

## Supporting Information

### Boranil Dye Based “Turn-on” Fluorescent Probes for Detection of Hydrogen Peroxide and Their Cell Imaging Application

Jayabalan Shanmugapriya,<sup>a‡</sup> Kandasamy Rajaguru,<sup>b‡</sup> Gandhi Sivaraman,<sup>c‡</sup> Shanmugam Muthusubramanian,<sup>\*b</sup> and Nattamai Bhuvanesh<sup>d</sup>

<sup>a</sup> *Department of Chemistry, Thiagarajar College of Engineering, Madurai – 625 01, India*

<sup>b</sup> *Department of Organic Chemistry, School of Chemistry, Madurai Kamaraj University, Madurai - 625 021, India*

<sup>c</sup> *Institute for Stem Cell Biology and Regenerative Medicine, National Centre for Biological Sciences, Bangalore – 560065, India*

<sup>d</sup> *X-ray Diffraction Laboratory, Department of Chemistry, Texas A & M University, College Station, Texas, 77842, USA*

<sup>‡</sup> *These authors have contributed equally*

*\*e-mail: [muthumanian2001@yahoo.com](mailto:muthumanian2001@yahoo.com)*

<b>Table of contents</b>	<b>Page No</b>
$^1\text{H}$ , $^{13}\text{C}$ , $^{11}\text{B}$ and $^{19}\text{F}$ NMR spectra of compound <b>SB-1</b>	S3
$^1\text{H}$ , $^{13}\text{C}$ , $^{11}\text{B}$ and $^{19}\text{F}$ NMR spectra of compound <b>SB-2</b>	S7
Table S1	S11
Optimized geometries of <b>SB-1</b> and <b>SB-2</b>	S21
Table S2	S25

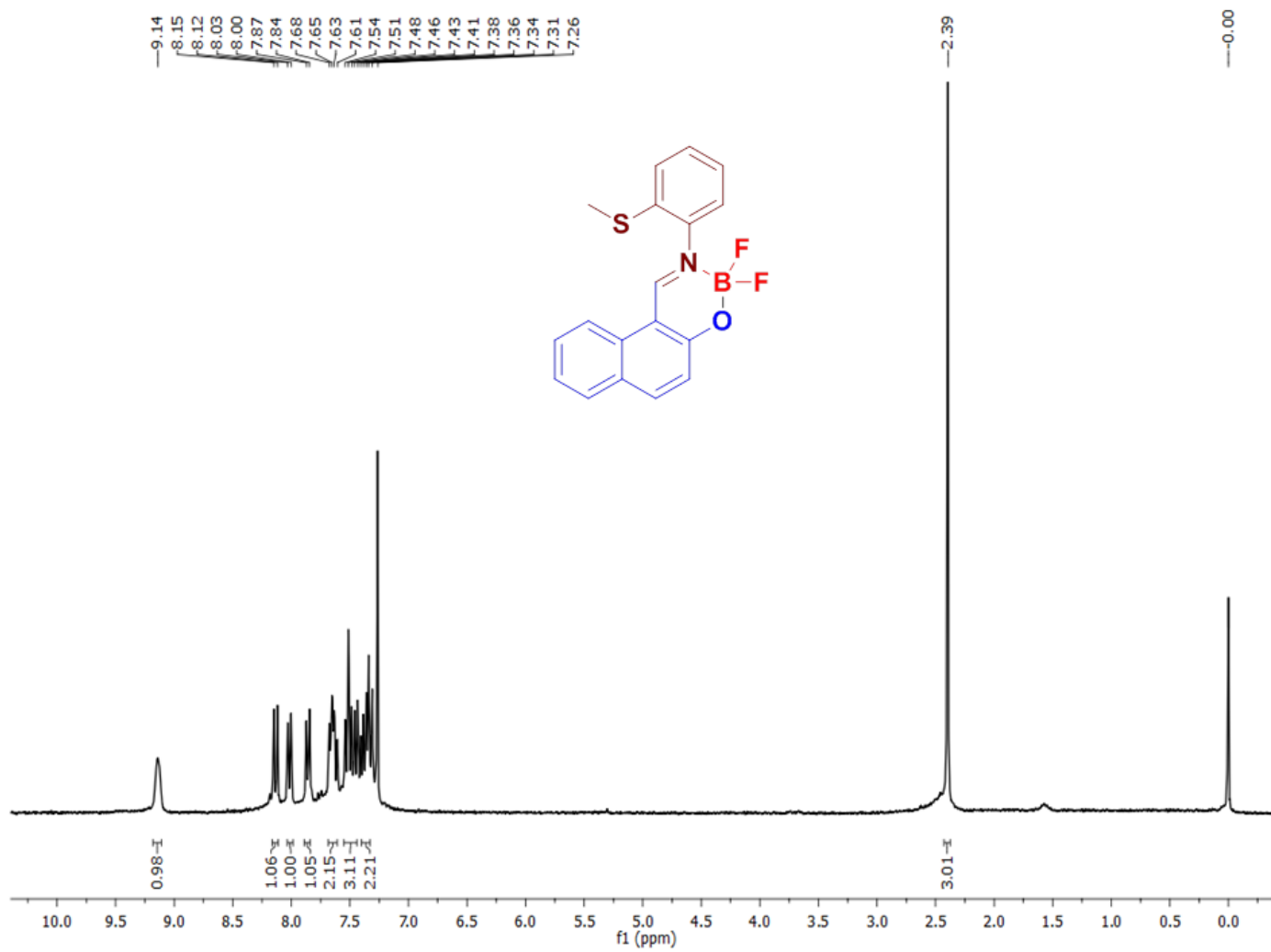


Figure S1. <sup>1</sup>H NMR spectrum of compound **SB-1**

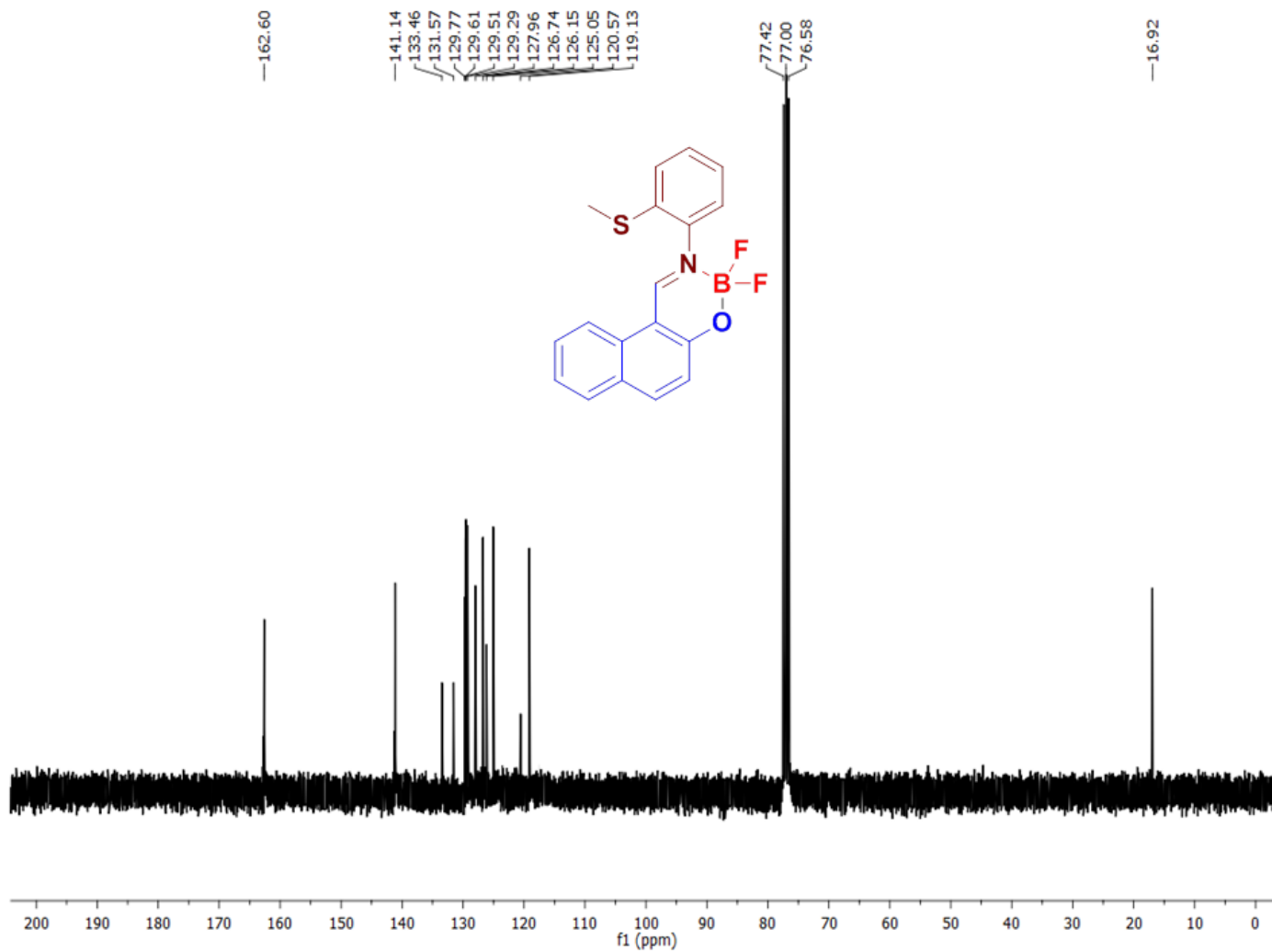


Figure S2.  $^{13}\text{C}$  NMR spectrum of compound SB-1

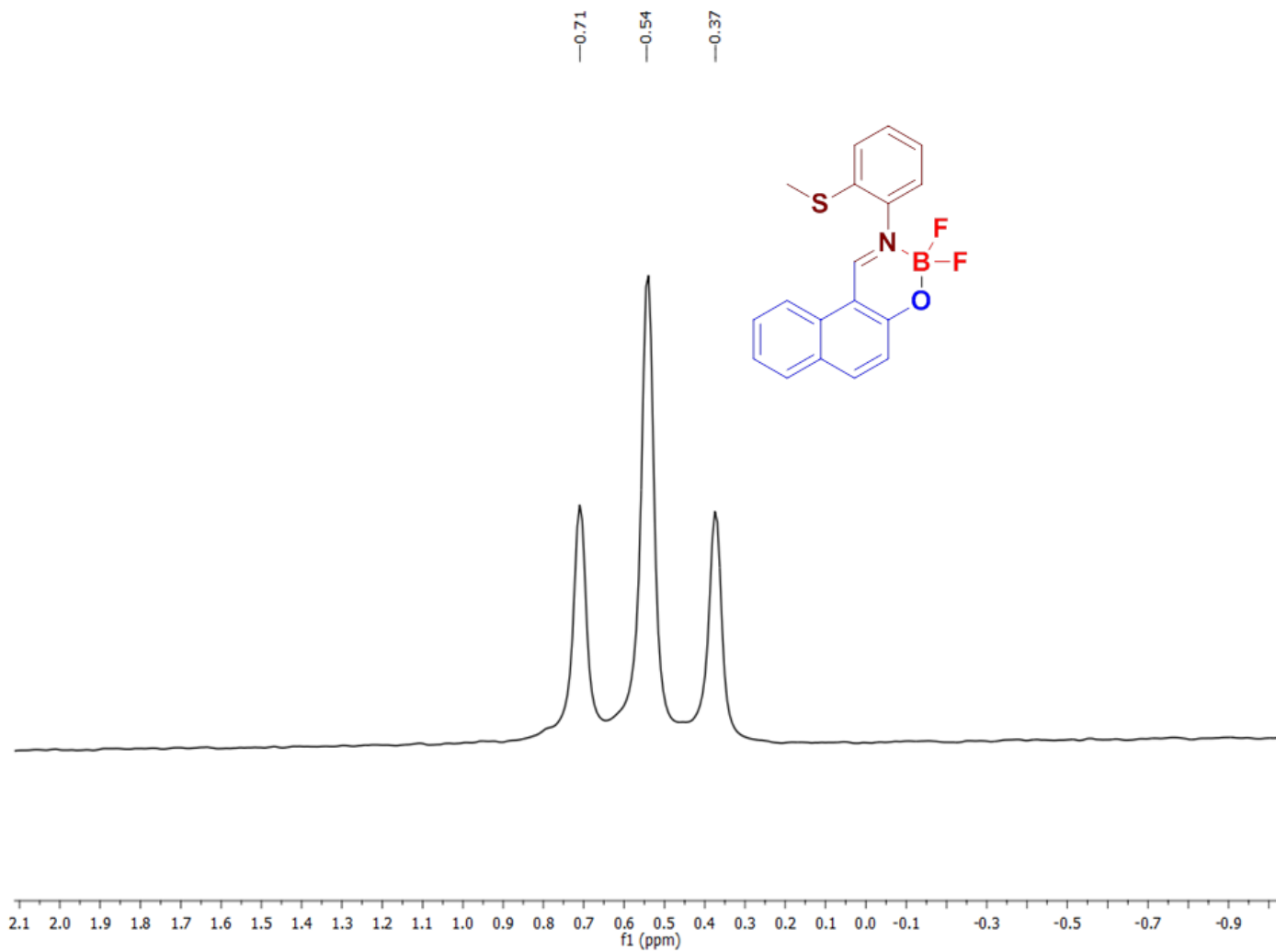


Figure S3.  $^{11}\text{B}$  NMR spectrum of compound **SB-1**

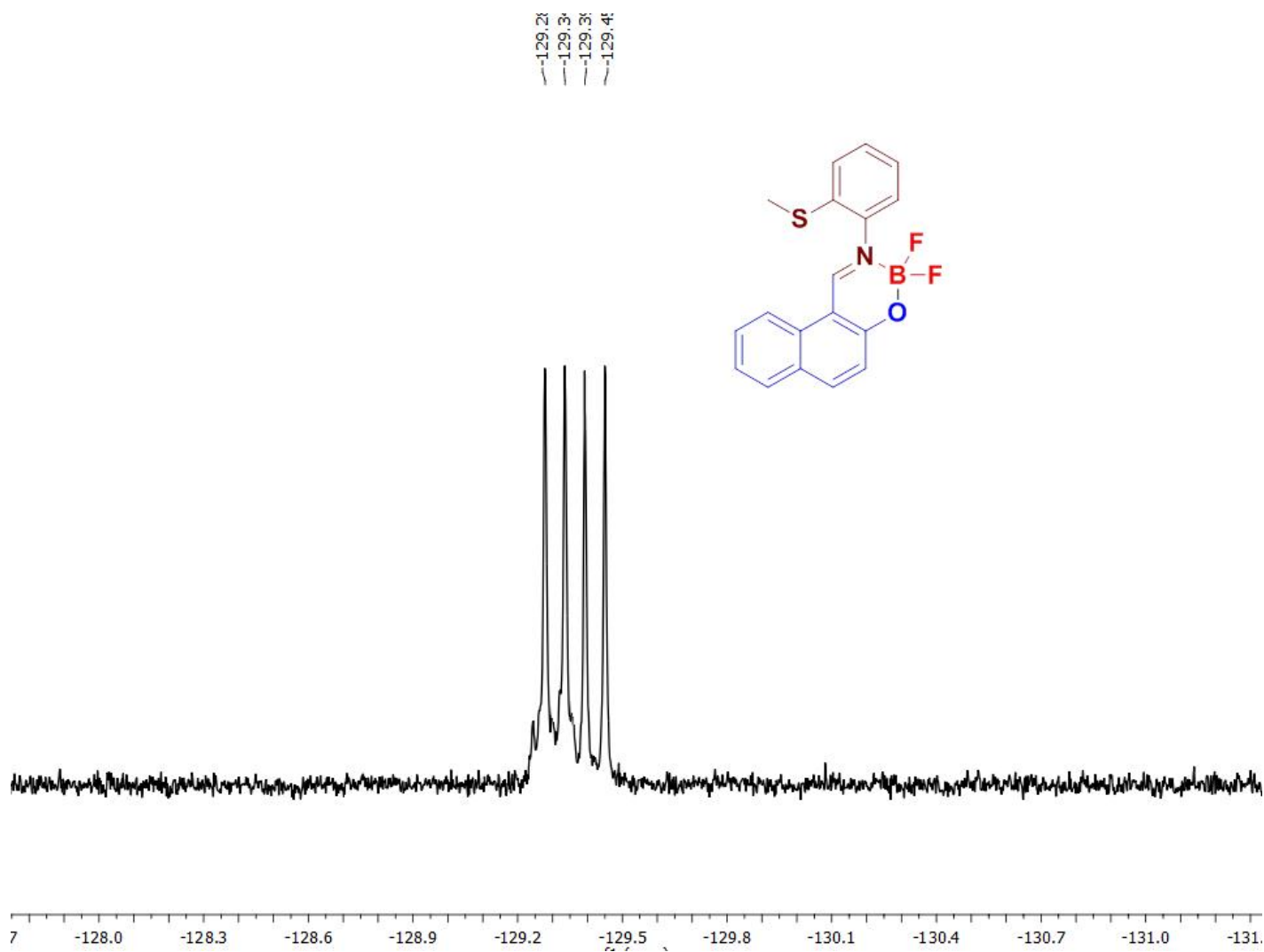


Figure S4.  $^{19}\text{F}$  NMR spectrum of compound **SB-1**

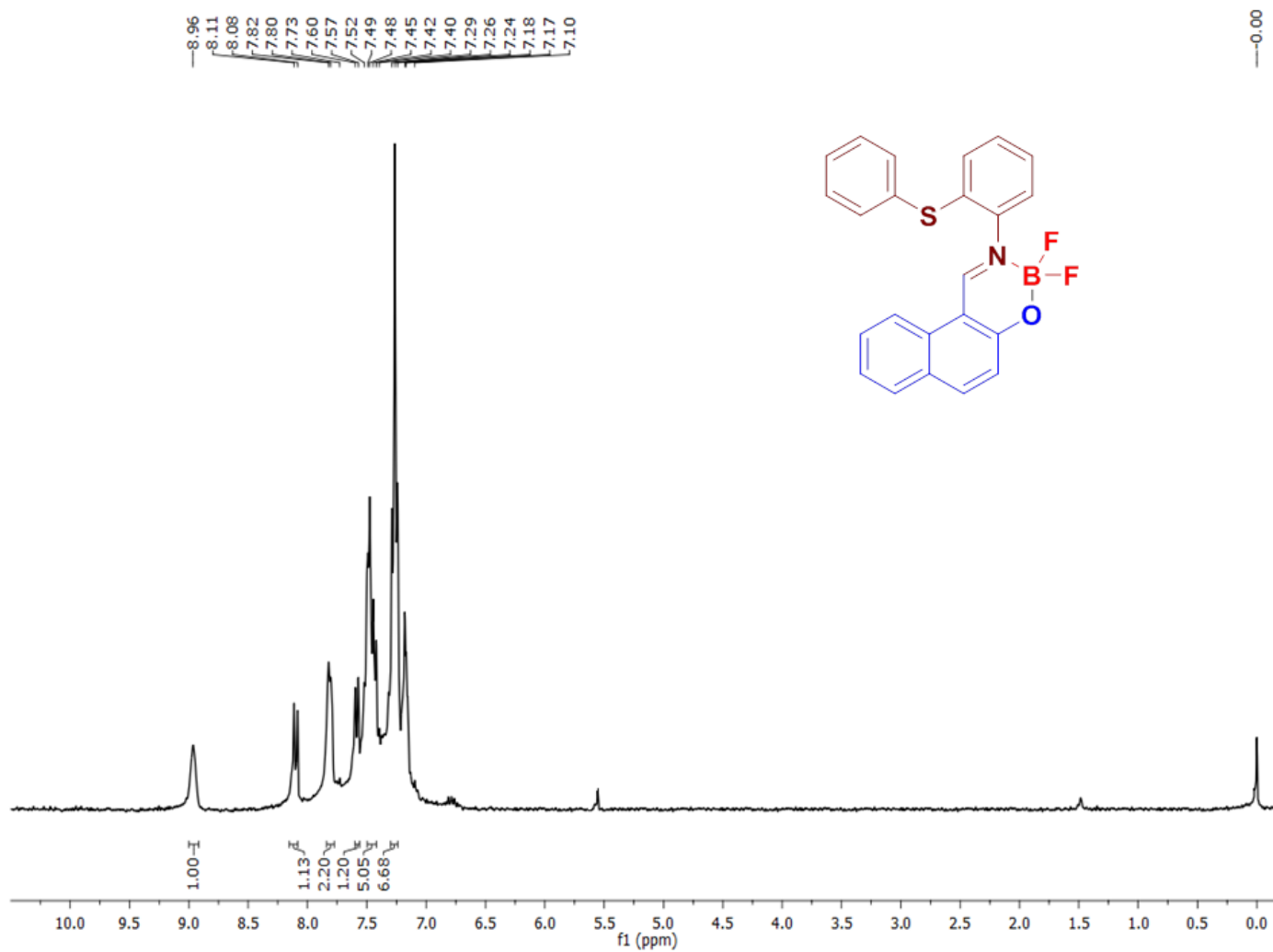


Figure S5. <sup>1</sup>H NMR spectrum of compound **SB-2**

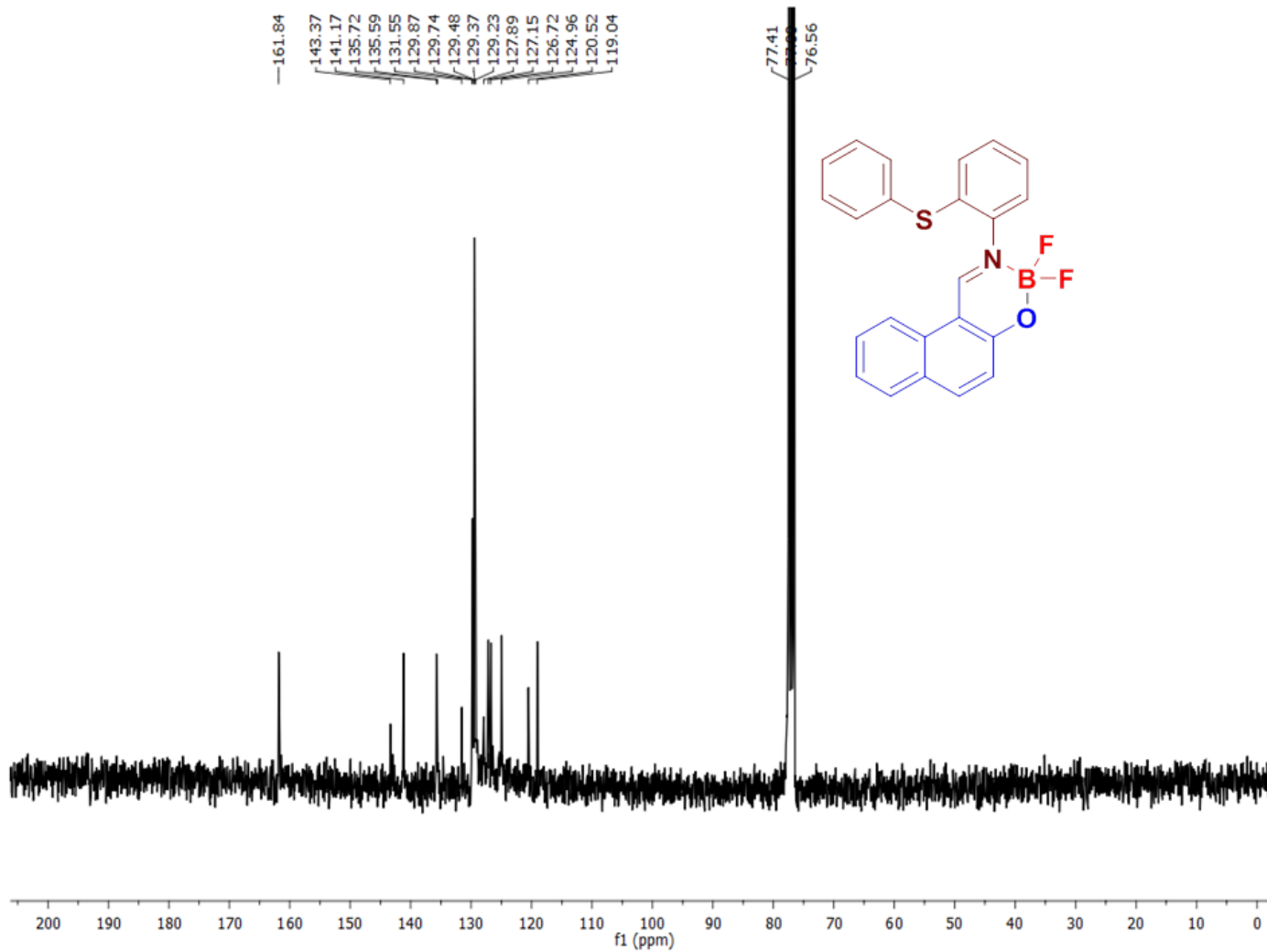


Figure S6.  $^{13}\text{C}$  NMR spectrum of compound **SB-2**



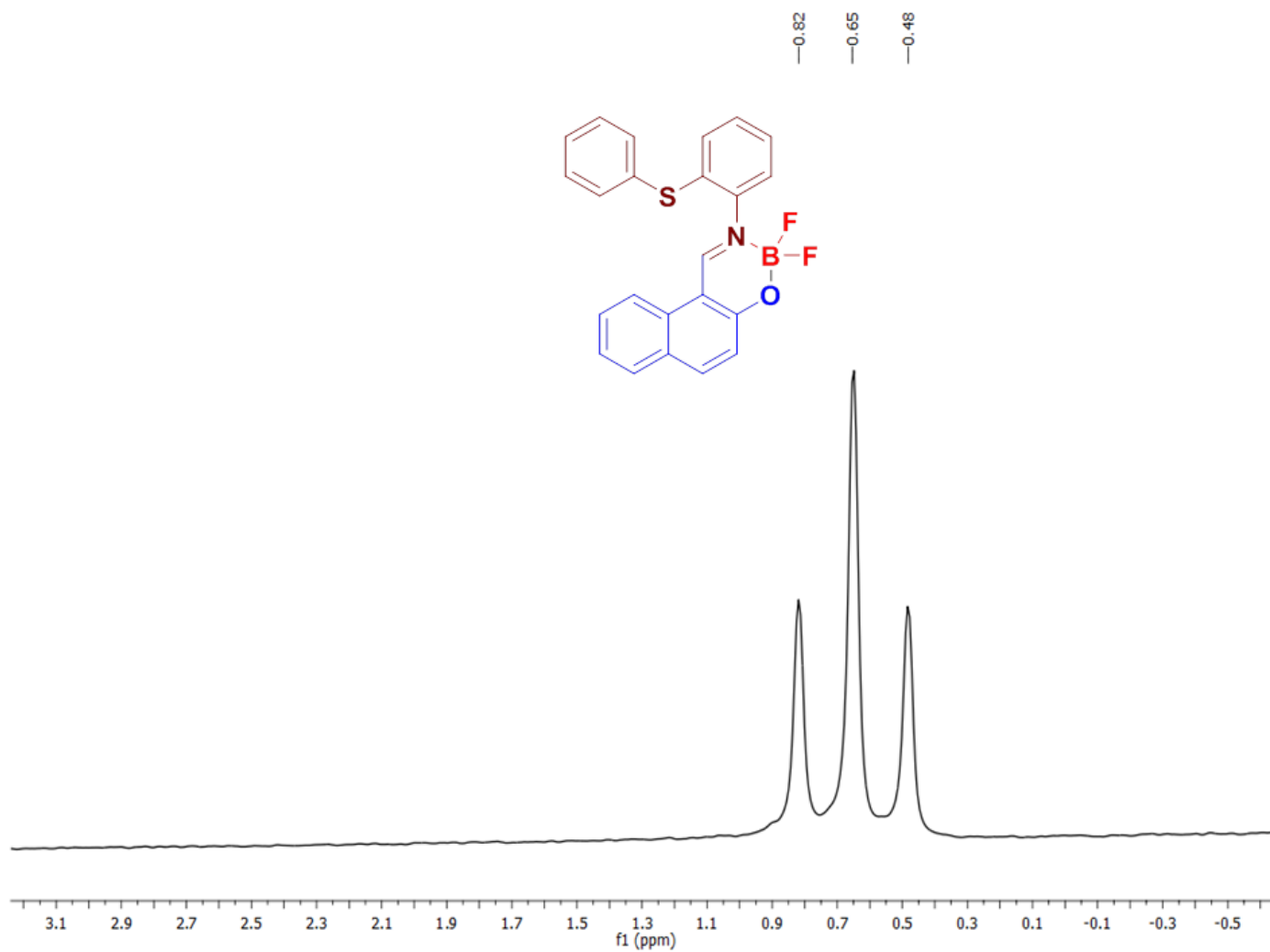


Figure S7.  $^{11}\text{B}$  NMR spectrum of compound **SB-2**

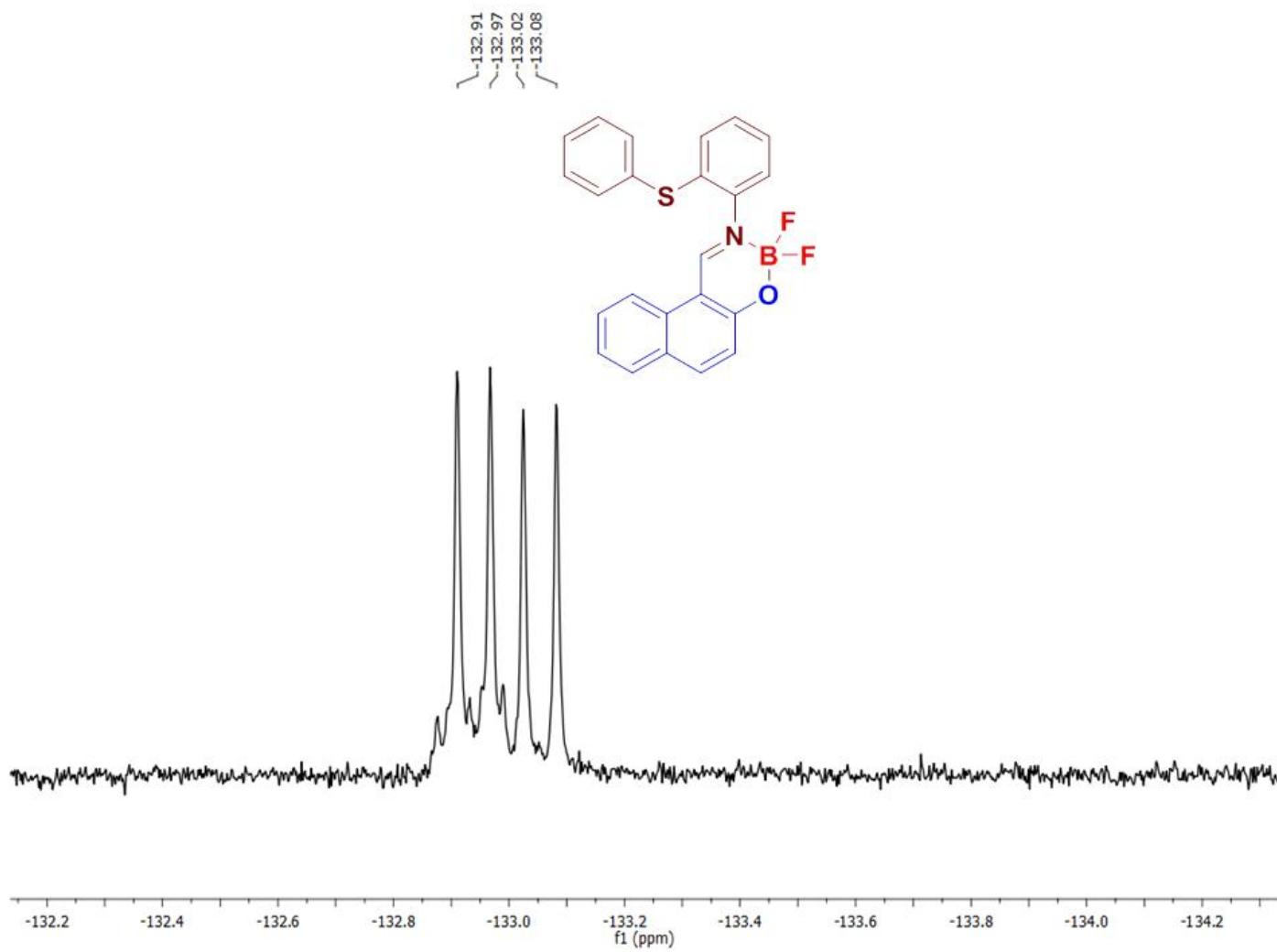


Figure S8.  $^{19}\text{F}$  NMR spectrum of compound **SB-2**

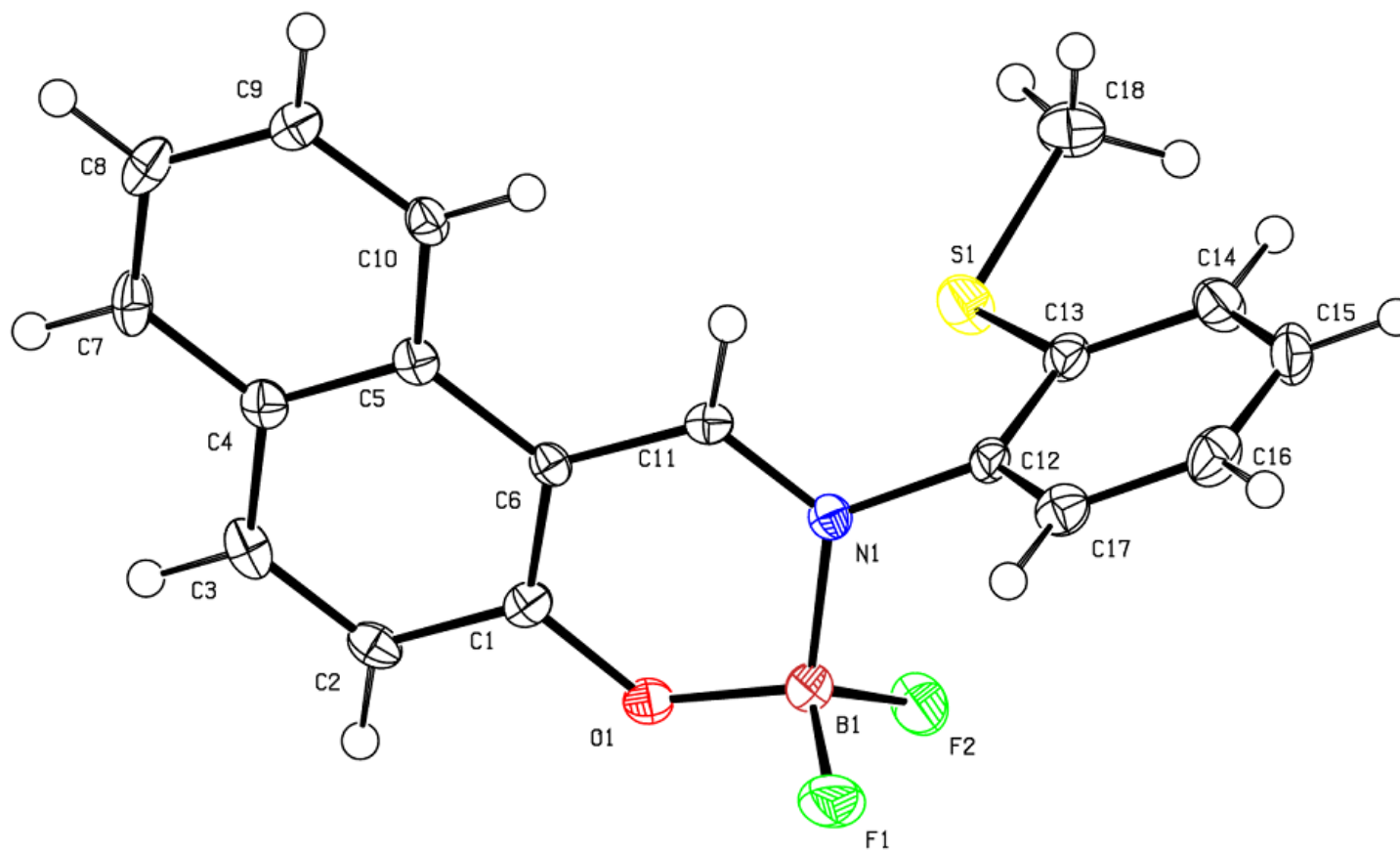
**Table S1. Selected Details about Data Collection and Crystal Refinement for boranil dyes SB-1 and SB-2**

	SB-1	SB-2
CCDC number	1456040	1456041
Empirical formula	C <sub>18</sub> H <sub>14</sub> BF <sub>2</sub> NOS	C <sub>23</sub> H <sub>16</sub> BF <sub>2</sub> NOS
Formula weight	341.17	403.24
Temperature	110.15 K	110.15 K
Wavelength	0.71073 Å	0.71073 Å
Crystal system	Orthorhombic	Monoclinic
Space group	Pbca	P121/c1
Unit cell dimensions	a = 10.661(2) Å; α = 90°. b = 11.234(2) Å; β = 90°. c = 25.712(5) Å; γ = 90°.	a = 15.06(2) Å; α = 90°. b = 11.446(16) Å; β = 101.569(16)°. c = 10.980(16) Å; γ = 90°.
Volume	3079.5(11) Å <sup>3</sup>	1855(5) Å <sup>3</sup>
Z	8	4
Density (calculated)	1.472 Mg/m <sup>3</sup>	1.444 Mg/m <sup>3</sup>
Absorption coefficient	0.236 mm <sup>-1</sup>	0.209 mm <sup>-1</sup>
F(000)	1408	832
Crystal size	0.38 x 0.127 x 0.104 mm <sup>3</sup>	0.522 x 0.148 x 0.074 mm <sup>3</sup>

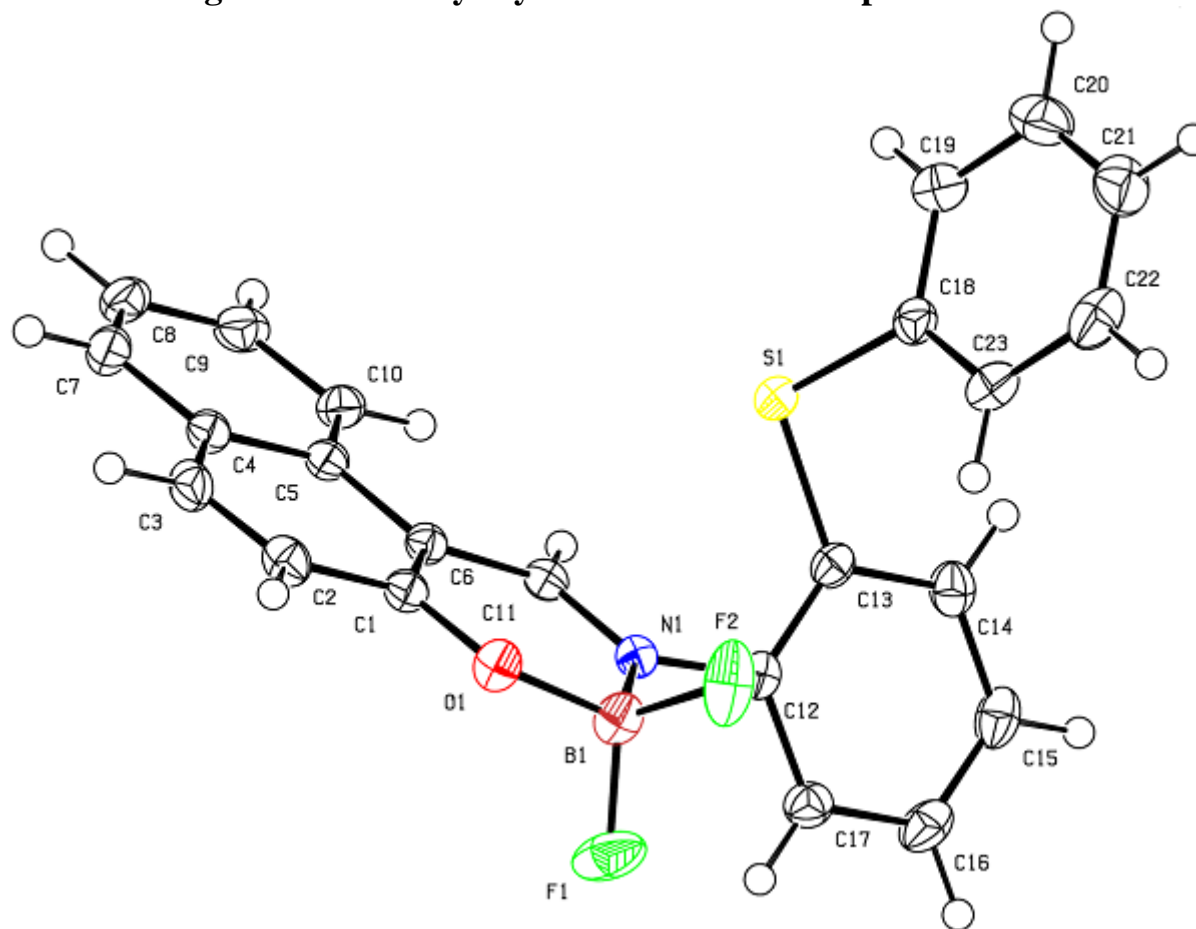
Theta range for data collection	2.482 to 24.998°.	2.252 to 27.550°.
Index ranges	-12<=h<=12, -13<=k<=13, -30<=l<=30	-19<=h<=19, -14<=k<=14, -14<=l<=14
Reflections collected	27275	21099
Independent reflections	2721 [R(int) = 0.0895]	4245 [R(int) = 0.0325]
Completeness to theta = 25.242°	97.8 %	99.9 %
Absorption correction	Semi-empirical from equivalents	Semi-empirical from equivalents
Max. and min. transmission	0.7456 and 0.6639	.7456 and 0.6822
Refinement method	Full-matrix least-squares on F <sup>2</sup>	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	2721 / 0 / 218	4245 / 0 / 262
Goodness-of-fit on F <sup>2</sup>	1.060	1.040
Final R indices [I>2sigma(I)]	R1 = 0.0458, wR2 = 0.0822	R1 = 0.0410, wR2 = 0.0910
R indices (all data)	R1 = 0.0675, wR2 = 0.0915	R1 = 0.0523, wR2 = 0.0972
Largest diff. peak and hole	0.277 and -0.304 e.Å <sup>-3</sup>	0.351 and -0.317 e.Å <sup>-3</sup>

---

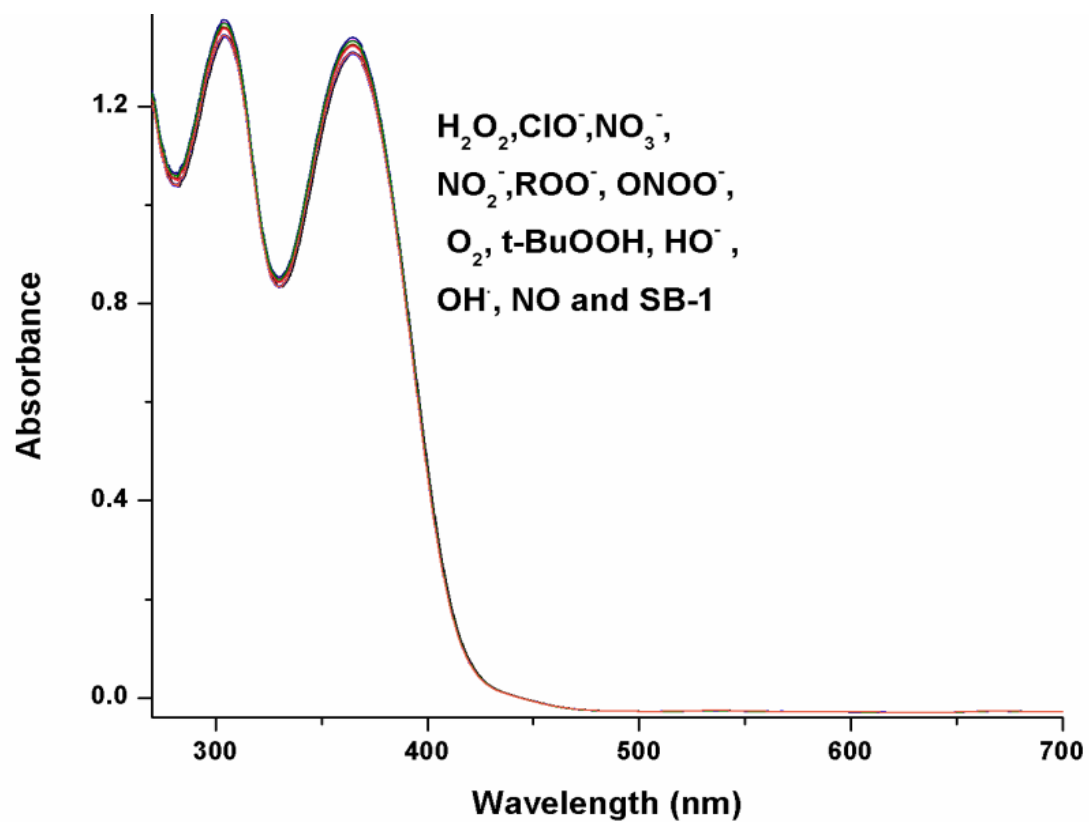
**Figure-S9: X-ray crystal structure of compound SB-1**



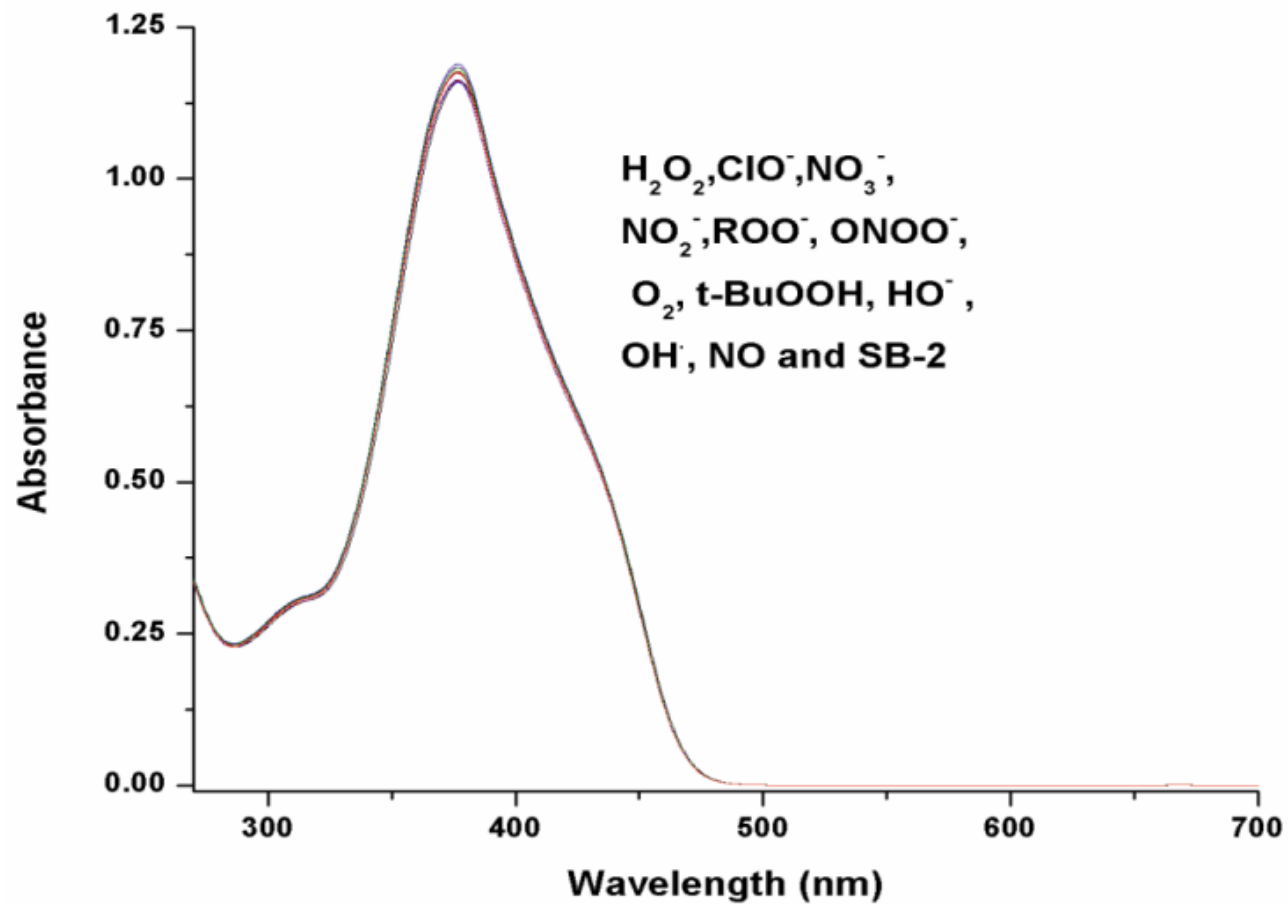
**Figure-S10: X-ray crystal structure of compound SB-2**



**Figure S11.** UV-Vis absorption spectra of **SB-1** (10  $\mu$ M) in 100 mM Phosphate buffer (pH 7.54) in the presence of various RNS and ROS such as NO, H<sub>2</sub>O<sub>2</sub>, ClO<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup>, ROO<sup>-</sup>, ONOO<sup>-</sup>, O<sub>2</sub>, t-BuOOH, ascorbic acid, GSH and HO<sup>-</sup>(1mM) .

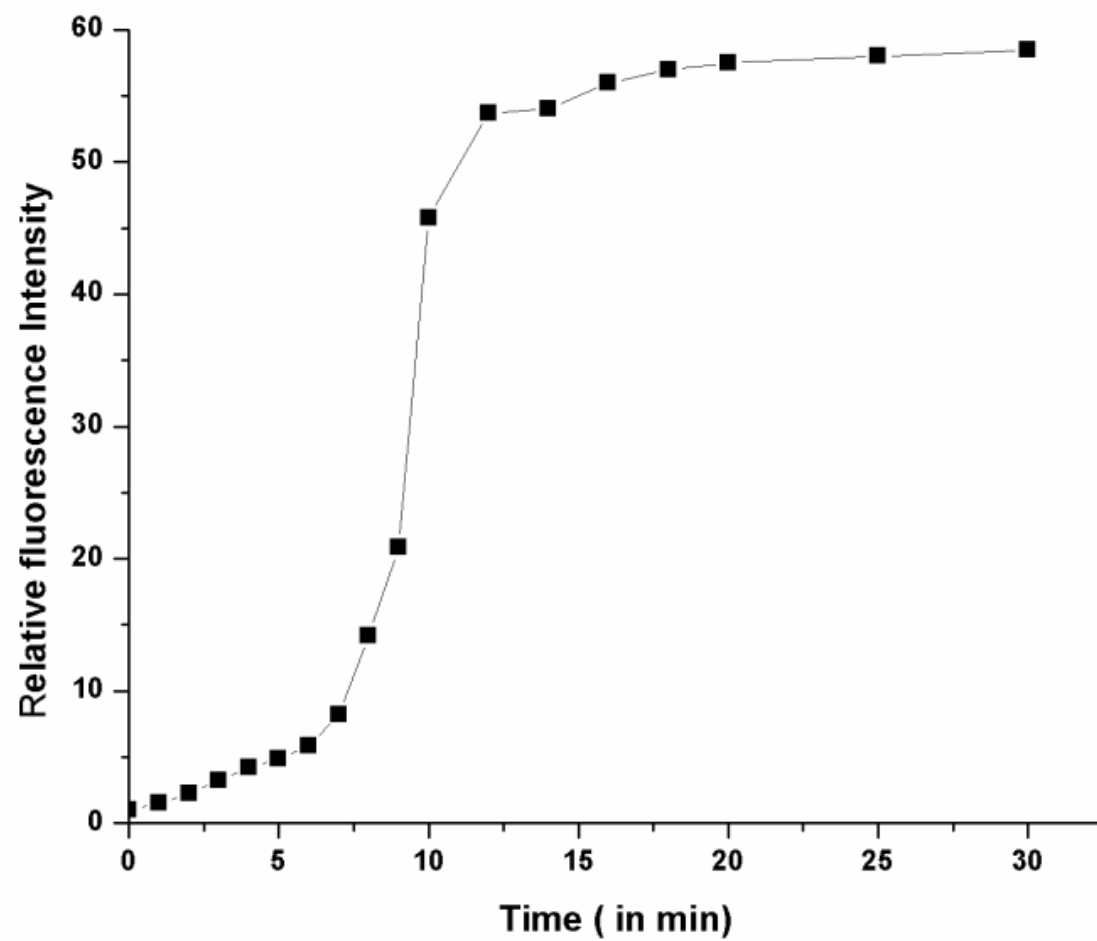


**Figure S12.** UV-Vis absorption spectra of **SB-2** (10  $\mu$ M) in 100 mM phosphate buffer (pH 7.54) in the presence of various RNS and ROS such as NO, H<sub>2</sub>O<sub>2</sub>, ClO<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup>, ROO<sup>-</sup>, ONOO<sup>-</sup>, O<sub>2</sub>, t-BuOOH, ascorbic acid, GSH and HO<sup>-</sup> (1mM) .

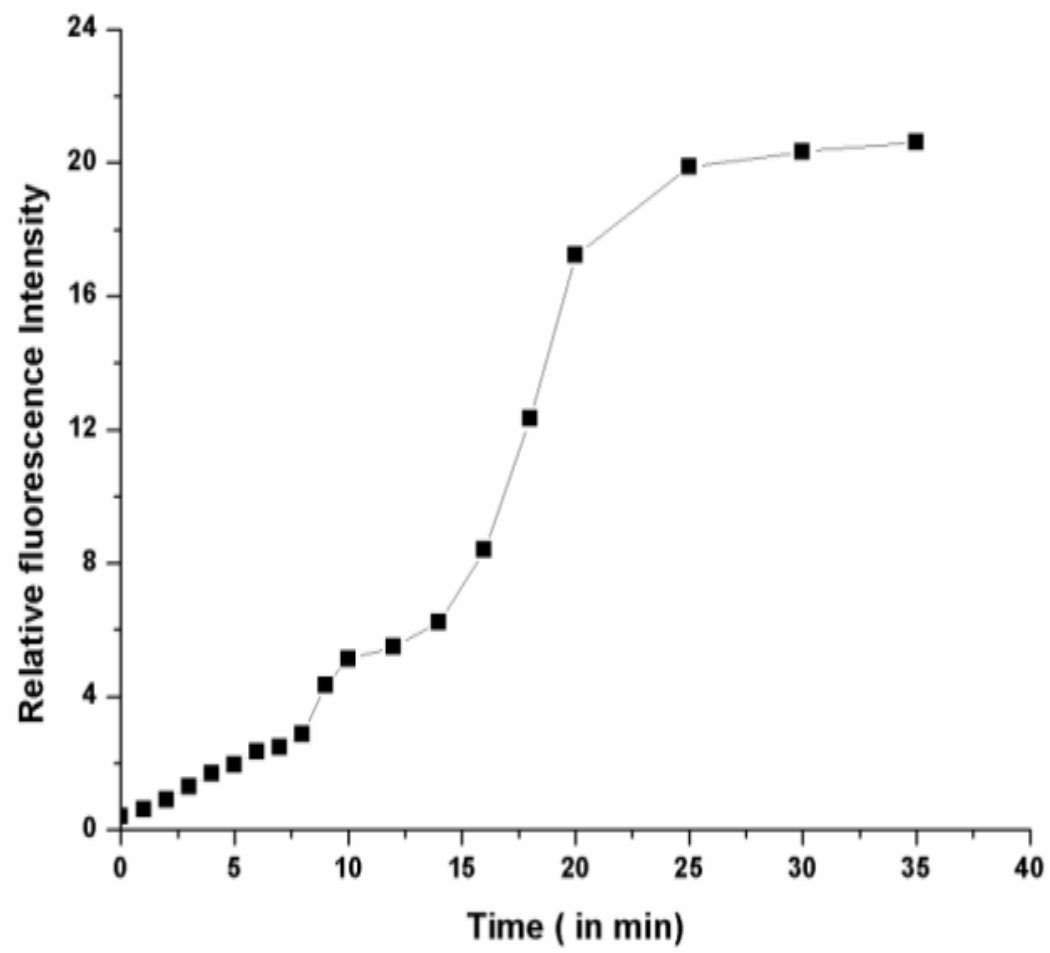




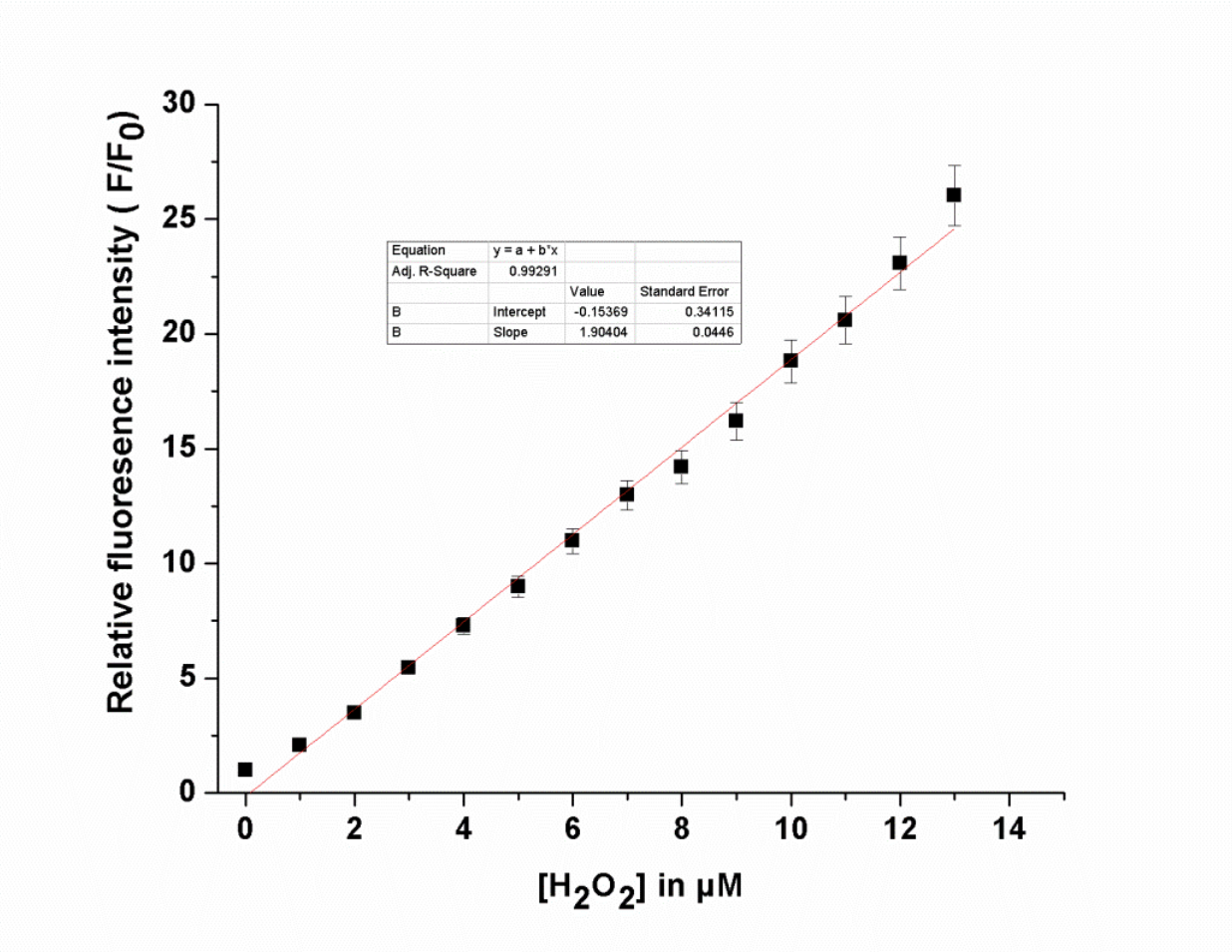
**Figure S13.** Time dependent fluorescence response of **SB-1** with  $\text{H}_2\text{O}_2$ .



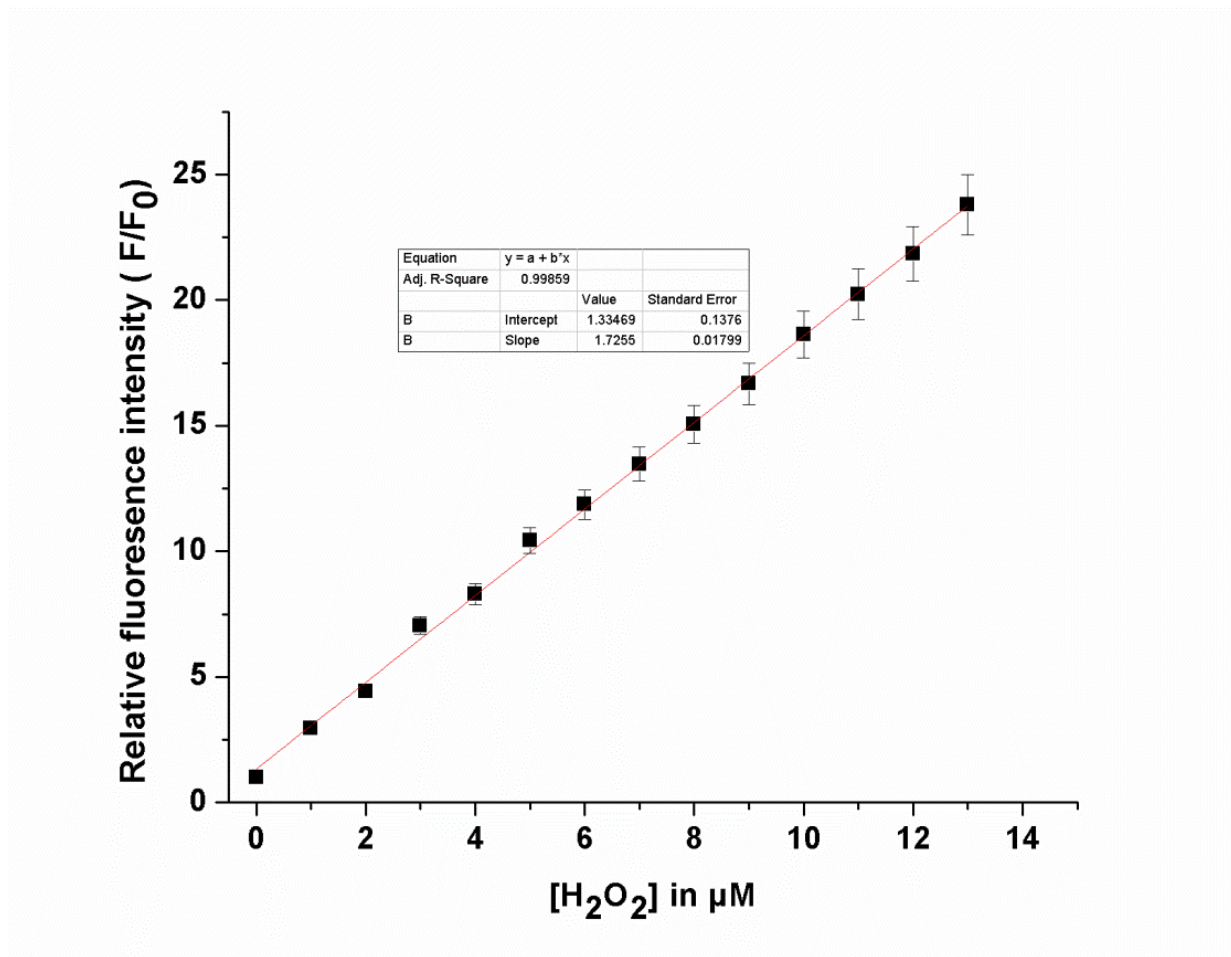
**Figure S14.** Time dependent fluorescence response of **SB-2** with  $\text{H}_2\text{O}_2$ .



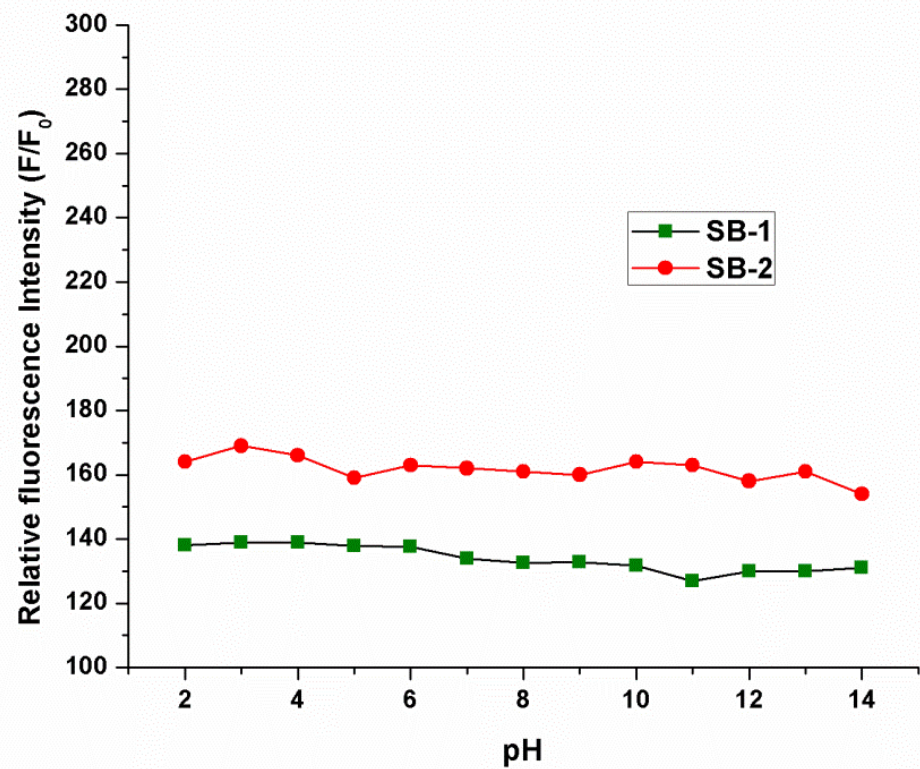
**Figure S15.** Linearity plot of change of fluorescence intensity at  $\lambda_{Emm} = 503$  vs concentration of hydrogen peroxide added to the probe **SB-1**.



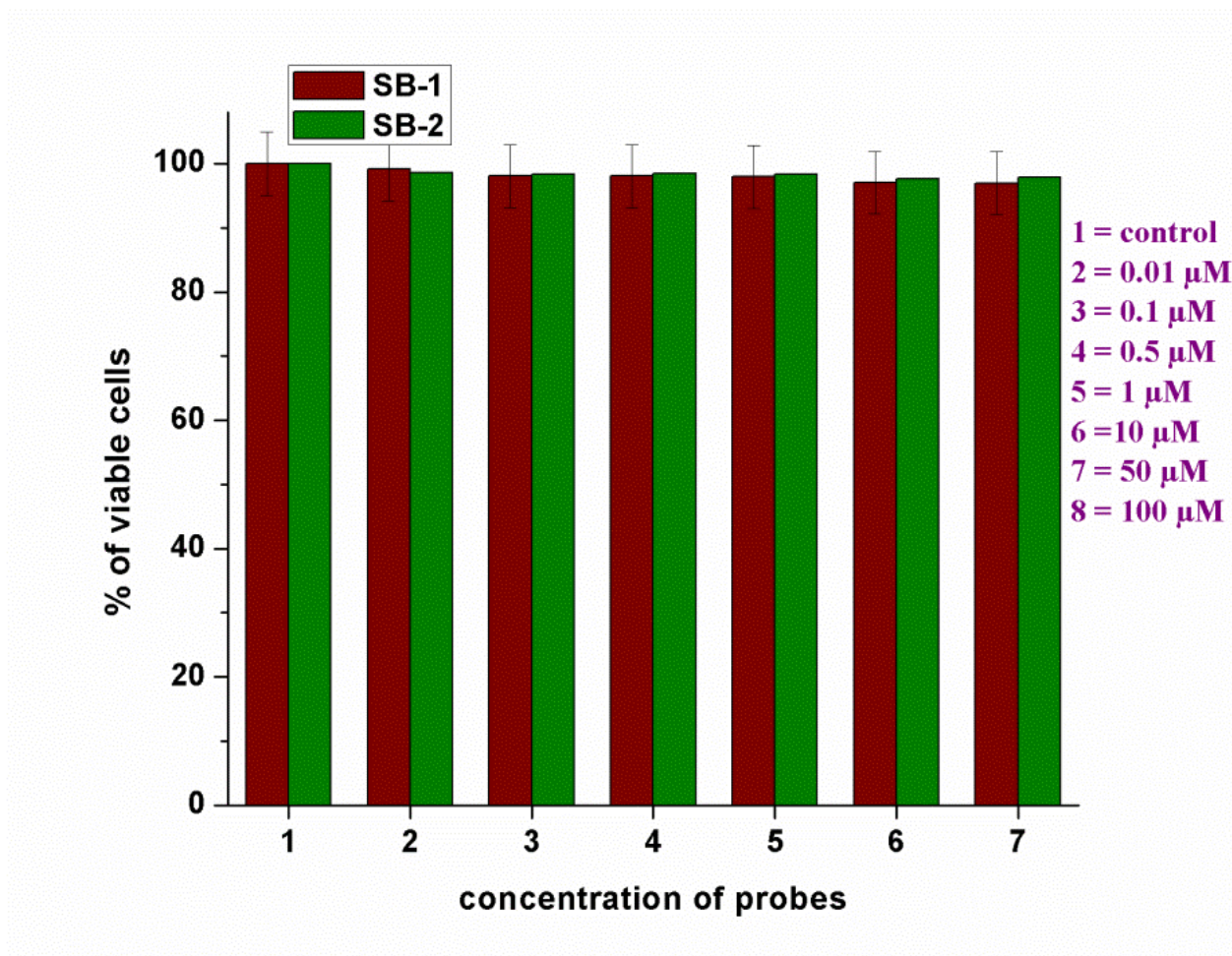
**Figure S16.** Linearity plot of change of fluorescence intensity at  $\lambda_{\text{Emm}} = 510$  vs concentration of Hydrogen peroxide added to the probe **SB-2**.



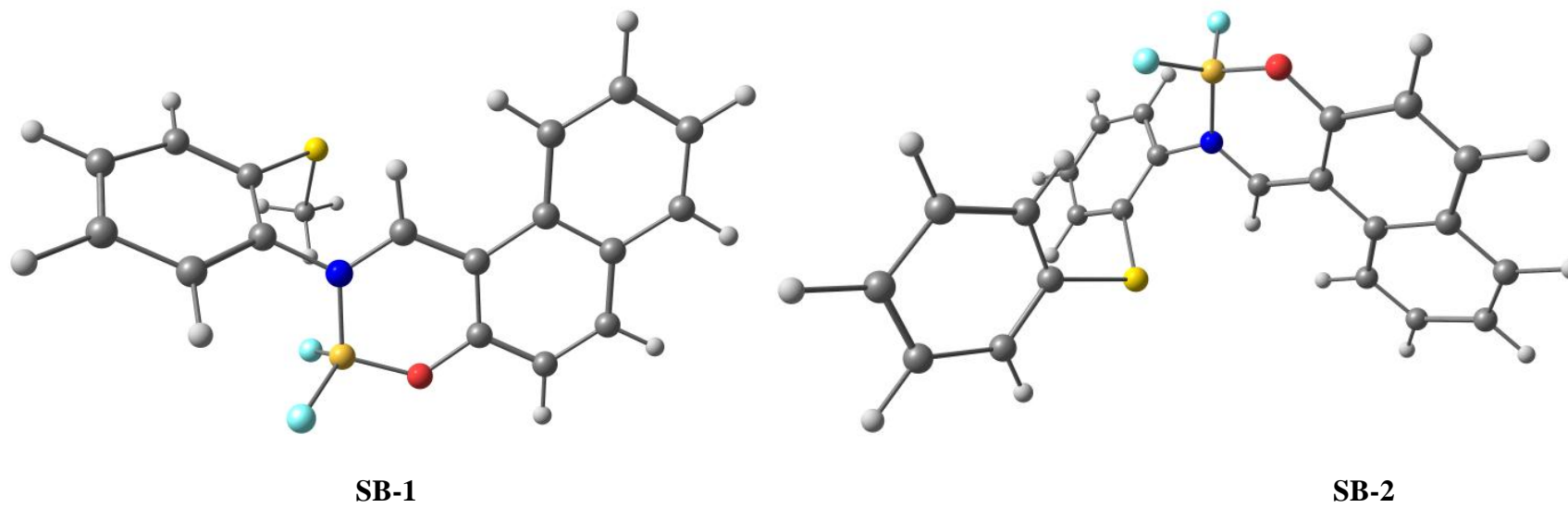
**Figure S17.** Effect of pH on the fluorescence of **SB-1** and SB-2



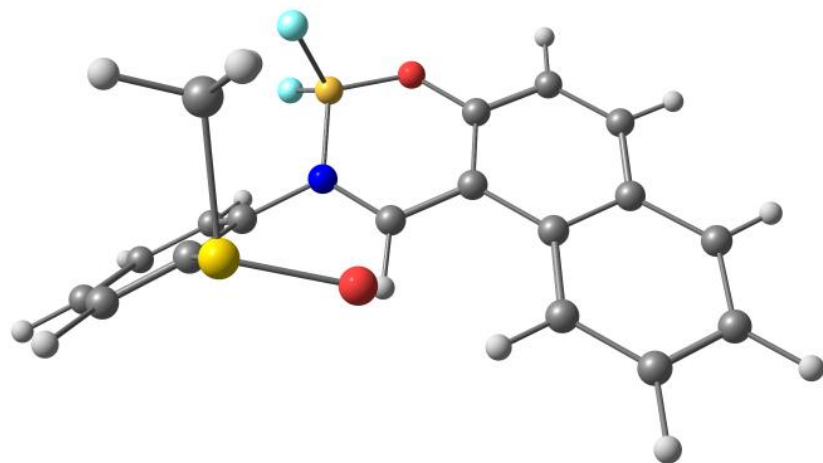
**Figure S18.** Cell viability assay- Plot of % of viable cells vs concentration of **SB-1** and **SB-2**:



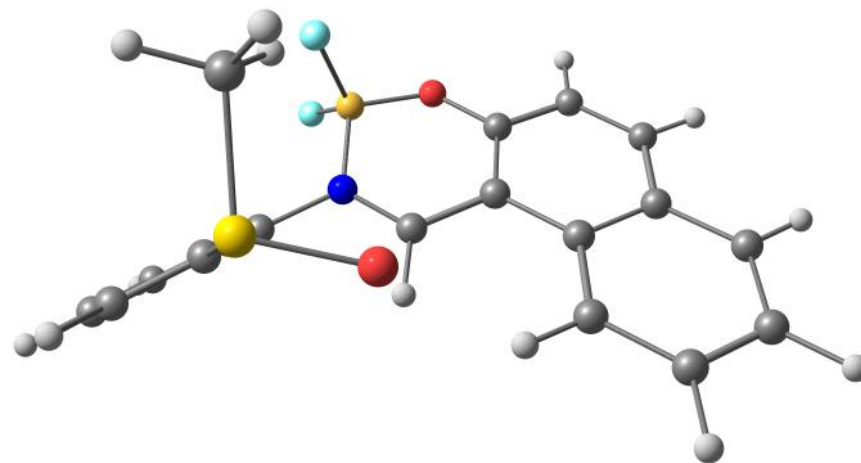
**Figure S19.** Optimized geometries of **SB-1** and **SB-2**



**Figure S20.** Optimized geometries of oxidized forms of **SB-1** and **SB-2**



**Oxidized form of SB-1**



**Oxidized form of SB-2**



**Table S2: The comparison of the present H<sub>2</sub>O<sub>2</sub> detection methods with the existing methods.**

<i>Boron based Probe</i>	<i>Emission Maximum (<math>\lambda</math> in nm)</i>	<i>Turn on/off (fluorescence)</i>	<i>Detection limit</i>	<i>Ref</i>
Peroxy Lucifer	475	ON	Ratiometric	1
Cyaninefluorochrome	715	ON	1 $\mu$ M	2
Mitochondria peroxy yellow 1	540	ON	NA	3
Ratio-Peroxyfluor-1	464	NA	Ratiometric	4
Prototype probe	440	NA	0.1 ~ 5 $\mu$ M	5
MitoBoronic acid	NA	NA	1 $\mu$ M	6
Tetraphenylethylene-Borolane (TPE-BO)	500	ON	0.52 $\mu$ M	7
Naphthalimide fluorophore	528	ON	NA	8
Carbazole-Quinoline cationic based	527	ON	0.04 $\mu$ M	9
<i>meso</i> -(4-pyridinyl)-substituted BODIPY	520	ON	0.1 - 40 $\mu$ M	10
Boranil Dye <b>SB-1/SB-2</b>	503/510	ON	70.27 nM /31.27 nM	Present work

*NA = Not available*

**References:**

1. D. Srikun, E. W. Miller, D. W. Domaille, and C. J. Chang, *J. Am. Chem. Soc.*, 2008, **130**, 4596.
2. N. K. Lifshin, E. Segal, L. Omer, M. Portnoy, R. S. Fainaro, and D. Shabat, *J. Am. Chem. Soc.*, 2011, **133**, 10960.
3. B. C. Dickinson and C. J. Chang, *J. Am. Chem. Soc.* 2008, **130**, 9638.
4. A. E. Albers, V. S. Okreglak and C. J. Chang, *J. Am. Chem. Soc.*, 2006, **128**, 9640.
5. L. -C. Lo and C. -Y Chu, *Chem. Commun.*, 2003, 2728.
6. H. M. Cochemé, A. Logan, T. A. Prime, I. Abakumova, C. Quin, S. J. McQuaker, J. V. Patel, I. M. Fearnley, A. M James, C. M. Porteous, R. A. J. Smith, R. C. Hartley, L. Partridge, and M. P. Murphy, *Nat. Protoc.*, 2012, 7, 946.
7. W. Zhang, W. Liu, P. Li, F. Huang, H. Wang and B. Tang. *Anal. Chem.* 2015, **87**, 9825.
8. D. Kim, G. Kim, S. -J. Nam, J. Yin and J. Yoon, *Sci. Rep.*, 2015, **5**, 8488.
9. J. Xu, Y. Zhang, H. Yu, X. Gao and S. Shao, *Anal. Chem.*, 2016, **88**, 1455.
10. J. Xu, Q. Li, Y. Yue, Y. Guo, S. Shao, *Biosens. Bioelectron.*, 2014, **56**, 58.