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Supplementary

Solvent tuned discriminant sensing of Al³⁺, Mg²⁺ and HF₂⁻ by vanilinylpicolinyl hydrazide Schiff base

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Experimental Section

Materials and methods

All the required inorganic salts and organic chemicals of AR quality were obtained from Sigma-Aldrich and Merck, and used without further purification. Aqueous solutions of the required salts were prepared using Milli-Q water (Millipore). Perkin-Elmer 2400 Series-II CHN analyzer, Perkin Elmer, USA elemental analyser was used for elemental analyses. UVvis spectra were taken from Perkin Elmer Lambda 25 spectrophotometer and fluorescence spectra were obtained by using a Perkin Elmer spectrofluorimeter model LS55; Fluorescence lifetime measurements were carried out by using time-correlated single photon counting set up from Horiba Jobin-Yvon; FT-IR spectra (KBr disk, 4000–400 cm⁻¹) were collected from a Perkin Elmer LX-1 FTIR spectrophotometer. ¹H NMR spectra were obtained on a Bruker (AC) 300 MHz FT-NMR spectrometer and TMS is used as an internal standard. ESI mass spectra were recorded from a Water HRMS model XEVO-G2QTOF#YCA351 spectrometer. All of the required experimental measurements were conducted at room temperature.

Synthesis of HVPh

Picolino hydrazide was prepared from Picolinic acid. With continuous stirring to the MeOH (7 ml) solution of Picolino hydrazide (137.13 mg, 1.0 mmol), 2-hydroxy-4-methoxy-benzaldehyde (152.14 mg, 1.0 mmol) in methanol solution (7 ml) was added drop-by-drop

and stirring was continued for 8 h. Then the resultant clear solution was kept and allowed for slow evaporation, crystalline product was obtained after two weeks in 94% yield. Pyridine-2carboxylic acid (2-hydroxy-4-methoxy-benzylidene)-hydrazide (**HVPh**) : M.P.>200 °C. The mass spectrum displays an ion peak at 272.1420 for [**HVPh**+H]⁺ and a base peak at 294.1283 which corresponds the formation of [**HVPh** +Na]⁺ (**Fig. S1**); Microanalytical data: $C_{14}H_{13}N_3O_3$ calcd (found): C, 61.99 (61.81); H, 4.83 (4.90); N, 15.49 (14.58); %. ¹H NMR (300 MHz, DMSO-d₆): δ 12.39 (s, 1H, NH), 11.74 (s, 1H, OH), 8.74 (s, 1H, imine-H), 8.72 (d, 1H, Pyridine), 8.14-8.03 (m, 2H, pyridine), 7.69-7.65 (m, 1H, pyridine), 7.34 (d, 1H, vaniline), 6.54 (d. 1H, vaniline), 6.50 (t, 1H, vaniline), 3.78 (s, 1H, OCH₃) (**Fig. S2**); IR: 3461 cm⁻¹ (phenolic-OH), 3467 cm⁻¹ (hydrazide -NH), 1664 cm⁻¹ (imines C=N). (**Fig. S3**).

UV-Vis and fluorescence spectral studies

The stock solution of **HVPh** ($1 \times 10^{-3} \text{ mol } L^{-1}$) was prepared in DMSO. For the fluorescence and UV-Visible spectral study, the solution of **HVPh** was diluted to 20×10^{-6} mol L^{-1} with DMSO and Water. For UV-vis and fluorescence experiments, the test samples were prepared by placing appropriate amounts of the stock solution of the respective cations into 2.0 mL of the probe solution. For fluorescence measurements, the excitation was provided at 340 nm, and emission was acquired from 370 nm to 660 nm (excitation slit = 15.0 and emission slit = 5.0). Spectral data were taken within 10 seconds after addition of the ions.



Fig S1 Mass spectrum of HVPh



Fig S2 ¹H NMR of HVPh in DMSO-d₆



Fig S3 IR spectrum of HVPh in KBr disk.

Table S1 Single Crystal X-ray diffraction data of HVPh

Empirical formula	C ₁₄ H ₁₃ N ₃ O ₃
Formula weight	271.27
Temperature (K)	296(2)
System	triclinic
Space group	P -1
a (Å)	13.956(9)
b (Å)	15.246(7)

15.246(7)
77.91
62.76
62.76
2564(2)
8
1.406
0.102
1.502-24.998
1136.0
730
7516
0.1076
0.3265
0.915

 ${}^{a}R_{1} = \Sigma ||F_{o}| - |F_{c}|| / \Sigma |F_{o}|; \ {}^{b}wR_{2} = \{\Sigma [w(F_{o}^{2} - F_{c}^{2})^{2}] / \Sigma [w(F_{o}^{2})^{2}] \}^{1/2}; \ w = [\sigma^{2}(F_{o})^{2} + (0.1003P)^{2} + (0.1003P)^{2}$

4.9693P]⁻¹ (F_o² + 2F_c²)/3; ^cGoodness-of-fit



Fig S4 Inter-molecular H-bonding in HVPh



Fig S5 Inter-molecular π ... π stacking in HVPh



Fig S6 UV-vis spectrum of HVPh in Water medium.



Fig S7 UV-vis spectra of HVPh upon addition of various metal ions in water medium.







Fig S8 Fluorescence titration determine binding constant using Benisi-Hildebrand method for Al³⁺ [(a) Linear fitting; (b) Non-linear fitting]





(a)



Fig S9 Fluorescence titration determines binding constant using Benisi-Hildebrand method for Mg²⁺ [(a) Linear fitting; (b) Non-linear fitting]

(b)



Fig S10 Calculation of limit of detection (LOD) for Al³⁺



Fig S11 Calculation of limit of detection (LOD) for Mg^{2+}



Fig S12 Studies of interference by various metal ions on Al³⁺ sensitivity



Fig S13 Studies of interference by various metal ions on Mg²⁺ sensitivity



Fig S14 pH variation sensitivity of HVPh towards Al³⁺



Fig S15 pH variation sensitivity of HVPh towards Mg^{2+}



Fig S16 Job's plot for Al^{3+} by fluorescence spectroscopy



Fig S17 Job's plot for Al³⁺ by UV-vis spectroscopy



Fig S18 Mass spectra of HVPh-Al complex



Fig S19 Mass spectra of HVPh-Mg complex



Fig S20 IR spectrum of HVPh-Al complex in KBr disk



Fig S21 ¹H NMR spectra of HVPh in DMSO-d₆ on gradual addition of Mg²⁺ salt.



Fig S22 IR spectrum of HVPh-Mg in KBr disk



Fig S23 DFT optimised structure with total energy of probable Mg²⁺ complex of **HVPh** (A) charged (+2); (B) Neutral; (C) Mg²⁺ Complex as **HVPh**-Al.



Fig S24 Interaction of HVPh-Al³⁺ with various anions in Fluorescence spectroscopy



Fig S25 Interaction of HVPh-Mg²⁺ with various anions in Fluorescence spectroscopy. Anions are $S_2O_3^{2-}$ (Na₂S₂O₃, 5H₂O), SCN⁻(NH₄SCN), PO₄³⁻(K₃PO₄, H₂O), OAc⁻ (NaOAc, 3H₂O), ClO₄⁻(NaClO₄), SO₄²⁻(Na₂SO₄), HSO₄⁻(NaHSO₄), Cl⁻ (NaCl), F⁻(NaF), HF₂⁻ (NH₄HF₂), NO₃⁻(KNO₃), Br⁻ (KBr), NO₂⁻(NaNO₂), N₃⁻(NaN₃), AsO₄³⁻(Na₃AsO₄), AsO₂⁻ (NaAsO₂).



Fig. S26. Changes observed on incremental addition of HF_2^- (0.003 ml × 10⁻³ M each) to in situ formed 1:1 **HVPh-**Al³⁺ in H₂O (pH 7.2, HEPES buffer) in (14) UV-vis spectra.



Fig. S27. Changes observed on incremental addition of HF_2^- (0.003 ml × 10⁻³ M each) to in situ formed **HVPh**-Mg²⁺ in DMSO (pH 7.2, HEPES buffer) in UV-vis spectra



Fig S28 Calculation of limit of detection (LOD) for HF_2^-



Fig S29 Calculation of limit of detection (LOD) for HF_2 -



Fig 30 DFT/B3LYP optimised structure of HVPh, HVPh-Al and HVPh-Mg

MOs	Energy (eV)	vanilin	picoyl hydrazide
LUMO+5	0.95	45	55
LUMO+4	0.9	7	93
LUMO+3	-0.04	99	1
LUMO+2	-1.1	34	66
LUMO+1	-1.46	17	83
LUMO	-2.16	17	83
НОМО	-5.81	68	32
HOMO-1	-6.33	98	2

Table S2	Contributions	in	MOs	of HVPh

НОМО-2	-7.23	1	99
НОМО-3	-7.44	56	44
HOMO-4	-7.74	26	74
НОМО-5	-7.92	3	97





Fig S31 Frontier molecular orbitals images of HVPh

MOs	Energy	vanilin	picoyl hydrazide	Al	ОН
LUMO+5	-2.89	12	65	22	1
LUMO+4	-2.99	4	22	73	1
LUMO+3	-3.43	98	0	2	0
LUMO+2	-4.37	0	100	0	0
LUMO+1	-5.4	42	58	1	0
LUMO	-6.18	40	59	1	0

НОМО	-9.55	75	23	1	0
HOMO-1	-9.92	97	1	1	1
НОМО-2	-10.97	3	97	0	0
НОМО-3	-11.0	17	80	0	3
HOMO-4	-11.33	7	10	5	79
НОМО-5	-11.42	19	55	1	25

HOMO-5	HOMO-4	НОМО-3
НОМО-2	HOMO-1	НОМО

LUMO	LUMO+1	LUMO+2
LUMO+3	LUMO+4	LUMO+5

Fig S32 Frontier molecular orbitals images of HVPh-Al

Table S4 Contributions in MOs of HVPh-Mg

MOs	Energy (eV)	picoyl	vanilin	Mg
		hydrazide		
LUMO+5	-5.75	98	2	0
LUMO+4	-5.85	3	97	0

LUMO+3	-6.68	73	27	0
LUMO+2	-6.75	80	20	0
LUMO+1	-7.64	52	47	1
LUMO	-7.81	47	52	1
НОМО	-11.29	25	75	0
HOMO-1	-11.44	26	74	0
НОМО-2	-12.33	74	26	0
НОМО-3	-12.34	77	23	0
HOMO-4	-12.34	70	30	0
HOMO-5	-12.4	14	86	0





Fig S33 Frontier molecular orbitals images of HVPh-Mg



Fig 34 Frontier molecular orbitals that represent band gap (HOMO-LUMO gap) of **HVPh**, **HVPh**-Al and (**HVPh**)₂-Mg.



Fig S35 Calibration plot for detection of Al³⁺ using fluorescence active probe HVPh.