

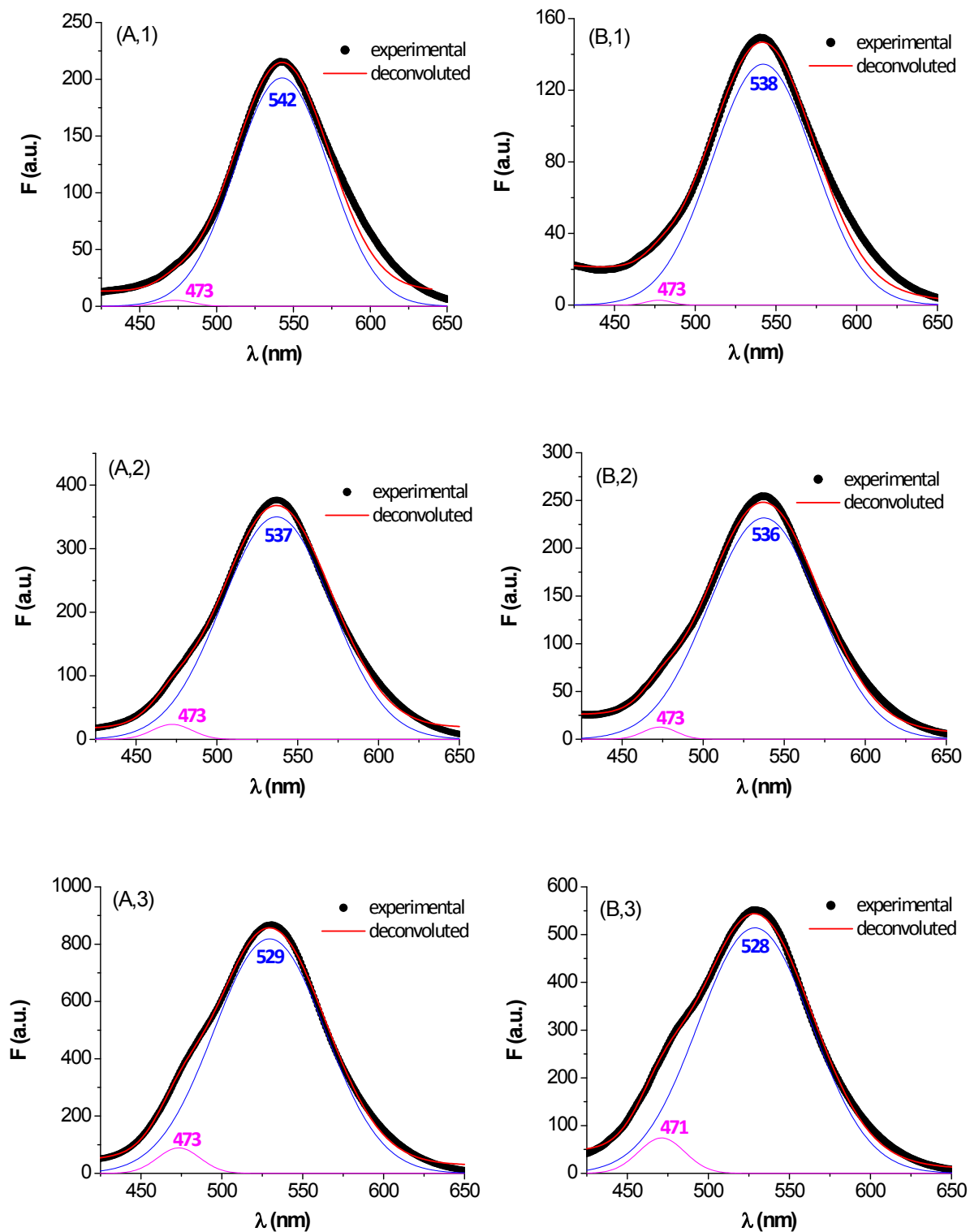
**New flexible molecular probes bearing dansyl and TEMPO moieties for host-guest interactions in solution and gels**

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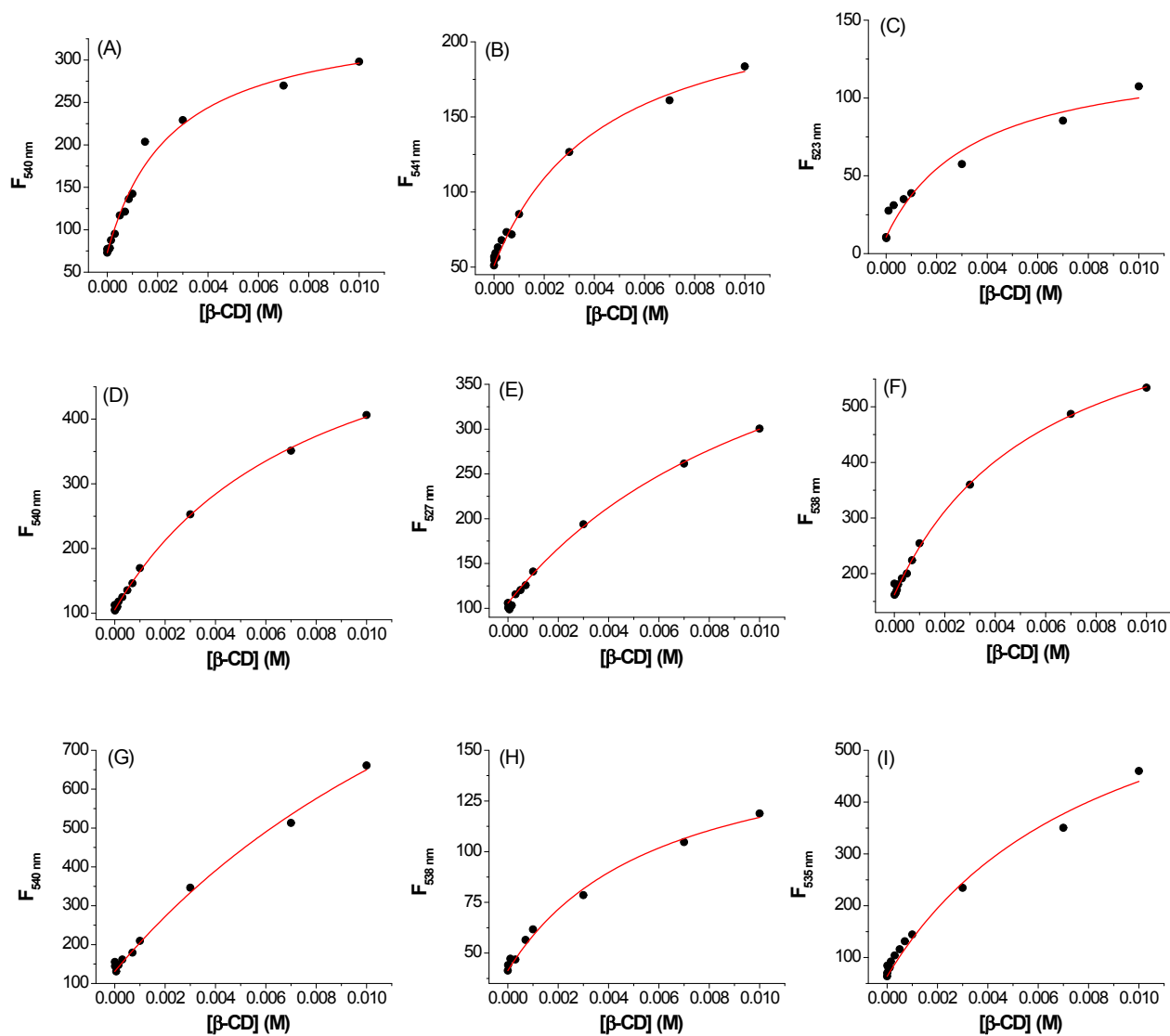
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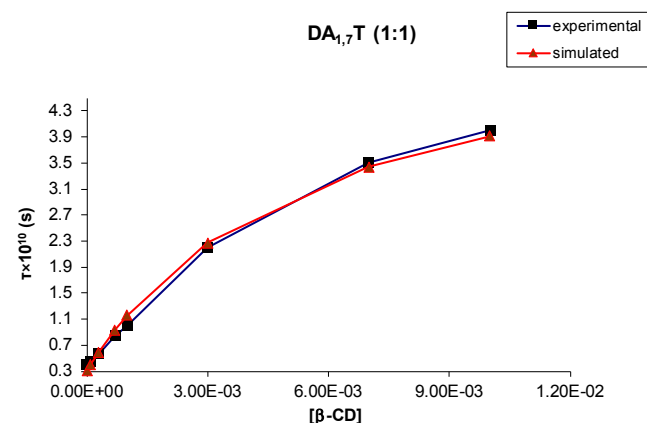
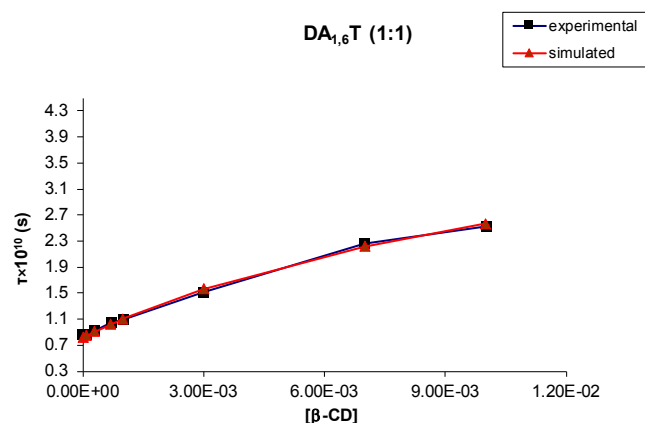
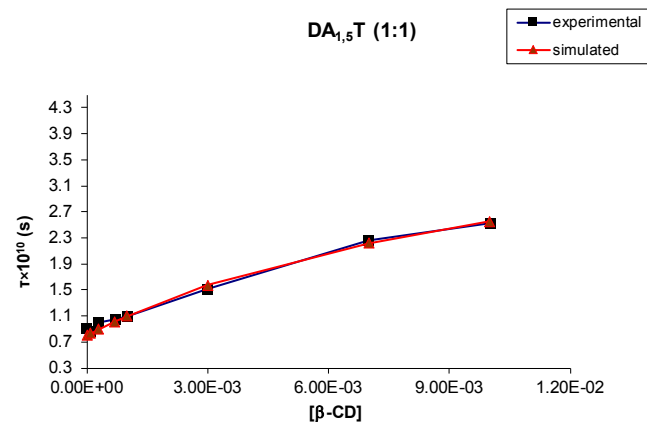
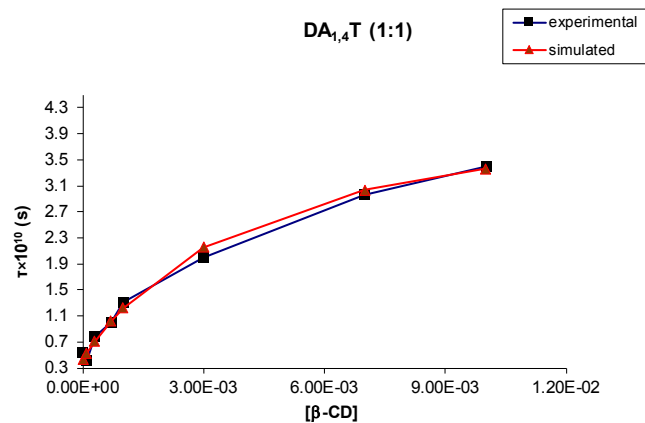
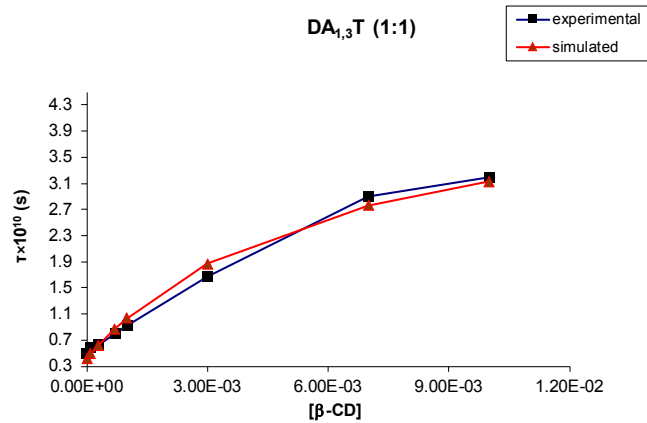
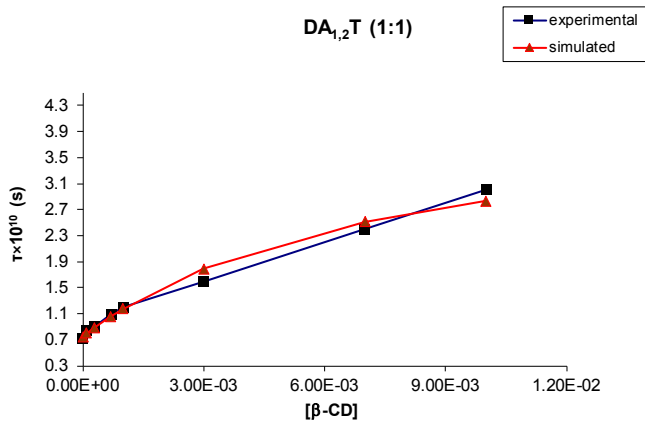
**Fig. S1.** Deconvolution of the fluorescence spectra of (A) DA<sub>1.7</sub> and (B) DA<sub>1.7</sub>T in water in the absence (1) and in the presence of  $\beta$ -CD at  $10^{-3}$  M (2) and  $10^{-2}$  M (3) concentrations.

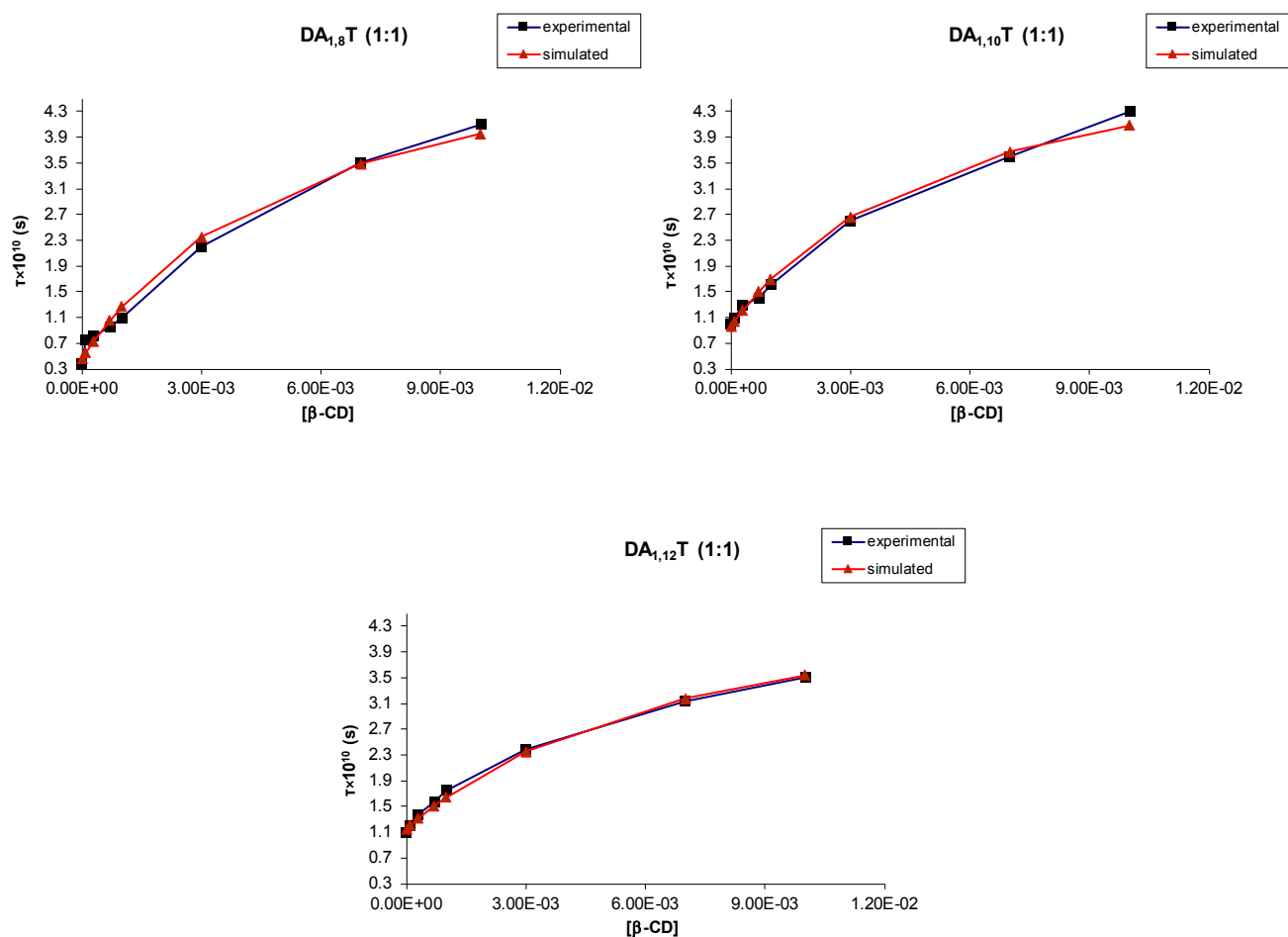
**Table S1.** Position ( $\lambda$ , in nm) and relative area (A, in %) of the two fluorescence bands obtained by deconvolution of the emission spectra of DA<sub>1,n</sub> and DA<sub>1,n</sub>T in the absence and in the presence of  $\beta$ -CD

[ $\beta$ -CD] (M)	DA <sub>1,n</sub>				DA <sub>1,n</sub> T			
	$\lambda_1$	A <sub>1</sub>	$\lambda_2$	A <sub>2</sub>	$\lambda_1$	A <sub>1</sub>	$\lambda_2$	A <sub>2</sub>
0	473	0.9	542	99.1	473	0.3	538	99.7
10 <sup>-3</sup>	473	2.4	537	97.6	473	1.7	536	98.3
10 <sup>-2</sup>	473	4.2	529	95.8	471	5.6	528	94.4



**Fig. S2.** Dependence of the normalized fluorescence emission of  $DA_{1,n}T$  probes on the cyclodextrrin concentration. The solid lines represent the best fits according to eq. (3) for (A)  $DA_{1.2}T$ , (B)  $DA_{1.3}T$ , (C)  $DA_{1.4}T$ , (D)  $DA_{1.5}T$ , (E)  $DA_{1.6}T$ , (F)  $DA_{1.7}T$ , (G)  $DA_{1.8}T$ , (H)  $DA_{1.10}T$ , (I)  $DA_{1.12}T$ .



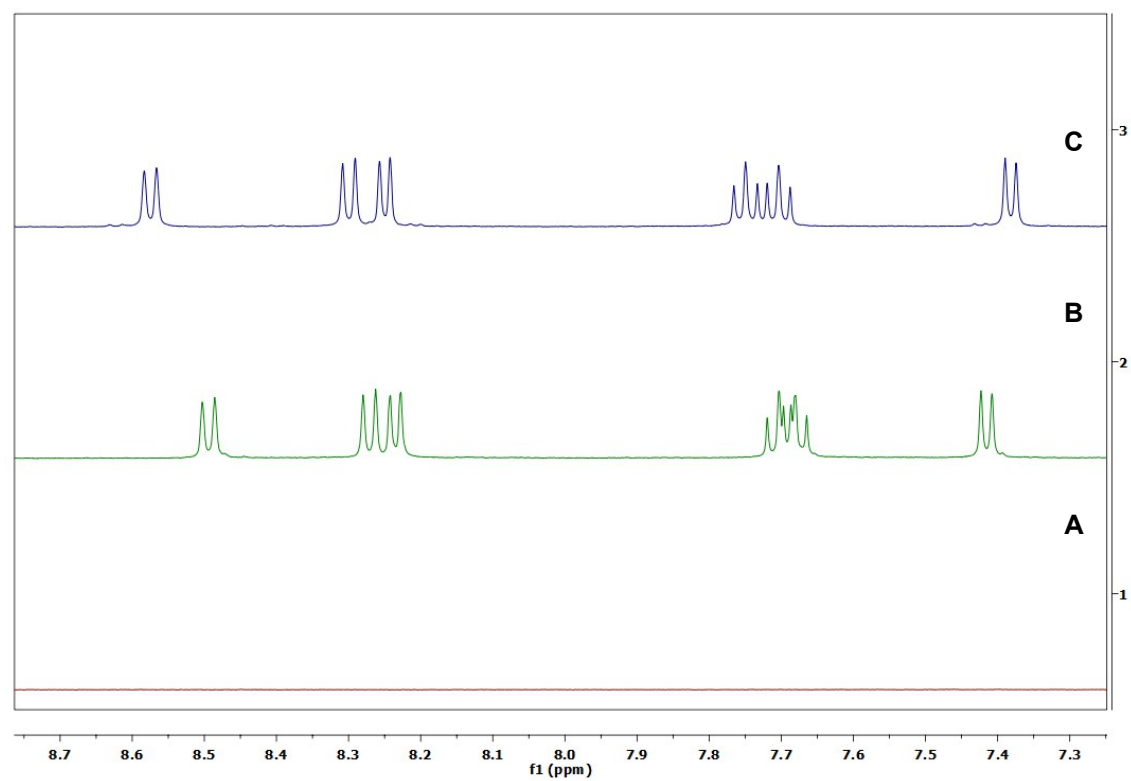
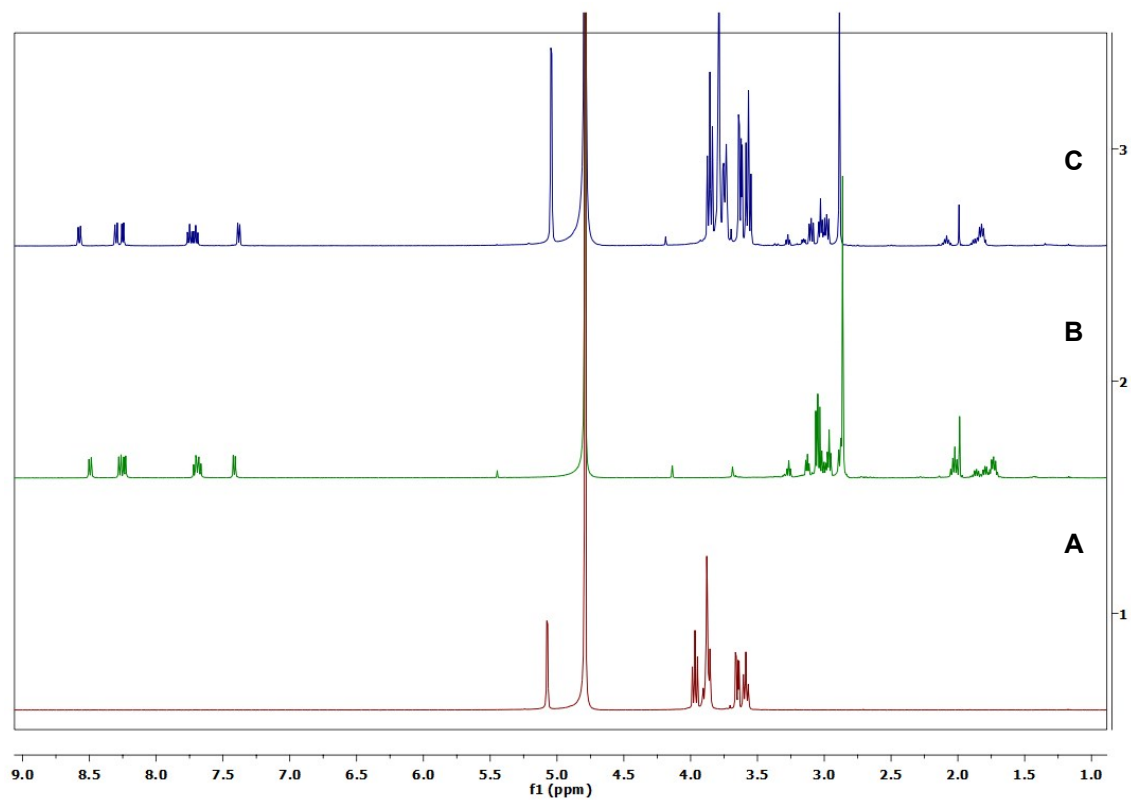


**Fig. S3.** Dependence of the rotational correlation time of DA<sub>1,n</sub>T probes on the cyclodextrin concentration.

**NMR spectra of DA<sub>1.3</sub> and DA<sub>1.3</sub>T (reduced with hydrazine)  
in the absence and in the presence of β-cyclodextrin**

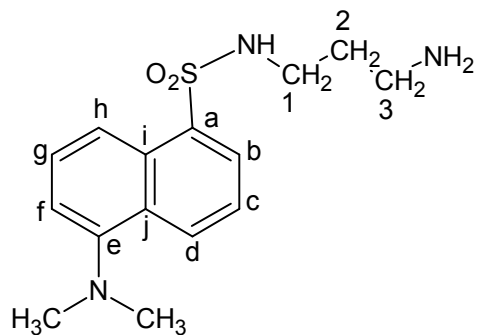
The NMR spectra of DA<sub>1.3</sub> and DA<sub>1.3</sub>T (reduced with hydrazine) have been recorded in the absence and in the presence of β-CD in deuterated water and in DMSO. This allowed us to evidence the differences in chemical shifts of the proton signals assigned to the dansyl group. Unfortunately, the corresponding dual probe, DA<sub>1.3</sub>T, is not soluble in deuterated water at concentrations that would allow us to record NMR spectra. While fluorescence and EPR measurements can be performed at concentrations of  $\sim 10^{-6}$ – $10^{-4}$  M, NMR measurements require higher concentrations of the probes ( $\sim 10^{-2}$  M). Moreover, while the complex of DA<sub>1.3</sub> with β-CD is soluble in deuterated water, the solid complex of DA<sub>1.3</sub>T separated from solution. Therefore, in this case both β-CD and DA<sub>1.3</sub>T were in too low concentration in solution to record the NMR spectra.

For this reason, we recorded the NMR spectra in another polar solvent that solubilizes both DA<sub>1.3</sub>T and the mixture DA<sub>1.3</sub>T/β-CD. As can be noticed from Figs. S4, S5 and Tables S1, S2, chemical shifts in the NMR spectrum of the DA<sub>1.3</sub>T/β-CD mixture are observed for protons of the dansyl group but not of the TEMPO moiety. This sustains our conclusion, based on fluorescence and EPR data, that the dansyl moiety is involved in the inclusion process.

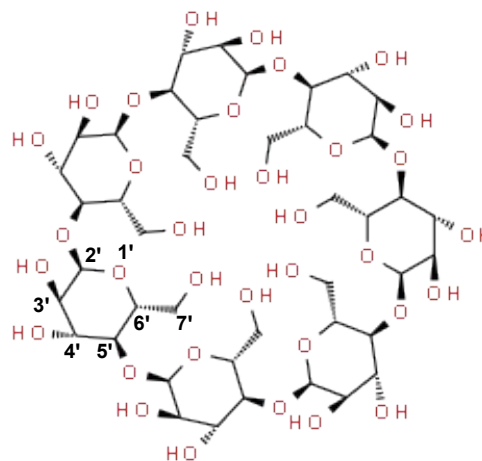


**Fig. S4.** <sup>1</sup>H-NMR spectra in D<sub>2</sub>O of: **A** – β-cyclodextrin, **B** – DA<sub>1,3</sub>; **C** - β-cyclodextrin + DA<sub>1,3</sub>.





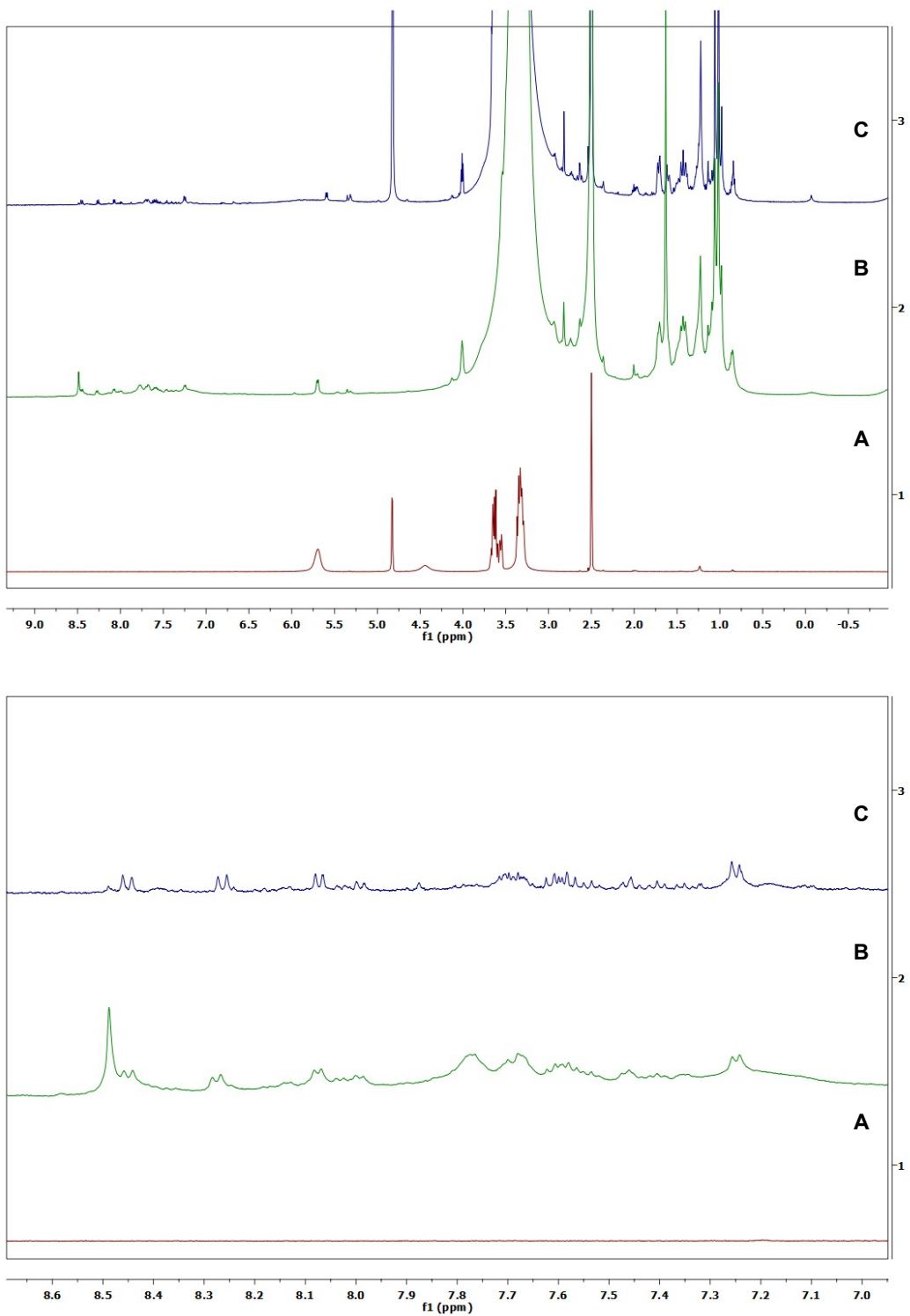
DA<sub>1.3</sub> structure



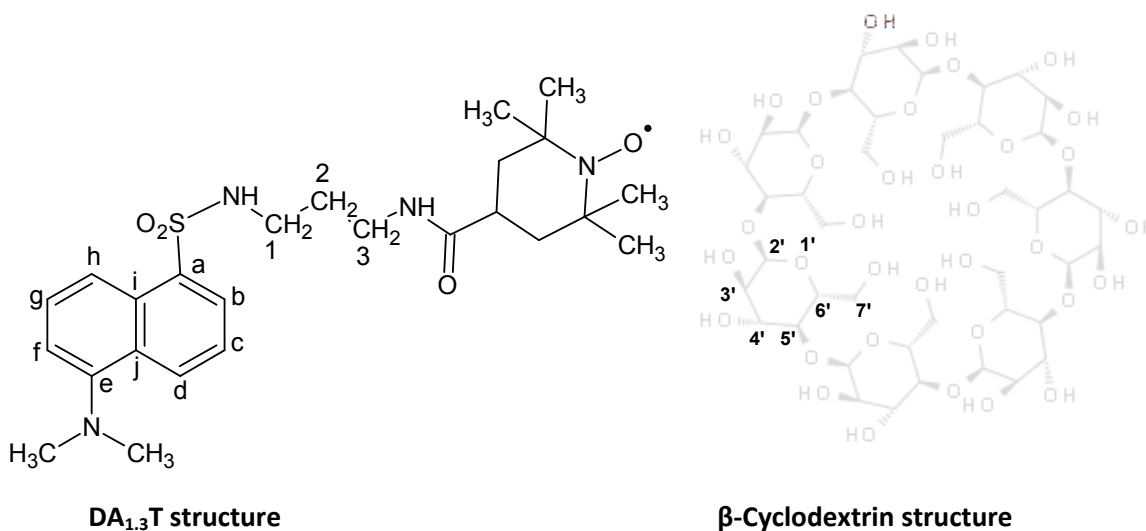
$\beta$ -Cyclodextrin structure

**Table S1.** Chemical shifts of DA<sub>1.3</sub> and  $\beta$ -CD protons in D<sub>2</sub>O

Compounds protons	Chemical shifts of $\beta$ -CD (ppm)	Chemical shifts of DA <sub>1.3</sub> (ppm)	Chemical shifts of $\beta$ -CD + DA <sub>1.3</sub> (ppm)	$\Delta\delta = \left  \delta_{BCD \text{ or } DA_{1.3}} - \delta_{BCD + DA_{1.3}} \right $	
				ppm	Hz
H-b	-	8.23	8.25	0.02	11.1
H-c	-	7.68	7.70	0.02	11.2
H-d	-	8.49	8.57	0.08	40.3
H-f	-	7.41	7.38	0.03	17.2
H-g	-	7.70	7.74	0.05	22.8
H-h	-	8.27	8.30	0.03	14.1
H-1	-	2.96	3.02	0.06	32.0
H-2	-	1.73	1.82	0.09	45.70
H-3	-	2.88	2.98	0.11	52.3
-N(CH <sub>3</sub> ) <sub>2</sub>	-	2.86	2.88	0.02	11.8
H-2 $\square$	5.07	-	5.04	0.03	15.2
H-3 $\square$	3.58	-	3.56	0.02	10.1
H-4 $\square$	3.97	-	3.85	0.12	55.8
H-5 $\square$	3.90-3.85	-	3.79-3.72	0.11-0.13	57.5-64.8
H-6 $\square$	3.65	-	3.63	0.02	12.0
H-7 $\square$	3.90-3.85	-	3.79-3.72	0.11-0.13	57.5-64.8



**Fig. S5.** <sup>1</sup>H-NMR spectra in DMSO of: **A** –  $\beta$ -cyclodextrin, **B** – DA<sub>1,3</sub>T; **C** -  $\beta$ -cyclodextrin + DA<sub>1,3</sub>T.

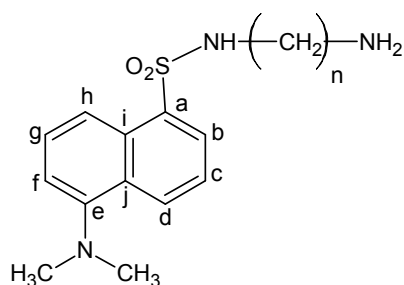


**Table S2.** Chemical shifts of DA<sub>1.3</sub>T and β-CD protons in DMSO

Compounds protons	Chemical shifts of β-CD (ppm)	Chemical shifts of DA <sub>1.3</sub> T (ppm)	Chemical shifts of β-CD + DA <sub>1.3</sub> T (ppm)	$\Delta\delta =  \delta_{BCD \text{ or } DA_{1.3}T} - \delta_{BCD + DA_{1.3}T} $	
				ppm	Hz
H-b	-	8.08	8.06	0.02	9.6
H-c	-	7.62-7.53	7.64-7.55	0.02	9.4
H-d	-	8.45	8.43	0.02	8.98
H-f	-	7.25	7.23	0.02	9.7
H-g	-	7.62-7.53	7.64-7.55	0.02	9.4
H-h	-	8.28	8.26	0.02	10.8
H-1	-	2.93	2.92	0.01	6.7
H-2	-	1.43	1.40	0.03	15.3
H-3	-	2.74	2.72	0.02	9.6
-N(CH <sub>3</sub> ) <sub>2</sub>	-	2.82	2.81	0.01	6.2
H-2 <sup>Ⓢ</sup>	4.83	-	4.82	0.01	4.5
H-3 <sup>Ⓢ</sup>	3.36-3.29	-	Overlapped with water signal from DMSO	-	-
H-4 <sup>Ⓢ</sup>	3.67-3.55	-	3.65-3.52	0.02-0.03	11.1-15.6
H-5 <sup>Ⓢ</sup>	3.67-3.55	-	3.65-3.52	0.02-0.03	11.1-15.6
H-6 <sup>Ⓢ</sup>	3.36-3.29	-	Overlapped with water signal from DMSO	-	-
H-7 <sup>Ⓢ</sup>	3.67-3.55	-	3.65-3.52	0.02-0.03	11.1-15.6
-CH (TEMPO)	-	2.63	2.63	0	0.0
-CH <sub>2</sub> - (TEMPO)	-	1.74	1.74	0	0.0
-CH <sub>3</sub> (TEMPO)	-	1.06 and 1.02	1.055-1.015	0.005	1.6

## Spectral characterization of DA<sub>1,n</sub>

### Notation of H atoms for assignment of NMR <sup>1</sup>H signals



n = 2,3,4,5,6,7,8,10,12

MAD-2, n=2, -NH-CH<sub>2</sub><sup>1</sup>-CH<sub>2</sub><sup>2</sup>-NH<sub>2</sub>

MAD-7, n=7, -NH-CH<sub>2</sub><sup>1</sup>-CH<sub>2</sub><sup>2</sup>-CH<sub>2</sub><sup>3</sup>-CH<sub>2</sub><sup>4</sup>-CH<sub>2</sub><sup>5</sup>-CH<sub>2</sub><sup>6</sup>-CH<sub>2</sub><sup>7</sup>-NH<sub>2</sub>

MAD-3, n=3, -NH-CH<sub>2</sub><sup>1</sup>-CH<sub>2</sub><sup>2</sup>-CH<sub>2</sub><sup>3</sup>-NH<sub>2</sub>

MAD-8, n=8, -NH-CH<sub>2</sub><sup>1</sup>-CH<sub>2</sub><sup>2</sup>-CH<sub>2</sub><sup>3</sup>-CH<sub>2</sub><sup>4</sup>-CH<sub>2</sub><sup>5</sup>-CH<sub>2</sub><sup>6</sup>-CH<sub>2</sub><sup>7</sup>-CH<sub>2</sub><sup>8</sup>-NH<sub>2</sub>

MAD-4, n=4, -NH-CH<sub>2</sub><sup>1</sup>-CH<sub>2</sub><sup>2</sup>-CH<sub>2</sub><sup>3</sup>-CH<sub>2</sub><sup>4</sup>-NH<sub>2</sub>

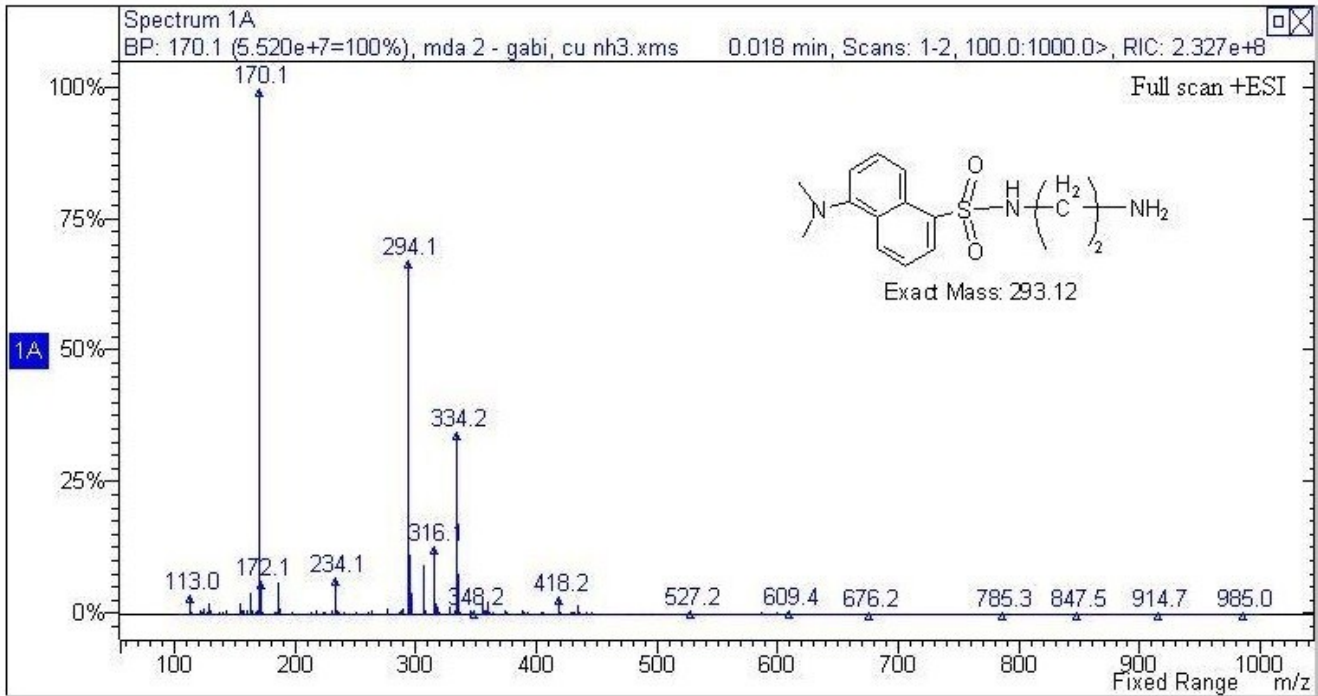
MAD-10, n=10, -NH-CH<sub>2</sub><sup>1</sup>-CH<sub>2</sub><sup>2</sup>-CH<sub>2</sub><sup>3</sup>-CH<sub>2</sub><sup>4</sup>-CH<sub>2</sub><sup>5</sup>-CH<sub>2</sub><sup>6</sup>-CH<sub>2</sub><sup>7</sup>-CH<sub>2</sub><sup>8</sup>-CH<sub>2</sub><sup>9</sup>-CH<sub>2</sub><sup>10</sup>-NH<sub>2</sub>

MAD-5, n=5, -NH-CH<sub>2</sub><sup>1</sup>-CH<sub>2</sub><sup>2</sup>-CH<sub>2</sub><sup>3</sup>-CH<sub>2</sub><sup>4</sup>-CH<sub>2</sub><sup>5</sup>-NH<sub>2</sub>

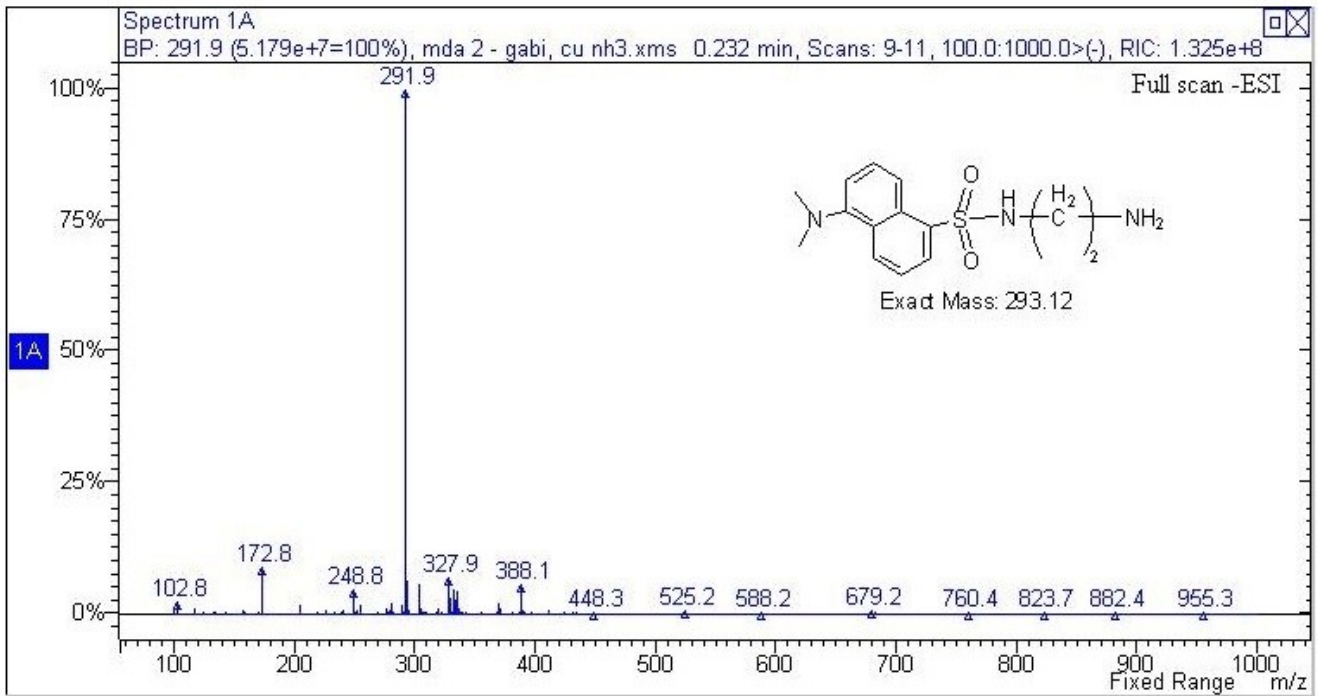
MAD-12, n=12, -NH-CH<sub>2</sub><sup>1</sup>-CH<sub>2</sub><sup>2</sup>-CH<sub>2</sub><sup>3</sup>-CH<sub>2</sub><sup>4</sup>-CH<sub>2</sub><sup>5</sup>-CH<sub>2</sub><sup>6</sup>-CH<sub>2</sub><sup>7</sup>-CH<sub>2</sub><sup>8</sup>-CH<sub>2</sub><sup>9</sup>-CH<sub>2</sub><sup>10</sup>-CH<sub>2</sub><sup>11</sup>-CH<sub>2</sub><sup>12</sup>-NH<sub>2</sub>

MAD-6, n=6, -NH-CH<sub>2</sub><sup>1</sup>-CH<sub>2</sub><sup>2</sup>-CH<sub>2</sub><sup>3</sup>-CH<sub>2</sub><sup>4</sup>-CH<sub>2</sub><sup>5</sup>-CH<sub>2</sub><sup>6</sup>-NH<sub>2</sub>

### DA<sub>1,2</sub> (dansyl ethylenediamine)

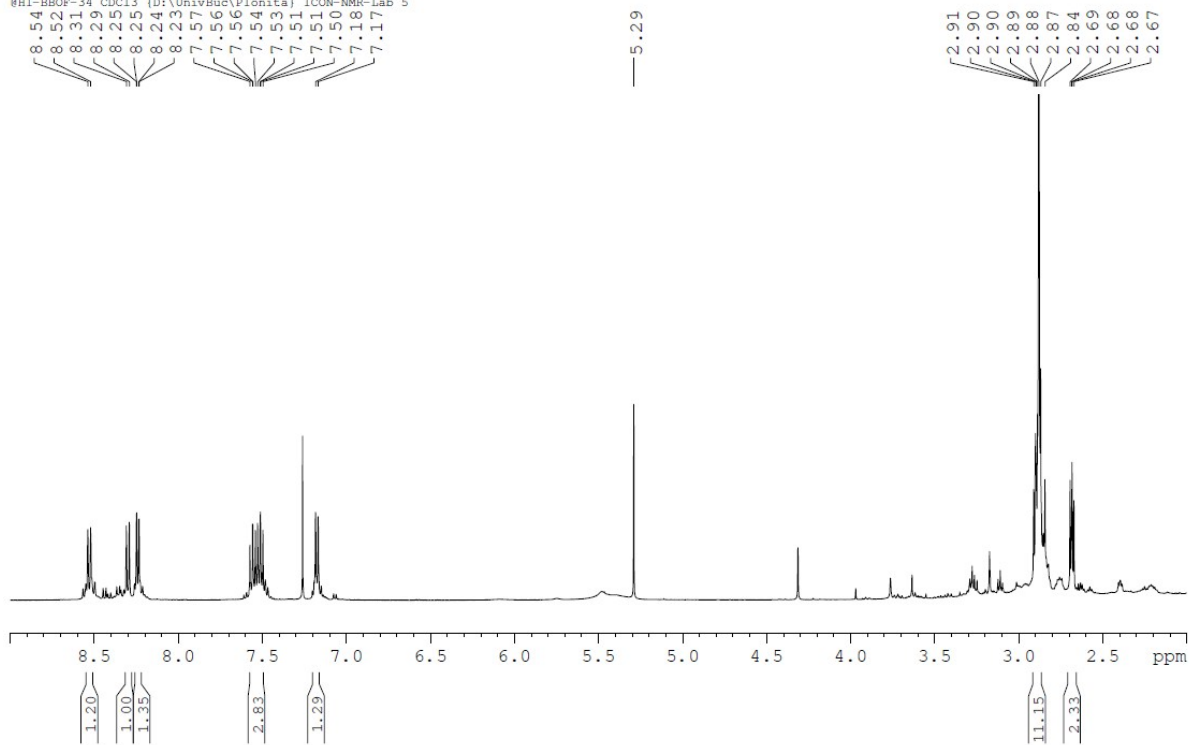


ESI-MS ( $m/z$ ): 294 (DA<sub>1,2</sub> + H<sup>+</sup>)



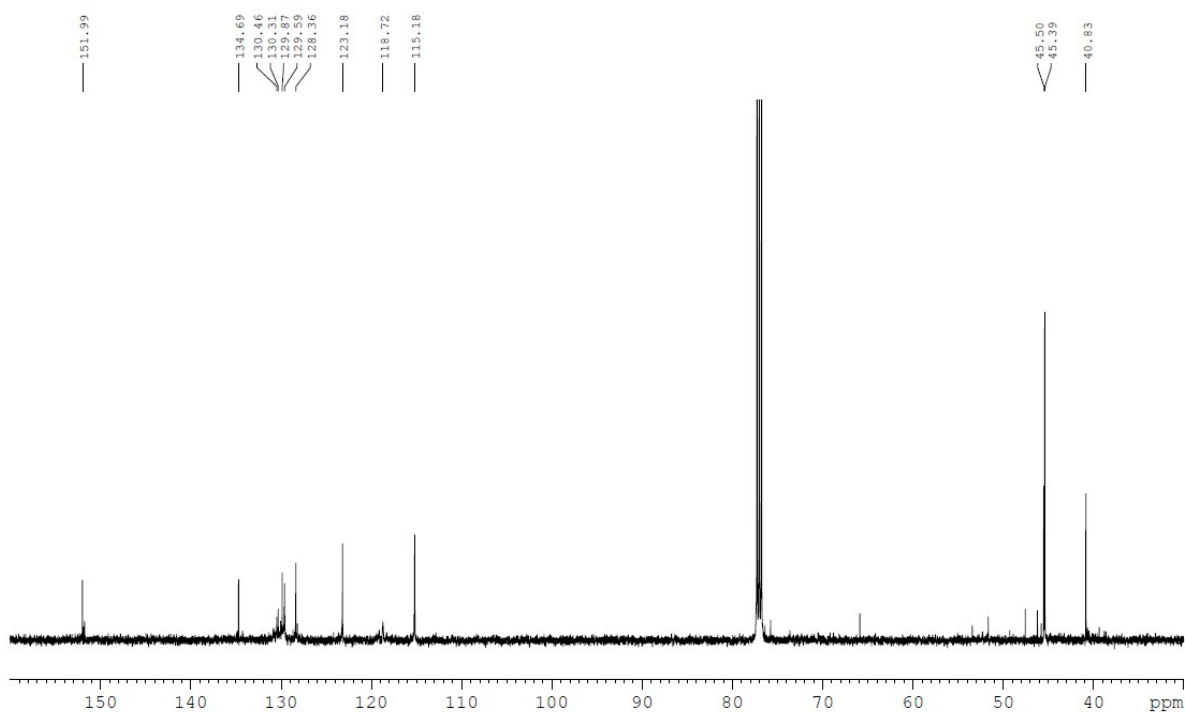
ESI-MS ( $m/z$ ): 292 (DA<sub>1,2</sub> - H<sup>+</sup>)

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Operator CS  
Registry No. 2185  
User G. Ionita  
Sample Changer Position 9  
Sample name: MAD-2  
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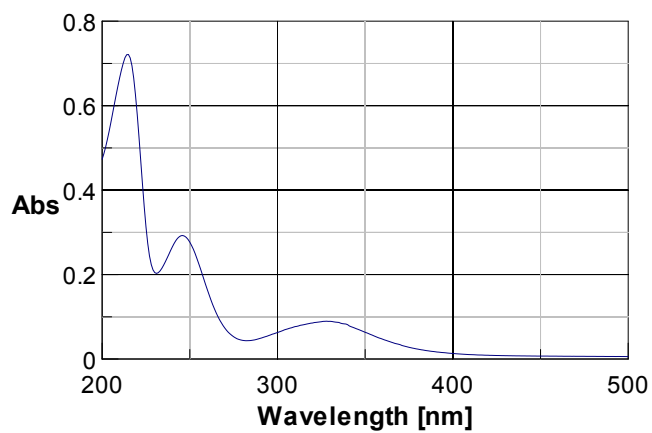


<sup>1</sup>H NMR spectrum of compound DA<sub>1,2</sub>

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
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User G. Ionita  
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Sample name: MAD-2  
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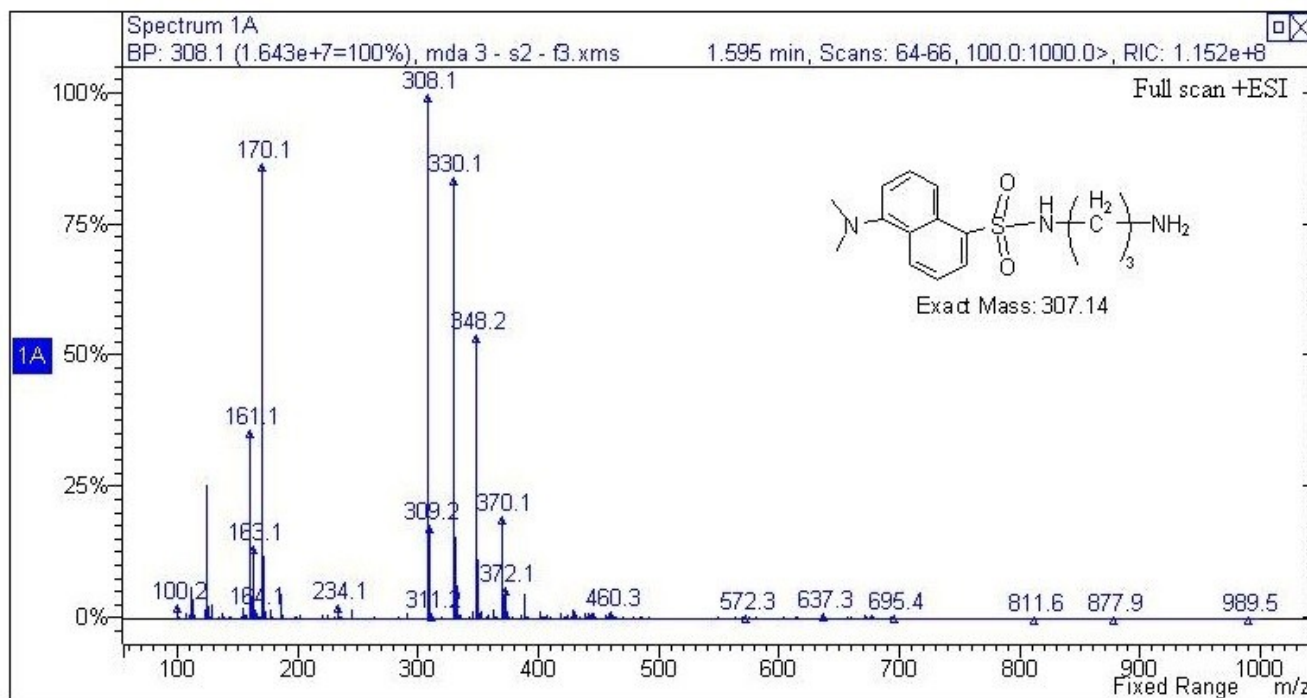


<sup>13</sup>C NMR spectrum of compound DA<sub>1.2</sub>

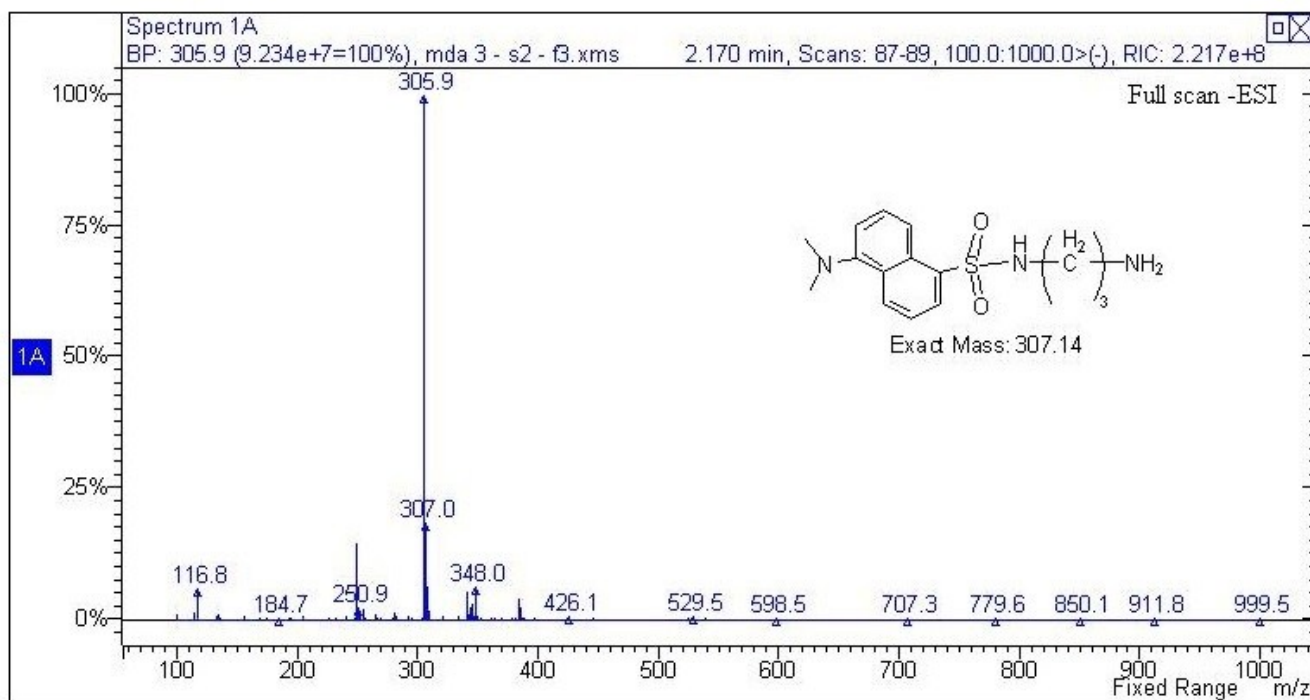


UV-Vis spectrum of compound DA<sub>1.2</sub>

### DA<sub>1.3</sub> (dansyl 1,3-diaminopropane)



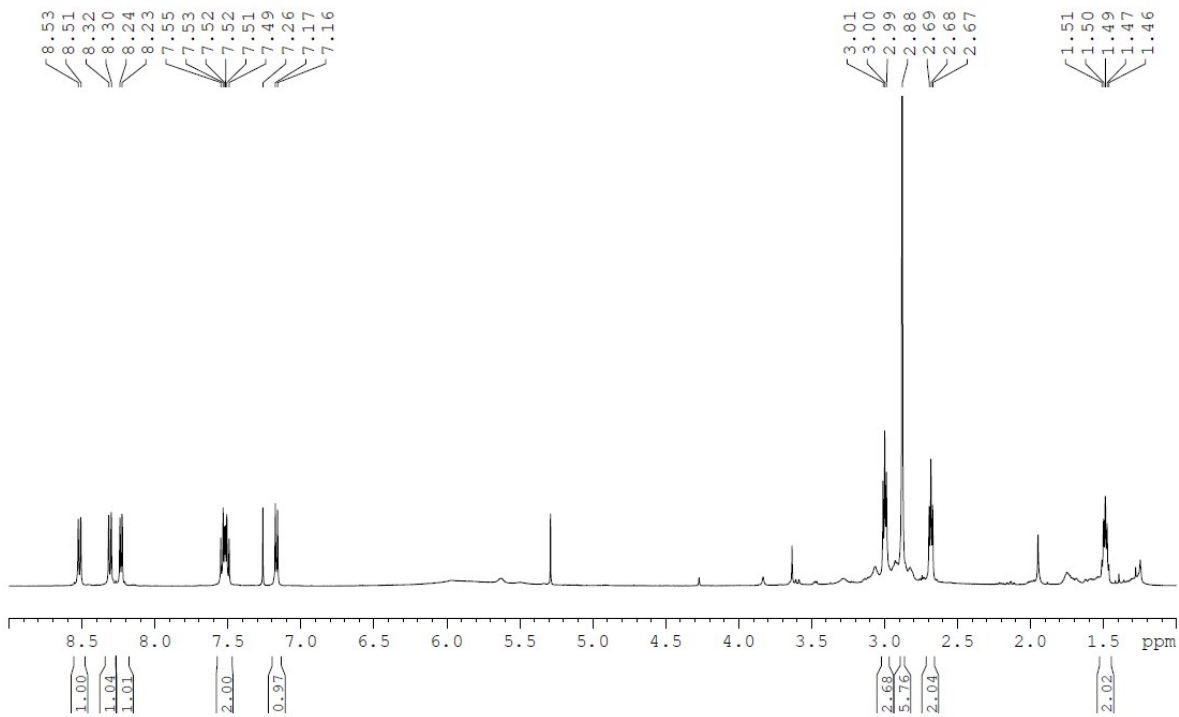
ESI-MS ( $m/z$ ): 308 (DA<sub>1.3</sub> + H<sup>+</sup>)



ESI-MS ( $m/z$ ): 306 (DA<sub>1.3</sub> - H<sup>+</sup>)

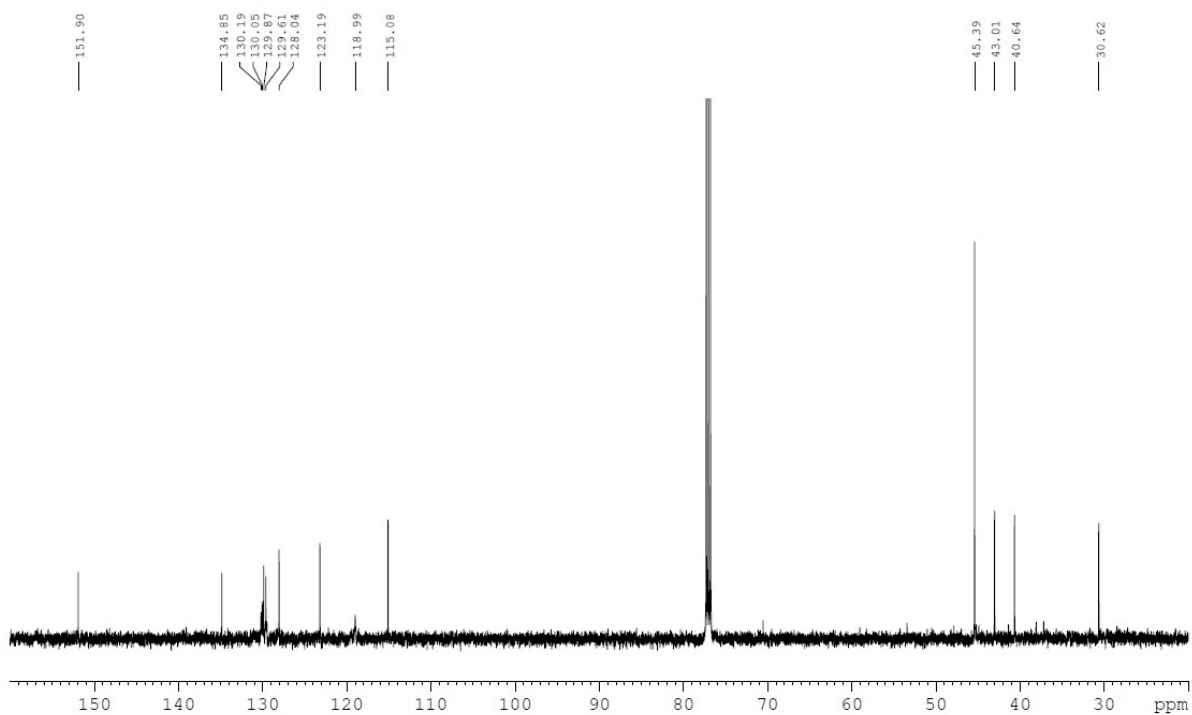


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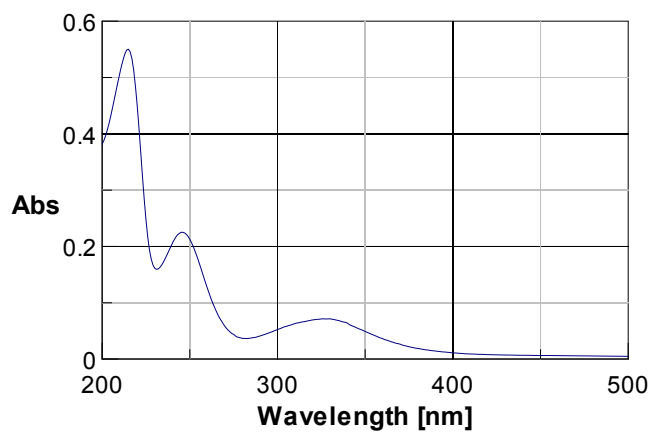


<sup>1</sup>H NMR spectrum of compound DA<sub>1.3</sub>

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
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User G. Ionita  
Sample Changer Position 4  
Sample name: MAD-3  
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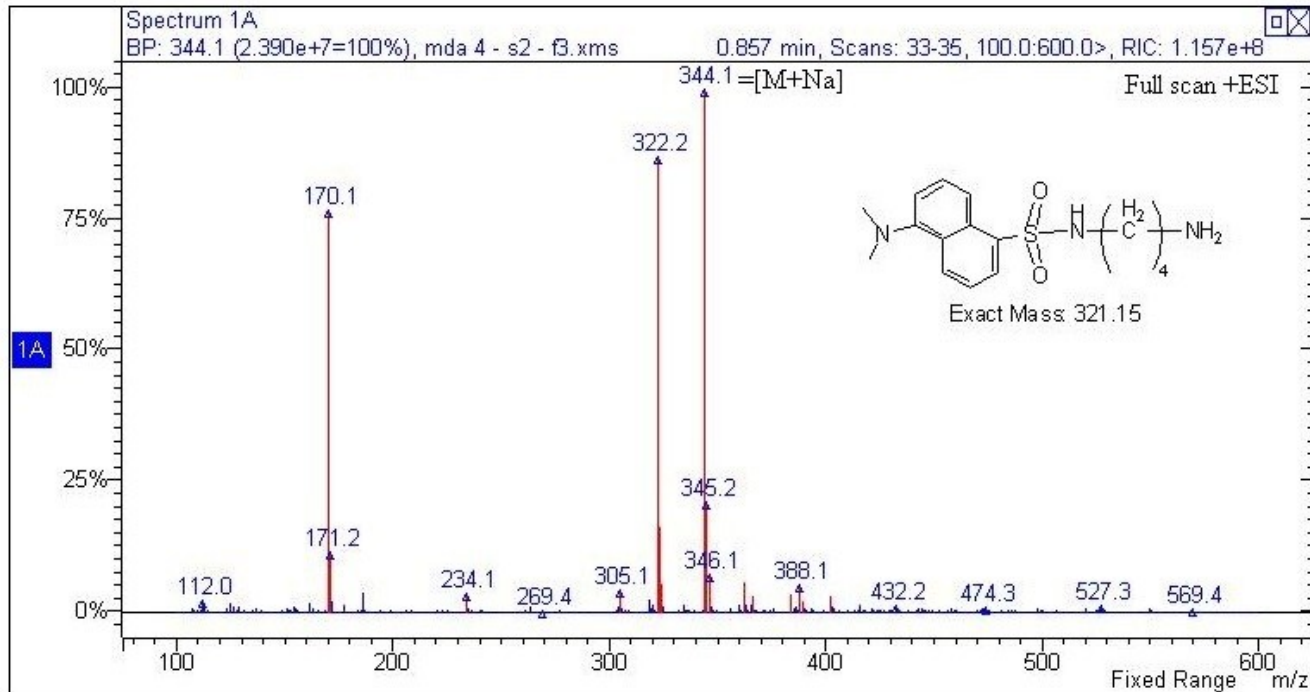


$^{13}\text{C}$  NMR spectrum of compound  $\text{DA}_{1.3}$

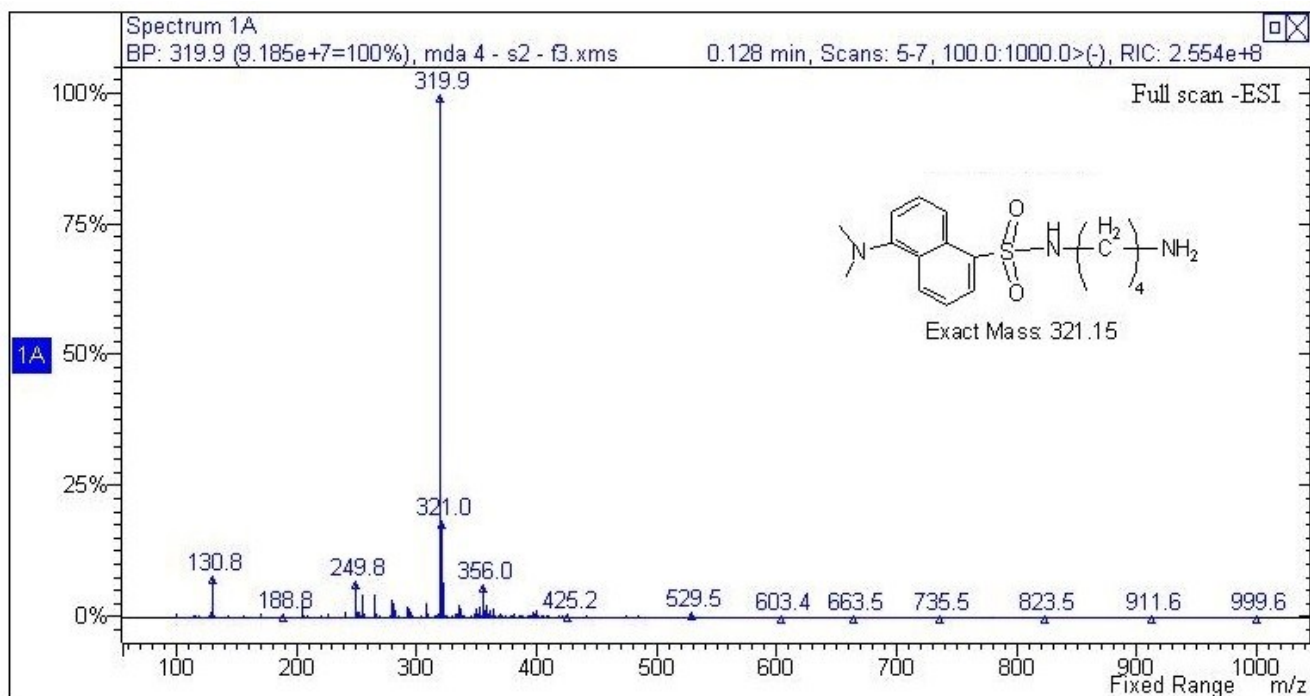


UV-Vis spectrum of compound  $\text{DA}_{1.3}$

### DA<sub>1,4</sub> (dansyl 1,4-diaminobutane)

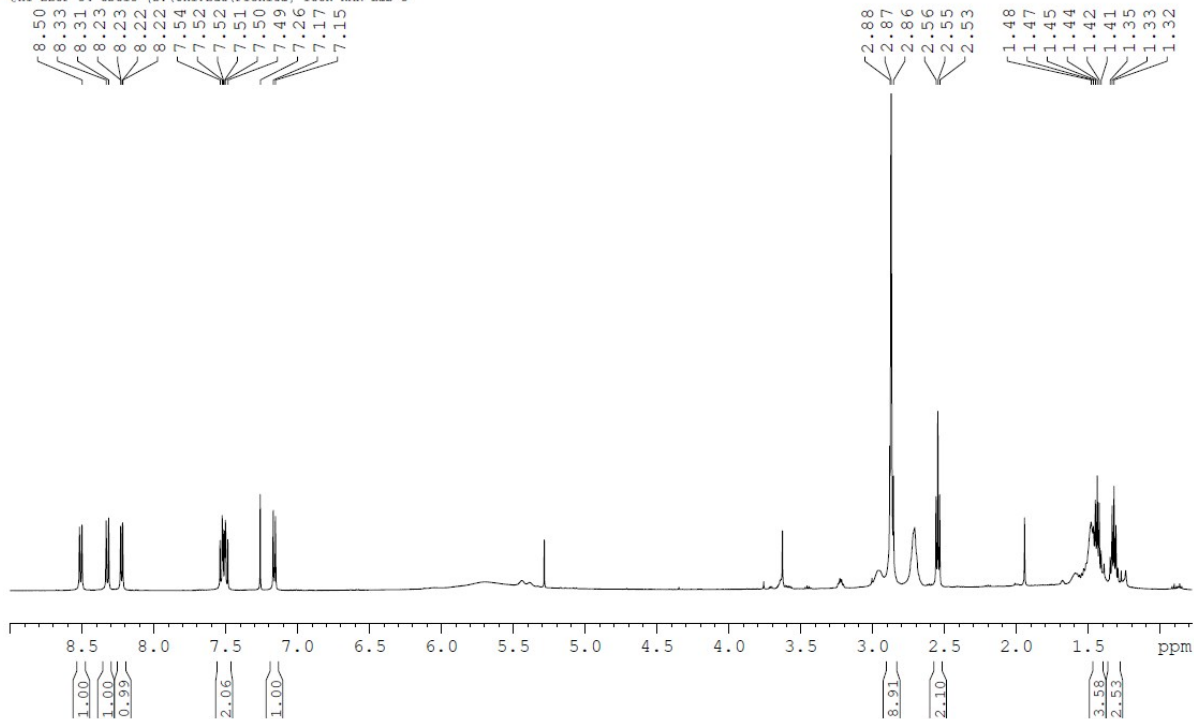


ESI-MS ( $m/z$ ): 321 (DA<sub>1,4</sub> + H<sup>+</sup>)



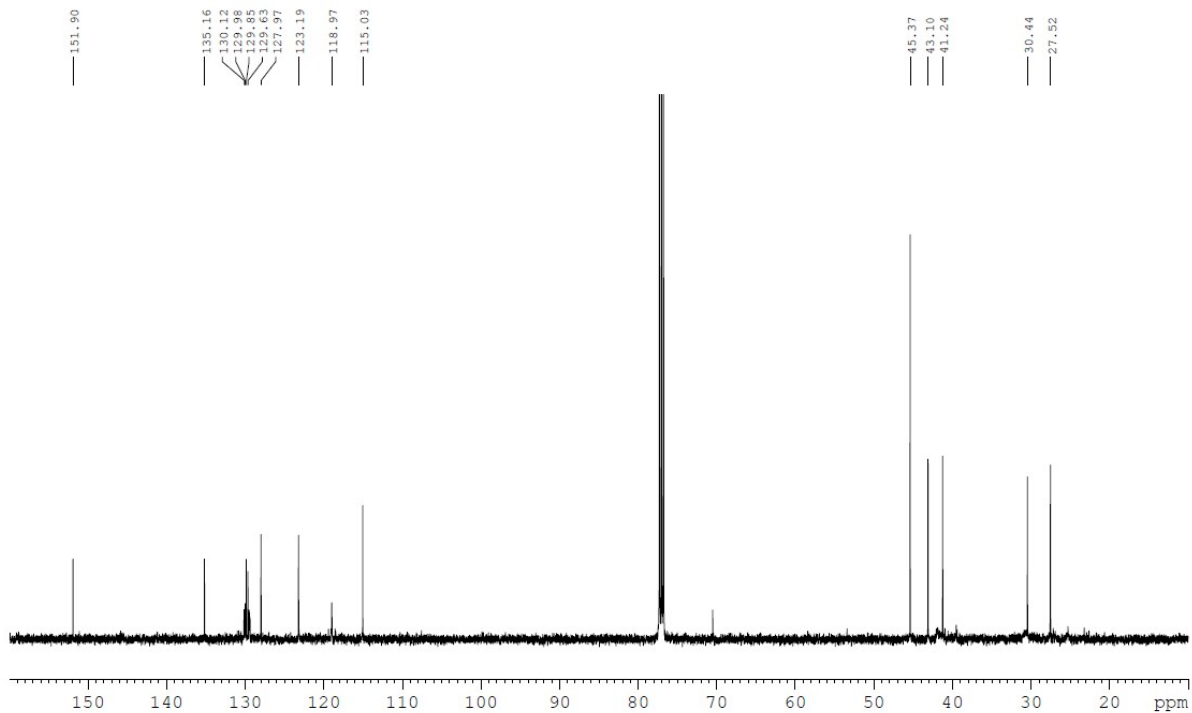
ESI-MS ( $m/z$ ): 320 (DA<sub>1,4</sub> - H<sup>+</sup>)

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
Operator CS  
Registry No. 2187  
User G. Ionita  
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8H1-BBOF-34 CDCl3 (D:\UnivBuc\Pionita) ICON-NMR-Lab 5

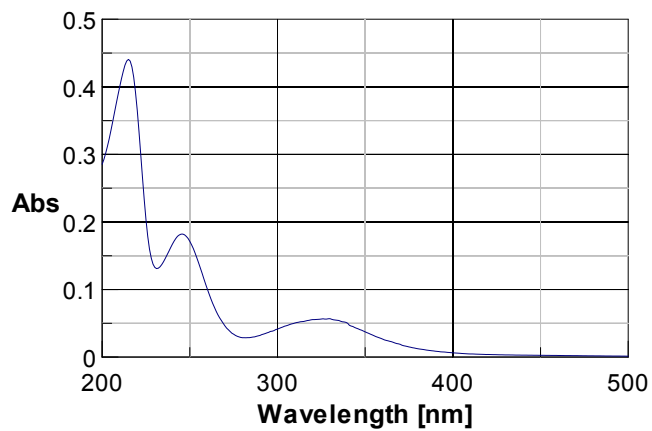


<sup>1</sup>H NMR spectrum of compound DA<sub>1.4</sub>

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
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User G. Ionita  
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2012-09-18 09:05:34

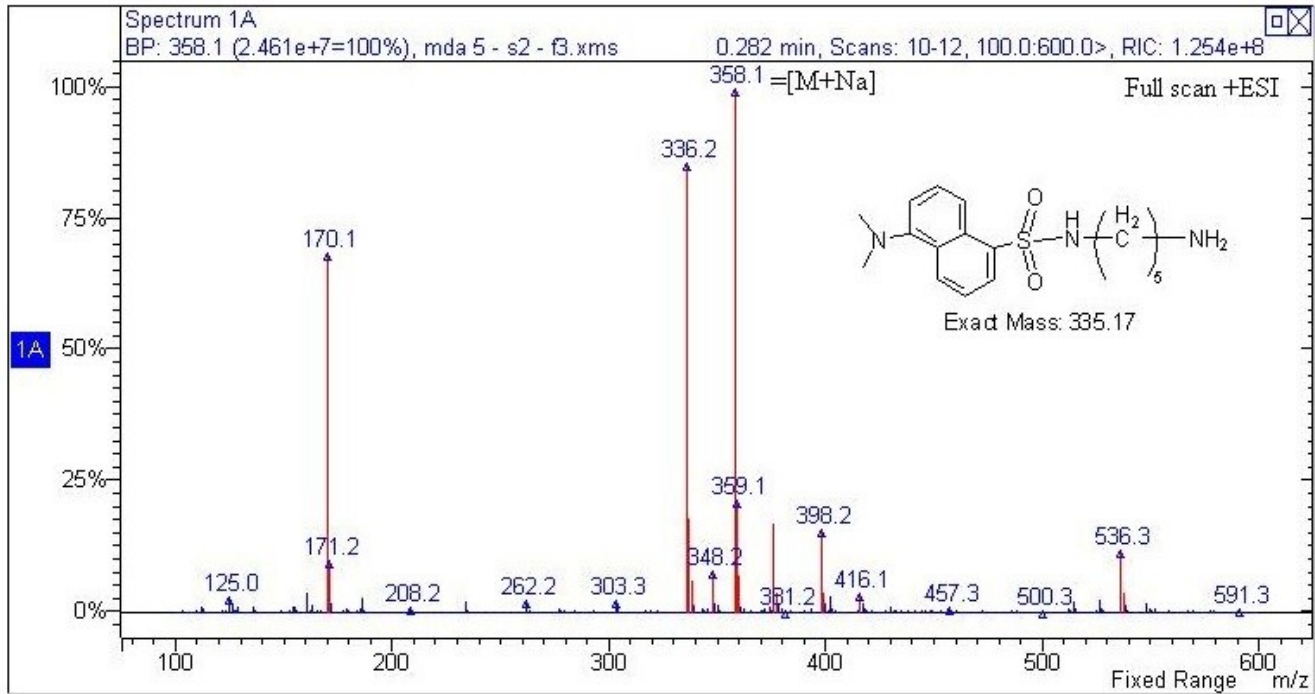


<sup>13</sup>C NMR spectrum of compound DA<sub>1.4</sub>

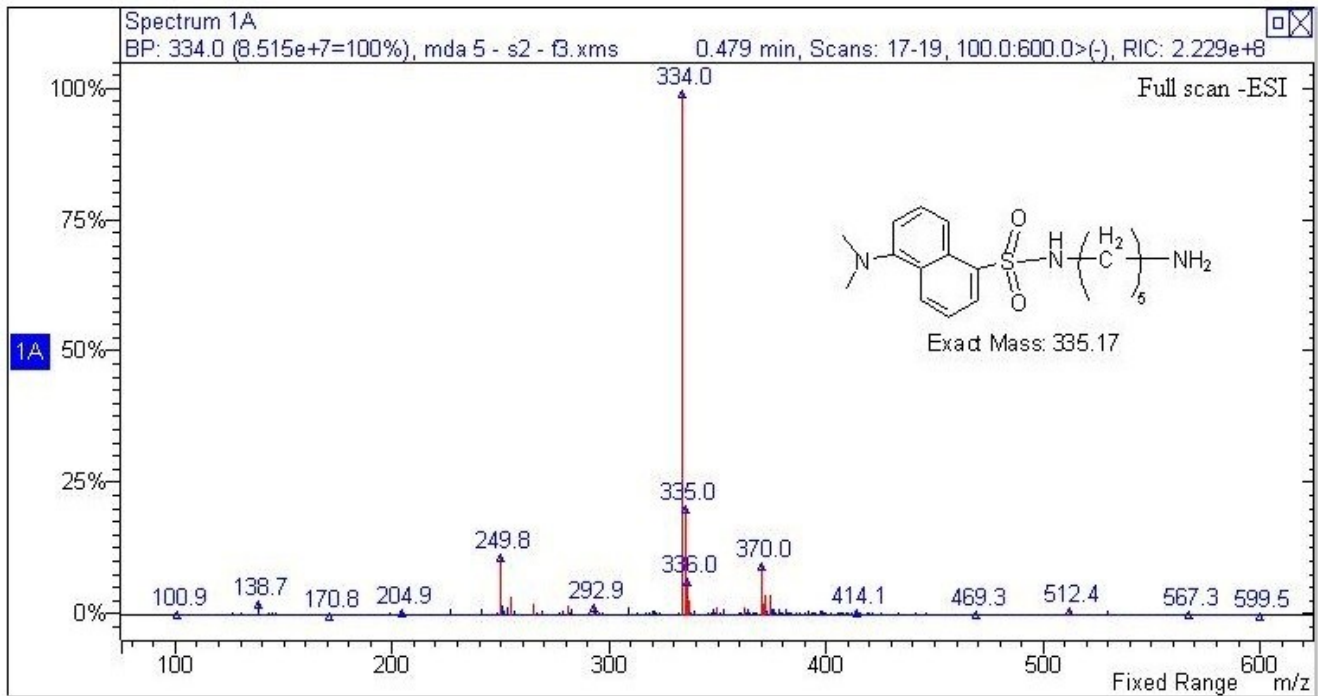


UV-Vis spectrum of compound DA<sub>1.4</sub>

### DA<sub>1.5</sub> (dansyl 1,5-diaminopentane)

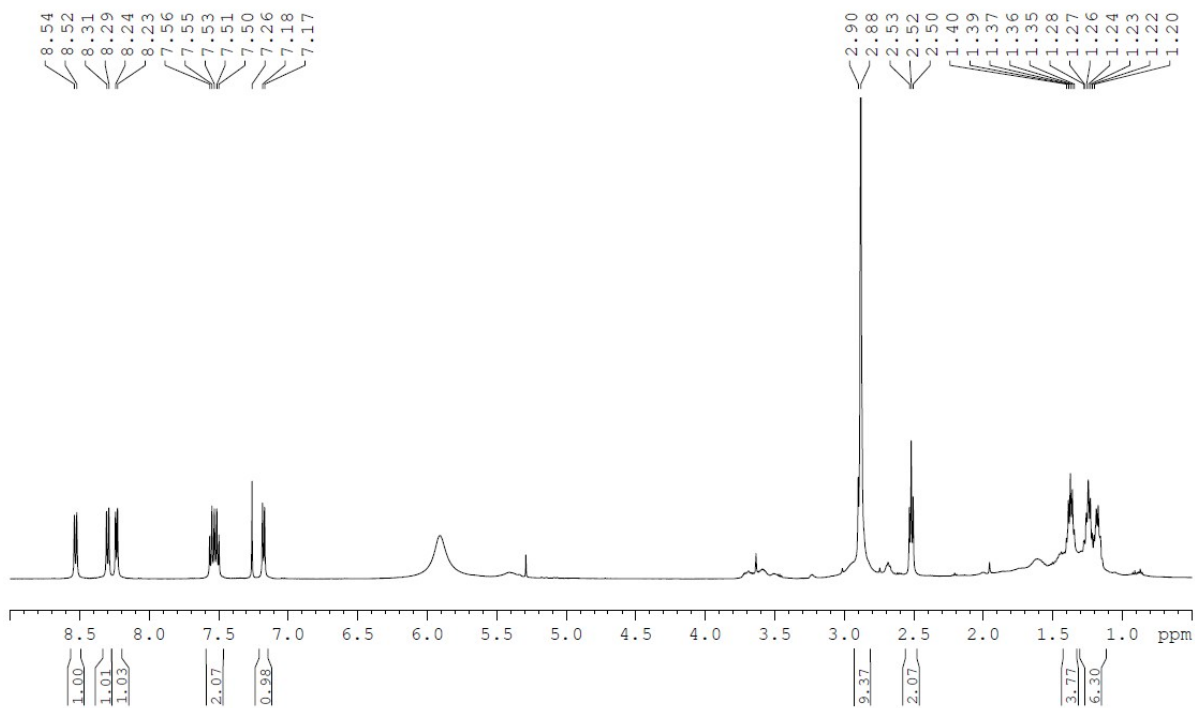


ESI-MS ( $m/z$ ): 335 (DA<sub>1.5</sub> + H<sup>+</sup>)



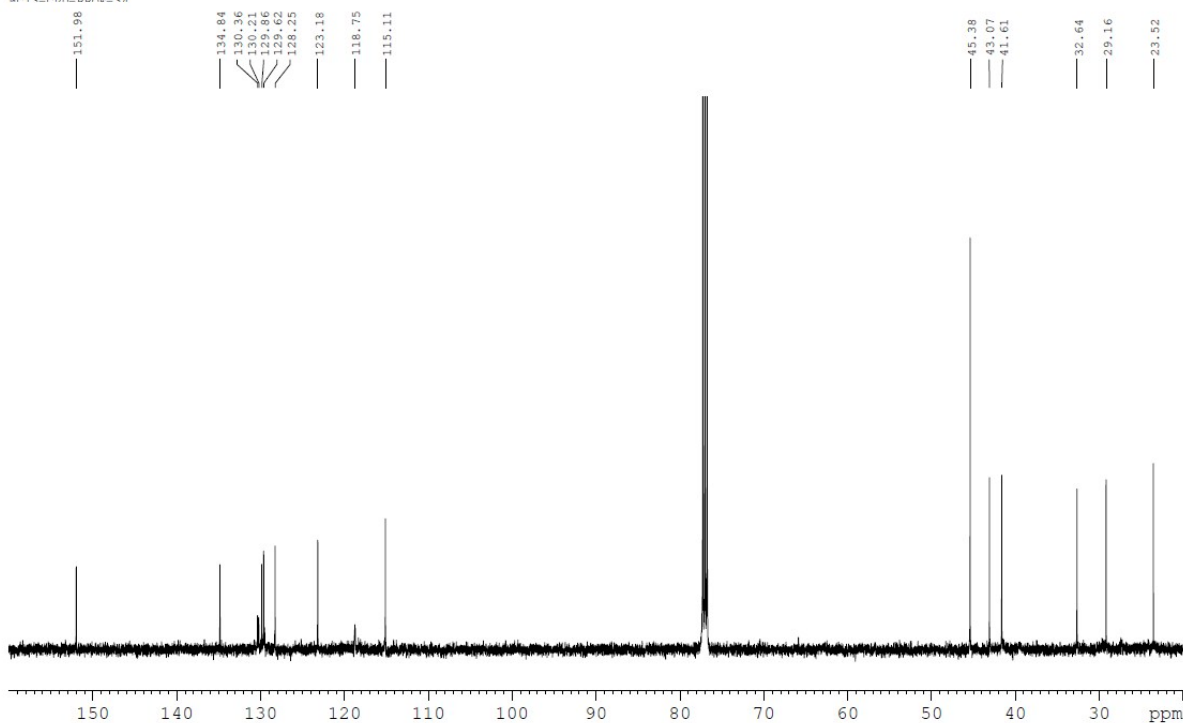
ESI-MS ( $m/z$ ): 334 (DA<sub>1.5</sub> - H<sup>+</sup>)

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
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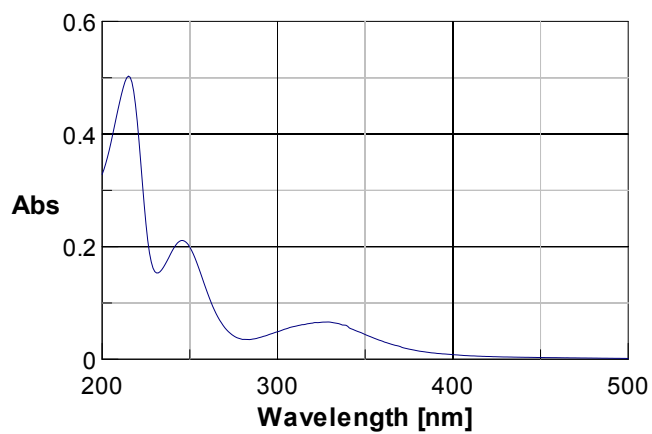


<sup>1</sup>H NMR spectrum of compound DA<sub>1.5</sub>

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
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User G. Ionita  
Sample Changer Position 13  
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CDCl3  
2013\_09\_18\_09:28:28



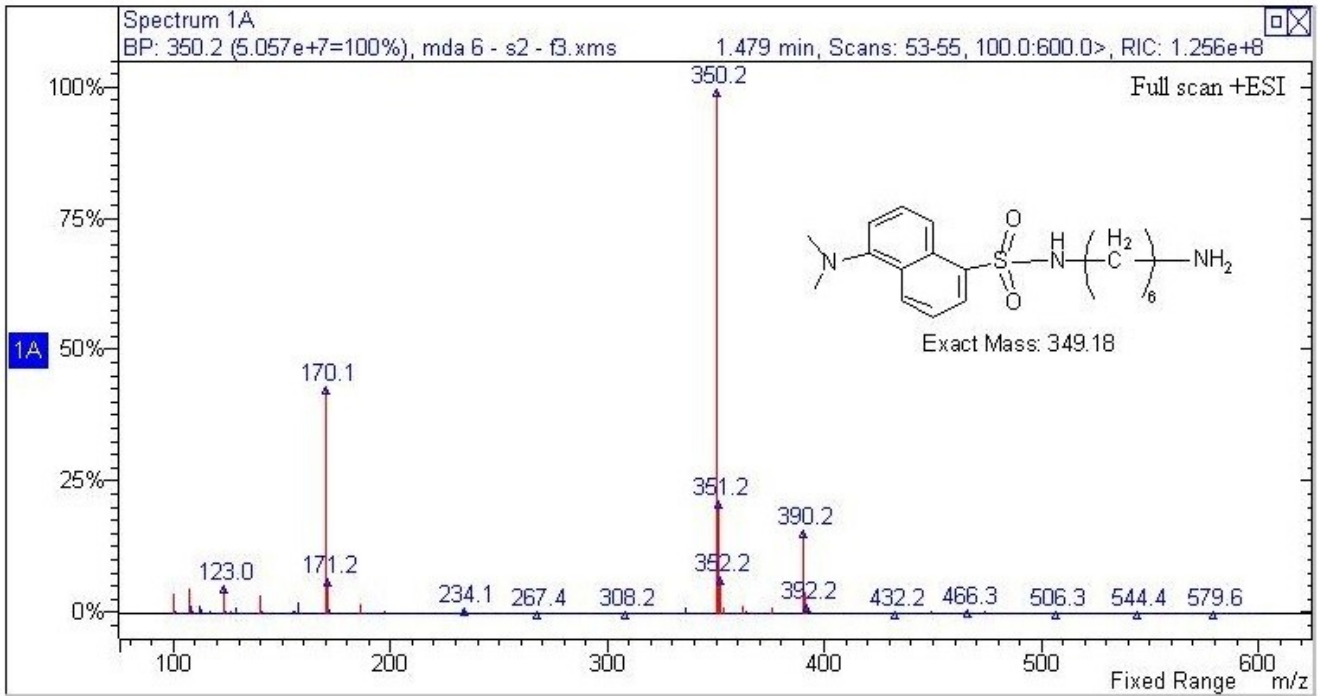
<sup>13</sup>C NMR spectrum of compound DA<sub>1.5</sub>



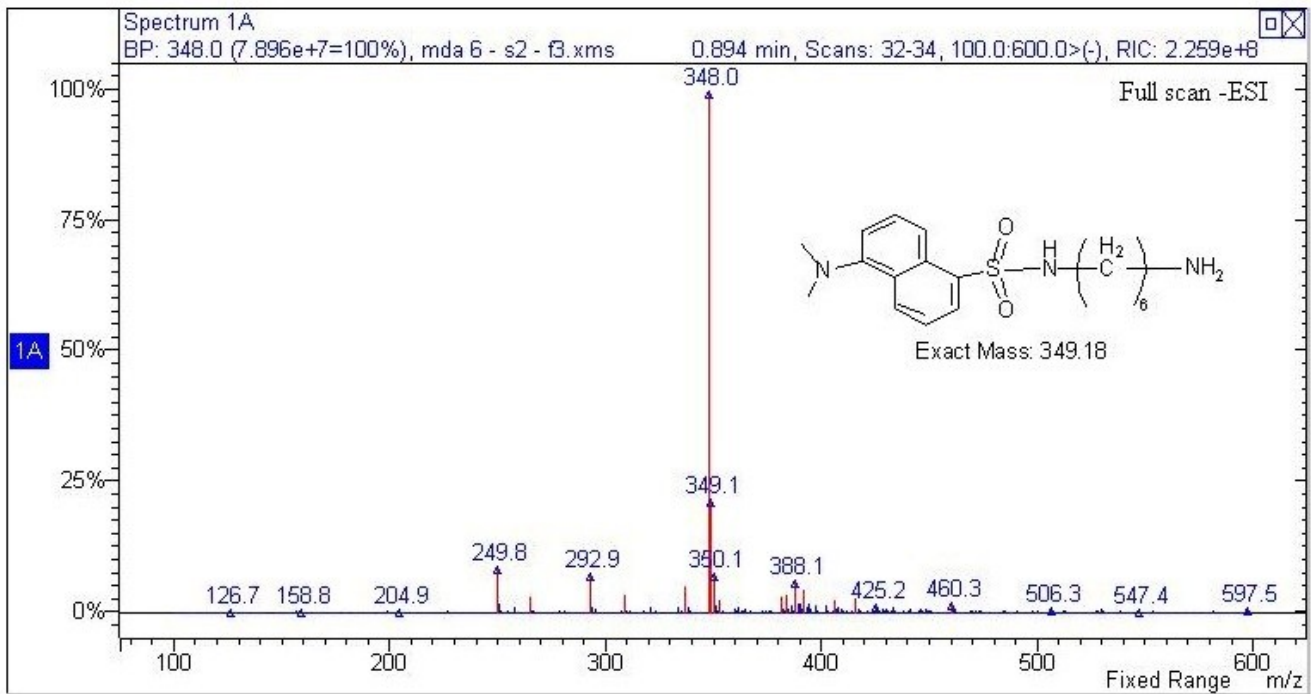
UV-Vis spectrum of compound DA<sub>1.5</sub>



# DA<sub>1.6</sub> (dansyl 1,6-diaminohexane)

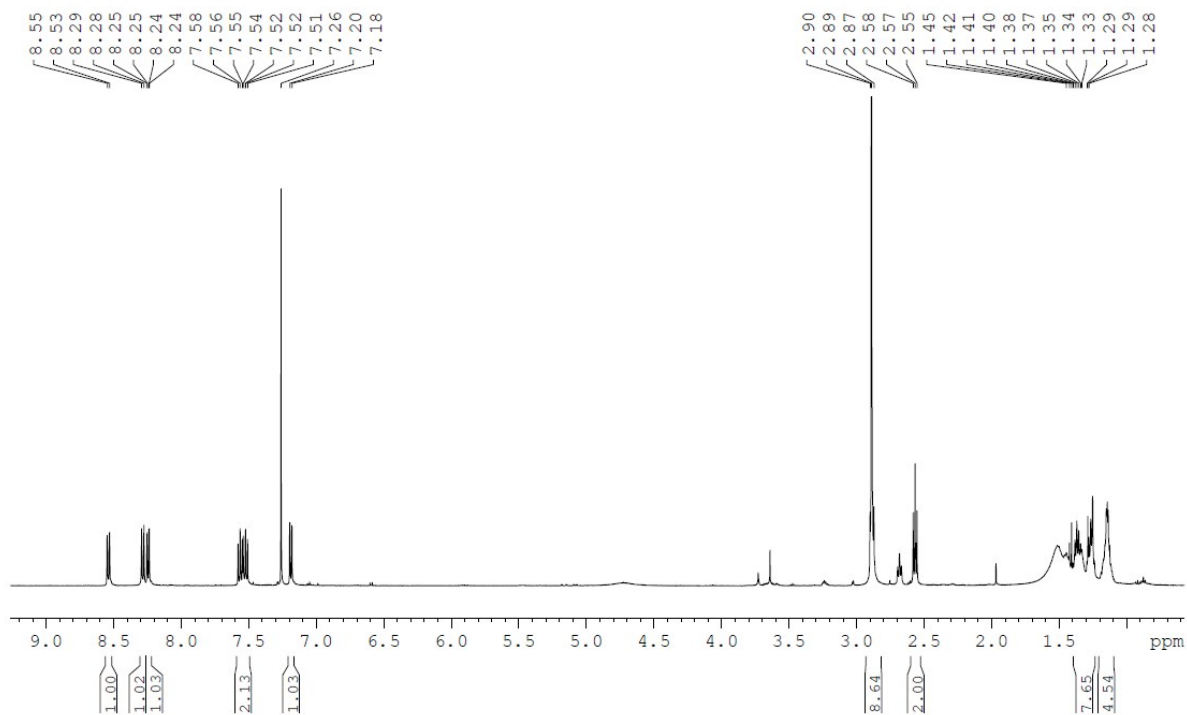


ESI-MS ( $m/z$ ): 350 (DA<sub>1.6</sub> + H<sup>+</sup>)



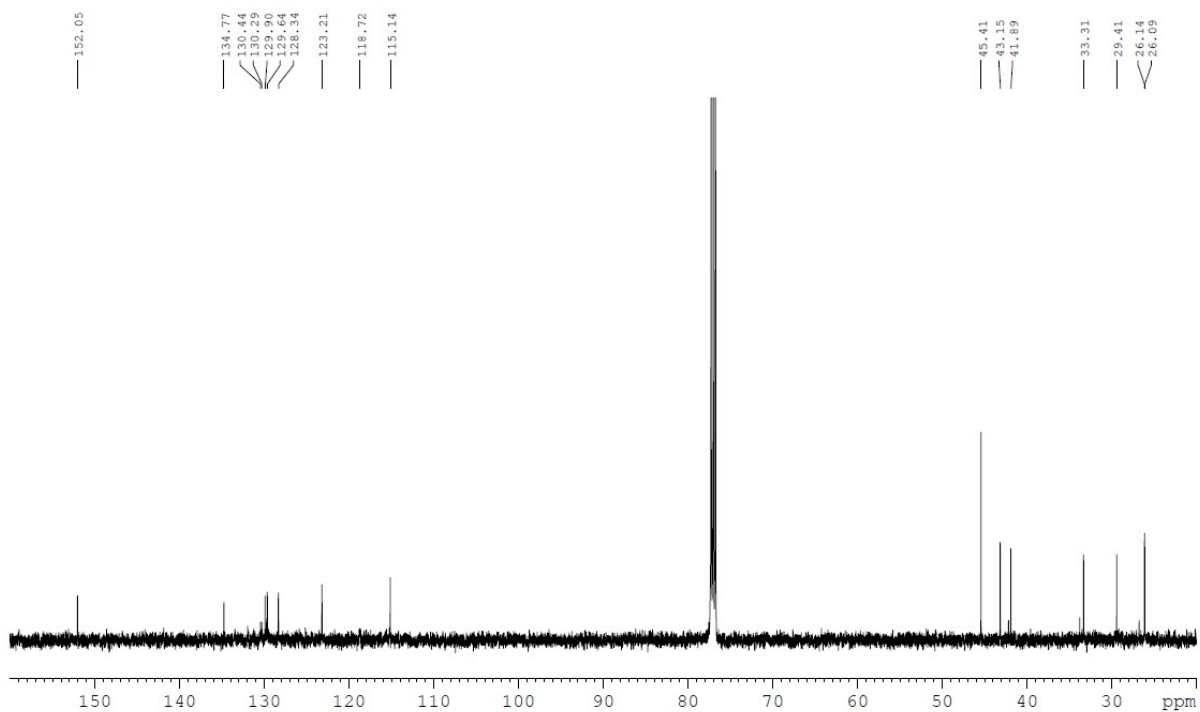
ESI-MS ( $m/z$ ): 348 (DA<sub>1.6</sub> - H<sup>+</sup>)

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
 Operator CS  
 Registry No. 2189  
 User G. Ionita  
 Sample Changer Position 6  
 Sample name: MAD-6  
 8H1-BBOP-34 CDCl3 (D:\UnivBuc\Pionita) ICON-NMR-Lab 5

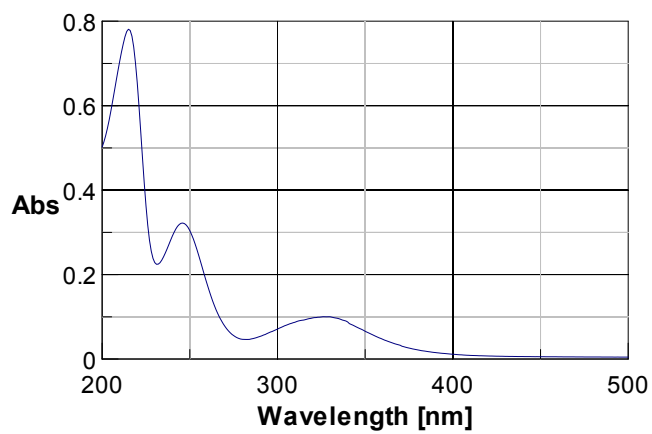


**<sup>1</sup>H NMR spectrum of compound DA<sub>1.6</sub>**

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
Operator CS  
Registry No. 2189  
User G. Ionita  
Sample Changer Position 6  
Sample name: MAD-6  
CDCl3  
9712\_PPM\_0000\_24

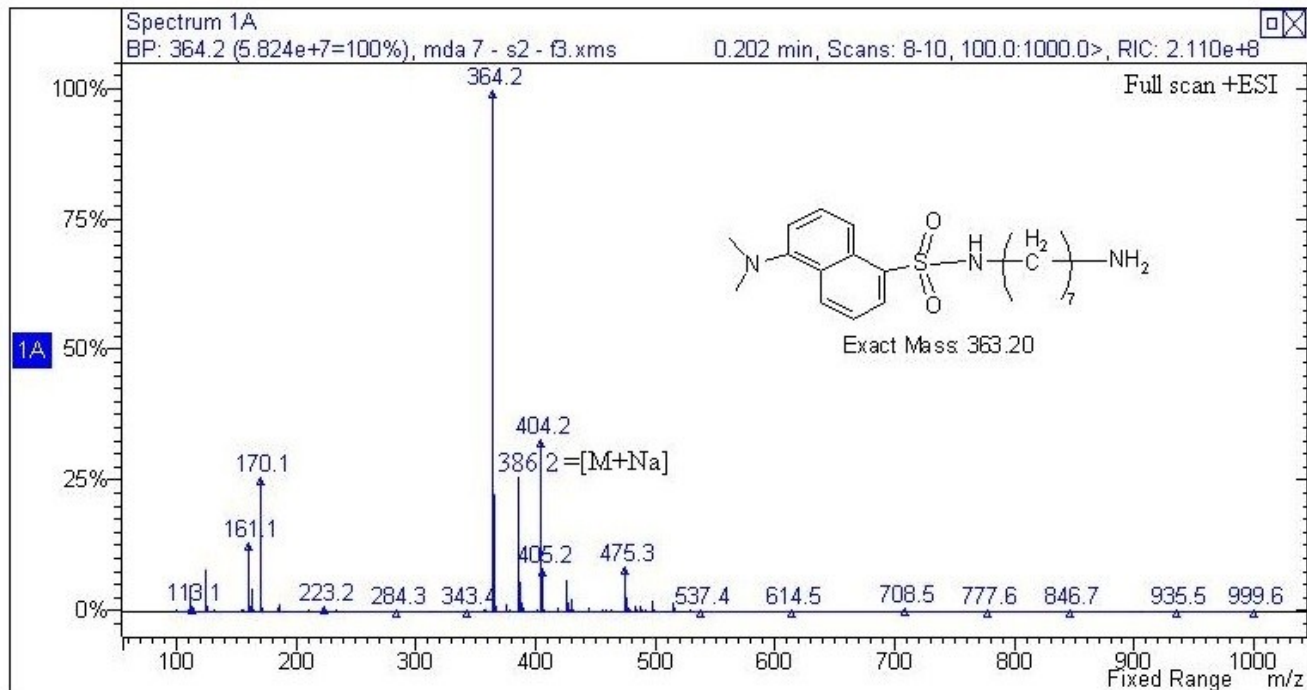


<sup>13</sup>C NMR spectrum of compound DA<sub>1.6</sub>

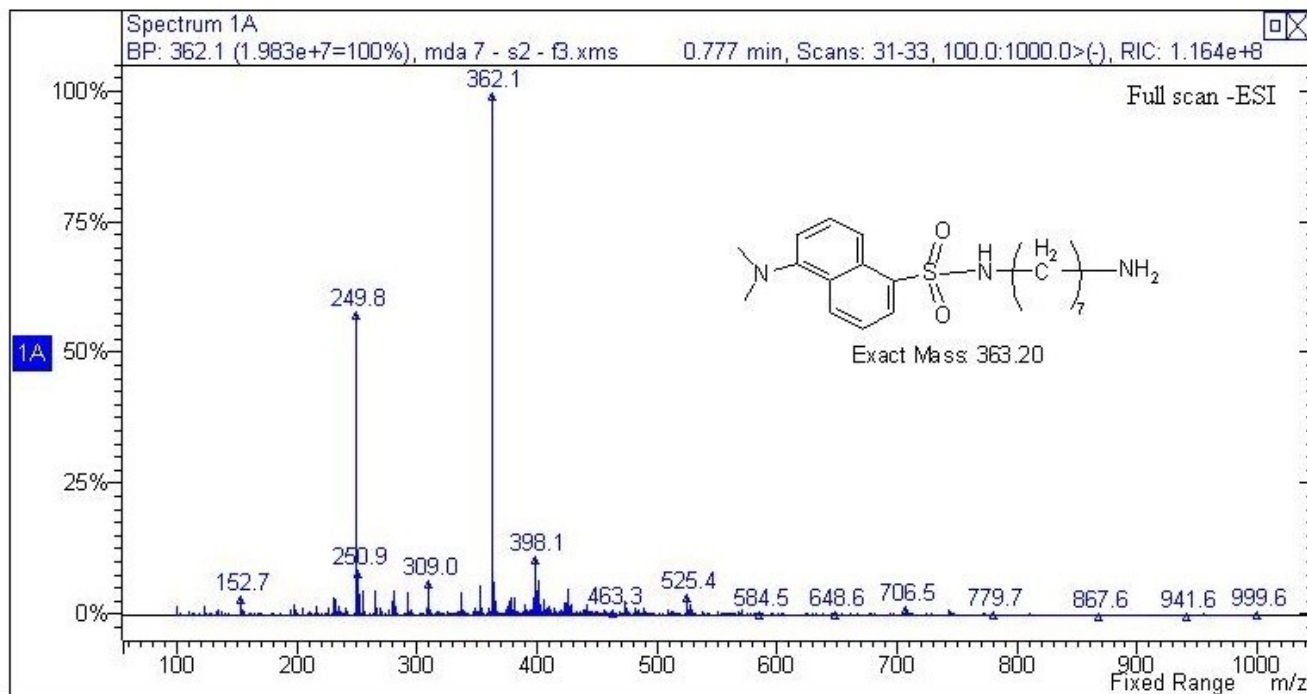


UV-Vis spectrum of compound DA<sub>1.6</sub>

### DA<sub>1,7</sub> (dansyl 1,7-diaminoheptane)

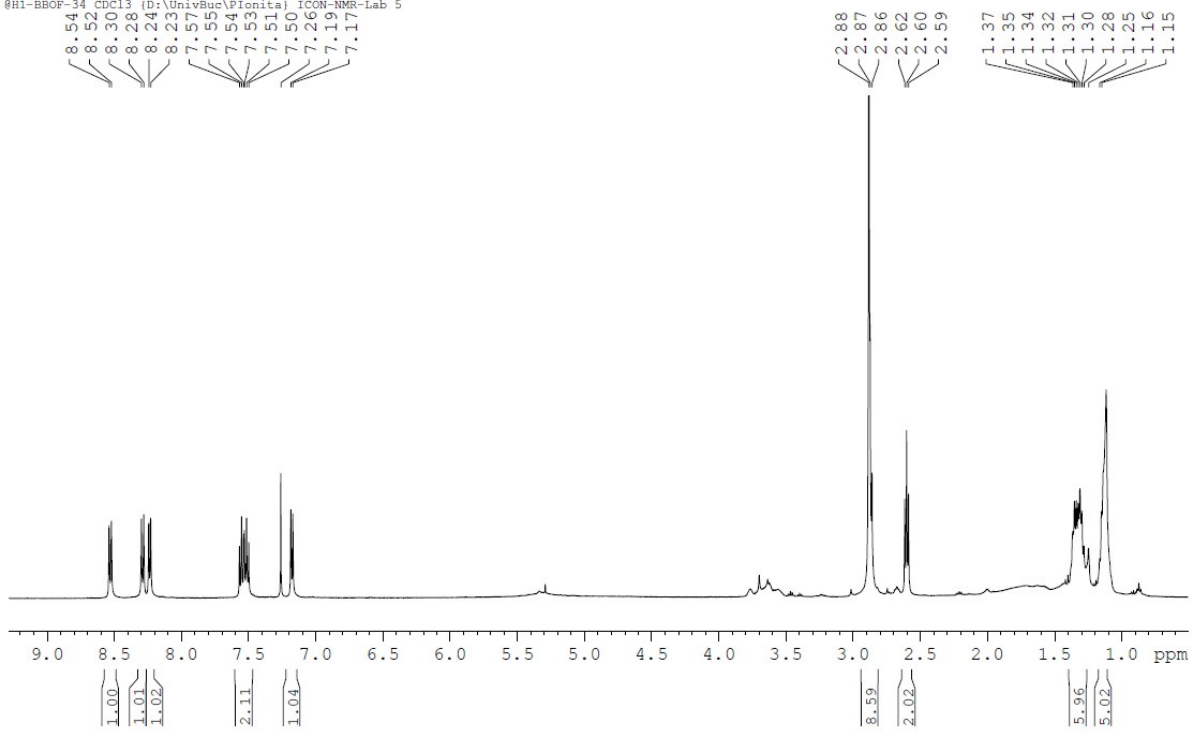


ESI-MS ( $m/z$ ): 364 (DA<sub>1,7</sub> + H<sup>+</sup>)



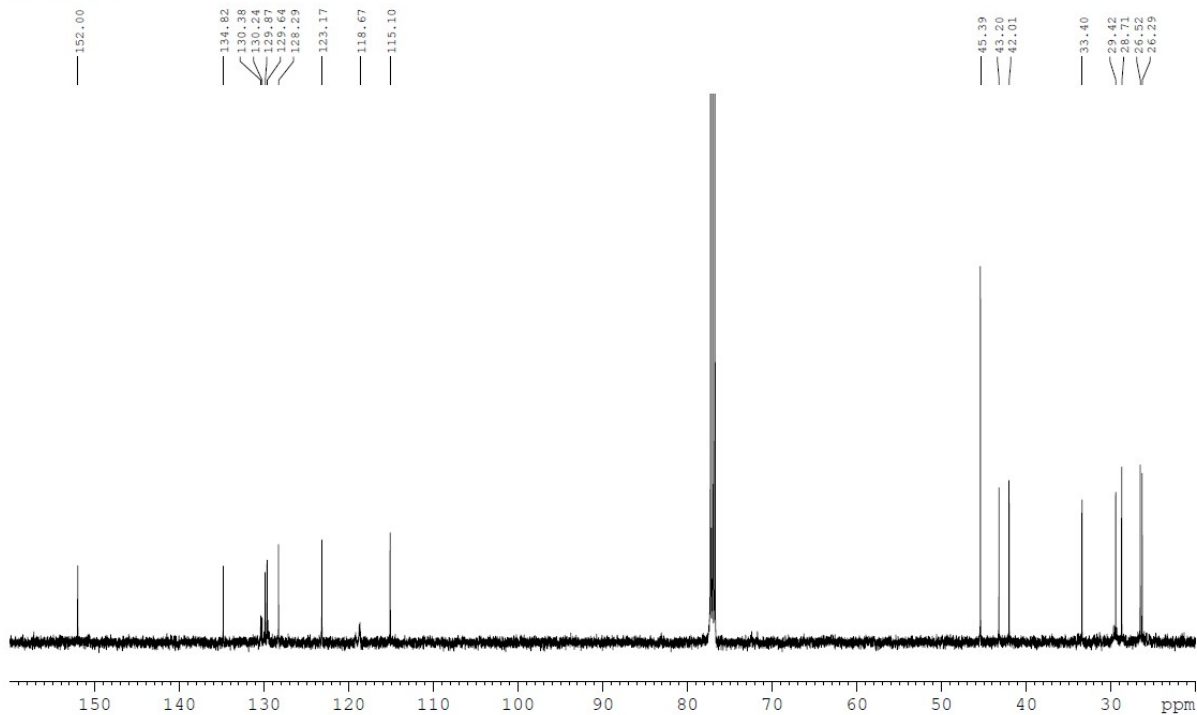
ESI-MS ( $m/z$ ): 362 (DA<sub>1,7</sub> - H<sup>+</sup>)

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
 Operator CS  
 Registry No. 2190  
 User G. Ionita  
 Sample Changer Position 11  
 Sample name: MAD-7  
 #H1-BBOP-34 CDCl3 (D:\UnivBuc\PIonita) ICON-NMR-Lab 5

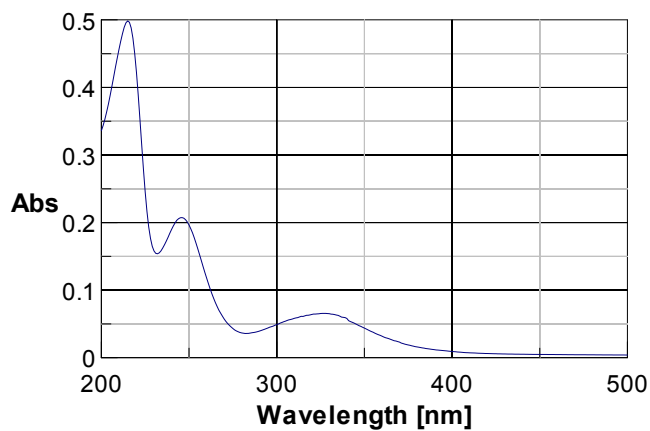


<sup>1</sup>H NMR spectrum of compound DA<sub>1.7</sub>

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
Operator CS  
Registry No. 2190  
User G. Ionita  
Sample Changer Position 11  
Sample name: MAD-7  
CDCl3  
2013\_09\_09\_09\_24

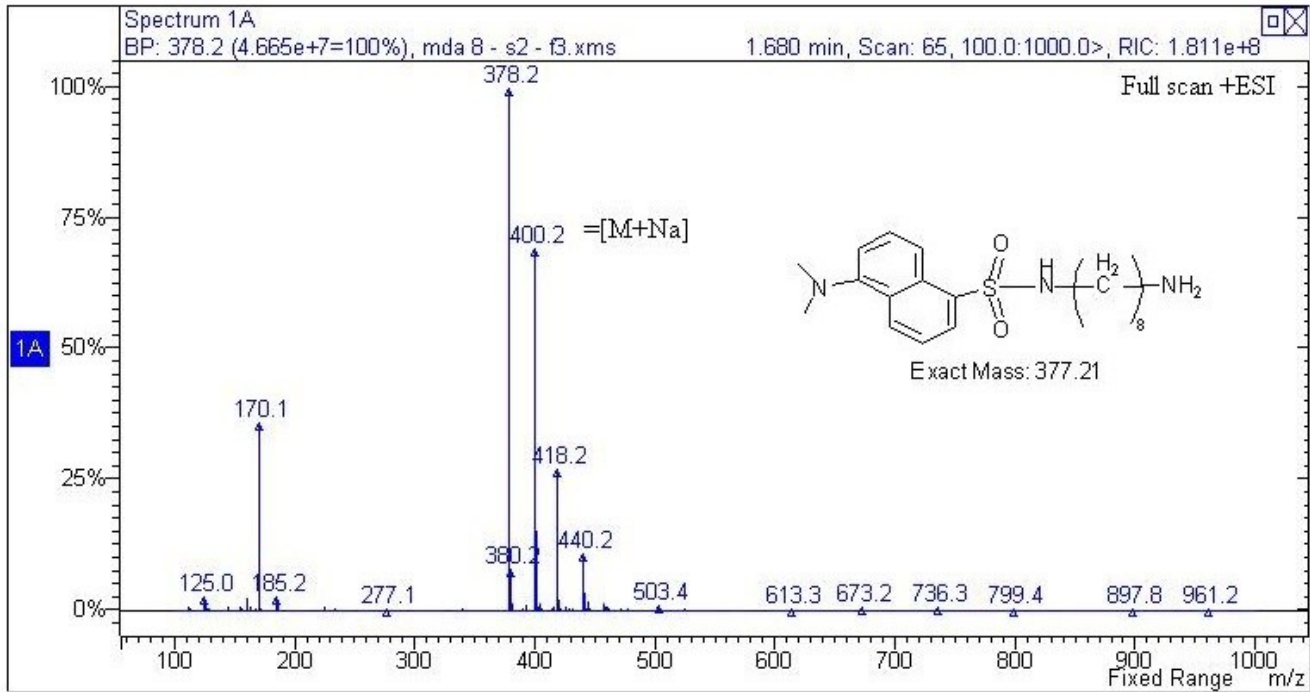


<sup>13</sup>C NMR spectrum of compound DA<sub>1.7</sub>

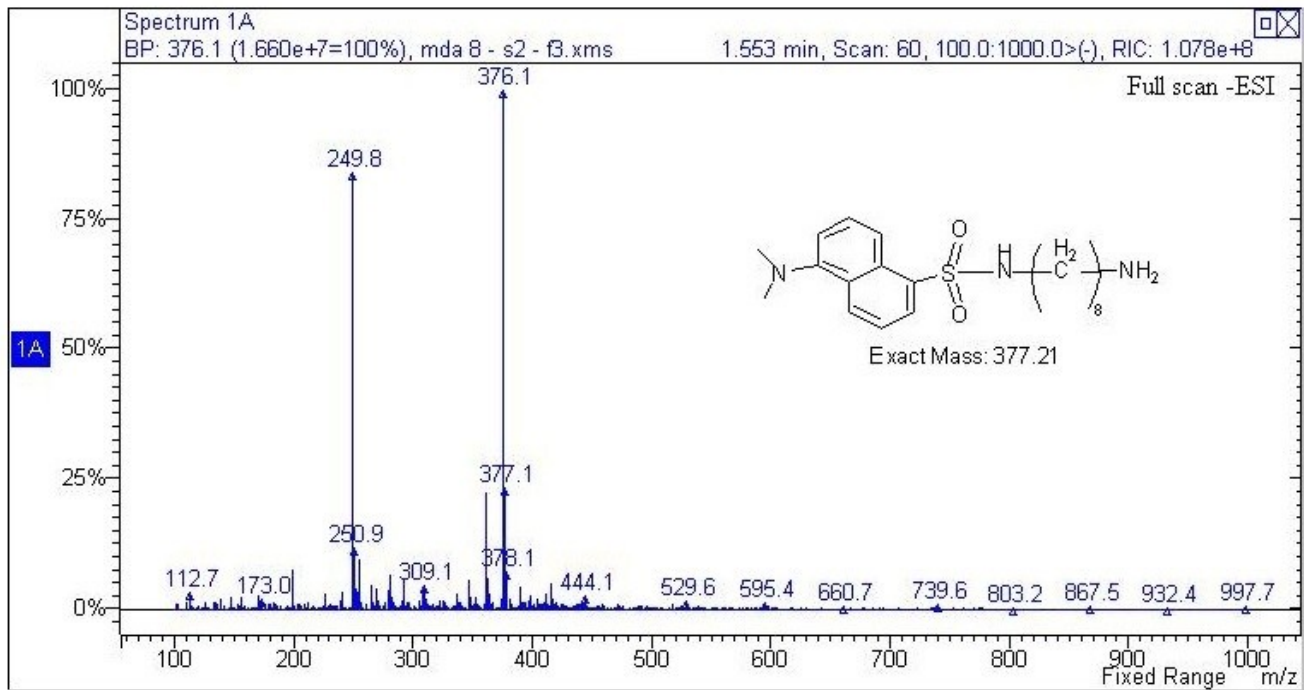


UV-Vis spectrum of compound DA<sub>1.7</sub>

# DA<sub>1,8</sub> (dansyl 1,8-diaminooctane)

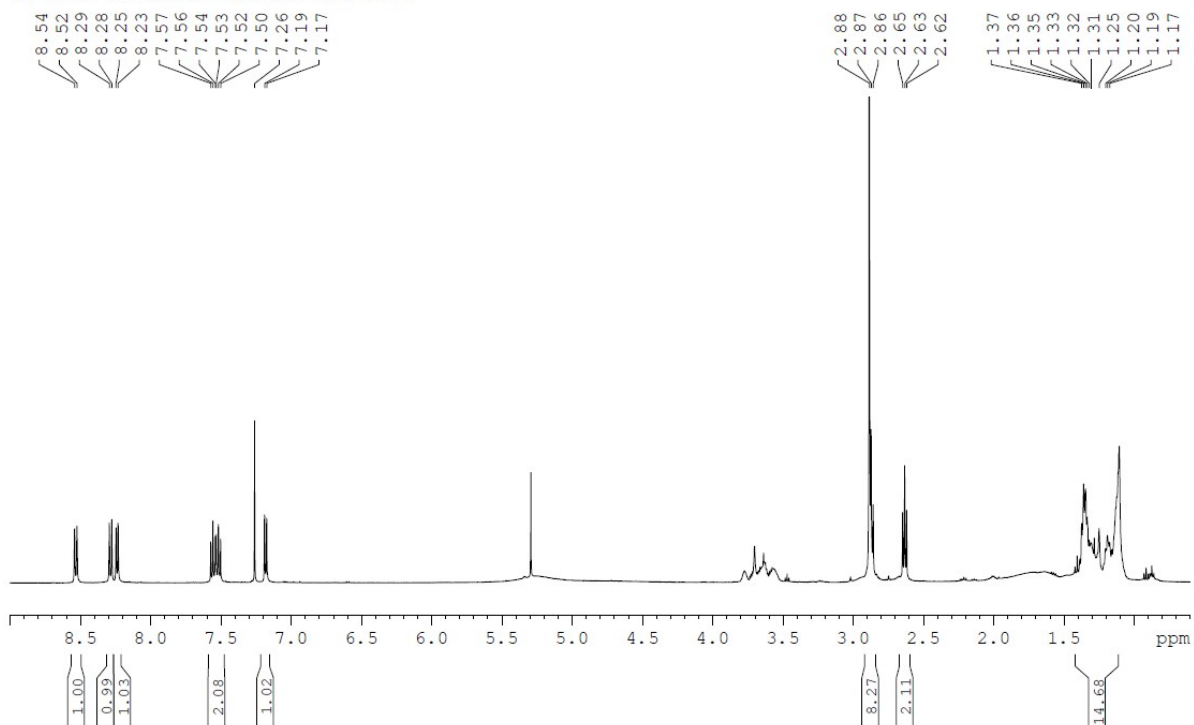


ESI-MS ( $m/z$ ): 378 (DA<sub>1,8</sub> + H<sup>+</sup>)



ESI-MS ( $m/z$ ): 376 (DA<sub>1,8</sub> - H<sup>+</sup>)

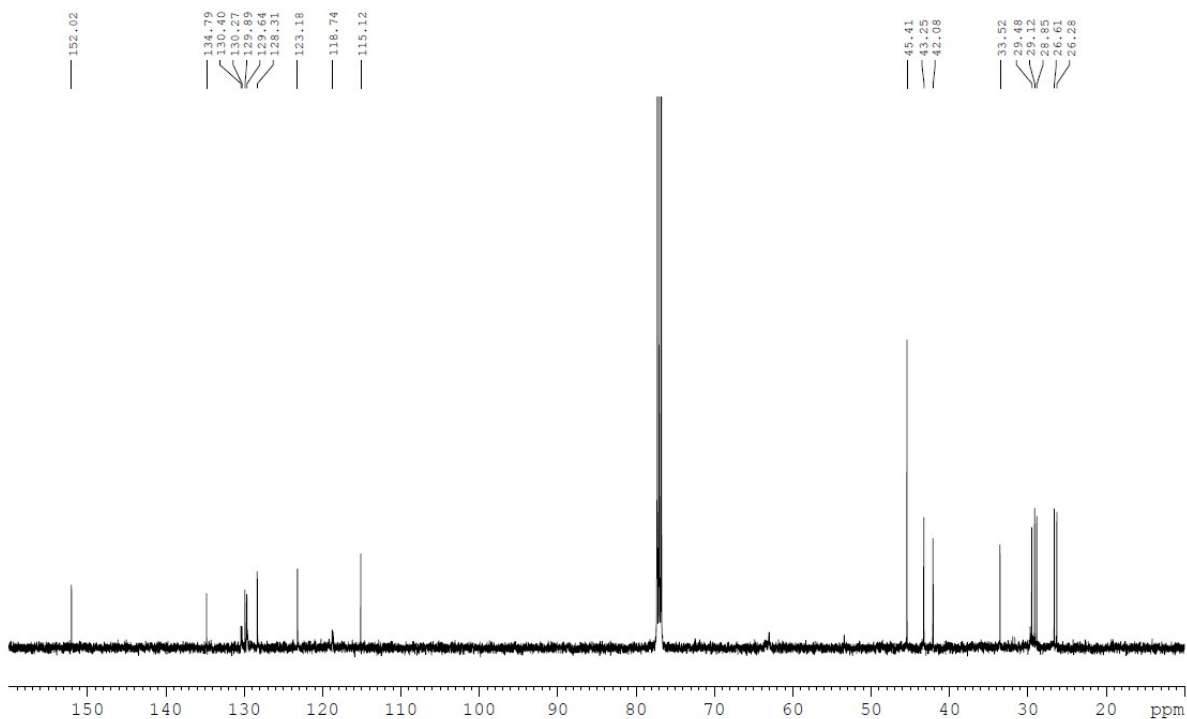
Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
Operator CS  
Registry No. 2191  
User C. Ionita  
Sample Changer Position 4  
Sample name: MAD-8  
@H1-BBOF-34 CDCl3 (D:\UnivBuc\Plonita) ICON-NMR-Lab 5



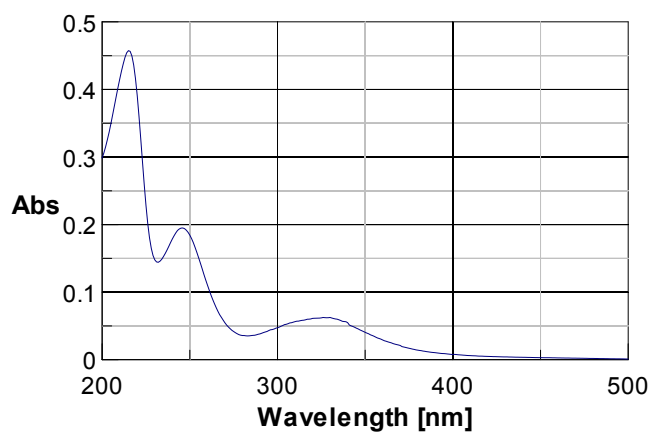
$^1\text{H}$  NMR spectrum of compound DA<sub>1.8</sub>



Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
Operator CS  
Registry No. 2191  
User G. Ionita  
Sample Changer Position 4  
Sample name: MAD-8  
CDCl3  
SMILES: O=C1C=CC(=O)C=C1

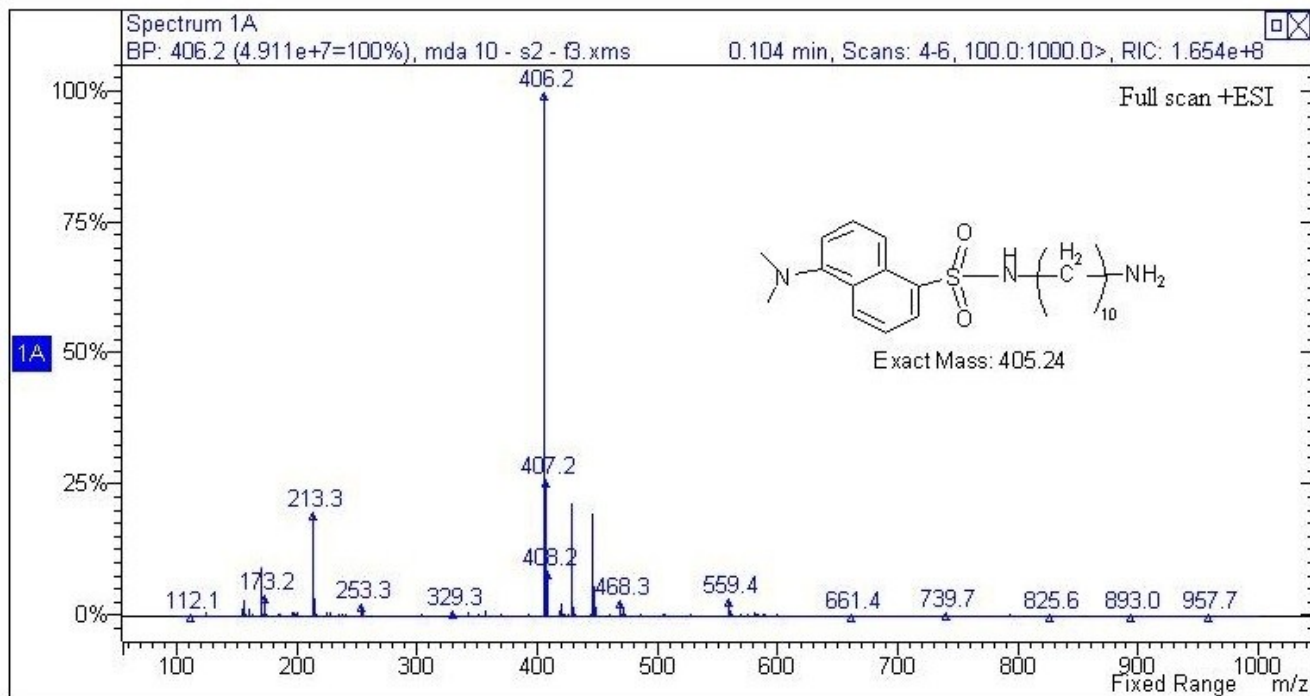


<sup>13</sup>C NMR spectrum of compound DA<sub>1.8</sub>

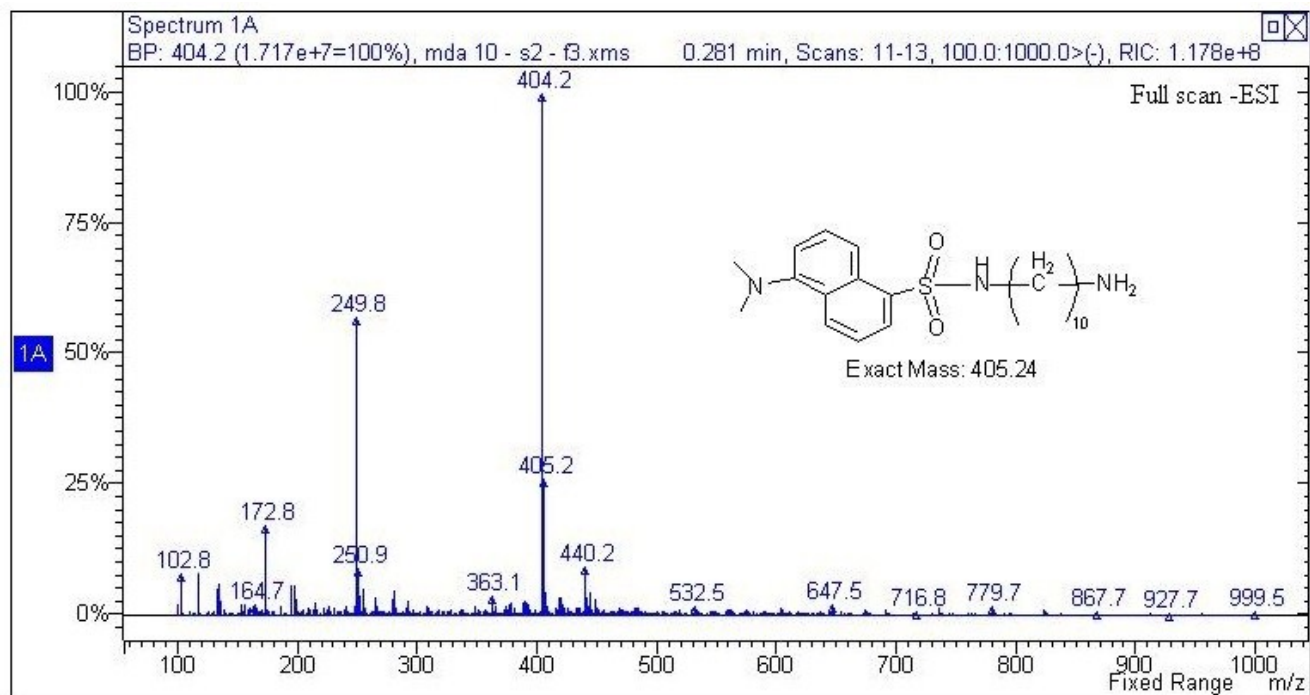


UV-Vis spectrum of compound DA<sub>1.8</sub>

### DA<sub>1.10</sub> (dansyl 1,10-diaminodecane)

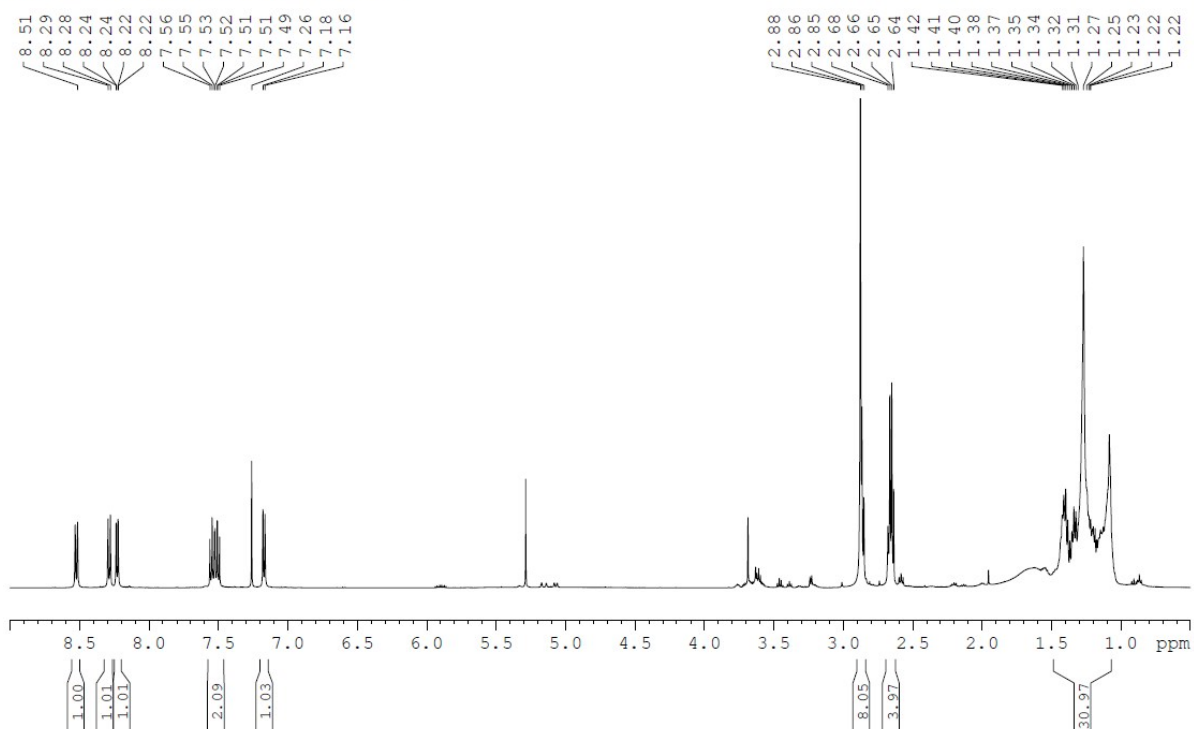


ESI-MS ( $m/z$ ): 406 (DA<sub>1.10</sub> + H<sup>+</sup>)



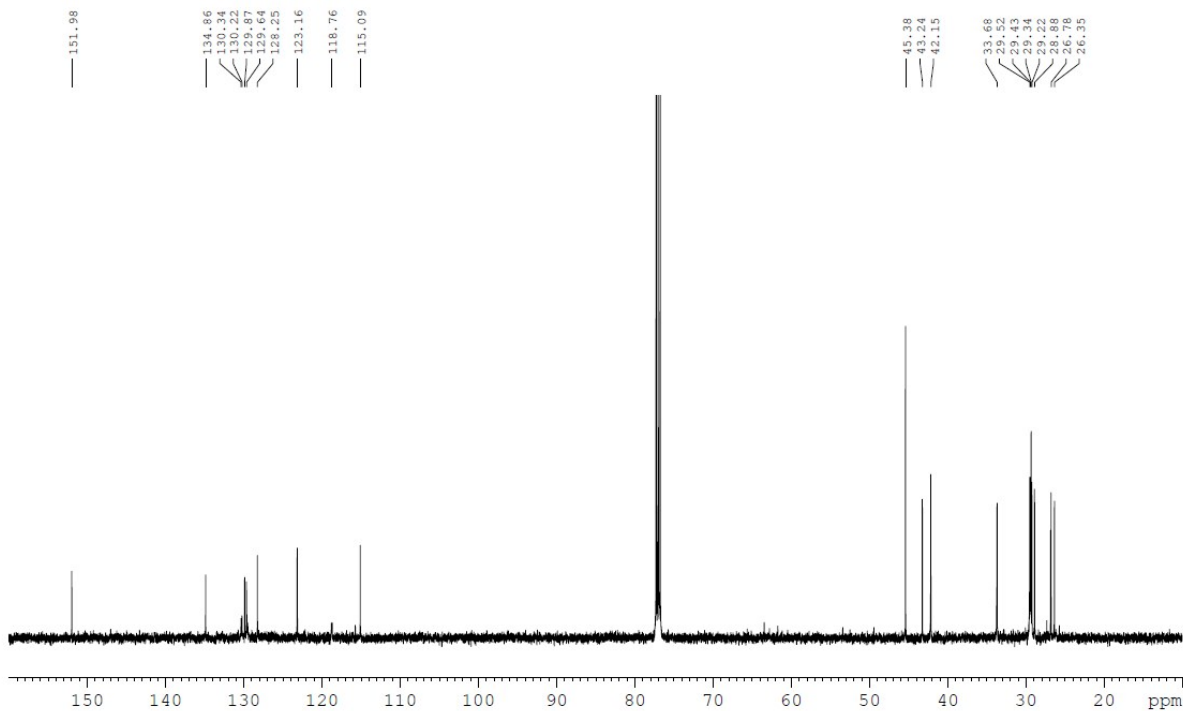
ESI-MS ( $m/z$ ): 404 (DA<sub>1.10</sub> - H<sup>+</sup>)

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
Operator CS  
Registry No. 2192  
User G. Ionita  
Sample Changer Position 9  
Sample name: MAD-10  
@H1-BBOF-34 CDCl3 (D:\UnivBuc\Pionita) ICON-NMR-Lab 5

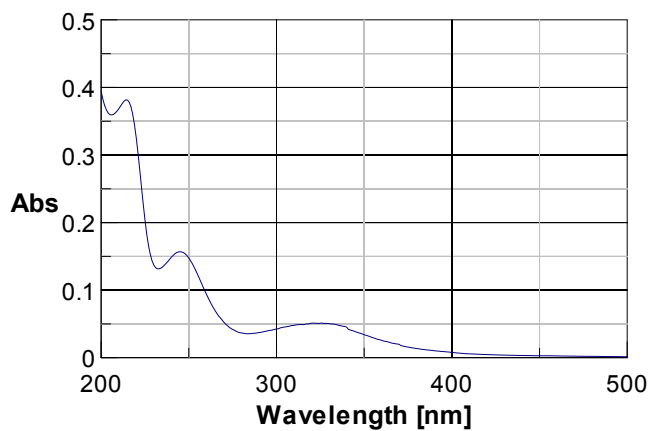


**<sup>1</sup>H NMR spectrum of compound DA<sub>1.10</sub>**

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
Operator CS  
Registry No. 2192  
User G. Ionita  
Sample Changer Position 9  
Sample name: MAD-10  
CDCl3  
@Cl3-CPD-BBOF-34

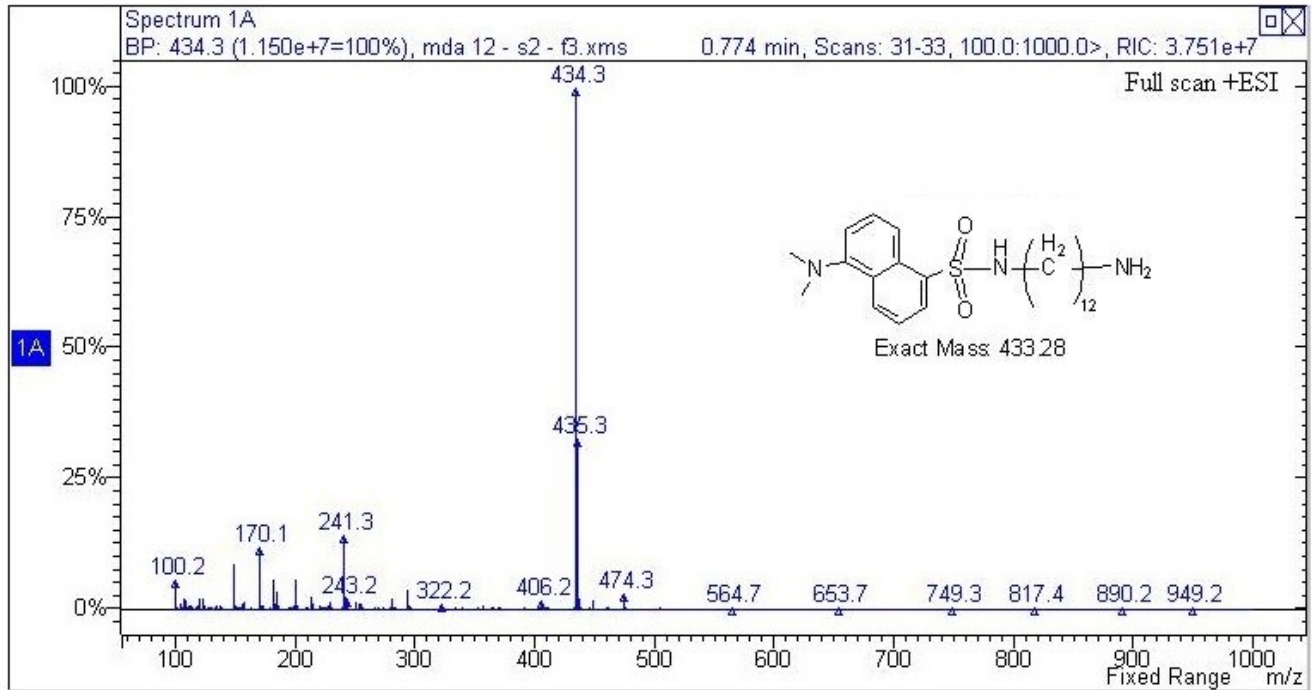


<sup>13</sup>C NMR spectrum of compound DA<sub>1.10</sub>

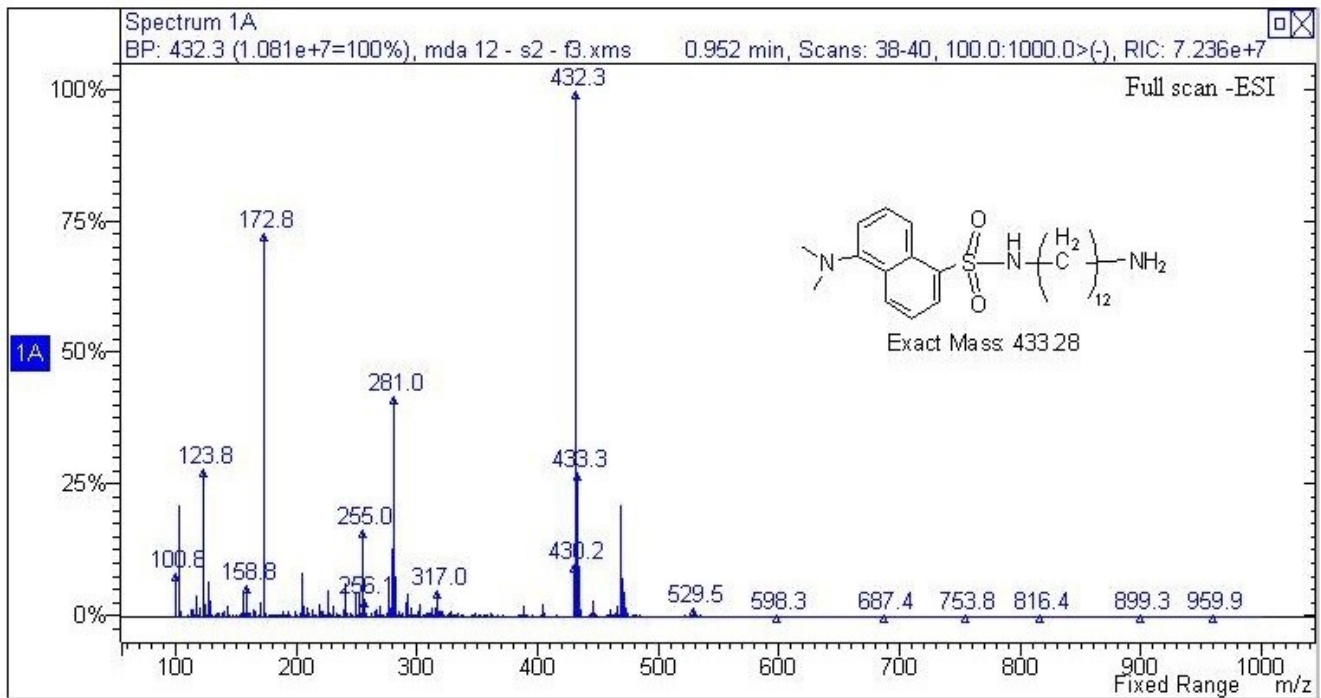


UV-Vis spectrum of compound DA<sub>1.10</sub>

# DA<sub>1.12</sub> (dansyl 1,12-diaminododecane)

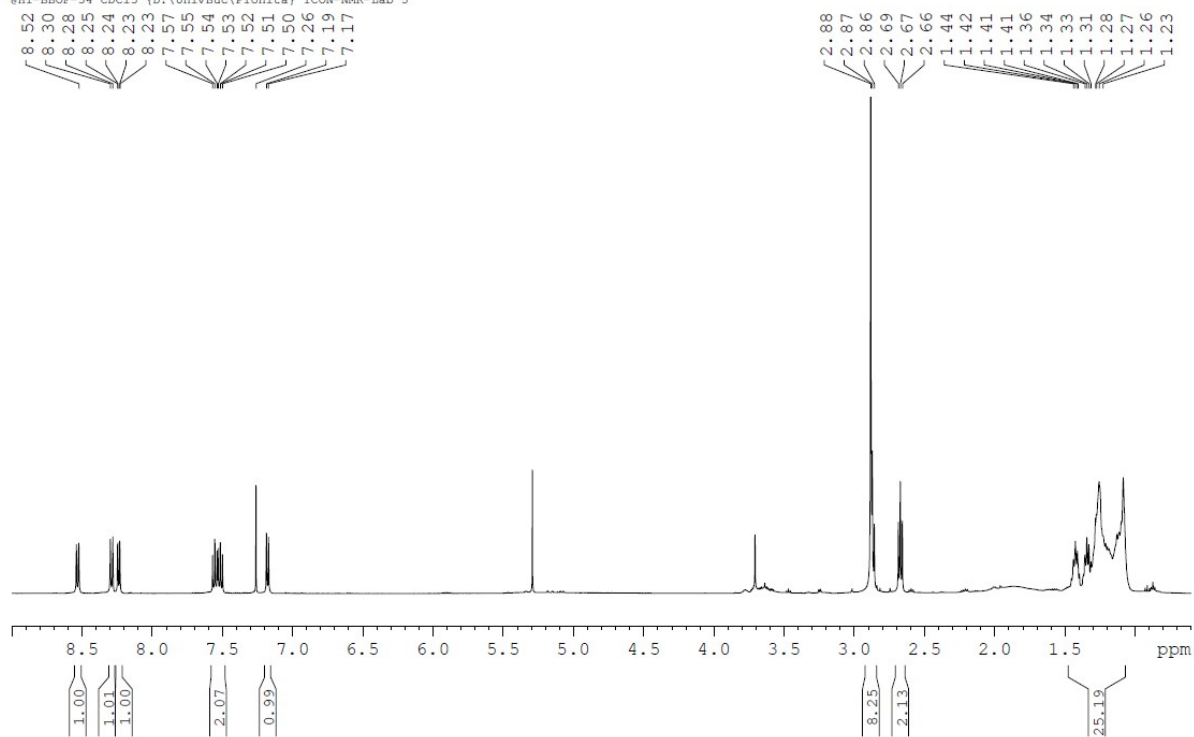


ESI-MS ( $m/z$ ): 434 (DA<sub>1.12</sub> + H<sup>+</sup>)



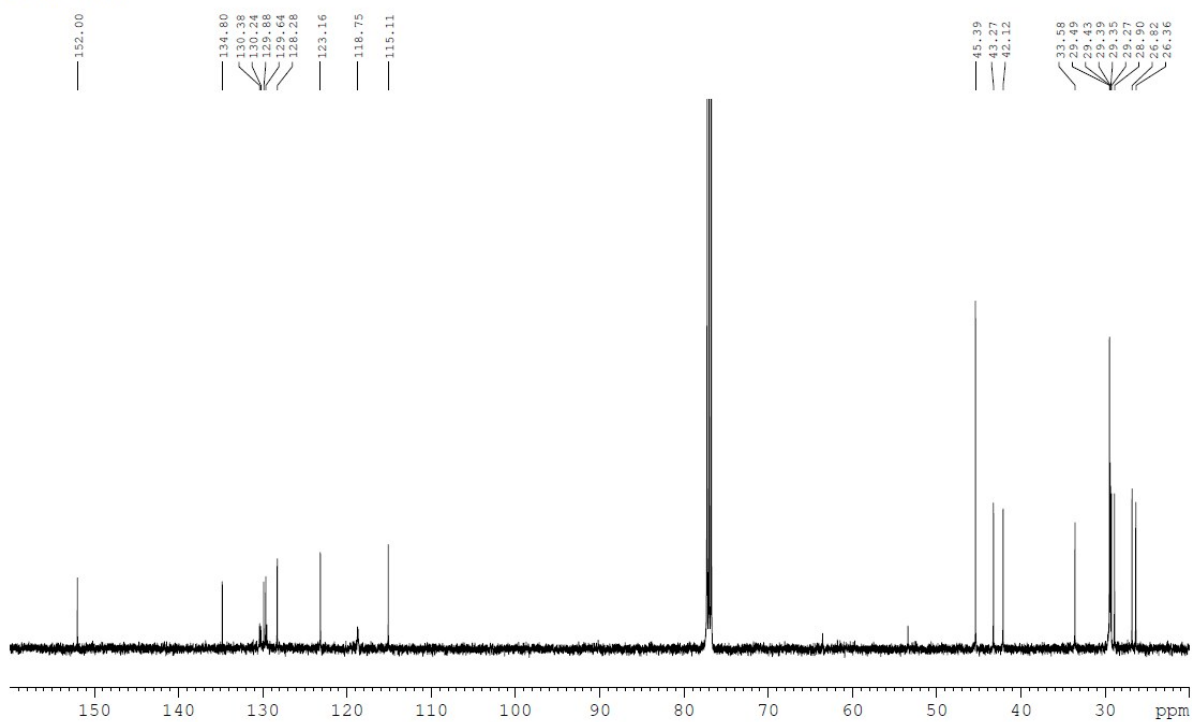
ESI-MS ( $m/z$ ): 432 (DA<sub>1.12</sub> - H<sup>+</sup>)

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
Operator CS  
Registry No. 2193  
User G. Ionita  
Sample Changer Position 13  
Sample name: MAD-12  
@H1-BBOF-34 CDCl3 (D:\UnivBuc\Pionita) ICON-NMR-Lab 5

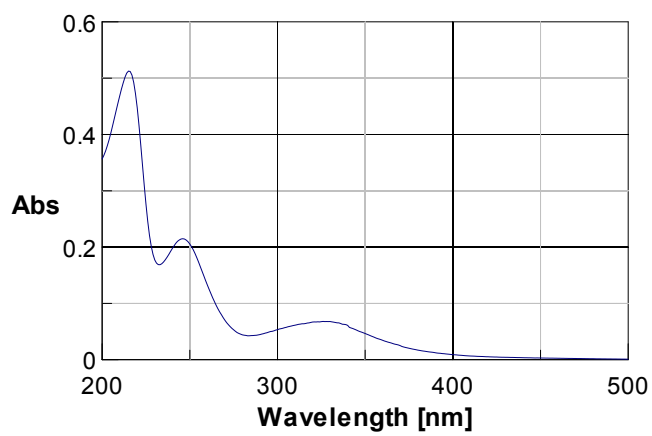


<sup>1</sup>H NMR spectrum of compound DA<sub>1.12</sub>

Instrument Bruker AvanceIII 500MHz (UnivBucuresti)  
Operator CS  
Registry No. 2193  
User G. Ionita  
Sample Changer Position 9  
Sample name: MAD-12  
CDCl3  
2013-09-10 09:28:24



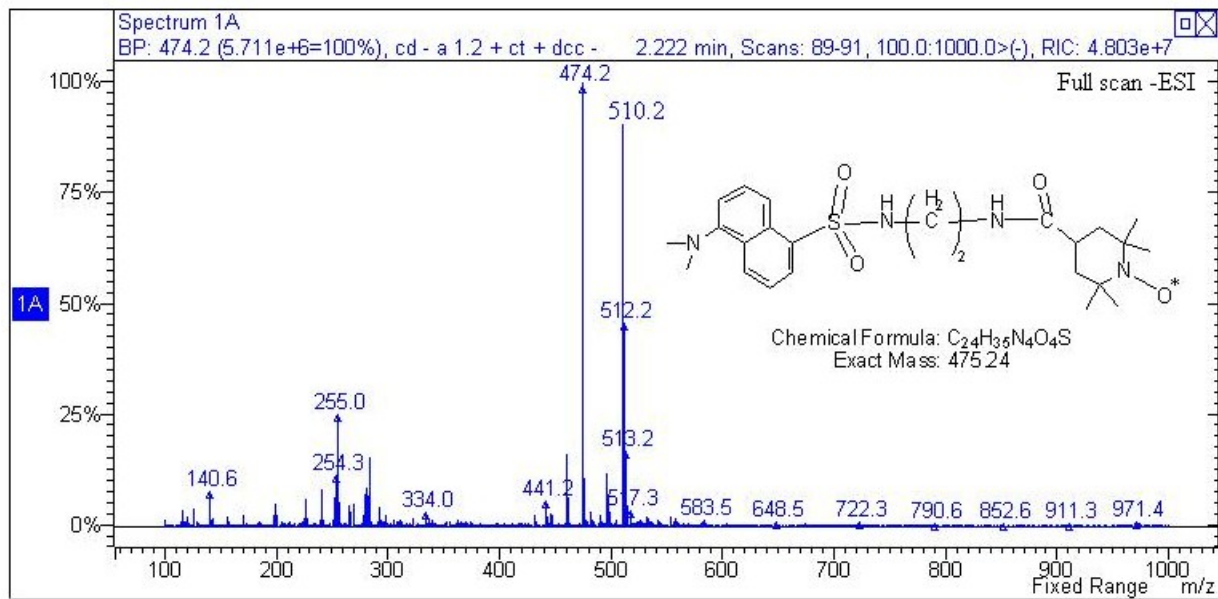
<sup>13</sup>C NMR spectrum of compound DA<sub>1.12</sub>



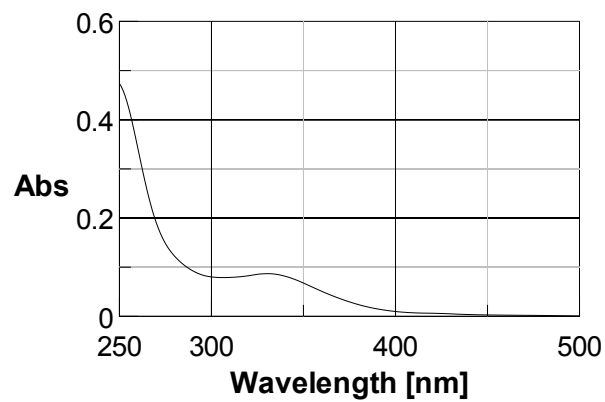
UV-Vis spectrum of compound DA<sub>1.12</sub>

## Spectral characterization of DA<sub>1,n</sub>T

### DA<sub>1,2</sub>T

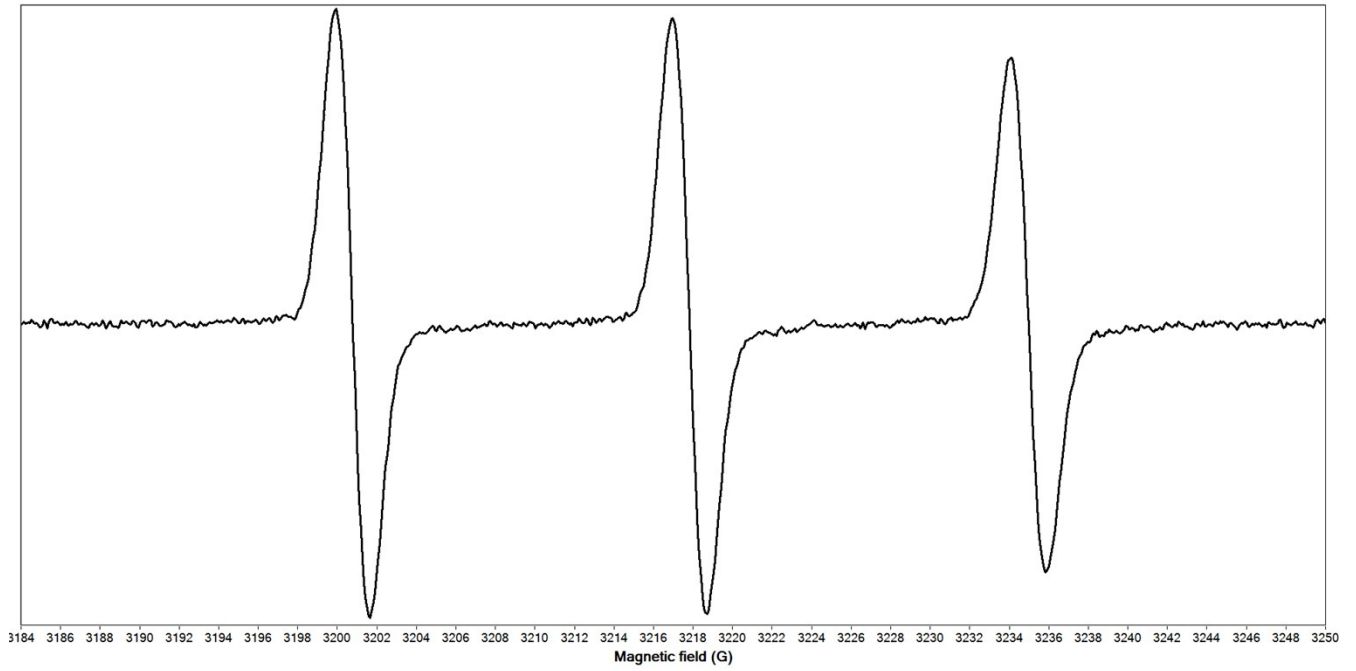


ESI-MS ( $m/z$ ): 474 (DA<sub>1,2</sub>T - H<sup>+</sup>)



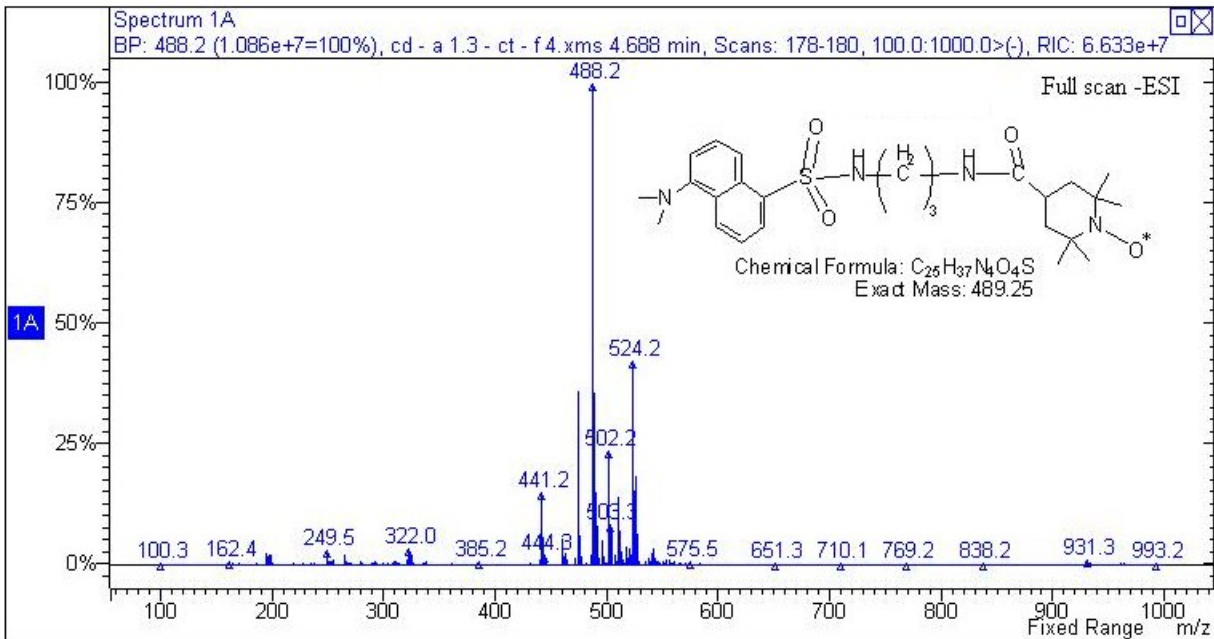
UV-Vis spectrum of compound DA<sub>1,2</sub>T



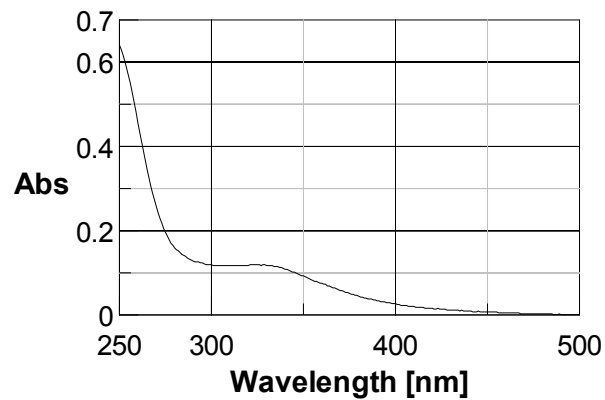


**EPR spectrum of DA<sub>1.2</sub>T**

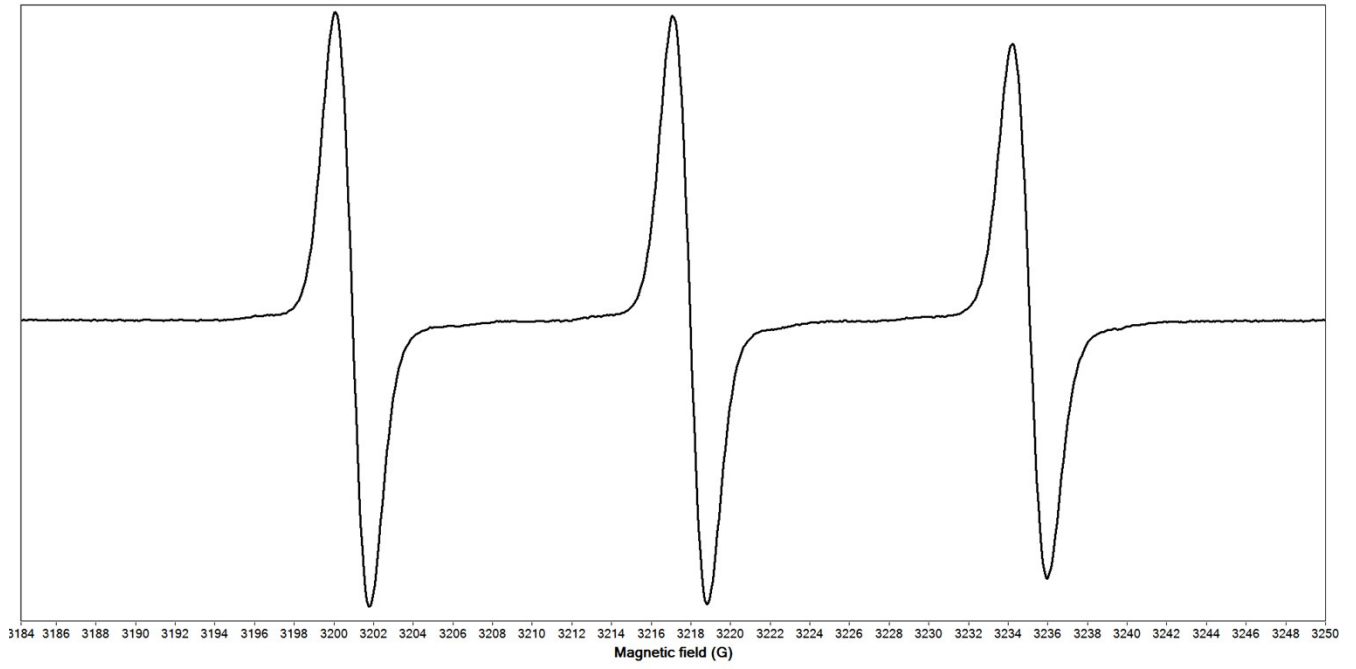
# DA<sub>1.3</sub>T



ESI-MS ( $m/z$ ): 488 (DA<sub>1.3</sub>T - H<sup>+</sup>)

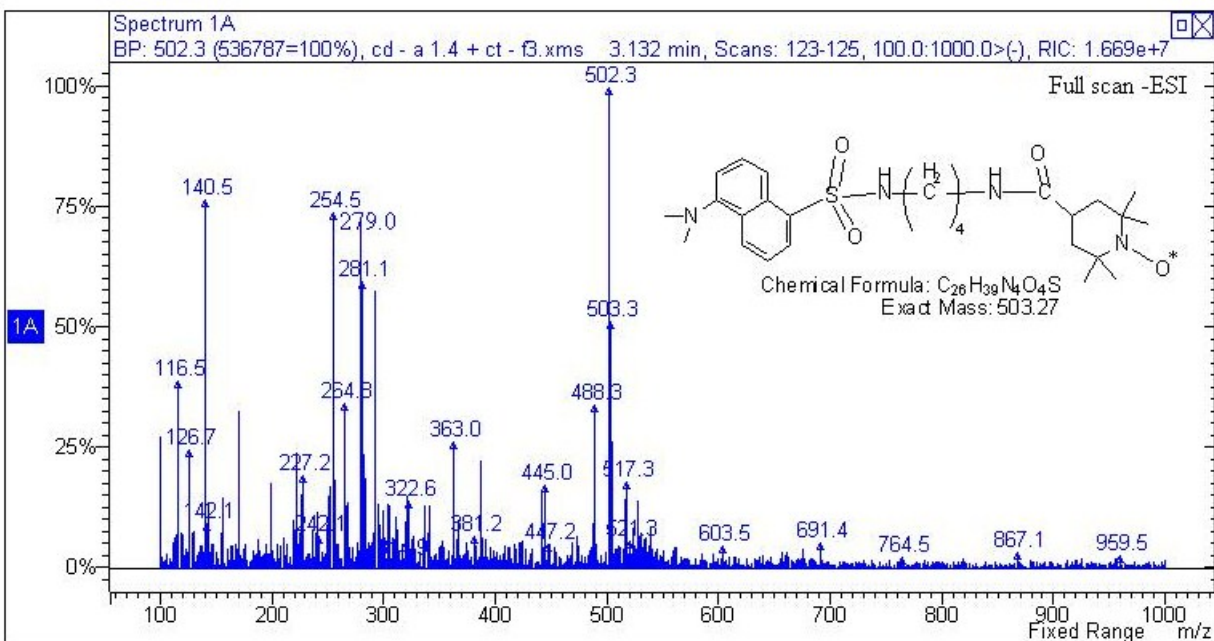


UV-Vis spectrum of compound DA<sub>1.3</sub>T

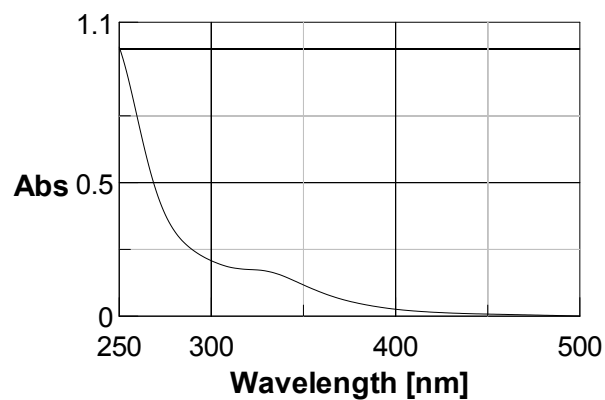


**EPR spectrum of DA<sub>1.3</sub>T**

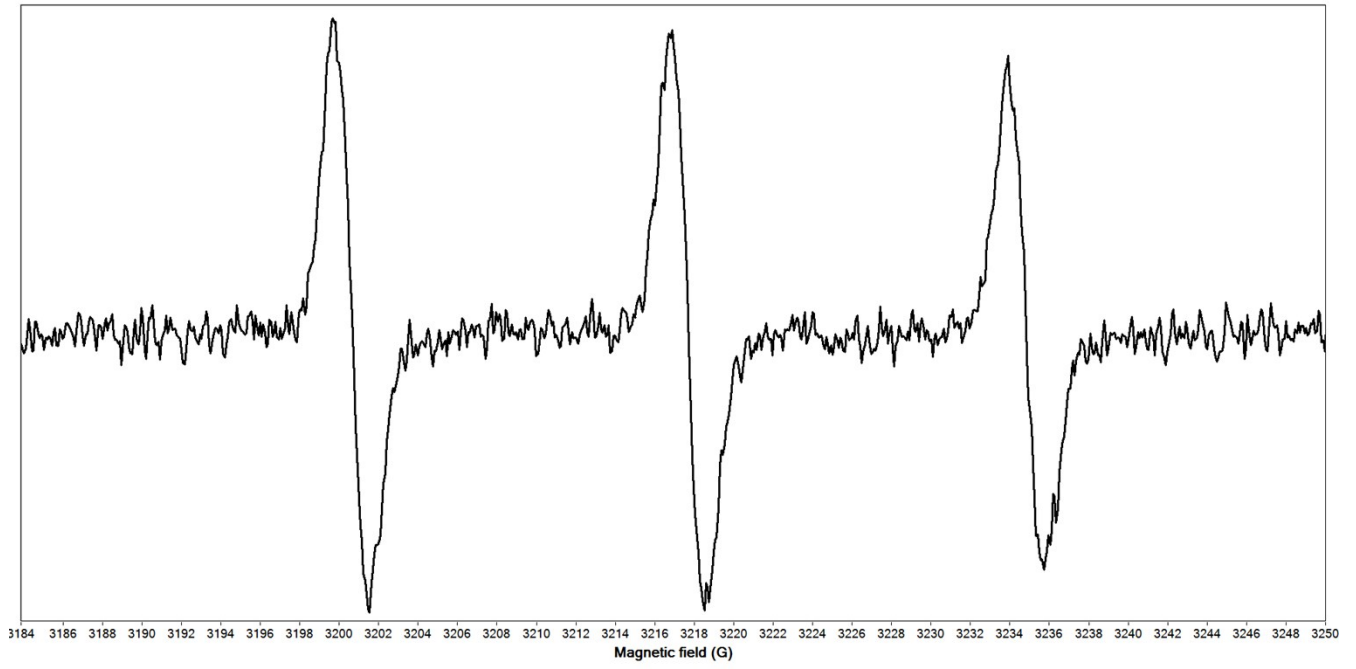
## DA<sub>1.4</sub>T



ESI-MS ( $m/z$ ): 502 (DA<sub>1.4</sub>T - H<sup>+</sup>)

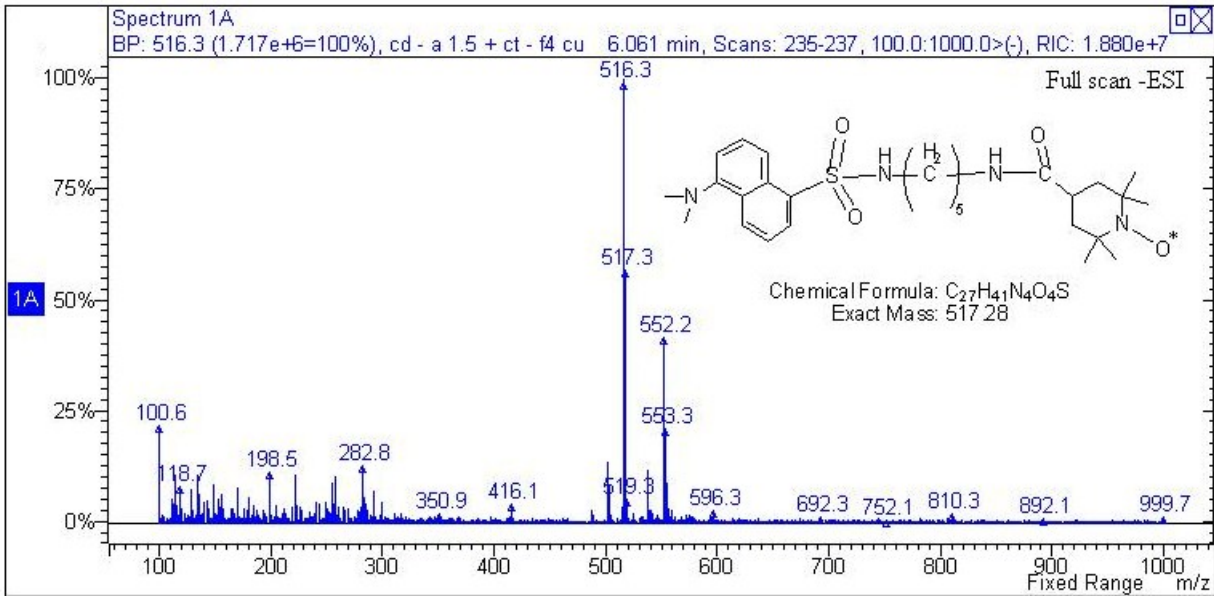


UV-Vis spectrum of compound DA<sub>1.4</sub>T

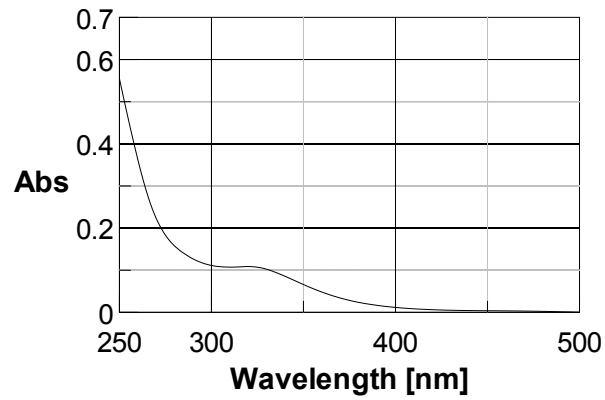


**EPR spectrum of DA<sub>1.4</sub>T**

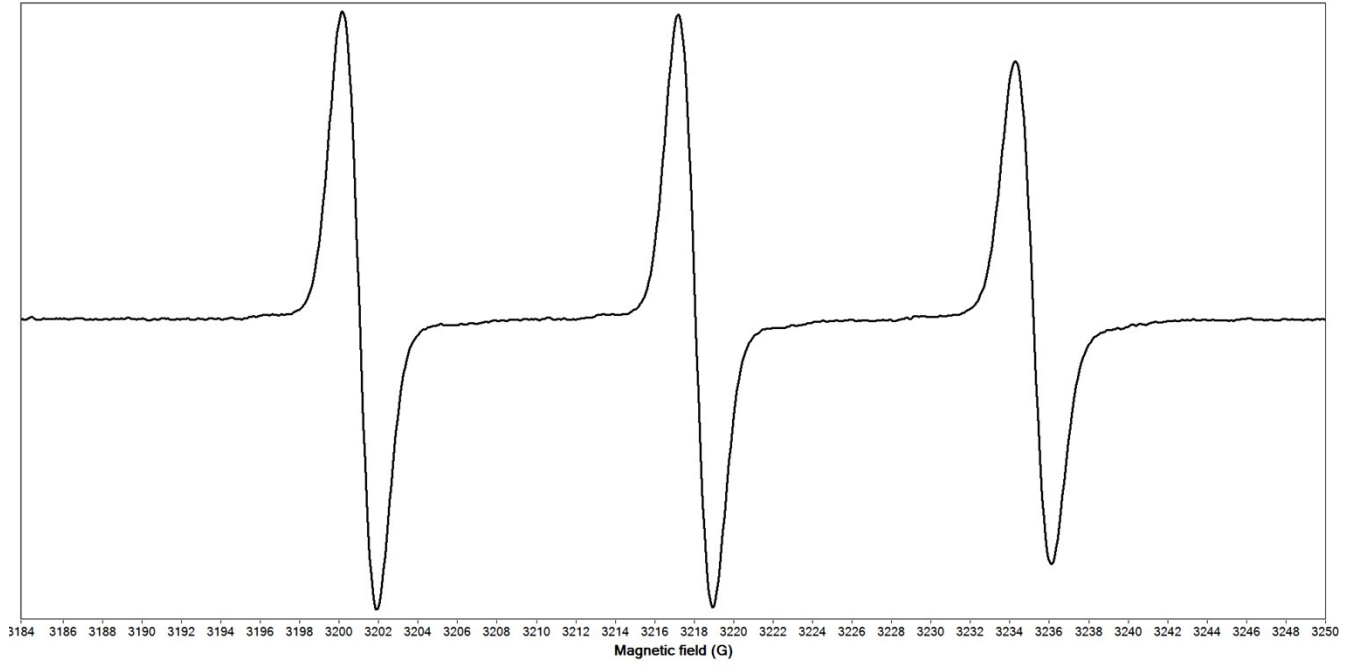
# DA<sub>1.5</sub>T



ESI-MS ( $m/z$ ): 516 (DA<sub>1.5</sub>T - H<sup>+</sup>)

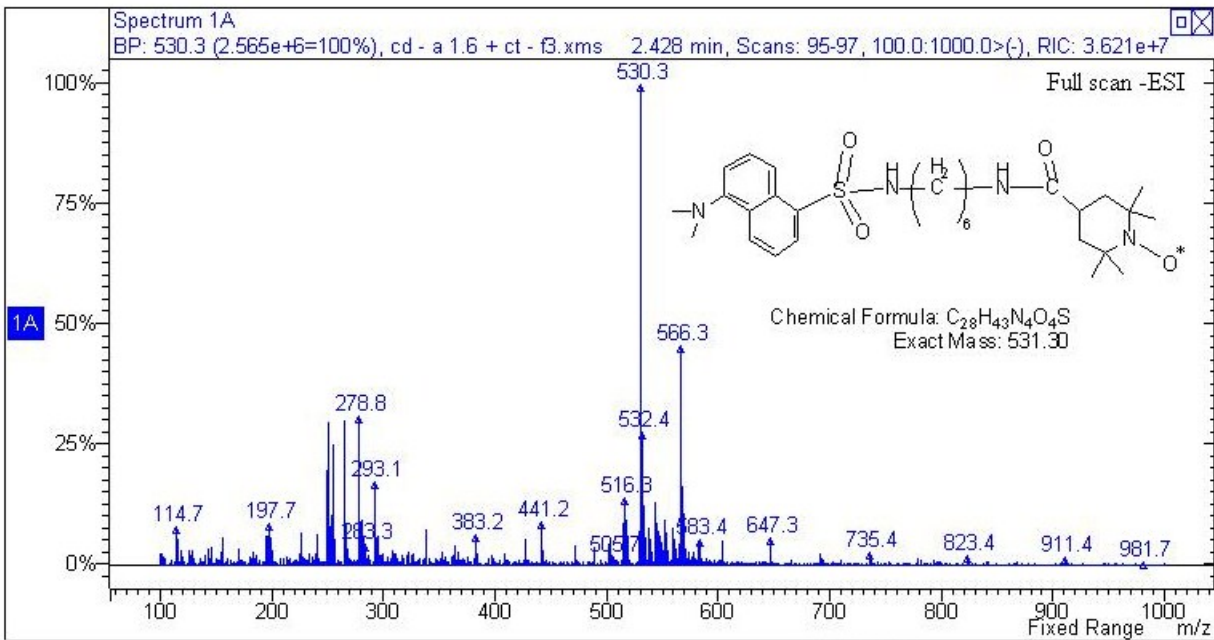


UV-Vis spectrum of compound DA<sub>1.5</sub>T

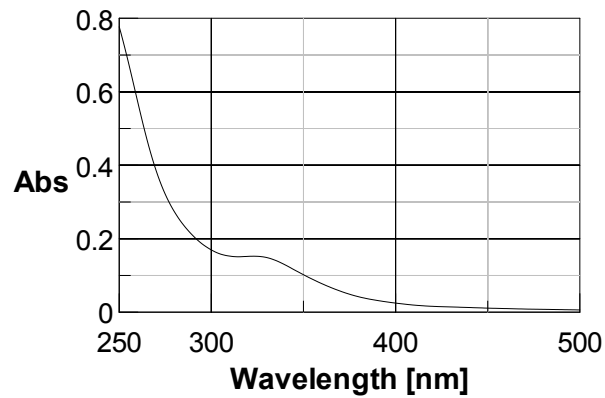


**EPR spectrum of DA<sub>1.5</sub>T**

# DA<sub>1.6</sub>T

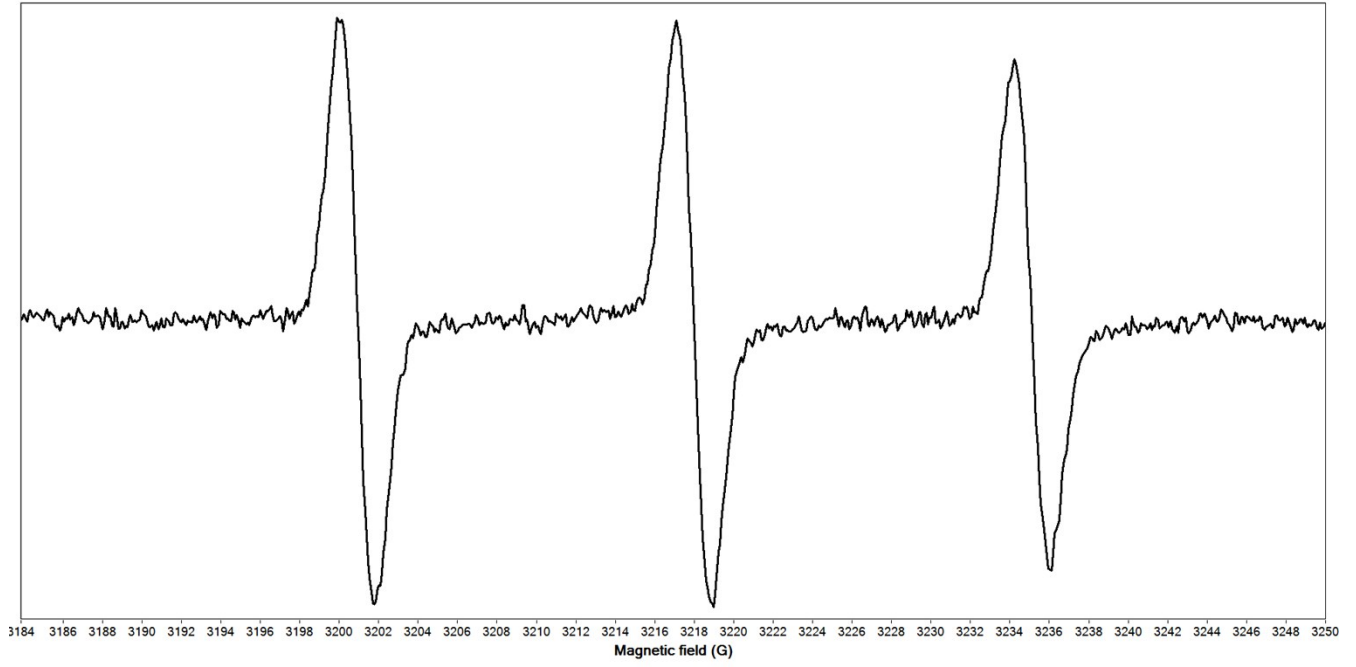


ESI-MS ( $m/z$ ): 530 (DA<sub>1.6</sub>T - H<sup>+</sup>)



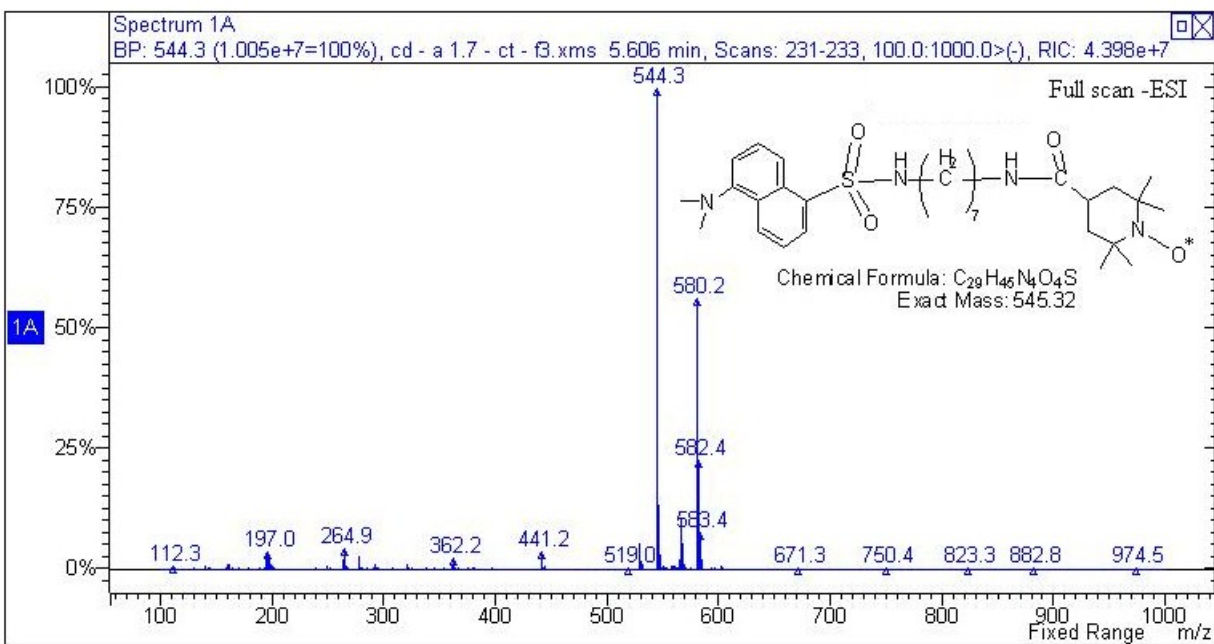
UV-Vis spectrum of compound DA<sub>1.6</sub>T



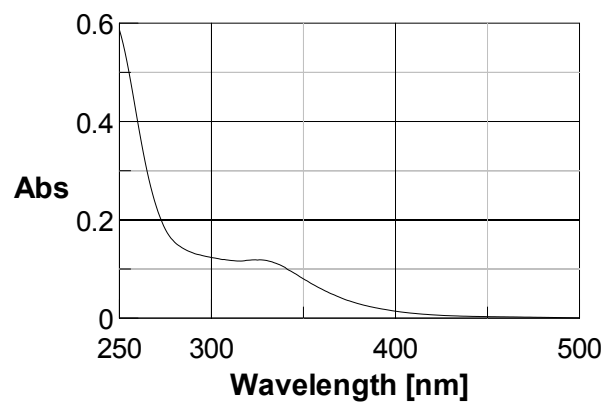


**EPR spectrum of DA<sub>1.6</sub>T**

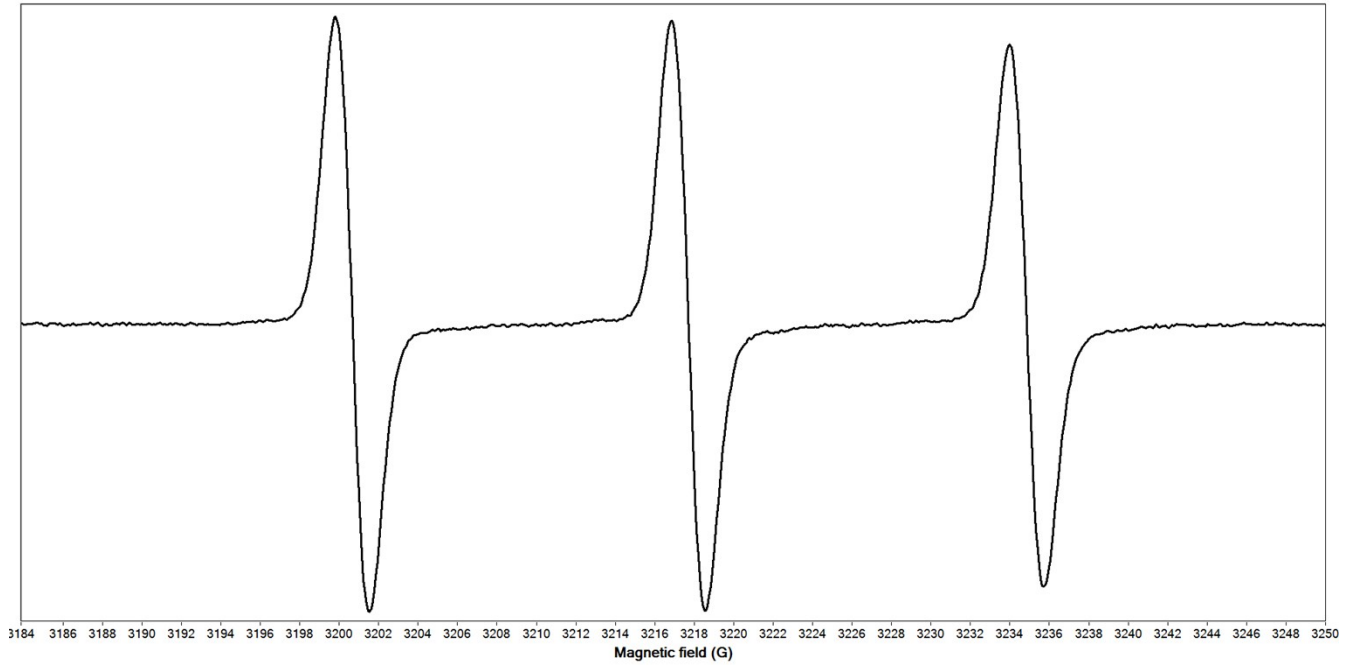
# DA<sub>1.7</sub>T



ESI-MS ( $m/z$ ): 544 (DA<sub>1.7</sub>T - H<sup>+</sup>)

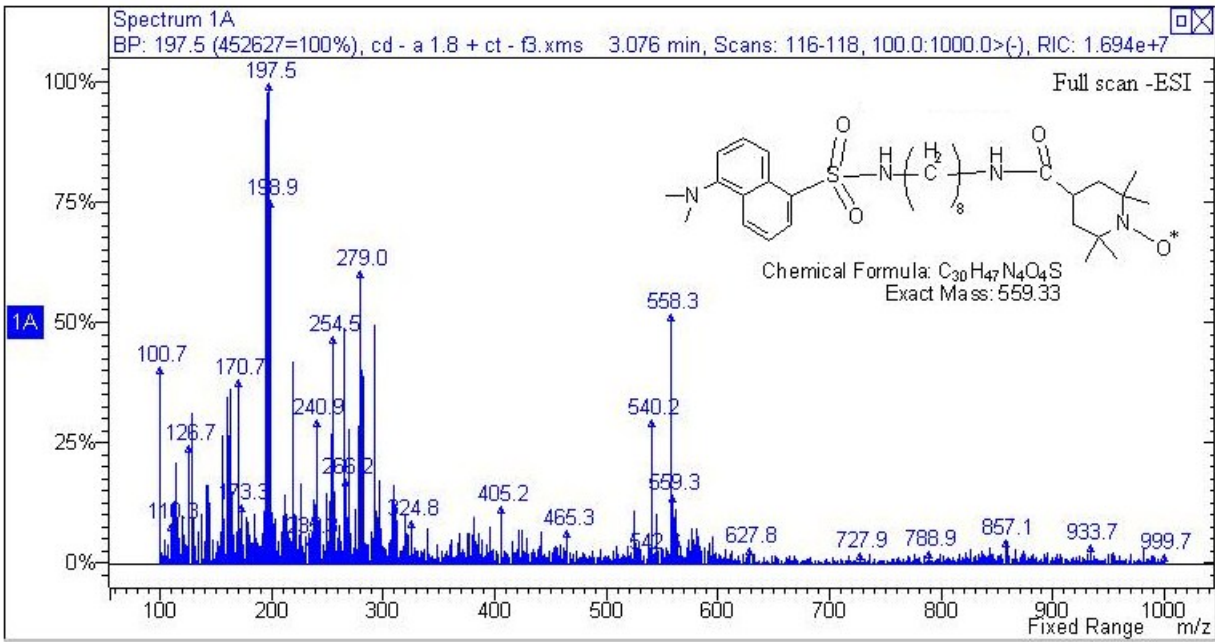


UV-Vis spectrum of compound DA<sub>1.7</sub>T

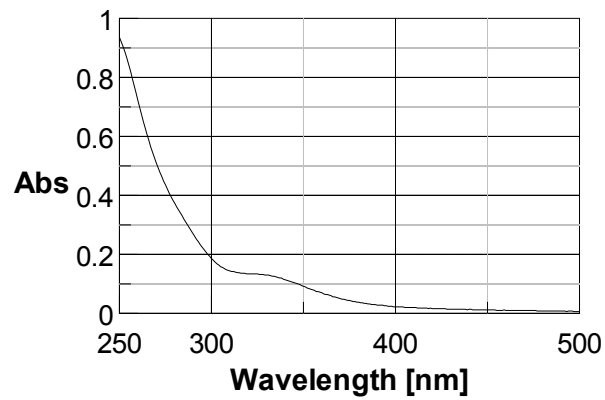


**EPR spectrum of DA<sub>1.7</sub>T**

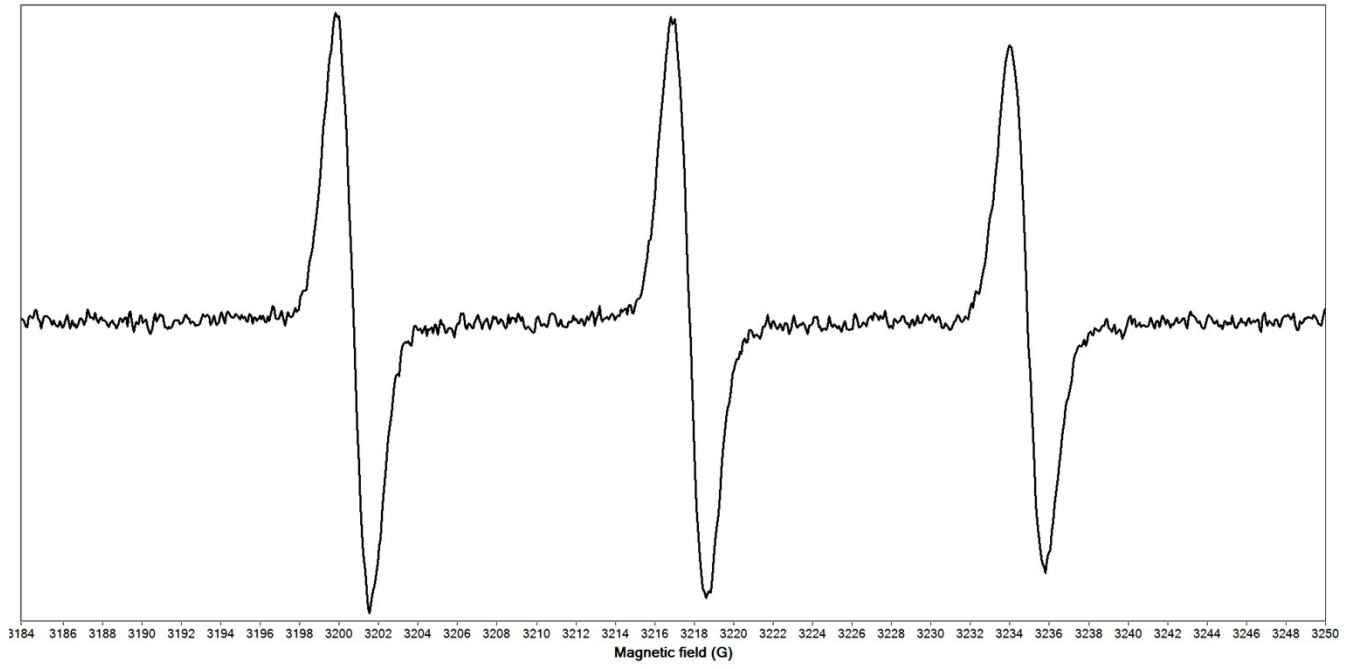
# DA<sub>1.8</sub>T



ESI-MS ( $m/z$ ): 558 (DA<sub>1.8</sub>T - H<sup>+</sup>)

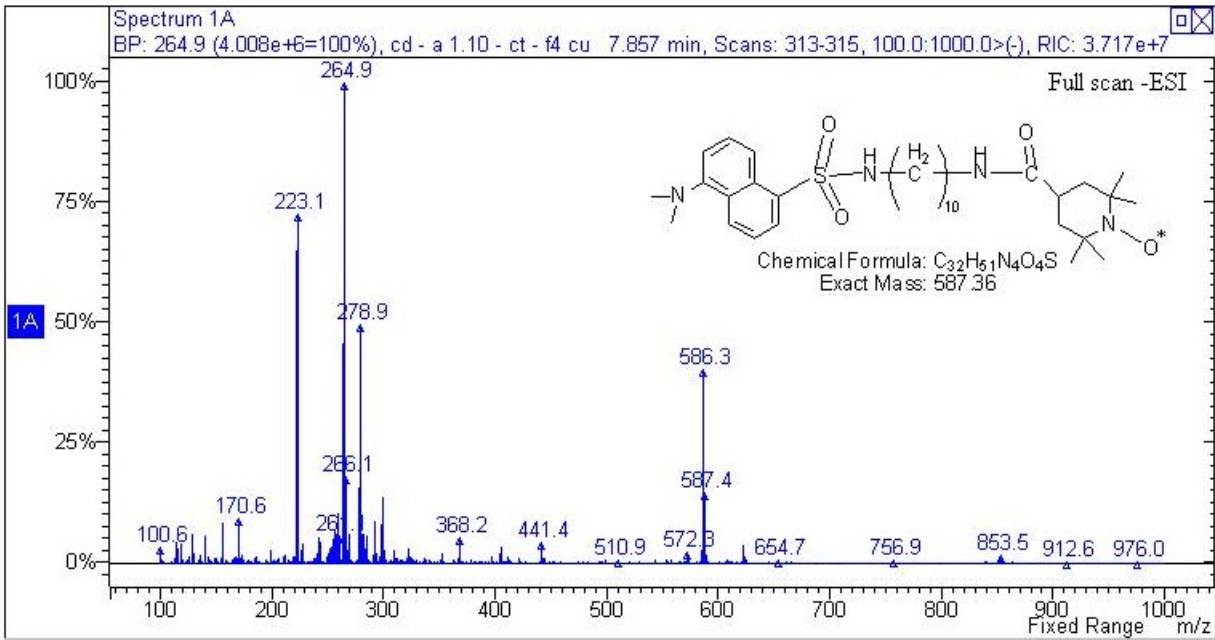


UV-Vis spectrum of compound DA<sub>1.8</sub>T

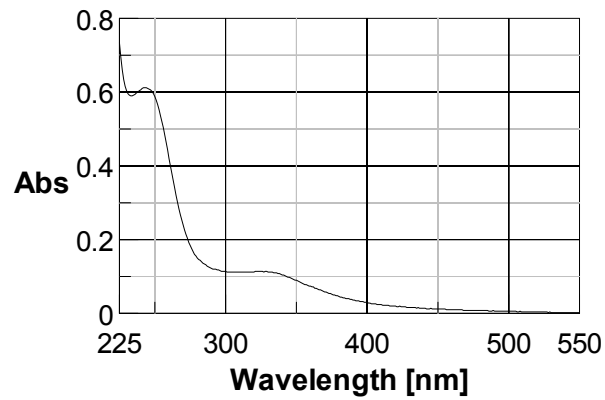


**EPR spectrum of DA<sub>1.8</sub>T**

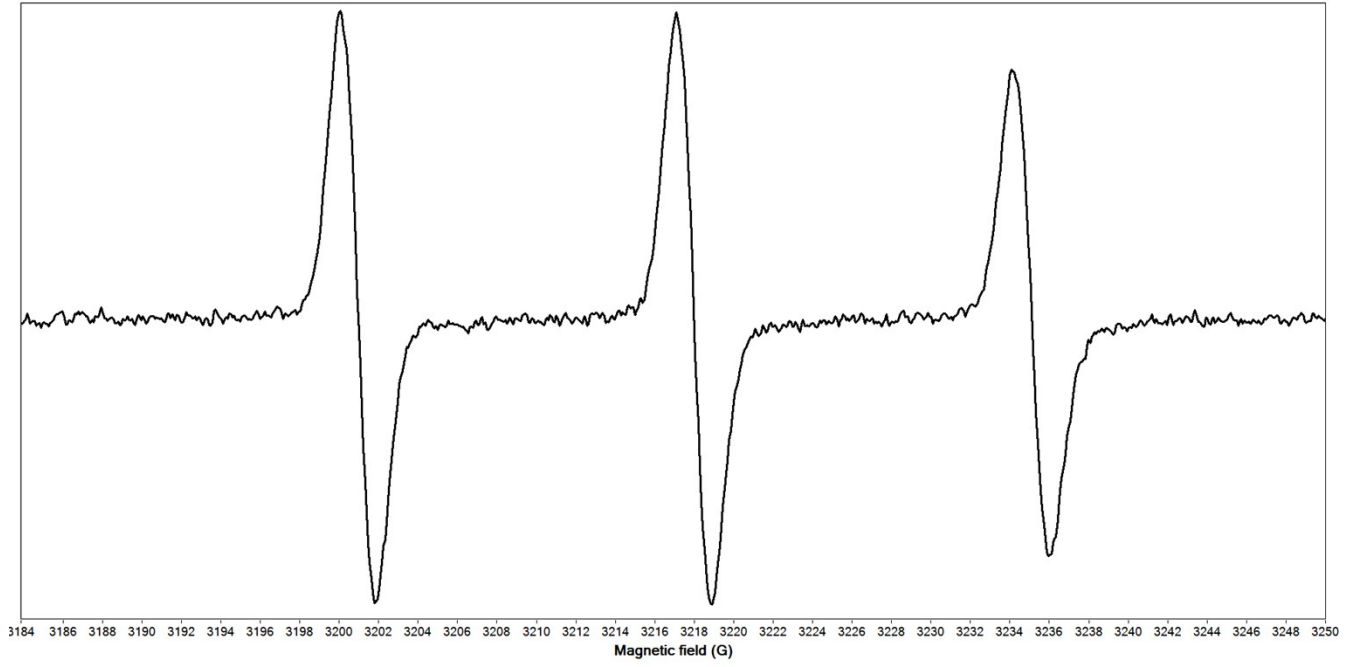
# DA<sub>1.10</sub>T



ESI-MS ( $m/z$ ): 586 (DA<sub>1.10</sub>T - H<sup>+</sup>)

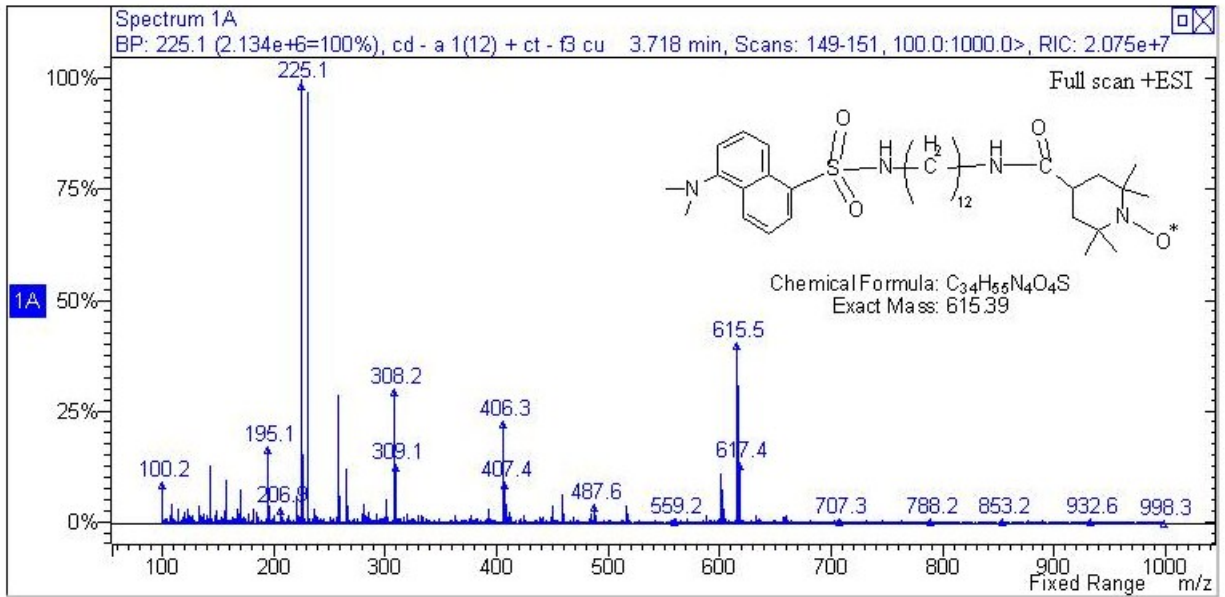


UV-Vis spectrum of compound DA<sub>1.10</sub>T

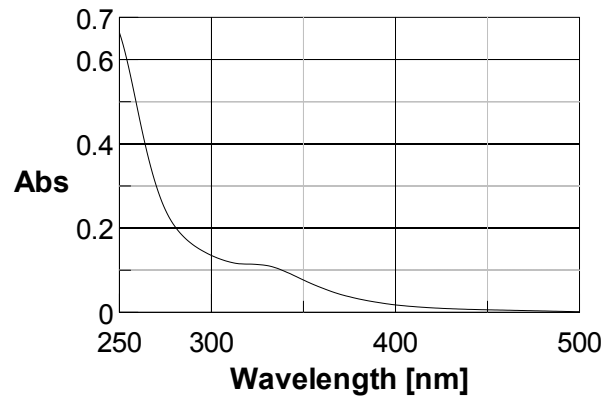


**EPR spectrum of DA<sub>1.10</sub>T**

# DA<sub>1.12</sub>T

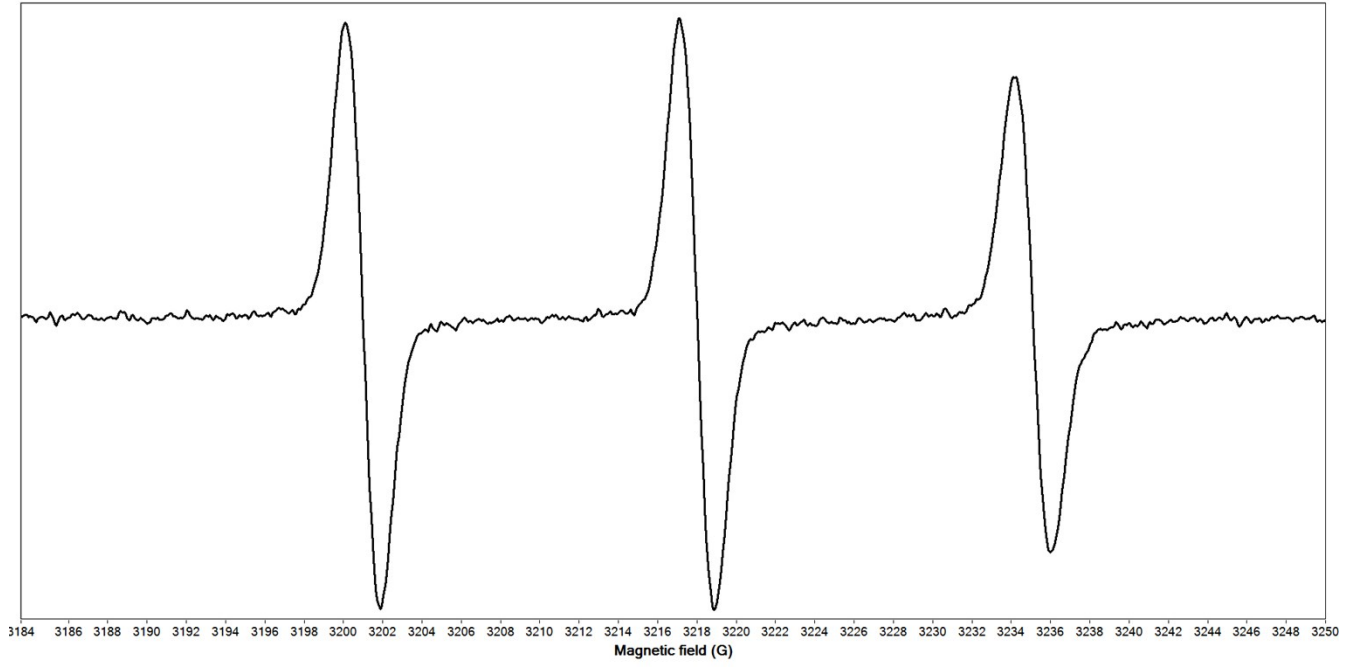


ESI-MS ( $m/z$ ): 615 (DA<sub>1.12</sub>T)



UV-Vis spectrum of compound DA<sub>1.12</sub>T





**EPR spectrum of DA<sub>1.12</sub>T**