

Supporting Information

Enhanced immunofluorescence detection of protein marker using PAA modified ZnO nanorod array-based microfluidic device

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Figures

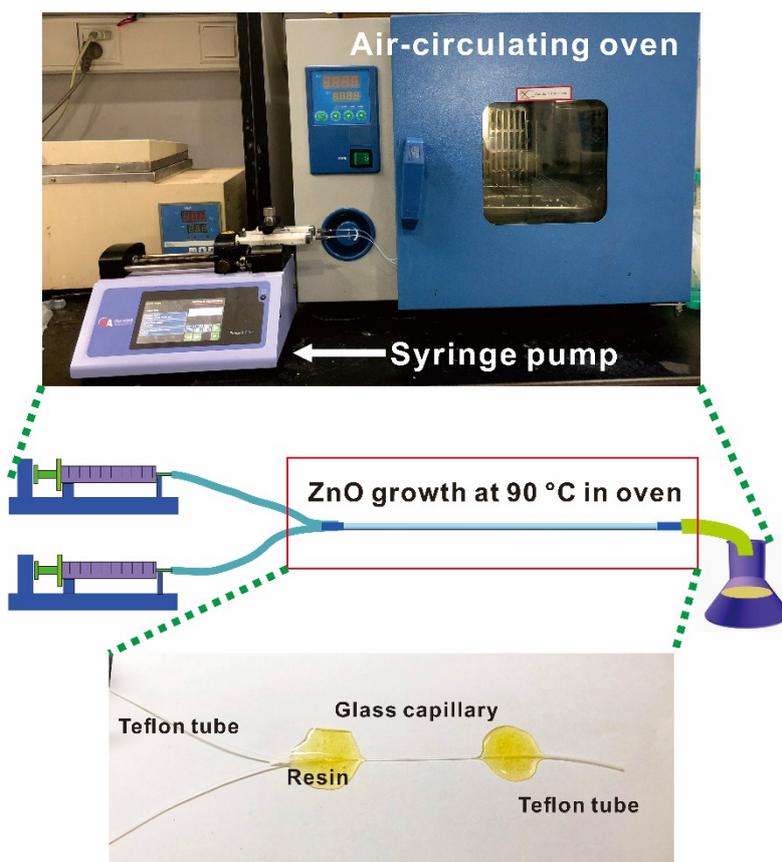


Fig. S1 Photographs of fabrication process and the microfluidic device. The diameter of the glass capillaries is about 500 μm . The whole length of capillaries is 10 cm, and the available part is 5-8 cm. As shown in Fig. S1, the capillary was placed in air-circulating oven to maintain the reaction temperature at 90°C. Glass capillary was sealed using resin to ensure the airtightness. Besides, ZnO nanorods can be constructed in the innerwall of glass with the capillary diameter of 100-900 μm .

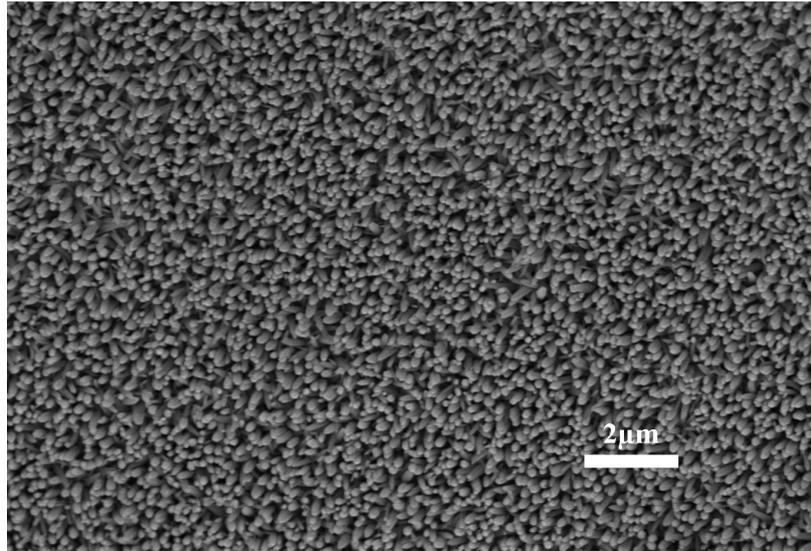


Fig. S2 Low-magnification SEM image of ZnO-III nanorod arrays.

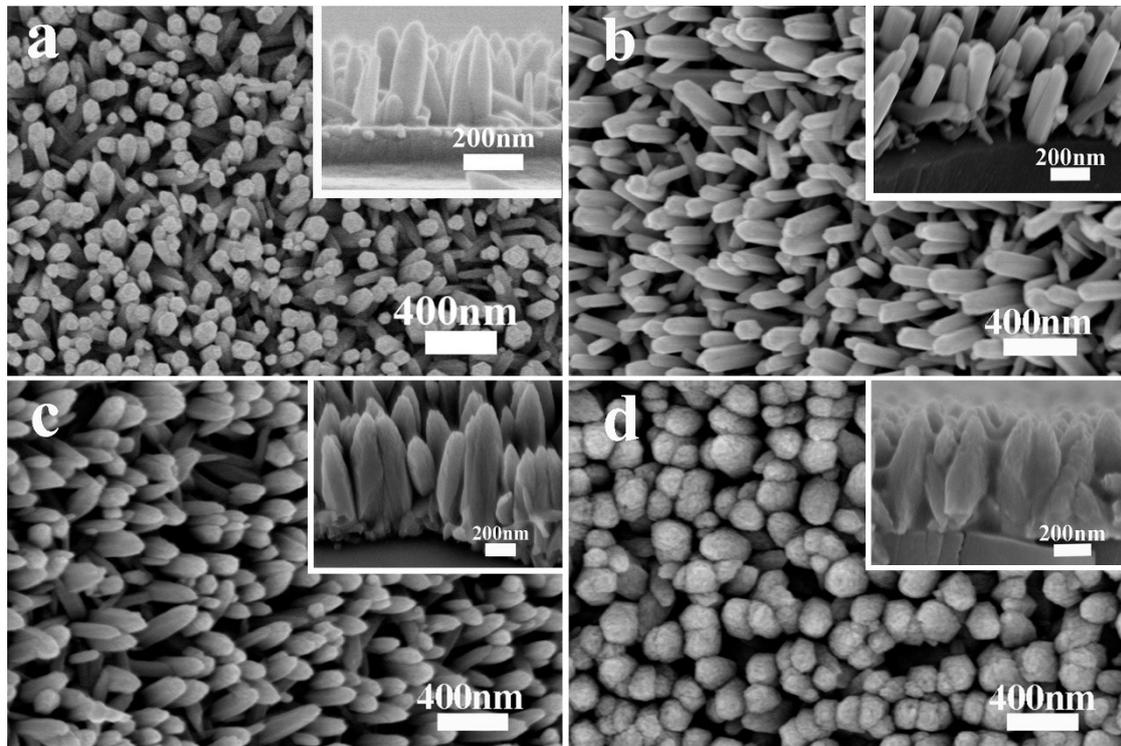


Fig. S3 SEM images of ZnO nanorods grown in capillaries at 90 °C with the Zn²⁺ concentration of 0.10 M, 5 min at the rate of 0 μL•min⁻¹ (a) (ZnO-V), repeat (a) for another 5 min (b) (ZnO-VI), 4 min at the rate of 25 μL•min⁻¹ (c) (ZnO-VII), and 2 min at the rate of 50 μL•min⁻¹ (d) (ZnO-VIII).

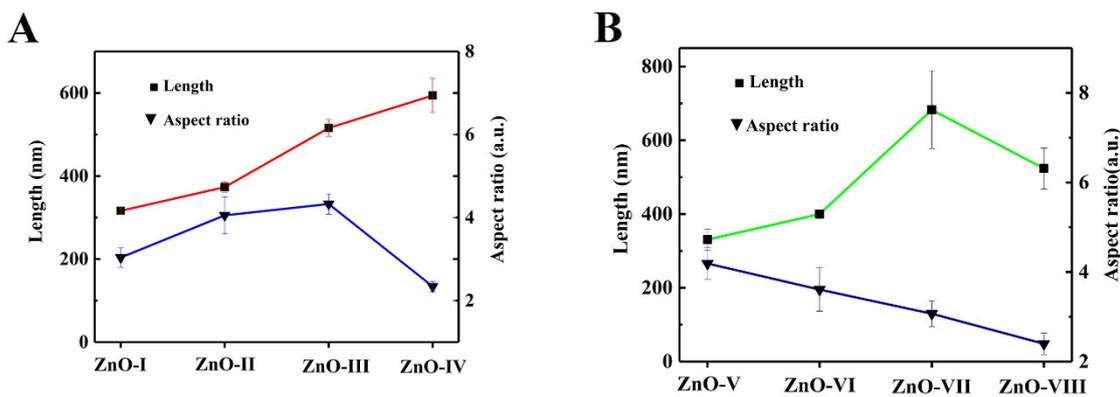


Fig. S4 Statistics of aspect ratio and length of ZnO nanorods prepared with different reaction solution concentration, (A) 0.05 M (ZnO- I ~ ZnO-IV), (B) 0.10 M (ZnO-V ~ ZnO-VIII).

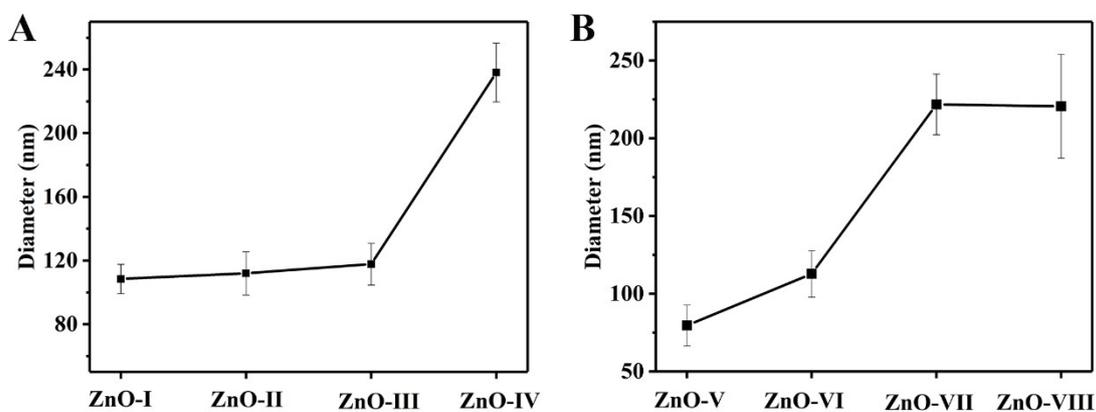


Fig. S5 Statistics of diameter of ZnO nanorods prepared with different reaction solution concentration, (A) 0.05 M (ZnO- I ~ ZnO-IV), (B) 0.10 M (ZnO-V ~ ZnO-VIII).

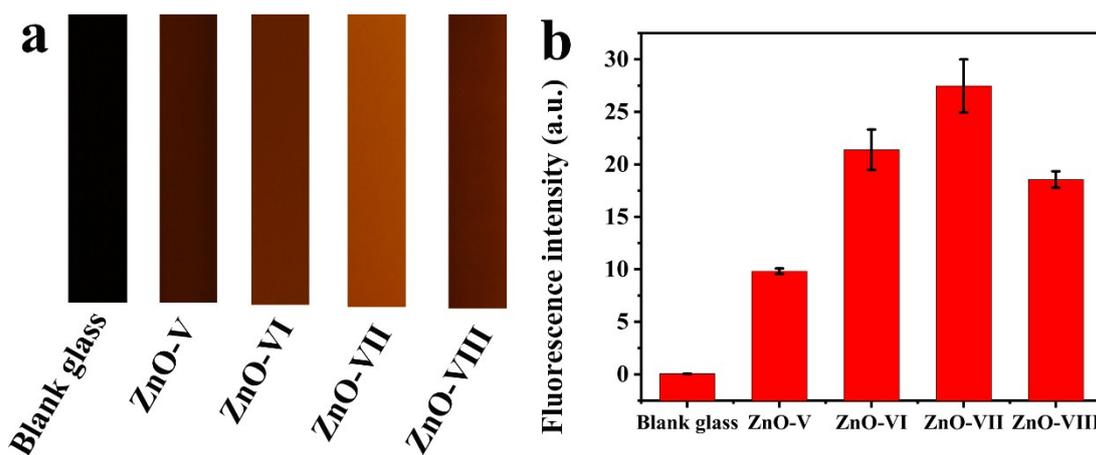


Fig. S6 Comparison of fluorescent enhancement of ZnO nanorods with with Zn^{2+} concentration of 0.10 M, (a) Fluorescent images, (b) the corresponding fluorescence intensity comparison of (a).

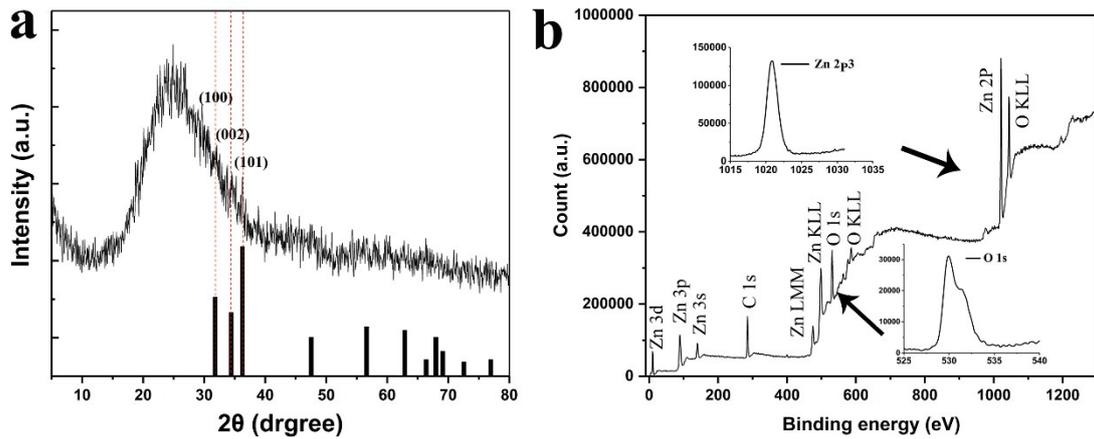


Fig. S7 (a) Powder-XRD pattern of ZnO nanorods. Due to the high content of glass in powder sample, it shows a broad diffraction peak positioned at 2-theta angles of 25, (b) XPS survey spectrum of ZnO nanorods.

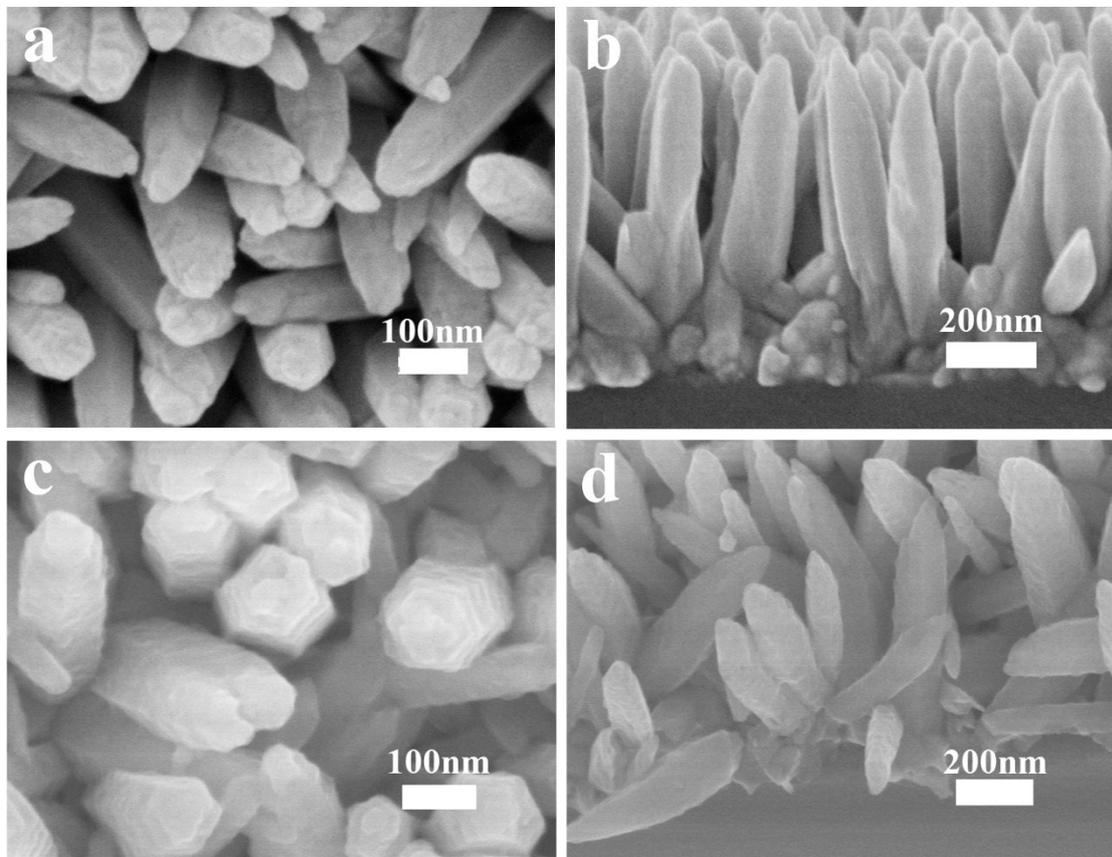


Fig. S8 Top view and cross-section view of SEM images of ZnO nanorods (reaction solution flowing through the capillary for 4 min at the rate of $25 \mu\text{L}\cdot\text{min}^{-1}$), (a) (b) before PAA modification, (c) (d) PAA modified ZnO nanorods.

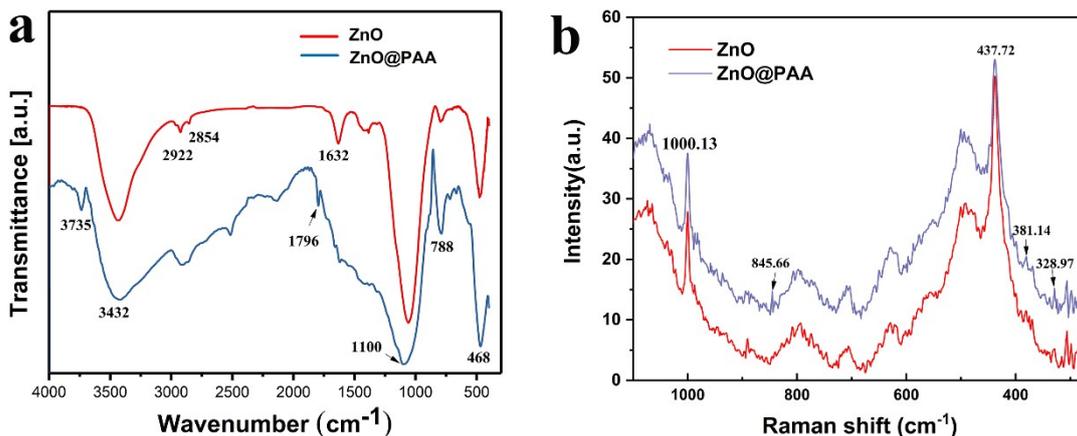


Fig. S9 Characterization of ZnO nanorods, (a) FTIR spectra of original ZnO and ZnO@PAA, (b) Raman spectrum of ZnO and ZnO@PAA. The characteristic Raman peak of ZnO is located at 437.72 cm^{-1} , which is ascribed to E_{2H} mode of ZnO. Besides, Raman peaks located at 381.14 and 328.97 cm^{-1} can be assigned to the $A_1(\text{TO})$ and $3E_{2H} - E_{2L}$ mode of ZnO (J. Phys. Chem. C 2008, 112, 7332-7336). And the peak centered at 1000.13 cm^{-1} could be assigned to $\delta(\text{COH})$ resulting from ethanolamine. (Carbohydrate research, 1996, 284(2): 145-157; J. Am. Chem. Soc., 1964, 86, 559-564). In the case of ZnO@PAA nanorods, the Raman peak located at 845.66 cm^{-1} corresponds to $\nu(\text{C-COO})$ stretching, which also indicates the successful modification of PAA (Polymer, 1998, 39(2): 267-273).

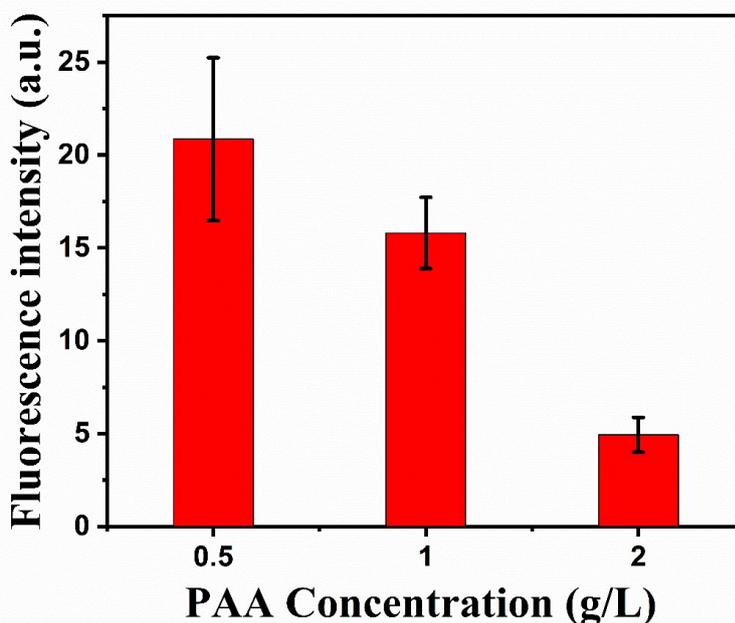


Fig. S10 Nonspecific protein adsorption on ZnO nanorod array with different contents of PAA coating.

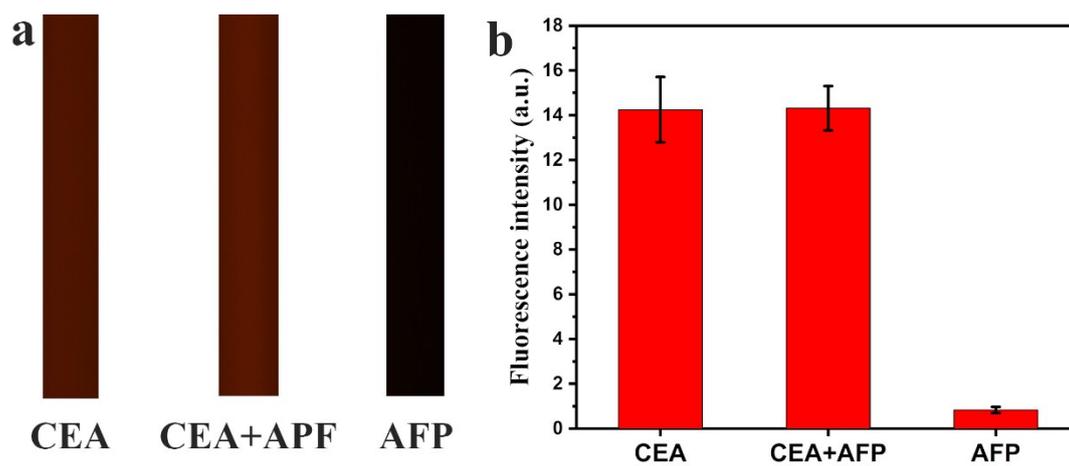


Fig. S11 Specific detection of CEA (the concentration of CEA and AFP is $1 \text{ ng}\cdot\text{mL}^{-1}$). (a) Fluorescent images of CEA, CEA+AFP, AFP, respectively, (b) Corresponding fluorescent intensity of each group.