Supporting Information

A Flexible Smart Membrane Consisting of GO Composite Fibres and

Upconversion MSNs for microRNA Detection

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Materials

Tetraethoxysilane (TEOS), cetyltrimethylammonium bromide (CTAB), ethanol, calcium acetate monohydrate (99%), anhydrous N,N-dimethyl-formamide (DMF), tetrahydrofuran(99%) were acquired from Sinopharm Chemical Reagent Co., Ltd. Lysine, n-octane (99%), 2, 2'-azobis(2-amidinopropane) dihydrochloride (AIBA), styrene (99%, contains 4-tert-butylcatechol as stabilizer), trifluoroacetic acid (TFA, 99.5%), 1-ethyl-3-(3-(dimethylamino)propyl) carbodiimide hydrochloride (EDC), N-hydroxysulfosuccinimide (sulfo-NHS), 3-aminopropyltriethoxysilane (APTES) were purchased from Sigma-Aladdin Industrial Co., Ltd. Ytterbium(III) acetate hydrate (99.9%), holmium(III) acetate hydrate (99.9%) were purchased from Thermo Fisher Scientific. All chemicals were used as received without further purification. TPU was purchased by Bayer A.G. (Germany).

DNA oligonucleotides with a concentration of 100 nM were ordered from Sangon Biotech (Shanghai) Co., Ltd., and the sequence are listed in Table 1.

Characterization

The morphology of materials was investigated via a Hitachi SU-70 field-emission scanning electron microscopy (FESEM) and a FEI Tecnai F20 high-resolution transmission electron microscope (HRTEM). The X-ray diffraction (XRD) pattern was recorded by using a Thermo ARL X'TRA powder diffractometer. Surface textural characterization was implemented according to the Brunauer–Emmett–Teller (BET) method by using a Coulter OMNISORP-100 apparatus. FTIR spectra were evaluated using a Perkin-Elmer 580B infrared spectrophotometer. UV/Vis absorption values were measured with a TU-1810 spectrophotometer. The upconversion photoluminescence spectra were recorded using a fluorescence spectrophotometer (PL, FLSP920, Edinburgh) under the excitation of a 980 nm laser.

Synthesis of CaF₂: Yb/Ho @MSN

100 mg CTAB were dissolved in 30 ml deionized water, and stirred at 60°C. After stirred magnetically for 30 min, 25.2 ml octane, 1000 mg TEOS, 22 mg lysine, styrene (30 mg/ml) and AIBA (0.84 mg/ml) were subsequently added to the system and stirred for 3 h. After 4 h, the solution was cooled down to the ambient temperature. The products were collected by centrifugation at 10000 rpm for 10 min and then washed 3 times. The template was completely removed by heat treatment at 550°C under atmospheric conditions. To incorporate CaF₂:RE³⁺ nanocrystals within mesoporous silica nanoparticles, a precursor solution should be prepared beforehand. Ca(OAc)₂, Yb(OAc)₃, and Ho(OAc)₃ were dissolved in deionized water to make a 0.5 M solution, and added with 12 ml TFA under stirring for 24 h. Subsequently, 300 mg MSN was added into 20 ml precursor solution and immersed for 24 h under stirring at 35°C. The particles were then centrifugated at 8000 rpm for 5 min, and stood for 4 h at room temperature. After being air-dried at 80°C overnights, the particles were further calcined at 600°C for 3 h.

Conjugation of CaF₂: Yb/Ho @MSN with probe oligonucleotides

100 mg CaF₂: Yb/Ho @MSN was dispersed in 100 ml ethanol under sonication for 10 min, and then 3 ml APTES was added under stirring at 50°C for 24 h. The samples were collected by centrifugation and dried at 80°C. 100 μ l ssDNA probe was dissolved in 5 ml PBS solution. 0.8 ml EDC (2 mg/ml) and 0.4 ml NHS (2 mg/ml) were added and the reaction was allowed to react for 30 min at 37°C under shaking. 10 mg CaF₂: Yb/Ho @MSN which modified with amino group was then added, and the reaction was incubated for 12 h. After reaction, the products were centrifuged, washed with PBS and then resuspended in 5 ml PBS solution.

Preparation of smart substrate

The precursor solution for electrospinning was prepared by a sol-gel method. In a typical process, 15 wt% TPU dissolved in 10 ml DMF and 10 ml THF, all the stuff had been stirring for 3 h at room temperature until got homogeneous spinnable precursor sols. For the electrospinning procedure, the electrospinning sol was fed into the conducting nozzle (2.5 mm ID) using an infusion pump (KDS-100, KD Scientific, USA) at a constant flow rate of 1 mL/h. The distance between the needle tip and the grounded aluminum foil was 12.5 cm. And the voltage was set to be 6 kV (PS/FC30P04.0-22, Glassman High voltage Inc., USA). The as-spun fibers were dried at 37 °C for 12 h.

The TPU fiber substrate was cut into the area of 0.8 cm x 0.5 cm small pieces, and added in 2 mg/ml GO solution under magnetic stirring for 24h at room temperature. The samples were dried at 37 °C. These small pieces were added in the solution which contained CaF₂: Yb/Ho @MSN-ssDNA probe as-prepared under gentle shaking for 12h at 37 °C. Finally, the substrate was washed with PBS solution for 3 times.

Hybridization assay and PL measurements

The smart substrates were incubated with different concentration of target analog miRNA at 37°C under shaking for 1 h in PBS and 10 vol% fetal bovine serum, and then washed with PBS solution gently. Photoluminescence spectrum measurements were done all these smart substrates at 980 nm excitation.

	Surface area [m²g ⁻¹]	Pore volume [cm³g⁻¹]	Pore size [nm]
MSN	747	1.54	8.27
CaF₂:Yb/Ho@MSN	454	0.98	8.63

Table S1. Structural parameters of nanoparticles before and after CaF₂: Yb/Ho crystals combination



Fig. S1 Elemental mapping images of an ultrathin section through part of CaF₂: Yb/Ho@MSN.



Fig. S2 TEM image of GO film.



Fig. S3 Optical photograph of TPU fiber membrane [A] and TPU@GO membrane [B]



Fig. S4 FTIR spectra of CaF₂: Yb/Ho@MSN and CaF₂: Yb/Ho@MSN modified with APTES.



Fig. S5 UV-vis absorption of CaF₂: Yb/Ho@MSN modified with NH₂ and DNA probe.



Fig. S6 ζ-potential of CaF₂: Yb/Ho@MSN, CaF₂: Yb/Ho@MSN modified with-NH₂ and DNA probe.



Fig. S7 Representation of the maximum fluorescence relative intensity of the smart membrane measured at (a) 540nm and (b) 647nm as a function of the target miR-21 concentration.



Fig. S8 (a) UCL spectra of the smart membrane after treating with solutions with different miR-21 concentration in 10 vol% fetal bovine serum, and the relationship between the relative intensity of UCL, at (b) 540 nm and (c) 647 nm, and miR-21 concentration at a certain range.



Fig. S9 Specific detection of miR-21, miR-21 with single base mismatch, miR-21 with three base mismatch and miR-195. (a) UCL spectra of the smart membrane after treating with different samples at 1 μ M. (b) The relationship between the relative intensity of UCL and samples at 647 nm.