

Supporting Information

Constructing Densely Functionalized Hajos–Parrish–type Ketones with Six Contiguous Stereogenic Centers and Two Quaternary Carbons in a formal [2 + 2 + 2] Cycloaddition Cascade.

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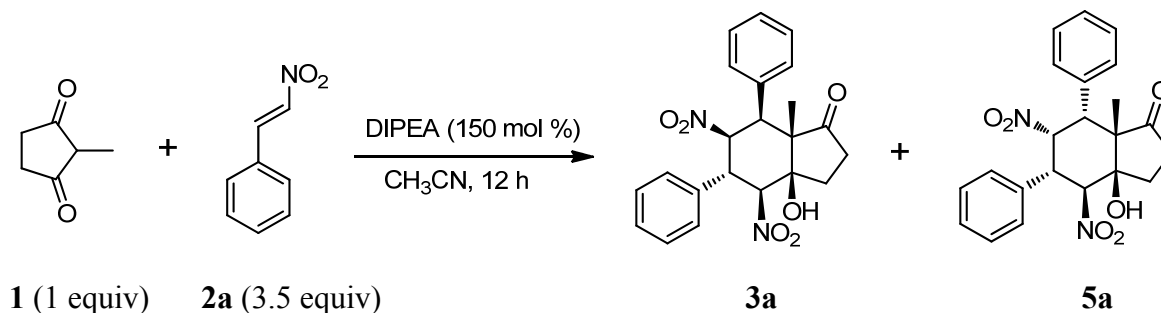
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General Procedure. All solvents were reagent grade. Reactions were normally carried out under nitrogen atmosphere in glassware. Merck silica gel 60 (particle size 0.04-0.063 mm) was employed for flash chromatography. Melting points are uncorrected. ¹H NMR spectra were obtained in CDCl₃ unless otherwise noted at 400 MHz (Bruker DPX-400) or 500 MHz (Varian-Unity INOVA-500). ¹³C

NMR spectra were obtained at 100 MHz or 125 MHz. *E.e.* values were measured by HPLC on a chiral column (chiralpak IC, 0.46 cm ID x 25 cm, particle size 5 μ) by elution with THF-hexane. The flow rate of the indicated elution solvent is maintained at 1 mL/min, and the retention time of a compound is recorded accordingly. HPLC was equipped with the ultraviolet and refractive index detectors. The melting point was recorded on a melting point apparatus (MPA100 – Automated melting point system, Stanford Research Systems, Inc.) and is uncorrected.

Preparation of adduct **3a** and **5a**.



To a solution of DIPEA (44 mg, 0.34 mmol, 1.5 equiv), 2-methyl-1,3-cyclopentanone (**1**, 25 mg, 0.22 mmol) in CH_3CN (1.1 mL) was added nitrostyrene (**2a**, 116.3 mg, 0.78 mmol, 3.5 equiv) and the resulting solution was stirred at room temperature for 12 h until the completion of the reaction, as monitored by TLC. Then the reaction mixture was directly filtered through a Celite bed (to remove insoluble polymer junk) and washed with EtOAc (15 mL). The filtrate was concentrated *in vacuo* and the crude product was purified by flash column chromatography with 20 % EtOAc–hexane to give **3a** (58.5 mg, 64% yield) as a pale yellow solid ($R_f = 0.23$ in 20% EtOAc–hexane developed two times on TLC) and **5a** (13.7 mg, 15%) as white solid ($R_f = 0.19$ in 20% EtOAc–hexane developed two times on TLC).

Select data for **3a**: m.p. 240–241 $^\circ\text{C}$; IR (neat): 3441, 2919, 1740, 1556, 1454, 1364, 1073, 756, 701, 463 cm^{-1} ; ^1H NMR (500 MHz, acetone- d_6): δ 7.50 – 7.42 (m, 2 H), 7.40 – 7.32 (m, 4 H), 7.28 – 7.15 (m, 4 H), 5.93 (s, 1 H), 5.21 (d, $J = 12.5$ Hz, 1 H), 5.14 (dd, $J = 12.5, 6.5$ Hz, 1 H), 4.50 (t, $J = 12.5$ Hz, 1 H), 4.15 (d, $J = 6.5$ Hz, 1 H), 2.91 – 2.80 (m, 2 H), 2.76 – 2.68 (m, 1 H), 2.34 – 2.25 (m, 1 H), 0.96 (s, 3 H); ^{13}C NMR (125 MHz, acetone- d_6): δ 214.5 (C), 137.1 (C), 135.6 (C), 129.3 (6 CH), 128.94 (2 CH), 128.90 (2 CH), 92.7 (CH), 89.3 (CH), 79.8 (C), 58.0 (C), 50.2 (CH), 41.5 (CH), 34.1 (CH_2), 28.9 (CH_2), 19.3 (CH_3); MS (m/z , relative intensity): 410 (M^+ , 18), 364 (15), 318 (52), 257 (81), 245 (45), 205 (21), 115 (51), 105 (35), 91 (100), 77 (34); exact mass calculated for $\text{C}_{22}\text{H}_{22}\text{N}_2\text{O}_6$ (M^+): 410.1478; found: 410.1477. Anal. Calcd for $\text{C}_{22}\text{H}_{22}\text{N}_2\text{O}_6$: C, 64.38; H, 5.40; N, 6.83; O, 23.39; found: C, 64.45; H, 5.36; N, 6.56; O, 23.26.

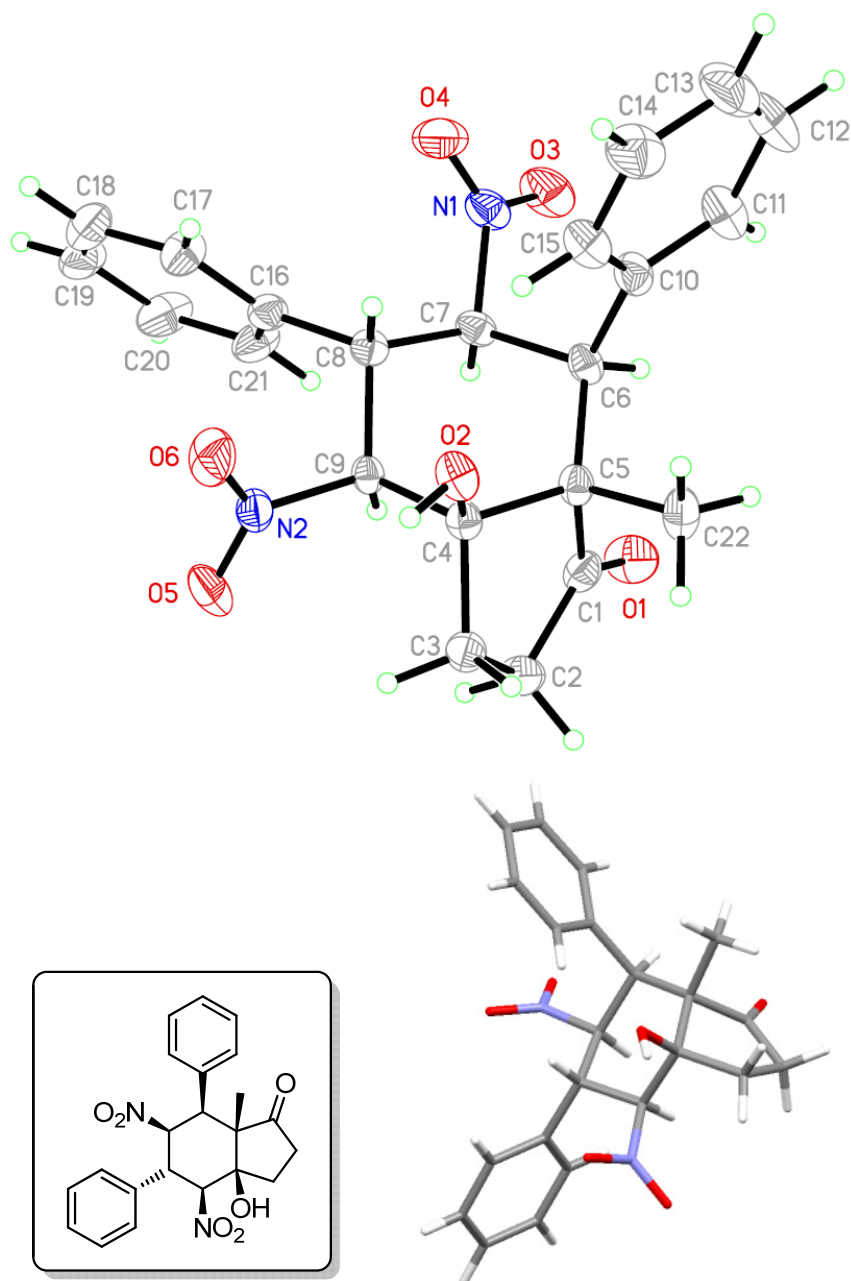


Figure S1. ORTEP and Stereo plots for X-ray crystal structures of (±)-**3a**.

CCDC 1474917 contains the supplementary crystallographic data for (±)-**3a**. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Table S1. Crystal data and structure refinement for (\pm)-**3a**.

Identification code	ic17001	
Empirical formula	C ₂₂ H ₂₂ N ₂ O ₆	
Formula weight	410.42	
Temperature	200(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P2(1)/c	
Unit cell dimensions	a = 11.4563(7) Å	a = 90°.
	b = 6.7710(4) Å	b = 93.8075(14)°.
	c = 25.4114(15) Å	g = 90°.
Volume	1966.8(2) Å ³	
Z	4	
Density (calculated)	1.386 Mg/m ³	
Absorption coefficient	0.102 mm ⁻¹	
F(000)	864	
Crystal size	0.40 x 0.27 x 0.05 mm ³	
Theta range for data collection	1.61 to 27.50°.	
Index ranges	-14 ≤ h ≤ 14, -8 ≤ k ≤ 8, -32 ≤ l ≤ 33	
Reflections collected	14689	
Independent reflections	4514 [R(int) = 0.0417]	
Completeness to theta = 27.50°	100.0 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.9949 and 0.9604	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	4514 / 0 / 273	
Goodness-of-fit on F ²	1.109	
Final R indices [I > 2σ(I)]	R1 = 0.0586, wR2 = 0.1365	
R indices (all data)	R1 = 0.0758, wR2 = 0.1460	
Largest diff. peak and hole	0.394 and -0.209 e.Å ⁻³	

Select data for **5a**: m.p. 184–185 °C; ^1H NMR (500 MHz, acetone- d_6): δ 7.56 – 7.54 (m, 2 H), 7.37 – 7.30 (m, 2 H), 7.29 – 7.22 (m, 6 H), 6.18 (d, J = 13.0 Hz, 1 H), 5.47 (s, 1 H), 5.18 (dd, J = 4.5 Hz, 4.0 Hz, 1 H), 4.58 (dd, J = 12.5 Hz, 4.5 Hz, 1 H), 3.74 (d, J = 4.0 Hz, 1 H), 3.08 – 3.01 (m, 1 H), 2.79 – 2.75 (m, 1 H), 2.65 (dd, J = 19.0 Hz, 9.0 Hz, 1 H), 2.31 – 2.24 (m, 1 H), 1.11 (s, 3 H); ^{13}C NMR (125 MHz, acetone- d_6): δ 211.8 (C), 136.8 (C), 136.7 (C), 132.2 (2 CH), 129.9 (2 CH), 129.2 (CH), 128.9 (2 CH), 128.7 (CH), 128.0 (2 CH), 93.5 (CH), 87.5 (CH), 80.2 (C), 55.8 (C), 50.3 (CH), 44.1 (CH), 35.7 (CH $_2$), 28.8 (CH $_2$), 21.6 (CH $_3$).

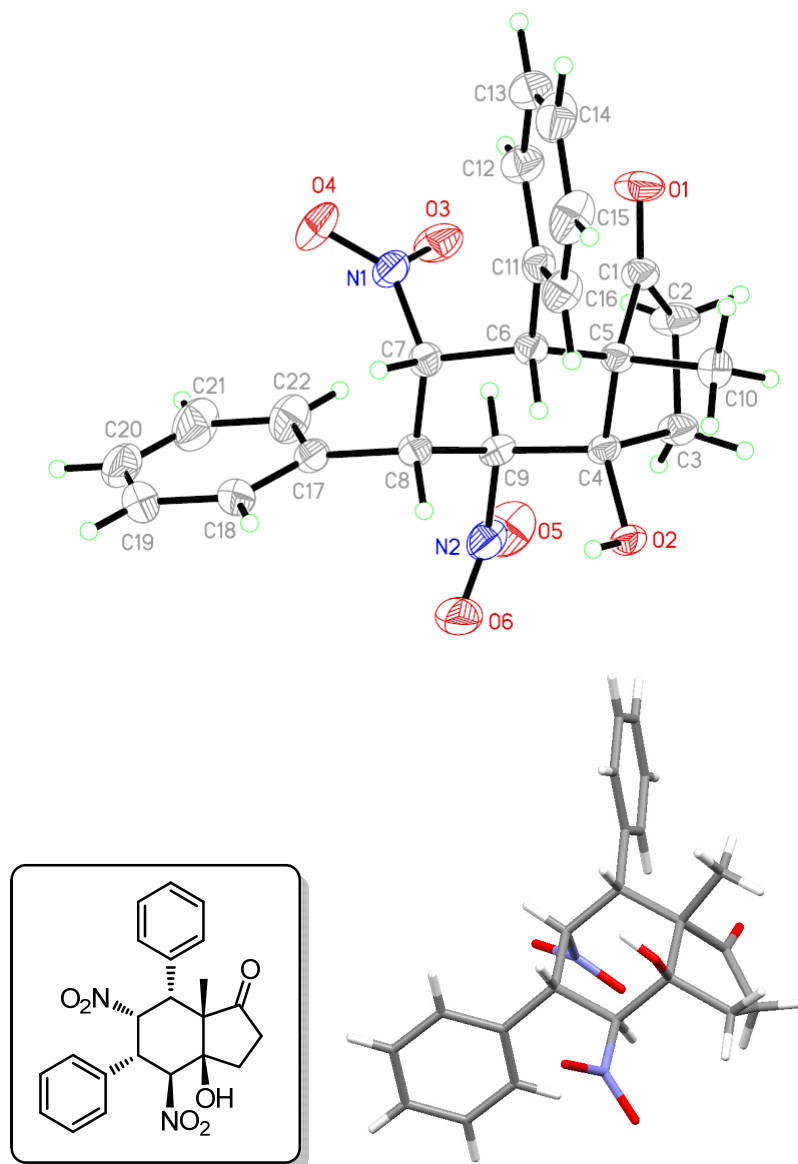
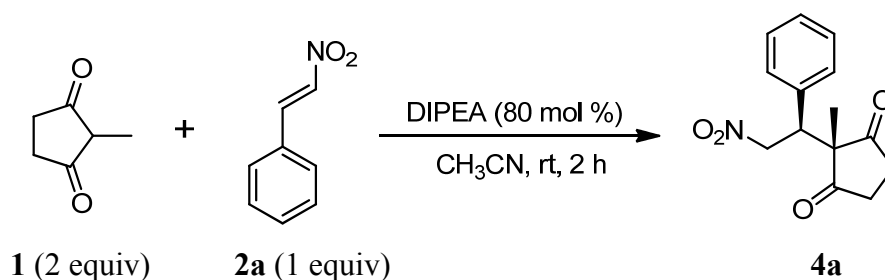


Figure S2. ORTEP and Stereo plots for X-ray crystal structures of (±)-**5a**.

CCDC 1495243 contains the supplementary crystallographic data for (±)-**5a**. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

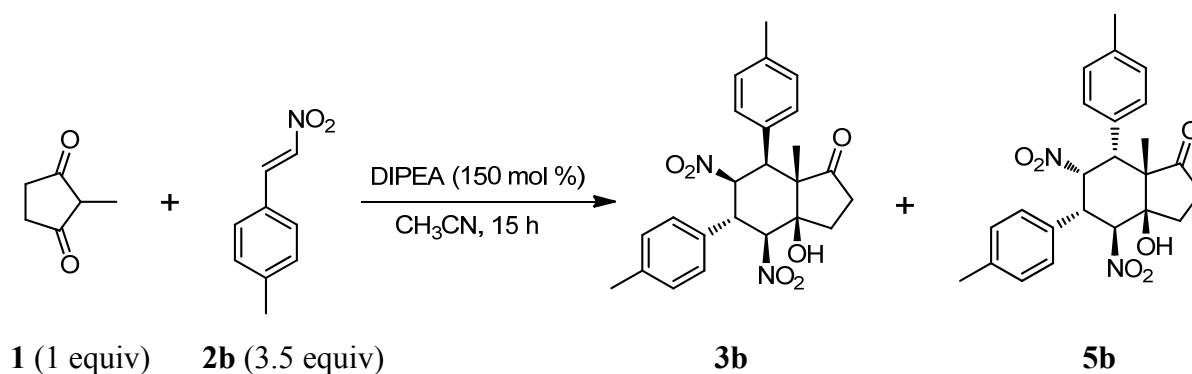
Table S2. Crystal data and structure refinement for (±)-**5a**.

Identification code	ic18058-1	
Empirical formula	C ₂₂ H ₂₂ N ₂ O ₆	
Formula weight	410.41	
Temperature	200(2) K	
Wavelength	1.54178 Å	
Crystal system	Orthorhombic	
Space group	P2 ₁ 2 ₁ 2 ₁	
Unit cell dimensions	a = 6.60530(10) Å	a = 90°.
	b = 16.3208(3) Å	b = 90°.
	c = 18.4301(3) Å	g = 90°.
Volume	1986.83(6) Å ³	
Z	4	
Density (calculated)	1.372 Mg/m ³	
Absorption coefficient	0.838 mm ⁻¹	
F(000)	864	
Crystal size	0.506 x 0.150 x 0.065 mm ³	
Theta range for data collection	3.617 to 69.976°.	
Index ranges	-7 ≤ h ≤ 8, -16 ≤ k ≤ 19, -22 ≤ l ≤ 22	
Reflections collected	9468	
Independent reflections	3755 [R(int) = 0.0198]	
Completeness to theta = 67.679°	99.9 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7533 and 0.5874	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	3755 / 0 / 273	
Goodness-of-fit on F ²	1.065	
Final R indices [I > 2σ(I)]	R1 = 0.0318, wR2 = 0.0793	
R indices (all data)	R1 = 0.0326, wR2 = 0.0800	
Absolute structure parameter	0.52(5)	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.151 and -0.225 e.Å ⁻³	

Isolation of the intermediate **4a**.

To a solution of DIPEA (62 mg, 0.48 mmol, 0.8 equiv), 2-methyl-1,3-cyclopentanone (**1**, 135 mg, 1.20 mmol, 2 equiv) in CH₃CN (6 mL) was added nitrostyrene (**2a**, 90 mg, 0.60 mmol) and the resulting solution was stirred at room temperature for 2 h. The reaction mixture was concentrated *in vacuo* to give the residue. The crude product was purified by flash column chromatography with 10 to 20 % EtOAc–hexane ($R_f = 0.29$ in 25 % EtOAc–hexane) to afford product **4a** (35 mg, 22% yield) as a yellow oil. Selected data for **4a**: IR (neat): 3035, 2973, 2924, 1722, 1555, 1454, 1416, 1379, 1344, 1076, 799, 762, 704 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.28–7.23 (m, 3 H), 7.16 – 7.13 (m, 2 H), 5.04 (dd, $J = 13.2, 9.6$ Hz, 1 H), 4.95 (d, $J = 13.2, 6.0$ Hz, 1 H), 3.92 (dd, $J = 9.7, 6.0$ Hz, 1 H), 2.72–2.51 (m, 2 H), 2.39–2.21 (m, 1 H), 2.10 – 1.90 (m, 1 H), 1.18 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃, ppm): δ 216.3 (C), 214.6 (C), 135.0 (C), 129.1 (2CH), 128.7 (2CH), 128.6 (CH), 74.8 (CH₂), 57.9 (C), 47.8 (CH), 35.5 (CH₂), 35.0 (CH₂), 19.4 (CH₃); MS (m/z , relative intensity): 261 (M^+ , 1), 215 (4), 186 (20), 171 (21), 149 (58), 131 (100), 104 (71), 103 (47), 102 (69), 91 (100), 77 (64); exact mass calculated for C₁₄H₁₅NO₄ (M^+): 261.1001; found: 261.1003.

Preparation of adduct **3b** and **5b**.



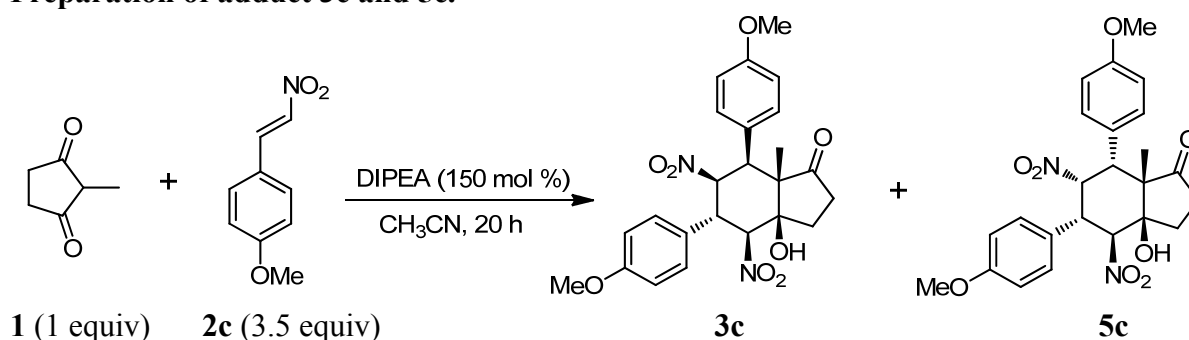
To a solution of DIPEA (44 mg, 0.34 mmol, 1.5 equiv), 2-methyl-1,3-cyclopentanone (**1**, 25 mg, 0.22 mmol) in CH₃CN (1.1 mL) was added nitrostyrene (**2b**, 127.3 mg, 0.78 mmol, 3.5 equiv) and the resulting solution was stirred at room temperature for 15 h until the completion of the reaction, as monitored by TLC. Then the reaction mixture was directly filtered through a Celite bed (to remove insoluble polymer junk) and washed with EtOAc (15 mL). The filtrate was concentrated *in vacuo* and the crude product was purified by flash column chromatography with 20 % EtOAc–hexane to afford **3b** (39.8 mg, 41% yield) as white solid ($R_f = 0.29$ in 20% EtOAc–hexane, developed two times on TLC) and **5b** (6.8 mg, 7%) as white solid ($R_f = 0.23$ in 20% EtOAc–hexane developed two times on TLC).

Select data for **3b**: m.p. 195–196 °C; IR (neat): 3550–3250, 2923, 2852, 1739, 1559, 1514, 1466, 1365, 1316, 1299, 1072, 809, 734 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ 7.23 (d, $J = 7.5$ Hz, 2 H), 7.23 – 7.10 (m, 4 H), 7.08 (d, $J = 7.5$ Hz, 2 H), 4.92 – 4.87 (m, 1 H), 4.79 (d, $J = 12.0$ Hz, 1 H), 4.55 – 4.53 (m, 1 H), 4.35 (dd, $J = 12.5, 12.5$ Hz, 1 H), 4.04 (d, $J = 6.5$ Hz, 1 H), 2.89 – 2.81 (m, 1 H), 2.80 – 2.70 (m, 1 H), 2.57 (dd, $J = 13.0$ Hz, 9.0 Hz, 1 H), 2.31 (s, 3 H), 2.20 (s, 3 H), 2.18 – 2.09 (m, 1 H), 0.89 (s, 3 H); ¹³C NMR (125 MHz, CD₃CN): δ 214.9 (C), 139.4 (C), 139.0 (C), 133.7 (C), 132.3 (C), 130.2 (4 CH), 130.1 (4 CH), 93.0 (CH), 89.5 (CH), 79.9 (C), 58.2 (C), 49.8 (CH), 41.1 (CH), 34.5 (CH₂), 28.9 (CH₂), 21.13 (CH₃), 21.09 (CH₃), 19.2 (CH₃); MS (m/z , relative intensity): 438 (M⁺, 2), 349 (9), 326 (5), 279 (9), 219 (73), 163 (32), 129 (33), 115 (100), 105 (83), 91 (79); exact mass calculated for C₂₄H₂₆N₂O₆ (M⁺): 438.1791; found: 438.1790.

Select data for **5b**: m.p. 164–165 °C; ¹H NMR (500 MHz, CD₃CN): δ 7.33 (d, $J = 8.5$ Hz, 2 H), 7.16 – 7.11 (m, 6 H), 6.01 (d, $J = 12.5$ Hz, 1 H), 4.94 (dd, $J = 4.5$ Hz, 4.5 Hz, 1 H), 4.30 (dd, $J = 13.0$ Hz, 4.5 Hz, 1 H), 4.22 (br.s, 1 H), 3.50 (d, $J = 4.0$ Hz, 1 H), 2.98 – 2.89 (m, 1 H),

2.66 – 2.56 (m, 2 H), 2.33 – 2.24 (m, 1 H), 2.30 (s, 3 H), 2.27 (s, 3 H), 1.0 (s, 3 H); ^{13}C NMR (125 MHz, CD_3CN): δ 212.7 (C), 139.7 (C), 138.9 (C), 133.5 (C), 133.4 (C), 132.1 (2 CH), 130.8 (2 CH), 129.8 (2 CH), 128.0 (2 CH), 93.6 (CH), 87.7 (CH), 80.4 (C), 55.8 (C), 49.6 (CH), 43.6 (CH), 35.9 (CH_2), 28.8 (CH_2), 21.6 (CH_3), 21.11 (CH_3), 21.09 (CH_3).

Preparation of adduct **3c** and **5c**.



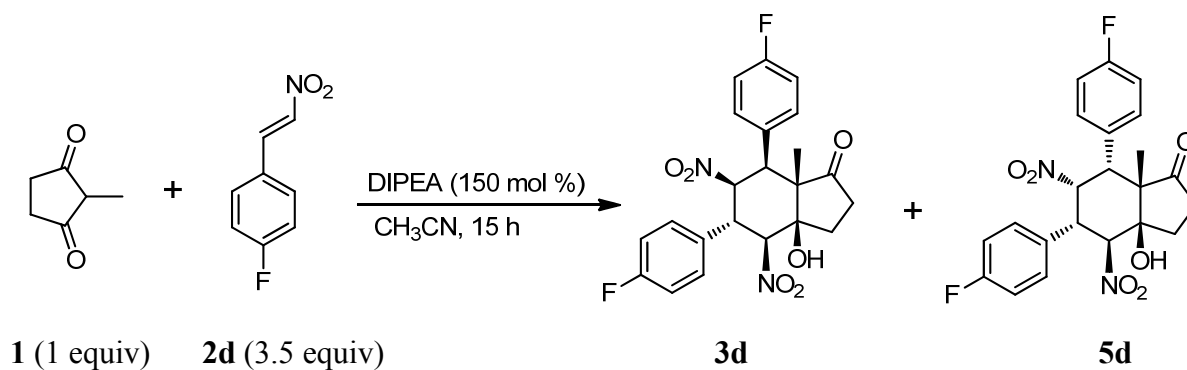
To a solution of DIPEA (44 mg, 0.34 mmol, 1.5 equiv), 2-methyl-1,3-cyclopentanedione (**1**, 25 mg, 0.22 mmol) in CH_3CN (1.1 mL) was added nitrostyrene (**2c**, 139 mg, 0.78 mmol, 3.5 equiv) and the resulting solution was stirred at room temperature for 20 h until the completion of the reaction, as monitored by TLC. Then the reaction mixture was directly filtered through a Celite bed (to remove insoluble polymer junk) and washed with EtOAc (15 mL). The filtrate was concentrated *in vacuo* and the crude product was purified by flash column chromatography with 20 % EtOAc–hexane to afford **3c** (21.5 mg, 20% yield) as white solid ($R_f = 0.13$ in 20% EtOAc–hexane, developed two times on TLC) and **5c** (4.2 mg, 4%) as a pale yellow solid ($R_f = 0.09$ in 20% EtOAc–hexane developed two times on TLC).

Select data for **3c**: m.p. 196–197 °C; IR (neat): 3550–3250, 2923, 2852, 1740, 1558, 1364, 1299, 1073, 1027, 811, 735 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3): δ 7.09 (d, $J = 8.5$ Hz, 2 H), 6.87 – 6.80 (m, 4 H), 6.76 (d, $J = 8.5$ Hz, 2 H), 4.83 (dd, $J = 12.0, 7.0$ Hz, 1 H), 4.48 (d, $J = 12.0$ Hz, 1 H), 4.36 (dd, $J = 12.5, 12.0$ Hz, 1 H), 4.16 (d, $J = 6.5$ Hz, 1 H), 3.78 (s, 3 H), 3.69 (s, 3 H), 3.46 (s, 1 H), 2.88 (dd, $J = 20.0$ Hz, 10.5 Hz, 1 H), 2.62 – 2.53 (m, 1 H), 2.31 (dd, $J = 13.5$ Hz, 9.5 Hz, 1 H), 2.22 – 2.14 (m, 1 H), 0.98 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3): δ 212.8 (C), 159.7 (C), 159.5 (C), 132.5 (2 CH), 125.6 (C), 125.3 (C), 114.5 (2 CH), 114.0 (2 CH), 92.7 (CH), 87.9 (CH), 78.2 (C), 57.2 (C), 55.2 (CH_3), 55.1 (CH_3), 47.5 (CH),

39.9 (CH), 33.9 (CH₂), 28.8 (CH₂), 18.3 (CH₃), some aryl carbons are broadened and disappeared due to the slow rotation and coalescence phenomenon; MS (*m/z*, relative intensity): 470 (M⁺, 11), 397 (14), 355 (19), 267 (17), 253 (17), 179 (19), 135 (100), 121 (75), 73 (60); exact mass calculated for C₂₄H₂₆N₂O₈ (M⁺): 470.1689; found: 470.1689.

Select data for **5c**: mp 103–104 °C; ¹H NMR (500 MHz, CDCl₃): δ 7.31 (d, *J* = 9.0 Hz, 2 H), 7.07 (d, *J* = 8.5 Hz, 2 H), 6.90 – 6.78 (m, 4 H), 6.13 (d, *J* = 12.5 Hz, 1 H), 4.84 (dd, *J* = 4.5 Hz, 4.0 Hz, 1 H), 4.11 (dd, *J* = 13.0 Hz, 4.5 Hz, 1 H), 3.77 (s, 3 H), 3.73 (s, 3 H), 3.36 (s, 1 H), 3.35 (d, *J* = 16.0 Hz, 1 H), 2.90 – 2.82 (m, 1 H), 2.70 – 2.60 (m, 1 H), 2.36 – 2.30 (m, 1 H), 2.20 – 2.14 (m, 1 H), 1.11 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃): δ 209.8 (C), 160.0 (C), 159.6 (C), 132.0 (2 CH), 128.4 (2 CH), 125.9 (C), 124.8 (C), 114.9 (2 CH), 113.8 (2 CH), 92.4 (CH), 87.1 (CH), 78.8 (C), 55.20 (CH₃), 55.19 (CH₃), 54.6 (C), 49.1 (CH), 43.6 (CH), 35.2 (CH₂), 29.0 (CH₂), 20.8 (CH₃).

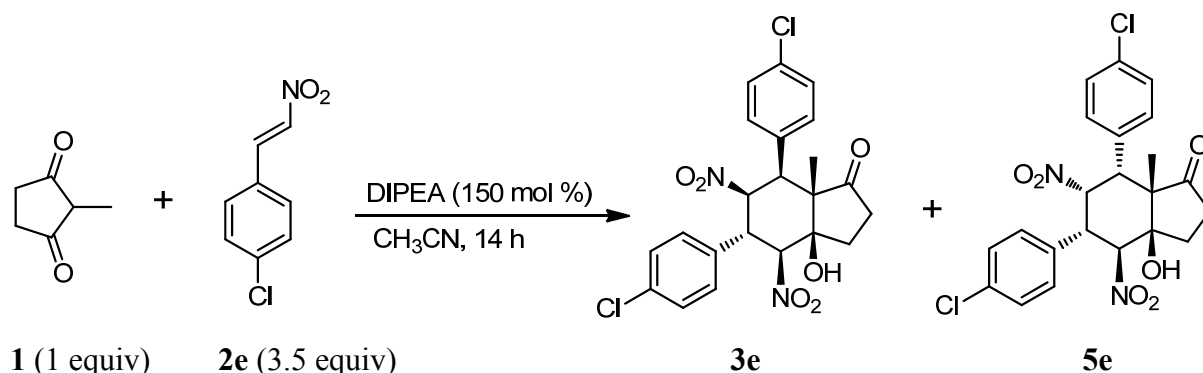
Preparation of adduct **3d** and **5d**.



To a solution of DIPEA (44 mg, 0.34 mmol, 1.5 equiv), 2-methyl-1,3-cyclopentanedione (**1**, 25 mg, 0.22 mmol) in CH₃CN (1.1 mL) was added nitrostyrene (**2d**, 130.2 mg, 0.78 mmol, 3.5 equiv) and the resulting solution was stirred at room temperature for 15 h until the completion of the reaction, as monitored by TLC. Then the reaction mixture was directly filtered through a Celite bed (to remove insoluble polymer junk) and washed with EtOAc (15 mL). The filtrate was concentrated *in vacuo* and the crude product was purified by flash column chromatography with 20 % EtOAc–hexane to afford **3d** (59.7 mg, 60% yield) as

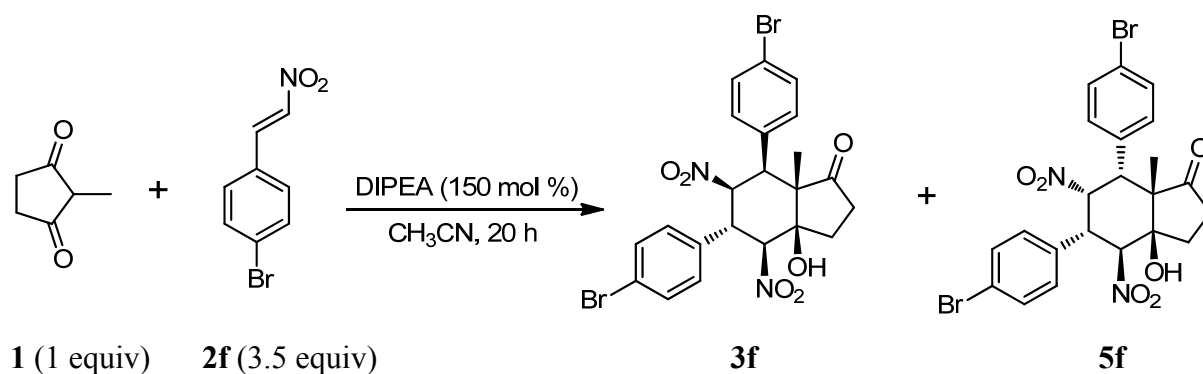
white solid ($R_f = 0.19$ in 20% EtOAc–hexane, developed two times on TLC) and **5d** (7.0 mg, 7% yield) as a pale yellow solid ($R_f = 0.13$ in 20% EtOAc–hexane developed two times on TLC). Select data for **3d**: m.p. 208–209 °C; IR (neat): 3550–3200, 2922, 2852, 1740, 1606, 1558, 1512, 1364, 1233, 1164, 840, 735 cm^{-1} ; ^1H NMR (500 MHz, CD_3CN): δ 7.42 – 7.39 (m, 2 H), 7.18 – 7.03 (m, 4 H), 7.01 (dd, $J = 9.0$ Hz, 9.0 Hz, 2 H), 4.91 (dd, $J = 12.5$, 6.5 Hz, 1 H), 4.80 (d, $J = 12.0$ Hz, 1 H), 4.55 (s, 1 H), 4.38 (dd, $J = 12.0$, 12.0 Hz, 1 H), 4.12 (d, $J = 6.5$ Hz, 1 H), 2.90 – 2.82 (m, 1 H), 2.80 – 2.70 (m, 1 H), 2.61 – 2.56 (m, 1 H), 2.20 – 2.11 (m, 1 H), 0.90 (s, 3 H); ^{13}C NMR (125 MHz, CD_3CN): δ 214.4 (C), 163.7 (d, $J = 243.9$ Hz, C), 163.4 (d, $J = 244.4$ Hz, C), 134.5 (2 CH), 132.6 (d, $J = 3.4$ Hz, C), 131.4 (d, $J = 3.4$ Hz, C), 116.4 (d, $J = 21.6$ Hz, 2 CH), 116.2 (d, $J = 21.3$ Hz, 2 CH), 92.6 (CH), 89.2 (CH), 80.0 (C), 58.1 (C), 49.3 (CH), 40.7 (CH), 34.4 (CH_2), 28.8 (CH_2), 19.1 (CH_3), two aryl carbons and two aryl protons are broadened and disappeared due to the slow rotation and coalescence phenomenon; MS (m/z , relative intensity): 446 (M^+ , 11), 400 (5), 354 (20), 293 (34), 281 (21), 227 (17), 133 (35), 123 (84), 109 (100); exact mass calculated for $\text{C}_{22}\text{H}_{20}\text{F}_2\text{N}_2\text{O}_6$ (M^+): 446.1289; found: 446.1287.

Selected data for **5d**: m.p. 170–171 °C; ^1H NMR (500 MHz, CD_3CN): δ 7.53 – 7.49 (m, 2 H), 7.34 – 7.30 (m, 2 H), 7.10 – 7.04 (m, 4 H), 6.00 (d, $J = 13.0$ Hz, 1 H), 4.97 (dd, $J = 4.0$ Hz, 4.5 Hz, 1 H), 4.38 (dd, $J = 13.0$ Hz, 4.5 Hz, 1 H), 4.23 (s, 1 H), 3.58 (d, $J = 4.0$ Hz, 1 H), 2.97–2.90 (m, 1 H), 2.70 – 2.59 (m, 2 H), 2.18 – 2.11 (m, 1 H) 1.03 (s, 3 H); ^{13}C NMR (125 MHz, CD_3CN): δ 212.8 (C), 163.6 (d, $J = 244.5$ Hz, C), 163.5 (d, $J = 243.6$ Hz, C), 134.2 (d, $J = 8.4$ Hz, 2 CH), 132.6 (d, $J = 3.4$ Hz, C), 132.4 (d, $J = 3.3$ Hz, C), 130.2 (d, $J = 8.4$ Hz, 2 CH), 117.1 (d, $J = 21.8$ Hz, 2 CH), 115.8 (d, $J = 21.1$ Hz, 2 CH), 93.3 (CH), 87.6 (CH), 80.4 (C), 55.8 (C), 49.1 (CH), 43.2 (CH), 35.8 (CH_2), 28.8 (CH_2), 21.5 (CH_3).

Preparation of adduct **3e** and **5e**.

To a solution of 2-methyl-1,3-cyclopentanedione (**1a**, 25.0 mg, 0.223 mmol, 1.0 equiv) and diisopropylethylamine (43.3 mg, 0.335 mmol, 1.5 equiv) in CH₃CN (1.1 mL) was added nitrostyrene (**2e**, 143.2 mg, 0.78 mmol, 3.5 equiv) and the resulting solution was stirred at room temperature for 14 h until the completion of the reaction, as monitored by TLC. Then the reaction mixture was directly filtered through a Celite bed (to remove insoluble polymer junk) and washed with EtOAc (15 mL). The filtrate was concentrated *in vacuo* and the crude product was purified by flash column chromatography with 20 % EtOAc-hexane to afford **3e** (60 mg, 56% yield) as white solid ($R_f = 0.21$ developed two times in 20 % EtOAc-hexane) and **5e** (7.5 mg, 7% yield) as a pale yellow solid ($R_f = 0.16$ in 20% EtOAc-hexane developed two times on TLC). Select data for **3e**: m.p. 231–232 °C; IR (neat): 3491, 2922, 2851, 1740, 1558, 1493, 1468, 1415, 1362, 1095, 836, 739 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ 7.41 – 7.38 (m, 4H), 7.27 (d, $J = 6.5$ Hz, 4H), 4.92 (dd, $J = 12.5$ Hz, 6.5 Hz, 1H), 4.80 (d, $J = 12.0$ Hz, 1H), 4.58 (s, 1H), 4.36 (dd, $J = 12.5$ Hz, 12.0 Hz, 1H), 4.11 (d, $J = 6.5$ Hz, 1H), 2.90 – 2.84 (m, 1H), 2.79 – 2.71 (m, 1H), 2.61 – 2.57 (m, 1H), 2.14 – 2.11 (m, 1H), 0.90 (s, 3H); ¹³C NMR (125 MHz, CD₃CN): δ 214.3 (C), 135.4 (C), 134.9 (C), 134.8 (C), 134.1 (C), 129.7 (4 CH), 129.6 (4 CH), 92.4 (CH), 89.0 (CH), 79.9 (C), 58.0 (C), 49.4 (CH), 40.9 (CH), 34.4 (CH₂), 28.7 (CH₂), 19.1 (CH₃); exact mass calculated for C₂₂H₂₀Cl₂N₂O₆ (M⁺): 478.0698; found: 478.0696.

Selected data for **5e**: m.p. 137–138 °C; ¹H NMR (500 MHz, CD₃CN): δ 7.49 (dd, $J = 11.5$ Hz, 3.0 Hz, 2 H), 7.36 – 7.32 (m, 4 H), 7.30 – 7.27 (m, 2 H), 5.99 (d, $J = 12.5$ Hz, 1 H), 4.97 (dd, $J = 4.5$ Hz, 4.0 Hz, 1 H), 4.38 (dd, $J = 13.0$ Hz, 4.5 Hz, 1 H), 4.32 (s, 1 H), 3.58 (d, $J = 4.0$ Hz, 1 H), 3.00 – 2.90 (m, 1 H), 2.70 – 2.59 (m, 2 H), 2.11–2.07 (m, 1 H), 1.03 (s, 3 H); ¹³C NMR (125 MHz, CD₃CN): δ 212.7 (C), 135.3 (C), 135.2 (C), 135.1 (C), 134.7 (C), 133.9 (2 CH), 130.3 (2 CH), 129.8 (2 CH), 129.2 (2 CH), 92.9 (CH), 87.3 (CH), 80.4 (C), 55.8 (C), 49.2 (CH), 43.3 (CH), 35.8 (CH₂), 28.7 (CH₂), 21.5 (CH₃).

Preparation of adduct **3f** and **5f**.

To a solution of DIPEA (44 mg, 0.34 mmol, 1.5 equiv), 2-methyl-1,3-cyclopentanedione (**1**, 25 mg, 0.22 mmol) in CH₃CN (1.1 mL) was added nitrostyrene (**2f**, 178 mg, 0.78 mmol, 3.5 equiv) and the resulting solution was stirred at room temperature for 20 h until the completion of the reaction, as monitored by TLC. Then the reaction mixture was directly filtered through a Celite bed (to remove insoluble polymer junk) and washed with EtOAc (15 mL). The filtrate was concentrated *in vacuo* and the crude product was purified by flash column chromatography with 20 % EtOAc–hexane to afford **3f** (57.8 mg, 46% yield) as white solid ($R_f = 0.20$ in 20% EtOAc–hexane, developed two times on TLC) and **5f** (12.7 mg, 10% yield) as a pale yellow solid ($R_f = 0.15$ in 20% EtOAc–hexane developed two times on TLC). Select data for **3f**: m.p. 232–233 °C; IR (neat): 3600–3400, 2922, 2852, 1740, 1558, 1487, 1362, 1068, 1013, 828, 736 cm⁻¹; ¹H NMR (500 MHz, CS₂/CDCl₃): δ 7.47 – 7.41 (m, 4 H), 7.39 (d, $J = 8.5$ Hz, 2 H), 7.05 (d, $J = 8.5$ Hz, 2 H), 4.82 (dd, $J = 12.0, 6.5$ Hz, 1 H), 4.46 (d, $J = 12.5$ Hz, 1 H), 4.35 (dd, $J = 12.5, 12.5$ Hz, 1 H), 4.17 (d, $J = 7.0$ Hz, 1 H), 3.46 (s, 1 H), 2.95 – 2.88 (m, 1 H), 2.64 – 2.53 (m, 1 H), 2.40 – 2.31 (m, 1 H), 2.23 – 2.15 (m, 1 H), 0.98 (s, 3 H); ¹³C NMR (125 MHz, CS₂/CDCl₃): δ 211.8 (C), 132.5 (C), 132.3 (4 CH), 132.1 (C), 131.9 (4 CH), 123.3 (C), 123.1 (C), 91.9 (CH), 87.2 (CH), 78.1 (C), 56.8 (C), 47.5 (CH), 40.0 (CH), 33.7 (CH₂), 28.5 (CH₂), 18.1 (CH₃); IR (neat): 3725, 3705, 3654, 3634, 2922, 2852, 1740, 1558, 1487, 1362, 1299, 1170, 1068, 1013, 828, 736 cm⁻¹; ¹H NMR (500 MHz, CDCl₃): δ 7.47 – 7.45 (m, 4 H), 7.39 (d, $J = 8.5$ Hz, 2 H), 7.06 (d, $J = 8.5$ Hz, 2 H), 4.82 (dd, $J = 5.0, 7.0$ Hz, 1 H), 4.46 (d, $J = 12.0$ Hz, 1 H), 4.35 (dd, $J = 12.5, 12.5$ Hz, 1 H), 4.18 (d, $J = 7.0$ Hz, 1 H), 3.50 (s, 1 H), 2.94 – 2.86 (m, 1 H), 2.63 – 2.55 (m, 1 H), 2.39 – 2.34 (m, 1 H), 2.22 – 2.16 (m, 1 H), 0.98 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃): δ 212.1 (C), 132.7 (C), 132.4 (C), 132.2 (4 CH), 132.0 (4 CH), 123.2 (C), 123.1 (C), 92.0 (CH), 87.3 (CH), 78.2 (C), 56.9 (C), 47.6 (CH), 40.1 (CH), 33.8 (CH₂), 28.5 (CH₂), 18.2 (CH₃); MS (m/z , relative intensity): 570 ($M^+ + 4$, 20), 568 ($M^+ + 2$, 40), 566 (M^+ , 20), 476 (51), 415 (39), 403 (28), 349 (34), 284 (32), 202 (57), 169 (72), 102 (100); exact mass calculated for C₂₂H₂₀Br₂N₂O₆ (M^+): 565.9688; found: 565.9688.

Select data for **5f**: m.p. 151–152 °C; ^1H NMR (500 MHz, CDCl_3): δ 7.44 (d, $J = 8.0$ Hz, 4 H), 7.29 (d, $J = 8.0$ Hz, 2 H), 7.04 (d, $J = 8.0$ Hz, 2 H), 6.10 (d, $J = 13.0$ Hz, 1 H), 4.83 (dd, $J = 4.5$ Hz, 4.0 Hz, 1 H), 4.15 (dd, $J = 12.5$ Hz, 4.5 Hz, 1 H), 3.39 (d, $J = 4.0$ Hz, 1 H), 3.30 (s, 1 H), 2.90 – 2.83 (m, 1 H), 2.72 – 2.66 (m, 1 H), 2.41 – 2.35 (m, 1 H), 2.20 – 2.15 (m, 1 H), 1.11 (s, 3 H); ^{13}C NMR (125 MHz, CDCl_3): δ 209.3 (C), 132.8 (2 CH), 132.7 (C), 132.5 (2 CH), 131.8 (2 CH), 128.8 (2 CH), 123.7 (C), 123.0 (C), 91.3 (CH), 86.5 (CH), 78.8 (C), 54.4 (C), 49.3 (CH), 43.7 (CH), 35.0 (CH_2), 30.9 (C), 28.8 (CH_2), 20.7 (CH_3).

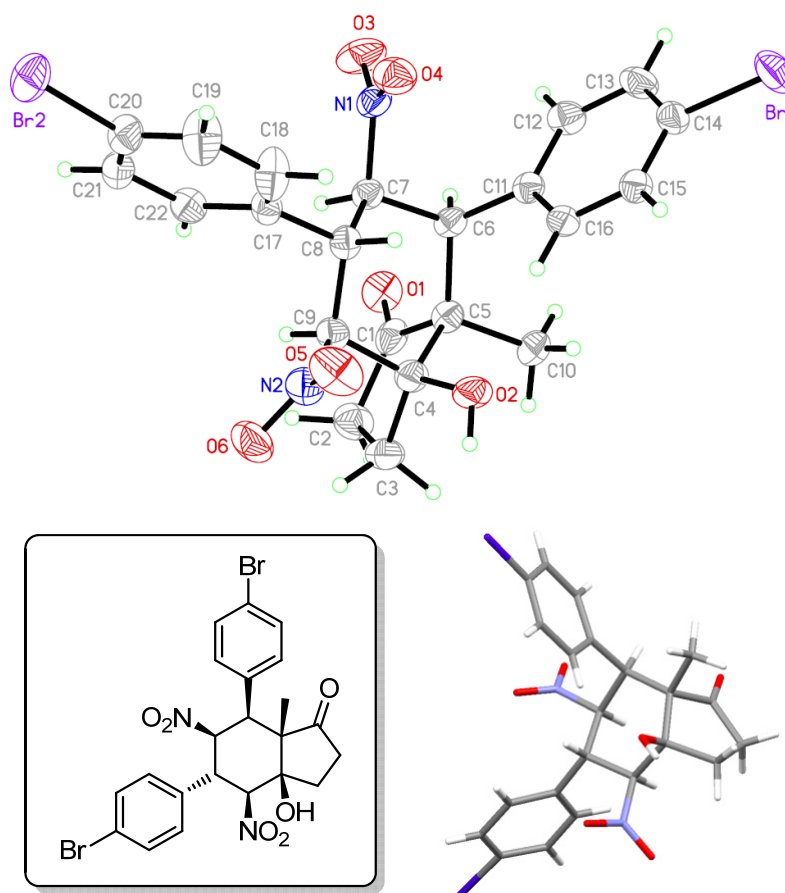


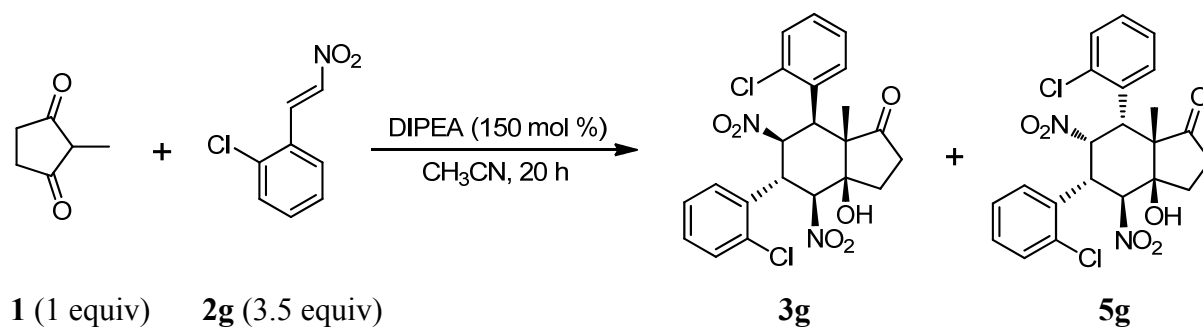
Figure S3. ORTEP and Stereo plots for X-ray crystal structures of (\pm)-**3f**.

CCDC 1474865 contains the supplementary crystallographic data for (\pm)-**3f**. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Table S3. Crystal data and structure refinement for (\pm)-**3f**.

Identification code	ic17312	
Empirical formula	$C_{25}H_{26}Br_2N_2O_7$	
Formula weight	626.30	
Temperature	150(2) K	
Wavelength	0.71073 Å	
Crystal system	Triclinic	
Space group	P-1	
Unit cell dimensions	a = 10.4202(7) Å	a = 61.4060(13)°.
	b = 12.1455(8) Å	b = 71.1351(14)°.
	c = 12.5464(8) Å	g = 78.7175(14)°.
Volume	1317.70(15) Å ³	
Z	2	
Density (calculated)	1.578 Mg/m ³	
Absorption coefficient	3.122 mm ⁻¹	
F(000)	632	
Crystal size	0.250 x 0.200 x 0.120 mm ³	
Theta range for data collection	1.912 to 27.498°.	
Index ranges	-13 ≤ h ≤ 13, -15 ≤ k ≤ 15, -16 ≤ l ≤ 16	
Reflections collected	17236	
Independent reflections	6063 [R(int) = 0.0345]	
Completeness to theta = 25.242°	100.0 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.706 and 0.509	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	6063 / 114 / 340	
Goodness-of-fit on F ²	1.035	
Final R indices [I > 2σ(I)]	R1 = 0.0430, wR2 = 0.0929	
R indices (all data)	R1 = 0.0579, wR2 = 0.0998	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.760 and -0.440 e.Å ⁻³	

Preparation of Compound **3g** and **5g**:

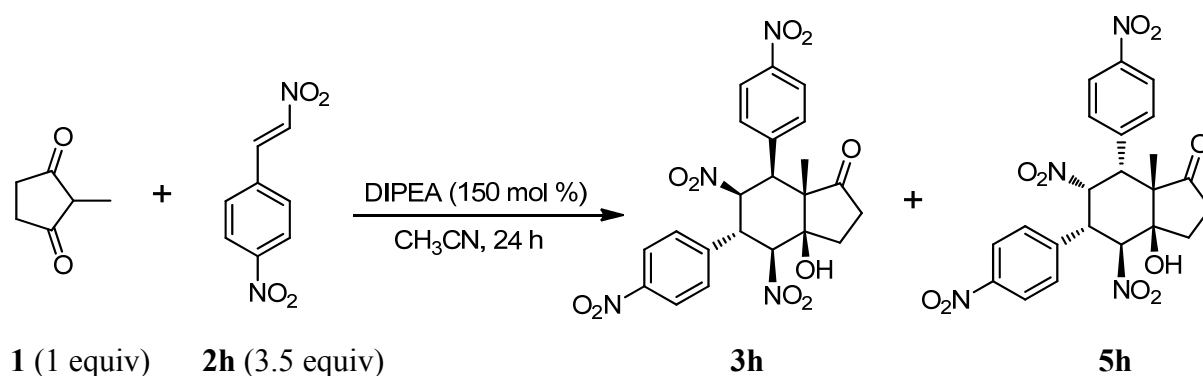


To a solution of 2-methyl-1,3-cyclopentanedione (**1**, 25.0 mg, 0.223 mmol, 1.0 equiv) and diisopropylethylamine (43.3 mg, 0.335 mmol, 1.5 equiv) in CH₃CN (1.1 mL) was added nitrostyrene (**2g**, 143.2 mg, 0.78 mmol, 3.5 equiv) and the resulting solution was stirred at room temperature for 20 h until the completion of the reaction, as monitored by TLC. The solution was diluted with EtOAc (10 mL), washed with water (5 mL) and brine (5 mL), dried over MgSO₄ and concentrated in *vacuo* to give the crude residue. The crude product was purified by flash column chromatography with 25 % EtOAc–hexane (R_f = 0.21 for **3g** in 20 % EtOAc–hexane developed two times on TLC) to afford **3g** (86.6 mg, 81% yield) as white solid (R_f = 0.21 for **3g** in 20 % EtOAc–hexane developed two times on TLC) and **5g** (11.8 mg, 11% yield) as white solid (R_f = 0.26 in 20% EtOAc–hexane developed two times on TLC); Select data for **3g**: m.p. 212–213 °C; IR (neat): 3550–3250, 3069, 3015, 2988, 2961, 1733, 1559, 1477, 1445, 1365, 1321, 1173, 1112, 1089, 1037, 980, 908, 811, 759 cm⁻¹; ¹H NMR (500 MHz, acetone-d₆): δ 8.67 (d, J = 8.0 Hz, 1 H), 7.89 – 7.86 (m, 1 H), 7.48 (dd, J = 8.0, 7.0 Hz, 1 H), 7.43 (d, J = 7.0 Hz, 1 H), 7.37 – 7.33 (m, 2 H), 7.26 – 7.22 (m, 2 H), 5.92 (s, 1 H), 5.42 (dd, J = 12.0, 12.0 Hz, 1 H), 5.33 (d, J = 6.5 Hz, 1 H), 5.28 (d, J = 12.0 Hz, 1 H), 5.19 (dd, J = 12.5 Hz, 6.5 Hz, 1 H), 2.92 – 2.86 (m, 2 H), 2.78 – 2.70 (m, 1 H), 2.32 (dd, J = 10.5 Hz, 3.0 Hz, 1 H), 0.94 (s, 3 H); ¹³C NMR (125 MHz, acetone-d₆): δ 214.0 (C), 137.7 (C), 137.2 (C), 134.8 (C), 134.5 (CH), 133.2 (C), 131.1 (CH), 130.8 (CH), 130.5 (CH), 130.4 (CH), 128.3 (CH), 128.1 (CH), 128.0 (CH), 92.4 (CH), 88.8 (CH), 80.2 (C), 58.4 (C), 43.6 (CH), 37.0 (CH), 34.0 (CH₂), 28.7 (CH₂), 18.8 (CH₃); MS (m/z , relative intensity): 480 (M⁺+2, 13), 478 (M⁺, 20), 443 (36), 367 (48), 325 (100), 202 (30), 165 (27), 139 (36), 125 (82); exact mass calculated for C₂₂H₂₀Cl₂N₂O₆ (M⁺): 478.0698; found: 478.0703.

Select data for **5g**: m.p. 99–100°C; ¹H NMR (500 MHz, acetone-d₆): δ 8.38 (dd, J = 8.0

Hz, 1.5 Hz, 1 H), 7.56 (d, $J = 8.0$ Hz, 1 H), 7.50 – 7.40 (m, 3 H), 7.35 – 7.28 (m, 3 H), 6.30 (d, $J = 12.0$ Hz, 1 H), 5.91 (s, 1 H), 5.02 – 4.98 (m, 2 H), 4.91 (d, $J = 2.5$ Hz, 1 H), 3.12 – 3.08 (m, 1 H), 2.98 – 2.93 (m, 1 H), 2.84 – 2.79 (m, 1 H), 2.51 – 2.48 (m, 1 H), 1.07 (s, 3 H); ^{13}C NMR (125 MHz, acetone- d_6): δ 213.5 (C), 135.73 (C), 135.68 (C), 134.9 (C), 133.9 (C), 132.4 (CH), 131.2 (CH), 131.1 (CH), 130.9 (CH), 130.6 (CH), 128.8 (CH), 128.4 (CH), 127.6 (CH), 89.9 (CH), 87.5 (CH), 80.1 (C), 56.5 (C), 43.4 (CH), 37.1 (CH), 34.8 (CH $_2$), 31.0 (CH $_2$), 18.9 (CH $_3$).

Preparation of adduct **3h** and **5h**.

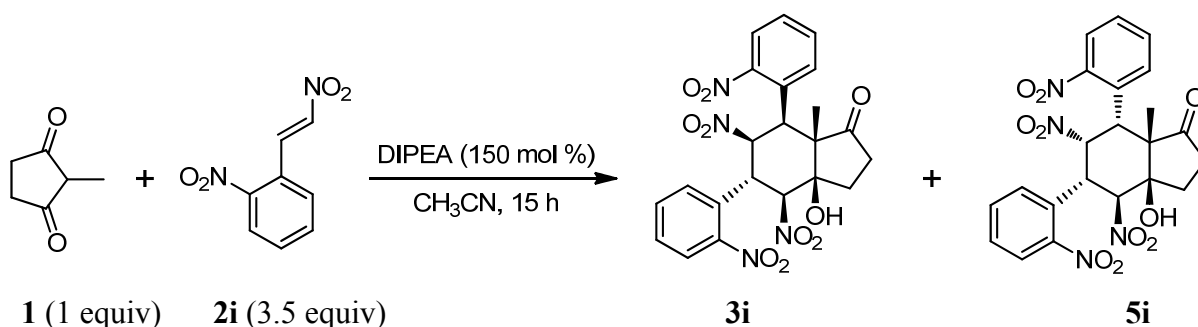


To a solution of DIPEA (44 mg, 0.34 mmol, 1.5 equiv), 2-methyl-1,3-cyclopentanedione (**1**, 25 mg, 0.22 mmol) in CH_3CN (1.1 mL) was added nitrostyrene (**2h**, 151.4 mg, 0.78 mmol, 3.5 equiv) and the resulting solution was stirred at room temperature for 24 h until the completion of the reaction, as monitored by TLC. Then the reaction mixture was directly filtered through a Celite bed (to remove insoluble polymer junk) and washed with EtOAc (15 mL). The filtrate was concentrated *in vacuo* and the crude product was purified by flash column chromatography with 30% EtOAc–hexane to afford **3h** (85.9 mg, 77% yield) as white solid ($R_f = 0.22$ in 30% EtOAc–hexane developed two times on TLC) and **5h** (19.0 mg, 17% yield) as pale yellow solid ($R_f = 0.17$ in 30% EtOAc–hexane developed two times on TLC). Select data for **3h**: m.p. 194 °C (decompose). IR (neat): 3550–3300, 3118, 3082, 2922, 2856, 1743, 1604, 1559, 1522, 1350, 1110, 1068, 858, 734 cm^{-1} ; ^1H NMR (500 MHz, CD_3CN): δ

8.40 – 8.30 (m, 4 H), 8.27 (d, $J = 8.5$ Hz, 2 H), 7.82 (d, $J = 8.5$ Hz, 2 H), 5.22 (dd, $J = 12.5$ Hz, 6.5 Hz, 1H), 5.08 (d, $J = 12.0$ Hz, 1 H), 4.98 (d, $J = 7.5$ Hz, 1 H), 4.73 (dd, $J = 12.0, 12.5$ Hz, 1 H), 4.47 (d, $J = 6.5$ Hz, 1 H), 3.07 (dd, $J = 20.5$ Hz, 9.0 Hz, 1 H), 3.00 – 2.90 (m, 1 H), 2.83 – 2.78 (m, 1 H), 1.08 (s, 3 H); ^{13}C NMR (125 MHz, CD_3CN): δ 213.7 (C), 149.1 (C), 143.7 (C), 142.5 (C), 133.9 (C), 124.7 (4 CH), 124.5 (4 CH), 92.0 (CH), 88.6 (CH), 80.0 (C), 57.9 (C), 49.5 (CH), 41.4 (CH), 34.4 (CH_2), 28.5 (CH_2), 19.0 (CH_3); MS (m/z , relative intensity): 500 (M^+ , 1), 134 (59), 104 (87), 92 (81), 89 (55), 77 (100), 76 (71); exact mass calculated for $\text{C}_{22}\text{H}_{20}\text{N}_4\text{O}_{10}$ (M^+): 500.1179; found: 500.1182.

Selected data for **5h**: mp. 150–151 °C; ^1H NMR (500 MHz, CD_3CN): δ 8.17 (d, $J = 8.5$ Hz, 2 H), 8.16 ($J = 8.5$ Hz, 2 H), 7.78 (d, $J = 8.5$ Hz, 2 H), 7.57 (d, $J = 8.5$ Hz, 2 H), 6.09 (d, $J = 12.5$ Hz, 1 H), 5.08 (dd, $J = 4.5$ Hz, 4.0 Hz, 1 H), 4.59 (dd, $J = 12.5$ Hz, 4.5 Hz, 1 H), 3.80 (d, $J = 4.0$ Hz, 1 H), 3.02 – 2.95 (m, 1 H), 2.78 (dd, $J = 13.5$ Hz, 9.0 Hz, 1 H), 2.67 (dd, $J = 20.0$ Hz, 9.0 Hz, 1 H), 2.23 – 2.08 (m, 1 H), 1.06 (s, 3 H); ^{13}C NMR (125 MHz, CD_3CN): δ 212.7 (C), 149.2 (C), 148.9 (C), 143.5 (2 C), 133.5 (2 CH), 129.5 (2 CH), 125.3 (2 CH), 124.1 (2 CH), 91.9 (CH), 87.1 (CH), 80.5 (C), 55.9 (C), 49.4 (CH), 43.7 (CH), 35.7 (CH_2), 28.6 (CH_2), 21.4 (CH_3).

Preparation of Compound **3i** and **5i**:



To a solution of DIPEA (44 mg, 0.34 mmol, 1.5 equiv), 2-methyl-1,3-cyclopentanedione (**1**, 25 mg, 0.22 mmol) in CH_3CN (1.1 mL) was added nitrostyrene (**2i**, 151.4 mg, 0.78 mmol, 3.5 equiv) and the resulting solution was stirred at room temperature for 15 h until the completion of the reaction, as monitored by TLC. The solution was diluted with EtOAc (10

mL), washed with water (5 mL) and brine (5 mL), dried over MgSO₄ and concentrated in *vacuo* to give the crude residue. The crude product was purified by flash column chromatography with 20% to 30 % EtOAc–hexane ($R_f = 0.14$ for **5i**, $R_f = 0.07$ for **3i**, in 20% EtOAc–hexane developed twice on TLC) to afford **5i** as pale yellow solid (33 mg, 30% yield) and **3i** as pale yellow solid (42 mg, 38% yield).

Selected data for **3i**: m.p. 184 °C (decomposed). IR (neat): 3550–3200, 2924, 2854, 1743, 1561, 1529, 1449, 1351, 1073, 855, 787, 735 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ 8.62 (d, $J = 7.0$ Hz, 1 H), 7.90 (d, $J = 8.5$ Hz, 1 H), 7.83 (dd, $J = 8.0$ Hz, 8.0 Hz, 1 H), 7.74 – 7.70 (m, 2 H), 7.64 – 7.56 (m, 2 H), 7.47 (dd, $J = 8.5$ Hz, 7.0 Hz, 1 H), 5.83 (dd, $J = 12.0$, 11.5 Hz, 1 H), 4.98 (dd, $J = 11.5$, 6.5 Hz, 1 H), 4.95 (d, $J = 6.5$ Hz, 1 H), 4.92 (brs, 1 OH), 4.89 (d, $J = 12.0$ Hz, 1 H), 2.91 – 2.88 (m, 1 H), 2.80 – 2.76 (m, 1 H), 2.64 – 2.60 (m, 1 H), 2.19 – 2.17 (m, 1 H), 1.00 (s, 3 H); ¹³C NMR (125 MHz, CD₃CN): δ 213.7 (C), 153.7 (C), 151.7 (C), 134.84 (CH), 134.77 (CH), 133.3 (CH), 130.9 (CH), 130.8 (CH), 130.7 (C), 128.4 (CH), 127.7 (C), 127.1 (CH), 125.5 (CH), 92.3 (CH), 88.6 (CH), 80.3 (C), 58.3 (C), 42.3 (CH), 34.3 (CH), 34.2 (CH₂), 28.3 (CH₂), 18.5 (CH₃); ¹H NMR (500 MHz, acetone-d₆): δ 8.81 (d, $J = 8.0$ Hz, 1 H), 8.08 (d, $J = 8.0$ Hz, 1 H), 7.98 (d, $J = 7.0$ Hz, 1 H), 7.85 (dd, $J = 8.0$, 1.5 Hz, 1 H), 7.76 (dd, $J = 8.0$ Hz, 1.5 Hz, 1 H), 7.67 – 7.62 (m, 2 H), 7.55 – 7.51 (m, 1 H), 6.15 (s, 1 H), 6.07 (dd, $J = 12.0$ Hz, 12.0 Hz, 1 H), 5.36 (d, $J = 12.0$ Hz, 1 H), 5.26 (dd, $J = 12.5$, 6.5 Hz, 1 H), 5.04 (d, $J = 6.5$ Hz, 1 H), 2.96 – 2.92 (m, 1 H), 2.88 – 2.72 (m, 2 H), 2.40 – 2.32 (m, 1 H), 1.09 (s, 3H); ¹³C NMR (125 MHz, acetone-d₆): δ 213.2 (C), 153.9 (C), 151.7 (C), 134.9 (CH), 134.6 (CH), 132.9 (CH), 131.5 (C), 130.6 (CH), 130.4 (CH), 128.4 (CH), 128.0 (C), 126.9 (CH), 125.2 (CH), 92.5 (CH), 88.7 (CH), 80.3 (C), 58.2 (C), 42.3 (CH), 34.4 (CH), 33.9 (CH₂), 28.5 (CH₂), 18.5 (CH₃); exact mass calculated for C₂₂H₂₀N₄O₁₀ (M⁺): 500.1179; found: 500.1182.

Selected data for **5i**: m.p 186–187 °C. IR (neat): 3550–3250, 2951, 2923, 2852, 1745, 1556, 1525, 1459, 1348, 1296, 857, 789, 739 cm⁻¹; ¹H NMR (500 MHz, CD₃CN): δ 8.17 (d, $J = 8.0$ Hz, 1 H), 7.99 (d, $J = 8.0$ Hz, 1 H), 7.91 (d, $J = 8.0$ Hz, 1 H), 7.79 (dd, $J = 8.0$ Hz, 6.0 Hz, 1 H), 7.64 – 7.60 (m, 2 H), 7.56 – 7.52 (m, 1 H), 7.36 (d, $J = 8.0$ Hz, 1 H), 6.04 (d, $J = 12.5$ Hz, 1 H), 5.54 (dd, $J = 6.0$ Hz, 5.5 Hz, 1 H), 4.99 (dd, $J = 12.5$ Hz, 6.0 Hz, 1 H), 4.56 (d, $J = 5.5$ Hz, 1 H), 2.95 – 2.90 (m, 1 H), 2.85 – 2.77 (m, 1 H), 2.75 – 2.68 (m, 1 H), 2.42 – 2.36 (m, 1 H), 1.03 (s, 3 H); ¹³C NMR (125 MHz, CD₃CN): δ 214.5 (C), 151.4 (C), 150.9 (C),

135.3 (CH), 134.2 (CH), 132.6 (CH), 131.1 (CH), 130.9 (CH), 130.7 (C), 130.6 (C), 127.9 (CH), 127.0 (CH), 126.1 (CH), 90.7 (CH), 88.8 (CH), 80.7 (C), 56.6 (C), 41.5 (CH), 36.4 (CH), 35.3 (CH₂), 31.6 (CH₂), 18.3 (CH₃).

A sample of 15 mg of racemic **3i**, prepared as described above, was dissolved with CH₂Cl₂ (15 mL) in a vial, followed by the slow addition of hexane (2 mL) down the inside of the vial and the two-layer solution was maintained throughout the addition process. The solution was capped and left to stand at room temperature for slow evaporation over 5 days to give some crystals (ca. 100 crystals) of the compound **3i**. A crystal was subjected for the single crystal X-ray analysis (crystal ID: ic17971, as shown in the **Figure S4** and **Table S4**). Surprisingly, a chiral space group: orthorhombic P2₁2₁2₁ was observed in the crystal! In order to understand the sign of its specific rotation and to probe the approximate frequency of occurrence of the SDE effect in the recrystallization process, another two crystals from the same batch were arbitrarily selected and subjected for single crystal X-ray analysis. Notably, each of them had the same chiral space group, orthorhombic P2₁2₁2₁, with the same absolute configuration as the previous crystal. The two single crystal X-ray analyses are shown in the **Figure S5**, **Table S5** (for crystal ID: ic18028), **Figure S6**, and **Table S6** (for crystal ID: ic18028_1). On the other hand, another batch of ca. 40 mg of racemic **3i** was subject to the same recrystallization process as described previously to give ca. 150 crystals. From them, 46 crystals were arbitrarily selected and individually subjected to the HPLC analysis with a chiral column (Chiralpack IC). Eluent: 25% THF–hexane, flow rate: 1.0 mL/min. The two enantiomers were separated at *R_t* 5.6 min and *R_t* 8.0 min. For the 46 crystals analyzed, 16 of them eluted with *R_t* 5.6 min (>99% *ee*), 14 of them eluted with *R_t* 8.0 min (>99% *ee*), 2 of them displayed >80% *ee*, 4 of them were almost a racemate, and 10 of them had low *ee*. The pure enantiomer was individually collected, checking with the aid of the HPLC analysis, and subjected to optical rotation analysis. For *R_t* 5.6 min: $[\alpha]_{\text{D}}^{29} +10.2$ (*c* 0.58, acetone); for *R_t* 8.0 min: $[\alpha]_{\text{D}}^{29} -9.1$ (*c* 0.36, acetone). Moreover, the above two crystals investigated by X-ray (CCDC 1488798 and CCDC 1488808) were individually subjected to HPLC analysis, comparing and co-injecting with the previous samples, with the conclusion that the chiral X-ray structures obtained herein are (–)-**3i**.

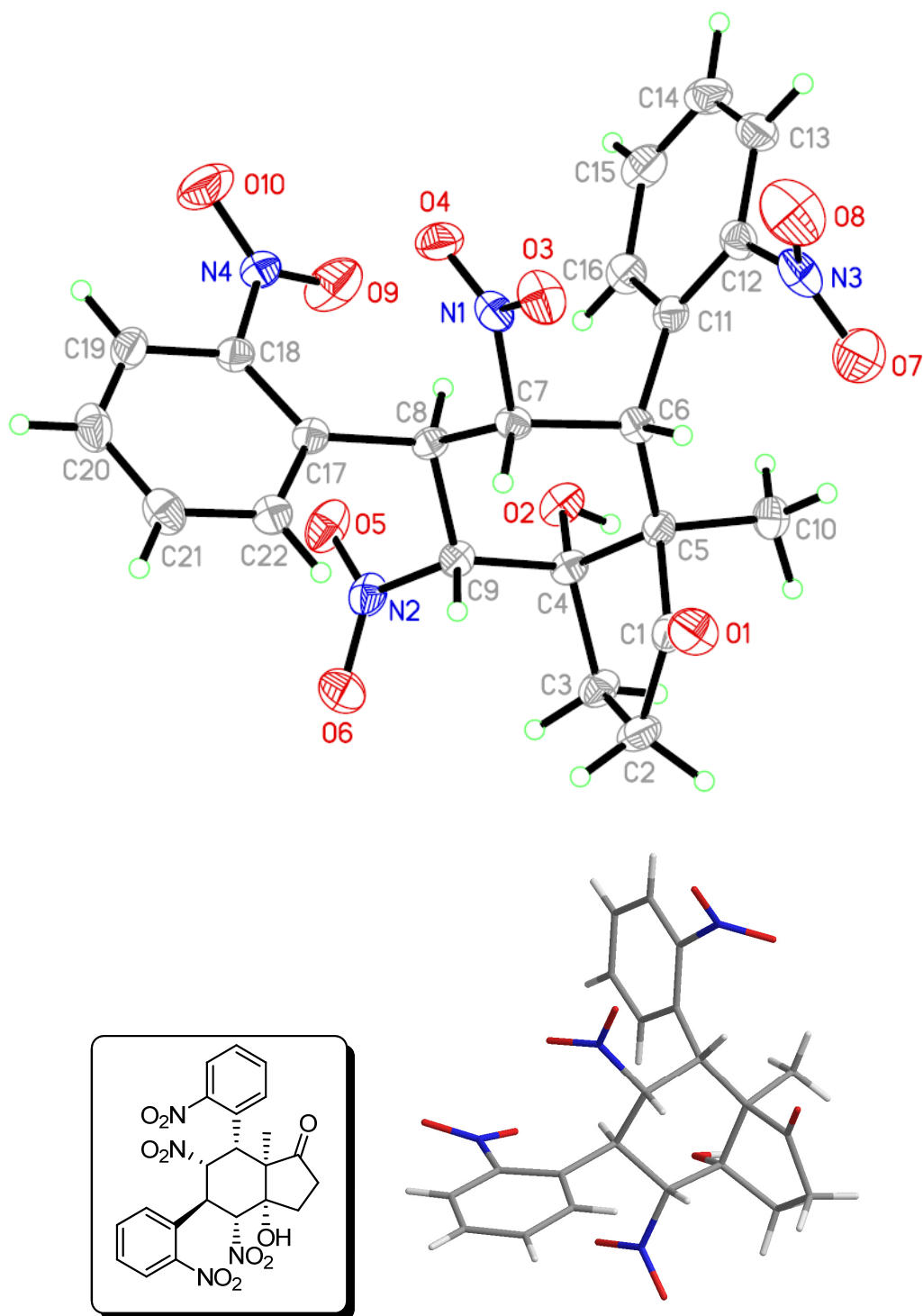


Figure S4. ORTEP, thermal ellipsoids drawn at the 50% probability level, and stereo plots for X-ray crystal structures of (-)-**3i**.

CCDC 1485986 contain the supplementary crystallographic data for this crystal. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Table S4. Crystal data and structure refinement for (-)-**3i**, first crystal (CCDC 1485986).

Identification code	ic17971	
Empirical formula	C ₂₂ H ₂₀ N ₄ O ₁₀	
Formula weight	500.42	
Temperature	200(2) K	
Wavelength	1.54178 Å	
Crystal system	Orthorhombic	
Space group	P2 ₁ 2 ₁ 2 ₁	
Unit cell dimensions	a = 9.3887(2) Å	$\alpha = 90^\circ$.
	b = 11.6113(3) Å	$\beta = 90^\circ$.
	c = 19.7433(5) Å	$\gamma = 90^\circ$.
Volume	2152.32(9) Å ³	
Z	4	
Density (calculated)	1.544 Mg/m ³	
Absorption coefficient	1.064 mm ⁻¹	
F(000)	1040	
Crystal size	0.222 x 0.182 x 0.166 mm ³	
Theta range for data collection	4.417 to 69.904°.	
Index ranges	-11 ≤ h ≤ 11, -14 ≤ k ≤ 14, -18 ≤ l ≤ 23	
Reflections collected	10046	
Independent reflections	4021 [R(int) = 0.0168]	
Completeness to theta = 67.679°	99.4 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7533 and 0.6531	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	4021 / 0 / 328	
Goodness-of-fit on F ²	1.047	
Final R indices [I > 2σ(I)]	R1 = 0.0255, wR2 = 0.0698	
R indices (all data)	R1 = 0.0258, wR2 = 0.0701	
Absolute structure parameter	0.06(3)	
Extinction coefficient	0.0039(3)	
Largest diff. peak and hole	0.227 and -0.158 e.Å ⁻³	

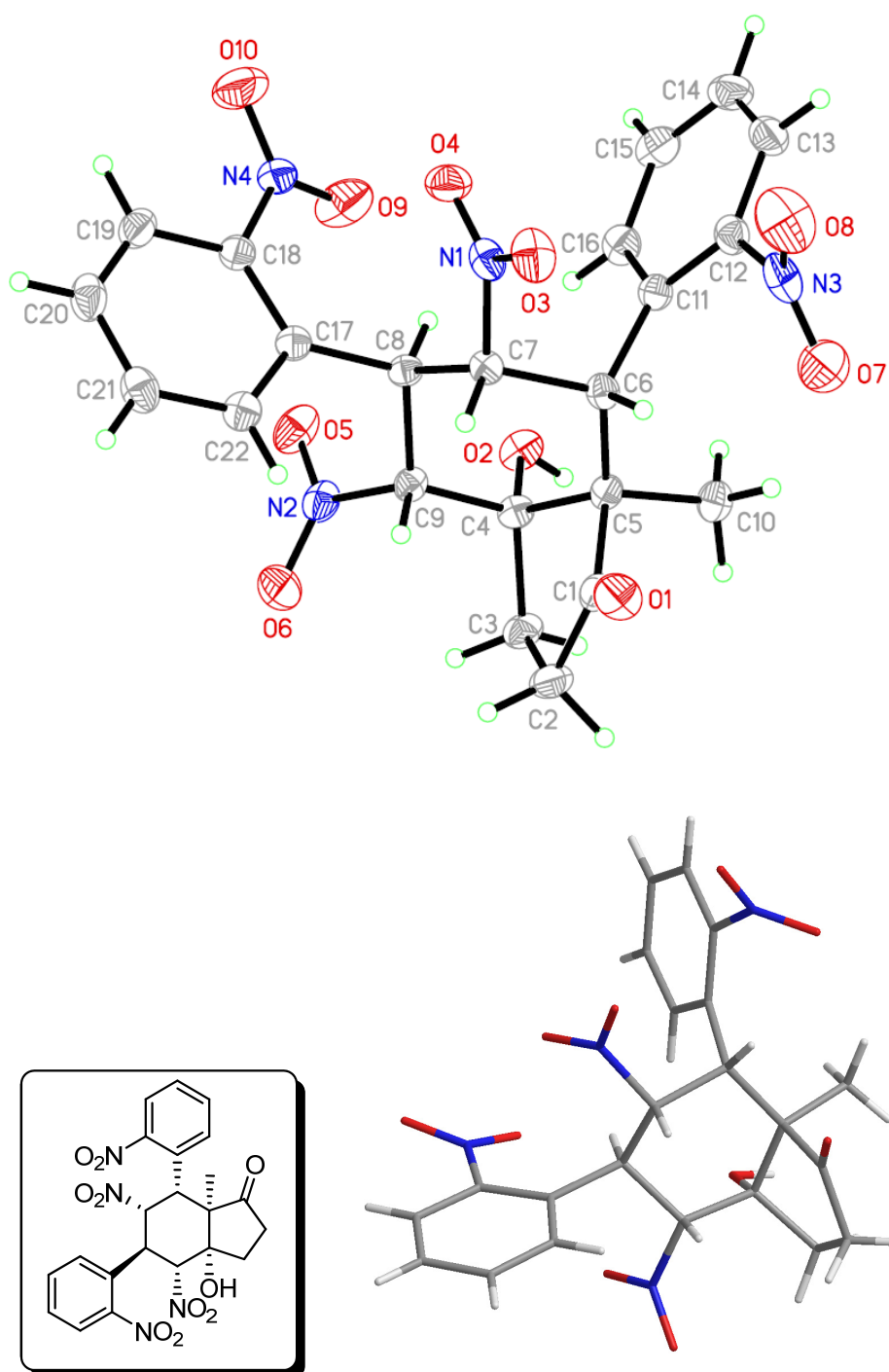


Figure S5. ORTEP, thermal ellipsoids drawn at the 50% probability level, and stereo plots for X-ray crystal structures of (-)-**3i**.

CCDC 1488798 contain the supplementary crystallographic data of this crystal data. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Table S5. Crystal data and structure refinement for (–)-**3i**, second crystal (CCDC 1488798).

Identification code	ic18028	
Empirical formula	C ₂₂ H ₂₀ N ₄ O ₁₀	
Formula weight	500.42	
Temperature	200(2) K	
Wavelength	1.54178 Å	
Crystal system	Orthorhombic	
Space group	P2 ₁ 2 ₁ 2 ₁	
Unit cell dimensions	a = 9.3839(2) Å	α = 90°.
	b = 11.6090(2) Å	β = 90°.
	c = 19.7514(4) Å	γ = 90°.
Volume	2151.67(7) Å ³	
Z	4	
Density (calculated)	1.545 Mg/m ³	
Absorption coefficient	1.064 mm ⁻¹	
F(000)	1040	
Crystal size	0.333 x 0.298 x 0.157 mm ³	
Theta range for data collection	4.418 to 69.962°.	
Index ranges	-10 ≤ h ≤ 11, -14 ≤ k ≤ 12, -24 ≤ l ≤ 23	
Reflections collected	12358	
Independent reflections	4066 [R(int) = 0.0174]	
Completeness to theta = 67.679°	100.0 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7533 and 0.6145	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	4066 / 0 / 328	
Goodness-of-fit on F ²	1.051	
Final R indices [I > 2σ(I)]	R1 = 0.0240, wR2 = 0.0617	
R indices (all data)	R1 = 0.0245, wR2 = 0.0621	
Absolute structure parameter	-0.01(3)	
Extinction coefficient	0.0047(3)	
Largest diff. peak and hole	0.224 and -0.128 e.Å ⁻³	

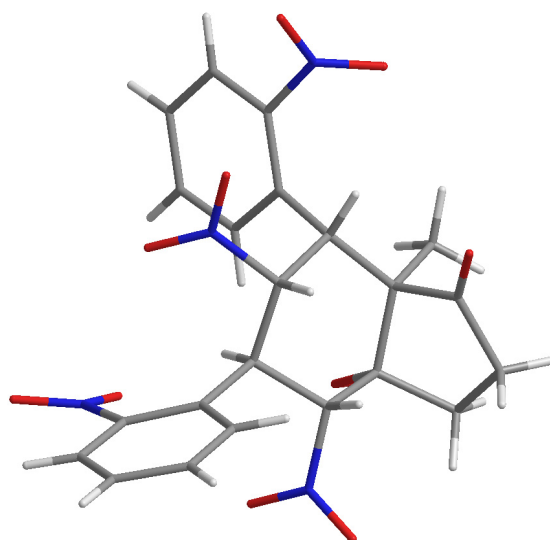
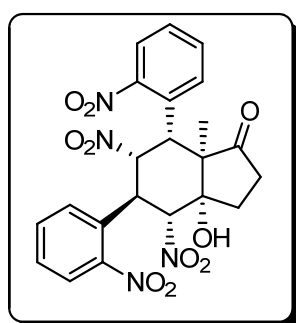
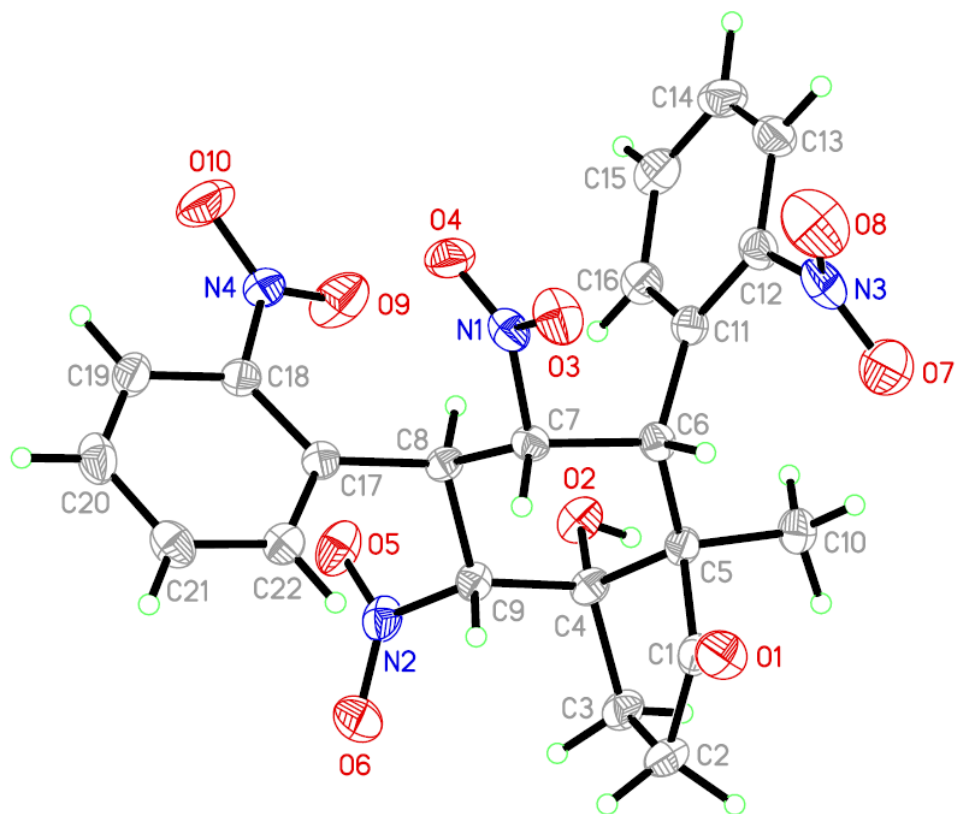


Figure S6. ORTEP, thermal ellipsoids drawn at the 50% probability level, and stereo plots for X-ray crystal structures of (-)-**3i**.

CCDC 1488808 contain the supplementary crystallographic data of this crystal. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Table S6. Crystal data and structure refinement for (-)-**3i**, third crystal (CCDC 1488808).

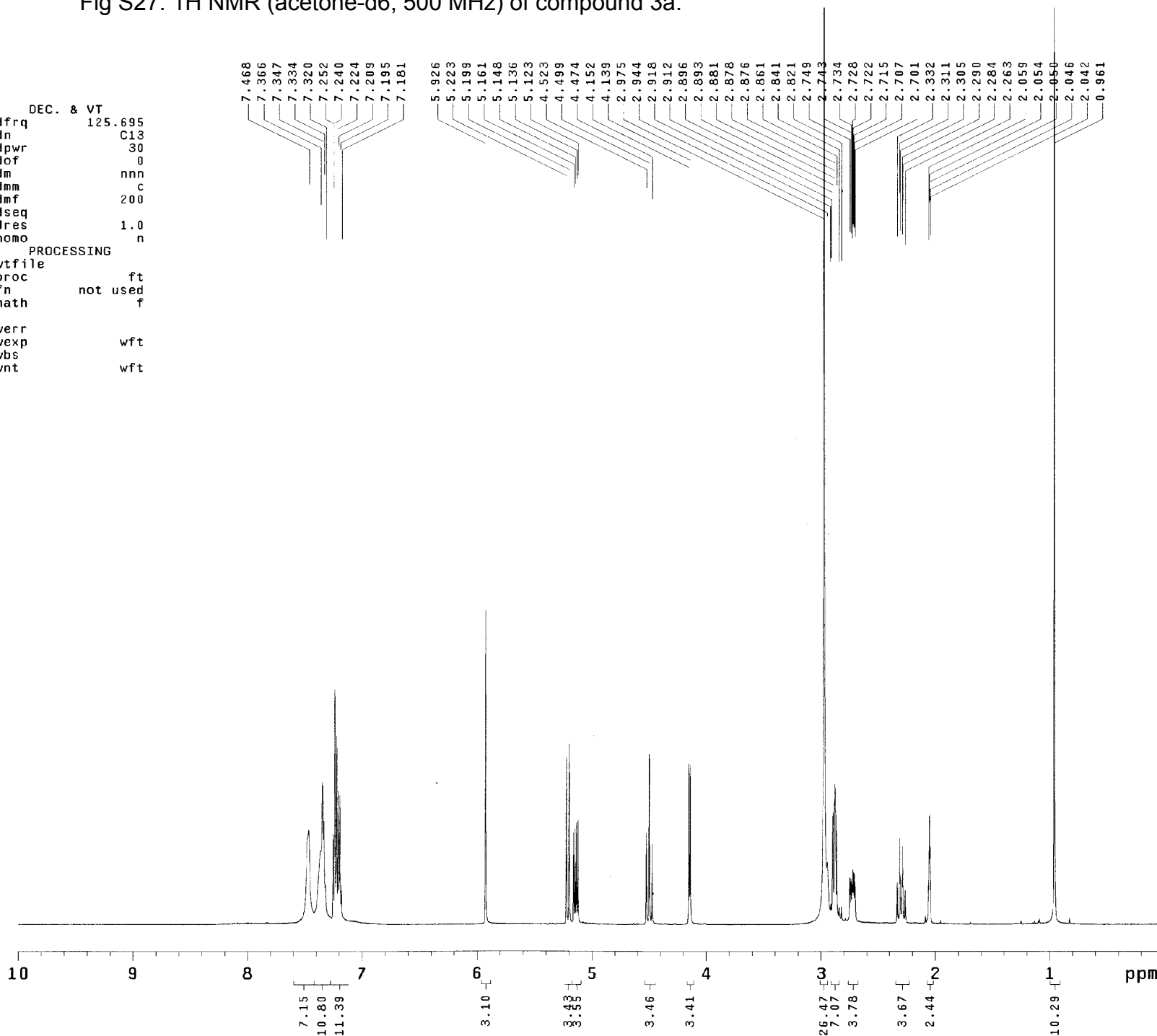
Identification code	ic18028_1	
Empirical formula	C ₂₂ H ₂₀ N ₄ O ₁₀	
Formula weight	500.42	
Temperature	200(2) K	
Wavelength	1.54178 Å	
Crystal system	Orthorhombic	
Space group	P2 ₁ 2 ₁ 2 ₁	
Unit cell dimensions	a = 9.38810(10) Å	α = 90°.
	b = 11.6070(2) Å	β = 90°.
	c = 19.7468(3) Å	γ = 90°.
Volume	2151.76(5) Å ³	
Z	4	
Density (calculated)	1.545 Mg/m ³	
Absorption coefficient	1.064 mm ⁻¹	
F(000)	1040	
Crystal size	0.333 x 0.298 x 0.157 mm ³	
Theta range for data collection	4.418 to 69.931°.	
Index ranges	-11 ≤ h ≤ 10, -13 ≤ k ≤ 14, -24 ≤ l ≤ 23	
Reflections collected	10354	
Independent reflections	4066 [R(int) = 0.0181]	
Completeness to theta = 67.679°	99.9 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7533 and 0.5782	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	4066 / 0 / 328	
Goodness-of-fit on F ²	1.031	
Final R indices [I > 2σ(I)]	R1 = 0.0247, wR2 = 0.0629	
R indices (all data)	R1 = 0.0253, wR2 = 0.0634	
Absolute structure parameter	-0.06(4)	
Extinction coefficient	0.0034(3)	
Largest diff. peak and hole	0.213 and -0.144 e.Å ⁻³	

Fig S27. ¹H NMR (acetone-d₆, 500 MHz) of compound 3a.

CJW-2-15-top

exp147 s2pu1

SAMPLE		DEC. & VT	
date	Dec 18 2014	dfrq	125.695
solvent	Acetone	dn	C13
file	exp	dpwr	30
ACQUISITION		dof	0
sfrq	499.836	dm	nnn
tn	H1	dmm	c
at	3.000	dmf	200
np	48000	dseq	
sw	8000.0	dres	1.0
fb	not used	homo	n
bs	4	PROCESSING	
tpwr	62	wtfile	
pw	4.8	proc	ft
d1	1.000	fn	not used
tof	499.7	math	f
nt	4		
ct	4	werr	
alock	y	wexp	wft
gain	not used	wbs	
	FLAGS	wnt	wft
il	n		
in	n		
dp	y		
hs	nn		
DISPLAY			
sp	-0.0		
wp	4998.3		
vs	156		
sc	0		
wc	210		
hzmm	23.80		
is	33.57		
rfl	2033.0		
rfp	1024.7		
th	2		
ins	100.000		
ai	cdc	ph	



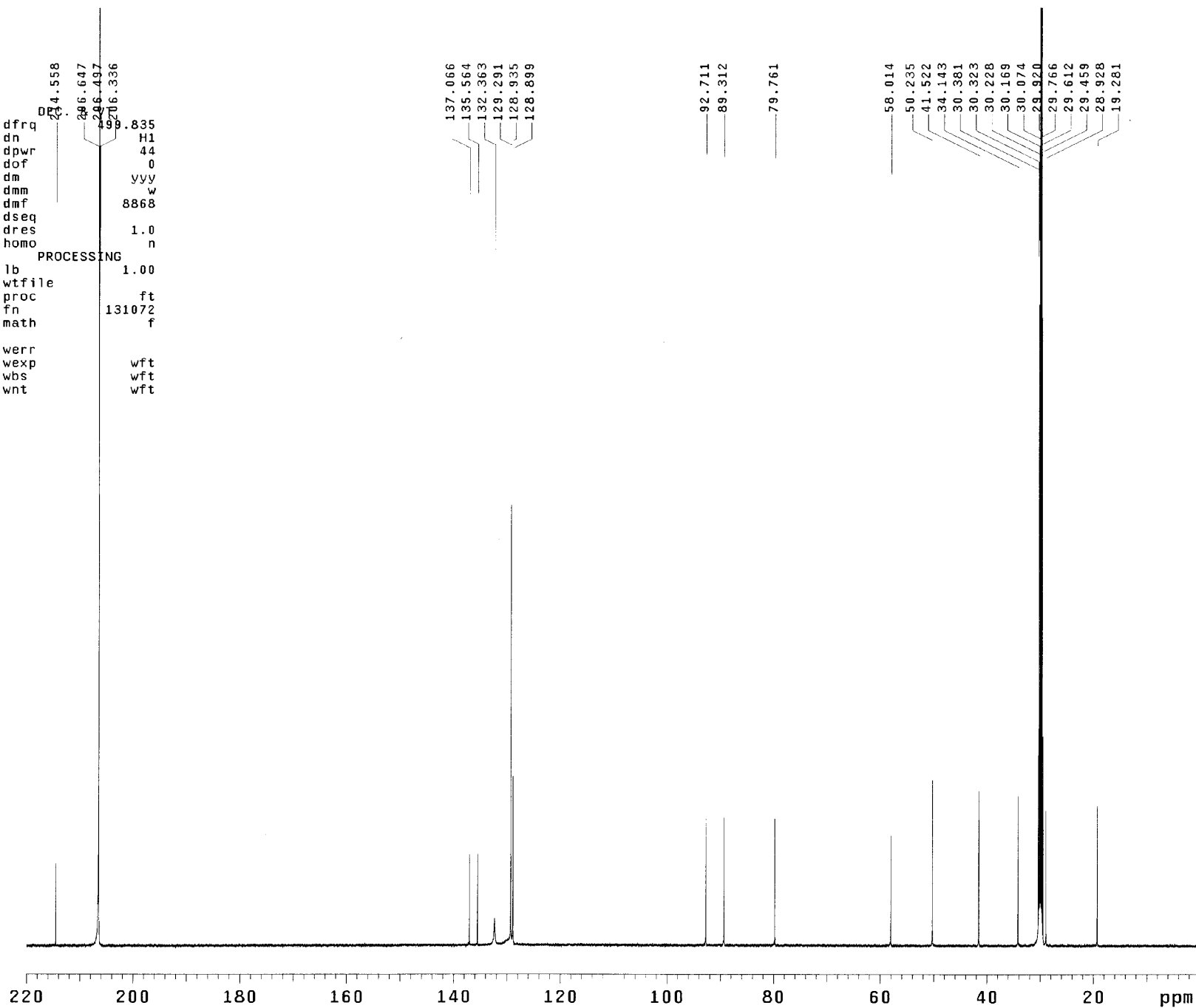
CJW-2-15-top

expi48 s2pul

```

SAMPLE
date Dec 19 2014 dfrq 499.835
solvent Acetone dn H1
file exp dpwr 44
ACQUISITION dof 0
sfrq 125.697 dm yyy
tn C13 dmm w
at 1.000 dmf 8868
np 60332 dseq
sw 30165.9 dres 1.0
fb not used homo n
bs 4 PROCESSING lb 1.00
tpwr 59 wtfile
pw 4.8 proc ft
d1 1.000 fn 131072
tof 1883.7 math f
nt 3000
ct 3000
alock y werr
gain not used wexp wft
FLAGS wbs wft
il n wnt
in n
dp y
hs nn
DISPLAY
sp -0.2
wp 27649.9
vs 295
sc 0
wc 210
hzmm 131.67
is 33.57
rfl 4931.1
rfp 3760.4
th 4
ins 100.000
nm cdc ph

```

Fig S28. ^{13}C NMR (acetone- d_6 , 125 MHz) of compound 3a.

CJW-2-15-top

exp149 DEPT

SAMPLE		DEPT	ACQUISITION ARRAYS	
date	Dec 19 2014	j1xh	140.0	array
solvent	Acetone	mult	arrayed	mult
sample	undefined	SPECIAL	arraydim	4
ACQUISITION		temp	not used	i
sw	30165.9	gain	28	mult
at	1.000	spin	0	1
np	60332	PROCESSING		
bs	4	lb	1.00	4
ss	-4	fn	131072	1.5
d1	1.000	SPECTRUM		
nt	1500	wp	27649.9	
ct	1500	sp	-0.2	
TRANSMITTER		rp	-169.4	
tn	C13	lp	242.8	
tof	1883.7	ai	cdc	ph
tpwr	59	REFERENCE		
pw	14.700	rfl	1171.2	
DECOUPLER		rfl	0	
dn	H1	PLOT		
dof	0	wc	210	
dpwr	44	sc	0	
dm	nny	vs	150	
dmm	ccw	hzmm	131.67	
dmf	8868	th	7	
pp1v1	59			
pp	21.200			

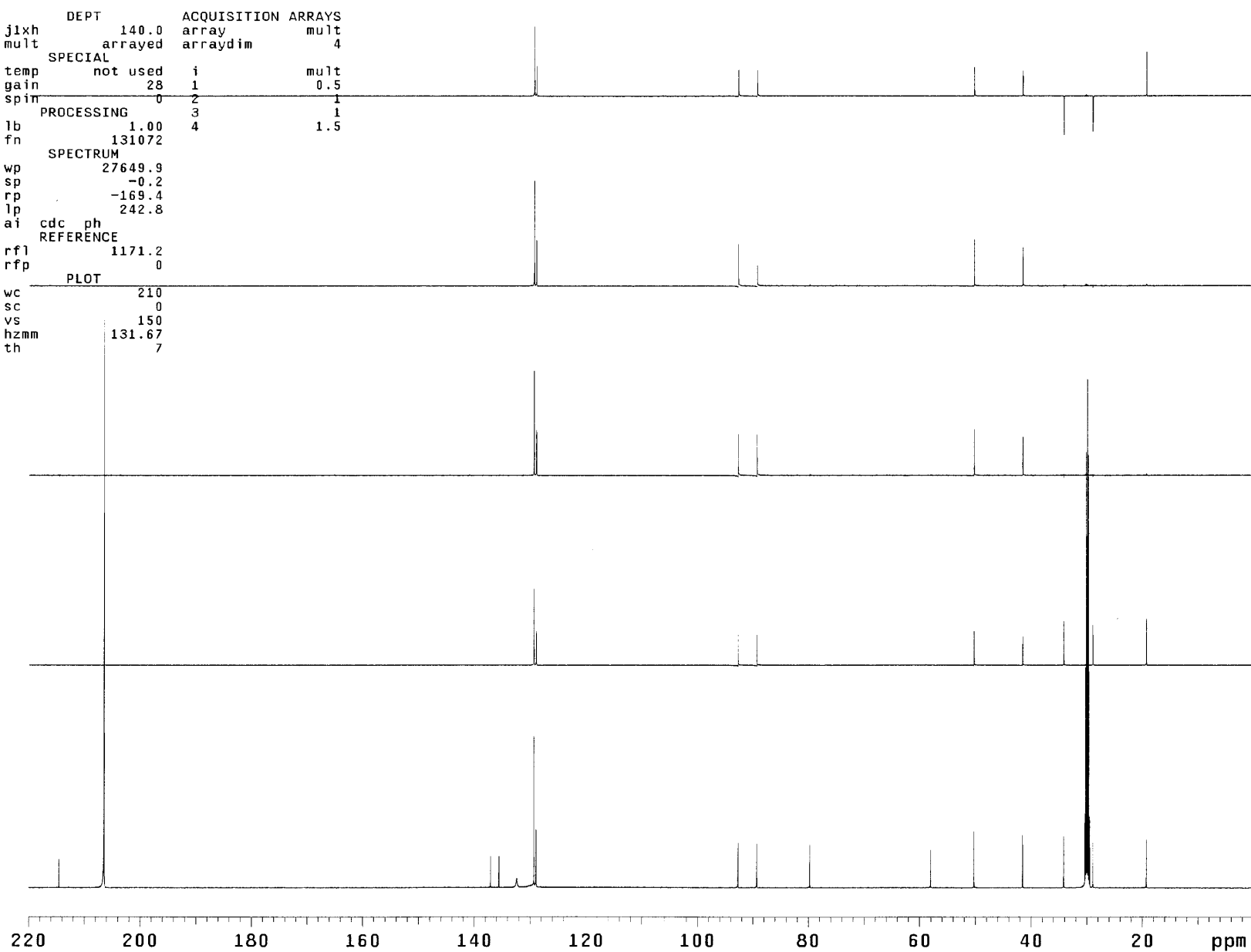


Fig S29. DEPT of compound 3a.

Fig S30. HSQC of compound 3a.

CJW-2-15-top
exp152 gHSQC

SAMPLE	FLAGS	ACQUISITION	ARRAYS
date Dec 19 2014	hs n	array	phase
solvent Acetone	sspul y	arraydim	256
sample undefined	PFGflg y		
ACQUISITION	hsglv1 1004	i	phase
sw 4743.8	SPECIAL	1	1
at 0.216	temp not used	2	2
np 2048	gain 48		
fb not used	spin 0		
ss 32	GRADIENTS		
d1 1.000	gzlv11 1004		
nt 8	gt1 0.002000		
2D ACQUISITION	gzlv13 505		
sw1 21367.5	gt3 0.001000		
ni 128	gstab 0.000500		
phase arrayed	F2 PROCESSING		
TRANSMITTER	gf 0.100		
tn H1	gfs not used		
sfrq 499.835	fn 2048		
tof -375.0	F1 PROCESSING		
tpwr 62	gf1 0.006		
pw 13.800	gfs1 not used		
DECOUPLER	proc1 lp		
dn C13	fn1 2048		
dof -2515.2	DISPLAY		
dm nny	sp 386.5		
dmm ccp	wp 3437.4		
dmf 32258	sp1 2029.2		
dpwr 43	wp1 14648.4		
pwxlvl 61	rfl 2501.6		
pw 11.000	rfl 2248.7		
HSQC	rfl1 6382.0		
j1xh 140.0	rfl1 5218.6		
nullflg y	PLOT		
mult 2	wc 150.0		
	sc 6.2		
	wc2 116.2		
	sc2 0		
	vs 736		
	th 4		
	ai cdc ph		

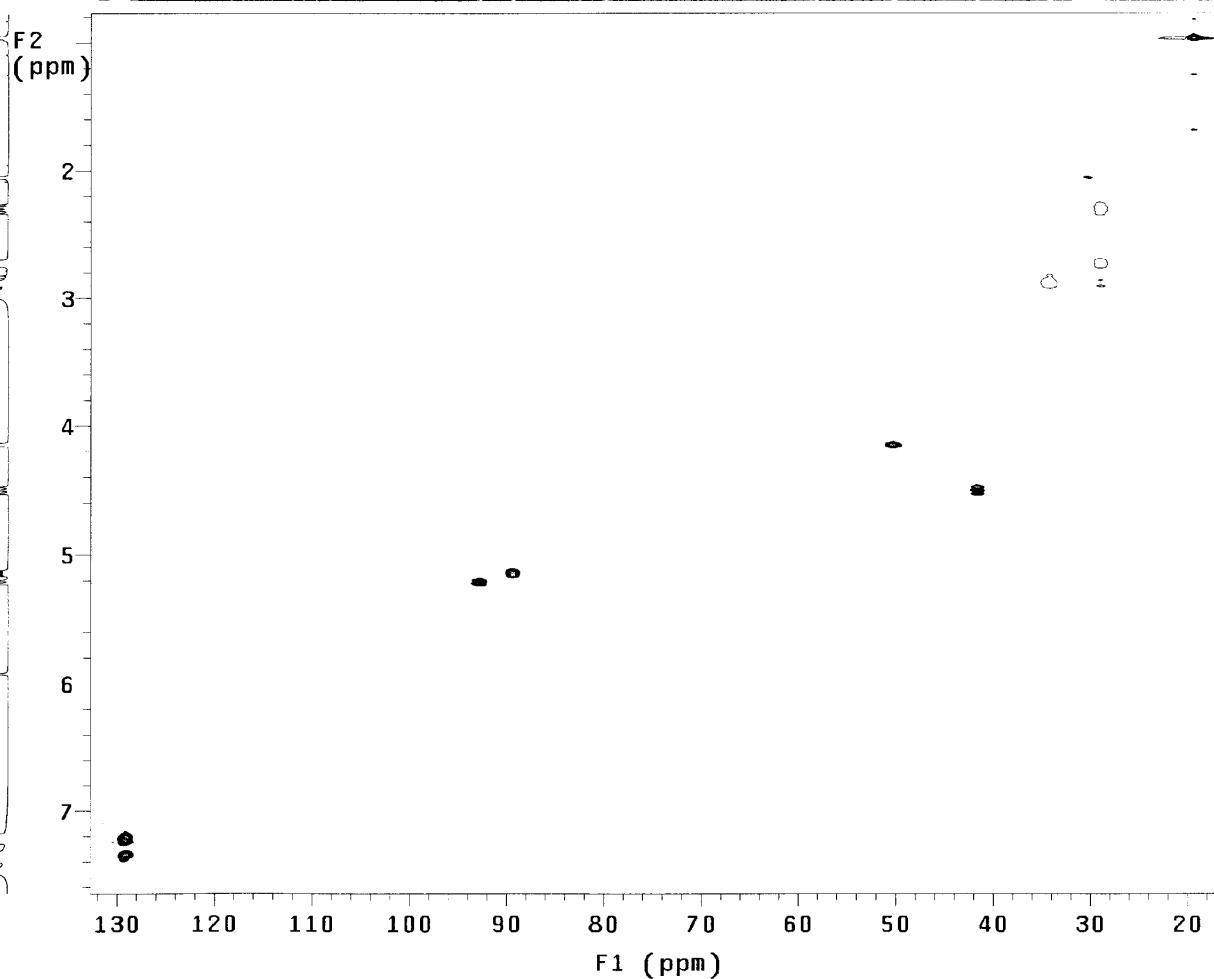


Fig S31. COSY of compound 3a.

CJW-2-15-top
exp150 gCOSY

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date	Dec 19 2014	hs	nn
solvent	Acetone	sspul	n
sample	undefined	hsglvl	1004
ACQUISITION		SPECIAL	
sw	4743.8	temp	not used
at	0.216	gain	28
np	2048	spin	0
fb	not used	F2 PROCESSING	
ss	16	sb	-0.108
d1	1.000	sbs	not used
nt	8	fn	2048
2D ACQUISITION		F1 PROCESSING	
sw1	4743.8	sb1	-0.027
n1	128	sbs1	not used
TRANSMITTER		PROC1	
tn	H1	fn1	2048
sfrq	499.835	DISPLAY	
tof	-375.0	sp	344.7
tpwr	62	wp	3775.6
pw	13.800	sp1	348.1
GRADIENTS		wp1	3775.6
gzlv11	1004	rfl	3214.9
gt1	0.001000	rfp	2962.0
gstab	0.000500	rfl1	3216.2
DECOUPLER		rfp1	2962.0
dn	C13	PLOT	
dm	nnn	wc	155.0
		sc	10.0
		wc2	155.0
		sc2	0
		vs	736
		th	4
	ai	cdc	av

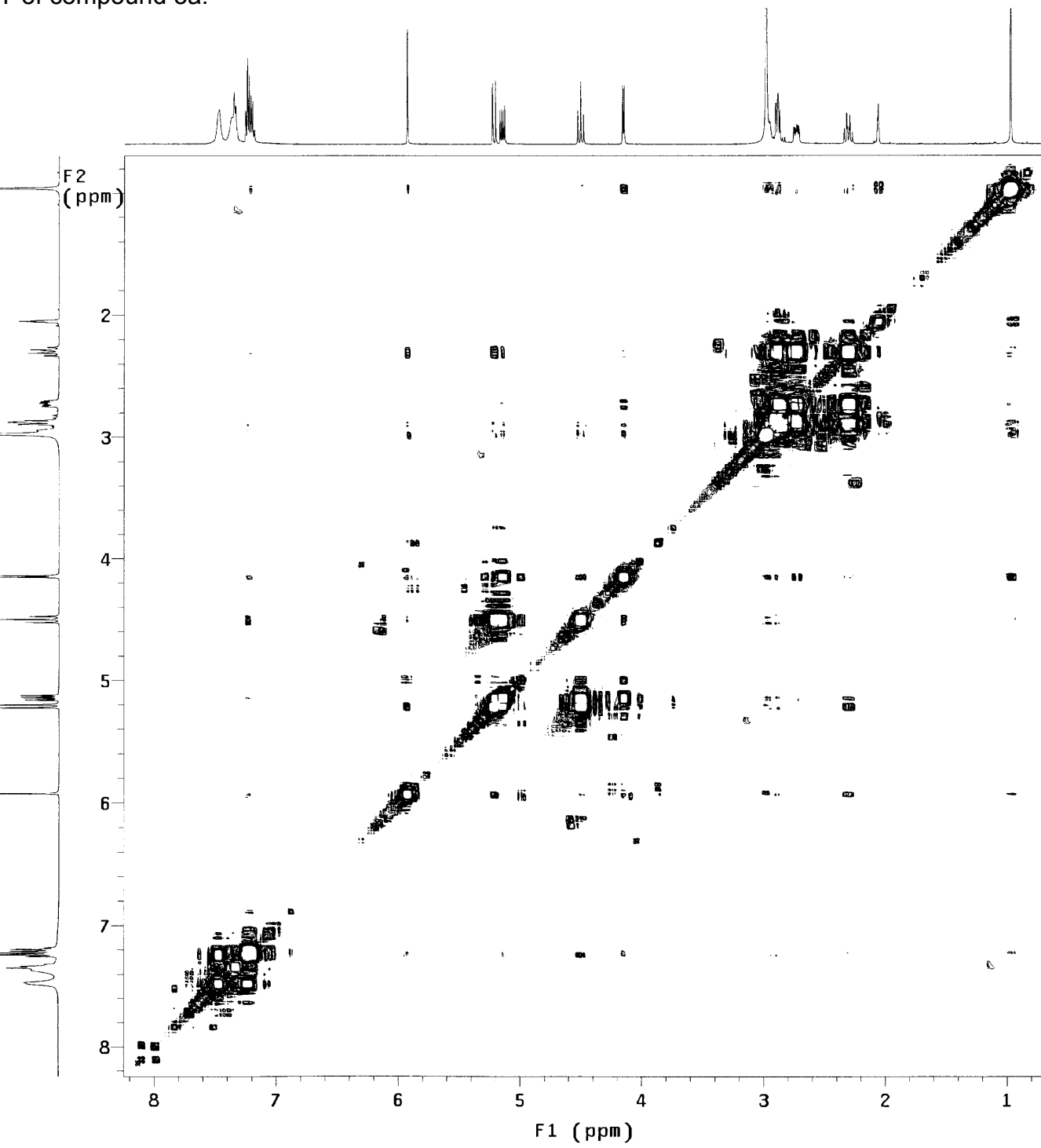


Fig S32. NOESY of compound 3a.

CJW-2-15-top
exp151 NOESY

SAMPLE		FLAGS	
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solvent	Acetone	sspul	y
sample	undefined	PFGflg	y
ACQUISITION		hsglv1	1004
sw	4743.8	SPECIAL	
at	0.216	temp	not used
np	2048	gain	28
fb	not used	spin	0
ss	32	F2 PROCESSING	
d1	1.000	gf	0.100
nt	16	gfs	not used
2D ACQUISITION		fn	2048
sw1	4743.8	F1 PROCESSING	
ni	200	gf1	0.039
TRANSMITTER		gfs1	not used
tn	H1	proc1	lp
sfrq	499.835	fn1	2048
tof	-375.0	DISPLAY	
tpwr	62	sp	312.0
pw	13.800	wp	3525.4
NOESY		sp1	312.4
mix	0.600	wp1	3525.4
PRESATURATION		rfl	3215.2
satmode	nnnn	rfp	2962.0
satpwr	0	rfl1	3214.8
satdly	0	rfp1	2962.0
satfrq	0	PLOT	
DECOUPLER		wc	155.0
dn	C13	sc	10.0
dm	nnn	wc2	155.0
		sc2	0
		vs	736
		th	1
		ai	ph

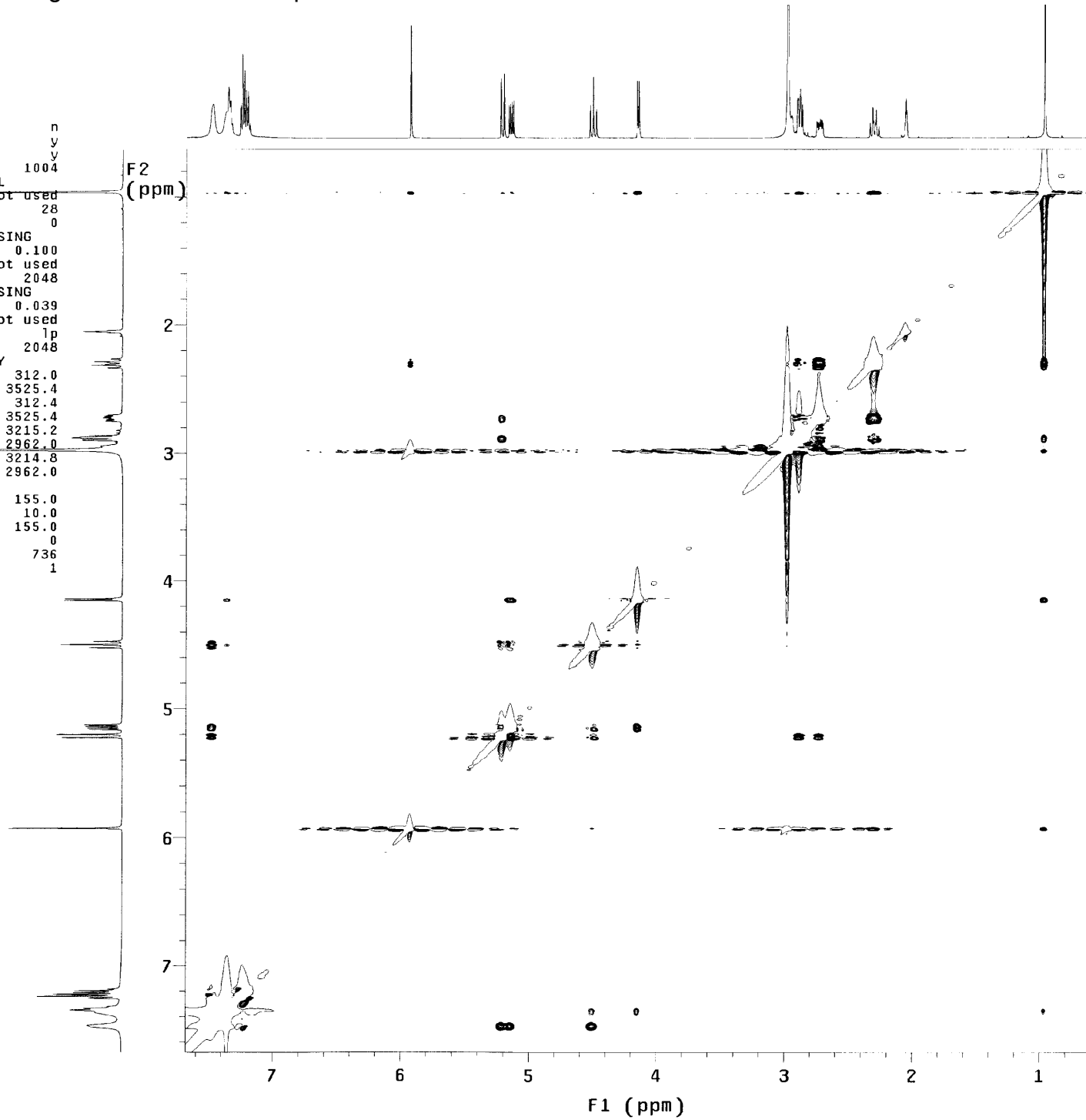


Fig S33. ¹H NMR (acetone-d₆, 500 MHz) of compound 5a.

CHP-8a-12

Sample Name **CHP-8a-12**
Date collected **2016-05-17**

Pulse sequence **PROTON**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

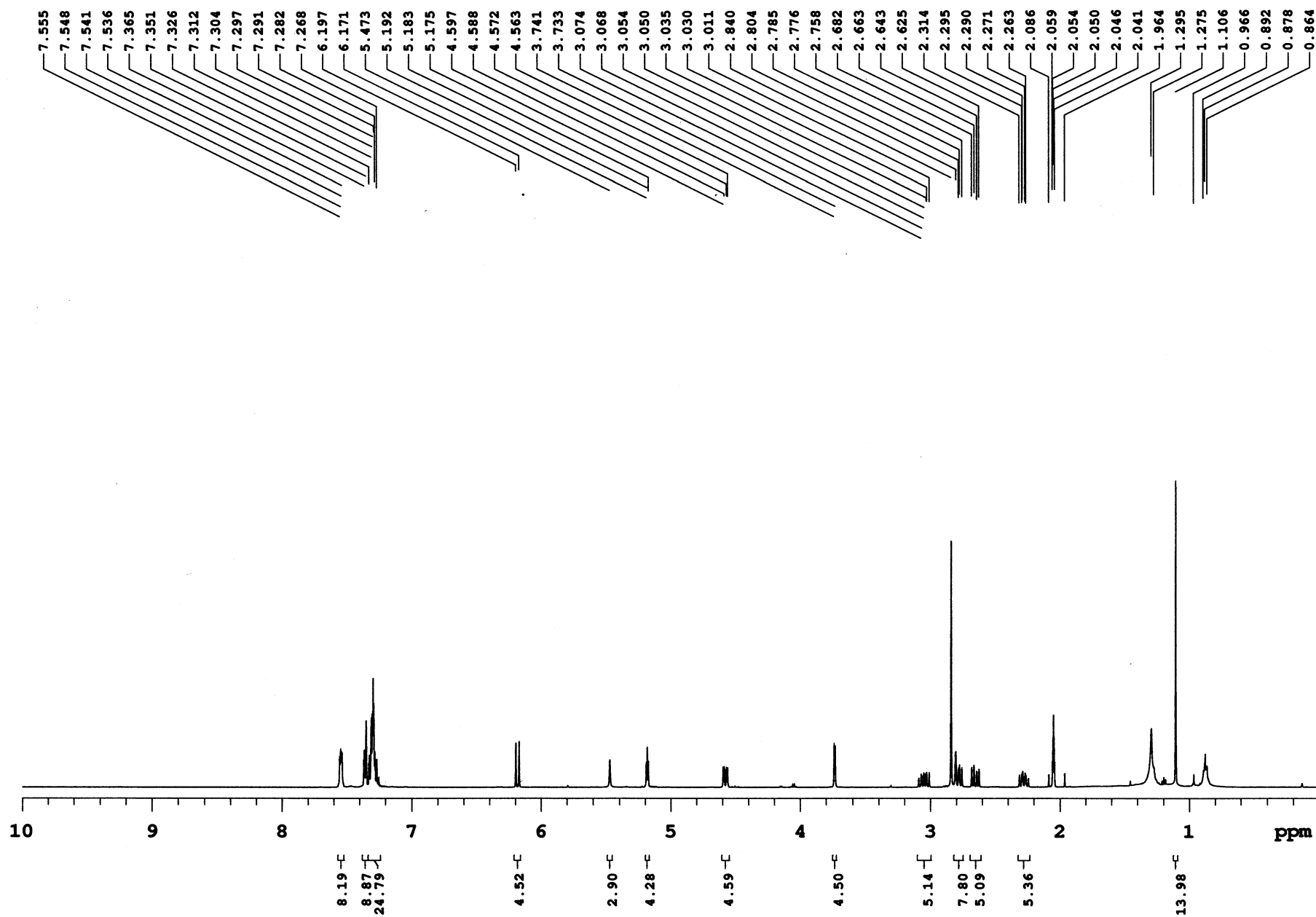


Fig S34. ¹³C NMR (acetone-d₆, 125 MHz) of compound 5a.

CHP-8a-f2

Sample Name **CHP-8a-f2**
Date collected **2016-05-17**

Pulse sequence **CARBON**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

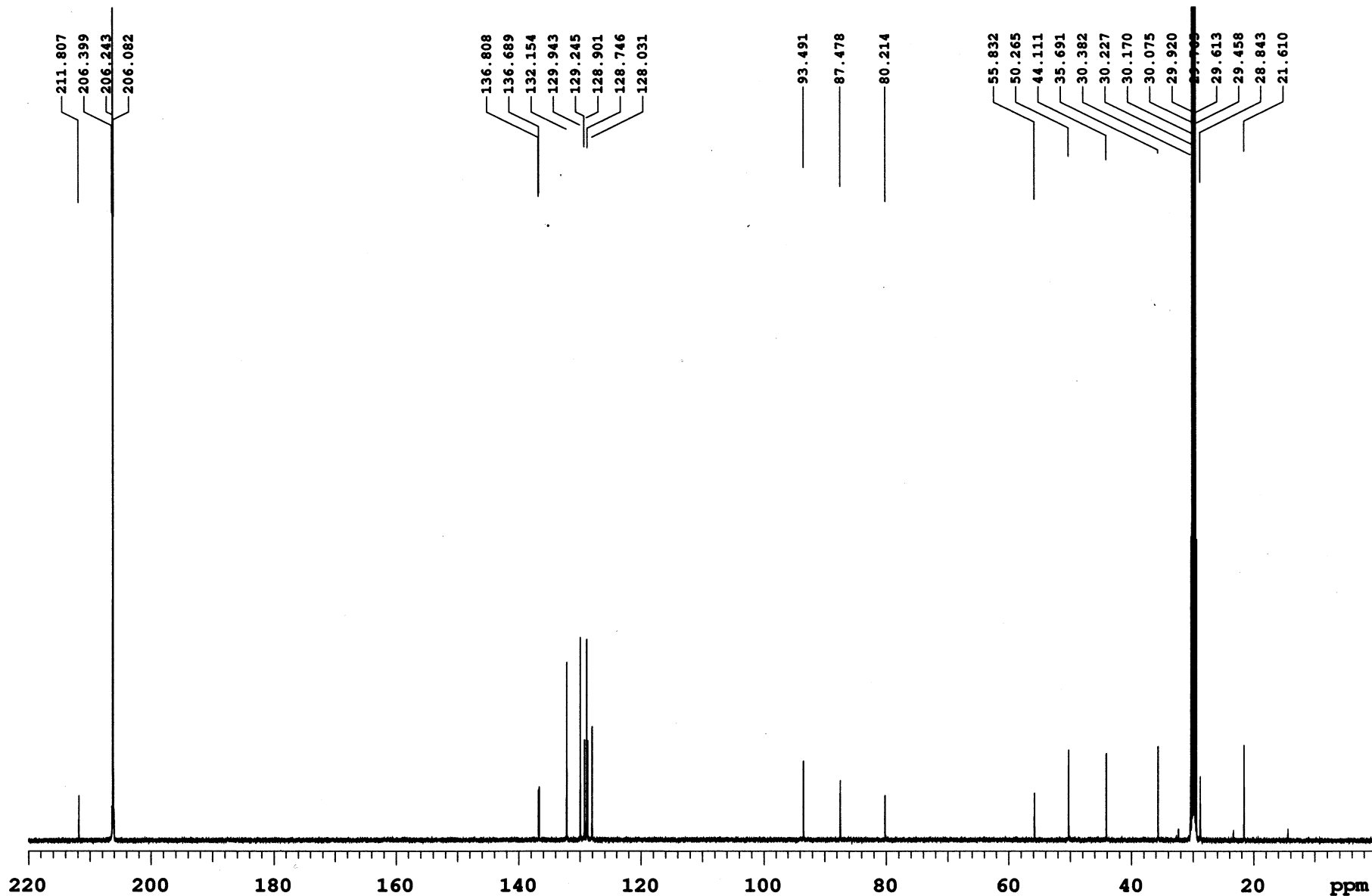


Fig S35. DEPT of compound 5a.

CHP-8a-f2

Sample Name **CHP-8a-f2**
Date collected **2016-05-18**

Pulse sequence **DEPT**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

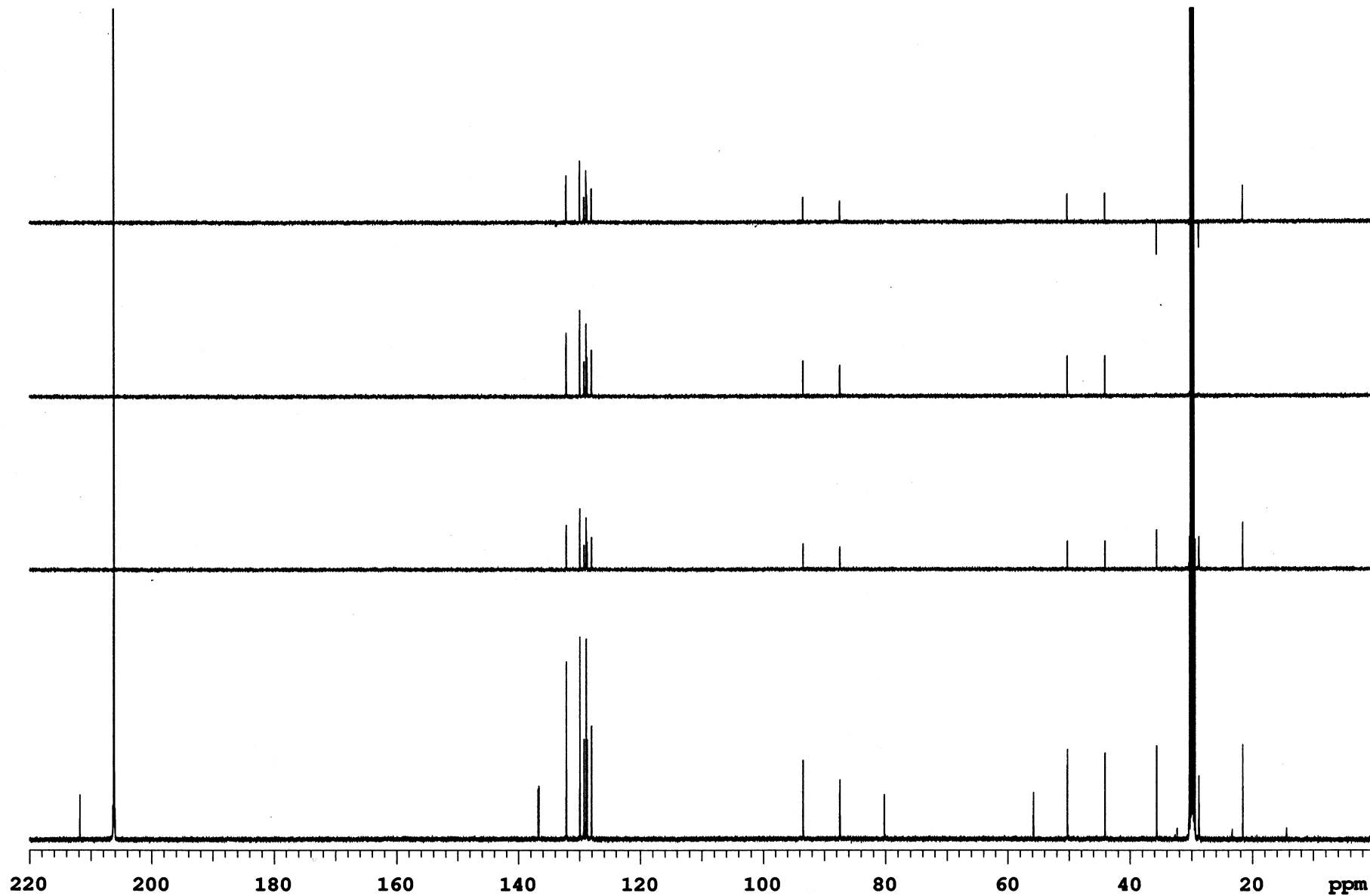


Fig S36. HSQC of compound 5a.

CHP-8a-f2

Sample Name **CHP-8a-f2**
Date collected **2016-05-18**

Pulse sequence **gHSQC**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

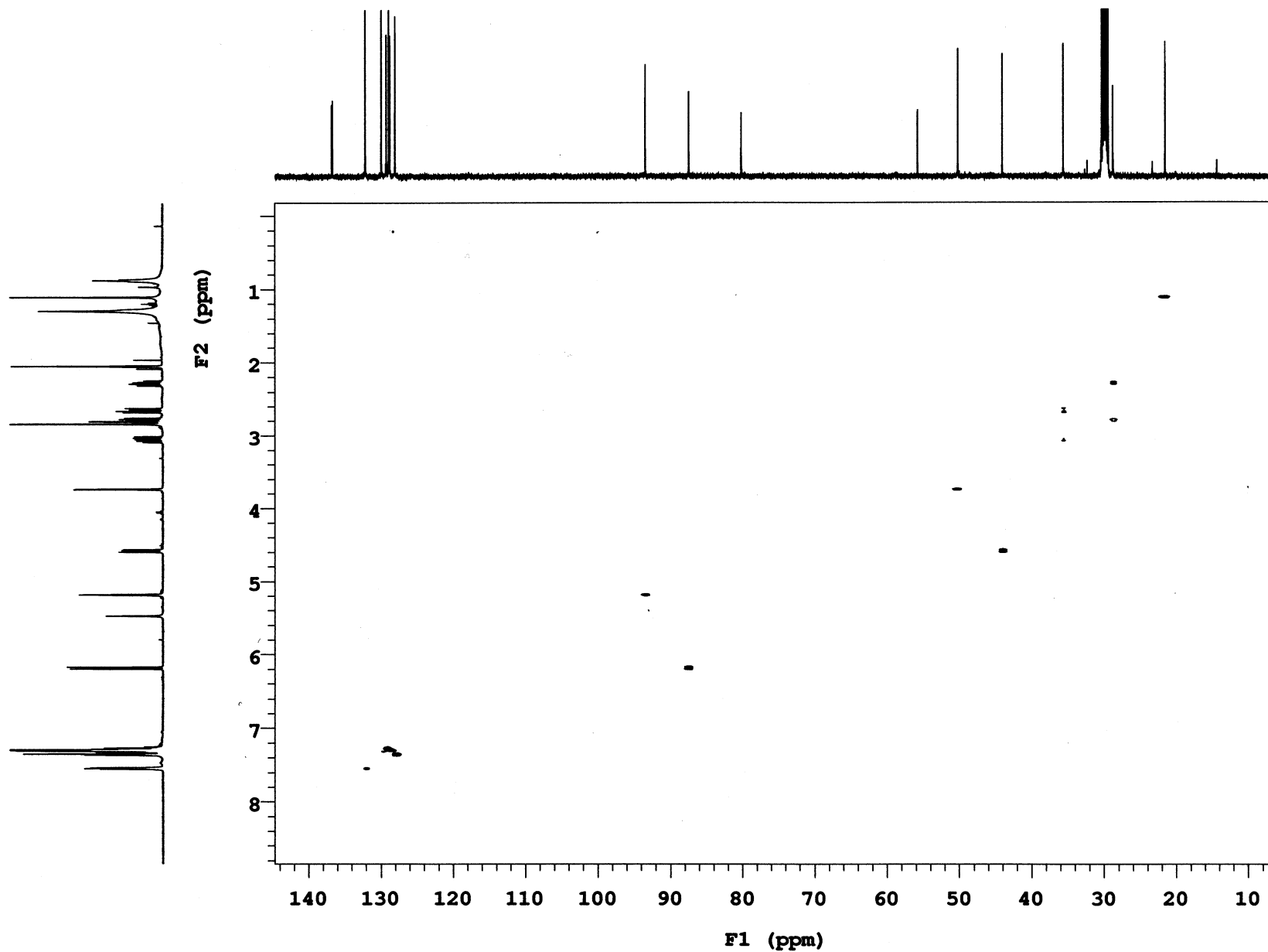


Fig S37. COSY of compound 5a.

CHP-8a-f2

Sample Name **CHP-8a-f2**
Date collected **2016-05-18**

Pulse sequence **gCOSY**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

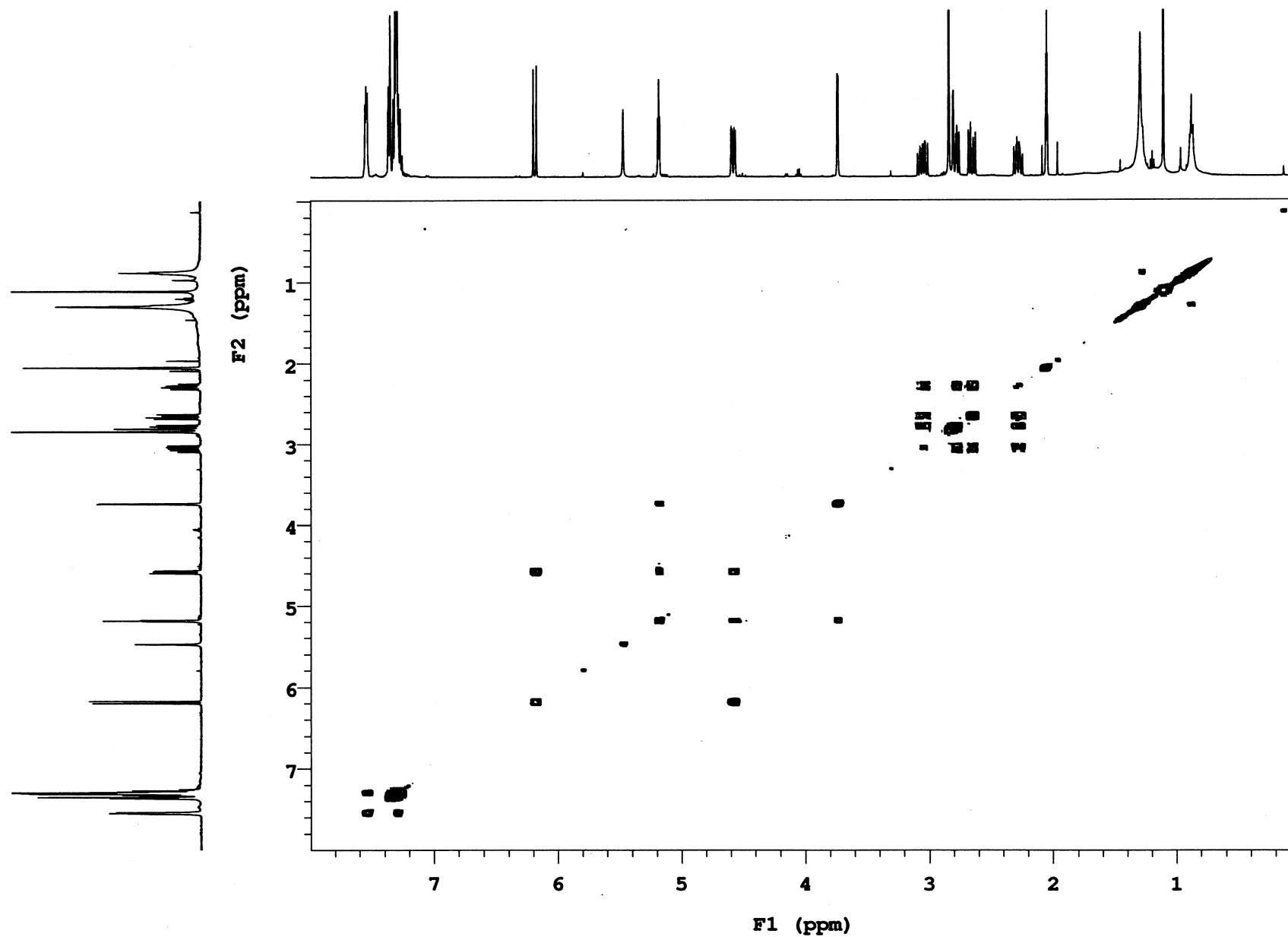


Fig S38. NOESY of compound 5a.

CHP-8a-f2

Sample Name **CHP-8a-f2**
Date collected **2016-05-18**

Pulse sequence **NOESY**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

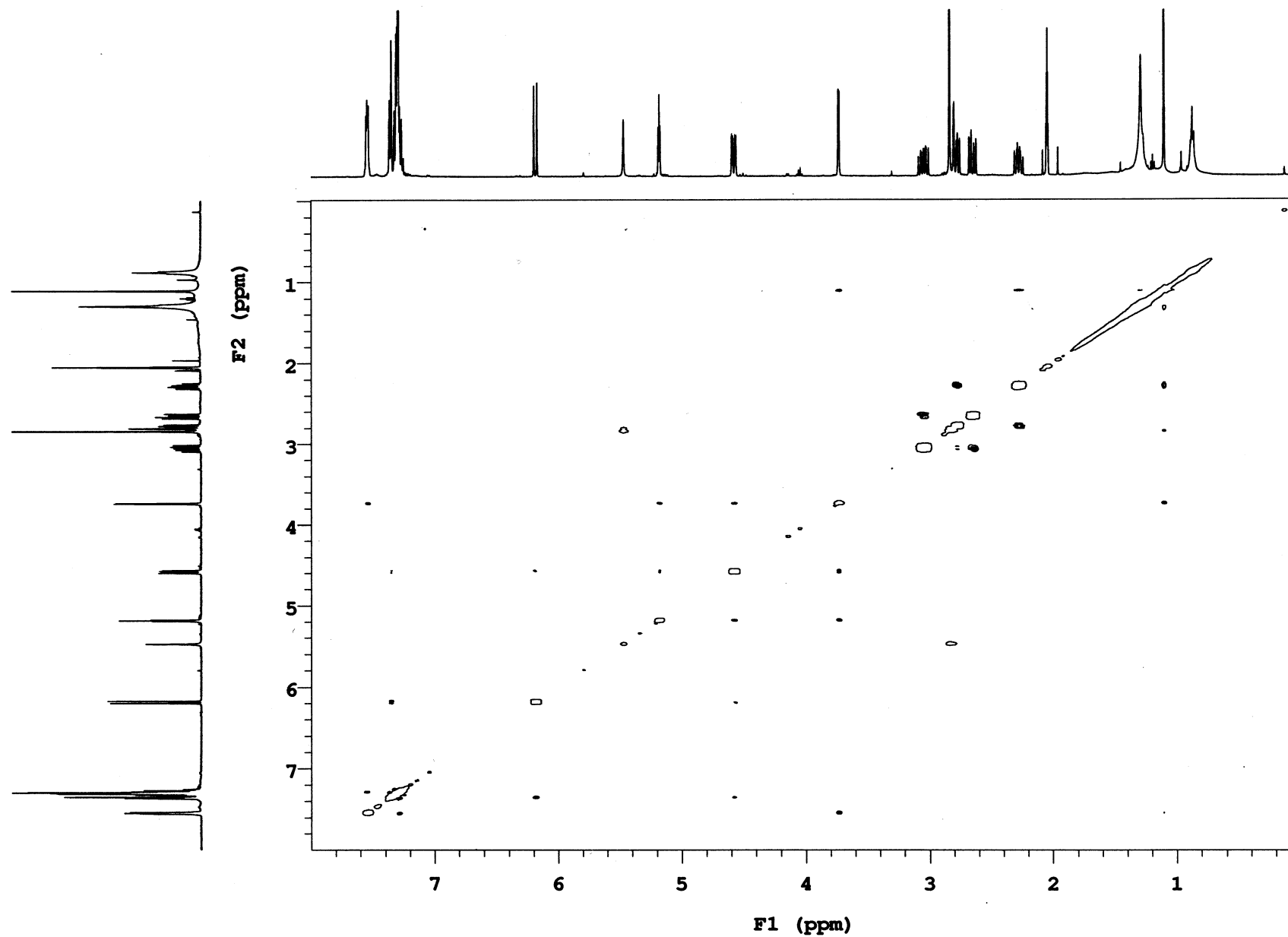


Fig S39. NOESY of compound 5a, expanded.

CHP-8a-f2

Sample Name **CHP-8a-f2**
Date collected **2016-05-18**

Pulse sequence **NOESY**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

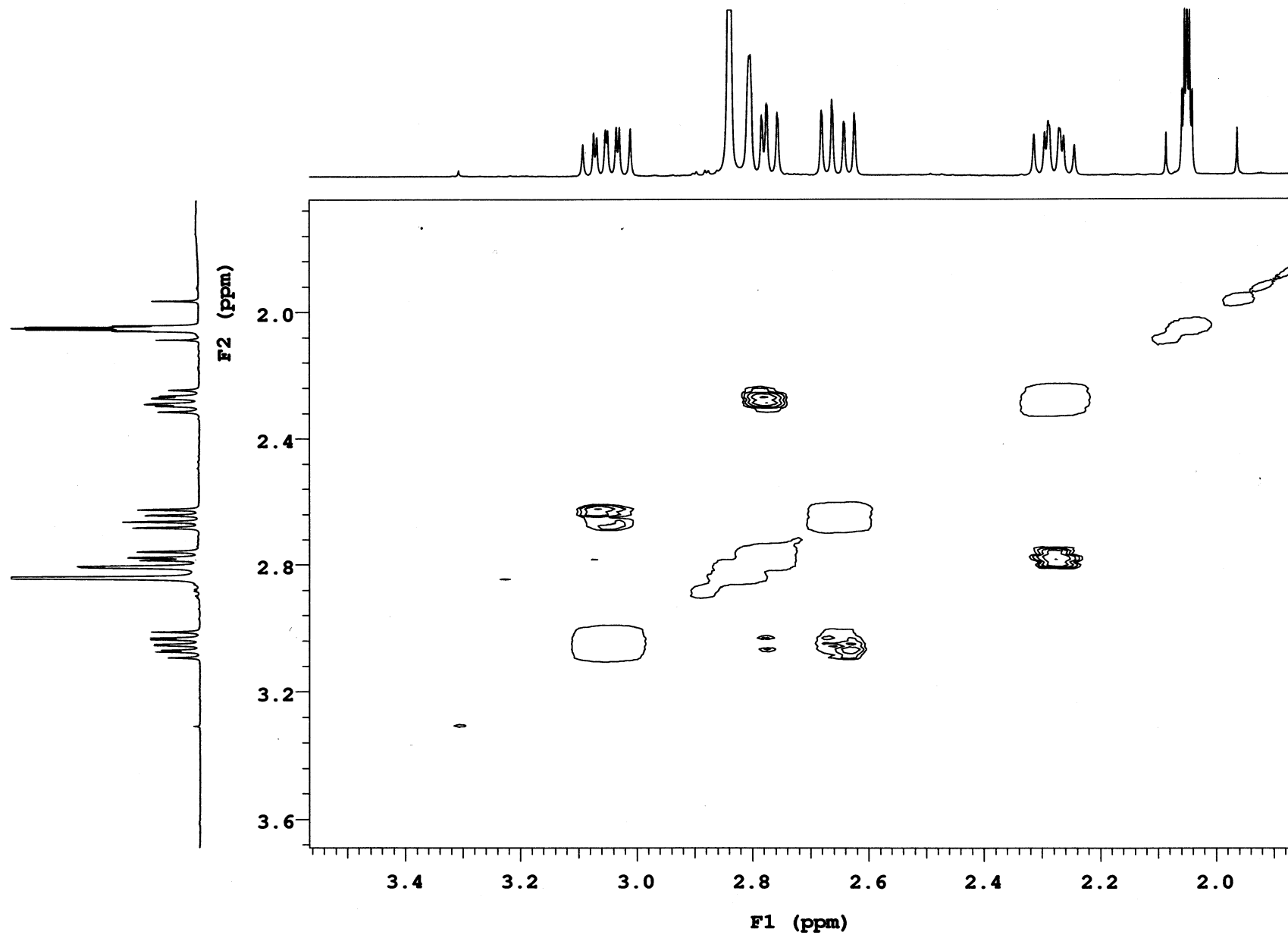


Fig S40. ¹H NMR (CD₃CN, 500 MHz) of compound 3b.

CHP-8b

Sample Name	CHP-8b	Pulse sequence	PROTON	Temperature	25	Study owner	vnmr2
Date collected	2016-03-25	Solvent	cd3cn	Spectrometer	Agilent-NMR-inova500	Operator	vnmr2

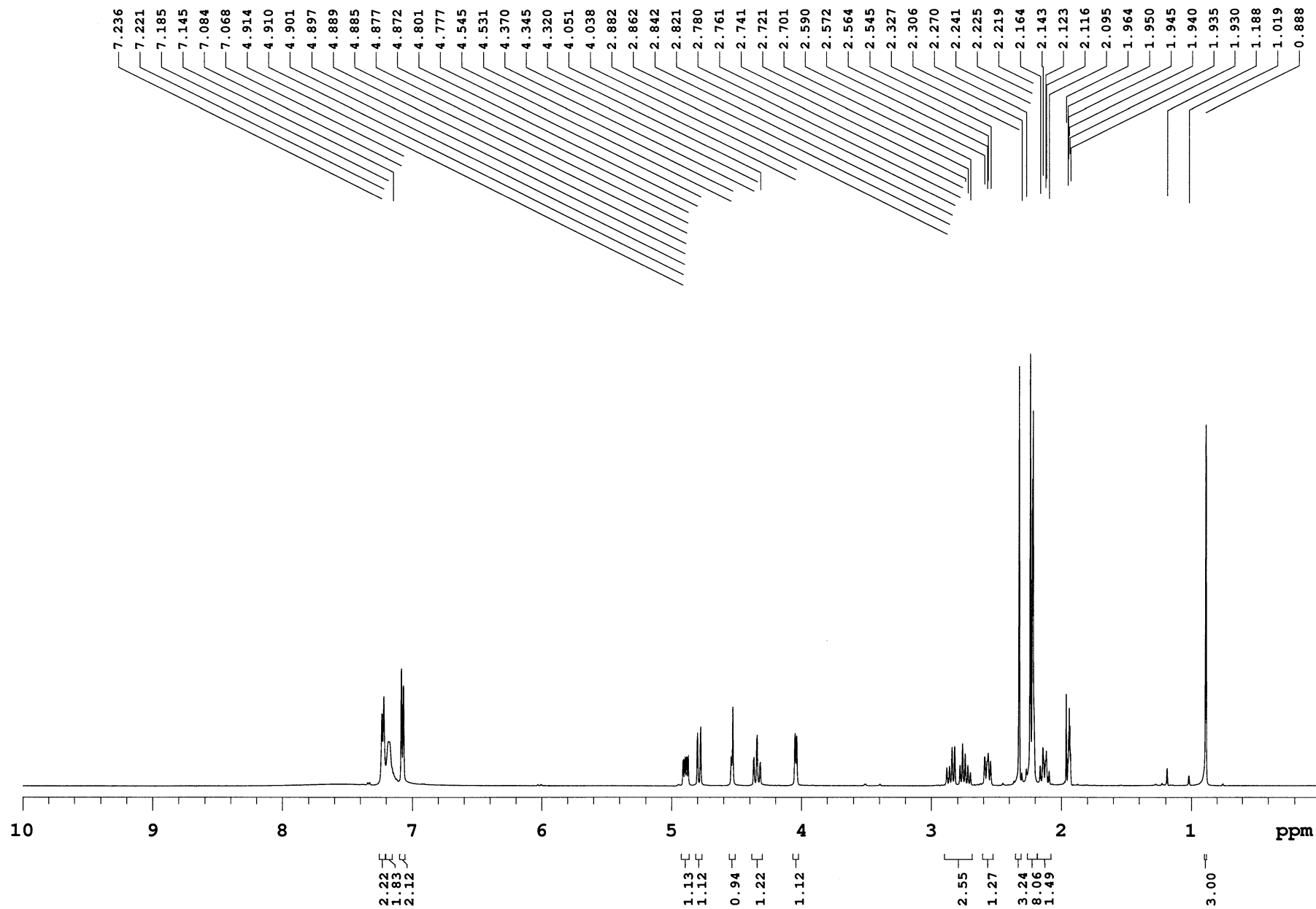


Fig S41. ¹³C NMR (CD₃CN, 125 MHz) of compound 3b.

CHP-8b

Sample Name **CHP-8b**
Date collected **2016-03-25**

Pulse sequence **CARBON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

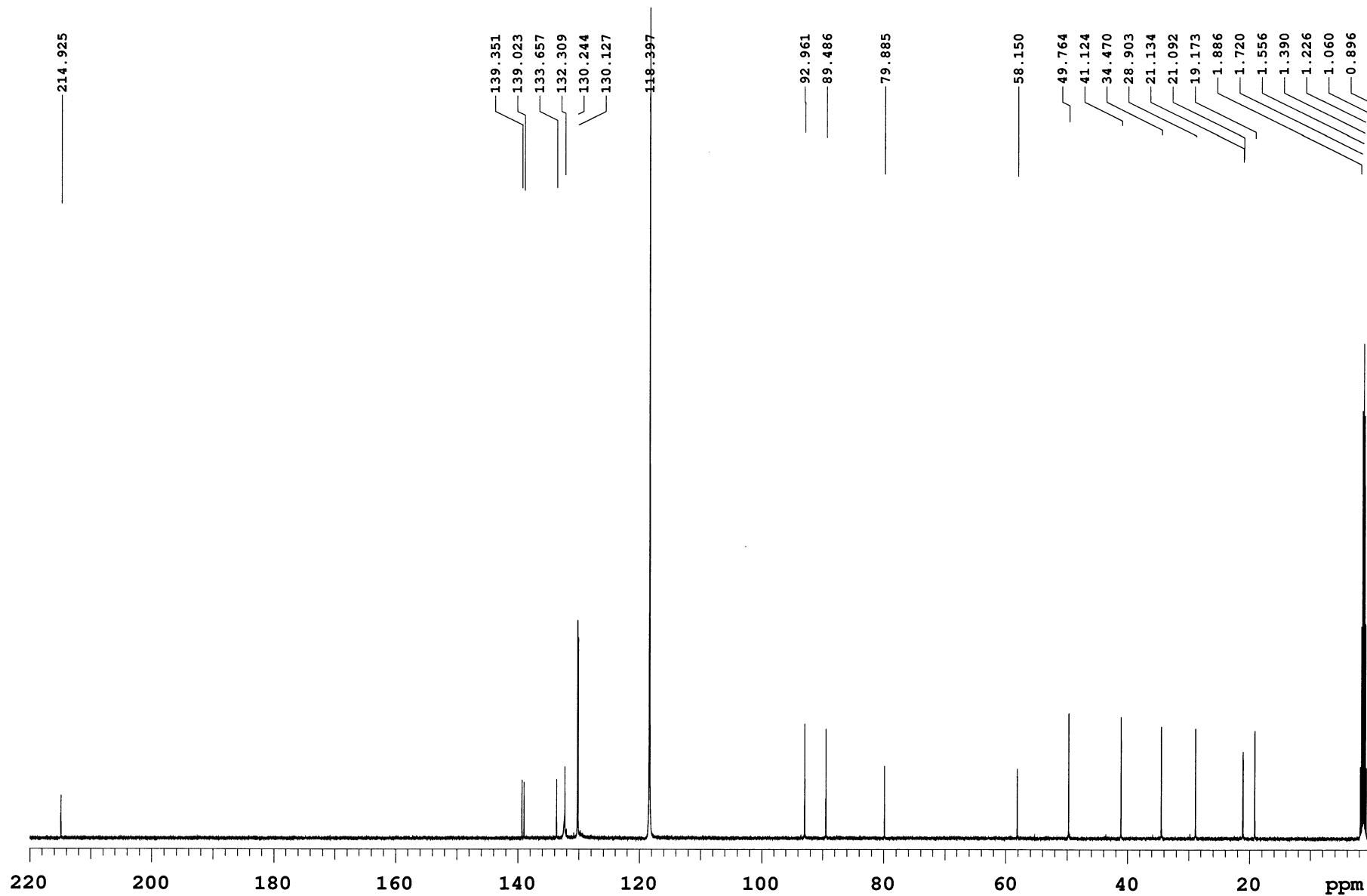


Fig S42. DEPT of compound 3b.

CHP-8b

Sample Name **CHP-8b**
Date collected **2016-03-25**

Pulse sequence **DEPT**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

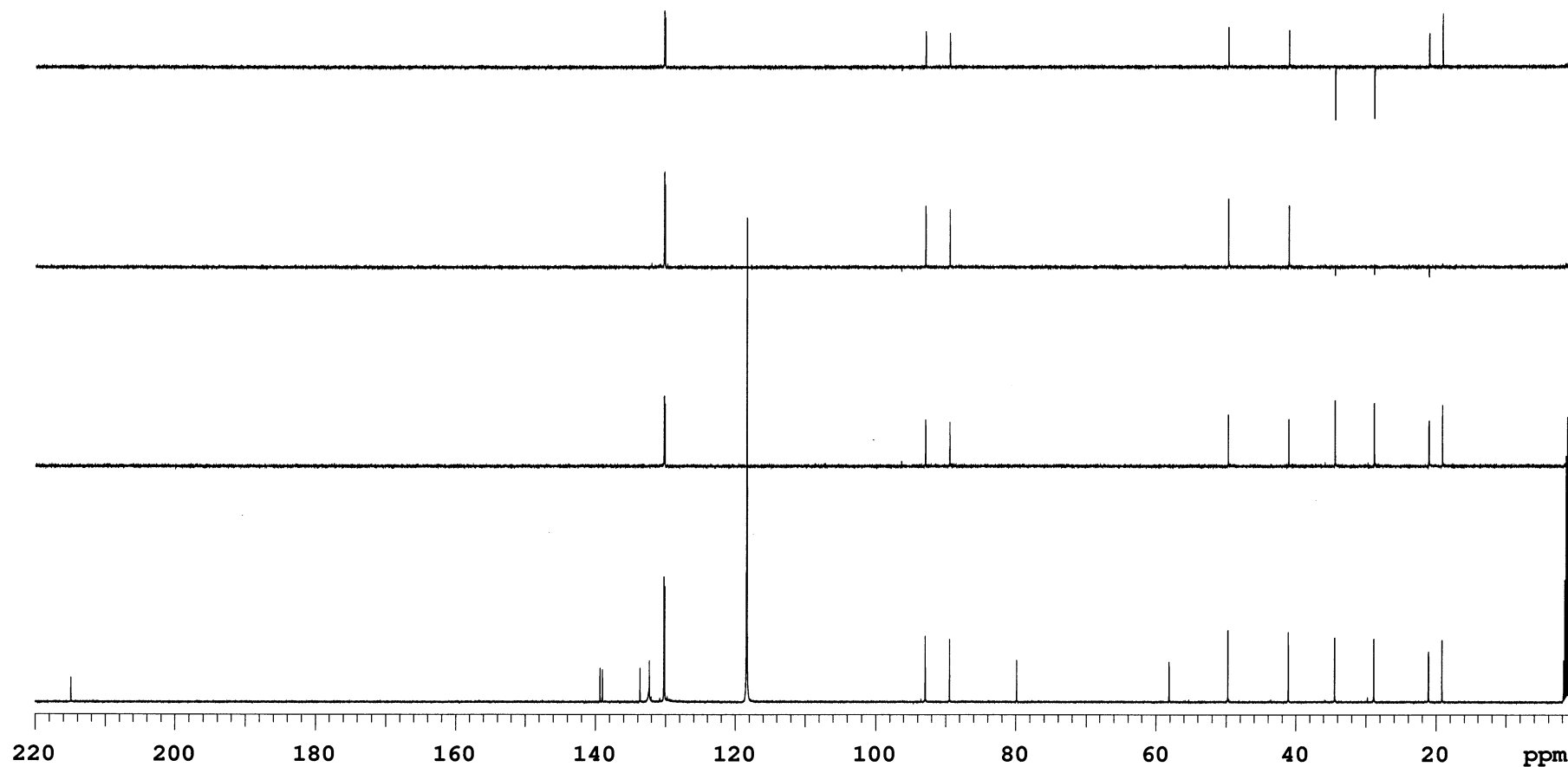


Fig S43. HSQC of compound 3b.

CHP-8b

Sample Name **CHP-8b**
Date collected **2016-03-25**

Pulse sequence **gHSQC**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

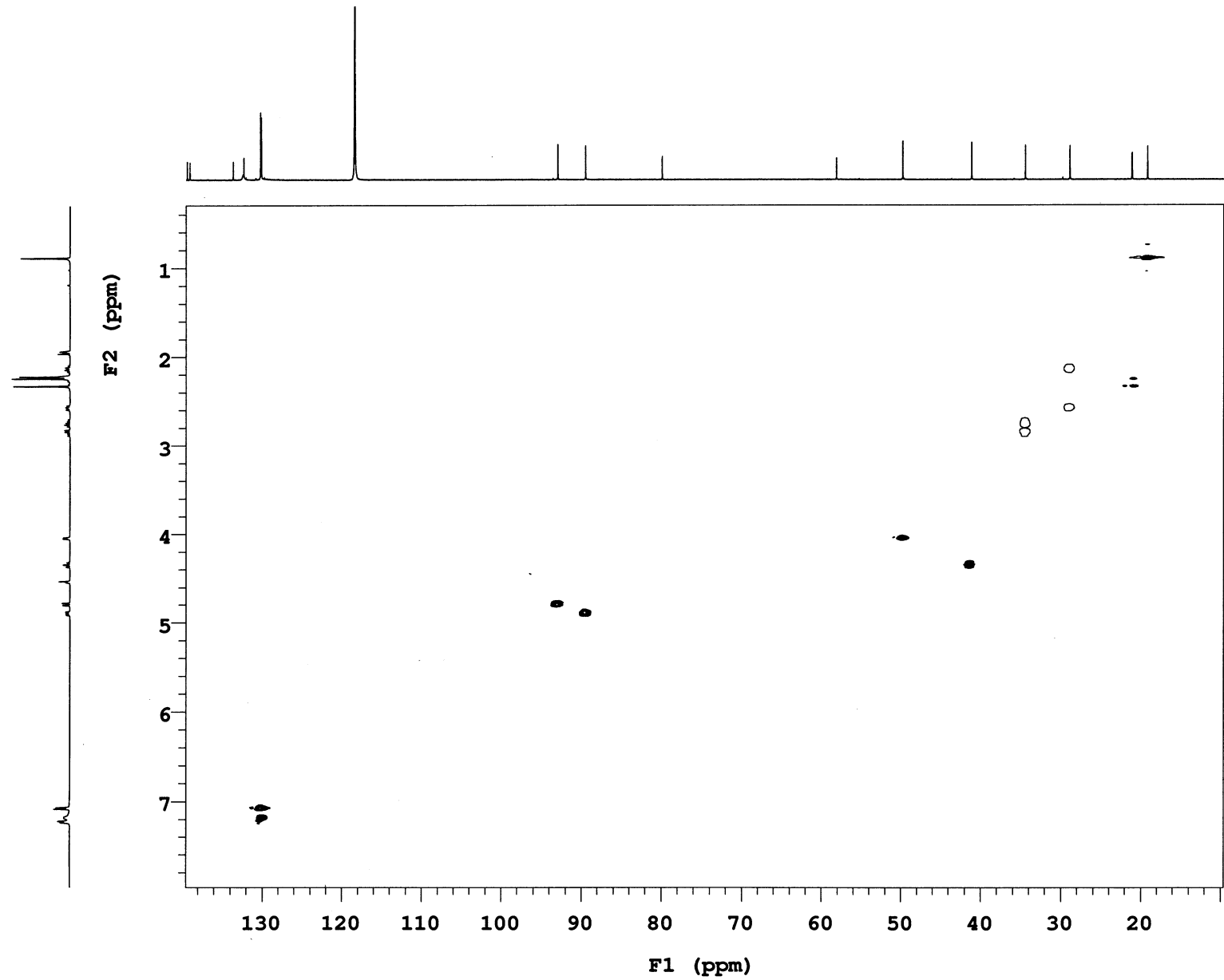


Fig S44. COSY of compound 3b.

CHP-8b

Sample Name **CHP-8b**
Date collected **2016-03-25**

Pulse sequence **gCOSY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

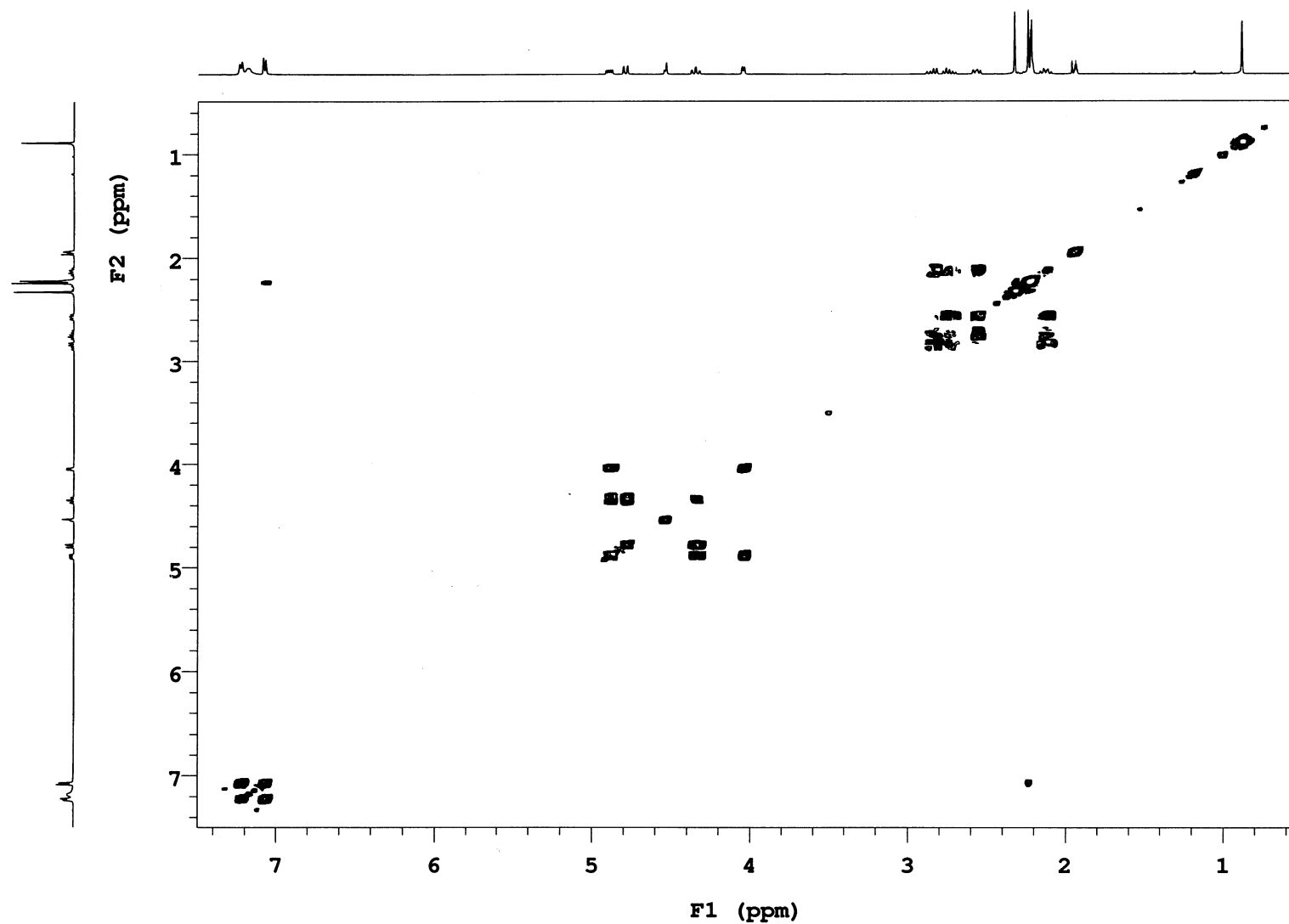


Fig S45. NOESY of compound 3b.

CHP-8b

Sample Name **CHP-8b**
Date collected **2016-03-25**

Pulse sequence **NOESY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

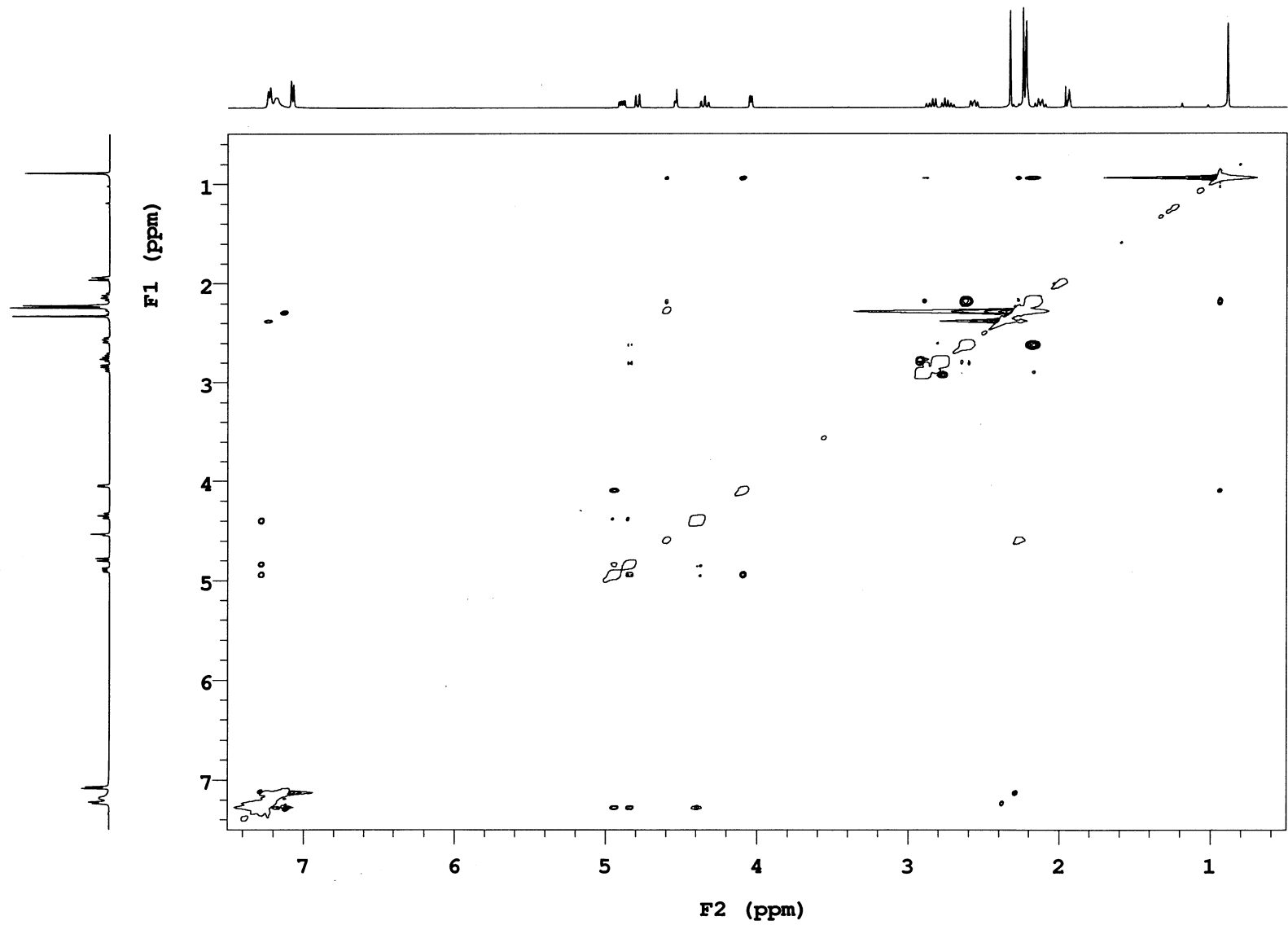


Fig S46. ¹H NMR (CD₃CN, 500 MHz) of compound 5b.

CHP-8b-12

Sample Name **CHP-8b-12**
Date collected **2016-05-20**

Pulse sequence **PROTON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

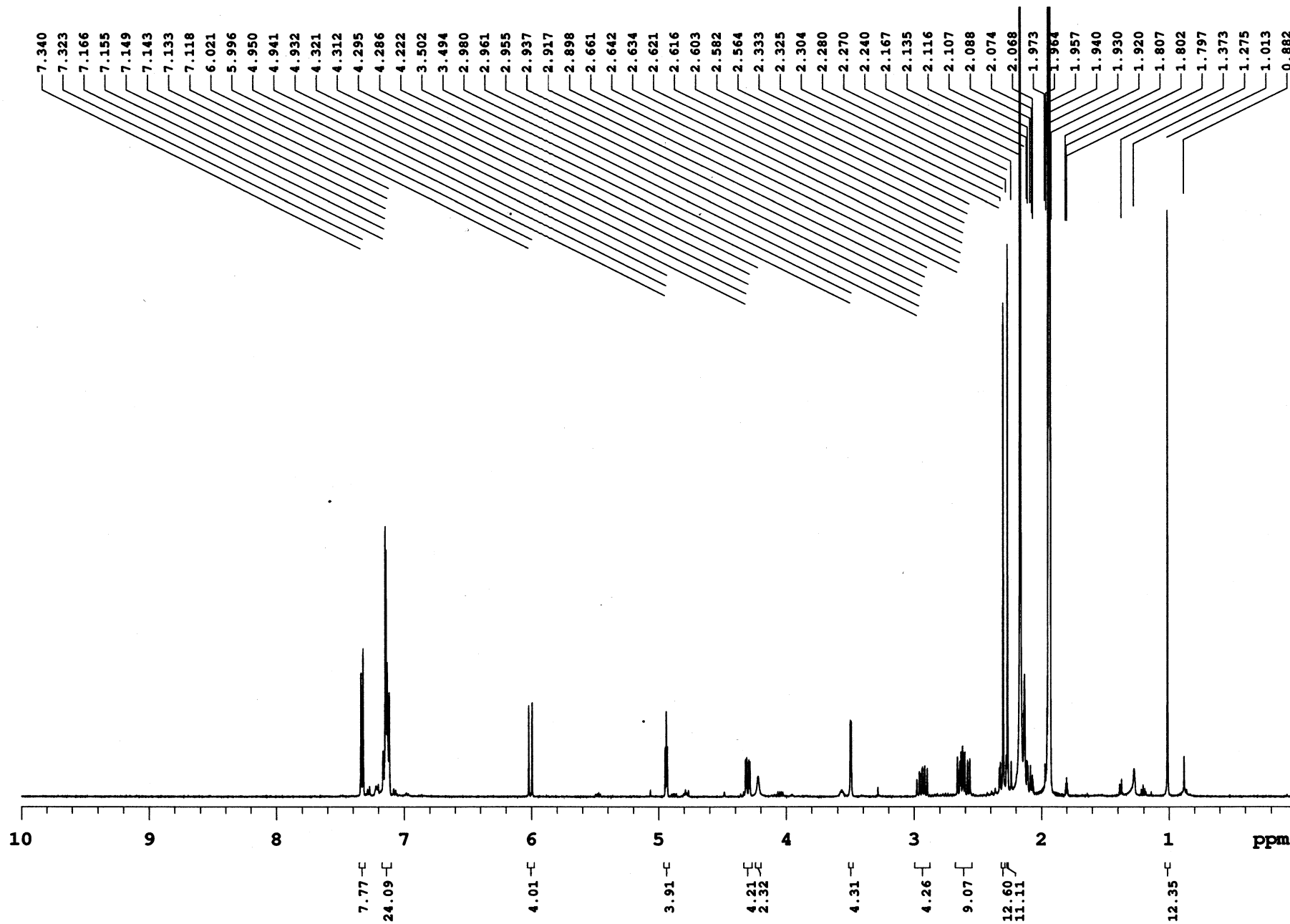


Fig S47. ¹³C NMR (CD₃CN, 125 MHz) of compound 5b.

CHP-8b-f2

Sample Name **CHP-8b-f2**
Date collected **2016-05-20**

Pulse sequence **CARBON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

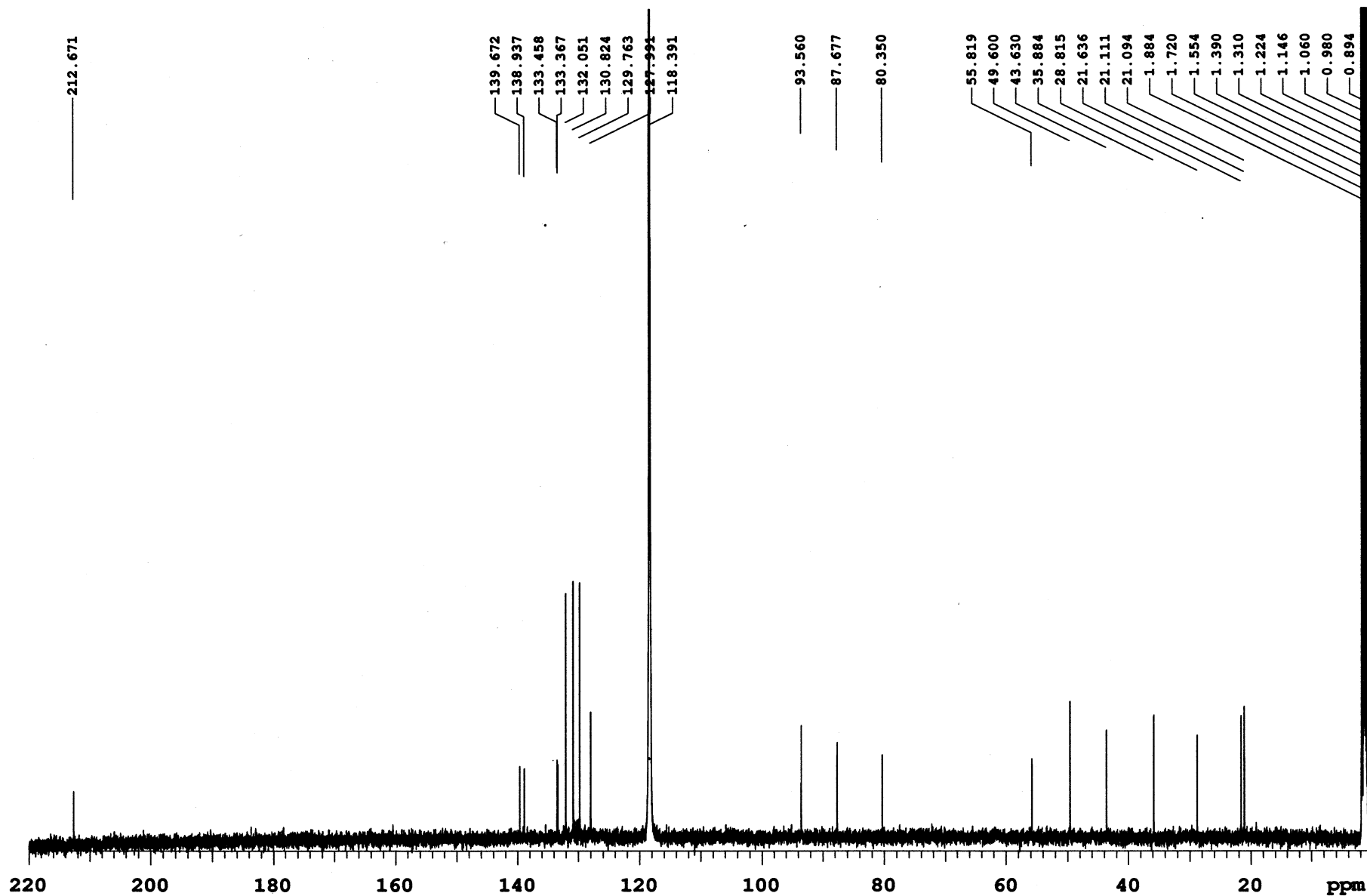


Fig S48. DEPT of compound 5b.

CHP-8b-12

Sample Name **CHP-8b-12**
Date collected **2016-05-20**

Pulse sequence **DEPT**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

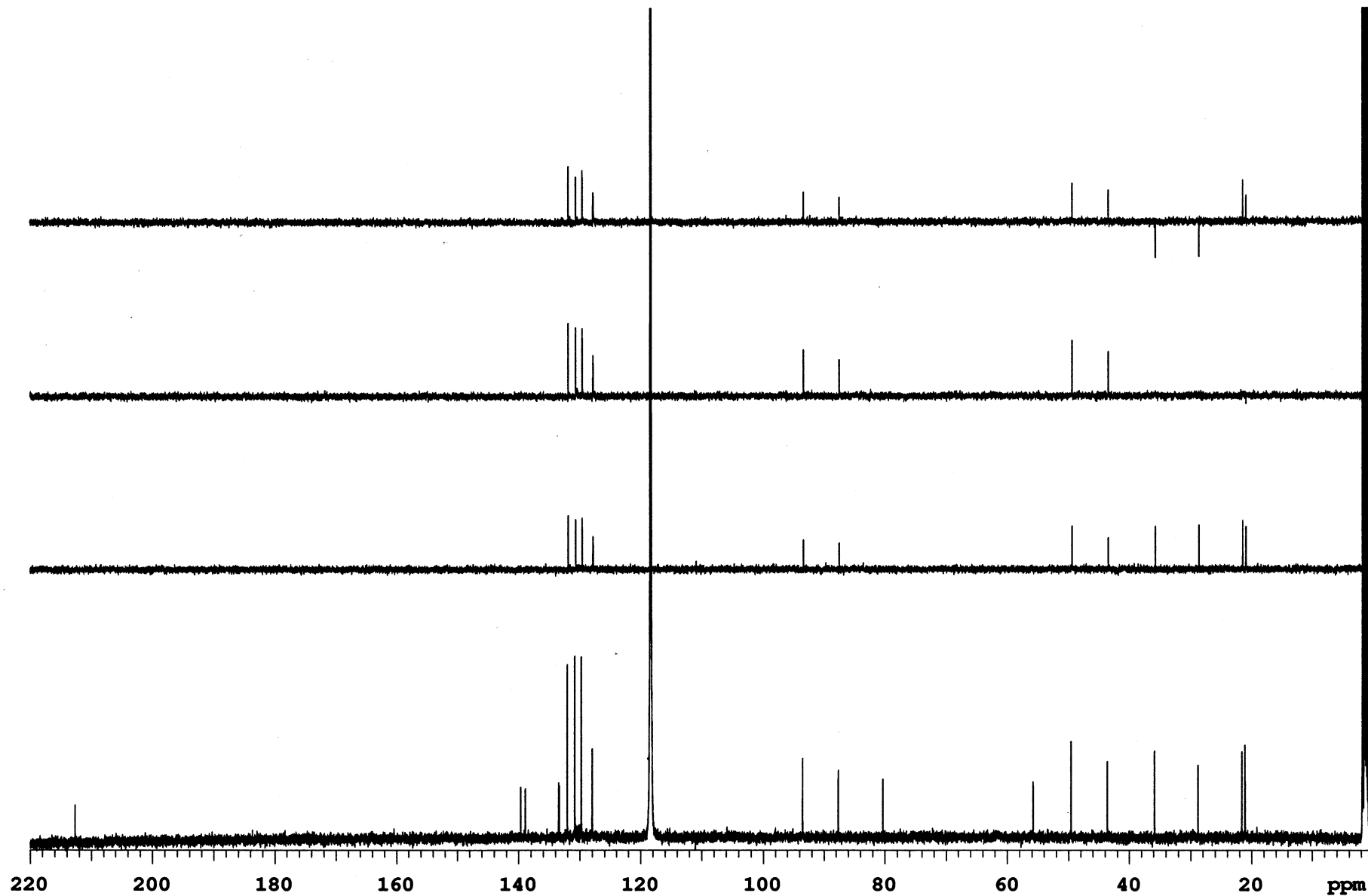


Fig S49. HSQC of compound 5b.

CHP-8b-f2

Sample Name **CHP-8b-f2**
Date collected **2016-05-20**

Pulse sequence **gHSQC**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

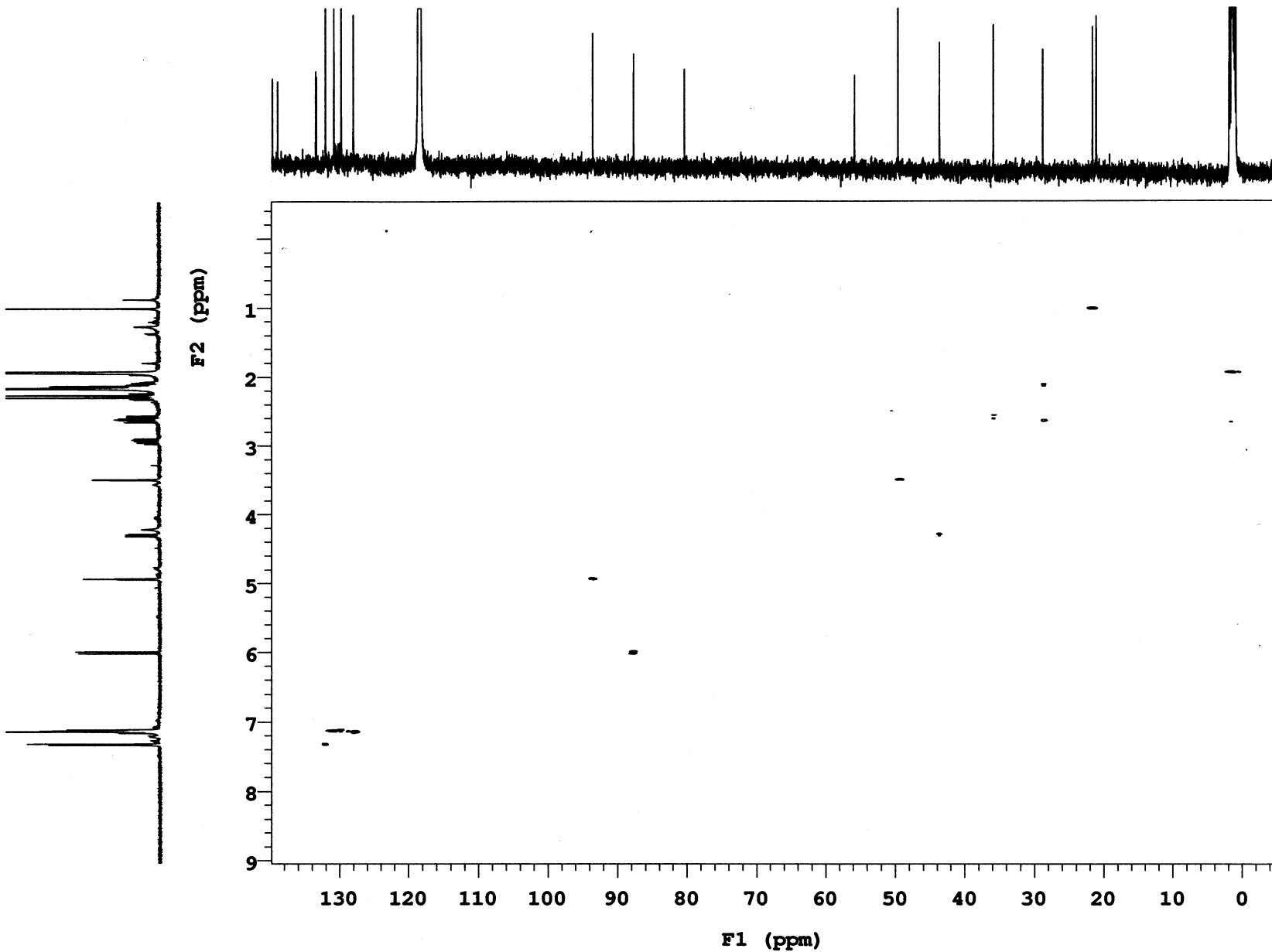


Fig S50. COSY of compound 5b.

CHP-8b-f2

Sample Name **CHP-8b-f2**
Date collected **2016-05-20**

Pulse sequence **gCOSY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

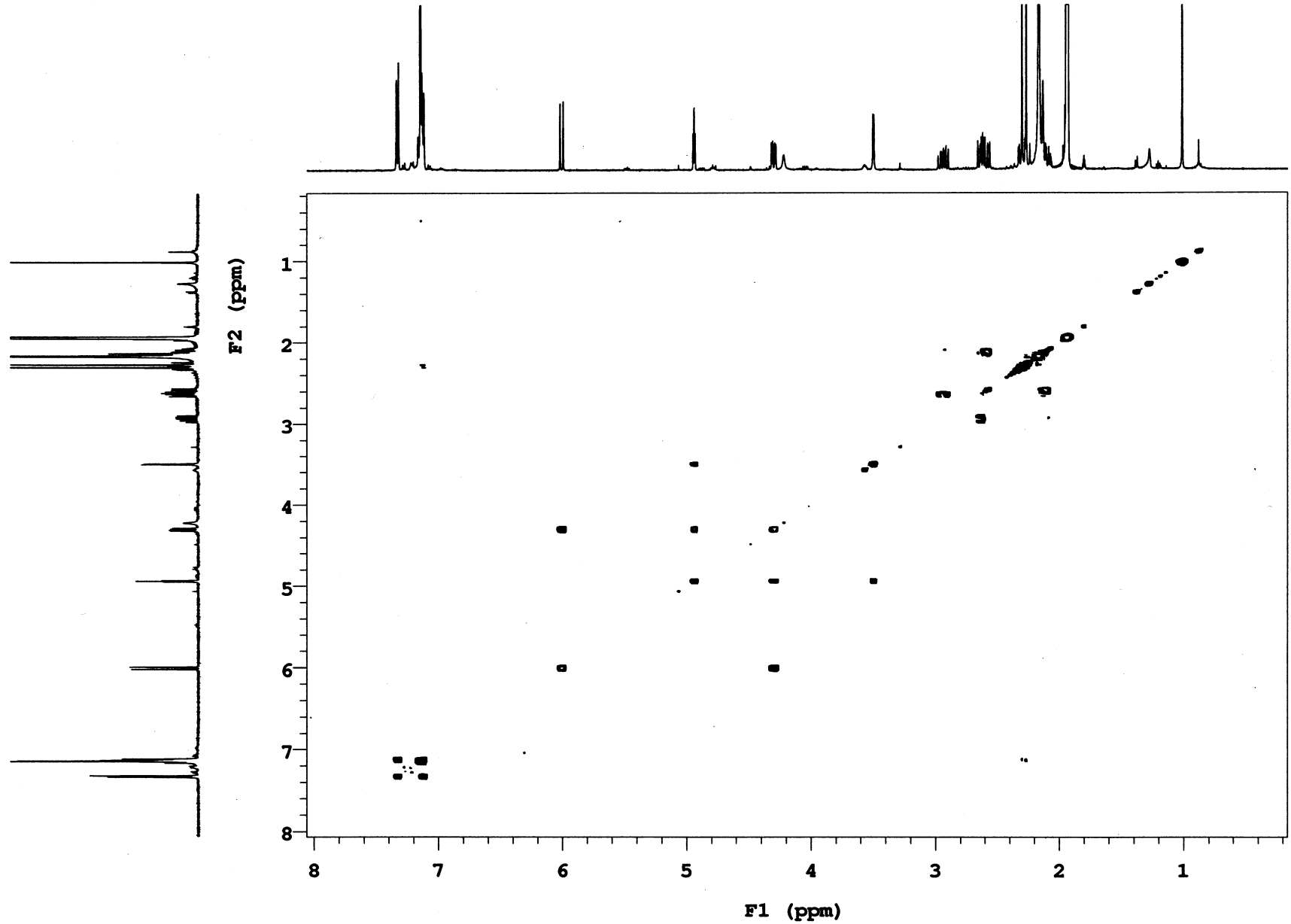


Fig S51. NOESY of compound 5b.

CHP-8b-12

Sample Name **CHP-8b-12**
Date collected **2016-05-20**

Pulse sequence **NOESY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

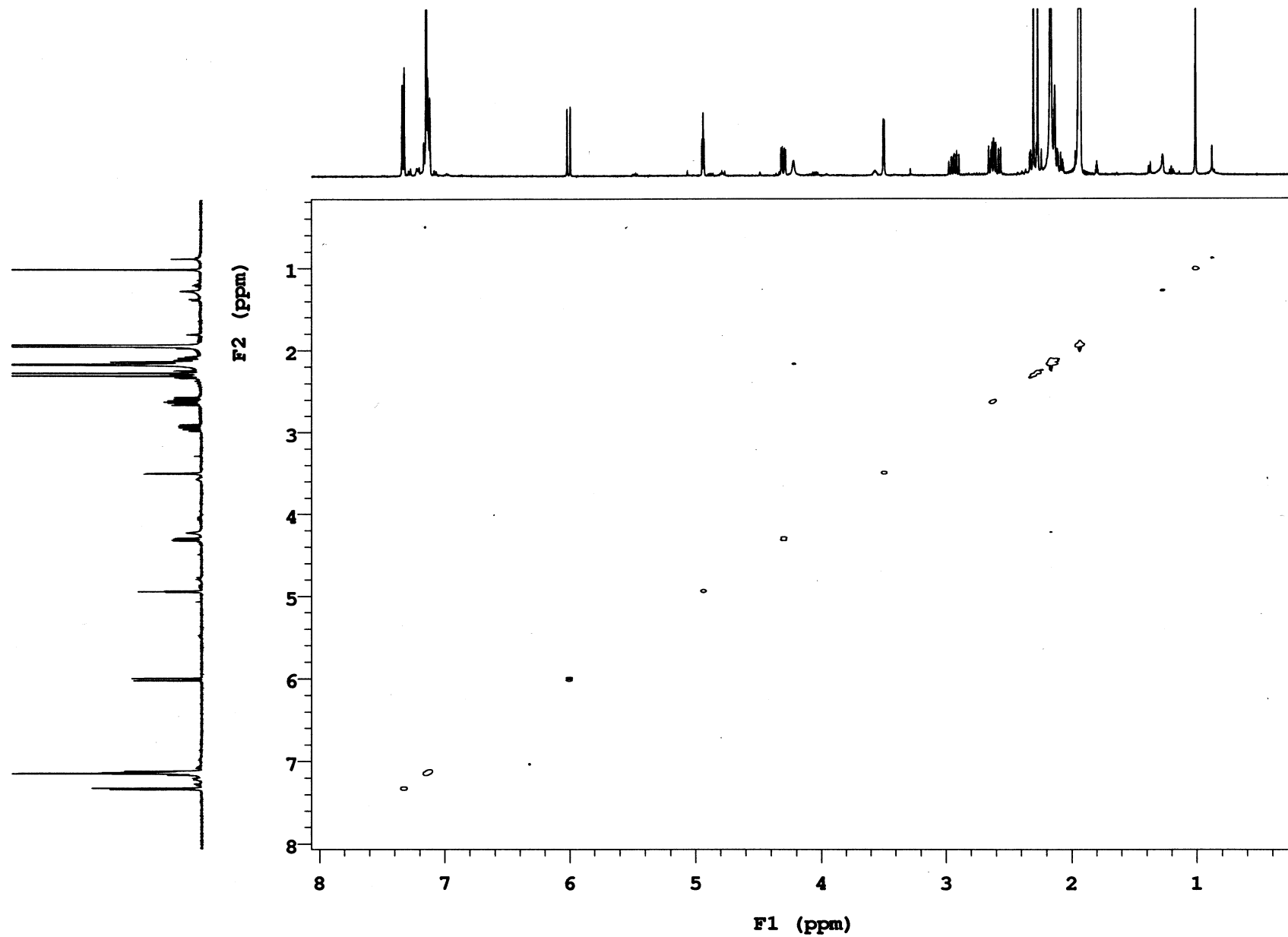


Fig S52. ¹H NMR (CDCl₃, 500 MHz) of compound 3c.

CHP-8C

Sample Name	CHP-8C	Pulse sequence	PROTON	Temperature	25	Study owner	vnmr2
Date collected	2016-03-26	Solvent	cdcl3	Spectrometer	Agilent-NMR-inova500	Operator	vnmr2

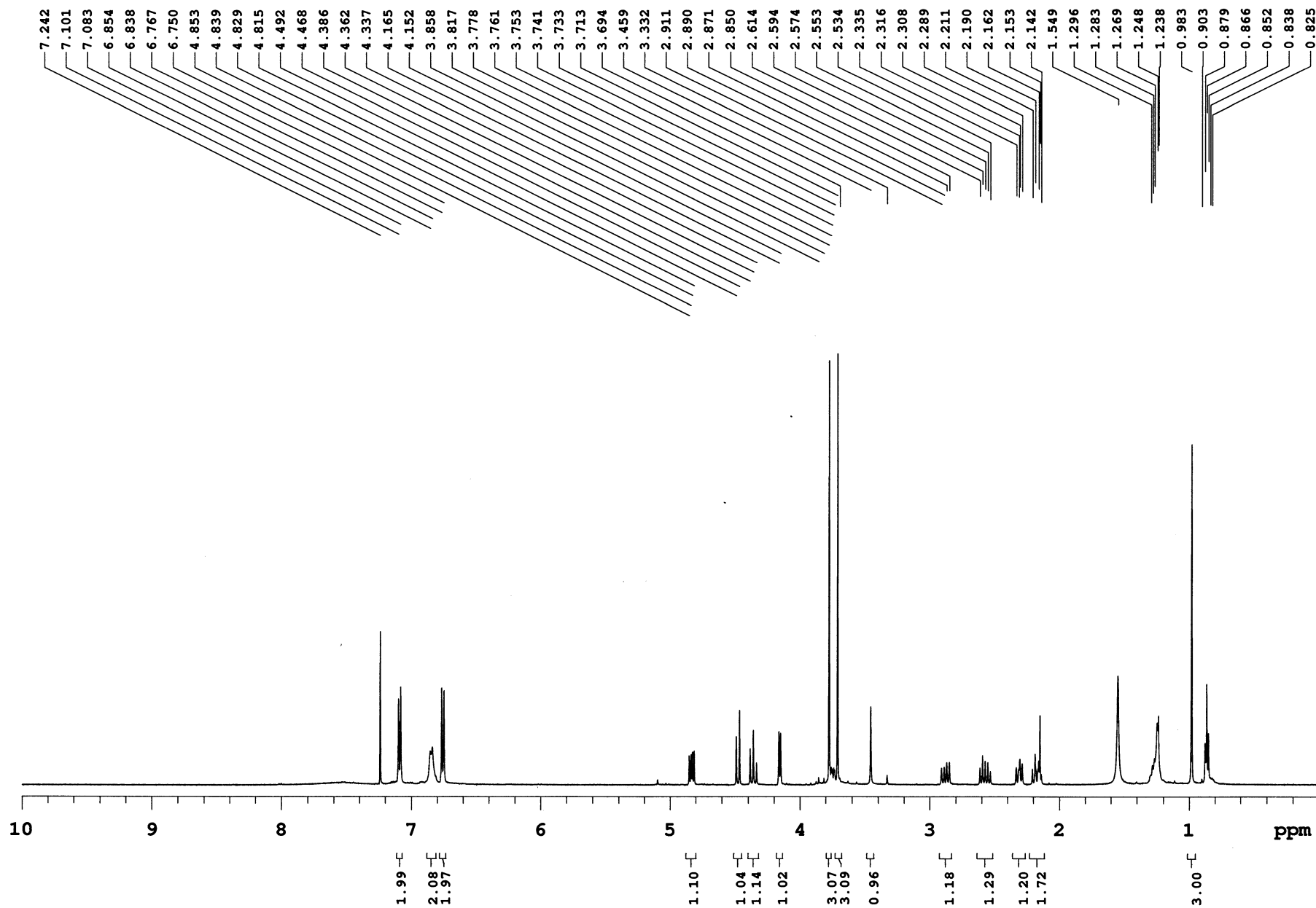


Fig S53. ¹³C NMR (CDCl₃, 125 MHz) of compound 3c.

CHP-8C

Sample Name **CHP-8C**
Date collected **2016-03-26**

Pulse sequence **CARBON**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

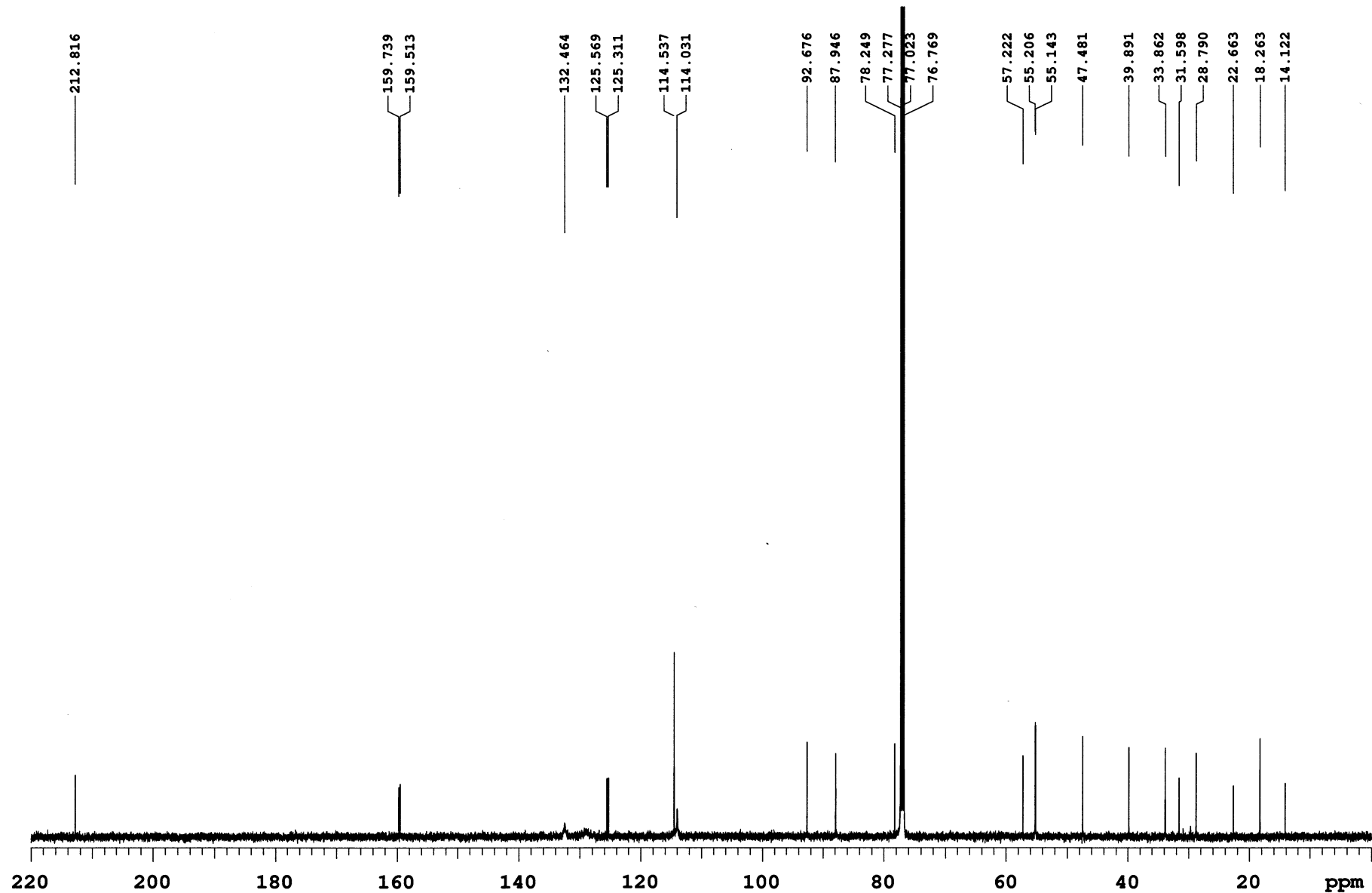


Fig S54. DEPT of compound 3c.

CHP-8C

Sample Name **CHP-8C**
Date collected **2016-03-26**

Pulse sequence **DEPT**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

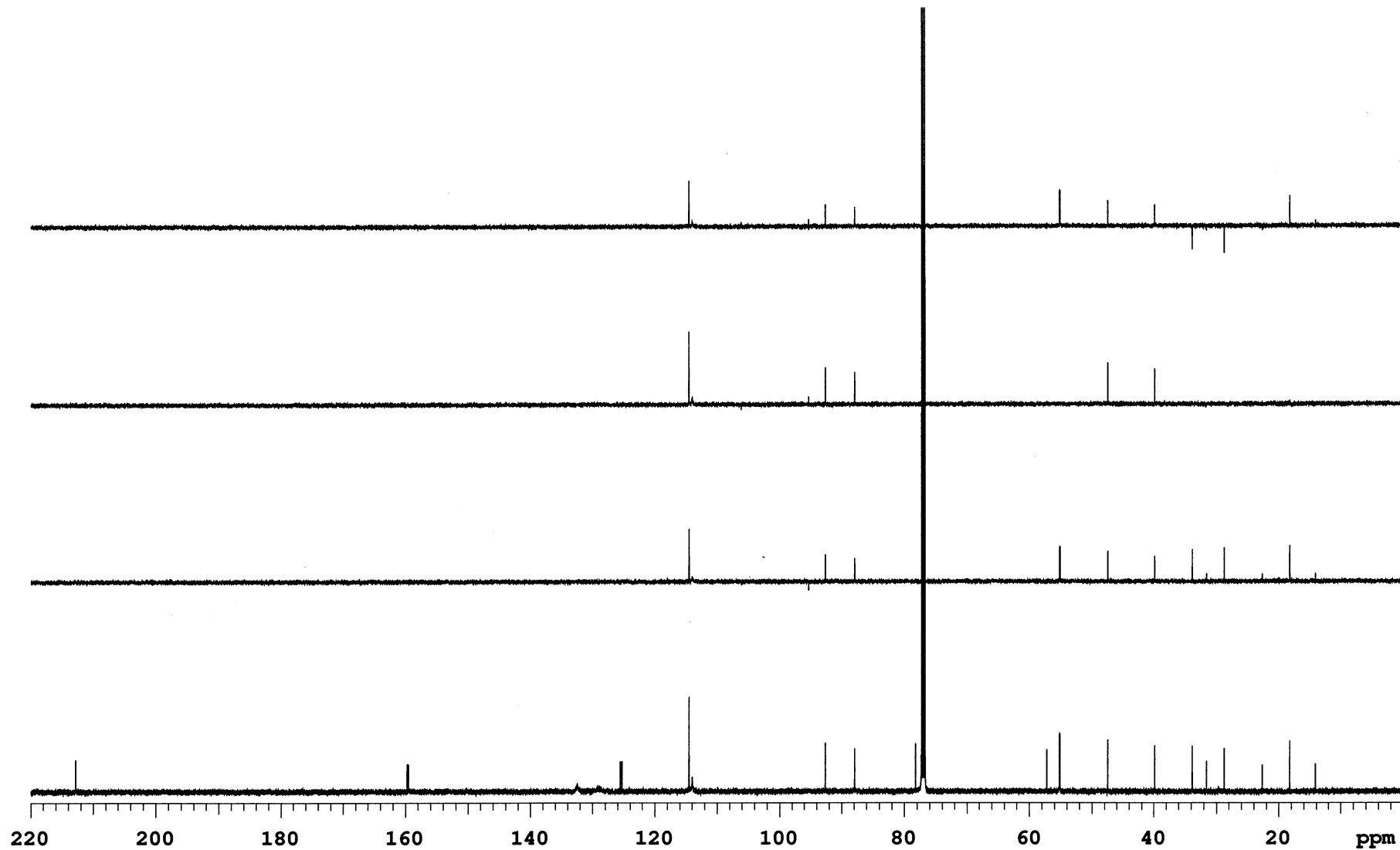


Fig S55. HSQC of compound 3c.

CHP-8C

Sample Name **CHP-8C**
Date collected **2016-03-26**

Pulse sequence **gHSQC**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

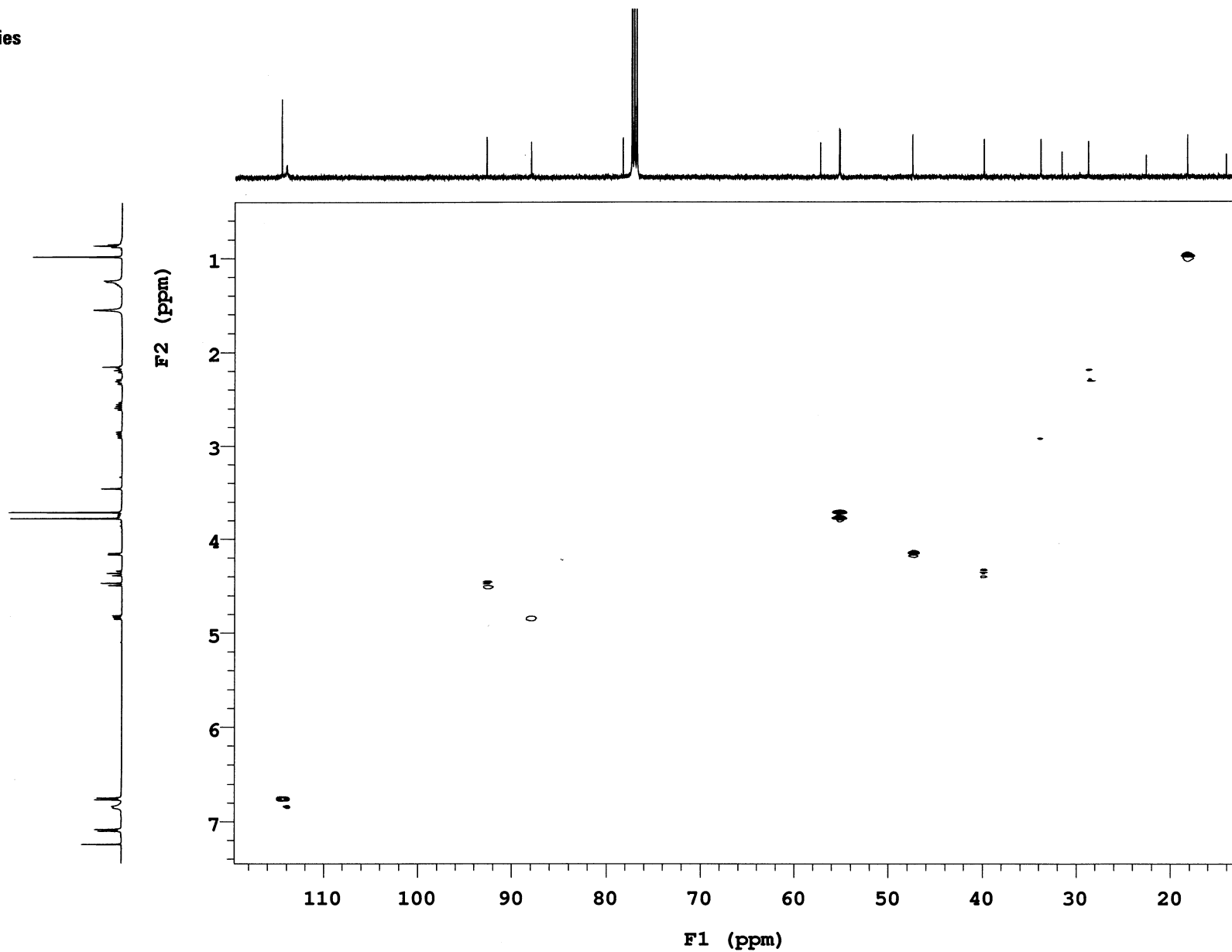


Fig S56. COSY of compound 3c.

CHP-8C

Sample Name **CHP-8C**
Date collected **2016-03-26**

Pulse sequence **gCOSY**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

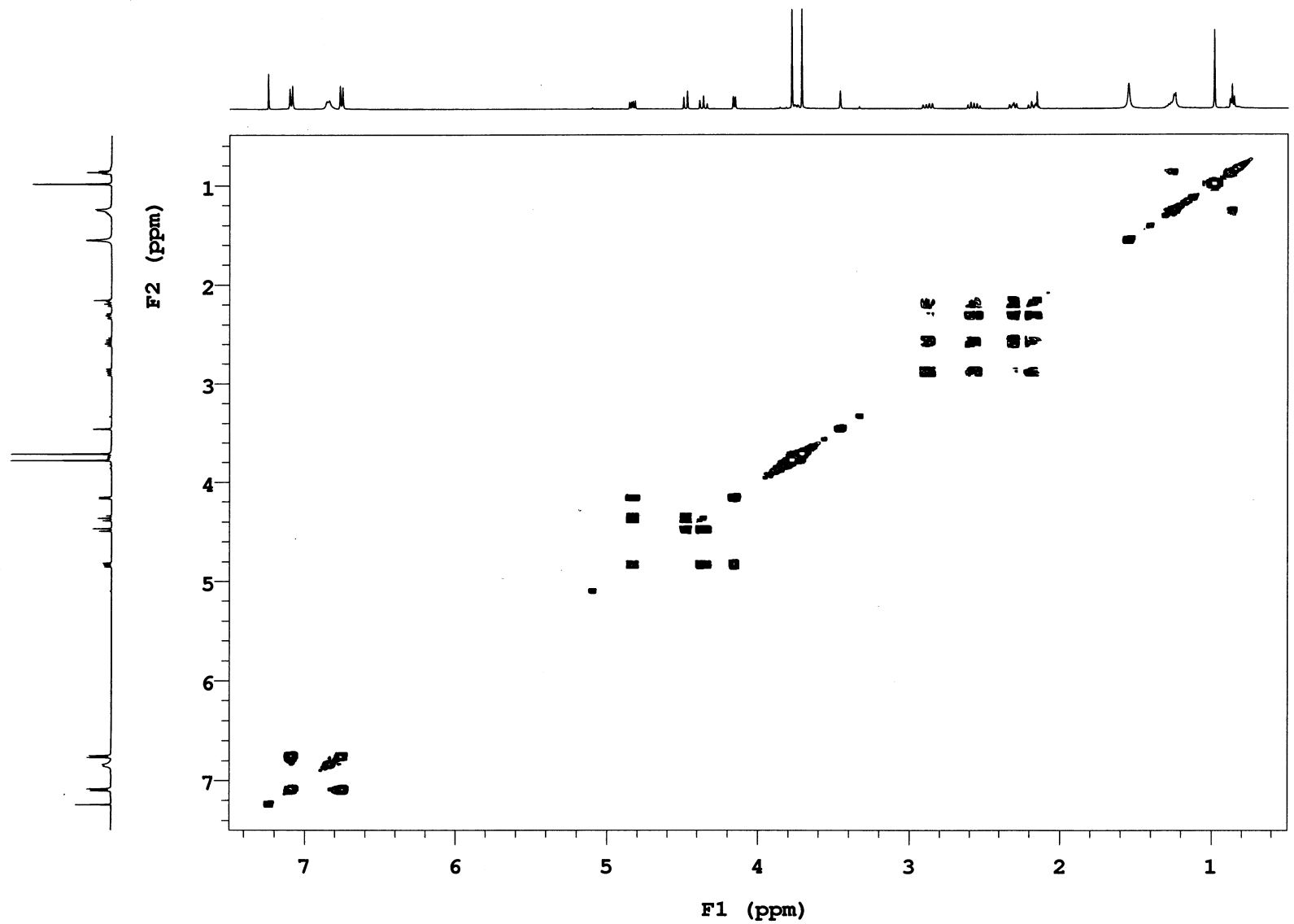


Fig S57. NOESY of compound 3c.

CHP-8C

Sample Name **CHP-8C**
Date collected **2016-03-26**

Pulse sequence **NOESY**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

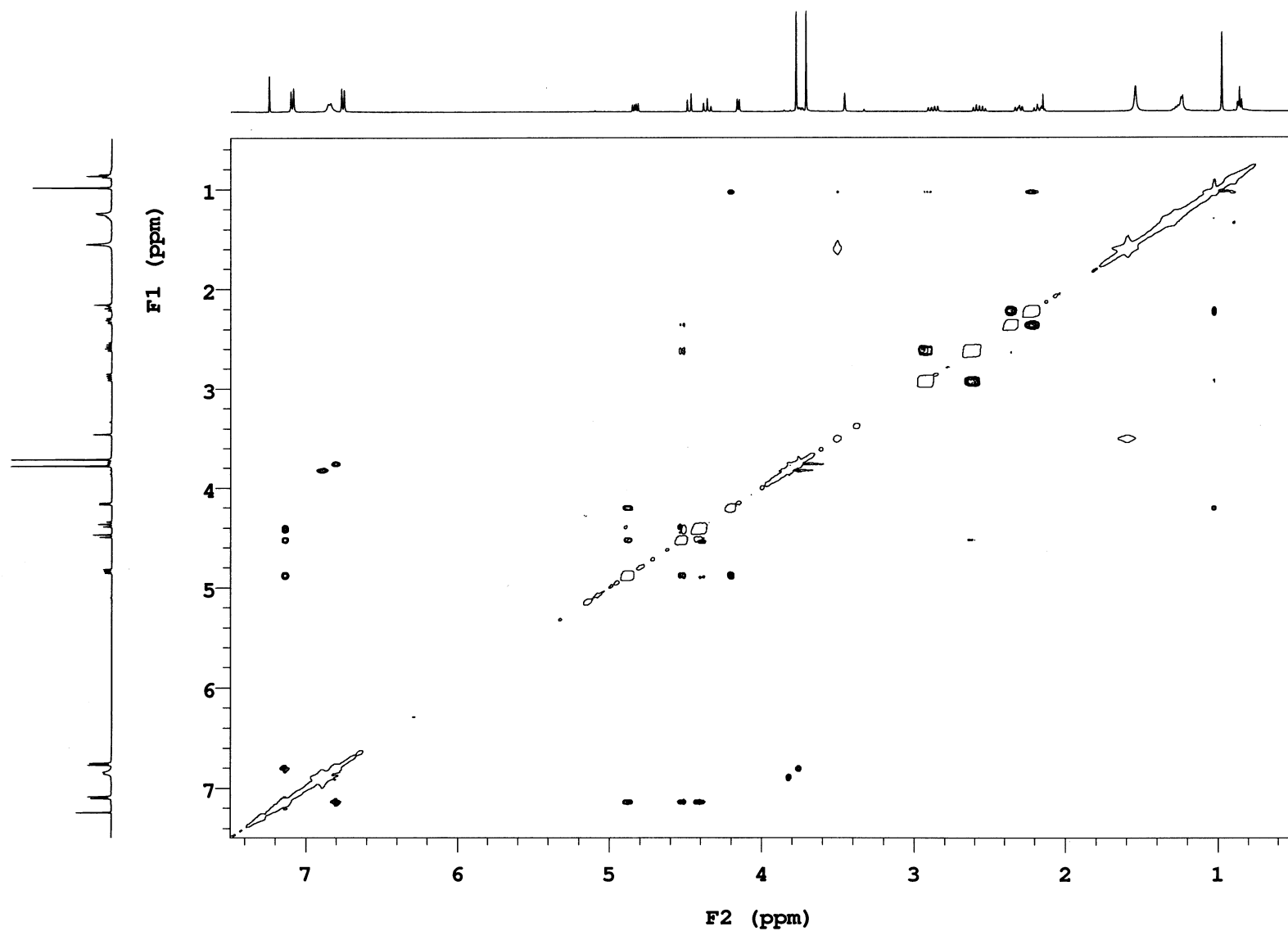


Fig S58. ¹H NMR (CDCl₃, 500 MHz) of compound 5c.

CHP-8c-f2

Sample Name	CHP-8c-f2	Pulse sequence	PROTON	Temperature	25	Study owner	vnmr2
Date collected	2016-05-28	Solvent	cdcl3	Spectrometer	Agilent-NMR-inova500	Operator	vnmr2

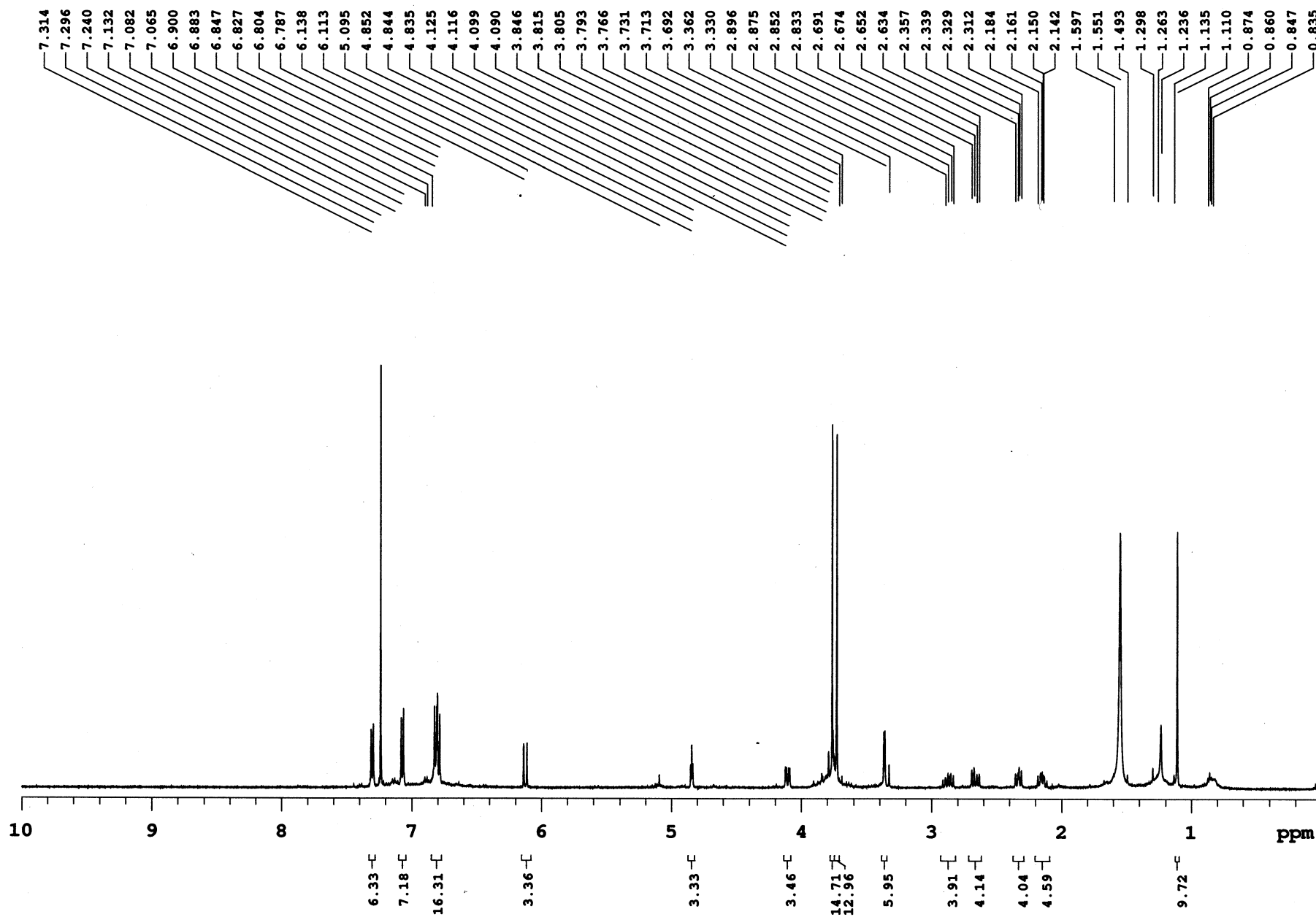


Fig S59. ¹³C NMR (CDCl₃, 125 MHz) of compound 5c.

CHP-8c-f2

Sample Name	CHP-8c-f2	Pulse sequence	CARBON	Temperature	25	Study owner	vnmr2
Date collected	2016-05-28	Solvent	cdcl3	Spectrometer	Agilent-NMR-inova500	Operator	vnmr2

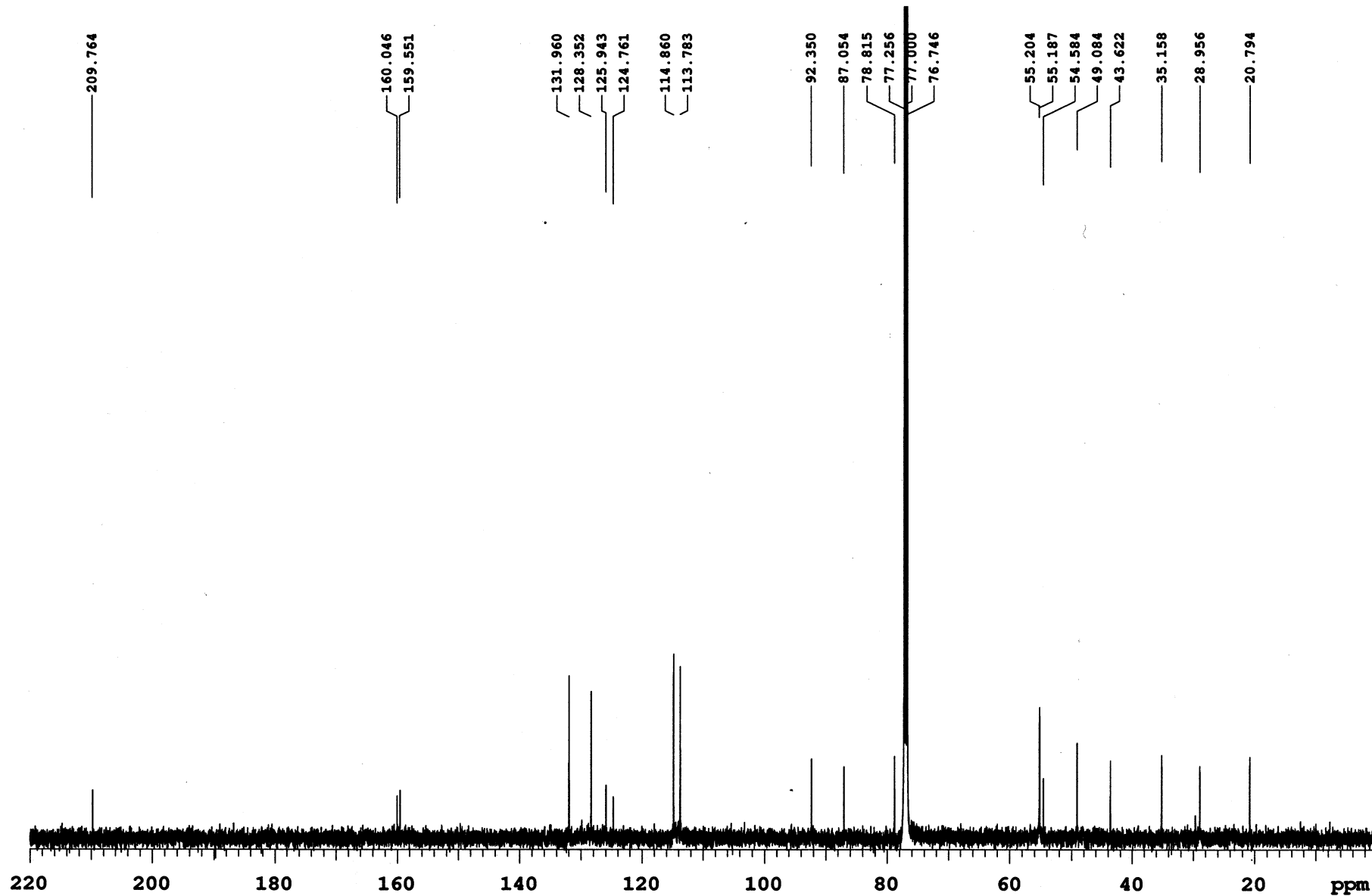


Fig S60. DEPT of compound 5c.

CHP-8c-f2

Sample Name **CHP-8c-f2**
Date collected **2016-05-28**

Pulse sequence **DEPT**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

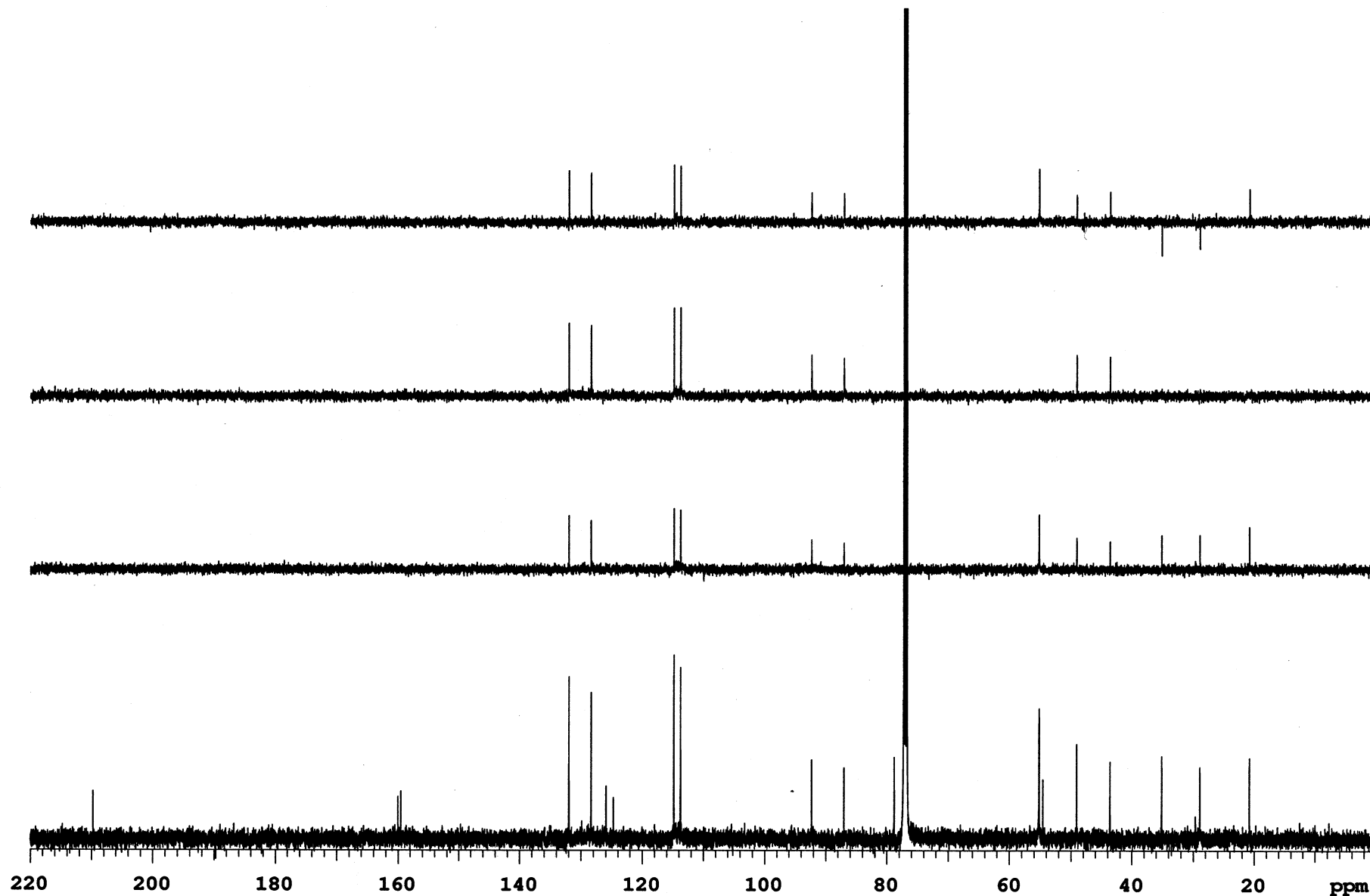


Fig S61. HSQC of compound 5c.

CHP-8c-f2

Sample Name **CHP-8c-f2**
Date collected **2016-05-28**

Pulse sequence **gHSQC**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

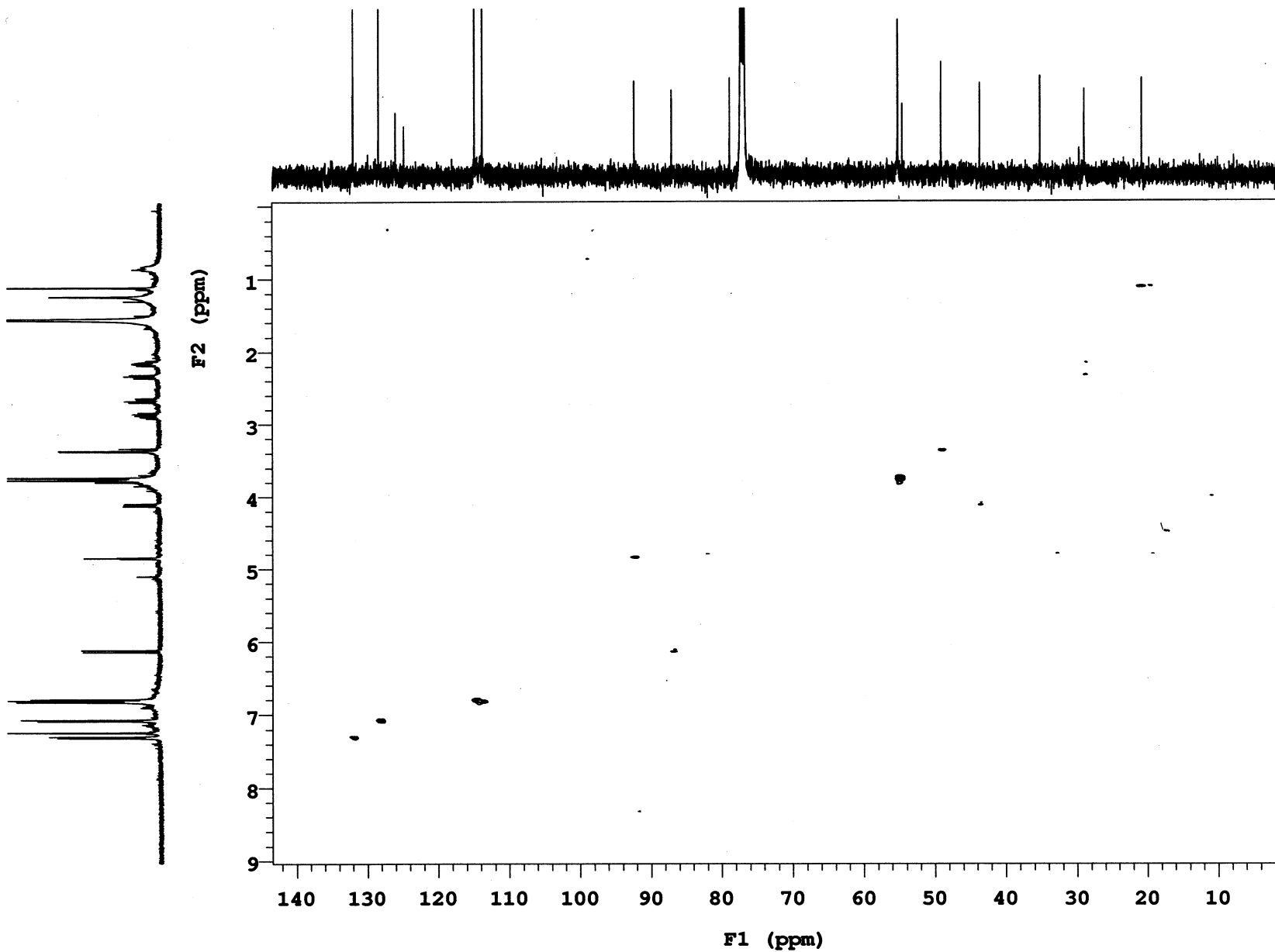


Fig S63. NOESY of compound 5c.

CHP-8c-f2

Sample Name **CHP-8c-f2**
Date collected **2016-05-28**

Pulse sequence **NOESY**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

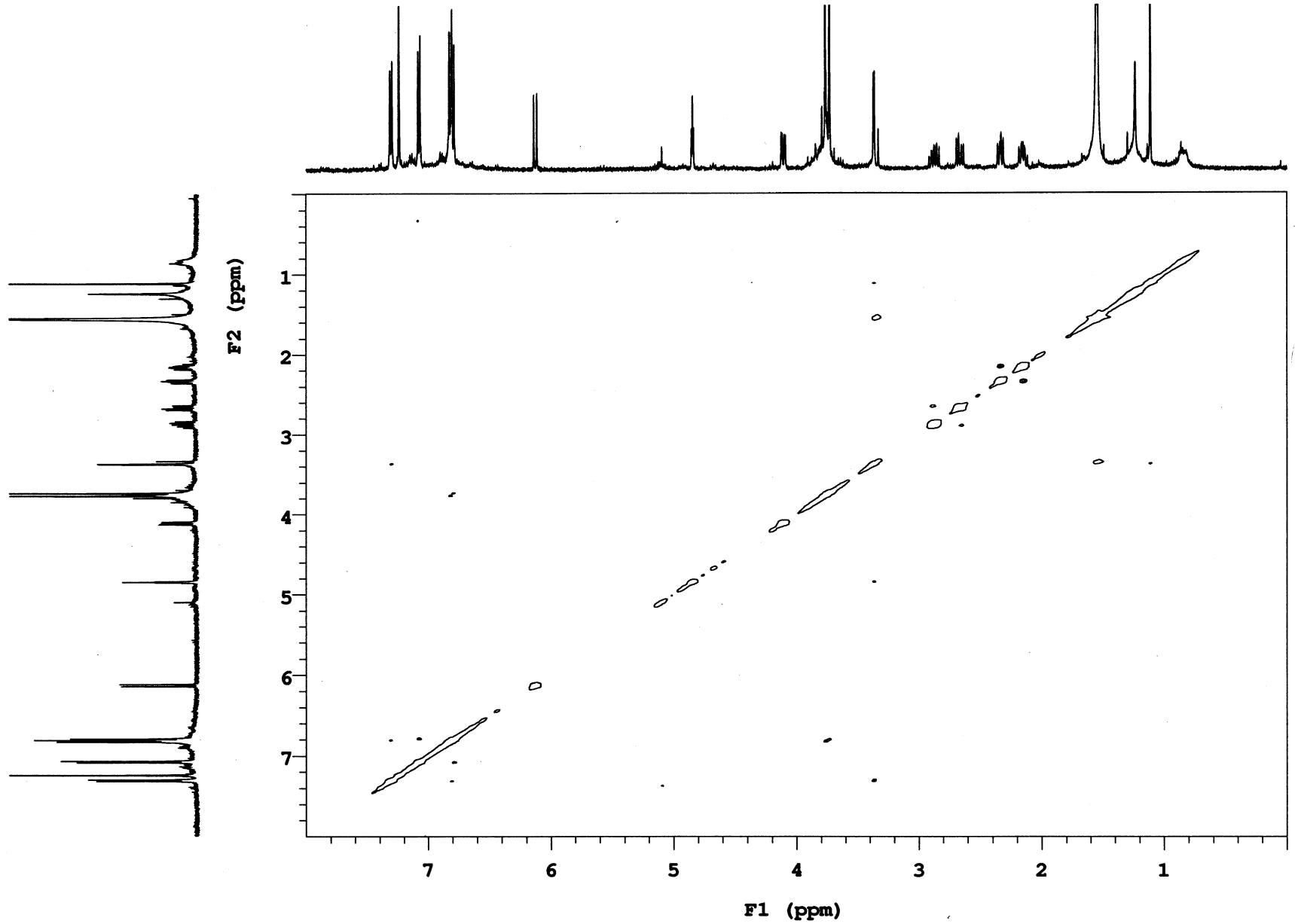


Fig S64. ¹H NMR (CD₃CN, 500 MHz) of compound 3d.

CHP-8e

Sample Name	CHP-8e	Pulse sequence	PROTON	Temperature	25	Study owner	vnmr2
Date collected	2016-03-24	Solvent	cd3cn	Spectrometer	Agilent-NMR-inova500	Operator	vnmr2

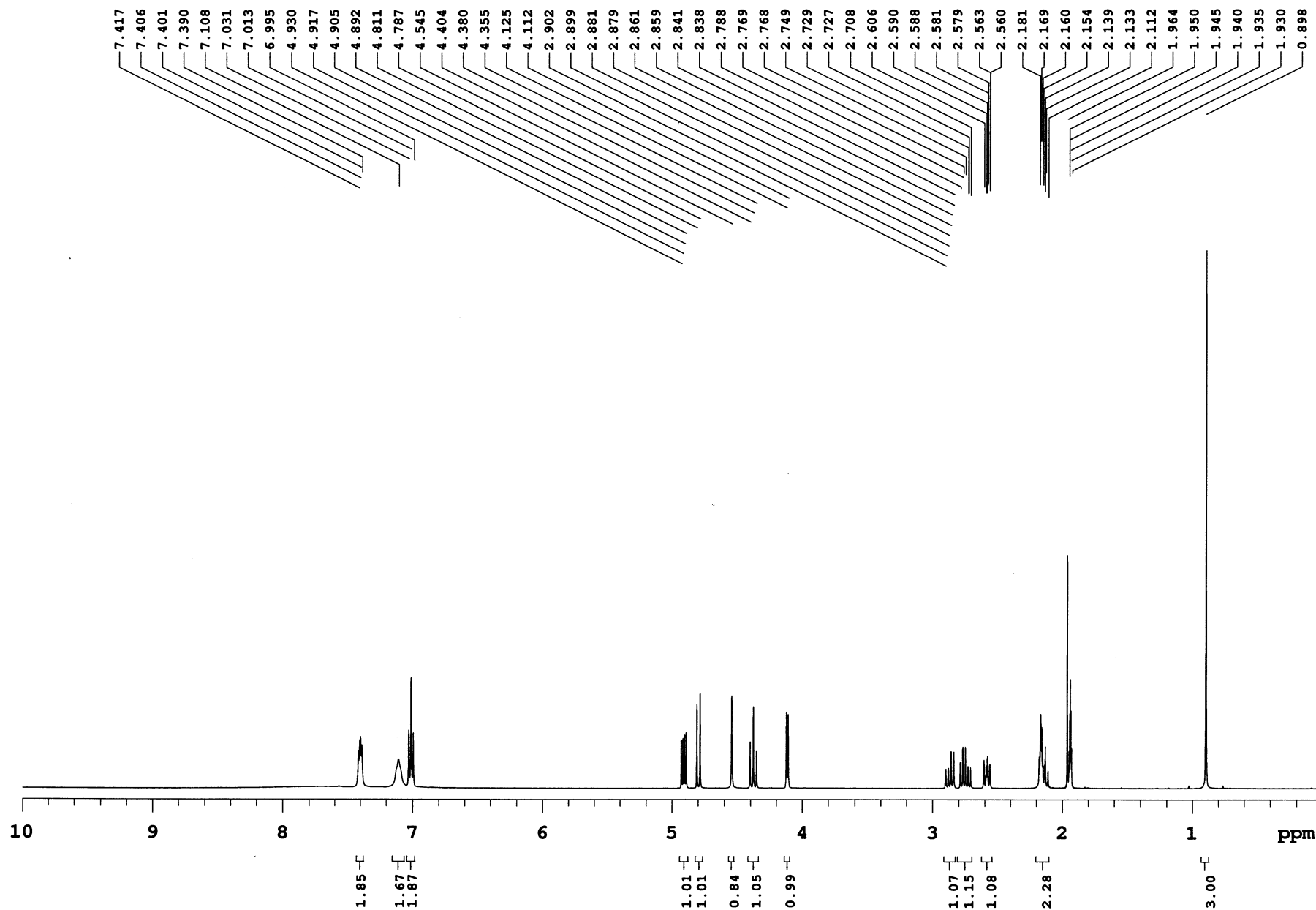


Fig S65. ¹³C NMR (CD₃CN, 125 MHz) of compound 3d.

CHP-8e

Sample Name **CHP-8e**
Date collected **2016-03-24**

Pulse sequence **CARBON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

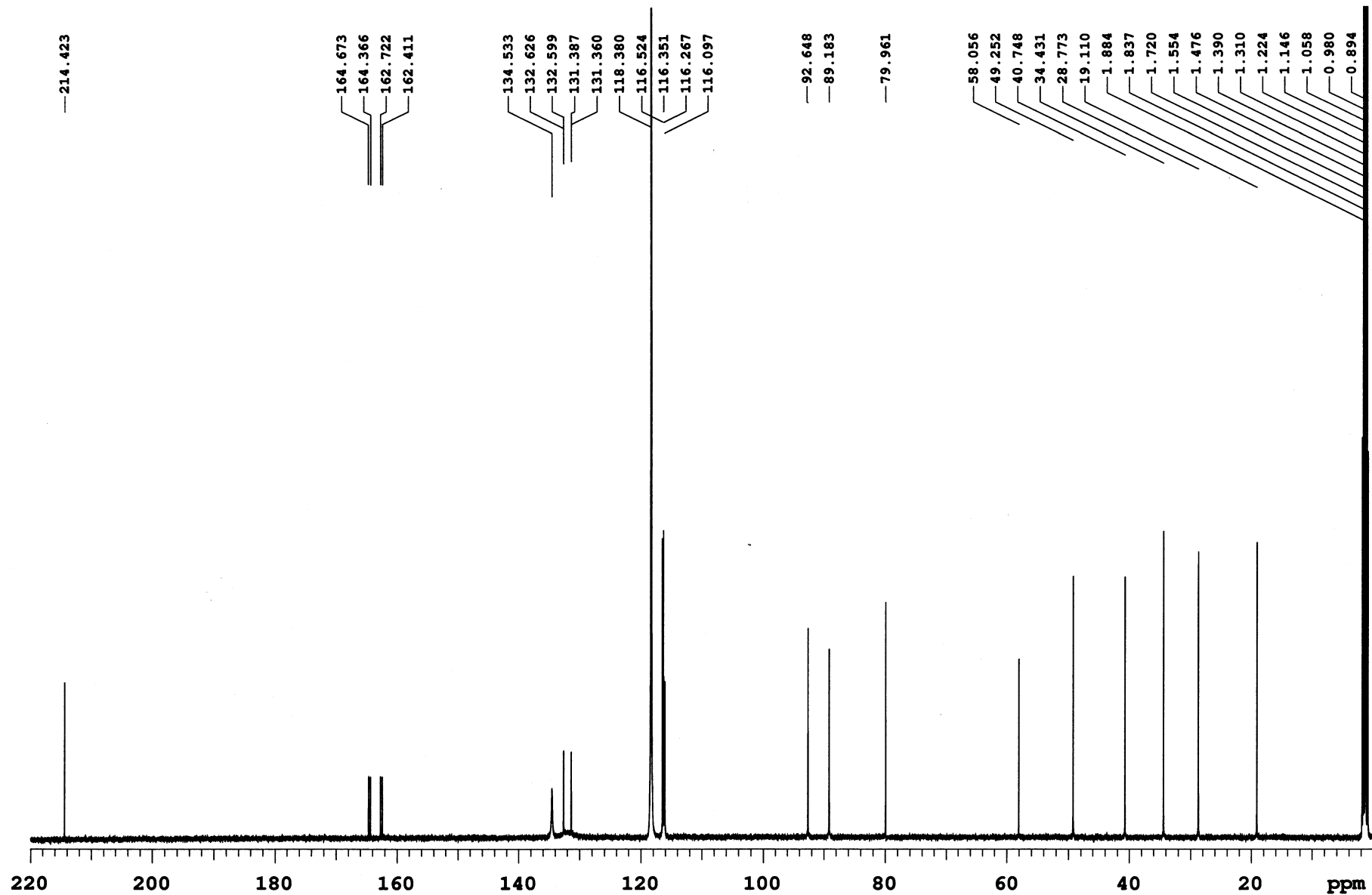


Fig S66. DEPT of compound 3d.

CHP-8e

Sample Name **CHP-8e**
Date collected **2016-03-24**

Pulse sequence **DEPT**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

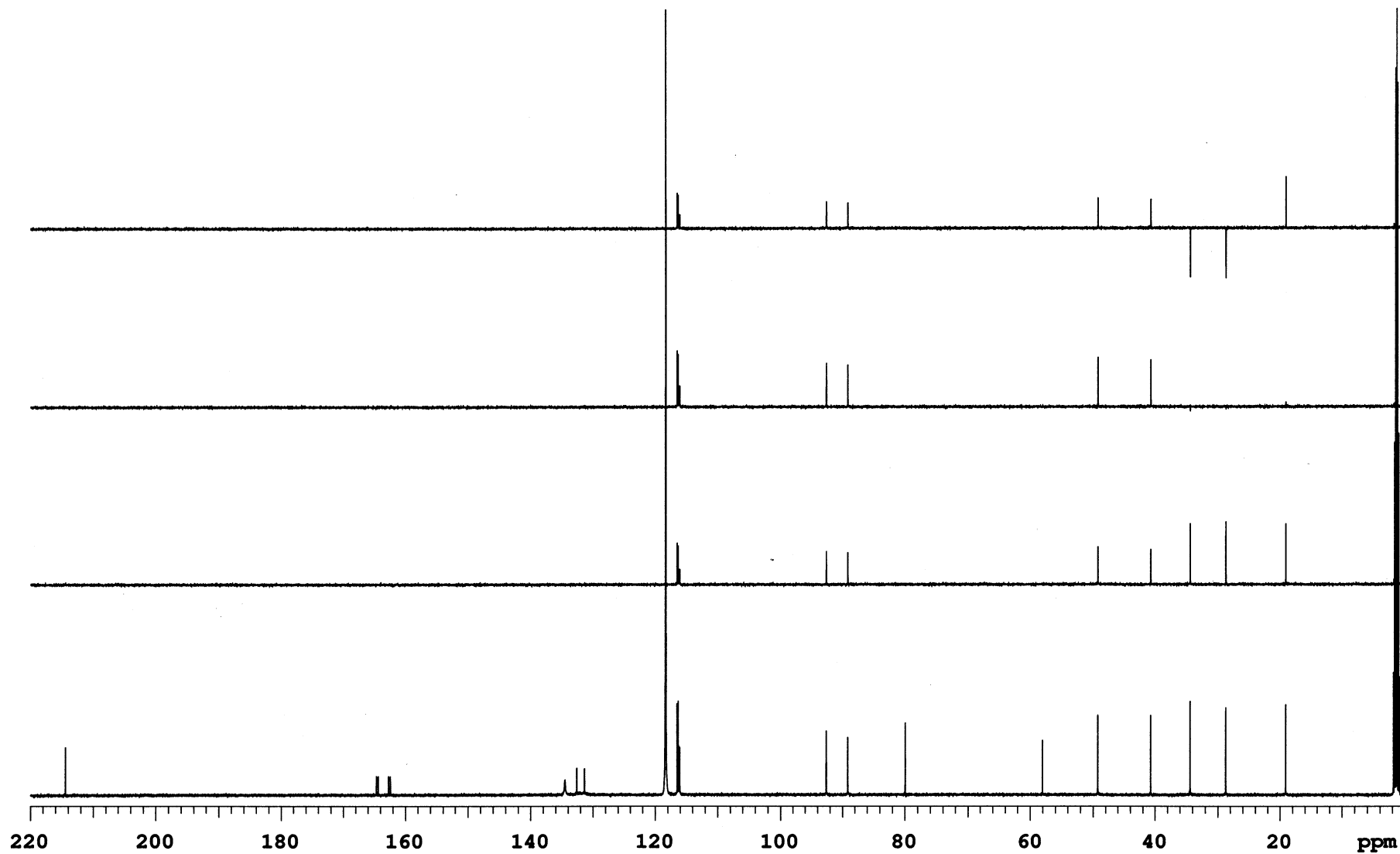


Fig S67. DEPT of compound 3d, expanded.

CHP-8e

Sample Name	CHP-8e	Pulse sequence	DEPT	Temperature	25	Study owner	vnmr2
Date collected	2016-03-24	Solvent	cd3cn	Spectrometer	Agilent-NMR-inova500	Operator	vnmr2

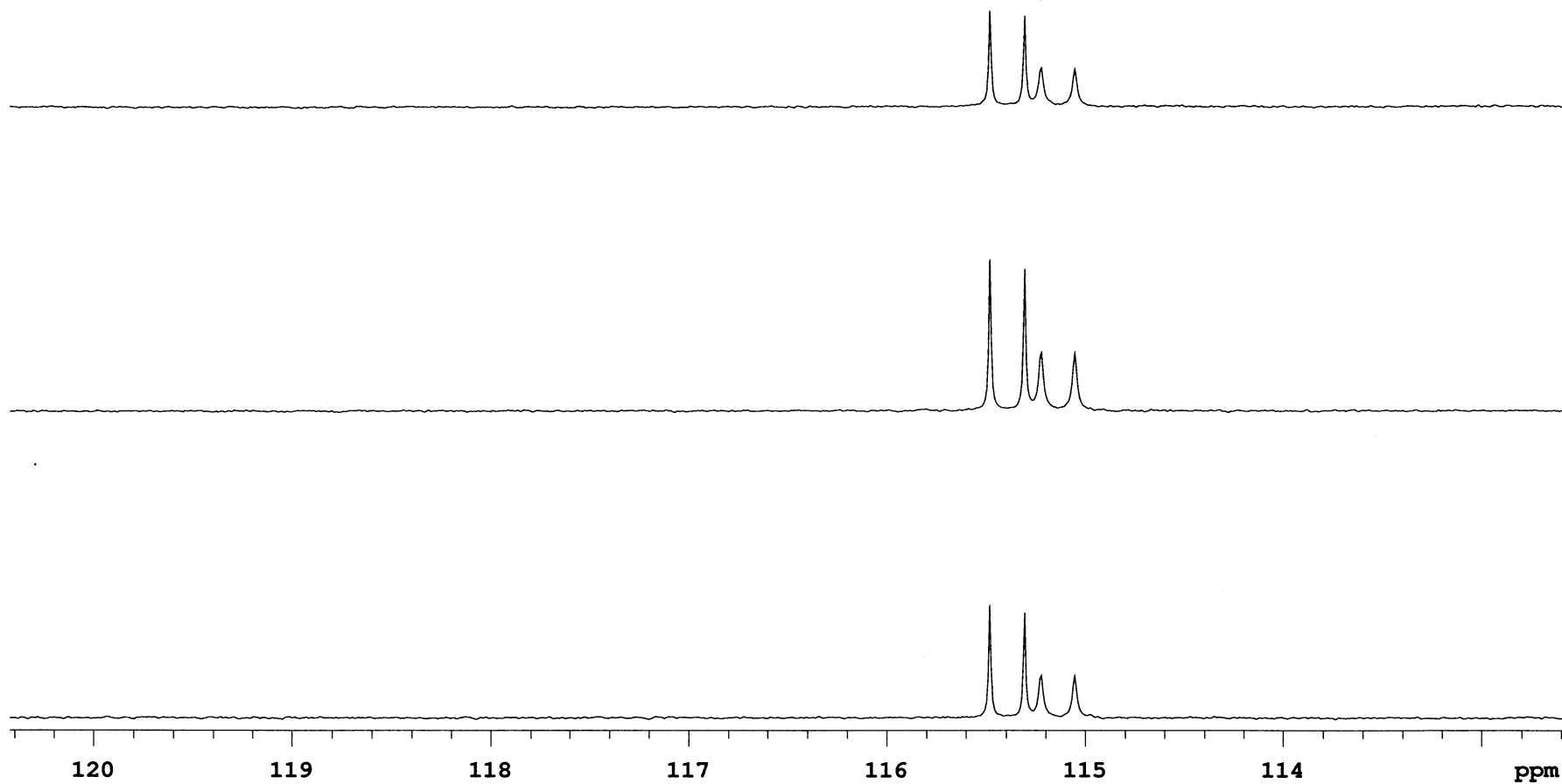


Fig S68. HSQC of compound 3d.

CHP-8e

Sample Name **CHP-8e**
Date collected **2016-03-24**

Pulse sequence **gHSQC**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

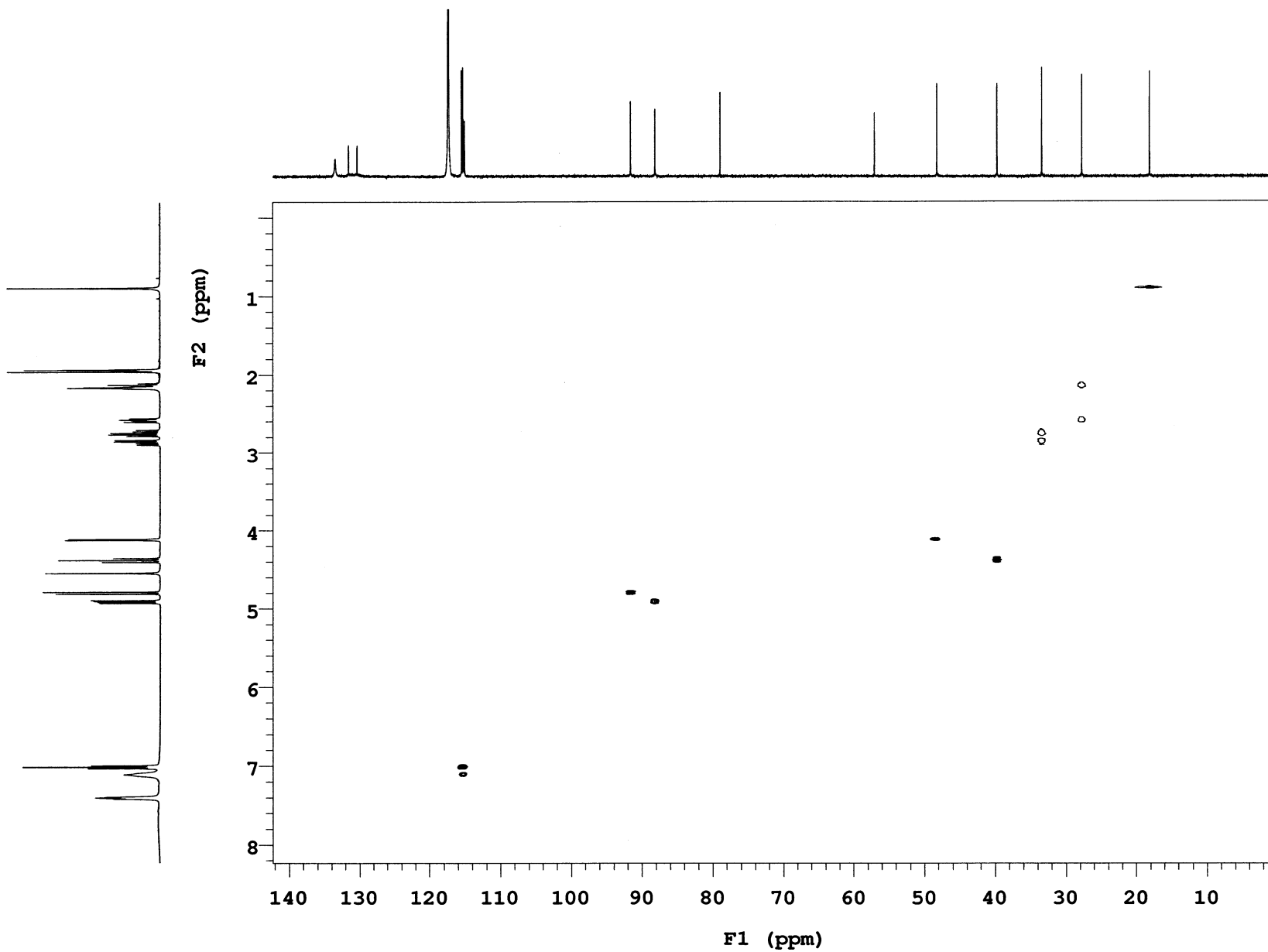


Fig S69. COSY of compound 3d.

CHP-8e

Sample Name **CHP-8e**
Date collected **2016-03-24**

Pulse sequence **gCOSY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

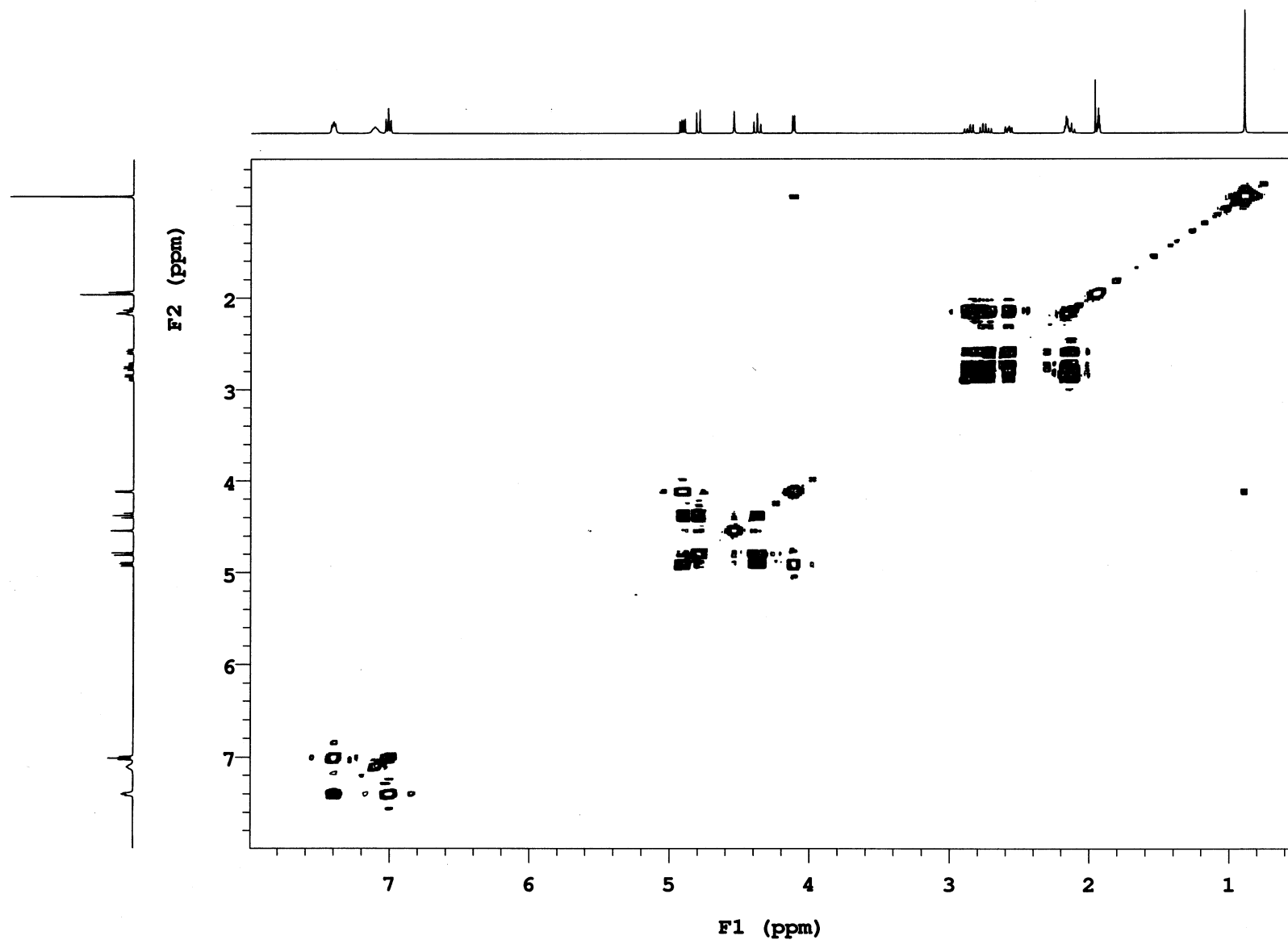


Fig S71. ¹H NMR (CD₃CN, 500 MHz) of compound 5d.

CHP-8e-f2

Sample Name **CHP-8e-f2**
Date collected **2016-05-11**

Pulse sequence **PROTON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

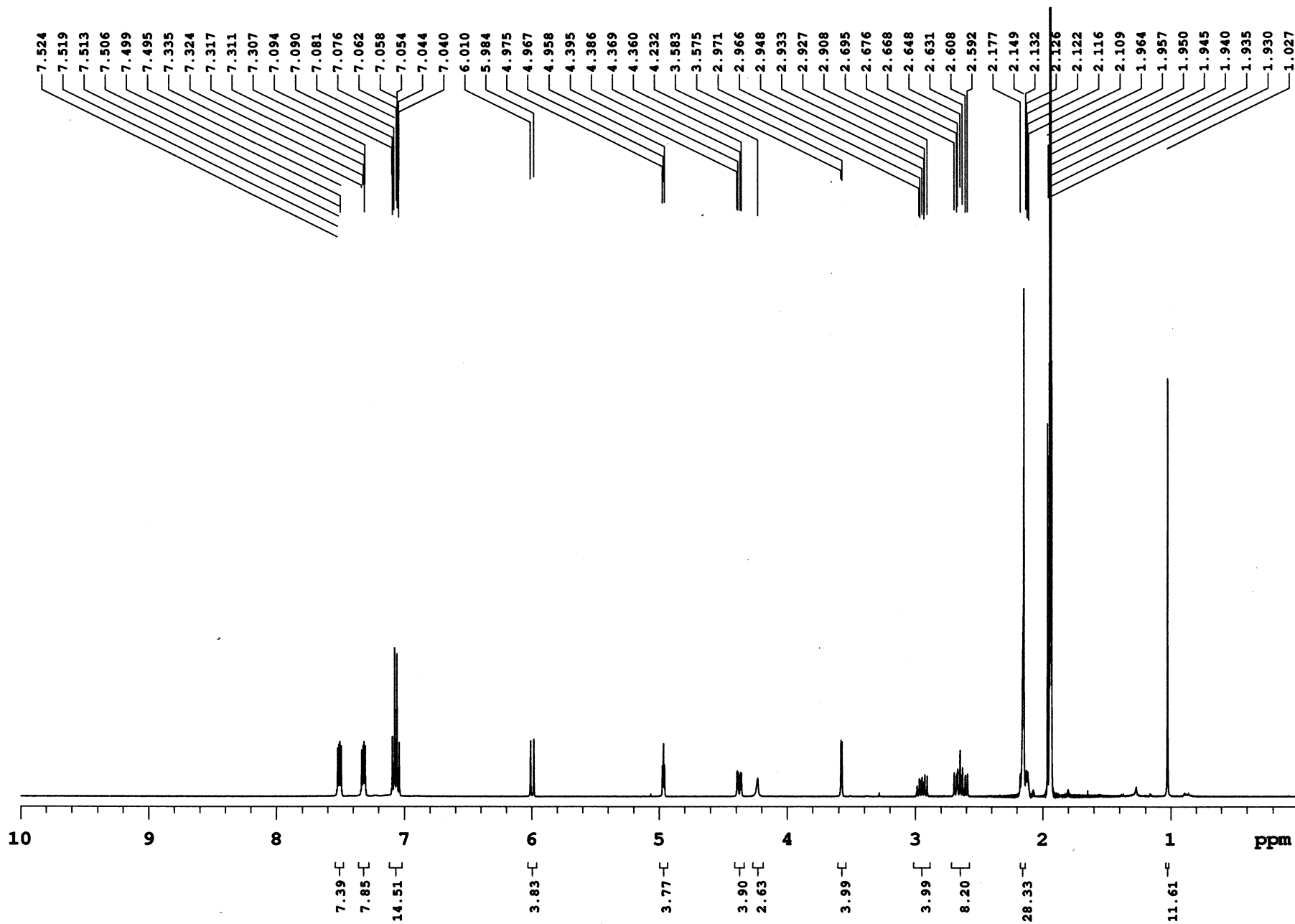


Fig S72. ¹³C NMR (CD₃CN, 125 MHz) of compound 5d.

CHP-8e-12

Sample Name **CHP-8e-12**
Date collected **2016-05-11**

Pulse sequence **CARBON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

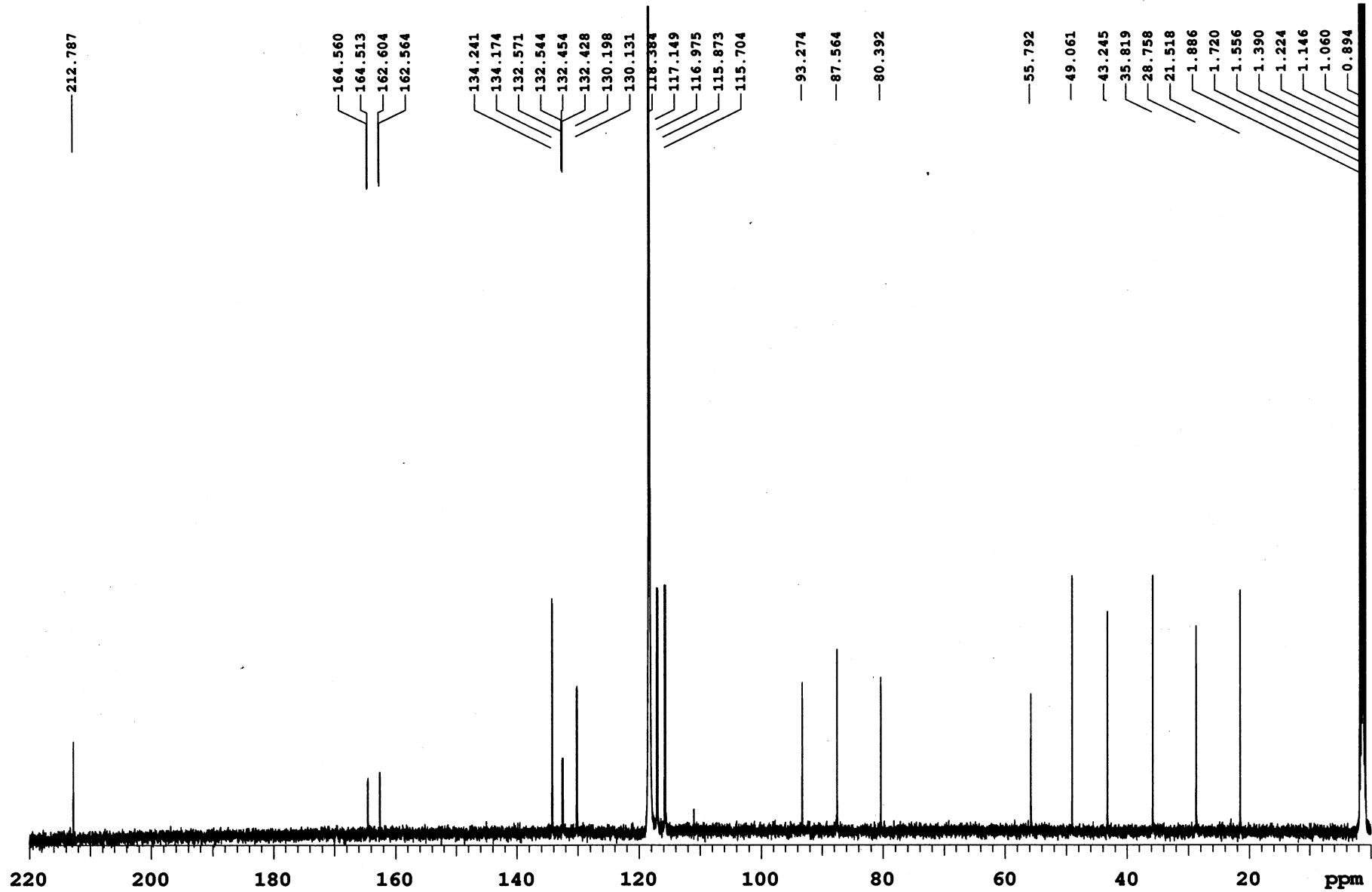


Fig S73. DEPT of compound 5d.

CHP-8e-12

Sample Name **CHP-8e-12**
Date collected **2016-05-12**

Pulse sequence **DEPT**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

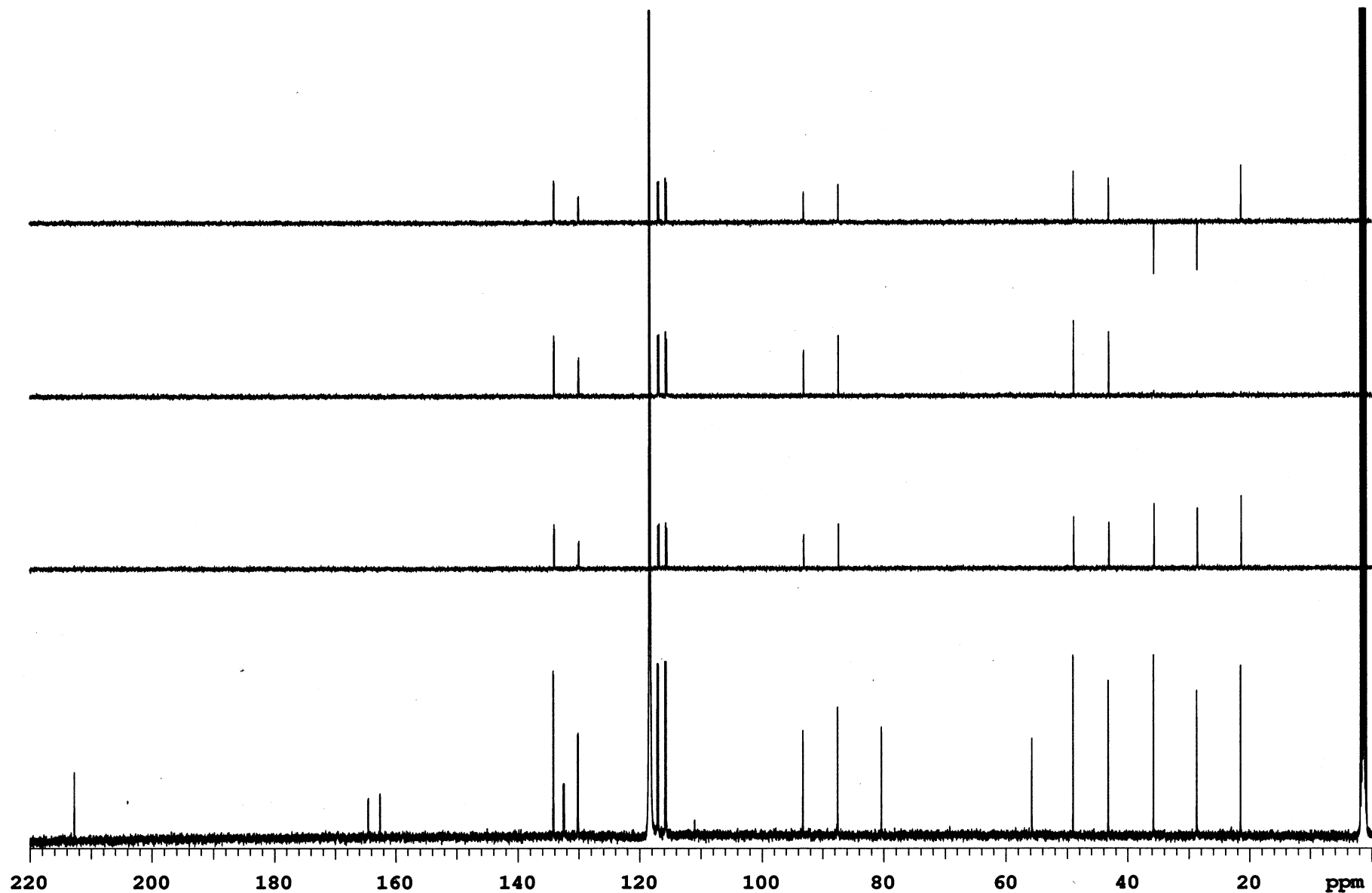


Fig S74. HSQC of compound 5d.

CHP-8e-f2

Sample Name **CHP-8e-f2**
Date collected **2016-05-12**

Pulse sequence **gHSQC**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

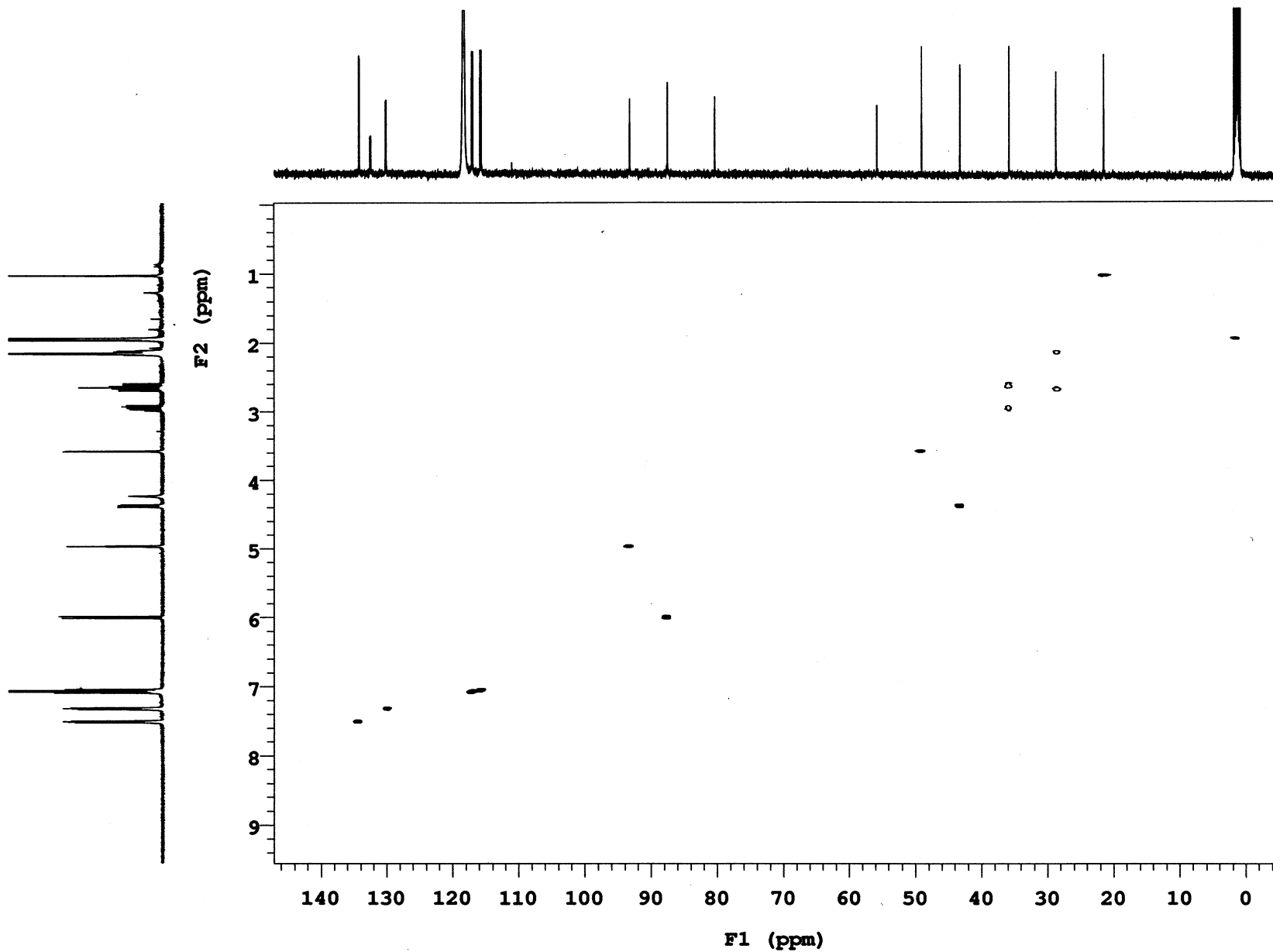


Fig S75. HSQC of compound 5d, expanded.

CHP-8e-f2

Sample Name **CHP-8e-f2**
Date collected **2016-05-12**

Pulse sequence **gHSQC**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

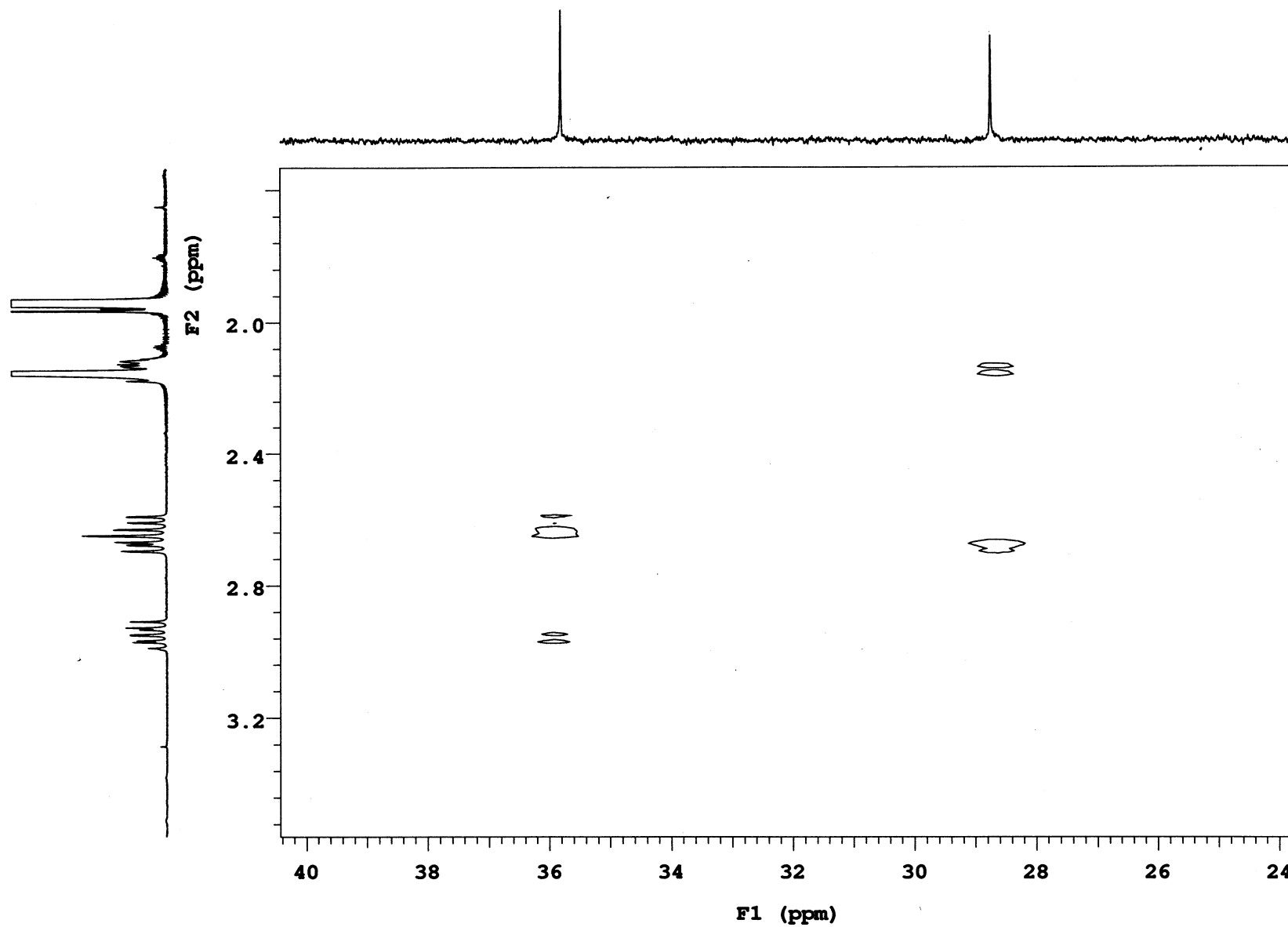


Fig S76. COSY of compound 5d.

CHP-8e-f2

Sample Name **CHP-8e-f2**
Date collected **2016-05-12**

Pulse sequence **gCOSY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

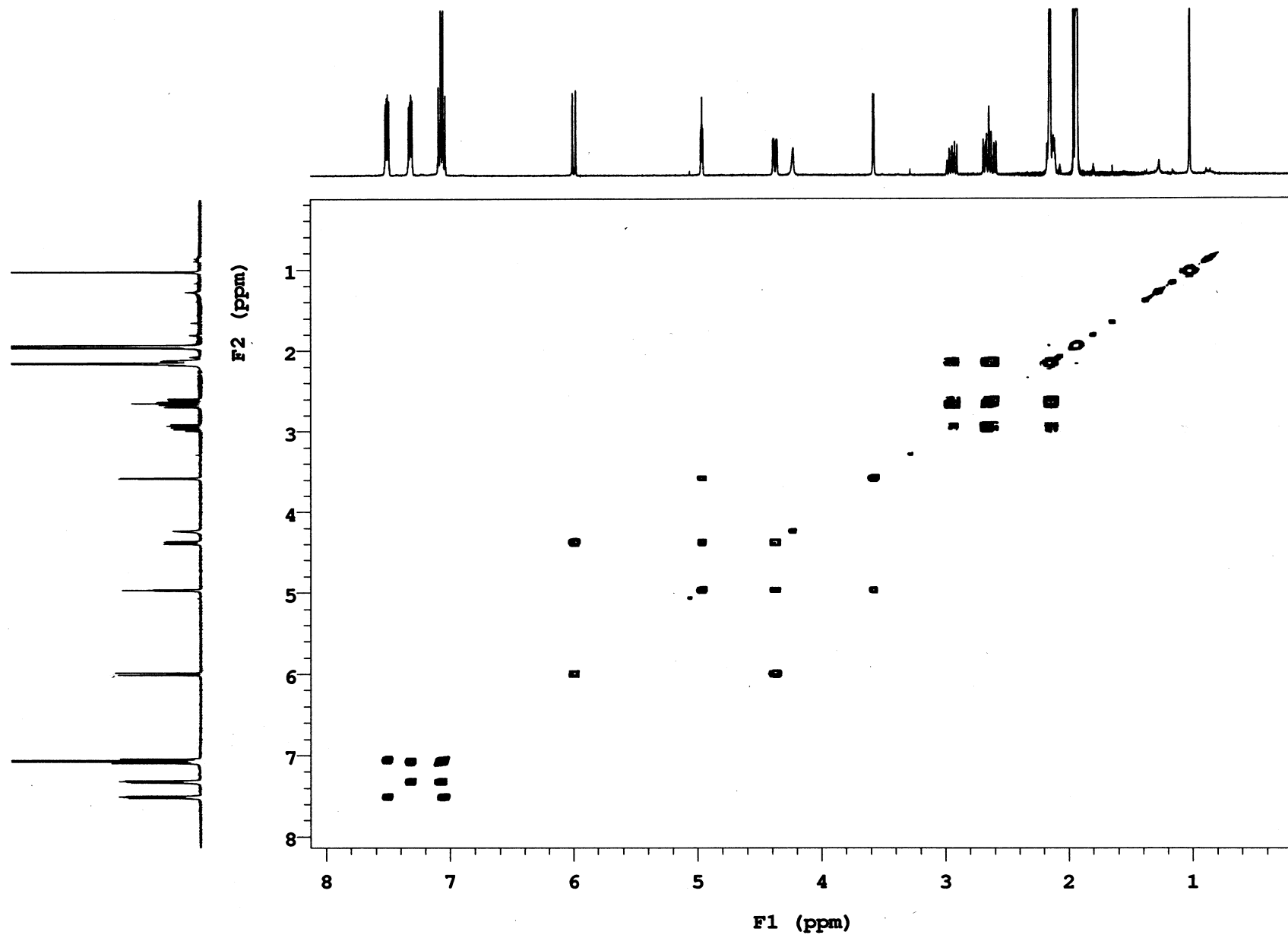


Fig S78. ¹H NMR (CD₃CN, 500 MHz) of compound 3e.

CHP-8o

Sample Name **CHP-8o**
Date collected **2016-05-27**

Pulse sequence **PROTON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

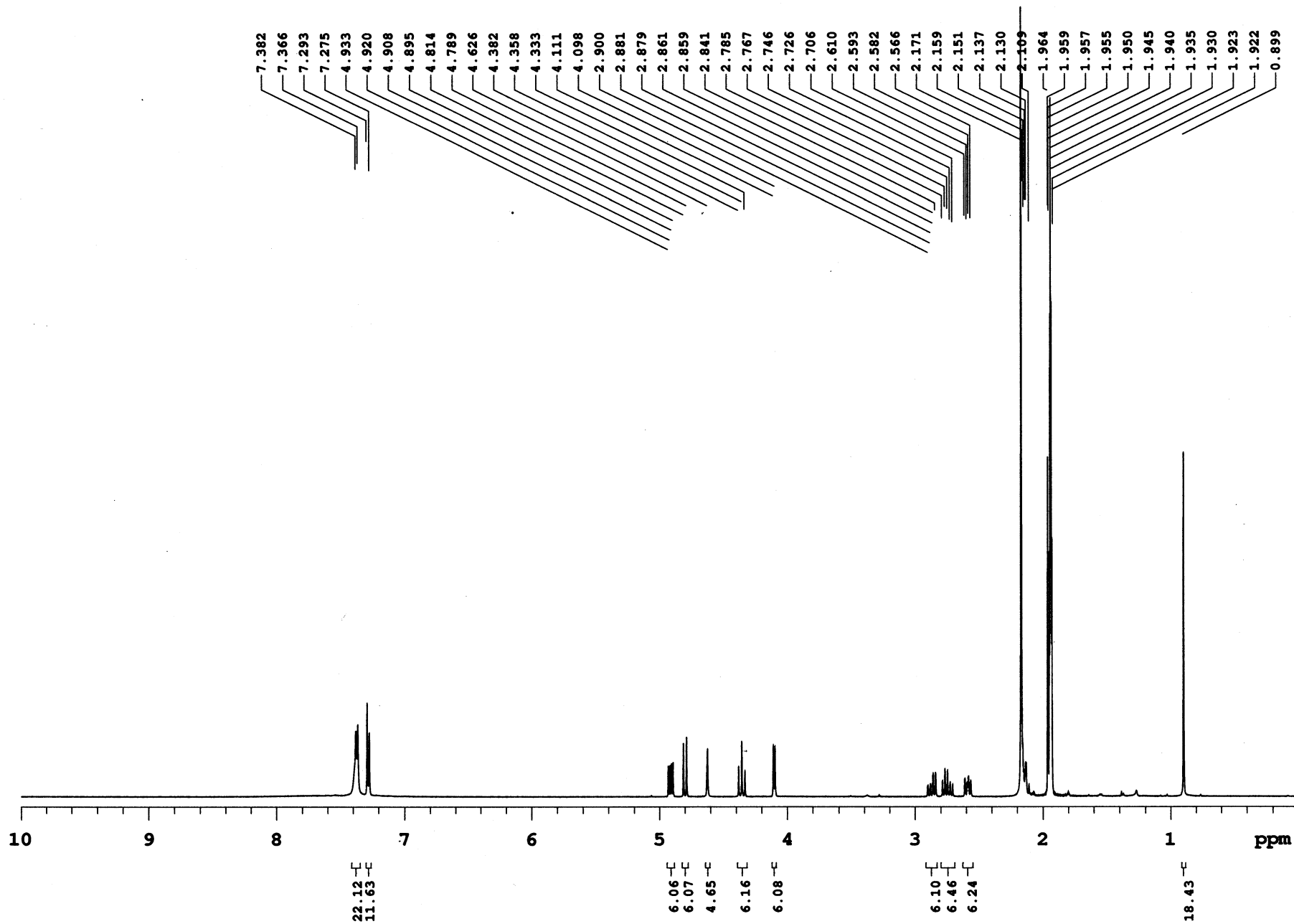


Fig S79. ¹³C NMR (CD₃CN, 125 MHz) of compound 3e.

CHP-80

Sample Name **CHP-80**
Date collected **2016-05-01**

Pulse sequence **CARBON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

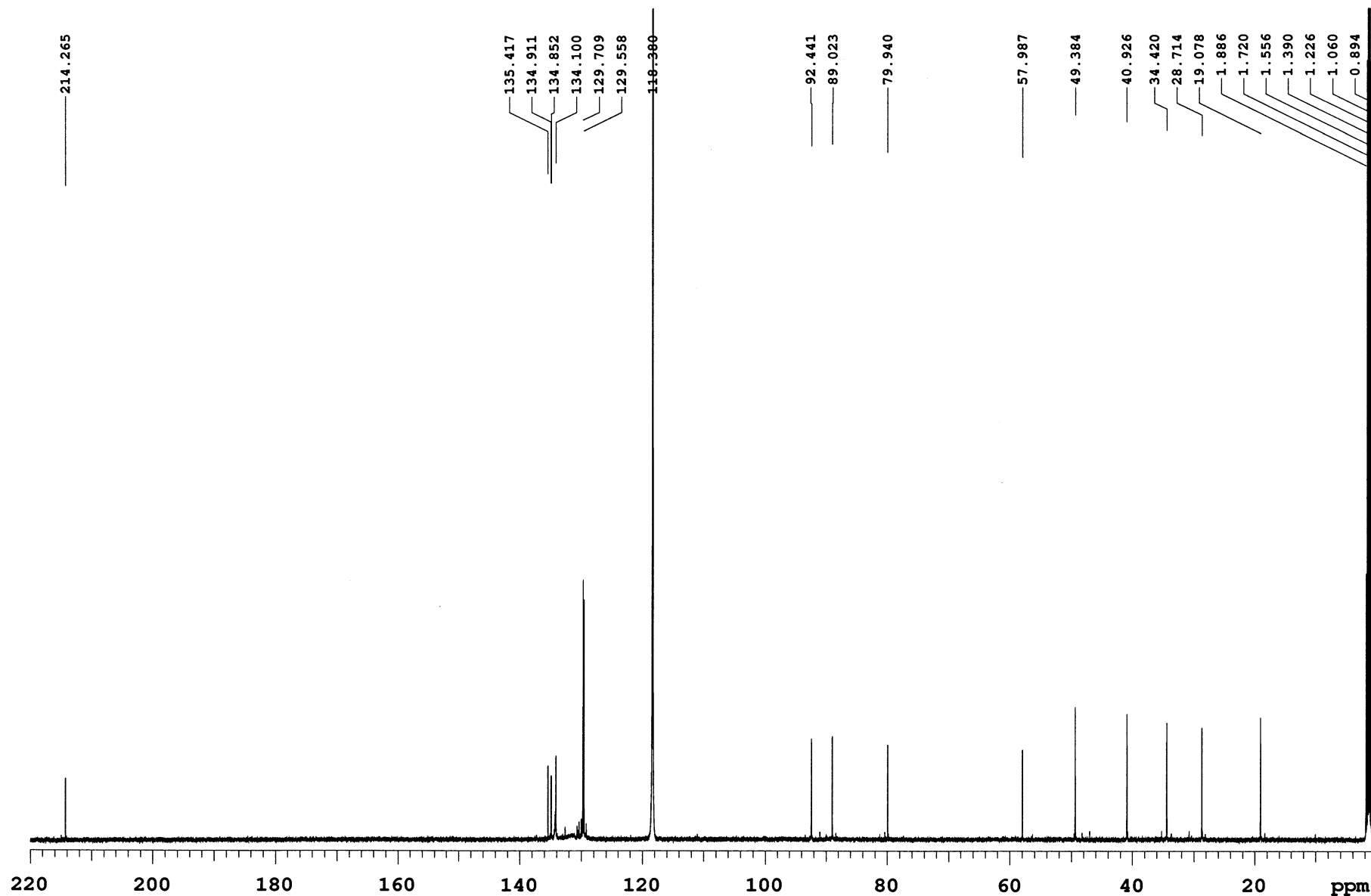


Fig S80. DEPT of compound 3e.

CHP-80

Sample Name **CHP-80**
Date collected **2016-05-01**

Pulse sequence **DEPT**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

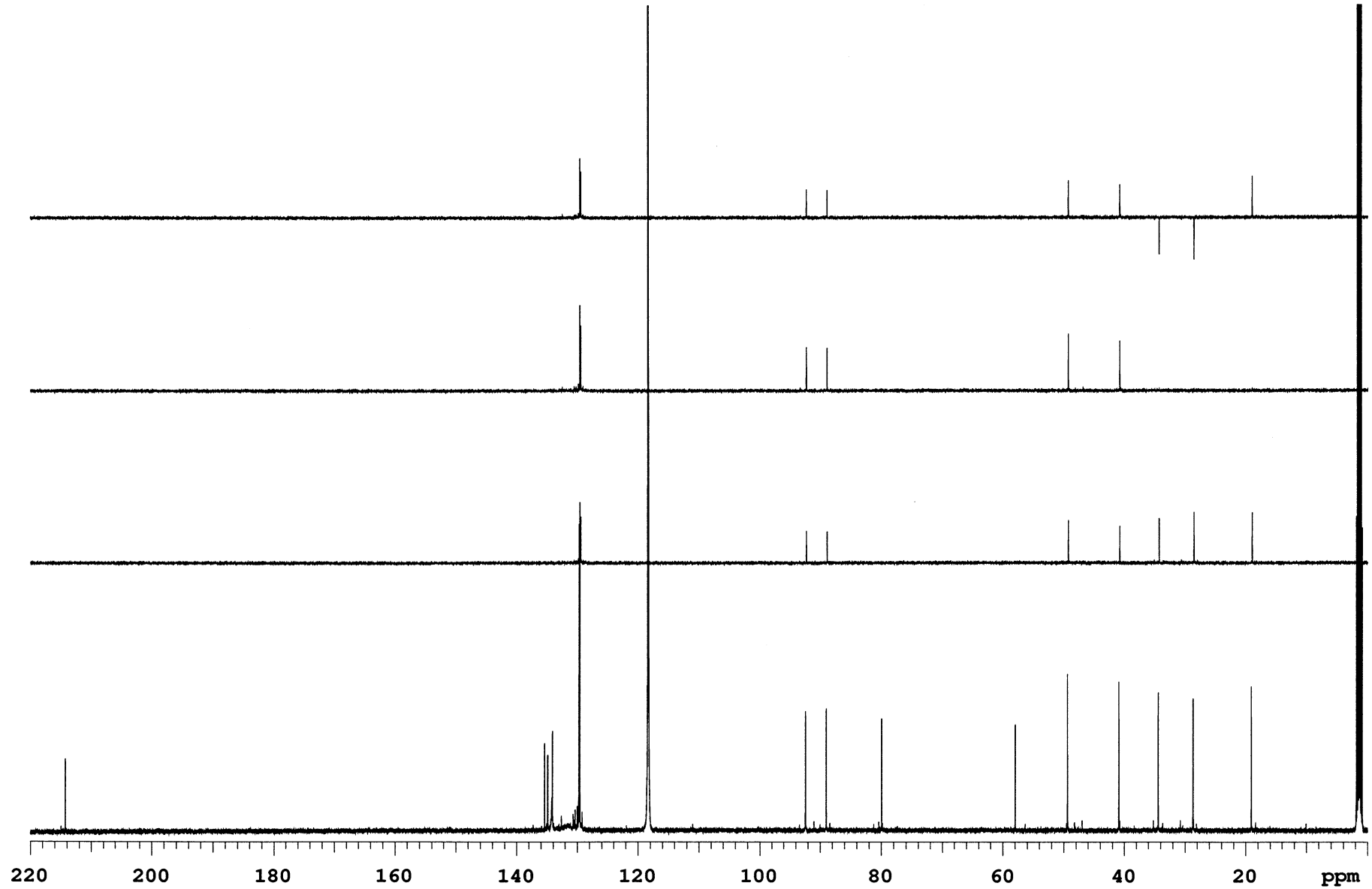


Fig S81. HSQC of compound 3e.

CHP-80

Sample Name **CHP-80**
Date collected **2016-05-01**

Pulse sequence **gHSQC**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

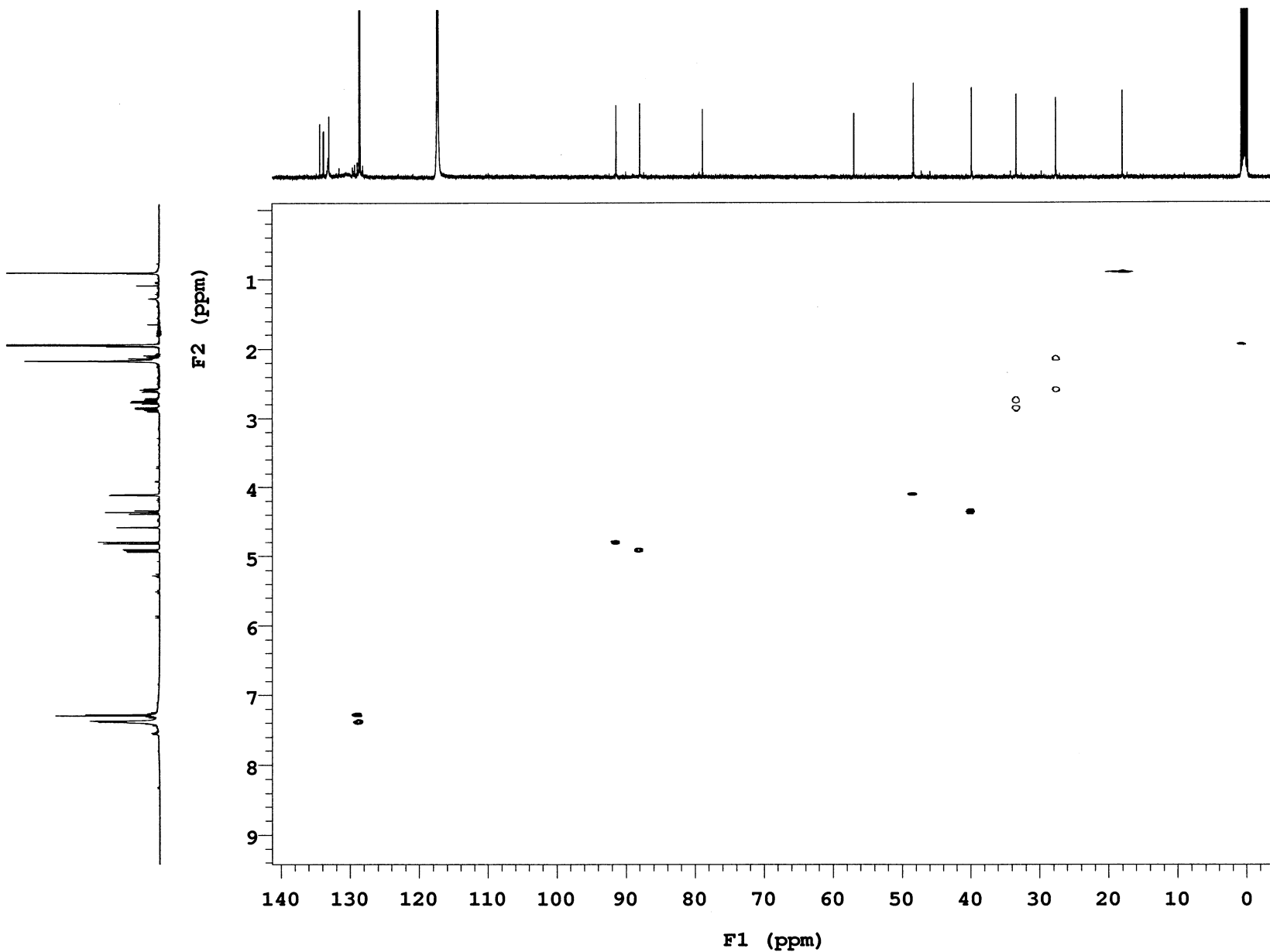


Fig S82. COSY of compound 3e.

CHP-80

Sample Name **CHP-80**
Date collected **2016-05-01**

Pulse sequence **gCOSY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

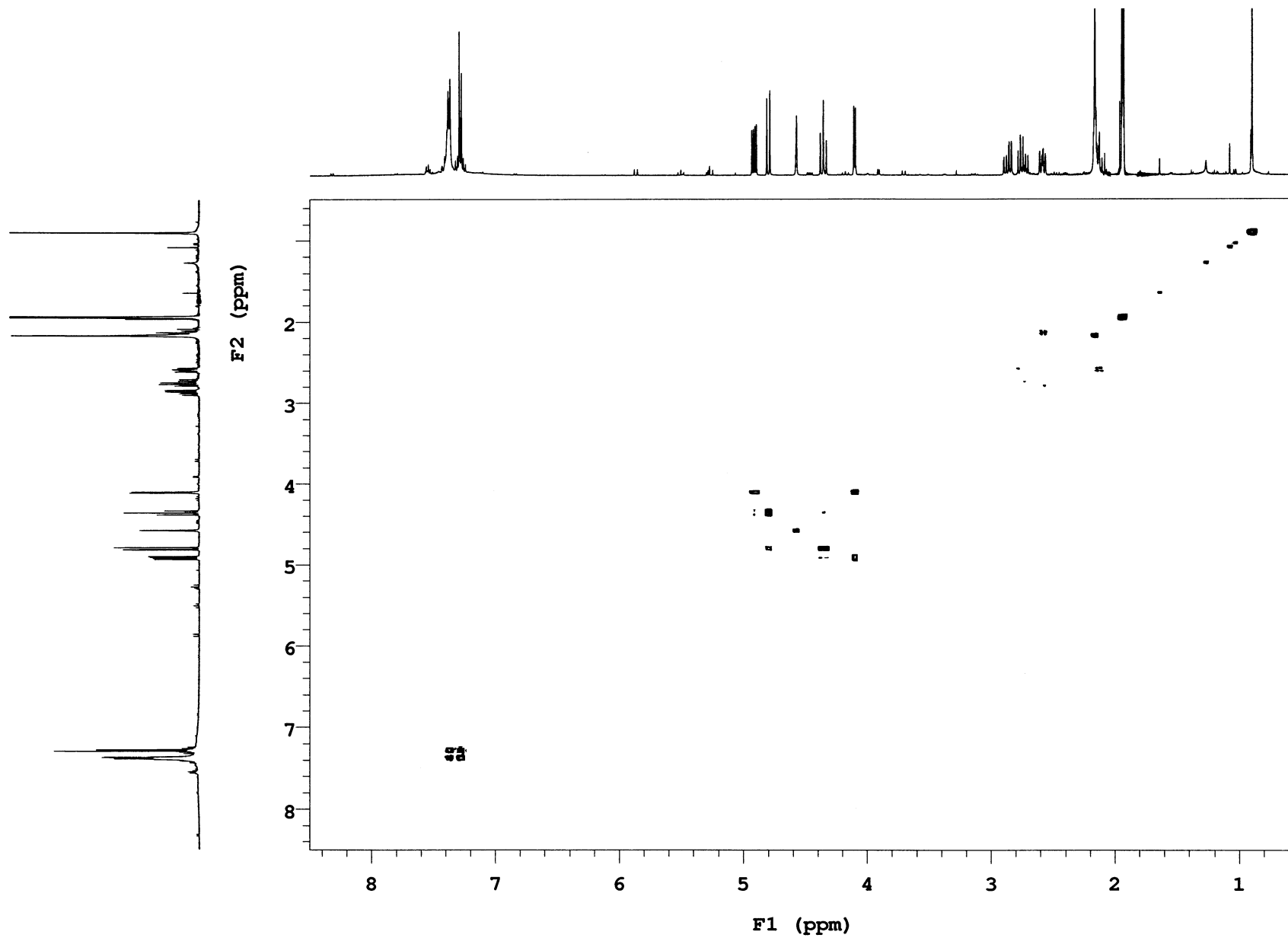


Fig S83. NOESY of compound 3e.

CHP-80

Sample Name **CHP-80**
Date collected **2016-05-01**

Pulse sequence **NOESY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

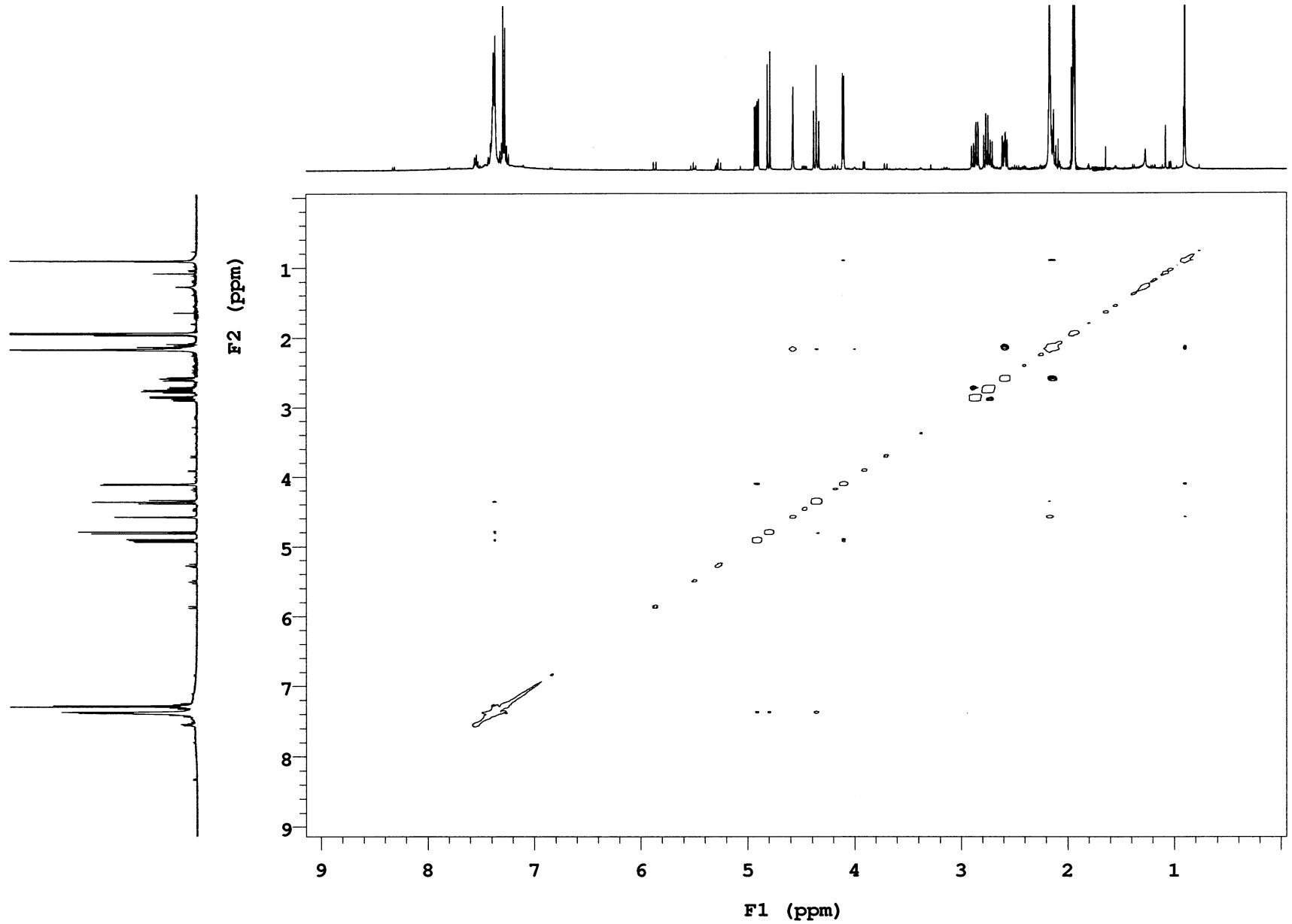


Fig S84. ¹H NMR (CD₃CN, 500 MHz) of compound 5e.

CHP-8o-12

Sample Name	CHP-8o-12	Pulse sequence	PROTON	Temperature	25	Study owner	vnmr2
Date collected	2016-05-22	Solvent	cd3cn	Spectrometer	Agilent-NMR-inova500	Operator	vnmr2

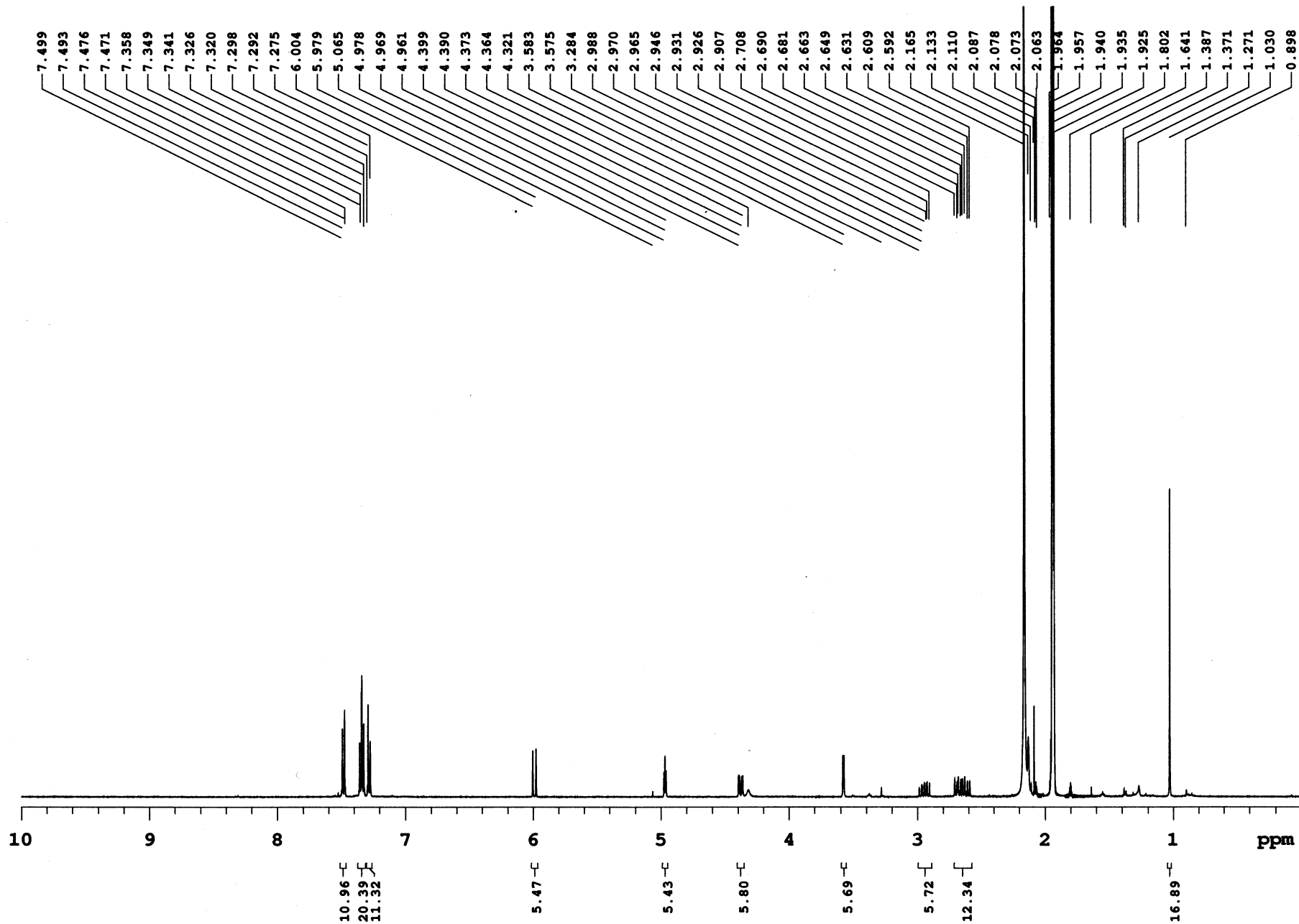


Fig S85. ¹³C NMR (CD₃CN, 125 MHz) of compound 5e.

CHP-8o-f2

Sample Name **CHP-8o-f2**
Date collected **2016-05-22**

Pulse sequence **CARBON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-Inova500**

Study owner **vnmr2**
Operator **vnmr2**

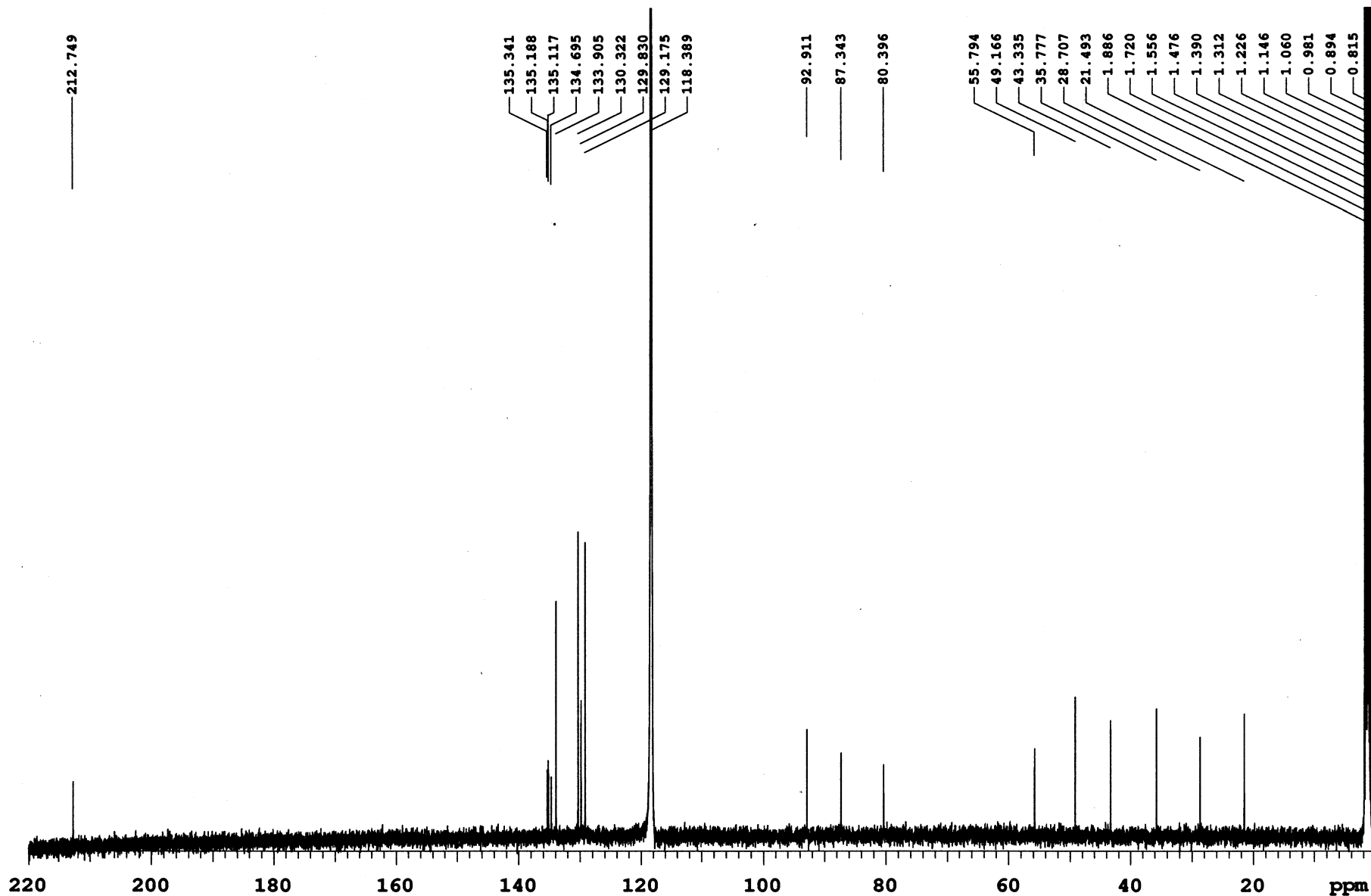


Fig S86. DEPT of compound 5e.

CHP-8o-f2

Sample Name **CHP-8o-f2**
Date collected **2016-05-23**

Pulse sequence **DEPT**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

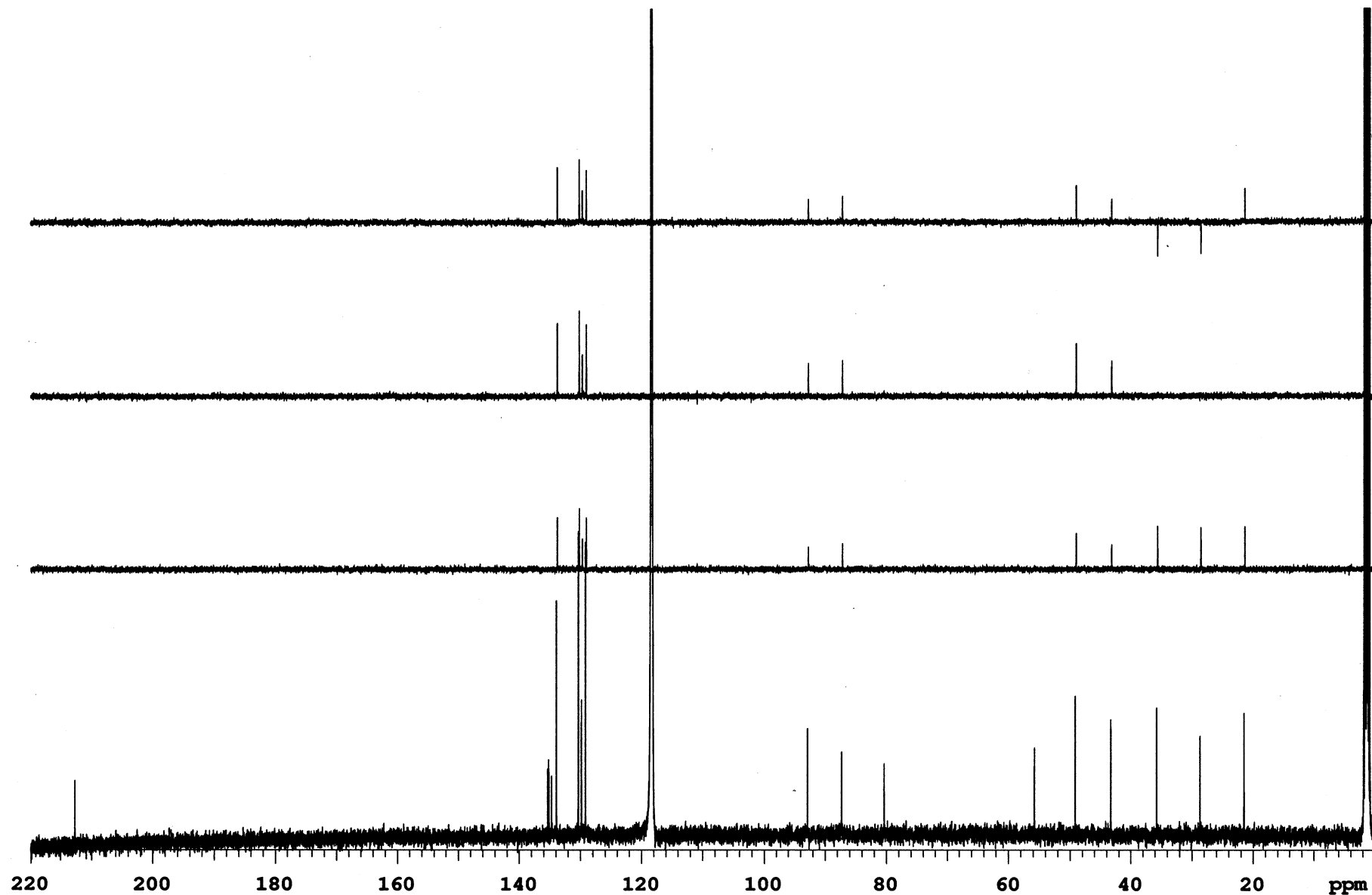


Fig S87. HSQC of compound 5e.

CHP-8o-f2

Sample Name **CHP-8o-f2**
Date collected **2016-05-23**

Pulse sequence **gHSQC**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

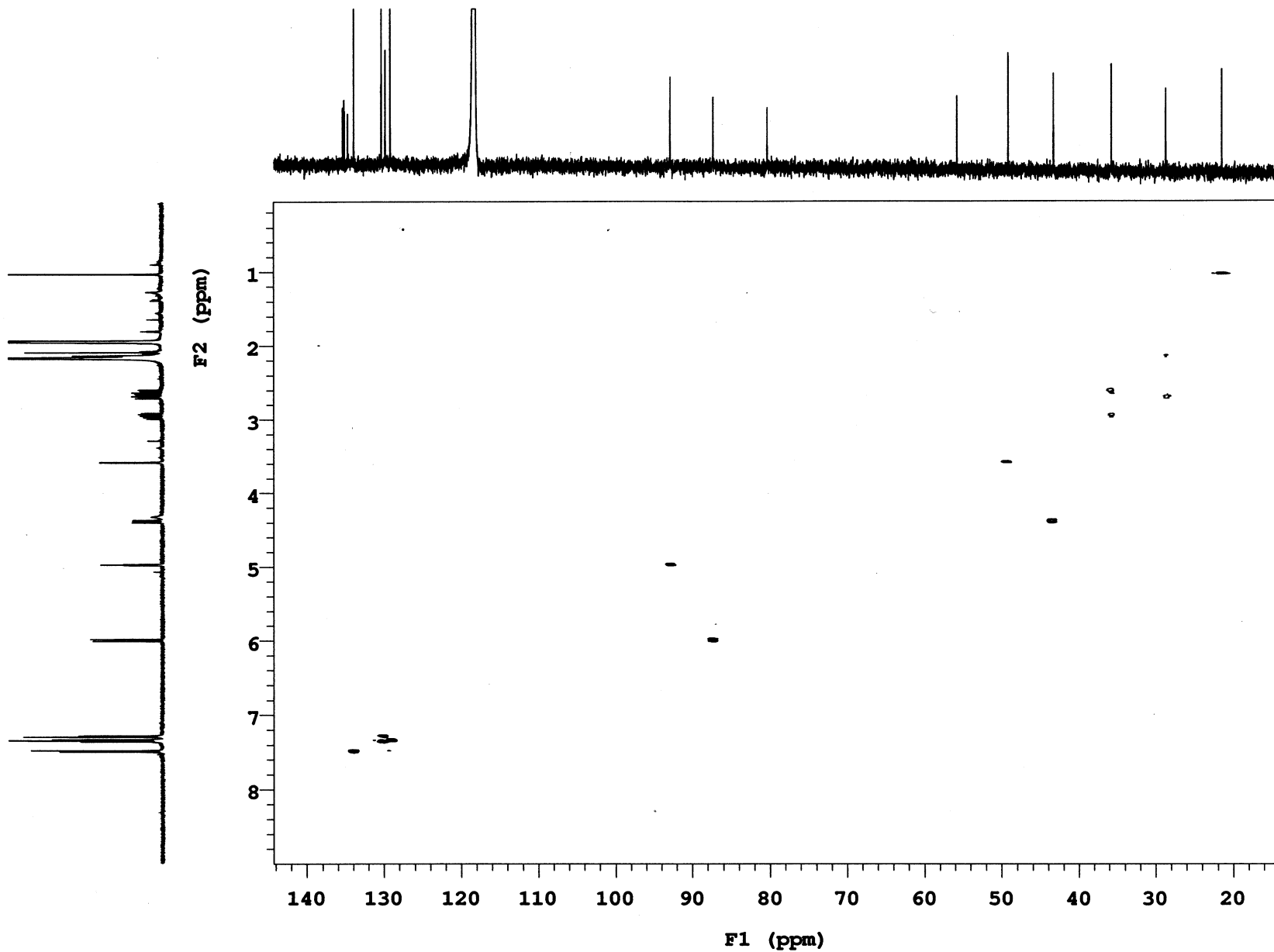


Fig S88. COSY of compound 5e.

CHP-8o-f2

Sample Name **CHP-8o-f2**
Date collected **2016-05-23**

Pulse sequence **gCOSY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

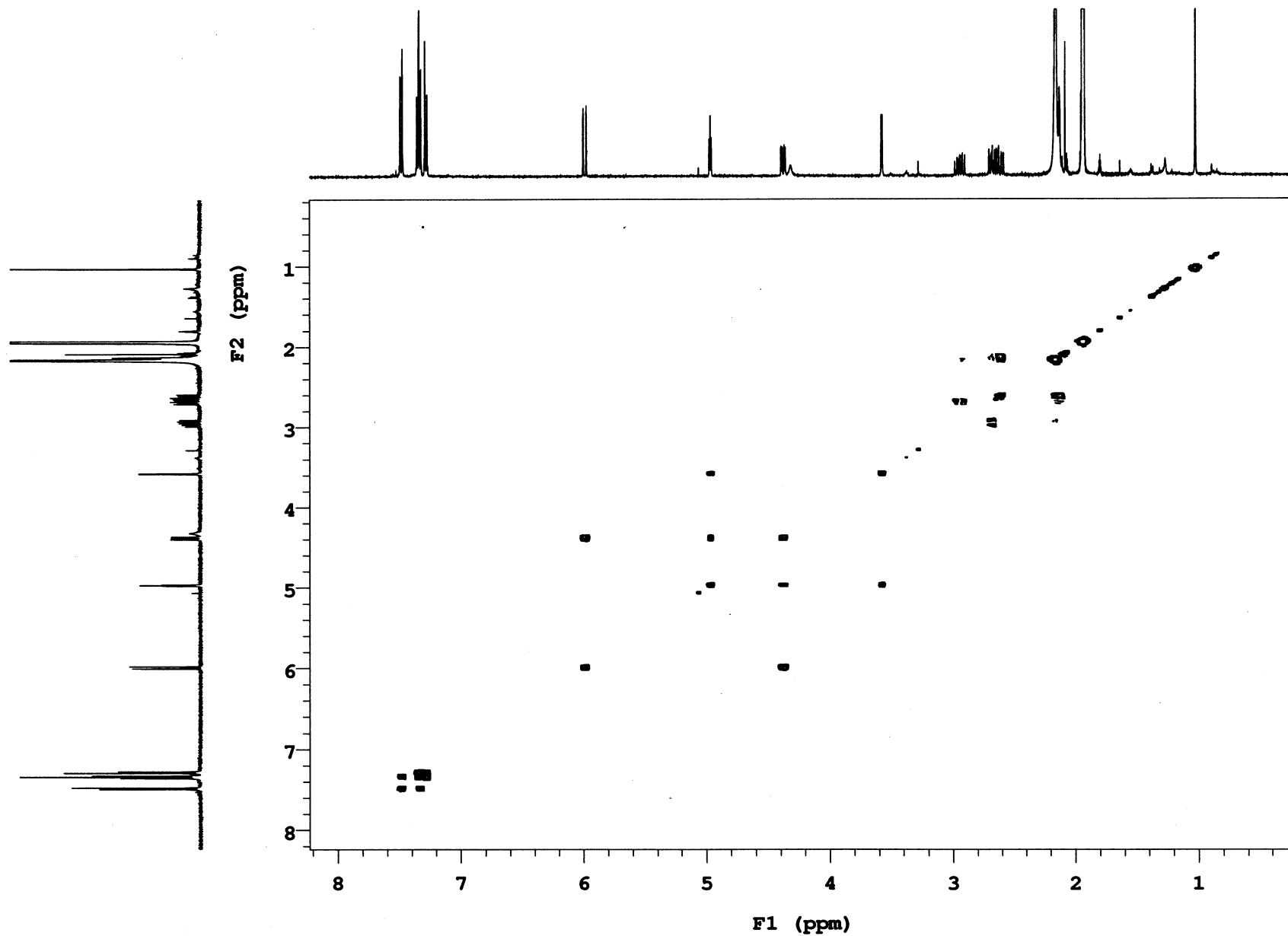


Fig S89. NOESY of compound 5e.

CHP-8o-f2

Sample Name **CHP-8o-f2**
Date collected **2016-05-23**

Pulse sequence **NOESY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

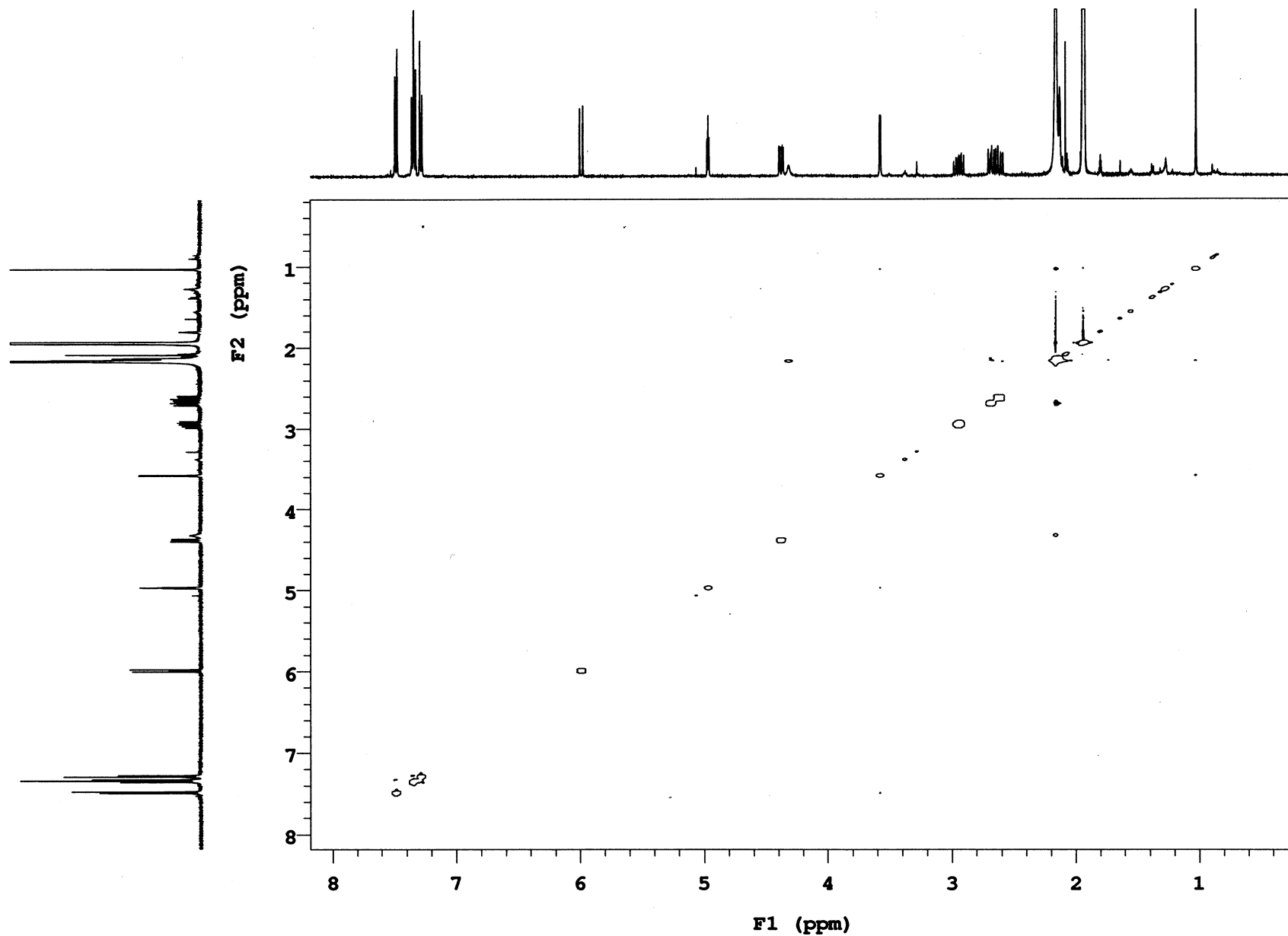


Fig S90. ¹H NMR (CDCl₃, 500 MHz) of compound 3f.

CHP-8d

Sample Name **CHP-8d**
Date collected **2016-05-10**

Pulse sequence **PROTON**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

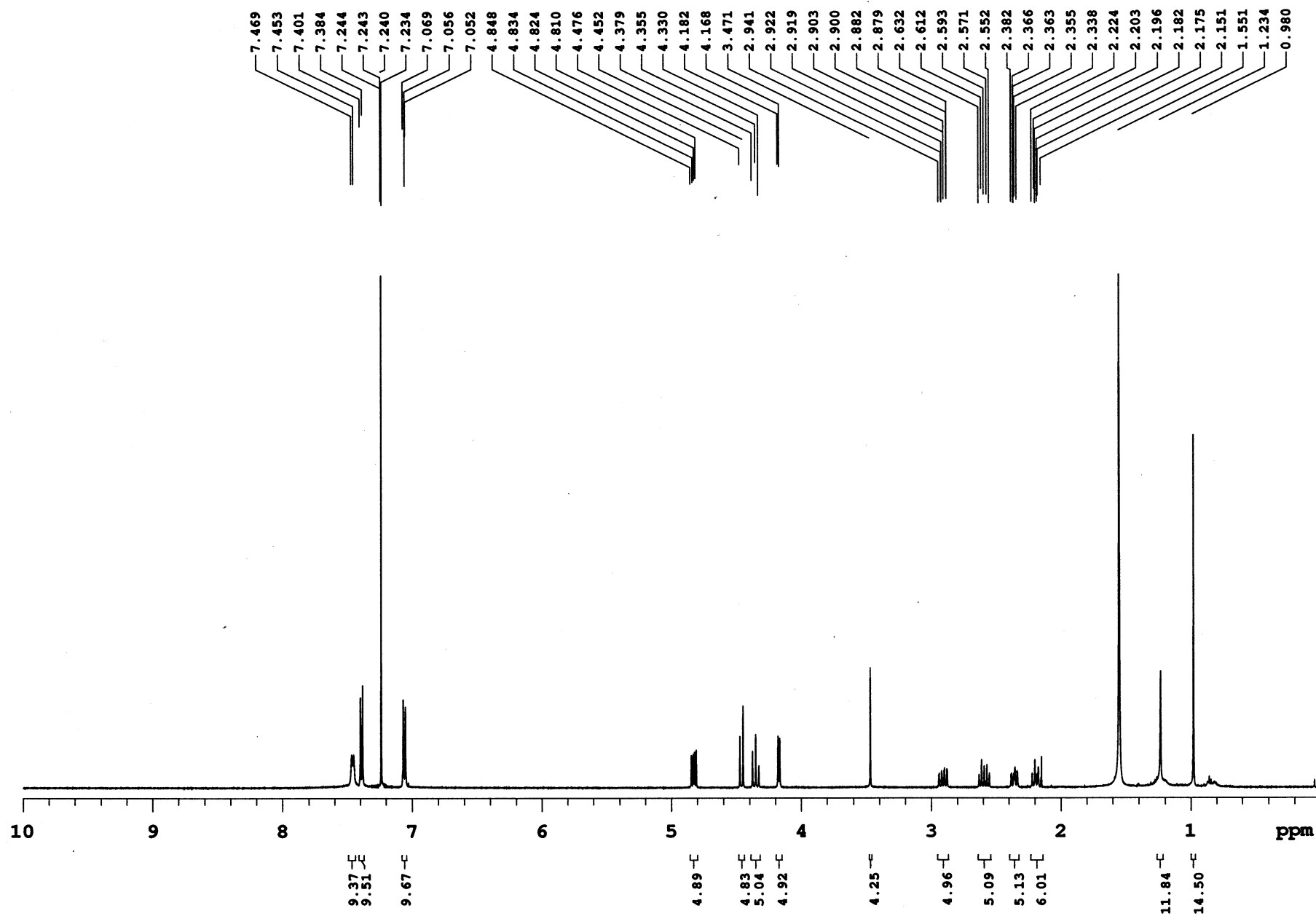


Fig S91. ¹³C NMR (CDCl₃, 125 MHz) of compound 3f.

CHP-8d

Sample Name **CHP-8d** Pulse sequence **CARBON** Temperature **25** Study owner **vnmr2**
Date collected **2016-03-27** Solvent **cdcl3** Spectrometer **Agilent-NMR-inova500** Operator **vnmr2**

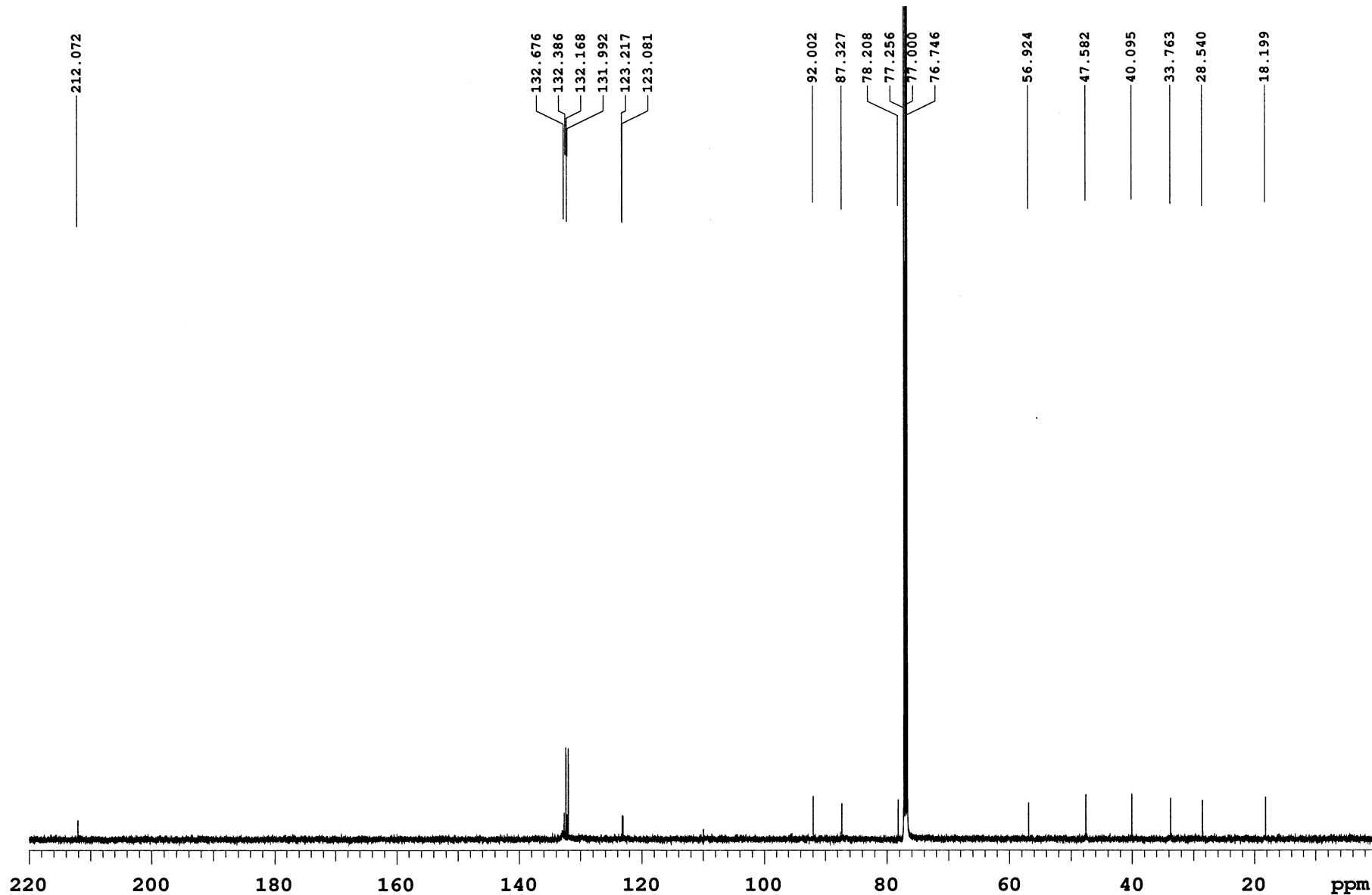


Fig S92. DEPT of compound 3f.

CHP-8d

Sample Name **CHP-8d**
Date collected **2016-03-27**

Pulse sequence **DEPT**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

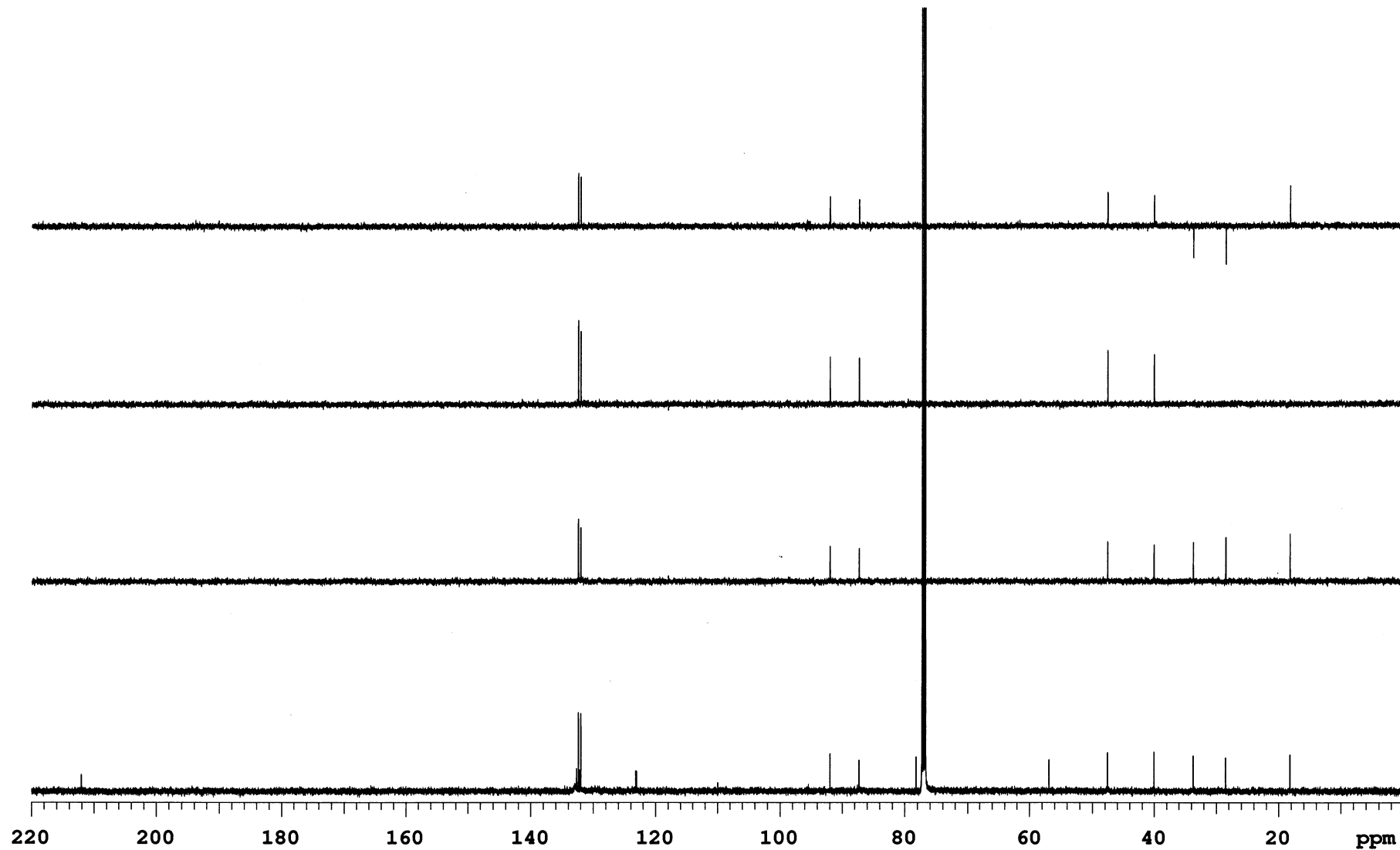


Fig S93. HSQC of compound 3f.

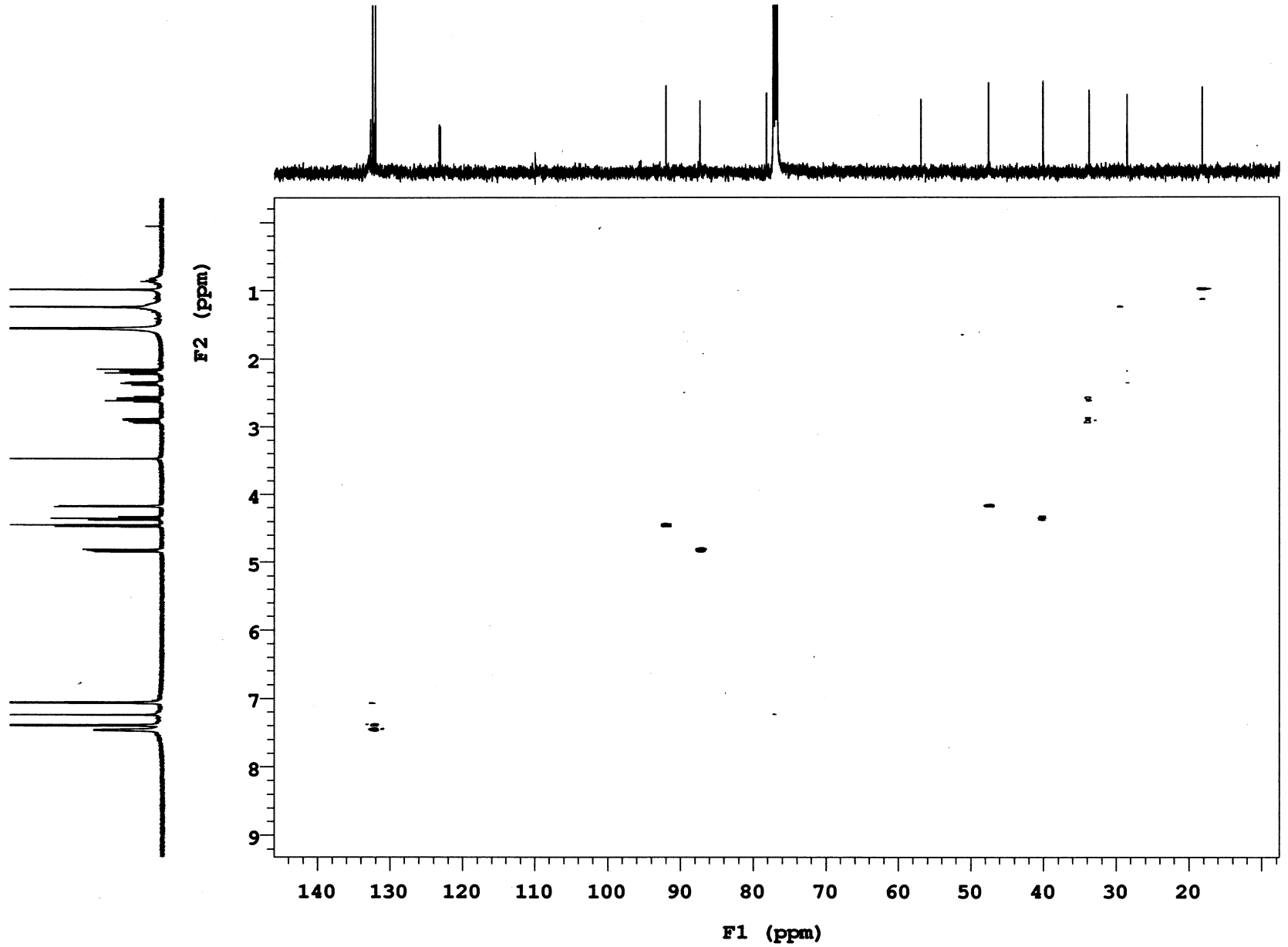
CHP-8d

Sample Name **CHP-8d**
Date collected **2016-05-10**

Pulse sequence **gHSQC**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**



CHP-8d

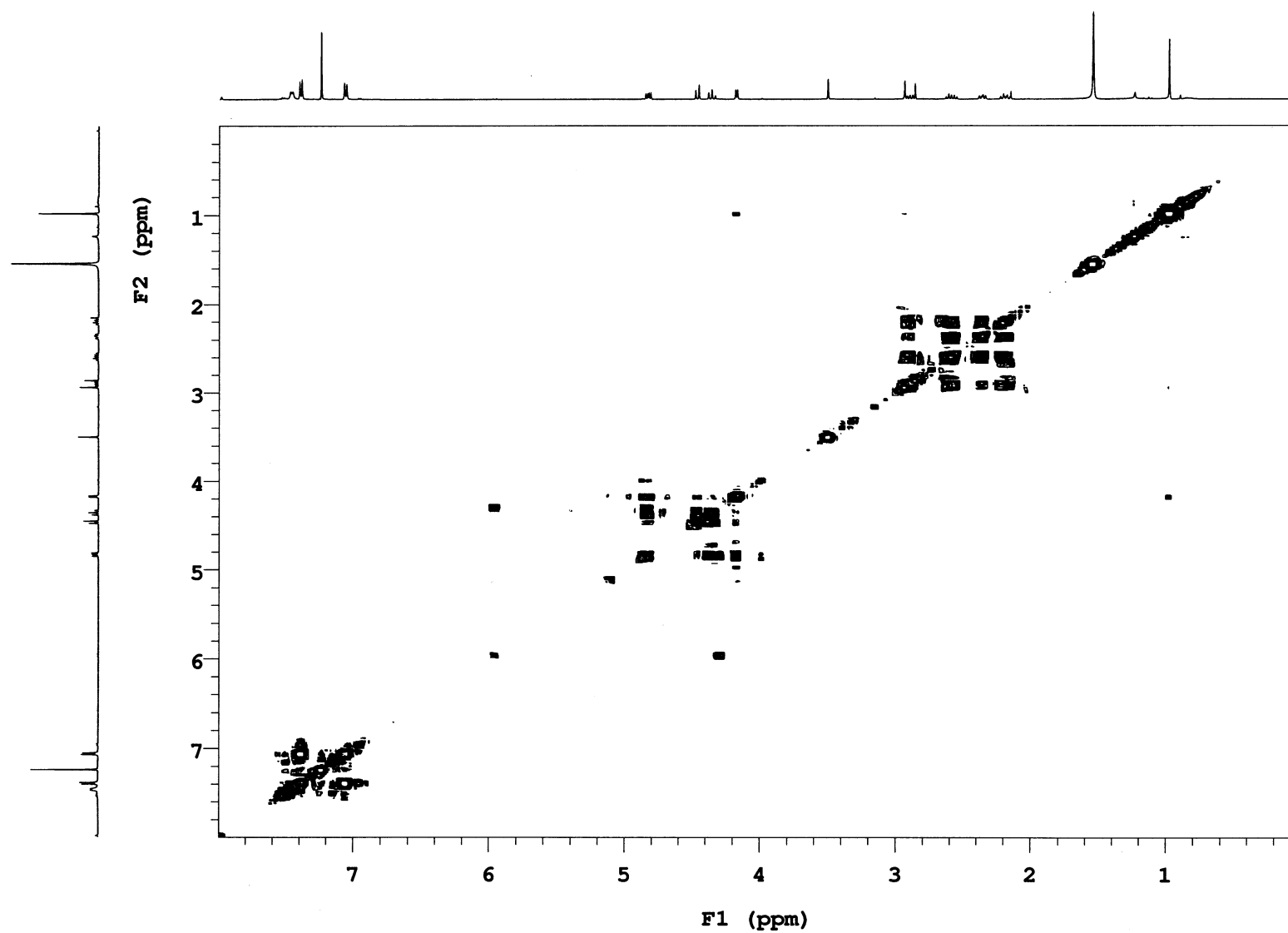
Sample Name **CHP-8d**
Date collected **2016-03-27**Pulse sequence **gCOSY**
Solvent **cdcl3**Temperature **25**
Spectrometer **Agilent-NMR-inova500**Study owner **vnmr2**
Operator **vnmr2**

Fig S96. ¹H NMR (CDCl₃, 500 MHz) of compound 5f.

CHP-8d-f2

Sample Name **CHP-8d-f2**
Date collected **2016-05-19**

Pulse sequence **PROTON**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-Inova500**

Study owner **vnmr2**
Operator **vnmr2**

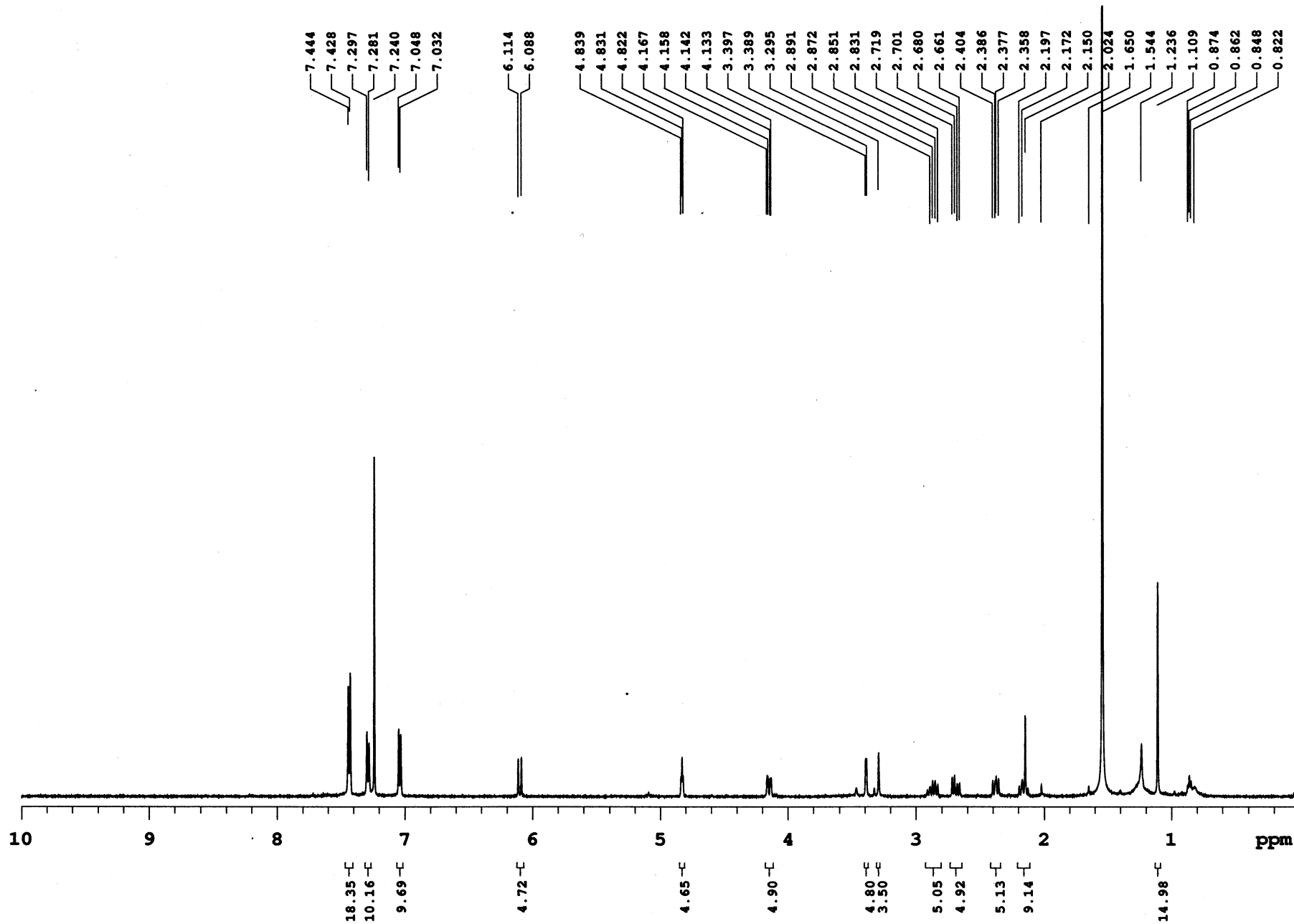


Fig S97 ¹³C NMR (CDCl₃, 125 MHz) of compound 5f.

CHP-8d-I2

Sample Name **CHP-8d-I2**
Date collected **2016-06-03**

Pulse sequence **CARBON**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

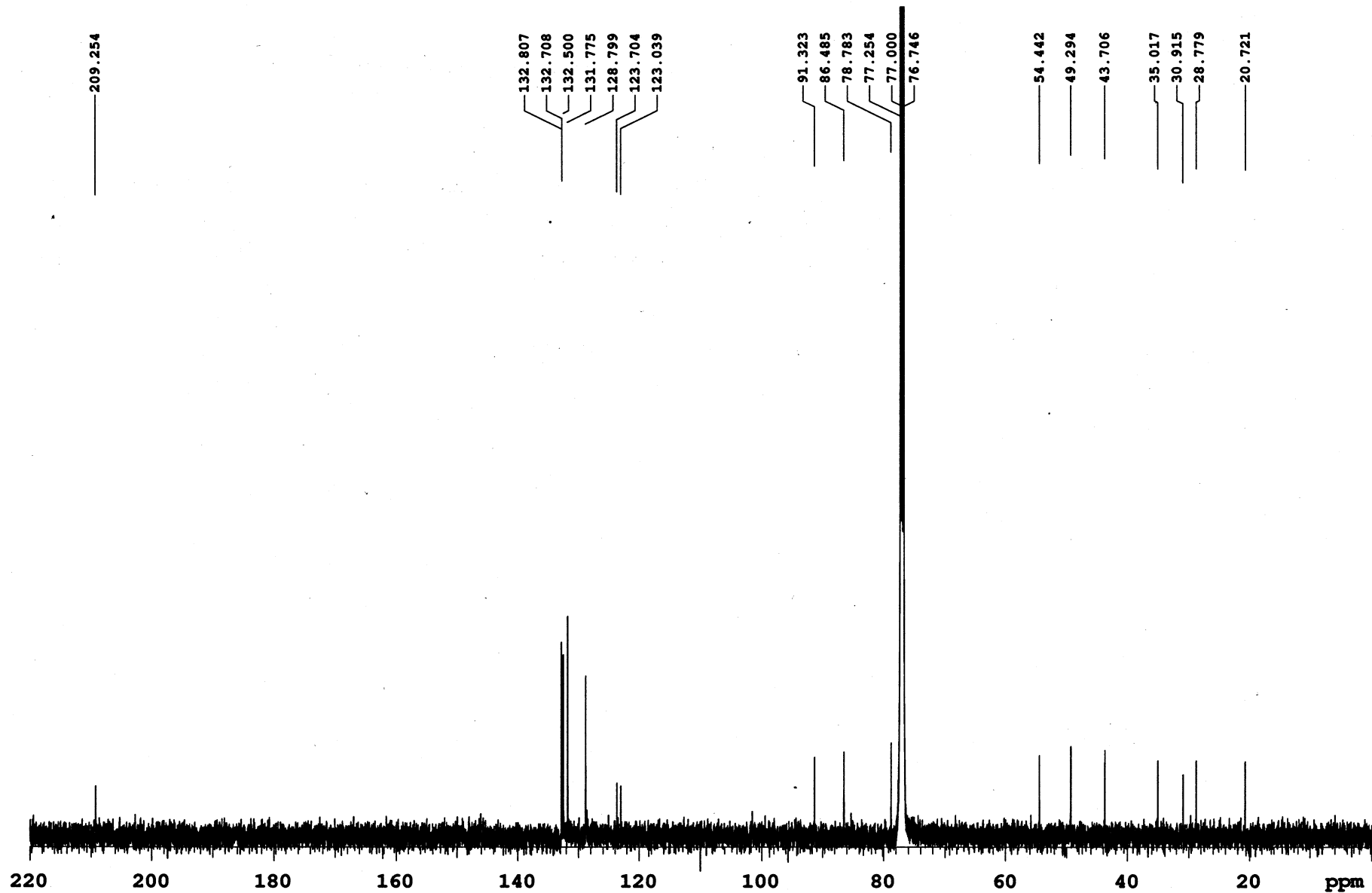


Fig S98. DEPT of compound 5f.

CHP-8d-f2

Sample Name **CHP-8d-f2**
Date collected **2016-06-04**

Pulse sequence **DEPT**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

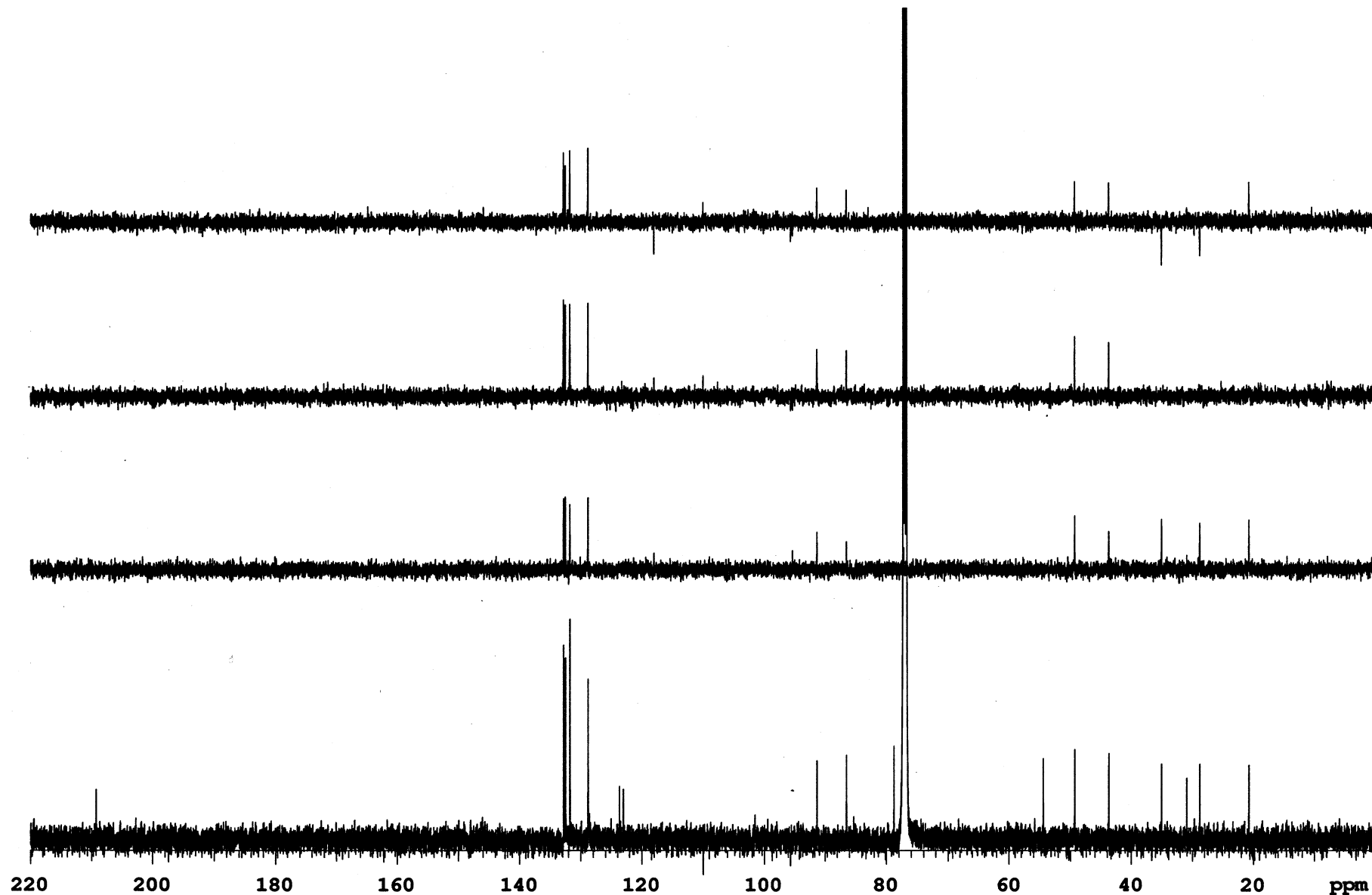


Fig S99. HSQC of compound 5f.

CHP-8d-f2

Sample Name **CHP-8d-f2**
Date collected **2016-06-18**

Pulse sequence **gHSQC**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

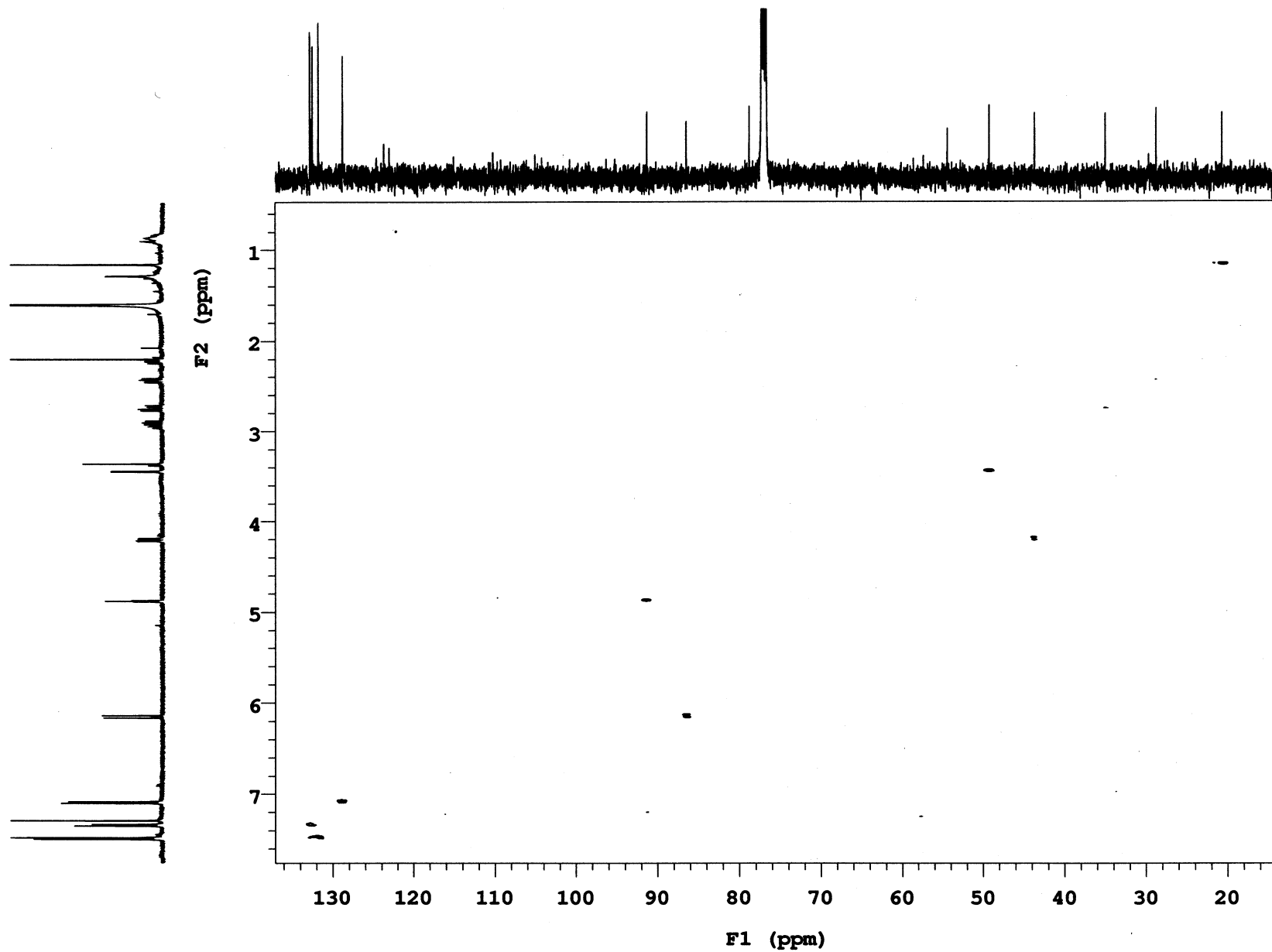


Fig S101. COSY of compound 5f, expanded.

S101

CHP-8d-12

Sample Name **CHP-8d-12**
Date collected **2016-05-19**

Pulse sequence **gCOSY**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

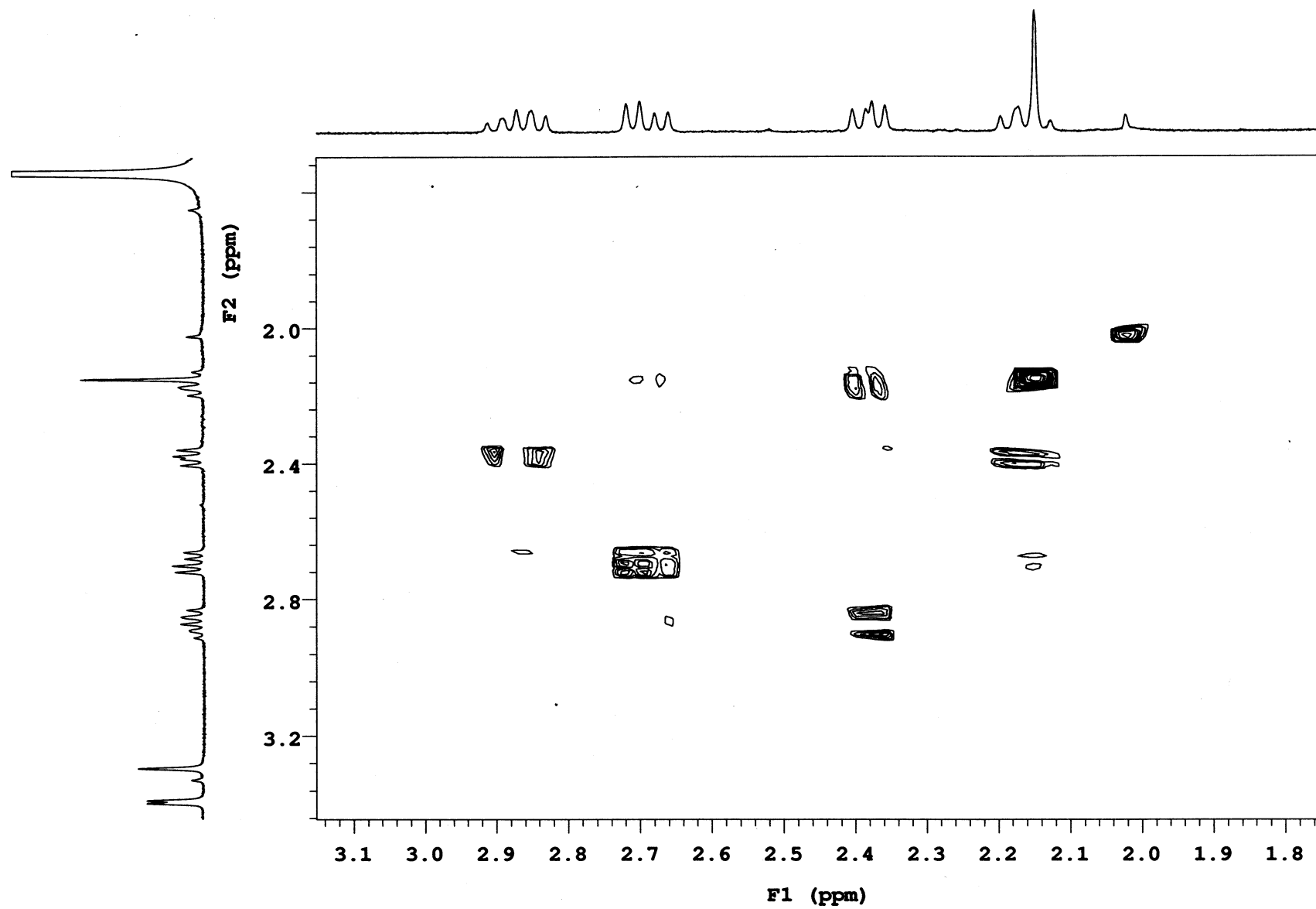


Fig S102. NOESY of compound 5f.

CHP-8d-f2

Sample Name **CHP-8d-f2**
Date collected **2016-05-19**

Pulse sequence **NOESY**
Solvent **cdcl3**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

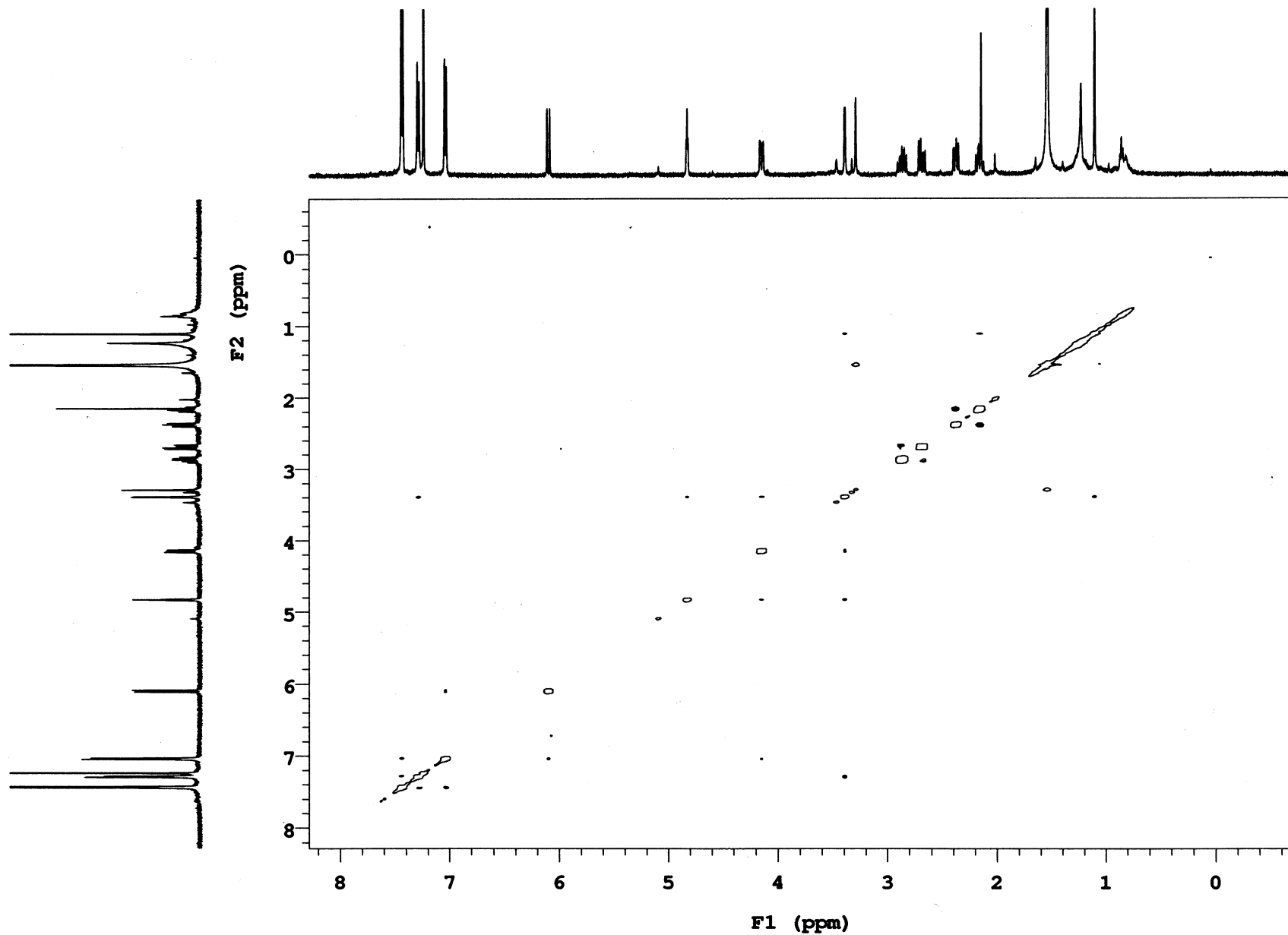


Fig S103. ¹H NMR (acetone-d₆, 500 MHz) of compound 3g.

CHP-081

Sample Name **CHP-081**
Date collected **2016-03-05**

Pulse sequence **PROTON**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner: **vnmr2**
Operator **vnmr2**

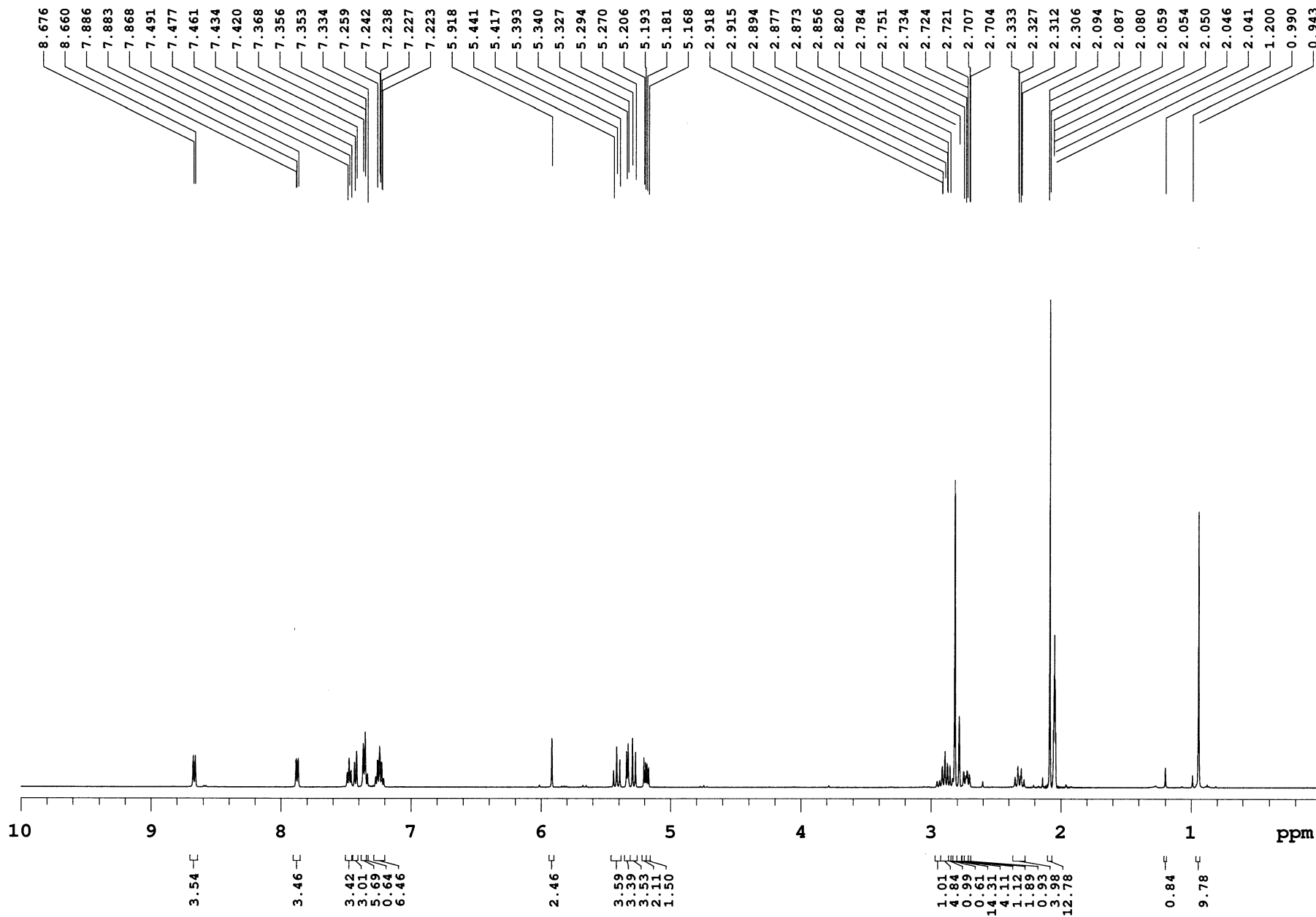


Fig S104. ¹³C NMR (acetone-d₆, 125 MHz) of compound 3g.

CHP-08I

Sample Name **CHP-08I**
Date collected **2016-03-05**

Pulse sequence **CARBON**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

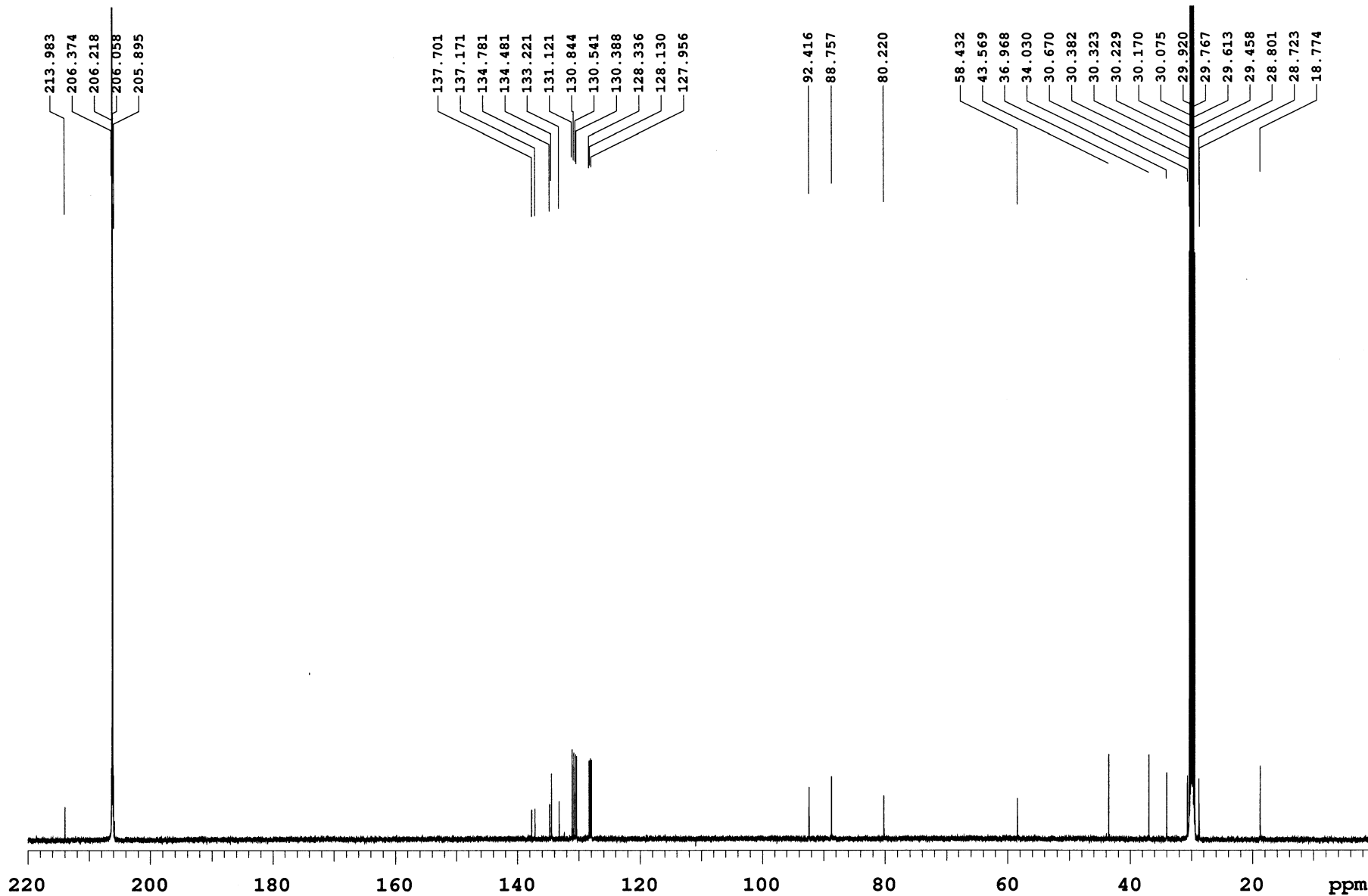


Fig S105. DEPT of compound 3g.

CHP-081

Sample Name **CHP-081**
Date collected **2016-03-05**

Pulse sequence **DEPT**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

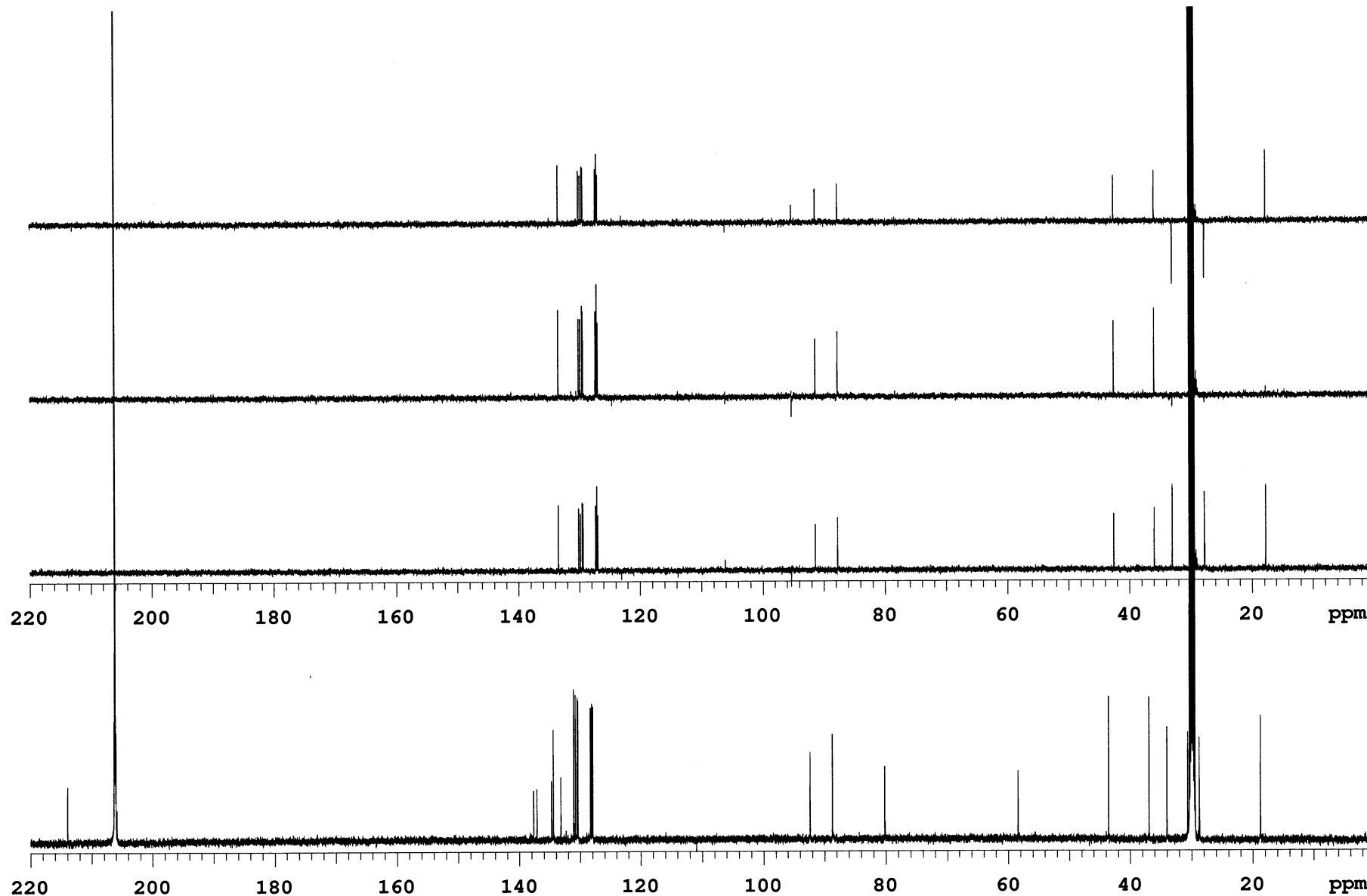


Fig S106. HSQC of compound 3g.

S106

CHP-081

Sample Name **CHP-081**
Date collected **2016-03-05**

Pulse sequence **gHSQC**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

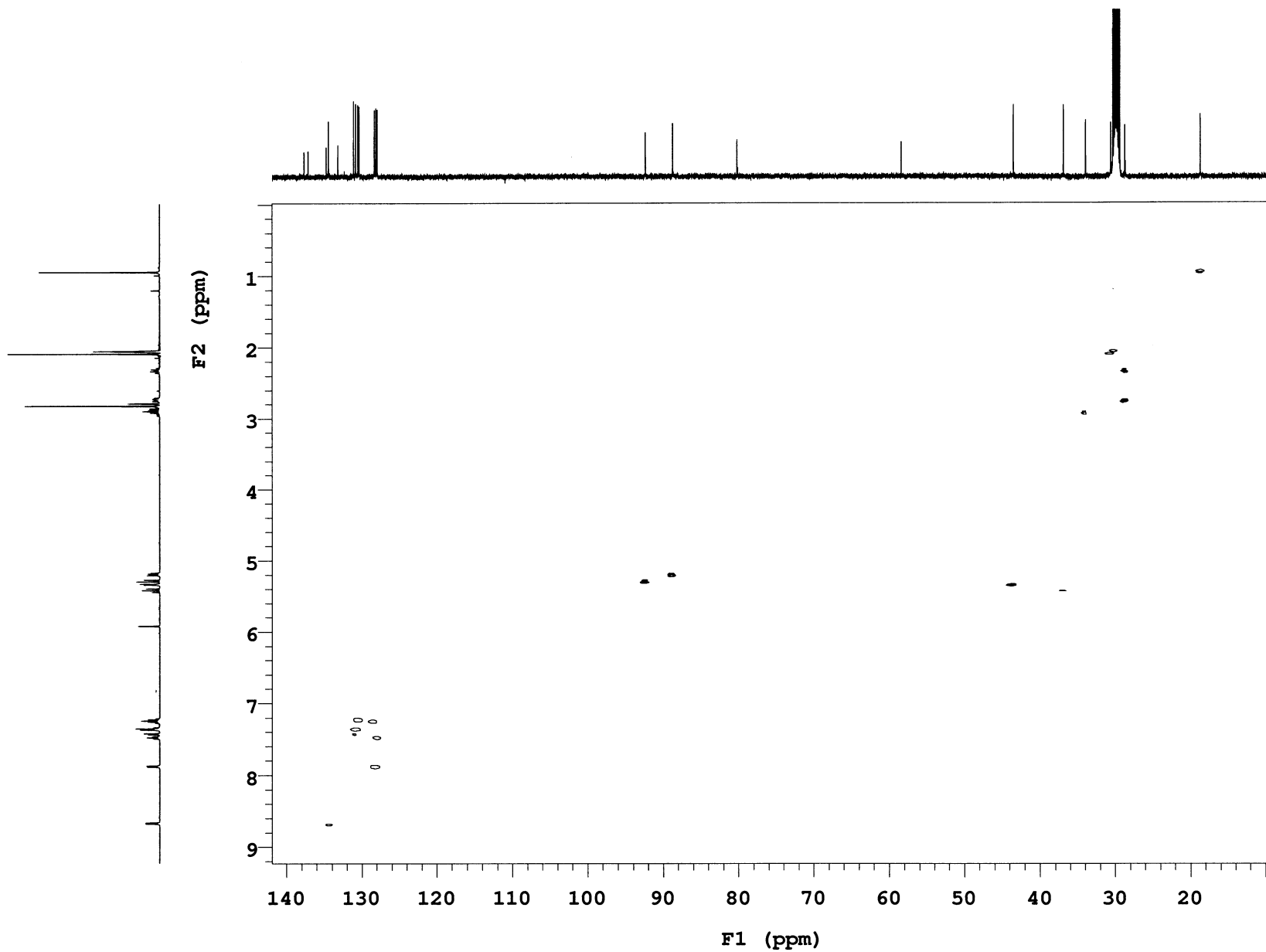


Fig S107. COSY of compound 3g.

CHP-08I

Sample Name **CHP-08I**
Date collected **2016-03-05**

Pulse sequence **gCOSY**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

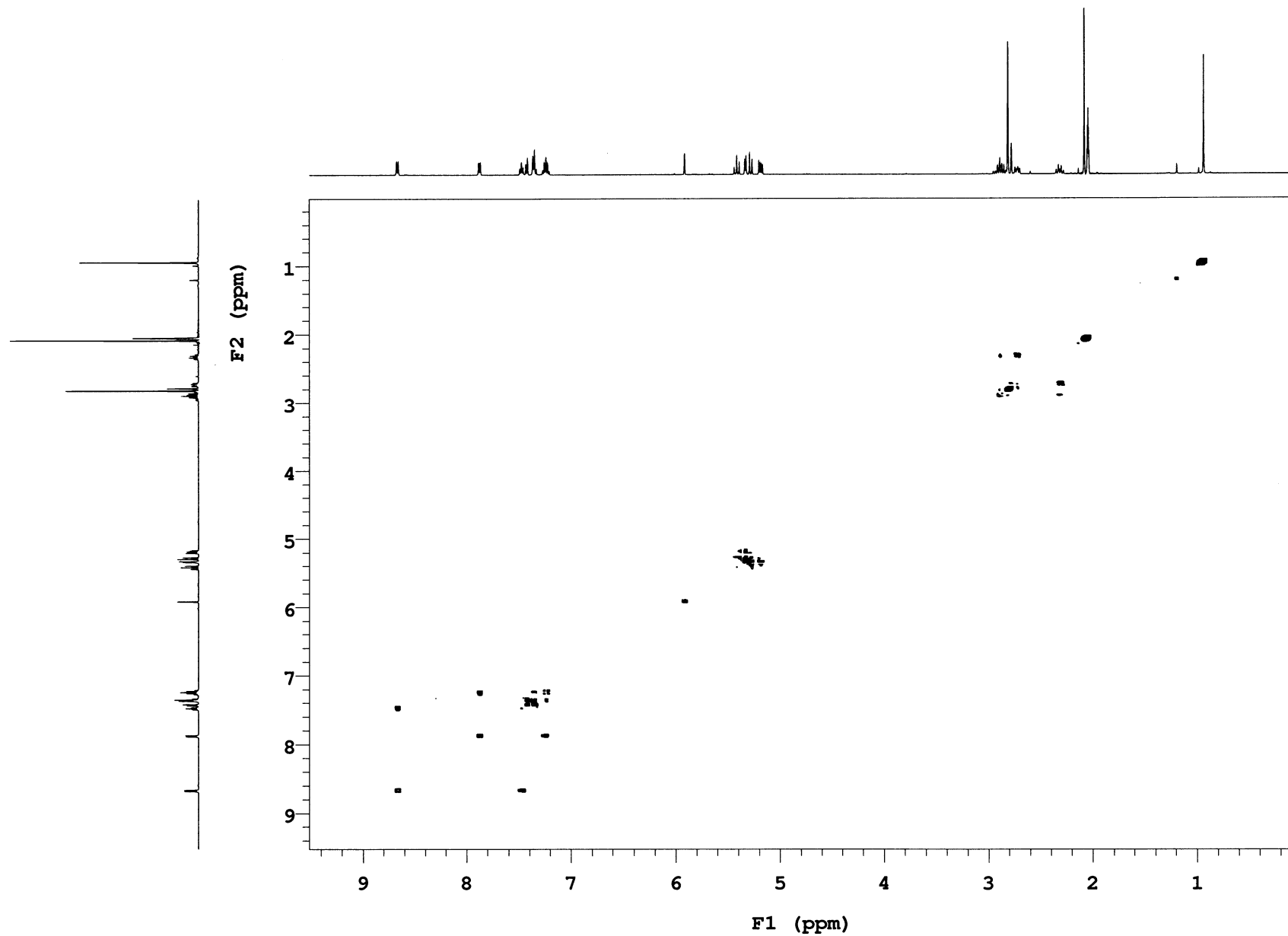


Fig S108. NOESY of compound 3g.

CHP-081

Sample Name **CHP-081**
Date collected **2016-03-05**

Pulse sequence **NOESY**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

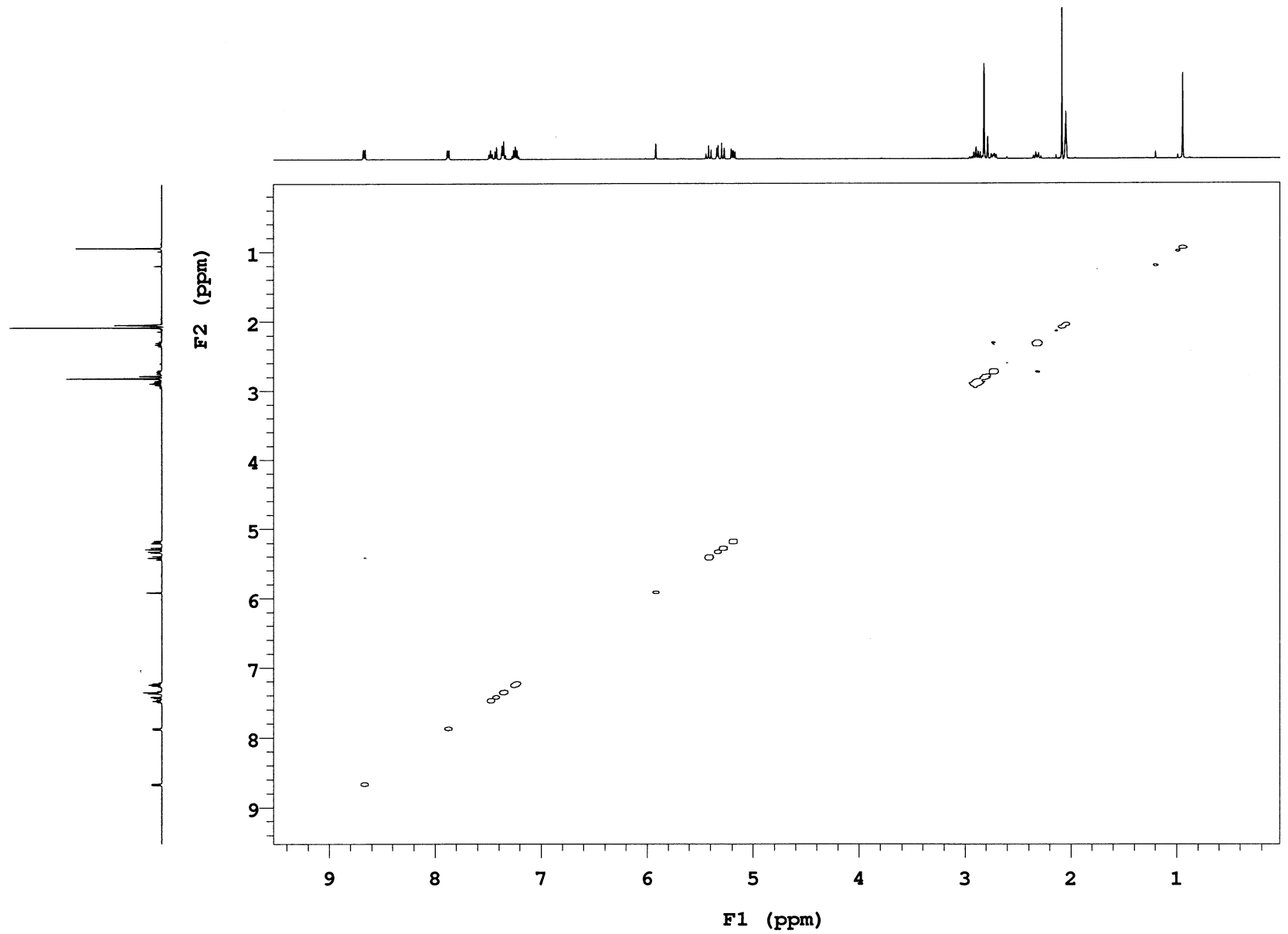


Fig S109. ¹H NMR (acetone-d₆, 500 MHz) of compound 5g.

CHP-81-f1

Sample Name **CHP-81-f1**
Date collected **2016-05-27**

Pulse sequence **PROTON**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

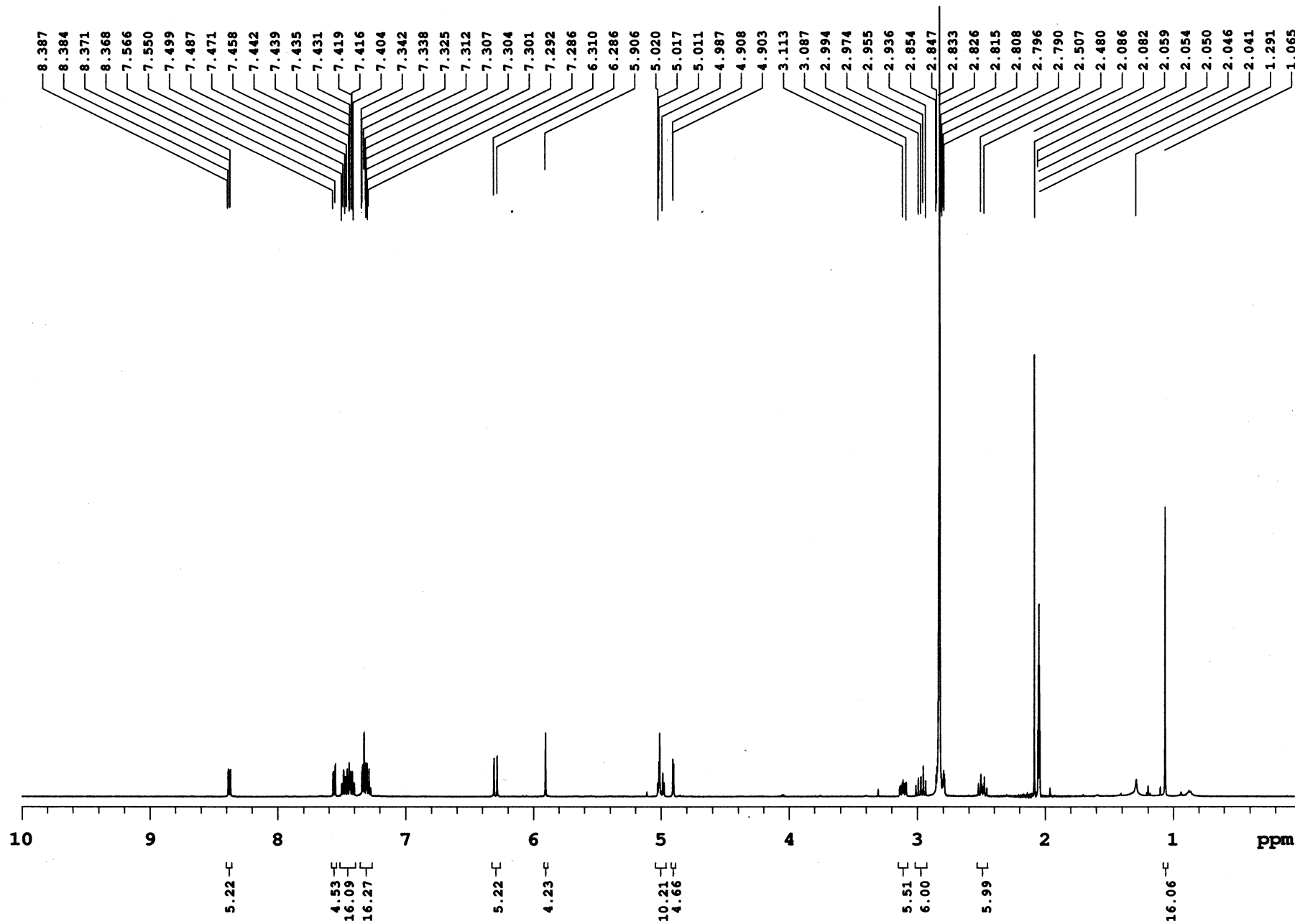


Fig S110. ¹³C NMR (acetone-d₆, 125 MHz) of compound 5g.

CHP-81-11

Sample Name **CHP-81-11**
Date collected **2016-05-27**

Pulse sequence **CARBON**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

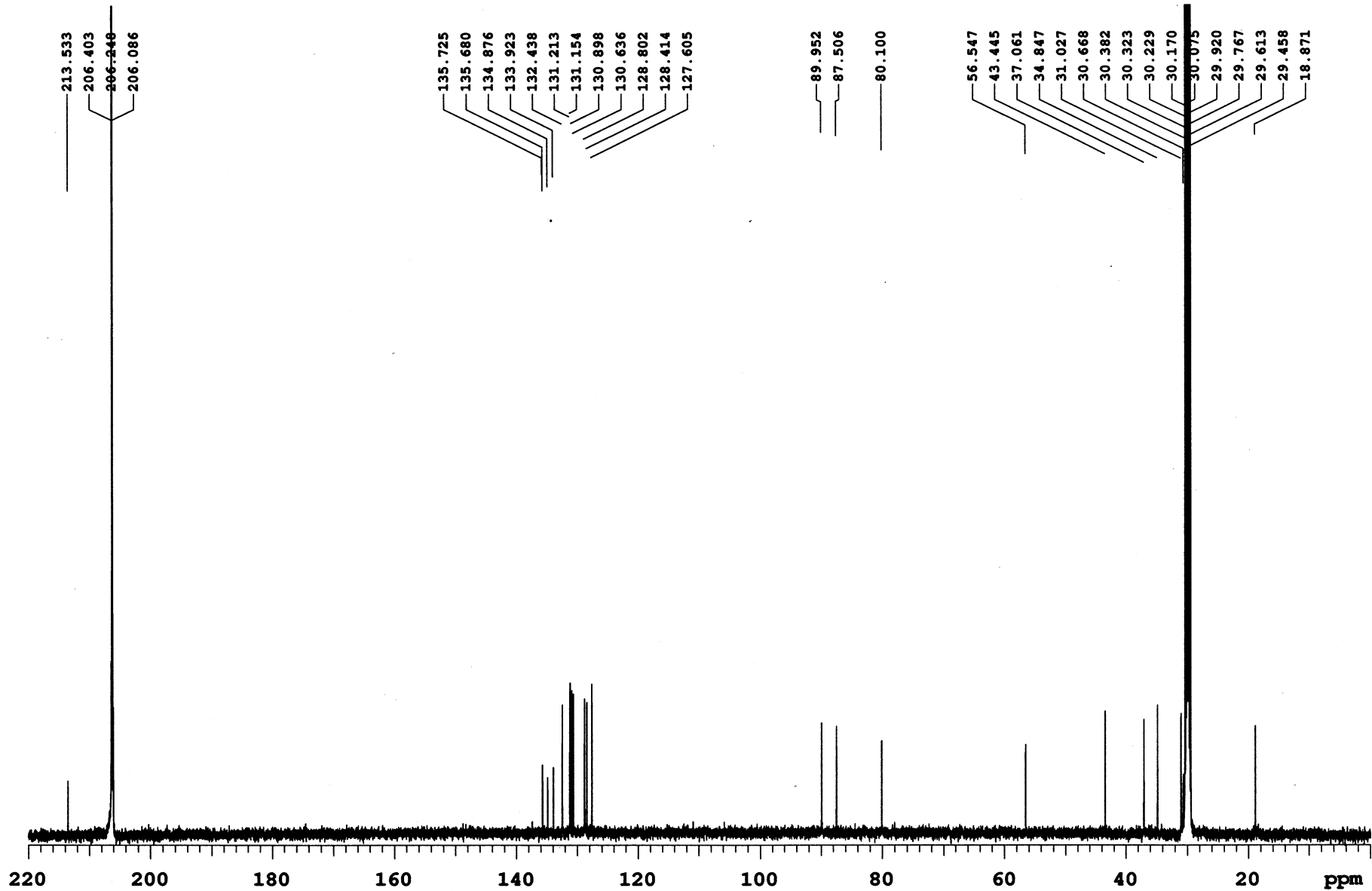


Fig S111. DEPT of compound 5g.

S111

CHP-81-11

Sample Name **CHP-81-11**
Date collected **2016-05-27**

Pulse sequence **DEPT**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

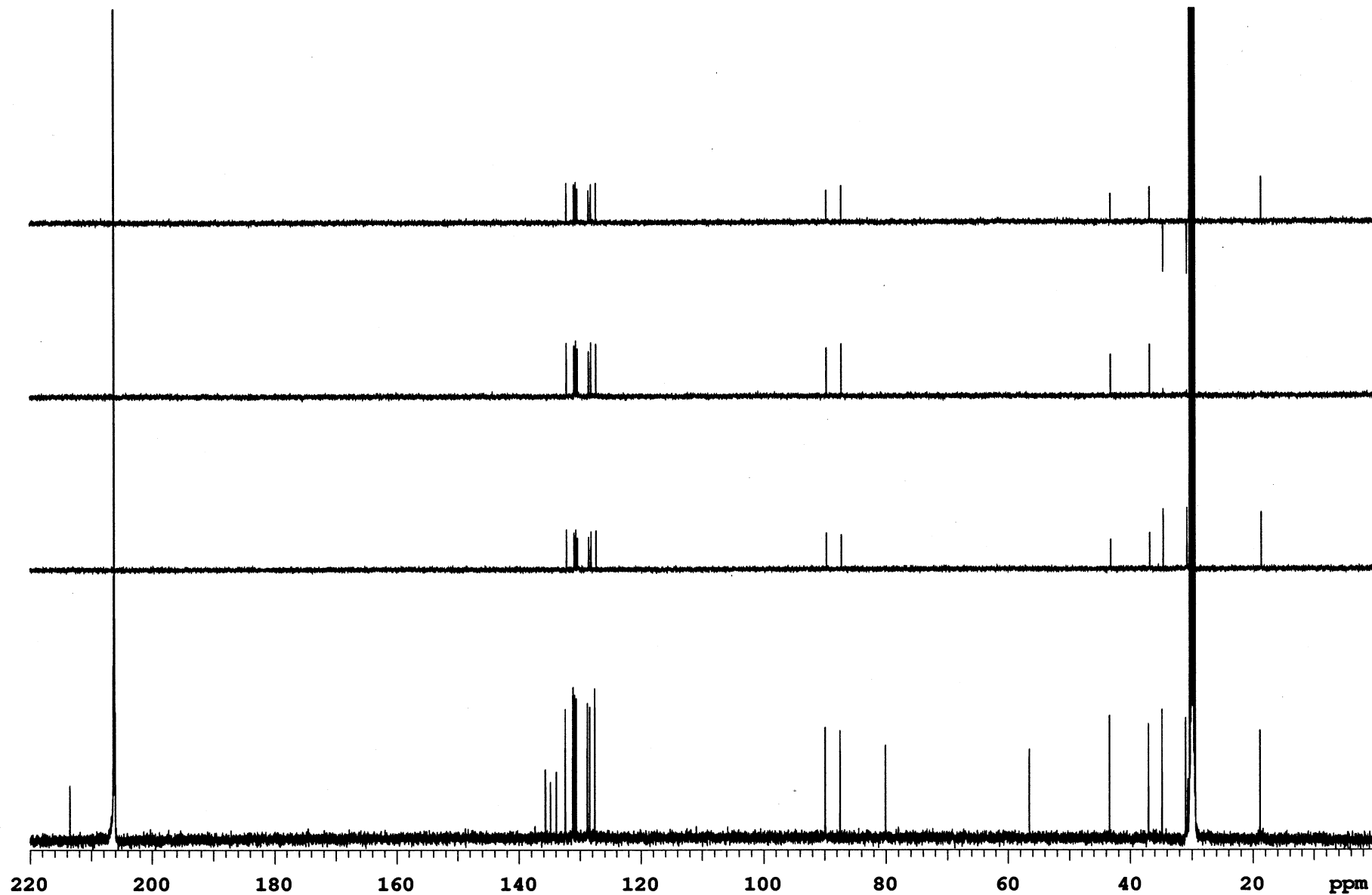


Fig S112. HSQC of compound 5g.

S112

CHP-81-f1

Sample Name **CHP-81-f1**
Date collected **2016-05-27**

Pulse sequence **gHSQC**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-Inova500**

Study owner **vnmr2**
Operator **vnmr2**

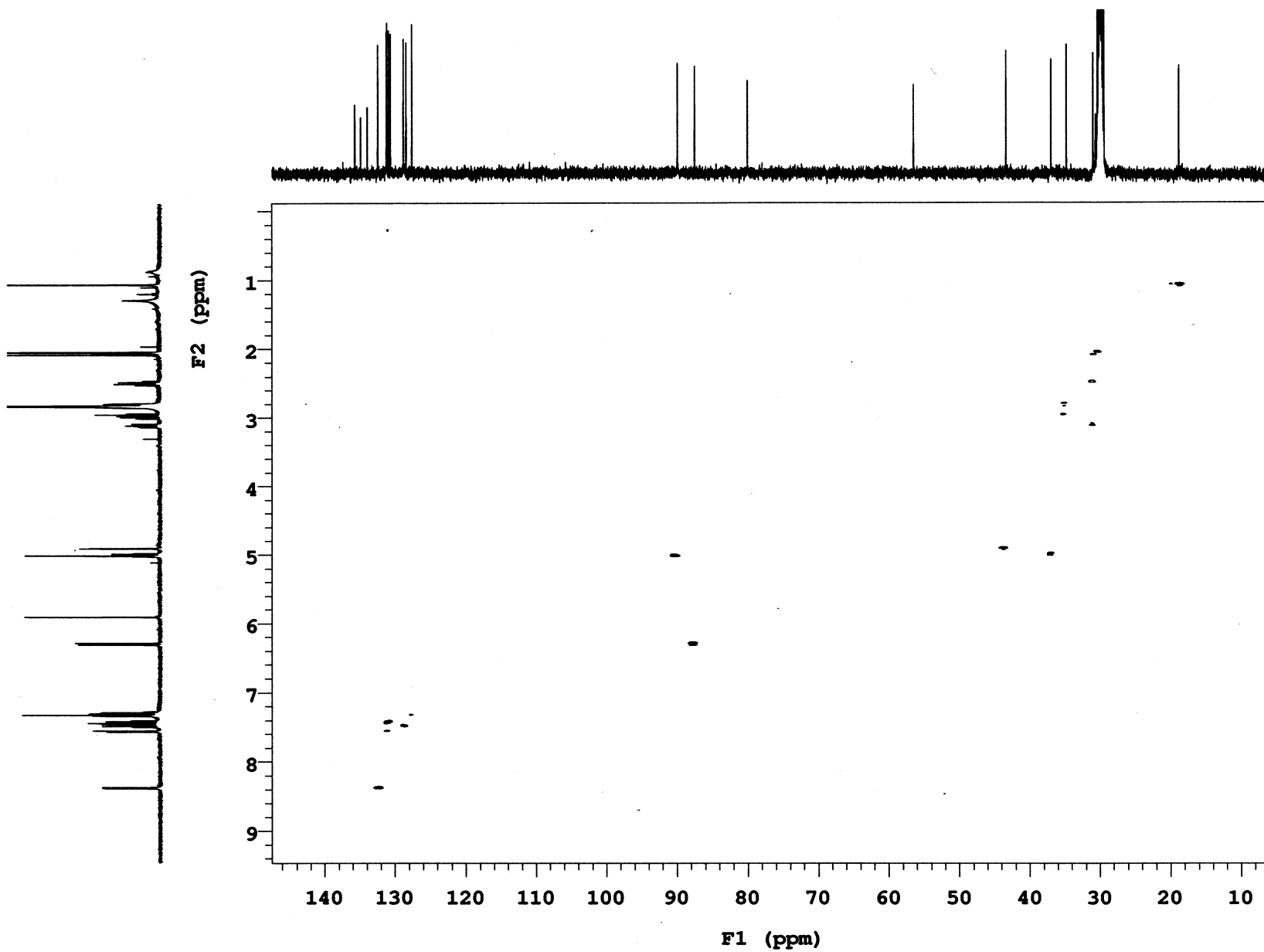


Fig S113. COSY of compound 5g.

S113

CHP-81-f1

Sample Name **CHP-81-f1**
Date collected **2016-05-27**

Pulse sequence **gCOSY**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

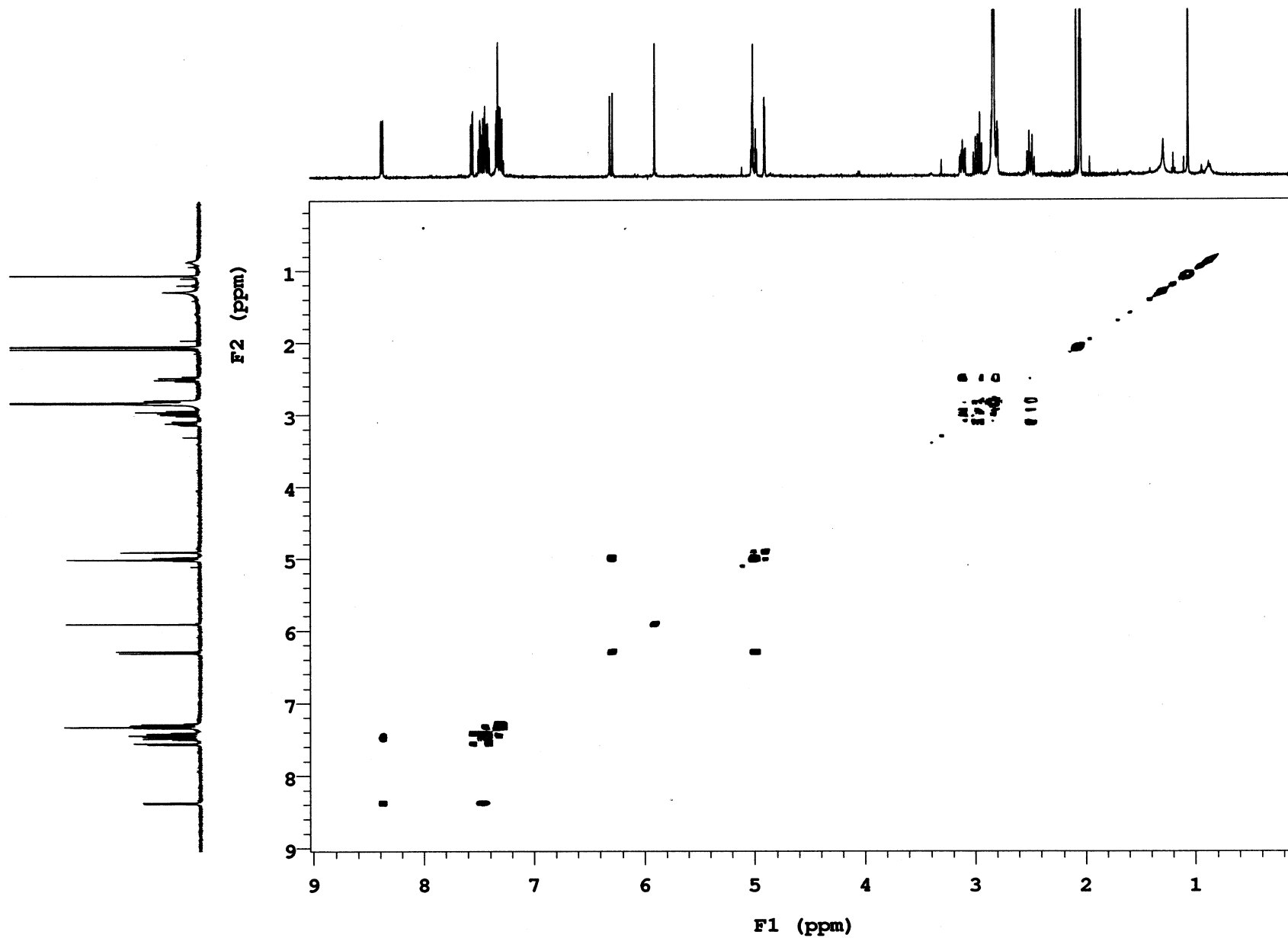


Fig S114. NOESY of compound 5g.

CHP-81-f1

Sample Name **CHP-81-f1**
Date collected **2016-05-27**

Pulse sequence **NOESY**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

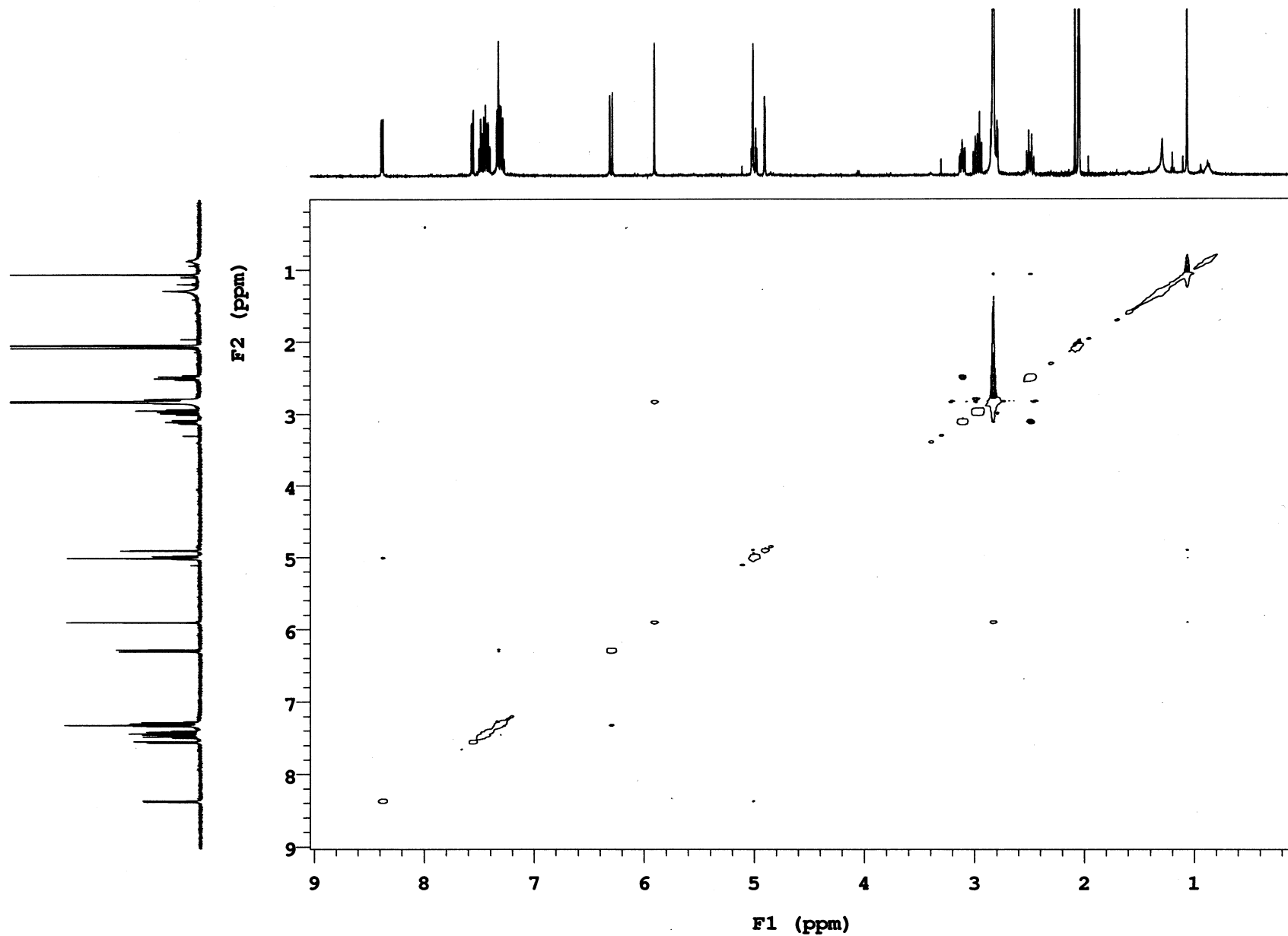
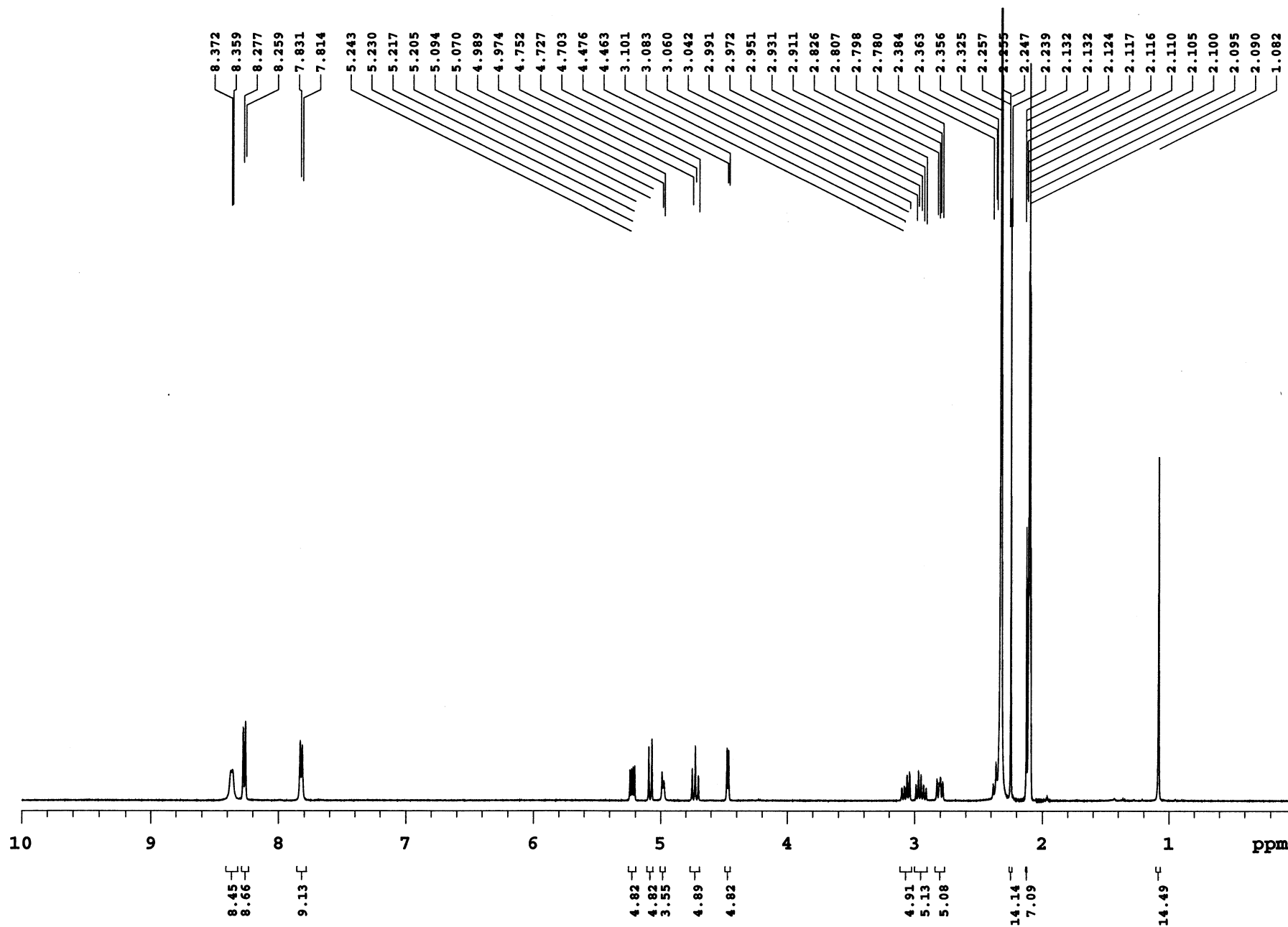
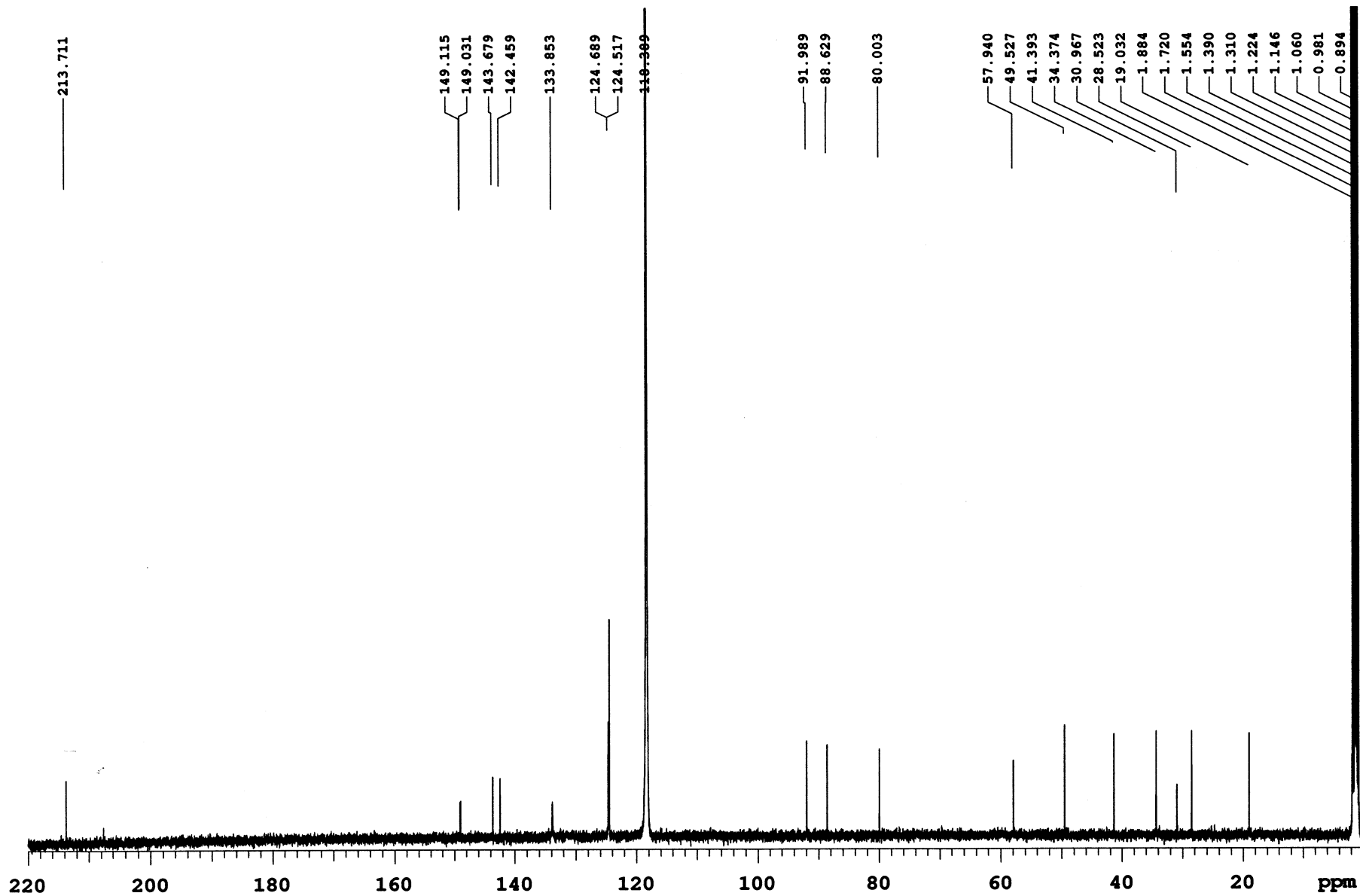


Fig S115. 1H NMR (CD3CN, 500 MHz) of compound 3h.

CHP-8f

Sample Name	CHP-8f	Pulse sequence	PROTON	Temperature	25	Study owner	vnmr2
Date collected	2016-02-03	Solvent	cd3cn	Spectrometer	Agilent-NMR-inova500	Operator	vnmr2



Sample Name **CHP-8f**
Date collected **2016-02-03**Pulse sequence **CARBON**
Solvent **cd3cn**Temperature **25**
Spectrometer **Agilent-NMR-inova500**Study owner **vnmr2**
Operator **vnmr2**Fig S116. ^{13}C NMR (CD_3CN , 125 MHz) of compound 3h.

Sample Name **CHP-8f**
Date collected **2016-02-04**

Pulse sequence **DEPT**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

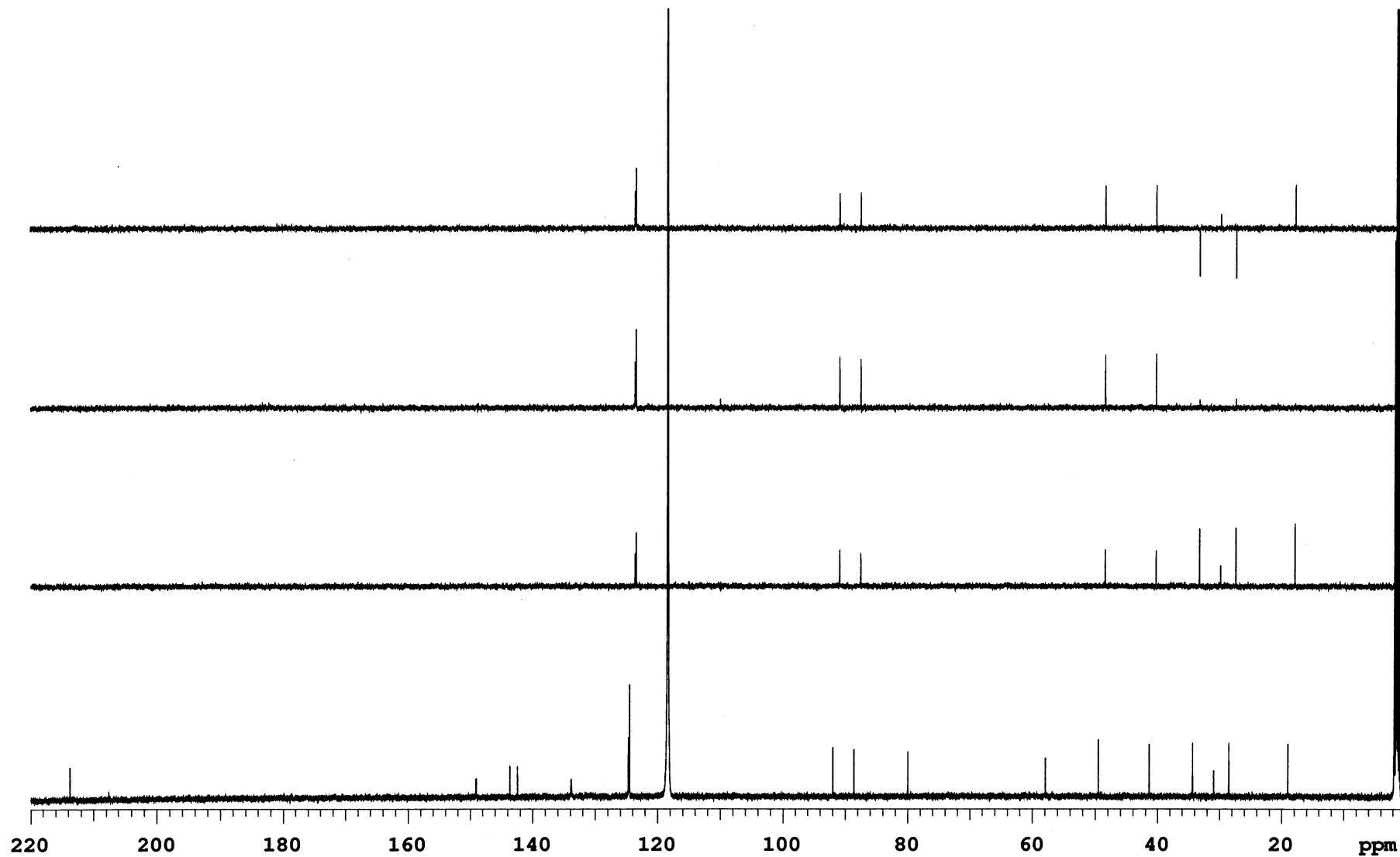


Fig S117. DEPT of compound 3h.

Sample Name **CHP-8f**
 Date collected **2016-02-04**

Pulse sequence **gHSQC**
 Solvent **cd3cn**

Temperature **25**
 Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
 Operator **vnmr2**

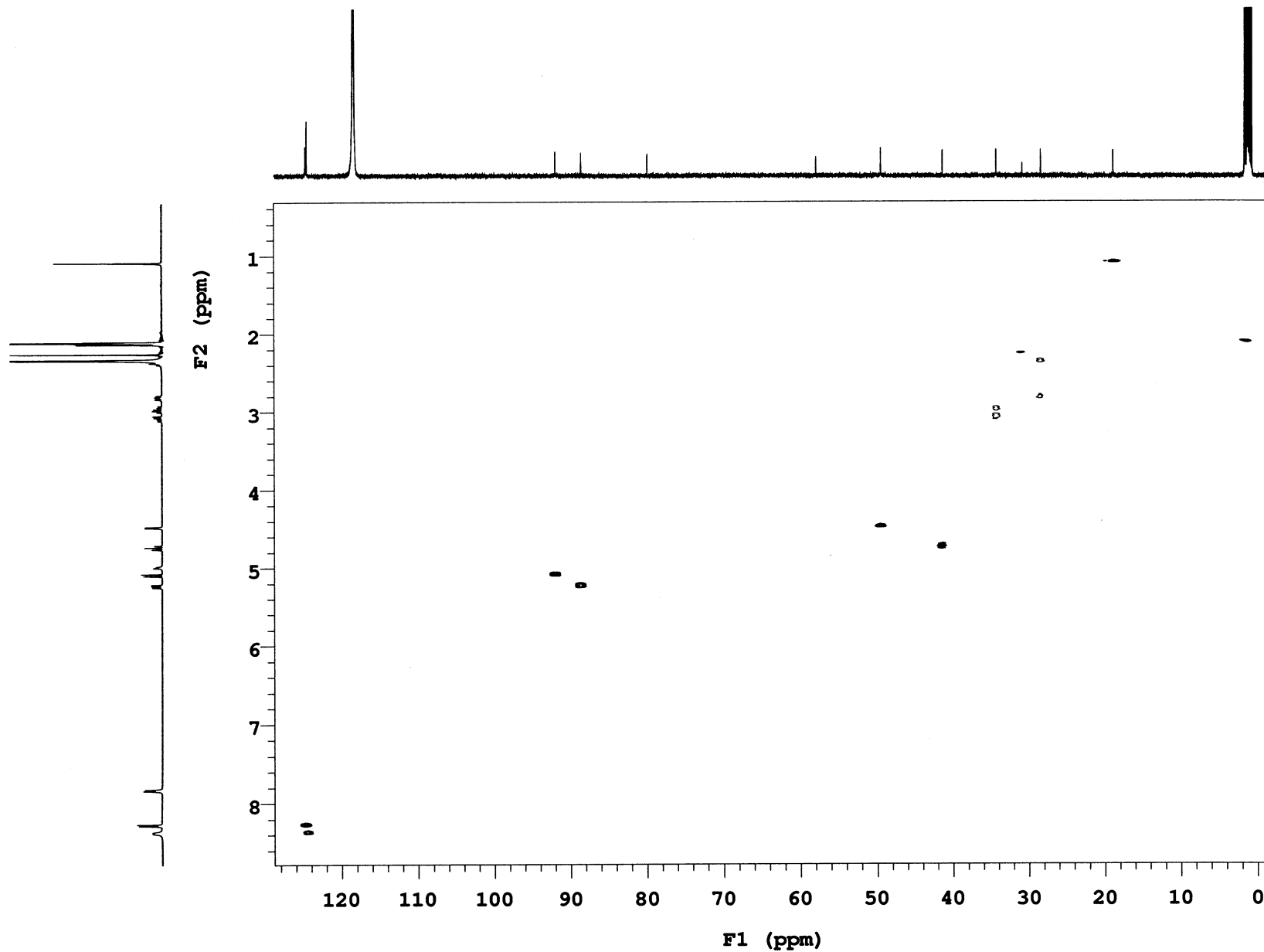


Fig S118. HSQC of compound 3h.

Sample Name **CHP-8f**
Date collected **2016-02-04**

Pulse sequence **NOESY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

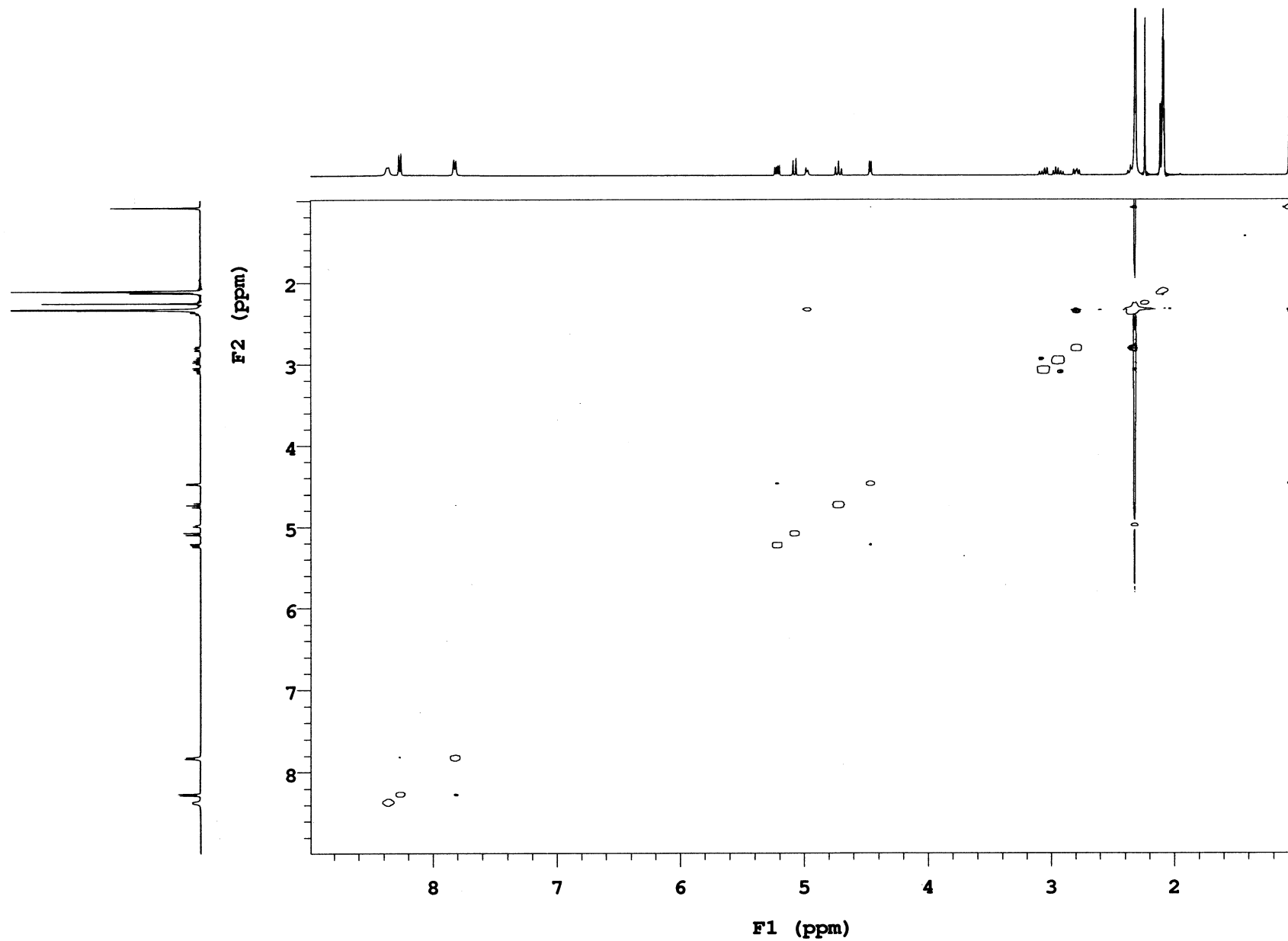


Fig S120. NOESY of compound 3h.

Fig S121. ¹H NMR (CH₃CN, 500 MHz) of compound 5h

S121

CHP-8f-f3

Sample Name	CHP-8f-f3	Pulse sequence	PROTON	Temperature	25	Study owner	vnmr2
Date collected	2016-05-20	Solvent	cd3cn	Spectrometer	Agilent-NMR-inova500	Operator	vnmr2

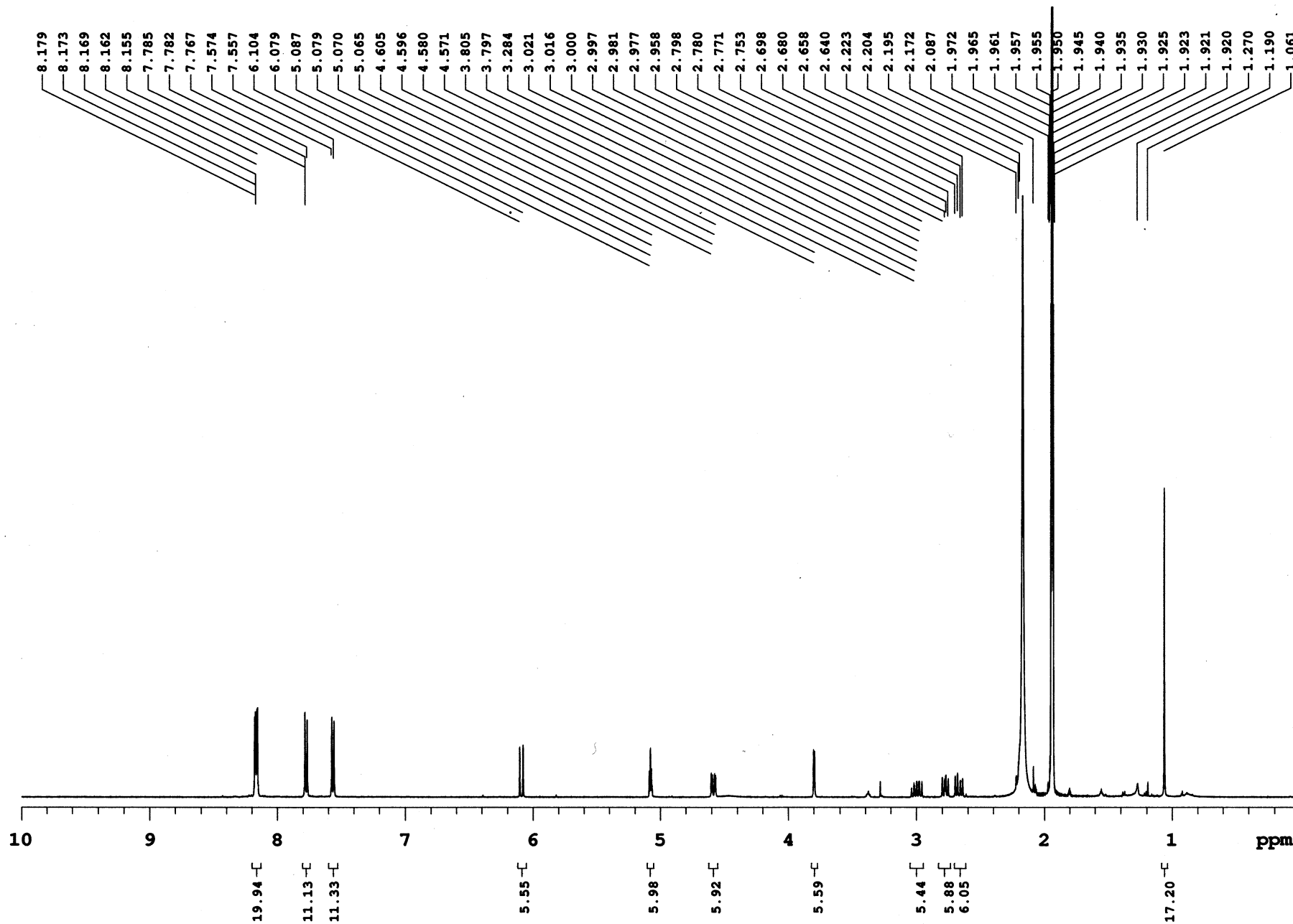


Fig S122. ¹³C NMR (CH₃CN, 125 MHz) of compound 5h

S122

CHP-8f-f3

Sample Name **CHP-8f-f3**
Date collected **2016-05-20**

Pulse sequence **CARBON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

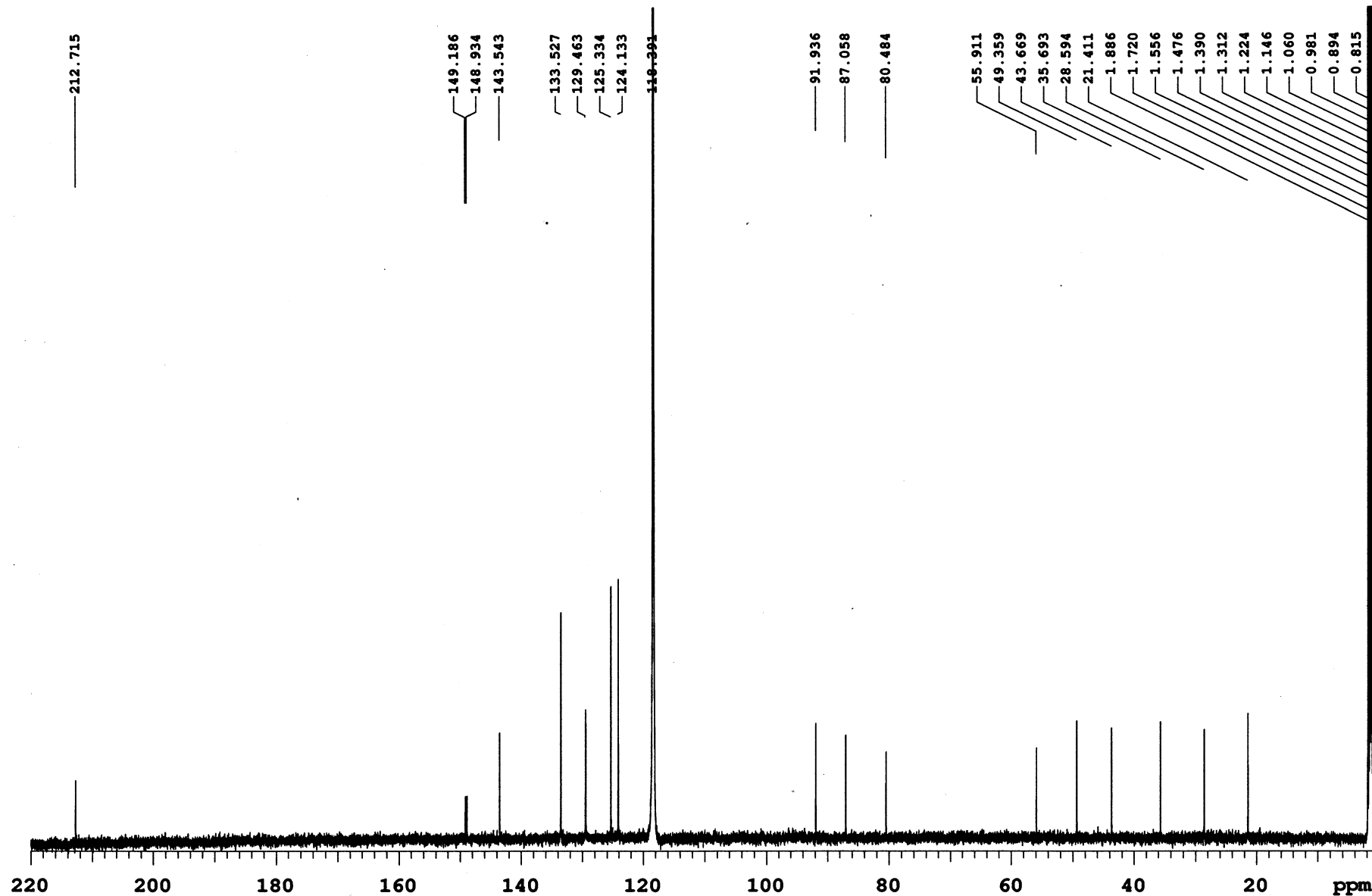


Fig S123. DEPT of compound 5h

S123

CHP-8f-f3

Sample Name **CHP-8f-f3**
Date collected **2016-05-21**

Pulse sequence **DEPT**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

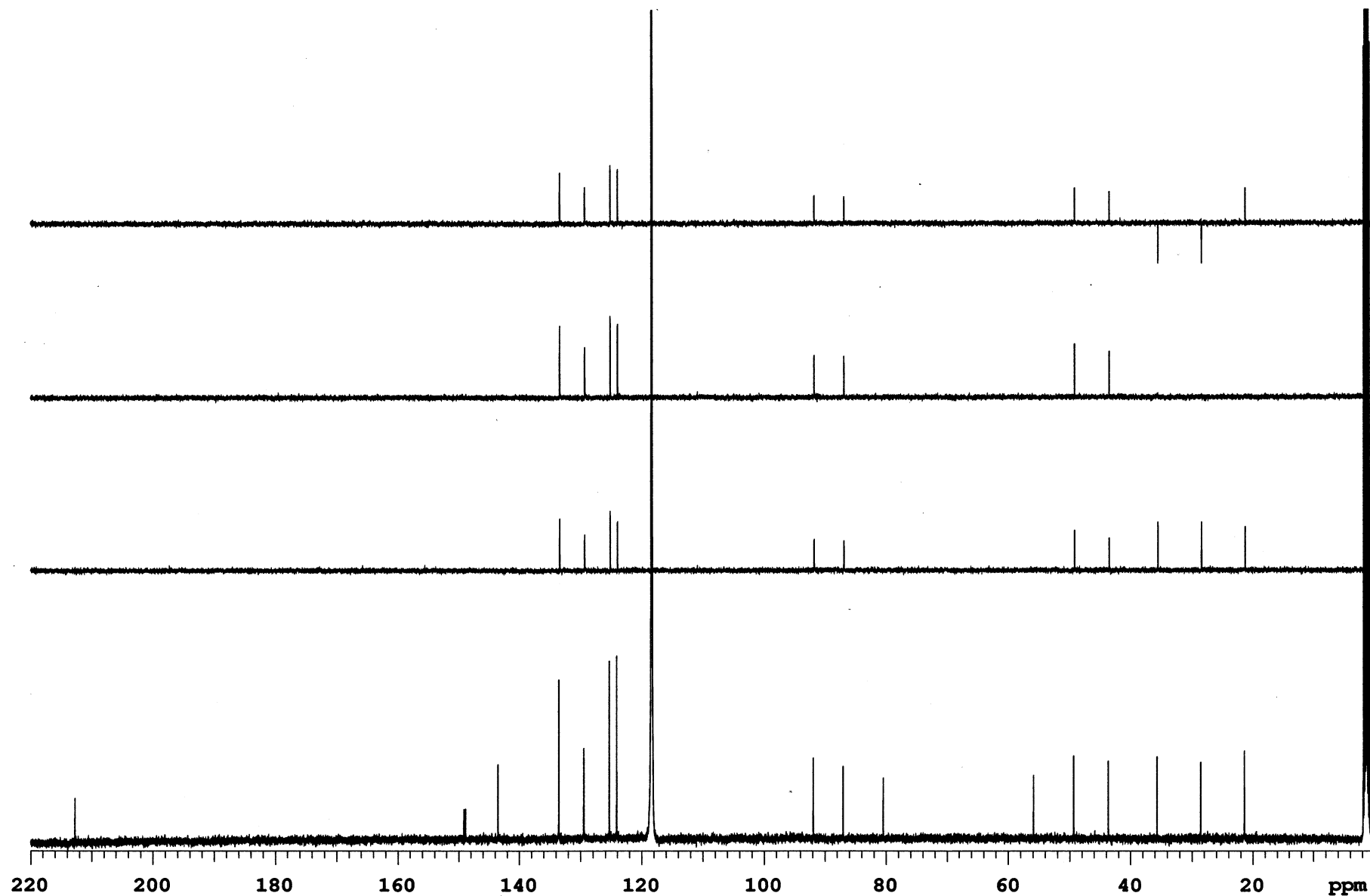


Fig S124. HSQC of compound 5h

S124

CHP-8f-f3

Sample Name **CHP-8f-f3**
Date collected **2016-05-21**

Pulse sequence **gHSQC**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

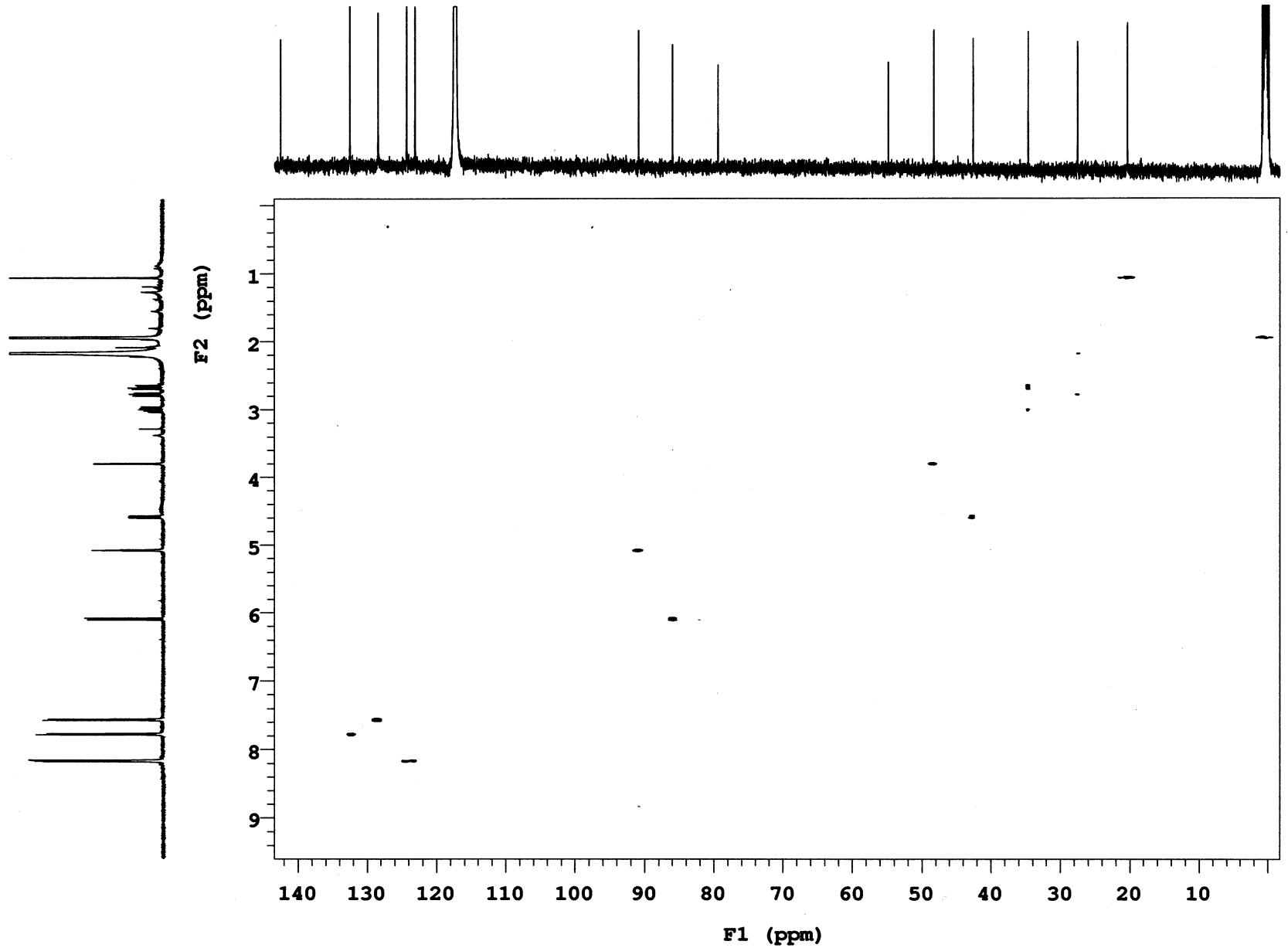


Fig S125. COSY of compound 5h

CHP-8f-f3

Sample Name **CHP-8f-f3**
Date collected **2016-05-21**

Pulse sequence **gCOSY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

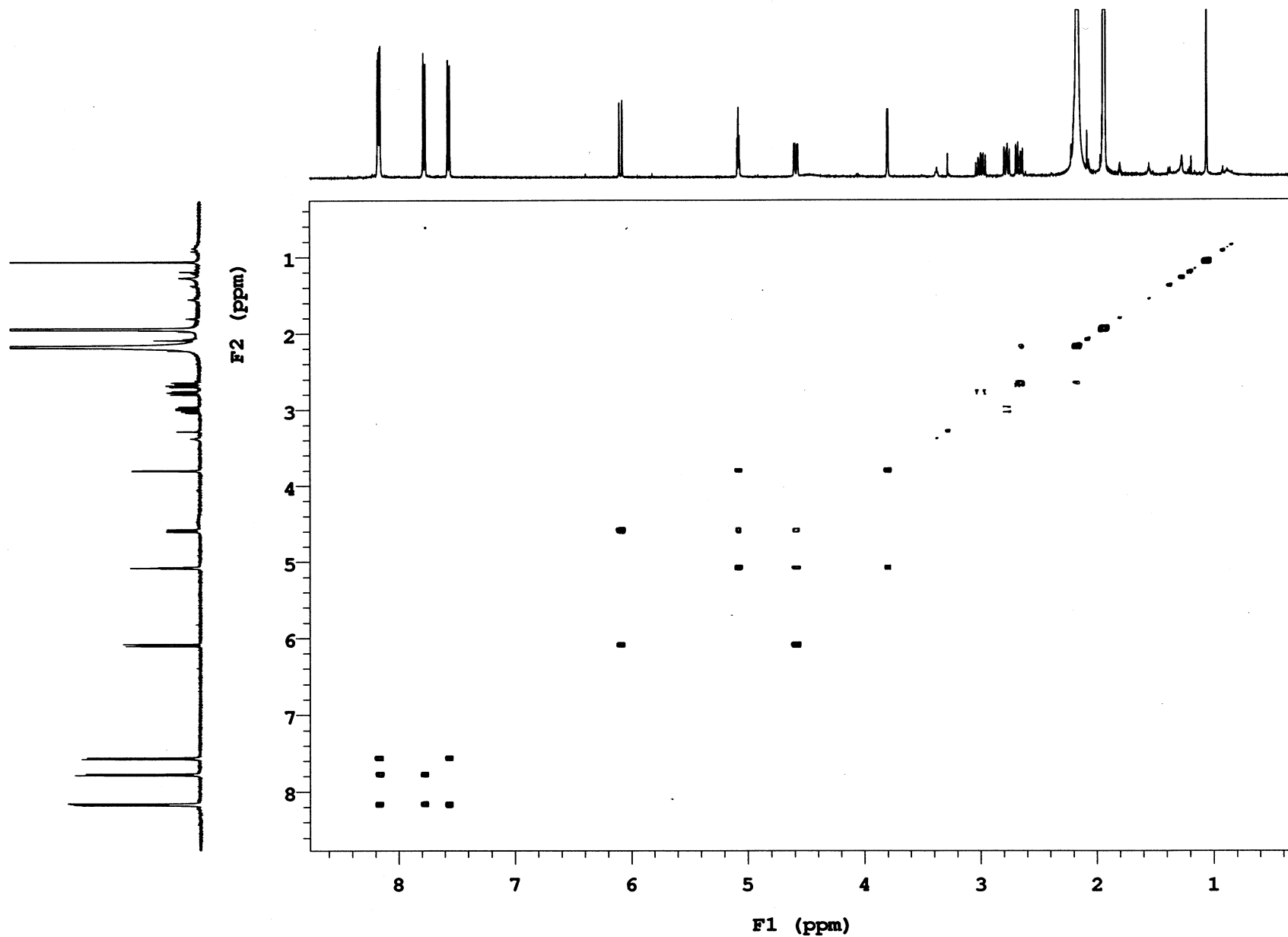


Fig S126. NOESY of compound 5h

S126

CHP-8f-f3

Sample Name **CHP-8f-f3**
Date collected **2016-05-21**

Pulse sequence **NOESY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

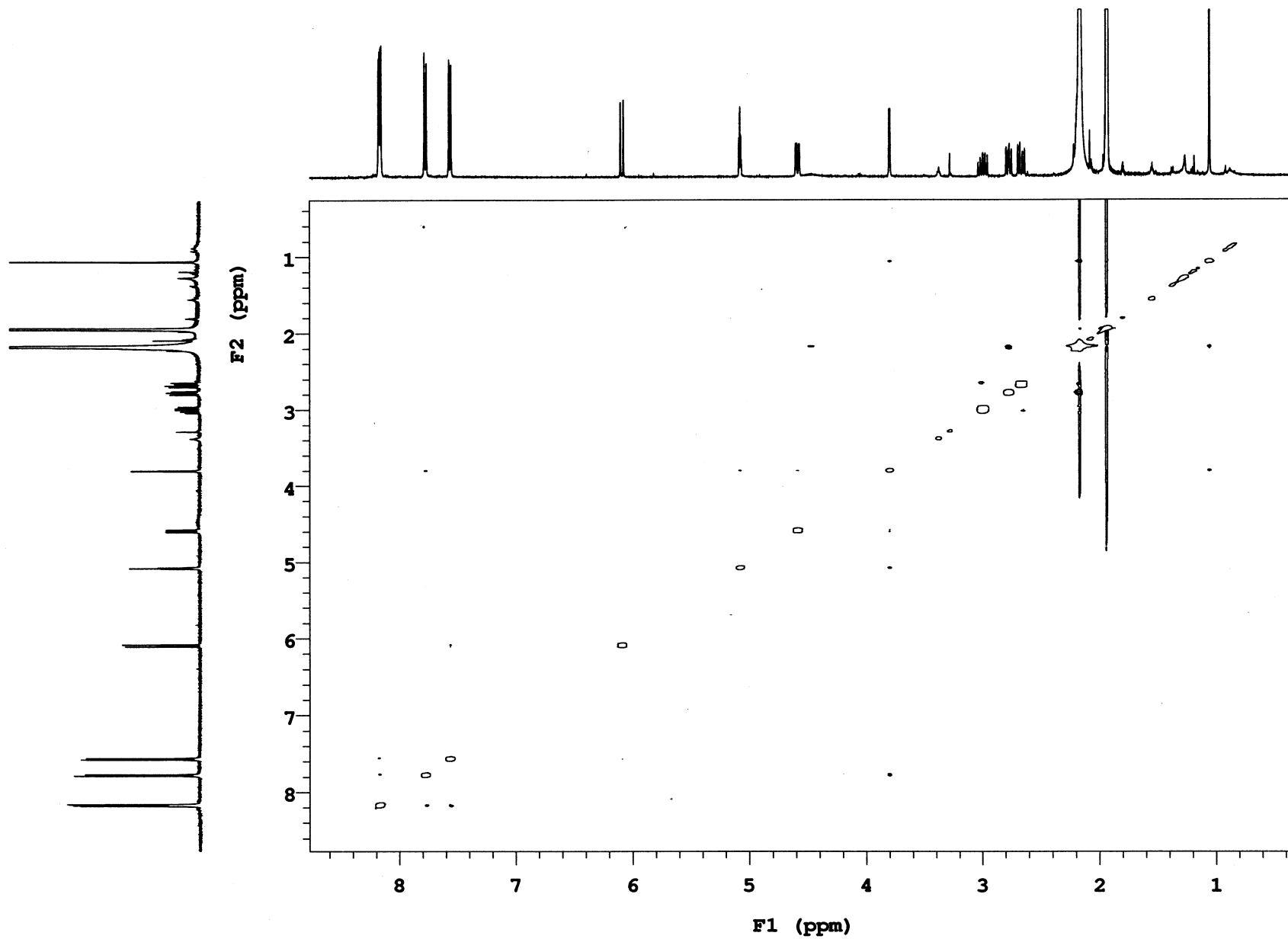


Fig S127. ¹H NMR (acetone-d₆, 500 MHz) of compound 3i.

S127

CHP-8g-F2

Sample Name **CHP-8g-F2**
Date collected **2016-05-12**

Pulse sequence **PROTON**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-Inova500**

Study owner **vnmr2**
Operator **vnmr2**

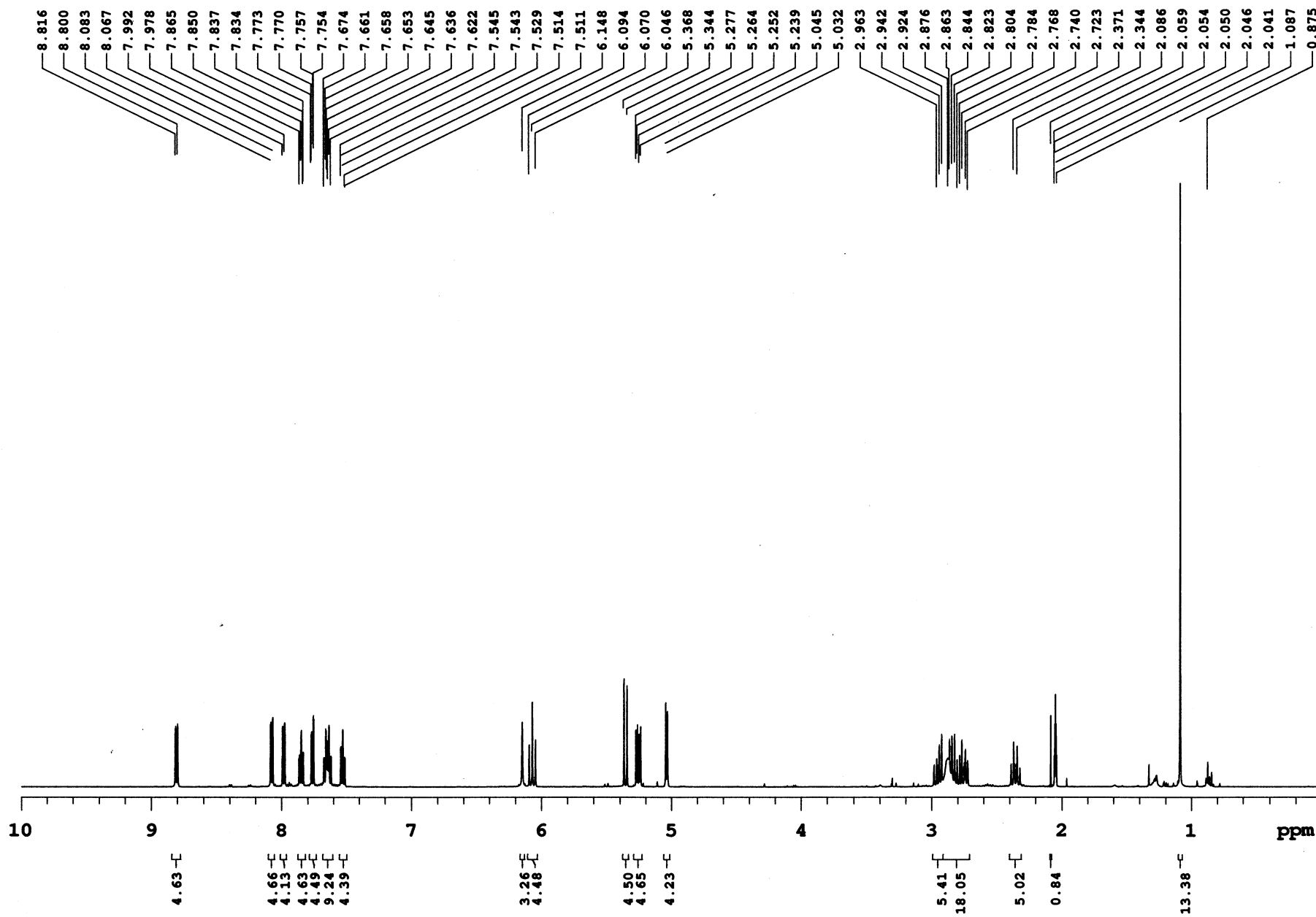


Fig S128. ¹³C NMR (acetone-d₆, 125 MHz) of compound 3i.

S128

CHP-8g-F2

Sample Name **CHP-8g-F2**
Date collected **2016-05-09**

Pulse sequence **CARBON**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

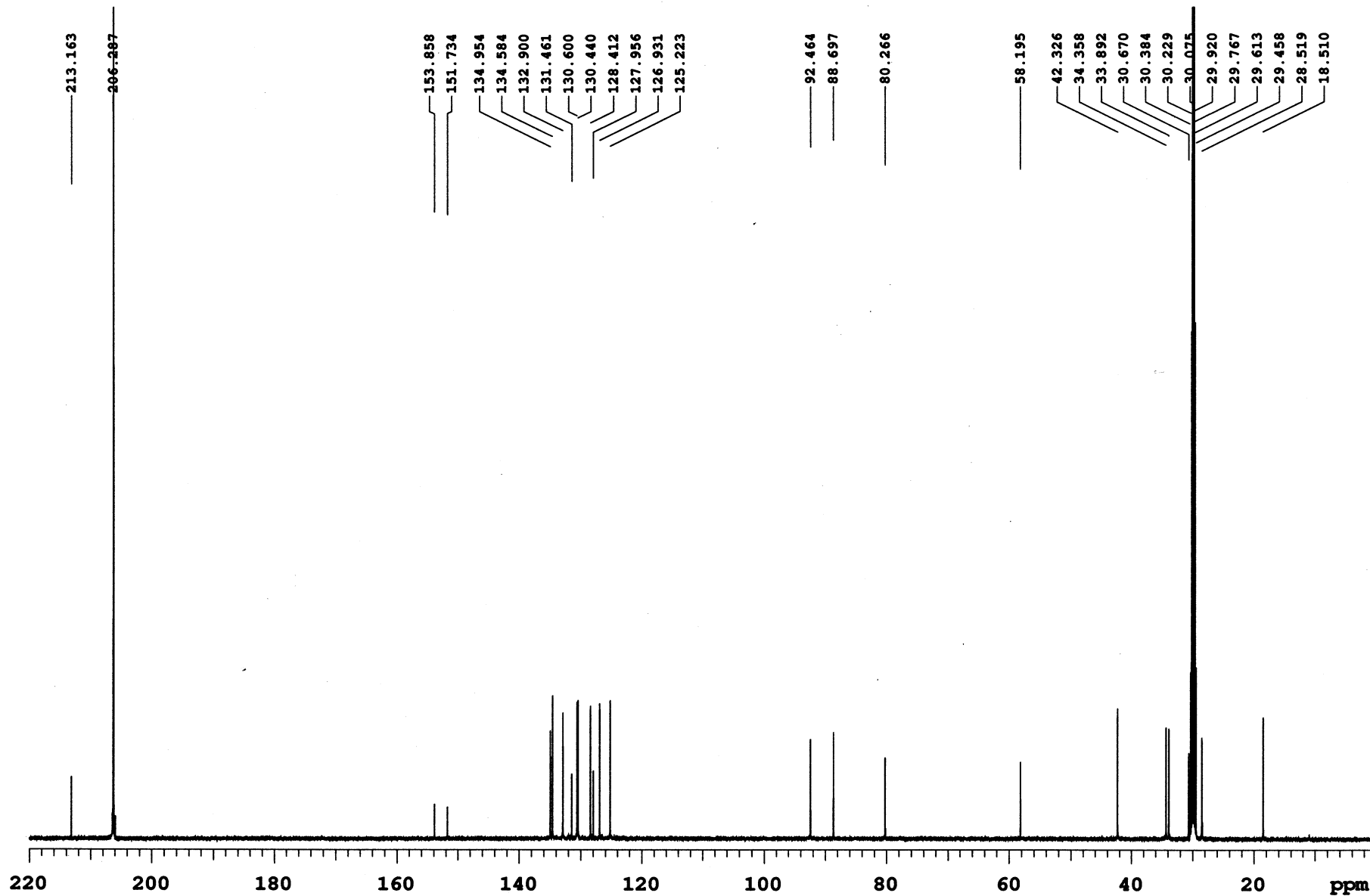


Fig S129. DEPT (acetone-d6) of compound 3i.

CHP-8g-F2

Sample Name **CHP-8g-F2**
Date collected **2016-05-10**

Pulse sequence **DEPT**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

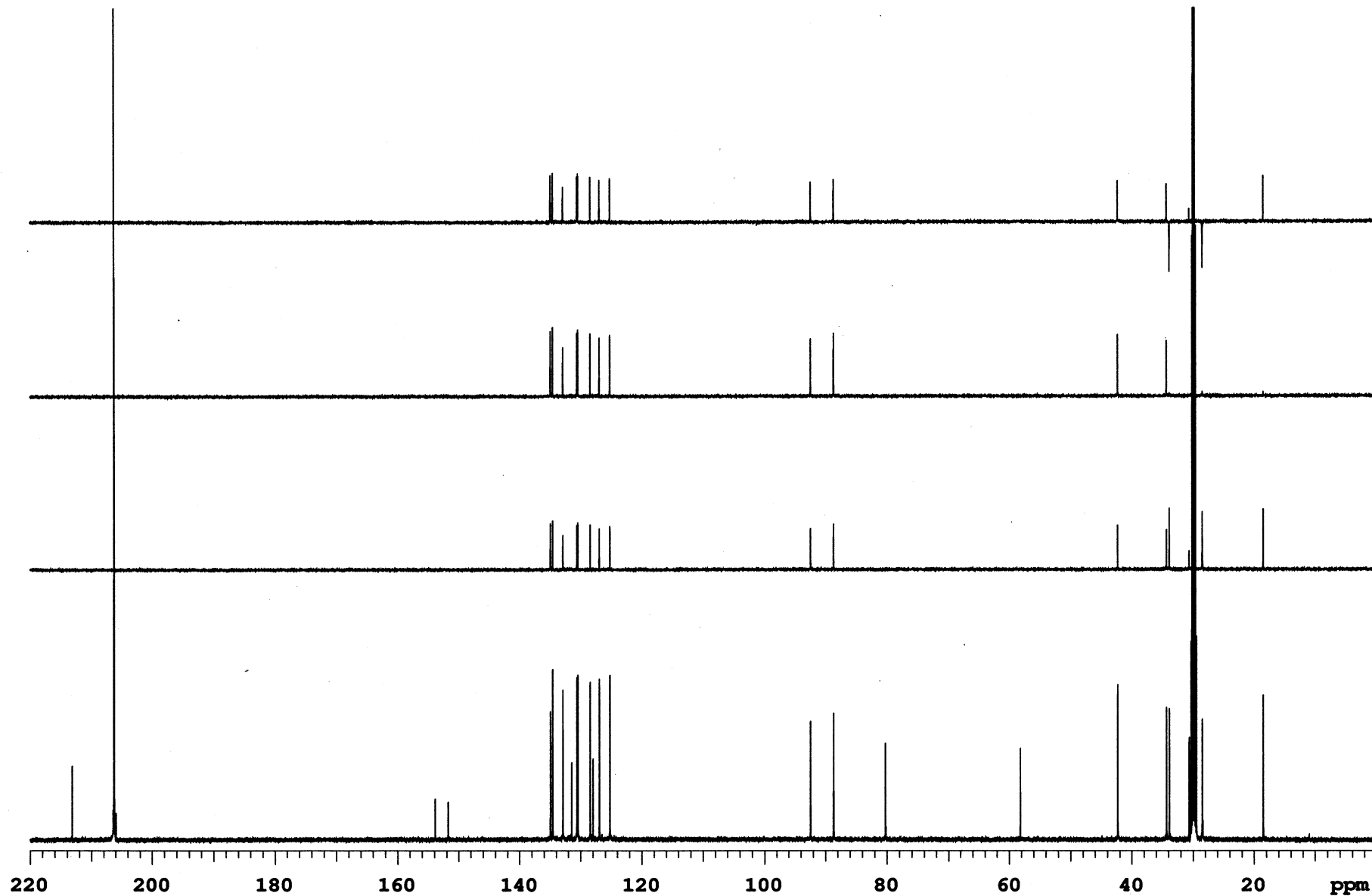


Fig S130. HSQC (acetone-d6) of compound 3i.

S130

CHP-8g-F2

Sample Name **CHP-8g-F2**
Date collected **2016-05-10**

Pulse sequence **gHSQC**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

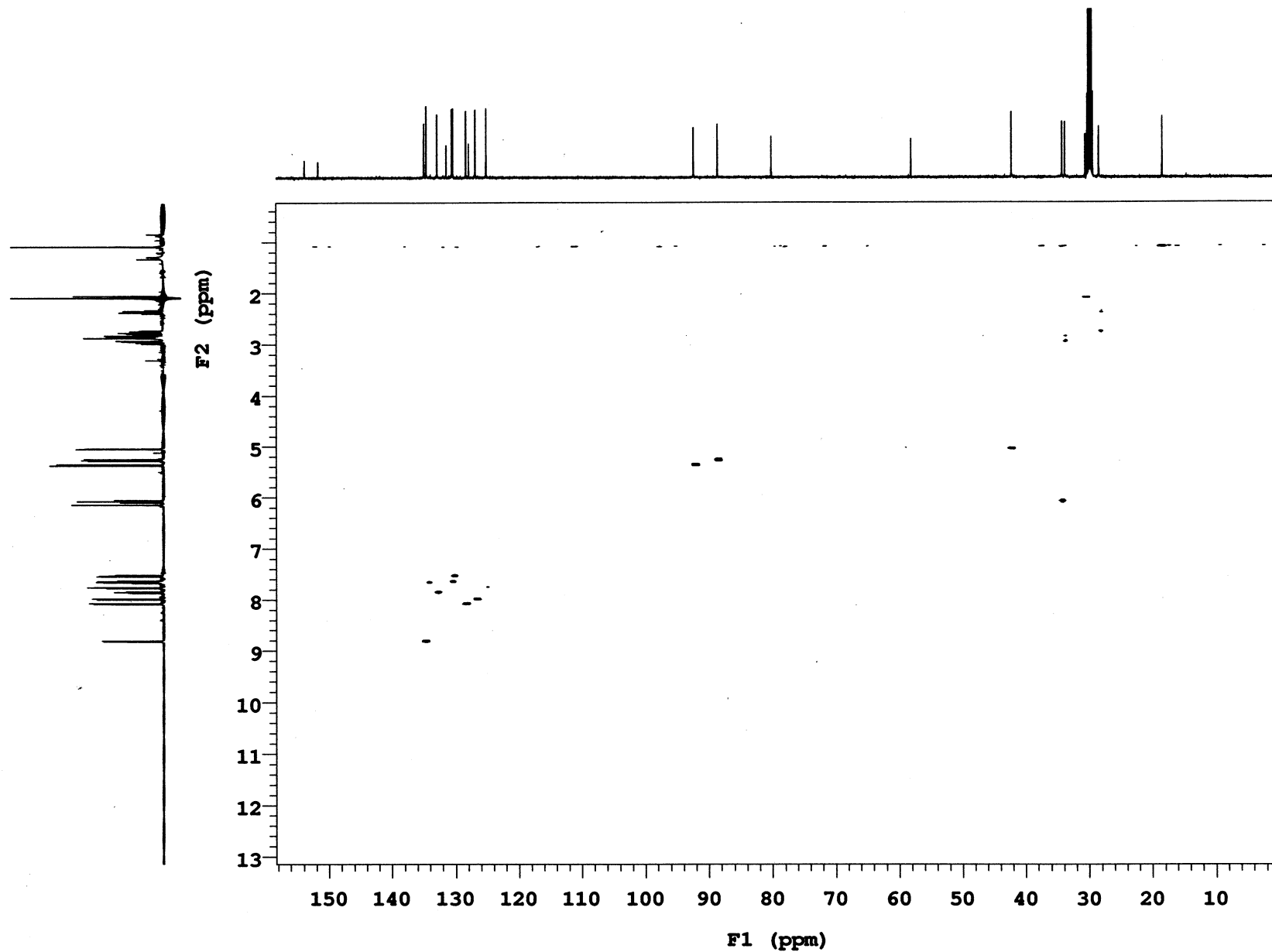


Fig S131. COSY (acetone-d6) of compound 3i.

CHP-8g-F2

Sample Name **CHP-8g-F2**
Date collected **2016-05-10**

Pulse sequence **gCOSY**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

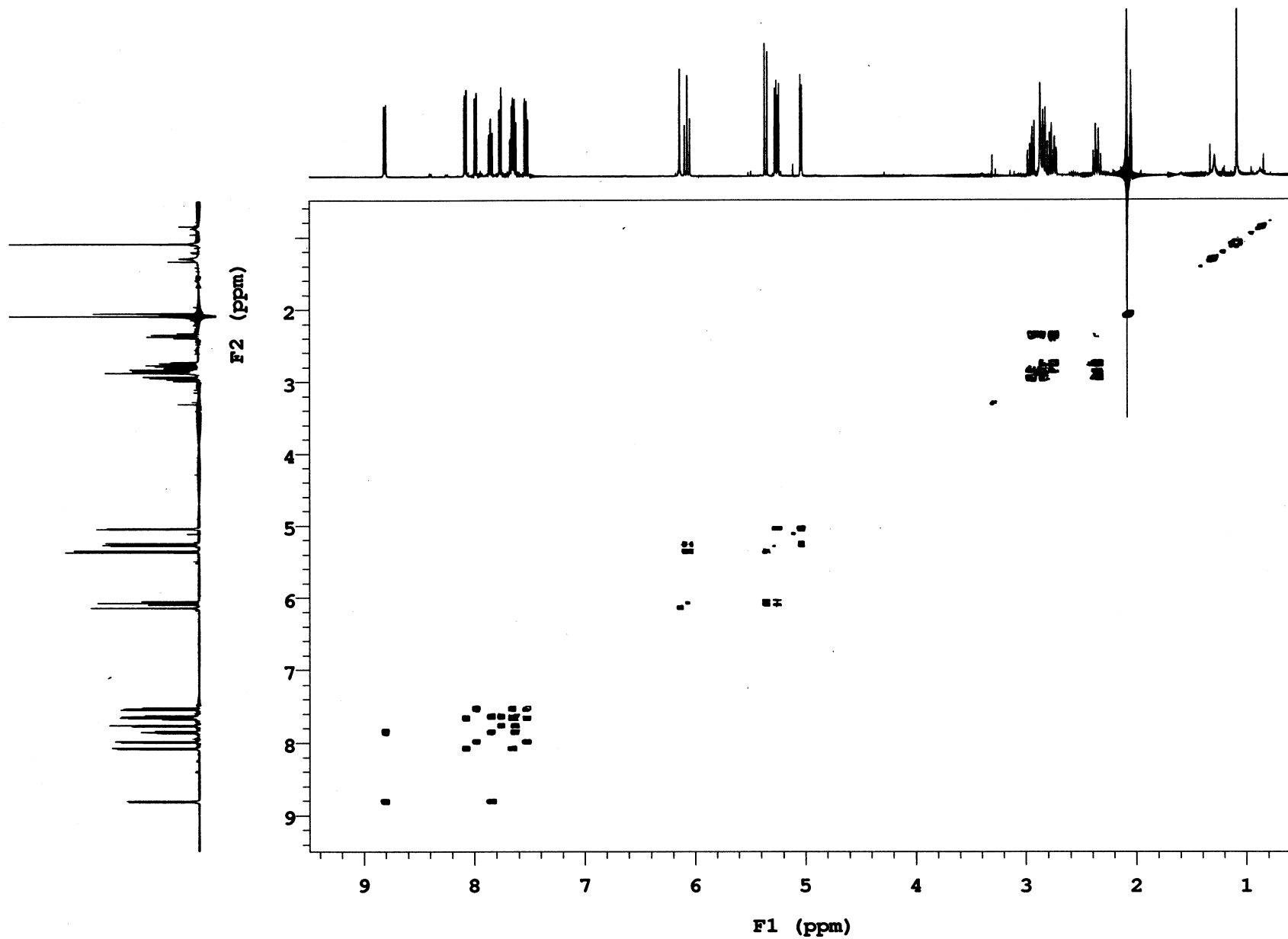


Fig S132. NOESY (acetone-d6) of compound 3i.

CHP-8g-F2

Sample Name **CHP-8g-F2**
Date collected **2016-05-10**

Pulse sequence **NOESY**
Solvent **acetone**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

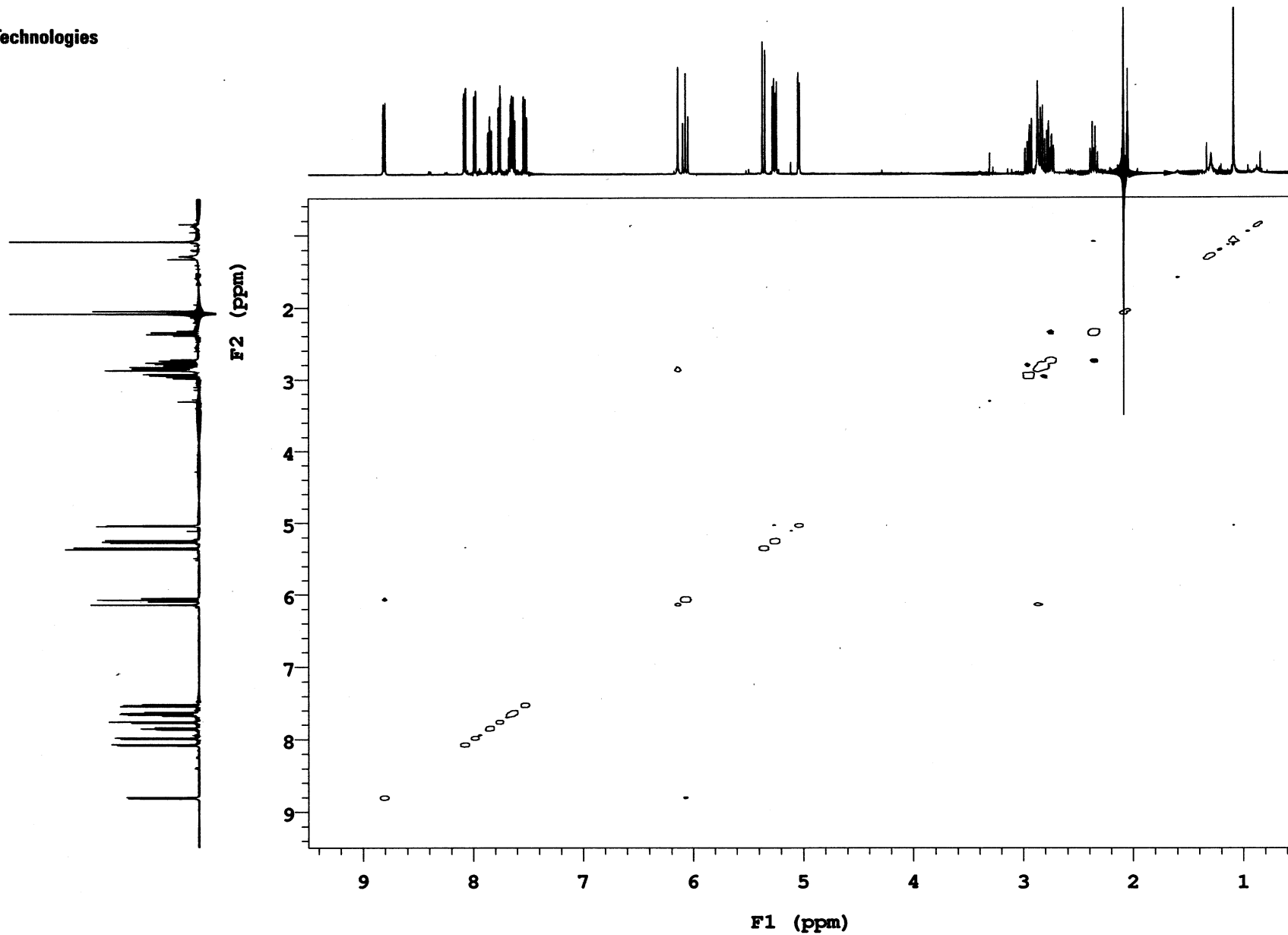


Fig S133. ¹H NMR (CD₃CN, 500 MHz) of compound 3i.

CHP-01-075-F5

Sample Name **CHP-01-075-F5**
Date collected **2016-05-02**

Pulse sequence **PROTON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

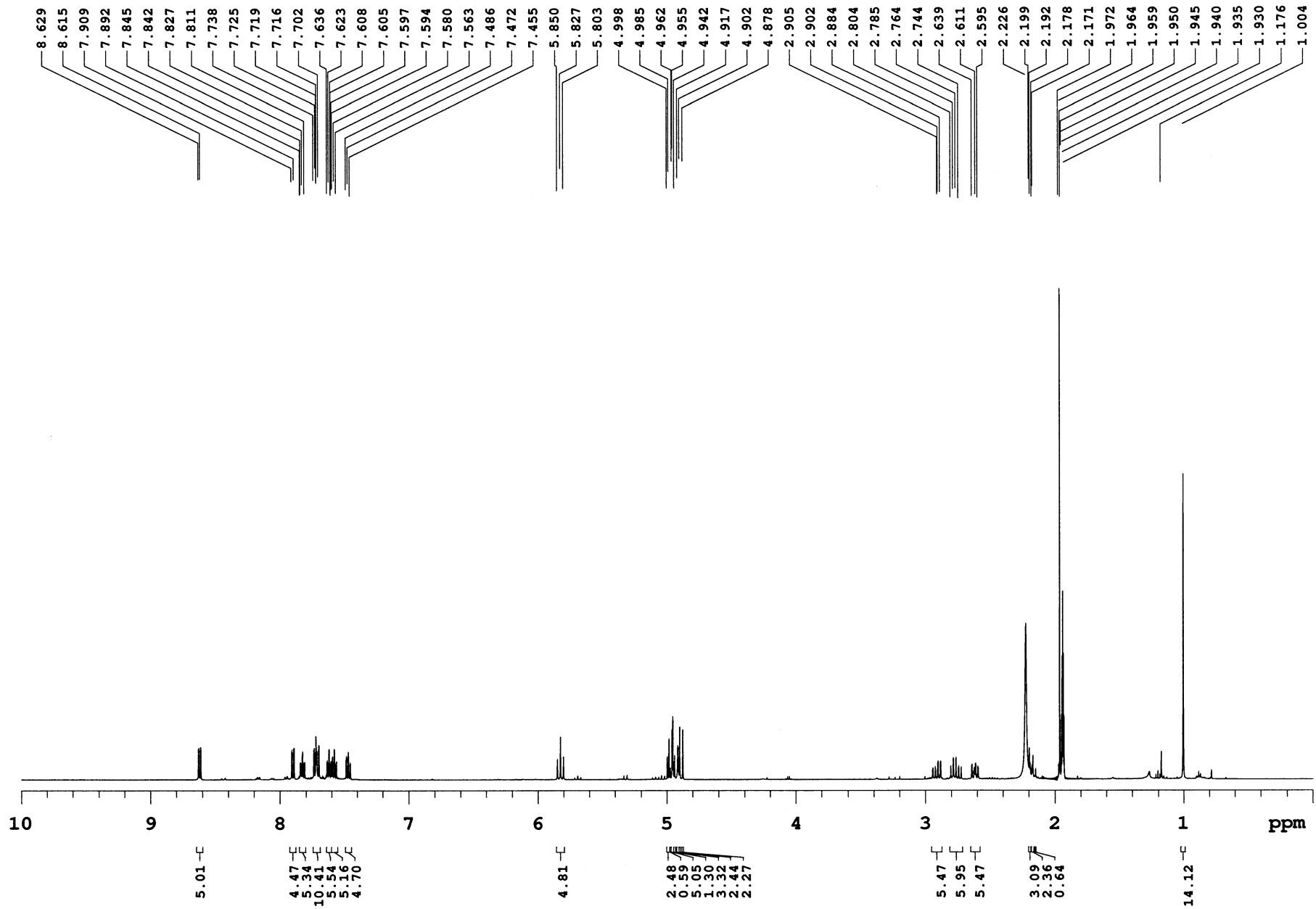


Fig S134. ¹³C NMR (CD₃CN, 125 MHz) of compound 3i.

S134

CHP-01-075-F5

Sample Name **CHP-01-075-F5**
Date collected **2016-05-02**

Pulse sequence **CARBON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

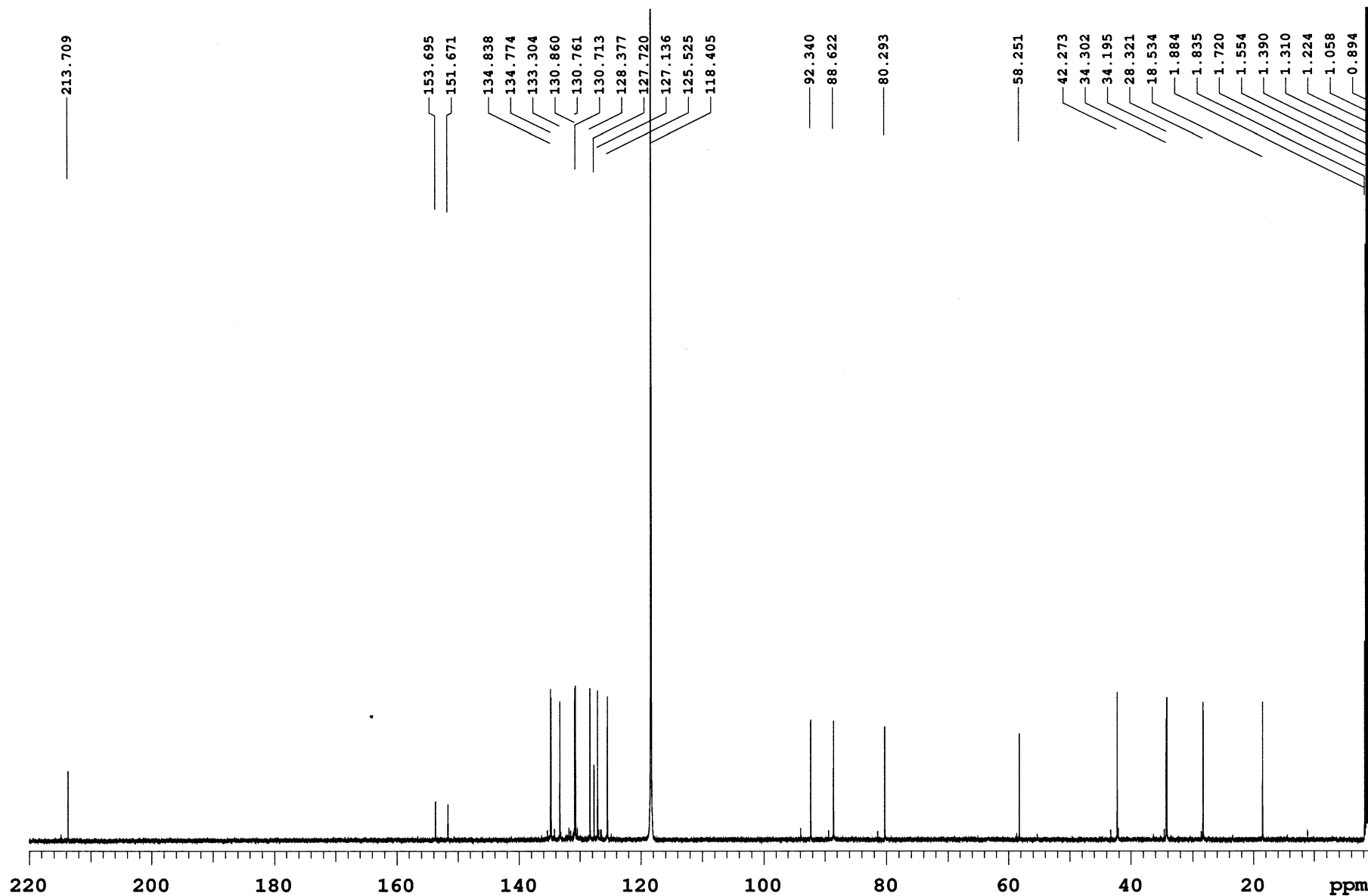


Fig S135. DEPT (CD3CN) of compound 3i.

S135

CHP-01-075-F5

Sample Name **CHP-01-075-F5**
Date collected **2016-05-03**

Pulse sequence **DEPT**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

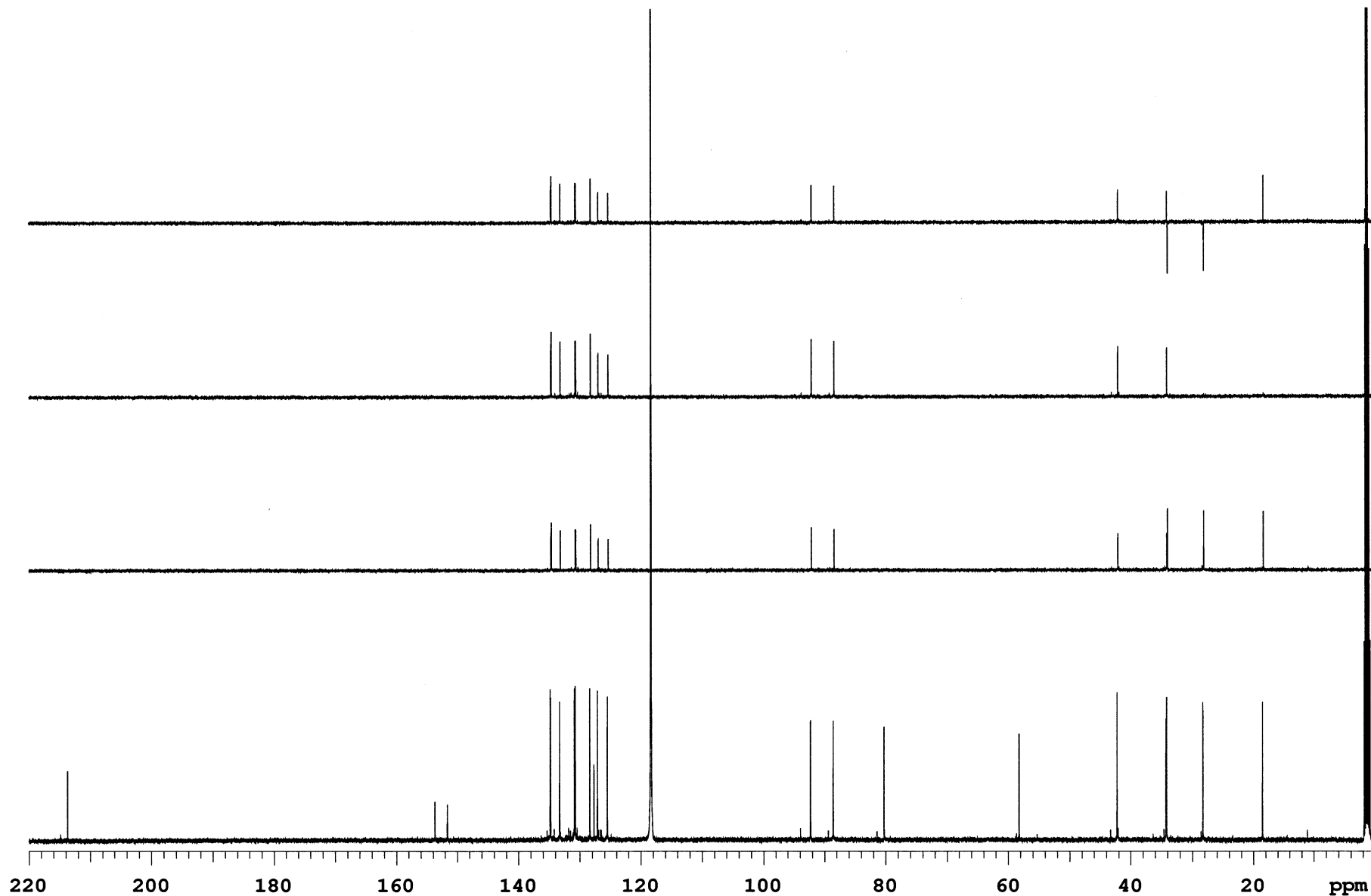


Fig S136. HSQC (CD3CN) of compound 3i.

S136

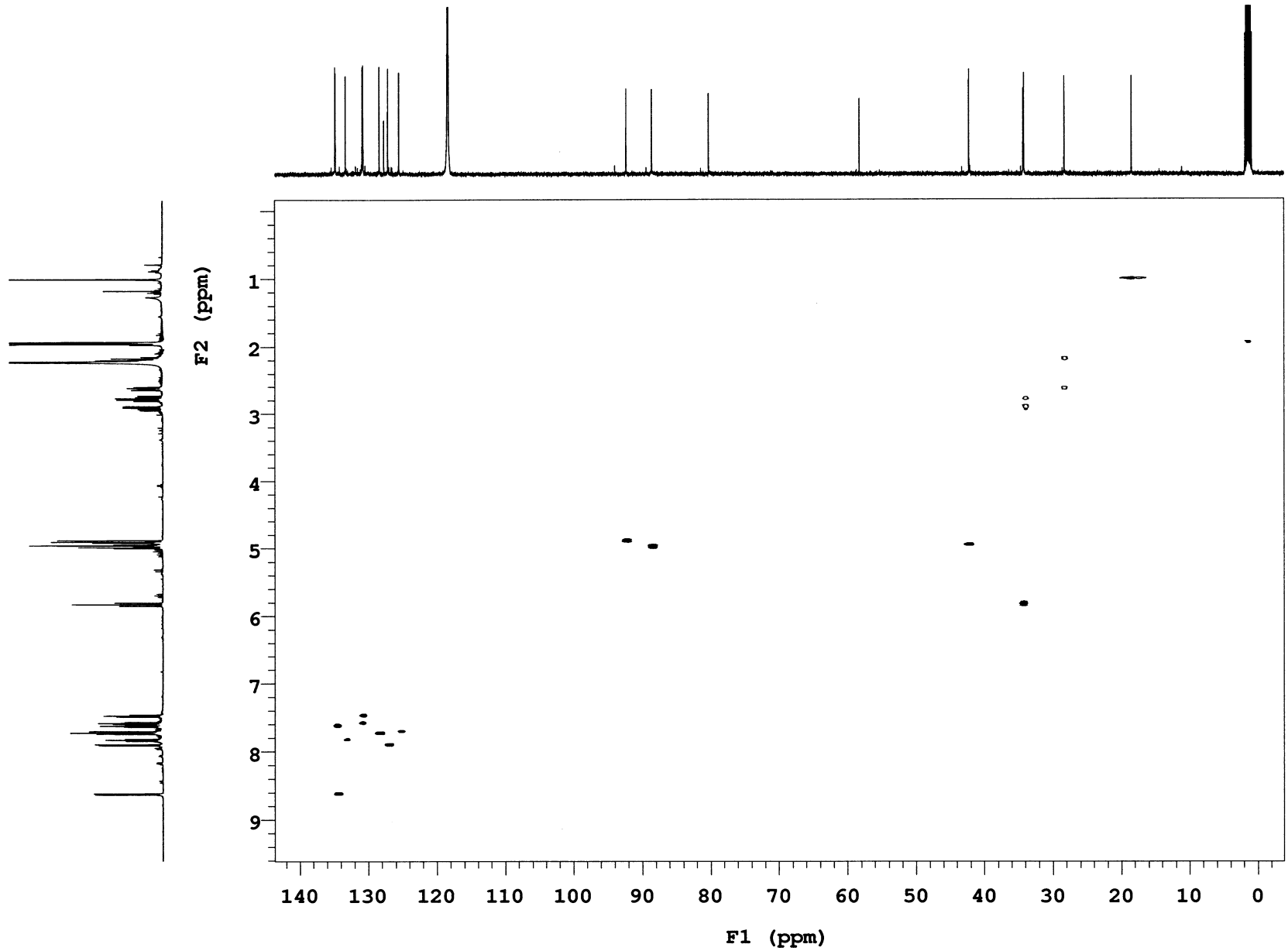
CHP-01-075-F5

Sample Name **CHP-01-075-F5**
Date collected **2016-05-03**

Pulse sequence **gHSQC**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**



CHP-01-075-F5

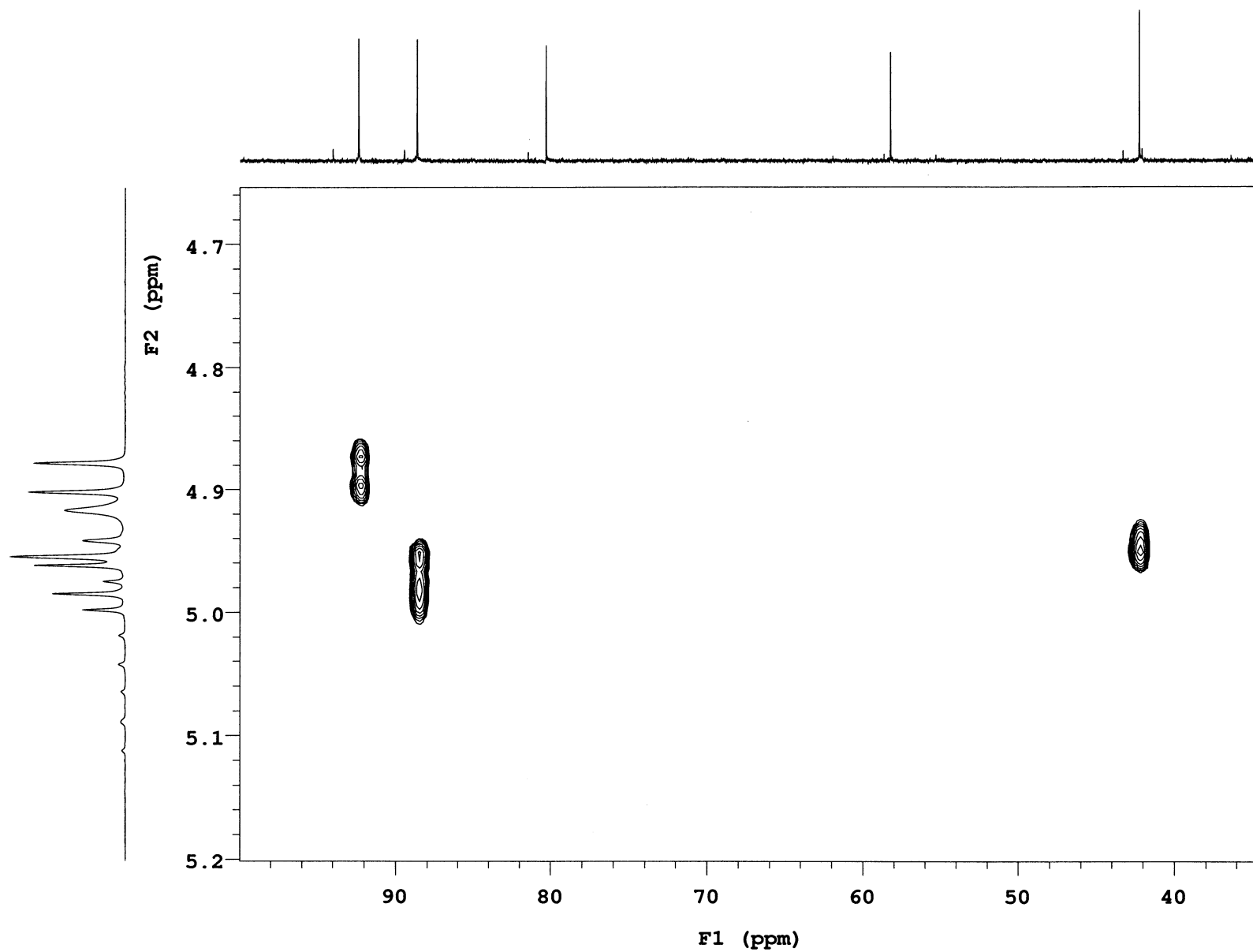
Sample Name **CHP-01-075-F5**
Date collected **2016-05-03**Pulse sequence **gHSQC**
Solvent **cd3cn**Temperature **25**
Spectrometer **Agilent-NMR-inova500**Study owner **vnmr2**
Operator **vnmr2**

Fig S138. COSY (CD3CN) of compound 3i.

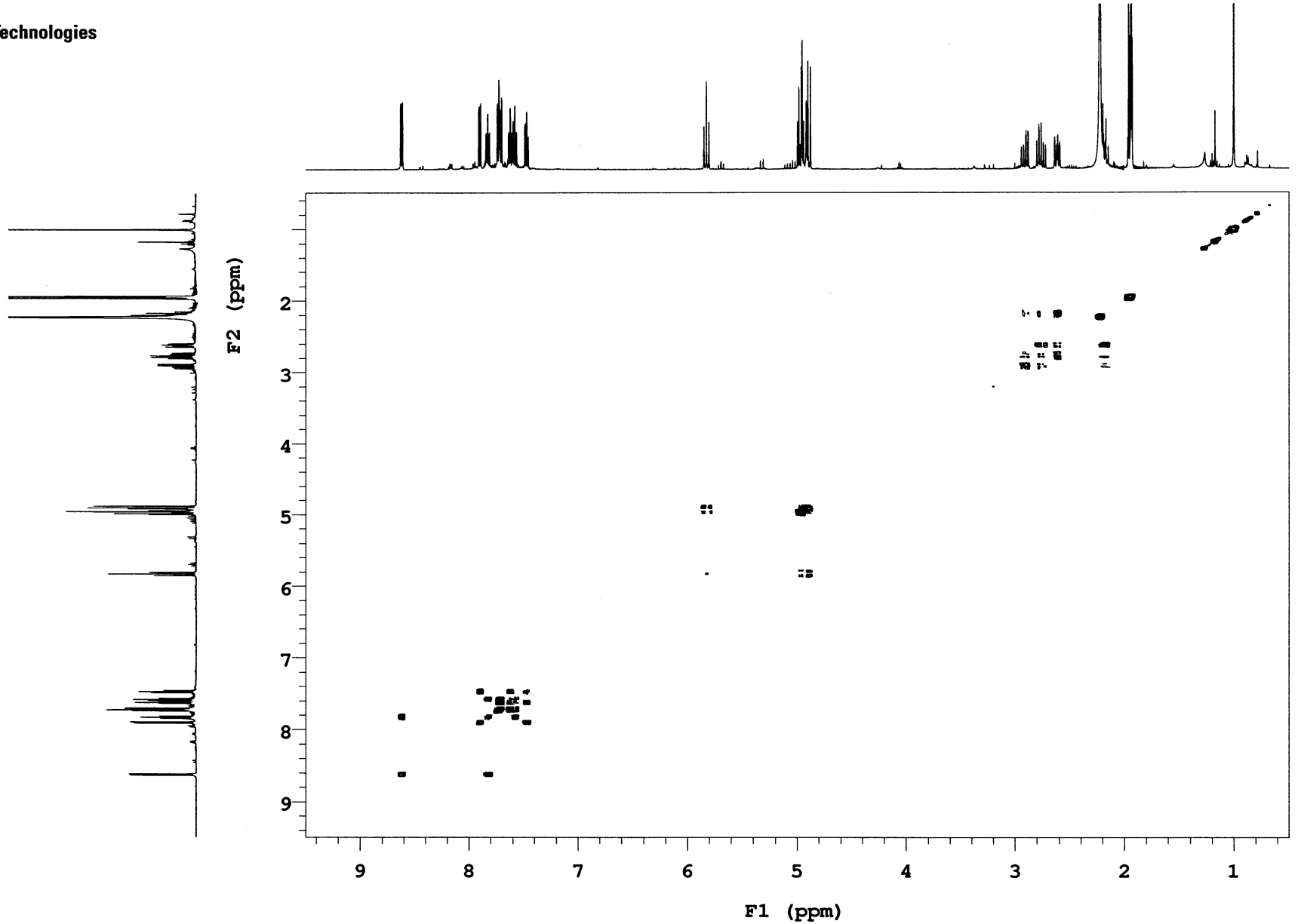
CHP-01-075-F5

Sample Name **CHP-01-075-F5**
Date collected **2016-05-03**

Pulse sequence **gCOSY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**



CHP-01-075-F5

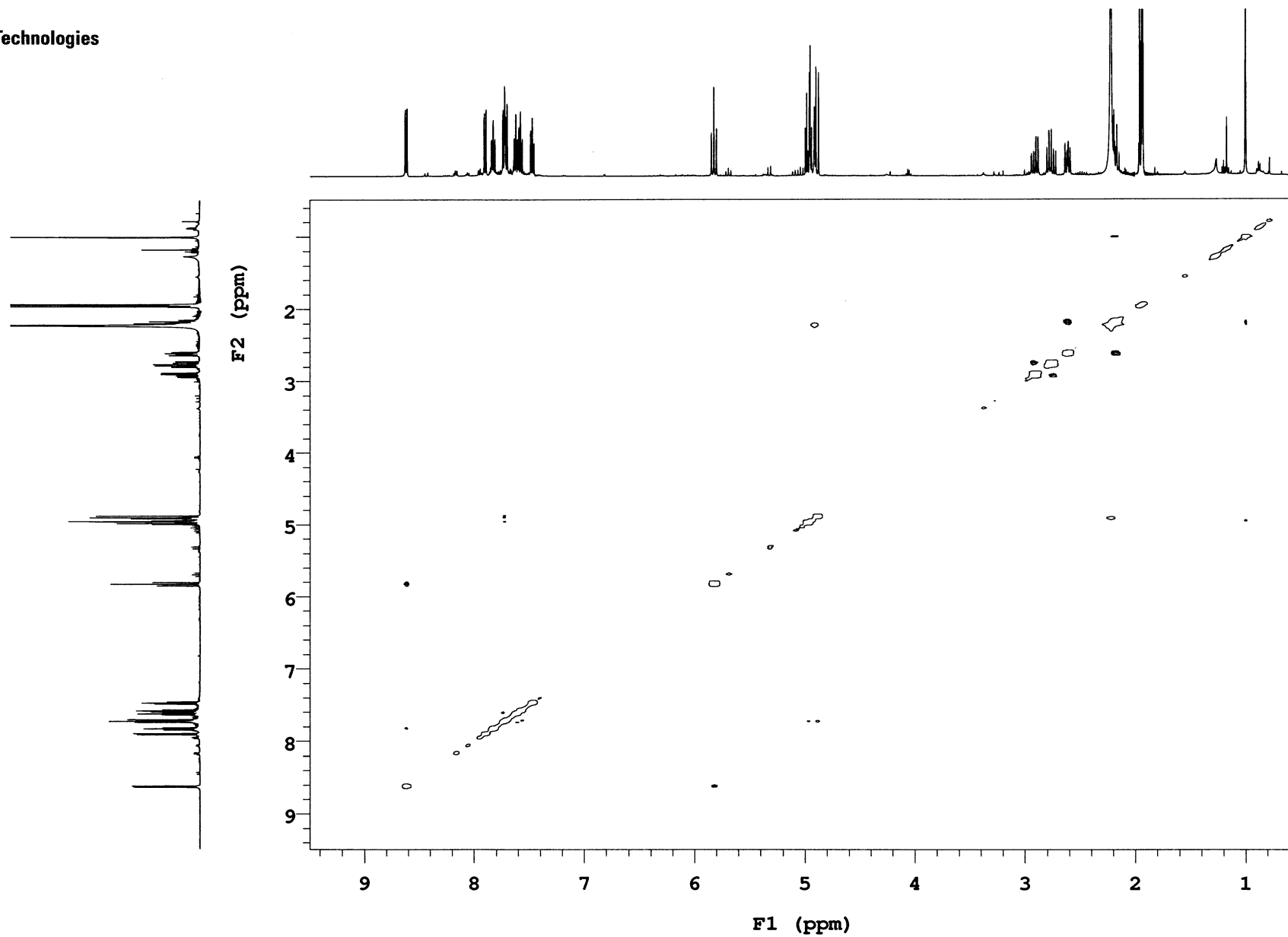
Sample Name **CHP-01-075-F5**
Date collected **2016-05-03**Pulse sequence **NOESY**
Solvent **cd3cn**Temperature **25**
Spectrometer **Agilent-NMR-inova500**Study owner **vnmr2**
Operator **vnmr2**

Fig S140. ¹H NMR (CD₃CN, 500 MHz) of compound 5i.

S140

CHP-8g

Sample Name **CHP-8g**
Date collected **2016-04-12**

Pulse sequence **PROTON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-Inova500**

Study owner **vnmr2**
Operator **vnmr2**

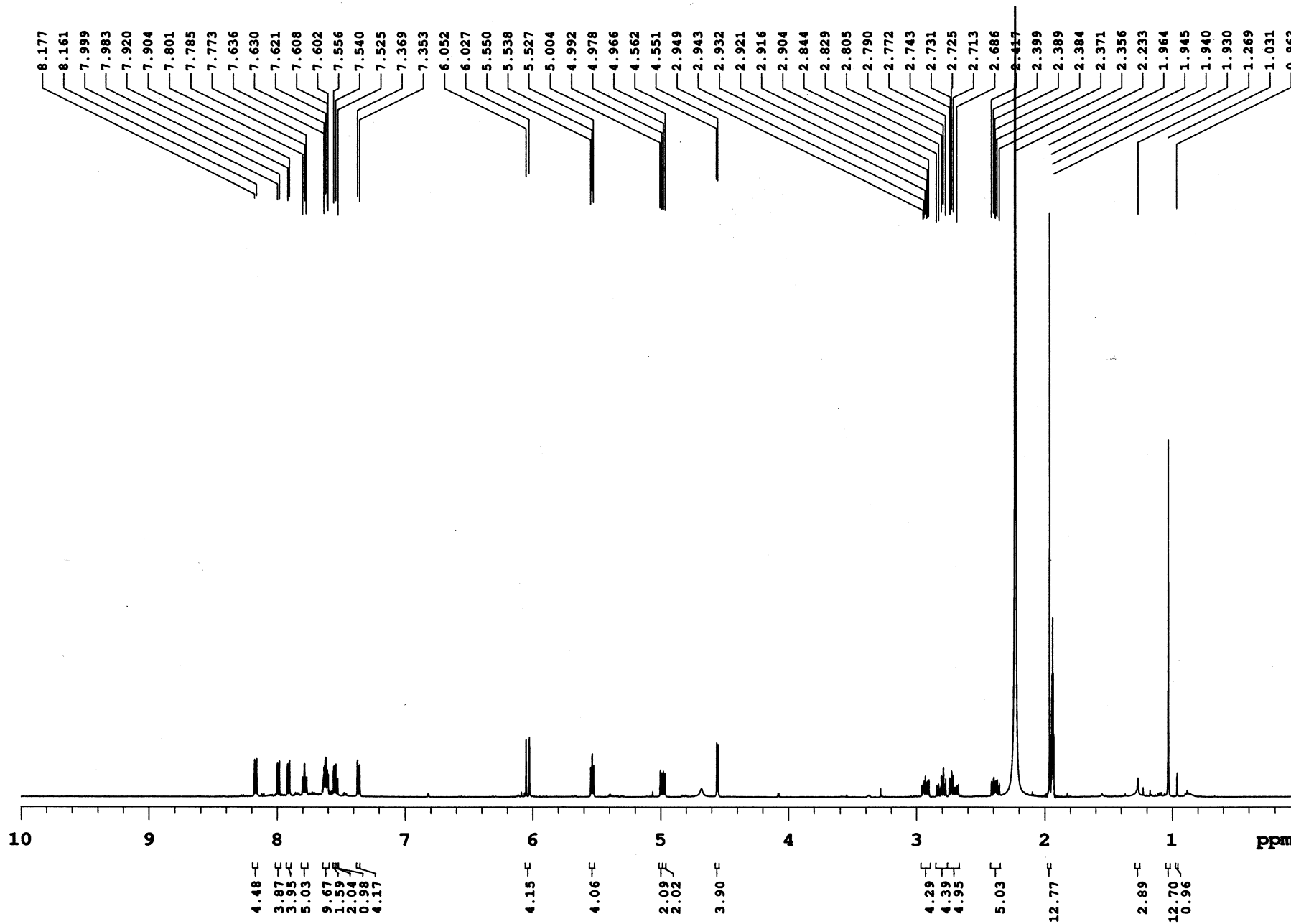


Fig S141. ¹³C NMR (CD₃CN, 125 MHz) of compound 5i.

S141

CHP-8g

Sample Name **CHP-8g**
Date collected **2016-04-12**

Pulse sequence **CARBON**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

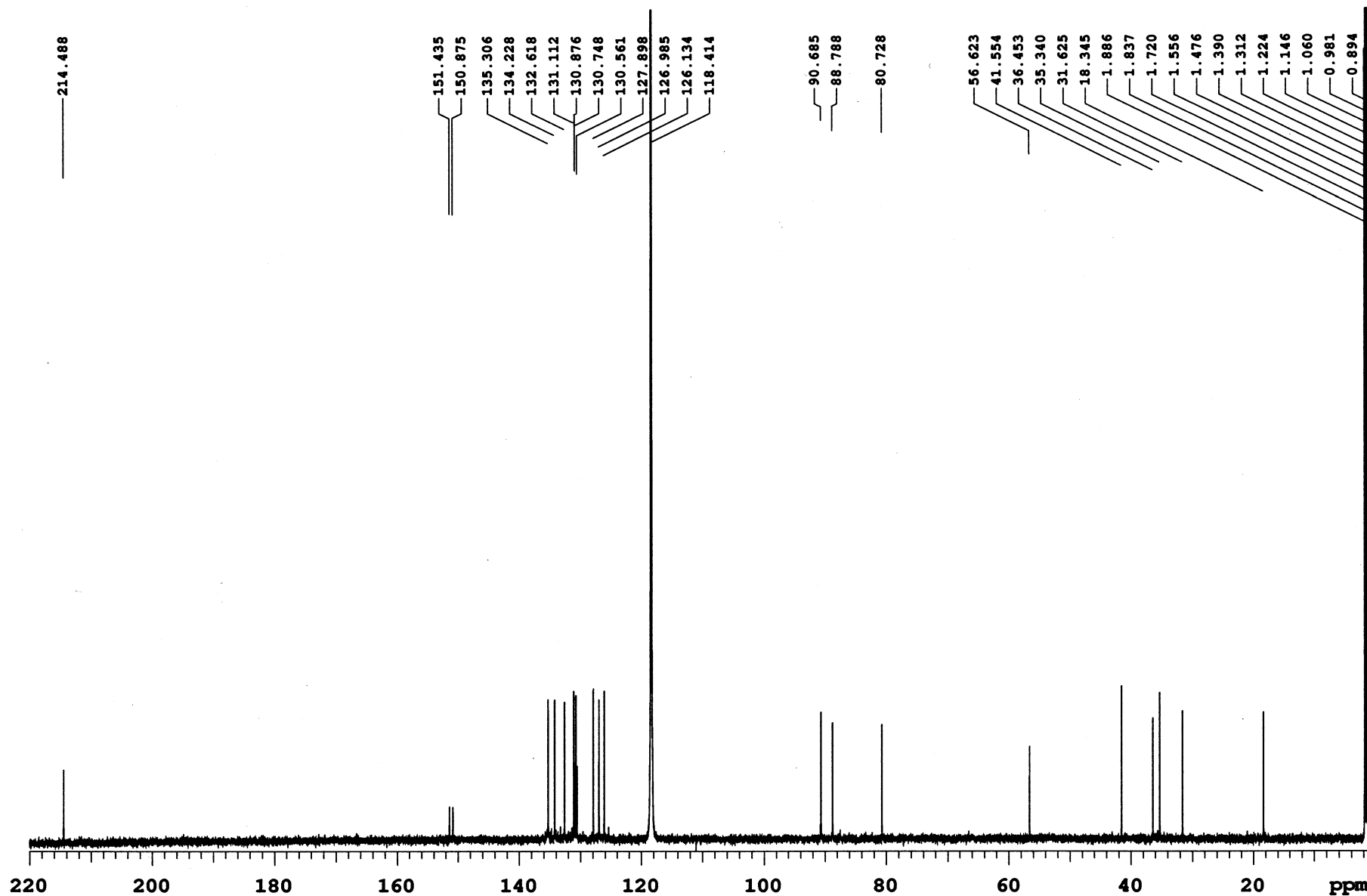


Fig S142. DEPT of compound 5i.

CHP-8g

Sample Name **CHP-8g**
Date collected **2016-04-13**

Pulse sequence **DEPT**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

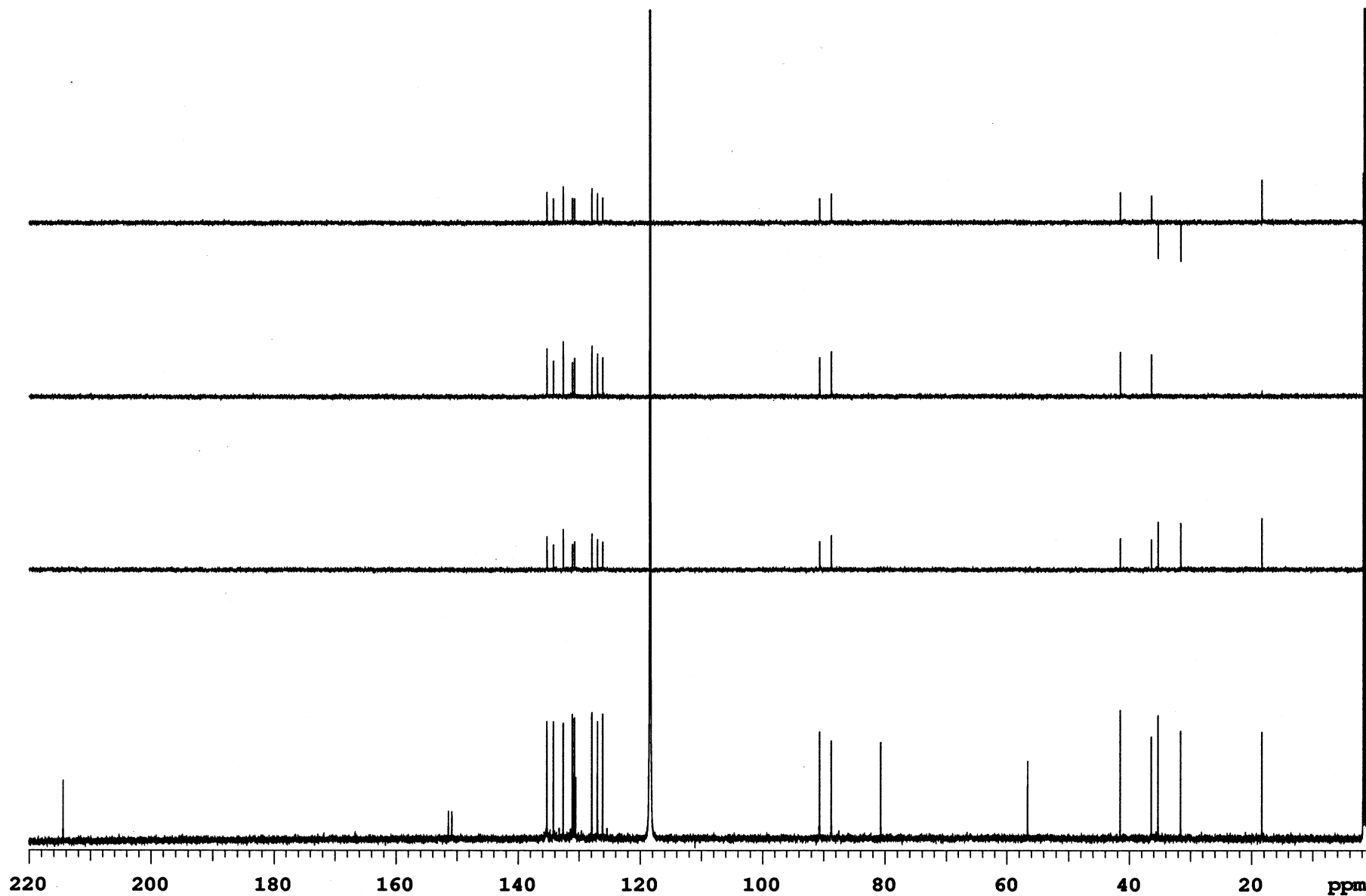


Fig S143. HSQC of compound 5i.

S143

CHP-8g

Sample Name **CHP-8g**
Date collected **2016-04-13**

Pulse sequence **gHSQC**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

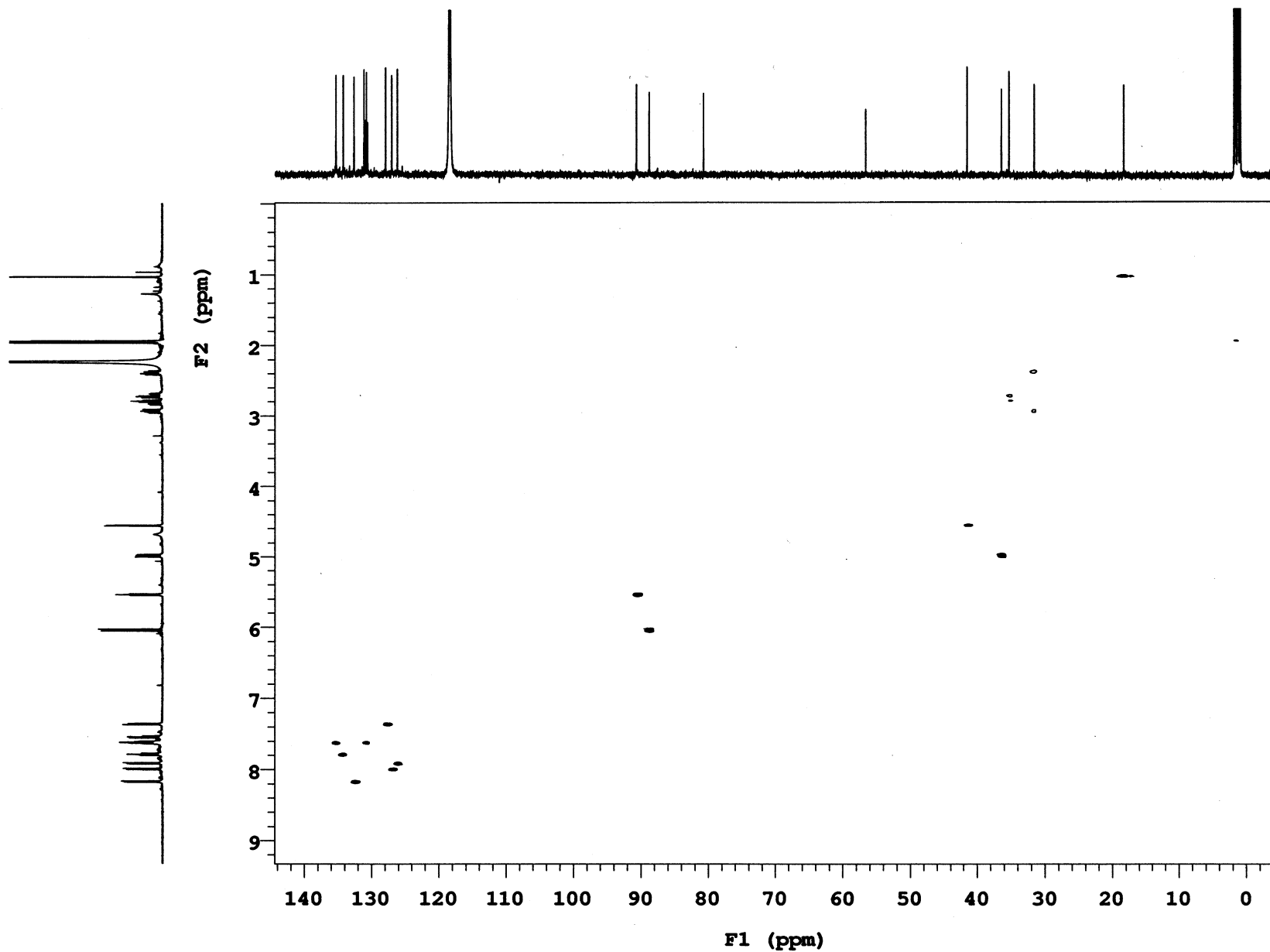


Fig S144. COSY of compound 5i.

S144

CHP-8g

Sample Name **CHP-8g**
Date collected **2016-04-13**

Pulse sequence **gCOSY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

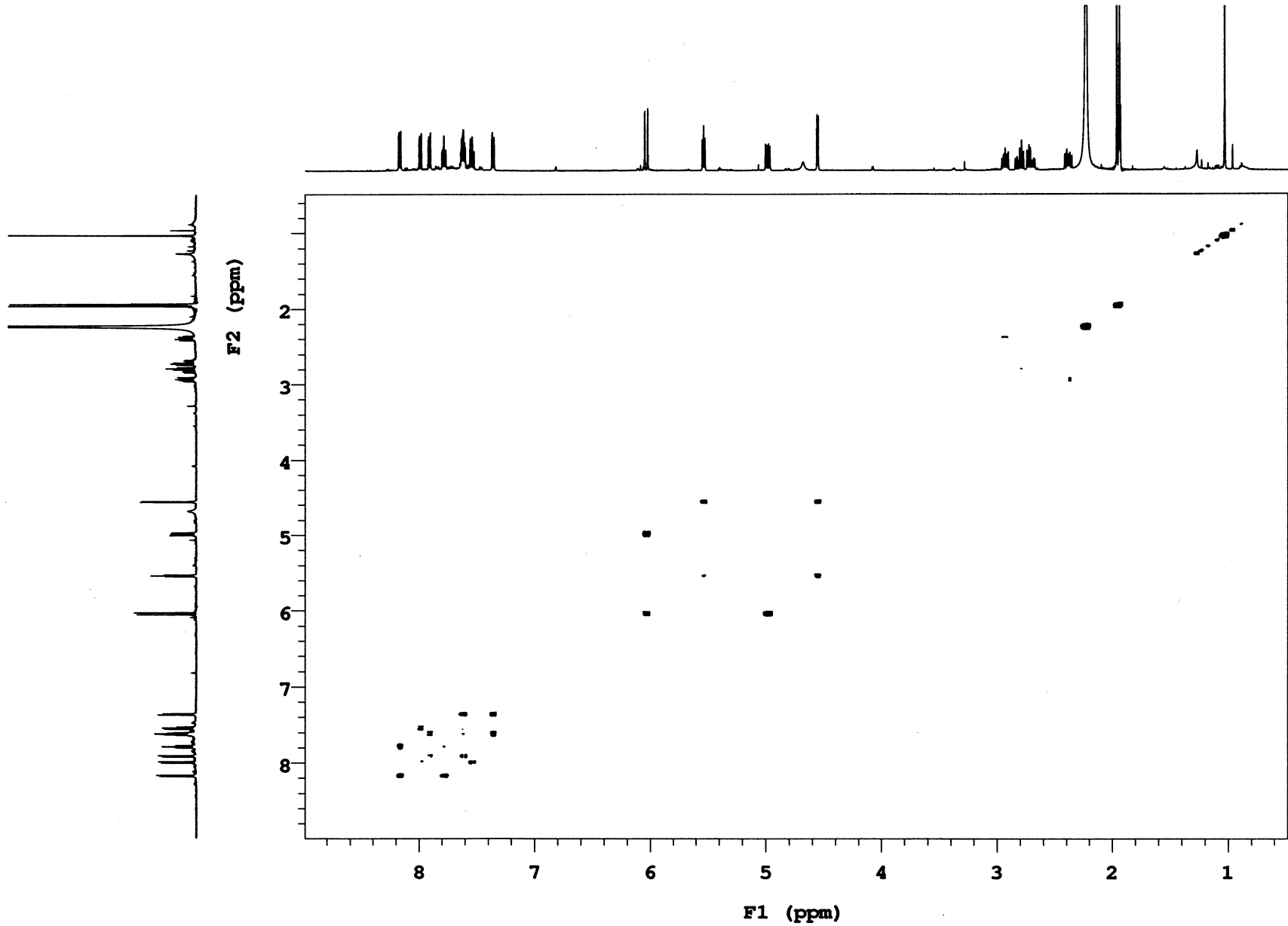


Fig S145. NOESY of compound 5i.

CHP-8g

Sample Name **CHP-8g**
Date collected **2016-04-13**

Pulse sequence **NOESY**
Solvent **cd3cn**

Temperature **25**
Spectrometer **Agilent-NMR-inova500**

Study owner **vnmr2**
Operator **vnmr2**

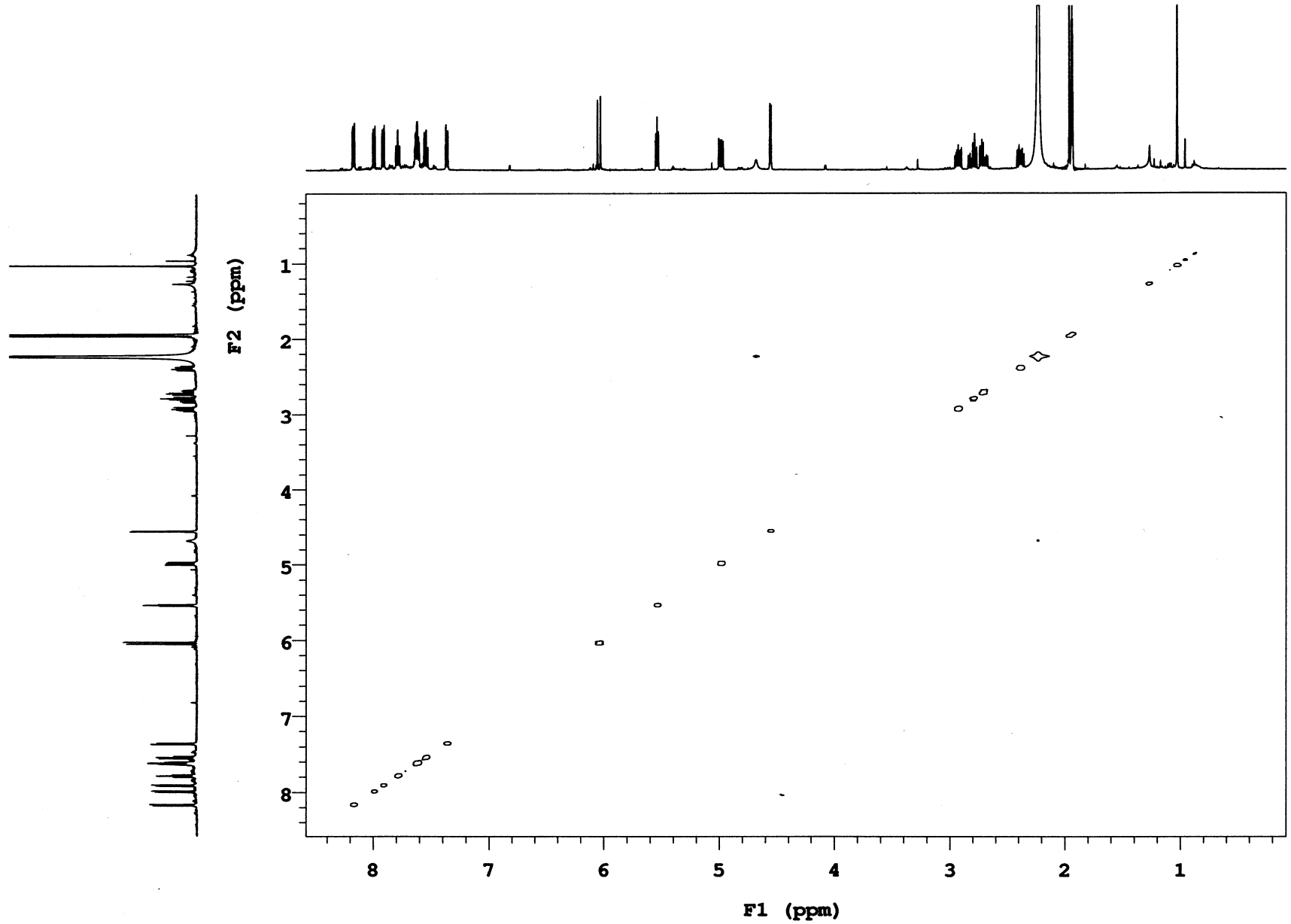
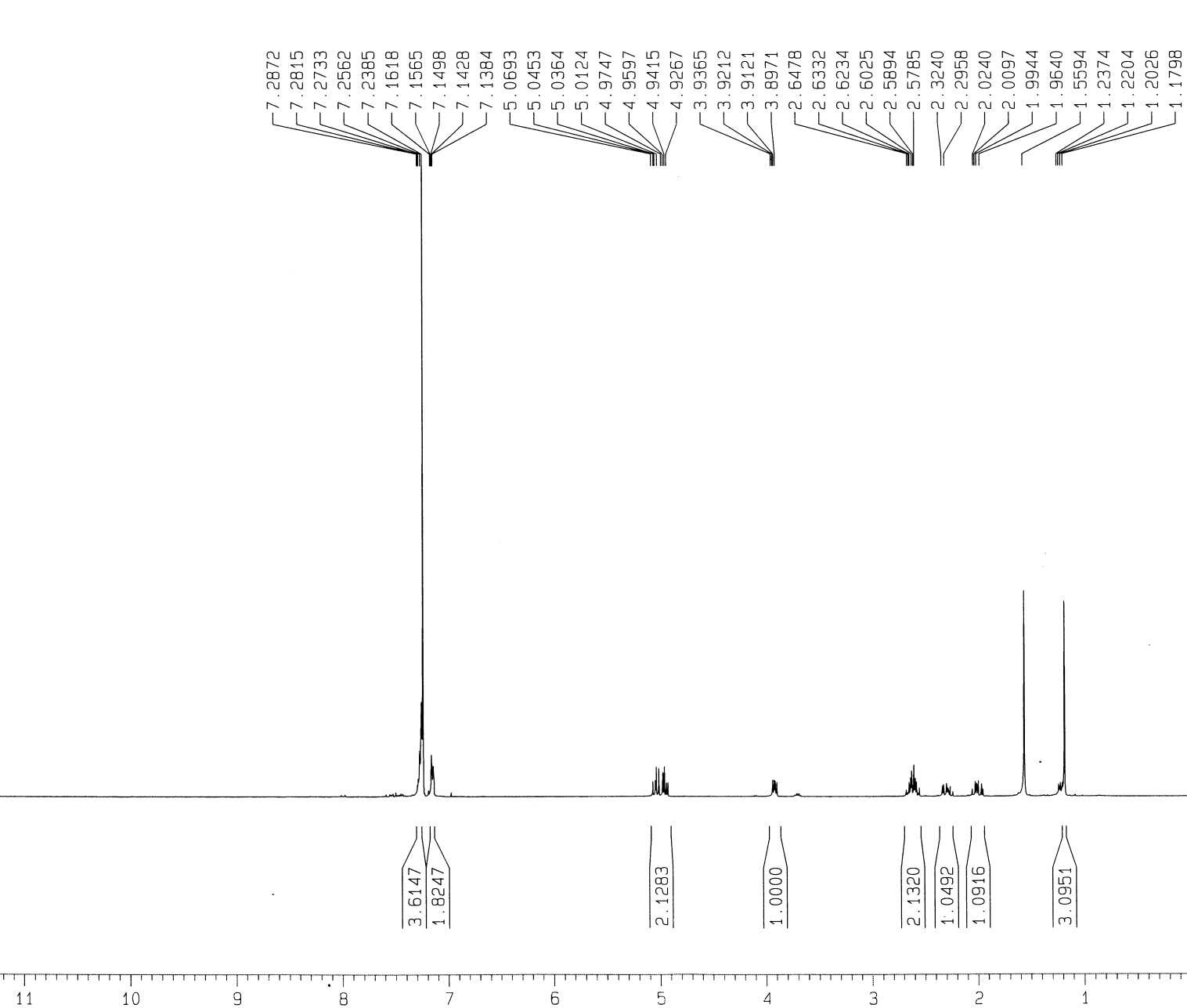


Fig S146. ¹H NMR (CDCl₃, 400 MHz) of compound 4a.

Current Data Parameters
 NAME CJW-2-20-top
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20141113
 Time 14.13
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zg30
 TD 16384
 SOLVENT CDCl3
 NS 32
 DS 0
 SWH 5995.204 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664756 sec
 RG 4597.6
 DW 83.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.50000000 sec

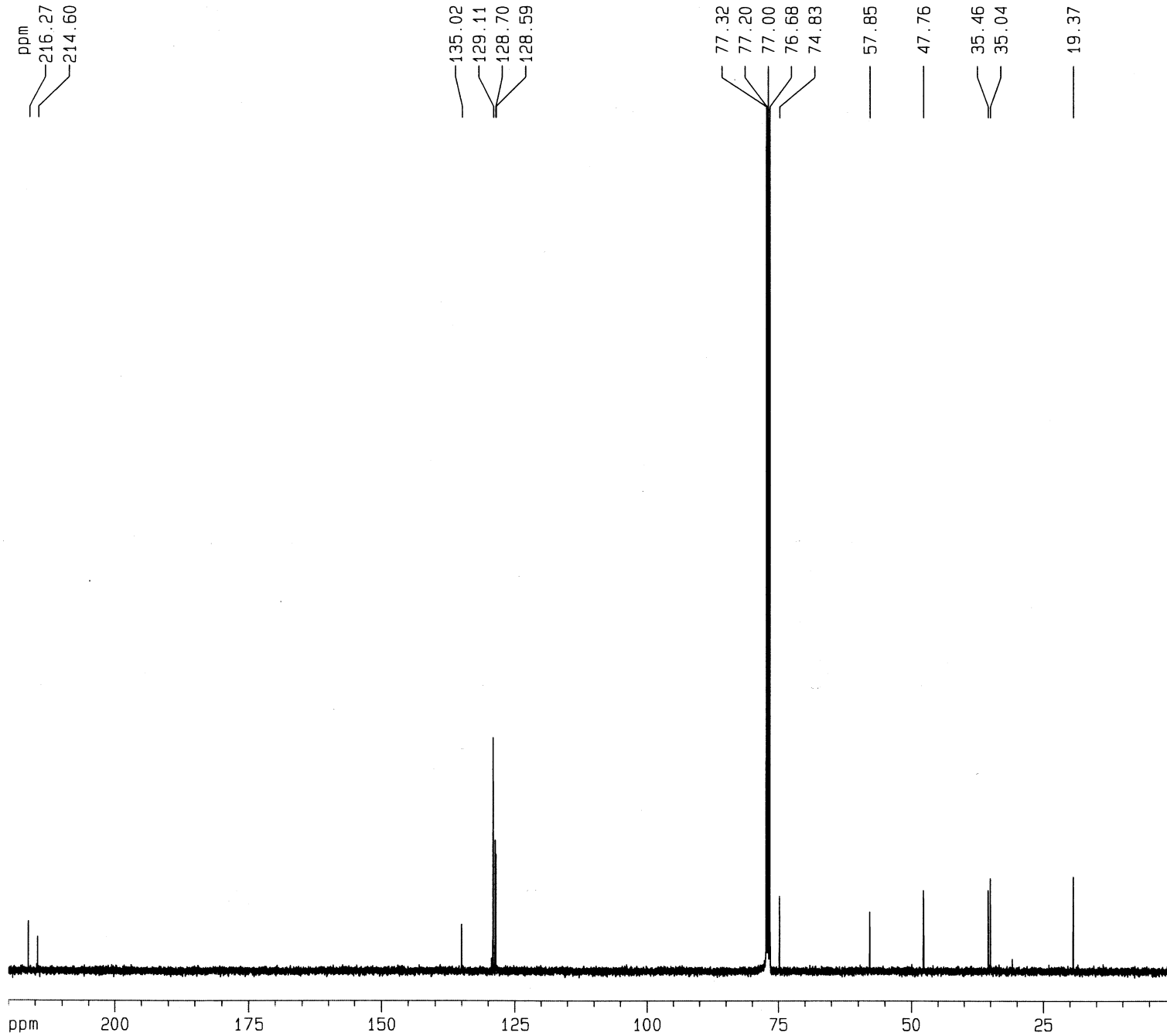
===== CHANNEL f1 =====
 NUC1 1H
 P1 11.90 usec
 PL1 -3.00 dB
 SF01 400.1326008 MHz

F2 - Processing parameters

SI 8192
 SF 400.1300179 MHz
 WDW EM
 SSB 0
 LB 0.10 Hz
 GB 0
 PC 1.00

1D NMR plot parameters

CX 21.50 cm
 F1P 12.000 ppm
 F1 4801.56 Hz
 F2P -0.000 ppm
 F2 -0.00 Hz
 PPMCM 0.55814 ppm/cm
 HZCM 223.32838 Hz/cm

Fig S147. ¹³C NMR (CDCl₃, 100 MHz) of compound 4a.

Current Data Parameters
 NAME CJW-2-20-top
 EXPNO 3
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20141114
 Time 4.37
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 5120
 DS 4
 SWH 25125.629 Hz
 FIDRES 0.383387 Hz
 AQ 1.3042164 sec
 RG 256
 DW 19.900 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

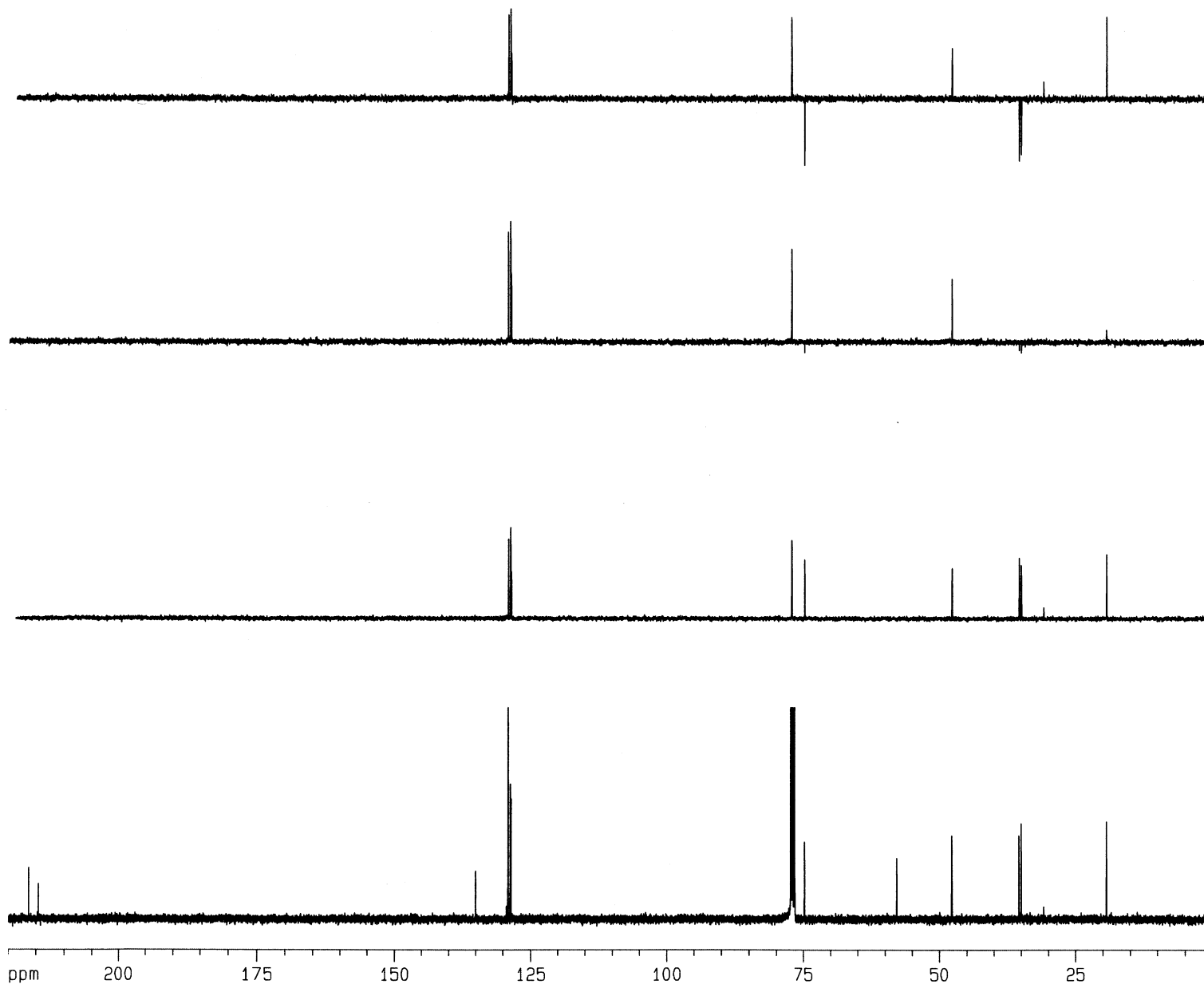
===== CHANNEL f1 =====
 NUC1 13C
 P1 10.20 usec
 PL1 0.00 dB
 SF01 100.6237959 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 -3.00 dB
 PL12 14.50 dB
 PL13 17.50 dB
 SF02 400.1326008 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127715 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 CY 20.00 cm
 F1P 220.000 ppm
 F1 22134.81 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 11.00000 ppm/cm
 HZCM 1106.74048 Hz/cm

Fig S148. DEPT of compound 4a.



Current Data Parameters

NAME CJW-2-20-top
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters

Date_ 20141114
Time 4.37
INSTRUM spect
PROBHD 5 mm GNP 1H
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 5120
DS 4
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 256
DW 19.900 usec
DE 6.50 usec
TE 300.0 K
D1 2.0000000 sec
d11 0.0300000 sec
d12 0.0000200 sec

===== CHANNEL f1 =====

NUC1 13C
P1 10.20 usec
PL1 0.00 dB
SF01 100.6237959 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 -3.00 dB
PL12 14.50 dB
PL13 17.50 dB
SF02 400.1326008 MHz

F2 - Processing parameters

SI 32768
SF 100.6127715 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.40

1D NMR plot parameters

CX 20.00 cm
CY 20.00 cm
F1P 220.000 ppm
F1 22134.81 Hz
F2P 0.000 ppm
F2 0.00 Hz
PPMCM 11.00000 ppm/cm
HZCM 1106.74048 Hz/cm

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 03/04/2016

Reported Date and Time: 03/06/2016

01:33 PM

02:14 PM

Processed Date and Time: 03/06/2016

02:13 PM

Data Path: D:\Arun\DATA\0427\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0427

Application(data): Arun

Vial Number: 1

Sample Name: CHP-01-50

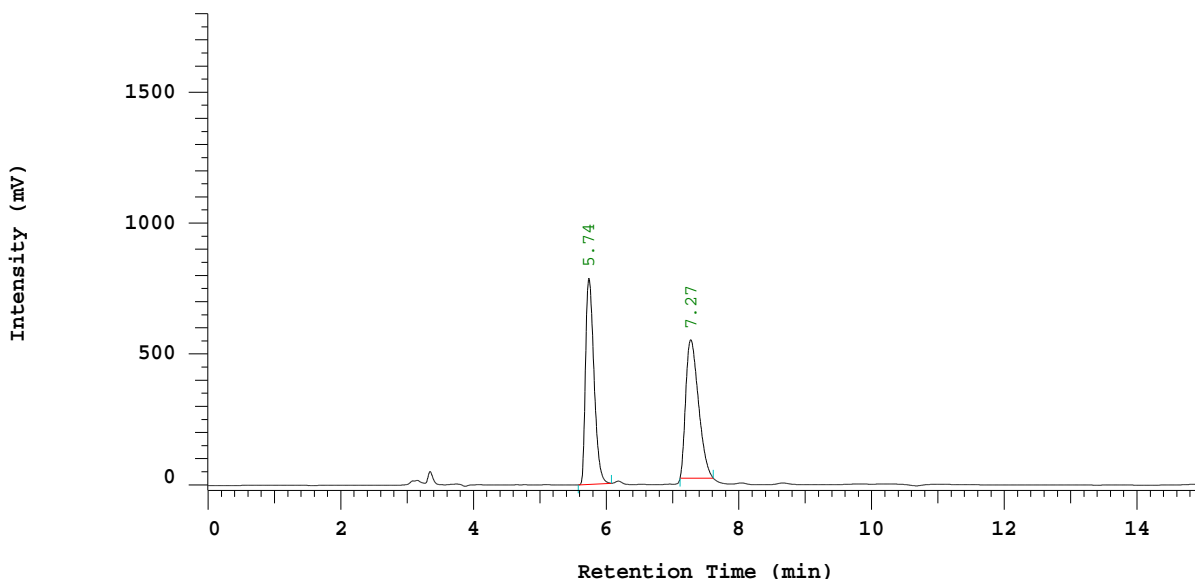
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 13%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 217 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 217 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.74	6951809	787157	49.727
2	7.27	7028108	528549	50.273
		13979917	1315706	100.000

Peak rejection level: 5000

Fig S149. HPLC analysis of the compound **3a** obtained, (Table 1, entry 6)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 03/04/2016
01:58 PMReported Date and Time: 03/06/2016
02:09 PMProcessed Date and Time: 03/06/2016
02:08 PM

Data Path: D:\Arun\DATA\0428\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0428

Application(data): Arun

Vial Number: 1

Sample Name: CHP-01-51

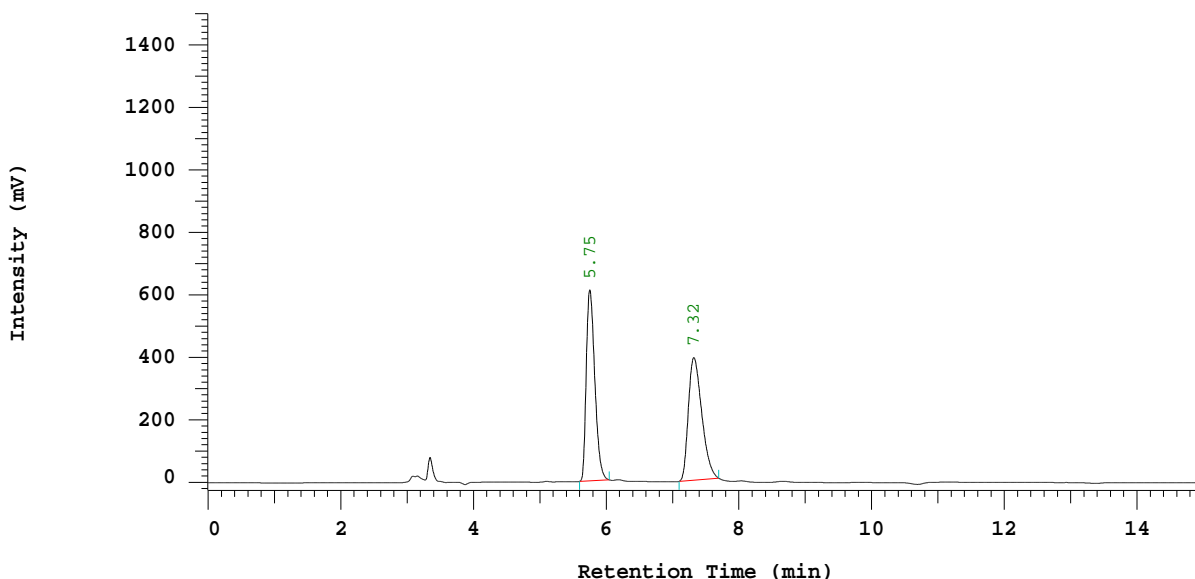
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 13%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 217 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 217 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.75	5453257	611259	49.736
2	7.32	5511103	392354	50.264
		10964360	1003613	100.000

Peak rejection level: 5000

Fig S150. HPLC analysis of the compound **3a** obtained, (Table 1, entry 7)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 03/04/2016
02:57 PMReported Date and Time: 03/06/2016
02:06 PMProcessed Date and Time: 03/06/2016
02:06 PM

Data Path: D:\Arun\DATA\0429\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0429

Application(data): Arun

Vial Number: 1

Sample Name: CHP-01-52

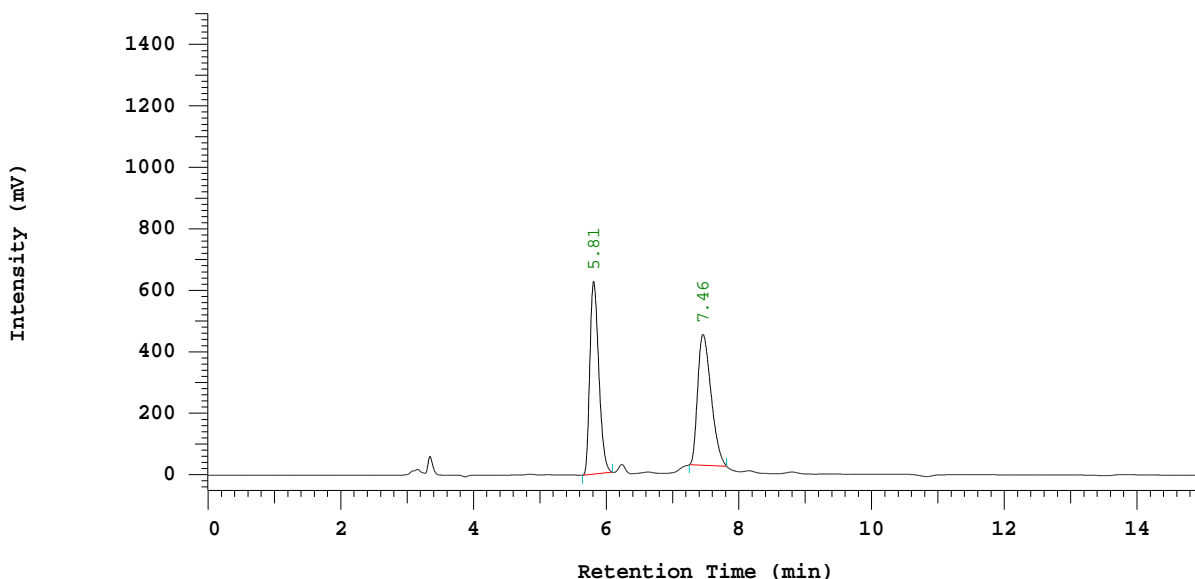
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 13%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 217 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 217 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.81	5885371	626405	49.589
2	7.46	5983016	425624	50.411
		11868387	1052029	100.000

Peak rejection level: 5000

Fig S151. HPLC analysis of the compound **3a** obtained, (Table 1, entry 8)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 03/03/2016
04:10 PMReported Date and Time: 03/03/2016
04:40 PMProcessed Date and Time: 03/03/2016
04:39 PM

Data Path: D:\Arun\DATA\0426\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0426

Application(data): Arun

Vial Number: 1

Sample Name: AR-03-173 (Racemic)

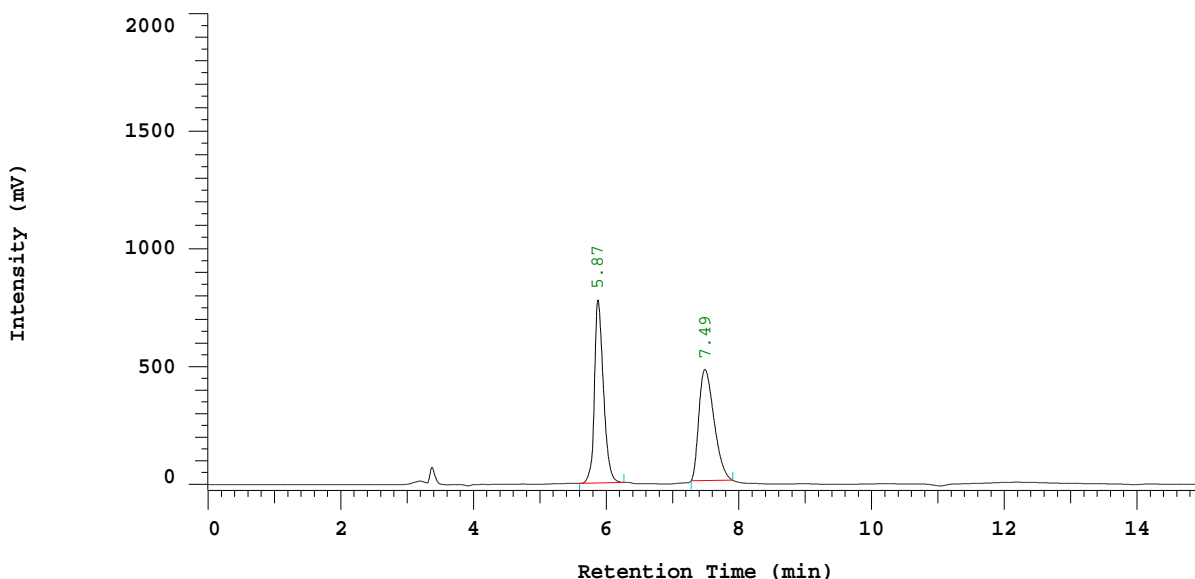
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 13%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 217 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 217 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.87	7350442	777507	49.556
2	7.49	7482187	472766	50.444
		14832629	1250273	100.000

Peak rejection level: 5000

Fig S152. HPLC analysis of the compound **3a** obtained, (Table 1, entry 9)

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 06/23/2016
05:55 PM

Reported Date and Time: 07/01/2016
01:13 PM

Processed Date and Time: 07/01/2016
01:13 PM

Data Path: D:\Arun\DATA\0469\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0469

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crystal 3)

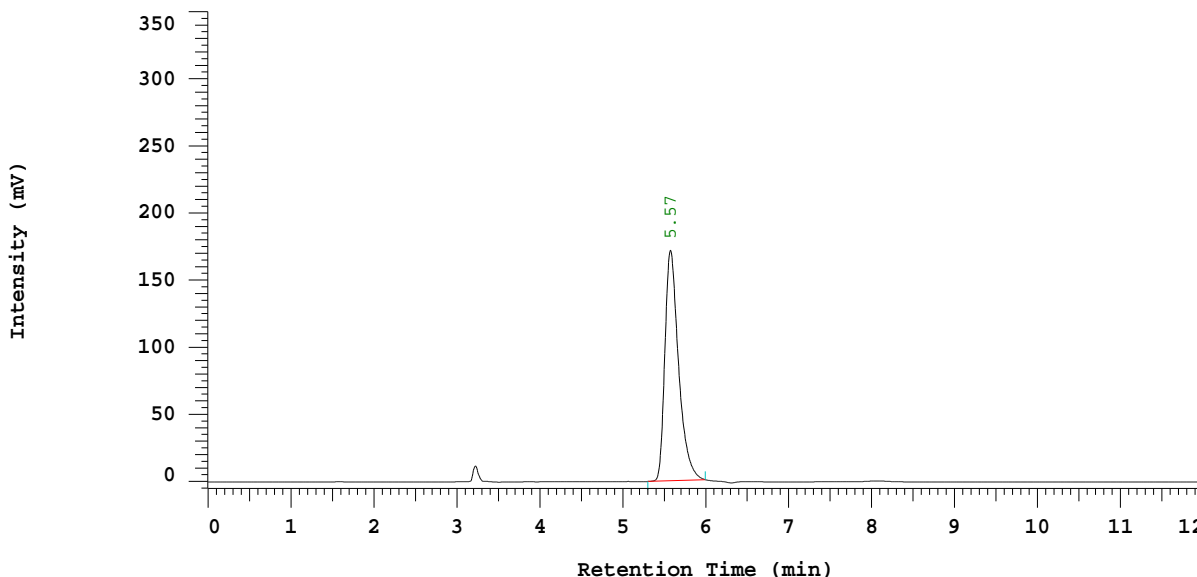
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.57	1965962	171710	100.000
		1965962	171710	100.000

Peak rejection level: 5000

Fig S153. HPLC analysis of a crystal of the (+)-3i obtained (crystal count number 1)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/24/2016
12:22 PMReported Date and Time: 07/01/2016
01:07 PMProcessed Date and Time: 07/01/2016
01:07 PM

Data Path: D:\Arun\DATA\0474\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0474

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crys 1)

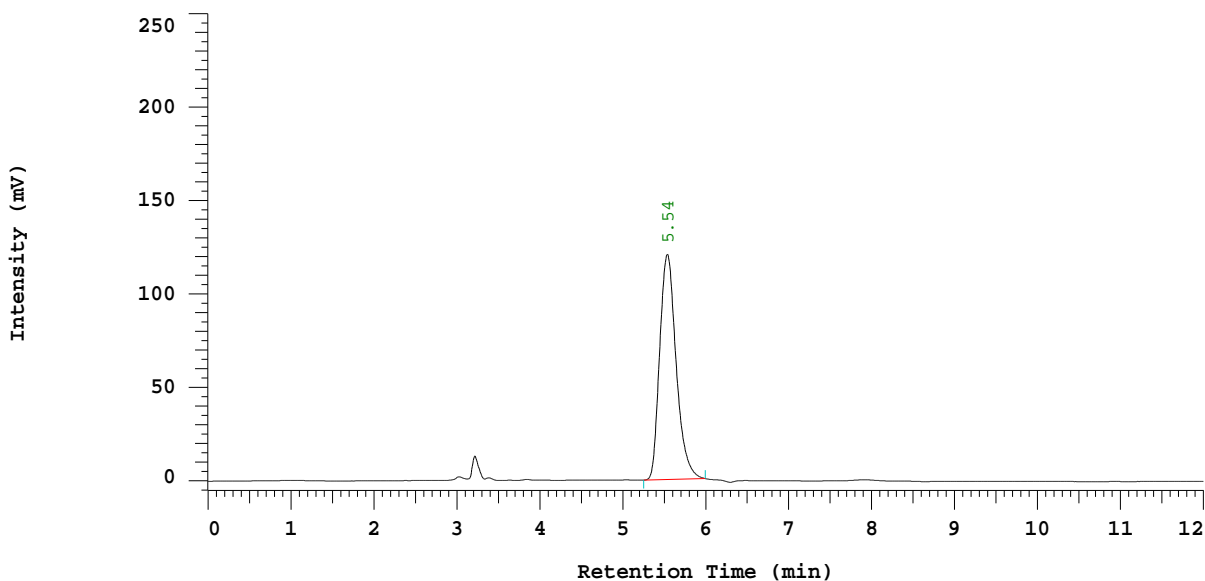
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.54	1660111	120449	100.000
		1660111	120449	100.000

Peak rejection level: 5000

Fig S154. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 2)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/24/2016
12:50 PMReported Date and Time: 07/01/2016
01:04 PMProcessed Date and Time: 07/01/2016
01:04 PM

Data Path: D:\Arun\DATA\0476\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0476

Application(data): Arun

Vial Number: 3

Sample Name: CHP-8g-F2 (Crys 3)

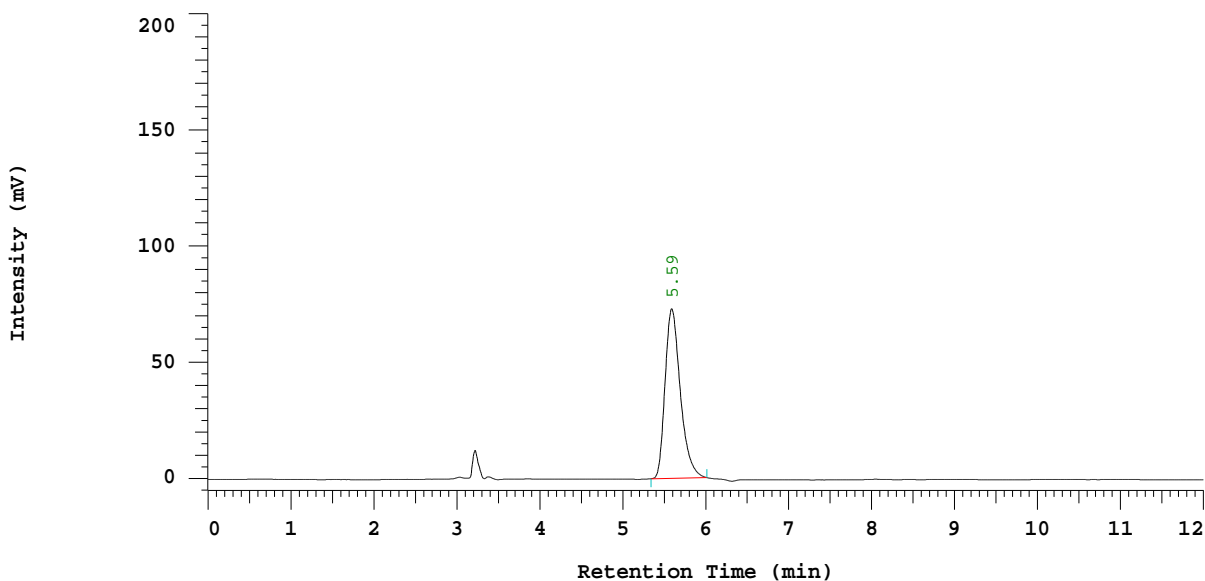
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.59	932401	72897	100.000
		932401	72897	100.000

Peak rejection level: 5000

Fig S155. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 3)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/26/2016
04:01 PMReported Date and Time: 07/01/2016
12:55 PMProcessed Date and Time: 07/01/2016
12:54 PM

Data Path: D:\Arun\DATA\0480\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0480

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crys 1)

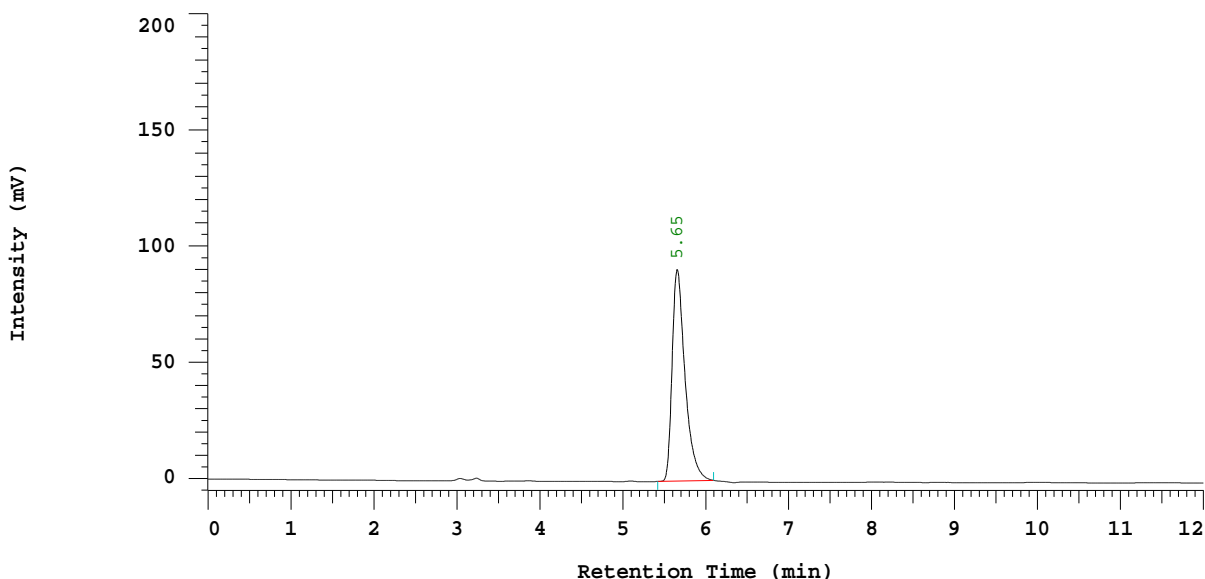
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.65	982521	90981	100.000
		982521	90981	100.000

Peak rejection level: 5000

Fig S156. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 4)

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 06/26/2016
04:15 PM

Reported Date and Time: 07/01/2016
12:53 PM

Processed Date and Time: 07/01/2016
12:53 PM

Data Path: D:\Arun\DATA\0481\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0481

Application(data): Arun

Vial Number: 2

Sample Name: CHP-8g-F2 (Crys 2)

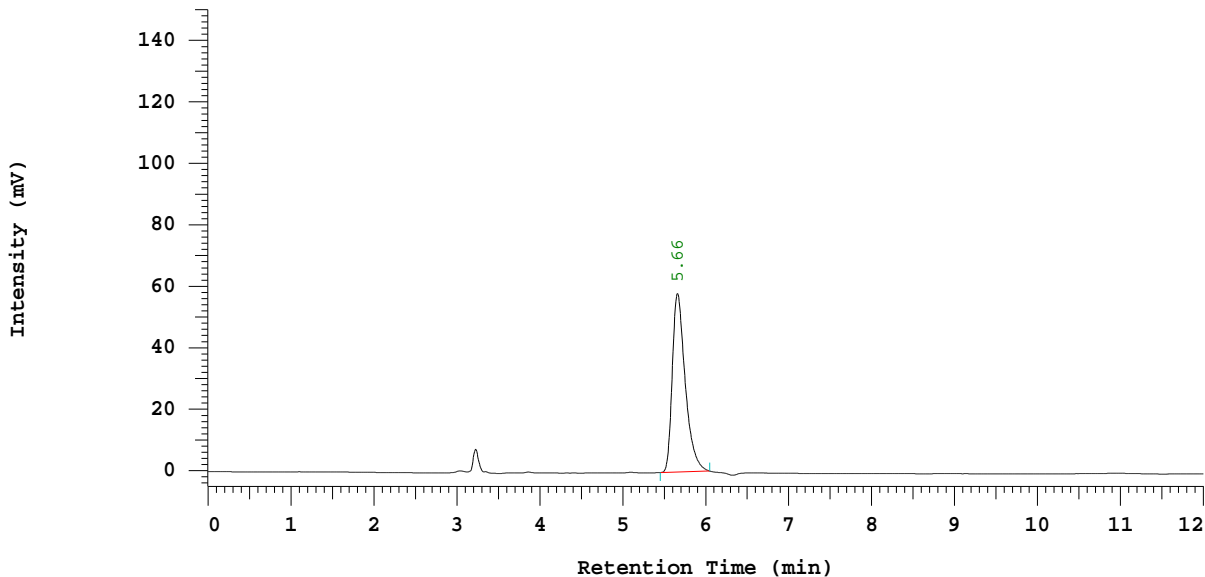
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.66	629116	58061	100.000
		629116	58061	100.000

Peak rejection level: 5000

Fig S157. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 5)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/26/2016
04:28 PMReported Date and Time: 07/01/2016
12:52 PMProcessed Date and Time: 07/01/2016
12:51 PM

Data Path: D:\Arun\DATA\0482\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0482

Application(data): Arun

Vial Number: 3

Sample Name: CHP-8g-F2 (Crys 3)

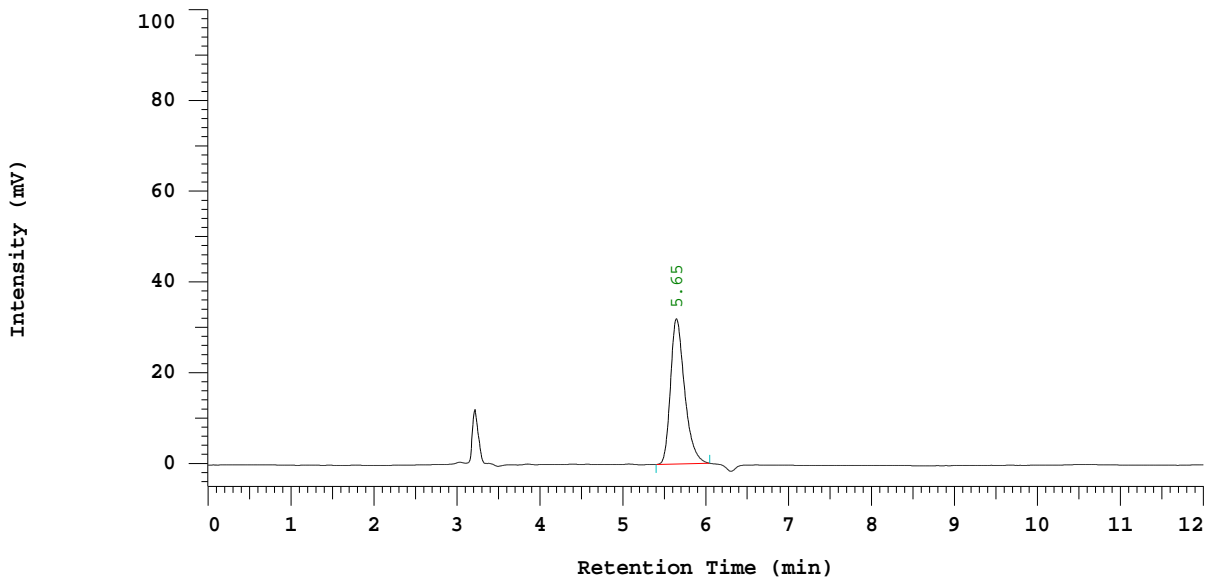
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.65	378230	32017	100.000
		378230	32017	100.000

Peak rejection level: 5000

Fig S158. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 6)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/27/2016
04:18 PMReported Date and Time: 07/01/2016
12:44 PMProcessed Date and Time: 07/01/2016
12:44 PM

Data Path: D:\Arun\DATA\0486\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0486

Application(data): Arun

Vial Number: 2

Sample Name: CHP-8g-F2 (Crys 2)

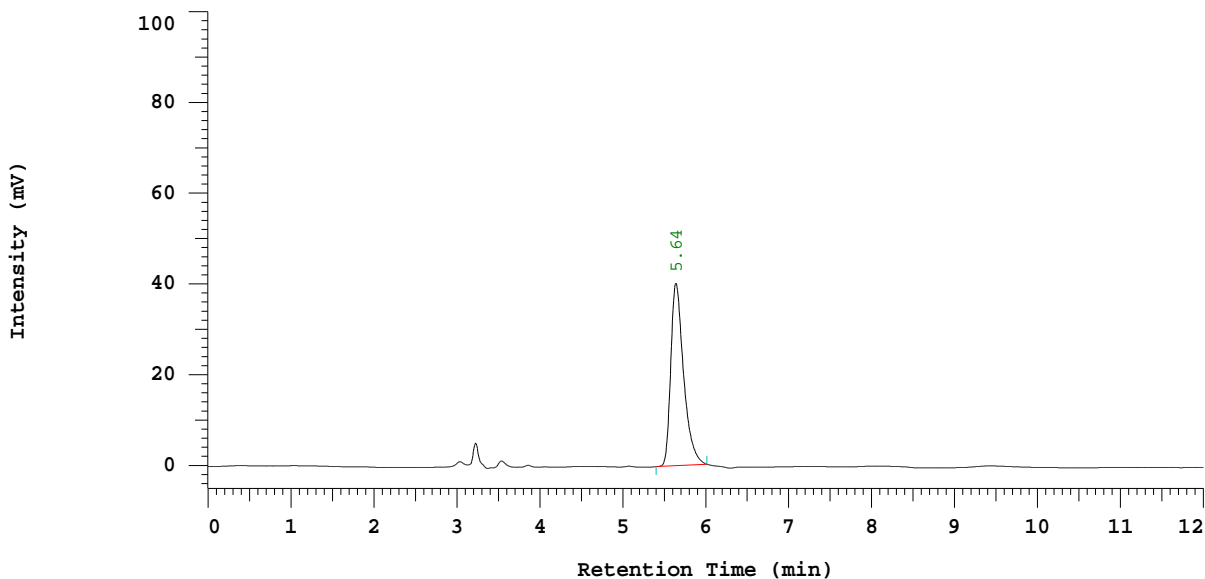
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.64	431230	40210	100.000
		431230	40210	100.000

Peak rejection level: 5000

Fig S159. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 7)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/27/2016
06:00 PMReported Date and Time: 07/01/2016
12:32 PMProcessed Date and Time: 07/01/2016
12:32 PM

Data Path: D:\Arun\DATA\0493\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0493

Application(data): Arun

Vial Number: 4

Sample Name: CHP-8g-F2 (Crys 4)

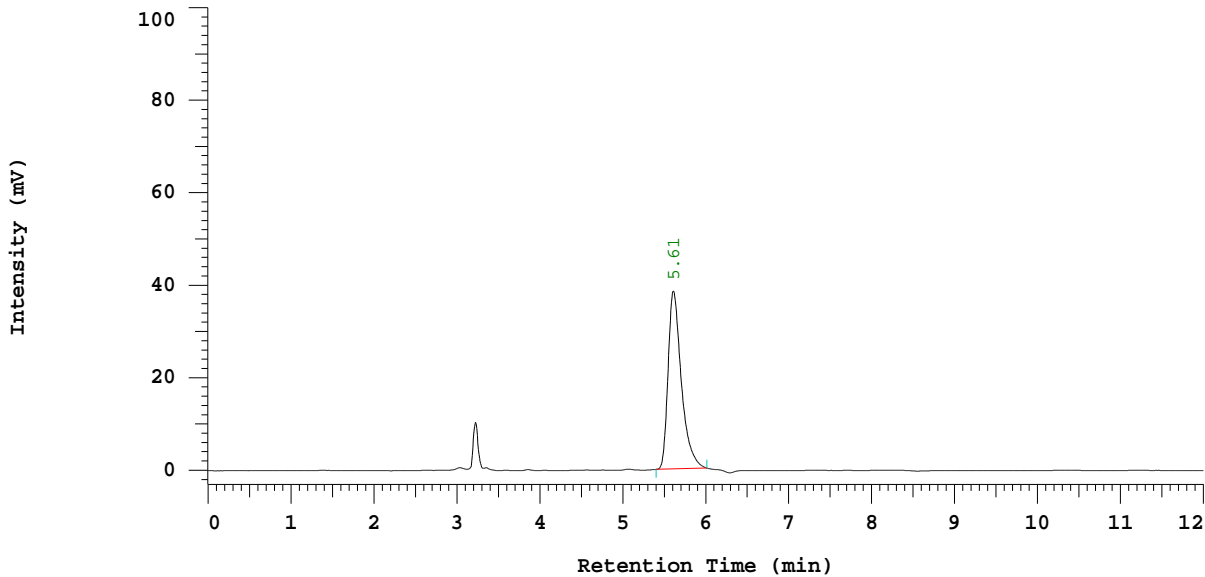
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.61	426716	38449	100.000
		426716	38449	100.000

Peak rejection level: 5000

Fig S160. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 8)

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 06/27/2016
06:16 PM

Reported Date and Time: 07/01/2016
12:30 PM

Processed Date and Time: 07/01/2016
12:29 PM

Data Path: D:\Arun\DATA\0494\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0494

Application(data): Arun

Vial Number: 5

Sample Name: CHP-8g-F2 (Crys 5)

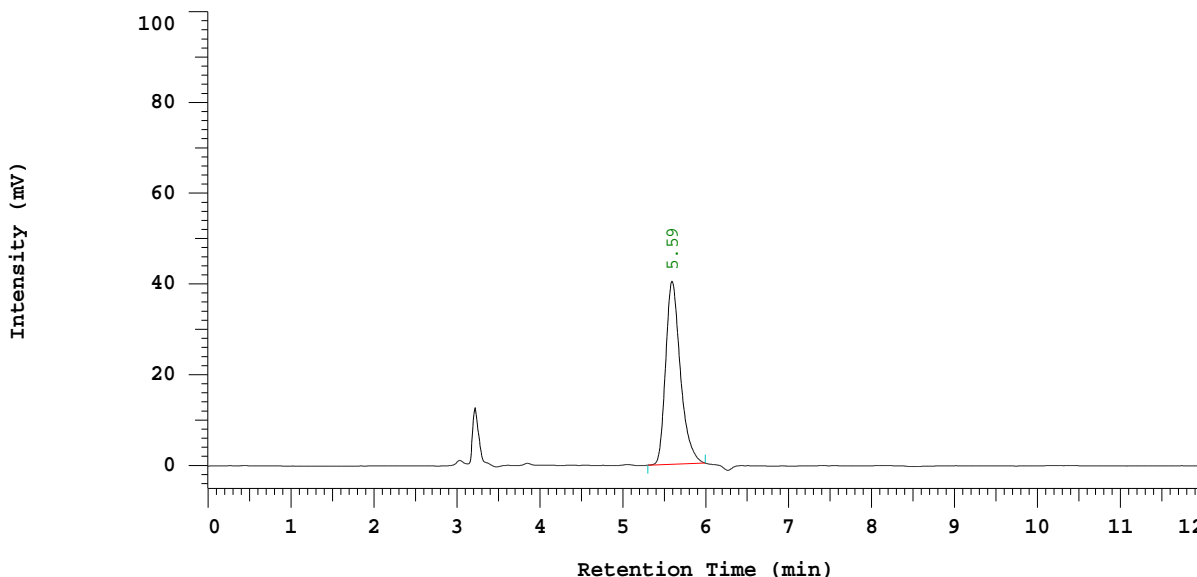
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.59	498296	40384	100.000
		498296	40384	100.000

Peak rejection level: 5000

Fig S161. HPLC analysis of a crystal of the (+)-3i obtained (crystal count number 9)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/29/2016
04:27 PMReported Date and Time: 07/01/2016
12:23 PMProcessed Date and Time: 07/01/2016
12:23 PM

Data Path: D:\Arun\DATA\0499\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0499

Application(data): Arun

Vial Number: 3

Sample Name: CHP-8g-F2 (Crys 3)

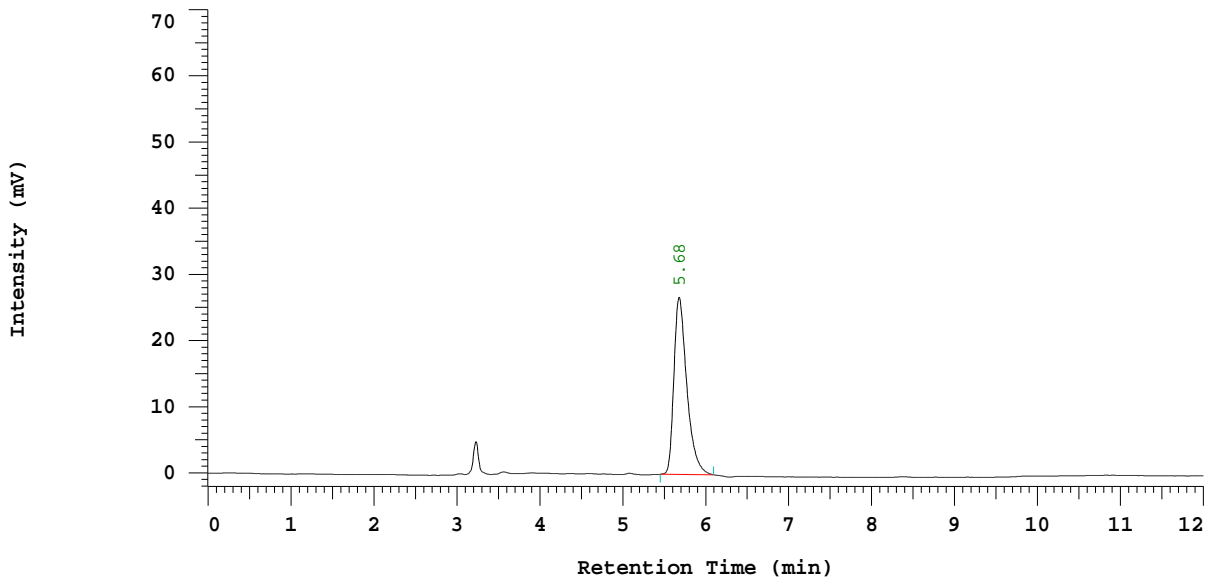
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.68	289629	26775	100.000
		289629	26775	100.000

Peak rejection level: 5000

Fig S162. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 10)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/29/2016
04:40 PMReported Date and Time: 07/01/2016
12:22 PMProcessed Date and Time: 07/01/2016
12:22 PM

Data Path: D:\Arun\DATA\0500\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0500

Application(data): Arun

Vial Number: 4

Sample Name: CHP-8g-F2 (Crys 4)

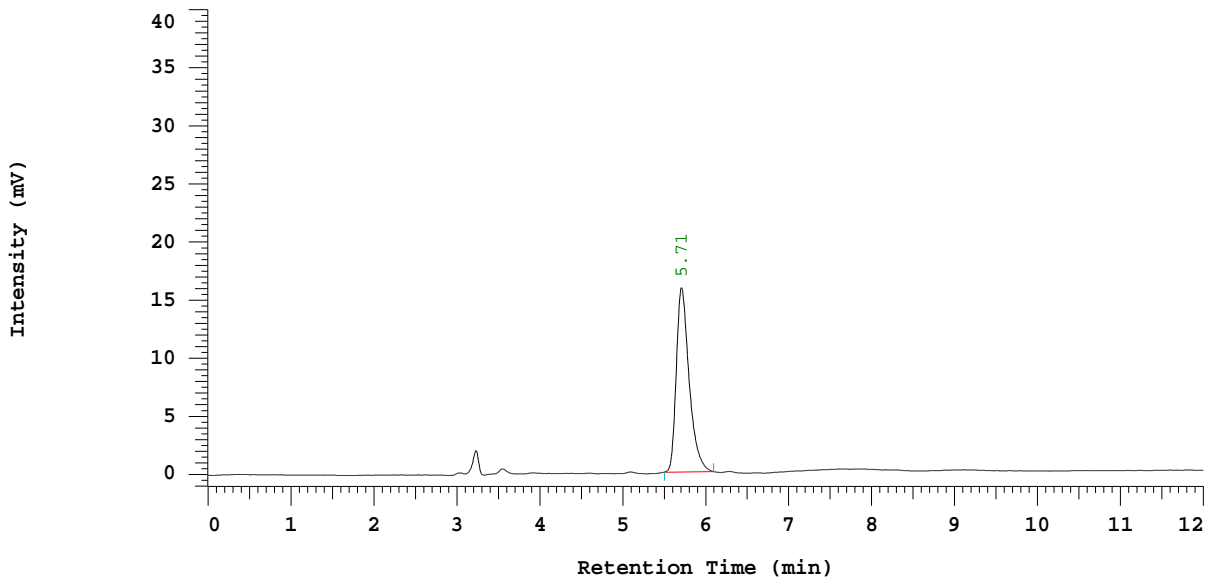
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.71	170488	15853	100.000
		170488	15853	100.000

Peak rejection level: 5000

Fig S163. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 11)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/29/2016
05:34 PMReported Date and Time: 07/01/2016
12:15 PMProcessed Date and Time: 07/01/2016
12:14 PM

Data Path: D:\Arun\DATA\0504\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0504

Application(data): Arun

Vial Number: 3

Sample Name: CHP-8g-F2 (Crys 3)

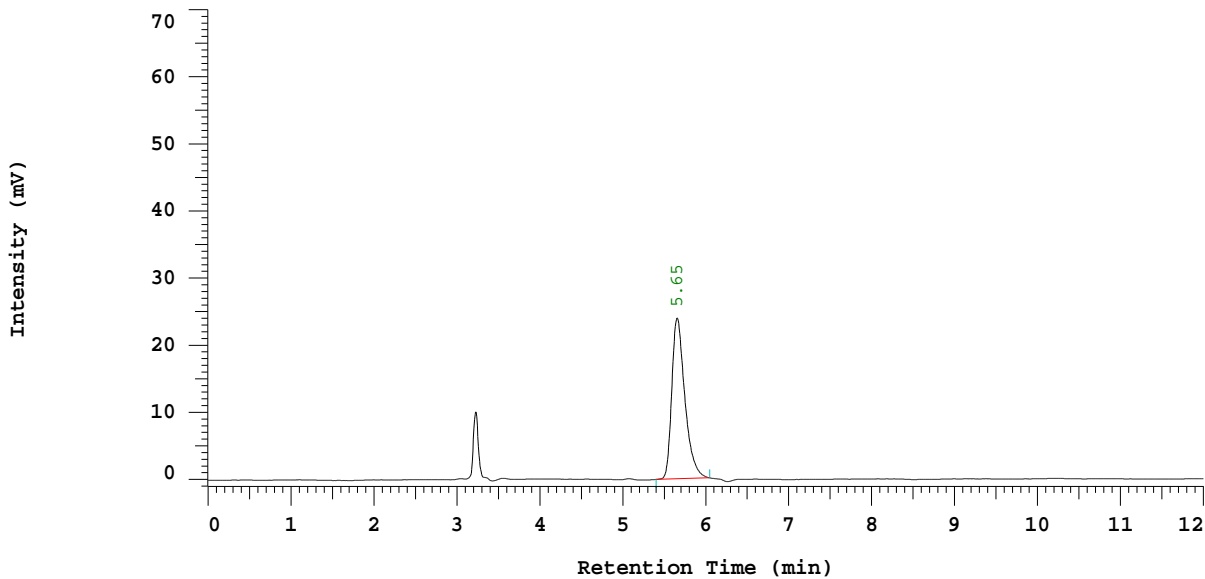
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.65	261816	23954	100.000
		261816	23954	100.000

Peak rejection level: 5000

Fig S164. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 12)

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 06/29/2016
05:47 PM

Reported Date and Time: 07/01/2016
12:13 PM

Processed Date and Time: 07/01/2016
12:12 PM

Data Path: D:\Arun\DATA\0505\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0505

Application(data): Arun

Vial Number: 4

Sample Name: CHP-8g-F2 (Crys 4)

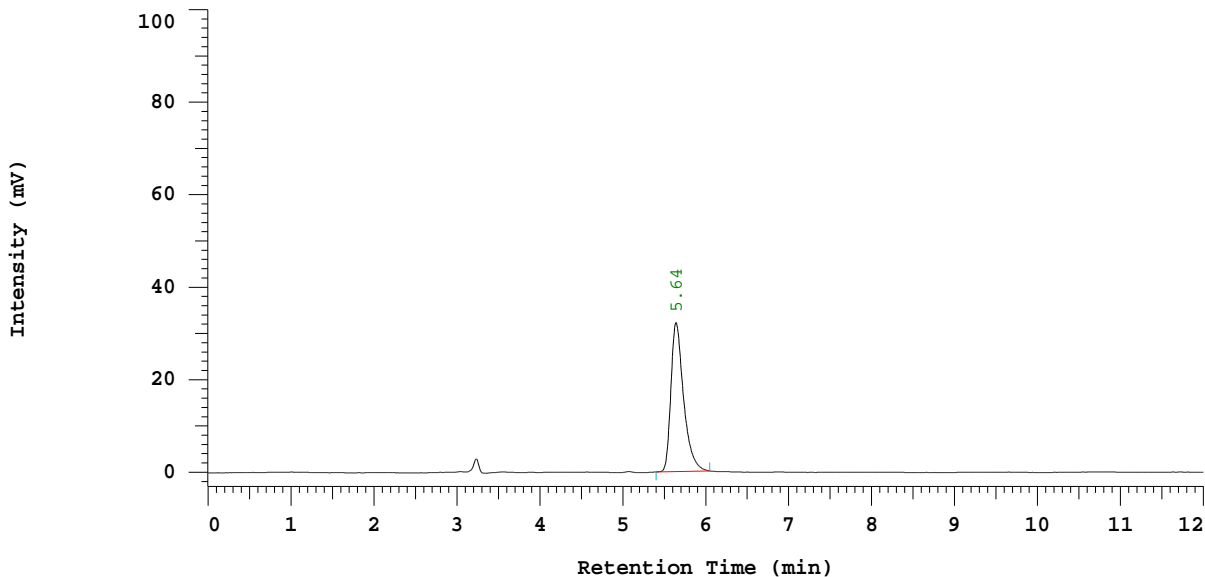
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.64	341129	32185	100.000
		341129	32185	100.000

Peak rejection level: 5000

Fig S165. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 13)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/30/2016
04:59 PMReported Date and Time: 07/01/2016
11:58 AMProcessed Date and Time: 07/01/2016
11:57 AM

Data Path: D:\Arun\DATA\0515\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0515

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crys 1)

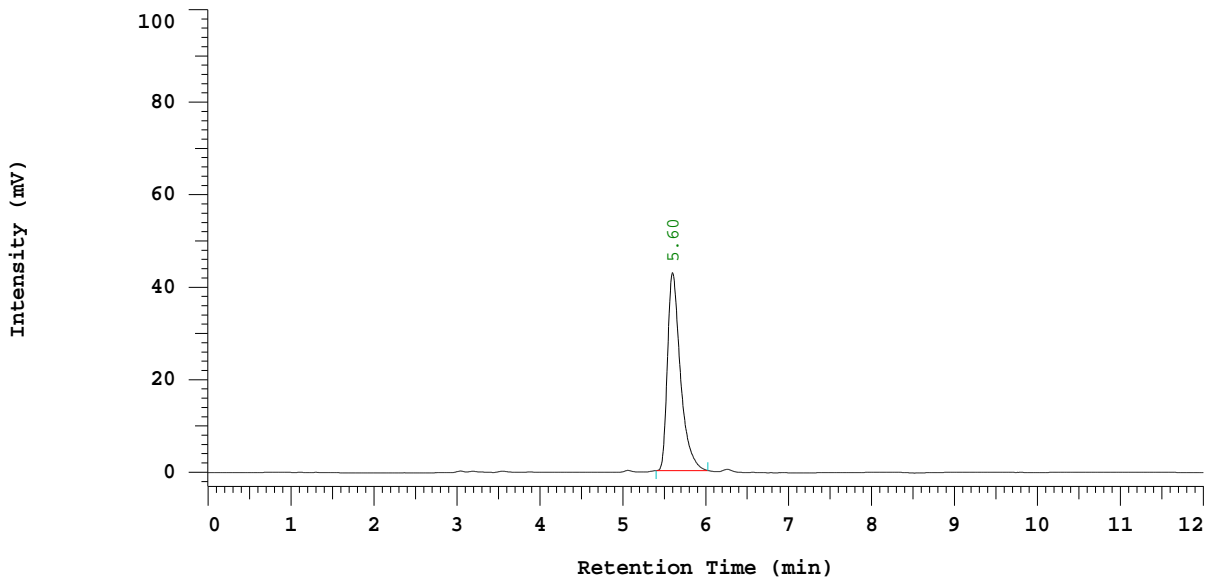
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.60	463194	42825	100.000
		463194	42825	100.000

Peak rejection level: 5000

Fig S166. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 14)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/30/2016
05:12 PMReported Date and Time: 07/01/2016
11:56 AMProcessed Date and Time: 07/01/2016
11:55 AM

Data Path: D:\Arun\DATA\0516\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0516

Application(data): Arun

Vial Number: 2

Sample Name: CHP-8g-F2 (Crys 2)

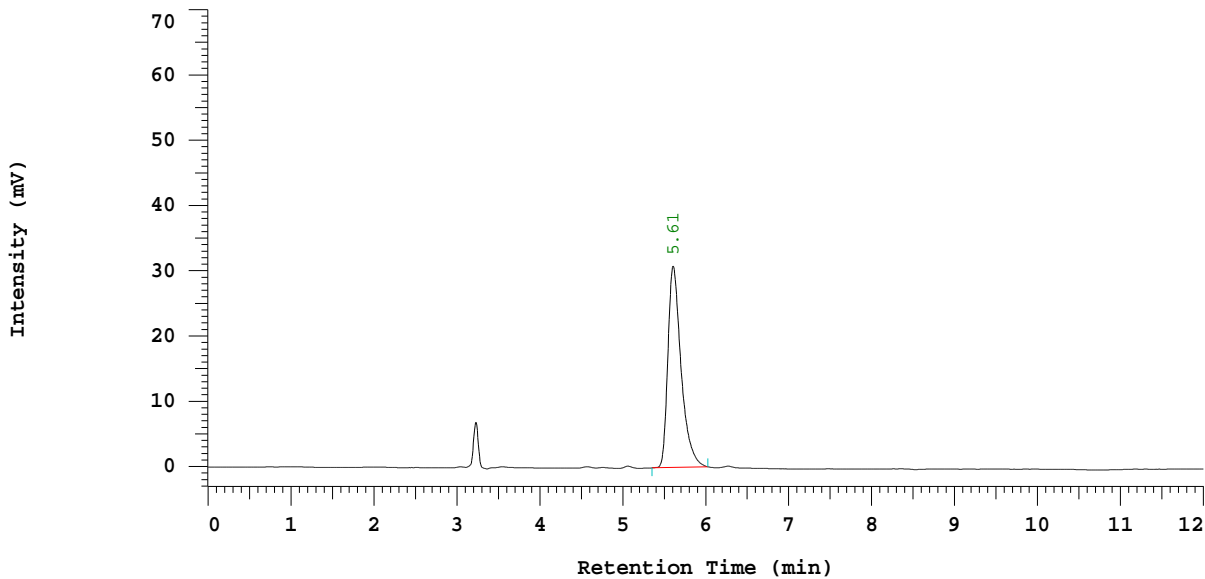
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.61	340930	30841	100.000
		340930	30841	100.000

Peak rejection level: 5000

Fig S167. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 15)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/30/2016
05:50 PMReported Date and Time: 07/01/2016
11:49 AMProcessed Date and Time: 07/01/2016
11:49 AM

Data Path: D:\Arun\DATA\0519\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0519

Application(data): Arun

Vial Number: 5

Sample Name: CHP-8g-F2 (Crys 5)

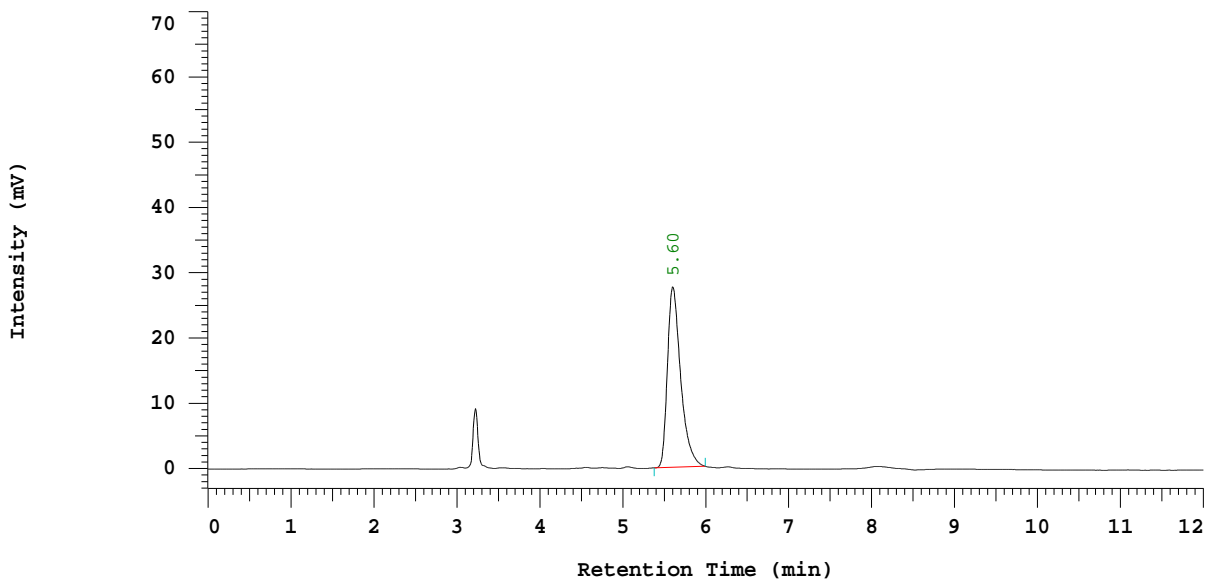
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.60	306252	27617	100.000
		306252	27617	100.000

Peak rejection level: 5000

Fig S168. HPLC analysis of a crystal of the (+)-**3i** obtained (crystal count number 16)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/23/2016
05:39 PMReported Date and Time: 07/01/2016
01:14 PMProcessed Date and Time: 07/01/2016
01:14 PM

Data Path: D:\Arun\DATA\0468\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0468

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crystal 2)

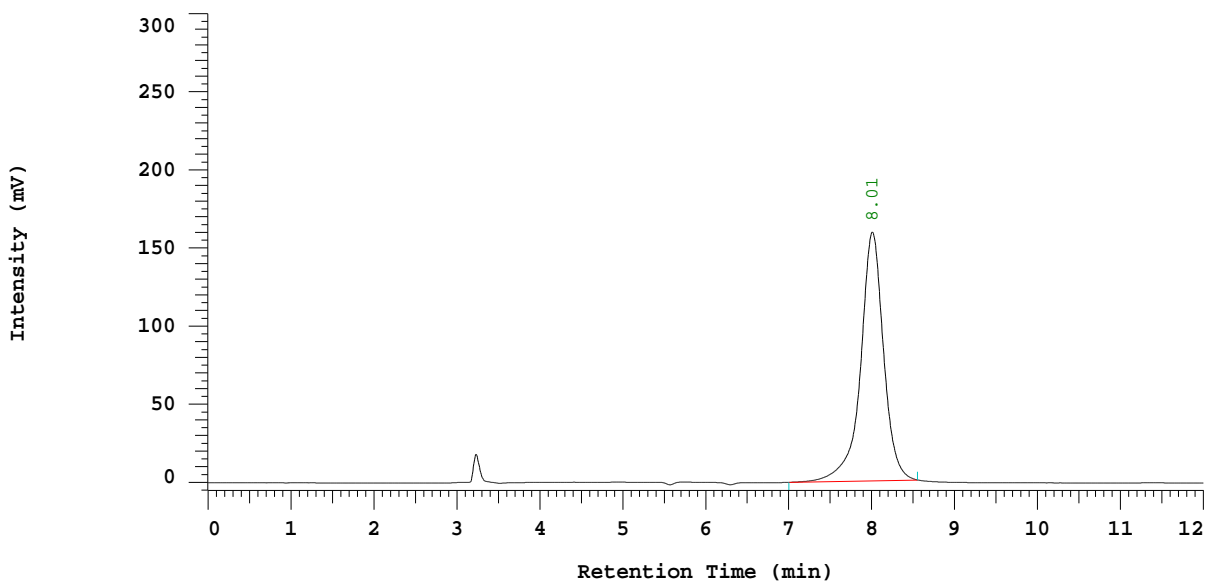
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	8.01	3072195	159115	100.000
		3072195	159115	100.000

Peak rejection level: 5000

Fig S169. HPLC analysis of a crystal of the (–)-**3i** obtained (crystal count number 1)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/23/2016
09:15 PMReported Date and Time: 07/01/2016
01:09 PMProcessed Date and Time: 07/01/2016
01:09 PM

Data Path: D:\Arun\DATA\0472\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0472

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crystal 5)

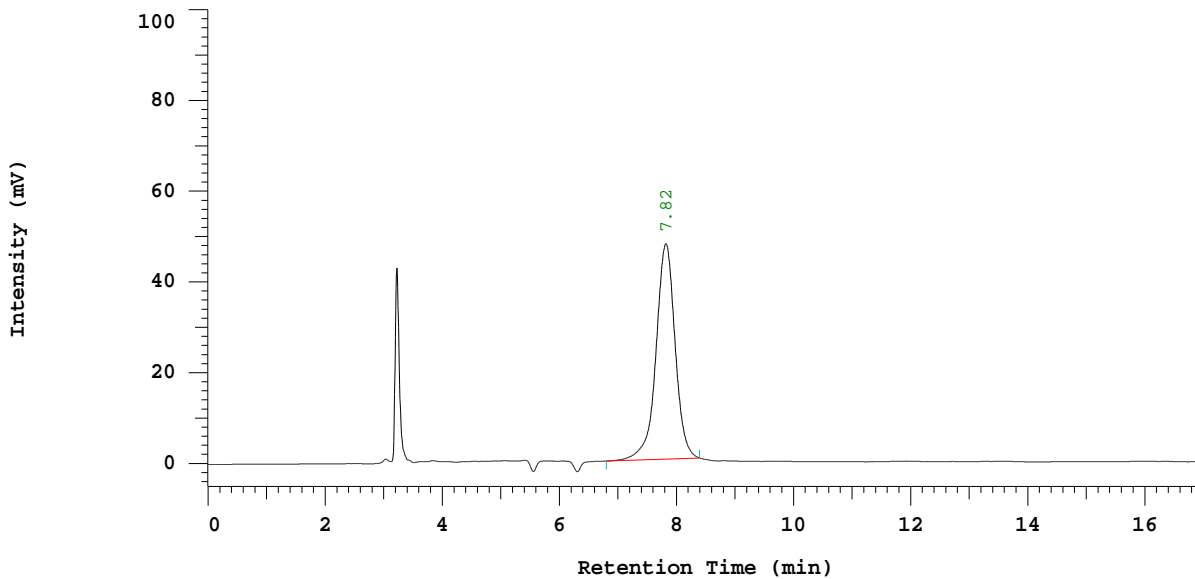
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	7.82	1075182	47439	100.000
		1075182	47439	100.000

Peak rejection level: 5000

Fig S170. HPLC analysis of a crystal of the (-)-**3i** obtained (crystal count number 2)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/24/2016
12:35 PMReported Date and Time: 07/01/2016
01:06 PMProcessed Date and Time: 07/01/2016
01:06 PM

Data Path: D:\Arun\DATA\0475\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0475

Application(data): Arun

Vial Number: 2

Sample Name: CHP-8g-F2 (Crys 2)

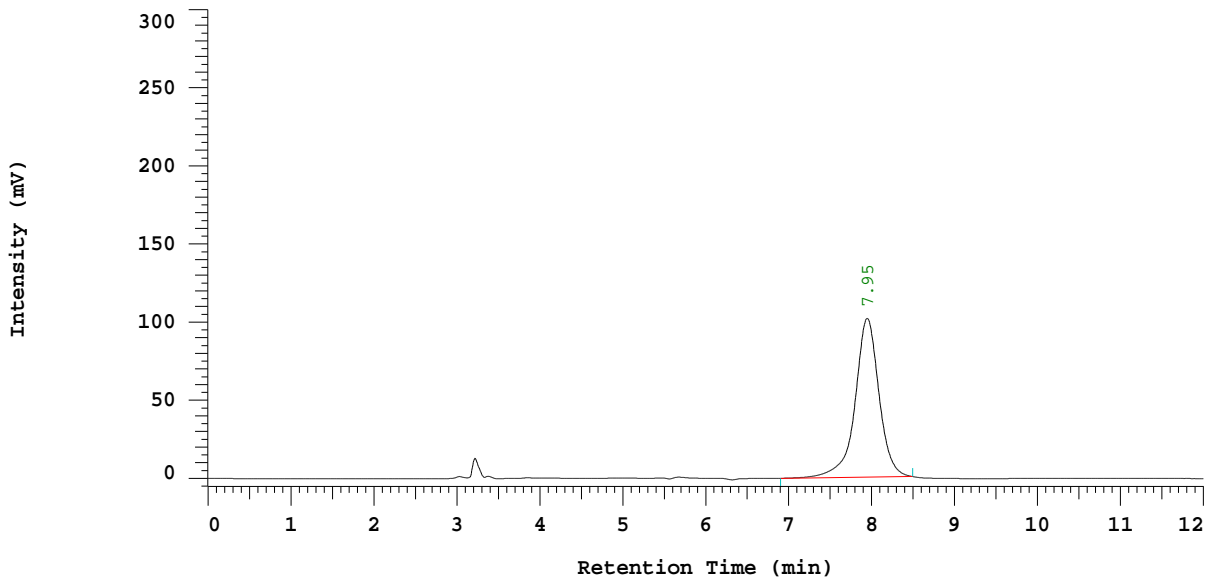
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	7.95	2042287	101574	100.000
		2042287	101574	100.000

Peak rejection level: 5000

Fig S171. HPLC analysis of a crystal of the (–)-**3i** obtained (crystal count number 3)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/27/2016
04:31 PMReported Date and Time: 07/01/2016
12:43 PMProcessed Date and Time: 07/01/2016
12:42 PM

Data Path: D:\Arun\DATA\0487\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0487

Application(data): Arun

Vial Number: 3

Sample Name: CHP-8g-F2 (Crys 3)

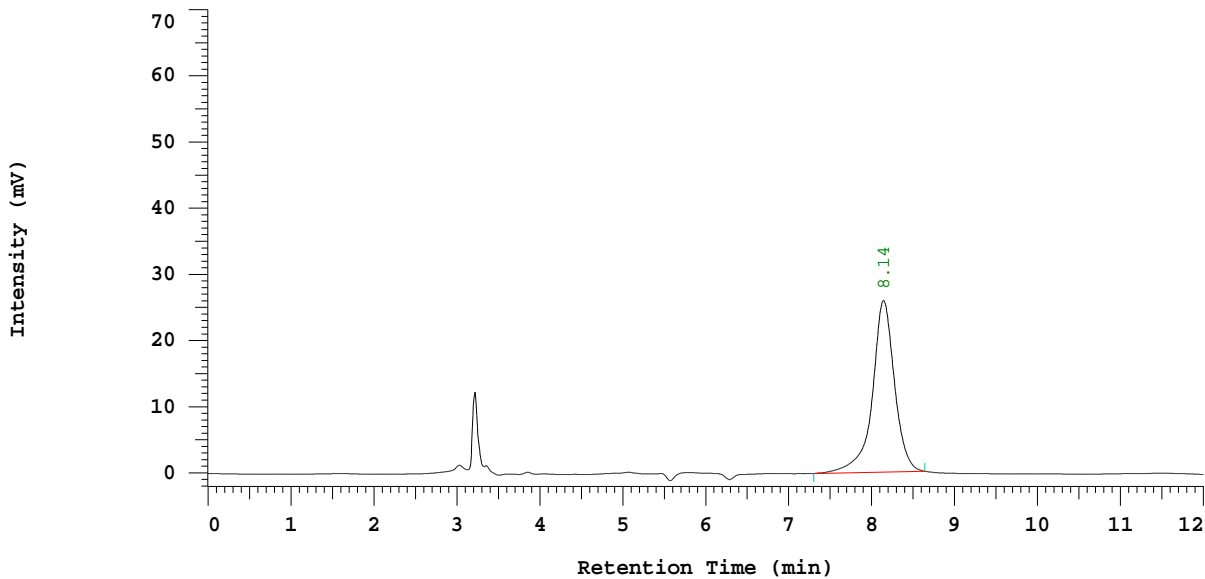
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	8.14	481174	25906	100.000
		481174	25906	100.000

Peak rejection level: 5000

Fig S172. HPLC analysis of a crystal of the (-)-**3i** obtained (crystal count number 4)

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 06/27/2016
04:44 PM

Reported Date and Time: 07/01/2016
12:41 PM

Processed Date and Time: 07/01/2016
12:41 PM

Data Path: D:\Arun\DATA\0488\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0488

Application(data): Arun

Vial Number: 4

Sample Name: CHP-8g-F2 (Crys 4)

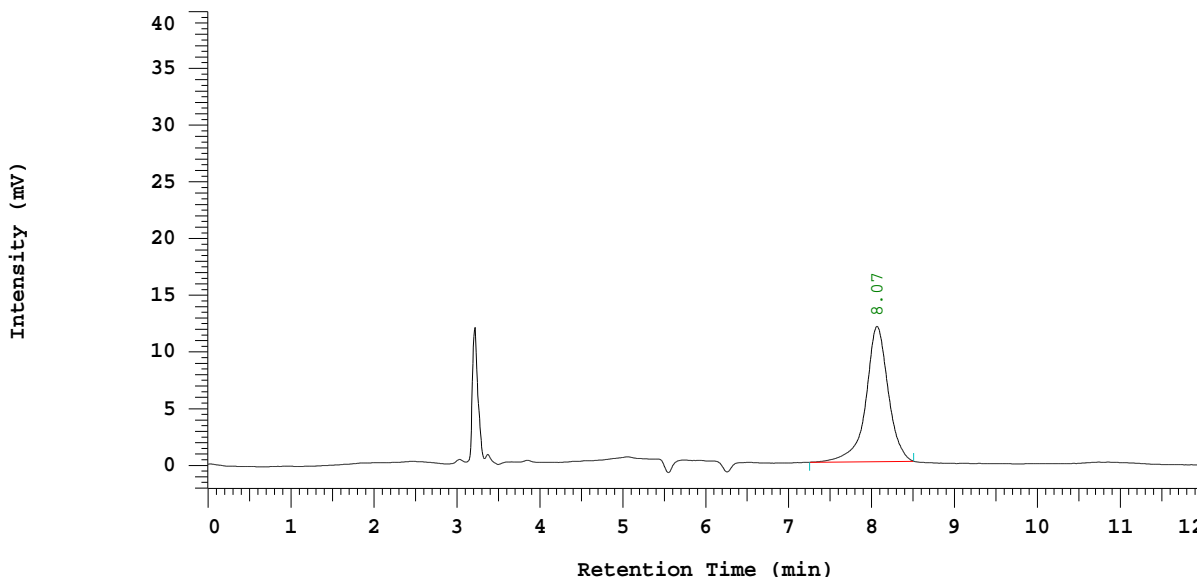
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	8.07	222480	11916	100.000
		222480	11916	100.000

Peak rejection level: 5000

Fig S173. HPLC analysis of a crystal of the (-)-3i obtained (crystal count number 5)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/27/2016
05:35 PMReported Date and Time: 07/01/2016
12:35 PMProcessed Date and Time: 07/01/2016
12:35 PM

Data Path: D:\Arun\DATA\0491\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0491

Application(data): Arun

Vial Number: 2

Sample Name: CHP-8g-F2 (Crys 2)

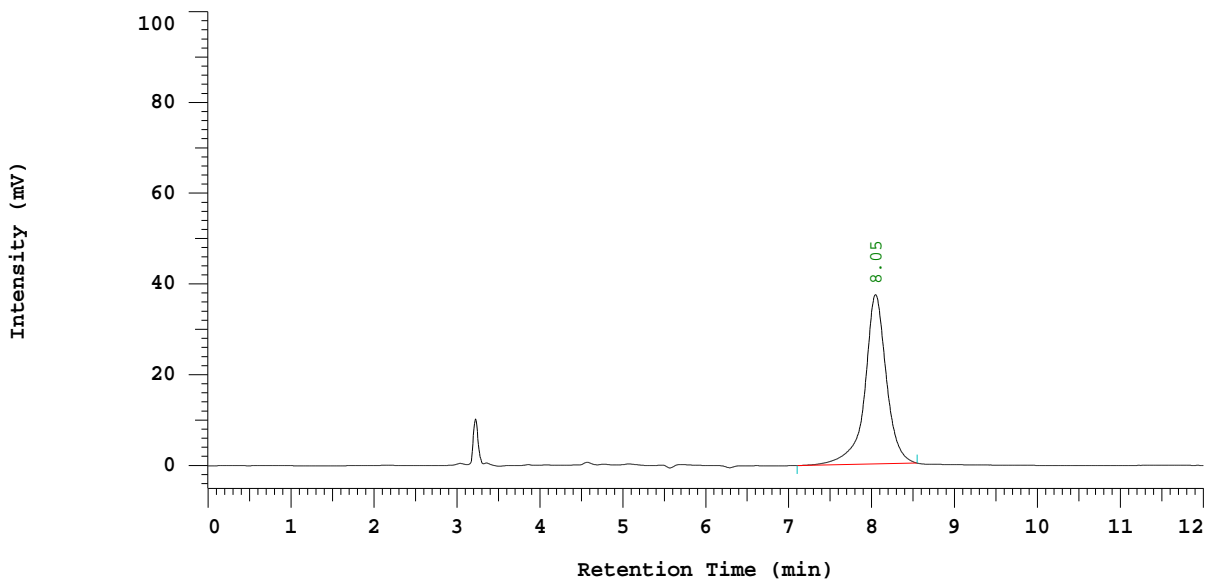
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	8.05	672817	37319	100.000
		672817	37319	100.000

Peak rejection level: 5000

Fig S174. HPLC analysis of a crystal of the (–)-**3i** obtained (crystal count number 6)

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 06/29/2016
04:14 PM

Reported Date and Time: 07/01/2016
12:25 PM

Processed Date and Time: 07/01/2016
12:25 PM

Data Path: D:\Arun\DATA\0498\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0498

Application(data): Arun

Vial Number: 2

Sample Name: CHP-8g-F2 (Crys 2)

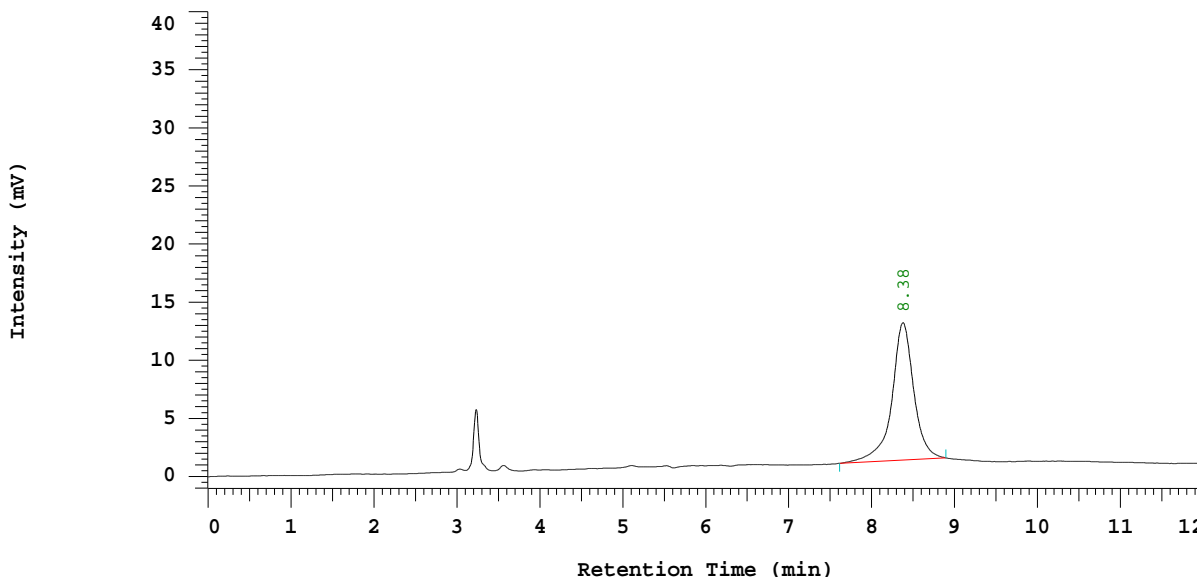
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	8.38	217094	11818	100.000
		217094	11818	100.000

Peak rejection level: 5000

Fig S175. HPLC analysis of a crystal of the (-)-3i obtained (crystal count number 7)

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 06/29/2016
04:53 PM

Reported Date and Time: 07/01/2016
12:21 PM

Processed Date and Time: 07/01/2016
12:20 PM

Data Path: D:\Arun\DATA\0501\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0501

Application(data): Arun

Vial Number: 5

Sample Name: CHP-8g-F2 (Crys 5)

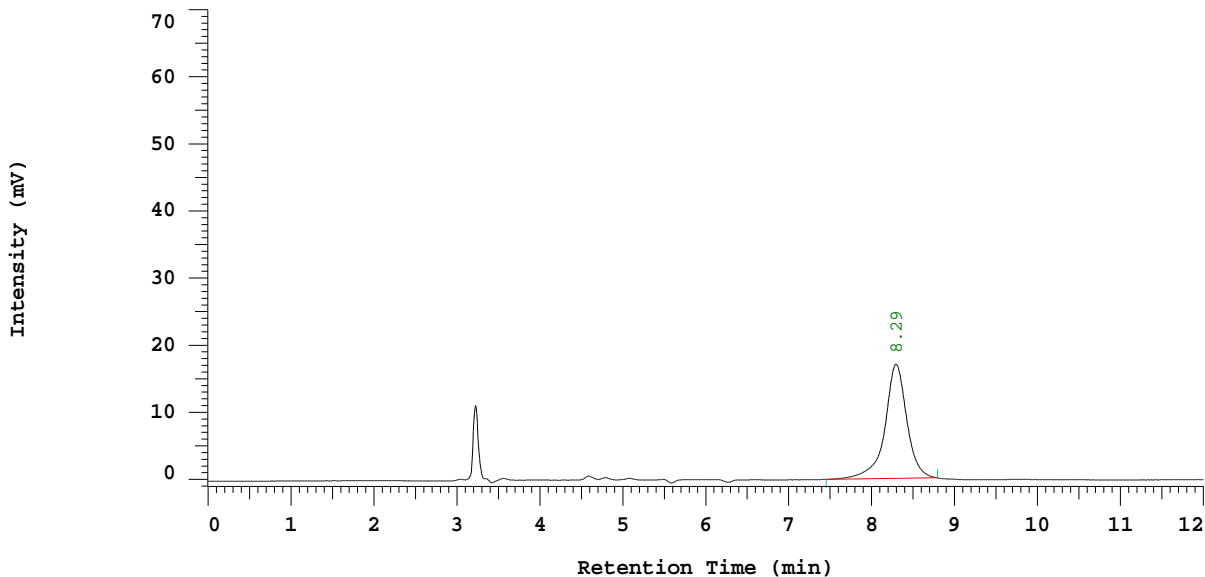
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	8.29	313751	17026	100.000
		313751	17026	100.000

Peak rejection level: 5000

Fig S176. HPLC analysis of a crystal of the (-)-**3i** obtained (crystal count number 8)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/29/2016
05:59 PMReported Date and Time: 07/01/2016
12:11 PMProcessed Date and Time: 07/01/2016
12:11 PM

Data Path: D:\Arun\DATA\0506\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0506

Application(data): Arun

Vial Number: 5

Sample Name: CHP-8g-F2 (Crys 5)

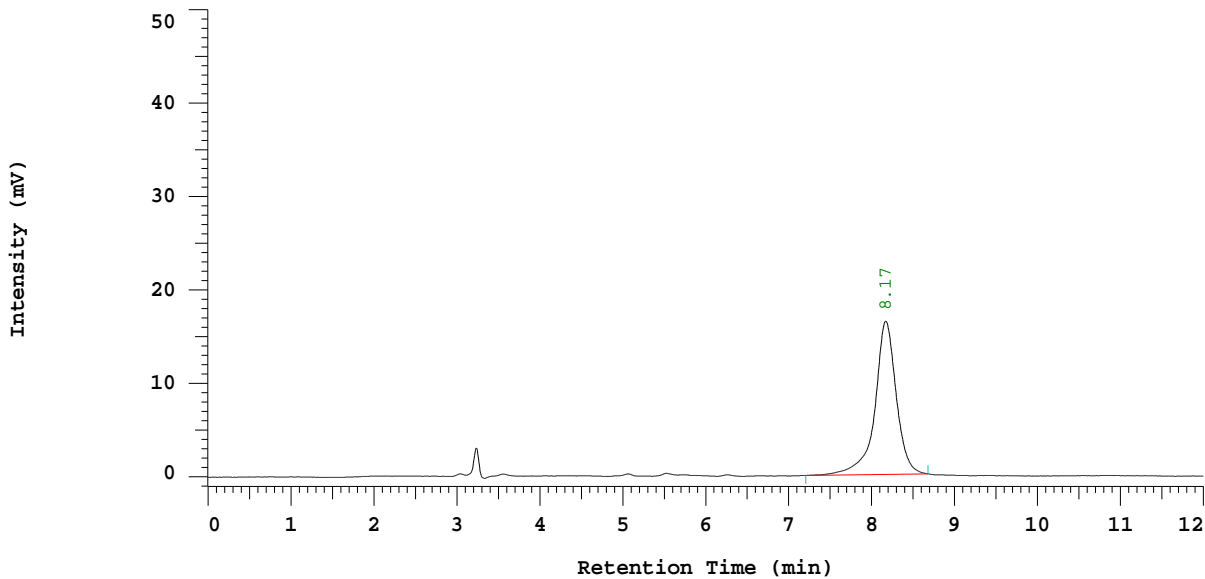
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	8.17	293692	16396	100.000
		293692	16396	100.000

Peak rejection level: 5000

Fig S177. HPLC analysis of a crystal of the (–)-**3i** obtained (crystal count number 9)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/30/2016
03:38 PMReported Date and Time: 07/01/2016
12:09 PMProcessed Date and Time: 07/01/2016
12:09 PM

Data Path: D:\Arun\DATA\0509\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0509

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crys 1)

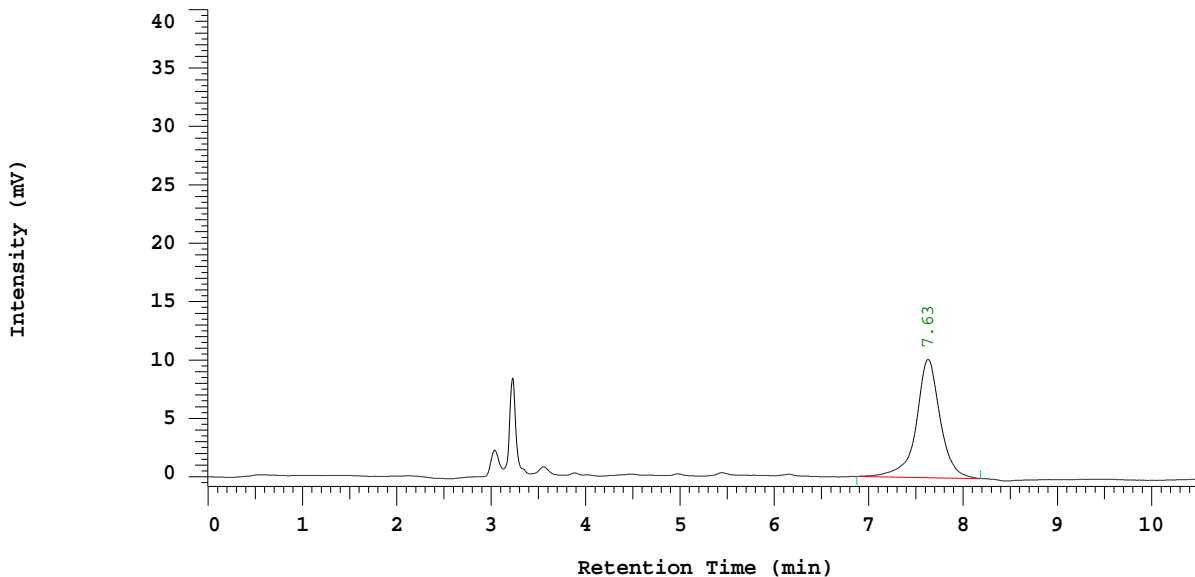
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	7.63	178613	10137	100.000
		178613	10137	100.000

Peak rejection level: 5000

Fig S178. HPLC analysis of a crystal of the (-)-**3i** obtained (crystal count number 10)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/30/2016
03:50 PMReported Date and Time: 07/01/2016
12:07 PMProcessed Date and Time: 07/01/2016
12:07 PM

Data Path: D:\Arun\DATA\0510\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0510

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crys 1)

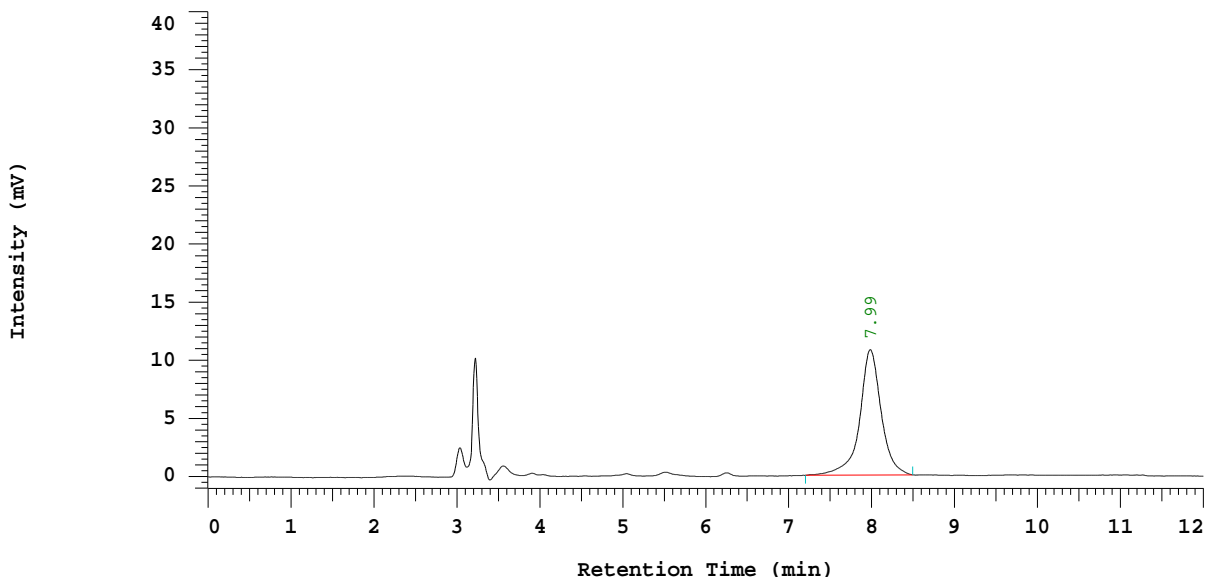
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	7.99	194411	10775	100.000
		194411	10775	100.000

Peak rejection level: 5000

Fig S179. HPLC analysis of a crystal of the (–)-**3i** obtained (crystal count number 11)

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 06/30/2016
04:03 PM

Reported Date and Time: 07/01/2016
12:05 PM

Processed Date and Time: 07/01/2016
12:05 PM

Data Path: D:\Arun\DATA\0511\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0511

Application(data): Arun

Vial Number: 2

Sample Name: CHP-8g-F2 (Crys 2)

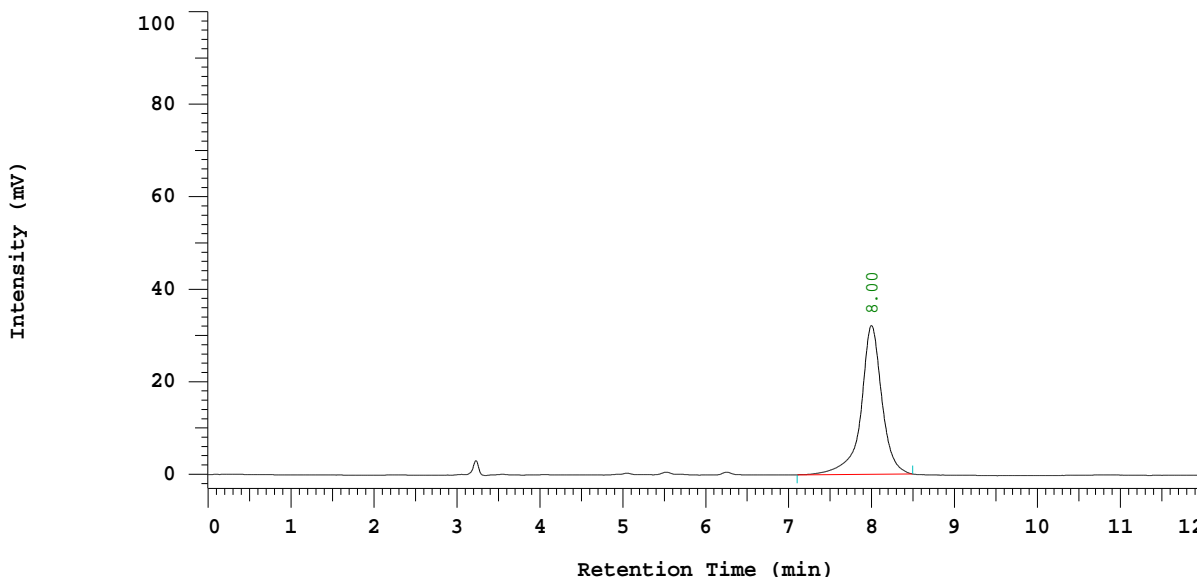
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	8.00	563043	32162	100.000
		563043	32162	100.000

Peak rejection level: 5000

Fig S180. HPLC analysis of a crystal of the (-)-**3i** obtained (crystal count number 12)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/30/2016
04:16 PMReported Date and Time: 07/01/2016
12:04 PMProcessed Date and Time: 07/01/2016
12:03 PM

Data Path: D:\Arun\DATA\0512\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0512

Application(data): Arun

Vial Number: 3

Sample Name: CHP-8g-F2 (Crys 3)

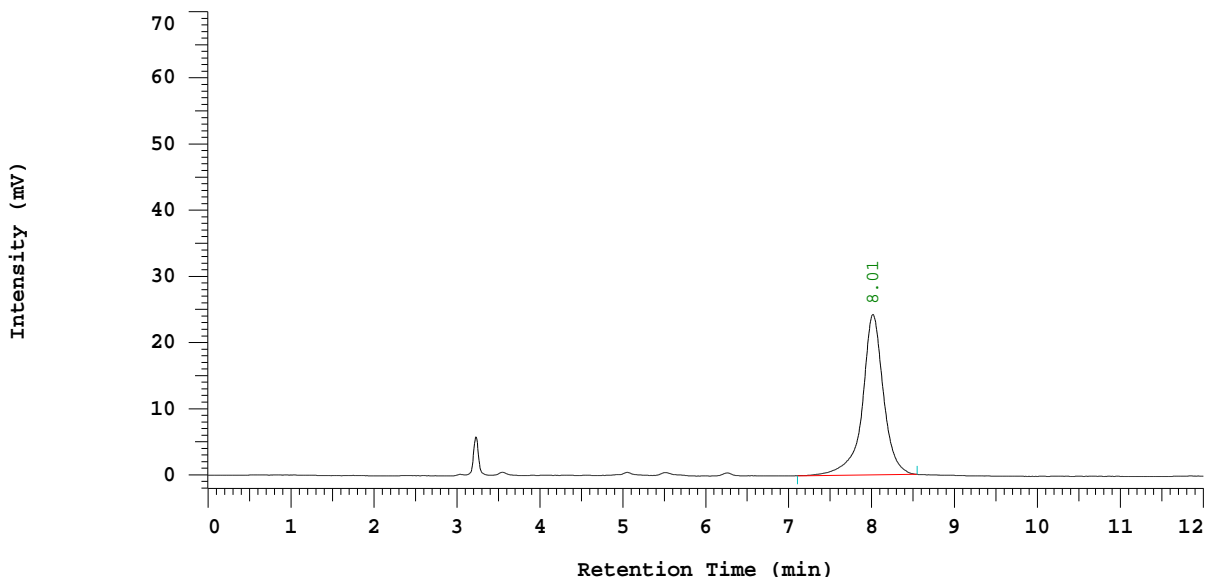
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	8.01	427998	24238	100.000
		427998	24238	100.000

Peak rejection level: 5000

Fig S181. HPLC analysis of a crystal of the (–)-**3i** obtained (crystal count number 13)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/30/2016
05:24 PMReported Date and Time: 07/01/2016
11:54 AMProcessed Date and Time: 07/01/2016
11:53 AM

Data Path: D:\Arun\DATA\0517\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0517

Application(data): Arun

Vial Number: 3

Sample Name: CHP-8g-F2 (Crys 3)

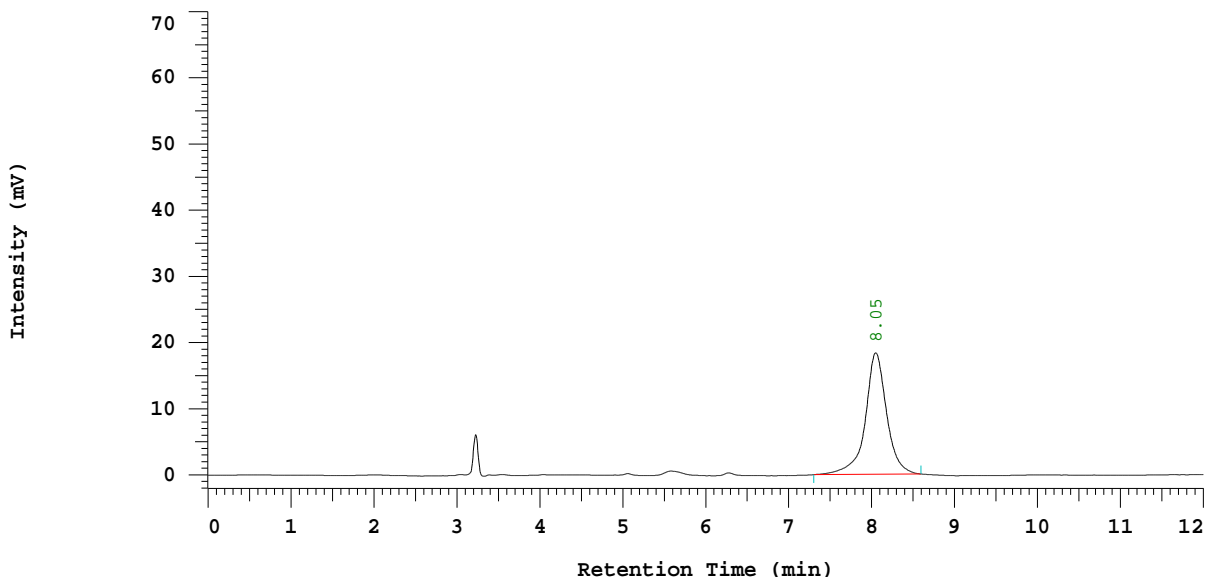
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	8.05	328706	18358	100.000
		328706	18358	100.000

Peak rejection level: 5000

Fig S182. HPLC analysis of a crystal of the (–)-**3i** obtained (crystal count number 14)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 07/04/2016
08:13 PMReported Date and Time: 07/04/2016
09:13 PMProcessed Date and Time: 07/04/2016
09:12 PM

Data Path: D:\Arun\DATA\0523\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0523

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (IC18028)

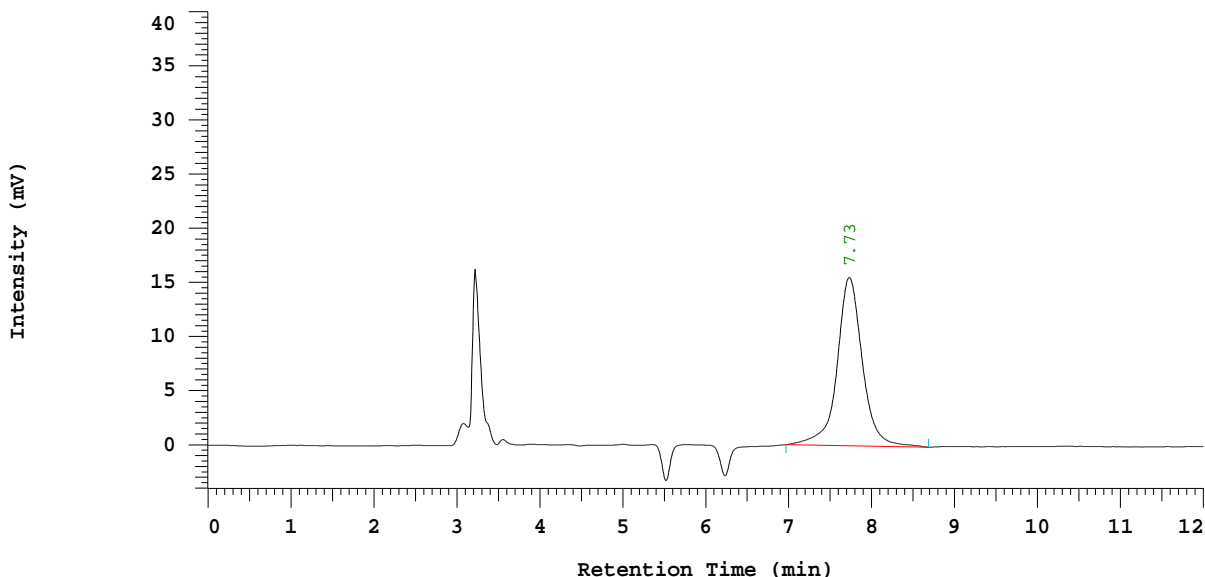
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	7.73	330698	15553	100.000
		330698	15553	100.000

Peak rejection level: 5000

Fig S183. HPLC analysis of the crystal recovered from the X-ray analysis (IC18028, CCDC 1488798)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 07/05/2016
02:39 PMReported Date and Time: 07/05/2016
03:00 PMProcessed Date and Time: 07/05/2016
02:55 PM

Data Path: D:\Arun\DATA\0527\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0527

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (IC18028-Co)

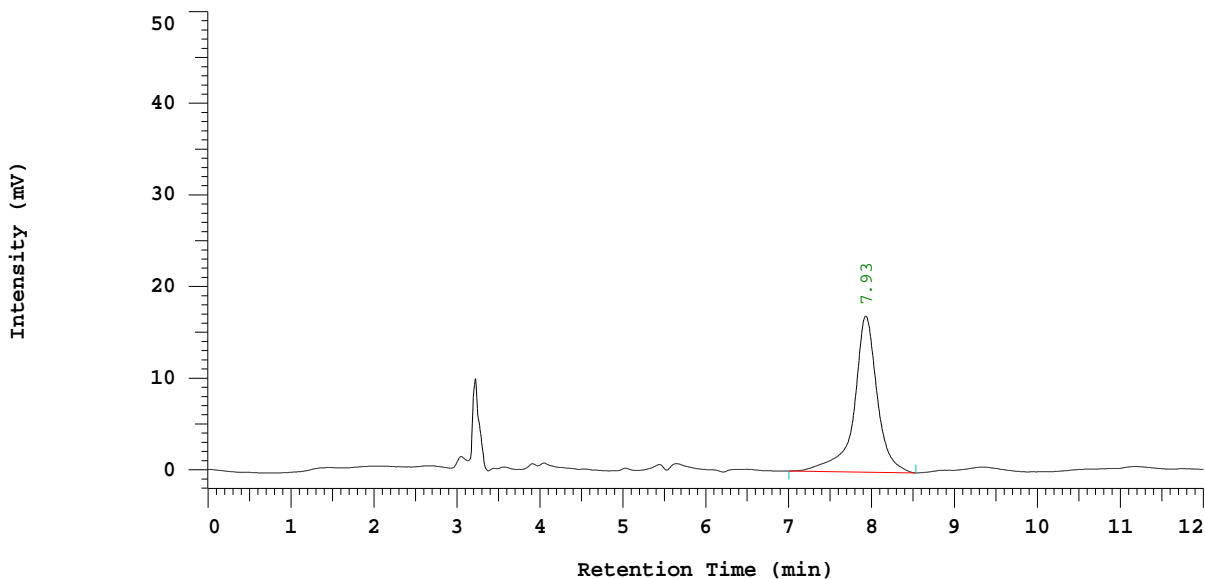
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	7.93	330565	17043	100.000
		330565	17043	100.000

Peak rejection level: 5000

Fig S184. HPLC analysis of the co-injection of the crystal recovered from the X-ray analysis (IC18028, CCDC 1488798) and the (–)-**3i**.

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 07/04/2016
08:39 PMReported Date and Time: 07/04/2016
09:07 PMProcessed Date and Time: 07/04/2016
09:06 PM

Data Path: D:\Arun\DATA\0524\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0524

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (IC18028-1)

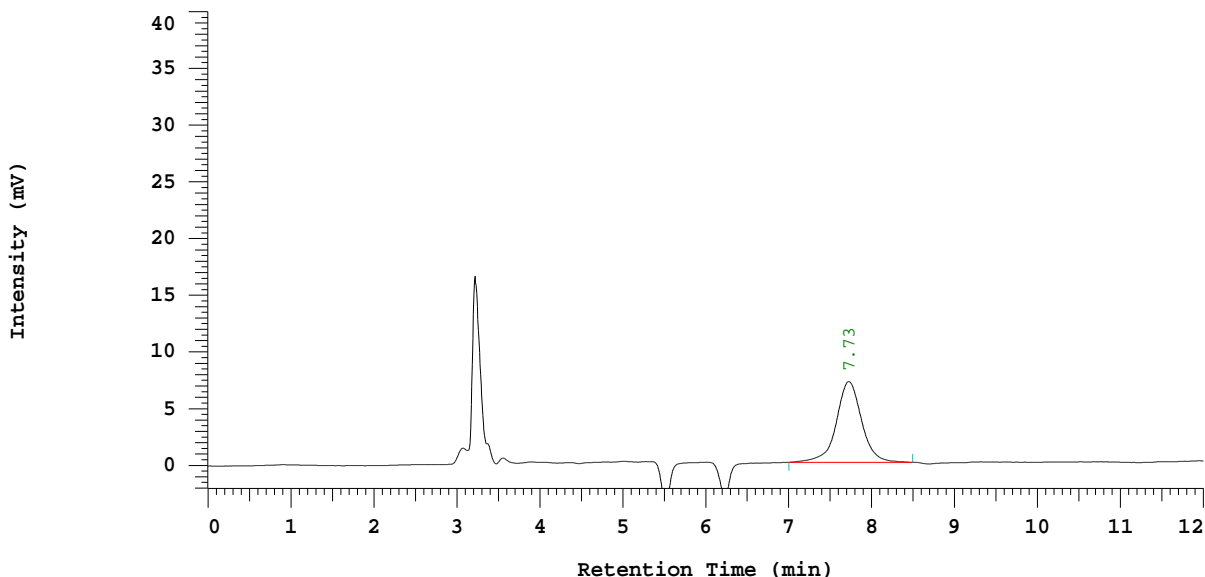
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	7.73	151145	7119	100.000
		151145	7119	100.000

Peak rejection level: 5000

Fig S185. HPLC analysis of the crystal recovered from the X-ray analysis (IC18028-1, CCDC 1488808)

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/26/2016
04:55 PMReported Date and Time: 07/01/2016
12:48 PMProcessed Date and Time: 07/01/2016
12:48 PM

Data Path: D:\Arun\DATA\0484\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0484

Application(data): Arun

Vial Number: 5

Sample Name: CHP-8g-F2 (Crys 5)

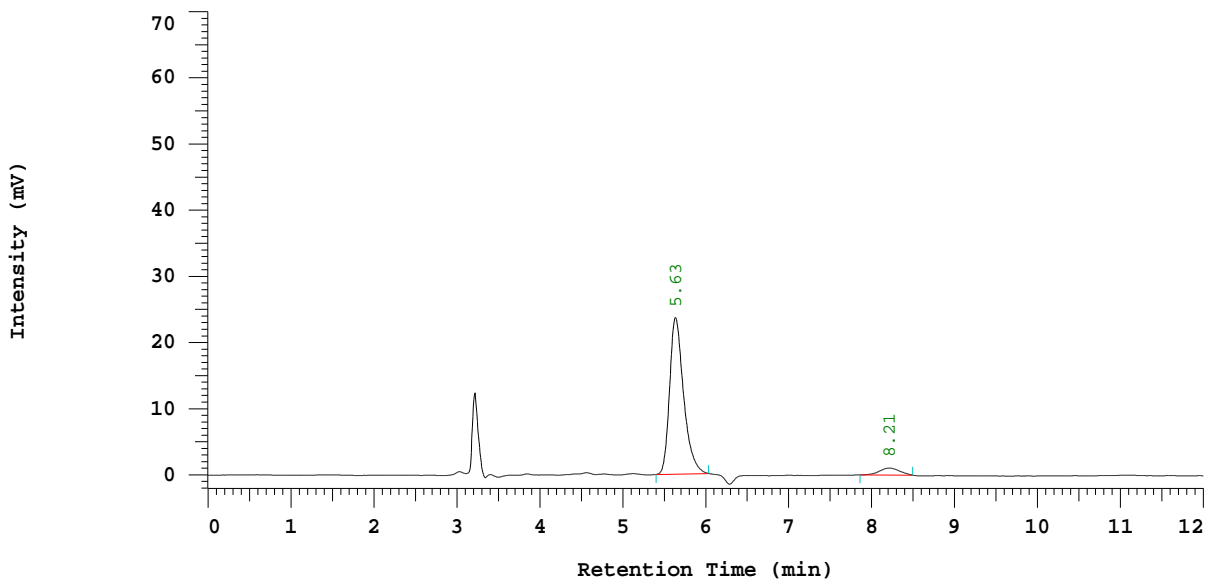
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.63	277123	23648	93.859
2	8.21	18130	1064	6.141
		295253	24712	100.000

Peak rejection level: 5000

Fig S186. HPLC analysis of a crystal **3i** obtained with chiral enriched (crystal count number 1).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/29/2016
05:08 PMReported Date and Time: 07/01/2016
12:19 PMProcessed Date and Time: 07/01/2016
12:19 PM

Data Path: D:\Arun\DATA\0502\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0502

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crys 1)

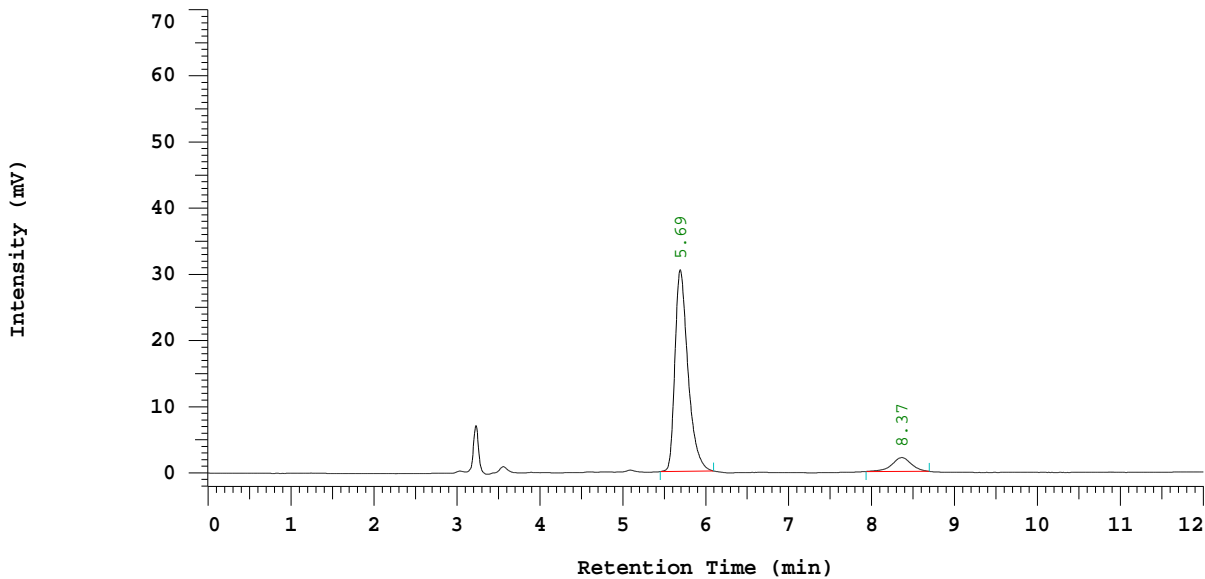
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.69	332157	30486	90.707
2	8.37	34029	2085	9.293
		366186	32571	100.000

Peak rejection level: 5000

Fig S187. HPLC analysis of a crystal 3i obtained with chiral enriched (crystal count number 2).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/23/2016
05:21 PMReported Date and Time: 07/01/2016
01:16 PMProcessed Date and Time: 07/01/2016
01:16 PM

Data Path: D:\Arun\DATA\0467\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0467

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crystal 1)

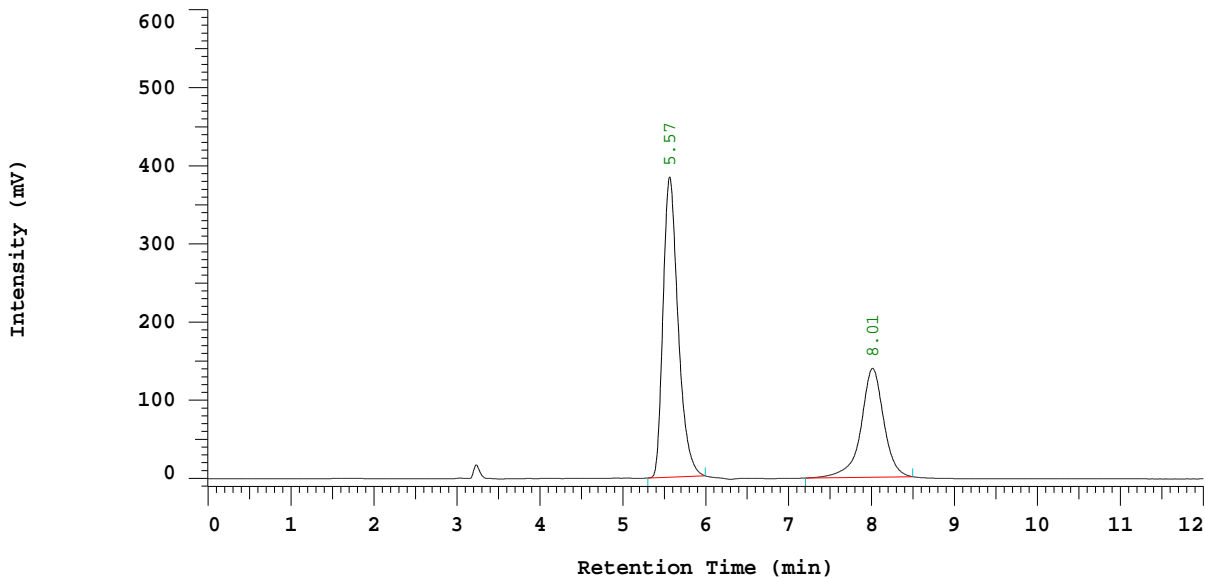
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.57	4845292	384221	64.647
2	8.01	2649724	139303	35.353
		7495016	523524	100.000

Peak rejection level: 5000

Fig S188. HPLC analysis of a crystal **3i** obtained with low ee (crystal count number 1).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/26/2016
04:41 PMReported Date and Time: 07/01/2016
12:50 PMProcessed Date and Time: 07/01/2016
12:50 PM

Data Path: D:\Arun\DATA\0483\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0483

Application(data): Arun

Vial Number: 4

Sample Name: CHP-8g-F2 (Crys 4)

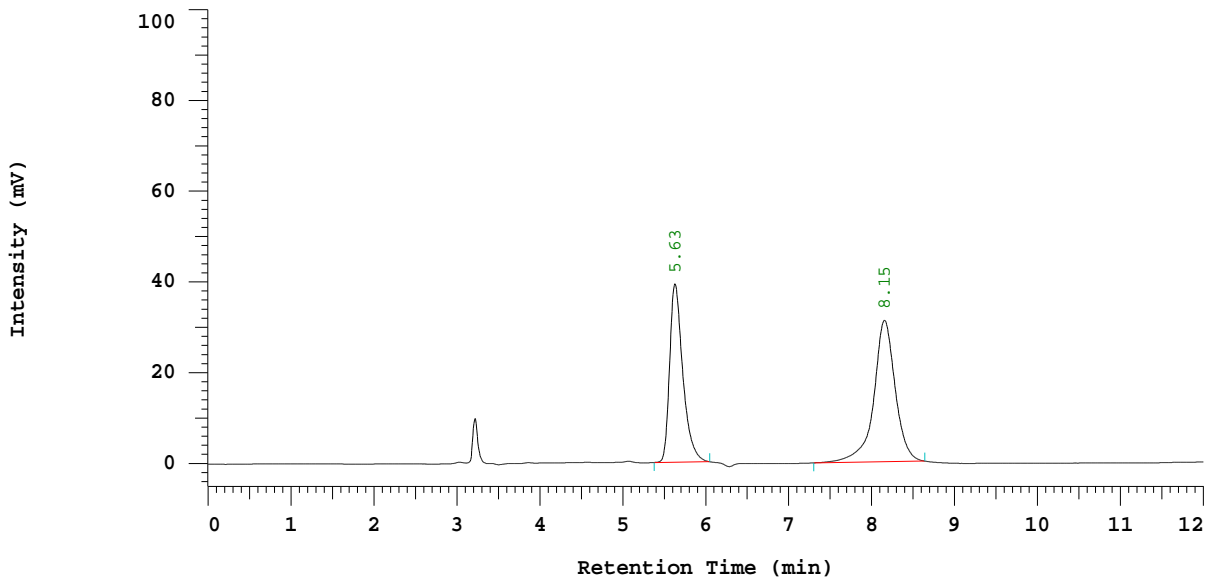
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.63	432836	39300	43.534
2	8.15	561421	31154	56.466
		994257	70454	100.000

Peak rejection level: 5000

Fig S189. HPLC analysis of a crystal **3i** obtained with low ee (crystal count number 2).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/27/2016
04:05 PMReported Date and Time: 07/01/2016
12:46 PMProcessed Date and Time: 07/01/2016
12:46 PM

Data Path: D:\Arun\DATA\0485\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0485

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crys 1)

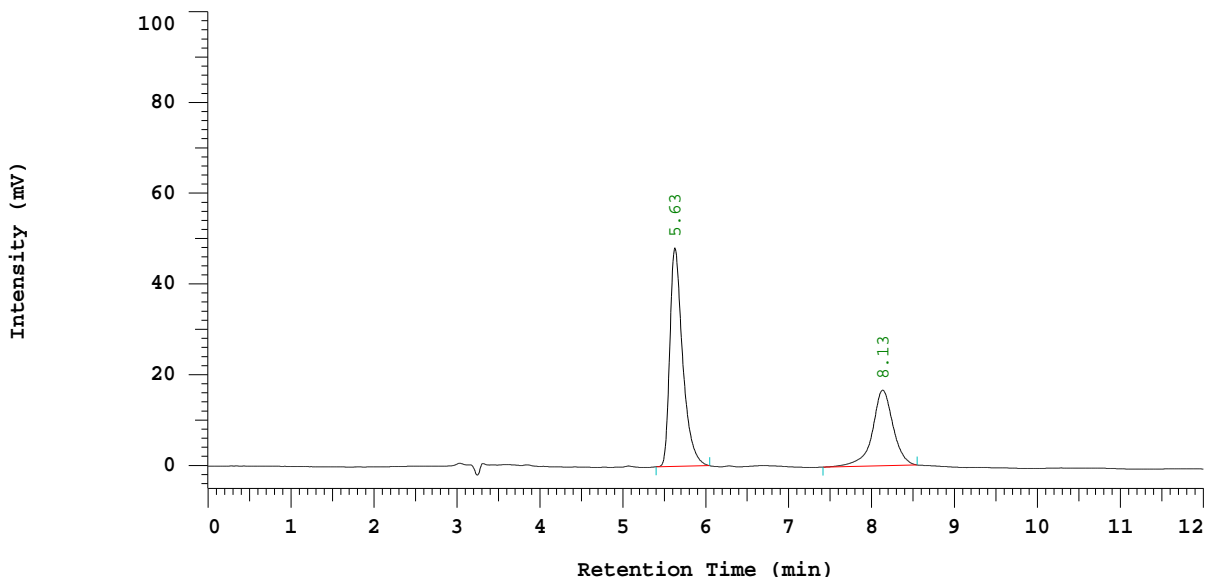
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.63	516846	48139	64.117
2	8.13	289258	16679	35.883
		806104	64818	100.000

Peak rejection level: 5000

Fig S190. HPLC analysis of a crystal **3i** obtained with low ee (crystal count number 3).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/27/2016
05:00 PMReported Date and Time: 07/01/2016
12:40 PMProcessed Date and Time: 07/01/2016
12:40 PM

Data Path: D:\Arun\DATA\0489\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0489

Application(data): Arun

Vial Number: 5

Sample Name: CHP-8g-F2 (Crys 5)

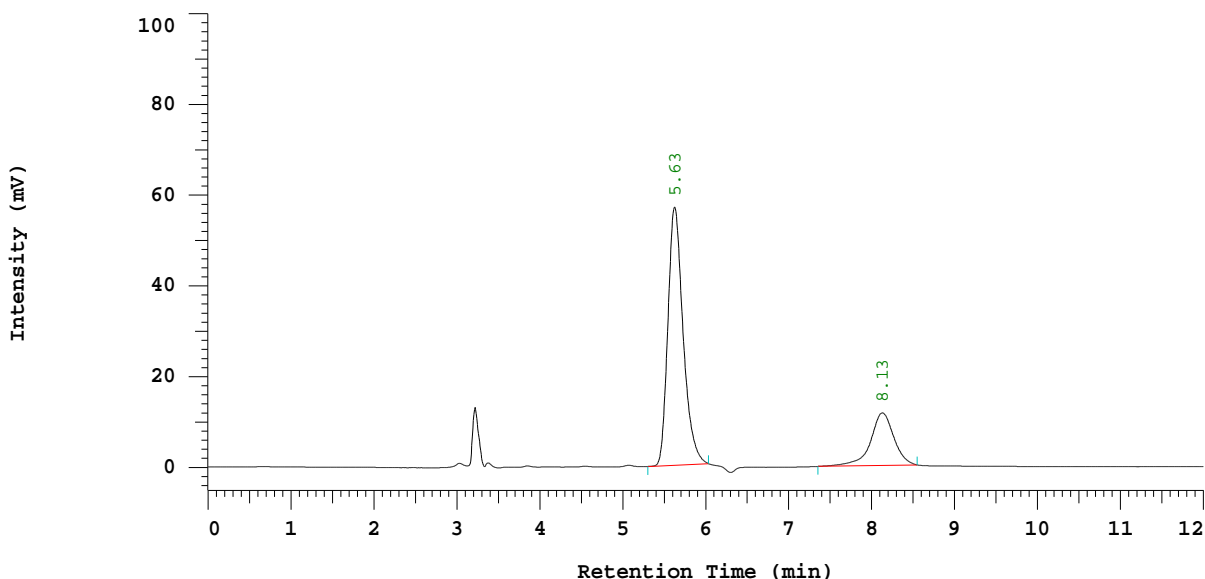
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.63	723288	56910	76.120
2	8.13	226909	11587	23.880
		950197	68497	100.000

Peak rejection level: 5000

Fig S191. HPLC analysis of a crystal **3i** obtained with low ee (crystal count number 4).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/27/2016
05:48 PMReported Date and Time: 07/01/2016
12:34 PMProcessed Date and Time: 07/01/2016
12:34 PM

Data Path: D:\Arun\DATA\0492\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0492

Application(data): Arun

Vial Number: 3

Sample Name: CHP-8g-F2 (Crys 3)

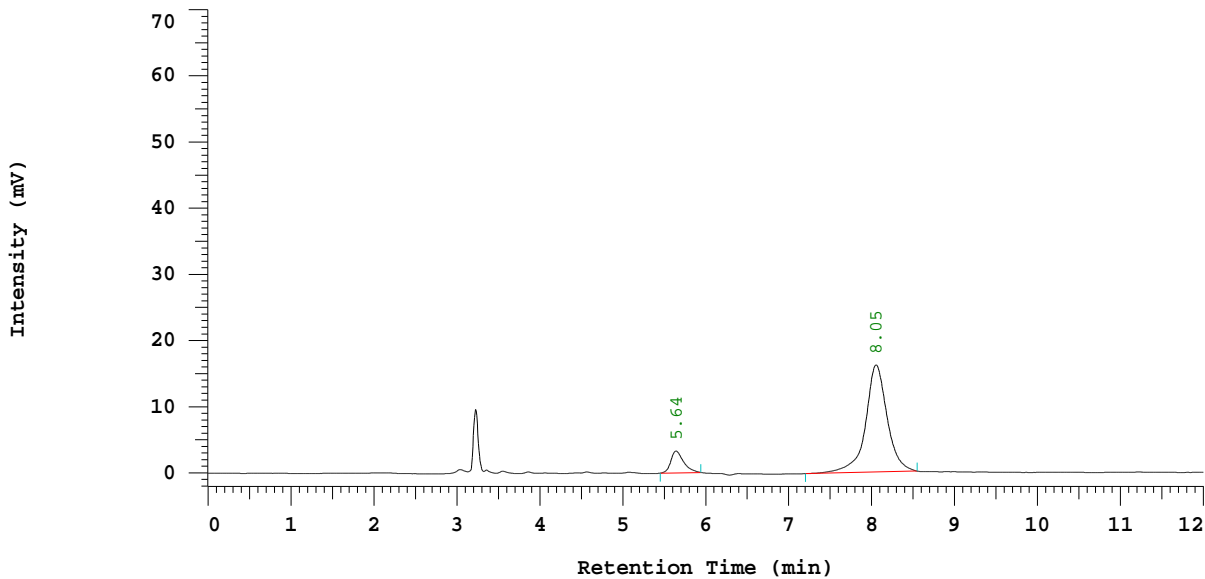
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.64	35093	3289	10.672
2	8.05	293735	16188	89.328
		328828	19477	100.000

Peak rejection level: 5000

Fig S192. HPLC analysis of a crystal **3i** obtained with low ee (crystal count number 5).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/29/2016
04:01 PMReported Date and Time: 07/01/2016
12:27 PMProcessed Date and Time: 07/01/2016
12:27 PM

Data Path: D:\Arun\DATA\0497\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0497

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crys 1)

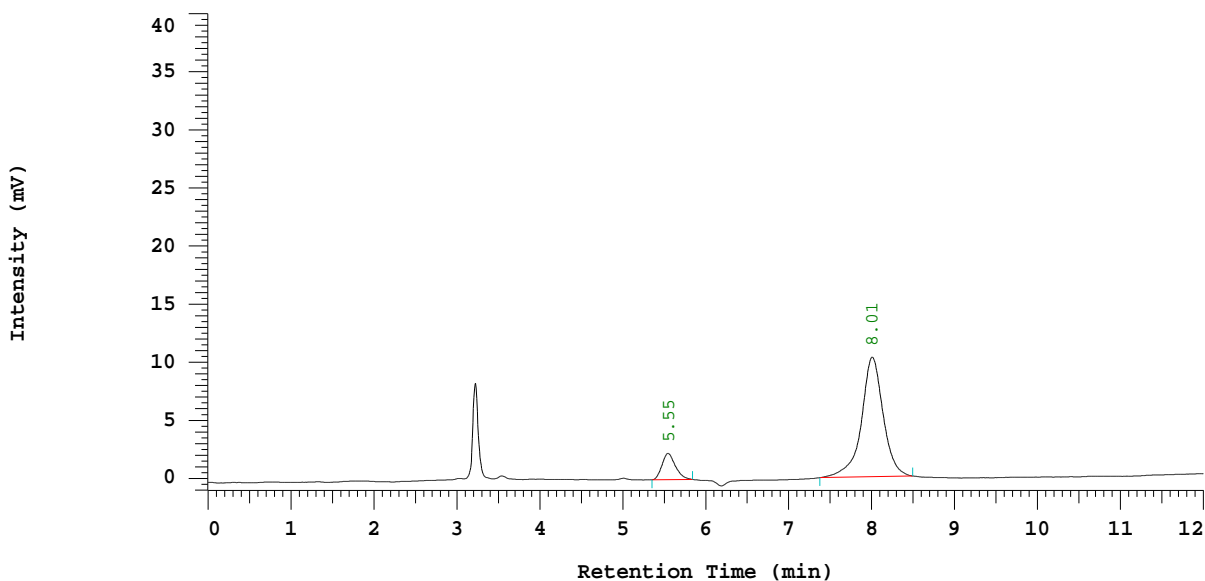
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.55	25275	2257	11.866
2	8.01	187727	10277	88.134
		213002	12534	100.000

Peak rejection level: 5000

Fig S193. HPLC analysis of a crystal **3i** obtained with low ee (crystal count number 6).

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 06/30/2016
04:30 PM

Reported Date and Time: 07/01/2016
12:02 PM

Processed Date and Time: 07/01/2016
12:01 PM

Data Path: D:\Arun\DATA\0513\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0513

Application(data): Arun

Vial Number: 4

Sample Name: CHP-8g-F2 (Crys 4)

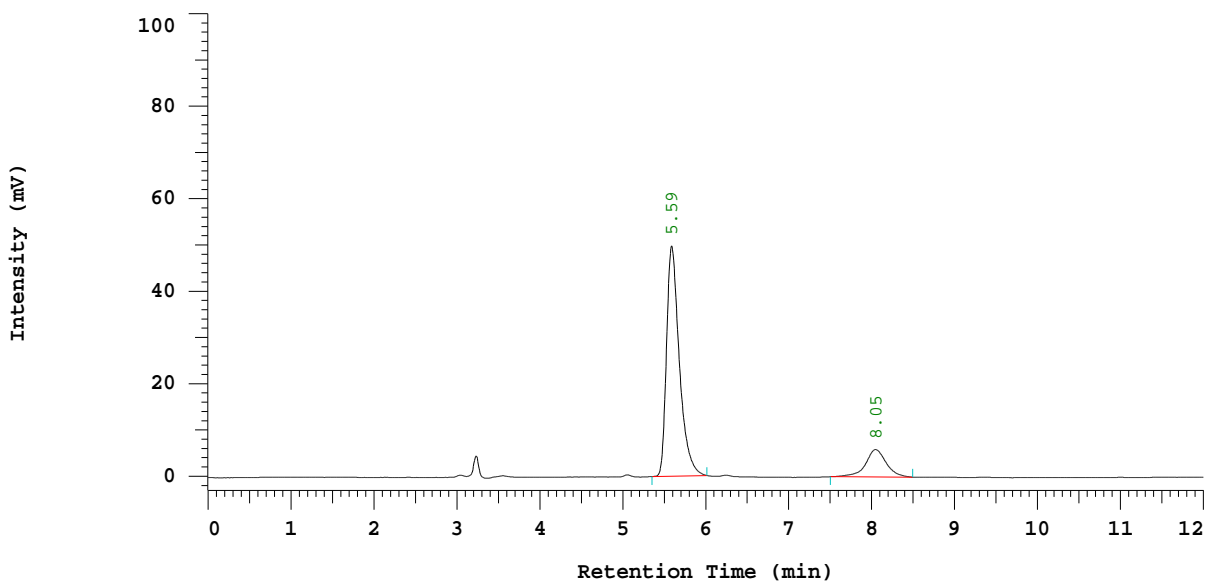
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.59	534229	49723	83.775
2	8.05	103463	5932	16.225
		637692	55655	100.000

Peak rejection level: 5000

Fig S194. HPLC analysis of a crystal **3i** obtained with low ee (crystal count number 7).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/29/2016
05:21 PMReported Date and Time: 07/01/2016
12:17 PMProcessed Date and Time: 07/01/2016
12:17 PM

Data Path: D:\Arun\DATA\0503\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0503

Application(data): Arun

Vial Number: 2

Sample Name: CHP-8g-F2 (Crys 2)

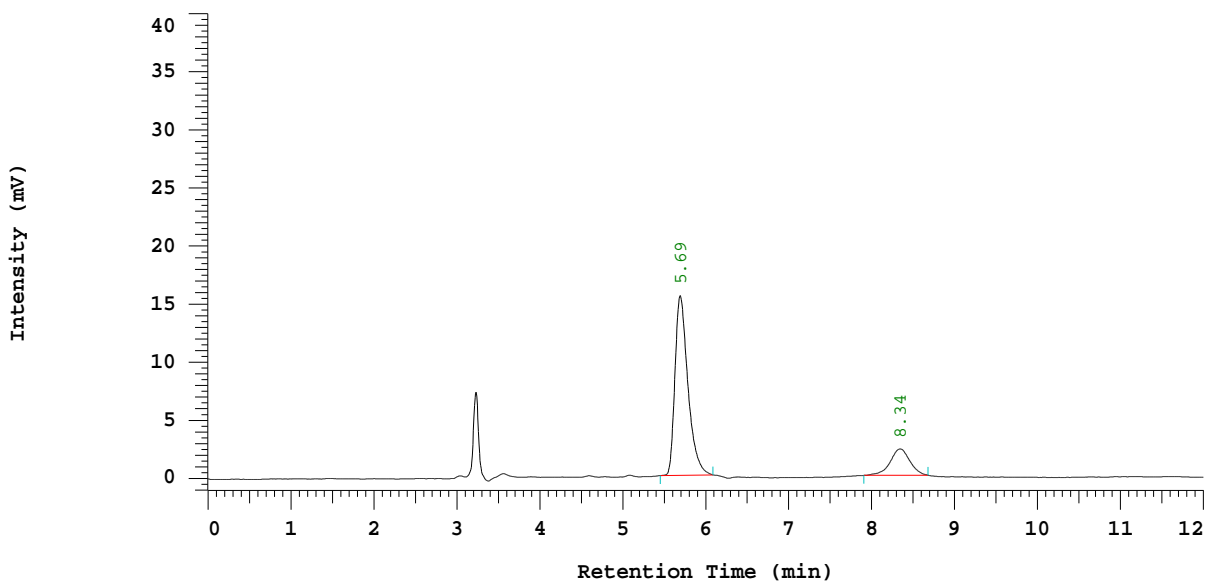
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.69	167329	15446	81.733
2	8.34	37396	2266	18.267
		204725	17712	100.000

Peak rejection level: 5000

Fig S195. HPLC analysis of a crystal **3i** obtained with low ee (crystal count number 8).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/24/2016
01:17 PMReported Date and Time: 07/01/2016
01:00 PMProcessed Date and Time: 07/01/2016
01:00 PM

Data Path: D:\Arun\DATA\0478\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0478

Application(data): Arun

Vial Number: 5

Sample Name: CHP-8g-F2 (Crys 5)

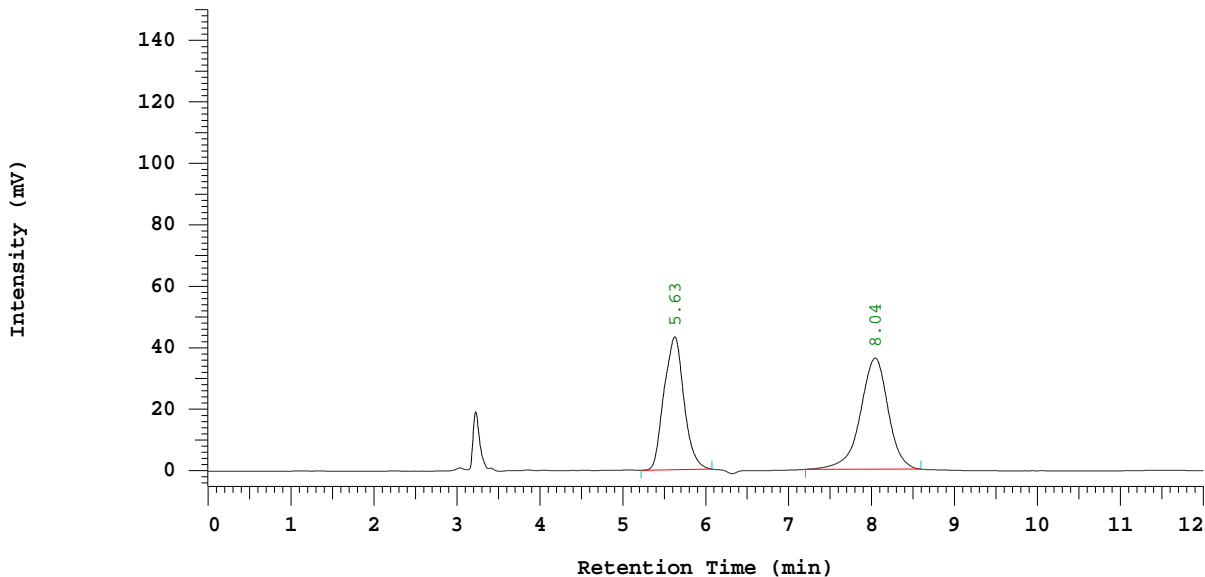
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.63	721768	43285	46.139
2	8.04	842566	36161	53.861
		1564334	79446	100.000

Peak rejection level: 5000

Fig S196. HPLC analysis of a crystal **3i** obtained with very low ee (crystal count number 1).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/23/2016
06:11 PMReported Date and Time: 07/01/2016
01:12 PMProcessed Date and Time: 07/01/2016
01:11 PM

Data Path: D:\Arun\DATA\0470\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0470

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crystal 4)

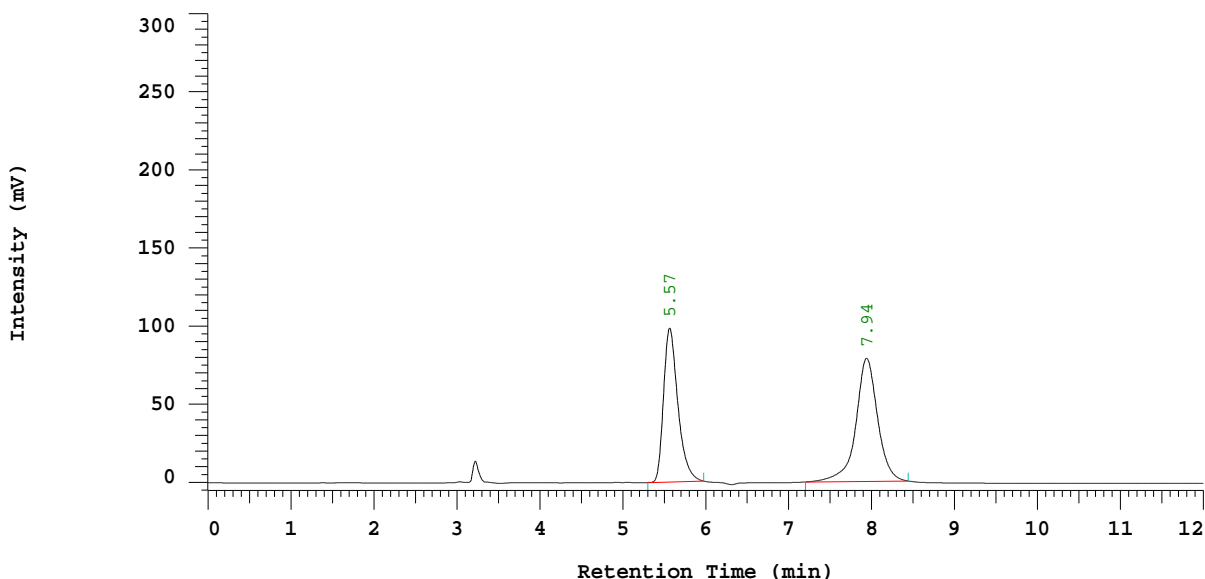
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.57	1180654	98373	45.116
2	7.94	1436277	78794	54.884
		2616931	177167	100.000

Peak rejection level: 5000

Fig S197. HPLC analysis of a crystal **3i** obtained with very low ee (crystal count number 2).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/30/2016
04:42 PMReported Date and Time: 07/01/2016
12:00 PMProcessed Date and Time: 07/01/2016
12:00 PM

Data Path: D:\Arun\DATA\0514\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0514

Application(data): Arun

Vial Number: 5

Sample Name: CHP-8g-F2 (Crys 5)

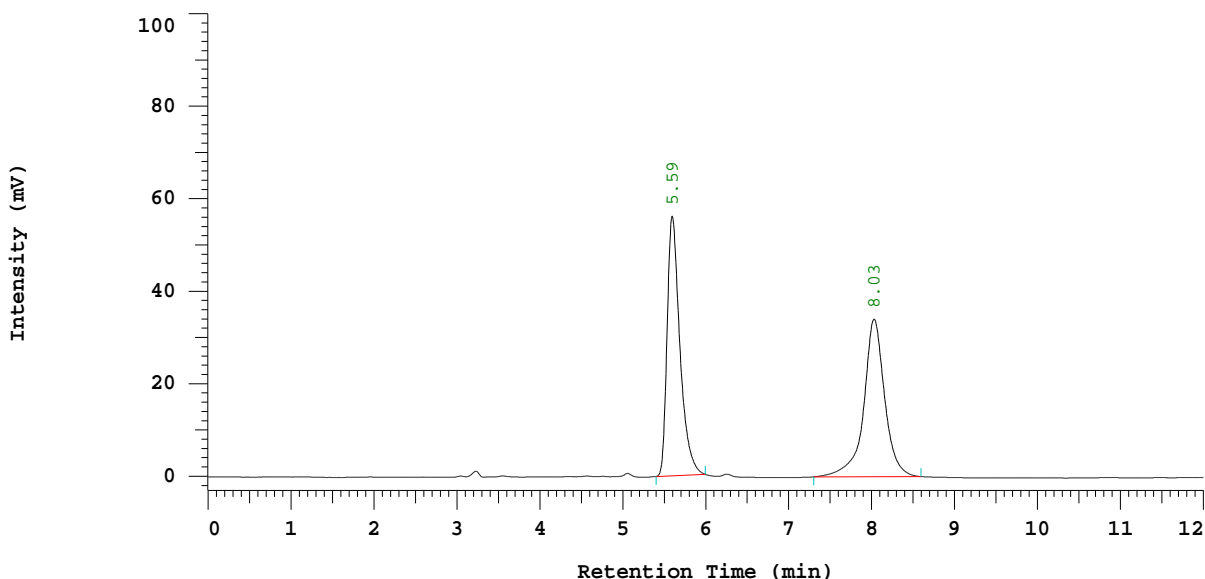
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.59	600148	56147	50.073
2	8.03	598406	34061	49.927
		1198554	90208	100.000

Peak rejection level: 5000

Fig S198. HPLC analysis of a crystal **3i** obtained with racemic mixture (crystal count number 1).

D-2000 Elite HPLC System Manager Report

Analyzed Date and Time: 06/30/2016
05:37 PM

Reported Date and Time: 07/01/2016
11:52 AM

Processed Date and Time: 07/01/2016
11:51 AM

Data Path: D:\Arun\DATA\0518\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0518

Application(data): Arun

Vial Number: 4

Sample Name: CHP-8g-F2 (Crys 4)

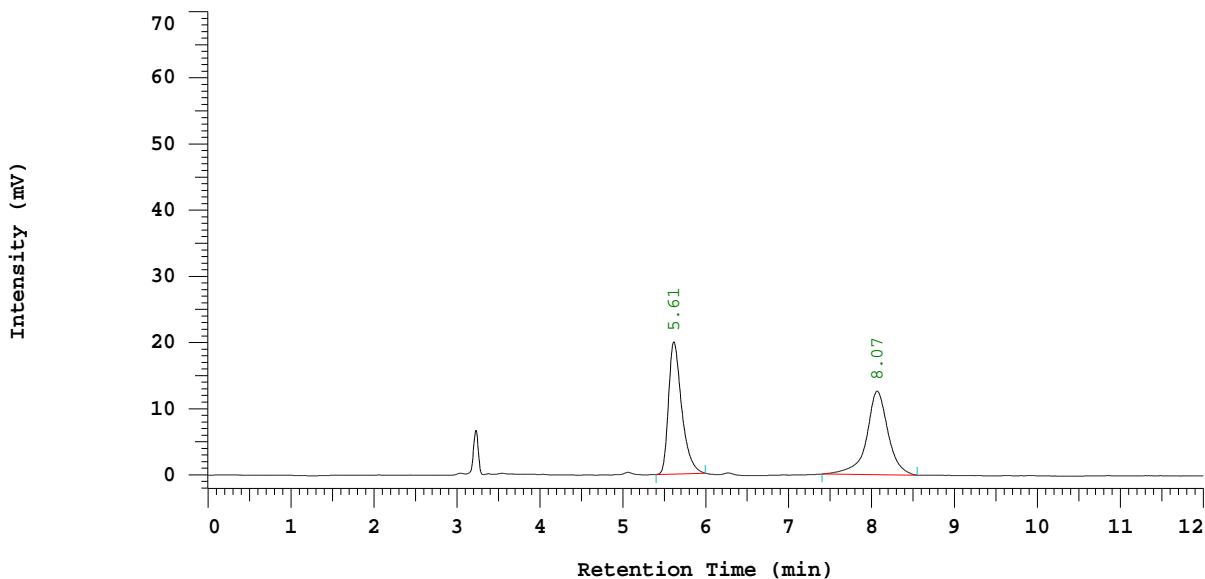
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.61	220086	19960	49.292
2	8.07	226410	12632	50.708
		446496	32592	100.000

Peak rejection level: 5000

Fig S199. HPLC analysis of a crystal **3i** obtained with racemic mixture (crystal count number 2).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/27/2016
05:22 PMReported Date and Time: 07/01/2016
12:38 PMProcessed Date and Time: 07/01/2016
12:37 PM

Data Path: D:\Arun\DATA\0490\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0490

Application(data): Arun

Vial Number: 1

Sample Name: CHP-8g-F2 (Crys 1)

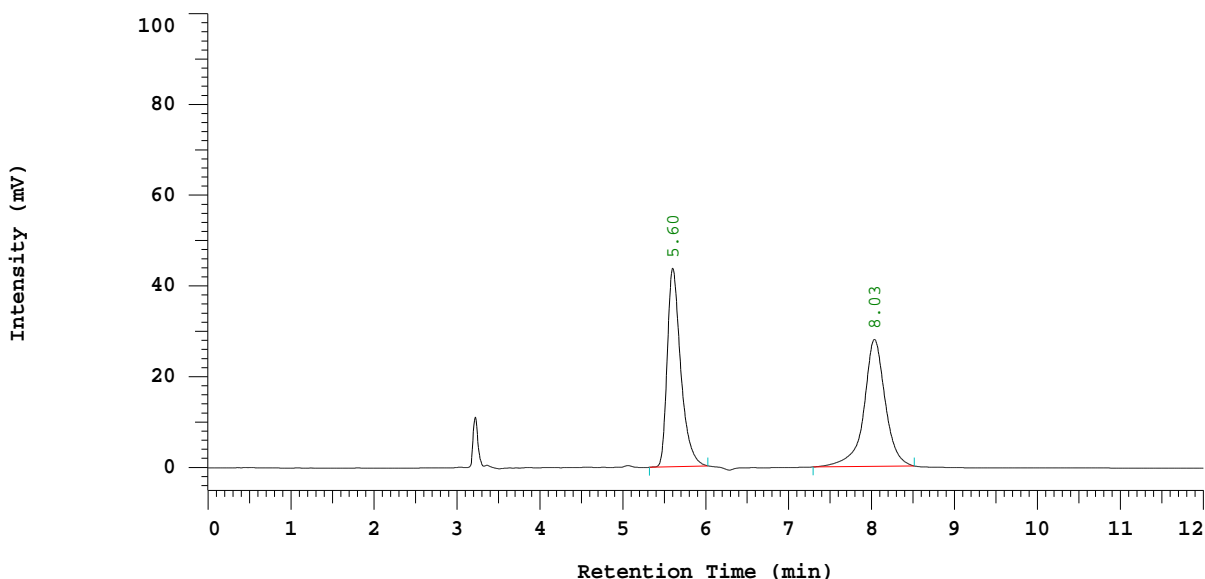
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.60	493609	43720	49.641
2	8.03	500748	27998	50.359
		994357	71718	100.000

Peak rejection level: 5000

Fig S200. HPLC analysis of a crystal **3i** obtained with racemic mixture (crystal count number 3).

D-2000 Elite HPLC System Manager ReportAnalyzed Date and Time: 06/24/2016
01:04 PMReported Date and Time: 07/01/2016
01:03 PMProcessed Date and Time: 07/01/2016
01:03 PM

Data Path: D:\Arun\DATA\0477\

Processing Method: test-THF/Hx-Final II

System (acquisition): Sys 1

Series: 0477

Application(data): Arun

Vial Number: 4

Sample Name: CHP-8g-F2 (Crys 4)

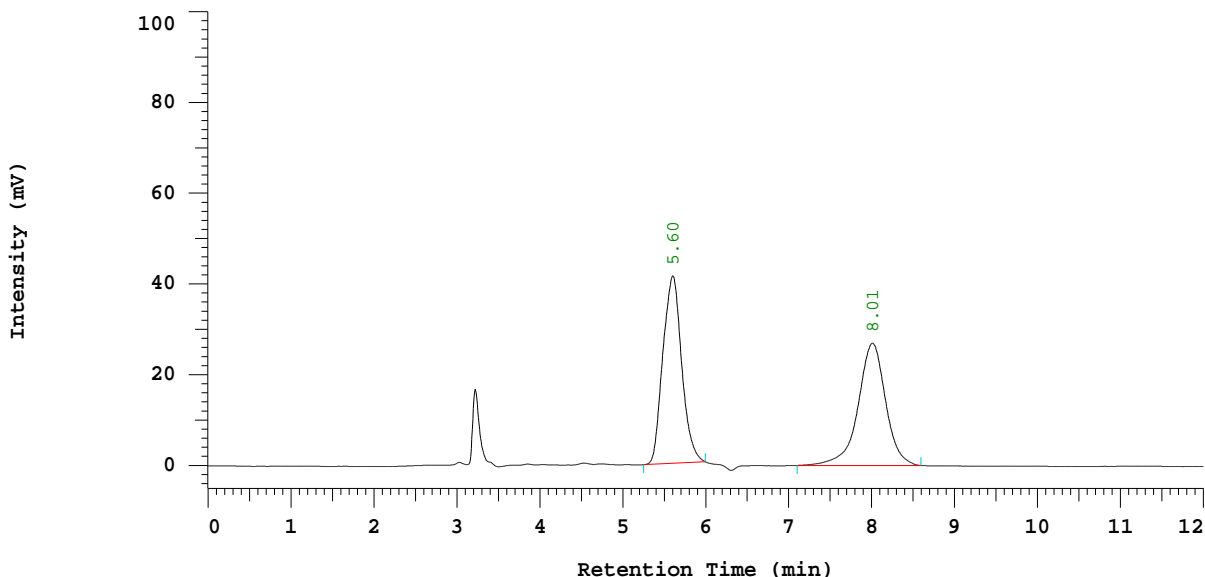
Vial Type: UNK

Injection from this vial: 1 of 1

Volume: 20.0 ul

Sample Description: 25%THF+HX 1.0 mL/MIN COL-IC

Chrom Type: Fixed WL Chromatogram, 254 nm



Processing Method: test-THF/Hx-Final II

Column Type: IC

Method Developer: ARUN

Method Description:

Chrom Type: Fixed WL Chromatogram, 254 nm

Peak Quantitation: AREA

Calculation Method: EXT-STD

Scale Factor 1: 1.000

No.	RT	Area	Height	Area %
1	5.60	650641	41299	51.122
2	8.01	622071	26912	48.878
		1272712	68211	100.000

Peak rejection level: 5000

Fig S201. HPLC analysis of a crystal **3i** obtained with racemic mixture (crystal count number 4).