

## Supporting information

### **Na<sub>2</sub>S promoted reduction of azides in water: Synthesis of pyrazolopyridines in one pot and evaluation of antimicrobial activity**

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## **Biological evaluation**

### ***Antimicrobial activity***

The antimicrobial activity of the derivatives was tested based on well diffusion method<sup>1</sup> against different pathogenic reference strains procured from the Microbial Type Culture Collection (MTCC), CSIR-Institute of Microbial Technology, Chandigarh, India. The pathogenic reference strains were seeded with 0.1 ml of previously prepared microbial suspensions individually containing  $1.5 \times 10^8$  cfu ml<sup>-1</sup> (equal to 0.5 McFarland) on the surface of Muller-Hinton agar Petri plates. Wells of 6.0 mm diameter were prepared in the media plates using a cork borer and the synthesized derivatives were added at a dose range of 150 - 1.17  $\mu\text{g well}^{-1}$  in each well under sterile conditions in a laminar air flow chamber. Standard antibiotic solutions of ciprofloxacin and miconazole at a dose range of 150 - 0.58  $\mu\text{g well}^{-1}$  and the well containing methanol served as positive and negative controls, respectively. The plates were incubated for 24 h at 37 °C for bacterial strains and 30 °C for *Candida albicans*. The well containing the least concentration showing the inhibition zone was considered as the minimum inhibitory concentration. All experiments were carried out in duplicates and mean values are represented.

### ***Minimum bactericidal concentration (MBC) assay***

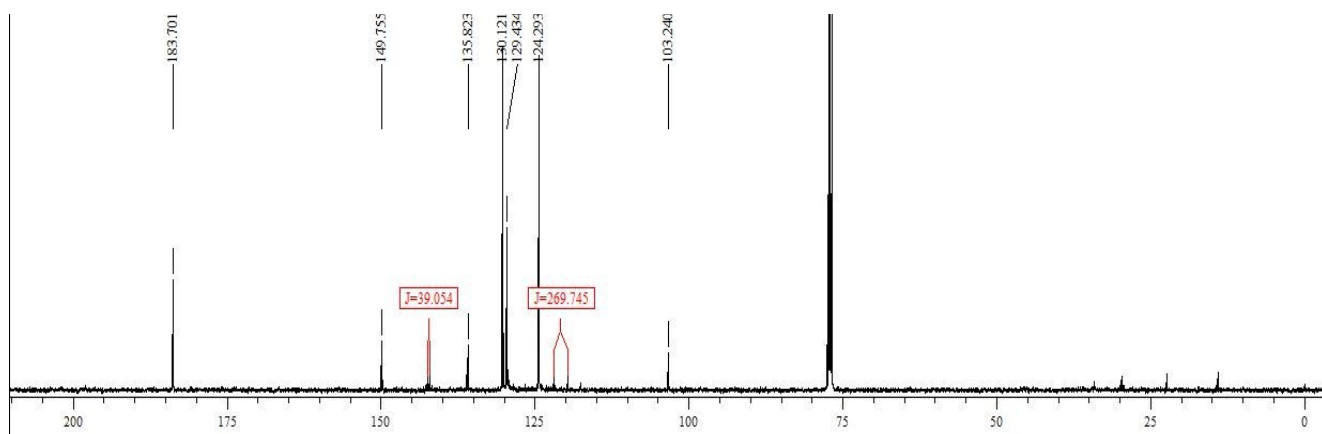
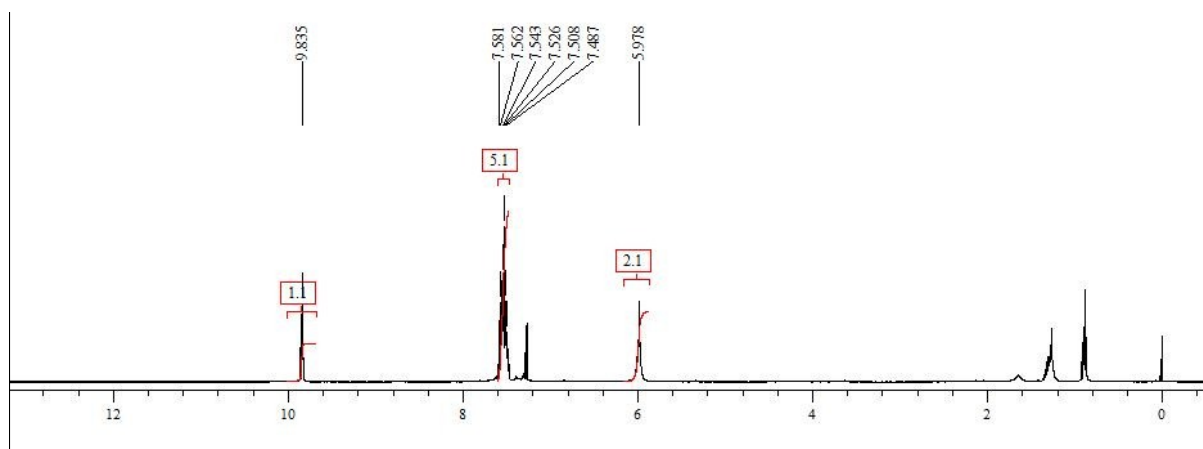
Bactericidal assay<sup>2</sup> (NCCLS, 2000) was performed in sterile 2.0 ml microfuge tubes against a panel of pathogenic bacterial strains, including *Micrococcus luteus* MTCC 2470, *Bacillus subtilis* MTCC 121, *Escherichia coli* MTCC 739, and *Klebsiella planticola* MTCC 530 which were cultured overnight in Mueller Hinton broth. Serial dilutions of test compounds were prepared in Mueller Hinton broth with different concentrations ranging from 0 to 250  $\mu\text{g mL}^{-1}$ . To the test compounds, 100  $\mu\text{L}$  of overnight cultured bacterial suspensions were added to reach a final concentration of  $1.5 \times 10^8$  cfu mL<sup>-1</sup> (equal to 0.5 McFarland) and incubated at 37 °C for 24 h. After 24 h of incubation, the minimum bactericidal concentration (MBC) was determined by sampling 10  $\mu\text{L}$  of suspension from the tubes onto Mueller Hinton agar plates and were incubated for 24 h at 37 °C to observe the growth of test organisms. MBC is the lowest concentration of compound required to kill a particular bacterium. All the experiments were carried in duplicates.

National Committee for Clinical Laboratory Standards, NCCLS (2000) Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard fifth edition, NCCLS: Wayne, PA.

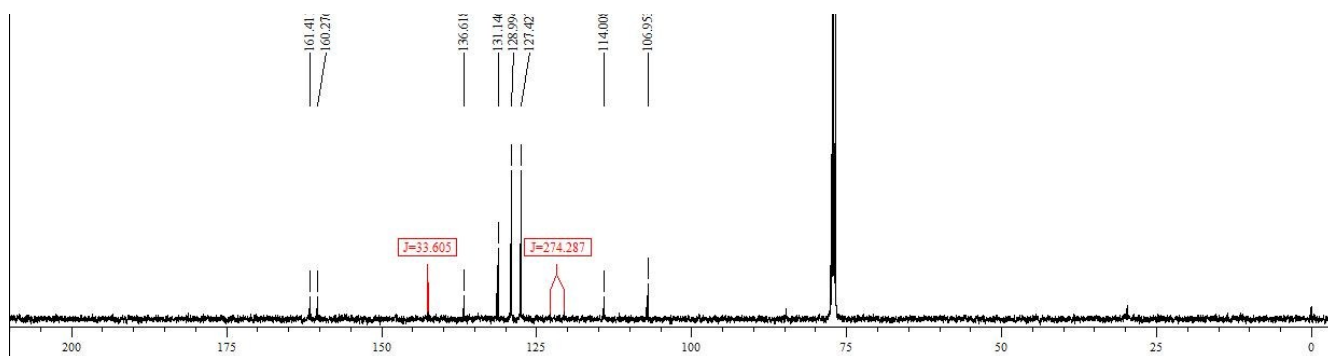
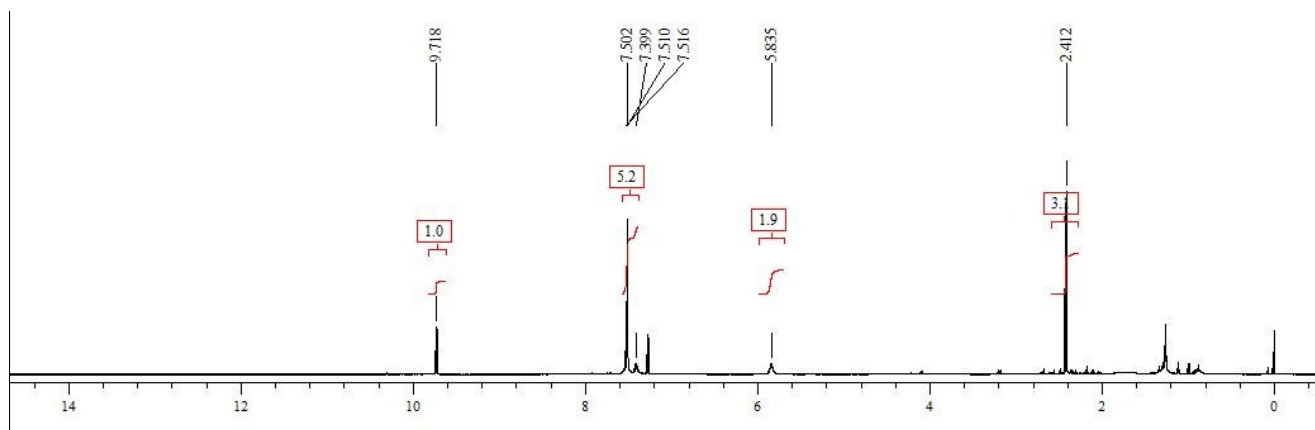
### **References:**

1. Amsterdam, D. Susceptibility testing of antimicrobials in liquid media. In: Loman, V. (Ed.) Antibiotics in Laboratory Medicine, 4<sup>th</sup> Edition, Williams and Wilkins, Baltimore, MD **1996**, pp. 52.
2. National Committee for Clinical Laboratory Standards, NCCLS Methods for dilution Antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard fifth edition, NCCLS: Wayne, PA, **2000**.

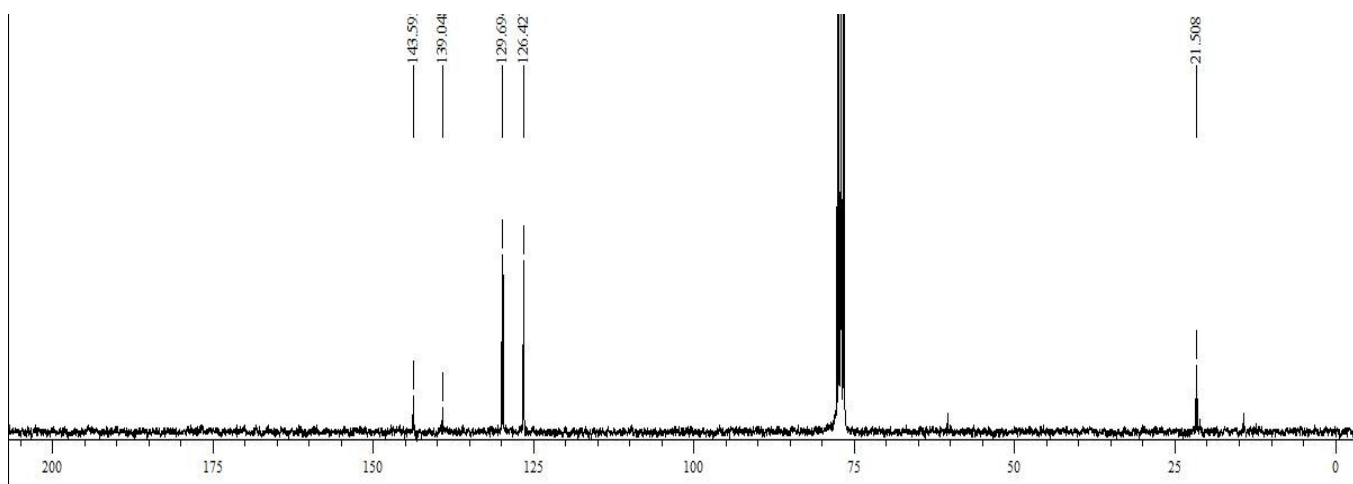
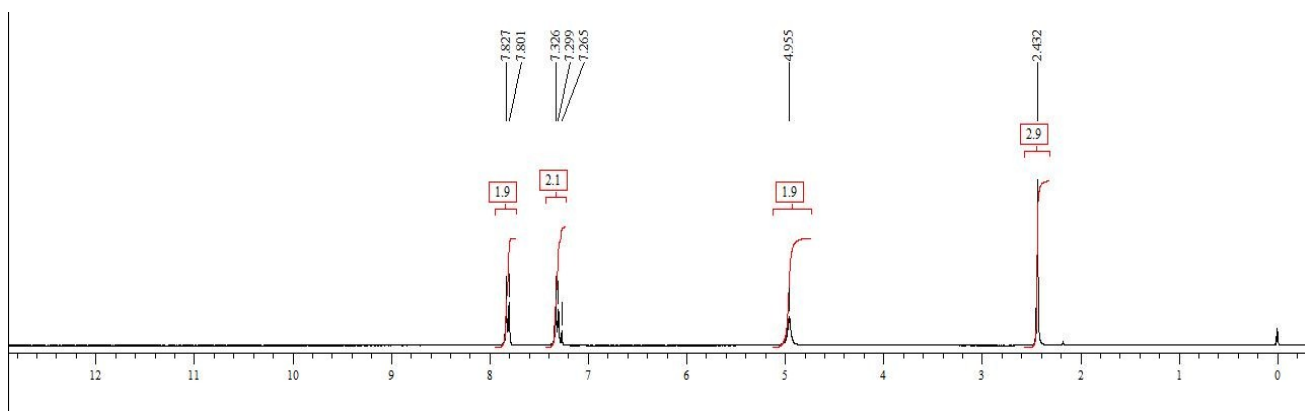
# $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR spectra of compound 2a



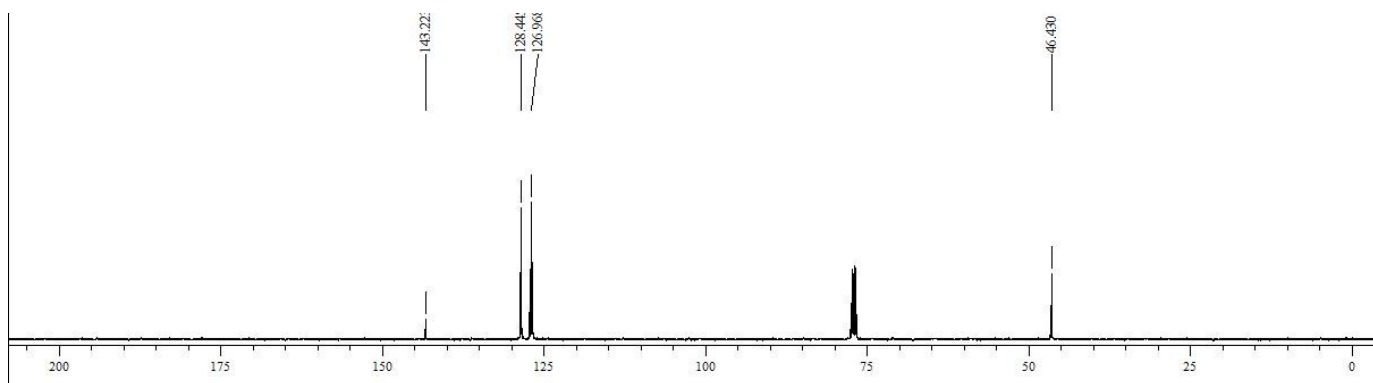
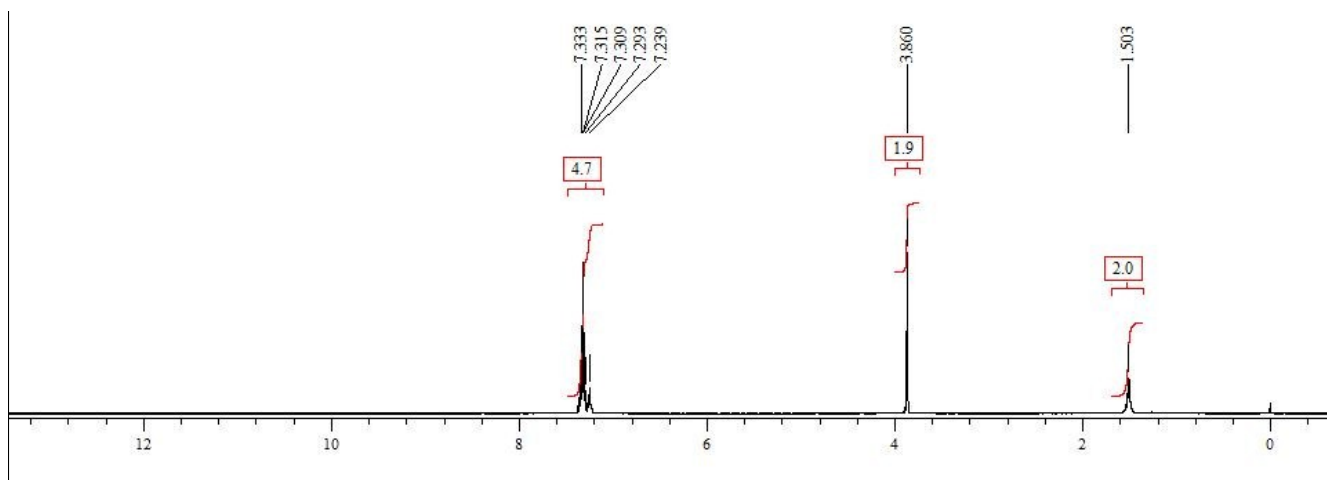
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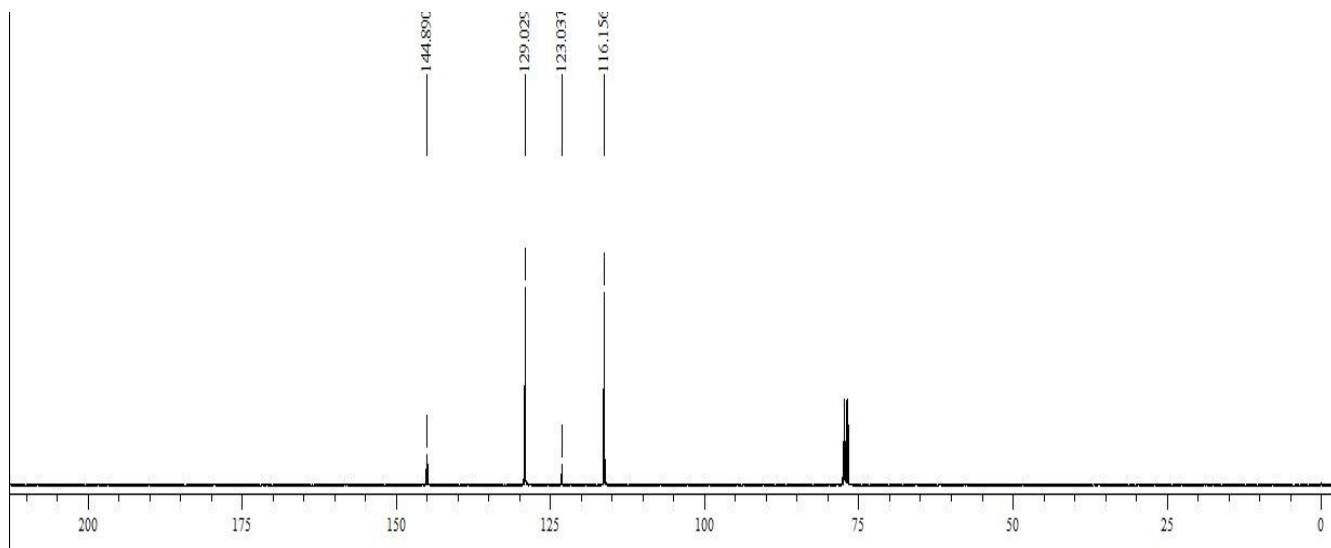
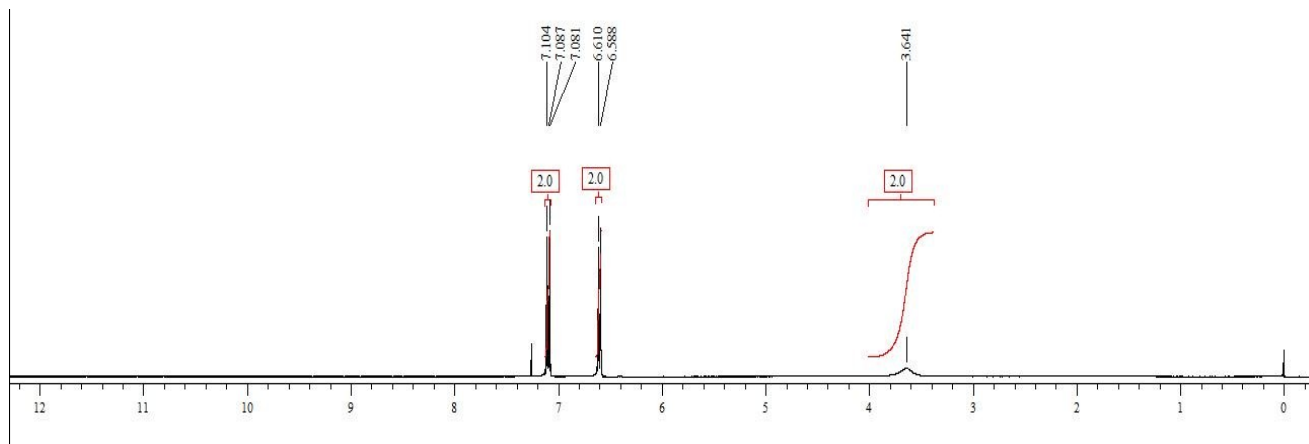
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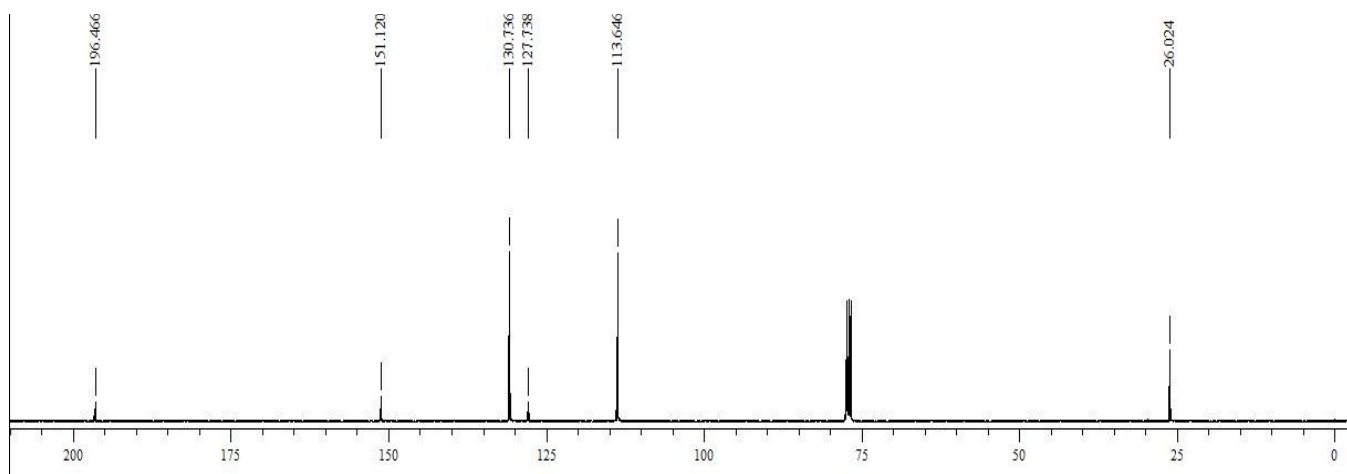
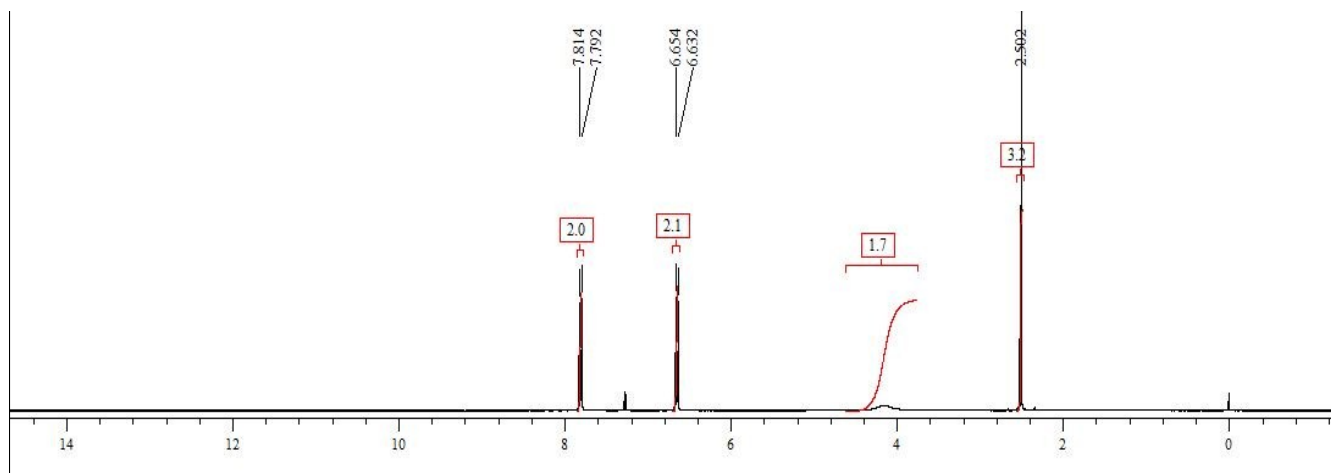
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# $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR spectra of compound 2e

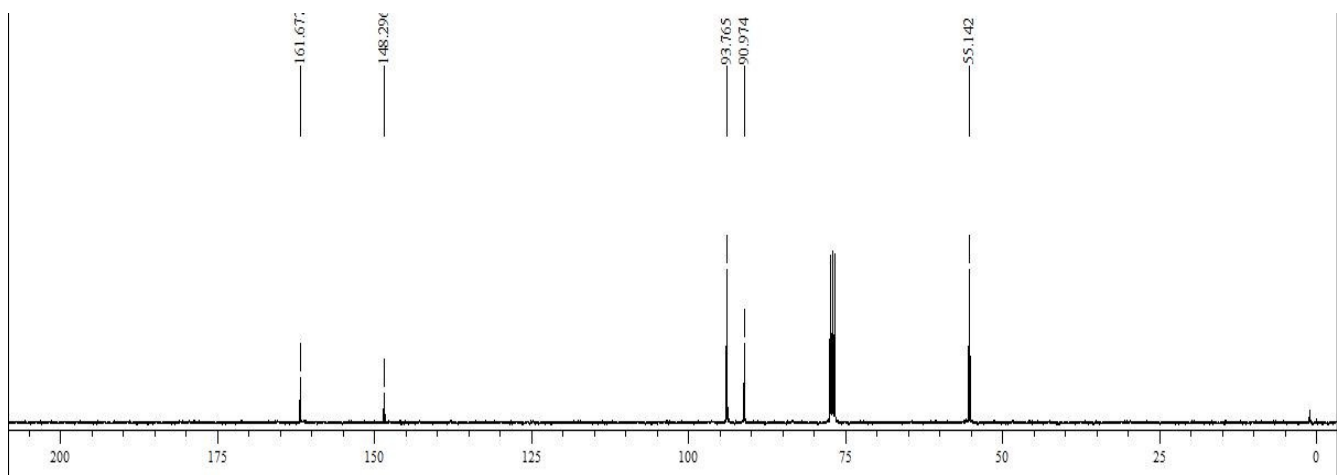
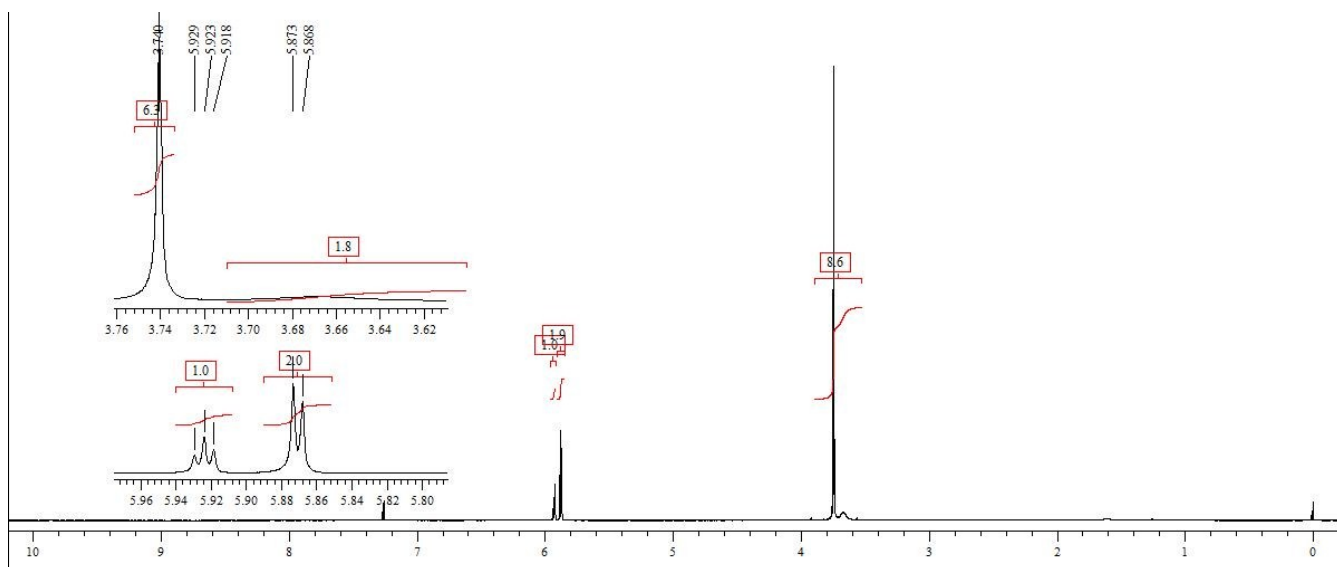


# $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR spectra of compound 2f

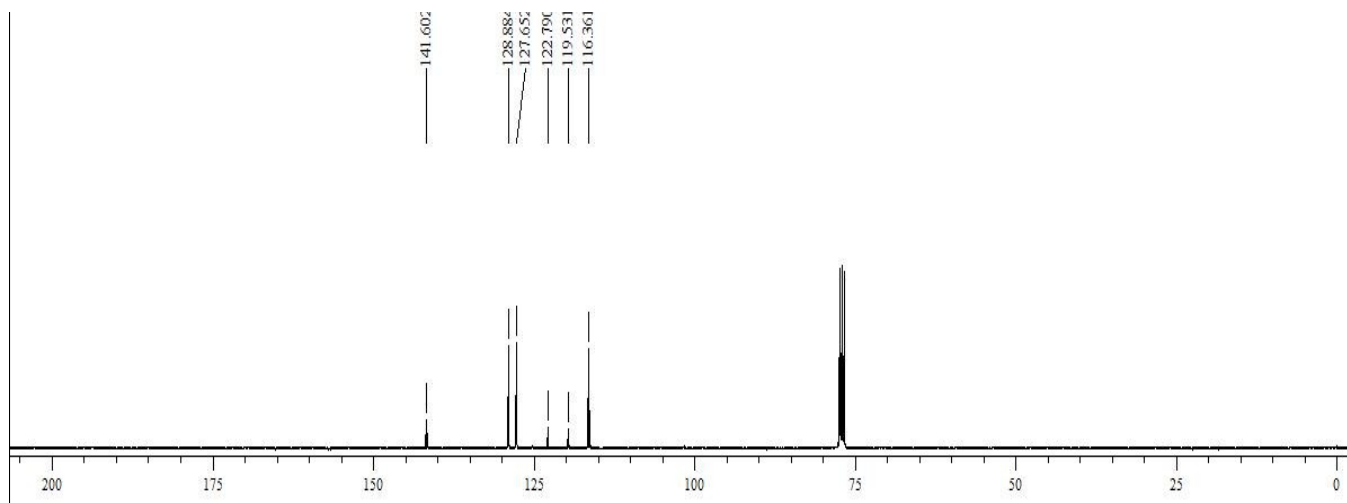
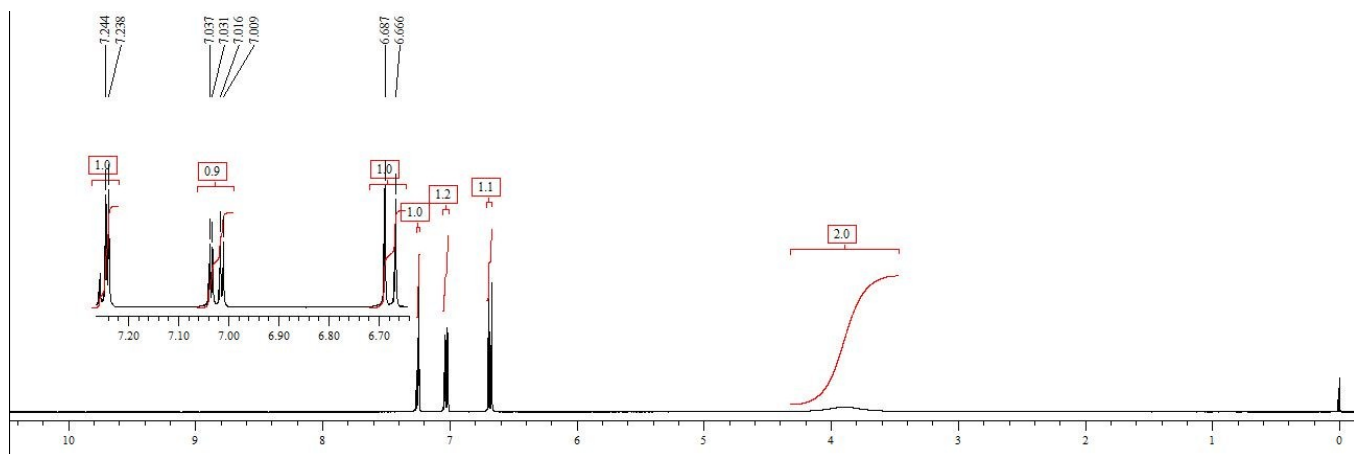




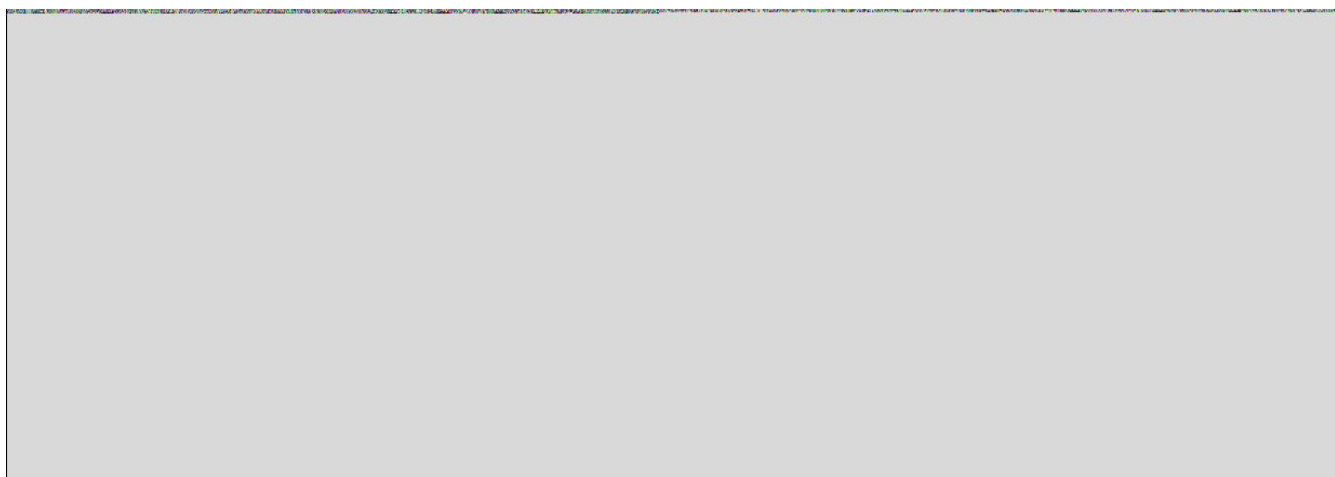
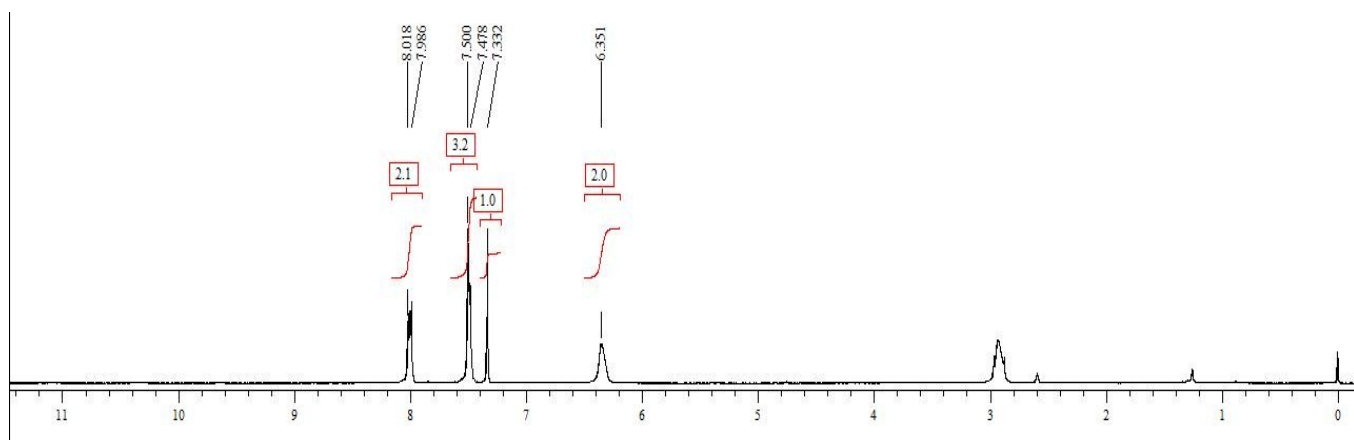
# <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra of compound 2g



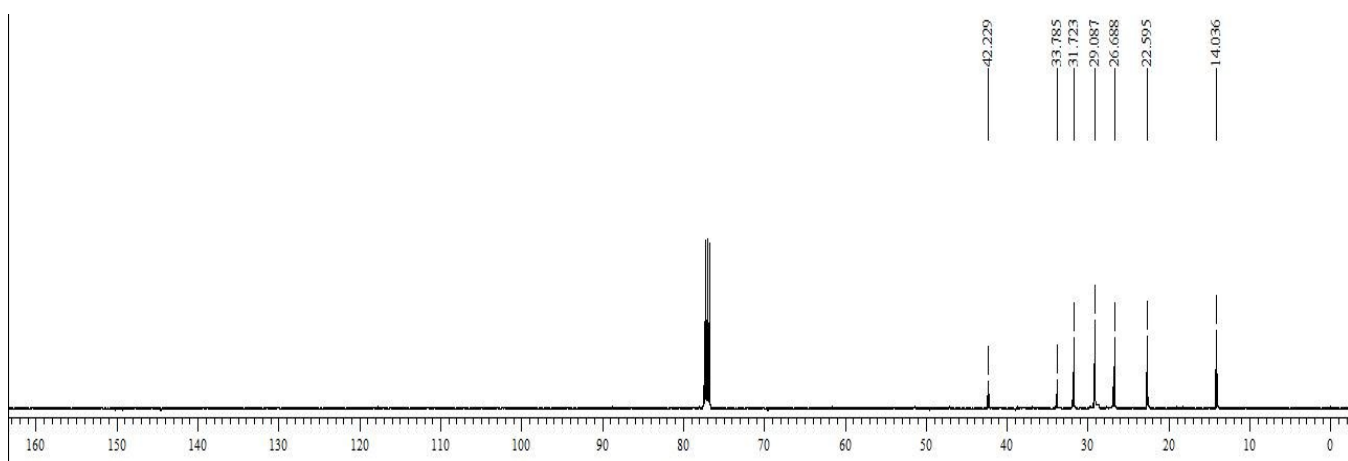
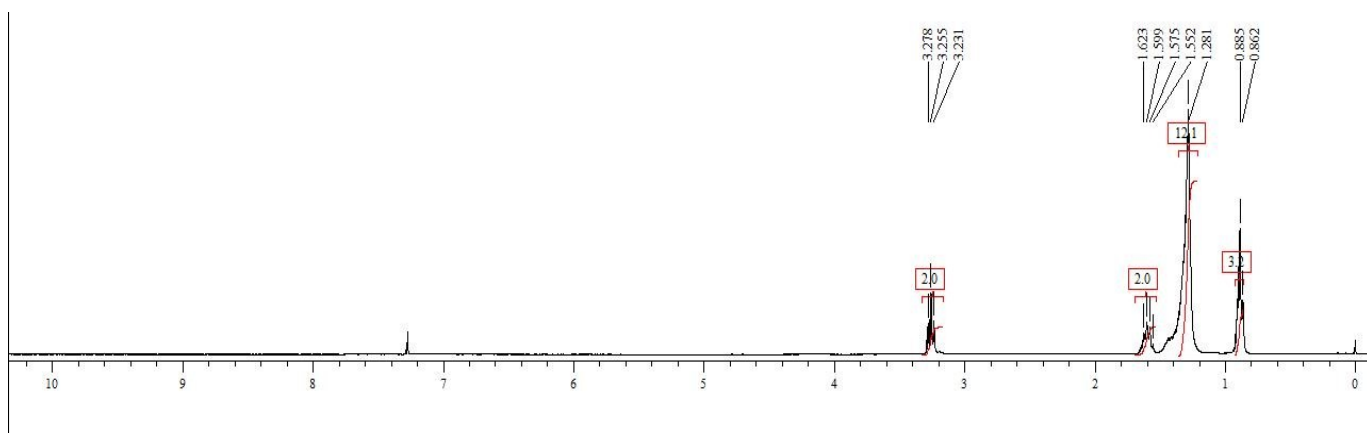
# $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectra of compound 2h



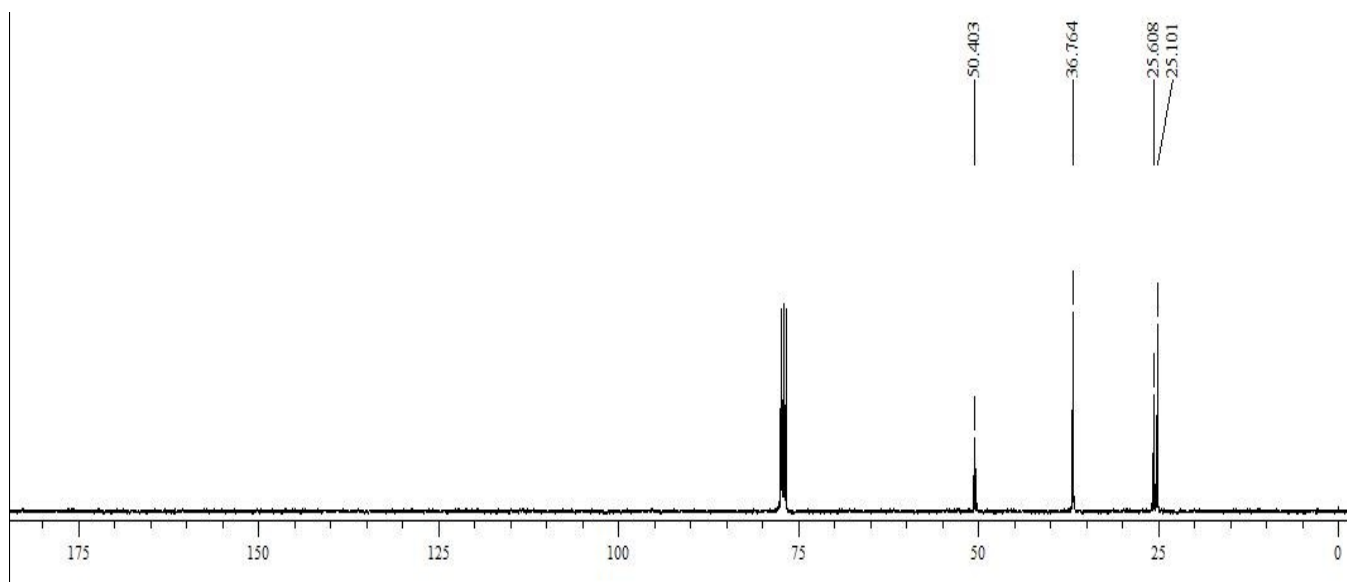
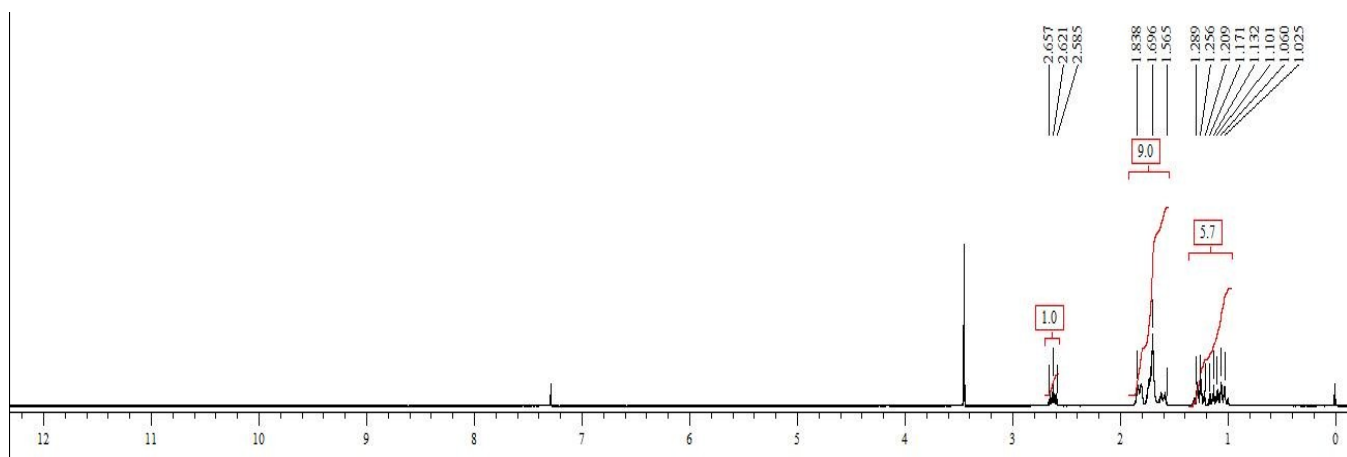
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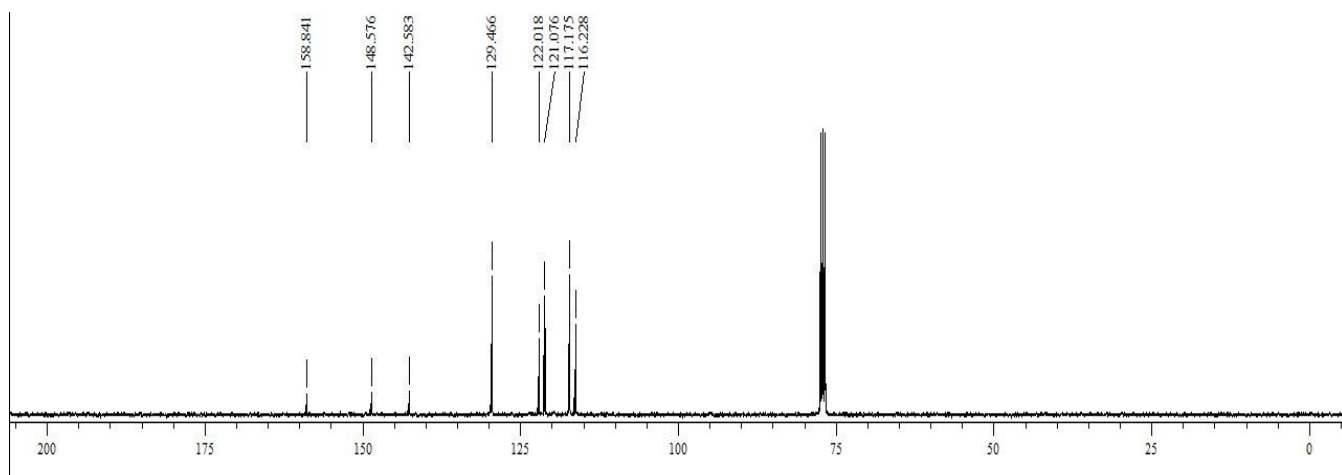
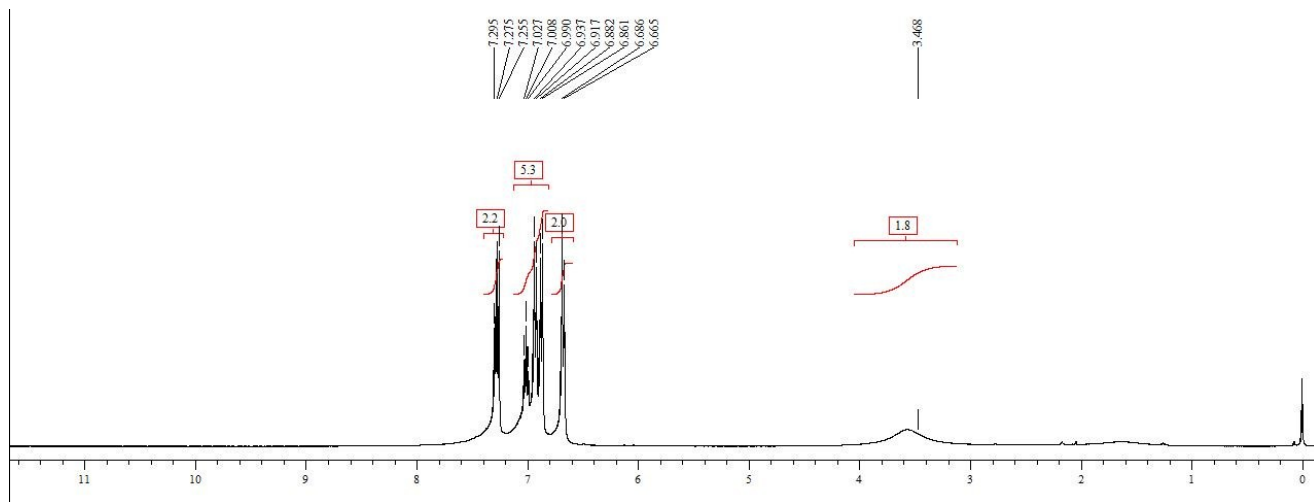
# $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR spectra of compound 2j



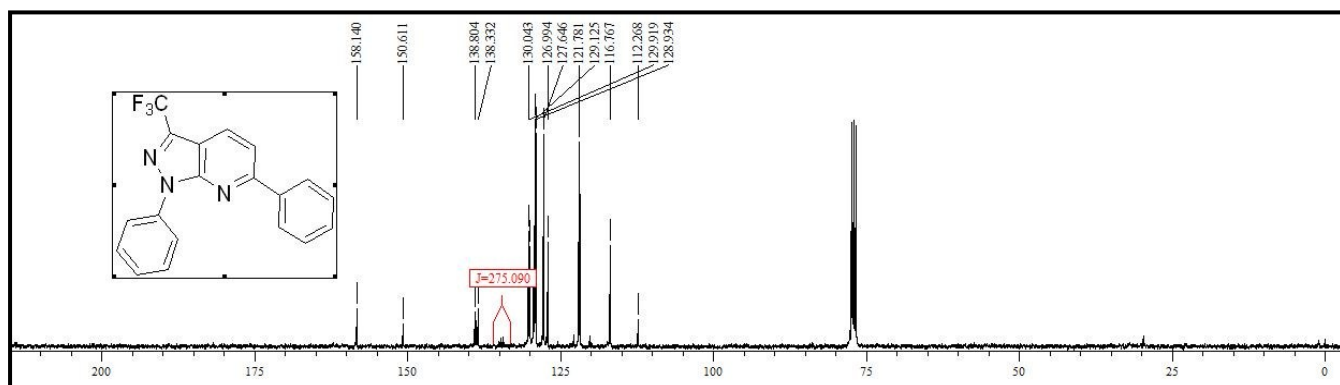
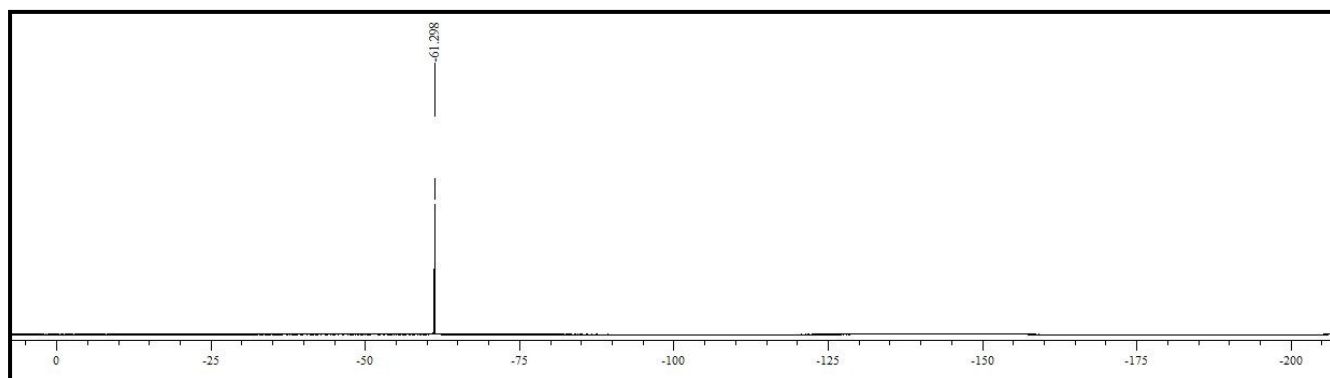
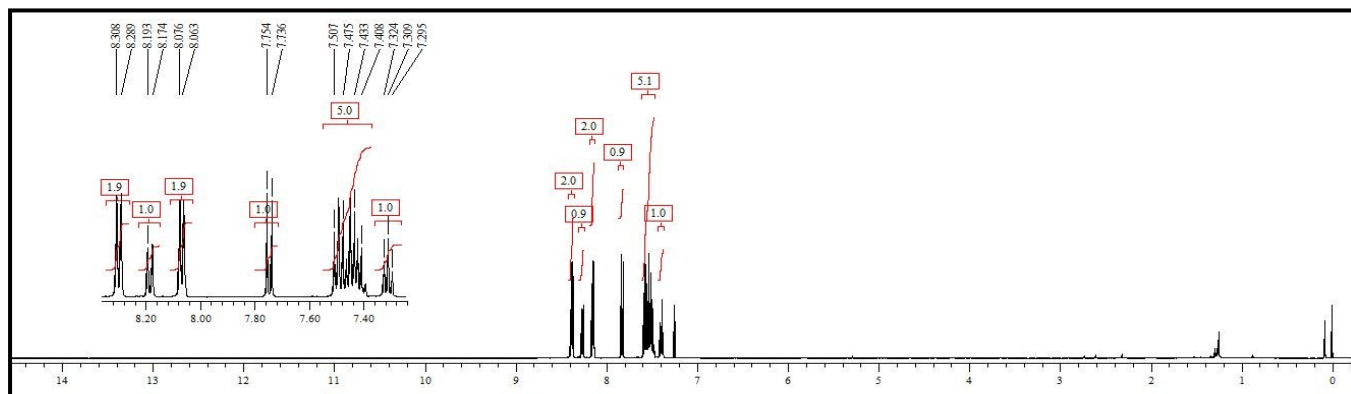
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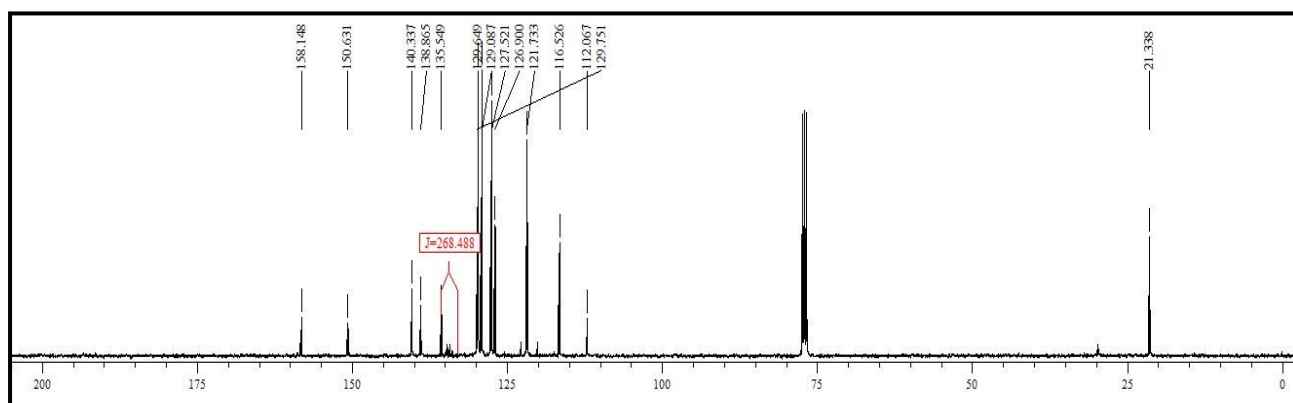
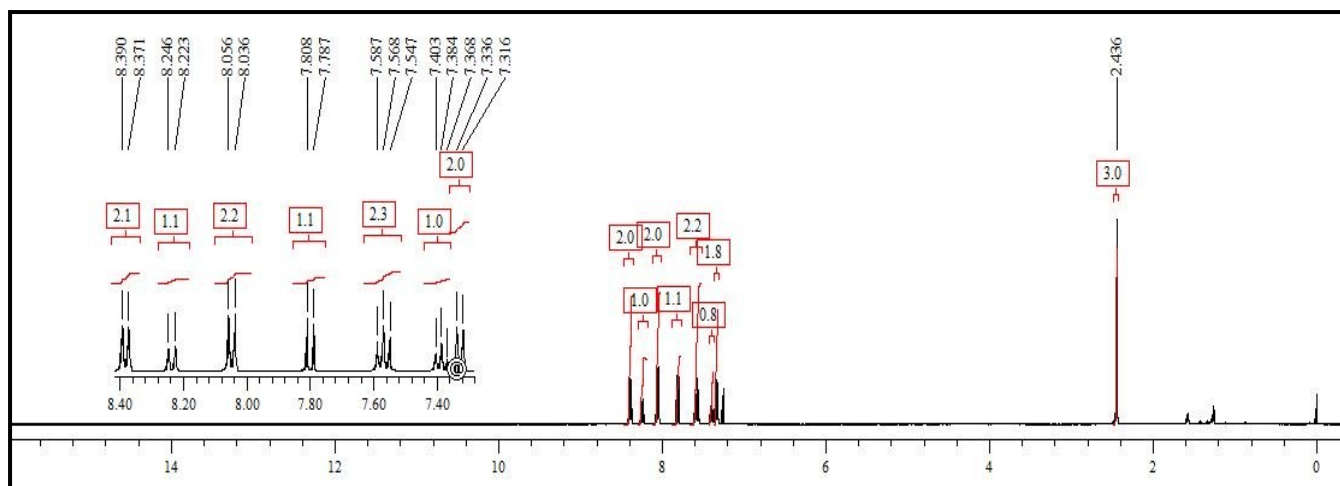
# $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR spectra of compound 2l



# <sup>1</sup>H-NMR, <sup>19</sup>F-NMR and <sup>13</sup>C-NMR spectra of compound 4a

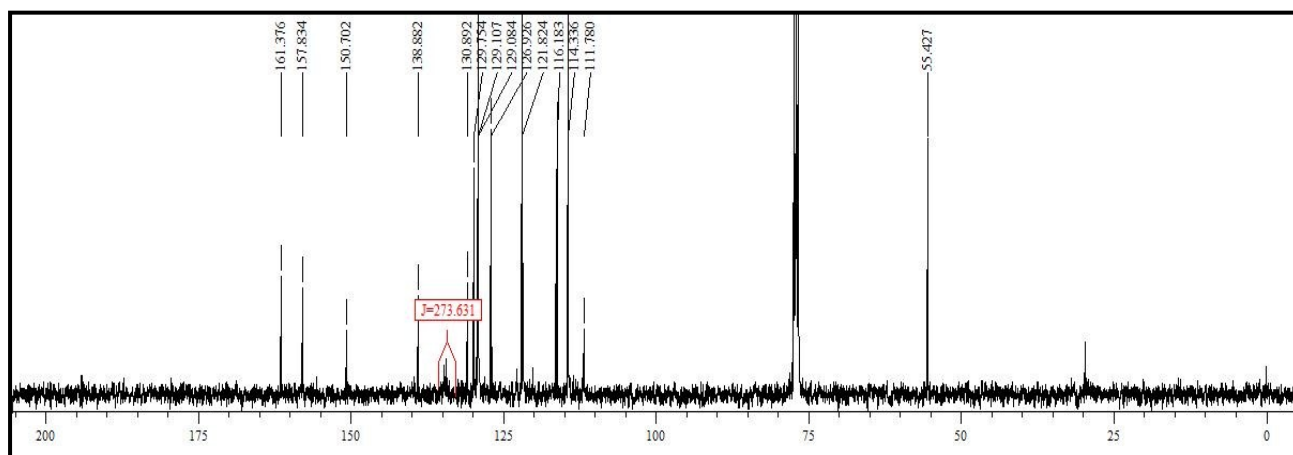
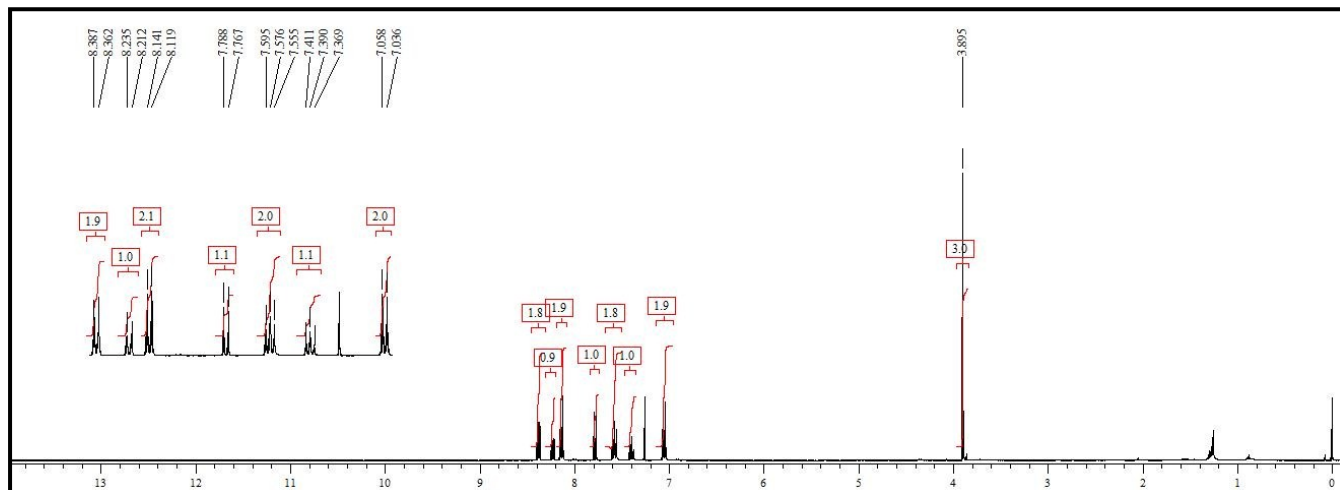


# <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra of compound 4b

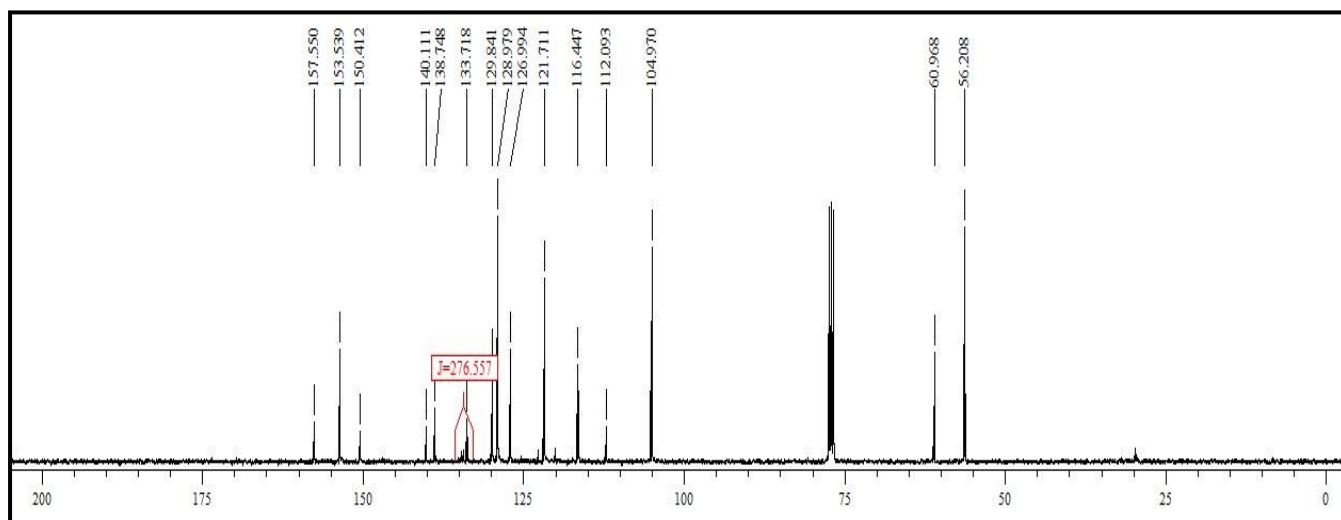
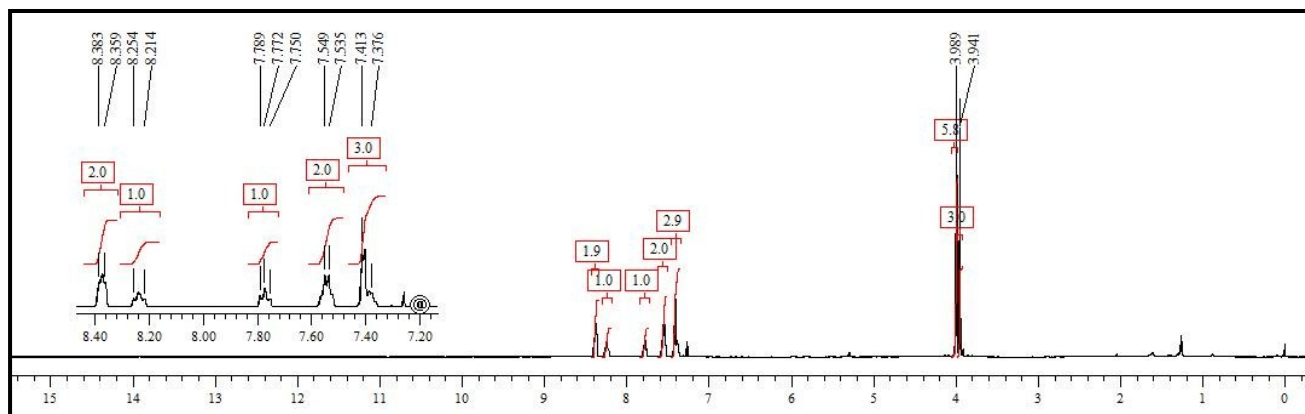




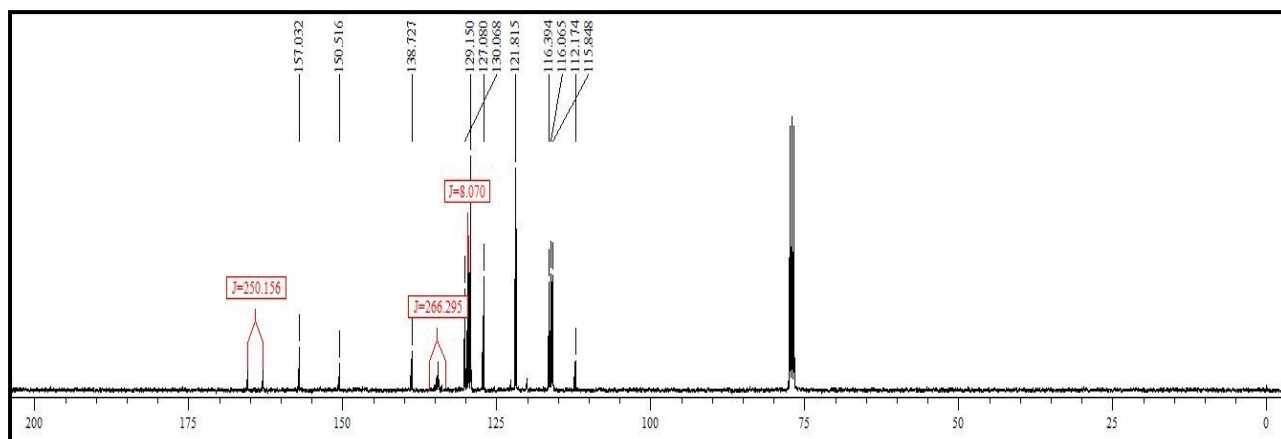
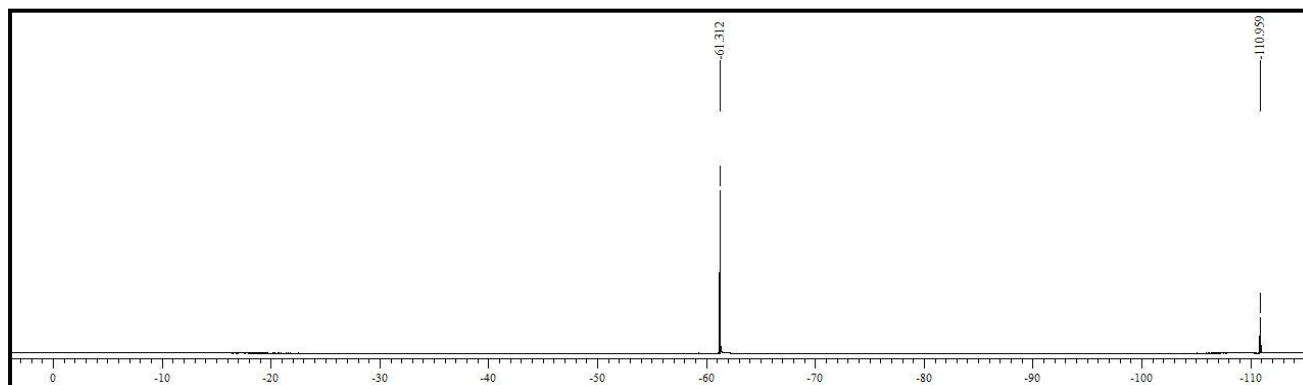
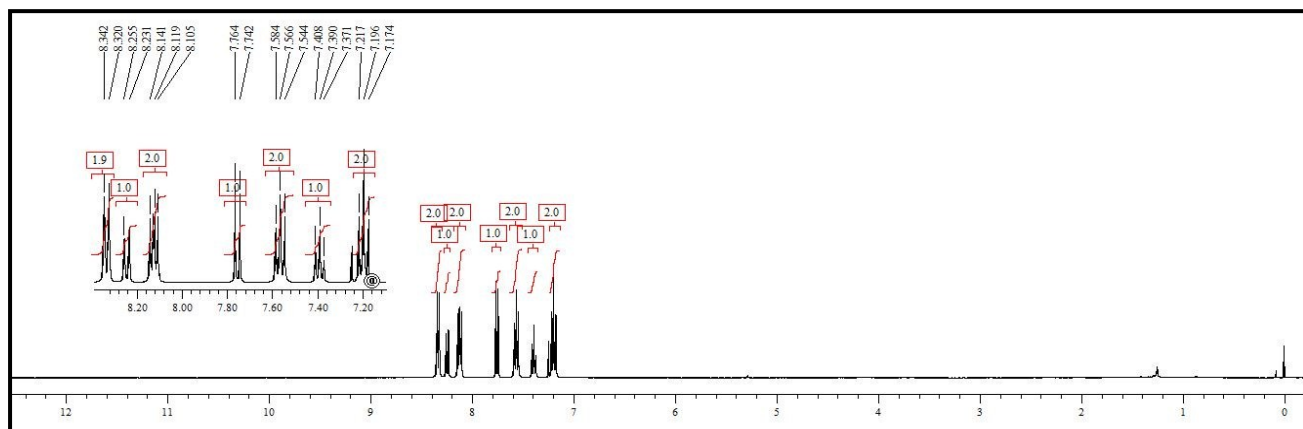
# <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra of compound 4c



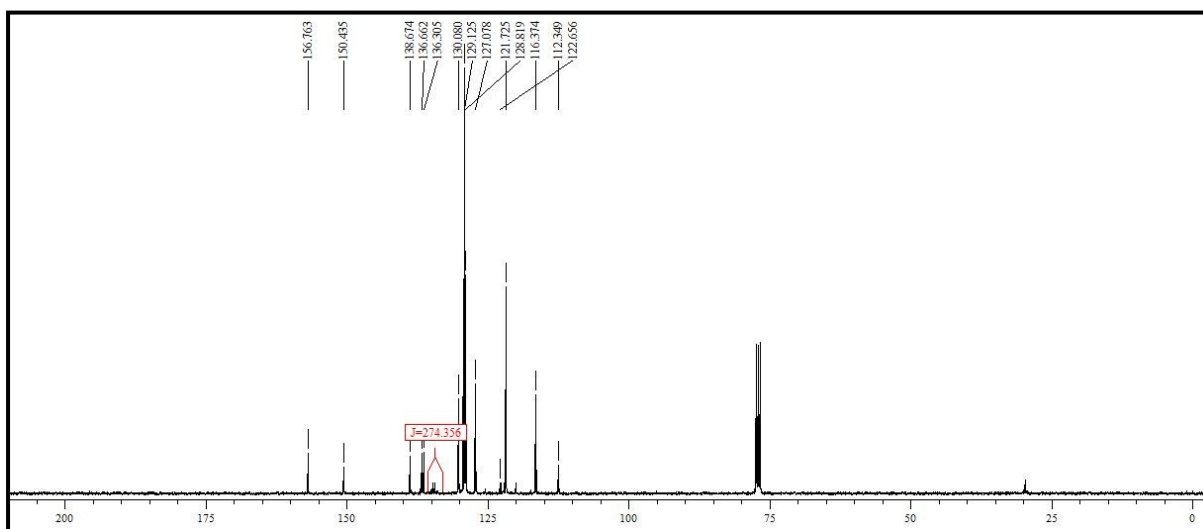
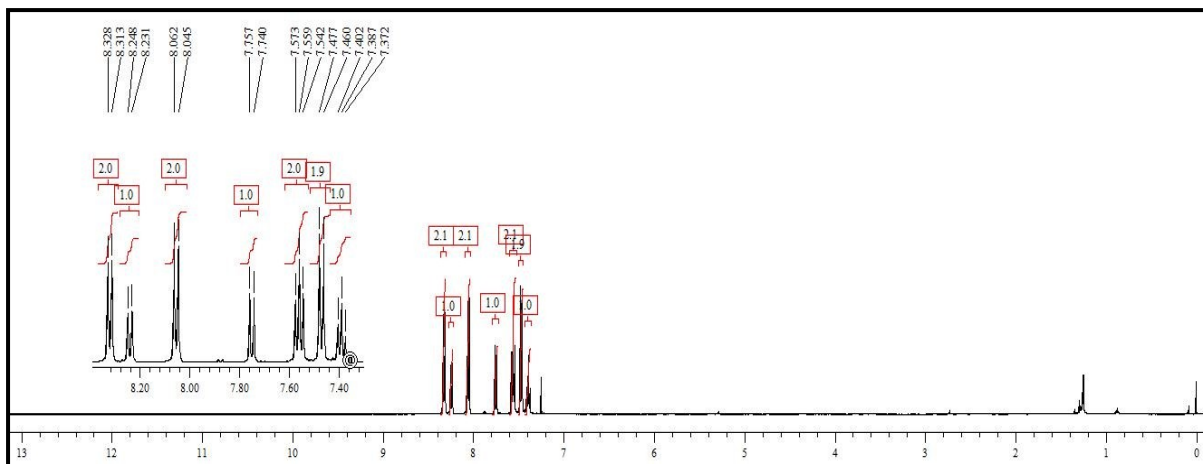
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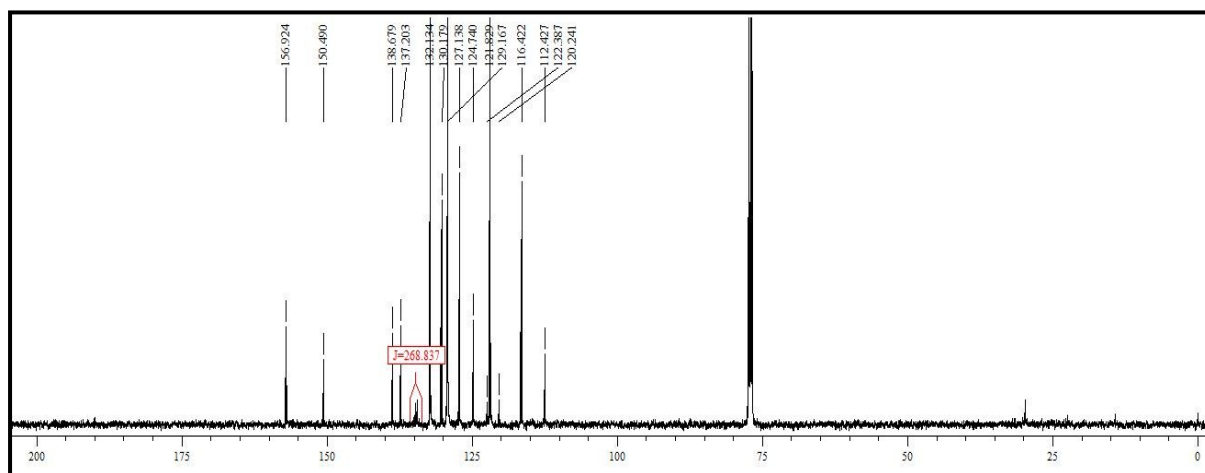
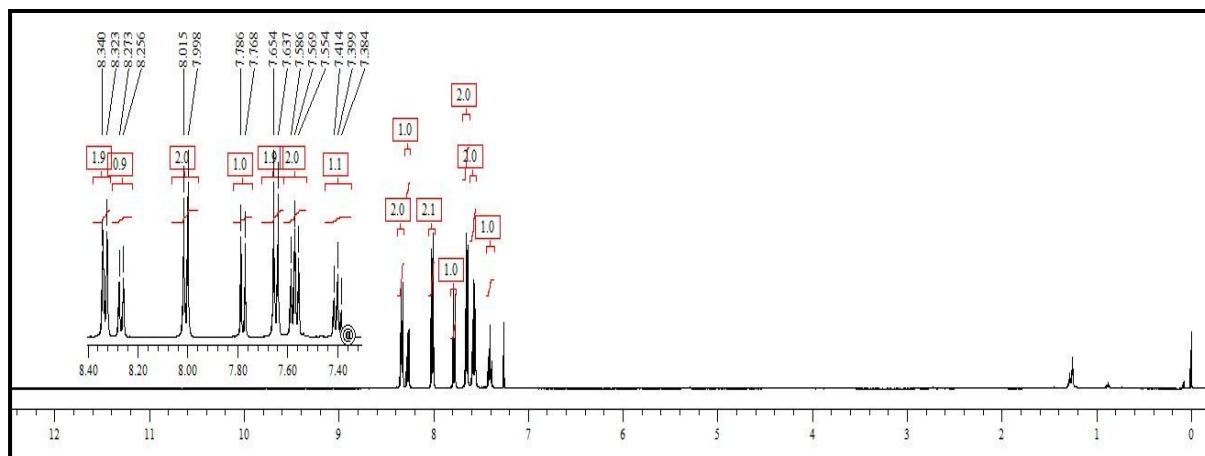
**<sup>1</sup>H-NMR, <sup>19</sup>F-NMR and <sup>13</sup>C-NMR spectra of compound 4e**



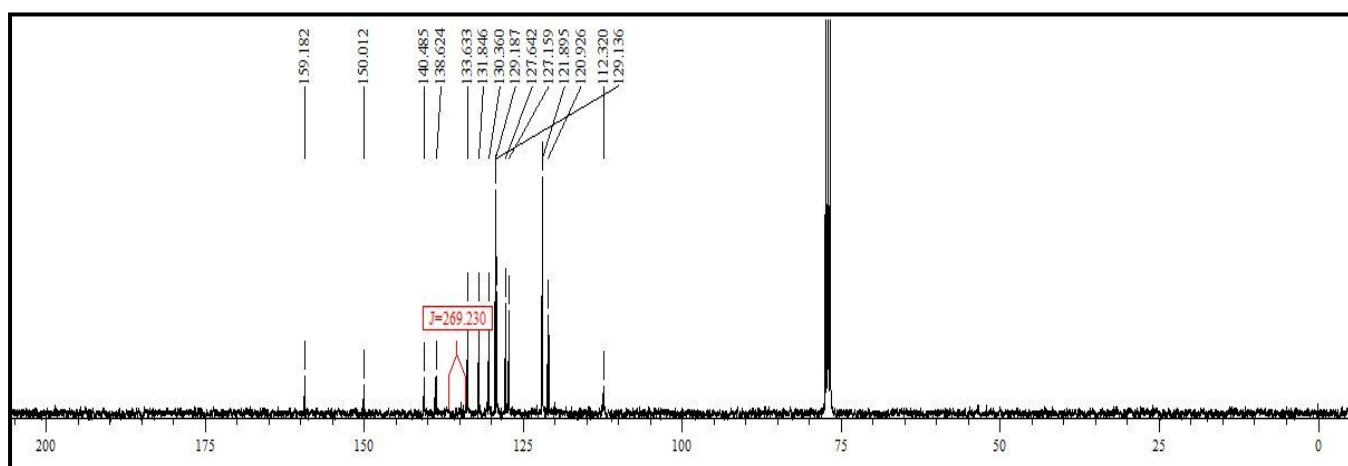
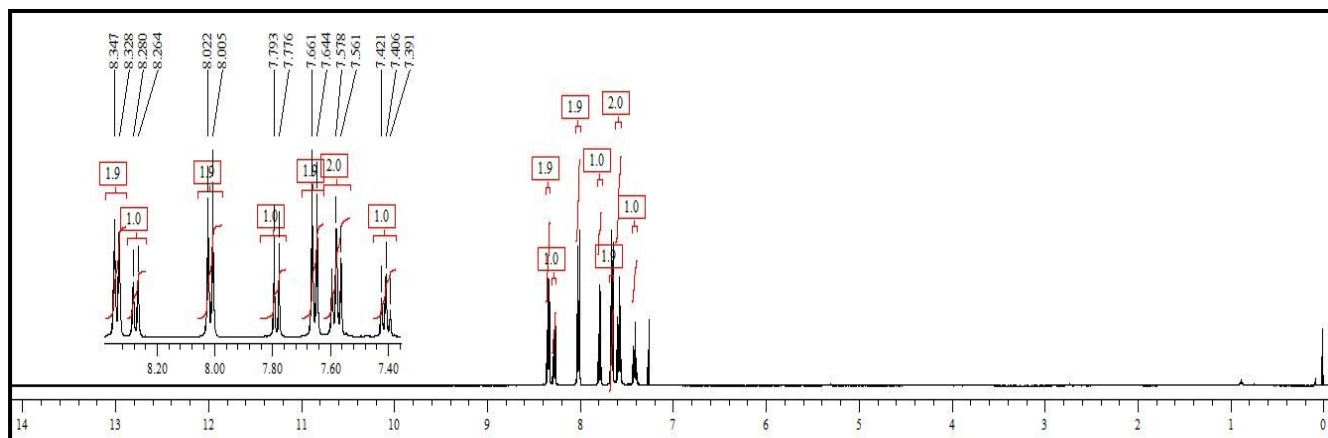
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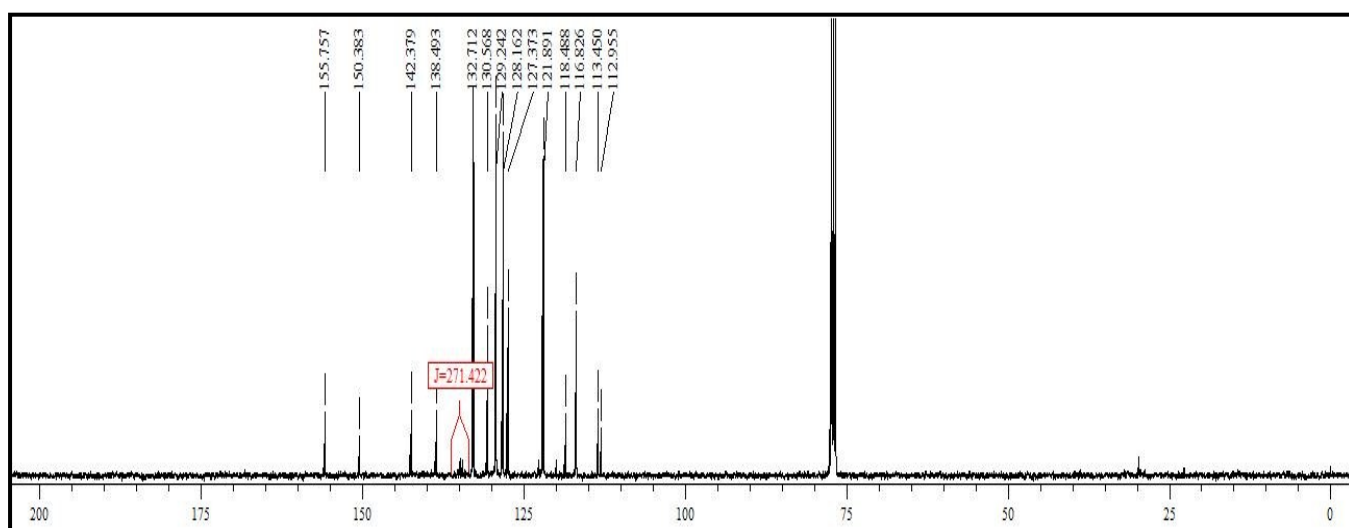
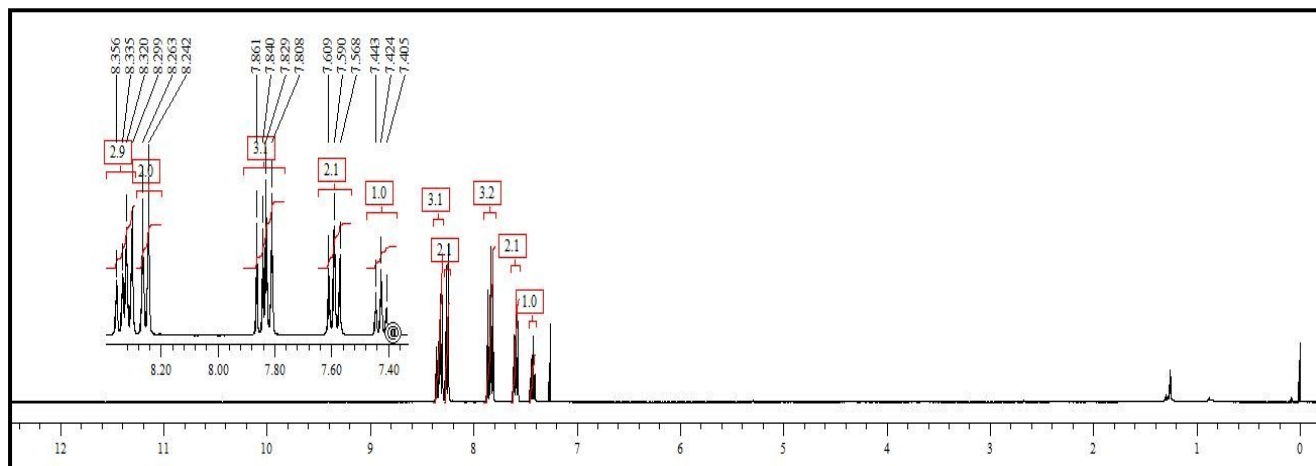
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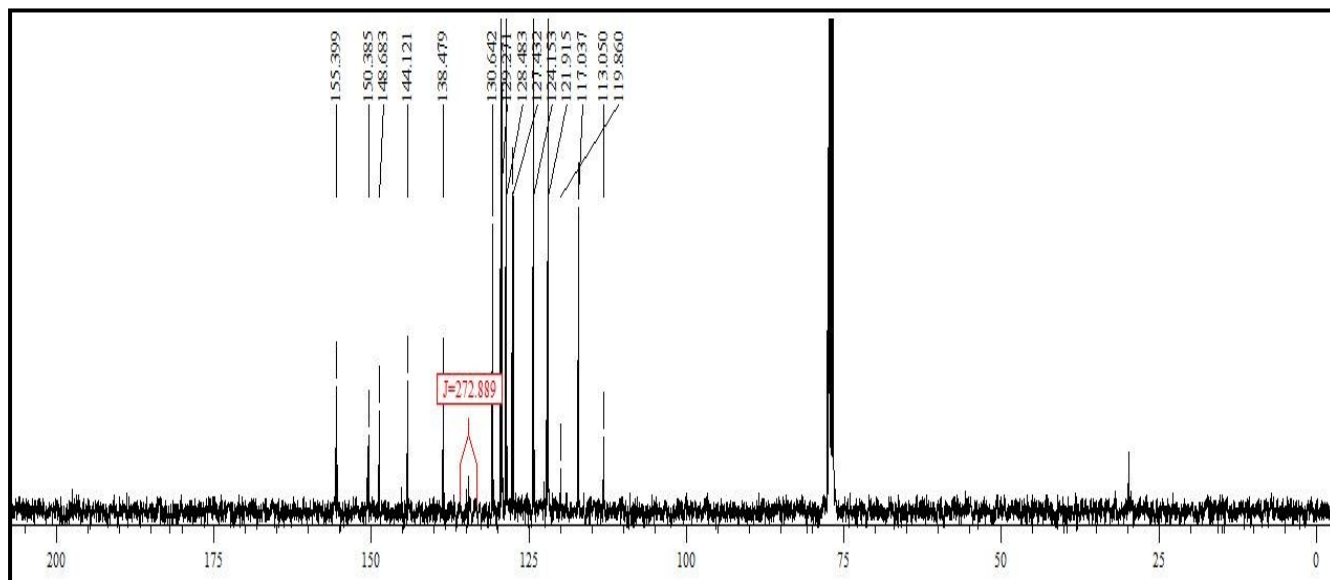
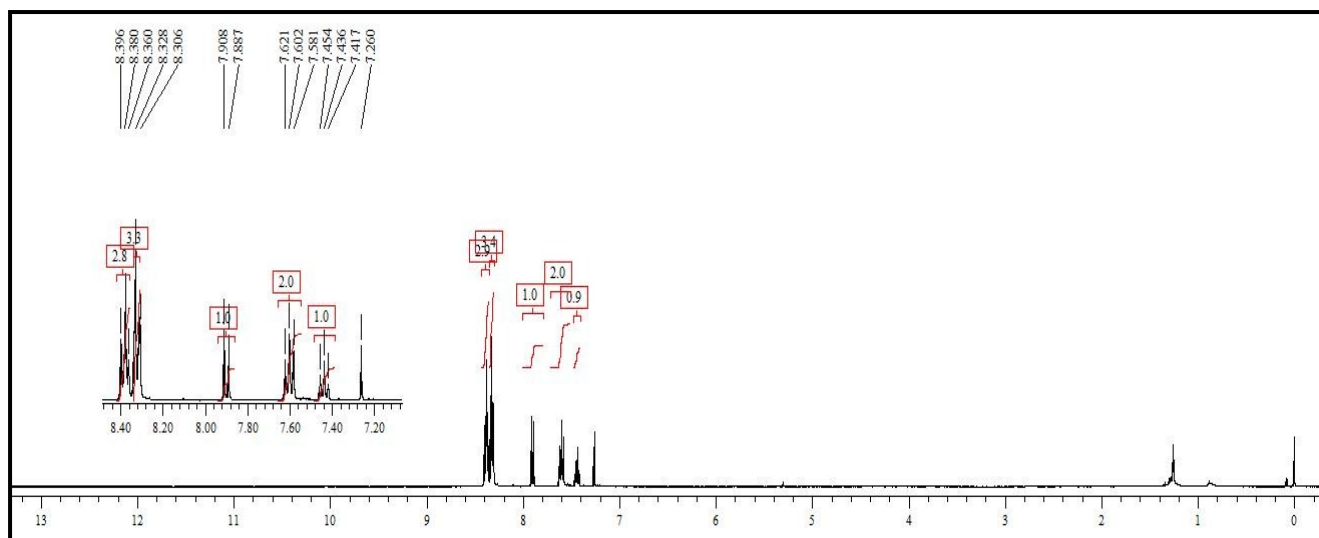
# <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra of compound 4h



# <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra of compound 4i

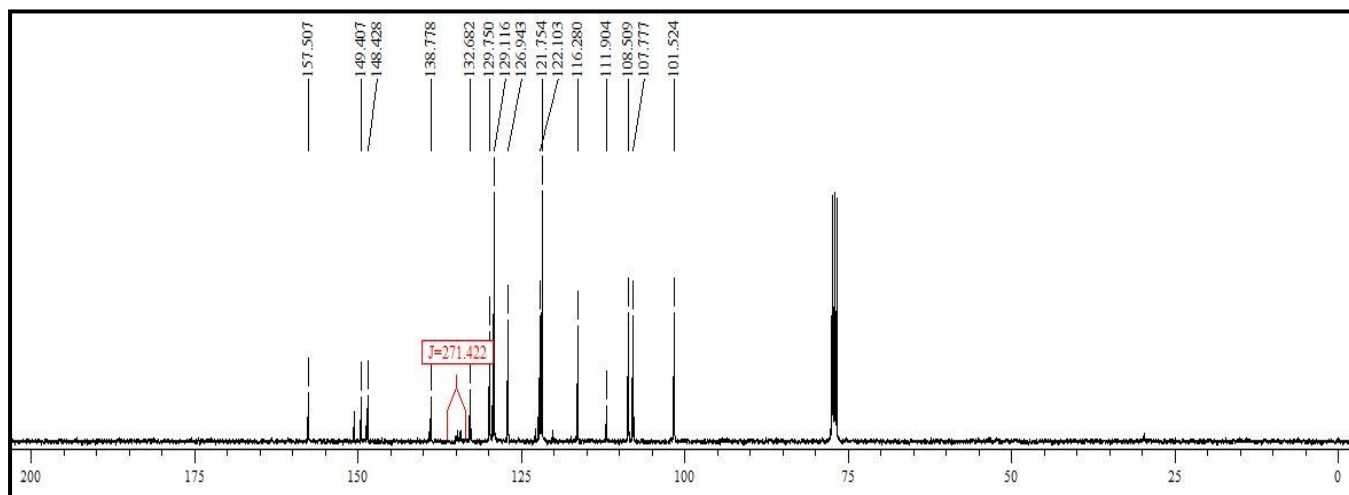
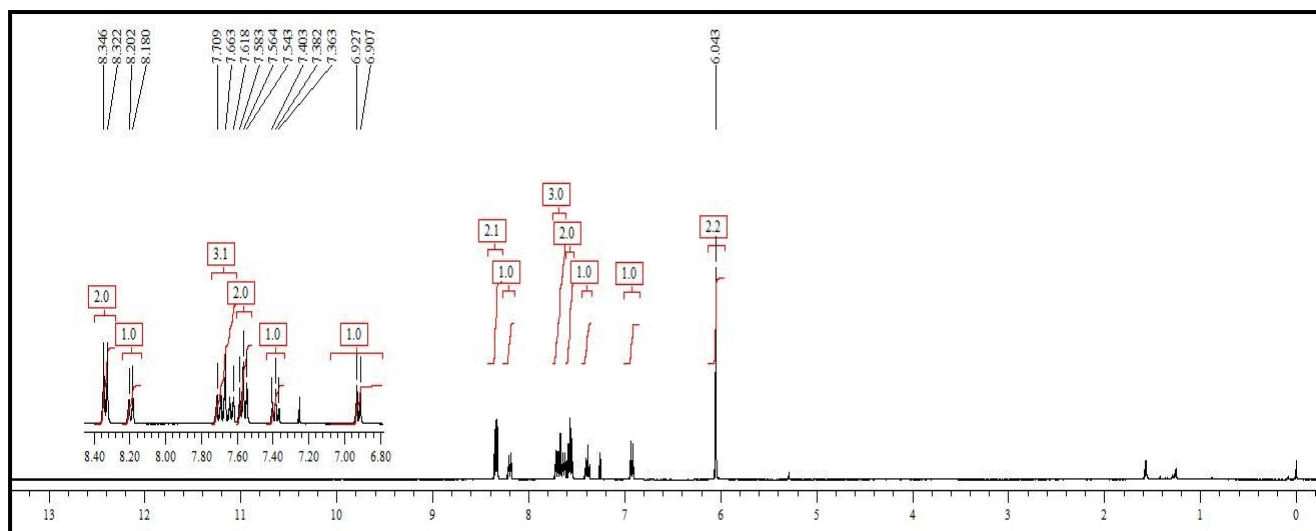


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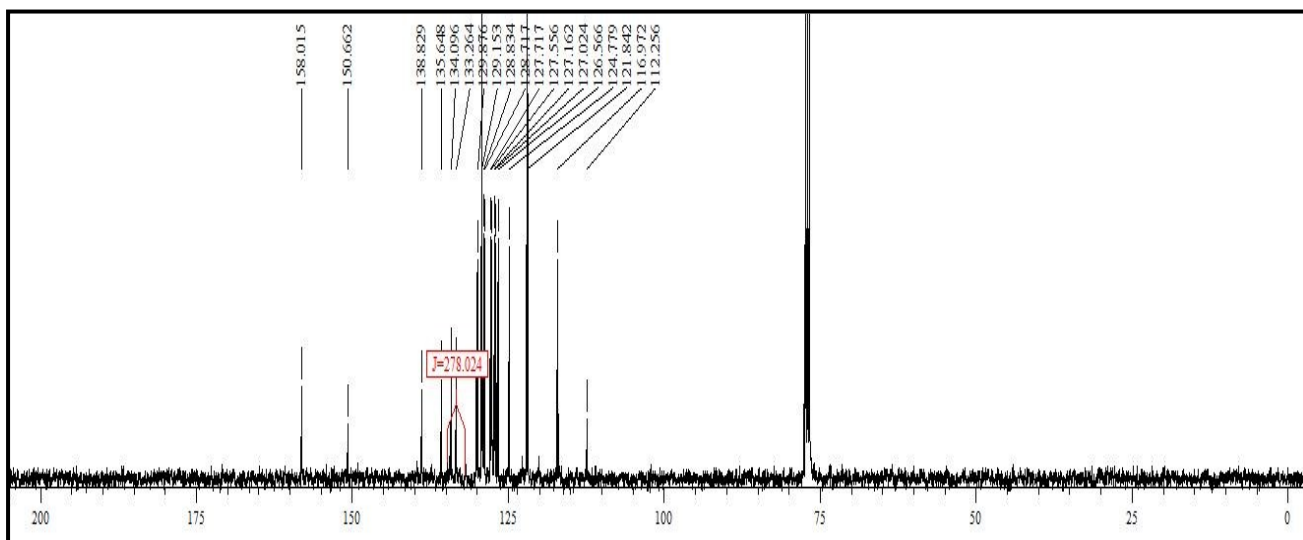
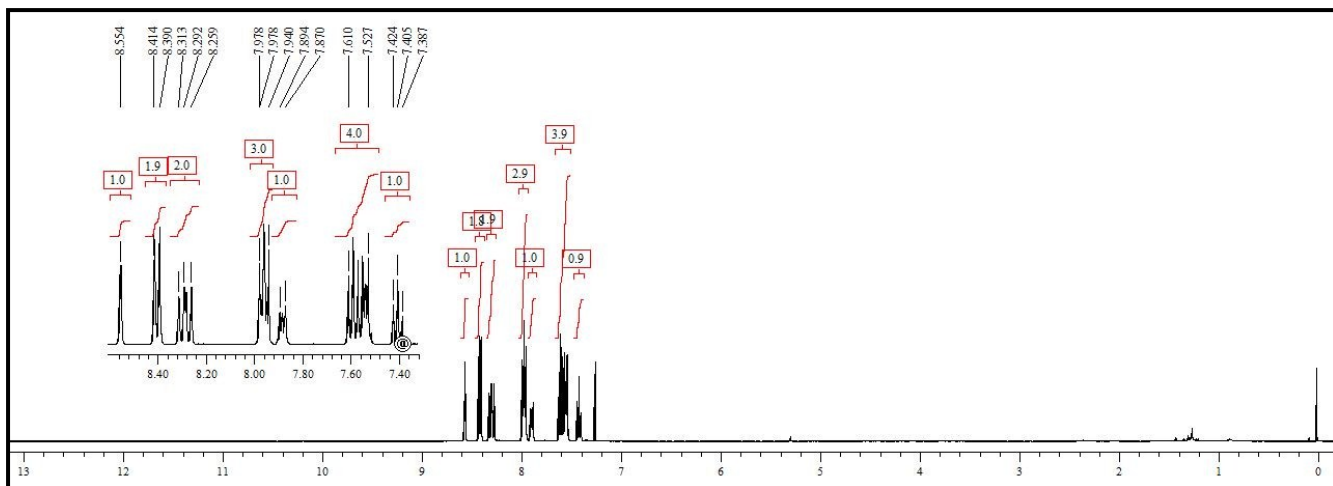




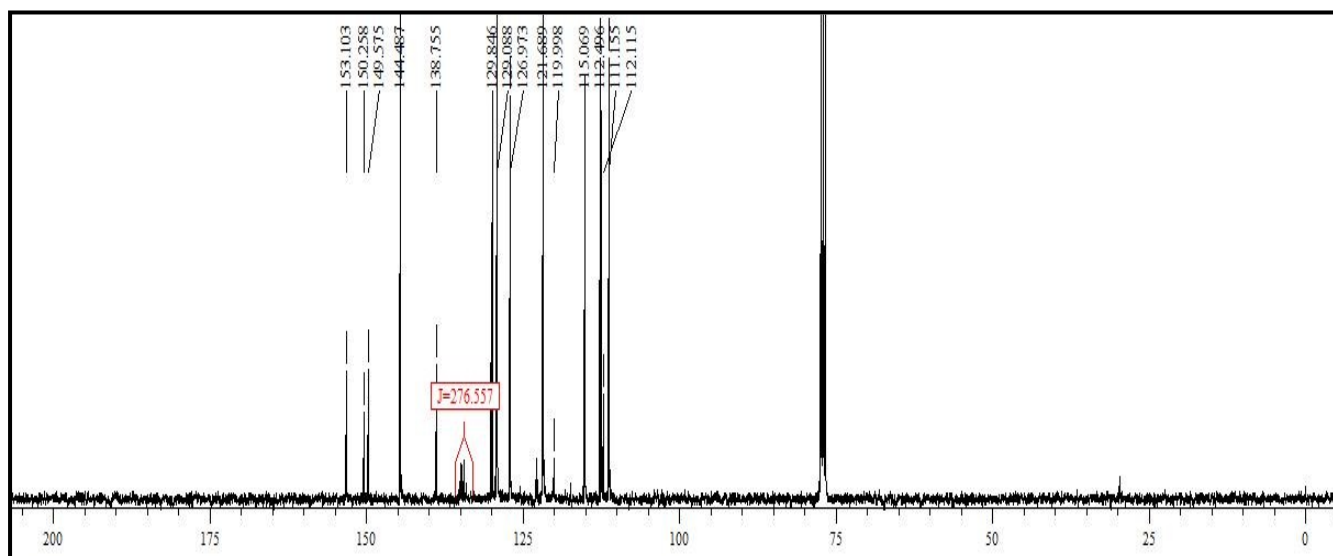
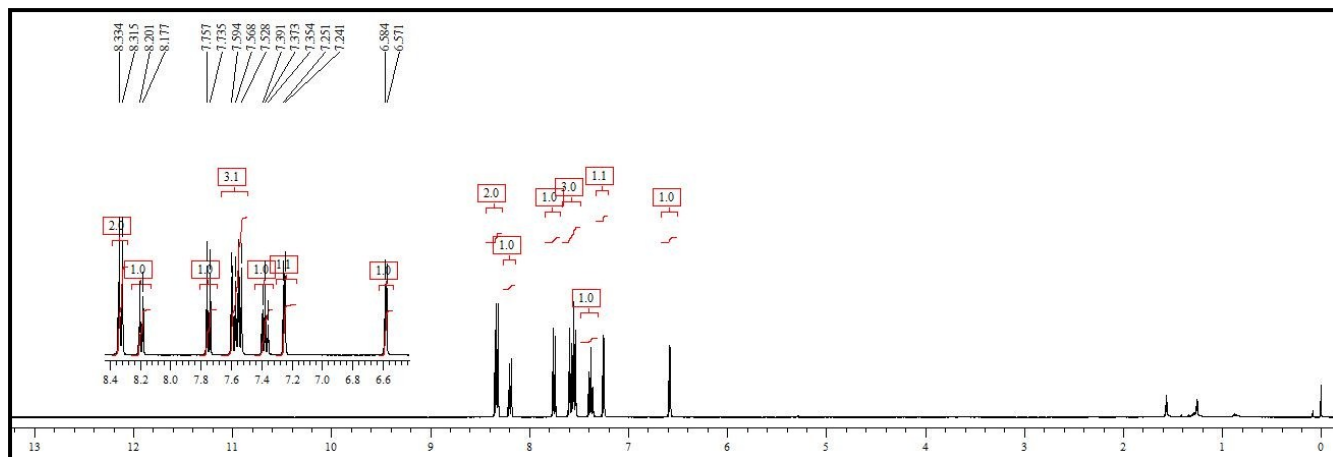
# <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra of compound 4k



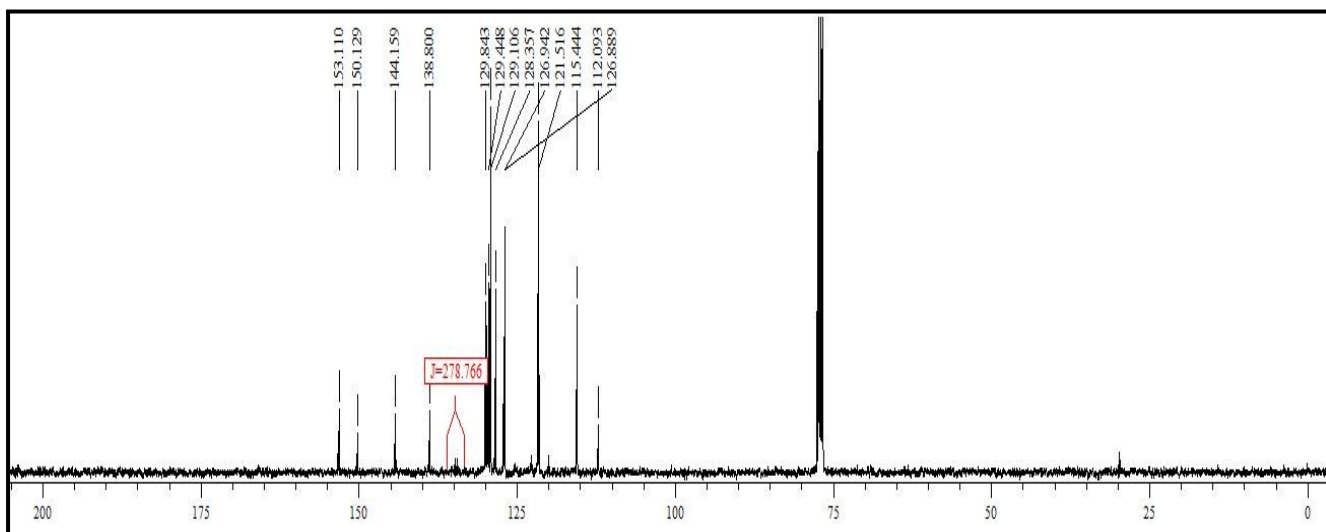
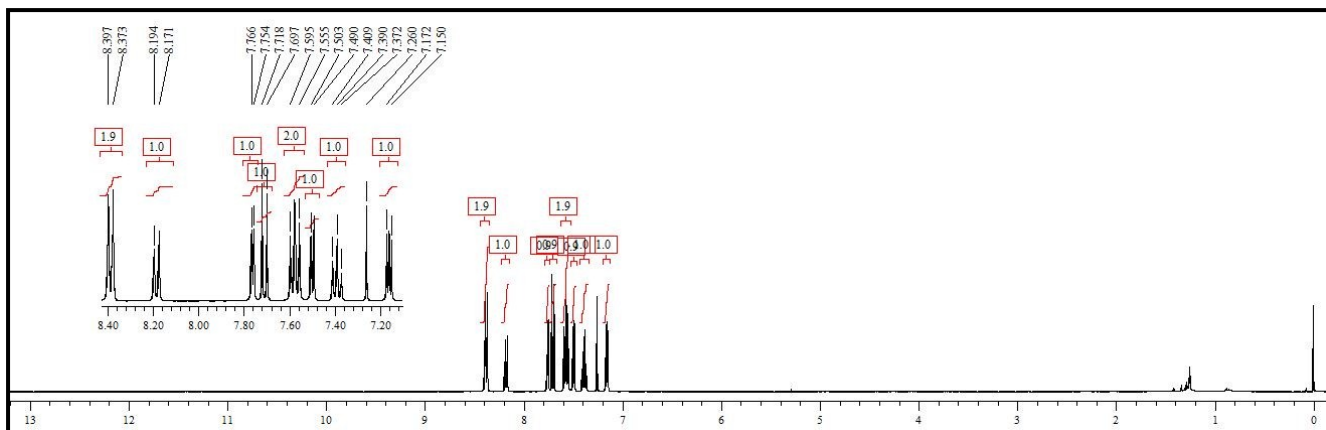
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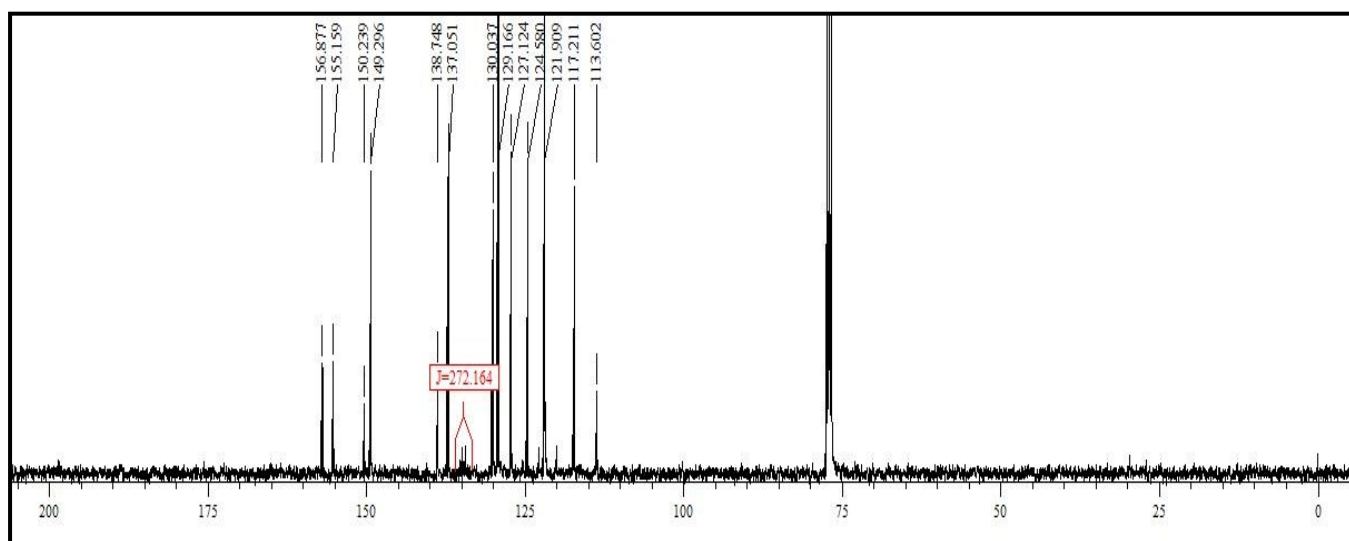
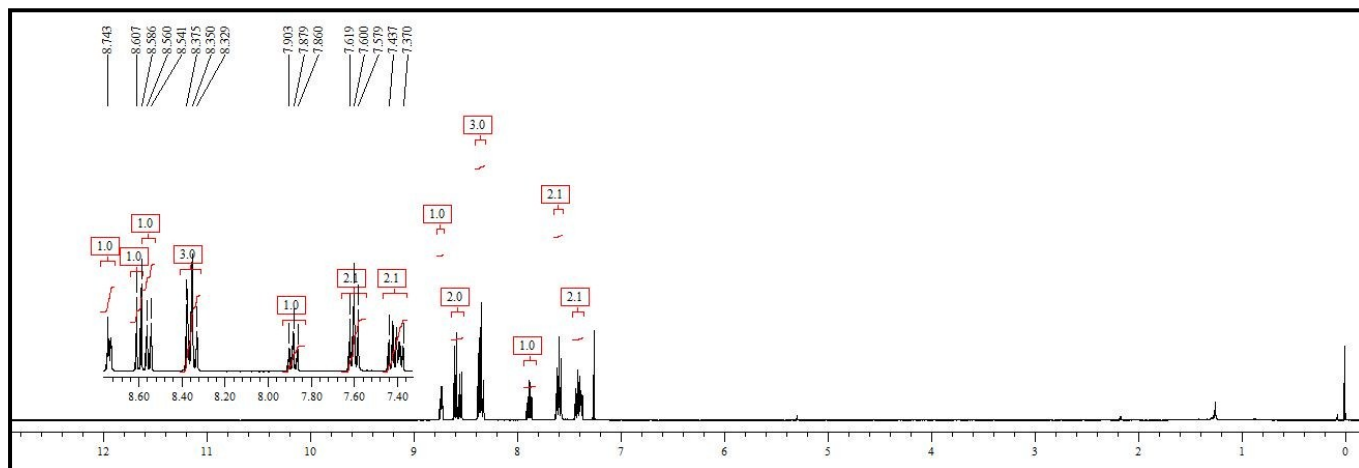
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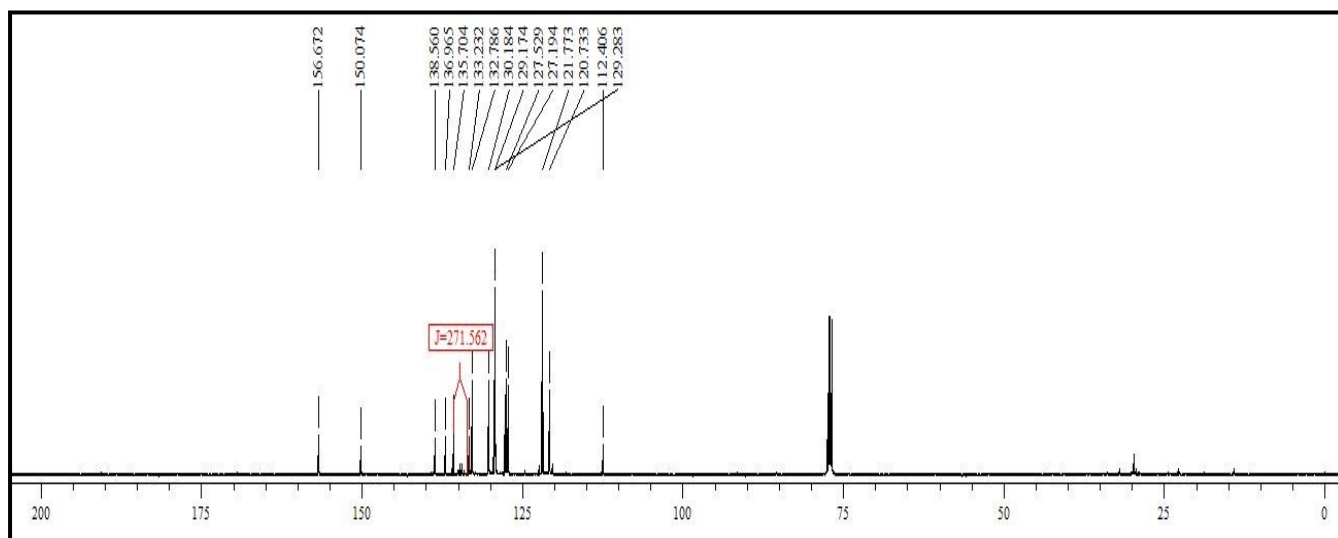
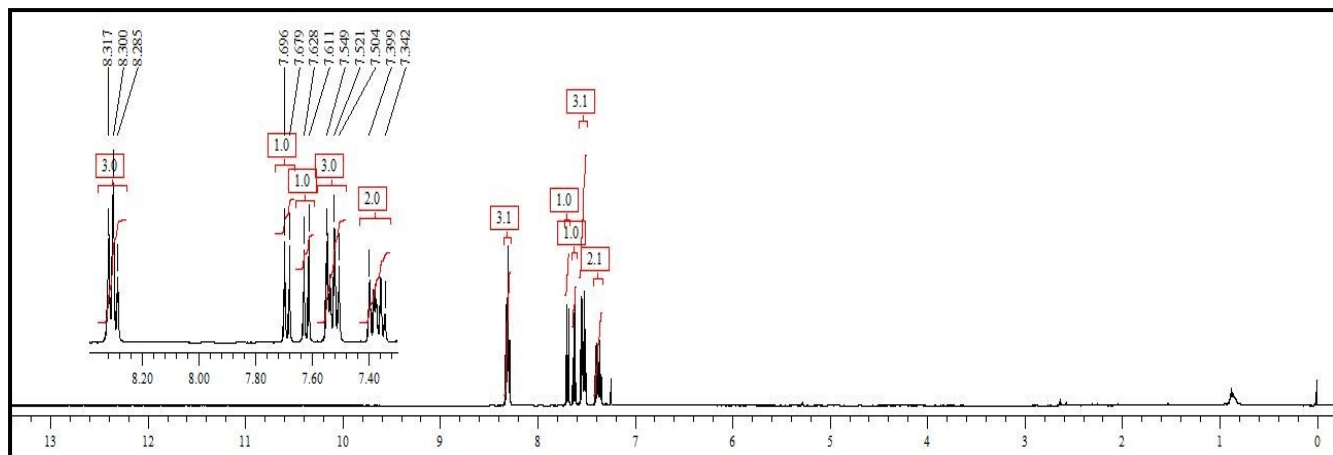
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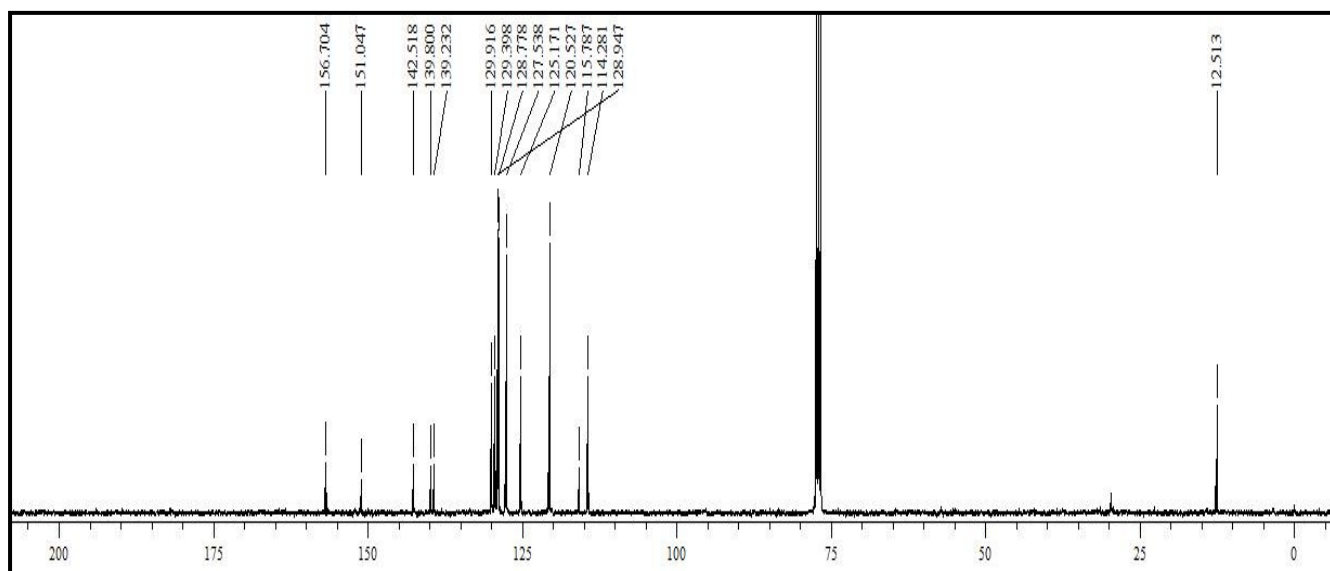
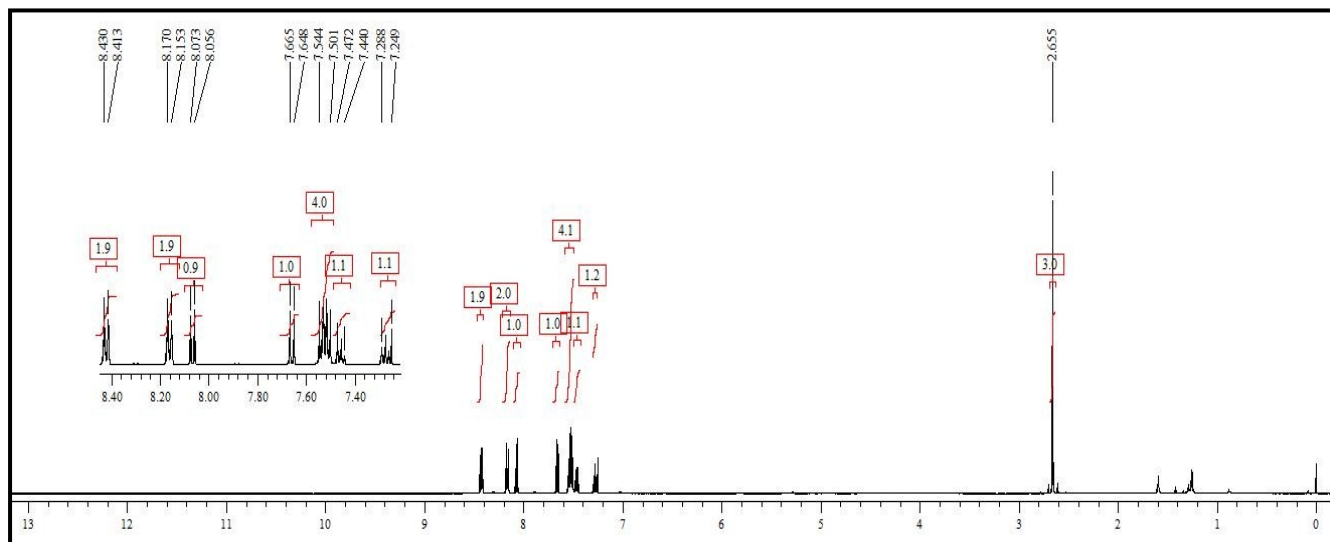
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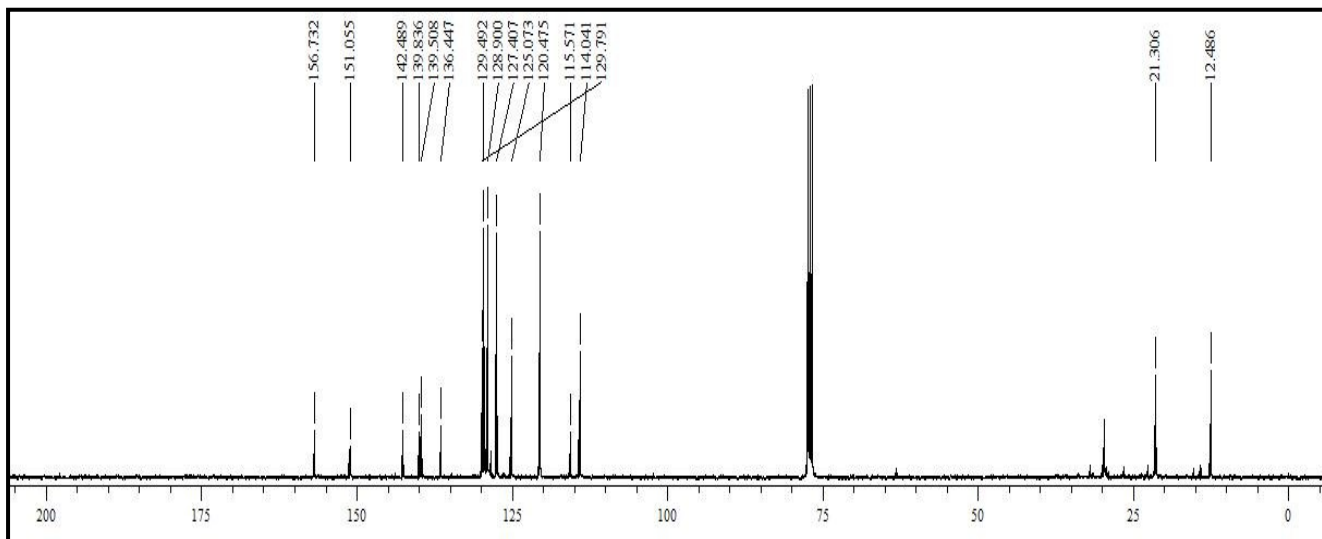
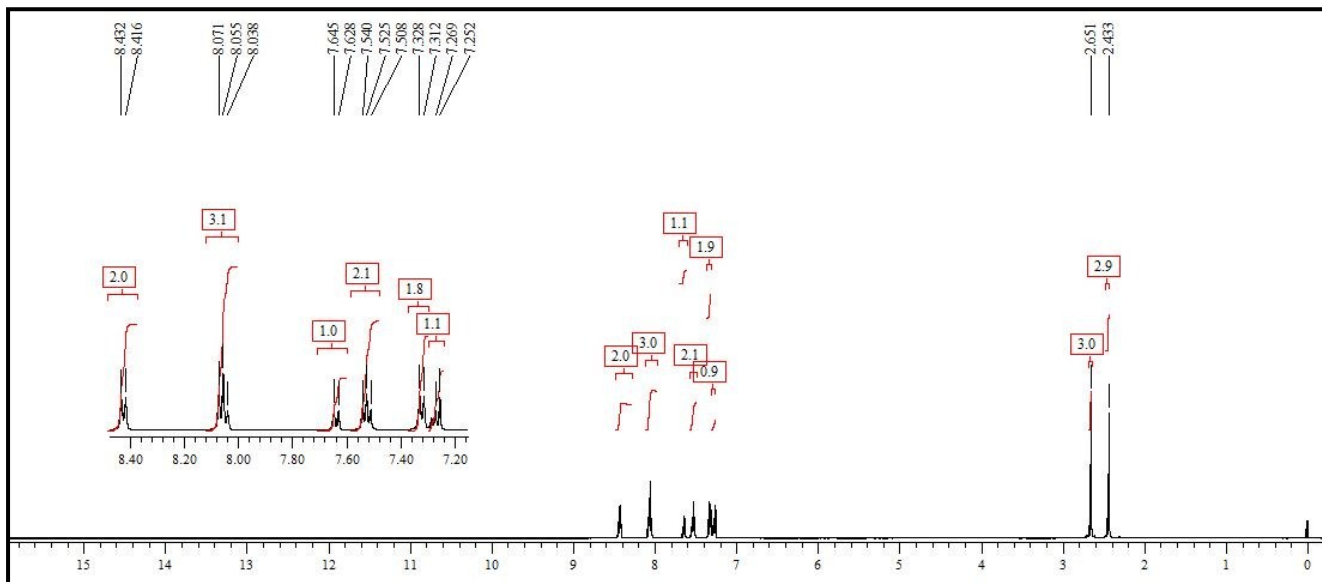
# <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra of compound 4p



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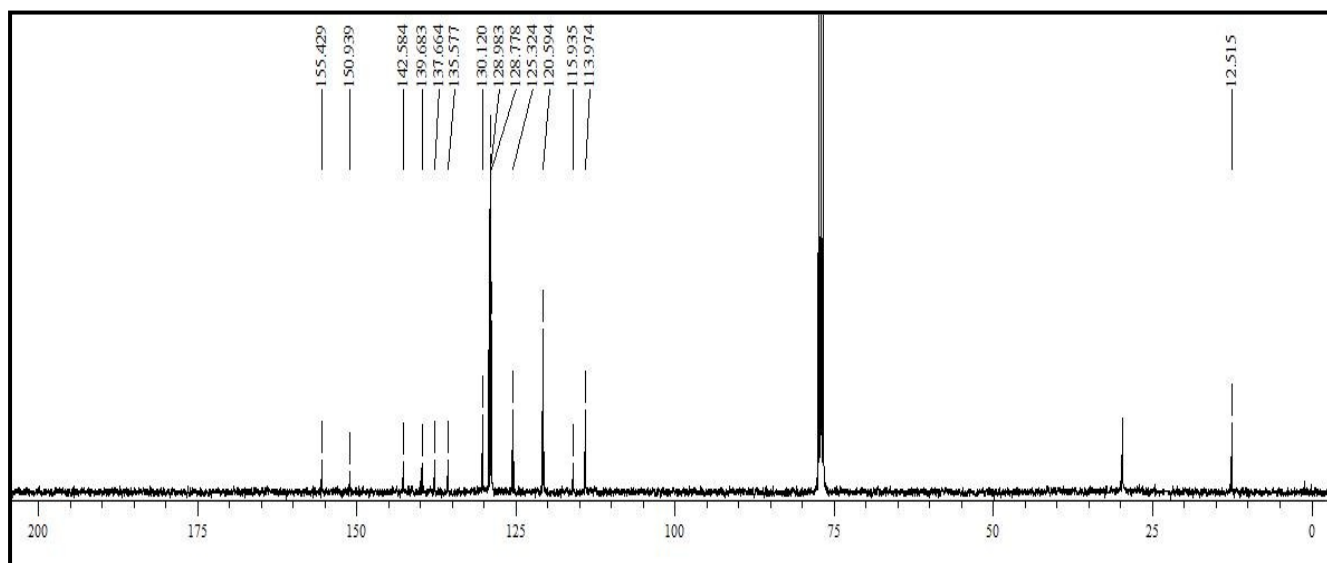
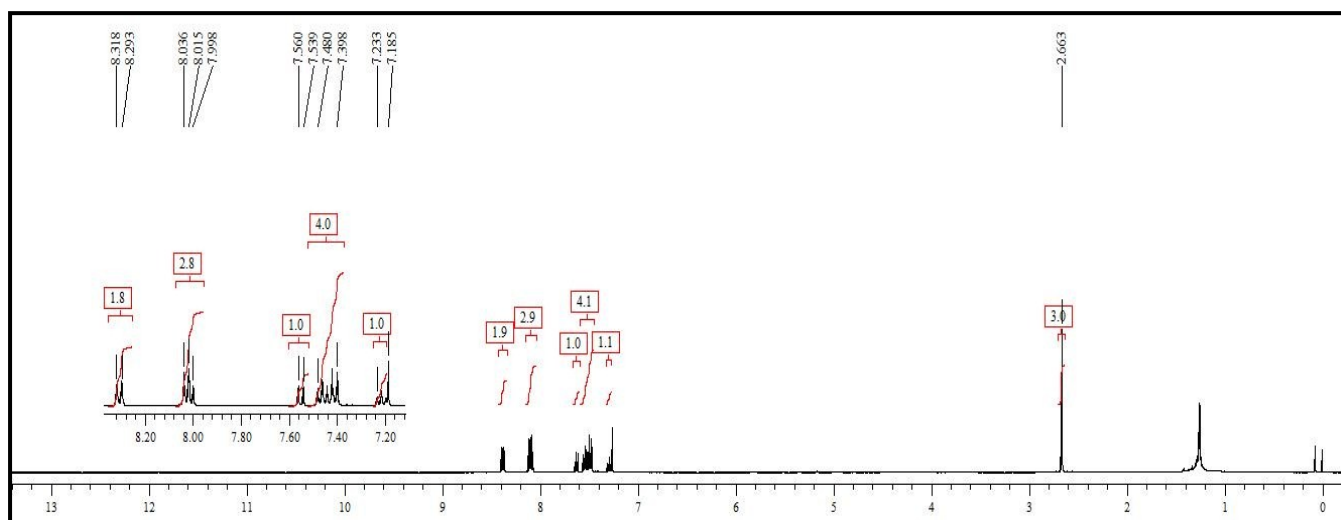


# $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectra of compound 4r





# $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR spectra of compound 4s



# <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra of compound 4t

