

Silver comes into play: Henry reaction and domino cycloisomerisation sequence catalysed by [Ag(I)(Pc-L)] complexes

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General experimental details. All of the reactions that involved the use of reagents sensitive to oxygen or hydrolysis were carried out under an inert atmosphere. The glassware was previously dried in an oven at 110 °C and was set with cycles of vacuum and nitrogen. Also syringes, used to transfer reagents and solvents, were previously set under a nitrogen atmosphere. The syntheses of the silver complexes were carried out under a nitrogen atmosphere by employing standard Schlenk techniques. All chemicals and solvents were commercially available and were used after distillation or treatment with drying agents. The chromatographic column separations were performed by a flash technique, using silica gel (pore size 60 Å, particle size 230–400 mesh, Merck grade 9385). For TLC, silica was used on TLC Alu foils with fluorescent indicator (254 nm) and the detection was performed by irradiation with UV light ($\lambda = 254$ nm or 366 nm). ^1H NMR analyses were performed with 200, 300, 400 or 600 MHz spectrometers at room temperature. The coupling constants (J) are expressed in hertz (Hz), and the chemical shifts (δ) in ppm. ^{13}C NMR analyses were performed with the same instruments at 75.5, 100 or 150 MHz, and attached proton test (APT) sequence was used to distinguish the methine and methyl carbon signals from those arising from methylene and quaternary carbon atoms. All ^{13}C NMR spectra were recorded with complete proton decoupling. The ^1H NMR signals of the ligand described in the following have been attributed by correlation spectroscopy (COSY) and nuclear Overhauser effect spectroscopy (NOESY) techniques. Assignments of the resonance in ^{13}C NMR were made using the APT pulse sequence and heteronuclear single quantum correlation (HSQC) and heteronuclear multiple bond correlation (HMBC) techniques. Low resolution MS spectra were recorded with instruments equipped with electron ionization (EI), ESI/ion trap (using a syringe pump device to directly inject sample solutions), or fast atom bombardment (FAB) (for Pc-L and metal complexes) sources. The values are expressed as mass–charge ratio and the relative intensities of the most significant peaks are shown in brackets. UV–vis spectra of the ligand and its silver complexes were recorded in CH_2Cl_2 . Elemental analyses were recorded in the analytical laboratories of Università degli Studi di Milano. Optical rotations were measured on a Perkin Elmer instruments model 343 plus; $[\alpha]_{\text{D}}$ values are given in 10^{-1} deg cm^2 g^{-1} . Silver complex **1**¹, *N*-tosyl-aziridine and copper complex **2**² and 2,6-pyridinedimethanol 2,6-dimesylate³ were synthesized as previously reported.

Henry reaction. Yields and ^1H spectra of products **4** and **9**

2-nitro-1-(4-nitrophenyl)ethanol **4a**. Yield 90% (62 mg); ^1H NMR (400 MHz, CDCl_3 , δ) 8.27 (d, $J = 8.6$ Hz, 2H, H_{Ar}), 7.63 (d, $J = 8.5$ Hz, 2H, H_{Ar}), 5.60 (dd, $J = 7.7, 3.4$ Hz, 1H, CH), 4.68 – 4.40 (m, 2H, CH_2), 3.12 (bs, 1H, OH). Spectral data are consistent with literature values.⁴

4-(1-hydroxy-2-nitroethyl)benzotrile **4b**. Yield 80% (49 mg); ^1H NMR (300 MHz, CDCl_3 , δ) 7.71 (d, $J = 8.4$ Hz, 2H, H_{Ar}), 7.56 (d, $J = 8.1$ Hz, 2H, H_{Ar}), 5.55 (dd, $J = 7.9, 3.8$ Hz, 1H, CH), 4.68 – 4.43 (m, 2H, CH_2), 3.11 (bs, 1H, OH). Spectral data are consistent with literature values.⁴

2-nitro-1-(4-(trifluoromethyl)phenyl)ethanol **4c**. Yield 92% (69 mg); ^1H NMR (300 MHz, CDCl_3 , δ) 7.68 (d, $J = 8.3$ Hz, 2H, H_{Ar}), 7.56 (d, $J = 8.2$ Hz, 2H, H_{Ar}), 5.56 (d, $J = 6.6$ Hz, 1H, CH), 4.70 – 4.45 (m, 2H, CH_2), 2.96 (bs, 1H, OH). Spectral data are consistent with literature values.⁵

1-(4-bromophenyl)-2-nitroethanol **4d**. Yield 82% (65 mg); ^1H NMR (300 MHz, CDCl_3 , δ) 7.54 (d, $J = 8.4$ Hz, 2H, H_{Ar}), 7.29 (d, $J = 8.2$ Hz, 2H, H_{Ar}), 5.44 (dd, $J = 9.2, 3.3$ Hz, 1H, CH), 4.61 – 4.46 (m, 2H, CH_2), 2.88 (bs, 1H, OH). Spectral data are consistent with literature values.⁴

1-(4-chlorophenyl)-2-nitroethanol **4e**. Yield 80% (52 mg); ^1H NMR (600 MHz, CDCl_3 , δ) 7.38 (d, $J = 8.6$ Hz, 2H, H_{Ar}), 7.34 (d, $J = 8.5$ Hz, 2H, H_{Ar}), 5.44 (d, $J = 9.5$ Hz, 1H, CH), 4.57 (dd, $J = 13.3, 9.5$ Hz, 1H, CH_2), 4.49 (dd, $J = 13.3, 3.1$ Hz, 1H, CH_2), 3.01 (d, $J = 3.6$ Hz, 1H, OH). Spectral data are consistent with literature values.⁴

1-(4-fluorophenyl)-2-nitroethanol **4f**. Yield 64% (38 mg); ^1H NMR (300 MHz, CDCl_3 , δ) 7.39 (dd, $J = 8.4, 5.2$ Hz, 2H, H_{Ar}), 7.10 (pst, $J = 8.6$ Hz, 2H, H_{Ar}), 5.46 (d, $J = 9.1$ Hz, 1H, CH), 4.63 – 4.47 (m, 2H, CH_2), 2.82 (bs, 1H, OH). Spectral data are consistent with literature values.⁴

2-nitro-1-phenylethanol **4g**. Yield 55% (29 mg); ^1H NMR (300 MHz, CDCl_3 , δ) 7.42 – 7.29 (m, 5H, H_{Ar}), 5.48 (dd, $J = 9.3, 3.2$ Hz, 1H, CH), 4.66 – 4.50 (m, 2H, CH_2), 1.56 (bs, 1H, OH exchanging with water). Spectral data are consistent with literature values.⁴

2-nitro-1-(perfluorophenyl)ethanol **4i**. Yield 68% (56 mg); ^1H NMR (300 MHz, CDCl_3 , δ) 5.87 (ddd, $J = 9.2, 5.8, 3.5$ Hz, 1H, CH), 5.02 (dd, $J = 13.9, 9.4$ Hz, 1H, CH_2), 4.61 (dd, $J = 14.0, 3.4$ Hz, 1H, CH_2), 3.08 (d, $J = 6.0$ Hz, 1H, OH). Spectral data are consistent with literature values.⁶

2-nitro-1-(2-nitrophenyl)ethanol **4k**. Yield 95% (65 mg); ^1H NMR (300 MHz, CDCl_3 , δ) 8.07 (d, $J = 8.2$ Hz, 1H, H_{Ar}), 7.95 (d, $J = 7.9$ Hz, 1H, H_{Ar}), 7.75 (pst, $J = 7.6$ Hz, 1H, H_{Ar}), 7.56 (pst, $J = 7.8$ Hz, 1H, H_{Ar}), 6.05 (dd, $J = 9.0, 2.0$ Hz, 1H, CH), 4.87 (dd, $J = 13.8, 2.3$ Hz, 1H, CH_2), 4.55 (dd, $J = 13.8, 9.1$ Hz, 1H, CH_2), 3.23 (bs, 1H, OH). Spectral data are consistent with literature values.⁴

1-(2-bromophenyl)-2-nitroethanol **4l**. Yield 65% (51 mg); ¹H NMR (300 MHz, CDCl₃, δ) 7.67 (d, *J* = 7.9 Hz, 1H, H_{Ar}), 7.57 (d, *J* = 8.0 Hz, 1H, H_{Ar}), 7.41 (pst, *J* = 7.1 Hz, 1H, H_{Ar}), 7.22 (pst, *J* = 7.7 Hz, 1H, H_{Ar}), 5.96 – 5.70 (m, 1H, CH), 4.70 (dd, *J* = 13.7, 2.3 Hz, 1H, CH₂), 4.44 (dd, *J* = 13.7, 9.6 Hz, 1H, CH₂), 2.94 (d, *J* = 4.0 Hz, 1H, OH). Spectral data are consistent with literature values.⁴

1-(2,6-dichlorophenyl)-2-nitroethanol **4m**. Yield 68% (51 mg); ¹H NMR (300 MHz, CDCl₃, δ) 7.37 (m, 2H, H_{Ar}), 7.32 – 7.20 (m, 2H, H_{Ar}), 6.38 – 6.21 (m, 1H, CH), 5.17 (dd, *J* = 13.1, 10.1 Hz, 1H, CH₂), 4.56 (dd, *J* = 13.1, 3.4 Hz, 1H, CH₂), 3.24 (d, *J* = 8.3 Hz, 1H, OH). Spectral data are consistent with literature values.⁵

1-(furan-2-yl)-2-nitroethanol **4o**. Yield 62% (31 mg); ¹H NMR (400 MHz, CDCl₃, δ) 7.40 (s, 1H, H_{Ar}), 6.55 – 6.23 (m, 2H, H_{Ar}), 5.51 – 5.38 (m, 1H, CH), 4.76 (dd, *J* = 13.4, 9.0 Hz, 1H, CH₂), 4.65 (dd, *J* = 13.4, 3.6 Hz, 1H, CH₂), 3.18 (bs, 1H, OH). Spectral data are consistent with literature values.⁴

2-nitro-1-(4-nitrophenyl)propan-1-ol **4q**. Yield 90% (65 mg); Diastereomeric ratio (*syn/anti* 60:40) determined by ¹H NMR. *Syn* isomer: ¹H NMR (400 MHz, CDCl₃, δ) 8.25 – 8.21 (m, 2H, H_{Ar}), 7.60 – 7.56 (m, 2H, H_{Ar}), 5.18 (d, *J* = 6.6 Hz, 1H, CH), 4.79 – 4.72 (m, 1H, CH), 3.15 (bs, 1H, OH), 1.37 (d, *J* = 6.6 Hz, 3H, CH₃). *Anti* isomer: ¹H NMR (400 MHz, CDCl₃, δ) 8.25 – 8.21 (m, 2H, H_{Ar}), 7.60 – 7.56 (m, 2H, H_{Ar}), 5.55 (m, 1H, CH), 4.79 – 4.72 (m, 1H, CH), 3.15 (bs, 1H, OH), 1.48 (d, *J* = 6.6 Hz, 3H, CH₃). Spectral data are consistent with literature values.⁴

2-nitro-1-(2-nitrophenyl)propan-1-ol **4r**. Yield 93% (67 mg); Diastereomeric ratio (*syn/anti* 45:55) determined by ¹H NMR. *Syn* isomer: ¹H NMR (400 MHz, CDCl₃, δ) 8.00 (dd, *J* = 8.2, 0.9 Hz, 1H, H_{Ar}), 7.73 – 7.69 (m, 2H, H_{Ar}), 7.56 – 7.53 (m, 1H, H_{Ar}), 5.71 (d, *J* = 6.6 Hz, 1H, CH), 5.01 – 4.95 (m, 1H, CH), 3.34 (bs, 1H, OH), 1.56 – 1.51 (m, 3H, CH₃). *Anti* isomer: ¹H NMR (400 MHz, CDCl₃, δ) 8.08 (dd, *J* = 8.2, 1.4 Hz, 1H, H_{Ar}), 7.93 (d, *J* = 7.9 Hz, 1H, H_{Ar}), 7.73 – 7.69 (m, 1H, H_{Ar}), 7.56 – 7.53 (m, 1H, H_{Ar}), 6.09 (d, *J* = 2.6 Hz, 1H, CH), 4.79 – 4.72 (m, 1H, CH), 3.15 (bs, 1H, OH), 1.56 – 1.51 (d, *J* = 6.0 Hz, 3H, CH₃). Spectral data are consistent with literature values.⁴

2-nitro-1-(4-(trifluoromethyl)phenyl)propan-1-ol **4s**. Yield 71% (89 mg); Diastereomeric ratio (*syn/anti* 51:49) determined by ¹H NMR. *Syn* isomer: ¹H NMR (600 MHz, CDCl₃, δ) 7.69 – 7.66 (m, 2H, H_{Ar}), 7.54 – 7.51 (m, 2H, H_{Ar}), 5.13 (d, *J* = 8.6 Hz, 1H, CH), 4.76 (dq, *J* = 8.6, 6.9 Hz, 1H, CH), 1.37 (d, *J* = 6.9 Hz, 3H, CH₃). *Anti* isomer: ¹H NMR (600 MHz, CDCl₃, δ) 7.69 – 7.66 (m, 2H, H_{Ar}), 7.54 – 7.51 (m, 2H, H_{Ar}), 5.50 (d, *J* = 3.4 Hz, 1H, CH), 4.70 (qd, *J* = 6.8, 3.4 Hz, 1H, CH), 1.50 (d, *J* = 6.8 Hz, 3H, CH₃). OH signal appeared as a very broad signal around 3.5 – 1.8 ppm. Spectral data are consistent with literature values.⁷

2-nitro-1-(4-chlorophenyl)propan-1-ol **4t**. Yield 75% (52 mg); Diastereomeric ratio (*syn/anti* 61:39) determined by ¹H NMR. *Syn* isomer: ¹H NMR (300 MHz, CDCl₃, δ) 7.39 – 7.35 (m, 2H, H_{Ar}), 7.33 – 7.26 (m, 2H, H_{Ar}), 5.02 (d, *J* = 8.9 Hz, 1H, CH), 4.76 – 4.70 (m, 1H, CH), 2.67 (br, 1H, OH), 1.33 (dd, *J* = 6.8, 1.6 Hz, 3H, CH₃). *Anti* isomer: ¹H NMR (300 MHz, CDCl₃, δ) 7.39 – 7.35 (m, 2H, H_{Ar}), 7.33 – 7.26 (m, 2H, H_{Ar}), 5.37 (br, 1H, CH), 4.68 – 4.62 (m, 1H, CH), 2.77 (br, 1H, OH), 1.49 (dd, *J* = 6.8, 1.5 Hz, 3H, CH₃). Spectral data are consistent with literature values.⁸

2-nitro-1-phenylpropan-1-ol **4u**. Yield 60% (35 mg); Diastereomeric ratio (*syn/anti* 55:45) determined by ¹H NMR. *Syn* isomer: ¹H NMR (400 MHz, CDCl₃, δ) 7.41 – 7.36 (m, 5H, H_{Ar}), 5.03 (dd, *J* = 9.0, 3.0 Hz, 1H, CH), 4.77 (dq, *J* = 9.0, 6.8 Hz, 1H, CH), 2.58 (d, *J* = 3.0 Hz, 1H, OH), 1.32 (d, *J* = 6.8 Hz, 3H, CH₃). *Anti* isomer: ¹H NMR (300 MHz, CDCl₃, δ) 7.41 – 7.36 (m, 5H, H_{Ar}), 5.40 (pst, *J* = 3.5 Hz, 1H, CH), 4.70 (qd, *J* = 6.8, 3.6 Hz, 1H, CH), 2.70 (d, *J* = 3.5 Hz, 1H, OH), 1.51 (d, *J* = 6.8 Hz, 3H, CH₃). Spectral data are consistent with literature values.⁸

3-hydroxy-3-(nitromethyl)indolin-2-one **9a**. Yield 80% (53 mg); ¹H NMR (300 MHz, DMSO-d₆, δ) 10.54 (s, 1H, NH), 7.40 (dd, *J* = 7.3, 1.2 Hz, 1H, H_{Ar}), 7.27 (m, 1H, H_{Ar}), 7.10 – 6.82 (m, 2H, H_{Ar}), 6.74 (d, *J* = 1.0 Hz, 1H, OH), 5.15 – 4.82 (m, 2H, CH₂). Spectral data are consistent with literature values.⁹

3-hydroxy-3-(nitroethyl)indolin-2-one **9b**. The product was not isolated; the yield and the diastereoisomeric ratio (6:4) were determined by ¹H NMR (internal standard 2,4-dinitrotoluene). Major isomer: ¹H NMR (300 MHz, DMSO-d₆, δ) 8.12 – 6.75 (m, 4H, H_{Ar}), 5.03 (m, 1H, CH), 1.34 (d, *J* = 6.8 Hz, 3H, CH₂). Minor isomer: ¹H NMR (300 MHz, DMSO-d₆, δ) 8.12 – 6.75 (m, 4H, H_{Ar}), 5.04 (m, 1H, CH), 1.66 (d, *J* = 6.8 Hz, 3H, CH₂). Spectral data are consistent with literature values.⁹

Henry reaction/cycloisomerisation sequence. Yields and ¹H spectra of products **11**, **12** and **13**.

1-(2-((4-methoxyphenyl)ethynyl)phenyl)-2-nitroethanol **11a**. Yield 18% (13 mg); ¹H NMR (300 MHz, CDCl₃, δ) 7.62 (d, *J* = 7.7 Hz, 1H, H_{Ar}), 7.55 – 7.44 (m, 3H, H_{Ar}), 7.37 (td, *J* = 7.6, 1.3 Hz, 1H, H_{Ar}), 7.31 (td, *J* = 7.5, 1.3 Hz, 1H, H_{Ar}), 6.88 (d, *J* = 8.9 Hz, 2H, H_{Ar}), 5.98 (dd, *J* = 9.8, 1.8 Hz, 1H, CH), 4.79 (dd, *J* = 13.0, 2.4 Hz, 1H, CH₂), 4.50 (dd, *J* = 13.0, 9.9 Hz, 1H, CH₂), 3.81 (s, 3H, CH₃), 2.94 (s, 1H, OH). Spectral data are consistent with literature values.¹⁰

3-(4-methoxyphenyl)-1-(nitromethyl)-1H-isochromene **12a**. Yield 30% (22 mg); ¹H NMR (300 MHz, CDCl₃, δ) 7.62 – 7.55 (m, 2H, H_{Ar}), 7.31 (td, *J* = 7.5, 1.1 Hz, 1H, H_{Ar}), 7.20 (td, *J* = 7.5, 1.1 Hz, 1H, H_{Ar}), 7.12 (dd, *J* = 11.4, 7.6 Hz, 2H, H_{Ar}), 6.93 – 6.86 (m, 2H, H_{Ar}), 6.37 (s, 1H, Csp²-H),

6.11 (dd, $J = 10.3, 3.2$ Hz, 1H, CH), 4.95 (dd, $J = 12.3, 10.3$ Hz, 1H, CH₂), 4.35 (dd, $J = 12.3, 3.2$ Hz, 1H, CH₂), 3.82 (s, 3H, CH₃). Spectral data are consistent with literature values.¹⁰

2-nitro-1-(2-(p-tolylolethynyl)phenyl)ethanol 11c. Yield 32% (23 mg) Table 5, entry 4; yield 60% (42 mg) Table 5, entry 5; ¹H NMR (300 MHz, CDCl₃, δ) 7.59 (d, $J = 7.6$ Hz, 1H, H_{Ar}), 7.49 (dd, $J = 7.6, 1.1$ Hz, 1H, H_{Ar}), 7.43 (d, $J = 8.1$ Hz, 2H, H_{Ar}), 7.35 (td, $J = 7.6, 1.3$ Hz, 1H, H_{Ar}), 7.28 (td, $J = 7.5, 1.2$ Hz, 1H, H_{Ar}), 7.14 (d, $J = 7.9$ Hz, 2H, H_{Ar}), 5.94 (dd, $J = 9.8, 2.2$ Hz, 1H, CH), 4.74 (dd, $J = 13.1, 2.4$ Hz, 1H, CH₂), 4.45 (dd, $J = 13.1, 9.9$ Hz, 1H, CH₂), 2.95 (s, 1H, OH), 2.34 (s, 3H, CH₃). Spectral data are consistent with literature values.¹⁰

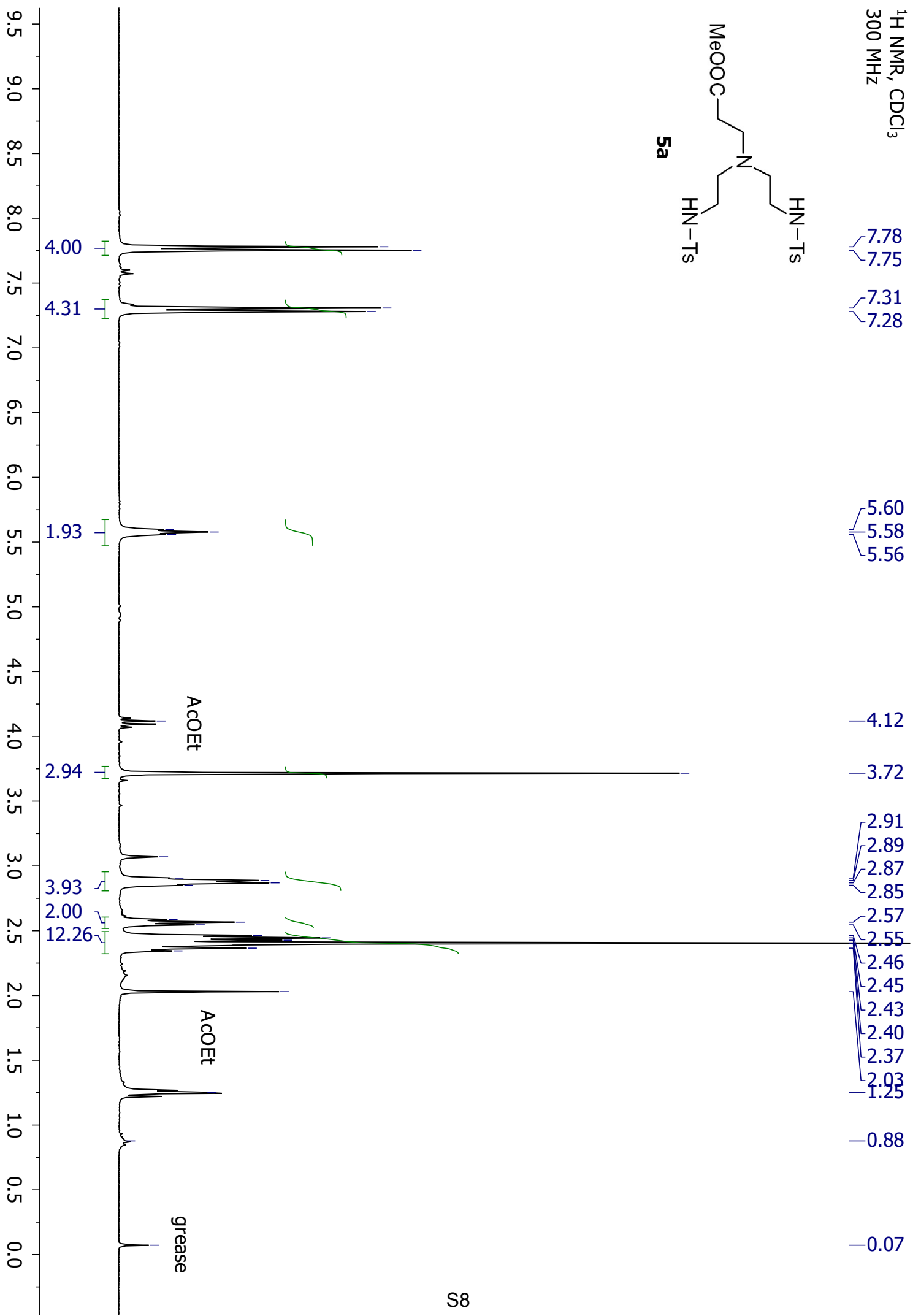
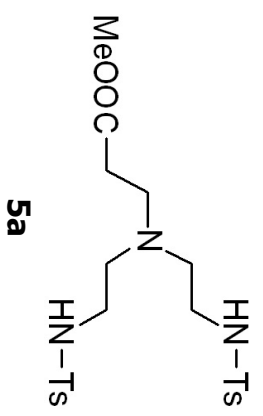
1-(nitromethyl)-3-(p-tolyl)-1H-isochromene 12c. Yield 31% (22 mg) Table 5, entry 4; yield 5% (4 mg) Table 5, entry 5; ¹H NMR (400 MHz, CDCl₃, δ) 7.55 (d, $J = 7.9$ Hz, 2H, H_{Ar}), 7.32 (t, $J = 7.5$ Hz, 1H, H_{Ar}), 7.27 – 7.04 (m, 5H, H_{Ar}), 6.45 (s, 1H, Csp²-H), 6.13 (dd, $J = 10.2, 2.7$ Hz, 1H, CH), 4.95 (dd, $J = 12.3, 7.2$ Hz, 1H, CH₂), 4.36 (dd, $J = 12.3, 3.0$ Hz, 1H, CH₂), 2.37 (s, 3H, CH₃). Spectral data are consistent with literature values.¹⁰

3-(4-methoxyphenyl)-1H-isochromen-1-one 13. Yield 29% (18 mg); ¹H NMR (200 MHz, CDCl₃, δ) 8.29 (d, $J = 9.2$ Hz, 1H, H_{Ar}), 7.83 (d, $J = 8.8$ Hz, 2H, H_{Ar}), 7.70 (td, $J = 7.7, 1.1$ Hz, 1H, H_{Ar}), 7.50 – 7.42 (m, 2H, H_{Ar}), 6.98 (d, $J = 8.8$ Hz, 2H, H_{Ar}), 6.84 (s, 1H, Csp²-H), 3.87 (s, 3H, CH₃). Spectral data are consistent with literature values.¹¹

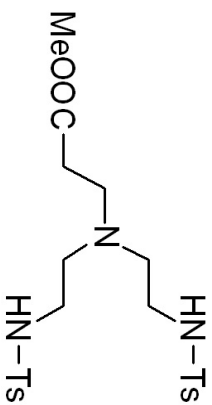
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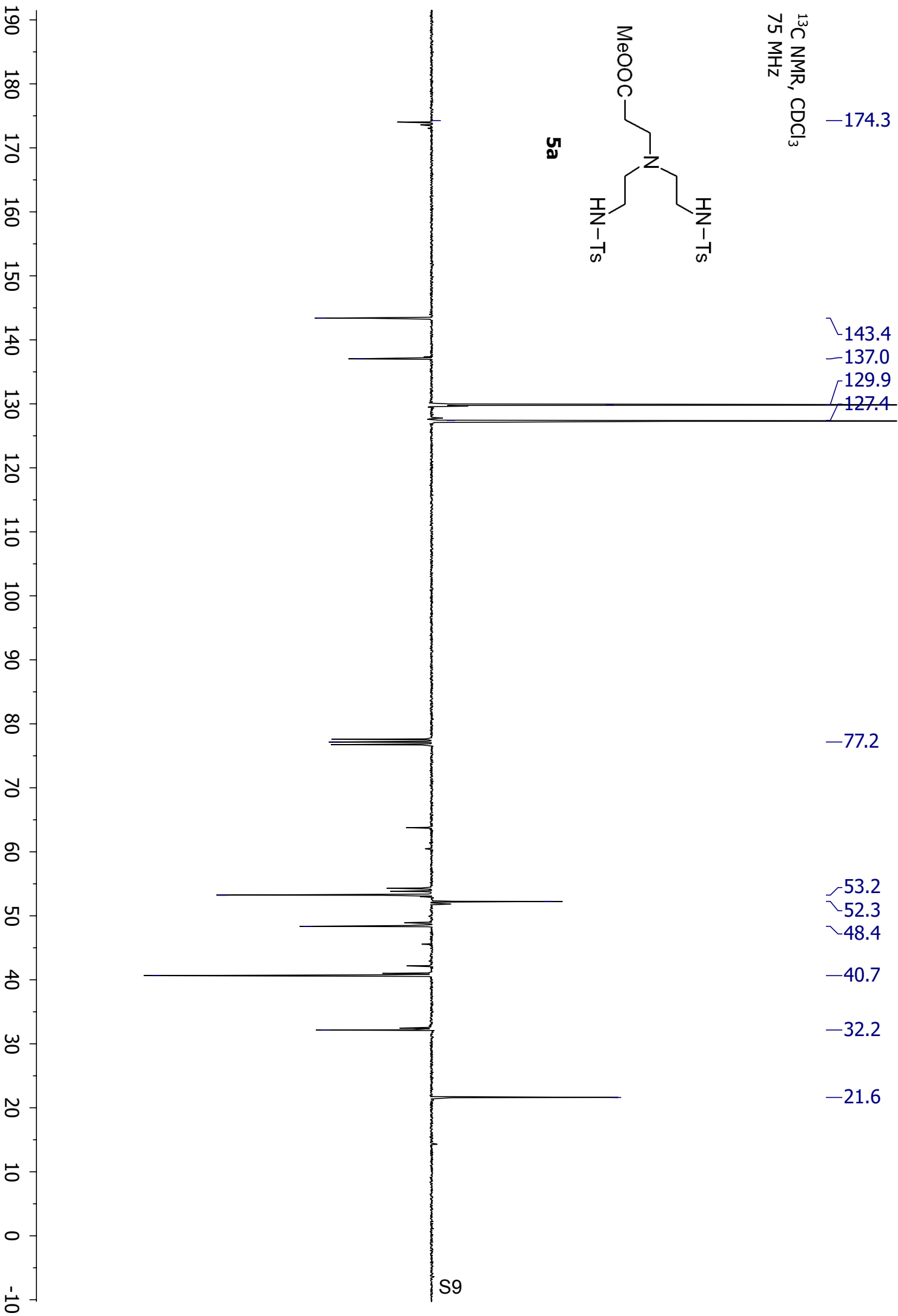
¹H NMR, CDCl₃
300 MHz



^{13}C NMR, CDCl_3
75 MHz

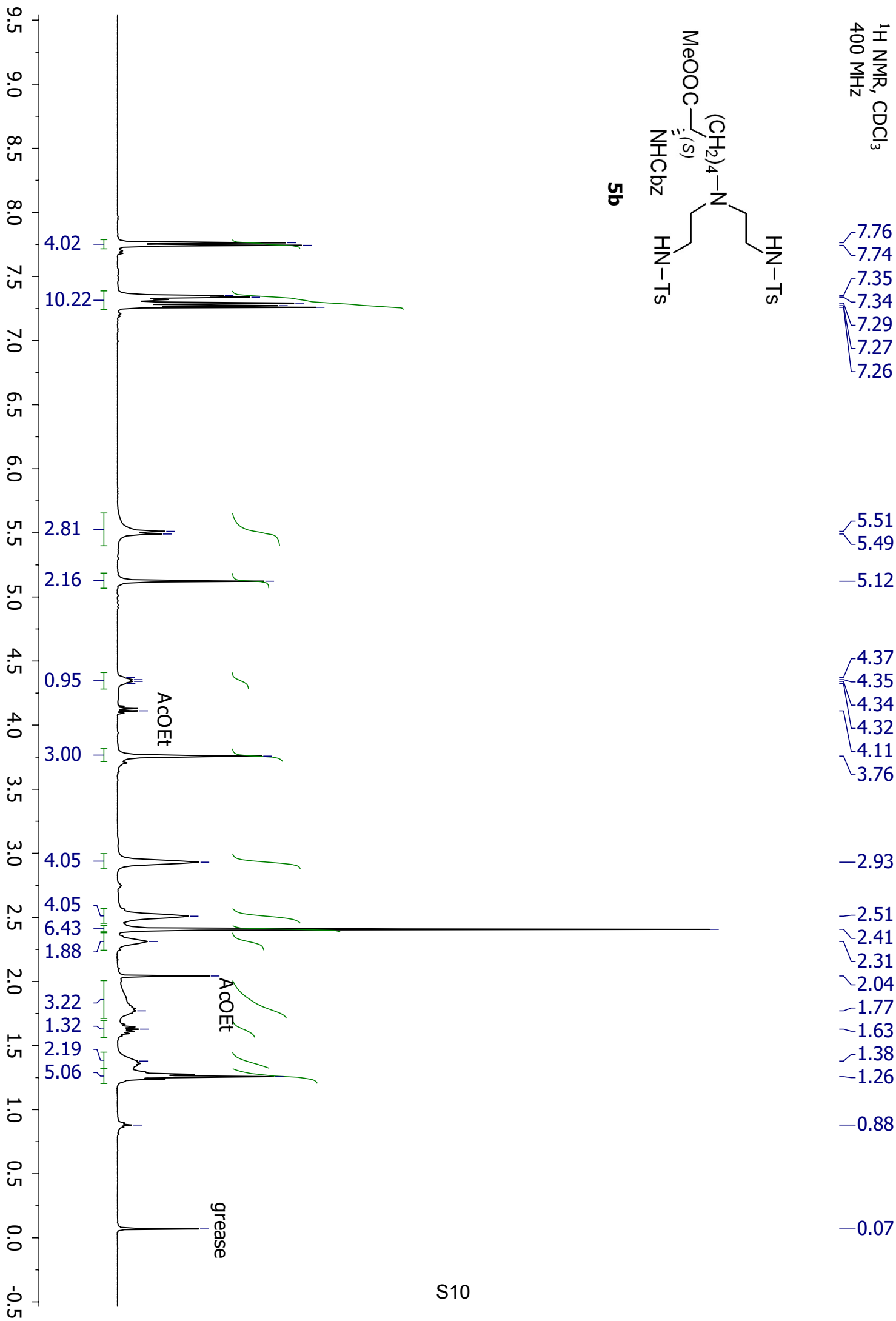
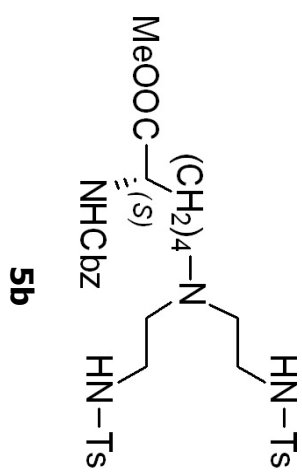


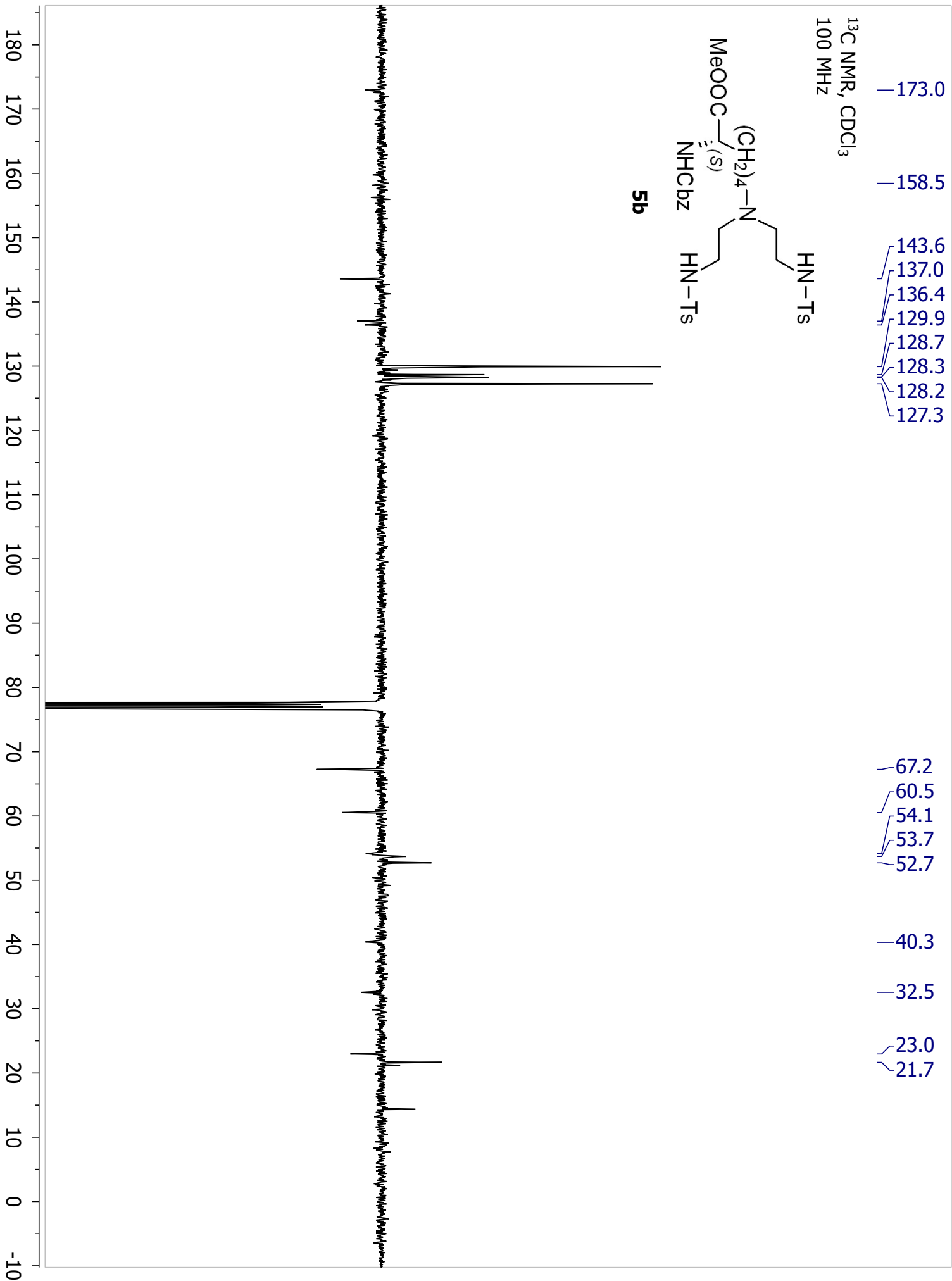
5a



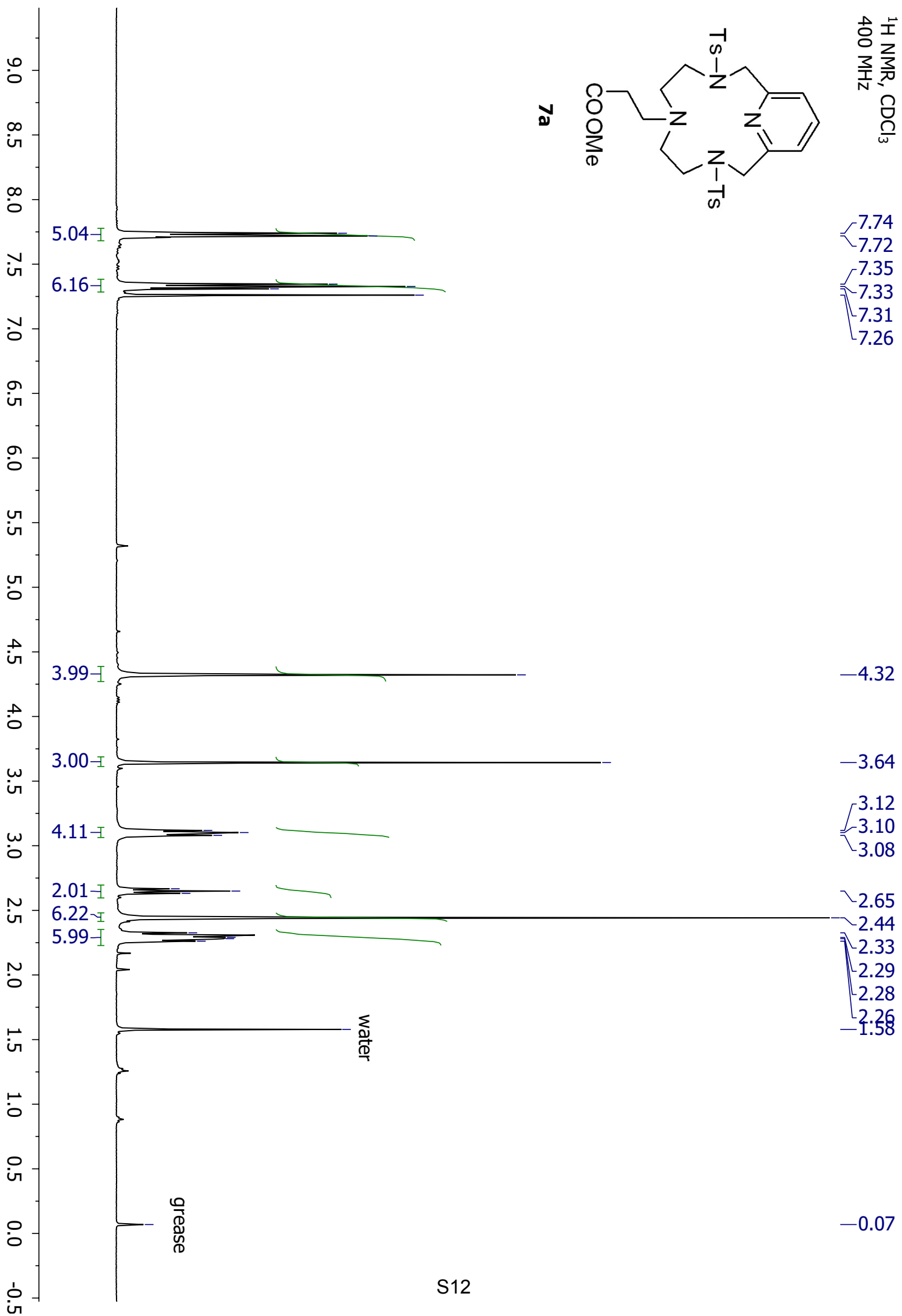
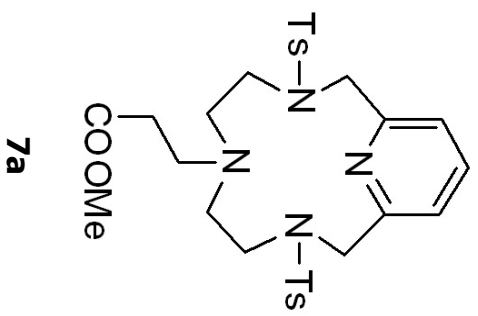
S9

¹H NMR, CDCl₃
400 MHz

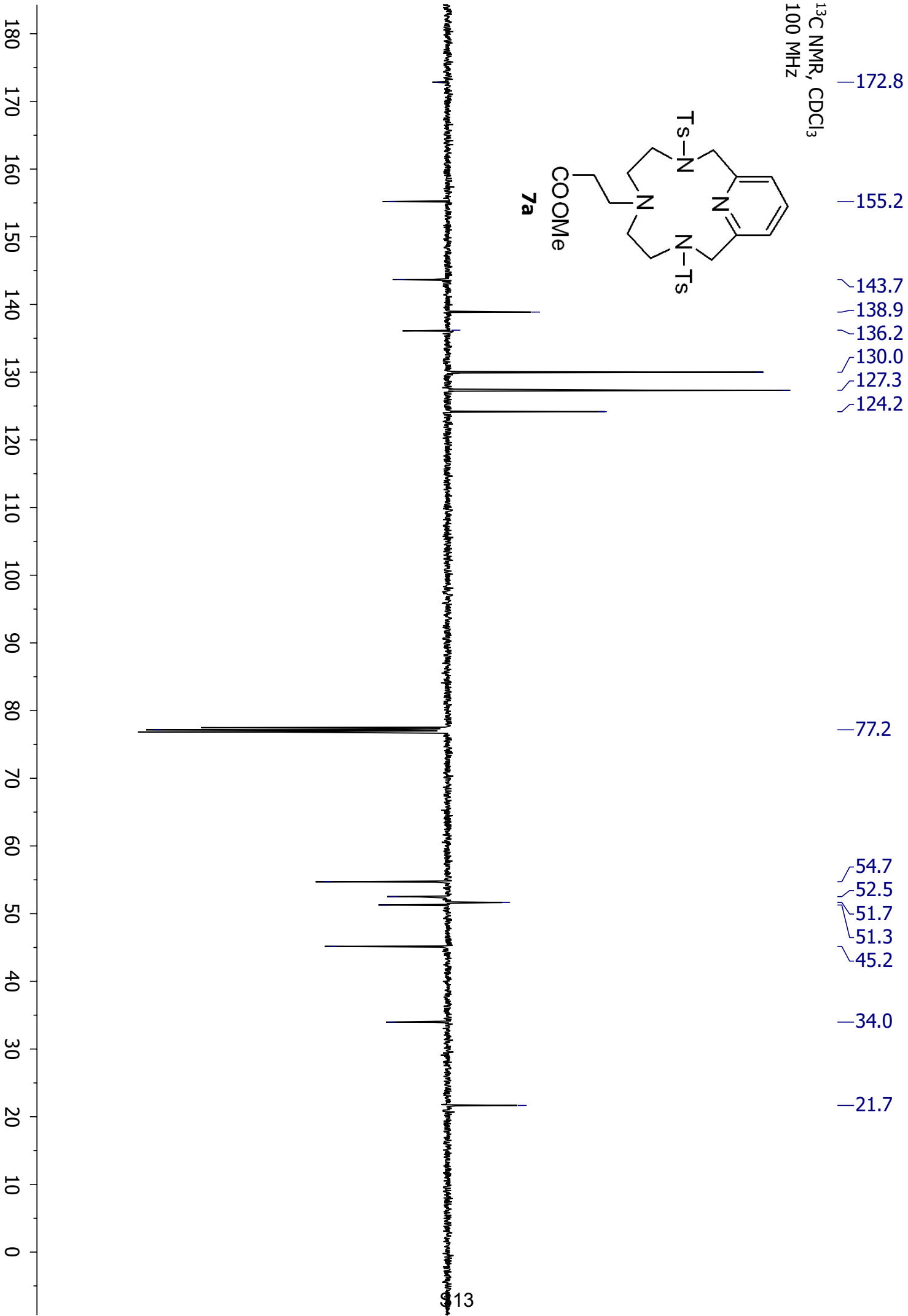
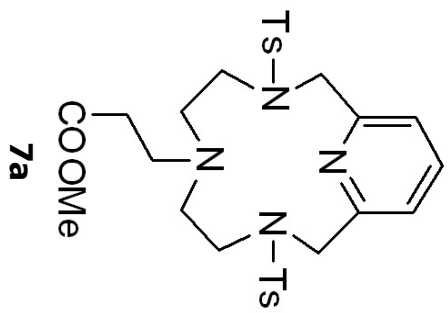




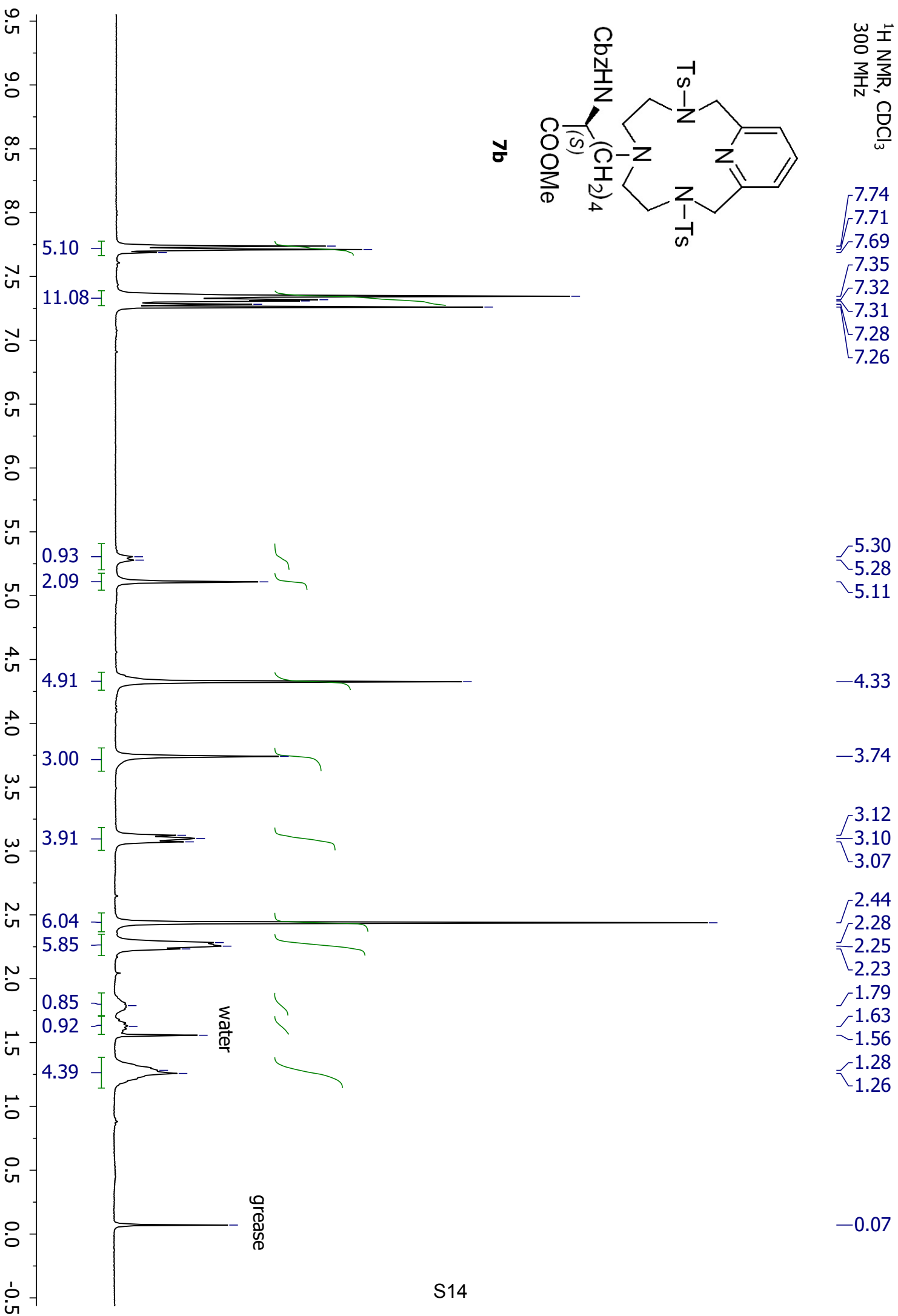
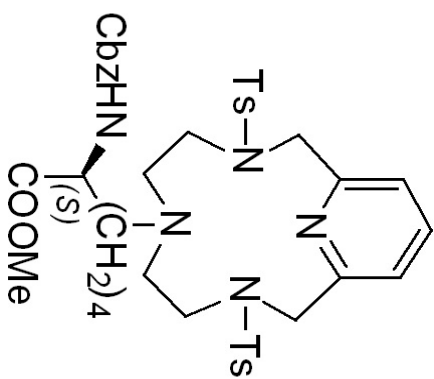
¹H NMR, CDCl₃
400 MHz



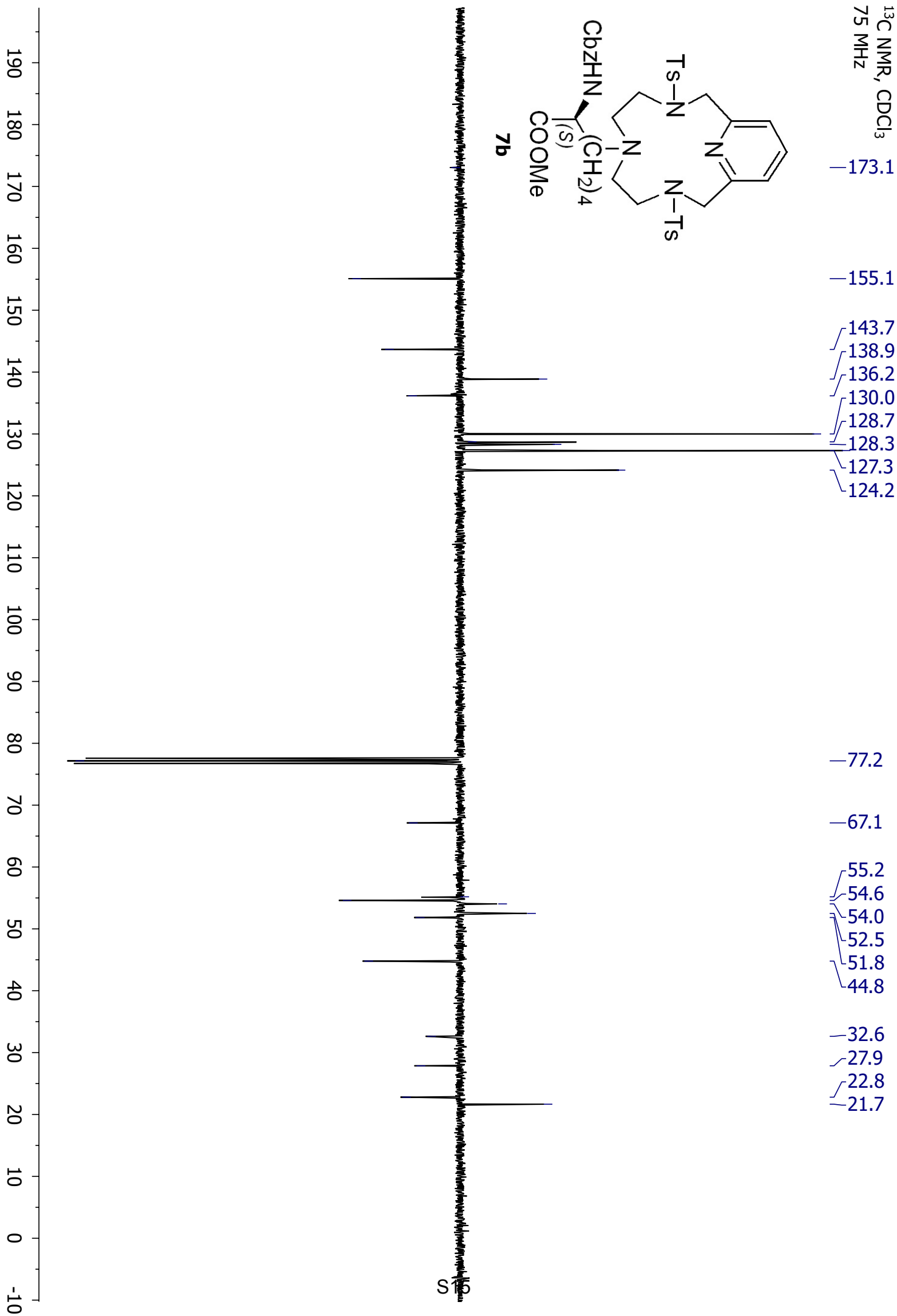
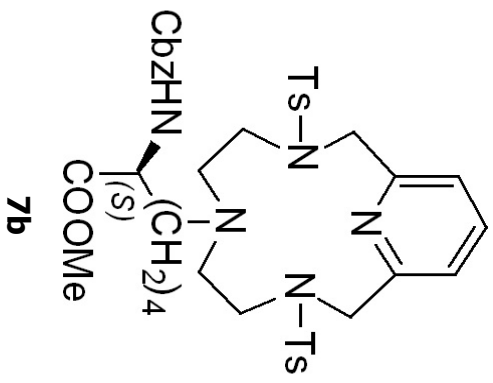
¹³C NMR, CDCl₃
100 MHz

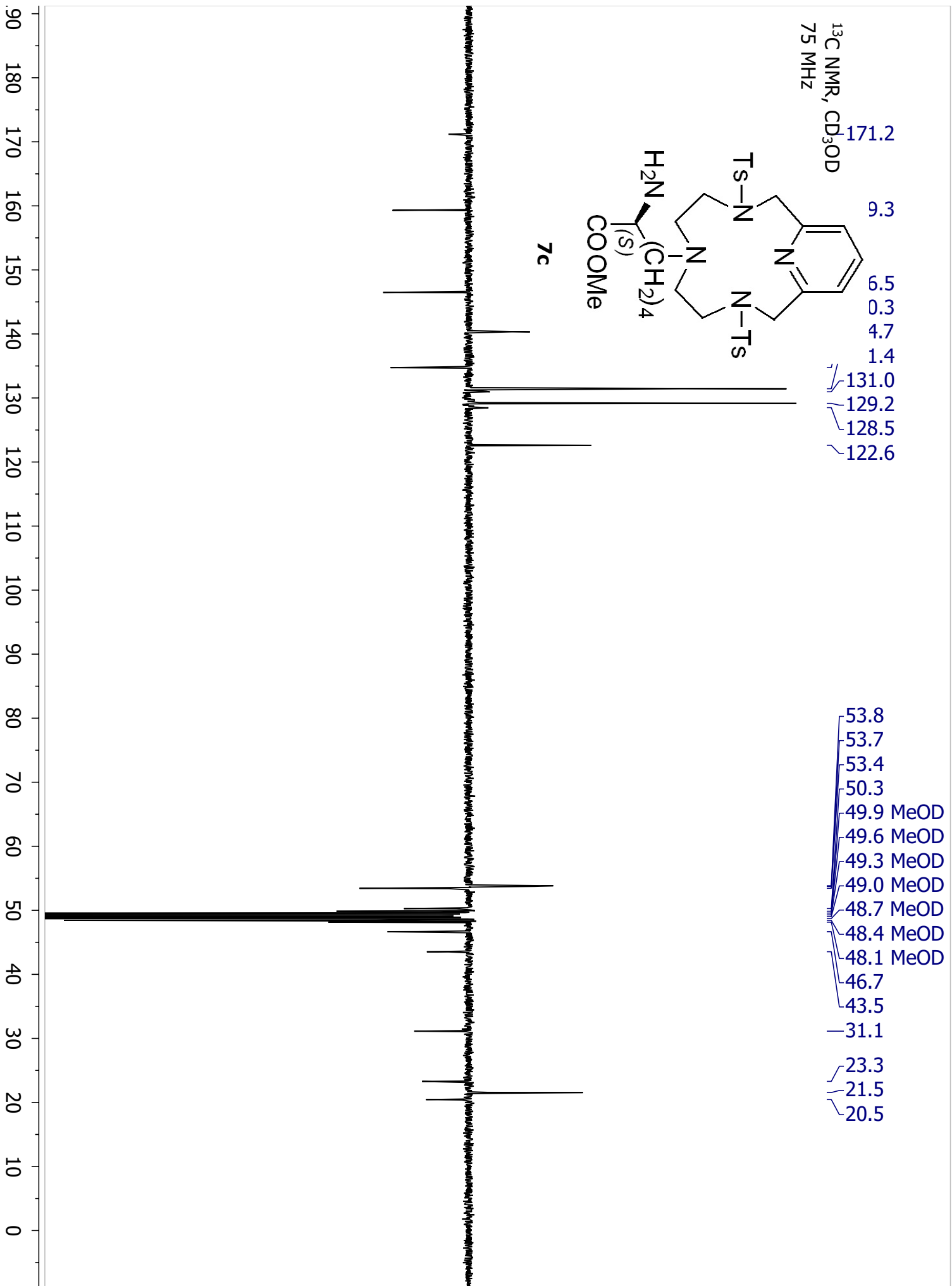


¹H NMR, CDCl₃
300 MHz

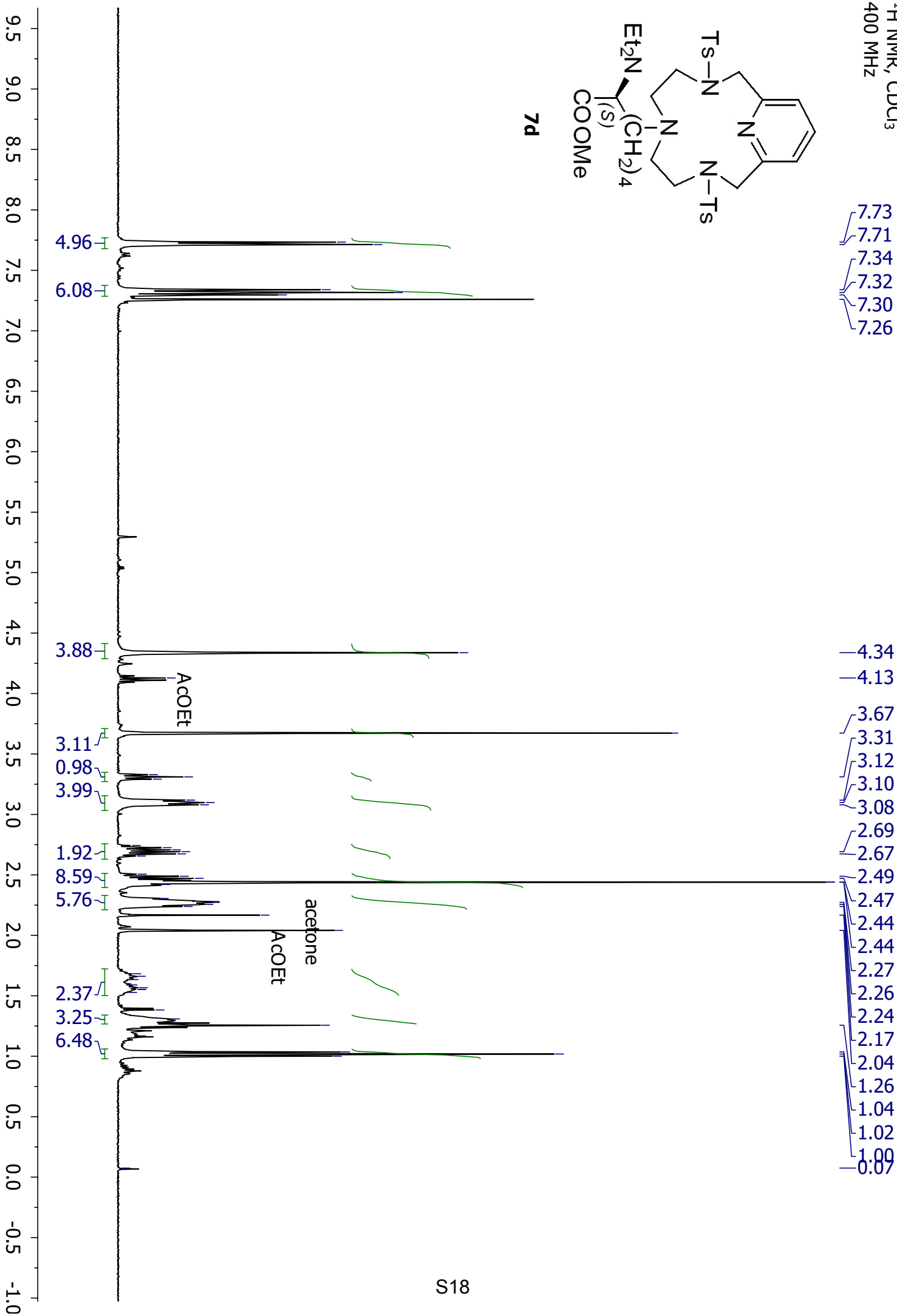
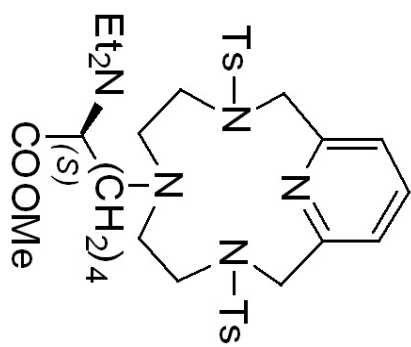


¹³C NMR, CDCl₃
75 MHz

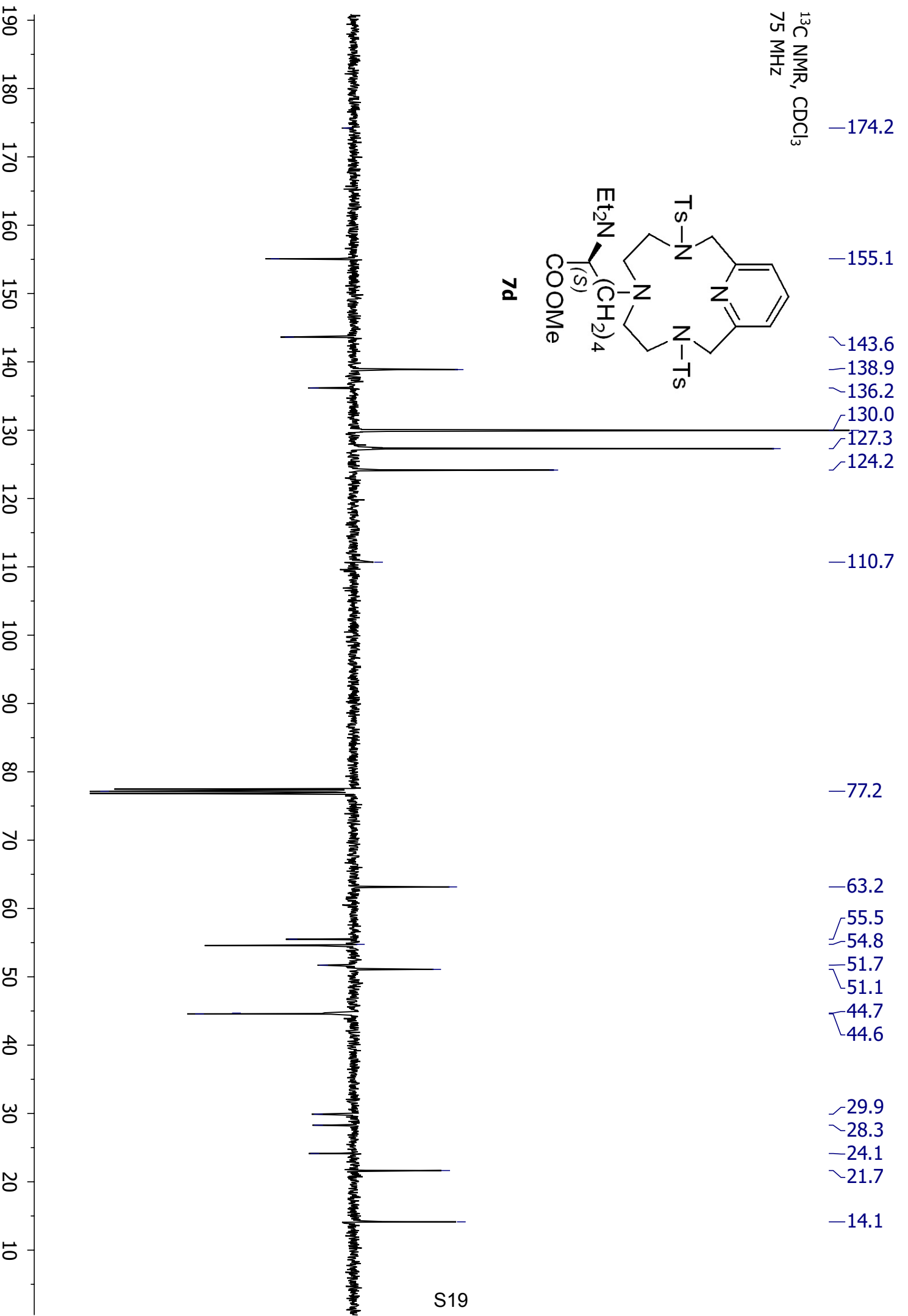
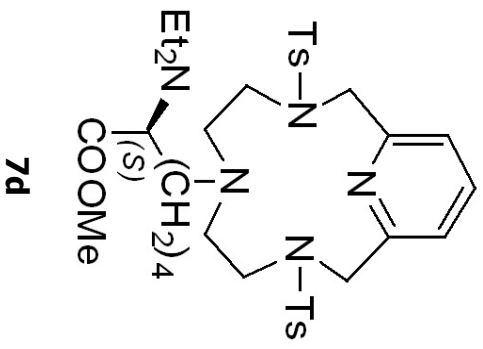




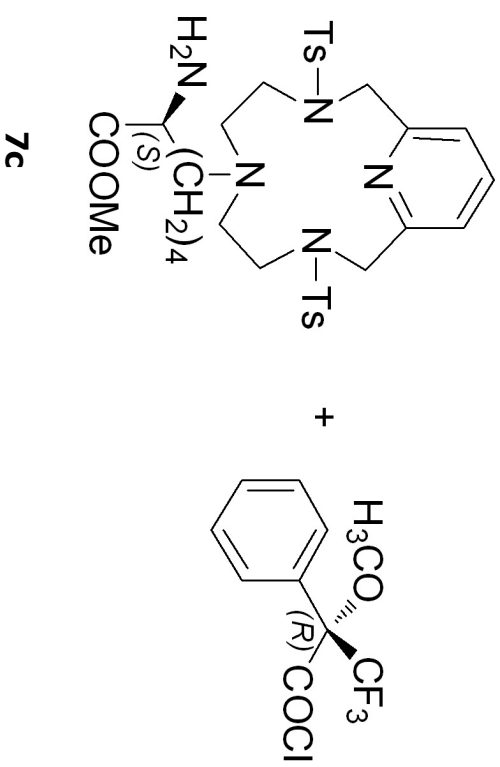
¹H NMR, CDCl₃
400 MHz



¹³C NMR, CDCl₃
75 MHz



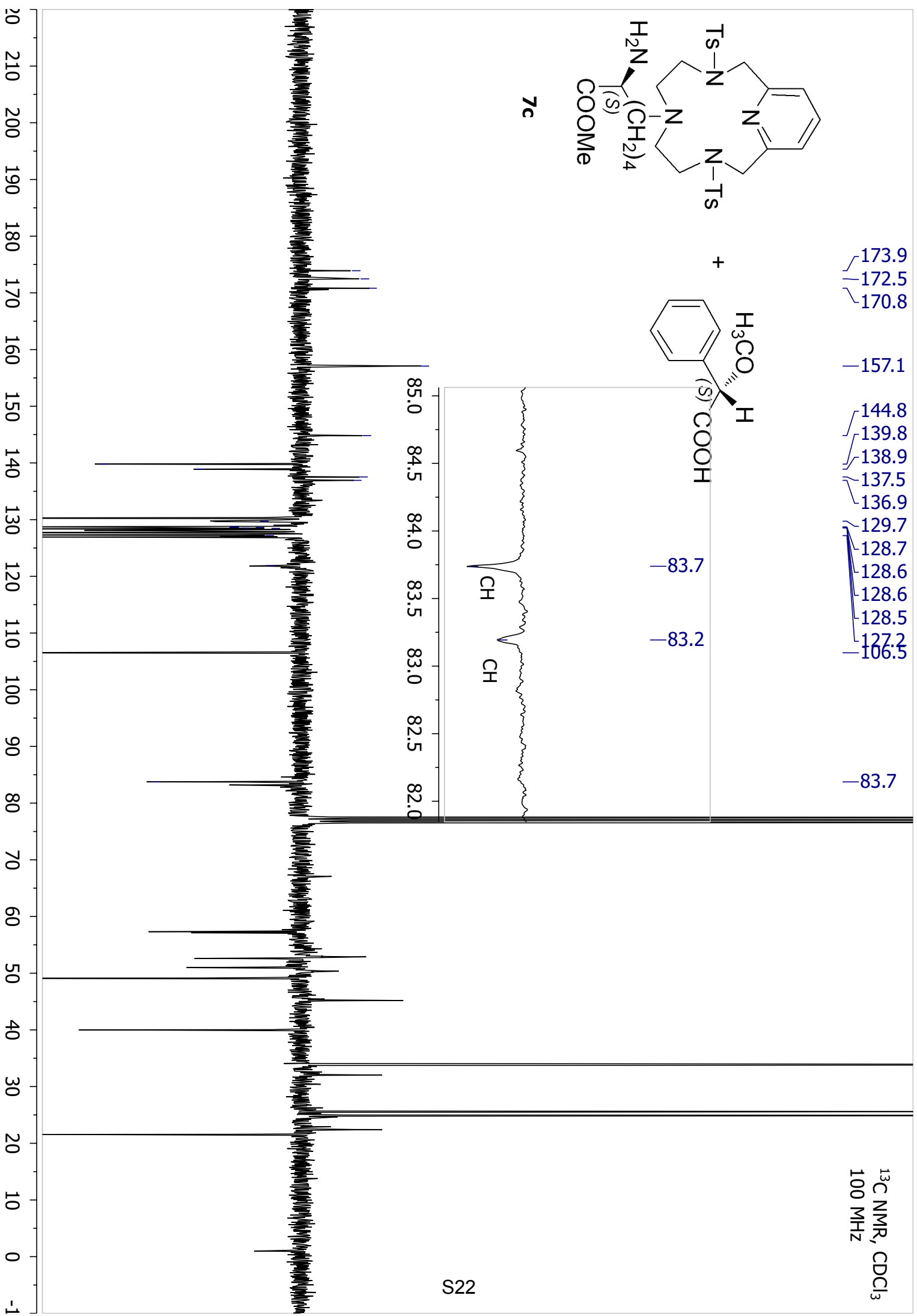
-71.24



S20

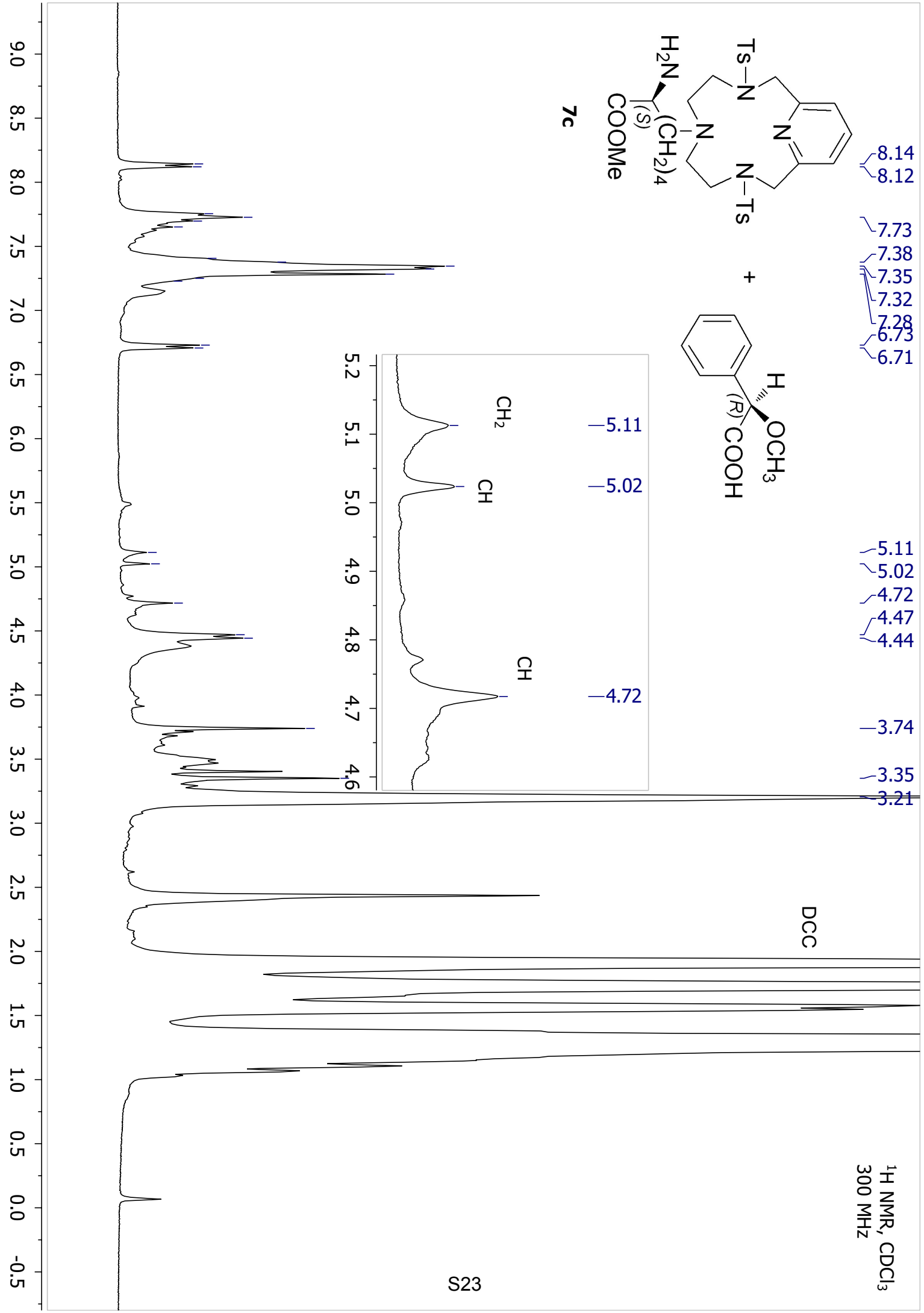
-63.5 -64.5 -65.5 -66.5 -67.5 -68.5 -69.5 -70.5 -71.5 -72.5 -73.5 -74.5 -75.5 -76.5 -77.5

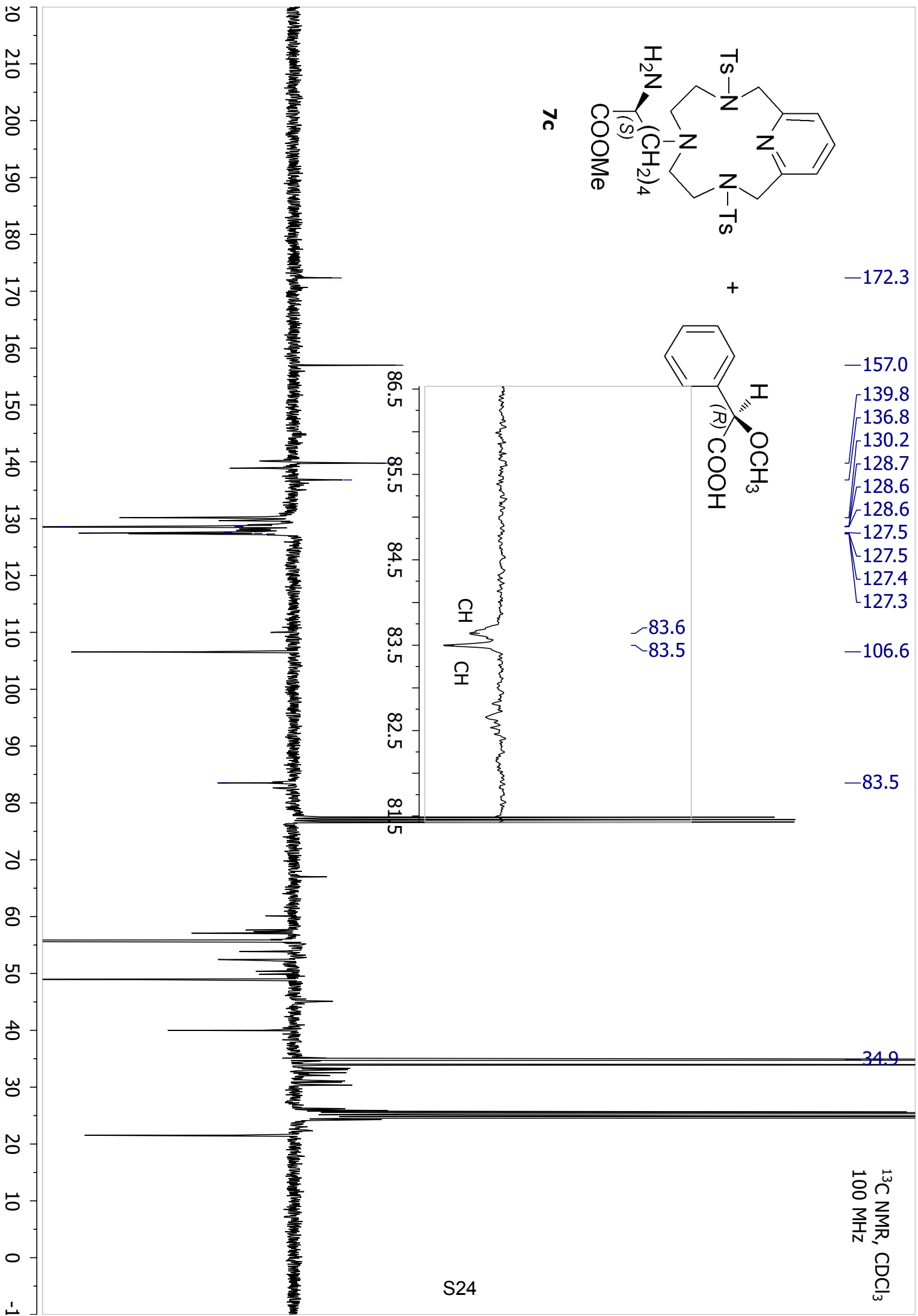
¹³C NMR, CDCl₃
100 MHz



¹H NMR, CDCl₃
300 MHz

S23





¹H NMR, CDCl₃
300 MHz

7.89
7.86
7.84
7.77
7.75
7.47
7.44
7.33
7.30
7.26

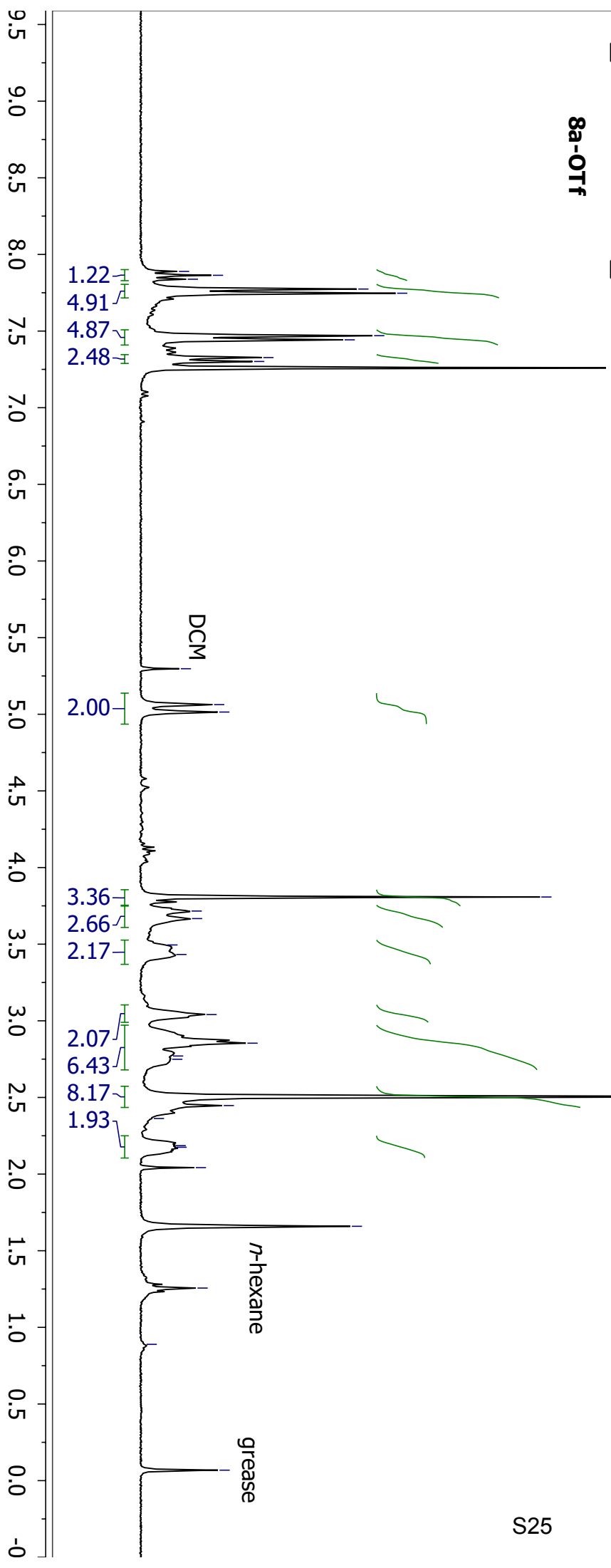
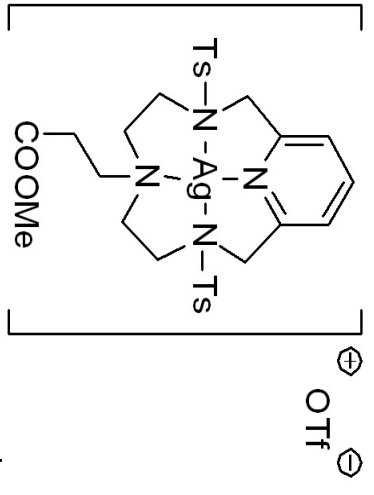
5.30
5.06
5.01

3.81
3.72
3.67
3.49
3.43

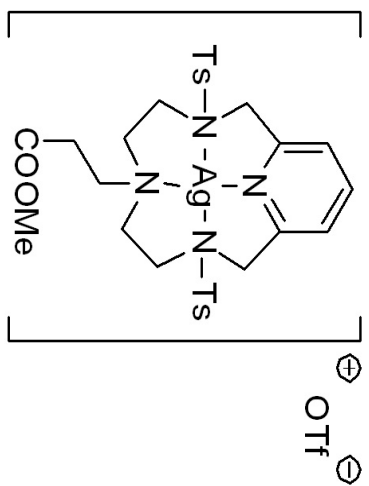
3.04
2.85
2.77
2.51
2.45
2.19
2.18
2.04
1.66

1.26
0.89

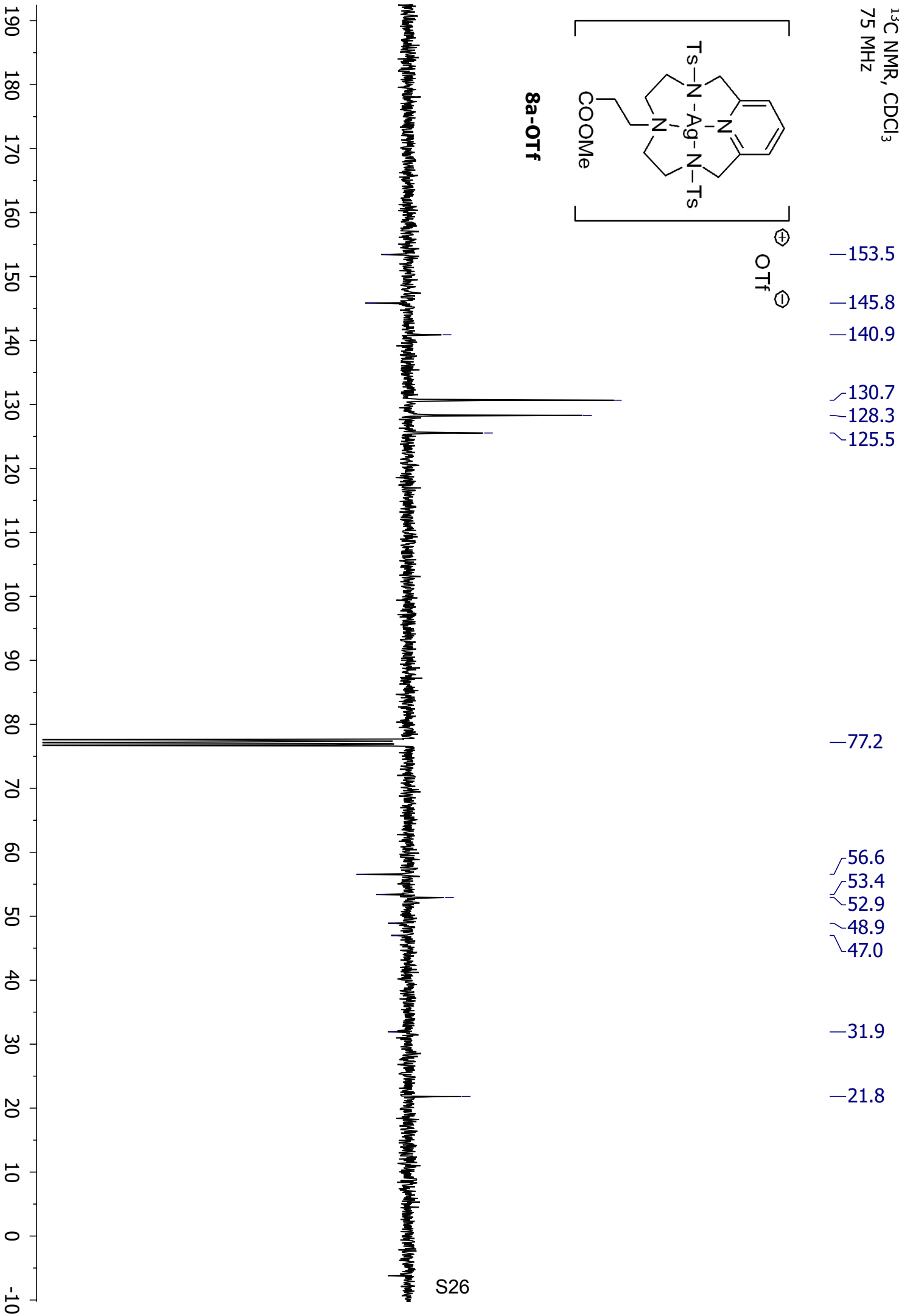
0.07



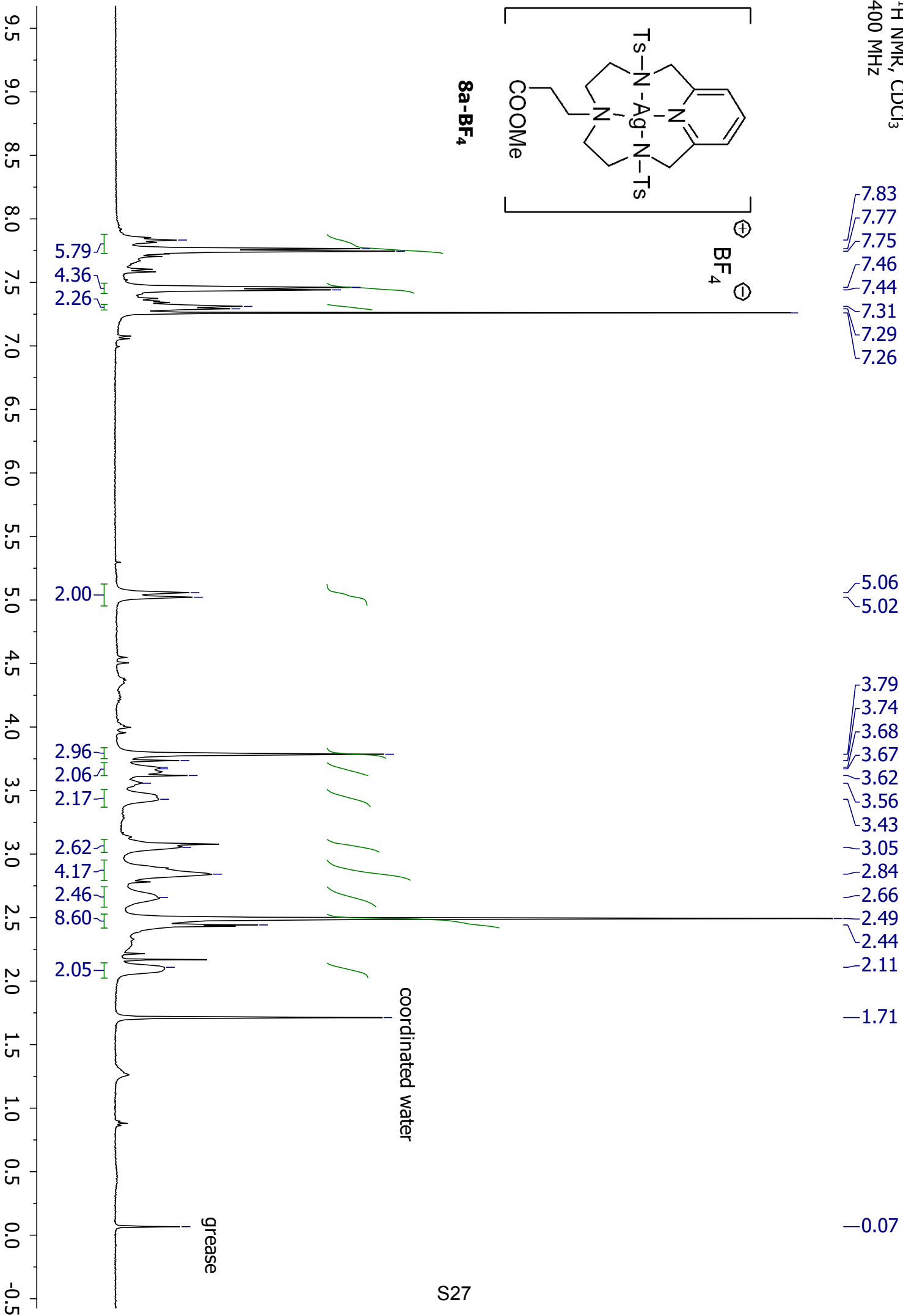
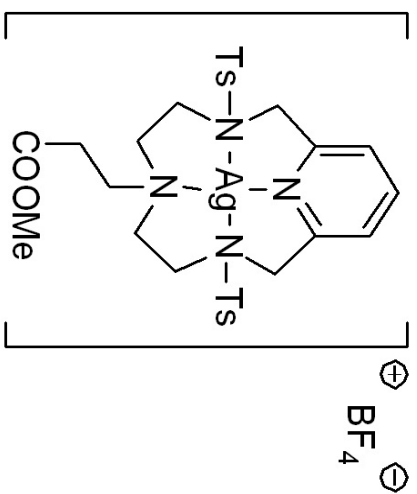
¹³C NMR, CDCl₃
75 MHz



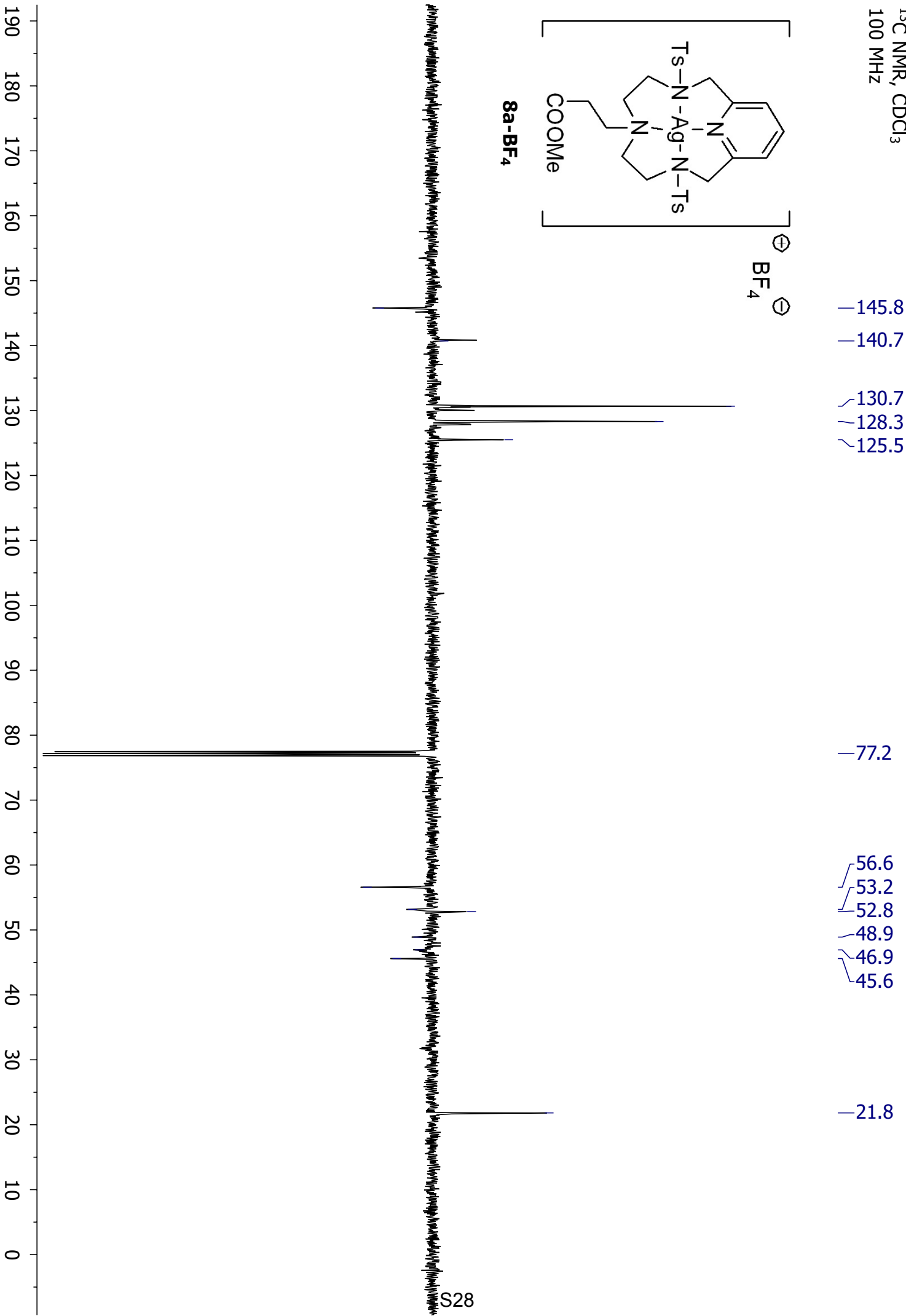
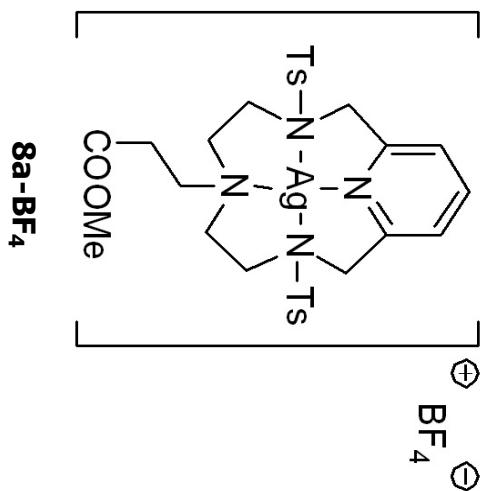
8a-OTf



¹H NMR, CDCl₃
400 MHz



¹³C NMR, CDCl₃
100 MHz



¹H NMR, CDCl₃
400 MHz

7.80
7.75
7.46
7.44
7.34
7.26

5.68

5.18
5.13
5.09

4.48

3.74
3.69
3.65

3.42
3.27
3.08

2.93
2.76

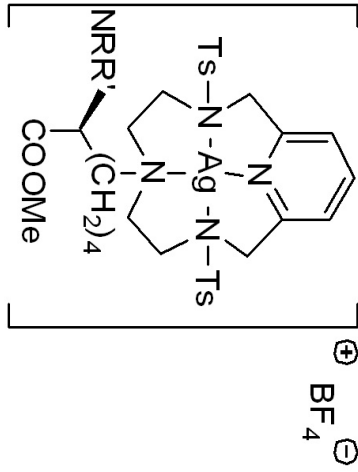
2.57
2.48
2.44

2.06
1.94
1.77

1.55
1.25

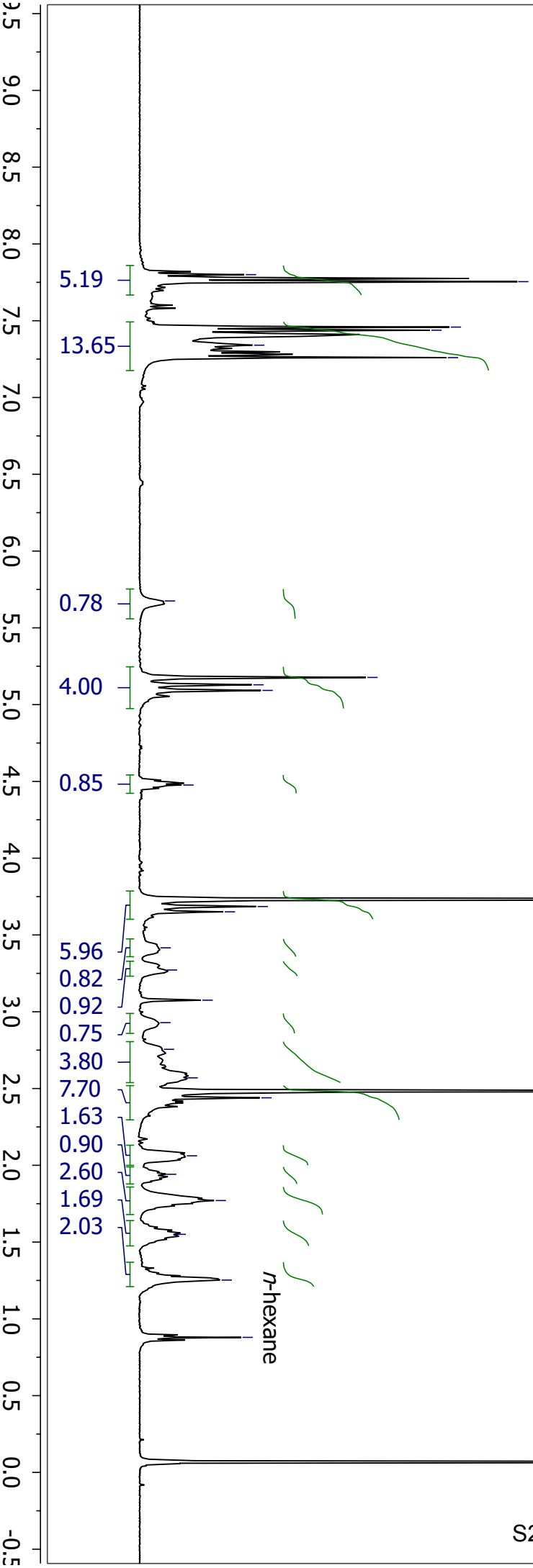
0.88

0.07

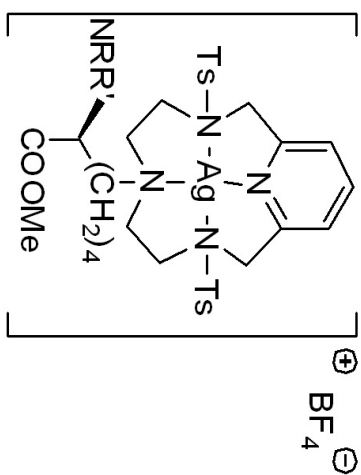


R = H, R' = Cbz

8b-BF₄

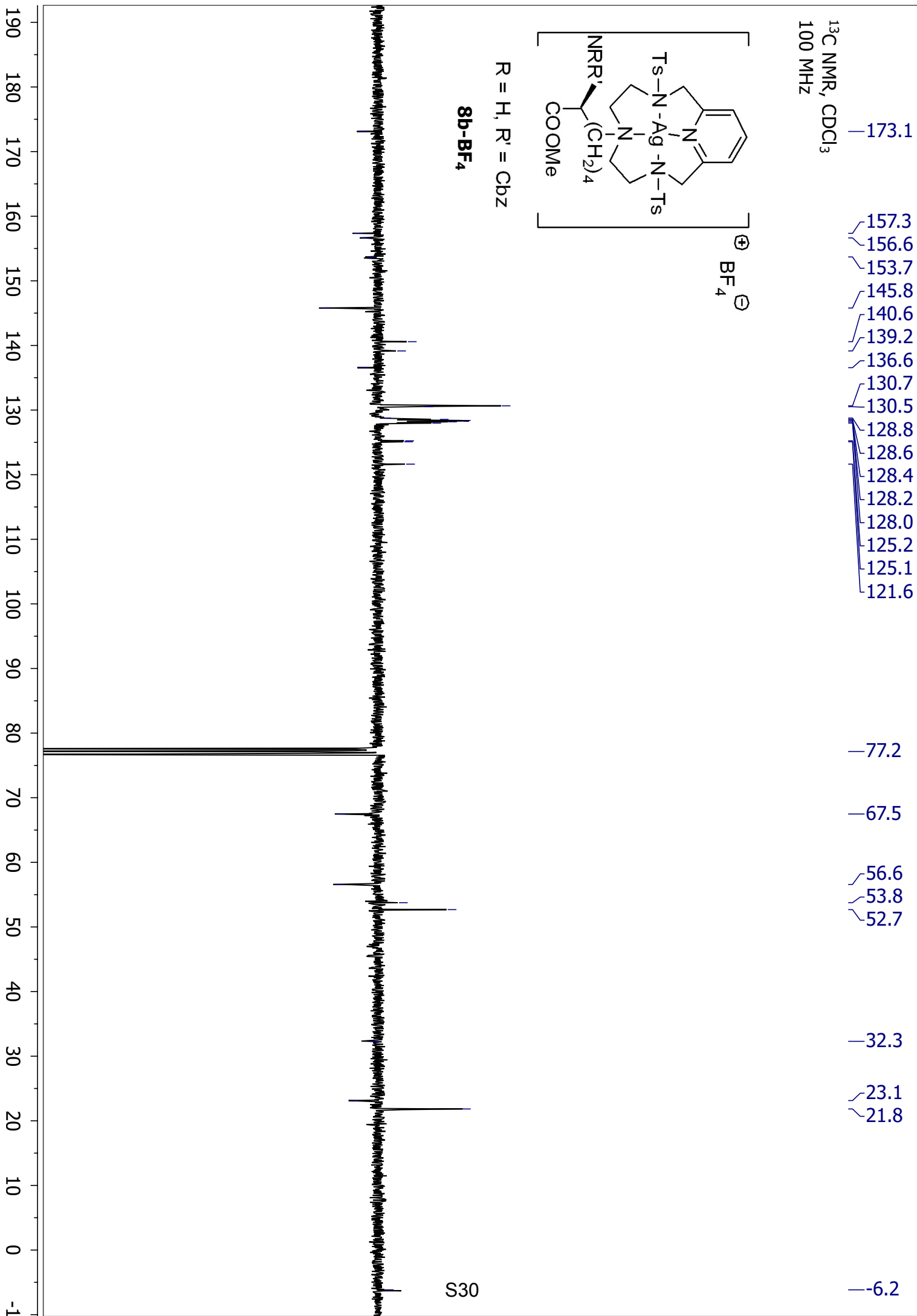


¹³C NMR, CDCl₃
100 MHz



R = H, R' = Cbz

8b-BF₄



¹H NMR, CDCl₃
300 MHz

7.85
7.78
7.75
7.47
7.41
7.39
7.36
7.26
7.08

5.58
5.55
5.17
5.13
5.08

4.53
4.50

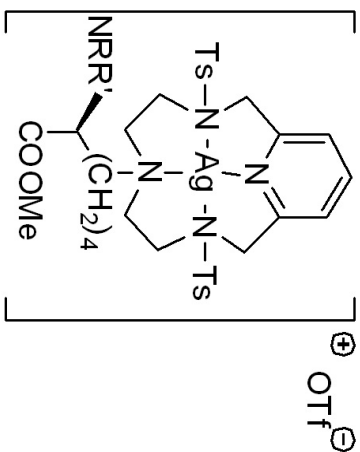
3.76
3.73
3.48
3.33
3.27
3.03

2.74
2.72
2.51
2.43
2.17
1.95

1.81
1.75
1.59
1.25

0.87

0.07



R = H, R' = Cbz

8b-OTf

9.5
9.0
8.5
8.0
7.5
7.0
6.5
6.0
5.5
5.0
4.5
4.0
3.5
3.0
2.5
2.0
1.5
1.0
0.5
0.0

1.16
5.30
19.96

1.16
1.99
2.66

1.31

6.47
0.31
0.21
0.37
3.59
0.90

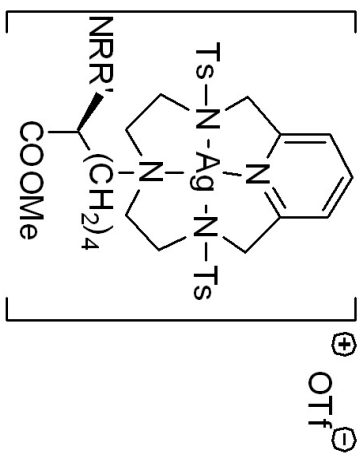
5.91
2.12
4.64
1.18
2.53

7.47
2.34
2.75

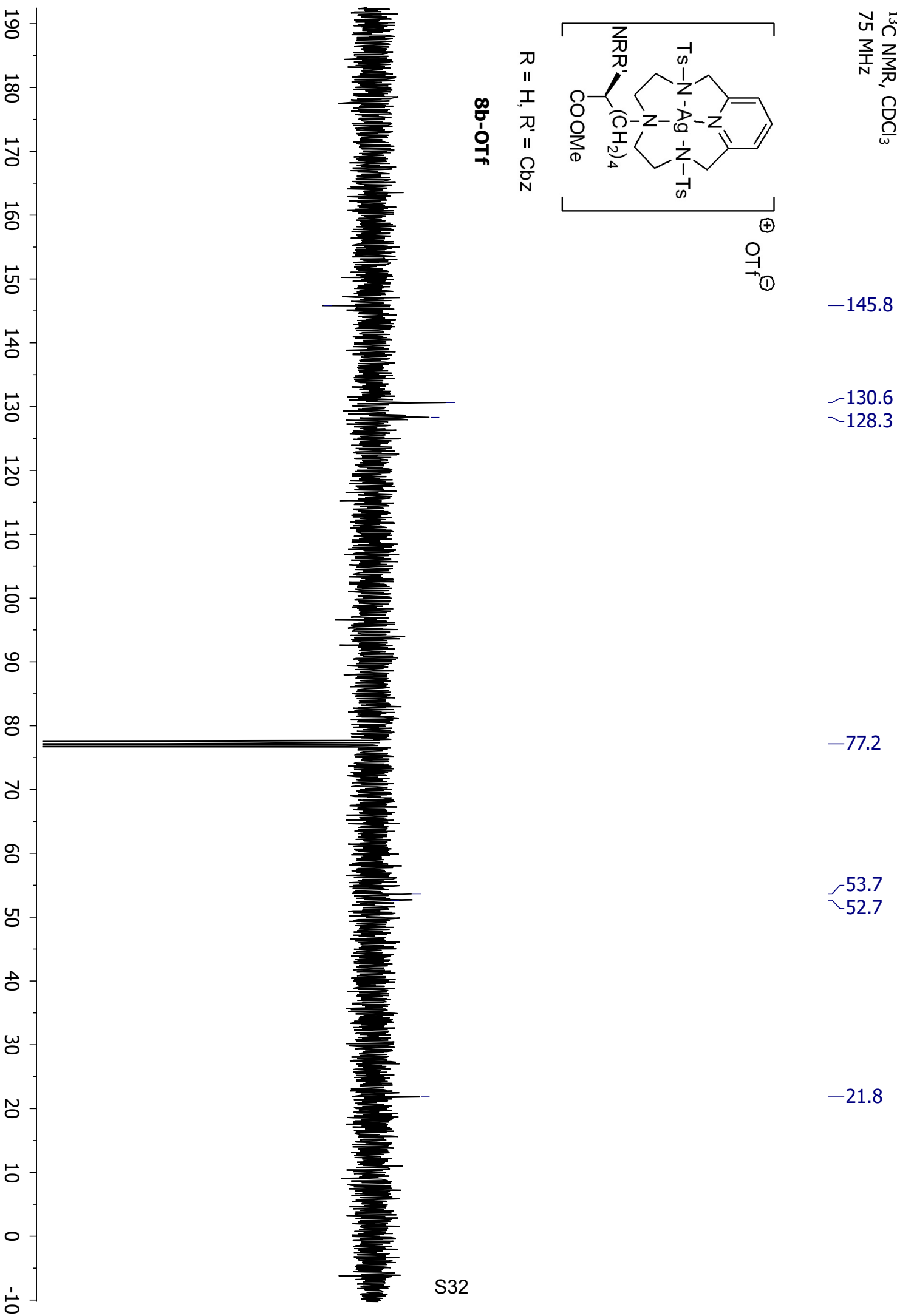
n-hexane

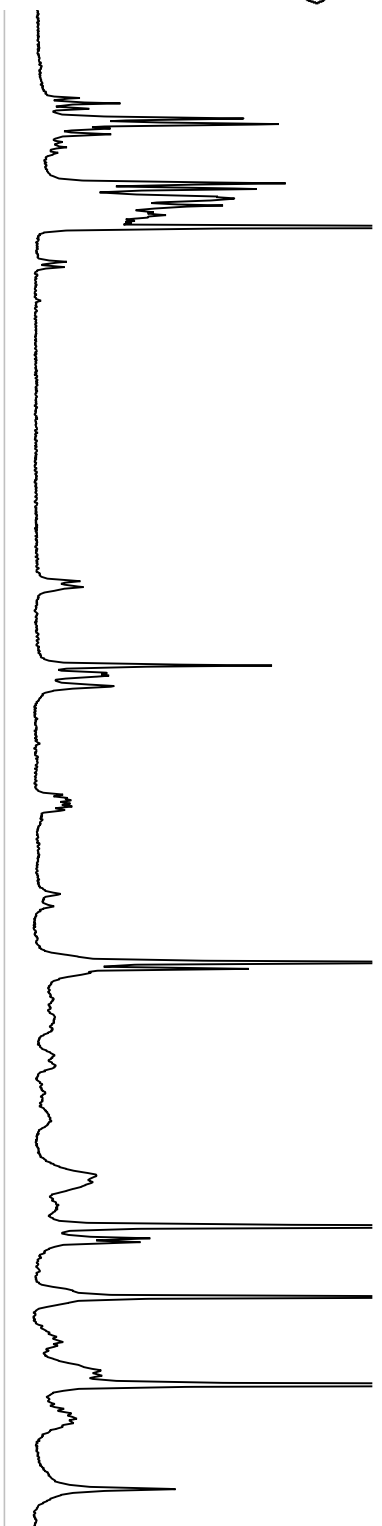
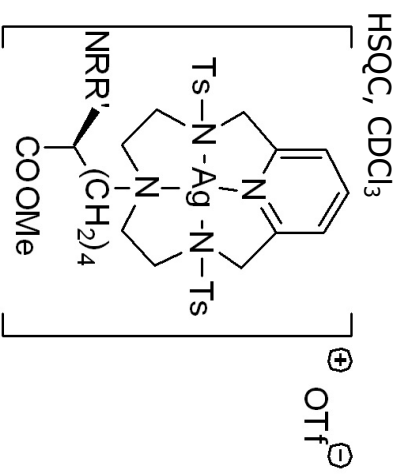
grease

¹³C NMR, CDCl₃
75 MHz



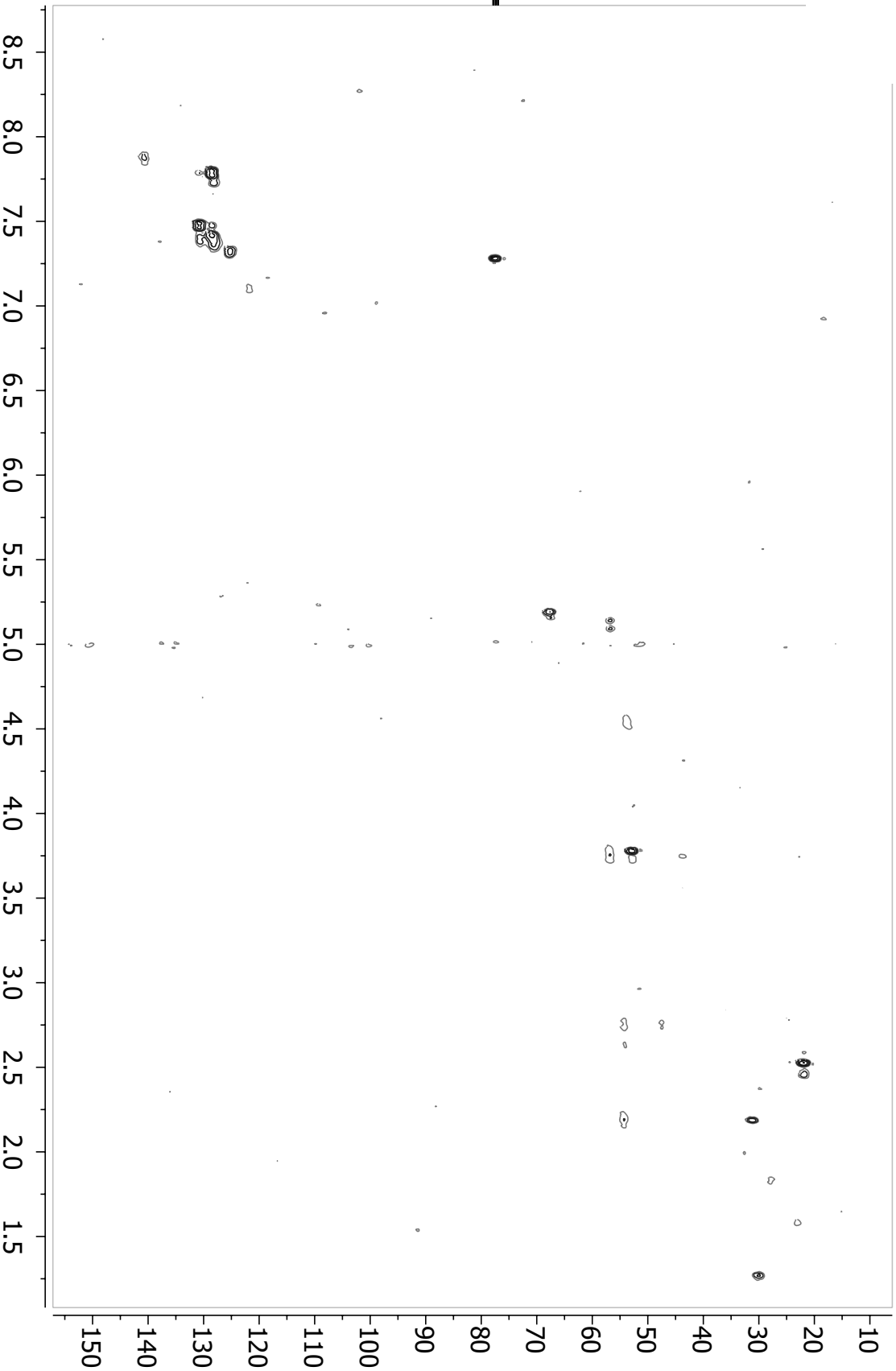
8b-OTf





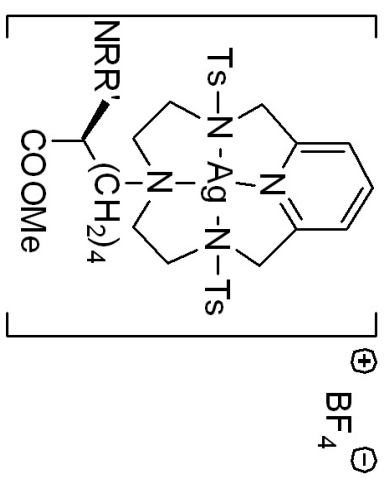
R = H, R' = Cbz

8b-OTf

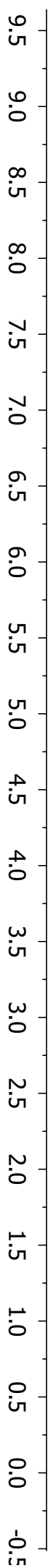


¹H NMR, CD₃OD
300 MHz

- 7.84
- 7.81
- 7.78
- 7.75
- 7.73
- 7.70
- 7.48
- 7.45
- 7.42
- 7.39
- 7.34
- 7.32
- 7.26
- 7.23
- 4.83
- 4.64
- 4.58
- 4.33
- 4.29
- 4.24
- 4.09
- 4.03
- 3.87
- 3.74
- 3.72
- 3.62
- 3.40
- 3.31
- 3.11
- 3.07
- 2.45
- 2.10
- 1.92
- 1.66
- 0.10



R = R' = H
8c-BF₄



¹³C NMR, CD₃OD
75 MHz

— 159.3

BF_4^{\ominus}

~ 146.4

~ 140.3

~ 134.7

~ 131.4

~ 131.0

~ 129.1

~ 128.2

~ 122.6

— 67.8

~ 53.9

~ 53.7

~ 53.4

~ 49.0

~ 46.7

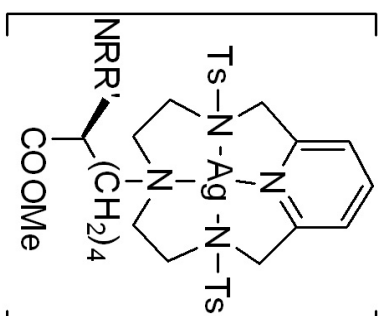
~ 43.6

— 31.1

~ 23.2

~ 21.5

~ 20.2



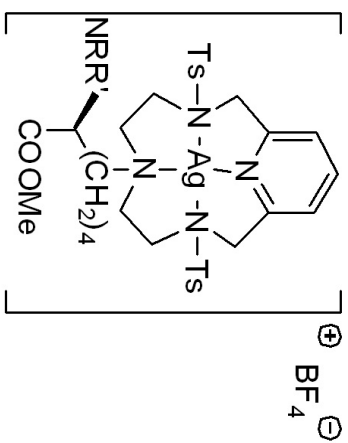
$\text{R} = \text{R}' = \text{H}$

8c-BF₄

190
180
170
160
150
140
130
120
110
100
90
80
70
60
50
40
30
20
10
0

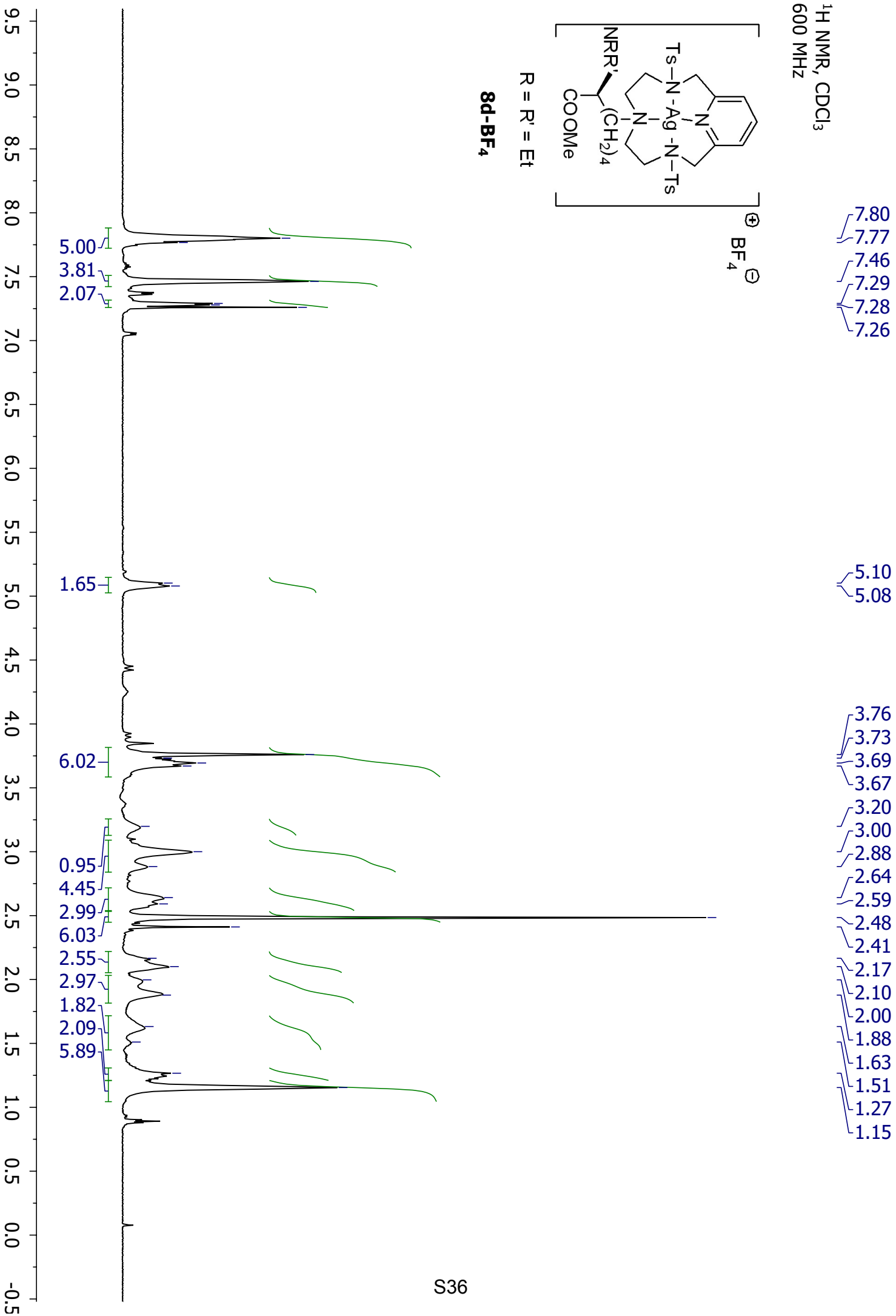
S35

¹H NMR, CDCl₃
600 MHz



R = R' = Et

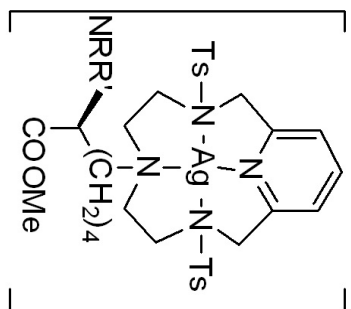
8d-BF₄



¹³C NMR, CDCl₃
151 MHz

— 153.7
— 145.8
— 140.6
— 130.7
— 128.7
— 128.6
— 125.0

BF_4^-



R = R' = Et

8d-BF₄

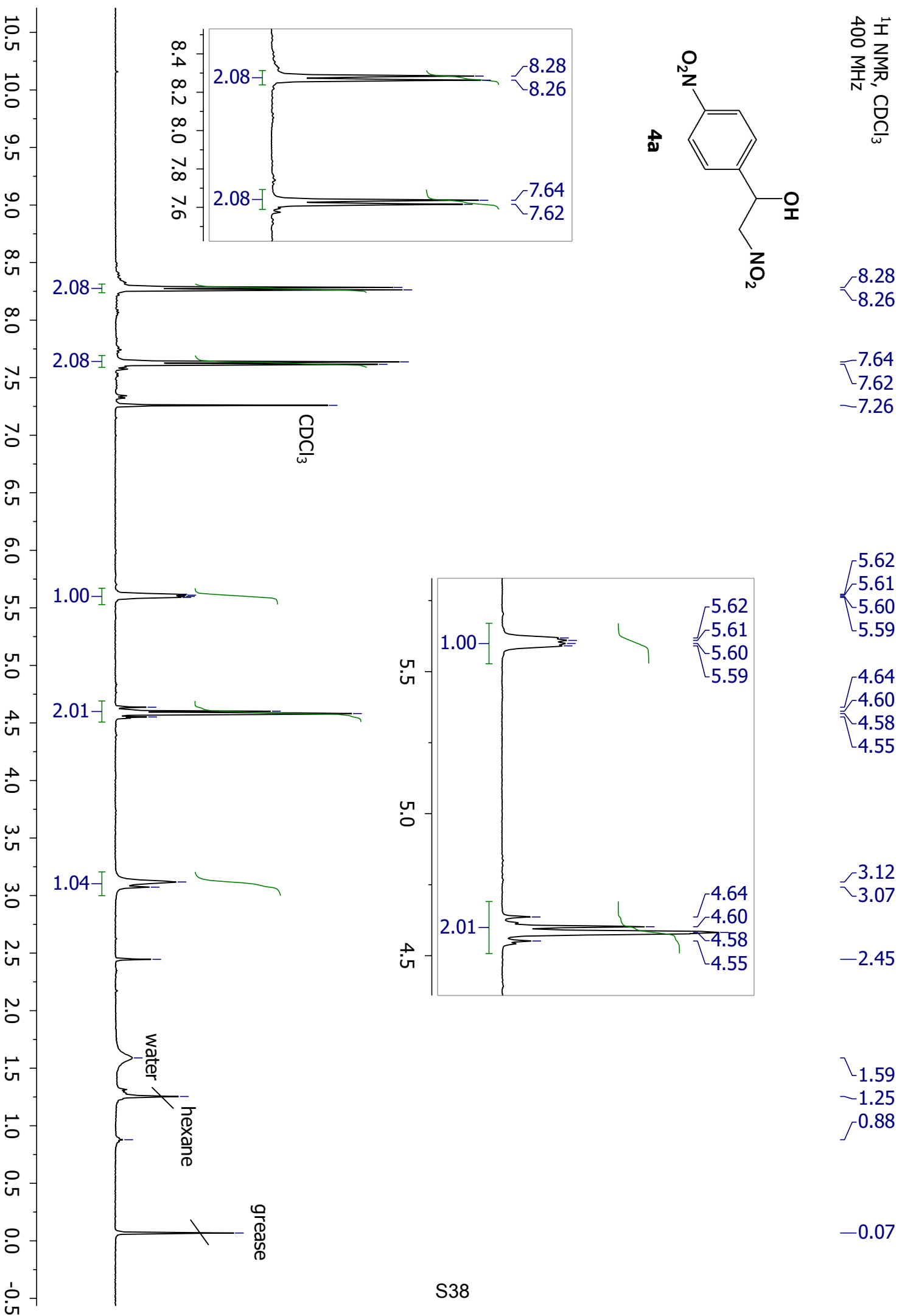
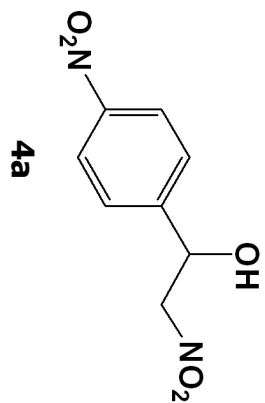
— 77.2

— 56.5

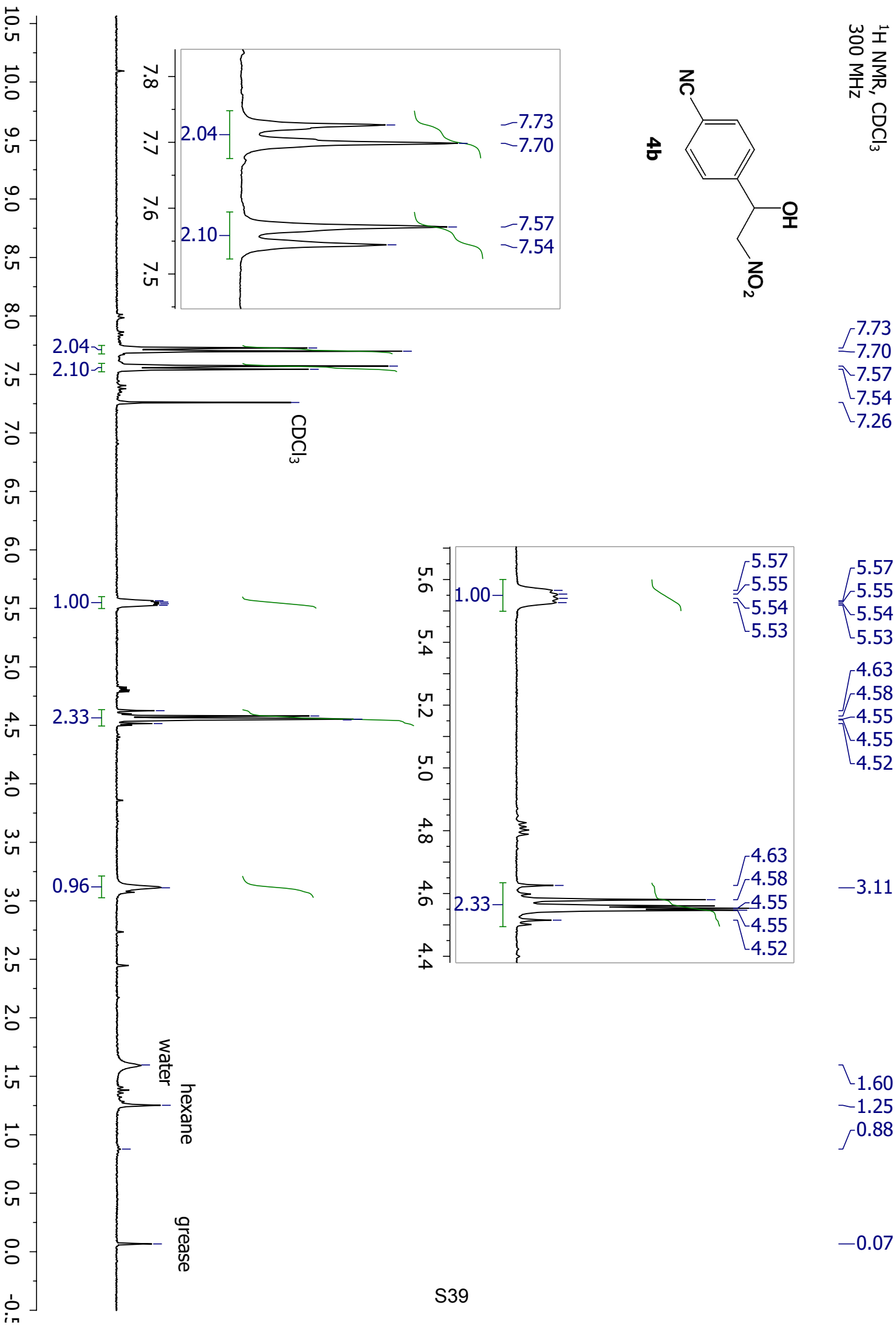
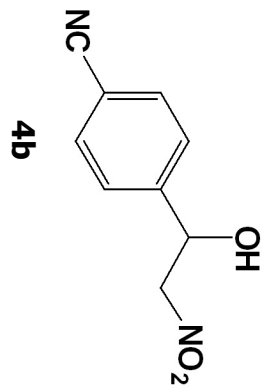
— 21.8

190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0

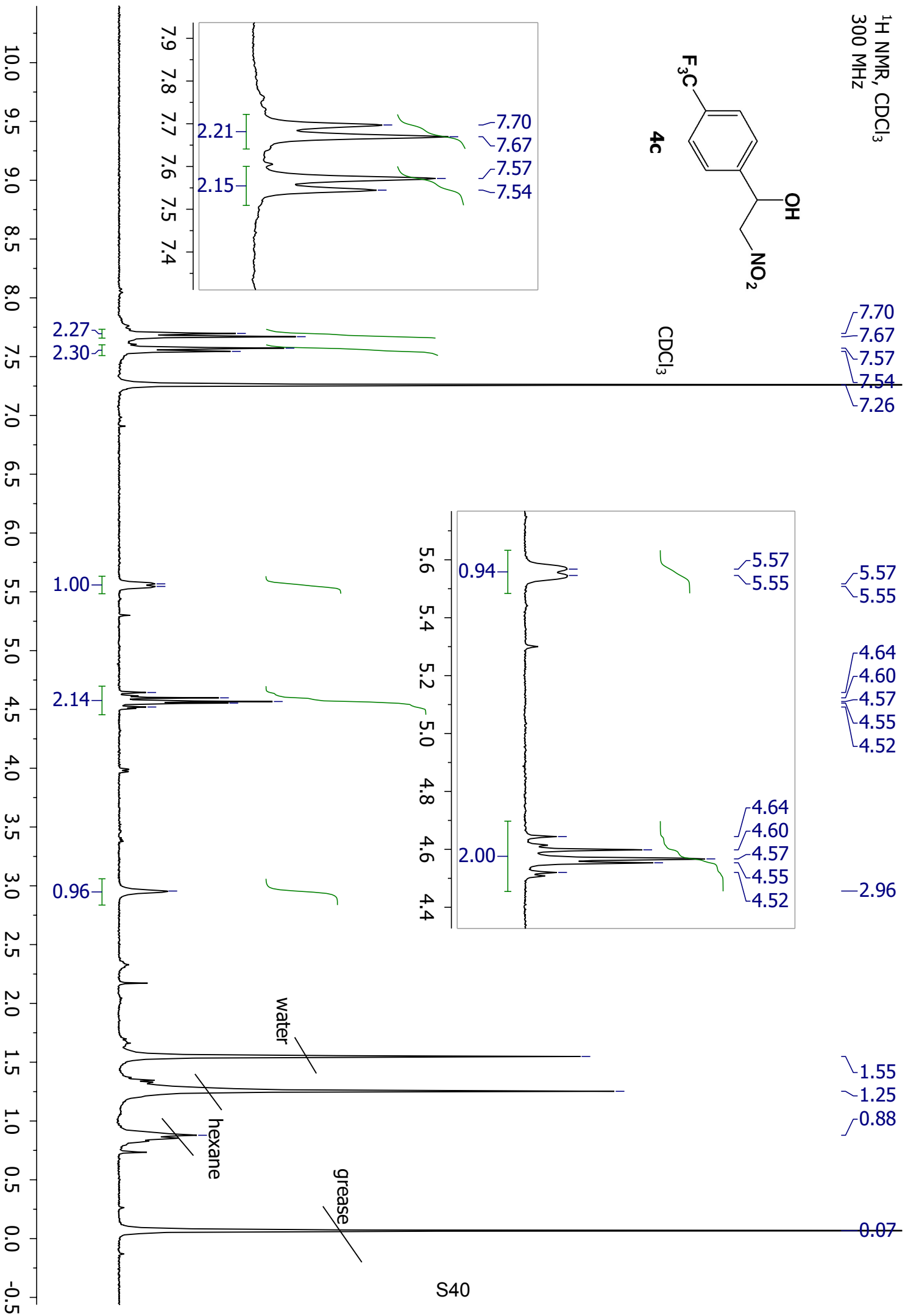
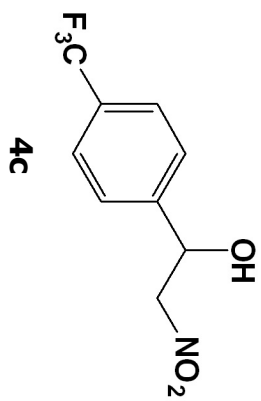
¹H NMR, CDCl₃
400 MHz



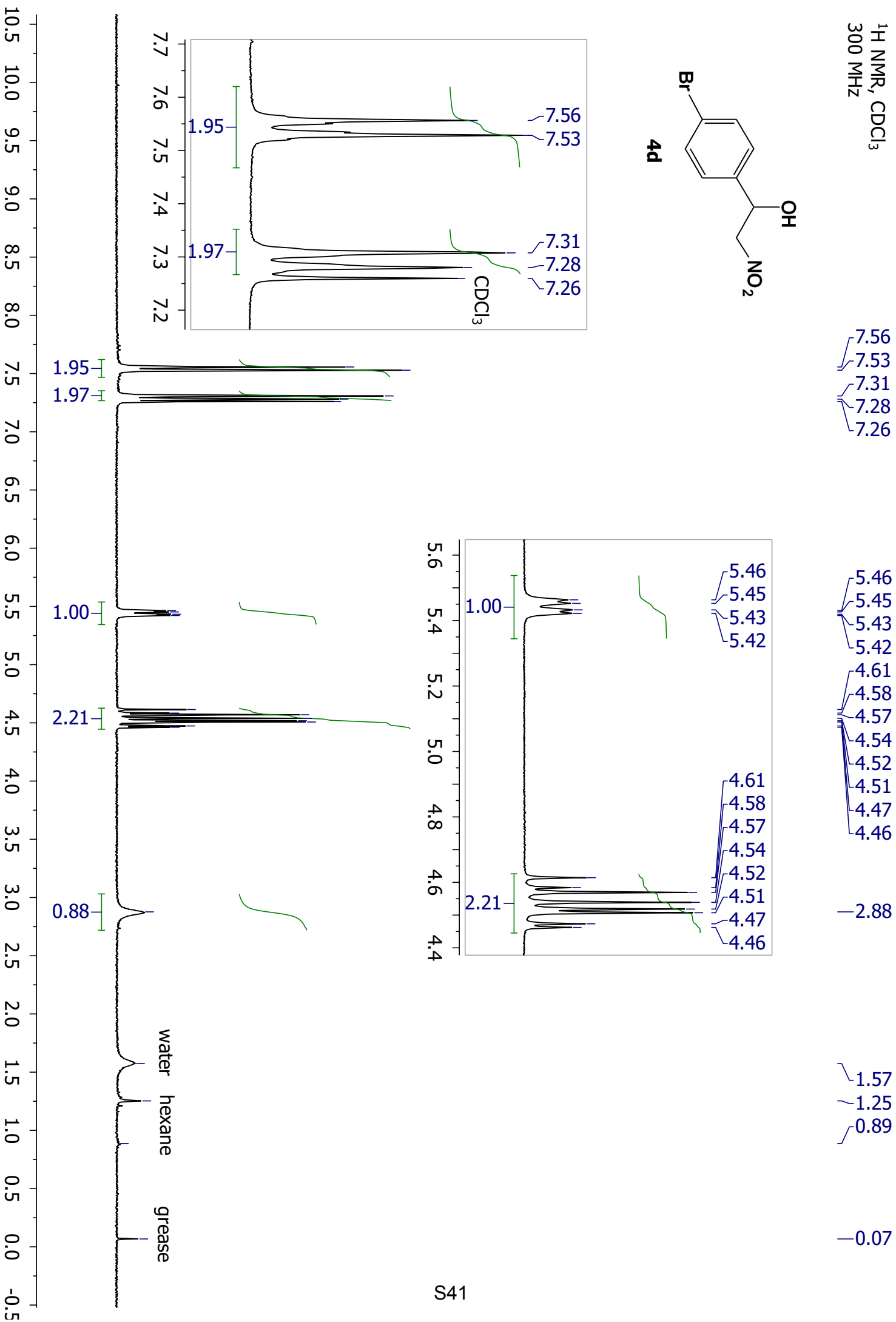
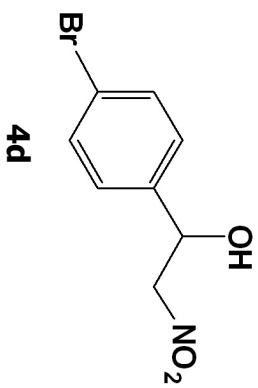
¹H NMR, CDCl₃
300 MHz



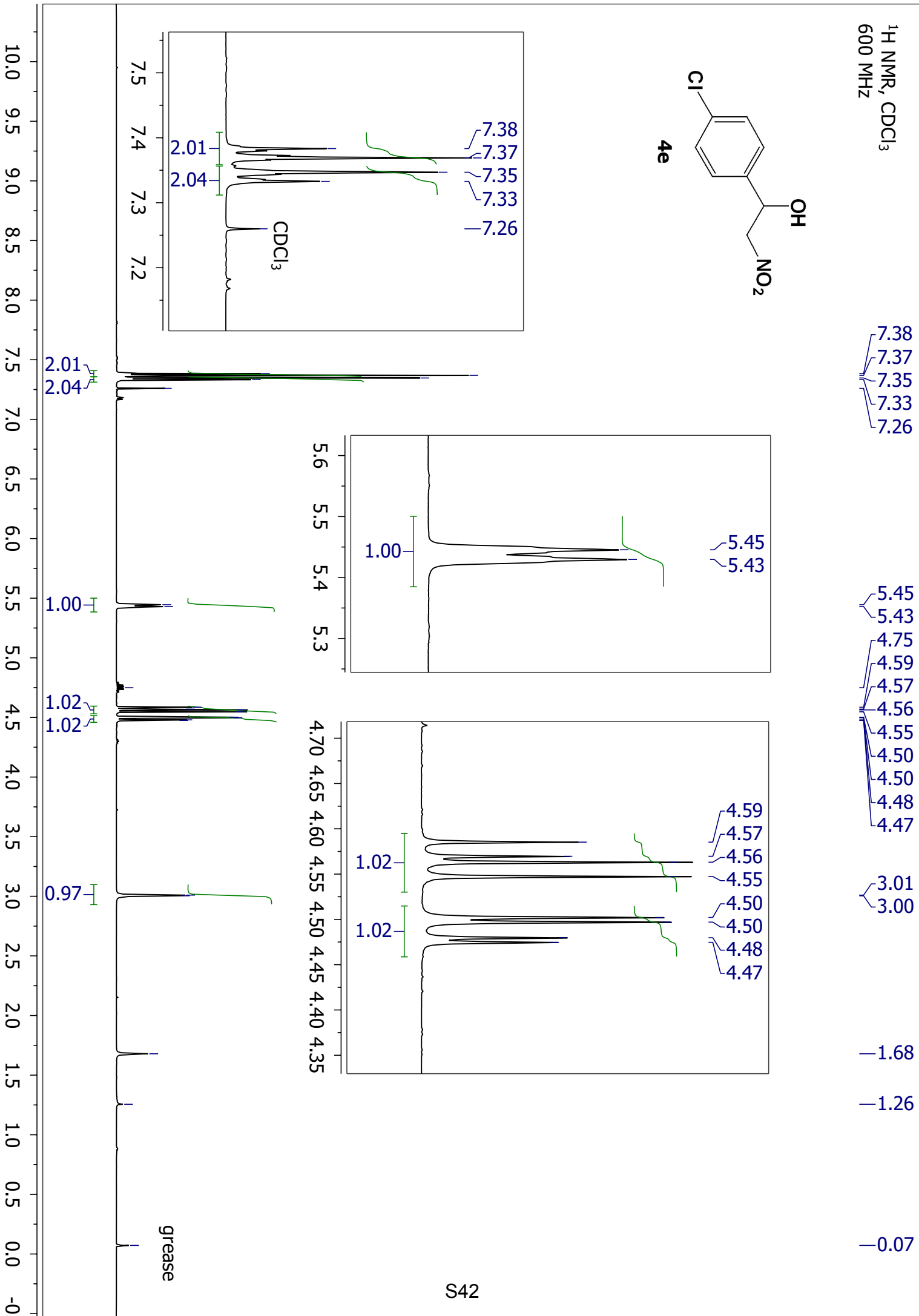
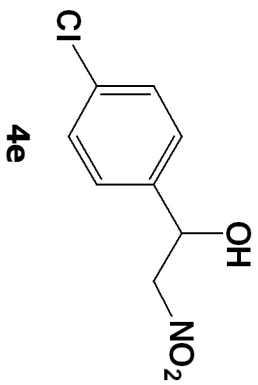
¹H NMR, CDCl₃
300 MHz



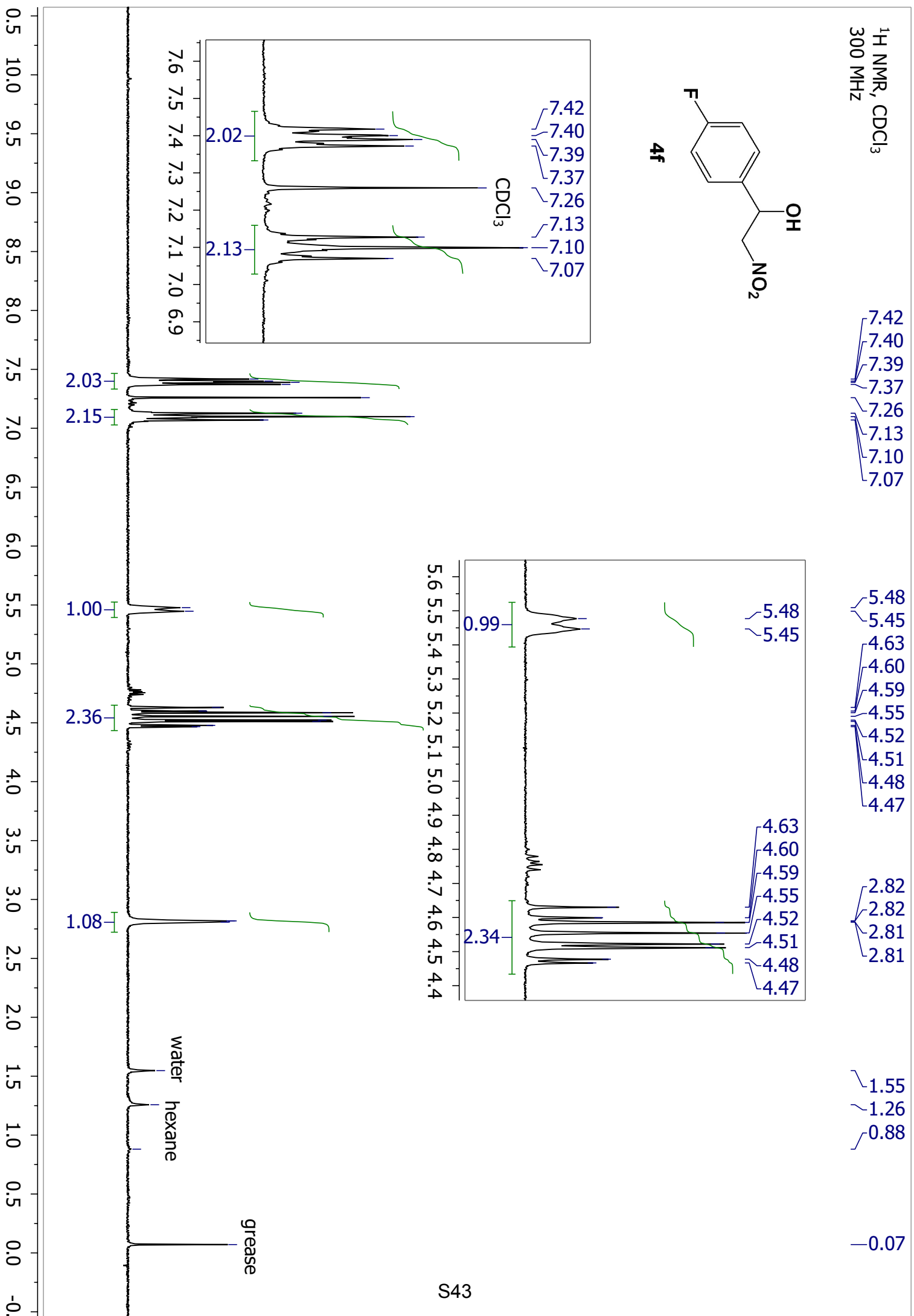
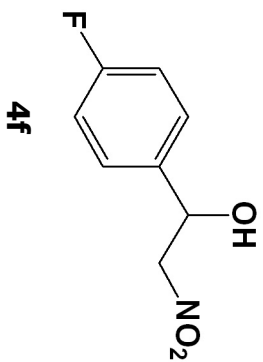
¹H NMR, CDCl₃
300 MHz



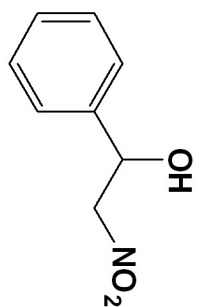
¹H NMR, CDCl₃
600 MHz



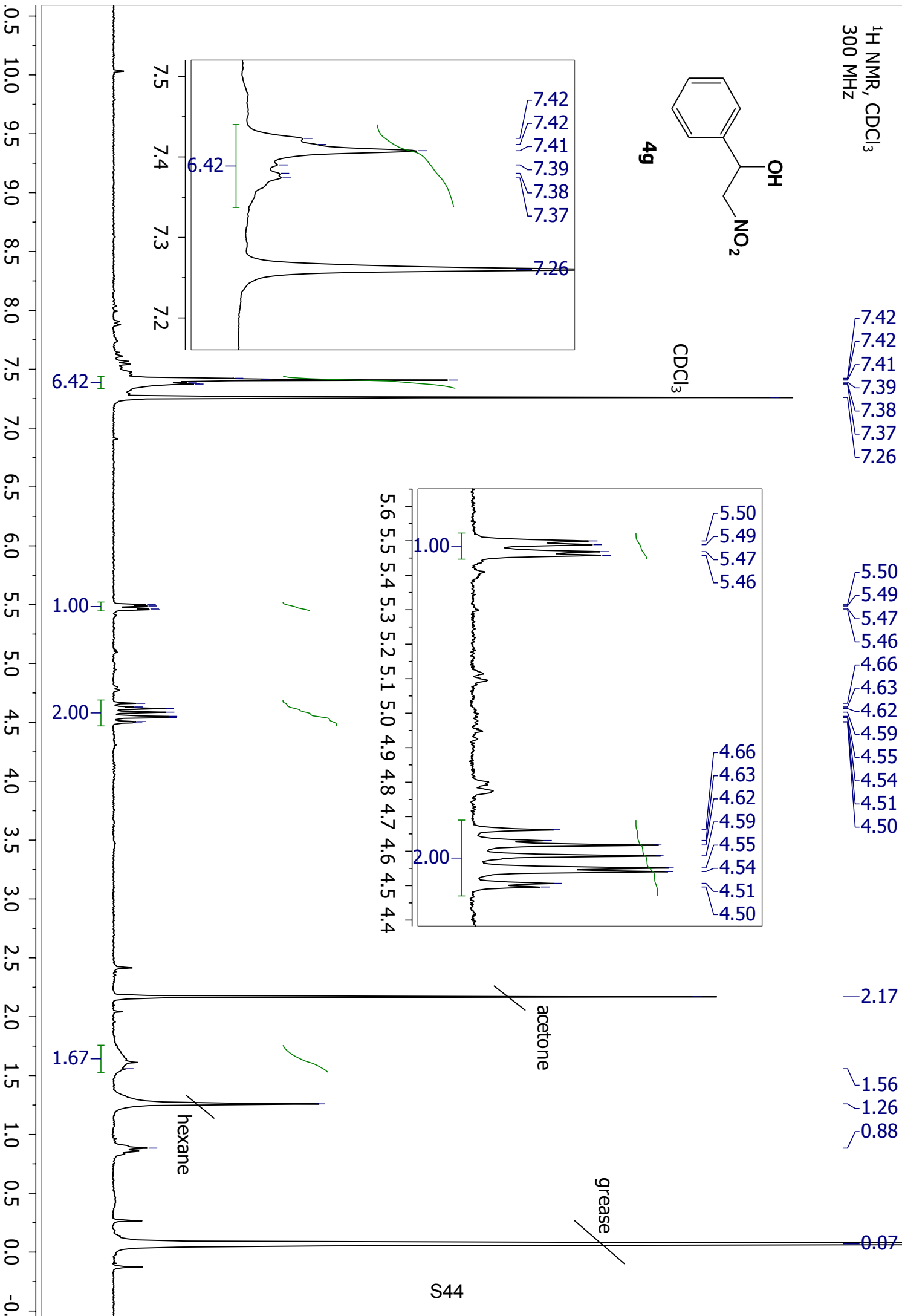
¹H NMR, CDCl₃
300 MHz



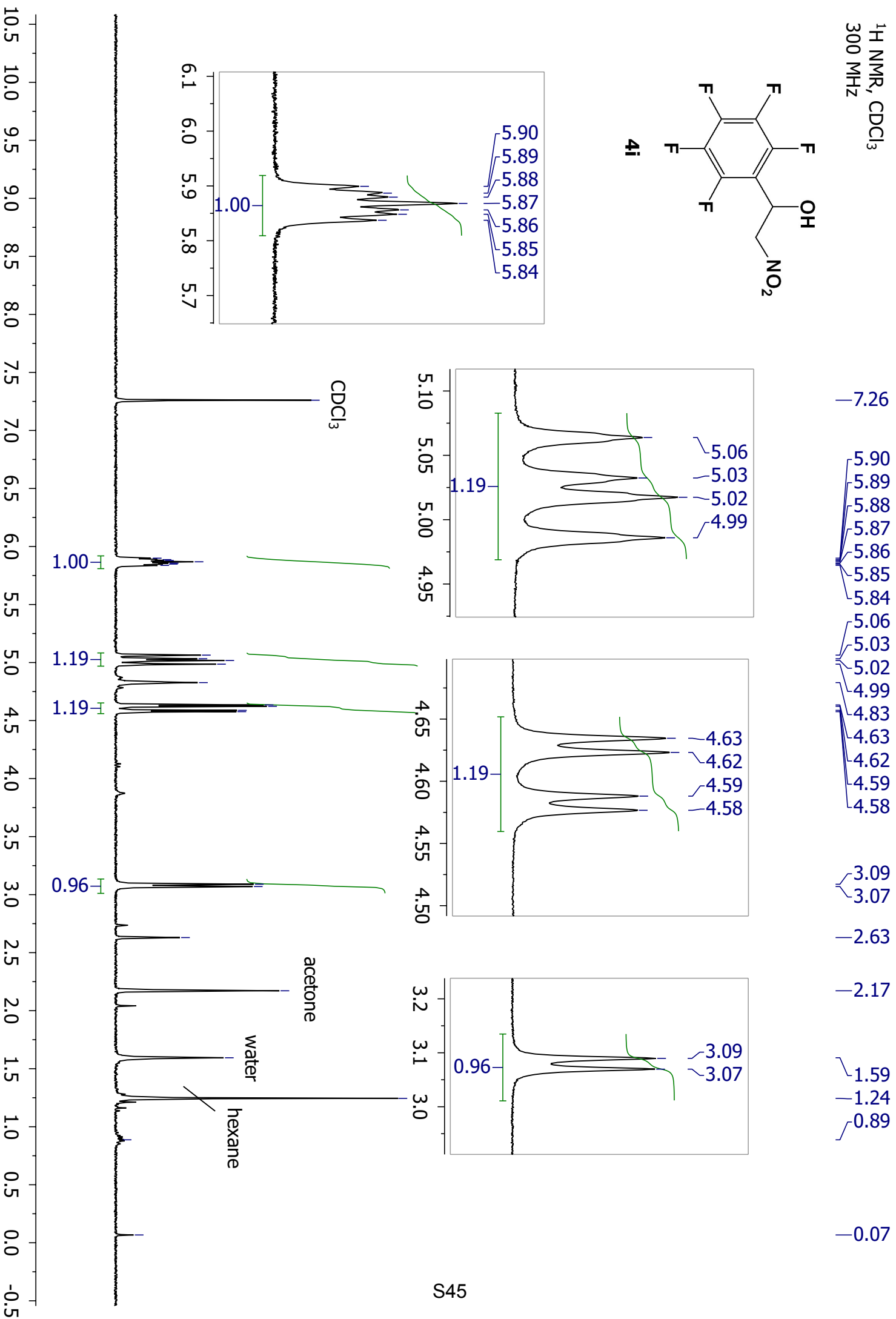
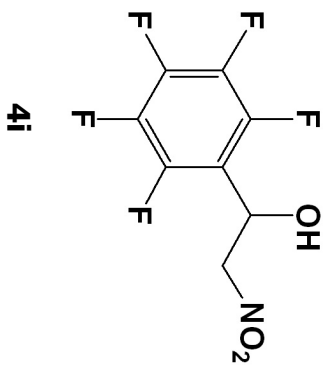
¹H NMR, CDCl₃
300 MHz



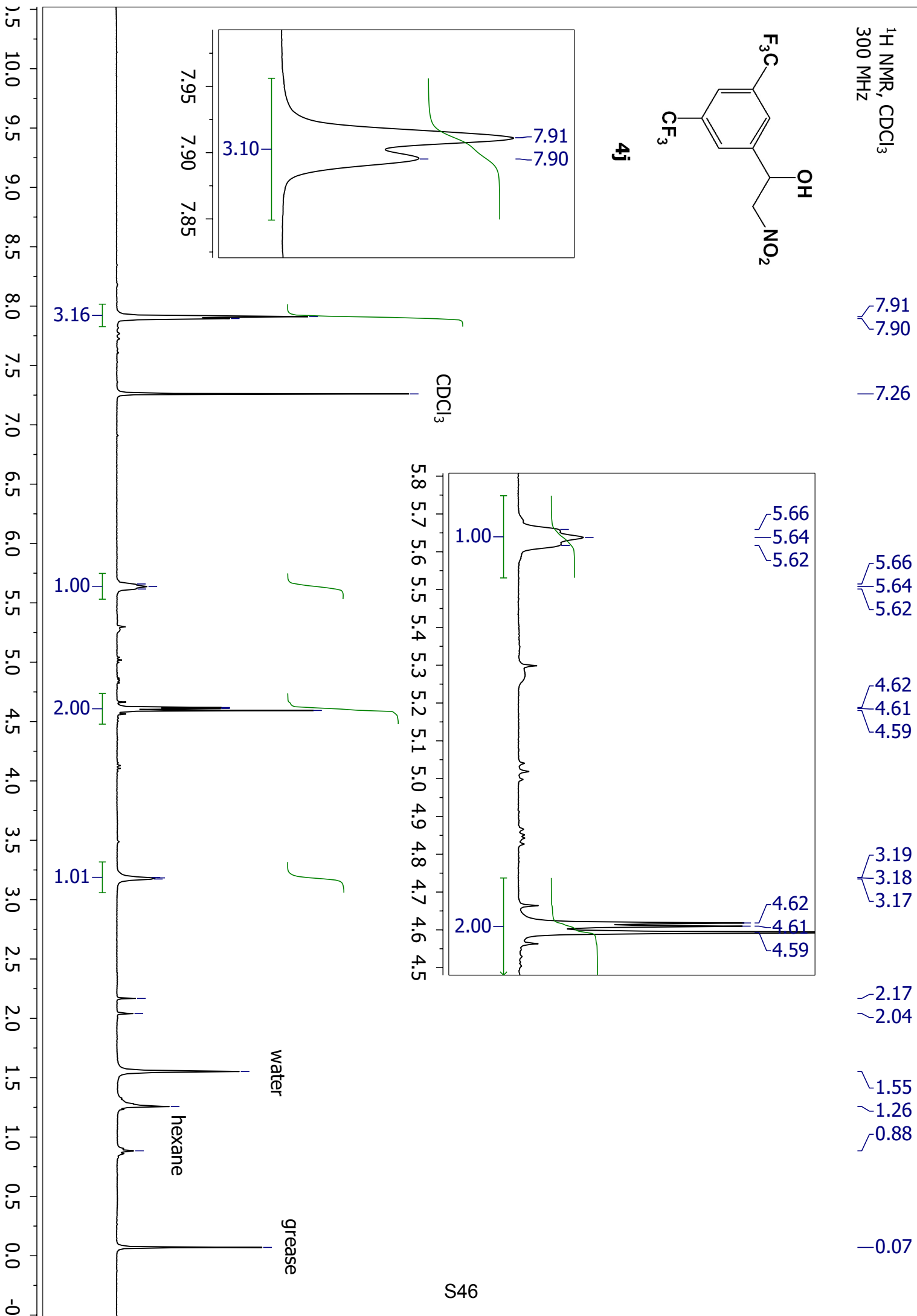
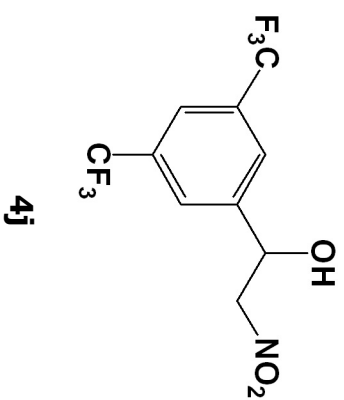
CDCl₃



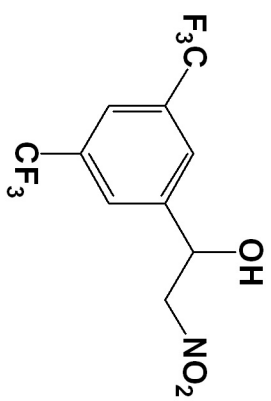
¹H NMR, CDCl₃
300 MHz



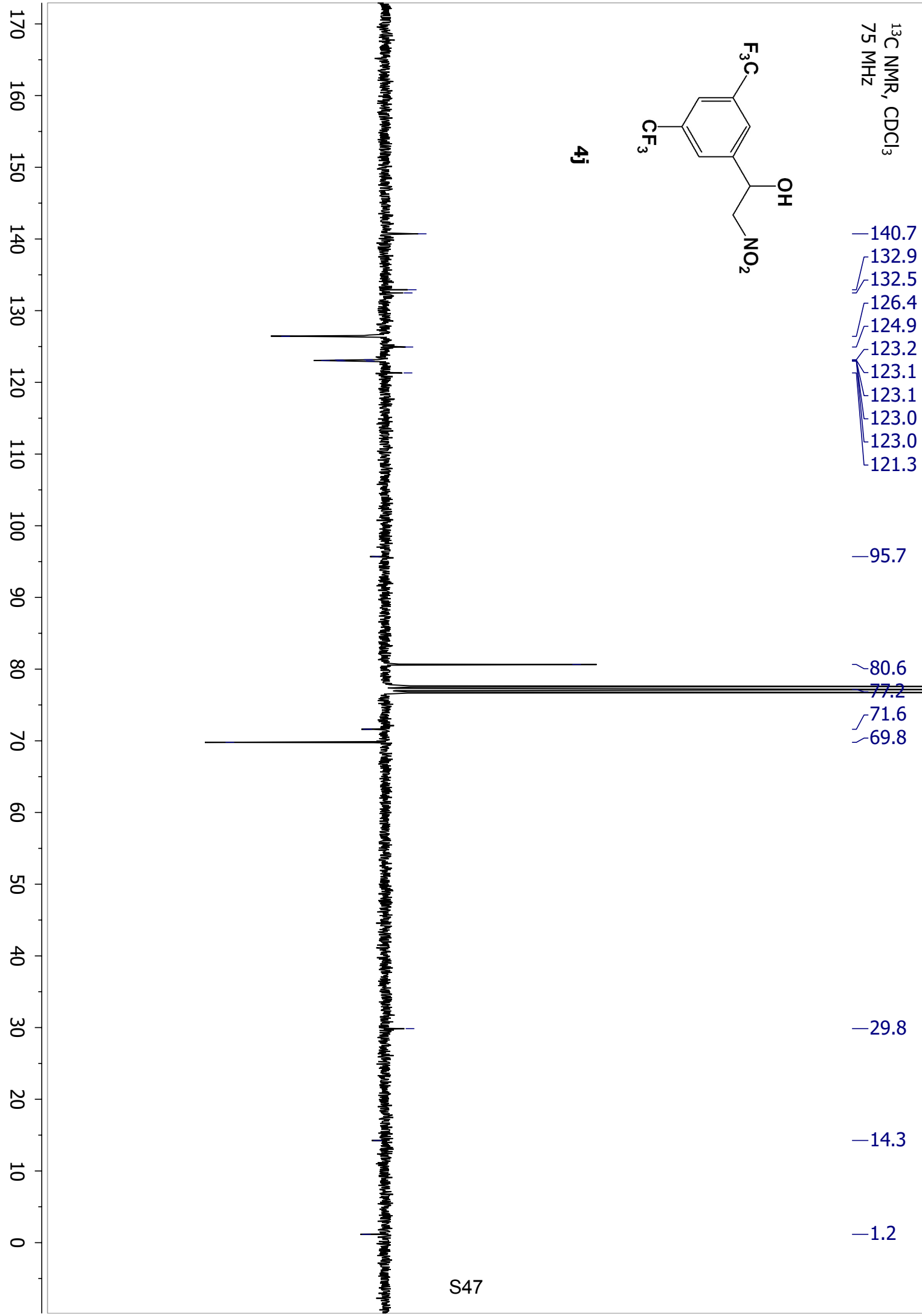
¹H NMR, CDCl₃
300 MHz



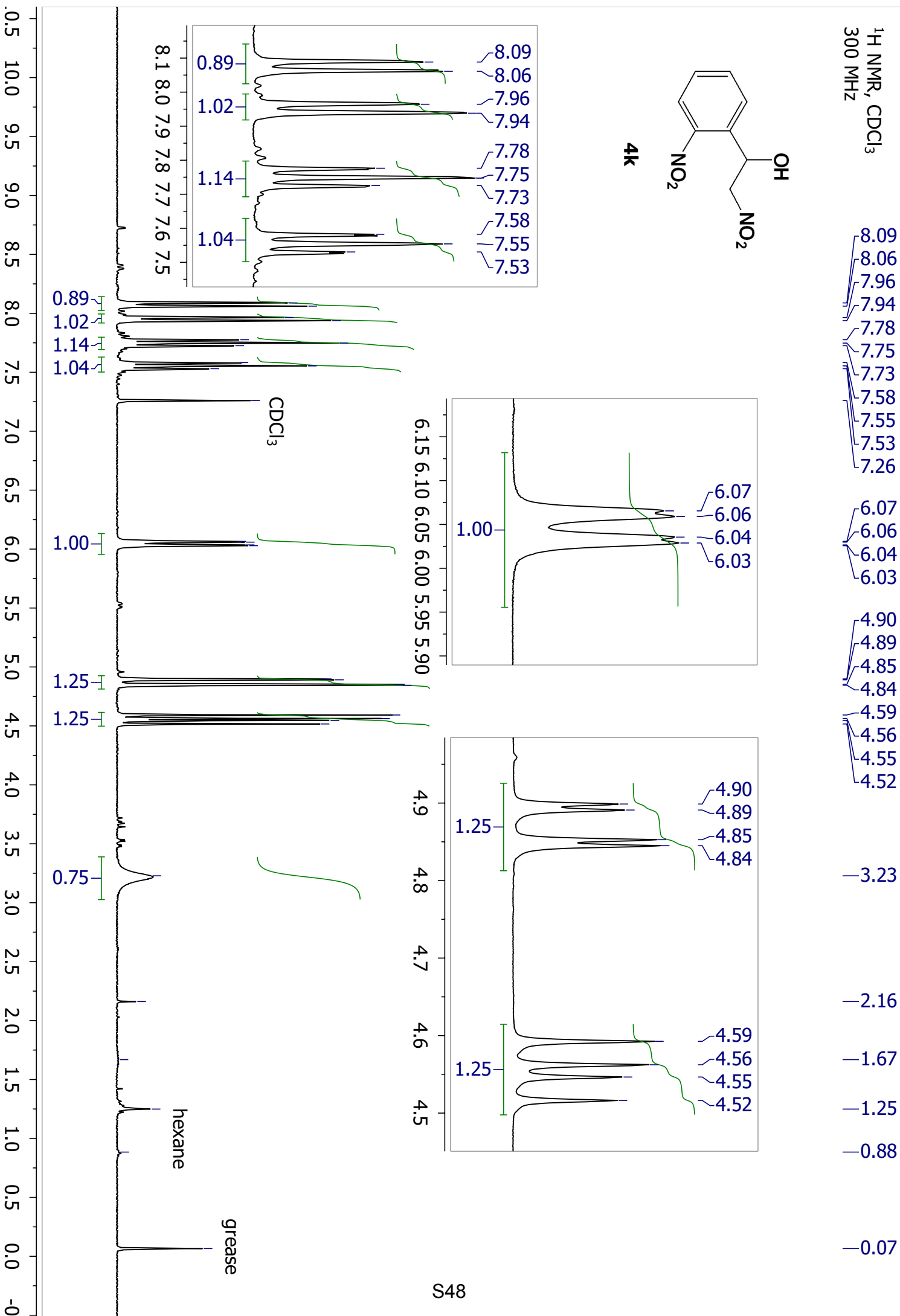
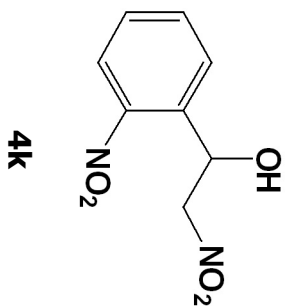
¹³C NMR, CDCl₃
75 MHz



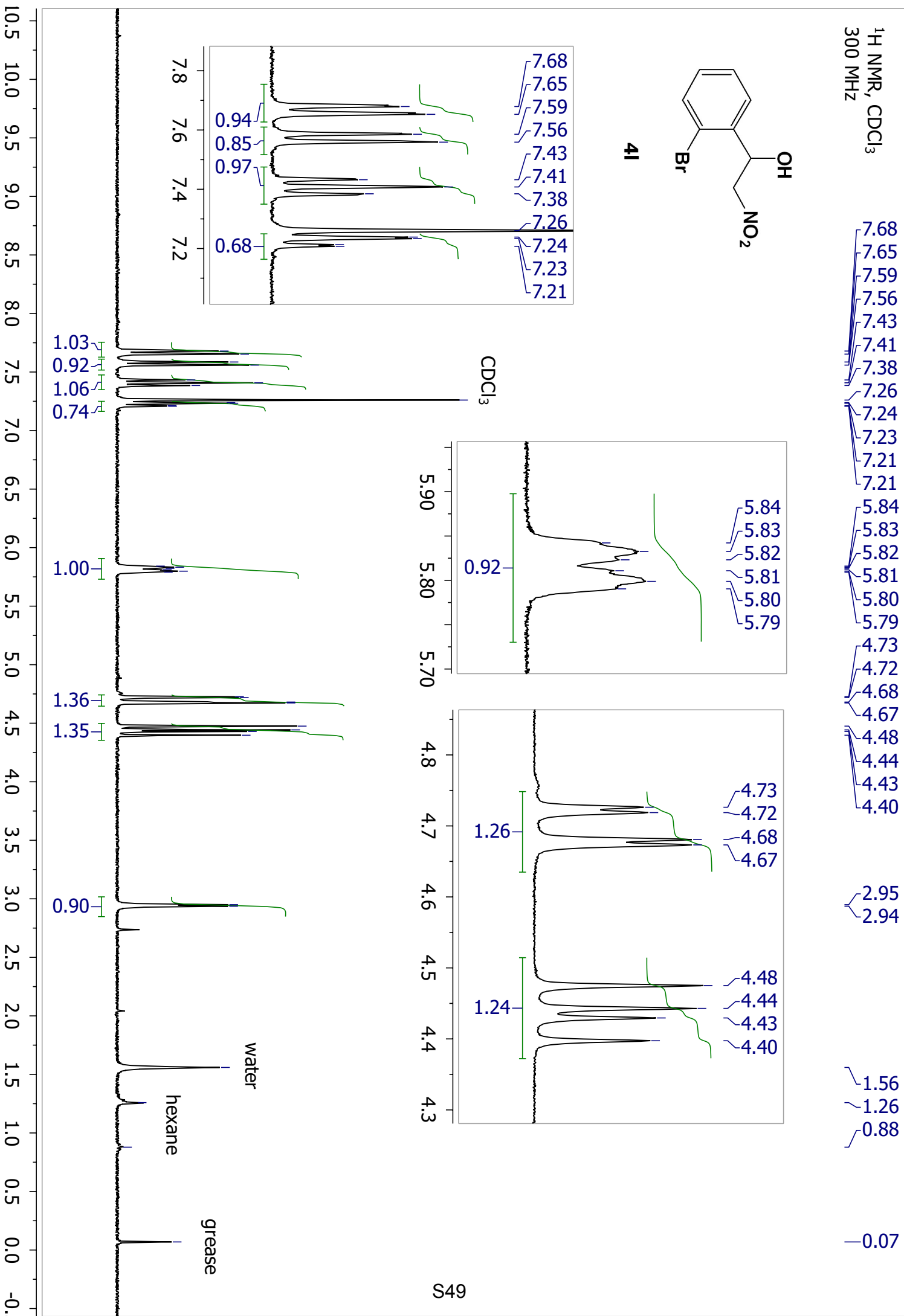
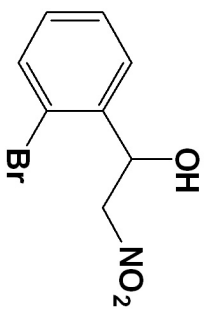
4j



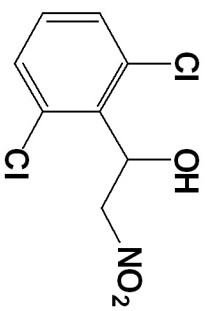
¹H NMR, CDCl₃
300 MHz



¹H NMR, CDCl₃
300 MHz

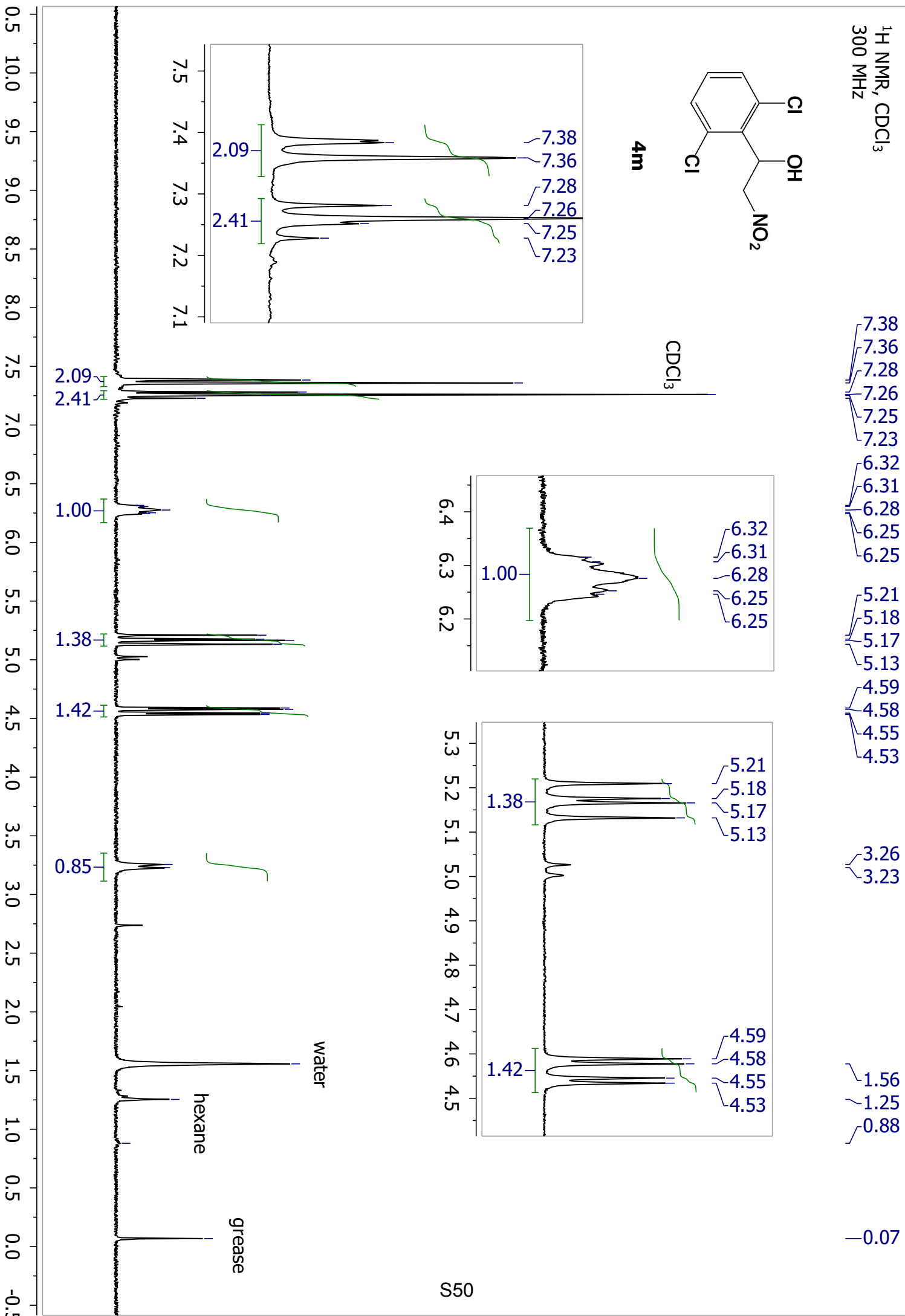


¹H NMR, CDCl₃
300 MHz

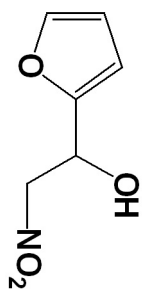


4m

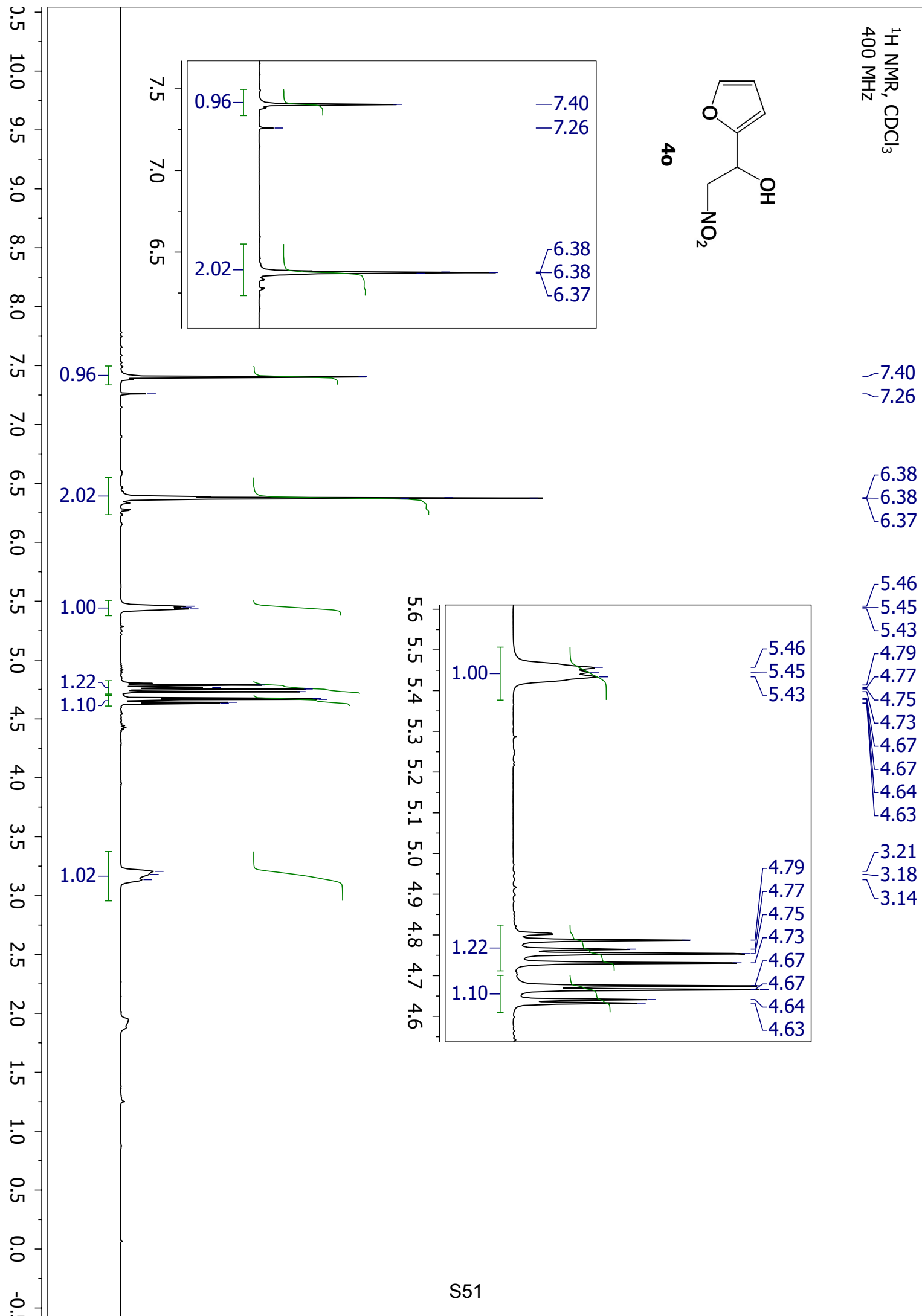
CDCl₃



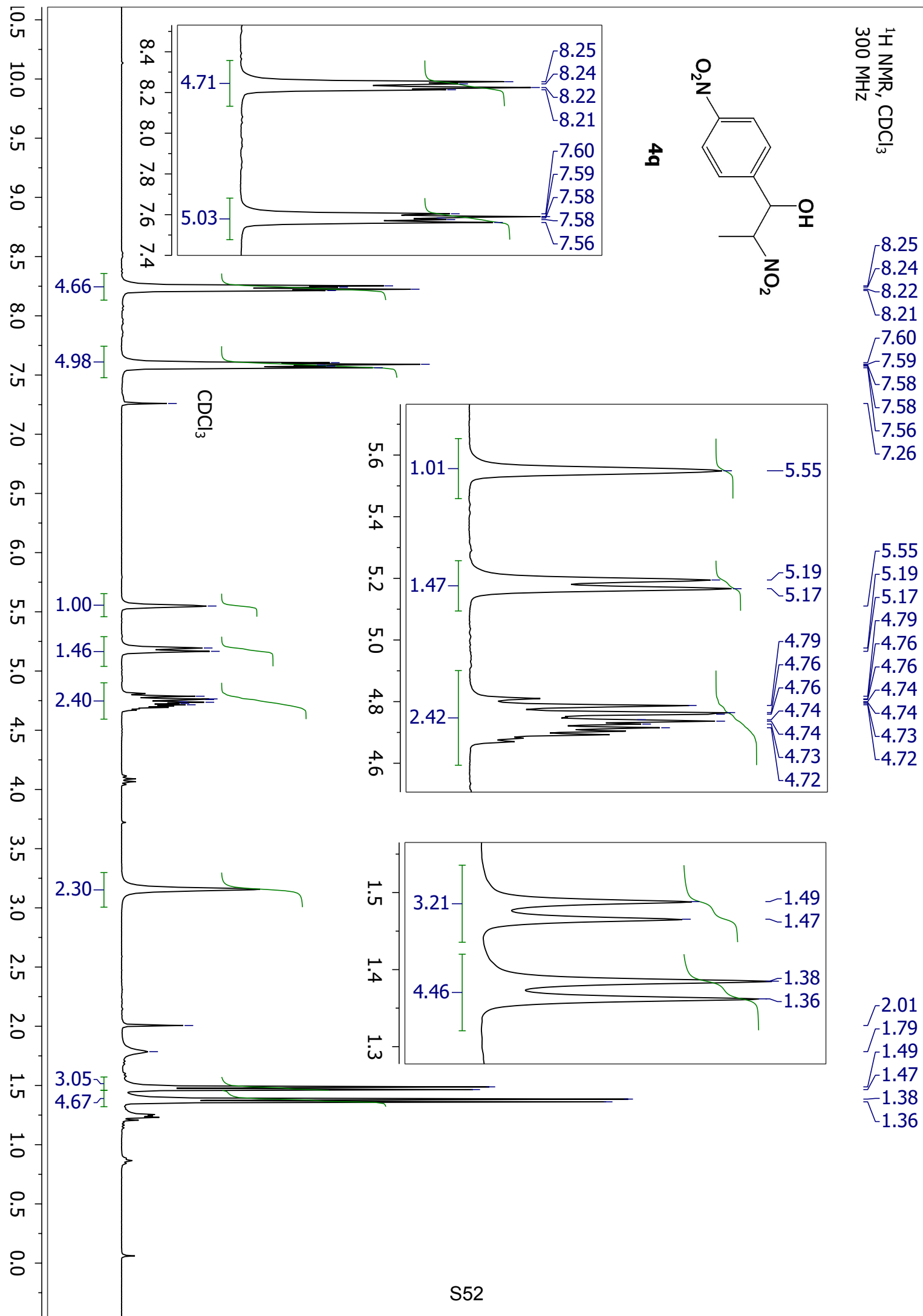
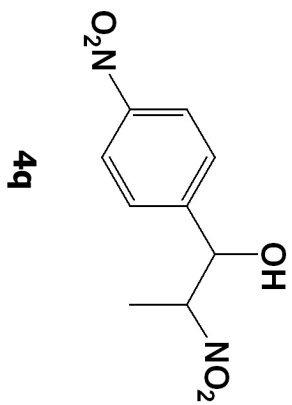
¹H NMR, CDCl₃
400 MHz



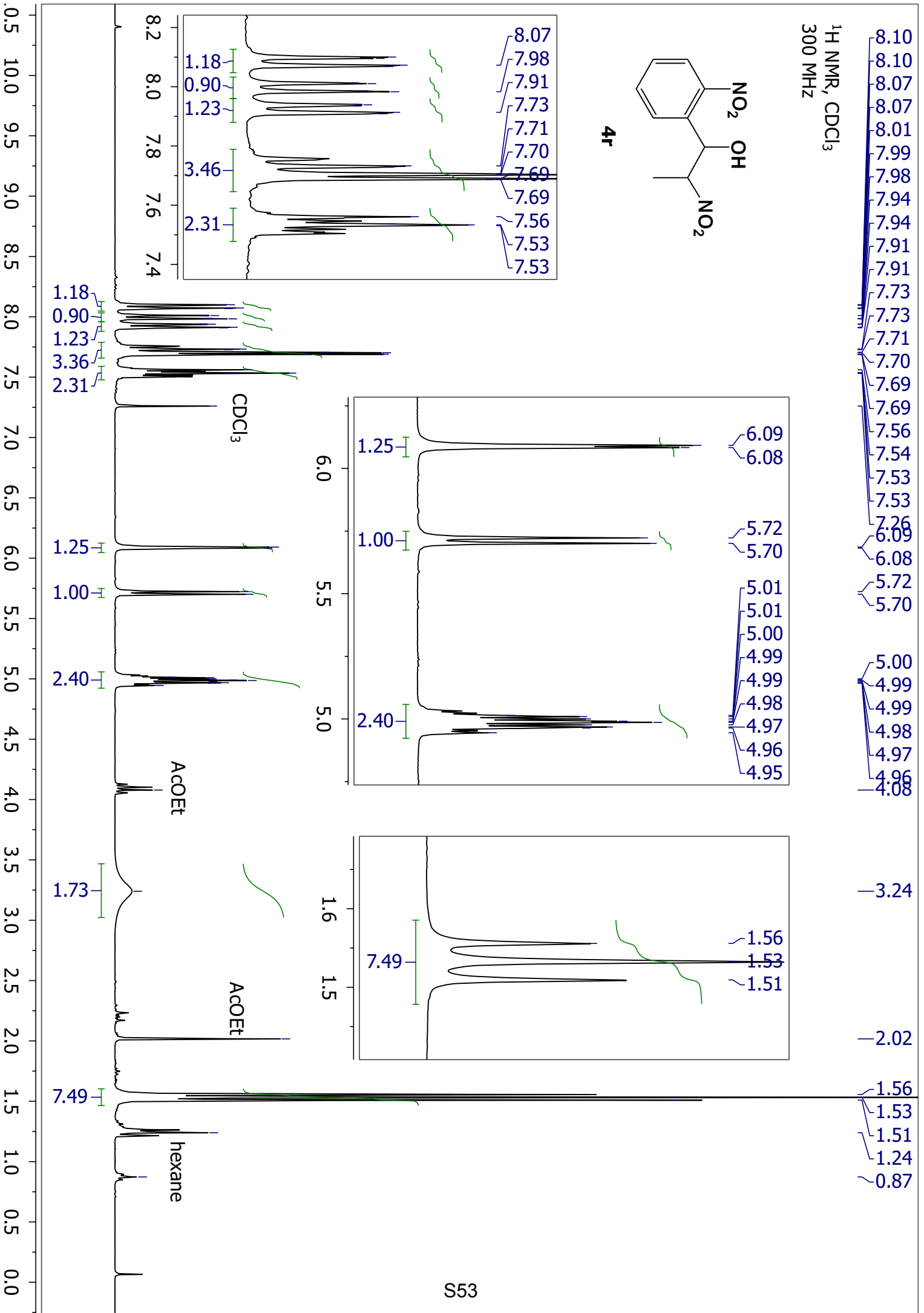
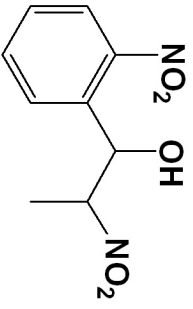
40



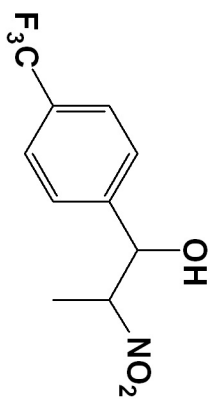
¹H NMR, CDCl₃
300 MHz



¹H NMR, CDCl₃
300 MHz



¹H NMR, CDCl₃
600 MHz



CDCl₃

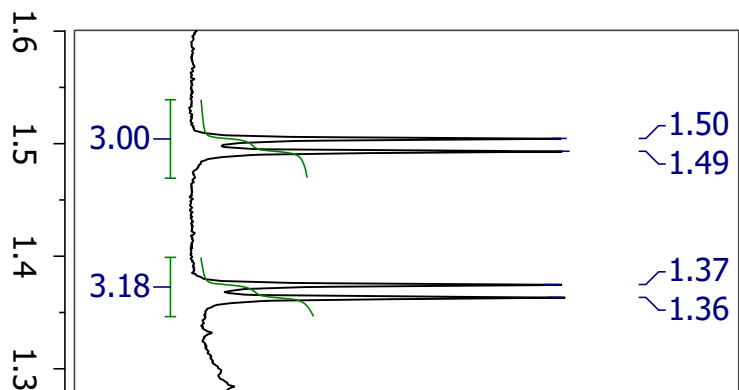
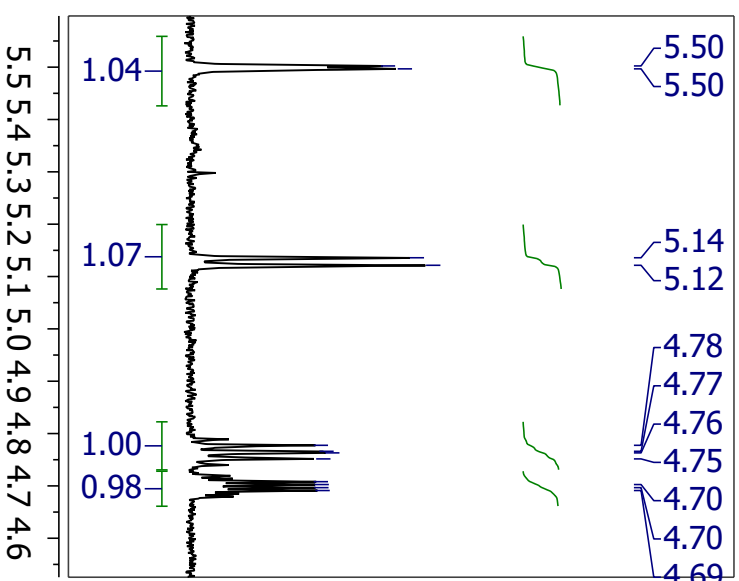
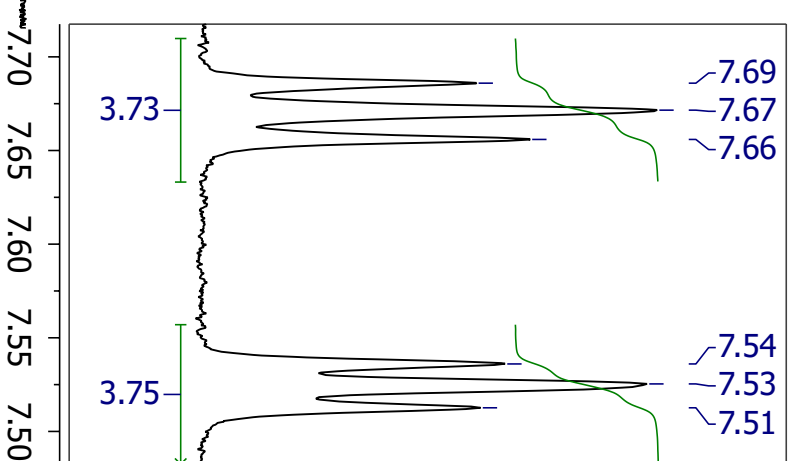
7.69
7.67
7.66
7.54
7.53
7.51
7.26

5.50
5.50
5.14
5.12
4.78
4.77
4.76
4.75
4.71
4.70
4.70
4.69

1.50
1.49
1.37
1.36
1.26
0.88

0.07

grease



3.73
3.75

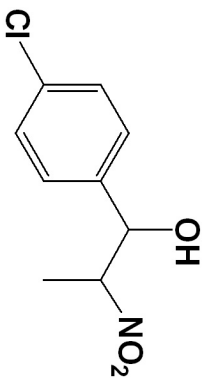
1.04
1.07
1.00
0.98

3.00
3.18

hexane

S54

¹H NMR, CDCl₃
300 MHz

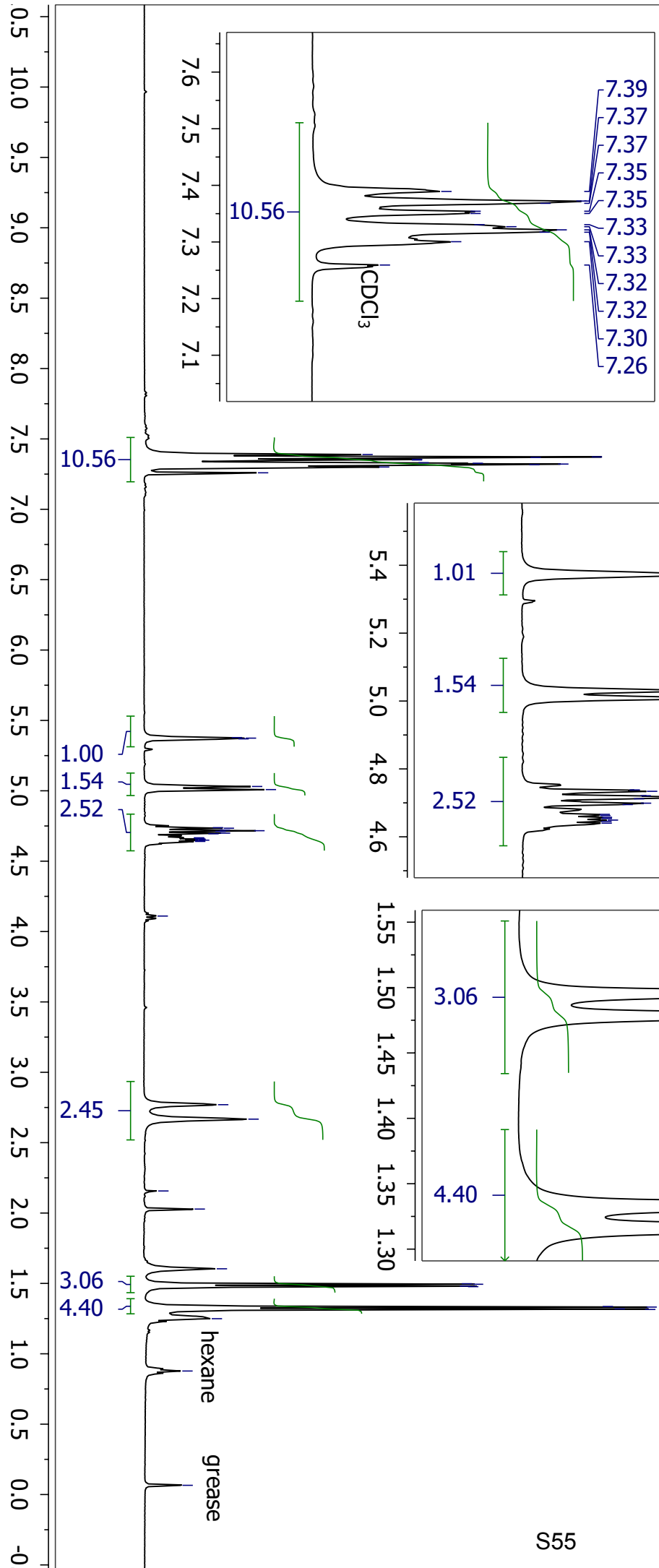


7.39
7.37
7.37
7.35
7.35
7.33
7.33
7.32
7.32
7.30
7.26

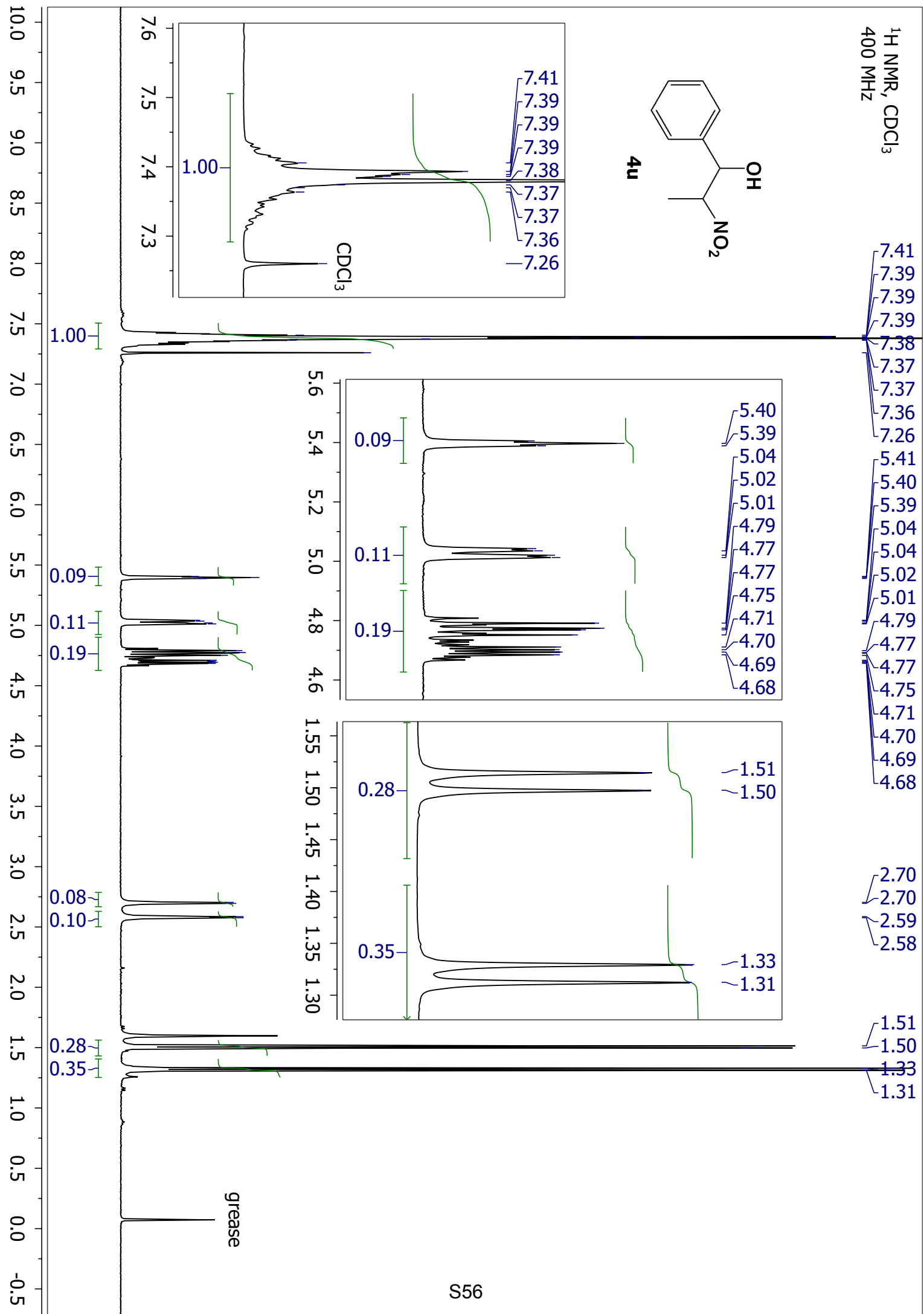
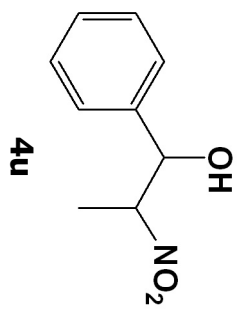
5.38
5.37
5.37
5.03
5.01
4.74
4.73
4.72
4.72
4.71
4.70
4.69
4.65

2.77
2.67
2.16
2.03

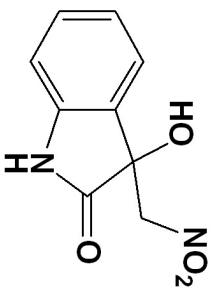
1.50
1.49
1.48
1.48
1.34
1.33
1.32
1.31
0.07



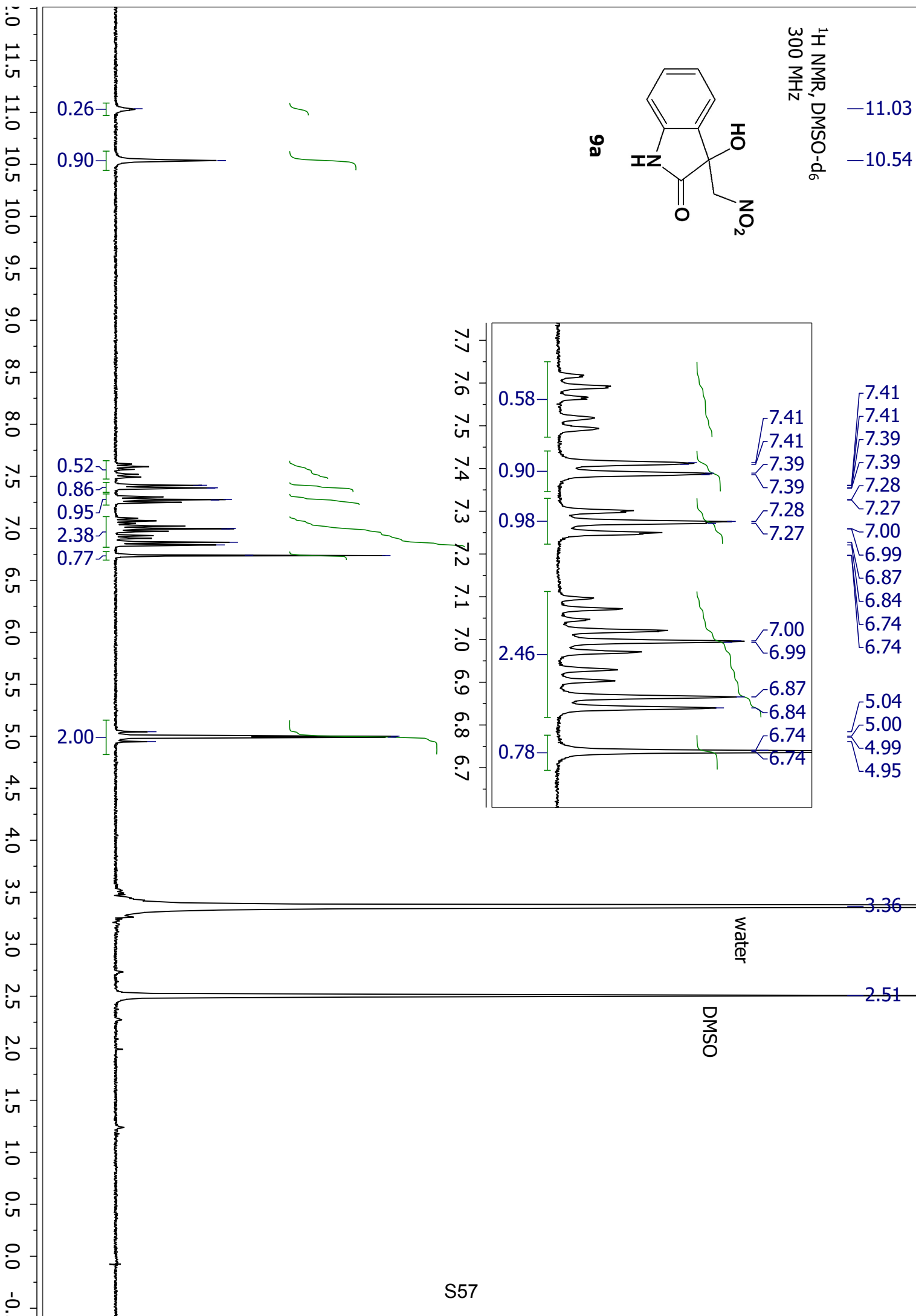
¹H NMR, CDCl₃
400 MHz

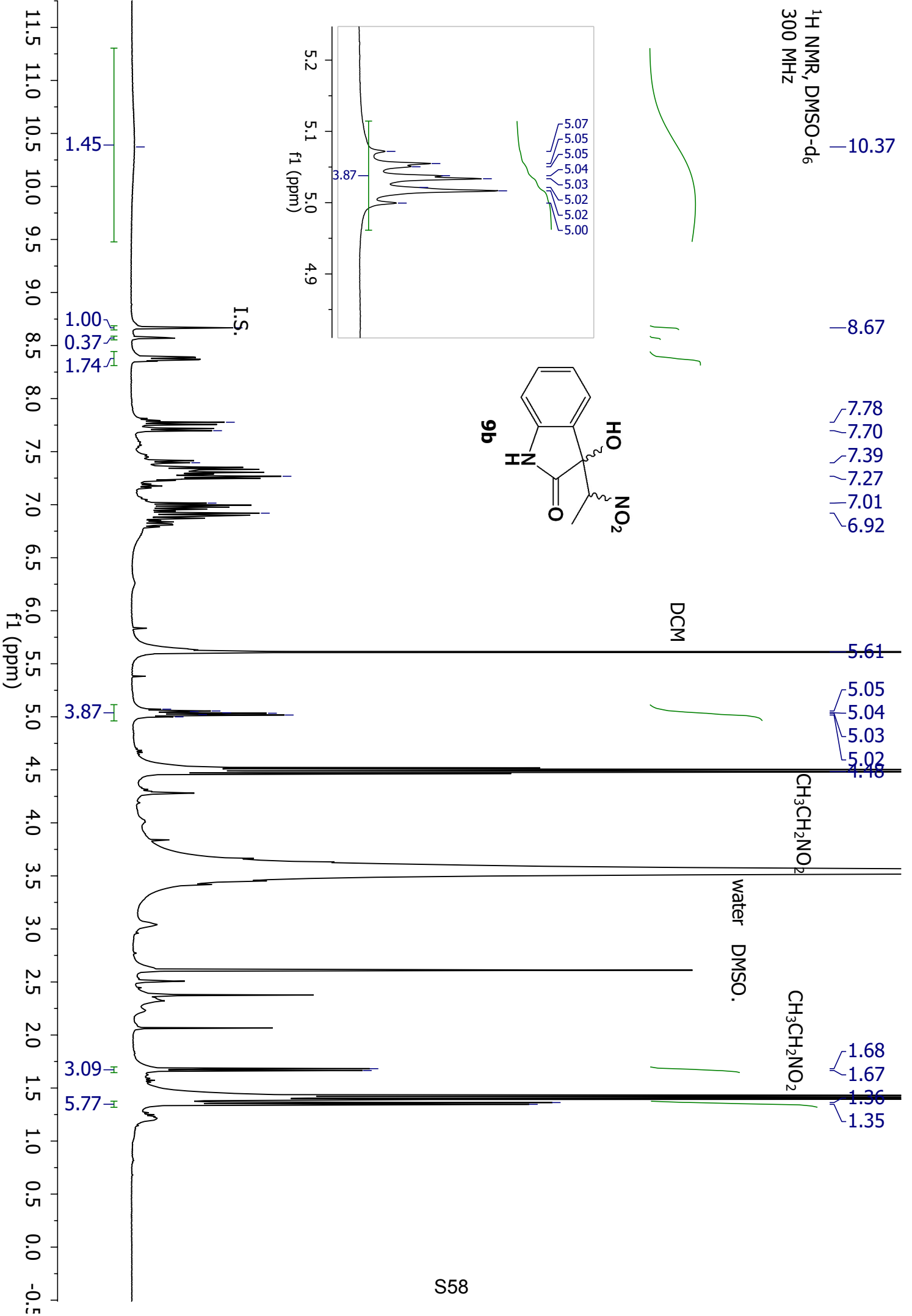


¹H NMR, DMSO-d₆
300 MHz

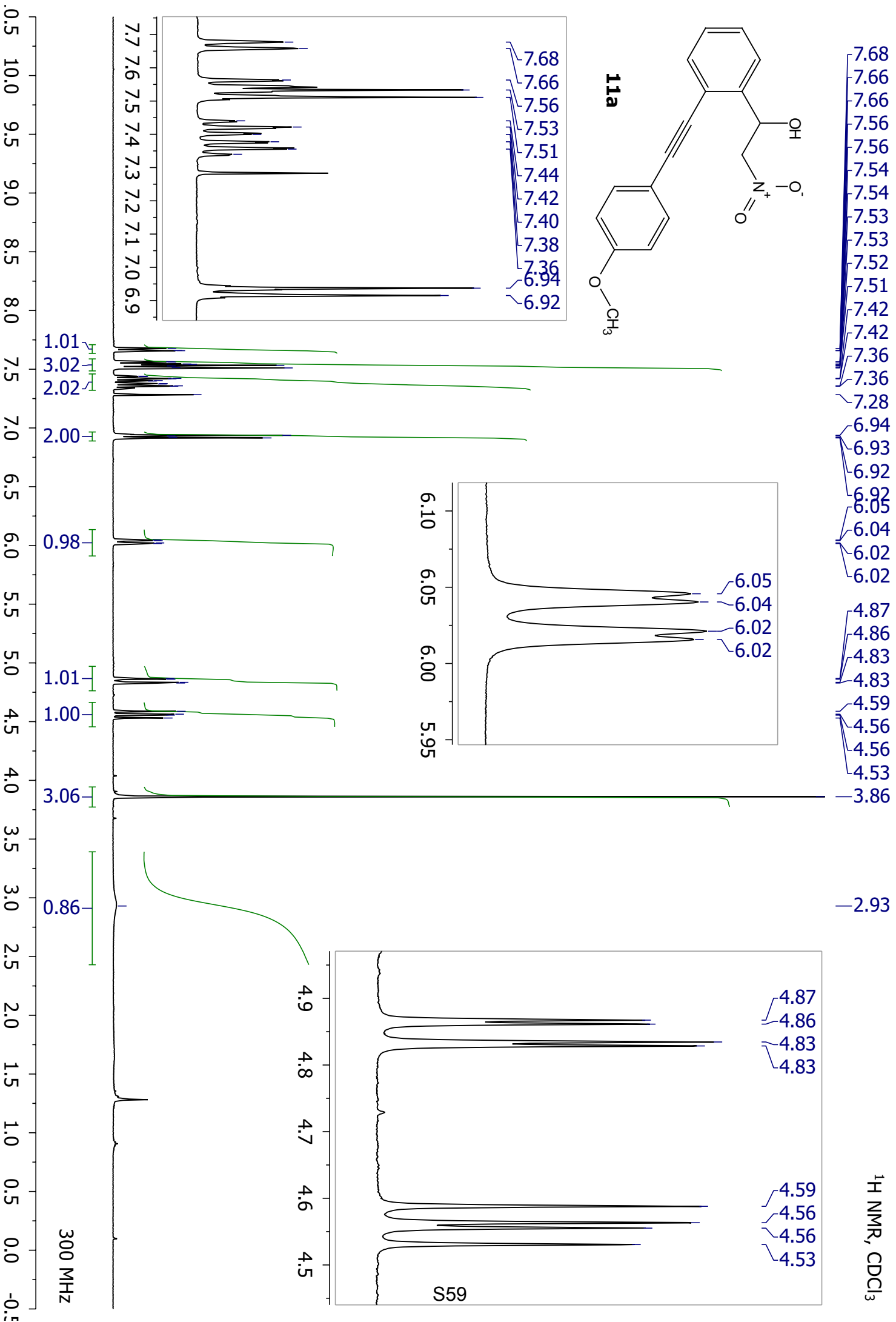


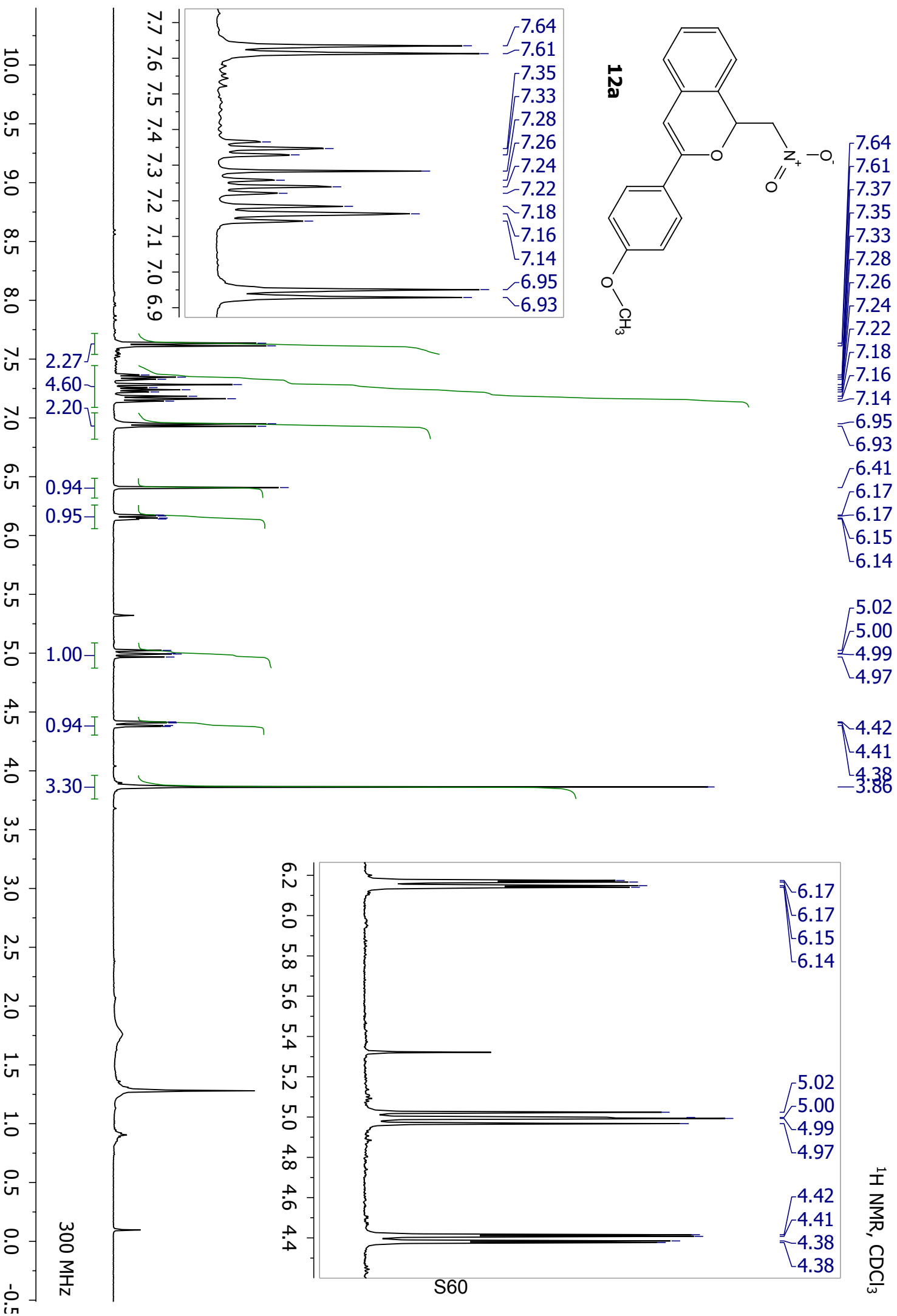
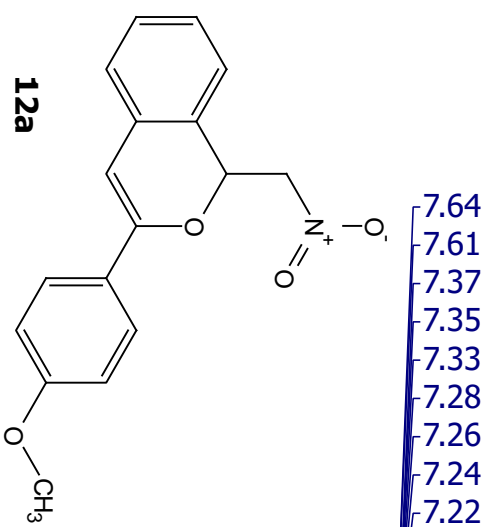
— 11.03
— 10.54

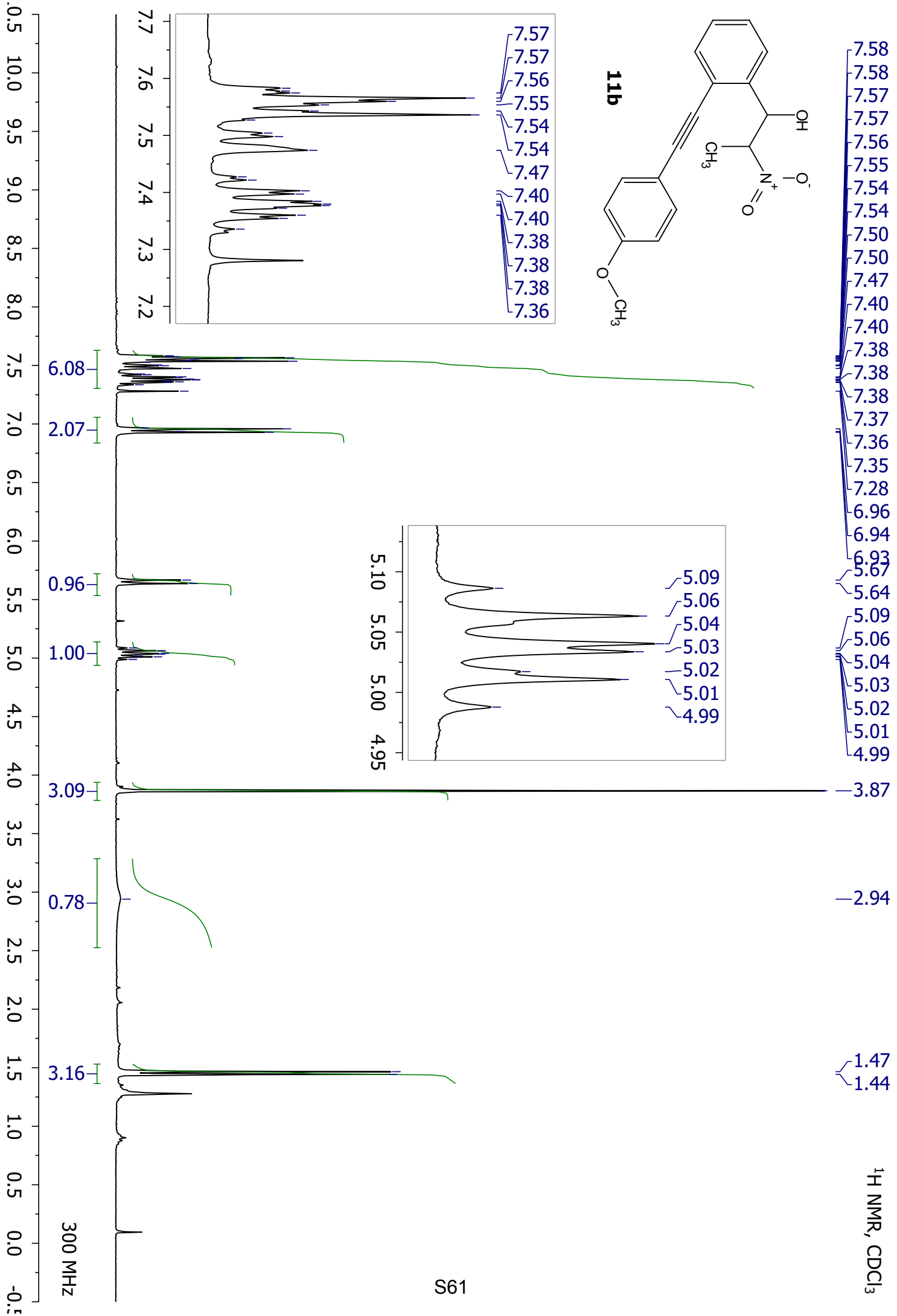
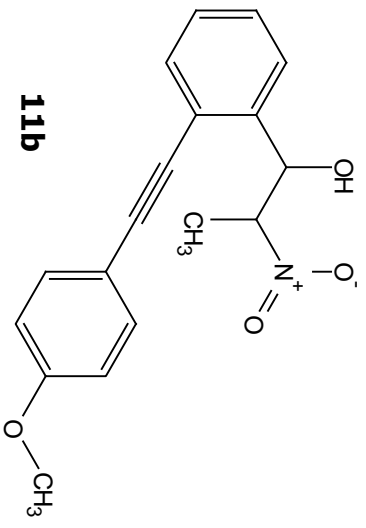




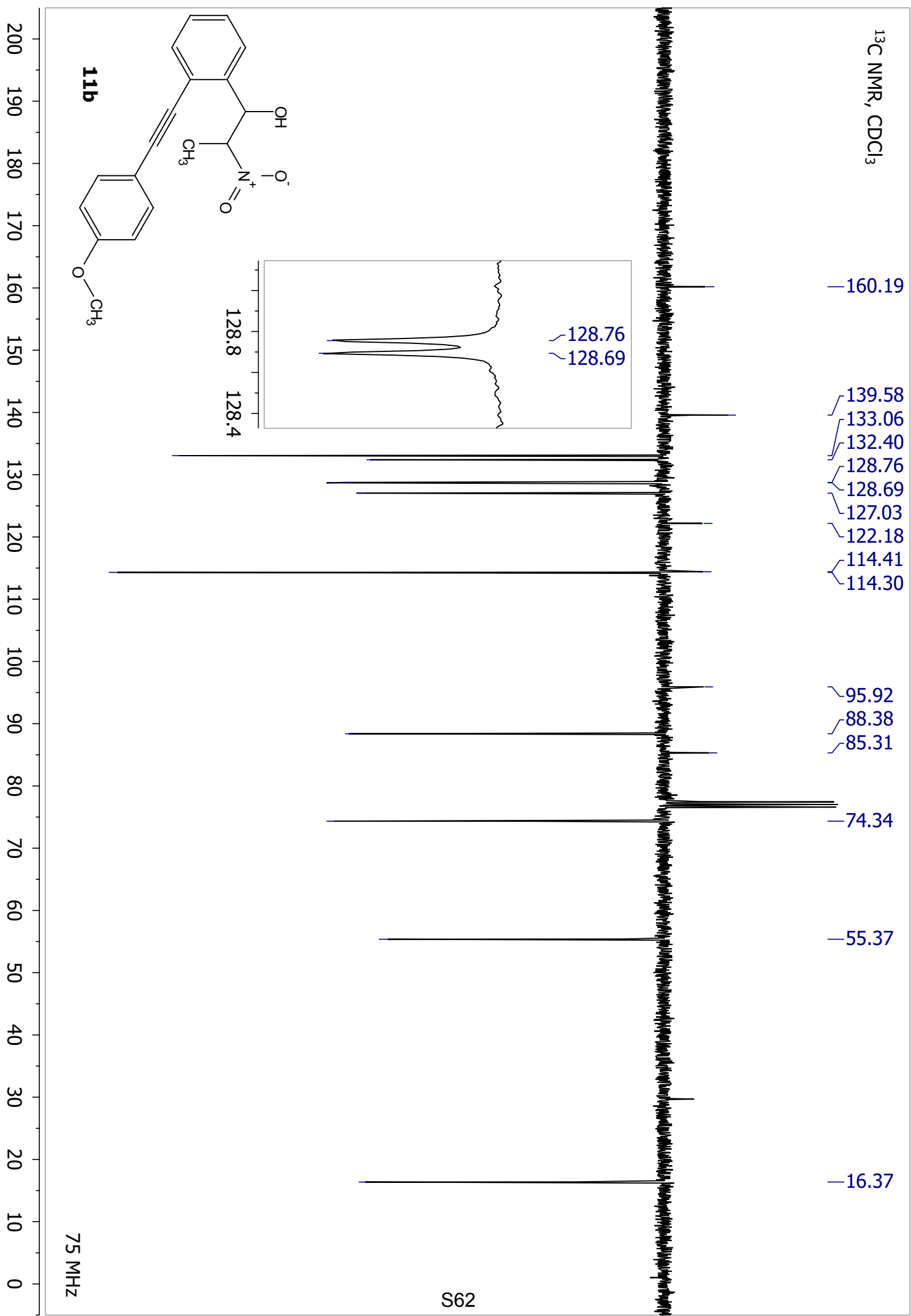
¹H NMR, CDCl₃





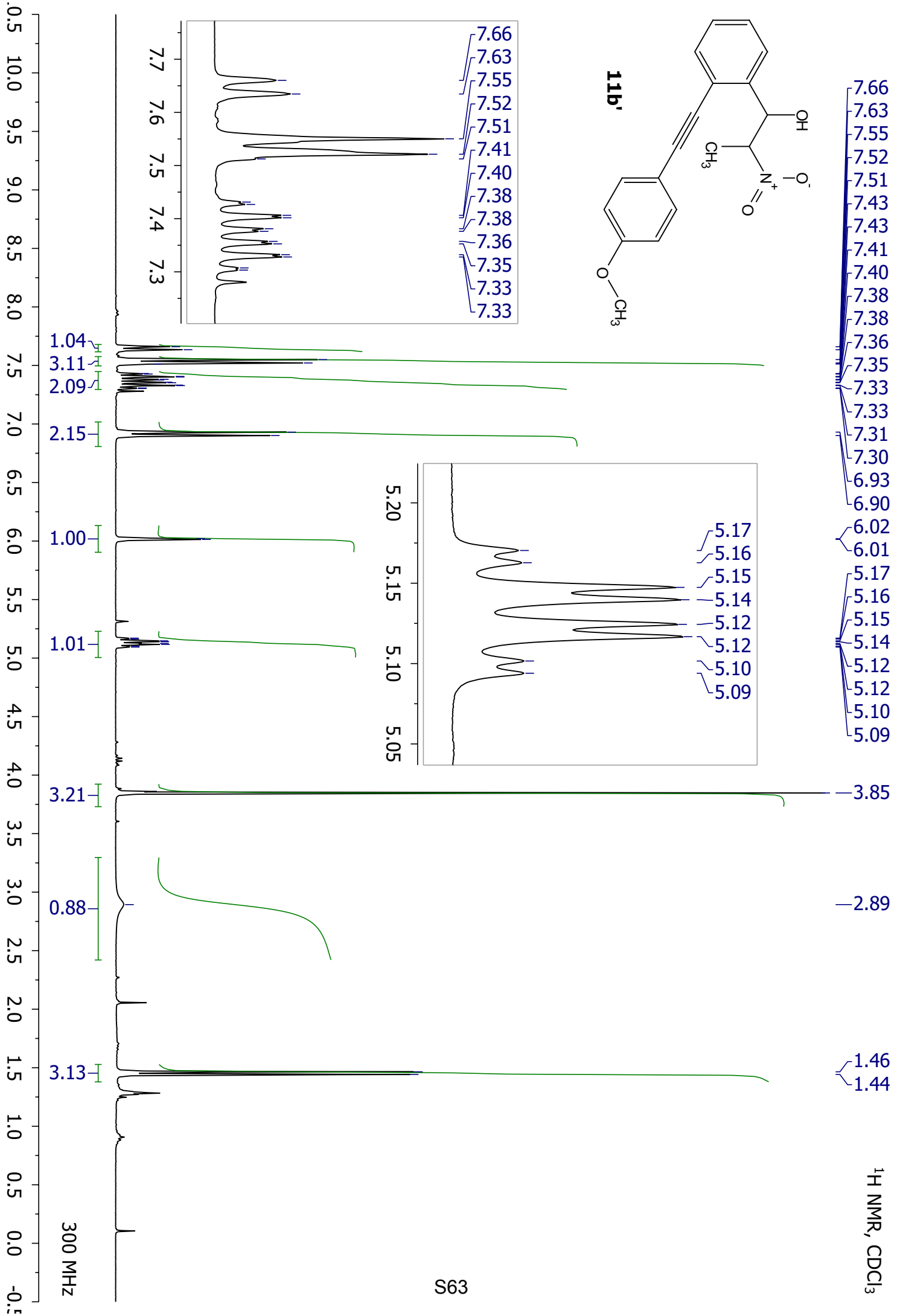
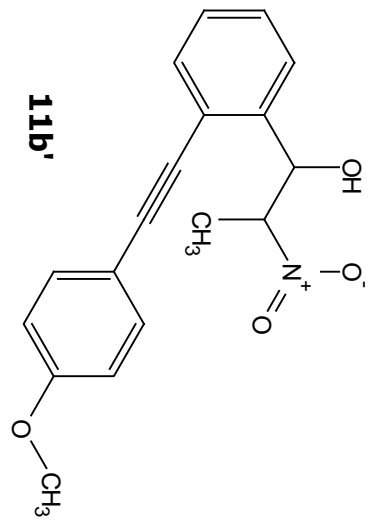


¹³C NMR, CDCl₃

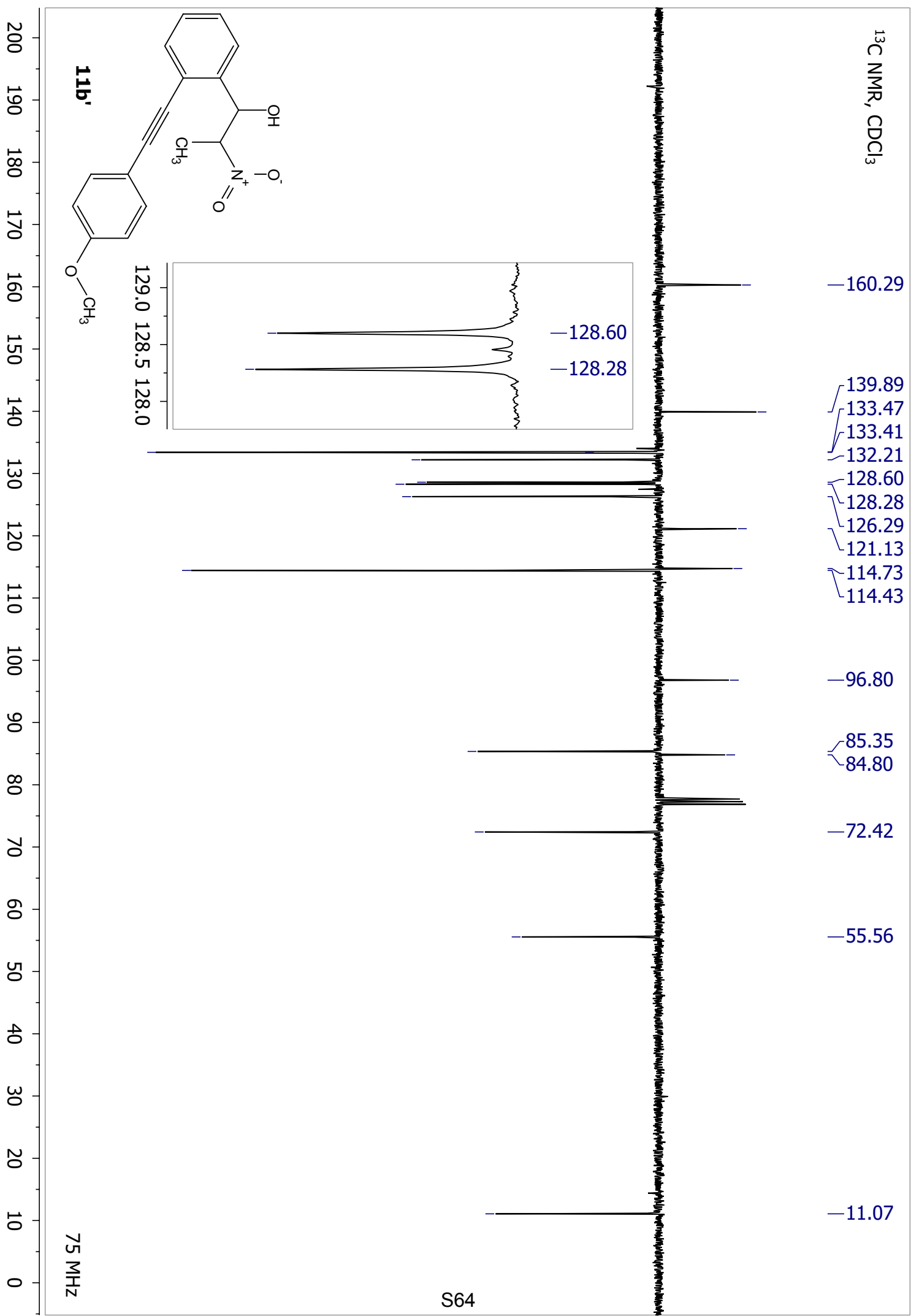


75 MHz

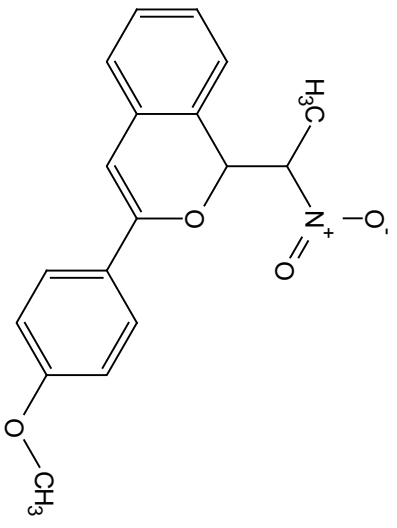
¹H NMR, CDCl₃



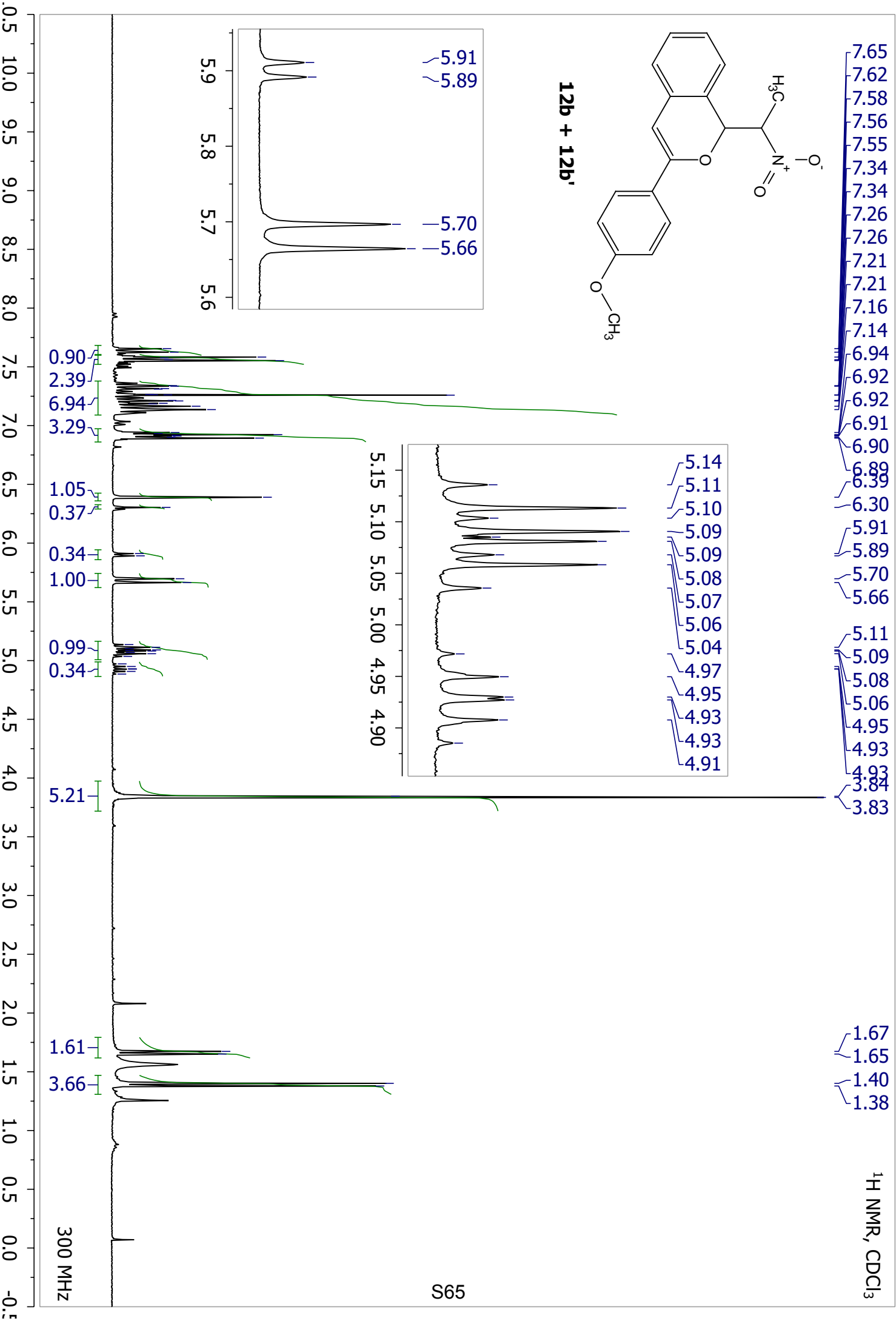
¹³C NMR, CDCl₃

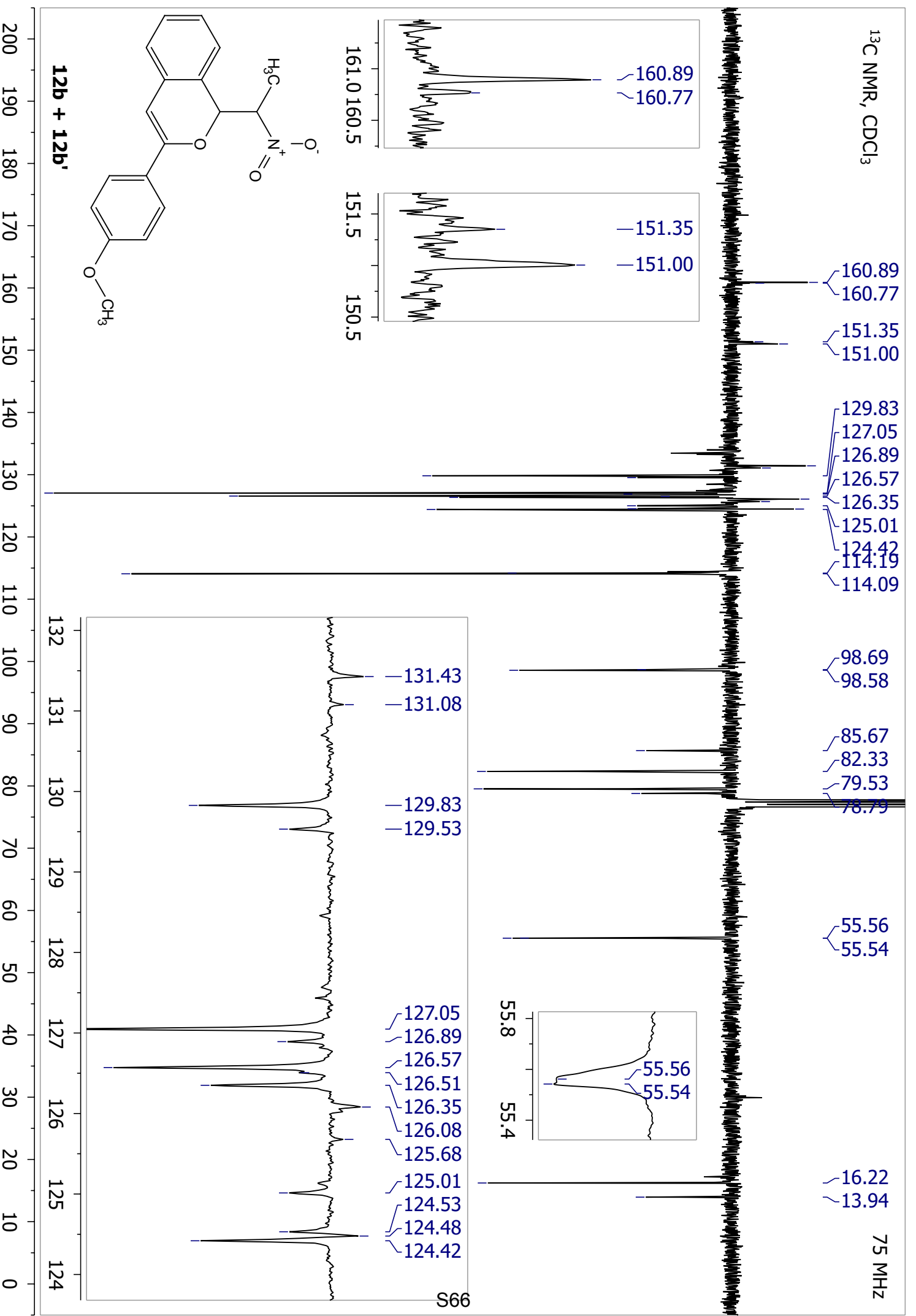


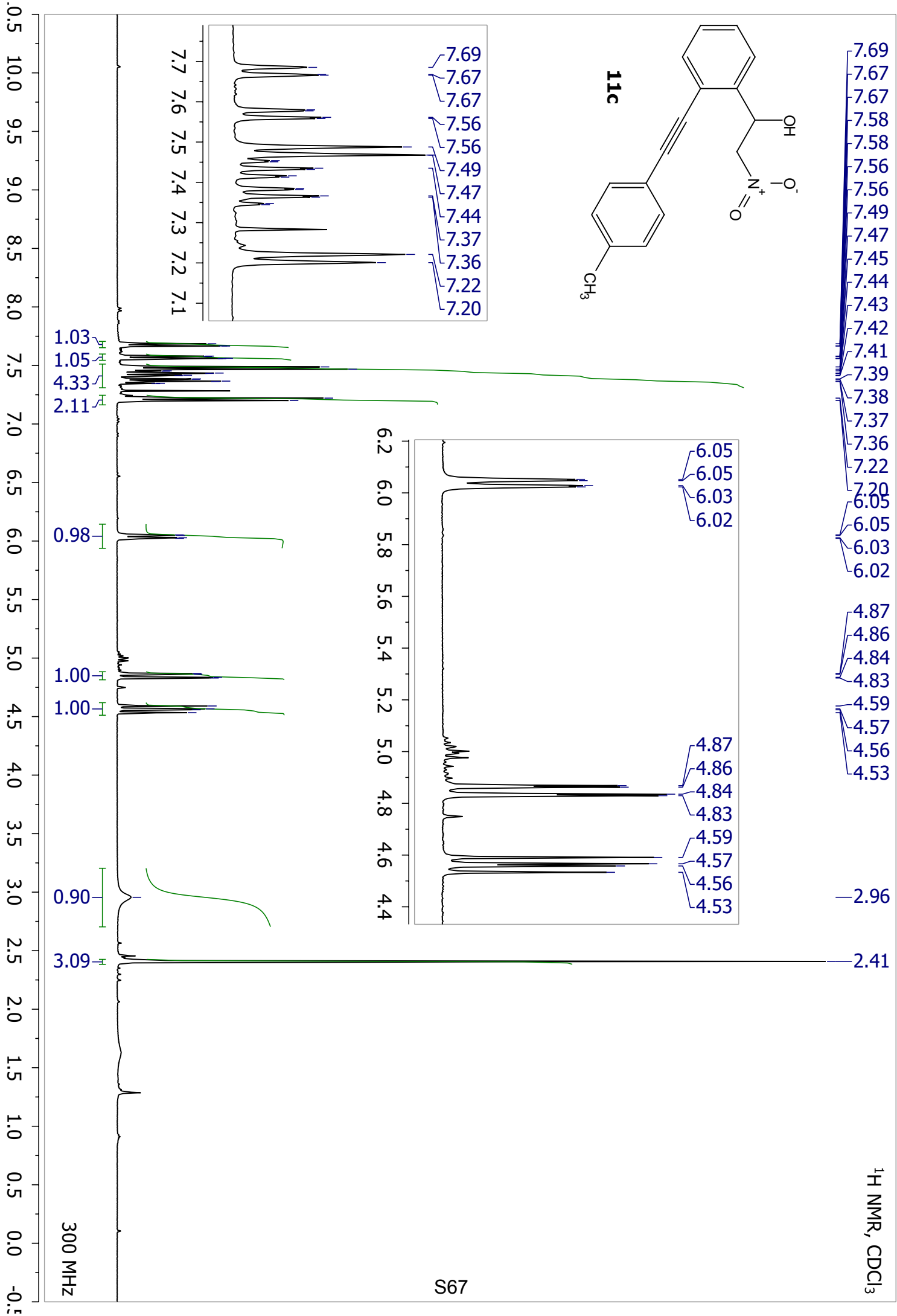
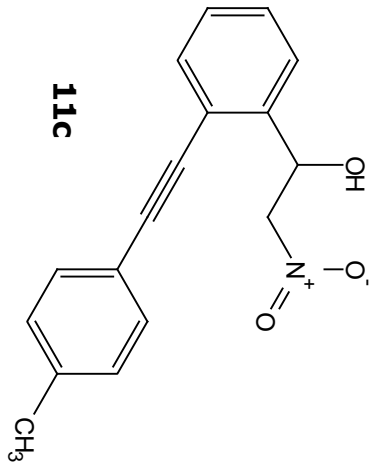
¹H NMR, CDCl₃

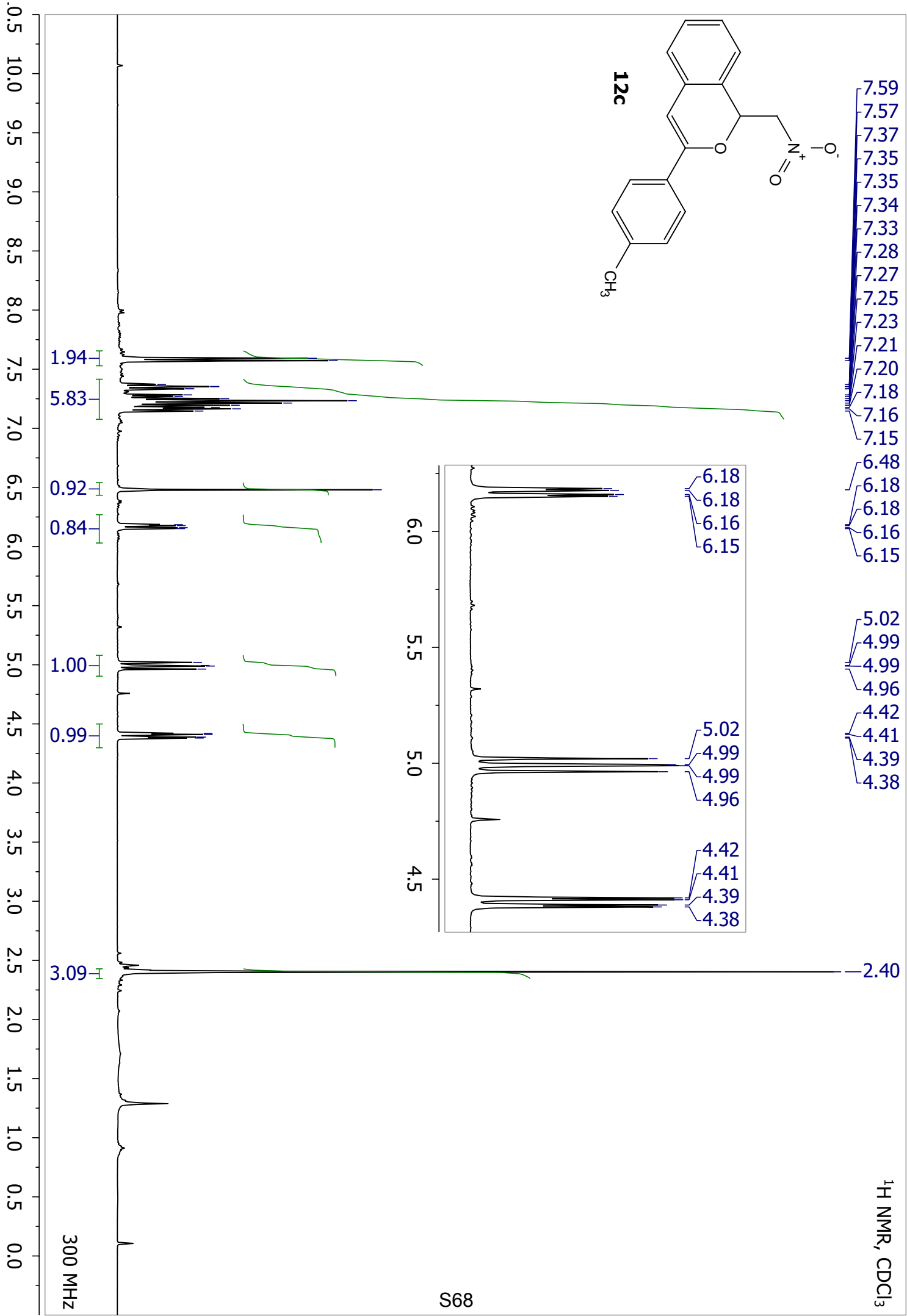
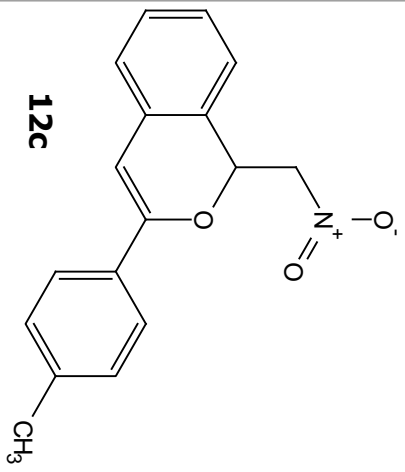


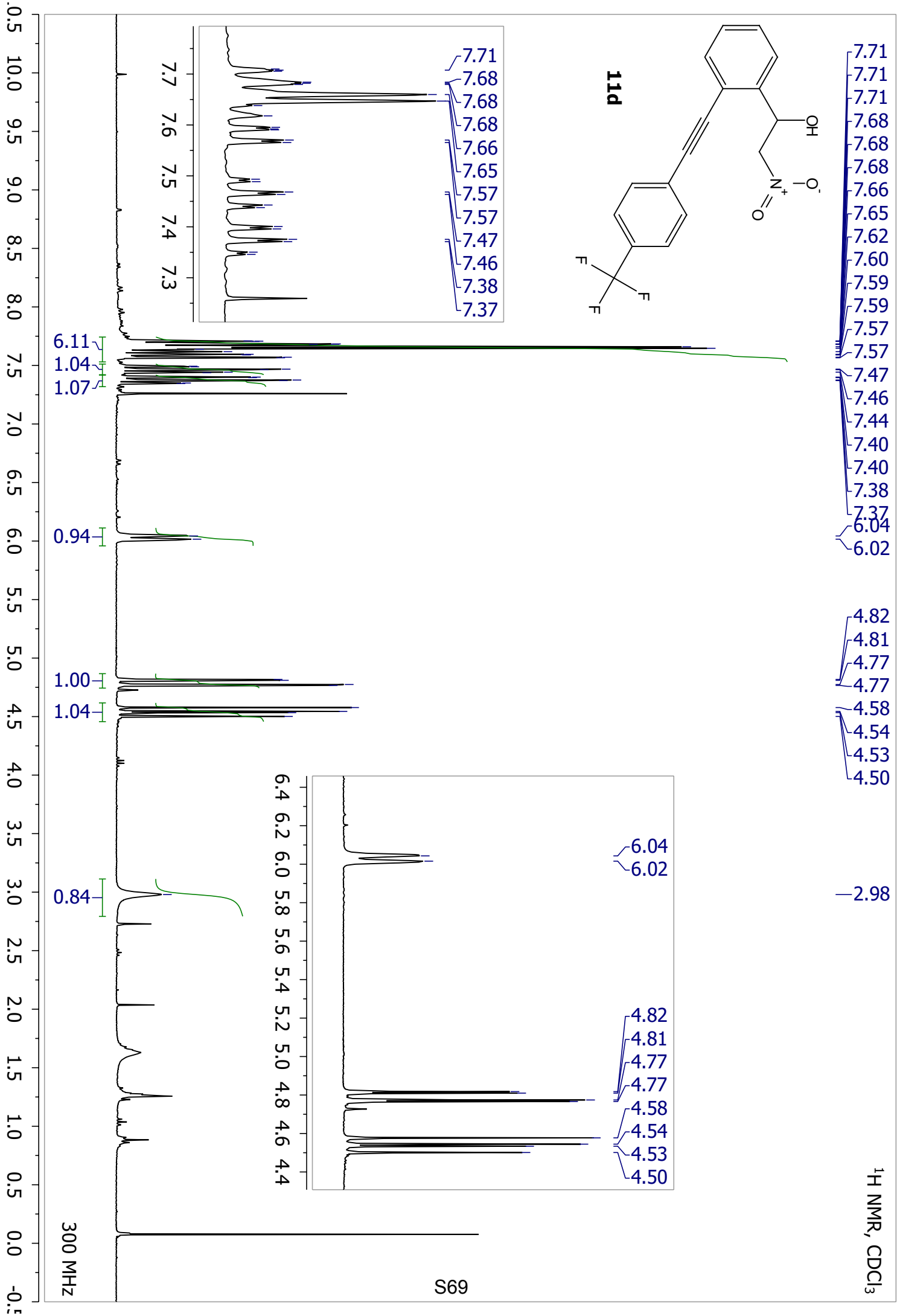
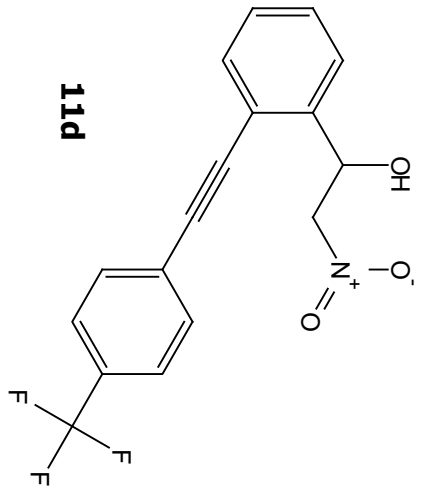
12b + 12b'

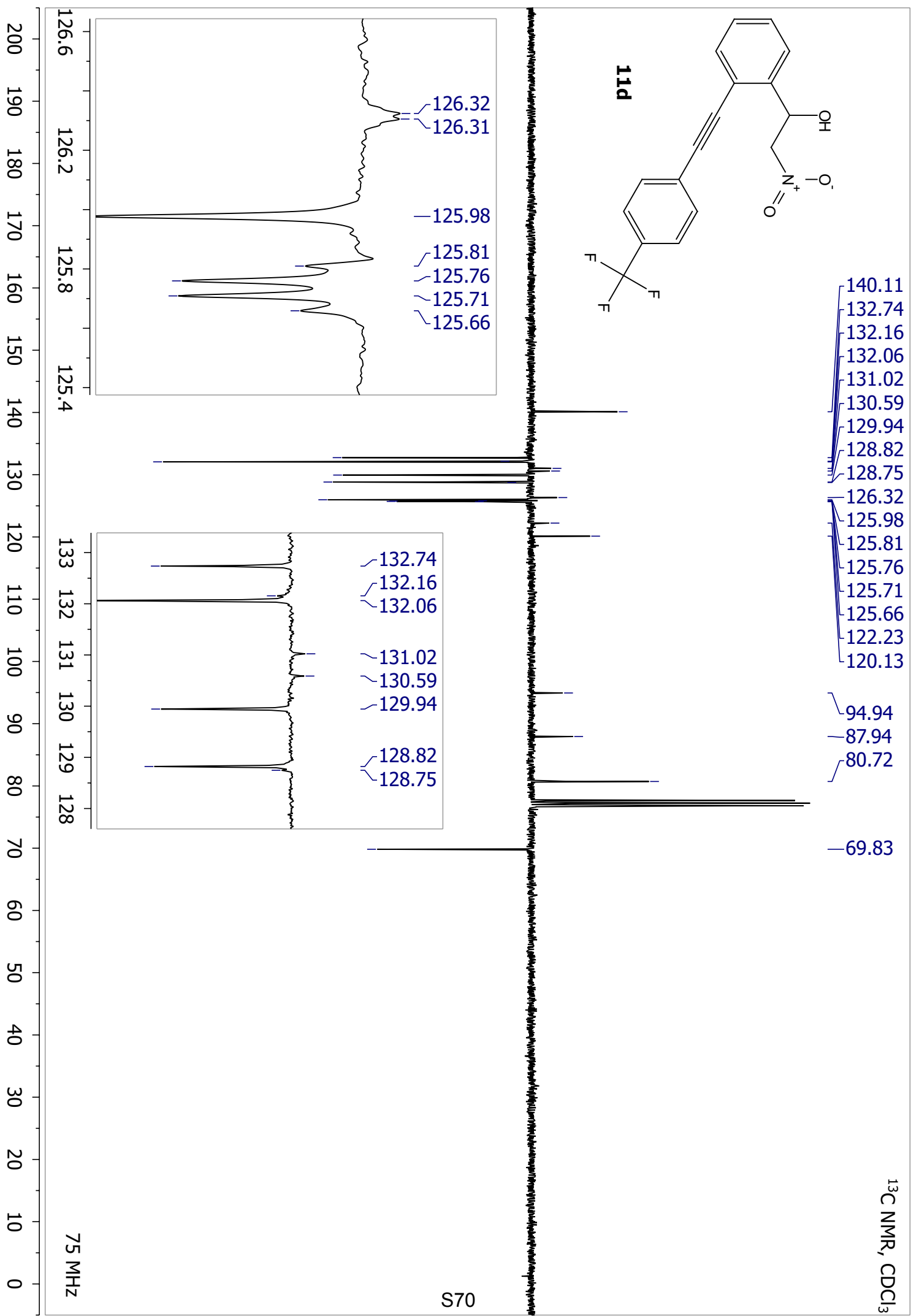
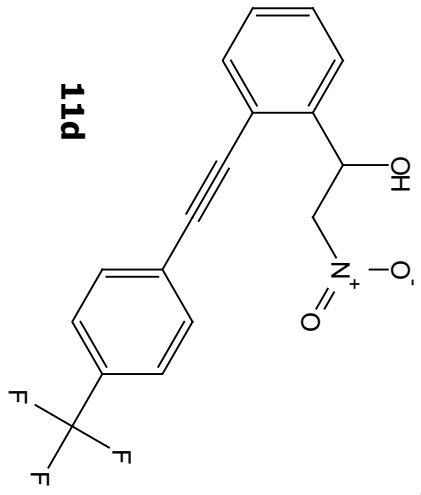


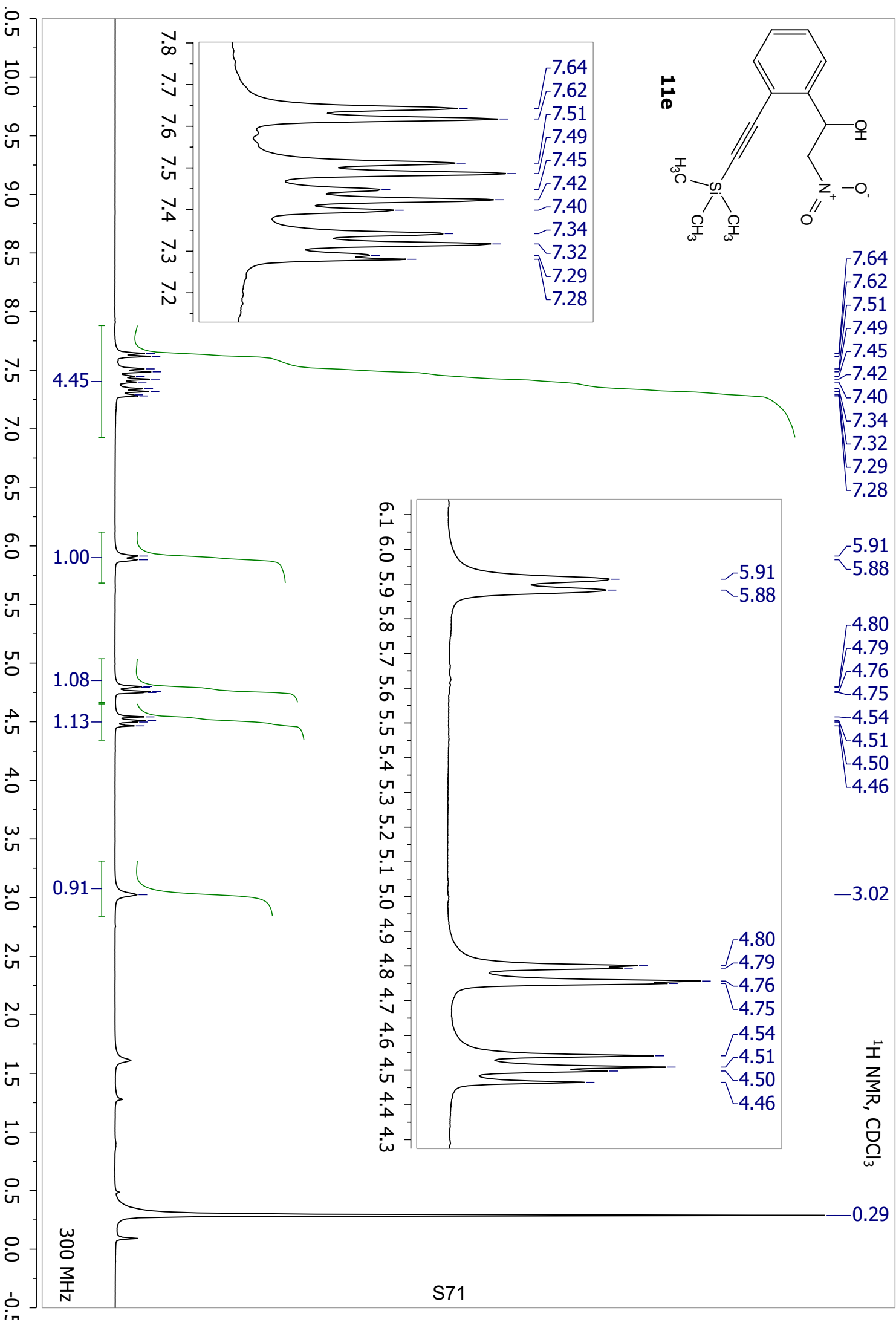
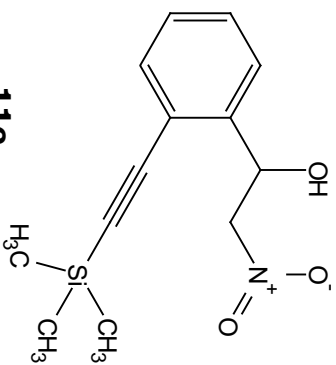




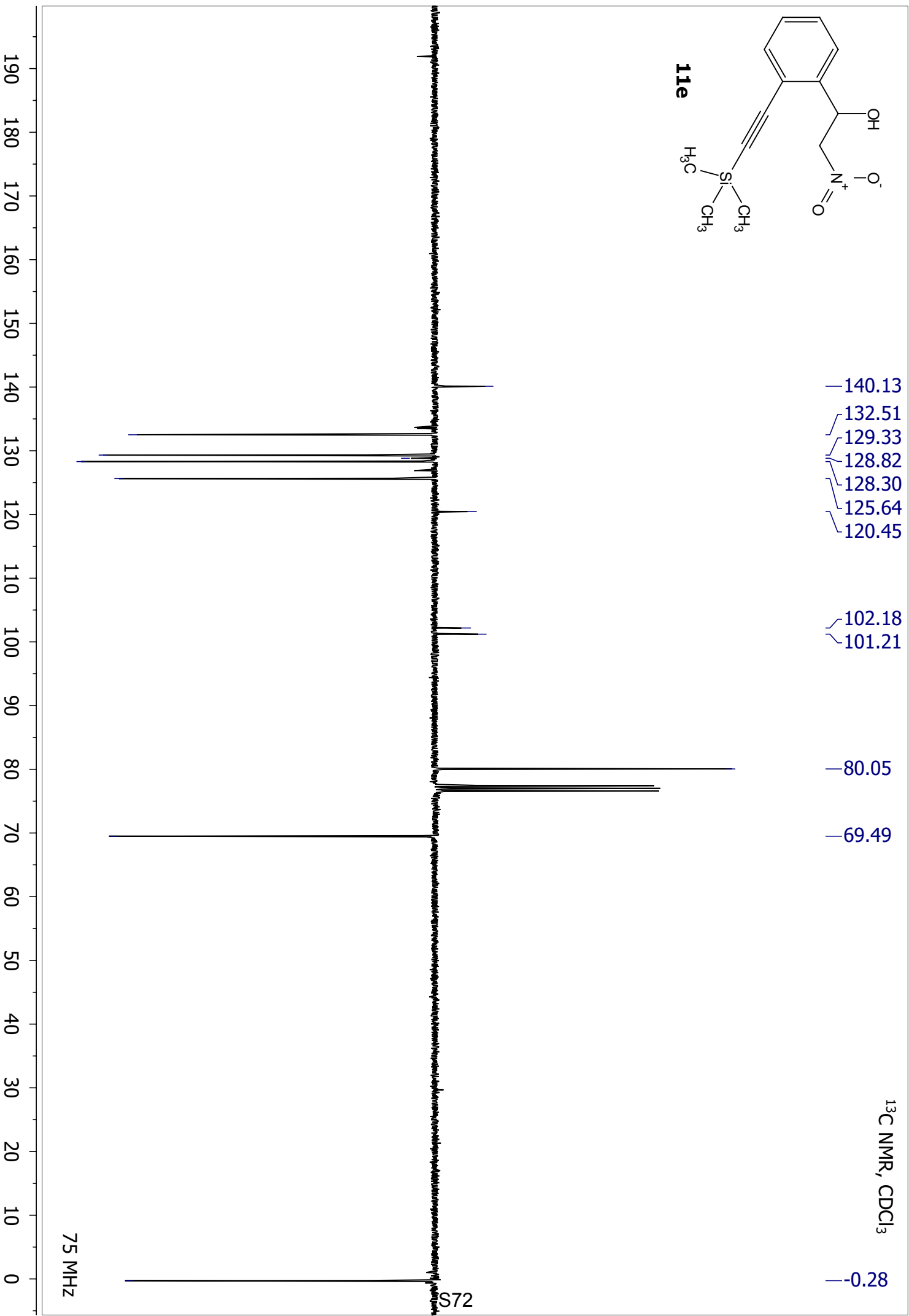
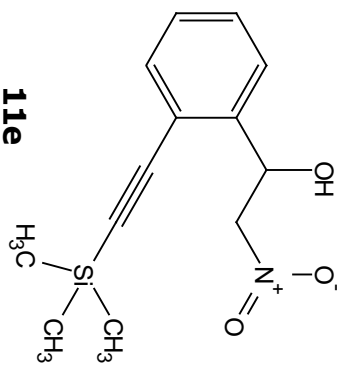


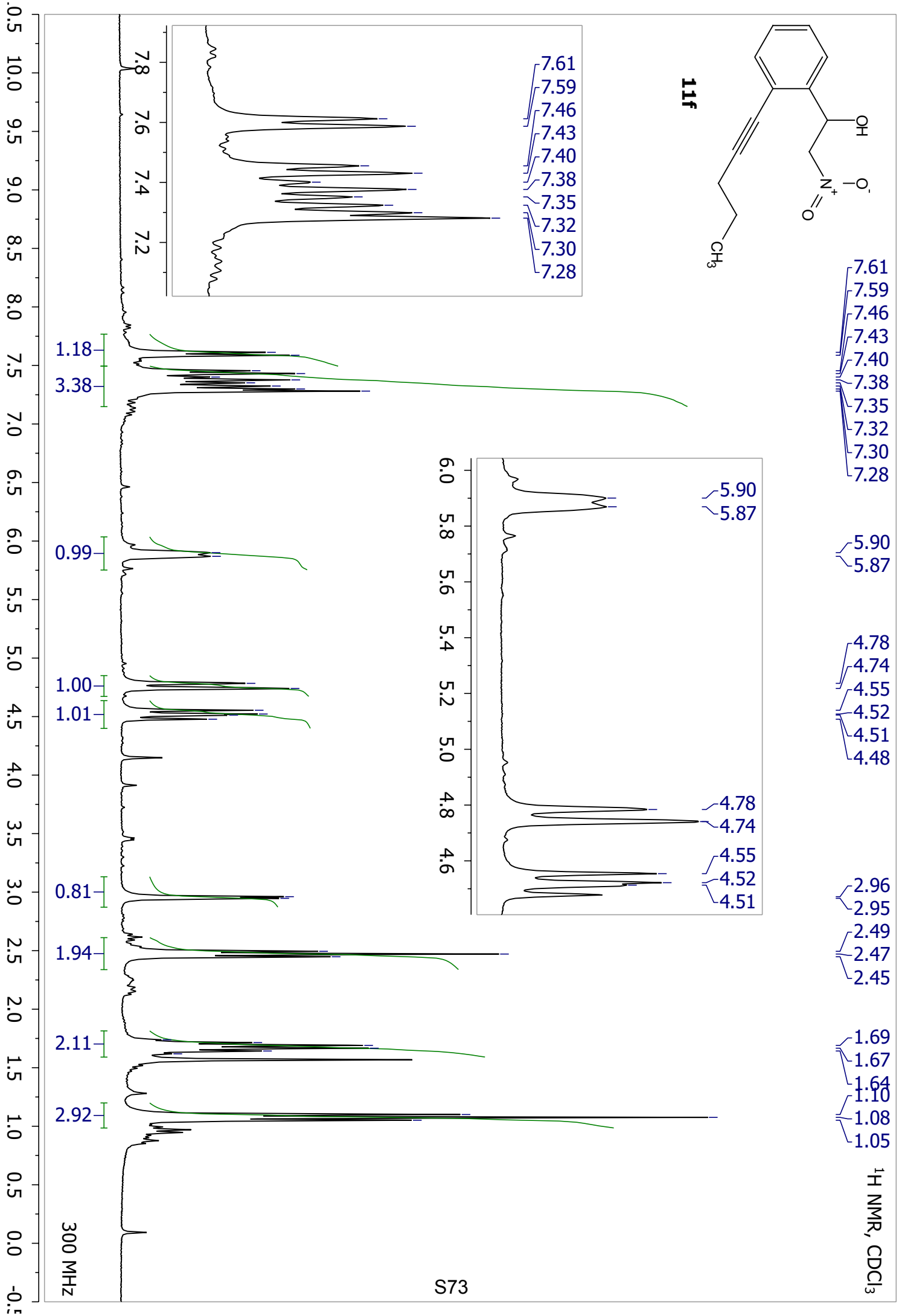
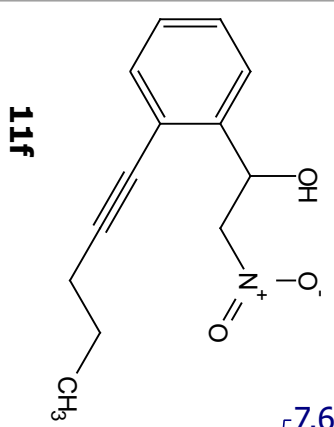


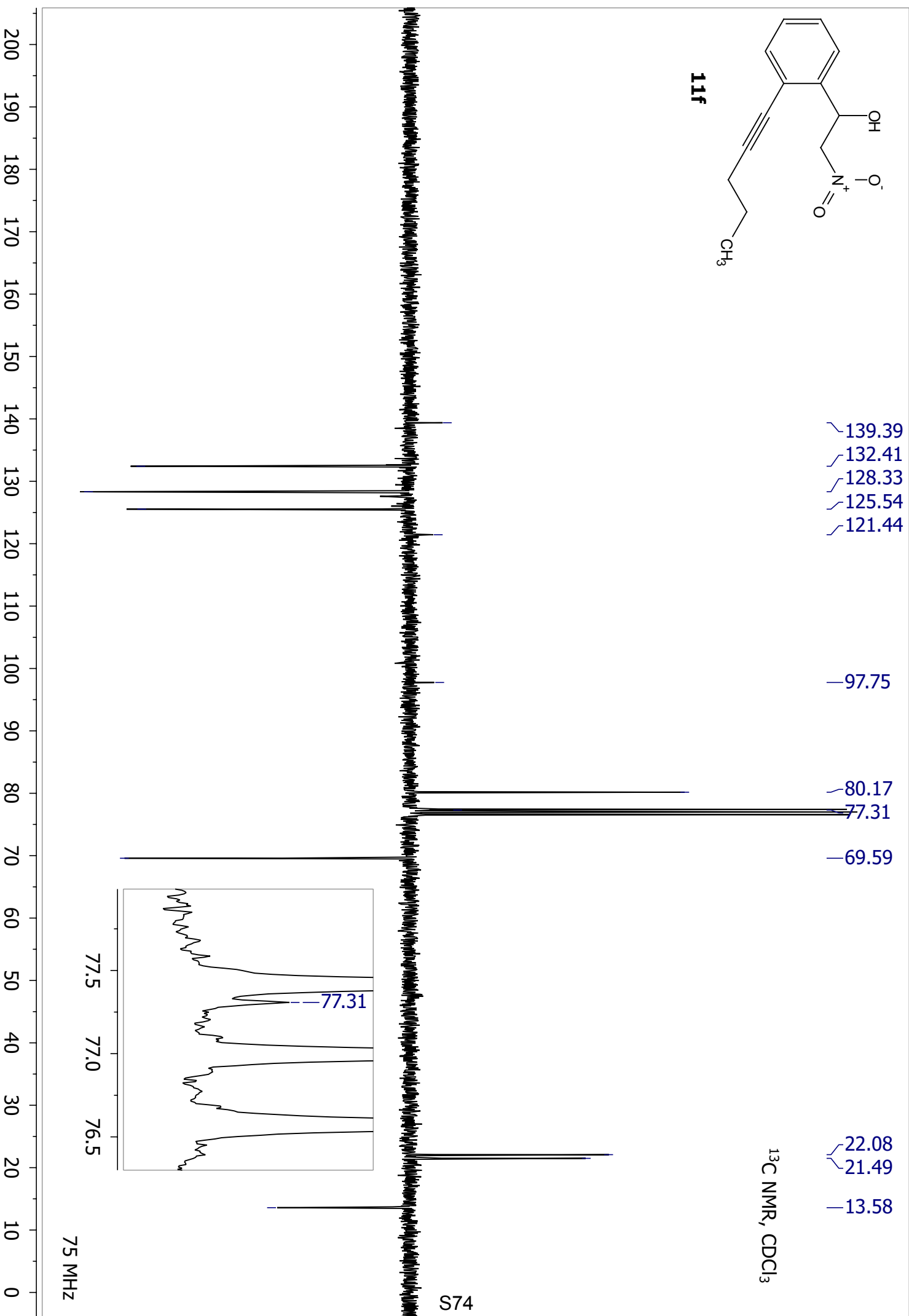
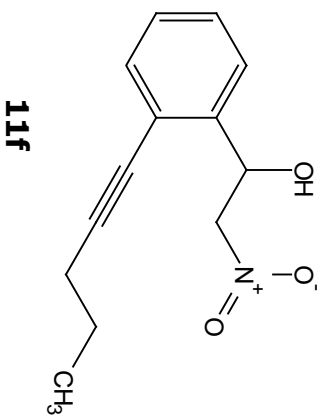


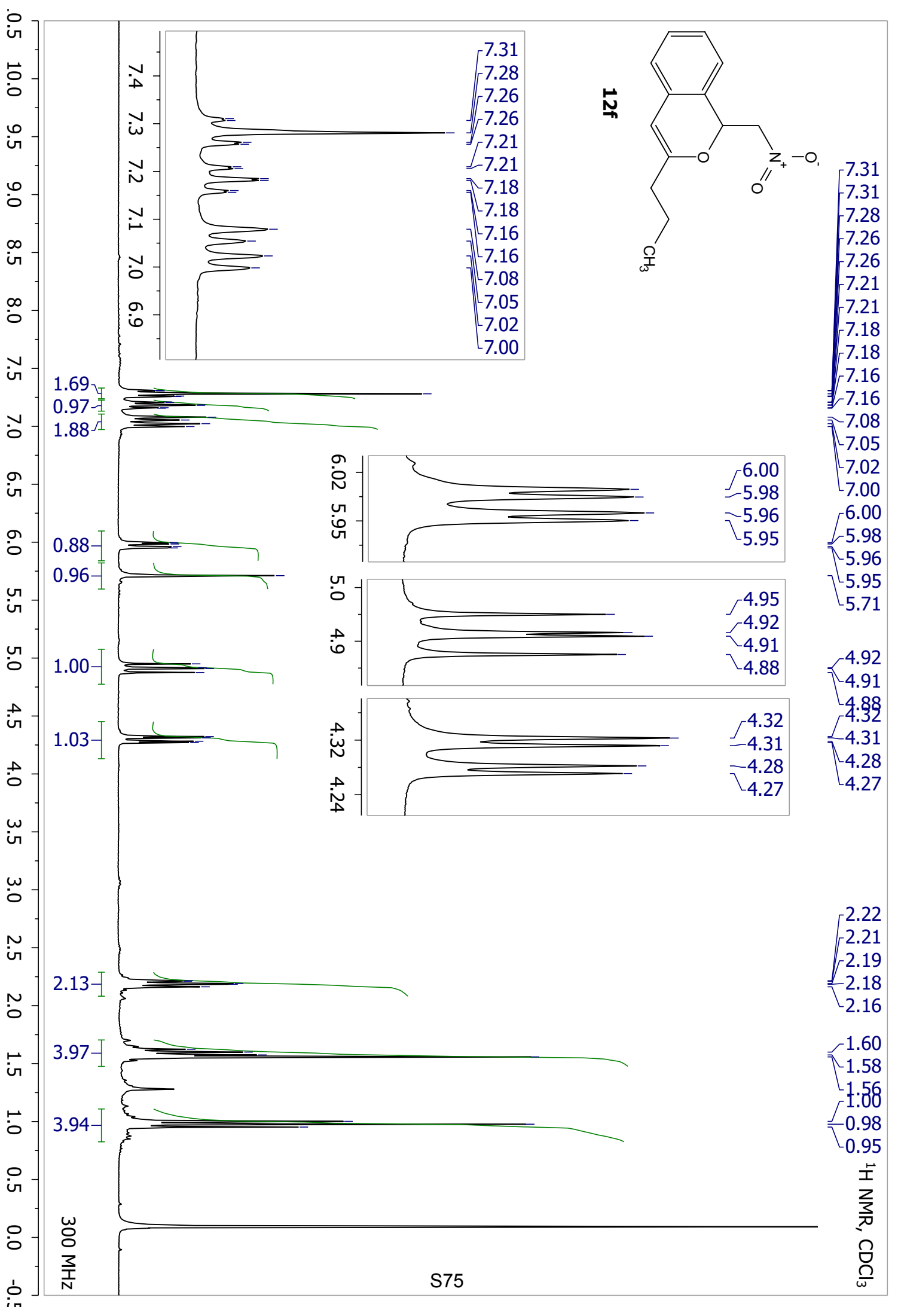


¹³C NMR, CDCl₃

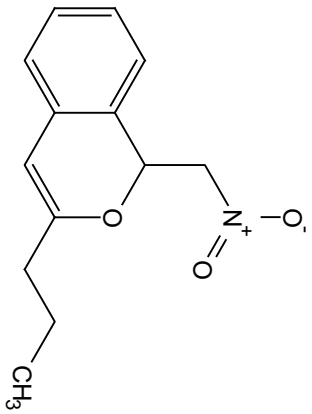




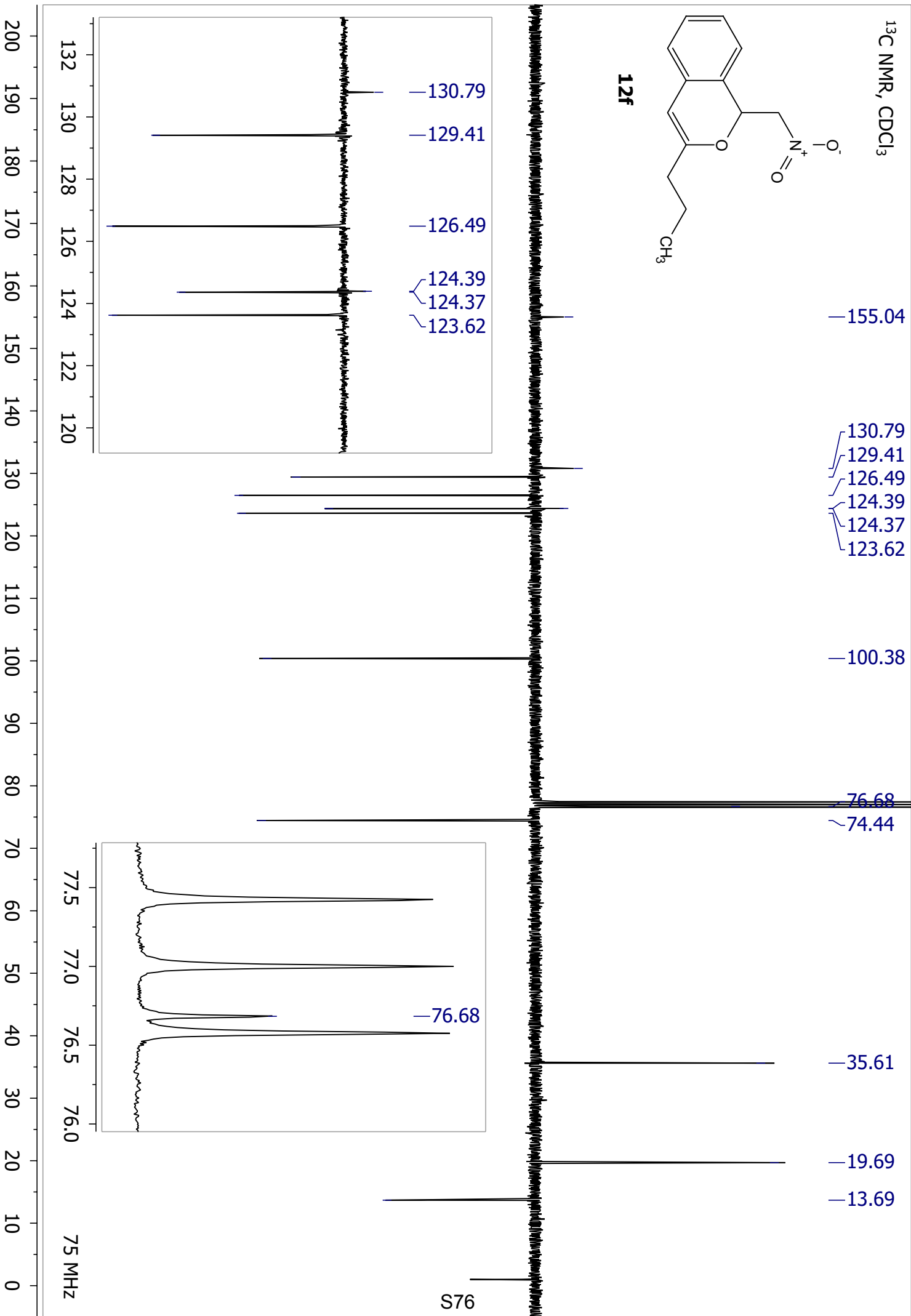


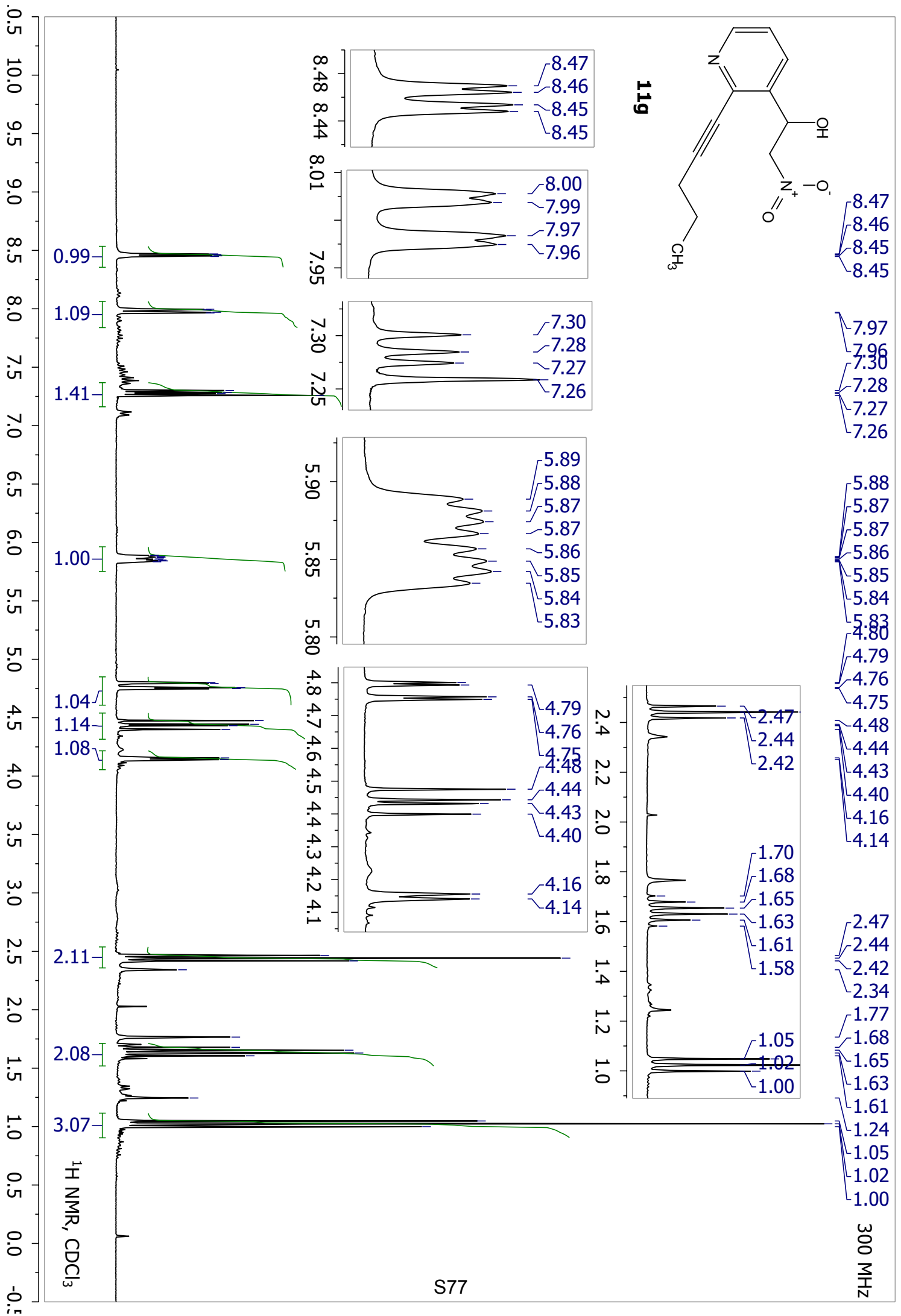


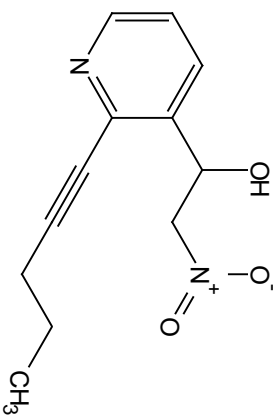
¹³C NMR, CDCl₃



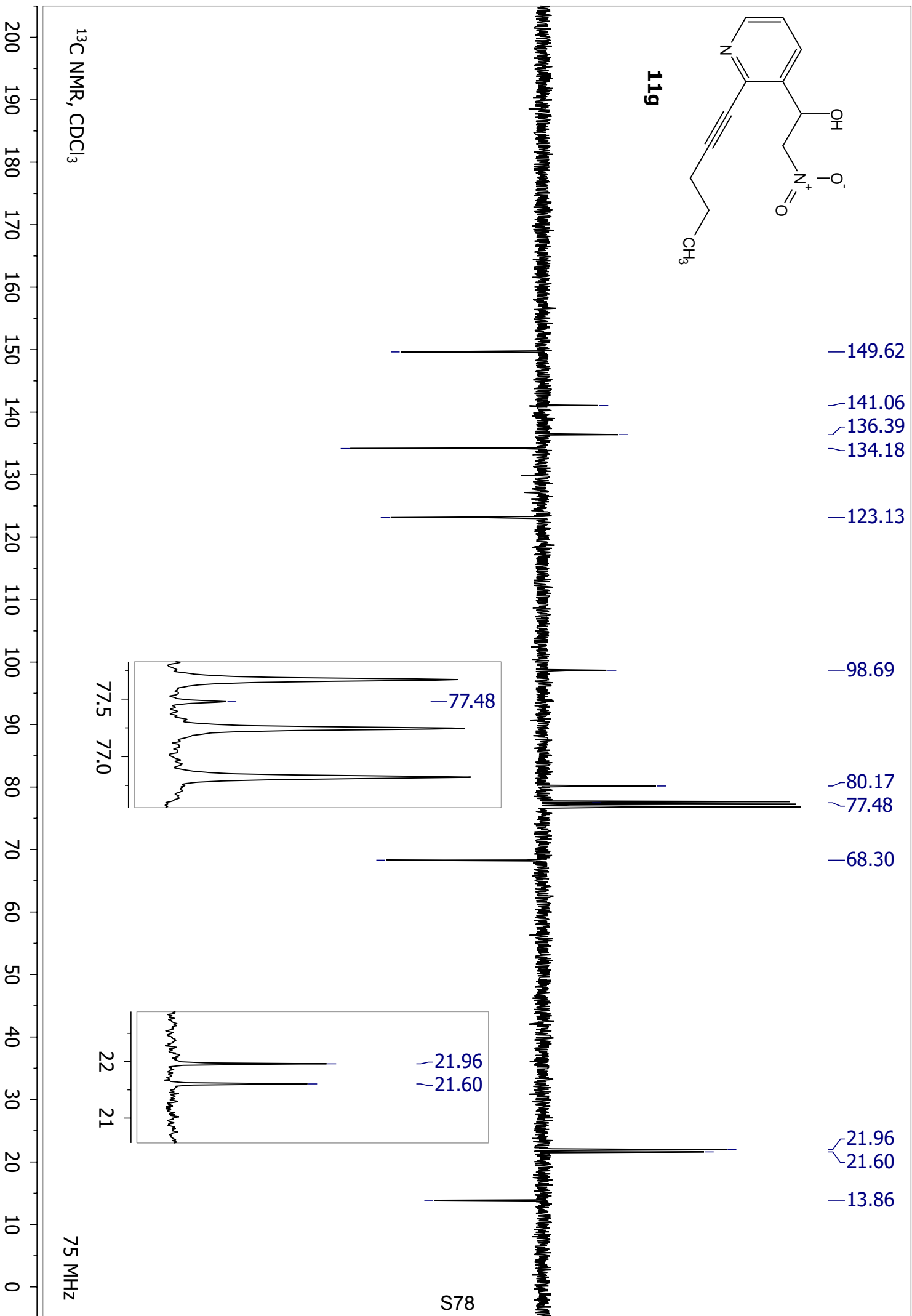
12f

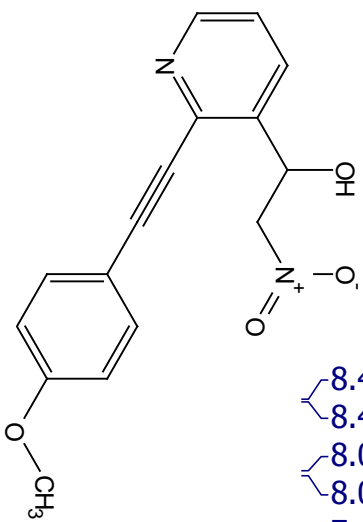




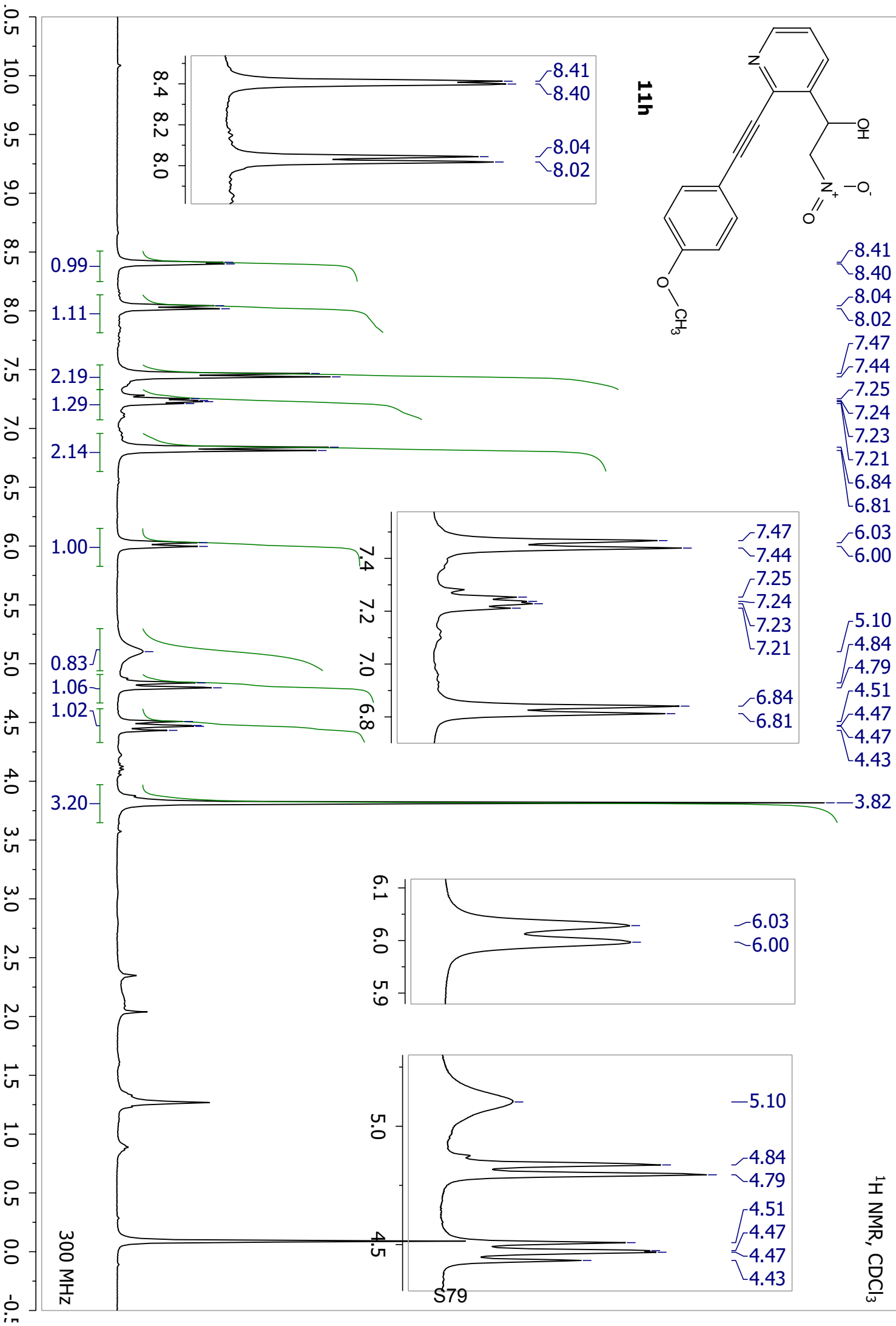


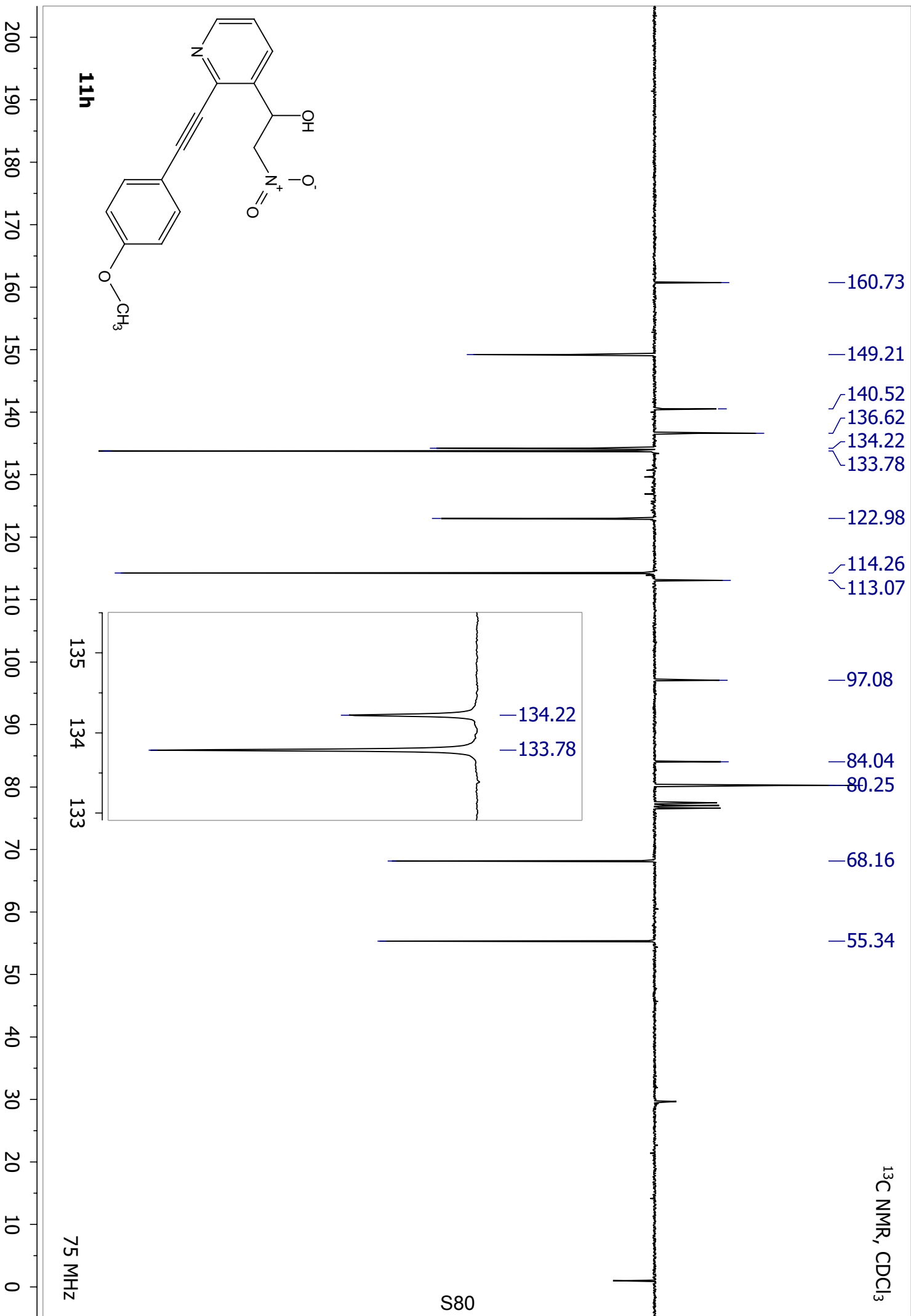
11g

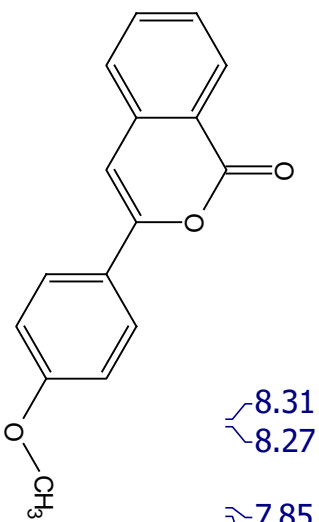




11h







13

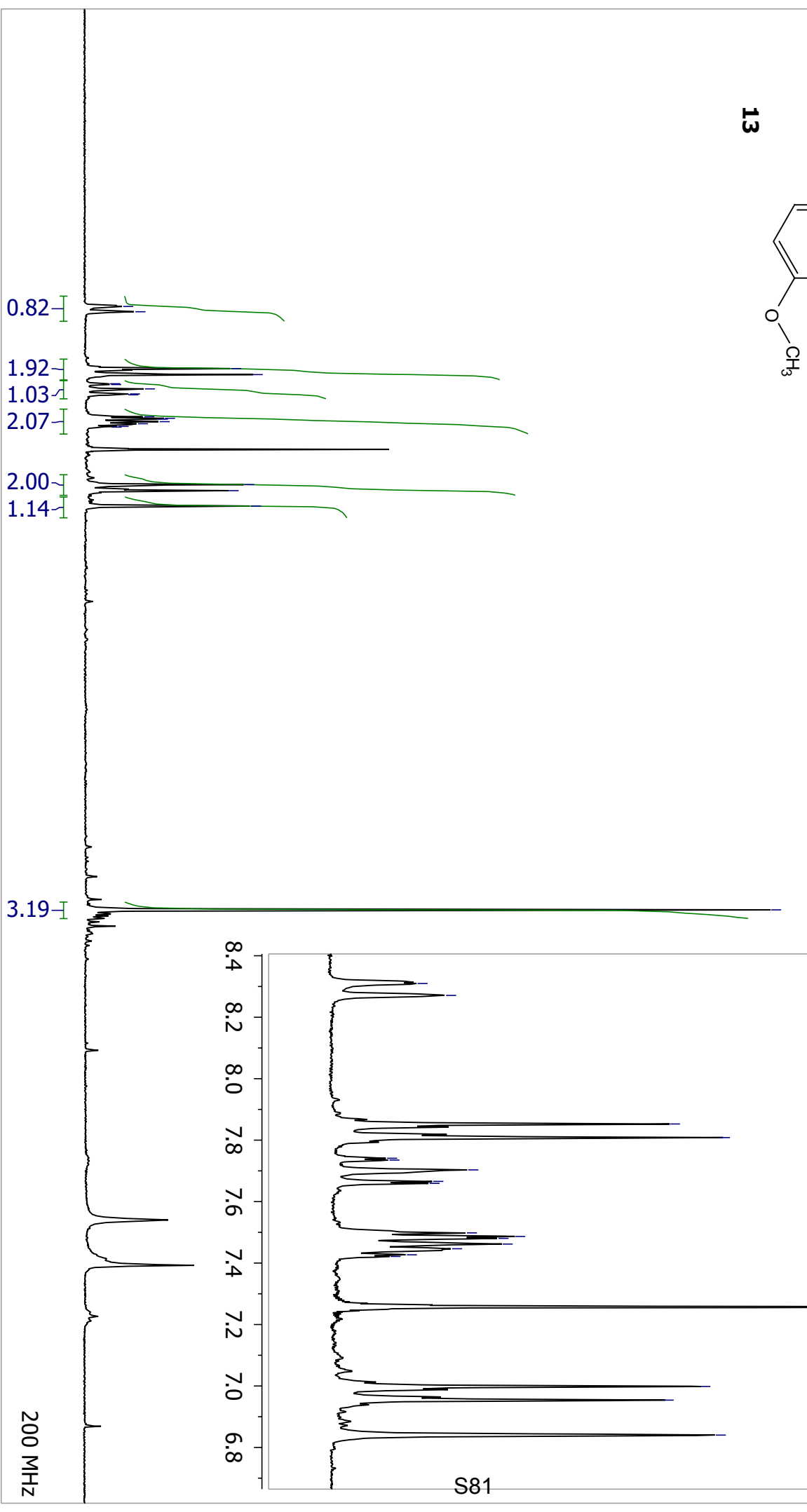
¹H NMR, CDCl₃

8.31
8.27
7.85
7.81
7.49
7.46
7.00
6.95
6.84

3.87

8.31
8.27
7.85
7.81
7.74
7.70
7.67
7.66
7.50
7.49
7.48
7.46
7.45
7.43
7.00
6.95
6.84

0.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5



200 MHz

S81