

Supporting Information

Nitro-functionalized Metal–organic Frameworks with Catalase Mimic Properties for Glutathione Detection

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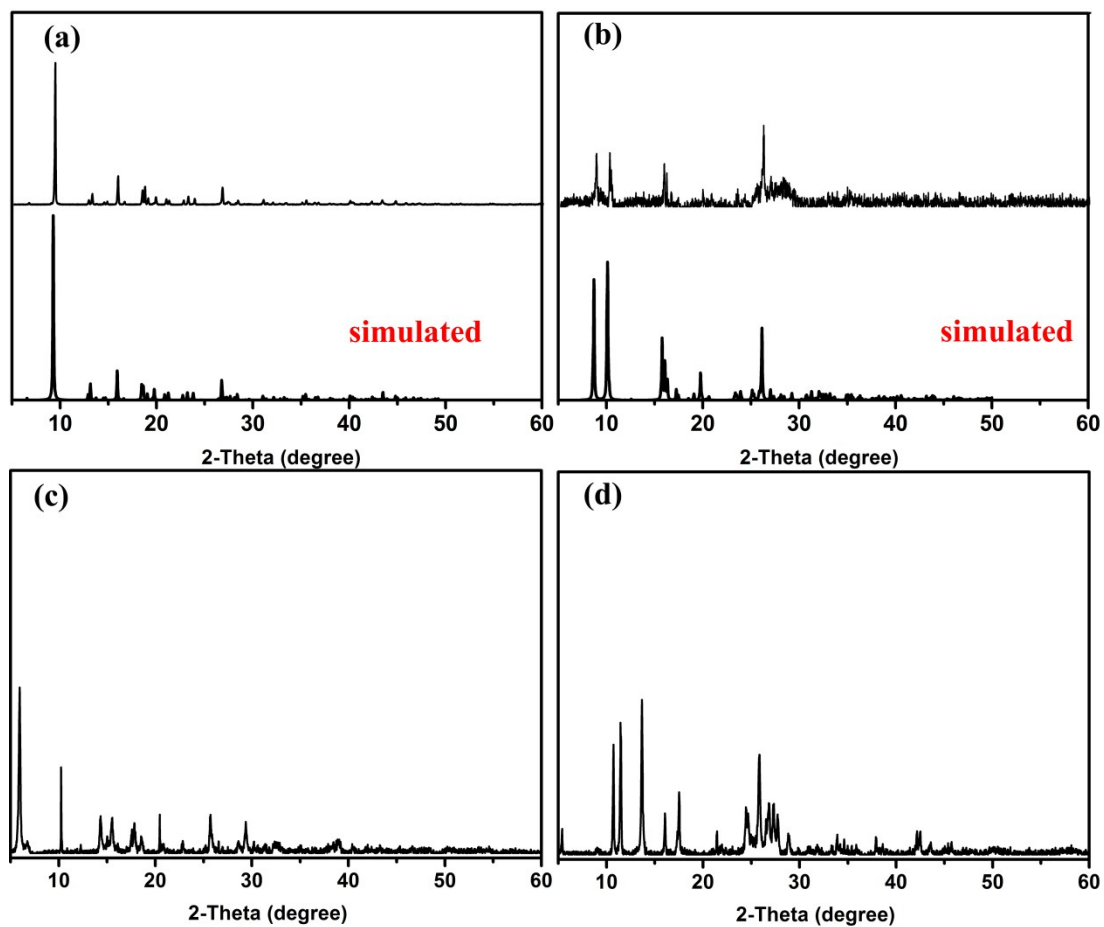


Figure S1. XRD patterns of (a) Cu-MOFs (i), (b) Cu-MOFs (ii), (c) Cu-MOFs (iii), (d) Cu-MOFs (iv).

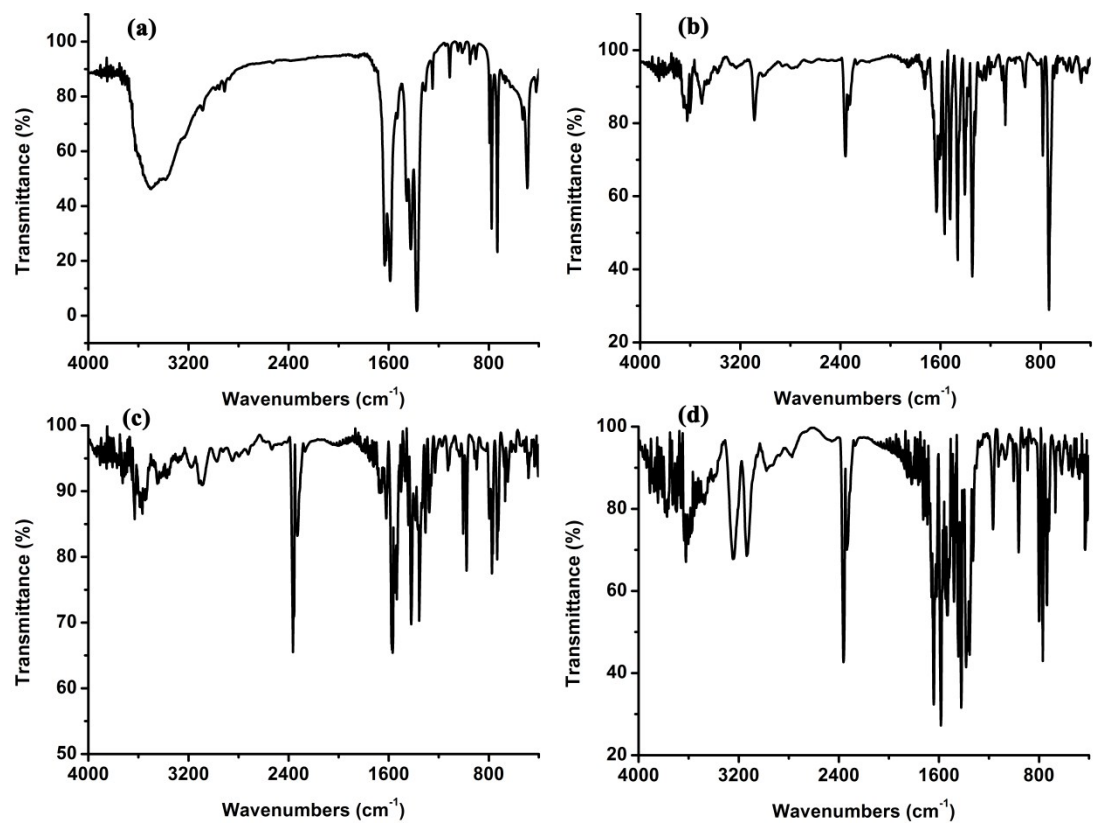


Figure S2. FTIR spectrum of (a) Cu-MOFs (i), (b) Cu-MOFs (ii), (c) Cu-MOFs (iii), (d) Cu-MOFs (iv).

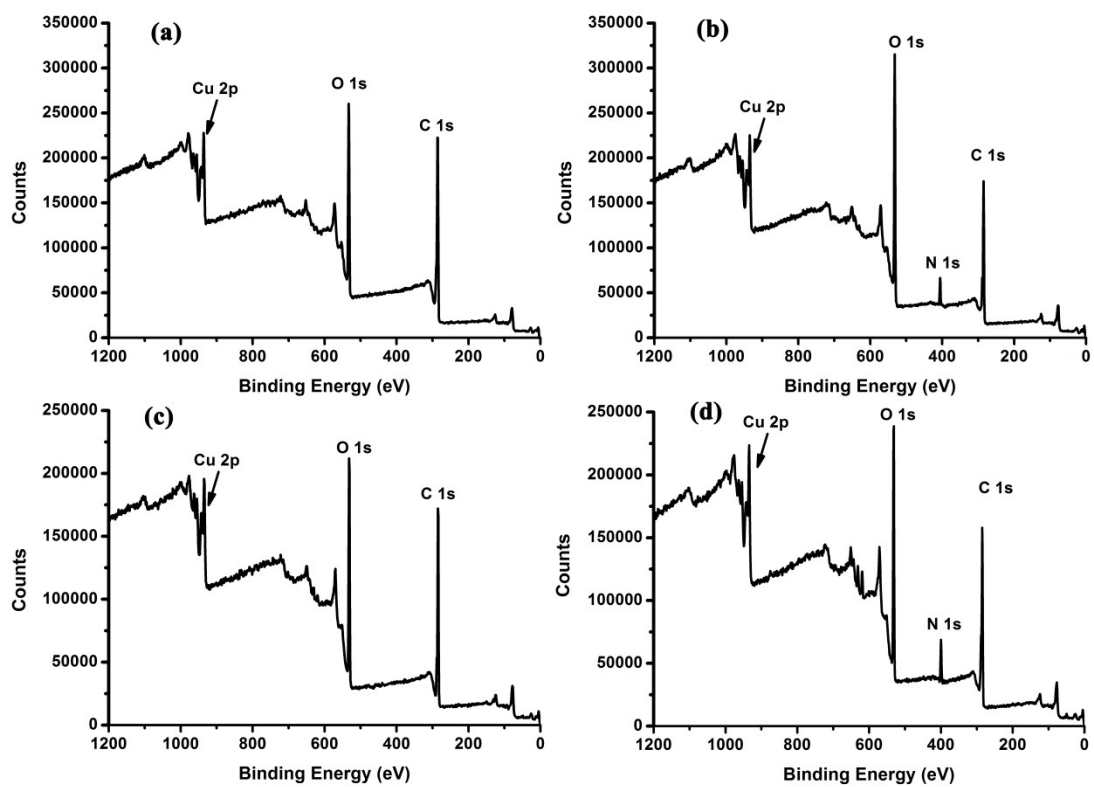


Figure S3. Survey XPS (a) Cu-MOFs (i), (b) Cu-MOFs (ii), (c) Cu-MOFs (iii), (d) Cu-MOFs (iv).

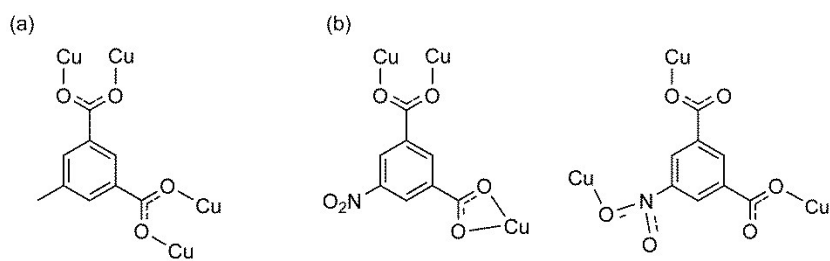


Figure S4 Coordination Modes of ligand i (a) and ligand ii (b) with Cu^{2+} .

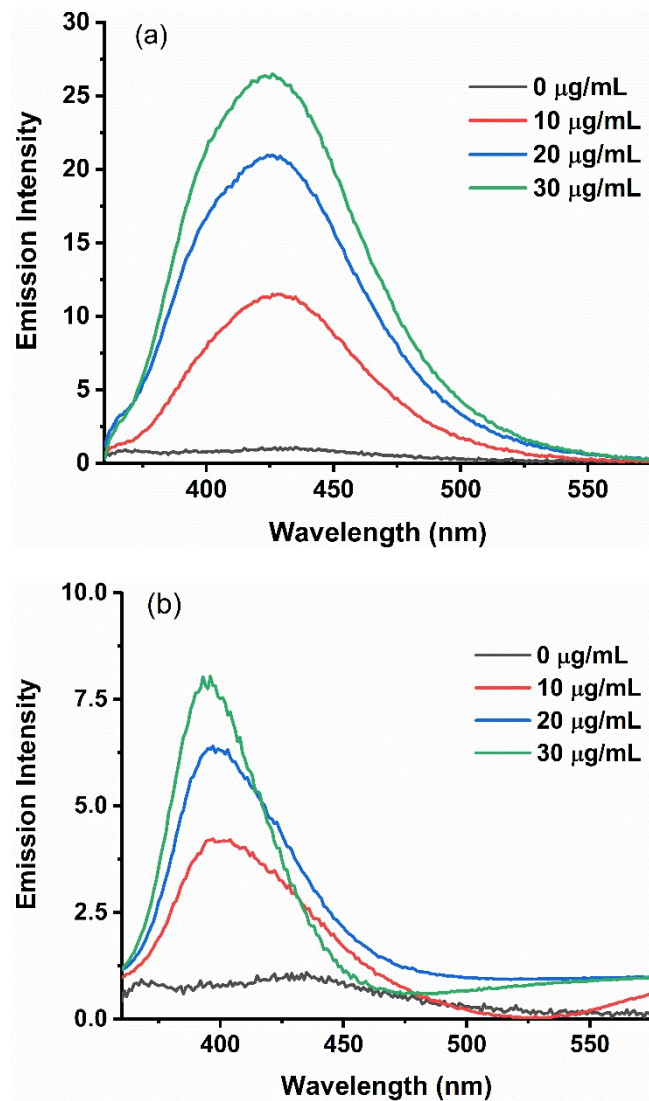


Figure S5. Effects of (a) Cu-MOFs (i) and (b) Cu-MOFs (iv) on the changes of hydroxyl radicals with terephthalic acid as the fluorescence probe. Assay conditions: 5 μM terephthalic acid, 4 mM H_2O_2 , pH 7.0, 5 min. The emission intensity of Cu-MOFs (iv) itself was subtracted.

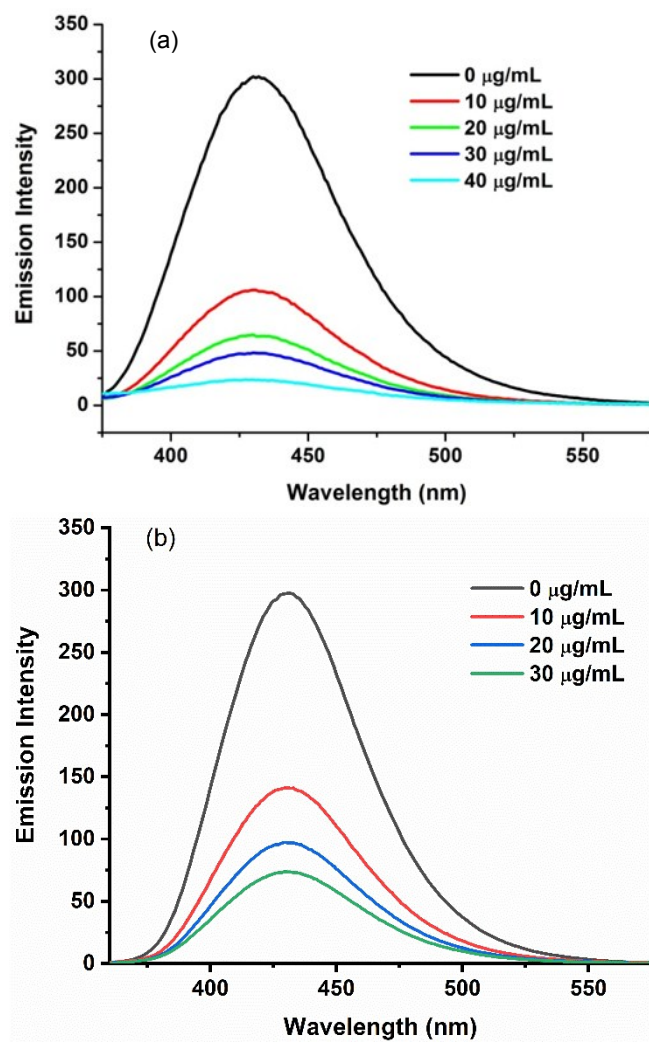


Figure S6. Effects of (a) Cu-MOFs (ii) or (b) Cu-MOFs (iii) on the changes of hydroxyl radicals with terephthalic acid as the fluorescence probe. Assay conditions: 5 µM terephthalic acid, 4 mM H₂O₂, pH 7.0, 5 min.

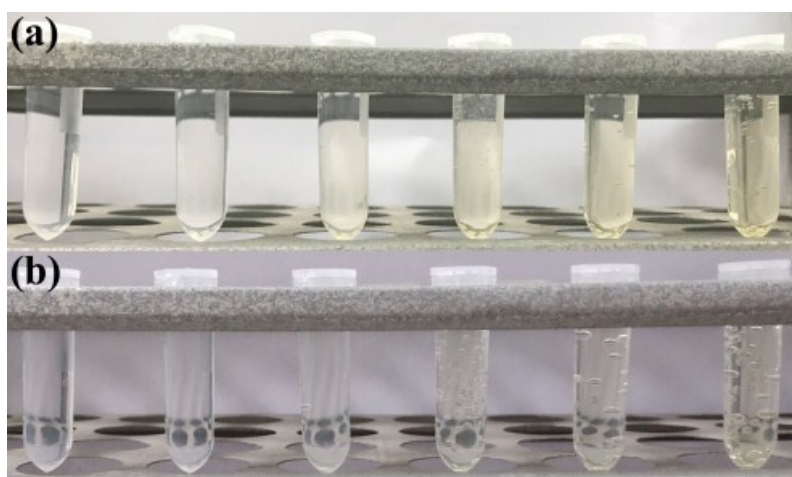


Figure S7. Photograph of gas bubble generated by the decomposition of H_2O_2 in pH 7.0. (a) Reaction for 30 minutes. (b) Reaction for 60 minutes. From left to right, the concentrations of Cu-MOFs (ii) were 0, 10, 20, 30, 40 and 50 $\mu\text{g}/\text{mL}$ respectively.

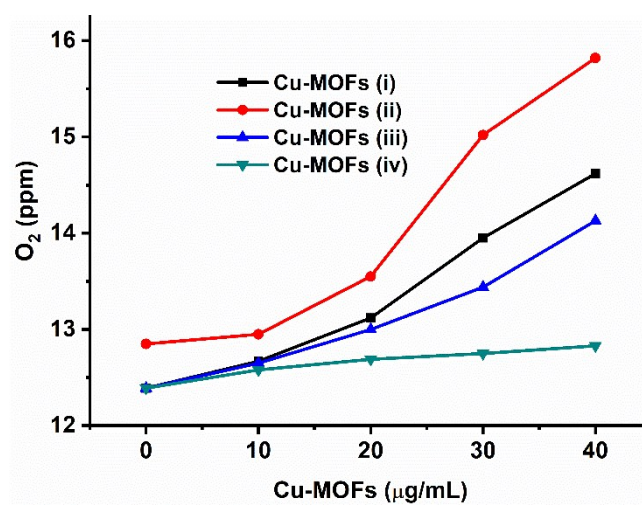


Figure S8. Effect of Cu-MOFs (ii) concentration on the generation of O_2 by the decomposition of H_2O_2 . Assay conditions: 4 mM H_2O_2 and different concentrations of Cu-MOFs (pH 7.0).

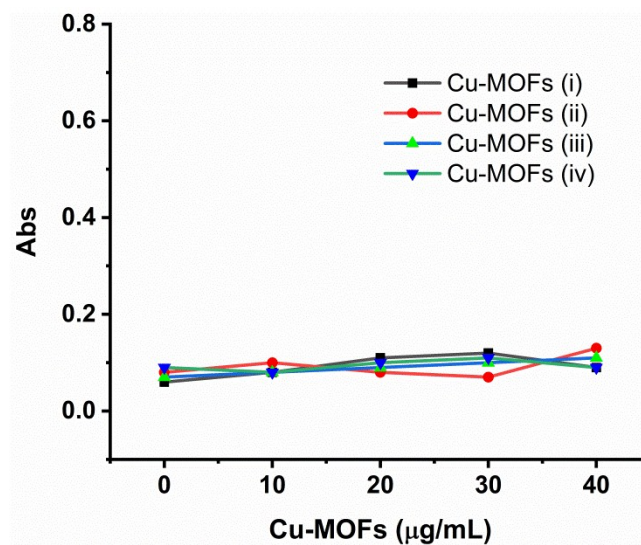


Figure S9. Study of oxidase-like properties of Cu-MOFs. Assay conditions: 0.5 mM TMB and different concentrations of Cu-MOFs (pH 7.0).

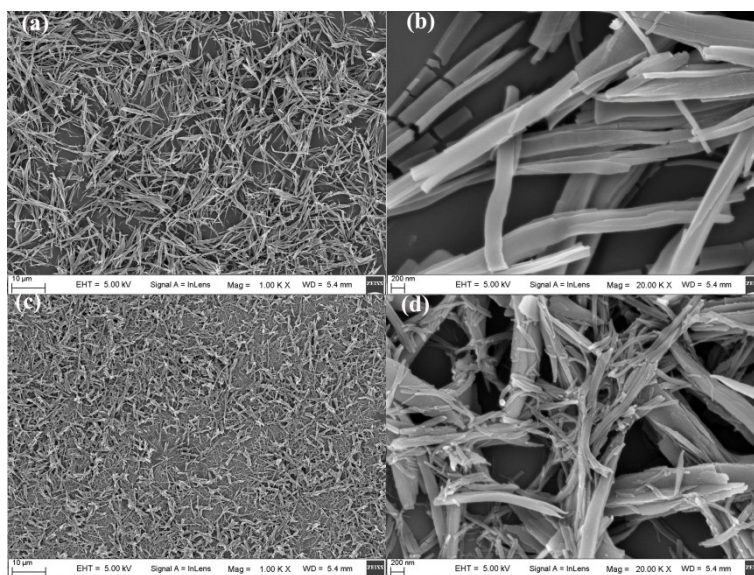


Figure S10. SEM images of Cu-MOFs (ii) before (a), (b) and after (c), (d) the catalytic reaction.

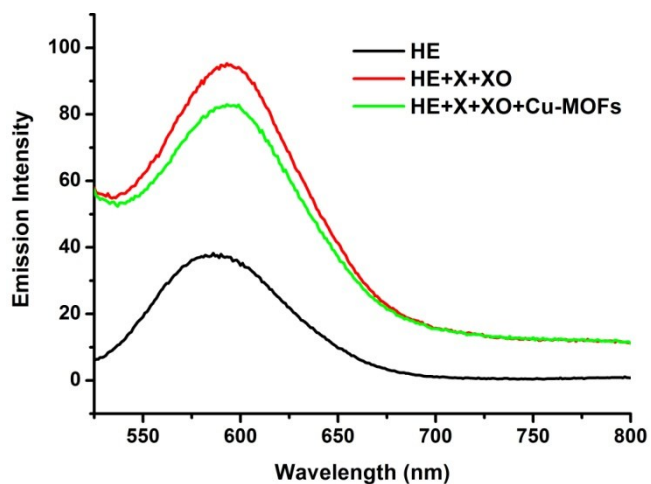


Figure S11. Effects of Cu-MOFs (ii) on the changes of $\cdot\text{O}_2^-$ using dihydroethidium as the fluorescence probe. Assay conditions: 50 μM dihydroethidium, 1 mM X, 1 U/mL XO, 0.1 mM DTPA, 30 $\mu\text{g/mL}$ Cu-MOFs (ii), excitation wavelength, 480 nm.

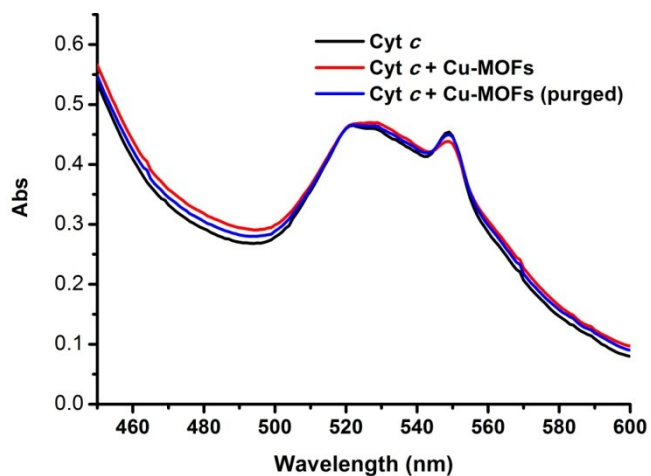


Figure S12. UV-vis spectrum of Cyt *c*, Cyt *c* reacted with Cu-MOFs (ii) and Cyt *c* reacted with Cu-MOFs (ii) under deoxygenated condition. Assay conditions: 0.5 mg/ mL Cyt *c*, 20 $\mu\text{g/mL}$ Cu-MOFs (ii).

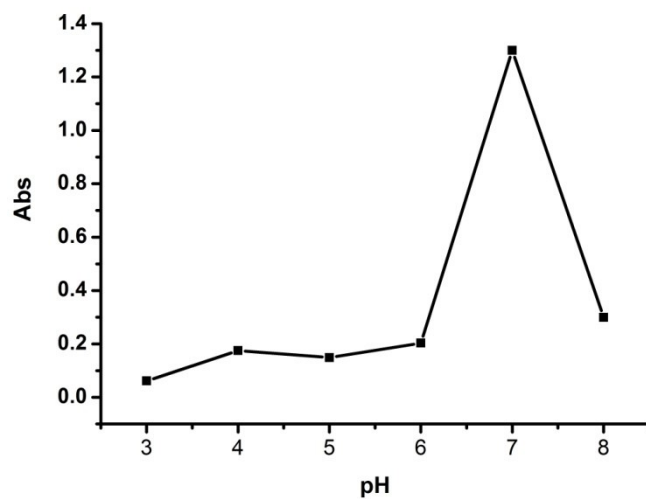


Figure S13. Effect of pH on catalytic activity of the Cu-MOFs (ii). Assay conditions: 0.5 mM TMB, 4 mM H₂O₂, 20 µg/mL Cu-MOFs (ii), 5 min.

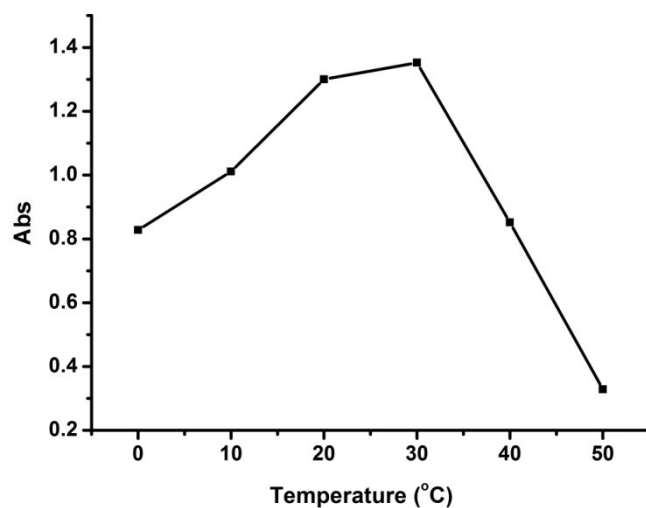


Figure S14. Dependence of the catalytic activity of Cu-MOFs (ii) on temperature. Assay conditions: 0.5 mM TMB, 4 mM H₂O₂, pH 7.0, 5 min.

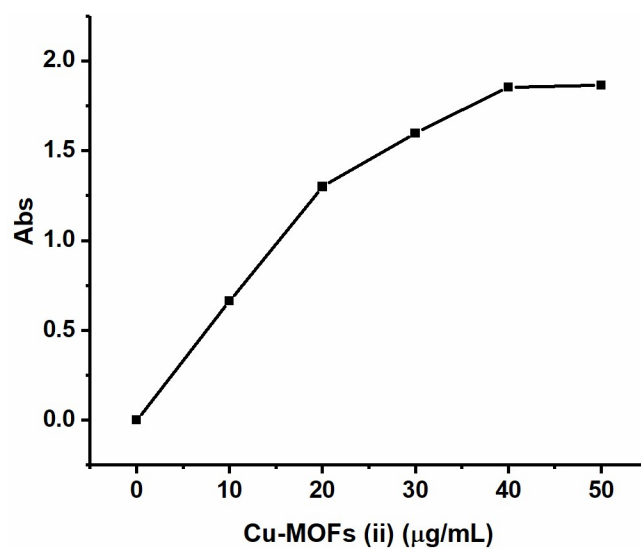


Figure S15. Effect of the amount of Cu-MOFs (ii). Assay conditions: 0.5 mM TMB, 4 mM H₂O₂, pH 7.0, 5 min.

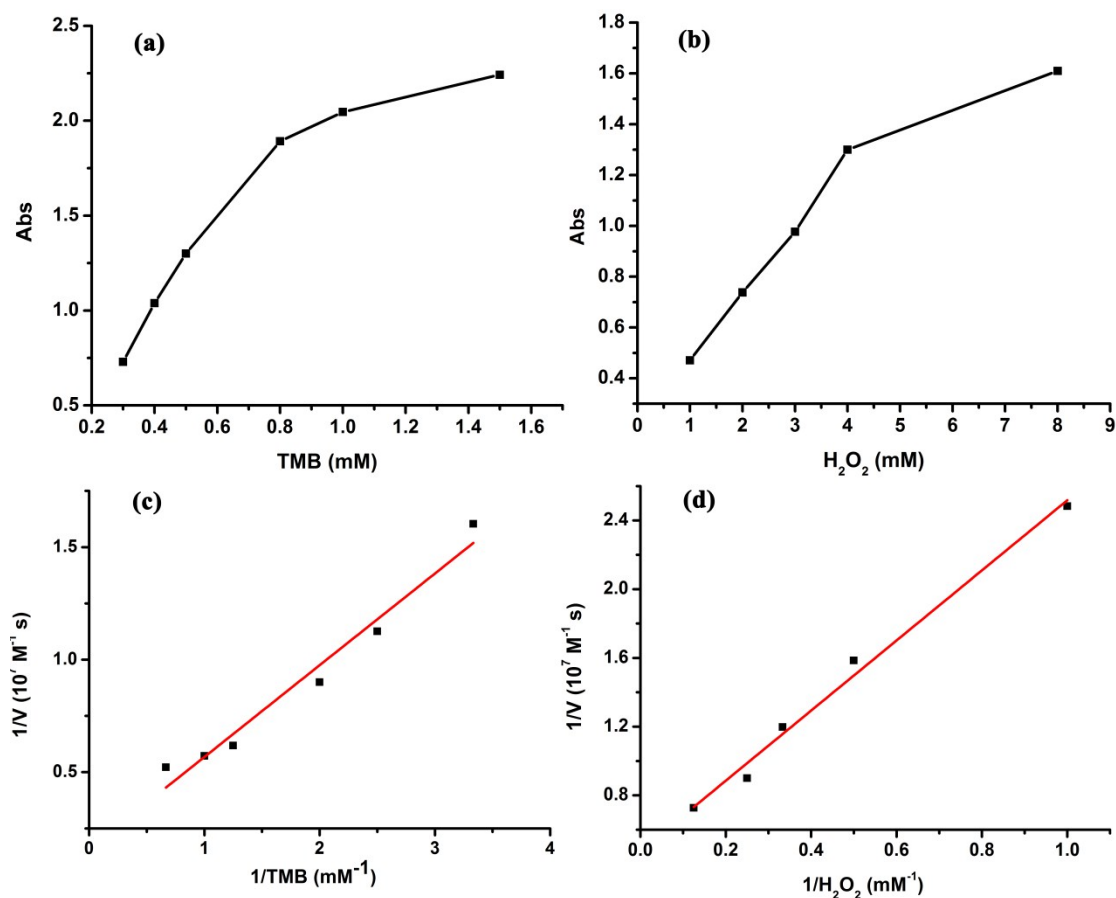


Figure S16. Steady-state kinetics measurements of the Cu-MOFs (ii). (a) The concentration of H₂O₂ was 5 mM and the TMB concentration varied. (b) The concentration of TMB was 0.5 mM and the H₂O₂ concentration varied. (c) Double reciprocal plots of activity of Cu-MOFs (ii) with the concentration of TMB varied. The TMB concentration was 0.3, 0.4, 0.5, 0.8, 1.0 and 1.5 mM, respectively. (d) Double reciprocal plots of activity of Cu-MOFs (ii) with the concentration of H₂O₂ varied. The H₂O₂ concentration was 1, 2, 3, 4 and 8 mM, respectively.

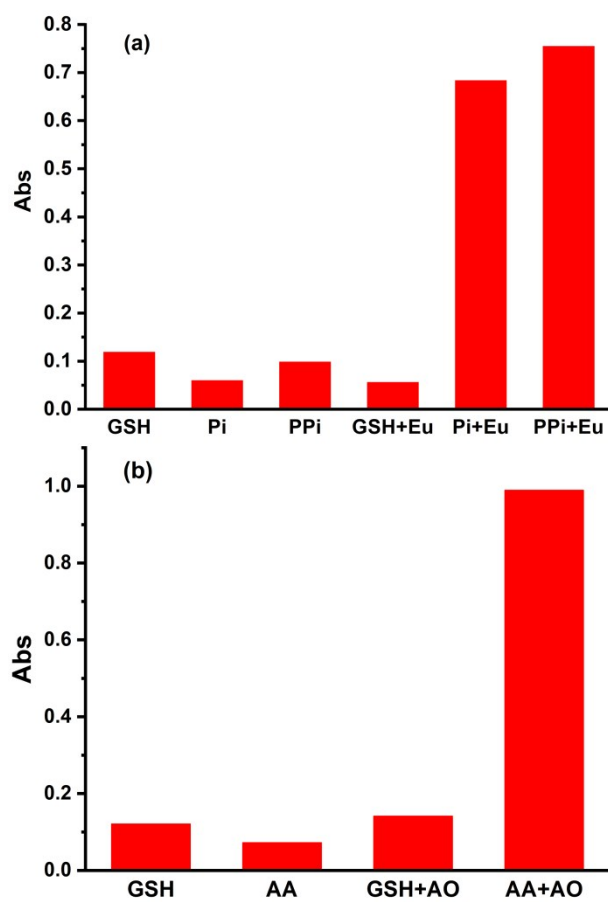


Figure S17. The effects of (a) Phosphate species or (b) ascorbic acid on GSH detection. Assay conditions: 0.5 mM TMB, 4 mM H₂O₂, 20 µg/mL Cu-MOFs (ii), 20 µM Eu³⁺, 1 U/mL AO, 5 min.

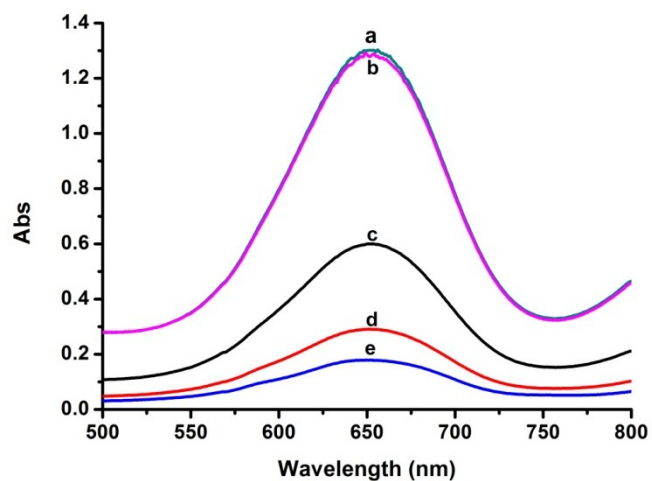


Figure S18 GSH detection in serum. line a: TMB+H₂O₂+ Cu-MOFs (ii); line b: TMB+H₂O₂+ Cu-MOFs (ii)+serum; line b: TMB+H₂O₂+ Cu-MOFs (ii)+serum+25 μ M GSH; line c: TMB+H₂O₂+ Cu-MOFs (ii)+serum+55 μ M GSH; line d: TMB+H₂O₂+ Cu-MOFs (ii)+serum; line e: TMB+H₂O₂+ Cu-MOFs(ii)+serum+75 μ M GSH.

Table S1. Comparison of the BET surface areas and pore volumes of different Cu-MOFs.

	(i)	(ii)	(iii)	(iv)
BET Surface Area (m ² /g)	5.09	1.81	14.07	8.56

Table S2. Zeta potential of the obtained Cu-MOFs.

	1	2	3	average
Cu-MOFs (i)	-1.96	-2.04	-1.9	-1.96
Cu-MOFs (ii)	-2.6	-1.71	-2.1	-2.13
Cu-MOFs (iii)	5.56	4.96	4.6	5.04
Cu-MOFs (iv)	-20.6	4.96	-19.7	-11.78

Table S3. Comparison of the Michaelis-Menten constant (K_m) and maximum reaction rate (V_m).

catalyst	K_m (mM)		ref
	TMB	H ₂ O ₂	
Catalase	-	4.1	37
Cu-MOFs (ii)	2.56	4.34	This work

Table S4. Absorbance of six blank samples and calculated Y axes signals

i	1	2	3	4	5	6
A_i	1.2997	1.2883	1.3021	1.2781	1.3103	1.3142
$y_i = \log(A_0/A)$	-0.000308	0.00352	-0.00111	0.00697	-0.00384	-0.00513

Note: i represented the number of six blank samples, and A_0 was the average of six absorbance values (652 nm). And the limit of detection of calculated as follows:

$$\sigma = \sqrt{\frac{1}{n-1} \sum_{i=1}^6 (y_i - y_{aver})^2} = 0.00454$$
$$\text{LOD} = 3 \times 0.00454 / 0.014 = 0.97 \text{ } (\mu\text{M})$$