

Assessing the performance and microbial structure of biofilms adhering on aerated membranes for domestic saline sewage treatment

Hailong Tian¹⁺ • **Jie Liu**²⁺ • **Tengteng Feng**³ • **Haifeng Li**¹ • **Xiaolei Wu**^{7*} •
Baoan Li^{4,5,6*}

¹ *College of Bioengineering, Henan University of Technology, Zhengzhou 450001, PR China*

² *College of Architecture and Urban Planning, Chongqing Jiaotong University, Chongqing 400074, PR China*

³ *Shandong Academy of Environmental Science, Jinan 250013, PR China*

⁴ *State Key Laboratory of Chemical Engineering, Tianjin University, Tianjin 300072, PR China*

⁵ *Collaborative Innovation Center of Chemical Science and Engineering, Tianjin 300072, PR China*

⁶ *Tianjin Key Laboratory of Membrane Science and Desalination Technology, Tianjin University, Tianjin 300072, PR China*

⁷ *Department of Energy and Resources Engineering, College of Engineering, Peking University, Beijing 100871, PR China*

⁺These authors contributed equally to this work.

***Corresponding author:**

Baoan Li (for wastewater treatment) Tel.: +86-22-27407854; Fax: +86-22-27404496;
E-mail: baoan.li@gmail.com; Postal address: 92 Weijin Road, Nankai District,
Tianjin, China.

Xiaolei Wu (for microbiology), Phone/Fax: +86-10-62759047. E-
mail: xiaolei_wu@pku.edu.cn

Supplementary

Table S1 Components of the synthetic wastewater

Synthetic wastewater	Concentration (mg/L)
CH ₃ COONa	See Table 1
NH ₄ Cl	See Table 1
NaHCO ₃	45
KH ₂ PO ₄	80
K ₂ SO ₄	45
MgSO ₄ •7H ₂ O	55
Na ₂ CO ₃	70
Fe ₂ (SO ₄) ₃	0.1
MnSO ₄	0.1
CuSO ₄	0.1
ZnSO ₄	0.1

Table S2 The PCR system for bacterial 16S rDNA

Bacteria	
PCR mixture	
5×Pfu Buffer	10 µl
2.5 µM dNTP	5 µl
341F 5 pmol/µl (forward primer)	1.25 µl
1073R 5 pmol/µl (reverse primer)	1.25 µl
FastPfu Polymerase	1 µl
Template DNA	40-50 ng
ddH ₂ O	29 µl
Total	50 µl
PCR conditions	
95 °C (initial denaturation)	2 min
95 °C (denaturation)	30 s
56 °C (annealing)	30 s
72 °C (elongation)	30 s
	} 24 cycles
72 °C (final elongation)	5 min

Table S3 The PCR system for archaeal 16S rDNA

Archaea	
First round	
PCR mixture	
5×Pfu Buffer	4 µL
2.5 µM dNTP	2 µL
109F 5 pmol/µl (forward primer)	0.5 µL
915R 5 pmol/µl (reverse primer)	0.5 µL
FastPfu Polymerase	0.4 µL
Template DNA	40-50 ng
ddH ₂ O	11.6 µL
Total	20 µL
PCR conditions	
95 °C (initial denaturation)	2 min
95 °C (denaturation)	30 s
54 °C (annealing)	30 s
72 °C (elongation)	30 s
72 °C (final elongation)	5 min
} 28 cycles	
Second round	
PCR mixture	
5×Pfu Buffer	10 µL
2.5 µM dNTP	5 µL
339F 5 pmol/µL (forward primer)	1.25 µL
1048R 5 pmol/µL (reverse primer)	1.25 µL
FastPfu Polymerase	1 µL
DNA	40-50 ng
ddH ₂ O	29 µL
Total	50 µL
PCR conditions	
95 °C (initial denaturation)	2 min
95 °C (denaturation)	30 s
54 °C (annealing)	30 s
72 °C (elongation)	30 s
72 °C (final elongation)	5 min
} 21 cycles	

Fig. S1 Variations in COD_{Cr} , $\text{NH}_4\text{-N}$, TN, $\text{NO}_2\text{-N}$, $\text{NO}_3\text{-N}$ and DO under three salinities in an 24 h HRT.

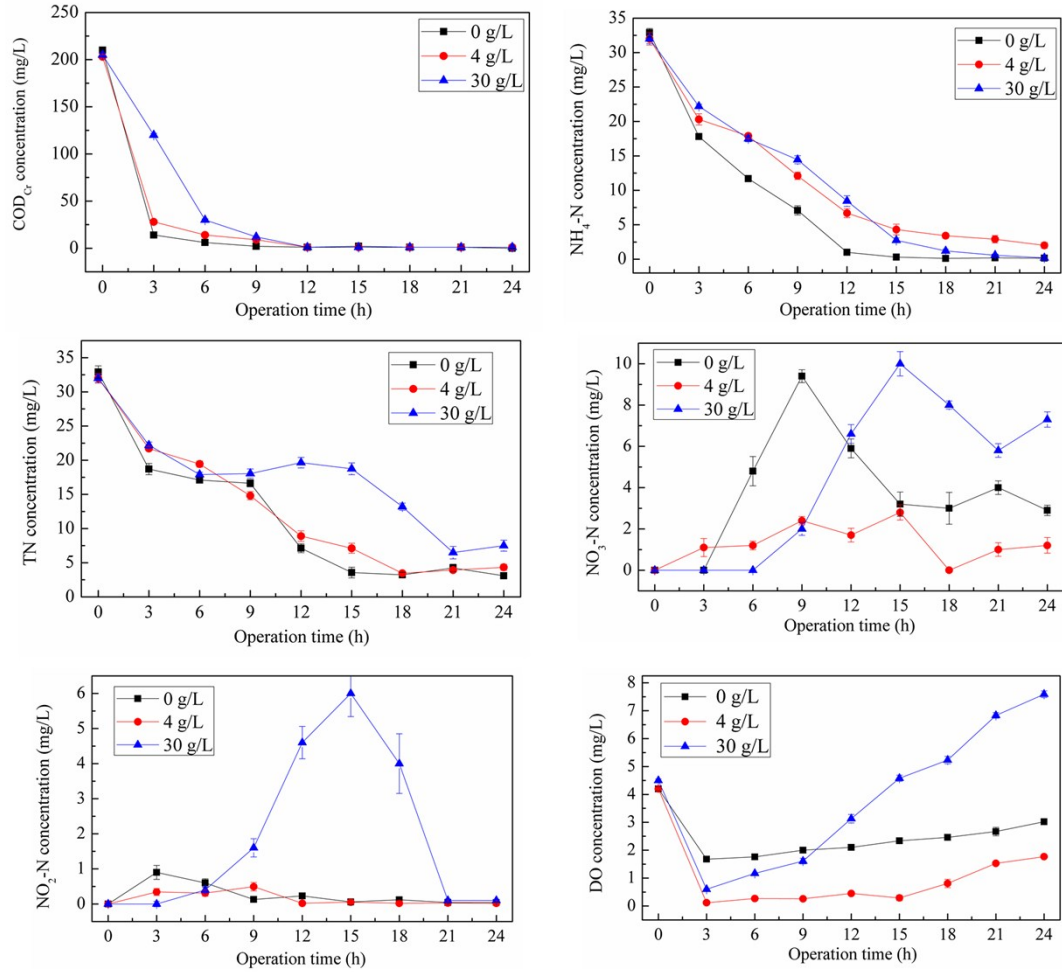


Fig. S2 The bacterial compositions in each biosample at different classification levels. The hot map (c) just listed the identified families no less than 1% in one sample, and the color bar represented the relative abundance in total reads.

