

Supporting Information

Textile supported silver nanoparticles as highly efficient and recyclable heterogeneous catalyst for nitroaromatics reduction at room temperature

Wei Feng,^c Tingting Huang,^b Liqian Gao,^c Xianfeng Yang,^d Wenbin Deng,^c Rui Zhou^{b,*} and Hongjun Liu^{a,*}

a Key Laboratory of Natural Medicine and Immuno-Engineering of Henan Province, Henan University, Kaifeng, Henan 475004, People's Republic of China 475004

b School of Aerospace Engineering, Xiamen University, 422 Siming South Road, Siming District, Xiamen, Fujian Province, People's Republic of China 361005

c School of Pharmaceutical Sciences, Sun Yat-sen University Shenzhen, Guang Dong Province, People's Republic of China 510006

d Analytical and Testing Center, South China University of Technology, Guang Zhou, People's Republic of China, 510640

a* email: hjliu@henu.edu.cn

b* email: rzhou2@xmu.edu.cn

Contents

1 General Procedures and Methods

2 Representative procedure for nanotextile catalyst preparation

3 Reaction optimization table

4 Representative procedure for nanotextile catalyzed nitroaromatic reduction

5 NMR and mass data for products

6 NMR Spectra

1 General Procedures and Methods

Proton nuclear magnetic resonance (^1H NMR) and carbon NMR (^{13}C NMR) spectra were recorded in CD_3OD and CDCl_3 unless otherwise stated. ^1H (400 MHz) and ^{13}C (101 MHz) with complete proton decoupling were performed on a 400 MHz Bruker Ultra Shield NMR spectrometer. Chemical shifts were reported as δ in units of parts per million (ppm) downfield from tetramethylsilane (δ 0.00), using the residual solvent signal as an internal standard: CD_3OD (^1H NMR, δ 3.31; ^{13}C NMR, δ 49.0), CDCl_3 (^1H NMR, δ 7.26 singlet; ^{13}C NMR, δ 77.0 triplet). Multiplicities were given as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (doublet of doublets), ddd (doublet of doublet of doublets), dt (doublet of triplets), dq (doublet of quartets), and br (broad). Coupling constants (J) were recorded in hertz (Hz). The number of proton atoms (n) for a given resonance was indicated by nH. MS was reported in units of mass to charge ratio (m/z). Mass samples were dissolved in MeCN (AR grade). X-Ray Photoelectron Spectroscopy (XPS) was performed on a PerkinElmer, PHI1600 spectrometer. Inductively Coupled Plasma (ICP) was carried out on a Shimadzu ICPE-9800 Series system.

All commercial reagents were purchased from Sigma- Aldrich, Fluka, Alfa Aesar, Merck, TCI, or Acros and were of the highest purity grade available. They were used without further purification unless specified. Reaction solvents were AR grade THF and deionized water.

2 Representative procedure for textile supported nanosilver catalyst preparation

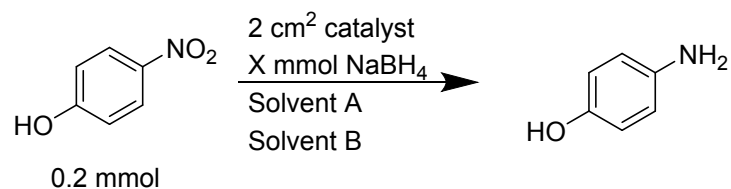
Commercially available white cotton textile was washed with deionized water and dried in oven at $60\text{ }^\circ\text{C}$. Silver nitrate (0.7 g) and (3-Aminopropyl) triethoxysilane (APTES) (10ml) were dissolved in deionized water (1L). The dried cotton textile (30g) was bathed in the solution at $60\text{ }^\circ\text{C}$ for 10 min. The textile was then washed thoroughly with deionized water and soaked for 10 min in a sodium borohydride (5 mM) solution at room temperature. A yellow color was formed on the textile due to the formation of silver nanoparticles on its surface. The textile was ready to use after rinsing with deionized water to remove any excess reactants and un-bonded silver nanoparticles. It was then dried at room temperature in drying cabinet.



Figure S1, Picture of textile before (white color) and after (yellow color) coating with silver nanoparticles.

3 Reaction optimizations

Table S1 Optimization of TsNS catalyzed 4-nitrophenol reduction.



Entry	Solvent A	Solvent B	X	Time (hours)	Yield (%) ^a
1 ^b	2 ml DCM		1	12	trace
2 ^b	2 ml MeCN		1	12	~8%
3 ^b	2ml THF		1	12	~10%
4	1ml THF	1ml H ₂ O	1	3	73%
5	1ml THF	1ml H ₂ O	0.5	3	46%
6	1ml THF	1ml H ₂ O	2	3	75%
7 ^b	1.5 ml THF	0.5 ml H ₂ O	1	3	50%
8 ^c	0.5	1.5	1	3	26%

Nitrophenol, catalyst and NaBH₄ were mixed in solvent and stirred at room temperature. ^a isolated yield. ^b NaBH₄ is not completely dissolved. ^c nitrophenol is not completely dissolved. Entry 4 is the best condition.

4 Representative procedure for nanotextile catalyzed nitroaromatic reduction

To a 25ml rbf was added a magnetic stirring bar, 2 cm² nanosilver textile catalyst, 0.2 mmol nitro compound and 1mmol NaBH₄, following this, 1ml THF and 1ml deionized water were added. The mixture was stirred at room temperature (25°C, 100 °C for certain substrate) until substrate was completely consume as indicated by TLC monitoring. Then the catalyst was taken out and washed thoroughly with deionized water, which was combined with the reaction mixture. Extract with Ether for three times and the organic layers were combined and dried with Na₂SO₄, concentrated and purified by flash chromatography.

5 NMR and Mass data for products

All compounds were purified by flashing silica chromatography with hexane and ethyl acetate (EA) mixture as eluent.

Compound 1: 4-aminophenol

¹H NMR (400 MHz, MeOD) δ 6.62 (td, *J* = 9.0, 6.4 Hz, 4H). ¹³C NMR (101 MHz, MeOD) δ 151.32, 140.22, 118.54, 116.74. LRMS (ESI) *m/z* 110.0 (M + H⁺). Eluent: Hexane/ EA = 5/1.

Compound 2: 2-methyl-1H-indol-5-amine

¹H NMR (400 MHz, MeOD) δ 7.05 (d, *J* = 8.4 Hz, 1H), 6.83 (d, *J* = 2.0 Hz, 1H), 6.57 (dd, *J* = 8.4, 2.1 Hz, 1H), 5.92 (s, 1H), 2.35 (s, 3H). ¹³C NMR (101 MHz, MeOD) δ 139.20, 137.04, 133.32, 131.34, 112.81, 111.55, 107.03, 99.49, 13.49. LRMS (ESI) *m/z* 147.2 (M + H⁺). Eluent: Hexane/ EA = 1/1.

Compound 3: indolin-5-amine

¹H NMR (400 MHz, CDCl₃) δ 6.61 – 6.56 (m, 1H), 6.52 (d, *J* = 8.1 Hz, 1H), 6.42 (dd, *J* = 8.1, 2.3 Hz, 1H), 3.49 (t, *J* = 8.3 Hz, 2H), 3.16 (br, 3H), 2.95 (t, *J* = 8.2 Hz, 2H). ¹³C NMR (101 MHz, CDCl₃) δ 144.11, 139.15, 131.27, 114.27, 113.15, 110.76, 47.57, 30.47. LRMS (ESI) *m/z* 135.0 (M + H⁺). Eluent: Hexane/ EA = 1/1.

Compound 4: aniline

¹H NMR (400 MHz, MeOD) δ 7.24 – 7.14 (m, 2H), 7.00 – 6.93 (m, 2H), 6.85 (tt, *J* = 7.5, 1.1 Hz, 1H). ¹³C NMR (101 MHz, MeOD) δ 152.69, 129.61, 121.97, 115.14. LRMS (ESI) *m/z* 93.9 (M + H⁺). Eluent: Hexane/ EA = 10/1.

Compound 5: 2-(2-aminophenyl) ethanol

^1H NMR (400 MHz, CDCl_3) δ 7.07 (t, $J = 8.2$ Hz, 2H), 6.85 – 6.61 (m, 2H), 3.90 (t, $J = 6.1$ Hz, 2H), 2.96 (br, 3H), 2.80 (t, $J = 6.1$ Hz, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 144.81, 130.60, 127.69, 124.40, 119.37, 116.39, 63.32, 34.75. LRMS (ESI) m/z 138.1 ($\text{M} + \text{H}^+$). Eluent: Hexane/ EA = 1/1.

Compound 6: 3-aminophenol

^1H NMR (400 MHz, MeOD) δ 7.70 (ddd, $J = 8.2, 2.2, 0.9$ Hz, 1H), 7.62 (t, $J = 2.3$ Hz, 1H), 7.42 (t, $J = 8.2$ Hz, 1H), 7.18 (ddd, $J = 8.2, 2.4, 0.9$ Hz, 1H). ^{13}C NMR (101 MHz, MeOD) δ 159.81, 150.65, 131.26, 122.92, 115.09, 110.87. LRMS (ESI) m/z 110.0 ($\text{M} + \text{H}^+$). Eluent: Hexane/ EA = 3/1.

Compound 7: 1-aminonaphthalen-2-ol

^1H NMR (400 MHz, MeOD) δ 7.86 (dd, $J = 8.5, 0.7$ Hz, 1H), 7.68 (d, $J = 8.2$ Hz, 1H), 7.36 (ddd, $J = 8.4, 6.8, 1.3$ Hz, 1H), 7.23 (ddd, $J = 8.8, 5.5, 2.1$ Hz, 2H), 7.08 (d, $J = 8.7$ Hz, 1H). ^{13}C NMR (101 MHz, MeOD) δ 141.79, 130.50, 129.16, 128.04, 126.61, 125.79, 123.68, 121.58, 120.14, 118.31. LRMS (ESI) m/z 182.3 ($\text{M} + \text{Na}^+$). Eluent: Hexane/ EA = 1/1.

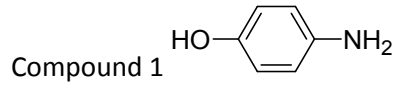
Compound 8: 3-chloroaniline

^1H NMR (400 MHz, MeOD) δ 7.14 (t, $J = 8.0$ Hz, 1H), 6.96 (t, $J = 2.1$ Hz, 1H), 6.84 – 6.77 (m, 2H). ^{13}C NMR (101 MHz, MeOD) δ 154.53, 135.54, 130.83, 121.12, 114.46, 112.90. LRMS (ESI) m/z 127.9 ($\text{M} + \text{H}^+$). Eluent: Hexane/ EA = 10/1.

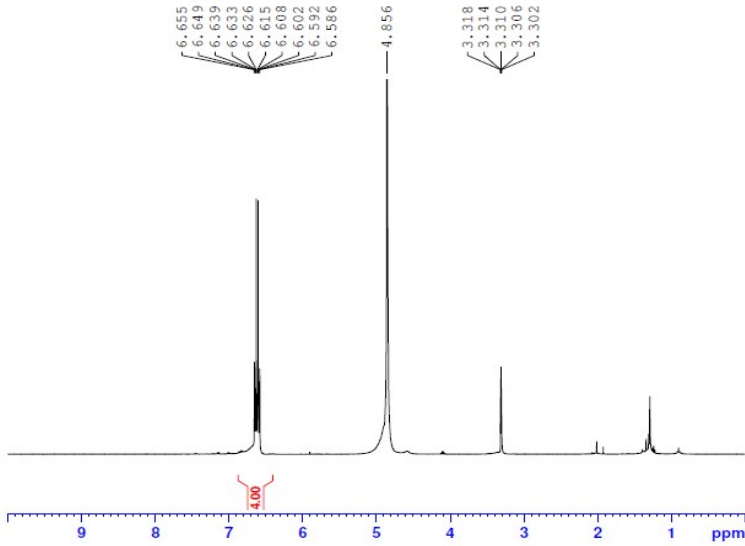
Compound 9: 4-methoxybenzene-1,2-diamine

^1H NMR (400 MHz, CDCl_3) δ 6.63 (d, $J = 8.4$ Hz, 1H), 6.31 (d, $J = 2.7$ Hz, 1H), 6.25 (dd, $J = 8.3, 2.7$ Hz, 1H), 3.72 (s, 3H), 3.29 (s, 4H). ^{13}C NMR (101 MHz, CDCl_3) δ 154.60, 137.09, 127.36, 118.34, 104.10, 102.96, 55.54. LRMS (ESI) m/z 139.0 ($\text{M} + \text{H}^+$). Eluent: Hexane/ EA = 3/1.

6 NMR Spectra



1001B



```

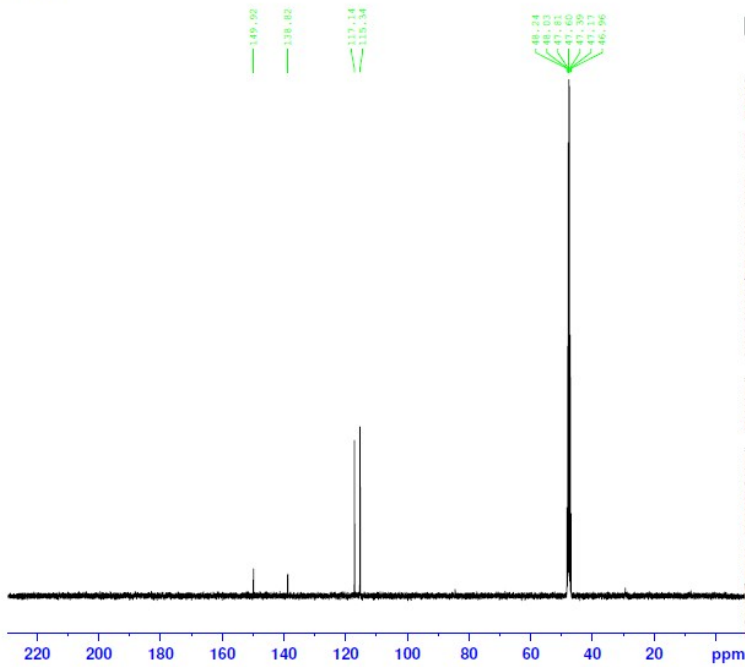
Current Data Parameters
NAME      Jan21fw1001B
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20140121
Time      12.17
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zg30
TD        32768
SOLVENT   MeOD
NS         8
DS         0
SWH       8223.685 Hz
FIDRES    0.250967 Hz
AQ        1.9922944 sec
RG        120.14
DW        60.800 usec
DE        6.50 usec
TE        299.8 K
D1        1.00000000 sec
TDO       1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1      1H
P1        13.35 usec
PLW1      17.20000076 W

F2 - Processing parameters
SI        65536
SF        400.1300078 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
    
```

1001B



```

Current Data Parameters
NAME      Jan21fw1001B
EXPNO     2
PROCNO    1

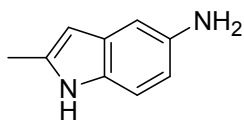
F2 - Acquisition Parameters
Date_     20140121
Time      12.21
INSTRUM   spect
PROBHD    5 mm PABBO BB-
PULPROG   zgpg30
TD        65536
SOLVENT   MeOD
NS         4
DS         4
SWH       24038.461 Hz
FIDRES    0.366798 Hz
AQ        1.3631488 sec
RG        194.02
DW        20.800 usec
DE        6.50 usec
TE        299.6 K
D1        1.50000000 sec
D11       0.03000000 sec
TDO       1

===== CHANNEL f1 =====
SFO1      100.6238364 MHz
NUC1      13C
P1        10.00 usec
PLW1      60.00000000 W

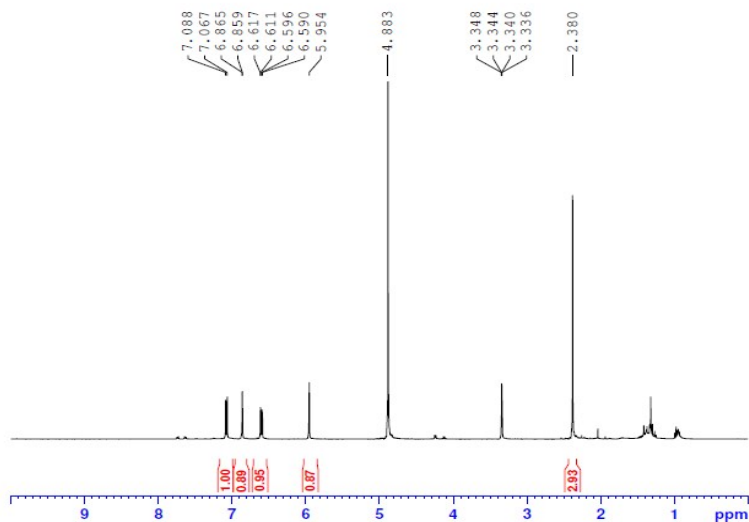
===== CHANNEL f2 =====
SFO2      400.1316005 MHz
NUC2      1H
CPDPRG2   waltz16
PCPD2     80.00 usec
PLW2      17.20000076 W
PLW12     0.47896389 W
PLW13     0.30654001 W

F2 - Processing parameters
SI        32768
SP        100.6127690 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
    
```

Compound 2



1003C



```

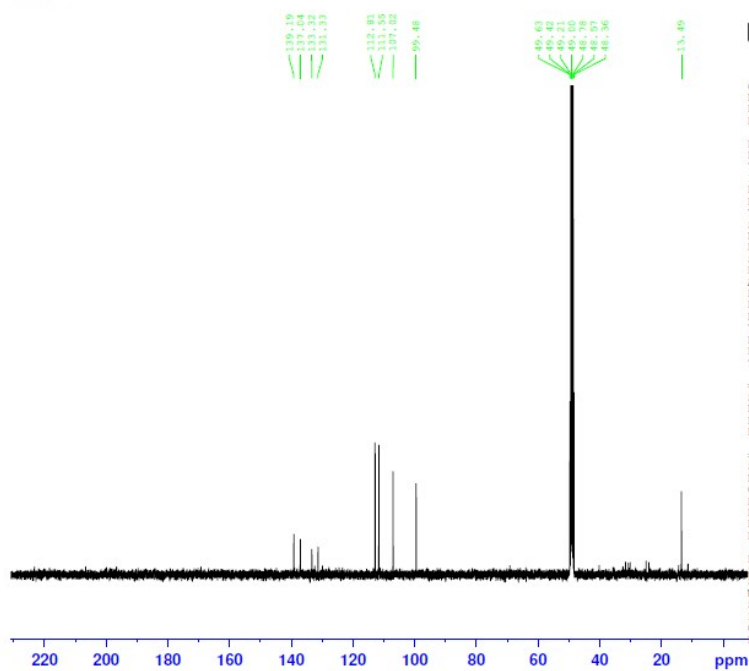
Current Data Parameters
NAME      Jan17fw1003C
EXPNO    3
PROCNO   1

F2 - Acquisition Parameters
Date_    20140117
Time     13.00
INSTRUM spect
PROBHD   5 mm PABBO BB-
PULPROG zg30
TD       32768
SOLVENT  MeOD
NS       8
DS       0
SWH      8223.685 Hz
FIDRES   0.250967 Hz
AQ       1.8922944 sec
RG       120.14
DW       60.800 usec
DE       6.50 usec
TE       298.7 K
D1       1.0000000 sec
TDO      1

===== CHANNEL f1 =====
SFO1    400.1324710 MHz
NUC1     1H
P1       13.35 usec
PLW1    17.20000076 W

F2 - Processing parameters
SI       65536
SF       400.1293897 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
```

1003C



```

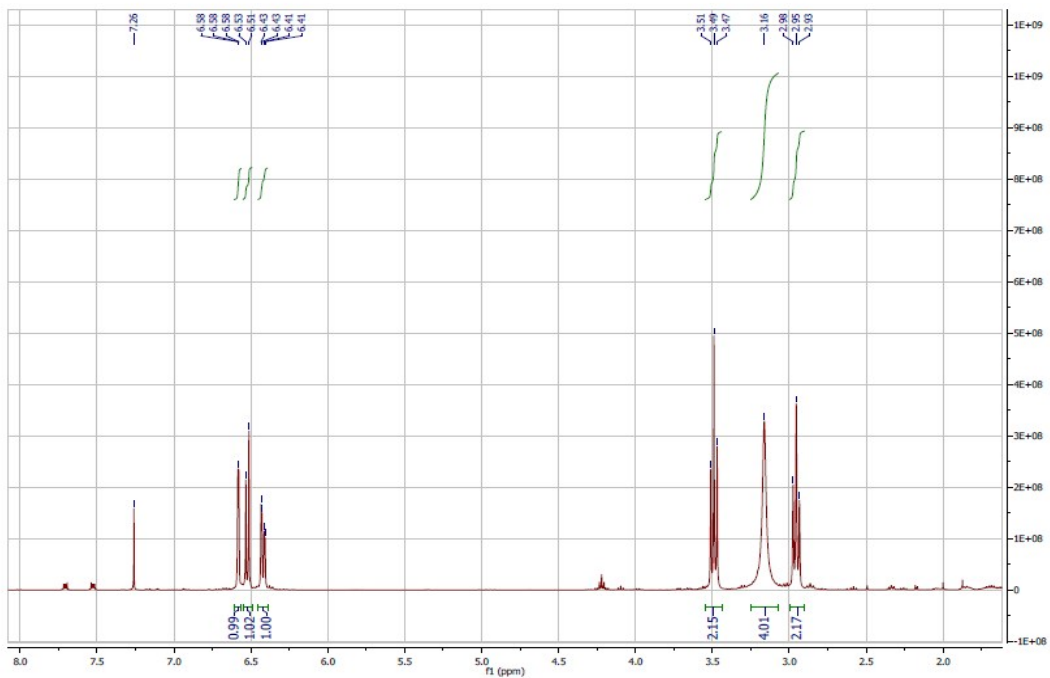
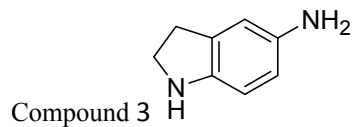
Current Data Parameters
NAME      Jan17fw1003C
EXPNO    5
PROCNO   1

F2 - Acquisition Parameters
Date_    20140117
Time     14.43
INSTRUM spect
PROBHD   5 mm PABBO BB-
PULPROG zgpg30
TD       65536
SOLVENT  MeOD
NS       300
DS       4
SWH      24038.461 Hz
FIDRES   0.366798 Hz
AQ       1.3631488 sec
RG       194.02
DW       20.800 usec
DE       6.50 usec
TE       299.7 K
D1       1.5000000 sec
D11      0.03000000 sec
TDO      1

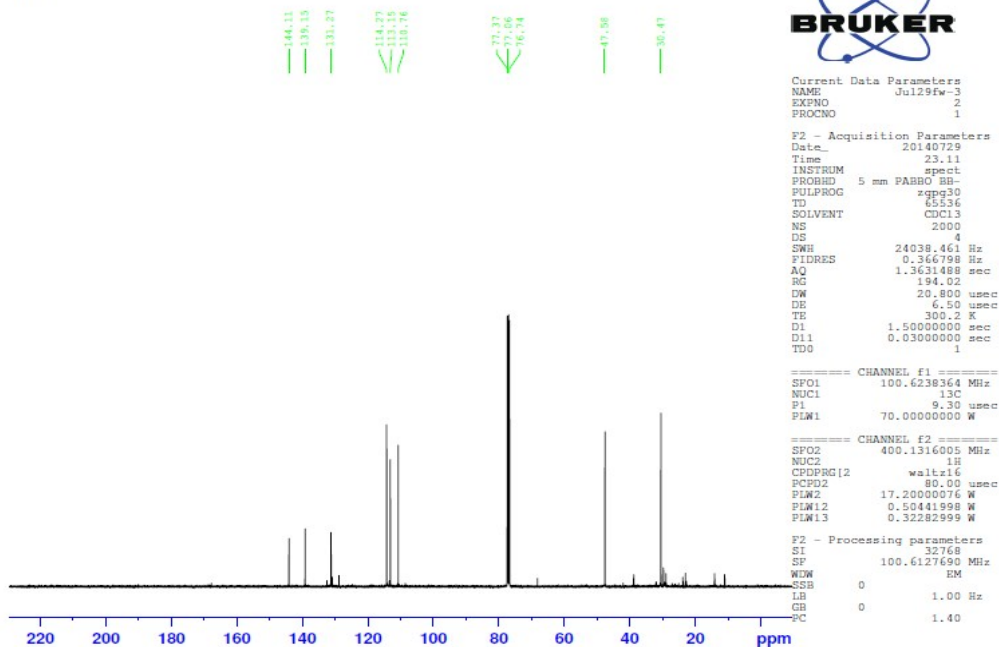
===== CHANNEL f1 =====
SFO1    100.6238364 MHz
NUC1     13C
P1       10.00 usec
PLW1    60.00000000 W

===== CHANNEL f2 =====
SFO2    400.1316005 MHz
NUC2     1H
CPDPRG2 waltz16
PCPD2   80.00 usec
PLW2    17.20000076 W
PLW12   0.47896999 W
PLW13   0.30654001 W

F2 - Processing parameters
SI       32768
SF       100.6126267 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
    
```

3 H



```

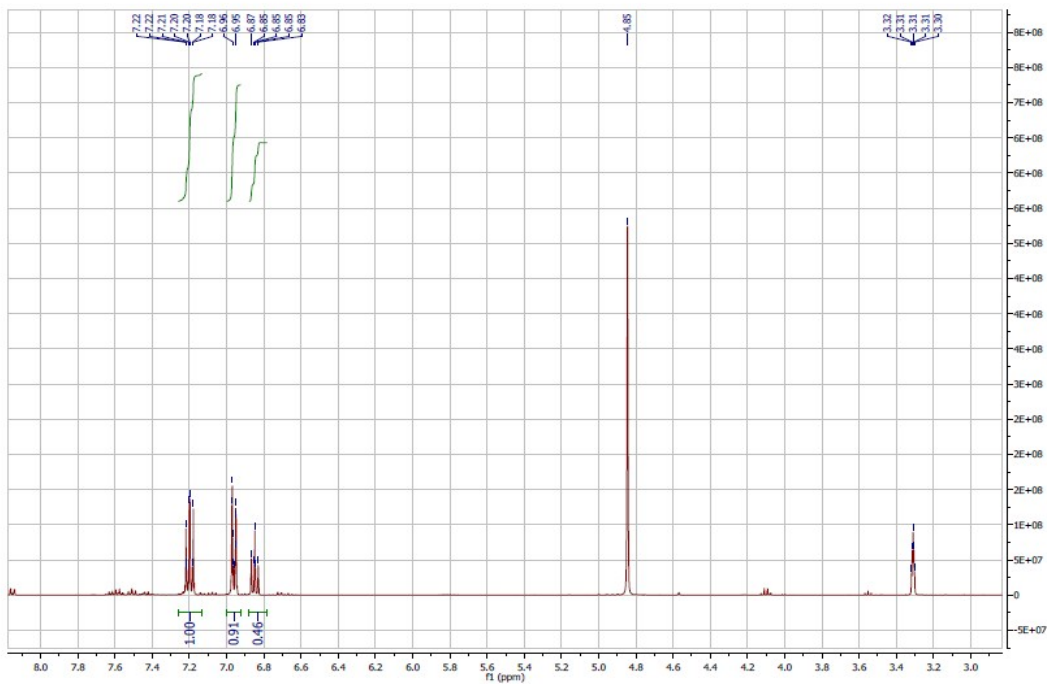
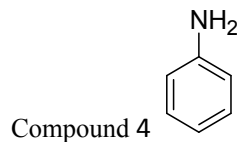
Current Data Parameters
NAME      Jul129fw-3
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20140729
Time     23.11
INSTRUM spect
PROBHD   5 mm PABBO BB-
PULPROG zgpg30
TD       65536
SOLVENT  CDCl3
NS       2000
DS       4
SWH      24038.461 Hz
FIDRES   0.366798 Hz
AQ       1.3631488 sec
RG       194.02
DW       20.800 usec
DE       6.50 usec
TE       300.2 K
D1       1.50000000 sec
D11      0.03000000 sec
TD0      1

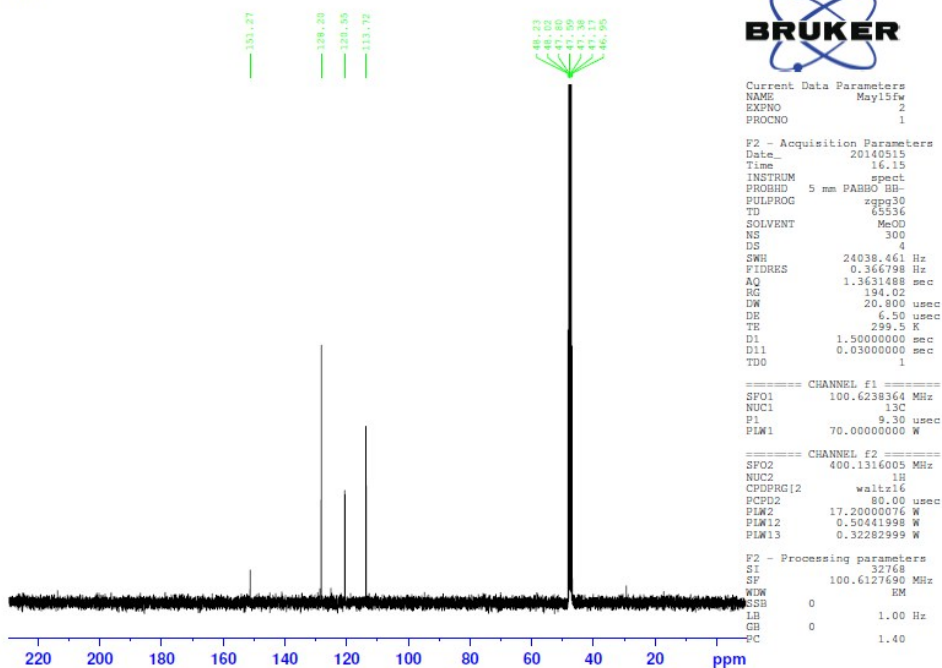
===== CHANNEL f1 =====
SFO1    100.628364 MHz
NUC1    13C
P1      9.30 usec
PLW1    70.0000000 W

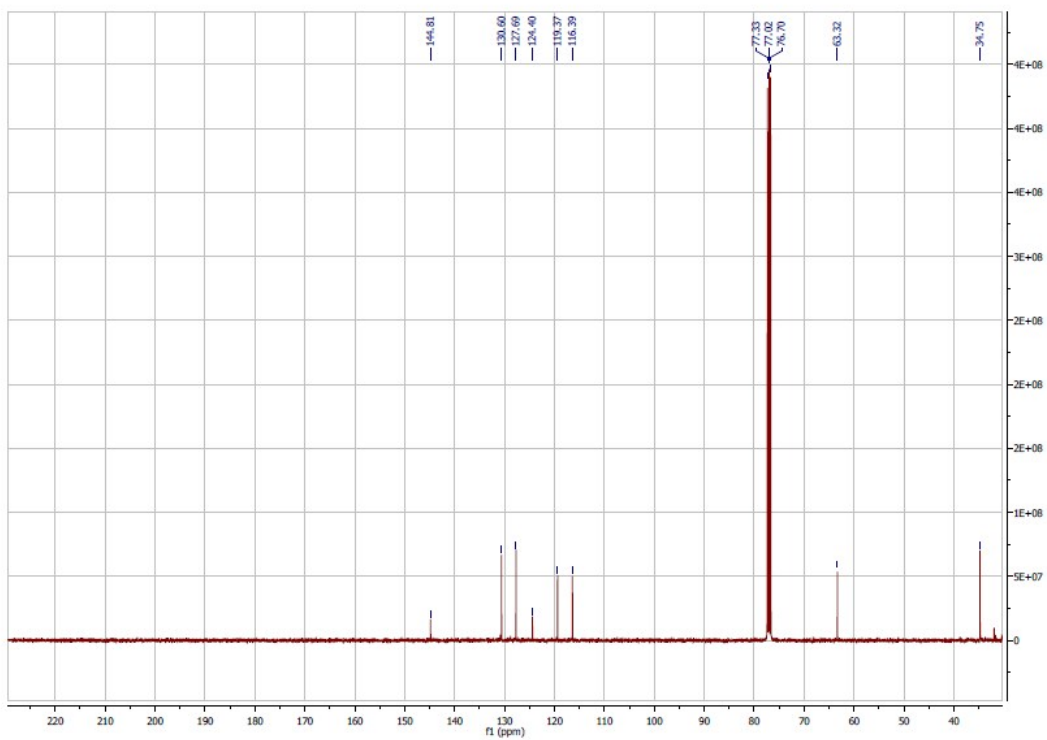
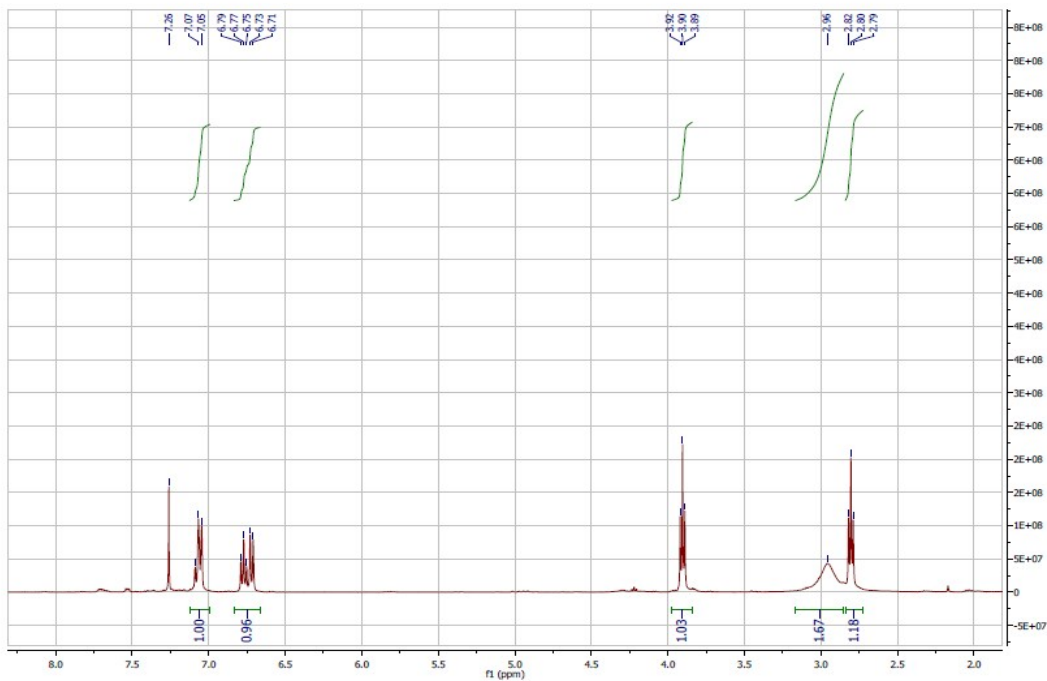
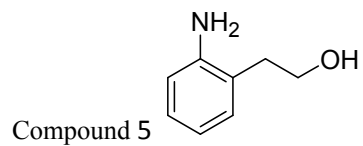
===== CHANNEL f2 =====
SFO2    400.1316005 MHz
NUC2    1H
CPDPRG2 waltz16
PCPD2   80.00 usec
PLW2    17.2000076 W
PLW12   0.5041998 W
PLW13   0.32282999 W

F2 - Processing parameters
SI      32768
SF      100.6127690 MHz
WDW     EM
SSB     0
LB      1.00 Hz
GB      0
PC      1.40
  
```

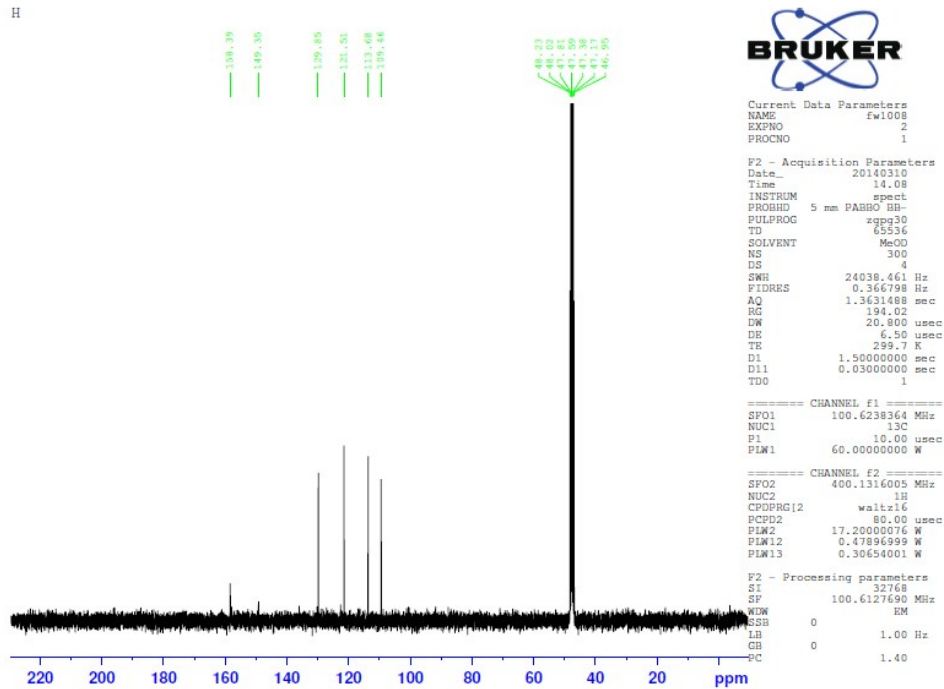
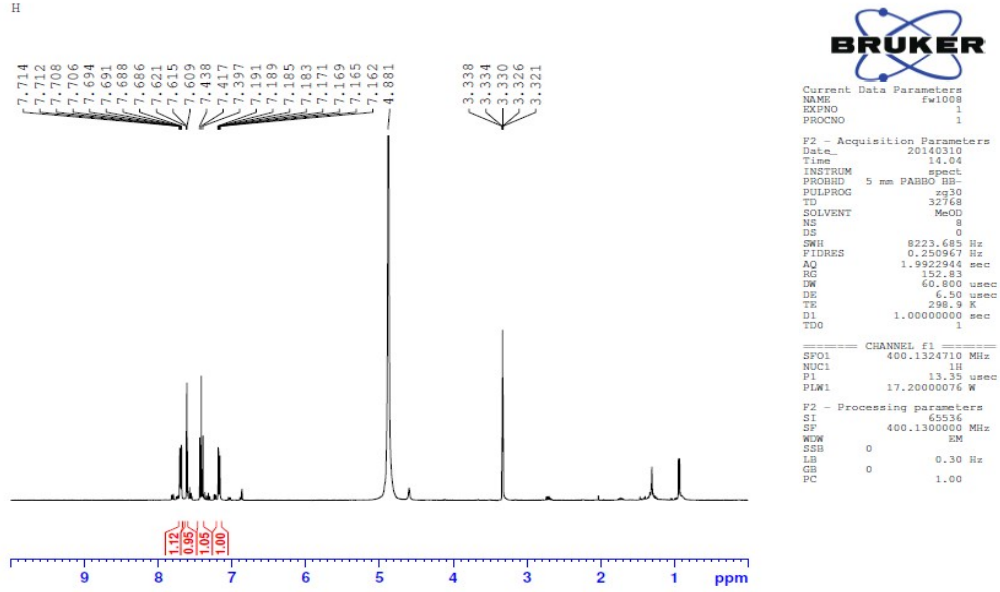
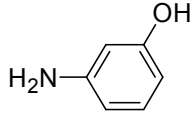


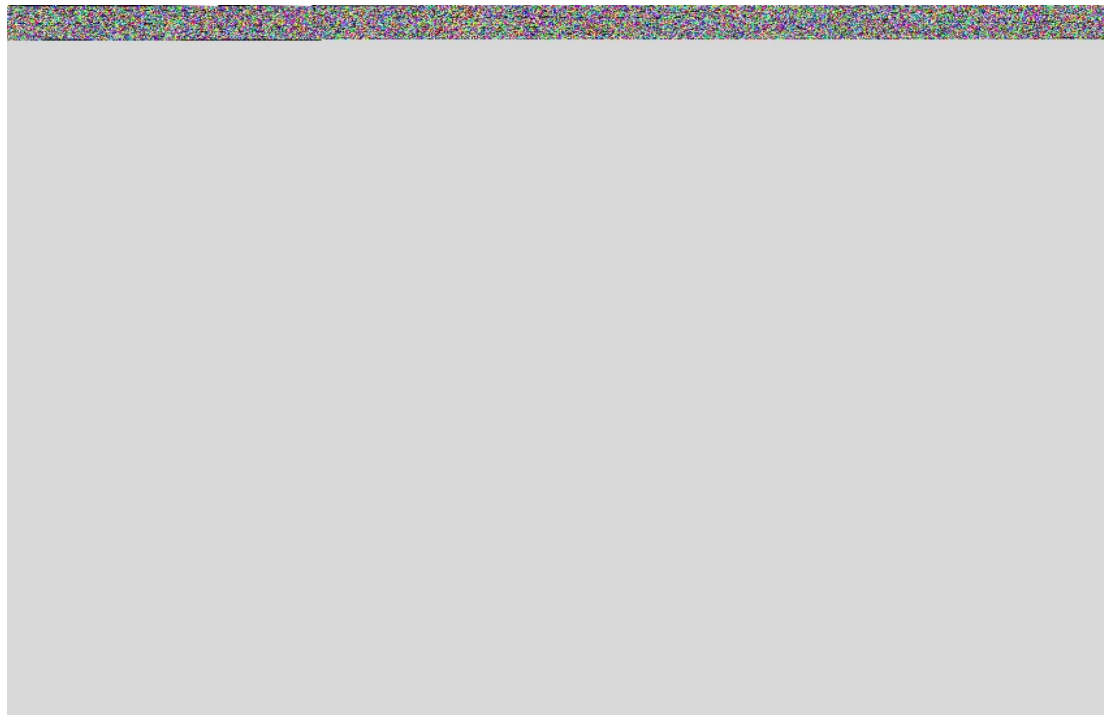
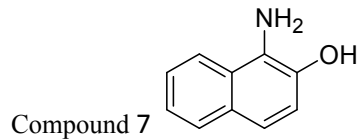
r4 H



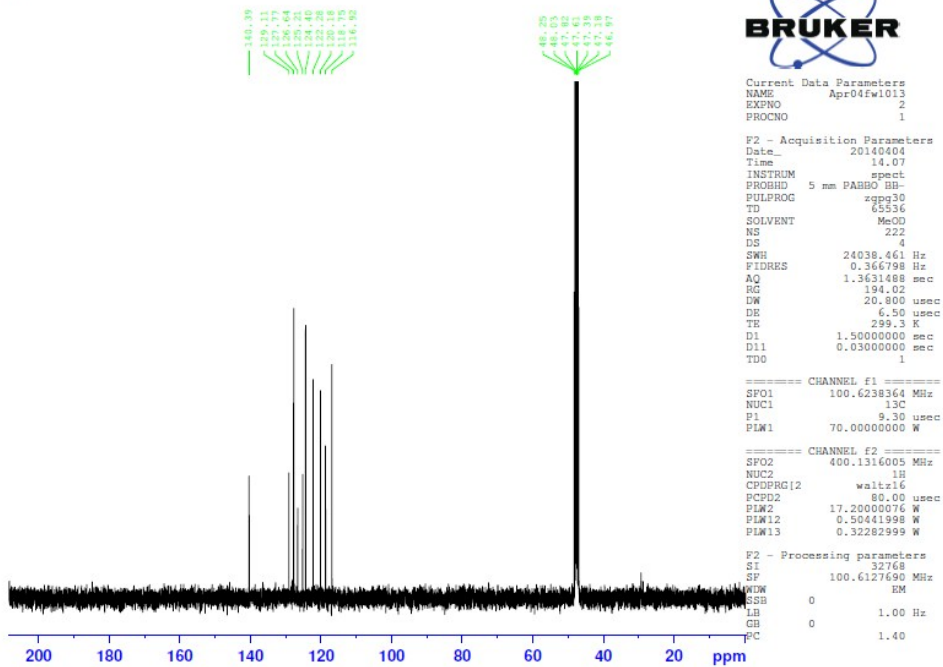


Compound 6

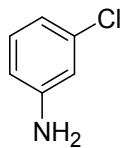




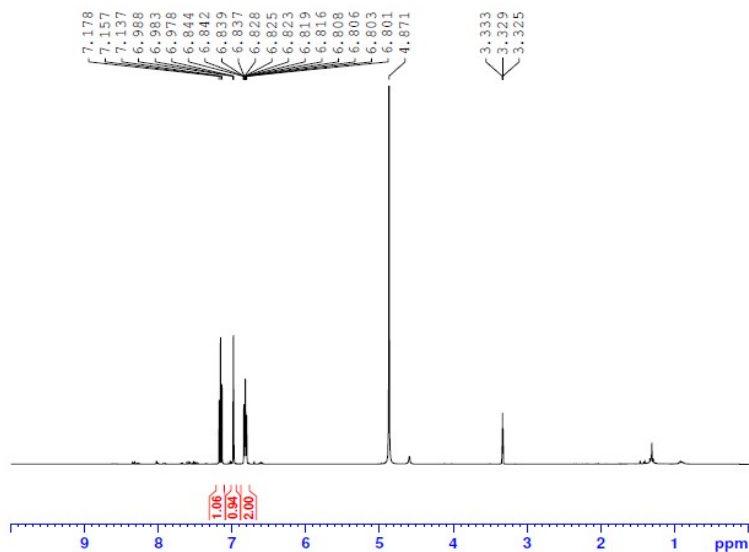
H



Compound 8



1014



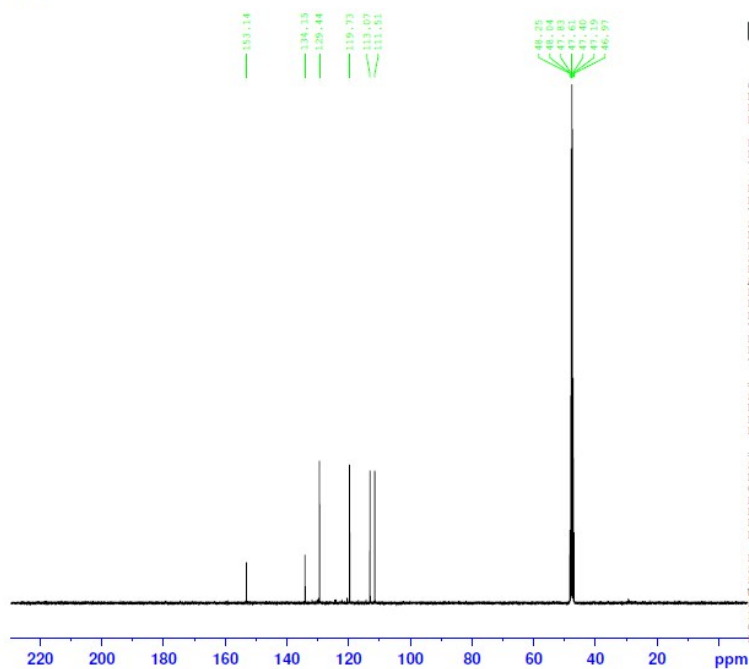
Current Data Parameters
 NAME Apr08fw
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20140408
 Time 22.09
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT MeOD
 NS 8
 DS 0
 SWH 8223.685 Hz
 FIDRES 0.250867 Hz
 AQ 1.9922944 sec
 RG 107.13
 DW 60.800 usec
 DE 6.50 usec
 TE 299.3 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 400.1324710 MHz
 NUC1 1H
 P1 13.70 usec
 PLW1 17.20000076 W

F2 - Processing parameters
 SI 65536
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1014



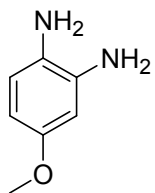
Current Data Parameters
 NAME Apr08fw
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20140408
 Time 23.00
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT MeOD
 NS 1024
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 194.02
 DW 20.800 usec
 DE 6.50 usec
 TE 300.2 K
 D1 1.5000000 sec
 D11 0.0300000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 100.6238364 MHz
 NUC1 13C
 P1 8.30 usec
 PLW1 70.00000000 W

===== CHANNEL f2 =====
 SFO2 400.1316005 MHz
 NUC2 1H
 CPDPRG2 waitz16
 PCPD2 80.00 usec
 PLW2 17.20000076 W
 PLW12 0.50441998 W
 PLW13 0.32282999 W

F2 - Processing parameters
 SI 32768
 SF 100.6127690 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



Compound 9

