Structural, kinetics and mechanistic studies of asymmetric transfer hydrogenation of

ketones catalyzed by chiral (pyridyl)imine nickel(II) complexes

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Synthesis and characterization of ligands

(S-)-1-phenyl-N-(pyridine-2-yl) ethylidine) ethanamine (L1)

Compound (*S*-)-1-phenyl-N-(pyridine-2-yl) ethylidine) ethanamine (**L1**) was synthesized by refluxing 2-acetylpyridine (2.00 g, 16.51 mmol), (*S*-)-methylbenzylamine (2.00 g, 16.51 mmol), and p-TsOH monohydrate (1.0 mol %) toluene (25 mL) for 12 h using Dean Stark apparatus. After 12 h of reaction time, the solvent was removed under vacuum to obtain a brown crude oil. This was dissolved in CH₂Cl₂ (15 mL) and washed with water (3x15 mL). The organic layer was separated followed by drying over anhydrous MgSO₄, filtered and solvent evaporated to obtain **L1** as a brown oil. Yield: 2.67 g (72%). ¹H NMR (CDCl3, δ , ppm): 1.56 (d, 3H, NCHCH₃ ³J_{HH} = 3.6), 2.24 (**s**, 3H, CCH₃N), 4.69 (q, 1H, NCH ³J_{HH} = 4.0), 8.82 (d, 1H, 5H-py, ³J_{HH} = 7.6), 8.86 (d, 1H, 6H-py, ³J_{HH}, = 7.6), 8.23 (dd, 1H, 4H-py, ³J_{HH} = 7.6), 7.98 (dd, 1H, 3H-py, ³J_{HH} = 7.6 Hz); 7.29 (dd, 2H, 3-bz, ³J_{HH}, 8.0Hz); 7.39 (dd, 1H, 4-bz, ³J_{HH} = 8.0 Hz); 7.29 (d, 2H, 2-bz, ³J_{HH} = 8.0 Hz). ¹³C NMR (CDCl₃): δ 24.59 (CH₃), 16.73 (CH₃), 160.48 (C=N), 154.87 (2-Py-C), 149.36 (6-Py-C), 144.63 (*0*-bz-C), 128.25 (*m*-bz-C), 127.03 (3-py-C), 121.47 (5-py-c), 69.57 (1C, =N-C). FT-IR (cm⁻¹): (v_{C=N})_{mine}=1697. ESI-MS: *m/z* (%) 247 [M⁺ + Na⁺, 100]: HRMS-ESI ([M+ Na⁺]): Anal. Calc: 247.1211; Found: 247.1206.

(R)-1-phenyl-N-(pyridine-2-yl) ethylidene) ethanamine (L2)

(R)-1-phenyl-N-(pyridine-2-yl) ethylidene) ethanamine (L2) was synthesised according to protocol described for L1 using (*R*)-1- phenyl ethanamine (2.00 g, 16.51mmol), and 2-acetylpyridine (2.00 g, 16.51mmol). Yield = 3.10 g (83 %). ¹H NMR (400 MHz, CDCl₃): $\delta 2.24$ (s, 3H, CCH₃N); 1.6 (d, 3H, NCCH ³J_{HH} = 3.6 Hz); 4.69 (q, 1H, N-CH, ³J_{HH} = 4.0 Hz), 8.82 (d, 1H, 5H-py, ³J_{HH} = 7.6 Hz); 8.67 (d, 1H, 5H-py, ³J_{HH} = 7.6 Hz); 8.23 (dd, 1H, 4-H-

py,³ $J_{HH} = 7.6$); 7.98 (dd, 1H, 3-H-py, ³ $J_{HH} = 7.6$ Hz); 7.29 (dd, 2H, 3H-bz, ³ $J_{HH} = 8.0$ Hz); 7.39 (dd, 1H, 4H-bz, ³ $J_{HH} = 8.0$ Hz); 7.29 (d, 2H, 2H-bz, ³ $J_{HH} = 8.0$ Hz). ¹³C NMR (CDCl₃): δ 24.59 (<u>C</u>H₃), 16.73 (<u>C</u>H₃), 160.48 (C=N), 154.87 (2-Py-C), 149.36 (6-Py-C), 144.63 (*o*-bz-<u>C</u>), 128.25 (*m*-bz-C), 136.26 (4-py-C), 127.03 (3-py-C), 121.47 (5-py-C), 69.57 (1C, =N-<u>C</u>). FT-IR (cm⁻¹): ^v(C=N)_{imine} = 1687.29. ESI- MS: *m*/*z* (%) 247 [M⁺⁺ Na⁺, 100]. HRMS-ESI ([M⁺ + Na⁺]): Anal Calc: 247.1211; Found: 247.1206.

(S)-1-phenyl-N-(pyridine-2-yl methylene) ethanamine (L3)

To a solution of (*S*)-2-methyl benzylamine (2.15 g, 17.8 mmol) in CH₂Cl₂ (10. mL) was added a solution of 2-pyridine carboxyaldehyde (1.91 g, 17.8 mmol) in CH₂Cl₂ (10. mL) and MgSO4 (0.50 g) and stirred at room temperature for 12 h. After the reaction period, the crude product was filtered and the solvent was removed under reduced pressure to afford **L2** as a brown oil. Yield = 3.27 g, (87 %). ¹H NMR (400 MHz, CDCl₃): δ 1.6 (d, 3H, NCCH₃ ³*J*_{HH} = 3.6); 4.69 (q, 1H, H(a), ³*J*_{HH} = 3.6), 8.65 (d, 1H, H_(d), ³*J*_{HH} = 7.6); 8.51 (*s*, 1H, HCN); 8.23 (dd, 1H, 3H-py, ³*J*_{HH} = 7.6); 7.74 (dd, 1H, 4H-py, ³*J*_{HH} = 7.6); 7.48 (dd, 2H, 5H-py, ³*J*_{HH}.= 8.0Hz); 7.39 (dd, 1H, 3H-bz, ³*J*_{HH} = 8.0 Hz); 7.29 (*d*, 2H, 2H-bz, ³*J*_{HH} = 8.0 Hz). ¹³C NMR (CDCl₃): δ 24.59 (CH₃), 16.73 (CH₃), 160.48 (C=N), 154.87 (5-Py-C), 149.36 (6-Py-C), 144.63 (*o*-bz-C), 128.25 (*m*-bz-C), 127.03 (5-py-C), 121.47 (3-py-c), 69.57 (1C, =N-C). FT-IR (cm⁻¹): ($v_{C=N}$)_{imine} = 1643, ESI-MS: *m/z* (%) 233 [(M+ Na)⁺, 100%]. HRMS-ESI: Anal. Calc: 211.1235; Found: 211.1232.

(R)-1-phenyl-N-(pyridine-2-yl methylene) ethanamine (L4)

(*S*)-2-methyl benzylamine (2.15 g, 17.8 mmol), 2-pyridine carboxyaldehyde (1.91 g, 17.8 mmol), and MgSO₄.H₂O (1.00 g). Yield = 3.10 g, (83%). ¹H NMR (400 MHz, CDCl₃): δ 1.6 (d, 3H, NCC<u>H</u>₃, ³*J*_{HH} = 3.6); 4.69 (q, 1H, NC<u>H</u>, ³*J*_{HH} = 4.0), 8.65 (d, 1H, 6H-py, ³*J*_{HH} = 7.6 Hz); 8.51 (s, 1H, H_(c)); 8.23 (dd, 1H, 3H-py, ³*J*_{HH} = 7.6); 7.98 (dd, 1H, 3H-py, ³*J*_{HH} = 7.6); 7.48 (dd, 2H, 5H-py, ³*J*_{HH} = 7.29Hz); 7.39 (dd, 1H, 3H-bz, ³*J*_{HH} = 8.0); 7.29 (*d*, 2H, 4H-bz, ${}^{3}J_{\text{HH}} = 8.0$); 7.28 (dd, 2H, 2H-bz ${}^{3}J_{\text{HH}} = 8.0$). 13 C NMR (CDCl₃): δ 24.59 (CH₃), 16.73 (CH₃), 160.48 (C=N), 154.87 (2-Py-C), 149.36 (6-Py-C), 144.63 (*o*-bz-C), 128.25 (*m*-bz-C), 127.03 (5-py-C), 121.47 (3-py-c), 137.25 (4-py-C), 69.57 (1C, =N-C). FT-IR (cm⁻¹): (v_{C=N})_{imine}=1642, ESI-MS: *m*/*z* (%) 233 [(M +Na)⁺, 100%]. HRMS-ESI: Anal. Calc. 211.1235; Found: 211.1233.



Fig S1. ¹H NMR spectrum of L1.



Fig S2. ¹H NMR spectrum of L2.



Fig S4. ¹H NMR spectrum of L4.





Single Mass Analysis Tolerance = 5.0 PPM / DBE: min = -1.5, max = 500.0 Element prediction: Off Number of isotope peaks used for i-FIT = 2

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Monoisotopic Mass, Even Electron Ions 8 formula(e) evaluated with 1 results within limits (up to 20 closest results for each mass) Elements Used: C: 15-20 H: 15-20 N: 0-5 Na: 0-1 RK-L1(S) 2 (0.034) Cm (1:61) TOF MS ES+







Fig. S8. HR MS of L2.

Single Mass Analysis Tolerance = 5.0 PPM / DBE: min = -1.5, max = 500.0 Element prediction: Off Number of isotope peaks used for i-FIT = 2

Monoisotopic Mass, Even Electron Ions 3 formula(e) evaluated with 1 results within limits (up to 20 closest results for each mass) Elements Used: C: 10-15 H: 10-15 N: 0-5 RK-L4(R) 3 (0.068) Cm (1:61) TOF MS ES+







Fig. S10. HR-MS of L4.







Fig. S12. FT-IR spectrum of L2.







Fig. S14. FT-IR spectrum of Ni1.



Fig. S15. FT-IR spectrum of Ni2.



Fig. S16. FT-IR spectrum of Ni3.



Fig. S17. FT-IR spectrum of Ni4.



Fig. S18: (a) ESI-MS (Low resolution) spectra of complex Ni1. (b) Experimental (green) and simulated (red) spectrum for signal at at $m/z = 589.1991(1/2M^+ - Br]$ in complex Ni1.



Fig. S19 (a) ESI-MS (Low resolution) spectrum of Ni2. (b) Experimental (green) and simulated (red) spectrum for molecular ion at m/z = 591.1953 in Complex Ni2



Fig. S20 (a) ESI-MS (low resolution) spectrum of Ni3. (b) Experimental (green) and simulated (red) spectrum for molecular ion at m/z = 558.9774 in Complex Ni3



Fig. S21 (a) ESI-MS (low resolution) spectrum of Ni4. Inset shows the mass spectrum acquired in negative mode of the $[NiBr_4]^{2-}$ (m/z= 373 amu) counterion. (b) Experimental (green) and simulated (red) spectrum for molecular ion at m/z = 559.1251 in Complex Ni4.



Fig. S22 (a) ESI-MS (low resolution) spectrum of Ni5. (b) Experimental (green) and simulated (red) spectrum for molecular ion at m/z = 483.1721 in Complex Ni5



Fig. S23. Typical ¹H NMR spectrum of crude product from ATH reaction of acetophenone using complex **4** after 24 h indicating the intensity of methyl groups of 1-phenylethanol and acetophenone respectively used for calculating percentage conversions.



Fig. S24. Typical GC spectrum of crude sampled from ATH reaction of acetophenone after 24 h indicating the intensity of (R-) and (S-)-1-phenylethanol and acetophenone.



Fig. S25. Time dependence transfer hydrogenation (TH) of acetophenone by the Ni(II)-complexes Ni1- Ni5.



Fig. S26. Graphs showing pseudo-first order/first-order exponential fits of kinetics for complexes **Ni1-Ni5**. The linear section (0-10 mins) of the plots was used in calculating the order of the ATH reactions with respect to acetophenone substrate.



Fig. S27. Plots of catalytic activity versus catalyst loading for complexes Ni4.



Fig. S28. ESI-MS (low resolution) showing molecular mass corresponding to fragments of reactive intermediates present in reacting mixture sampled after 12 h of ATH of acetophenone.



Fig. S29. Mercury (3, 5, and 10 drops) and PPh₃ (100 mol%) poisoning tests. The observed catalytic activities of **Ni4** in the presence of excess mercury and 100% PPh₃ (55%) poison agents support that presence of largely homogeneous active species in the transfer hydrogenation reactions.