

Supplementary Information

Metal-free quinolylation of the primary amino groups of amino acid derivatives and peptides with dihydrooxazolo[3,2-a]quinoliniums

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1. Supporting figure

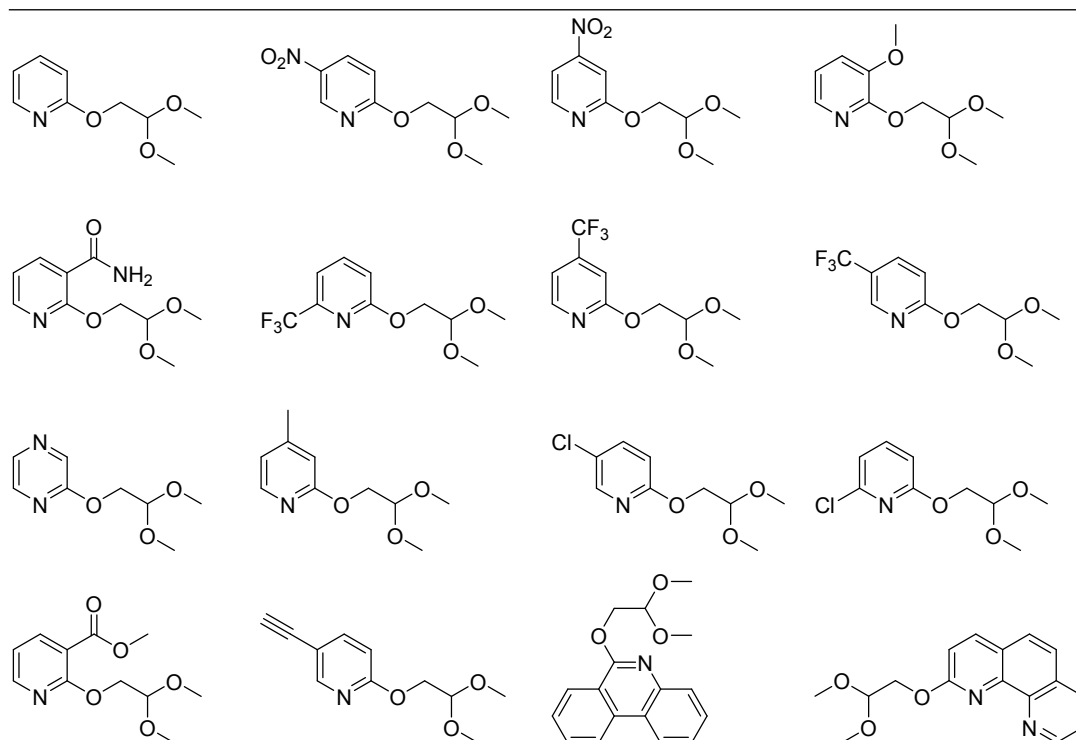


Figure S1. Screened substrates that can't react with N-terminus of amino acid

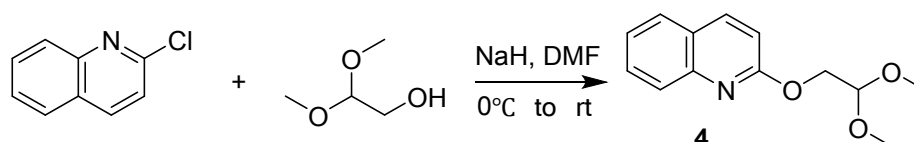
2. Chemical experiments

2.1 General information

All reactions were carried out at room temperature under anhydrous atmosphere. Yields were determined by HPLC. All materials and solvents were purchased from commercial suppliers and used without further purification. The LRMS and HRMS were recorded on Finnigan LCQ/DECA and Micromass Ultra Q-TOF (ESI) spectrometer, respectively.

The ^1H and ^{13}C spectra were taken on Bruker Avance - 400 and 500 NMR spectrometer operating at 400 MHz for ^1H NMR and 125 MHz for ^{13}C NMR, respectively, using TMS as the internal standard and CDCl_3 , $\text{MeOD-}d_4$, $\text{Acetone-}d_6$ or $\text{DMSO-}d_6$ as the solvent. Chemical shifts are given in δ values of ppm. The abbreviations s is singlet, d is doublet, t is triplet and m is multiplet. Coupling constants (J) were measured in hertz (Hz).

2.2 Substrate synthesis



2-(2,2-Dimethoxyethoxy)quinoline (4). 2,2-dimethoxyethan-1-ol (3.89 g, 36.6 mmol) was charged into a 250 ml round-bottom flask containing anhydrous DMF (50 ml). Sodium hydride (1.76 g, 73.2 mmol) was added. After 30 minutes, 2-chloroquinoline (4 g, 24.4 mmol) was added and the reaction was stirred at room temperature for 24 h. The grey solution was quenched with

ice water (200 ml), extracted with EtOAc (200 ml × 2), washed with brine (200 ml × 2) and dried over Na₂SO₄, and purified by column chromatography to afford **4** as a white solid in 81% yield (4.608 g). ¹H NMR (400 MHz, CDCl₃) δ 8.02 (d, *J* = 8.8 Hz, 1H), 7.85 (d, *J* = 8.5 Hz, 1H), 7.74 (d, *J* = 7.9 Hz, 1H), 7.64 (ddd, *J* = 8.3, 6.8, 1.5 Hz, 1H), 7.43 – 7.38 (m, 1H), 6.99 (d, *J* = 8.8 Hz, 1H), 4.86 (t, *J* = 5.3 Hz, 1H), 4.58 (d, *J* = 5.3 Hz, 2H), 3.51 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 161.41, 146.29, 138.81, 129.46, 127.37, 127.26, 125.18, 124.11, 113.07, 101.84, 64.58, 53.95. HRMS (ESI) [M+H]⁺ found 234.1127, calcd for C₁₃H₁₆NO₃ 234.1125.

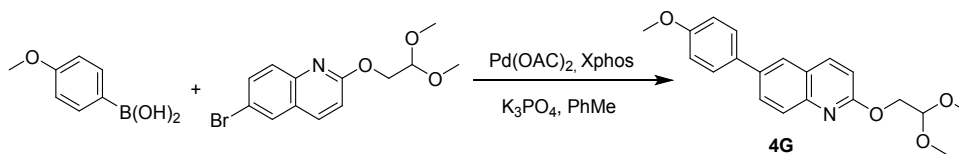
Alternative preparation method for compound **4**: quinolin-2-ol (10.0 mmol) was charged into a round-bottom flask containing anhydrous acetone (50 ml). Sodium carbonate (12.0 mmol) was added. After 30 minutes, 2-bromo-1,1-dimethoxyethane (11.0 mmol) was added and the reaction was stirred at reflux temperature for 48 h. The reaction mixture was quenched with ice water and filtered. After washing by water, the precipitate was dried to give the compound **4** in almost quantitative yield.

2-(2,2-Dimethoxyethoxy)-4-methylquinoline (4A). Following the procedure of **4** starting from 2-chloro-4-methylquinoline, the product **4A** was obtained as a white solid in 39% yield. ¹H NMR (400 MHz, CDCl₃) δ 7.89 (d, *J* = 8.2 Hz, 1H), 7.85 (d, *J* = 8.3 Hz, 1H), 7.63 (ddt, *J* = 8.2, 7.0, 1.3 Hz, 1H), 7.42 (ddt, *J* = 8.0, 7.0, 1.2 Hz, 1H), 6.85 (d, *J* = 1.3 Hz, 1H), 4.85 (td, *J* = 5.2, 1.0 Hz, 1H), 4.56 (dd, *J* = 5.3, 1.0 Hz, 2H), 3.50 (d, *J* = 1.0 Hz, 6H), 2.64 (d, *J* = 1.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 161.26, 146.92, 146.28, 129.20, 127.74, 125.48, 123.86, 123.61, 112.98, 101.87, 64.30, 53.93, 18.65. HRMS (ESI) [M+H]⁺ found 248.1277, calcd for C₁₄H₁₈NO₃ 248.1281.

2-(2,2-Dimethoxyethoxy)-8-methylquinoline (4B). Following the procedure of **4** starting from 2-chloro-8-methylquinoline, the product **4B** was obtained as a white solid in 64% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.00 (d, *J* = 8.8 Hz, 1H), 7.59 (d, *J* = 8.1 Hz, 1H), 7.53 – 7.49 (m, 1H), 7.33 – 7.29 (m, 1H), 6.99 (d, *J* = 8.8 Hz, 1H), 4.92 (t, *J* = 5.4 Hz, 1H), 4.59 (d, *J* = 5.4 Hz, 2H), 3.52 (s, 6H), 2.71 (s, 3H). HRMS (ESI) [M+H]⁺ found 248.1277, calcd for C₁₄H₁₈NO₃ 248.1281.

6-Chloro-2-(2,2-dimethoxyethoxy)quinoline (4C). Following the procedure of **4** starting from 2,6-dichloroquinoline, the product **4C** was obtained as a white solid in 81% yield. ¹H NMR (400 MHz, CDCl₃) δ 7.93 (d, *J* = 8.8 Hz, 1H), 7.78 (d, *J* = 8.9 Hz, 1H), 7.72 (d, *J* = 2.3 Hz, 1H), 7.57 (dd, *J* = 8.9, 2.4 Hz, 1H), 7.01 (d, *J* = 8.9 Hz, 1H), 4.85 (t, *J* = 5.3 Hz, 1H), 4.55 (d, *J* = 5.3 Hz, 2H), 3.50 (s, 6H). HRMS (ESI) [M+H]⁺ found 268.0734, calcd for C₁₃H₁₅ClNO₃ 268.0735.

8-Bromo-2-(2,2-dimethoxyethoxy)quinoline (4D). Following the procedure of **4** starting from 8-bromo-2-chloroquinoline and heating to 80 °C, the product **4D** was obtained as a white solid in 50% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.04 – 8.00 (m, 1H), 7.98 – 7.96 (m, 1H), 7.73 – 7.70 (m, 1H), 7.27 – 7.25 (m, 1H), 7.04 (d, *J* = 8.8 Hz, 1H), 4.99 (d, *J* = 5.4 Hz, 1H), 4.66 (d, *J* = 5.4 Hz, 2H), 3.53 (s, 6H). HRMS (ESI) [M+H]⁺ found 312.016, calcd for C₁₃H₁₅BrNO₃ 312.018.



2-(2,2-Dimethoxyethoxy)-6-(4-methoxyphenyl)quinoline (4G). (4-methoxyphenyl)boronic acid (73 mg, 0.64 mmol), 6-bromo-2-(2,2-dimethoxyethoxy)quinoline (100 mg, 0.32 mmol), Pd(OAc)₂ (1.5 mg, 0.64 mmol%), Xphos (12 mg, 2.56 mmol%) and K₃PO₄ (136 mg, 0.64 mmol)

were charged into a 50 ml round-bottom flask containing toluene (20 ml). The reaction was nitrogen-flushed and then stirred at 100 °C overnight. Upon completion, the reaction mixture was cooled and evaporated to dryness. The residue was diluted in water and extracted twice with EtOAc, washed with brine and dried over Na₂SO₄, and purified by column chromatography to afford **4G** as white solid in 67% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.02 (d, *J* = 8.8 Hz, 1H), 7.89 – 7.83 (m, 3H), 7.62 (d, *J* = 8.8 Hz, 2H), 7.02 (d, *J* = 8.8 Hz, 2H), 6.98 (d, *J* = 8.8 Hz, 1H), 4.85 (t, *J* = 5.3 Hz, 1H), 4.57 (d, *J* = 5.3 Hz, 2H), 3.87 (s, 3H), 3.50 (s, 6H). HRMS (ESI) [M+H]⁺ found 340.147, calcd for C₂₀H₂₂NO₄ 340.150.

2-(2,2-Dimethoxyethoxy)-6-(4-(trifluoromethyl)phenyl)quinoline (4H). Following the procedure of **4G**, the product **4H** was obtained as a white solid in 90% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.11 – 8.01 (m, 1H), 7.97 – 7.83 (m, 3H), 7.83 – 7.68 (m, 4H), 7.07 – 6.97 (m, 1H), 4.90 – 4.80 (m, 1H), 4.63 – 4.52 (m, 2H), 3.52 (s, 3H), 3.49 (s, 3H). HRMS (ESI) [M+H]⁺ found 378.1315, calcd for C₂₀H₁₉F₃NO₃ 378.1312.

4-(2-(2,2-Dimethoxyethoxy)quinolin-6-yl)benzonitrile (4I). Following the procedure of **4G**, the product **4I** was obtained as a white solid in 85% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.05 (d, *J* = 8.9 Hz, 1H), 7.93 (s, 1H), 7.92 (d, *J* = 4.8 Hz, 1H), 7.87 – 7.83 (m, 1H), 7.80 – 7.73 (m, 4H), 7.02 (d, *J* = 8.8 Hz, 1H), 4.85 (t, *J* = 5.3 Hz, 1H), 4.57 (d, *J* = 5.3 Hz, 2H), 3.50 (s, 6H). HRMS (ESI) [M+H]⁺ found 335.1324, calcd for C₂₀H₁₉N₂O₃ 335.1326.

Methyl 4-(2-(2,2-dimethoxyethoxy)quinolin-6-yl)benzoate (4J). Following the procedure of **4G**, the product **4J** was obtained as a white solid in 88% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.14 (d, *J* = 8.4 Hz, 2H), 8.05 (d, *J* = 8.9 Hz, 1H), 7.95 (s, 1H), 7.92 – 7.89 (m, 2H), 7.75 (d, *J* = 8.4 Hz, 2H), 7.01 (d, *J* = 8.8 Hz, 1H), 4.85 (t, *J* = 5.3 Hz, 1H), 4.57 (d, *J* = 5.3 Hz, 2H), 3.95 (s, 3H), 3.50 (s, 6H). HRMS (ESI) [M+H]⁺ found 368.1423, calcd for C₂₁H₂₂NO₅ 368.1424.

2-(2,2-Dimethoxyethoxy)-7-(2-fluorophenyl)-4-methylquinoline (4K). Following the procedure of **4G**, the product **4K** was obtained as a white solid in 87% yield. ¹H NMR (400 MHz, CDCl₃) δ 8.05 (s, 1H), 7.94 (d, *J* = 8.5 Hz, 1H), 7.65 (d, *J* = 8.5 Hz, 1H), 7.60 (t, *J* = 7.7 Hz, 1H), 7.42 – 7.35 (m, 1H), 7.29 (d, *J* = 7.3 Hz, 1H), 7.25 – 7.19 (m, 1H), 6.87 (s, 1H), 4.87 (t, *J* = 5.2 Hz, 1H), 4.58 (d, *J* = 5.2 Hz, 2H), 3.51 (s, 6H), 2.66 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 157.3, 156.5, 142.4, 142.0, 132.5, 126.6, 124.9, 124.3, 123.5, 120.6, 120.4, 120.1, 119.3, 111.7, 108.9, 97.5, 60.1, 49.6, 14.2. HRMS (ESI) [M+H]⁺ found 342.1499, calcd for C₂₀H₂₁FNO₃ 342.15.

2.3 Experimental data

N-methyl-2-(quinolin-2-ylamino)acetamide (6). 2-(2,2-dimethoxyethoxy)quinoline (200 mg, 0.86 mmol) reacted with hydrochloric acid in diethyl ether (5 mL). After that the solvent was evaporated under anhydrous atmosphere. The residue was dissolved in anhydrous n-butanol (10 mL) directly without further purification and treated with 2-amino-N-methylacetamide (151 mg, 1.71 mmol) and Et₃N (237 μL, 1.71 mmol). After stirring overnight at room temperature, the reaction mixture was concentrated. H₂O was added to the reaction mixture and the mixture was extracted twice with EtOAc. The combined organic layer was washed with brine, dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by reverse-phase silica gel column chromatography (MeOH/H₂O = 0% to 80%) to give **6** as a white solid in 95% yield (172 mg). ¹H NMR (400 MHz, CDCl₃) δ 7.86 (d, *J* = 8.9 Hz, 1H), 7.72 (d, *J* = 8.4 Hz, 1H), 7.63 (d, *J* = 8.0 Hz, 1H), 7.57 (t, *J* = 7.5 Hz, 1H), 7.31 – 7.25 (m, 1H), 6.96 (s, 1H), 6.71 (d, *J* = 8.9 Hz, 1H), 5.67 (s, 1H), 4.24 (s, 2H), 2.84 (d, *J* = 4.9 Hz, 3H), 2.24 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 171.3,

156.0, 147.4, 137.8, 129.8, 127.5, 126.2, 123.7, 122.8, 111.9, 45.9, 26.2. HRMS (ESI) [M+H]⁺ found m/z 216.1006, calcd for C₁₂H₁₄N₃O 215.1058.

Alternative quaternary ammonium salt preparation method: Concentrated sulfuric acid is added dropwise to the sodium chloride solid to produce hydrogen chloride gas. This gas is then passed to the acetone solution containing the 2-(2,2-dimethoxyethoxy)quinoline for reaction. After the reaction is completed, the solvent is distilled for recovery. The obtained residue is a quaternary ammonium salt for further amino coupling reaction.

(R)-N-methyl-2-(quinolin-2-ylamino)propanamide (6a). Following the procedure of **6** starting from (S)-2-amino-N-methylpropanamide, the product **6a** was obtained as a white solid (yield 92% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.80 (d, *J* = 8.8 Hz, 1H), 7.67 (d, *J* = 8.3 Hz, 1H), 7.59 (d, *J* = 7.9 Hz, 1H), 7.53 (t, *J* = 7.3 Hz, 1H), 7.27 – 7.21 (m, 1H), 7.10 (s, 1H), 6.61 (d, *J* = 8.8 Hz, 1H), 5.28 (d, *J* = 5.9 Hz, 1H), 4.72 (p, *J* = 6.7 Hz, 1H), 2.78 (d, *J* = 4.9 Hz, 3H), 1.52 (d, *J* = 7.0 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 174.3, 155.8, 147.4, 137.6, 129.7, 127.5, 126.1, 123.6, 122.6, 112.0, 50.7, 26.1, 18.1. HRMS (ESI) [M+H]⁺ found m/z 230.1284, calcd for C₁₃H₁₆N₃O 230.1288. [α]_D²⁰ -141.5 (c = 0.188, CH₃OH).

(S)-N,3,3-trimethyl-2-(quinolin-2-ylamino)butanamide (6b). Following the procedure of **6** starting from 2-amino-N,3,3-trimethylbutanamide, the product **6b** was obtained as a white solid (yield 66% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.82 (d, *J* = 8.9 Hz, 1H), 7.69 (d, *J* = 8.4 Hz, 1H), 7.61 (s, 1H), 7.57 – 7.52 (m, 1H), 7.25 (t, *J* = 8.0 Hz, 1H), 6.70 (d, *J* = 8.9 Hz, 1H), 6.44 (s, 1H), 5.68 (s, 1H), 4.50 (d, *J* = 7.8 Hz, 1H), 2.80 (d, *J* = 4.9 Hz, 3H), 1.14 (s, 9H). ¹³C NMR (125 MHz, CDCl₃) δ 172.6, 156.3, 147.1, 137.8, 129.7, 127.5, 125.7, 123.6, 122.5, 112.0, 34.4, 27.0, 26.1. HRMS (ESI) [M+H]⁺ found m/z 272.1756, calcd for C₁₆H₂₂N₃O 272.1757. [α]_D²⁰ -178.7 (c = 0.075, CH₃OH).

(S)-N-methyl-2-phenyl-2-(quinolin-2-ylamino)acetamide (6c). Following the procedure of **6** starting from (S)-2-amino-N-methyl-2-phenylacetamide, the product **6c** was obtained as a white solid (yield 63% for 2 steps). ¹H NMR (400 MHz, MeOD-*d*₄) δ 7.88 (d, *J* = 8.9 Hz, 1H), 7.66 – 7.60 (m, 2H), 7.57 – 7.47 (m, 3H), 7.42 – 7.36 (m, 2H), 7.36 – 7.30 (m, 1H), 5.69 (s, 1H), 2.77 (s, 3H). ¹³C NMR (125 MHz, MeOD-*d*₄) δ 173.8, 156.1, 147.5, 138.6, 136.8, 128.9, 128.3, 127.7, 127.5, 127.1, 125.6, 123.7, 122.0, 112.4, 59.7, 25.1. HRMS (ESI) [M+H]⁺ found m/z 292.145, calcd for C₁₈H₁₈N₃O 292.144. [α]_D²⁰ -4.0 (c = 0.121, CH₃OH).

(S)-N-methyl-3-phenyl-2-(quinolin-2-ylamino)propanamide (6d). Following the procedure of **6** starting from (S)-2-amino-N-methyl-3-phenylpropanamide, the product **6d** was obtained as a white solid (yield 72% for 2 steps). ¹H NMR (400 MHz, Acetone-*d*₆) δ 7.86 (s, 1H), 7.63 (d, *J* = 8.3 Hz, 2H), 7.50 (t, *J* = 7.5 Hz, 1H), 7.41 (s, 1H), 7.33 (d, *J* = 7.5 Hz, 2H), 7.25 (t, *J* = 7.5 Hz, 2H), 7.17 (s, 2H), 6.88 (d, *J* = 8.9 Hz, 1H), 6.50 (s, 1H), 5.11 – 5.02 (m, 1H), 3.30 (dd, *J* = 13.8, 5.9 Hz, 1H), 3.13 (dd, *J* = 13.7, 7.8 Hz, 1H), 2.70 (d, *J* = 4.7 Hz, 3H). ¹³C NMR (125 MHz, Acetone-*d*₆) δ 205.2, 172.3, 156.2, 147.8, 138.5, 136.7, 129.3, 129.0, 128.1, 127.4, 126.2, 126.1, 123.6, 121.8, 113.1, 56.0, 38.0, 25.2. HRMS (ESI) [M+H]⁺ found m/z 306.1597, calcd for C₁₉H₂₀N₃O 306.1601. [α]_D²⁰ 29.9 (c = 0.126, CH₃OH).

Methyl quinolin-2-ylphenylalaninate (6e). Following the procedure of **6** starting from methyl L-phenylalaninate, the product **6e** was obtained as a white solid (yield 61% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.82 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.75 (d, *J* = 8.1 Hz, 1H), 7.62 (d, *J* = 7.7 Hz, 1H), 7.60 – 7.54 (m, 1H), 7.35 – 7.26 (m, 4H), 7.21 (d, *J* = 7.3 Hz, 2H), 6.65 (dd, *J* = 8.8, 2.7 Hz, 1H), 5.27 – 5.21 (m, 1H), 5.19 (s, 1H), 3.77 (d, *J* = 2.8 Hz, 3H), 3.39 (ddd, *J* = 13.7, 5.5, 2.4 Hz, 1H), 3.27 (ddd, *J* = 13.9, 5.7, 2.5 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 173.3, 155.1, 147.1, 137.6, 136.6, 130.4, 129.6, 129.4, 128.5, 127.4, 126.9, 126.2, 123.6, 122.6, 112.1, 55.1, 20

52.2. HRMS (ESI) [M+H]⁺ found *m/z* 307.1437, calcd for C₁₉H₁₉N₂O₂ 307.1441. [α]_D -2.9 (c = 0.105, CH₃OH).

(R)-2-(quinolin-2-ylamino)butanamide (6f). Following the procedure of **6** starting from 2-aminobutanamide, the product **6f** was obtained as a white solid (yield 85% for 2 steps). ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.85 (d, *J* = 8.9 Hz, 1H), 7.62 (d, *J* = 7.9 Hz, 1H), 7.48 – 7.43 (m, 3H), 7.15 (ddd, *J* = 8.1, 4.8, 3.2 Hz, 1H), 7.06 (d, *J* = 7.8 Hz, 1H), 6.99 (s, 1H), 6.93 (d, *J* = 8.9 Hz, 1H), 4.54 (q, *J* = 6.9 Hz, 1H), 1.90 – 1.76 (m, 1H), 1.76 – 1.63 (m, 1H), 0.94 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 175.1, 157.0, 148.0, 136.6, 129.4, 127.9, 126.0, 123.5, 121.7, 113.9, 55.4, 25.8, 10.8. HRMS (ESI) [M+H]⁺ found *m/z* 230.1288, calcd for C₁₃H₁₆N₃O 230.1288. [α]_D 20

-66.3 (c = 0.080, CH₃OH).

Methyl quinolin-2-ylglycinate (6g). Following the procedure of **6** starting from methyl glycinate, the product **6g** was obtained as a white solid (yield 95% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.83 (d, *J* = 8.9 Hz, 1H), 7.71 (d, *J* = 8.4 Hz, 1H), 7.60 (d, *J* = 7.9 Hz, 1H), 7.57 – 7.51 (m, 1H), 7.23 (t, 1H), 6.72 (d, 1H), 5.24 (s, 1H), 4.38 (d, *J* = 5.1 Hz, 2H), 3.80 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 171.8, 155.7, 147.6, 137.4, 129.5, 127.4, 126.4, 123.7, 122.5, 112.1, 52.3, 43.3, 29.7. HRMS (ESI) [M+H]⁺ found *m/z* 217.0973, calcd for C₁₂H₁₃N₂O₂ 217.0972.

Methyl quinolin-2-yl-L-alaninate (6h). Following the procedure of **6** starting from methyl D-alaninate, the product **6h** was obtained as a white solid (yield 93% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.81 (d, *J* = 8.8 Hz, 1H), 7.68 (d, *J* = 8.3 Hz, 1H), 7.59 (d, *J* = 8.8 Hz, 1H), 7.53 (t, *J* = 7.7 Hz, 1H), 7.25 – 7.21 (m, 1H), 6.67 (d, *J* = 8.8 Hz, 1H), 5.27 (s, 1H), 4.90 (p, *J* = 7.1 Hz, 1H), 3.77 (s, 3H), 1.56 (d, *J* = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 175.0, 155.3, 147.4, 137.4, 129.5, 127.4, 126.4, 123.6, 122.5, 112.0, 52.3, 49.7, 18.6. HRMS (ESI) [M+H]⁺ found *m/z* 20

231.1129, calcd for C₁₃H₁₅N₂O₂ 231.1128. [α]_D -23.7 (c = 0.114, CH₃OH).

Methyl quinolin-2-yl-L-valinate (6i). Following the procedure of **6** starting from methyl D-valinate, the product **6i** was obtained as a white solid (yield 91% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.76 (d, *J* = 8.8 Hz, 1H), 7.67 (d, *J* = 8.4 Hz, 1H), 7.56 (d, *J* = 8.8 Hz, 1H), 7.54 – 7.49 (m, 1H), 7.24 – 7.19 (m, 1H), 6.67 (d, *J* = 8.8 Hz, 1H), 5.21 (d, *J* = 7.5 Hz, 1H), 4.87 (dd, *J* = 8.3, 5.4 Hz, 1H), 3.75 (s, 3H), 1.05 (dd, *J* = 6.8, 3.4 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 174.1, 156.1, 147.7, 137.2, 129.4, 127.3, 126.5, 123.7, 122.3, 112.1, 59.0, 51.9, 31.3, 19.1, 18.5. [α]_D 20

-145.6 (c = 0.148, CH₃OH).

Methyl quinolin-2-yl-L-isoleucinate (6j). Following the procedure of **6** starting from methyl D-isoleucinate, the product **6j** was obtained as a white solid (yield 82% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.84 (dd, *J* = 8.9, 2.2 Hz, 1H), 7.71 (d, *J* = 8.4 Hz, 1H), 7.59 (d, *J* = 7.9 Hz, 1H),

7.54 (t, $J = 7.7$ Hz, 1H), 7.23 (t, $J = 7.3$ Hz, 1H), 6.71 (dd, $J = 8.9, 2.0$ Hz, 1H), 5.02 (s, 1H), 4.83 (s, 1H), 3.77 (s, 3H), 2.14 – 2.01 (m, 1H), 1.70 – 1.53 (m, 1H), 1.03 (d, $J = 3.0$ Hz, 3H), 1.02 – 0.99 (m, 3H). HRMS (ESI) $[M+H]^+$ found m/z 273.1603, calcd for $C_{16}H_{21}N_2O_2$ 273.1598. $[\alpha]_{D}^{20}$ -8.1 (c = 0.078, CH_3OH).

Methyl quinolin-2-yl-L-serinate (6k). Following the procedure of **6** starting from methyl D-serinate, the product **6k** was obtained as a white solid (yield 88% for 2 steps). 1H NMR (400 MHz, $CDCl_3$) δ 7.86 (d, $J = 8.9$ Hz, 1H), 7.68 (d, $J = 8.4$ Hz, 1H), 7.62 (d, $J = 7.9$ Hz, 1H), 7.59 – 7.53 (m, 1H), 7.28 – 7.25 (m, 1H), 6.77 (d, $J = 8.9$ Hz, 1H), 5.87 (s, 1H), 4.97 (s, 1H), 4.26 (dd, $J = 10.9, 2.6$ Hz, 1H), 3.98 (dd, $J = 10.9, 6.2$ Hz, 1H), 3.85 (s, 3H). ^{13}C NMR (125 MHz, $CDCl_3$) δ 171.8, 155.6, 146.2, 138.2, 130.0, 127.5, 125.6, 123.5, 123.0, 112.5, 65.5, 58.0, 52.9. HRMS (ESI) $[M+H]^+$ found m/z 247.1081, calcd for $C_{13}H_{15}N_2O_3$ 247.1077. $[\alpha]_{D}^{20}$ 5.8 (c = 0.136, CH_3OH).

Methyl quinolin-2-yl-L-threoninate (6l). Following the procedure of **6** starting from methyl L-threoninate, the product **6l** was obtained as a white solid (yield 66% for 2 steps). 1H NMR (400 MHz, $CDCl_3$) δ 7.86 (d, $J = 8.8$ Hz, 1H), 7.68 (d, $J = 8.3$ Hz, 1H), 7.62 (dd, $J = 8.0, 1.2$ Hz, 1H), 7.55 (ddd, $J = 8.4, 7.0, 1.5$ Hz, 1H), 7.28 – 7.24 (m, 1H), 6.77 (d, $J = 8.8$ Hz, 1H), 5.51 (s, 1H), 4.96 (dd, $J = 7.3, 3.8$ Hz, 1H), 4.40 (qd, $J = 6.4, 3.7$ Hz, 1H), 3.82 (s, 3H), 1.35 (d, $J = 6.4$ Hz, 3H). ^{13}C NMR (125 MHz, $CDCl_3$) δ 172.8, 156.0, 147.1, 137.6, 129.6, 127.4, 126.3, 123.8, 122.7, 112.4, 69.3, 59.9, 52.5, 20.4. HRMS (ESI) $[M+H]^+$ found m/z 261.1228, calcd for $C_{14}H_{17}N_2O_3$ 261.1234. $[\alpha]_{D}^{20}$ -113.8 (c = 0.052, CH_3OH).

Methyl quinolin-2-yl-L-tyrosinate (6m). Following the procedure of **6** starting from methyl L-tyrosinate, the product **6m** was obtained as a white solid (yield 63% for 2 steps). 1H NMR (400 MHz, $CDCl_3$) δ 7.85 (d, $J = 8.9$ Hz, 1H), 7.73 (d, $J = 8.5$ Hz, 1H), 7.61 (dd, $J = 8.0, 1.2$ Hz, 1H), 7.54 (ddd, $J = 8.4, 7.0, 1.5$ Hz, 1H), 7.26 (ddd, $J = 8.0, 7.0, 1.1$ Hz, 1H), 6.97 (d, $J = 8.5$ Hz, 2H), 6.68 (d, $J = 8.5$ Hz, 2H), 6.64 (d, $J = 8.9$ Hz, 1H), 5.47 (s, 1H), 5.03 (s, 1H), 3.75 (s, 3H), 3.25 (dd, $J = 13.9, 5.3$ Hz, 1H), 3.13 (dd, $J = 13.9, 6.4$ Hz, 1H). ^{13}C NMR (125 MHz, $CDCl_3$) δ 173.6, 155.5, 155.5, 147.0, 138.1, 130.4, 129.9, 127.5, 127.3, 125.8, 123.6, 122.7, 115.6, 111.5, 55.5, 52.3, 37.2. HRMS (ESI) $[M+H]^+$ found m/z 323.1388, calcd for $C_{19}H_{19}N_2O_3$ 323.1390. $[\alpha]_{D}^{20}$ 12.4 (c = 0.106, CH_3OH).

Methyl quinolin-2-yl-L-tryptophanate (6n). Following the procedure of **6** starting from methyl L-tryptophanate, the product **6n** was obtained as a white solid (yield 70% for 2 steps). 1H NMR (400 MHz, $CDCl_3$) δ 8.27 (s, 1H), 7.80 (d, $J = 8.9$ Hz, 1H), 7.76 (d, $J = 8.4$ Hz, 1H), 7.61 (d, $J = 7.9$ Hz, 2H), 7.59 – 7.54 (m, 1H), 7.35 (d, $J = 8.1$ Hz, 1H), 7.28 – 7.24 (m, 1H), 7.20 (t, $J = 7.1$ Hz, 1H), 7.13 (t, $J = 7.5$ Hz, 1H), 7.01 (d, $J = 2.1$ Hz, 1H), 6.57 (d, $J = 8.8$ Hz, 1H), 5.34 – 5.29 (m, 1H), 5.27 (d, $J = 8.2$ Hz, 1H), 3.73 (s, 3H), 3.54 (dd, $J = 14.6, 5.2$ Hz, 1H), 3.45 (dd, $J = 14.6, 5.6$ Hz, 1H). ^{13}C NMR (125 MHz, $CDCl_3$) δ 173.9, 155.4, 147.5, 137.4, 136.2, 129.5, 127.8, 127.4, 126.4, 123.6, 122.9, 122.5, 122.1, 119.6, 118.8, 112.2, 111.2, 110.7, 54.6, 52.2, 27.9. HRMS (ESI) $[M+H]^+$ found m/z 346.1560, calcd for $C_{21}H_{20}N_3O_2$ 346.1550. $[\alpha]_{D}^{20}$ 2.5 (c = 0.120, CH_3OH).

Methyl quinolin-2-yl-D-methioninate (6p). Following the procedure of **6** starting from methyl L-methioninate, the product **6p** was obtained as a white solid (yield 96% for 2 steps). 1H

NMR (400 MHz, CDCl₃) δ 7.82 (d, J = 8.8 Hz, 1H), 7.68 (d, J = 8.4 Hz, 1H), 7.59 (d, J = 7.9 Hz, 1H), 7.53 (t, J = 7.7 Hz, 1H), 7.24 (t, J = 7.4 Hz, 1H), 6.70 (d, J = 8.8 Hz, 1H), 5.42 (s, 1H), 5.06 (q, J = 7.2 Hz, 1H), 3.78 (s, 3H), 2.65 (td, J = 7.9, 3.4 Hz, 2H), 2.35 (dq, J = 13.5, 7.4 Hz, 1H), 2.16 (dd, J = 14.3, 7.5 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 173.9, 155.4, 147.3, 137.5, 129.6, 127.4, 126.4, 123.7, 122.6, 112.0, 53.2, 52.4, 32.0, 30.3, 15.5. HRMS (ESI) [M+H]⁺ found m/z 291.1167, calcd for C₁₅H₁₉N₂O₂S 291.1162. [α] -4.2 (c = 0.095, CH₃OH).

Methyl N⁶-(quinolin-2-yl)lysinate (6r). Following the procedure of **6** starting from methyl lysinate, the product **6r** was obtained as a white solid (yield 53% for 2 steps). ¹H NMR (400 MHz, D₂O) δ 7.94 (s, 1H), 7.63 (t, 1H), 7.59 (d, J = 7.5 Hz, 1H), 7.53 (s, 1H), 7.35 (t, J = 7.6 Hz, 1H), 6.76 (s, 1H), 4.06 (t, J = 6.4 Hz, 1H), 3.70 (s, 3H), 3.33 (t, J = 6.7 Hz, 2H), 2.00 – 1.80 (m, 2H), 1.67 (p, J = 7.2 Hz, 2H), 1.55 – 1.34 (m, 2H). ¹³C NMR (125 MHz, D₂O) δ 170.6, 152.5, 141.9, 135.6, 132.5, 128.7, 125.4, 121.1, 117.0, 113.8, 53.6, 52.7, 41.6, 29.4, 26.9, 21.7. HRMS (ESI) [M+H]⁺ found m/z 288.17, calcd for C₁₆H₂₂N₃O₂ 288.1707.

2-Amino-N-methyl-6-(quinolin-2-ylamino)hexanamide (6s). Following the procedure of **6** starting from 2-(2,2-dimethoxyethoxy)-4-methylquinoline, the product **6A** was obtained as a white solid (yield 50% for 2 steps). ¹H NMR (500 MHz, CDCl₃) δ 7.80 (d, J = 8.9 Hz, 1H), 7.65 (d, J = 8.4 Hz, 1H), 7.57 (d, J = 7.9 Hz, 1H), 7.54 – 7.46 (m, 1H), 7.19 (t, J = 7.7 Hz, 1H), 6.63 (d, J = 8.9 Hz, 1H), 3.56 – 3.49 (m, 2H), 3.40 – 3.33 (m, 1H), 2.80 (d, J = 4.9 Hz, 3H), 1.73 – 1.67 (m, 2H), 1.61 – 1.57 (m, 2H), 1.54 – 1.48 (m, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 175.5, 156.9, 147.8, 137.3, 129.5, 127.4, 125.8, 123.2, 121.9, 111.2, 77.2, 76.9, 76.7, 55.0, 41.3, 34.7, 29.3, 25.7, 23.1. HRMS (ESI) [M+H]⁺ found m/z 287.1864, calcd for C₁₆H₂₃N₄O 287.1866.

N-methyl-2-((4-methylquinolin-2-yl)amino)acetamide (6A). Following the procedure of **6** starting from 2-(2,2-dimethoxyethoxy)-4-methylquinoline, the product **6A** was obtained as a white solid (yield 91% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.81 (s, 1H), 7.73 (s, 1H), 7.58 (s, 1H), 7.30 (s, 1H), 7.05 (s, 1H), 6.55 (s, 1H), 5.63 (s, 1H), 4.21 (s, 2H), 2.83 (d, J = 4.5 Hz, 3H), 2.56 (s, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.9, 156.7, 147.8, 144.1, 129.3, 126.4, 124.2, 123.8, 121.9, 113.5, 44.3, 26.0, 18.7. HRMS (ESI) [M+H]⁺ found m/z 230.1291, calcd for C₁₃H₁₆N₃O 230.1288.

N-methyl-2-((8-methylquinolin-2-yl)amino)acetamide (6B). Following the procedure of **6** starting from 2-(2,2-dimethoxyethoxy)-8-methylquinoline, the product **6B** was obtained as a white solid (yield 45% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.85 (d, J = 8.8 Hz, 1H), 7.50 (dd, J = 8.0, 1.4 Hz, 1H), 7.46 (d, J = 7.1 Hz, 1H), 7.24 (s, 2H), 7.19 (dd, J = 7.9, 7.1 Hz, 1H), 6.72 (d, J = 8.8 Hz, 1H), 5.50 (s, 1H), 4.22 (d, J = 5.2 Hz, 2H), 2.83 (d, J = 4.9 Hz, 3H), 2.65 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 171.9, 155.2, 146.1, 138.0, 134.0, 130.1, 125.5, 123.4, 122.5, 111.6, 46.5, 26.0, 17.9. HRMS (ESI) [M+H]⁺ found m/z 230.1282, calcd for C₁₃H₁₆N₃O 230.1288.

2-((6-Chloroquinolin-2-yl)amino)-N-methylacetamide (6C). Following the procedure of **6** starting from 6-chloro-2-(2,2-dimethoxyethoxy)quinoline, the product **6C** was obtained as a white solid (yield 53% for 2 steps). ¹H NMR (400 MHz, MeOD-*d*₄) δ 7.86 (d, J = 9.0 Hz, 1H), 7.65 (s, 1H), 7.59 (d, J = 9.0 Hz, 1H), 7.46 (d, J = 8.9 Hz, 1H), 6.88 (d, J = 8.8 Hz, 1H), 4.13 (s, 2H), 2.76 (s, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.6, 157.3, 146.6, 135.9, 129.6, 128.0, 126.6, 125.6, 124.4, 114.9, 44.3, 26.0. HRMS (ESI) [M+H]⁺ found m/z 250.0739, calcd for C₁₂H₁₃ClN₃O 250.0742.

2-((8-Bromoquinolin-2-yl)amino)-N-methylacetamide (6D). Following the procedure of **6** starting from 8-bromo-2-(2,2-dimethoxyethoxy)quinoline, the product **6D** was obtained as a white solid (yield 42% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.90 (dd, *J* = 7.6, 1.2 Hz, 1H), 7.81 (d, *J* = 8.9 Hz, 1H), 7.59 (dd, *J* = 7.9, 1.1 Hz, 1H), 7.13 (t, *J* = 7.8 Hz, 1H), 6.76 (d, *J* = 8.9 Hz, 1H), 6.10 (s, 1H), 4.23 (d, *J* = 6.0 Hz, 2H), 2.85 (d, *J* = 4.9 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 171.5, 156.7, 144.4, 137.9, 133.1, 127.3, 124.8, 123.0, 121.2, 113.2, 46.8, 26.0. HRMS (ESI) [M+H]⁺ found *m/z* 294.0245, calcd for C₁₂H₁₃BrN₃O 294.0237.

2-((3-Chloroisoquinolin-1-yl)amino)-N-methylacetamide (6E). Following the procedure of **6** starting from 3-chloro-1-(2,2-dimethoxyethoxy)isoquinoline, the product **6E** was obtained as a white solid (yield 52% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.86 (d, *J* = 8.4 Hz, 1H), 7.65 – 7.58 (m, 2H), 7.50 – 7.42 (m, 1H), 7.01 (s, 1H), 6.49 (s, 1H), 6.39 (s, 1H), 4.29 (d, *J* = 5.0 Hz, 2H), 2.91 (d, *J* = 4.8 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 170.5, 154.7, 143.8, 138.7, 130.8, 126.4, 126.2, 122.0, 116.6, 109.4, 45.4, 26.3. HRMS (ESI) [M+H]⁺ found *m/z* 250.0739, calcd for C₁₂H₁₃ClN₃O 250.0742.

N-methyl-2-((6-phenylquinolin-2-yl)amino)acetamide (6F). Following the procedure of **6** starting from 2-(2,2-dimethoxyethoxy)-6-phenylquinoline, the product **6F** was obtained as a white solid (yield 75% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.90 (d, *J* = 8.8 Hz, 1H), 7.82 (t, 2H), 7.76 (d, *J* = 9.1 Hz, 1H), 7.67 (d, *J* = 8.0 Hz, 2H), 7.47 (t, *J* = 7.6 Hz, 2H), 7.36 (t, *J* = 7.4 Hz, 1H), 6.83 (s, 1H), 6.72 (d, *J* = 8.8 Hz, 1H), 5.50 (s, 1H), 4.23 (d, *J* = 5.1 Hz, 2H), 2.84 (d, *J* = 4.9 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 171.2, 156.0, 146.7, 140.7, 138.0, 135.7, 129.3, 128.9, 127.1, 127.1, 126.5, 125.4, 123.9, 112.3, 45.9, 26.2. HRMS (ESI) [M+H]⁺ found *m/z* 292.1449, calcd for C₁₈H₁₈N₃O 292.1444.

2-((6-(4-Methoxyphenyl)quinolin-2-yl)amino)-N-methylacetamide (6G). Following the procedure of **6** starting from 2-(2,2-dimethoxyethoxy)-6-(4-methoxyphenyl)quinoline, the product **6G** was obtained as a white solid (yield 54% for 2 steps). ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.91 (d, *J* = 9.0 Hz, 1H), 7.87 (d, *J* = 1.9 Hz, 1H), 7.75 (dd, *J* = 8.7, 2.0 Hz, 1H), 7.65 (d, *J* = 8.7 Hz, 2H), 7.51 (d, 1H), 7.28 (t, *J* = 5.5 Hz, 1H), 7.01 (d, *J* = 8.7 Hz, 2H), 6.89 (d, *J* = 8.9 Hz, 1H), 4.00 (d, *J* = 5.5 Hz, 2H), 3.78 (s, 3H), 2.60 (d, *J* = 4.0 Hz, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.8, 159.0, 157.0, 147.1, 136.9, 133.3, 132.8, 128.1, 128.0, 126.6, 124.7, 123.8, 114.8, 114.1, 55.6, 44.5, 26.0. HRMS (ESI) [M+H]⁺ found *m/z* 322.1549, calcd for C₁₉H₂₀N₃O₂ 322.155.

N-methyl-2-((6-(4-(trifluoromethyl)phenyl)quinolin-2-yl)amino)acetamide (6H). Following the procedure of **6** starting from 2-(2,2-dimethoxyethoxy)-6-(4-(trifluoromethyl)phenyl)quinoline, the product **6H** was obtained as a white solid (yield 65% for 2 steps). ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.06 (d, *J* = 1.8 Hz, 1H), 7.96 (t, 3H), 7.86 (dd, *J* = 8.7, 2.0 Hz, 1H), 7.80 (d, *J* = 8.3 Hz, 2H), 7.57 (d, 1H), 7.41 (t, *J* = 5.8 Hz, 1H), 6.93 (d, *J* = 8.9 Hz, 1H), 4.02 (d, *J* = 5.4 Hz, 2H), 2.60 (d, *J* = 3.9 Hz, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.6, 157.5, 148.1, 144.4, 137.1, 131.7, 128.3, 127.5, 126.8, 126.3, 126.2, 126.2, 123.8, 114.5, 44.4, 26.0. HRMS (ESI) [M+H]⁺ found *m/z* 360.1317, calcd for C₁₉H₁₇FN₃O 360.1318.

2-((6-(4-Cyanophenyl)quinolin-2-yl)amino)-N-methylacetamide (6I). Following the procedure of **6** starting from 4-(2-(2,2-dimethoxyethoxy)quinolin-6-yl)benzotrile, the product **6I** was obtained as a white solid (yield 78% for 2 steps). ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.11 (s, 1H), 7.94 (dt, *J* = 19.1, 9.6 Hz, 7H), 7.59 (d, *J* = 8.8 Hz, 1H), 7.49 (t, *J* = 5.6 Hz, 1H), 6.96 (d, *J* = 9.1 Hz, 1H), 4.04 (d, *J* = 5.2 Hz, 2H), 2.62 (d, *J* = 4.4 Hz, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ

170.6, 157.6, 148.3, 144.9, 137.1, 133.3, 131.3, 128.2, 127.6, 126.9, 126.5, 123.7, 119.5, 114.5, 109.7, 44.4, 26.0. HRMS (ESI) [M+H]⁺ found m/z 317.1401, calcd for C₁₉H₁₇N₄O 317.1397.

Methyl 4-(2-((2-(methylamino)-2-oxoethyl)amino)quinolin-6-yl)benzoate (6J). Following the procedure of **6** starting from methyl 4-(2-(2,2-dimethoxyethoxy)quinolin-6-yl)benzoate, the product **6J** was obtained as a white solid (yield 57% for 2 steps). ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.09 (d, *J* = 2.0 Hz, 1H), 8.05 (d, *J* = 8.4 Hz, 2H), 7.98 (d, *J* = 8.9 Hz, 1H), 7.94 – 7.88 (m, 4H), 7.59 (d, *J* = 8.7 Hz, 1H), 7.44 (t, *J* = 5.5 Hz, 1H), 6.95 (d, *J* = 8.9 Hz, 1H), 4.04 (d, *J* = 5.6 Hz, 2H), 3.88 (s, 3H), 2.62 (d, *J* = 4.6 Hz, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.7, 166.6, 157.5, 148.1, 145.0, 137.1, 131.9, 130.3, 128.3, 128.2, 127.0, 126.8, 126.3, 123.8, 114.4, 56.5, 52.6, 26.0. HRMS (ESI) [M+H]⁺ found m/z 350.1501, calcd for C₂₀H₂₀N₃O₃ 350.1499.

2-((7-(2-Fluorophenyl)-4-methylquinolin-2-yl)amino)-N-methylacetamide (6K). Following the procedure of **6** starting from 2-(2,2-dimethoxyethoxy)-7-(2-fluorophenyl)-4-methylquinoline, the product **6K** was obtained as a white solid (yield 70% for 2 steps). ¹H NMR (400 MHz, MeOD-*d*₄) δ 7.83 (d, *J* = 8.5 Hz, 1H), 7.80 (s, 1H), 7.54 (t, *J* = 7.8 Hz, 1H), 7.41 (d, *J* = 8.5 Hz, 1H), 7.39 – 7.35 (m, 1H), 7.27 (d, *J* = 7.5 Hz, 1H), 7.25 – 7.20 (m, 1H), 6.70 (s, 1H), 4.14 (s, 2H), 2.76 (s, 3H), 2.56 (s, 3H). ¹³C NMR (126 MHz, MeOD) δ 172.4, 160.3, 158.4, 156.5, 146.9, 144.3, 136.1, 130.1, 128.6, 125.4, 123.8, 123.0, 122.7, 122.3, 115.3, 112.2, 43.7, 24.5, 16.8. HRMS (ESI) [M+H]⁺ found m/z 324.1507, calcd for C₁₉H₁₉FN₃O 324.1507.

N-(2-(2,5-dimethoxyphenyl)-2-hydroxyethyl)-2-(quinolin-2-ylamino)acetamide (8a). Following the procedure of **6** starting from midodrine hydrochloride, the product **8a** was obtained as a white solid (yield 72% for 2 steps). ¹H NMR (400 MHz, MeOD-*d*₄) δ 7.88 (d, *J* = 8.9 Hz, 1H), 7.62 (t, *J* = 8.0 Hz, 2H), 7.53 – 7.48 (m, 1H), 7.24 (t, 1H), 7.00 (d, 1H), 6.81 (d, 1H), 6.75 (d, 1H), 6.70 (dd, *J* = 8.9, 3.1 Hz, 1H), 5.08 (dd, *J* = 7.1, 4.3 Hz, 1H), 4.12 (s, 2H), 3.72 (s, 3H), 3.68 (s, 3H), 3.56 (dd, 1H), 3.39 (dd, 1H). ¹³C NMR (125 MHz, MeOD-*d*₄) δ 172.5, 156.8, 153.8, 150.3, 147.3, 137.2, 131.1, 129.1, 127.2, 125.4, 123.7, 122.0, 112.5, 112.3, 111.1, 66.7, 54.9, 54.6, 45.1, 44.6. HRMS (ESI) [M+H]⁺ found m/z 382.1761, calcd for C₂₁H₂₄N₃O₄ 382.1761.

(1S,3S,5S)-2-((1S,3S,5R,7S)-3-hydroxyadamantan-1-yl)-2-(quinolin-2-ylamino)acetyl)-2-azabicyclo[3.1.0]hexane-3-carbonitrile (8b). Following the procedure of **6** starting from saxagliptin, the product **8b** was obtained as a white solid (yield 46% for 2 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.58 – 7.46 (m, 5H), 7.22 (t, *J* = 7.3 Hz, 1H), 6.65 (d, *J* = 8.7 Hz, 1H), 5.94 (d, *J* = 9.5 Hz, 1H), 5.27 (d, *J* = 9.4 Hz, 1H), 5.00 (dd, *J* = 10.6, 2.4 Hz, 1H), 4.63 (td, *J* = 6.2, 2.6 Hz, 1H), 2.75 (s, 1H), 2.51 (ddd, *J* = 13.8, 10.7, 5.8 Hz, 1H), 2.36 (dd, *J* = 13.7, 2.4 Hz, 1H), 2.32 – 2.23 (m, 3H), 2.07 (d, *J* = 11.6 Hz, 1H), 2.00 – 1.92 (m, 2H), 1.86 (dd, *J* = 25.7, 12.0 Hz, 2H), 1.73 (q, *J* = 14.1 Hz, 6H), 1.59 (d, *J* = 14.3 Hz, 3H), 1.28 (s, 2H), 1.18 (s, 1H), 1.17 – 1.11 (m, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 171.7, 156.6, 147.5, 136.9, 129.2, 127.5, 126.1, 123.8, 122.2, 119.5, 113.2, 68.7, 58.9, 46.2, 45.3, 44.6, 44.2, 40.7, 38.3, 38.0, 37.7, 35.5, 30.6, 30.4, 30.3, 17.5, 13.7. HRMS (ESI) [M+H]⁺ found m/z 443.2432, calcd for C₂₇H₃₁N₄O₂ 443.2442. [α]_D²⁰ -

111.0 (c = 0.091, CH₃OH).

Methyl quinolin-2-yl-L-tyrosyl-L-seryl-L-leucinate (8c). Following the procedure of **6** starting from methyl tyrosyleutide, the product **8c** was obtained as a white solid (yield 56% for 2 steps). ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.16 (s, 1H), 8.20 (d, *J* = 7.9 Hz, 1H), 7.98 (d, *J* = 7.9 Hz, 1H), 7.84 (d, *J* = 8.9 Hz, 1H), 7.60 (d, *J* = 7.9 Hz, 1H), 7.50 – 7.42 (m, 2H), 7.21 (d, *J* = 7.6 Hz, 1H), 7.17 (s, 1H), 7.13 (d, *J* = 7.6 Hz, 2H), 6.85 (d, *J* = 8.9 Hz, 1H), 6.62 (d, *J* = 7.6 Hz, 2H), 4.84

(t, $J = 5.0$ Hz, 1H), 4.76 (s, 1H), 4.31 (dd, $J = 14.0, 7.0$ Hz, 2H), 4.15 (d, $J = 5.0$ Hz, 1H), 3.61 (s, 3H), 3.19 – 3.17 (m, 3H), 1.58 – 1.44 (m, 2H), 1.41 – 1.35 (m, 2H), 0.81 – 0.75 (m, 6H). ^{13}C NMR (125 MHz, DMSO- d_6) δ 173.1, 173.1, 170.5, 156.8, 156.1, 147.9, 136.8, 130.6, 129.4, 129.0, 127.9, 126.2, 123.5, 121.9, 115.3, 113.6, 62.1, 56.7, 55.3, 52.3, 50.6, 49.1, 37.2, 24.5, 23.1, 21.7.

HRMS (ESI) $[\text{M}+\text{H}]^+$ found m/z 523.2554, calcd for $\text{C}_{28}\text{H}_{35}\text{N}_4\text{O}_6$ 523.2551. $[\alpha]_D^{20}$ -7.2 (c = 0.107, CH_3OH).

(S)-N-(4-(hydroxymethyl)phenyl)-2-((S)-3-methyl-2-(quinolin-2-ylamino)butanamido)-5-ureidopentanamide (8d). Following the procedure of **6** starting from Val-Cit-PAB-OH, the product **8d** was obtained as a white solid (yield 56% for 2 steps). ^1H NMR (400 MHz, MeOD- d_4) δ 7.87 (d, $J = 8.9$ Hz, 1H), 7.63 – 7.58 (m, 2H), 7.47 (ddd, $J = 8.4, 7.0, 1.4$ Hz, 1H), 7.39 (d, $J = 8.5$ Hz, 2H), 7.25 (d, $J = 8.6$ Hz, 2H), 7.22 – 7.17 (m, 1H), 6.91 (d, $J = 8.9$ Hz, 1H), 4.58 – 4.53 (m, 3H), 4.49 (d, $J = 7.5$ Hz, 1H), 3.37 (s, 2H), 3.09 (dt, $J = 13.3, 6.7$ Hz, 1H), 2.99 (dt, $J = 13.5, 6.7$ Hz, 1H), 2.30 – 2.19 (m, 1H), 1.94 – 1.84 (m, 1H), 1.80 – 1.68 (m, 1H), 1.60 – 1.43 (m, 2H), 1.11 (d, $J = 6.9$ Hz, 3H), 1.09 (d, $J = 6.8$ Hz, 3H). ^{13}C NMR (125 MHz, MeOD- d_4) δ 174.4, 170.9, 160.8, 157.0, 147.4, 137.4, 137.0, 136.9, 129.0, 127.1, 127.1, 125.4, 123.6, 121.9, 120.0, 112.6, 63.4, 60.9, 53.4, 48.4, 30.4, 29.3, 26.3, 18.7, 17.9. HRMS (ESI) $[\text{M}+\text{H}]^+$ found m/z 507.2724,

calcd for $\text{C}_{27}\text{H}_{35}\text{N}_6\text{O}_4$ 507.2714. $[\alpha]_D^{20}$ -186.3 (c = 0.096, CH_3OH).

(S)-2-((S)-2-((7-(2-fluorophenyl)-4-methylquinolin-2-yl)amino)-3-methylbutanamido)-N-(4-(hydroxymethyl)phenyl)-5-ureidopentanamide (8e). Following the procedure of **6** starting from Val-Cit-PAB-OH, the product **8e** was obtained as a white solid (yield 52% for 2 steps). ^1H NMR (500 MHz, MeOD- d_4) δ 7.88 (d, $J = 8.4$ Hz, 1H), 7.80 (s, 1H), 7.51 (t, $J = 7.8$ Hz, 1H), 7.44 (d, $J = 8.4$ Hz, 1H), 7.42 – 7.37 (m, 1H), 7.33 (d, $J = 8.5$ Hz, 2H), 7.28 (t, $J = 7.5$ Hz, 1H), 7.24 – 7.21 (m, 1H), 7.20 (d, $J = 8.4$ Hz, 2H), 6.82 (s, 1H), 4.56 (dd, $J = 9.0, 5.1$ Hz, 1H), 4.52 (s, 2H), 4.48 (d, $J = 7.4$ Hz, 1H), 3.11 – 3.04 (m, 1H), 3.01 – 2.94 (m, 1H), 2.61 (s, 3H), 2.31 – 2.22 (m, 1H), 1.89 (dt, $J = 14.3, 6.6$ Hz, 1H), 1.78 – 1.68 (m, 1H), 1.57 – 1.47 (m, 2H), 1.13 (d, $J = 6.8$ Hz, 3H), 1.11 (d, $J = 6.8$ Hz, 3H). ^{13}C NMR (125 MHz, MeOD- d_4) δ 175.8, 172.3, 162.2, 160.3, 158.7, 148.8, 146.2, 138.8, 138.3, 137.9, 132.1, 130.5, 130.4, 128.4, 127.4, 125.7, 124.8, 124.6, 124.1, 121.5, 117.2, 117.0, 114.2, 64.8, 62.4, 54.8, 31.7, 30.7, 27.7, 20.1, 19.3, 18.7. HRMS (ESI)

$[\text{M}+\text{H}]^+$ found m/z 615.3081, calcd for $\text{C}_{34}\text{H}_{40}\text{FN}_6\text{O}_4$ 615.309. $[\alpha]_D^{20}$ -0.8 (c = 0.105, CH_3OH).

(S)-1-((4R,7S,10S,13S,16S,19S)-7-(2-amino-2-oxoethyl)-10-(3-amino-3-oxopropyl)-13-((S)-sec-butyl)-16-(4-hydroxybenzyl)-6,9,12,15,18-pentaazo-19-(quinolin-2-ylamino)-1,2-dithia-5,8,11,14,17-pentaazacycloicosane-4-carbonyl)-N-((S)-1-((2-amino-2-oxoethyl)amino)-4-methyl-1-oxopentan-2-yl)pyrrolidine-2-carboxamide (8f). Following the procedure of **6** starting from oxytocin, the product **8f** was obtained as a white solid (yield 42% for 2 steps). ^1H NMR (400 MHz, MeOD- d_4) δ 8.36 (d, $J = 9.2$ Hz, 1H), 7.96 (d, $J = 7.8$ Hz, 1H), 7.89 (t, $J = 7.8$ Hz, 1H), 7.74 (d, $J = 8.3$ Hz, 1H), 7.61 (t, $J = 7.4$ Hz, 1H), 7.00 (d, 4H), 6.18 (s, 1H), 5.17 (s, 1H), 4.87 (d, $J = 6.8$ Hz, 2H), 4.50 – 4.42 (m, 1H), 4.31 (dd, $J = 9.4, 5.4$ Hz, 1H), 4.07 (d, $J = 6.5$ Hz, 1H), 3.95 (d, $J = 7.6$ Hz, 1H), 3.91 (d, $J = 17.1$ Hz, 1H), 3.81 (d, $J = 8.4$ Hz, 1H), 3.76 (d, $J = 17.0$ Hz, 1H), 3.68 (dt, $J = 10.4, 5.7$ Hz, 1H), 3.42 (dt, $J = 13.6, 4.3$ Hz, 2H), 3.37 (s, 4H), 2.91 (s, 1H), 2.71 (s, 1H), 2.39 (t, $J = 6.9$ Hz, 2H), 2.31 – 2.14 (m, 3H), 2.03 – 1.83 (m, 4H), 1.68 (qd, $J = 9.7, 9.3, 5.1$ Hz, 4H), 1.37 – 1.24 (m, 1H), 1.05 (d, $J = 6.7$ Hz, 3H), 1.00 (t, $J = 7.4$ Hz, 3H), 0.94 (d, $J = 6.3$ Hz,

3H), 0.91 (d, $J = 6.1$ Hz, 3H). ^{13}C NMR (125 MHz, MeOD- d_4) δ 176.6, 173.7, 173.5, 173.2, 172.9, 172.1, 168.4, 167.7, 157.7, 157.4, 155.6, 133.1, 129.8, 128.7, 127.2, 125.9, 121.8, 117.7, 115.8, 114.4, 113.5, 61.0, 60.8, 55.8, 54.9, 53.6, 53.0, 52.3, 50.6, 48.5, 48.2, 41.9, 39.7, 37.7, 35.7, 31.4, 29.0, 25.6, 24.5, 24.5, 22.1, 20.5, 14.6, 10.2. HRMS (ESI) $[\text{M}+\text{H}]^+$ found m/z 1132.4742, calcd for **10**

$\text{C}_{52}\text{H}_{70}\text{N}_{13}\text{O}_{12}\text{S}_2$ 1132.4742. $[\alpha]_D$ -110.2 ($c = 0.077$, CH_3OH).

(S)-N-((S)-1-amino-3-hydroxy-1-oxopropan-2-yl)-1-(quinolin-2-yl-D-tyrosyl-L-alanyl-D-phenylalanyl-glycyl-D-tyrosyl)pyrrolidine-2-carboxamide (8g). Following the procedure of **6** starting from dermorphin, the product **8g** was obtained as a white solid (yield 47% for 2 steps). ^1H NMR (400 MHz, DMSO- d_6) δ 9.25 (s, 1H), 9.17 (s, 1H), 8.27 (d, $J = 8.0$ Hz, 1H), 8.20 (s, 1H), 8.12 (d, $J = 8.6$ Hz, 1H), 8.00 (d, $J = 6.9$ Hz, 1H), 7.82 (d, $J = 8.9$ Hz, 1H), 7.75 (d, $J = 7.4$ Hz, 1H), 7.59 (d, $J = 7.6$ Hz, 1H), 7.49 (d, $J = 7.8$ Hz, 1H), 7.46 – 7.38 (m, 1H), 7.18 (dd, $J = 13.4, 5.1$ Hz, 5H), 7.12 (d, $J = 7.5$ Hz, 3H), 7.10 – 7.05 (m, 3H), 6.85 (d, $J = 8.9$ Hz, 1H), 6.66 (d, $J = 8.2$ Hz, 2H), 6.62 (d, $J = 8.0$ Hz, 2H), 4.91 (t, $J = 5.3$ Hz, 1H), 4.74 (d, $J = 4.4$ Hz, 1H), 4.60 (s, 1H), 4.47 (s, 1H), 4.35 (d, $J = 6.0$ Hz, 1H), 4.26 – 4.04 (m, 2H), 3.78 – 3.69 (m, 1H), 3.68 – 3.54 (m, 4H), 2.97 (d, $J = 10.9$ Hz, 3H), 2.87 – 2.76 (m, 1H), 2.69 – 2.60 (m, 1H), 2.57 (d, $J = 12.8$ Hz, 1H), 1.93 (d, $J = 51.9$ Hz, 4H), 0.82 (d, $J = 6.9$ Hz, 3H). ^{13}C NMR (125 MHz, DMSO- d_6) δ 172.5, 172.3, 172.3, 171.6, 171.6, 171.0, 168.8, 156.6, 156.3, 156.1, 147.9, 138.2, 136.8, 130.7, 130.6, 129.6, 129.5, 128.8, 128.4, 128.1, 127.8, 126.6, 126.2, 123.5, 121.9, 115.5, 115.3, 113.6, 62.0, 60.4, 56.5, 55.5, 54.1, 53.1, 48.5, 47.4, 42.0, 38.1, 37.5, 36.7, 29.3, 24.9, 18.8. HRMS (ESI) **10**

$[\text{M}+\text{H}]^+$ found m/z 928.3993, calcd for $\text{C}_{49}\text{H}_{54}\text{N}_9\text{O}_{10}$ 928.3999. $[\alpha]_D$ -19.9 ($c = 0.098$, CH_3OH).

(4R,7S,10S,13R,16S,19R)-13-((1H-indol-3-yl)methyl)-10-(4-aminobutyl)-16-benzyl-N-(1,3-dihydroxybutan-2-yl)-7-((R)-1-hydroxyethyl)-6,9,12,15,18-pentaoxo-19-((R)-3-phenyl-2-(quinolin-2-ylamino)propanamido)-1,2-dithia-5,8,11,14,17-pentaazacycloicosane-4-carboxamide (8h). Following the procedure of **6** starting from otreotide, the product **8h** was obtained as a white solid (yield 41% for 2 steps). ^1H NMR (500 MHz, Methanol- d_4) δ 8.27 (d, $J = 9.4$ Hz, 1H), 7.85 (d, $J = 7.9$ Hz, 1H), 7.78 (dd, $J = 14.4, 7.1$ Hz, 2H), 7.52 (t, $J = 7.8$ Hz, 1H), 7.48 (d, $J = 7.9$ Hz, 1H), 7.40 – 7.35 (m, 3H), 7.27 – 7.22 (m, 3H), 7.19 (t, $J = 7.9$ Hz, 4H), 7.16 – 7.10 (m, 3H), 7.04 (t, $J = 7.4$ Hz, 1H), 7.00 (s, 1H), 5.25 (d, $J = 9.4$ Hz, 2H), 4.68 (dd, $J = 8.8, 6.2$ Hz, 1H), 4.49 (d, $J = 5.6$ Hz, 1H), 4.38 – 4.32 (m, 1H), 4.23 (dd, $J = 11.0, 5.3$ Hz, 1H), 4.17 (dd, $J = 6.4, 2.8$ Hz, 1H), 4.02 (dd, $J = 10.9, 3.2$ Hz, 1H), 3.89 – 3.85 (m, 1H), 3.74 (s, 2H), 3.50 (d, $J = 9.2$ Hz, 1H), 3.33 (s, 2H), 3.18 – 2.90 (m, 8H), 2.86 (dd, $J = 13.8, 5.2$ Hz, 1H), 2.61 (dq, $J = 18.7, 12.4$ Hz, 2H), 1.35 – 1.23 (m, 9H), 0.62 – 0.43 (m, 2H). ^{13}C NMR (125 MHz, MeOD- d_4) δ 173.9, 173.2, 171.2, 171.0, 170.9, 168.8, 152.3, 136.3, 136.2, 135.4, 129.0, 128.7, 128.2, 128.1, 127.6, 126.9, 126.7, 126.1, 125.3, 123.1, 121.1, 120.8, 118.3, 117.7, 110.7, 108.4, 66.8, 61.2, 59.3, 57.0, 56.1, 55.8, 54.5, 53.3, 53.0, 52.2, 48.2, 38.8, 38.6, 38.5, 29.5, 26.0, 25.5, 21.5, 18.7, 18.4. **10**

HRMS (ESI) $[\text{M}+\text{H}]^+$ found m/z 1146.4891, calcd for $\text{C}_{58}\text{H}_{72}\text{N}_{11}\text{O}_{10}\text{S}_2$ 1146.49. $[\alpha]_D$ -31.8 ($c = 0.067$, CH_3OH).

(4R,7S,10S,13R,16S,19R)-13-((1H-indol-3-yl)methyl)-10-(4-aminobutyl)-16-benzyl-N-(1,3-dihydroxybutan-2-yl)-7-((R)-1-hydroxyethyl)-19-((R)-2-((6-methoxyquinolin-2-yl)amino)-3-phenylpropanamido)-6,9,12,15,18-pentaoxo-1,2-dithia-5,8,11,14,17-pentaazacycloicosane-4-carboxamide (8i). Following the procedure of **6** starting from otreotide, the product **8i** was

obtained as a white solid (yield 46% for 2 steps). ¹H NMR (500 MHz, Methanol-*d*₄) δ 8.35 (d, *J* = 9.4 Hz, 1H), 8.23 (d, *J* = 8.7 Hz, 1H), 8.14 – 8.00 (m, 2H), 7.84 (d, *J* = 27.0 Hz, 1H), 7.71 – 7.64 (m, 2H), 7.51 (d, *J* = 8.0 Hz, 1H), 7.42 (d, *J* = 7.6 Hz, 2H), 7.39 (d, *J* = 8.1 Hz, 1H), 7.31 (t, *J* = 7.5 Hz, 2H), 7.27 – 7.19 (m, 5H), 7.18 – 7.13 (m, 2H), 7.08 (dd, *J* = 8.4, 6.7 Hz, 3H), 7.02 (s, 1H), 5.30 (t, *J* = 12.2 Hz, 2H), 4.71 (dd, *J* = 8.9, 6.0 Hz, 1H), 4.54 – 4.47 (m, 1H), 4.44 – 4.33 (m, 1H), 4.25 (dd, *J* = 10.9, 5.4 Hz, 1H), 4.23 – 4.16 (m, 1H), 4.06 (dd, *J* = 11.0, 3.5 Hz, 1H), 3.89 (s, 4H), 3.77 (d, *J* = 5.2 Hz, 2H), 3.54 (dd, *J* = 14.0, 5.0 Hz, 1H), 3.39 (s, 2H), 3.23 – 2.94 (m, 8H), 2.89 (dd, *J* = 13.8, 5.3 Hz, 1H), 2.65 (tt, *J* = 20.1, 10.6 Hz, 2H), 1.28 (m, 9H), 0.69 – 0.50 (m, 2H). ¹³C NMR (125 MHz, MeOD-*d*₄) δ 174.2, 173.6, 171.6, 171.3, 169.2, 160.0, 152.5, 138.6, 136.6, 136.5, 135.8, 131.2, 129.5, 129.4, 129.3, 129.0, 128.4, 128.0, 127.8, 127.2, 127.1, 126.4, 125.2, 123.4, 122.0, 121.1, 118.6, 118.0, 114.2, 110.9, 108.8, 67.1, 59.7, 57.3, 56.4, 54.8, 54.4, 53.5, 53.3, 52.6, 48.4, 38.8, 31.6, 28.9, 26.3, 25.5, 22.3, 19.0, 13.0. HRMS (ESI) [M+H]⁺ found *m/z* 1252.5299, **20**

calcd for C₆₅H₇₈N₁₁O₁₁S₂ 1252.5318. [α]_D 1.8 (c = 0.055, CH₃OH).

3. Biological studies

3.1 Cell proliferation inhibition assay

The cancer cell lines BEL-7402 and SMMC-7721 were purchased from the American Type Culture Collection (Manassas, VA, USA). All cell lines were maintained in DMEM containing 10% fetal bovine serum at 37 °C and 5% CO₂ in a humid environment.

Tumor cells were seeded in the 96-wells plates overnight and then were treated with increasing doses of compounds in triplicate for 72 h. The anti-proliferative activities were accessed by SRB assay. Then the OD was measured at 510 nm wavelength using Synergy H4 Hybrid reader (BioTek, Winooski, VT, USA) using Gen5.0 software (BioTek). The IC₅₀ values were calculated using the software Prism 5 (GraphPad Software, Inc). The test results are shown in the following table.

Table S1. Liver cancer cell proliferation inhibition assays of selected compounds

Compound	Proliferation inhibition IC ₅₀ (μM)	
	BEL-7402	SMMC-7721
YSL	>100	>100
YSL-M	>100	>100
8c	52.57±2.47	63.30±6.35

3.2 [³⁵S]GTP_γS Binding Assay

Transfer 10 μl standard protein to a centrifuge tube (final concentration 0.5 mg/ml). Transfer the sample and 0, 1, 2, 4, 8, 12, 16, 20 μl bovine serum albumin (BSA) standard protein into the wells of a 96-well plate. The solution was supplemented to 20 μl using diluted standard. 200 μl of BCA working solution was added to each well then the plate was placed at 37 °C for 30 min. Meanwhile, set the plate reader to read absorption at 560 nm. The protein concentration was calculated from the standard curve.

The prepared membrane receptor is diluted to the desired concentration with a reaction buffer (R.B). Load according to the following table (unit: μl).

	R.B	[³⁵ S]GTP _γ S 0.1-0.2nM	GTP _γ S 20μM	GDP 40μM	Agonist	Protein 20-30μg
NS binding	30	10	10	0	0	50
Basal	30	10	0	10	0	50
Agonist	20	10	0	10	10	50

The reaction tube was incubated for 1 hour in a 27 °C water bath, filtered under reduced pressure on a glass fiber membrane, and flash-counted. Calculate according to the following formula: [³⁵S]GTP_γS binding rate = $100 \times (\text{cpm}_{\text{sample}} - \text{cpm}_{\text{non-specific}}) / (\text{cpm}_{\text{basal}} - \text{cpm}_{\text{non-specific}})$

Table S2. μ-opioid receptor (MOR) agonist activity assays

Compound	(MOR) (mean±sem)	
	EC ₅₀ (nM)	E _{max} (%)
Dermorphin	61.3±10.6	218.8±7.1
8g	223.9±25.1	202.7±5.1

3.3 *In vitro* liver stability assessment

Adhering fat and connective tissue were removed from 5 g fresh pig liver. Wash it with cold saline, blot water on the surface with filter paper and weigh it. Add appropriate PBS buffer, cut the pig liver with scissor and make a 20 ml homogenate with a homogenizer with a frequency of 30/s for 4 min. Tested compound (final concentration 1 mg/ml and co-solvent (1% DMSO)) were added to 20 ml of pig liver homogenate, stir well and stand. Take 1 ml of the sample twice in the six time periods of 0.25 h, 0.5 h, 1 h, 1.5 h, 2 h and 3 h respectively and methanol (500μl) was added to terminate the reaction. After centrifugation (1000 rpm, 6 min), the supernatant was separated and extracted twice with 750 μl of water-saturated n-butanol. Then extracted samples were analyzed by HPLC. Reversed phase HPLC was carried out on an Agilent Zorbax SB-C18 column (5 μm, 4.6 × 250 mm) from Agilent, with a flow rate of 1 mL/min at 25 °C. The gradient conditions used are 50% A (MeOH), 50% B (H₂O + 0.2% CH₃COOH), to 95% A and 5% B in 16 min. Acquisition for the UV-DAD detector was set to 210, 254, and 280 nm.

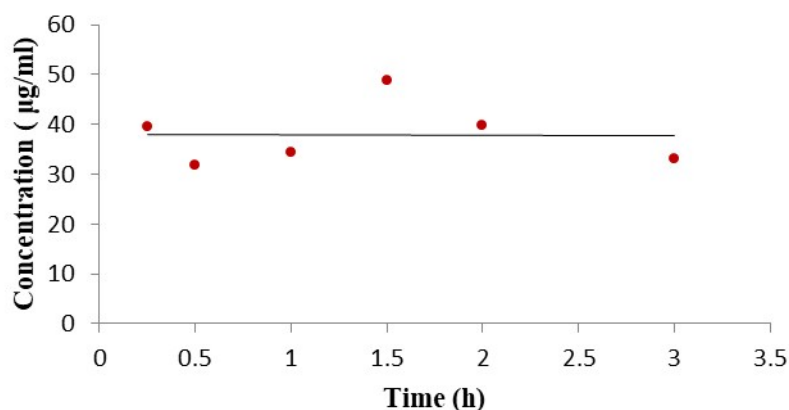


Figure S2. The concentration of 8e changes with time

4. DFT Studies

4.1 Computational Methods

All computations were performed with the Gaussian 09 ^[1] series of programs. All the structures were optimized using the density functional M06-2X methods ^[2] at the 6-311+G(d) level. A Solvent effects of 1-butanol were considered by using the SMD model ^[3]. Frequencies calculations were performed to judge them as local minima or transition states and obtain the thermal corrections to the Gibbs free energies and enthalpies. The Gibbs free energies were used to describe the reaction energies.

4.2 M06-2X calculated energies for reported complexes and transition states

Geometry	E ¹	H ²	G ³	IF ⁴
5a	-669.441609	-669.204898	-669.255131	
7	-303.8408662	-303.71088	-303.749907	
TS	-973.2815993	-972.914091	-972.981862	-297.80
9	-973.288612	-972.918828	-972.986512	

¹Single point energies calculated by M06-2X at the 6-311+g(d) level in 1-butanol solvent.

²Enthalpies calculated by M06-2X at the 6-311+g(d) level in 1-butanol solvent.

³Gibbs free energies calculated by M06-2X at the 6-311+g(d) level in 1-butanol solvent.

⁴M06-2X/6-311+g(d) calculated imaginary frequencies for transition states.

The unit of energy values in the above table are Hartree/Particle (a.u.).

4.3 Cartesian coordinates of minimum energy structures

5a	H -3.777000 -1.961000 -0.237000
C 2.385000 -2.003000 0.303000	H -2.900000 -2.794000 -1.543000
C 1.076000 -1.576000 0.371000	H -2.398000 -3.028000 0.154000
C 0.807000 -0.217000 0.166000	H -1.635000 -1.131000 1.294000
C 1.840000 0.700000 -0.121000	
C 3.166000 0.225000 -0.184000	7
C 3.434000 -1.107000 0.028000	C 0.062000 0.527000 -0.008000
H 2.606000 -3.053000 0.463000	O 0.642000 1.615000 0.081000
H 0.274000 -2.277000 0.572000	N 0.707000 -0.641000 -0.051000
C 1.510000 2.070000 -0.345000	H 0.141000 -1.474000 -0.146000
H 3.962000 0.930000 -0.400000	C 2.154000 -0.724000 -0.004000
H 4.453000 -1.472000 -0.019000	H 2.443000 -1.772000 -0.026000
C 0.224000 2.521000 -0.284000	H 2.604000 -0.214000 -0.859000
C -0.768000 1.573000 0.020000	H 2.538000 -0.270000 0.911000
H 2.314000 2.763000 -0.568000	C -1.452000 0.462000 -0.096000
H -0.060000 3.551000 -0.449000	H -1.844000 1.175000 0.637000
O -2.047000 1.838000 0.119000	H -1.715000 0.848000 -1.085000
C -2.725000 0.649000 0.608000	N -1.977000 -0.892000 0.044000
C -1.714000 -0.486000 0.415000	H -2.116000 -1.116000 1.024000
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H -3.627000 0.507000 0.018000	
N -0.479000 0.291000 0.239000	TS
O -1.894000 -1.226000 -0.750000	C -4.323000 1.047000 -0.497000
C -2.798000 -2.314000 -0.572000	C -3.012000 1.247000 -0.097000

C -2.222000 0.136000 0.204000
C -2.737000 -1.171000 0.103000
C -4.069000 -1.339000 -0.302000
C -4.856000 -0.243000 -0.602000
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H -2.601000 2.248000 -0.028000
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C -0.585000 -2.120000 0.758000
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H 0.082000 -2.931000 1.022000
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H 1.983000 1.404000 1.202000
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H 6.027000 -0.880000 0.712000
H 5.718000 0.296000 -0.581000
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H 2.279000 -1.767000 -2.244000
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9

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C -3.043000 1.228000 -0.171000
C -2.230000 0.136000 0.136000
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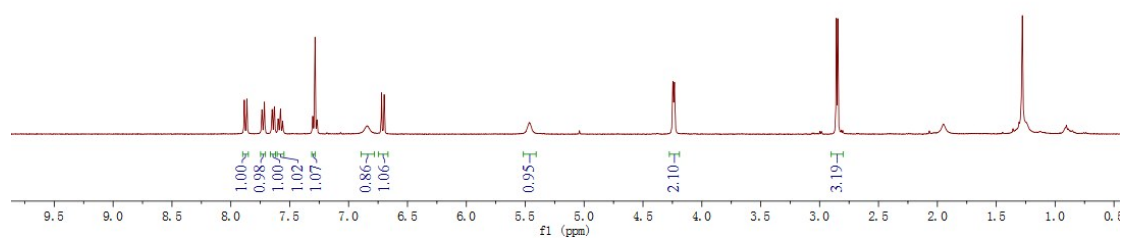
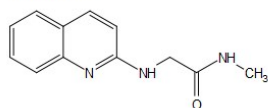
5. References

- [1] Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Keith, T.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, Ö.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. *Gaussian 09*, Revision D.01; Gaussian, Inc.: Wallingford, CT, 2013.
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6. NMR spectra of products

EO-LP-01-44-11

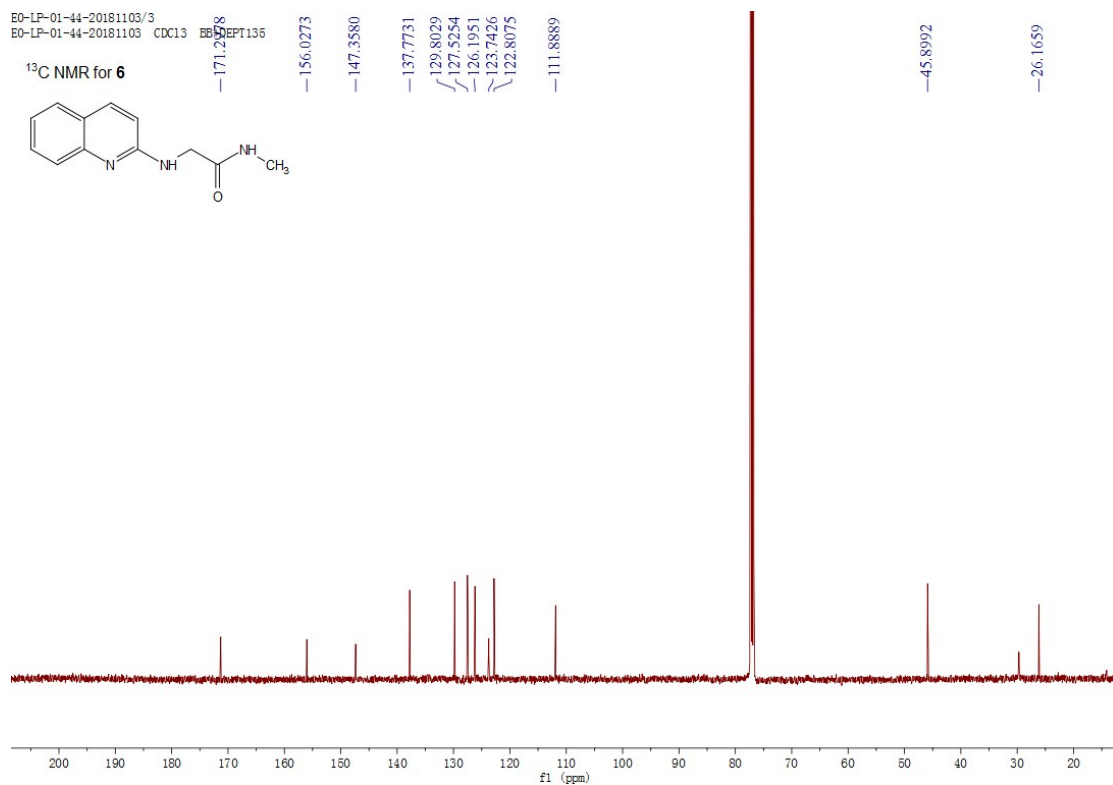
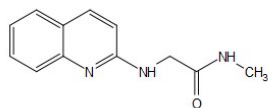
¹H NMR for **6**

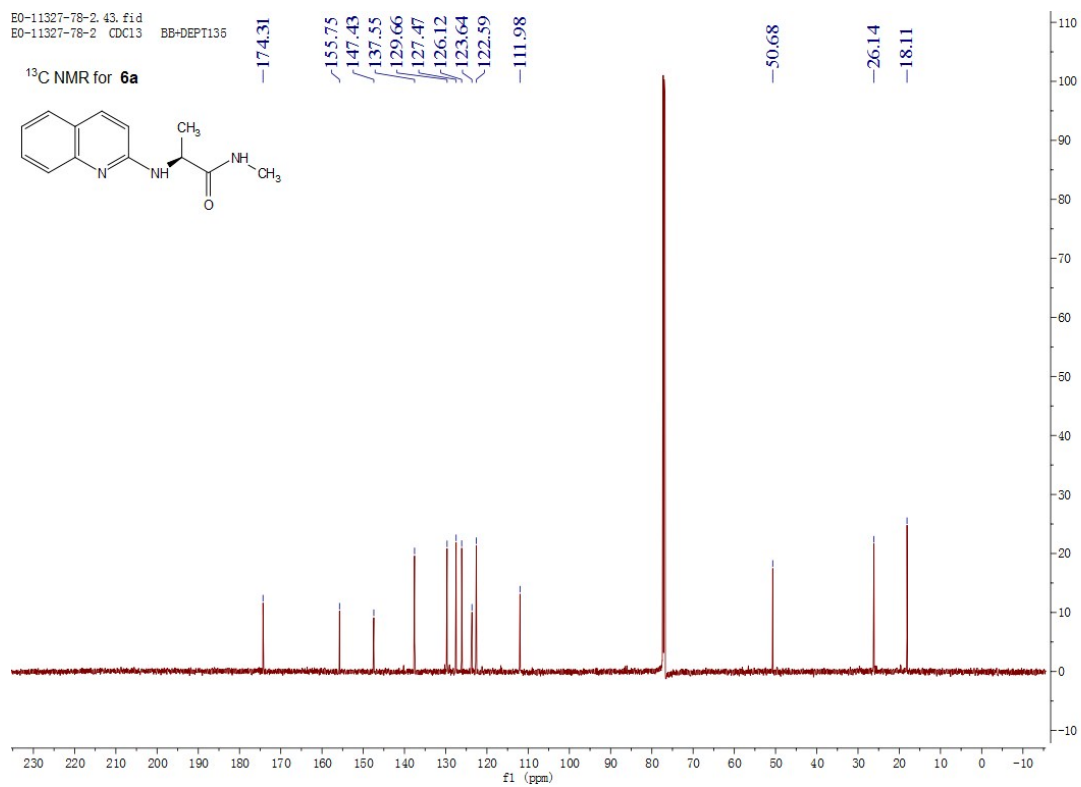
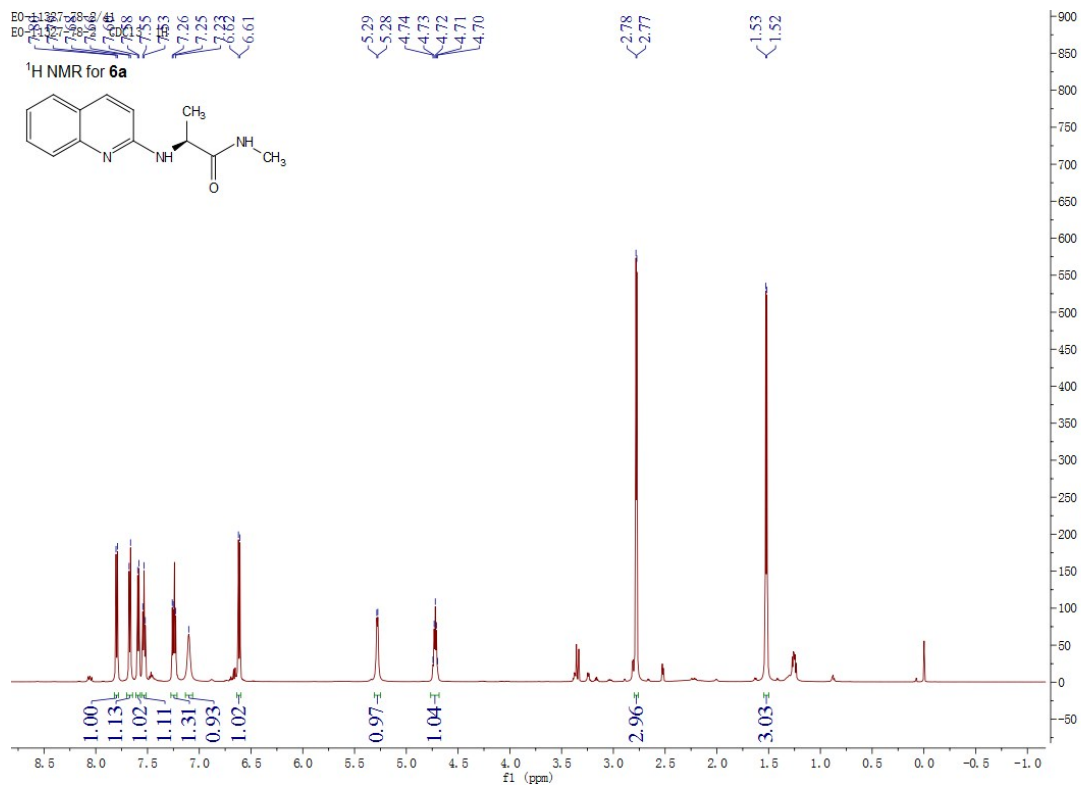


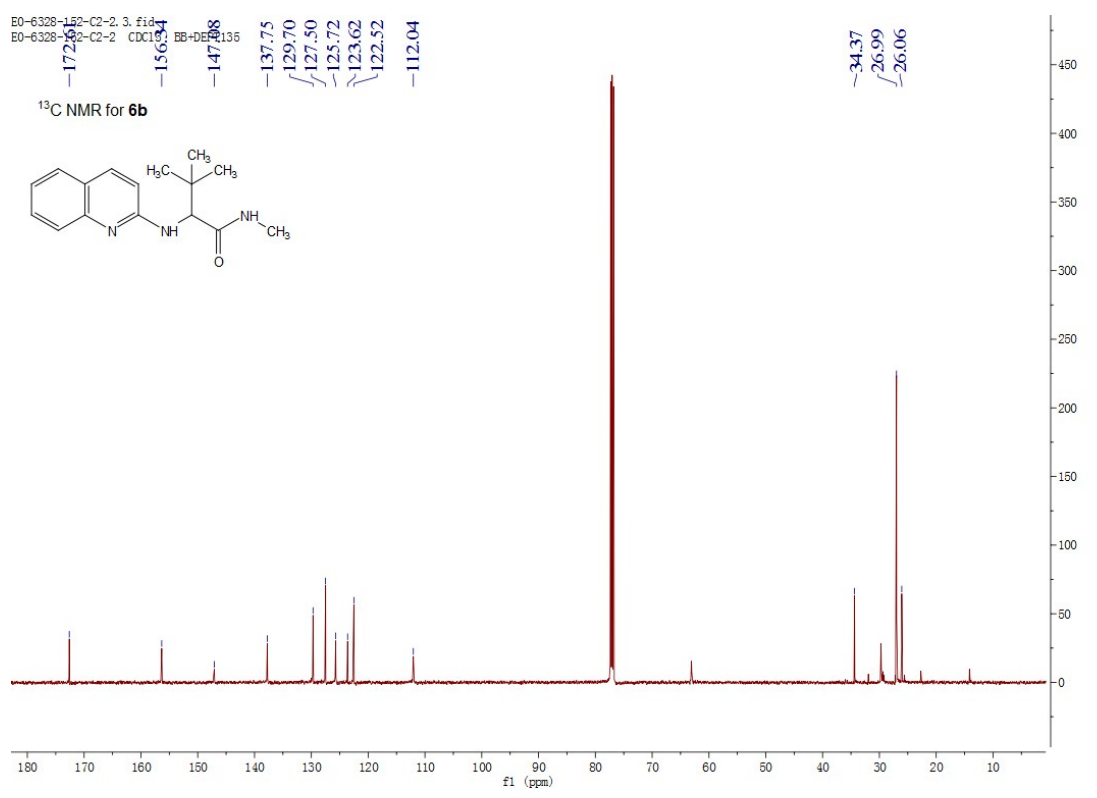
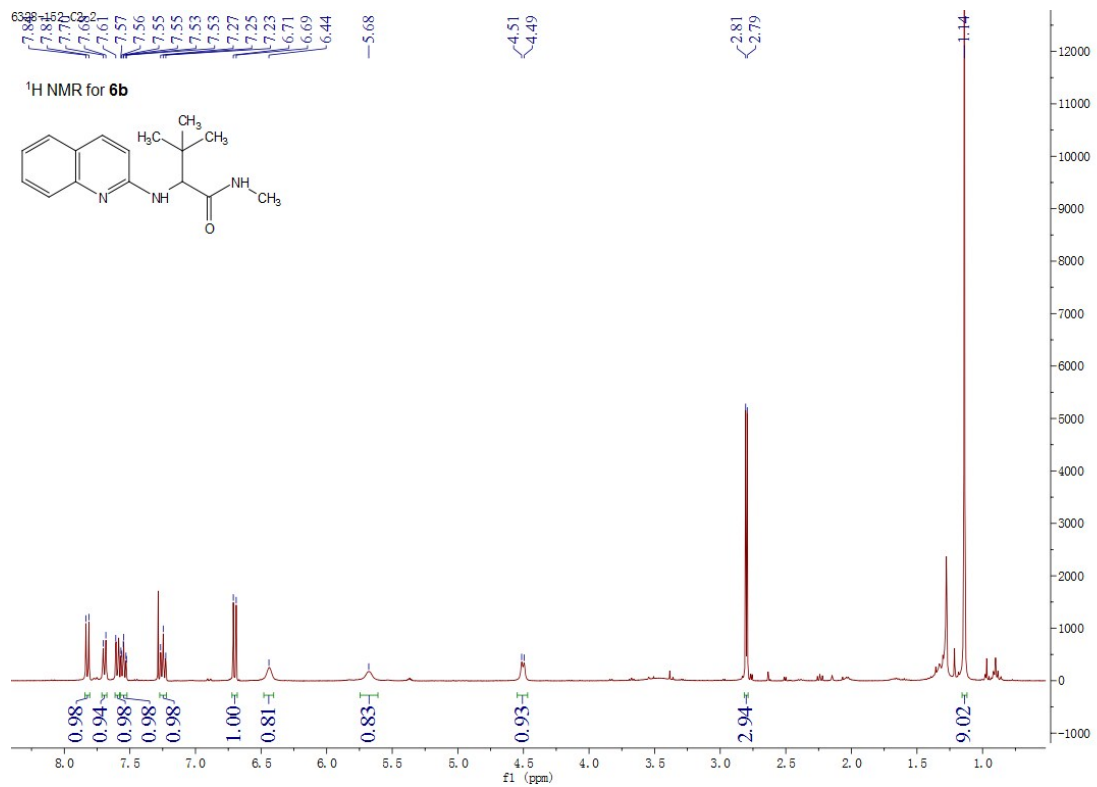
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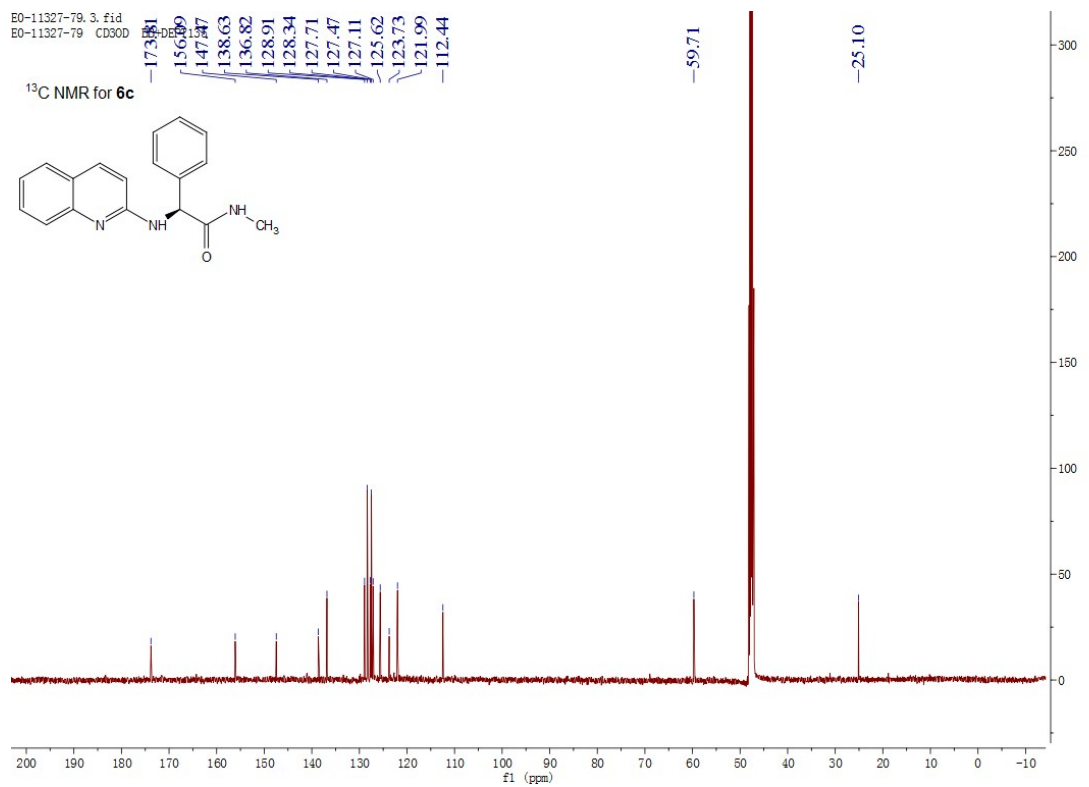
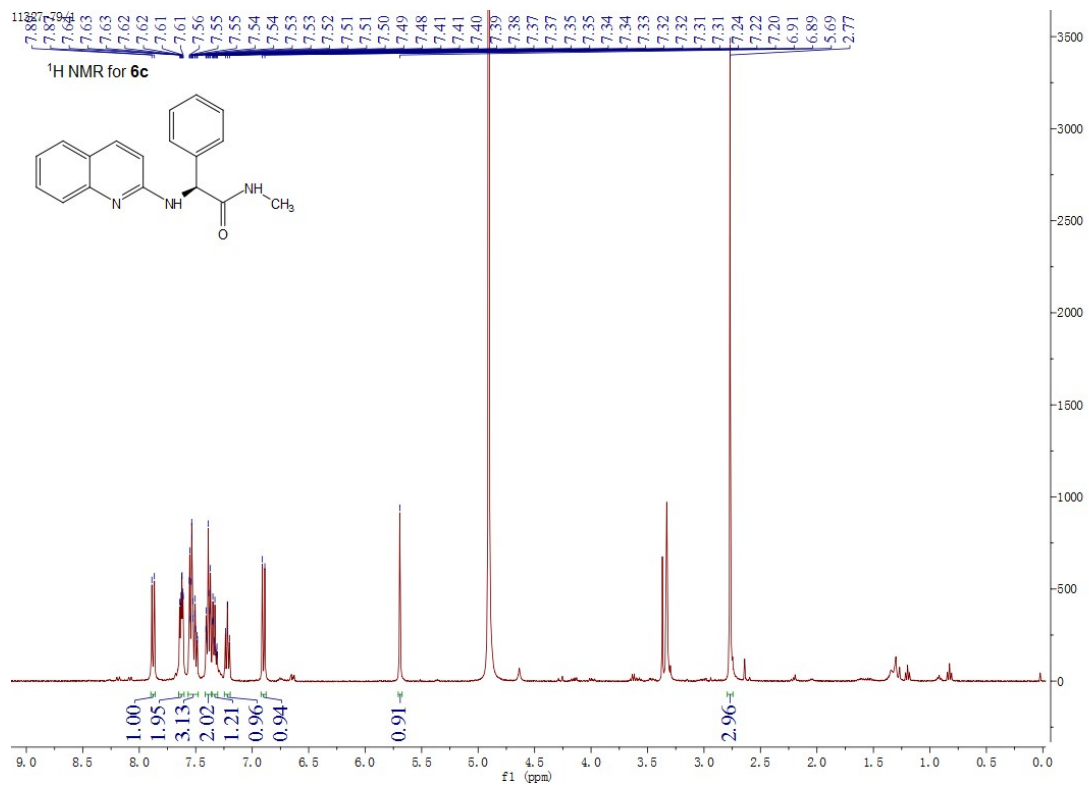
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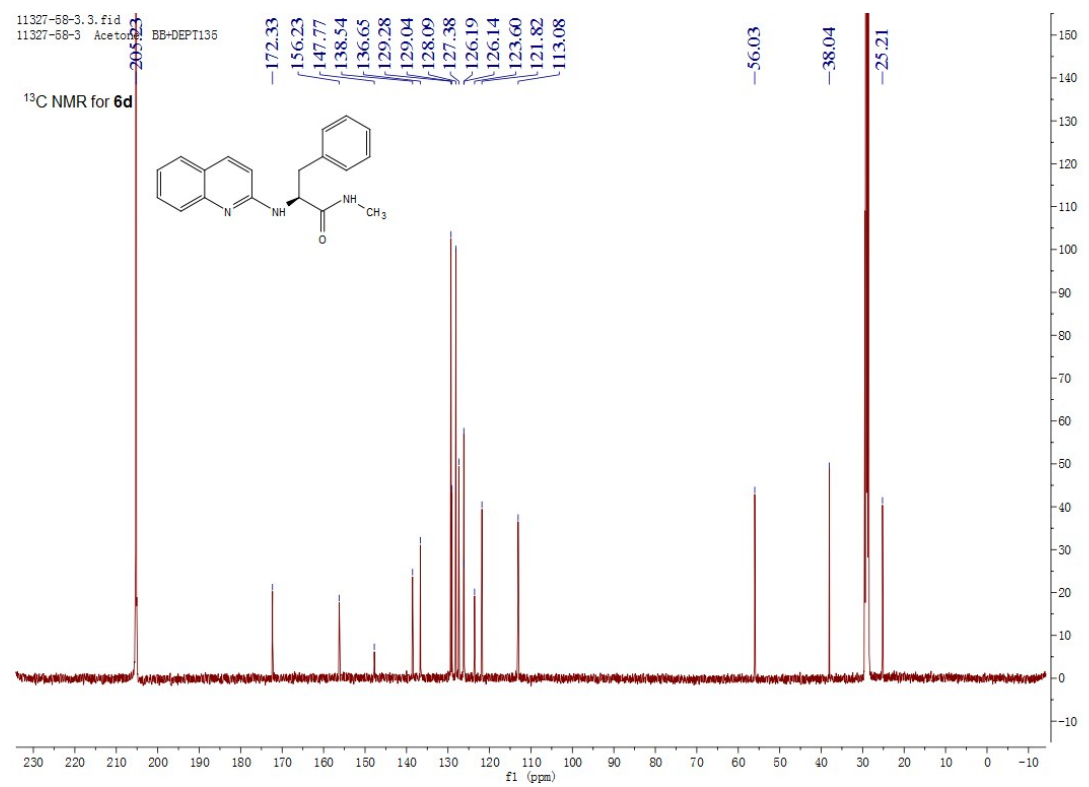
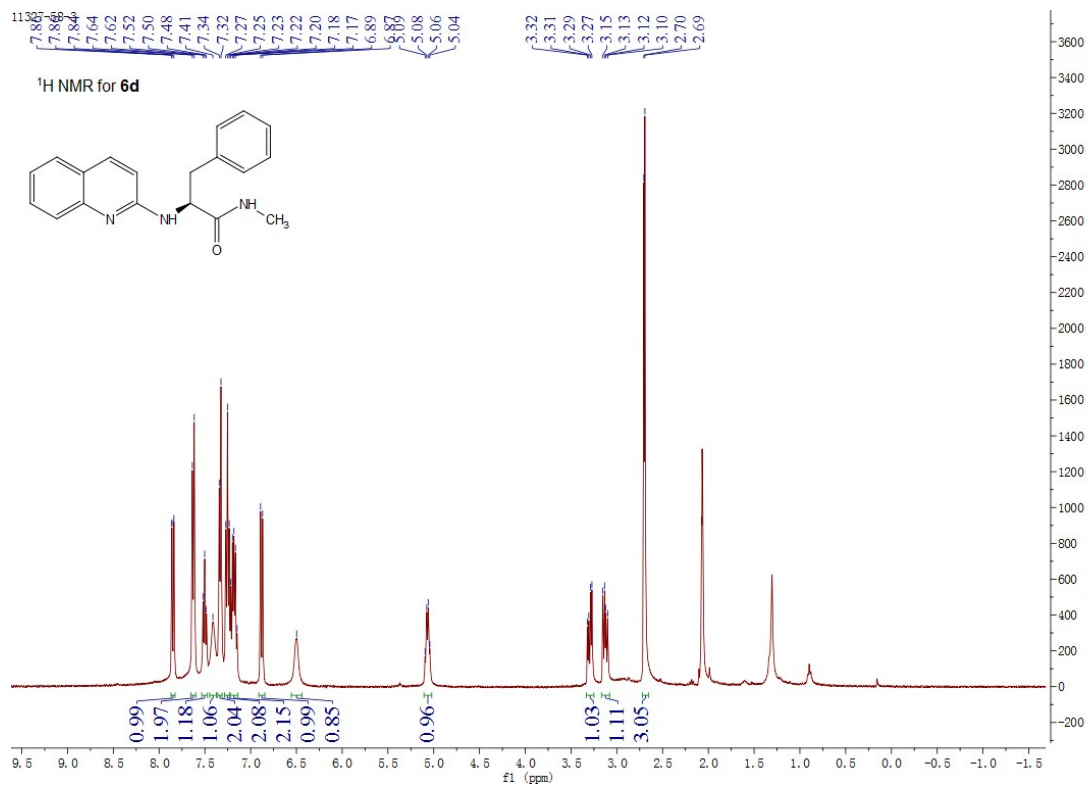
¹³C NMR for **6**

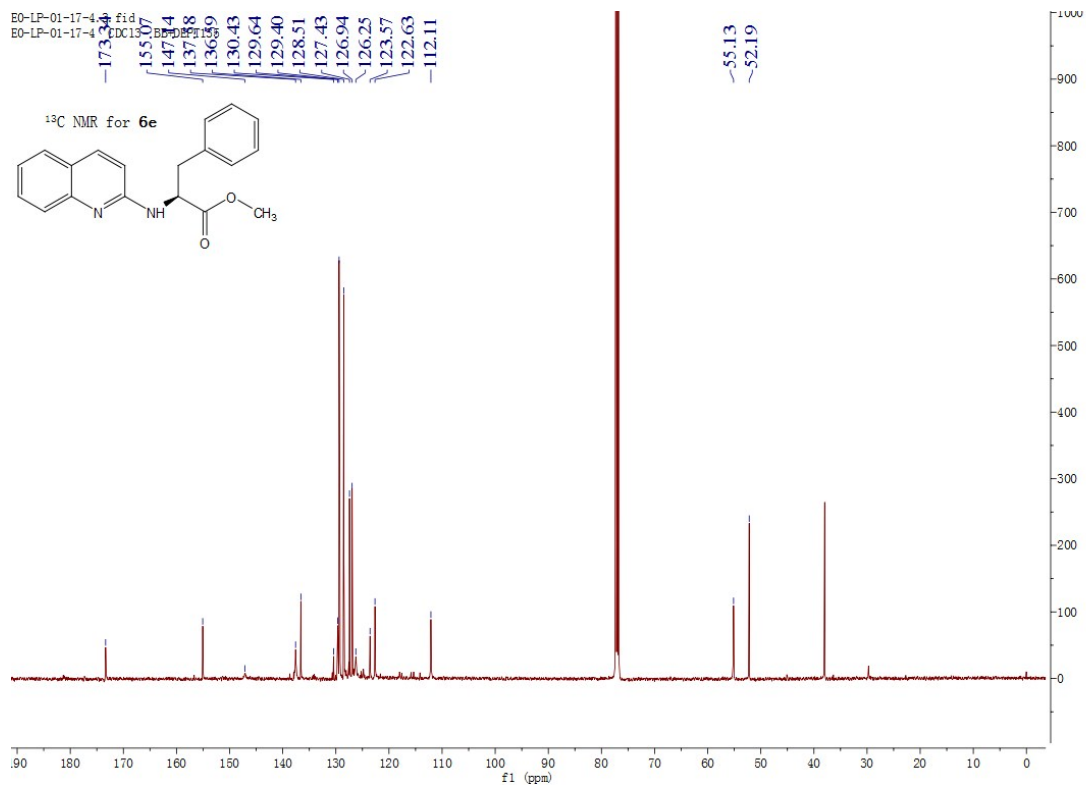
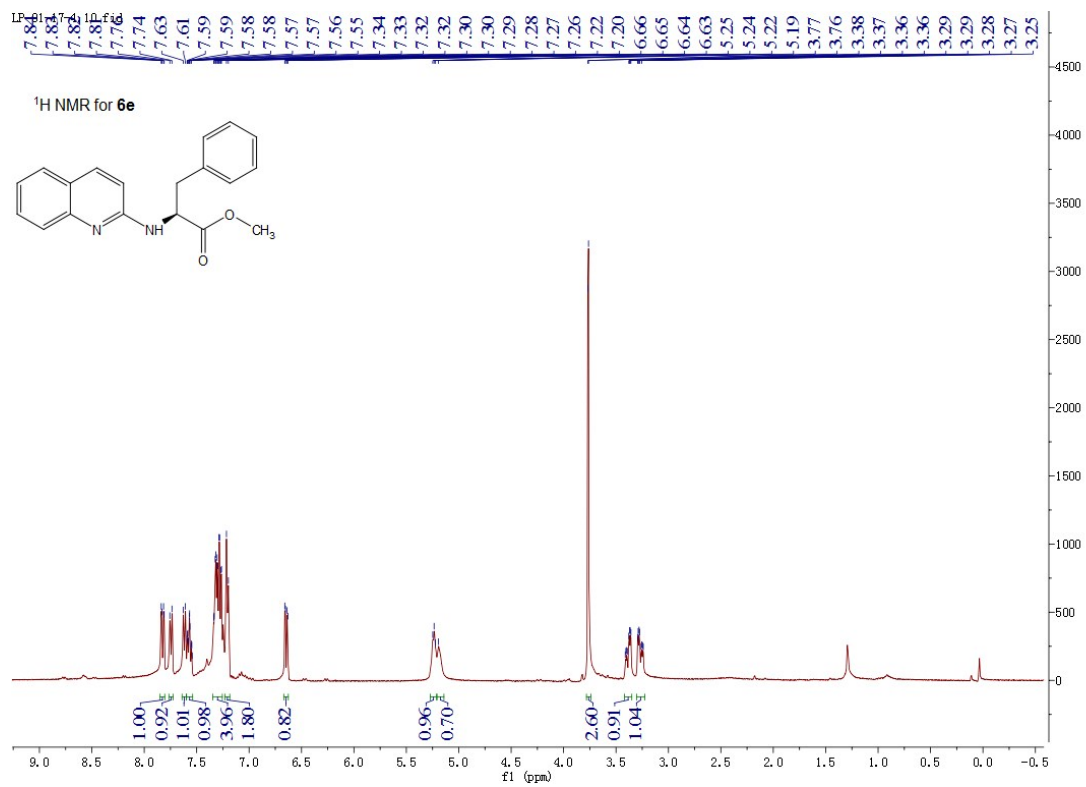


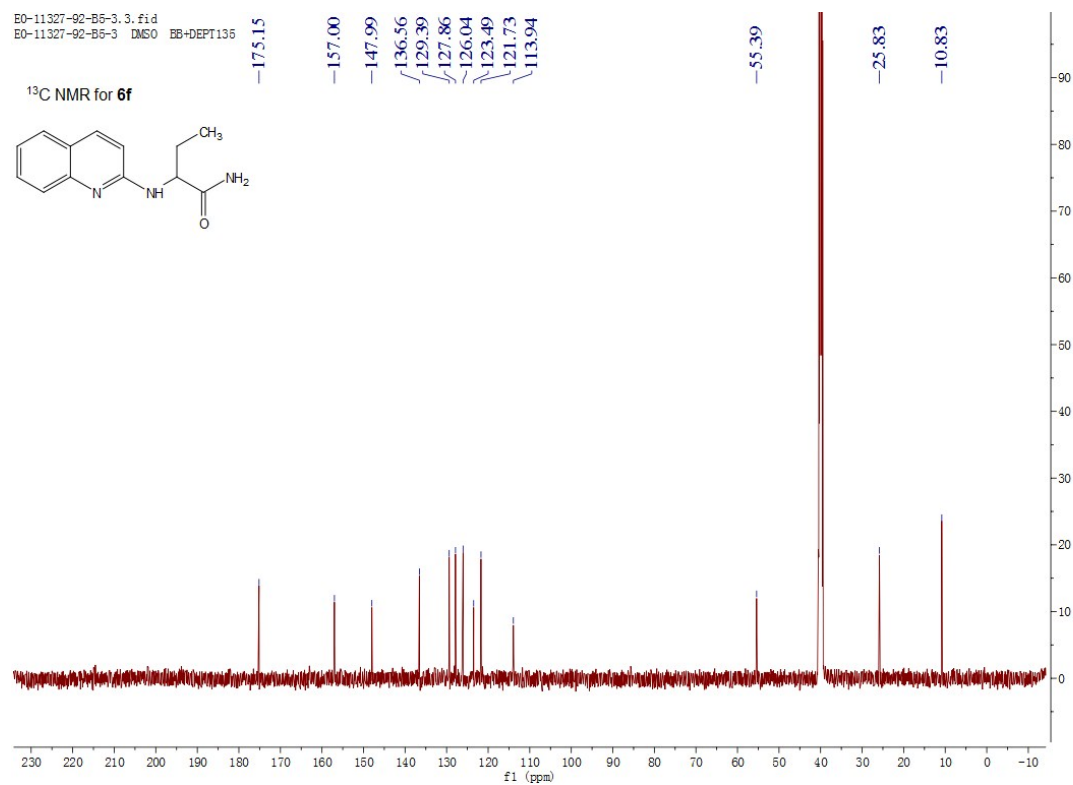
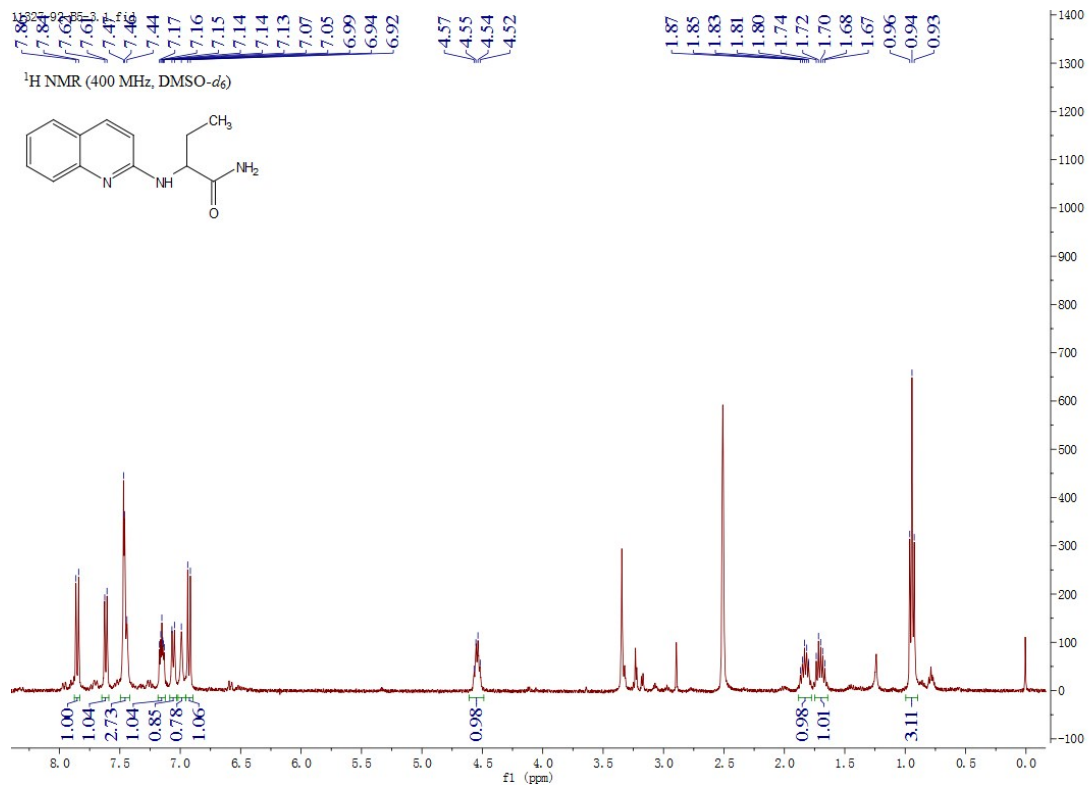


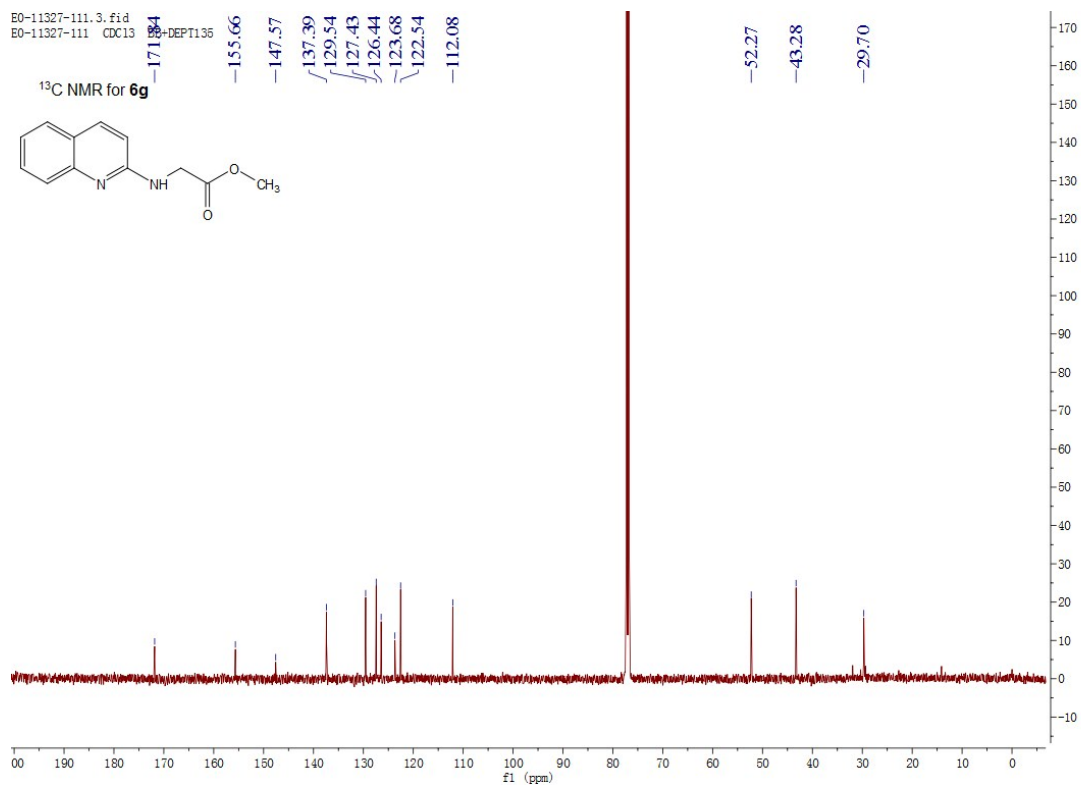
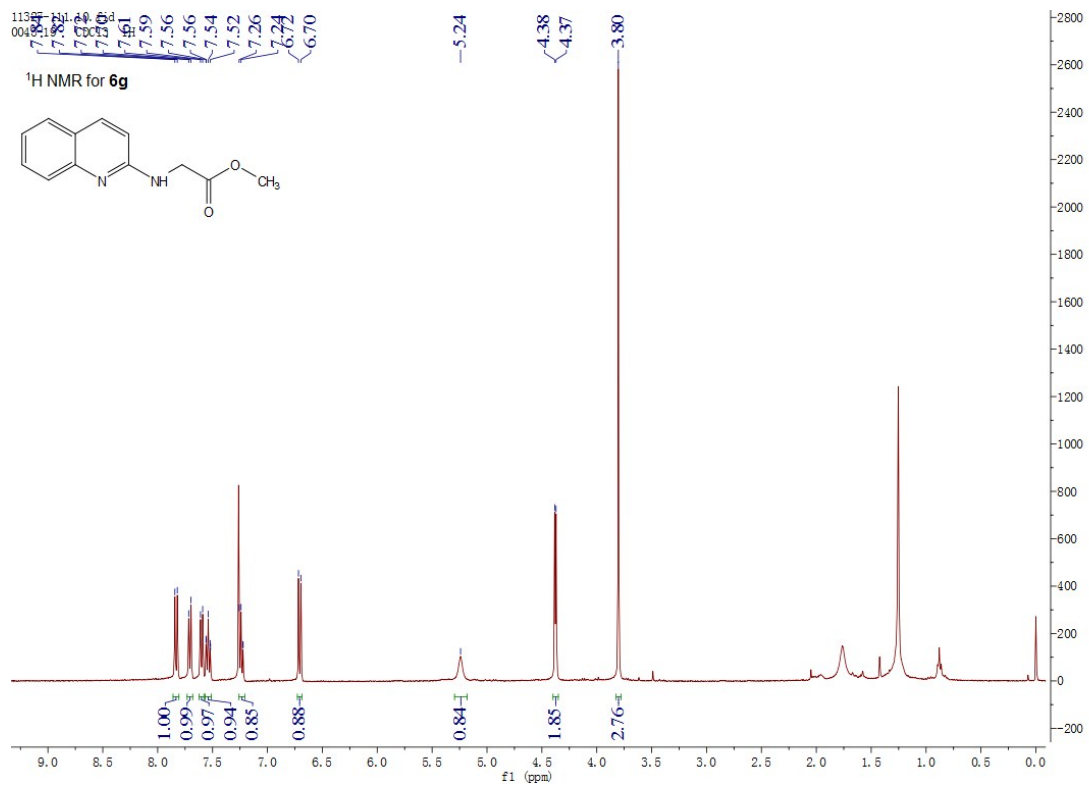


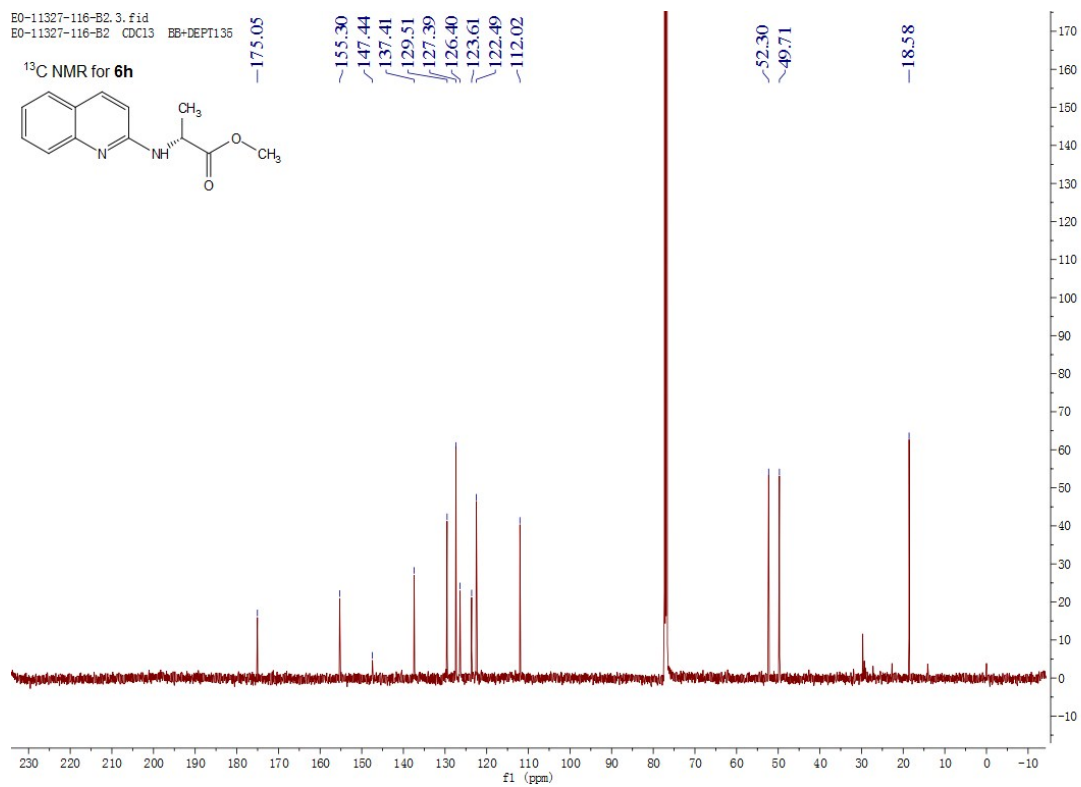
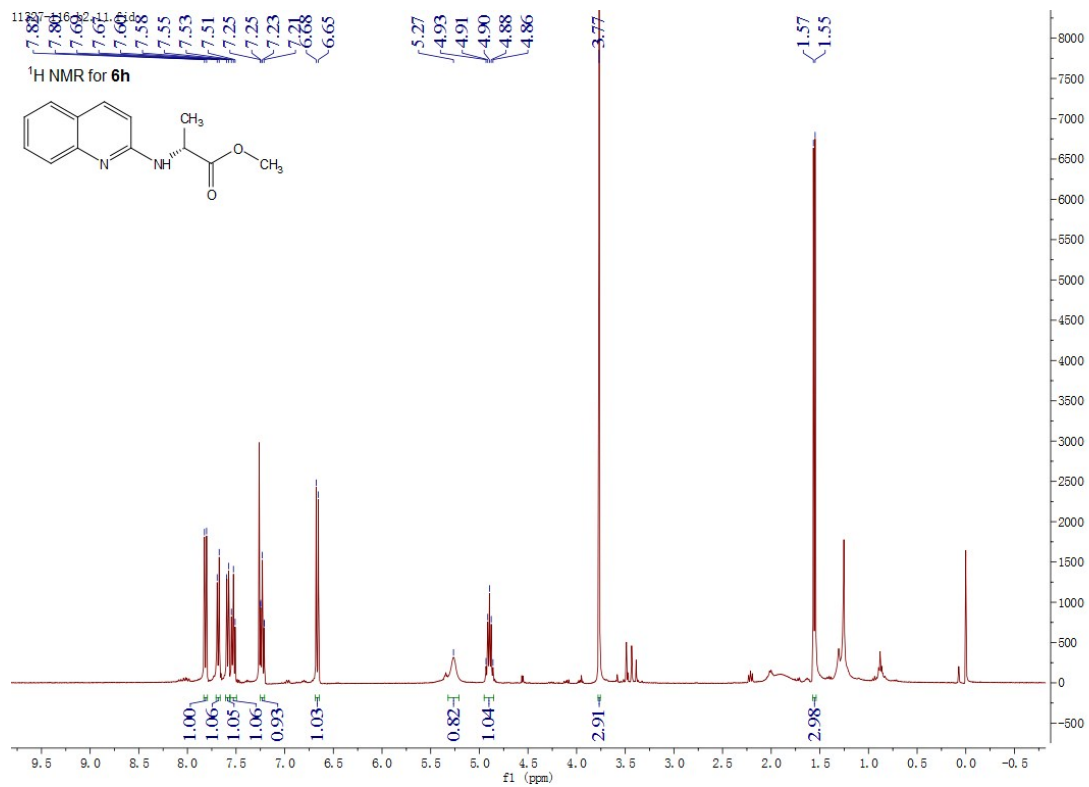


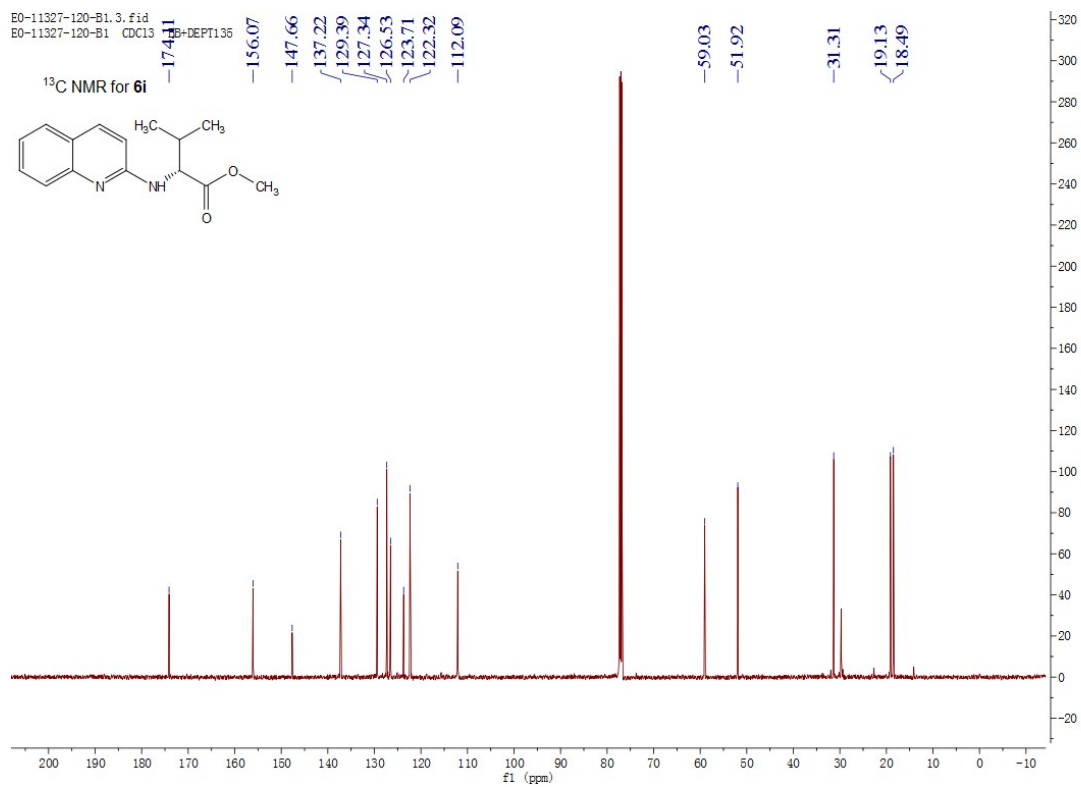
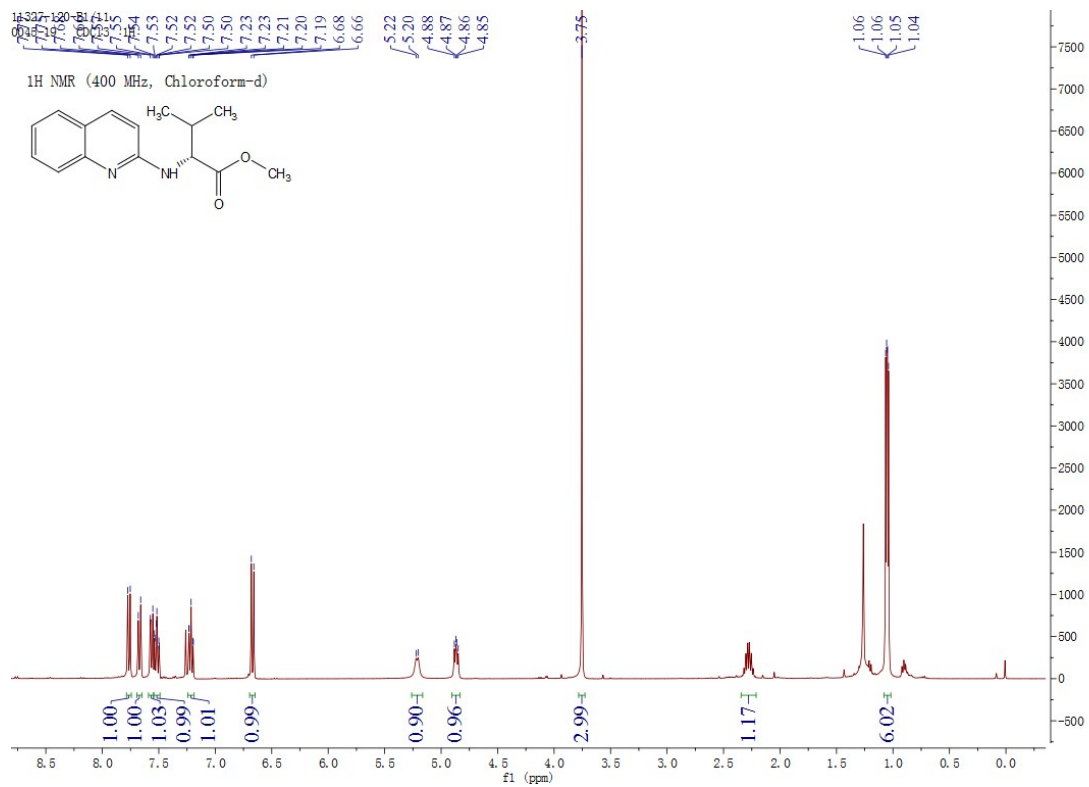


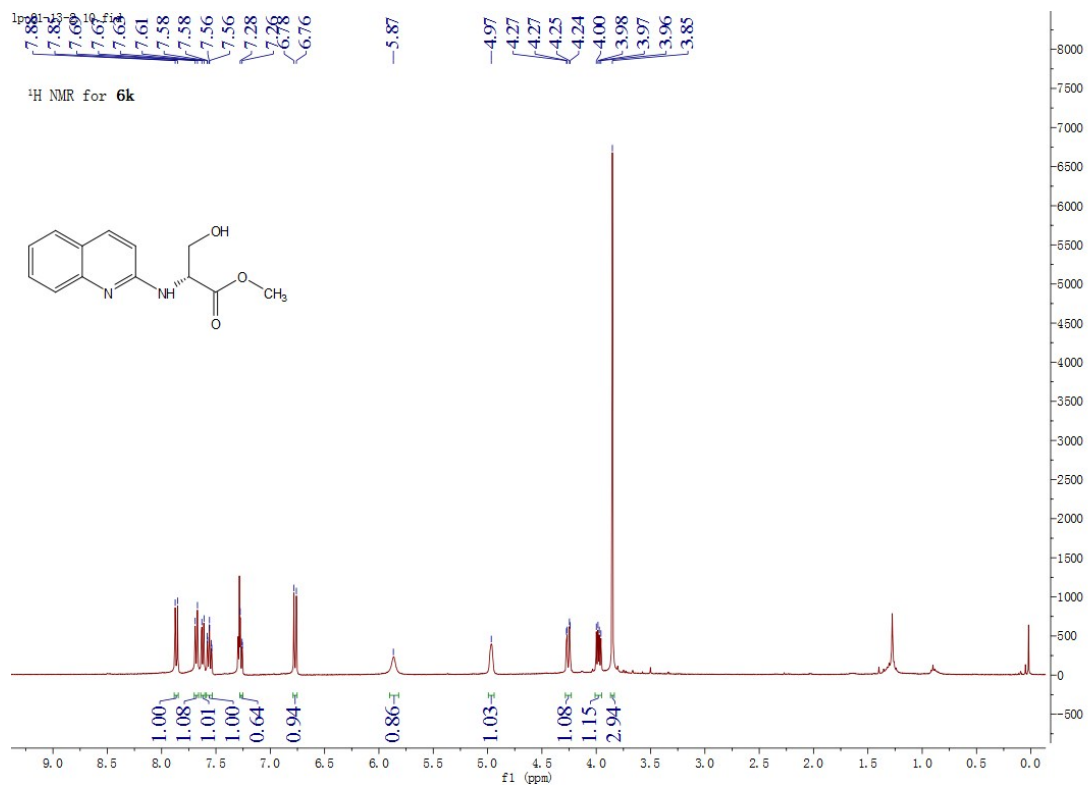
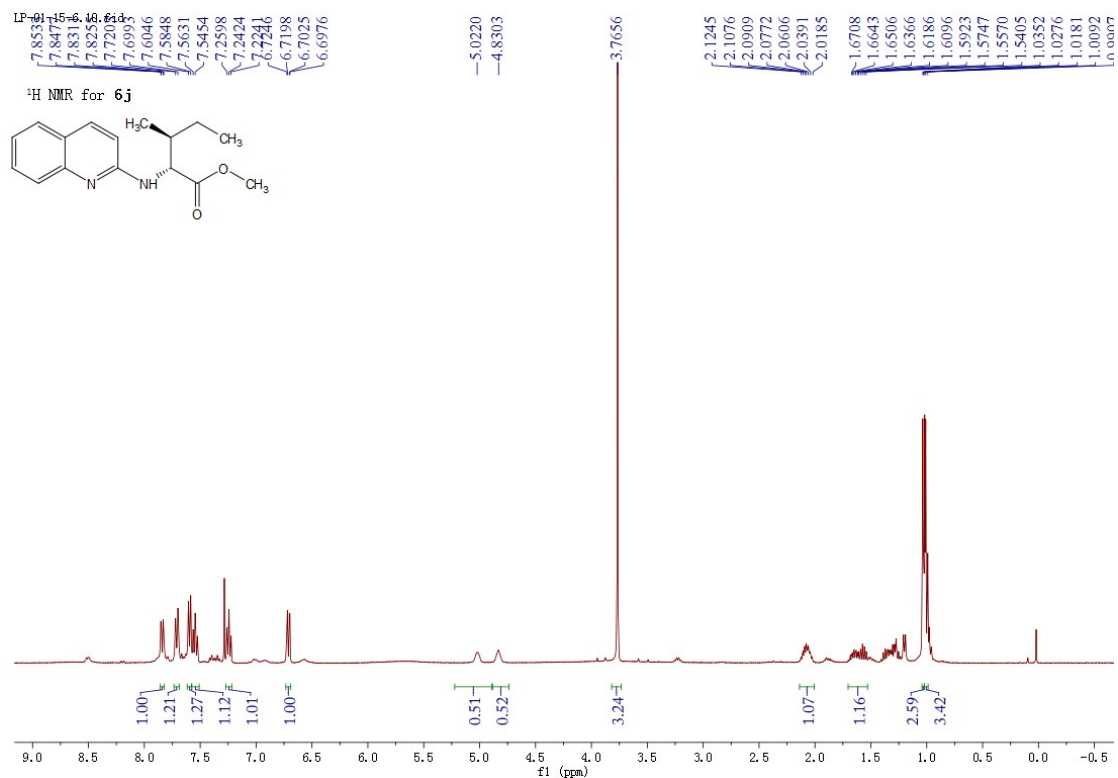






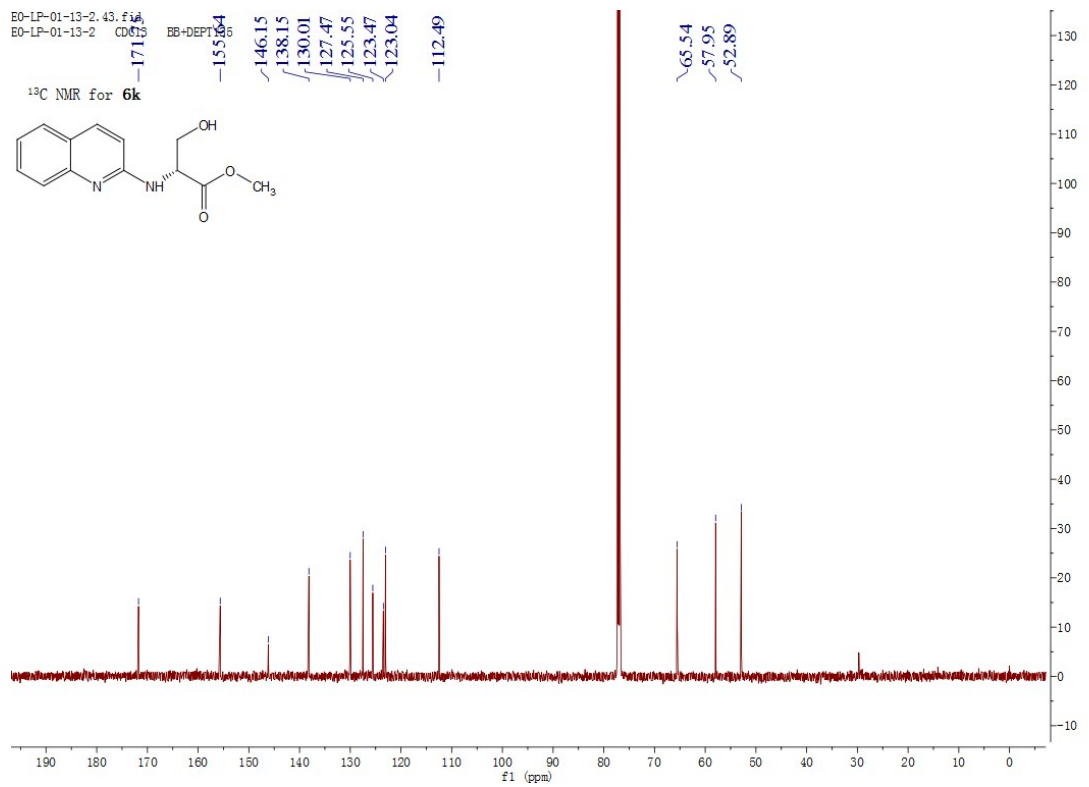
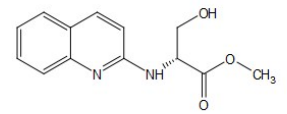




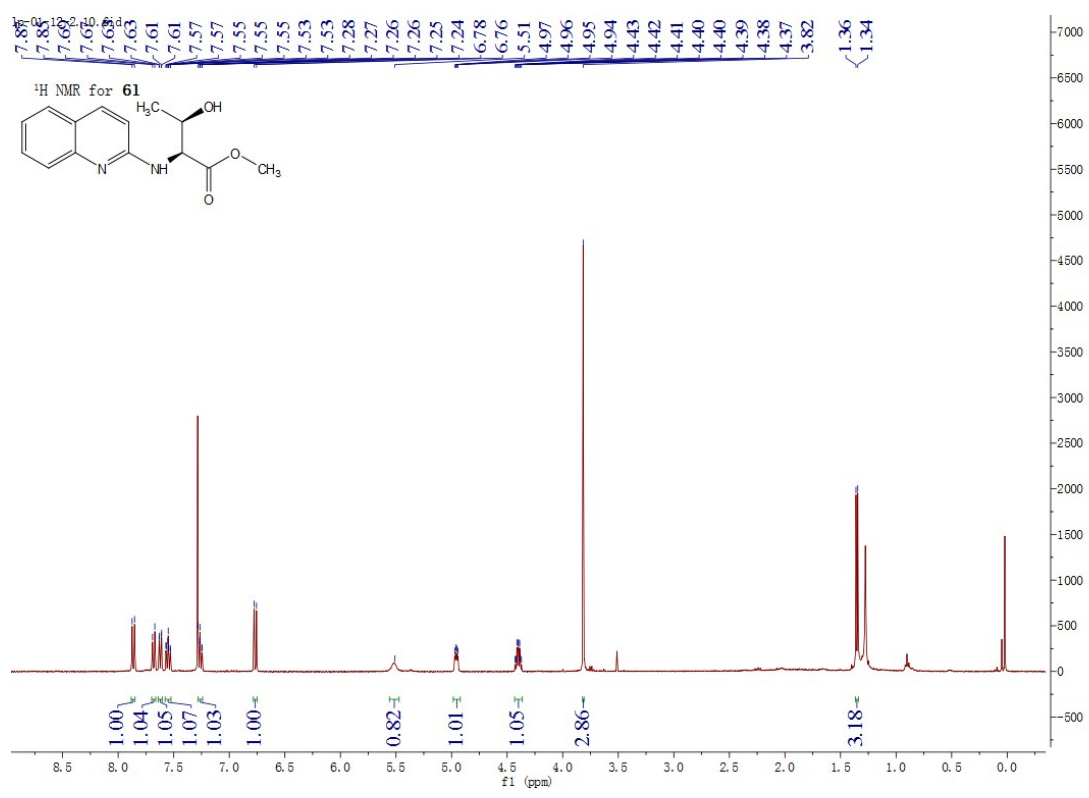
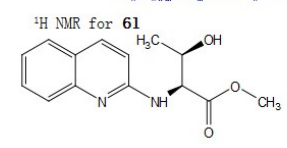


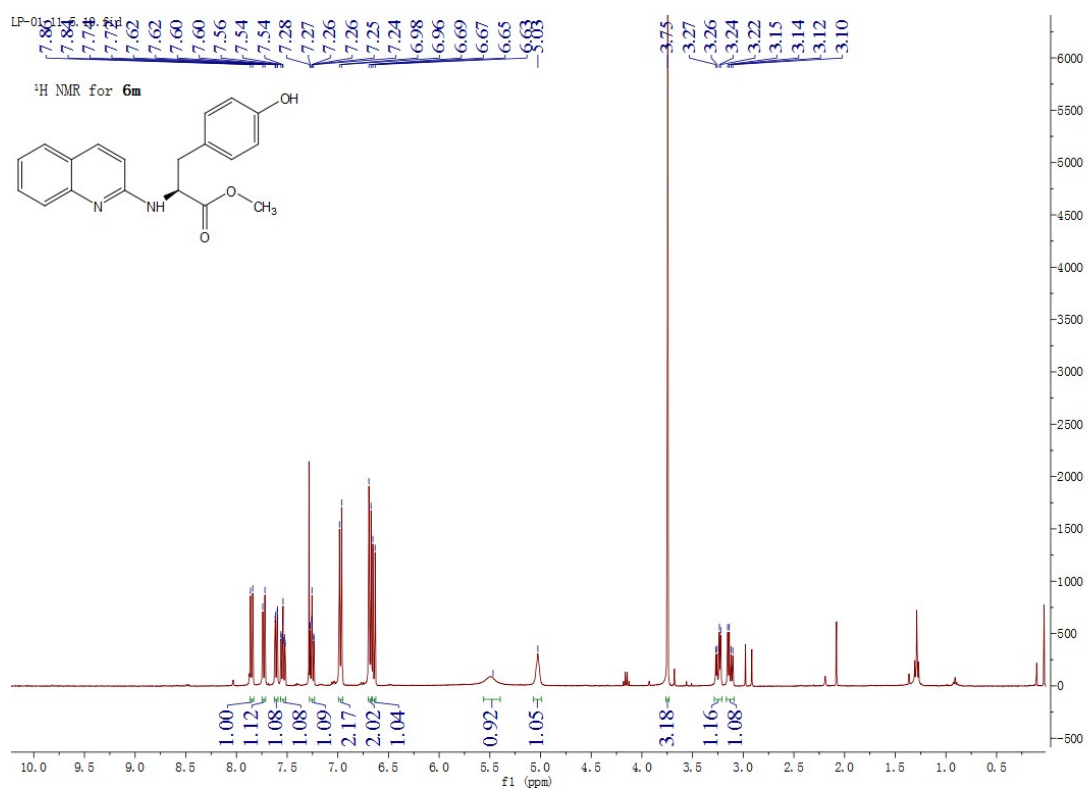
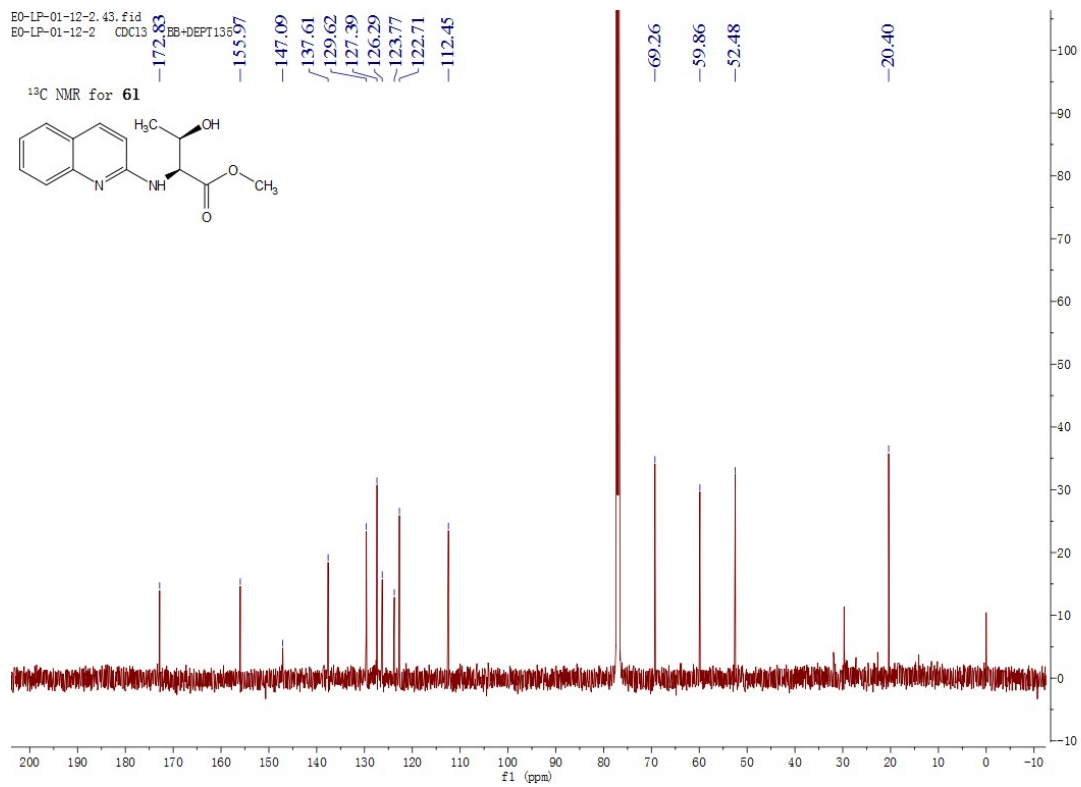
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EO-LP-01-13-2 CDCl3

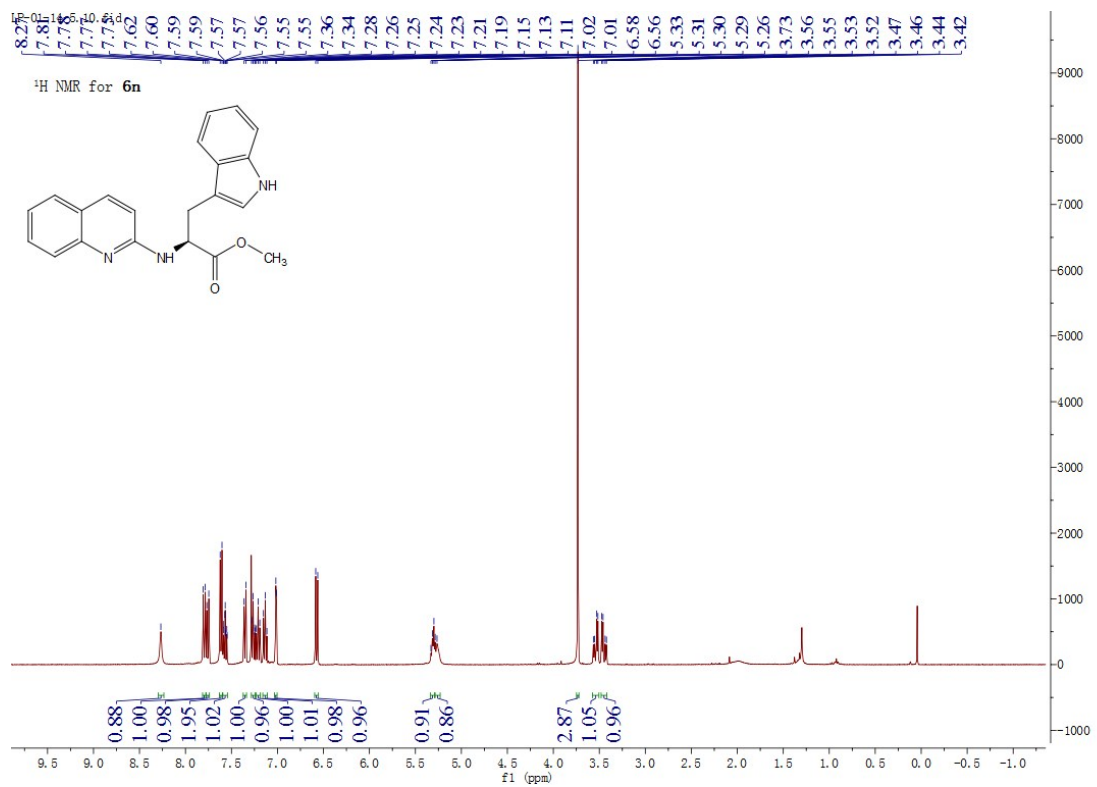
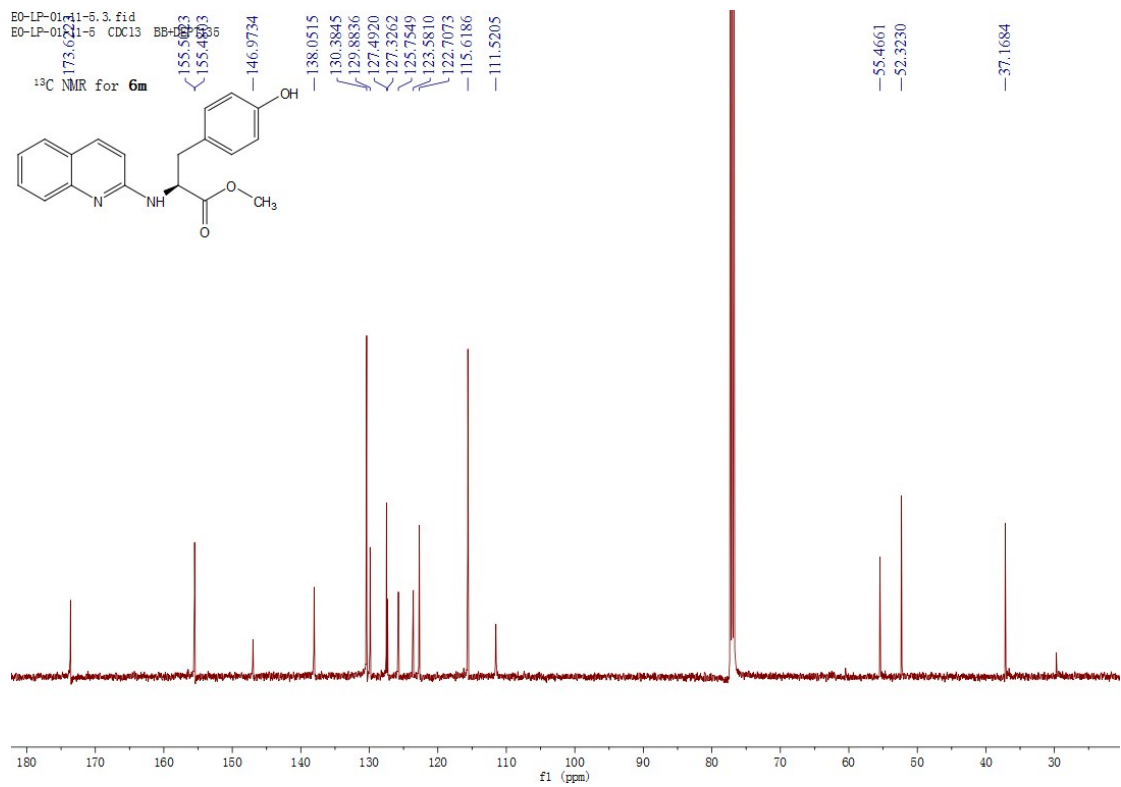
¹³C NMR for **6k**

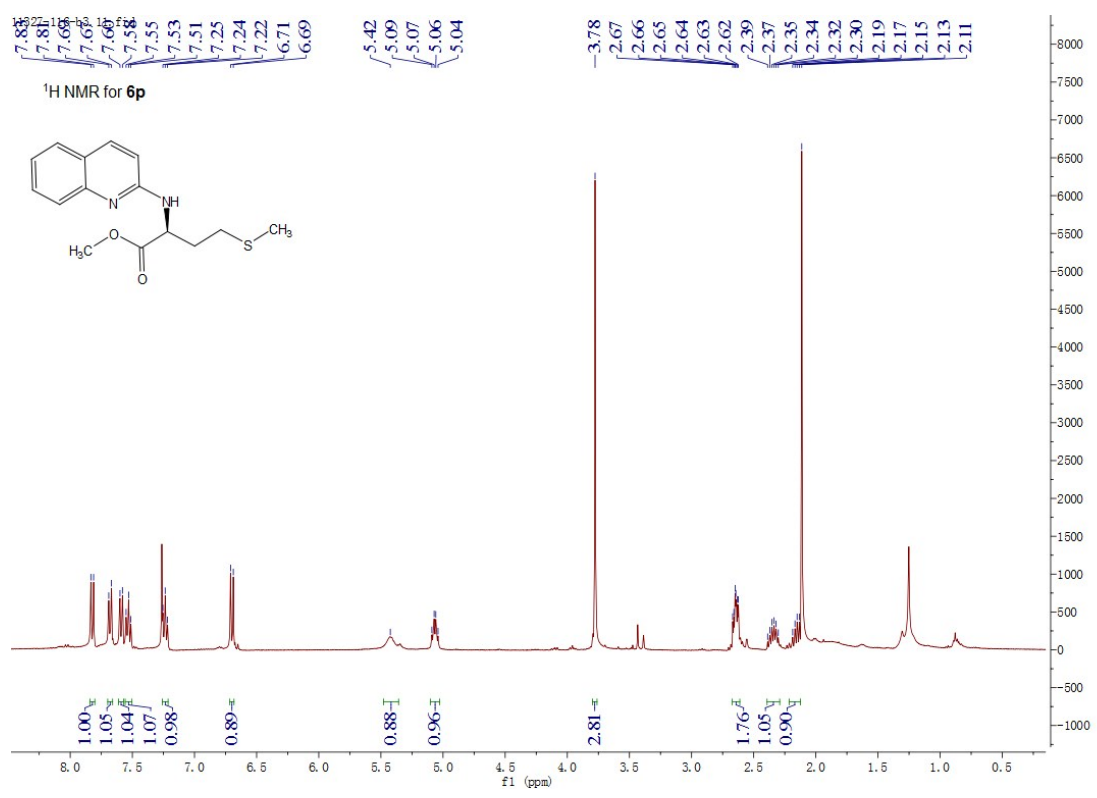
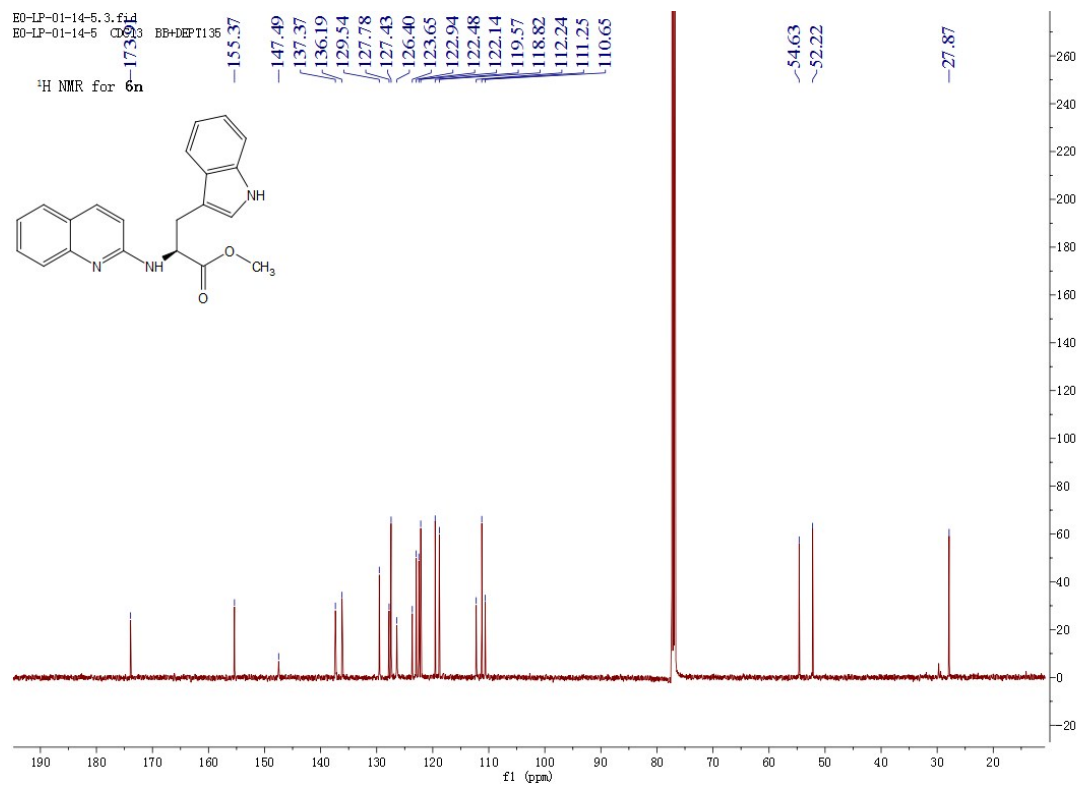


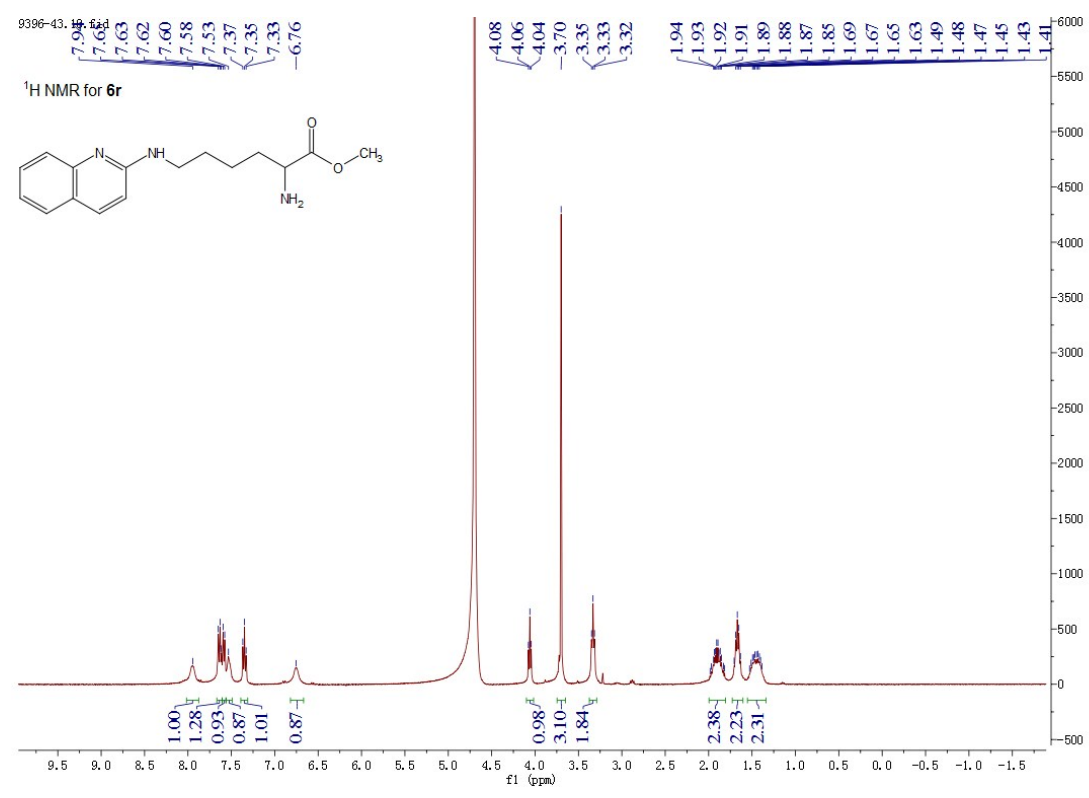
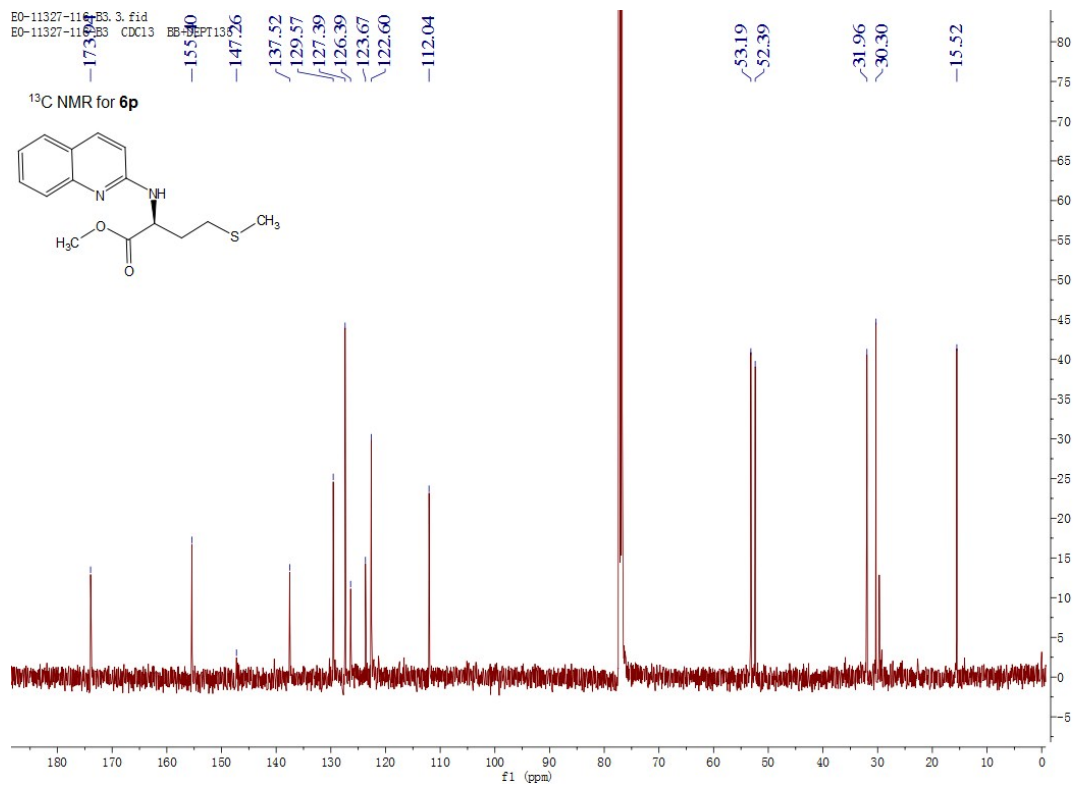
¹H NMR for **6l**









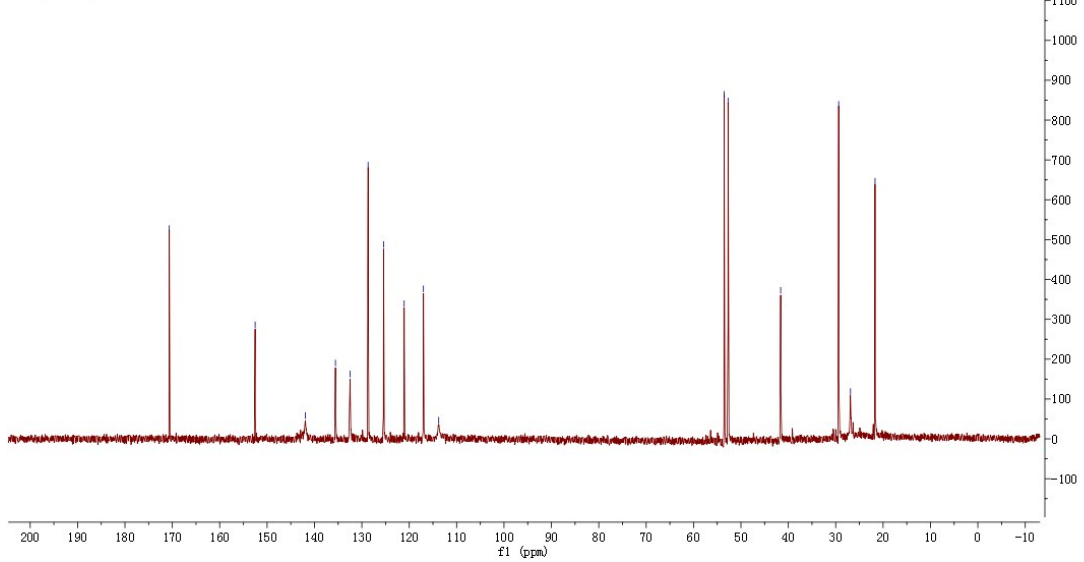
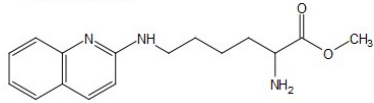


E0-9396-43.3.fid
E0-9396-34.D20 BB+DEPT135

170.95
152.53
141.93
135.57
132.50
128.68
125.42
121.07
117.01
113.78

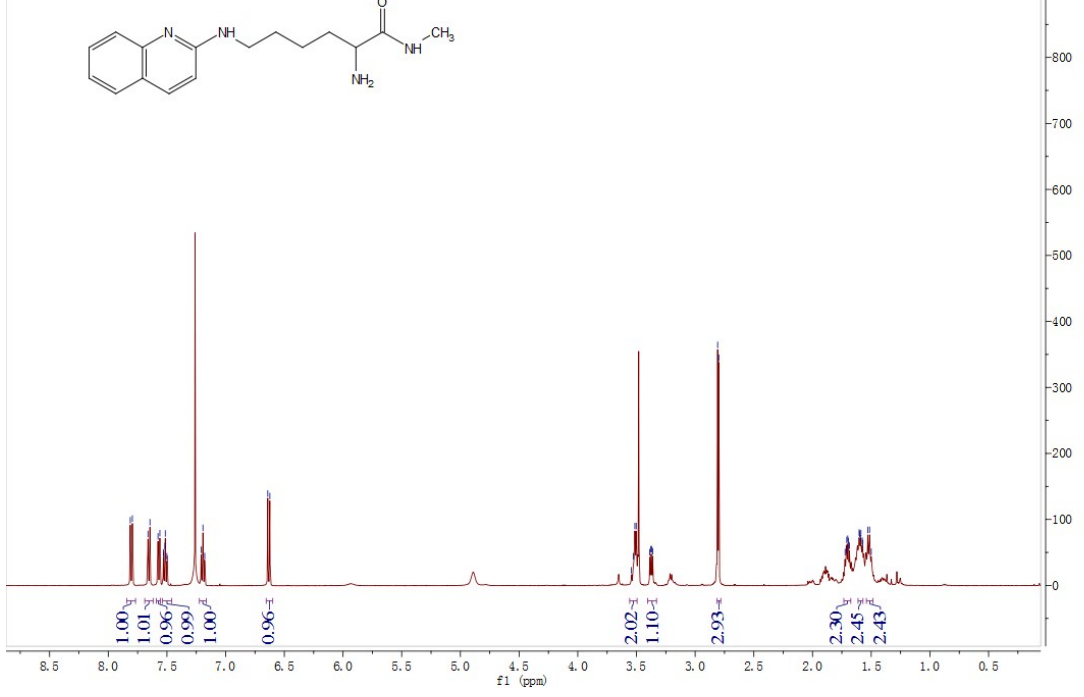
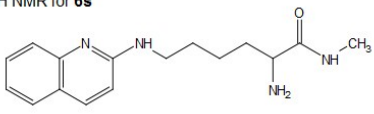
53.56
52.72
41.62
29.37
26.88
21.69

¹³C NMR for 6r



7.73, 7.72, 7.68, 7.66, 7.64, 7.53, 7.52, 7.51, 7.50, 7.50, 7.21, 7.19, 7.18, 6.64, 6.62, 3.54, 3.53, 3.51, 3.50, 3.39, 3.38, 3.37, 3.36, 2.81, 2.80, 1.71, 1.71, 1.70, 1.69, 1.69, 1.60, 1.60, 1.59, 1.59, 1.58, 1.57, 1.53, 1.51

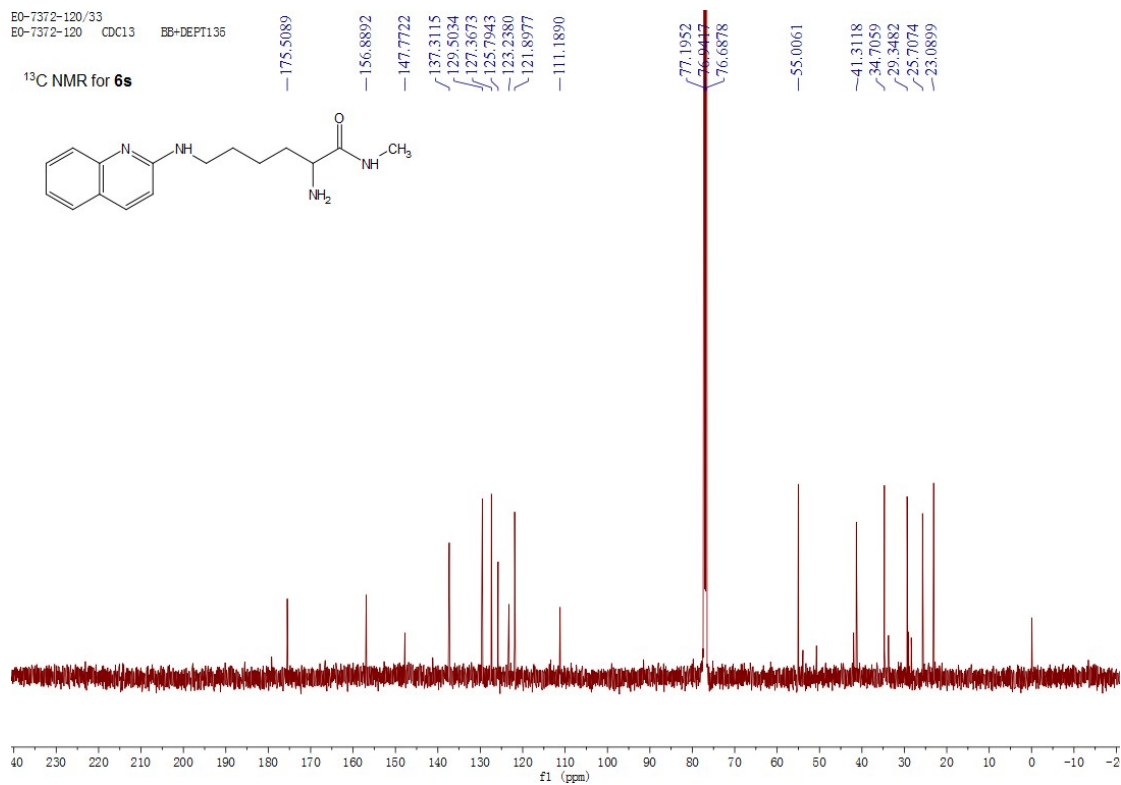
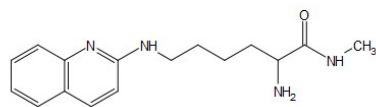
¹H NMR for 6s



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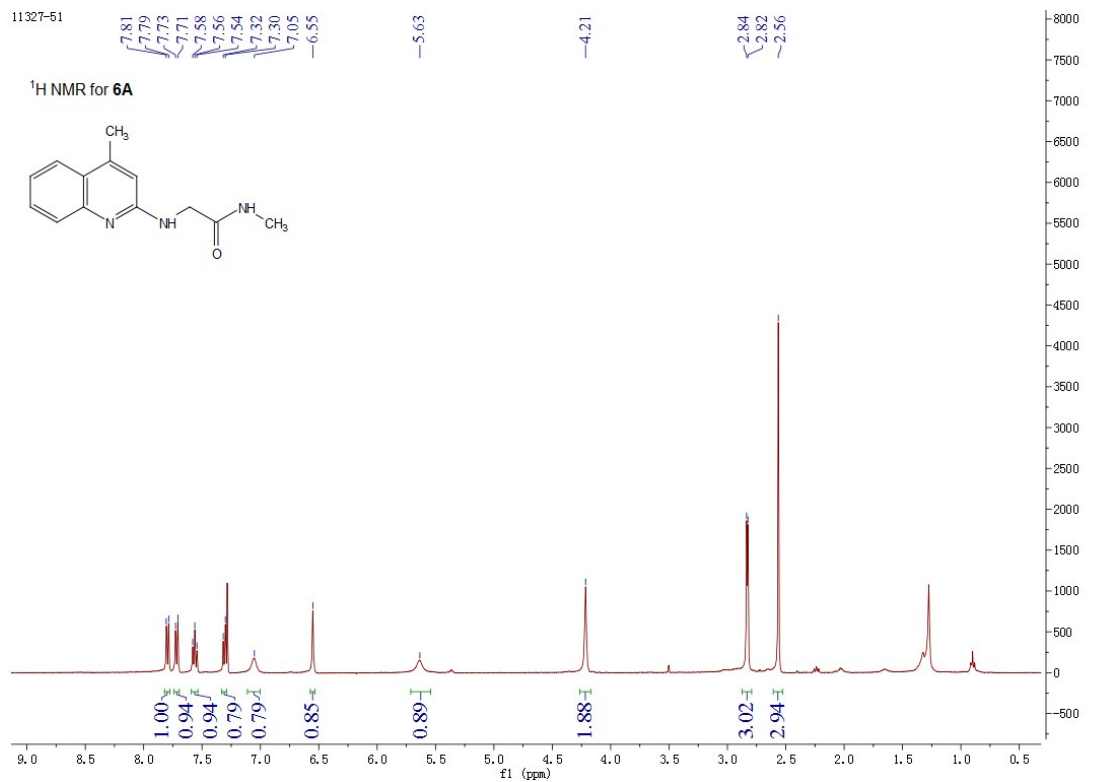
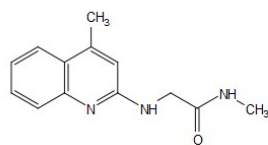
BB+DEPT135

¹³C NMR for 6s



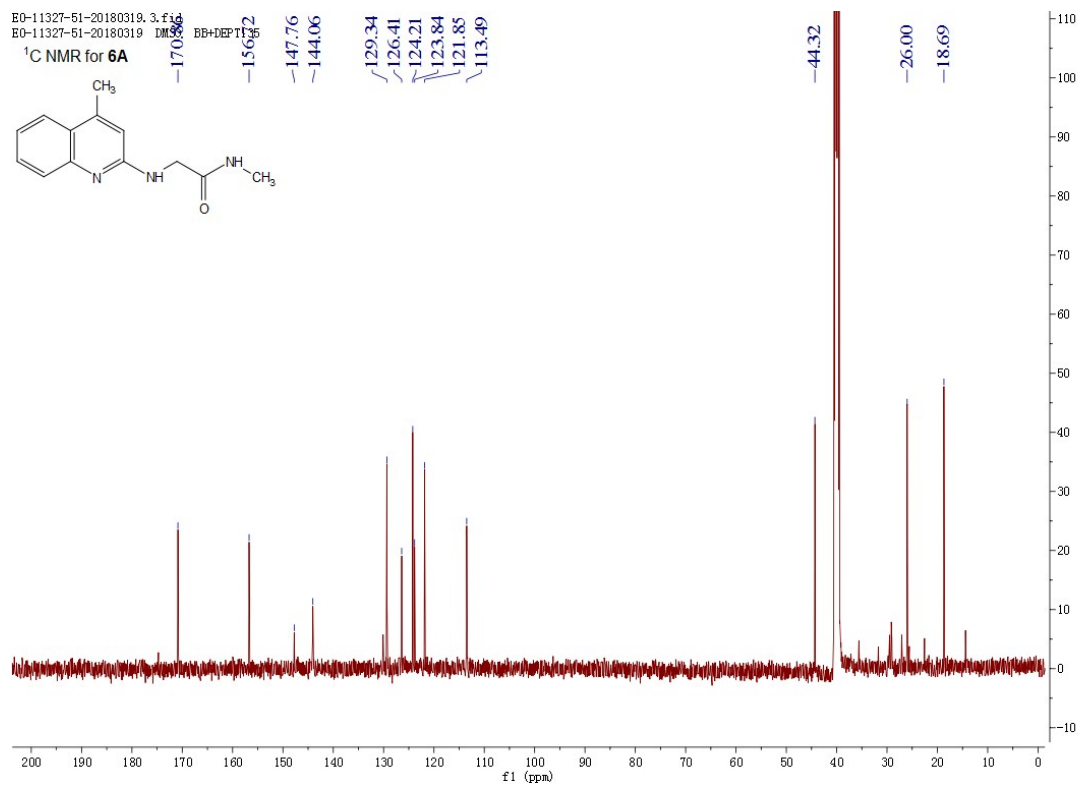
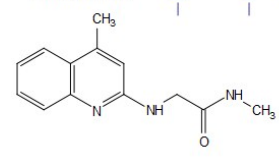
11327-51

¹H NMR for 6A

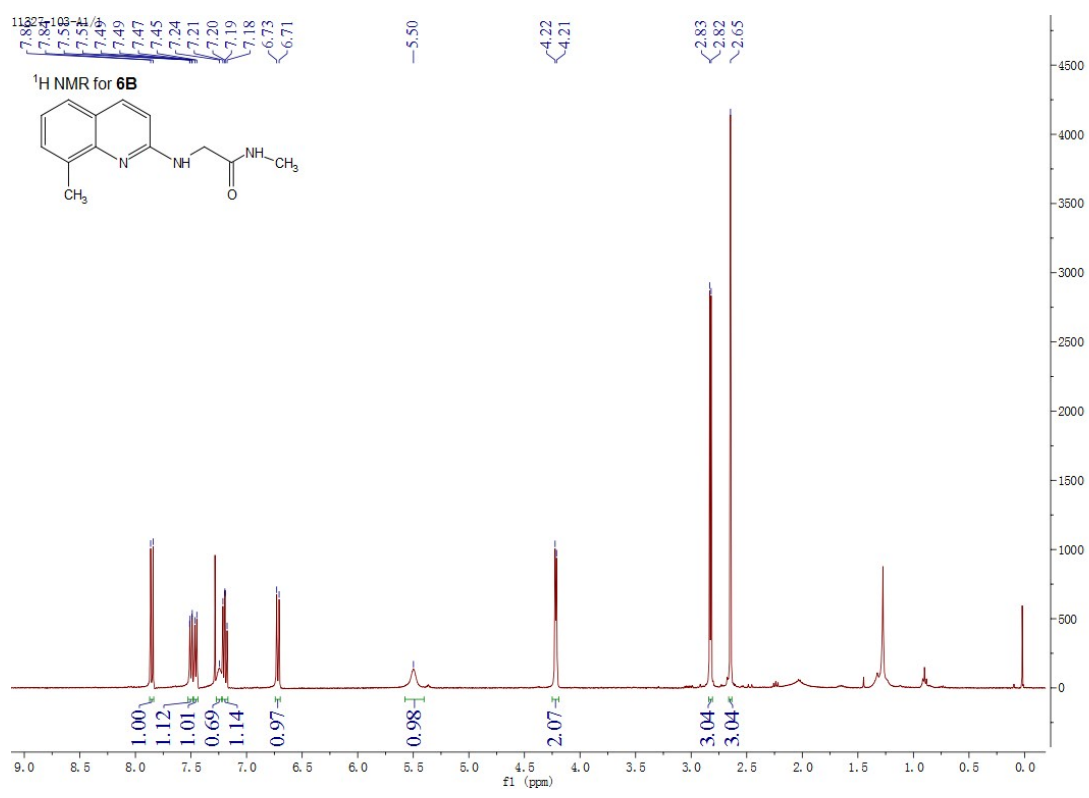
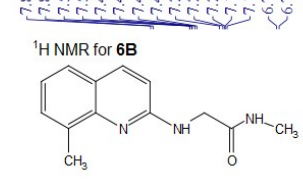


EO-11327-51-20180319_3.fid
 EO-11327-51-20180319.DM3

¹³C NMR for 6A

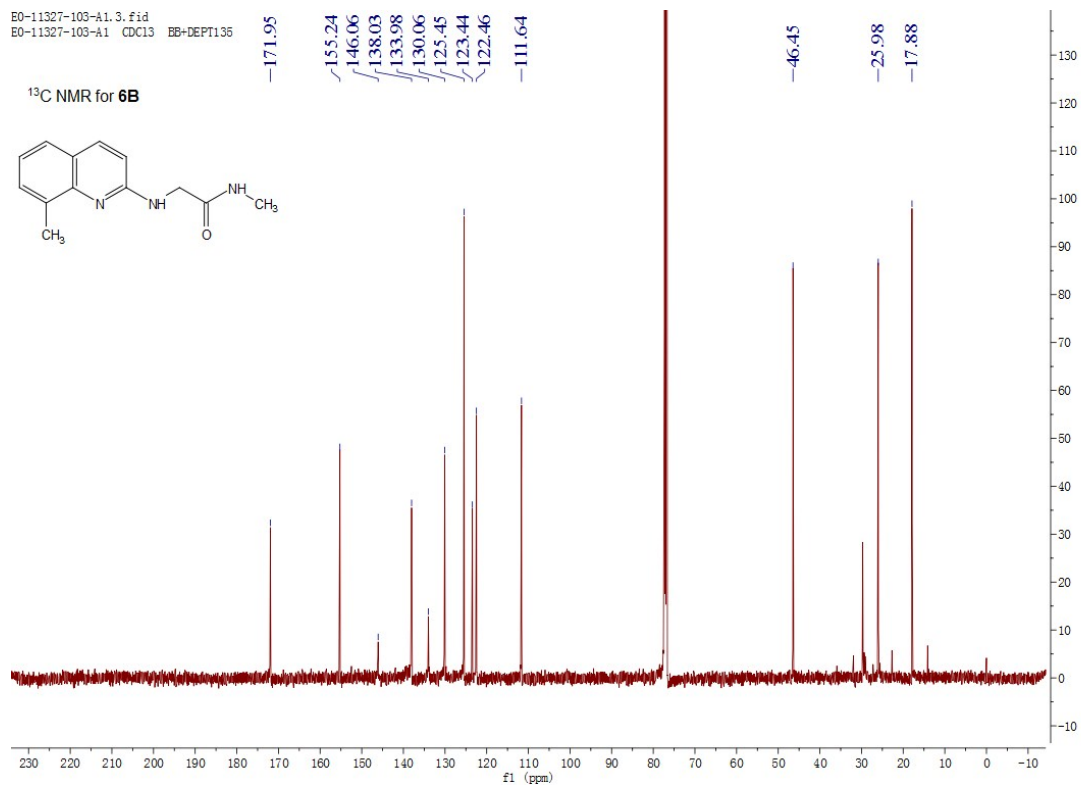
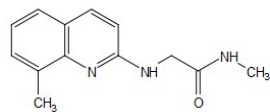


¹H NMR for 6B



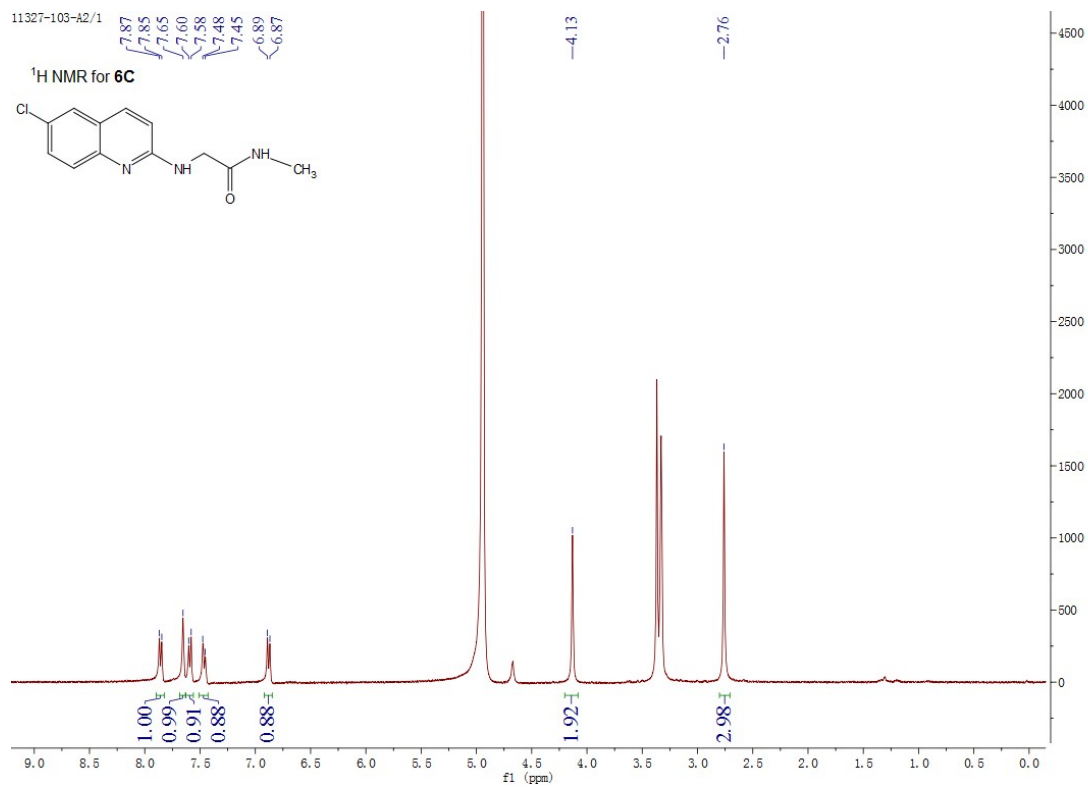
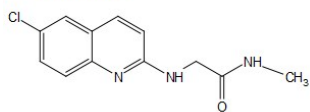
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¹³C NMR for 6B



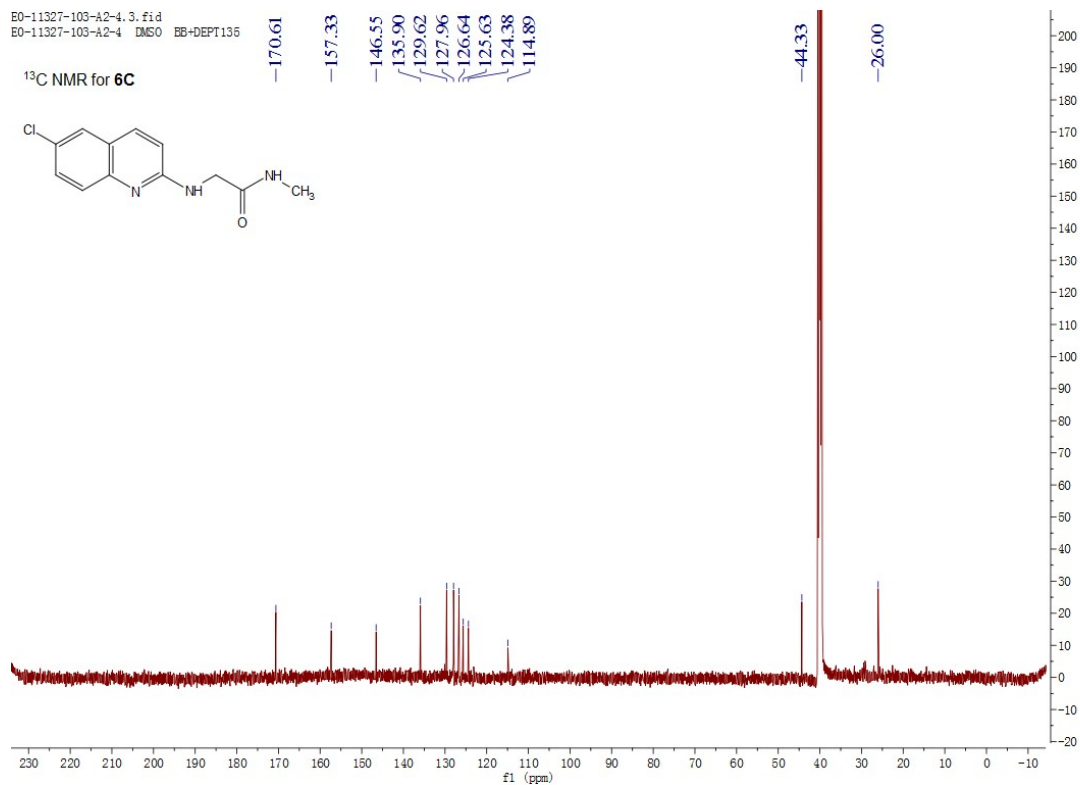
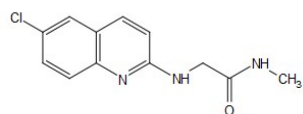
11327-103-A2/1

¹H NMR for 6C



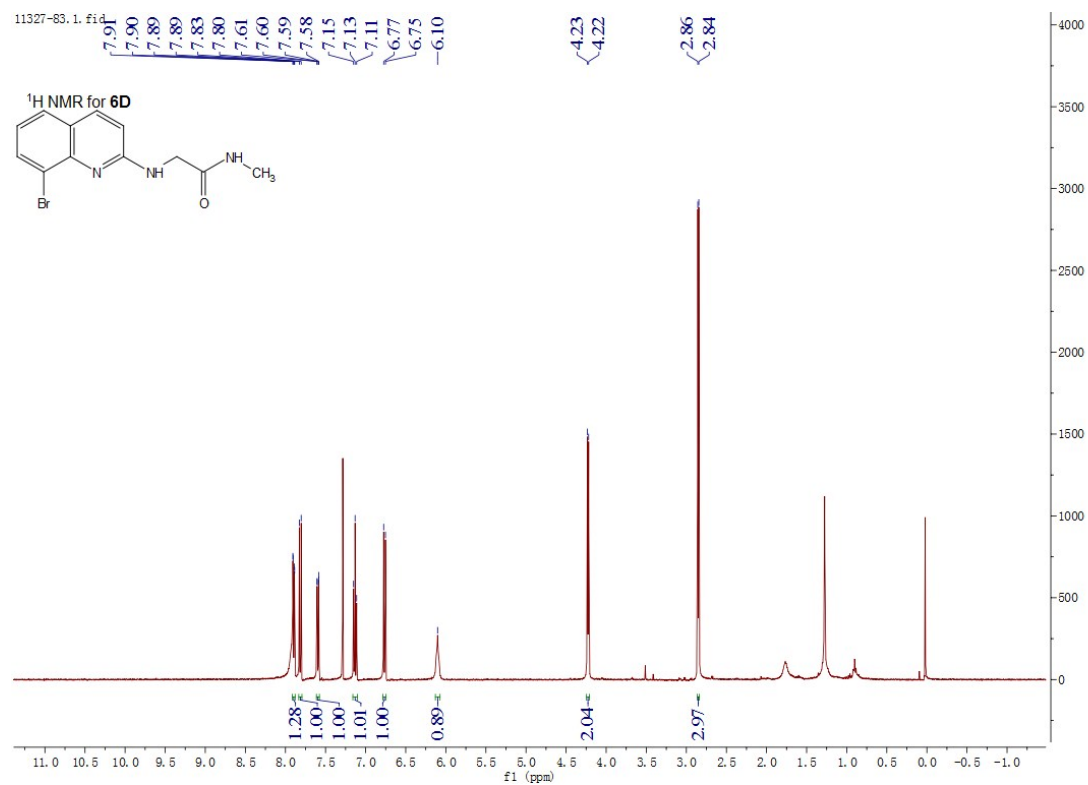
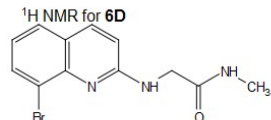
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¹³C NMR for 6C



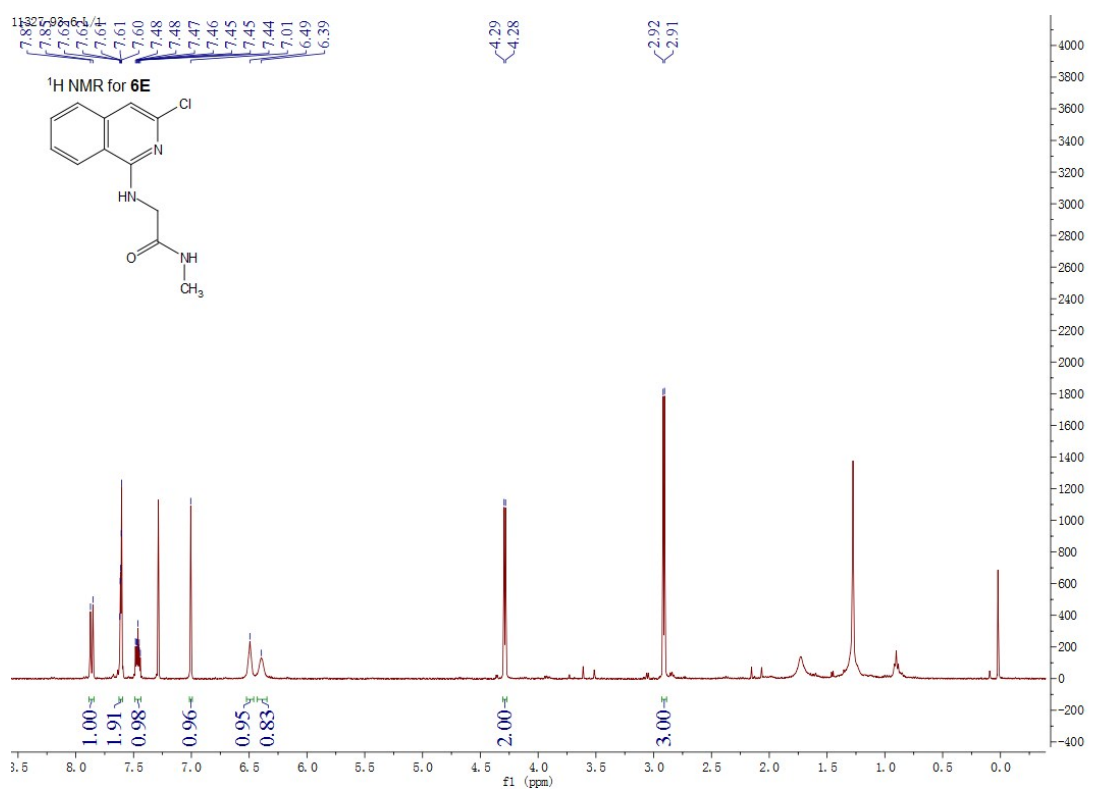
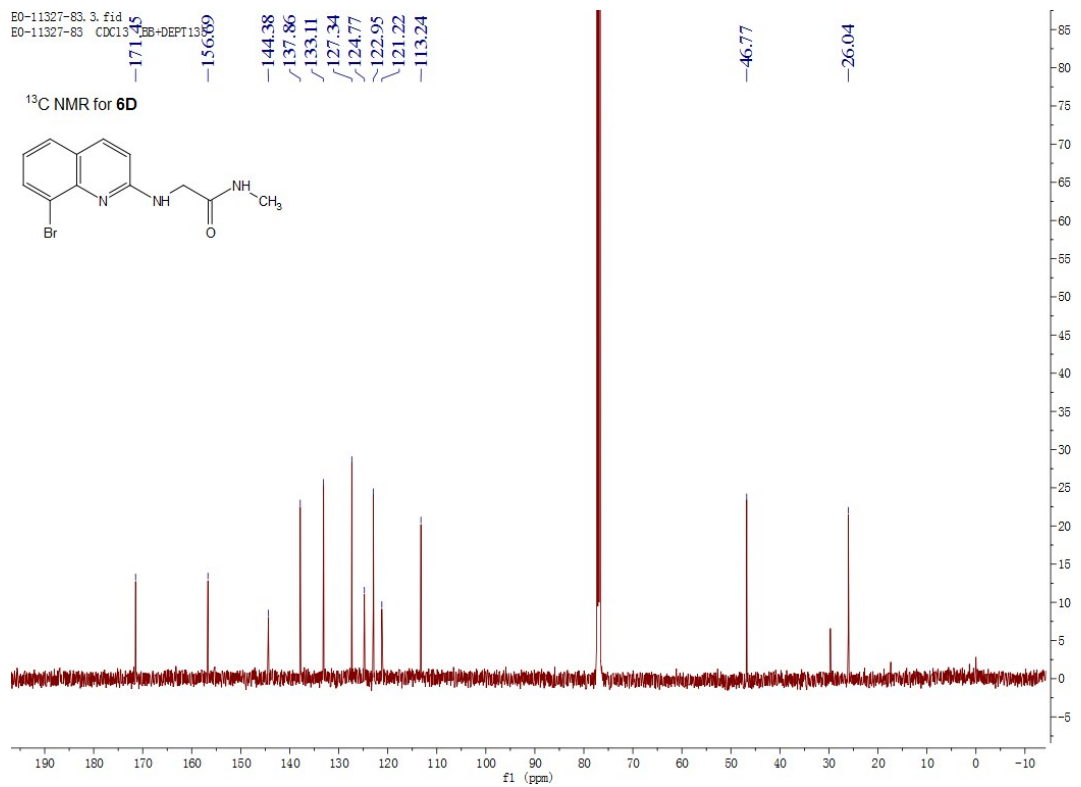
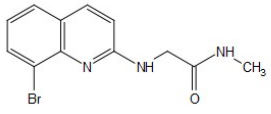
11327-83.1.fid

¹H NMR for 6D



EO-11327-83, 3. fid
EO-11327-83 CDC13

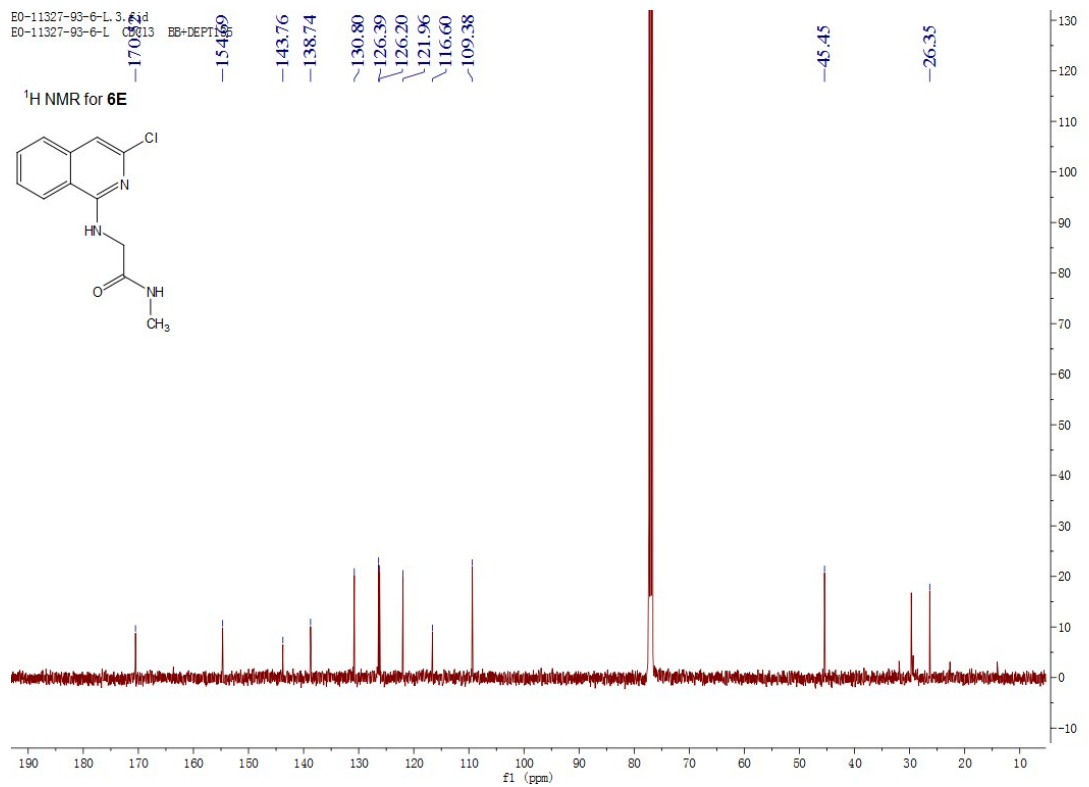
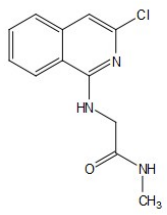
¹³C NMR for 6D



EO-11327-93-6-L-3-6-13
EO-11327-93-6-L-3-6-13

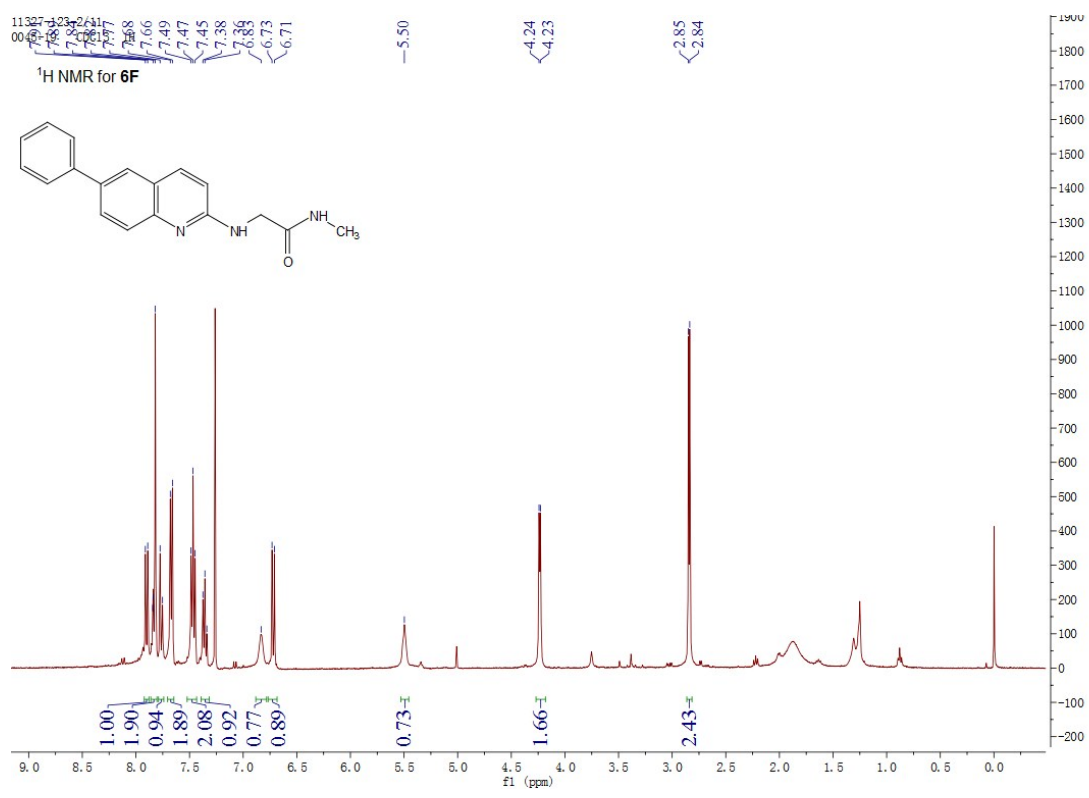
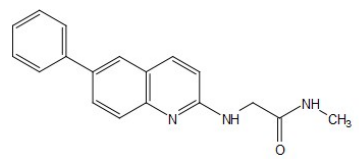
EB-DEPT

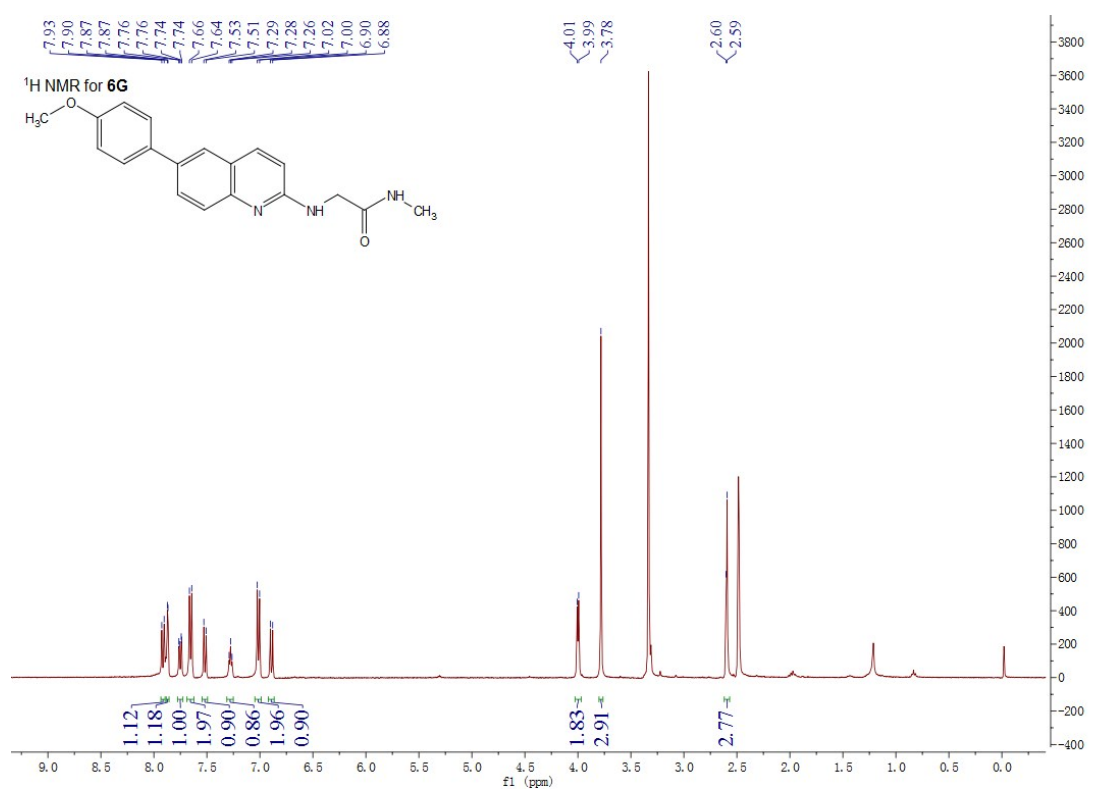
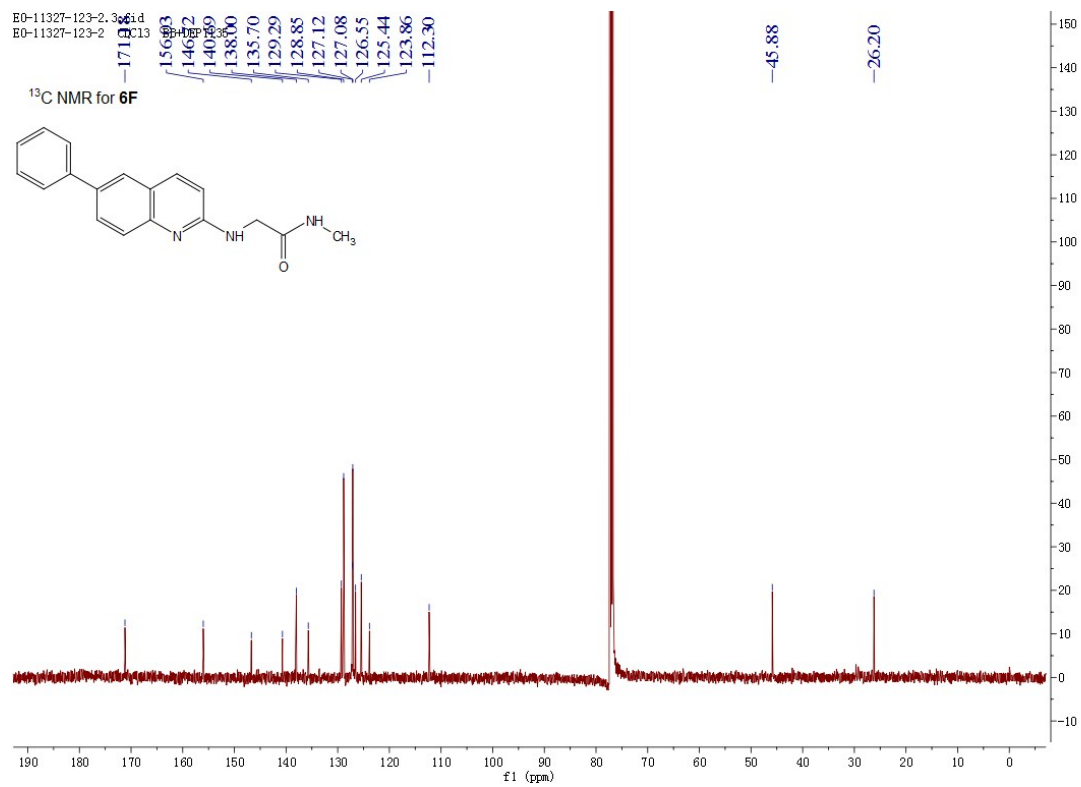
¹H NMR for 6E



11327-93-6-L-3-6-13

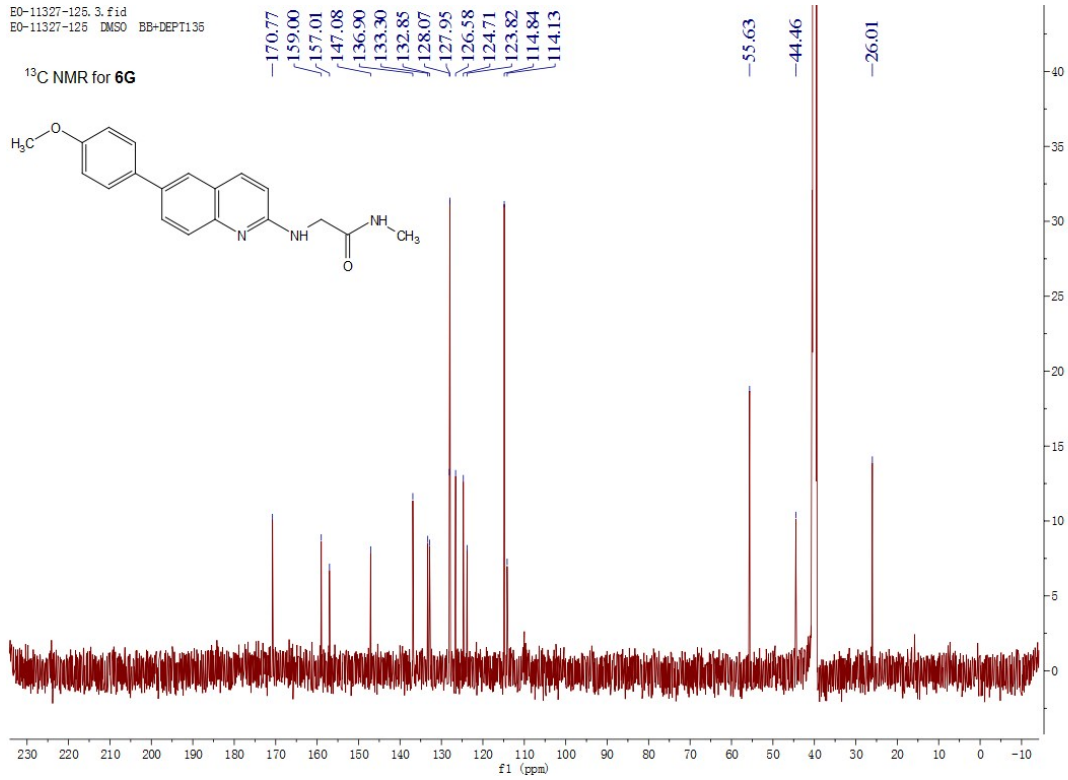
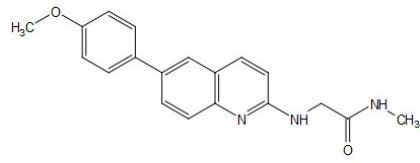
¹H NMR for 6F





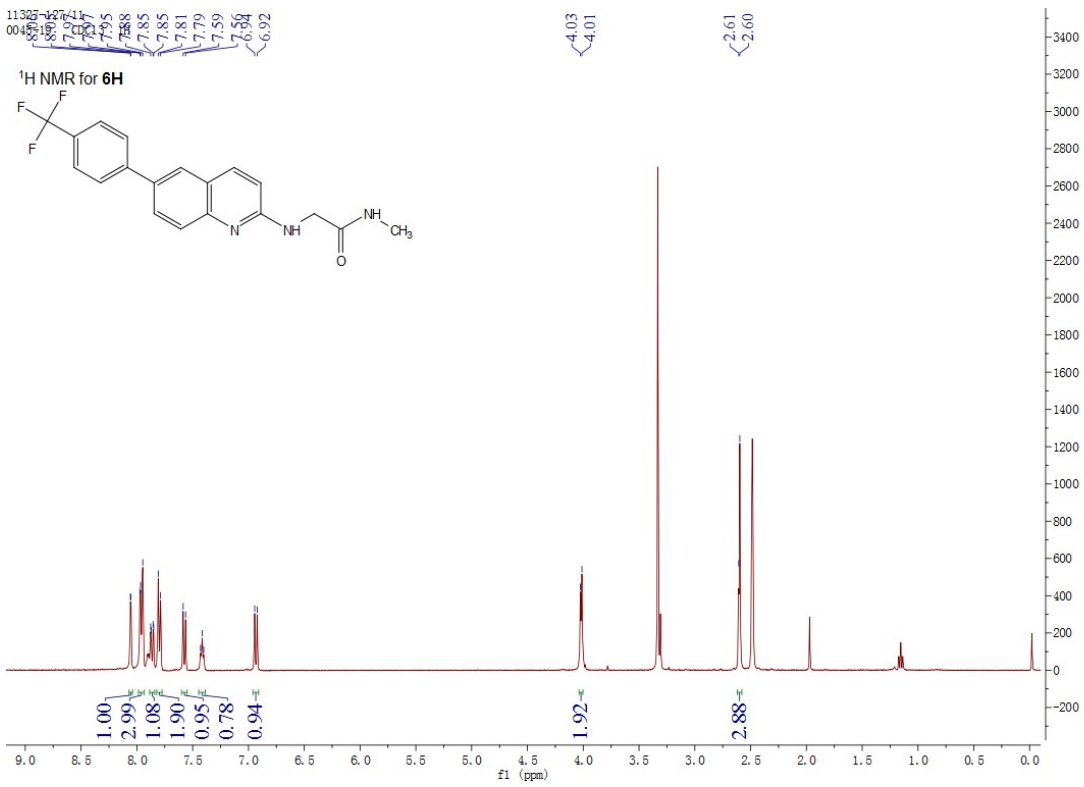
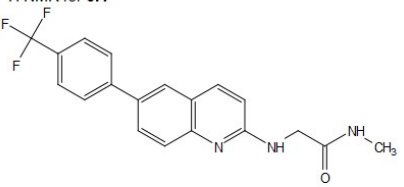
EO-11327-125.3.fid
EO-11327-125 DMSO BB+DEPT135

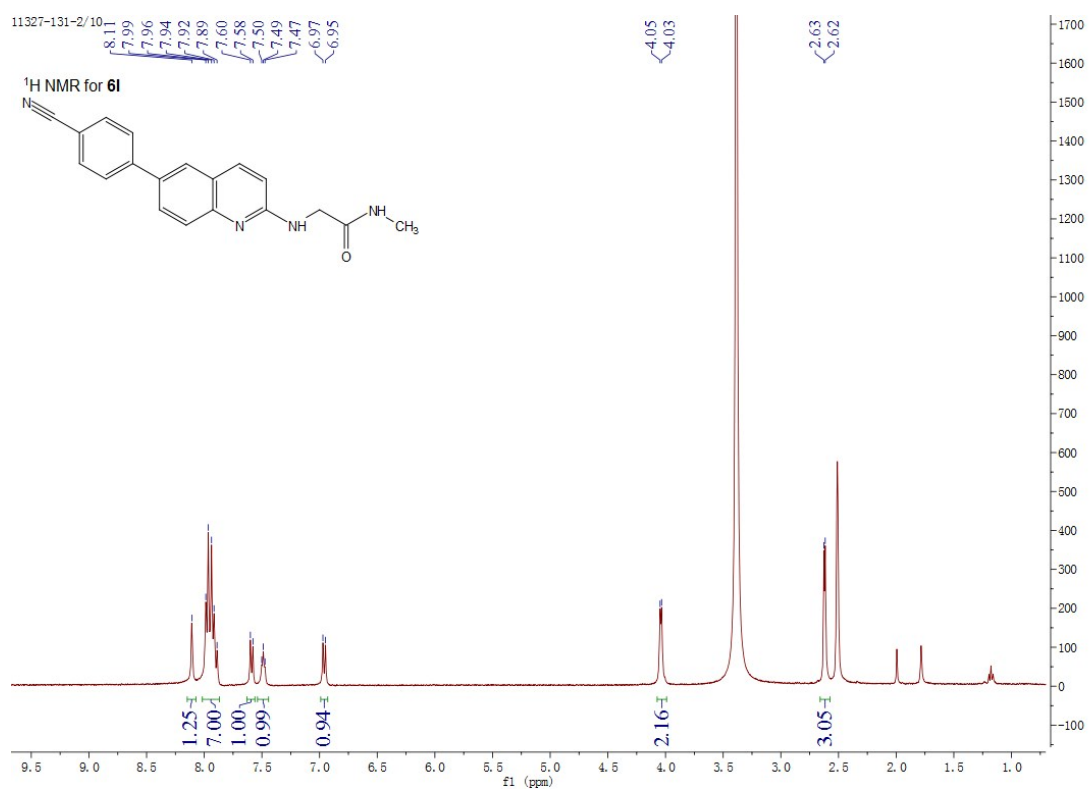
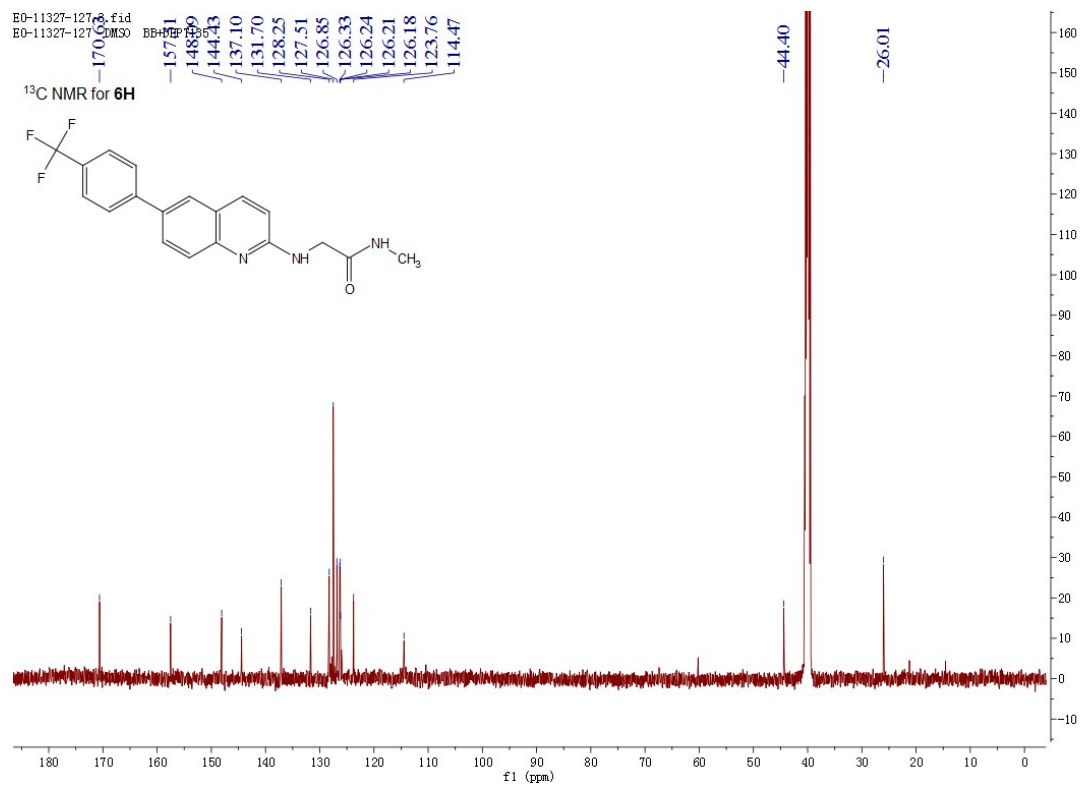
¹³C NMR for 6G



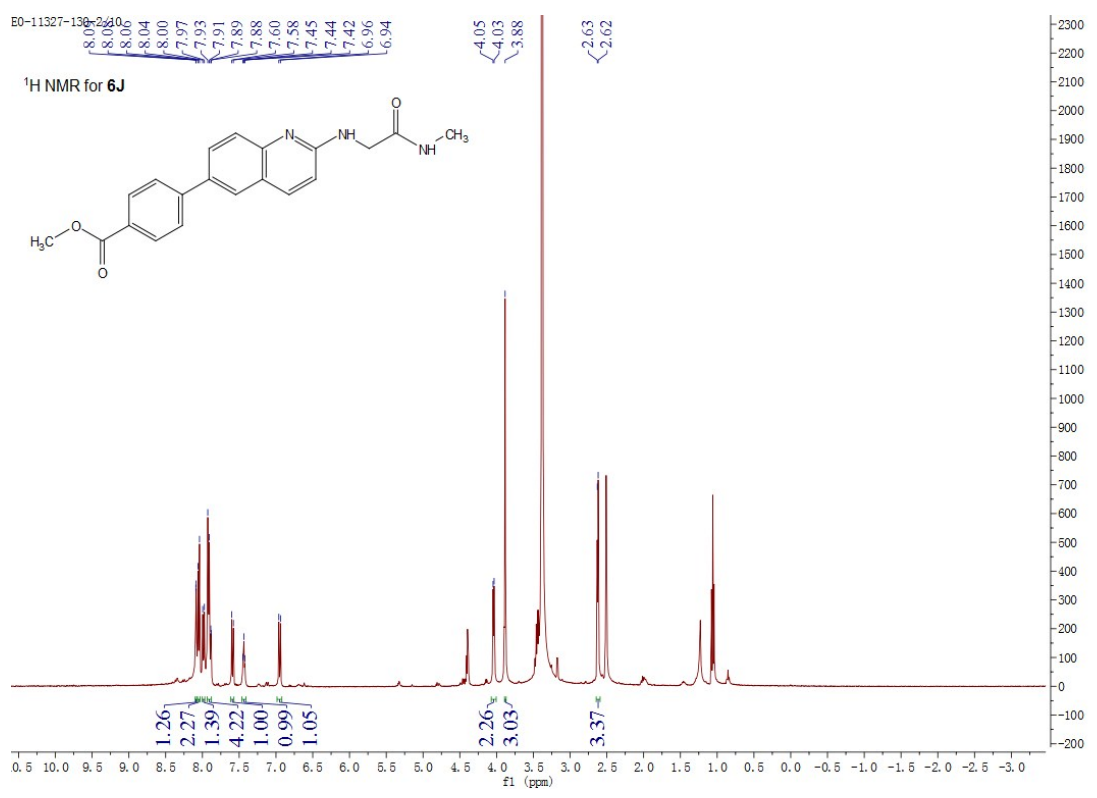
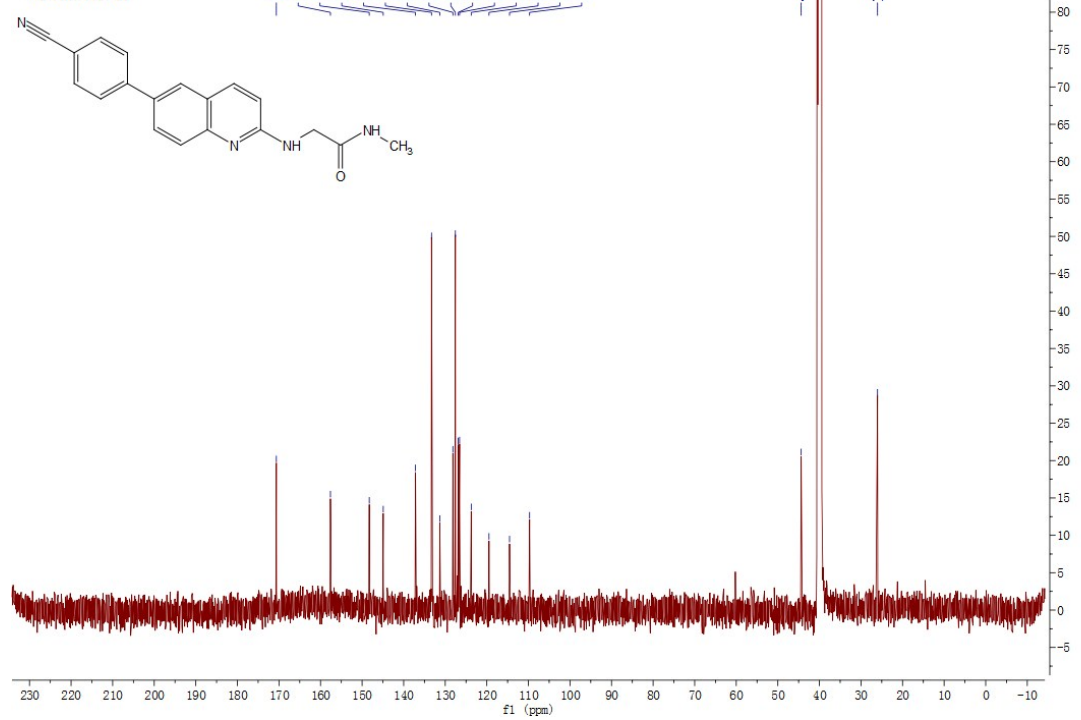
11325 107.41
0045 107.41
7.95
7.95
7.88
7.85
7.81
7.79
7.59
6.94
6.92

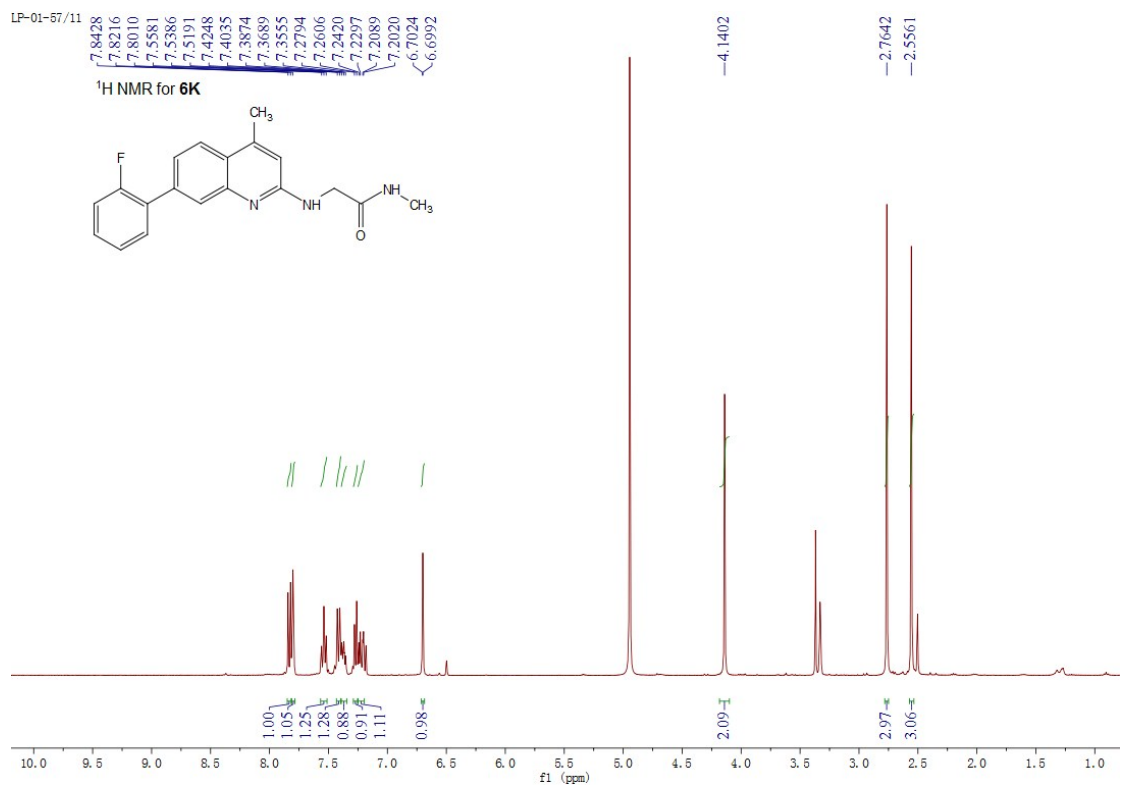
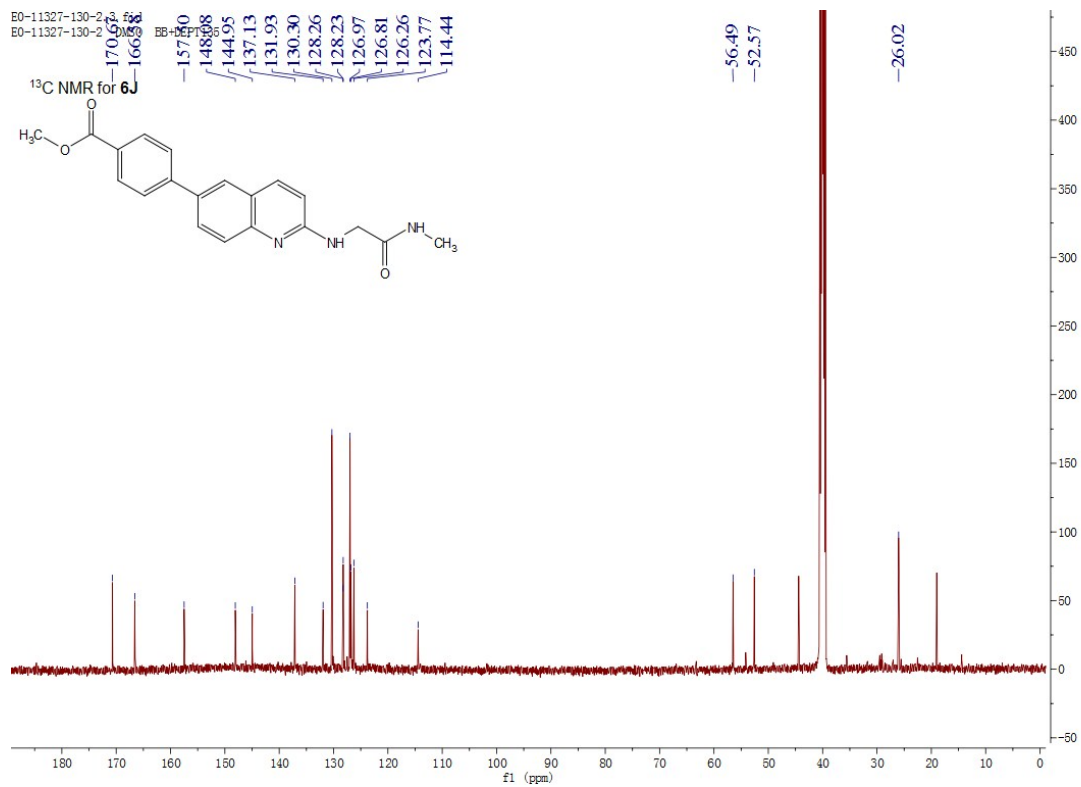
¹H NMR for 6H

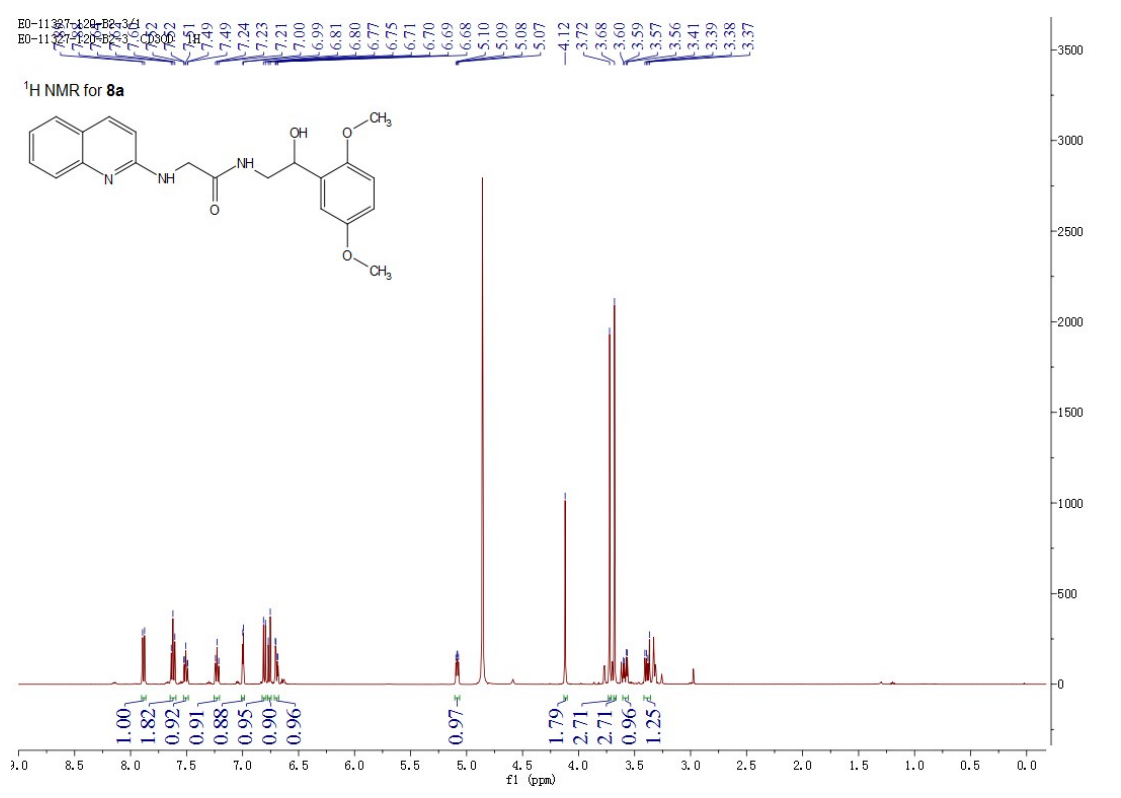
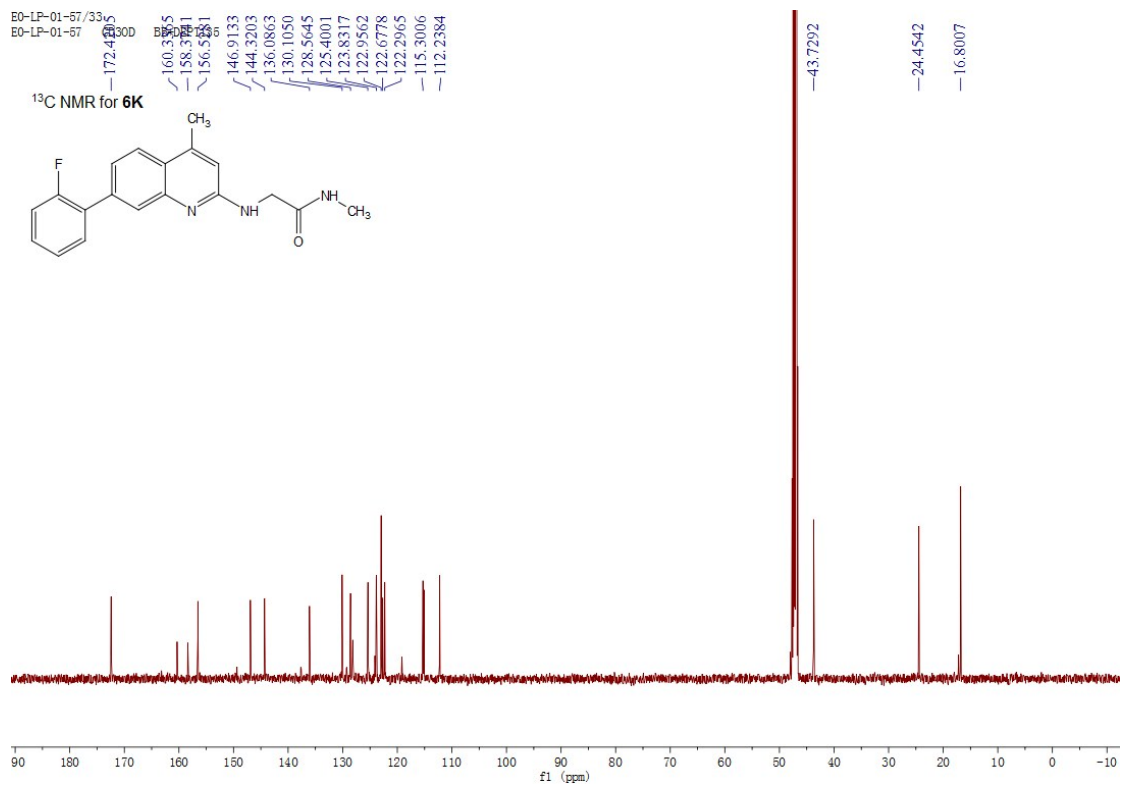




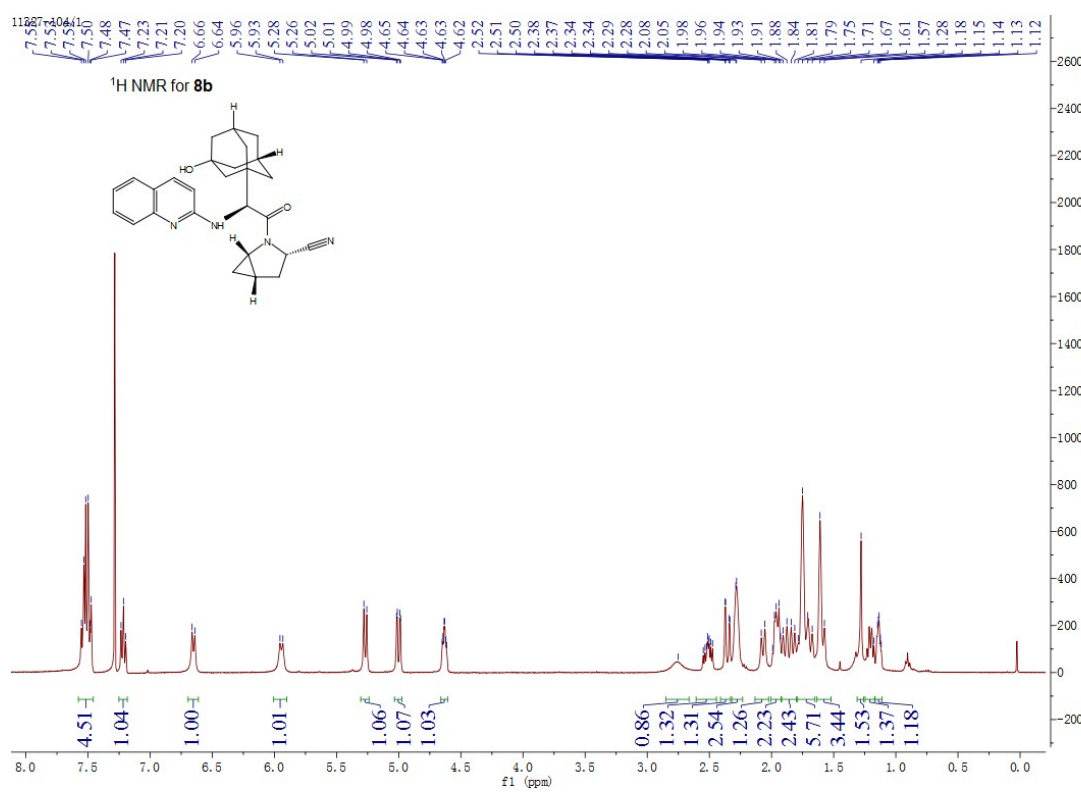
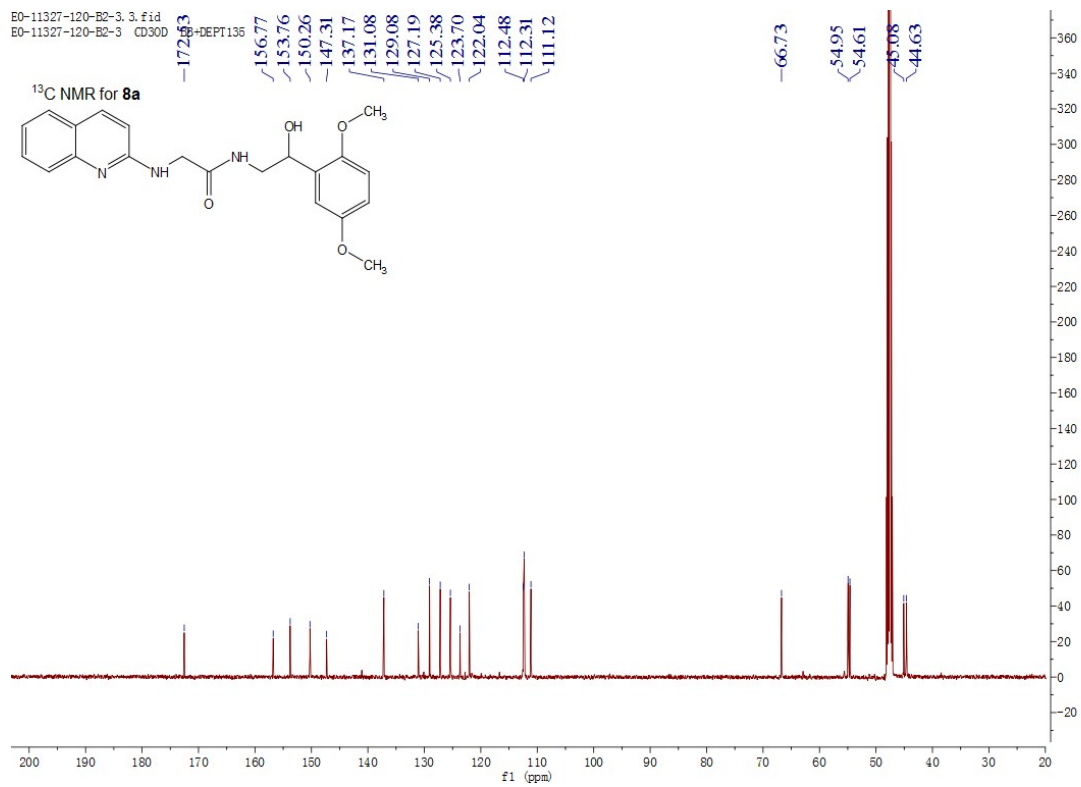
E0-11327-131-2_3.fid
E0-11327-131-2 DMSO BB+DEPT135
¹³C NMR for 6I

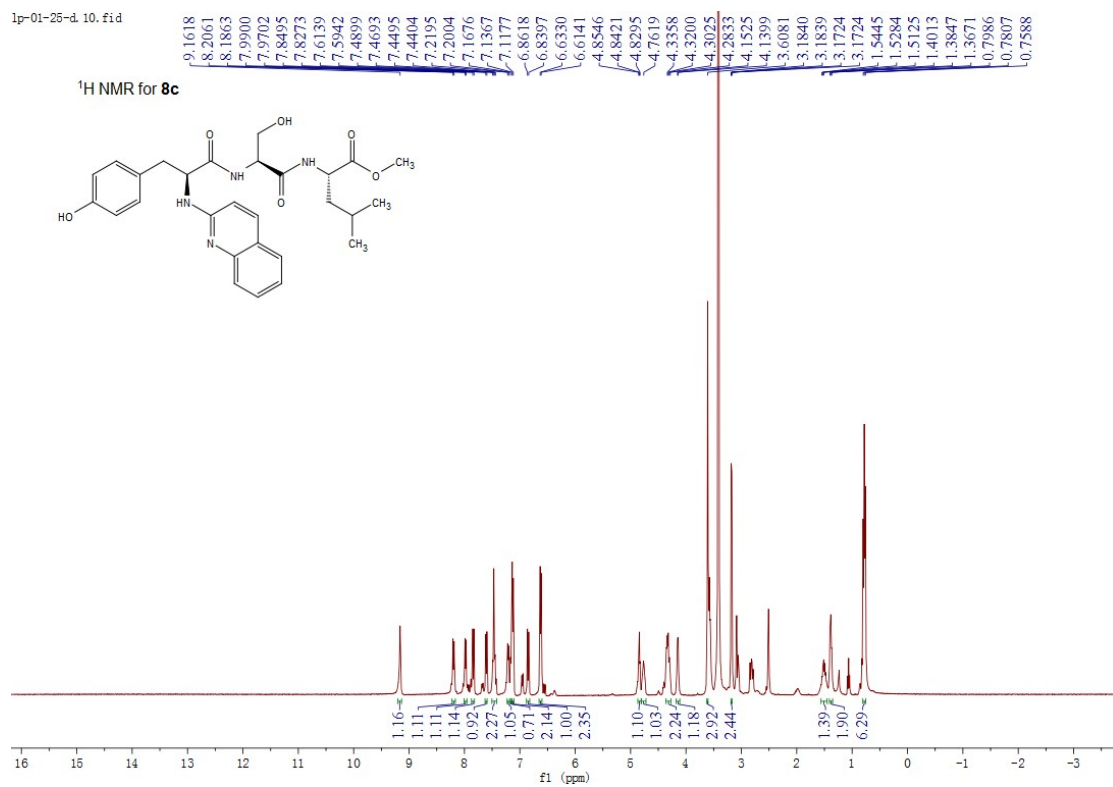
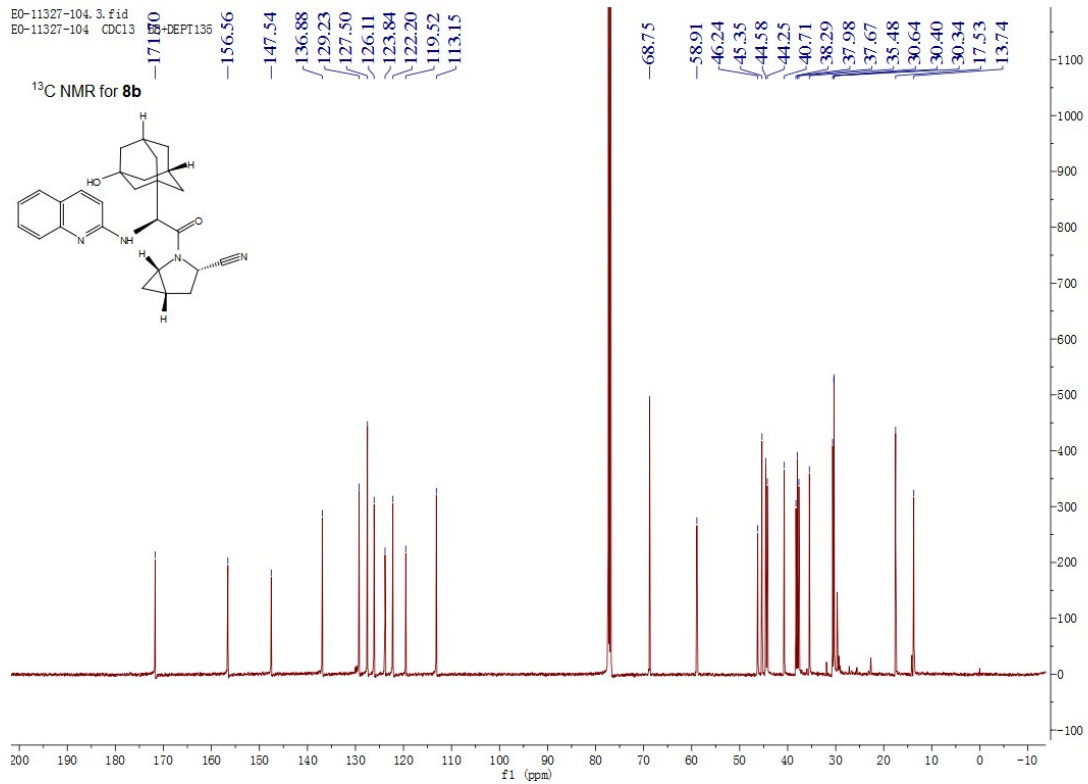




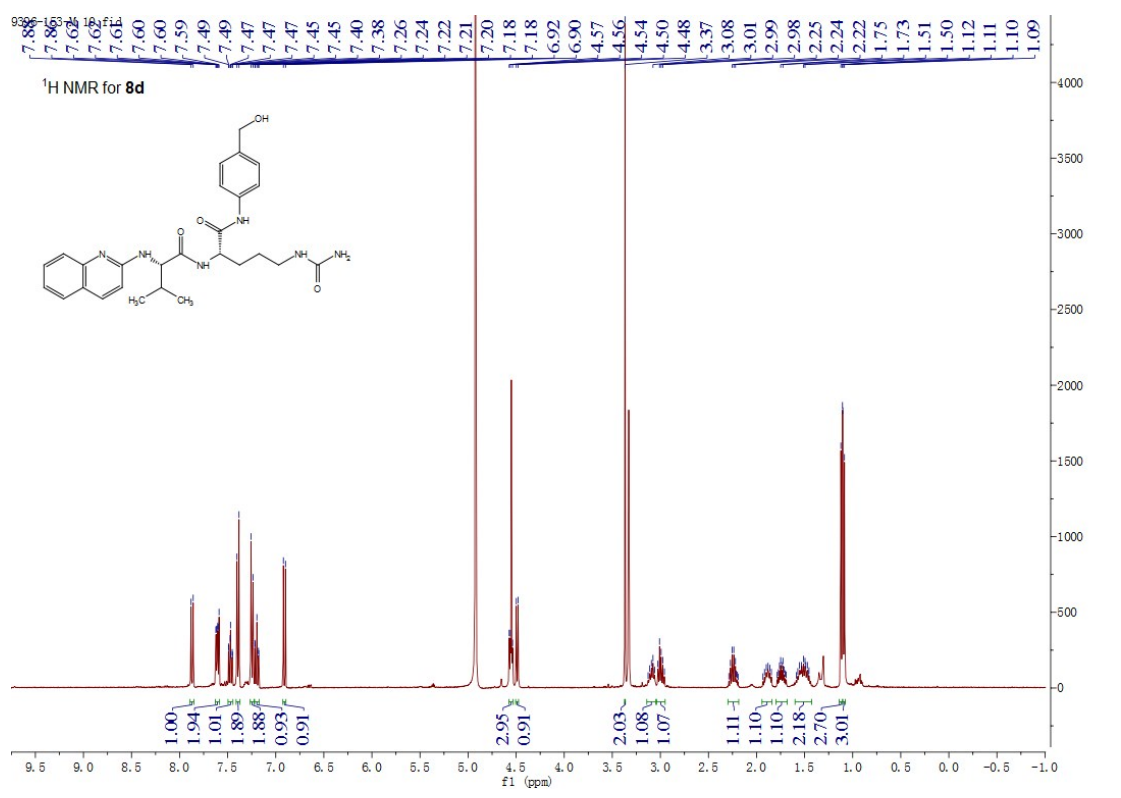
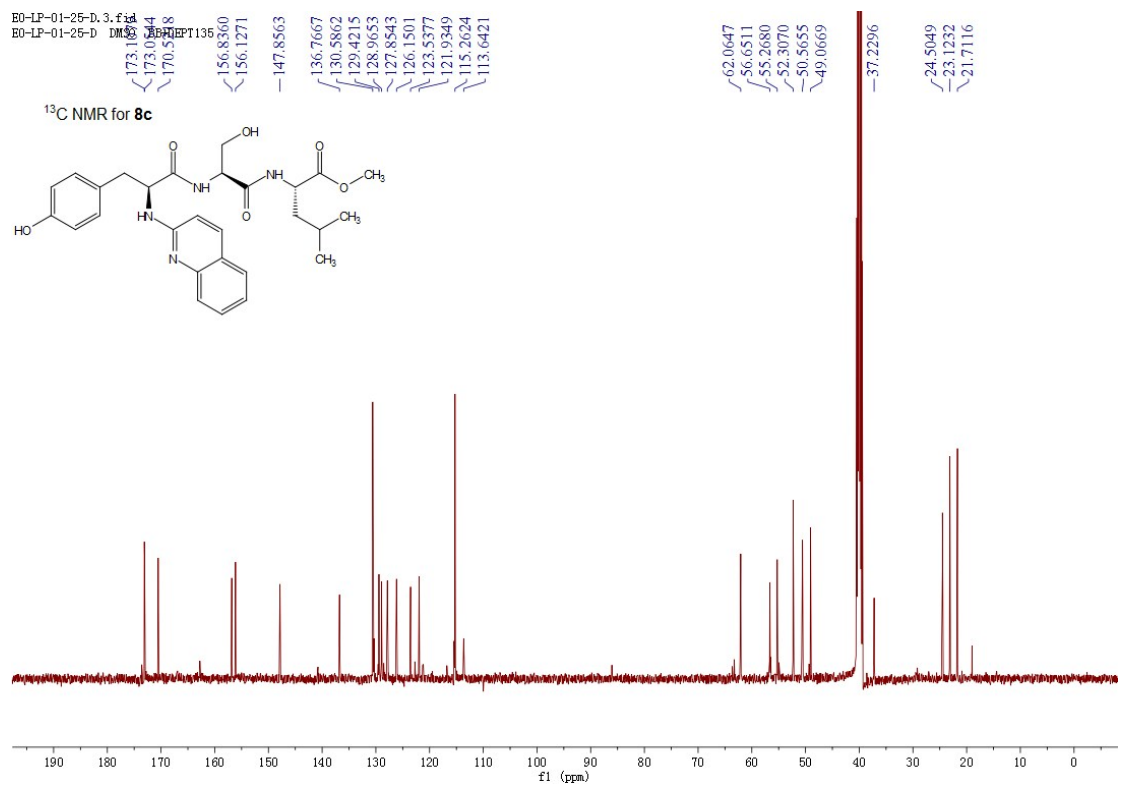


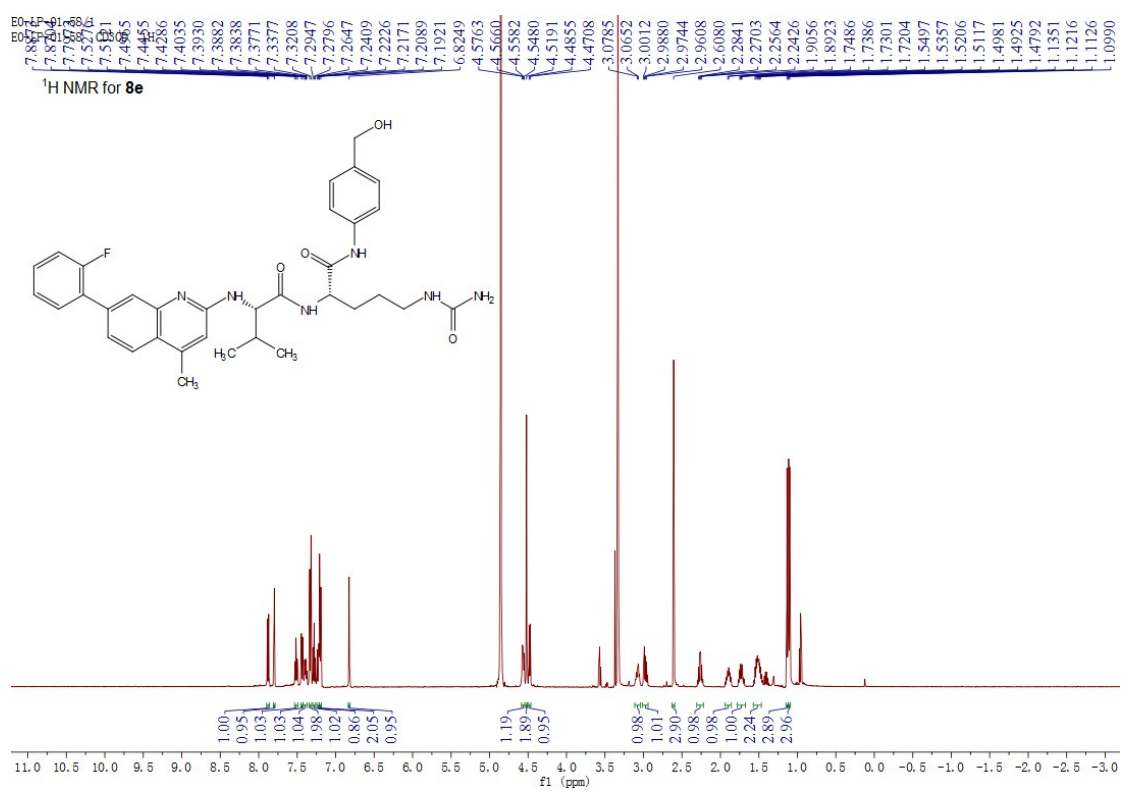
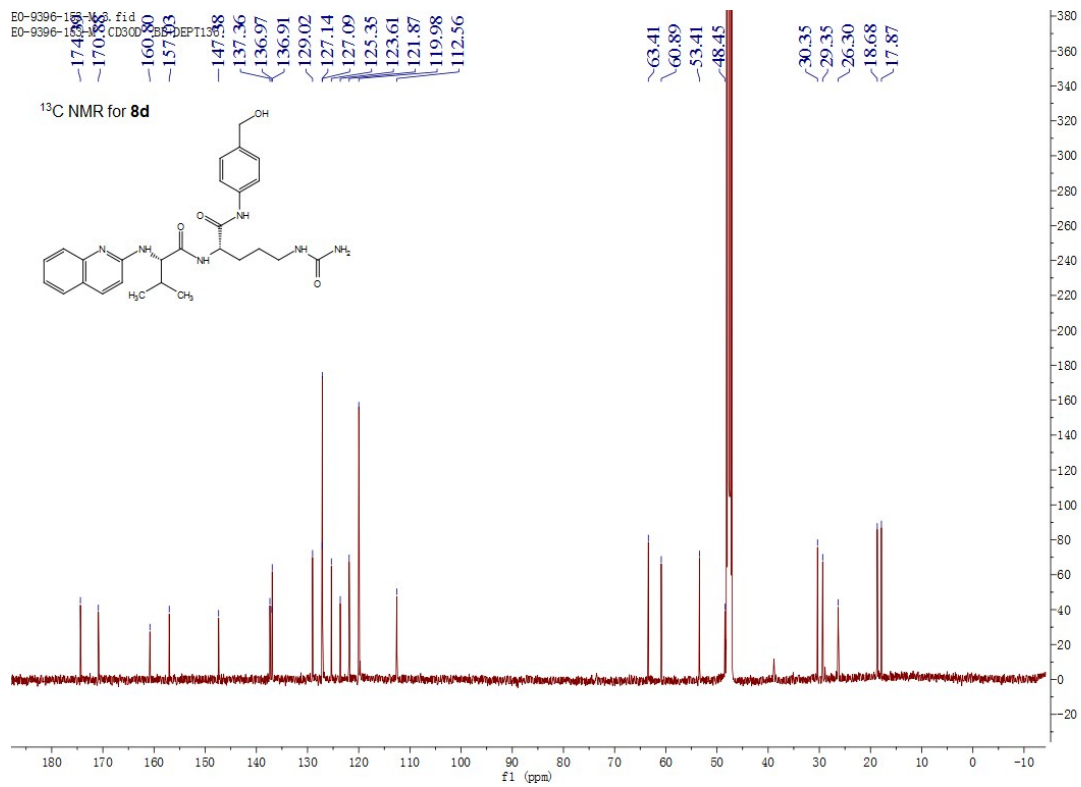
EO-11327-120-B2-3, 3.fid
EO-11327-120-B2-3 CD30D



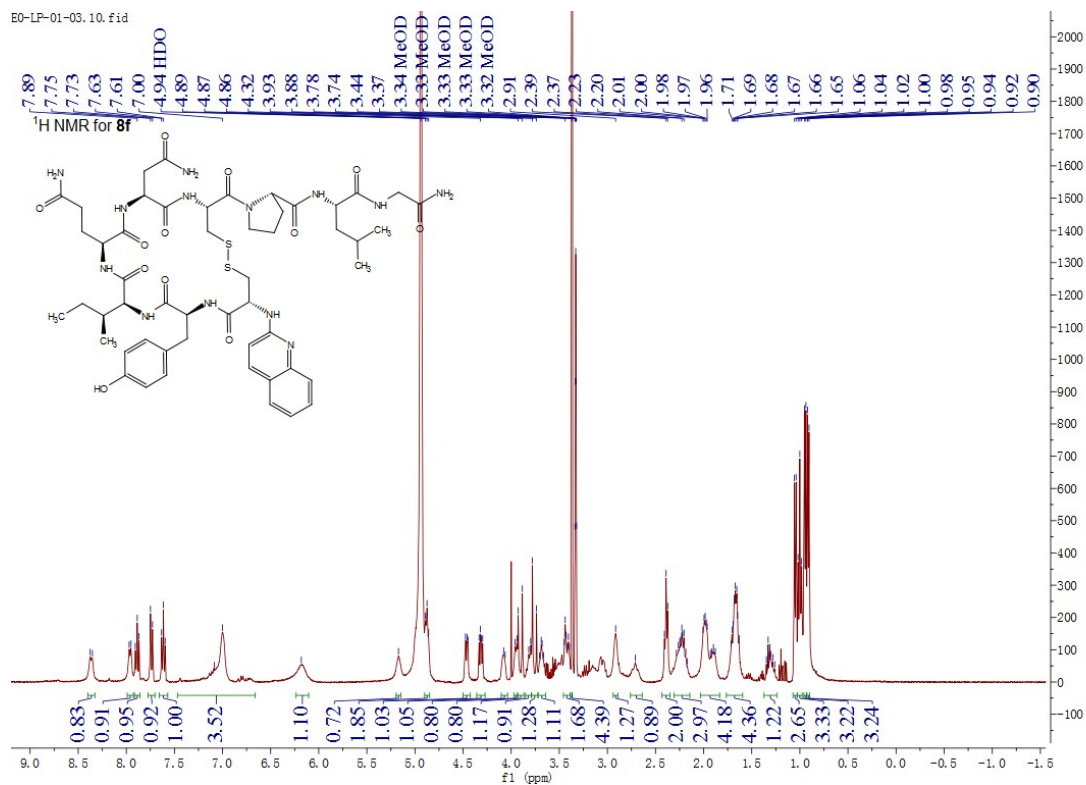
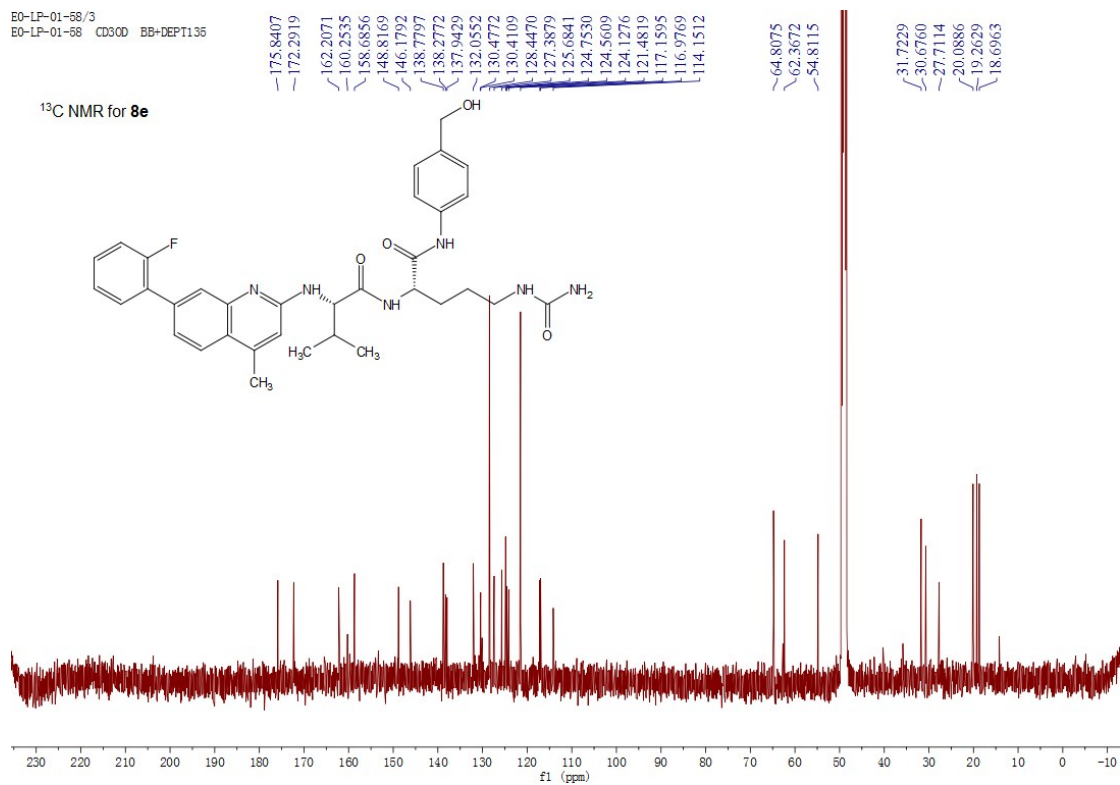


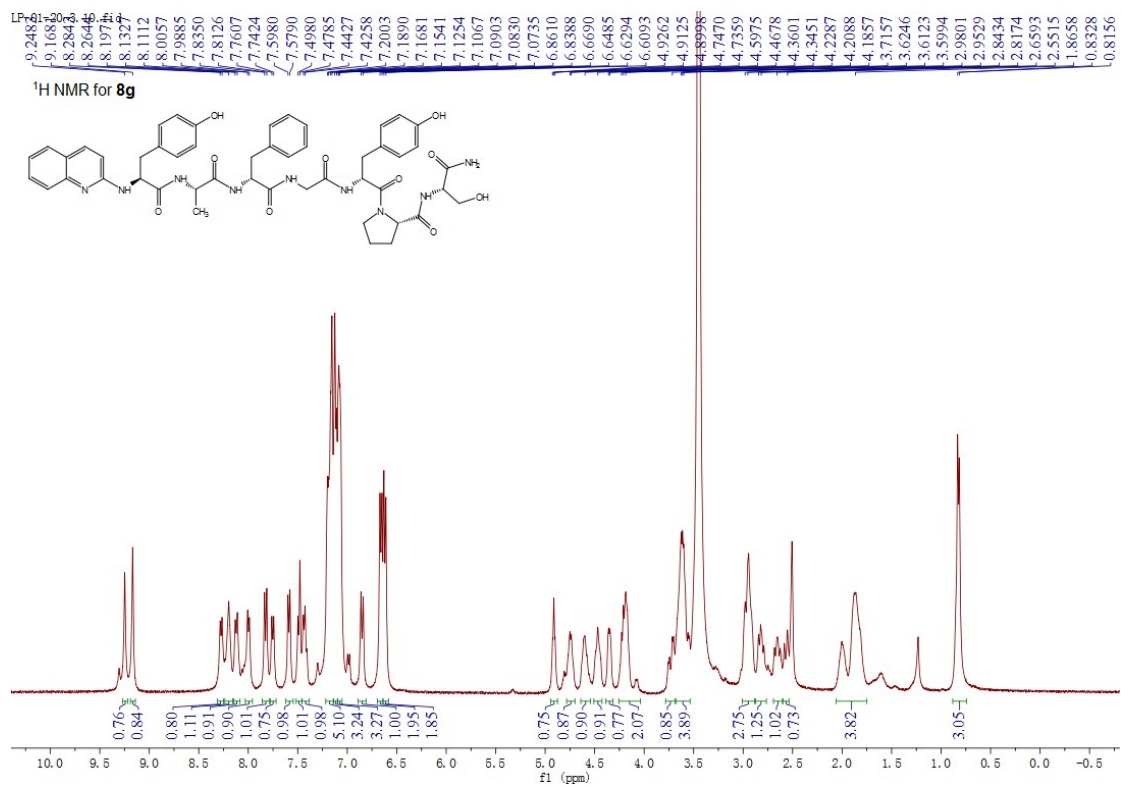
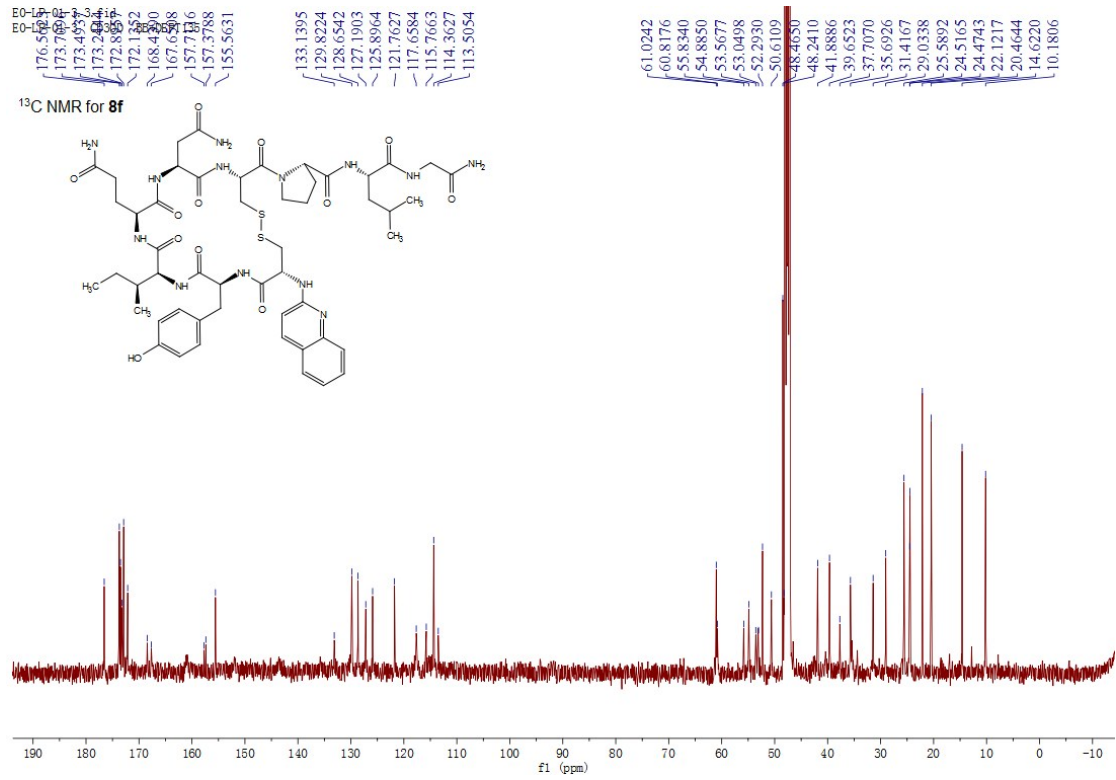
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EO-IP-01-25-D.DMSO-d6

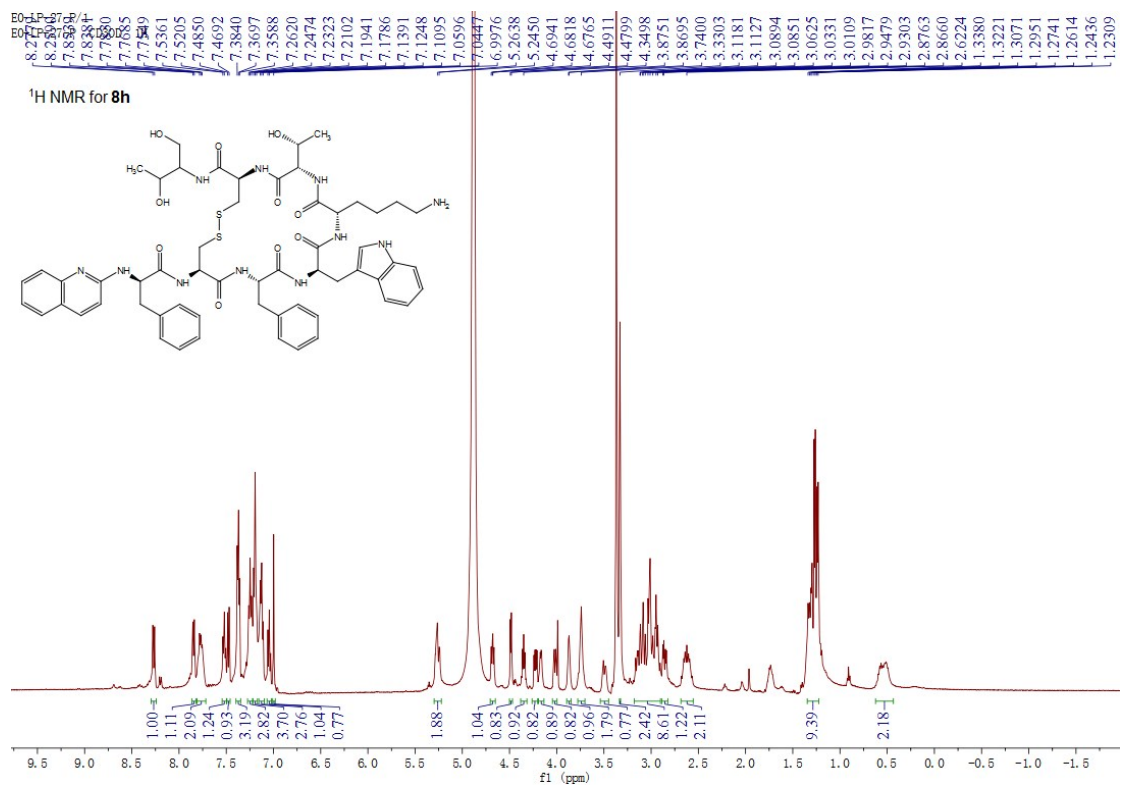
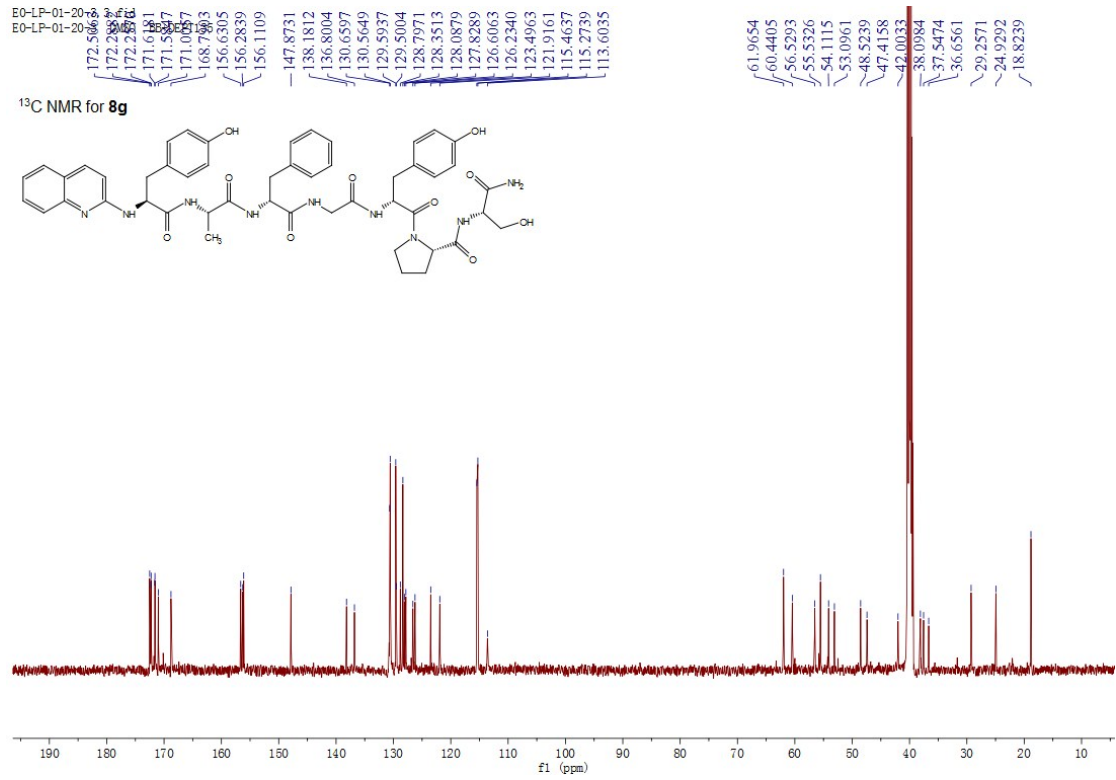


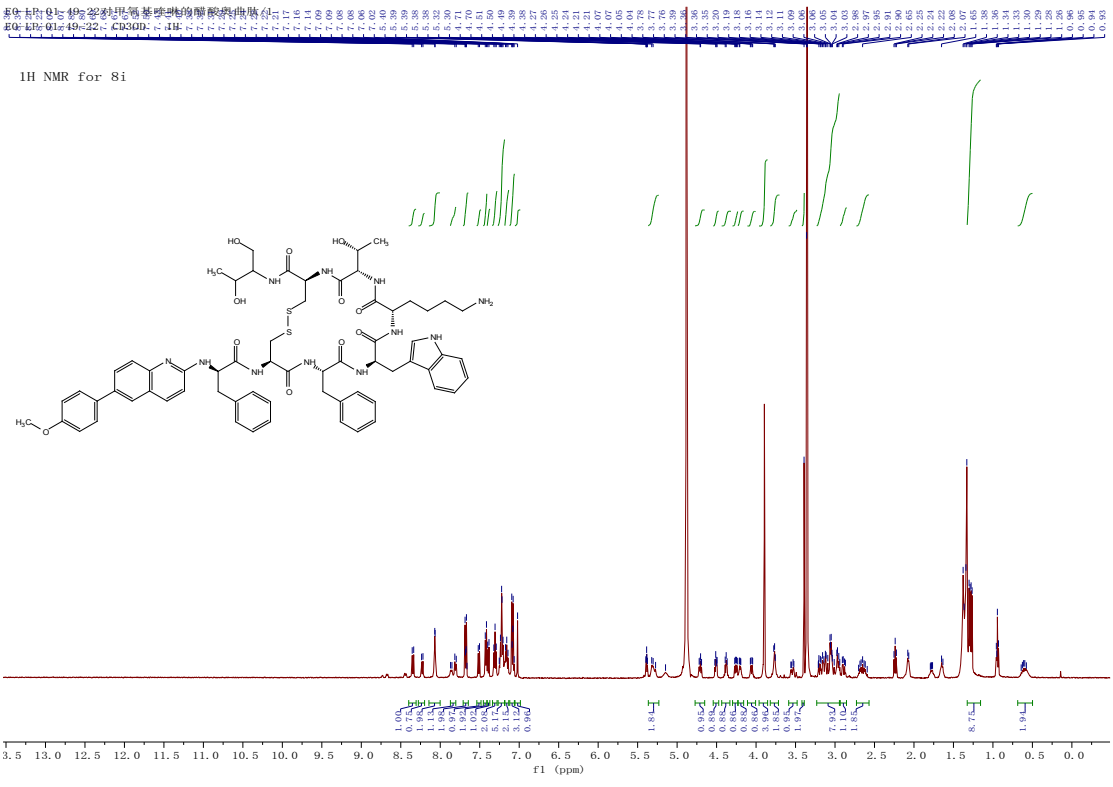
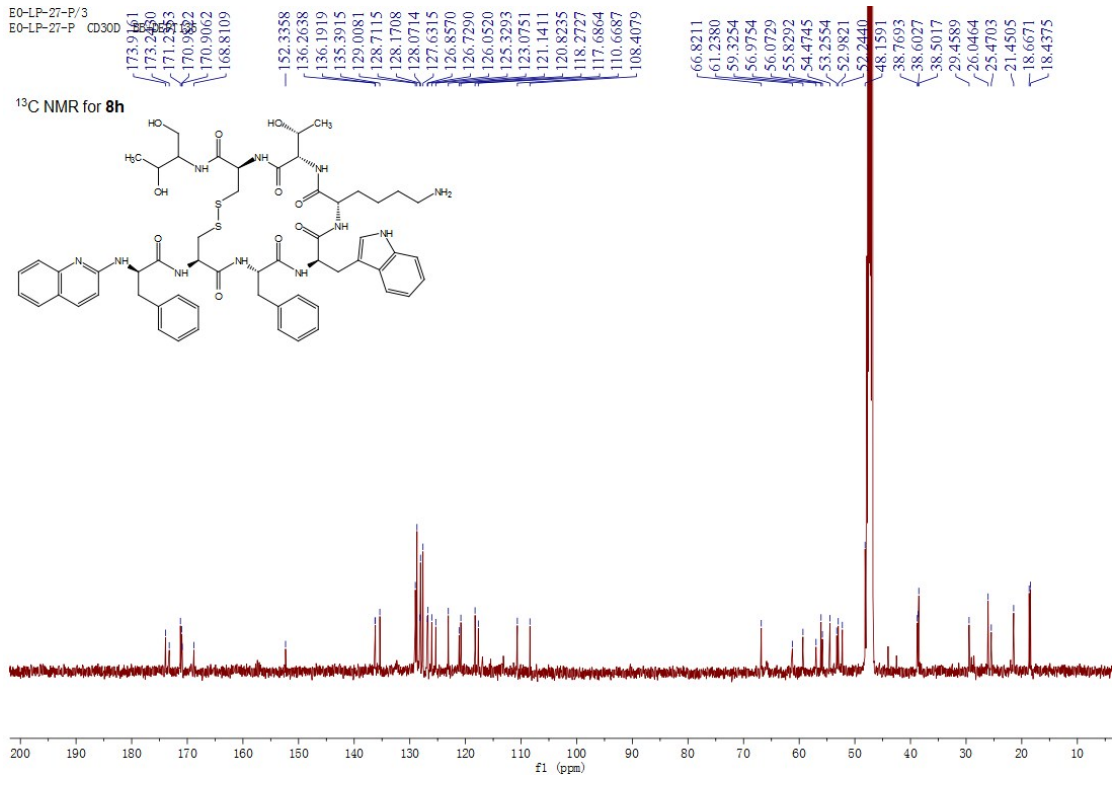


E0-LP-01-58/3
E0-LP-01-58 CD3OD BB-DEPT135









EO-LP-01-49-22/3
EO-LP-01-49-22
¹³C NMR for 8i

