# Organocatalytic Diastereoselective [3+2] Cyclization of MBH Carbonates with Dinucleophiles: Synthesis of Bicyclic Imidazoline Derivatives that Inhibit MDM2-p53 Interaction

Hong-Ping Zhu,<sup>a,c</sup> Ke Xie,<sup>c</sup> Xiang-Hong He,<sup>a</sup> Wei Huang,<sup>a</sup> Rong Zeng,<sup>c</sup> Yang Fan,<sup>c</sup> Cheng Peng,<sup>a</sup> Gu He,<sup>\*b</sup> and Bo Han<sup>\*a</sup>

<sup>a</sup>State Key Laboratory of Southwestern Chinese Medicine Resources, School of Pharmacy, Chengdu University of Traditional Chinese Medicine, Chengdu, 611137, China. E-mail: hanbo@cdutcm.edu.cn

<sup>b</sup>State Key Laboratory of Biotherapy and Cancer Center, West China Hospital, Sichuan University, Chengdu 610041, China. E-mail: hegu@scu.edu.cn

<sup>c</sup>Antibiotics Research and Re-evaluation Key Laboratory of Sichuan Province, Sichuan Industrial Institute of Antibiotics, Chengdu University, Chengdu 610052, China

### **Supplementary Information**

### **Table of Contents**

1.	General Information2
2.	Optimization of the annulation of HKA 1 with MBH carbonate 2a (Table S1)3
3.	Optimization of the Asymmetric cycloaddition of chiral 1 with 2a (Table S2)4
4.	General Procedure for the Preparation of the products 3 and 55
5.	Crystal Data and Structure Refinement for the Representative Product 5g20
6.	References and Notes
7.	NMR Spectra22
8.	Cell culture and cellular proliferation assay47
9.	HTRF based MDM2-p53 interaction assay47
10.	Molecular docking48
11.	Western blot analysis48
12.	Immunofluorescence assay48
13.	Superposition of compound 5c binding conformers and p53 substrate peptide
(Fig	gure S1)49
14.	Inhibition of MDM2 activity and of proliferation of HCT116 and MDA-MB231 cells
by	synthetic imidazoline derivatives (Table S3)50

### 1. General Information

### **General Procedures**

- All reactions were performed in oven-dried or flame-dried reaction vessels, modified Schlenk flasks, or round-bottom flasks. The flasks were fitted with Teflon screw caps and reactions were conducted under an atmosphere of argon if needed. Gas-tight syringes with stainless steel needles were used to transfer air- and moisture-sensitive liquids. All moisture and/or air sensitive solid compounds were manipulated inside normal desiccators. Flash column chromatography was performed using silica gel (40–63 µm, 230–400 mesh).
- Analytical thin layer chromatography (TLC) was performed on silica gel 60 F<sub>254</sub> aluminum plates (Merck) containing a 254 nm fluorescent indicator. TLC plates were visualized by exposure to short wave ultraviolet light (254 nm) and I<sub>2</sub>.
- Organic solutions were concentrated at 30-50 °C on rotary evaporators at ~10 torr followed by drying on vacuum pump at ~1 torr. Reaction temperatures are reported as the temperature of the bath surrounding the vessel unless otherwise stated.

### <u>Materials</u>

• Commercial reagents and solvents were were purchased from Adamas-beta, Aldrich Chemical Co., Alfa Aesar, Macklin and Energy Chemical and used as received with the following exceptions: THF, Et<sub>2</sub>O and toluene were purified by refluxing over Na-benzophenone under positive argon pressure followed by distillation.<sup>1</sup> The HKA 1<sup>2</sup> and MBH carbonates 2<sup>3</sup> were prepared according to literature procedure.

#### **Instrumentation**

- Proton nuclear magnetic resonance (<sup>1</sup>H NMR) spectra were recorded with JEOL-600M. Proton chemical shifts are reported in parts per million (δ scale), and are referenced using residual protium in the NMR solvent (CDCl<sub>3</sub>: δ 7.26 (CHCl<sub>3</sub>)). Data are reported as follows: chemical shift [multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br s = broad singlet), coupling constant(s) (Hz), integration].
- Carbon-13 nuclear magnetic resonance (<sup>13</sup>C NMR) spectra were recorded with JEOL 150 MHz spectrometers. Carbon chemical shifts are reported in parts per million ( $\delta$  scale), and are referenced using the carbon resonances of the solvent ( $\delta$  77.0 (CHCl<sub>3</sub>)). Data are reported as follows: chemical shift [multiplicity (if not singlet), assignment (C<sub>q</sub> = fully substituted carbon)].
- High resolution mass spectra (HRMS) were recorded on a Waters SYNAPT G2 using an electrospray (ESI) ionization source.
- Melting points were recorded on WRX-X-4A melting point apparatus.

#### 2. Optimization of the annulation of HKA 1 with MBH carbonate 2a

	$\begin{bmatrix} H \\ N \\ N \\ H \\ H \\ 1a \end{bmatrix} + H$	OBoc Ph CO <sub>2</sub> Me 2a	$\xrightarrow{\text{Lewis base}}_{\text{solvent, rt, 6 h}} \xrightarrow{O_2N}_{N} \xrightarrow{H}_{N}$	$MeO_2C Ph 4a$
entry	catalyst	solvent	yield of $3a/4$ (%) <sup>b</sup>	dr of $3a^c$
1	DABCO	DCM	40/53	14:1
2	DABCO	toluene	<5/12	/
3	DABCO	MeCN	63/<5	12:1
4	DABCO	THF	<5/36	/
5	DMAP	MeCN	<5/<5	
6	PPh <sub>3</sub>	MeCN	<5/<5	
$7^d$	DABCO	DCM	89/<5	14:1
$8^e$	DABCO	DCM	36/49	13:1
<b>9</b> <sup>f</sup>	DABCO	DCM	mess	/
10 <sup>g</sup>	DABCO	DCM	mess	/

**Table S1.** Optimization of the reaction of **1** with  $2a^a$ 

<sup>*a*</sup> Unless noted otherwise, the reactions were carried out with **1a** (0.12 mmol), **2a** (0.1 mmol) and Lewis base (20 mol %) in solvent (1 mL) at room temperature for 6 h. <sup>*b*</sup> Isolated yield. <sup>*c*</sup> *d.r.* was determined by <sup>1</sup>H-NMR analysis of the crude reaction mixture. <sup>*d*</sup> K<sub>2</sub>CO<sub>3</sub>(1 equiv.) was added. <sup>*e*</sup> DIPEA (1 equiv.) was added. <sup>*f*</sup> DBU (1 equiv.) was added.

Initially, heterocyclic ketene aminal (HKA) **1**a and the readily available Morita-Baylis-Hillman (MBH) carbonate 2a were chosen as model substrates to investigate the feasibility of desired [3+3] cyclization conditions that should deliver **3a** as the target product. As shown in Table S1, solvents were screened in the presence of DABCO as a Lewis base catalyst, (entries 1-4) and DCM was found able to afford the desired adduct 3a, along with an unclosed intermediate 4 in 53% yield (entry 1). This intermediate was not detected when MeCN as the solvent, but afford moderate yield (entry 3). Then, we screened other Lewis base catalysts, such as DMAP and PPh<sub>3</sub>; however, no desired products were observed (entries 5 and 6). To improve the yield of this reaction, we envisioned that the intermediate 4 in the reaction in DCM might convert to the desired product 3 by the addition of a Brønsted base. Fortunately, after screening several Brønsted bases in DCM, we found K2CO3 could promote the cyclization to deliver 3a in excellent yield without the observation of 4 (entries 7–10).

#### 3. Optimization of the Asymmetric cycloaddition of HKA chiral 1 with 2a

	$\begin{array}{c} Ph_{M_{H_{H_{H_{H_{H_{H_{H_{H_{H_{H_{H_{H_{H_$	Boc CO <sub>2</sub> Me 2a CO <sub>2</sub> Me CO <sub>2</sub> Me CO <sub>2</sub> Me CO <sub>2</sub> Me CO <sub>2</sub> Me CO <sub>2</sub> Me	$\rightarrow$	Ph
entry	catalyst	solvent	yield $(\%)^b$	dr <sup>c</sup>
1	DABCO	toluene	52	10:1
2	DABCO	DCM	90	17:1
3	DABCO	MeCN	33	13:1
4	DABCO	THF	75	17:1
5	DABCO	DMF	18	/
6	DMAP	DCM	<5	/
7	PPh <sub>3</sub>	DCM	<5	/
$8^d$	DABCO	DCM	72	16:1
9 <sup>e</sup>	DABCO	DCM	33	16:1

н

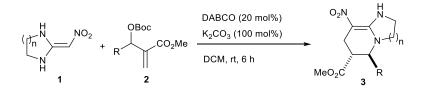
**Table S2.** Optimization of the reaction of 1c with  $5a^a$ 

<sup>*a*</sup> Unless noted otherwise, the reactions were carried out with **chiral 1** (0.10 mmol), **2a** (0.12 mmol), catalyst (20 mol %) and in solvent (1.0 mL) at room temperature for 2 h. <sup>*b*</sup> Isolated yield. <sup>*c*</sup> Dr was determined by <sup>1</sup>H-NMR analysis of the crude reaction mixture. <sup>*d*</sup> 10 mol % of DABCO was used. <sup>*e*</sup> at 0 °C.

To prove the feasibility of our proposal, readily available HKA chiral 1 and MBH carbonate 2a were chosen as substrates. The reaction was conducting in toluene in the presence of 20 mol% DABCO, providing desired chiral bicyclic imidazoline derivative 5a in 52% yield with 10:1 *d.r.* (entry 1). Different solvents and Lewis bases were screened to improve the yield and diastereoselectivity. As shown in Table S2, dichloromethane was the best solvent for this [3+3] cycloaddition, delivering adduct 5a in 90% yield with 17:1 *d.r.*, while other solvents gave inferior results (entry 2-5). Then, we screened other Lewis base catalysts, such as DMAP and PPh<sub>3</sub>; however, no desired products were observed (entries 6 and 7). Reducing the catalyst loading or lowering the reaction temperature had negative effects on the yields (entries 8 and 9). Thus, the optimal conditions were identified as 20 mol% DABCO in CH<sub>2</sub>Cl<sub>2</sub> at room temperature.

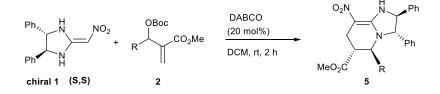
#### 4. General Procedure for the Preparation of the products 3 and 5

General procedure A for the synthesis of products 3



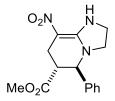
A glass tube was charged with heterocyclic ketene aminal **1** (0.1 mmol), MBH carbonate **2** (0.12 mmol), DABCO (0.02 mmol) and  $K_2CO_3$  (0.1 mmol) in DCM (1 mL). The mixture was stirred at room temperature for 6 hour. Then the mixture was directly purified by column chromatography on silica gel (DCM/ethyl acetate = 10/1 to 3/1) to afford the corresponding product **3**.

General procedure B for the asymmetric synthesis of products 5



A glass tube was charged with chiral heterocyclic ketene aminal **chiral 1**(*S*, *S*) (0.1 mmol), MBH carbonate **2** (0.12 mmol) and DABCO (0.02 mmol) in DCM (1 mL). The mixture was stirred at room temperature for 2 hour. Then the mixture was directly purified by column chromatography on silica gel (petroleum ether/ethyl acetate = 5/1 to 2/1) to afford the corresponding product **5**.

#### methyl 8-nitro-5-phenyl-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6-carboxylate 3a



Prepared according to the general procedure A to afford **3a** (26.3 mg) in 87% yield as white solid. The diastereomeric ratio was determined to be 14:1 by crude <sup>1</sup>H NMR analysis; the relative configuration was determined by NOEDS analysis; m.p. 183 - 188 °C.

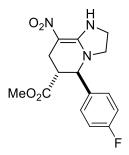
*NMR and HRMS data for the product* **3a**:

<sup>1</sup>**H NMR (600 MHz, CDCl**<sub>3</sub>)  $\delta$  (ppm): 8.69 (brs, 1H), 7.41 – 7.34 (m, 3H), 7.19 (d, *J* = 7.2 Hz, 2H), 4.74 (d, *J* = 4.8 Hz, 1H), 3.86 – 3.75 (m, 2H), 3.65 (s, 3H), 3.61 – 3.54 (m, 2H), 3.25 (dd, *J* = 15.6 Hz, *J* = 5.4 Hz, 1H), 3.01 (dd, *J* = 16.8 Hz, *J* = 5.4 Hz, 1H), 2.83 (dd, *J* = 15.6 Hz, *J* = 5.4 Hz, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 172.2, 157.0, 137.2, 129.2, 128.7, 126.5, 102.1, 59.2, 52.5, 48.2, 45.4, 42.3, 22.5.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{15}H_{17}N_3O_4Na^+$ : 326.1111, found: 326.1115.

# methyl 5-(4-fluorophenyl)-8-nitro-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6carboxylate 3b



Prepared according to the general procedure A to afford **3b** (23.4 mg) in 73% yield as white solid. The diastereomeric ratio was determined to be 10:1 by crude <sup>1</sup>H NMR analysis; m.p. 192 – 194 °C.

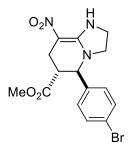
*NMR and HRMS data for the product* **3b**:

<sup>1</sup>**H NMR (600 MHz, CDCl**<sub>3</sub>)  $\delta$  (ppm): 8.68 (brs, 1H), 7.20 – 7.18 (m, 2H), 7.11 – 7.07 (m, 2H), 4.70 (d, J = 5.4 Hz, 1H), 3.86 – 3.81 (m, 1H), 3.77 (dd, J = 19.8 Hz, J = 9.6 Hz, 1H), 3.64 (s, 3H), 3.56 – 3.50 (m, 2H), 3.21 (dd, J = 15.6 Hz, J = 6.0 Hz, 1H), 2.98 (dd, J = 11.4 Hz, J = 5.4 Hz, 1H), 2.87 (dd, J = 15.6 Hz, J = 6.0 Hz, 1H).

<sup>13</sup>**C NMR (150 MHz, CDCl**<sub>3</sub>)  $\delta$  (ppm): 172.1, 162.7 (d,  $J_{C-F} = 246.9$  Hz), 156.9, 133.0, 128.4 (d,  $J_{C-F} = 7.2$  Hz), 116.2 (d,  $J_{C-F} = 21.6$  Hz), 102.0, 58.8, 52.5, 48.2, 45.6, 42.3, 22.8.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{15}H_{16}FN_3O_4Na^+$ : 344.1017, found: 344.1014.

## methyl 5-(4-bromophenyl)-8-nitro-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6carboxylate 3c



Prepared according to the general procedure A to afford 3c (29.0 mg) in 76% yield as white solid. The diastereometric ratio was determined to be 12:1 by crude <sup>1</sup>H NMR analysis; m.p. 105

– 109 °C.

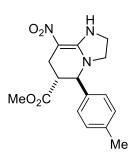
NMR and HRMS data for the product **3c**:

<sup>1</sup>**H NMR** (**600 MHz**, **CDCl**<sub>3</sub>)  $\delta$  (ppm): 8.68 (brs, 1H), 7.53 (d, J = 8.4 Hz, 2H), 7.10 (d, J = 8.4 Hz, 2H), 4.69 (d, J = 4.8 Hz, 1H), 3.86 – 3.82 (m, 1H), 3.78 (dd, J = 19.8 Hz, J = 9.6 Hz, 1H), 3.65 (s, 3H), 3.57 – 3.50 (m, 2H), 3.22 (dd, J = 16.2 Hz, J = 6.0 Hz, 1H), 2.98 (dd, J = 17.4 Hz, J = 5.4 Hz, 1H), 2.84 (dd, J = 16.2 Hz, J = 6.0 Hz, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 172.0, 156.8, 136.3, 132.3, 128.3, 122.7, 101.9, 58.8, 52.5, 48.2, 45.3, 42.3, 22.7.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{15}H_{16}^{79}BrN_3O_4Na^+$ : 404.0222, found: 404.0218; calculated for  $C_{15}H_{16}^{81}BrN_3O_4Na^+$ : 406.0201, found: 406.0199.

#### methyl 8-nitro-5-(p-tolyl)-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6-carboxylate 3d



Prepared according to the general procedure A to afford **3d** (28.2 mg) in 89% yield as white solid. The diastereomeric ratio was determined to be 4:1 by crude <sup>1</sup>H NMR analysis; m.p. 93 – 98 °C.

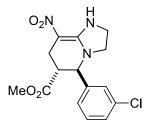
*NMR and HRMS data for the product* **3d**:

<sup>1</sup>**H NMR (600 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm): 8.70 (brs, 1H), 7.19 (d, *J* = 7.8 Hz, 2H), 7.07 (d, *J* = 7.8 Hz, 2H), 4.69 (d, *J* = 4.8 Hz, 1H), 3.82 – 3.72 (m, 2H), 3.65 (s, 3H), 3.57 – 3.53 (m, 2H), 3.24 (dd, *J* = 16.2 Hz, *J* = 5.4 Hz, 1H), 2.98 (dd, *J* = 10.8 Hz, *J* = 5.4 Hz, 1H), 2.85 (dd, *J* = 16.2 Hz, *J* = 5.4 Hz, 1H), 2.36 (s, 3H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 172.3, 157.0, 138.7, 134.1, 129.9, 126.5, 102.1, 59.0, 52.5, 48.2, 45.5, 42.3, 22.6, 21.1.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{16}H_{19}N_3O_4Na^+$ : 340.1268, found: 340.1270.

<u>methyl 5-(3-chlorophenyl)-8-nitro-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6-</u> <u>carboxylate 3e</u>



Prepared according to the general procedure A to afford 3e (32.0 mg) in 95% yield as white solid. The diastereomeric ratio was determined to be 14:1 by crude <sup>1</sup>H NMR analysis; m.p. 164 – 167 °C.

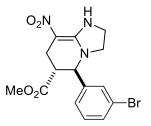
NMR and HRMS data for the product **3e**:

<sup>1</sup>**H NMR (600 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm): 8.68 (brs, 1H), 7.35 – 7.34 (m, 2H), 7.19 (s, 1H), 7.10 – 7.08 (m, 1H), 4.73 (d, J = 4.2 Hz, 1H), 3.87 – 3.78 (m, 2H), 3.67 (s, 3H), 3.62 – 3.56 (m, 2H), 3.28 (dd, J = 16.2 Hz, J = 5.4 Hz, 1H), 2.98 (dd, J = 10.8 Hz, J = 5.4 Hz, 1H), 2.84 (dd, J = 16.2 Hz, J = 5.4 Hz, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 172.0, 156.7, 139.5, 135.2, 130.5, 129.0, 126.7, 124.7, 101.8, 58.8, 52.6, 48.3, 45.2, 42.3, 22.3.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{15}H_{16}^{35}ClN_3O_4Na^+$ : 360.0722, found: 360.0724; calculated for  $C_{15}H_{16}^{37}ClN_3O_4Na^+$ : 362.0698, found: 362.0693.

# methyl 5-(3-bromophenyl)-8-nitro-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6carboxylate 3f



Prepared according to the general procedure A to afford **3f** (30.9 mg) in 81% yield as white solid. The diastereomeric ratio was determined to be 8:1 by crude <sup>1</sup>H NMR analysis; m.p. 113 – 117 °C.

NMR and HRMS data for the product **3f**:

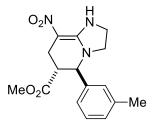
<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 8.68 (brs, 1H), 7.50 (d, J = 8.4 Hz, 1H), 7.34 (s, 1H), 7.28 (t, J = 7.8 Hz, 1H), 7.14 (d, J = 7.8 Hz, 1H), 4.72 (d, J = 4.2 Hz, 1H), 3.88 – 3.78 (m, 2H), 3.67 (s, 3H), 3.62 – 3.55 (m, 2H), 3.28 (dd, J = 16.2 Hz, J = 5.4 Hz, 1H), 2.99 (dd, J = 10.2 Hz, J = 5.4 Hz, 1H), 2.81 (dd, J = 16.2 Hz, J = 5.4 Hz, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 172.0, 156.8, 139.8, 131.9, 130.8, 129.6, 125.1, 123.3, 101.9, 58.8, 52.6, 48.4, 45.3, 42.4, 22.4.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{15}H_{16}^{79}BrN_3O_4Na^+$ : 404.0222, found:

404.0218; calculated for C<sub>15</sub>H<sub>16</sub><sup>81</sup>BrN<sub>3</sub>O<sub>4</sub>Na<sup>+</sup>: 406.0201, found: 406.0195.

#### methyl 8-nitro-5-(m-tolyl)-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6-carboxylate 3g



Prepared according to the general procedure A to afford **3g** (24.4 mg) in 77% yield as white solid. The diastereomeric ratio was determined to be 4:1 by crude <sup>1</sup>H NMR analysis; m.p. 94 – 98 °C.

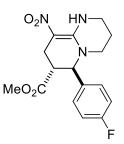
*NMR and HRMS data for the product* **3g**:

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 8.68 (brs, 1H), 7.28 (t, *J* = 7.8 Hz, 1H), 7.16 (d, *J* = 7.2 Hz, 1H), 6.98 (s, 1H), 6.97 (d, *J* = 7.8 Hz, 1H), 4.72 (d, *J* = 4.8 Hz, 1H), 3.84 – 3.76 (m, 2H), 3.66 (s, 3H), 3.60 – 3.57 (m, 2H), 3.27 (dd, *J* = 16.2 Hz, *J* = 5.4 Hz, 1H), 3.00 (dd, *J* = 10.2 Hz, *J* = 5.4 Hz, 1H), 2.82 (dd, *J* = 15.6 Hz, *J* = 5.4 Hz, 1H), 2.37 (s, 3H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 172.3, 156.9, 139.0, 137.2, 129.4, 129.0, 127.0, 123.5, 102.0, 59.1, 52.4, 48.2, 45.3, 42.3, 22.4, 21.4.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{16}H_{19}N_3O_4Na^+$ : 340.1268, found: 340.1269.

# methyl 6-(4-fluorophenyl)-9-nitro-2,3,4,6,7,8-hexahydro-1H-pyrido[1,2-a]pyrimidine-7carboxylate 3h



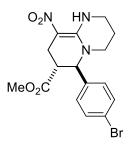
Prepared according to the general procedure A to afford **3h** (25.0 mg) in 79% yield as white solid. The diastereomeric ratio was determined to be 13:1 by crude <sup>1</sup>H NMR analysis; m.p. 169 – 172 °C.

NMR and HRMS data for the product **3h**:

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 11.79 (brs, 1H), 7.16 – 7.07 (m, 4H), 4.84 – 4.82 (m, 1H), 3.73 (s, 3H), 3.56 – 3.47 (m, 4H), 3.32 – 3.28 (m, 1H), 2.92 – 2.90 (m, 1H), 2.59 (dd, J = 16.8 Hz, J = 6.0 Hz, 1H), 2.10 – 2.06 (m, 1H), 2.00 – 1.95 (m, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 172.3, 162.6 (d,  $J_{C-F} = 247.1$  Hz), 152.8, 133.3, 127.7 (d,  $J_{C-F} = 8.7$  Hz), 116.3 (d,  $J_{C-F} = 21.6$  Hz), 103.6, 63.1, 52.7, 47.0, 43.1, 38.3, 21.8, 20.1. HRMS (ESI-TOF) m/z: [M + Na]<sup>+</sup> calculated for C<sub>16</sub>H<sub>18</sub>FN<sub>3</sub>O<sub>4</sub>Na<sup>+</sup>: 358.1174, found: 358.1174.

methyl 6-(4-bromophenyl)-9-nitro-2,3,4,6,7,8-hexahydro-1H-pyrido[1,2-a]pyrimidine-7carboxylate 3i



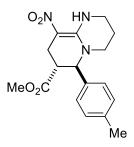
Prepared according to the general procedure A to afford **3i** (32.8 mg) in 83% yield as white solid. The diastereomeric ratio was determined to be 9:1 by crude <sup>1</sup>H NMR analysis; m.p. 105 – 108 °C.

NMR and HRMS data for the product **3i**:

<sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ (ppm): 11.77 (brs, 1H), 7.53 (d, J = 7.8 Hz, 2H), 7.04 (d, J = 9.6 Hz, 2H), 4.82 – 4.81 (m, 1H), 3.74 (s, 3H), 3.57 – 3.47 (m, 4H), 3.31 – 3.24 (m, 1H), 2.93 – 2.90 (m, 1H), 2.57 (dd, J = 16.8 Hz, J = 6.0 Hz, 1H), 2.10 – 2.06 (m, 1H), 2.00 – 1.96 (m, 1H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm):172.2, 152.7, 136.6, 132.4, 127.7, 122.5, 103.6, 63.2, 52.7, 47.1, 42.9, 38.3, 21.8, 20.1.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{16}H_{18}^{79}BrN_3O_4Na^+$ : 418.0373, found: 418.0373; calculated for  $C_{16}H_{18}^{81}BrN_3O_4Na^+$ : 420.0358, found: 420.0356.

#### methyl 8-nitro-5-(p-tolyl)-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6-carboxylate 3j



Prepared according to the general procedure A to afford **3j** (28.5 mg) in 86% yield as white solid. The diastereomeric ratio was determined to be 3:1 by crude <sup>1</sup>H NMR analysis; m.p. 98 - 102 °C.

*NMR and HRMS data for the product* **3***j*:

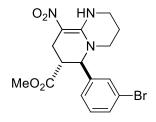
<sup>1</sup>**H NMR (600 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm): 11.80 (brs, 1H), 7.19 (d, J = 8.4 Hz, 2H), 7.02 (d, J = 8.4 Hz, 2H), 4.80 (brs, 1H), 3.72 (s, 3H), 3.56 – 3.45 (m, 4H), 3.38 – 3.28 (m, 2H), 2.93 – 2.90

(m, 1H), 2.60 (dd, *J* = 16.2 Hz, *J* = 5.4 Hz, 1H), 2.35 (s, 3H), 2.08 – 2.05 (m, 1H), 1.99 – 1.92 (m, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 172.5, 152.8, 138.4, 134.4, 129.9, 125.8, 103.9, 63.5, 52.5, 47.0, 43.2, 38.3, 21.9, 21.0, 20.0.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{17}H_{21}N_3O_4Na^+$ : 354.1424, found: 354.1423.

### <u>methyl 6-(3-bromophenyl)-9-nitro-2,3,4,6,7,8-hexahydro-1H-pyrido[1,2-a]pyrimidine-7-</u> <u>carboxylate 3k</u>



Prepared according to the general procedure A to afford **3k** (29.2 mg) in 74% yield as white solid. The diastereomeric ratio was determined to be 12:1 by crude <sup>1</sup>H NMR analysis; m.p. 83 - 87 °C.

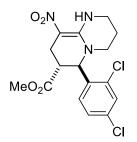
*NMR and HRMS data for the product* **3k**:

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 11.79 (brs, 1H), 7.50 (d, J = 7.8 Hz, 2H), 7.29 – 7.28 (m, 2H), 7.08 (d, J = 7.8 Hz, 2H), 4.82 – 4.81 (m, 1H), 3.74 (s, 3H), 3.59 – 3.48 (m, 4H), 3.33 – 3.29 (m, 1H), 2.96 – 2.94 (m, 1H), 2.57 (dd, J = 17.4 Hz, J = 6.0 Hz, 1H), 2.11 – 2.07 (m, 1H), 2.03 – 1.98 (m, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm):172.2, 152.7, 140.0, 131.8, 130.9, 129.1, 124.4, 126.4, 103.5, 63.2, 52.7, 47.2, 43.0, 38.3, 21.8, 20.1.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{16}H_{18}^{79}BrN_3O_4Na^+$ : 418.0373, found: 418.0374; calculated for  $C_{16}H_{18}^{81}BrN_3O_4Na^+$ : 420.0358, found: 420.0363.

### methyl 6-(2,4-dichlorophenyl)-9-nitro-2,3,4,6,7,8-hexahydro-1H-pyrido[1,2-a]pyrimidine-7-carboxylate 31



Prepared according to the general procedure A to afford **31** (26.9 mg) in 70% yield as white solid. The diastereomeric ratio was determined to be 9:1 by crude <sup>1</sup>H NMR analysis; m.p. 217

– 219 °C.

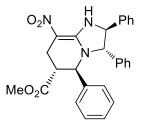
NMR and HRMS data for the product **31**:

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 11.78 (brs, 1H), 7.47 (s, 1H), 7.30 (d, J = 8.4 Hz, 1H), 7.02 (d, J = 8.4 Hz, 1H), 5.19 – 5.18 (m, 1H), 3.76 (s, 3H), 3.59 – 3.49 (m, 4H), 3.27 – 3.22 (m, 1H), 3.07 – 3.05 (m, 1H), 2.46 (dd, J = 17.4 Hz, J = 6.0 Hz, 1H), 2.12 – 2.08 (m, 1H), 2.00 – 1.94 (m, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 172.0, 153.0, 135.2, 133.3, 133.0, 130.4, 127.9, 127.9, 103.2, 60.8, 52.8, 47.3, 39.9, 38.3, 21.8, 20.1.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{16}H_{17}Cl_2N_3O_4Na^+$ : 408.0488, found: 408.0490.

## (2S,3S,5R,6R)-methyl 8-nitro-2,3,5-triphenyl-1,2,3,5,6,7-hexahydroimidazo[1,2-a] pyridine-6-carboxylate 5a



Prepared according to the general procedure B to afford **5a** (40.9 mg) in 90% yield as white solid. The diastereomeric ratio was determined to be 17:1 by crude <sup>1</sup>H NMR analysis; m.p. 96 -100 °C.

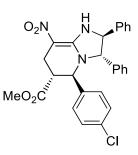
NMR and HRMS data for the product **5a**:

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 9.12 (brs, 1H), 7.44 – 7.38 (m, 3H), 7.36 – 7.31 (m, 6H), 7.22 – 7.19 (m, 2H), 7.11 (d, J = 6.6 Hz, 2H), 7.02 – 7.01 (m, 2H), 4.97 (d, J = 9.0 Hz, 1H), 4.56 (d, J = 3.6 Hz, 1H), 4.41 (d, J = 9.0 Hz, 1H), 3.69 (s, 3H), 3.41 (dd, J = 16.2 Hz, J = 3.6 Hz, 1H), 2.98 – 2.96 (m, 1H), 2.82 (dd, J = 16.8 Hz, J = 6.0 Hz, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm):171.7, 156.1, 137.9, 136.6, 135.5, 129.6, 129.5, 129.3, 129.2, 129.1, 128.9, 128.7, 127.9, 126.5, 126.3, 102.2, 71.9, 67.8, 56.4, 52.4, 44.6, 21.1.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for C<sub>27</sub>H<sub>25</sub>N<sub>3</sub>O<sub>4</sub>Na<sup>+</sup>: 478.1737, found: 478.1733.

### (2*S*,3*S*,5*R*,6*R*)-methyl 5-(4-chlorophenyl)-8-nitro-2,3-diphenyl-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6-carboxylate 5b

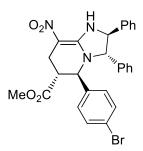


Prepared according to the general procedure B to afford **5b** (45.0 mg) in 92% yield as white solid. The diastereomeric ratio was determined to be 16:1 by crude <sup>1</sup>H NMR analysis; m.p. 205 -210 °C.

*NMR and HRMS data for the product* **5b**:

<sup>1</sup>**H NMR** (**600 MHz**, **CDCl**<sub>3</sub>) δ (ppm): 9.10 (brs, 1H), 7.43 – 7.39 (m, 3H), 7.37 – 7.35 (m, 3H), 7.31 (d, J = 8.4 Hz, 2H), 7.20 – 7.18 (m, 2H), 7.10 (d, J = 6.0 Hz, 2H), 6.96 (d, J = 8.4 Hz, 2H), 4.96 (d, J = 8.4 Hz, 1H), 4.53 (d, J = 3.6 Hz, 1H), 4.34 (d, J = 7.2 Hz, 1H), 3.68 (s, 3H), 3.39 (dd, J = 16.2 Hz, J = 3.6 Hz, 1H), 2.96 – 2.93 (m, 1H), 2.84 (dd, J = 16.8 Hz, J = 6.0 Hz, 1H). <sup>13</sup>**C NMR** (**150 MHz**, **CDCl**<sub>3</sub>) δ (ppm): 171.5, 155.9, 137.8, 135.3, 135.1, 134.7, 129.6, 129.5, 129.2, 129.2, 129.0, 128.0, 127.8, 126.2, 102.0, 71.8, 67.7, 56.0, 52.5, 44.6, 21.3. **HRMS** (**ESI-TOF**) m/z: [**M** + **Na**]<sup>+</sup> calculated for C<sub>27</sub>H<sub>24</sub><sup>35</sup>ClN<sub>3</sub>O<sub>4</sub>Na<sup>+</sup>: 512.1348, found: 512.1349; calculated for C<sub>27</sub>H<sub>24</sub><sup>37</sup>ClN<sub>3</sub>O<sub>4</sub>Na<sup>+</sup>: 514.1324, found: 514.1320.

# (2S,3S,5R,6R)-methyl 5-(4-bromophenyl)-8-nitro-2,3-diphenyl-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6-carboxylate 5c



Prepared according to the general procedure B to afford **5c** (50.1 mg) in 94% yield as white solid. The diastereomeric ratio was determined to be >19:1 by crude <sup>1</sup>H NMR analysis; m.p. 105 - 110 °C.

*NMR and HRMS data for the product* **5c**:

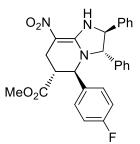
<sup>1</sup>**H NMR (600 MHz, CDCl**<sub>3</sub>) δ (ppm): 9.10 (brs, 1H), 7.47 – 7.38 (m, 5H), 7.36 – 7.32 (m, 3H), 7.20 – 7.18 (m, 2H), 7.10 (d, *J* = 6.0 Hz, 2H), 6.90 (d, *J* = 9.0 Hz, 2H), 4.96 (d, *J* = 8.4 Hz, 1H),

4.51 (d, *J* = 3.6 Hz, 1H), 4.34 (d, *J* = 8.4 Hz, 1H), 3.68 (s, 3H), 3.39 (dd, *J* = 16.2 Hz, *J* = 3.6 Hz, 1H), 2.96 – 2.93 (m, 1H), 2.84 (dd, *J* = 16.8 Hz, *J* = 5.4 Hz, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 171.5, 155.9, 137.8, 135.7, 135.3, 132.4, 129.6, 129.3, 129.2, 129.0, 128.3, 127.8, 126.2, 122.8, 102.0, 71.8, 67.7, 56.0, 52.5, 44.6, 21.3.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{27}H_{24}^{79}BrN_3O_4Na^+$ : 556.0842, found: 556.0844; calculated for  $C_{27}H_{24}^{81}BrN_3O_4Na^+$ : 558.0827, found: 558.0825.

### (2*S*,3*S*,5*R*,6*R*)-methyl 5-(4-fluorophenyl)-8-nitro-2,3-diphenyl-1,2,3,5,6,7-hexahydro imidazo[1,2-a]pyridine-6-carboxylate 5d



Prepared according to the general procedure B to afford **5d** (41.1 mg) in 87% yield as white solid. The diastereomeric ratio was determined to be 15:1 by crude <sup>1</sup>H NMR analysis; m.p. 236 -239 °C.

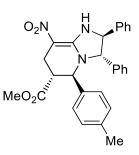
NMR and HRMS data for the product **5d**:

<sup>1</sup>**H NMR (600 MHz, CDCl**<sub>3</sub>)  $\delta$  (ppm): 9.10 (brs, 1H), 7.46 – 7.35 (m, 6H), 7.20 – 7.19 (m, 2H), 7.10 (d, J = 7.8 Hz, 2H), 7.05 – 6.97 (m, 4H), 4.96 (d, J = 7.8 Hz, 1H), 4.53 (d, J = 3.0 Hz, 1H), 4.34 (d, J = 8.4 Hz, 1H), 3.68 (s, 3H), 3.38 (dd, J = 16.2 Hz, J = 3.0 Hz, 1H), 2.96 – 2.85 (m, 2H).

<sup>13</sup>**C NMR (150 MHz, CDCl**<sub>3</sub>) δ (ppm): 171.6, 156.0, 137.9, 135.4, 132.3, 129.6, 129.2, 129.2, 129.0, 128.4, 128.4, 127.8, 126.2, 116.3 (*J*<sub>C-F</sub> = 21.6 Hz), 102.0, 71.8, 67.7, 56.0, 52.4, 44.8, 21.4.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for C<sub>27</sub>H<sub>24</sub>FN<sub>3</sub>O<sub>4</sub>Na<sup>+</sup>: 496.1643, found: 496.1644.

(2S,3S,5R,6R)-methyl 8-nitro-2,3-diphenyl-5-(p-tolyl)-1,2,3,5,6,7-hexahydroimidazo [1,2-a]pyridine-6-carboxylate 5e



Prepared according to the general procedure B to afford **5e** (38.0 mg) in 81% yield as white solid. The diastereomeric ratio was determined to be 16:1 by crude <sup>1</sup>H NMR analysis; m.p. 215 -218 °C.

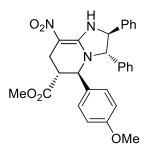
*NMR and HRMS data for the product* **5e**:

<sup>1</sup>**H NMR** (**600 MHz**, **CDCl**<sub>3</sub>)  $\delta$  (ppm): 9.12 (brs, 1H), 7.42 – 7.35 (m, 6H), 7.21 – 7.19 (m, 2H), 7.14 – 7.10 (m, 4H), 6.90 (d, J = 7.2 Hz, 2H), 4.95 (d, J = 9.0 Hz, 1H), 4.52 (d, J = 3.6 Hz, 1H), 4.40 (d, J = 9.0 Hz, 1H), 3.68 (s, 3H), 3.39 (dd, J = 16.2 Hz, J = 3.6 Hz, 1H), 2.96 – 2.93 (m, 1H), 2.83 (dd, J = 16.2 Hz, J = 6.0 Hz, 1H), 2.32 (s, 3H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 171.8, 156.1, 138.5, 138.0, 135.6, 133.5, 129.9, 129.4, 129.1, 129.1, 128.9, 127.8, 126.4, 126.3, 102.2, 71.8, 67.7, 56.2, 52.3, 44.7, 21.1, 21.0.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{28}H_{27}N_3O_4Na^+$ : 492.1894, found: 492.1899.

# (2S,3S,5R,6R)-methyl 5-(4-methoxyphenyl)-8-nitro-2,3-diphenyl-1,2,3,5,6,7-hexahydro imidazo[1,2-a]pyridine-6-carboxylate 5f



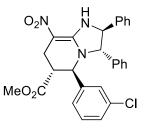
Prepared according to the general procedure B to afford **5f** (41.2 mg) in 85% yield as white solid. The diastereomeric ratio was determined to be 12:1 by crude <sup>1</sup>H NMR analysis; m.p. 100 -104 °C.

NMR and HRMS data for the product **5f**:

<sup>1</sup>**H NMR (600 MHz, CDCl**<sub>3</sub>) δ (ppm): 9.11 (brs, 1H), 7.43 – 7.33 (m, 6H), 7.20 – 7.19 (m, 2H), 7.11 – 7.09 (m, 2H), 6.93 (d, *J* = 9.0 Hz, 2H), 6.84 (d, *J* = 9.0 Hz, 2H), 4.94 (d, *J* = 8.4 Hz, 1H),

4.49 (d, J = 3.6 Hz, 1H), 4.38 (d, J = 9.0 Hz, 1H), 3.79 (s, 3H), 3.68 (s, 3H), 3.38 (dd, J = 16.2 Hz, J = 3.6 Hz, 1H), 2.96 – 2.93 (m, 1H), 2.89 (dd, J = 16.2 Hz, J = 6.0 Hz, 1H). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 171.8, 159.7, 156.1, 138.0, 135.7, 129.4, 129.2, 129.1, 128.9, 128.3, 127.8, 127.8, 126.3, 114.6, 102.2, 71.8, 67.7, 56.1, 55.3, 52.3, 44.9, 21.5. HRMS (ESI-TOF) m/z: [M + Na]<sup>+</sup> calculated for C<sub>28</sub>H<sub>27</sub>N<sub>3</sub>O<sub>5</sub>Na<sup>+</sup>: 508.1843, found: 508.1846.

### (2*S*,3*S*,5*R*,6*R*)-methyl 5-(3-chlorophenyl)-8-nitro-2,3-diphenyl-1,2,3,5,6,7-hexahydroimidazo[1,2-a]pyridine-6-carboxylate 5g



Prepared according to the general procedure B to afford **5g** (40.0 mg) in 90% yield as white solid. The diastereomeric ratio was determined to be 17:1 by crude <sup>1</sup>H NMR analysis; m.p. 115 – 120 °C.

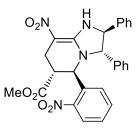
Notably, according to the same reaction condition, a 2.5mmol-scale reaction has been performed, which afford 5g (1.05 grams) in 86% yield as white solid.

*NMR and HRMS data for the product* **5g**:

<sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 9.11 (brs, 1H), 7.47 – 7.35 (m, 7H), 7.27 – 7.26 (m, 1H), 7.22 (d, J = 7.8 Hz, 2H), 7.13 (d, J = 6.6 Hz, 2H), 6.99 (s, 1H), 6.94 – 6.93 (m, 1H), 4.98 (d, J = 9.0 Hz, 1H), 4.54 – 4.53 (m, 1H), 4.39 (d, J = 8.4 Hz, 1H), 3.69 (s, 3H), 3.42 (d, J = 16.2 Hz, 1H), 2.98 – 2.97 (m, 1H), 2.79 (dd, J = 16.8 Hz, J = 4.8 Hz, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 171.4, 155.9, 138.8, 137.9, 135.4, 135.3, 130.6, 129.6, 129.3, 129.2, 129.0, 128.9, 127.8, 126.7, 126.2, 124.4, 101.9, 71.9, 67.8, 55.9, 52.5, 44.5, 21.0. HRMS (ESI-TOF) m/z:  $[M + Na]^+$  calculated for C<sub>27</sub>H<sub>24</sub><sup>35</sup>ClN<sub>3</sub>O<sub>4</sub>Na<sup>+</sup>: 512.1348, found: 512.1352; calculated for C<sub>27</sub>H<sub>24</sub><sup>37</sup>ClN<sub>3</sub>O<sub>4</sub>Na<sup>+</sup>: 514.1324, found: 514.1318.

# (2S,3S,5R,6R)-methyl 8-nitro-5-(2-nitrophenyl)-2,3-diphenyl-1,2,3,5,6,7-hexahydro imidazo[1,2-a]pyridine-6-carboxylate 5h



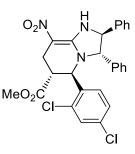
Prepared according to the general procedure B to afford **5h** (39.5 mg) in 79% yield as white solid. The diastereomeric ratio was determined to be 13:1 by crude <sup>1</sup>H NMR analysis; m.p. 130 -133 °C.

### *NMR and HRMS data for the product* **5h**:

<sup>1</sup>**H NMR (600 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm): 9.06 (brs, 1H), 7.98 (d, J = 7.8 Hz, 1H), 7.73 (t, J = 7.8 Hz, 1H), 7.55 (t, J = 7.8 Hz, 1H), 7.44 – 7.32 (m, 7H), 7.20 – 7.18 (m, 2H), 7.12 (d, J = 6.6 Hz, 2H), 5.18 – 5.17 (m, 1H), 5.07 (d, J = 9.6 Hz, 1H), 4.27 (d, J = 10.2 Hz, 1H), 3.74 (s, 3H), 3.62 (d, J = 10.8 Hz, 1H), 3.27 – 3.25 (m, 1H), 2.69 (dd, J = 16.8 Hz, J = 6.0 Hz, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm): 171.1, 156.0, 148.2, 137.4, 134.0, 133.9, 132.1, 130.0, 129.7, 129.3, 129.1, 129.0, 128.2, 127.7, 126.3, 125.9, 101.7, 72.0, 67.7, 52.6, 51.3, 42.4, 20.4. HRMS (ESI-TOF) m/z: [M + Na]<sup>+</sup> calculated for C<sub>27</sub>H<sub>24</sub>N<sub>4</sub>O<sub>6</sub>Na<sup>+</sup>: 523.1588, found: 523.1584.

# (2*S*,3*S*,5*R*,6*R*)-methyl 5-(2,4-dichlorophenyl)-8-nitro-2,3-diphenyl-1,2,3,5,6,7-hexahydro imidazo[1,2-a]pyridine-6-carboxylate 5i



Prepared according to the general procedure B to afford **5i** (40.0 mg) in 86% yield as white solid. The diastereomeric ratio was determined to be 17:1 by crude <sup>1</sup>H NMR analysis; m.p. 195 – 198 °C.

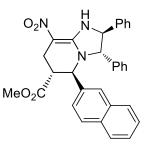
NMR and HRMS data for the product **5i**:

<sup>1</sup>**H NMR (600 MHz, CDCl**<sub>3</sub>) δ (ppm): 9.06 (brs, 1H), 7.46 – 7.30 (m, 8H), 7.22 – 7.20 (m, 2H), 7.17 – 7.14 (m, 2H), 7.04 (s, 1H), 5.05 – 4.99 (m, 1H), 5.04 (d, *J* = 9.6 Hz, 1H), 4.36 (d, *J* = 9.6 Hz, 1H), 3.71 (s, 3H), 3.54 – 3.50 (m, 1H), 3.03 – 2.98 (m, 1H), 2.61 – 2.57 (m, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 171.1, 1 56.0, 137.6, 135.2, 134.6, 133.5, 132.5, 130.6, 129.7, 129.7, 129.3, 129.2, 129.1, 128.1, 127.8, 127.7, 127.6, 126.2, 101.8, 71.9, 67.8, 52.5, 52.4, 41.7, 20.3.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for C<sub>27</sub>H<sub>23</sub>Cl<sub>2</sub>N<sub>3</sub>O<sub>4</sub>Na<sup>+</sup>: 546.0958, found: 546.0963.

### (2S,3S,5R,6R)-methyl 5-(naphthalen-2-yl)-8-nitro-2,3-diphenyl-1,2,3,5,6,7-hexahydro imidazo[1,2-a]pyridine-6-carboxylate 5j



Prepared according to the general procedure B to afford **5j** (42.9 mg) in 85% yield as white solid. The diastereomeric ratio was determined to be 17:1 by crude <sup>1</sup>H NMR analysis; m.p. 133 -138 °C.

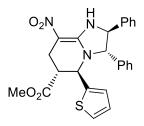
NMR and HRMS data for the product 5j:

<sup>1</sup>**H NMR** (**600 MHz**, **CDCl**<sub>3</sub>)  $\delta$  (ppm): 9.12 (brs, 1H), 7.88 (d, J = 8.4 Hz, 1H), 7.83 (d, J = 7.8 Hz, 1H), 7.62 (d, J = 8.4 Hz, 1H), 7.48 (t, J = 7.8 Hz, 2H), 7.45 – 7.34 (m, 7H), 7.28 (d, J = 6.6 Hz, 2H), 7.23 (d, J = 7.2 Hz, 1H), 7.11 (d, J = 6.6 Hz, 2H), 5.48 – 5.47 (m, 1H), 5.07 (d, J = 9.6 Hz, 1H), 4.60 (d, J = 9.0 Hz, 1H), 3.78 (s, 3H), 3.48 (d, J = 16.2 Hz, 1H), 3.18 – 3.17 (m, 1H), 2.49 (dd, J = 16.8 Hz, J = 6.0 Hz, 1H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 171.7, 156.6, 137.9, 135.6, 134.2, 131.4, 129.7, 129.6, 129.3, 129.2, 129.1, 129.0, 127.9, 127.1, 126.3, 126.2, 125.1, 122.8, 121.5, 102.0, 71.8, 67.9, 52.5, 42.7, 20.2.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for C<sub>27</sub>H<sub>25</sub>N<sub>3</sub>O<sub>4</sub>Na<sup>+</sup>: 478.1737, found: 478.1733.

# (2S,3S,5R,6R)-methyl 8-nitro-2,3-diphenyl-5-(thiophen-2-yl)-1,2,3,5,6,7-hexahydro imidazo[1,2-a]pyridine-6-carboxylate 5k



Prepared according to the general procedure B to afford **5**k (35.0 mg) in 76% yield as white solid. The diastereomeric ratio was determined to be 10:1 by crude <sup>1</sup>H NMR analysis; m.p. 115 – 117 °C.

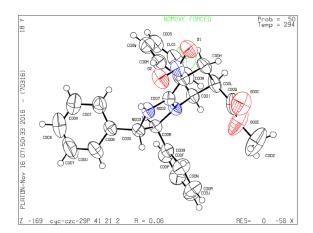
*NMR and HRMS data for the product* **5k**:

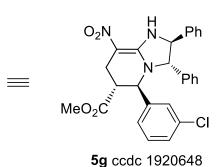
<sup>1</sup>**H NMR (600 MHz, CDCl**<sub>3</sub>)  $\delta$  (ppm): 9.06 (brs, 1H), 7.45 – 7.40 (m, 3H), 7.34 – 7.30 (m, 3H), 7.30 (d, J = 4.2 Hz, 1H), 7.18 – 7.14 (m, 4H), 6.95 (dd, J = 6.0 Hz, J = 3.0 Hz, 1H), 6.78 (d, J = 2.4 Hz, 1H), 4.90 – 4.87 (m, 2H), 4.41 (d, J = 9.0 Hz, 1H), 3.72 (s, 3H), 3.50 (d, J = 16.2 Hz, 1H), 3.09 – 3.02 (m, 2H).

<sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ (ppm): 171.4, 155.4, 139.1, 137.6, 135.3, 129.6, 129.5, 129.2, 129.1, 129.0, 129.0, 127.9, 127.0, 126.8, 126.6, 126.5, 126.3, 102.2, 71.8, 67.7, 52.5, 52.5, 44.5, 21.3.

**HRMS (ESI-TOF)** m/z:  $[M + Na]^+$  calculated for  $C_{25}H_{23}N_3O_4SNa^+$ : 484.1301, found: 484.1302.

### 5. Crystal Data and Structure Refinement for the Representative Product 5g





Identification code Empirical formula Formula weight Temperature/K Crystal system Space group a/Å b/Å c/Å  $\alpha/^{\circ}$ ß/°  $\gamma/^{\circ}$ Volume/Å<sup>3</sup> Ζ pcalcg/cm<sup>3</sup>  $\mu$ /mm-1 F(000) Crystal size/mm<sup>3</sup> Radiation

 $2\Theta$  range for data collection/° Index ranges Reflections collected Independent reflections Data/restraints/parameters Goodness-of-fit on F2 Final R indexes  $[I \ge 2\sigma(I)]$ Final R indexes [all data] Largest diff. peak/hole / e Å<sup>-3</sup> Flack parameter

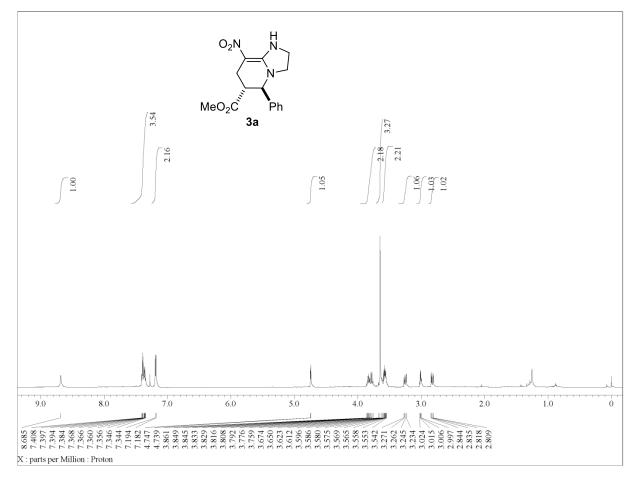
5g

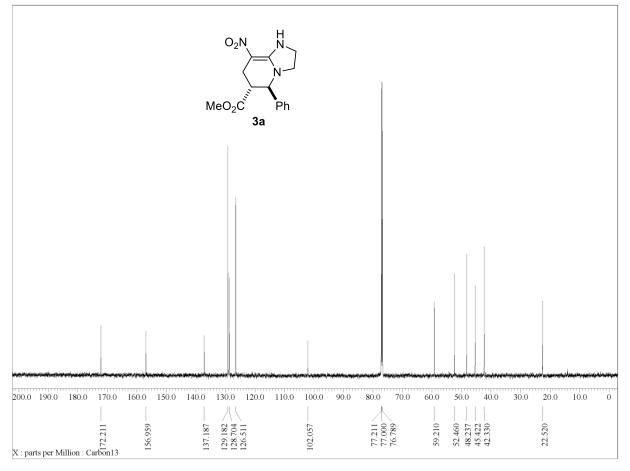
 $C_{27}H_{24}ClN_3O_4$ 489.94 293.9(3) tetragonal  $P4_{1}2_{1}2$ 10.62565(16) 10.62565(16) 43.3437(17) 90 90 90 4893.7(2) 8 1.330 1.703 2048.0  $0.7 \times 0.5 \times 0.3$ CuKa ( $\lambda = 1.54184$ ) 8.16 to 134.156  $\text{-}12 \leq h \leq 10, \, \text{-}12 \leq k \leq 6, \, \text{-}50 \leq l \leq 51$ 11517  $4209 [R_{int} = 0.0351, R_{sigma} = 0.0313]$ 4209/0/317 1.085  $R_1 = 0.0647, wR_2 = 0.1701$  $R_1 = 0.0682, wR_2 = 0.1743$ 0.40/-0.37 0.009(12)

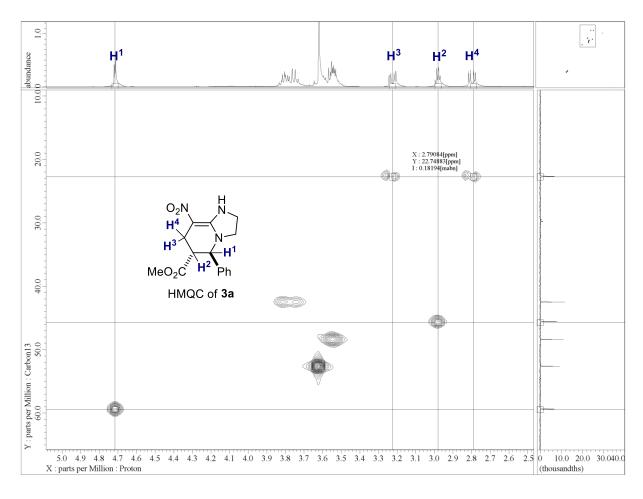
### 6. References and Notes

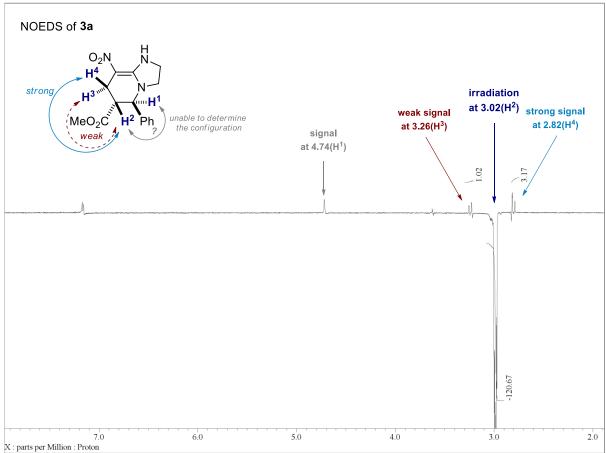
- 1 (a) E. Krell, Handbook of Laboratory Distillation, Elseriver Publishing Company, Amsterdam-London-New York, 1963; (b) M. J. Rosengart, *The Technique of Distillation* and Rectification in the Laboratory, VEB Verlag Technik, Berlin, 1954; (c) F. Stage, Angew. Chem. 1947, **19**, 175.
- 2 E. Safari, A. Maryamabadi and A. Hasaninejad, *RSC Adv.* 2017, 7, 39502.
- 3 X. Li, J. Su, Z. Liu, Y. Zhu, Z. Dong, S. Qiu, J. Wang, L. Lin, Z. Shen, W. Yan, K. Wang and R. Wang, *Org. Lett.* 2016, **18**, 956.

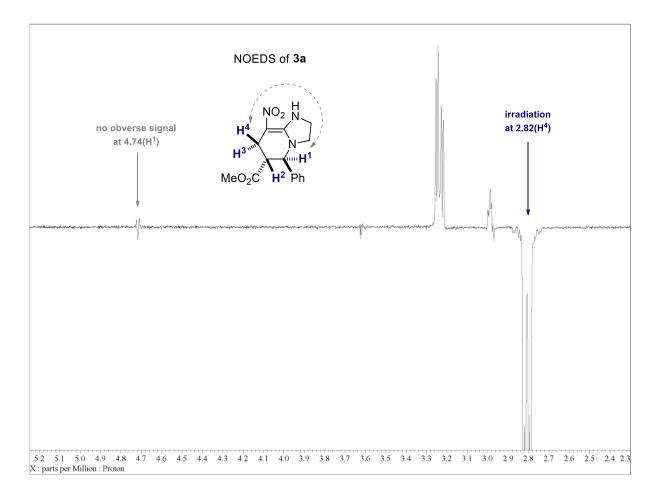
### 7. NMR Spectra

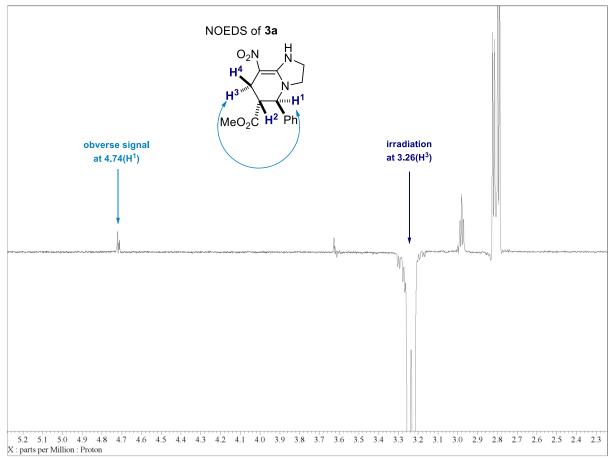


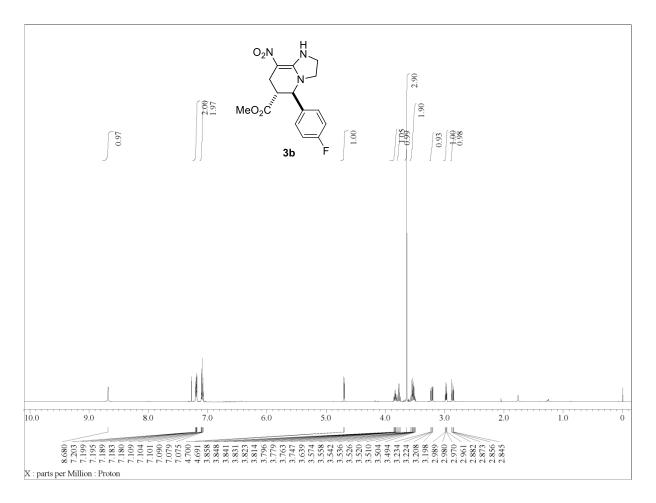


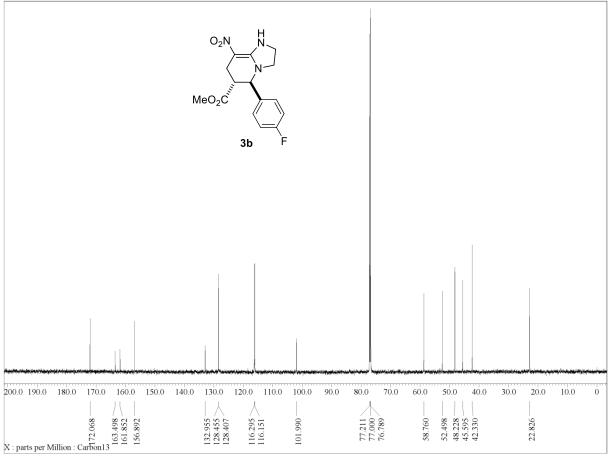


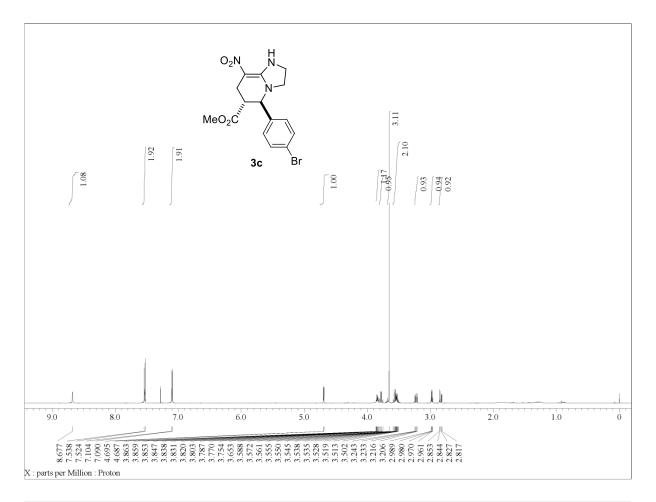


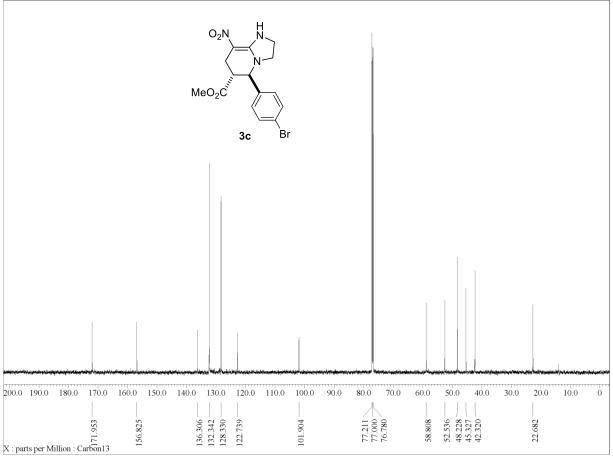


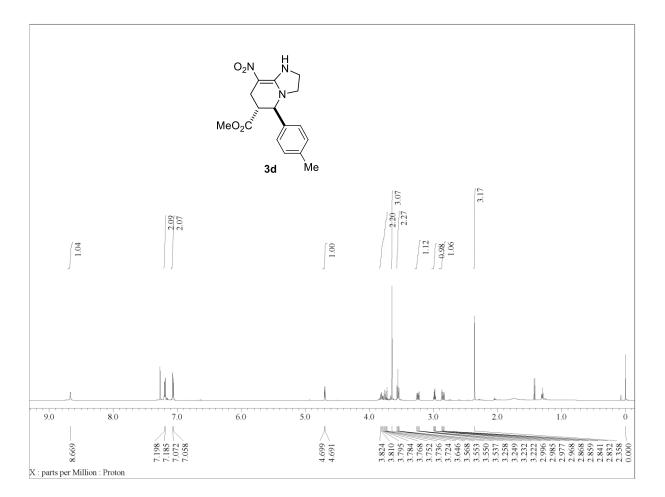


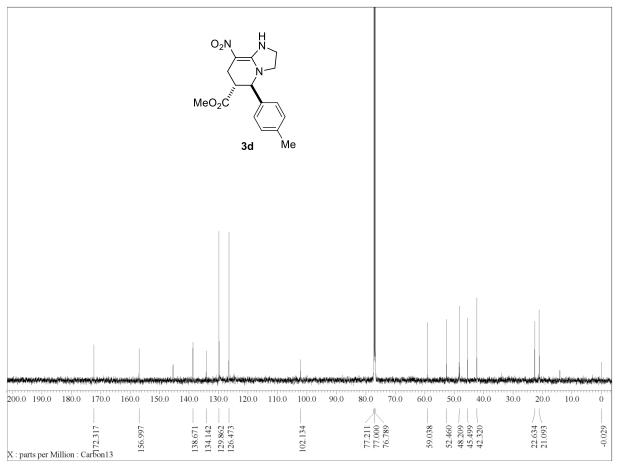


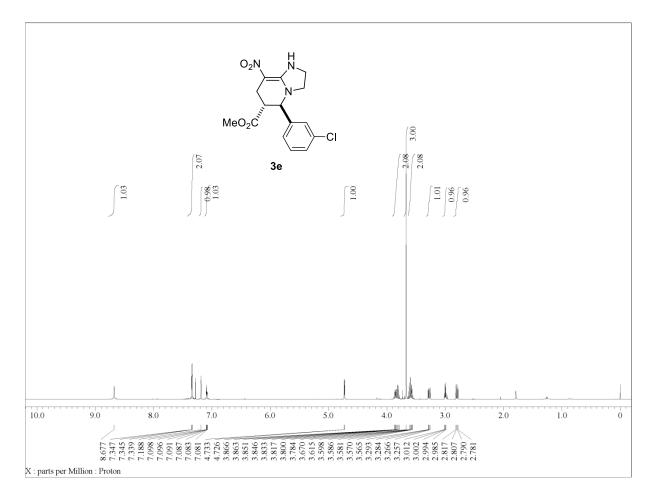


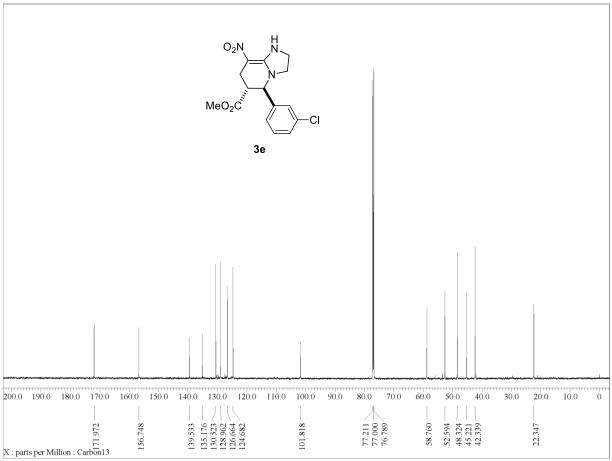


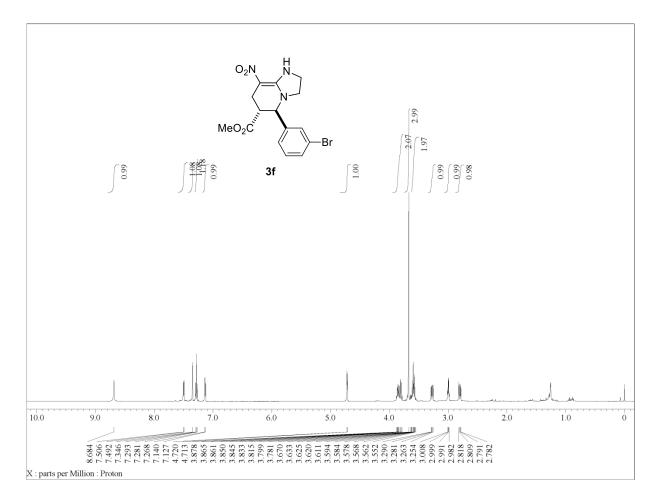


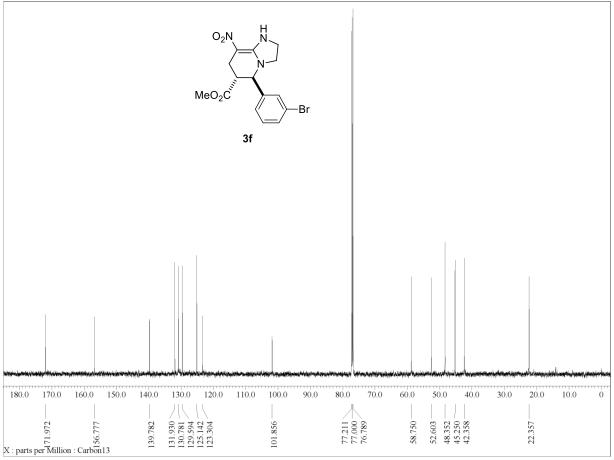


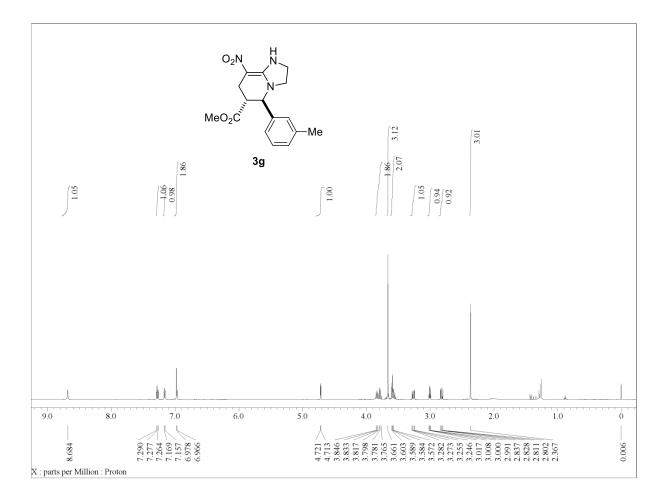


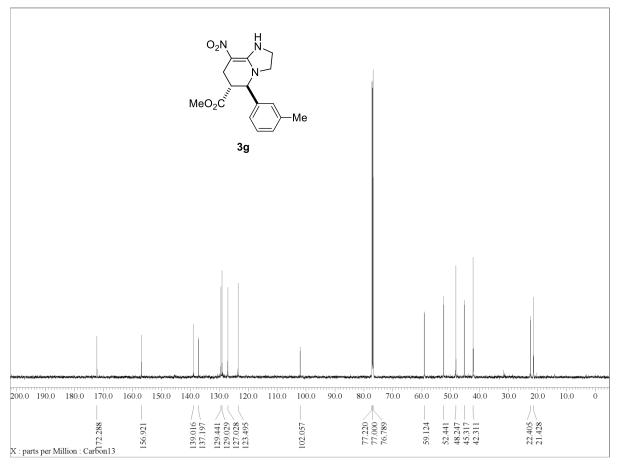


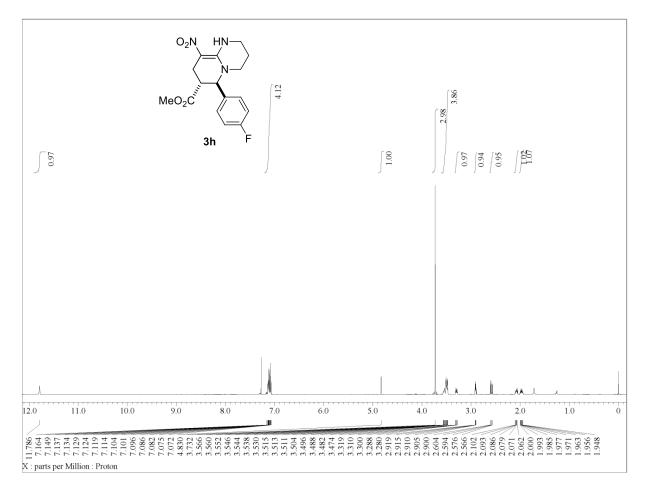


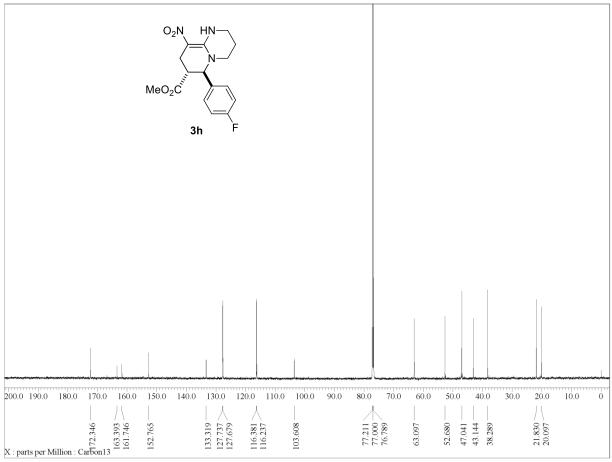


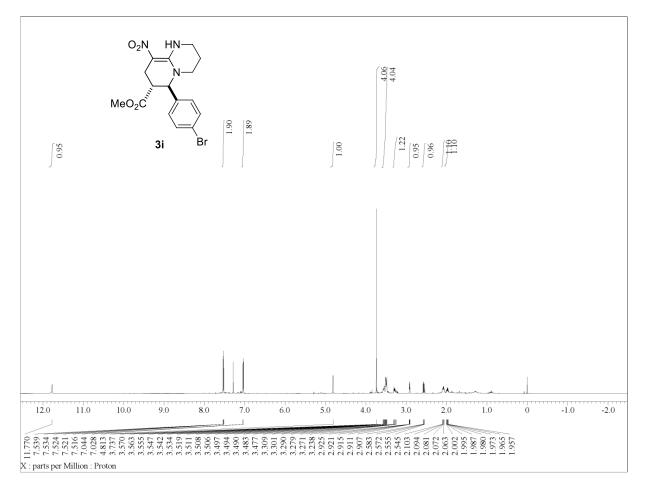


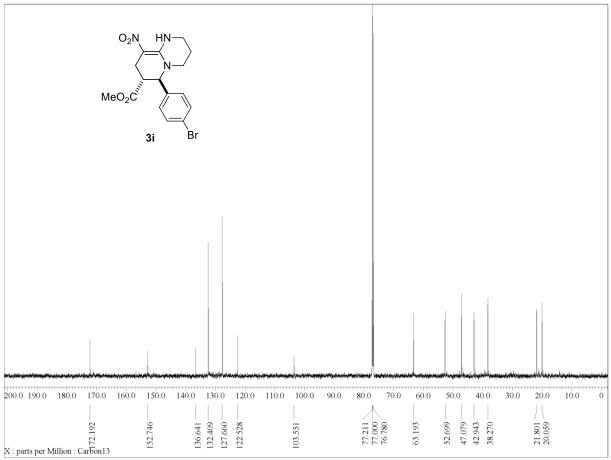


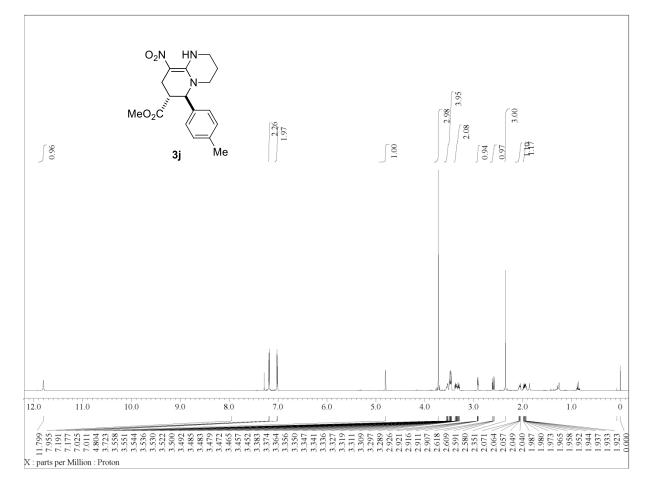


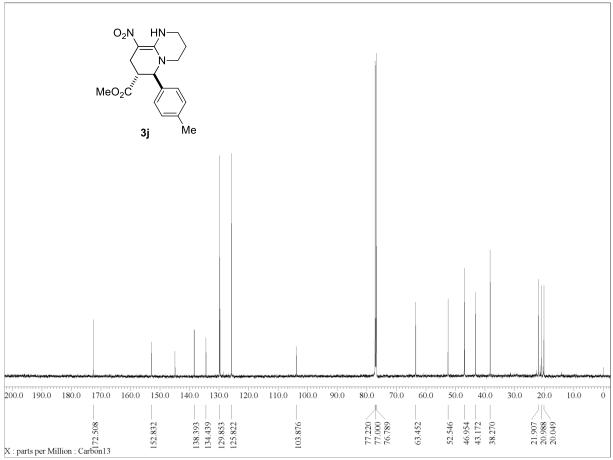


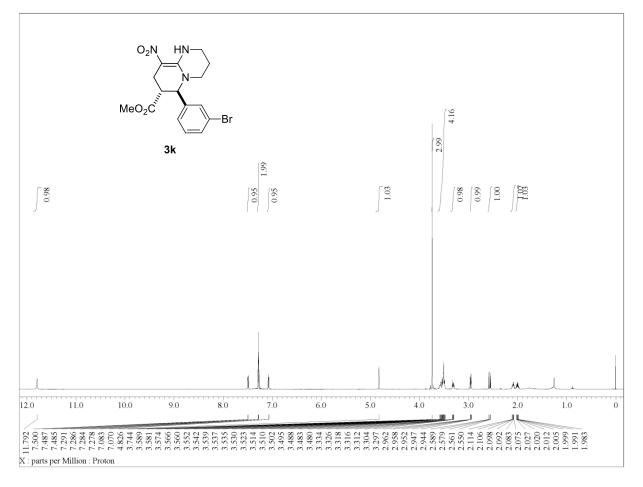


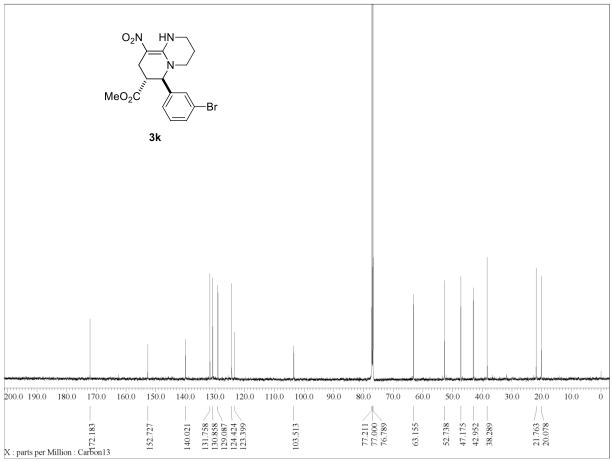


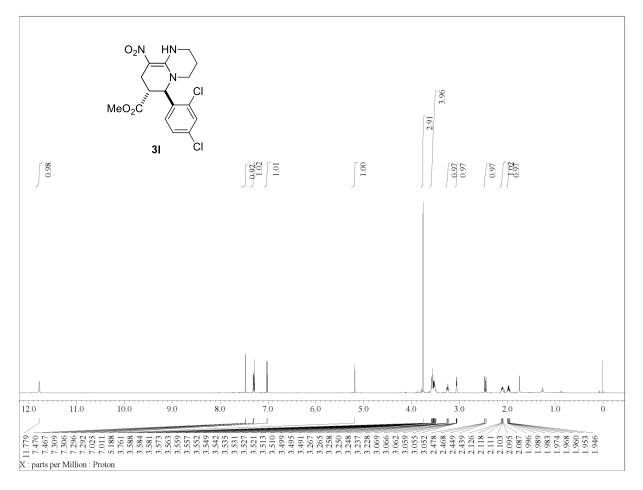


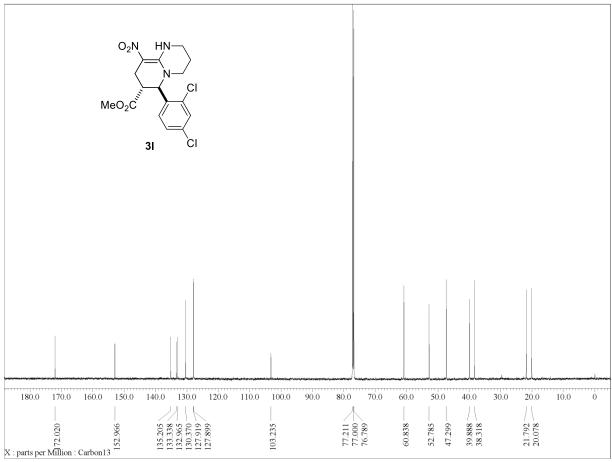


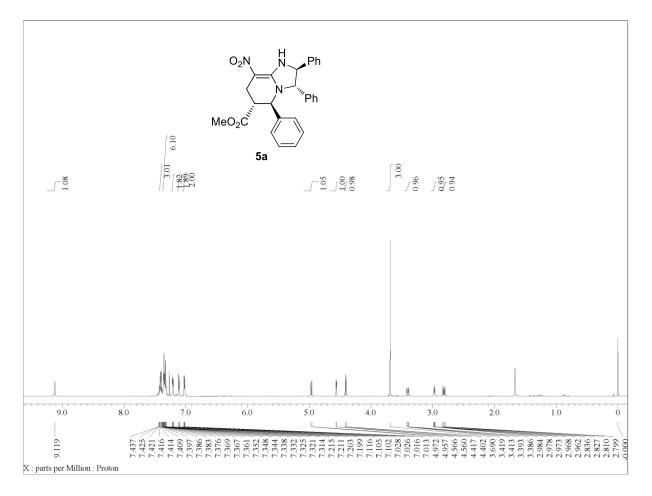


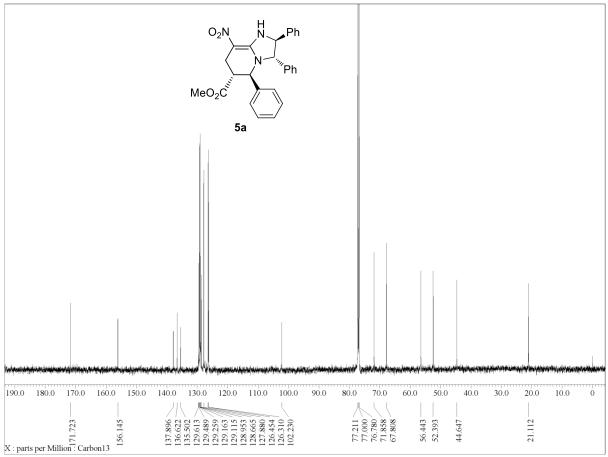


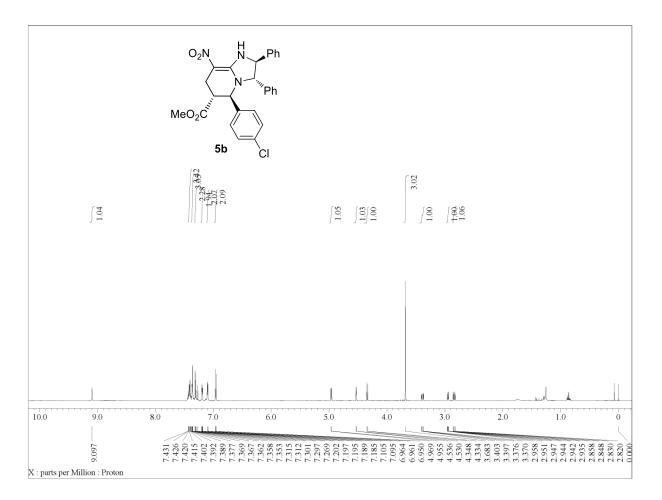


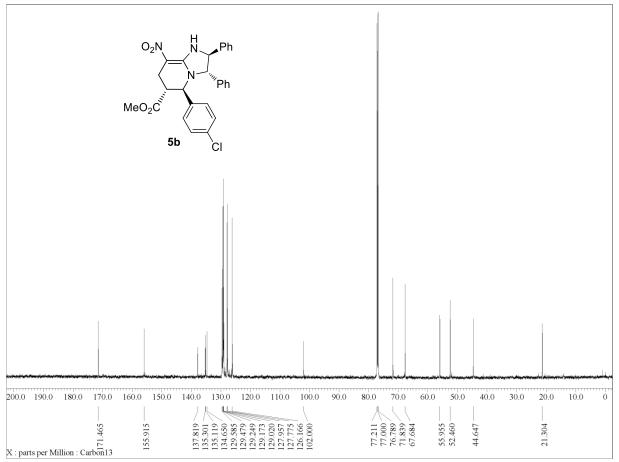


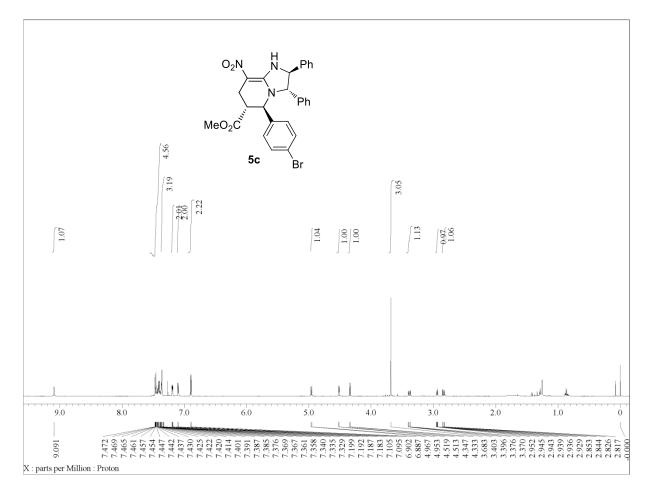


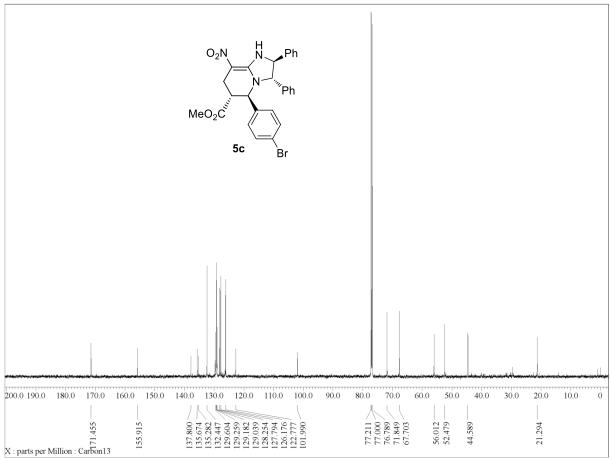


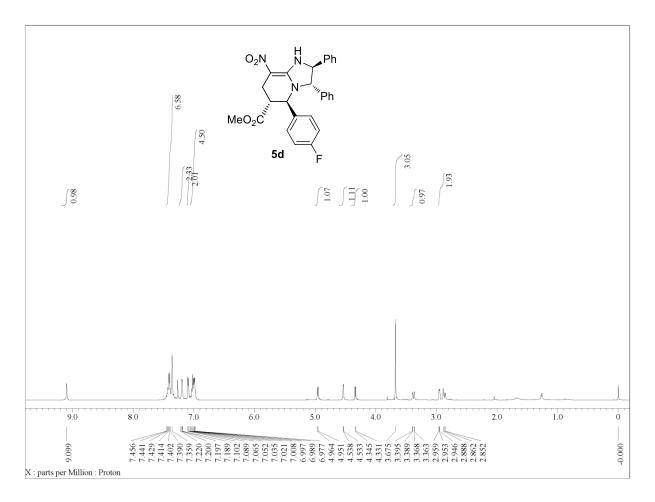


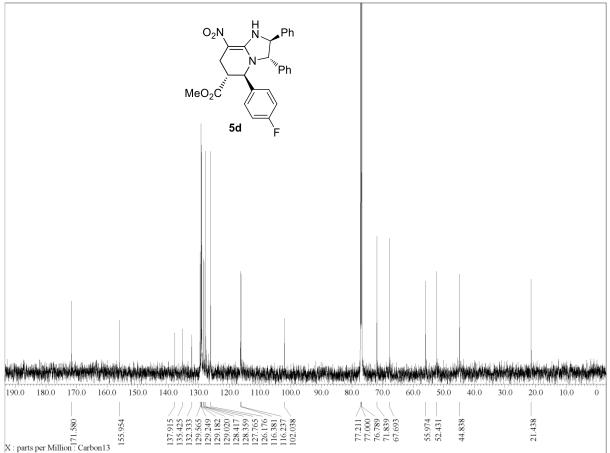


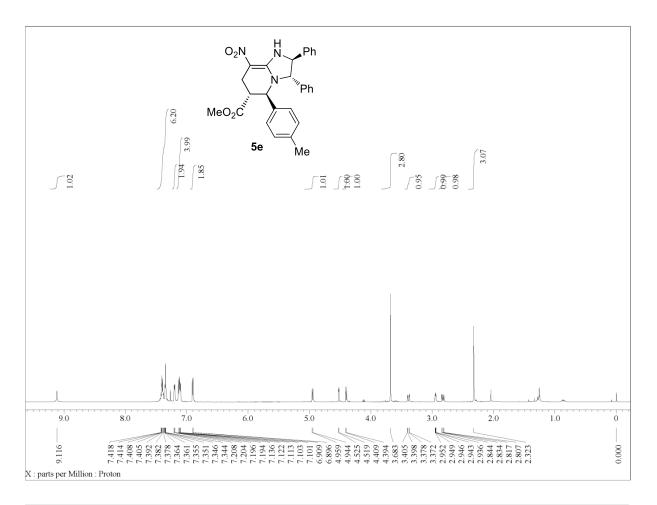


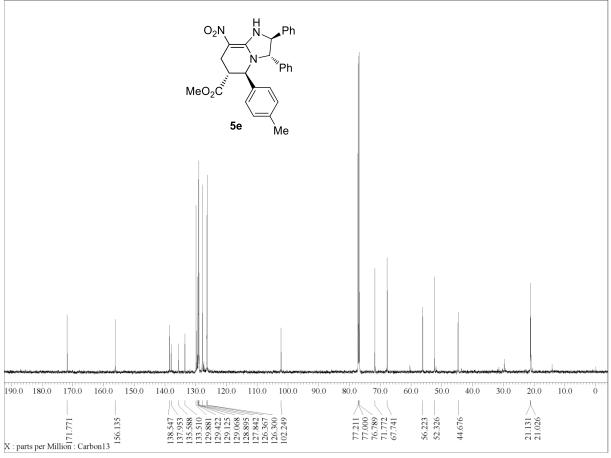


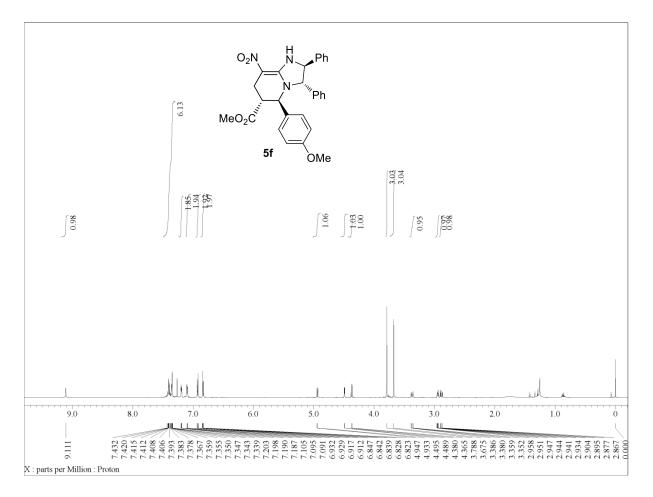


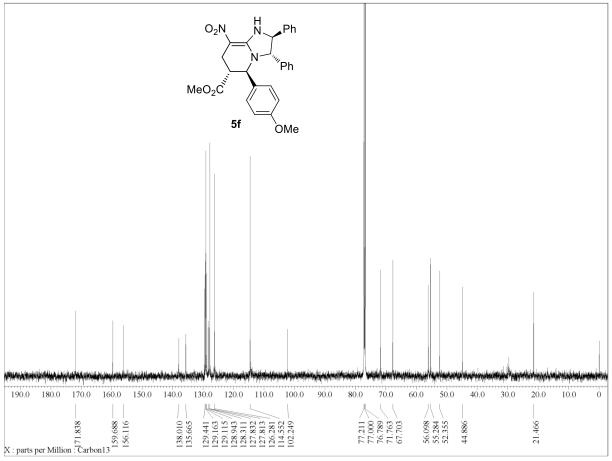


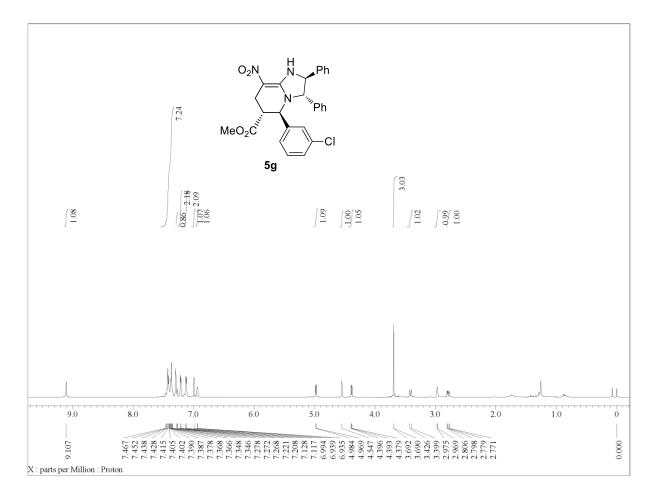


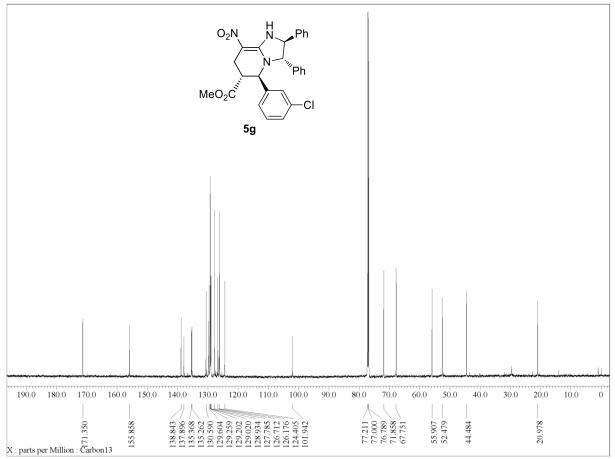


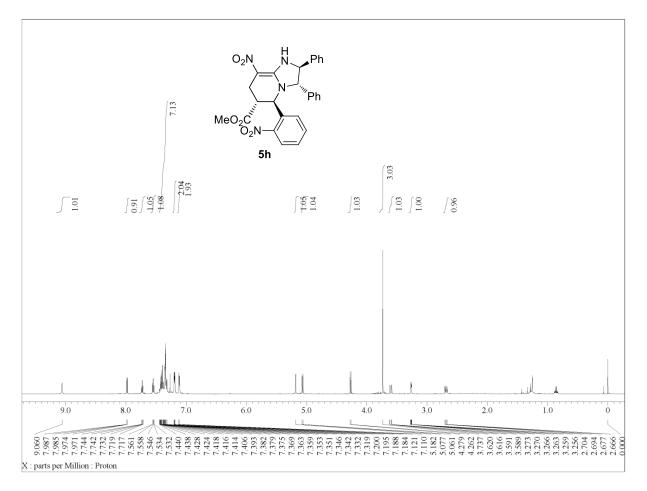


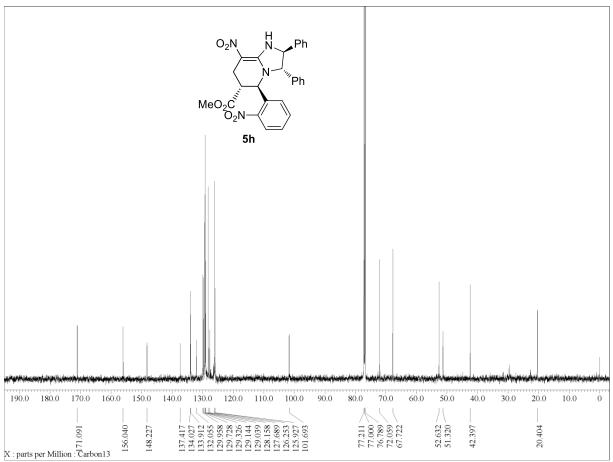


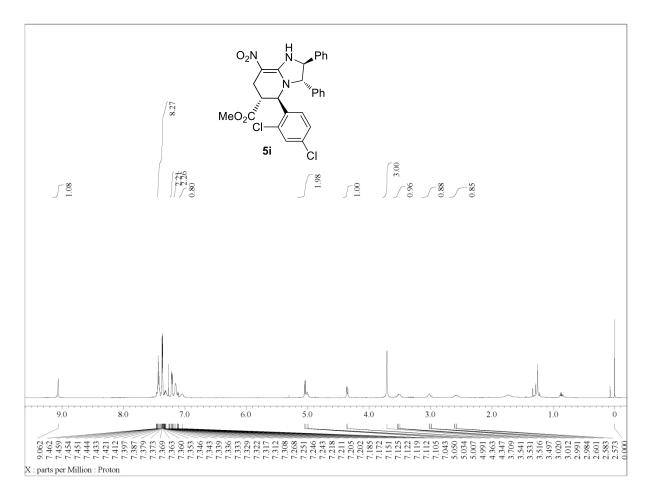


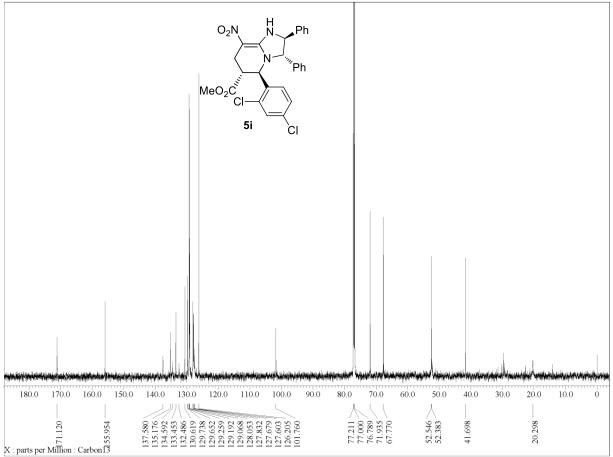


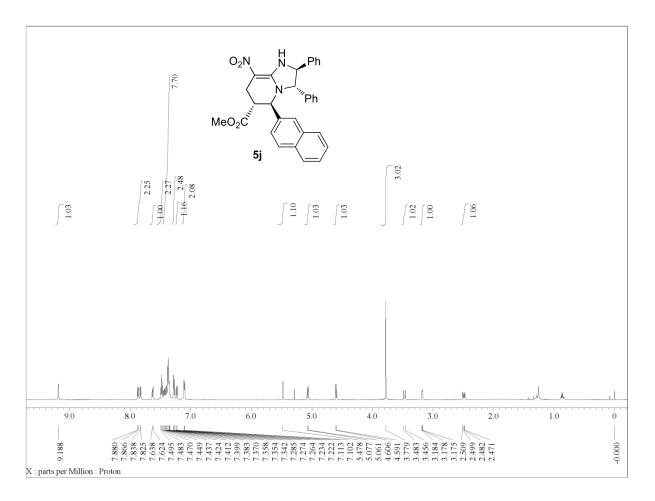


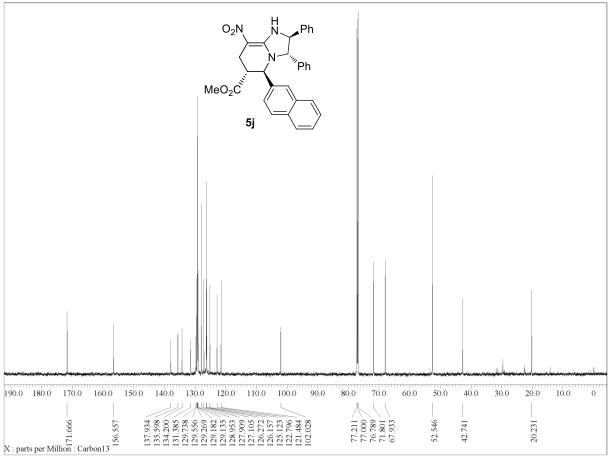


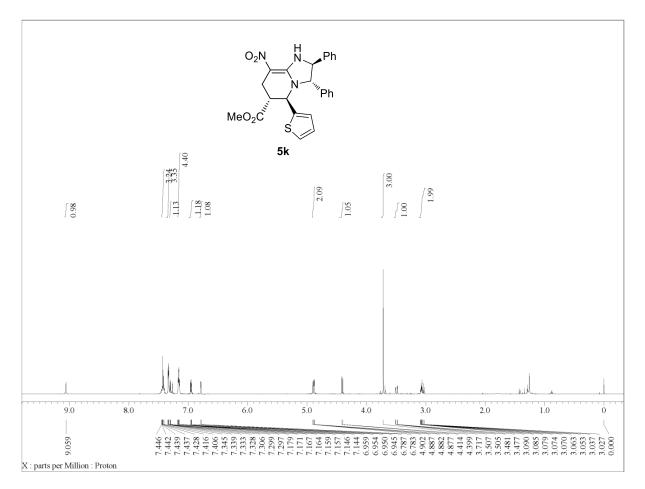


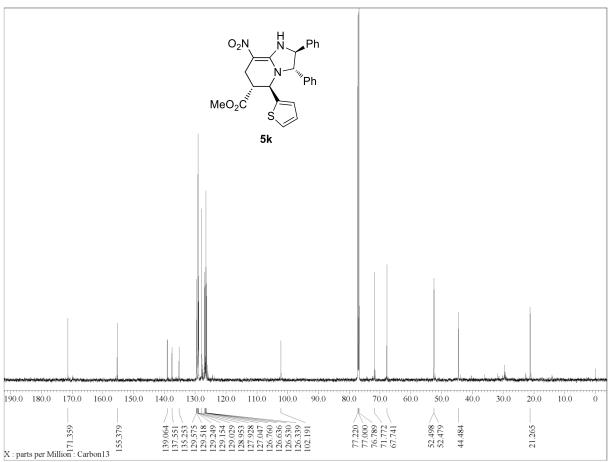












## 8. Cell culture and cellular proliferation assay

The human breast cancer cell line MDA-MB-231 and human colorectal carcinoma cell HCT116 were obtained from American Type Culture Collection (ATCC), the cells were incubated under sterile conditions at 37°C and were maintained in a humidified atmosphere 5% (v/v) CO<sub>2</sub> with RPMI-1640 or DMEM medium containing 10% fetal bovine serum (GIBCO, Waltham, MA, USA). MTT assay was performed to evaluate the cellular proliferation inhibitory activities of test compounds by cancer cells. In general, cells were seeded into 96-well plates and treated with a series of concentration of test drugs for 24 h. The MTT reagent (5 mg/ml) was added per well for 3 h at 37°C. After that, the MTT was removed and 150 µl DMSO was added to dissolve the formazan crystals. Then, optical density (OD) was measured at 570 nm of the solution. The control group consisted of untreated cells. The percentage of cell viability averaged from three individual experiments.

## 9. HTRF based MDM2-p53 interaction assay

The enzymatic assay of test compounds was using the HTRF based method provided by Cisbio Co. Ltd., in brief, the HTRF assay used a GST-tagged kinase domain of the MDM2, and then the biotinylated substrate peptide of p53 and two HTRF detection reagent are added. The HTRF signal is proportional to the amount of interactions between GST-tagged MDM2 protein and the biotinylated substrate p53 peptide, the detailed experimental procedures are according to the manufacturer's protocols and were reported in previous articles by us and other groups.<sup>[ref]</sup> These experiments involve the transfer of energy from europium pyridine-bis-bipyridine cryptate (Eu-PBBP) as donor fluorophore to Alexa 647 as acceptor fluorophore. We used the ligand labeled with Eu-PBBP and the monoclonal anti-MDM2 antibody labeled with Alexa 647 provided by Cisbio international research group. Cells or membranes were incubated with 2 nM Eu-PBBP labeled p53 substrate peptide, 3 nM Alexa 647-labeled anti-MDM2 antibody, and increasing concentrations (10 pM to 10 µM) of test compounds. As a negative control, cells or membranes were incubated only with the donor fluorophore-labeled antibody. Competition experiments in a 384-well plate were performed in a final volume of 50 µL. After an incubation of 16h at 4 °C, preparations were excited at 337 nm and fluorescence emissions were measured on 620 nm and 665 nm, wavelengths which correspond to the total europium cryptate emission and to the FRET signal, respectively. The specific signal was calculated using the following equation: DeltaF = (R - Rneg)/(Rneg)where R is the ratio (fluorescence 665 nm/fluorescence 620 nm) calculated for each assay and Rneg is the same ratio for the negative control.

#### **References:**

[1] M.-C. Yang, C. Peng, H. Huang, L. Yang, X.-H. He, W. Huang, H.-L. Cui, G. He and B. Han, *Org. Lett.* 2017, **19**, 6752.

[2] L. Albizu, G. Teppaz, R. Seyer, H. Bazin, H. Ansanay, M. Manning, B. Mouillac and T. Durroux, *J. Med. Chem.* 2007, **50**, 4976.

[3] M. Gicquel, C. Gomez, M. C. G. Alvarez, O. Pamlard, V. Guérineau, E. Jacquet, J. Bignon, A. Voituriez and A. Marinetti, *J. Med. Chem.* 2018, **61**, 9386.

# 10. Molecular docking

The CDOCKER module in Discovery Studio 3.5 were employed for molecular docking in the current study. The crystal structure of MDM2 complexed with RG7112 (PDB ID: 4IPF) was chosen as the reference structure since it has the highest resolution (1.7 Å) among all the imidazole-based MDM2 inhibitors co-crystal structures. We adjusted the docking parameters until the docked pose is as close as possible to the original crystallized structure in the p53 substrate binding site of MDM2. The docking parameters of CDOCKER used default settings, unless for special statements.

## 11. Western blot analysis

HCT116 Cells were treated with 0.5, 2.0  $\mu$ M of compound **5c** or 2.0  $\mu$ M of RG7112 as described above, collected and washed twice with ice-cold PBS, then lysed in RIPA buffer (20 mM Tris-HCl, 150 mM NaCl, 0.5% sodium deoxycholate, 5 mM EDTA, 1% Nonidet P-40, 0.1% SDS) supplemented with protease and phosphatase inhibitor cocktails (Sigma-Aldrich, MA, USA). Protein concentration was measured using the bicinchoninic acid assay (Keygen, Nanjing, China). Total protein (approximately 50  $\mu$ g) was fractionated on a 10-15% sodium dodecyl sulfate polyacrylamide gel, then electrophoretically transferred to a polyvinylidene difluoride membrane (Millipore, CA, USA). Membranes were incubated at 4 °C overnight with primary antibodies against MDM2, p53, p21 and GAPDH. Membranes were then incubated with alkaline phosphatase-conjugated secondary antibody, antibody binding was visualized using enhanced chemiluminescence, and bands were quantitated using an imaging system.

## 12. Immunofluorescence assay

The HCT116 cells were seeded and cultured in 6-well dishes, HCT116 cells were treated with 2.0  $\mu$ M of compound **5c** or RG7112 for 24 hours, then washed and fixed by 10% formalin. The fixed samples were treated by TritonX-100, incubated overnight at 4 °C with the anti-p53 primary antibodies, followed by incubation with the secondary antibody for 1 h in the dark, and observed using fluorescence microscopy (Axio Observer A1, Zeiss, Germany).

13. Superposition of compound 5c binding conformers and p53 substrate peptide (Figure S1).

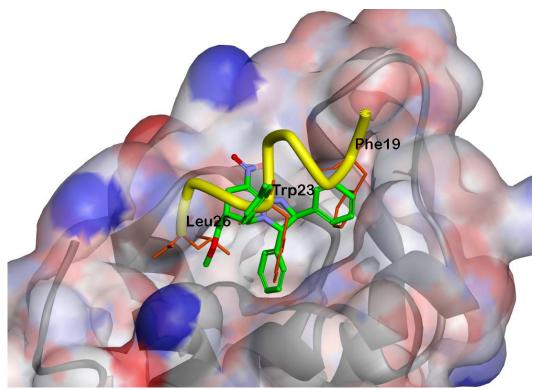


Figure S1. Comparison of compound 5c to p53 substrate peptide, green: compound 5c, yellow: main chain of p53 substrate peptide, orange: the interaction residues of p53 substrate peptide to MDM2, Phe19, Trp23 and Leu26.

Compound -	MDM2		HCT116		MDA-MB-231	
	Mean	SD	Mean	SD	Mean	SD
<b>3</b> a	7.79	0.51	16.24	1.46	21.82	1.22
<b>3</b> b	8.78	0.83	18.37	1.69	28.90	2.75
3c	4.34	0.41	21.98	2.31	12.60	0.84
3d	15.54	1.43	18.65	1.42	16.93	1.35
3e	5.78	0.53	25.02	1.73	26.34	2.55
<b>3f</b>	16.43	1.71	29.31	1.67	17.70	1.56
3g	13.91	1.86	12.05	1.42	13.21	1.57
3h	2.04	0.20	24.81	2.65	29.78	2.00
<b>3i</b>	11.12	1.42	28.79	3.60	17.61	0.99
3ј	8.97	1.18	28.5	1.69	29.84	2.28
3k	14.51	1.19	23.54	2.14	28.26	1.42
31	11.21	0.95	28.52	2.07	23.66	2.38
<b>5</b> a	72.57	9.83	68.46	9.17	76.93	6.17
5b	76.43	4.74	85.32	9.73	77.69	6.29
5c	80.99	7.87	91.99	6.91	89.28	4.35
5d	61.25	6.37	86.59	11.50	75.77	4.39
5e	72.17	6.64	86.35	4.49	82.06	7.83
<b>5f</b>	79.66	5.02	82.71	10.67	81.96	6.88
5g	71.29	4.78	79.23	6.51	84.75	10.42
5h	62.92	4.98	61.25	3.92	57.12	5.43
5i	65.06	6.63	63.69	5.99	62.86	6.13
5ј	69.65	6.11	73.21	5.22	64.25	6.94
5k	72.31	6.94	84.51	12.47	85.21	4.86
<b>RG7112</b>	85.42	3.08	83.66	6.31	80.83	4.59

14. Inhibition of MDM2 activity and of proliferation of HCT116 and MDA-MB231 cells by synthetic imidazoline derivatives (Table S3).