

SiCl₄ mediated one-pot synthesis of novel spirobibenzopyrans as potent anticancer agents

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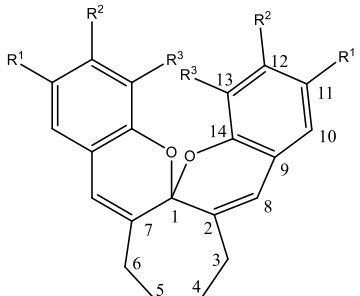
Supporting information:

1. Synthesis

The spirobibenzopyrans were synthesized by stirring a mixture of the ketone (0.01 mol), the required substituted salicylaldehyde (0.02 mol), and SiCl₄ (silicon tetrachloride, 0.03 mol) in absolute ethanol (20ml) under nitrogen atmosphere. The above reaction mixture was stirred till the completion of the reaction indicated by formation of solid (generally white in color) in the reaction vessel. After completion the contents were poured into ice to quench the catalyst and stirred for 15 minutes. The solid was filtered, washed with water, dried and recrystallized from distilled chloroform. If there was formation of a colored solid in the reaction vessel, the reaction was worked up in the similar manner as mentioned before. The coloured solid obtained was then suspended in diethyl ether (20 mL) and to it 20 mL of aqueous ammonia was added. The suspension was shaken vigorously and the ether layer was separated, dried over anhydrous Na₂SO₄. Then the ether layer was evaporated to obtain a crude solid which was recrystallized from distilled acetone or chloroform. The compounds synthesized along with their sample code and yield shown in **SD table I** below.

SD Table I: Structure, IUPAC names, % yield and melting points of the synthesized spirobibenzopyrans.

CODE	IUPAC NAME	SUBSTITUENT	YIELD (%)
CHP-SAL	6,7,8,9-tetrahydrocyclohepta[1,2-b:1,7-b']dichromene	R ¹ = -H R ² = -H	90



CHP-4MO-SAL	2,13-dimethoxy-6,7,8,9-tetrahydrocyclohepta[1,2-b:1,7-b']dichromene	R ¹ = -H R ² = -OCH ₃	64
CHP-5MO-SAL	3,12-dimethoxy-6,7,8,9-tetrahydrocyclohepta[1,2-b:1,7-b']dichromene	R ¹ = -OCH ₃ R ² = -H	84
CHP-5Cl-SAL	3,12-dichloro-6,7,8,9-tetrahydrocyclohepta[1,2-b:1,7-b']dichromene	R ¹ = -Cl R ² = -H	88
CHP-5Br-SAL	3,12-dibromo-6,7,8,9-tetrahydrocyclohepta[1,2-b:1,7-b']dichromene	R ¹ = -Br R ² = -H	86
CHP-DiCl-SAL	1,3,12,14-tetrachloro-6,7,8,9-tetrahydrocyclohepta[1,2-b:1,7-b']dichromene	R ¹ = -Cl R ² = -H R ³ = -Cl	69
CHX-SAL	7,8-dihydro-6H-chromeno[3,2-d]xanthene	R ¹ = -H R ² = -H	40
CHX-4MO-SAL	2,12-dimethoxy-7,8-dihydro-6H-chromeno[3,2-d]xanthene	R ¹ = -H R ² = -OCH ₃	33
CHX-5MO-SAL	3,11-dimethoxy-7,8-dihydro-6H-chromeno[3,2-d]xanthene	R ¹ = -OCH ₃ R ² = -H	81
CHX-5Cl-SAL	3,11-dichloro-7,8-dihydro-6H-chromeno[3,2-d]xanthene	R ¹ = -Cl R ² = -H	86
CHX-5Br-SAL	3,11-dibromo-7,8-dihydro-6H-chromeno[3,2-d]xanthene	R ¹ = -Br R ² = -H	78
CHX-DiCl-SAL	1,3,11,13-tetrachloro-7,8-dihydro-6H-chromeno[3,2-d]xanthene	R ¹ = -Cl R ² = -H R ³ = -Cl	75

2. Instrumentation

The ¹H NMR spectra were obtained on VARIAN 200 MHz, ¹³C NMR on VARIAN 50 MHz for CHP-SAL, CHP-4MO-SAL, CHP-5Cl-SAL, and CHP-5Br-SAL, for CHP-5MO-SAL and CHX-4MO-SAL the ¹H NMR spectra were obtained on VARIAN 500 MHz, and ¹³C NMR on VARIAN 125 MHz and finally the ¹H NMR spectra were obtained on VARIAN 400 MHz, and ¹³C NMR on VARIAN 100 MHz for CHP-DiCl-SAL and all the remaining CHX derivatives. TMS was used as the internal standard and CDCl₃ as the solvent. For mass spectra AGILENT 6430 Triple Quad LC/MS was employed. The FT-IR spectra were recorded between 400 and 4000 cm⁻¹ using KBr

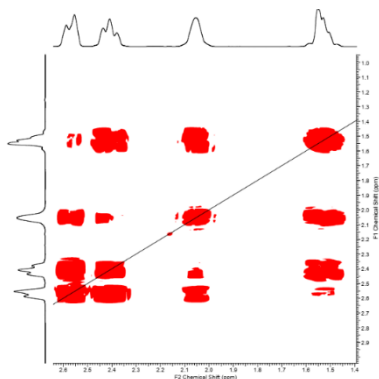
pellets employing Thermo-Nicolet Avatar 370 spectrophotometer. UV-Vis spectra in methanol were recorded in the wavelength range 200-600 nm using Shimadzu 2450 spectrophotometer.

For HPLC Agilent 1260 Infinity high performance liquid chromatography system, equipped with a quaternary solvent deliver system, inline degasser, autosampler and photo diode array detector, was used. A Waters Nova - Pak C18 (3.9mm X 150mm) connected with a Zorbax C18 guard column (20 mm × 4 mm, 5 μm) was applied for all analyses. Detection wavelengths were set at the respective λ_{max} for each compound (characterization data). The mobile phase A consisted of water and mobile phase B was acetonitrile. A flow rate of 1 ml/min was maintained and the elution was conducted using a linear gradient mode as shown in supplementary (SD table 1 and 2). The separation was carried out at 25 °C with an injection volume of 20 μl and samples were analyzed in triplicate. Spectral characteristics of eluted peaks were recorded using diode array detector from 200 to 700 nm.

Single crystals of suitable dimensions were chosen carefully for X-ray diffraction studies. The X-ray intensity data were collected at a temperature of 293(2) K on a Bruker Proteum2 CCD diffractometer equipped with an X-ray generator operating at 45 kV and 10 mA, using $\text{CuK}\alpha$ radiation of wavelength 1.54178 Å. Data were collected for 24 frames per set with different settings of φ (0° and 90°), keeping the scan width of 0.5°, exposure time of 2 s, the sample to detector distance of 45.10 mm and 2θ value at 46.6°. The complete data sets were processed using *SAINT PLUS*¹. The structures were solved by direct methods and refined by full-matrix least squares method on F^2 using *SHELXS* and *SHELXL* programs². The geometrical calculations were carried out using the program *PLATON*³. The molecular and packing diagrams were generated using the software *MERCURY*⁴.

3. Characterization

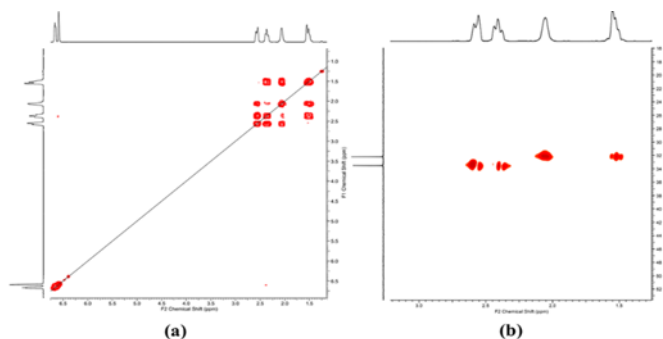
The ¹H-COSY for the –CH₂- protons is shown in figure I.



SD Figure I. 2D-COSY of CHP-SAL zoomed to view the –CH₂- protons

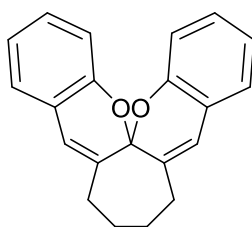
From the contour of the signal it is evident that the signals at δ 2.56 and δ 2.41 are the geminal hydrogens H-3a, H-3b / H-6a, H-6b. Also the fact that both the signals couple with the other signals appearing at δ 2.05 and δ 1.53 which are H-4 and H-5 respectively indicate that they are indeed geminal hydrogens. It is further confirmed by two more facts: one, there is a small amount of

allylic coupling observed with the signal at $\delta 2.41$ with the H-7 at $\delta 6.70$ which is absent for the signal at $\delta 2.56$ (shown in figure IIa). This tells us that one of the hydrogens is projecting outward the cycloheptanone ring with the allylic coupling (Ha) and the other hydrogen is inwards having no allylic coupling (Hb). Fact two, from the HMQC spectra shown in figure IIb it is clear that the signals at $\delta 2.56$ and $\delta 2.41$ are for 2 hydrogens each H-3a, H-6a and H-3b, H-6b corresponding to C3 and C6 which appear at the same value of $\delta 32$ in the ^{13}C -NMR spectrum. Additionally, from the Gaussian studies we obtained the theoretical NMR values which upon analysis showed that indeed the protons H-3a and H-3b, H-6a and H-6b to have different chemical shift values. The calculation table is given in supplementary section as SD table 4. Our theory of the difference in chemical environment of H-3a and H-3b is finally confirmed by the single crystal XRD structure shown in **figure III** for **CHP-5Cl-SAL** where the H-3a and H-6a protons are outward and the H3b and H6b protons are inwards. Similar observation can be made from the single crystal XRD structure of **CHP-5MO-SAL** shown in figure IV. We thus understand the non-equivalence of the protons at H3/H6 in the **CHP** system using the spectroscopic techniques.

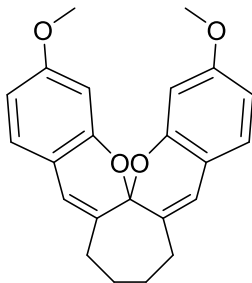


SD Figure II. (a) 2D-COSY of CHP-SAL zoomed to view the allylic coupling. (b) 2D-HMQC of CHP-SAL zoomed to view the $-\text{CH}_2-$ region

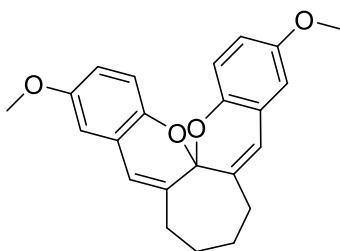
Characterization data of the synthesized spirobibenzopyrans:



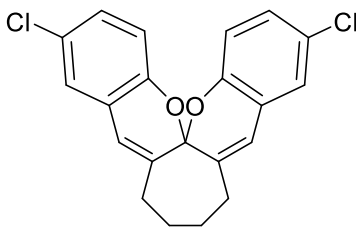
CHP-SAL: Colorless solid. M.P.: 215-216 °C; $^1\text{H-NMR}$ (200 MHz, CDCl_3) δ , ppm = 7.20 (t, H-12, $J=7.0$ Hz), 7.10 (d, H-10, $J=7.0$ Hz), 6.95 (t, H-11, $J=7.0$ Hz), 6.8(d, H-13, $J=7.0$ Hz), 6.70 (s, H-8), 2.60 (m, H-3b and H-6b, $J=6.4$ Hz), 2.40 (m, H-3a and H-6a), 2.10 (m, H-4), and 1.55 (m, H-5); $^{13}\text{C-NMR}$ (150 MHz, CDCl_3) δ , ppm = 150(C14), 136 (C2, C7), 130(C12), 126 (C10), 124 (C11), 122 (C8), 121 (C9), 116(C13), 107 (C1), 34(C4, C5), and 32 (C3, C6); IR (KBr disc): 3061-w (alkene C-H stretch), 2920-m (aliphatic C-H stretch), 1604-s (C=C stretch), 1575, 1557-m (aromatic skeletal bands), 1437 (aromatic C-H bend), 1374-m, 1247-s, 1141-s (C—O stretch), 795-s, 753-s (C-H oop bend); UV-vis (CHCl_3) λ_{max} nm: 241, 298; Mass: m/z $[\text{M}]^+= 302.0$



CHP-4MO-SAL: Pale colored solid. M.P.: 168.2-169.1 °C; ¹H-NMR(200 MHz, CDCl₃) δ, ppm = 7.25 (s, H-8), 7.06 (d, H-10), 6.61 (m, H-11), 6.40 (d, H-13), 2.52-2.39 (m, H-3 and H-6), 2.04 (m, H-4), 1.50 (m, H-5), and 3.71 (s, -OCH₃); ¹³C-NMR(50 MHz, CDCl₃) δ, ppm = 160(C12), 151 (C14), 132(C2, C7), 126 (C8), 123 (C1), 114 (C9), 108 (C11), 102 (C13), 101 (C9), 33 (C4, C5), and 32 (C3, C6); IR (KBr disc): 3065-w (alkene C-H stretch), 2919-m (aliphatic C-H stretch), 1614-s (C=C stretch), 1572-m, 1463-s (aromatic skeletal bands), 1438-s, 1372-s (aromatic C-H bend), 1325-s, 1272-s, 1117-s (C—O stretch), 908-s, 864-s, 768s (C-H oop bend); UV-vis (CHCl₃) λ_{max} nm: 240, 278; Mass: m/z [M]⁺= 362.0

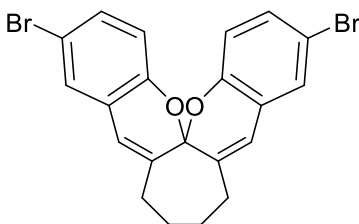


CHP-5MO-SAL: Pale yellow solid. M.P.: 180.2-180.9 °C; ¹H-NMR(500 MHz, CDCl₃) δ, ppm = 7.20 (s, H-8), 6.71 (d, H-13, J=7.8 Hz), 6.69 (d, H-12, J=7.8 Hz), 6.61 (s, H-10), 2.55 (m, H-3b and H-6b), 2.40 (m, H-3a and H-6a), 2.05 (m, H-4), 1.52 (m, H-5), and 3.77 (s, -OCH₃); ¹³C-NMR(101 MHz, CDCl₃) δ, ppm = 154 (C11), 144 (C14), 136 (C2, C7), 122 (C8), 121 (C9), 119 (C12), 117 (C13), 111 (C10), 102 (C1), 34 (C4, C5), 32 (C3, C6), and 56 (-OCH₃); IR (KBr disc): 3051-w (alkene C-H stretch), 2927-m (aliphatic C-H stretch), 1616-m (C=C stretch) 1581-m, 1487-s (aromatic skeletal bands), 1451-s 1433-s (aromatic C-H bend), 1311-s, 1200-s, 1141-s (C—O stretch), 955-s, 880-s, 750-s (C-H oop bend); UV-vis (CHCl₃) λ_{max} nm: 243, 321; Mass: m/z [M+H]⁺ = 363.1592

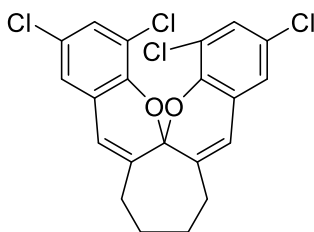


CHP-5Cl-SAL: White solid. M.P.: 127-128 °C; ¹H-NMR (200 MHz, CDCl₃) δ, ppm = 7.13 (s, H-10), 7.09 (d, H-12, J=8 Hz), 6.71 (d, H-13, J=8 Hz), 6.60(s, H-8), 2.55 (m, H-3b and H-6b), 2.37 (m, H-3a and H-6a), 2.06 (m, H-4), and 1.55 (m, H-5); ¹³C-NMR(50 MHz, CDCl₃) δ, ppm = 148 (C14), 137 (C2, C7), 129 (C12), 127 (C11), 126 (C10), 123 (C9), 118 (C13), 114 (C8), 101

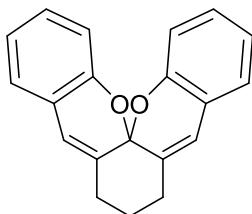
(C1), 34 (C4, C5), and 32 (C3, C6); IR (KBr disc): 3060-w (alkene C-H stretch), 2926-m (aliphatic C-H stretch), 1634-m (C=C stretch), 1560-m (aromatic skeletal bands), 1479-s, 1445-s (aromatic C-H bend), 1362-s, 1229-s, 1181-s (C—O stretch), 1077-m, 1052-s (Aromatic -Cl stretch), 875-s, 749-s, 716-s (C-H oop bend); UV-vis (CHCl₃) λ_{max} nm: 242, 308; Mass: m/z [M-H]⁺ = 370.0, [M-H+2]⁺ = 372.0



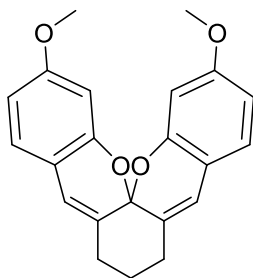
CHP-5Br-SAL: White solid. M.P.: 120-122 °C; ¹H-NMR (200 MHz, CDCl₃) δ , ppm = 7.40 (s, H-10), 7.30 (d, H-12, J=7.7 Hz), 6.70 (d, H-13, J=7.7 Hz), 6.60 (s, H-8), 2.60 (m, H-3b and H-6b), 2.40 (m, H-3a and H-6a), 2.00 (m, H-4), and 1.60 (m, H-5); ¹³C-NMR (50 MHz, CDCl₃) δ , ppm = 148 (C14), 136 (C2, C7), 132 (C12), 128 (C10), 125 (C1), 123 (C11), 119 (C13), 117 (C8), 114 (C9), 34 (C4, C5), and 32 (C3, C6); IR (KBr disc): 3059-w (aromatic C-H stretch); 2927-s (aliphatic C-H), 1633-m (C=C stretch); 1476 - s, 1419 -m (aromatic C-H bend), 1339-m (CH₃ bend); 1286-m, 1228-m, 1179-m, 1126-m (C—O stretch); 1067-m, 1054-m (Ar-Br stretch); 852-s, 682-s (C-H oop). UV-vis (CHCl₃) λ_{max} nm: 242, 309; Mass: m/z [M]⁺ = 460.0 and [M+2]⁺ = 462.0



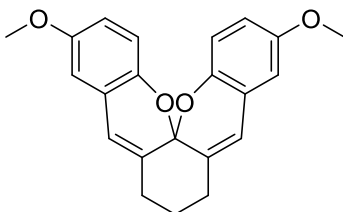
CHP-DiCl-SAL: Pale white solid. ¹H-NMR (400 MHz, CDCl₃) δ , ppm = 7.20 (d, H-10, J=2.8Hz), 7.06 (d, H-12, J=2.8Hz), 6.63 (s, H-8), 2.63 (m, H-3b and H-6b), 2.39 (m, H-3a and H-6a), 2.09 (m, H-4), and 1.54 (m, H-5); ¹³C-NMR (100 MHz, CDCl₃) δ , ppm = 144 (C14), 137 (C2, C7), 129 (C12), 128 (C11), 125 (C13), 124 (C9), 123 (C10), 122 (C8), 101 (C1), 33 (C4, C5), and 31 (C3, C6); IR (KBr disc): 3040-w (alkene C-H stretch), 2920-m (aliphatic C-H stretch), 1610-s (C=C stretch), 1577-m, 1465-s (aromatic skeletal bands), 1440-s, 1373-s (aromatic C-H bend), 1320-s, 1275-s, 1110-s (C—O stretch), 910-s, 866-s, 770s (C-H oop bend); UV-vis (CHCl₃) λ_{max} nm: 240, 278; Mass: m/z [M - H]⁺ = 439, [M - H + 2]⁺ = 441



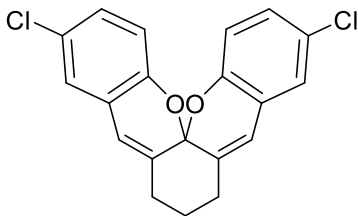
CHX-SAL: White solid. M.P.: 160-161 °C; ¹H-NMR (400 MHz, CDCl₃) δ, ppm = 7.22 (2H, d, H-9, J=7.4 Hz), 7.16 (2H, t, H-11, J=7.4 Hz), 7.02 (2H, t, H-10, J=7.4 Hz), 6.92 (2H, d, H-12, J=7.4 Hz), 6.69(2H, s, H-7), 2.53 (4H, t, H-3 and H-5, J=6.5 Hz), and 1.82 (2H, m, H-4, J=6.5 Hz); ¹³C-NMR (100 MHz, CDCl₃) δ, ppm = 150.0 (C13), 132.0 (C2, C6), 129.0 (C11), 126.0 (C9), 122.0 (C10), 121.8 (C8), 121.3 (C7), 117.0 (C12), 96.0 (C1), 26.0 (C3, C5), and 20.0 (C4); IR (KBr disc): 3150 cm⁻¹ -w (sp² C-H stretch); 2920-m (aliphatic C-H), 1606-m, (C=C stretch); 1572 - s, 1481 -m, 1455 -m (aromatic C-H bend), 1219-m, 1124-m, (C—O stretch); 751-s (C-H oop). UV-vis (CHCl₃) λ_{max} nm: 259, 296; Mass: m/z [M]⁺ = 288.0



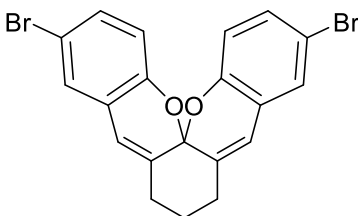
CHX-4MO-SAL: White solid. M.P.: 168.2-169.1 °C; ¹H-NMR(500 MHz, CDCl₃) δ, ppm = 7.20 (s, H-7), 6.71 (d, H-9, J=7.8 Hz), 6.69 (d, H-10, J=7.8 Hz), 6.61 (s, H-12), 2.49 (t, H-3 and H-5), 1.78 (quintet, H-4), and 3.72 (s, -OCH₃); ¹³C-NMR (125 MHz, CDCl₃) δ, ppm = 160.0 (C13), 151.0 (C11), 129.0 (C10), 126.2 (C2, C6), 121.4 (C9), 115.8 (C8), 108.3 (C7), 102.0 (C12), 96.0 (C1), 55.4 (-OCH₃), 26.0 (C3, C5), and 20.0 (C4); IR (KBr disc): 3065-w (alkene C-H stretch), 2919-m (aliphatic C-H stretch), 1614-s (C=C stretch), 1572-m, 1463-s (aromatic skeletal bands), 1438-s, 1372-s (aromatic C-H bend), 1325-s, 1272-s, 1117-s (C—O stretch), 908-s, 864-s, 768s (C-H oop bend); UV-vis (CHCl₃) λ_{max} nm: 240, 278; Mass: m/z [M + H]⁺ = 349.1



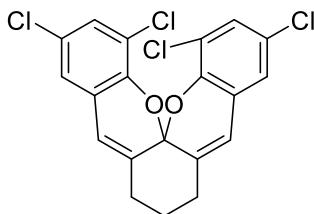
CHX-5MO-SAL: White solid. ¹H-NMR(400 MHz, CDCl₃) δ, ppm = 7.20 (s, H-7), 7.06 (d, H-12, J=6 Hz), 6.74 (d, H-11, J=6 Hz), 6.62 (s, H-9), 2.39 (t, H-3 and H-5), 1.79 (quintet, H-4), and 3.78 (s, -OCH₃); ¹³C-NMR (100 MHz, CDCl₃) δ, ppm = 154.5 (C13), 143.9 (C10), 136.9 (C11), 132.7.2 (C2, C6), 128.4 (C9), 124.3 (C8), 123.2 (C7), 12.07.0 (C12), 95.9 (C1), 55.7 (-OCH₃), 32.0 (C3, C5), and 33.0 (C4); IR (KBr disc): 3065-w (alkene C-H stretch), 2919-m (aliphatic C-H stretch), 1614-s (C=C stretch), 1572-m, 1463-s (aromatic skeletal bands), 1438-s, 1372-s (aromatic C-H bend), 1325-s, 1272-s, 1117-s (C—O stretch), 908-s, 864-s, 768s (C-H oop bend); UV-vis (CHCl₃) λ_{max} nm: 240, 278; Mass: m/z [M + H]⁺ = 349.1



CHX-5Cl-SAL: White solid. $^1\text{H-NMR}$ (200 MHz, CDCl_3) δ , ppm = 7.17 (d, H-9, $J=2\text{Hz}$), 7.10 (dd, H-11, $J=2\text{Hz}$, $J=8\text{Hz}$), 6.73 (d, H-12, $J=8\text{Hz}$), 6.61 (s, H-7), 2.51 (t, H-3 and H-5), and 1.81 (quintet, H-4); $^{13}\text{C-NMR}$ (50 MHz, CDCl_3) δ , ppm = 148.2 (C13), 133.1 (C10), 128.5 (C2, C6), 126.9 (C11), 125.8 (C9), 123.1 (C8), 121.4 (C7), 118.1 (C12), 96.0 (C1), 26.4 (C3, C5), and 20.3 (C4); IR (KBr disc): 3065-w (alkene C-H stretch), 2919-m (aliphatic C-H stretch), 1614-s (C=C stretch), 1572-m, 1463-s (aromatic skeletal bands), 1438-s, 1372-s (aromatic C-H bend), 1325-s, 1272-s, 1117-s (C—O stretch), 908-s, 864-s, 768s (C-H oop bend); UV-vis (CHCl_3) λ_{max} nm: 240, 278; Mass: m/z $[\text{M} - \text{H}]^+ = 356$, $[\text{M} - \text{H} + 2]^+ = 358$



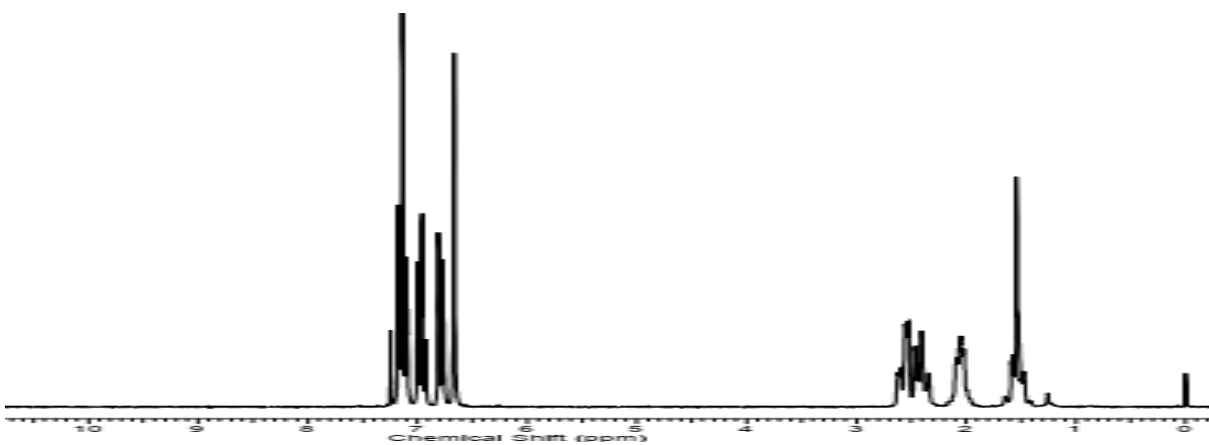
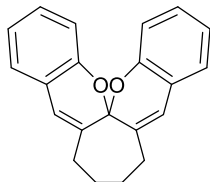
CHX-5Br-SAL: Pale yellow solid. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ , ppm = 7.40 (s, H-9), 7.30 (d, H-11, $J=7.7$ Hz), 6.70 (d, H-12, $J=7.7$ Hz), 6.60(s, H-7), 2.51 (t, H-3 and H-5), and 1.81 (quintet, H-5); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ , ppm = 149.2 (C13), 133.4 (C10), 128.9 (C2, C6), 126.3 (C11), 125.2 (C9), 123.1 (C8), 121.5 (C7), 118.4 (C12), 96.8 (C1), 26.4 (C3, C5), and 20.3 (C4); IR (KBr disc): 3059-w (aromatic C-H stretch); 2927-s (aliphatic C-H), 1633-m (C=C stretch); 1476 – s, 1419 -m (aromatic C-H bend), 1339-m (CH3 bend); 1286-m, 1228-m, 1179-m, 1126-m (C—O stretch); 1067-m, 1054-m (Ar-Br stretch); 852-s, 682-s (C-H oop). UV-vis (CHCl_3) λ_{max} nm: 242, 309; Mass: m/z $[\text{M}]^+ = 446.0$ and $[\text{M}+2]^+ = 448.0$



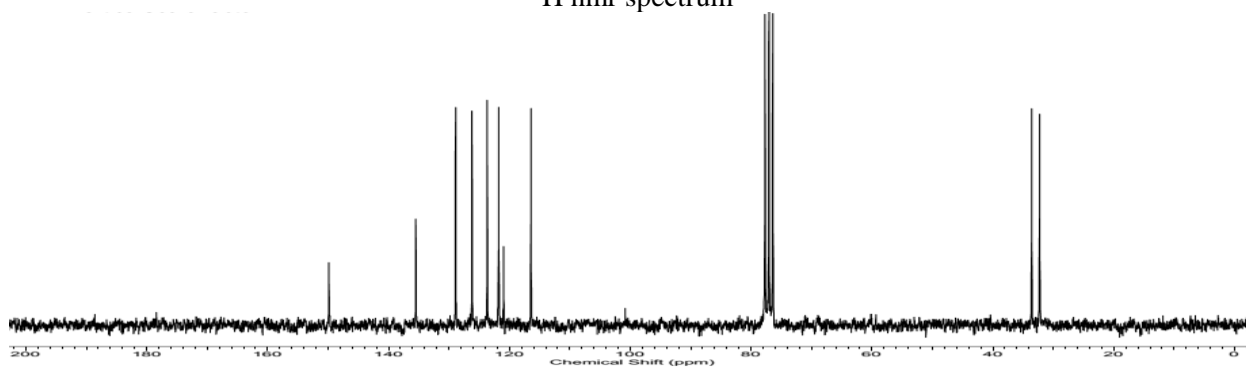
CHX-DiCl-SAL: Yellow solid. $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ , ppm = 7.20 (d, H-11, $J=2.8\text{Hz}$), 7.06 (d, H-9, $J=2.8\text{Hz}$), 6.63 (s, H-7), 2.53 (4H, t, H-3 and H-5, $J=6.5$ Hz), and 1.82 (2H, m, H-4, $J=6.5$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ , ppm = 144.0 (C13), 137.1 (C2, C6), 128.7 (C12), 126.8 (C10), 124.2 (C11), 123.3 (C9), 123.2 (C8), 122.4 (C7), 101.7 (C1), 33.4 (C3, C5), and 31.9 (C4); IR (KBr disc): 3040-w (alkene C-H stretch), 2920-m (aliphatic C-H stretch), 1610-s (C=C stretch), 1577-m, 1465-s (aromatic skeletal bands), 1440-s, 1373-s (aromatic C-H bend), 1320-s, 1275-s, 1110-s (C—O stretch), 910-s, 866-s, 770s (C-H oop bend); UV-vis (CHCl_3) λ_{max} nm: 240, 278; Mass: m/z $[\text{M} - \text{H}]^+ = 439$, $[\text{M} - \text{H} + 2]^+ = 441$

^1H -NMR, ^{13}C -NMR and Mass spectra of the spirobenzopyrans

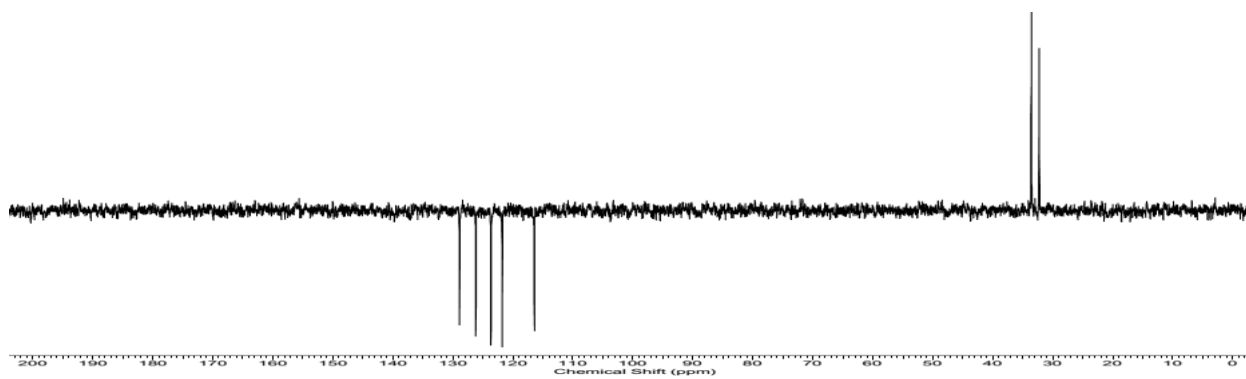
CHP-SAL



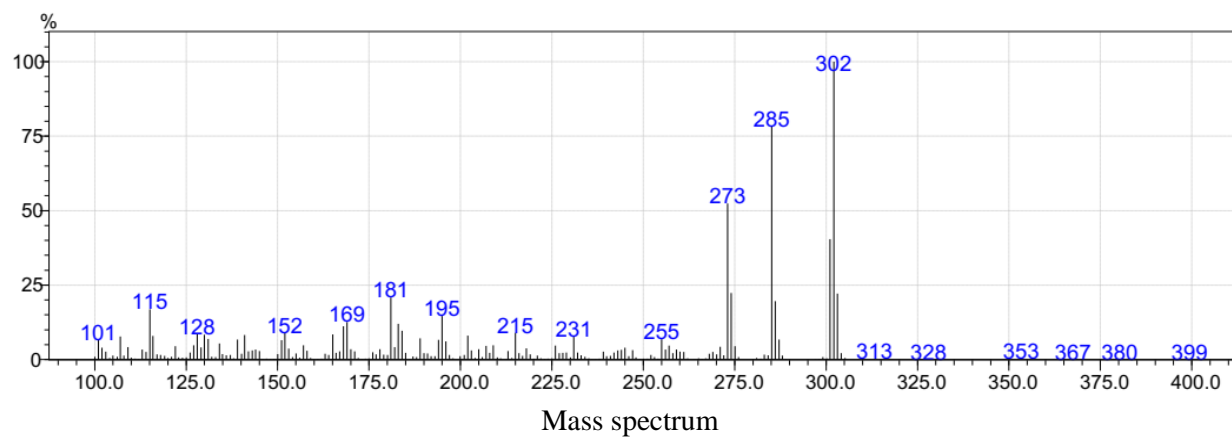
^1H nmr spectrum



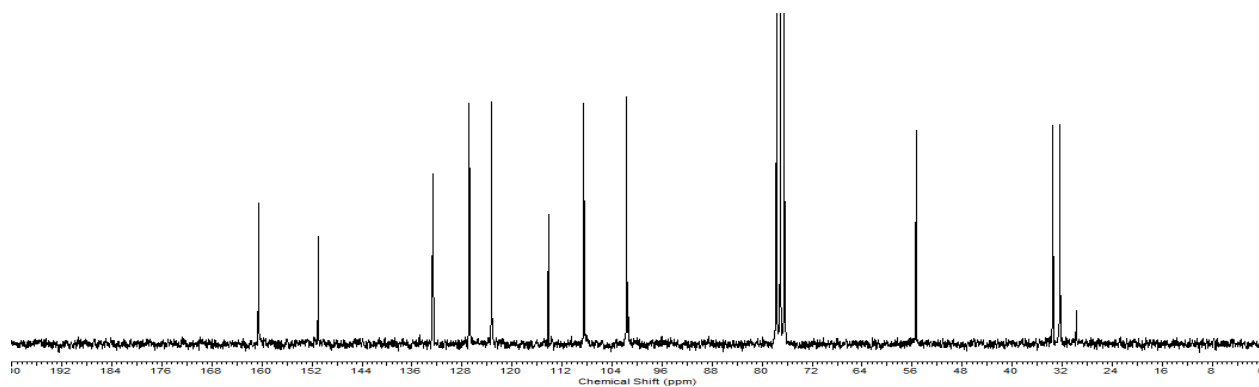
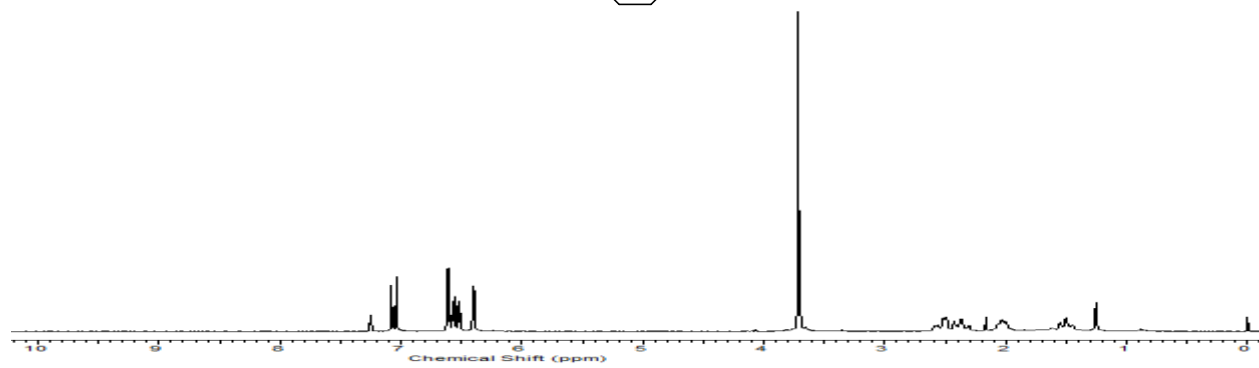
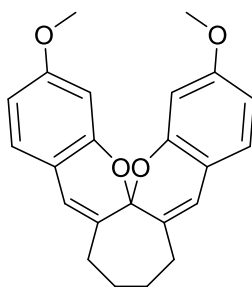
^{13}C nmr spectrum

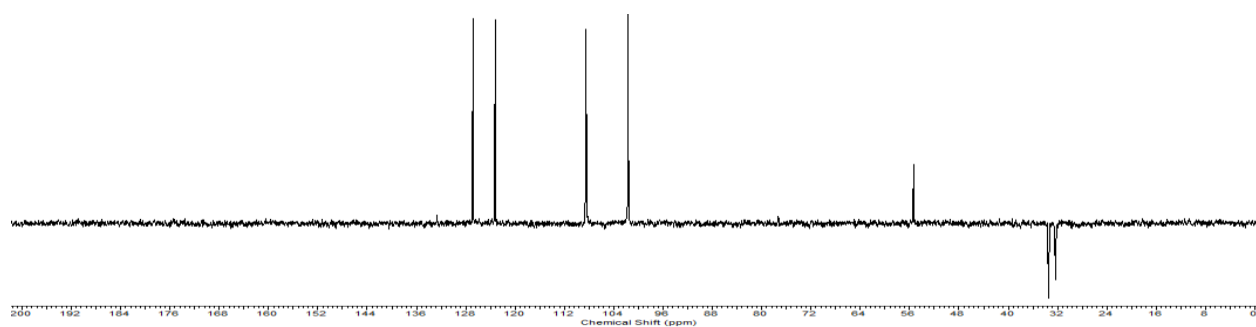


DEPT-135 ^{13}C nmr spectrum

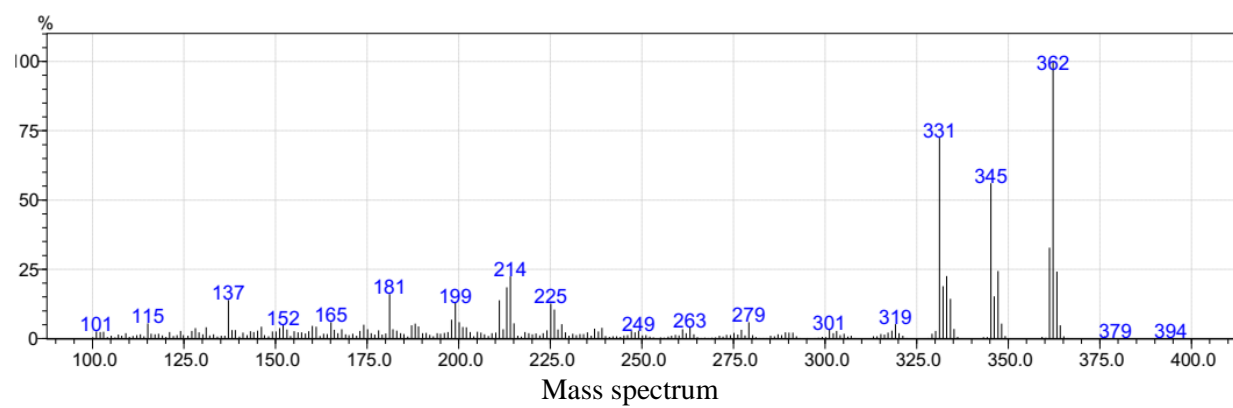


CHP-4MO-SAL

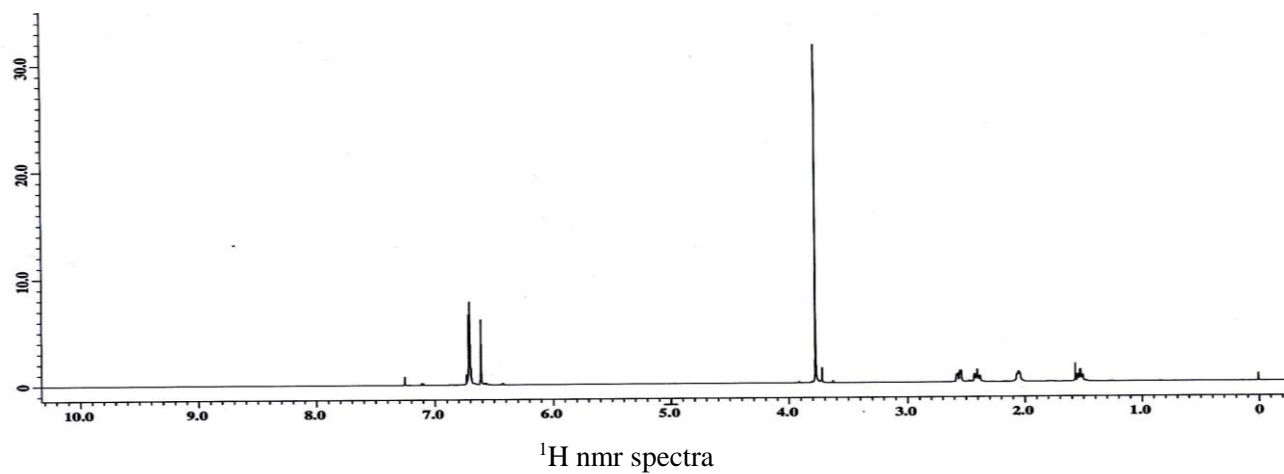
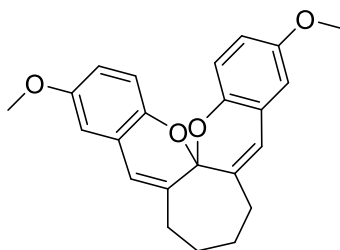


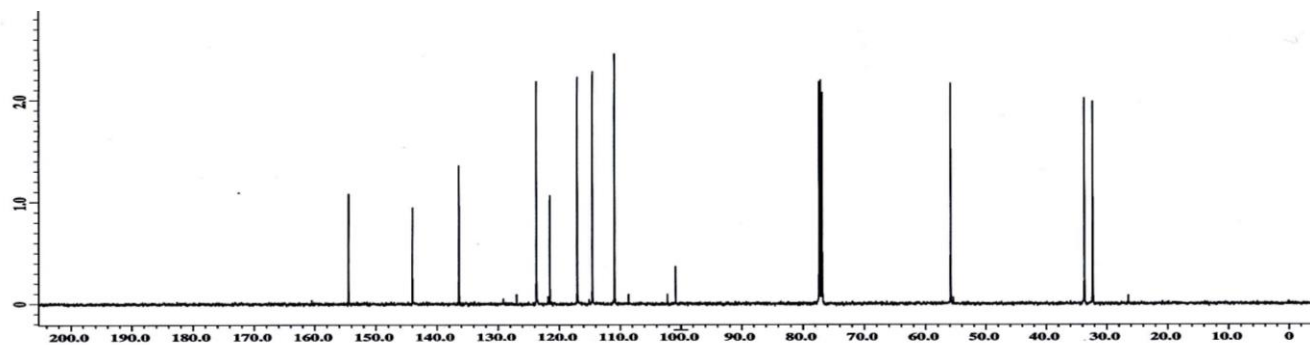


DEPT-135 ¹³C nmr spectrum

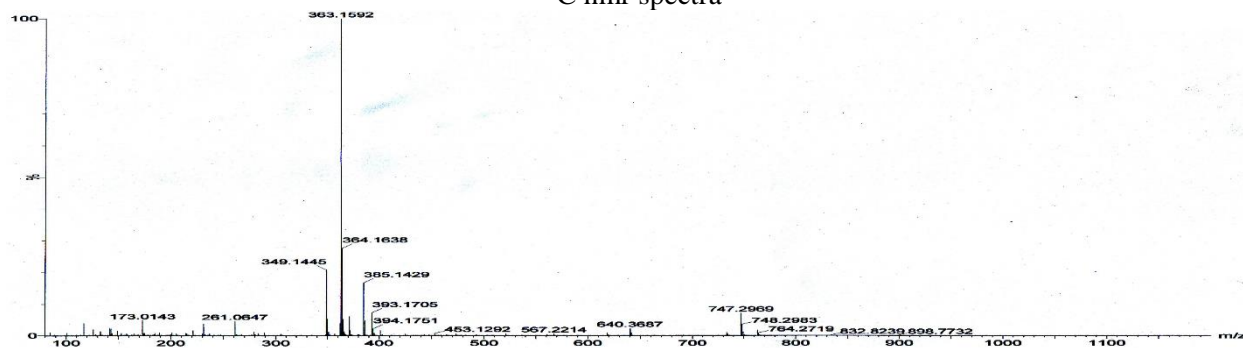


CHP-5MO-SAL



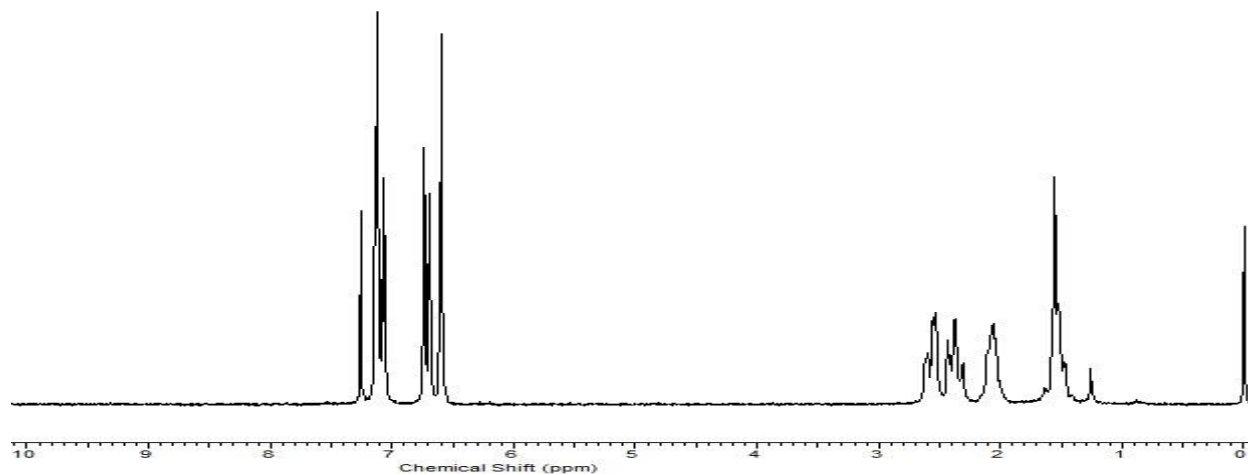
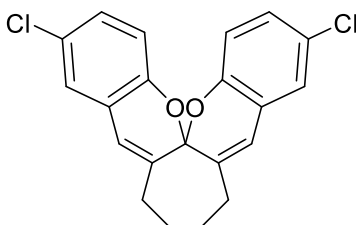


^{13}C nmr spectra

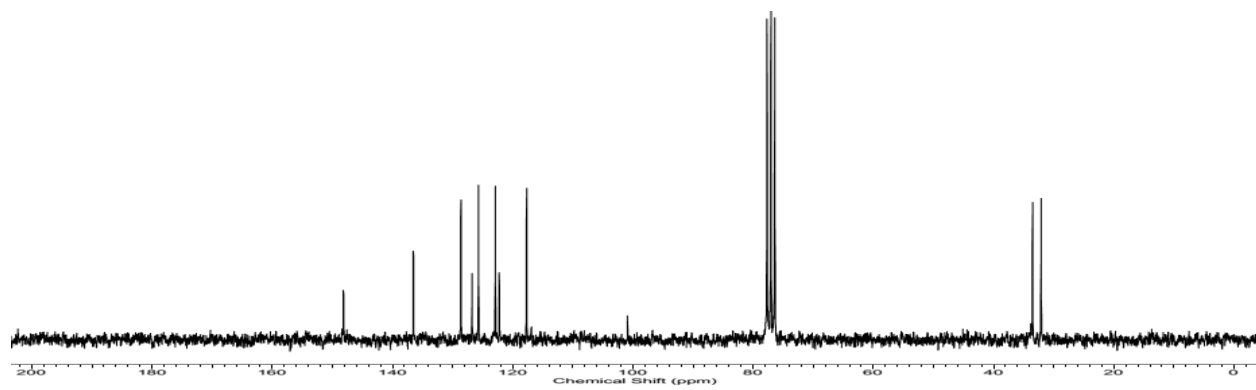


Mass spectrum

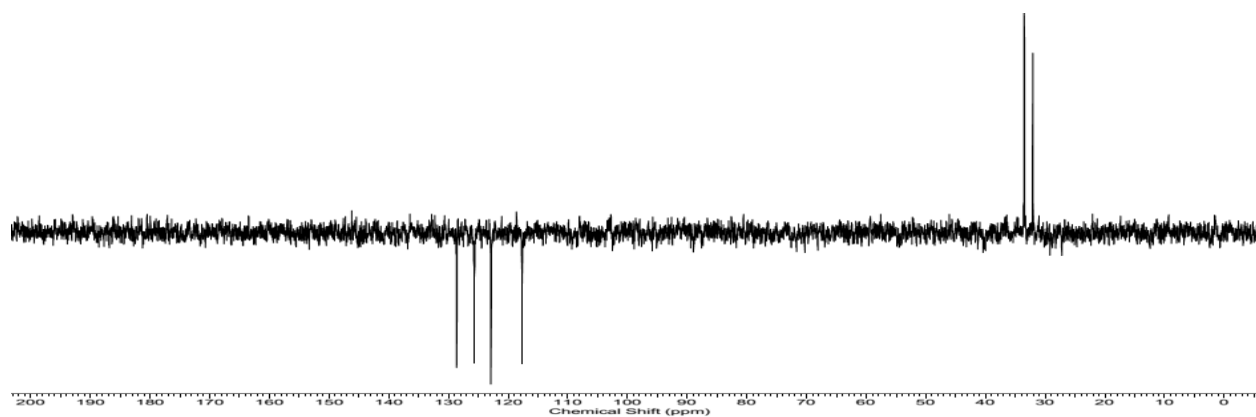
CHP-5Cl-SAL



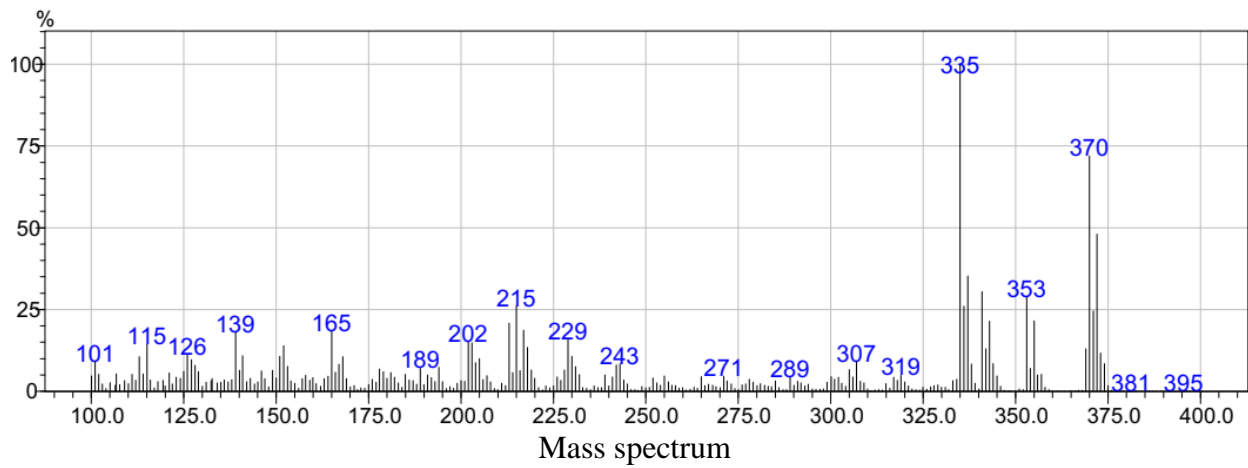
^1H nmr spectrum

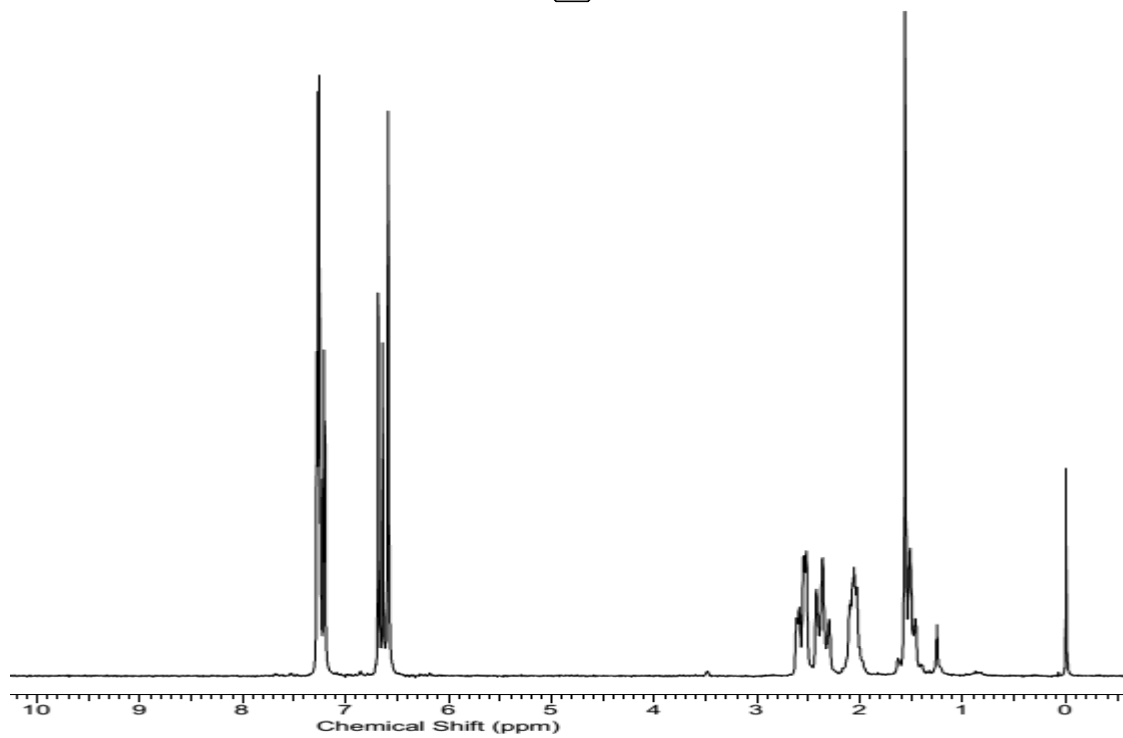
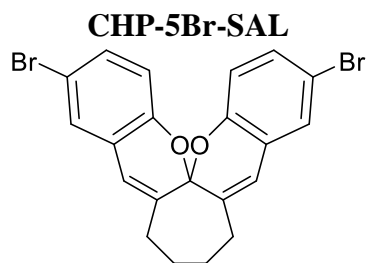


^{13}C nmr spectrum

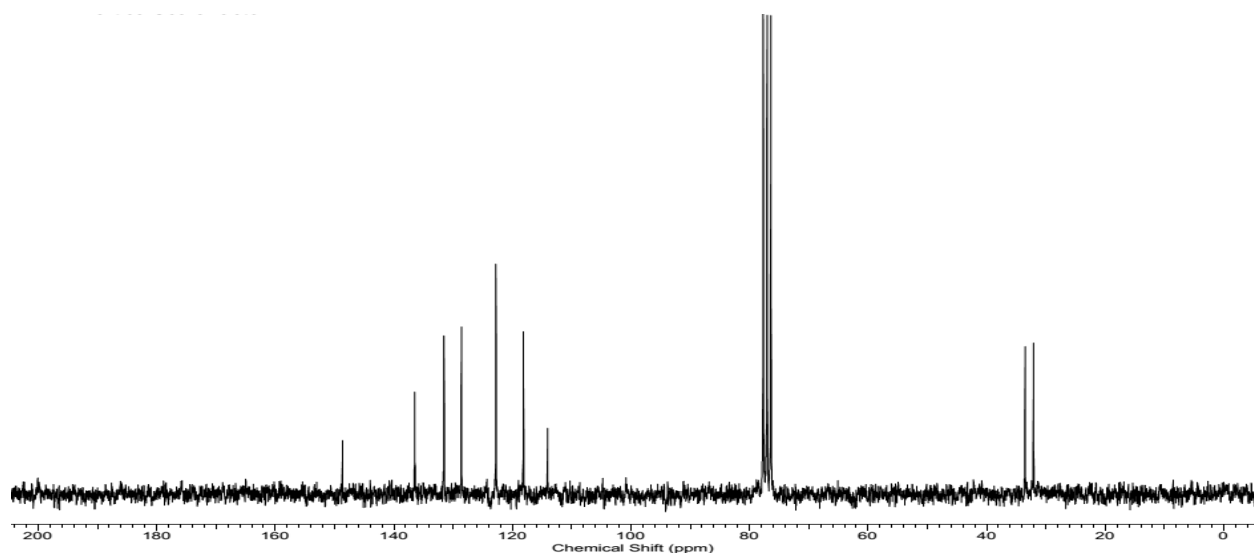


DEPT-135 ^{13}C nmr spectrum

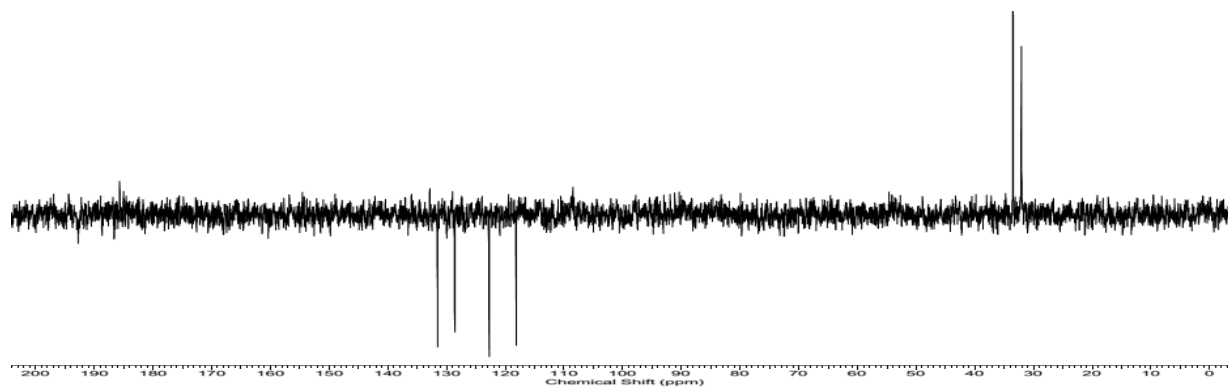




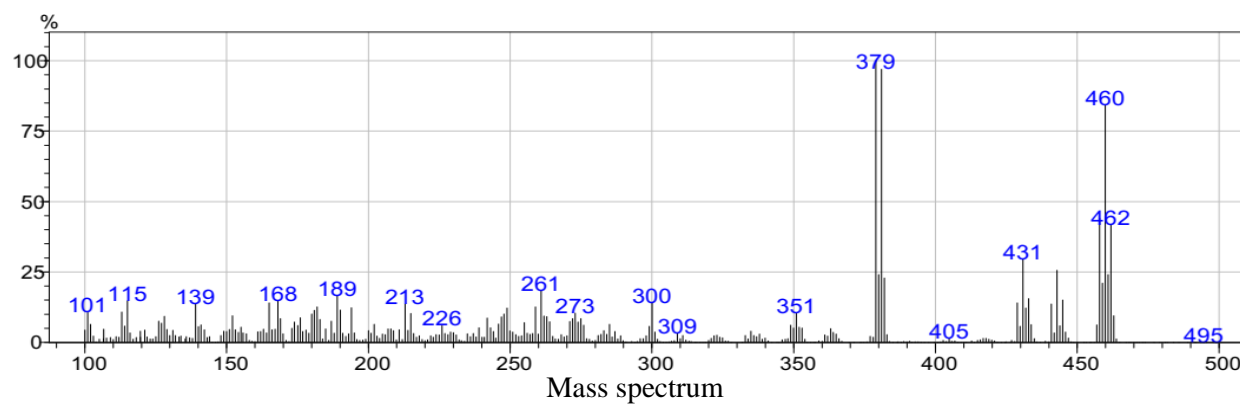
^1H nmr spectra



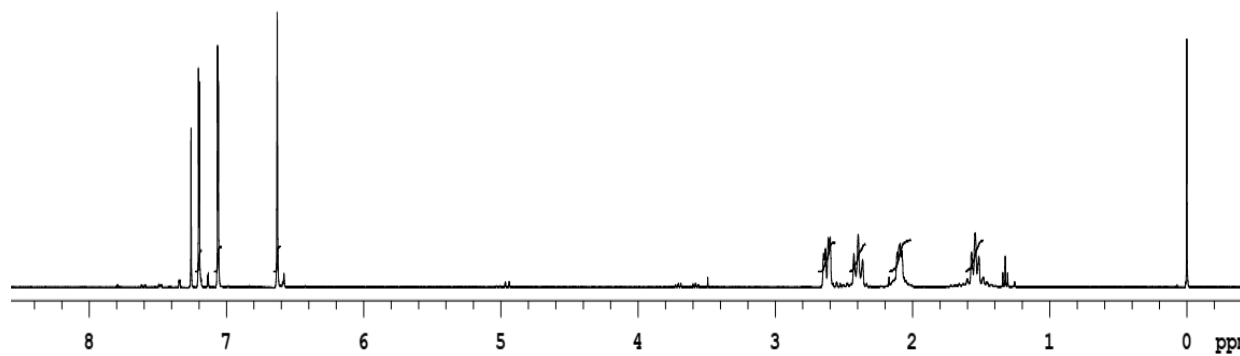
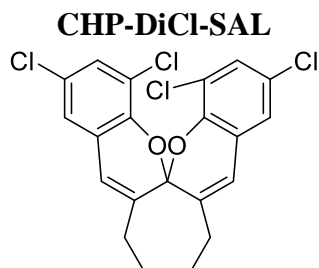
^{13}C nmr spectra



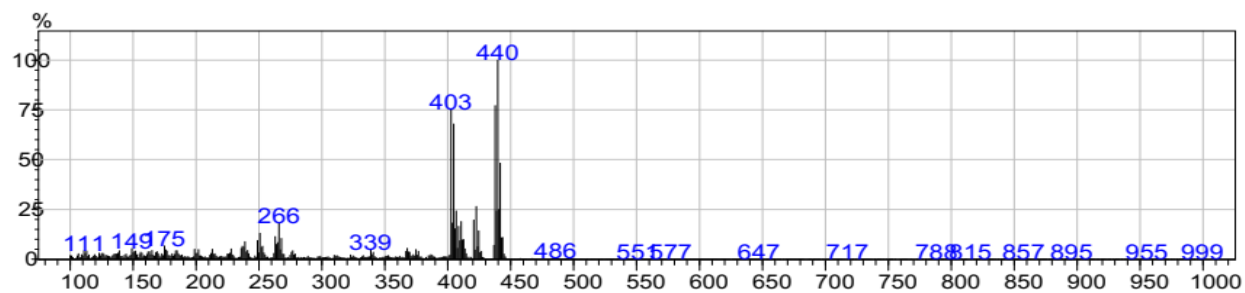
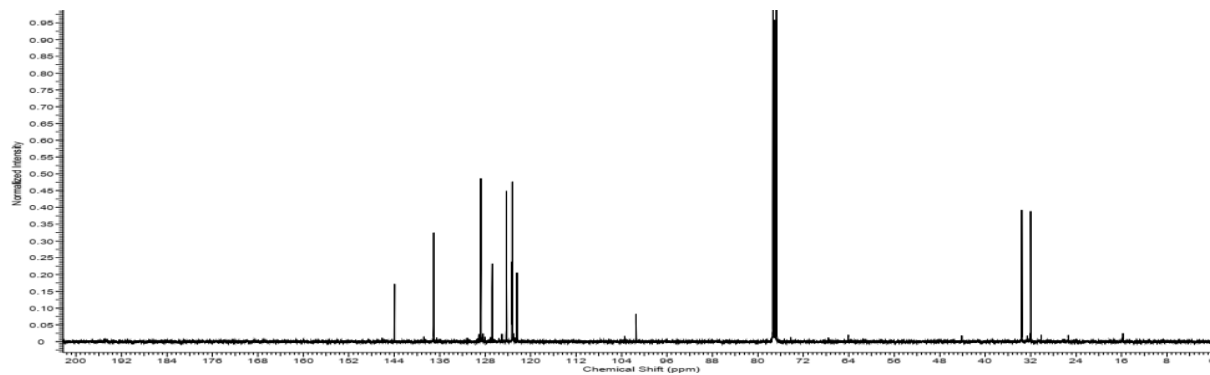
DEPT-135 ^{13}C nmr spectra



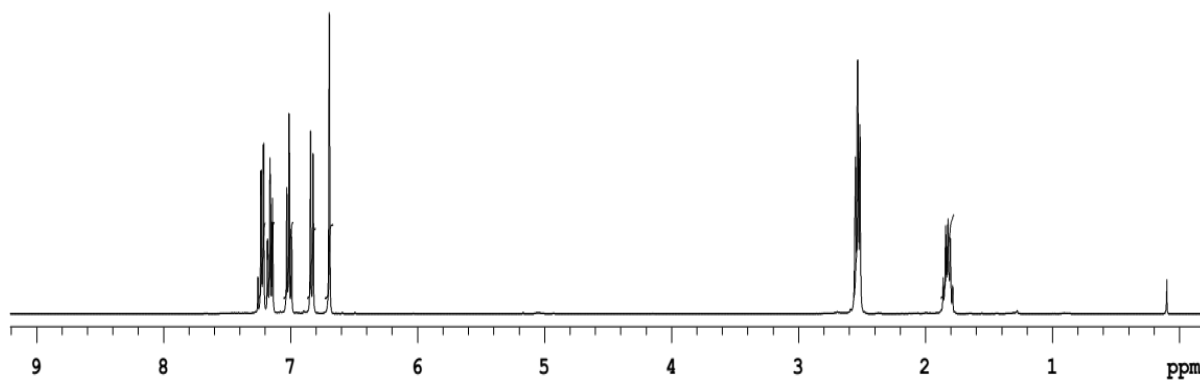
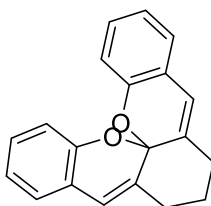
Mass spectrum

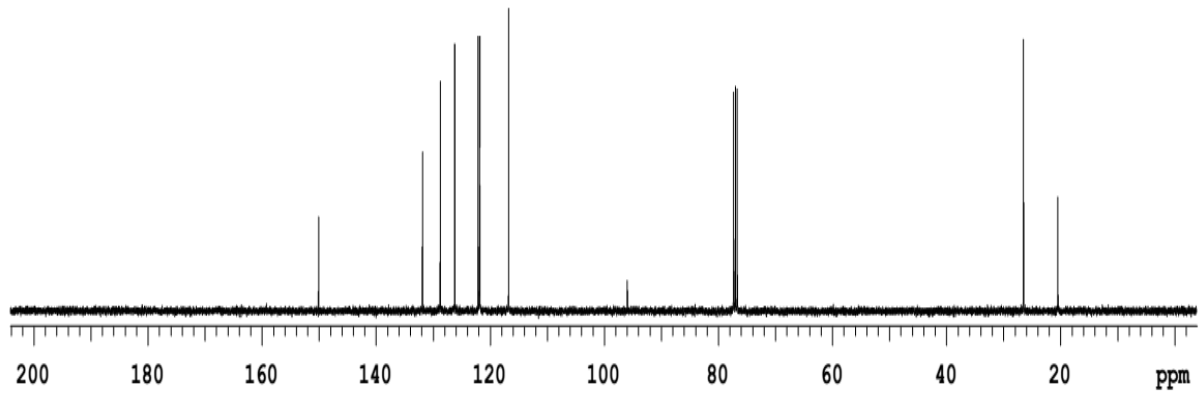


^1H nmr spectrum

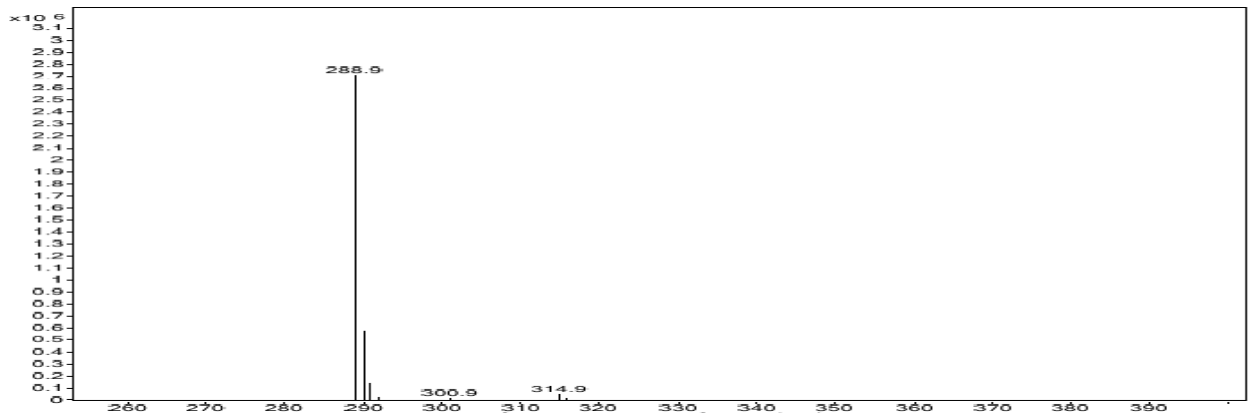


CHX-SAL



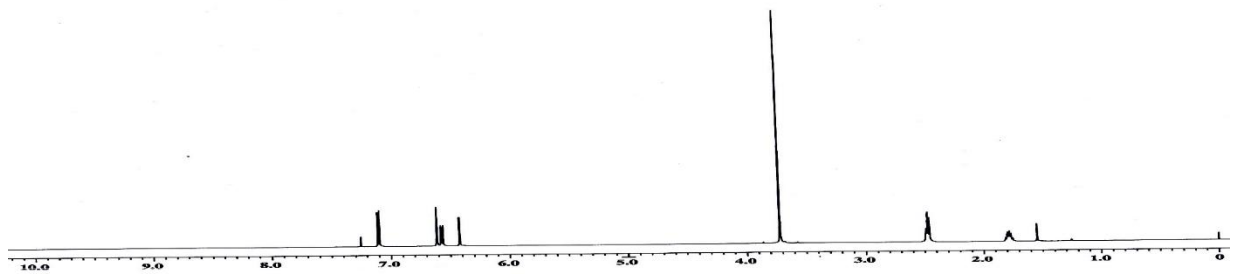
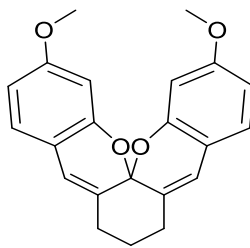


^{13}C nmr spectrum

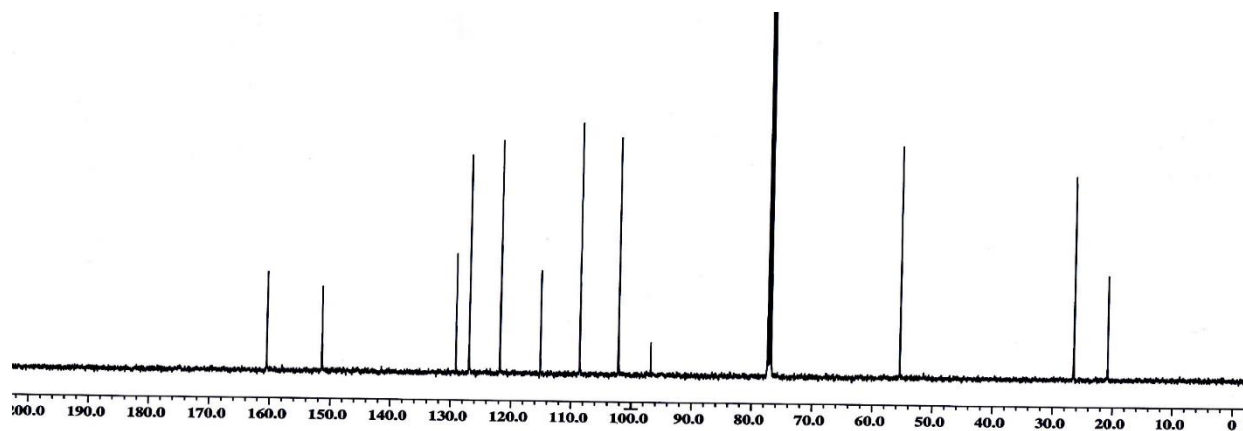


Mass spectrum

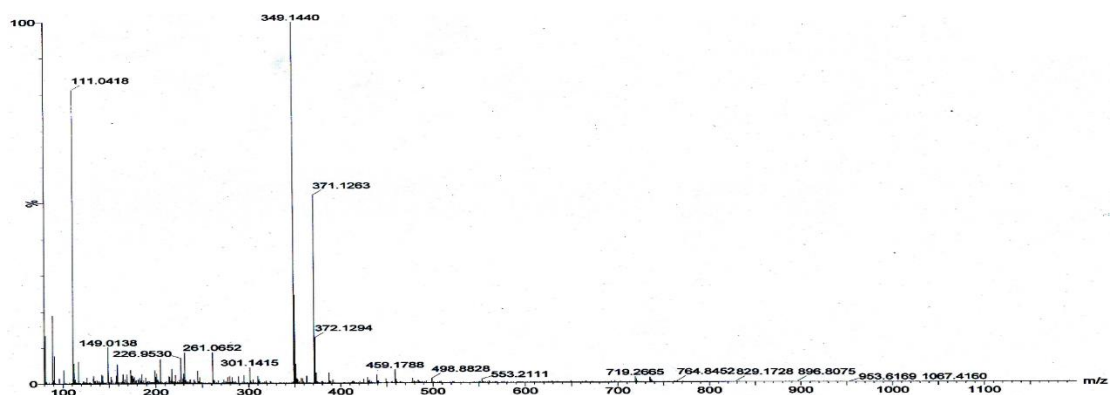
CHX-4MO-SAL



^1H nmr spectrum

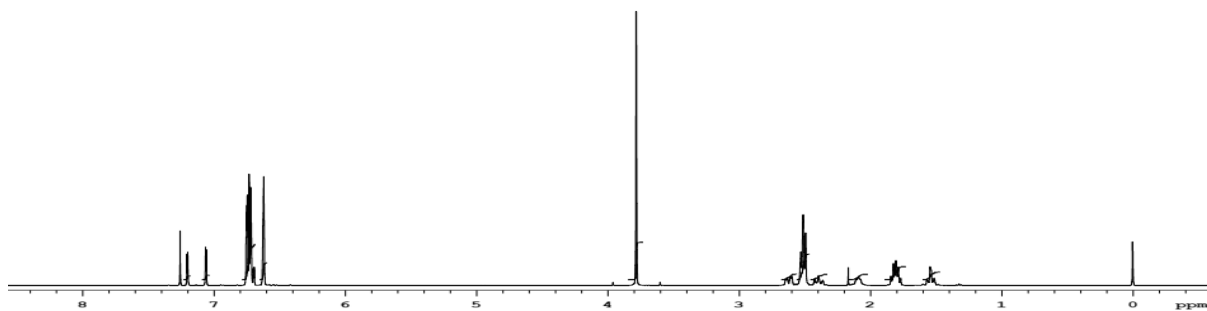
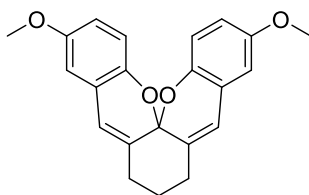


^{13}C nmr spectrum

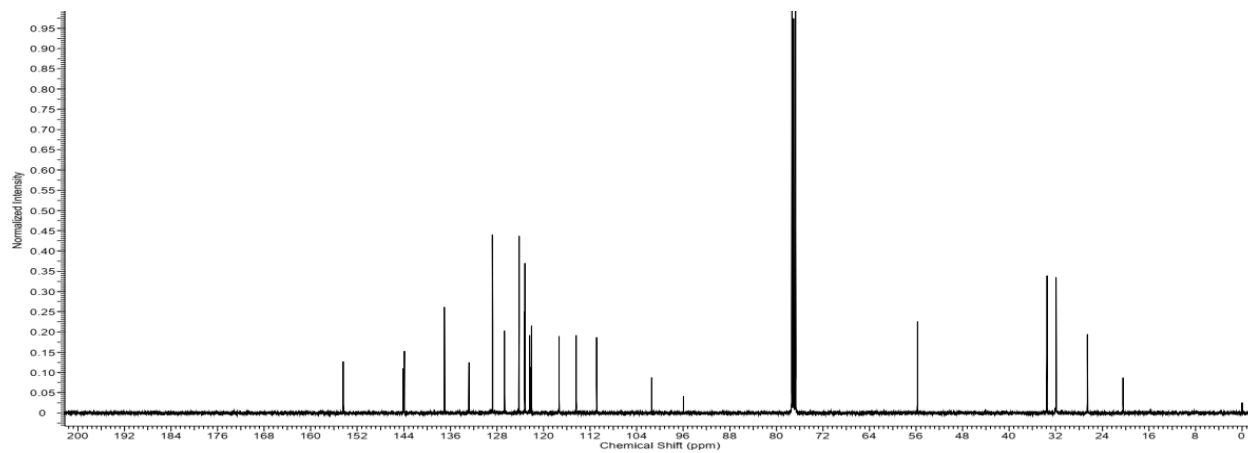


Mass spectra

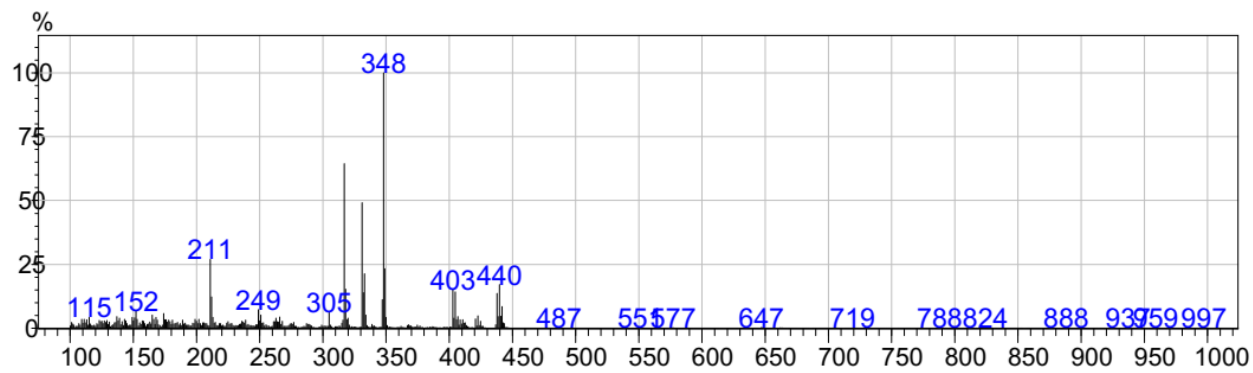
CHX-5MO-SAL



^1H nmr spectrum

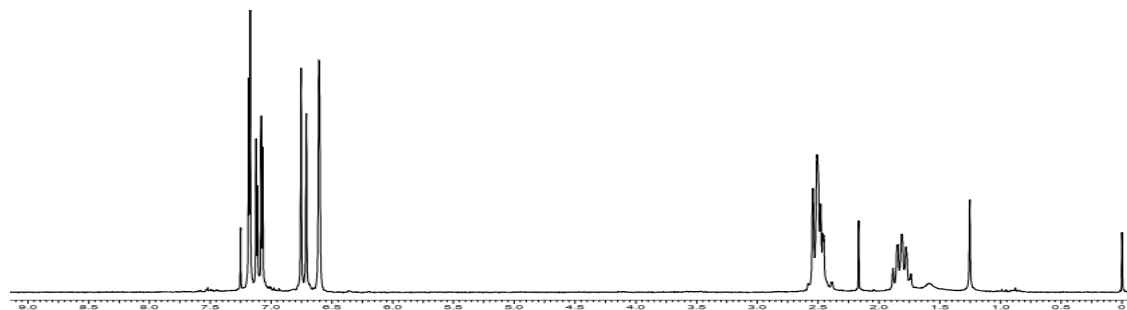
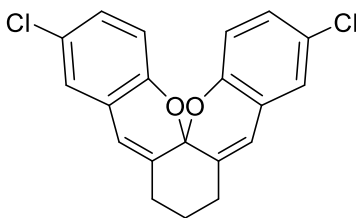


^{13}C nmr spectrum

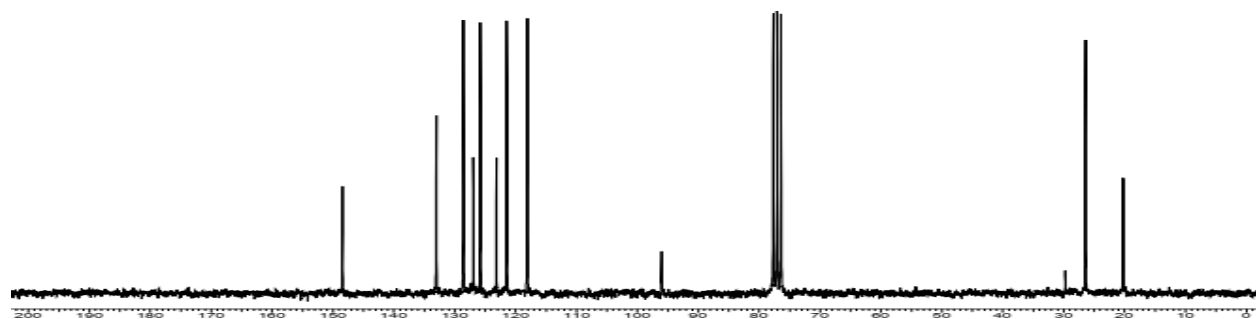


Mass spectrum

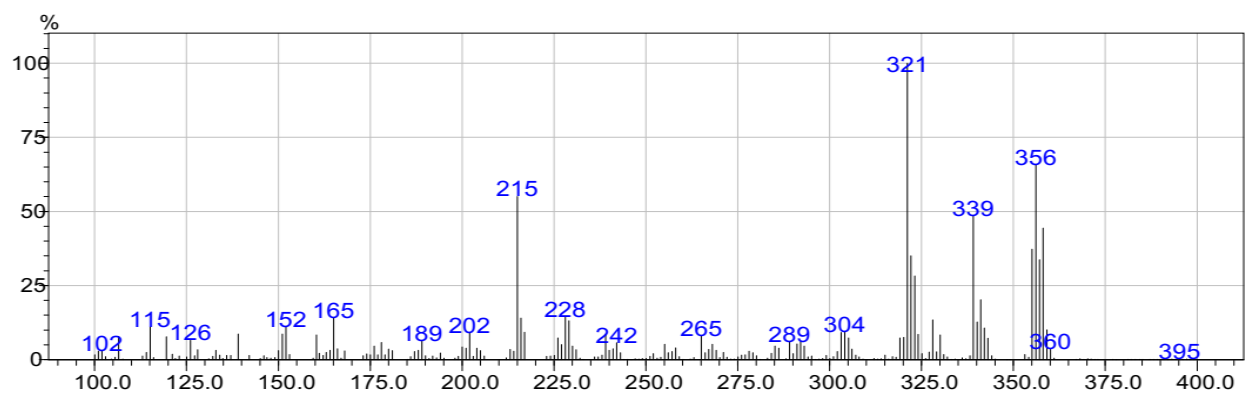
CHX-5Cl-SAL



^1H nmr spectrum

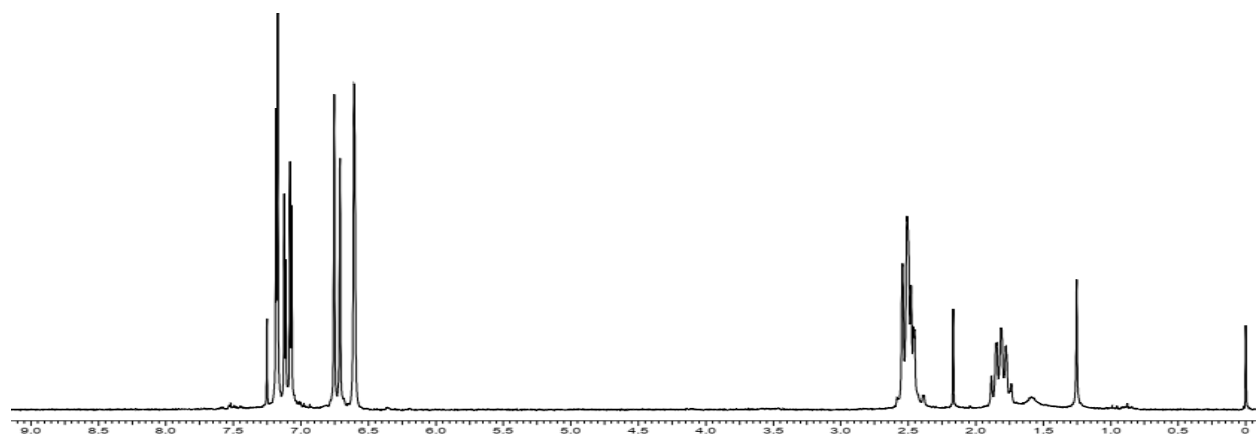
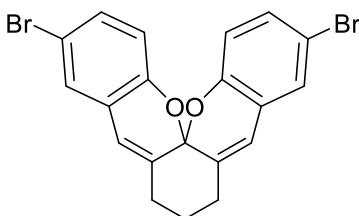


^{13}C nmr spectrum

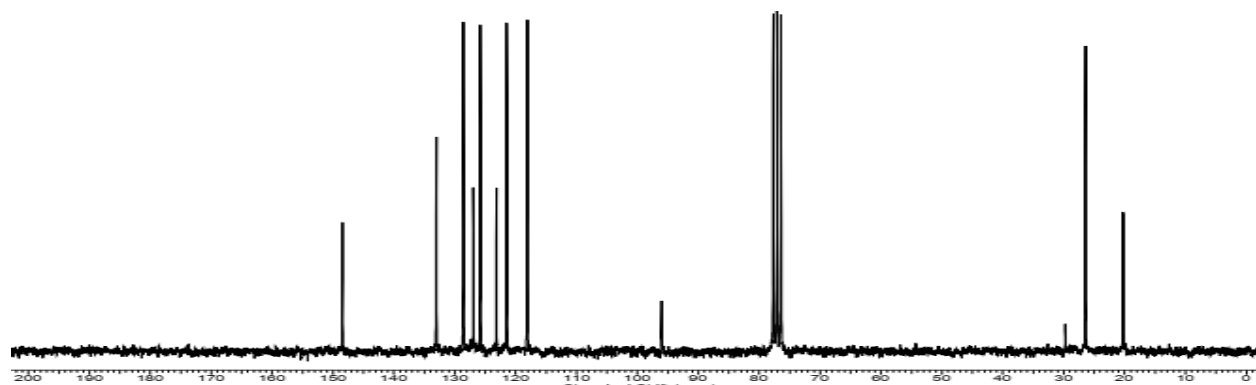


Mass spectrum

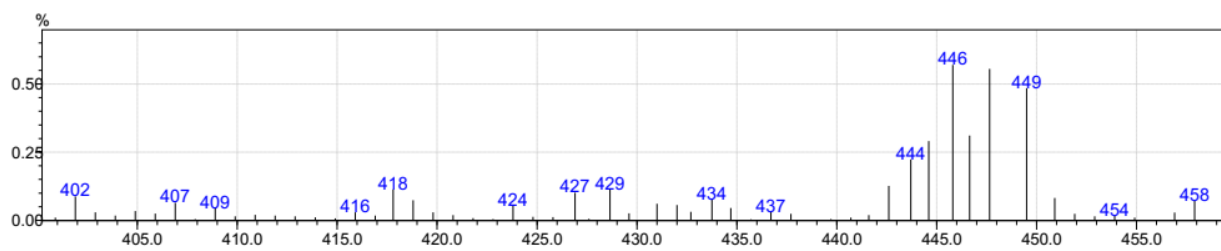
CHX-5Br-SAL



^1H nmr spectrum

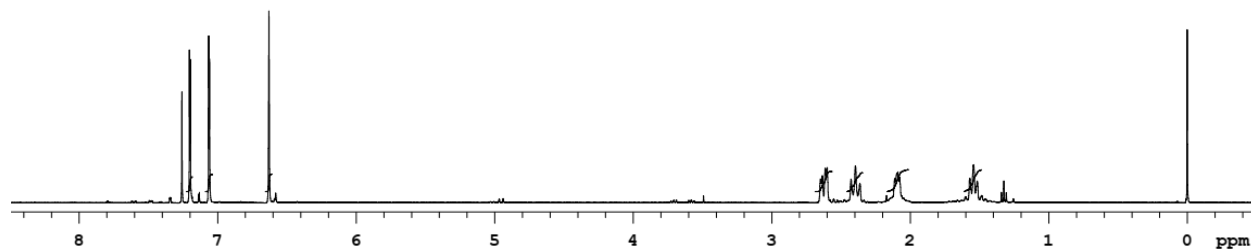
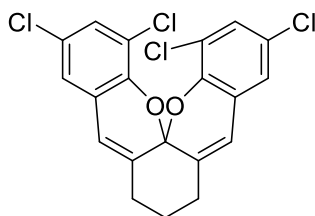


^{13}C nmr spectrum

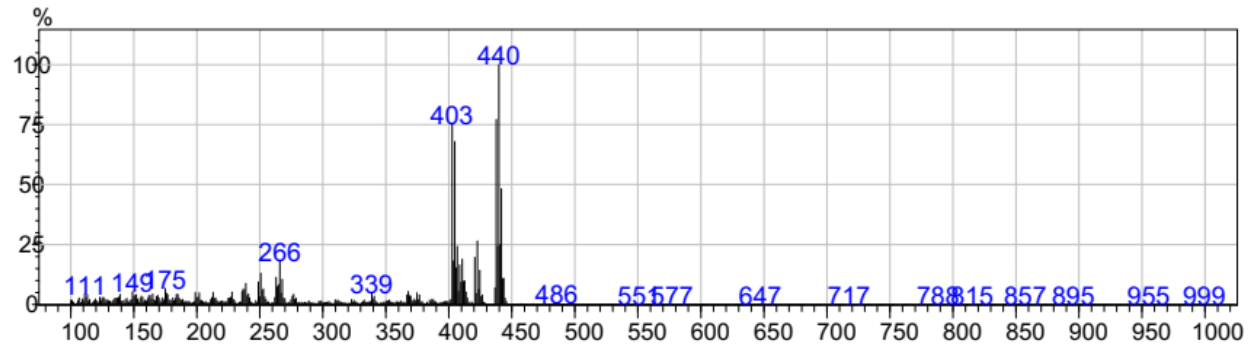


Mass spectrum

CHX-DiCl-SAL



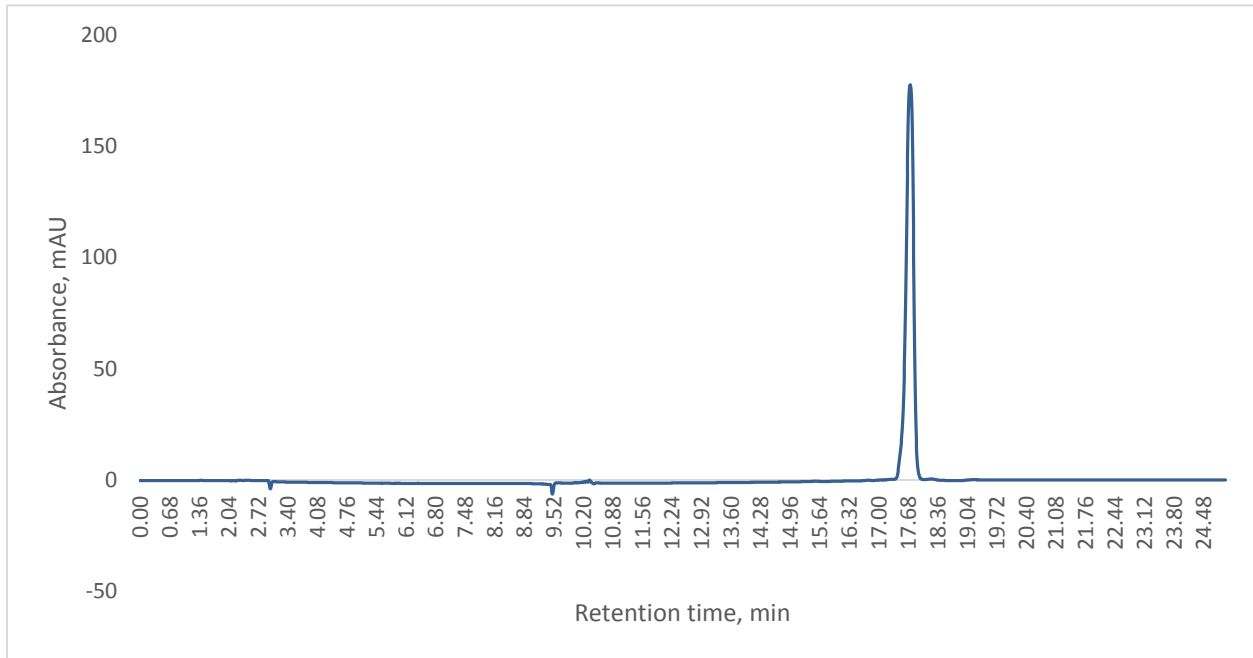
^1H nmr spectrum



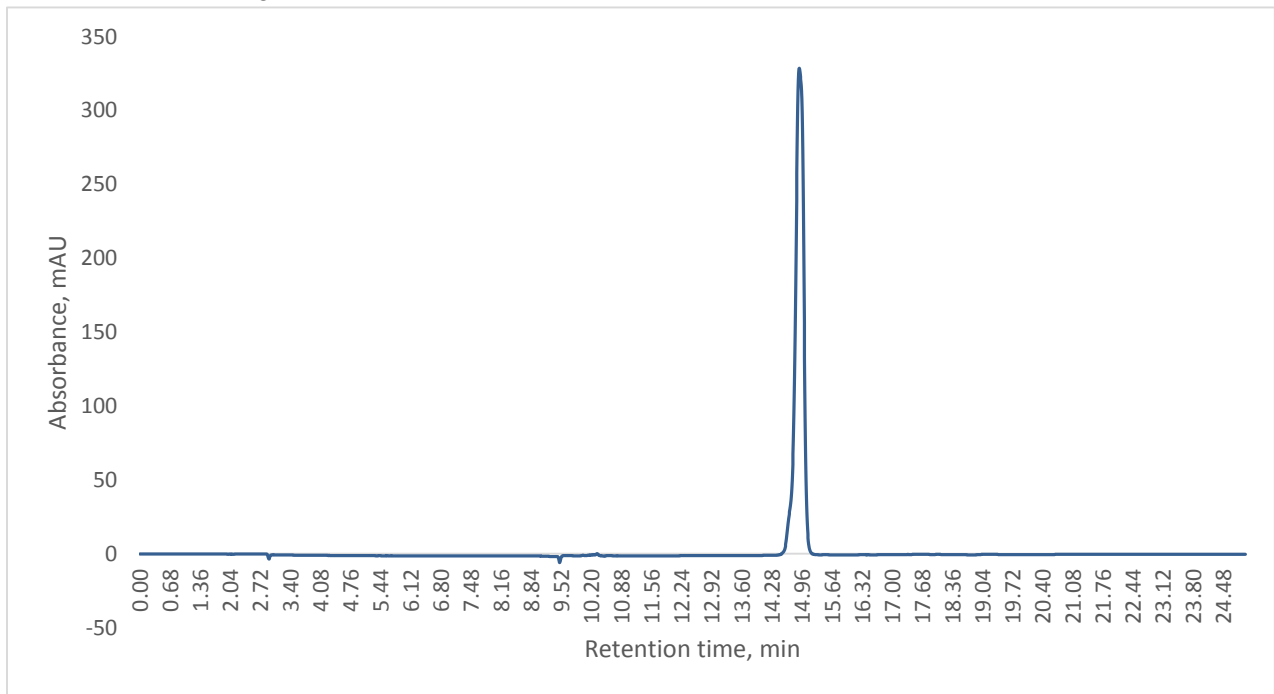
Mass Spectrum

HPLC DATA

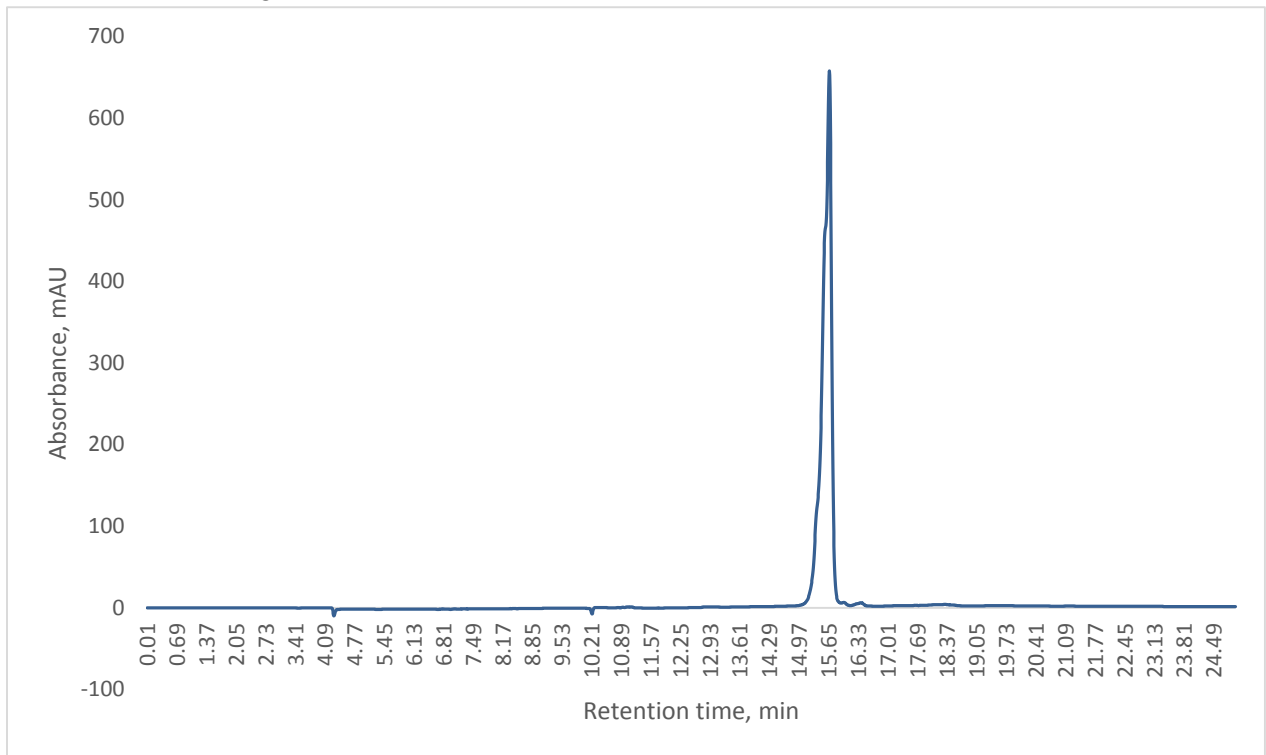
CHP-5Br-SAL, Signal at λ_{\max} nm = 308



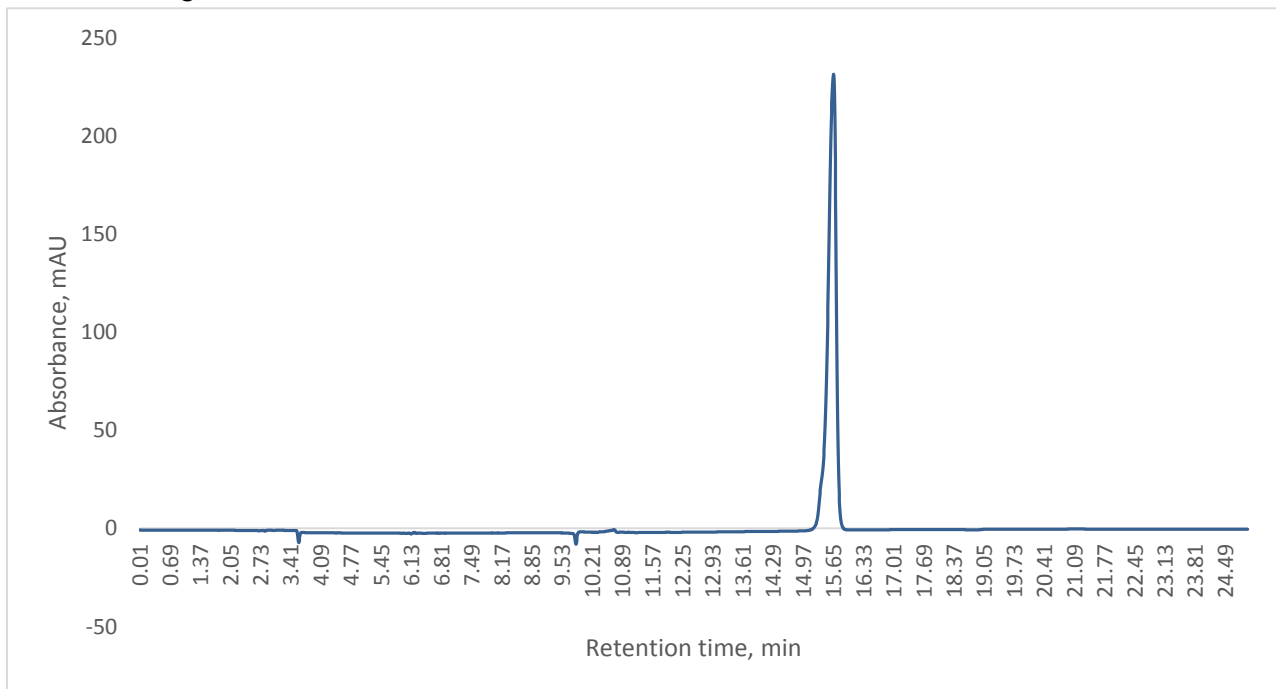
CHP-5MO-SAL, Signal at λ_{\max} nm = 321



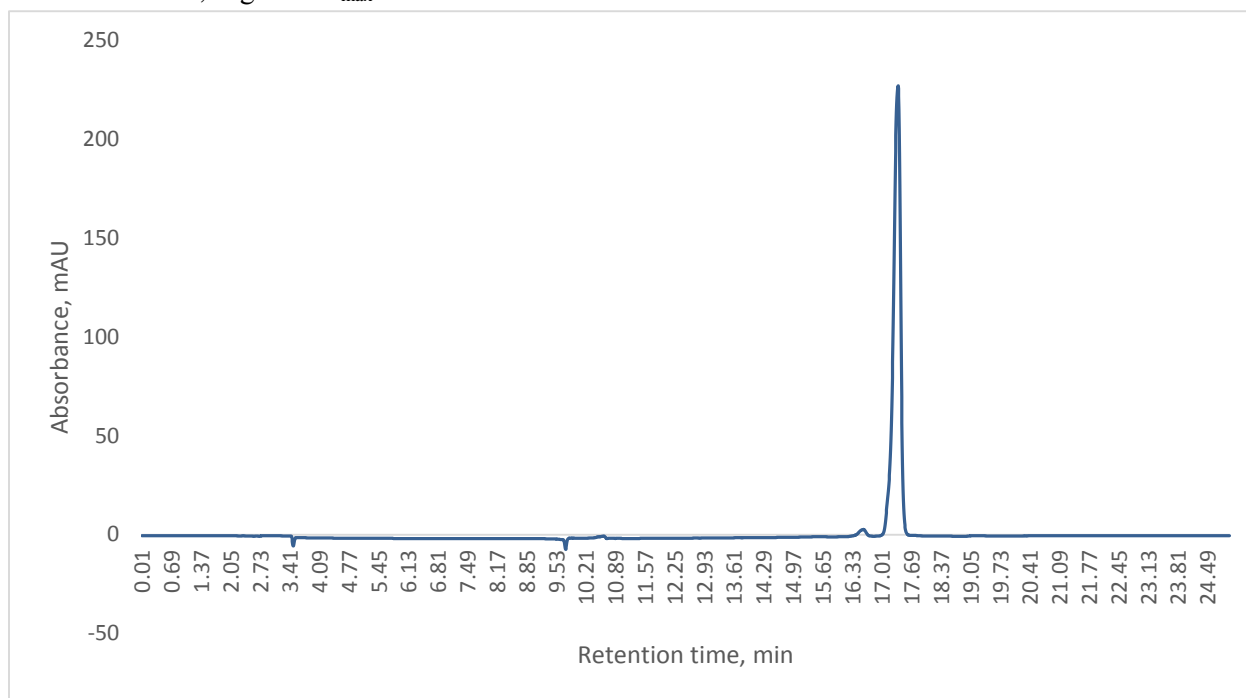
CHP-4MO-SAL, Signal at λ_{\max} nm = 278



CHP-SAL, Signal at λ_{\max} nm = 298



CHP-5Cl-SAL, Signal at λ_{\max} nm = 298



Sample	Retention time
CHP-SAL	15.65
CHP-4MO-SAL	15.67
CHP-5MO-SAL	14.91
CHP-5Cl-SAL	17.34
CHP-5Br-SAL	17.74

SD Table II. Retention time of each spirobibenzopyran in minutes

Time (min)	Solvent A (%) (Water)	Solvent B (%) (ACN)
0	100	0
15	5	95
17	100	0

SD Table III: Mobile Phase program for Gradient Elution

XRD DATA

CHP-5Cl-SAL

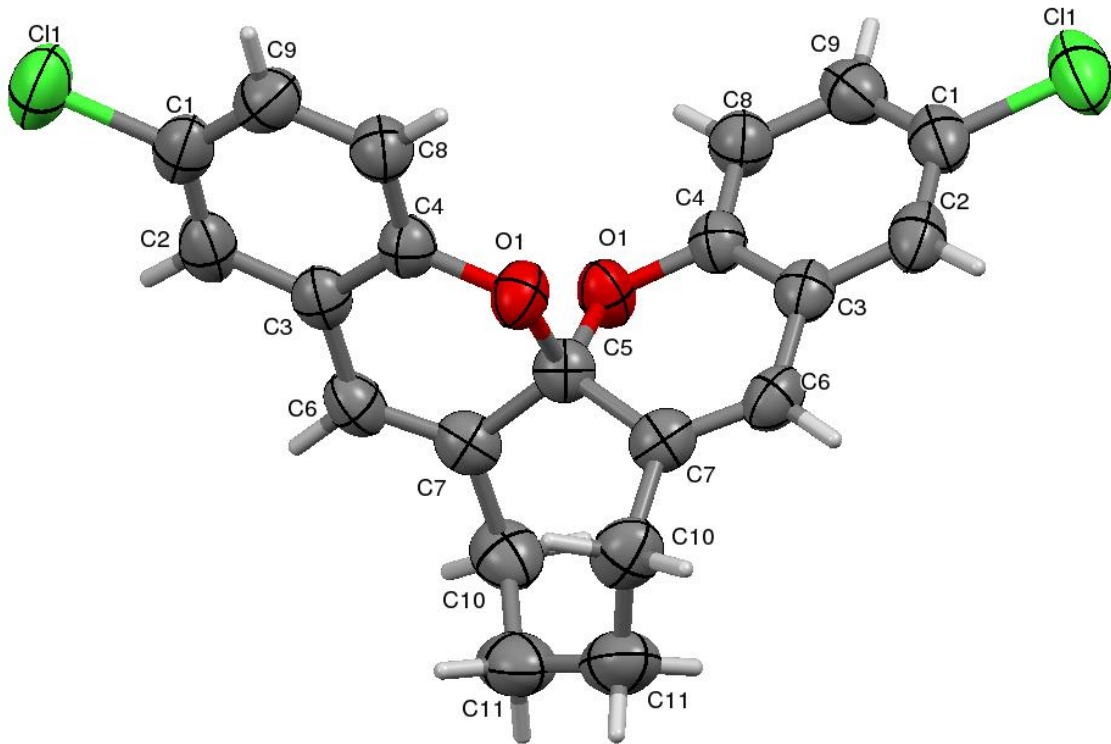


Table 1: Crystal data and structure refinement details.

CCDC Number	CCDC 1504427	
Empirical formula	$C_{21}H_{16}Cl_2O_2$	
Formula weight	185.62	
Temperature	296(2) K	
Wavelength	1.54178 Å	
Refins. for cell determination	1150	
θ range for above	4.61° to 63.68°	
Crystal system	Orthorhombic	
Space group	<i>Pbcn</i>	
Cell dimensions		
$a = 11.1071(6)$ Å	$b = 8.2385(4)$ Å	$c = 19.1755(9)$ Å
$\alpha = 90.00^\circ$	$\beta = 90.00^\circ$	$\gamma = 90.00^\circ$
Volume	1754.67(15) Å ³	
<i>Z</i>	8	
Density(calculated)	1.405 Mg m ⁻³	
Absorption coefficient	3.416 mm ⁻¹	
F_{000}	768	
Crystal size	0.29 × 0.26 × 0.23 mm	
θ range for data collection	4.61° to 63.68°	
Index ranges	-12 ≤ <i>h</i> ≤ 12	
	-9 ≤ <i>k</i> ≤ 7	
	-21 ≤ <i>l</i> ≤ 22	
Reflections collected	7530	
Independent reflections	1423 [<i>R</i> _{int} = 0.0561]	
Absorption correction	multi-scan	
Refinement method	Full matrix least-squares on <i>F</i> ²	
Data / restraints / parameters	1423 / 0 / 115	
Goodness-of-fit on <i>F</i> ²	1.121	
Final [<i>I</i> > 2σ(<i>I</i>)]	<i>R</i> 1 = 0.0729, <i>wR</i> 2 = 0.1888	
<i>R</i> indices (all data)	<i>R</i> 1 = 0.0820, <i>wR</i> 2 = 0.2022	
Extinction coefficient	0.0132(19)	
Largest diff. peak and hole	0.440 and -0.481 e Å ⁻³	

Table 2: Bond lengths (Å).

Atoms	Length	Atoms	Length
C11-C1	1.746(3)	C5-O1	1.440(3)
O1-C4	1.374(3)	C5-C7	1.505(3)
O1-C5	1.440(3)	C5-C7	1.505(3)
C1-C9	1.360(5)	C6-C7	1.327(4)
C1-C2	1.372(5)	C7-C10	1.509(4)
C2-C3	1.397(4)	C8-C9	1.388(4)
C3-C4	1.390(4)	C10-C11	1.529(5)
C3-C6	1.448(4)	C11-C11	1.510(7)
C4-C8	1.377(4)		

Table 3: Bond angles ($^{\circ}$).

Atoms	Angle	Atoms	Angle
C4-O1-C5	118.88(16)	O1-C5-C7	103.38(13)
C9-C1-C2	121.8(3)	O1-C5-C7	103.38(13)
C9-C1-C11	119.2(3)	O1-C5-C7	113.19(13)
C2-C1-C11	119.0(3)	C7-C5-C7	116.2(3)
C1-C2-C3	119.7(3)	C7-C6-C3	122.0(3)
C4-C3-C2	118.0(3)	C6-C7-C5	119.2(2)
C4-C3-C6	117.5(2)	C6-C7-C10	123.8(3)
C2-C3-C6	124.4(3)	C5-C7-C10	116.9(2)
O1-C4-C8	117.1(2)	C4-C8-C9	119.2(3)
O1-C4-C3	121.2(2)	C1-C9-C8	119.6(3)
C8-C4-C3	121.6(2)	C7-C10-C11	113.1(3)
O1-C5-O1	107.5(3)	C11-C11-C10	115.2(2)
O1-C5-C7	113.19(13)		

CHP-5MO-SAL

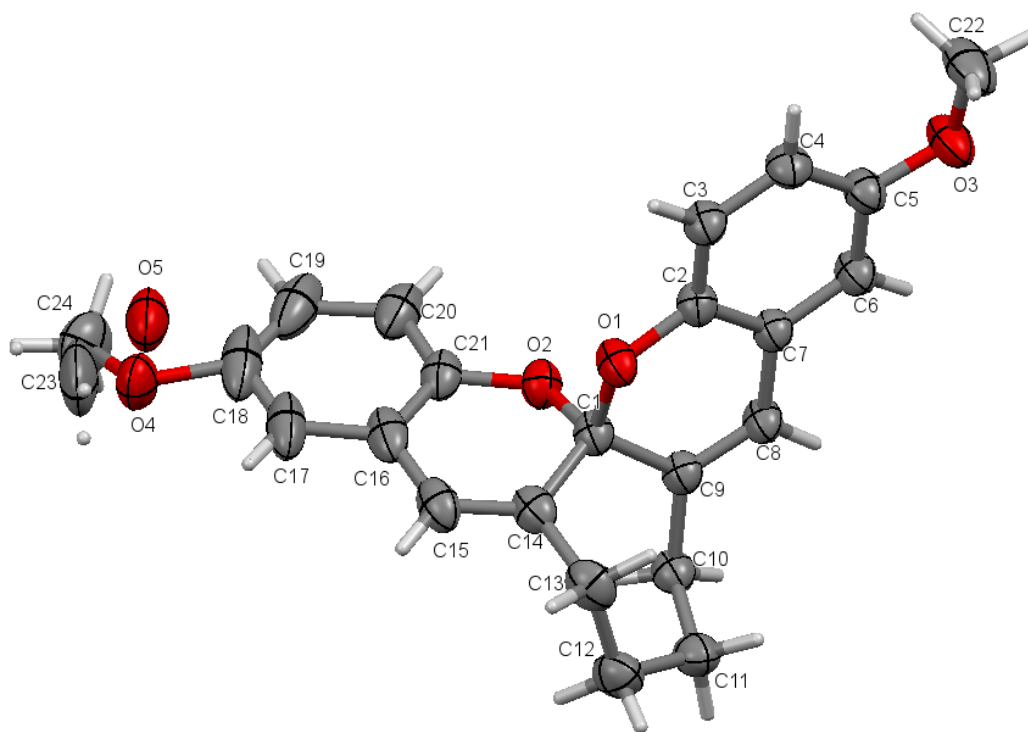


Table 1: Crystal data and structure refinement details.

CCDC Number	CCDC 1504532		
Empirical formula	$C_{23}H_{22}O_4$		
Formula weight	362.41		
Temperature	296(2) K		
Wavelength	1.54178 Å		
Refins. for cell determination	1926		
θ range for above	5.76° to 64.30°		
Crystal system	Monoclinic		
Space group	Cc		
Cell dimensions			
$a = 8.029(2)$ Å	$b = 21.258(5)$ Å	$c = 11.349(3)$ Å	
$\alpha = 90.00^\circ$	$\beta = 101.660(19)^\circ$	$\gamma = 90.00^\circ$	
Volume	1897.0(9) Å ³		
Z	4		
Density(calculated)	1.269 Mg m ⁻³		
Absorption coefficient	0.695 mm ⁻¹		
F_{000}	768		
Crystal size	0.29 × 0.28 × 0.26 mm		
θ range for data collection	5.76° to 64.30°		
Index ranges	$-9 \leq h \leq 7$ $-22 \leq k \leq 24$ $-13 \leq l \leq 13$		
Reflections collected	6231		
Independent reflections	2376 [$R_{\text{int}} = 0.0453$]		
Absorption correction	multi-scan		
Refinement method	Full matrix least-squares on F^2		
Data / restraints / parameters	2376 / 8 / 266		
Goodness-of-fit on F^2	1.049		
Final [$I > 2\sigma(I)$]	$R1 = 0.0588$, $wR2 = 0.1552$		
R indices (all data)	$R1 = 0.0712$, $wR2 = 0.1659$		
Largest diff. peak and hole	0.204 and $-0.208 e \text{ \AA}^{-3}$		

Table 2: Bond lengths (Å).

Atoms	Length	Atoms	Length
O2-C21	1.380(4)	C21-C16	1.408(7)
O2-C1	1.436(4)	C14-C15	1.319(6)
O1-C2	1.376(4)	C14-C13	1.507(6)
O1-C1	1.433(4)	C3-C4	1.390(5)
O3-C5	1.371(4)	C11-C12	1.518(6)
O3-C22	1.383(5)	C11-C10	1.525(6)
C8-C9	1.314(4)	C20-C19	1.413(8)
C8-C7	1.446(5)	C15-C16	1.439(7)
C7-C6	1.382(5)	C16-C17	1.396(6)
C7-C2	1.395(4)	C12-C13	1.541(6)
C5-C6	1.380(5)	C17-C18	1.368(11)
C5-C4	1.386(5)	C18-C19	1.393(12)
C2-C3	1.374(5)	C18-O5	1.459(13)
C9-C1	1.504(5)	C18-O4	1.482(12)
C9-C10	1.512(5)	O4-C23	1.42(2)
C1-C14	1.518(5)	O5-C24	1.29(4)
C21-C20	1.381(7)		

Table 3: Bond angles (°).

Atoms	Angle	Atoms	Angle
C21-O2-C1	120.0(3)	C20-C21-C16	123.3(4)
C2-O1-C1	119.4(2)	C15-C14-C13	123.7(4)
C5-O3-C22	118.3(3)	C15-C14-C1	119.7(4)
C9-C8-C7	122.4(3)	C13-C14-C1	116.6(3)
C6-C7-C2	119.1(3)	C2-C3-C4	119.7(3)
C6-C7-C8	123.6(3)	C5-C4-C3	120.4(3)
C2-C7-C8	117.1(3)	C12-C11-C10	114.5(3)
O3-C5-C6	115.7(3)	C21-C20-C19	116.9(6)
O3-C5-C4	125.1(3)	C9-C10-C11	112.4(3)
C6-C5-C4	119.2(3)	C14-C15-C16	121.6(4)
C5-C6-C7	121.0(3)	C17-C16-C21	117.6(5)
C3-C2-O1	118.1(2)	C17-C16-C15	123.2(5)
C3-C2-C7	120.4(3)	C21-C16-C15	119.2(3)
O1-C2-C7	121.2(3)	C11-C12-C13	114.1(3)
C8-C9-C1	119.6(3)	C14-C13-C12	113.2(3)
C8-C9-C10	123.1(3)	C18-C17-C16	120.7(7)
C1-C9-C10	117.3(3)	C17-C18-C19	120.9(5)
O1-C1-O2	108.0(3)	C17-C18-O5	142.5(10)
O1-C1-C9	113.1(3)	C19-C18-O5	96.2(10)
O2-C1-C9	103.7(2)	C17-C18-O4	101.6(8)
O1-C1-C14	104.1(2)	C19-C18-O4	137.5(8)
O2-C1-C14	114.1(3)	O5-C18-O4	42.0(5)
C9-C1-C14	114.1(3)	C18-C19-C20	120.6(7)
O2-C21-C20	116.4(4)	C23-O4-C18	105.7(13)
O2-C21-C16	120.1(4)	C24-O5-C18	109(2)

4. *In-vitro* anti-cancer activity protocol

The human tumor cell lines of the cancer screening panel were grown in RPMI 1640 medium containing 5% fetal bovine serum and 2 mM L-glutamine. For a typical screening experiment, cells were inoculated into 96 well microtiter plates in 100 μ L at plating densities ranging from 5,000 to 40,000 cells/well depending on the doubling time of individual cell lines. After cell inoculation, the microtiter plates were incubated at 37° C, 5 % CO₂, 95 % air and 100 % relative humidity for 24 h prior to addition of experimental drugs. After 24 h, two plates of each cell line were fixed *in situ* with TCA, to represent a measurement of the cell population for each cell line at the time of drug addition (Tz). Experimental drugs were solubilized in dimethyl sulfoxide at 400-fold the desired final maximum test concentration and stored frozen prior to use. At the time of drug addition, an aliquot of frozen concentrate was thawed and diluted to twice the desired final maximum test concentration with complete medium containing 50 μ g/ml gentamicin. Additional four, 10-fold or ½ log serial dilutions were made to provide a total of five drug concentrations plus control. Aliquots of 100 μ l of these different drug dilutions were added to the appropriate microtiter wells already containing 100 μ l of medium, resulting in the required final drug concentrations. Following drug addition, the plates were incubated for an additional 48 h at 37°C, 5 % CO₂, 95 % air, and 100 % relative humidity. For adherent cells, the assay was terminated by the addition of cold TCA. Cells were fixed *in situ* by the gentle addition of 50 μ l of cold 50 % (w/v) TCA (final concentration, 10 % TCA) and incubated for 60 minutes at 4°C. The supernatant was discarded, and the plates were washed five times with tap water and air dried. Sulforhodamine B (SRB) solution (100 μ l) at 0.4 % (w/v) in 1 % acetic acid was added to each well, and plates were incubated for 10 minutes at room temperature. After staining, unbound dye was removed by washing five times with 1 % acetic acid and the plates were air dried. Bound stain was subsequently solubilized with 10 mM trizma base, and the absorbance was read on an automated plate reader at a wavelength of 515 nm. For suspension cells, the methodology was the same except that the assay was terminated by fixing settled cells at the bottom of the wells by gently adding 50 μ l of 80 % TCA (final concentration, 16 % TCA). Using the seven absorbance measurements [time zero, (Tz), control growth, (C), and test growth in the presence of drug at the five concentration levels (Ti)], the percentage growth was calculated at each of the drug concentrations levels [1, 2].

Percentage growth inhibition was calculated as:

$$[(Ti-Tz)/(C-Tz)] \times 100 \text{ for concentrations for which } Ti \geq Tz$$

$$[(Ti-Tz)/Tz] \times 100 \text{ for concentrations for which } Ti < Tz.$$

Results for each compound (One dose data) are reported as mean graph of the percent growth of the treated cells when compared with untreated control cells [1, 2]. The One-dose data (given below) is reported as a mean graph of the percent growth of treated cells. The number reported for the One-dose assay is growth relative to the no-drug control, and relative to the time zero number of cells. This allows detection of both growth inhibition (values between 0 and 100) and lethality

(values less than 0). For example, a value of 100 means no growth inhibition. A value of 40 would mean 60% growth inhibition. A value of 0 means no net growth over the course of the experiment. A value of -40 would mean 40% lethality. A value of -100 means all cells are dead.

SD Table IV. The *in vitro* anticancer analysis results for the CHP series

CANCER CELL	SUB PANEL	Growth Percentage				
		CHP-SAL	CHP-4MO-SAL	CHP-5MO-SAL	CHP-5CI-SAL	CHP-DiCI-SAL
Leukemia	K-562	<u>58.35</u>	88.68	94.10	103.71	85.98
Non-Small Cell Lung Cancer	NCI-H460	<u>66.95</u>	94.39	113.07	105.60	105.82
	NCI-H522	82.42	<u>66.21</u>	96.24	76.64	95.79
Colon Cancer	HCT-116	<u>53.13</u>	73.44	100.36	97.30	78.22
	HCT-15	<u>45.96</u>	92.62	104.58	104.14	92.22
CNS Cancer	U251	<u>68.50</u>	89.65	103.20	93.56	96.84
Prostate Cancer	PC-3	<u>56.97</u>	69.59	102.01	84.41	92.15
Breast Cancer	T-47D	91.92	<u>67.62</u>	109.60	102.04	83.58

The results of the single dose study for the 5 selected spirobibenzopyrans from the CHX series have been tabulated below in SD table V.

CANCER CELL	SUB PANEL	Growth Percentage				
		CHX-SAL	CHX-4MO-SAL	CHX-5MO-SAL	CHX-5CI-SAL	CHX-DiCI-SAL
Leukemia	CCRF-CEM	<u>18.28</u>	<u>5.97</u>	88.31	96.34	<u>45.36</u>
	HL-60(TB)	<u>8.47</u>	<u>7.56</u>	99.59	94.36	87.67
	K-562	<u>15.32</u>	<u>23.50</u>	94.85	87.10	75.88
	MOLT-4	<u>17.51</u>	<u>-3.56</u>	89.70	84.23	80.56
	RPMI-8226	<u>26.01</u>	<u>-20.58</u>	93.55	107.15	79.43
	SR	<u>6.44</u>	<u>4.94</u>	97.09	100.20	<u>63.19</u>
Non-Small Cell Lung Cancer	A549/ATCC	<u>32.65</u>	75.02	96.33	88.36	100.56
	EKVX	<u>57.76</u>	<u>39.65</u>	103.01	104.23	89.44
	HOP-62	<u>41.57</u>	<u>41.40</u>	101.21	105.83	104.91
	HOP-92	<u>61.42</u>	<u>26.64</u>	105.10	90.98	<u>55.63</u>
	NCI-H226	<u>56.44</u>	<u>64.09</u>	97.30	99.10	97.10
	NCI-H23	<u>42.30</u>	<u>48.27</u>	97.16	98.66	105.59
	NCI-H322M	<u>50.79</u>	80.84	106.32	108.23	87.65
	NCI-H460	<u>13.47</u>	<u>14.25</u>	105.25	103.73	108.43
Colon Cancer	NCI-H522	<u>3.90</u>	<u>20.45</u>	101.58	76.84	102.48
	COLO 205	<u>25.44</u>	<u>-9.95</u>	111.27	104.18	115.35
	HCC-2998	<u>58.40</u>	<u>57.94</u>	102.97	103.67	100.49

	HCT-116	<u>23.99</u>	<u>19.97</u>	94.74	93.46	96.55
	HCT-15	<u>20.14</u>	<u>12.83</u>	104.56	100.79	76.56
	HT29	<u>5.93</u>	<u>8.16</u>	97.57	88.84	112.44
	KM12	<u>19.78</u>	<u>-5.69</u>	91.22	102.25	96.39
	SW-620	<u>18.11</u>	<u>16.61</u>	94.58	98.47	99.99
CNS Cancer	SF-268	<u>31.56</u>	<u>38.52</u>	97.90	100.10	77.33
	SF-295	<u>11.30</u>	<u>78.63</u>	99.15	100.22	98.49
	SF-539	<u>7.81</u>	84.49	114.62	100.50	90.77
	SNB-19	<u>27.44</u>	<u>54.71</u>	105.09	102.91	88.84
	SNB-75	<u>22.99</u>	74.56	107.33	82.64	98.36
	U251	<u>26.46</u>	<u>34.95</u>	102.25	94.84	86.99
Melanoma	LOX IMVI	<u>22.99</u>	<u>-32.06</u>	97.19	97.45	83.75
	MALME-3M	<u>56.77</u>	<u>70.32</u>	111.16	109.43	103.68
	M14	<u>19.15</u>	<u>31.51</u>	96.01	102.54	98.79
	MDA-MB-435	<u>-41.60</u>	<u>-10.37</u>	103.06	109.31	105.19
	SK-MEL-2	<u>36.05</u>	<u>15.69</u>	117.14	89.48	108.02
	SK-MEL-28	<u>54.24</u>	<u>62.98</u>	97.48	114.26	95.27
	SK-MEL-5	<u>19.96</u>	<u>40.27</u>	98.77	99.67	97.86
	UACC-257	<u>40.07</u>	<u>52.52</u>	89.70	97.04	99.51
	UACC-62	<u>14.54</u>	<u>14.82</u>	103.55	99.18	95.13
Ovarian Cancer	IGROV1	<u>44.53</u>	<u>41.44</u>	109.88	104.31	88.19
	OVCAR-3	<u>3.51</u>	<u>5.62</u>	100.27	109.66	82.04
	OVCAR-4	<u>52.59</u>	<u>55.15</u>	105.13	112.87	90.73
	OVCAR-5	71.12	<u>47.18</u>	102.00	105.05	109.16
	OVCAR-8	<u>38.51</u>	<u>47.71</u>	99.51	96.66	89.11
	NCI/ADR-RES	<u>11.57</u>	<u>21.65</u>	102.73	105.49	95.20
	SK-OV-3	<u>30.85</u>	<u>60.13</u>	105.27	103.45	107.84
Renal Cancer	786-0	<u>26.44</u>	<u>38.70</u>	97.32	97.54	102.42
	A498	<u>5.61</u>	69.76	102.61	99.78	89.89
	ACHN	<u>47.34</u>	<u>25.30</u>	95.73	110.56	96.41
	CAKI-1	<u>27.21</u>	-	94.96	-	98.36
	RXF 393	<u>9.77</u>	<u>15.96</u>	101.32	96.52	106.92
	SN12C	<u>34.38</u>	<u>44.06</u>	110.33	100.10	96.28
	TK-10	<u>55.40</u>	<u>46.97</u>	112.95	91.51	111.64
	UO-31	<u>43.31</u>	<u>36.31</u>	95.56	86.54	84.11
Prostate Cancer	PC-3	<u>34.65</u>	<u>23.55</u>	93.86	86.42	69.95
	DU-145	<u>28.07</u>	<u>33.91</u>	109.02	108.75	105.70
Breast Cancer	MCF7	<u>13.51</u>	<u>8.63</u>	94.21	94.43	84.02
	MDA-MB-231/ATCC	<u>43.53</u>	<u>32.03</u>	121.22	101.04	98.52
	HS 578T	<u>33.37</u>	<u>45.03</u>	104.52	98.47	94.21
	BT-549	<u>26.39</u>	<u>35.03</u>	115.48	108.48	98.59

	T-47D	31.86	44.67	95.98	101.93	80.58
	MDA-MB-468	25.53	56.33	81.77	106.79	90.19

SD Table V. The *in vitro* anticancer analysis results for the CHX series.

The GI50 value (growth inhibitory activity) corresponds to the concentration of the compound causing 50% decrease in net cell growth, the TGI value (cytostatic activity) is the concentration of the compound resulting in total growth inhibition and LC50 value (cytotoxic activity) is the concentration of the compound causing net 50% loss of initial cells at the end of the incubation period of 48 h.

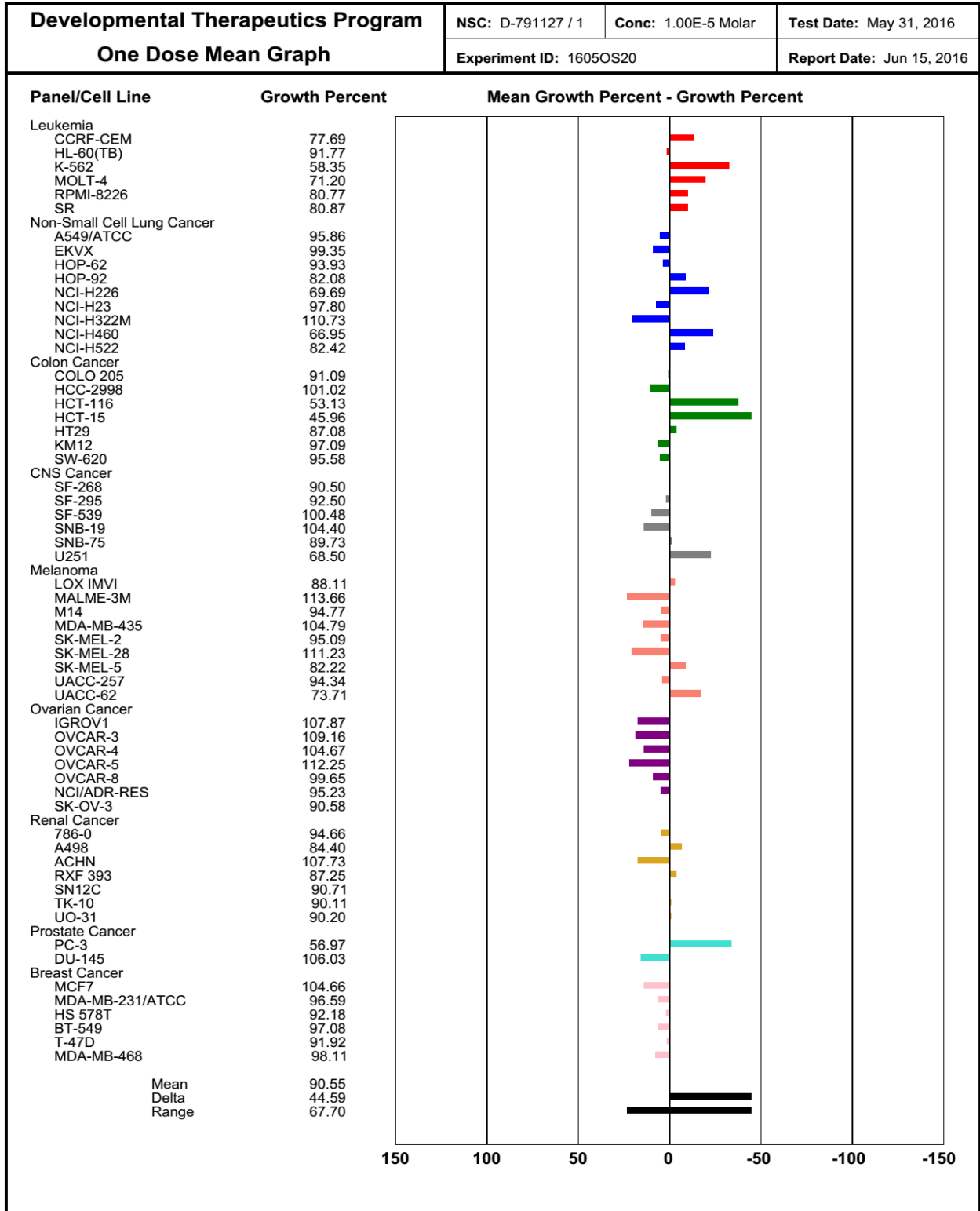
SD Table VI. The *in vitro* 5 dose analysis results for CHX-SAL and CHX-4MO-SAL.

		CHX-SAL	CHX-4MO-SAL	CHX-SAL	CHX-4MO-SAL	CHX-SAL	CHX-4MO-SAL
PANEL	CELL	GI ₅₀		TGI		LC ₅₀	
Leukemia	CCRF-CEM	3.47	5.88	25.90	>100	>100	>100
	HL-60(TB)	3.24	>100	>100	>100	>100	>100
	K-562	3.54	>100	>100	>100	>100	>100
	MOLT-4	3.65	5.81	>100	>100	>100	>100
	RPMI-8226	3.38	6.12	15.00	>100	>100	>100
	SR	2.82	4.89	>100	>100	>100	>100
Non-Small Cell Lung Cancer	A549/ATCC	5.87	>100	>100	>100	>100	>100
	EKVX	5.49	>100	>100	>100	>100	>100
	HOP-62	3.80	>100	>100	>100	>100	>100
	NCI-H226	>100	4.46	>100	>100	>100	>100
	NCI-H23	6.20	>100	>100	>100	>100	>100
	NCI-H322M	3.92	>100	>100	>100	>100	>100
	NCI-H460	3.47	>100	16.40	>100	>100	>100
	NCI-H522	2.30	>100	6.20	>100	>100	>100
Colon Cancer	COLO 205	2.24	>100	5.04	>100	>100	>100
	HCC-2998	1.03	>100	60.70	>100	>100	>100
	HCT-116	4.34	-	>100	>100	>100	>100
	HCT-15	3.68	>100	>100	>100	>100	>100
	HT29	3.52	>100	17.90	>100	>100	>100
	KM12	3.61	18.70	56.70	>100	>100	>100
	SW-620	3.35	>100	53.60	>100	>100	>100
CNS Cancer	SF-268	4.74	>100	>100	>100	>100	>100
	SF-295	3.99	>100	>100	>100	>100	>100
	SF-539	1.92	>100	5.01	>100	>100	>100
	SNB-19	4.01	>100	>100	>100	>100	>100
	SNB-75	1.64	-	5.84	>100	>100	>100
	U251	4.54	>100	>100	>100	>100	>100
Melanoma	LOX IMVI	3.88	>100	28.00	>100	>100	>100

	MALME-3M	7.17	>100	>100	>100	>100	>100
	M14	3.37	>100	>100	>100	>100	>100
	MDA-MB-435	1.64	5.52	4.67	>100	>100	>100
	SK-MEL-2	4.31	>100	25.60	>100	>100	>100
	SK-MEL-28	5.22	>100	>100	>100	>100	>100
	SK-MEL-5	3.37	13.60	24.20	>100	>100	>100
	UACC-257	5.49	>100	>100	>100	>100	>100
	UACC-62	2.91	4.50	82.50	>100	>100	>100
Ovarian Cancer	IGROV1	5.36	>100	>100	>100	>100	>100
	OVCAR-3	3.06	-	>100	>100	>100	>100
	OVCAR-4	6.12	>100	>100	>100	>100	>100
	OVCAR-5	3.46	>100	>100	>100	>100	>100
	OVCAR-8	4.04	>100	>100	>100	>100	>100
	NCI/ADR-RES	3.18	>100	55.70	>100	>100	>100
	SK-OV-3	3.52	>100	>100	>100	>100	>100
Renal Cancer	786-0	6.41	>100	>100	>100	>100	>100
	A498	2.76	>100	13.10	>100	>100	>100
	ACHN	5.06	>100	>100	>100	>100	>100
	CAKI-1	3.71	>100	>100	>100	>100	>100
	RXF 393	2.85	>100	8.77	>100	>100	>100
	SN12C	4.42	>100	>100	>100	>100	>100
	TK-10	9.87	>100	>100	>100	>100	>100
	UO-31	4.86	>100	>100	>100	>100	>100
Prostate Cancer	PC-3	4.00	>100	>100	>100	>100	>100
	DU-145	3.73	>100	69.10	>100	>100	>100
Breast Cancer	MCF7	3.03	>100	60.60	>100	>100	>100
	MDA-MB-231/ATCC	4.23	4.80	45.50	>100	>100	>100
	HS 578T	3.03	>100	-	>100	>100	>100
	BT-549	3.73	>100	73.70	>100	>100	>100
	T-47D	2.46	4.70	>100	>100	>100	>100
	MDA-MB-468	5.42	>100	69.10	>100	>100	>100

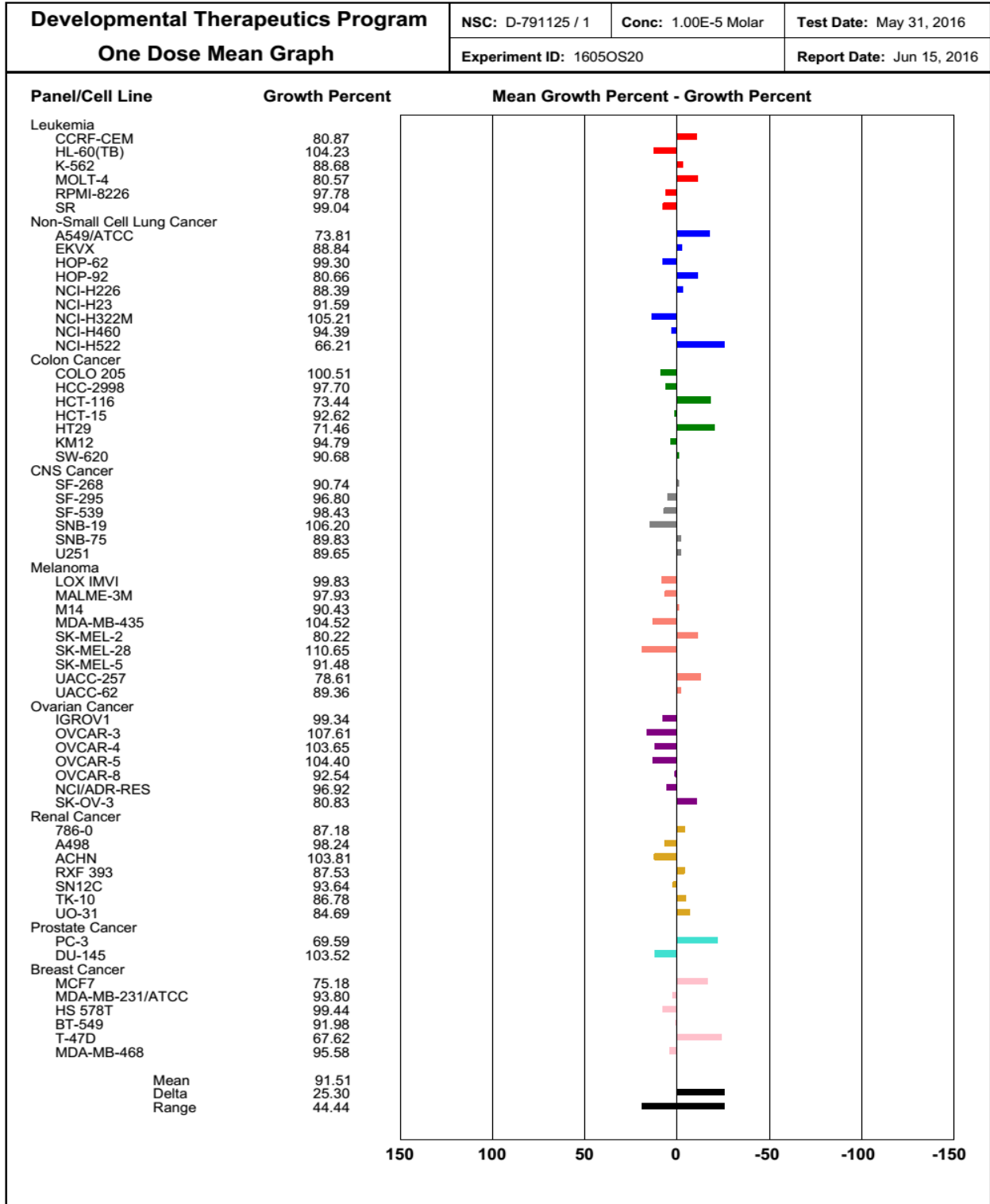
***In-vitro* anti-cancer activity at 10µM against NCI 60 cell line panel.(One dose study)**

CHP-SAL



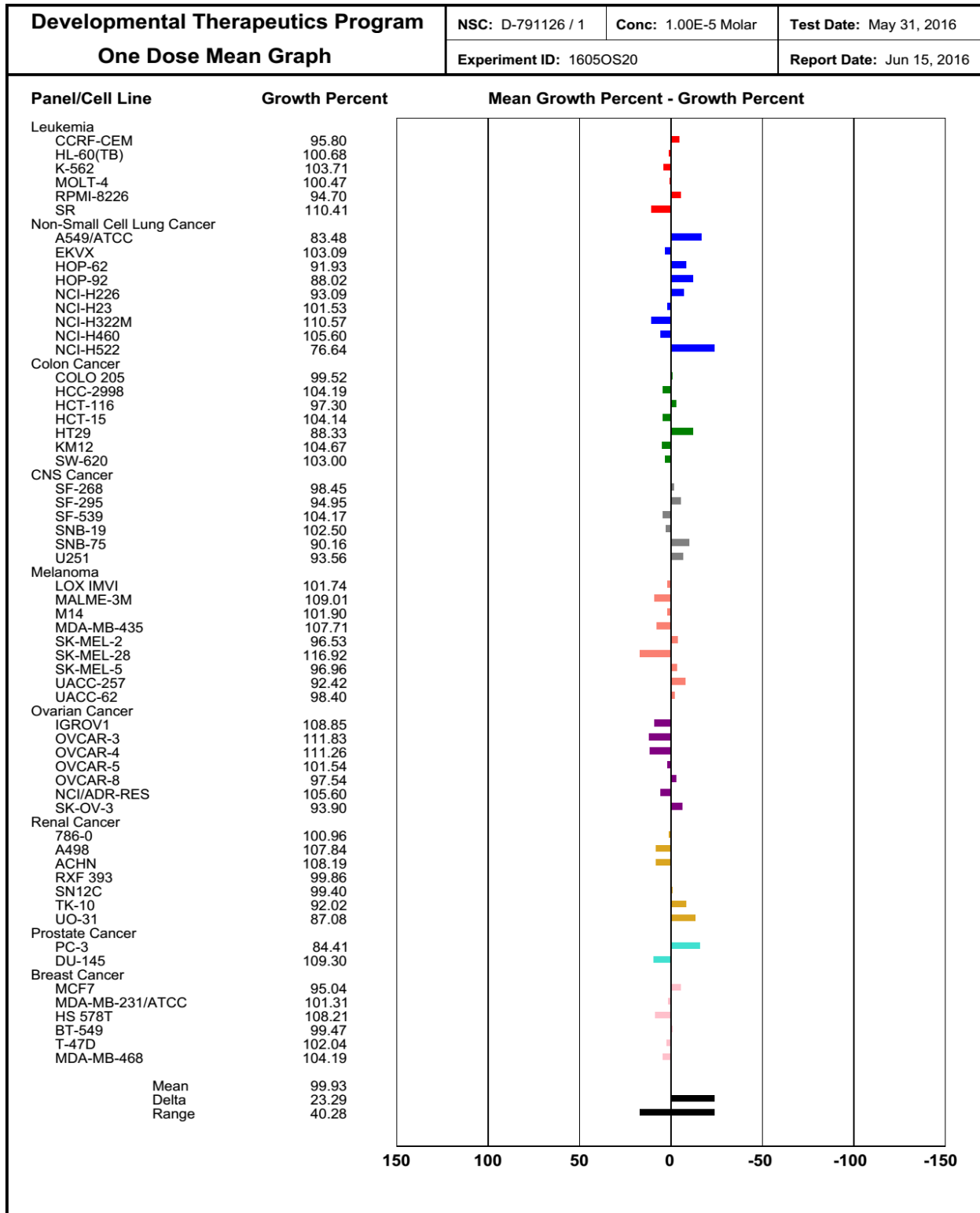
In-vitro anti-cancer activity at 10µM against NCI 60 cell line panel.

CHP-4MO-SAL



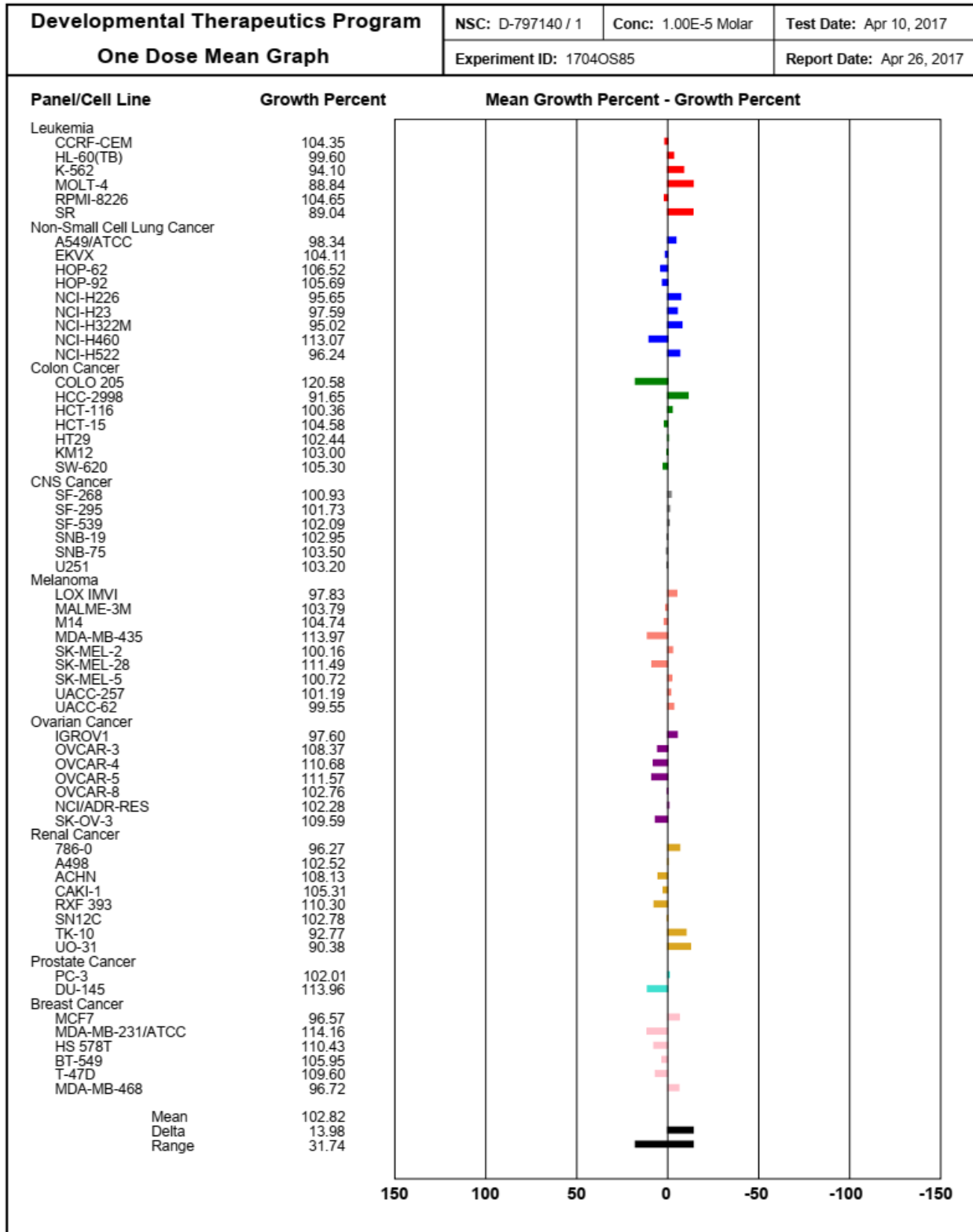
***In-vitro* anti-cancer activity at 10µM against NCI 60 cell line panel.**

CHP-5Cl-SAL



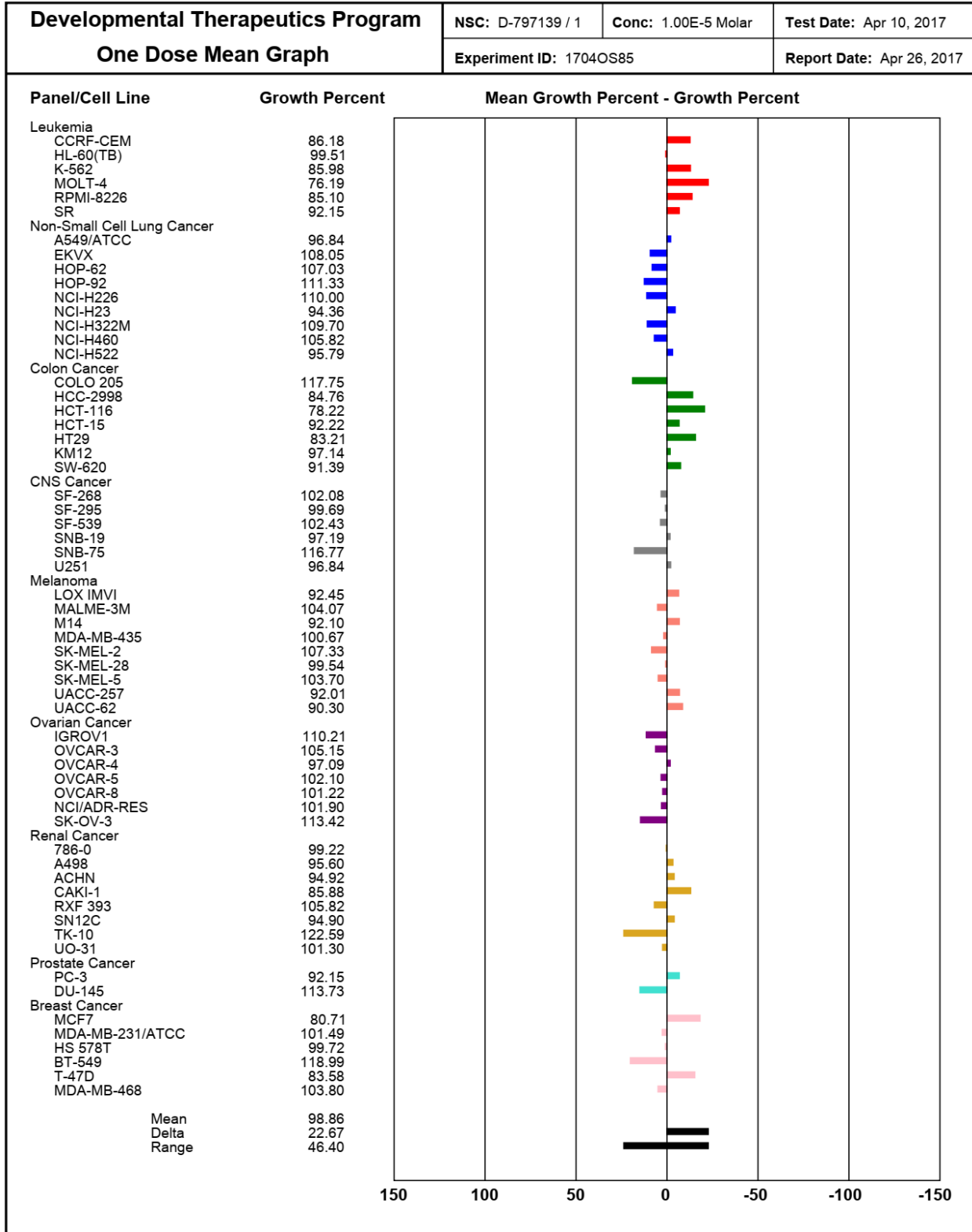
In-vitro anti-cancer activity at 10µM against NCI 60 cell line panel.

CHP-5MO-SAL



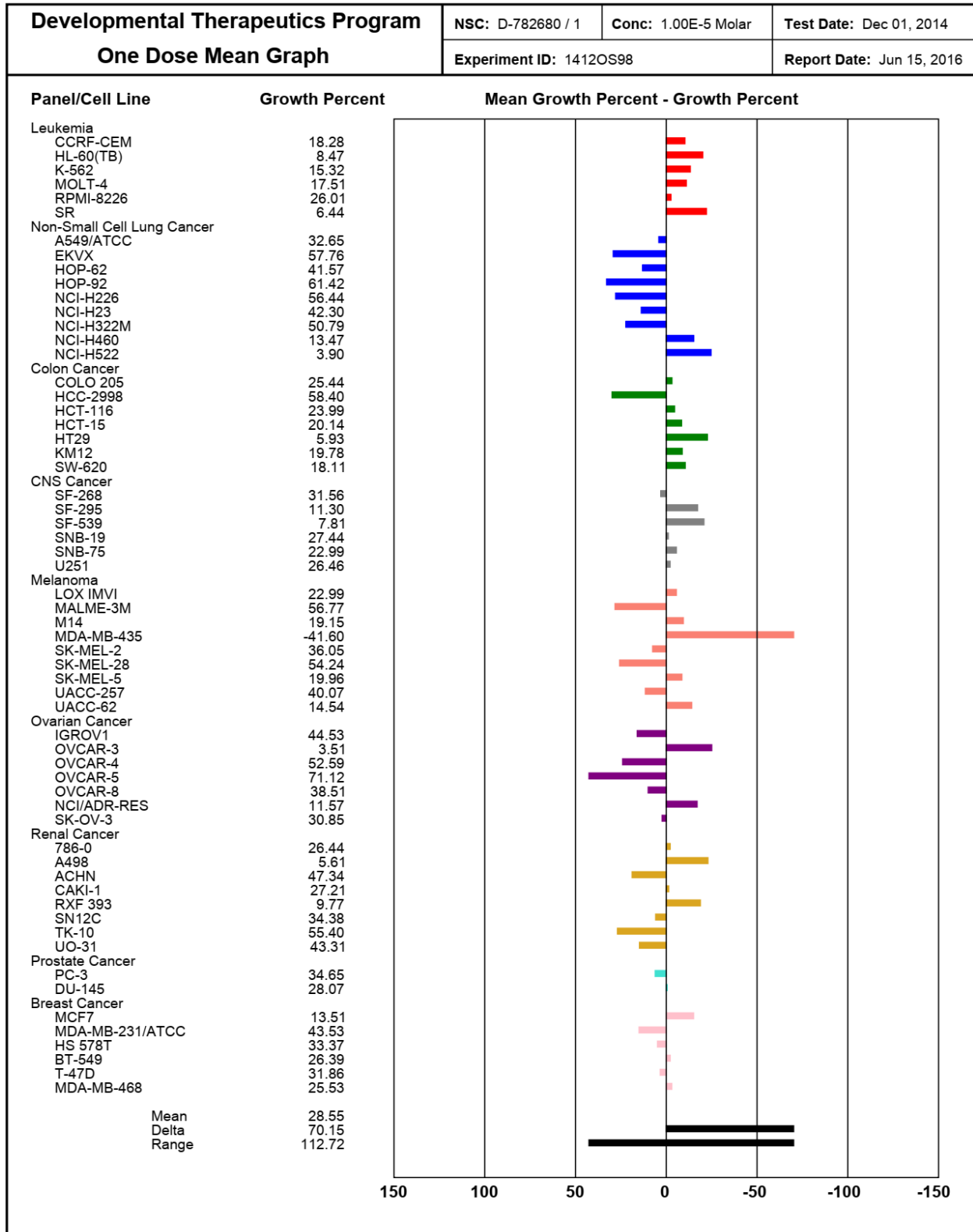
***In-vitro* anti-cancer activity at 10µM against NCI 60 cell line panel.**

CHP-DiCl-SAL



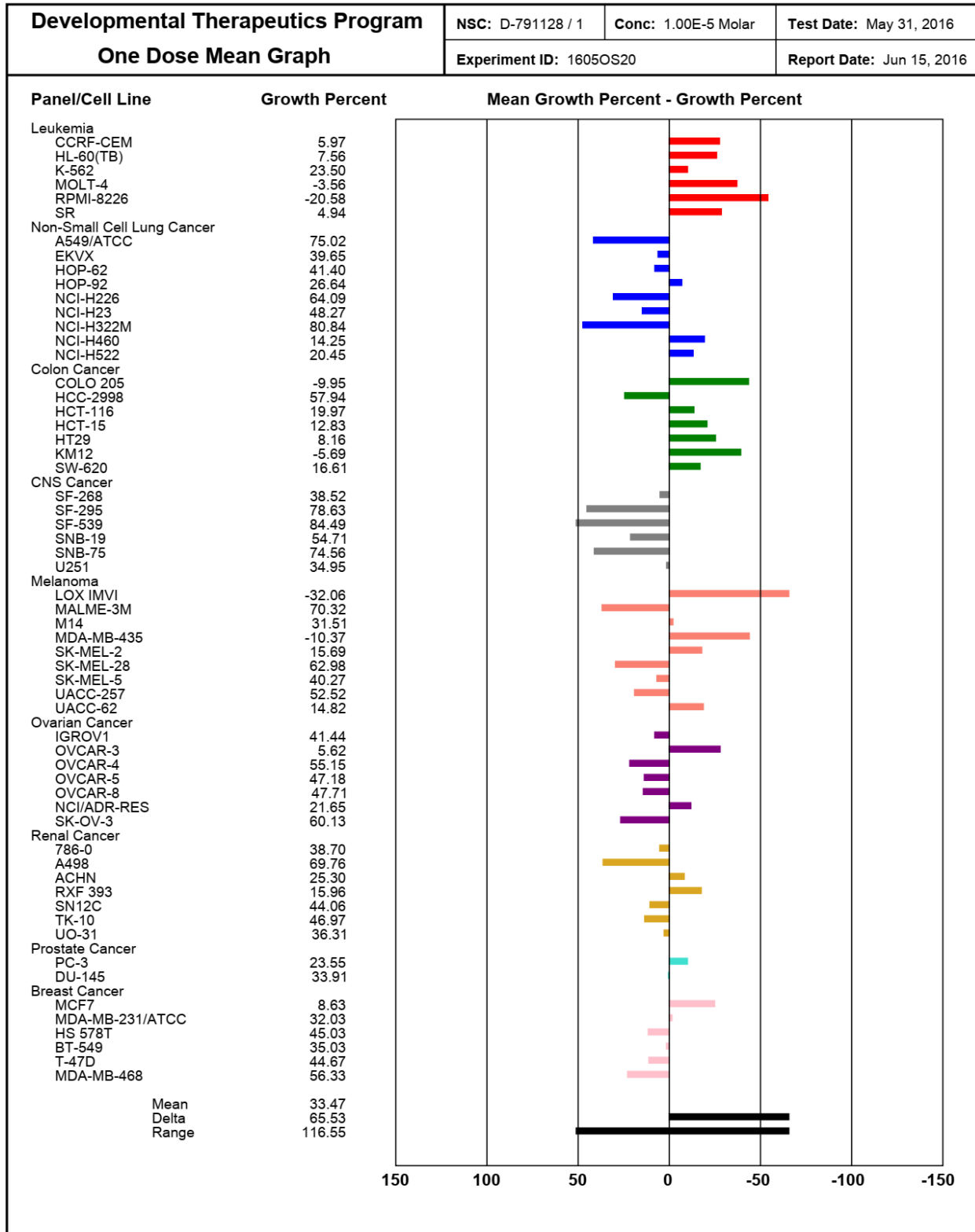
In-vitro anti-cancer activity at 10µM against NCI 60 cell line panel.

CHX-SAL



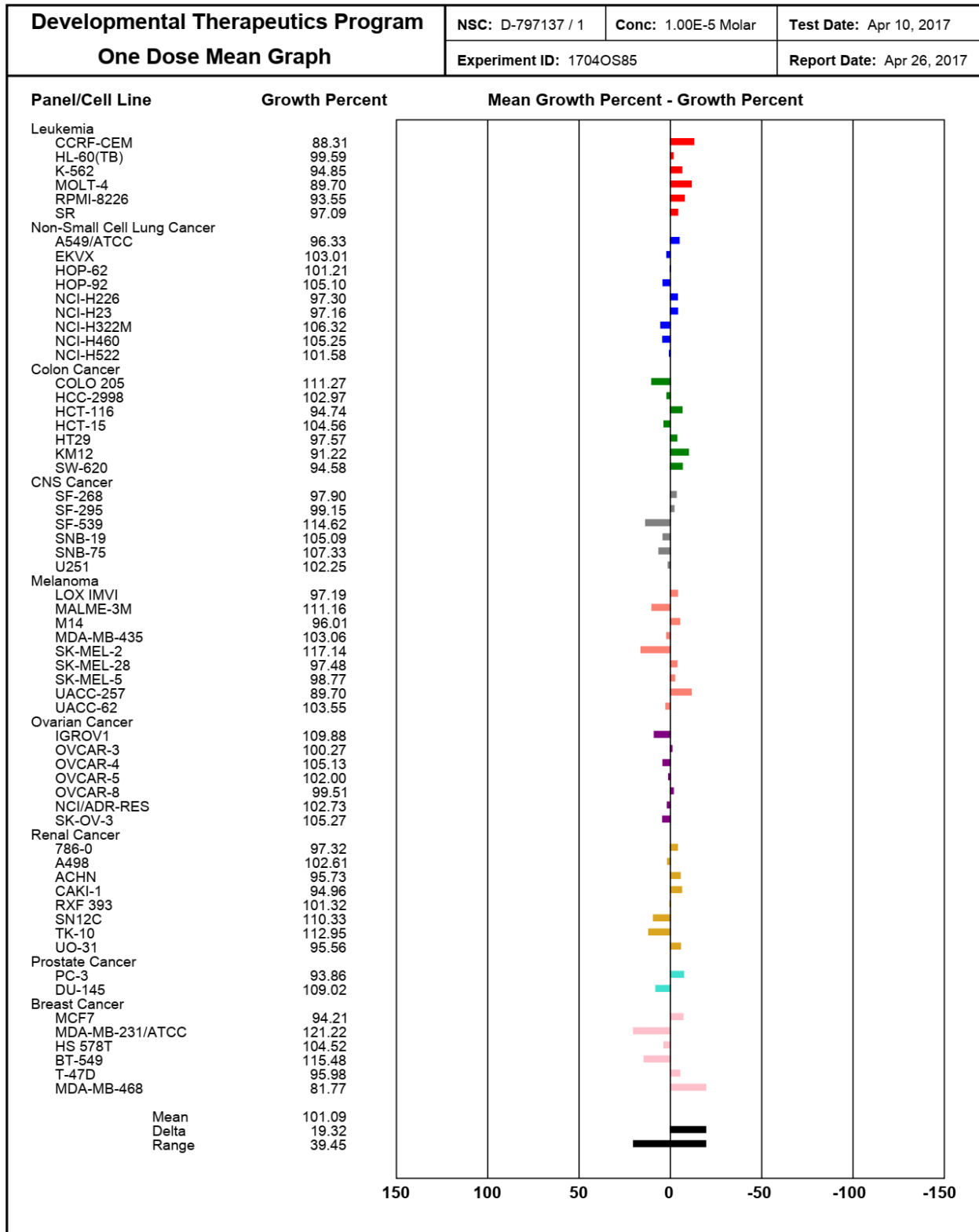
In-vitro anti-cancer activity at 10µM against NCI 60 cell line panel.

CHX-4MO-SAL



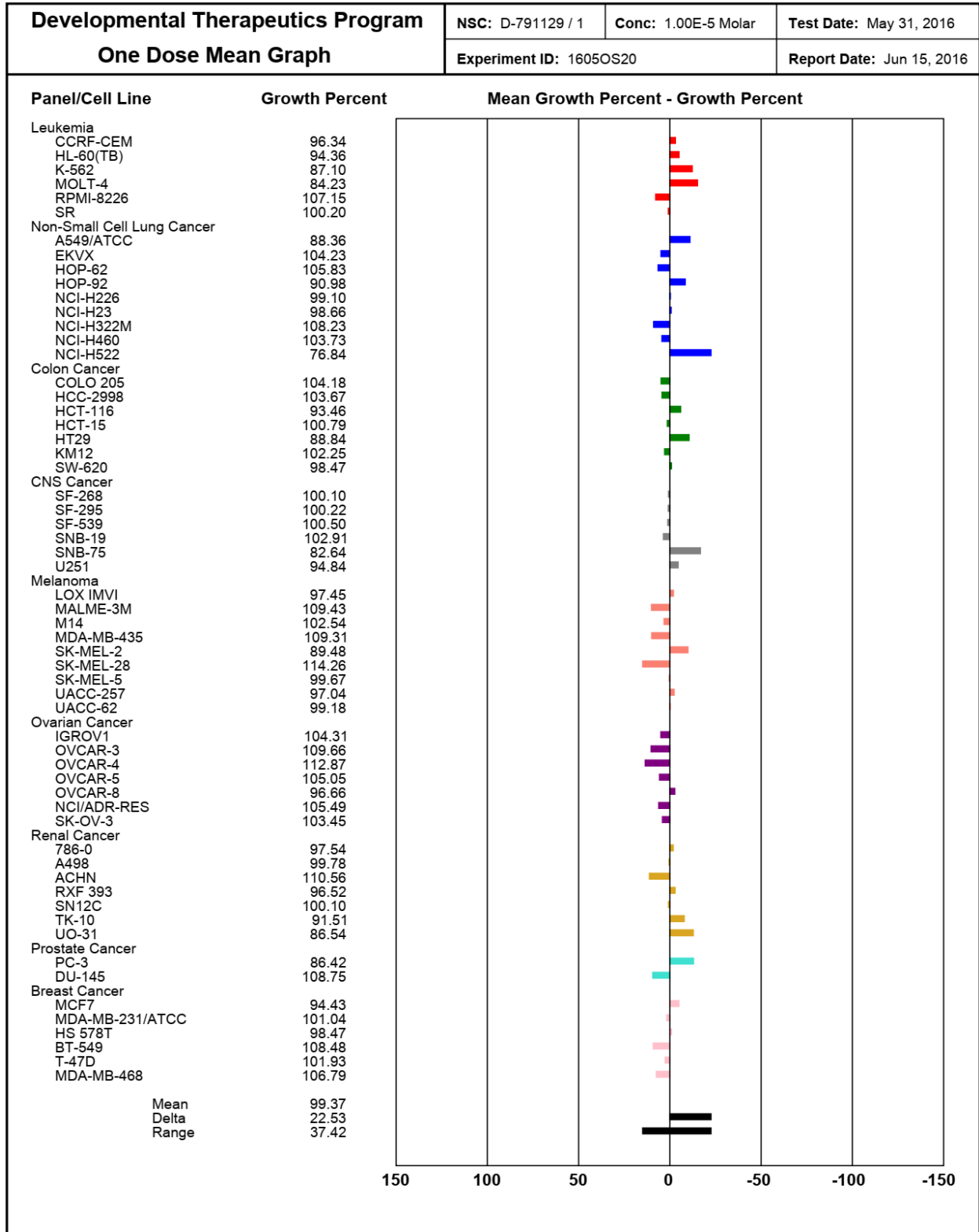
In-vitro anti-cancer activity at 10µM against NCI 60 cell line panel.

CHX-5MO-SAL



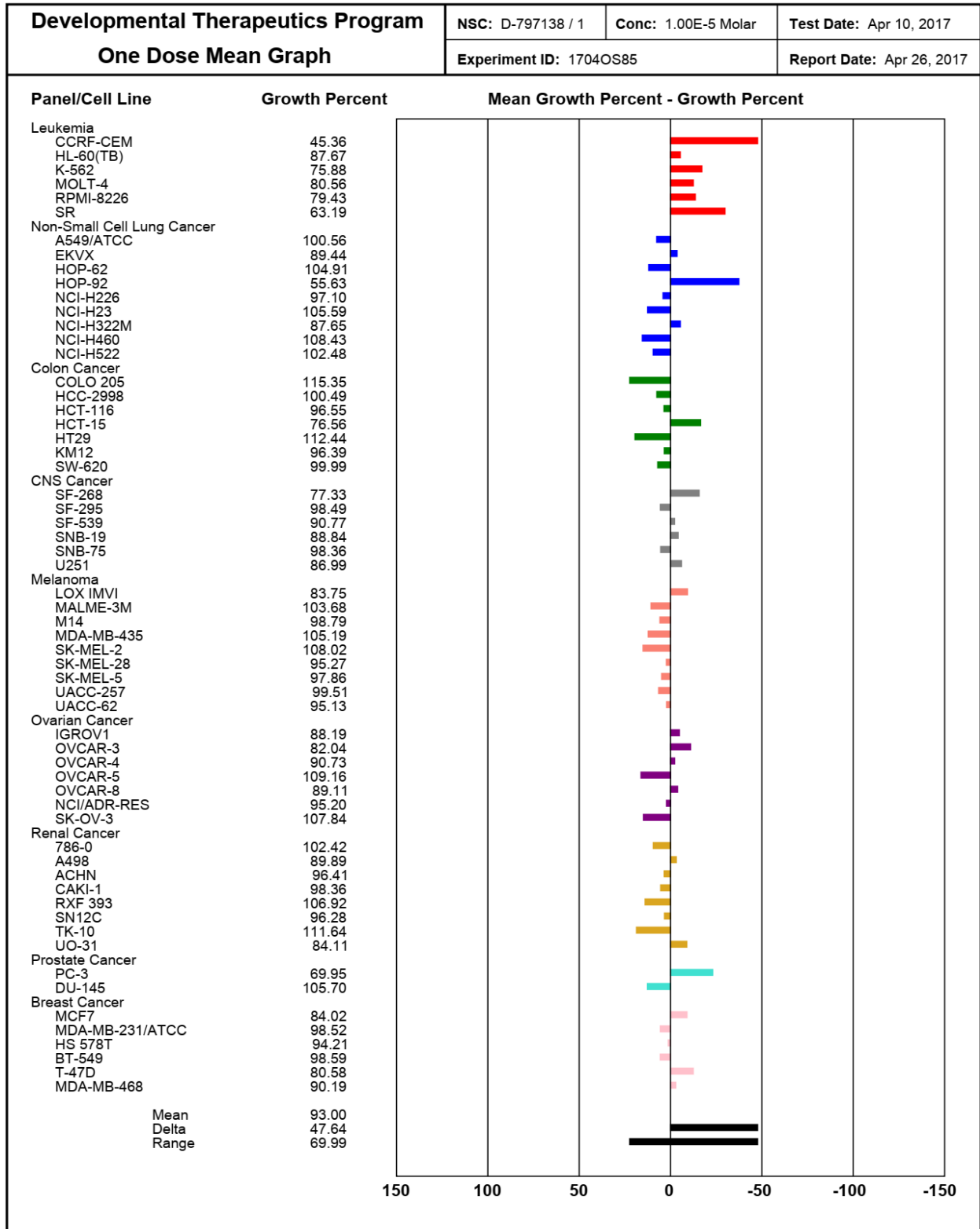
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CHX-5CI-SAL



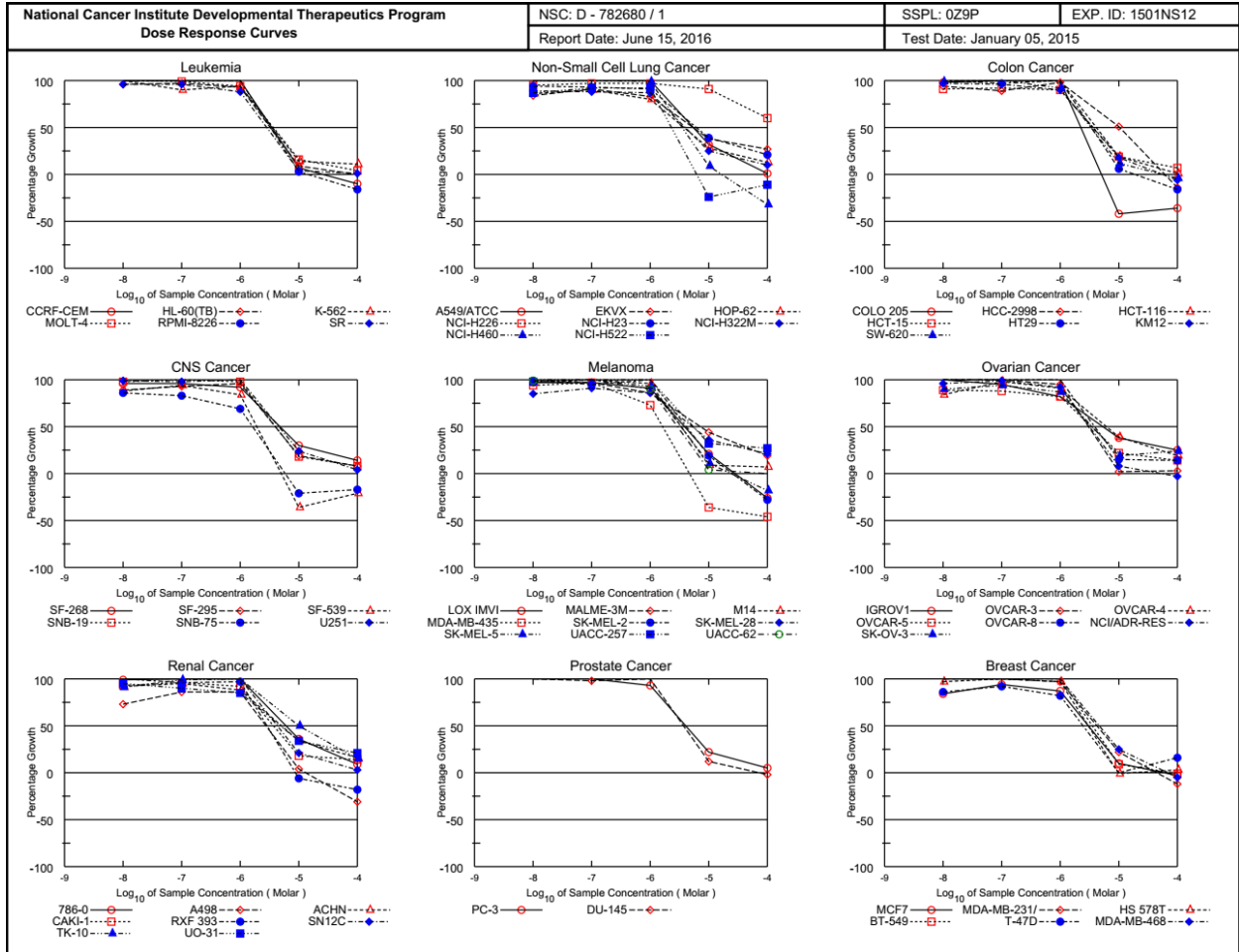
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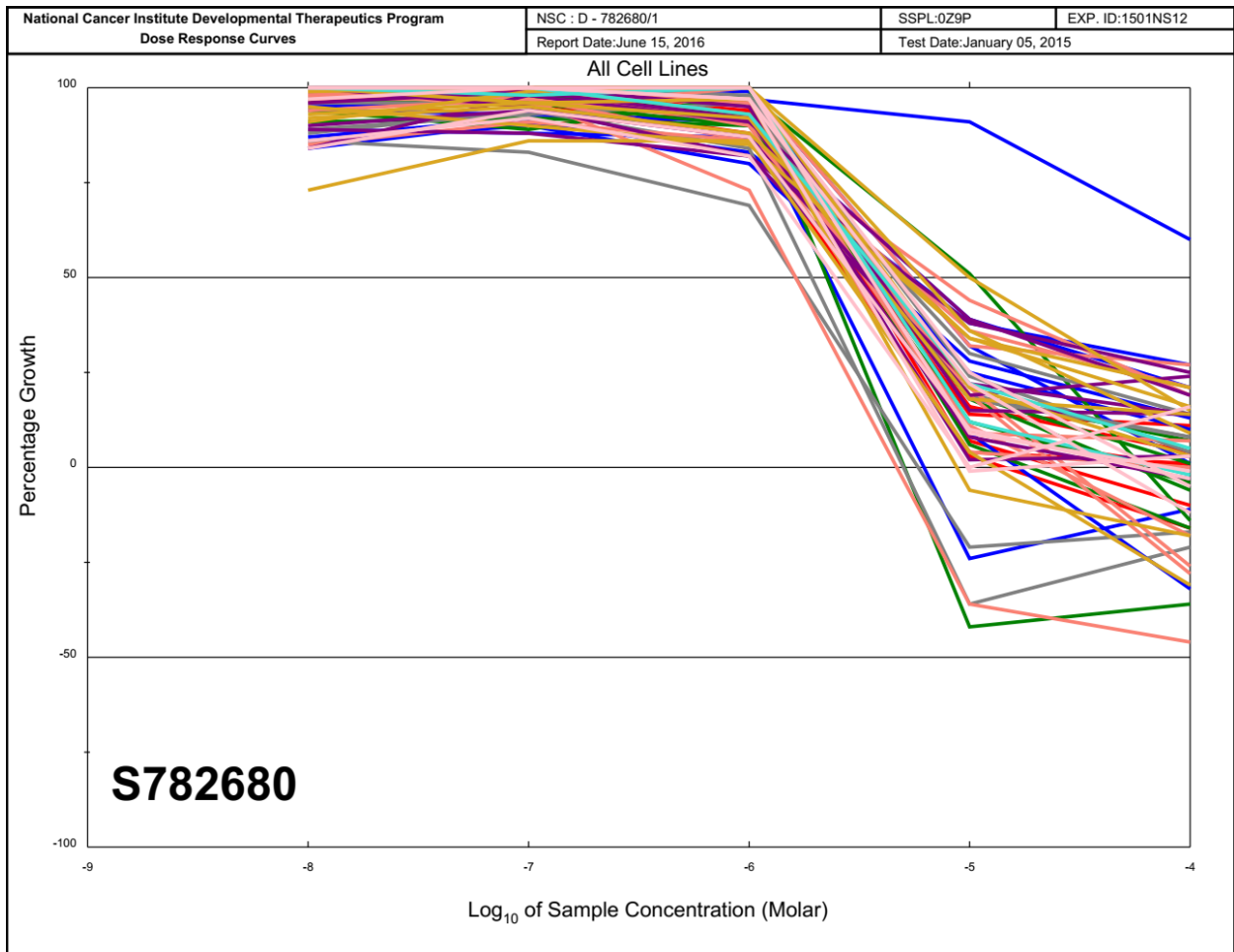
CHX-DiCl-SAL



***In-vitro* anti-cancer activity against NCI 60 cell line panel – five dose study.**

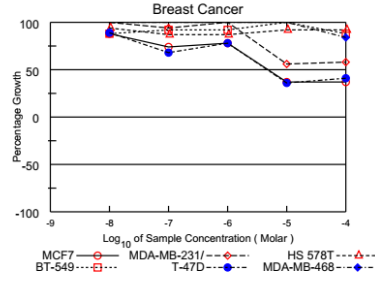
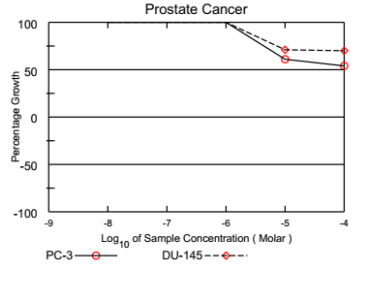
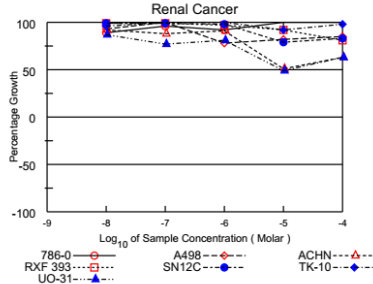
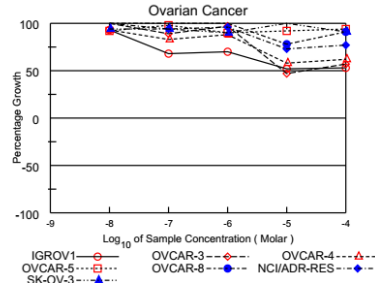
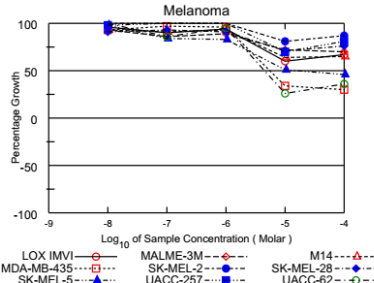
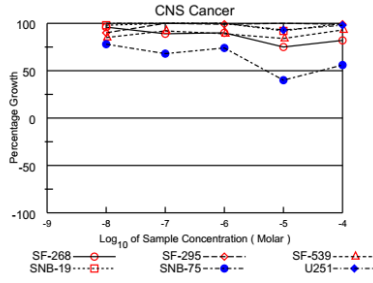
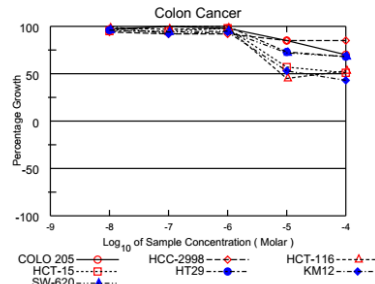
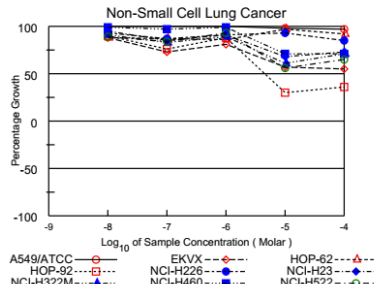
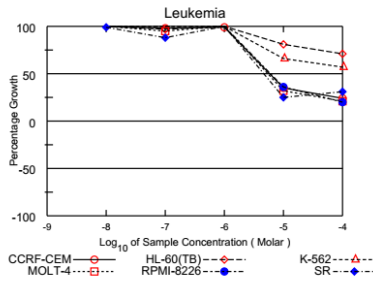
CHX-SAL

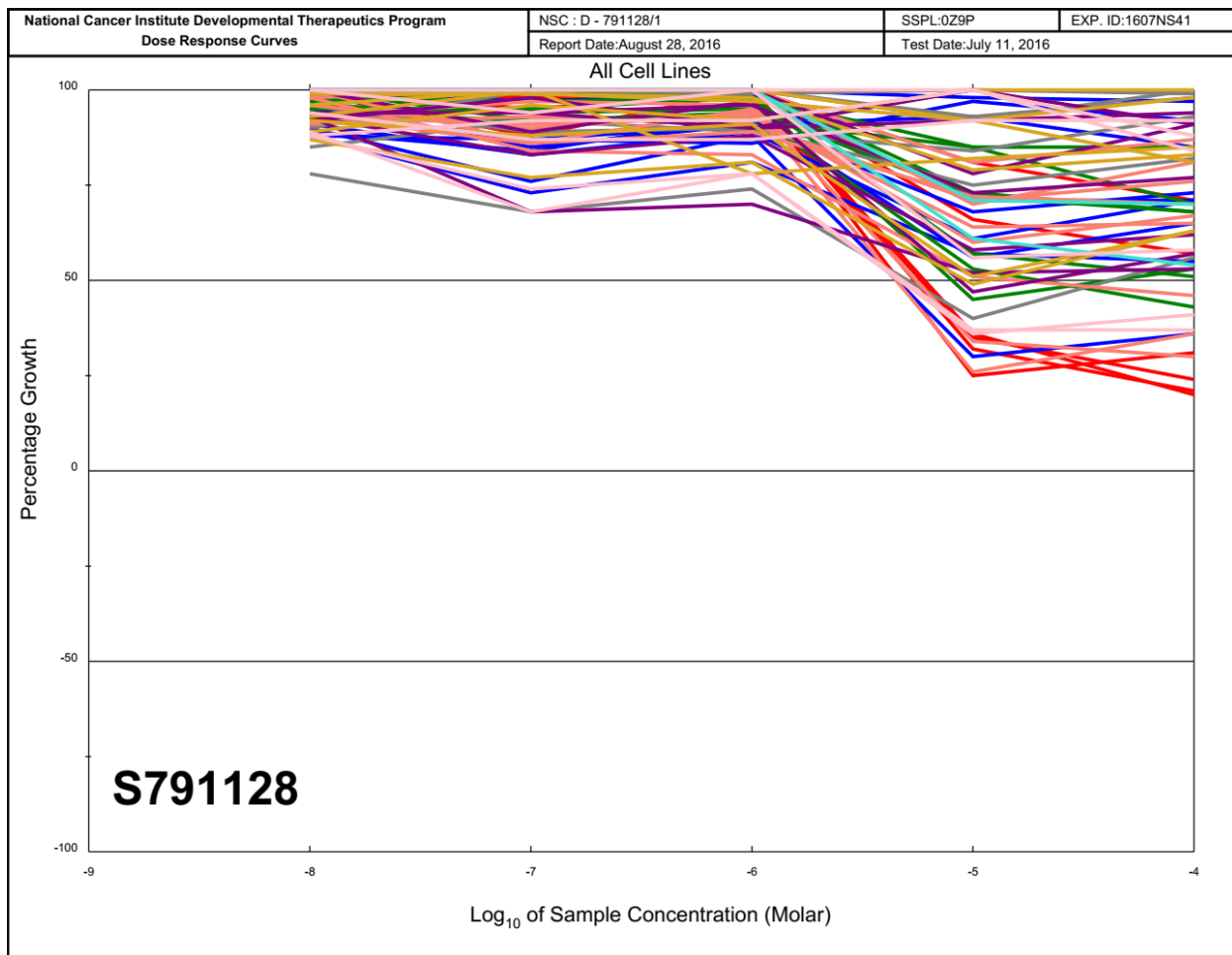




In-vitro anti-cancer activity against NCI 60 cell line panel – five dose study.

CHX--4MO-SAL





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5. Computational studies

The spirobibenzopyrans used for the studies namely: CHP-5Cl-SAL, and CHP-5MO-SAL were modelled using GaussView (5.0.8) ⁵. The energy minimization of these molecules has been carried out using DFT theory in Gaussian 09 ⁶, along with frequency studies. The 10 spirobibenzopyrans selected for the *in vitro* analysis were then evaluated for their drug likeness by testing their Lipinski's parameters at:

<http://www.scfbioitd.res.in/software/drugdesign/lipinski.jsp> ^{7,8}.

Label	Experimental	Gaussian	
H-12	7.20	7.54	
H-10	7.10	7.47	
H-11	6.95	7.36	
H-13	6.80	7.15	
H-8	6.70	7.05	
H-6	2.40	2.70	3.28
H-3	2.60	2.57	2.88
H-4	2.10	1.97	
H-5	1.55	1.62	

SD table VII: ¹H-NMR comparison for CHP-SAL

Bond lengths (in Å)	EXPT	Gaussian
C11-C1	1.75	1.76
O1-C4	1.37	1.37
O1-C5	1.44	1.44
C1-C9	1.36	1.39
C1-C2	1.37	1.39
C2-C3	1.40	1.40
C3-C4	1.39	1.40
C3-C6	1.45	1.45
C4-C8	1.38	1.39
C5-C7	1.51	1.52
C6-C7	1.33	1.34
C7-C10	1.51	1.51
C8-C9	1.33	1.39
C10-C11	1.53	1.56
C11-C11	1.51	1.53

SD table VIIIa: Bond lengths comparison for CHP-5Cl-SAL

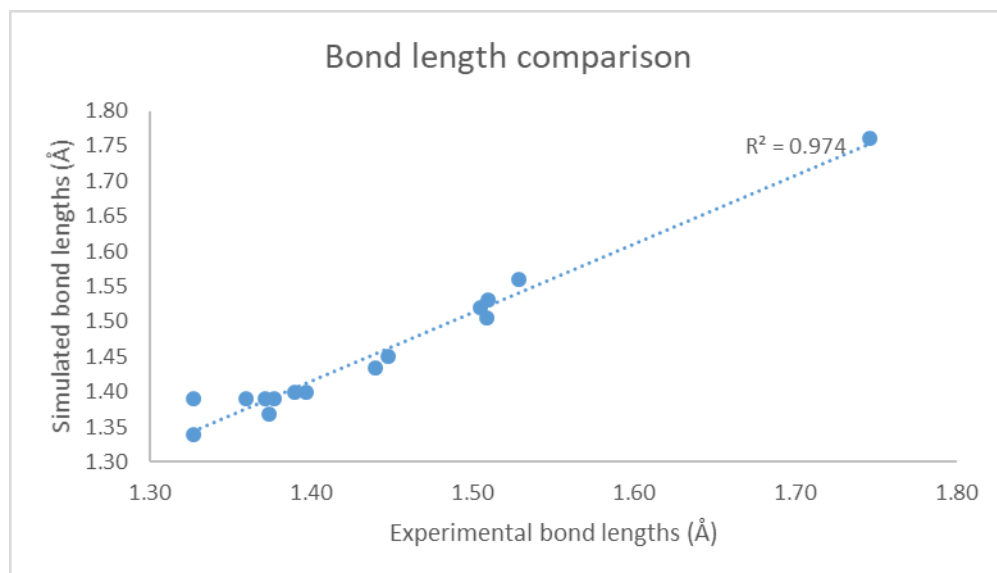


Figure III: Bond lengths comparison for CHP-5Cl-SAL with regression analysis

Bond angles (in degrees)	EXPT	Gaussian
C4-O1-C5	118.9	119.2
C9-C1-C2	121.8	121.2
C9-C1-C11	119.2	119.4
C2-C1-C11	119.0	119.4
C1-C2-C3	119.7	119.8
C4-C3-C2	118.0	118.7
C4-C3-C6	117.5	117.7
C2-C3-C6	124.4	123.5
O1-C4-C8	117.1	118.0
O1-C4-C3	121.2	120.7
C8-C4-C3	121.6	121.2
O1-C5-O1	107.5	108.3
O1-C5-C7	113.2	112.8
O1-C5-C7	103.4	104.6
C7-C5-C7	116.2	114.3
C7-C6-C3	122.0	121.7
C6-C7-C5	119.2	118.5
C6-C7-C10	123.8	124.0
C5-C7-C10	116.9	117.2
C4-C8-C9	119.2	119.7
C1-C9-C8	119.6	119.4
C7-C10-C11	113.1	114.9
C11-C11-C10	115.2	113.8

SD table VIIIb: Bond angles comparison for CHP-5Cl-SAL

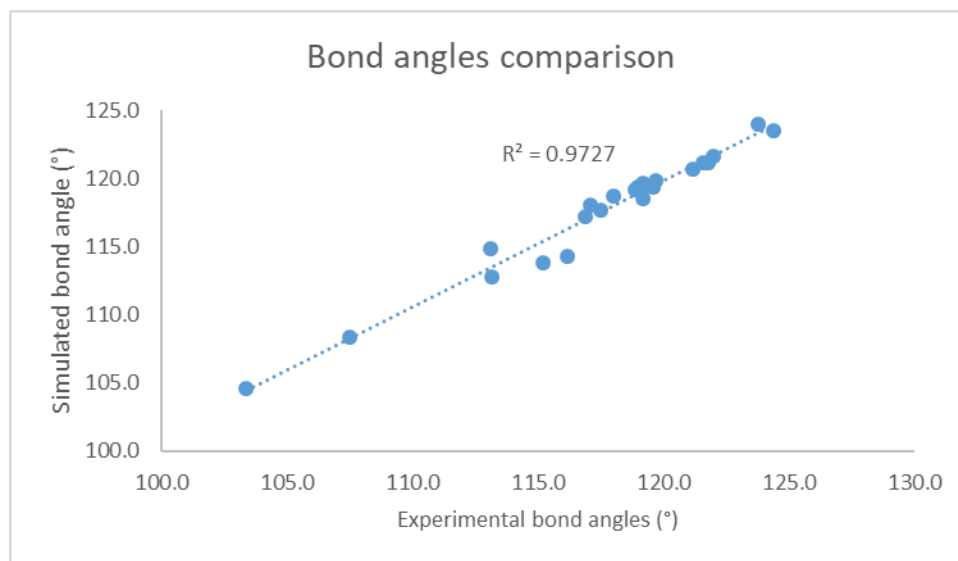


Figure IV: Bond angles comparison for CHP-5Cl-SAL with regression analysis

Bond lengths (in Å)	EXPT	Gaussian
O2-C21	1.38	1.38
O2-C1	1.44	1.43
O1-C2	1.38	1.37
O1-C1	1.43	1.43
O3-C5	1.37	1.37
O3-C22	1.38	1.41
C8-C9	1.31	1.34
C8-C7	1.45	1.45
C7-C6	1.38	1.40
C7-C2	1.40	1.40
C5-C6	1.38	1.39
C5-C4	1.39	1.40
C2-C3	1.37	1.39
C9-C1	1.50	1.51
C9-C10	1.51	1.51
C1-C14	1.52	1.52
C21-C20	1.38	1.39
C21-C16	1.41	1.40
C14-C15	1.32	1.34
C14-C13	1.51	1.50
C3-C4	1.39	1.39
C11-C12	1.52	1.53
C11-C10	1.53	1.56
C20-C19	1.41	1.39
C15-C16	1.44	1.45

C16-C17	1.40	1.40
C12-C13	1.54	1.56
C17-C18	1.37	1.39
C18-C19	1.39	1.40
C18-O4	1.48	1.37
O4-C23	1.42	1.41

SD table IXa: Bond lengths comparison for CHP-5MO-SAL

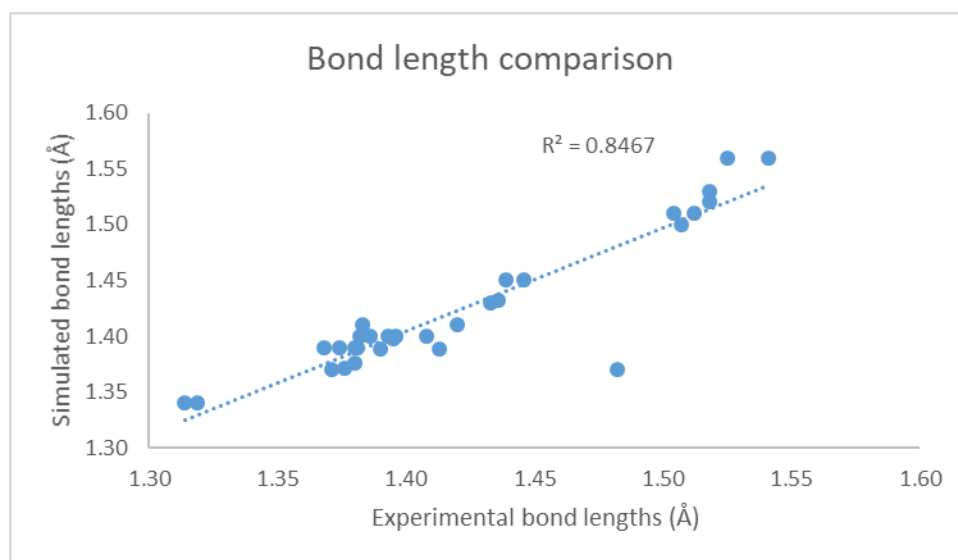


Figure V: Bond lengths comparison for CHP-5MO-SAL with regression analysis

Bond angles (in degrees)	EXPT	Gaussian
C21-O2-C1	120.0	121.0
C2-O1-C1	119.4	117.9
C5-O3-C22	118.3	118.0
C9-C8-C7	122.4	121.3
C6-C7-C2	119.1	119.1
C6-C7-C8	123.6	123.3
C2-C7-C8	117.1	117.5
O3-C5-C6	115.7	115.5
O3-C5-C4	125.1	124.7
C6-C5-C4	119.2	119.7
C5-C6-C7	121.0	120.3
C3-C2-O1	118.1	118.2
C3-C2-C7	120.4	120.7
O1-C2-C7	121.2	120.9
C8-C9-C1	119.6	119.1
C8-C9-C10	123.1	125.1
C1-C9-C10	117.3	116.5
O1-C1-O2	108.0	108.8

O1-C1-C9	113.1	112.2
O2-C1-C9	103.7	103.0
O1-C1-C14	104.1	105.4
O2-C1-C14	114.1	112.9
C9-C1-C14	114.1	114.4
O2-C21-C20	116.4	118.2
O2-C21-C16	120.1	120.8
C20-C21-C16	123.3	120.7
C15-C14-C13	123.7	121.1
C15-C14-C1	119.7	119.2
C13-C14-C1	116.6	116.5
C2-C3-C4	119.7	119.7
C5-C4-C3	120.4	120.4
C12-C11-C10	114.5	114.0
C21-C20-C19	116.9	119.6
C9-C10-C11	112.4	113.0
C14-C15-C16	121.6	122.7
C17-C16-C21	117.6	119.3
C17-C16-C15	123.2	123.4
C21-C16-C15	119.2	117.2
C11-C12-C13	114.1	115.6
C14-C13-C12	113.2	113.0
C18-C17-C16	120.7	120.1
C17-C18-C19	120.9	119.7
C18-C19-C20	120.6	120.5

SD table IXb: Bond angles comparison for CHP-5MO-SAL

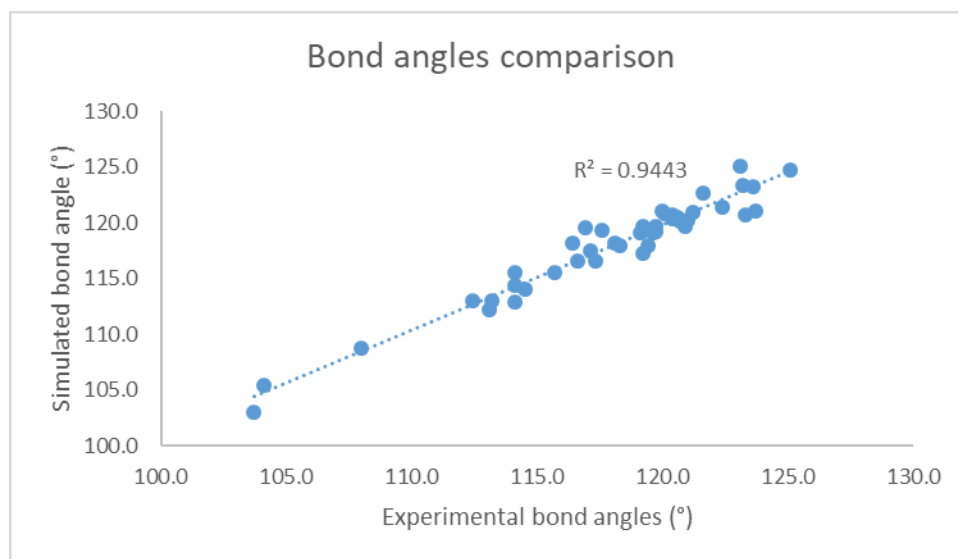


Figure VI: Bond angles comparison for CHP-5MO-SAL with regression analysis

SD Table X. Regression analysis of the observed IR values vs the computed IR values

Molecule	R ² value for Observed IR vs Computed IR
CHP-5MO-SAL	0.8176
CHP-5Cl-SAL	0.7940

SD Table XI. Drug likeness score of the spirobibenzopyrans on the basis of Lipinski parameters

Spirobibenzopyrans	Molecular mass	No. of hydrogen bond donors	No. of hydrogen bond acceptors	Log P	Molar refractivity
CHP-SAL	302	0	2	5.21	91.29
CHP-4MO-SAL	362	0	4	5.23	104.40
CHP-5Cl-SAL	371	0	2	4.97	95.45
CHP-5MO-SAL	362	0	4	5.23	104.40
CHP-DiCl-SAL	440	0	2	4.73	99.61
CHX-SAL	288	0	2	4.82	86.68
CHX-4MO-SAL	348	0	4	4.84	99.78
CHX-5MO-SAL	348	0	4	4.84	99.78
CHX-5Cl-SAL	357	0	2	4.58	90.83
CHX-DiCl-SAL	426	0	2	4.34	94.99

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