

**Supporting information for:**

**Thiol-Ene Photoimmobilization of Chymotrypsin on  
Polysiloxane Gels for Enzymatic Peptide Synthesis**

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## General Considerations

Poly[3-mercaptopropylmethylsiloxane] (PMMS, SMS-992,  $M_w$  4000-7000, 95 cst) was purchased from Gelest Inc. Poly(ethylene glycol)diacrylate (average  $M_n \sim 700$ ) was purchased from Aldrich Inc. 2,2-Dimethoxy-2-phenylacetophenone (DMPA), chymotrypsin (from bovine pancreas), *N*-benzoyl-*L*-tyrosine ethyl ester (Bz-Tyr-OEt), *N*-acetyl-*L*-phenylalanine ethyl ester (Ac-Phe-OEt), *N*-acetyl-*L*-tryptophan ethyl ester (Ac-Trp-OEt), *L*-phenylalaninamide (Phe-NH<sub>2</sub>), *L*-tyrosine amide (Tyr-NH<sub>2</sub>), glycylamide (Gly-NH<sub>2</sub>), alaninamide (Ala-NH<sub>2</sub>), anhydrous organic solvents (hexane, THF, toluene, DMF, DMSO) were purchased from Aladdin (Shanghai) Inc. All flash chromatography were performed using Macherey-Nagel MN Kieselgel 60 (0.063-1.2 mm).

All <sup>1</sup>H NMR spectra were obtained using a Bruker HW500 MHz spectrometer (AVANCE AV-500) and recorded in DMSO-*d*<sub>6</sub> (internal reference 2.50 ppm).

Fourier transform infrared (FT-IR) spectra were obtained on a Nicolet 5700 FT-IR spectrometer.

Matrix-assisted laser desorption ionization time-of-flight (MALDI-TOF) mass spectrometry was performed on an AB Sciex 5800 MALDI-TOF spectrometry. Protein solution (0.5-1.0 mg/ mL) was mixed with an equal volume of matrix (0.5 mL of water, 0.5 mL of acetonitrile, 2 μL of trifluoroacetic acid, and 8.0 mg of 4-hydroxy-3,5-dimethoxycinnamic acid), and 2 μL of the resulting mixture was spotted on the plate target.

An ultraviolet (UV) lamp (230V, 0.20A,  $\lambda = 365$  nm, LUYOR-3109, Shanghai LUYOR Instrument Co, Ltd) was used to irradiate the samples to perform the photocrosslinking reaction.

### **Enzyme activity assay**

Chymotrypsin (from bovine pancreas, lyophilized powder) activity is determined by hydrolyzing *N*-benzoyl-*L*-tyrosine ethyl ester (BTEE). Unit of enzyme activity (U)<sup>[1]</sup> is defined as: 1 mg of protein hydrolyzes 1.0 μmol of BTEE per min at pH 7.8 at 25 °C. The assay mixture was composed of 1.42 mL of Tris–HCl buffer (80 mM, pH 7.8), 1.4 mL of 1.18 mM BTEE and 0.08 ml of 2 M CaCl<sub>2</sub>. After addition of 0.1 mL of enzyme solution, the reaction was carried out at 25 °C for 3 min. The suspension was immediately separated by an external magnetic field of 0.5 T and the absorbance of the solution was measured at 256 nm. The specific activity was calculated as follows:

$$\text{Specific activity (U/mg min)} = \frac{\Delta A}{0.964 * Ew * 3 * 3} = 48 \text{ units/mg protein}$$

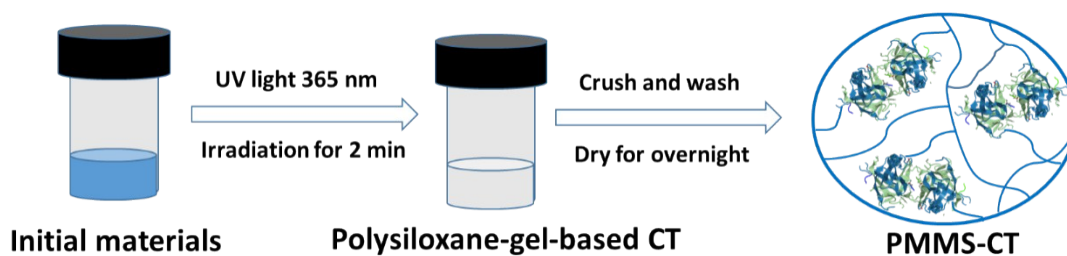
where ΔA was the absorbance change of the solution at 256 nm, Ew represented the amount of enzyme contained in 0.1 ml of enzyme solution, 0.964 was the molar extinction coefficient of *N*-benzoyl-*L*-tyrosine at 256 nm.

### **Enzyme modification**

Chymotrypsin (2.00 g, 0.08 mmol of protein, 1.04 mmol of –NH<sub>2</sub> groups) was dissolved in 200 mL phosphate buffer (0.1 M, pH 8.0). 10-Undecenoyl chloride (3.24 g, 16 mmol, the molar ratio of enzyme to 10-undecenoyl chloride is 1:200) dissolved in dichloromethane (20 mL) was added dropwise to the above phosphate buffer solution.<sup>[2]</sup> The two-phase reaction mixture was vigorously stirred at 25 °C and the pH value was maintained at 7.5~8.0 (pH 7.5~8.0 was chosen to increase the nucleophilicity of the amino groups of lysine fragments in the protein towards 10-undecenoyl chloride) by the addition of 0.1 M sodium hydroxide solution. After 36 h, the vinyl-modified proteins were purified by dialysis against phosphate buffer (0.1 M, pH 8.0) for 48 h using a dialysis cube with molecular weight cut-off (MWCO) of 5000 Da, followed by lyophilization for 48 h.

### **Typical preparation procedure of PMMS-CT**

As shown in Figure S1, vinyl-modified proteins chymotrypsin (50.00 mg), PMMS (100.00 mg, 0.75 mmol of –SH groups), poly (ethylene glycol) diacrylate (140.00 mg, 0.2 mmol, as cross-linker), DMPA (10 mg, as photo-initiator), and dichloromethane (2 mL) were added in a 10 mL transparent vial tube.<sup>[3,4]</sup> The mixture was ultrasonicated for 2 min to ensure a homogeneous dispersion, and then UV irradiated at 365 nm at room temperature for 2 min. The biocatalyst gel was carefully crushed to small particles, washed 3 times with dichloromethane to remove any unreacted chymotrypsin and DMPA, and dried in a fume hood overnight, to provide the polysiloxane-based biocatalyst gel PMMS-CT (274.00 mg) which was stored for future uses.



**Figure S1.** Typical preparation procedure of biopolymer PMMS-CT catalyst.

## Enzymatic synthesis of dipeptide Bz-Tyr-Phe-NH<sub>2</sub>

In a 10 mL round-bottom flask, acyl donor *N*-benzoyl-*L*-tyrosine ethyl ester (Bz-Tyr-OEt, 250.8 mg, 0.8 mmol), and acyl acceptor *L*-Phenylalaninamide (Phe-NH<sub>2</sub>, 262.4 mg, 1.6 mmol), distilled water (40  $\mu$ L, 1.0 vol%), PMMS-CT (100.00 mg, ca. 17.2 mg chymotrypsin immobilized) were added in 4 mL hexane/THF (v/v= 1:1). The reaction mixture was stirred at 37  $^{\circ}$ C for 3 h.<sup>[5]</sup> The crude product mixture was dissolved in MeOH (40 mL) and PMMS-CT was separated by filtration. The resulting solvent was evaporated and the crude product was first well-dispersed in benzene, then filtered, washed with benzene and water, and finally dried under vacuum to give the dipeptide Bz-Tyr-Phe-NH<sub>2</sub> (337.9 mg, Yield: 98%, m.p. = 256-259  $^{\circ}$ C) as a white solid. Bz-Tyr-Phe-NH<sub>2</sub>: <sup>1</sup>H-NMR (300 MHz, DMSO)  $\delta_{\text{H}}$  8.51 (1 H, d,  $J$  = 8.2 Hz), 8.06 (1 H, d,  $J$  = 8.1 Hz), 7.77 (2 H, d,  $J$  = 7.2 Hz), 7.54 – 7.38 (3 H, m), 7.28 – 7.10 (5 H, m), 7.06 (2 H, d,  $J$  = 8.4 Hz), 6.61 (2 H, d,  $J$  = 8.3 Hz), 5.33 (1 H, s), 4.46 (2 H, m), 3.15 – 2.65 (5 H, m). <sup>13</sup>C NMR (75 MHz, DMSO):  $\delta$  173.16, 171.66, 166.80, 156.13, 138.22, 134.52, 131.80, 130.51, 129.71, 128.82, 128.66, 128.47, 127.87, 126.69, 115.32, 55.80, 54.21, 38.10, 36.46. ESI-MS  $m/z$ : 430.2 [M - H]<sup>-</sup>, calculated for Bz-Tyr-Phe-NH<sub>2</sub>, 431.2.

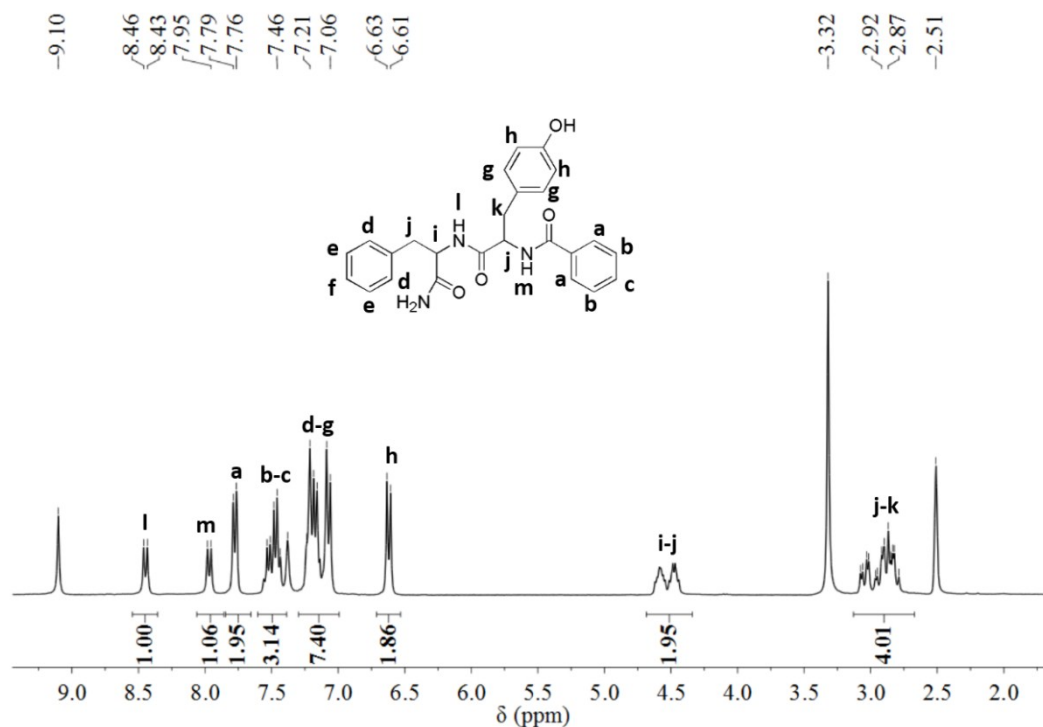
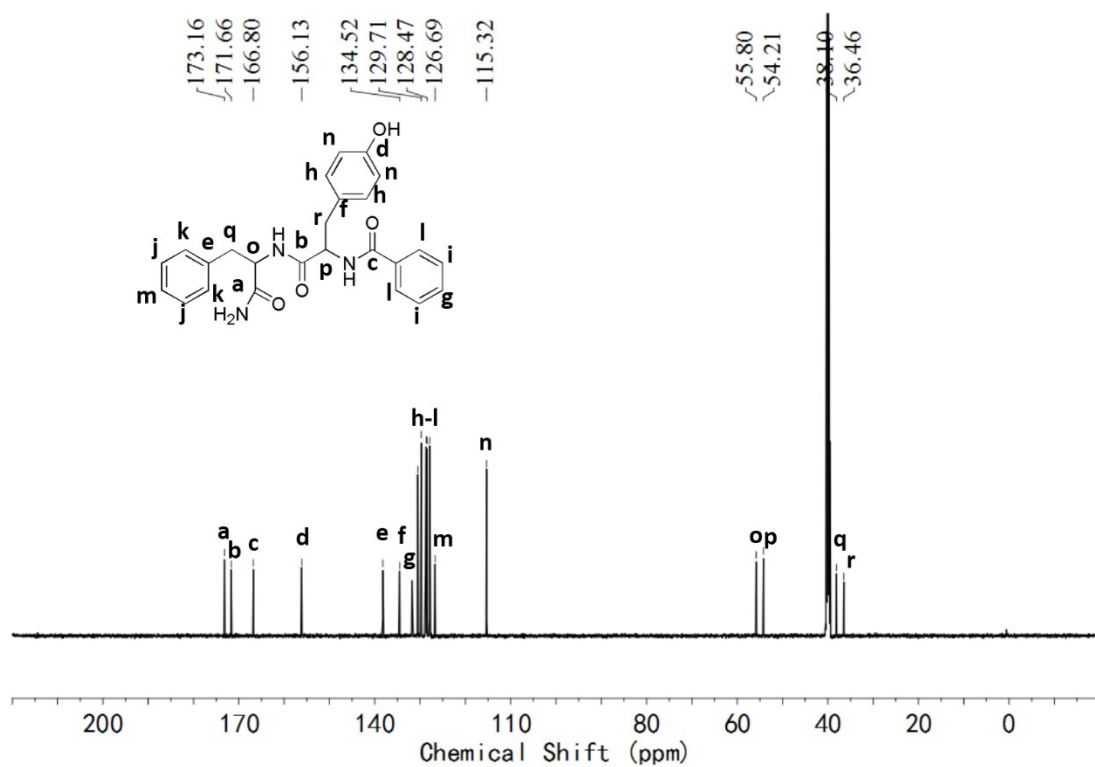
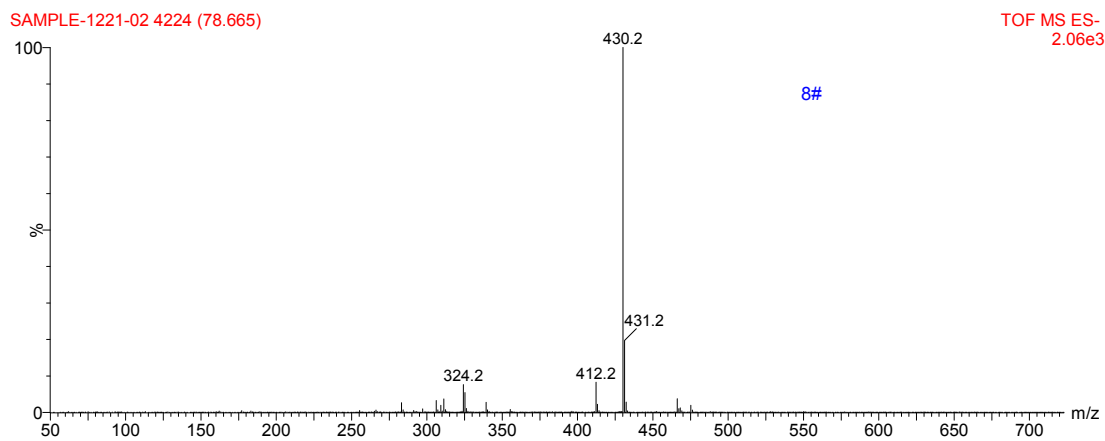


Figure S2. <sup>1</sup>H-NMR spectrum of Bz-Tyr-Phe-NH<sub>2</sub> in DMSO.



**Figure S3.** <sup>13</sup>C-NMR spectrum of Bz-Tyr-Phe-NH<sub>2</sub> in DMSO.



**Figure S4.** ESI-MS data of Bz-Tyr-Phe-NH<sub>2</sub>.

### Peptide synthesis catalyzed by PMMS-CT in low-water organic solvent

In a 10 mL round-bottom flask, distilled water (40  $\mu$ L, 1.0 vol%), PMMS-CT (100 mg, ca. 17.2 mg chymotrypsin immobilized), 0.8 mmol acyl donor (Bz-Tyr-OEt, Ac-Phe-OEt, or Ac-Trp-OEt) and 1.6 mmol acyl acceptor (Phe-NH<sub>2</sub> or Tyr-NH<sub>2</sub>) were added into 4 mL hexane/ tetrahydrofuran (v/v= 1:1). The reaction mixture was stirred at 37  $^{\circ}$ C for 3 h. The crude product mixture was dissolved in MeOH (40 mL) and PMMS-CT was separated by filtration. The resulting solvent was evaporated and the crude product was purified by flash column chromatography to give the desired product.

Ac-Trp-Phe-NH<sub>2</sub> (257.40 mg, yield: 82%, m.p. = 206-208  $^{\circ}$ C) as a white solid, purified by flash column chromatography using ethyl acetate/petroleum ether (v/v= 1/2) as the eluent. <sup>1</sup>H-NMR (300 MHz, DMSO)  $\delta_{\text{H}}$  8.03 (1 H, d, *J* 7.8), 7.83 (1 H, d, *J* 8.1), 7.55 (0 H, d, *J* 7.7), 7.36 – 7.20 (2 H, m), 7.13 – 6.94 (5 H, m), 6.64 (2 H, d, *J* 8.2), 5.75 (0 H, s), 4.47 (1 H, s), 4.34 (1 H, d, *J* 5.1), 2.88 (4 H, ddd, *J* 43.3, 34.5, 6.4), 2.09 (2 H, s). <sup>13</sup>C NMR (75 MHz, DMSO):  $\delta$  174.62, 173.22, 171.25, 139.64, 137.87, 131.16, 131.02, 130.03, 129.82, 129.11, 128.26, 128.02, 125.23, 122.63, 120.11, 120.01, 113.09, 112.03, 63.07, 55.57, 55.48, 39.19, 29.15, 24.32. ESI-MS *m/z*: 391.2 [M - H]<sup>-</sup>, calculated for Ac-Trp-Phe-NH<sub>2</sub>, 392.2.

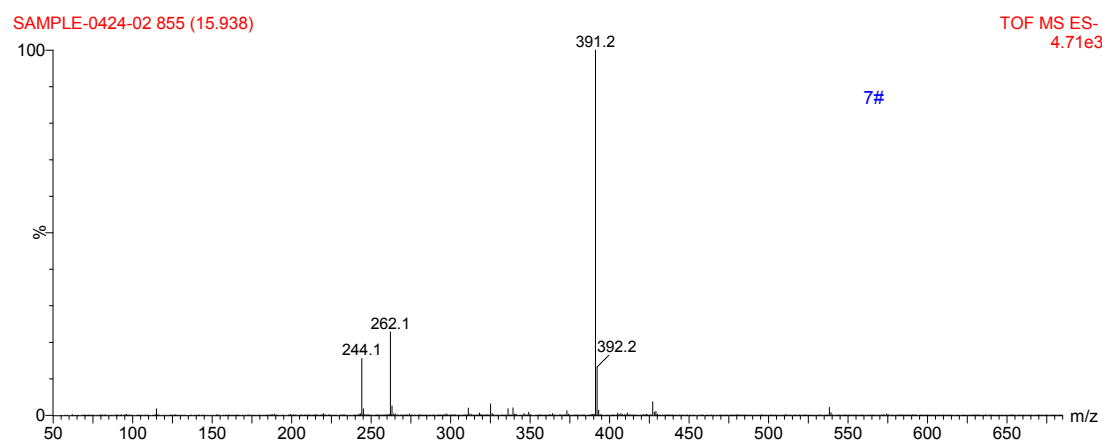
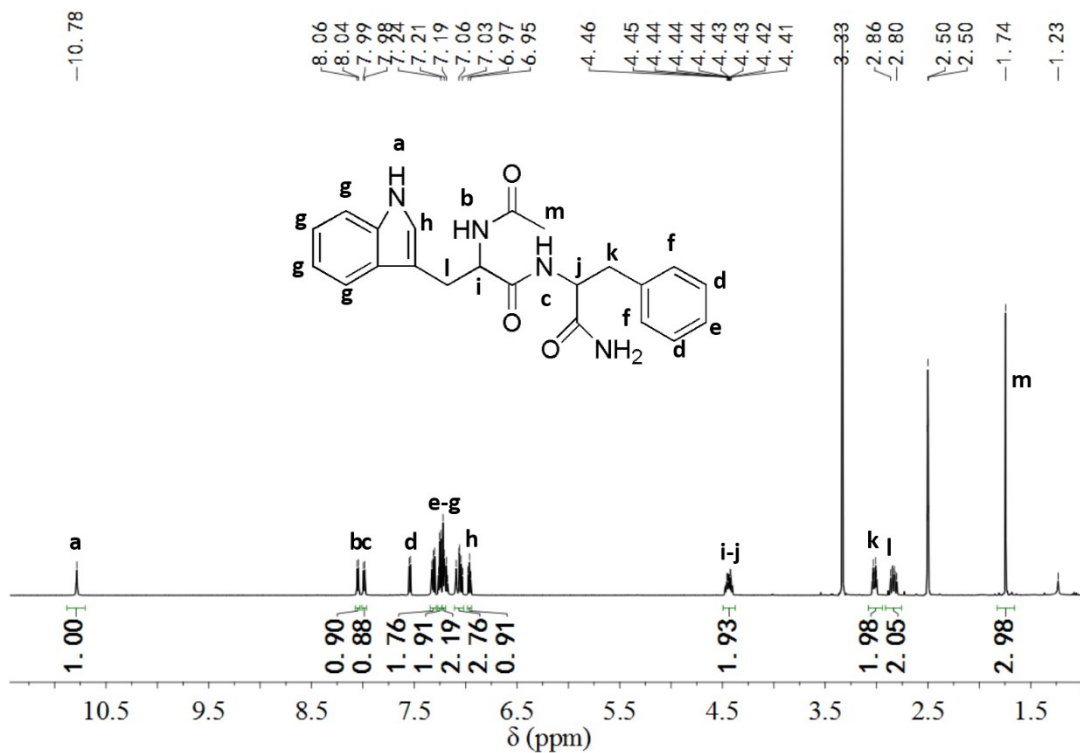
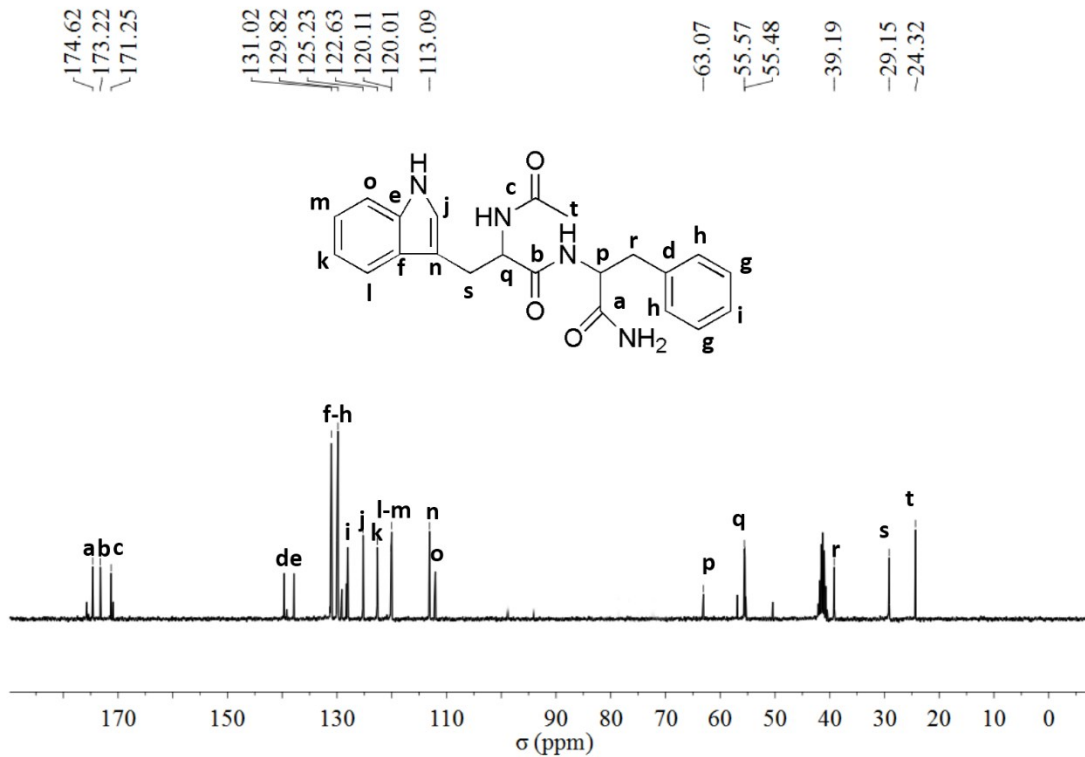


Figure S5. ESI-MS data of Ac-Trp-Phe-NH<sub>2</sub>.



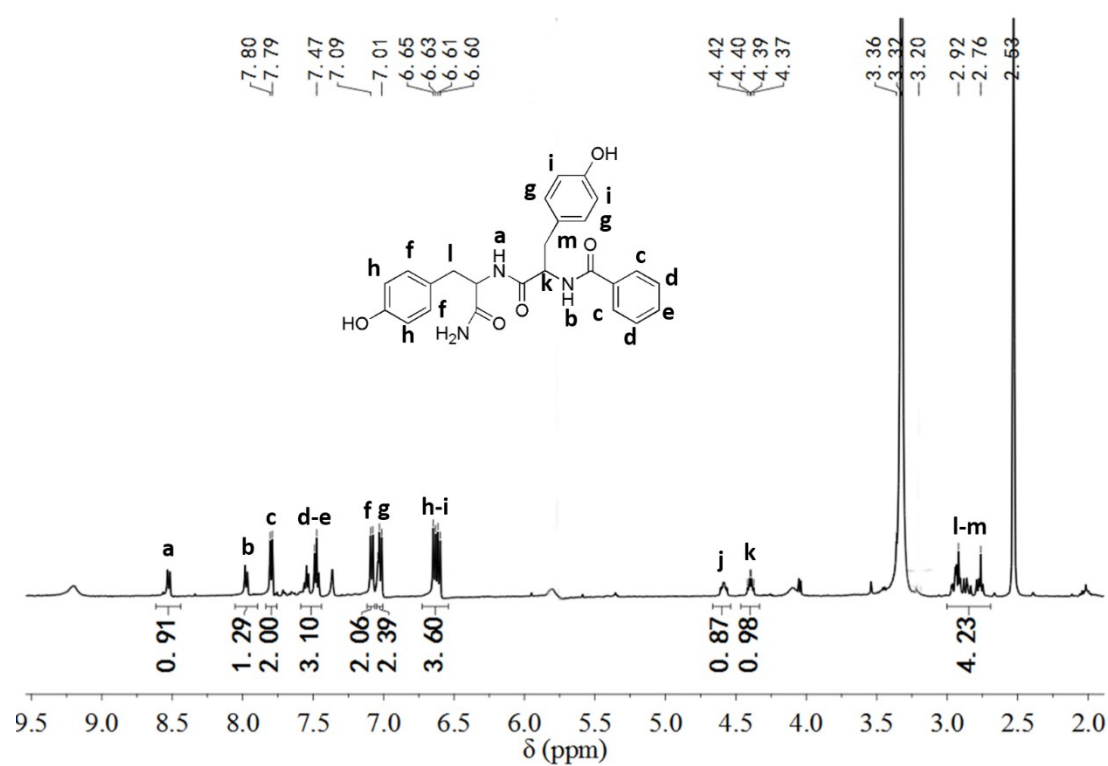
**Figure S6.**  $^1\text{H-NMR}$  spectrum of Ac-Trp-Phe- $\text{NH}_2$ .



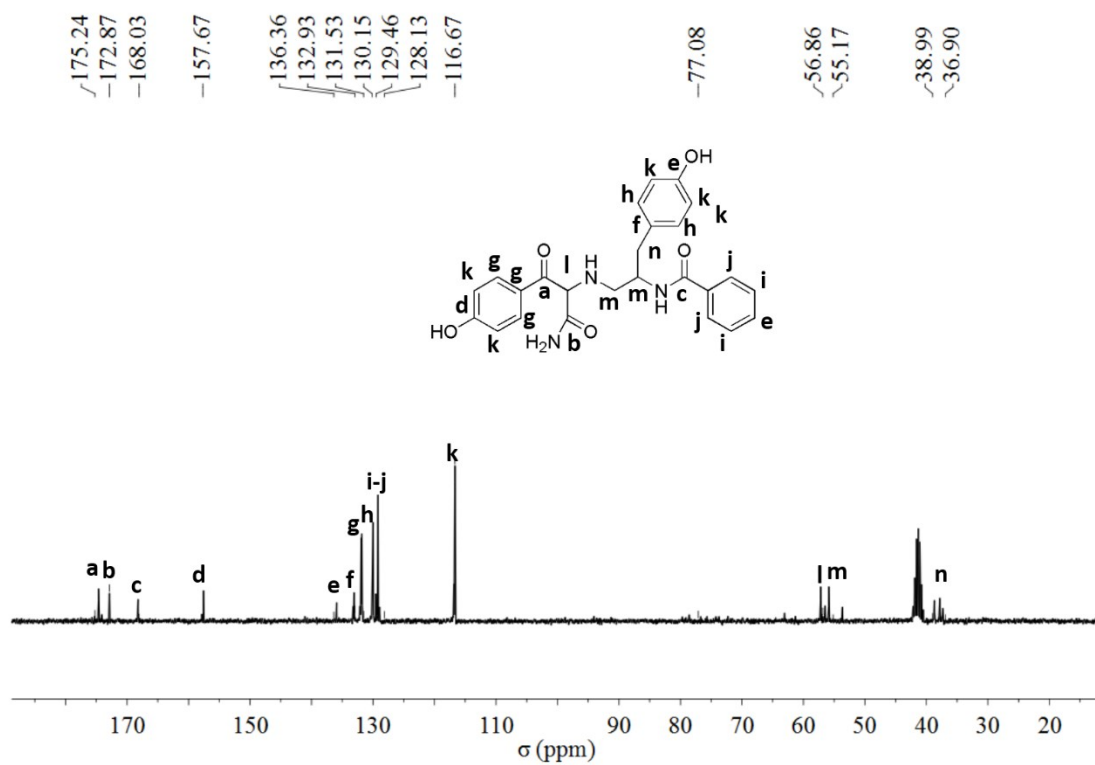
**Figure S7.**  $^{13}\text{C-NMR}$  spectrum of Ac-Trp-Phe- $\text{NH}_2$ .



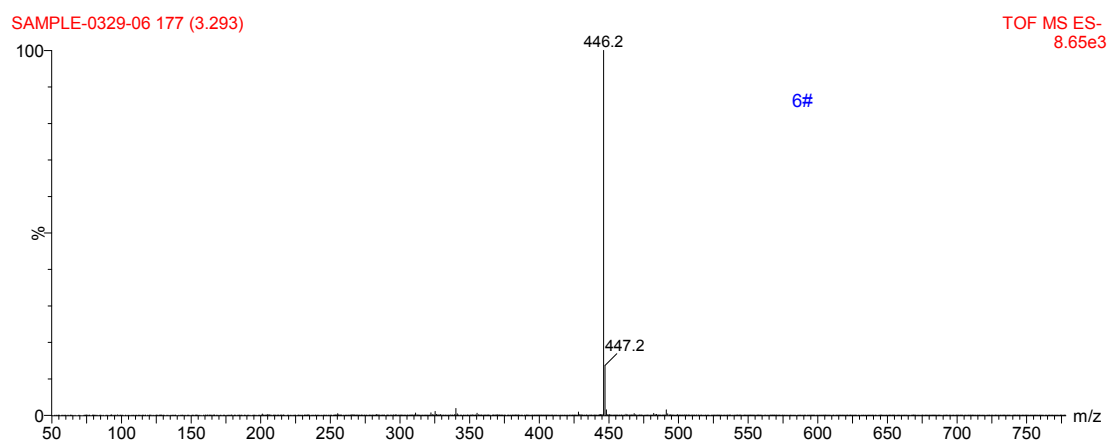
Bz-Tyr-Tyr-NH<sub>2</sub> (311.40 mg, yield: 87%, m.p. = 250-252 °C) as a white solid, purified by flash column chromatography using ethyl acetate as the eluent. <sup>1</sup>H-NMR (500 MHz, DMSO) δ<sub>H</sub> 8.54 (1 H, t, *J* 11.8), 8.05 – 7.89 (1 H, m), 7.80 (2 H, d, *J* 7.5), 7.48 (3 H, d, *J* 7.6), 7.08 (2 H, d, *J* 8.3), 7.05 – 7.00 (2 H, m), 6.62 (4 H, dd, *J* 16.6, 8.3), 4.66 – 4.54 (1 H, m), 4.40 (1 H, dd, *J* 13.4, 8.3), 2.84 (4 H, d, *J* 79.1). <sup>13</sup>C NMR (75 MHz, DMSO): δ 174.64, 172.87, 168.20, 157.58, 157.47, 135.92, 133.24, 133.08, 129.98, 129.17, 116.67, 57.20, 55.87, 37.83, 37.34. ESI-MS *m/z*: 446.2 [*M* - H]<sup>-</sup>, calculated for Bz-Tyr-Tyr-NH<sub>2</sub>, 447.2.



**Figure S8.** <sup>1</sup>H-NMR spectrum of Bz-Tyr-Tyr-NH<sub>2</sub> in DMSO.

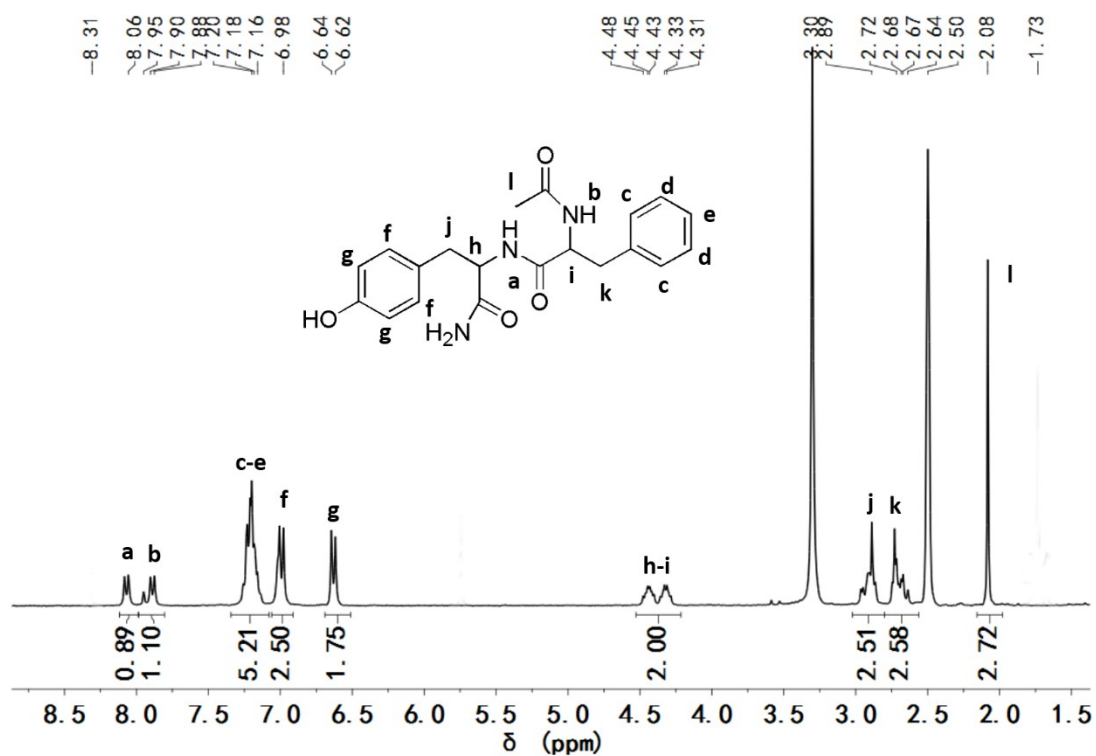


**Figure S9.**  $^{13}\text{C}$ -NMR spectrum of Bz-Tyr-Tyr-NH<sub>2</sub> in DMSO.

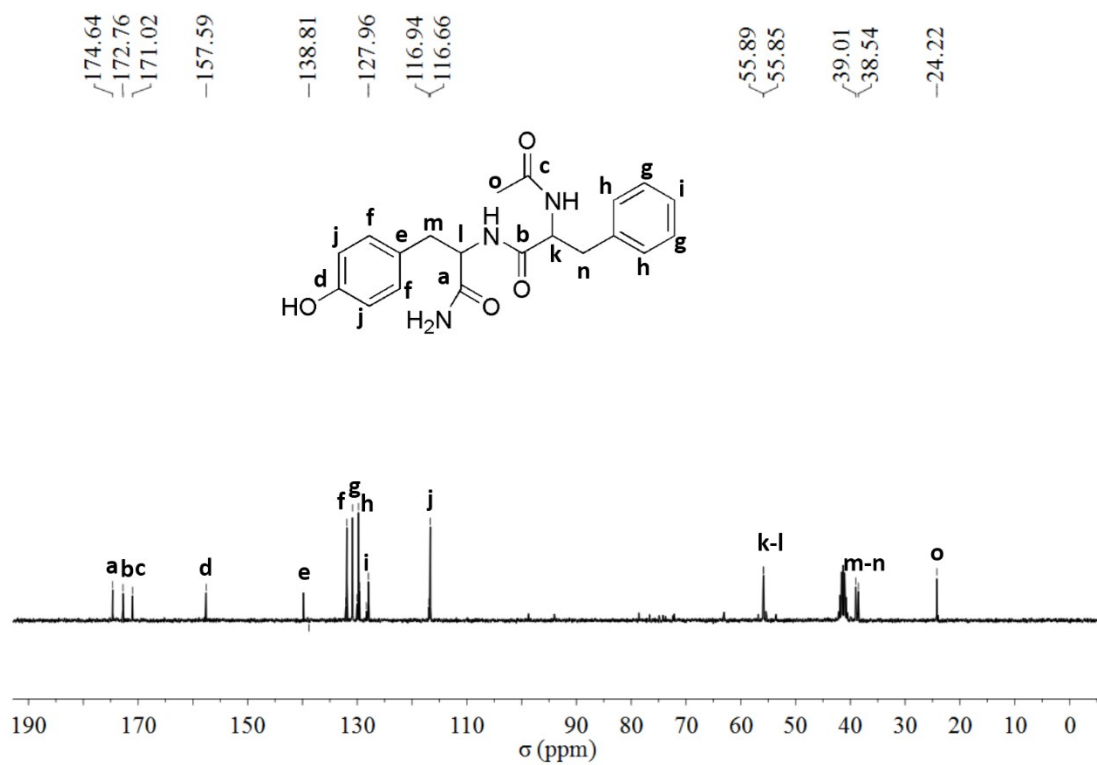


**Figure S10.** ESI-MS data of Bz-Tyr-Tyr-NH<sub>2</sub>.

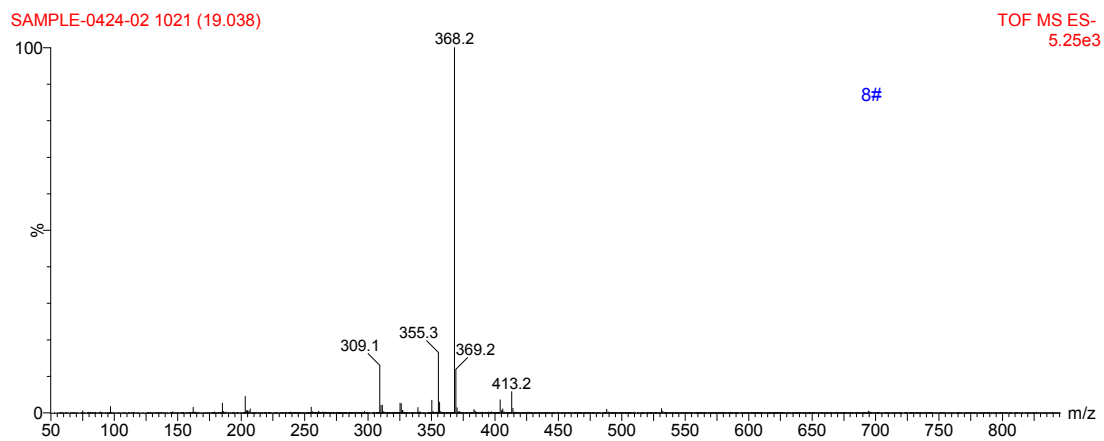
Ac-Phe-Tyr-NH<sub>2</sub> (251.20 mg, yield: 85%, m.p. =249-252 °C) as a white solid, purified by flash column chromatography using ethyl acetate/petroleum ether (v/v= 1/2) as the eluent. <sup>1</sup>H-NMR (300 MHz, DMSO) δ<sub>H</sub> 8.07 (1 H, d, *J* 8.2), 7.99 – 7.80 (1 H, m), 7.34 – 7.08 (5 H, m), 6.99 (3 H, d, *J* 8.4), 6.63 (2 H, d, *J* 8.3), 5.74 (0 H, s), 4.53 – 4.21 (2 H, m), 2.91 (3 H, dt, *J* 11.6, 5.8), 2.80 – 2.56 (3 H, m), 2.08 (3 H, s). <sup>13</sup>C NMR (75 MHz, DMSO): δ 174.64, 172.76, 171.02, 157.59, 139.82, 131.90, 130.88, 129.78, 127.96, 116.66, 55.89, 55.85, 38.54, 24.22. ESI-MS *m/z*: 368.2 [M - H]<sup>-</sup>, calculated for Ac-Phe-Tyr-NH<sub>2</sub>, 369.2.



**Figure S11.** <sup>1</sup>H-NMR spectrum of Ac-Phe-Tyr-NH<sub>2</sub> in DMSO.

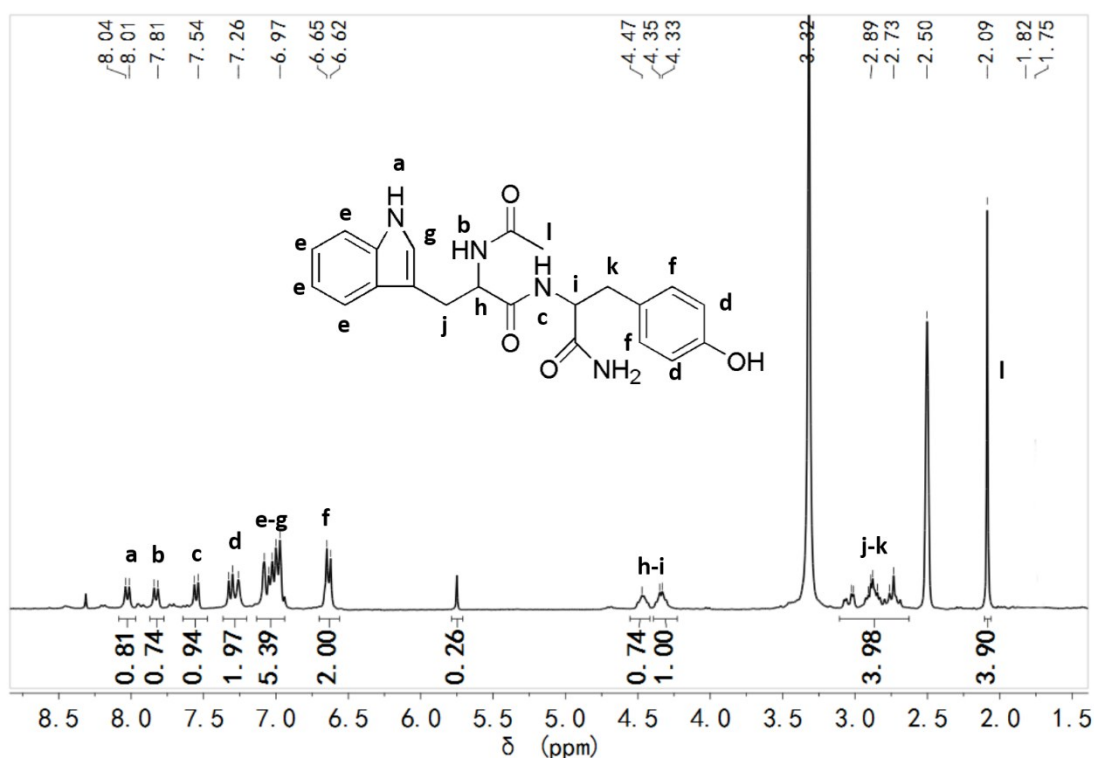


**Figure S12.** <sup>13</sup>C-NMR spectrum of Ac-Phe-Tyr-NH<sub>2</sub> in DMSO.

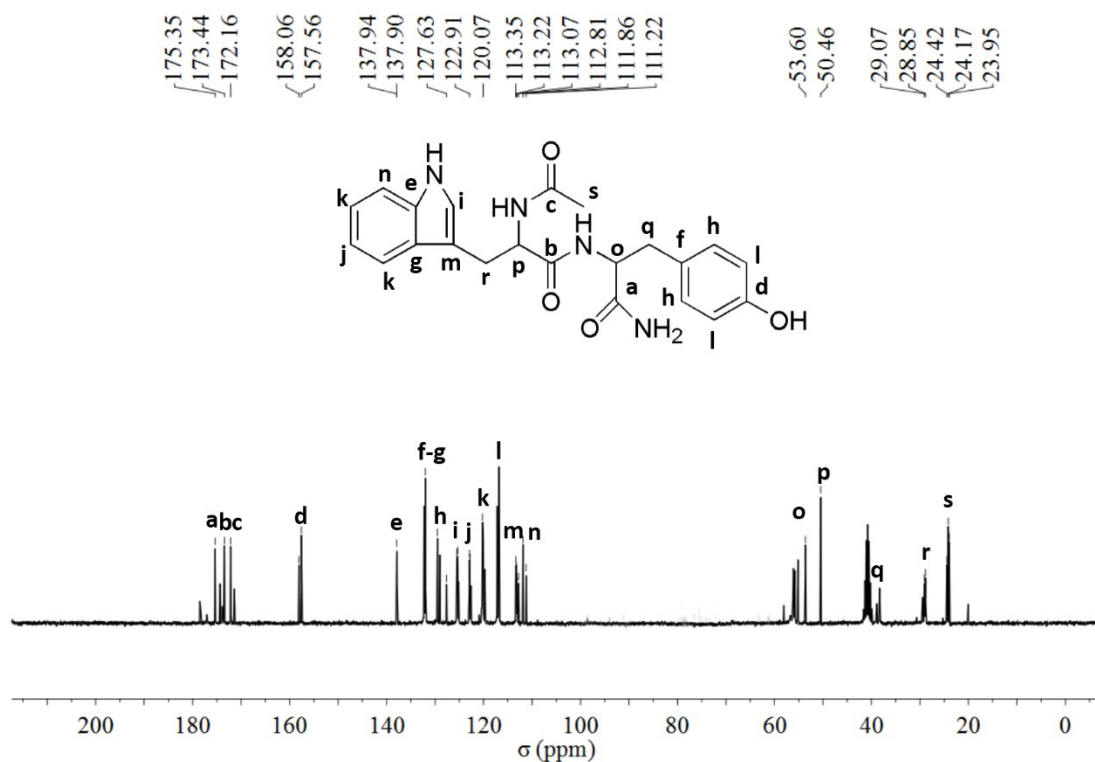


**Figure S13.** ESI-MS data of Ac-Phe-Tyr-NH<sub>2</sub>.

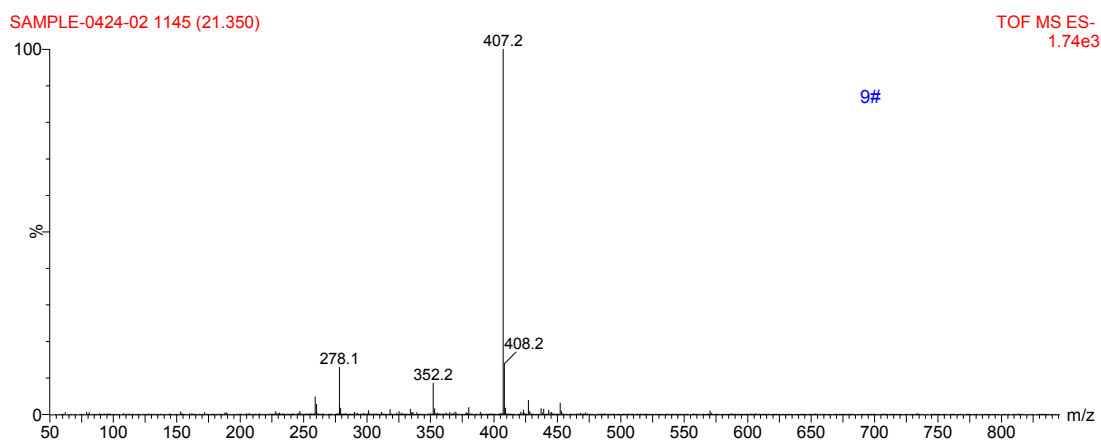
Ac-Trp-Tyr-NH<sub>2</sub> (267.90 mg, yield: 82%) as a yellow oil, purified by flash column chromatography using ethyl acetate/petroleum ether (v/v= 1/1) as the eluent. <sup>1</sup>H-NMR (300 MHz, DMSO) δ<sub>H</sub> 8.03 (1 H, d, *J* = 7.8 Hz), 7.83 (1 H, d, *J* = 8.1 Hz), 7.55 (1 H, d, *J* = 7.7 Hz), 7.36 – 7.20 (2 H, m), 7.13 – 6.94 (5 H, m), 6.64 (2 H, d, *J* = 8.2 Hz), 4.47 (1 H, s), 4.34 (1 H, d, *J* = 5.1 Hz), 2.88 (4 H, m), 2.09 (2 H, s). <sup>13</sup>C NMR (75 MHz, DMSO): δ 175.35, 173.44, 172.16, 157.56, 137.90, 132.00, 129.52, 129.50, 129.02, 128.85, 125.48, 125.34, 122.91, 122.81, 117.17, 116.83, 53.60, 50.46, 29.07, 28.85, 24.42, 24.17, 23.95.ESI-MS *m/z*: 407.2 [*M* - H]<sup>-</sup>, calculated for Ac-Trp-Tyr-NH<sub>2</sub>, 408.2.



**Figure S14.** <sup>1</sup>H-NMR spectrum of Ac-Trp-Tyr-NH<sub>2</sub> in DMSO.



**Figure S15.** <sup>13</sup>C-NMR spectrum of Ac-Trp-Tyr-NH<sub>2</sub> in DMSO.



**Figure S16.** ESI-MS data of Ac-Trp-Tyr-NH<sub>2</sub>.

Ac-Phe-Phe-NH<sub>2</sub> (223.30 mg, yield: 79%) as a white solid, purified by flash column chromatography using ethyl acetate/petroleum ether (v/v= 1/2) as the eluent. <sup>1</sup>H-NMR (600 MHz, DMSO) δ<sub>H</sub> 8.12 (1 H, m), 8.08 (1 H, m), 7.24 (6 H, m), 7.21 – 7.15 (4 H, m), 4.47 – 4.37 (2 H, m), 2.97 (2 H, m), 2.75 (3 H, m), 1.72 (3 H, s). <sup>13</sup>C NMR (75 MHz, DMSO): δ 174.46, 172.84, 170.99, 139.80, 139.63, 131.00, 130.87, 129.82, 129.78, 128.02, 55.83, 55.55, 39.28, 39.02, 24.22. ESI-MS m/z: 352.2 [M - H]<sup>-</sup>, calculated for Ac-Phe-Phe-NH<sub>2</sub>, 353.2.

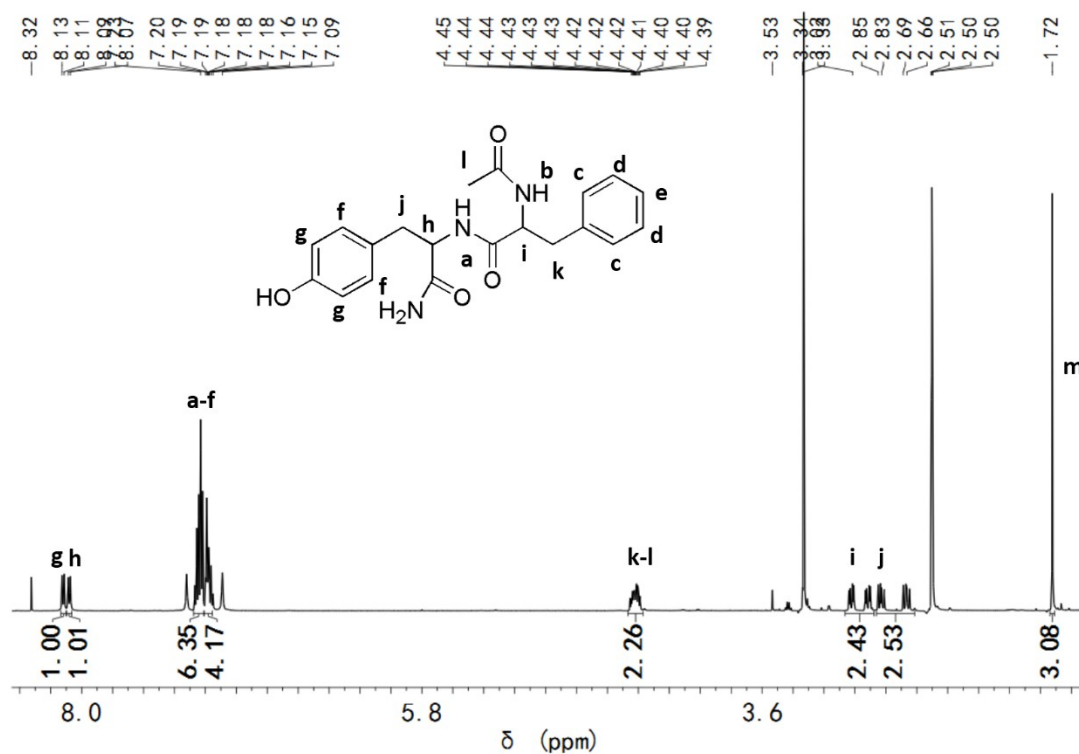
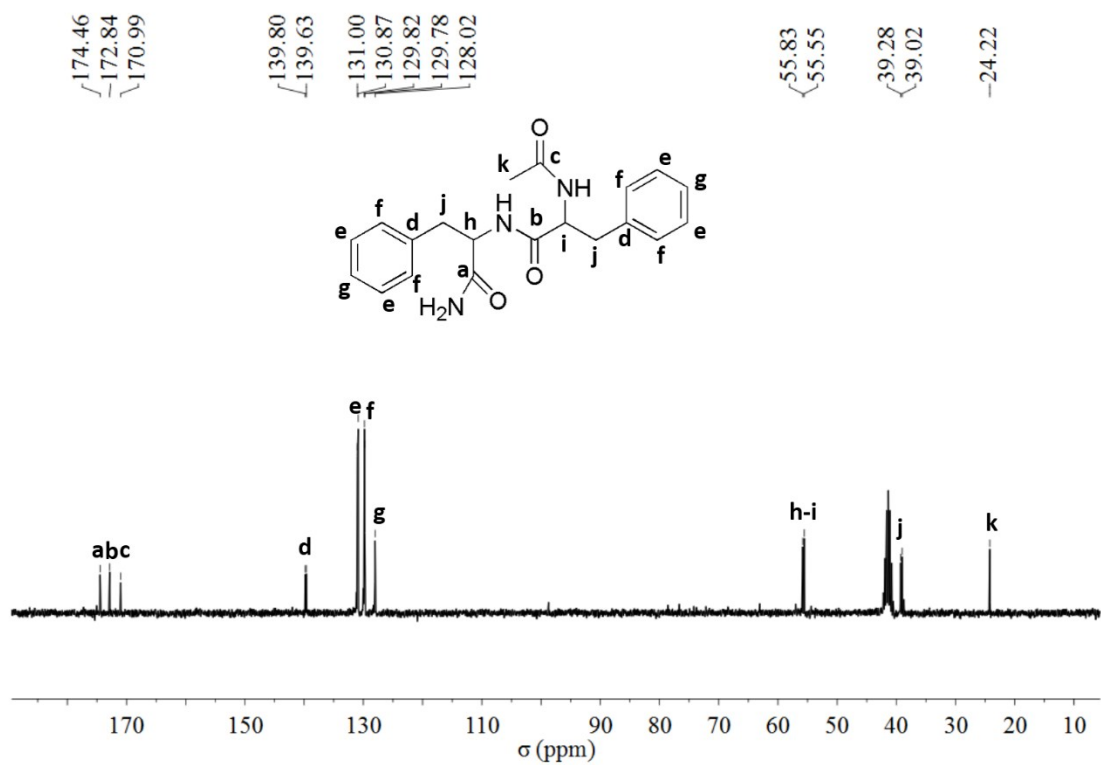
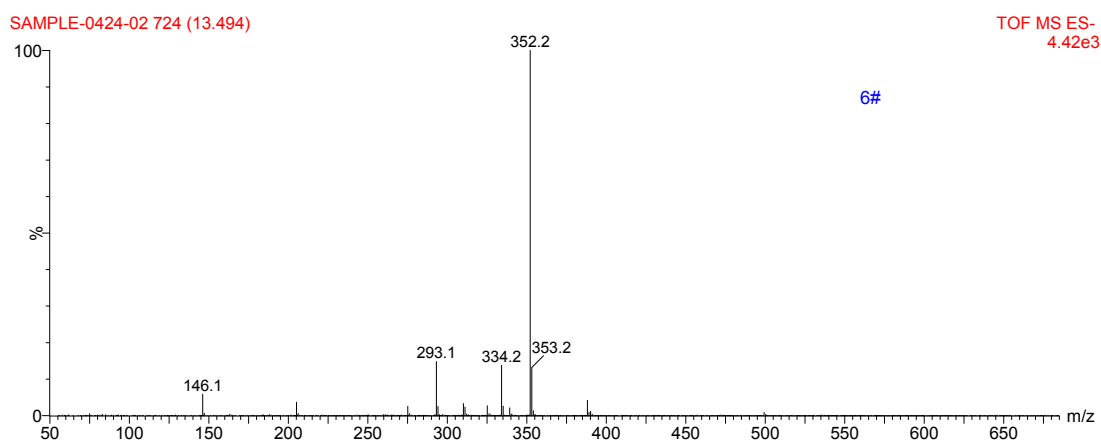


Figure S17. <sup>1</sup>H-NMR spectrum of Ac-Phe-Phe-NH<sub>2</sub> in DMSO.



**Figure S18.** <sup>13</sup>C-NMR spectrum of Ac-Phe-Phe-NH<sub>2</sub> in DMSO.



**Figure S19.** ESI-MS data of Ac-Phe-Phe-NH<sub>2</sub>.



## References

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