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Highly-selective recognition of latent fingermarks by La -sensitized

Ce nanocomposite via electrostatic binding

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Experimental section

Materials:

Terbium (III) oxide (CeO₂) and lanthanum (III) oxide (La₂O₃), at 99.99% purity, were purchased from Shanghai Yuelong Rare Earth New Materials Co., Ltd., China. sulfosalicylic acid (SSA, AR), 1,10-phenanthroline (Phen, AR), dimethyl sulfoxide (DMSO, AR), absolute ethyl alcohol (EA, AR), acetone (AR), sodium hydroxide (NaOH, AR), concentrated hydrochloric acid (HCl, AR) and potassium bromide (KBr, AR) were provided by Sinopharm Chemical Reagent Co., Ltd., China. Supercoiled pBR322 plasmid DNA (3 kb) was purchased from Shanghai Sangon Biotech Co., Ltd. All the chemicals were used without further purification, and ultrapure millipore water (18.2 M Ω cm) was prepared throughout all the experiments.

Synthesis:

The synthesis procedure for Ce_xLa_{1-x} (SSA)₃Phen complexes was carried out by an optimized route based on our previous work. Different calculated weights of CeO₂ and La₂O₃ (Ce_xLa_{1-x} (SSA)₃Phen, (x=1, 0.875, 0.75, 0.5, 0.25, 0.125, 0)) were dissolved in excess concentrated HCl and were evaporated to near dryness at 100 °C for 11 h. Then, 80 mL of ethanol was added to the crystal with stirring after cooling. A homogeneous solution of Ce_xLa_{1-x}Cl₃ in ethanol was then obtained. SSA (12 mmol) and Phen (4 mmol) were dissolved in 30 mL 95% ethanol in a three-necked flask. The mixture solution was adjusted to pH 6.5 with NaOH (6 mmol). Then, 40 mL of Ce_xLa_{1-x}Cl₃ solution (4 mmol) was slowly dropped into the previous solution with constant stirring at 60 °C for 30 minutes. The mixture was quickly transferred to a polytetrafluoroethylene reactor and incubated at 140 °C for 4 h and was then precipitated, filtered, washed with ethanol three times, and dried in a vacuum at 60 °C to dryness. The nanopowder was collected, ground, and stored away from light for characterization and application.

Characterization of the Ce_xLa_{1-x}(SSA)₃Phen complexes

The morphology and size of the $Ce_xLa_{1-x}(SSA)_3$ Phen complexes were characterized by SEM (NanoLab 600i, FEI Corp.). A total of 10 mg sample was dissolved in 5 mL of ethanol, and the supernatant was placed on a special conductive adhesive that was dried in air. The SEM images were acquired at an electron acceleration voltage of 5 kV.

The contents of carbon, hydrogen, oxygen, and nitrogen in the complexes were characterized using an elemental analyzer (PE2400, Perkin Elmer, USA). The content of RE(III) was determined from ethylenediaminetetraacetic acid (EDTA) titration. The mass spectrometry (Bruker autoflex III, USA) was performed to analyze the final composition of complexes at positive ion mode using acetonitrile as the solvent. The structure and thermal behaviors of the complexes were studied with a thermogravimetric analyzer (TGA/DSC 1, Mettler, Switzerland) under nitrogen atmosphere with the rate of temperature rise at 10 °C/min from room temperature to 900 °C/min.

The state-solid photoluminescence excitation (PLE), photoluminescence (PL) spectra, quantum yield (QY) and luminescence decay curves were measured at room temperature on a fluorescence spectrometer (Hitachi F-700, Hitachi High-Technologies Corporation, Tokyo, Japan), with an integrating sphere, and a 450 W xenon lamp was used as the light source. UV-vis absorption spectra were obtained with a UV-vis spectrophotometer (UV-3600, Shimada UV, Japan).

Latent fingermark recognition

The latent fingermark samples were from the right thumb of the same male donor and were prepared by lightly rubbing the finger on the nose and forehead before touching the surface. Various substrates, including transparent glass, plastic sheets, aluminum alloys, ceramic tiles, painted wood, artificial leather and coated paper were used for depositing latent fingermarks. The powder dusting method was employed to develop the latent fingermarks using a soft feather brush. In contrast, some traditional

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developing methods, e.g., commercial phosphor, cyanoacrylate fuming methodology, and magnetic powder were chosen as controls. The fingermark images were detected under day-light or 312 nm irradiation detection, and were then recorded with a digital camera (EDS 6D, Canon, Japan). More ridge details were captured using a digital microscope (VHX-6000, Keyence, Japan). Finally, an automatic fingermark identification system (AFIS) was applied for the evaluation of the image quality of the developed fingermarks.

Interaction experiment between fingermark residues and Ce_xLa_{1-x}(SSA)₃Phen complexes

The transparent glass was cleaned up with ultrapure water and was dried and prepared as the substrate for the interaction experiment. Particularly, stained fingermarks using $Ce_{0.125}La_{0.875}$ (SSA)₃Phen complex as the developing powder were scraped from the glass and were collected as the sample for the FTIR and zeta potential analyses. Fourier transform infrared spectra (FTIR) were performed on a PerkinElmer Spectrum (PerkinElmer, USA) using KBr pellets. Zeta potentials of the $Ce_{0.125}La_{0.875}$ (SSA)₃Phen and $Ce_{0.125}La_{0.875}$ (SSA)₃Phen / fingermark residues were examined with a Malvern Nano-ZS, Model ZEN3600 (Malvern Instruments, Malvern, UK).

Interaction experiment between DNA and Ce_xLa_{1-x}(SSA)₃Phen complexes

An agarose gel electrophoresis system was used to analyze the interaction mode of our complex and DNA. The complexes/plasmid DNA were prepared freshly prior to use. The pure plasmid DNA was prepared with concentrations of 3.5×10^{-6} mol/L in ultrapure water, and Ce_{0.125}La_{0.875} (SSA)₃Phen complexes was added to the DNA solution with the concentrations of 5.0×10^{-6} , 10.0×10^{-6} , 15.0×10^{-6} , 20.0×10^{-6} mol/L, respectively. For each lane, a 20 µL volume of sample was loaded containing same concentration of DNA and different ratio of complexes, and additional ultrapure water are needed. The mixed samples were allowed to incubate at 37 °C for 20 min, and were then separated on a 1.5% agarose gel for 1 h at 120 V in Tris-acetate-EDTA (TAE) buffer, containing 1.5% agarose gel. Ethidium bromide (0.5 µg/mL) was then

used to stain gels, which were imaged under UV light.

For Circular dichroism (CD) spectra measurements (CHIRASCAN, Applied Photophysics, GB), moderate amounts of DNA and $Ce_{0.125}La_{0.875}$ (SSA)₃Phen were mixed in a 25 mL tube and diluted to 10 mL volume by ultrapure water. The pure plasmid DNA and mixed sample and with the concentration of 3.5×10^{-5} and 3.8×10^{-5} mol/L were respectively analyzed by CD spectrum using 1 cm quartz absorption cell, with a 220 \sim 320 nm scan range and ultrapure water as a blank.

V	weight (wt%)									
X values	Ln		С		Н		N		S	
	Found	Calc.	Found	Calc.	Found	Calc.	Found	Calc.	Found	Calc.
1	14.59	14.37	40.89	40.63	2.72	2.67	2.90	2.87	9.82	9.85
0.875	14.54	14.36	40.91	40.63	2.72	2.67	2.94	2.87	9.83	9.85
0.75	14.52	14.34	40.88	40.63	2.71	2.67	2.94	2.87	9.82	9.85
0.5	14.48	14.32	40.88	40.64	2.74	2.67	2.93	2.87	9.81	9.85
025	14.47	14.29	40.90	40.66	2.71	2.67	2.89	2.87	9.88	9.86
0.125	14.47	14.28	40.93	40.67	2.72	2.67	2.95	2.88	9.85	9.86
0	14.40	14.26	40.99	40.67	2.75	2.67	2.91	2.88	9.86	9.86

Table S1 Elemental percentages of the $Ce_xLa_{1-x}(SSA)_3$ Phen complexes.





Fig. S1 MS spectra of (a) Ce(SSA)₃Phen and (b) Ce_{0.125}La_{0.875} (SSA)₃Phen.

The mass spectrometry was performed to analysis the final composition of complexes. As shown in Fig. S1a, it could be observed that the m/z=218.659 matched well with the ligand SSA (M.W.=218.21), since the two H₂O combined would be removed when ionization. The m/z=180.680 matched well with the ligand Phen (M.W.=180.2). Based on formula of (m+z)/z, it an be deduced that the peak at 289.701 can be assigned to the Ce(SSA)₂ ((140.116+218.21×2+2)/2=289.268). The m/z=488.946 could be extrapolated to be the Ce(SSA)₃Phen ((974.946+2)/2=488.473). Similar results could be observed in Fig. S1b. For sample of Ce_{0.125}La_{0.875}(SSA)₃Phen, the m/z=180.691 and 218.659 matched well with the ligand Phen (M.W.=138.21), respectively. In addition, the m/z=138.586 matched well with the La (M.W.=138.9). Based on previous formula, the peak at 288.834 could be assigned to the Ce_{0.125}La_{0.875}(SSA)₂ ((140.116×0.125+138.9055×0.875+218.21×2+2)/2=288.738). We also extrapolated

that the peak at 487.990 would be the $Ce_{0.125}La_{0.875}(SSA)_3Phen$ ((973.887+2)/2=487.943).

Solid-state Quantum efficiency of Ce_xLa_{1-x}(SSA)₃Phen complexes

For luminescence, the quantum yield is an important parameter for a phosphor and determines its performance directly. The inner quantum yield is defined by:

$$\eta_{in} = \int L_S / (\int E_R - \int E_S)$$

where L_S is the emission intensity of the sample, and E_S and E_R are the spectra of the excitation light with and without the sample in the integrating sphere, respectively.

X values	Quantum yields (%)
1	38.8
0.875	9.5
0.75	9.9
0.5	12.7
0.25	44.8
0.125	51.2
0	20.1

Table S2 Solid-state quantum yields of $Ce_xLa_{1-x}(SSA)_3$ Phen complexes.



Fig. S2 Luminescence photographs of $Ce_xLa_{1-x}(SSA)_3$ Phen complexes (from left to right refers to different samples when x=0.875, 0.75, 0.5, 0, 1, 0.25 and 0.125, respectively).



Fig. S3 PL decay of Ce(SSA)₃Phen and Ce_{0.5}La_{0.5}(SSA)₃Phen at 297 K. a₀

The energy transfer efficiency (*E*p) between La^{3+} and Ce^{3+} cations is estimated based on the equation $Ep = 1 - \tau_{DA}/\tau_D$, where τ_{DA} and τ_D are the lifetime values in the presence and absence of La^{3+} cation, which provides an *Ep* of 29.2%.



Fig. S4 Stained fingermarks on various surfaces: (a, a') transparent plastic sheet, (b, b') ceramic tile, (c, c') aluminum alloy, (d, d') coated paper, (e, e') artificial leather, and (f, f') painted wood in (a–f) bright field and then developed by (a'– f') $Ce_{0.125}La_{0.875}$ (SSA)₃Phen nanopowder by 312 nm UV light in a dark field.



Fig. S5 EDS analysis of a developed fingermark on transparent glass by $Ce_{0.125}La_{0.875}$ (SSA)₃Phen nanopowder: (Spectrum 1) ridge, (Spectrum 2) furrow.



Fig. S6 3D images of a developed fingermark on PVC sheet by $Ce_{0.125}La_{0.875}$ (SSA)₃Phen nanopowder (a ~ d were captured at different angles of view).

Table S3 Wavenumbers of main peaks of FTIR spectra of two samples (cm⁻¹).

Samples	<i>0</i> 0-н	<i>D</i> N=С	<i>г</i> с-н	ŨС=О	<i>D</i> С=С	ØO→RE	ØN→RE
	(cm ⁻¹)	(cm ⁻¹)	(cm ⁻¹)	(cm ⁻¹)	(cm ⁻¹)	(cm ⁻¹)	(cm ⁻¹)
Ce0.125La0.875(SSA)3Phen	2224	1540	947 716	1660	1589,	591	464
	3224	1542	847,710	1660	1472		
Ce0.125La0.875(SSA)3Phen					1587		
reacted with fingermark	3215	1545	846,717	1661	1472	589	464
residues					14/5		



Fig. S7 FTIR spectrum of the pure and the $Ce_{0.125}La_{0.875}$ (SSA)₃Phen reacted with fingermark residues at the region of 450 ~ 1800 cm⁻¹.



Fig. S8 Electrophorogram for cleavage of plasmid DNA in the presence of complex. Lane 1: DNA only; Lane 2–5 in the different concentrations of complex: 5.0×10^{-6} mol/L, 10.0×10^{-6} mol/L, 15.0×10^{-6} mol/L, 20.0×10^{-6} mol/L.

Methods	Ce0.125La0.875	Commercial	superglue	magnetic ferric
Time	(SSA) ₃ Phen complex	phosphor	fuming	powder
1 d	98.6	97.3	97.3	95.9
7 d	97.3	93.2	86.3	79.5
15 d	95.9	84.9	75.3	63.0
24 d	91.7	75.3	64.4	47.9
32 d	90.4	65.8	58.9	41.1

Table S4 Comparation of second-level feature recognition rate of four developing method on

different aging time of latent fingermarks (unit: %)



Fig. S9 AFIS analysis of a fresh fingermark developed by $Ce_{0.125}La_{0.875}$ (SSA)₃Phen nanopowder.