Effect of shearing stress on the radial heterogeneity and chromatographic performance of styrene-based polymerised high internal phase emulsions prepared in capillary format

Christopher T. Desire,^a R. Dario Arrua,^b Fotouh R. Mansour,^c Stefan A.F. Bon^d and Emily F. Hilder*^b

^aAustralian Centre for Research on Separation Science (ACROSS), School of Physical Sciences, University of Tasmania, Hobart, Australia

^bFuture Industries Institute, University of South Australia, Adelaide, Australia

^cDepartment of Pharmaceutical Analytical Chemistry, Tanta University, Tanta, Egypt

^dDepartment of Chemistry, The University of Warwick, Coventry, CV4 7AL, United Kingdom

Email: Emily.Hilder@unisa.edu.au; Tel: +(61) 883026292

Supporting Information



1. Column Permeabilities

Fig. S1 Plot of column back pressure (MPa/m) against flow rate (μ L/min) for three poly(Sty-*co*-DVB) poly(HIPE)s prepared from the same batch, using a shear rate of 300 rpm, in 540 μ m i.d. silica capillaries using: **A**) MeOH; **B**) H₂O.



Fig. S2 Plot of column back pressure (MPa/m) against flow rate (μ L/min) for three poly(Sty-*co*-DVB) poly(HIPE)s prepared from the same batch, using a shear rate of 300 rpm, in 250 μ m i.d. silica capillaries using: **A**) MeOH; **B**) H₂O.



Fig. S3 Plot of column back pressure (MPa/m) against flow rate (μ L/min) for three poly(Sty-*co*-DVB) poly(HIPE)s prepared from the same batch, using a shear rate of 300 rpm, in 150 μ m i.d. silica capillaries using: **A**) MeOH; **B**) H₂O.



Fig. S4 Plot of column back pressure (MPa/m) against flow rate (μ L/min) for three poly(Sty-*co*-DVB) poly(HIPE)s prepared from the same batch, using a shear rate of 14 000 rpm, in 540 μ m i.d. silica capillaries using: **A**) MeOH; **B**) H₂O.



Fig. S5 Plot of column back pressure (MPa/m) against flow rate (μ L/min) for three poly(Sty-*co*-DVB) poly(HIPE)s prepared from the same batch, using a shear rate of 14 000 rpm, in 250 μ m i.d. silica capillaries using: **A**) MeOH; **B**) H₂O.



Fig. S6 Plot of column back pressure (MPa/m) against flow rate (μ L/min) for three poly(Sty-*co*-DVB) poly(HIPE)s prepared from the same batch, using a shear rate of 14 000 rpm, in 150 μ m i.d. silica capillaries using: **A)** MeOH; **B)** H₂O.

2. Preparation in capillary format



Fig. S7 SEM images of poly(HIPE)s prepared from emulsions emulsified at 300 rpm or 14 000 rpm in fused silica capillaries of different i.d.: **A)** 540 μ m; **B)** 250 μ m; **C)** 150 μ m. Scale bar is 15 μ m. Images obtained at 500 × magnification.

3. Longitudinal Heterogeneity Study



Fig. S8 SEM images of poly(HIPE)s obtained by curing emulsions which had been emulsified at 300 rpm and passed through the syringe and/or needle. **A)** Bulk; **B)** Syringe; **C)** Syringe and needle. Scale bar is 20 μ m. Images were obtained at 500 × magnification.

Table S1 Porous properties of cured emulsions which had been passed through the syringe and thesyringe and needle.

Sample	V ª / μm	W ^b / μm	D ^c / μm
Bulk	30 ± 10	4 ± 2	20 ± 10
Syringe	30 ± 10	4 ± 2	13 ± 8
Syringe + Needle	20 ± 10	4 ± 2	12 ± 6

^a Average void diameter as determined from SEM. ^b Average window diameter as determined from SEM. ^c Average droplet diameter immediately after preparation as determined from optical microscopy.



Fig. S9 SEM images of poly(HIPE)s obtained from emulsions which had been emulsified at 300 rpm and cured in 20 cm of a 540 μ m i.d. capillary then cut at different lengths from the capillary inlet. **A)** 2 cm; **B)** 5 cm; **C)** 10 cm; **D)** 15 cm; **E)** 18 cm. Scale bar is 50 μ m. Images obtained at 150 × magnification.



Fig. S10 SEM images of poly(HIPE)s obtained from emulsions which had been emulsified at 300 rpm and cured in 20 cm of a 250 μ m i.d. capillary then cut at different lengths from the capillary inlet. **A)** 2 cm; **B)** 5 cm; **C)** 10 cm; **D)** 15 cm; **E)** 18 cm. Scale bar is 25 μ m. Images obtained at 300 × magnification.



Fig. S11 SEM images of poly(HIPE)s obtained from emulsions which had been emulsified at 300 rpm and cured in 20 cm of a 150 μ m i.d. capillary then cut at different lengths from the capillary inlet. **A)** 2 cm; **B)** 5 cm; **C)** 10 cm; **D)** 15 cm; **E)** 18 cm. Scale bar is 15 μ m. Images obtained at 500 × magnification.

540 μm i.d.	V ª / μm	W ^b / μm
2 cm	12 ± 5	3 ± 1
5 cm	12 ± 5	3 ± 1
10 cm	14 ± 6	3 ± 2
15 cm	12 ± 4	4 ± 2
18 cm	12 ± 7	2 ± 1
250 μm i.d.	V ª / μm	W ^b / μm
2 cm	12 ± 4	3 ± 2
5 cm	9 ± 3	3 ± 2
10 cm	9 ± 4	2 ± 1
15 cm	8 ± 4	2 ± 1
18 cm	9 ± 7	3 ± 1
150 μm i.d.	V ª / μm	W ^b / μm
2 cm	9 ± 4	3 ± 1
5 cm	8 ± 6	3 ± 1
10 cm	9 ± 4	2 ± 1
15 cm	9 ± 3	2.3 ± 0.9
18 cm	8 ± 6	2 ± 1

Table S2 Porous properties of poly(HIPE)s obtained from emulsions which had been emulsified at 300 rpm and cured in 20 cm of different i.d. capillaries then cut at different lengths from the capillary inlet.

^a Average void diameter as determined from SEM. ^b Average window diameter as determined from SEM.

4. Influence of Filling Rate

Table S3 Porous properties of poly(HIPE)s obtained from emulsions which had been emulsified at 300 rpm and passed through 20 cm of 250 or 150 μ m i.d. capillaries at different rates.

Filling	250 μm			150 μm		
Rate / µLmin ⁻¹	V ª / μm	W ^b / μm	D ^c / μm	V ª / µm	W ^b / μm	D ^c / μm
10	30 ± 10	5 ± 3	10 ± 5	21 ± 7	4 ± 2	9 ± 5
25	30 ± 10	4 ± 2	11 ± 6	30 ± 10	5 ± 4	8 ± 4
50	30 ± 10	4 ± 2	8 ± 5	30 ± 10	4 ± 2	9 ± 5
100	21 ± 9	3 ± 2	9 ± 5	19 ± 8	3 ± 1	9 ± 5

 $^{\rm a}$ Average void diameter as determined from SEM. $^{\rm b}$ Average window diameter as determined from SEM.

^c Average droplet diameter immediately after preparation as determined from optical microscopy.



Fig. S12 SEM images of poly(HIPE)s obtained from emulsions which had been emulsified at 300 rpm, passed through 20 cm of 250 μ m i.d. capillary at different rates and cured. **A)** 10 μ L/min; **B)** 25 μ L/min; **C)** 50 μ L/min; **D)** 100 μ L/min. Scale bar is 15 μ m. Images obtained at 500 × magnification.



Fig. S13 SEM images of poly(HIPE)s obtained from emulsions which had been emulsified at 300 rpm, passed through 20 cm of 150 μ m i.d. capillary at different rates and cured. **A)** 10 μ L/min; **B)** 25 μ L/min; **C)** 50 μ L/min; **D)** 100 μ L/min. Scale bar is 15 μ m. Images obtained at 500 × magnification.

5. Influence of Capillary Length



Fig. S14 SEM images of poly(HIPE)s obtained from emulsions which had been emulsified at 300 rpm and passed through different lengths of $250 \ \mu m$ i.d. capillary and then cured. A) 5 cm; B) 10 cm; C) 20

cm; **D)** 30 cm; **E)** 40 cm; **F)** 50 cm; **G)** 60 cm; **H)** Bulk. Scale bar is 15 μ m. Images obtained at 500 × magnification.



Fig. S15 SEM images of poly(HIPE)s obtained from emulsions which had been emulsified at 300 rpm and passed through different lengths of 150 μ m i.d. capillary and then cured. **A)** 5 cm; **B)** 10 cm; **C)** 20

cm; **D**) 30 cm; **E**) 40 cm; **F**) 50 cm; **G**) 60 cm; **H**) Bulk. Scale bar is 15 μ m. Images obtained at 500× magnification.



6. Chromatography

Fig. S16 The separation of ribonuclease A (1), lysozyme (2) and α -chymotrypsinogen A (3) under reversed-phase conditions. Conditions: 18 cm of different i.d. columns: **(A)** 540 μ m i.d., **(B)** 250 μ m i.d., **(C)** 150 μ m i.d., prepared with different emulsification energies. Eluent A was 0.1 vol% formic acid in Milli-Q H₂O, and eluent B was 0.1 vol% formic acid in acetonitrile; injection volume, 1 μ L; protein concentration, 0.05 mg/mL for **(B)** and **(C)** and 0.3 mg/mL for **(A)**. Gradient: linear gradient 15 to 70% B in 15 min and then isocratic elution at 70% B for 5 min before returning to 15% B in 5 min; flow rate, 4.0 μ L/min. UV detection at 214 nm.



Fig. S17 The separation of ribonuclease A (1), lysozyme (2) and α -chymotrypsinogen A (3) under reversed-phase conditions. Conditions: 18 cm of different i.d. columns: **(A)** 540 µm i.d., **(B)** 250 µm i.d., **(C)** 150 µm i.d., prepared with different emulsification energies. Eluent A was 0.1 vol% formic acid in Milli-Q H₂O, and eluent B was 0.1 vol% formic acid in acetonitrile; injection volume, 1 µL; protein concentration, 0.3 mg/mL. Gradient: linear gradient 15 to 50% B in 15 min and then isocratic elution at 50% B for 5 min before returning to 15% B in 5 min; flow rate, 8.0 µL/min. UV detection at 214 nm.

7. Optical Microscopy



Fig. S18 Optical microscopy images of emulsions which had been emulsified at 300 rpm and passed through the syringe and/or needle. **A)** Bulk; **B)** Syringe; **C)** Syringe and needle. Scale bar is 50 µm.



Fig. S19 Optical microscopy images of emulsions which had been emulsified at 300 rpm and passed through 20 cm of various i.d. capillary columns: **A)** Bulk; **B)** 540 μ m i.d.; **C)** 250 μ m i.d.; **D)** 150 μ m i.d. Scale bar is 50 μ m.



Fig. S20 Optical microscopy of emulsions which had been emulsified at 300 rpm and passed through 20 cm of 250 μ m i.d. capillary at different rates. **A)** 10 μ L/min; **B)** 25 μ L/min; **C)** 50 μ L/min; **D)** 100 μ L/min. Scale bar is 50 μ m.



Fig. S21 Optical microscopy of emulsions which had been emulsified at 300 rpm and passed through 20 cm of 150 μ m i.d. capillary at different rates. **A)** 10 μ L/min; **B)** 25 μ L/min; **C)** 50 μ L/min; **D)** 100 μ L/min. Scale bar is 50 μ m.



Fig. S22 Optical microscopy images of emulsions which had been emulsified at 300 rpm and passed through different lengths of 250 μm i.d. capillary. **A)** 5 cm; **B)** 10 cm; **C)** 20 cm; **D)** 30 cm; **E)** 40 cm; **F)** 50 cm; **G)** 60 cm; **H)** Bulk. Scale bar is 50 μm.



Fig. S23 Optical microscopy images of emulsions which had been emulsified at 300 rpm and passed through different lengths of 150 μm i.d. capillary. **A)** 5 cm; **B)** 10 cm; **C)** 20 cm; **D)** 30 cm; **E)** 40 cm; **F)** 50 cm; **G)** 60 cm; **H)** Bulk. Scale bar is 50 μm.



Fig. S24 Optical microscopy images of emulsions which had been emulsified at 14 000 rpm and passed through 20 cm of various i.d. capillary columns: **A)** Bulk; **B)** 540 μ m i.d.; **C)** 250 μ m i.d.; **D)** 150 μ m i.d. Scale bar is 50 μ m.