

Supporting Information for:

Carboallylation of Electron-Deficient Alkenes by Palladium/Copper Catalysis

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General. All manipulations of oxygen- and moisture-sensitive materials were conducted with a standard schlenk technique under an argon atmosphere or in a glove box under a nitrogen atmosphere. Medium pressure liquid chromatography (MPLC) was performed using Kanto Chemical silica gel 60 (spherical, 40–50 μm) or Biotage[®] SNAP Ultra. Analytical thin layer chromatography (TLC) was performed on Merck TLC silica gel 60 F₂₅₄ (0.25 mm) plates. Visualization was accomplished with UV light (254 nm) and/or an aqueous alkaline KMnO₄ solution followed by heating.

Apparatus. Proton, carbon, and fluorine nuclear magnetic resonance spectra (¹H, ¹³C, and ¹⁹F NMR) were recorded on a JEOL ECS-400 (¹H NMR, 400 MHz; ¹³C NMR 101 MHz; ¹⁹F NMR 376 MHz) spectrometer with solvent resonance as the internal standard (¹H NMR, CHCl₃ at 7.26 ppm, C₆H₆ at 7.16 ppm; ¹³C NMR, CDCl₃ at 77.0 ppm, C₆D₆ at 128.0 ppm; ¹⁹F NMR, C₆F₆ at –162.0 ppm). NMR data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, quint = quintet, sext = sextet, sept = septet, br = broad, m = multiplet), coupling constants (Hz), and integration. High-resolution mass spectra were obtained with Thermo Scientific Exactive (ESI or APCI) and JEOL JMX-SX102A (EI). Medium pressure liquid chromatography (MPLC) was performed with a Yamazen EPLC-W-Prep 2XY. Preparative recycling silica gel chromatography was performed on a Shimadzu Prominence chromatograph equipped with COSMOSIL 5SL-II (Nacalai Tesque, 20 mm x 250 mm, spherical, 5 μm). GC analysis was performed on a Shimadzu GC-2014 equipped with a BP1 column (SGE Analytical Science, 0.25 mm x 30 m, pressure = 149.0 kPa, detector = FID, 290 °C) with helium gas as a carrier. Chiral HPLC analyses were performed with a Shimadzu Prominence chromatograph. Optical rotations were measured on a HORIBA SEPA-200.

Chemicals. Unless otherwise noted, commercially available chemicals were used without purification. Alkenes (**1b**,^{1b} **1c**,^{1b} **1d**,^{1a} **1e**,^{1c} **1f**^{1d}), HOMS[®] reagents (**2a**,^{2a} **2b**,^{2b} **2c**,^{2b} **2d**,^{2c} **2e**,^{2a} **2f**,^{2d} **2g–2i**,^{2a} **2j**,^{2e} **2k**^{2e}), allylic carbonates (**3a**,^{3b} **3b**,^{3d} **3c**,^{3b} **3d**,^{3d} **3e**,^{3a} **3f**^{3c}), (IPr)CuCl₄,⁴ (SIPr)CuCl₄,⁴ (IMes)CuCl₄,⁴ and (SIMes)CuCl₄ were prepared according to the literature. Superdehydrated 1,4-dioxane was purchased from Wako Chemical and used as received.

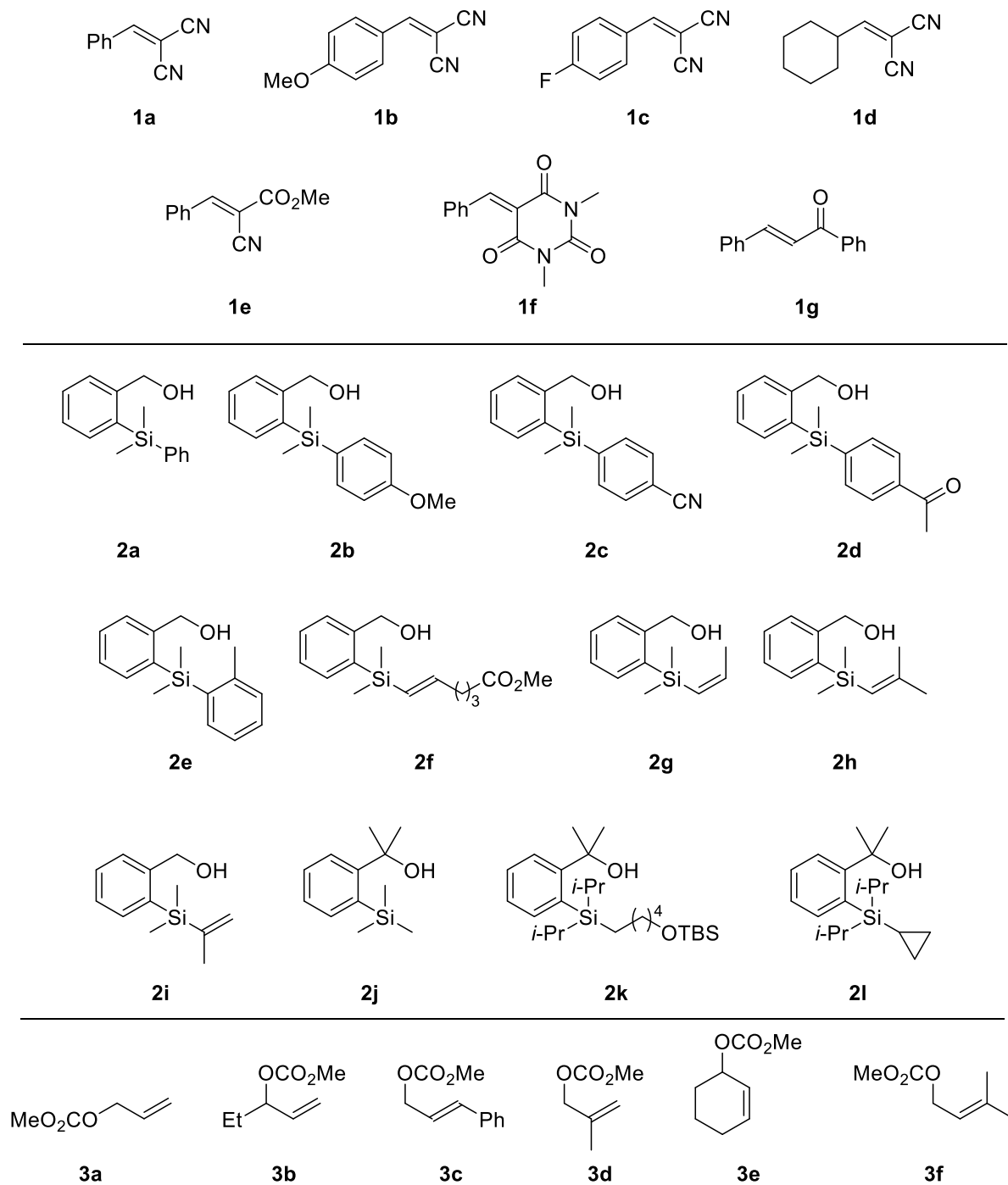
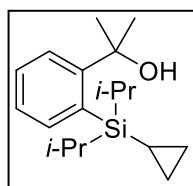
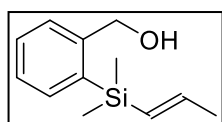


Figure S1. List of alkenes, HOMSi[®] reagents and allylic carbonates

Preparation of the substrate.

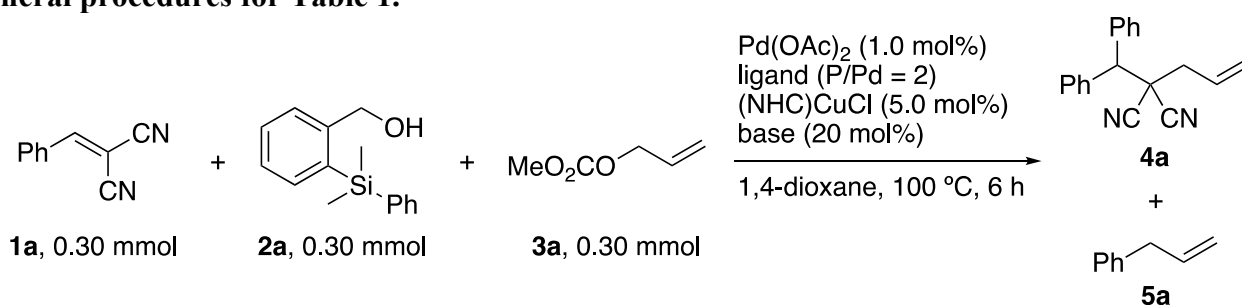


Preparation of 21. Following the reported procedure,^{2d} a 0.56 M solution of cyclopropyllithium in Et₂O (10 mL, 5.6 mmol) was added dropwise to a THF (5.0 mL) solution of 3,3-dimethyl-1,1-diisopropyl-1,3-dihydrobenzo[*c*][1,2]oxasilole (1.24 g, 5.0 mmol) at $-78\text{ }^{\circ}\text{C}$, and the resulting mixture was stirred at $-78\text{ }^{\circ}\text{C}$. After stirring for 1 h, the reaction mixture was quenched with a 2M HCl aqueous solution and saturated NaHCO₃ aqueous solution. The aqueous phase was extracted with Et₂O, and the combined organic phases were washed with brine, dried over anhydrous MgSO₄, and concentrated in *vacuo*. The residue was purified by flash column chromatography on silica gel (*n*-hexane:ethyl acetate = 30:1) to give the title compound (1.07 g, 74%) as a light yellow solid (mp = 46.4–49.7 °C), R_f 0.50 (*n*-hexane:ethyl acetate = 20:1). ¹H NMR (CDCl₃, 400 MHz): δ 8.07 (d, *J* = 7.3 Hz, 1H), 7.31 (t, *J* = 7.3 Hz, 1H), 7.24–7.17 (m, 2H), 2.21 (s, 1H), 1.61 (s, 6H), 1.34–1.21 (m, 2H), 1.02 (d, *J* = 7.3 Hz, 6H), 0.98 (d, *J* = 7.3 Hz, 6H), 0.82–0.75 (m, 2H), 0.51–0.44 (m, 2H); ¹³C NMR (CDCl₃, 101 MHz): δ 154.9, 137.9, 132.0, 128.3, 125.6, 125.1, 74.1, 33.7, 19.9, 19.3, 12.6, 3.4, –4.8; HRMS [APCI(–)] calcd for C₁₈H₂₉OSi [M–H][–]: 289.1993. Found: *m/z* 289.1994.



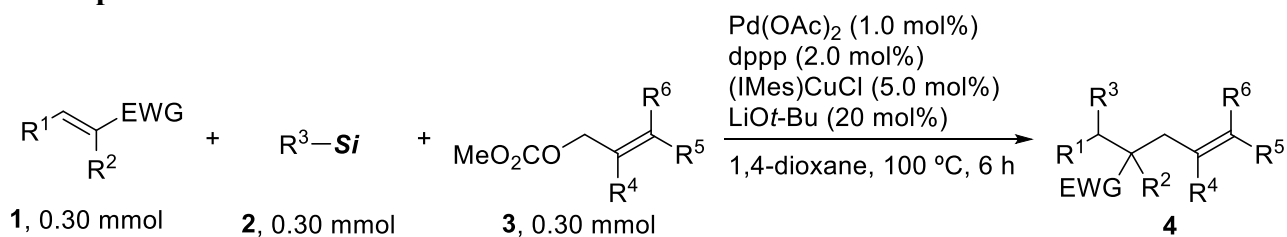
Preparation of (*E*)-2-(hydroxymethyl)phenyl]dimethyl(propen-1-yl)silane. Following the reported procedure,^{2a} a solution of (*E*)-propen-1-ylmagnesium bromide in THF (8.0 mL) [prepared from (*E*)-1-bromo-1-propene (0.69 g, 5.7 mmol) and Mg (0.17 g, 6.9 mmol)] was added to a THF (3.0 mL) solution of 1,1-dimethyl-1,3-dihydrobenzo[*c*][1,2]oxasilole (0.67 g, 4.1 mmol) at 0 °C, and the resulting mixture was stirred at room temperature for 12 h. The reaction mixture was quenched with a saturated NH₄Cl aqueous solution and the organic layer washed with a brine and dried over anhydrous MgSO₄. After removal of the solvent in *vacuo*, the residue was purified by flash chromatography on silica gel to give the title compound (0.20 g, 24%, *E*:*Z* = 81:19 as estimated by ¹H NMR analysis) as a colorless oil, R_f 0.26 (*n*-hexane:ethyl acetate = 7:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.55 (d, *J* = 6.9 Hz, 1H), 7.46 (d, *J* = 7.3 Hz, 1H), 7.40 (dt, *J* = 1.2 Hz, 7.5 Hz, 1H), 7.30 (d, *J* = 7.3 Hz, 1H), 6.17 (dq, *J* = 6.1 Hz, 19 Hz, 1H), 5.87 (dd, *J* = 1.6 Hz, 19 Hz, 1H), 4.74 (s, 2H), 1.86 (dd, *J* = 1.6 Hz, 6.2 Hz, 3H), 0.39 (s, 6H); ¹³C NMR (CDCl₃, 101 MHz): δ 146.4, 144.2, 137.0, 135.1, 130.2, 129.6, 128.0, 127.0, 65.4, 22.7, –1.3; Anal. Calcd for C₁₂H₁₈OSi; C, 69.84; H, 8.79. Found: C, 69.64; H, 8.85.

General procedures for Table 1.



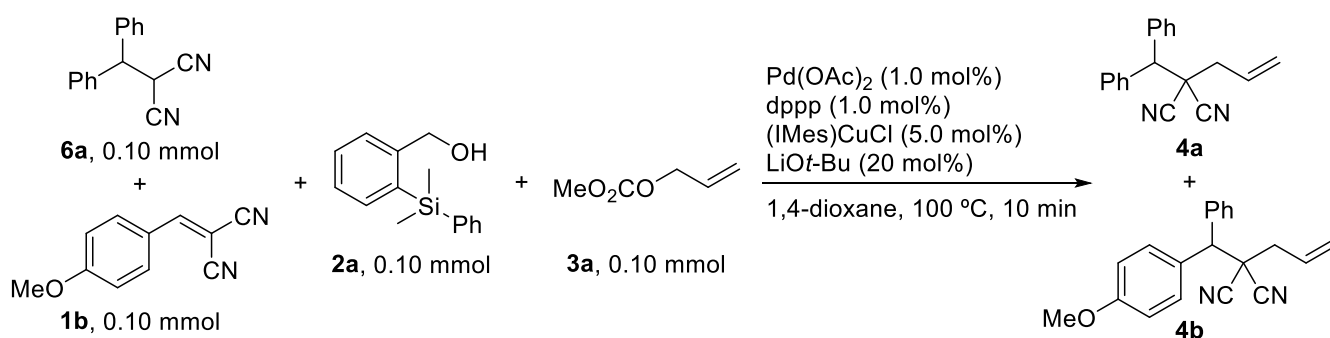
In the air, a vial (Vial A) was charged with (NHC)CuCl (15 μmol). A vial (Vial B) was charged with 1a (46 mg, 0.30 mmol) and 2a (73 mg, 0.30 mmol). A vial (Vial C) was charged with Pd(OAc)₂ (0.7 mg, 3.0 μmol) and phosphine ligand (P/Pd = 2). These vials were transferred into a glove box. In the glove box, base (60 μmol) and *n*-tridecane (50 μL) were added to Vial A. 1.95 mL of 1,4-dioxane was added into Vial B and 0.60 mL of 1,4-dioxane was added into Vial C. 3a (35 mg, 0.30 mmol) was added to Vial C. Then, the solution in Vial B and C were added to Vial A. The resulting mixture was stirred for 6 h at 100 °C. After the reaction, the yield of 4a and 5a were determined by GC with *n*-tridecane as an internal standard.

General procedures for Tables 2 and 3.



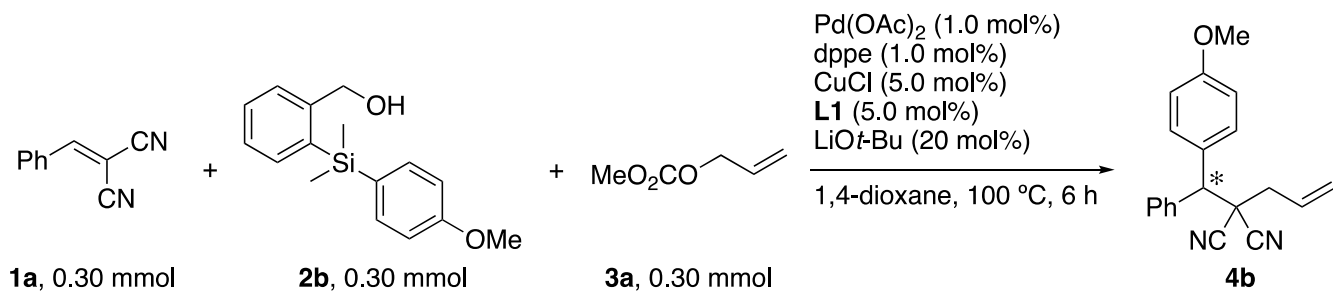
In the air, a vial (Vial **A**) was charged with (IMes)CuCl (20 mg, 50 μmol). A vial (Vial **B**) was charged with alkene (1.0 mmol) and HOMSi[®] reagent (1.0 mmol). A vial (Vial **C**) was charged with Pd(OAc)₂ (2.3 mg, 10 μmol) and dppp (4.1 mg, 10 μmol). These vials were transferred into a glove box. In the glove box, LiOt-Bu in 1.0 M THF solution (200 μL , 0.20 mmol) was added to Vial **A**. 6.5 mL of 1,4-dioxane was added into Vial **B** and 2.0 mL of 1,4-dioxane was added into Vial **C**. Allylic carbonate (1.0 mmol) was added to Vial **C**. Then, the solution in Vial **B** and **C** were added to Vial **A**. The resulting mixture was stirred for 6 h at 100 $^\circ\text{C}$. After the reaction, the mixture was filtered through a short pad of silica gel and Celite[®], and all of the volatiles were removed in *vacuo*. The product was purified by MPLC on silica gel to give the corresponding products.

Procedures for Eq. 4.



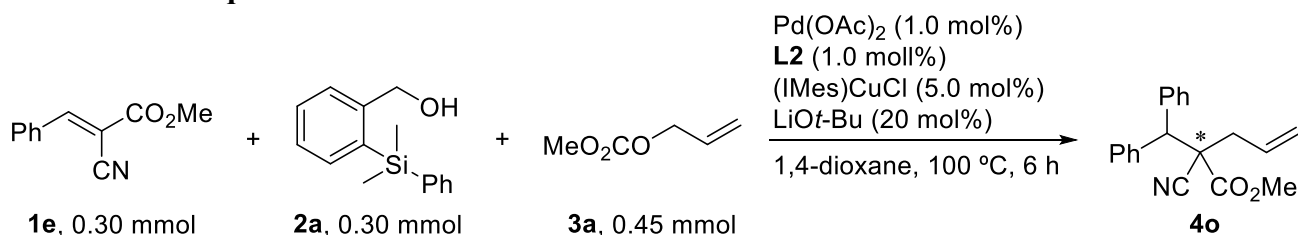
In the air, a vial (Vial **A**) was charged with (IMes)CuCl (2.0 mg, 5.0 μmol). A vial (Vial **B**) was charged with **1b** (18.4 mg, 0.10 mmol), **2a** (24 mg, 0.10 mmol) and 1,1-dicyano-2,2-diphenylethane (**6a**) (24 mg, 0.10 mmol). A vial (Vial **C**) was charged with Pd(OAc)₂ (0.2 mg, 1.0 μmol) and dppp (0.4 mg, 1.0 μmol). These vials were transferred into a glove box. In the glove box, LiOt-Bu in 1.0 M THF solution (20 μL , 20 μmol) and *n*-tridecane (30 μL) were added to Vial **A**. 0.65 mL of 1,4-dioxane was added into Vial **B** and 0.20 mL of 1,4-dioxane was added into Vial **C**. **3a** (11.6 mg, 0.10 mmol) was added to Vial **C**. Then, the solution in Vial **B** and **C** were added to Vial **A**. The resulting mixture was stirred for 10 min at 100 $^\circ\text{C}$. After the reaction, the yields of **4a** and **4b** were determined by GC with *n*-tridecane as an internal standard.

Procedures for Eq. 5.



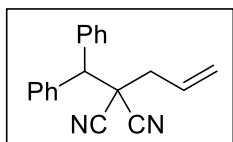
In the air, a vial (Vial **A**) was charged with **1a** (46 mg, 0.30 mmol) and **2b** (82 mg, 0.30 mmol). A vial (Vial **B**) was charged with Pd(OAc)₂ (0.7 mg, 3.0 μmol) and dppe (1.2 mg, 3.0 μmol). These vials were transferred into a glove box. In the glove box, CuCl (1.5 mg, 15 μmol), **L1** (10.5 mg, 15 μmol) in 1,4-dioxane (0.50 mL) and LiOt-Bu in 1.0 M THF solution (60 μL, 60 μmol) were added to a vial (Vial **C**). 1.45 mL of 1,4-dioxane was added into Vial **A** and 0.60 mL of 1,4-dioxane was added into Vial **B**. **3a** (35 mg, 0.30 mmol) was added to Vial **B**. Then, the solution in Vial **A** and **B** were added to Vial **C**. The resulting mixture was stirred for 6 h at 100 °C. After the reaction, the mixture was filtered through a short pad of silica gel and Celite[®], and all of the volatiles were removed in *vacuo*. The product was purified by MPLC on silica gel to give **4b** (17.2 mg, 57 μmol, 19%).

Procedures for Eq. 6.

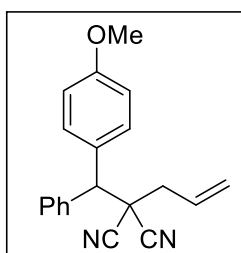


In the air, a vial (Vial **A**) was charged with (IMes)CuCl (6.1 mg, 15 μmol). A vial (Vial **B**) was charged with **1a** (46 mg, 0.30 mmol) and **2a** (73 mg, 0.30 mmol). A vial (Vial **C**) was charged with Pd(OAc)₂ (0.7 mg, 3.0 μmol). These vials were transferred into a glove box. In the glove box, LiOt-Bu in 1.0 M THF solution (60 μL, 60 μmol) was added to Vial **A**. **L2** (2.1 mg, 3.0 μmol) was added to the Vial **C**. 1.95 mL of 1,4-dioxane was added into Vial **B** and 0.60 mL of 1,4-dioxane was added into Vial **C**. **3a** (52 mg, 0.45 mmol) was added to Vial **C**. Then, the solution in Vial **B** and **C** were added to Vial **A**. The resulting mixture was stirred for 6 h at 100 °C. After the reaction, the mixture was filtered through a short pad of silica gel and Celite[®], and all of the volatiles were removed in *vacuo*. The product was purified by MPLC on silica gel to give **4n** (74 mg, 0.24 mmol, 81%).

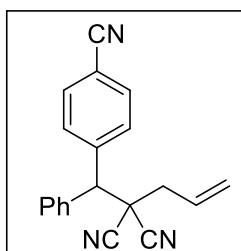
Characterization of the products



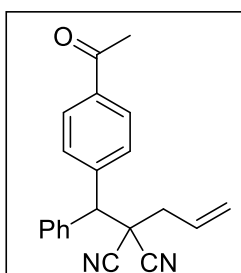
2,2-dicyano-1,1-diphenyl-4-pentene (4a). The reaction of **1a** (154 mg, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 96:4 to 84:16) gave the title compound (0.25 g, 0.92 mmol, 92%) as white solid (mp 88.9 °C), *R*_f 0.43 (*n*-hexane:ethyl acetate = 5:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.60 (d, *J* = 7.4 Hz, 4H), 7.47–7.33 (m, 6H), 6.05–5.92 (m, 1H), 5.45 (d, *J* = 10.0 Hz, 1H), 5.35 (dd, *J* = 1.3 Hz, 16.8 Hz, 1H), 4.34 (s, 1H), 2.70 (d, *J* = 7.4 Hz, 2H); ¹³C NMR (CDCl₃, 101 MHz): δ 136.6, 129.0, 128.8, 128.4, 123.3, 115.0, 56.4, 42.4, 41.4, a signal for sp²-carbon overlap with others; All the resonances of ¹H NMR spectrum were consist with reported values.⁵ HRMS [APCI(+)] calcd for C₁₉H₁₇N₂ [M+H]⁺: 273.1386. Found: *m/z* 273.1380.



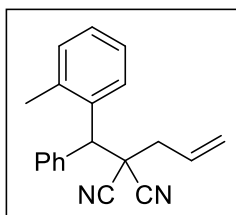
2,2-dicyano-1-phenyl-1-(4-methoxyphenyl)-4-pentene (4b). The reaction of **1a** (154 mg, 1.0 mmol), **2b** (0.27 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 94:6 to 76:24) gave the title compound (191 mg, 0.63 mmol, 63%) as light yellow solid (mp 67.4 °C), *R*_f 0.43 (*n*-hexane:ethyl acetate = 4:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.59 (d, *J* = 7.4 Hz, 2H), 7.51 (d, *J* = 8.7 Hz, 2H), 7.42 (t, *J* = 7.4 Hz, 2H), 7.38–7.28 (m, 1H), 6.94 (d, *J* = 8.7 Hz, 2H), 6.05–5.90 (m, 1H), 5.44 (d, *J* = 10.1 Hz, 1H), 5.34 (d, *J* = 16.8 Hz, 1H), 4.30 (s, 1H), 3.80 (s, 3H), 2.68 (d, *J* = 6.7 Hz, 2H); ¹³C NMR (CDCl₃, 101 MHz): δ 159.5, 137.0, 130.0, 129.0, 128.6, 128.53, 128.49, 128.3, 123.2, 115.1, 114.3, 55.7, 55.1, 42.7, 41.3; HRMS [APCI(+)] calcd for C₂₀H₁₉N₂O [M+H]⁺: 303.1492. Found: *m/z* 303.1481.



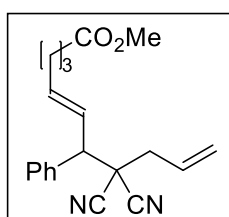
2,2-dicyano-1-phenyl-1-(4-cyanophenyl)-4-pentene (4c). The reaction of **1a** (77 mg, 0.50 mmol), **2c** (134 mg, 0.50 mmol) and **3a** (58 mg, 0.50 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 92:8 to 68:32) gave the title compound (59 mg, 0.20 mmol, 40%) as colorless oil, *R*_f 0.46 (*n*-hexane:ethyl acetate = 5:2). ¹H NMR (CDCl₃, 400 MHz): δ 7.73–7.64 (m, 4H), 7.53–7.46 (m, 2H), 7.46–7.35 (m, 3H), 6.00–5.86 (m, 1H), 5.46 (d, *J* = 10.1 Hz, 1H), 5.33 (d, *J* = 16.9 Hz, 1H), 4.34 (s, 1H), 2.68 (d, *J* = 7.3 Hz, 2H); ¹³C NMR (CDCl₃, 101 MHz): δ 141.9, 135.0, 132.8, 129.6, 129.5, 129.1, 128.8, 127.9, 123.9, 118.0, 114.6, 114.5, 112.6, 56.1, 41.9, 41.2; HRMS [APCI(+)] calcd for C₂₀H₁₆N₃ [M+H]⁺: 298.1339. Found: *m/z* 298.1337.



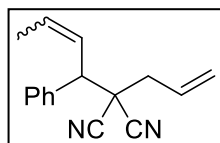
2,2-dicyano-1-phenyl-1-(4-acetylphenyl)-4-pentene (4d). The reaction of **1a** (77 mg, 0.50 mmol), **2d** (142 mg, 0.50 mmol) and **3a** (58 mg, 0.50 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 92:8 to 68:32) gave the title compound (57 mg, 0.18 mmol, 36%) as yellow oil, *R*_f 0.19 (*n*-hexane:ethyl acetate = 5:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.99 (d, *J* = 8.2 Hz, 2H), 7.67 (d, *J* = 8.7 Hz, 2H), 7.53 (d, *J* = 6.9 Hz, 2H), 7.45–7.33 (m, 3H), 6.01–5.88 (m, 1H), 5.45 (d, *J* = 10.1 Hz, 1H), 5.33 (dd, *J* = 1.1 Hz, 16.7 Hz, 1H), 4.36 (s, 1H), 2.68 (d, *J* = 7.3 Hz, 2H), 2.60 (s, 3H); ¹³C NMR (CDCl₃, 101 MHz): δ 197.2, 141.7, 137.0, 135.7, 129.3, 129.14, 129.07, 128.9, 128.2, 123.8, 114.8, 114.7, 56.3, 42.1, 41.4, 26.6; HRMS [APCI(+)] calcd for C₂₁H₁₈N₂O [M+H]⁺: 315.1492. Found: *m/z* 315.1498..



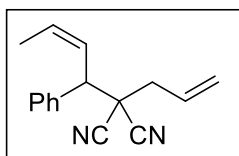
2,2-dicyano-1-phenyl-1-(2-methylphenyl)-4-pentene (4e). The reaction of **1a** (154 mg, 1.0 mmol), **2e** (0.26 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) using dppe (1,2-Bis(diphenylphosphino)ethane) as a ligand followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 100:0 to 94:6) gave the title compound (0.21 g, 0.72 mmol, 72%) as white solid (mp 83.0 °C), *R_f* 0.57 (*n*-hexane:ethyl acetate = 5:1). ¹H NMR (CDCl₃, 400 MHz): δ 8.00 (d, *J* = 7.8 Hz, 1H), 7.48–7.44 (m, 2H), 7.40–7.30 (m, 4H), 7.24 (dd, *J* = 0.9 Hz, 7.3 Hz, 1H), 7.18 (d, *J* = 7.3 Hz, 1H), 6.01–5.89 (m, 1H), 5.43 (d, *J* = 9.6 Hz, 1H), 5.29 (dd, *J* = 0.9 Hz, 16.9 Hz, 1H), 4.52 (s, 1H), 2.79 (dd, *J* = 7.6 Hz, 14.0 Hz, 1H), 2.73 (dd, *J* = 7.1 Hz, 14.0 Hz, 1H), 2.25 (s, 3H); ¹³C NMR (CDCl₃, 101 MHz): δ 136.6, 135.6, 131.7, 129.9, 129.1, 128.71, 128.66, 128.3, 126.9, 126.6, 123.6, 115.5, 115.3, 51.7, 42.7, 41.5, 20.1; HRMS [APCI(+)] calcd for C₂₀H₁₉N₂ [M+H]⁺: 287.1543. Found: *m/z* 287.1536.



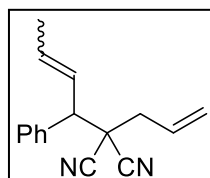
methyl (*E*)-8,8-dicyano-7-phenylundeca-5,10-dienoate (4f). The reaction of **1a** (77 mg, 0.50 mmol), **2f** (146 mg, 0.50 mmol) and **3a** (58 mg, 0.50 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 95:5 to 80:20) gave the title compound (77 mg, 0.24 mmol, 48%) as light yellow oil, *R_f* 0.33 (*n*-hexane:ethyl acetate = 5:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.44–7.33 (m, 5H), 5.99–5.84 (m, 2H), 5.79 (dt, *J* = 7.2 Hz, 15.5 Hz, 1H), 5.41 (d, *J* = 10.1 Hz, 1H), 5.34 (d, *J* = 16.8 Hz, 1H), 3.65 (s, 3H), 3.61 (d, *J* = 9.4 Hz, 1H), 2.61 (dd, *J* = 7.1, 13.8 Hz, 1H), 2.48 (dd, *J* = 7.1, 13.8 Hz, 1H), 2.32 (t, *J* = 7.4 Hz, 2H), 2.18 (q, *J* = 7.2 Hz, 2H), 1.75 (quintet, *J* = 7.2 Hz, 2H); ¹³C NMR (CDCl₃, 101 MHz): δ 173.6, 137.0, 136.1, 129.0, 128.7, 128.6, 128.4, 125.4, 123.0, 114.7, 114.3, 54.3, 51.4, 43.6, 40.2, 33.1, 31.7, 23.9; HRMS [APCI(+)] calcd for C₂₀H₂₃N₂O₂ [M+H]⁺: 323.1754. Found: *m/z* 323.1743.



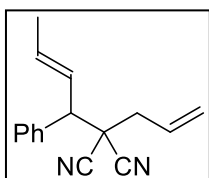
4,4-dicyano-5-phenyl-oct-1,6-diene (4g). The reaction of **1a** (77 mg, 0.50 mmol), **2g** (103 mg, 0.50 mmol) and **3a** (58 mg, 0.50 mmol) using dppe as a ligand followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 99:1 to 96:4) gave the title compound (79 mg, 0.34 mmol, 67%, *E:Z* = 4:96) as colorless oil, *R_f* 0.24 (*n*-hexane:ethyl acetate = 98:2). The ratio of *E/Z* isomers was determined by ¹H NMR analysis.



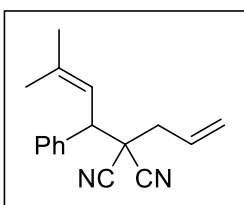
(*Z*)-4,4-dicyano-5-phenyl-oct-1,6-diene. Purification by HPLC (*n*-hexane:ethyl acetate = 10:1) gave the title compound as colorless oil, *R_f* 0.24 (*n*-hexane:ethyl acetate = 98:2). ¹H NMR (CDCl₃, 400 MHz): δ 7.48–7.32 (m, 5H), 6.05–5.84 (m, 3H), 5.42 (d, *J* = 10.1 Hz, 1H), 5.37 (dd, *J* = 1.4 Hz, 16.9 Hz, 1H), 4.01 (d, *J* = 10.1 Hz, 1H), 2.68 (dd, *J* = 6.9 Hz, 13.7 Hz, 1H), 2.53 (dd, *J* = 7.3 Hz, 14.2 Hz, 1H), 1.70 (dd, *J* = 1.4 Hz, 6.9 Hz, 3H); ¹³C NMR (CDCl₃, 101 MHz): δ 136.5, 130.6, 129.1, 128.8, 128.75, 128.66, 125.1, 123.1, 114.7, 114.5, 48.3, 43.8, 40.4, 13.5; HRMS [EI(+)] calcd for C₁₆H₁₆N₂ [M]⁺: 236.1313. Found: *m/z* 236.1306. The (*Z*)-configuration was determined by NOE experiments.



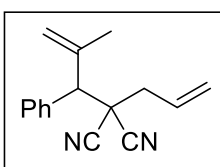
4,4-dicyano-5-phenyl-oct-1,6-diene. The reaction of **1a** (77 mg, 0.50 mmol), (*E*)-2-(hydroxymethyl)phenyl]dimethyl(propen-1-yl)silane (103 mg, 0.50 mmol) and **3a** (58 mg, 0.50 mmol) using dppe as a ligand followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 99:1 to 96:4) gave the title compound (46 mg, 0.19 mmol, 39%, *E:Z* = 66:34) as colorless oil, *R_f* 0.24 (*n*-hexane:ethyl acetate = 98:2). The ratio of *E/Z* isomers was determined by ¹H NMR analysis.



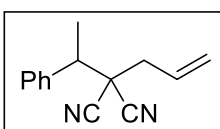
(E)-4,4-dicyano-5-phenyl-oct-1,6-diene. Purification by HPLC (*n*-hexane:ethyl acetate = 10:1) gave the title compound as colorless oil, R_f 0.24 (*n*-hexane:ethyl acetate = 98:2). ^1H NMR (CDCl_3 , 400 MHz): δ 7.44–7.32 (m, 5H), 5.98–5.87 (m, 2H), 5.82 (dq, $J = 6.4$ Hz, 15.1 Hz, 1H), 5.41 (d, $J = 10.1$ Hz, 1H), 5.35 (d, $J = 16.9$ Hz, 1H), 3.60 (d, $J = 9.2$ Hz, 1H), 2.62 (dd, $J = 7.1$ Hz, 14.0 Hz, 1H), 2.48 (dd, $J = 7.3$ Hz, 14.2 Hz, 1H), 1.79 (d, $J = 6.4$ Hz, 3H); ^{13}C NMR (CDCl_3 , 101 MHz): δ 136.5, 133.2, 129.1, 128.8, 128.7, 128.5, 125.7, 123.0, 114.9, 114.5, 54.6, 43.7, 40.5, 18.1; HRMS [APCI(+)] calcd for $\text{C}_{16}\text{H}_{17}\text{N}_2$ $[\text{M}+\text{H}]^+$: 237.1386. Found: m/z 237.1390. The (*E*)-configuration was determined by a coupling constant of a vinylic proton and NOE experiments.



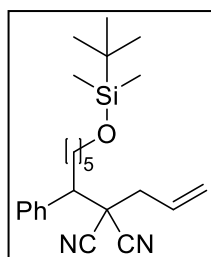
5,5-dicyano-2-methyl-4-phenyl-oct-2,7-diene (4h). The reaction of **1a** (154 mg, 1.0 mmol), **2h** (0.22 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 98:2 to 92:8) gave the title compound (0.20 g, 0.81 mmol, 81%) as white solid (mp 81.5 °C), R_f 0.65 (*n*-hexane:ethyl acetate = 4:1). ^1H NMR (CDCl_3 , 400 MHz): δ 7.44–7.32 (m, 5H), 5.98–5.86 (m, 1H), 5.72 (d, $J = 10.1$ Hz, 1H), 5.41 (d, $J = 10.1$ Hz, 1H), 5.35 (d, $J = 16.8$ Hz, 1H), 3.89 (d, $J = 10.1$ Hz, 1H), 2.65 (dd, $J = 7.1$ Hz, 13.8 Hz, 1H), 2.50 (dd, $J = 7.4$ Hz, 14.1 Hz, 1H), 1.84 (s, 3H), 1.70 (s, 3H); ^{13}C NMR (CDCl_3 , 101 MHz): δ 138.9, 136.9, 128.8, 128.4, 128.3, 122.7, 119.3, 114.7, 114.5, 49.4, 43.8, 40.2, 25.9, 18.4; HRMS [APCI(+)] calcd for $\text{C}_{17}\text{H}_{19}\text{N}_2$ $[\text{M}+\text{H}]^+$: 251.1543. Found: m/z 251.1537.



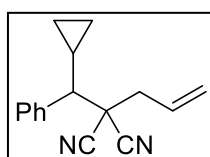
4,4-dicyano-2-methyl-3-phenyl-hept-1,6-diene (4i). The reaction of **1a** (154 mg, 1.0 mmol), **2i** (0.21 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 98:2 to 92:8) gave the title compound (159 mg, 0.67 mmol, 67%) as pale yellow oil, R_f 0.63 (*n*-hexane:ethyl acetate = 4:1). ^1H NMR (CDCl_3 , 400 MHz): δ 7.46–7.35 (m, 5H), 6.00–5.87 (m, 1H), 5.55 (s, 1H), 5.43 (d, $J = 10.1$ Hz, 1H), 5.35 (d, $J = 16.8$ Hz, 1H), 5.30 (s, 1H), 3.56 (s, 1H), 2.65 (dd, $J = 7.4$ Hz, 14.1 Hz, 1H), 2.58 (dd, $J = 7.1$ Hz, 13.8 Hz, 1H), 1.72 (s, 3H); ^{13}C NMR (CDCl_3 , 101 MHz): δ 140.9, 134.3, 129.2, 128.90, 128.86, 128.5, 123.2, 115.1, 114.8, 114.7, 56.8, 41.4, 40.5, 23.1; HRMS [APCI(+)] calcd for $\text{C}_{16}\text{H}_{17}\text{N}_2$ $[\text{M}+\text{H}]^+$: 237.1386. Found: m/z 237.1382.



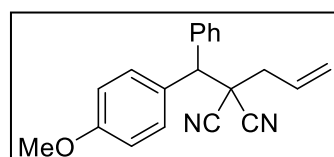
3,3-dicyano-2-phenyl-5-hexene (4j). The reaction of **1a** (154 mg, 1.0 mmol), **2j** (0.21 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 99:1 to 96:4) gave the title compound (116 mg, 0.55 mmol, 55%) as yellow oil, R_f 0.59 (*n*-hexane:ethyl acetate = 5:1). ^1H NMR (CDCl_3 , 400 MHz): δ 7.43–7.34 (m, 5H), 5.97–5.83 (m, 1H), 5.40 (dd, $J = 0.9$ Hz, 10.1 Hz, 1H), 5.34 (dd, $J = 0.9$ Hz, 16.9 Hz, 1H), 3.21 (q, $J = 7.0$ Hz, 1H), 2.54 (dd, $J = 7.3$ Hz, 13.7 Hz, 1H), 2.43 (dd, $J = 7.1$ Hz, 14.0 Hz, 1H), 1.71 (d, $J = 6.9$ Hz, 3H); ^{13}C NMR (CDCl_3 , 101 MHz): δ 137.5, 129.1, 129.00, 128.97, 128.5, 123.1, 115.3, 114.6, 45.8, 44.2, 40.4, 17.8; All the resonances of ^1H NMR spectrum were consistent with reported values.⁵ HRMS [APCI(+)] calcd for $\text{C}_{14}\text{H}_{15}\text{N}_2$ $[\text{M}+\text{H}]^+$: 211.1230. Found: m/z 211.1224.



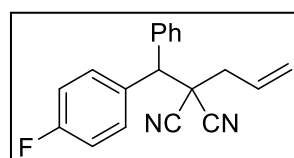
2-allyl-2-(6-((tert-butyldimethylsilyloxy)-1-phenylhexyl)malononitrile (4k). The reaction of **1a** (154 mg, 1.0 mmol), **2k** (0.45 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 97:3 to 88:12) gave the title compound (0.30 g, 0.76 mmol, 76%) as colorless oil, R_f 0.77 (*n*-hexane:ethyl acetate = 4:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.43–7.36 (m, 3H), 7.36–7.30 (m, 2H), 5.94–5.81 (m, 1H), 5.38 (d, J = 10.1 Hz, 1H), 5.30 (d, J = 16.8 Hz, 1H), 3.53 (t, J = 6.0 Hz, 2H), 2.98 (t, J = 7.7 Hz, 1H), 2.47 (dd, J = 7.4 Hz, 13.4 Hz, 1H), 2.35 (dd, J = 6.7 Hz, 14.1 Hz, 1H), 2.15 (d, J = 8.1 Hz, 1H), 2.11 (d, J = 8.1 Hz, 1H), 1.49–1.21 (m, 4H), 1.21–1.09 (m, 2H), 0.87 (s, 9H), 0.01 (s, 6H); ¹³C NMR (CDCl₃, 101 MHz): δ 135.4, 128.9, 128.8, 128.6, 122.7, 115.3, 114.4, 62.6, 51.1, 43.4, 40.2, 32.2, 31.5, 26.7, 25.8, 25.2, 18.1, –5.5; HRMS [APCI(+)] calcd for C₂₄H₃₇N₂OSi [M+H]⁺: 397.2670. Found: m/z 397.2663.



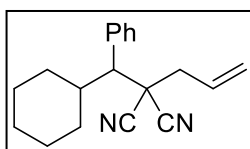
2,2-dicyano-1-cyclopropyl-1-phenyl-4-pentene (4l). The reaction of **1a** (154 mg, 1.0 mmol), **2l** (0.29 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 97:3 to 88:12) gave the title compound (162 mg, 0.69 mmol, 69%) as yellow oil, R_f 0.56 (*n*-hexane:ethyl acetate = 5:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.45–7.33 (m, 5H), 5.98–5.83 (m, 1H), 5.40 (d, J = 10.8 Hz, 1H), 5.33 (d, J = 16.8 Hz, 1H), 2.63 (dd, J = 7.4 Hz, 13.4 Hz, 1H), 2.50 (dd, J = 7.1 Hz, 13.8 Hz, 1H), 2.27 (d, J = 10.8 Hz, 1H), 1.60–1.48 (m, 1H), 1.08–0.97 (m, 1H), 0.77–0.67 (m, 1H), 0.65–0.54 (m, 1H), 0.16–0.06 (m, 1H); ¹³C NMR (CDCl₃, 101 MHz): δ 137.0, 128.8, 128.7, 128.6, 128.5, 122.8, 115.1, 114.6, 55.6, 43.8, 40.5, 13.4, 8.5, 3.3. HRMS [APCI(+)] calcd for C₁₆H₁₇N₂ [M+H]⁺: 237.1386. Found: m/z 237.1382.



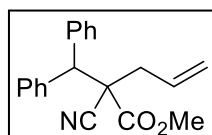
2,2-dicyano-1-phenyl-1-(4-methoxyphenyl)-4-pentene (4b). The reaction of **1b** (92 mg, 0.50 mmol), **2a** (121 mg, 0.50 mmol) and **3a** (58 mg, 0.50 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 94:6 to 76:24) gave the title compound (97 mg, 0.32 mmol, 64%) as white solid, R_f 0.43 (*n*-hexane:ethyl acetate = 4:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.54 (d, J = 7.8 Hz, 2H), 7.46 (d, J = 8.2 Hz, 2H), 7.39 (t, J = 7.6 Hz, 2H), 7.36–7.30 (m, 1H), 6.91 (d, J = 8.3 Hz, 2H), 6.01–5.88 (m, 1H), 5.43 (d, J = 10.1 Hz, 1H), 5.32 (d, J = 16.9 Hz, 1H), 4.24 (s, 1H), 3.80 (s, 3H), 2.66 (d, J = 6.9 Hz, 2H); ¹³C NMR (CDCl₃, 101 MHz): δ 159.6, 137.0, 130.1, 129.1, 128.72, 128.66, 128.6, 128.4, 123.4, 115.1, 114.5, 56.0, 55.3, 42.8, 41.5.; HRMS [APCI(+)] calcd for C₂₀H₁₉N₂O [M+H]⁺: 303.1492. Found: m/z 303.1480.



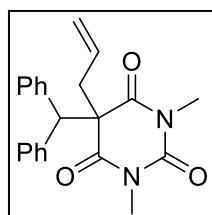
2,2-dicyano-1-phenyl-1-(4-fluorophenyl)-4-pentene (4m). The reaction of **1c** (172 mg, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 96:4 to 84:16) gave the title compound (0.21 g, 0.73 mmol, 73%) as pale yellow oil, R_f 0.38 (*n*-hexane:ethyl acetate = 5:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.56–7.49 (m, 4H), 7.44–7.33 (m, 3H), 7.12–7.05 (m, 2H), 6.00–5.88 (m, 1H), 5.44 (d, J = 10.1 Hz, 1H), 5.32 (d, J = 16.1 Hz, 1H), 4.28 (s, 1H), 2.66 (d, J = 7.4 Hz, 2H); ¹³C NMR (CDCl₃, 101 MHz): δ 162.4 (d, J = 249 Hz), 136.3, 132.5 (d, J = 3.8 Hz), 130.6 (d, J = 8.6 Hz), 129.1, 128.61, 128.55, 128.3, 123.4, 116.0 (d, J = 22 Hz), 114.9, 114.8, 55.5, 42.5, 41.2; ¹⁹F NMR (CDCl₃, 376 MHz): δ –113.2. HRMS [EI(+)] calcd for C₁₉H₁₅FN₂ [M]⁺: 290.1214. Found: m/z 290.1218.



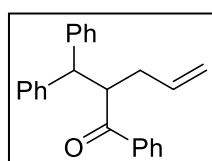
2,2-dicyano-1-cyclohexyl-1-phenyl-4-pentene (4n). The reaction of **1d** (160 mg, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 98:2 to 92:8) gave the title compound (145 mg, 0.52 mmol, 52%) as white solid (mp 60.1 °C), *R*_f 0.80 (*n*-hexane:ethyl acetate = 5:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.42–7.23 (m, 5H), 5.92–5.78 (m, 1H), 5.34 (d, *J* = 10.1 Hz, 1H), 5.22 (d, *J* = 16.8 Hz, 1H), 2.83 (d, *J* = 8.1 Hz, 1H), 2.35–2.07 (m, 4H), 1.90–1.81 (m, 1H), 1.70–1.56 (m, 3H), 1.44–1.31 (m, 1H), 1.31–1.03 (m, 3H), 0.97–0.85 (m, 1H); ¹³C NMR (CDCl₃, 101 MHz): δ 136.4, 129.0, 128.8, 128.4, 122.9, 116.3, 115.1, 57.3, 42.1, 42.0, 39.7, 32.4, 30.9, 26.2, 26.0, 25.8; All the resonances of ¹H NMR spectrum were consist with reported values.⁵ HRMS [APCI(+)] calcd for C₁₉H₂₃N₂ [M+H]⁺: 279.1856. Found: *m/z* 279.1850.



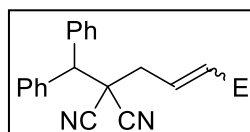
methyl 2-cyano-2-(diphenylmethyl)pent-4-enoate (4o). The reaction of **1e** (187 mg, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 96:4 to 84:16) gave the title compound (0.21 g, 0.70 mmol, 70%) as white solid (mp 86.5 °C), *R*_f 0.27 (*n*-hexane:ethyl acetate = 9:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.56 (d, *J* = 7.8 Hz, 2H), 7.51 (d, *J* = 7.8 Hz, 2H), 7.36 (t, *J* = 7.6 Hz, 2H), 7.33–7.20 (m, 4H), 5.84–5.69 (m, 1H), 5.21–5.12 (m, 2H), 4.41 (s, 1H), 3.59 (s, 3H), 2.65 (dd, *J* = 8.2 Hz, 13.7 Hz, 1H), 2.50 (dd, *J* = 6.2 Hz, 13.5 Hz, 1H); ¹³C NMR (CDCl₃, 101 MHz): δ 168.6, 139.0, 137.8, 130.4, 129.5, 128.8, 128.7, 128.3, 127.9, 127.6, 120.8, 118.6, 57.0, 54.7, 53.1, 42.1; HRMS [APCI(+)] calcd for C₂₀H₂₀NO₂ [M+H]⁺: 306.1489. Found: *m/z* 306.1488.



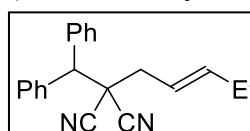
5-allyl-1,3-dimethyl-5-(diphenylmethyl)pyrimidine-2,4,6(1H,3H,5H)-trione (4p). The reaction of **1f** (0.24 g, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 96:4 to 84:16) gave the title compound (151 mg, 0.42 mmol, 42%) as white solid (mp 146.2 °C), *R*_f 0.52 (*n*-hexane:ethyl acetate = 5:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.41–7.36 (m, 4H), 7.32–7.22 (m, 6H), 5.51–5.39 (m, 1H), 5.05 (dd, *J* = 1.4 Hz, 16.9 Hz, 1H), 4.99 (dd, *J* = 1.6 Hz, 10.3 Hz, 1H), 4.56 (s, 1H), 3.12 (s, 6H), 2.71 (d, *J* = 7.3 Hz, 2H); ¹³C NMR (CDCl₃, 101 MHz): δ 170.7, 150.2, 137.9, 131.2, 129.4, 128.3, 127.7, 120.5, 61.7, 61.2, 41.0, 28.2; HRMS [APCI(+)] calcd for C₂₂H₂₃N₂O₃ [M+H]⁺: 363.1703. Found: *m/z* 363.1693.



1-phenyl-2-diphenylmethyl-4-pentene-1-one (4q). The reaction of **1g** (0.21 g, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3a** (116 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 99:1 to 96:4) and preparative recycling HPLC (*n*-hexane:ethyl acetate = 95:5) gave the title compound (21 mg, 0.064 mmol, 6%) as white solid (mp 97.1 °C), *R*_f 0.64 (*n*-hexane:ethyl acetate = 5:1). ¹H NMR (CDCl₃, 400 MHz): δ 7.86–7.76 (m, 2H), 7.53–7.45 (m, 1H), 7.43–7.28 (m, 6H), 7.25–7.17 (m, 3H), 7.11–7.02 (m, 2H), 7.01–6.93 (m, 1H), 5.68–5.54 (m, 1H), 4.91–4.82 (m, 2H), 4.54–4.45 (m, 1H), 4.41 (d, *J* = 11.5 Hz, 1H), 2.49–2.28 (m, 2H); ¹³C NMR (CDCl₃, 101 MHz): δ 203.1, 142.8, 142.5, 138.3, 134.6, 132.7, 128.7, 128.4, 128.3, 128.0, 127.8, 126.6, 126.2, 117.3, 53.7, 50.1, 36.4, a signal for sp²-carbon overlap with others; HRMS [APCI(+)] calcd for C₂₄H₂₃O [M+H]⁺: 327.1743. Found: *m/z* 327.1727.

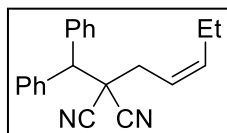


2,2-dicyano-1,1-diphenyl-4-heptene (4r). The reaction of **1a** (154 mg, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3b** (144 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 96:4 to 84:16) gave the title compound (0.21 g, 0.69 mmol, 69%, *E:Z* = 91:9) as colorless oil, *R*_f 0.48 (*n*-hexane:ethyl acetate = 5:1). The ratio of *E/Z* isomers was determined by ¹H NMR analysis.

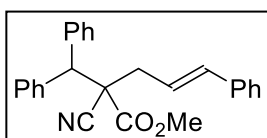


trans-2,2-dicyano-1,1-diphenyl-4-heptene. Purification by HPLC (*n*-hexane:ethyl acetate = 8:1) gave the title compound as colorless oil, *R*_f 0.48 (*n*-

hexane:ethyl acetate = 5:1). ^1H NMR (CDCl_3 , 400 MHz): δ 7.55 (d, $J = 7.8$ Hz, 4H), 7.44–7.30 (m, 6H), 5.75 (dt, $J = 6.0$ Hz, 15.6 Hz, 1H), 5.54 (dt, $J = 7.3$ Hz, 15.1 Hz, 1H), 4.27 (s, 1H), 2.61 (d, $J = 6.9$ Hz, 2H), 2.12 (quint, $J = 7.1$ Hz, 2H), 1.02 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (CDCl_3 , 101 MHz): δ 141.7, 136.8, 129.07, 128.88, 128.4, 118.8, 115.3, 56.5, 43.0, 40.6, 25.6, 13.3.; HRMS [APCI(+)] calcd for $\text{C}_{21}\text{H}_{21}\text{N}_2$ $[\text{M}+\text{H}]^+$: 301.1699. Found: m/z 301.1694.

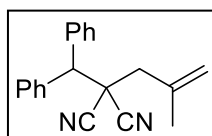


cis-2,2-dicyano-1,1-diphenyl-4-heptene. Purification by HPLC (*n*-hexane:ethyl acetate = 8:1) gave the title compound as colorless oil, R_f 0.48 (*n*-hexane:ethyl acetate = 5:1). ^1H NMR (CDCl_3 , 400 MHz): δ 7.55 (d, $J = 7.8$ Hz, 4H), 7.43–7.30 (m, 6H), 5.80 (dt, $J = 7.8$ Hz, 10.1 Hz, 1H), 5.53 (dt, $J = 8.0$ Hz, 9.2 Hz, 1H), 4.30 (s, 1H), 2.71 (d, $J = 7.3$ Hz, 2H), 1.92 (quint, $J = 7.4$ Hz, 2H), 0.95 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (CDCl_3 , 101 MHz): δ 139.7, 136.8, 129.10, 128.86, 128.5, 118.5, 115.3, 56.4, 42.6, 35.2, 21.0, 13.8.; HRMS [APCI(+)] calcd for $\text{C}_{21}\text{H}_{21}\text{N}_2$ $[\text{M}+\text{H}]^+$: 301.1699. Found: m/z 301.1693.



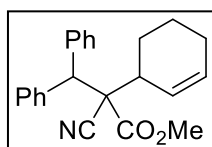
trans-2-cyano-2-methoxycarbonyl-1,1,5-triphenyl-4-pentene (4s). The reaction of **1e** (187 mg, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3c** (192 mg, 1.0 mmol) using dppe as a ligand followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 96:4 to 84:16) gave the title compound (0.26 g, 0.67 mmol, 67%) as white solid (mp 120.7 °C), R_f 0.45 (*n*-hexane:ethyl acetate =

5:1). ^1H NMR (CDCl_3 , 400 MHz): δ 7.60 (d, $J = 8.1$ Hz, 2H), 7.54 (d, $J = 7.4$ Hz, 2H), 7.37 (t, $J = 7.4$ Hz, 2H), 7.34–7.27 (m, 7H), 7.26–7.19 (m, 2H), 6.46 (d, $J = 16.1$ Hz, 1H), 6.19–6.08 (m, 1H), 4.49 (s, 1H), 3.53 (s, 3H), 2.81 (dd, $J = 8.7$ Hz, 13.4 Hz, 1H), 2.67 (dd, $J = 6.4$ Hz, 13.8 Hz, 1H); ^{13}C NMR (CDCl_3 , 101 MHz): δ 168.6, 138.9, 137.7, 136.3, 135.5, 129.5, 128.8, 128.6, 128.4, 128.3, 127.9, 127.8, 127.5, 126.4, 121.3, 118.7, 56.9, 54.8, 53.2, 41.4; HRMS [APCI(+)] calcd for $\text{C}_{26}\text{H}_{24}\text{NO}_2$ $[\text{M}+\text{H}]^+$: 382.1802. Found: m/z 382.1792.



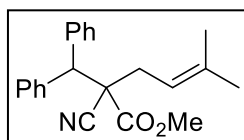
2,2-dicyano-4-methyl-1,1-diphenyl-4-pentene (4t). The reaction of **1a** (154 mg, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3d** (130 mg, 1.0 mmol) followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 96:4 to 84:16) gave the title compound (0.20 g, 0.71 mmol, 71%) as white solid (119.7 °C), R_f 0.52 (*n*-

hexane:ethyl acetate = 5:1). ^1H NMR (CDCl_3 , 400 MHz): δ 7.57 (d, $J = 7.4$ Hz, 4H), 7.43–7.32 (m, 6H), 5.14 (s, 1H), 5.03 (s, 1H), 4.29 (s, 1H), 2.61 (s, 2H), 1.95 (s, 3H); ^{13}C NMR (CDCl_3 , 101 MHz): δ 137.2, 136.7, 129.0, 128.8, 128.4, 118.6, 115.4, 58.0, 45.0, 41.8, 23.1; HRMS [APCI(+)] calcd for $\text{C}_{20}\text{H}_{19}\text{N}_2$ $[\text{M}+\text{H}]^+$: 287.1543. Found: m/z 287.1536.



methyl 2-cyano-2-(cyclohex-2-en-1-yl)-3,3-diphenylpropanoate (4u). The reaction of **1e** (187 mg, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3e** (156 mg, 1.0 mmol) using dppe as a ligand for 20 h at 130 °C followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 96:4 to 84:16) gave the title compound (130 mg, 0.38 mmol, 38%) as pale yellow oil, R_f 0.60 (*n*-hexane:ethyl acetate = 5:1). ^1H NMR

(CDCl_3 , 400 MHz): δ 7.64 (d, $J = 7.8$ Hz, 2H), 7.50 (d, $J = 7.8$ Hz, 2H), 7.35 (t, $J = 7.56$ Hz, 2H), 7.31–7.18 (m, 4H), 5.85–5.78 (m, 1H), 5.71–5.65 (m, 1H), 4.68 (s, 1H), 3.58 (s, 3H), 2.86–2.77 (m, 1H), 1.98–1.90 (m, 2H), 1.88–1.79 (m, 1H), 1.79–1.69 (m, 2H), 1.38–1.24 (m, 1H); ^{13}C NMR (CDCl_3 , 101 MHz): δ 167.4, 139.3, 138.2, 131.2, 129.0, 128.81, 128.77, 128.5, 127.6, 127.3, 125.1, 119.6, 57.6, 53.2, 52.8, 42.0, 24.6, 23.2, 21.6; HRMS [APCI(+)] calcd for $\text{C}_{23}\text{H}_{24}\text{NO}_2$ $[\text{M}+\text{H}]^+$: 346.1802. Found: m/z 346.1794.



2-cyano-2-methoxycarbonyl-5-methyl-1,1-diphenyl-4-hexene (4v). The reaction of **1e** (187 mg, 1.0 mmol), **2a** (0.24 g, 1.0 mmol) and **3f** (144 mg, 1.0 mmol) using dppe as a ligand for 20 h at 130 °C followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 96:4 to 84:16) gave the title compound (179 mg, 0.54 mmol, 54%) as colorless oil, *R_f* 0.56 (*n*-hexane:ethyl acetate = 5:1).

¹H NMR (CDCl₃, 400 MHz): δ 7.58 (d, *J* = 7.8 Hz, 2H), 7.50 (d, *J* = 8.2 Hz, 2H), 7.36 (t, *J* = 7.79 Hz, 2H), 7.33–7.19 (m, 4H), 5.12 (t, *J* = 7.1 Hz, 1H), 4.42 (s, 1H), 3.57 (s, 3H), 2.67 (dd, *J* = 8.2 Hz, 14.2 Hz, 1H), 2.46 (dd, *J* = 6.6 Hz, 14.0 Hz, 1H), 1.70 (s, 3H), 1.54 (s, 3H); ¹³C NMR (CDCl₃, 101 MHz): δ 168.9, 139.1, 137.93, 137.87, 129.3, 128.6, 128.5, 128.2, 127.7, 127.4, 119.0, 115.9, 56.8, 54.5, 53.0, 36.5, 25.8, 17.8; HRMS [APCI(+)] calcd for C₂₂H₂₄NO₂ [M+H]⁺: 334.1802. Found: *m/z* 334.1792.

Procedure for time-course study. Under air, a vial (Vial A) was charged with (IMes)CuCl (6.1 mg, 15 μmol) and fluorene (16.7 mg, 0.10 mmol) as an internal standard. A vial (Vial B) was charged with **1a** (46 mg, 0.30 mmol) and **2a** (73 mg, 0.30 mmol). A vial (Vial C) was charged with Pd(OAc)₂ (0.7 mg, 3.0 μmol) and dppp (1.2 mg, 3.0 μmol). These vials were transferred into a glove box. In the glove box, LiOt-Bu in 1.0 M THF solution (60 μL, 60 μmol) was added to Vial A. 1.95 mL of 1,4-dioxane was added into Vial B and 0.60 mL of 1,4-dioxane was added into Vial C. **3a** (35 mg, 0.30 mmol) was added to Vial C. Then, the solution in Vial B and C were added to Vial A. The resulting mixture was stirred at 100 °C. 100 μL of sample was taken every 10 minutes from 0 to 1 h and every 30 minutes from 1 to 6 h. All of the volatiles in the samples were removed in *vacuo* and filtered through a short pad of silica gel and Celite[®]. The resulting samples were measured by ¹H NMR spectroscopy to calculate yields of the hydroarylation product **5a** and **4a**.

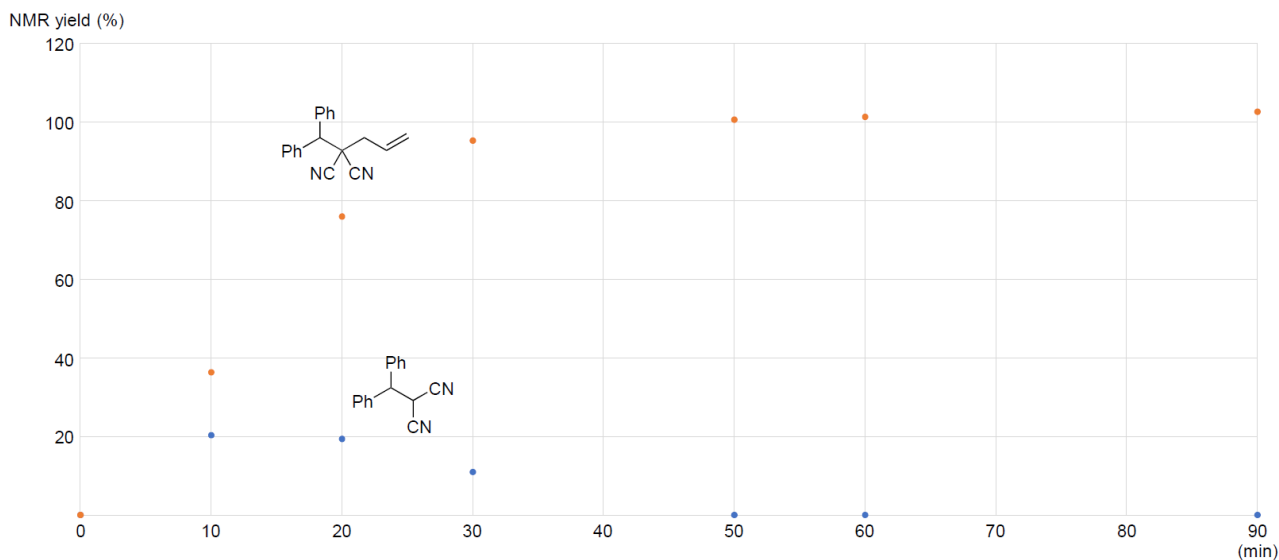
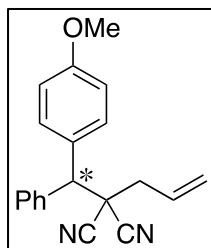
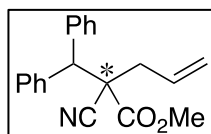


Figure S2. A time-course of the allylarylation of **1a** with **2a** and **3a**.



Enantioselective allylarylation of 4b. The reaction of **1a** (56 mg, 0.30 mmol), **2b** (82 mg, 0.30 mmol) and **3a** (35 mg, 0.30 mmol) using **L1** as a ligand followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 95:5 to 80:20) gave the title compound (17.2 mg, 57 μ mol, 19%) as pale yellow oil. The resonance is of ¹H and ¹³C NMR spectra of the product were consist with those of **4b** in Table 2. The ee was determined on a Daicel Chiralpak AS-H column with *n*-hexane–2-propanol = 99:1, flow = 1.0 mL/min, detection at 220 nm. Retention times: 9.2 min (major enantiomer), 11.1 min (minor enantiomer). $[\alpha]^{19}_{\text{D}} -0.746$ (*c* 0.0067, CH₂Cl₂).

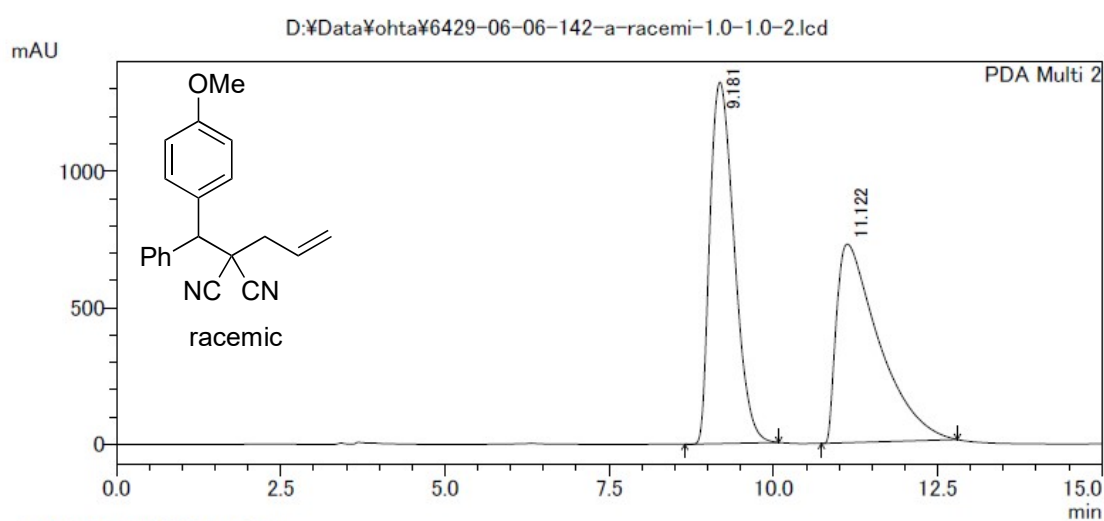


Enantioselective allylarylation of 4o. The reaction of **1e** (56 mg, 0.30 mmol), **2a** (73 mg, 0.30 mmol) and **3a** (52 mg, 0.45 mmol) using **L2** as a ligand followed by purification by MPLC (25 g Biotage[®] SNAP Ultra, *n*-hexane:ethyl acetate = 95:5 to 80:20) gave the title compound (74 mg, 0.24 mmol, 81%) as white solid. The resonance is of ¹H and ¹³C NMR spectra of the product were consist with those of **4o** in Table 3. The ee was determined on a Daicel Chiralpak AD-H column with *n*-hexane–2-propanol = 99:1, flow = 1.0 mL/min, detection at 254 nm. Retention times: 8.7 min (major enantiomer), 10.2 min (minor enantiomer). $[\alpha]^{19}_{\text{D}} 6.47$ (*c* 0.0056, CH₂Cl₂).

==== Shimadzu LCsolution 分析レポート ====

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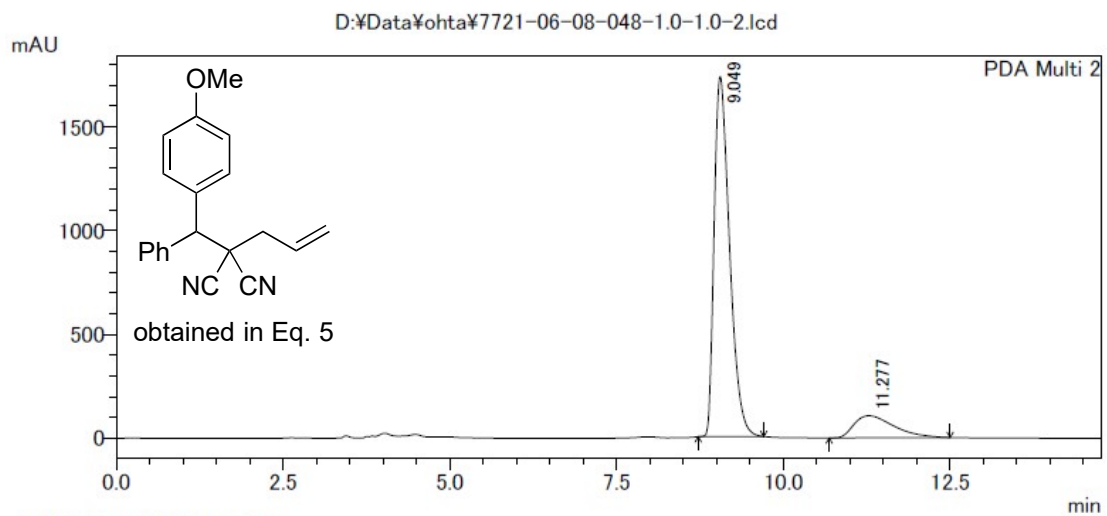
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==== Shimadzu LCsolution 分析レポート ====

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1 PDA Multi 2/220nm 4nm

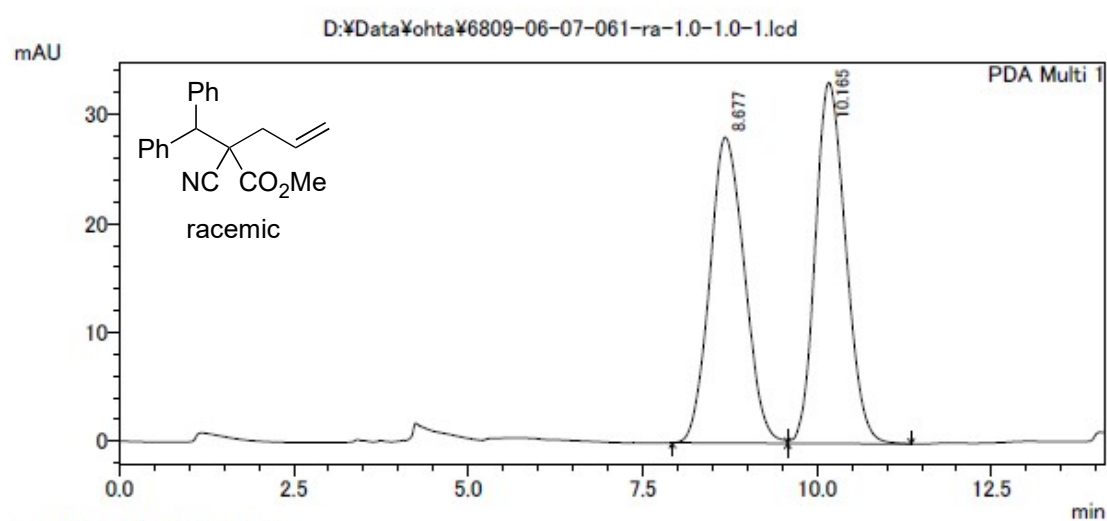
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==== Shimadzu LCsolution 分析レポート ====

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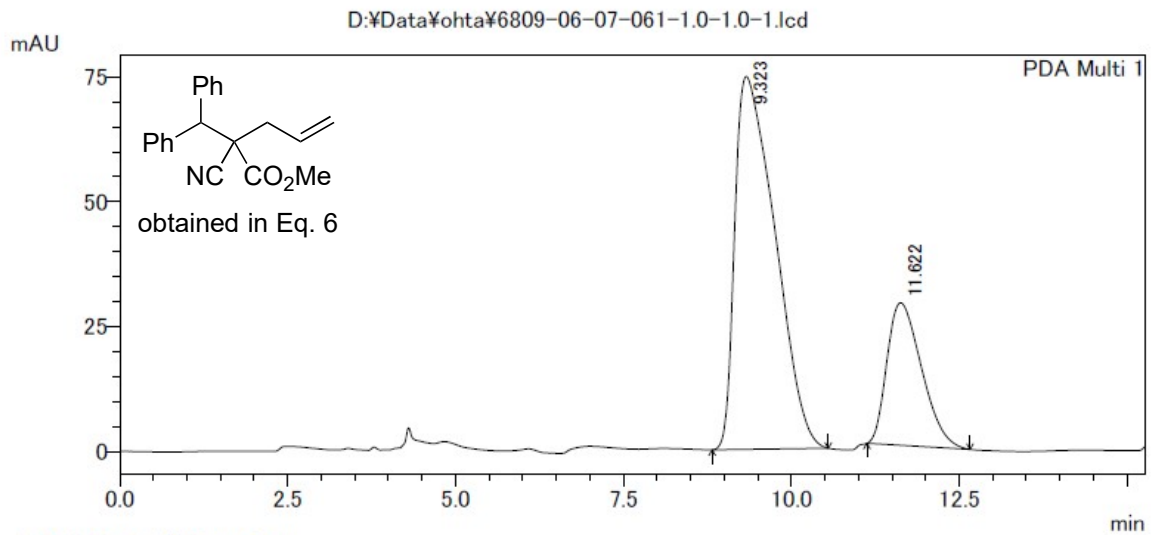
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合計		1977063	61218	100.000	100.000

==== Shimadzu LCsolution 分析レポート ====

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1 PDA Multi 1/254nm 4nm

ピークテーブル

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2	11.622	1025065	28543	24.826	27.649
合計		4129002	103233	100.000	100.000

References

- ¹ (a) G. B. Kharas, K. Watson, *Macromolecules* 1989, **22**, 3871–3877; (b) D. E. Q. Jimenez, I. M. Ferreira, W. G. Birolli, L. P. Fonseca, A. L. M. Porto, *Tetrahedron* 2016, **72**, 7317–7322; (c) M. A. Camerino, M. Liu, S. Moriya, T. Kitahashi, A. Mahgoub, S. J. Mountford, D. K. Chalmers, T. Soga, I. S. Parhar, P. E. Thompson, *J. Pept. Sci.* 2016, **22**, 406–414; (d) Y. Liu, W. Yang, Y. Wu, B. Mao, X. Gao, H. Liu, Z. Sun, Y. Xiao, H. Guo, *Adv. Synth. Catal.* 2016, **358**, 2867–2872.
- ² (a) Y. Nakao, H. Imanaka, A. K. Sahoo, A. Yada, T. Hiyama, *J. Am. Chem. Soc.* 2005, **127**, 6952–6953; (b) Y. Nakao, J. Chen, H. Imanaka, T. Hiyama, Y. Ichikawa, W.-L. Duan, R. Shintani, T. Hayashi, *J. Am. Chem. Soc.* 2007, **129**, 9137–9143; (c) M. Iizuka, Y. Kondo, *Eur. J. Org. Chem.* 2008, 1161–1163; (d) Y. Nakao, H. Imanaka, J. Chen, A. Yada, T. Hiyama, *J. Organomet. Chem.* 2007, **692**, 585–603; (e) Y. Nakao, M. Takeda, T. Matsumoto, T. Hiyama, *Angew. Chem., Int. Ed.* 2010, **49**, 4447–4450.
- ³ (a) T. R. Ramadhar, J. Kawakami, A. J. Lough, R. A. Batey, *Org. Lett.* 2010, **12**, 4446–4449; (b) X. Qian, A. Auffrant, A. Felouat, C. Gosmini, *Angew. Chem., Int. Ed.* 2011, **50**, 10402–10405; (c) S. Ghosh, S. Chaudhuri, A. Bisai, *Org. Lett.* 2015, **17**, 1373–1376; (d) H. Chen, X. Jia, Y. Yu, Q. Qian, H. Gong, *Angew. Chem., Int. Ed.* 2017, **56**, 13103–13106.
- ⁴ O. Santoro, A. Collado, A. M. Z. Slawin, S. P. Nolan, C. S. J. Cazin, *Chem. Commun.* 2013, **49**, 10483–10485.
- ⁵ K. Mizuno, M. Ikeda, S. Toda, Y. Otsuji, *J. Am. Chem. Soc.* 1988, **110**, 1288–1290.

