

## Supporting Information

### Visible light driven metal-free intramolecular cyclization: a facile synthesis to 3-substituted 3,4-dihydroisoquinolin-1(2H)-one

Shuai Zou, Shu Geng, Lina Chen, Haitao Wang and Feng Huang\*

*State Key Laboratory of NBC Protection for Civilian, Beijing 102205, P. R. China.*

\*E-mail: john\_hf@163.com

#### Table of Contents

1. General Information	S2
2. Experimental Section	
2.1 Synthesis and Characterization of Substrates	S2-S10
2.2 Synthesis and Characterization of Products	S11-S16
2.3 Mechanistic Studies	S17-S19
2.4 Gram-scale Reaction	S19
2.5 UV-Vis Spectral Analysis	S19-S22
2.6 Comparison Experiment between Blue and Green Light Source	S22-S25
3. References	S26
4. NMR Spectra	S27-S70

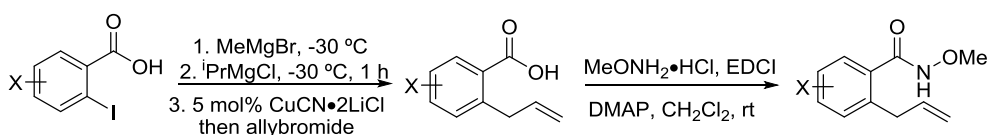
## 1. General information

All reactions were carried out under nitrogen atmosphere using Schlenk techniques. Reagents were purchased at the highest commercial quality and used without further purification, unless otherwise noted. EosinY Na was purchased from Solarbio (China). DBU was purchased from J&K and transferred under a nitrogen atmosphere. Solvents were dried and purified according to the procedure from "Purification of Laboratory Chemicals book". Analytical thin layer chromatography (TLC) was performed on pre-coated silica gel HSGF254 plates. Flash column chromatography was performed using Yantai silica gel (60, particle size 0.040-0.063 mm). Visualization on TLC was achieved by use of UV light (254 nm), iodine or  $\text{KMnO}_4$  solution. HPLC analysis was completed by Waters 2998 Photodiode Array Detector, Waters 1525 Binary HPLC Pump and Waters 2707 Autosampler. NMR spectra were recorded on Bruker AVANCE-300 spectrometer at 300 MHz for  $^1\text{H}$  NMR, 75 MHz for  $^{13}\text{C}$  NMR in  $\text{CDCl}_3$  with tetramethylsilane (TMS) as internal standard. The chemical shifts are expressed in ppm and coupling constants are given in Hz. Data for  $^1\text{H}$  NMR are recorded as follows: chemical shift (ppm), multiplicity (s, singlet; d, doublet; t, triplet; q, quarter; p, pentet; m, multiplet; br, broad), coupling constant (Hz), integration. Data for  $^{13}\text{C}$  NMR are reported in terms of chemical shift ( $\delta$ , ppm). High resolution mass spectrometry (HRMS) spectra were obtained on a Agilent 6520Q-TOF instrument.

## 2. Experimental Section

### 2.1 Synthesis and Characterization of Substrates

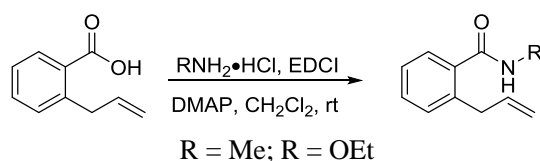
General synthesis of substrates **1a-1f**, **1h** and **1k**<sup>1,2</sup>:



1. To a stirred solution of 2-iodobenzoic acid (2.48 g, 10 mmol) and THF (30.4 mL) at  $-30\text{ }^{\circ}\text{C}$  in an oven-dried flask under argon was added MeMgBr (10 mL, 1 equiv., 1 M in THF) and stirred for 5 minutes. *i*PrMgCl (6.0 mL, 1.2 equiv., 2 M in THF) was added slowly and the reaction was stirred at  $-30\text{ }^{\circ}\text{C}$  for 1 hour or until the reaction was complete by GC/MS (an aliquot was quenching with water before analysis). The reaction was then cooled to  $-40\text{ }^{\circ}\text{C}$  and a solution of CuCN 2LiCl in THF (0.09 mL, 5 mol%, 0.34 M,) was added slowly and stirred for 10 minutes while warming to  $-30\text{ }^{\circ}\text{C}$ . Allyl bromide (3 equiv. 2.6 mL) was added at once and the reaction was allowed to warm to ambient temperature overnight. The reaction was diluted with EtOAc, acidified with 1 M HCl to a pH of 3, and extracted with EtOAc. The combined organic layers were washed with brine and dried over  $\text{Na}_2\text{SO}_4$ . The crude mixture was concentrated and purified by column chromatography on silica gel to yield 2-allylbenzoic acid (white solid 94%).

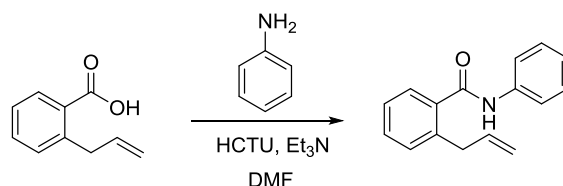
2. To a solution of 2-allylbenzoic acid (1.62 g, 10 mmol) in  $\text{CH}_2\text{Cl}_2$  (50 mL) was added MeONH<sub>2</sub> HCl (1.26 g, 1.5 equiv., 15 mmol), EDCI (3.85 g, 2.0 equiv. 20 mmol) and DMAP (1.25 g, 2 equiv., 20 mmol) successively. The resulting mixture was stirred at room temperature overnight. The reaction was quenched by adding HCl (2 M, 25 mL). The organic layer was separated and the aqueous layer was extracted with  $\text{CH}_2\text{Cl}_2$ . The combined extracts were washed with brine, dried over  $\text{Na}_2\text{SO}_4$ , and filtrated. The filtrate was concentrated and purified by column chromatography on silica gel to yield 2-allyl-*N*-methoxybenzamide (white solid 81%).

### General synthesis of substrates **1g**<sup>2</sup>:



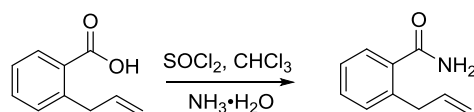
To a solution of 2-allylbenzoic acid (1.62 g, 10 mmol) in  $\text{CH}_2\text{Cl}_2$  (50 mL) was added  $\text{MeNH}_2\cdot\text{HCl}$  (1.03 g, 1.5 equiv., 15 mmol), EDCI (3.85 g, 2.0 equiv. 20 mmol) and DMAP (1.25 g, 2 equiv. 20 mmol) successively. The resulting mixture was stirred at room temperature overnight. The reaction was quenched by adding HCl (2 M, 25 mL). The organic layer was separated and the aqueous layer was extracted with  $\text{CH}_2\text{Cl}_2$ . The combined extracts were washed with brine, dried over  $\text{Na}_2\text{SO}_4$ , and filtrated. The filtrate was concentrated. Purification by column chromatography gave the product.

### Synthesis of substrates 2-allyl-N-phenylbenzamide<sup>3</sup>:



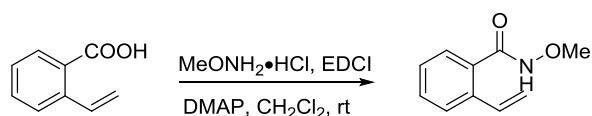
To a solution of 2-allylbenzoic acid (487 mg, 3.0 mmol), HCTU (1.37 g, 3.3 mmol) in 6 mL DMF was added  $\text{Et}_3\text{N}$  (459  $\mu\text{L}$ , 3.3 mmol). The reaction mixture was stirred at room temperature for 10 minutes. Then aniline (302  $\mu\text{L}$ , 3.3 mmol) was added. The resulting reaction mixture was stirred at room temperature for 4 h, and diluted with  $\text{CH}_2\text{Cl}_2$  (30 mL) and water (30 mL). The organic layer was separated and the aqueous layer was extracted with  $\text{CH}_2\text{Cl}_2$  (20 mL  $\times$  3). The combined organic layers were dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated in vacuo. The residue was purified by column chromatography on silica gel to yield the compound (472 mg, 67%) as a white solid.

### Synthesis of substrates 2-allylbenzamide<sup>4</sup>:



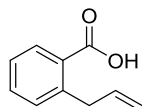
The solution of 2-allylbenzoic acid (324 mg, 2 mmol) and thionyl chloride (286 mg, 2.4 mmol) in  $\text{CHCl}_3$  was stirred at 70  $^\circ\text{C}$  for 2 h, then cooled to room temperature. The mixture was added slowly to aqueous ammonia (3 mL). The mixture was extracted with  $\text{CHCl}_3$  and the combined organic layer was washed with brine. The solvent was removed under reduce pressure to afford pure product as a white solid.

### General synthesis of substrates **1i-1j**<sup>2</sup>:



To a solution of 2-allylbenzoic acid (1.48 g, 10 mmol) in  $\text{CH}_2\text{Cl}_2$  (50 mL) was added  $\text{MeONH}_2\cdot\text{HCl}$  (1.26 g, 1.5 equiv. 15 mmol), EDCI (3.85 g, 2.0 equiv. 20 mmol) and DMAP (1.25

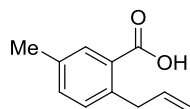
g, 2 equiv. 20 mmol) successively. The resulting mixture was stirred at room temperature overnight. The reaction was quenched by adding HCl (2M, 25 mL). The organic layer was separated and the aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined extracts were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and filtrated. The filtrate was concentrated. Purification by column chromatography gave the product.



**2-allylbenzoic acid<sup>5</sup>, white solid.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 12.16 (s, 1H), 8.07 (dd, *J* = 8.1, 1.3 Hz, 1H), 7.51 (td, *J* = 7.7, 1.4 Hz, 1H), 7.39 – 7.29 (m, 2H), 6.06 (ddt, *J* = 15.6, 11.3, 6.5 Hz, 1H), 5.11 – 4.95 (m, 2H), 3.85 (d, *J* = 6.5 Hz, 2H).

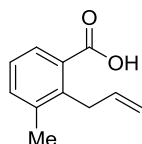
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 142.8, 137.3, 131.7, 131.2, 128.2, 126.4, 115.8, 38.6.



**2-allyl-5-methylbenzoic acid<sup>5</sup>, colorless oil.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.89 (s, 1H), 7.32 (d, *J* = 7.8 Hz, 1H), 7.21 (d, *J* = 7.8 Hz, 1H), 6.05 (dq, *J* = 15.1, 7.3 Hz, 1H), 5.05 (d, *J* = 12.7 Hz, 2H), 3.80 (d, *J* = 6.3 Hz, 2H), 2.38 (s, 3H).

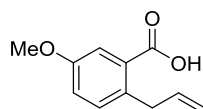
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 173.5, 139.8, 137.6, 136.0, 133.9, 132.1, 131.1, 128.1, 115.5, 38.2.



**2-allyl-3-methylbenzoic acid<sup>5</sup>, white solid.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.86 (d, *J* = 7.8 Hz, 1H), 7.37 (d, *J* = 7.5 Hz, 1H), 7.22 (t, *J* = 7.7 Hz, 1H), 5.99 (ddt, *J* = 16.2, 10.6, 5.7 Hz, 1H), 4.97 (dd, *J* = 33.0, 13.6 Hz, 2H), 3.82 (d, *J* = 5.5 Hz, 2H), 2.38 (s, 3H).

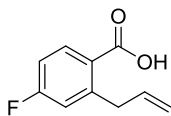
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 173.8, 139.8, 138.5, 136.3, 134.9, 129.4, 126.1, 115.1, 34.1, 19.9.



**2-allyl-5-methoxybenzoic acid<sup>5</sup>, yellow oil.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.58 (s, 1H), 7.22 (d, *J* = 8.5 Hz, 1H), 7.06 (d, *J* = 8.5 Hz, 1H), 6.03 (dq, *J* = 15.8, 6.6 Hz, 1H), 5.08 – 4.96 (m, 2H), 3.85 (s, 3H), 3.77 (d, *J* = 6.1 Hz, 2H).

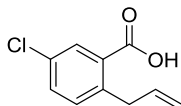
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 172.7, 157.8, 137.7, 134.9, 132.3, 128.9, 119.6, 115.8, 115.4, 55.5, 37.8.



**2-allyl-4-fluorobenzoic acid<sup>5</sup>, white solid.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.11 (dd, *J* = 8.5, 6.0 Hz, 1H), 7.01 (ddd, *J* = 9.7, 7.0, 3.9 Hz, 2H), 6.02 (ddt, *J* = 16.8, 10.2, 6.6 Hz, 1H), 5.17 – 5.02 (m, 2H), 3.84 (d, *J* = 6.5 Hz, 2H).

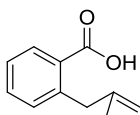
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 171.9, 167.2, 163.8, 146.7, 136.3, 134.5, 124.1, 118.0, 117.7, 116.6, 113.6, 113.3, 38.5.



**2-allyl-5-chlorobenzoic acid<sup>5</sup>, baby blue oil.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 9.84 (s, 1H), 8.04 (s, 1H), 7.44 (s, 1H), 7.31 (s, 1H), 6.01 (s, 1H), 5.04 (t, *J* = 11.5 Hz, 2H), 3.82 (s, 2H).

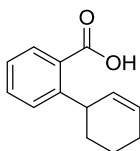
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 136.7, 133.1, 116.2.



**2-(2-methylallyl)benzoic acid<sup>6</sup>, colorless oil.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.05 (d, *J* = 7.8 Hz, 1H), 7.51 (t, *J* = 7.0 Hz, 1H), 7.32 (t, *J* = 7.8 Hz, 2H), 4.81 (s, 1H), 4.48 (s, 1H), 3.80 (s, 2H), 1.77 (s, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 145.4, 142.2, 132.8, 131., 131.5, 128.9, 126.3, 111.6, 41.8, 23.0.



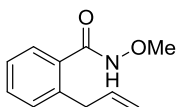
**1',2',3',4'-tetrahydro-[1,1'-biphenyl]-2-carboxylic acid, white solid.** *R<sub>f</sub>* = 0.44 (PE/EA = 5:1)

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.00 (d, *J* = 7.8 Hz, 1H), 7.51 (t, *J* = 7.4 Hz, 1H), 7.43 (d, *J* = 7.8 Hz, 1H), 7.30 (d, *J* = 7.5 Hz, 1H), 5.94 (d, *J* = 9.9 Hz, 1H), 5.67 (d, *J* = 10.0 Hz, 1H), 4.46 (s, 1H), 2.27 – 2.03 (m, 3H), 1.82 – 1.60 (m, 2H), 1.51 (q, *J* = 9.9 Hz, 1H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 173.3, 148.9, 132.7, 131.2, 130.3, 128.6, 128.2, 125.9, 37.8, 32.3, 25.0, 21.3.

IR(neat): *v*<sub>max</sub> 2932, 1691, 1573, 1412, 1315, 1307, 931, 762, 649 cm<sup>-1</sup>.

HRMS (ESI) *m/z* calcd for C<sub>13</sub>H<sub>14</sub>O<sub>2</sub> [M-H]<sup>-</sup>: 201.0921, found: 201.0963.

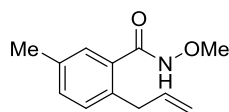


**1a: 2-allyl-N-methoxybenzamide<sup>2b, 7</sup>, white solid.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.43 (s, 1H), 7.46 – 7.33 (m, 2H), 7.27 (d, *J* = 1.5 Hz, 1H),

7.23 (d,  $J = 7.6$  Hz, 1H), 6.00 (ddt,  $J = 16.6, 10.1, 6.3$  Hz, 1H), 5.15 – 4.95 (m, 2H), 3.88 (s, 3H), 3.56 (d,  $J = 6.3$  Hz, 2H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform- $d$ )  $\delta$  138.4, 137.4, 132.6, 130.9, 127.7, 126.4, 116.38, 64.7, 37.3.



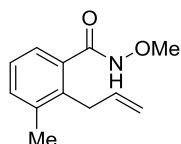
**1b: 2-allyl-*N*-methoxy-5-methylbenzamide, white solid.**  $R_f = 0.31$  (PE/EA= 3:1 )

$^1\text{H NMR}$  (300 MHz, Chloroform- $d$ )  $\delta$  8.38 (s, 1H), 7.20 (s, 2H), 7.14 (d,  $J = 8.1$  Hz, 1H), 5.99 (ddt,  $J = 16.5, 11.2, 6.5$  Hz, 1H), 5.04 (dd,  $J = 22.5, 13.6$  Hz, 2H), 3.87 (s, 3H), 3.52 (d,  $J = 6.1$  Hz, 2H), 2.33 (s, 3H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform- $d$ )  $\delta$  137.7, 136.1, 135.2, 132.5, 131.5, 130.6, 128.3, 116.1, 64.6, 37.0, 20.8.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  3143, 2978, 1645, 1607, 1320, 1049, 948, 944, 830  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{12}\text{H}_{15}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 206.1176, found: 206.0818.



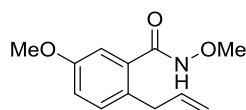
**1c: 2-allyl-*N*-methoxy-3-methylbenzamide, white solid.**  $R_f = 0.18$  (PE/EA= 3:1 )

$^1\text{H NMR}$  (300 MHz, Chloroform- $d$ )  $\delta$  8.49 (s, 1H), 7.22 (d,  $J = 7.8$  Hz, 2H), 7.19 – 7.11 (m, 1H), 5.99 (ddt,  $J = 15.8, 10.7, 5.6$  Hz, 1H), 5.06 (dq,  $J = 10.2, 1.7$  Hz, 1H), 4.86 (dd,  $J = 17.2, 1.7$  Hz, 1H), 3.86 (s, 3H), 3.52 (dt,  $J = 5.5, 1.7$  Hz, 2H), 2.31 (s, 3H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform- $d$ )  $\delta$  168.2, 138.2, 136.5, 135.5, 133.7, 132.6, 126.4, 125.6, 115.7, 64.6, 34.1, 19.6.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  3143, 2974, 2822, 1636, 1446, 1324, 1058, 806, 784, 602  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{12}\text{H}_{15}\text{NO}_2$   $[\text{M}-\text{OMe}]^+$ : 174.0913, found: 174.0562.



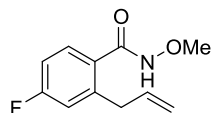
**1d: 2-allyl-*N*,5-dimethoxybenzamide, orange solid.**  $R_f = 0.13$  (PE/EA= 3:1 )

$^1\text{H NMR}$  (300 MHz, Chloroform- $d$ )  $\delta$  8.44 (d,  $J = 30.1$  Hz, 1H), 7.16 (d,  $J = 8.6$  Hz, 1H), 6.93 (s, 2H), 6.01 (dt,  $J = 14.7, 7.2$  Hz, 1H), 5.03 (dd,  $J = 28.1, 13.5$  Hz, 2H), 3.87 (s, 3H), 3.80 (s, 3H), 3.48 (d,  $J = 4.3$  Hz, 2H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform- $d$ )  $\delta$  157.9, 133.6, 131.8, 129.9, 116.6, 116.0, 113.1, 64.7, 55.5, 36.5.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  3143, 2974, 2822, 1636, 1446, 1324, 1058, 806, 784, 602  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{12}\text{H}_{15}\text{NO}_3$   $[\text{M}+\text{H}]^+$ : 222.1125, found: 222.1251.



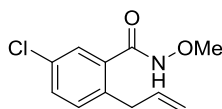
**1e: 2-allyl-4-fluoro-N-methoxybenzamide, white solid.**  $R_f = 0.14$  (PE/EA= 3:1 )

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.41 (s, 1H), 7.39 (dd,  $J = 8.4, 5.7$  Hz, 1H), 6.95 (qd,  $J = 9.1, 8.2, 2.5$  Hz, 2H), 5.97 (ddt,  $J = 16.6, 10.1, 6.4$  Hz, 1H), 5.17 – 4.99 (m, 2H), 3.87 (s, 3H), 3.56 (d,  $J = 6.4$  Hz, 2H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform-*d*)  $\delta$  141.9, 141.8, 136.4, 129.7, 128.8, 117.7, 117.4, 117.1, 113.5, 113.2, 64.7, 37.2.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  3426, 2323, 2940, 1653, 1607, 1505, 1248, 1041, 927, 822, 585  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{11}\text{H}_{12}\text{FNO}_2$   $[\text{M}+\text{H}]^+$ : 210.0925, found: 210.0929.



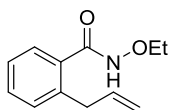
**1f: 2-allyl-5-chloro-N-methoxybenzamide, white solid.**  $R_f = 0.44$  (PE/EA= 3:1 )

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.51 (s, 1H), 7.43 – 7.30 (m, 2H), 7.20 (d,  $J = 7.7$  Hz, 1H), 6.04 – 5.86 (m, 1H), 5.05 (dd,  $J = 28.5, 13.6$  Hz, 2H), 3.86 (s, 3H), 3.51 (d,  $J = 6.0$  Hz, 2H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform-*d*)  $\delta$  136.9, 136.7, 134.1, 132.2, 132.0, 130.8, 127.7, 116.8, 64.7, 36.7.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  3231, 2978, 1666, 1505, 1315, 1054, 1037, 893, 767, 662  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{11}\text{H}_{12}\text{ClNO}_2$   $[\text{M}+\text{H}]^+$ : 226.0629, found: 226.0648.

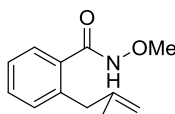


**1g: 2-allyl-N-ethoxybenzamide, yellow oil.**  $R_f = 0.38$  (PE/EA= 3:1 )

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.56 (s, 1H), 7.36 (d,  $J = 7.8$  Hz, 2H), 7.23 (dd,  $J = 11.6, 7.9$  Hz, 2H), 6.08 – 5.90 (m, 1H), 5.03 (dd,  $J = 20.0, 13.7$  Hz, 2H), 4.20 – 3.92 (m, 2H), 3.54 (d,  $J = 5.8$  Hz, 2H), 1.31 (s, 3H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform-*d*)  $\delta$  167.7, 138.4, 137.4, 132.9, 130.7, 130.5, 127.7, 126.3, 116.3, 37.3, 13.5.

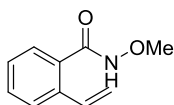
$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{12}\text{H}_{15}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 206.1176, found: 206.1194.



**1h: N-methoxy-2-(2-methylallyl)benzamide<sup>7</sup>, colorless oil.**

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.97 (s, 1H), 7.40 – 7.29 (m, 2H), 7.25 – 7.15 (m, 2H), 4.82 (s, 1H), 4.48 (s, 1H), 3.78 (s, 3H), 3.44 (s, 2H), 1.71 (s, 3H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform-*d*)  $\delta$  137.8, 133.1, 130.5, 128.0, 112.4, 64.4, 41.1, 22.8.

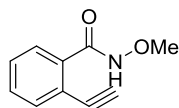


**1i: N-methoxy-2-vinylbenzamide<sup>8</sup>, white solid.**

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.36 (s, 1H), 7.62 – 7.55 (m, 1H), 7.49 – 7.38 (m, 2H), 7.34

– 7.27 (m, 1H), 7.03 (dd,  $J = 17.4, 11.0$  Hz, 1H), 5.74 (dd,  $J = 17.4, 0.9$  Hz, 1H), 5.39 (dd,  $J = 11.0, 0.9$  Hz, 1H), 3.89 (s, 3H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  133.9, 130.9, 127.8, 127.7, 126.4, 117.4, 64.7.

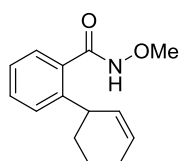


**1j: 2-ethynyl-N-methoxybenzamide, yellow oil.**  $R_f = 0.23$  (PE/EA= 3:1 )

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  9.74 (s, 1H), 7.83 (s, 1H), 7.58 – 7.48 (m, 1H), 7.47 – 7.36 (m, 2H), 3.86 (s, 3H), 3.47 (s, 1H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  164.8, 133.8, 130.9, 129.5, 129.4, 118.8, 83.5, 81.8, 64.5.

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{10}\text{H}_9\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 176.0706, found: 176.0714.



**1k: N-methoxy-1',2',3',4'-tetrahydro-[1,1'-biphenyl]-2-carboxamide, colorless oil.**

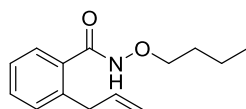
$R_f = 0.20$  (PE/EA= 3:1 )

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  8.46 (s, 1H), 7.35 (td,  $J = 20.7, 17.6, 8.1$  Hz, 3H), 7.20 (t,  $J = 6.7$  Hz, 1H), 5.90 (d,  $J = 9.8$  Hz, 1H), 5.62 (d,  $J = 10.0$  Hz, 1H), 3.88 (s, 3H), 3.82 (s, 1H), 2.07 (s, 3H), 1.66 (d,  $J = 26.4$  Hz, 2H), 1.57 – 1.40 (m, 1H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  168.1, 145.4, 130.6, 129.8, 128.8, 127.1, 125.9, 64.6, 38.2, 32.6, 24.9, 21.3.

IR(neat): $\nu_{\text{max}}$  3181, 2936, 1662, 1505, 1438, 1033, 881, 762, 657  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{14}\text{H}_{17}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 232.1332, found: 232.1329.



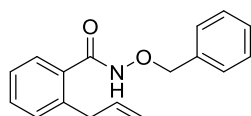
**1l: 2-allyl-N-butoxybenzamide, colorless oil.**  $R_f = 0.36$  (PE/EA= 3:1 )

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  8.33 (s, 1H), 7.38 (d,  $J = 7.3$  Hz, 2H), 7.23 (d,  $J = 8.2$  Hz, 1H), 6.00 (ddd,  $J = 16.3, 8.2, 4.1$  Hz, 1H), 5.05 (dd,  $J = 22.9, 13.6$  Hz, 2H), 4.03 (s, 2H), 3.56 (d,  $J = 5.9$  Hz, 2H), 1.79 – 1.63 (m, 2H), 1.53 – 1.34 (m, 2H), 0.95 (t,  $J = 6.6$  Hz, 3H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  138.3, 137.4, 132.9, 130.8, 127.7, 126.4, 116.3, 37.3, 30.1, 19.1, 13.9.

IR(neat): $\nu_{\text{max}}$  3185, 2966, 2873, 1653, 1514, 1315, 1159, 1033, 914, 754  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{14}\text{H}_{19}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 234.1489, found: 234.1490.



**1m: 2-allyl-N-(benzyloxy)benzamide, white solid.**  $R_f = 0.33$  (PE/EA= 3:1 )

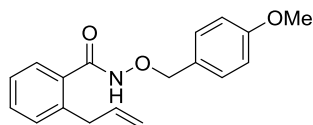
$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  8.22 (s, 1H), 7.44 (s, 2H), 7.42 – 7.28 (m, 5H), 7.23 (d,  $J =$



7.8 Hz, 2H), 5.91 (ddt,  $J = 16.7, 11.3, 6.6$  Hz, 1H), 5.17 – 4.85 (m, 4H), 3.51 (d,  $J = 6.0$  Hz, 2H).  
 $^{13}\text{C}$  NMR (75 MHz, Chloroform- $d$ )  $\delta$  138.3, 137.4, 132.7, 130.8, 130.6, 129.3, 128.9, 128.7, 127.8, 116.3, 37.3.

IR(neat): $\nu_{\text{max}}$  3181, 2974, 1640, 1501, 1455, 1307, 1159, 1024, 1020, 910, 750, 699, 509  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{17}\text{H}_{17}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 268.1332, found: 268.1321.



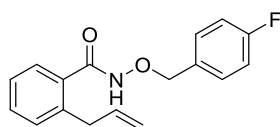
**1n: 2-allyl-*N*-((4-methoxybenzyl)oxy)benzamide, white solid.**  $R_f = 0.22$  (PE/EA = 3:1)

$^1\text{H}$  NMR (300 MHz, Chloroform- $d$ )  $\delta$  8.18 (s, 1H), 7.46 – 7.34 (m, 3H), 7.32 (d,  $J = 7.8$  Hz, 1H), 7.21 (d,  $J = 9.1$  Hz, 2H), 6.91 (d,  $J = 7.6$  Hz, 2H), 6.02 – 5.86 (m, 1H), 5.00 (q,  $J = 17.2, 13.7$  Hz, 4H), 3.82 (s, 3H), 3.52 (d,  $J = 5.8$  Hz, 2H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform- $d$ )  $\delta$  160.1, 138.4, 137.4, 132.8, 131.0, 130.8, 130.6, 127.7, 127.3, 126.4, 116.3, 114.0, 55.3, 37.3.

IR(neat): $\nu_{\text{max}}$  3185, 2953, 1649, 1615, 1518, 1256, 1176, 1033, 914, 826, 754, 526  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{18}\text{H}_{19}\text{NO}_3$   $[\text{M}+\text{Na}]^+$ : 320.1257, found: 320.1250.



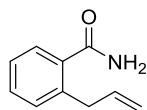
**1o: 2-allyl-*N*-((4-fluorobenzyl)oxy)benzamide, white solid.**  $R_f = 0.26$  (PE/EA = 3:1)

$^1\text{H}$  NMR (300 MHz, Chloroform- $d$ )  $\delta$  8.21 (s, 1H), 7.40 (dd,  $J = 15.2, 6.9$  Hz, 3H), 7.33 (d,  $J = 7.6$  Hz, 1H), 7.22 (d,  $J = 8.2$  Hz, 2H), 7.08 (t,  $J = 8.3$  Hz, 2H), 5.94 (ddt,  $J = 15.9, 11.1, 5.9$  Hz, 1H), 5.12 – 4.86 (m, 4H), 3.52 (d,  $J = 6.1$  Hz, 2H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform- $d$ )  $\delta$  164.7, 161.4, 132.6, 131.3, 131.2, 130.9, 130.7, 127.7, 126.4, 116.35, 115.8, 115.5, 37.3.

IR(neat): $\nu_{\text{max}}$  3189, 2982, 1649, 1598, 1514, 1227, 1159, 1020, 914, 830, 754, 513  $\text{cm}^{-1}$ .

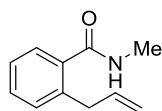
HRMS (ESI)  $m/z$  calcd for  $\text{C}_{17}\text{H}_{16}\text{FNO}_2$   $[\text{M}+\text{H}]^+$ : 286.1238, found: 286.1240.



**1p: 2-allylbenzamide<sup>9</sup>, colorless oil.**

$^1\text{H}$  NMR (300 MHz, Chloroform- $d$ )  $\delta$  8.03 (dd,  $J = 7.8, 1.2$  Hz, 1H), 7.56 (td,  $J = 7.6, 1.4$  Hz, 1H), 7.40 – 7.31 (m, 2H), 6.06 (ddt,  $J = 17.0, 10.7, 6.5$  Hz, 1H), 5.14 – 5.00 (m, 2H), 3.86 (d,  $J = 6.5$  Hz, 2H).

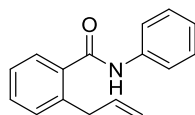
$^{13}\text{C}$  NMR (75 MHz, Chloroform- $d$ )  $\delta$  162.6, 144.0, 136.7, 133.7, 131.5, 131.4, 127.6, 126.5, 116.2, 38.4.



**1q: 2-allyl-*N*-methylbenzamide<sup>2</sup>, white solid.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.43 – 7.31 (m, 2H), 7.26 (s, 1H), 7.22 (d, *J* = 6.7 Hz, 1H), 6.01 (ddd, *J* = 16.5, 8.2, 4.4 Hz, 1H), 5.88 (s, 1H), 5.13 – 4.95 (m, 2H), 3.56 (d, *J* = 6.1 Hz, 2H).

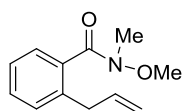
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 170.6, 137.7, 137.5, 136.6, 130.4, 130.0, 127.2, 126.3, 115.9, 37.6, 26.6.



**1r: 2-allyl-*N*-phenylbenzamide<sup>3</sup>, white solid.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.58 (dd, *J* = 15.4, 7.7 Hz, 4H), 7.46 – 7.27 (m, 5H), 7.20 – 7.12 (m, 1H), 6.09 (ddt, *J* = 16.4, 10.2, 6.1 Hz, 1H), 5.18 – 4.97 (m, 2H), 3.63 (d, *J* = 6.1 Hz, 2H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 137.9, 137.4, 136.7, 130.5, 129.1, 127.5, 126.6, 124.5, 119.8, 116.4, 37.6.



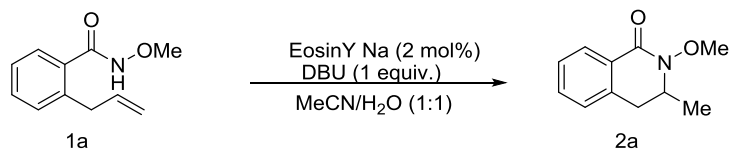
**2-allyl-*N*-methoxy-*N*-methylbenzamide<sup>10</sup>, white solid.**

<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.38 – 7.29 (m, 1H), 7.27 (s, 1H), 7.21 (d, *J* = 8.9 Hz, 2H), 6.02 – 5.84 (m, 1H), 5.14 – 5.01 (m, 2H), 3.80 – 3.07 (m, 8H).

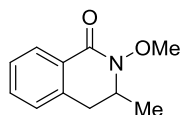
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 137.2, 136.6, 135.0, 129.5, 129.4, 126.5, 125.9, 77.4, 61.0, 37.4.

## 2.2 Synthesis and Characterization of Products

### I. Procedure for 3-methyl-3,4-dihydroisoquinolin-1(2H)-one



To a Schlenk tube were added **1a** (38.2 mg, 0.2 mmol), EosinY Na (3.14 mg, 0.004 mmol, 2 mol%) and charged with nitrogen for 15 minutes, anhydrous MeCN (1.5 mL) and H<sub>2</sub>O (1.5 mL) were added before DBU (0.3 mL, 0.2 mmol) via a syringe. The mixture was then placed around the 10 W Blue LEDs with a distance of 3 centimeters, and was stirred under visible light irradiation until the substrate was consumed (monitored by TLC), filtered, the solvent was removed by rotary evaporation and the resulting residue was purified directly by flash column chromatograph to give the desired product **2a** 86% yield.



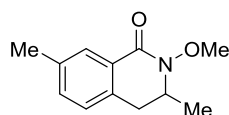
#### **2a: 2-methoxy-3-methyl-3,4-dihydroisoquinolin-1(2H)-one. White solid.**

$R_f$  = 0.20 (PE/EA = 3:1), Yield = 86%.

**<sup>1</sup>H NMR** (300 MHz, Chloroform-*d*)  $\delta$  8.12 (dd,  $J$  = 7.7, 1.5 Hz, 1H), 7.44 (td,  $J$  = 7.4, 1.5 Hz, 1H), 7.37 – 7.30 (m, 1H), 7.19-7.14 (m, 1H), 4.11 (p,  $J$  = 6.1 Hz, 1H), 3.90 (d,  $J$  = 2.1 Hz, 3H), 3.33 (dd,  $J$  = 16.0, 5.5 Hz, 1H), 2.90 (dd,  $J$  = 16.1, 5.8 Hz, 1H), 1.32 (dd,  $J$  = 6.6, 2.1 Hz, 3H).

**<sup>13</sup>C NMR** (75 MHz, Chloroform-*d*)  $\delta$  163.8, 136.0, 132.3, 128.3, 128.2, 127.7, 127.1, 77.5, 77.1, 76.7, 63.0, 54.9, 36.3, 17.6.

**HRMS** (ESI)  $m/z$  calcd for C<sub>11</sub>H<sub>14</sub>NO<sub>2</sub> [M+H]<sup>+</sup>: 192.1019, found: 192.1025.



#### **2b: 2-methoxy-3,7-dimethyl-3,4-dihydroisoquinolin-1(2H)-one. colorless oil.**

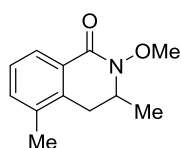
$R_f$  = 0.36 (PE/EA = 3:1), Yield = 72%.

**<sup>1</sup>H NMR** (300 MHz, Chloroform-*d*)  $\delta$  7.93 (s, 1H), 7.23 (s, 1H), 7.05 (d,  $J$  = 7.6 Hz, 1H), 4.09 (dt,  $J$  = 11.7, 6.5 Hz, 1H), 3.89 (s, 3H), 3.28 (dd,  $J$  = 16.0, 5.1 Hz, 1H), 2.85 (dd,  $J$  = 15.9, 5.5 Hz, 1H), 2.37 (s, 3H), 1.31 (d,  $J$  = 6.3 Hz, 3H).

**<sup>13</sup>C NMR** (75 MHz, Chloroform-*d*)  $\delta$  163.9, 136.8, 133.1, 133.0, 128.5, 128.1, 62.9, 55.0, 36.0, 21.0, 17.6.

**IR**(neat): $\nu_{\text{max}}$  2978, 2970, 1670, 1607, 1467, 1383, 1298, 1016, 741 cm<sup>-1</sup>.

**HRMS** (ESI)  $m/z$  calcd for C<sub>12</sub>H<sub>16</sub>NO<sub>2</sub> [M+H]<sup>+</sup>: 206.1176, found: 206.1373.



**2c: 2-methoxy-3,5-dimethyl-3,4-dihydroisoquinolin-1(2H)-one. colorless oil.**

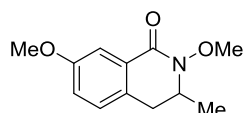
$R_f = 0.36$  (PE/EA = 3:1), Yield = 73%.

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.01 (d,  $J = 7.4$  Hz, 1H), 7.31 (d,  $J = 6.7$  Hz, 1H), 7.23 (d,  $J = 7.6$  Hz, 1H), 4.08 (h,  $J = 6.3$  Hz, 1H), 3.89 (s, 3H), 3.16 (d,  $J = 5.6$  Hz, 1H), 2.86 (dd,  $J = 16.4, 6.2$  Hz, 1H), 2.28 (s, 3H), 1.34 (d,  $J = 6.4$  Hz, 3H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform-*d*)  $\delta$  164.4, 135.3, 134.5, 133.9, 128.4, 126.7, 126.17, 63.0, 54.3, 33.5, 19.1, 18.1.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  2978, 2974, 2274, 1670, 1615, 1425, 1037, 919, 729  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{12}\text{H}_{16}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 206.1176, found: 206.1349.



**2d: 2,7-dimethoxy-3-methyl-3,4-dihydroisoquinolin-1(2H)-one. colorless oil.**

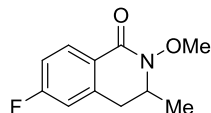
$R_f = 0.24$  (PE/EA = 3:1), Yield = 63%.

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  7.63 (s, 1H), 7.07 (d,  $J = 8.2$  Hz, 1H), 6.99 (d,  $J = 8.3$  Hz, 1H), 4.06 (p,  $J = 5.6$  Hz, 1H), 3.89 (s, 3H), 3.84 (s, 3H), 3.25 (dd,  $J = 15.8, 5.1$  Hz, 1H), 2.82 (dd,  $J = 15.9, 5.5$  Hz, 1H), 1.31 (d,  $J = 6.2$  Hz, 3H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform-*d*)  $\delta$  163.7, 129.3, 128.9, 128.1, 119.9, 111.2, 63.0, 55.6, 55.1, 35.5, 17.6.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  2923, 2852, 1674, 1594, 1383, 1303, 1024, 746  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{12}\text{H}_{16}\text{NO}_3$   $[\text{M}+\text{H}]^+$ : 222.1125, found: 222.1059.



**2e: 6-fluoro-2-methoxy-3-methyl-3,4-dihydroisoquinolin-1(2H)-one. colorless oil.**

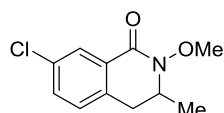
$R_f = 0.36$  (PE/EA = 3:1), Yield = 78%.

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.12 (dd,  $J = 8.7, 5.8$  Hz, 1H), 7.01 (td,  $J = 8.6, 2.5$  Hz, 1H), 6.86 (dd,  $J = 8.8, 2.5$  Hz, 1H), 4.09 (h,  $J = 6.0$  Hz, 1H), 3.89 (s, 3H), 3.31 (dd,  $J = 16.2, 5.4$  Hz, 1H), 2.88 (dd,  $J = 16.2, 5.8$  Hz, 1H), 1.32 (d,  $J = 6.4$  Hz, 3H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform-*d*)  $\delta$  166.8, 163.5, 163.1, 139.0, 138.8, 131.1, 130.9, 124.7, 124.7, 114.6, 114.5, 114.3, 114.3, 63.1, 54.8, 36.3, 17.6.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  2974, 2847, 2248, 1716, 1674, 1615, 1505, 1429, 1277, 1028, 914, 729  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{12}\text{H}_{13}\text{FNO}_2$   $[\text{M}+\text{H}]^+$ : 210.0925, found: 210.1299.



**2f: 7-chloro-2-methoxy-3-methyl-3,4-dihydroisoquinolin-1(2H)-one. colorless oil.**

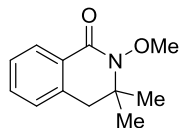
$R_f = 0.37$  (PE/EA = 3:1), Yield = 62%.

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.08 (s, 1H), 7.39 (d,  $J = 8.1$  Hz, 1H), 7.11 (d,  $J = 7.9$  Hz, 1H), 4.09 (h,  $J = 6.8, 6.3$  Hz, 1H), 3.89 (s, 3H), 3.29 (dd,  $J = 16.0, 5.1$  Hz, 1H), 2.86 (dd,  $J = 15.9, 5.2$  Hz, 1H), 1.31 (d,  $J = 6.2$  Hz, 3H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  162.4, 133.3, 132.3, 129.2, 128.1, 63.0, 54.8, 35.7, 17.6.

IR(neat): $\nu_{\text{max}}$  2978, 2974, 2847, 1678, 1611, 1497, 1260, 1024, 754  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{11}\text{H}_{13}\text{ClNO}_2$   $[\text{M}+\text{H}]^+$ : 226.0629, found: 226.0804.



**2h: 2-methoxy-3,3-dimethyl-3,4-dihydroisoquinolin-1(2H)-one. colorless oil.**

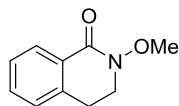
$R_f$  = 0.43 (PE/EA = 3:1), Yield = 66%.

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  8.12 (dd,  $J$  = 7.7, 1.3 Hz, 1H), 7.44 (td,  $J$  = 7.4, 1.5 Hz, 1H), 7.35 (d,  $J$  = 7.5 Hz, 1H), 7.14 (d,  $J$  = 7.5 Hz, 1H), 3.93 (s, 3H), 3.06 (s, 2H), 1.38 (s, 6H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  136.1, 132.4, 128.2, 127.6, 126.9, 64.4, 62.0, 43.3, 24.8.

IR(neat): $\nu_{\text{max}}$  2982, 2936, 1683, 1611, 1471, 1463, 1033, 952, 750, 708  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{12}\text{H}_{16}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 206.1176, found: 206.1095.



**2i: 2-methoxy-3,4-dihydroisoquinolin-1(2H)-one, colorless oil.**

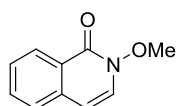
$R_f$  = 0.27 (PE/EA = 2:1), Yield = 49%.

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  8.14 (dd,  $J$  = 7.7, 1.3 Hz, 1H), 7.44 (td,  $J$  = 7.4, 1.5 Hz, 1H), 7.39 – 7.32 (m, 1H), 7.19 (d,  $J$  = 7.5 Hz, 1H), 3.90 (s, 3H), 3.83 (t,  $J$  = 6.7 Hz, 2H), 3.17 (t,  $J$  = 6.7 Hz, 2H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  163.8, 137.3, 132.2, 128.7, 128.4, 127.2, 62.1, 47.9, 29.2.

IR(neat): $\nu_{\text{max}}$  2978, 2932, 1678, 1611, 1463, 1332, 1083, 902, 746, 695, 686  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{10}\text{H}_{12}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 178.0863, found: 178.1065.



**2j: 2-methoxyisoquinolin-1(2H)-one. colorless oil.**

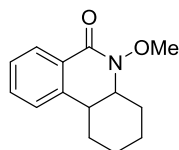
$R_f$  = 0.24 (PE/EA = 3:1), Yield = 28%.

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  8.47 (d,  $J$  = 8.0 Hz, 1H), 7.66 (t,  $J$  = 7.5 Hz, 1H), 7.52 (dd,  $J$  = 14.1, 7.3 Hz, 2H), 7.34 (d,  $J$  = 7.2 Hz, 1H), 6.48 (d,  $J$  = 7.6 Hz, 1H), 4.11 (s, 3H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  158.2, 136.2, 132.3, 129.4, 126.9, 126.2, 105.60, 64.4, 29.7.

IR(neat): $\nu_{\text{max}}$  2919, 2847, 1678, 1311, 1235, 1020, 935, 927, 746  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{10}\text{H}_{10}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 176.0706, found: 176.0877.



**2k: 5-methoxy-1,3,4,4a,5,10b-hexahydrophenanthridin-6(2H)-one. white solid.**

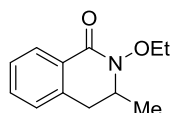
$R_f = 0.40$  (PE/EA = 3:1), Yield = 48%.

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.15 (d,  $J = 7.7$  Hz, 1H), 7.48 (t,  $J = 7.5$  Hz, 1H), 7.35 (t,  $J = 7.5$  Hz, 1H), 7.23 (s, 1H), 3.99 (d,  $J = 5.8$  Hz, 1H), 3.92 (s, 3H), 3.29 (s, 1H), 2.17 (s, 1H), 1.91 – 1.65 (m, 4H), 1.56 – 1.41 (m, 3H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform-*d*)  $\delta$  163.7, 132.5, 128.6, 128.5, 126.9, 125.8, 62.4, 59.3, 40.0, 29.7, 28.2, 26.5, 22.6.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  2919, 2852, 1391, 1020, 872, 674, 475  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{14}\text{H}_{18}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 232.1332, found: 232.1928.



**2g: 2-ethoxy-3-methyl-3,4-dihydroisoquinolin-1(2H)-one, colorless oil.**

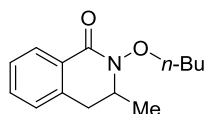
$R_f = 0.56$  (PE/EA = 3:1), Yield = 61%.

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.11 (d,  $J = 7.7$  Hz, 1H), 7.43 (t,  $J = 7.4$  Hz, 1H), 7.33 (t,  $J = 7.5$  Hz, 1H), 7.16 (d,  $J = 7.4$  Hz, 1H), 4.10 (ddd,  $J = 14.0, 10.1, 6.8$  Hz, 3H), 3.34 (dd,  $J = 15.9, 5.2$  Hz, 1H), 2.87 (dd,  $J = 16.1, 5.0$  Hz, 1H), 1.36 – 1.29 (m, 6H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform-*d*)  $\delta$  163.9, 132.2, 128.5, 128.1, 127.7, 127.0, 76.6, 70.8, 55.4, 36.2, 17.6.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  2982, 2978, 2847, 2252, 1678, 1598, 1573, 1429, 1235, 1024, 914, 733  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{12}\text{H}_{16}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 206.1176, found: 206.1434.



**2l: 2-n-butoxy-3-methyl-3,4-dihydroisoquinolin-1(2H)-one, colorless oil.**

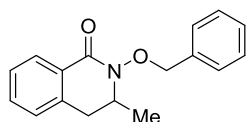
$R_f = 0.56$  (PE/EA = 3:1), Yield = 60%.

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.11 (d,  $J = 7.6$  Hz, 1H), 7.44 (t,  $J = 7.3$  Hz, 1H), 7.34 (t,  $J = 7.4$  Hz, 1H), 7.17 (d,  $J = 7.3$  Hz, 1H), 4.06 (t,  $J = 6.2$  Hz, 3H), 3.35 (dd,  $J = 16.0, 5.1$  Hz, 1H), 2.88 (dd,  $J = 16.0, 4.8$  Hz, 1H), 1.72 (p,  $J = 6.8$  Hz, 2H), 1.47 (q,  $J = 7.3$  Hz, 2H), 1.31 (d,  $J = 6.2$  Hz, 3H), 0.96 (t,  $J = 7.2$  Hz, 3H).

$^{13}\text{C NMR}$  (75 MHz, Chloroform-*d*)  $\delta$  163.7, 135.9, 132.2, 128.5, 128.1, 127.0, 75.1, 55.3, 36.2, 30.3, 19.3, 17.6, 14.0.

$\text{IR}(\text{neat})$ :  $\nu_{\text{max}}$  2962, 2936, 2873, 1678, 1467, 1379, 1298, 1016, 746  $\text{cm}^{-1}$ .

$\text{HRMS}$  (ESI)  $m/z$  calcd for  $\text{C}_{14}\text{H}_{19}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 234.1489, found: 234.1512.



**2m: 2-(benzyloxy)-3-methyl-3,4-dihydroisoquinolin-1(2H)-one, colorless oil.**

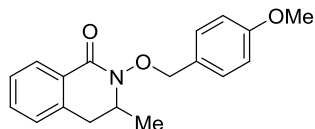
$R_f = 0.50$  (PE/EA = 3:1), Yield = 47%.

$^1\text{H NMR}$  (300 MHz, Chloroform-*d*)  $\delta$  8.14 (d,  $J = 7.7$  Hz, 1H), 7.51 (t,  $J = 3.9$  Hz, 2H), 7.48 – 7.30 (m, 5H), 7.15 (d,  $J = 7.5$  Hz, 1H), 5.09 (s, 2H), 3.79 (q,  $J = 6.0$  Hz, 1H), 3.19 (dd,  $J = 16.1, 5.7$  Hz, 1H), 2.74 (dd,  $J = 16.0, 4.9$  Hz, 1H), 1.28 – 1.24 (m, 3H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  163.9, 136.0, 135.7, 132.3, 129.8, 128.7, 128.5, 128.1, 127.8, 127.0, 56.0, 35.9, 29.7, 17.4.

IR(neat): $\nu_{\text{max}}$  2915, 2847, 1674, 1463, 910, 733, 522  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{17}\text{H}_{17}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 268.1332, found: 268.1355.



**2n: 2-((4-methoxybenzyl)oxy)-3-methyl-3,4-dihydroisoquinolin-1(2H)-one, colorless oil.**

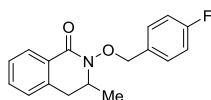
$R_f$  = 0.39 (PE/EA = 3:1), Yield = 45%.

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  8.14 (d,  $J$  = 7.7 Hz, 1H), 7.48 – 7.39 (m, 3H), 7.35 (t,  $J$  = 7.6 Hz, 1H), 7.14 (d,  $J$  = 7.5 Hz, 1H), 6.90 (dd,  $J$  = 8.7, 2.2 Hz, 2H), 5.09 – 4.96 (m, 2H), 3.82 (d,  $J$  = 2.3 Hz, 3H), 3.75 (q,  $J$  = 6.1 Hz, 1H), 3.18 (dd,  $J$  = 16.0, 5.6 Hz, 1H), 2.73 (dd,  $J$  = 16.0, 4.8 Hz, 1H), 1.25 (s, 3H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  163.7, 159.9, 136.0, 132.3, 131.4, 128.5, 128.1, 127.9, 127.8, 127.0, 113.8, 56.0, 55.3, 35.8, 29.7, 17.4.

IR(neat): $\nu_{\text{max}}$  3455, 1661, 1611, 1518, 1252, 910, 733, 644  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{18}\text{H}_{19}\text{NO}_3$   $[\text{M}+\text{Na}]^+$ : 320.1257, found: 320.1250.



**2o: 2-((4-fluorobenzyl)oxy)-3-methyl-3,4-dihydroisoquinolin-1(2H)-one, colorless oil.**

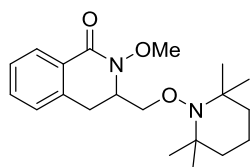
$R_f$  = 0.49 (PE/EA = 3:1), Yield = 43%.

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  8.18 – 8.08 (m, 1H), 7.57 – 7.41 (m, 3H), 7.36 (t,  $J$  = 7.7 Hz, 1H), 7.16 (d,  $J$  = 7.6 Hz, 1H), 7.07 (td,  $J$  = 7.7, 6.8, 1.9 Hz, 2H), 5.12 – 4.97 (m, 2H), 3.80 (p,  $J$  = 5.9 Hz, 1H), 3.21 (dd,  $J$  = 16.1, 5.6 Hz, 1H), 2.78 (dd,  $J$  = 16.1, 5.1 Hz, 1H), 1.25 (dd,  $J$  = 6.2, 2.1 Hz, 3H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  164.0, 136.0, 132.4, 131.7, 131.6, 128.4, 128.1, 127.8, 127.1, 56.03, 35.9, 17.5.

IR(neat): $\nu_{\text{max}}$  2919, 2852, 1670, 1607, 1509, 1463, 1374, 1227, 1155, 1016, 830, 737, 535, 530  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{17}\text{H}_{16}\text{FNO}_2$   $[\text{M}+\text{H}]^+$ : 286.1238, found: 286.1240.



**2-methoxy-3-(((2,2,6,6-tetramethylpiperidin-1-yl)oxy)methyl)-3,4-dihydroisoquinolin-1(2H)-one, white solid.**

$R_f$  = 0.27 (PE/EA = 3:1).

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  8.11 (dd,  $J$  = 7.7, 1.1 Hz, 1H), 7.40 (td,  $J$  = 7.4, 1.4 Hz, 1H),

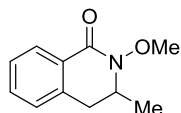
7.29 (t,  $J = 7.5$  Hz, 1H), 7.17 (d,  $J = 7.5$  Hz, 1H), 4.07 (ddd,  $J = 6.5, 4.7, 2.6$  Hz, 1H), 4.01 (dd,  $J = 8.9, 5.8$  Hz, 1H), 3.90 (s, 3H), 3.86 (dd,  $J = 8.8, 2.5$  Hz, 1H), 3.50 (dd,  $J = 16.3, 6.5$  Hz, 1H), 3.14 (dd,  $J = 16.3, 2.4$  Hz, 1H), 1.48 – 1.22 (m, 6H), 1.03 (d,  $J = 19.3$  Hz, 6H), 0.87 (s, 3H), 0.64 (s, 3H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  163.4, 136.3, 131.9, 128.9, 127.7, 127.0, 126.7, 75.1, 62.6, 59.9, 57.5, 39.6, 32.8, 32.8, 32.6, 20.0, 19.7, 16.9.

IR(neat): $\nu_{\text{max}}$  2923, 2847, 1383, 1020, 876, 674, 475, 471  $\text{cm}^{-1}$ .

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{20}\text{H}_{31}\text{N}_2\text{O}_3$   $[\text{M}+\text{H}]^+$ : 347.2329, found: 347.2160.

The purified product of  $\text{D}_2\text{O}$  experiment:



**2-methoxy-3-methyl-3,4-dihydroisoquinolin-1(2H)-one. White solid.**  $R_f = 0.20$  (PE/EA= 3:1 )

$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  8.12 (dd,  $J = 7.7, 1.3$  Hz, 1H), 7.44 (td,  $J = 7.4, 1.5$  Hz, 1H), 7.34 (t,  $J = 7.5$  Hz, 1H), 7.17 (d,  $J = 7.4$  Hz, 1H), 4.10 (h,  $J = 6.2$  Hz, 1H), 3.90 (d,  $J = 2.1$  Hz, 3H), 3.33 (dd,  $J = 16.0, 5.5$  Hz, 1H), 2.90 (dd,  $J = 16.0, 5.8$  Hz, 1H), 1.32 (d,  $J = 6.4$  Hz, 3H).

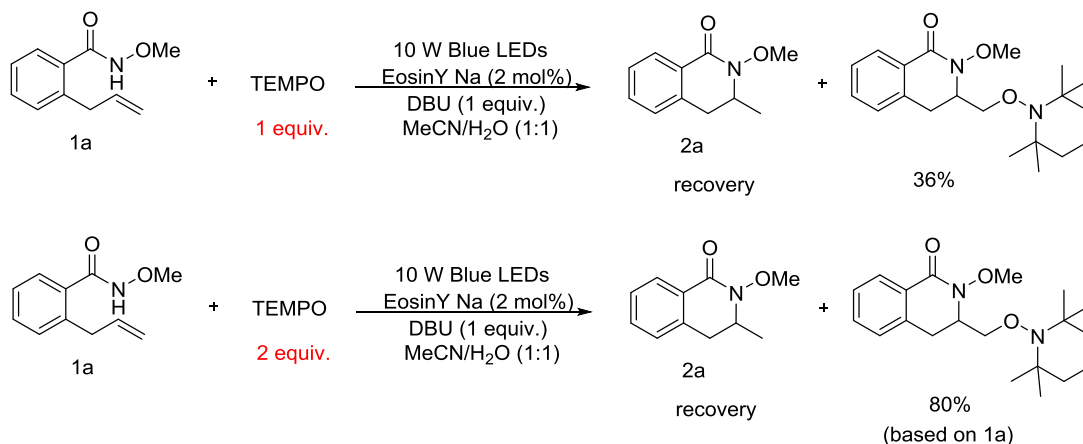
$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  163.8, 136.0, 132.3, 128.3, 128.2, 127.7, 127.0, 63.0, 54.9, 36.3, 17.7.

HRMS (ESI)  $m/z$  calcd for  $\text{C}_{11}\text{H}_{14}\text{NO}_2$   $[\text{M}+\text{H}]^+$ : 192.1019, found: 192.1020.



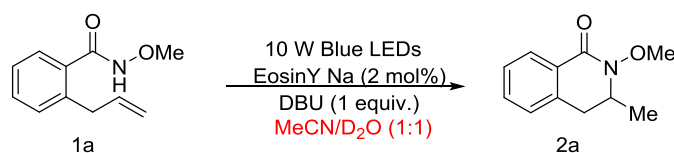
### 2.3. Mechanism Investigation.

#### (1) Radical trapping experiment



To a Schlenk tube were added **1a** (38.2 mg, 0.2 mmol), EosinY Na (3.14 mg, 0.004 mmol, 2 mol%), TEMPO (31.3 mg, 0.2 mmol, 1 equiv.) and charged with nitrogen for 15 minutes, anhydrous MeCN (1.5 mL) and H<sub>2</sub>O (1.5 mL) were added before DBU (0.03 mL, 0.2 mmol) via a syringe. The mixture was then placed around the 10 W Blue LEDs with a distance of 3 centimeters, and was stirred under visible light irradiation until the substrate was consumed (monitored by TLC), filtered, the solvent was removed by rotary evaporation and the resulting residue was purified directly by flash column chromatograph to give the 2-methoxy-3-(((2,2,6,6-tetramethylpiperidin-1-yl)oxy)methyl)-3,4-dihydroisoquinolin-1(2H)-one product as a white solid (25 mg, 36% yield);  $R_f = 0.27$ (Pet:EtOA = 3:1).

#### (2) D<sub>2</sub>O experiment



To a Schlenk tube were added **1a** (38.2 mg, 0.2 mmol), EosinY Na (3.14 mg, 0.004 mmol, 2 mol%) and charged with argon for 15 minutes, anhydrous MeCN (1.5 mL) and D<sub>2</sub>O (1.5 mL) were added before DBU (0.03 mL, 0.2 mmol) via a syringe. The mixture was then placed around the 10 W Blue LEDs with a distance of 3 centimeters, and was stirred under visible light irradiation until the substrate was consumed (monitored by TLC), filtered, the solvent was removed by rotary evaporation and the resulting residue was purified directly by flash column chromatograph to give the desired product **2a** 78%.  $M(\text{product}): 191.0946$

We provided an exact mass spectra of the purified product (Figure 1) and the reaction solution (Figure 2). of the D<sub>2</sub>O experiment. We carefully examined the mass spectrum of the mixture and don't found molecular weight of the various complexes such as the substrate and DBU+H.

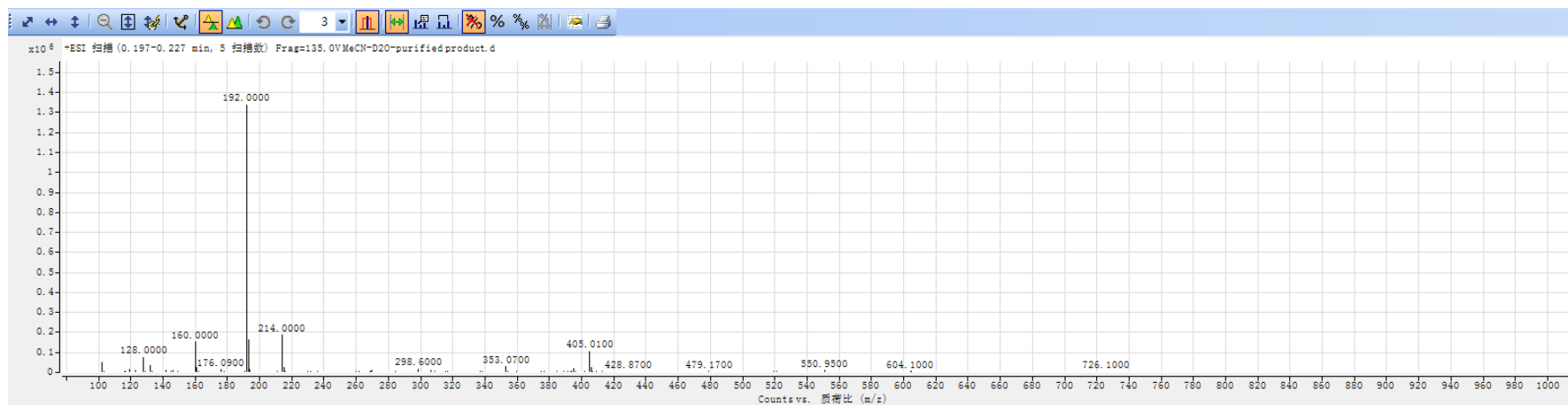


Figure 1. The mass spectra of the purified product

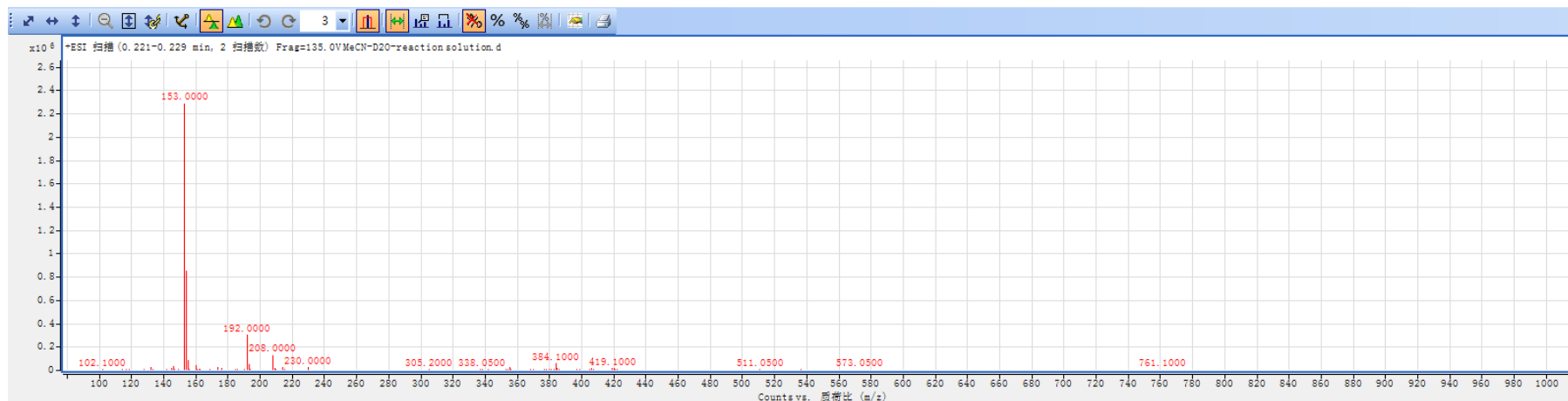
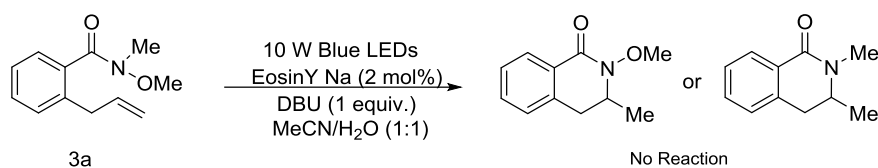


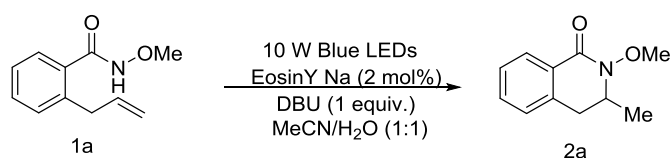
Figure 2. The mass spectra of the reaction solution

### (3) Proton-free experiment on nitrogen



To a Schlenk tube were added **3a** (41 mg, 0.2 mmol), EosinY Na (3.14 mg, 0.004 mmol, 2 mol%) and charged with nitrogen for 15 minutes, anhydrous MeCN (1.5 mL) and H<sub>2</sub>O (1.5 mL) were added before DBU (0.03 mL, 0.2 mmol) via a syringe. The mixture was then placed around the 10 W Blue LEDs with a distance of 3 centimeters, and was stirred under visible light irradiation (monitored by TLC), filtered, the solvent was removed by rotary evaporation and the resulting residue was purified directly by flash column chromatograph, the raw materials **3a** were almost completely recovered.

### 2.4. Gram-scale Reaction



To a Schlenk tube were added **1a** (0.573 g, 3 mmol), EosinY Na (47.1 mg, 0.06 mmol, 2 mol%) and charged with nitrogen for 15 minutes, anhydrous MeCN (10 mL) and H<sub>2</sub>O (10 mL) were added before DBU (0.45 mL, 3 mmol) via a syringe. The mixture was then placed around the two 10 W Blue LEDs with a distance of 3 centimeters, and was stirred under visible light irradiation until the substrate was consumed (monitored by TLC), filtered, the solvent was removed by rotary evaporation and the resulting residue was purified directly by flash column chromatograph to give the desired product **2a** 70%.

### 2.5 UV-Vis Spectral Analysis

(a) The emission spectra of the 10 W Blue LEDs

We measured the emission spectra of the 10 W Blue LEDs used in our experiments by AULTT-P series fiber optic spectrometer (Figure 3.). As we can see from Figure 3, the Blue LED used in the experiment has a maximum emission wavelength of 554 nm and is located in the blue region (435-480nm).

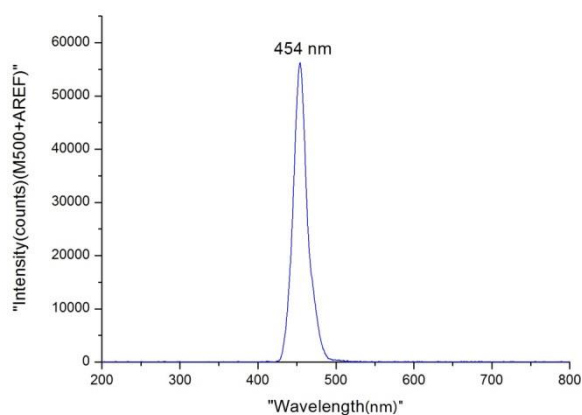


Figure 3. The emission spectra of 10 W Blue LEDs

(b) In order to understand the light absorption characteristics of this reaction system, the UV-Vis spectrum of the photocatalyst EosinY Na in MeCN/H<sub>2</sub>O (1:1), the UV-Vis spectrum of the reaction solution, the UV-Vis spectrum of substrate (1a) in MeCN/H<sub>2</sub>O (1:1) and the UV-Vis spectrum of DBU in MeCN/H<sub>2</sub>O (1:1) were measured by an UV-Vis spectrophotometer.

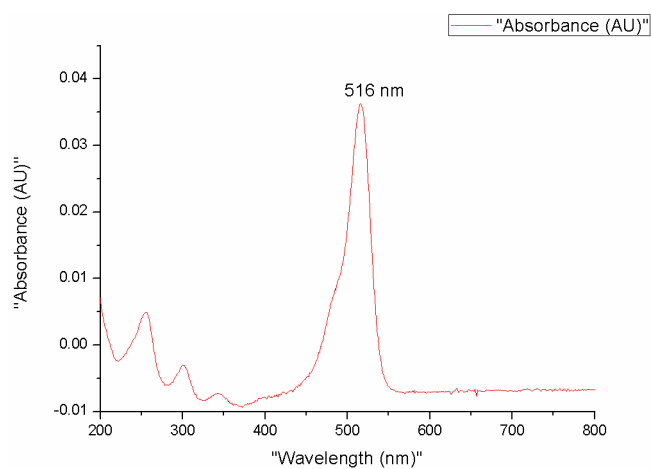


Figure 4. UV-Vis spectrum of EosinY Na in MeCN/H<sub>2</sub>O (1:1)

As shown in the figure, the UV-visible absorption spectrum of EosinY Na, which is easily soluble in MeCN/H<sub>2</sub>O (1:1). Its characteristic absorption wavelength is between 400-550 nm and the maximum absorption wavelength is 516 nm, located in the green area (492-560 nm).

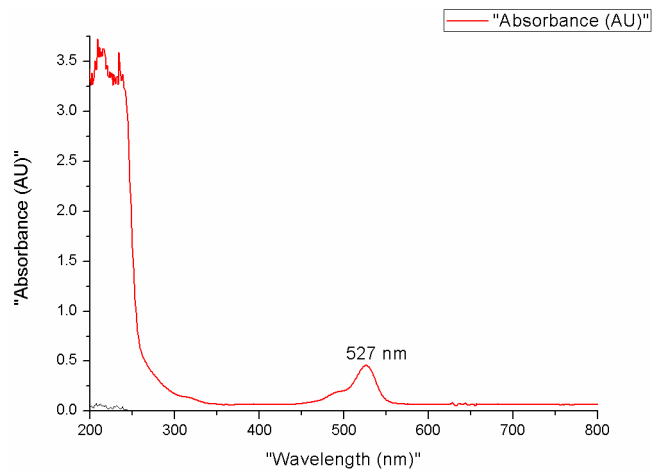


Figure 5. UV-Vis spectrum of the reaction solution

As shown in the figure, the characteristic absorption wavelength of the reaction solution is between 450-550 nm and the maximum absorption wavelength is 527 nm, located in the green area as well.

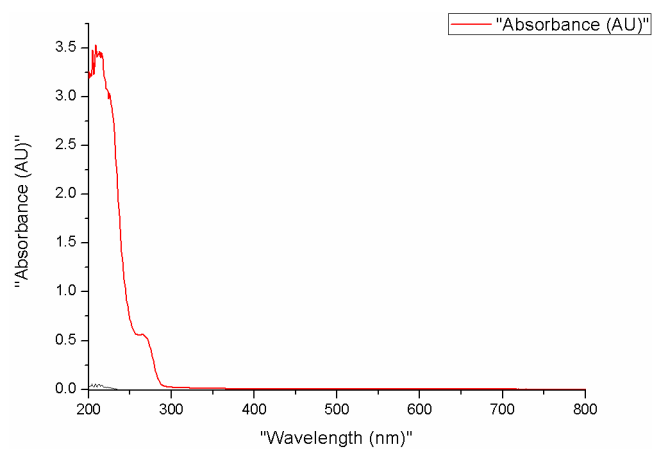


Figure 6. UV-Vis spectrum of substrate (1a) in MeCN/H<sub>2</sub>O (1:1)

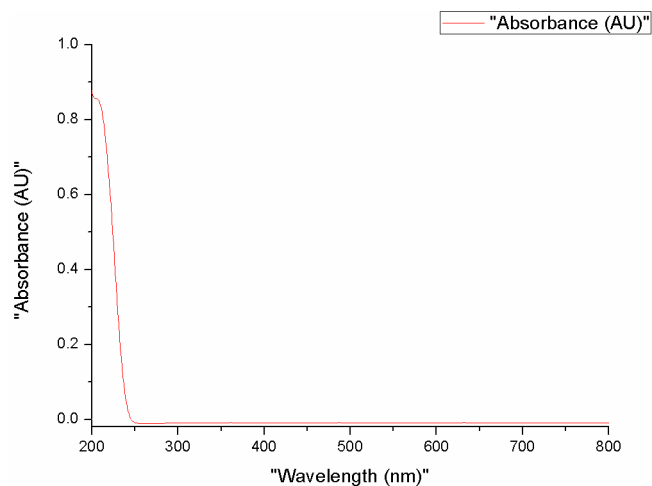


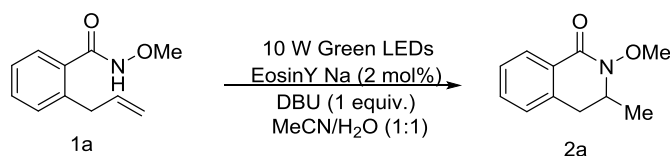
Figure 7. UV-Vis spectrum UV-Vis spectrum of DBU in MeCN/H<sub>2</sub>O(1:1)

Using 2-allyl-*N*-methoxybenzamide **1a** as a benchmark substrate. As shown in Figures 4-7 above, we examined the UV-visible absorption spectra of EosinY Na, reaction solution, substrate (**1a**) and DBU in the same solvent MeCN/H<sub>2</sub>O (1:1). The maximum absorption wavelength of EosinY Na in MeCN/H<sub>2</sub>O (1:1) is 516 nm (Figure 4). the maximum absorption wavelength of the reaction solution is 527 nm, substrate (**1a**) and DBU only absorb in the near-ultraviolet region in MeCN/H<sub>2</sub>O (1:1) (Figure 6 and 7). This indicates that only EosinY Na absorbs visible light in the our system to induce the reaction.

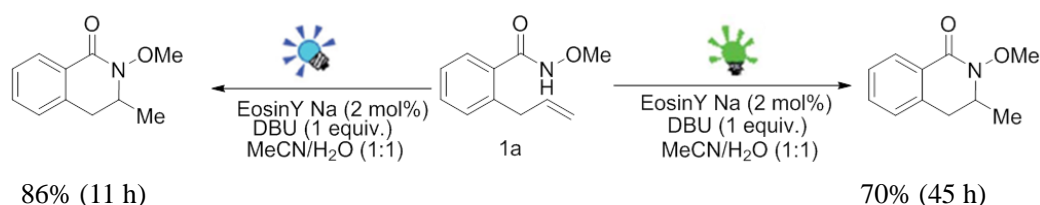
## 2.6 Comparison Experiment between Blue and Green Light Source

The characteristic absorption of the reaction system mainly occurs in the blue and green light range of the visible light region. Therefore, the effects of light sources provided by blue and green light on the reaction system are compared.

### (1) Green Light Experiment



To a Schlenk tube were added **1a** (38.2 mg, 0.2 mmol), EosinY Na (3.14 mg, 0.004 mmol, 2 mol%) and charged with argon for 15 minutes, anhydrous MeCN (1.5 mL) and H<sub>2</sub>O (1.5 mL) were added before DBU (0.03 mL, 0.2 mmol) via a syringe. The mixture was then placed around the 10 W Green LEDs with a distance of 3 centimeters, and was stirred under visible light irradiation until the substrate was consumed (monitored by TLC), filtered, the solvent was removed by rotary evaporation and the resulting residue was purified directly by flash column chromatograph to give the desired product **2a** (70% in 45 h vs 86% in 11 h).



### (2) Comparison Experiment between Blue and Green Light Source by HPLC

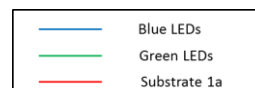
In order to more intuitively show the effect of blue and green light sources on the reaction, using HPLC time-lapse monitoring reaction.

#### HPLC spectrum

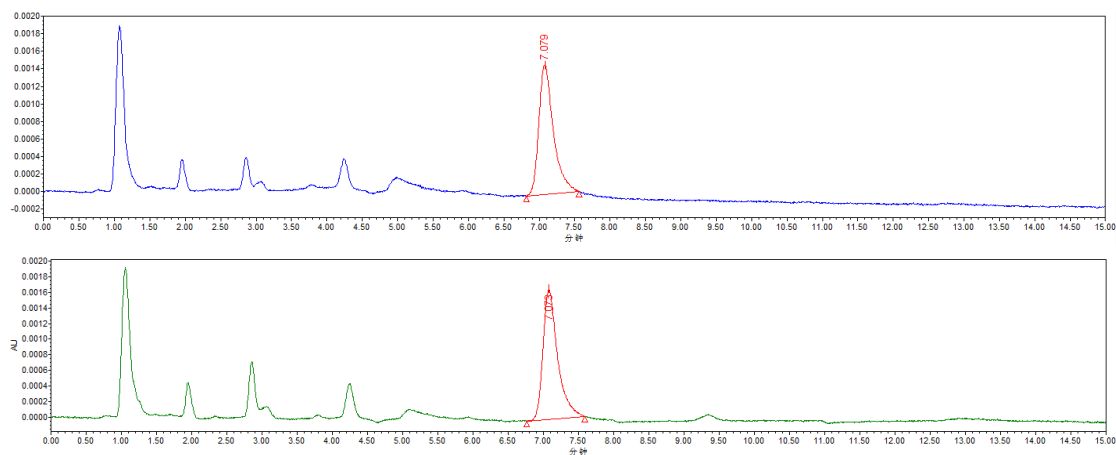
To two Schlenk tubes were added **1a** (38.2 mg, 0.2 mmol), EosinY Na (3.14 mg, 0.004 mmol, 2 mol%) and charged with argon for 15 minutes, anhydrous MeCN (1.5 mL) and H<sub>2</sub>O (1.5 mL) were added before DBU (0.03 mL, 0.2 mmol) via a syringe separately. The mixture was then placed separately around the 10 W Blue LEDs and 10 W Green LEDs with a distance of 3 centimeters, and was stirred under visible light irradiation until the substrate was consumed (monitored by HPLC, 30% MeCN in H<sub>2</sub>O as the eluent). Taking the reaction solution every two hours and observing the chromatogram change (As shown below).

Compared the liquid crystal spectra, it can be intuitively seen that the reaction rate of blue light as a light source is significantly higher than that of a green light source. At 11 hours, the reaction material for blue light irradiation is almost completely consumed, while the reaction for green light irradiation takes 45 hours.

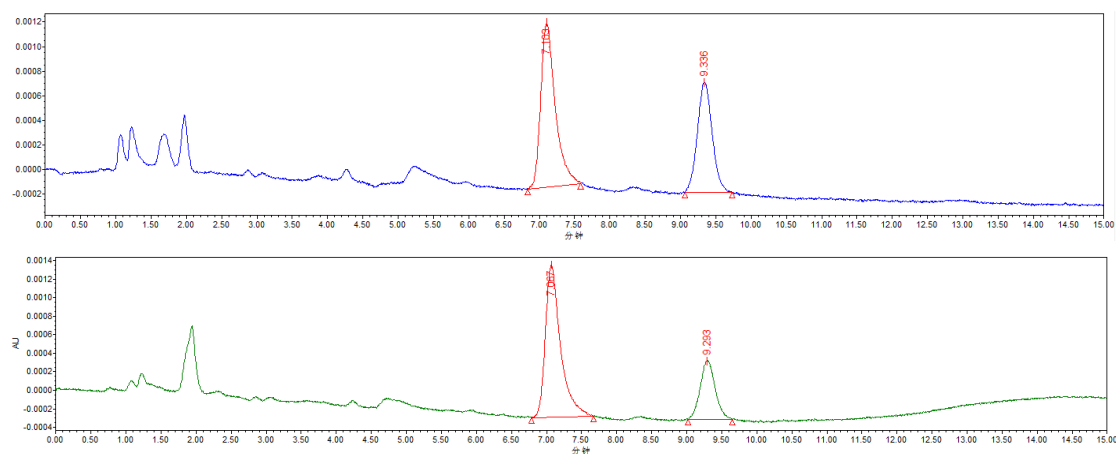
In summary, blue light is the most suitable light source in the reaction system we have reported.



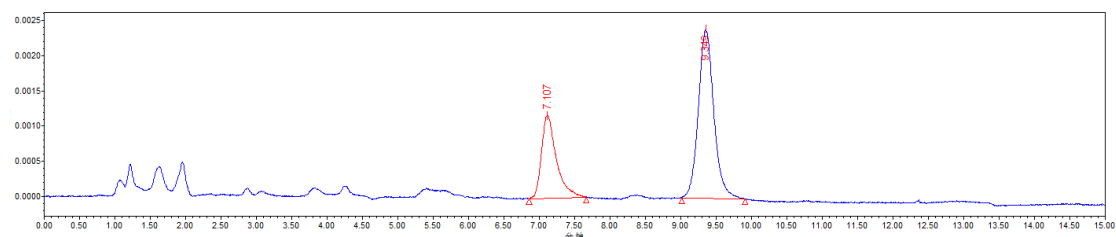
0 h:

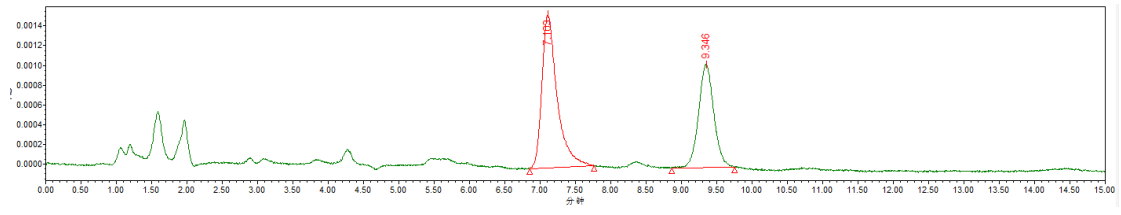


2 h:

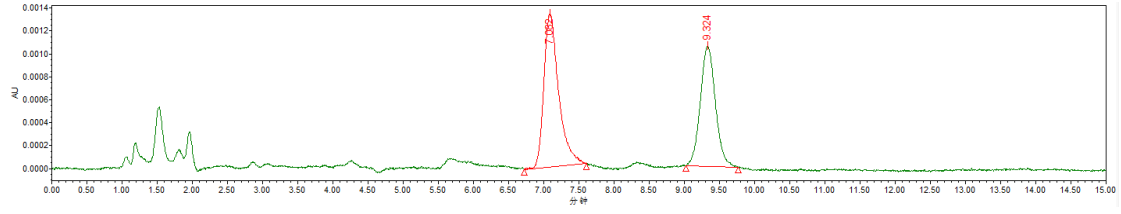
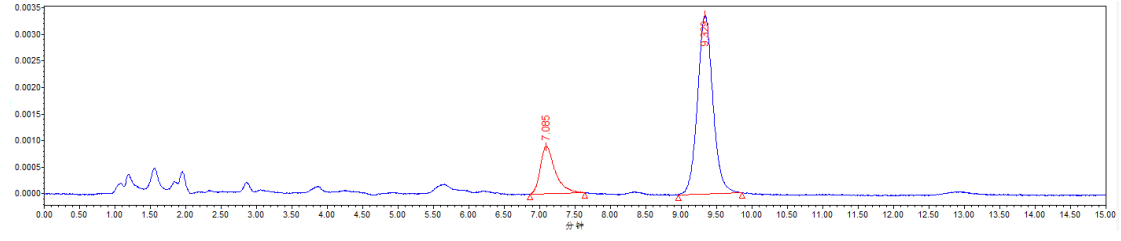


4 h:

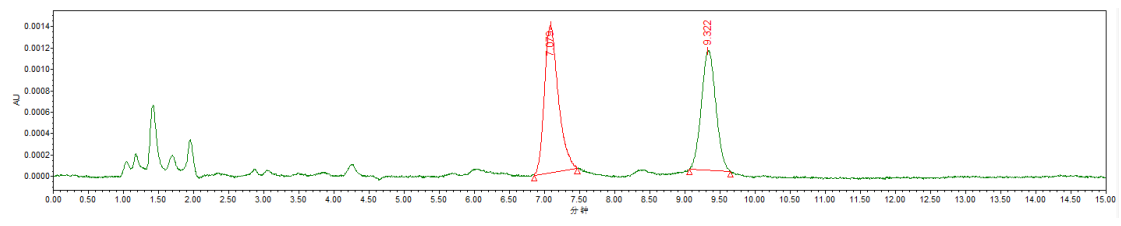
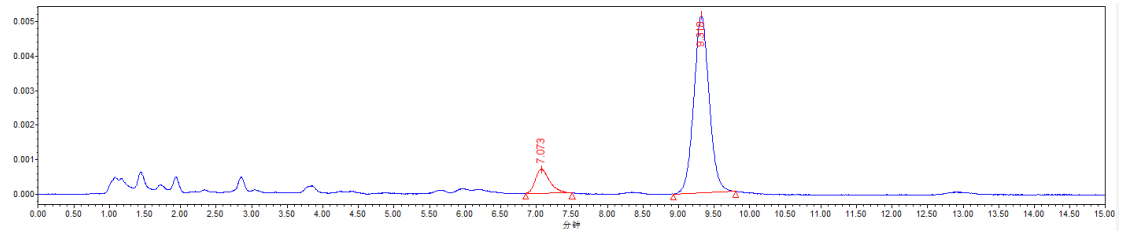




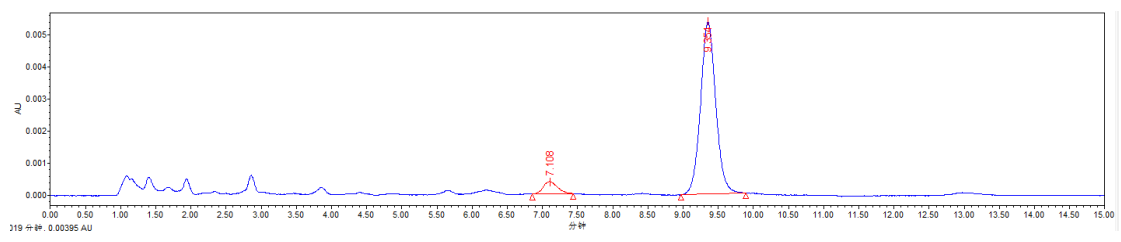
6 h:



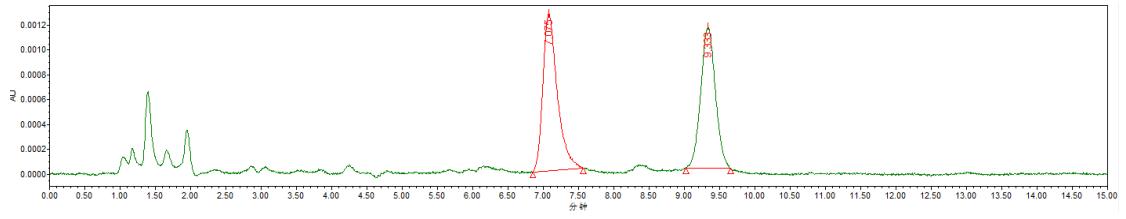
8 h:



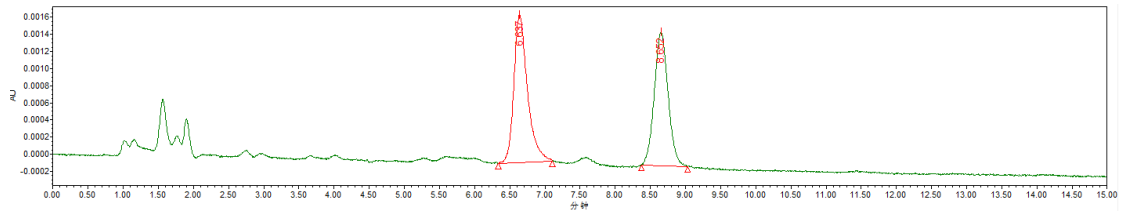
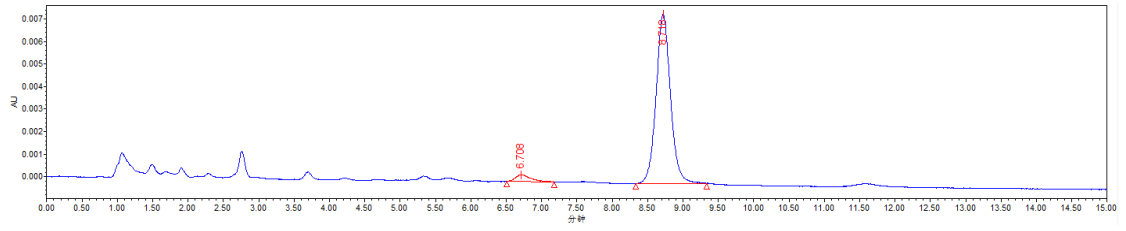
10 h:







11 h:



45 h:

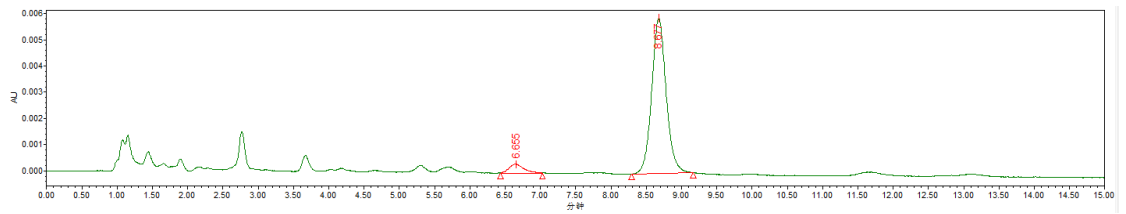


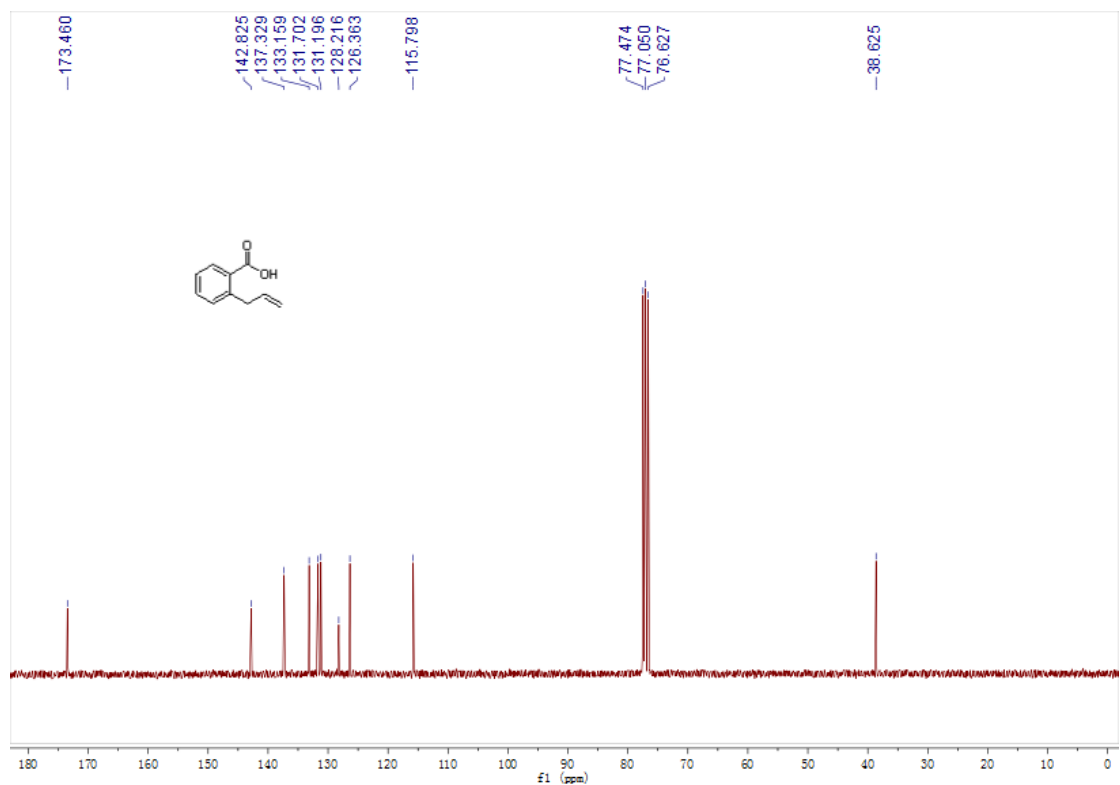
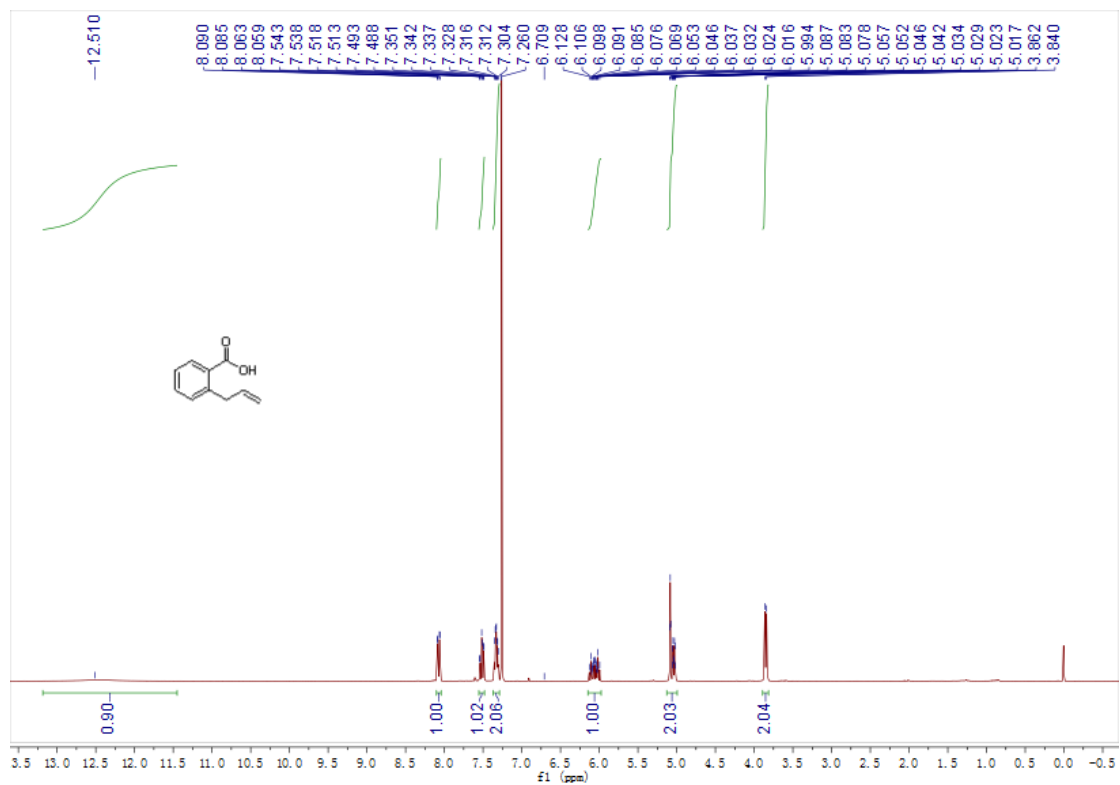
Figure 8. Comparison between Blue and Green Light Source by HPLC

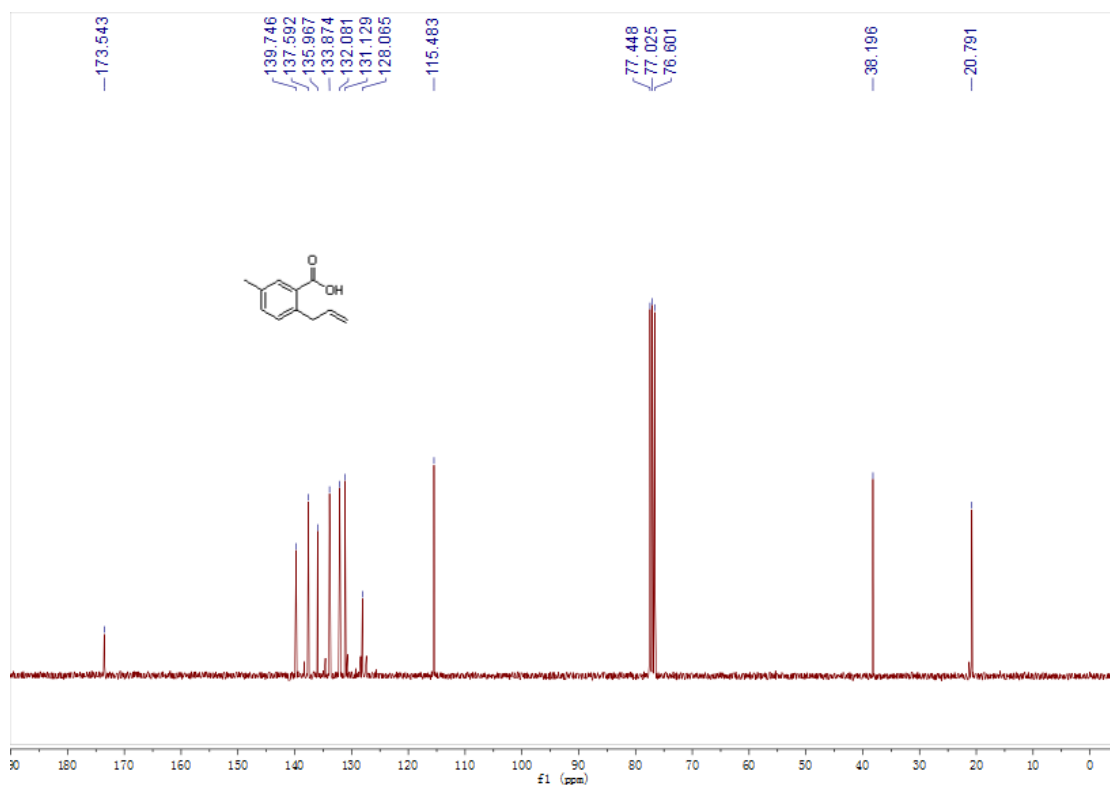
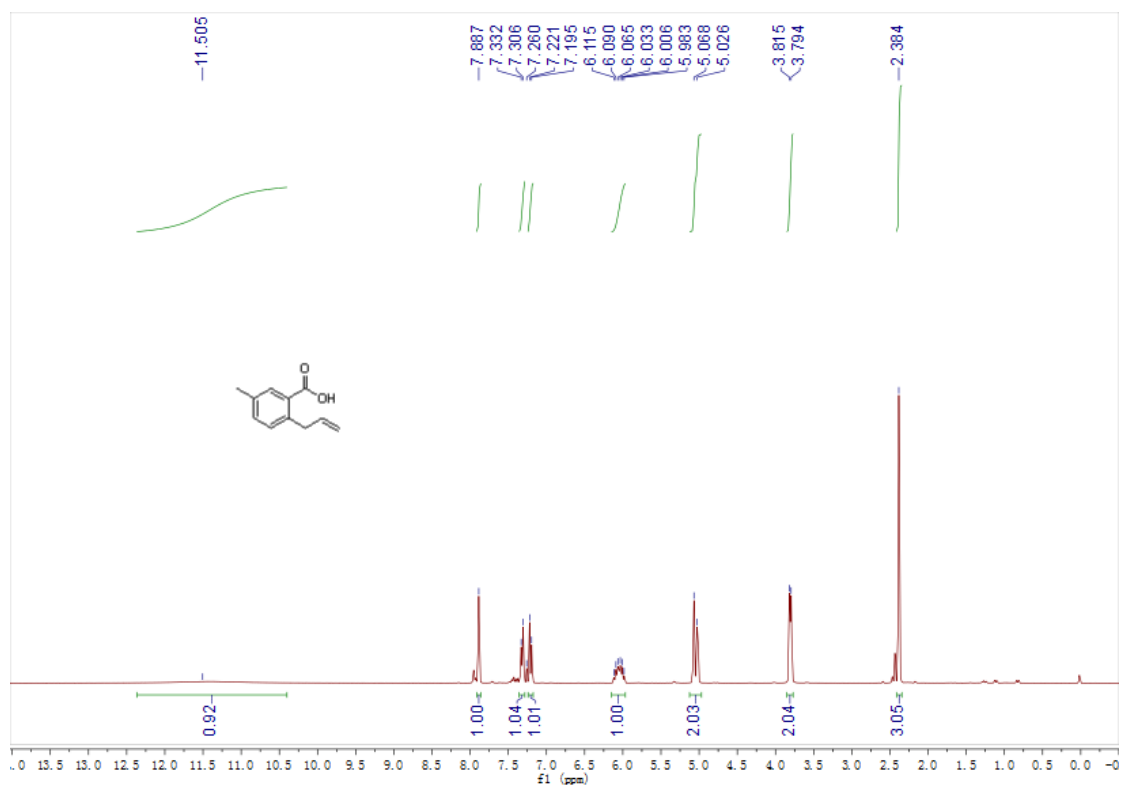
### 3. References

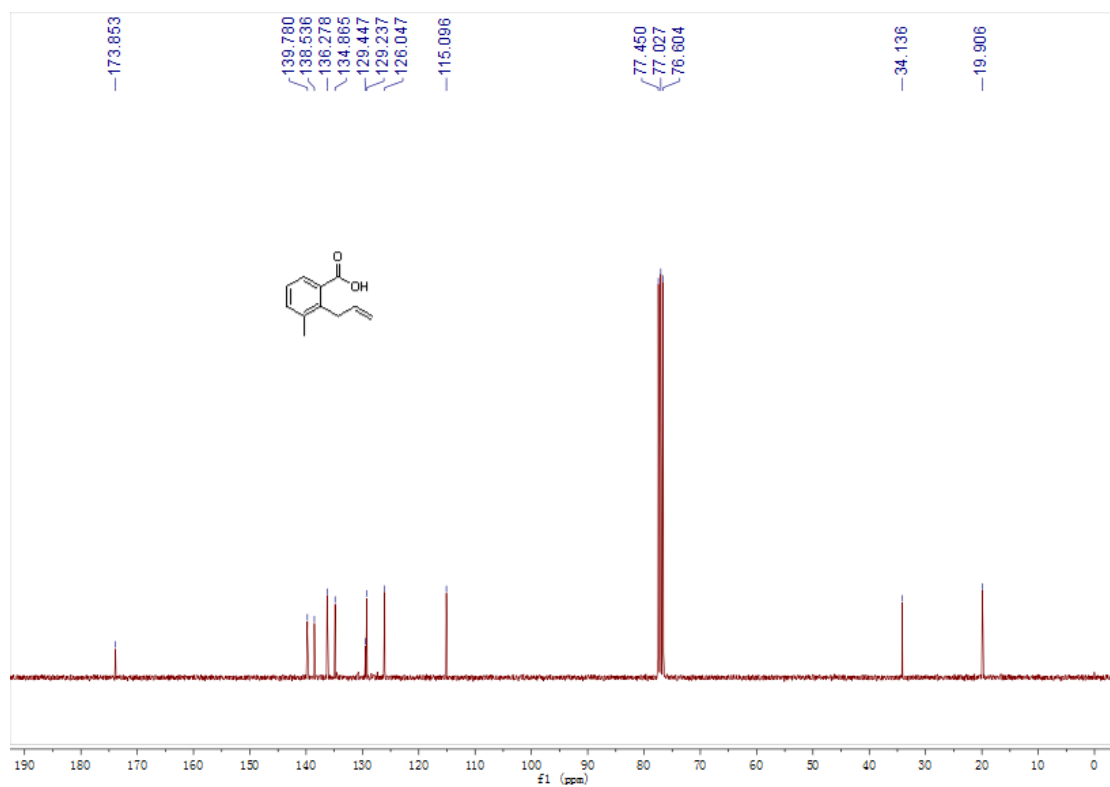
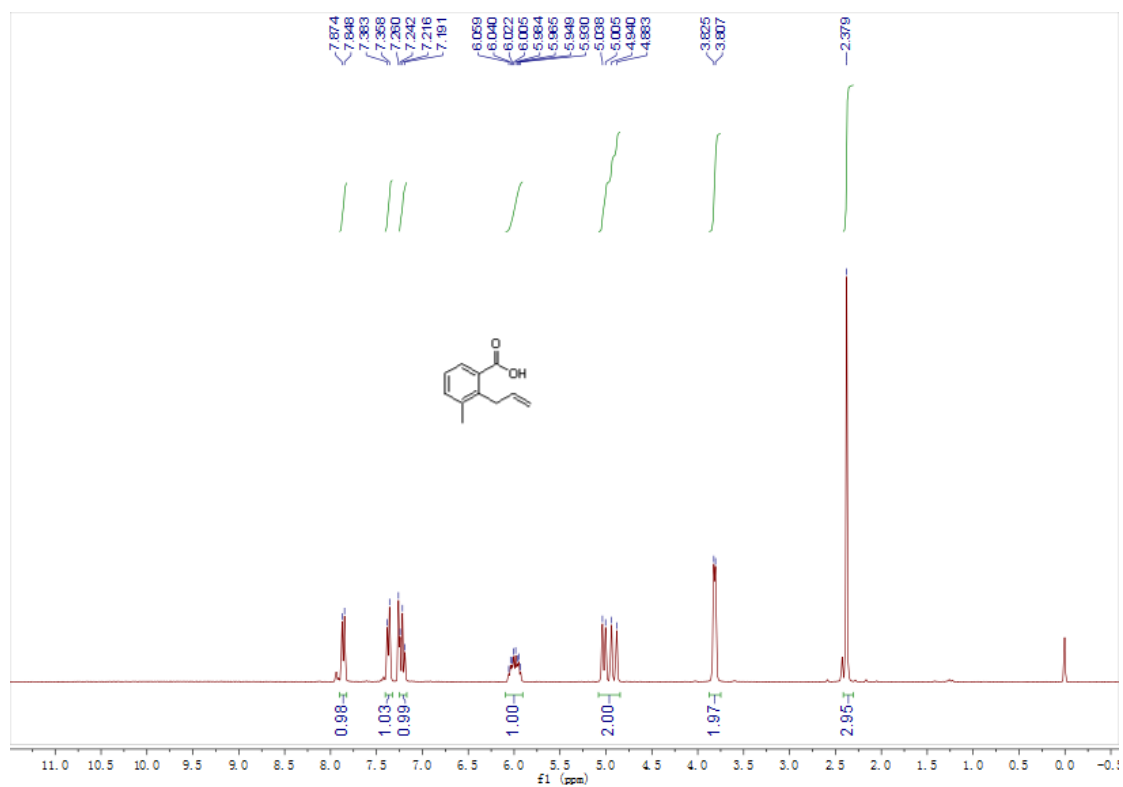
- 1 F. Kopp, S. Wunderlich and P. Knochel, *Chem. Commun.* 2007, **20**, 2075-2077.
- 2 (a) K. Shen and Q. Wang, *Chem. Sci.* 2015, **6**, 4279-4283; (b) K. Shen and Q. Wang, *Org. Chem. Front.* 2016, **3**, 222-226; (c) D. J. Wardrop, E. G. Bowen, R. E. Forslund, A. D. Sussman and S. L. Weerasekera, *J. Am. Chem. Soc.* 2010, **132**, 1188-1189.
- 3 Z.-Y. Chen, L.-Y. Wu, H.-S. Fang, T. Zhang, Z.-F. Mao, Y. Zou, X.-J. Zhang and M. Yan, *Adv. Synth. Catal.* 2017, **359**, 3894-3899.
- 4 (a) D. E. Korte, L. S. Hegedus and R. K. Wirth, *J. Org. Chem.* 1977, **42**, 1329-1336; (b) L. E. Fisher, J. M. Muchowski, and R. D. Clark, *J. Org. Chem.* 1992, **57**, 2700-2705; (c) C. F. Bender, and R. A. Widenhoefer, *Chem. Commun.* 2008, **24**, 2741-2743.
- 5 K. C. Miles, C. C. Le and J. P. Stambuli, *Chemistry*, 2014, **20**, 11336-11339.
- 6 B. N. Hemric, K. Shen, and Q. Wang, *J. Am. Chem. Soc.* 2016, **138**, 5813-5816.
- 7 K. Shen, Q. Wang, *J Am Chem Soc* **2017**, *139*, 13110-13116.
- 8 (a) T. Takesue, M. Fujita, T. Sugimura and H. Akutsu, *Org. Lett.* 2014, **16**, 4634-4637; (b) Z. Q. Zhang and F. Liu, *Org. Biomol. Chem.* 2015, **13**, 6690-6693.
- 9 N. Okamoto, K. Takeda and R. Yanada, *J. Org. Chem.* 2010, **75**, 7615-7625.
- 10 J. Einhorn, C. Einhorn and J.-L. Luche, *Synthetic Commun.*, 1990, **20**, 1105-1112.

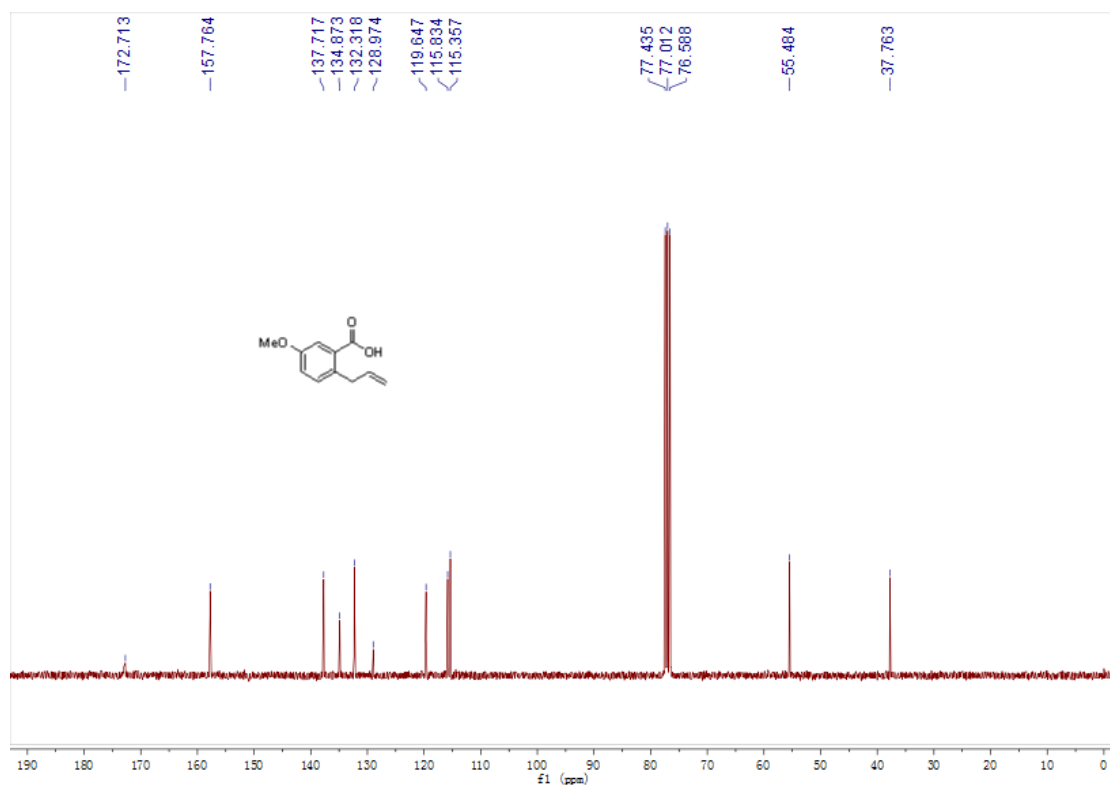
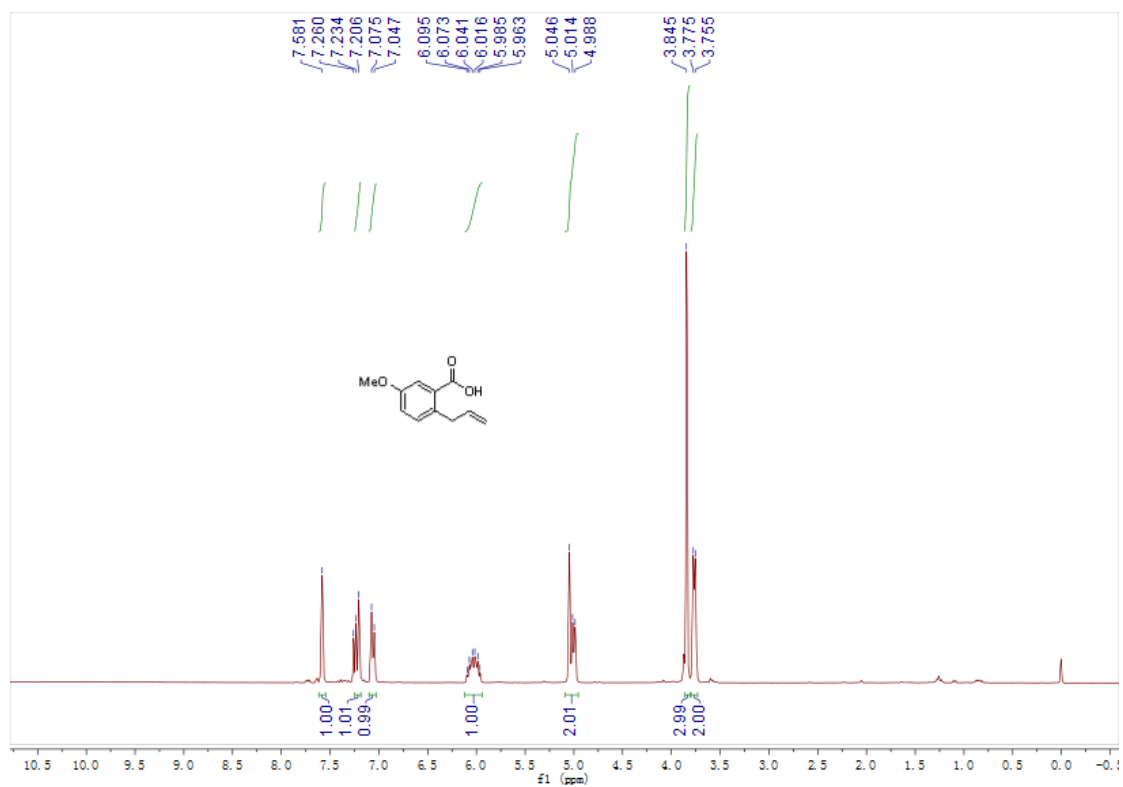
## 4. NMR Spectra

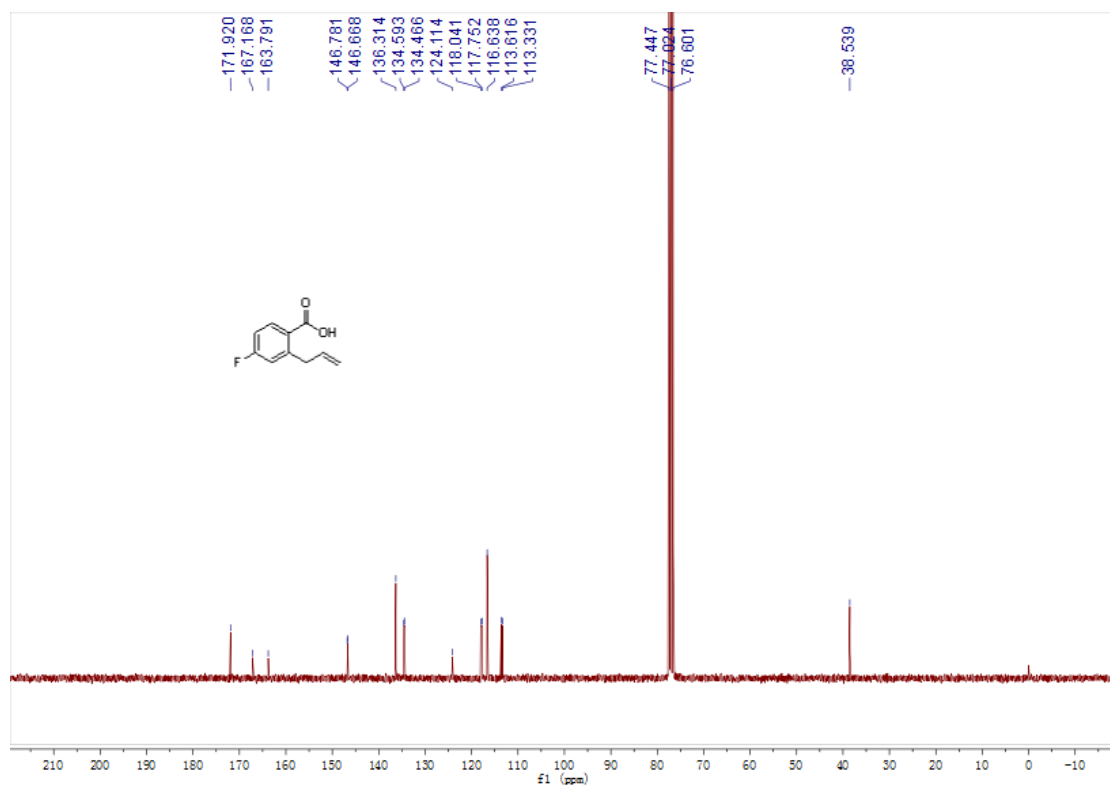
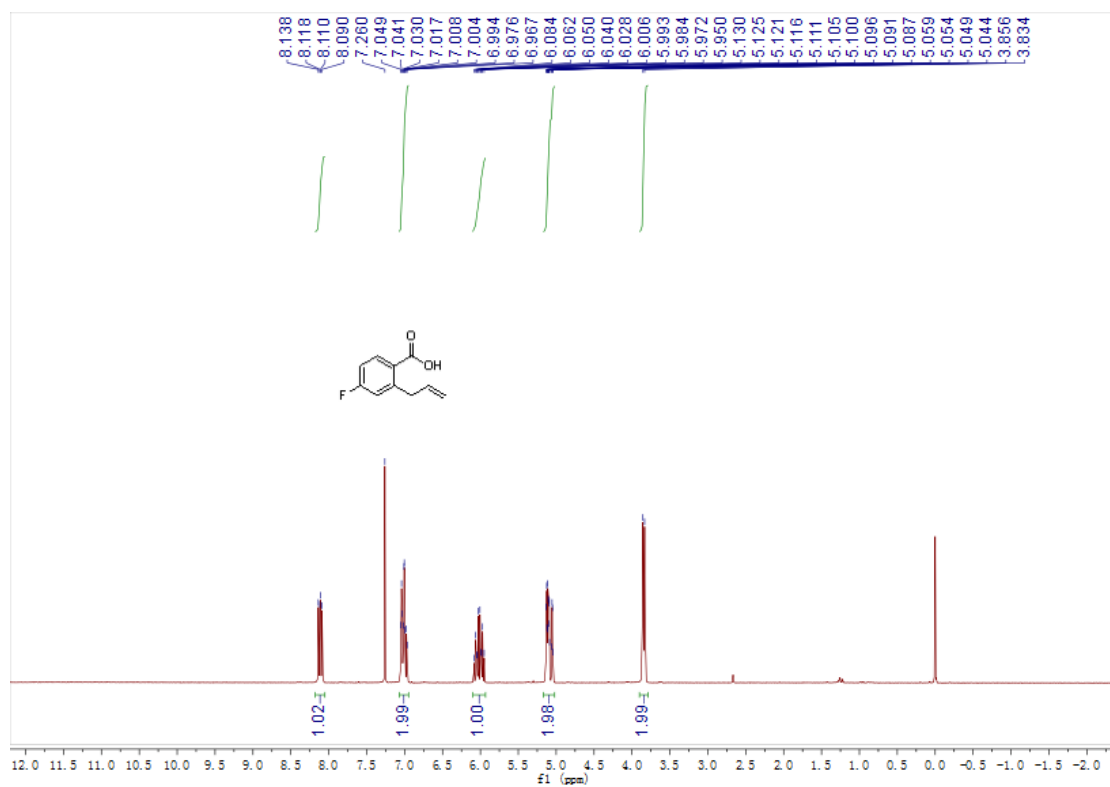
$^1\text{H}$  NMR,  $^{13}\text{C}$  NMR spectra of Substrates:

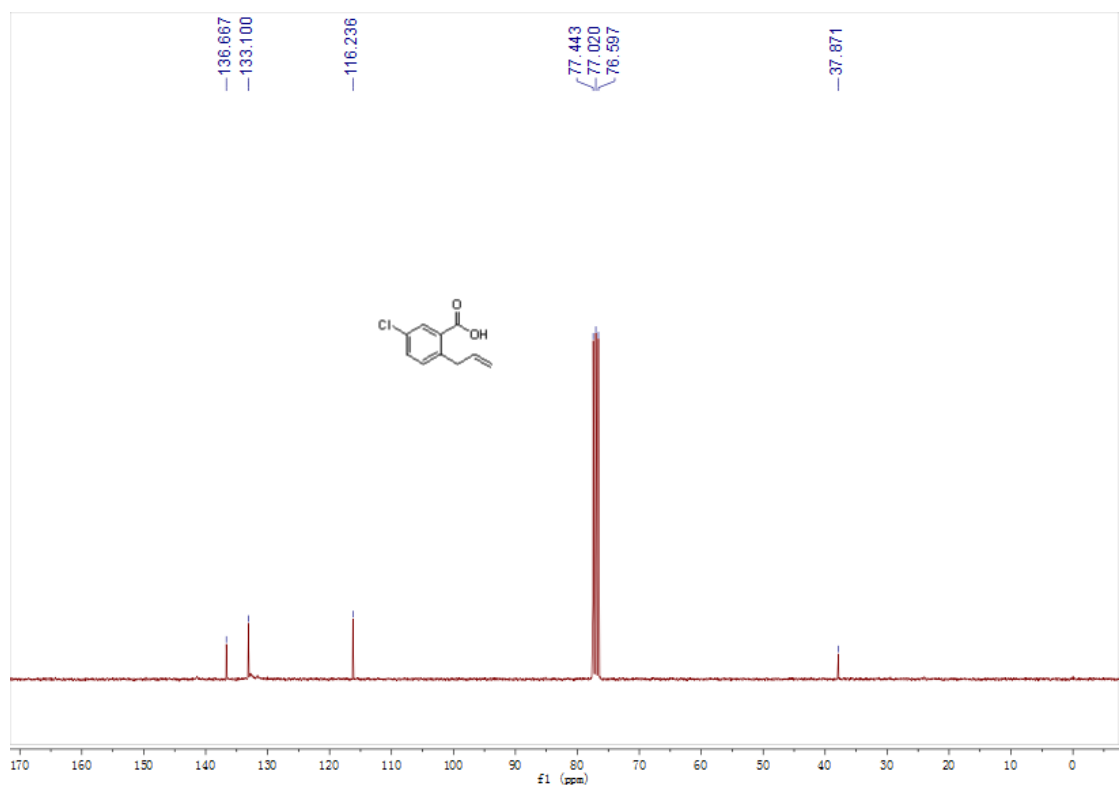
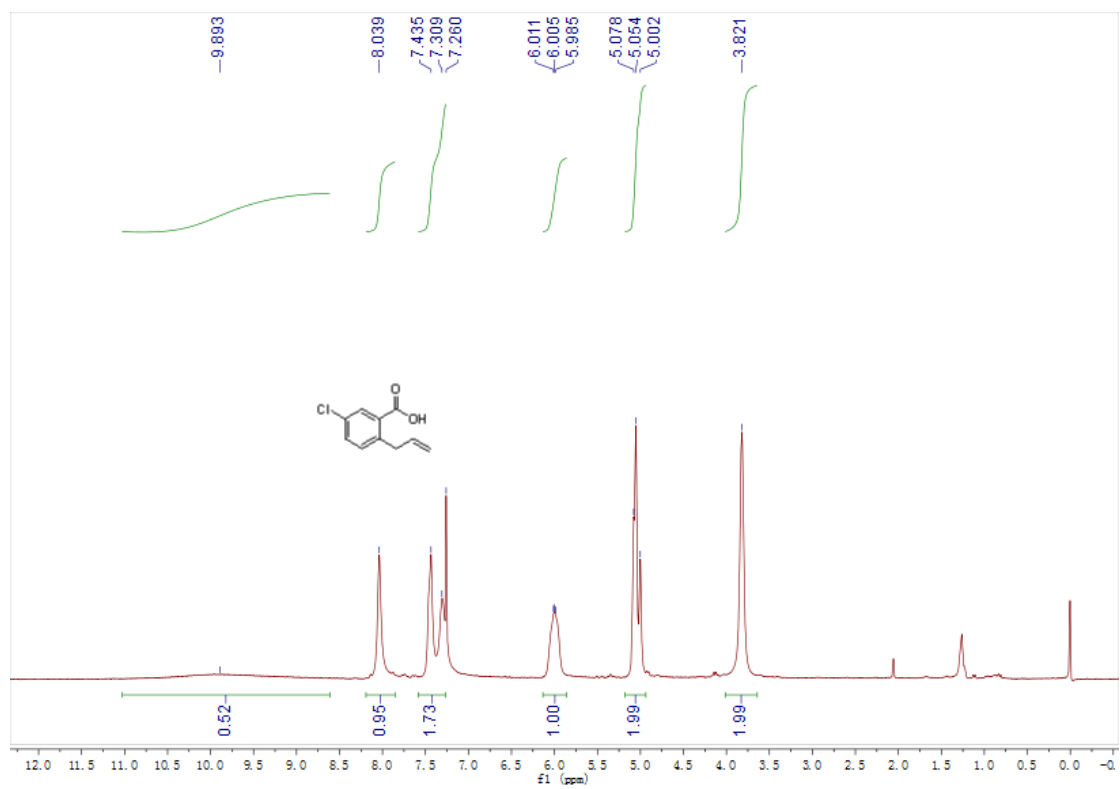




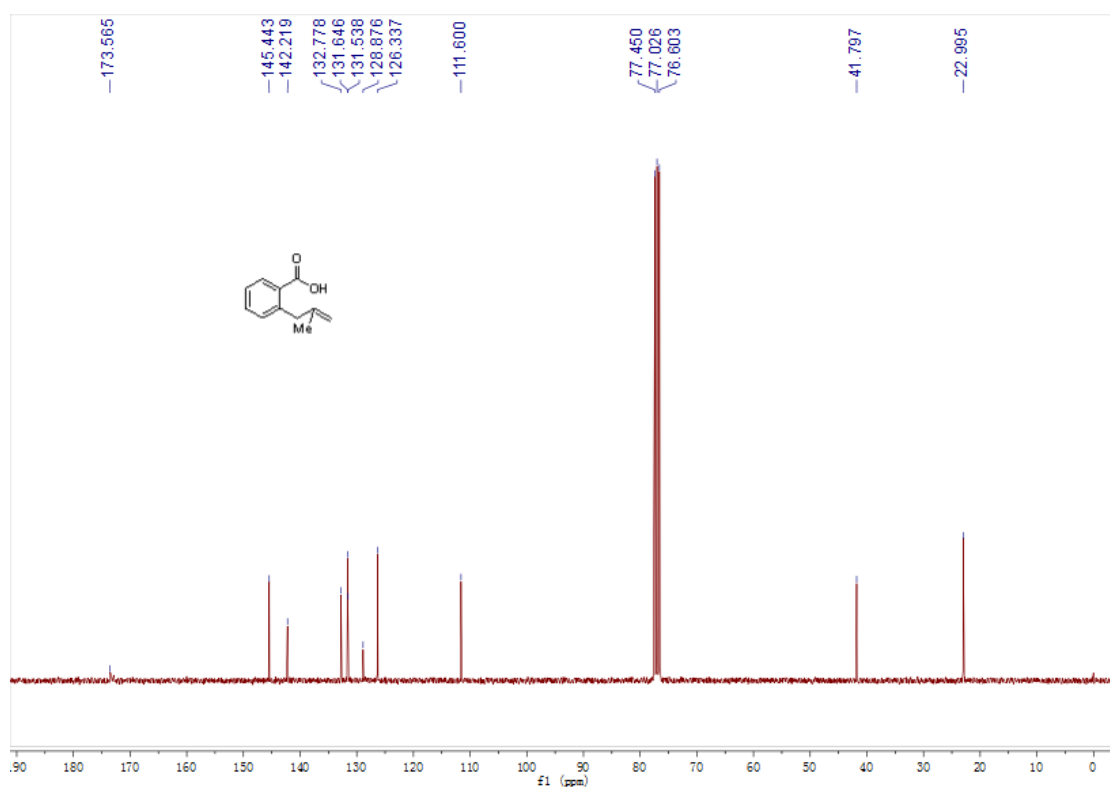
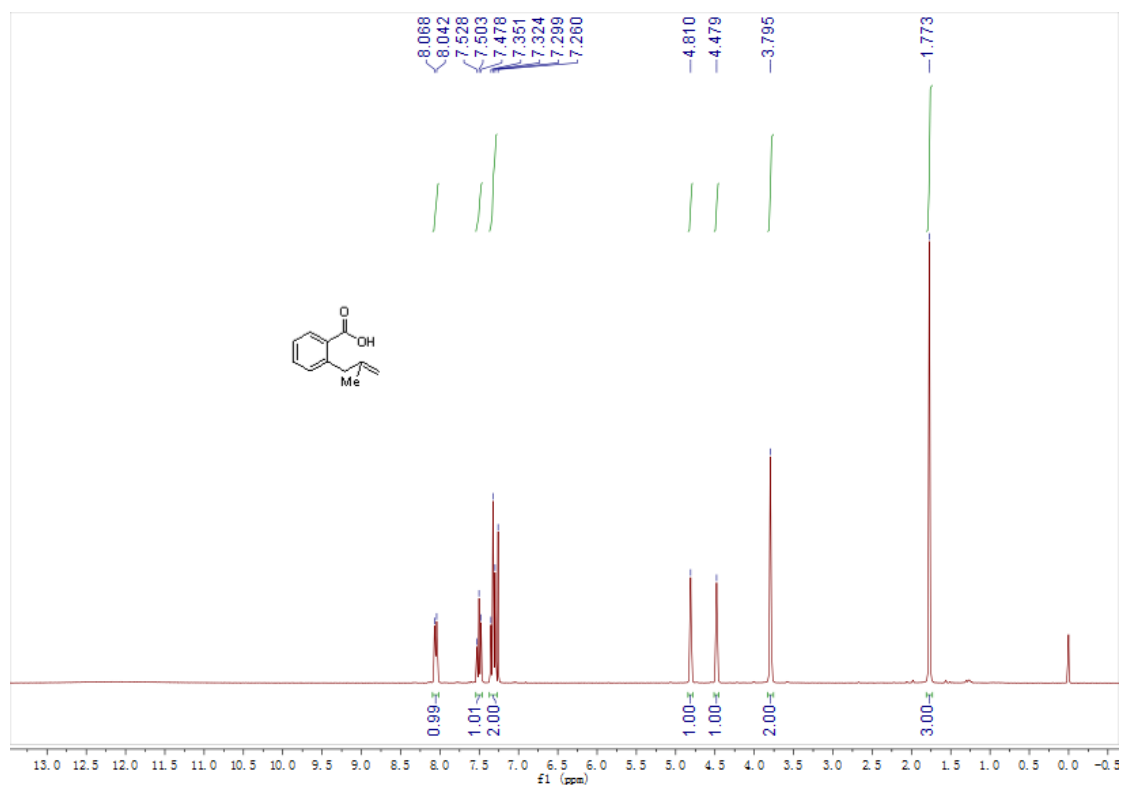


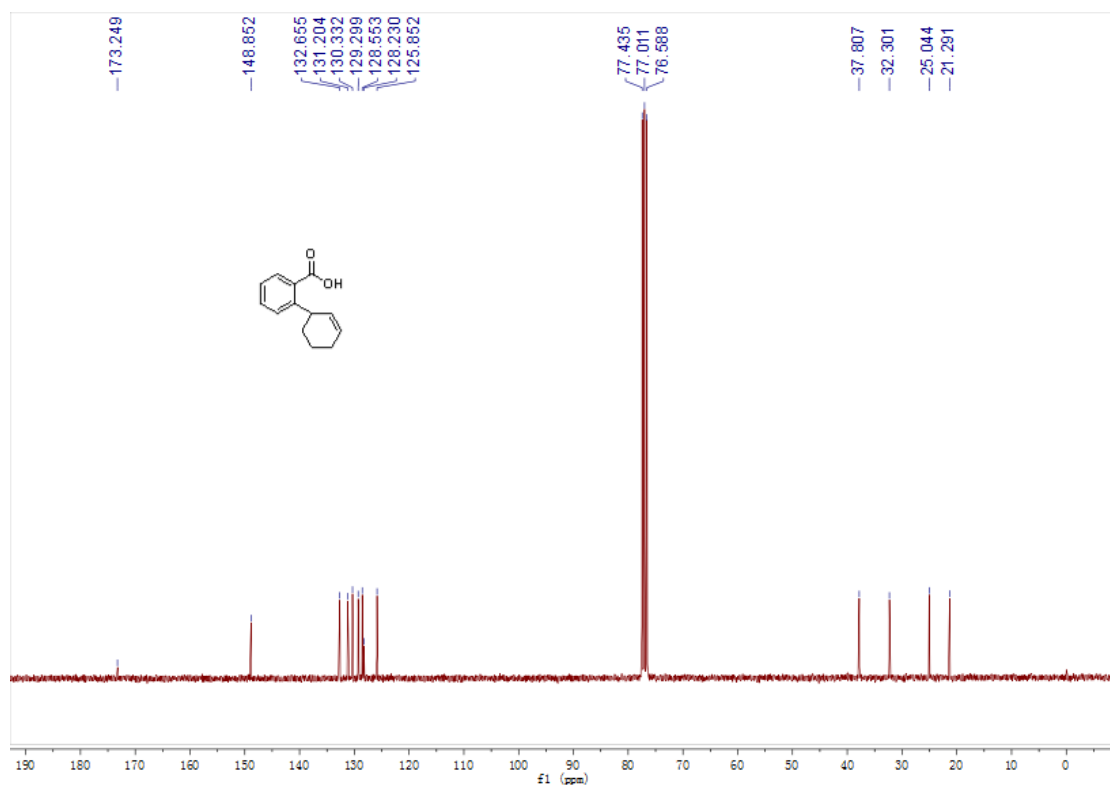
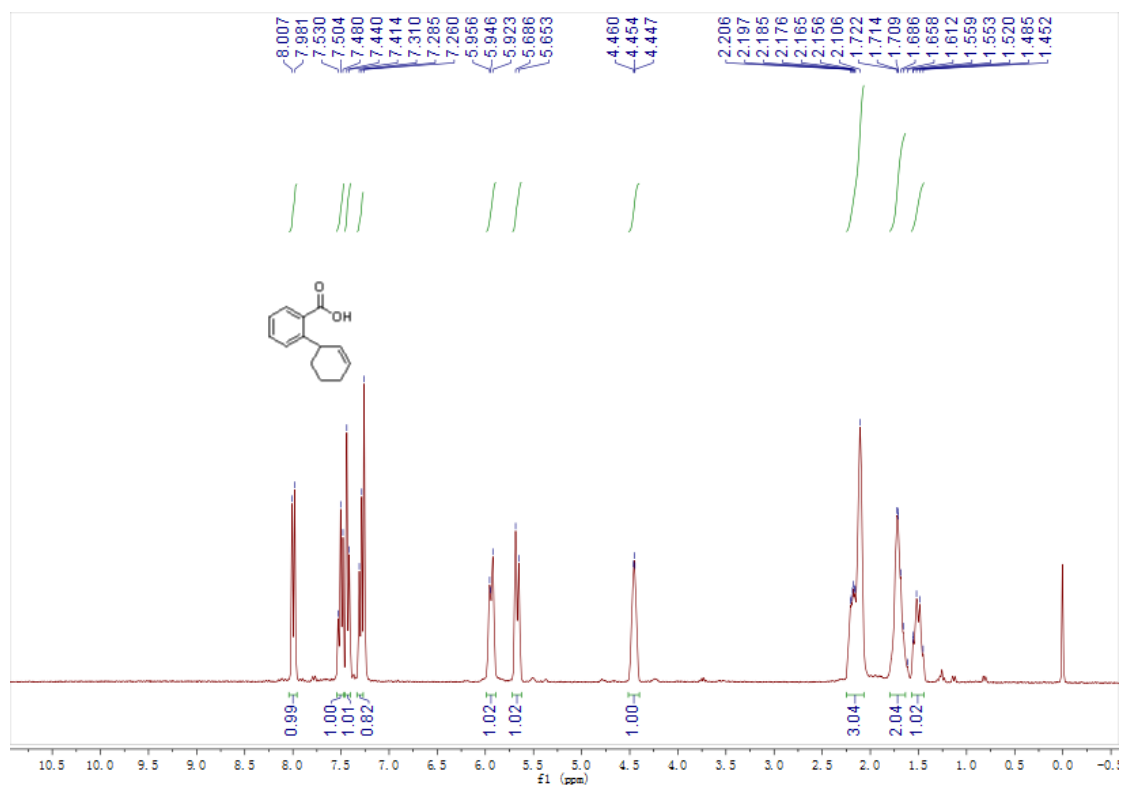


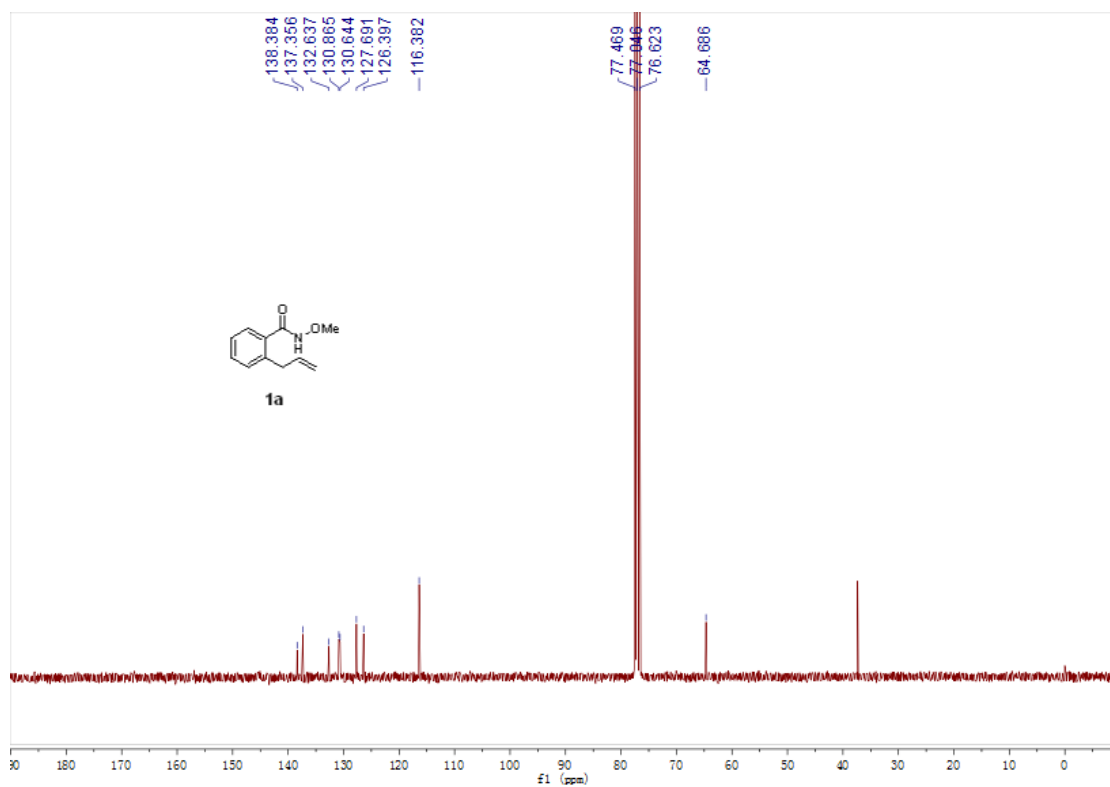
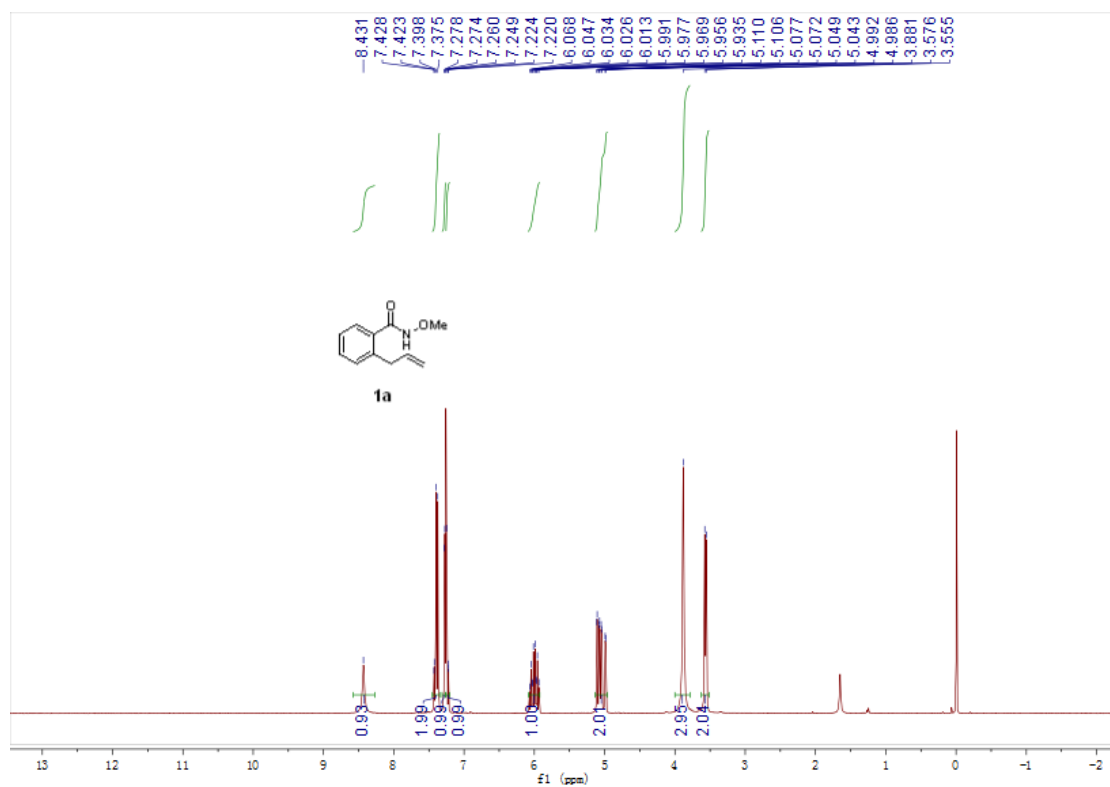


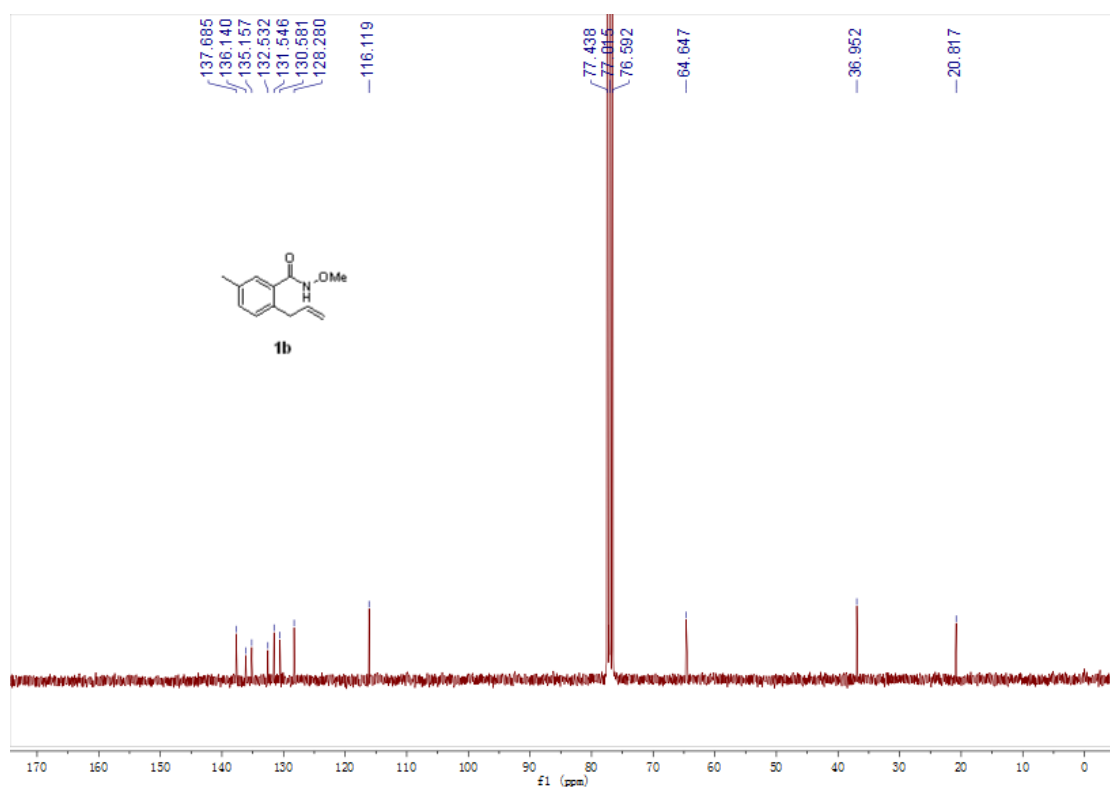
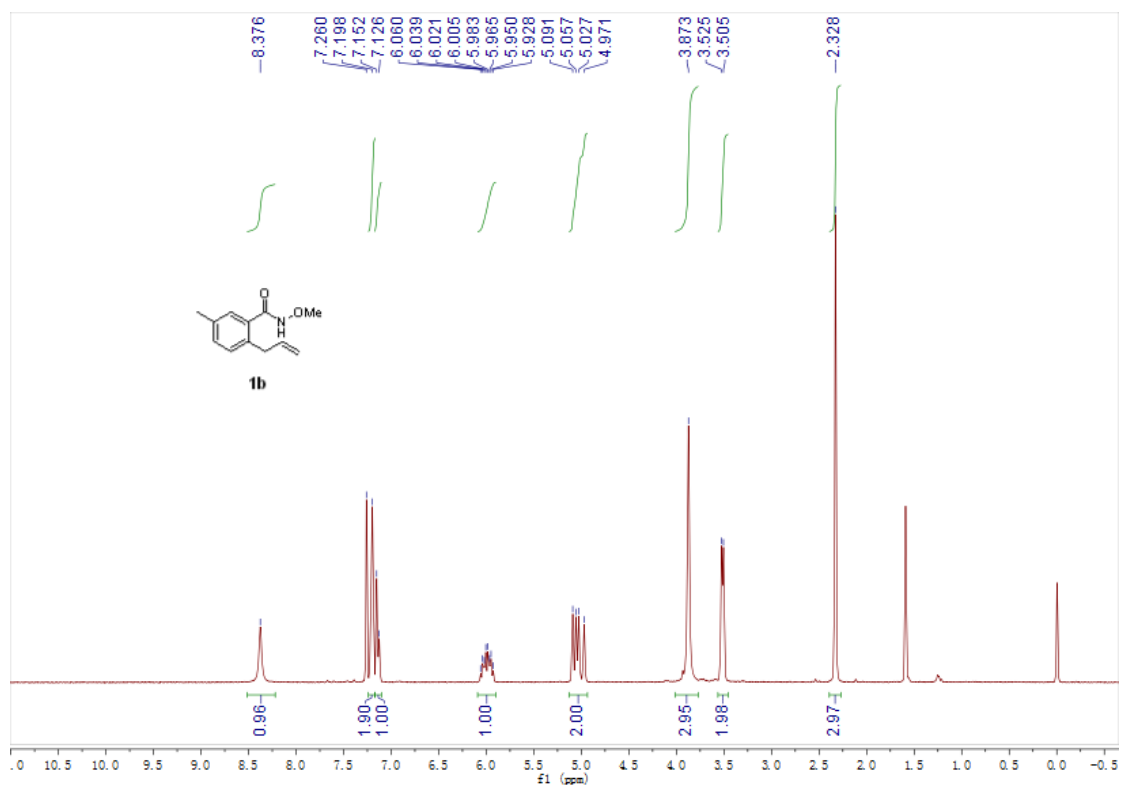


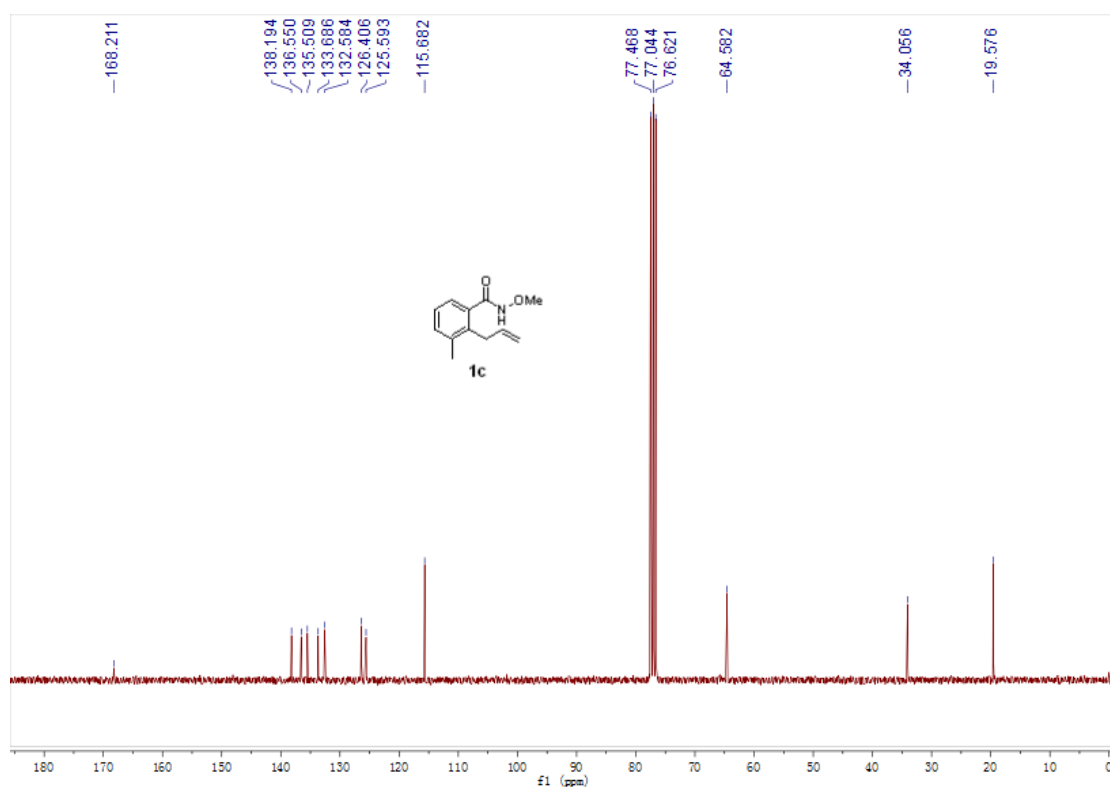
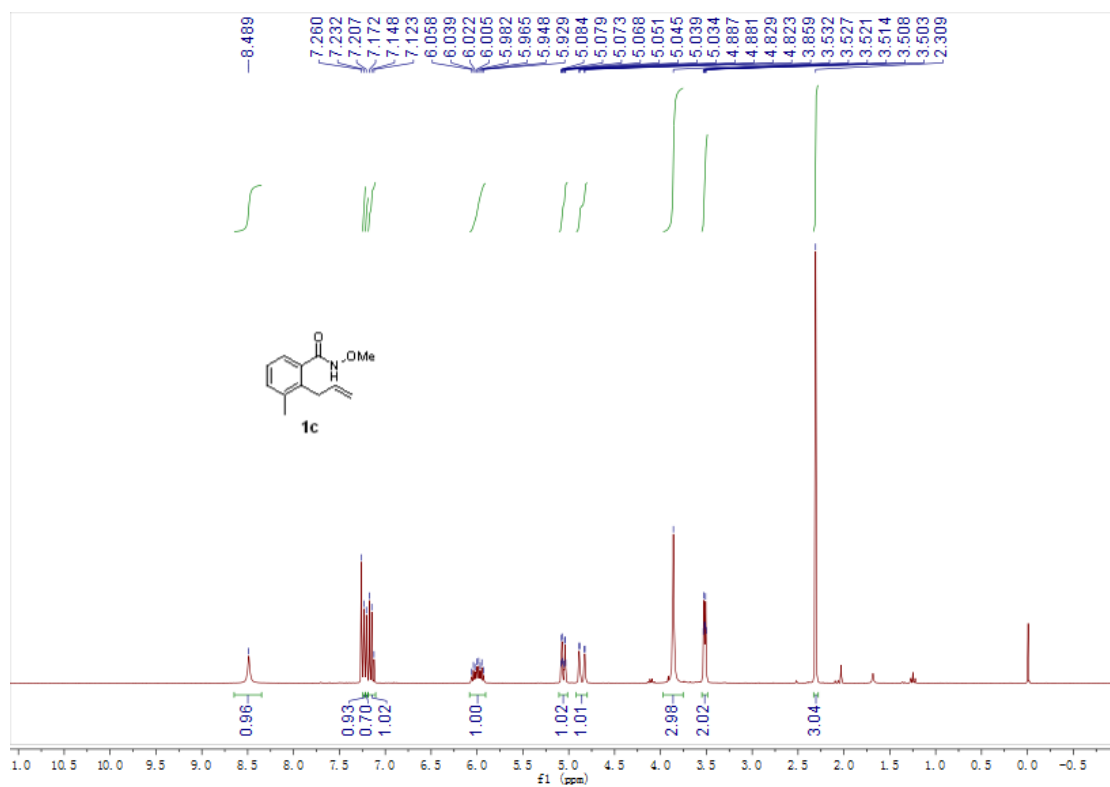


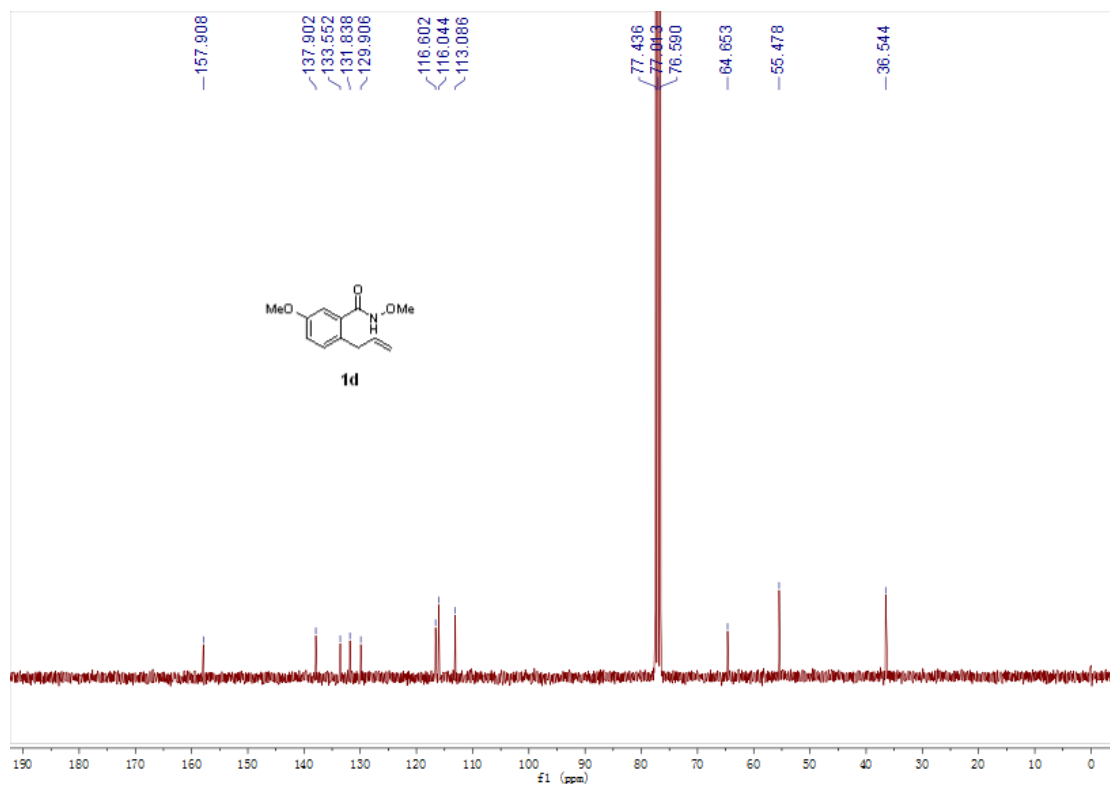
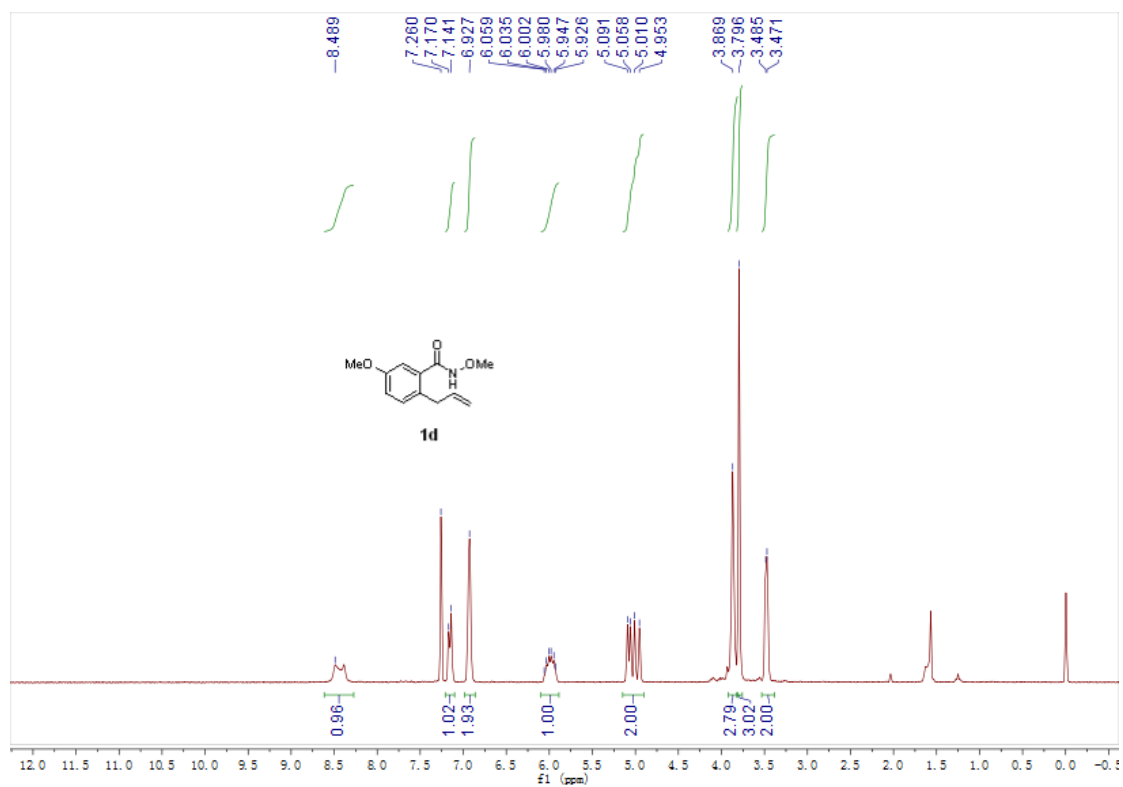


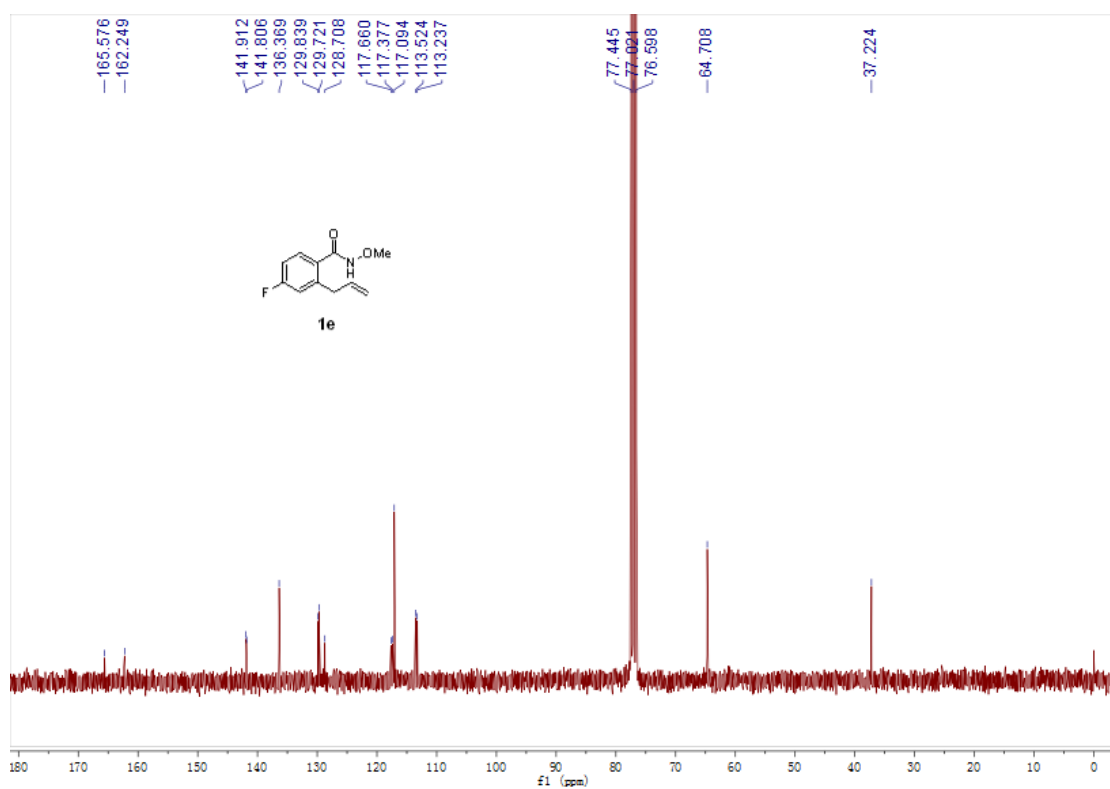
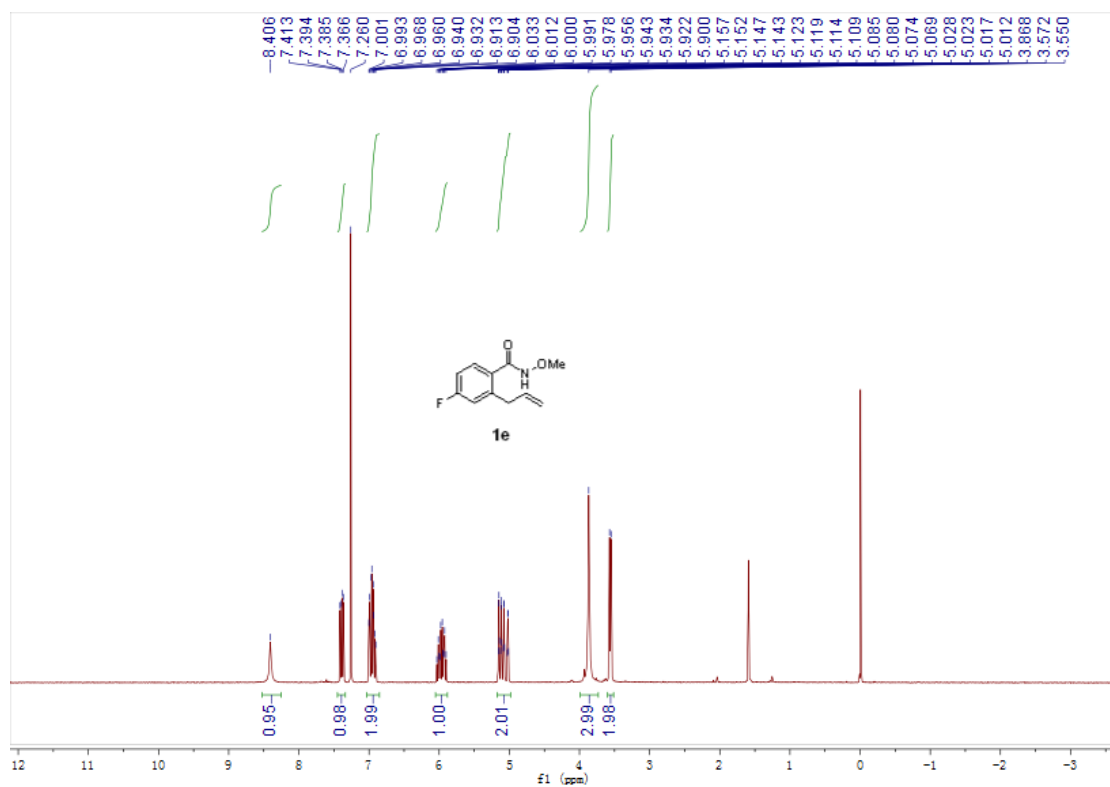


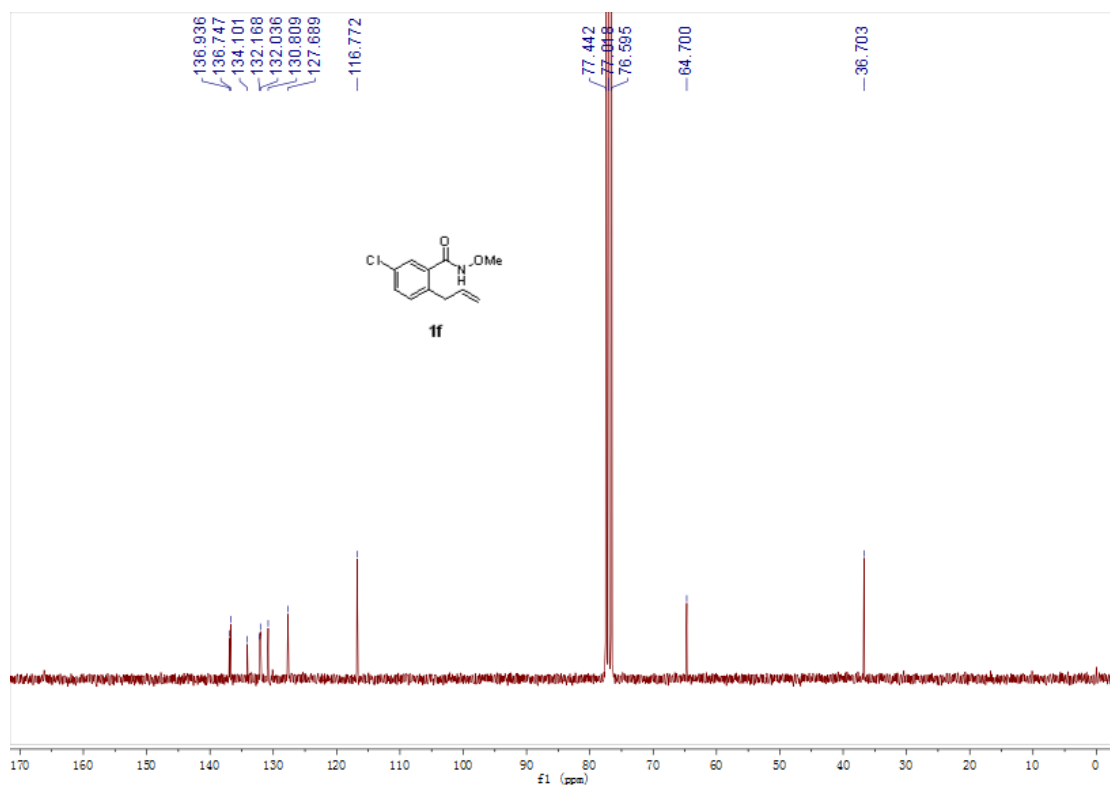
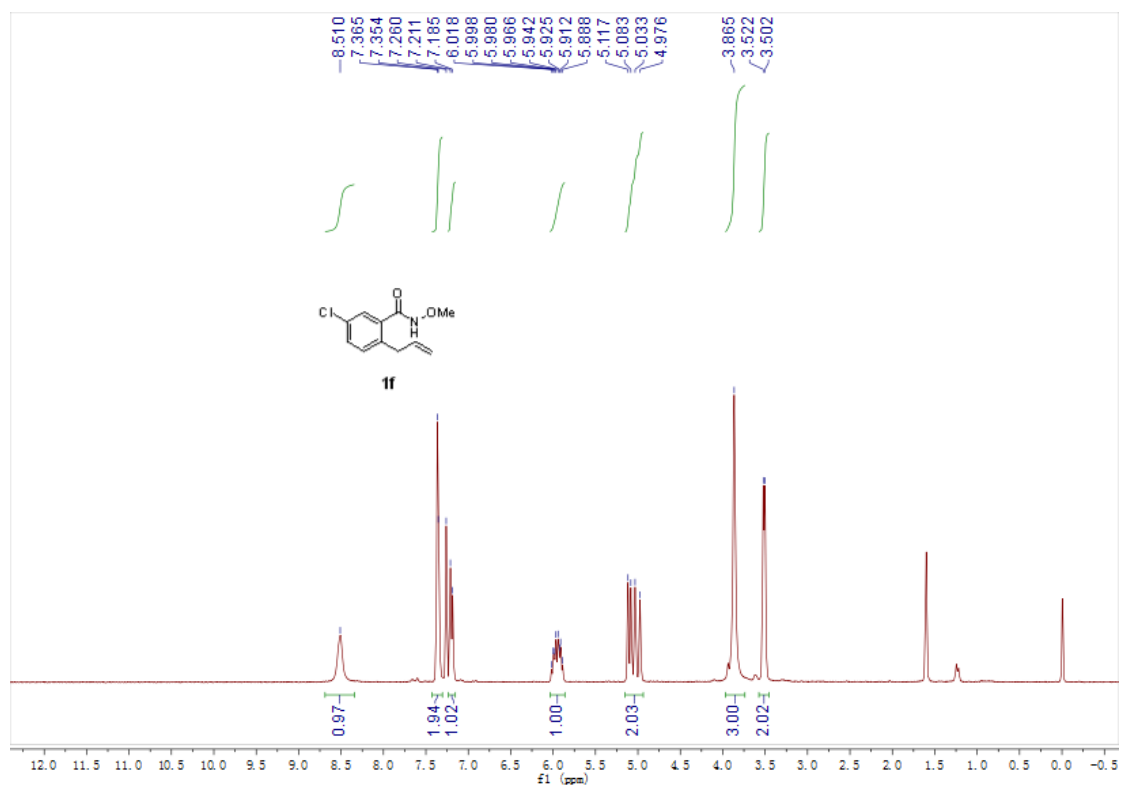




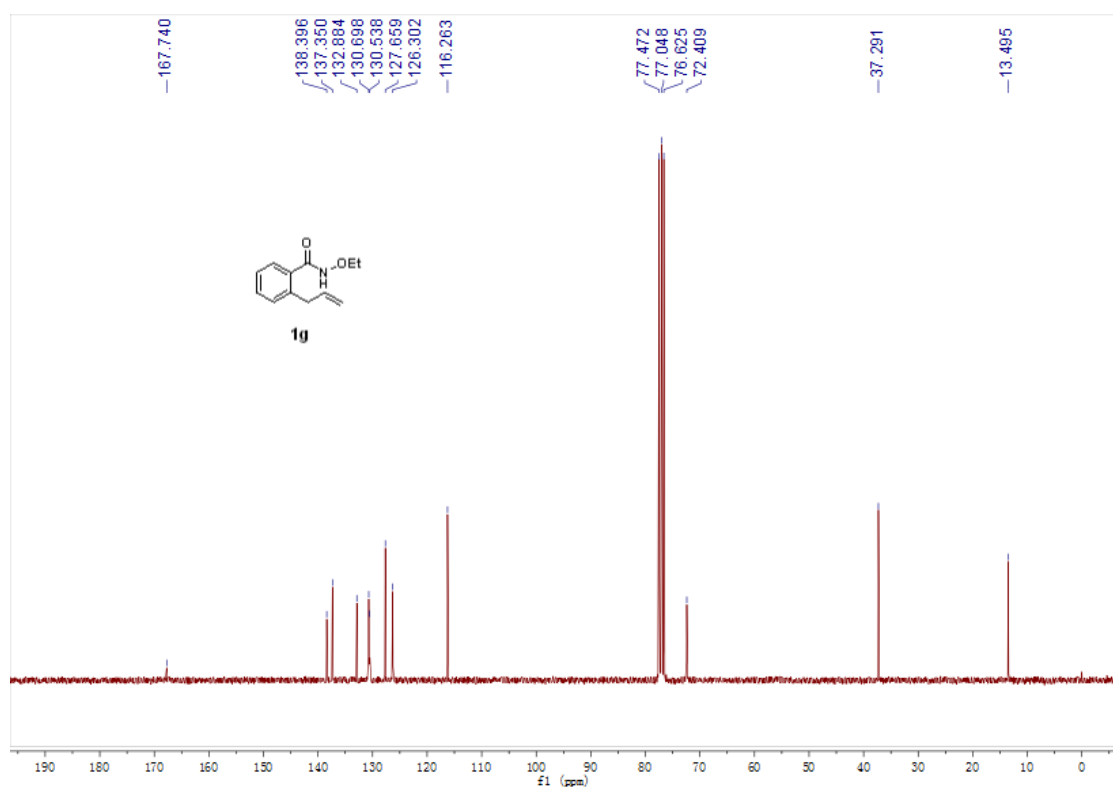
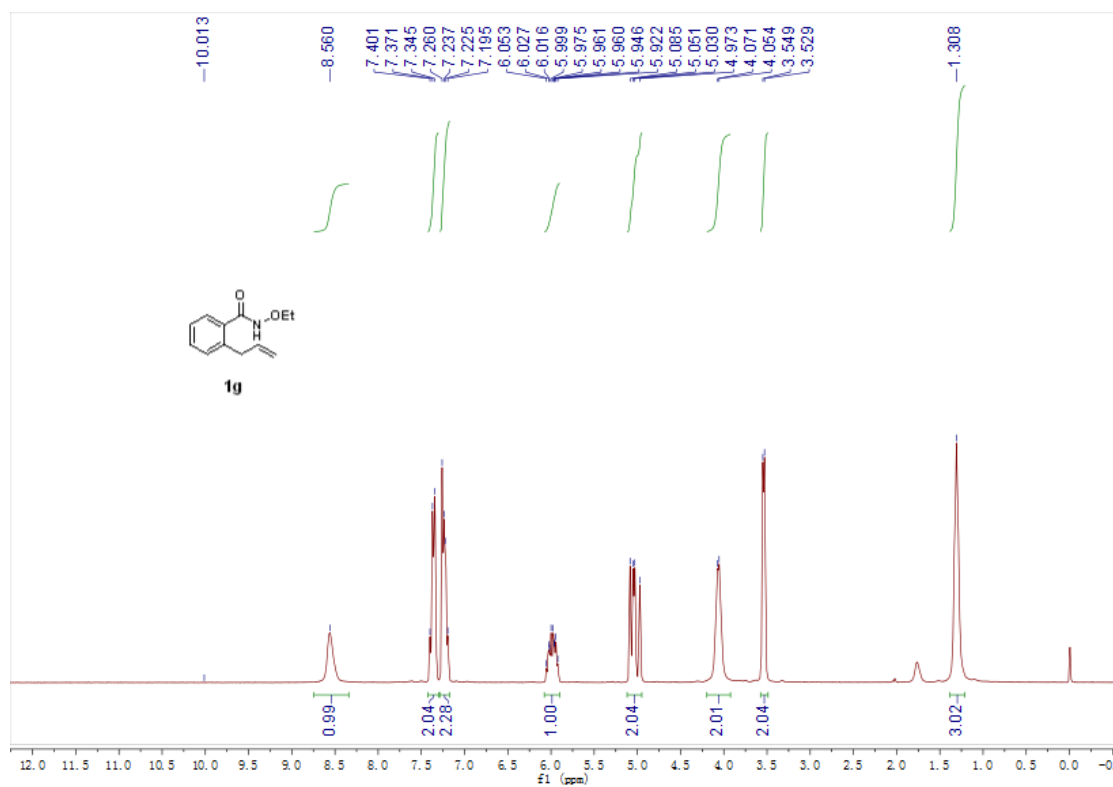


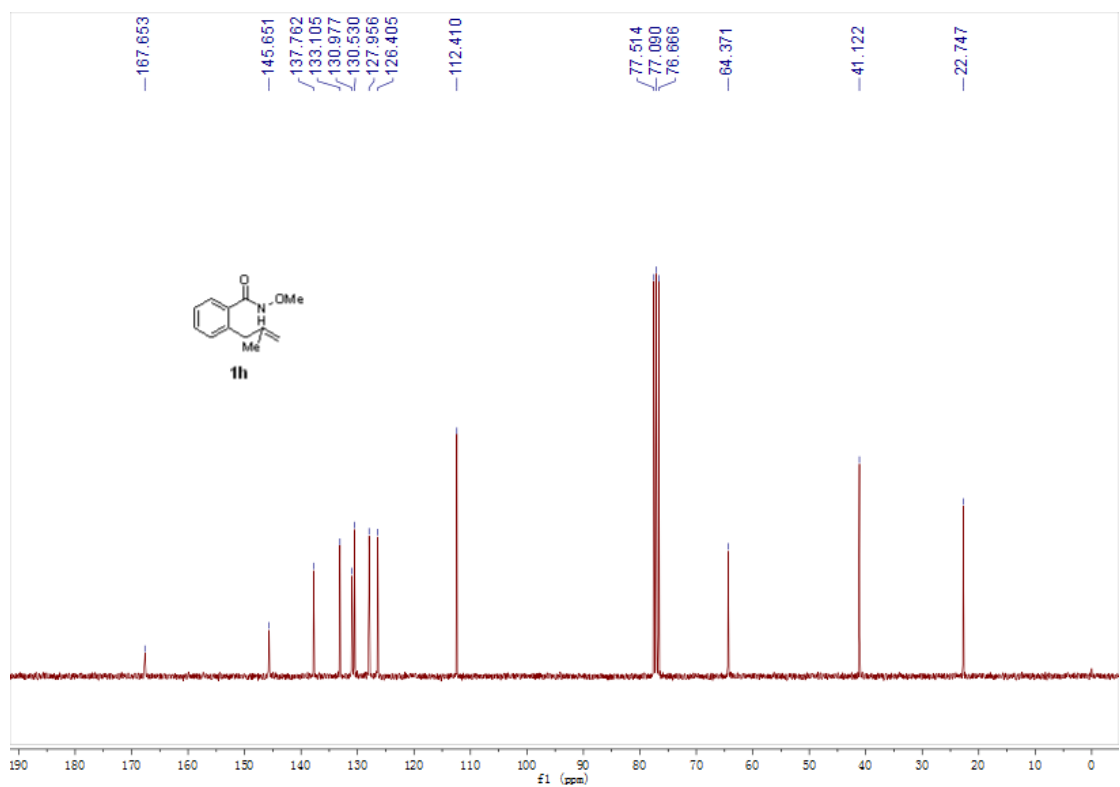
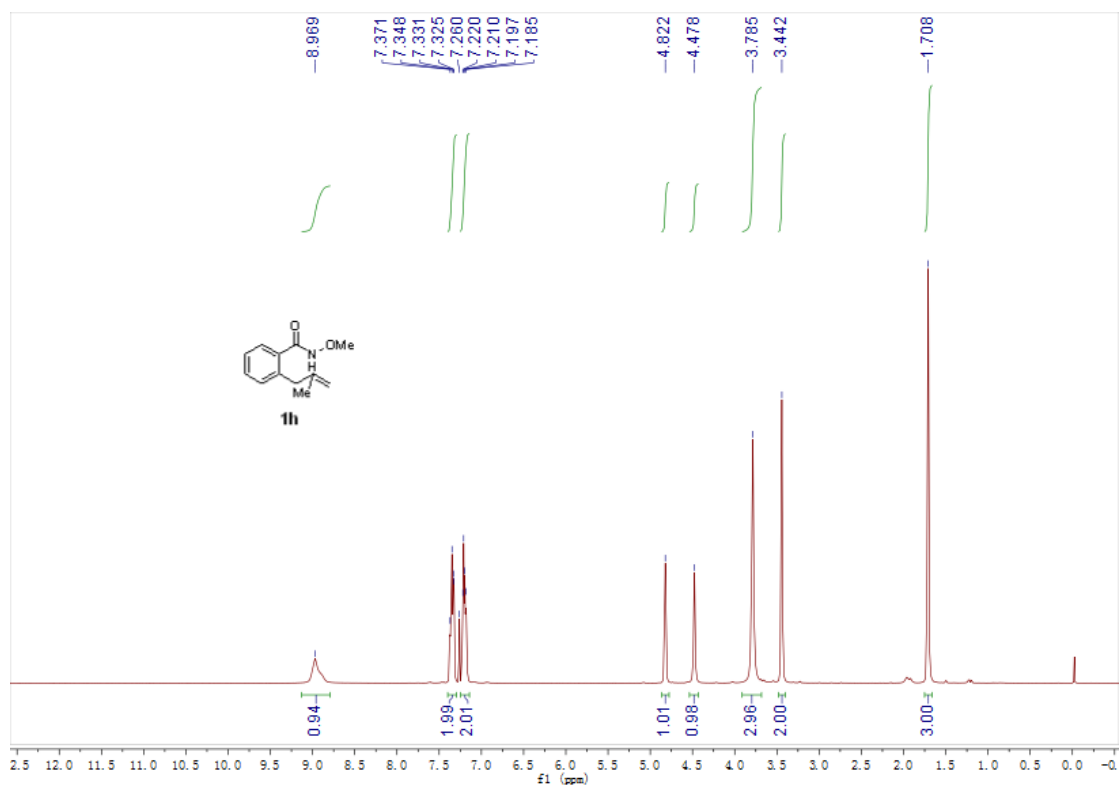


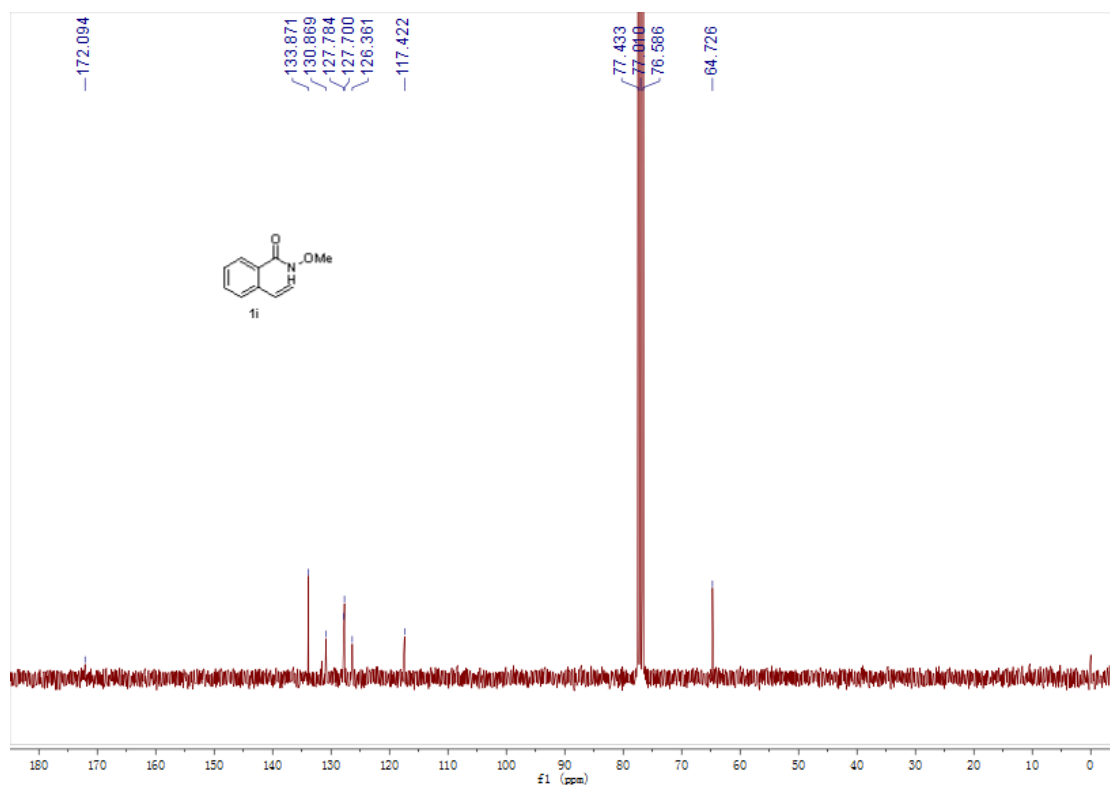
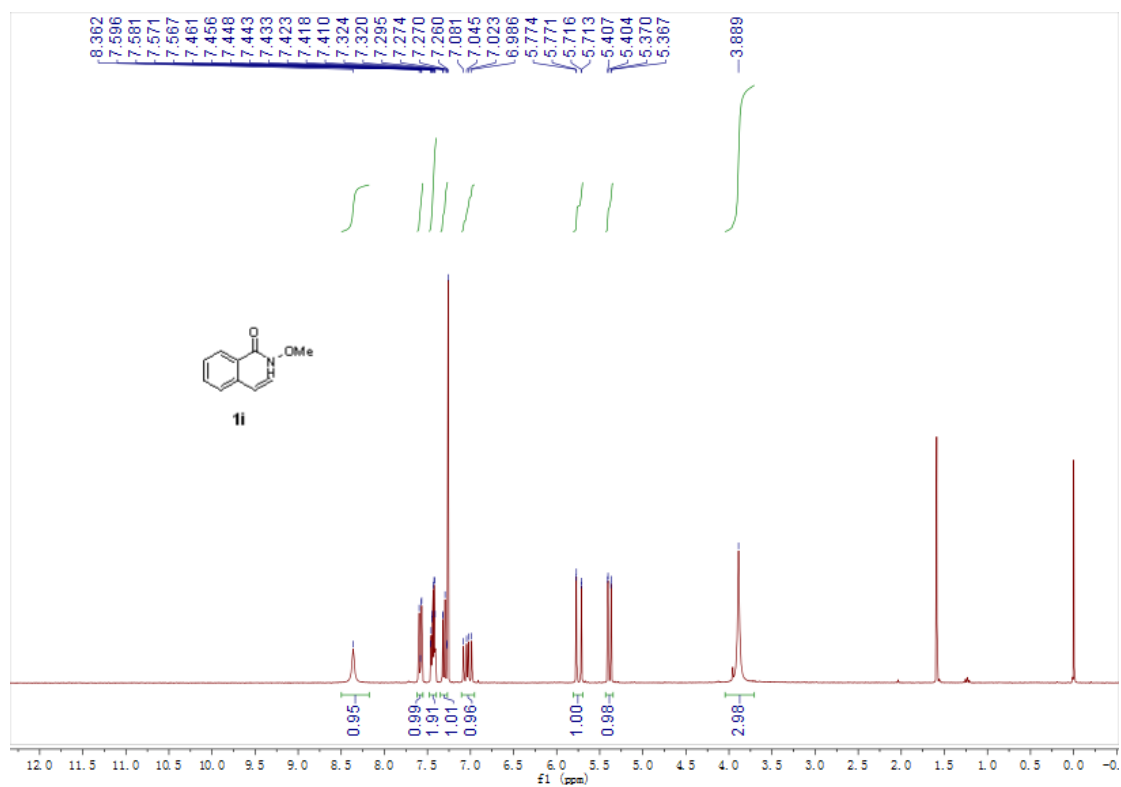


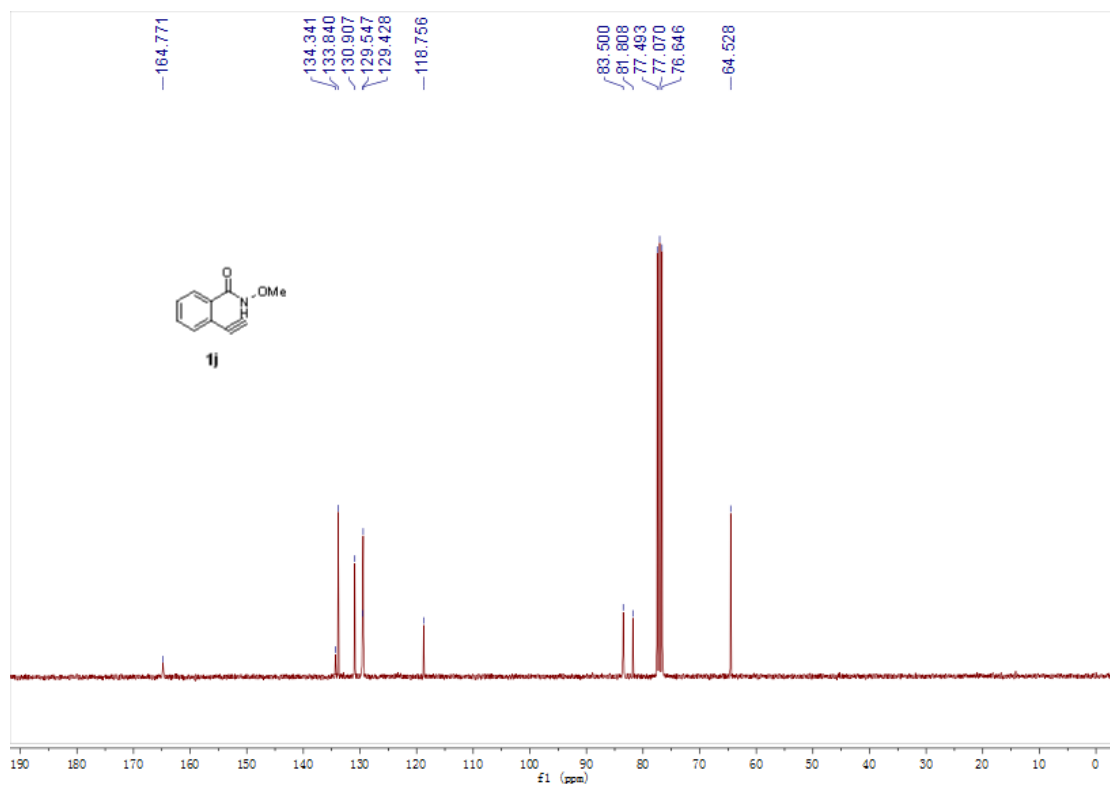
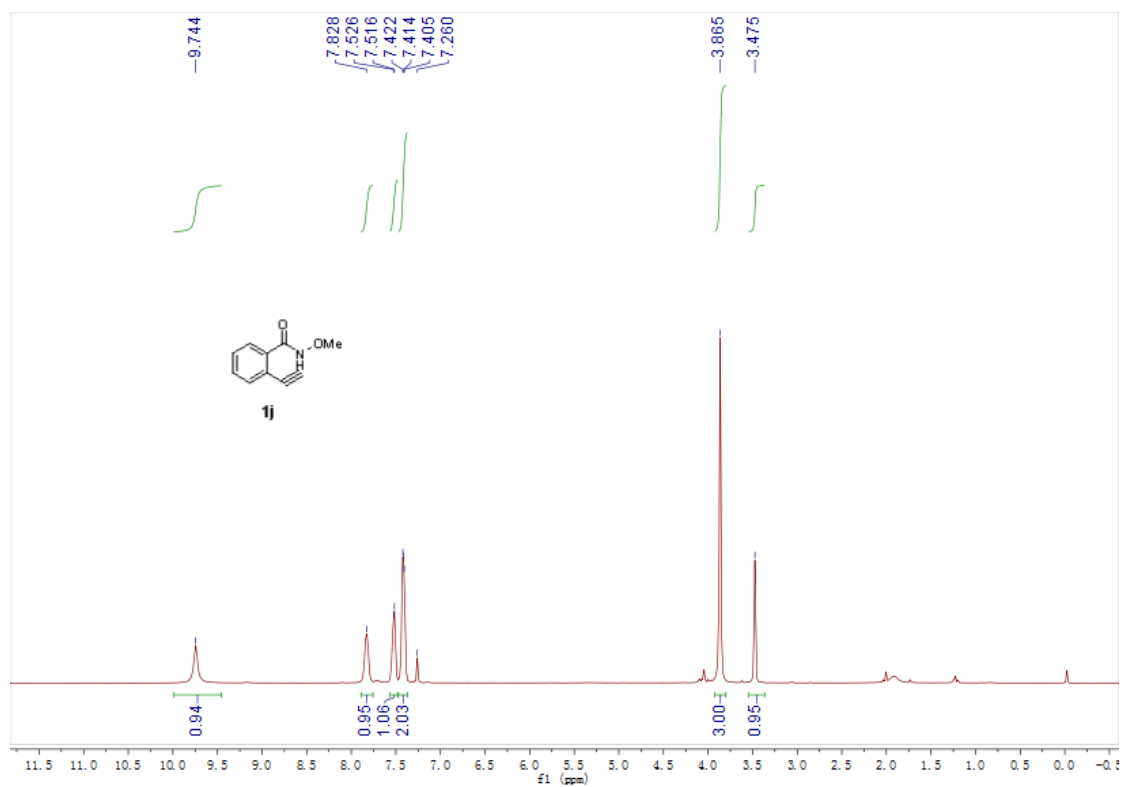


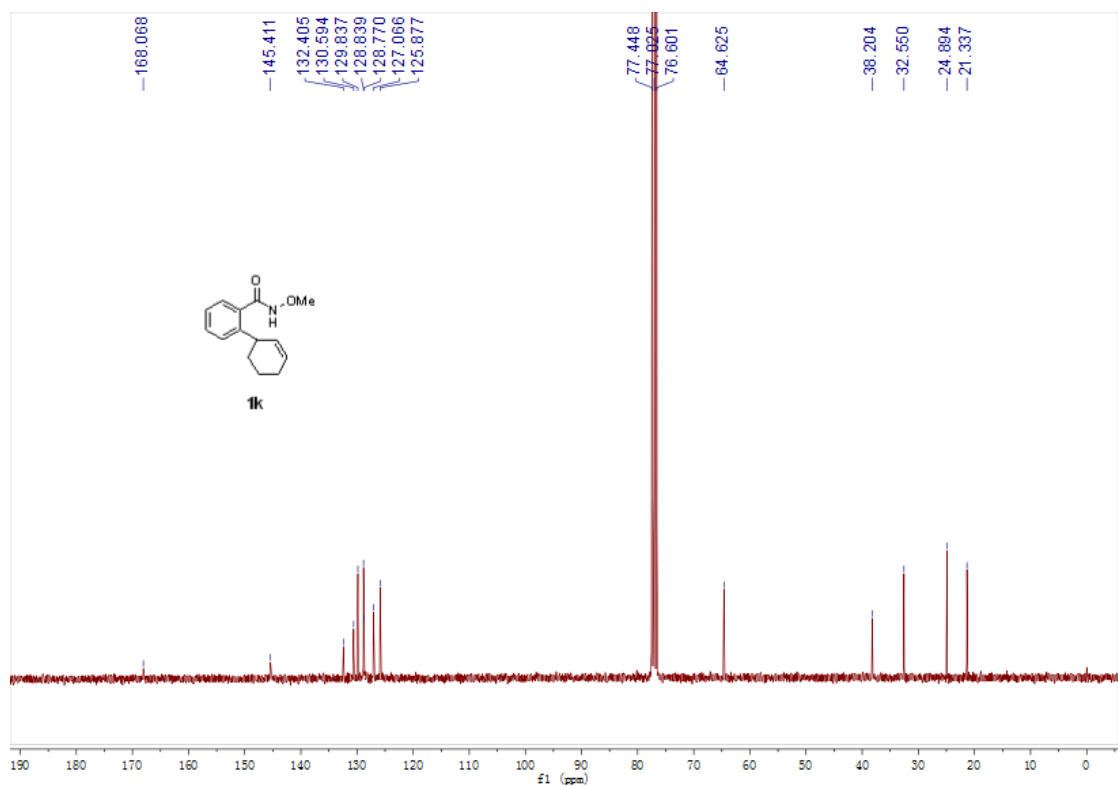
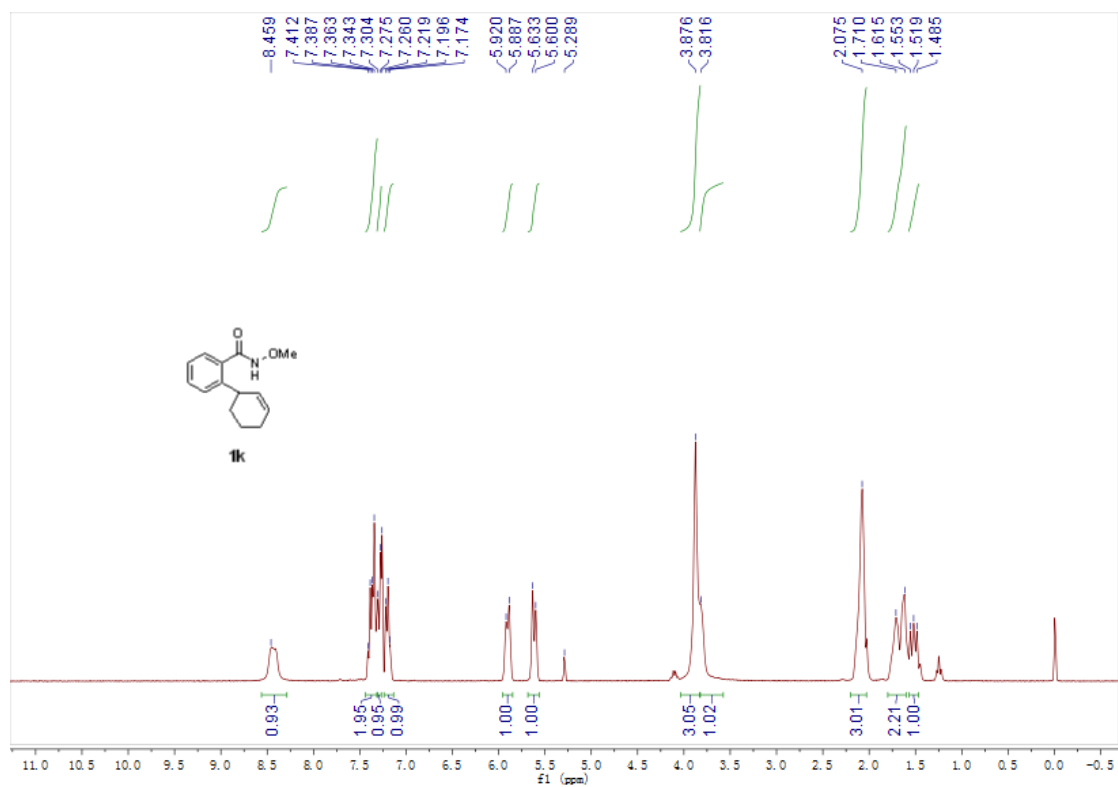


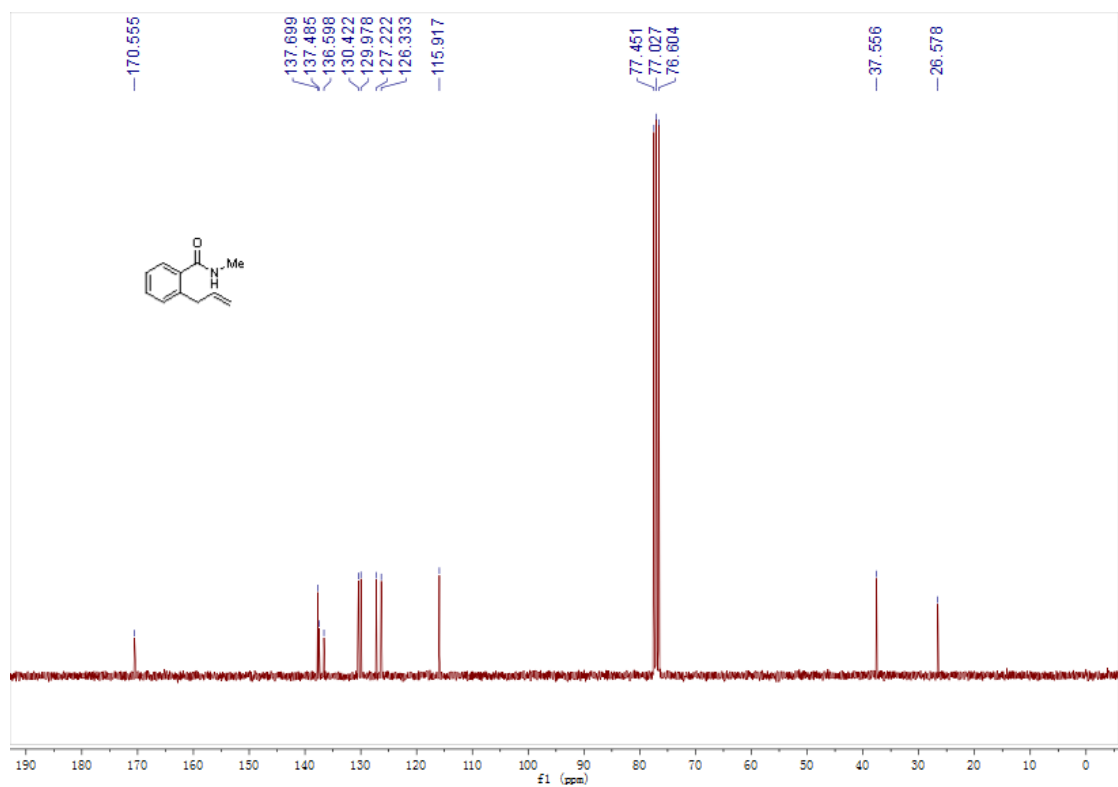
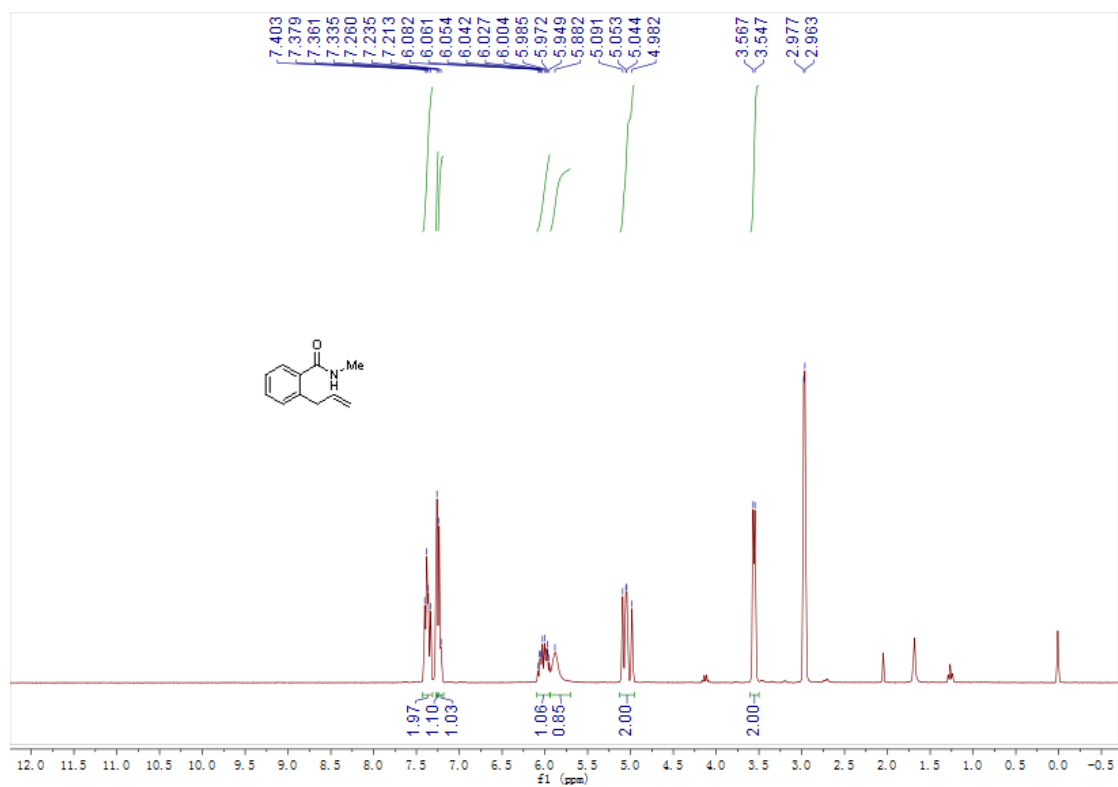


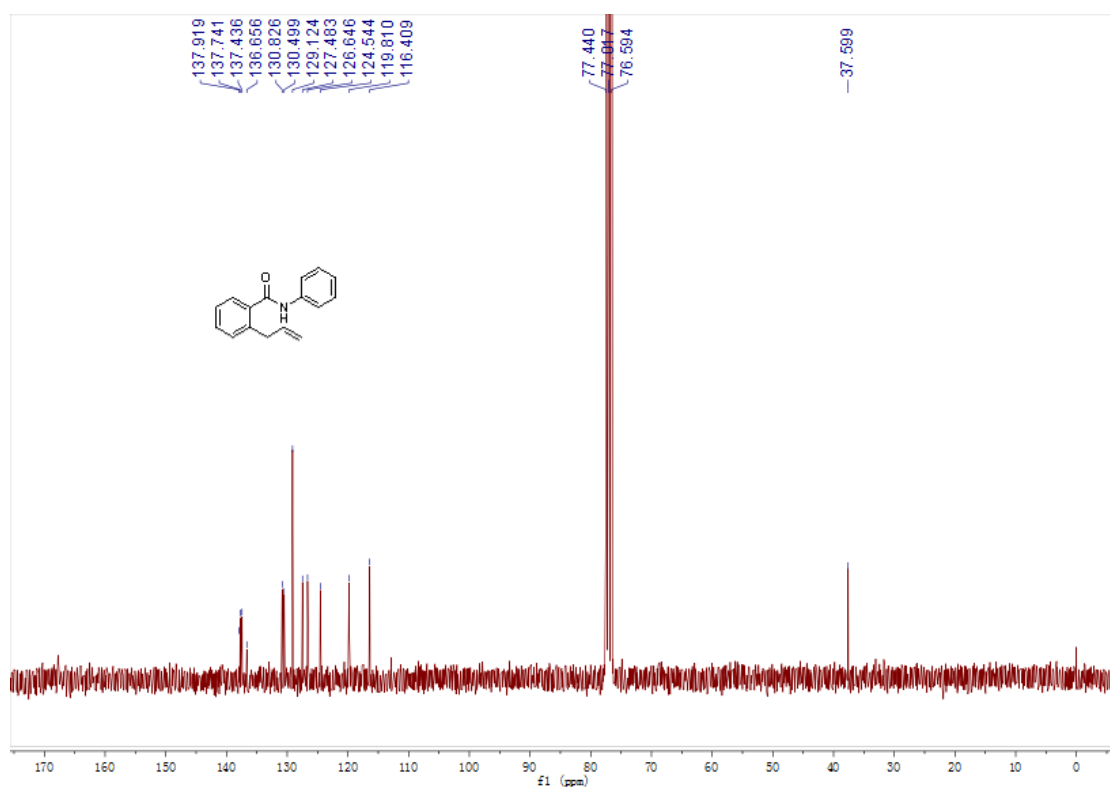
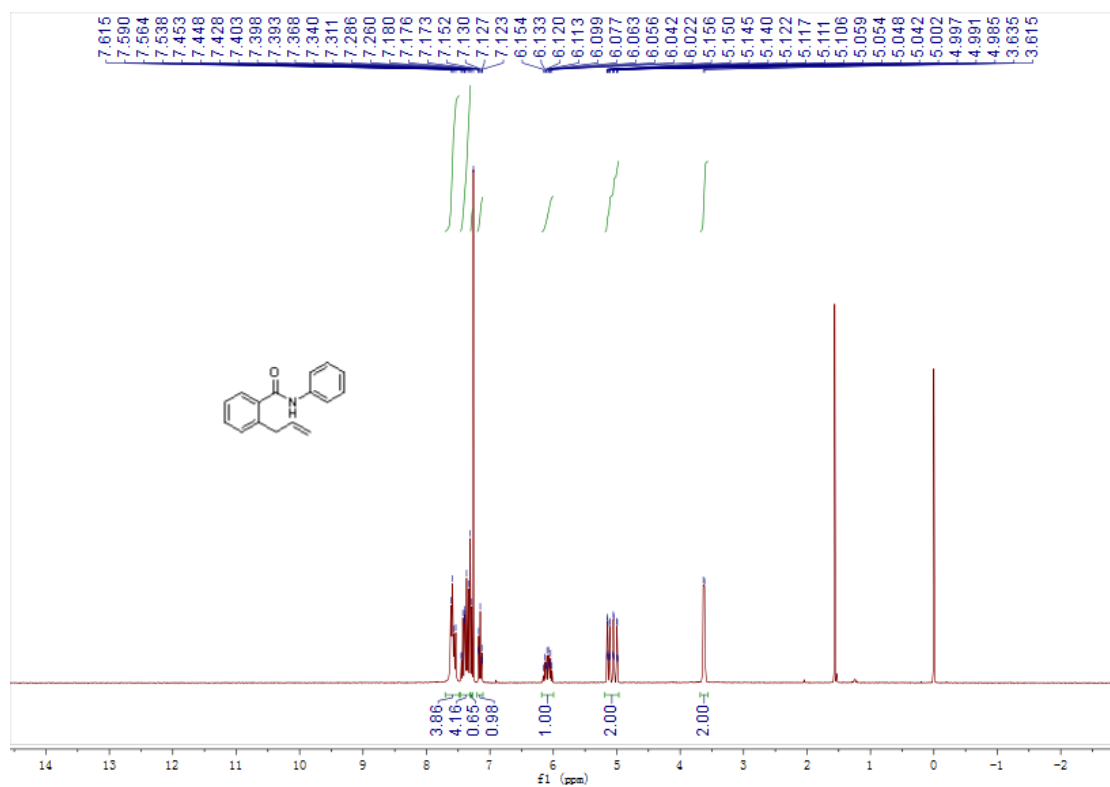


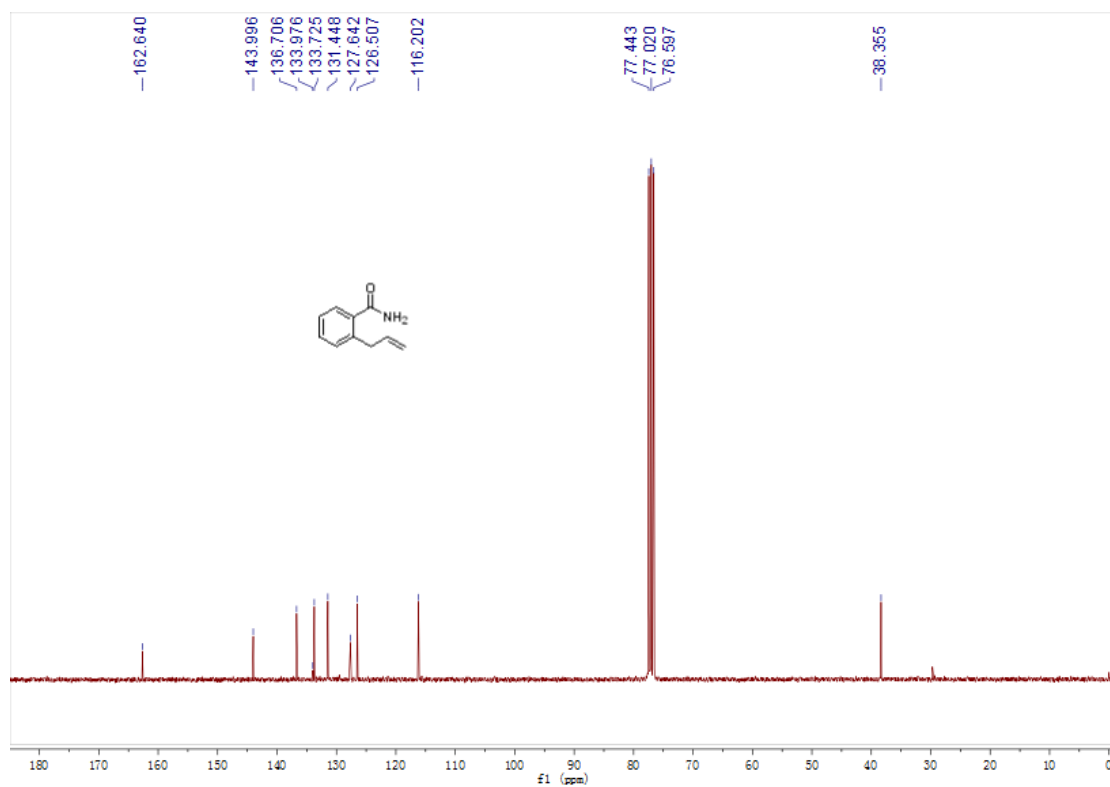
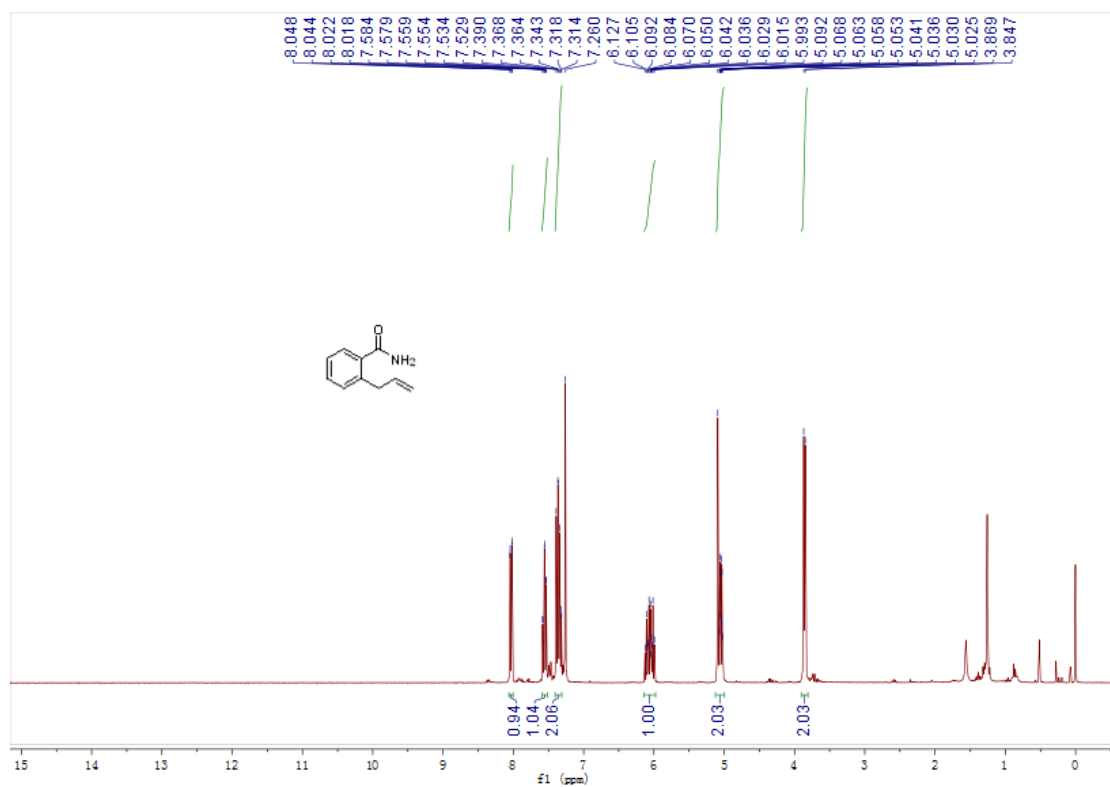




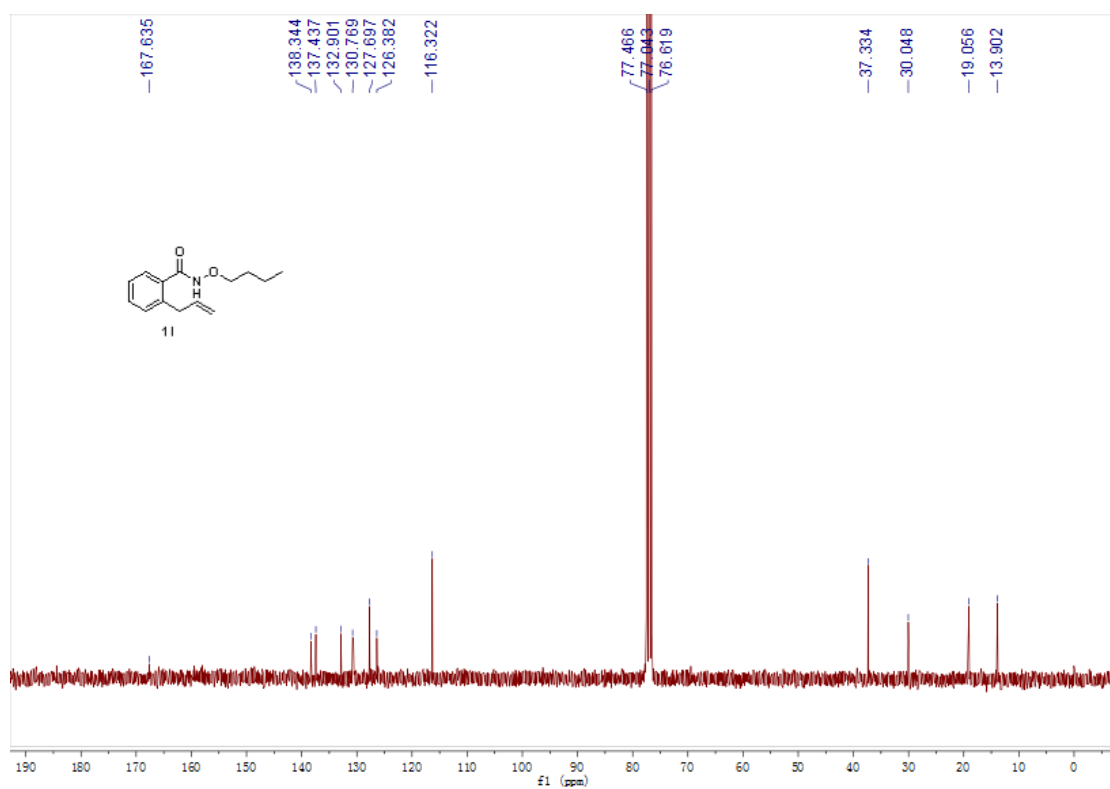
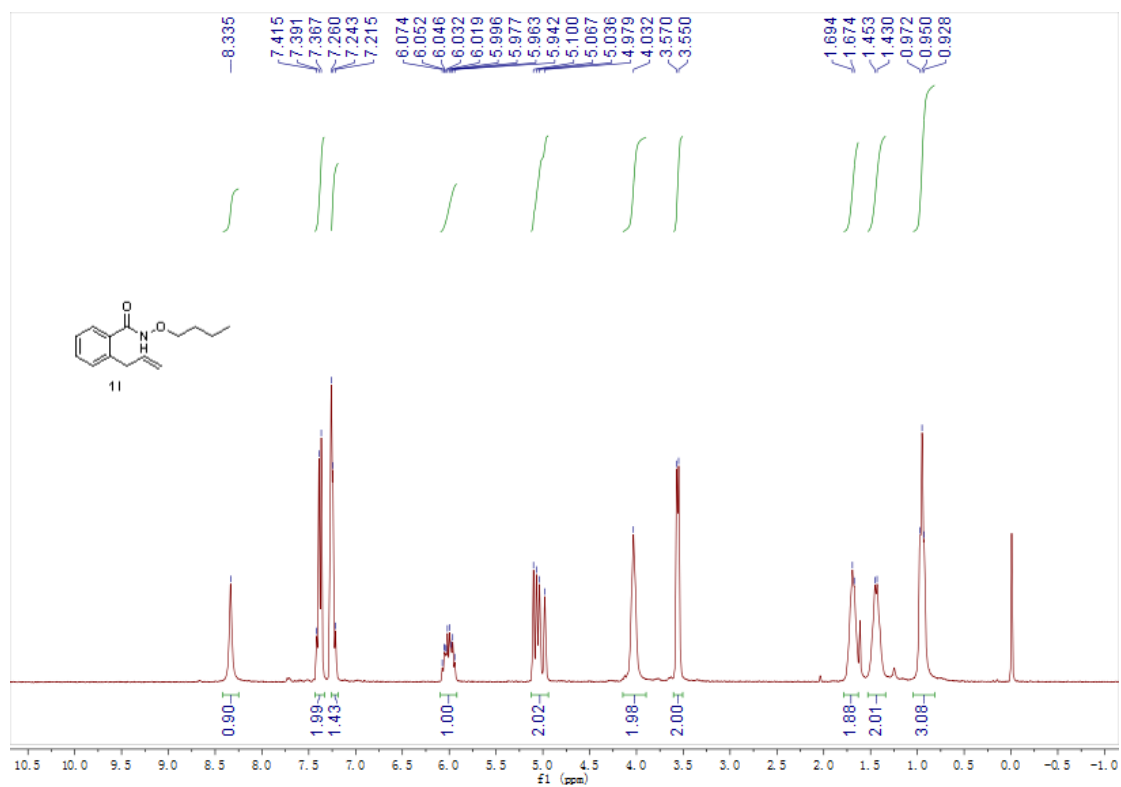


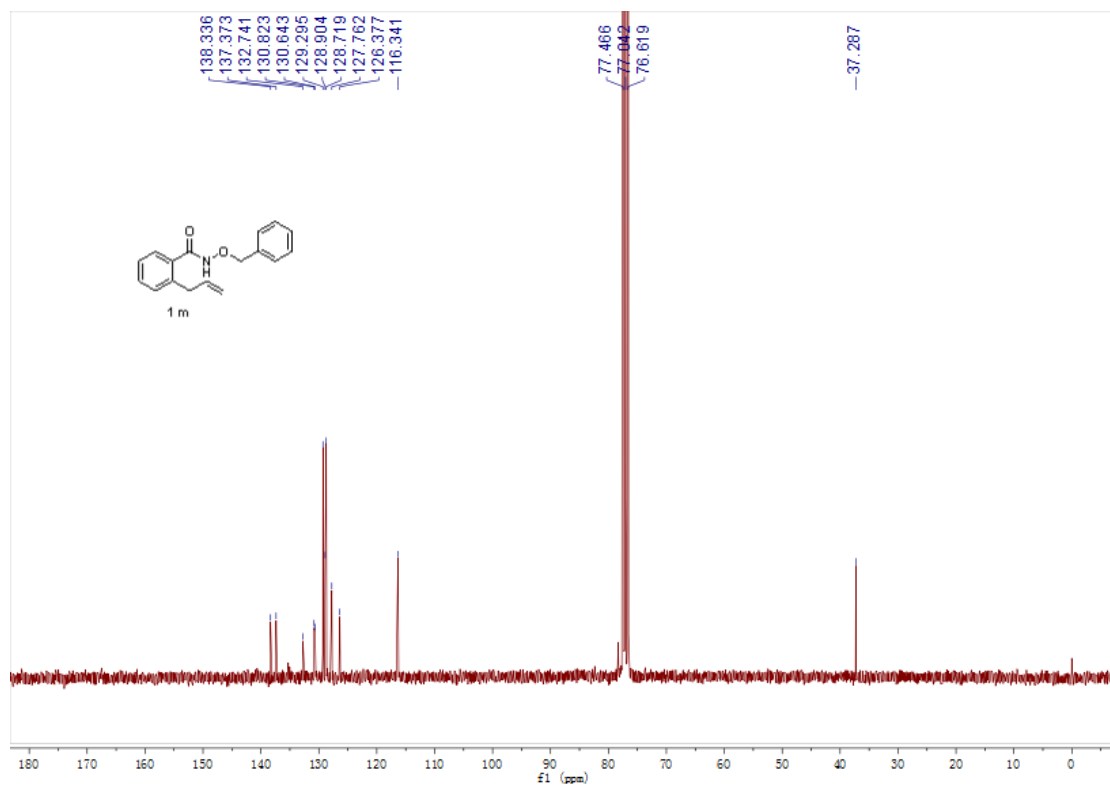
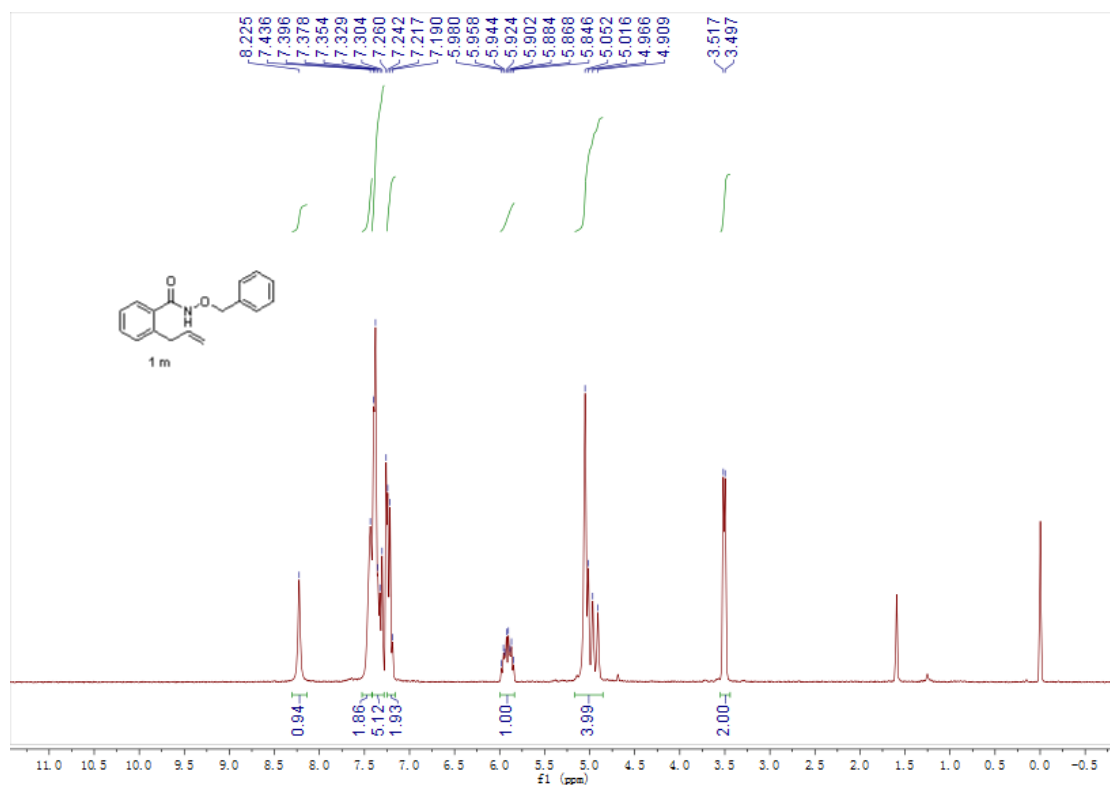


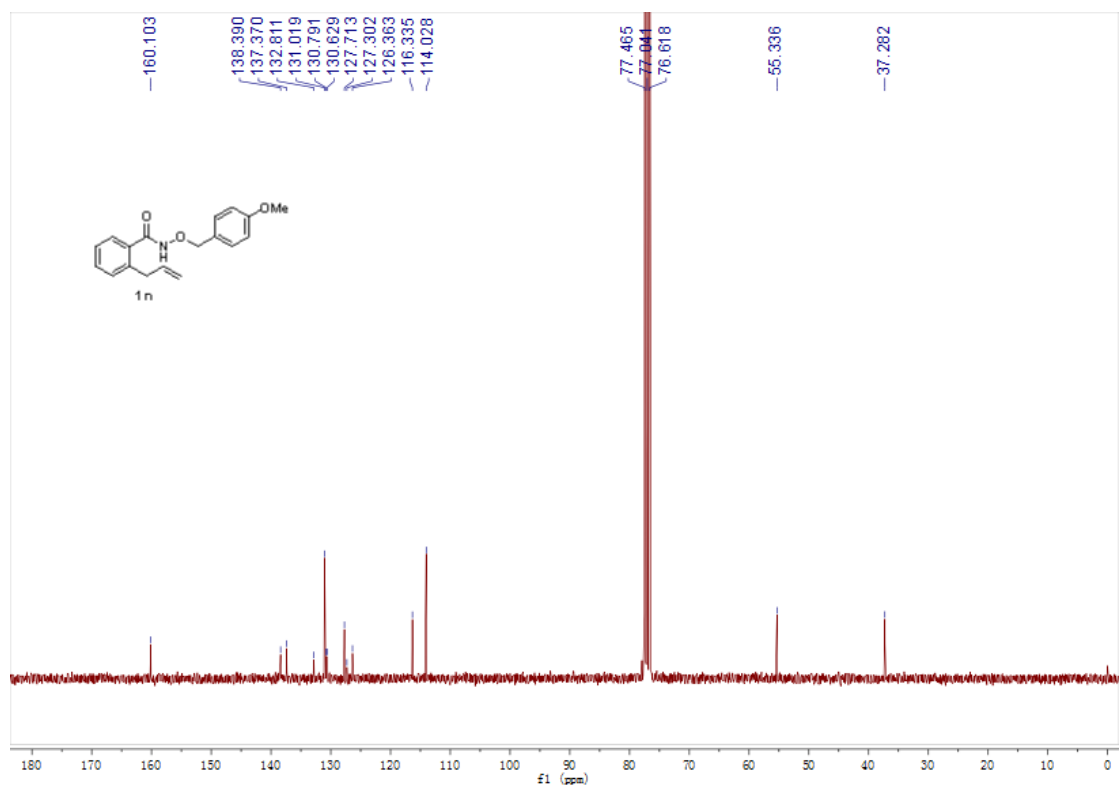
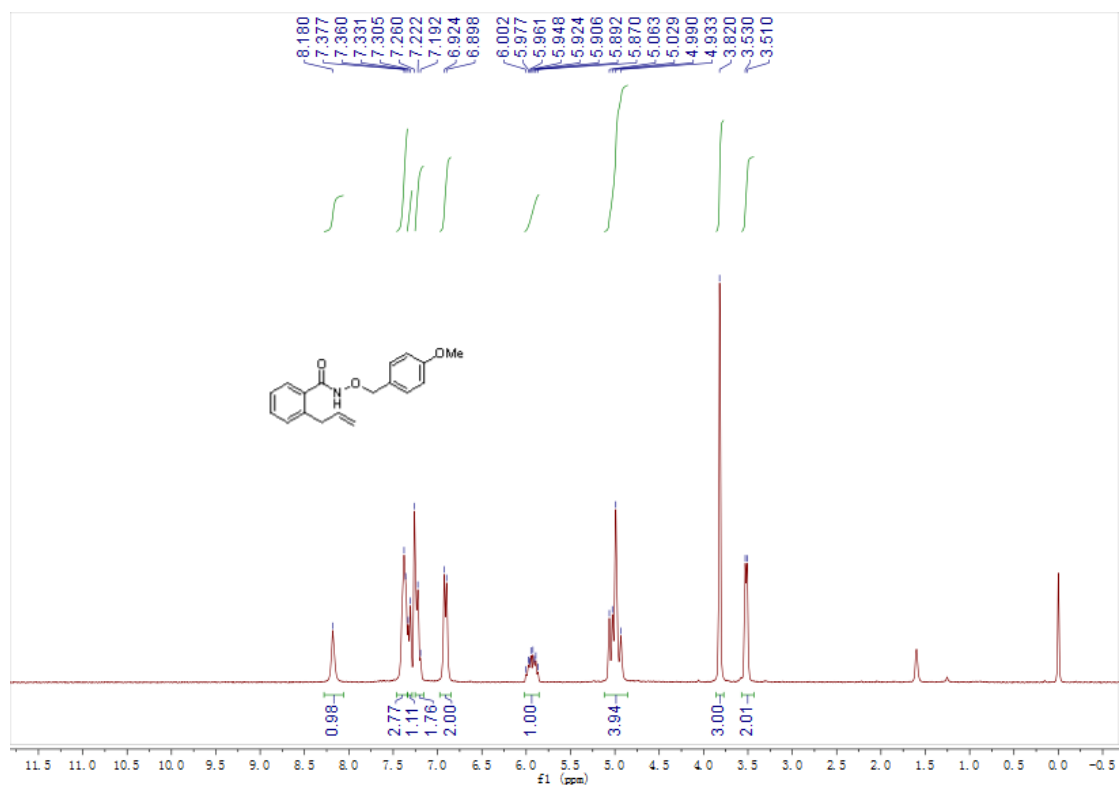


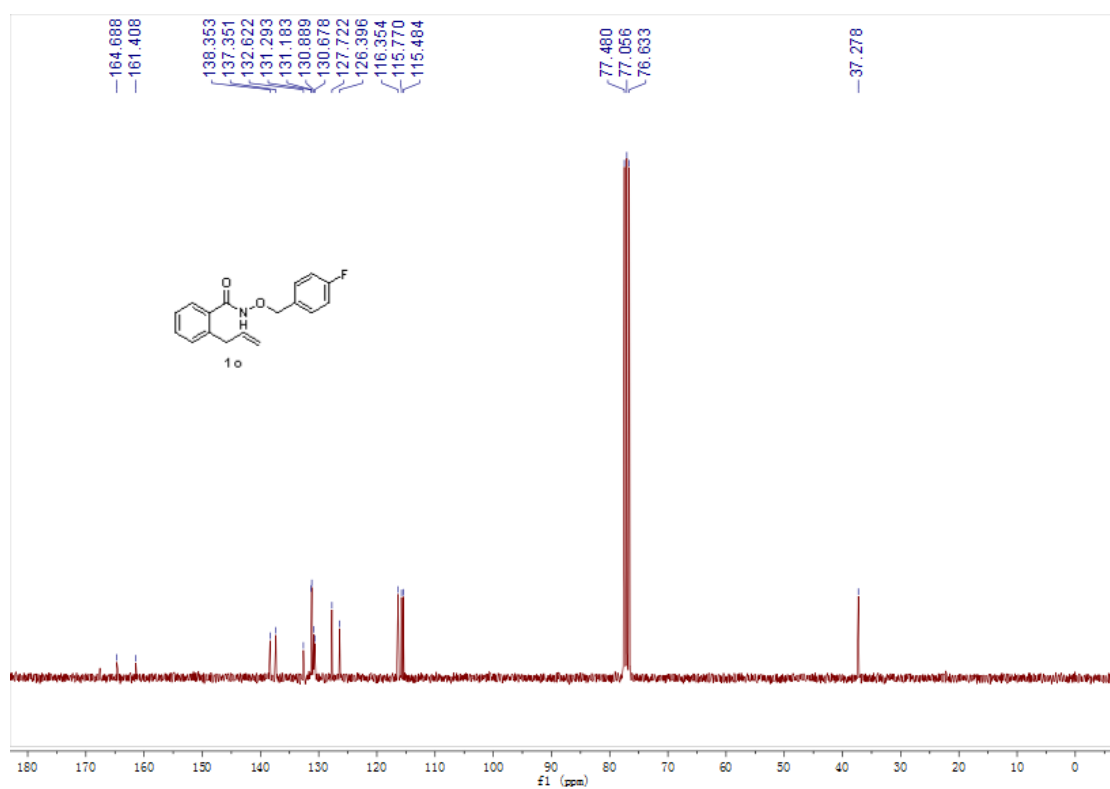
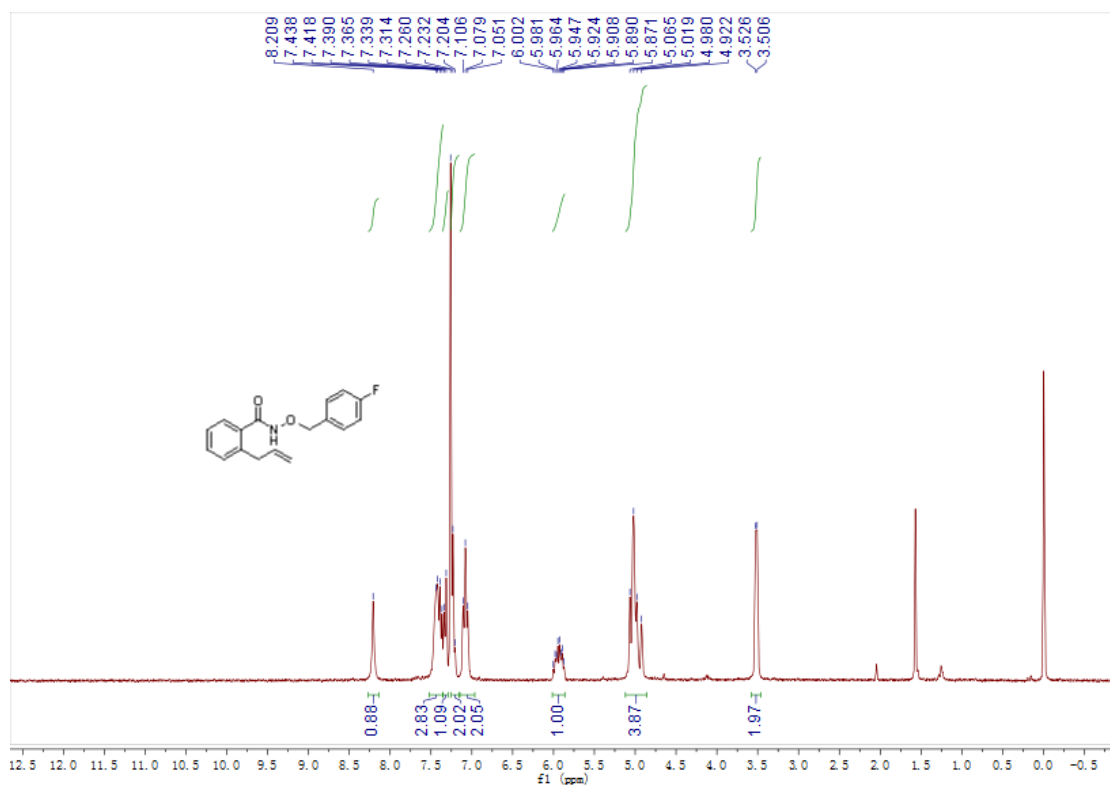


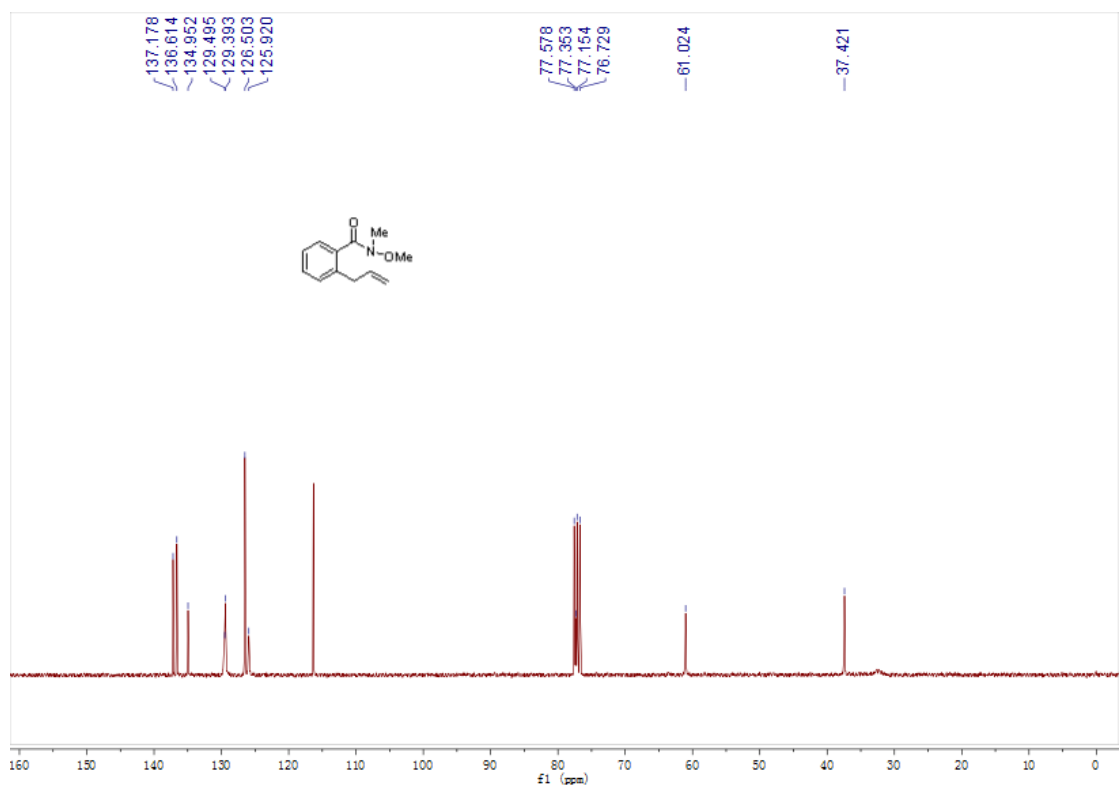
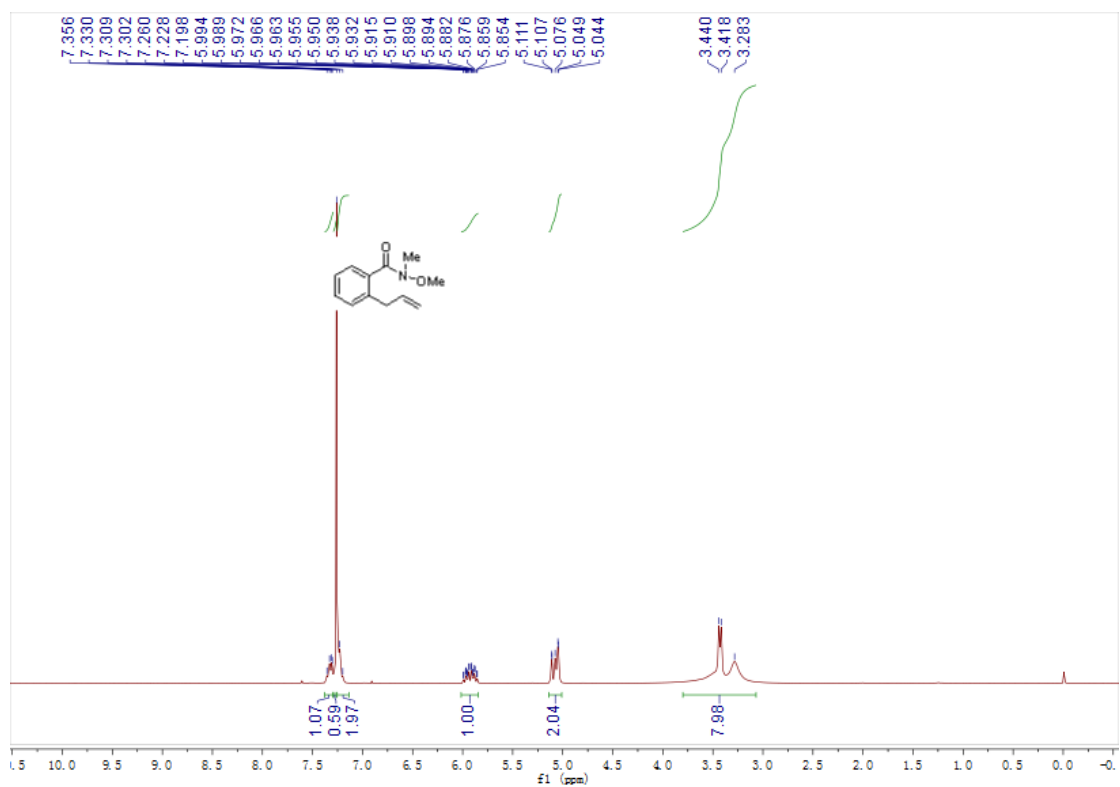




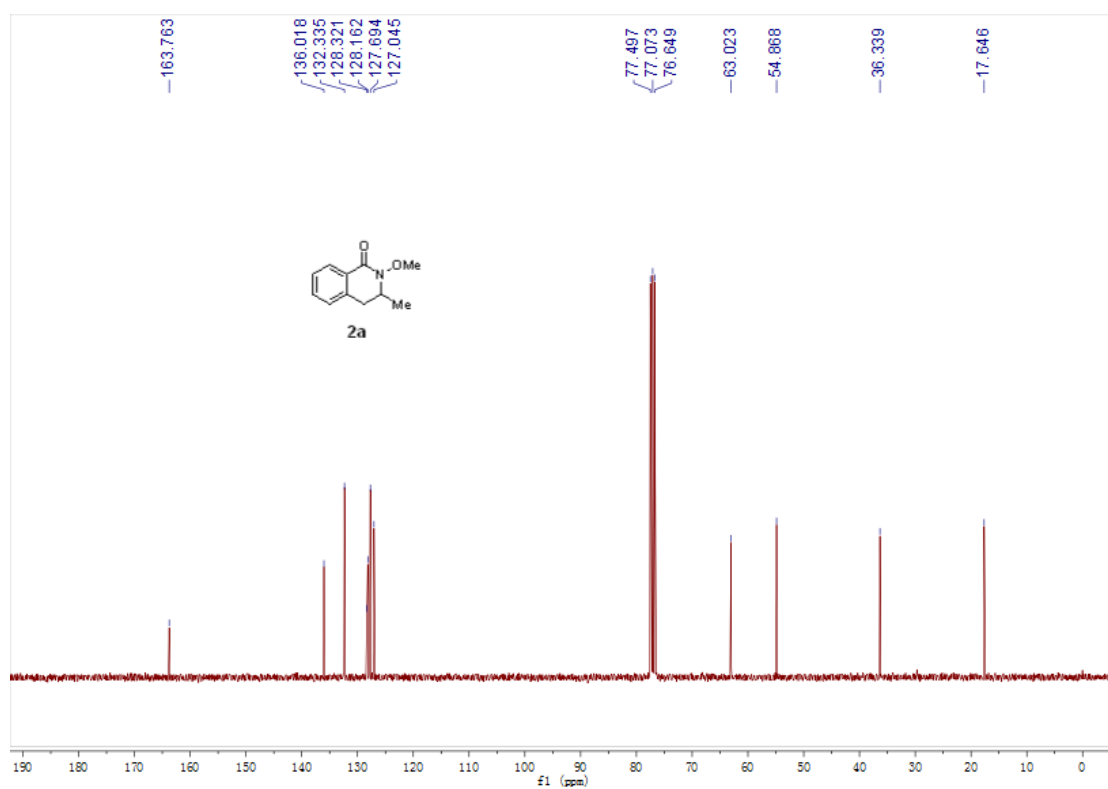
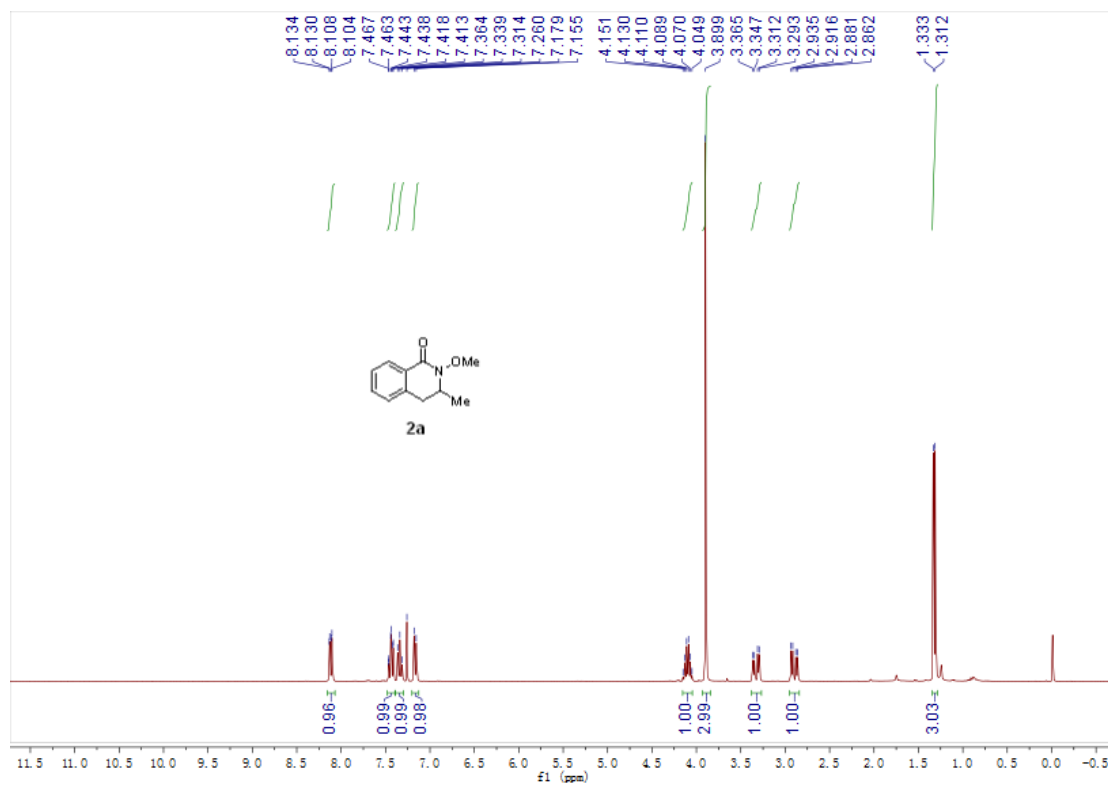


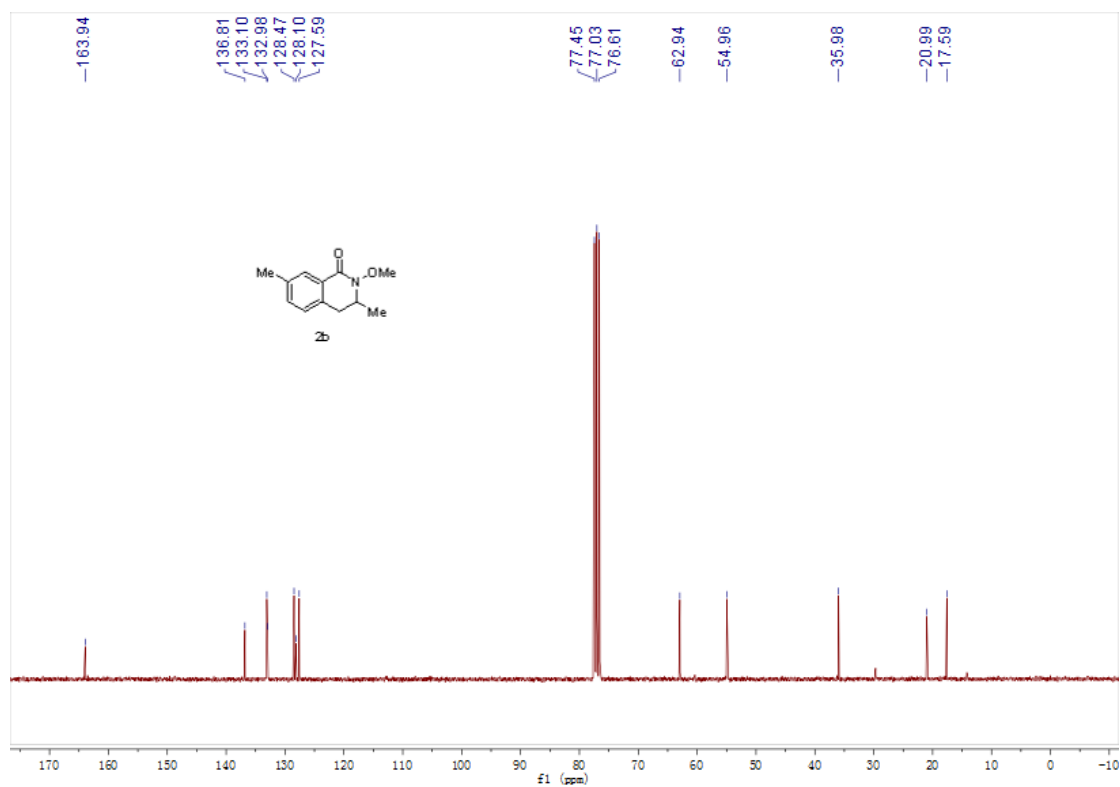
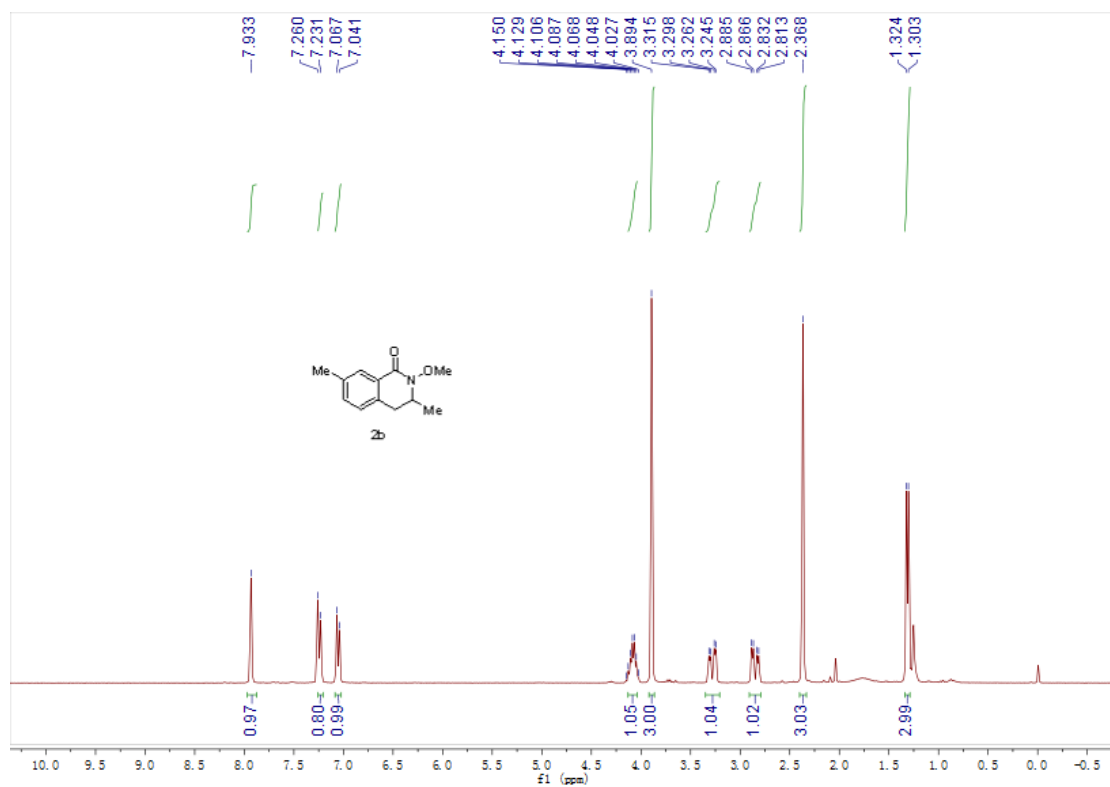


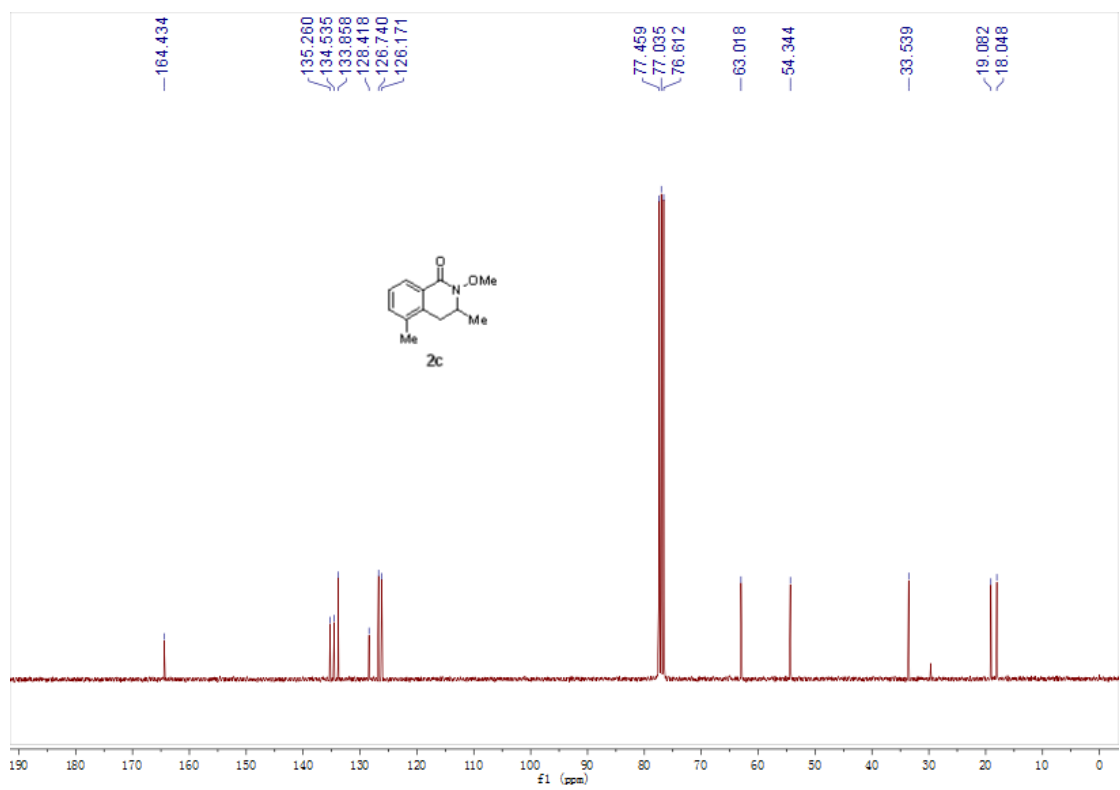
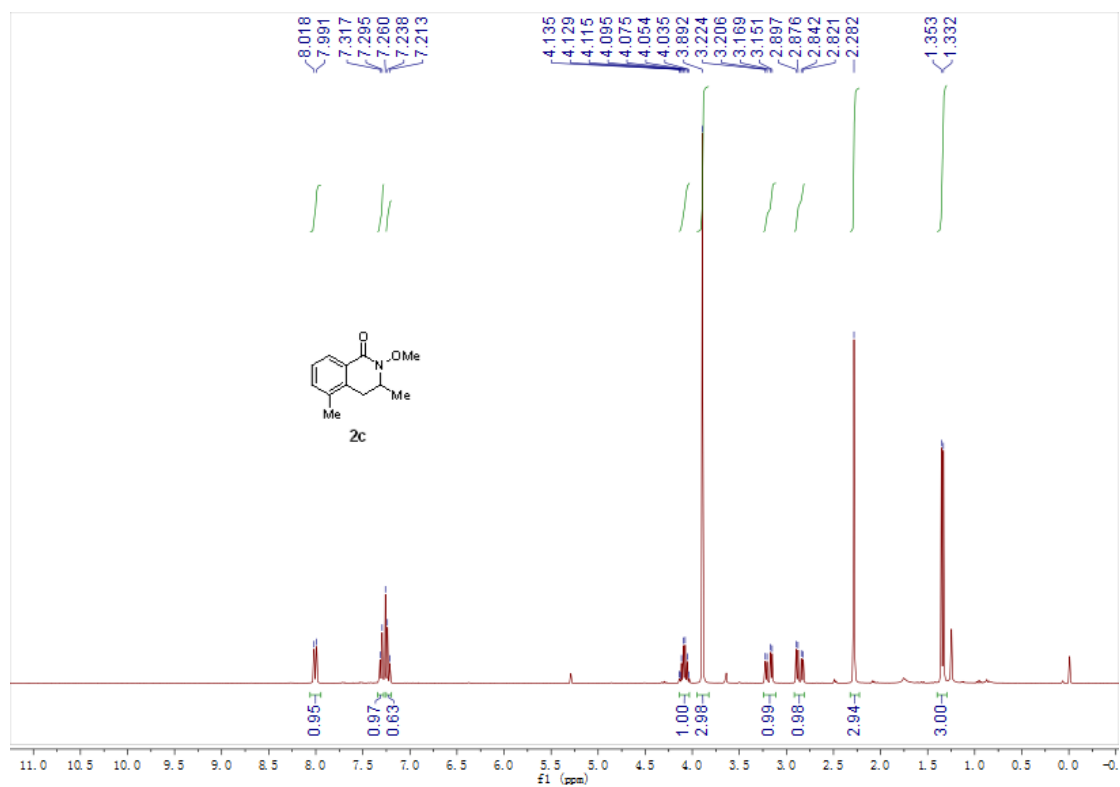




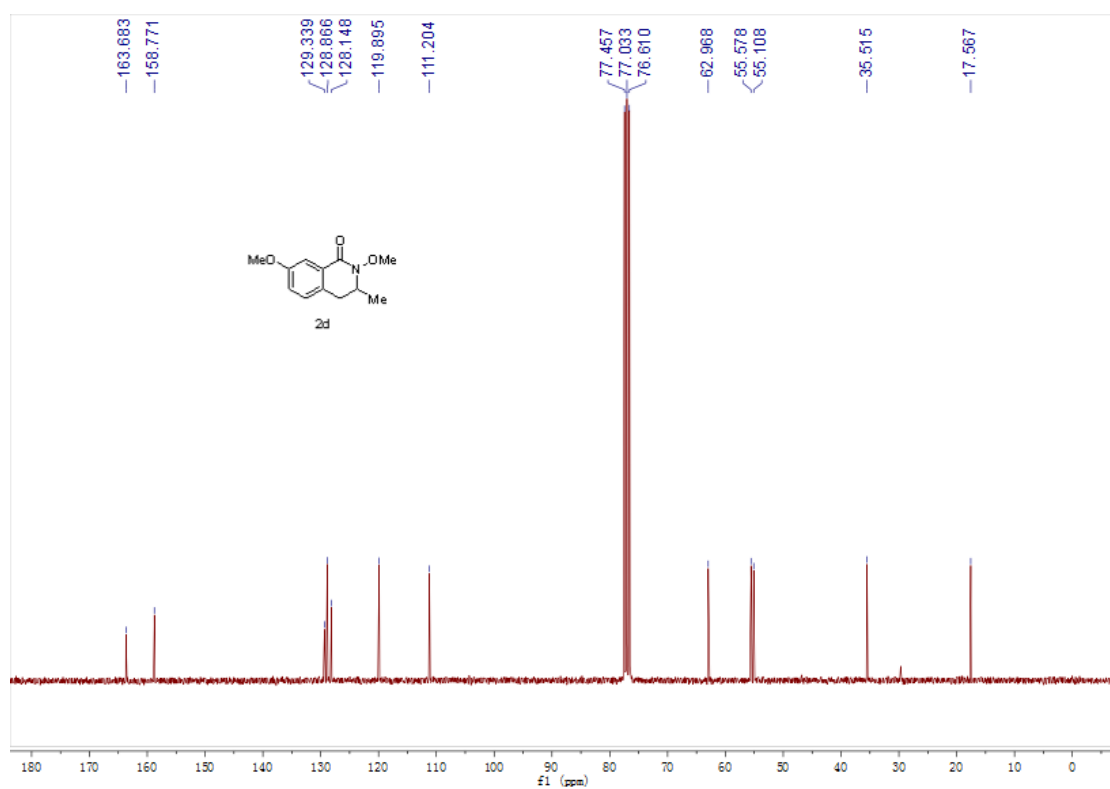
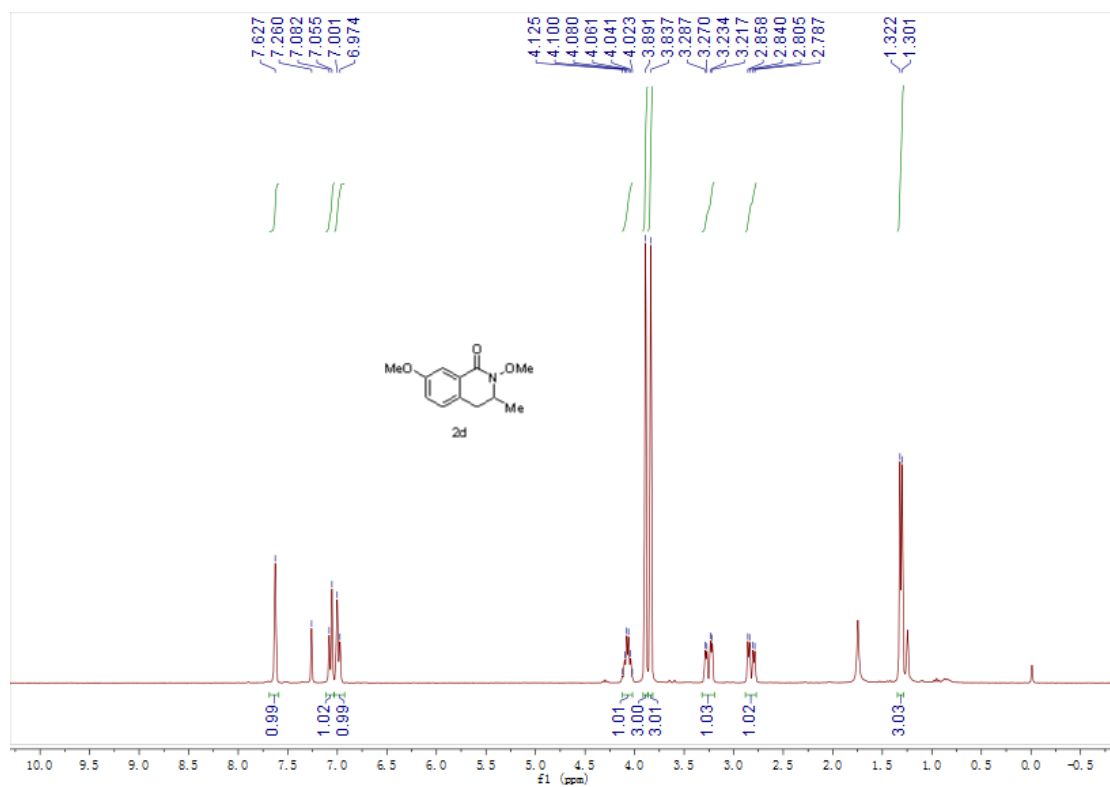
<sup>1</sup>H NMR, <sup>13</sup>C NMR spectra of Products:

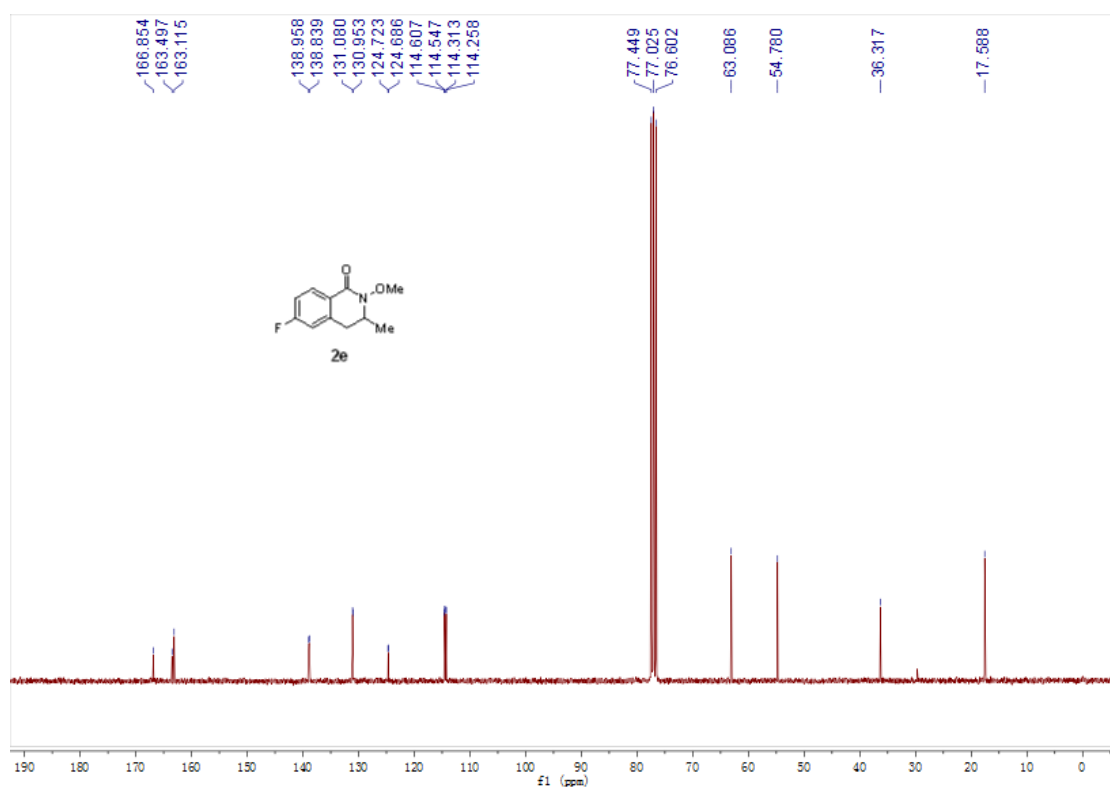
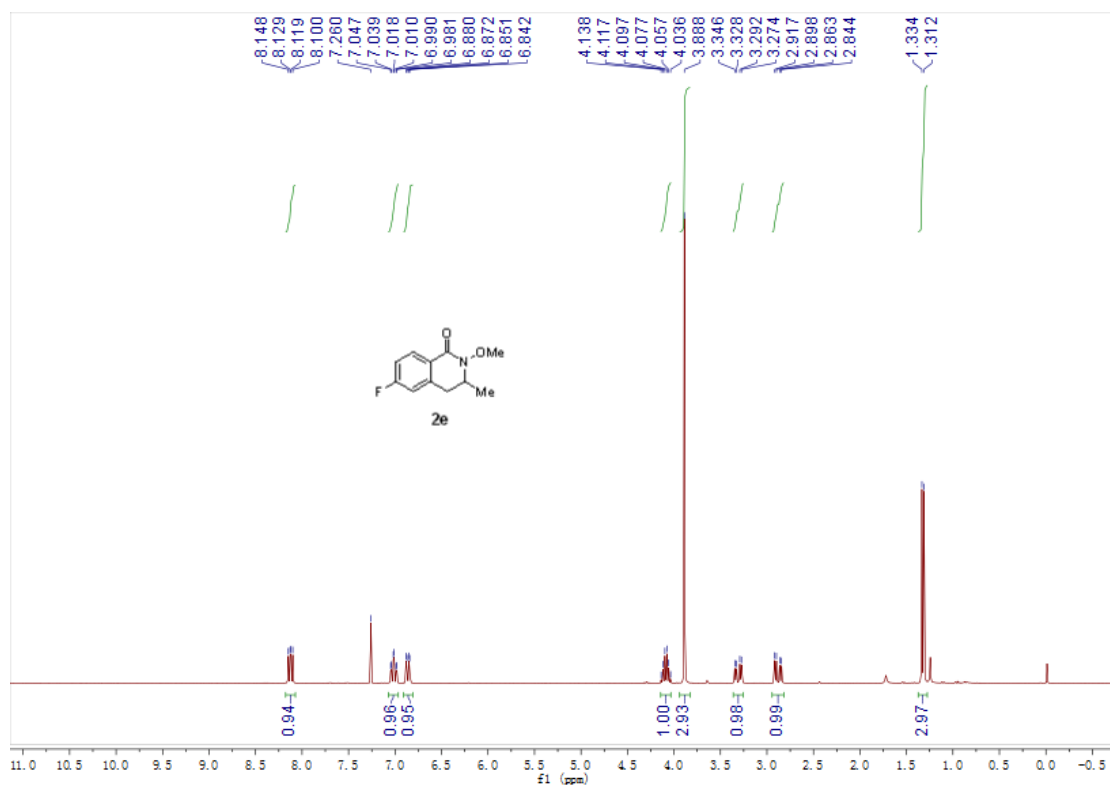


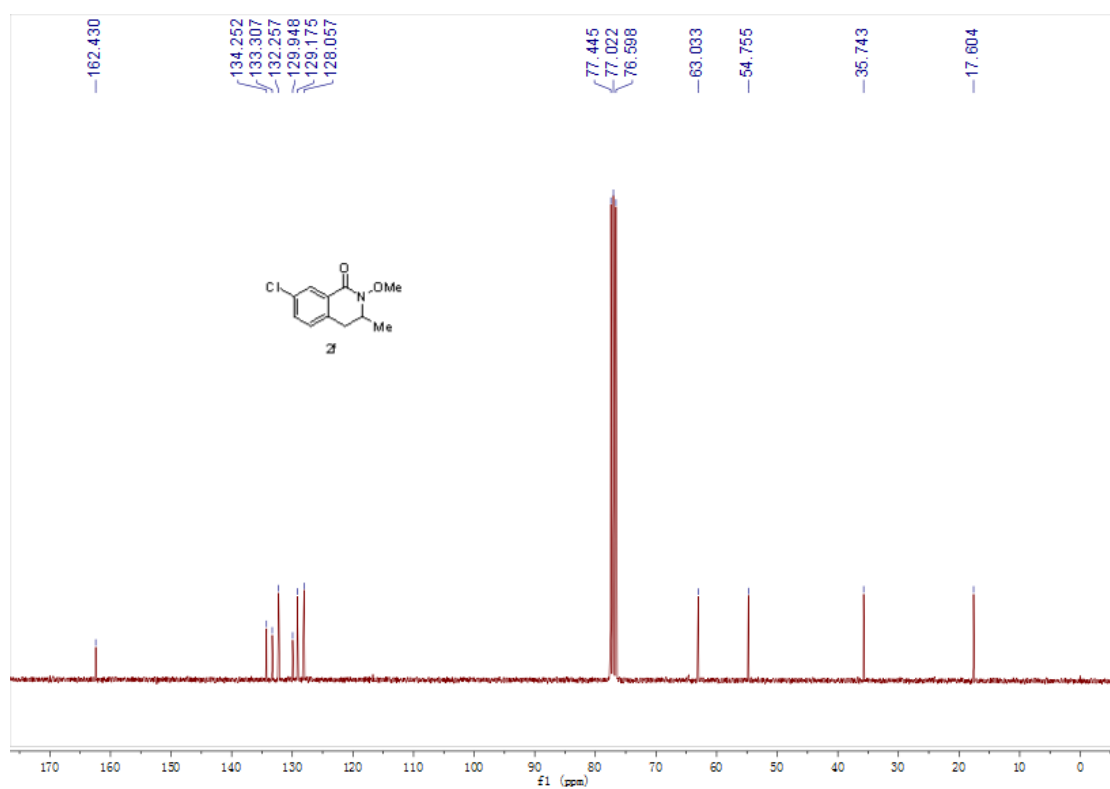
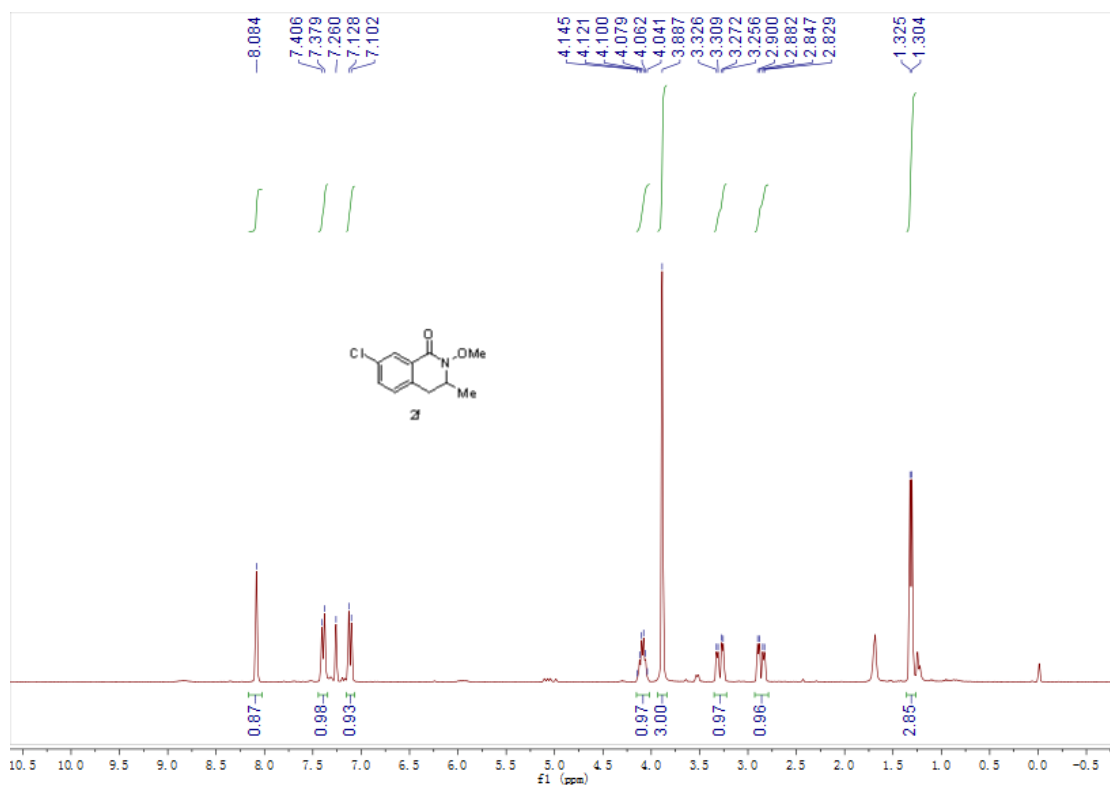


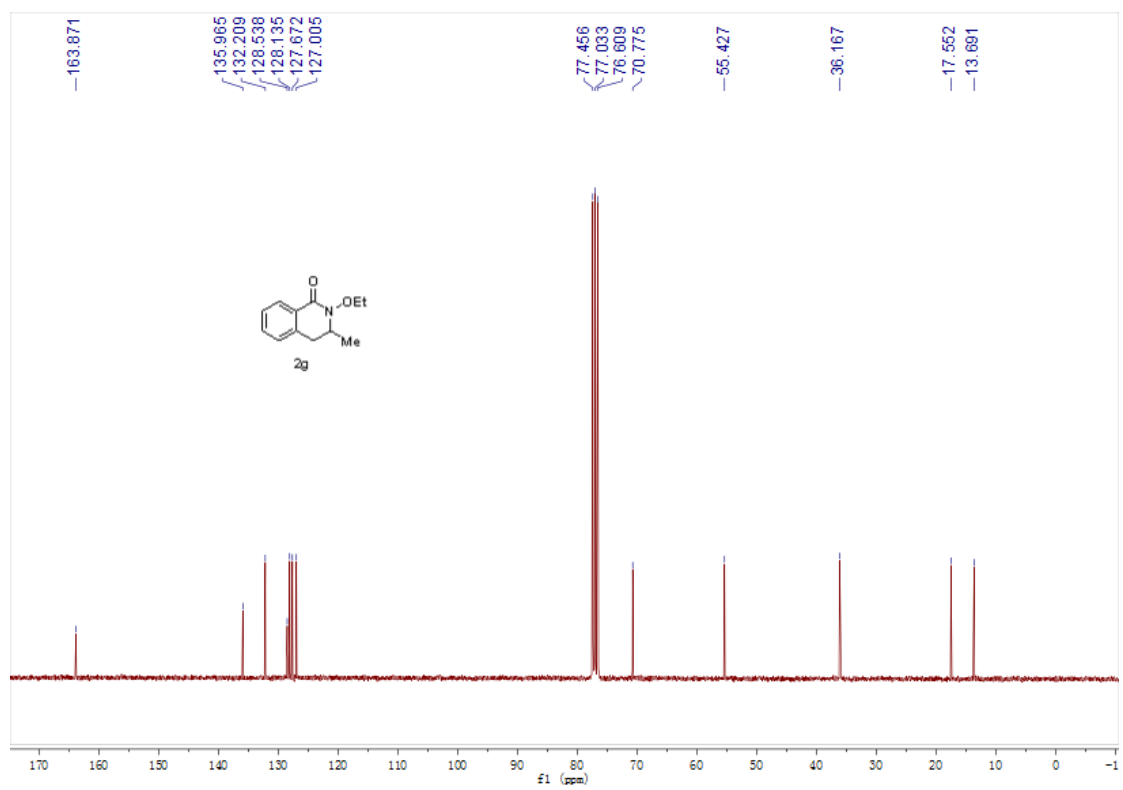
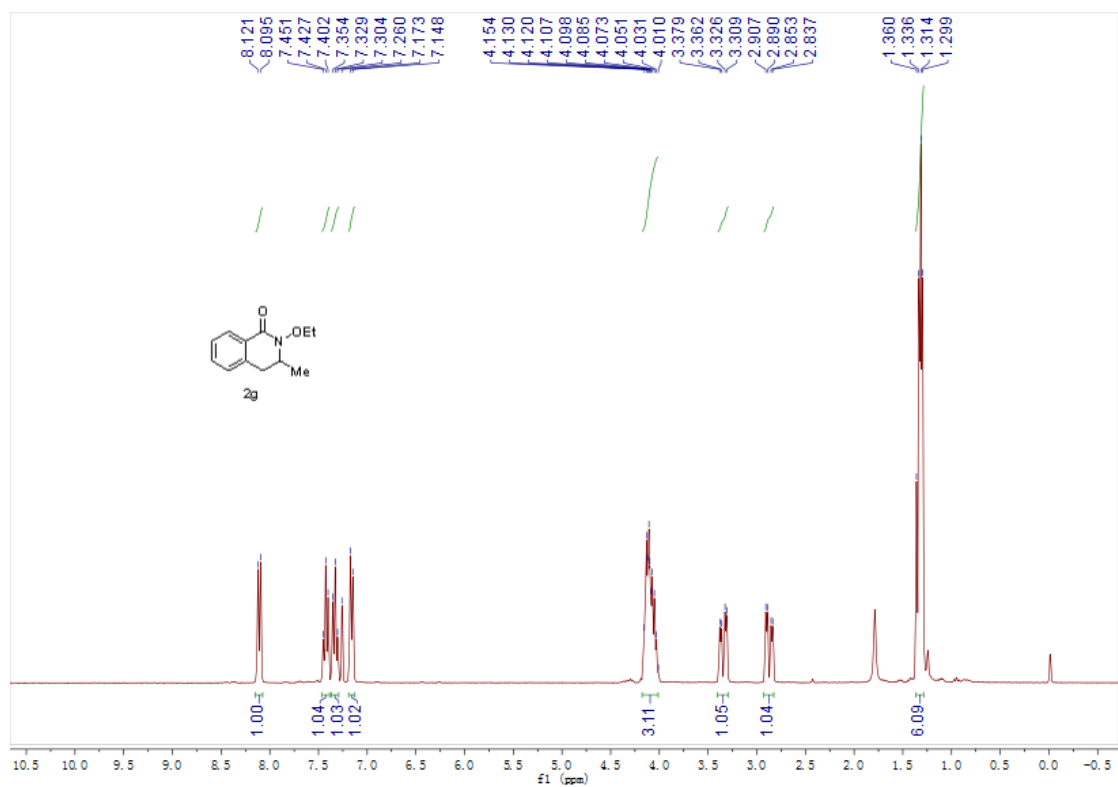


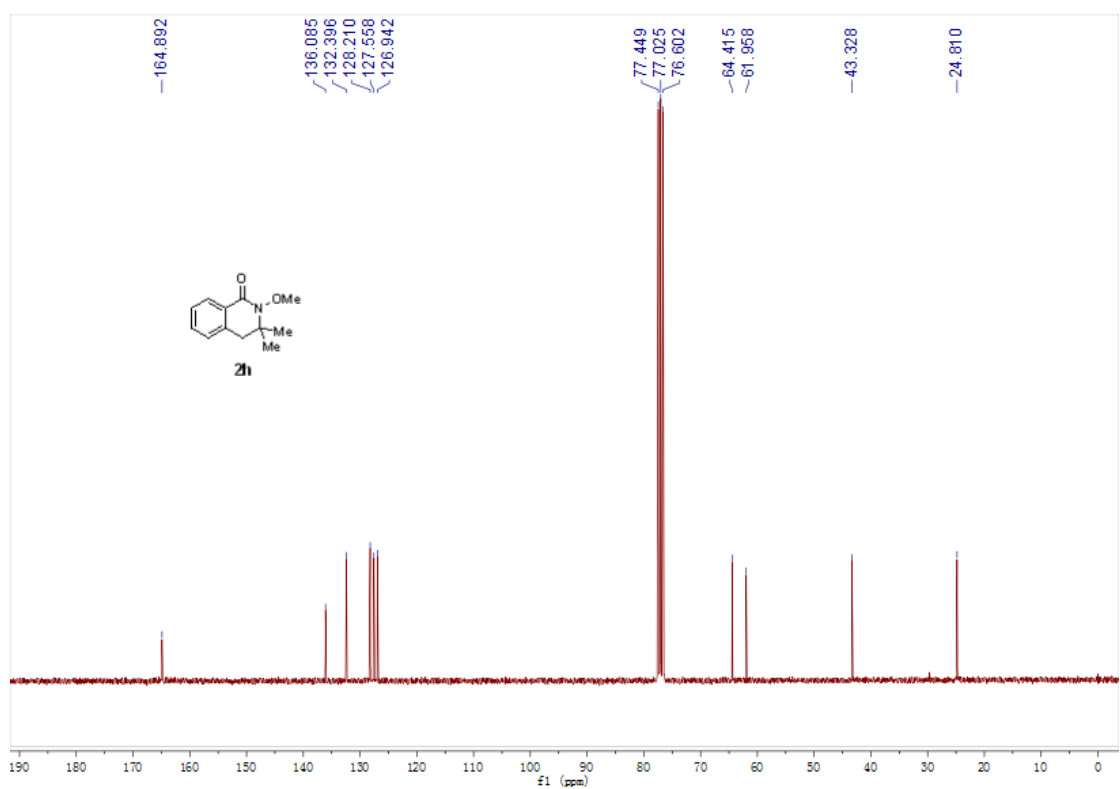
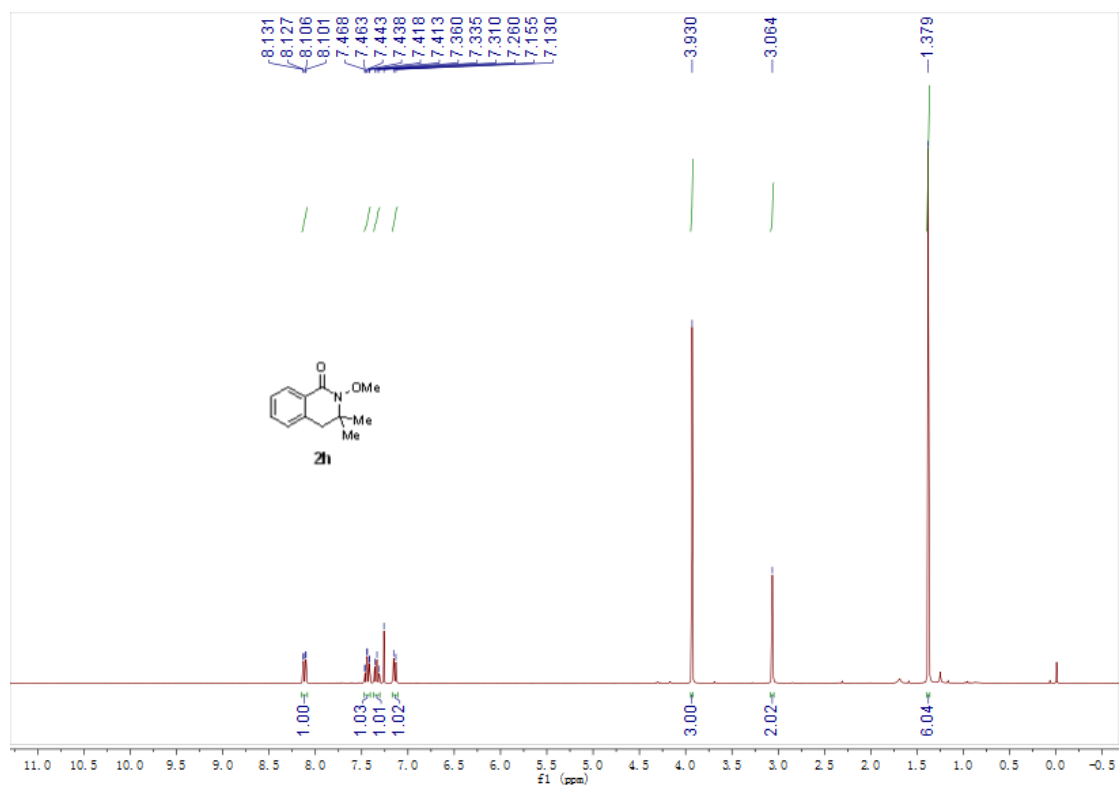


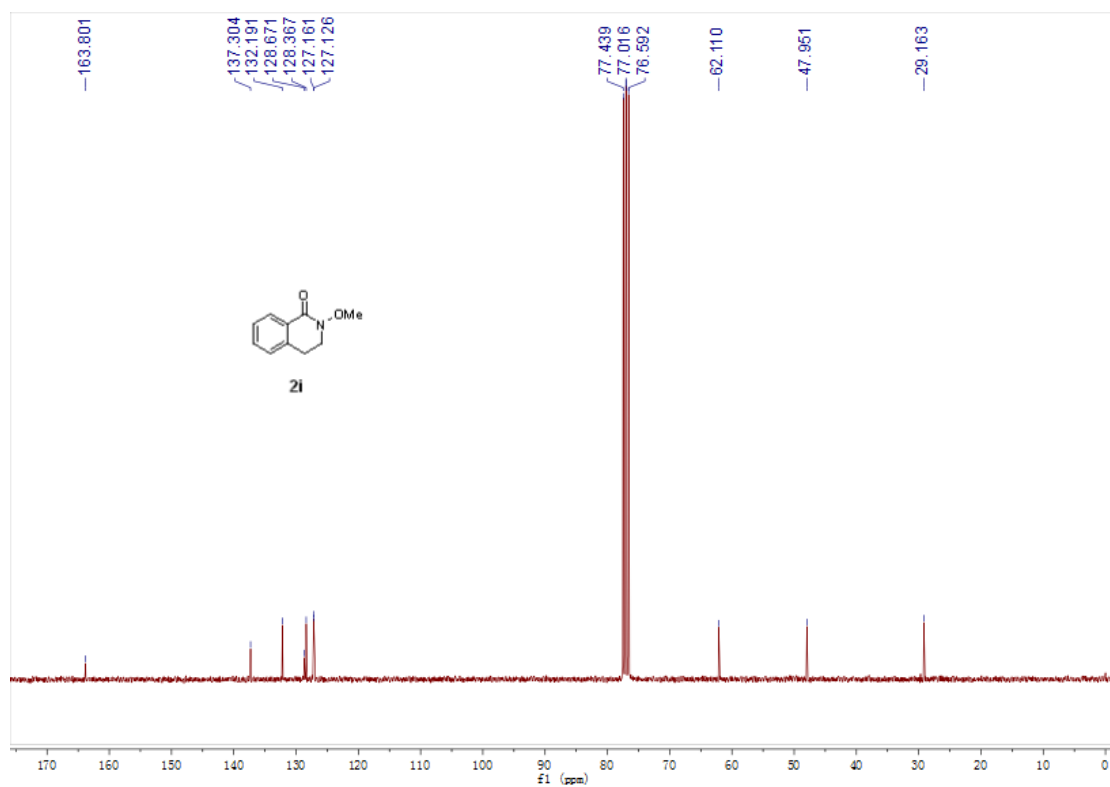
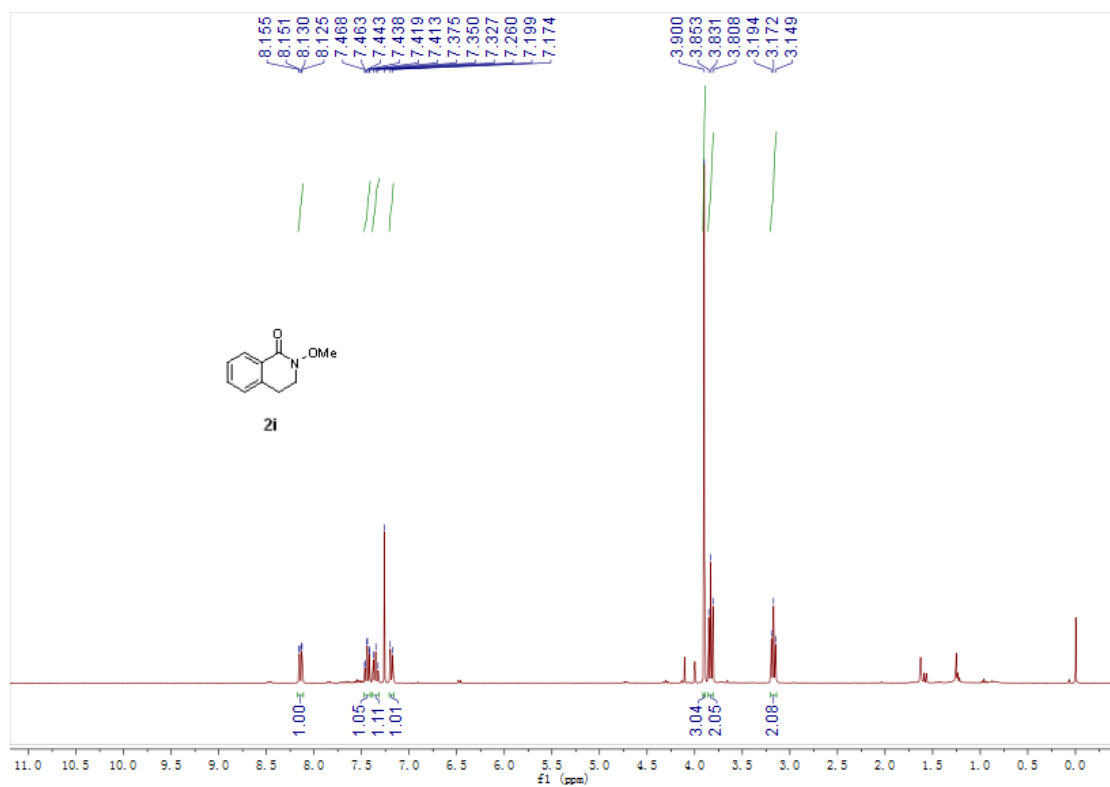


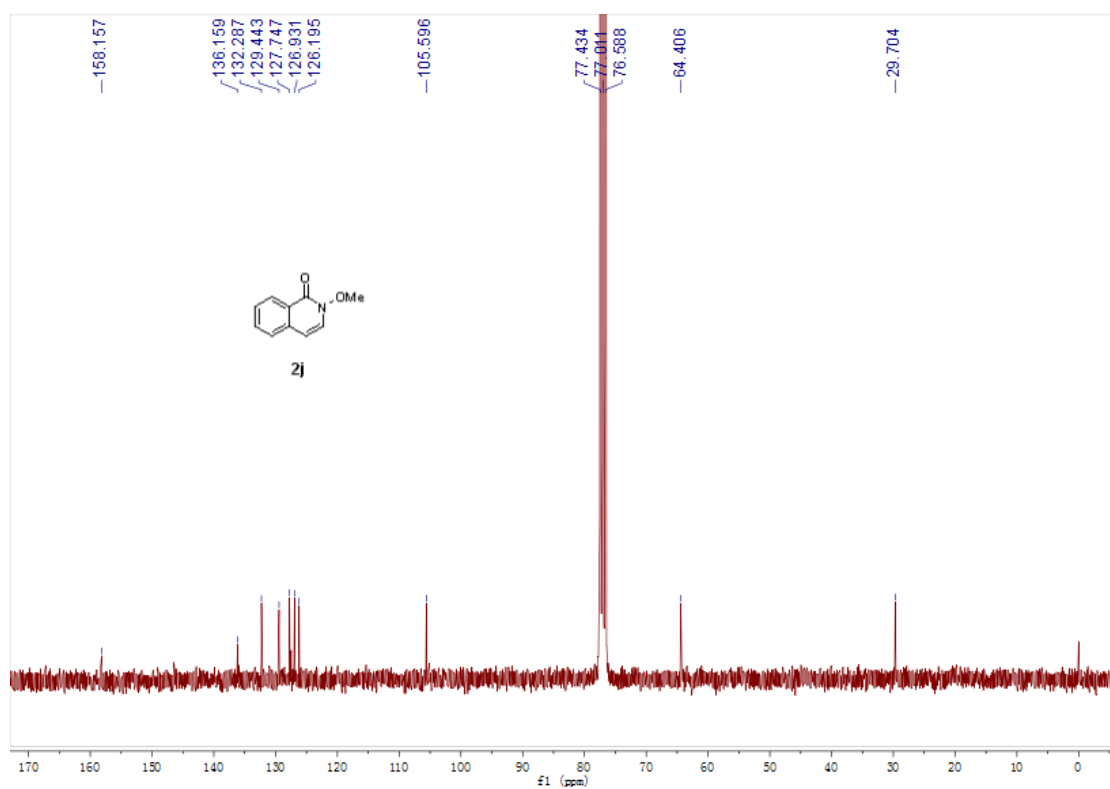
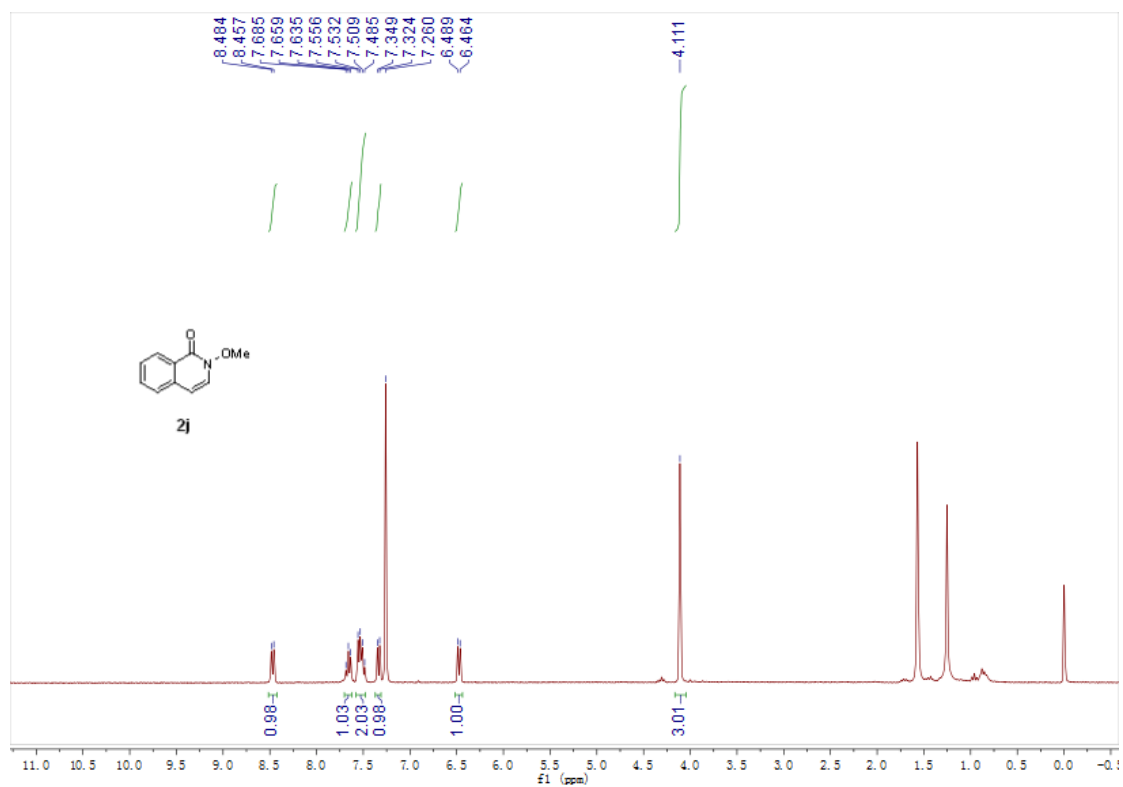


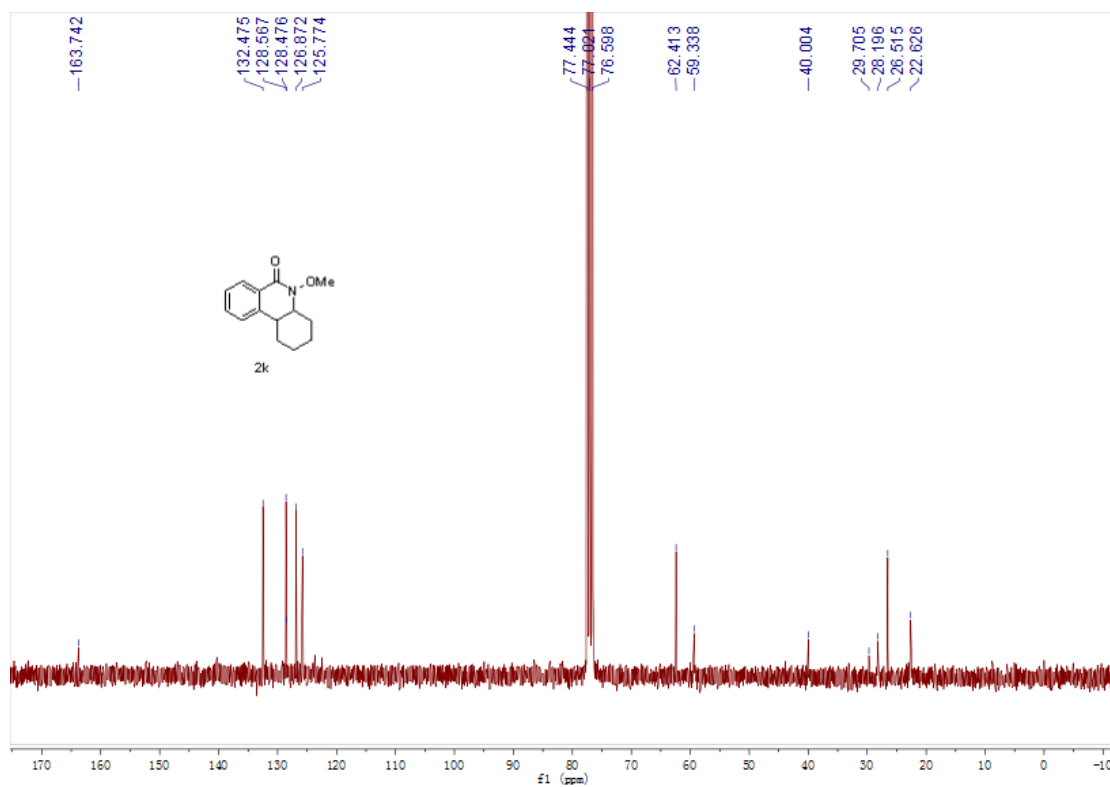
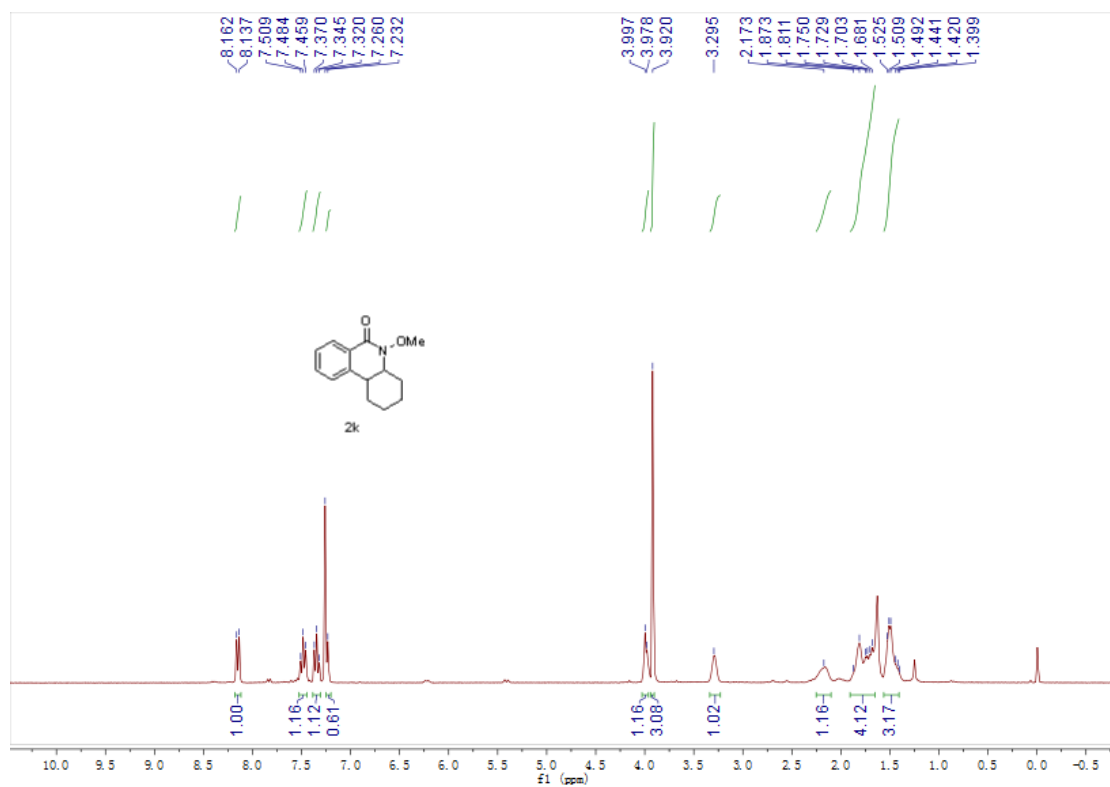




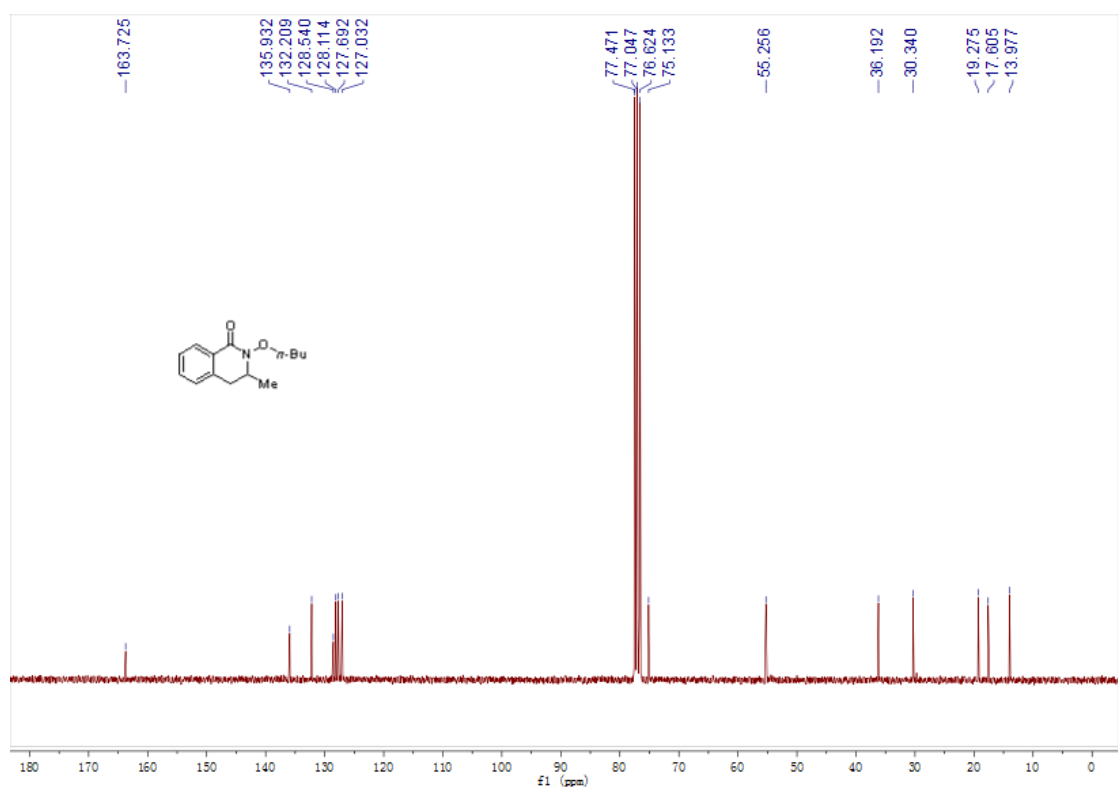
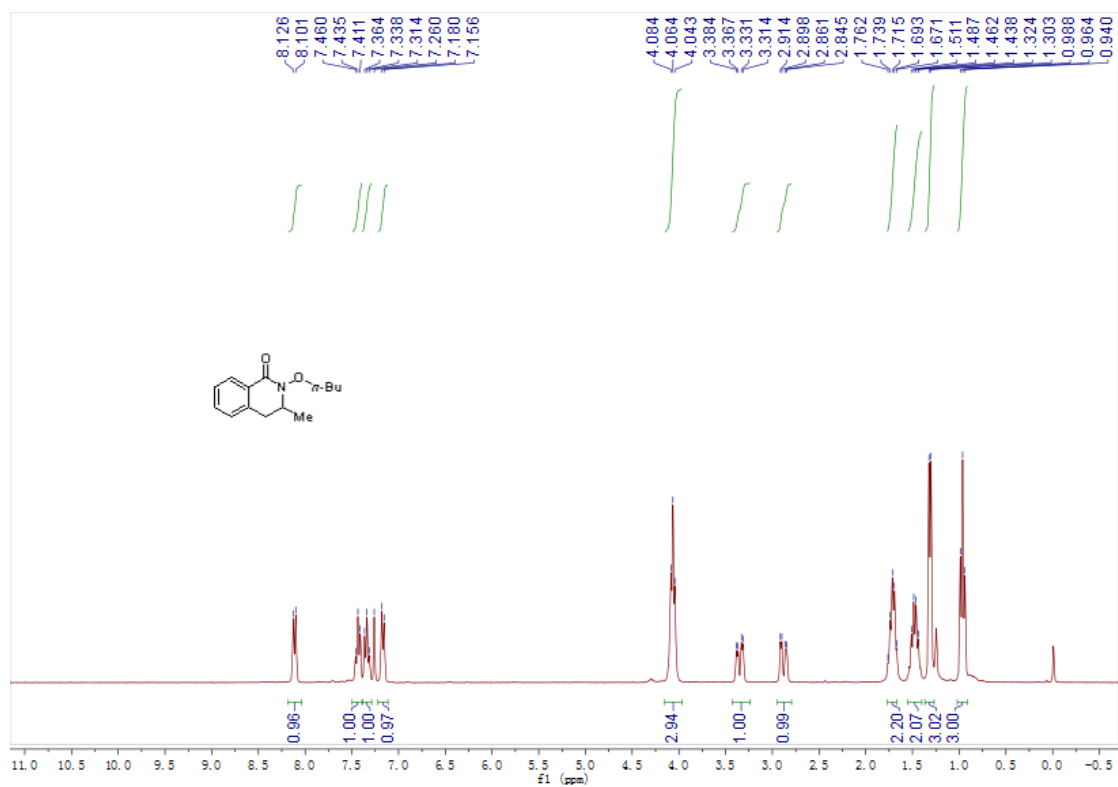


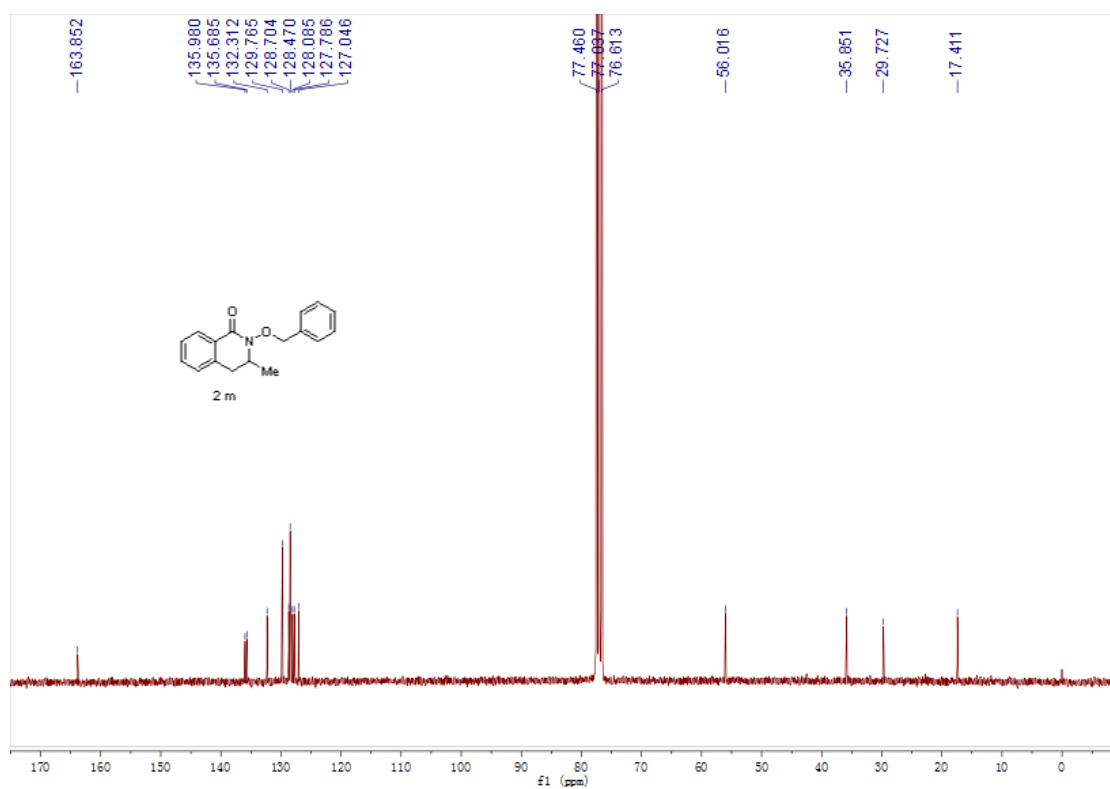
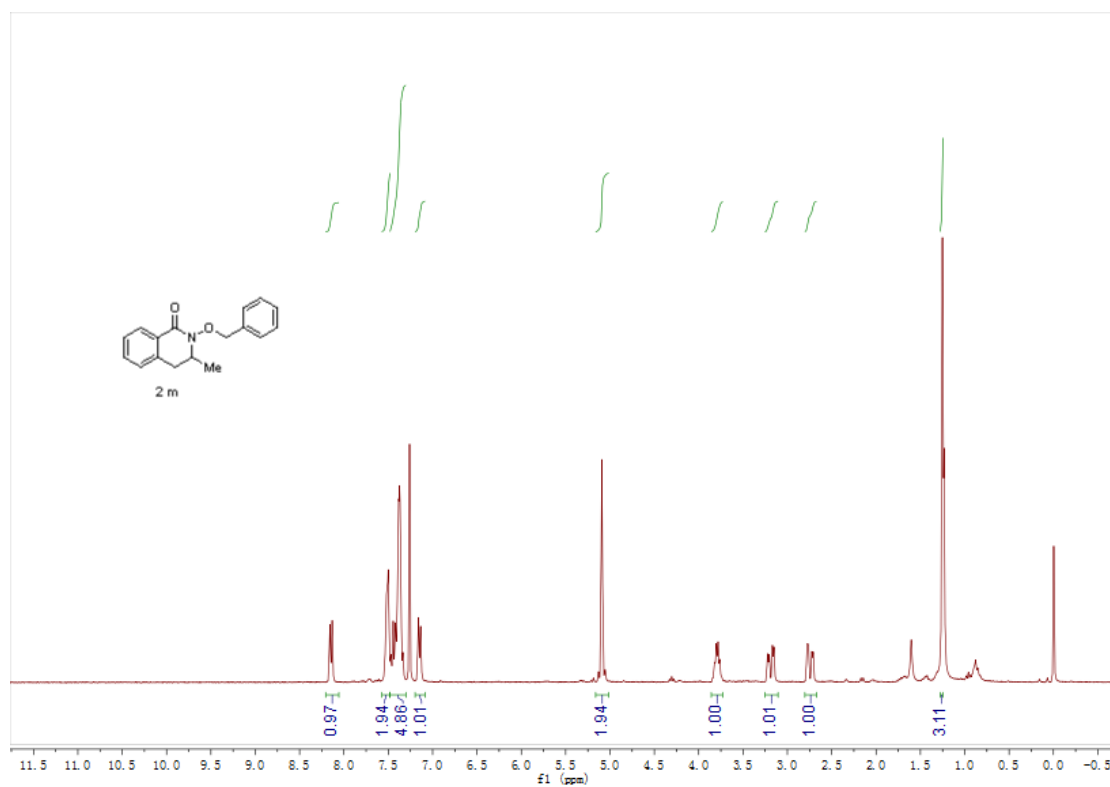


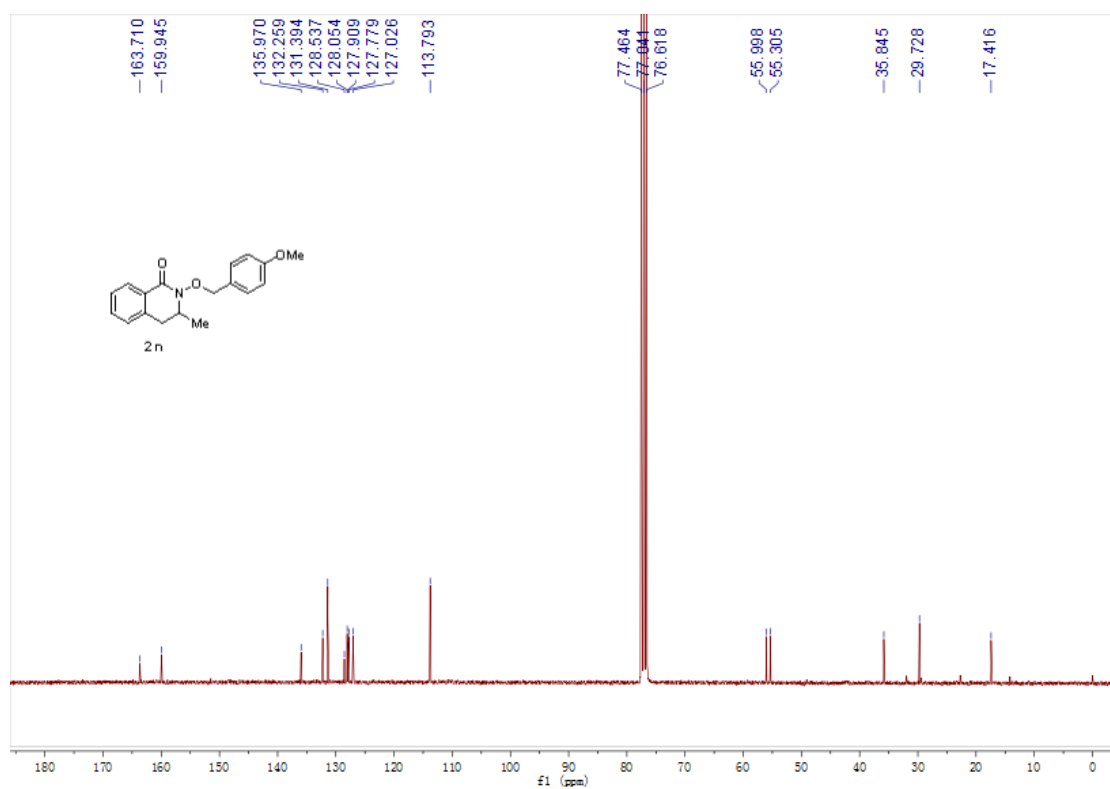
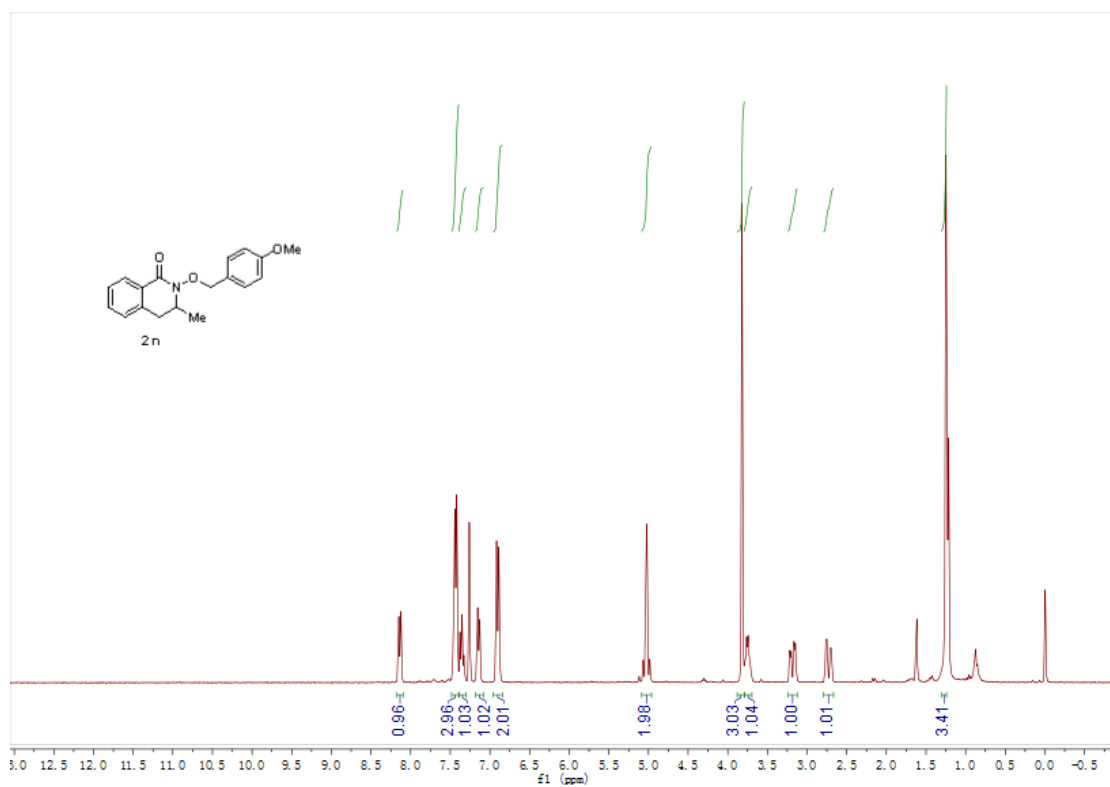


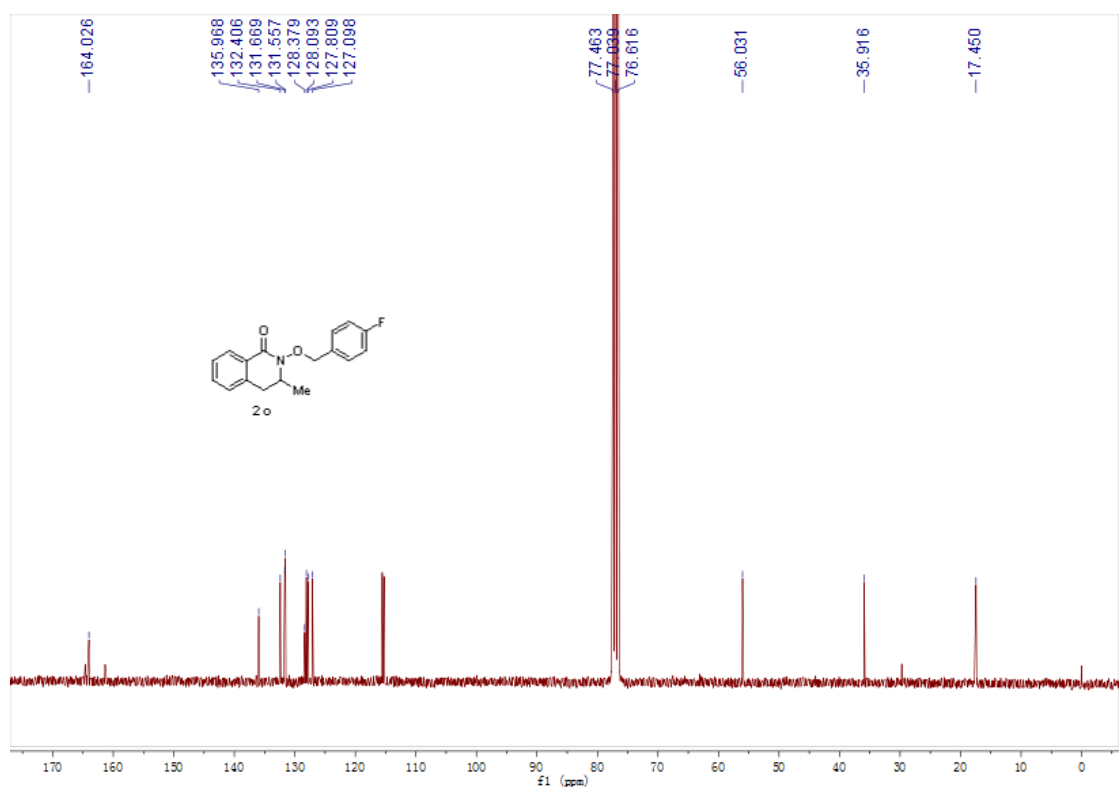
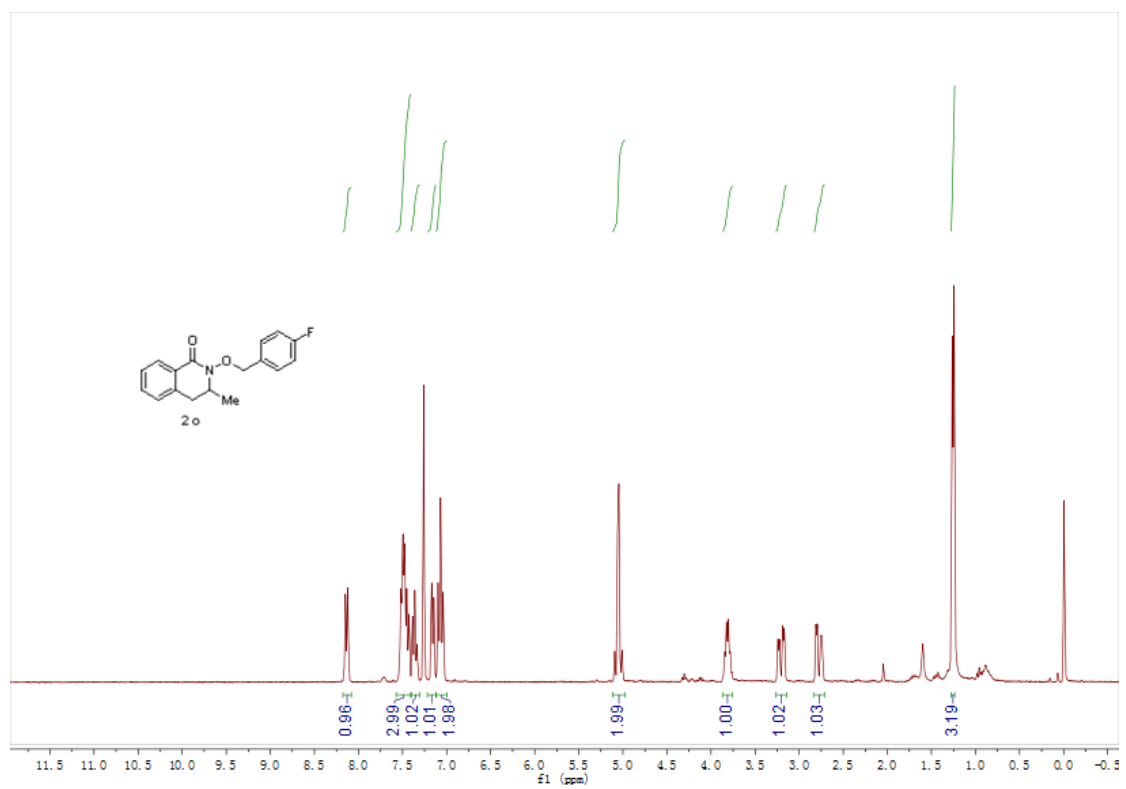


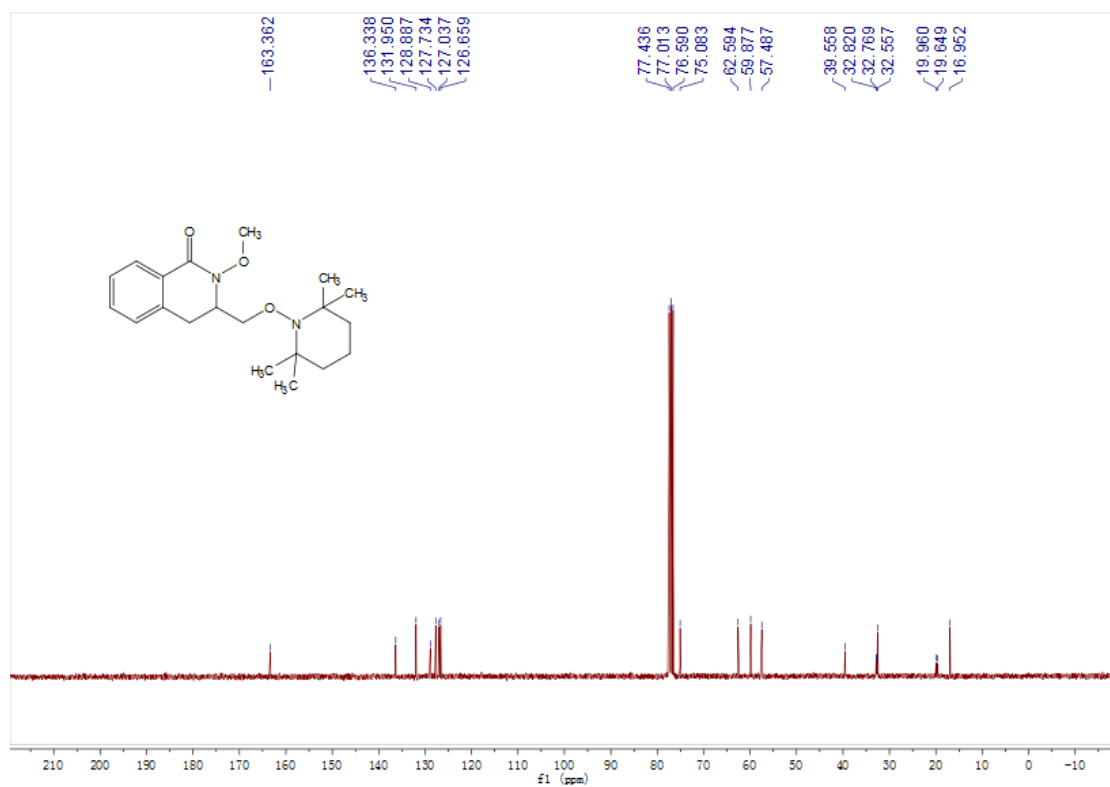
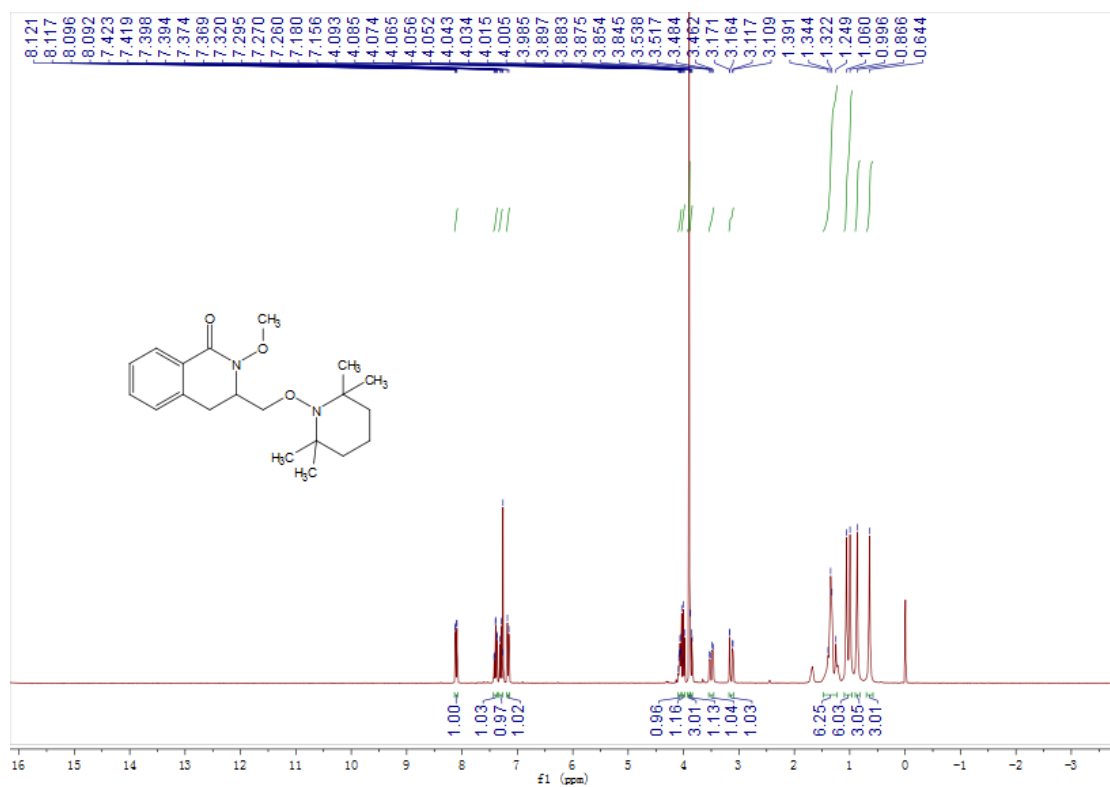












**<sup>1</sup>H NMR, <sup>13</sup>C NMR spectra of the purified Product in D<sub>2</sub>O Experiment:**

