Q bands for free TCPP molecules (10 mg L^{-1}) in DMF/HEPES buffer solution (v/v = 1:1, pH = 7) upon the addition of various concentrations of Hg^{2+} . In the inset, schematic representations of the interaction mechanisms between the porphyrin molecule and Hg^{2+} ions in the PCN-224 probe, which is a “sitting-atop” (SAT) structure.

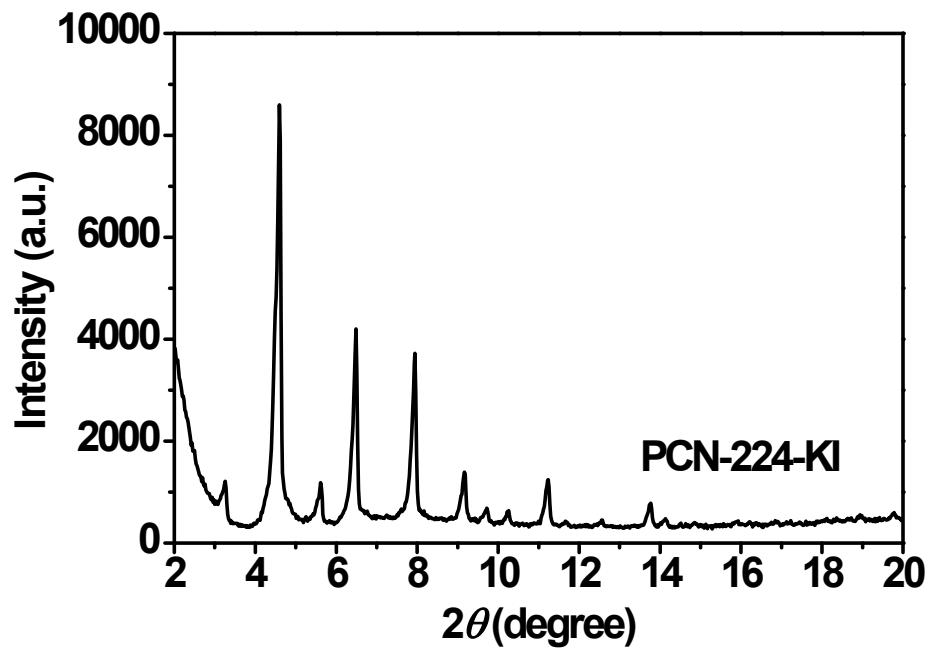


Fig. S7 Powder XRD patterns of the PCN-224-KI particles regenerated by KI solution.

Table S1. The Comparison of Sensing Features between the Current PCN-224 Sensor and Other Reported Probes for the Detection of Hg^{2+} .

Probe	Material	Detection range (μM)	LOD (nM)	Response Time	Regeneration	References
MOF-based sensors	$\{\text{Eu}_2(\text{bqdc})_3(\text{H}_2\text{O})(\text{DMF})_3\} \cdot 0.5 \text{ DMF} \cdot \text{H}_2\text{O}\}_n$	10-1000	10	4 h	—	1
	$\text{TbL}_{1.5}(\text{H}_2\text{O})_2 \cdot \text{H}_2\text{O}$	1-1000	—	—	EDTA	2
	$\text{Zr}_6\text{O}_4(\text{OH})_4(\text{BDC})_6$	0.001-0.5	52	10 s	ClO_4^-	3
	$\{\text{Cd}_{1.5}(\text{C}_{18}\text{H}_{10}\text{O}_{10})\} \cdot (\text{H}_3\text{O})(\text{H}_2\text{O})_3\}_n$	4-25	2	15s	—	4
	UiO-66-NH ₂ @DNA	0.1-10	17.6	—	—	5
Other porphyrin-based sensors	COF-LZU8	0.33-33.3	125	Real-time	Na_2S	6
	H ₂ TPP	0.04-450	40	4 min	3-MPA	7
	TDMAPP	0.04-4	8	—	HCl	8
	naphthalimide-porphyrin	0.1-50	20	2 min	EDTA	9
	DTPP	0.5-310	—	9 min	HCl	10
	MTHNP	0.005-12.5	3	3 min	buffer ^a	11
	cationic porphyrin ^b	0.0001-1	0.1	—	—	12
PCN-224 sensor	TPPS@SBA-15	0.025-0.5	17.5	4 min	—	13
	$\text{Zr}_6\text{O}_4(\text{OH})_4(\text{TCPP})_{1.5}$	0.1-10	6	2 min	KI	This work

^a the blank phosphate buffer of pH 7.5

^bthe cationic 5,15-(*p*-(9,9-bis(6-trimethylammoniumhexyl)fluorenylethynyl)phenyl)-porphyrin tetrabromide

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